

The complete mitogenome of *Helix pomatia* and the basal phylogeny of Helicinae (Gastropoda, Stylommatophora, Helicidae)

Ondřej Korábek¹, Adam Petrusek¹, Michail Rovatsos¹

¹ Department of Ecology, Faculty of Science, Charles University, Viničná 7, CZ-12844 Prague 2, Czechia

Corresponding author: Ondřej Korábek (korabeko@natur.cuni.cz)

Academic editor: E. Neubert | Received 16 January 2019 | Accepted 27 January 2019 | Published 5 March 2019

<http://zoobank.org/D32B999B-FEB7-43CC-A481-E89C3AC76160>

Citation: Korábek O, Petrusek A, Rovatsos M (2019) The complete mitogenome of *Helix pomatia* and the basal phylogeny of Helicinae (Gastropoda, Stylommatophora, Helicidae). ZooKeys 827: 19–30. <https://doi.org/10.3897/zookeys.827.33057>

Abstract

A complete mitochondrial genome of the Roman snail *Helix pomatia* Linnaeus, 1758 has been sequenced. The length and gene order correspond to that of other available helicid mitogenomes. We used the mitogenome sequence to reappraise the relationships among the four presumed principal groups of the helicid subfamily Helicinae. The results support the idea that the subfamily is divided between two western Palaearctic diversification centres: Iberian Peninsula and western Maghreb in the west, and Anatolia, the Aegean and Caucasus in the east. One group, the tribe Helicini, diversified in the east and the remaining three currently recognised tribes in the west. However, the exact relationships among lineages of the non-Helicini tribes could not be resolved.

Keywords

Allognathini, biogeography, Helicini, land snail, Mediterranean, mitochondrial genome, Otalini, phylogeny, Thebini

Introduction

Phylogenetic research on land snails has been thus far dominated by studies based on mitochondrial markers, mostly partial sequences of the *cox1* (COI) and *rrnL* (16S rRNA) genes, not exceeding 2000 bp in total. These genes dominate not only in terms of the amount of sequences generated, but usually also by the variability of the se-

quences used if combined with sequences of nuclear loci. Although modern techniques targeting many loci simultaneously begin to advance also into the phylogenetics of pulmonate land snails (Teasdale et al. 2016), the mitochondrial markers will likely continue to be used not only due to relatively low costs and accessibility of the methods, but especially thanks to the wealth of previously published data. Mitochondrial genome sequences may then become handy for primer design, to facilitate assembly of further mitochondrial genomes from outputs of high-throughput sequencing, or directly as the data for phylogenetic analyses. In fact, one of the best resolved backbone phylogenies of Stylommatophora to date was reconstructed from mitochondrial genome sequences (Groenenberg et al. 2017).

We sequenced a complete mitogenome of *Helix pomatia* Linnaeus, 1758, a large and common European edible land snail species, to provide a reference for further work focused on the family Helicidae Rafinesque, 1815. We then used other helicid mitogenomic sequences to evaluate the support for the basal branching order in the subfamily Helicinae, which was reported by Neiber and Hausdorf (2015) from analyses based on partial sequences of *rrnL* and *cox1* genes along with part of the nuclear rRNA gene cluster (5.8S rRNA, internal transcribed spacer 2, 28S rRNA).

The subfamily Helicinae includes several charismatic species, which are large, common, and edible. As the most prominent, we may mention *Cepaea nemoralis* (Linnaeus, 1758), famous for evolutionary studies of its colour polymorphism (e.g. Cain and Sheppard 1950, Jones et al. 1977, Silvertown et al. 2011), *Cornu aspersum* (Müller, 1774), the most commonly reared and consumed helicid species which became a pest in several regions around the globe (e.g. Rees 1955, Ansart et al. 2009), and *Helix pomatia*, a characteristic snail of Central Europe and the type species of the family's type genus. Each of these three species represents a different lineage of the three presumed clades constituting the Helicinae, which were proposed and assigned a tribe status in the first comprehensive molecular phylogenetic study of Helicoidea by Razkin et al. (2015). *Cepaea* represents Allognathini Westerlund, 1902, *Helix* Helicini Rafinesque, 1815, and *Cornu* Otalini Pfeffer, 1930. Razkin et al. (2015) also recognized a fourth tribe, the monotypic Thebini Wenz, 1923 represented by *Theba* Risso, 1826. The phylogenetic position of *Theba* has been contentious (Perrot 1939, Gittenberger and Ripken 1987, Bouchet and Rocroi 2005, Schileyko 2013, Razkin et al. 2015), but *Helix*, *Cornu* and *Cepaea* were generally considered closer to each other than to *Theba*, for which a separate tribe or even subfamily has been erected.

