# Review of the genus Tinissa Walker, 1864 (Lepidoptera, Tineidae, Scardiinae) from China, with description of five new species 

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#### Abstract

The genus Tinissa Walker is reviewed for China. Seven species are recognized, of which T. apicimaculata $\mathbf{s p} . \mathbf{n} .$, T. conchata sp. n., T. connata sp. n., T. leguminella $\mathbf{s p}$. n. and T. spirella $\mathbf{s p}$. $\mathbf{n}$. are described as new; and T. insularia Robinson, 1976 is newly recorded from China. Photographs of the adults and illustrations of the genitalia are given. A key to all the known Chinese species and a distribution map of Tinissa in China are included.


## Keywords

Lepidoptera, Tineidae, Tinissa, new species, China

## Introduction

The genus Tinissa was established by Walker (1864) with T. torvella Walker, 1864 as the type species. It was once included in the subfamily Tinissinae, which was established by Gozmány and Vári (1973) for Tinissa and Leptozancla Meyrick, 1920. Robinson (1976) revised the Tinissinae on a worldwide basis, including 32 Tinissa species. Subse-
quently, Robinson (1981) described two more species from New Guinea and Borneo, and proposed a phylogenetic classification for Tinissa. Robinson (1986) synonymized Tinissinae with the subfamily Scardiinae. Following this treatment, Robinson and Tuck (1998) described one species from Brunei; and Robinson (2008) elevated the subspecies Tinissa torvella mysorensis Robinson, 1976 to species level. Currently, Tinissa comprises 36 named species, 14 of which are described from the Australian Region, 17 from the Oriental Region and five from the Afrotropical Region (Meyrick 1910, 1916, 1917, 1926, 1927, 1928, 1932; Gozmány and Vári 1973; Robinson 1976, 1981, 2008; Zagulajev 1972; Robinson and Tuck 1998).

Prior to this study, T. indica Robinson, 1976 was recorded in Taiwan (Robinson 1976), but none were recorded from Mainland China. The aim of the present paper is to review Tinissa in China, describe five new species, and record one species as new for the Chinese fauna. The distribution of Tinissa in China is shown by map (Fig. 1).

## Material and methods

The specimens examined were collected using light traps. Morphological terms in the descriptions follow Robinson (1976). Whole body dissections were carried out following the methods described by Lee and Brown (2006), and genitalia dissection and mounting methods follow Li (2002). Photographs of adults were taken with a Nikon D300, and the genitalia were photographed with an Olympus C7070WZ digital camera. All the studied specimens, including the types, are deposited in the Insect Collection, College of Life Sciences, Nankai University, Tianjin, China.

## Taxonomic accounts

## Tinissa Walker, 1864

http://species-id.net/wiki/Tinissa
Tinissa Walker, 1864: 780. Type-species: Tinissa torvella Walker, 1864: 780, by monotypy. Polymnestra Meyrick, 1927: 331. Type species: Polymnestra perilithas Meyrick, 1927: 331, by monotypy. [Synonymized by Gozmány and Vári 1973: 85.]

Generic characters. Medium to large tineid moths.
Head (Figs 2-3): Vertex and frons white of various shade, usually mixed with grayish brown scales (Fig. 2a). Compound eye large (Fig. 3a). Antenna about $0.5 \times$ length of forewing; scape with pecten more than 10 bristles; flagellum with elongate cilia (Fig. 3b). Labial palpus segmental ratio 1:2:2 (Fig. 3c), second segment with a few bristles and dense tuft, third segment upturned to about $60^{\circ}$ (Fig. 2b). Maxillary palpus (Fig. 3d) with three to five segments.


Figure I. Distribution of Chinese Tinissa spp. • T. indica Robinson, 1976 ■ T. insularia Robinson, 1976 © T. apicimaculata sp. n. $\star$ T. conchata sp. n. \& T. leguminella sp. n. $\downarrow$. connata sp. n. $\downarrow$. spirella sp. n.

Forewing (Figs 4-5): Index 0.25-0.30, somewhat rectangular, with termen slightly concave inward, or lanceovate, with termen obliquely blunt; color brown in general, with a purplish sheen, with scattered white dots (Fig. 4a); M absent or with weak trace in cell, $\mathrm{R}_{5}$ to costa or apex or just on to termen $\left(\mathrm{R}_{5}\right.$ to costa near apex in all the seven Chinese species), $\mathrm{R}_{4}$ and $\mathrm{R}_{5}$ separate, stalked or completely fused (Fig. 5); with elongate oval patch of small, flat, ovate scales on ventral surface between $A_{1+2}$ and dorsum (Fig. 4b). Hindwing (Figs 4-5) index 0.3-0.35, costa with distal half slightly concave; grayish brown, shining purplish; with a patch of rough, pale scales opposite forewing patch, anterior to $\mathrm{Sc}+\mathrm{R}_{1}$ (Fig. 4c); all veins present (Fig. 5), M stem usually present, branched or not, weak or conspicuous. Male frenulum with one slender bristle, female frenulum with two or three bristles (female frenulum with two bristles in all Chinese species). Legs yellowish white to white; mid tibia usually with a brown spot at base and two oblique brown bands on outer surface; hind leg elongate, tibia bearing large tufts of erect scales at apex.

Abdomen (Fig. 6): Second sternite elongate, twice length of other sternites, with sclerotized eyepatch-shaped trace on anterior half, with small tuft of forwarddirected setae medially (Fig. 6a); corema present or absent in eighth segment in male, eighth sternite with (Fig. 6b) or without (Fig. 6c) processes; corethrogyne present or absent in seventh segment in female. Male genitalia (Fig. 7) with tegumen and gnathos absent; uncus bilobate, being a pair of lobes of highly interspecific


Figures 2-3. Head structure of Tinissa indica Robinson, ${ }^{\text {on }} \mathbf{2}$ Head with scales: $\mathbf{2 a}$ dorsal view $\mathbf{2 b}$ lateral view $\mathbf{3}$ Head scales removed: $\mathbf{3} \mathbf{a}$ head, slide No. NKYLL012 3b antenna $\mathbf{3 c}$ labial palpus $\mathbf{3 d}$ maxillary palpus.
diversity attached to vinculum by membrane or fused with vinculum; subscaphium elongate and conspicuous, diagnostic at species level; saccus broad and triangular, or slightly narrow and rodlike; juxta large, closely appressed to valva, diversely


Figures 4-5. Wings of Tinissa indica Robinson, ${ }^{\lambda} .4$ Wings and patches: $\mathbf{4 a}$ wings $\mathbf{4 b}$ oval patch $4 \mathbf{c}$ rough scale patch 5 Venation, slide No. NKYLL012.
modified; valva usually short and conical; a pair of variously shaped processes arising from membrane between valva and juxta; labides present, dorsal to aedeagus, usually a pair of lobes, sometimes fused. Aedeagus of various shape and size, with or without carina, cornuti absent. Female genitalia with variously shaped eighth sternite, usually ventrally protuberant; ostium similarly diverse; antrum usually present, often divided by a narrow, oblique membranous ring at point of junction with ductus seminalis; ductus bursae usually with transverse, regular constrictions. Corpus bursae with or without signum.


Figures 6-7. Male abdomen of Tinissa indica Robinson. 6 Pregenital abdomen: 6a setae on second sternite 6b-6c eighth sternite $\mathbf{7}$ male genitalia ( $\mathbf{6 a} \mathbf{- b}, \mathbf{7}$ slide No. NKYLL012; $\mathbf{6 c}$ Tinissa connata sp. n., slide No. YLL11167).

Diagnosis. Members of Tinissa are medium to large sized robust tineid moths that can be easily recognized by having an elongate, brown-colored forewing with purple sheen and patterned with faint white dots; the hind legs are elongate and bear large tufts of erect scales at tibia apex; the male genitalia are characterized by the bilobate uncus, and the enlarged juxta being a pair of processes, forming a complex
with the valva. Tinissa shares the above characters with Leptozancla, but differs from Leptozancla by the antenna about $0.5 \times$ length of forewing, the forewing with elongate oval patch of small, flat, ovate scales, the hindwing with patch of rough, pale scales opposite to the forewing patch; the male genitalia with reduced valva, and the labides present as a pair of lobes. In Leptozancla, the antenna is as long as the forewing, the forewing lacks the ovate patch, the hindwing lacks the patch opposite to the forewing patch, the valva is absent, the labides are present as elongate, posteriorly directed spines in the male genitalia.

Biology. One species was reared from fungus on bamboo (Robinson 1976).
Distribution. Afrotropical, Oriental and Australian regions, as far east as the Solomon Islands, and as far south as Queensland (Robinson 1986).

Remarks. Robinson (1976) described that the maxillary palpus has three or five segments. Our whole body dissection of T. indica shows that the maxillary palpus has four segments in this species (Fig. 3d).

## Key to Chinese Tinissa species based on males

(Excluding T. apicimaculata sp. n. of which the male is unknown)
1 Wingspan $>15.0 \mathrm{~mm}$; first hind tarsal segment with smooth scales............ 2

- Wingspan $<15.0 \mathrm{~mm}$; first hind tarsal segment with rough, raised scales (Fig. 11)
T. connata sp. n.

2 Saccus broad, triangular; transtilla present; aedeagus stout, with carina....... 3

- Saccus elongate, rodlike; transtilla absent; aedeagus slender, without carina..... 5

3 Forewing scattered with conspicuous white spots throughout, arranged regularly along margins as well as between veins; transtilla inverted peach-shaped; aedeagus complete dorsally and ventrally .................................................... 4

- Forewing diffused with irregular ochreous white dots forming indistinct transverse striae (Fig. 8); transtilla being two small, sclerotized plates; aedeagus dorsally complete, ventrally with narrow slit from apex to middle (Fig. 14) T. insularia

4 Uncus lobe crescent; subscaphium triangular; juxta fist-shaped (Fig. 7) ........ T. indica

- Uncus lobe beanpod-shaped; subscaphium bifurcate posteriorly; juxta hornshaped (Fig. 17). $\qquad$ T. leguminella sp. n.

5 Uncus lobe ovate, with shallow pocket posterolaterally; subscaphium clubbed, with elongate setae posteriorly; juxta rectangular in basal 2/3, scallop-shaped in distal $1 / 3$; valva flask-shaped; aedeagus clubbed, straight, with a row of small spinules (Fig. 15) T. conchata sp. n .

- Uncus lobe L-shaped, with slender hornlike thorn at apex; subscaphium bulletlike, with slender fingerlike process on each side; juxta spiral; valva pyramidical; aedeagus needlelike, curved dorsad, without spinule (Fig. 18)...T. spirella sp. n.


Figures 8-13. Adults of Tinissa spp. 8 T. insularia Robinson, male 9 T. apicimaculata sp. n., holotype, female IO T. conchata sp. n., holotype, male II T. connata sp. n., holotype, male $\mathbf{I} 2$ T. leguminella sp. n., holotype, male 13 T. spirella sp. n., holotype, male.

## Key to Chinese Tinissa species based on females

(Excluding T. leguminella sp. n. and T. spirella sp. n., of which the females are unknown)
1 Wingspan $>15.0 \mathrm{~mm}$; first hind tarsal segment with rough, raised scales.... 2

- Wingspan < 15.0 mm ; first hind tarsal segment with smooth scales............ 3

2 Forewing with an ovate, large, blackish brown spot near apex (Fig. 9); corethrogyne present. $\qquad$ T. apicimaculata sp. n.

Forewing without spot near apex (Fig. 11); corethrogyne absent...................
$\quad$...................................................................................... T. connata sp. n.
3 Posterior margin of eighth tergite deeply concave at middle........................ 4
－Posterior margin of eighth tergite slightly sinuate（Fig．19）．．．．．．．．．．．．T．indica
4 Antrum almost as long as sternite，junction with ductus seminalis at 2／3； ductus bursae with conspicuously coarse transverse constrictions（Fig．21）．．． T．conchata sp． n ．
－Antrum shorter than sternite，junction with ductus seminalis at one－half， ductus bursae with very fine transverse constrictions（Robinson 1976： Fig．87）

Tinissa indica Robinson， 1976
http：／／species－id．net／wiki／Tinissa＿indica
Figs 1－7， 19
Tinissa indica Robinson，1976： 282.

Material examined．CHINA，Hainan Province： 1 q，Mt．Diaoluo（ $18^{\circ} 28^{\prime} \mathrm{N}, 109^{\circ} 31^{\prime} \mathrm{E}$ ）， 940 m，31．v．2007，leg．Zhiwei Zhang and Weichun Li，genitalia slide No．ZL09028； $1 \sigma^{\top}$ ，Mt．Yingge（ $19^{\circ} 01^{\prime} \mathrm{N}, 109^{\circ} 33^{\prime} \mathrm{E}$ ），30．ix．2011，leg．Bingbing Hu，genitalia slide No．YLL11135m，YLL11135w； $1 \widehat{J}^{\top}$ ，Nancha River，Mt．Bawang（ $19^{\circ} 04^{\prime} \mathrm{N}$ ， $109^{\circ} 02^{\prime} \mathrm{E}$ ）， 600 m，9．vi．2007，leg．Zhiwei Zhang and Weichun Li； 2 ふす，East first Protection Sta－ tion，Mt．Bawang， 650 m，7．iv．2008，leg．Bingbing Hu and Haiyan Bai，genitalia slide No．NKYLL012； $1 \jmath^{\top}$ ，East Administration，Mt．Bawang，8．v．2011，leg．Dandan Zhang and Lifeng Yang； $1 \delta^{\top}$ ，Mt．Wuzhi（ $18^{\circ} 46^{\prime} \mathrm{N}, 109^{\circ} 30^{\prime} \mathrm{E}$ ）， $650 \mathrm{~m}, 15 . \mathrm{v} .2007$ ，leg．Zhiwei Zhang and Weichun Li； $2 \widehat{\delta す}^{\star}$ ，Mt．Limu（ $19^{\circ} 09^{\prime} \mathrm{N}, 109^{\circ} 28^{\prime} \mathrm{E}$ ），5．v．2011，leg．Dandan Zhang and Lijun Yang．Yunnan Province： $1 \delta^{\lambda}$ ，Bubang（ $21^{\circ} 36^{\prime} \mathrm{N}, 101^{\circ} 35^{\prime} \mathrm{E}$ ），Mengla， 650 m，25．vii．2008，leg．Yingdang Ren，genitalia slide No．XYL05049．

Diagnosis．Adult（Figs 2－5）with wingspan 24．0－28．0 mm in male， 30.0 mm in fe－ male．Tinissa indica can be easily recognized from its congeners by the male genitalia hav－ ing a pair of crescent－shaped uncus lobes，the triangular subscaphium with wide，fingerlike process posterolaterally，and the fist－shaped juxta（Figs 6－7），and by the female genitalia having the eighth sternite ventrally protuberant and the hemispherical antrum（Fig．19）．

Distribution．China（Hainan，Yunnan，Taiwan），India，Sikkim，Bhutan．

## Tinissa insularia Robinson， 1976

http：／／species－id．net／wiki／Tinissa＿insularia
Figs 1，8， 14
Tinissa insularia Robinson，1976：285；Robinson，1981： 371.

Material examined．CHINA，Yunnan Province： $1 \delta^{\lambda}$ ，Baka Village，Menglun Town （ $21^{\circ} 56^{\prime} \mathrm{N}, 101^{\circ} 15 \mathrm{E}$ ），Mengla County， $620 \mathrm{~m}, 6$. VIII．2010，leg．Yinghui Sun and Lixia Li，genitalia slide No．YLL11138．


Figures 14-18. Male genitalia of Tinissa spp. 14 T. insularia Robinson, slide No. YLL11138 I5 T. conchata sp. n., paratype, slide No. YLL10196 16 T. connata sp. n., paratype, slide No. YLL11167 17 T. leguminella sp. n., paratype, slide No. XYL05048 18 T. spirella sp. n., holotype, slide No. XYL05050.

Diagnosis. Adult (Fig. 8) with male wingspan 17.0 mm . Tinissa insularia is close to T. spirella sp. n., but differs as noted in the description of the new species.

Distribution. China (Yunnan), Malaysia, Borneo, Indonesia (Sumatra, Java, Celebes, Moluccas), Philippines (Luzon, Mindanao, Palawan, Balabac, Tawi Tawi), New Guinea (Papua, Karkar I., New Britain), Solomon Is.

Notes. This species is recorded for the first time in China.

## Tinissa apicimaculata sp. n.

urn:Isid:zoobank.org:act:DA24EC3C-3D69-4107-8D07-79A9F1B8841A
http://species-id.net/wiki/Tinissa_apicimaculata
Figs 1, 9, 20
Type material. Holotype $q$ - CHINA, Guangxi Zhuang Autonomous Region: Jinxiu County ( $24^{\circ} 08^{\prime} \mathrm{N}, 110^{\circ} 11^{\prime} \mathrm{E}$ ), $550 \mathrm{~m}, 15$.IV. 2002, leg. Shulian Hao and Huaijun Xue, genitalia slide No. YLL11136.

Diagnosis. The new species is a distinctive species: the forewing markings are diagnostic, with an ovate, large, blackish brown spot near apex. It is small-sized as Tinissa connata sp. n., but can be easily recognized by the superficial characters.

