

Chordodes ferox, a new record of horsehair worms (Nematomorpha, Gordiida) from South Africa

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Abstract

Three females and one male specimen of a previously unconfirmed species of horsehair worms (Nematomorpha) from South Africa are described using Scanning Electron Microscopy. The females correspond to the description of *Chordodes ferox* Camerano, 1897, a species previously described from the Democratic Republic of the Congo (Congo-Kinshasa) and an adjacent, not further specified region of the Republic of Congo (Congo-Brazzaville). Characteristic is the presence of enlarged and elevated simple areoles around the base of a thorn areole, in combination with further cuticular characters. This is the latest of a total of six species of horsehair worms reported from South Africa so far. Two species of praying mantids, *Poly-spilota aeruginosa* (Goeze, 1778) and *Sphodromantis gastrica* Stål, 1858, have been identified as hosts of *C. ferox*, while its distribution range in the region and the period of adult emergence from the host remain largely unknown.

Keywords

Nematomorpha, *Chordodes ferox*, praying mantid hosts, new records, South Africa

Introduction

A number of horsehair worms (Nematomorpha) are known and have been described from Africa, but most are occasional records, with few results from extensive sampling. Although still far from systematic sampling, Sciacchitano's (1933, 1937, 1955, 1958,

1961) reports from the Democratic Republic of the Congo (then the “Belgian Congo”) are the most comprehensive records yet reported for an African country. For most other countries, only few species are known, which certainly underestimates the diversity of horsehair worms in most cases. From South Africa, six species have tentatively been reported thus far: *Chordodes capensis* Camerano, 1895 from the Cape of Good Hope; *Chordodes hawkeri* Camerano, 1902 from Grahamstown (erroneously reported as “Grahamstow”) in the Eastern Cape; *Chordodes ferox* Camerano, 1897 from Pietermaritzburg, KwaZulu-Natal; *Paragordius areolatus* Linstow, 1906 from Botshabelo, *Paragordius cinctus* Linstow, 1906 from the Mpumalanga Province; and *Beatogordius regularius* Heinze, 1934 probably from the Cape of Good Hope, as the location was only given in German as “Cap” (= Cape) (Camerano 1895, 1908, Linstow 1906, Heinze 1934).

Nematomorpha are long and slender worms, which develop parasitically in terrestrial insects (mainly praying mantids, carabid beetles, crickets and cockroaches) and a few other arthropods. Eventually, they emerge from these hosts into water for reproduction (see e.g. Hanelt et al. 2005, Schmidt-Rhaesa 2013). Early larval development takes place in water and larvae infect various aquatic animals as intermediate/paratenic hosts. This early phase of the life cycle and the transmission to the final host are not completely understood yet (see Hanelt et al. 2005, Schmidt-Rhaesa 2013 for reviews). About 360 species of nematomorphs are currently known, five from the marine environment (genus *Nectonema*) and the remaining (Gordiida) from freshwater.

Gordiids have comparably few diagnostic features. These are the shape of the posterior end and cuticular structures, such as bristles, spines and variously shaped elevations called areoles. Traditionally, cuticular samples were removed from the animals and investigated with the microscope, but now scanning electron microscopical (SEM) investigation has become the standard method, because with this technique characters can be documented at higher magnification and better quality. A number of African species have been reinvestigated using SEM (e.g. Schmidt-Rhaesa and De Villalobos 2002, Zanca et al. 2006a, b, De Villalobos et al. 2007, 2009) and recent species descriptions always include SEM analyses (e.g. Bolek et al. 2010, 2013, Hanelt et al. 2012).

We report here the determination of four specimens of horsehair worms, as *Chordodes ferox* Camerano, 1897 (at least the females). This species has been reported once before from Pietermaritzburg, KwaZulu-Natal, based on an old and poorly preserved specimen deposited in the Natural History Museum, London (see Schmidt-Rhaesa and Ehrmann 2001). However, the correctness of this identification remains questionable. More than 20 specimens of horsehair worms from different locations in Africa carry the label “*Chordodes ferox*” but it has not been established yet how many of these were adequately investigated and identified.

Methods

A total of four specimens were found in the Baviaanskloof Valley at Kudu Kaya Farm (river at campsite, 33°39'12.4"S, 24°34'59.7"E) in the Eastern Cape Province of South

Africa, on 22 March 2015, while undertaking preliminary observations in the area. Two specimens emerged from the abdomen of a praying mantid, identified as *Poly-spilota aeruginosa* (Figure 1a), attracted to the light at night. The other two specimens were lying free underwater (approximately 5–10 cm deep) on rocky substrata (Figure 1b), one in close proximity to an inactive, but still alive specimen of the praying mantid species *Sphodromantis gastrica*. Unfortunately, specimens were initially pooled together in the same container and, therefore, it cannot be established with certainty which of the specimens were the ones that emerged from the mantid *P. aeruginosa*. All specimens were taken alive to a laboratory for close observation under a dissecting microscope and later preserved in 99% ethanol.

Specimens are deposited in the Zoological Museum in the Centrum für Naturkunde of the University Hamburg under the numbers ZMH13363-ZMH13366.



Figure 1. *Chordodes ferox*: **a** two specimens emerging from the posterior end of the praying mantid *Poly-spilota aeruginosa* **b** free-living specimen on rocky substrata underwater showing a curled posterior end. Photos: Lynette Clennell, March 2015.

For Scanning Electron Microscopy (SEM), both anterior and posterior ends of the body as well as a 2–3 mm long section from the middle region of the body were cut and dehydrated in an increasing ethanol:water gradient, then critically point dried, and coated with gold in a sputter coater. Observations were made using JSM-6360 (JEOL) SEM operating at 15 kV. Digital images were taken. Specimens comprise three females and one male, which are described separately here. All females are similar in their diagnostic characters, while the male differs slightly from the females in some characters. Figures used here come from the male (ZMH13363) and two females (ZMH13364 and ZMH13367), because in the third female (ZMH13366) the cuticle was dirty and less well preserved. For the description of cuticular structures (areoles) the terminology of Schmidt-Rhaesa et al. (2008) is used.

Description

Female

The posterior end of the females appears slightly swollen because the diameter of the body decreases slightly about 1 mm from the posterior tip (Figure 2a). The posterior surface is free of cuticular structures (areoles) and the cloacal opening is terminal (Figure 2a).

Five different types of areoles were observed on the cuticle of the median piece. Simple areoles are roundish and have an almost smooth surface (Figures 2d, f, 3b–d). In the ventral midline they are slightly smaller than in other regions (Figure 2d). Crowned areoles occur in two forms, with long apical filaments along the ventral and dorsal midline and with short apical filaments in the lateral regions of the body (Figure 2e). Apical filaments of lateral crowned areoles are short to medium in length (about 10 µm), while the long filaments are up to 150 µm long. Crowned areoles occur in clusters with two central crowned areoles surrounded by circum-cluster areoles. These areoles have short bristles on top (Figure 3b). They decrease in size towards the periphery, which makes their number difficult to count. They are rarely few (ca. 14, Figure 3b), but usually between 18 and 20 (Figure 2f). Typical tubercle areoles were not observed, but rarely a tubercle areole with a strongly eccentric tubercle was found between simple areoles (Figure 3e). Few thorn areoles were found in the ventral region. Solid thorns are surrounded by 2–3 areoles, which appear sometimes (Figure 3f), but not always (Figure 3d) larger or more elevated than surrounding ones.

Male

The posterior end exhibits the typical characteristics of the genus *Chordodes* (Figure 2b). The cloacal opening is ventral, about 250 µm from the posterior tip of the body. Its shape or the presence of circum-cloacal spines could not be observed due to a plug

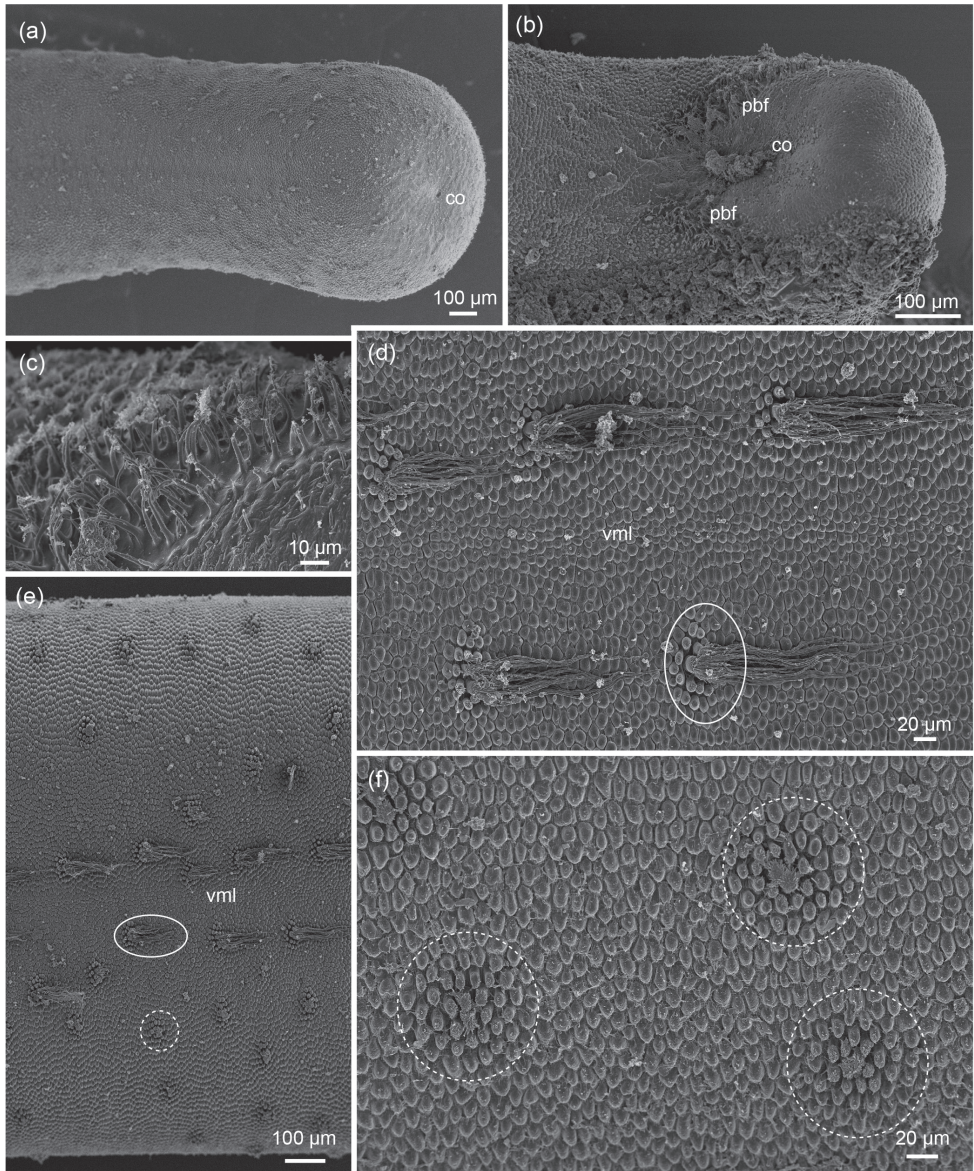


Figure 2. *Chordodes ferox*: **a** posterior end of female with terminal cloacal opening (co) **b** ventral view of posterior end of male with ventral cloacal opening (co) and precloacal bristlefields (pbf) **c** enlargement of bristles in the precloacal bristlefields **d** clusters of crowned areoles with long apical filaments (encircled) along both sides of the ventral midline (vml) in a female **e** overview of the ventral side of a female showing the distribution of clusters of crowned areoles with long (oval with unbroken line) and short (circle with broken line) apical filaments **f** clusters of crowned areoles in center and surrounding circum-cluster areoles (entire cluster encircled). SEM images: Andreas Schmidt-Rhaesa **a, f** from ZMH13366 **b, c** from ZMH13363 **d, e** from ZMH13364.

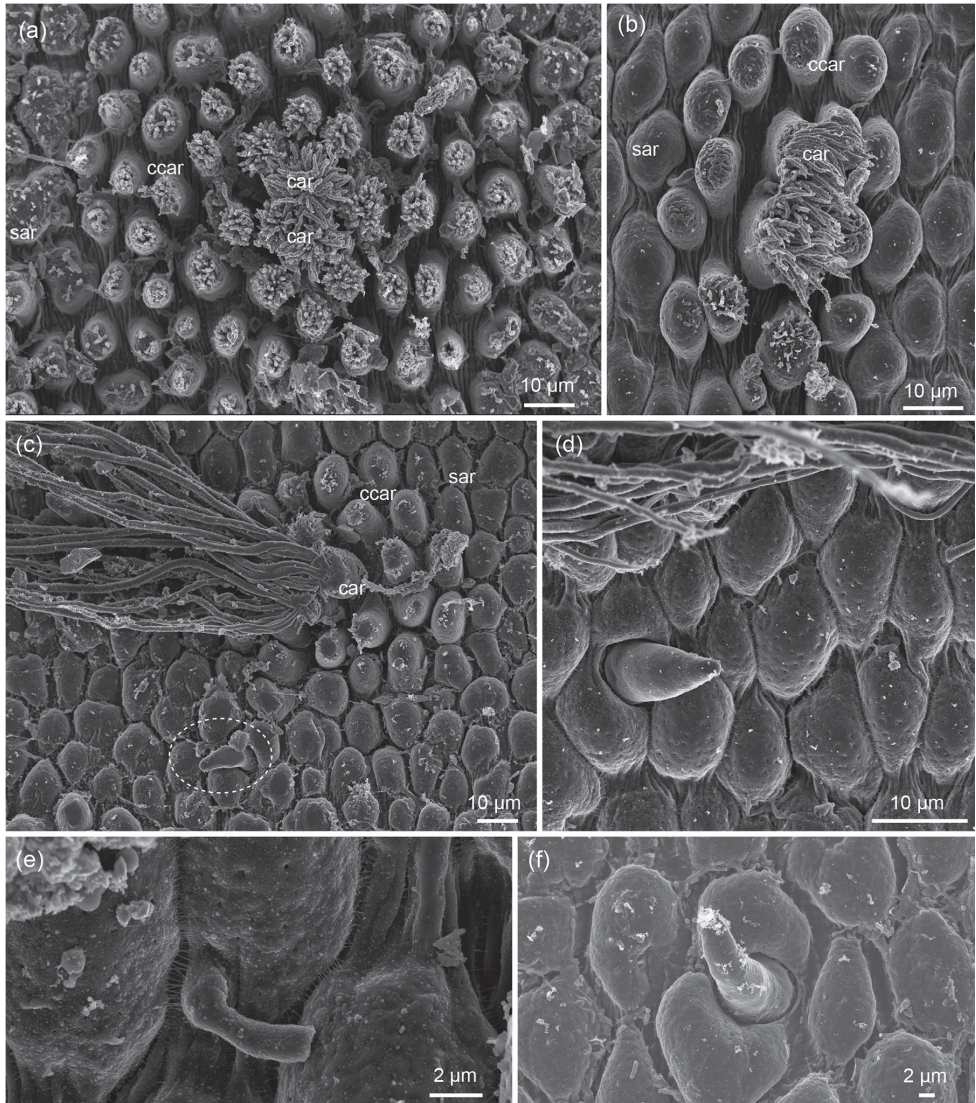


Figure 3. *Chordodes ferox*, higher magnification of cuticular characters: **a** cluster composed of two central crowned areoles (car) and surrounding circumcluster areoles (ccar) **b** cluster with fewer circumcluster areoles (Note transition between circumcluster areoles and simple areoles, sar) **c** cluster of crowned areoles with long apical filaments (encircled is a thorn areoles) **d, f** thorn areoles **e** isolated tubercle next to simple areoles. SEM images: Andreas Schmidt-Rhaesa **a** from ZMH13363 **b, d, e** from ZMH13364 **c, f** from ZMH13366.

of dirt or sperm covering the opening. Antero-lateral of the cloacal opening are paired fields of spines (Figure 2c).

Simple areoles are roundish and their apical surface has small “knobs” (Figure 3a). Tubercle areoles were not seen, thorn areoles were found very rarely, while the surrounding simple areoles are not conspicuously larger than the others (not figured).

Crowned areoles occur only with short apical filaments. Clusters of crowned and circum-cluster areoles are larger than in females and comprise > 40 circum-cluster areoles (Figure 3a).

Discussion

Simple areoles, tubercle areoles, thorn areoles, crowned areoles in clusters surrounded by circum-cluster areoles and the presence of crowned areoles with conspicuously long apical filaments are typical characters found in *Chordodes* species (Schmidt-Rhaesa et al. 2008). The two other *Chordodes* species described from South Africa, *C. capensis* Camerano, 1895 and *C. hawkeri* Camerano, 1902 differ substantially from the specimens reported here. Both of these species have been reinvestigated recently using SEM techniques (Zanca et al. 2006b, De Villalobos et al. 2007). In *C. capensis* both sexes are similar in their cuticular characters, with the exception that crowned areoles with long filaments are present in females, but absent in males (De Villalobos et al. 2007). The main characters of *C. capensis* which differ from the specimens reported here are the presence of fine bristles on the surface of the simple areoles, the scattered presence of tubercle areoles and the absence of thorn areoles. In *C. hawkeri*, both sexes are consistent in their cuticular characters, but the cuticle of the specimens analysed so far (lectotype male and paralectotype female) is in a relatively poor state and misinterpretations cannot be excluded (Zanca et al. 2006b). Crowned areoles with long filaments and thorn areoles were not reported in the specimens investigated by SEM. Other differences to the specimens reported here are the rough surface and irregular shape of simple areoles, isolated occurrence of crowned areoles (rather than in clusters), the scattered presence of tubercle areoles and the presence of another type of areoles, i.e. bulging areoles.

The characters of the female specimens investigated here correspond well with those reported in the description of *Chordodes ferox* Camerano, 1893. The species has been described based on one female specimen from the Republic of Congo ("French Congo") found in an unidentified mantid species (Camerano 1893). According to Camerano (1893), *Gordius verrucosus* is, at least in part, a synonym for *C. ferox*. This species has been described by Baird (1853) from a specimen of unknown locality. Subsequent authors have mentioned the location "Africa australis" (Diesing 1861, Villot 1874) or South Africa and Ceylon (Örley 1881), but while Diesing and Villot seem to relate to Baird's specimen only, it is not clear whether Örley investigated further specimens. However, none of the descriptions was detailed enough or sufficiently well illustrated for a proper species description and the name was eventually regarded as "species inquirendae" (Camerano 1897).

The holotype of this species as well as further specimens have been investigated using SEM by De Villalobos et al. (2009). The described characters correspond to the characters described here. The combination of simple areoles with a smooth surface, crowned areoles with very short apical filaments, the presence of thorn areoles and tubercle areoles with an eccentric tubercle are regarded as characteristic for *C. ferox*.

In addition, one potential further character of the specimens reported here is that the thorn areoles appear to be surrounded by a few large simple areoles. In most other species, thorn areoles are composed of a base, which resembles a “usual” areole and the thorn itself (compare, e.g. with figure 2C in Schmidt-Rhaesa et al. 2008). In the specimens described here, a basal structure is not recognized and instead the thorn is closely surrounded by simple areoles, which appear in some, but not all cases to be larger than the remaining ones. This arrangement has not yet been recognized as a character of taxonomic importance, but is, according to published figures, present in the species *C. ferox*, *C. maculatus* Sciacchitano, 1958 and *C. madagascariensis* (Camerano, 1893). *Chordodes maculatus* is a species from the Democratic Republic of Congo (Sciacchitano 1958) and a SEM reinvestigation shows elevated simple areoles surrounding the base of thorn areoles (Figure 6B in De Villalobos et al. 2009). *Chordodes madagascariensis* is a species reported from Angola, the Democratic Republic of Congo, Guinea and Madagascar (see De Villalobos et al. 2009) and the shape of thorn and surrounding areoles can be seen in the original drawings (Figure 23 in Camerano 1897), but most clearly in the SEM reinvestigation (Figure 3D in De Villalobos et al. 2009). The main differences of these two species compared to *C. ferox* are that *C. madagascariensis* has high and slender crowned areoles (De Villalobos et al. 2009), while *C. maculatus* has additional bulging areoles (De Villalobos et al. 2009). Therefore the cuticular characters of the female specimens reported here correspond best to *C. ferox*, thus extending the distribution of this species to the southern African region.

The single male specimen available differs in its cuticular characters from the female specimens. The most conspicuous difference is the lack of crowned areoles with long apical filaments, which is a well-known sexually dimorphic character in the genus *Chordodes* (e.g. Schmidt-Rhaesa et al. 2008). However, as other differences concerning the surface of the simple areoles, the arrangement of areoles around thorn areoles and the number of circum-cluster areoles are also present, it is not clear whether this male, belongs to *C. ferox*, despite having been found next to one of the females. All previous specimens reported to date are females, so it is not known how extensive sexually dimorphic characters are. There is no other species, which corresponds well to the characters of the male, therefore it cannot be assigned to any species currently known.

Praying mantids are the dominant host insects of horsehair worms of the genus *Chordodes* (Schmidt-Rhaesa and Ehrmann 2001). Both species recognized here, *Poly-spilota aeruginosa* and *Sphodromantis gastrica*, have been recorded once before for worm specimens identified as “*C. ferox*”, from Tanzania and from Pietermaritzburg, South Africa, respectively (Schmidt-Rhaesa and Ehrmann 2001). Both records were not from fresh specimens adequately documented in the literature, but from poorly preserved museum specimens. As the methods to investigate horsehair worms, as well as the awareness of diagnostic characters most suitable for their identification have changed and/or developed during the past decades, there is now a need for all old museum specimens to be re-investigated, at least those that have not yet been included in the peer-reviewed literature.

Although direct evidence of worm emergence was only observed from the abdomen of *P. aeruginosa*, the second mantid species, *S. gastrica*, was found floating on the water

in close proximity to one of the free-living worms, still alive but poorly active. There is little doubt, therefore, that it was involved in the release of the worm. Regarding timing and locality of the finding of the *C. ferox* specimens, it is not known yet whether the period of emergence from the host is restricted to the austral autumn, as all specimens were collected towards the end of March. Searches during other seasons of the year are currently in progress but have so far yielded no further specimens. Regarding locality, all specimens were found in a relatively small riverine area within the Baviaanskloof valley. Although in a privately owned farm, the area is completely surrounded by the Baviaanskloof Wilderness Area, which is part of the Cape Floral Kingdom World Heritage Site. It is virtually certain that *C. ferox* occurs through much of this area, particularly in the smaller water courses that act as tributaries to the main river.

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Review of species of the genus *Adelurola* Strand, 1928, with a key to species (Hymenoptera, Braconidae, Alysiinae)

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Abstract

The alysiine genus *Adelurola* Strand, 1928 (Hymenoptera, Braconidae) is revised. Illustrated re-descriptions and a key to all known species of this genus are given. The following new combination is proposed: *Dapsilarthra eurys* (Chen & Wu, 1994), **comb. n.** *Adelurola amplidens* (Fischer, 1966) and *A. asiatica* Telenga, 1935 are recorded for the first time from Iran and Kyrgyzstan, respectively.

Keywords

Hymenoptera, Alysiinae, *Adelurola*, fly endoparasitoid, new record, new combination, redescription, Iran

Introduction

Adeluroloa Strand, 1928 is a small Palaearctic genus of the braconid subfamily Alysiinae that currently contains five recognised species (Yu et al. 2012). Traditionally, most species of *Adeluroloa* were included within *Dapsilarthra* (e.g. Wharton 1980). Van Achterberg (1983) clarified the status of both genera and found valuable differences between them, including the presence of a ventral lamelliform lobe on the mandible (Fig. 1), the second flagellar segment usually subequal or slightly longer than the first segment, and the precoxal sulcus more or less sculptured.

Our current taxonomic research on the braconid wasps of the subfamily Alysiinae in Iran resulted in the first record of a species of *Adeluroloa* in this country. The difficulty for the identification of species of *Adeluroloa* has led to a major revision of this genus. In this work, all the currently recognised species *Adeluroloa* are re-described and an identification key is provided.

Material and methods

Sampling in Iran was carried out by sweeping with a standard net in Kermanshah province (western part of Iran) in 2013. Specimens were subsequently prepared using the AXA method (van Achterberg 2009). For terminology of the morphological features and sculpture, measurements and wing venation nomenclature see van Achterberg (1993). Photographs were taken with a Digital Microscope VHX-2000 and with a Nikon® D700 mounted on a Leica® S8APO microscope, with images combined using Helicon Focus® and edited using Adobe Photoshop® imaging system.

Specimens examined are deposited in the collection of the Zoological Institute of the Russian Academy of Sciences, St Petersburg, Russia (ZISP), and in the collection of Naturalis Biodiversity Center, Leiden, the Netherlands (RMNH). Type specimen of *Alysia florimela* Haliday was studied by the third author in the Haliday Collection (Dublin, Ireland) (van Achterberg 1993); the type material of *Neocarpa amplidens* Fischer is missing in the Zoologische Sammlung (Munich, Germany) and its current location is unclear.

Results

Genus *Adeluroloa* Strand, 1928

Adelura Foerster 1863: 267 (not Bonaparte 1854); Shenefelt 1974: 986; van Achterberg 1993: 4; Yu et al. 2012.

Adeluroloa Strand 1928: 51 (nom. n. for *Adelura* Foerster); Shenefelt 1974: 986; van Achterberg 1993: 4; Yu et al. 2012.

Neocarpa Fischer 1966: 185; Shenefelt 1974: 987; van Achterberg 1993: 4; Yu et al. 2012.

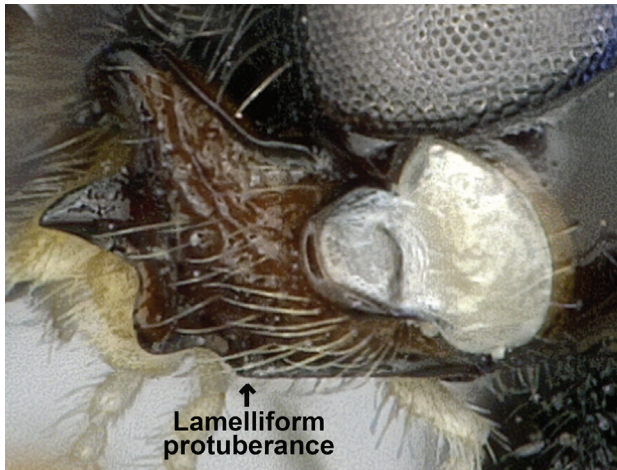


Figure 1. Mandible in *Adeluroloa*.

Type species. *Alysia florimela* Haliday, 1838.

Diagnosis. Mandibles distinctly broadened apically, with additional ventral wide fourth denticle (lobe). Eyes glabrous. Second flagellar segment not longer than first segment. Pterostigma short and wide. Radial vein (r) originating almost from its middle. Brachial (first subdiscal) cell of fore wing closed apically. Pronope of mesosoma small or absent. Precoxal sulcus distinctly sculptured. Dorsople of first metasomal tergite distinct. Second tergite smooth. Ovipositor short, shorter than apical height of metasoma.

Remarks. This genus is similar to *Dapsilarthra* but differs from it in having the wide ventral lamelliform lobe on the mandible (Fig. 1) (absent in *Dapsilarthra*), the first flagellar segment not shorter than second segment (shorter in *Dapsilarthra*), precoxal suture sculptured (usually smooth in *Dapsilarthra*), and radial vein (r) arising submedially from pterostigma (usually before pterostigma in *Dapsilarthra*).

The observation of these characters in studied images of the holotype of *Adeluroloa euryis* Chen & Wu, 1994 showed that this species is better placed under genus *Dapsilarthra* Foerster, 1863 (comb. n.).

Hosts. Cyclorrhaphous Diptera (Tephritidae and Anthomyiidae).

***Adeluroloa amplidens* (Fischer, 1966)**

Fig. 2

Neocarpa amplidens Fischer 1966: 85.

Dapsilarthra amplidens: Shenefelt 1974: 987; Wharton et al. 2006: 325.

Adeluroloa amplidens: van Achterberg 1983: 5; Yu et al. 2012.

Material examined. **Iran:** 2 females, Kermanshah Province, Kermanshah, 16.iv.2013, swept on *Medicago sativa* L. (Z. Sharifi coll.) (ZISP, RMNH); 1 male, Iran, Hormozgan

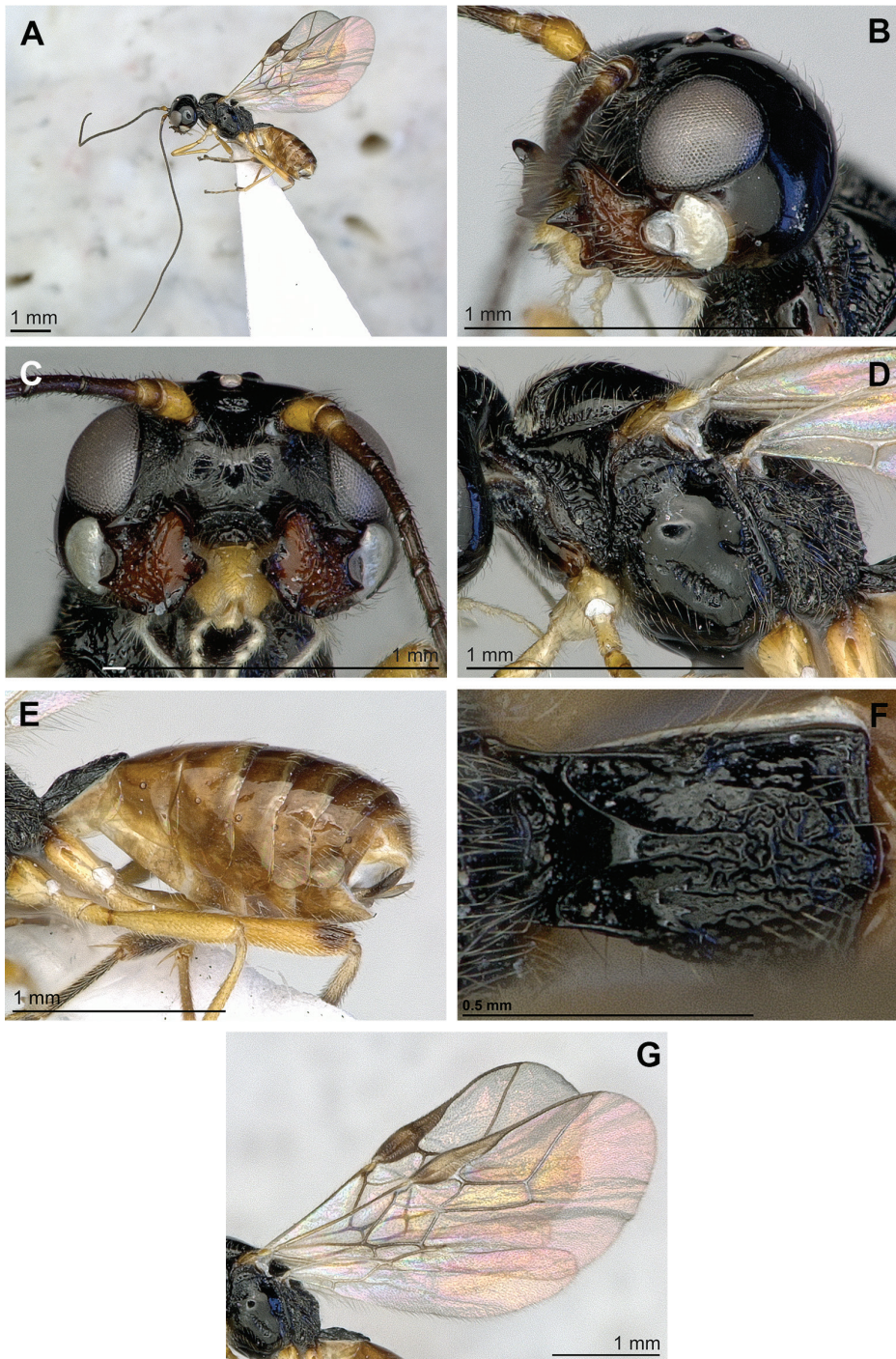


Figure 2. *Adeluraola amplidens* (Fischer) (female). **A** Habitus, lateral view **B** Head (lateral view) and mandible **C** Face, front view **D** Mesosoma, lateral view **E** Hind femur, metasoma and ovipositor, lateral view **F** First metasomal tergite **G** Fore and hind wings.

Province, Harsin, 16.iv.2013, swept on *Medicago sativa* L. (S. Sharifi coll.) (ZISP). **Iraq:** 2 females, Baghdad, em. 10.iv.[19]80 and 13.iv.[19]80; L. Jabbar; on *Beta vulgaris*; ex *Pegomyia hyoscyami*; *Dapsilarthra amplidens* ♀ det. Papp J. 1981" (RMNH).

Description. Female.

Head entirely smooth; in dorsal view twice as wide as median length, 1.5 times as wide as mesoscutum, with rounded temples behind eye. Eye in lateral view 1.5 times as high as wide and 0.8 times as wide as temple medially. POL 1.5 times OD; OOL 3.5 times OD. Face slightly punctate, with scattered short setae, without middle vertical protuberance in upper half, 1.9 times as wide as high; inner margins of eyes subparallel. Clypeus slightly curved ventrally, 1.9 times as wide as high. Mandible widened towards apex, 1.3 times as long as its maximum width. Upper tooth of mandible broadened towards subapex, longer than lower tooth; middle tooth wide basally and narrowed towards apex, rounded apically; lower tooth rounded apically. Antenna thick, 37-segmented. Scape 1.5 times as long as pedicel. First flagellar segment 3.0 times as long as its apical width; second segment 4.4 times as long as its maximum width, about as long as first segment. Third flagellar segment 4.0 times as long as its maximum width. Penultimate segment 2.0 times and apical segment 4.0 times as long as their maximum widths, respectively.

Mesosoma 1.5 times as long as high (lateral view). Mesoscutum smooth, punctate in antero-dorsal area, with numerous scattered setae, about as long as maximum width. Notauli present, punctate, reaching half part of mesoscutum, not reaching with mesoscutal pit. Mesoscutal pit present, elongate. Scutellar sulcus rugose-striate; with median and lateral carinae. Sides of pronotum sculptured. Precoxal suture present, widely rugose-crenulate, not reaching anterior and posterior margins of mesopleuron. Posterior mesopleural furrow crenulated. Propodeum completely rugose-reticulate, with numerous scattered setae. Propodeal spiracle relatively small.

Wings. Length of fore wing 2.3 times its maximum width. Pterostigma cuneate. Marginal cell ending before apex of wing, 2.5 times as long as its maximum width. Vein 3-SR 2.0 times as long as vein 2-SR. Vein SR1 1.8 times as long as vein 3-SR. Second submarginal cell 3.3 times as long as its maximum width. Vein cu-a postfurcal. Subdiscal cell closed, 3.8 times as long as its maximum width. Hind wing 4.6 times as long as its maximum width.

Legs. Hind femur 5.2 times as long as its maximum width. Hind tibia slightly widened towards apex, about 9.7 times as long as its maximum subapical width, 1.1 times as long as hind tarsus. First segment of hind tarsus 1.8 times as long as second segment.

Metasoma slightly compressed laterally. First tergite rugose-reticulate in apical half, without median carinae, slightly widened towards apex, 1.3 times as long as its apical width. Second metasomal tergite smooth. Ovipositor sheath 0.6 times as long as first tergite, 0.4 times as long as hind femur.

Colour. Body brown to dark brown. First metasomal tergite paler than second and third tergites, apical segments dark. Legs yellow, apical part of the tibia and hind tarsus darker than femur. Wings hyaline. Pterostigma brown.

Body length 3.9 mm; fore wing length 4.3 mm.

Male. Body length 3.5 mm; fore wing length 4.0 mm. Eye in lateral view 1.3 times as high as wide. Mandible 1.1 times as long as its maximum width. First flagellar segment 3.2 times as long as its apical width. Hind femur 4.8 times as long as its maximum width. Otherwise differs from female.

Differences of male types (according to original description: Fischer 1966). Fore wing length 4.4 mm. Mandible 1.3 times as long as its maximum width. Antenna 38–39-segmented. First flagellar segment 3.0 times as long as its apical width. Mesoscutum about as long as its maximum width. Hind femur 4.5 times as long as its maximum width.

Comparative diagnosis. This species is similar to *A. asiatica* Telenga, 1935 and *A. florimela* (Haliday, 1838). *Adeluroloa amplidens* differs from *A. asiatica* in having the eye in lateral view 0.8 times as wide as temple medially (1.2 times in *A. asiatica*), marginal cell 2.5 times as long as its maximum width (3.8 times in *A. asiatica*), and precoxal suture not reaching anterior and posterior margins of mesopleuron (reaching anterior and posterior margins in *A. asiatica*). *A. amplidens* differs from *A. florimela* in having the eye in lateral view 0.85 times as wide as temple medially (about 1.2 times in *A. florimela*), first metasomal tergite without median carinae (with median carinae in *A. florimela*), vein 3-SR 2.0 times as long as vein 2-SR (1.1–1.3 times in *A. florimela*), vein SR1 1.8 times as long as veins 3-SR (2.2–2.6 times in *A. florimela*), and marginal cell 2.5 times as long as its maximum width (2.8–3.2 times in *A. florimela*).

Distribution. Iraq, Iran (new record).

Adeluroloa asiatica Telenga, 1935

Fig. 3

Adeluroloa asiatica Telenga 1935: 186; van Achterberg 1983: 6; Tobias 1986: 236; Yu et al. 2012.

Dapsilarthra asiatica: Königsmann 1959: 599; Fischer 1970: 11; 1971: 78; Shenefelt 1974: 987.

Type material. Holotype: female (nearly entirely destroyed; head, antennae, all wings, metasoma, right fore and hind legs missing), **Uzbekistan**, silver circle, “Yargak (Jargak), Khatvirg. r. (?), 24.iv.[19]28, L. Zimin” (ZISP).

Additional material. **Kyrgyzstan:** 1 female, “20 km S of Toktogul, gorge of Karasu River, forest, 25.vii.1982, S. Belokobylskij coll.” (ZISP). **Turkmenistan:** 1 female, “between Sumbar and Chandyr Rivers, Monzhukly Mountain Range, Kara-Yantam gorge, 2 km E of Karakel Aul, 28.iv.1993, V. Perepechaenko coll.” (ZISP).

Description. Female.

Head entirely smooth; in dorsal view 1.8 times as wide as median length, 1.45 times as wide as mesoscutum, with convex rounded temples behind eye. Eye in lateral view 1.35 times as high as wide and 0.9 times as wide as temple medially. POL 0.8 times OD; OOL 2.8 times OD. Face smooth, with very fine reticulation, with sca-

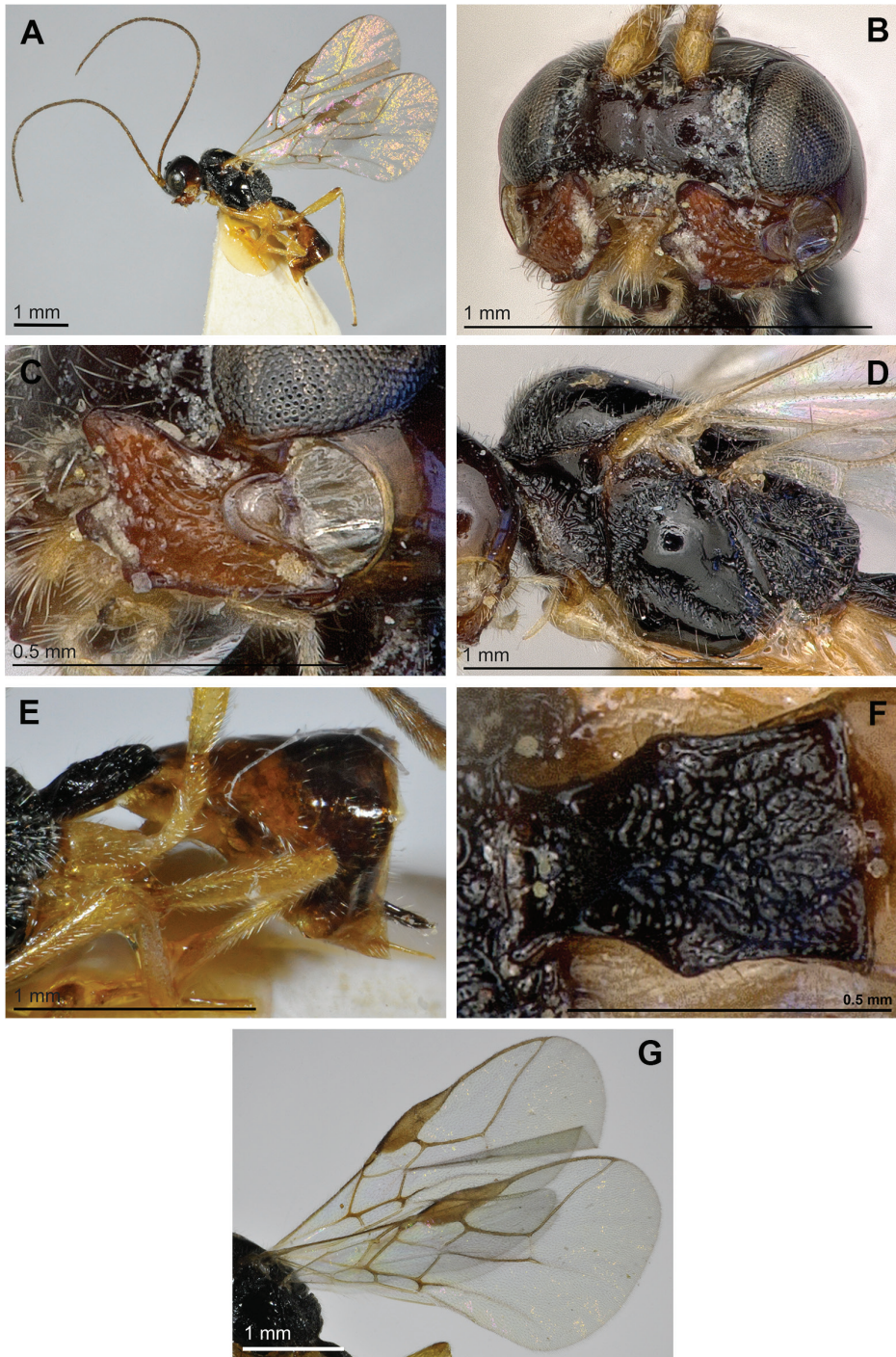


Figure 3. *Adeluroloa asiatica* Telenga (female). **A** Habitus, lateral view **B** Face, front view **C** Mandible **D** Mesosoma, lateral view **E** Metasoma and ovipositor, lateral view **F** First metasomal tergite **G** Fore and hind wings.

tered short setae in lateral areas, with low middle vertical protuberance, 2.1 times as wide as high; inner margins of eyes subparallel. Clypeus slightly convex ventrally, about twice as wide as high. Mandible broadened towards subapex, 0.9 times as long as its maximum width. Upper tooth of mandible broadened sideward, much longer than lower tooth; middle tooth wide basally and narrowed towards apex, rounded apically; lower tooth short, rounded apically. Antenna rather slender, 37-segmented. Scape 1.15 times as long as pedicel. First flagellar segment 3.1 times as long as its apical width; second segment 3.7 times as long as its maximum width, 1.2 times as long as first segment. Third flagellar segment about 3.0 times as long as its maximum width. Penultimate segment 2.2 times and apical segment 4.2 times as long as their maximum widths, respectively.

Mesosoma 1.5 times as long as high (lateral view). Mesoscutum smooth, punctate and densely setose in anterior part, 0.9 times as long as maximum width. Notauli coarsely crenulate, present in anterior half of mesoscutum, not reaching with mesoscutal pit. Mesoscutal pit present, very long, sparsely crenulate. Scutellar sulcus distinctly crenulate, with median carina but without lateral carinae. Sides of pronotum smooth in anterodorsal area, mainly rugose-reticulate. Precoxal suture present, wide, reaching anterior margin of mesopleuron but absent posteriorly. Posterior mesopleural furrow sparsely and widely crenulate below and shortly and densely crenulate in upper half. Propodeum completely rugose-reticulate, with numerous scattered setae. Propodeal spiracle relatively small.

Wings. Length of fore wing 2.3 times its maximum width. Pterostigma cuneate. Marginal cell distinctly shortened, reaching distinctly before apex of wing, 2.8 times as long as its maximum width. Vein 3-SR 1.7 times as long as vein 2-SR. Vein SR1 1.9 times as long as vein 3-SR. Second submarginal cell 3.3 times as long as maximum width. Vein cu-a distinctly postfurcal. Subdiscal cell closed, 2.3 times as long as its maximum width. Hind wing 4.0 times as long as its maximum width.

Legs. Hind femur 4.7 times as long as its maximum width. Hind tibia slightly widened towards apex, about 10.0 times as long as its maximum subapical width, as long as hind tarsus. First segment of hind tarsus 1.9 times as long as second segment.

Metasoma compressed laterally. First tergite completely and densely rugose-reticulate, without median carinae, slightly widened towards apex, 1.4 times as long as its apical width. Second metasomal tergite smooth. Ovipositor sheath 0.6 times as long as first tergite, 0.3 times as long as hind femur.

Colour. Body reddish brown to dark brown or black. Metasoma medially light reddish brown, apical segments dark brown. Legs mainly yellow, hind femur and tibia dark. Wings very faintly infusate. Pterostigma brown.

Body length 3.2 mm; fore wing length 3.0 mm.

Variation. Body length 3.4 mm; fore wing length 3.3 mm. Mandible 0.8 times as long as its maximum width; middle teeth distinctly reduced. Antenna 39-segmented. First flagellar segment 3.3 times as long as its apical width; second segment 3.4 times as long as its maximum width, 1.1 times as long as first segment. Mesoscutum about as long as its maximum width. Length of fore wing 2.3 times its maximum width. Marginal

cell 3.0 times as long as its maximum width. Vein 3-SR 1.55 times as long as vein 2-SR. Vein SR1 2.0 times as long as veins 3-SR. Second submarginal cell 3.4 times as long as its maximum width. Hind wing 4.3 times as long as its maximum width. Hind femur 5.3 times as long as its maximum width. First metasomal tergite 1.5 times as long as its apical width.

Male. Unknown.

Diagnosis. This species is similar to *A. amplidens* (Fischer, 1966) and *A. florimela* (Haliday, 1838). *Adeluroloa asiatica* differs from *A. florimela* by having the mandible 1.25 times as long as its maximum width (1.0 times in *A. florimela*), first metasomal tergite without median carinae (with median carinae in *A. florimela*), vein 3-SR 1.9 times as long as vein 2-SR (1.2 times in *A. florimela*), vein SR1 1.8 times as long as vein 3-SR (2.4 times in *A. florimela*), and precoxal suture reaching anterior and posterior margins of mesopleuron (not reaching anterior and posterior margins in *A. florimela*). Differences between *A. asiatica* and *A. amplidens* are in the re-description of the latter species.

Distribution. Uzbekistan, Turkmenistan, Kyrgyzstan (new record).

Adeluroloa florimela (Haliday, 1838)

Fig. 4

Alysia florimela Haliday 1838: 239; 1839: 25.

Adelura florimela: Foerster 1863: 267; Marshall 1894: 420; Dalla Torre 1901: 38; Lyle 1933: 74; Morley 1933: 183; Stelfox 1941: 2.

Alysia (Adelura) florimela: Thomson 1895: 2287.

Dapsilarthra florimela: Kloet and Hincks 1945: 239; Königsmann 1959: 589; Fischer 1970: 13; Shenefelt 1974: 988; van Achterberg 1997: 40; Wharton et al. 2006: 325.

Adeluroloa florimela: van Achterberg 1983: 5; Gurasashvili 1983: 784; Tobias 1986: 236; Belokobyl'skij 1998: 284; 2003: 356; Yu et al. 2012; Broad et al. 2012: 7; Riedel and Hansen 2014: 148.

Phaenocarpa multiarticulata Marshall 1898: 245.

Dapsilarthra multiarticulata: Shenefelt 1974: 989.

Dapsilarthra pentapleuroides Fischer 1971: 85 (as synonym of *A. multiarticulata*).

Material examined. Germany: 1 female, “*Adelura florimela* Hal. ♀”, “Schmiedeknecht dt.”; 2 males, without geographical labels, from Schmiedeknecht Collection. **Latvia:** 1 female, Valmier Region, Draudzība, ex larva of *Pegomyia hyoscyami* (Panzer) on beet, 6.vii.1962, V. Ozolinsh coll. (ZISP); 1 female, same label, but 14.07.1962 (ZISP); 1 female, same label, but 21.viii.1962 (ZISP). **Russia:** 1 female, 2 males, Leningrad Province, Kingisepp, 20 and 22.v.1904, Vinogradov-Nikitin coll. (ZISP); 1 female, Yaroslavl' Province, Bykovo, 25.v.1891, N. Kokuev coll. (ZISP); 1 female, Yamalo-Nenetsk Autonomous Region, Krasnosel'kup, Taz River, terrace, 15–17.viii.1992, D. Kasparyan coll. (ZISP); 1 male, Primorskiy Territory, 12 km S Khorol', forest, 4.vi.1979, S. Belokobyl'skij coll. (ZISP); 1 male, Primorskiy Territory, 8 km from Brovnichi, Ser-

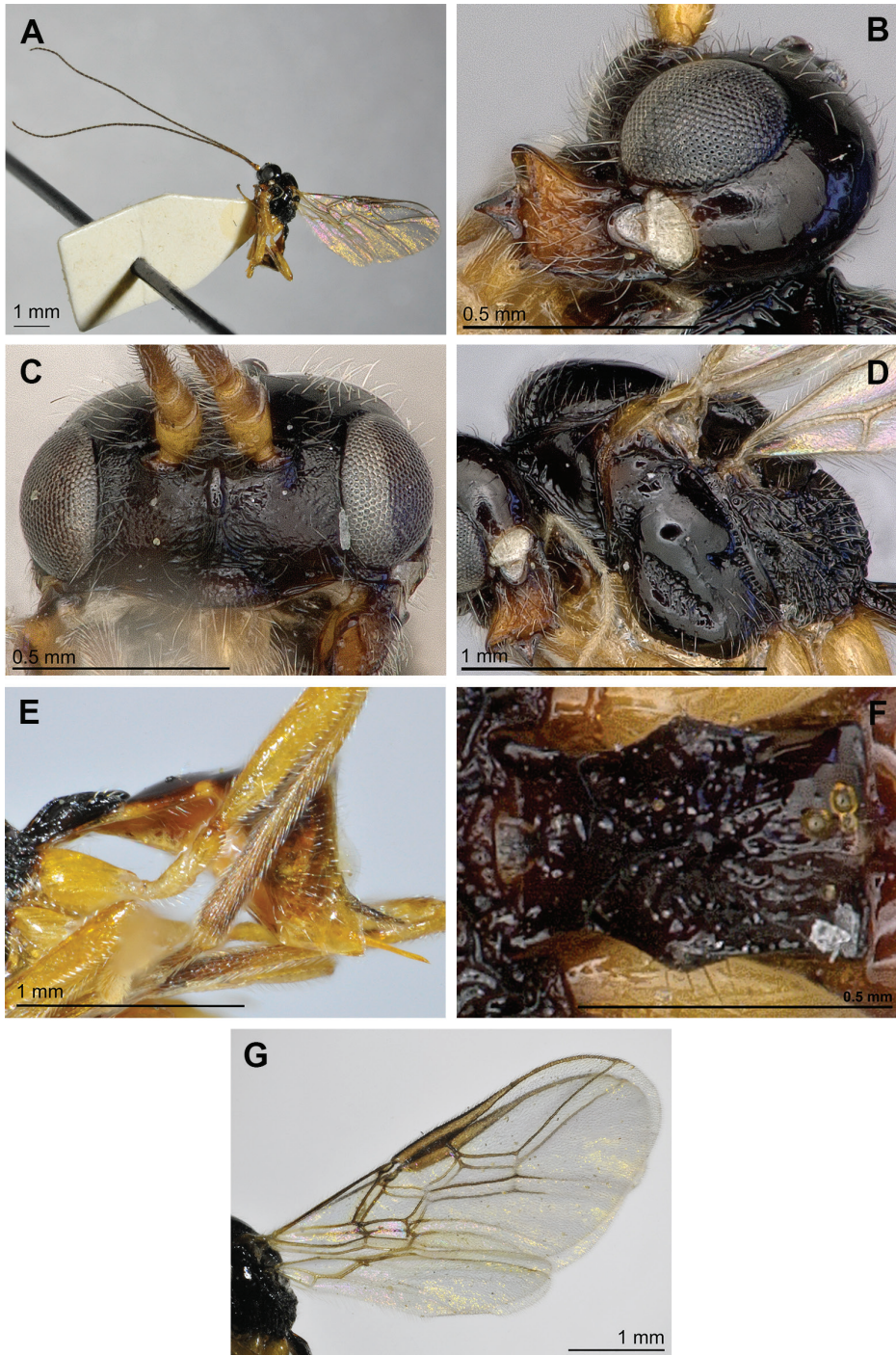


Figure 4. *Adelura florimela* (Haliday) (female). **A** Habitus, lateral view **B** Head (lateral view) and mandible **C** Face, front view **D** Mesosoma, lateral view **E** Metasoma and ovipositor, lateral view **F** First metasomal tergite **G** Fore and hind wings.

ebryanoe, 9.vi.1978, A. Kupyanskaya coll. (ZISP); 1 male, Primorskiy Territory, Vladivostok, Sedanka, 3.vi.1978, S. Belokobylskij coll. (ZISP); 1 female, Primorskiy Territory, 30 km SE Ussuriysk, Ussuriysk Nature Reserve forest, 10–11.vi.1993, S. Belokobylskij coll. (ZISP); 1 female, Sakhalin Island, Yuzhno-Sakhalinsk environs, Chekhov Mountain, 900 m, 28.vii.1988, A. Kotenko coll. (ZISP). **Georgia:** 1 female, Kazbegi, 2300 m, meadow, 16.viii.1982, M. Gurasashvili coll. (ZISP); for material in RMNH from the Netherlands, Germany and Bulgaria, see van Achterberg (1983).

Description. Female.

Head entirely smooth; in dorsal view 2.0 times as wide as median length, 1.5 times as wide as mesoscutum, with not convex rounded temples behind eye. Eye in lateral view 1.3 times as high as wide and 1.15 times as wide as temple medially. POL about as long as OD; OOL 3.0 times OD. Face rugulose medially, with scattered setae, with distinct and complete middle prominence, 2.0 times as wide as high; inner margins of eyes subparallel. Clypeus slightly curved ventrally, 3.0 times as wide as high. Mandible broadened towards subapex, 1.2 times as long as its maximum width, rugose. Upper tooth of mandible broadened sideward, distinctly longer than lower tooth; middle tooth wide basally and narrowed towards apex, pointed apically; lower tooth rounded apically. Antenna rather slender, 45-segmented. Scape 1.25 times as long as pedicel. First flagellar segment 3.3 times as long as its apical width; second segment 3.8 times as long as its maximum width, 1.1 times as long as first segment. Third flagellar segment 3.5 times as long as its maximum width. Penultimate segment about 2.0 times and apical segment 3.3 times as long as their maximum widths, respectively.