Neiber and Hausdorf (2015) used a more informative dataset than Razkin et al. (2015) and found *Cepaea*, *Cornu* and *Theba* to be more closely related to each other than to *Helix* and some other eastern Mediterranean genera. Their results suggest a principal biogeographic division within Helicinae, running approximately along the Apennine Peninsula and separating two equally old clades with centre of diversification in the western (*Cornu*, *Cepaea* and *Theba*, with their respective relatives) and the eastern (*Helix* and relatives; Korábek et al. 2015) Mediterranean basin, respectively. Considering the Paleogene/Neogene palaeogeographic evolution of the western Mediterranean (Giusti and Manganello 1984, Rosenbaum et al. 2002, Advokaat et al. 2014, van Hinsbergen et al. 2014), it may be expected that the western taxa are indeed more closely related to each other.

Alternatively, the eastern group may be an offshoot of the western diversity, if the western group is paraphyletic. The systematic concept and monophyly of *Otulini* remains an open issue, as they (in the sense of Neiber and Hausdorf 2015) may be paraphyletic to both *Thebini* and *Allognathini*, and the east-west divide has not been unambiguously supported. The absence of an east-west divide in the phylogeny would lend support to a Western European origin of the whole subfamily *Helicinae* and to the north-western Maghreb being a refugium of its oldest and perhaps phylogenetically most distinctive lineages. Apart from biogeographic implications, the branching order at the base of the *Helicinae* has for us also a practical significance for the selection of appropriate outgroup taxa for analyses of relationships between *Helix* and related genera.

There are already published complete mitochondrial genome sequences of *Ce. nemoralis* (Terrett et al. 1996; NCBI accession number NC_001816), *Co. apersum* (Gaitán-Espitia et al. 2013; NCBI accession number NC_021747) and *Theba pisana* (Müller, 1774) (Wang et al. 2018b; Genbank accession number MH362760), as well as of a convenient outgroup species *Cylindrus obtusus* (Draparnaud, 1805) (Groenenberg et al. 2012; NC_017872). Combined with the newly sequenced mitogenome of *Helix pomatia* they offered an opportunity to verify the relationships between the four *Helicinae* tribes.

Methods

Sequencing and annotation

As a starting point we utilised mitochondrial sequences obtained from transcriptome sequencing (mRNA-Seq) performed for a different project, which included also a single *H. pomatia* individual (from Rožmitál pod Třemšínem, Czechia). In detail, 20 mg of foot tissue was homogenised with MagNA Lyser (Roche) and total RNA was extracted using the standard Trizol reagent protocol (Thermo Fisher Scientific). The barcoded and stranded mRNA-sequencing libraries were prepared using the Illumina TruSeq mRNA v2 sample preparation kit (Illumina, San Diego, CA, USA). The libraries were loaded on an Illumina NextSeq 500 sequencer and 75 bp were sequenced uni-directionally, resulting in approx. 87 million reads. The raw Illumina reads were trimmed for adapters and low-quality bases in GENEIOUS R7.1 (Kearse et al. 2012) with the trim utility using default parameters. Reads with a length of less than 50 bp were removed from the dataset. The trimmed reads were checked for quality in FASTQC (Andrews 2010) and MULTIQC (Ewels et al. 2016), and mapped with GENEIOUS R7.1 to the mitogenome of *Co. aspersum*. The partial sequences of 12 protein coding genes (all but *atp8*) of *H. pomatia* (GenBank acc. nos. MK400678-MK400689) were extracted from the alignment and were used to design primers for PCR amplification of the mitogenome of *H. pomatia* in few long overlapping fragments.

For amplification we used another individual sharing an identical *cox1* sequence with the transcriptome data. The sample originated from Huldessen, Bavaria, Germany (48.3978N, 12.7084E) and the shell voucher is deposited in the National Museum,

Prague, lot P6M 29637. We designed specific primers with Primer BLAST (Ye et al. 2012) based on the transcriptome sequences and previous 16S data. Then, we amplified the mitogenome in several overlapping segments of ca. 4000–6000 bp with Platinum SuperFi proof-reading DNA polymerase (Invitrogen, Carlsbad, CA, USA) and sequenced ends of the fragments using the primers used for PCR. Resulting gaps were iteratively filled in further rounds of sequencing using new individual-specific primers flanking the gaps. The reads were aligned to the *Co. aspersum* sequence as reference and combined into a single 14070 bp contig. The sequence is available at GenBank (MK347426).