Description. Adult (Fig. 9): Wingspan 12.0 mm in female. Vertex and frons creamy white. Antenna with scape and pecten white, pecten about 10-20 bristles; flagellum yellowish white. Labial palpus creamy white, second segment and tuft yellowish brown on outer surface. Thorax and tegula creamy white, anterior margin dark brown. Forewing index 0.27 , lanceovate, apex protruded roundly, termen oblique; ground color creamy white on basal $1 / 3$, yellowish brown on distal $2 / 3$, gradually darker to apex, shining copperish violet; patterned with indistinct, faint transverse striae, with large, blackish brown ovate spot near apex; $M$ absent in cell, $\mathrm{R}_{4}$ and $\mathrm{R}_{5}$ separated; fringe brown, pale yellowish brown at middle, forming a line parallel with pale yellowish brown basal line. Hindwing index 0.3; grayish brown, yellowish brown near apex, shining copperish violet; M stem conspicuous, not branched; fringe yellowish brown. Legs dominantly creamy white; fore femur with narrow, blackish brown spot on ventral surface, tibia blackish brown on outer surface, tarsus with fifth segment dark brown; mid tibia distally with oblique dark brown band on outer surface, spurs with oblique black band before apex, tarsus with middle portion of first and second segments dark brown, lighter on inner surface; hind tibial tuft creamy white on basal half, blackish brown on distal half, spurs with oblique blackish bands before apex, tarsus with basal half of first segment grayish brown, with rough, raised scales on dorsal surface, second and third segments blackish brown.

Male. Unknown.
Female genitalia (Fig. 20). Corethrogyne present in seventh segment. Eighth tergite rectangular, with sparse long setae on posterior margin; sternite hemicyclic, ventrally protuberant in short, tubular shape, with a pair of small, setose, mastoid processes at middle on posterior margin. Apophysis anterior $0.5 \times$ length of apophysis posterior. Ostium situated at middle of eighth sternite on posterior margin. Antrum absent; ductus bursae $3.5 \times$ length of corpus bursae, posterior $1 / 4$ narrow, anterior $3 / 4$ broader, $2.0 \times$ width of posterior $1 / 4$. Corpus bursae ovate, without signum.

Distribution. China (Guangxi).
Etymology. The specific name is derived from the Latin prefix apici- (= apex) and maculatus ( = macula), referring to the ovate, blackish brown spot near apex of forewing.

Remarks. In Tineidae, new species are usually described on the basis of male specimens. However, Tinissa is an exception. Among the 36 nominal species, seven are based


Figures 19-22. Female genitalia of Tinissa spp. 19 T. indica Robinson, slide No. ZL09028 20 T. apicimaculata sp. n., holotype, slide No. YLL11136 21 T. conchata sp. n., paratype, slide No. YLL11140 22 T. connata sp. n., paratype, slide No. YLL11137.
on both male and female， 14 on male and 15 on female only．As there is no sexual dimorphism in this genus，Tinissa apicimaculata $\mathrm{sp} . \mathrm{n}$ ．is distinctive from other species －most likely also in the male－by the forewing having an ovate blackish spot near apex．

## Tinissa conchata sp． $\mathbf{n}$ ．

urn：lsid：zoobank．org：act：1A49EDE3－1411－45DB－99AA－EF22FBA1201E http：／／species－id．net／wiki／Tinissa＿conchata
Figs 1，10，15， 21

Type material．Holotype $\widehat{\delta}$－CHINA，Fujian Province：Mt．Wuyi $\left(27^{\circ} 45^{\prime} \mathrm{N}\right.$ ， $118^{\circ} 02^{\prime} \mathrm{E}$ ）， $740 \mathrm{~m}, 25 . v i i .2008$ ，leg．Weichun Li，Yongling Sun and Haiyan Bai．Parat－ ypes： 2 ぶ $^{\text {ぶ，}}$ ，same data as holotype except dated 27．vii．2008．Guangdong Province： 1 ô，Nanling Mt．Babao（ $24^{\circ} 24^{\prime} \mathrm{N}, 113^{\circ} 08^{\prime} \mathrm{E}$ ）， $1070 \mathrm{~m}, 23 . v i i i .2010$ ，leg．Shulian Hao， genitalia slide No．YLL10196．Guangxi Zhuang Autonomous Region： 1 §，Qinmu－ cun，Yongfu County（ $24^{\circ} 58^{\prime} \mathrm{N}, 109^{\circ} 58^{\prime} \mathrm{E}$ ）， $160 \mathrm{~m}, 1 . \mathrm{v} .2008$ ，leg．Hui Zhen and Li Zhang； $1 \delta^{\top}$ ，Yachang Forest，Leye County（ $24^{\circ} 47^{\prime} \mathrm{N}, 106^{\circ} 33^{\prime} \mathrm{E}$ ）， 910 m ，28．vii．2004， leg．Jiasheng Xu，genitalia slide No．MYH11089； 1 \＆，Dongzhong Forest（ $21^{\circ} 37^{\prime}$ N， $107^{\circ} 32^{\prime} \mathrm{E}$ ），Fangchenggang City， 370 m ，9．iv．2002，leg．Shulian Hao and Huaijun Xue，genitalia slide No．YLL11140； $1 \delta^{\top}$ ，Hongqi Forest，Shangsi County（ $22^{\circ} 09^{\prime}$ N， $\left.107^{\circ} 59^{\prime} E\right), 260 \mathrm{~m}, 2 . I V .2002$ ，leg．Shulian Hao and Huaijun Xue．

Diagnosis．The new species is similar to T．amboinensis Robinson，1976，T． cinerascens Meyrick， 1910 and T．distracta Meyrick， 1916 in the forewing having fine transverse striae；the ovate uncus lobe with shallow pocket posterolaterally，the clubbed subscaphium setose posteriorly，the rodlike saccus expanded anteriorly，and the slender aedeagus in male genitalia；and the antrum divided by a membranous ring at point of junction with ductus seminalis in the female genitalia．However， the shapes of the valva，the juxta and the aedeagus in the male genitalia are differ－ ent among the four species：in the new species，the valva is flask－shaped，the juxta is rectangular basally and scallop－shaped distally，with dentation on posterior margin， and the aedeagus has small spinules；in T．amboinensis，the valva is triangular，the juxta is roundly lobe－shaped，with smooth margins，the aedeagus lacks the carina or spinule（Robinson 1976，Fig．46）；in T．cinerascens，the valva is triangular，the juxta is somewhat triangular，with smooth margins，the aedeagus lacks carina or spinule （Robinson 1976，Fig．48）；in T．distracta，the valva is triangular，the juxta has two small conical projections on inner surface，the aedeagus bears two small carinae be－ fore apex（Robinson 1976，Fig．45）．Besides，the structures of the eighth segment， the position of the membranous ring on the antrum and the shape of the antrum in the female genitalia are also different among the four species．

Description．Adult（Fig．10）：Wingspan $16.0-20.0 \mathrm{~mm}$ in male， 22.0 mm in fe－ male．Vertex yellowish white；frons with creamy white scales directed forward，tinged with dark brown scales laterally．Antenna with scape and pecten white，pecten more than 20 bristles；flagellum ochreous white，first segment dark brown above．Labial pal－
pus creamy white; second segment and tuft blackish brown above, mixed with black scales on outer surface and distal half of inner surface; third segment with oblique, blackish brown band near middle. Thorax and tegula creamy white, anterior margin dark brown. Forewing index 0.25 , subrectangular, apex bluntly rounded, termen oblique; ground color yellowish white to yellowish brown, shining bluish violet, scattered with unconspicuous, transverse, fine grayish brown striae, with large dark brown spot at distal $3 / 5$ and $2 / 3$ of costa as well as at basal $1 / 4$ near fold; $M$ absent in cell, $\mathrm{R}_{4}$ and $\mathrm{R}_{5}$ separated; fringe brown. Hindwing index 0.32 ; pale grayish brown, shining bluish violet, distal $1 / 5$ with faint, grayish brown striae; all veins present, M stem distinct in cell, branched at middle; fringe yellowish brown. Legs creamy white; fore femur with narrow, blackish brown spot on ventral surface, tibia blackish brown on outer surface, first segment of tarsus with large, blackish spot on outer surface, third and fourth segments mixed with dark brown; mid tibia with three broad oblique dark brown bands on outer surface, spurs with oblique dark brown band before apex, first to fourth segments of tarsus with large, blackish brown spot on outer surface; hind tibial tuft pale yellowish brown, forming two pale grayish brown clusters, blackish and shining purple at apex, tarsus and spurs of same pattern with mid leg.

Male genitalia (Fig. 15). Corema present; eighth sternite slightly convex at middle on posterior margin. Uncus lobe ovate, heavily sclerotized, with long setae on dorsal surface; apex pointed, setose, with shallow pocket posterolaterally. Subscaphium clubbed, $0.6 \times$ length of saccus, anterior $4 / 5$ smooth, posterior $1 / 5$ with elongate setae. Saccus elongate, rodlike, expanded anteriorly. Juxta heavily sclerotized, each lobe rectangular in basal $2 / 3$, scallop-shaped in distal $1 / 3$, with dentation and fine setae on posterior margin. Valva flask-shaped, basal $3 / 5$ about $2.0 \times$ width of distal $2 / 5$; process from membrane between valva and juxta papillary, with short setae on posterior margin. Transtilla absent. Labides with anterior $4 / 5$ slender, posterior $1 / 5$ umbrellashaped. Aedeagus slender, clubbed, $1.4 \times$ length of saccus, straight, pointed at apex, complete ventrally and dorsally, with a row of 6 or 7 small spinules.

Female genitalia (Fig. 21). Corethrogyne present in seventh segment. Eighth tergite shield-shaped, with sparse short setae, lateral margins slightly concave at anterior $1 / 3$, anterior margin concave at middle, anterolateral side protruding triangularly, posterior margin incised, deeply concave at middle, forming a pair of small papillary lateral processes; sternite tapered, posterior margin concave at middle, forming two overlapped plates, between two plates with a tubular chunnel. Ostium at middle of middle concavity on posterior margin of eighth sternite. Apophysis anterior $0.3 \times$ length of apophysis posterior. Antrum heavily sclerotized, divided at posterior $2 / 3$ by a narrow, oblique membranous ring at point of junction with ductus seminalis, posterior $2 / 3$ broader and less sclerotized than anterior $1 / 3$; ductus bursae pipe-shaped, with coarse, transverse, regular constrictions, posterior $4 / 5$ slightly sclerotized, anterior $1 / 5$ slightly expanded and incurvate. Corpus bursae as long as ductus bursae, without signum.

Distribution. China (Fujian, Guangdong, Guangxi).
Etymology. The specific name is derived from the Latin conchatus (= conchoidal), referring to the scallop-shaped distal half of the juxta.

## Tinissa connata sp. n.

urn:lsid:zoobank.org:act:F528F4DB-C10C-470B-A2DA-B95CD21E454B
http://species-id.net/wiki/Tinissa_connata
Figs 1, 6c, 11, 16, 22
Type material. Holotype $\boldsymbol{o}^{\lambda}$ - CHINA, Fujian Province: Guadun ( $27^{\circ} 44^{\prime} \mathrm{N}$, $117^{\circ} 38^{\prime} \mathrm{E}$ ), Mt. Wuyi, $1100 \mathrm{~m}, 29 . \mathrm{VII} .2008$, leg. Weichun Li, Yongling Sun and Haiyan Bai, genitalia slide No. XYL08114. Paratypes: Guangdong Province: $1 \jmath^{\lambda}$, Nanling, Shaoguan City ( $24^{\circ} 48^{\prime} \mathrm{N}, 113^{\circ} 35^{\prime} \mathrm{E}$ ), 22-28.VI.2008, leg. Liusheng Chen, genitalia slide No. YLL11167. Guangxi Zhuang Autonomous Region: 1 \& , Mt. Daming ( $23^{\circ} 09^{\prime} \mathrm{N}, 108^{\circ} 16^{\prime} \mathrm{E}$ ), Nanning City, 1200 m , 5.VIII.2011, leg. Shulian Hao and Yinghui Sun, genitalia slide No. YLL11137.

Diagnosis. The new species can be easily recognized by the small-sized body and the rough, raised scales on the first hind tarsal segment. These characters are also present in T. apicimaculata sp. n., but the forewing of T. apicimaculata sp. n. is darker in color, and has an ovate blackish brown spot near apex that is absent in T. connata sp. n.

Description. Adult (Fig. 11): Wingspan 12.5 mm . Vertex and frons ochreous white, tinged with grayish brown around antenna. Antenna with scape and pecten ochreous white, pecten more than 20 bristles; flagellum yellowish brown, first segment with blackish brown above. Labial palpus white, second segment and tuft blackish brown above, mixed with blackish brown on distal half of inner surface and on outer surface, third segment with oblique, blackish brown band before apex. Thorax and tegula white, anterior margin dark brown. Forewing index 0.26 , lanceolate, apex protruded triangularly; ground color pale yellowish brown, gradually darker from base to apex, shining bluish violet, with scattered dark brown dots throughout, concentrated from costal $1 / 4$ to dorsal $1 / 6$, forming a discontinuous oblique stria; costal margin with conspicuous creamy white dot at distal $1 / 3$ and $1 / 4 ; \mathrm{M}$ with weak trace in cell, $\mathrm{R}_{4}$ and $\mathrm{R}_{5}$ separated; fringe brown. Hindwing index 0.32 ; yellowish brown, shining bluish violet; M stem conspicuous in cell, branched at middle; fringe pale yellowish brown. Legs ochreous white; fore coxa and femur mixed with brown on ventral surface, tibia blackish brown ventrally, tarsus with first segment on outer surface as well as fourth and fifth segments blackish brown; mid tibia with blackish brown spot at base, with a narrow, blackish brown band at middle, with an oblique dark brown band before apex, spurs with oblique dark band before apex, first segment of tarsus with blackish brown spot at base and middle, fourth segment dark brown; hind tibial tuft pale yellowish brown, distal scales with dark brown tips, tarsus with first segment dark brown on distal half, bearing rough, raised scales on dorsal surface, second segment with small blackish brown dot near base dorsally, fourth segment dark brown.

Male genitalia (Fig. 16). Corema present; eighth sternite straight on posterior margin. Uncus lobe deeply emarginated posteriorly, forming two processes: inner process short, bears stub spiculas; outer process $2.5 \times$ length of inner one, with spiculas varying in length, pectinated; with a large rounded flap arising from near posterior margin. Subscaphium cheliform, anterior $3 / 4$ slender, slightly curved ventrad, poste-
rior $1 / 3$ broadened, with deeply U-shaped concavity at middle on posterior margin. Saccus broad triangular, narrowed anteriorly, rounded apically. Juxta sclerotized, each lobe triangular, pointed at apex, serrate along inner margin. Valva fully fused with juxta; process from membrane between valva and juxta foliole-shaped, bearing a small apical process and some short setae before apex. Transtilla trapezoidal, fully fused with labides. Labides peach-shaped. Aedeagus stout, as long as saccus, broadened to flared apex, slightly curved ventrad, ventrally complete, with large, triangular flap at base, dorsally with a deep cleft, without carina or spinule.

Female genitalia (Fig. 22). Corethrogyne absent, with deciduous setae on posterior margin of seventh segment. Eighth tergite semicircularly concave on anterior margin, and forming rounded plate posterolaterally, with a few short setae on posterior margin; eighth sternite semicircularly concave on anterior margin, forming semicylindrical plate anterolaterally extending to tergite, attached with apophysis posterior, setose and slightly concave at middle on posterior margin, with ovate ventral protuberance on ventral margin of ostium. Apophysis anterior $0.3 \times$ length of apophysis posterior. Antrum short, divided at posterior $2 / 3$ by a membranous ring at point of junction with ductus seminalis on right; ductus bursae $3.0 \times$ length of corpus bursae, posterior $1 / 5$ with transverse constrictions, anterior $4 / 5$ smooth. Corpus bursae pear-shaped, with large, heavily sclerotized, inverted funnel-shaped signum.

Distribution. China (Fujian, Guangdong, Guangxi).
Etymology. The specific name is derived from the Latin connatus (= connate), referring to the fusion of the juxta and valva.

Remarks. The single female specimen was not collected in the same locality as the male specimens, but it has the same forewing patterns as the males do, and its first hind tarsal segment has the rough, raised scales, which are characteristic for this species. The rough, raised scales on the first hind tarsal segment are also present in T. apicimaculata sp. n. and T. spaniastra Meyrick, 1932, but the forewing of T. apicimaculata sp. n. has an ovate blackish brown spot near apex, and the members of T. spaniastra are larger in size ( 25 mm in male, $20-27 \mathrm{~mm}$ in female (Robinson 1976)). In addition, the genitalia structures of the above mentioned species are also different.

## Tinissa leguminella sp. n .

urn:lsid:zoobank.org:act:D9425924-508D-475F-B2A6-A21B746E84DC
http://species-id.net/wiki/Tinissa_leguminella
Figs 1, 12, 17

Type material. Holotype $\widehat{\diamond}$ - CHINA, Yunnan Province: Rare Botanical Garden, Ruili ( $24^{\circ} 00^{\prime} \mathrm{N}, 97^{\circ} 50^{\prime} \mathrm{E}$ ), 1000 m , 5.VIII. 2005, leg. Yingdang Ren, genitalia slide No. YLL11139. Paratype: $\widehat{J}^{\top}$, same data as holotype except dated 7.VIII.2005, genitalia slide No. XYL05048.