Mesosoma 1.4 times as long as high (lateral view). Mesoscutum entirely smooth, with dense setae along notauli and scattered setae laterally, as long as its maximum width. Notauli present in anterior half and absent in posterior half, crenulate. Mesoscutal pit present, short, elongate. Scutellar sulcus finely and sparsely rugulose, with distinct median carina but without lateral carinae. Sides of pronotum mainly smooth. Precoxal suture rather wide and rugulose, reaching anterior margin of mesopleuron, but absent posteriorly. Posterior mesopleural furrow completely crenulate. Propodeum completely rugose-reticulate. Propodeal spiracle small.

Wings. Length of fore wing 2.6 times its maximum width. Pterostigma cuneate. Marginal cell just not reaching apex of wing, 3.2 times as long as its maximum width. Vein 3-SR 1.3 times as long as vein 2-SR. Vein SR1 2.2 times as long as veins 3-SR. Second submarginal cell 3.6 times as long as its maximum width. Vein cu-a distinctly postfurcal. Subdiscal cell closed, 2.5 times as long as its maximum width. Hind wing 4.3 times as long as its maximum width.

Legs. Hind femur about 5.0 times as long as its maximum width. Hind tibia slightly widened towards apex, about 10.0 times as long as its maximum subapical width, 0.9 times as long as hind tarsus. First segment of hind tarsus 2.0 times as long as second segment.

Metasoma depressed dorso-ventrally. First tergite completely rugose-reticulate with median carina, hardly widened towards apex (subparallel), 1.6 times as long as its

apical width. Second metasomal tergite smooth. Ovipositor sheath 0.3 times as long as first tergite, 0.2 times as long as hind femur.

Colour. Body brown to dark reddish brown. Second metasomal tergite reddish brown, paler than first and apical tergites. Legs yellowish brown, hind tibia apically and most part of hind tarsus distinctly infusate. Wings almost hyaline. Pterostigma brown.

Body length 3.4 mm; fore wing length 3.9 mm.

Variation. Body length 3.1–4.2 mm; fore wing length 3.5–4.3 mm. Antenna 43–49-segmented. First flagellar segment 3.0–3.4 times as long as its apical width; second segment 3.7–4.2 times as long as its maximum width, 1.10–1.15 times as long as first segment. Third flagellar segment 3.5–4.0 times as long as its maximum width. Marginal cell of fore wing 2.8–3.1 times as long as its maximum width. Vein 3-SR 1.1–1.3 times as long as vein 2-SR. Vein SR1 2.2–2.6 times as long as vein 3-SR. Second submarginal cell 3.0–3.5 times as long as its maximum width. Subdiscal cell 2.1–3.0 times as long as its maximum width. Hind femur 4.7–5.0 times as long as its maximum width. First tergite 1.3–1.6 times as long as its apical width. Ovipositor sheath 0.3–0.6 times as long as first tergite, 0.2–0.4 times as long as hind femur.

Male. Body length 3.2–4.0 mm; fore wing length 3.6–4.6 mm. Mandible often brown to dark brown. Veins of fore wing more or less widened; pterostigma distinctly thickened and completely black or dark brown. Hind femur 5.0 times as long as its maximum width. First metasomal tergite narrow, 1.6–1.9 times as long as its apical width.

Diagnosis. *Adeluroloa florimela* (Haliday, 1838) differs from *A. amplidens* (Fischer, 1966), *A. asiatica* Telenga, 1935 and *A. kamtschatica* Belokobylskij, 1998 by the features listed in the diagnoses of each of these species, as listed above.

Hosts. *Acidia cognata* (Wiedemann, 1817) (Tephritidae), *Pegomya hyoscyami* (Panzer, 1809), *P. nigritarsis* (Zetterstedt, 1838) and *P. solennis* (Meigen, 1826) (Anthomyiidae) (van Achterberg 1983; Belokobylskij 1998; Yu et al. 2012).

Distribution. Austria, former Czechoslovakia, Finland, Georgia, Germany, Hungary, Ireland, Italy, Japan, Latvia, Lithuania, Netherlands, Poland, Russia, Slovenia, Spain, Sweden, Switzerland, United Kingdom, former Yugoslavia.

Adeluroloa kamtschatica Belokobylskij, 1998

Fig. 5

Adeluroloa kamtschatica Belokobylskij 1998: 285; Yu et al. 2012.

Type material. Holotype: female, Kamchatka, saddle of Avacha and Koryaka Volcanoes, 1000 m, mountain tundra, 27.vii.1985, S. Belokobylskij coll. (ZISP); 1 female (paratype), same label, but 26.vii.1985 (ZISP).

Description. Female.

Head entirely smooth; in dorsal view 2.0 times as wide as median length, 1.5 times as wide as mesoscutum, with rounded temples behind eye. Eye in lateral view 1.2 times as high as wide and 0.9 times as wide as temple medially. POL as long

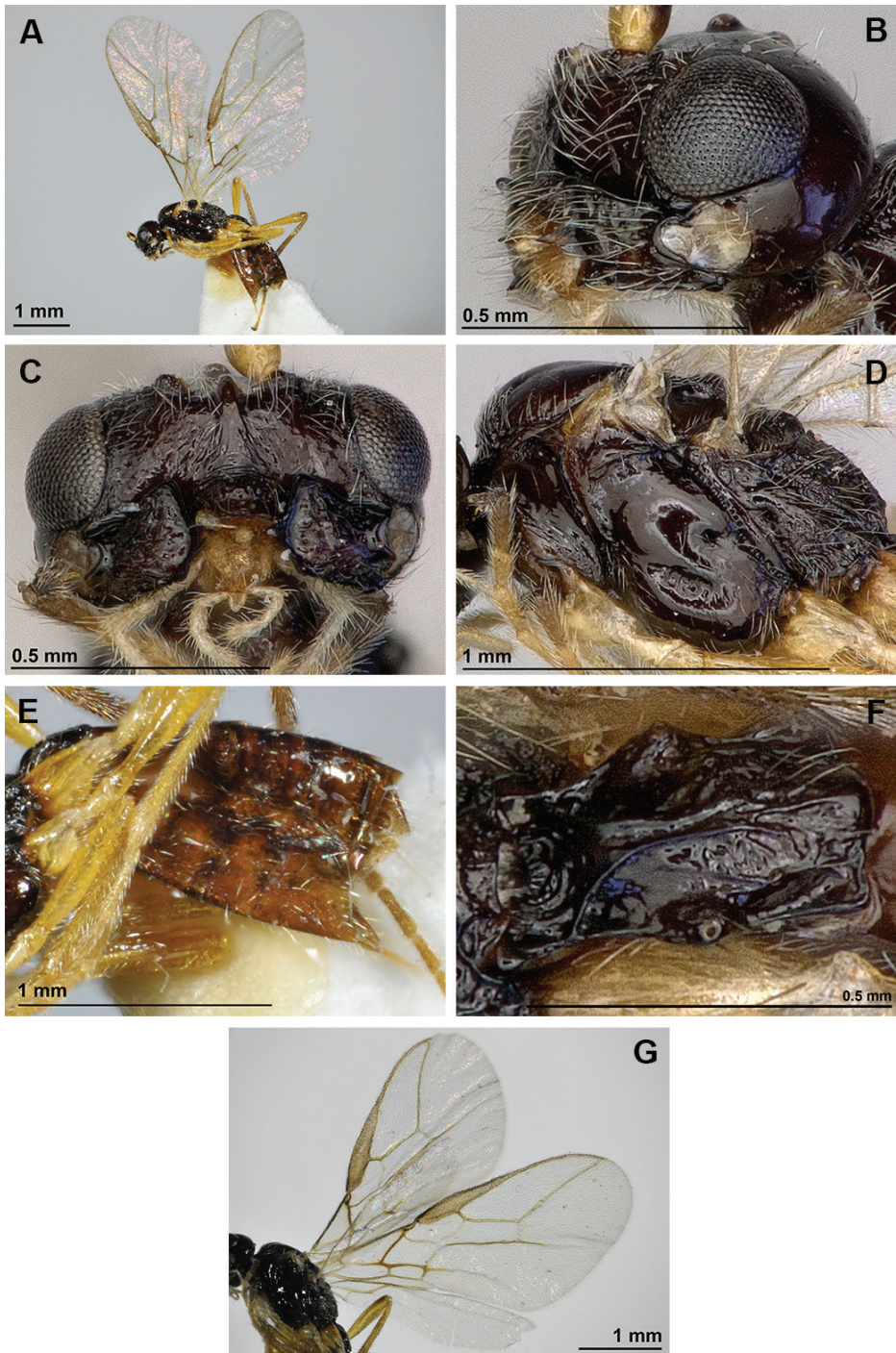


Figure 5. *Adeluroloa kamtschatica* Belokobylskij (female, paratype). **A** Habitus, lateral view **B** Head (lateral view) and mandible **C** Face in front view **D** Mesosoma, lateral view **E** Metasoma and ovipositor, lateral view **F** First metasomal tergite **G** Fore and hind wings.

as OD; OOL 4.3 times OD. Face faintly rugulose-striate, with rather dense setae, with complete middle prominence, 2.4 times as wide as high; inner margins of eyes subparallel. Clypeus distinctly curved ventrally, 2.0 times as wide as high. Mandible not strongly broadened towards subapex, 1.3 times as long as its maximum width, rugose. Upper tooth of mandible broadened sideward, distinctly longer than lower tooth; middle tooth wide basally and strongly narrowed towards apex, pointed apically; lower tooth rounded apically. Antenna slender, 45-segmented. Scape 1.9 times as long as pedicel. First flagellar segment 2.8 times as long as its apical width; second segment 3.6 times as long as its maximum width, 1.25 times as long as first segment. Third flagellar segment 3.1 times as long as its maximum width. Penultimate segment 2.4 times and apical segment 3.0 times as long as their maximum widths, respectively.

Mesosoma 1.4 times as long as high (lateral view). Mesoscutum smooth, its upper part with dense setae latero-anteriorly, 1.1 times as long as maximum width. Notauli absent in posterior half. Mesoscutal pit present, elongate. Scutellar sulcus smooth, without median and lateral carinae. Sides of pronotum mainly smooth. Precoxal suture present, not reaching anterior and posterior margins of mesopleuron, slightly crenulate. Posterior mesopleural furrow completely and shortly crenulate. Propodeum completely rugose-reticulate, with numerous scattered setae. Propodeal spiracle small.

Wings. Length of fore wing 2.2 times its maximum width. Pterostigma cuneate. Marginal cell reaching just before apex of wing, 3.5 times as long as its maximum width. Vein 3-SR 1.3 times as long as vein 2-SR. Vein SR1 2.6 times as long as vein 3-SR. Second submarginal cell 3.1 times as long as its maximum width. Vein cu-a postfurcal. Subdiscal cell closed, 2.6 times as long as its maximum width. Hind wing 4.3 times as long as its maximum width.

Legs. Hind femur 4.7 times as long as its maximum width. Hind tibia slightly widened towards apex, about 10.0 times as long as its maximum subapical width, as long as hind tarsus. First segment of hind tarsus 1.9 times as long as second segment.

Metasoma compressed laterally. First tergite slightly rugose-reticulate in apical half, with several striae, with median carinae, not widened towards apex (parallel subparallel), 1.6 times as long as its apical width. Second metasomal tergite smooth. Ovipositor sheath 0.7 times as long as first tergite, 0.4 times as long as hind femur.

Colour. Body dark brown dark reddish brown. Second and third metasomal tergites light reddish brown, apical tergites faintly pale. Legs yellowish brown, hind femur apically, hind tibia in apical half and most part of hind tarsus distinctly infusate. Wings hyaline. Pterostigma brown basally and pale brown apically.

Body length 2.9 mm; fore wing length 3.5 mm.

Variation. Fore wing length 3.4 mm. Head in dorsal view 1.9 times as wide as median length POL 0.8 times as long as OD. Face about twice as wide as high. Clypeus 2.5 times as wide as high. Mesosoma 1.5 times as long as high (lateral view). Length of fore wing 2.4 times its maximum width. Vein 3-SR 1.4 times as long as vein 2-SR. Vein SR1 twice as long as veins 3-SR. Second submarginal cell 3.4 times as long as

maximum width. Subdiscal cell 2.3 times as long as its maximum width. Hind femur 4.6 times as long as its maximum width. First tergite 1.5 times as long as its apical width. Ovipositor sheath 0.8 times as long as first tergite.

Male. Unknown.

Diagnosis. This species is similar to *A. florimela* (Haliday, 1838), but differs from it in having the scutellar sulcus smooth and without complete median carina (sculptured and with complete median carina in *A. florimela*), first flagellar segment shorter (longer in *A. florimela*), mandible slightly widened towards apex and its upper tooth smaller and less protruding upwards (distinctly widened and with large upper tooth in *A. florimela*), precoxal suture finely and narrow rugulose (distinctly and widely rugose in *A. florimela*), pterostigma paler, pale brown (darker, brown in *A. florimela*).

Distribution. Russia (Far East).

Key to the world species of *Adeluroloa*

- 1 Mandible slightly widened towards apex (Fig. 5B), upper tooth smaller and less distant up (Fig. 5B). Prescutellar sulcus without median carina and almost smooth. Precoxal suture finely and narrowly rugulose (Fig. 5D). Body length 2.8–2.9 mm ***A. kamtschatica* Belokobylskij**
- Mandible distinctly widened towards apex (Figs 2B, 3C, 4B), upper tooth larger and more distant up (Figs 2B, 3B, 4B). Prescutellar sulcus with median carina and sculptured. Precoxal suture distinctly and widely rugose (Figs 2D, 3D, 4D)..... **2**
- 2(1) First flagellar segment 0.8 times as long as second segment. Precoxal suture reaching anterior and usually posterior margins of mesopleuron (Fig. 3D). Third mandibular tooth small or very small (Fig. 3C). Body length 3.2–3.4 mm..... ***A. asiatica* Telenga**
- First flagellar segment 0.95–1.05 times as long as second segment. Precoxal suture short, not reaching anterior and posterior margins of mesopleuron or reaching only anterior margin (Fig. 2D, 4D). Third mandibular tooth rather large (Figs 2B, 4B) **3**
- 3(2) Eye in lateral view 1.2 times as wide as temple medially (Fig. 4B). Metasoma mainly depressed dorso-ventrally (Fig. 4E). Marginal cell almost reaching apex of wing, 2.8–3.2 times as long as its maximum width (Fig. 4F). Vein 3-SR 1.1–1.3 times as long as vein 2-SR (Fig. 4F). Vein SR1 2.2–2.6 times as long as vein 3-SR (Fig. 4F). Body length 3.1–4.2 mm..... ***A. florimela* (Haliday)**
- Eye in lateral view 0.8 times as wide as temple medially (Fig. 2B). Metasoma more or less compressed laterally (Fig. 2E). Marginal cell remaining far from apex of wing, 2.5 times as long as its maximum width (Fig. 2F). Vein 3-SR 2.0 times as long as vein 2-SR (Fig. 2F). Vein SR1 1.8 times as long as vein 3-SR (Fig. 2F). Body length 3.5–3.9 mm ***A. amplidens* (Fischer)**

Excluded species

Dapsilarthra euryis (Chen & Wu, 1994), comb. n.

Adeluroloa euryis Chen and Wu 1994: 19, 154; Yu et al. 2012.

Type material. Holotype: female (studied the images), China, “Xianfengling, Mt. Wuyi, Fujian, 2.viii.1986, Liu Minghui” (Beneficial Insects Institute, Fujian Agricultural University, Fuzhou, China).

Remarks. After examining of the holotype images of *A. euryis* we surely considered that this species actually belongs to the genus *Dapsilarthra* by the absence of a ventral lamelliform lobe on mandible and elongated the first flagellar segment of antenna.

Discussion

Adeluroloa is exclusively Palaearctic small genus of parasitoid wasps of subfamily Alysiinae closely related to *Dapsilarthra*. If *A. florimela* (Haliday) is very widely distributed in the Palaearctic Region (from U.K. till Russian Far East), then other three species have local Asian distribution on the territories of Middle East [*A. amplidens* (Fischer)], Central Asia (*A. asiatica* Telenga) and Kamchatka Peninsula (*A. kamtschatica* Belokobylskij). Additionally this paper includes the first records of *Adeluroloa* species, *A. amplidens* and *A. asiatica* Telenga, from Iran and Kyrgyzstan.

Unfortunately, the hosts of *Adeluroloa* taxa are unknown yet. However published in this paper information is valuable one owing to significant role of many Alysiinae taxa in the regulation of the natural dipterans populations mainly from families Anthomyiidae and Tephritidae.

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Revision of the genus *Menevia* Schaus, 1928 (Lepidoptera, Mimallonoidea, Mimallonidae) with the description of 11 new species

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Abstract

The Neotropical genus *Menevia* Schaus, 1928 is revised to include 18 species, 11 of which are new. Two species, *M. ostia* **comb. n.** and *M. parostia* **comb. n.** are transferred from *Pamea* Walker, 1855 to *Menevia*. Four species-groups are diagnosed for the first time based on external characters and male genitalia morphology. The following new species are described: *M. rosea* **sp. n.**, *M. torvameatoria* **sp. n.**, *M. magna* **sp. n.**, *M. menapia* **sp. n.**, *M. mielkei* **sp. n.**, *M. australis* **sp. n.**, *M. vulgaris* **sp. n.**, *M. franclemonti* **sp. n.**, *M. vulgaricula* **sp. n.**, *M. cordillera* **sp. n.**, and *M. delphinus* **sp. n.**. A neotype is designated for *Mimallo plagiata* Walker, 1855, which has since been placed in *Menevia*. *Mimallo saturata* Walker, 1855 is interpreted to be a *nomen dubium*.

Keywords

Mimallo, Neotropical, neotype, *Pamea*

Introduction

Until recently, very little revisionary work has been done with the family Mimallonidae. Schaus (1928) was the last to revise the family completely, describing most of the genera that are currently recognized. In addition to organizing the family by erecting numerous genera, Schaus (1928) also separated Mimallonidae into two subfamilies. This subfam-

ily arrangement, however, has been deemed by most contemporary authors (Pearson 1951, 1984, Franclemont 1973, Herbin 2012, St Laurent and Dombroskie 2015, but see Becker 1996) not to reflect their phylogeny. We continue to recognize the lack of a clear subfamily arrangement awaiting a higher-level treatment of the family.

Adding to the poor understanding of the higher-level arrangement of the family are a few weakly diagnosed genera currently persisting as catch-alls to subsume numerous recently described species (Herbin 2012, Herbin and Mielke 2014, Herbin 2015). The present taxonomic treatment examines one of the more morphologically distinct genera, *Menevia* Schaus, 1928, and provides autapomorphies for the genus so that new species described herein can be accurately placed.

The genus *Menevia* currently consists of five Central and South American species: *M. plagiata* (Walker, 1855), *M. lantona* (Schaus, 1905), *M. lucara* (Schaus, 1905), *M. alurca* Herbin & Mielke, 2014, and *M. pallida* Herbin & Mielke, 2014. No synonyms or other names have been formerly assigned to *Menevia*. We diagnose the genus *Menevia* based on external characters, re-describe all five currently known species, describe each previously undescribed female, describe 11 additional new species, and move two species currently assigned to the genus *Pamea* Walker, 1855 into *Menevia*.

Methods

Dissections were performed as described by Lafontaine (1987) except when same-day analysis was required, whereby abdomens were heated in a 10% KOH solution for 20–30 minutes. Not all genitalia were prepared on slides to allow for three-dimensional analysis of the complex male genitalia. Genitalia and abdomens, when not slide mounted, are preserved in glycerol filled microvials. Morphological, including genitalia, terminology follows Lemaire and Minet (1999).

Costa Rican material of *M. ostia* was obtained from the biodiversity inventory of the Área de Conservación Guanacaste (ACG) (Janzen et al. 2009, Janzen and Hallwachs 2011). Some specific localities within the ACG that do not appear in gazetteers are local names, but the provided GPS coordinates give the true location. Full information content for all ACG specimens can be found at <http://janzen.sas.upenn.edu> by searching the voucher code (Janzen and Hallwachs 2009).

The holotypes and one neotype were all dissected or, when present, previously made genitalia preparations were examined.

Specimens from the following collections were examined:

AMNH	American Museum of Natural History, New York, New York, USA
CGCM	Collection of Carlos G. C. Mielke, Curitiba, Paraná, Brazil
CMNH	Carnegie Museum of Natural History, Pittsburgh, Pennsylvania, USA
CNC	Canadian National Collection of Insects, Arachnids and Nematodes, Ottawa, Ontario, Canada

CPAC	Coleção Embrapa Cerrados, Planaltina, Distrito Federal, Brazil
CUIC	Cornell University Insect Collection, Ithaca, New York, USA
DZUP	Collection of Pe. Jesus S. Moure, Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil
FSCA	Florida State Collection of Arthropods, Gainesville, Florida, USA
MEM	Mississippi Entomological Museum, Mississippi State, Mississippi, USA
MGCL	McGuire Center for Lepidoptera & Biodiversity, Gainesville, Florida, USA
MNHN	Muséum nationale d'Histoire naturelle de Paris, France
MNHU	Museum für Naturkunde der Humboldt-Universität zu Berlin, Germany
NHMUK	Natural History Museum [formerly British Museum (Natural History)], London, U.K.
OM	Collection of Olaf Hermann Hendrik Mielke, Curitiba, Paraná, Brazil
RAS	Research collection of Ryan A. St. Laurent, Ithaca, New York, USA
USNM	National Museum of Natural History [formerly United States National Museum], Washington D.C., USA

The symbol ‡ will be used to represent unavailable names in the text (Fletcher and Nye 1982).

Figures were manipulated with Adobe Photoshop CS4 (Adobe 2008). Male genitalia are figured in natural color with CS4 “auto color” used to improve white backgrounds. Female genitalia were treated with “auto tone” in CS4 to darken characters; insets, however, are manipulated only with “auto color.” Most adult specimens were photographed in natural light with an Apple iPhone 5S, additional adult photos were provided by CGCM and NHMUK. Genitalia were photographed with a Macroscopic Solutions Macropod Pro and Canon EOS 6D DSLR camera body using the Macro Photo MP-E 65mm f/2.8 1–5× Manual Focus Lens for EOS. Thirty (3×) photographs were taken of each specimen in ethanol under glass, and stacked using Zerene Stacking Software. Maps were created with SimpleMappr (Shorthouse 2010) and edited with CS4. All geographical coordinates are approximate, and are based on the localities provided on specimen labels. GPS data were acquired with Google Earth.

Results and discussion

Menevia Schaus, 1928

Type species. *Cicinnus lantona* Schaus, 1905; Schaus 1928: 665, by original designation.

Diagnosis. *Menevia* can be recognized by the contrast between the usually gray submarginal area and the darker gray, brown, yellowish, or rarely pink medial area of the forewing and the presence of a white apical dash. This apical dash becomes the “postmedial lunule” (see Fig. 1), which in most species-groups consists of a variably distinct white line originating from the apical dash. The white mark follows the postmedial line from near the apex to one quarter or to one half the length of the postmedial

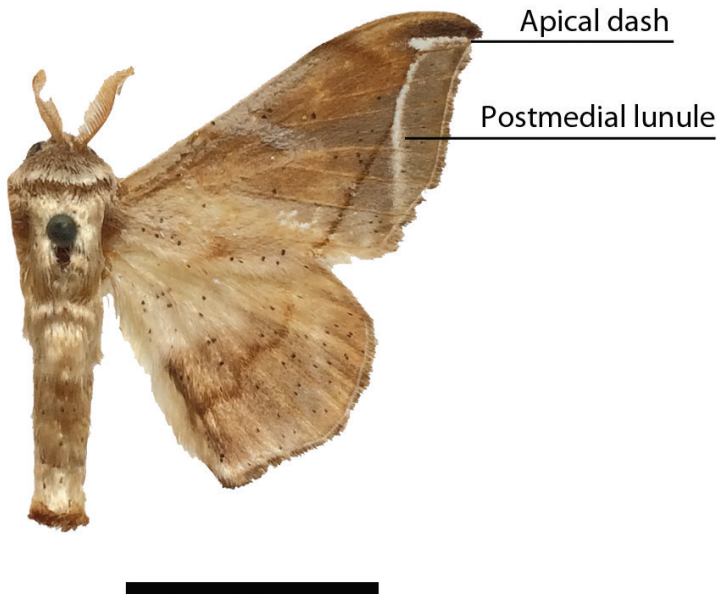


Figure 1. Important forewing characters. *Menevia lantona*, male, Suriname, Moengo, Boven Cottica River (CUIC). Scale bar = 1 cm.

line until sweeping outward toward the wing margin, either at an acute angle or nearly perpendicular to the postmedial line. In the *plagiata* species-group only does the postmedial lunule not sweep outward from the postmedial line, but follows it as a white outlining band that may be interrupted. In other genera where markings similar to the postmedial lunule exist, they are not white and contrasting against a gray submarginal area. Finally, the male genitalia are also unique among Mimallonidae. They are complex structures (see Fig. 2) with a pair of distinct, elongated tusks pointing outwards, originating from the modified transtilla that itself extends inward into the body from attachments on either side of the inner costal apodemes of the valves. The elongated tusks pass outward between a pair of weakly sclerotized setose flaps. The juxta is fused to the phallus, encircling it, and is without clear form except for a pair of juxtal processes, which curve toward the distal end of the phallus and are superior to it. Phallic shape is diverse, but always recognizable by the presence of the attached juxtal processes dorsally. The phallus is longitudinally rolled, roughly forming a “U” in cross section, and is open lengthwise along the dorsum where the edges of the rolled phallic structure do not meet. The dorsal, left edge of the phallus is usually uneven, with extensive ridges or protuberances of varying size and shape.

Description. Male. Head: Small, scales on frons swept ventrad, either the same color as vertex or darker ventrally, eyes large comprising about half to two-thirds of head area, eyes usually bordered posteriorly by darker collar of scales reaching labial palpi, labial palpi small, segments variably defined ventrally depending on thickness of vestiture, incrementally smaller in length distally, dorsally and laterally with darker

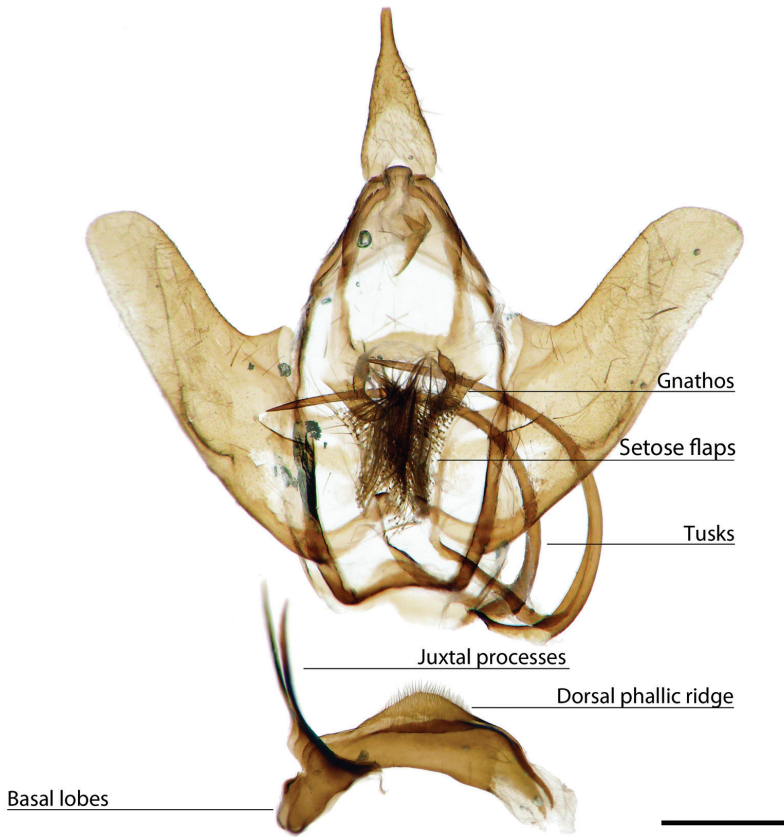


Figure 2. Important male genitalia characters. *Menevia mielkei*, holotype male, Brazil, Minas Gerais, Estação Biológica de Caratinga, Caratinga [CGCM diss.: OM 61.563] (DZUP). Photo courtesy of CGCM. Scale bar = 1 mm.

scales contrasting with overall lighter coloration of head. Antenna bipectinate to tip, scape and pedicel tufted. Ocelli and chaetosemata absent. *Thorax*: Tan, gray, or straw colored. Densely covered in scales of varying widths with interspersed darker petiolate scales, scales of prothoracic collar finer, lighter, overlapping scales of mesothorax. *Legs*: Vestiture thick, scales long, especially on femur and tibia, coloration as for thorax, petiolate scales present. Tibial spurs often scaled, about one fourth length of tibia, apex may be somewhat hooked. *Forewing dorsum*: Forewing length: 14–28 mm. Triangular, outer margins concave on apical half, apex usually falcate. Ground color yellow, brown, gray, or gray-brown, lightly or moderately speckled by dark petiolate scales. Discal spot absent or faintly marked by light gray, no hyaline patches present. Dark postmedial line always present, either straight or with slight undulations. Gray submarginal area usually contrasting with medial area, submarginal area with a variably distinct white line originating from apical dash, white mark follows postmedial line from apex to one quarter to one half the length of postmedial line until sweeping outward toward wing margin,

either at an acute angle or nearly perpendicular to postmedial line. In some species, white line wider, forming a complete or interrupted band following postmedial line, not sweeping toward wing margin. Antemedial line, if present, faint and undulated. *Forewing venter*: As in forewing dorsum but postmedial line may be fainter, antemedial line absent, discal spots may be much darker. *Hindwing dorsum*: Rounded or subtriangular, anal angle often accentuated, similar coloration and patterning as forewings, vague postmedial lunules present but undulated or sharply zigzagged, never dramatically swept to wing margin, antemedial line absent. *Hindwing venter*: Following similar pattern as forewing venter, usually lighter, frenulum with single bristle. *Wing venation*: As for *Cicinnus melsheimeri* (Harris, 1841) in Franclemont (1973) but M_2 originating closer to M_3 near posterior, distal corner of cell. *Abdomen*: Short, subtriangular, reaching just barely beyond anal margin of hindwing, depth equal to that of thorax, truncated to slightly upturned posterior tip, coloration a continuation of thoracic color, which varies from yellowish to brown, generally matching the ground color of wings. Longitudinal midventral stripe present or absent. *Genitalia*: Complex; tegumen variable in shape, subtriangular or broader and more rectangular or ovoid, often constricted near base of gnathos. Vinculum usually quadrate ventrally, variable in thickness. Transtilla as rectangular frame extending backward from attachments on either side of the inner costal apodemes of valves, elongated tusks extend outward originating from complex transtilla, passing between pair of setae covered flaps. Valves simple, narrow or broad, triangular or ovoid, with or without projections from saccular edge and with or without similar mesal costal projections, projections of left valve usually larger than those of right valve. Setae covered uncus teardrop or bottle-shaped, or acutely triangular, extended apically to sharp, rounded, or quadrate tip. Gnathos as two prominent outward facing or upturned extensions, variable in shape, thickness. Gnathos extensions thick and boxing glove shaped, flattened and triangular, subtriangular, cupped, or ovoid. Anal tube lightly sclerotized, with apex nearly reaching base of uncus. Juxta fused to phallus, enveloping proximal quarter of phallus, pair of juxtal processes curve toward the distal end of phallus and are superior to it. Juxtal processes variable in length, from roughly three quarters the length of the phallus to slightly longer. Juxtal processes vary in sclerotization; processes flattened, rounded or sharp distally. Processes always enveloped in membrane, usually covered in fine setae. Base of phallus with paired, elongated, short, or peg-like diverging lobes. Phallus usually complicated, formed by singular structure rolled into a cylindrical shape, dorsal edges of phallus do not meet, gap exists between edges, exposing hollow cavity within containing the vesica. Left edge of rolled phallus variously shaped, either flat and simple or with ridge or protuberance dorsally. Distal tip of phallus separated into two distinct points. Vesica very weak, bag-like or somewhat elongate and tapered apically, cornuti absent. **Female**. Generally similar to male, degree of sexual dimorphism variable. *Head*: As in male, antennae sometimes smaller overall. *Thorax*: As in male. *Legs*. As in in males except tibial spurs sometimes more heavily scaled. *Forewing dorsum*: Forewing length: 15–39 mm. Elongated or subtriangular, outer margins concave near apex, convex near tornus, apex usually falcate. Coloration and markings as in corresponding males of each species. *Forewing venter*: As in forewing

dorsum but postmedial line may be fainter, antemedial line absent, discal spots may be much darker than in males. *Hindwing dorsum*: As in males but always more rounded, broader. *Hindwing venter*: Following similar pattern as forewing venter, frenulum present with multiple bristles or highly reduced. *Abdomen*: As in male but stouter, presence of midventral stripe corresponds to presence/absence in conspecific males, sternites of VIII as pair of elongated sclerotized bands of varying width and length. *Genitalia*: Very simple; VIII prominently sclerotized laterally, sometimes with appendicular apophyses dorsolaterally in addition to apophyses anteriores. Tergite of VIII upturned mesally, posteriorly pointed, smoothly curving, or replaced by bulbous sac-like sclerotized structure. Apophyses anteriores slightly shorter or same length as apophyses posteriores. Lamella antevaginalis of varying width and shape, usually forming a semicircle or "V". Ostium bursae unsclerotized. Ductus bursae usually short. Corpus bursae bag-like, without any sclerotized structures, rarely preserved, appendix bursae elongated. Papillae anales quadrate or slightly elongated, subtriangular, covered in fine setae.

Remarks. The genus *Menevia* consists of four clear species-groups, which are delineated here for the first time. The species-groups are readily differentiated by the adult morphology; namely the degree of sexual dimorphism, size, forewing maculation and shape, ground color, and male genitalia characteristics.

Key to species-groups of *Menevia*

- 1 Forewing submarginal area with white, curved, lunule originating from apical dash (Fig. 1)..... **2**
- Forewing postmedial lunule replaced by continuous or interrupted white band along exterior edge of postmedial line ***plagiata* species-group**
- 2 Ground color light tan, yellow, or orange, contrasting with gray submarginal area, postmedial lunule usually very weakly curved toward wing margin, forewings weakly falcate..... **3**
- Ground color mixture of deep red-brown and dark gray, not strongly contrasting with gray submarginal area except for when medial area more red-brown than gray, postmedial lunule sharply curved toward wing margin, forewings falcate ***lucara* species-group**
- 3 Sexual dimorphism pronounced. Gray submarginal area highly contrasting with yellowish or golden medial area, submarginal area always with small accessory white mark near tornus, hindwing postmedial line mostly straight. Female: forewings broad, nearly ovoid; male: phallus always with dorsal ridge or variously shaped protuberance ***ostia* species-group**
- Male and female hardly differentiable. Gray submarginal area moderately or sometimes only weakly contrasting with light tan, yellowish, orange, or rarely pinkish medial area, small white accessory mark near tornus absent in most species, hindwing postmedial line usually wavy, especially near anal margin. Male: phallus irregularly shaped, usually without dorsal ridge, but when present, anteriorly situated, triangular..... ***lantona* species-group**

***lantona* species-group**

The *lantona* species-group, containing the type species *M. lantona* by original designation, is the only species-group with very weak sexual dimorphism. This group includes the following species: *M. lantona*, *M. rosea* sp. n., *M. torvameessoria* sp. n., and *M. magna* sp. n. The females of two species belonging to this group were available for study, and both are exceptionally reminiscent of the males, with only slightly more elongated forewings and meagerly larger size. Species belonging to the *lantona* species-group are generally small for the genus, containing some of the smallest species of *Menevia*. The shape of the forewings of both males and females are not overly elongated, barely falcate, with weakly curving to almost straight postmedial lunules, and with a light ground color, which is usually yellowish with light shades of brown and tan. One species however, is mostly pink. The flat, triangular, or cupped processes of the gnathos and the usually broad phallus characterize male genitalia of this species-group.

Key to *lantona* species-group

- 1 Ground color orange, yellow-tan, or dark tan, never with any pink suffusion, dorsum of phallus variable **2**
- Ground color suffused with pink, phallus with well-defined dorsal protuberance ***M. rosea* sp. n.**
- 2 Forewing postmedial line straight or convex, faint or very thick and contrasting, abdomen never with midventral stripe. Panama and northern South America.. **3**
- Forewing postmedial line with slight inward kink on inferior half, phallus with triangular dorsal ridge, venter of abdomen with quarter-length or complete midventral stripe. Southeastern Brazil ***M. magna* sp. n.**
- 3 Ground color orange-yellow, forewing postmedial line very dark and contrasting, genitalia with tegumen nearly circular, uncus hooked, phallus pistol shaped and bent mesally, tubular and elongated distally..... ***M. torvameessoria* sp. n.**
- Ground color light tan, fading to yellow in older specimens, forewing postmedial line usually thin and not highly contrasting against surrounding color, tegumen rectangular or somewhat ovoid, especially when prominently constricted at base of gnathos, uncus barely hooked, phallus irregularly shaped but not bent mesally or tubular distally..... ***M. lantona***

***Menevia lantona* (Schaus, 1905)**

Figs 1, 6–10, 72, 92; Map 1

Cicinnus lantona Schaus, 1905: 327–328

Cicinnus lantona; Dyar 1914

Cicinnus lantona; Lucas 1915

Menevia lantona; Schaus 1928: fig. ♂ 88f

Menevia lantona; Forbes 1942: Plate 14, fig. 102

Menevia lantona; Becker 1996

Menevia lantona; Herbin and Mielke 2014: figs ♂ 57, 58

Type material. Holotype, ♂: **FRENCH GUIANA**: St. Jean, Maroni, F. Guiana/ Collection Wm. Schaus/ *Perophora lantona* type Schaus/ Type No.: 8894 U.S.N.M./ USNM-Mimal: 1122/ St. Laurent diss.: 3-7-15:1/ (USNM) [examined]. No paratypes. Type locality: French Guiana: St. Jean du Maroni.

Additional specimens examined. (65 ♂, 1 ♀ total) **BRAZIL: Amazonas:** 1 ♂, Reserva Ducke, km. 26 Manaus-Itacoatiara Highway: 19.V.1972, E.G., I. & E.A. Munroe, St. Laurent diss.: 3-9-15:1 (CNC). 1 ♂, Codajás: IV.1907, S.M. Klages, Rothschild Bequest, BM 1939-1 (NHMUK). 2 ♂, Fonte Boa: V.1906, IX.1906, S.M. Klages, Rothschild Bequest, BM 1939-1 (NHMUK). **COLOMBIA:** 2 ♂, Muzo, 400–800 m: Coll. Fassl, ex. Joicey Coll. Brit. Mus, 1925–157, BMNH(E) 1378744, St. Laurent diss.: 6-29-15:7 (NHMUK); Dognin Collection, USNM-Mimal: 2569, St. Laurent diss.: 3-7-15:7 (USNM). 1 ♂, East Cordillera, Muzo: A. & E. Fassl, 1911 (MNHU). **ECUADOR:** 2 ♂, Napo, Rio Napo, Biol. Sta. Jatun Satcha, 380–400 m: 12–16. IV.1990, S.J. Weller, P. Batra, & M.J. Ryan, USNM-Mimal: 2727, St. Laurent diss.: 3-7-15:8, 7-7-15:4 (USNM). 1 ♂, Napo, 33 km. N. Tena, 27 km. E. on Loreto Rd., 3600 ft: 2.XI.1998, black light, leg. J.S. Miller, St. Laurent diss.: 7-7-15:3 (AMNH). **FRENCH GUIANA:** 10 ♂, 1 ♀, St. Jean du Maroni: Collection Wm Schaus, one male with label: “*M. lantona* Schaus topotype”, female with label: USNM-Mimal: 2574 and St. Laurent diss.: 7-8-15:1 (USNM); 16.I.1978, Porion (MNHN); Received from E. LeMoult, Rothschild Bequest, BM 1939-1 (NHMUK); VII–VIII.1904, E. LeMoult, Rothschild Bequest, BM 1939–1 (NHMUK); “1905-14”, Rothschild Bequest, BM 1939-1 (NHMUK). 1 ♂, Pied Saut, Oyapok River: XII.1917, S.M. Klages, C.M. Acc. 6111, St. Laurent diss.: 3-7-15:2 (CMNH). 2 ♂, St. Laurent du Maroni: IV, Dognin Collection, USNM-Mimal: 2571, St. Laurent diss.: 3-7-15:4 (USNM); II.1906, E. LeMoult, Rothschild Bequest, BM 1939–1 (NHMUK). 1 ♂, Regine Rte de l’Est. km 65, el. 100 m: 11.II.1991, at lights, Coll. C. Snyder (AMNH). 1 ♂, Piste Coralie, PK 2: 11.VI.1993, L. & A. Sénécaux (MNHN). 1 ♂, Route de Regina, PK 62, Piste de Coralie, PK 2.2: 24.VII.1998, H. de Toulgoët & J. Navatte (MNHN). 1 ♂, Route de Regina, PK 62, Auberge des Orpailleurs: 23.VII.1998, H. de Toulgoët & J. Navatte, Mus. nat. Hist. nat. don de H. de Toulgoët (MNHN). 1 ♂, Piste de Kaw, PK 7: 4.VIII.1996, H. de Toulgoët & J. Navatte réc. (MNHN). 1 ♂, Piste de Nancibo: 5.XI.1983, UV, J. Minet & D. Dauthuille (MNHN). 2 ♂, Roura, Route de Kaw, Near Fourgassie, N04°38.643', W52°17.988', 280 m: 26.X.2014, 28.X.2014, P. Sammut leg. (CUIC). 2 ♂, Cayenne, Patawa, N04°32.024', W52°07.684', 275 m: 27.X.2014, P. Sammut leg. (CUIC; RAS). **GUYANA:** 1 ♂, Tumatumari, Rio Potaro, Br. Guiana: Ac. 5615 (AMNH). 1 ♂, Tumatumari: XII.1907, S.M. Klages, Rothschild Bequest, BM 1939–1 (NHMUK). **PANAMA:** 2 ♂, La Cabima: V.1911 [1931?], August Busck, USNM-Mimal: 2567, 2568, St. Laurent diss.: 3-7-15:10



Figure 3. *Menevia* in resting position. *Menevia australis* sp. n., Brazil, Santa Catarina, Laurentino. Photo credit: Miguel Angelo Biz, used with permission.

(USNM). 1 ♂, La Chorrera: “May 12”, Aug. Busck, USNM-Mimal: 2566, St. Laurent diss.: 3-7-15:11 (USNM). 21 ♂, Barro Colorado Island, Canal Zone: 16.IV.1941, X.1941, 10.X.1941, 19.X.1941, [other data illegible], at light, J. Zetek collector, Nos. 4884, 4884A,B, USNM-Mimal: 2716–2721 (USNM); 28–30.IV.1964, 1–9.V.1964, 10–17.V.1964, W.D. & S.S. Duckworth, USNM-Mimal: 2709–2715, St. Laurent diss.: 3-7-15:13 (USNM); 22.IV.1979, at light, Silberglied/Aiello, USNM-Mimal: 2724, 2725, 2726, St. Laurent diss.: 3-7-15:14 (USNM); 8.V.1935, A. Friedman (CUIC); 25.X, 31.X, 3.XI, 7–XI, M. Bates coll., St. Laurent diss.: 3-7-15:12 (CUIC). 2 ♂, “Rep. de Panama”: XII.1935–I.1936, L.M. Smith, J.G. Franclemont diss.: 1770 (CUIC). **SURINAME:** 1 ♂, Moengo, Boven Cottica River: 19.V.1927, Cornell Univ. Lot 760, Sub 67, St. Laurent diss.: 3-7-15:3 (CUIC). 2 ♂, Aroewarwa Creek, Maroewym Valley: III.1905, IV.1905, S.M. Klages, Rothschild Bequest, BM 1939–1 (NHMUK). **VENEZUELA:** 1 ♂, Caripito: 14.V.1942, Gift of New York Zoo. Soc. Dept. Tropical Research, William Beebe, Dir., St. Laurent diss.: 3-7-15:6 (AMNH).



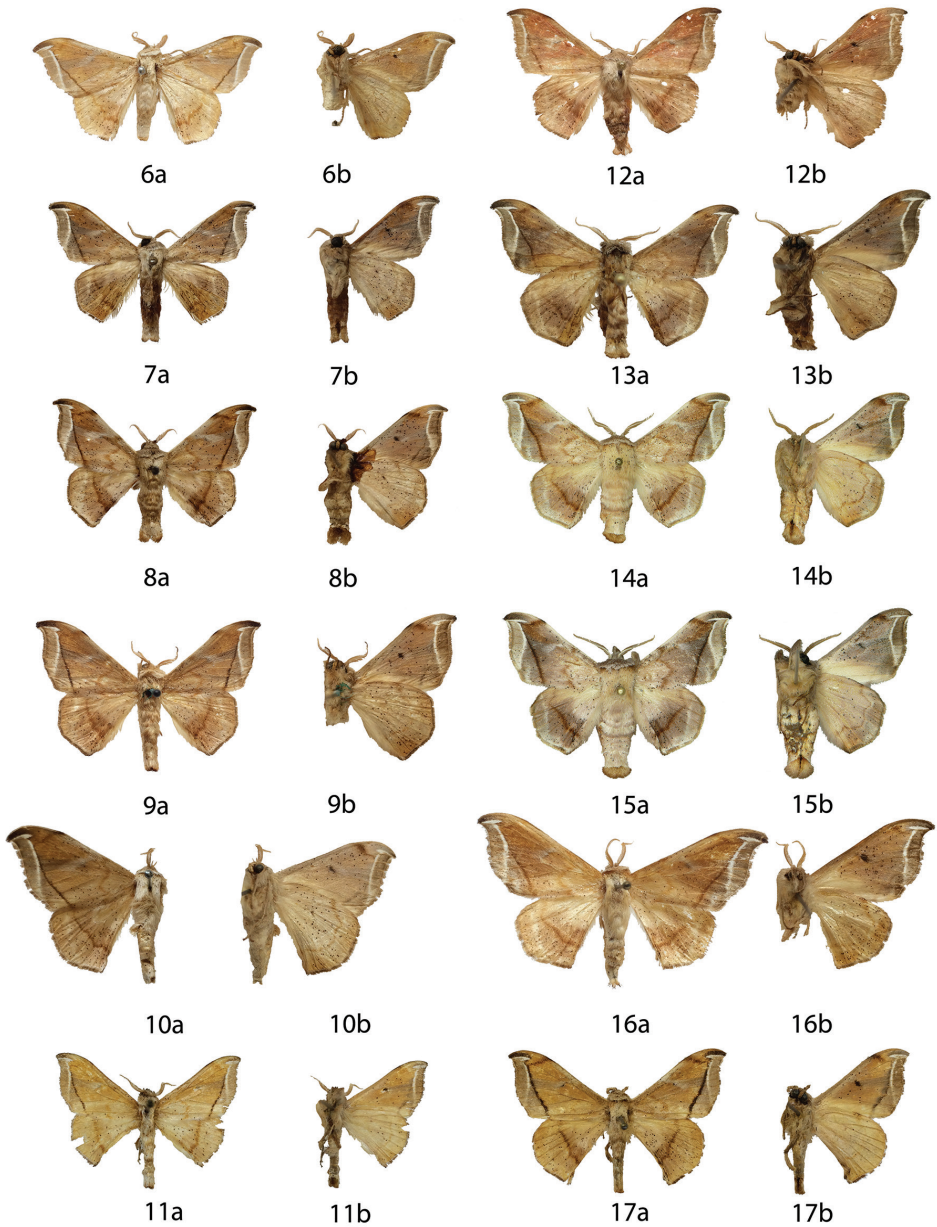
Figure 4. *Menevia ostia* larva with case, lateral view. On unknown host plant, larval length = 33 mm, 13-SRNP-71165. Image courtesy of D. H. Janzen & W. Hallwachs, used with permission.



Figure 5. *Menevia ostia* larva with case, anterior view. As for Fig. 4.

Diagnosis. Sexual dimorphism is weak; the females are only slightly larger. Thus both sexes of *M. lantona* are recognizable by their small to moderate size, only slightly falcate forewing apices, and yellowish tan to gold ground color with gray highlights, especially near the discal region. The postmedial lunule is bright white and only barely curved outward toward the forewing margin, not sharply curving as in other species-groups. The phallus is broad, usually with a small protuberance dorsally but always without a dorsal ridge. The gnathos processes are unique in that they are very broad, flat, and subtriangular, not oblong or thick as in most other species-groups. Female genitalia are separable from those of the similar female of *M. magna* sp. n. by the tergite of VIII, which is thin and upturned mesally rather than being replaced by a lightly sclerotized sac as in *M. magna* sp. n.

Description. Male. *Head:* Straw or tan colored, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally due to ventral tufts, dorsally with darker scales contrasting with overall straw coloration. Scape and pedicel weakly tufted. *Thorax:* As for genus but tan or gold, fading to straw. *Legs:* As for genus. Tibial spurs thin apically, terminal third not scaled, especially ventrally. *Forewing dorsum:* Forewing length: 15–18 mm, avg.: 16.2 mm, n = 28. Triangular, apical half of outer margins concave, convex near tornus, apex slightly falcate. Ground color yellowish tan to gold with varying degrees of gray, especially near discal region, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above apical dash. Straight or slightly undulating postmedial line black or brown. Antemedial area lighter, submarginal area gray with tan coloration near tornus, postmedial lunule originating from near where apical dash meets postmedial line, lunule follows postmedial line from apex to one third length of postmedial line where lunule smoothly curves outward toward wing margin, forming roughly 45 degree angle with postmedial line. Antemedial line very faint or absent, if present, brown, undulating. *Forewing venter:* As in forewing dorsum but postmedial line fainter, antemedial line absent; usually rounded discal spot present, small, black. *Hindwing dorsum:* Rounded with margin weakly pointed mesally, anal angle weakly accentuated, similar coloration and patterning as forewings, vague postmedial lunule originating near anterior margin undulating, not steeply swept to margin, antemedial line absent, postmedial line undulating, especially near anal angle. *Hindwing venter:* Following similar pattern as forewing venter but discal mark not always present. *Abdomen:* As for genus. Coloration a continuation of tan or golden thoracic color. Midventral stripe absent. *Genitalia:* (Fig. 72) n = 16. Tegumen rectangular or somewhat ovoid, especially when prominently constricted at base of gnathos. Vinculum narrow, somewhat quadrate ventrally. Valves asymmetrical, relatively narrow, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth much reduced in size, both valves with smaller mesal costal teeth immediately above saccular edge teeth, apex of mesal tooth pointed toward saccular edge. Valves rounded apically. Uncus handbell-shaped, truncated apically, apex rounded. Gnathos as two prominent, moderately flattened, subtriangular outward facing flaps, upturned where they converge over phallus. Juxtal processes shorter than phallus, flattened, slightly curved, covered in short setae. Base of phallus with



Figures 6–17. *Menevia lantona* species-group adults, **a** recto, **b** verso. **6** *M. lantona* holotype ♂, French Guiana, St. Jean du Maroni (USNM) **7** *M. lantona* ♂, French Guiana, Cayenne, Patawa, 275 m (RAS) **8** *M. lantona* ♂, Panama (CUIC) **9** *M. lantona* ♂, Colombia, Muzo, 800 m (USNM) **10** *M. lantona* ♀, French Guiana, St. Jean du Maroni (USNM) **11** *M. torvameatoria* holotype ♂, Peru, Puno, La Unión, 2000 ft (NHMUK) **12** *M. rosea* holotype ♂, Ecuador, Napo, Simón Bolívar, 1200 m (CMNH) **13** *M. magna* holotype ♂, Brazil, Santa Catarina, São Bento do Sul, Rio Natal, 450 m (DZUP) **14** *M. magna* paratype ♂, Brazil, São Paulo, Guapiara, Paivinha, 800 m [photo courtesy CGCM] (CGCM) **15** *M. magna* paratype ♂, Brazil, São Paulo, Apiaí, 750 m [photo courtesy CGCM] (CGCM) **16** *M. magna* paratype ♀, Brazil, Santa Catarina, Dalbérgia (CUIC) **17** *M. torvameatoria* paratype ♂, Peru, Puno, S. Domingo, 6500 ft (NHMUK). Scale bar = 1 cm.

paired, somewhat elongated, rounded, diverging, backwards facing fingerlike lobes. Phallus broad, widened mesally, usually with a small protuberance dorsally but extended dorsal ridge absent. Left edge of rolled phallus simple, without ridge-like process, distal tip of phallus separated into two distinct, bent points. Vesica small, bag-like.

Female. *Head:* As in male, antennae slightly smaller overall. *Thorax:* As in male. *Legs:* As in male. *Forewing dorsum:* Forewing length: 21 mm, $n = 1$. As in male but barely more elongated, only slightly broader, less falcate, postmedial line bent mesally. Faint brown antemedial line present, undulated. *Forewing venter:* As in forewing dorsum but postmedial line fainter, antemedial absent, small black discal spot present. *Hindwing dorsum:* As in male but slightly broader. *Hindwing venter:* Following similar pattern as forewing venter except lighter, frenulum reduced. *Abdomen:* As in male but stouter. Sternites of VIII as pair of elongated sclerotized bands widening toward anterior margin of VIII. *Genitalia:* (Fig. 92) $n = 1$. Tergite of VIII very thin, converging mesally to form anteriorly directed point. Apophyses anteriores slightly shorter than apophyses posteriores. Lamella antevaginalis ribbon-like, weakly concave mesally. Ductus bursae thin. Papillae anales rectangular when viewed ventrally, covered in setae.

Distribution (Map 1). *Menevia lantona* is found throughout the Amazonian rainforest in the Guianas, Suriname, northern Venezuela, the Brazilian state of Amazonas, as well as in central Colombia and northcentral Ecuador. This species also ranges into central Panama.