We used MITOS (Bernt et al. 2013b) to annotate the genome sequence, which identified the full set of the expected 2 rRNA, 22 tRNA, and 13 protein-coding genes (Table 1). However, we made manual adjustments to the MITOS output regarding the limits of rRNA and protein-coding genes, which were inconsistent between the MITOS annotation and the *Ce. nemoralis*, *Co. aspersum*, and *Cy. obtusus* RefSeq entries. To be more specific, we looked for start- and stop-codons whose positions would be compatible between *H. pomatia* and the other three species, and considered also the positions of adjacent genes. However, as gene overlap and alternative start- and incomplete stop-codons may occur in invertebrate mitogenomes (Bernt et al. 2013a) including land snails (Gaitán-Espitia et al. 2013), the start and end positions of the genes remain tentative until sequences of the coded proteins and rRNAs are known. The 16S gene was not recovered in one piece by MITOS, probably because the structure towards the 3' end was too variable; so we combined the two fragments.

Phylogenetic analyses

For the phylogenetic analysis we used a concatenated alignment of 12 protein-coding genes, excluding the short and fast evolving *atp8*. From annotated, previously published sequences (accession numbers NC_001816, NC_021747 and NC_017872), we extracted the individual genes following the annotation in the NCBI RefSeq database. However, previous studies, which used the sequence of *Ce. nemoralis* in mitogenomic phylogenetic analyses, repeatedly resulted in a spuriously long branch for this species (Knudsen et al. 2006, González et al. 2016, Minton et al. 2016, Groenenberg et al. 2017). A closer examination of the mitogenome sequence and translation of its protein coding regions revealed a very low quality of the sequence (see also White et al. 2011). Compared to newer *Ce. nemoralis* sequences there seemed to be frequent indels and suspect amino-acid changes were found in translated sequences. The very high number of errors in the sequence probably stems from the complex way it was obtained, which involved two rounds of cloning, and generally the immature methods of that time (Terrett et al. 1996). Therefore, we replaced the sequences of protein-coding genes of *Ce. nemoralis* by data retrieved from a transcriptome shotgun assembly (TSA) (Kerkvliet et al. 2017; NCBI BioProject PRJNA377398; see Table 1). To extract the mitochondrial genes from the transcriptome data, we performed for each gene a BLASTn search limited to *Ce. nemoralis* with the respective part of NC_001816 as the query in the NCBI TSA database. The sequence of *T. pisana* (MH362760) was not

Table 1. NCBI Transcriptome Shotgun Assembly database accession numbers of the *Cepaea nemoralis* sequences used in analysis.

<i>cox1</i>	GFLU01084822
<i>nd6</i>	GFLU01007552
<i>nd5</i>	GFLU01092360
<i>nd1</i>	GFLU01092360
<i>nd4L</i>	GFLU01092360
<i>cyb</i>	GFLU01076837
<i>cox2</i>	GFLU01076837
<i>atp6</i>	GFLU01122686
<i>nd3</i>	GFLU01014131
<i>cox3</i>	GFLU01122685
<i>nd4</i>	concatenated from GFLU01122684, GFLU01122687, GFLU01122688

annotated, so we identified the individual genes by alignment with the extracted gene sequences of the other four species.

We performed the phylogenetic analyses in three stages, starting with the analysis of the nucleotide sequences. The protein coding genes were aligned with TRANSLATORX (Abascal et al. 2010), which aligns nucleotide sequences based on their amino acid translations. TRANSLATORX was run with the MAFFT 5 aligner (Kato et al. 2005) and the invertebrate mitochondrial genetic code; other settings were left at their defaults. The resulting alignments were trimmed to a common length of the sequences. We specified a partitioning scheme consisting of the three codon positions and used IQTREE 1.6.5 to test the scheme against simpler scenarios, select substitution models (Kalyaanamoorthy et al. 2017) and to perform a maximum likelihood phylogenetic analysis (Nguyen et al. 2015, Chernomor et al. 2016). The model selection suggested the TIM2+F+I+Γ4, TVM+F+Γ4, and TIM2+F+Γ4 models for the three partitions. We used standard bootstrap and Shimodaira-Hasegawa-like approximate likelihood ratio tests (SH-aLRT; Guindon et al. 2010; both with 1000 replicates) to calculate branch supports. Then we translated the alignment into amino acid sequences, leaving it a single partition with 417 parsimony informative sites out of the total of 3489 sites, and run the model selection (selecting mtZoa+F+Γ4 as the best fit substitution model; Rota-Stabelli et al. 2009) and phylogenetic analysis as above.

The resulting topologies showed substantial differences in branch lengths between taxa. To explore if the resulting topology was influenced by long-branch attraction (LBA), we employed a site-heterogeneous mixture model using posterior mean site frequencies (PMSF; Wang et al. 2018a). This approach has been shown to effectively overcome LBA, and even to be somewhat prone to long-branch repulsion. Thus, if a grouping is due to LBA, this approach is likely to reveal that. We calculated the PMSF under the mtZoa+C60+F+Γ4 mixture model. As a guide tree we used successively all the three alternative topologies in the western group: *Cornu+(Theba+Cepaea)*, *(Cornu+Theba)+Cepaea*, and *(Cornu+Cepaea)+Theba*.