Diagnosis. Tinissa leguminella sp. n. is similar to T. indica in having a similar forewing pattern, a broad and triangular saccus and a short and conical valva in the
male genitalia. However, the new species can be recognized from the latter by the beanpod-shaped uncus lobe, the bifurcate subscaphium, the horn-shaped juxta, the process from the membrane between the valva and the juxta with basal $3 / 5$ nearly parallel dorso-ventrally, widended at distal $2 / 5$, then narrowed to melanised and setose apex, and the aedeagus with short carina in the male genitalia. In T. indica, the uncus lobe is crescent, the subscaphium is triangular, the juxta is fist-shaped, the process from the membrane between the valva and the juxta is fingerlike, and the aedeagus has long carina in the male genitalia.

Description. Adult (Fig. 12): Male wingspan 16.5-19.0 mm. Vertex ochreous yellow, tinged with blackish brown near eyes; frons ochreous yellow, with blackish brown scales laterally. Antenna with scape and pecten ochreous white, pecten more than 20 bristles; flagellum yellowish brown, first two segments blackish brown above. Labial palpus creamy white; second segment brown on outer surface, mixed with creamy white at middle and apex, tuft black; third segment with dark brown spot at base and distal $1 / 3$ on outer surface. Thorax ochreous white, posterior $1 / 3$ grayish brown; tegula creamy white, anterior $1 / 3$ dark brown, posterior $1 / 3$ mixed with yellowish brown. Forewing index 0.25 , rectangular, apex protruded triangularly, termen slightly concave inward at about anterior $1 / 3$; ground color brown, shining dark purplish, scattered with conspicuous white spots throughout, regularly arranged along margins as well as between veins, more white spots concentrated in basal $1 / 5$, near fold and at upper angle of cell; $M$ absent in cell, $R_{4}$ and $R_{5}$ separated; fringe brown. Hindwing index 0.32; pale grayish brown, shining dark purplish, with small pale dots apically; M stem conspicuous in cell, branched at middle; fringe yellowish brown. Fore leg yellowish brown, femur with narrow, dark brown spot on ventral surface, tibia blackish brown, tarsus blackish brown except apex of first segment as well as fifth segment ochreous white; mid leg ochreous yellow, tibia with three oblique, blackish brown bands on outer surface, broader near apex, shorter spur with oblique blackish brown band on outer surface, longer spur yellowish brown on outer surface, tarsus with first segment dark brown at base and middle, third and fourth segments dark brown; hind leg pale yellowish brown, tibia ochreous white at basal $2 / 3$ ventrally, tuft dark grayish brown, forming two clusters at spurs, spurs ochreous white ventrally, dark brown dorsally but yellowish at apex, first segment of tarsus ochreous white ventrally, with dark brown spot at base on outer surface, with large, dark brown spot from basal $1 / 3$ to before apex on outer surface, other tarsal segments yellowish brown ventrally, third and fourth segments blackish brown dorsally.

Male genitalia (Fig. 17). Corema present; eighth sternite straight on posterior margin. Uncus lobe beanpod-shaped, hornlike and heavily sclerotized, sparsely setose on distal half, with shallow pocket distally. Subscaphium fused anteriorly, bifurcate from $1 / 4$, forming long band-shaped lobe on each side, gradually narrowed to blunt apex. Saccus broad triangular. Juxta heavily sclerotized, each lobe stout, narrow basally, dilated distally; apex straight, setose, and melanised; basally fused and protruded ventrad, forming a plate with a vertical ridge at middle. Valva short, conical, apex narrowly rounded, with long distal setae; process from membrane between valva and juxta
with basal $3 / 5$ nearly parallel dorso-ventrally, widended at distal $2 / 5$, then narrowed to melanised and setose apex. Transtilla broad, inverted peach-shaped. Labides concave at middle on posterior margin, with mastoid process posterolaterally. Aedeagus stout, clubbed, $1.5 \times$ length of saccus, gently curved dorsad, complete dorsally and ventrally, with a short carina arising from distal $1 / 4$ ventrally.

Female. Unknown.
Distribution. China (Yunnan).
Etymology. The specific name is derived from the Latin legumin- (= legume) and the postfix-ellus, referring to the beanpod-shaped uncus lobe.

## Tinissa spirella sp. n. <br> urn:lsid:zoobank.org:act:151565EB-709E-4628-AAD4-81CCDE40A961 <br> http://species-id.net/wiki/Tinissa_spirella

Figs 1, 13, 18

Type material: Holotype $\begin{gathered} \\ \text { - CHINA, Sichuan Province: Wolong Nature Re- }\end{gathered}$ serves $\left(31^{\circ} 01^{\prime} \mathrm{N}, 103^{\circ} 10^{\prime} \mathrm{E}\right), 1900 \mathrm{~m}, 7 . \mathrm{VIII} .2004$, leg. Yingdang Ren, genitalia slide No. XYL05050.

Diagnosis. The new species is similar to T. conchata sp. n. by having an elongate, rodlike saccus and a slender aedeagus. It can be recognized from the latter by the Lshaped uncus lobe with an apical thorn, the subovate subscaphium with slender fingerlike process on each side, the spiral juxta, the pyramidical valva, and the needlelike aedeagus curved dorsad, without spinule. In T. conchata sp. n., the uncus lobe is ovate, with shallow pocket posterolaterally; the subscaphium is clubbed, with elongate setae posteriorly, the juxta is rectangular in basal $2 / 3$ and scallop-shaped in distal $1 / 3$, the valva is flask-shaped, and the clubbed aedeagus is straight, with a row of small spinules.

Description. Adult (Fig. 13): Male wingspan 20.0 mm . Vertex ochreous yellow, posterior margin ochreous white; frons ochreous white, tinged with ochreous yellow. Antenna with scape and pecten ochreous white, pecten about 10-20 bristles; flagellum pale yellowish brown, first segment blackish brown above. Labial palpus with first and second segments creamy white on inner surface, dark brown mixed yellowish brown on outer surface; third segment ochreous yellow, with yellowish brown spot at base and before apex on outer surface. Thorax creamy white mixed with dark grayish brown scales; tegula with anterior half dark grayish brown, posterior half creamy white but pale gray at middle. Forewing index 0.27 , rectangular, apex protruded triangularly, termen slightly concave inward at about $2 / 5$; yellowish brown mixed with grayish brown, shining bluish violet, with scattered faint white dots, large and conspicuous at base, along costa, termen and dorsum; M absent in cell, $\mathrm{R}_{4}$ and $\mathrm{R}_{5}$ separated. Hindwing index 0.32; grayish brown, shining bluish violet; $M$ stem conspicuous in cell, branched at middle; fringe pale grayish brown. Legs yellowish brown; fore femur dark brown on ventral surface, tarsus with first segment blackish brown on outer surface, with faint dark brown spots on outer surface of second to fifth segments; mid tibia with blackish
brown spot at base, with one narrow, blackish brown band at middle, with one broad, oblique, dark brown band before apex on outer surface, tarsus with small, blackish brown spot at base and middle on outer surface of first segment; hind tibial tuft pale yellowish brown, blackish brown before apex on outer surface, tarsus with first segment dark brown at base and apex on dorsal surface, with long, fine scales dorsally, third segment dark brown dorsally.

Male genitalia (Fig. 18). Corema present; eighth sternite straight on posterior margin. Uncus lobe L-shaped, widely spaced to each other, completely fused with vinculum; distal half setose dorsally, with a large apical thorn. Subscaphium bulletlike, with a pair of small and narrow triangular protuberances at middle on anterior margin, with a slender fingerlike process arising from anterior $2 / 5$ of each side, $0.5 \times$ length of subscaphium. Saccus rodlike. Juxta heavily sclerotized, each lobe spiral, with long setae ventrally, fused on inner margin. Valva short, heavily sclerotized, more or less triangular, apex pointed, dorsal margin with long setae; process from membrane between valva and juxta mastoid, with short setae at apex. Transtilla absent. Labides small, triangular, heavily sclerotized. Aedeagus very slender, needlelike, $1.2 \times$ length of saccus, curved dorsad, complete dorsally and ventrally, roundly protruded at base ventrally, without carina or spinule.

Female. Unknown.
Distribution. China (Sichuan).
Etymology. The specific name is derived from the Latin spirellus (= spiral), referring to the small, spiral, whorl-shaped juxta.

## Discussion

The genus Tinissa is unique among the 30 genera of the subfamily Scardiinae. It is highly diagnostic by the hind legs bearing large tufts of erect scales at apex of the tibia. Based on this character, we assign five new species to this genus.

As all the nominal species were well described and illustrated by previous taxonomists (Robinson 1976, 1981; Robinson and Tuck 1998; Gozmány and Vári 1973; Zagulajev 1972), we examined only the holotypes of T. araucariae Robinson, 1976, T. bakeri Robinson, 1976, T. insignis Zagulajev, 1972, T. parallela Robinson, 1976, T. polysema Zagulajev, 1972) as well as the paratype of T. insularia Robinson, 1976 that were available during this study. Besides, the shape of valva + juxta complex and the uncus lobes in the male genitalia as well as the eighth sternite and the ostium in the female genitalia are highly diverse among species, which makes the species identification more effective.

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## References

Gozmány LA, Vári L (1973) The Tineidae of the Ethiopian Region. Transvaal Museum Memoir, Pretoria 18: i-vi, 1-238, figs 1-570.
Lee S, Brown RL (2006) A new method for preparing slide mounts of whole bodies of Microlepidoptera. Journal of Asia-Pacific Entomology 9(3), 249-253. doi: 10.1016/S1226-8615(08)60299-X
Li HH (2002) The Gelechiidae of China (I) (Lepidoptera: Gelechioidea). Nankai University Press, Tianjin, 504 pp .
Meyrick E (1910) Descriptions of Microlepidoptera from Mauritius and the Chagos Isles. Transactions of the Entomological Society of London 1910: 430-478.
Meyrick E (1916) Exotic Microlepidoptera 1: 609-640.
Meyrick E (1917) Exotic Microlepidoptera 2: 65-96.
Meyrick E (1920) Microlepidoptera. In: Alluaud C, Jeannel R, Voyage de Ch. Alluaud et R. Jeannel en Afrique Orientale (1911-1912). Resultats scientifiques. Insectes Lépidoptères. 2: 33-120.
Meyrick E (1926) Exotic Microlepidoptera 3: 289-320.
Meyrick E (1927) Exotic Microlepidoptera 3: 321-352.
Meyrick E (1928) Exotic Microlepidoptera 3: 417-448.
Meyrick E (1932) Entomological Expedition to Abyssinia, 1826-27, Microlepidoptera. Transactions of the Entomological Society of London 80: 107-120. doi: 10.1111/j.13652311.1932.tb00085.x

Robinson GS (1976) A taxonomic revision of the Tinissinae of the world (Lepidoptera: Tineidae). Bulletin of the British Museum (Natural History), Entomology 32(7): 253-300.
Robinson GS (1981) Remarks on the classification of the fungivorous Tineidae with special reference to the Tinissinae (Lepidoptera). Entomologica Scandinavica 12: 363-380.
Robinson GS (1986) Fungus moths: a review of the Scardiinae (Lepidoptera: Tineidae). Bulletin of the British Museum (Natural History) Entomology 52(2): 37-181.
Robinson GS (2008) Global taxonomic database of Tineidae (Lepidoptera) (v.8.0). http:// www.nhm.ac.uk/entomology/tineidae/index.html
Robinson GS, Tuck KR (1998) A spectacular new montane species of Tinissa (Lepidoptera: Tineidae, Scardiinae) from Borneo and Sulawesi. Tinea 15: 338-341.
Walker F (1864) Tineites. List of the specimens of the lepidopterous insects in the collection of the British Museum 29: 564-835.
Zagulajev AK (1972) New and little known species of Tineidae, Deuterotineidae, Ochsenheimeriidae (Lepidoptera). Trudy Zoologicheskogo Instituta, Leningrad 52: 332-356.

# Forever in the dark: the cave-dwelling azooxanthellate reef coral Leptoseris troglodyta sp. n. (Scleractinia, Agariciidae) 

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#### Abstract

The coral species Leptoseris troglodyta sp. n. (Scleractinia, Agariciidae) is described as new to science. It is the first known azooxanthellate shallow-water agariciid and is recorded from the ceilings of caves at 5-35 $m$ depth in West Pacific coral reefs. The corals have monocentric cup-shaped calices. They may become colonial through extramural budding from the basal coenosteum, which may cause adjacent calices to fuse. The size, shape and habitat of $L$. troglodyta are unique compared to other Leptoseris species, many of which have been recorded from mesophotic depths. The absence of zooxanthellae indicates that it may survive well in darkness, but endolithic algae in some corals indicate that they may be able to get some light. The presence of menianes on the septal sides, which may help to absorb light at greater depths in zooxanthellate corals, have no obvious adaptive relevance in the new species and could have been inherited from ancestral species that perhaps were zooxanthellate. The new species may be azooxanthellate as derived through the loss of zooxanthellae, which would be a reversal in Leptoseris phylogeny.


## Keywords

Cavernicolous, colonial, dwarfism, extramural budding, monocentric, skiophilous, solitary, troglobiotic

## Introduction

Reef-dwelling species of the genus Leptoseris Milne-Edwards and Haime, 1849 (Scleractinia: Agariciidae) consist of foliaceous corals that are common in poorly illuminated environments, such as the deepest parts of reef slopes and vertical rocky walls with crevices, caves, tunnels and overhangs (Dinesen 1980, 1982, 1983; Waheed and Hoeksema in press). They are considered skiophilous (shade-loving) or cavernicolous, i.e., living in caves (Dinesen 1982, 1983). Because they appear to show more preference for dark habitats than many other reef corals, Leptoseris species constitute an important component of zooxanthellate scleractinian coral communities at mesophotic depths (30-150 m) (Kahng and Kelley 2007, Chan et al. 2009, Bare et al. 2010, Bongaerts et al. 2010, Kahng et al. 2010, Rooney et al. 2010, Dinesen et al. 2012). They may even occur deeper, with records of 153 and 165 m by Leptoseris hawaiiensis Vaughan, 1907, in the Pacific Ocean (Maragos and Jokiel 1986, Kahng and Maragos 2006), and 145 m by L. fragilis Milne Edwards and Haime, 1849, in the Red Sea (Fricke and Schuhmacher 1983, Fricke et al. 1987, Kaiser et al. 1993).

Because some Leptoseris species inhabit deep and poorly accessible habitats, they may not all be well known. An example is the recently discovered $L$. kalayaanensis Licuanan \& Aliño, 2009, which so far has only been recorded from rocky substrates at 13-28 m depth in the South China Sea basin (Licuanan and Aliño 2009, Hoeksema et al. 2010). It shows a distinct brown and white coloured pattern on its upper surface, consisting of areas that are either with or without zooxanthellae.

The Agariciidae were not known to include true deep-sea species but according to recent phylogenetic studies, the solitary attached deep-water coral Dactylotrochus cervicornis (Moseley, 1881), which was originally classified with the Caryophylliidae, is also a member of the Agariciidae (Kitahara et al. 2010, 2012). This species has a recorded depth range of $73-852 \mathrm{~m}$, is therefore considered ahermatypic and probably azooxanthellate (for terminology see Schuhmacher and Zibrowius 1985, Cairns and Kitahara 2012). It is monocentric and has smooth-edged septa that bear $2-5$ elongate ridges (menianes or latera), which are considered characteristic for the Agariciidae (Kitahara et al. 2010, 2012).

Because D. cervicornis is predomintly from deep water, it is considered the first known extant azooxanthellate agariciid. D. cervicornis holds a basal position in a recent phylogeny reconstruction of extant Agariciidae and because extinct solitary agariciids from the Middle Cretaceous were also solitary, it is assumed that the ancestor of the Agariciidae, which nowadays predominantly consist of colonial and zooxanthellate species, was also solitary and azooxanthellate (Kitahara et al. 2012).

In the present paper a new agariciid coral species is described that is entirely azooxanthellate and dwells on ceilings of caves in steep reef slopes and walls. No co-occurrence with any zooxanthellate scleractinians was observed. Although its calices are relatively small, cup-shaped and predominantly monocentric, it resembles species of Leptoseris, which otherwise is known to consist of zooxanthellate species
with polycentric ("circumoral") calices (Wells 1956). It is furthermore unique among extant reef-dwelling Agariciidae because it is modular (colonial) through extramural budding by growing new calices from a basal coenosteum, which eventually may fuse. It has been found in the western Pacific, including eastern Indonesia, central Philippines, Papua New Guinea, Palau and Guam. Most of its presently known distribution range overlaps with the centre of maximum marine species richness, the so-called Coral Triangle (Hoeksema 2007).

## Methods

Specimens were observed, photographed and collected while diving with the help of SCUBA. All specimens were encountered below 5 m depth on the ceilings of caves inside steep reef slopes and walls, usually in areas with limestone outcrops. The caves measured one to several meters in width and several meters in length, enabling easy access and maneuvering for observations. Use of an underwater torch was indispensable to locate the corals. Collected specimens were soaked in fresh water or in sodium hypochlorite solution for cleaning. They were deposited in the Coelenterata collection (RMNH Coel.) of Naturalis Biodiversity Center, Leiden (formerly known as Rijksmuseum van Natuurlijke Historie). Specimens from Cebu were already available in the RMNH collection before they were photographed and collected for the present research. SEM photographs were taken with a Jeol 6480LV electron microscope operated at 10 kV .

## Systematic section

Order Scleractinia Bourne, 1905
Family Agariciidae Gray, 1847
Genus Leptoseris Milne Edwards \& Haime, 1849

## Leptoseris troglodyta sp. n.

urn:lsid:zoobank.org:act:DB802B45-E18D-4F55-9D23-5401660DD94E
http://species-id.net/wiki/Leptoseris_troglodyta
Figures 1-8
Leptoseris sp. Hoeksema and Van Ofwegen 2004.