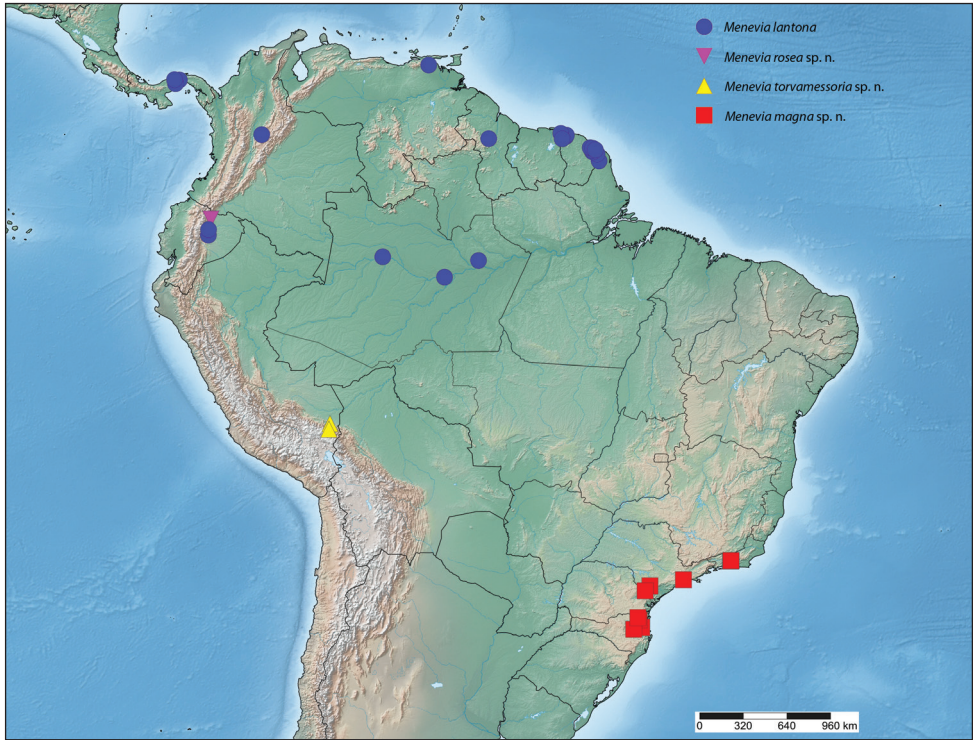
Remarks. As a wide-ranging species, *M. lantona* expresses some geographic variation. Most notably, specimens from Colombia are larger (Fig. 9) with somewhat broader forewings, but genitalia show no remarkable differences. Specimens from Panama (Fig. 8) are slightly grayer and a bit darker overall, but like the Colombian populations, show no genitalia differences specifically applicable to this geographic form. Additionally, in specimens from Ecuador, the dorsal point of the phallus is somewhat more pronounced than in other populations, and one specimen from a relatively high elevation (1097 m) in Ecuador has much darker postmedial lines than most other individuals of *M. lantona*, somewhat reminiscent of *M. torvameatoria* sp. n., to be described below. The genitalia of this unique higher elevation individual however, are completely in line with those of *M. lantona*, and cannot be mistaken for the highly remarkable and distinct genitalia of *M. torvameatoria* sp. n.

***Menevia rosea* sp. n.**

<http://zoobank.org/5E2B3DC6-6B92-48EF-958D-6F67D144563F>

Figs 12, 73; Map 1

Type material. **Holotype**, ♂: **ECUADOR:** Ecuador: Napo, Simon Bolivar [Simón Bolívar], Coca River canyon, 1200 m, 16 Aug 1996, Jan Hillman, undisturbed wet forest/ St. Laurent diss.: 3-7-15:9/ HOLOTYPE male *Menevia rosea* St Laurent and Dombroskie, 2016 [handwritten red label]/ (CMNH). No paratypes. Type locality: Ecuador: Napo: Simón Bolívar.



Map I. *Menevia lantona* species-group.

Diagnosis. *Menevia rosea* is distinguishable from all other species in the *lantona* species-group by the pink coloration of the forewings, especially medially and proximal to the apical region. The postmedial lunule is also more weakly curved than in the most similar species, *M. lantona*. Genitalia characters should also readily distinguish *M. rosea* from other species in the *lantona* species-group. The particularly short phallus has a small, but obvious dorsal ridge, which is lacking in *M. lantona*. The phallic ridge of *M. rosea* should not be confused with the dorsal protuberance on the phallus of some *M. lantona*, as this protuberance is closer to mid-length of the phallus while the phallic ridge of *M. rosea* is more distal. The juxtal processes are more curved toward the distal end of the phallus. The valves are more triangular and are particularly broad proximal to the vinculum, the saccular edges of the valves are also particularly straight.

Description. Male. Head: Brownish gray with pinkish hue, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally due to ventral tufts, dorsally with darker scales contrasting with overall pinkish gray coloration. Scape and pedicel weakly tufted. **Thorax:** As for genus but light tan, scales of prothoracic collar pinker, tipped with white. **Legs:** As for genus. Tibial spurs very thin, relatively long, terminal third not scaled, especially ventrally. **Forewing dorsum:** Forewing length: 16 mm, n = 1. Triangular, apical half of outer margins concave, apex slightly falcate. Ground color light tan with excessive pink scal-

ing, especially medially and nearing apex before postmedial line, very sparsely speckled by dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above small apical dash. Dark brown postmedial line mostly straight, somewhat undulating. Antemedial area lighter, less pink, submarginal area faint gray to more tan near tornus, postmedial lunule originating perpendicular to where apical dash meets postmedial line, lunule follows postmedial line from apex to nearly half length of postmedial line where lunule barely curves outward toward wing margin, forming very acute angle with postmedial line. Antemedial line absent. *Forewing venter*: As in forewing dorsum, pink coloration widespread but postmedial line fainter, more curved, antemedial line absent, small black discal spot present. *Hindwing dorsum*: Rounded with margin weakly pointed mesally, anal angle very weakly accentuated, similar coloration and patterning as forewings but with more petiolate scales, vague postmedial lunule originating near anterior margin undulating, not steeply swept to margin, antemedial line absent, postmedial line mostly straight, brown, surrounded by pink. *Hindwing venter*: Following similar pattern as forewing venter, but lighter. *Abdomen*: As for genus. Coloration a continuation of tan coloration of thorax with pink hue. *Genitalia*: (Fig. 73) $n = 1$. Tegumen rectangular, not constricted at base of gnathos. Vinculum moderately broad, somewhat quadrate ventrally. Valves asymmetrical, triangular, very broad at base, with very straight saccular edge except for large triangular tooth proximal to transtilla on left saccular edge, right valve with tooth much reduced in size, both valves with smaller mesal costal projection immediately above saccular edge teeth, apex of mesal projection pointed outwards. Valves rounded apically. Uncus handbell-shaped, truncated apically, apex rounded. Gnathos as two prominent, flattened, vaguely subtriangular outward facing flaps, upturned where flaps converge over phallus. Juxtal processes shorter than phallus, curved, creased along length, covered in moderately long setae. Base of phallus with paired, somewhat elongated, rounded, diverging, backwards facing fingerlike lobes. Phallus broad, short, widened mesally, with small dorsal ridge anteriorly. Left edge of rolled phallus simple, with small ridge-like process, distal tip of phallus separated into two distinct, straight points. Vesica small, sac-like. **Female**. Unknown.

Distribution (Map 1). *Menevia rosea* is so far known only from the type locality in Ecuador, Napo, Simón Bolívar, at a rather high elevation for the genus, 1200 m.

Etymology. *Menevia rosea* is named for the unique pink coloration of this species, unlike other species of the *lantona* species-group, which are usually tan.

Remarks. This new species is unique externally and is the only species in the *lantona* species-group with pink scales on the wings. Pink scaling is seen in many other *Menevia* species, particularly those in the *lucara* species-group. However, all other characteristics of patterning and the genitalia of *M. rosea* perfectly match those characters typical of the *lantona* species-group.

Apart from the interesting external coloration and genitalia characters, the type locality of this species is noteworthy mostly due to the relatively high elevation, 1200 m. The most similar species, *M. lantona*, has been collected in the Napo province just 80 km south of the type locality of *M. rosea*. Furthermore, the specimen collected 80

km south of the *M. rosea* type locality was collected at 1097 m elevation, also high for the genus. We have attributed this particular specimen to *M. lantona* due to the complete lack of pink and the genitalia characters. It is worth noting however, that this specimen has much darker postmedial lines on the fore and hindwings than typical *M. lantona*. Although the external characters and elevation of the collecting site are somewhat unique, the genitalia characters (St. Laurent diss.: 7-7-15:3) are completely typical of *M. lantona*, and are unlike either *M. rosea* or the other suspected possibility, *M. torvameatoria* sp. n. described below. Other Ecuadorian *M. lantona* are from much lower altitudes, 380–400 m. The differences in genitalia and the fact that *M. lantona* has been recorded from rather high elevations suggest that *M. rosea* is not a mere high elevation form of *M. lantona*.

***Menevia torvameatoria* sp. n.**

<http://zoobank.org/68607BD2-04A6-41E5-92F5-C8F709869950>

Figs 11, 17, 74; Map 1

Type material. Holotype, ♂: **PERU**: La Union [La Unión], R. Huacamayo, Carabaya [Puno], 2000 ft., wet s., Nov. 1904 (G. Ockenden)/ Rothschild Bequest BM 1939–1/ St. Laurent diss.: 6-29-15:4/ BMNH(E) 1378762/ HOLOTYPE male *Menevia torvameatoria* St Laurent and Dombroskie, 2016 [handwritten red label] (NHMUK). Type locality: Peru: Puno: Carabaya: La Unión.

Paratypes, 2 ♂: **BRAZIL: Pará**: 1 ♂, Monte Cristo, Rio Tapajós: Dognin Collection, USNM-Mimal: 2576, St. Laurent diss.: 3-7-15:5 (USNM). **PERU**: 1 ♂, S. Domingo, Carabaya [Puno], 6500 ft.: XII.1902, wet s., “591”, Rothschild Bequest BM 1939–1, St. Laurent diss.: 6-29-15:5, BMNH(E) 1378761 (NHMUK). – All paratypes with the following yellow label: PARATYPE male *Menevia torvameatoria* St Laurent and Dombroskie, 2016.

Diagnosis. Externally, *M. torvameatoria* is similar to *M. lantona*, but can be distinguished by the darker, yellow-orange ground color (in well-preserved specimens); *M. lantona* is lighter, more yellow-tan. Additionally, the postmedial line is very dark and contrasting in *M. torvameatoria* and there is a roughly rectangular gray patch of scales that extends from the discal region to the postmedial line. There is a similar gray patch in *M. lantona* but is not so well defined. The most outstanding diagnostic features of this new species are in the male genitalia. The phallus is unlike any other in the genus, it is almost pistol shaped and sharply bent halfway along its length. The dorsal surface of the proximal end of the phallus bears a distinct triangular or rounded ridge while the remainder of the length of the phallus is smooth, elongated, and tubular. The juxtal processes are very thin, shorter and more curved in other species in the *lantona* species-group. The gnathos processes are unique in that they are cupped and circular. The valves are unlike the previous species in that they are symmetrical and do not bear teeth on the saccular edge, but instead have distinct sclerotized inward facing lobes at the base of the valves, which conceal the gnathos. The sides of the tegumen are greatly

bowed outwards, causing it to appear almost circular. Finally, the scythe-like uncus is acutely triangular and sharply hooked.

Description. Male. *Head:* Brownish tan or almost black, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments moderately well defined ventrally due to ventral tufts, dorsally with darker scales contrasting with overall lighter coloration. Scape and pedicel thinly tufted. *Thorax:* As for genus but light tan, fading to straw. *Legs:* As for genus. Tibial spurs relatively small, only lightly scaled, especially proximally. *Forewing dorsum:* Forewing length: 14–16.5 mm, avg.: 14.8 mm, $n = 3$. Triangular, apical half of outer margin concave, apex slightly falcate. Ground color orange-yellow with elongate, gray rectangle of scales extending from discal region to postmedial line, overall very sparsely speckled by dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above scythe-like apical dash. Slightly undulating postmedial line black, strongly contrasting. Antemedial area lighter, submarginal area gray with slight invasion of medial area coloration near tornus, postmedial lunule originating from near where apical dash meets postmedial line, forming scythe-like dash, lunule follows postmedial line from apex to one third length of postmedial line where lunule smoothly curves outward toward wing margin, forming roughly 45 degree angle with postmedial line. Antemedial line very faint or absent, if present, brown, undulating, bowed out near anal margin. *Forewing venter:* As in forewing dorsum but postmedial line fainter, undulations more distinct, antemedial line absent, small black, rounded or oblong discal spot present. *Hindwing dorsum:* Rounded, anal angle weakly accentuated, similar coloration and patterning as forewings, but postmedial lunule almost nonexistent, antemedial line absent, postmedial line more undulated and brown, not black and contrasting, undulations prominent, especially near anal angle. *Hindwing venter:* Following similar pattern as forewing venter, but lighter, discal spot much less distinct or absent. *Abdomen:* As for genus. Coloration a continuation of tan thoracic color. Midventral stripe absent. *Genitalia:* (Fig. 74) $n = 3$. Tegumen almost circular, sides bowed out dramatically. Vinculum somewhat broad, quadrate ventrally. Transtilla tusks relatively short, thick, bent. Valves symmetrical, base of valves each with bulbous lobe pointed inward, partially covering gnathos. Valves bent mesally. Uncus extremely truncated apically, apex hooked, scythe-like. Gnathos projections as pair of cupped, rounded flaps, situated behind inward facing extensions at base of valves. Juxtal processes shorter than phallus, thin, curved. Base of phallus with paired, short, rounded, diverging, ventrally angled lobes. Phallus pistol shaped, sharply bent mesally, dorsal surface of proximal end of phallus with distinct triangular or rounded ridge, remainder of phallus thin, elongated, and tubular. Left edge of rolled phallus simple, without ridge-like process except for rounded or triangular ridge proximally, distal tip of phallus separated into two distinct points. Vesica small, sac-like. **Female.** Unknown.

Distribution (Map 1). *Menevia torvameatoria* is known from only three locations, one of them being questionable. Two specimens, including the holotype, come from two nearby localities in the Carabaya Province of Peru, in the Cordillera Oriental. The type locality, La Unión, is at about 609 m elevation, while the paratype from nearby

Santo Domingo was collected at a rather high elevation for the genus, about 1981 m. From these two data points it seems logical to infer that *M. torvamelessoria* is a species of moderate elevation, apparently from the Andean Cordillera Oriental. However, a third specimen questionably from Brazil (not shown on Map 1), is discussed in the remarks below.

Etymology. *Menevia torvamelessoria* is named for the unique hooked uncus, reminiscent of the Grim Reaper's (=torva messor Latin) scythe. The name is doubly appropriate to describe the apical dash, which combined with the postmedial lunule of the forewing, is scythe-like, a character seen in all *Menevia* species.

Remarks. Although externally *M. torvamelessoria* is rather similar to *M. lantona*, this new species is wholly unlike any other in the genus when taking into account male genitalia. Every aspect of the genitalia, particularly the hooked uncus, circular tegumen, cupped gnathos processes, bulbous projections at the base of the valves, and the shape of the phallus, are all unique to this species. *Menevia torvamelessoria* belongs in the genus *Menevia* due to the presence of the general structure of the genitalia such as the paired gnathos, juxtal processes, and outward facing tusks; but it is difficult to assign this taxon to a species-group based on genitalia alone. External characters, however, such as the size, orange-yellow coloration, and weakly falcate forewings, tentatively allow placement of *M. torvamelessoria* in the *M. lantona* species-group.

In addition to the unusual genitalia, the distribution is also strange, but this may be due to one specimen being incorrectly labeled. One paratype bears a nearly illegible label reading "Monte Cristo, Rio Tapajós, Amazonas" and seems to be from Monte Cristo, in the Brazilian state of Pará, on the Tapajós River. This particular location is very low in elevation, with some hills only as high as about 300 m nearby (as determined from Google Earth), which is quite divergent from the Andean foothill localities of the holotype and paratype. Either the specimen is mislabeled or *M. torvamelessoria* is very widespread in South America, and apparently very rare. Regardless of the uncertainty of the collecting locality of this paratype, its genitalia display the very unique and bizarre characteristics of the specimens from southern Peru and the ground color of this specimen is the same distinctive orange-yellow of *M. torvamelessoria*, thus we include this individual in the type series given that there are so few examples of this species available.

***Menevia magna* sp. n.**

<http://zoobank.org/2B8D7774-B967-46AB-A825-32D9D12F0727>

Figs 13–16, 75, 93; Map 1

Type material. Holotype, ♂: **BRAZIL: Santa Catarina:** BRAZIL – SC, São Bento do Sul, Rio Natal, 450 m. I. Rank leg, 27.VI.2014, S 26°20'2", W 49°18'30" / St. Laurent diss.: 6-16-15:1/ DZ 32.694/ HOLOTYPE male *Menevia magna* St Laurent and Dombroskie, 2016 [handwritten red label]/ (DZUP). Type locality: Brazil: Santa Catarina: São Bento do Sul.

Paratypes, 22 ♂, 3 ♀: **BRAZIL: Santa Catarina:** 1 ♂, Blumenau: X, Pohl, “696”, USNM-Mimal: 2582, St. Laurent diss.: 3-7-15:16 (USNM). 1 ♂, Corupá: VIII.1958, Maller col., No. HRP 889, “*Menevia lanthona*? Schaus Pearson det.”, USNM-Mimal: 2416, St. Laurent diss.: 3-7-15:17 (USNM). 1 ♂, 1 ♀, Nova Bremen [Dalbérgia]: 4.X.1935, 8.VII.1936, Fritz Hoffmann, St. Laurent diss.: 9-7-14:3, 3-7-15:15 (CUIC). 1 ♂, Rio Vermelho [São Bento do Sul], 830 m: VI.1936, A. Maller (NHMUK). 3 ♂, São Bento do Sul, Rio Natal, 550 m: XI.2013, A. Rank leg., Col. C. Mielke 28.001, 28.020, 28.040 (CGCM). 2 ♂, São Bento do Sul, Rio Natal, S 26°20'2", W 49°18'30", 450 m: 27.VI.2014, I. Rank leg., Col. C. Mielke 29.334, 29.422 (CGCM). **São Paulo:** 1 ♂, Alto da Serra: VI.1926, R. Spitz, Rothschild Bequest BM 1939–1, St. Laurent diss.: 6-29-15:6, BMNH(E) 1378759 (NHMUK). 3 ♂, Guapiara, Paivinha, 800 m: 18.VII.2007, C. Mielke leg., Col. C. Mielke 25.826, 26.555, St. Laurent diss.: 6-16-15:2 (CGCM). 3 ♂, Apiaí, 750 m: 18–21.VI.2007, C. Mielke leg., Col. C. Mielke 20.950, 21.350, 21.382 (CGCM). **Rio de Janeiro:** 3 ♂, 1 ♀, Petrópolis: 10.X.1961, 19.II.1963, 25.II.1963, 19.XII.1965, Gagarin leg., ex. col. Gagarin, CGCM diss.: DZ 32.695, DZ 32.695–32.697, 32.722 (DZUP). 2 ♂, 1 ♀, Petrópolis, Independência, 900 m: 19.VI.1934, 16.V.1935, 23.XI.1939, Gagarin leg., ex. col. Gagarin, DZ 32.698–32.700 (DZUP). 1 ♂, Petrópolis, Parque São Vicente: 5.XI.1958, DZ 32.701 (DZUP). – All paratypes with the following yellow label: PARATYPE male/female *Menevia magna* St Laurent and Dombroskie, 2016.

Diagnosis. *Menevia magna* is similar to other species in the *lanthona* species-group, except *M. rosea*, due to the tan ground color, but can be easily recognized by the larger size on average, the darker gray and brown scaling overall (especially in fresh specimens), and the usually mesally kinked forewing postmedial line. Additionally, the male genitalia are unique in that the valves are very broad and ovoid, the paired processes of the gnathos very thin and fingerlike or acutely triangular, and the smoothly curved phallus with a prominent, sharp, triangular dorsal ridge. Female genitalia are unique in the genus in having a bulbous, sac-like, sclerotized structure dorsally, replacing the usual sclerotization of tergite VIII, which also distinguishes females of *M. magna* from the very similar female of *M. lanthona*.

Description. Male. Head: Tan to gray, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally due to ventral tufts, dorsally with very dark scales contrasting with overall tan-gray coloration. Scape and pedicel weakly tufted. **Thorax:** As for genus, but tan, fading to straw. **Legs:** As for genus. Tibial spurs short, mostly scaled. **Forewing dorsum:** Forewing length: 17–19.5 mm, avg.: 17.8 mm, n = 6. Triangular, apical half of outer margin concave, apex falcate. Ground color dark tan-brown fading to lighter yellowish, grayer medially, overall lightly speckled with dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above apical dash. Dark brown postmedial line straight with slight inward kink mesally. Antemedial area lighter, submarginal area gray with very little tan coloration near tornus, postmedial lunule originating from near where apical dash meets postmedial line, lunule follows postmedial line from apex to one third length of postmedial line where lunule angled outward toward

wing margin, forming roughly 45 degree angle with postmedial line. Antemedial line usually present, brown, undulated. *Forewing venter*: As in forewing dorsum but postmedial nearly absent, may be more heavily speckled with petiolate scales, antemedial line absent, small black, rounded or oblong discal spot present. *Hindwing dorsum*: Rounded with margin weakly pointed mesally, anal angle accentuated, similar coloration and patterning as forewings, vague postmedial lunules originating near anterior margin undulating or nearly straight, antemedial line absent, postmedial line undulating, especially near anal angle. *Hindwing venter*: Following similar pattern as forewing venter but lighter, discal spot absent. *Abdomen*: As for genus. Coloration a continuation of tan thoracic color. Midventral stripe weak, either extending from posterior tip to about one quarter length of abdomen or until thorax. *Genitalia*: (Fig. 75) $n = 7$. Tegumen gumdrop shaped, not constricted at base of gnathos. Vinculum narrow, somewhat quadrate ventrally. Valves very broad, ovoid, asymmetrical, saccular edge of left valve with triangular tooth proximal to transtilla, right valve with tooth much reduced in size. Valves rounded apically. Uncus somewhat triangular, truncated apically, apex slightly hooked. Gnathos as two, fingerlike or acutely triangular outward facing extensions. Juxtal processes shorter than phallus, flattened, slightly curved, setae absent. Base of phallus with paired, short, rounded or pointed, diverging, backwards facing lobes. Phallus broad, widened mesally, with prominent, sharp or subtriangular dorsal ridge. Left edge of rolled phallus with triangular, ridge-like process, distal tip of phallus separated into two distinct, bent points. Vesica somewhat elongated, bag-like. **Female.** *Head*: As in male. *Thorax*: As in male. *Legs*: As in male. *Forewing dorsum*: Forewing length: 20.5 mm, $n = 1$. As in male but longer, only slightly broader, postmedial line barely kinked mesally. Faint antemedial line present, brown, bowed. *Forewing venter*: As in forewing dorsum but postmedial line mostly absent, antemedial absent, small black discal spot present. *Hindwing dorsum*: As in male. *Hindwing venter*: Following similar pattern as forewing venter except somewhat lighter, postmedial line faint. *Abdomen*: As in male but stouter. Sternite VIII as pair of elongated sclerotized bands widening toward anterior margin of VIII where they converge. *Genitalia*: (Fig. 93) $n = 1$. Tergite of VIII very broad, thin, weakly sclerotized, transparent, sac-like (see Fig. 93b). Robust apophyses anteriores shorter and wider than apophyses posteriores. Lamella antevaginalis relatively thin, weakly concave mesally. Ductus bursae elongate, tubular. Papillae anales somewhat rectangular, covered in setae.

Distribution (Map 1). This new species is apparently endemic to southeastern Brazil in the states of Rio de Janeiro, São Paulo, and Santa Catarina. The habitat at each of the collecting localities falls within the Brazilian Atlantic Forest biome (IBGE 2004).

Etymology. *Menevia magna* is named for the very broad valves of the male genitalia and also for the overall size, which is quite large on average for the *lantona* species-group.

Remarks. *Menevia magna* appears to be closely related to the much more widespread *M. lantona* of northern and central South America. Based on our present understanding of the distribution of this species in São Paulo, Santa Catarina, and Rio de Janeiro, it can be reasonably inferred that *M. magna* is probably found in eastern Paraná as well.

***lucara* species-group**

Species of the *lucara* species-group are similar to those of the *lantona* species-group and includes *M. lucara*, *M. menapia* sp. n., and *M. mielkei* sp. n. Sexual dimorphism is more pronounced than in the previous group, but still not as strong as in the next two species-groups, with females having broader, more ovoid forewings than males. The size of the species belonging to this group are on the smaller side for the genus, being only slightly larger than those of the *lantona* group overall. Forewing shape is stout and triangular as in the *lantona* group, but with more acutely falcate forewings and more steeply curved postmedial lunules. The ground color is dark, generally slate gray with varying degrees of brown and red in the medial area. The lobes of the gnathos in this species-group are unique among the genus, rather than being flattened, cupped, triangular or subtriangular in shape, they are thick, usually upturned, well-sclerotized structures reminiscent of boxing gloves. The phallus is rather tubular and simple, with only one species having a noticeable dorsal phallic ridge.

Key to *lucara* species-group

- 1 Forewing not acutely falcate, phallus without dorsal ridge. Central America to northern South America **2**
- Forewing very acutely falcate, narrow, phallus with dorsal ridge. Southeastern Brazil ***M. mielkei* sp. n.**
- 2 Juxtal processes pointed. Belize and Guatemala ***M. menapia* sp. n.**
- Juxtal processes terminally spatulate. Northern South America ***M. lucara***

***Menevia lucara* (Schaus, 1905)**

Figs 18–23, 76, 94; Map 2

Cicinnus lucara Schaus, 1905: 328

Menevia lucara; Schaus 1928: fig. ♂ 88f

Menevia lucara; Forbes 1942

Menevia lucara; Becker 1996

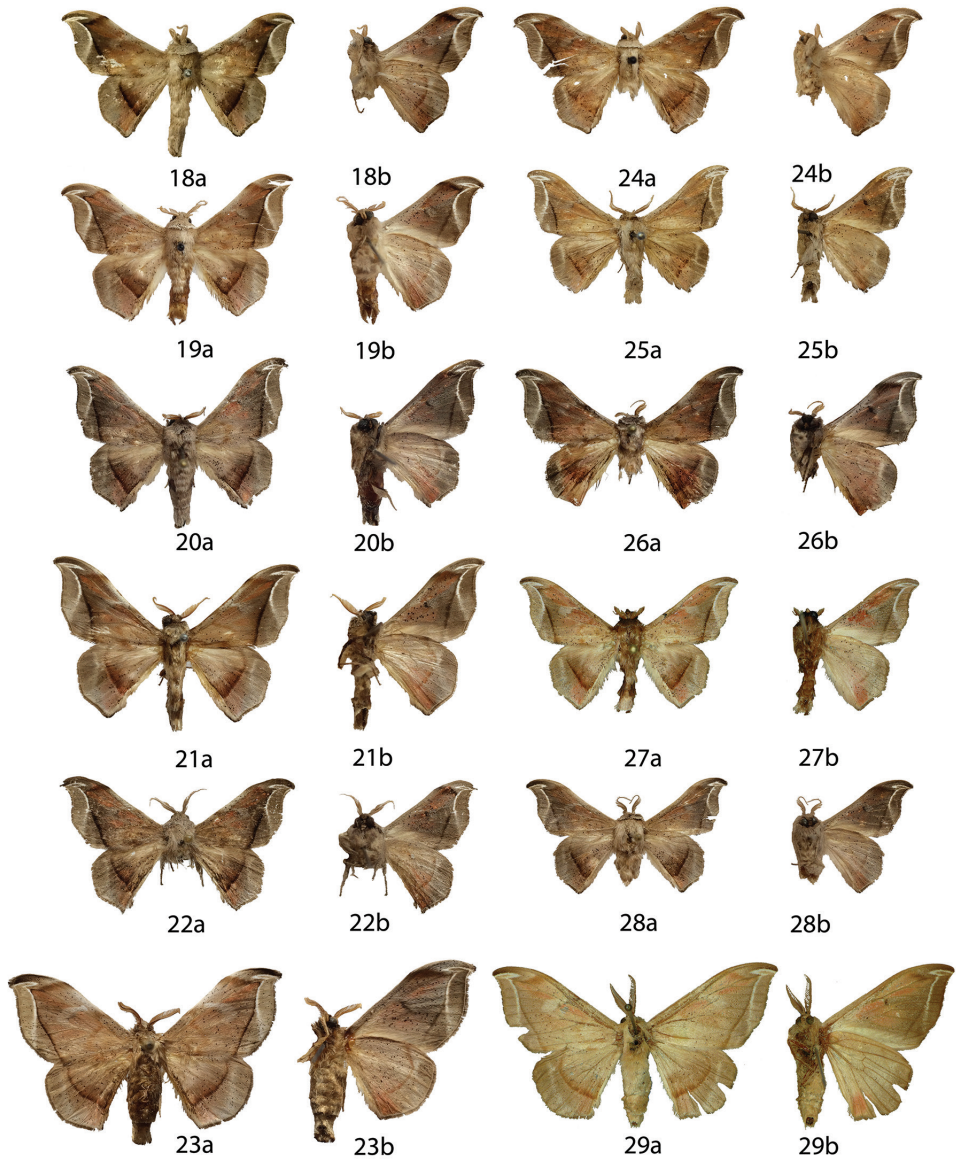
Menevia lucara; Herbin and Mielke 2014: fig. ♂ 51

Type material. Holotype, ♂: FRENCH GUIANA: St. Jean, Maroni, Fr. Guiana/ Collection Wm. Schaus/ *Perophora lucara* type Schaus/ Type No.: 8895 U.S.N.M./ USNM-Mimal: 1123/ St. Laurent diss.: 2-5-15:3/ (USNM) [examined]. No paratypes. Type locality: French Guiana: St. Jean du Maroni.

Additional specimens examined. (60 ♂, 4 ♀ total) **BRAZIL: Amazonas:** 4 ♂, Hyutanahan [Huitanaã], Rio Purus: II.1922, III.1922, S.M. Klages, Carn. Mus. Accs. 6963, 7088, 8840, St. Laurent diss.: 2-5-15:5 (CMNH). 2 ♂, Nova Olinda, Rio Pu-

rus: V.1922, VI.1922, S.M. Klages, Carn. Mus. Acc. 7088, St. Laurent diss.: 2-5-15:9 (CMNH). 1 ♂, Miracema, Rio Purus: IV.1922, S.M. Klages, Carn. Mus. Acc. 6960, St. Laurent diss.: 2-7-15:1 (CMNH). 1 ♂, “en remontant l’Amazones de Tefé à Tonantins,” [between Tefé and Tonantins]: XI.1921, Dognin Collection, USNM-Mimal: 2584 (USNM). 1 ♂, 2 km. N. of Itacoatiara-Manaus Highway, 11 km. W. of Itacoatiara, “Canadian Fathers’ Pool”: 11.V.1972, E.G., I. & E.A. Munroe (CNC). 3 ♂, Reserva Ducke, km. 26 Manaus-Itacoatiara Highway: 16.V.1972, 20.IV.1972, E.G., I. & E.A. Munroe, St. Laurent diss.: 7-7-15:2 (CNC). 5 ♂, Fonte Boa: V.1906, VI.1906, S.M. Klages, Rothschild Bequest, BM 1939–1 (NHMUK). **Pará:** 1 ♂, Ponte Nova, Rio Xingu: Dognin Collection, USNM-Mimal: 2575 (USNM). 3 ♂, 1 ♀, Likely Belém: A.M. Moss (NHMUK). **COLOMBIA:** 1 ♂, Antioquia, Nari [Nare?] River: Collection Frank Johnson, USNM-Mimal: 2580, St. Laurent diss.: 2-5-15:6 (USNM). 1 ♂, Muzo, 400–800 m: Coll. Fassl, ex. Joicey Coll. Brit. Mus, 1925—157, BMNH(E) 1378765, St. Laurent diss.: 6-29-15:1 (NHMUK). 1 ♂, Villavicencio, 400 m: Coll. Fassl, ex. Joicey Coll. Brit. Mus, 1925—157 (NHMUK). 1 ♀, “Interior of Colombia”: Wheeler, ex. Joicey Coll. Brit. Mus, 1925—157, BMNH(E) 1378760 [abdomen missing, no dissection prep.] (NHMUK). 2 ♂, “W. Columb.”, Rio Dagua, 600–1000 m: “2–5,” W. Hopp S., St. Laurent diss.: 7-7-15:1 (MNHU). **ECUADOR:** 2 ♂, Esmeraldas, 27 km. W. Alto Tambo, 200 m, timber tract at forest edge: 19.VIII.1996, Jan Hillman, St. Laurent diss.: 2-7-15:3, 9-7-14:2 (CMNH). **FRENCH GUIANA:** 13 ♂, St. Jean du Maroni: Collection Le Moul, ex. Dognin Collection, “topotype”, USNM-Mimal: 2578, St. Laurent diss.: 2-7-15:2 (USNM); VII-VIII.1904, E. Le Moul, Rothschild Bequest, BM 1939–1 (NHMUK); 1905–14, Rothschild Bequest, BM 1939-1 (NHMUK). 2 ♂, Mana River: V.1917, Acc. 6008, St. Laurent diss.: 2-5-15:4 (CMNH). 1 ♀, Kourou Forest: 25.VIII.1975, Mission M. Boulard and P. Pompanon, Muséum Paris, St. Laurent diss.: 5-22-15:1 (MNHU). 3 ♂, St. Laurent du Maroni: 1920–1932, ex. coll. L. & J. de Joannis (MNHU); XI, Collection Le Moul, ex. coll. Ed. Brabant, ex. Joicey Coll. Brit. Mus, 1925–157 (NHMUK); I.1906, E. Le Moul, Rothschild Bequest, BM 1939–1 (NHMUK). 1 ♂, Säul: 7.III.1978, Th. Porion (MNHU). **GUYANA:** 5 ♂, Potaro: V.1908, S.M. Klages, Rothschild Bequest, BM 1939–1 (NHMUK). 1 ♀, Bartica: W.J. Kaye, 1904-25, BMNH(E) 1378763, St. Laurent diss.: 7-4-15:1 (NHMUK). 1 ♂, Montsinery-Tonnegrande Nr. Tonnegrande [D5], N04°50.772', W52°31.165', 12 m: 11.III.2011, P. Sammut (RAS). **PANAMA:** 1 ♂, Darien vic. Cerro Pirre, Rancho Frio, N 08°01'11.3", W 077°43'57.0", 100 m: 19–24.VII.2013, J.R. MacDonald, St. Laurent diss.: 9-7-15:1 (MEM). **PERU:** 3 ♂, Carabaya, La Union, Huacamayo River, 2000 ft.: “wet s.,” XII.1904, G. Ockenden, Rothschild Bequest, BM 1939–1, BMNH(E) 1378764, St. Laurent diss.: 6-29-15:2 (NHMUK). **SURINAME:** 2 ♂, Moengo, Boven Cottica River: 25.V.1927, Cornell Univ. Lot 760, Sub 79, Lot 672, Sub 382, St. Laurent diss.: 12-10-13:1 (CUIC). 1 ♂, Aroewarwa Creek, Maroewym Valley: IV.1905, S.M. Klages, Rothschild Bequest, BM 1939–1 (NHMUK).

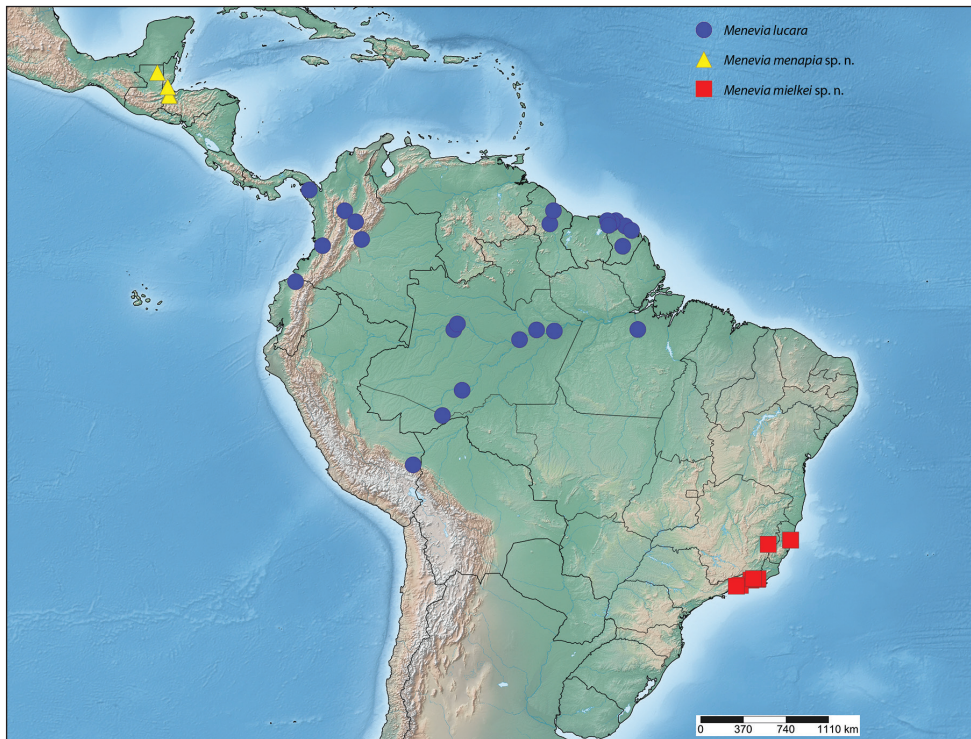
Diagnosis. Both males and females are recognizable by their moderate size and gray ground color with pinkish highlights especially concentrated near the apical angle of the contrasting black postmedial line and near the anal angle of hindwings. The postmedial



Figures 18–29. *Menevia lucara* species-group adults, **a** recto, **b** verso. **18** *M. lucara* holotype ♂, French Guiana, St. Jean du Maroni (USNM) **19** *M. lucara* ♂, Brazil, Nova Olinda, Rio Purus (CMNH) **20** *M. lucara* ♂, Panama, Darien vic. Cerro Pirre, Rancho Frio (MEM) **21** *M. lucara* ♂, Colombia, Rio Dagua, 600–1000 m (MNHU) **22** *M. lucara* ♂, Ecuador, Esmeraldas, 27 km W Alto Tambo, 200 m (CMNH) **23** *M. lucara* ♀, French Guiana, Kourou Forest (MNHN) **24** *M. menapia* holotype ♂, Guatemala, Cayuga (USNM) **25** *M. menapia* paratype ♂, Belize, Punta Gorda (NHMUK) **26** *M. menapia* paratype ♂, Guatemala, Izabal, Finca la Firmeza (CUIC) **27** *M. mielkei* holotype ♂, Brazil, Minas Gerais, Estação Biológica de Caratinga, Caratinga [photo courtesy CGCM] (DZUP) **28** *M. mielkei* paratype ♂, Brazil, Rio de Janeiro, Cachoeiras de Macacu, 700 m (USNM) **29** *M. mielkei* paratype ♀, Brazil, Rio de Janeiro, Sahy-Ramal de Mangaratiba [photo courtesy CGCM] (DZUP). Scale bar = 1 cm.

lunule is bright white and sharply curved outward toward the forewing margin, nearly forming a semicircle. The phallus/juxta combination is unique in the genus in that the phallus is smooth and cylindrical, without a dorsal phallic ridge. The terminal ends of the juxtal processes are spatulate, not pointed, and curved inward toward each other, near the terminal quarter of their length. The gnathos processes are highly distinct; they are thick, upturned, heavily sclerotized, somewhat boxing glove in shape.

Description. Male. *Head:* Light gray, straw colored in old specimens, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally due to ventral tufts, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. *Thorax:* As for genus. Grayish. *Legs:* As for genus. Tibial spurs thin apically, terminal third not scaled, especially ventrally, weakly hooked. *Forewing dorsum:* Forewing length: 16.5–21 mm, avg.: 17.3 mm, $n = 22$. Triangular, apical half of outer margin concave, apex falcate. Ground color gray with pink hue especially near apical point of postmedial line and throughout medial area, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above apical dash. Straight postmedial line black or brown, usually strongly contrasting. Antemedial area lighter, submarginal area gray without pink hue, somewhat contrasting with medial area, postmedial lunule originating from apical dash, lunule follows postmedial line from apex to one third length of postmedial line where lunule sharply sweeps outward toward wing margin, forming acute angle with postmedial line. Antemedial line usually absent, if present, faint, brown, undulating. *Forewing venter:* As in forewing dorsum but postmedial line fainter, antemedial line absent, small black discal spot present. *Hindwing dorsum:* Somewhat rounded with margin weakly pointed mesally, anal angle accentuated, similar coloration and patterning as forewings, vague postmedial lunule originating near anterior margin, sweeping outward to marginal point, antemedial line absent, postmedial line straight or slightly curved, especially near anterior wing margin. *Hindwing venter:* Following similar pattern as forewing venter, but slightly lighter, discal mark absent. *Abdomen:* As for genus. Coloration a continuation of grayish thoracic color. Midventral stripe absent. *Genitalia:* (Fig. 76) $n = 14$. Tegumen subtriangular, not constricted near base of gnathos. Vinculum broad, somewhat quadrate ventrally. Valves simple, relatively narrow, saccular and costal edges of valves with sharp tooth proximal to transtilla. Valves rounded apically. Uncus bottle-shaped, apex quadrate. Gnathos as two prominent, thick, heavily sclerotized, boxing glove shaped, upturned, outward facing extensions of varying width, occasionally broken dorsally. Juxtal processes roughly phallus length, heavily sclerotized, curving inward toward each other. Processes spatulate distally, setae absent. Base of phallus with robust, paired, somewhat elongated, rounded, diverging lobes, angled ventrally. Phallus simple, cylindrical, thin, but expanded distally. Left edge of rolled phallus simple, ridge-like process absent, distal tip of phallus separated into two distinct, often curled, points of varying length. Vesica elongated. **Female.** *Head:* As in male, labial palpi slightly longer. *Thorax:* As in male. *Legs:* As in male. *Forewing dorsum:* Forewing length: 15–20.5 mm, avg. 18.5 mm, $n = 3$. As in male but broader, less falcate, pinkish hue more prominent,



Map 2. *Menevia lucara* species-group.

postmedial line bowed out slightly mesally. *Forewing venter*: As in forewing dorsum but postmedial line fainter, antemedial line absent, small black discal spot present. *Hindwing dorsum*: As in male but slightly more rounded, broader. *Hindwing venter*: Following similar pattern as forewing venter except lighter. *Abdomen*: As in male but stouter. Sternite of VIII as pair of elongated sclerotized bands converging near anterior margin of VIII forming a “V”. *Genitalia*: (Fig. 94) $n = 2$. Tergite of VIII smoothly curved, very wide, with rounded edges, membranous gap mesally. Apophyses anteriores slightly shorter than apophyses posteriores. Lamella antevaginalis bent mesally, forming sharp “V” with ostium bursae at apex. Ductus bursae elongate. Papillae anales subtriangular when viewed ventrally, base widened, covered in relatively long setae especially at apex.

Distribution (Map 2). *Menevia lucara* is found throughout the Amazonian rainforest in the Guianas, Suriname, the Brazilian states of Pará and Amazonas, as well as in western and central Colombia, Panama, northwestern Ecuador, and southern Peru.

Remarks. Like *M. lantona*, *M. lucara* is a wide-ranging Amazonian species found throughout northern South America and is sympatric with *M. lantona* throughout most of its range.

Some geographical variation has been noted among the material examined. Specimens from Panama and Colombia (Figs 20 and 21 respectively) average slightly larger than those from the rest of the species range. Similarly, *M. lantona* showed parallel

geographic variation, with specimens from Colombia, and some from Panama also being larger on average. Additional geographic variation is present in the shape of the phallus. In some of the examined specimens from Colombia, and one from Ecuador, the phallus is slightly thicker and more robust overall than in specimens from other Amazonian localities. Also, the apical tips of the phallus are very short in specimens from French Guiana, Guyana, Suriname, and Peru, whereas they are longer in specimens from Ecuador and those from some Colombian localities.

***Menevia menapia* sp. n.**

<http://zoobank.org/290C69E5-B0E0-4C36-B4B6-57D71ECAB807>

Figs 24–26, 77; Map 2

Type material. Holotype, ♂: **GUATEMALA**: Cayuga Guat/ June/ Schaus and Barnes coll/ USNM-Mimal: 2565/ St. Laurent diss.: 9-7-14:1/ HOLOTYPE male *Menevia menapia* St Laurent and Dombroskie, 2016 [handwritten red label] (USNM). Type locality: Guatemala: Cayuga.

Paratypes, 8 ♂: **BELIZE**: 2 ♂, Punta Gorda, Brit. Honduras: VI.1933, J.J. White, Rothschild Bequest BM 1939-1, St. Laurent diss.: 6-29-15:3, BMNH(E) 1378758 (NHMUK); same data as previous Belize specimen, but no BMNH(E) number and not dissected, this specimen is missing the abdomen with no genitalia prep. (NHMUK). **GUATEMALA**: 3 ♂, Cayuga: Schaus and Barnes coll., Dognin Collection, “*Cicinnus lucara* Schaus 1920, Schaus,” USNM-Mimal: 2579 (USNM); VI, Schaus and Barnes coll., USNM-Mimal: 2564 (USNM); IV, “*Cicinnus lucara* I Schaus,” Carn. Mus. Acc. 6540, St. Laurent diss.: 2-5-15:7 (CMNH). 2 ♂, Tikal: 10.I.1980, R. Holland (AMNH); 11.I.1980, R. Holland, UV light, St. Laurent diss. 2-7-15:4 (AMNH). 1 ♂, Izabal, Finca la Firmeza 15.407, -88.696: 1.III.2014, J.J. Dombroskie, T. McCabe, J. Monzón, MV/UV light, St. Laurent diss.: 2-5-15:8 (CUIC). – All paratypes with the following yellow label: PARATYPE male *Menevia menapia* St Laurent and Dombroskie, 2016.

Diagnosis. Externally *M. menapia* is nearly identical to *M. lucara*, albeit this new species is slightly smaller on average. The most significant differences in external characters are present in the postmedial lunules of the fore- and hindwings. The postmedial lunule is less sharply swept to the forewing margin in *M. menapia*, and on the hindwing the lunule is more distinct, a brighter white rather than extremely faded as in *M. lucara*. The male genitalia of *M. menapia* are recognizable by the weaker sclerotization; smaller, slightly thinner processes of the gnathos, and the sharp, rather than spatulate, apices of the juxtal processes. Furthermore, the lobes at the base of the phallus are much shorter and stouter than in *M. lucara*. Geography is perhaps the easiest way to differentiate *M. menapia* from other species in the *lucara* species-group, as it is the only representative of the group from Central America north of Panama.

Description. Male. Head: Gray, fading to straw colored in old specimens, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by dark

brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally. Scape and pedicel tufted. *Thorax*: As for genus. Gray fading to straw in old specimens. *Legs*: As for genus. Tibial spurs thin apically, terminal third not scaled, weakly hooked. *Forewing dorsum*: Forewing length: 15–17 mm, avg.: 16 mm, $n = 7$. Triangular, apical half of outer margin concave, apex falcate. Ground color gray with pink hue medially, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray. Apex marked by black scales above apical dash. Weakly concave postmedial line black, contrasting. Antemedial area lighter, submarginal area gray without pink hue, contrasting with pinkish medial area, postmedial lunule originating from apical dash, lunule follows postmedial line from apex to one third length of postmedial line where lunule sweeps outward toward wing margin, roughly forming 45 degree angle with postmedial line. Antemedial line faint or absent, if present, brown, undulating. *Forewing venter*: As in forewing dorsum but postmedial line fainter, antemedial line absent, small black discal spot present. *Hindwing dorsum*: Rounded with margin weakly pointed mesally, anal angle accentuated, similar coloration and patterning as forewings, postmedial lunule present, originating near anterior margin, sweeping outward and fading to marginal point, antemedial line absent, postmedial line straight except near anterior margin. *Hindwing venter*: Following similar pattern as forewing venter. *Abdomen*: As for genus. Coloration a continuation of grayish thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 77) $n = 5$. Tegumen subtriangular, not constricted near base of gnathos. Vinculum broad, somewhat quadrate ventrally. Valves simple, relatively narrow, saccular and dorsal edges of valves with sharp tooth proximal to transtilla. Valves rounded or somewhat quadrate apically. Uncus somewhat triangular, apex quadrate. Gnathos as two sclerotized, somewhat boxing glove shaped, upturned, outward facing extensions. Juxtal processes roughly phallus length, not inwardly curved, parallel, curved toward phallus apex, flattened, lightly covered in short setae, pointed apically. Base of phallus with paired, stout, rounded, diverging lobes angled ventrally. Phallus simple, cylindrical, thin but somewhat engorged distally. Left edge of rolled phallus simple, without ridge like process, distal tip of phallus separated into two distinct points of varying length. Vesica bag-like. **Female**. Unknown.

Distribution (Map 2). *Menevia menapia* is so far known only from the Petén and Izabal Departments of eastern Guatemala and an adjacent area of Belize.

Etymology. *Menevia menapia* is named for the likely derivation of *Menevia*, Menapia. The etymology of Menapia, however, is less clear and may refer to an ancient Roman settlement supposed to have existed in Pembrokeshire, Wales or to a settlement inhabited by the Menapii people in Belgica.

Remarks. This species, although very similar in external appearance and genitalia characteristics to the South American representatives of the *lucara* species-group, especially the Amazonian *M. lucara*, is separated by well over 2000 km land distance. Because of the extreme allopatry of *M. menapia*, it is surprising that this new species differs so little from the wide-ranging, South American *M. lucara*. Despite these similarities, the allopatry combined with the overall slightly smaller size, minor external differences, and the distinct genitalia differences, warrant the separation of these two similar species.

***Menevia mielkei* sp. n.**

<http://zoobank.org/A8588B6A-EE9D-4D08-8ADD-ADAF7C90AA45>

Figs 2, 27–29, 78; Map 2

Type material. Holotype, ♂: **BRAZIL: Minas Gerais:** 29-I--3-II-2003, Estação Biológica de Caratinga, Caratinga, MG, 400 m, Mielke & Casagrande leg./ OM 61.563/ CGCM diss.: OM 61.563/ HOLOTYPE male *Menevia mielkei* St Laurent and Dombroskie, 2016 [handwritten red label] (donated to DZUP by OM). Type locality: Brazil: Minas Gerais: Caratinga.

Paratypes, 13 ♂, 1 ♀: **BRAZIL: Espírito Santo:** 1 ♂, Linhares, 40 m: 05–09.IV.1992, V.O. Becker col., USNM-Mimal: 2345, St. Laurent diss.: 2-5-15:12 (USNM). **Rio de Janeiro:** 4 ♂, Angra-Jussara: 25.XI.1935, coll. D’Almeida, No. 19.179 (DZUP); 20.II.1936, 24.II.1936, D’Almeida, Oiticica & A. Costa, ex. coll. D’Almeida, No. 19.176, 19.177, 19.178, DZ 32.702–32.705 (DZUP). 1 ♂, Petrópolis, 650 m: 10–20.X.1985, V.O. Becker col., col. Becker 64840, USNM-Mimal: 2344, St. Laurent diss.: 2-5-15:10 (USNM). 2 ♂, Petrópolis: 7.IX.1928, 12.XI.1928, Gagarin leg., ex. col. Gagarin, DZ 32.706–32.707 (DZUP). 3 ♂, Teresópolis, Barreira: 11.X.1955, 15.X.1955, 18.X.1955, ex. col. Gagarin, DZ 32.708–32.710 (DZUP). 1 ♂, Cachoeiras de Macacu, 700 m: 23.I.1998, V.O. Becker col., col. Becker 113168, USNM-Mimal: 2036, St. Laurent diss.: 2-5-15:11 (USNM). 1 ♂, Cachoeiras de Macacu, Boca do Mato: 18.I.1998, N. Tangerini col., ex. coleção Nirton Tangerini, DZ 32.711 (DZUP). 1 ♀, Sahy-Ramal de Mangaratiba [Sa]: 25.X.1932, coll. Ferr. d’Almeida, ex. coll. D’Almeida, No. 19.180, DZ 32.712 (DZUP). – All paratypes with the following yellow label: PARATYPE male/female *Menevia mielkei* St Laurent and Dombroskie, 2016.

Diagnosis. Externally *M. mielkei* is nearly identical to both *M. lucara* and *M. menapia*. The forewings of *M. mielkei* are slightly narrower than in the other two species, most notably by the somewhat more acute apices. Genitalia are very useful however, in the diagnosis of *M. mielkei*. In *M. mielkei*, the uncus is more slender, the processes of the gnathos are extremely atrophied and thin, the phallus has a distinct dorsal ridge, the processes of the juxta are nearly vertically extended above the base of the phallus, not curved over its length, and the valves lack the saccular edge tooth. Furthermore, the lobes at the base of the phallus are much shorter and stouter than in *M. lucara*, and in that way are more similar to *M. menapia*.

Description. Male. *Head:* Light gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by somewhat reduced dark brown collar of scales reaching labial palpi, labial palpi small, segments weakly defined ventrally. Scape and pedicel tufted. *Thorax:* As for genus. Grayish. *Legs:* As for genus. Tibial spurs thin apically, terminal third not scaled, especially ventrally, weakly hooked. *Forewing dorsum:* Forewing length: 16–17 mm, avg.: 16.6 mm, n = 4. Triangular, apical half of outer margin deeply concave, slightly convex near tornus, apex acutely falcate. Ground color gray with pinkish-red to salmon hue especially near apical point of postmedial line and near discal region, overall lightly speckled by dark petiolate scales.

Discal spot faintly marked by light gray. Apex marked by black scales above apical dash. Postmedial line straight, black-brown. Antemedial area lighter than medial area, submarginal area gray without pink hue, postmedial lunule originating from apical dash, lunule follows postmedial line from apex to roughly one third length of postmedial line where lunule sharply sweeps outwards toward wing margin, forming acute angle with postmedial line. Antemedial line very faint, brown, undulating. *Forewing venter*: As in forewing dorsum but postmedial line fainter, pinkish-red hue concentrated near costa and discal region, antemedial line absent, small black discal spot present. *Hindwing dorsum*: Rounded with margin weakly pointed mesally, anal angle accentuated, similar coloration and patterning as forewings, small black discal mark usually present, very vague postmedial lunules originating near anterior margin sweeping outward to marginal point, antemedial line absent, postmedial line straight or slightly concave. *Hindwing venter*: Following similar pattern as forewing venter, pinkish-red hue concentrated in anal region. *Abdomen*: As for genus. Coloration a continuation of grayish thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 78) $n = 4$. Tegumen subtriangular, quadrate at base. Vinculum broad, quadrate ventrally. Valves simple, relatively narrow, saccular edge smooth. Valves rounded apically. Uncus elongated, apex quadrate. Gnathos as two thin, atrophied, lightly sclerotized, outward facing extensions. Juxtal processes angled nearly perpendicular to dorsal surface of phallus. Juxtal processes sharply tipped, lightly covered in setae. Base of phallus with paired, backwards facing, rounded, diverging lobes. Phallus thick, expanded mesally. Left edge of rolled phallus forms distinct ridge of varying shape, always covered in setae, distal tip of phallus separated into two distinct points of varying length. Vesica elongate, sac-like. **Female.** *Head*: As in male. *Thorax*: As in male. *Legs*: As in male. *Forewing dorsum*: Forewing length: about 21 mm, $n = 1$. As in male but broader, less falcate, pinkish hue more prominent, postmedial line bowed out slightly mesally. *Forewing venter*: As in forewing dorsum but postmedial line fainter, antemedial line absent, small black discal spot present. *Hindwing dorsum*: As in male but slightly more rounded, broader. *Hindwing venter*: Following similar pattern as forewing venter except lighter. *Abdomen*: As in male but stouter. *Genitalia*: Not examined.

Distribution. *Menevia mielkei* is found in a small region of southeastern Brazil in the states of Rio de Janeiro, Espírito Santo, and adjacent eastern Minas Gerais. Whether or not this species is more widely distributed in the Brazilian Atlantic Forest is yet to be determined.

Etymology. *Menevia mielkei* is named after Carlos G. C. Mielke who provided extensive support and data throughout the process of writing this revision.