Finally, to account for differences in nucleotide composition, we performed a Bayesian phylogenetic analysis of the nucleotide data with a branch-heterogeneous model al-

lowing branches to have distinct compositions as implemented in P4 1.0 (Foster 2004; <http://p4.nhm.ac.uk/>). We performed the analysis in two runs of 1,000,000 generations with four heated chains each, using the same partitioning scheme as with IQTREE and the closest possible substitution models. For each data partition we allowed for three different nucleotide compositions to be assigned to tree branches randomly. We sampled each 1000th generation and discarded the first 20% of samples of each run as burn-in. To account for differences in results caused by the use of different inference algorithms in the methods, we repeated the analysis in P4 also assuming composition homogeneity.

Table 2. Annotation of the mitogenome of *Helix pomatia*. The plus/minus strand refers to the position of the *cox1* gene.

Gene	Start position	Start codon	End position	Stop codon	Strand
<i>cox1</i>	1	ATG	1530	TAA	plus
<i>trnV</i>	1527		1589		plus
<i>rrnL (16S)</i>	1657		2555		plus
<i>trnL1</i>	2566		2626		plus
<i>trnA</i>	2630		2691		plus
<i>nd6</i>	2692	TTG	3180	TAA	plus
<i>trnP</i>	3177		3239		plus
<i>nd5</i>	3285	ATG	4955	TAA	plus
<i>nd1</i>	4970	ATA	5842	TAG	plus
<i>nd4L</i>	5842	GTG	6118	T--	plus
<i>cyb</i>	6119	ATG	7190	T--	plus
<i>trnD</i>	7234		7285		plus
<i>trnC</i>	7286		7345		plus
<i>trnF</i>	7346		7404		plus
<i>cox2</i>	7405	ATG	8071	T--	plus
<i>trnY</i>	8073		8119		plus
<i>trnW</i>	8121		8180		plus
<i>trnG</i>	8186		8233		plus
<i>trnH</i>	8237		8296		plus
<i>trnQ</i>	8296		8355		minus
<i>trnL2</i>	8356		8411		minus
<i>atp8</i>	8405	ATA	8578	TAG	minus
<i>trnN</i>	8579		8638		minus
<i>atp6</i>	8637	ATG	9290	T--	minus
<i>trnR</i>	9291		9349		minus
<i>trnE</i>	9349		9411		minus
<i>rrnS (12S)</i>	9479		10173		minus
<i>trnM</i>	10173		10235		minus
<i>nd3</i>	10241	TTG	10581	T--	minus
<i>trnS2</i>	10579		10629		minus
<i>trnT</i>	10632		10693		minus
<i>cox3</i>	10693	ATG	11472	TAA	minus
<i>trnS1</i>	11664		11719		plus
<i>nd4</i>	11720	ATA	13027	TAA	plus
<i>trnI</i>	13030		13090		plus
<i>nd2</i>	13091	GTG	14021	T--	plus
<i>trnK</i>	14022		14070		plus

Results

We have sequenced a complete mitogenome of *H. pomatia*, from a specimen representative of a common central-European lineage (Korábek et al. 2018) of this broadly distributed snail species. The length and gene content and order (Table 2) correspond with those of mitogenomes of *Ce. nemoralis*, *T. pisana* and *Co. aspersum*, the other Helicinae species with available mitogenome sequences.

The phylogenetic analyses in all cases recovered the expected split between the eastern *Helix* and the western group of *Theba*, *Cepaea*, *Cornu*, assuming *Cylindrus* as an outgroup (Figure 1). This grouping received equivocal support in the analysis based on the nucleotide data, but it increased with amino acid data and further when using site-heterogeneous model. The support thus increased as the analyses got less sensitive to substitution saturation and among site heterogeneity. In the western group, the results united *Theba* and *Cepaea* to the exclusion of *Cornu*, but this branch received positive support only from bootstrap with the nucleotide data. Especially analyses of the amino-acid alignment with the site-heterogeneous model based on alternative guide trees resulted in very low support for the branch uniting *Cepaea* and *Theba*. The Bayesian analysis of nucleotide data with P4 recovered the same topology as the maximum likelihood analysis but with full support, regardless whether homo- or heterogeneous model was used.