Type material. Holotype. RMNH Coel. 40138 (1 specimen, dry, Figure 3a), Palau, W of Ulong Island (Rattakadokoru Island), W off barrier reef, $07^{\circ} 18^{\prime} 40^{\prime \prime} \mathrm{N}, 134^{\circ} 13^{\prime} 30^{\prime \prime} \mathrm{E}$, Tsey's tunnel ceiling at 32 m depth, 28 July 2002, coll. B.W. Hoeksema. Paratypes: RMNH Coel. 40139 ( 15 specimens, dry), same collection data (Figures 2d, 3b-d).


Figure I. Living specimens of Leptoseris troglodyta sp. n. a Philippines, Cebu Strait, W of Bohol, NW of Cabilao Island, 10-30 m depth (7 November 1999) b Indonesia, NE Kalimantan, Berau Islands, S of Derawan Island, 7-10 m depth (4 October 2003).


Figure 2. Living specimens of Leptoseris troglodyta sp. n. a Philippines, Cebu Strait, W of Bohol, NW of Cabilao Island, $10-30 \mathrm{~m}$ depth ( 7 November 1999) b Indonesia, Tukang Besi Islands (Wakatobi), Binongko, 20 m depth ( 10 May 2003) c Indonesia, North Sulawesi, S of Bunaken Island, 17 m depth ( 19 December 2008; photo B.T. Reijnen) d Palau, W of Ulong Island (Rattakadokoru Island), W off barrier reef, 32 m depth ( 28 July 2002) e Papua New Guinea, Misima Island, 6-10 m depth (31 May 1998; photo G. Paulay) f Guam, Blue Hole, 35 m depth (1 June 2000; photo G. Paulay).

Other material examined. Philippines. RMNH Coel. 24187 ( 3 specimens, dry), RMNH Coel 24195 ( 36 specimens, dry), Cebu Strait, W of Bohol, NW side of Cabilao Island, $09^{\circ} 53^{\prime} 12^{\prime \prime} \mathrm{N}, 123^{\circ} 45^{\prime} 32^{\prime \prime} \mathrm{E}$, vertical wall with caves, $10-30 \mathrm{~m}$ depth, 7 and 17 November 1999, coll. B.W. Hoeksema (Figures 1a, 2a, 7, 8). RMNH Coel. 40151 (2 specimens, dry), Philippines, Cebu, Mactan Island, Lapu-Lapu City, Marigondon


Figure 3. Holotype (RMNH Coel. 40138) and three paratypes (RMNH Coel. 40139) of Leptoseris troglodyta sp. n. from Palau. Scale bars: 1 cm . a Holotype consists of four calices: one (most left) has fused mid-height its calyx with two totally fused calices (centre), while another (right) has fused only with its corallum margin to those at the centre and the rest of its calyx has remained separate $\mathbf{b}$ Paratype: two separate calyces $\mathbf{c}$ Paratype: single calyx d Paratype: two fused calices. Scale bars: 1 cm .


Figure 4. Specimens of Leptoseris troglodyta sp. n. from Indonesia, Wakatobi (RMNH Coel. 40152) a Single specimen from the side showing cup-shaped calyx shape and costae with fine granular spines (scale bar: 1 cm ) b Close-up of large calyx showing septa with wide menianes along their sides (see Figure 5; scale bar: 5 mm ).


Figure 5. SEM photographs of Leptoseris troglodyta sp. n. from Wakatobi, Indonesia (RMNH Coel. 40152). a Upper side of septa showing wide menianes (scale bar: 1 mm ; insert: Figure 5b) b Close-up (insert) of Figure 5a (arrows: menianes; scale bar: 0.5 mm ) c Cross-section of septa showing multiple menianes along their sides (scale bar: 0.5 mm ; insert: Figure 5d) d Close-up (insert) of Figure 5c (arrows: menianes; scale bar: 0.5 mm ).


Figure 6. Specimens of Leptoseris troglodyta sp. n. from the Philippines, Cebu, Mactan Island, (RMNH Coel. 40151). Scale bars: 1 cm . a Upper side of a specimen showing four separate calices $\mathbf{b}$ Three calices that are partly fused at their sides.


Figure 7. Specimens of Leptoseris troglodyta sp. n. from the Philippines, W of Bohol, NW side of Cabilao Island (RMNH Coel 24195). Scale bars: 1 cm . a Cluster of calices formed by extra-calicular budding showing costae with small granular spines $\mathbf{b}$ Idem.


Figure 8. Specimens of Leptoseris troglodyta sp. n. from the Philippines, W of Bohol, NW side of Cabilao Island (RMNH Coel 24195). Scale bars: 5 mm a Single calyx from above, showing nearly solid columella in the fossa $\mathbf{b}$ Two totally fused monocentric calices showing, each with its own fossa $\mathbf{c}$ Upper side of calyx showing clearly visible granulation on septal sides $\mathbf{d}$ Calyx showing solid corallum wall covered by costae with fine granular spines.

Cave, $10^{\circ} 15^{\prime} 33^{\prime \prime} \mathrm{N}, 123^{\circ} 59^{\prime} 07^{\prime \prime} \mathrm{E}$, ceiling of cave at $25-30 \mathrm{~m}$ depth, May 1981 , coll. M.B. Best (Figure 6). Indonesia. RMNH Coel. 40152 (7 specimens, dry), Tukang Besi Islands (Wakatobi), Binongko, SW Bay, $05^{\circ} 59^{\prime} 47^{\prime \prime} \mathrm{S}, 124^{\circ} 02^{\prime} 55^{\prime \prime} \mathrm{E}$, cave in steep wall 20 m depth, 10 May 2003, coll. B.W. Hoeksema (Figures 2b, 4-5). RMNH Coel. 40150 (25 specimens, ethanol), Tukang Besi Islands (Wakatobi), NE Hoga Island, $05^{\circ} 28^{\prime} \mathrm{S}, 123^{\circ} 47^{\prime} \mathrm{E}$, cave in steep reef slope 5 m depth, 12 July 2011, coll. B.W. Hoeksema. RMNH Coel. 40137 ( 2 specimens, ethanol), North Sulawesi, South of Bunaken Island, Alung Banua village, $01^{\circ} 37^{\prime} 07^{\prime \prime N}, 124^{\circ} 45^{\prime} 30^{\prime \prime} \mathrm{E}$, tunnel in reef wall 17 m depth, 19 December 2008, coll. B.T. Reijnen and S.E.T van der Meij (Figure 2c).

Additional photographic records: Indonesia, NE Kalimantan, Berau Islands, S of Derawan Island, jetty Derawan Dive Resort, $02^{\circ} 17^{\prime} 03^{\prime \prime N}, 118^{\circ} 14^{\prime} 49^{\prime \prime} \mathrm{E}$, ceiling of caves, $7-10 \mathrm{~m}$ deep ( 4 October 2003; Figure 1b). Papua New Guinea, Off SE point, Misima Island, Pt. Ebola, 6-10 m depth (31 May 1998; Figure 2e). Guam, Blue Hole, 35 m depth ( 1 June 2000; Figure 2f).

Description. Corallum attached and solitary or colonial by budding from basal coenosteum (Figures 3-4, 6-8). Calices predominantly monocentric, very thin, cup-shaped to foliaceous, height $<15 \mathrm{~mm}$, outline irregularly circular, $\varnothing<30 \mathrm{~mm}$ (Figures 1-4, 6-8). They are usually separate from each other above the interconnecting basal plate (Figures 3b, 6, 7), but can also be fused at their margins or lateral sides (Figures 3a, 8b). Corallum wall massive. Costae equal and well defined, with small spiny protuberances (Figures 4a, $7 \mathrm{a}, 8 \mathrm{~d})$. Septa approximately equal in size, with smooth upper edges and parallel ridges (menianes) on their sides (Figures 3, 5, 8). Septal sides may show evenly distributed granulations where menianes are absent (Figures 8b-c). Columella nearly solid (Figures 3a, 3c-d, 8a-b). Living animals azooxanthellate; corals are white, or partly green or red (Figures 1-2), owing to the presence of endolithic algae (Kühl and Polerecky 2008).

Diagnosis. Corals cave-dwelling, azooxanthellate. Calices small, cup-shaped, monocentric or fused, forming buds at basal coenosteum.

Etymology. The epithet troglodyta (noun) means cave dweller in Latin, derived from ancient Greek for "one who dwells in holes".

Distribution. Records are from coral reefs, usually in areas with limestone outcrops: Indonesia (East Kalimantan, North Sulawesi, Southeast Sulawesi), the central Philippines (Cebu, Bohol), Palau, eastern Papua New Guinea, and the Marianas (Guam) (Figure 9).

## Discussion

## Systematics

Leptoseris troglodyta sp. n. has a habitat and growth form unlike any other known Leptoseris; its corals are cavernicolous and azooxanthellate, and have small, monocentric calices that may multiply by extramural budding and fuse. Other Leptoseris species are


Figure 9. Distribution map of Leptoseris troglodyta sp. n. showing records at (I) Palau, (2) East Kalimantan, (3) North Sulawesi, (4) Wakatobi, (5) Bohol, (6) Guam, (7), eastern Papua New Guinea.
polycentric by circumoral and marginal budding (Dinesen 1980, Veron and Pichon 1980, Veron 2000, Licuanan and Aliño 2009). All congeneric species are larger, which may be related to their symbiosis with zooxanthellae. Polycentric species are usually bigger than close relatives with monocentric calices, as demonstrated for mushroom corals (Hoeksema 1991b, Gittenberger et al. 2011).

The new species lacks pigments of its own, like many cavernicolous (= troglobiotic) animals. Although there are no zooxanthellae in its soft tissue, it usually contains green or red shade-adapted endolithic cyanobacteria imbedded in the skeleton, which have also been reported from Leptoseris fragilis Milne-Edwards and Haime, 1849 (Schlichter et al. 1997).

Dinesen (1983) mentions the occurrence of "numerous Leptoseris cf. scabra (G. Hodgson, M. Ross, pers. comm.)", which were observed in 1981 on the ceiling inside the large Marigondon Cave (Mactan Island, Cebu), the Philippines. They were found "further back in the cave, in gloomier conditions" than the cave entrance at ca 30 m depth. It is likely that this record pertains to specimens of the new species. Two corals of the present study were collected at that site in 1981 and were available for study in the RMNH collection in Leiden. These specimens confirm that the new species was present at that locality at that time. Museum collections may indeed help to retrieve information on the earlier occurrence of coral species that have not been recorded before (Van der Meij et al. 2010, Hoeksema et al. 2011, Van der Meij and Visser 2011). However, without field observations it would not have been possible to know that the new species lacks zooxanthellae.

Despite its unique small monocentric corallites and lack of zooxanthellae, the new species is classified with Leptoseris because of similarly shaped septa and costae, and by its thin, cup-shaped or foliaceous coralla somewhat resembling those of L. fragilis. The latter species features corals that can be monocentric, but its calices grow larger ( $\varnothing$ > 50 mm ) and may eventually form secondary stomata by intracalicular, circumoral budding. Other extinct agariciid genera predominantly consist of encrusting and massive, polystomatous species (Wells 1954, Veron 2000).

Support from molecular analyses (Stefani et al. in prep) would be needed to justify the position of the new species in a separate genus instead of Leptoseris. In that case, it could be appropriate to classify it with Trochoseris Milne Edwards \& Haime, 1849, an extinct genus (Mid Cretaceous - Oligocene) consisting of corals that are solitary, attached and turbinate or trochoid (Wells 1956). However, molecular analyses cannot support such a transition because no live material is available of this genus.

The corals have a basic growth form like that of Cladopsammia gracilis (Milne Edwards and Haime 1848) (Dendrophylliidae), i.e., by extramural budding from the basal coenosteum (see Cairns 1991). They are not distinctly reptoid as described for some Rhizopsammia species of the same family (Cairns and Zibrowius 1997) because there are no clear basal costate stolons involved. Extramural budding is also shown by fossils of the extinct genus Brachyphyllia Reuss, 1854 (Agariciidae), which has thick, low plocoid corallites (Wells 1956) instead of the thin cup-shaped corallites shown by the new species. Compared to the extant solitary agariciid deep-water species Dactylotrochus cervicornis (see Wells 1954, Cairns 1999, Kitahara et al. 2012, Cairns and Kitahara 2012), L. troglodyta differs by the capacity to become colonial and by having a circular corallum outline instead of a periphery with thecal extensions. Only a few specimens of $D$. cervicornis are known to show an 'aberrant' tendency for coloniality (Kitahara et al. 2012).

## Evolution of symbiosis with zooxanthellae

Leptoseris troglodyta is the first extant shallow-water agariciid known known to be reef-dwelling and azooxanthellate. The extinct agariciid genera, Trochoseris Milne Edwards \& Haime, 1849 and Vaughanoseris Wells, 1934, also consists of monocentric species; the first being attached and turbinate or trochoid, and the second being free-living and discoid (Wells 1956). According to Kitahara et al. (2012) they could have been azooxanthellate because the extant Dactylotrochus cervicornis, which shows a basal position in the phylogeny reconstruction of the Agariciidae, is considered both azooxanthellate and monocentric. However, the phylogenetic positions of the two extinct taxa are unknown and although they share a (supposedly plesiosmorph) solitary growth form with Dactylotrochus, this does not necessarily imply that they are phylogenetically closely related to each other. For example, several phylogenetic lineages within the reef-dwelling mushroom coral family Fungiidae show an evolution from monocentric to polycentric zooxanthellate corals, implying that not all
monocentric fungiid species are directly related to each other (Hoeksema 1989, 1991b, Gittenberger et al. 2011, Benzoni et al. 2012).

Coincidently, the Fungiidae show many examples of solitary free-living species that resemble the extinct Vaughanoseris. So, regarding their lifestyle, Vaughanoseris species could be reef-dwelling and zooxanthellate as well. Moreover, the attached, monocentric dendrophylliid Balanophyllia europaea (Risso, 1826) is an example of a single zooxanthellate species (Schuhmacher and Zibrowius 1985) among a majority of congeners without zooaxanthellae (Cairns et al. 1999). Its growth form resembles that of Trochoseris, for which the absence of zooxanthellae may therefore perhaps be less certain. Nevertheless, a solitary growth form, an old fossil record, and a possible ancestral position in the phylogeny may not be sufficient to predict whether an extinct coral taxon may have been azooxanthellate. Its habitat (especially depth) may be more indicative, especially if species were shallow reef-dwellers.

The deep-water species Dactylotrochus cervicornis and the cave-dweller Leptoseris troglodyta posses parallel ridges on the sides of their septa, which are called menianes (Kitahara et al. 2010, 2012). These are probably the same structures (compare Figure 5, Kitahara et al. 2012 figure 2, and Kahng et al. 2012 figure 9) that help zooxanthellate Leptoseris corals to absorb sunlight more efficiently at greater depths (Kahng et al. 2012). The bathymetric records of $D$. cervicornis (Kitahara et al. 2012) partly coincide with depth ranges of zooxanthellate Leptoseris species (see Dinesen et al. 2012). Depth may therefore not always be indicative for the absence of zooxanthellae.

All live specimens of L. troglodyta, which were observed in their habitat (Figures 1-2), were clearly azooxanthellate by lacking a brown colour, like completely bleached corals (Hoeksema 1991a, Hoeksema and Matthews 2011). In the case of dredged deep sea corals in collections, which are either dry or being preserved in ethanol, pigments of any present zooaxanthellae might have dissolved, which makes it difficult to see whether they were present. If $D$. cervicornis corals lack zooxanthellae, like those of Leptoseris troglodyta, the presence of menianes at least suggests that their ancestors might have been zooxanthellate and that the loss of zooxanthellae may be an evolutionary novelty related to life in deep water or in caves. Consequently, ancestral agariciids, along with Trochoseris and Vaughanoseris, were perhaps also zooxanthellate like many modern monocentric zooxanthellate reef corals in illuminated habitats. On the other hand, a preceding presence of menianes in agariciid corals may also have facilitated the development of symbiosis with zooxanthellae, which implies that early agariciids may have been azooxanthellate as suggested by Kitahara et al. (2012), presenting a "chicken or the egg" causality dilemma.

The deep-water species Dactylotrochus cervicornis and the cave-dweller Leptoseris troglodyta both live in dark environments. The latter has been observed to be azooxanthellate in caves at various localities. D. cervicornis specimens may have to be examined for the absence of zooxanthellae to be sure that this species is always azooxanthellate. If so, its menianes have no use in connection to light absorption, like in $L$. troglodyta. L. troglodyta shows that the evolutionary relation between scleractinian reef corals and their algal symbionts is not fixed and that it may be difficult to deduct such a relation
based on coral growth forms and their possible position in phylogeny reconstructions, especially if no molecular data can be obtained, as is the case for fossil corals. According to a recent molecular study, coloniality may have become lost at least six times and symbiosis with zooxanthellae at least three times in the phylogeny of the Scleractinia (Barbeitos et al. 2010). However, these numbers are based on a subset of species and may have to be revised if additional species are involved (see Gittenberger et al. 2011).