Remarks. Like *M. menapia*, *M. mielkei* is very similar to the wide-ranging *M. lucara*. Both *M. menapia* and *M. mielkei* are apparently restricted in distribution, both species being widely allopatric with *M. lucara*. Of the three species in the *lucara* species-group, *M. mielkei* has the most distinct genitalia characters as outlined in the diagnosis. The dorsal ridge of the phallus is a common character in many other species of *Menevia*, but within the *lucara* species-group, this ridge is only present in *M. mielkei*.

***ostia* species-group**

The following group contains three species, *M. ostia* comb. n., *M. parostia* comb. n., and *M. pallida*. The former two species were assigned to the genus *Pamea* Walker, 1855 and the latter species was recently described in *Menevia* (Herbin and Mielke 2014). The species belonging to the *ostia* species-group are appropriately placed in *Menevia* due to the presence of the white postmedial lunule and the complex genus-specific male genitalia. In this species-group, the sexual dimorphism is pronounced. The species belonging to the *ostia* species-group are similar in size to larger species of the *lantona* and *lucara* species-groups, although those of the *ostia* species-group average slightly larger. The females of this group, namely those of *M. ostia*, can be quite large for the genus. The forewing shape of females are characteristic of this species-group, males however, are much more similar to those of the *lantona* group. Specifically, the forewings are triangular in both species-groups and only weakly falcate, especially in the *ostia* species-group. The species belonging to the *ostia* group are light yellow to dark yellow or somewhat gold, without a brownish cast to the medial area as in the *lantona* species-group. The stark yellowish coloration of the medial area is highly contrasting with the gray submarginal area, which is typical of the genus. The male genitalia are characteristic of this species-group; the *ostia* group is the only group in which every examined specimen has a prominent phallic ridge. The paired gnathos is similar to that of the *lantona* group, but in the *ostia* group it is more spade-shaped rather than simply triangular.

Key to *ostia* species-group

- 1 Coloration washed out yellow-tan, hindwing with very faint zigzagged lunule; females: hindwing postmedial line closer to wing margin than to middle of wing **2**
- Coloration more gold yellow-orange, hindwing with usually prominent zigzagged lunule; females: hindwing postmedial line closer to thorax than to middle of wing. Distribution wide but usually in wet forests from Costa Rica to throughout Amazonia and into the Brazilian Atlantic Forest ***M. ostia* comb. n.**
- 2 Female forewing less than 21 mm, male unknown..... ***M. parostia* comb. n.**
- Female forewing greater than or equal to 21 mm. Brazilian Cerrado, possibly also south to Paraguay ***M. pallida***

***Menevia ostia* (Druce, 1898), comb. n.**

Figs 4, 5, 30–33, 36, 37, 79–81, 95; Map 3

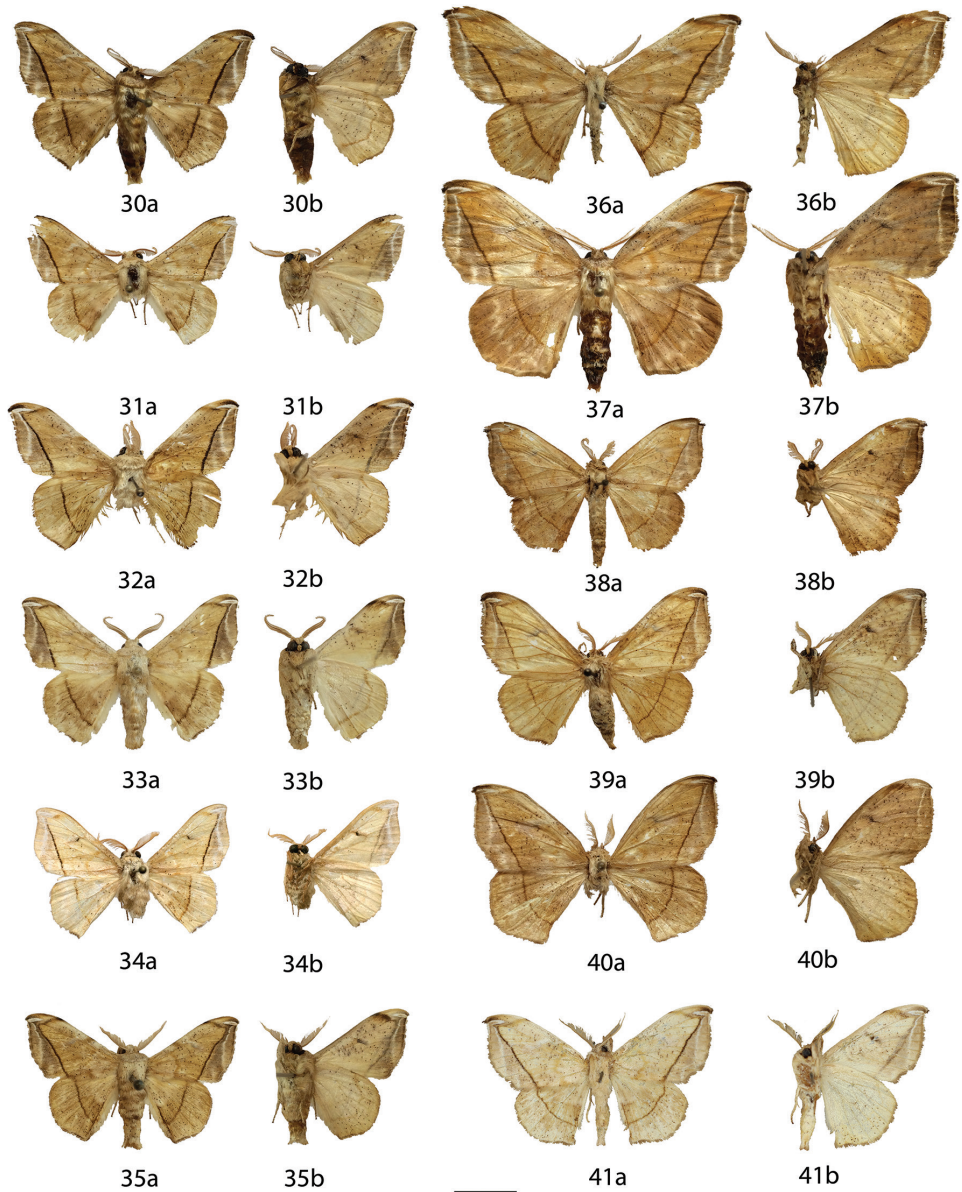
Perophora ostia Druce, 1898: 447; Tab. LXXXVIII, fig. 18 ♀

Pamea ostia; Schaus 1928: fig. ♀ 88g

Pamea ostia; Becker 1996

Type material. Holotype, ♀: **PANAMA**: Chiriqui [Chiriquí]/ 855/ Coll. Staudinger/ Type/ *Perophora ostia* ♀ type Druce/ Coll. Staudinger II.1178./ St. Laurent diss.: 7-14-15:1/ (MNHU) [examined]. Type locality: Panama: Chiriquí.

Additional specimens examined. (31 ♂, 15 ♀ total) **BRAZIL: Espírito Santo:** 9 ♂, Linhares, 40 m: 20–29.II.1992, 06.III.1993, 25–30.I.1998, V.O. Becker Col., Col. Becker 80930, 82945, 113493, USNM-Mimal: 2053, 2054, 2310–2316, St. Laurent diss.: 3-24-15:9, 3-24-15:10, 3-24-15:11, 7-14-14:5, 7-14-15:6 (USNM). **Minas Gerais:** 1 ♂, Estação Biológica de Caratinga, Caratinga, 400 m: 29.I.2003–3.II.2003, Mielke & Casagrande leg. (OM). **Pará:** 2 ♀, Likely Belém: A.M. Moss, Rothschild Bequest 1939–1, BMNH(E) 1377143 (NHMUK). **Rio de Janeiro:** 3 ♂, Petrópolis: 7.IV.1928, 6.IX.1959, 15.X.1963, Gagarin leg., ex. coll. Gagarin (DZUP). **COSTA RICA** (all Costa Rican specimens ex. D. H. Janzen & W. Hallwachs, Área de Conservación Guanacaste): **Alajuela:** 1 ♂, 1 ♀, Rio Blanco Abajo, 10.90037, -85.37254, 500 m: Voucher ♂: 05-SRNP-7880 [barcoded], collected: 18.XII.2005, emerged: 15.III.2006, food plant: *Terminalia oblonga* (USNM); Voucher ♀: 12-SRNP-4882 [barcoded], collected: 15.XI.2012, emerged: 12.V.2013, food plant: *Terminalia oblonga* (USNM). 1 ♂, Casa Leiva, 10.94314, -85.31808, 454 m: 17.IX.2009, light trap, Voucher: 09-SRNP-108159, St. Laurent diss.: 4-20-15:13 (USNM). 1 ♂, “Jabalina, Manta Pizote,” 10.97325, -85.31542, 288 m: 30.VIII.2008, light trap, Voucher: 08-SRNP-105449 (USNM). 1 ♂, Gallinazo, 11.01825, -85.37199, 360 m: Voucher: 11-SRNP-65684 [barcoded], collected: 21.VII.2011, emerged: 22.VIII.2011, food plant: *Terminalia amazonia* (USNM). 2 ♂, 1 ♀, Puente Palma, 10.9163, -85.37869, 460 m: Voucher ♂: 03-SRNP-34660 [barcoded], collected: 1.XII.2003, emerged: 16.I.2004, food plant: *Terminalia oblonga* (USNM); Voucher ♂: 03-SRNP-34659, collected: 1.XII.2003, emerged: 13.I.2004, food plant: *Terminalia oblonga* (USNM); Voucher ♀: 03-SRNP-34661, collected: 1.XII.2003, emerged: 20.I.2004, food plant: *Terminalia oblonga* (USNM). 1 ♂, 1 ♀, Tomatera: Voucher ♂: 12-SRNP-65485 [barcoded], collected: 16.VII.2012, emerged: 15.IX.2012, food plant: *Terminalia amazonia*, St. Laurent diss.: 4-20-15:12 (USNM); Voucher ♀: 12-SRNP-65488 [barcoded], collected: 16.VII.2012, emerged: 8.IX.2012, food plant: *Terminalia amazonia*, St. Laurent diss.: 4-20-15:11 (USNM). **Guanacaste:** 1 ♂, Sendero Puertas, 11.01087, -85.48817, 400 m: Voucher: 03-SRNP-18623, collected: 5.VIII.2003, emerged: 28.IX.2003, food plant: *Terminalia amazonia*, St. Laurent diss.: 7-14:15:3 (USNM). 1 ♂, 1 ♀, Pasmompa, 11.01926, -85.40997, 440 m: Voucher ♂: 07-SRNP-31642 [barcoded], collected: 8.III.2007, emerged: 1.VI.2007, food plant: *Terminalia amazonia* (USNM); Voucher ♀: 07-SRNP-31860 [barcoded], food plant: *Terminalia amazonia*, St. Laurent diss.: 4-20-15:8 (USNM). 1 ♂, Monte Cristo, 11.01373, -85.42531, 525 m: Voucher: 10-SRNP-21610 [barcoded], collected: 17.VII.2010, emerged: 14.IX.2010, food plant: *Terminalia amazonia* (USNM). 1 ♂, Tajo Angeles, 10.86472, -85.41531, 540 m: Voucher: 10-SRNP-4309 [barcoded], collected: 8.VIII.2010, emerged: 9.IX.2010, food plant: *Terminalia oblonga*, St. Laurent diss.: 7-14-15:2 (USNM). 1 ♂, Metereologico, 11.00199, -85.46166, 590 m: Voucher: 10-SRNP-21714 [barcoded], collected: 26.VII.2010, emerged: 29.VIII.2010, food plant: *Terminalia amazonia*, St. Laurent diss.: 4-20-15:14 (USNM). 1 ♂, 1 ♀, Casa Roberto,



Figures 30–41. *Menevia ostia* species-group adults, **a** recto, **b** verso. **30** *M. ostia* ♂, Costa Rica, Guanacaste, Tajo Angeles, 540 m, 10-SRNP-4309 (USNM) **31** *M. ostia* ♂, Costa Rica, Guanacaste, Sendero Puertas, 400 m, 03-SRNP-18623 (USNM) **32** *M. ostia* ♂, French Guiana, Sinnamary, Piste a St. Elie, km. 24, 50 m (AMNH) **33** *M. ostia* ♂, Brazil, Espírito Santo, Linhares, 40 m (USNM) **34** *M. pallida* holotype ♂, Brazil, Maranhão, Feira Nova do Maranhão, 480 m (DZUP) **35** *M. pallida* paratype ♂, Brazil, Maranhão, Feira Nova do Maranhão, 480 m (CGCM) **36** *M. ostia* holotype ♀, Panama, Chiriquí (MNHU) **37** *M. ostia* ♀, Costa Rica, Guanacaste, Sendero Rotulo, 510 m, 04-SRNP-34126 (USNM) **38** *M. parostia* holotype ♀, unknown locality (USNM) **39** *M. pallida* ♀ [questionable], Brazil, Minas Gerais, Lassance (CUIC) **40** *M. pallida* ♀, Brazil, Maranhão, Caixas, Reserva Ecol. Inhamum (CPAC) **41** *M. pallida* ♀ [questionable], Paraguay, Villarrica [photo courtesy NHMUK] (NHMUK). Scale bar = 1 cm.

11.01095, -85.42094, 520 m: Voucher ♂: 03-SRNP-20585 [barcoded], collected: 11.VIII.2003, food plant: *Terminalia amazonia* (USNM); Voucher ♀: 03-SRNP-20690, collected: 11.VIII.2003, emerged: 17.V.2004, food plant: *Terminalia amazonia* (USNM). 1 ♀, Mena Central, 11.02991, -85.45364, 345 m: Voucher: 01-SRNP-24319 [barcoded], collected: 29.XI.2001, St. Laurent diss.: 4-20-15:9 (USNM). 1 ♀, Sendero Rotulo, 11.01355, -85.42406, 510 m: Voucher: 04-SRNP-34126 [barcoded], collected: 27.VII.2004, emerged: 5.IX.2004, food plant: *Terminalia amazonia* (USNM). 1 ♀, Camino Mangos, 11.00766, -85.47926, 480 m: Voucher: 03-SRNP-19176, collected: 12.VIII.2003, emerged: 13.IX.2003, food plant: *Terminalia amazonia* (USNM). 1 ♀, Maderos, 11.00494, -85.47491, 510 m: Voucher: 11-SRNP-20049 [barcoded], collected: 4.I.2011, emerged: 15.V.2004, food plant: *Terminalia amazonia*, St. Laurent 4-20-15:10 (USNM). **FRENCH GUIANA:** 1 ♂, Sinnamary, Piste a St. Elie, km. 24, 50 m: 20.II.1991, at lights, coll. C. Snyder, St. Laurent diss.: 3-24-15:6 (AMNH). 1 ♂, Nouveau Chantier: Collection Le Moul, Dognin Collection, USNM-Mimal: 2572, St. Laurent diss.: 3-24-15:7 (USNM). 1 ♂, Piste Coralie PK 2: IV.1993, J. Navatte H. de Toulgoët (MNHN). 1 ♀, Godebert Maroni: VII, Collection Le Moul, Ex. Coll. Ed. Brabant 1920, ex. Joicey Coll., Brit. Mus. 1925–157, BMNH(E) 1377145 (NHMUK). 1 ♀, no additional locality data: Collection C. Bar, Ex. Oberthür Coll., Brit. Mus. 1927–3, BMNH(E) 1377140 (NHMUK). **PERU:** 1 ♀, Junín, Chanchamayo: I–VIII.1901, Hoffmann, Rothschild Bequest 1939–1, BMNH(E) 1377144 (NHMUK). **SURINAME:** 1 ♀, Paramaribo: 7.I.1955, Rf. [reared] *Terminalia catappa* L., J.B.M van Dinther, USNM-Mimal: 2612, St. Laurent diss.: 4-20-15:5 (USNM). **VENEZUELA:** 1 ♂, Aragua, Rancho Grande, 1100 m: 22-31.VII.1967, R.W. Poole, USNM-Mimal: 2739, St. Laurent diss.: 3-24-15:8 (USNM). **UNKNOWN:** 1 ♂ specimen without locality label, but bears label with red “1” written in colored pencil, there was a similar label on an examined female from Suriname, USNM-Mimal: 2613, St. Laurent diss.: 7-14-15:4 (USNM).

Diagnosis. *Menevia ostia* can be differentiated from all other species-groups by the gold-yellow to pale yellow ground color with the contrasting silvery gray submarginal area with a small white accessory mark near the tornus. These traits combined with overall very weakly falcate forewings, distinguish *M. ostia* from the similar *M. lantona* but not necessarily from the similar *M. pallida* and *M. parostia* comb. n. to be diagnosed below. The width of the submarginal area of the hindwings of female *M. ostia* is wider than in both *M. pallida* and *M. parostia* females. The postmedial line is situated at about midway along the length of the hindwing in *M. ostia*, but is slightly closer to the wing margin in the other two species. Male genitalia are distinct, except when in comparison with *M. pallida*, in that the apical end of the phallus is sharply downturned and the dorsal ridge of the phallus is highly variable, though always present. In females, the lateral portion of the prominently sclerotized VIII has appendicular apophyses dorsolaterally in addition to the apophyses anteriores, distinguishing the females from a questionable female of *M. pallida*, where the appendicular apophyses are absent.

Description. Male. *Head:* Straw or tan-gold colored, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments somewhat well defined ventrally, dorsally with darker scales contrasting with overall lighter coloration. Scape and pedicel weakly tufted. *Thorax:* As for genus. Gold to pale, fading to straw. *Legs:* As for genus. Tibial spurs thin apically lightly to almost completely scaled. *Forewing dorsum:* Forewing length: 12.5–18.5 mm, avg.: 16.8 mm, n = 23. Triangular, apical half of outer margin weakly concave, apex slightly falcate. Ground color gold-yellow to pale yellow, overall very lightly speckled by dark petiolate scales. Discal spot faintly marked by silvery white. Apex marked by black scales above apical dash. Black postmedial line slightly concave, sometimes weakly undulating. Submarginal area silvery gray, postmedial lunule originating from near where apical dash meets postmedial line, white mark follows postmedial line from apex to one quarter length of postmedial line where mark smoothly curves outwards toward wing margin becoming diffuse, forming acute angle with postmedial line. White accessory mark present near tornus. Antemedial line very faint or absent, if present, brown, undulating. *Forewing venter:* As in forewing dorsum but postmedial line fainter, convex near tornus, antemedial line absent, small black, somewhat rounded or elongated discal spot present. *Hindwing dorsum:* Rounded, anal angle weakly accentuated, similar coloration and patterning as forewings, postmedial lunule vague or well defined, zigzagged, originating near anterior margin, following curvature of wing margin, not steeply swept to margin, antemedial area lighter, postmedial line straight, sometimes undulating near anal angle. *Hindwing venter:* Following similar pattern as forewing venter but lighter, discal mark absent, marginal area usually browner than surrounding area. *Abdomen:* As for genus. Coloration a continuation of thoracic color. Midventral stripe absent. *Genitalia:* (Figs 79–81) n = 14. Tegumen subtriangular. Vinculum narrow, somewhat quadrate. Valves asymmetrical, moderate in width, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth much reduced in size to nearly absent, both valves with smaller mesal costal tooth immediately above saccular edge teeth. Valves somewhat indented mesally, rounded apically. Uncus teardrop-shaped, truncated apically, base variable in width, apex somewhat hooked. Gnathos as two, flattened, spade-shaped outward facing flaps, bent toward each other, tips nearly converging. Juxtal processes shorter than phallus, thin, flattened, slightly curved, smooth. Base of phallus with paired, rounded, diverging, backwards facing fingerlike lobes. Phallus variable, broad, dorsal ridge of varying shape. Left edge of phallus forms distinct ridge, either a rounded or rectangular hump mesally or extended along phallus length but always quadrate anteriorly forming distinct edge, distal tip of phallus downturned, separated into two distinct, bent points. Vesica elongated. **Female.** *Head:* As in male but scales more grayish, labial palpi longer, thinner, dorsally covered in fewer dark scales. *Thorax:* As in male. *Legs:* As in male but tibial spurs broader, more triangular. *Forewing dorsum:* Forewing length: 17.5–26 mm, avg.: 22.8 mm, n = 11. Coloration and patterning as in male but maculation stronger, silvery discal mark wider, wing shape broader, more elongate, ovoid. *Forewing venter:* As in forewing dorsum but usually darker, markings subdued, postmedial line much fainter, bent near tornus; antemedial line not present;

elongate black discal spot present. *Hindwing dorsum*: As in male but broader, post-medial lunule proportionally larger, zigzagged pattern more obvious. Postmedial line situated midway along length of wing. *Hindwing venter*: Following similar pattern as forewing venter but lighter. *Abdomen*: As in male but stouter. Sternite of VIII as pair of elongated sclerotized bands sometimes widening toward posterior margin of VIII converging near anterior margin. Sclerotized bands may be parallel or bowed out midway along length. *Genitalia*: (Fig. 95) $n = 6$. VIII prominently sclerotized laterally, with appendicular apophyses dorsolaterally in addition to apophyses anteriores. Tergite of VIII arch-like, converging mesally to form posteriorly directed point. Apophyses anteriores slightly shorter than apophyses posteriores. Lamella antevaginalis as wide, semicircular, sclerotized band. Ductus bursae short. Papillae anales rectangular or subtriangular when viewed ventrally, covered in short setae.

Distribution (Map 3). *Menevia ostia* is a widespread species, found in wet forests from Costa Rica and Panama, Peru, Suriname, French Guiana and the Brazilian Amazon in the state of Pará (the latter Brazilian state is not marked on Map 3 due to uncertainty of a specific locale, but which was probably Belém (I. Kitching pers. comm.)). There is also one record from northern Venezuela. This species is apparently found in the Brazilian Atlantic Forest, but see remarks below. *Menevia ostia* may range as far north as Belize as per a single report from Pook's Hill, Belize. This interesting record is treated in more detail in the remarks.

Natural history. *Menevia ostia* is one of the few species of *Menevia* with known host plant associations. Rearing records from D. Janzen & W. Hallwachs show that *M. ostia* feeds on both *Terminalia amazonia* and *T. oblonga* (Combretaceae). Additionally, a female specimen from Suriname at the USNM bears a label referencing *T. catappa*, a known host plant of *M. plagiata* (Raymundo 1919).

Furthermore, D. Janzen provided us access to photos of reared *M. ostia*, published here for the first time (see Figs 4, 5). Larval coloration is quite striking, but shows the usual morphology and one of a number of case structures typical of Mimallonidae.

Remarks. Druce described *Menevia ostia* at a time when most new Mimallonidae that were being described were being placed in the now invalid, preoccupied genus *Perophora* Harris, 1841. Later, Schaus (1928) moved *Perophora ostia* to the genus *Pamea* Walker, 1855, apparently based on wing venation, which along with other wing characteristics used by Schaus, are unreliable in assigning species to genera (St Laurent and Dombroskie 2015). Schaus made no mention of the striking similarity in appearance of *P. ostia* to species placed in the genus *Menevia*.

Schaus's assignment of *M. ostia* (and *M. parostia* comb. n.) to the genus *Pamea* is strange considering the resemblance of these two species to *M. lantona*, a species that he described and designated as type species of *Menevia*, a genus that he also described. In comparing the genitalia of *M. ostia* with the type species of *Pamea*, *P. albistriga* Walker, 1855, the first author found the male genitalia of *M. ostia* completely unlike those of *P. albistriga*. The male genitalia of *P. albistriga* are very simple in comparison with those of *M. ostia* and *Menevia* as a whole. The phallus of *P. albistriga* is simple and truncated apically, and lacking a fused juxta with extended superior processes as in



Map 3. *Menevia ostia* species-group.

M. ostia and all other *Menevia*. In *P. albistriga* the vinculum is much more elongated, valves extremely simple, and the gnathos are atrophied. Genitalia alone offer enough support for the reassignment of *P. ostia* to *Menevia* based on the presence of all generic autapomorphies. The external characters of *M. ostia* are also sufficient for assigning this species to *Menevia*, namely the postmedial lunule and apical dash, both of which are readily apparent in *M. ostia*.

Of the four *Menevia* species-groups, the *ostia* species-group is the most difficult group to tease apart into different morphologically separable species. Phallic structure is an important character for species identification in *Menevia*, but each specimen of *M. ostia* examined ($n = 14$) had a uniquely shaped dorsal phallic ridge, varying in structure from a singular rounded or rectangular hump, to a crest that follows the length of the phallus (compare Figs 79c, 80, and 81). There is some degree of geographic association with the phallus structure. For example, specimens from Costa Rica generally have a more hump-like phallic ridge whereas those from northern South America have more elongated crests along the length of the phallus. There is also a great deal of variation within a single given locality. Specimens from the state of Espírito Santo, Brazil, for example, seem to have dorsal phallic ridges almost intermediate between those found in Costa Rica and northern South America. Furthermore, specimens from the Atlantic Forest biome are paler than those from the rest of the distribution. However,

a small number of specimens from Rio de Janeiro contradicted this apparent trend as they were darker and much more similar to Costa Rican populations.

This issue of blurred species boundaries is non-existent in the *lantona* and *lucara* species-groups. Both of these species-groups each have one species endemic to the Brazilian Atlantic Forests, and these endemic species each have unique genitalia and display minor but consistent external differences. The high degree of genitalia and external variation in *M. ostia*, even from a single location, impedes our ability to locate species-specific traits. Molecular evidence may be able to offer more conclusive insights, but even then, material is greatly lacking for the *ostia* species-group. Until more data is made available, it is more parsimonious to consider *M. ostia* to be a wide-ranging, phenotypically homogeneous species with variable male genitalia.

Menevia ostia, the very similar *M. parostia* comb. n., and *M. pallida*, are apparently distinct on the basis of female morphology and environment, with the latter species being restricted to drier regions of central South America. Only one record of *M. ostia* exists from a dry region, a specimen from northern Venezuela. The genitalia of this specimen are surprisingly very similar to those of *M. pallida*. A similar issue exists for the specimens from Espírito Santo. Although these specimens are consistently larger and paler than Costa Rican specimens, some specimens from Costa Rica are in fact, larger and paler than most others from similar localities.

Further complicating our understanding of the distributional boundaries of true *M. ostia* is a male specimen resembling *M. ostia* reported from Pook's Hill, Belize (M. J. C. Barnes pers. comm.; color photo examined). This record is much farther north in Central America than any other records of the *ostia* group. Only one other species of *Menevia* is known from Belize: *M. menapia*. Unfortunately, this specimen is inaccessible to us and it cannot be dissected to determine if it is conspecific with *M. ostia*, or if it perhaps represents an undescribed species. Regardless of the specific identity of this specimen, the *ostia* species-group ranges at least as far north as Belize, perhaps making the extreme allopatry of *M. menapia* less of a biogeographic oddity relative to the genus *Menevia* as a whole.

***Menevia parostia* (Schaus, 1928), comb. n.**

Figs 38, 96

Pamea parostia Schaus, 1928: 667

Pamea perostia; Becker 1996, misspelling

Type material. Holotype, ♀: **UNKNOWN**: Type No. 33592 U.S.N.M./ *Pamea parostia* type Schaus/ USNM-Mimal: 1107/ St. Laurent diss.: 4-20-15:7/ (USNM) [examined]. Type locality: Unknown.

Diagnosis. *Menevia parostia* can be differentiated from *M. ostia* by the placement of the postmedial line of the hindwing, which is roughly midway along the length of the hindwing in *M. ostia* and closer to the wing margin in *M. parostia*. Furthermore,

the sclerotized bands on the venter of the VIII abdominal segment are very thin in *M. parostia*. Lack of material and variability of this structure, however, belies its diagnostic capability. Additionally, most (93%, $n = 15$) female specimens of *M. ostia* are much larger than those of *M. parostia*. This holotype of *M. parostia* does not differ remarkably from the single definitive female of *M. pallida* (see remarks).

Description. Male. Unknown. **Female.** *Head:* Straw colored, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi moderately long, reaching beyond frons, segments somewhat well defined ventrally, dorsally with darker scales contrasting with overall lighter coloration. Scape and pedicel weakly tufted. *Thorax:* As for genus. Straw colored. *Legs:* As for genus. Tibial spurs relatively thick, long, almost completely scaled except ventrally. *Forewing dorsum:* Forewing length: 18 mm, $n = 1$. Subtriangular, rounded, apical quarter of outer margins weakly concave, apex slightly falcate. Ground color pale tan-yellow, moderately speckled by dark petiolate scales. Discal spot faintly marked by gray. Apex marked by black scales near tip of apical dash. Postmedial line brown, mostly straight. Submarginal area pale gray, postmedial lunule originating from near where apical dash meets postmedial line, lunule follows postmedial line from apex to one quarter length of postmedial line where lunule smoothly curves outward toward wing margin becoming somewhat diffuse, forming acute angle with postmedial line. Faint white accessory mark present near tornus. Antemedial line very faint, brown, undulating. *Forewing venter:* As in forewing dorsum but more heavily speckled, postmedial line bent outwards near tornus, antemedial line absent, discal spot present, small, black. *Hindwing dorsum:* Rounded, similar coloration and patterning as forewings, postmedial lunule very vague, wavy, not zigzagged, originating near anterior wing margin, following curvature of wing margin, not steeply swept to margin, antemedial area lighter, postmedial line weakly curved, closer to wing margin than midway along wing length. *Hindwing venter:* Following similar pattern as forewing venter but discal mark absent, marginal area color as surrounding area. *Abdomen:* As for genus but stouter. Coloration a continuation of thoracic color. Midventral stripe absent. Sternite of VIII as pair of thin sclerotized bands not touching near anterior margin, bowed out slightly mesally. *Genitalia:* (Fig. 96) $n = 1$. VIII prominently sclerotized laterally, appendicular apophyses present. Tergite of VIII arch-like, converging mesally to form posteriorly directed point. Apophyses anteriores slightly shorter than apophyses posteriores. Lamella antevaginalis as semicircular, sclerotized band. Ductus bursae short. Papillae anales rectangular when viewed ventrally, covered in short setae.

Distribution. Unfortunately the holotype is without locality information, furthermore, Schaus's original (1928) description listed the "habitat" as "unknown."

Remarks. Schaus (1928) described *Pamea parostia*, known only from the female holotype, based on its resemblance to *M. ostia*, differing only by its smaller size, "reduced" markings, and the "more developed" frenulum. Upon examining the holotype and comparing it with the much larger female holotype of *M. ostia* and a series of mostly larger females from Costa Rica, we found that Schaus was incorrect in his assertion that the frenulum of *M. parostia* is more developed, when in fact the frenulum appears to be of the same size and arrangement in examined *M. ostia* females. Schaus

frequently failed to locate the frenulum despite its presence, as shown by previous authors (Pearson 1951, 1984, St Laurent and Dombroskie 2015). Furthermore, the size difference between *M. parostia* and *M. ostia* is certainly not enough evidence to maintain *M. parostia* as a valid species. Among the *M. ostia* specimens from Costa Rica that were examined in this work, one female specimen from Tomatera was actually smaller than the holotype of *M. parostia*, along with a similarly very small male. Public barcode data shows this smaller pair of *M. ostia* display no differences whatsoever from regularly sized *M. ostia* from nearby locations (BOLD). Many species of Mimallonidae frequently display dwarfed specimens (R. A. St. Laurent pers. obs.), perhaps due to poor host plant assimilation or other environmental factors, potentially explaining the small size of the single pair of *M. ostia*.

The presence of such small specimens of *M. ostia* originally lead us to believe that *M. parostia* must be just another small example of this species, well within the natural size range and we were prepared to synonymize *M. parostia* with *M. ostia*. However, additional examination of the holotype of *M. parostia* revealed characters of the hindwing maculation and genitalia that were not seen in any examined *M. ostia*. The paired sternites of VIII in *M. parostia* are slightly thinner overall in comparison to those of *M. ostia*, but this character is rather variable in general. A more dramatic difference is found in the arrangement of the hindwing postmedial line between the two species (see Figs 36, 37 compared with 38). The holotype of *M. parostia* is strikingly similar to a female specimen of *M. pallida* in relative size, coloration, and arrangement of the hindwing postmedial line. Unfortunately, the abdomen of the female *M. pallida* is missing and thus a genitalia comparison cannot be performed. It is quite possible that *M. pallida* is a junior synonym of *M. parostia*, but without locality data, the abdomen of the female *M. pallida*, or true males of *M. parostia*, we cannot render this conclusion definitive. Pending further data, we therefore retain both *M. parostia* and *M. pallida* as valid species.

***Menevia pallida* Herbin & Mielke, 2014**

Figs 34, 35, 39, 40, 41, 82, 97; Map 3

Menevia pallida Herbin & Mielke, 2014: 147–149; figs ♂ 52–54, ♂ genitalia 55, 56

Type material. Holotype, ♂: BRAZIL: Maranhão: genitalia prep. D. Herbin, ref H. 986 [blue label]/ BRESIL Maranhao [Maranhão], Feira Nova do Maranhão, Retiro, 480 m. 21-25.II.2012, -07°00'31"S, -46°26'41"W, C. Mielke leg. Coll. D. Herbin / DZ 15.734/ Holotype ♂, *Menevia pallida* Herbin & Mielke det., Antenor 2014 [red label]/ (DZUP) [examined]. Type locality: Brazil: Maranhão: Feira Nova do Maranhão.

Paratype, 1 ♂, BRAZIL: Maranhão: Feira Nova do Maranhão, Retiro, 46°26'41"W, -07°00'31"S, 480 m: 16–17.II.2013, C. Mielke leg., Paratypus *Menevia pallida* Herbin & C. Mielke det., 2014 [green label], Col. C. Mielke 26.349, St. Laurent diss.: 6-16-15:3 (CGCM).

Additional specimen examined. BRAZIL: Maranhão: 1 ♀, Caixas, Reserva Ecol. Inhamum, 13°12'S, 45°27'W [given coordinates inaccurate], 110 m: 27.II.2006–1. III.2006, lençol e luz mista [sheet and mixed light], F. Limeira-de-Oliveira & J.C. Silva cols, Coleção Embrapa-CPAC No. 20053, [missing abdomen, no genitalia prep.] (CPAC).

Additional questionable specimens examined. (3 ♀ total) **BRAZIL: Minas Gerais:** 1 ♀, Lassance: 12.XI.1919, Cornel Univ. Expedition Lot 569, Sub 116, Cornell U. Lot 672, Sub 385 Det. W. Schaus [*ostia* ♀], St. Laurent diss.: 4-20-15:6 (CUIC). **PARAGUAY:** 1 ♀, Amambay, Parque Nacional Cerro Corá, 22°39'S, 56°01'W: 7–10. IV.1986, M. Pogue & M. Solis, St. Laurent diss.: 5-8-15:1 (USNM). 1 ♀, Guairá, Villarrica: 21.IX.1926, F. Schade, ex. Joicey coll. 1925–157, BMNH(E) 1377142 (NHMUK).

Diagnosis. *Menevia pallida* can be differentiated from the somewhat larger, but very similar *M. ostia* by the pale tan to yellowish brown ground color as opposed to gold or pale yellow. Additionally, the dark speckling is usually heavier due to the presence of more petiolate scales. The hindwings are without bright, obvious, zigzagged postmedial lunules as in *M. ostia*. In both *M. pallida* and *M. parostia*, the placement of the postmedial line on the hindwing is closer to the wing margin than to midway along the length of the wing as it is in *M. ostia*. The phallic ridge is more smoothly curved and less quadrate terminally than in *M. ostia*, with the front edge of the phallic ridge evenly sloped rather than squared. The female of *M. pallida* is smaller than those of *M. ostia*. We are unable to provide characters to differentiate the unique female of *M. parostia* from female *M. pallida*, although the females of *M. pallida* at our disposal (both true and questionable specimens) are always slightly larger than the unique specimen of *M. parostia*.

Description. Male. Head: Pale tan, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi small, segments somewhat well defined ventrally, dorsally with darker scales contrasting with overall lighter coloration. Scape and pedicel weakly tufted. **Thorax:** As for genus. Pale gold-tan. **Legs:** As for genus. Tibial spurs relatively thick. **Forewing dorsum:** Forewing length: 15.5–17 mm, avg.: 16.3 mm, n = 2. Triangular, apical half of outer margins weakly concave, apex slightly falcate. Ground color pale tan to yellowish, moderately speckled by dark petiolate scales. Discal spot faintly marked by gray. Apex marked by black scales above apical dash. Black postmedial line mostly straight, sometimes weakly undulating or kinked. Submarginal area pale gray, postmedial lunule originating from near where apical dash meets postmedial line, lunule follows postmedial line from apex to one quarter length of postmedial line where lunule smoothly curves outward toward wing margin becoming somewhat diffuse, forming acute angle with postmedial line. White accessory mark present near tornus. Antemedial line faint, brown, undulating. **Forewing venter:** As in forewing dorsum but two postmedial lines present, both much fainter than single line on dorsum, one line convex near tornus and slightly undulating, the other straight, following the postmedial line of wing dorsum, antemedial line absent, small black elongated discal spot present. **Hindwing dorsum:** Rounded, anal angle weakly accentu-

ated, similar coloration and patterning as forewings, postmedial lunule very vague, wavy, not zigzagged, originating near anterior wing margin, following curvature of wing margin, not steeply swept to margin, antemedial area lighter, postmedial line straight, weakly undulating near anal angle. *Hindwing venter*: Following similar pattern as forewing venter but discal mark absent, marginal area color as surrounding area. *Abdomen*: As for genus. Coloration a continuation of thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 82) $n = 2$. Tegumen subtriangular to nearly ovoid. Vinculum narrow, somewhat accentuated quadrate corners. Valves slightly asymmetrical, short, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth much reduced in size, both valves with smaller central ridge immediately above saccular edge teeth. Valves somewhat indented mesally, rounded apically. Uncus triangular, apex sharp, moderately hooked. Gnathos as two, flattened, spade-shaped outward facing flaps, bent inward, tips nearly meeting. Juxtal processes shorter than phallus, narrow, flattened, slightly curved, smooth. Base of phallus with paired, rounded, diverging, backwards facing fingerlike lobes. Phallus curved, broad, lengthwise dorsal ridge present. Left edge of phallus forms distinct setae covered ridge, extended along phallus length, smoothly sloped at anterior terminus, distal tip of phallus weakly downturned separated into two distinct, bent points. Vesica bag-like. **Female.** *Head*: As in male but scales paler, labial palpi longer, thinner, dorsally covered in less dark scales. *Thorax*: As in male. *Legs*: As in male. *Forewing dorsum*: Forewing length: 21 mm, $n = 1$. Coloration and patterning exactly as in male, wing shape broader. *Forewing venter*: As in forewing dorsum but lighter, markings subdued, postmedial line much fainter, weakly bent near tornus; antemedial line not present; faint, elongated black discal spot present, darkest mesally. *Hindwing dorsum*: As in male but slightly broader, larger, submarginal area grayish. *Hindwing venter*: Following similar pattern as forewing venter except lighter with very little maculation. [Abdomen and genitalia based on a questionable specimen from Minas Gerais that may not be attributable to *M. pallida*, the single Maranhão female is missing an abdomen and thus genitalia are unavailable]. *Abdomen*: As in male, but stouter. Sternite of VIII as pair of elongated sclerotized bands, bowed out midway along length. *Genitalia*: (Fig. 97) $n = 1$. VIII prominently sclerotized laterally, appendicular apophyses absent. Tergite of VIII arch-like, converging mesally to form posteriorly directed point. Apophyses anteriores shorter than apophyses posteriores. Lamella antevaginalis as semicircular, sclerotized band. Ductus bursae short. Papillae anales subtriangular when viewed ventrally, covered in setae.

Distribution (Map 3). *Menevia pallida* is potentially restricted to drier Cerrado habitat. Reliable records exist only from the Brazilian state of Maranhão. We questionably consider a female specimen from eastern Minas Gerais, Brazil to be this species due to the presence of Cerrado at that locality (IBGE 2004). Additional records from Paraguay are herein provisionally considered *M. pallida*, but likely represent an additional undescribed species, see remarks below.

Remarks. The recently described *M. pallida* is not very distinct from the widespread *M. ostia* or from the unique specimen of *M. parostia*. The differences between

M. pallida and *M. ostia*, while present, provide only a weak basis on which to consider these taxa as separate species. The original description of *M. pallida* was based on comparisons with *M. lantona*, which was considered the most similar species by Herbin and Mielke (2014). *Menevia ostia* was not mentioned by these authors, despite the almost exact same external and genitalia morphologies of the males of the two species. In addition, the authors reported that the genitalia of *M. pallida* are “the same” as *M. lantona*, which we have found not to be the case. The genitalia of the holotype of *M. pallida* in Herbin and Mielke (2014) displays very little resemblance to any *M. lantona* dissections that we have reviewed (compare Figs 72 and 82). Most notably, the uncus of *M. lantona* is handbell shaped rather than triangular, the valves narrower, the gnathos processes more flattened and triangular, and finally the phallus of *M. lantona* is of an entirely different shape. The phallus of *M. lantona* lacks a dorsal ridge, which is a prominent character of the entire *ostia* species-group. Apparently the phallus of *M. pallida* was not fully examined during the description, as it was not figured separately from the rest of the genitalia nor was it removed from the genitalia when one of us examined the genitalia preparation.

Despite the issues with the original description of *M. pallida*, we still consider it a valid species based primarily on environmental differences, consistently smaller size, paler tan rather than golden coloration, the position of the hindwing postmedial line in females, and the potential difference in female genitalia compared to female *M. ostia*. The female from Caixas, Maranhão, Brazil, was collected about 440 km north-east of the type locality of *M. pallida*, and from the same Cerrado habitat (Silva et al. 2012), and thus is the single female specimen most likely associated with *M. pallida*. We consider this association reliable because the size and maculation agree perfectly with the examined holotype and paratype males of *M. pallida*.

The single female from Lassance, Minas Gerais, Brazil, which we associate here with *M. pallida*, albeit questionably, is relatively small and pale compared to *M. ostia* females from Central America and northern South America, and consequently seems more in line with *M. pallida*. However, as explained in the remarks to *M. ostia*, some populations from Atlantic Forest localities are similarly pale. Pending upon the availability of additional specimens of both sexes from Minas Gerais and Espírito Santo, we are unable to conclusively allocate these populations to either species.

We consider two females from Paraguay to be *M. pallida* due to their localities near the Cerrado, small size, and pale coloration. However, the forewing shape is less rounded than in female *M. ostia* and *M. pallida*, making the Paraguayan specimens appear rather distinct (see Fig. 41). Because of this morphological difference, these specimens were not included when writing the female description for *M. pallida*. Due to the fact that we lack males from Paraguay, we are unable to determine conclusively whether the specimens from Paraguay represent *M. pallida* or a distinct, undescribed species. Regardless, the specimens from Paraguay certainly belong to the *ostia* species-group and are treated here in the present study as they represent the only records of *Menevia* from Paraguay and are therefore significant.

***plagiata* species-group**

The *plagiata* species-group contains the largest *Menevia*, the females of some species are among the largest Mimallonidae. Sexual dimorphism is very pronounced in this group. Forewings of females are longer and broader than those of the males, which are conversely very falcate. Unlike in the three previous species-groups, the species belonging to the *plagiata* group do not have the curved postmedial lunules, but instead have distinct white bands that border the outer margin of the postmedial line, either along the complete length of the line as in the *vulgaris* subgroup containing: *M. vulgaris* sp. n., *M. franclemonti* sp. n., *M. vulgaricula* sp. n., *M. cordillera* sp. n., and *M. delphinus* sp. n.; or are interrupted midway along the line as in the *plagiata* subgroup, which contains: *M. plagiata*, *M. australis* sp. n., and *M. alurca*. The lack of the lunules immediately distinguishes this species-group from the others. However, the presence of the apical dash is very distinctive and thus the species in this group are easily identifiable as *Menevia*. Ground color is also darker for this group, most similar to the *lucara* group, being primarily gray, brown, or some combination. The male genitalia are more robust than in other species-groups. The paired gnathos is flat and oblong, either somewhat triangular or partly ovoid, usually with extreme truncation distally. The phallus is diverse in form, ranging from elongated and smooth to almost triangular due to an exaggerated dorsal projection when viewed laterally. The juxtal arms are generally very flat and wide, not sharp apically.

Key to *plagiata* species-group

- 1 Forewing with continuous white band along exterior edge of postmedial line, midventral abdominal stripe absent **4 [*vulgaris* subgroup]**
- Forewing with interrupted white band along exterior edge of postmedial line, midventral abdominal stripe present **2 [*plagiata* subgroup]**
- 2 Postmedial line usually straight, brown with little red coloration **3**
- Postmedial line usually bent outwards mesally, coloration dark gray with predominance of red suffusion. Central South America, especially in the Brazilian Cerrado ***M. alurca***
- 3 Wings relatively narrow, usually grayish, white band of forewing postmedial line usually reaching apical tip of wing, especially prominent in females, phallus with elongated, thin, dorsal projection. Vicinity of the state of Rio de Janeiro.. ***M. plagiata***
- Wings broader, usually brown, white band of forewing postmedial line usually intercepting apical dash mesally, or dissipating before reaching apex, especially prominent in females, phallus with triangular or rounded dorsal hump. Southeastern Brazil, especially Santa Catarina north to São Paulo ***M. australis* sp. n.**
- 4 Large, forewing length males: 22–28 mm, females: 27.5–39 mm, phallus smoothly curving, dorsally smooth or with irregular edge, but never with distinct bulge **5**

- Small, forewing length males: 17–23 mm, females: 27 mm, phallus with dorsal bulge anteriorly or central protuberance..... **6**
- 5 Phallus dorsum smooth, evenly edged, terminal sclerotized edge that becomes vesica, diagonal; ventral bump of phallus angled away from phallus terminus. Southeastern Brazil ***M. franclemonti* sp. n.**
- Phallus dorsum with irregular edge, terminal sclerotized edge that becomes vesica nearly vertical, ventral bump of phallus absent or indistinct. Northern South America ***M. vulgaris* sp. n.**
- 6 Lobes at base of phallus rounded, not peg-like. Andean or Brazilian Cerrado ... **7**
- Lobes at base of phallus peg-like. Amazonian..... ***M. vulgaricula* sp. n.**
- 7 Forewings elongate, very acute apically, forewing postmedial line mostly straight only slightly curved. Andean Cordillera Oriental, Peru to Bolivia
..... ***M. cordillera* sp. n.**
- Forewings stouter, less acute apically, forewing postmedial line slightly undulated. Endemic to the Brazilian Cerrado ***M. delphinus* sp. n.**

***plagiata* subgroup**

The *plagiata* subgroup contains *M. plagiata*, *M. australis* sp. n., and *M. alurca* and is diagnosed by the discontinuous white band along the outer margin of the postmedial line and by the presence of a midventral abdominal stripe. The species boundaries in the *plagiata* subgroup are unclear and thus we consider the three species in this subgroup as part of a species complex hereby-considered the “*Menevia plagiata* species complex” and may contain additional species not formally recognized herein.

***Menevia plagiata* (Walker, 1855)**

Figs 42–44, 52, 53, 83, 84, 98; Map 4

Mimallo plagiata Walker, 1855: 1341

Perophora plagiata; Raymundo 1919

Menevia plagiata; Schaus 1928: fig. ♀ 88g

Perophora plagiata; Monte 1934

Menevia plagiata; Lima 1950

Menevia plagiata; Silva et al. 1968

Menevia plagiata; Becker 1996

Menevia plagiata; Mecke et al. 2000

Menevia plagiata; Pastrana 2004

Menevia plagiata; Herbin and Mielke 2014

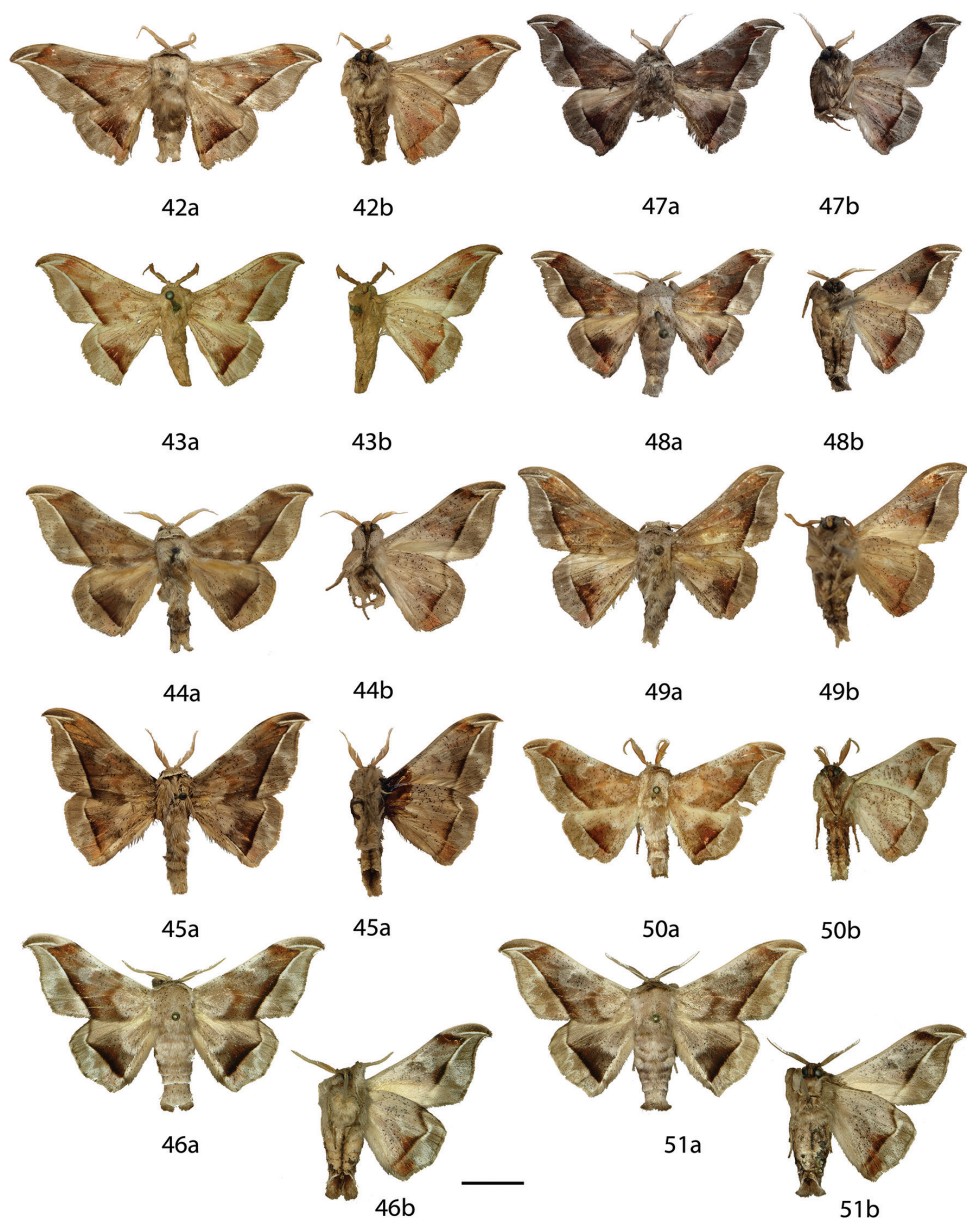
Type material. Holotype, ♂, presumed lost/destroyed. Type locality: Brazil: Rio de Janeiro (see remarks).

Neotype (here designated), ♂: BRAZIL: Rio de Janeiro: BRAZIL: Rio de Janeiro Ste., Teresopolis [Teresópolis], 13–22.iii.1958, H.B.D. Kettlewell, B.M. 1958–273/ NEOTYPE male *Mimallo plagiata* designated by St Laurent and Dombroskie 2016/ BMNH(E) 1378747/ St. Laurent diss.: 9-2-15:1/ (NHMUK). New type locality: Brazil: Rio de Janeiro: Teresópolis.

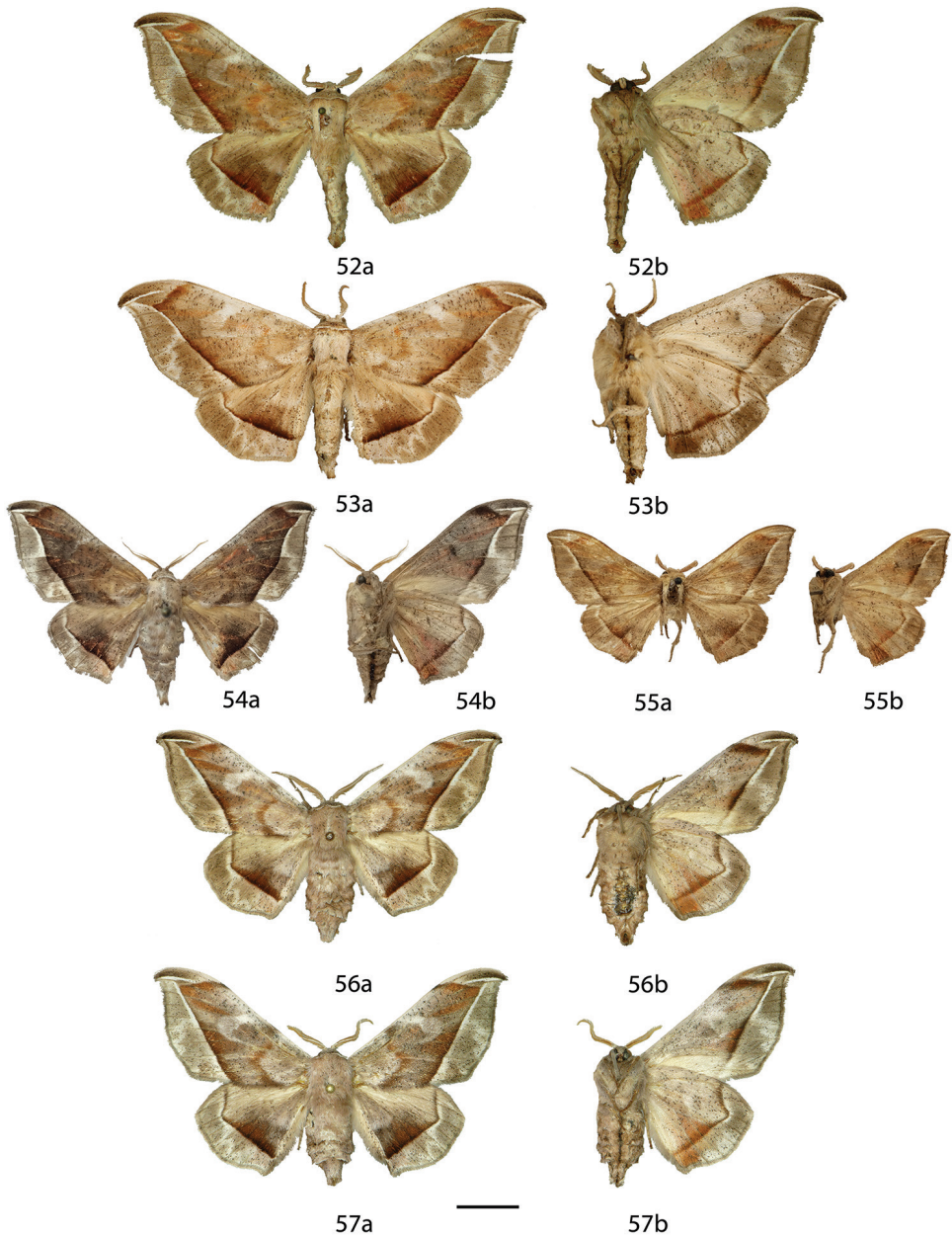
Additional specimens examined. (10 ♂, 19 ♀ total) **BRAZIL: Espírito Santo:** 3 ♀, Santa Teresa: 20.XI.1966, 18.XII.1966, Elias leg. (DZUP); XII.1966, C. & C.T. Elias (DZUP). **Pernambuco:** 1 ♂, Serra de Communaty [southeastern Pernambuco]: 1.II.1893, E. Gounelle, Ex. Oberthür Coll., Brit. Mus. 1927–3, ex. Joicey Coll. Brit. Mus. 1925–157, BMNH(E) 1378748, St. Laurent diss.: 6-29-15:12 (NHMUK). 1 ♀, Canhotinho: 19.VII.1991, Cardoso leg., ex. coleção A. Cardoso, No. 5467 (DZUP). **Rio de Janeiro:** 3 ♂, Teresópolis: 13–22.III.1958, H.B.D. Kettlewell, B.M. 1958–273, BMNH(E) 1378746, 1378751, 1378752, St. Laurent diss.: 6-29-15:13, 6-29-15:14 (NHMUK). 3 ♂, 4 ♀, Petrópolis: 2.XII.1875, 10.XII.1875, 29.III.1876, Joicey Bequest, Brit. Mus. 1934–120, BMNH(E) 1378749, St. Laurent diss.: 6-29-15:15 (NHMUK); 10.IV.1928, 6.IV.1959, 26.IV.1960, 23.X.1964, Gagarin leg., ex. col. Gagarin (DZUP). 1 ♂, Petrópolis, Independência: 21.IX.1939, Gagarin leg., ex. col. Gagarin (DZUP). 1 ♀, Mendes, 92 km from Rio de Janeiro: Collection Le Moul, Joicey Coll. Brit. Mus. 1925–157 (NHMUK). 2 ♀, Corcovado, 800 ft: II.1910, 2.II.1910, E.D. Jones, E.D. Jones coll. 1919–295, [1 ♀ is the “type” of manuscript name *Perophora ‡superba* D. Jones, BMNH(E) 805425] (NHMUK). 2 ♀, Eugenio de Dentro, E.F.C.B.: 1.I.1947, 4.I.1947, ex. pupa, Nelson Almeida, No. 19.174, 19.175 ex. coll. D’Almeida (DZUP). 1 ♀, Três Rios, Jacarepagua [additional illegible data]: 17.IX.1929, Ferr. d’Almeida, ex. coll. D’Almeida, No. 19.173 (DZUP). 1 ♀, “Penedo, Rezende” [either Penedo or Resende]: 4.X.1955, Coleção Richard Frey (DZUP). 2 ♂, 3 ♀, no additional locality data: Collection Wm Schaus, USNM-Mimal: 1202, St. Laurent diss.: 6-19-15:6 (USNM); 10.IX.1912, M.J. Holland, Carn. Mus. Acc. 4770, St. Laurent diss.: 4-25-15:2 (CMNH); Ex. Coll. J. Doll, Ac. 24352, St. Laurent diss.: 7-28-15:1 (AMNH); 12.IX.1943, Gagarin leg. ex. col. Gagarin (DZUP); IX.1944, ex. coleção A. Cardoso (DZUP). 1 ♀, additional locality information illegible: 5.I(?).1930, Ferreira d’Almeida, ex. coll. d’Almeida (DZUP).