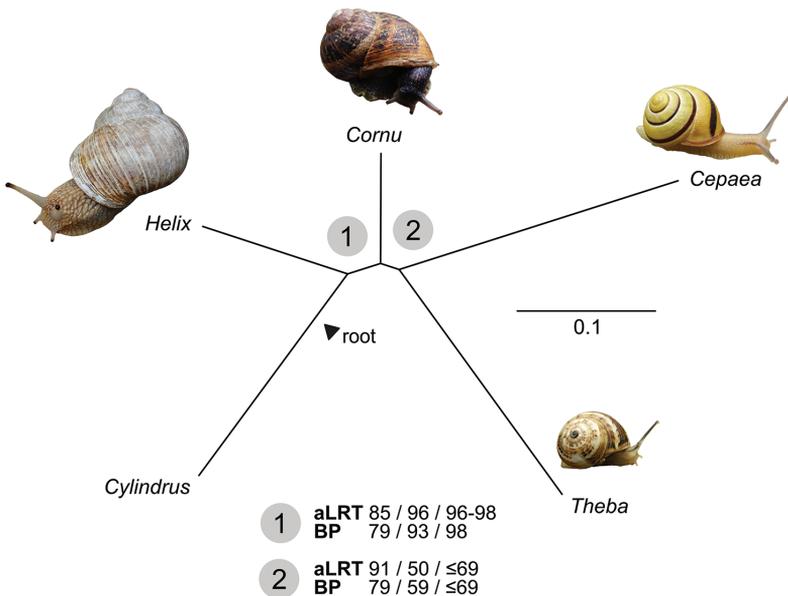


Figure 1. The inferred phylogenetic relationships between the five helicid mitogenomes available. Branch supports are given for maximum likelihood analyses based on nucleotide data, amino acid data under homogeneous model, and amino acid data under site-heterogeneous model (in this order). The latter was run under alternative settings, see details in the text. Shimodaira–Hasegawa-like approximate likelihood ratio test (SH-aLRT) and standard bootstrap percentages (BP) are reported. Values of SH-aLRT >90 % and BP >75 % are considered positive support. *Cylindrus* is included as an outgroup.

Discussion

Because *Theba* and *Cepaea* had the longest branches in the tree, which had short internal branches, and there is no independent indication of the relationship between *Theba* and *Cepaea*, we suspected the result to be caused by the LBA (Bergsten 2005). *Theba* is annual to biannual and semelparous (Heller 1982, Cowie 1984), unlike the other helicids, which are more long-lived (Pollard 1975, Bisenberger et al. 1999; Ansart et al. 2009). The genus may generally cause problems in phylogenetic inference if the difference in life history results in difference in substitution rate (Thomas et al. 2010; cf. Saclier et al. 2018). In addition, analysis of the nucleotide composition of the five taxa revealed substantial differences, where *Cornu* (GC 30.5%) and *Cepaea* (GC 41.6%) differed the most, which could obscure potential relationship between the two species. The low support with presumably more robust methods suggests that the relationship between *Cepaea* and *Theba* could be an artefact.

Our results are consistent with the hypothesis that Helicinae are principally divided into a western (mainly Iberia, western Maghreb, Macaronesia) and an eastern (Caucasus, Anatolia, Greece) group (Neiber and Hausdorf 2015). The split between these two lineages probably occurred no later than during the Late Eocene–Early Oligocene (Neiber and Hausdorf 2015), but the east-west pattern in the distribution of the two lineages persists despite 30 million years having elapsed since the split. Only two *Helix* species naturally represent the eastern group west of ca 9°E (one in Europe, one in Africa; Neubert 2014, Korábek et al. 2018), and two species of *Cepaea* (Europe; Welter-Schultes 2012) and few species of *Eremina* Pfeiffer, 1855 (northern Africa; Ali et al. 2016) represent the western group east of 18°E.

Despite analysing a substantially higher number of genes than Razkin et al. (2015) and Neiber and Hausdorf (2015), we also could not resolve the relationships between *Cornu*, *Cepaea* and *Theba*. We assume that additional lineages should be analysed in order to resolve all the major phylogenetic and biogeographic problems within the western branch of Helicinae. These include *Macularia* Albers, 1850, whose phylogenetic position is equally problematic as that of *Theba* (Nordsieck 1987; Schileyko 2006; Neiber and Hausdorf 2015), the closest relative of *Theba* (perhaps *Eremina*, but see Holyoak et al. 2018), and the type genera of the tribes Otalini and Allognathini. Nevertheless, it is likely that only analyses of multiple nuclear loci will yield robust estimates of the basal relationships of Helicinae due to high and lineage-specific evolutionary rates of the mtDNA and the saturation of nucleotide substitutions.

Acknowledgements

We thank Tereza Kosová for help in the lab, Lucie Juříčková for financial support, and Marco T. Neiber for review and language polishing. Access to computing and storage facilities owned by parties and projects contributing to the National Grid Infrastructure MetaCentrum provided under the programme “Projects of Large Research, Development, and Innovations Infrastructures” (CESNET LM2015042) is greatly appreciated. MR was supported by Charles University Research Centre program (204069).