## Eco-morphological considerations

Leptoseris troglodyta $\mathrm{sp} . \mathrm{n}$. is the first reef-dwelling agariciid coral without zooxanthellae. As a small cave-dwelling species it can live without sunlight. It has not been observed to co-occur with any zooxanthellate scleractinians (only azooxanthellate species), although in small and poorly illuminated cavities a variety of zooxanthellate scleractinians can be discerned (Dinesen 1983). Other Leptoseris species are zooxanthellate and most of them are able to live on deep, poorly illuminated reefs or rocky substrates. Therefore, from an evolutionary perspective, the new species may have lost the capacity to live in symbiosis with zooxanthellae and it may have obtained a smaller corallum size (dwarfism). By their small and thin corolla the corals have little weight. Consequently, with their wide basal plate they may not easily break off from their substrate, which consist of porous limestone cave ceilings where settlement space is limited. Without zooxanthellae, they may not easily reach large sizes, whatsoever. Owing to its modular growth form a $L$. troglodyta coral may risk losing a few expendable calices while other Leptoseris corals may harm and lose their entire corallum in unfavourable conditions. Other examples of zooxanthellae loss in relation to a cavernicolous lifestyle are so far unknown among scleractinian families. However, among shallow-water brachycnemic zoanthids an undescribed cave-dwelling Palythoa species has been recorded that also lacks zooxanthellae, while its congeners are known to be zooxanthellate (Reimer 2010).

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## References

Barbeitos MS, Romano SL, Lasker HR (2010) Repeated loss of coloniality and symbiosis in scleractinian corals. Proceedings of the National Academy of Sciences of the United States of America 107: 11877-11882. doi: 10.1073/pnas. 0914380107
Bare AY, Grimshaw KL, Rooney JJ, Sabater MG, Fenner D, Carroll B (2010) Mesophotic communities of the insular shelf at Tutuila, American Samoa. Coral Reefs 29: 369-377. doi: 10.1007/s00338-010-0600-y

Benzoni F, Arrigoni R, Stefani F, Reijnen BT, Montano S, Hoeksema BW (2012) Phylogenetic position and taxonomy of Cycloseris explanulata and C. wellsi (Scleractinia: Fungiidae): lost mushroom corals find their way home. Contributions to Zoology 81: 125-146.
Bongaerts P, Ridgway T, Sampayo EM, Hoegh-Guldberg O (2010) Assessing the 'deep reef refugia' hypothesis: focus on Caribbean reefs. Coral Reefs 29: 309-327. doi: 10.1007/ s00338-009-0581-x
Bourne GC (1905) Report on the solitary corals collected by Professor Herdmann, at Ceylon, in 1902. Ceylon Pearl Oyster Fisheries, Part 4, Supplementary Report 29: 187-242, pls. 1-4.

Cairns SD (1991) A revision of the ahermatypic Scleractinia of the Galapagos and Cocos Islands. Smithsonian Contributions to Zoology 504: 1-32, pls 1-12. doi: 10.5479/ si. 00810282.504
Cairns SD (1999) Cnidaria Anthozoa: deep-water azooxanthellate Scleractinia from Vanuatu, and Wallis and Futuna islands. Mémoires du Muséum national d'Histoire naturelle 180: 31-167.
Cairns SD, Kitahara MV (2012) An illustrated key to the genera and subgenera of the Recent azooxanthellate Scleractinia (Cnidaria: Anthozoa), with an attached glossary. ZooKeys 227: 1-47. doi: 10.3897/zookeys.227.3612
Cairns SD, Zibrowius H (1997) Cnidaria Anthozoa: Azooxanthellate Scleractinia from the Philippine and Indonesian regions. Mémoires du Muséum national d'Histoire naturelle 172: 27-243.
Cairns SD, Hoeksema BW, Van der Land J (1999) List of extant stony corals. Atoll Research Bulletin 459: 13-46.
Chan YL, Pochon X, Fisher MA, Wagner D, Concepcion GT, Kahng SE, Toonen RJ, Gates RD (2009) Generalist dinoflagellate endosymbionts and host genotype diversity detected from mesophotic (67-100 m depths) coral Leptoseris. BMC Ecology 2009, 9: 21 doi: 10.1186/1472-6785-9-21

Dinesen ZD (1980) A revision of the coral genus Leptoseris (Scleractinia: Fungina: Agariciidae). Memoirs of the Queensland Museum 20: 182-235.
Dinesen ZD (1982). Regional variation in shade-dwelling coral assemblages of the Great Barrier Reef Province. Marine Ecology Progress Series 7: 117-123. doi: 10.3354/meps007117

Dinesen ZD (1983) Shade-dwelling corals of the Great Barrier Reef. Marine Ecology Progress Series 10: 173-185. doi: 10.3354/meps010173
Dinesen ZD, Bridge TCL, Luck DG, Kahng SE, Bongaerts P (2012) Importance of the coral genus Leptoseris to mesophotic coral communities in the Indo-Pacific. Poster $12^{\text {th }}$ International Coral Reef Symposium, Cairns, 2012: P101.
Fricke HW, Schuhmacher H (1983) The depth limits of Red Sea stony corals: an ecophysiological problem (a deep diving survey by submersible). PSZNI: Marine Ecology 4: 163-194. doi: 10.1111/j.1439-0485.1983.tb00294.x
Fricke HW, Vareschi E, Schlichter D (1987) Photoecology of the coral Leptoseris fragilis in the Red Sea twilight zone (an experimental study by submersible). Oecologia 73: 371-381. doi: 10.1007/BF00385253
Gittenberger A, Reijnen BT, Hoeksema BW (2011) A molecularly based phylogeny reconstruction of mushroom corals (Scleractinia: Fungiidae) with taxonomic consequences and evolutionary implications for life history traits. Contributions to Zoology 80: 107-132.
Gray JE (1847) An outline of an arrangement of stony corals. Annals and Magazine of Natural History, Ser. 1, 19: 120-128. doi: 10.1080/037454809496460
Hoeksema BW (1989) Taxonomy, phylogeny and biogeography of mushroom corals (Scleractinia: Fungiidae). Zoologische Verhandelingen Leiden 254: 1-295.
Hoeksema BW (1991a) Control of bleaching in mushroom coral populations (Scleractinia: Fungiidae) in the Java Sea: stress tolerance and interference by life history strategy. Marine Ecology Progress Series 74: 225-237. doi: 10.3354/meps074225
Hoeksema BW (1991b) Evolution of body size in mushroom corals (Scleractinia: Fungiidae) and its ecomorphological consequences. Netherlands Journal of Zoology 41: 122-139.
Hoeksema BW (2007) Delineation of the Indo-Malayan centre of maximum marine biodiversity: the Coral Triangle. In: Renema W (Ed) Biogeography, Time and Place: Distributions, Barriers and Islands. Springer, Dordrecht, 117-178. doi: 10.1007/978-1-4020-6374-9_5
Hoeksema BW, Matthews JL (2011) Contrasting bleaching patterns in mushroom coral assemblages at Koh Tao, Gulf of Thailand. Coral Reefs 30: 95. doi: 10.1007/s00338-010-0675-5
Hoeksema BW, Van Ofwegen LP (2004) Indo-Malayan reef corals: a generic overview. World Biodiversity database, CD-ROM Series ETI, Amsterdam.
Hoeksema BW, Dautova TN, Savinkin OV, Tuan VS, Ben HX, Hoang PK, Du HT (2010) The westernmost record of the coral Leptoseris kalayaanensis in the South China Sea. Zoological Studies 49: 325.
Hoeksema BW, Van der Land J, Van der Meij SET, Van Ofwegen LP, Reijnen BT, Van Soest RWM, De Voogd NJ (2011) Unforeseen importance of historical collections as baselines to determine biotic change of coral reefs: the Saba Bank case. Marine Ecology 32: 135-141. doi: 10.1111/j.1439-0485.2011.00434.x
Kahng SE, Kelley C (2007) Vertical zonation of megabenthic taxa on a deep photosynthetic reef ( $50-140 \mathrm{~m}$ ) in the Au'au Channel, Hawaii. Coral Reefs 26: 679-687. doi: 10.1007/ s00338-007-0253-7

Kahng SE, Maragos JE (2006) The deepest zooxanthellate, scleractinian corals in the world? Coral Reefs 25: 254. doi: 10.1007/s00338-006-0098-5

Kahng SE, Garcia-Sais JR, Spalding HL, Brokovich E, Wagner D, Weil E, Hinderstein L, Toonen RJ (2010) Community ecology of mesophotic coral reef ecosystems. Coral Reefs 29: 255-275. doi: 10.1007/s00338-010-0593-6
Kahng SE, Hochberg EJ, Aprill A, Wagner D, Luck DG, Perez D, Bidigare RR (2012) Efficient light harvesting in deep-water zooxanthellate corals. Marine Ecology Progress Series 455: 65-77. doi: 10.3354/meps09657
Kaiser P, Schlichter D, Fricke HW (1993) Influence of light on algal symbionts of the deep coral Leptoseris fragilis. Marine Biology 117: 45-52. doi: 10.1007/BF00346424
Kitahara MV, Cairns SD, Stolarski J, Blair D, Miller DJ (2010) A comprehensive phylogenetic analysis of the Scleractinia (Cnidaria, Anthozoa) based on mitochondrial CO1 sequence data. PLoS ONE 5(7): e11490. doi: 10.1371/journal.pone. 0011490
Kitahara MV, Stolarski J, Miller DJ, Benzoni F, Stake J, Cairns SD (2012) The first modern solitary Agariciidae (Anthozoa, Scleractinia) revealed by molecular and microstructural analysis. Invertebrate Systematics 26: 303-315. doi: 10.1071/IS11053
Kühl M, Polerecky L (2008) Functional and structural imaging of phototrophic microbial communities and symbioses. Aquatic Microbial Ecology 53: 99-118. doi: 10.3354/ame01224
Licuanan WY, Aliño PM (2009) Leptoseris kalayaanensis (Scleractinia: Agariciidae), a new coral species from the Philippines. Raffles Bulletin of Zoology 57: 1-4.
Maragos JE, Jokiel P (1986) Reef corals of Johnston Atoll: one of the world's most isolated reefs. Coral Reefs 4: 141-150. doi: 10.1007/BF00427935
Milne Edwards H, Haime J (1848) Recherches sur les polypiers. Troisième mémoire. Monographie des Eupsammides. Annales des Sciences Naturelles, Zoologie, Ser. 3, 10: 65-114, pl. 1.
Milne Edwards H, Haime J (1849) Recherches sur les polypiers. Quatrième mémoire. Monographie des Astréides. Annales des Sciences Naturelles, Zoologie, Ser. 3, 11: 235-312.
Moseley HN (1881) Report on certain hydroid, alcyonarian, and madreporarian corals procured during the voyage of H.M.S. Challenger, in the years 1873-1876. Challenger Reports, Zoology 2: 1-248, pls. 1-32.
Reimer JD (2010) Key to field identification of shallow water brachycnemic zoanthids (Order Zoantharia: Suborder Brachycnemina) present in Okinawa. Galaxea, Journal of Coral Reef Studies 12: 23-29. doi: 10.3755/galaxea.12.23
Reuss AE (1854) Beiträge zur Characteristik der Kreideschichten in den Ostalpen besonders im Gosauthale und am Wolfgangsee. Denkschriften der Mathematisch-Naturwissenschaftlichen Klasse der Kaiserlichen Akademie der Wissenschaften, Wien 7 (1): 1-157, pls. 1-31.
Risso A (1826) Tableau des zoophytes les plus ordinaires qui existent ou qui ont existé dans les Alpes Maritimes. In: Risso A, Histoire naturelle des principales productions de l'Europe méridionale et particulièrement de celles des environs de Nice et des Alpes Maritimes. Levrault, Paris, 5: 307-383, pl. 3. doi: 10.5962/bhl.title. 58984
Rooney J, Donham E, Montgomery A, Spalding H, Parrish F, Boland R, Fenner D, Gove J, Vetter O (2010) Mesophotic coral ecosystems in the Hawaiian Archipelago. Coral Reefs 29: 361-367. doi: 10.1007/s00338-010-0596-3

Schlichter D, Kampmann H, Conrady S (1997) Trophic potential and photoecology of endolithic algae living within coral skeletons. PSZNI: Marine Ecology 18: 299-317. doi: 10.1111/j.1439-0485.1997.tb00444.x

Schuhmacher H, Zibrowius H (1985) What is hermatypic? A redefinition of ecological groups of corals and other organisms. Coral Reefs 4: 1-9. doi: 10.1007/BF00302198
Vaughan TW (1907) Recent Madreporaria of the Hawaiian Islands and Laysan. Bulletin of the United States National Museum 59: i-iv, 1-427, pls. 1-96. doi: 10.5479/si.03629236.59.i
Van der Meij SET, Visser RR (2011) The Acropora bumilis group (Scleractinia) of the Snellius expedition (1929-30). Raffles Bulletin of Zoology 59: 9-17.
Van der Meij SET, Suharsono, Hoeksema BW (2010) Long-term changes in coral assemblages under natural and anthropogenic stress in Jakarta Bay (1920-2005). Marine Pollution Bulletin 60: 1442-1454. doi: 10.1016/j.marpolbul.2010.05.011
Veron JEN (2000) Corals of the world. Volume 2. Australian Institute of Marine Science, Townsville, 429 pp .
Veron JEN, Pichon M (1980) Scleractinia of Eastern Australia III. Families Agariciidae, Siderastreidae, Fungiidae, Oculinidae, Merulinidae, Mussidae, Pectiniidae, Caryophylliidae, Dendrophylliidae. Australian Institute of Marine Science Monograph Series 4: 1-422. doi: 10.5962/bhl.title. 60646

Waheed Z, Hoeksema BW (in press) A tale of two winds: species richness patterns of reef corals around the Semporna peninsula, Malaysia. Marine Biodiversity. doi: 10.1007/s12526-012-0130-7
Wells JW (1934) Some fossil corals from the West Indies. Proceedings of the United States National Museum, Washington 83: 71-110, pls. 1-5. doi: 10.5479/si.00963801.83-2975.71
Wells JW (1954) Recent corals of the Marshall Islands. United States Geological Survey, Professional Paper 260-I: i-iv, 385-486, pls. 94-187.
Wells JW (1956) Scleractinia. In Moore RC (Ed) Treatise on invertebrate paleontology F. Coelenterata. Geologial Society of America and University of Kansas Press, 328-440.

# First record of the ant subfamily Aenictinae (Hymenoptera, Formicidae) from Saudi Arabia, with the description of a new species 

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#### Abstract

The ant subfamily Aenictinae is recorded for the first time from the Kingdom of Saudi Arabia and for the second time from the Arabian Peninsula. A new species Aenictus arabicus sp. n., is described from the worker caste. Aenictus arabicus belongs to the $A$. wroughtonii-group and appears to be most closely related to $A$. rhodiensis Menozzi, but can be easily distinguished from the latter by the following characters: overall smaller size; cephalic index (head width/head length) small; occipital corners in lateral view rounded; antennal scape when laid back surpassing approximately two-thirds of head length; funicular segments $2-8$ each at least $2 \times$ as long as broad; subpetiolar process well developed; petiole and postpetiole distinctly imbricate; gaster and clypeus entirely yellow, teeth of mandibles reddish- brown. Aenictus arabicus was collected from leaf litter, next to a tree of Psidium guajava L. The new species also is similar to $A$. sagei and A. wroughtonii. Affinities and a key to related species of the species group are given.


## Keywords

Aenictus, taxonomy, Arabian Peninsula, Saudi Arabia, Al Sarawat Mountains, ants, Palaearctic region

## Introduction

The subfamily Aenictinae Emery, 1901, was elevated to the rank of subfamily by Bolton (1990), and includes a single genus, Aenictus Schuckard, 1840. The genus presently has 177 species and subspecific forms (Bolton 2012), distributed through the East Mediterranean, Afrotropical, Oriental, Indo-Australian, and Australian regions (Gotwald 1995, Brown 2000, Aktaç et al. 2004, and Jaitrong and Yamane 2012). Most of the species are tropical (Brown 2000), with terrestrial habitats, foraging in soil, leaf litter, most of the Southeast Asian species forage on the ground, and some on trees (e.g., Hirosawa et al. 2000) and hunting other ant species and termites (Gotwald 1995, Rosciszewski and Maschwitz 1994).

The subfamily Aenictinae is characterized by having:- a waist of two segments, with the spiracle of the postpetiole set behind the midlength of the tergite; all gastral spiracles circular; and the first gastral segment with a narrow, neck-like constriction behind the articulation with the postpetiole, 8-10 antennal segments, the frontal lobes reduced with the antennal sockets completely exposed, and the promesonotal suture absent (Bolton 1994). Species of Aenictus are generally small, monomorphic and yellow to dark brown. Members of the Aenictus wroughtonii-group share the following characters (Jaitrong et al. 2010): head narrow; posterior margin of head lacking collar; antennae long, 10 -segmented; with long scape reaching or surpassing posterolateral corners of head; anterior clypeal margin bearing 5-10 denticles; mandibles subtriangular, with masticatory margin bearing 8-12 minute teeth in addition to a large apical tooth with a sharp apex; frontal carinae short; mesosoma narrow and elongate; legs thin and long; head entirely smooth and shiny; almost entire body clear yellow to yellowish brown.

Since Wilson's (1964) revision, several authors have published taxonomic papers dealing with particular areas or species groups, e.g., Zhou (2001) (South China), Shattuck (2008) (Australia), Terayama (2009) (Taiwan), Zettel and Sorger (2010) (Borneo and the Philippines), Jaitrong and Yamane (2012) (Southeast Asia), and Bharti et al. (2012) (India). A catalogue of the 35 known Afrotropical members can be accessed on the website of Taylor (2012), with photographs also on Fisher's Antweb.org (2012).