Diagnosis. *Menevia plagiata* is recognizable from all previous species in both sexes by the replacement of the wing margin-swept postmedial lunule with a white band along the length of the postmedial line, which is interrupted midway and resumes near the inner margin. The female of *M. plagiata* is differentiated from the following two similar species by the presence of a straight or only weakly undulated postmedial line, which, along with the white accessory band, curves toward the apex of the forewing, sometimes sharply, usually to the wingtip, rather than ending at or before reaching the apical dash. Male genitalia are unlike any other species except *M. alurca*, in that the phallus bears a prominent, elongated, pointed projection from the dorsal surface and is not smooth or ridged as in all other previously diagnosed species. The nearly straight forewing postmedial lines (except near the apex) distinguish both male and female *M. plagiata* from *M. alurca*, whereas male genitalia characters and the apical curve of the female forewing postmedial line distinguish *M. plagiata* from *M. australis* sp. n.

Description. Male. *Head:* Gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by darker gray collar of scales reaching labial palpi, labial palpi very small, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. *Thorax:* As for genus. Light gray-brown. *Legs:* As for genus. Tibial spurs small to moderate in length, almost entirely scaled. *Forewing dorsum:* Forewing length: 22–24.5 mm, avg.: 22.4 mm, $n = 7$. Triangular, apical half of outer margins concave, apex falcate. Ground color gray-brown with darker gray, brown suffusion especially near interior edge of postmedial line and medial area, reddish coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong shape, thin gray mark connecting discal spot to costa. Apex marked by black scales above apical dash, especially near apical tip. Postmedial line straight or weakly undulated, line black, strongly contrasting. Submarginal area light gray with whitish suffusion mesally, postmedial lunule as white band originating from apical dash, white band follows postmedial line from apex to midway along postmedial line, resuming near anal margin. Antemedial line faint, brown, curved outwards. *Forewing venter:* As in forewing dorsum but grayer rather than brownish, sometimes with pinkish hue, black portion of postmedial line mostly absent except medially where very dark, white outer band of postmedial line as in dorsum, antemedial line absent. *Hindwing dorsum:* Subtriangular, anal angle weakly accentuated, reddish suffusion near anal angle, similar coloration and patterning as forewings, except postmedial lunule present as zigzagged mark, originating from white outer band along first quarter of postmedial line, postmedial line usually sharply bent toward anterior wing margin, sometimes weakly concave mesally. *Hindwing venter:* Following similar pattern as forewing venter, but red coloration near anal angle and medial area much darker. *Abdomen:* As for genus but elongated, nearly sphingiform, reaching beyond anal margin of hindwing. Coloration a continuation of gray thoracic color. Dark, contrasting, mid-ventral stripe present along entire length. *Genitalia:* (Figs 83, 84) $n = 7$. Somewhat variable; tegumen ovoid or rounded rectangular, sometimes weakly constricted near base of gnathos. Vinculum narrow, somewhat quadrate ventrally. Valves short, stocky, bent outwards or more elongated; saccular edge of left valve with large triangular tooth proximal to transtilla, sometimes notched mesally, right valve with tooth slightly reduced in size, both valves with prominent mesal costal projection originating from central ridge of valve, projection immediately above saccular edge teeth, apex of mesal projection pointed toward saccular edge. Valves triangular, rounded, or somewhat pointed apically. Uncus truncated apically, apex rounded. Gnathos as two prominent flattened, moderately sclerotized, flap-like, somewhat triangular, outward-facing extensions with truncated apices. Apices usually form fingerlike projections of varying length. Juxtal processes roughly phallus length, moderately sclerotized, curving toward apex of phallus. Juxtal processes very thin, long, widening distally, covered in fine setae, especially apically. Base of phallus with paired, backwards facing, elongated, rounded, diverging lobes sometimes with pointed tips on one or both lobes. Phallus irregularly shaped, unevenly edged dorsum lacking an extensive dorsal ridge but with prominent, elongated or sharply triangular, pointed protuberance, covered in setae, apical tip usually bent back-



Figures 42–51. *Menevia plagiata* species-group [*plagiata* subgroup] male adults, **a** recto, **b** verso. **42** *M. plagiata* neotype ♂, Brazil, Rio de Janeiro, Teresópolis (NHMUK) **43** *M. plagiata* ♂, Brazil, Rio de Janeiro, Petrópolis, Independência, 900 m [photo courtesy CGCM] (DZUP) **44** *M. plagiata* ♂, Brazil, Rio de Janeiro (CMNH) **45** *M. australis* holotype ♂, Brazil, Santa Catarina, Jaraguá do Sul (CUIC) **46** *M. australis* paratype ♂, Brazil, São Paulo, Guapiara, Paivinha, 800 m [photo courtesy CGCM] (CGCM) **47** *M. alurca* holotype ♂, Brazil, Maranhão, Feira Nova do Maranhão, 480 m (DZUP) **48** *M. alurca* paratype ♂, Brazil, Maranhão, Feira Nova do Maranhão, 480 m (CGCM) **49** *M. alurca* [transitional population] ♂, Brazil, Minas Gerais, Sete Lagoas, 720 m (USNM) **50** *M. alurca* [transitional population] ♂, Brazil, Minas Gerais, Paraopeba [photo courtesy CGCM] (OM) **51** *M. australis* paratype ♂, Brazil, São Paulo, Guapiara, Paivinha, 800 m [photo courtesy CGCM] (CGCM). Scale bar = 1 cm.



Figures 52–57. *Menevia plagiata* species-group [*plagiata* subgroup] female adults, **a** recto, **b** verso. **52** *M. plagiata* ♀, Brazil, Rio de Janeiro, Petrópolis [photo courtesy CGCM] (DZUP) **53** *M. plagiata* ♀ [“holotype” of manuscript name ‡*superba* Jones, photo courtesy NHMUK], Brazil, Rio de Janeiro, Corcovado, 800 ft. (NHMUK) **54** *M. alurca* ♀, Brazil, Maranhão, Feira Nova do Maranhão, 480 m (CGCM) **55** *M. alurca* ♀ [questionable], Argentina, Formosa (USNM) **56** *M. australis* paratype ♀, Brazil, Santa Catarina, São Bento do Sul, Rio Natal, 450 m [photo courtesy CGCM] (CGCM) **57** *M. australis* paratype ♀, Brazil, São Paulo, Guapiara, Paivinha, 800 m [photo courtesy CGCM] (CGCM). Scale bar = 1 cm.

wards. Left edge of rolled phallus uneven, forming extended protuberance, right edge usually with setae covered bulge laterally; base of sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of phallus. Distal tip of phallus separated into two distinct points of varying length. Vesica elongated. **Female.** *Head:* As in male. *Thorax:* As in male. *Legs:* As in male. *Forewing dorsum:* Forewing length: 27.5 mm, n = 1. Maculation as in male, wing broader, more ovoid, less triangular, postmedial line may be bent slightly outward mesally, outer white band of postmedial line curved toward apex, continuously to wingtip, forming very acute angle at junction with apical dash, dark scaling above apical dash usually concentrated near apical tip. *Forewing venter:* As in forewing dorsum but grayer. *Hindwing dorsum:* As in male but more rounded, less triangular, postmedial line straight except for sharp turn towards anterior wing margin, not concave mesally. *Hindwing venter:* Following similar pattern as forewing venter, reddish-brown suffusion near anal angle much darker. *Abdomen:* As in male but more robust. Sternite of VIII as pair of elongated sclerotized bands curving toward each other near anterior edge of VIII segment, but never converge. *Genitalia:* (Fig. 98) n = 1. Tergite of VIII forms triangular, posteriorly directed arc. Apophyses anteriores shorter than apophyses posteriores, apophyses very thin. Lamella antevaginalis thin, C-shaped, weakly notched mesally near ostium bursae. Ductus bursae short. Papillae anales subtriangular, covered in relatively long setae.

Distribution (Map 4). *Menevia plagiata* is found in the Brazilian Atlantic Forest. Most records come from the state of Rio de Janeiro, but records also exist from farther north in Espírito Santo and Pernambuco states. The distribution of this species probably extends along the entire coast of Brazil north from the state of Rio de Janeiro.

Natural history. A number of host records exist in the literature for *M. plagiata sensu lato*, but due to the uncertainty of the identification of this taxon in the past (see remarks below), we cannot be certain that all of the following records pertain to *M. plagiata sensu stricto*. Perhaps the most reliable record comes from Raymundo (1919) because this author figures the adult of true *M. plagiata* and describes its distribution as only including the Brazilian states of Rio de Janeiro and Espírito Santo, which eliminates the more southerly distributed *M. australis* sp. n. and the primarily Amazonian *M. vulgaris* sp. n., both described below. Raymundo (1919), Monte (1934), and Lima (1950) mention only *Terminalia catappa* (Combretaceae) as a host of *M. plagiata*, a species we previously showed to be a host of *M. ostia*. Additional, less verifiable host plant records include: *Psidium guajava* (Myrtaceae), *Licania tomentosa* (Chrysobalanaceae), and even *Araucaria angustifolia* (Araucariaceae) all cited in Silva et al. (1968). Silva et al. (1968) refers to “pinheiro” (Pine), which we interpret to mean *Araucaria angustifolia* because this host is listed by Mecke et al. (2000) for *M. plagiata*, citing Silva et al. (1968). Additionally, Pastrana (2004) mentions *Prunus amygdalus* (Rosaceae) as a host for *M. plagiata*, but this is probably an erroneous misinterpretation of Lima (1950) who had listed “amendoeira (*Terminalia catappa*)” as the host of *M. plagiata*, referring to amendoeira da praia (*T. catappa*), not true amendoeira (almond, *P. amygdalus*).

Remarks. *Menevia plagiata* is the most problematic taxon in the genus, largely due to the unavailability of the holotype, which is presumed to be lost. The holotype of *M.*



Map 4. *Menevia plagiata* species-group [*plagiata* subgroup].

plagiata originated from Fry's collection, and was collected in Rio de Janeiro. Becker (2001) provided information pertaining to the history of the many specimens from the Fry Collection, collected in Rio de Janeiro, later described by Walker, that were subsequently damaged, and are apparently now lost. We attempted to locate the holotype of *M. plagiata* at the NHMUK but were unsuccessful. Contacting the University Museum, Oxford in an effort to locate the holotype there was also unsuccessful. Although it is not impossible that this type remains undiscovered somewhere in these or other collections, we consider it unlikely to be located, thus we here designate a neotype for this species based on information discussed below.

Walker's (1855) original description of *M. plagiata* is rather vague and could arguably be attributed to either *M. plagiata* or *M. franclemonti* sp. n. as both of these species are found in Rio de Janeiro and are somewhat similar in appearance. The most important line in Walker's description is that relating to the white band, which follows the postmedial line in both *M. plagiata* and *M. franclemonti* sp. n. This band is interrupted in the former and continuous in the latter. Walker (1855) states in his original description of *M. plagiata* that there is a "very oblique slender white band at three-fourths of the length, forming a very acute angle near the tip," which we interpret to mean the white band following the exterior of the postmedial line. However, Walker does not mention whether the white band is continuous or interrupted along

the length of the postmedial line, which is necessary information to attribute this description definitively to either species. Walker's description of *Mimallo saturata* Walker, 1855 from the same work offers some characters that could be attributed to what we consider *M. plagiata*, in which the most important again is the white band along what is assumed to be the postmedial line: "a whitish slender slightly oblique band, which has a blackish border on its outer side, and extends from near the tip of the costa to three-fourths of the length of the interior border" (Walker 1855). The holotype of *M. saturata* is also presumed lost due to it originating from Fry's collection and having a type locality of Rio de Janeiro.

An additional taxonomic issue was created by the presence of the "holotype" of *Perophora* ‡*superba* Jones in the NHMUK. The name ‡*superba* is a manuscript name and was never published by Jones. It is possible that Jones realized the similarity of ‡*superba* to *M. plagiata* as described by Walker, and hence did not describe it. This assumption is an additional piece of evidence supporting our concept of *M. plagiata* because the "holotype" of ‡*superba* matches it. If a validly published description using the name ‡*superba* is located, it would have to be treated as a junior synonym of *M. plagiata*.

After visiting the NHMUK and reviewing the specimens belonging to the *M. plagiata* species-group, it was clear that all Rio de Janeiro specimens matched our concept of *M. plagiata*, including a number of specimens from the late 1800's. The other species that the name *plagiata* could be associated with, which we describe below as *M. franclemonti* sp. n., is rarer relative to *M. plagiata* and was not present in the NHMUK. Schaus (in Seitz 1928) illustrated our concept of *M. plagiata* as this species, and did not figure anything resembling *M. franclemonti* sp. n. Therefore; it seems most probable that Walker had a specimen matching our concept of *M. plagiata* at his disposal when writing his description. Finally, one of the oldest *M. plagiata* determinations in the NHMUK, from 1889 by C. Berg, was a female specimen matching our concept of *M. plagiata*. This provides an indication that as early as 1889 the name *plagiata* was being associated with the species that we consider to be *M. plagiata*.

The taxonomic history of the name *plagiata* is surely complicated, and unfortunately, without seeing the holotype, we might never be completely sure that our concept of the species coincides with that of Walker's (1855). Complications compound further when considering the great deal of variation, both externally and in male genitalia morphology, that *M. plagiata* displays within the state of Rio de Janeiro. The considerable amount of variation suggests that *M. plagiata sensu lato* may represent a species complex. However, the four examined NHMUK specimens from Teresópolis, Rio de Janeiro, including the neotype, represent a cohesive series in terms of external characteristics and genitalia, with very little variation. The valves of these specimens are much stouter and the projection of the phallus shorter than in other *M. plagiata* from nearby locations in the state of Rio de Janeiro. Therefore, to stabilize the nomenclature, we here designate the neotype, chosen from this series. A specimen from Petrópolis, Rio de Janeiro at DZUP also greatly resembles the series of four *M. plagiata* from Teresópolis (see Fig. 43).

***Menevia alurca* Herbin & Mielke, 2014**

Figs 47–50, 54, 55, 85, 99; Map 4

Menevia alurca Herbin & Mielke, 2014: 146–147; figs ♂ 48, 49, ♂ genitalia 50*Menevia ulcara*; Herbin and Mielke 2014: 147 misspelling

Type material. Holotype, ♂: **BRAZIL: Maranhão**: genitalia prep. D. Herbin, ref H. 1008 [blue label]/ BRESIL Maranhao [Maranhão], Feira Nova do Maranhão, Retiro, 480 m. 4–12.XI.2011, -07°00'31"S, -46°26'41"W, C. Mielke leg. Coll. D. Herbin / Holotype ♂, *Menevia alurca* Herbin & Mielke det., Antenor 2014 [red label]/ DZ 15.727/ (DZUP) [examined]. Type locality: Brazil: Maranhão: Feira Nova do Maranhão.

Paratypes, 7 ♂ (6 specimens fit our concept of *M. alurca*, 1 specimen fits our concept of *M. delphinus* sp. n., see remarks under *M. delphinus* sp. n.), **BRAZIL: Maranhão**: 1 ♂, Feira Nova do Maranhão, Retiro, 46°26'41"W, -07°00'31"S, 480 m: 16–17. II.2013, C. Mielke leg., Paratypus *Menevia alurca* Herbin & C. Mielke det., 2014 [green label], Col. C. Mielke 26.882, St. Laurent diss.: 6-16-15:4 (CGCM) [examined]. Photos of all additional paratypes, each from the same locality and collector as the holotype and single physically examined paratype, were provided by Herbin and Mielke.

Additional specimens examined. (4 ♂, 1 ♀ total) **BRAZIL: Distrito Federal**: 4 ♂, Estação Florestal, Cabeça do Veado, 1100 m: 17.X.1971, 19.X.1971, 21.X.1971, 24.X.1971, E.G., I. & E.A. Munroe, St. Laurent diss.: 5-13-15:2 (CNC). **Maranhão**: 1 ♀, Feira Nova do Maranhão, Retiro, W 46°26'41", S -07°00'31", 480 m: 23–24. XI.2013, C. Mielke leg., St. Laurent diss.: 7-28-15:2 (CGCM).

Additional transitional and questionable specimens examined. (3 ♂, 1 ♀ total) **ARGENTINA**: 1 ♀, Formosa: data illegible except for locality, USNM-Mimal: 1203, St. Laurent diss.: 5-13-15:1 (USNM). **BRAZIL: Minas Gerais**: 2 ♂, Paraopeba: 27.II.1966 (OM). 1 ♂, Sete Lagoas, 720 m: 16.III.1974, V.O. Becker col., Col. Becker No. 413, USNM-Mimal: 2342, St. Laurent diss.: 5-13-15:3 (USNM).

Diagnosis. *Menevia alurca* is distinguishable from all previous species in both sexes by the slate gray coloration suffused with deep blood-red, and the postmedial line, which is usually bent outward toward the wing margin at about three-quarters of its length. Additionally, the white band along the exterior of the postmedial line does not curve sharply toward the wing apex as it does in *M. plagiata*, but instead ends where it meets the apical dash. In this respect, *M. alurca* is similar to *M. australis* sp. n.; however, the white band usually juts out sharply just before approaching the apical dash in *M. alurca*, which is not seen in most other *Menevia*. The male genitalia however, should immediately distinguish this species from *M. australis* sp. n.; the phallus bears a prominent, elongated, pointed protuberance from the dorsal surface, not a rounded hump as in *M. australis* sp.n. The female genitalia differ from those of both *M. plagiata* and *M. australis* sp. n. by the small size, the very thin abdominal sclerotizations, and the mesally creased lamella antevaginalis.

Description. Male. Head: Gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by darker gray collar of scales reaching labial palpi, labi-

al palpi very small, dorsally with darker scales contrasting with overall gray coloration. Antenna yellowish, scape and pedicel weakly tufted. *Thorax*: As for genus. Gray. *Legs*: As for genus. Tibial spurs moderate in length, scaled except for distal quarter. *Forewing dorsum*: Forewing length: 18–21.5 mm, avg.: 19.7 mm, n = 5. Triangular, apical half of outer margin concave, apex falcate. Ground color dark gray with predominance of deep red-brown or blood-red throughout medial area, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray, oblong shape; thin gray mark connects discal spot to costa. Apex marked by black scales above apical dash. Postmedial line usually bent outwards along three-fourths of its length, rarely nearly straight [especially in transitional population]. Submarginal area light gray, contrasting with much darker medial area, with whitish suffusion mesally forming a faint zigzag, postmedial lunule as distinct white band originating from apical dash, white band follows postmedial line from apex to roughly midway along postmedial line, resuming near anal margin. Antemedial line faint, brown, curved outwards. *Forewing venter*: As in forewing dorsum but generally much grayer, sometimes with pinkish hue, antemedial line absent, small black discal mark occasionally present. *Hindwing dorsum*: Subtriangular, anal angle accentuated, reddish coloration usually present near anal angle, bleeding into medial area, similar coloration and patterning overall as forewings, except postmedial lunule present as zigzagged mark, originating from white outer band outlining anterior bend of postmedial line, postmedial line weakly curved toward anterior wing margin, sometimes weakly concave mesally. *Hindwing venter*: Following similar pattern as forewing venter, but red coloration near anal angle much darker. *Abdomen*: As for genus, but somewhat stouter. Coloration a continuation of gray thoracic color. Dark, contrasting midventral stripe present. *Genitalia*: (Fig. 85) n = 4. Somewhat variable; tegumen ovoid or somewhat rectangular, sometimes weakly constricted near base of gnathos. Vinculum broad, somewhat quadrate ventrally. Valves broad at base, triangular, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth slightly reduced in size, both valves with smaller mesal costal projection originating from central ridge of valve, mesal costal projection immediately above saccular edge teeth, apex of projection pointed toward saccular edge. Valves may be truncated distally, rounded apically. Uncus truncated apically, apex rounded. Gnathos as two prominent flattened, moderately sclerotized, flap-like, barely triangular, upward facing extensions with truncated apices. Apices usually form fingerlike projections of varying length. Juxtal processes roughly phallus length, moderately sclerotized, curving toward apex of phallus. Juxtal processes very thin, widening distally, covered in fine setae, especially apically. Base of phallus with paired, backwards facing, elongated, rounded, diverging fingerlike lobes. Phallus irregularly shaped, unevenly edged dorsum lacking an extensive dorsal ridge but with prominent, elongated, pointed setae covered projection, tip bent backwards. Left edge of rolled phallus uneven, forming extended projection, right edge usually with setae covered bulge laterally, base of sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of phallus, distal tip of phallus separated into two distinct points of varying length. Vesica somewhat bag-like. **Female**. *Head*: As in male, dark gray scales surrounding

eyes reduced. *Thorax*: As in male. *Legs*: As in male, tibial spurs longer. *Forewing dorsum*: Forewing length: 19.5–26 mm, $n = 2$. Maculation as in male but grayer with less red-brown, wing broader but still subtriangular. *Forewing venter*: As in forewing dorsum but lighter gray, dark discal mark present. *Hindwing dorsum*: As in male but more rounded, less triangular, postmedial line bent more sharply toward anterior wing margin. *Hindwing venter*: Following similar pattern as forewing venter, reddish-brown suffusion near anal angle much darker, contrasting. *Abdomen*: As in male but more robust. Sternite of VIII as pair of thin, nearly parallel, elongated, sclerotized bands. *Genitalia*: (Fig. 99) $n = 2$. Tergite of VIII robust, forming triangular, posteriorly directed arc. Apophyses anteriores shorter, thicker than apophyses posteriores. Lamella antevaginalis moderate in thickness, with mesal crease at ostium bursae. Ductus bursae short. Papillae anales elongated, subtriangular, covered in relatively long setae.

Distribution (Map 4). *Menevia alurca* is primarily found in the Brazilian Cerrado, with records from Maranhão, Distrito Federal, and central Minas Gerais. The Minas Gerais population may represent an introgression zone with the similar *M. plagiata* of the Atlantic Forest. This species may also be present in Argentina, as suggested by the single questionable specimen discussed below.

Remarks. Herbin and Mielke (2014) considered this species to be of the same size as *M. lucara*, although we found *M. alurca* larger on average. *Menevia plagiata*, which we show to be the closest species to *M. alurca* based on external characters and genitalia, was indeed mentioned in their diagnosis of *M. alurca*, but the authors did not provide differentiating characters other than that *M. plagiata* is larger. *Menevia alurca* is easily distinguished from *M. lucara* by all characters presented by Herbin and Mielke (2014). However, the male genitalia of *M. alurca* are nearly indistinguishable from those of *M. plagiata* and individuals of *M. alurca* that have straighter postmedial lines are externally very similar to *M. plagiata*.

Upon reviewing each of the seven paratypes of *M. alurca*, it became apparent that one paratype in the collection of Herbin (Bc-Her4848) does not belong to this species and was incorrectly included in the paratype series. The specimen in question has a continuous white band along the forewing postmedial line and lacks a midventral stripe on the abdomen, characters that allow us to identify this specimen as *M. delphinus* sp. n., another Cerrado species, described below.

External characteristics and male genitalia, namely, the phallus, of *M. alurca* is extremely similar to that of *M. plagiata*, thus these species are not readily differentiable based on male genitalia alone, as are most *Menevia* species. Furthermore, three specimens from two nearby localities in central Minas Gerais that we questionably determined as *M. alurca*, are somewhat intermediate in habitus between *M. alurca* from farther north and west (central Cerrado), and *M. plagiata* from farther east and south-east (Brazilian Atlantic Forest). These specimens display the predominance of darker coloration and the arrangement of markings near the forewing apices that we attribute to *M. alurca*. However, the forewing postmedial line is only weakly curved outwards, not bent so dramatically as in the types of *M. alurca*. These specimens are also quite large, much more in line with those of *M. plagiata*. The population from Minas Gerais

is transitional in appearance between *M. alurca* and *M. plagiata*, and it is worth noting that the Cerrado and Atlantic Forest biomes converge there (IBGE 2004).

A female that we questionably assign to this species, from Formosa, Argentina, is extremely small and has a nearly straight forewing postmedial line. Although the size and postmedial line are not comparable to the other female *M. alurca*, the markings at the forewing apex, the predominance of red coloration, and genitalia (namely the mesally creased lamella antevaginalis), are all highly suggestive of *M. alurca*. Without more material from this locality, particularly males, it is impossible to reach conclusive determination about its identity. We decided to include this specimen in our material examined due to its collecting locality, which represents the only *Menevia* record from Argentina.

***Menevia australis* sp. n.**

<http://zoobank.org/F0C3F909-9FBF-4CBE-976E-FA9C903FE77D>

Figs 45, 46, 51, 56, 57, 86, 100; Map 4

Type material. Holotype, ♂: **BRAZIL: Santa Catarina:** Jaragua [Jaraguá do Sul], Santa Catharina, Brazil, 5 Sept 1934, Fritz Hoffmann/ **PARATYPE** *Menevia elegans* J. G. Franclemont/ St. Laurent diss.: 6-19-15:4/ Franclemont's label refers to a MS name./ **HOLOTYPE** male *Menevia australis* St Laurent and Dombroskie, 2016 [hand-written red label]/ (CUIC). Type locality: Brazil: Santa Catarina: Jaraguá do Sul.

Paratypes, 19 ♂, 9 ♀: **BRAZIL: Santa Catarina:** 3 ♂, São Bento do Sul, Rio Vermelho, 850 m: 16.IX.1985, Mielke, Rank & Casagrande leg., DZ 32.713–32.715 (DZUP). 1 ♀, São Bento do Sul, Rio Natal, 26°20'2"S, 49°18'30"W, 450 m: 2.X.2014, I. Rank leg, Col. Mielke 28.976 (CGCM). 5 ♂, 2 ♀, Jaraguá do Sul: 13.IX.1934, 14.IX.1934, 15.IX.1934, 19.IX.1934, 6.X.1934, 8.X.1934, Fritz Hoffmann, Franclemont diss.: 1761, 1762, St. Laurent diss.: 4-25-15:3, 6-19-15:5 (CUIC). 1 ♂, 1 ♀, Nova Bremen [Dalbérgia], 250 m: 17.IX.1933, 26.III.1934, Carn. Mus. Acc. 11040, St. Laurent diss.: 7-30-15:1 (CMNH). 1 ♂, 1 ♀, no additional locality data: USNM-Mimal: 2434, 2851, St. Laurent diss.: 4-25-15:1 (USNM). **São Paulo:** 1 ♀, Campinas: 1.IV.1902, Coll. A. Hempel, Holland Collection, St. Laurent diss.: 4-25-15:4 (CMNH). 9 ♂, 3 ♀, Guapiara, Paivinha, 800 m: 12.IX.2007, 16–19.IX.2005, C. Mielke leg, 25.777, 25.811, 26.498, 26.567, 26.573, 27.126, 27.134, 28.833, 28.841, 28.869, 28.880, 28.906 Col. C. Mielke, C. Mielke diss.: 25.811 (CGCM). – All paratypes with the following yellow label: **PARATYPE** male/female *Menevia australis* St Laurent and Dombroskie, 2016.

Diagnosis. *Menevia australis* is similar to *M. plagiata* in both sexes, but can be distinguished in both sexes by slightly broader wings, deeper brown coloration, and by the white band along the postmedial line, which terminates at the apex somewhat mesally along the length of the apical dash. The white band is either nearly perpendicular to the apical dash or it forms a roughly 45 degree angle with it. The white band may be somewhat curved toward the apex as in *M. plagiata*, but does not reach the apical tip of the wing. Male genitalia are easily recognized by the rounded hump or short triangular

protuberance on the dorsal surface of the phallus, easily distinguishing the male genitalia from those of *M. plagiata* and *M. alurca*, which both have very elongated dorsal protuberances on the phallus. Additionally, the uncus is narrower and more triangular in *M. australis*. The VIII tergite in the female genitalia of *M. australis* forms a rounded arc and is not triangular as in *M. plagiata* and *M. alurca*.

Description. Male. *Head:* Gray-brown or light brown, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by darker brown collar of scales reaching labial palpi, labial palpi very small, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. *Thorax:* As for genus. Light gray-brown to light brown. *Legs:* As for genus. Tibial spurs small to moderate in length, almost entirely scaled. *Forewing dorsum:* Forewing length: 21.5–23 mm, avg.: 22.4 mm, $n = 7$. Triangular, apical half of outer margin concave, apex falcate. Ground color gray-brown with caramel brown or almost slate gray suffusion throughout medial area, reddish coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong shape, thin gray mark connecting discal spot to costa. Apex marked by black scales above apical dash, especially near apical tip. Postmedial line straight or weakly undulated, line black, strongly contrasting. Submarginal area light gray with whitish suffusion mesally forming faint or conspicuous zigzag, postmedial lunule as white band originating mesally from apical dash, white band follows postmedial line from apex to midway along postmedial line becoming zigzagged diffusion, white band resumes near anal margin. Antemedial line faint, brown, curved outwards. *Forewing venter:* As in forewing dorsum but grayer rather than brownish, antemedial line absent. *Hindwing dorsum:* Subtriangular, anal angle weakly accentuated, reddish coloration usually present near anal angle, similar coloration and patterning as forewings, except postmedial lunule present as zigzagged mark similar to zigzagged diffusion on forewing, mark originating from white outer band along first quarter of postmedial line, postmedial line sharply bent toward anterior wing margin, sometimes weakly concave mesally. *Hindwing venter:* Following similar pattern as forewing venter, but red coloration near anal angle much darker, almost brown. *Abdomen:* As for genus but somewhat elongated, nearly sphingiform. Coloration a continuation of gray thoracic color. Dark, contrasting midventral stripe present. *Genitalia:* (Fig. 86) $n = 6$. Tegumen ovoid, weakly constricted near base of gnathos. Vinculum rectangular, somewhat quadrate ventrally. Valves triangular, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth slightly reduced in size, both valves with central tooth originating from central ridge of valve, tooth immediately above saccular edge teeth, apex of central tooth pointed toward saccular edge. Valves truncated distally, bent slightly outward near apex, rounded apically. Uncus narrow, triangular, apex rounded. Gnathos as two prominent flattened, moderately sclerotized, flap-like, somewhat triangular, outward-facing extensions with truncated apices. Apices usually form fingerlike projections of varying length. Juxtal processes roughly phallus length, moderately sclerotized, curving toward apex of phallus. Juxtal processes very thin, flattened, covered in fine setae. Base of phallus with paired, backwards facing, moderately elongated, rounded, diverging

lobes. Phallus irregularly shaped, unevenly edged dorsum with prominent rounded, triangular, or somewhat rectangular setae covered hump. Left edge of rolled phallus uneven, forming hump; right edge usually with setae covered bulge laterally, base of sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of phallus, distal tip of phallus separated into two distinct points of varying length. Vesica small, sac-like. **Female.** *Head:* As in male. *Thorax:* As in male. *Legs:* As in male, tibial spurs stouter. *Forewing dorsum:* Forewing length: 29.5–32 mm, avg: 30.3 mm, $n = 1$. Maculation as in male, wing broader, more ovoid, less triangular, outer white band of postmedial line intercepts apical dash mesally or dissipates before reaching mark, dark scaling above apical dash spread over length of apical dash. *Forewing venter:* As in forewing dorsum but usually grayer. *Hindwing dorsum:* As in male but more rounded, less triangular. *Hindwing venter:* Following similar pattern as forewing venter, reddish-brown suffusion near anal angle much darker, contrasting. *Abdomen:* As in male but more robust. Sternite of VIII as pair of elongated, broad or very narrow sclerotized bands curving toward each other near anterior edge of VIII segment, but not converging. *Genitalia:* (Fig. 100) $n = 2$. Tergite of VIII forms curved, rounded, posteriorly directed arc. Apophyses anteriores shorter than apophyses posteriores. Lamella antevaginalis thin, C-shaped, weakly notched mesally near ostium bursae. Ductus bursae narrow. Papillae anales subtriangular, covered in setae.

Distribution (Map 4). This new species is so far known only from southeastern Brazil in the northeast of Santa Catarina state and eastern São Paulo state. *Menevia australis* is likely present in intervening eastern Paraná as well.

Etymology. *Menevia australis* is named for its southerly distribution, which among *Menevia*, is only shared with *M. magna*. Additionally, *M. australis* seems to represent the southeast most extension of the *M. plagiata* species complex, replacing *M. plagiata* farther southeast.

Remarks. *Menevia australis* is the southeasternmost species of the *M. plagiata* complex and is quite difficult to separate from true *M. plagiata* without a genitalia dissection or geographic information. However, upon thorough examination of the male and female genitalia, external diagnostic characters became readily apparent and have been presented above in the diagnosis. Additionally, the allopatric distribution of these species suggests that they are two separate species, albeit very closely related. The allopatric distribution patterns of *M. australis* and *M. plagiata* are not unique, similar allopatry was shown in two closely related Saturniidae by Mielke et al. (2012) wherein there is a distinct gap in eastern São Paulo state. This gap is probably not due to lack of collecting as the region has been extensively sampled (C. Mielke pers. comm.).

Both *M. australis* and *M. franclemonti* sp. n. described below, were originally recognized as distinct by J. G. Franclemont, and given manuscript names, but never formally described. The holotype and some paratypes of *M. australis* (all from Jaraguá do Sul, Santa Catarina, Brazil) bear labels reading “PARATYPE,” “HOLOTYPE,” or “ALLOTYPE” with Franclemont’s manuscript name *Menevia ‡elegans*. In addition to our holotype and paratype labels, we have placed labels on these specimens stating that Franclemont’s labels represent a manuscript name.

***vulgaris* subgroup**

The *vulgaris* subgroup contains *M. vulgaris* sp. n., *M. franclemonti* sp. n., *M. vulgaricula* sp. n., *M. cordillera* sp. n., and *M. delphinus* sp. n. and is diagnosed by the continuous white band along the outer margin of the postmedial line and by the lack of a midventral abdominal stripe. The species boundaries in the *vulgaris* subgroup are clearer than in the previous subgroup.

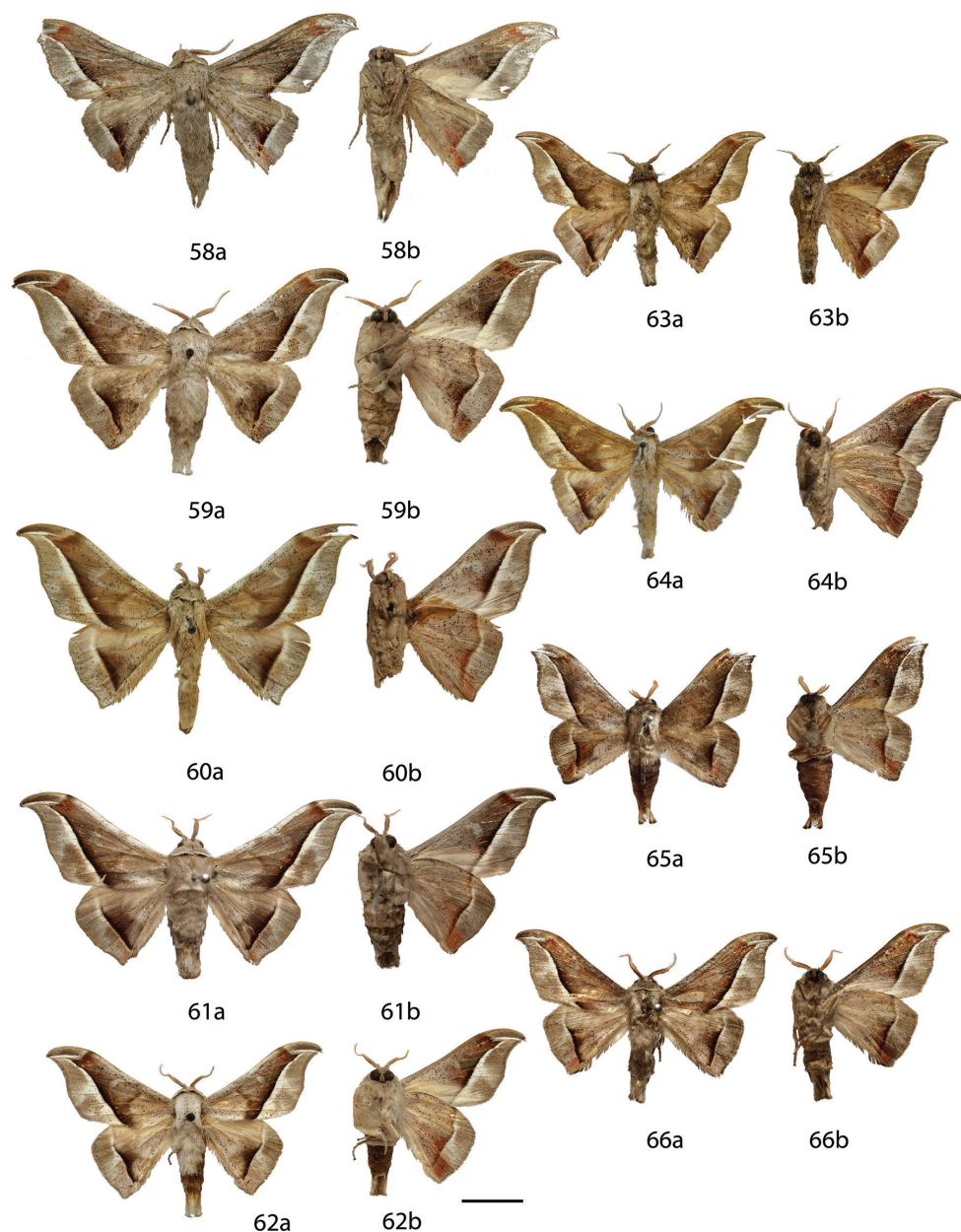
***Menevia vulgaris* sp. n.**

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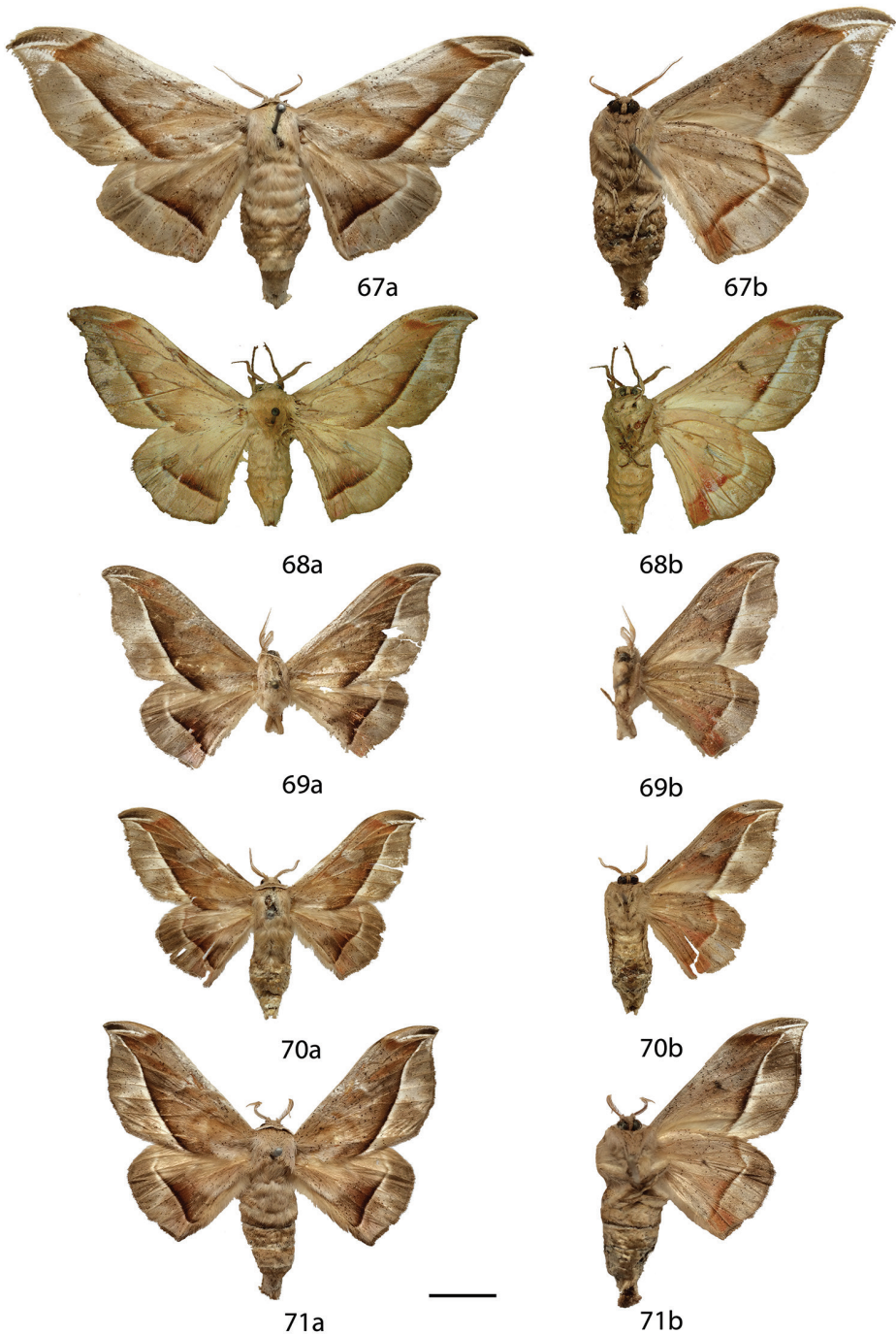
Figs 58, 59, 67, 87, 101; Map 5

Type material. **Holotype**, ♂: **FRENCH GUIANA:** GUYANE, St. Laurent-du-Maroni, Piste de Paul Isnard, N05°22.562', W53°57.678', 8.iii.2011, 52 m., P. Sammut/ *MENEVIA plagiata plagiata* (Walker, 1855)/ St. Laurent diss.: 9-2-15:2/ **HOLOTYPE** male *Menevia vulgaris* St Laurent and Dombroskie, 2016 [handwritten red label]/ (CUIC). Type locality: French Guiana: St. Laurent du Maroni.

Paratypes, 30 ♂, 10 ♀: **BRAZIL: Amazonas:** 8 ♂, Reserva Ducke, km. 26 Manaus-Itacoatiara Highway: 15.IV.1972, 16.IV.1972, 19.IV.1972, 15.V.1972, 16.V.1972, E.G., I. & E.A. Munroe, St. Laurent diss.: 4-25-15:8, 8-3-15:1 (CNC). 1 ♂, Itacoatiara Airport: 6.V.1972, E.G., I. & E.A. Munroe (CNC). 1 ♂, Mirapinima, Rio Negro: 8.IV.1972, E.G., I. & E.A. Munroe (CNC). 7 ♂, Hyutanahan [Huitanaã], Rio Purus: II.1922, III.1922, S.M. Klages, Carn. Mus. Acc. 6963, 7088, St. Laurent diss.: 4-25-15:10, 7-20-15:1, 7-20-15:2, 7-23-15:1 (CMNH). **Goiás:** 1 ♂, Campinas [Goiânia]: I.1934, Coll. R. Spitz, Rothschild Bequest BM 1939-1, BMNH(E) 1378754 St. Laurent diss.: 6-29-15:9 (NHMUK). **Pará:** 1 ♂, 1 ♀, Likely Belém: Collector Moss, USNM-Mimal: 2592, 2597, St. Laurent diss.: 9-7-14:4, 4-25-15:12 (USNM). **COLOMBIA:** 3 ♂, Antioquia, Nari [Nare?] River: Collection Frank Johnson, USNM-Mimal: 2586, St. Laurent diss.: 4-25-15:11 (USNM); USNM-Mimal: 2587, St. Laurent diss.: 6-19-15:3 (USNM); USNM-Mimal: 2588 (USNM). 1 ♀, "Colombia, S.A.": Felipe Ovalle, Q., Ac. 33501 (AMNH). **ECUADOR:** 1 ♂, Napo [Orellana], near Pompeya (Yasuni Nat. Pk.), 00°38–40'S, 76°22–27'W, 280 m: 17–30.X.1998, D. Robacker, M.H. Evans Collection, St. Laurent diss.: 9-7-14:5 (CUIC). 1 ♂, Napo [Orellana], Parque Nacional Yasuni, 1 km. SE PUCE station, edge of virgin forest: 13.V.1996, Jan Hillman (CMNH). 1 ♂, Napo [Orellana], Parque Nacional Yasui, Rio Natali, 20 km. S PUCE station, near edge of virgin forest: 16.V.1996, Jan Hillman, St. Laurent diss.: 7-20-15:3 (CMNH). 1 ♀, Napo, Rio Arajuno, Camp Dayuma, 1°05'35"S, 77°35'07"W, 390–420 m: 19–22.IV.1990, S.J. Weller, P. Batra, & M.J. Ryan, St. Laurent diss.: 7-21-15:3 (USNM). **FRENCH GUIANA:** 1 ♂, 1 ♀, Mana River: V.1917, Acc. 6008, St. Laurent diss.: 4-25-15:9 (CMNH). 1 ♀, St. Laurent du Maroni: Collection Wm Schaus, USNM-Mimal: 2591, St. Laurent diss.: 4-25-15:13 (USNM). 1 ♀, Pied Saut, Oyapok River: III.1918, S.M. Klages, C.M.



Figures 58–66. *Menevia plagiata* species-group [*vulgaris* subgroup] male adults, **a** recto, **b** verso. **58** *M. vulgaris* holotype ♂, French Guiana, St. Laurent du Maroni, Piste de Paul Isnard, 52 m (CUIC) **59** *M. vulgaris* paratype ♂, Brazil, Huitanaã, Rio Purus (CMNH) **60** *M. franclemonti* holotype ♂, Brazil, Santa Catarina, Jaraguá do Sul (CUIC) **61** *M. franclemonti* paratype ♂, Brazil, Rio de Janeiro, Teresópolis, 350 m (USNM) **62** *M. vulgaricula* holotype ♂, Brazil, Huitanaã, Rio Purus (CMNH) **63** *M. cordillera* holotype ♂, Peru, San Gabán, 2500 ft (NHMUK) **64** *M. cordillera* paratype ♂, Bolivia, Río Zongo, 750 m (USNM) **65** *M. delphinus* holotype ♂, Brazil, Distrito Federal, Planaltina, 1000 m (CPAC) **66** *M. delphinus* paratype ♂, Brazil, Distrito Federal, Planaltina, 1000 m (USNM). Scale bar = 1 cm.



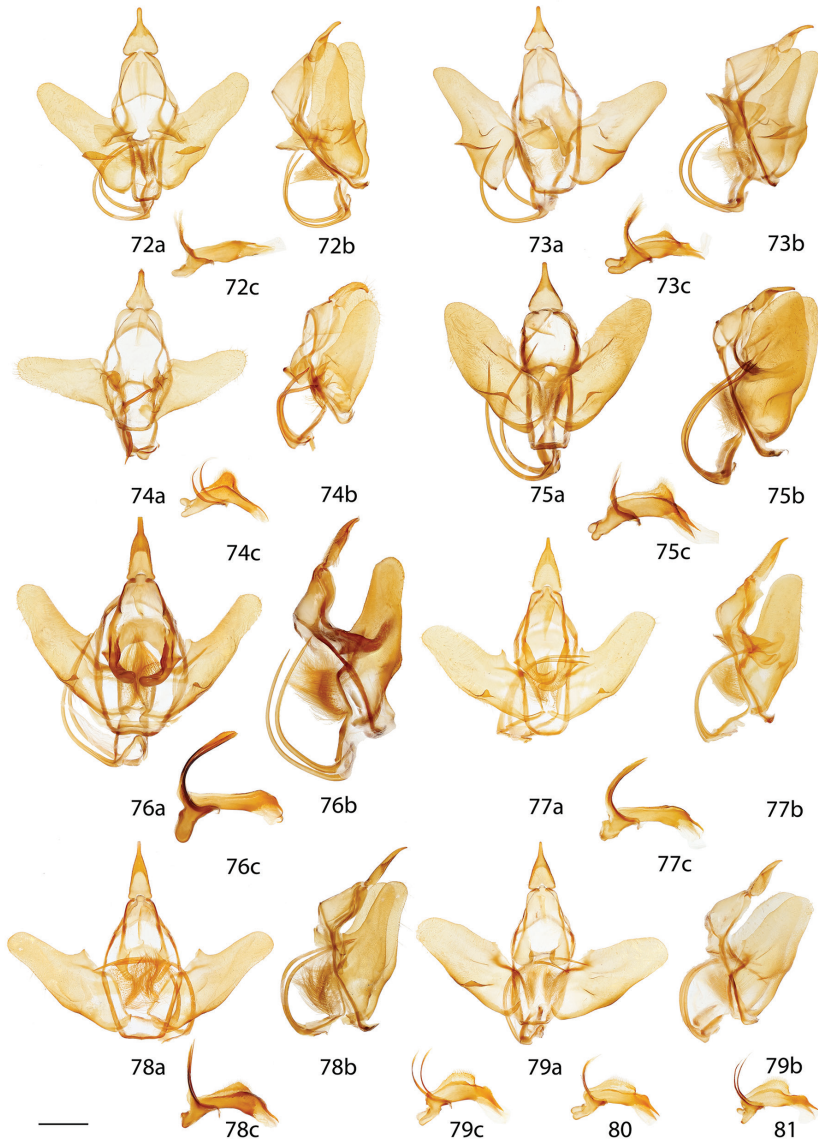
Figures 67–71. *Menevia plagiata* species-group [*vulgaris* subgroup] female adults, **a** recto, **b** verso. **67** *M. vulgaris* paratype ♀, Guyana, Omai (USNM) **68** *M. franclemonti* paratype ♀, Brazil, Rio de Janeiro, Teresópolis, Barreira [photo courtesy CGCM] (DZUP) **69** *M. vulgaricula* paratype ♀, Brazil, Amazonas, Rio Madeira (USNM) **70** *M. vulgaricula* ♀ [questionable], Brazil, Pará (NHMUK) **71** *M. delphinus* paratype ♀, Brazil, Distrito Federal, Planaltina, 1000 m (CPAC). Scale bar = 1 cm.

Acc. 6173, St. Laurent diss.: 7-21-15:1 (CMNH). 1 ♀, St. Jean du Maroni: II, Collection Le Moul, Dognin Collection, USNM-Mimal: 2590 (USNM). 1 ♀, Camp Patawa, Kaw Mountains, 36 km. E. of Roura: 21.XII.1997, leg. I.L. Finkelstein (MGCL). **GUYANA:** 1 ♀, Omai, Br. Guiana: Collection Wm Schaus, USNM-Mimal: 2589 (USNM). **PERU:** 1 ♀, Iquitos [Iquitos]: Dr. Luka Kassarov donation to FSCA collection (FSCA). **SURINAME:** 3 ♂, Moengo, Boven Cottica River: 25.V.1927, Cornell Univ. Lot 760, Sub 79, St. Laurent diss.: 12-10-13:2 (CUIC); 26.V.1927, Cornell Univ. Lot 760, Sub 80 (CUIC). **BRAZIL-PERU BORDER:** 1 ♂, “Amazons, Peru-Brazil”: 1930, H.S. Parish (CUIC). – All paratypes with the following yellow label: PARATYPE male/female *Menevia vulgaris* St Laurent and Dombroskie, 2016.