References

- Abascal F, Zardoya R, Telford MJ (2010) TranslatorX: multiple alignment of nucleotide sequences guided by amino acid translations. *Nucleic Acids Research* 38: 7–13. <https://doi.org/10.1093/nar/gkq291> [Web issue]
- Advokaat EL, van Hinsbergen DJJ, Maffione M, Langereis CG, Vissers RLM, Cherchi A, Schroeder R, Madani H, Columbu S (2014) Eocene rotation of Sardinia, and the paleogeography of the western Mediterranean region. *Earth and Planetary Science Letters* 401: 183–195. <https://doi.org/10.1016/j.epsl.2014.06.012>
- Ali RF, Neiber MT, Walther F, Hausdorf B (2016) Morphological and genetic differentiation of *Eremina desertorum* (Gastropoda, Pulmonata, Helicidae) in Egypt. *Zoologica Scripta* 45: 48–61. <https://doi.org/10.1111/zsc.12134>
- Andrews S (2010) FastQC: A quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>
- Ansart A, Madec L, Guiller A (2009) Datasheet: *Cornu aspersum* (common garden snail). Invasive Species Compendium. <http://www.cabi.org/isc> [updated by Cowie R (2015)]
- Bergsten J (2005) A review of long-branch attraction. *Cladistics* 21: 163–193. <https://doi.org/10.1111/j.1096-0031.2005.00059.x>
- Bernt M, Braband A, Schierwater B, Stadler PF (2013a) Genetic aspects of mitochondrial genome evolution. *Molecular Phylogenetics and Evolution* 69: 328–338. <https://doi.org/10.1016/j.ympev.2012.10.020>
- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsche G, Pütz J, Middendorf M, Stadler PF (2013b) MITOS: improved de novo metazoan mitochondrial genome annotation. *Molecular Phylogenetics and Evolution* 69: 313–319. <https://doi.org/10.1016/j.ympev.2012.08.023>
- Bisenberger A, Baumgartner G, Kleewein D, Sattmann H (1999) Untersuchungen zur Populationsökologie von *Cylindrus obtusus* (Draparnaud, 1805) (Pulmonata, Helicidae). *Annalen des Naturhistorischen Museums in Wien* 101B: 453–464.
- Bouchet P, Rocroi J-P (2005) Classification and nomenclator of gastropod families. *Malacologia* 47: 1–397.
- Cain AJ, Sheppard PM (1950) Selection in the polymorphic land snail *Cepaea nemoralis*. *Heredity* 4: 275–294. <https://doi.org/10.1038/hdy.1950.22>
- Chernomor O, von Haeseler A, Minh BQ (2016) Terrace aware data structure for phylogenomic inference from supermatrices. *Systematic Biology* 65: 997–1008. <https://doi.org/10.1093/sysbio/syw037>
- Cowie RH (1984) The life-cycle and productivity of the land snail *Theba pisana* (Mollusca: Helicidae). *Journal of Animal Ecology* 53: 311–325. <https://doi.org/10.2307/4359>
- Ewels P, Magnusson M, Lundin S, Käller M (2016) MultiQC: Summarize analysis results for multiple tools and samples in a single report. *Bioinformatics* 32: 3047–3048. <https://doi.org/10.1093/bioinformatics/btw354>
- Foster PG (2004) Modelling compositional heterogeneity. *Systematic Biology* 53: 485–495. <https://doi.org/10.1080/10635150490445779>
- Gaitán-Espitia JD, Nespolo RF, Opazo JC (2013) The complete mitochondrial genome of the land snail *Cornu aspersum* (Helicidae: Mollusca): intraspecific divergence of protein-coding