Ten species of Aenictus have been reported from the Palaearctic, nine of which are distributed in the Southwestern part of the region, Morocco in the west to Afghanistan in the east (Aktaç et al. 2004). Country records include $A$. rhodiensis from Greece (Menozzi 1936), Turkey (Aktaç et al. 2004), Iran (Radchenko and Alipanah, 2004) and Israel (Kugler 1988); A. fuscovarius fuscovarius Gerstäcker, $A$. fuscovarius sagittarius Santschi and $A$. hamifer Emery from Egypt (Sharaf 2006); and, an unidentified species from Yemen (Collingwood and van Harten 2001). The $A$. wroughtonii-group was revised for the Oriental and Indo-Australian regions (Jaitrong et al. 2010) giving seven species. The new species, $A$. arabicus belongs to this species group with resemblance to $A$. sagei and $A$. wroughtonii described by Forel from India.

Here the subfamily Aenictinae is recorded for the first time from Saudi Arabia and for the second time from the Arabian Peninsula. A new species, A. arabicus sp. n., is described based on the worker caste. The queen and male are unknown. A key to the related species within the $A$. wroughtonii-group is given.

## Materials and methods

The following abbreviations are used for particular morphological features and metrics:
TL Total length; the outstretched body length from the mandibular apex to the gastral apex.
HW Head width; the maximum width of the head in full-face view.
HL Head length; the maximum length of the head, excluding the mandibles.
CI Cephalic index (HW $\times 100 / \mathrm{HL})$.
SL Scape length, excluding condylar bulb.
SI $\quad$ Scape index (SL $\times 100 / \mathrm{HW})$.
ML Mesosoma length; the length of the mesosoma in lateral view, from the point at which the pronotum meets the cervical shield to the posterior base of the propodeal lobes or teeth.
PRW Pronotal width; the maximum pronotal width in dorsal view.
PL Petiole length; the maximum length of petiole measured in dorsal view, from the anterior margin to the posterior margin.
PW Petiole width; the maximum petiolar width measured in dorsal view.
PPL Postpetiole length; the maximum postpetiolar length measured in dorsal view.
PPW Postpetiole width; the maximum postpetiolar width measured in dorsal view.
All measurements are expressed in millimeters. Images were taken with a scanning electron microscope ((SEM) JSM-6380 LA).

## Depositories of type material

BMNH Natural History Museum, London, United Kingdom.
CASC California Academy of Science Collection, San Francisco, California, USA.
KSMA King Saud Museum of Arthropods, King Saud University, Riyadh, Kingdom of Saudi Arabia (Holotype depository).
MCZC Museum of Comparative Zoology, Harvard University, Cambridge, MA, USA.
MHNG Muséum d'Histoire Naturelle, Geneva, Switzerland.
NHMB Naturhistorisches Museum, Basel, Switzerland.
SEMC Division of Entomology (Snow Entomological Collections), University of Kansas Natural History Museum, Lawrence, Kansas, USA.
WMLC World Museum Liverpool, Liverpool, United Kingdom.

## Results

Aenictus arabicus Sharaf \& Aldawood, sp. n.

urn:lsid:zoobank.org:act:347C091D-1E98-4765-AEF5-10C4CACE8DDE
http://species-id.net/wiki/Aenictus_arabicus
Figs 1-12

Holotype worker. Saudi Arabia, Al Baha-Mukhwah Aqaba RD,19.IV.2012, $20.00000^{\circ} \mathrm{N}, 41.43758^{\circ} \mathrm{E}, 1300 \mathrm{~m}$, 19.IV. 2012 (M. R. Sharaf leg.); deposited in the KSMA.

Paratype workers. 21 workers, same data as holotype; 1 deposited in MHNG (Dr Bernhard Merz); 1 deposited in NHMB (Mrs. Isabelle Zürcher-Pfander); 2 deposited in CASC (Dr Brian Fisher); 2 deposited in MCZC (Prof. E. O. Wilson); 2 deposited in SEMC (Prof. Michael S. Engel); 1 deposited in WMLC (Mr. Tony Hunter), 1 deposited in BMNH (Mr. Barry Bolton); the remaining specimens in KSMA (M. R. Sharaf).

Measurements. Holotype: TL 3.0, HL 0.65 , HW 0.52 , SL 0.50 , PRW 0.35, ML 0.95 , PL 0.22, PW 0.15, PPL 0.17, PPW 0.15. Indices: SI 96, CI 80.

Paratypes. TL 2.75-3.12, HL 0.60-0.72, HW 0.42-0.55, SL $0.40-0.52$, PRW $0.20-0.35$, ML $0.77-1.00$, PL $0.22-0.27$, PW $0.12-0.15$, PPL $0.15-0.20$, PPW $0.12-$ 0.17. Indices: SI 77-104, CI 70-92. ( $\mathrm{n}=11$ ).

Description of worker. Head entirely smooth and shining. In full-face view head distinctly longer than broad, with convex sides and nearly straight posterior margin; occipital corners in lateral view rounded; anterior clypeal margin with six small denticles; masticatory margin of mandibles armed with a large apical tooth followed by five smaller subequal teeth and a relatively larger basal tooth; when laid back, antennal scapes surpassing about two thirds of head length; all funicular segments at least twice as long as broad; terminal funicular segment about $2.5 \times$ as long as the proceeding segment; mandibles dull with longitudinal striations; whole head dorsum and antennae with stiff scattered long hairs. Mesosoma in dorsal view broader anteriorly than posteriorly; promesonotum in profile distinctly convex, bearing many pairs of hairs; metanotal groove distinct; mesopleuron faintly but distinctly imbricate; propodeum bare or in some individuals with very sparse decumbent pubescence; propodeal dorsum long, about $4 \times$ as long as declivity; propodeum in profile slightly lower than promesonotum and almost flat dorsally; propodeal junction rounded. Petiole longer than broad in dorsal view with node clearly convex in lateral view; subpetiolar process triangular with convex ventral margin and blunt anteriorly. Postpetiole distinctly smaller than petiole, its node roundly convex, and its anteroventral edge sharp and bearing many hairs; both petiole and postpetiole distinctly imbricate and equipped dorsally with several pairs of backward directed long hairs. Gaster smooth and shining with abundant pairs of hairs. Color uniformly yellow.

Etymology. This species is named after the type locality.


Figures I-8. SEM of Aenictus arabicus sp. n. paratype I body in profile $\mathbf{2}$ mesosoma in profile $\mathbf{3}$ imbricate sculpture of mesopleuron $\mathbf{4}$ petiole and postpetiole in profile $\mathbf{5}$ antennal sockets and anterior clypeal margin $\mathbf{6}$ antenna $\mathbf{7}$ head in full-face view $\mathbf{8}$ anterior part of head.


Figures 9-I 2. Automontage of Aenictus arabicus sp. n. paratype $\mathbf{9}$ body in profile $\mathbf{I} \mathbf{0}$ body in dorsal view I I head in full-face view $\mathbf{I}$ label. ( CASENT0280972).

## Discussion

Affinities. Aenictus arabicus is similar to $A$. rhodiensis Menozzi, 1936 from Greece; and $A$. sagei and $A$. wroughtonii described by Forel from India. All the three species are members of the $A$. wroughtonii-group as defined by Jaitrong et al. (2010) and sharing the following characters: head narrow, entirely smooth and shining; occipital margin lacking collar; antennae long, 10 -segmented; anterior clypeal margin convex, rounded, with 5-10 denticles; mandibles subtriangular, with masticatory margin bearing 8-12 minute teeth in addition to a large apical tooth with a sharpe apex; mesosoma narrow and elongate; subpetiolar process weakly developed or almost absent; body clear yellow to yellowish brown.

Comparing $A$. arabicus with $A$. rhodiensis, both species have a similar general morphology, notably the shape of the mesosoma, petiole and postpetiole, a similar body pilosity; also both have a peculiar subpetiolar process which is somewhat wide and blunt anteriorly and the anterior clypeal margin is equipped with six small denticles. From the more accurate description in Aktaç et al. (2004), A. arabicus can be separated readily from $A$. rhodiensis. The former has a small relatively long, narrow head
(HL 0.60-0.72, HW 0.42-0.55, CI 70-92) and long scapes, when laid back surpassing about two-thirds of the head length (SI 77-104) while the latter has a shorter head (HL 1.23, HW 1.02) and shorter scapes, which just surpass the midpoint of the head. Aenictus arabicus has a nearly straight posterior margin of the head whereas it is weakly concave in A. rhodiensis. The funicular segments 2-8 are at least twice as long as broad in the former, while they are as long as broad in the latter. Aenictus arabicus has an entirely yellow clypeus and reddish-brown mandibular teeth while the sides of the clypeus and mandibular teeth are reddish brown in $A$. rhodiensis. The gaster of $A$. arabicus is entirely yellow, whereas in $A$. rhodiensis, the middle of the third gastral tergite has two longitudinal brownish lines which diverge forward, sometimes reducing to small points. Aenictus dlusskyi Arnoldi, known only from the type series from Armenia, also resembles $A$. arabicus but is of a similar size to rhodiensis (Aktaç et al. 2004).

Comparing $A$. arabicus with the Asian species $A$. sagei (CASENT0281958) and $A$. wroughtonii (lectotype images are given in Jaitrong et al. 2010:35), A. arabicus has the anterior clypeal margin bearing six small denticles; A. sagei has 9-10 denticles; whereas A. wroughtonii has 8-10 denticles. In addition, A. arabicus has the subpetiolar process well developed, triangular, with convex ventral margin and blunt anteriorly and body pilosity fewer and shorter; $A$. sagei has a weakly developed subpetiolar process, with its ventral outline nearly straight; its anteroventral corners obtusely angulate and body pilosity distinctly long and abundant (length of the longest pronotal hair $0.20-0.25 \mathrm{~mm}$, Jaitrong et al. 2010); whereas $A$. wroughtonii has an undeveloped subpetiolar process, with its ventral outline feebly convex and without anterior angle and relatively sparse standing hairs which are shorter than in $A$. sagei.

## Habitat and biology

Al-Baha Province is divided by massive and steep rocky mountains into the lowland coastal plain to the west, known as "Tihama", and the mountainous area ranging 1500 - 2450 m above sea level to the east, known as "Al-Sarat or Al-Sarah" which forms part of Al-Sarawat Mountains. The type locality (Fig. 13) is a small farm at the beginning of a narrow valley isolated between the mountains and the plain with a few native shrubs and trees at 1300 m . The farm is planted with Annona squamosa L. (Annonaceae), Prunus persica (L.), P. Amigdalus (Mill.) (Rosaceae), Psidium guajava L. (Family: Myrtaceae), Zea mays ssp. mays L. (Family: Poaceae), in addition to banana, and mango. The new species was found foraging on the ground under leaf litter and next to a tree of Psidium guajava L. The soil, at the time of collection was well saturated through irrigation and accumulation of organic matter.

The climate in Al-Baha Province is greatly influenced by its varying topography. It is generally moderate in summer and cold in winter with average temperatures ranging between $12-23^{\circ} \mathrm{C}$. In Tihama, the climate is hot in summer, warm in spring and mild in winter, with humidity ranging between $52 \%-67 \%$, and a rainfall less than 100 mm annually. While in the mountainous area, Al-Sarah, The climate is greatly different from


Figure I3. Type locality, Al Bahah, Mukhwah Aqaba RD. (photo M. R. Sharaf).
that in Tihama although they are separated by no more than 30 km . The weather is cooler in summer and winter due to its high altitude. Al-Sarah is exposed to the formation of clouds and fog, and this often happens in winter because of air masses coming from the Red Sea, accompanied by thunderstorms. In spring and summer, the climate is mild and pleasant. Also, rainfall is higher with falls in the range of $229-581 \mathrm{~mm}$. The average rain falls throughout the whole province is $100-250 \mathrm{~mm}$ annually

Collingwood and van Harten (2001) recorded Aenictinae for the first time from the Arabian Peninsula based on an unidentified species, from workers collected in Yemen among leaf litter of banana plantations in Khamis Bani Sa'd and Lahj. The diagnostic characters given for this species indicate a similarity in general habitus to $A$. arabicus but in two characters mentioned by them, a broadly emarginate head and unique crenulation of the anterior clypeal border, their species disagrees with the present new species. Future collecting in Yemen is needed to clarify the status of this taxon.

The presence of an Aenictus species in the Southwestern part of Saudi Arabia is not surprising as the area is regarded as being Afrotropical (Bodenheimer 1937; Nayman 1972; Sharaf et al. 2012; El-Hawagry et al. unpubl. data), and it is likely that more Afrotropical ants are awaiting discovery in the area.

Despite the Afrotropical nature of the type locality we found it important to give a key to the closely related species in the $A$. wroughtonii-group.

## Key to species of the Aenictus wroughtonii-group related to A. arabicus based on worker

| 1 | Subpetiolar process almost absent, anteroventrally not angulate (India) ........ |
| :---: | :--- |
|  | .................................................................................................................. 2 |

2 Anterior clypeal margin bearing 9-10 denticles; subpetiolar process weakly developed (India)

- Anterior clypeal margin bearing six denticles; subpetiolar process well developed 3
3 Funicular segments 2-8 as long as broad; middle of third gastral tergite with two longitudinal brownish lines, sometimes reducing to small points; scapes when laid back just surpass the midpoint of head (Greece) $\qquad$ rhodiensis
- Funicular segments 2-8 at least twice as long as broad; gaster entirely yellow; scapes when laid back surpassing about two-thirds of head length (Saudi Arabia) arabicus sp. n.


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## References

Aktaç N, Radchenko AG, Kiran K (2004) On the taxonomy of the west Palaearctic Aenictinae (Hymenoptera: Formicidae). Annales Zoologici 54(2): 361-364.
Bharti H, Wachkoo AA, Kumar R (2012) Two remarkable new species of Aenictus (Hymenoptera: Formicidae) from India. Journal of Asia-Pacific Entomology 15: 291-294. doi: 10.1016/j.aspen.2012.02.002

Bodenheimer FS (1937) Problems of animal distribution in Arabia. Proceedings of the Linnean Society of London 148: 47-48.
Bolton B (1990) Army ants reassessed: the phylogeny and classification of the Doryline section (Hymenoptera, Formicidae). Journal of Natural History 24: 1339-1364. doi: 10.1080/00222939000770811

Bolton B (1994) Identification guide to the ant genera of the world. Harvard University Press, Cambridge, Mass., 222 pp.
Bolton B (2012) Barry Bolton's Synopsis of the Formicidae and Catalogue of Ants of the World:1 January 2012. http://www.gap.entoclub.org/ [accessed 17 January 2012]
Brown WL Jr. (2000) Diversity of ants. In: Agosti et al. (Eds) Ants. standard methods for measuring and monitoring biodiversity. Biological diversity hand book series. Smithsonian Institution Press, Washington and London, 280 pp.
Collingwood CA, Harten AV (2001) Additions to the ant fauna of Yemen (Hymenoptera: Formicidae). Esperiana 8: 559-568.
Emery C (1901) Notes sur les sous-familles des Dorylines et Ponérines (Famille des Formicides). Annales de la Société Entomologique de Belgique 45: 32-54.
Fisher BL. Antweb. The California Academy of Sciences, San Francisco, U.S.A. http://www. antweb.org [accessed 17 January 2012]
Gotwald WH (1995) Army ants. The biology of social predation. Cornell University Press, New York, 320 pp.
Hirosawa H, Higashi S, Mohamed M (2000) Food habits of Aenictus army ants and their effects on ant community in a rain forest of Borneo. Insectes Sociaux 47, 42-49. doi: 10.1007/s000400050007

Jaitrong W, Yamane S, Wiwatwitaya D (2010) The army ant Aenictus wroughtonii (Hymenoptera: Formicidae: Aenictinae) and related species in the Oriental Region, with description of two new species. Japanese Journal of Systematic Entomology 16, 33-46.
Jaitrong W, Yamane S (2012) Review of the Southeast Asian species of the Aenictus javanus and Aenictus philippinensis species groups (Hymenoptera, Formicidae, Aenictinae). ZooKeys 192: 49-78. doi: 10.3897/zookeys.193.2768
Kugler J (1988) The zoogeography of social insects of Israel and Sinai. Pp. 251-275. In: Yom-Tov Y, Tchernov E (Eds) The zoogeography of Israel. Dr. W. Junk, Dordrecht. 600pp.
Menozzi C (1936) Nuovi contributi alla conoscenza della fauna delle Isole italiane dell'Egeo. VI. Hymenoptera - Formicidae. Boll. Lab. Zool. Gen. Agrar. R. Sc. Super. Agric. 29: 262-311. Nayman J (1972) Atlas of Wildlife. Heinenmann, London, 124 pp.
Radchenko AG, Alipanah H (2004) The first record of the subfamily Aenictinae (Hymenoptera: Formicidae) from Iran. Vestnik zoologii, 38(4): 75-78.
Rosciszewski K, Maschwitz U (1994) Prey specialization of army ants of the genus Aenictus in Malaysia. Andrias 13, 179-187.
Sharaf MR (2006) Taxonomic and ecological studies on family Formicidae (Order: Hymenoptera) in Egypt including some protectorates with a study of some insect fauna associated with ant species. Ain Shams University, Faculty of Science, Entomology Department, Cairo (unpubl. thesis), 340 pp .
Sharaf MR, Aldawood AS, Taylor B (2012) A new ant species of the genus Tetramorium Mayr, 1855 (Hymenoptera: Formicidae) from Saudi Arabia, Including a revised key to the Arabian species. PLoS one 7 (2) e30811: (1-9).
Shattuck SO (2008) Review of the ant genus Aenictus (Hymenoptera: Formicidae) in Australia with notes on A. ceylonicus (Mayr). Zootaxa 1926, 1-19.