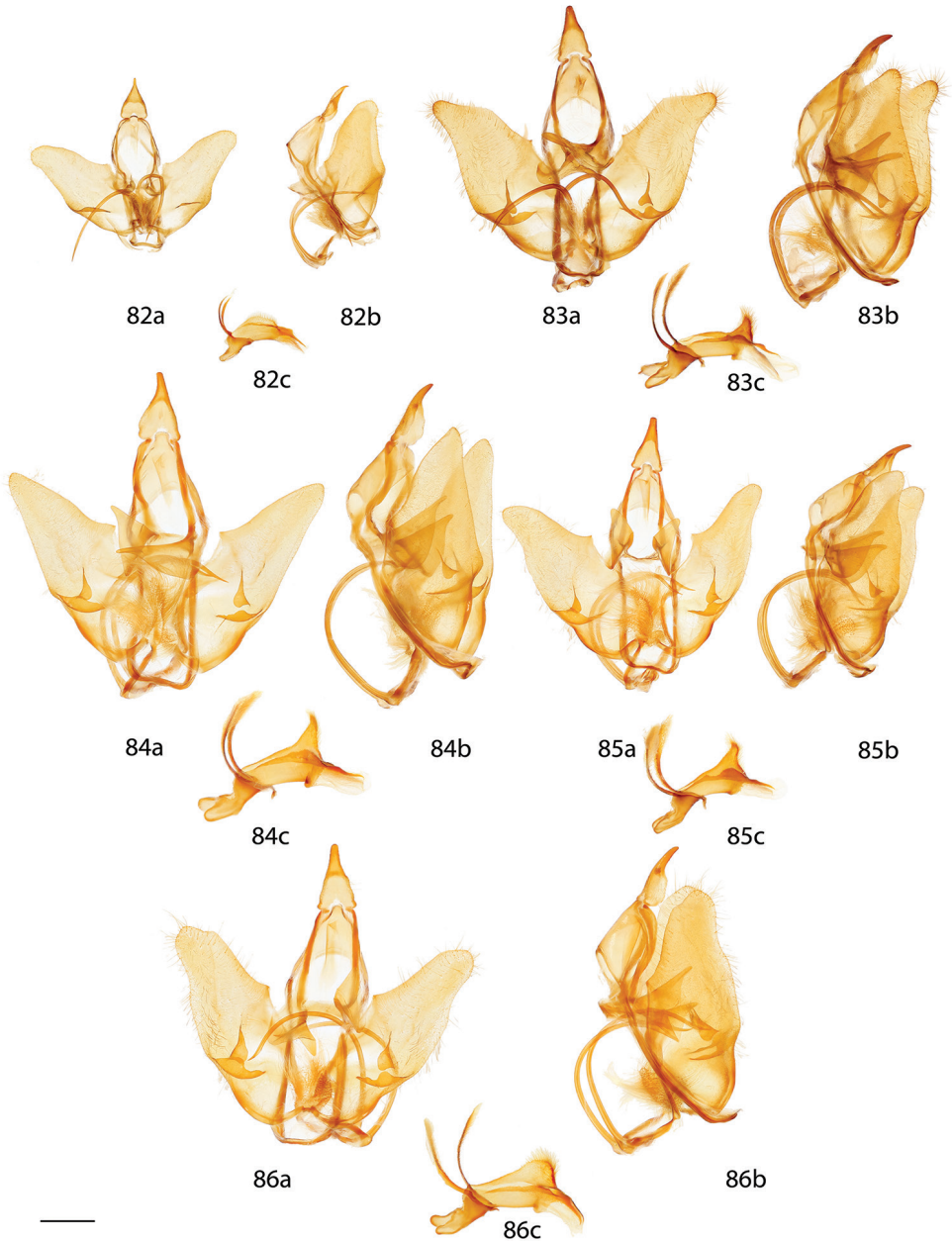
Diagnosis. *Menevia vulgaris* is recognizable from all previous species by the replacement of the wing margin swept postmedial lunule with a continuous white band along the entire length of the postmedial line. This species is quite large for the genus, with highly elongated acutely triangular forewings and triangular hind wings. Sexual dimorphism is well developed, with females having broader, less triangular, but still highly elongated forewings. Male genitalia are easily recognized by the somewhat cylindrical shape of the phallus with an irregularly edged dorsum lacking an extensive dorsal ridge, but usually with a weak anteriorly situated bulge. The juxtal processes are wide and flattened, but only weakly sclerotized; the proximal lobes of the phallus are also very broad, not elongated or fingerlike as in most other species. The combination of these genitalia characters also distinguish *M. vulgaris* from the following four new species described below. Size should also be sufficient in separating *M. vulgaris* from all sympatric species, as *M. vulgaris* is always the largest species within its range. *Menevia vulgaris* is most similar in appearance and size to *M. franclemonti* sp. n. described below, but is easily differentiated from this species by the more sharply angled hindwing postmedial line and by the more uneven dorsum of the phallus. Geography should also be sufficient for separating *M. vulgaris* from *M. franclemonti* sp. n., as the latter species is restricted to southeastern Brazil, outside the largely Amazonian distribution of *M. vulgaris*.

Description. Male. *Head:* Gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by darker gray collar of scales reaching labial palpi, labial palpi very small, segments weakly defined ventrally, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. *Thorax:* As for genus. Light gray. *Legs:* As for genus. Tibial spurs very small, short, almost entirely scaled. *Forewing dorsum:* Forewing length: 22–28 mm, avg.: 25.3 mm, n = 24. Elongated, acutely triangular, apical half of outer margin concave, apex falcate. Ground color gray with darker gray, brown, or reddish brown suffusion throughout medial area, especially near interior edge of postmedial line, pinkish red to blood red coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong shape, thin gray mark connecting discal spot to costa. Apex marked by black scales above extended apical dash. Postmedial line weakly curved to follow outline of wing margin, line black, strongly contrasting. Submarginal area light gray with whitish suffusion mesally, postmedial lunule as dis-

tinct white band originating from apical dash, white band follows postmedial line from apex to posterior wing margin. Antemedial line faint, brown, curved outwards. *Forewing venter*: As in forewing dorsum but antemedial area lighter gray, more contrasting, sometimes with blood-red suffusion, antemedial line absent, small black discal spot occasionally present. *Hindwing dorsum*: Triangular, anal angle weakly accentuated, reddish coloration near anal angle, similar coloration and patterning as forewings, antemedial line absent, postmedial line sharply bent toward anterior wing margin, concave mesally. *Hindwing venter*: Following similar pattern as forewing venter, but red coloration near anal angle much darker, discal mark sometimes present. *Abdomen*: As for genus, but somewhat elongated, nearly sphingiform. Coloration a continuation of gray thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 87) $n = 15$. Tegumen elongated, ovoid or rounded rectangular, constricted near base of gnathos. Vinculum broad, rounded ventrally. Valves relatively narrow, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth slightly reduced in size, both valves with smaller central tooth originating from central ridge of valve, tooth immediately above saccular edge teeth, apex of central tooth pointed toward saccular edge. Valves rounded or somewhat pointed apically. Uncus truncated apically, apex rounded. Gnathos as two prominent flattened, lightly sclerotized, flap-like, somewhat triangular, outward facing extensions with highly truncated apices. Apices usually form fingerlike projections of varying length. Juxtal processes roughly phallus length, lightly sclerotized, curving toward apex of phallus. Juxtal processes very thin, sclerotization weakening to become more membranous, covered in fine setae. Base of phallus with paired, backwards facing, short, rounded, diverging lobes. Phallus cylindrical, irregularly edged dorsum lacking an extensive dorsal ridge, covered in setae. Left edge of rolled phallus uneven but without ridge-like process, usually with weak, anteriorly situated setae covered bulge, distal tip of phallus separated into two distinct points of varying length. Vesica elongate, sac-like, originating from progressively weakened sclerotization of nearly vertical edge of phallus. **Female**. *Head*: As in male but collar of scales bordering eyes and palpi much lighter, less contrasting. *Thorax*: As in male. *Legs*: As in male, tibial spurs very small, only distal tip without scales. *Forewing dorsum*: Forewing length: 27.5–39 mm, avg. 34.6 mm, $n = 9$. Maculation as in male, wing broader and more elongate, less triangular, less falcate, pinkish hue may be replaced by deeper red brown, postmedial line usually straighter, antemedial line fainter. *Forewing venter*: As in forewing dorsum but sometimes with blood-red suffusion, antemedial line absent, thin black discal mark occasionally present. *Hindwing dorsum*: As in male but more rounded, less triangular, postmedial line straight, not concave mesally, but still sharply bent toward anterior wing margin. *Hindwing venter*: Following similar pattern as forewing venter except lighter, reddish suffusion near anal angle much darker, contrasting. *Abdomen*: As in male but more robust. Sternite of VIII as pair of elongated sclerotized bands converging into thicker, irregularly shaped sclerotization near anterior margin of VIII, forming a “V” or “U.” *Genitalia*: (Fig. 101) $n = 4$. Tergite of VIII smoothly curved or weakly triangular, with or without membranous gap mesally, which may be accentuated as posteriorly directed arc. Apophyses anteriores



Figures 72–81. *Menevia* male genitalia, **a** ventral, **b** lateral, **c** phallus [except Figures 80, 81]. **72** *M. lantona*, French Guiana, Pied Saut, Oyapok River [St. Laurent diss.: 3-7-15:2] (CMNH) **73** *M. rosea* holotype, Ecuador, Napo, Simón Bolívar, 1200 m [St. Laurent diss.: 3-7-15:9, 73b inverted horizontally] (CMNH) **74** *M. torvameatoria* holotype, Peru, Puno, La Unión, 2000 ft [St. Laurent diss.: 6-29-15:4, 74b inverted horizontally] (NHMUK) **75** *M. magna* holotype, Brazil, Santa Catarina, São Bento do Sul, Rio Natal, 450 m [St. Laurent diss.: 6-16-15:1] (DZUP) **76** *M. lucara*, French Guiana, Mana River [St. Laurent diss.: 2-5-15:4] (CMNH) **77** *M. menapia* holotype, Guatemala, Cayuga [St. Laurent diss.: 9-7-14:1] (USNM) **78** *M. mielkei* paratype, Brazil, Rio de Janeiro, Cachoeiras de Macacu, 700 m [St. Laurent diss.: 2-5-15:11] (USNM) **79** *M. ostia*, Costa Rica, Guanacaste, Tajo Angeles, 540 m, 10-SRNP-4309 [St. Laurent diss.: 7-14-15:2] (USNM) **80** *M. ostia* phallus, French Guiana, Nouveau Chantier [St. Laurent 3-24-15:7] (USNM) **81** *M. ostia* phallus, Brazil, Espírito Santo, Linhares [St. Laurent 3-24-15:11] (USNM). Scale bar = 1 mm.



Figures 82–86. *Menevia* male genitalia, **a** ventral, **b** lateral, **c** phallus. **82** *M. pallida* paratype, Brazil, Maranhão, Feira Nova do Maranhão, 480 m [St. Laurent diss.: 6-16-15:3] (CGCM) **83** *M. plagiata* neotype, Brazil, Rio de Janeiro, Teresópolis [St. Laurent diss.: 9-2-15:1] (NHMUK) **84** *M. plagiata*, Brazil, Rio de Janeiro [St. Laurent diss.: 4-25-15:2] (CMNH) **85** *M. alurca* paratype, Brazil, Maranhão, Feira Nova do Maranhão, 480 m [St. Laurent diss.: 6-16-15:4] (CGCM) **86** *M. australis* holotype, Brazil, Santa Catarina, Jaraguá do Sul [St. Laurent diss.: 6-19-15:4] (CUIC). Scale bar = 1 mm.



Map 5. *Menevia plagiata* species-group [*vulgaris* subgroup].

shorter or about same length as apophyses posteriores. Lamella antevaginalis quadrate, notched mesally near ostium bursae, anterior edge somewhat irregular. Ductus bursae moderately long. Papillae anales elongated, covered in relatively long setae.

Distribution (Map 5). *Menevia vulgaris* is found throughout northern South America, in the Guyanas, Suriname, Colombia, Ecuador, Peru and the Brazilian states of Amazonas, Pará, and Goiás. This species is predominantly Amazonian in distribution, but the record from the Cerrado in Goiás suggests it is more widespread in various habitats.

Etymology. *Menevia vulgaris* (=vulgaris Latin, meaning commonplace) is named for its wide distribution and apparent commonness.

Remarks. This species is usually the most frequently represented *Menevia* in collections and apparently quite common throughout its broad distribution. This new species is usually misidentified as *M. plagiata* and has long been considered conspecific with the southeastern Brazilian species redescribed above. The taxonomic issues surrounding the name *plagiata* have been explained in the remarks of that species, and the designation of the neotype of *M. plagiata* resolves any previous identification problems and allows recognition of these two very distinct taxa.

Walker's (1855) vague original description of *M. plagiata* could arguably have been associated with the external characters of our concept of *M. plagiata*, *M. vulgaris*,

M. australis, or what we describe below as *M. franclemonti* sp. n. However, we have resolved any ambiguity regarding the application of the name by designating the neotype of *M. plagiata* above. The wide-ranging Amazonian species, *M. vulgaris*, which is apparently absent in the biome inhabited by the other three species, therefore remained undescribed until now.

Currently, the name *M. vulgaris* can now only be associated with the large species present in the Amazonian and Cerrado regions. Diagnostic characters given before, particularly size and male genitalia, are adequate for identifying this species. Other names, including *plagiata* and the various new species described below, are associated with either allopatric or much smaller species.

As with other wide-ranging *Menevia* species, such as *M. lantona* and *M. lucara*, there is a degree of geographic variation, although this variation is less obvious than in these two species. As in other species-groups, *M. vulgaris* from Colombia have more robust male genitalia, except that the juxtal processes are thinner, the valve teeth smaller, and the gnathos elongations shorter. Externally, however, the examined males do not differ from other populations except for their overall slightly larger size. Similar shortened gnathos elongations were found in the males from Ecuador. These males and a single examined female from Ecuador were all smaller than males and females from other populations. However, the lack of differences of the phallus in Ecuadorian males compared to those from other locations suggests that the smaller size and some noted differences found in the single female dissection, namely the shape of the highly variable ventral sclerotized bands on VIII, are not grounds to consider the Ecuadorian population a distinct taxon. Another variation worth noting is in the single Peruvian female specimen, which has the reddish maculation on all wings replaced by deeper brownish-red.

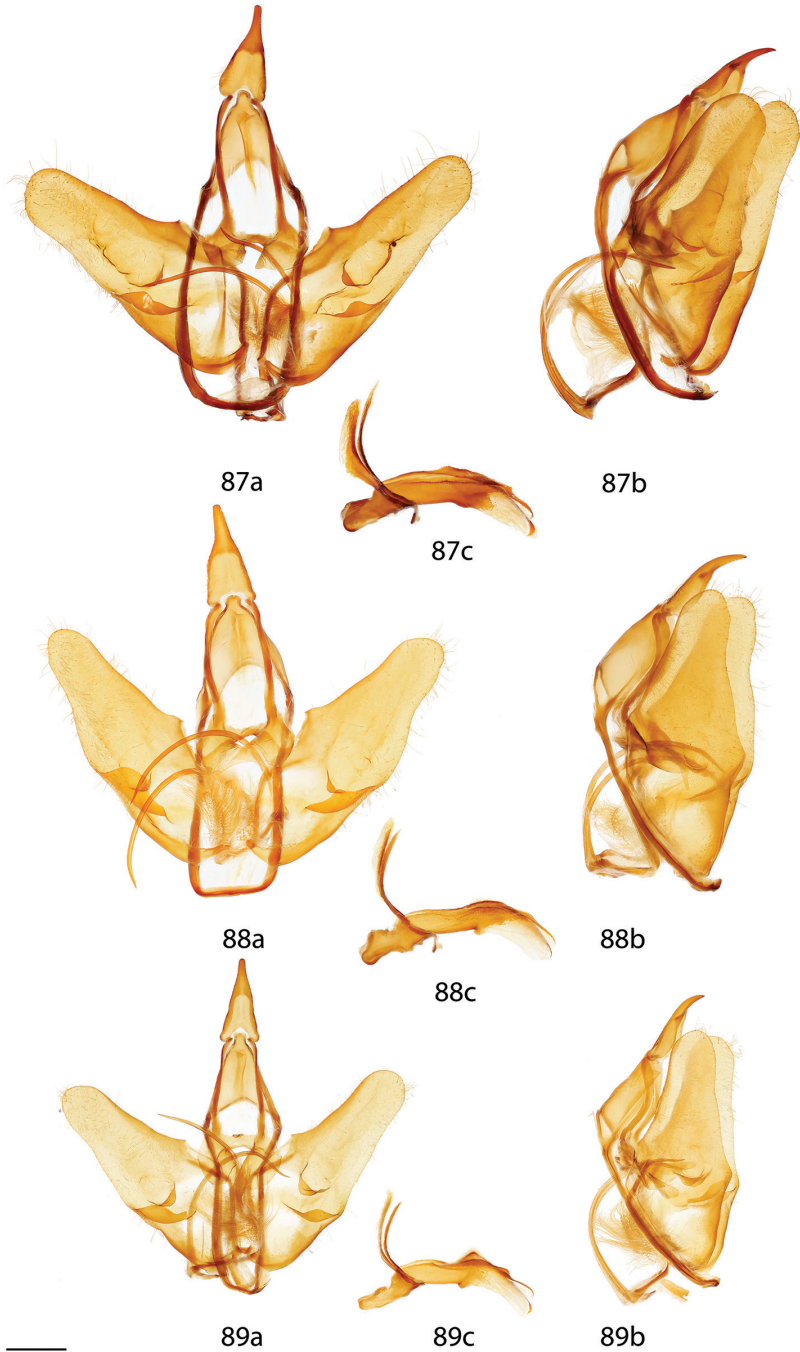
***Menevia franclemonti* sp. n.**

<http://zoobank.org/2CB186E2-CBFA-46B9-94A4-4FA8A3056F9E>

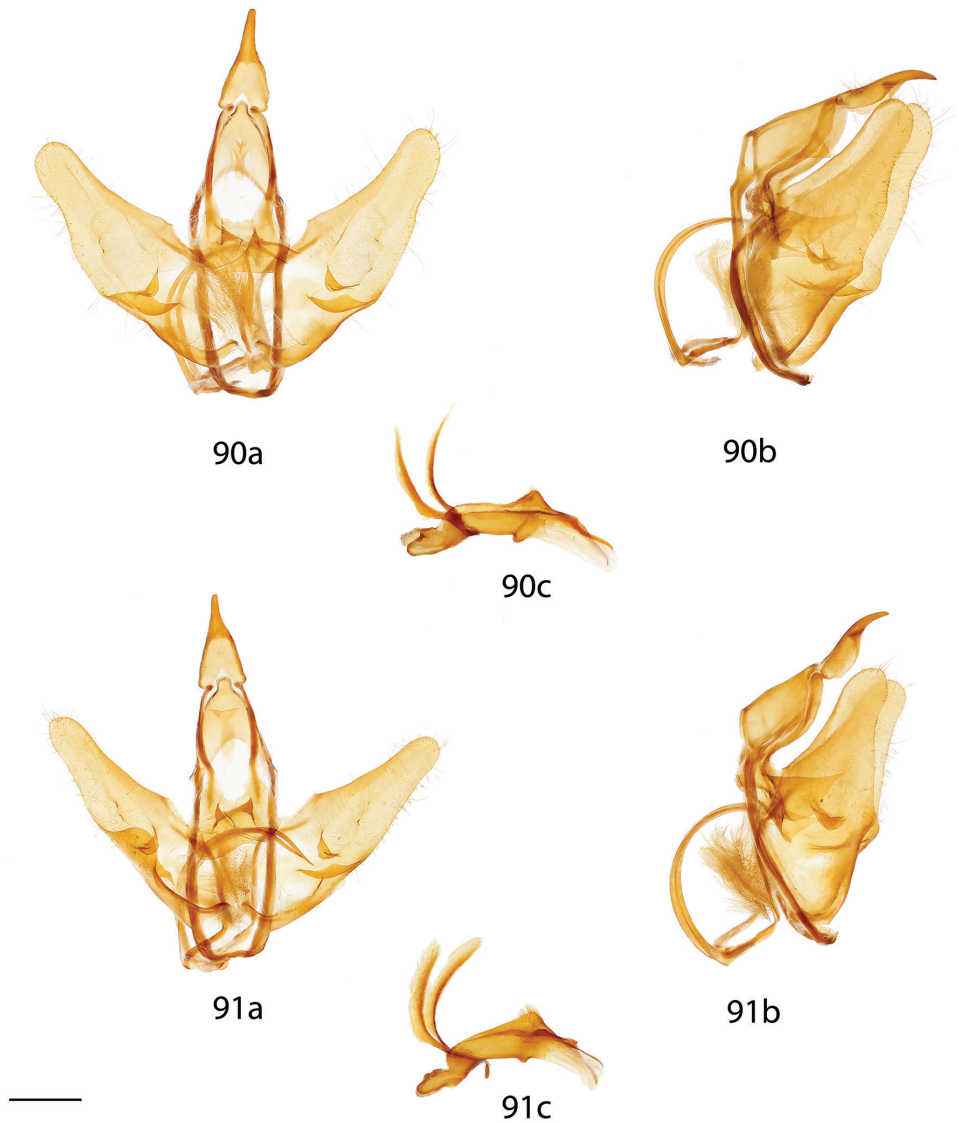
Figs 60, 61, 68, 88; Map 5

Type material. Holotype, ♂: **BRAZIL: Santa Catarina:** Jaragua [Jaraguá do Sul], Santa Catharina, Brazil, 10 Nov 1934, Fritz Hoffmann/ PARATYPE *Menevia falco* J. G. Franclemont/ St. Laurent diss.: 4-25-15:5/ Franclemont's label refers to a MS name./ HOLOTYPE male *Menevia franclemonti* St Laurent and Dombroskie, 2016 [handwritten red label]/ (CUIC). Type locality: Brazil: Santa Catarina: Jaraguá do Sul.

Paratypes, 10 ♂, 1 ♀: **BRAZIL: Rio de Janeiro:** 1 ♂, Petrópolis, Independência, 900 m: 15.XI.1939, Gagarin leg., ex. col. Gagarin, DZ 32.717 (DZUP). 1 ♂, Petrópolis: 17.X.1960, Gagarin leg., ex. coll Gagarin, DZ 32.718 (DZUP). 2 ♂, Teresópolis, Barreira, 350 m: 30.X.1956–3.XI.1956, 1.XII.1956, Pearson H&G, HRP No. 1070, USNM-Mimal: 2272, 2273, St. Laurent diss.: 4-25-15:6 (USNM). 2 ♂, 1 ♀, Teresópolis, Barreira: 12.XI.1955, 17.XI.1955, 19.XI.1955, ex. col. Gagarin, DZ 32.719–32.721 (DZUP). **Santa Catarina:** 2 ♂, Jaraguá do Sul: 17.XI.1934,



Figures 87–89. *Menevia* male genitalia, **a** ventral, **b** lateral, **c** phallus. **87** *M. vulgaris* holotype, French Guiana, St. Laurent du Maroni, Piste de Paul Isnard, 52 m [St. Laurent diss.: 9-2-15:2] (CUIC) **88** *M. franclemonti* holotype, Brazil, Santa Catarina, Jaraguá do Sul [St. Laurent diss.: 4-25-15:5] (CUIC) **89** *M. vulgaricula* holotype, Brazil, Huitanaã, Rio Purus [St. Laurent diss.: 6-19-15:1] (CMNH). Scale bar = 1 mm.



Figures 90–91. *Menevia* male genitalia, **a** ventral, **b** lateral, **c** phallus. **90** *M. cordillera* holotype, Peru, San Gabán, 2500 ft [St. Laurent diss.: 6-29-15:8] (NHMUK) **91** *M. delphinus* holotype, Brazil, Distrito Federal, Planaltina, 1000 m [St. Laurent diss.: 9-3-15:1] (CPAC). Scale bar = 1 mm.

20.XI.1934, Fritz Hoffmann, Franclemont diss.: 1760 (CUIC). **São Paulo:** 1 ♂, Juqueí: 20.III.1976, ex. coleção A. Cardoso, DZ 32.716 (DZUP). 1 ♂, Ubatuba, 10 m: 13-20.XII.1963, Prof. Heinz Ebert, St. Laurent diss.: 4-25-15:7 (CMNH). – All paratypes with the following yellow label: PARATYPE male/female *Menevia franclemonti* St Laurent and Dombroskie, 2016.

Diagnosis. Very similar to *M. vulgaris*, but distinguished by the usually warmer brown hue rather than gray, more sharply falcate forewings, and a more smoothly curving postmedial line on the hindwing, which does not sharply bend toward the anterior wing margin. The male genitalia are also unique in that the phallus is thin and smooth, not irregularly shaped dorsally, with the sclerotized portion of the distal end of the phallus diagonal rather than vertical as in *M. vulgaris*. Furthermore, the base of the sclerotized terminus of the phallus, on the venter, has a prominent bump, angled away from the distal end of the phallus. *Menevia franclemonti* replaces *M. vulgaris* in the Brazilian Atlantic Forest.

Description. Male. Head: Gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by light brownish gray collar of scales reaching labial palpi, labial palpi very small, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. **Thorax:** As for genus. Light gray to brown. **Legs:** As for genus. Tibial spurs small, short, almost entirely scaled, except distal tip. **Forewing dorsum:** Forewing length: 25–27 mm, avg.: 26.1 mm, n = 6. Elongated, acutely triangular, apical half of outer margin highly concave, apex very falcate. Ground color gray or warm brown with darker gray, brown, or reddish brown suffusion, especially near interior edge of postmedial line and medial area; brownish red to blood red coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong shape, thin gray mark connecting discal spot to costa. Apex marked by black scales above extended apical dash. Postmedial line smooth, mostly straight, line black, strongly contrasting. Submarginal area light gray with small grayish-white suffusion mesally, postmedial lunule as white band originating from apical dash, white band follows postmedial line from apex to anal wing margin. Antemedial line faint, brown, curved outwards. **Forewing venter:** As in forewing dorsum but antemedial area lighter gray, more contrasting, blood-red suffusion more pronounced, antemedial line absent, small black discal mark occasionally present. **Hindwing dorsum:** Triangular, anal angle usually sharply accentuated, reddish coloration that bleeds into medial area originates near anal angle, overall similar coloration and patterning as forewings, antemedial line absent, postmedial line weakly curved toward anterior wing margin, not concave mesally. **Hindwing venter:** Following similar pattern as forewing venter, but red coloration near anal angle much darker, discal mark sometimes present. **Abdomen:** As for genus, but somewhat elongated, nearly sphingiform. Coloration a continuation of gray thoracic color. Midventral stripe absent. **Genitalia:** (Fig. 88) n = 3. Tegumen elongated, ovoid, constricted near base of gnathos. Vinculum broad, somewhat quadrate ventrally. Valves triangular, saccular edge of left and right valves with triangular tooth proximal to transtilla, both valves with smaller mesal costal projection originating from central ridge of valve, projection immediately above saccular edge teeth, apex of mesal costal projection pointed toward saccular edge. Valves rounded apically. Uncus truncated apically, apex somewhat quadrate. Gnathos as two prominent flattened, moderately sclerotized, flap-like, somewhat rectangular, outward facing extensions with highly truncated apices. Apices form fingerlike projections of moderate length. Juxtal processes roughly phallus length, lightly sclerotized, curving toward apex of phallus. Juxtal processes very thin,

sclerotization fading to more membranous portion, covered in fine setae. Base of phallus with paired, backwards facing, very short, rounded, diverging lobes. Phallus cylindrical, smooth dorsum lacking an extensive dorsal ridge, covered in setae. Left edge of rolled phallus smooth, without ridge-like process, base of sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of phallus, distal tip of phallus separated into two distinct points. Vesica somewhat elongated, covered in setae laterally, originating from progressively weakened sclerotization of diagonal edge of phallus. **Female.** *Head:* As in male. *Thorax:* As in male. *Legs:* As in male. *Forewing dorsum:* Forewing length: about 30 mm, $n = 1$. Maculation as in male, wing broader, less triangular, less falcate, antemedial line fainter. *Forewing venter:* As in forewing dorsum but with blood-red suffusion, antemedial line absent, thin black discal mark present. *Hindwing dorsum:* As in male but more rounded, less triangular, postmedial line straight, not concave mesally, but still only weakly curved toward anterior wing margin. *Hindwing venter:* Following similar pattern as forewing venter except lighter, reddish suffusion near anal angle much darker, contrasting. *Abdomen:* As in male but more robust. *Genitalia:* Not examined.

Distribution (Map 5). *Menevia franclemonti* is found only in the Brazilian Atlantic Forest, in the states of Rio de Janeiro, São Paulo, and Santa Catarina. This species is likely endemic to this region, where it replaces the similar, more widespread *M. vulgaris*.

Etymology. *Menevia franclemonti* is named after the lepidopterist J. G. Franclemont, who originally recognized the uniqueness of this species. He also wrote an important fascicle on the Mimallonidae of North America north of Mexico (Franclemont 1973).

Remarks. This new species is the Brazilian Atlantic Forest component of the *vulgaris* subgroup of the *plagiata* species-group in much the same way that *M. mielkei* and *M. magna* are the Atlantic Forest components of the *lucara* and *lantona* species-groups respectively. Although *M. franclemonti* is not remarkably distinct from *M. vulgaris*, it is readily differentiated by the external and genitalia diagnostic characters presented above. Additionally, this species seems to be allopatric to all other species in this subgroup. Despite the differences, *M. franclemonti*, together with the similar *M. vulgaris*, have both been misidentified as *M. plagiata* for the reasons explained in the remarks of *M. plagiata* and *M. vulgaris*. The allopatry of *M. vulgaris* and *M. franclemonti* certainly affords that *M. plagiata sensu stricto* could not be applied to the Amazonian *M. vulgaris*, but it is certainly plausible that Walker's (1855) description of *M. plagiata* could have applied to *M. franclemonti* due to its presence at the type locality of *M. plagiata*. However, due to the apparent rarity of *M. franclemonti* in collections, compared to the sympatric *M. plagiata* and the complete lack of *M. franclemonti* from the NHMUK, it is more plausible that the original material from Rio de Janeiro that Walker had at his disposal was in fact the species that we consider *M. plagiata* in this present work, and not *M. franclemonti*. We have resolved the ambiguity surrounding the application of the name *plagiata* by designating the neotype of this taxon above. If the holotype of *M. plagiata* is discovered in the future, and is found to be what we consider *M. franclemonti*, then it would be taxonomically simple to set aside the neotype under Article 75.8 of the ICZN (1999), synonymize this new species with *M. plagiata*, and redescribe what we currently consider to be *M. plagiata* as a new species.



Figures 92–97. *Menevia* female genitalia, **a** ventral, **b** VIII tergite detail. **92** *M. lantona*, French Guiana, St. Jean du Maroni, [St. Laurent diss.: 7-8-15:1] (USNM) **93** *M. magna* paratype, Brazil, Santa Catarina, Dalbérgia [St. Laurent diss.: 3-7-15:15] (CUIC) **94** *M. lucara*, French Guiana, Kourou Forest [St. Laurent diss.: 5-22-15:1] (MNHN) **95** *M. ostia*, Costa Rica, Guanacaste, Mena Central, 345 m, Voucher: 01-SRNP-24319 [St. Laurent diss.: 4-20-15:9] (USNM) **96** *M. parostia* holotype, unknown locality [St. Laurent diss.: 4-20-15:7] (USNM) **98** *M. pallida* [questionable], Brazil, Minas Gerais, Las-sance [St. Laurent diss.: 4-20-15:6] (CUIC). Scale bar = 1 mm.

As in the case of *M. australis*, J. G. Franclemont recognized this species as distinct and labeled specimens as holotype and paratypes (all from Jaraguá do Sul, Santa Catarina, Brazil) under his manuscript name *Menevia* ‡*falco*. Rather than naming this species *M. ‡falco* as he originally intended, we decided to honor J. G. Franclemont by naming this species after him for his work on *Menevia* and Mimallonidae in general.

***Menevia vulgaricula* sp. n.**

<http://zoobank.org/02587433-09B5-4632-AE4A-D40EB1D8651B>

Figs 62, 69, 70, 89, 102; Map 5

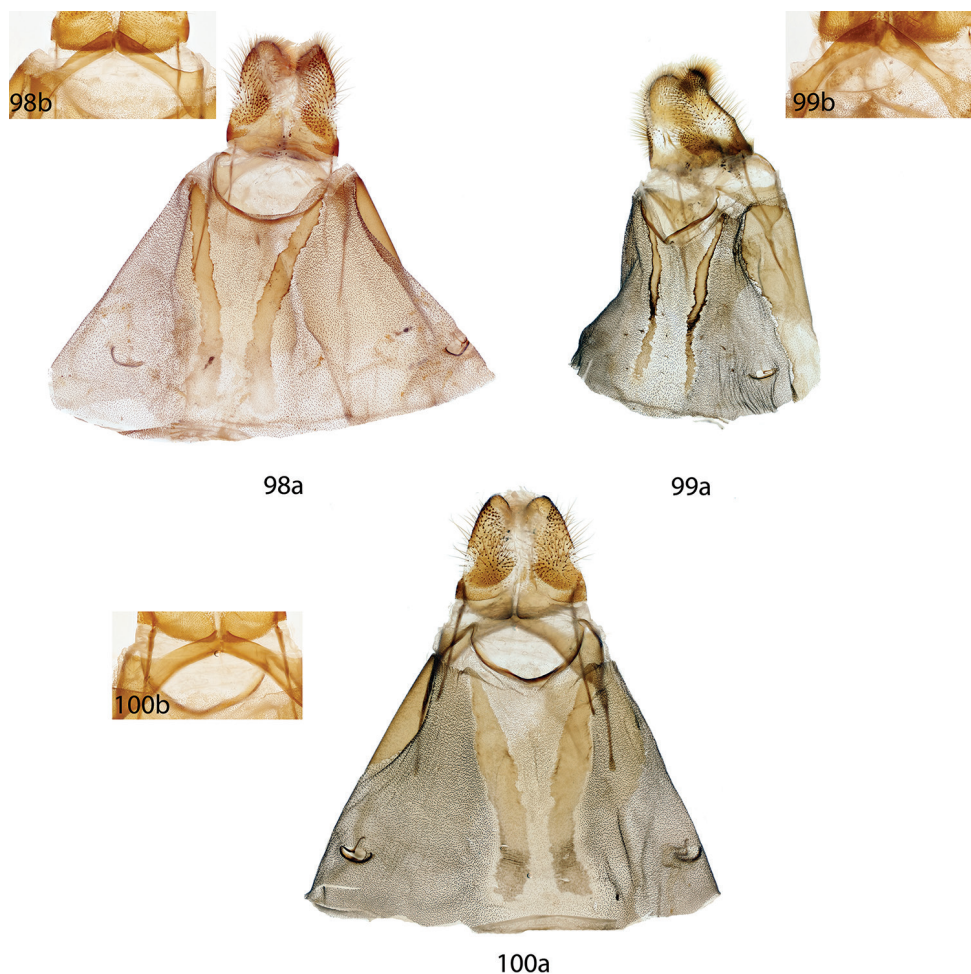
Type material. Holotype, ♂: **BRAZIL: Amazonas:** Hyutanahan [Huitanaã], Rio Purus, Brazil, S.M. Klages/ Jan. 1922/ Carn. Mus. Acc. 6963/ St. Laurent diss.: 6-19-15:1/ HOLOTYPE male *Menevia vulgaricula* St Laurent and Dombroskie, 2016 [handwritten red label]/ (CMNH). Type locality: Brazil: Amazonas: Huitanaã.

Paratypes, 2 ♂, 1 ♀: **BRAZIL: Amazonas:** 1 ♂, Hyutanahan [Huitanaã], Rio Purus: II.1922, S.M. Klages, Carn. Mus. Acc. 6963, St. Laurent diss.: 4-25-15:14 (CMNH). 1 ♂, Nova Olinda, Rio Purus: V.1922, S.M. Klages, St. Laurent diss.: Carn. Mus. Acc. 6962, St. Laurent diss.: 6-19-15:2 (CMNH). 1 ♀, Rio Madeira: Juillet-Août (Fassl), Dognin Collection, USNM-Mimal: 2595, St. Laurent diss.: 4-25-15:15 (USNM). – All paratypes with the following yellow label: PARATYPE male/female *Menevia vulgaricula* St Laurent and Dombroskie, 2016.

Additional questionable specimens examined. (3 ♀ total) [not included in type series] **BRAZIL: Pará:** 1 ♀, Likely Belém: A.M. Moss, Rothschild Bequest 1939–1, St. Laurent 7-21-15:2, BMNH(E) 1378755 (NHMUK). **FRENCH GUIANA:** 1 ♀, RN 2, Km 48: 14.VIII.1985, J. Haxaire, Bc-Her 2991 (Collection of Daniel Herbin, France). 1 ♀, Saül: 1.VIII.2011, Ph. Collet leg. (RAS).

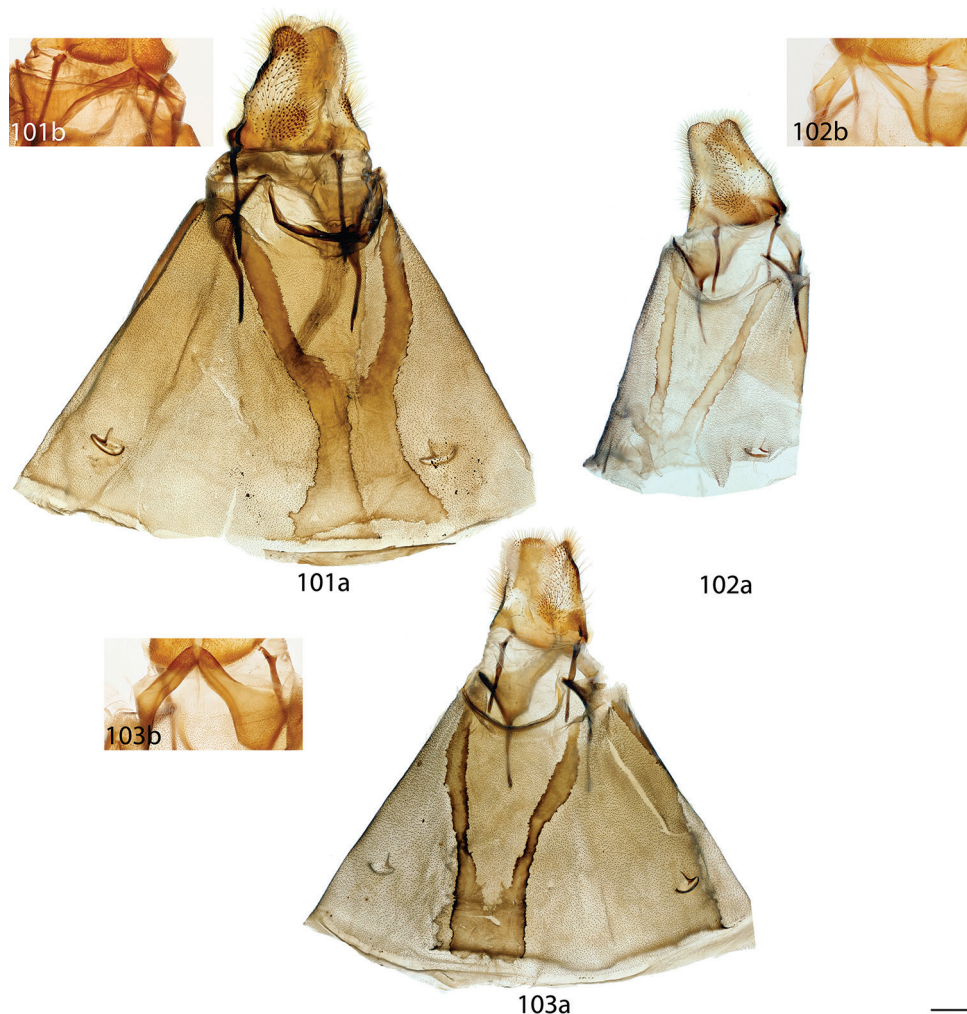
Diagnosis. *Menevia vulgaricula* is similar to the previous two species, but much smaller, both males and females are notably smaller than the respective sexes of *M. vulgaris*. The female of *M. vulgaricula* is easily differentiated from females of *M. vulgaris* by the width of the submarginal area, which is broader and decreases in width toward the apex much less gradually than the rapidly narrowing submarginal area of *M. vulgaris*. The genitalia of both sexes can be used to differentiate *M. vulgaricula* from similar species (except from the unexamined female of *M. franclemonti*). In males, the phallus has a very prominent setae covered dorso-anterior bulge reminiscent of a dorsal phallic ridge, the tegumen, vinculum, and acutely triangular uncus are all very narrow and elongated, and the paired processes of the gnathos converge and bend upwards. Additionally, the divergent lobes at the proximal end of the phallus are not broad, but thin and peg-like. In the female, the lamella antevaginalis is very thin, unlike the usually broad, quadrate lamella of *M. vulgaris*.

Description. Male. Head: Gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by brownish gray collar of scales reaching labial palpi, labial palpi very small, segments weakly defined ventrally, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel tufted. **Thorax:** As for genus. Light gray. **Legs:** As for genus. Tibial spurs moderate length, thin, somewhat hooked distally. **Forewing dorsum:** Forewing length: 19–22 mm, avg.: 21 mm, n = 3. Acutely triangular, apical half of outer margin concave, apex falcate. Ground color gray with brown or reddish brown suffusion, especially near interior edge of postmedial line and medial area, reddish coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong



Figures 98–100. *Menevia* female genitalia, **a** ventral, **b** VIII tergite detail. **98** *M. plagiata*, Brazil, Rio de Janeiro [St. Laurent diss.: 7-28-15:1] (AMNH) **99** *M. alurca*, Brazil, Maranhão, Feira Nova do Maranhão [St. Laurent diss.: 7-28-15:2] (CGCM) **100** *M. australis* paratype, Brazil, Santa Catarina, Jaraguá do Sul [St. Laurent diss.: 4-25-15:3] (CUIC). Scale bar = 1 mm.

shape, thin gray mark connecting discal spot to costa. Apex marked by black scales above extended apical dash. Postmedial line mostly straight except when approaching apex, line black, strongly contrasting. Submarginal area light gray with whitish suffusion mesally, sometimes appearing as a faint zigzag, postmedial lunule as white band originating from apical dash, white band follows postmedial line from apex to anal wing margin. Antemedial line faint, brown, curved outwards. *Forewing venter*: As in forewing dorsum but antemedial area lighter gray, more contrasting, blood red suffusion more expansive, antemedial line absent, small black discal spot occasionally present. *Hindwing dorsum*: Triangular, anal angle weakly accentuated, reddish coloration near anal angle, similar



Figures 101–103. *Menevia* female genitalia, **a** ventral, **b** VIII tergite detail. **101** *M. vulgaris* paratype, French Guiana, St. Laurent du Maroni [St. Laurent diss.: 4-25-15:13] (USNM) **102** *M. vulgaricula* paratype, Brazil, Amazonas, Rio Madeira [St. Laurent diss.: 4-25-15:15] (USNM) **103** *M. delphinus* paratype, Brazil, Distrito Federal, Planaltina [St. Laurent diss.: 4-2-15:4] (CPAC). Scale bar = 1 mm.

coloration and patterning as forewings, antemedial line absent, postmedial line moderately bent toward anterior wing margin, weakly concave mesally. *Hindwing venter*: Following similar pattern as forewing venter, but red coloration near anal angle much darker, discal mark absent or very faint. *Abdomen*: As for genus but somewhat elongated, nearly sphingiform. Coloration a continuation of gray thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 89) $n = 3$. Tegumen elongated, narrow, somewhat constricted near base of gnathos. Vinculum elongated, narrow. Valves relatively narrow, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth

slightly reduced in size, both valves with smaller mesal costal projection originating from central ridge of valve, mesal costal projection immediately above saccular edge teeth, apex of projection pointed toward saccular edge. Valves rounded apically. Uncus very narrow, acutely triangular. Gnathos as two prominent, converging, flattened, sclerotized, flap-like, somewhat triangular, upward facing extensions with somewhat truncated apices. Juxtal processes shorter than phallus, curving toward apex of phallus. Juxtal processes thin, covered in fine setae. Base of phallus with paired, backwards facing, short, peg-like, diverging lobes. Phallus cylindrical, irregularly edged dorsum with accentuated bulging ridge-like projection situated anteriorly, covered in setae. Left edge of rolled phallus uneven but forming anteriorly situated bulge, base of the sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of phallus, distal tip of phallus separated into two distinct points. Vesica elongate, covered in setae laterally, originating from progressively weakened sclerotization. **Female.** *Head:* As in male but with light brown tint. *Thorax:* As in male but with brownish tint. *Legs:* As in male, tibial spurs shorter or about same length. *Forewing dorsum:* Forewing length: 27 mm, $n = 1$. Maculation as in male, wing broader, barely more elongate, less triangular, less falcate, postmedial line straighter, nearly parallel to wing margin until just before apex, submarginal area rectangular not triangular, antemedial line fainter. *Forewing venter:* As in forewing dorsum but lighter gray, with pinkish suffusion, antemedial line absent, thin black discal mark present. *Hindwing dorsum:* As in male but more rounded, less triangular, postmedial line concave mesally, but still moderately bent toward anterior wing margin. *Hindwing venter:* Following similar pattern as forewing venter, reddish suffusion near anal angle much darker, contrasting. *Abdomen:* As in male but more robust. Sternite of VIII with pair of thin sclerotized bands converging near anterior margin of VIII forming a “V”. *Genitalia:* (Fig. 102) $n = 1$. Tergite of VIII forming posteriorly directed triangle, without membranous gap mesally. Apophyses anteriores about same length as apophyses posteriores. Lamella antevaginalis very thin, indistinct, curved. Ductus bursae short. Papillae anales elongated, covered in relatively long setae.

Distribution (Map 5). This new species is restricted to the Amazon region, specifically in the vicinity of Rio Madeira and Rio Purus, but may be more widespread, see remarks. This species is sympatric with *M. vulgaris*.

Etymology. *Menevia vulgaricula* is named for its appearance as a diminutive *M. vulgaris*.

Remarks. *Menevia vulgaricula* is an interesting species due to its remarkable external resemblance to *M. vulgaris*, with which *M. vulgaricula* is sympatric. Despite notable similarity in external characters, *M. vulgaricula* is much smaller and displays distinct genitalia characters, in both the male and the female.

Both *M. vulgaris* and *M. vulgaricula* were collected by S. M. Klages during the same period at both Huitanaã and Nova Olinda, Amazonas, Brazil (CMNH). Therefore, not only are these two species sympatric, but are also apparently both active at the same time of year. Size and distinct differences in genitalia likely afford some form of a prezygotic barrier. The last two new species described below are very similar to *M. vulgaricula*, in that each species is very small relative to the larger *M. vulgaris* and *M. franclemonti*, and both display prominent anterior phallic bulges. These species

however are widely allopatric. Therefore, the modes of isolation between sympatric *M. vulgaris* and *M. vulgarcicula* certainly warrants future investigation.

Additional females from Pará, Brazil and French Guiana were examined, but cannot be included in the type series because we lack the more easily identifiable males from these localities. Also, these specimens display deeper red coloration and wavier forewing postmedial lines, characters not seen in all other examined *M. vulgarcicula*. Although the small size of these females and the genitalia of the Pará specimen suggest that they are probably *M. vulgarcicula*, Pará and French Guiana are distant from both Rio Madeira and Rio Purus, the two relatively nearby localities known to support *M. vulgarcicula*.

***Menevia cordillera* sp. n.**

<http://zoobank.org/DE2B8D54-5671-4603-A9ED-7CC342CB46D5>

Figs 63, 64, 90; Map 5

Type material. **Holotype**, ♂: **PERU**: 1584, San Gaban [San Gabán], Peru, 2500 ft, March–April 1913/ Joicey Coll. Brit. Mus. 1925–157/ St. Laurent diss.: 6-29-15:8/ BMNH(E) 1378757/ HOLOTYPE male *Menevia cordillera* St Laurent and Dombroskie, 2016 [handwritten red label]/ (NHMUK). Type locality: Peru: Puno: Carabaya: San Gabán.

Paratype, 1 ♂: **BOLIVIA**: Rio Songo [Río Zongo], 750 m: Coll. Fassl, Dognin Collection, USNM-Mimal: 2599, St. Laurent diss.: 4-25-15:17 (USNM). – Paratype with the following yellow label: PARATYPE male *Menevia cordillera* St Laurent and Dombroskie, 2016.

Diagnosis. This new species, like *M. vulgarcicula*, is quite small in comparison with the widespread *M. vulgaris* and the southeast Brazilian *M. franclemonti*. Due to the small size of *M. cordillera*, it may be confused with the allopatric *M. vulgarcicula* but can easily be differentiated by the deeper reddish brown coloration, more sharply acute apices of the more elongated forewings, straighter hindwing margins, and by the male genitalia. The phallus of *M. cordillera* is somewhat reminiscent of that of *M. vulgarcicula*, but with a more triangularly shaped anterior dorsal bulge and rounded, not peg-like, lobes at the base of the phallus. Overall, the phallus of *M. cordillera* is broader than that of *M. vulgarcicula*. No other *Menevia* species are known from the Cordillera Oriental, besides the clearly distinct *M. torvamesoria*.

Description. Male. **Head:** Light brown-gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by dark brown collar of scales reaching labial palpi, labial palpi large, robust for genus, dorsally with darker scales contrasting with overall gray coloration. Scape and pedicel weakly tufted. **Thorax:** As for genus. Light tan. **Legs:** As for genus. Tibial spurs short, stout. **Forewing dorsum:** Forewing length: 22–23 mm, avg.: 22.5 mm, n = 2. Very acutely triangular, apical half of outer margin deeply concave, apex very falcate. Ground color gray with deep reddish brown suffusion throughout medial area, brighter reddish coloration near apex along apical interior of postmedial line, overall lightly speckled by dark petiolate scales. Discal spot faintly marked by light gray oblong

shape, gray mark connecting discal spot to costa. Apex marked by black scales above extended apical dash. Black postmedial line mostly straight except when approaching apex where sharply curved, strongly contrasting. Submarginal area light gray with whitish suffusion mesally, sometimes appearing as faint zigzag, submarginal area with distinct white band originating from apical dash, white band follows postmedial line from apex to anal wing margin. Antemedial line very faint, brown, curved outwards. *Forewing venter*: As in forewing dorsum but antemedial area lighter gray, more contrasting, antemedial line absent, small black discal mark present. *Hindwing dorsum*: Triangular, outer margin very straight, anal angle weakly accentuated, reddish coloration near anal angle, bleeding into medial area, similar coloration and patterning as forewings, antemedial line absent, postmedial line sharply bent toward anterior wing margin, weakly concave mesally. *Hindwing venter*: Following similar pattern as forewing venter, but red coloration near anal angle much darker, discal mark absent. *Abdomen*: As for genus, but elongated, nearly sphingiform. Midventral stripe absent. *Genitalia*: (Fig. 90) $n = 2$. Tegumen elongated, moderately narrow, weakly constricted near base of gnathos. Vinculum elongated, narrow, ovoid, somewhat rounded ventrally. Valves relatively narrow, rounded, saccular edge of left valve with large triangular tooth proximal to transtilla, right valve with tooth slightly reduced in size, both valves with smaller mesal costal tooth originating from central ridge of valve, mesal costal projection immediately above saccular edge teeth, apex of projection pointed toward saccular edge. Valves truncated apically. Uncus very narrow, acutely triangular, quadrate or rounded apically. Gnathos as two prominent, converging, flattened, sclerotized, flap-like, somewhat triangular, upward facing extensions with truncated apices. Juxtal processes shorter than phallus, curving toward apex of phallus. Juxtal processes thin, covered in fine setae. Base of phallus with paired, backwards facing, short, rounded, diverging lobes. Phallus cylindrical, dorsum with accentuated triangular bulging projection situated anteriorly, covered in setae. Left edge of rolled phallus uneven but forming anteriorly situated, setae covered, triangular bulge, base of sclerotized terminus of phallus with prominent ventral bump, angled away from distal end of the phallus, distal tip of phallus separated into two, elongated, distinct points. Vesica elongated, bag-like, covered in setae laterally, originating from progressively weakened sclerotization. **Female**. Unknown.

Distribution (Map 5). *Menevia cordillera* is apparently restricted to the Cordillera Oriental of Peru and Bolivia at moderate elevations, from 750–760 m in elevation.

Etymology. This new species is named for the Andean Cordillera Oriental, to which this species is endemic.

Remarks. *Menevia cordillera* is very closely related to both *M. vulgaris* and *M. vulgarecula* based on general external characters and the genitalia morphology. These three species may represent taxa of a species complex that spans throughout northern South America.

An additional specimen from the Yungas of Bolivia, in the collection of Daniel Herbin (Bc-Her2532) as seen in the BOLD database, almost certainly belongs to this new species. However, we were unable to examine this specimen and thus it cannot be included in the type series, but we report it here as it provides additional distributional data.

***Menevia delphinus* sp. n.**

<http://zoobank.org/12D5AF97-8DE4-43EA-902A-26E96BB24065>

Figs 65, 66, 71, 91, 103; Map 5.

Type material. **Holotype**, ♂: **BRAZIL: Distrito Federal:** Coleção EMBRAPA-CPAC No. 9401/ Planaltina, DF, 1000 m, 15°35'S, 47°42'W, 12.XI.1982/ V.O. Becker Col./ St. Laurent diss.: 9-3-15:1/ HOLOTYPE male *Menevia delphinus* St Laurent and Dombroskie, 2016 [handwritten red label]/ (CPAC). Type locality: Brazil: Distrito Federal: Planaltina.

Paratypes, 6 ♂, 2 ♀: **BRAZIL: Distrito Federal:** 3 ♂, 2 ♀, Planaltina, 15°35'S, 47°42'W, 1000 m: 11.XI.1977, 15.XI.1977, V.O. Becker col., Col. Becker No. 22269, 22304, St. Laurent diss.: 4-25-15:16, USNM-Mimal: 2341, 2343 (USNM); 10.XI.1975, 3.XI.1982, 4.XI.1982, V.O. Becker col., Coleção EMBRAPA-CPAC No. 79, 9401, St. Laurent diss.: 4-2-15:4 (CPAC). 1 ♂, Planaltina, 15°36'S, 47°44'W, 960 m: 30.XI.1992, No. 2135, Coleção EMBRAPA-CPAC No. 12816, St. Laurent diss.: 9-2-15:3 (CPAC). **Goiás:** 1 ♂, Leop. Bulhoes [Leopoldo de Bulhões]: XI.1935, Coll. R. Spitz, Rothschild Bequest BM 1939–1, St. Laurent diss.: 6-29-15:10, BMNH(E) 1378753 (NHMUK). **Minas Gerais:** 1 ♂, Sabara-Belo Horizonte, Rio das Velhas [Sabará-Belo Horizonte, Rio das Velhas]: A.G.N. Chalmers, B.M. 1932–11, St. Laurent diss.: 6-29-15:11, BMNH(E) 1378756 (NHMUK). – All paratypes with the following yellow label: PARATYPE male/female *Menevia delphinus* St Laurent and Dombroskie, 2016.

Additional specimen examined from photo. [not included in type series] **BRAZIL: Maranhão:** 1 ♂, Feira Nova do Maranhão, Retiro, 46°26'41"W, -07°00'31"S, 480 m: C. Mielke leg., Paratype, *Menevia alurca* Herbin & Mielke, 2014 (Collection of Daniel Herbin, France).