- genes and phylogenetic considerations within Euthyneura. PLoS ONE 8: e67299. <https://doi.org/10.1371/journal.pone.0067299>
- Gittenberger E, Ripken TEJ (1987) The genus *Theba* (Mollusca: Gastropoda: Helicidae), systematics and distribution. Zoologische Verhandlungen 241: 1–59.
- Giusti F, Manganeli G (1984) Relationships between geological land evolution and present distribution of terrestrial gastropods in the western Mediterranean area. In: Solem A, van Bruggen AC (Eds) World-wide snails: Biogeographical studies on non-marine Mollusca. Brill Archive, Leiden, 70–92.
- González VL, Kayal E, Halloran M, Shrestha Y, Harasewych MG (2016) The complete mitochondrial genome of the land snail *Cerion incanum* (Gastropoda: Stylommatophora) and the phylogenetic relationships of Cerionidae within Panpulmonata. Journal of Molluscan Studies 82: 525–533. <https://doi.org/10.1093/mollus/eyw017>
- Groenenberg DSJ, Harl J, Duijijm E, Gittenberger E (2017) The complete mitogenome of *Oracula dolium* (Draparnaud, 1801); ultra-deep sequencing from a single long-range PCR using the Ion-Torrent PGM. Hereditas 154: 7. <https://doi.org/10.1186/s41065-017-0028-2>
- Groenenberg DSJ, Pirovano W, Gittenberger E, Schilthuizen M (2012) The complete mitogenome of *Cylindrus obtusus* (Helicidae, Ariantinae) using Illumina next generation sequencing. BMC Genomics 13: 114. <https://doi.org/10.1186/1471-2164-13-114>
- Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Systematic Biology 59: 307–321. <https://doi.org/10.1093/sysbio/syq010>
- Heller J (1982) Natural history of *Theba pisana* in Israel (Pulmonata: Helicidae). Journal of Zoology 196: 475–487. <https://doi.org/10.1111/j.1469-7998.1982.tb03519.x>
- Holyoak DT, Holyoak GA, Chueca LJ, Gómez Moliner BJ (2018) Evolution and taxonomy of the populations of *Eremina* (Gastropoda, Pulmonata, Helicidae) in Morocco. Journal of Conchology 43: 17–57.
- Jones JS, Leith BH, Rawlings P (1977) Polymorphism in Cepaea: a problem with too many solutions? Annual Review of Ecology and Systematics 8: 109–143. <https://doi.org/10.1146/annurev.es.08.110177.000545>
- Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermini LS (2017) ModelFinder: Fast model selection for accurate phylogenetic estimates. Nature Methods 14: 587–589. <https://doi.org/10.1038/nmeth.4285>
- Katoh K, Kuma KI, Toh H, Miyata T (2005) MAFFT version 5: improvement in accuracy of multiple sequence alignment. Nucleic Acids Research 33: 511–518. <https://doi.org/10.1093/nar/gki198>
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A (2012) Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28: 1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>
- Kerkvliet J, de Boer T, Schilthuizen M, Kraaijeveld K (2017) Candidate genes for shell colour polymorphism in *Cepaea nemoralis*. PeerJ 5: e3715. <https://doi.org/10.7717/peerj.3715>

- Knudsen B, Kohn AB, Nahir B, McFadden CS, Moroz LL (2006) Complete DNA sequence of the mitochondrial genome of the sea-slug, *Aplysia californica*: conservation of the gene order in Euthyneura. *Molecular Phylogenetics and Evolution* 38: 459–469. <https://doi.org/10.1016/j.ympev.2005.08.017>
- Korábek O, Petrušek A, Juříčková L (2018) Glacial refugia and postglacial spread of an iconic large European land snail, *Helix pomatia* (Pulmonata: Helicidae). *Biological Journal of the Linnean Society* 123: 218–234. <https://doi.org/10.1093/biolinnean/blx135>
- Korábek O, Petrušek A, Neubert E, Juříčková L (2015) Molecular phylogeny of the genus *Helix* (Pulmonata: Helicidae). *Zoologica Scripta* 44: 263–280. <https://doi.org/10.1111/zsc.12101>
- Minton RL, Martinez Cruz MA, Farman ML, Perez KE (2016) Two complete mitochondrial genomes from *Praticolella mexicana* Perez, 2011 (Polygyridae) and gene order evolution in Helicoidea (Mollusca, Gastropoda). *ZooKeys* 626: 137–154. <https://doi.org/10.3897/zookeys.626.9633>
- Neiber MT, Hausdorf B (2015) Molecular phylogeny reveals the polyphyly of the snail genus *Cepaea* (Gastropoda: Helicidae). *Molecular Phylogenetics and Evolution* 93: 143–149. <https://doi.org/10.1016/j.ympev.2015.07.022>
- Neubert E (2014) Revision of *Helix* Linnaeus, 1758 in its eastern Mediterranean distribution area, and reassignment of *Helix godetiana* Kobelt, 1878 to *Maltzanella* Hesse, 1917 (Gastropoda, Pulmonata, Helicidae). *Contributions to Natural History* 26: 1–200.
- Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ (2015) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology and Evolution* 32:268–274. <https://doi.org/10.1093/molbev/msu300>
- Nordsieck H (1987) Revision des Systems der Helicoidea (Gastropoda: Stylommatophora). *Archiv für Molluskenkunde* 118: 9–50.
- Perrot M (1939) Sur la position systématique d'*Euparypha pisana* Muller, d'après l'étude chromosomique. *Archives des Sciences physiques et naturelles* 21 (Supplément: Compte Rendu des Séances de la Société de Physique et d'Histoire naturelle de Genève) 56: 32–35.
- Pollard E (1975) Aspects of the ecology of *Helix pomatia* L. *Journal of Animal Ecology* 44: 305–329. <https://doi.org/10.2307/3865>
- Razkin O, Gómez-Moliner BJ, Prieto CE, Martínez-Ortí A, Arrébola JR, Muñoz B, Chueca LJ, Madeira MJ (2015) Molecular phylogeny of the western Palaearctic Helicoidea (Gastropoda, Stylommatophora). *Molecular Phylogenetics and Evolution* 83: 99–117. <https://doi.org/10.1016/j.ympev.2014.11.014>
- Rees WJ (1955) Escaped escargots. *The Nautilus* 68: 90–94.
- Rosenbaum G, Lister GS, Duboz C (2002) Reconstruction of the tectonic evolution of the western Mediterranean since the Oligocene. In: Rosenbaum G, Lister GS (Eds) Reconstruction of the evolution of the Alpine-Himalayan Orogen. *Journal of the Virtual Explorer* 8: 107–130. <https://doi.org/10.3809/jvirtex.2002.00053>
- Rota-Stabelli O, Yang Z, Telford MJ (2009) MtZoa: A general mitochondrial amino acid substitutions model for animal evolutionary studies. *Molecular Phylogenetics and Evolution* 52: 268–272. <https://doi.org/10.1016/j.ympev.2009.01.011>