Taylor B (2012) The ants of (sub-Saharan) Africa. Aenictus catalogue. http://www.webarchive. org.uk/wayback/archive/20101217230548/http://antbase.org/ants/africa/aenictus/aenictus/aenictus.htm [accessed January 2012]
Terayama M (2009) A synopsis of the Family Formicidae of Taiwan (Insecta, Hymenoptera). Liberal Arts, Bulletin of Kanto Gakuen University 17, 81-266.
Wilson EO (1964) The true army ants of the Indo-Australian area (Hymenoptera: Formicidae: Dorylinae). Pacific Insects 6: 427-483.
Zettel H, Sorger DM (2010) Three new species of the army ant genus Aenictus Shuckard, 1840 (Hymenoptera: Formicidae: Aenictinae) from Borneo and the Philippines. Zeitschr. der Arbeitsgemeinschaft österr. Entomologen 62, 115-125.
Zhou S (2001) Ants of Guangxi. Guangxi Normal University, Guilin.

# A new genus of the tribe Issini (Hemiptera, Fulgoromorpha, Issidae) from China 

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#### Abstract

Macrodarumoides petalinus, a new genus and species of Issidae (Hemiptera) are described based on specimens from Yunnan and Guangxi, China.


## Keywords

Taxonomy, Fulgoromorpha, Macrodaruma, new species

## Introduction

Issidae are small insects (about 5 mm to 20 mm ) generally with a stocky body and usually brownish in color, few bright; and some have pronounced maculae. Issid planthoppers and ladybirds exhibit considerable similarity in general especially in the stocky body form. Issids are all plant feeders, with both nymphs and adults intaking phloem fluid from young branches and sometimes roots. Feeding of issids can result in the yellowing of plant foliage. At high population, issids can seriously affect plant
growth (personal observation). Although they often have developed wings, some of them like to crawl and jump in shrubs, rather than to fly (personal observation). The family includes 973 species in 155 genera (Gnezdilov 2010).

Gnezdilov (2009) considered that the presence of the trilobed hind wing was an insufficient character for defining the tribe Thioniini, and therefore treated Thioniini Melichar, 1906 as a junior synonym of Issini Spinola, 1839. According to Gnezdilov (2003a, 2009), the subfamily Issinae consists of four tribes: Hemisphaeriini Melichar; Parahiraciini Cheng \& Yang; Issini Spinola and Colpopterini Gnezdilov. In the present paper, a new genus and species, Macrodarumoides petalinus gen. et sp. n., from China is described and illustrated. Based on the hemispherical body, the claval suture on tegmen, the trilobed hind wings and the not-dilated legs, Macrodarumoides gen. n. is placed in the tribe Issini, which has the only species Macrodarumoides petalinus sp. n. from China.

## Materials and methods

The terminology of the head, body and male genitalia follows Chan and Yang (1994), and the terminology of the female genitalia follows Gnezdilov (2003b). The genital segments of the examined specimens were macerated in $10 \% \mathrm{KOH}$ and observed in glycerin jelly using a Leica MZ125 stereomicroscope. Photographs of the specimens were made using a Nikon SMZ1500 stereomicroscope with a Q-image CCD. Images were produced using the software Synoptics Automontage. All the specimens studied are deposited in the Entomological Museum of Northwest Agriculture and Forestry University of (NWAFU) or the College of Life Sciences, Nankai University (NKU), as indicated.

## Macrodarumoides gen. n.

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http://species-id.net/wiki/Macrodarumoides

## Type species. Macrodarumoides petalinus sp. n.

Description. Head (including eyes) distinctly narrower than pronotum (Fig. 1). Vertex long and horizontal, approximately triangular, disc depressed, with 2 depressions near hind margin; anteriorly strongly angularly convex and posteriorly slightly angularly concave, lateral margins carinate; width at apex distinctly shorter than length in midline (Fig. 1). In dorsal view, vertex and frons extending far beyond eyes (Fig. 1). Ocelli present. Frons long and nearly triangular, disc obviously elevated, median carina present (Fig. 2); in lateral view frons curved towards apex (Fig. 3). Clypeus elevated with central carina, situated the same plane as frons (Fig. 2). Rostrum long, reaching to hind-trochanter. Pronotum short laterally, anterior margin convex and arched, posterior margin horizontal to slightly convex; disc elevated with pits (Fig. 1). Mesonotum nearly triangular, with 2 pits along lateral margins; disc slightly elevated, with or without carina (Fig. 1). Tegmen (Figs 1, 3, 7) leathery and approxi-


Figures I-6. Macrodarumoides petalinus sp. n., male I Holotype, dorsal view 2 Holotype, head, ventral view 3 Holotype, lateral view; Macrodaruma pertinax Fennah, male: 4 Dorsal view 5 Head, ventral view 6 Lateral view.
mately elliptical, claval suture present; longitudinal veins prominent, between them with a number of obscure veinlets, rendering the whole surface faintly reticulate. Wing (Fig. 8) large, veins distinct and netlike, longer than half of tegmen, apically forming 3 lobes. Legs (Fig. 3) relatively long, not dilated; lateral margin of hind tibia with 2 teeth. Spinal formula of hind leg (5-6)-11-2, indicating number of spines at apex of hind tibia and tarsomeres I and II.

Male genitalia symmetrical; anal segment (Fig. 10) in dorsal view longer than greatest width; pygofer (Fig. 9) without spines; aedeagus (Figs 11-13) tubular, symmetrical and shallowly U-shaped, divided distally into a dorsal and ventral lobe, the latter sometimes split, a pair of elongate lateral lobes and a pair of spiniform processes lying ventrolaterally and directed either cephalad or caudad. Genital style (Fig. 9) subtrianglar, apical margin curved and arched, basal margin convex near apex and dorsal margin produced into a single process.

Female genitalia with anal segment (Fig. 15) in dorsal view elliptical, length nearly equal to the widest part. Apex of endogonocoxal process without lobe and anterior connective lamina of gonapophyse VIII with 3 teeth in lateral group. First valvula (Fig. 14) with teeth, ninth tergum and third valvula subquadrate. Pregenital sternite (Fig. 16) with apical margin convex at mid.

Diagnosis. This genus is similar to Eusarima Yang, 1994 according to body shape and trilobed wings, but can be differentiated by the following characteristics: 1) vertex long, approximately triangular, the latter with vertex short and subquadrangular; 2) wings narrow, netlike and anal lobe rudimentary, the latter, wings broad, longitudinal veins distinct with little cross veins, and anal lobe developed.

This genus resembles Macrodaruma Fennah, 1978 (Figs 4-6), but can be distinguished from the latter by: 1) lateral margins of vertex and anterior margin of pronotum not carinated, the latter, distinctly and foliately carinated; 2) tegmen with claval suture present, the latter, claval suture absent; 3) wing trilobed, the latter, not bilobed.

Etymology. This generic name "-oides" from Greece suffix refers to the resemblance of this genus to Macrodaruma Fennah. The gender is masculine.

Distribution. China (Guangxi, Yunnan).

## Macrodarumoides petalinus sp. n.

urn:lsid:zoobank.org:act:6D96BEEB-3639-41BC-88EA-43A0FD893343
http://species-id.net/wiki/Macrodarumoides_petalinus
Figs 1-3, 7-16

Description. Length, male (including tegmen): 8.1 mm , length of tegmen: 6.0 mm ; female (including tegmen): 8.3 mm , length of tegmen: 6.1 mm .

Body brown (Fig. 1). Vertex with disc brown and lateral margins black. Eyes black brown. Frons brown with median carina paler and lateral margin black at apical half. Clypeus brown, rostrum pale brown. Pronotum, mesonotum, tegmen and legs brown; wing dark brown. Abdomen ventrally and dorsally brown, disc dark brown.

Vertex long and horizontal, approximately triangular, disc depressed, with 2 despressions near hind margin (Fig. 1); vertex $1.8 \times$ wider at apex than length in midline. Frons with disc distinctly elevated, with median carina (Fig. 2); oblique between median carina and lateral margin, frons curved towards apex in lateral view (Fig. 3), 1.1× wider at widest part than at base, $2.8 \times$ longer in midline than at widest part. Frontoclypeal suture nearly straight (Fig. 2). Thorax (Fig.1): disc of pronotum with 2 pits; mesonotum short and broad, greatest width $1.7 \times$ medial length. Tegmen $2.5 \times$ longer than widest part; Sc long, reaching beyond midlength; Sc and R forked near apex, M 4-branched, Cu not forked, claval suture only reaching to middle of sutural margin (Figs $1,3,7$ ). Wing $0.7 \times$ length of tegmen, veins distinctly reticulate (Fig. 8). Spinal formula of hind leg 11-(5-6)-2.

Male genitalia. Anal segment in dorsal view (Fig. 10) distinctly longer than greatest width apically, apical margin strongly convex, lateral margin strongly divergent from base to apex, anal tube situated near middle; in lateral view ventral margin convex at midlength, nearly truncate in distal half (Fig. 9). Pygofer in lateral view with hind margin evenly convex near middle and slightly convex at base (Fig. 9). Phallus in profile shallowly curved with 2 long spiniform processes directed cephalad at midlength. Aedeagus in profile with apex bifurcated (Fig. 11); dorsal lobe (Figs 11, 13) in dorsal


Figures 7-16. Macrodarumoides petalinus sp. n. 7 Tegmen $\mathbf{8}$ Wing 9 Male genitalia, left side 10 Male anal segment, dorsal view II Aedeagus, lateral view $\mathbf{I 2}$ Apex of aedeagus, ventral view $\mathbf{1 3}$ Apex of dorsal lobe, dorsal view 14 Female genitalia, left side 15 Female anal segment, dorsal view 16 Pregenital sternite, ventral view. Scale bars $=1.0 \mathrm{~mm}$ (Figs 7-8), 0.5 mm (Figs 9-16)
view tri-lobed near apex, with apical margin slightly concave at mid, lateral margins curved downward and encasing lateral and ventral lobe; lateral lobes in lateral view divided and tapered into fingers (Fig. 11); ventral lobe with apical margin convex and arched at mid, and lateroapical angle rounded in ventral view (Fig. 12). Genital styles in lateral view nearly triangular, apical margin curved and arched, dorsal margin produced into one large process near apex; base of process acuminate and apex obtusely rounded in caudal view (Fig. 9).

Female genitalia. Anal segment in dorsal view (Fig. 15) slightly longer than greatest width with lateral margins convex, apical margin convex and arched, anal tube situated at basal half; in lateral view ventral margin concave at base, nearly truncate in distal half (Fig. 14). Ovipositor with anterior connective lamina of gonapophyse VIII
curved dorsally, with 5 nearly parallel spines; tooth near lateral margin larger. Gonoplac stout, strongly convex and subquadrate, with apical margin polished (Fig. 14). Pregenital sternite with apical margin distinctly convex at midlength (Fig. 16).

Material examined. Holotype, male, China: Yunnan, Mt. Baoshan, 1900m, 20 November 1999, coll. Qin Daozheng (NWAFU). Paratypes, one male, one female, China: Guangxi, Leye, Tonglelinchang, 15 September 1980, coll. Lu Junsheng (NWAFU); one female China: Guangxi, Leye, Yachangyanpeng, 24 September 1980, coll. Lu Junsheng (NWAFU); one female, same data as holotype (NWAFU); two males, one female, China: Yunnan, Mt. Baoshan, 22 August 1979, coll. Cui Jianxin (NKU).

Remarks. This species can be differentiated from Eusarima contorta Yang, 1994 by the following characteristics: 1) vertex long, approximately triangular, the latter with vertex short and subquadrangular; 2) frons only with median carina, the latter with median and lateral carinae; 3) wings narrow, netlike and anal lobe rudimentary, the latter, wings broad, longitudinal veins distinct with little cross veins, and anal lobe developed.

This species resembles Macrodaruma pertinax Fennah, 1978 (Figs 4-6) in shape, but differs from the latter in the following characteristics: 1) lateral margins of vertex and lateral margins of pronotum not elevated, the latter with lateral margins of vertex and lateral margins of pronotum elevated foliately, 2) claval suture present, the latter without claval suture, 3) wing with 3 lobes, the latter with wing not split.

Etymology. The specific name is derived from the Latin word "petalinus", referring to the dorsal lobe of aedeagus being concave with the lateral margin distinctly reflected as a petal.

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## Reference

Chan ML, Yang CT (1994) Issidae of Taiwan (Homoptera: Fulgoroidea). Chen Chung Book, Taichung, 188 pp .
Fennah RG (1978) Fulgoroidea (Homoptera) from Vietnam. Annales Zoologici 34 (9): 207-279.
Gnezdilov VM (2003a) A new tribe of the family Issidae with comments on the family as a whole (Homoptera: Cicadina). Zoosystematica Rossica 11: 305-309.

Gnezdilov VM (2003b) Review of the family Issidae (Homptera: Cicadina) of the European fauna with notes on the structure of ovipositor in the planthoppers. Chteniya Pamyati Nikolaya Aleksandrovicha Kholodkovskogo 56(1): 1-145.
Gnezdilov VM (2009) Revisionary notes on some tropical Issidae and Nogodinidae (Hemiptera: Fulgoroidea). Acta Entomological Musei Nationalis Pragae 49: 75-92.
Gnezdilov VM (2010) Composition and distribution of the family Issidae (Hemiptera, Fulgoroidea). 13th International Auchenorrhyncha congress, Vaison-la-Romaine, France 28 June - 2 July 2010. Abstracts: talks and posters: 87-88.

# A review of the genus Megapulvinaria Young (Hemiptera, Coccoidea, Coccidae) from China, with a description of a new species 

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#### Abstract

Prior to this study, only Megapulvinaria maxima (Green) was known from China. However, a new species M. beibaiensis Wang \& Feng, sp. n. is described below and M. maxima is redescribed. A key is provided for the five species now placed in this genus.


## Keywords

Hemiptera, Coccoidea, soft scale, taxonomy, China

## Introduction

Soft scale or Coccidae is the third largest family after Diaspididae and Pseudococcidae within the superfamily Coccoidea (Ben-Dov 2012). Soft scale insects have a world-wide distribution and many of them are important pests on agricultural and horticultural crops and amenity plantings (Henderson and Hodgson 2005). China has a fauna of soft scale insects with a total of at least 125 species belonging to 46 genera (Tang 1991, Tao 1999, Wu 1999, Martin and Lau 2011). Some species that occur in China, such as Ceroplastes rubens and C. japonicus, can cause deformation or death of plants shoots and lead to great economic losses due to their feeding. However, some
species present in China can be considered beneficial, such as Ericerus pela, whose wax provides an important raw material for many industries (Tang 1991).

The genus Megapulvinaria was erected by Young (1982), with Pulvinaria maxima Green, 1904 as its type species, and belongs to the tribe Pulvinariini, subfamily Coccinae. Three more species M. burkilli (Green, 1908), M. orientalis (Reyne, 1963) and M. maskelli (Olliff, 1891) have been subsequently added (Avasthi and Shafee 1991, Ben-Dov 1993, Hodgson 1994).

Previously, only $M$. maxima was known from China but a new species has now been discovered. The adult female of $M$. maxima is redescribed, the adult female of the new species $M$. beihaiensis Wang \& Feng sp. n. is described and a key is provided for separation of the five species now known in this genus.

## Materials and methods

Specimens were slide mounted using the method recommended by Hodgson and Henderson (2000). The morphological terminology of the mounted specimens used in the descriptions mainly follows Hodgson (1994). Characters were examined under a Nikon microscope. Illustrations were drawn from mounted adult female specimens, with the dorsum depicted on the left side and the venter on the right side, and with enlargements of important characters shown around the main illustration. All measurements were given in micrometers ( $\mu \mathrm{m}$ ) or millimeters ( mm ).

All specimens are deposited in the Entomological Museum of Northwest A \& F University, Yangling, Shaanxi, China (NWAFU).

## Checklist of known species of the genus Megapulvinaria Young

Megapulvinaria maxima (Green, 1904); China (Guangxi, Yunnan, Taiwan), Thailand, India, Indonesia, Philippines, Sri Lanka, Vietnam, Papua New Guinea, Chuuk Islands. Megapulvinaria burkilli (Green, 1908); India.
Megapulvinaria orientalis (Reyne, 1963); Thailand.
Megapulvinaria maskelli (Olliff, 1891); Australia.
Megapulvinaria beihaiensis sp. n.; China (Guangxi).

## Taxonomy

Genus Megapulvinaria Young, 1982
http://species-id.net/wiki/Megapulvinaria
Megapulvinaria Young, 1982: 162. Type species: Pulvinaria maxima Green, 1904. By original designation and monotype.