Diagnosis. *Menevia delphinus*, like the previous two species, is quite small in comparison with the widespread *M. vulgaris* and the southeast Brazilian *M. franclemonti*, but males and females can easily be differentiated from all others belonging to the *plagiata* species-group by the relatively stout forewings and by the genitalia. The phallus of *M. delphinus* is most similar to that of *M. cordillera*, but with the dorsal bulge being situated more mesally along the length of the phallus and usually much more pronounced as a singular, blunt, protuberance, not a pointed or triangular bulge as in other species. Additionally, the lobes at the base of the phallus in *M. delphinus* are very elongated and tubular, almost fingerlike, not rounded or peg-like as in the previous similar species. In females, the VIII tergite is very robust, forming a distinct triangle, and is not rounded and arc-like as in some similar species. *Menevia delphinus* may also be confused with *M. alurca* due to the similar size and sympatry; however, *M. delphinus* can be straightforwardly recognized by the lack of a midventral abdominal stripe and a continuous white band along the outer edge of the postmedial line, which is discontinuous midway along the postmedial line in *M. alurca*. Additionally, the genitalia easily differentiate *M. delphinus* from *M. alurca*. The dorsal protuberance of the phallus of *M. delphinus* is much smaller and blunter in comparison with the extremely

elongate, curved, and sharply pointed dorsal phallic protuberance of *M. alurca*. The only other sympatric species besides *M. alurca* is the much larger *M. vulgaris*.

Description. Male. *Head*: Light brown or gray, eyes large comprising about two-thirds of head area, eyes bordered posteriorly by brownish collar of scales reaching labial palpi, labial palpi very small, short, covered in dark scales. Scape and pedicel weakly tufted. *Thorax*: As for genus. Light gray. *Legs*: As for genus. Tibial spurs moderate length, thin, scaled except for distal tip. *Forewing dorsum*: Forewing length: 17–23 mm, avg.: 20.5 mm, $n = 6$. Triangular, not overly elongated, apical half of outer margin concave, apex falcate. Ground color gray with reddish brown suffusion, especially near interior edge of postmedial line, reddish coloration near apex along apical interior of postmedial line, overall moderately speckled by dark petiolate scales. Discal spot very faintly marked by light gray oblong shape. Apex marked by black scales above apical dash. Black postmedial line mostly straight except when very near apex, strongly contrasting. Submarginal area light gray with whitish suffusion mesally, postmedial lunule as white band originating from apical dash, white band follows postmedial line from apex to posterior wing margin. Antemedial line brown, almost nonexistent. *Forewing venter*: As in forewing dorsum but antemedial area lighter gray, blood red suffusion present, especially along interior edge of postmedial line and near apex, antemedial line absent, small black discal mark occasionally present. *Hindwing dorsum*: Triangular, anal angle weakly accentuated with reddish coloration, similar coloration and patterning as forewings, antemedial line absent, postmedial line weakly bent toward anterior wing margin, weakly concave or straight mesally. *Hindwing venter*: Following similar pattern as forewing venter, but red coloration near anal angle much darker, discal mark absent or very faint. *Abdomen*: As for genus but somewhat elongated, nearly sphingiform. Coloration a continuation of gray thoracic color. Midventral stripe absent. *Genitalia*: (Fig. 91) $n = 5$. Tegumen ovoid, constricted near base of gnathos. Vinculum rectangular, somewhat quadrate ventrally. Valves somewhat triangular, narrow, saccular edge of left and right valves with triangular tooth proximal to transtilla, both valves with smaller mesal costal projection originating from central ridge of valve, mesal costal projection immediately above saccular edge teeth, apex of projection toward saccular edge. Valves rounded or nearly pointed apically. Uncus truncated apically, apex rounded. Gnathos as two prominent flattened, moderately sclerotized, flap-like, somewhat triangular, upward facing extensions with highly truncated apices. Apices form elongated fingerlike projections. Juxtal processes roughly phallus length, weakly sclerotized, curving toward apex of phallus. Juxtal processes thin, covered in fine setae. Base of phallus with paired, backwards facing, elongated, fingerlike, diverging lobes. Phallus broad, irregularly edged dorsum usually with accentuated, rounded mesal protuberance but sometimes much reduced and flattened, always covered in setae. Left edge of rolled phallus forming mesally situated protuberance, base of sclerotized terminus of phallus with weak ventral bump, angled ventrally or away from end of phallus, distal tip of phallus separated into two distinct points. Vesica somewhat elongated, bag-like, covered in setae laterally, originating from progressively weakened scleroti-

zation of diagonal edge of phallus. **Female.** *Head:* As in male. *Thorax:* As in male. *Legs:* As in male. *Forewing dorsum:* Forewing length: 26 mm, $n = 1$. Maculation as in male, wing slightly broader, barely more elongate, less triangular, less falcate. *Forewing venter:* As in forewing dorsum but lighter gray, antemedial line absent, thin black discal mark present. *Hindwing dorsum:* As in male but more rounded, less triangular. *Hindwing venter:* Following similar pattern as forewing venter. *Abdomen:* As in male but more robust. Sternite of VIII as pair of thin sclerotized bands converging near anterior margin of VIII forming well-defined rectangular sternite. *Genitalia:* (Fig. 103) $n = 1$. Tergite of VIII robust, forming posteriorly directed triangle, membranous gap mesally. Apophyses anteriores about same length as apophyses posteriores, apophyses posteriores slightly thicker. Lamella antevaginalis thin, curved, slightly indented mesally near ostium bursae. Ductus bursae short. Papillae anales elongated, covered in relatively long setae.

Distribution. This new species is a resident of the Brazilian Cerrado in the states of Goiás and Minas Gerais, as well as in Distrito Federal. An additional record from Maranhão will be discussed below in the remarks.

Etymology. This new species is named for the phallus, which, due to the dorsal protuberance, bears the likeness of a dolphin (=delphinus Latin) when viewed laterally.

Natural history. Dr. A. Camargo (CPAC) kindly provided additional information pertaining to one of the paratypes (specimen number 12816), and is thus the only available natural history information for this new species. The pupa of this specimen was collected on *Miconia albicans* (Melastomataceae) on 8.VI.1992 and the subsequent adult eclosed on 30.X.1992. This is the only record of *Menevia* found on Melastomataceae, but cannot be definitively considered a host record without determining if this species was feeding on the plant prior to pupation.

Remarks. *Menevia delphinus* represents another species similar to the large, widespread *M. vulgaris* and its Brazilian Atlantic Forest counterpart, *M. franclemonti*. *Menevia delphinus*, like three other similar species described as new in the present work, is much smaller and with much more complicated phallic structures than *M. vulgaris* and *M. franclemonti*.

Due to the lack of data regarding the distribution of this species, other than it clearly being found in the Brazilian Cerrado, we consider it is necessary to report another state record for this species, despite our inability to gain access to the specimen in question. The recently described, and very distinct, *M. alurca*, was described from eight males (Herbin and Mielke 2014). The two authors of *M. alurca* were kind enough to supply us with specimens or photos of specimens such that the holotype and all seven paratypes could be examined. Upon close examination, it was discovered that a single undissected paratype, from the type locality of *M. alurca*, fits our concept of *M. delphinus* based on the continuous white band along the external edge of the postmedial line, and the lack of a ventral abdominal line. *Menevia alurca* has a discontinuous white band and a very prominent dark, ventral, abdominal line. We therefore consider *M. delphinus* to be present in Maranhão, which is highly likely given the Cerrado habitat there (Herbin and Mielke 2014).

The single reared specimen of *M. delphinus* was much smaller than other *M. delphinus* specimens and its genitalia differed in the shape of the dorsal protuberance of the phallus, which was flattened rather than distinctly raised. However, other genital characteristics, such as the highly elongated lobes at the base of the phallus, were consistent with *M. delphinus*. Given the consistency in other characters and the close proximity of this specimen's locale to the type locality of *M. delphinus*, we include this specimen in the type series and attribute the different phallic structure to the overall small size.

Mimallo saturata* Walker, 1855, *nomen dubium

Type material. **Holotype**, ♀, presumed lost/destroyed. Type locality: Brazil: Rio de Janeiro (see remarks of *Menevia plagiata*).

The unknown taxon *Mimallo saturata* was briefly discussed above in the remarks relating to *Menevia plagiata* because some specimens of *Menevia* had previously been attributed to *Mimallo saturata* (USNM; BOLD database). Given that the original description of *Mimallo saturata* includes characters that are not known in any *Menevia*, or even in any Mimallonidae, such as a red abdomen with yellow hairs and an orange stripe along each side, we treat *Mimallo saturata* as a *nomen dubium* until specimens from near the type locality matching this description can be located.

Additional discussion

Menevia is a wide-ranging genus displaying distinct patterns of speciation. For example, both the *lantona* and *lucara* species-groups are widely sympatric, with exceptions only in Central America. In both species-groups we discovered taxa distinct from the nominotypical species on the peripheries of those species' distributions, in southeastern Brazil, Central America, and the Andean Cordillera Oriental. The *plagiata* species-group also showed similar patterns of speciation in the Andean Cordillera Oriental and southeastern Brazil. Additionally, the *plagiata* and *ostia* species-groups both have unique Brazilian Cerrado species. The broad overlap in the distributions of the four species-groups suggests that they may have originated from the same geographic region, with various degrees of parallel speciation.

The distinct dorsal phallic ridge or projection of the male genitalia is an important character used to differentiate species in this genus, and interestingly, all species from southeastern Brazil, except *M. franclemonti*, display this character, whereas all Amazonian and Central American species, except *M. ostia*, do not. Additionally, the mid-ventral abdominal stripe is present only in central and southeastern Brazilian species, namely in *M. plagiata*, *M. alurca*, *M. australis*, and *M. magna*. This trait is not present in Amazonian or Central American species. Most species displaying the midventral

abdominal stripe belong to a closely-knit section of the *M. plagiata* subgroup of the *plagiata* species-group, therefore its presence in *M. magna* of the *lantona* species-group implies that this trait may be plesiomorphic along with dorsal ridges/projections of the phallus which this species also displays. Furthermore, St Laurent and Dombroskie (2015) showed that *Eadmuna pulverula* (Schaus, 1896), also of southeastern Brazil, possesses a midventral abdominal stripe as well, which may be a character useful to aid in determining the phylogenetic relationships among these taxa.

A clearer understanding of the evolutionary origins of *Menevia*, and Mimallonidea as whole, would be particularly interesting because recent higher-level phylogenetic studies of Lepidoptera continue to demonstrate not only the uniqueness of Mimallonidae as the only family in Mimallonidea, but also its key phylogenetic role as a potential sister lineage to the Macrolepidoptera (Timmermans et al. 2014, Kawahara and Breinholt 2014).

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Three new genera of Neotropical Mimallonidae (Lepidoptera, Mimallonoidea, Mimallonidae) with descriptions of three new species

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Abstract

Three new genera of Mimallonidae are described. The monotypic genus *Tostallo* **gen. n.** is erected to contain “*Perophora*” *albescens* Jones, 1912, which was previously placed in the preoccupied genus *Perophora* Harris, 1841 and was never formally moved to a valid genus. *Perophora* is a junior homonym of *Cicinnus* Blanchard, 1852, but the name *albescens* is not appropriately placed in *Cicinnus* due to external and genitalia characteristics entirely unique to the species *albescens*. The female of *Tostallo albescens* **comb. n.** is described and both sexes are figured for the first time. *Auroriana* **gen. n.** is erected to contain *A. florianensis* (Herbin, 2012), **comb. n.** previously described as *Cicinnus florianensis*, and two new species: *A. colombiana* **sp. n.** from Colombia and *A. gemma* **sp. n.** from southeastern and southern Brazil. The female of *A. florianensis* is described and figured for the first time. Finally, the monotypic genus *Micrallo* **gen. n.** is erected to include a new species, *M. minutus* **sp. n.** described from northeastern Brazil.

Keywords

Auroriana, biodiversity, *Cicinnus*, *Micrallo*, Neotropical, taxonomy, *Tostallo*

Introduction

Schaus (1928) described 16 of the 28 currently recognized Mimallonidae genera, with only *Naniteta* Franclemont, 1973 and *Arianula* Herbin, 2012 having been described since then. Franclemont (1973) recognized strong affinities between all of the genera and speculated that the family must be recently diversified. The number of genera in Mimallonidae is high relative to the small size of the family, wherein only a little more than 200 species have been described (Becker 1996, Herbin 2012, 2015, Herbin and Mielke 2014, Herbin and Monzón 2015, St Laurent and Dombroskie 2015). Therefore, many of the genera are represented by three or fewer species (St Laurent and Dombroskie 2015), including the three new genera described here.

Currently there is no accepted subfamily arrangement for the family Mimallonidae. Schaus (1928) proposed two subfamilies based primarily on the presence/absence of the frenulum. Schaus frequently failed to recognize the presence of vestigial frenula when they were in fact present, hence his subfamily arrangement has been rejected by most contemporary authors (Pearson 1951, 1984, Franclemont 1973, Herbin 2012, St Laurent and Dombroskie 2015, but see Becker 1996 and Herbin and Monzón 2015). We continue to recognize the lack of a clear subfamily arrangement awaiting a higher-level treatment of the family.

Recent interest in the taxonomy of Mimallonidae has resulted in numerous new species (Herbin 2012, 2015, Herbin and Mielke 2014, Herbin and Monzón 2015, St Laurent and Dombroskie 2015) described over a short period of time, displaying the previous lack of work on the group. Recent efforts by Herbin (2012) and on-going investigations by the first author of the present article have resulted in new genera, three of which are described herein.

Methods

Dissections were performed as in Lafontaine (2004). Morphological, including genitalia, terminology follows Kristensen (2003). Figures were manipulated with Adobe Photoshop CS4 (Adobe 2008).

Male genitalia are figured in natural color with CS4 “auto color” used to improve white backgrounds. Most genitalia were photographed with a Macroscopic Solutions Macropod Pro and Canon EOS 6D DSLR camera body using the Macro Photo MP-E 65mm f/2.8 1–5× Manual Focus Lens for EOS except when received from other individuals or institutions. Thirty (3×) photographs were taken of each specimen in ethanol under glass, and stacked using Zerene Stacking Software.

Maps were created with SimpleMappr (Shorthouse 2010) and edited with CS4. All geographical co-ordinates are approximate, and are based on the localities provided on specimen labels. Co-ordinate data were acquired with Google Earth (2015).

Specimens from the following collections were examined:

CGCM Coll. Carlos G. C. Mielke, Curitiba, Paraná, Brazil

COM	Coll. Olaf H. H. Mielke, Curitiba, Paraná, Brazil
CPC	Coll. Philippe Collet, Caen, France
DZUP	Coll. Pe. Jesus S. Moure, Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil
MNHN	Muséum nationale d'Histoire naturelle de Paris, France
MZSP	Coll. Museu de Zoologia, Universidade de São Paulo, São Paulo, São Paulo, Brazil
BMNH	The Natural History Museum [statutorily British Museum (Natural History)], London, U.K.
RAS	Research collection of Ryan A. St. Laurent, Ithaca, New York, USA
USNM	National Museum of Natural History [formerly United States National Museum], Washington D.C., USA

Results and discussion

Tostallo St Laurent & C. Mielke, gen. n.

<http://zoobank.org/5C06B802-4FA9-4DDA-B6E6-B4D422292E0E>

Type species. *Perophora albescens* Jones, 1912.

Etymology. The genus name is derived from the toasted (*tostus* Latin) appearance of the white forewings, which is unique among Mimallonidae and reminiscent of toasted marshmallows; + the ending -llo, which is shared with *Mimallo* Hübner, [1820], the type genus of Mimallonidae. The genus name is masculine.

Diagnosis. The monotypic *Tostallo* is remarkable among Mimallonidae in that the ground color is nearly white and the forewings rounded, a combination of characters seen nowhere else in the family. Male genitalia are unique in that the gnathos is formed by two large columnar structures with multiple invaginations and internal wrinkles. Female genitalia can be recognized by the wrinkled, setae covered accessory part of segment VIII, which encircles the papillae anales, as well as by the apophyses posteriores, which are hollow, almost balloon-like lobes, rather than thin rods as in other genera.

Description. Male. *Head:* Width half that of thorax, light brown, eyes bordered posteriorly by dark-brown scales encircling head, forming dark mane; palpus not extending beyond frons, covered dorsally in darker brown scales; antenna opaque yellow, bipectinate to tip, rami increasing in length from antennal base to roughly middle of antennal length where rami length continuously decrease until terminus. *Thorax:* Pale tan brown, lighter near wing base, darker mesally, overall lightly speckled with darker petiolate scales. *Legs:* Color as for thorax, but tarsus light brown, petiolate scales present. Tibial spurs very thin, elongate, clothed in fine white scales. *Forewing dorsum:* Forewing length: 12–13 mm, avg. 12.5 mm, Wingspan: 23–26.5, n = 3. Subtriangular, outer margin convex, apex not pronounced. Ground color cream to white, overall lightly speckled by dark petiolate scales; outer margin fringed, grey. Antemedial line faint pale brown, somewhat wavy, especially

near costa. Postmedial line dark brown, waxy, convex and mostly evident from costa to Rs3. Basal and medial areas proximally concolorous with thorax; medial area distally gray reaching inner margin towards wing base. Submarginal band darker, compressed, sometimes absent between postmedial line and marginal band. The latter grayish brown, delineated by an undulating line suffused with white distally. Discal spot marked by small, tan, crescent-shaped mark. Black patch of scales at apex extends to Rs3. *Forewing ventrum*: Similar to dorsum but more yellow brown, white and gray suffusions absent, submarginal and marginal bands lighter, postmedial line continuous, submarginal and antemedial lines absent. *Hindwing dorsum*: Coloration as for forewing dorsum, but lighter. Antemedial line brown, partially visible near inner margin. Postmedial line faintly marked, bordered by white distally. Submarginal band grayish brown. Marginal band lighter. *Hindwing ventrum*: Follows same pattern as forewing ventrum but discal mark absent. Frenulum a single, relatively-thick bristle. *Venation*: Typical of Mimallonidae, very similar to *Cicinnus melsheimeri* (Harris, 1841), but forewing M_1 and M_2 originate from cell nearer to R and CuA_1 respectively, than to each other. *Abdomen*: Short, not exceeding hindwing tornus, stout, coloration as for thorax, ventrally pale. *Genitalia*: (Fig. 4) $n = 4$. Simple; tegumen reduced to slender rod. Vinculum box-like, not projected anteriorly. Uncus simple, subtriangular, truncated apically. Gnathos as two unfused, heavily-sclerotized, multiply-invaginated columnar structures with multiple internal folds. Valves short, irregularly shaped, somewhat triangular, truncated apically forming lobe, sclerotization weak mesally with heavily sclerotized fingerlike projection subapically. Juxta partially fused to phallus, separate portion of juxtal plate embedded in anellus. Phallus short, pistol shaped; flattened, open dorsally; base of phallus with short, anteriorly-curved lobe, vesica small, somewhat bag-like. **Female.** *Head*: As for male. *Thorax*: As for male. *Legs*: As for male, but tibial spurs slightly longer. *Forewing dorsum*: Forewing length: 14–16.5 mm, avg. 15.25 mm, Wingspan: 26–35 mm, $n = 2$. As for male but slightly longer, black patch of scales at forewing apex extended to Rs4 rather than Rs3. *Forewing ventrum*: Similar to dorsum but yellower brown. Antemedial line absent. Postmedial line continuous, submarginal band lighter. *Hindwing dorsum*: As for male but somewhat broader. *Hindwing ventrum*: Follows same pattern as forewing ventrum but discal mark absent. Frenulum apparently absent or highly reduced. *Abdomen*: As in male but more robust, sclerotized bands present ventrally on segment VIII. *Genitalia*: (Figs 5, 6) $n = 2$. Papillae anales setose, somewhat flattened ventrally, covered in short denticles from which very short setae originate. Apophyses anteriores very short, robust, flattened; apophyses posteriores as hollow, rounded, balloon-like lobes. Ductus bursae relatively long, corpus bursae bag-like with slight sclerotization terminally. Dorsal sclerotization of tergite VIII thin, folded inward, arc-like, with asymmetrical ridges on lateral sides of central arc. VIII encircles genitalia as thick, moderately sclerotized, setae covered, finely wrinkled structure, invaginated around entire circumference. Ostium unsclerotized. Lamella antevaginalis reduced, subtriangular, small notch apically.

Remarks. *Perophora albescens* Jones, 1912 was described in the preoccupied genus *Perophora* Harris, 1841 (*Perophora* Wiegmann, 1835 [Tunicata]) despite the fact that another name, *Ptochopsyche* Grote, 1896, was proposed as a replacement well before Jones's description. Both *Perophora* and *Ptochopsyche* are currently treated as synonyms of *Cicinnus* Blanchard, 1852 (Becker 1996). However, *T. albescens* comb. n. lacks the external and genital characters that are "characteristic" of the poorly defined genus *Cicinnus* (sensu Franclemont 1973, Herbin 2012, 2015, Herbin and Mielke 2014, Herbin and Monzón 2015), namely in that the forewings are not falcate and the valves of the male genitalia are simple rather than variously reduced as in *C. melsheimeri*, the type species of both *Perophora* and *Ptochopsyche*. Thus it is not appropriate to merely consider *albescens* within *Cicinnus*.

Schaus (1928), the last to completely revise the family, was unable to "identify" "*Perophora albescens*" and only re-iterated Jones's description without assigning the species to a valid genus. Additionally, Becker (1996) did not list this species. Further compounding the ambiguity of the taxon *albescens*, neither sex of this species has been previously figured. While the validity of the species has never been questioned, Schaus's inability to locate the holotype likely has led to this taxon being ignored or overlooked in all subsequent literature. Given the opportunity to re-examine Jones's original specimens, including the holotype, and perform comparisons to all known Mimallonidae genera, it is readily apparent that *T. albescens* comb. n. is unique among Mimallonidae and should be transferred to a new, monotypic genus. We therefore make this long overdue change in the present work, moving the name *albescens* from the preoccupied *Perophora* to a new genus, to solidify its uniqueness in the family. We provide figures of the holotype male, an additional male and female, as well as the genitalia of both sexes to allow for better recognition of this rarely reported enigmatic species.

***Tostallo albescens* (Jones, 1912), comb. n.**

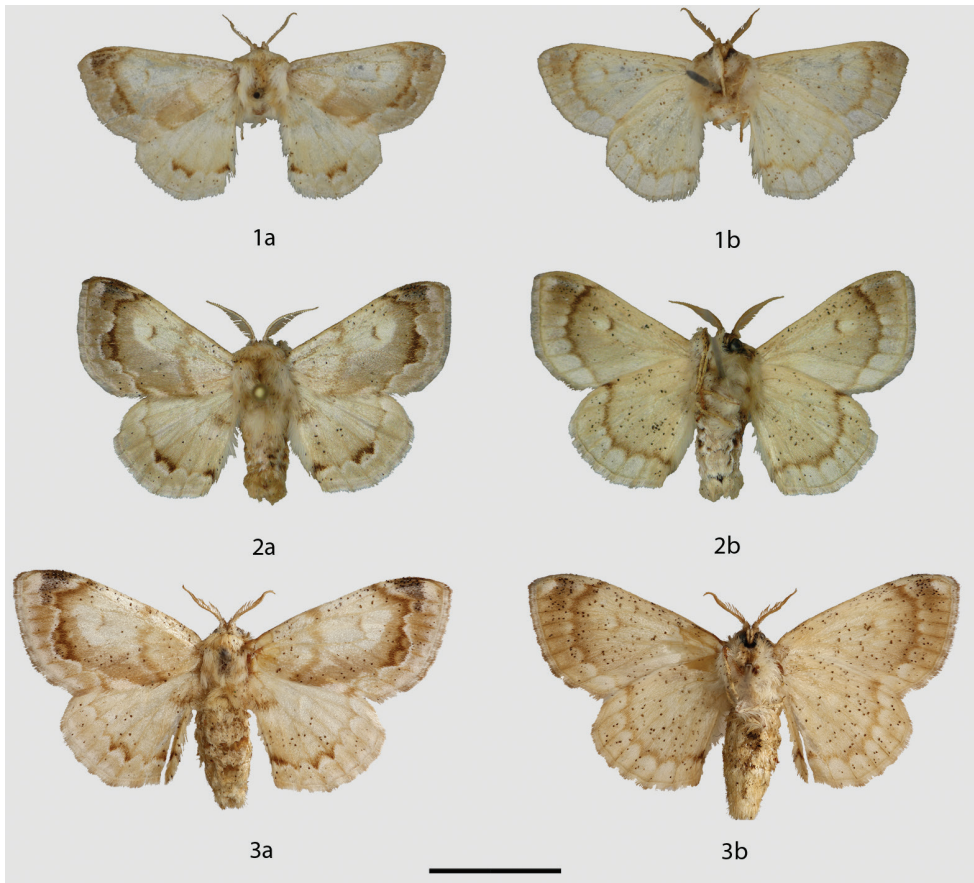
Figs 1–7

Perophora albescens Jones, 1912: 435

Perophora albescens Jones; Schaus 1928: 671–672

Type material. **Holotype**, ♂: S. Paulo, S. E. Brazil./ *Perophora albescens*, Type ♂ .D. Jones/ E. D. Jones Coll., Brit. Mus. 1919–295/ BMNH(E) #805419/ Mimallonidae BMNH(E) Slide #001/ (BMNH, examined). Type locality: Brazil: São Paulo.

Additional specimens examined. (5 ♂, 2 ♀) **ARGENTINA: Misiones:** 1 ♂, Posadas: 26.IX.1921, BMNH(E) 1378118, St. Laurent diss.: 10-21-15:5 (BMNH). **BRAZIL: Paraná:** 1 ♂, 1 ♀, Tirol das Torres, Uberaba, Curitiba, 900 m: 2.III.2000, 2.I.2001, O. Mielke leg., C. Mielke diss.: 51.381, 53.261 (COM 51.381, 53.261). **Rio Grande do Sul:** 1 ♀, illegible collector and date: Joicey Coll. Brit. Mus. 1925–157, BMNH(E) 1378120, St. Laurent diss.: 10-21-15:6 (BMNH). **São Paulo:** 1 ♂, Guapiara, Paivinha, 800 m: 16–19.IX.2005, C. Mielke leg., (CGCM 28.855). 1 ♂, Alto da Serra: IX.1935, Coll. R. Spitz, Brit. Mus-112(?), BMNH(E) 1378119, St.



Figures 1–3. *Tostallo albescens* adults, **a** recto, **b** verso. **1** Holotype ♂, Brazil, São Paulo (BMNH) **2** ♂, Brazil, São Paulo, Guapiara, Paivinha, 800 m (CGCM) **3** ♀, Brazil, Rio Grande do Sul (BMNH). Scale bar = 1 cm.

Laurent diss.: 10-21-15:4 (BMNH). 1 ♂, “S.E. São Paulo, 750 m”: E.D. Jones, E.D. Jones Coll. Brit. Mus. 1919–295, BMNH(E) 1378117 (BMNH).

Diagnosis. See genus diagnosis.

Description. See genus description.

Distribution (Fig. 7). *Tostallo albescens* primarily inhabits the Brazilian Atlantic Forest in the states of São Paulo and Paraná. A single record from Rio Grande do Sul lacks more specific data, thus we are unable to determine if this record comes from the Atlantic Forest region in the northeastern part of the state or from elsewhere. A single record from Misiones, Argentina, suggests that this species is more widespread in the inland ecoregions of the Atlantic Forest biome.

Remarks. As mentioned in the genus remarks above, this species is unique among Mimallonidae, namely because of the white ground color and rounded forewings. This species is poorly represented in collections.

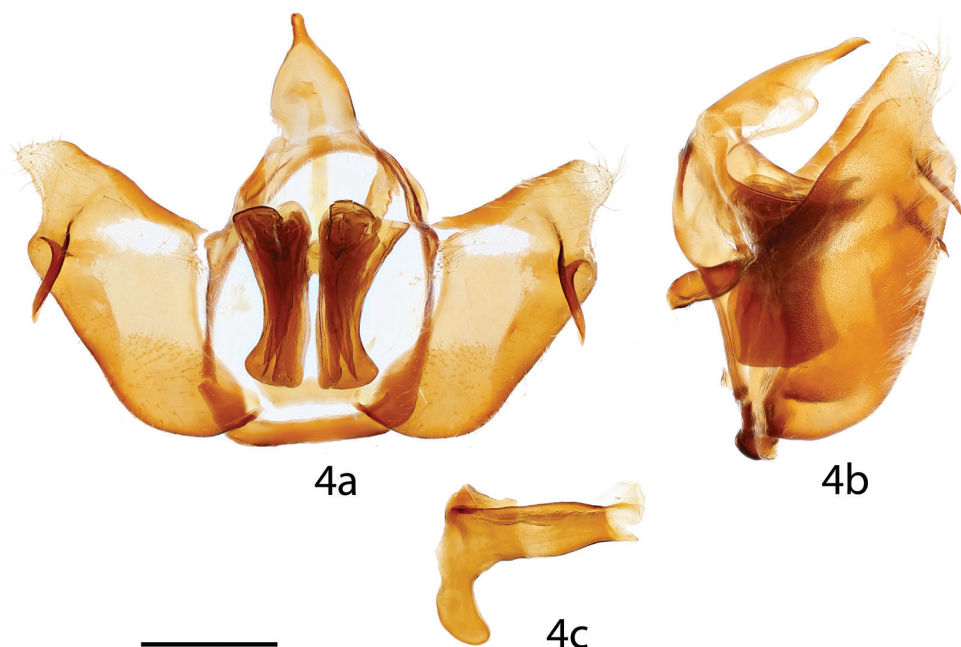


Figure 4. *Tostallo albescens* male genitalia, **a** ventral, **b** lateral, **c** phallus. Brazil, São Paulo, Alto da Serra [St. Laurent diss.: 10-21-15:4] (BMNH). Scale bar = 1 mm.

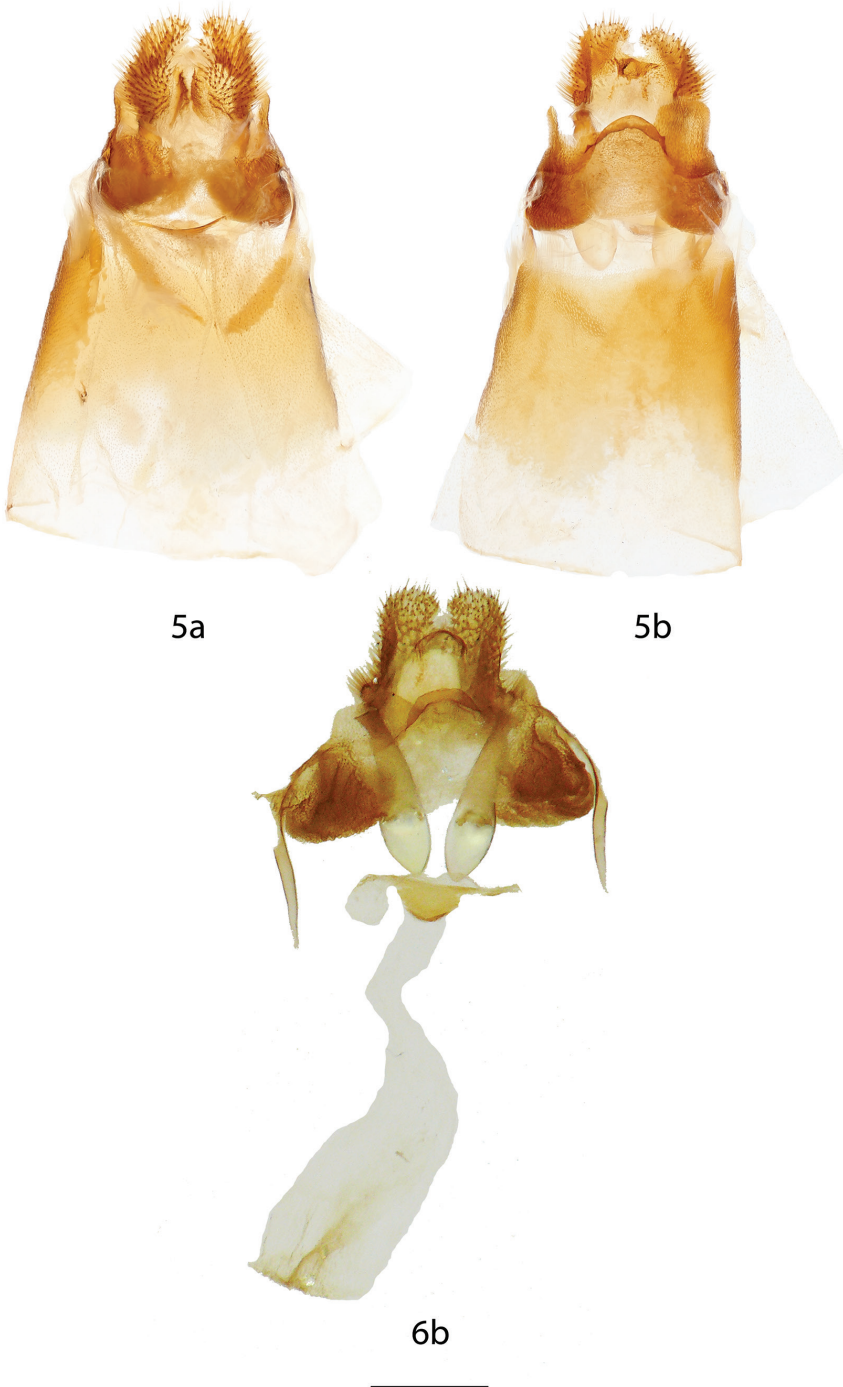
***Auroriana* St Laurent & C. Mielke, gen. n.**

<http://zoobank.org/31C8B6C2-3B3D-4CF4-94EB-BBC312041FDF>

Type species. *Auroriana colombiana* St Laurent & C. Mielke, sp. n.

Etymology. This genus is named for the sunrise (*aurora* Latin) because of the pink and orange coloration of all species in the genus. These colors are reminiscent of the light scattering phenomenon of the rising sun. The genus name is feminine.

Diagnosis. *Auroriana* can be distinguished from all currently described Mimallonidae by the orange and diffuse pink coloration on all wings, and by the pink and beige head and thorax. Male genitalia are unique in the combination of the following characters: variably-shaped, heavily-sclerotized, unfused gnathos processes, relatively-simple valves, and downward-curved phallus with a distinct lobe-like process at the base. Only a few other similarly-sized, orange and pink Mimallonidae species are known: *Druentica fatella* (Schaus, 1905), *Trogoptera semililacea* (Dognin, 1916), and the species of *Reinmara* Schaus, 1928. These species, while superficially similar to *Auroriana*, can be easily differentiated by the straight forewing postmedial line and by the stark contrast between the completely or nearly completely pink medial region and the orange postmedial region rather than the much more diffuse pink coloration in *Auroriana*. Additionally, the dark brown wing fringes accent the notch at the tornus of both *D. fatella* and *T. semilacea*, a character not seen in *Auroriana*. *Druentica fatella* has



Figures 5, 6. *Tostallo albescens* female genitalia, **a** ventral, **b** dorsal. **5** Brazil, Rio Grande do Sul [St. Laurent diss.: 10-21-15:6] (BMNH) **6** Brazil, Paraná, Curitiba [C. Mielke diss.: 51.381] (COM). Scale bar = 1 mm.



Figure 7. Distribution of *Tostallo albescens*. The square represents a generalized (center of the state) locality for Rio Grande do Sul, a state for which detailed locality data is lacking.

a characteristic dark tuft of scales at the terminus of the abdomen, which is absent in *Auroriana*. Furthermore, the genitalia of these species are entirely distinct from those of *Auroriana*, where all major characters pertinent to the diagnosis of *Auroriana* are absent. Interestingly, the present study reveals that *D. fatella* likely does not belong to its present genus due to a combination of external and genitalia characters unlike what is seen in other species of *Druentica*.

Description. Male. *Head:* Width roughly two thirds that of thorax, pinkish; labial palpus not extending beyond frons, pink; antenna bipectinate to terminus; rami increasing in length from antennal base to roughly one fifth of antennal length where rami length is nearly constant for following fifth of antennal length after which rami continuously decrease in length until terminus. *Thorax:* Pink and beige, thickly covered in long scales, darker petiolate scales absent. *Legs:* Light pink, vestiture long, tarsus usually tan, tibial spurs similar to those of *Eadmuna* Schaus, 1928. *Forewing dorsum:* Forewing length: 16–21 mm. Somewhat triangular, slight inward notch at tornus, margin convex mesally. Ground color light orange brown, overall very sparsely speckled by dark petiolate scales. Antemedial line faint pink, nearly straight or bowed slightly. Postmedial line slightly or moderately bowed inward, dark brown, outward edge lined with very pale pink band of varying width, postmedial line angled sharply to-

wards costa after passing Rs₄. Discal spot a small black mark. Antemedial area usually more solid pink than diffusely shaded medial area. Sub- and marginal areas darker than medial area. *Forewing ventrum*: As for forewing dorsum but antemedial and postmedial lines faint or absent; pink suffuse line near apical quarter extending from costa to postmedial suffusion. *Hindwing dorsum*: Coloration as for forewing dorsum, but less pink, postmedial line may lack outer pink edging. *Hindwing ventrum*: As in for forewing ventrum but postmedial line better developed, lobed outward mesally. Frenulum a single bristle. *Venation*: Typical of Mimallonidae, very similar to *C. melsheimeri* but CuA₁ bent more posteriorly. *Abdomen*: Short, not extending beyond hindwing tornus, stout, coloration as for thorax but with more beige than pink. *Genitalia*: Somewhat complex; tegumen very broad to narrow, subtriangular, or more rectangular. Vinculum ovoid or irregularly shaped, lobes present mesally below gnathos, or when not present, elongated, trumpet-like structures present at base of valves instead; uncus tubular or triangular, in most species hardly differentiated from tegumen; gnathos as two unfused heavily-sclerotized processes, either roughly triangular or two-pronged. Valves triangular, with triangular or rounded saccular edge lobe, valve with mesal tooth present in one species; saccular edge of valve with longer, heavier setae in Amazonian species. Juxta fused to phallus, with elongate ventral process connecting phallus at base of valves. Phallus variable in length, but always curved downward; basally with ventrally angled elongation, terminus with hook-like process or simple. **Female.** *Head*: As for male, but antennal rami shorter. *Thorax*: As for male, but with more beige scales. *Legs*: As for male. *Forewing dorsum*: Forewing length: 17 mm. As for male, but broader, pink suffusions reduced, ground color more olive green than orange. *Forewing ventrum*: As for forewing dorsum but pinkish suffusions nearly absent, postmedial line absent, discal mark more pronounced. *Hindwing dorsum*: As for male but slightly broader, dark petiolate scales more numerous, especially antemedially. *Hindwing ventrum*: Following same pattern as forewing ventrum. *Abdomen*: As for male, but broader. Sternite VIII with pair of short sclerotized bands posteriorly. *Genitalia*: Papillae anales broad, somewhat rectangular, covered in setae, increasing in length at base. Apophyses anteriores with curved tips; apophyses posteriores robust, wide, slightly longer and broader than apophyses anteriores. Ductus bursae very long, somewhat sclerotized near ostium, ductus not differentiated from long, narrow corpus bursae. Dorsal sclerotization of tergite VIII as narrow band with posteriorly directed mesal arc with membranous center. Lamella antevaginalis very large, broad, trapezoidal, notched mesally.

Remarks. This new genus is erected to include three South American species. All three species are known from very few specimens, being poorly represented in collections. Herbin (2012) was the first to recognize the uniqueness of *A. florianensis* at the time of its original description, but did not describe a new genus in which to place it. Upon the discovery of two additional species similar to *A. florianensis*, the first author determined that external morphology and male genital characteristics united these three species and set them apart from all other described Mimallonidae genera. We hereby describe a new genus in which to place the previously described *A. florianensis* and the two new species described below.

Key to species of *Auroriana**

- 1 Forewing postmedial line nearly straight (Figs 8, 12) **2**
- Forewing postmedial line bowed inward toward thorax (Figs 9–11)
..... ***A. florianensis***
- 2 Hindwing postmedial line thin, well defined, pale diffuse outer band absent..
..... ***A. colombiana***
- Hindwing postmedial line as pale, pink, diffuse band ***A. gemma***

*Note: the females of *A. colombiana* and *A. gemma* are unknown.

***Auroriana colombiana* St Laurent & C. Mielke, sp. n.**

<http://zoobank.org/3BDB9C32-B1A1-43F1-A5D5-EE3815262CE3>

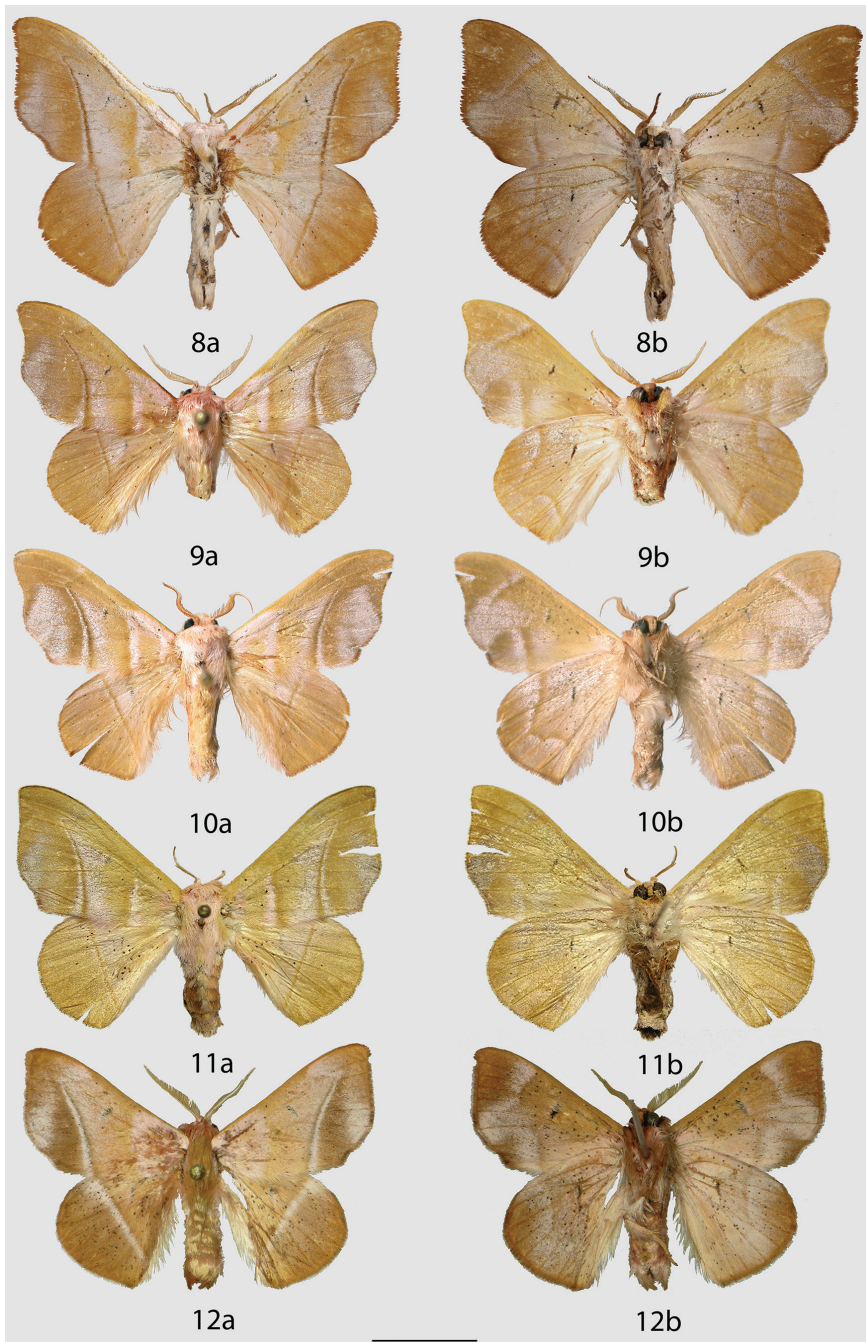
Figs 8, 13, 14, 19

Type material. **Holotype**, ♂: Ob. Rio Negro [Upper Río Negro], Ost Colombia 800 m, Coll. Fassl/ Joicey Coll., Brit. Mus. 1925–157/ BMNH(E) 1378121/ St. Laurent diss.: 11-3-15:1/ HOLOTYPE male *Auroriana colombiana* St Laurent and C. Mielke, 2016 [handwritten red label]/ (BMNH). Type locality: Colombia: Meta: Rio Negro.

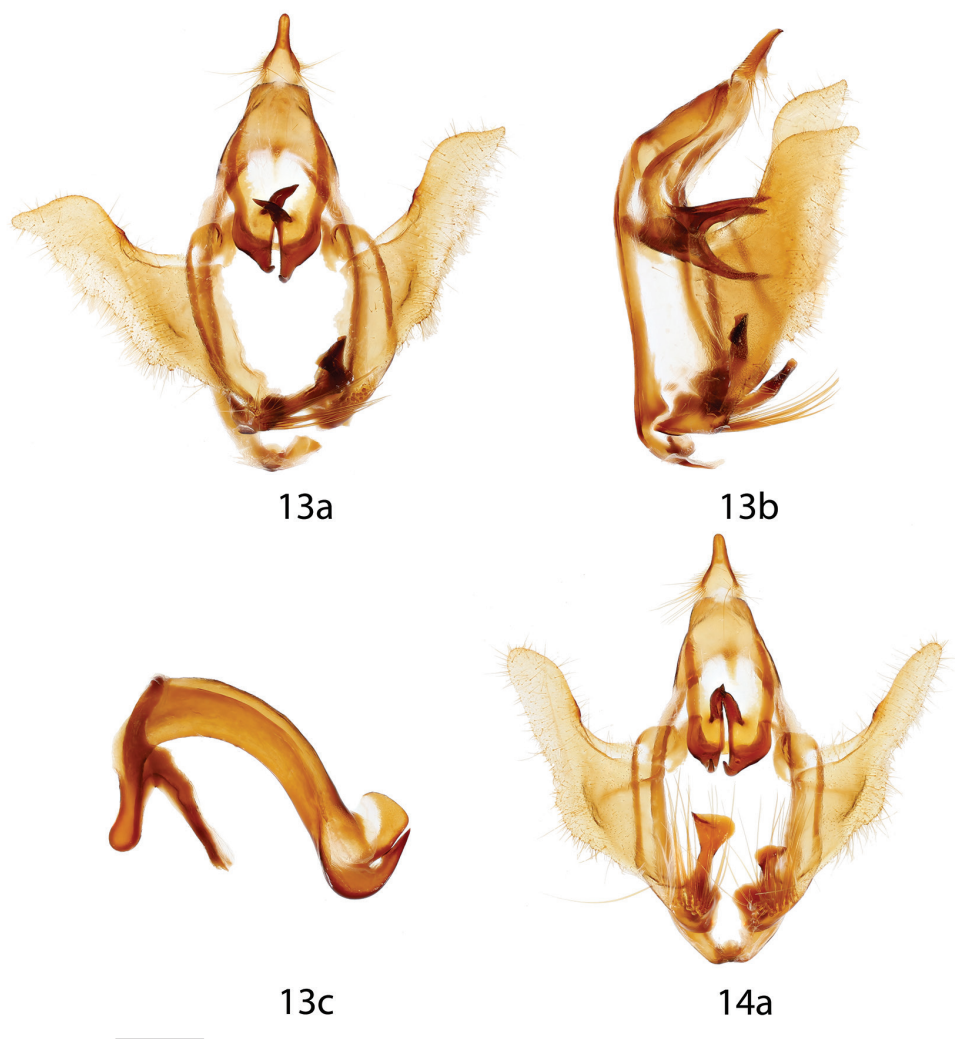
Paratype, 1 ♂: **COLOMBIA:** Upper Río Negro [Meta], 800 m: Coll. Fassl, Joicey Coll., Brit. Mus. 1925–57, BMNH(E) 1378116, St. Laurent diss.: 10-21-15:3 (BMNH). Paratype with the following yellow label: PARATYPE male *Auroriana colombiana* St Laurent and C. Mielke, 2016.

Diagnosis. This new species is most similar to *A. florianensis* in adult habitus but can easily be differentiated by the larger size, darker, straighter, better defined post-medial lines on all wings, and the more solid pink medial area. The genitalia also immediately distinguish these two species: in *A. colombiana* the valves are narrower and less triangular, and the paired processes of the gnathos are two-pronged structures, rather than thick, triangular, basally-spined structures as in *A. florianensis*. The phallus is similar in both species, but in *A. colombiana* it is larger, more deeply curved, and the distal curve is more developed and backwardly angled.

Description. Male. **Head:** Light pink, eyes bordered posteriorly by black scales; antenna opaque yellow; labial palpus small, dorsally with darker scales. **Thorax:** Pink, interspersed with beige scales. **Legs:** Light pink, vestiture long, tarsus pale brown, tarsal spurs relatively long, clothed in pink scales except naked, finely-pointed apical tip. **Forewing dorsum:** Forewing length: 20–21 mm, avg. 20.5 mm, Wingspan: 36–38 mm, avg. 37 mm, n = 2. Triangular, slight inward notch at tornus, margin convex mesally. Ground color light orange brown, medial region very sparsely speckled by dark petiolate scales. Antemedial line diffuse, faint pink, nearly straight. Postmedial line bowed inward slightly, dark brown, outward edge finely lined with pale pink, especially near tornus, postmedial line angled sharply towards costa after passing Rs₄. Antemedial area pink. Postmedial area darker brown orange with pale



Figures 8–12. *Auroriana* adults, **a** recto, **b** verso. **8** *A. colombiana* holotype ♂, Colombia, Meta, Río Negro, 800 m (BMNH) **9** *A. florianensis* holotype ♂, French Guiana, Piste du Dégrad Florian, PK 12 [photo courtesy of MNHN] (MNHN) **10** *A. florianensis* ♂, French Guiana, St. Jean du Maroni, Plateau des Mines, PK 1.3 [photo courtesy of MNHN] (MNHN) **11** *A. florianensis* ♀, French Guiana, Réserve des Nouragues, Inselberg [photo courtesy of CPC] (MNHN) **12** *A. gemma* holotype ♂, Brazil, Santa Catarina, São Bento do Sul, Rio Natal, 550 m (DZUP). Scale bar = 1 cm.



Figures 13, 14. *Auroriana colombiana* male genitalia, **a** ventral, **b** lateral, **c** phallus. **13** Holotype, Colombia, Meta, Río Negro, 800 m [St. Laurent diss.: 11-3-15:1] (BMNH) **14** Paratype, Colombia, Meta, Río Negro, 800 m [St. Laurent diss.: 10-21-15:3] (BMNH). Scale bar = 1 mm.

pink-gray suffusion near wing margin, medial area lighter orange with pink suffusion, especially along postmedial line. Discal spot a small black, elongated mark. *Forewing ventrum*: Similar to dorsum but postmedial line very faint; antemedial line absent; deeper orange overall with reduced pink suffusion. *Hindwing dorsum*: Coloration as for forewing dorsum; well-defined postmedial line lacks outer pink edging. *Hindwing ventrum*: Follows same pattern as forewing ventrum but pinker, lighter overall. *Abdomen*: As for genus, coloration as for thorax, but less pink. *Genitalia*: (Figs 13, 14) $n = 2$. Tegumen narrow, rectangular. Vinculum ovoid with small pointed projection ventrally between saccular bases of valves. Uncus small, triangular. Gnathos as two

unfused heavily-sclerotized, two-pronged processes. Valves simple, bent upward, saccular edge with mesal lobe, base of valve with elongated, trumpet-shaped processes, process of left valve longer than that of right; base of valves with long, thick deciduous setae. Juxta fused to phallus, with elongate, flattened ventral process connecting phallus-juxta complex to base of valves. Phallus very elongated, curved, downturned, terminus with hook-like backwardly-angled process, base of phallus with elongated lobe. **Female.** Unknown.

Distribution (Fig. 19). *Auroriana colombiana* is so far known only from the type locality in central Colombia.

Etymology. This species is named for Colombia, as it is the only species in the genus known from this country.

Remarks. Although this species resembles *A. florianensis*, the striking genital differences clearly differentiate the two. *Auroriana florianensis* and *A. colombiana* seem to represent an Amazonian lineage of *Auroriana*, unified by the presence of heavier setae on the saccular basal edge of the valves and by the relatively large, elongate phallus, characters that are not present in *A. gemma* sp. n. described below.

***Auroriana florianensis* (Herbin, 2012), comb. n.**

Figs 9–11, 15, 17–19

Cicinnus florianensis Herbin, 2012: 23–25, figs 17, 18.

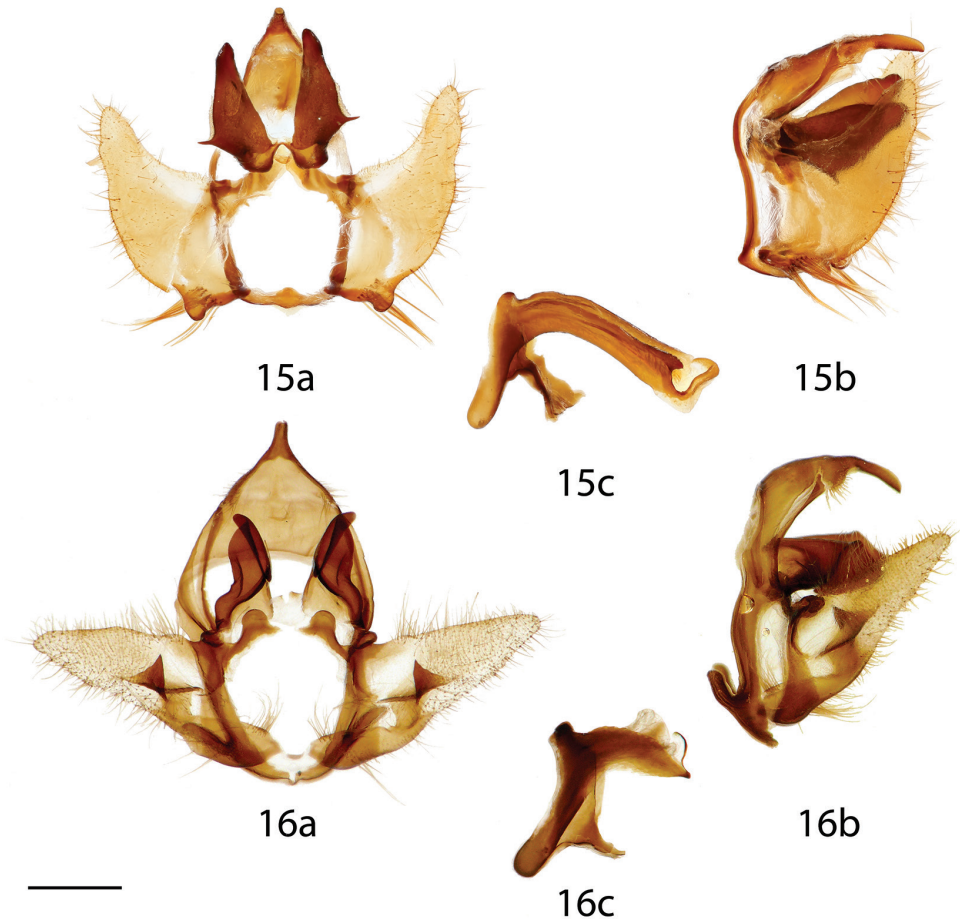
Type material. **Holotype**, ♂: 29-VII-2001, Piste du Dégrad Florian, PK 12, GUYANE Fcse, Coll. M. LAGUERRE, M. Laguerre leg./ genitalia, prep. D. Herbin, H. 674/ BC-Her1657/ *Cicinnus florianensis*, Herbin, 2012, The European entomologist, Vol. 4, N° 1, Holotype ♂/ (MNHN, examined). Type locality: French Guiana: Piste du Dégrad Florian, PK 12.