- Saclier N, François CM, Konecny-Dupré L, Lartillot N, Guéguen L, Duret L, Malard F, Douady CJ, Lefébure T (2018) Life history traits impact the nuclear rate of substitution but not the mitochondrial rate in isopods. *Molecular Biology and Evolution* 35: 2900–2912. <https://doi.org/10.1093/molbev/msy184>
- Schileyko AA (2006) Treatise on recent terrestrial pulmonate molluscs. Part 13. Helicidae, Pleurodontidae, Polygyridae, Ammonitellidae, Oreohelicidae, Thysanophoridae. *Ruthenica Supplement 2*: 1035–1166.
- Schileyko AA (2013) Family Helicidae excluding Helicinae (Gastropoda Pulmonata): morphology, taxonomy, and a catalogue of taxa. *Ruthenica* 23: 127–162.
- Silvertown J, Cook L, Cameron R, Dodd M, McConway K, Worthington J, Skelton P, Anton C, Bossdorf O, Baur B, Schilthuizen M, Fontaine B, Sattmann H, Bertorelle G, Correia M, Oliveira C, Pokryszko B, Ozgo M, Stalaž A, Gill E, Rammul Ü, Sólymos P, Féher Z, Juan X (2011) Citizen science reveals unexpected continental-scale evolutionary change in a model organism. *PLoS ONE* 6: e18927. <https://doi.org/10.1371/journal.pone.0018927>
- Teasdale LC, Koehler F, Murray KD, O'Hara T, Moussalli A (2016) Identification and qualification of 500 nuclear, single-copy, orthologous genes for the Eupulmonata (Gastropoda) using transcriptome sequencing and exon capture. *Molecular Ecology Resources* 16: 1107–1123. <https://doi.org/10.1111/1755-0998.12552>
- Terrett JA, Miles S, Thomas RH (1996) Complete DNA Sequence of the Mitochondrial Genome of *Cepaea nemoralis* (Gastropoda: Pulmonata). *Journal of Molecular Evolution* 42: 160–168. <https://doi.org/10.1007/BF02198842>
- Thomas AJ, Welch JJ, Lanfear R, Bromham L (2010) A generation time effect on the rate of molecular evolution in invertebrates. *Molecular Biology and Evolution* 27: 1173–1180. <https://doi.org/10.1093/molbev/msq009>
- van Hinsbergen DJJ, Vissers RLM, Spakman W (2014) Origin and consequences of western Mediterranean subduction, rollback, and slab segmentation. *Tectonics* 33: 393–419. <https://doi.org/10.1002/2013TC003349>
- Wang H-C, Minh BQ, Susko E, Roger AJ (2018a) Modeling site heterogeneity with posterior mean site frequency profiles accelerates accurate phylogenomic estimation. *Systematic Biology* 67: 216–235. <https://doi.org/10.1093/sysbio/syx068>
- Wang P, Yang S-P, Lin J-H, Zhang M-Z, Zhou W-C (2018b) The mitochondrial genome of the land snail *Theba pisana* (Müller, 1774) (Stylommatophora: Helicidae): the first complete sequences in the genus *Theba*. *Mitochondrial DNA Part B: Resources* 3: 798–800. <https://doi.org/10.1080/23802359.2018.1491341>
- Welter-Schultes FW (2012) European non-marine molluscs, a guide for species identification. Planet Poster Editions, Göttingen.
- White TR, Conrad MM, Tseng R, Balayan S, Golding R, de Frias Martins AM, Dayrat BA (2011) Ten new complete mitochondrial genomes of pulmonates (Mollusca: Gastropoda) and their impact on phylogenetic relationships. *BMC Evolutionary Biology* 11: 295. <https://doi.org/10.1186/1471-2148-11-295>
- Ye J, Coulouris G, Zaretskaya I, Cutcutache I, Rozen S, Madden TL (2012) Primer-BLAST: a tool to design target-specific primers for polymerase chain reaction. *BMC Bioinformatics* 13: 134. <https://doi.org/10.1186/1471-2105-13-134>