Additional specimens examined. (4 ♂, 1 ♀) **FRENCH GUIANA:** 1 ♂, St. Jean du Maroni, Plateau des Mines, PK 1,3: 29.VII.1993, L. Sènécaux réc. (MNHN). 1 ♂, Mont Mitaraka [Massif du Mitaraka], 300 m: 19.VIII.2015, La Planète revisitée, MNHN/PNI, APA 973-I, Ph. Collet leg., at mercury vapor light, St. Laurent diss.: 11-19-15:1 (RAS). 1 ♂, Saül, Point de vue: 30.VII.2011, Eddy Poirier leg., at UV light (CPC). 1 ♂, Réserve des Nouragues, Inselberg: 5.VIII.2010, Ph. Collet leg., at UV light (CPC). 1 ♀, Réserve des Nouragues, Inselberg, 4° 5'15" N, 52° 40'48" W, 110 m: 8.VIII.2010, at UV light, Eddy Poirier leg., Chr. Gibeaux diss. prep: 7759 (MNHN).

Diagnosis. *Auroriana florianensis* is similar to the previous species; see the diagnosis of *A. colombiana* for differentiating characters between both species. *Auroriana florianensis* can be distinguished from *A. gemma* sp. n. by the more strongly curved forewing postmedial line, straighter forewing antemedial line, and by the more expansive pink suffusion in the postmedial region. The genitalia can also be used to differentiate the two: in *A. florianensis* the tegumen is narrower, the gnathos processes larger, with

a sharp tooth basally, and the valves are bent upward, with heavier setae on the basal saccular edge. *Auroriana florianensis* lacks the mesal valve tooth present in *A. gemma* sp. n. Finally, the phallus of *A. florianensis* is much longer than in *A. gemma* sp. n.

Description. Male. *Head:* Pink, eyes bordered posteriorly by dark-brown scales; antenna opaque yellow; labial palpus small, dorsally with darker scales. *Thorax:* Pink, interspersed with beige scales, prothorax darker pink. *Legs:* Light pink, vestiture long, tarsus beige, tarsal spurs relatively long, finely pointed apical tips. *Forewing dorsum:* Forewing length: 17 mm, avg. 17 mm, n = 3, Wingspan: 32 mm. Triangular, slight inward notch at tornus, margin convex mesally. Ground color light orange brown, overall very sparsely speckled by dark petiolate scales. Antemedial line faint pink, nearly straight. Postmedial line bowed inward, dark brown, outward edge lined with pale pink, especially approaching tornus, postmedial line angled sharply towards costa immediately after passing Rs4. Antemedial area pink. Medial area lighter orange with pink suffusion. Submarginal area darker brown, marginal area with variably sized pink-gray suffusion. Discal spot a small black mark. *Forewing ventrum:* Similar to dorsum but ante- and postmedial lines absent, pink suffusion reduced antemedially and medially, discal mark more visible. *Hindwing dorsum:* Coloration as for forewing dorsum, but less pink; postmedial line lacks outer pink edging. *Hindwing ventrum:* Follows same pattern as forewing ventrum, but postmedial line diffuse, brown, outwardly lobed mesally. *Abdomen:* As for genus but coloration more beige than pink. *Genitalia:* (Fig. 15) n = 1. Tegumen broad, subtriangular. Vinculum somewhat rectangular. Uncus moderate length, tubular, hardly differentiated from tegumen. Gnathos as two unfused, heavily-sclerotized, somewhat triangular processes with sharp basal tooth. Simple triangular valves bent upward, narrowed distally. Base of valves with heavier, differentiated setae, heavily sclerotized lobes. Juxta fused to phallus, with wide ventral process connecting phallus to base of vinculum. Phallus elongated, curved, downturned, terminus with weak hook-like curl and heavily sclerotized forward pointing process. Base of phallus with elongated lobe. Vesica thick, bag-like. **Female.** *Head:* As for male, but antennal rami shorter. *Thorax:* As for male, but with more beige scales. *Legs:* As for male. *Forewing dorsum:* Forewing length: 17 mm, Wingspan: ~33.5 mm, n = 1. As for male, but broader, pink suffusions reduced, ground color more olive green than orange [possible artifact of photography]. *Forewing ventrum:* As for forewing dorsum but pinkish suffusions nearly absent, postmedial line absent, discal mark more pronounced. *Hindwing dorsum:* As for male but slightly broader, dark petiolate scales more numerous, especially antemedially. *Hindwing ventrum:* Following same pattern as forewing ventrum. *Abdomen:* As for male, but slightly broader. Sternite VIII with pair of short sclerotized bands posteriorly. *Genitalia:* (Figs 17, 18), n = 1. Papillae anales broad, somewhat rectangular, covered in setae, longer anteriorly. Apophyses anteriores with curved tips; apophyses posteriores robust, wide, slightly longer and broader than apophyses anteriores. Ductus bursae very long, somewhat sclerotized near ostium, ductus not differentiated from long, narrow corpus bursae. Appendix bursae present, elongate, fingerlike. Tergite VIII as narrow band with posteriorly directed mesal arc with membranous center. Lamella antevaginalis very large, broad, trapezoidal, notched mesally.



Figures 15, 16. *Auroriana* male genitalia, **a** ventral, **b** lateral, **c** phallus. **15** *A. florianensis*, French Guiana, Massif du Mitaraka, 300 m [St. Laurent diss.: 11-19-15:1] (RAS) **16** *A. gemma* holotype, Brazil, Santa Catarina, São Bento do Sul, Rio Natal, 550 m [C. Mielke diss.: 27.473] (DZUP). Scale bar = 1 mm.

Distribution (Fig. 19). *Auroriana florianensis* is so far known only from French Guiana, but is widely distributed in the region.

Remarks. At the time of its original description, *A. florianensis* was first suggested by Herbin (2012) to be unique among Mimallonidae and possibly belonging to a new genus. Despite this opinion, with which we agree, Herbin placed this species in the “catch-all” genus *Cicinnus*, which is currently being used to subsume many newly described taxa that show no obvious generic associations (Herbin 2012, 2015, Herbin and Mielke 2014, Herbin and Monzón 2015).

After reading the description of *Cicinnus florianensis*, the first author recognized that two undescribed species under study were congeneric with *C. florianensis*, but none of them belonged in *Cicinnus*. We further support Herbin’s (2012) original as-



Figure 17. *Auroriana florianensis* female genitalia. French Guiana, Réserve des Nouragues, Inselberg, 110 m [C. Gibeaux diss. prep: 7759] (MNHN). Photo courtesy of C. Gibeaux. Scale bar = 1 mm.

section that *C. florianensis* belonged to a new genus. This realization was solidified when genitalia dissections revealed that the characters previously thought wholly unique to *A. florianensis* were indeed shared with both *A. gemma* sp. n. and *A. colombiana*, and with no other described Mimallonidae.

Until now, *A. florianensis* was only known from the holotype male. The female and its genitalia are figured for the first time. Additional distributional and temporal data are presented for this species.

The apparent rarity of *A. florianensis* is noteworthy, especially considering that this species is known from only five males and one female, despite intensive collecting ef-

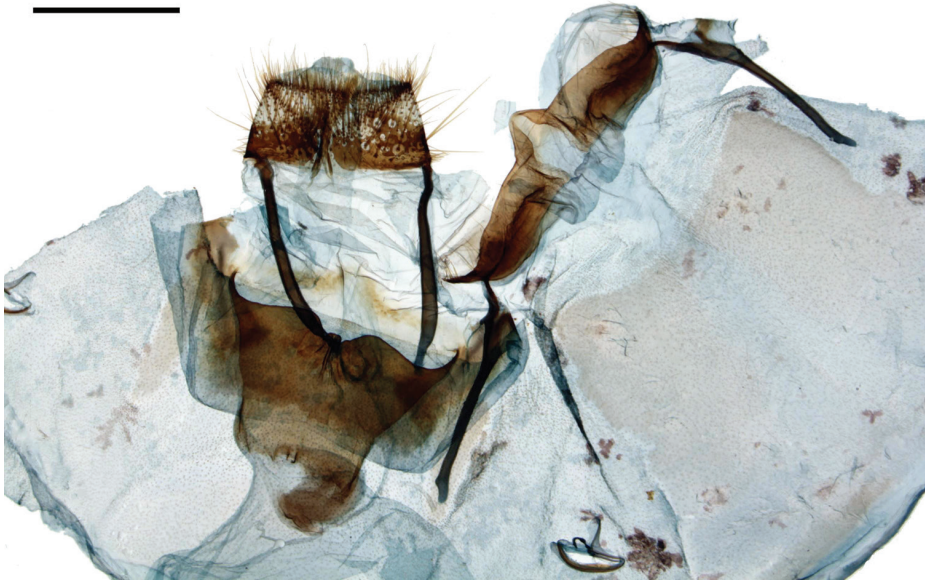


Figure 18. *Auroriana florianensis* female genitalia, detail. As for Figure 17. Scale bar = 1 mm.

forts in French Guiana. All specimens were collected in the dry season, the last days of July until late August, displaying a rather restricted flight period of less than a month. However, there are less collecting efforts during the dry season (C. G. Mielke pers. obs.), thus it is not possible to determine if the few records are due to this season being poorly collected or true rarity of the species.

***Auroriana gemma* St Laurent & C. Mielke, sp. n.**

<http://zoobank.org/CFB7093A-C529-439F-867C-3756B16ACB46>

Figs 12, 16, 19

Type material. **Holotype**, ♂: BRAZIL – SC [Santa Catarina], São Bento do Sul, Rio Natal, 550 m. XI.2013. A. Rank leg/ 27.473 Col. C. Mielke / C. Mielke diss.: 27.473/ DZ 32.729/ **HOLOTYPE** male *Auroriana gemma* St Laurent and C. Mielke, 2016 [handwritten red label]/ (DZUP). Type locality: Brazil: Santa Catarina: São Bento do Sul, Rio Natal.

Paratype, 1 ♂: **BRAZIL: Santa Catarina:** Neu Bremen [Dalbérgia]: III.1938, B. Pohl (MZSP). Paratype with the following yellow label: PARATYPE male *Auroriana gemma* St Laurent and C. Mielke, 2016.

Diagnosis. *Auroriana gemma* can be easily distinguished from its two congeners by the reduction of the pinkish suffusion postmedially and by the lighter medial and darker postmedial regions. Additionally, the forewing postmedial line is outwardly lined with a thicker pale-pink band than in other species in the genus. The hindwing

postmedial line in *A. gemma* is diffuse rather than thin and dark as in the other two species. The male genitalia also readily differentiate this species from the others. In *A. gemma* the tegumen is broader, gnathos processes shorter, somewhat wrinkled, and without a sharp tooth proximally. Additionally, the valves bear a unique, distinct, triangular mesal tooth and lack differentiated setae at the base of the valves. Finally, the phallus of *A. gemma* is much shorter than in either of the other *Auroriana* species.

Description. Male. *Head:* Light pink, eyes bordered posteriorly by dark scales; antenna opaque yellow. *Thorax:* Light tan brown, interspersed with pink scales, especially near wing base, prothorax pink. *Legs:* Light pink, but tarsus light brown. *Forewing dorsum:* Forewing length: 16 mm, avg. 16 mm, Wingspan: 30 mm, n = 2. Short, triangular, inward notch at tornus reduced, margin convex mesally. Ground color amber, orange brown, overall lightly speckled by dark petiolate scales. Antemedial line very faint pink. Postmedial line slightly curved, gray, outward edge lined with very pale pink, postmedial line angled sharply towards costa immediately after passing Rs₄, becoming diffuse pink suffusion. Antemedial area pink. Medial area lighter orange with pink suffusion. Postmedial area darker brown with pale pink-gray suffusion near wing margin; medial area lighter orange with pink suffusion; antemedial area pink. Discal spot a small dark-gray line across width of discal cell. *Forewing ventrum:* Similar to dorsum but with more speckling; ante- and postmedial lines absent; deeper orange overall with more expansive, defined pink suffusion postmedially. *Hindwing dorsum:* Coloration as for forewing dorsum, but less pink; postmedial line only as pale pink suffusion. *Hindwing ventrum:* Follows same pattern as forewing ventrum but more pink overall, orange coloration restricted to marginal region. *Abdomen:* As for genus, coloration as for thorax but more golden beige dorsally, pinker ventrally. *Genitalia:* (Fig. 16) n = 2. Tegumen broad, subtriangular. Vinculum irregular with paired, anterior process, rounded lobes present mesally below gnathos. Uncus short, tubular, hardly differentiated from tegumen. Gnathos as two unfused, heavily sclerotized, wrinkled, somewhat triangular processes. Triangular valves small relative to tegumen, simple, with sharp triangular mesal tooth. Juxta fused to phallus, with acute, pointed process. Phallus very short, simple, downturned, pointed with thin, sclerotized accessory extension terminally. Vesica short, bag-like, bulbous ejaculatoris twice length of phallus, bag-like. **Female.** Unknown.

Distribution (Fig. 19). *Auroriana gemma* is known from only two specimens: one at the type locality at 550 m in northeastern Santa Catarina state, Brazil, and the other from about 80 km farther south in the same state.

Etymology. This species is named for the amber (*gemma* Latin) ground color. The name is doubly appropriate because *gemma* also translates to gem, which refers to the beauty and rarity of this species.

Remarks. This new species is known from only two specimens, both of which surprisingly come from very well collected regions of southeastern Brazil (R. A. St. Laurent & C. G. Mielke pers. obs.). Given that one specimen was collected in March and the other in November, we cannot consider a short flight period as the reason for the apparent rarity of this species, as we mentioned for *A. florianensis*. All *Auroriana*



Figure 19. Distribution of *Auroriana*.

species are known from very few specimens, suggesting that *Auroriana* in general is not an easily collected genus or one that is merely overlooked by lepidopterists, as is much of the family.

***Micrallo* St Laurent & C. Mielke, gen. n.**

<http://zoobank.org/56E27AA5-B571-4CB7-8223-1DCD3AC7E381>

Type species. *Micrallo minutus* St Laurent & C. Mielke, sp. n.

Etymology. The genus name is based on the fact that the type species is remarkably small (*micro-* Latin), being one of the smallest in the family; + the ending *-llo*, which is shared with *Mimallo* Hübner, [1820], the type genus of Mimallonidae. The genus name is masculine.

Diagnosis. *Micrallo minutus* is immediately distinguished from all described Mimallonidae by the small size (forewing length: 11.5 mm) combined with the silvery-gray ground color and smooth wing margins. Genitalia are also unique in both sexes. Males have extremely complex genitalia with semi-membranous valves and lateral pockets encasing the valves. The pockets are filled with thick, elongate deciduous setae. Female genitalia have bent apophyses anteriores and long, robust apophyses poste-

riores that are connected to paired pouches that open on either side of the papillae anales. In the single examined female, one (the left hand pouch when viewed ventrally) of the two pouches was completely filled with elongate, thick setae that appear to originate from the male. *Cicinnus acuta* (Schaus, 1892) is superficially similar, but much larger and lacks the club-like valves and differentiated setae in lateral pouches of the male genitalia.

Description. Male. *Head:* Width roughly two thirds that of thorax, antenna bipectinate to tip; basal three rami pairs increasing in length from antennal base to roughly one fifth of antennal length where rami length is nearly constant for remainder of antenna until the final distal quarter, after which rami continuously decrease in length until terminus. Frons tan brown, labial palpus reduced, not extending beyond frons, second segment tufted, third segment highly reduced, indistinct. *Thorax:* Light gray, interspersed with darker petiolate scales, especially anteriorly. *Legs:* Gray brown, but tarsus darker brown, tibial spurs thin. *Forewing dorsum:* Forewing length: 11.5 mm, Wingspan: 23 mm, $n = 1$. Somewhat narrow, acutely triangular, apical third slightly concave, somewhat convex before tornus. Ground color light silvery gray, overall lightly speckled by dark petiolate scales. Antemedial line absent. Postmedial line light brown, angled outward from M_3 to Rs_3 , angled sharply towards costa immediately after passing Rs_4 , line straight from M_3 until posterior wing margin, outer edge of postmedial line lined with pale coloration, becoming darker at costa. Postmedial area darker graphite colored with pale pink-gray suffusion mesally. Discal mark black, oblique, somewhat ovoid in shape. *Forewing ventrum:* Similar to dorsum but browner medially, especially on costa; inner side of postmedial line with black suffusion, postmedial line more undulate than on dorsum. *Hindwing dorsum:* Coloration as for forewing dorsum, but browner, especially anteriorly; postmedial line more undulate than on forewing. *Hindwing ventrum:* Follows same pattern as forewing ventrum. Frenulum apparently absent or vestigial. *Venation:* Typical of Mimallonidae, very similar to *C. melsheimeri* but $Rs_3 + Rs_4$ much longer stalked. *Abdomen:* Short, extending slightly beyond hindwing tornus, almost tubular, coloration as for thorax but lighter gray ventrally. *Genitalia:* (Fig. 22), $n = 1$. Extremely complex; tegumen reduced, narrow, but with robust sclerotized margins. Vinculum circular, but heavily modified, variously connected to valves; pair of heavily sclerotized, curved, tusk-like structures originate near base of vinculum. Uncus reduced, bottle shaped. Gnathos as two unfused, outwardly sclerotized, inwardly membranous, elongated, tubular, mesally bent processes with two pairs of teeth mesally, two pairs basally. Valves highly modified, nearly absent, valves membranous proximal to vinculum, transitioning into club-shaped lobes distally, lobes extend nearly to apex of uncus. Base of valve with elongate, thick, deciduous, specialized setae, most of which contained in membranous fold laterally encasing valve-vinculum complex. Juxta partially fused to phallus, juxtal plate a reduced, roughly T-shaped structure embedded in anellus. Phallus simple, spade shaped (viewed dorsally), open dorsally, base with elongated, downward-angled process. **Female.** *Head:* As for male but antenna smaller overall with shorter rami. *Thorax:* As for male. *Legs:* As for male. *Forewing dorsum:* Forewing length: 11.5 mm,

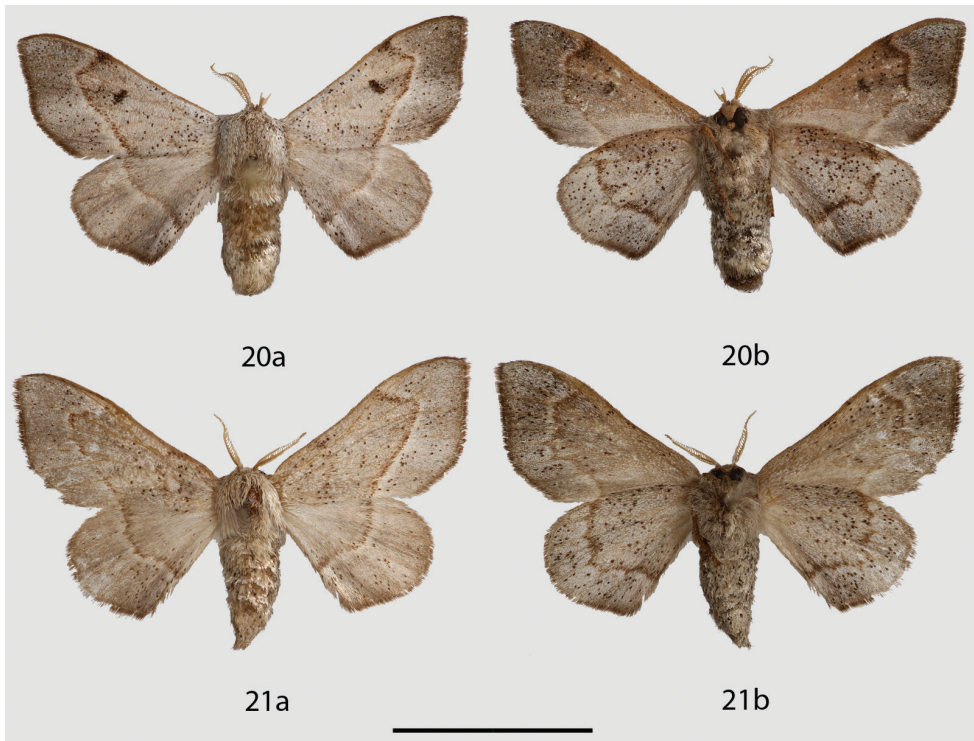


Figure 20, 21. *Micrallo minutus* adults, **a** dorsal, **b** ventral. **20** Holotype ♂, Brazil, Piauí, Oeiras, 200 m (DZUP) **21** Paratype ♀, Brazil, Piauí, Oeiras, 200 m (USNM). Scale bar = 1 cm.

Wingspan: 22 mm, n = 1. As for male but browner, discal spot absent. *Forewing ventrum*: Similar to dorsum but browner, especially anteriorly and medially; inner side of postmedial line with black suffusion, postmedial line more undulate than on dorsum. *Hindwing dorsum*: As for male but browner. *Hindwing ventrum*: Follows same pattern as forewing ventrum. Frenulum apparently absent or vestigial. *Abdomen*: As for male but slightly more robust; sternite VIII with quadrate U-shape formed by thin sclerotized band spanning width and length of segment. *Genitalia*: (Figs 23, 24), n = 1. Papillae anales small, rounded, covered in fine setae. Apophyses anteriores thin, distal third angled dorsally; apophyses posteriores, robust, nearly straight, twice length and width of apophyses anteriores. Ductus bursae same length as segment VIII, ostium unsclerotized. Corpus bursae bag-like, without any sclerotized structures. Dorsal sclerotization of tergite VIII complex, ridged, forming posteriorly directed point. Lamella antevaginalis a very large plate, nearly of equal width as segment VIII, with deep, wide, mesal indentation forming ostium. Pair of specialized, lateral pouches on either side of papillae anales, pouches connected to apophyses posteriores. Viewed ventrally, right pouch filled with elongated, deciduous setae, left pouch empty.

Remarks. An examination of *Cicinnus acuta* genitalia preparations (Franclemont genitalia prep. 1768 (CUIC) and St. Laurent diss.: 10-25-15:1) reveals minor simi-

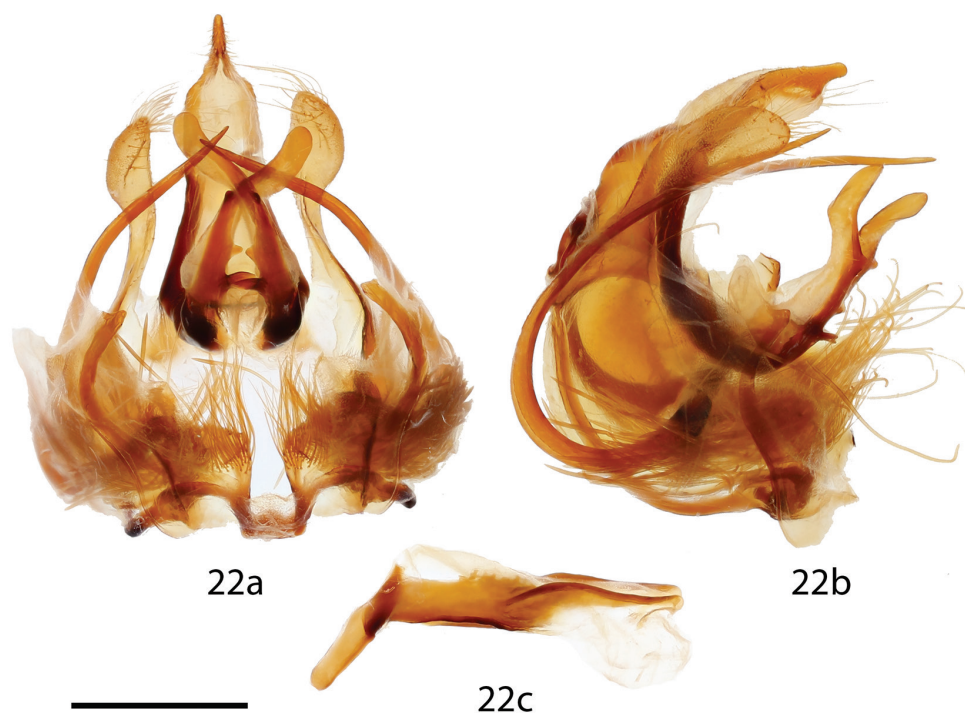


Figure 22. *Micrallo minutus* male genitalia, **a** ventral, **b** lateral, **c** phallus. Holotype ♂, Brazil, Piauí, Oeiras, 200 m [St. Laurent diss.: 10-21-15:1] (DZUP). Scale bar = 1 mm.

larities in the shape of the phallus, wherein both species the phallus is rather flattened and spade shaped when viewed dorsally. *Cicinnus acuta* differs, however, in all other respects of genitalia structure, namely in the presence of well-defined valves, much shorter gnathos processes, and in the presence of sharp, almost pincer-like structures at the base of the valves.

***Micrallo minutus* St Laurent & C. Mielke, sp. n.**

<http://zoobank.org/B04B1CB2-8C94-4323-AD47-13A6A4404975>

Figs 20–25

Type material. **Holotype**, ♂: BRASIL: PI [Piauí], Oeiras. 200 m, 12.iv.1994, V.O. Becker Col/ Col. Becker 92248/ USNM-Mimal: 2376/ St. Laurent diss.: 10-21-15:1/ DZ 32.730/ HOLOTYPE male *Micrallo minutus* St Laurent and C. Mielke, 2016 [handwritten red label]/ (ex. USNM, to be deposited in DZUP). Type locality: Brazil: Piauí: Oeiras.

Paratype, 1 ♀: **BRAZIL: Piauí:** Oeiras, 200 m: 12.IV.1994, V.O. Becker Col., Col. Becker 92248, USNM-Mimal: 2377, St. Laurent diss.: 10-21-15:2 (USNM).

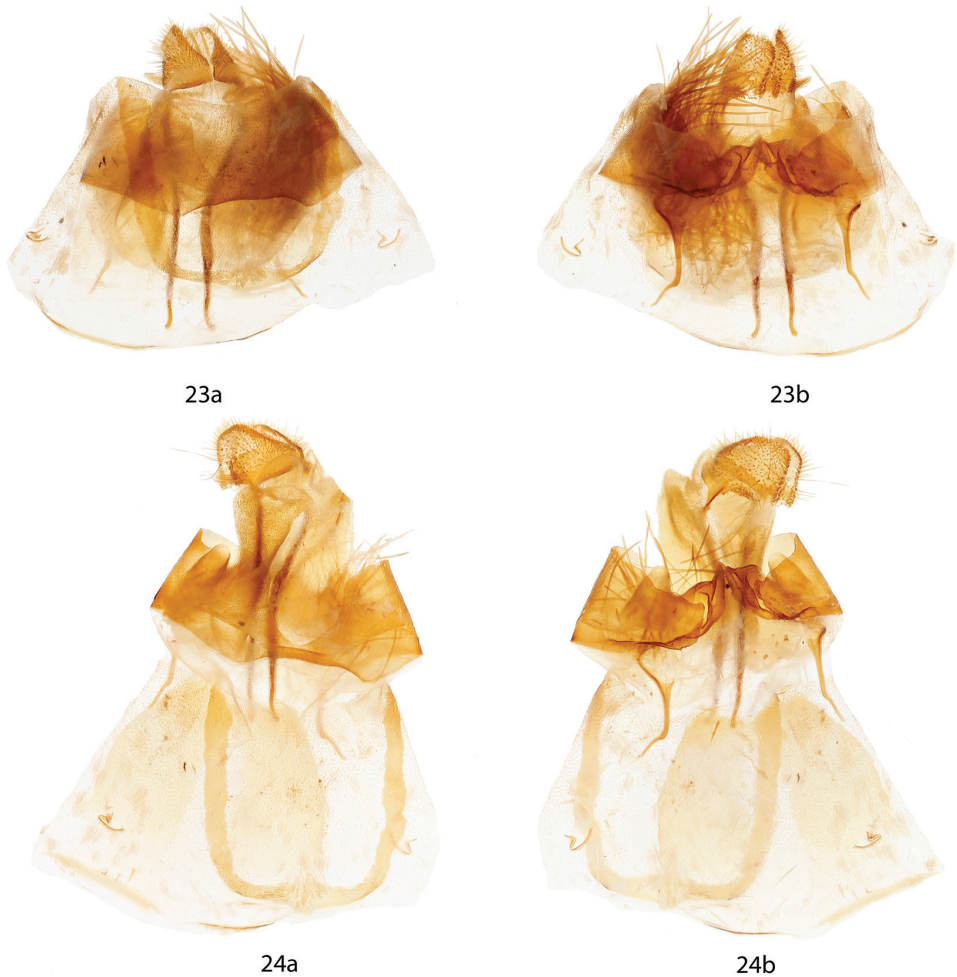


Figure 23, 24. *Micrallo minutus* female genitalia, **a** ventral, **b** dorsal. **23** Paratype ♀, Brazil, Piauí, Oeiras, 200 m, before extension [St. Laurent diss.: 10-21-15:2] (USNM) **24** As for Figure 23, after extension. Scale bar = 1 mm.

Paratype with the following yellow label: PARATYPE female *Micrallo minutus* St Laurent and C. Mielke, 2016.

Diagnosis. See genus diagnosis.

Description. See genus description.

Distribution (Fig. 25). The unique species in the genus *Micrallo* is so far known only from the type locality at Oeiras, Piauí, Brazil. This location is interesting because it lies on the edge of both Cerrado and Caatinga biomes (IBGE 2004).

Etymology. *Micrallo minutus* is named for its minute (*minutus* Latin) size, making it one of the smallest described species in the family.



Figure 25. Distribution of *Micrallo minutus*.

Remarks. Due to the interesting habitat at the type locality of *M. minutus*, it is not surprising that this taxon represents a previously undescribed species in a new genus. Both Caatinga and Cerrado biomes are incredibly undersampled (C. G. Mielke & R. A. St. Laurent pers. obs.), with the latter biome recently proven to support many new and endemic Mimallonidae species (Herbin and Mielke 2014). As of yet, no study has been published to determine the degree of Mimallonidae endemism to Caatinga.

The highly specialized genitalia in both sexes are unique among Mimallonidae. The complicated male genitalia bear two pouches on either side of the semi-membranous valves that contain many thick, specialized setae that are prone to falling out when the genitalia are examined. The female, likewise has rather complicated genitalia for female Mimallonidae, and also has specialized pouches, one on each lateral side of the papillae anales. In the single available female specimen, the right (when viewed ventrally) pouch contained an abundance of the same setae, the pouch was so completely filled that many of the setae are extending outside the pouch. See Fig. 23 for the genitalia before extension of the final abdominal segment. After extension (Fig. 24), most of the setae were expelled from the pouch. Close examination shows that the female genitalia are not asymmetrical, but that the second pouch was simply never filled with setae, or if it did contain setae at one time, they were completely lost before our examination. It is possible that these setae have characteristics pertinent to copulation, although we are unable to determine their

exact purpose. It is interesting that only one pouch was filled, suggesting that females of *M. minutus* are capable of mating two or more times. Perhaps even more intriguing is the fact that males may be able to fill only one female pouch at a time, despite possessing two complimentary pouches of their own on either side of the genital capsule.

Acknowledgements

We would like to thank John Brown (USNM) for access to specimens and facilitating the donation of the holotype of *Micrallo minutus* to DZUP. Alessandro Giusti at The Natural History Museum, London, provided access to important specimens and performed the necessary dissection and photography of the *Tostallo albescens* holotype. We would also like to thank Joël Minet (MNHN) for access to specimens and photographs of *Auroriana florianensis*, Philippe Collet (France) for photos of *A. florianensis* and the donation of a specimen to the first author for detailed examination, and Christian Gibeaux (France) for kindly preparing and photographing the female genitalia of *A. florianensis*. Marcelo Duarte (MZSP) and Rafael Dell'Erba facilitated the loan and examination of the *A. gemma* paratype. Ana P. S. Carvalho (University of Florida) helped name *A. gemma* and offered valuable suggestions to the manuscript. Jason Dombroskie (CUIC) provided access to the collection's camera equipment and stacking software, as well as helpful feedback on the manuscript. Finally, we would like to thank the Hunter R. Rawlings III Cornell Presidential Research Scholars program for providing funding to the first author for this and ongoing research with Mimallonidae.

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Taxonomic hypotheses regarding the genus *Gerbillus* (Rodentia, Muridae, Gerbillinae) based on molecular analyses of museum specimens

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Abstract

Methodological improvements now allow routine analyses of highly degraded DNA samples as found in museum specimens. Using these methods could be useful in studying such groups as rodents of the genus *Gerbillus* for which i) the taxonomy is still highly debated, ii) collection of fresh specimens may prove difficult. Here we address precise taxonomic questions using a small portion of the cytochrome *b* gene obtained from 45 dry skin/skull museum samples (from 1913 to 1974) originating from two African and three Asian countries. The specimens were labelled *G. gerbillus*, *G. andersoni*, *G. nanus*, *G. amoenus*, *G. perpallidus* and *G. pyramidum*, and molecular results mostly confirmed these assignments. The close relationship between *G. nanus* (Asian origin) and *G. amoenus* (African origin) confirmed that they represent vicariant sibling species which differentiated in allopatry on either side of the Red Sea. In the closely related *G. perpallidus* and *G. pyramidum*, specimens considered as belonging to one *G. pyramidum* subspecies (*G. p. floweri*) appeared closer to *G. perpallidus* suggesting that they (*G. p. floweri* and *G. perpallidus*) may represent a unique species, distributed on both sides of the Nile River, for which the correct name should be *G. floweri*. Furthermore, the three other *G. pyramidum* subspecies grouped together with no apparent genetic structure suggesting that they may not yet represent genetically differentiated lineages. This study confirms the importance of using these methods on museum samples, which can open new perspectives in this particular group as well as in other groups of interest.

† William Stanley passed away in October 2015. This paper is dedicated to his memory.

Keywords

Cytochrome b, degraded DNA, synonymy, systematics, vicariance

Introduction

DNA sequences have proven useful in taxonomic studies, and they now represent a primary source of information when it comes to the delimitation of species (Wiens 2007). Used in combination with other sources of data in the frame of integrative taxonomy (Dayrat 2005), they often provide convincing arguments for, or against the recognition of taxa as distinct species. DNA is especially useful in the case of cryptic species, where morphological criteria fail to unambiguously identify specific taxa (Knowlton 1986). For more than 25 years now, the recovery of DNA from ancient paleontological, archaeological and historic study specimens is routinely conducted (Pääbo 1989, Cooper 1994). The analysis of such DNA sequences has been instrumental in clarifying the systematics of extinct taxa, but it can also be of help in modern taxa that are difficult to sample today. This may happen when they are endangered or vulnerable in the wild, but also when their distribution interferes with sensible human activities and / or is situated in areas of conflict. In these cases, the use of museum specimens dating from periods and coming from areas where collection activities were easier could represent a convenient way of getting molecular information from particular species / populations. However, there are some limits to using such materials. The primary concern is that the sequences obtained are usually of relatively small size, due to the degraded state of the DNA of museum specimens. Given this limitation, the choice of the gene that will be targeted is of special concern: it has to be sufficiently variable to contain enough information, even in a small fragment, to allow distinguishing a particular species from its sister and other closely allied ones. Sequences as short as one hundred base pairs long have been shown to meet these requirements, for genes like cytochrome oxidase 1 (CO1) in insects, or cytochrome *b* (*cytb*) in rodents (Hajibabaei et al. 2006, Galan et al. 2012).

Gerbils of the genus *Gerbillus* represent a good example where such an approach can be expected to bring significant information. The systematics of this genus, as well as the one of the Gerbillinae subfamily to which it belongs, is still intensely debated, at various taxonomic levels (Chevret and Dobigny 2005, Abiadh et al. 2010, Alhajeri et al. 2015, Ndiaye et al. in review). At the specific level, many species still await confirmation of their taxonomic status, being based on very few specimens coming from localized areas (e.g. *G. agag*, *G. burtoni*, *G. grobbeni*, *G. jamesi*, *G. muriculus*, *G. principulus*, *G. syticus*, *G. vivax*, see Musser and Carleton 2005, Granjon 2013). A number of these areas are currently difficult to access due to political instability and regional insecurity, especially in the Saharo–Sahelian area where the majority of the *Gerbillus* diversity occurs (Brito et al. 2014). Conversely, some of these areas have been sampled quite extensively for rodents in the second half of the 20th century, and important collections have been gathered during this period. Examples include areas such as Sudan

(Setzer 1956), Libya (Ranck 1968), and Egypt (Osborn and Helmy 1980). The last-named country is of special interest to *Gerbillus* evolutionary history. First, it is located at the junction of Africa and Asia, the two continents over which the genus *Gerbillus* is distributed. Second, it is crossed by the Nile River that may represent a significant barrier to rodent, and especially gerbil, dispersal, thus promoting potential differentiation between species or subspecies. As the collection built by Osborn and Helmy (1980) proved to be especially rich in gerbilline rodent specimens, we tried to address the following questions on the basis of partial cytochrome *b* sequences obtained from a selected sample of *Gerbillus* museum specimens:

- Is the differentiation between *Gerbillus amoenus* (from Africa) and *Gerbillus nanus* (from Asia), recently evidenced by Ndiaye et al. (2013) based on complete sequences of *cytb*, found when using shorter sequences? If it is, then do the Egyptian specimens belong to *G. amoenus*, as should be the case? This question was addressed using museum specimens from Egypt, Pakistan and Afghanistan.
- What are the evolutionary relationships between various purported *G. pyramidum* subspecies and other *Gerbillus* species, such as *G. perpallidus*, *G. andersoni* and *G. gerbillus*?
- Do the subspecies of *Gerbillus pyramidum* listed by Osborn and Helmy (1980), based on morphological attributes (*G. p. elbaensis*, *G. p. floweri*, *G. p. gedeedus* and *G. p. pyramidum*), correspond to unique genetic clusters?

Material and methods

Forty-five tissue samples were obtained from dry fragments that were still present on the skulls and skins of *Gerbillus* specimens from Egypt and Asia, that are housed at the Field Museum of Natural History, Chicago, USA (Suppl. material 1). These samples represent six species and were labeled as: *G. amoenus amoenus* (N = 6), *G. andersoni andersoni* (N = 5), *G. gerbillus gerbillus* (N = 4), *G. nanus* (N = 6), *G. perpallidus* (N = 5) and *G. pyramidum* (N = 19). The latter was represented in our sample by the subspecies *G. p. elbaensis* (N = 5), *G. p. floweri* (N = 4), *G. p. gedeedus* (N = 5) and *G. p. pyramidum* (N = 5). These specimens were collected from 1913 to 1974 in Egypt (N = 38), Sudan (N = 1), Afghanistan, India and Pakistan (N = 2 for each; Suppl. material 1).

DNA was extracted in the Labex CeMEB degraded DNA platform (Montpellier, France) using the QiaAmp DNA micro kit (Qiagen). Due to the degradation of DNA in museum samples, we amplified a short fragment of *cytb* by designing two new primers named GERBCYTB-F2 (5'- GCA AAC GGA GCC TCA ATA TT - 3') and GERBCYTB-R3 (5'-CAT TCT ACR ATT GTT GGG CCA - 3'). These primers are respectively located at positions 250 and 488 of the *cytb* gene, delimiting a 239 base pair (bp) fragment. The 25µl reaction solution was prepared by mixing 14.5µl of DNase-RNase free water (Qiagen), 2.5µl of buffer (1X concentration), 2µl MgCl₂ (2mM), 2.5µl dNTP (250µM; Sigma), 0.5µl of each primer (0.5µM), 0.5µl

of AmpliTaq Gold (2.5 units; Applied biosystems). 1 µl and 2 µl of DNA aliquots of the extracted samples were amplified separately, and used for further comparisons. The *cytb* amplification was done at the CBGP molecular biology platform (Montferrier-sur-Lez, France) using PCR programs on a Master Cycle rep Gradient (eppendorf), including an activation step of 95 °C for 10 min followed by 55 cycles comprising a first denaturation at 94 °C for 30 s, hybridization at 50 °C for 30 s and elongation at 72 °C for 45s. The last step was a final extension at 72 °C for 7min. Three negative controls were used to check for contamination during DNA extraction, preparation of the mix and DNA distribution. In the first control (extraction control), no tissue was added to the tube; the second control (PCR mix control) was a closed tube, with only the PCR mix; the last control (DNA distribution control) was a tube with only the PCR mix, which was open during the entire process of DNA distribution, in order to check for the presence of DNA in the air. We verified the size and quality of each amplified DNA sequence fragment by performing an electrophoretic migration on a 2% agarose gel. The PCR products obtained at both DNA concentrations were sent to Eurofins MWG (Germany) for sequencing, and the results were compared among individuals to ensure that we obtained the same amplified sequence fragment.

The sequences were then checked, aligned and edited with BIOEDIT v.7.1.3.0 (Hall 1999) and we added 40 sequences downloaded from GenBank (www.ncbi.nlm.nih.gov/genbank) of various well-characterized *Gerbillus* species as a reference, and *Sekeetamys calurus* was used as an outgroup (see Suppl. material 1 for details). Phylogenetic reconstructions were carried out via Neighbor-Joining (NJ) and Bayesian inference (BI) using SEAVIEW v.4.2.12 (Gouy et al. 2010) and MRBAYES v.3.1.2 (Ronquist et al. 2012), with bootstrap values (BP) and posterior probabilities (PP) used as node support in respective analyses. The best fit models for Neighbor-Joining and Bayesian reconstructions were K2P and GTR+I+G, respectively. We tested it using jModeltest v2.1.4 (Darriba et al. 2012), with default settings (11 number of substitution schemes corresponding to 88 models to test, base frequencies and rate variation with 4 categories, ML optimized for the likelihood calculations). In the latter, two independently Markov chain Monte Carlo (MCMC) runs were carried out for one million generations each. Trees were sampled every 100 generations and convergence was reached when the average standard deviation of split frequencies remained under 0.01, thus reflecting the fact that the two tree samples become increasingly similar. Finally we applied a 25% burn-in. Pairwise Kimura 2-Parameter genetic distances were obtained for our *cytb* dataset under MEGA v6 (Tamura et al. 2013) with an associated standard error estimated based on 1000 bootstrap replicates. All codon positions were kept for analyses and no positions containing gaps and / or missing data were observed.

Results

No contamination was recorded during this series of experiments, as testified by examination of the content of the control tubes. DNA could not be amplified from only

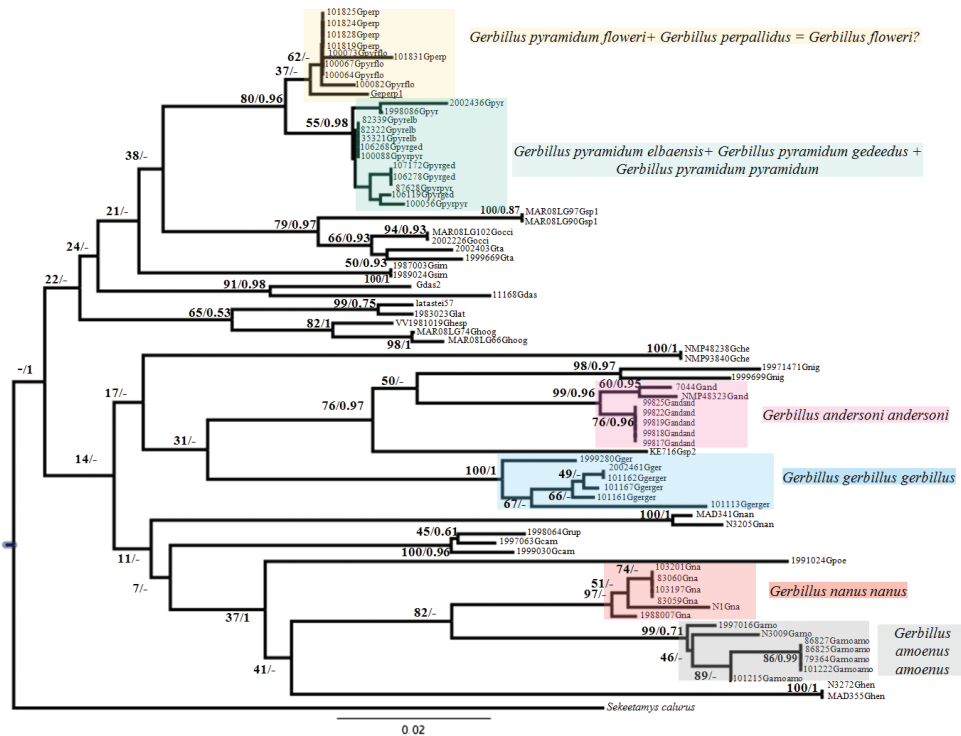


Figure 1. Phylogenetic reconstruction based on 239bp sequences of the cytochrome *b* gene using Neighbor-Joining. Values on nodes correspond to bootstraps / posterior probabilities respectively while “-” refer to places where both methods of reconstruction did not agree. Colored clades include museum specimens for which original sequences were obtained for the purpose of this study.

one of the samples. Additionally, a comparison of the results obtained using amplified PCR products from two DNA concentrations showed that the obtained sequences were identical in all but seven individuals, making us suspect the presence of nuclear copies of mitochondrial DNA (*Numt*). Unambiguous sequences (239 bp) of 37 individuals were obtained, to which we added the sequences of 40 “reference” individuals of different, well-characterized, *Gerbillus* species taken from GenBank and a representative of the outgroup *Sekeetamys calurus*, leading to a final *cytb* dataset comprising 77 individuals.

The phylogenetic tree (Fig. 1) obtained using NJ (with the bootstrap / posterior probability values indicated on nodes, respectively) presented here shows a similar topology to the BI tree with our museum specimens distributed in four main clades. The first two are unambiguously identified as *G. andersoni* (98/0.97) and *G. gerbillus* (99/1). The other two correspond to a *G. nanus* / *G. amoenus* (82/-), and a *G. perpallidus* / *G. pyramidum* (80/0.96) clade, respectively, each of which appears structured in two sub-clades. In the former, specimens referable to *G. nanus* from Asia (Pakistan and Afghanistan) cluster together (97/-) as a sister group to specimens referable to *G.*

Table 1. K2P genetic distances between and within (in italic) taxa based on *cyt b* sequences.

	<i>G. pyramidum floweri</i> + <i>G. perpallidus</i>	<i>G. pyramidum</i>	<i>G. gerbillus</i>	<i>G. amoenus</i>	<i>G. nanus</i>	<i>G. andersoni</i>
<i>G. pyramidum floweri</i> + <i>G. perpallidus</i>	0.004					
<i>G. pyramidum</i>	0.017	0.004				
<i>G. gerbillus</i>	0.105	0.109	0.013			
<i>G. amoenus</i>	0.135	0.139	0.146	0.011		
<i>G. nanus</i>	0.112	0.116	0.138	0.037	0.025	
<i>G. andersoni</i>	0.113	0.114	0.126	0.103	0.106	0.006

amoenus from Africa (Egypt, Niger, Mauritania; 99/0.71). The latter is divided into two sub-clades (separated by a K2P genetic distance = 0.017, see Table 1), one with *G. perpallidus* and *G. pyramidum floweri* (37/-; mean intragroup K2P distance = 0.004, Table 1), the other including the specimens attributed to the three other subspecies of *G. pyramidum* (i.e. *G. p. pyramidum*, *G. p. gedeedus* and *G. p. elbaensis*; 55/0.98; mean intragroup K2P distance = 0.004, Table 1).

Discussion

We removed 7 individuals from the analyses, for which minor differences between sequences obtained from the amplified DNA coming from the two different concentrations of extracted DNA were observed. This observation strongly suggests the presence of nuclear copies of the target sequence, as is sometimes recorded in gerbilline rodents (Dobigny 2002, Ndiaye 2013).

Overall, the labelling of the museum samples was remarkably in line with current taxonomy, and all corresponding specimens appear coherently placed in the phylogenetic tree produced (but see below for further details). This validates *a posteriori* the “mini-barcode” method used. Indeed, the sequences obtained, although short (239bp) made it possible to obtain generally robust reconstructions of the phylogenetic relationships between the study specimens, thus confirming the findings of Hajibabaei et al. (2006) and Galan et al. (2012) on the usefulness of small fragments in molecular taxonomy. This also means that the *ad hoc* primers designed for this particular experiment are well-adapted for a molecular barcoding approach based on potentially degraded DNA in the study group.

Museum specimens of *Gerbillus gerbillus* and *G. andersoni* from Egypt clustered unambiguously with “fresh” specimens of the same species from other origins. As for the distinction between Asian *G. nanus* and African *G. amoenus*, even if supported here by non-optimal BP values, it confirms the findings of Ndiaye et al. (2013) and the hypothesis of two vicariant sibling species that could have differentiated in allopatry on both sides of the Red Sea. The museum specimens used here were labelled as *G. amoenus* following Osborn and Helmy (1980) who treated them (= *Dipodillus amoenus*)

as a distinct species from *G. nanus* (= *D. nanus*). This taxonomy was not followed by many subsequent studies (see details in Ndiaye et al. 2013), but it now unambiguously appears that these two taxa have to be considered as distinct species. The inclusion of Egyptian specimens in the present study (clearly classified as *G. amoenus*) and the presence among the *G. nanus* sample of one reference specimen from Israel reinforce this conclusion, and confirmed that the Red Sea probably represents the geographical limit separating these two vicariant species.

The series of museum specimens of large-sized and hairy-footed gerbils referred to as *G. perpallidus* and *G. pyramidum* ssp. were distributed into two moderately well supported genetic clades: the first one includes all *G. pyramidum floweri* and *G. perpallidus* samples, together with a reference specimen of *G. perpallidus* (namely Gperp1, underlined in Fig. 1 and Suppl. material 1). The sequences of these 10 specimens show a very high degree of similarity. *Gerbillus perpallidus*, described by Setzer (1958) in Egypt, is currently considered as endemic to Egypt, where it is distributed in a relatively small area west of the Nile delta (Happold 2013). It is listed as a valid species by most recent authors (Osborn and Helmy 1980, Lay 1983, Pavlinov et al. 1990; Musser and Carleton 2005) whereas Cockrum (1977) and Petter (1975) considered it as synonymous with *G. latastei* and *G. pyramidum*, respectively. Osborn and Helmy (1980) considered *G. floweri* (Thomas, 1919) as a subspecies of *G. pyramidum*, but it is generally listed as a valid species following the review of Lay (1983). It also has a relatively small distribution range in Northern Egypt, east of the Nile delta and in most of the Sinai Peninsula (Happold 2013). Interestingly, Osborn and Helmy (1980: 114) insisted on the morphological similarity between the two taxa, both based on body and skull characteristics: “*Gerbillus p. floweri* and *G. perpallidus* are strikingly similar in color, bulla shape and, in some individuals, posterior margin of nasals”. Our results confirm these observations and suggest that these two taxa may represent a single species, the name of which should be *G. floweri*, according to his first description. This species would be characterized by a karyotype with a diploid number of chromosomes of $2n = 40$, and an autosomal fundamental number $NFa = 76$, as described in Lay et al. (1975) and Aniskin et al. (2006) under *G. perpallidus*. The distribution of this species would then encompass both sides of the Nile delta in Northern Egypt, and extend through the whole of Northern Sinai (Fig. 2). Its sister species would be *G. pyramidum*, which confirms what was found in other recent studies. The genetic distance between these taxa, as shown by this study, appears to be very small ($K2P = 0.017$). It was larger in two other studies based on complete *cytb* sequences of samples of different individuals of *G. pyramidum* and *G. perpallidus* only ($K2P$ distance = 0.029 and 0.031, respectively, in Ndiaye et al. 2012; in review).

In addition to *G. p. floweri*, Osborn and Helmy (1980) recognized three other subspecies in Egypt, namely *G. p. pyramidum*, *G. p. elbaensis* and *G. p. gedeedus*. The museum samples that refer to these three subspecies cluster together in our analyses (with two reference samples of *G. pyramidum*), but they were intermixed in this clade, suggesting that these subspecies do not represent genetic lineages with independent evolutionary histories, at least based on partial sequences of *cytb*. Their current allopat-

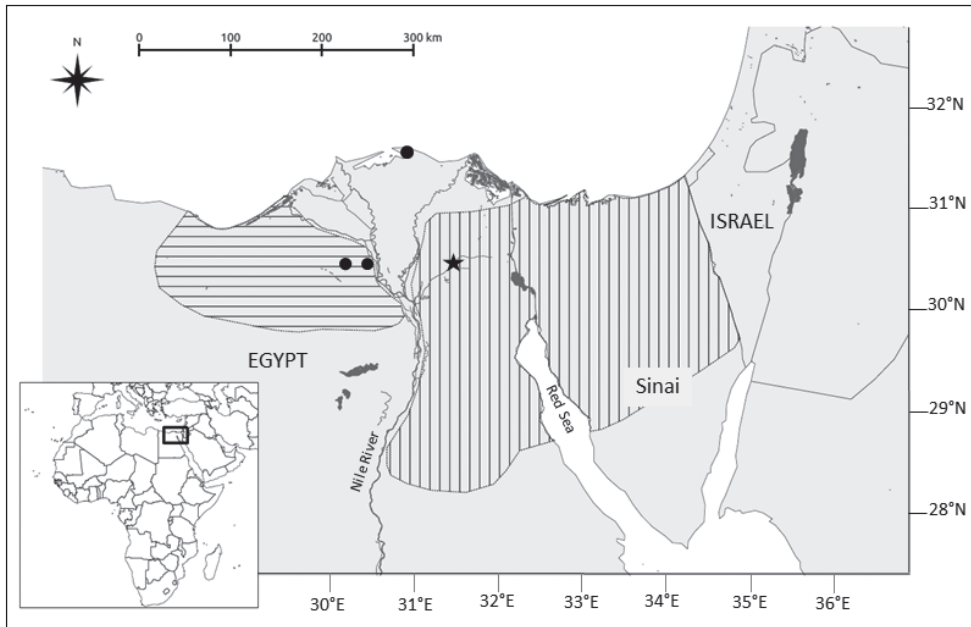


Figure 2. Reevaluated *Gerbillus floweri* distribution area based on the results presented here (see text; horizontal lines: *G. perpallidus*; vertical lines: *G. floweri* distributions according to the IUCN Red List of Threatened Species. Version 2015.2. www.iucnredlist.org). Black circles and black star: specimens of *G. perpallidus* and *G. p. floweri*, respectively, used in the present study.

ric distributions may be of relatively recent origin, following the last episode of drying of the Sahara, between 6,000 and 4,000 years ago (Holmes 2008). The geographic and / or adaptive morphological differentiation that has resulted from this separation may thus also be of recent origin, and the *cytb* fragments sequenced here may not reflect this differentiation yet.

In conclusion, we show here that molecular analysis of historic museum samples of the genus *Gerbillus*, up to more than 100 years after their collection, may give useful information, and address testable hypotheses, about the systematics of the genus. This could aid in the completion of the taxonomic understanding of this complex and speciose genus, which is well represented in museum collections worldwide. The new primers specifically designed here, may prove useful for this purpose.

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Supplementary material I

List of study specimens with identification numbers, geographic origin, collector name and collection date

Authors: Arame Ndiaye, Caroline Tatard, William Stanley, Laurent Granjon

Data type: Information of study specimens and cytb sequences

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