Generic diagnosis. Adult female. Body elongate oval to broad oval; stigmatic clefts distinct. Dorsum. Dorsal setae spinose or conical. Dorsal submarginal tubercles absent. Preopercular pores present or absent. Dorsal tubular ducts present or absent. Eyespots generally displaced onto dorsum (marginal on M. maxima). Anal plates together quadrate, each plate with 2 spinose and/or truncate setae along inner margin, a similar seta on apex and a spinose seta present in discal position (possibly on outer margin of M. maskelli). Anal ring with 6 setae. Margin. Marginal setae stout, apex truncate or bidentate, and with 2 types present, one shorter and broader than other (about same length and one slightly broader than other both in $M$. maskelli and $M$. beihaiensis); broader setae on head and posterior margins of abdomen ( $0-3$ broader setae present between two stigmatic clefts in $M$. beihaiensis). Stigmatic clefts deep or shallow, each with 3-12 stigmatic spines. Venter. Antennae 7-9 (mostly 8) segmented. Legs well-developed, each with a tibio-tarsal articulation and an articulatory sclerosis, each claw with a denticle on the widest part. Pregenital setae 2 pairs. Spiracular disc-pores each mainly with 5 loculi. Pregenital disc-pores each mainly with 10 loculi, restricted to abdominal segments. Ventral tubular ducts of three types, with a submarginal band of small tubular ducts; median area of head, thorax, and anterior 1-3 abdominal segments with large ducts each with both outer and inner ductules broad or stout (anterior submargin and all median area in M. maskelli); posterior abdominal segments of moderately tubular ducts.

Distribution. Oriental and Australian regions.

## Key to all adult females of Megapulvinaria

| 1 | Dermal areolations absent.................................................................................................................................... 3 |
| :---: | :--- |

## Megapulvinaria maxima (Green, 1904)

http://species-id.net/wiki/Megapulvinaria_maxima
Figure 1
Pulvinaria maxima Green, 1904: 206.
Pulvinaria thespesiae Green, 1909: 259. Syn. by Takahashi 1935: 10.
Eriochiton formosae Takahashi, 1929: 64. Syn. by Takahashi 1935: 10.
Megapulvinaria maxima (Green), Young 1982: 162.

Material examined. 5 adult females, CHINA, Yunnan, Jingdong, 18. x. 1976 on Pigeonpea (Cajanus cajan (L.) Millsp., Leguminosae), Xiao-Ze Chen (NWAFU).

Note. The measurements are based on all 5 specimens.
Diagnosis. Adult female. Mounted material. Body elongate oval, about 4.2-6.2 mm long and $2.7-3.8 \mathrm{~mm}$ wide. Anal cleft approximately $1 / 7$ of the body length. Stigmatic clefts deep.

Dorsum. Derm membranous. Dermal areolations well developed, each with 1 or 2 dorsal microducts. Dorsal setae conical, with a well-developed basal socket, each 8-16 $\mu \mathrm{m}$ long, scattered throughout. Dorsal simple pores each with a slightly sclerotized margin, randomly distributed. Dorsal microducts each with a very short outer ductule and a longer, fairly broad inner filamentous ductule, sparsely located in dorsal areaolations. Dorsal tubular ducts each with a short outer ductule and a fine inner ductule with a minute terminal gland, sparsely distributed. Preopercular pores absent. Anal plates together quadrate; posterior margin slightly longer than anterior margin, outer angle slightly obtuse; each plate with a large cylindrical seta in discal position, each 34-50 $\mu \mathrm{m}$ long, a large spatulate seta apically, each $52-64 \mu \mathrm{~m}$ long, and with 2 spinose and/ or spatulate setae along posterior $1 / 3$ rd of inner margin, each $40-56 \mu \mathrm{~m}$ long. Anogenital fold with 1 pair of long setae and 1 pair of short setae along anterior margin and 2 or 3 pairs lateral margin. Anal ring subcircular, with 2 or 3 rows of translucent pores and 6 anal ring setae. Eyespots present some way onto dorsum, each $80-96 \mu \mathrm{~m}$ wide.

Margin. Marginal setae of 2 types: 1) large and stout setae, each $17-38 \mu \mathrm{~m}$ long, with nearly parallel sides, and with either a truncate or a bifid apex, all with well-developed basal sockets, each socket with 1 or 2 small pores; with 96-110 setae between anterior clefts, $36-46$ setae on each side between stigmatic clefts, and $84-98$ setae between each posterior stigmatic cleft and anal cleft; and 2) quite broad and short setae, each 14-24 $\mu \mathrm{m}$ long, with parallel sides and a truncate, flattened apex, and with a larger basal socket about twice as broad as that of type 1), each socket with 3-8 small pores; latter type of marginal setae only distributed on anterior and posterior ends, with 16-22 setae anteriorly on head and prothorax, 5-12 setae on either side of abdomen near anal cleft. Stigmatic clefts deep; stigmatic spines bluntly spinose and mostly straight, with 4-8 spines in each anterior cleft and 5-10 in each posterior cleft; length of each $42-96 \mu \mathrm{~m}$, with median 1-3 spines much longer than the lateral spines.

Venter. Derm membranous. Antennae 8 segmented, each $505-586 \mu \mathrm{~m}$ long; third segment longest; with 2 pairs of long setae and 1-3 pairs of short interantennal setae. Clypeolabral shield 198-232 $\mu \mathrm{m}$ long, 205-240 $\mu \mathrm{m}$ wide; labium 90-106 $\mu \mathrm{m}$ long, 113-144 $\mu \mathrm{m}$ wide. Legs well-developed, each with a tibio-tarsal articulation and articulatory sclerosis; claws with a denticle on widest part, claw digitules broad and expanded apically, tarsal digitules slender, knobbed and longer than claw digitules; trochanter+femur 239-405 $\mu \mathrm{m}$, tibia $180-245 \mu \mathrm{~m}$ and tarsus $96-122 \mu \mathrm{~m}$. With 2 pairs of long pregenital setae present in both segments VI \& VII; submarginal setae present in a single row; other setae slender, each $4-10 \mu \mathrm{~m}$ long, quite sparsely distributed. Spiracles normal, spiracular disc-pores each with 5 loculi, present in a broad band between stigmatic cleft and each spiracle. Pregenital disc-pores each mainly with


Figure I. Adult female of $M$. maxima $G$ reen, $\mathbf{A}$ body derm BI, B2 two kinds of marginal setae $\mathbf{C}$ stigmatic spine $\mathbf{D}$ dorsal microduct $\mathbf{E}$ dorsal pore $\mathbf{F}$ dorsal tubular duct $\mathbf{G}$ anal plates $\mathbf{O}$ ano-genital fold $\mathbf{H}$ pregenital disc-pore $\mathbf{I}$ tibio-tarsus of hind leg JI, J2, J3 ventral tubular ducts $\mathbf{K}$ spiracle disc-pore $\mathbf{L}$ ventral microduct $\mathbf{M}$ antenna $\mathbf{N}$ dorsal seta.

10 loculi, present around the vulva and on posterior 4 abdominal segments. Ventral microducts scattered. Ventral tubular ducts of 3 types present: 1) a duct with a short outer ductule and a fine inner filament, with a minute terminal gland, present in a
complete submarginal band; 2) a duct with outer and inner ductules both broad and with a well-developed terminal gland, present medially on head, thorax and anterior 1 or 2 abdominal segments; and 3) a duct with a moderately long outer ductule and a thin inner ductule slightly longer than outer ductule, with a flower-shaped terminal gland, present medially on posterior abdominal segments and extending and mingling with marginal band of type 1) ducts.

Distribution. China (Guangxi, Yunnan, Taiwan), Thailand, India, Indonesia, Philippines, Sri Lanka, Vietnam, Papua New Guinea, Chuuk Islands.

Comments. Since Green (1904) originally described M. maxima, it had been described and illustrated by many authors, such as Green (1904, 1909), Takahashi (1929), Tang (1991), Hodgson (1994). Our observations agree well with these in descriptions in most respects. Tang (1991) and Hodgson (1994) pointed out the presence of dorsal areolations and denticles on widest part of claws, which Green (1909) and Takahashi (1929) failed to observe respectively. The outer angles of anal plates were obtuse or broadly rounded rather than at a right-angle, as shown by Hodgson (1994). We have confirmed the occurrence of dorsal areolations and denticles, and the outer angles are slightly obtuse in our examined specimens. Moreover, Hodgson (1994) described four types of dorsal pores, but we just observed just two types.

This species is close to M. burkilli (Green) (data from Green, 1908), but it can be distinguished from the latter by the following features (character states of M. burkilli in brackets): (1) the much larger body size in comparison to the latter ( 4 mm long, 2 mm wide); and (2) with well-developed dermal areolations present (absent).

## Megapulvinaria beibaiensis Wang \& Feng, sp. n.

urn:lsid:zoobank.org:act:56E4CA5F-6C56-431C-AD2B-6A54776BC16B
http://species-id.net/wiki/Megapulvinaria_beihaiensis
Figure 2

Material examined. Holotype: adult female. CHINA, Guangxi, Beihai, Haibin Park. 26. vii. 2010, on Cinnamomum sp., (Lauraceae), Bin Zhang (NWAFU)

Paratypes. 3 adult females, the data same as holotype.
Note. The measurements are based on all 4 specimens.
Description. Adult female. Unmounted material. Adult female yellowish brown or dark brown, elongate oval and with a longitudinal dorsal ridge in dorsal straight median area (materials examined were all immersed in $75 \%$ ethanol, and the ovisac was not seen). The specimens collected on the lamina of the host plant.

Mounted material. Body elongate oval, about 2.1-3.2 mm long, $1.3-1.7 \mathrm{~mm}$ wide. Anal cleft approximately $1 / 8$ of the body length. Stigmatic clefts deep.

Dorsum. Derm membranous. Dermal areolations well-developed, each with a dorsal microduct. Dorsal setae conical, with a well-developed basal socket, each 6-11 $\mu \mathrm{m}$ long, scattered throughout. Dorsal simple pores each with a slightly sclerotized margin, randomly distributed. Dorsal microducts each with a very short outer duct-


Figure 2. Adult female of $M$. beihaiensis sp. n., A body derm BI, B2 two kinds of marginal setae $\mathbf{C}$ stigmatic spine $\mathbf{D}$ dorsal microduct $\mathbf{E}$ dorsal pore $\mathbf{F}$ dorsal tubular duct $\mathbf{G}$ anal plates $\mathbf{O}$ ano-genital fold $\mathbf{H}$ pregenital disc-pore $\mathbf{I}$ tibio-tarsus of hind $\operatorname{leg} \mathbf{J} \mathbf{I}, \mathbf{J} \mathbf{2}, \mathbf{J} \mathbf{3}$ ventral tubular ducts $\mathbf{K}$ spiracle disc-pore $\mathbf{L}$ ventral microduct $\mathbf{M}$ antenna $\mathbf{N}$ dorsal seta.
ule and a long, fairly broad inner filamentous ductule, sparsely located in each dorsal areaolation. Dorsal tubular ducts each with a short outer ductule and a fine inner ductule with a minute terminal gland, sparsely distributed. Preopercular pores absent.

Anal plates together quadrate, dorsal surface with reticulations on anterior two-thirds; posterior margin subequal to or slightly longer than anterior margin, outer angle a right-angle; each plate with a blunt spinose seta in discal position, each $34-42 \mu \mathrm{~m}$ long, a large spinose or spatulate seta apically, each $48-54 \mu \mathrm{~m}$ long, and with 2 spinose setae along posterior $1 / 3 \mathrm{rd}$ of the inner margin, each $32-44 \mu \mathrm{~m}$ long, length of plates 146-167 $\mu \mathrm{m}$, width of single plate $74-88 \mu \mathrm{~m}$. Ano-genital fold with 1 pair of long setae and 1 pair of short setae along anterior margin and 2 or 3 pairs lateral margin. Anal ring subcircular, with 2 or 3 rows of translucent pores and 6 anal ring setae. Eyespots present some way onto dorsum, each $42-60 \mu \mathrm{~m}$ wide.

Margin. Marginal setae of 2 types: 1) stout setae, each 18-30 $\mu \mathrm{m}$ long; each seta with nearly parallel sides and with either a truncate or a bifid apex, all with well-developed basal sockets, each socket with 1 or 2 small pores; with $101-111$ setae between anterior clefts, 34-42 setae on each side between stigmatic clefts, and $74-85$ setae between each posterior stigmatic cleft and anal cleft; 2) quite strong setae, subequal in length with type 1) but slightly broader; each seta with parallel sides, with a truncate and flattened apex, and with a large basal socket about twice as broad as that of type 1), each socket with $2-8$ small pores; with 10-16 setae anteriorly on head and prothorax, $0-3$ setae between stigmatic clefts, and $4-10$ setae on either side of abdomen near anal cleft. Stigmatic clefts deep; stigmatic spines bluntly spinose and mainly curved apically, with 4 or 5 spines in each anterior cleft and 5-8 spines in each posterior cleft; length of each 34-62 $\mu \mathrm{m}$, and the median 1-3 spines longer than the lateral spines.

Venter. Derm membranous. Antennae 8 segmented, each 346-378 $\mu \mathrm{m}$ long, the third segment longest; with 2 pairs of long setae and 2 or 3 pairs of short interantennal setae. Clypeolabral shield $138-160 \mu \mathrm{~m}$ long, $160-172 \mu \mathrm{~m}$ wide; labium $96-112 \mu \mathrm{~m}$ long, $84-112 \mu \mathrm{~m}$ wide. Legs well-developed, each with a tibio-tarsal articulation and articulatory sclerosis; claws with a denticle on widest part, claw digitules both broad and expanded apically; tarsal digitules slender, knobbed and longer than claw digitules; trochanter+femur $212-245 \mu \mathrm{~m}$, tibia $136-188 \mu \mathrm{~m}$ and tarsus $54-75 \mu \mathrm{~m}$. With 2 pairs of long pregenital setae present in both segments VI \& VII; submarginal setae present in a single row; other setae slender, 6-20 $\mu \mathrm{m}$ long, quite sparsely distributed. Spiracles normal; spiracular disc-pores each mainly with 5 loculi, present in a broad band between stigmatic cleft and each spiracle. Pregenital disc-pores each mainly with 10 loculi, present around the vulva and on posterior 5 abdominal segments but becoming progressively less frequent anteriorly. Ventral microducts scattered. Ventral tubular ducts of 3 types present: 1) a duct with a short outer ductule and a fine inner filament with a minute terminal gland, present in a complete submarginal band; 2) a duct with a broad outer ductule, a stout inner ductule (as broad as outer ductule in some specimens) and with a welldeveloped terminal gland, present medially on thorax and anterior abdominal segments; and 3) a duct with a moderately long outer ductule, a thin inner ductule slightly longer than outer ductule, with a flower-shaped terminal gland, present medially on posterior abdominal segments and extending and mingling with marginal band of type 1) ducts.

Distribution. China (Guangxi).
Etymology. The specific epithet is taken from the type locality Beihai.

Comments. This new species resembles M. maskelli (Olliff) (data from Qin and Gullan 1992) in having: (1) dorsal reticulations on the anal plates, and (2) 2 types of marginal setae of about same length. However, M. beihaiensis can be distinguished by following features (character states of M. maskelli in brackets): (1) more than 3 stigmatic spines in each stigmatic cleft (only 3); (2) having dorsal tubular ducts (absent); (3) lacking preopercular pores (present); (4) eyespots displaced onto the dorsum (on the margin); and (5) the marginal setae of much broader basal socket often present between stigmatic clefts (absent).
M. maskelli, currently only known from the Australian region, is the only nonOriental species in this genus and has some distinctive characteristics within Megapulvinaria. It differs from other species in having: (1) only 3 stigmatic spines in each stigmatic cleft; (2) eyespots located on margin; and (3) the discal setae possibly on outer margin of anal plates.

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## References

Avasthi RK, Shafee SA (1991) Classification of Indian Coccidae (Homoptera: Coccoidea). Indian Journal of Systematic Entomology 8: 7-26.
Ben-Dov Y (1993) A Systematic Catalogue of the Soft Scale Insects of the World (Homoptera: Coccoidea: Coccidae) with data on geographical distribution, host plants, biology and economic importance. Flora \& Fauna Handbook, No. 9. Sandhill Crane Press, Gainesville, 536 pp.
Ben-Dov Y (2012) ScaleNet: a database of the scale insects of the world. http://www.sel.barc. usda.gov/scalenet/scalenet.htm [accessed 18 March 2012]
Green EE (1904) On some Javanese Coccidae: with descriptions of new species. Entomologist's Monthly Magazine 40: 204-210.
Green EE (1908) Remarks on Indian Scale Insects (Coccidae), Part III. With a catalogue of all species hitherto recorded from the Indian continent. Memoirs of the Department of Agriculture in India, Entomology Series 2: 15-46.
Green EE (1909) The Coccidae of Ceylon, Part IV. Dulau \& Co., London, 250-344.
Henderson RC, Hodgson CJ (2005) Two new species of Umbonichiton (Hemiptera: Sternorrhyncha: Coccoidea: Coccidae) from New Zealand. Zootaxa 854: 1-11.
Hodgson CJ (1994) The Scale Insect Family Coccidae: An Identification Manual to Genera. CAB International, Wallingford, Oxon, UK, 639 pp.

Hodgson CJ, Henderson RC (2000) Coccidae (Insecta Hemiptera: Coccoidea). Fauna of New Zealand 41: 1-264.
Martin JH, Lau CSK (2011) The Hemiptera-Sternorrhycha (Insecta) of Hong Kong, China-an annotated inventory citing voucher specimens and published records. Zootaxa 2847: 1-122.
Olliff AS (1891) A new scale insect destroying saltbush. Agricultural Gazette of New South Wales 2: 667-669.
Qin TK, Gullan PJ (1992) A revision of the Australian pulvinariine soft scales (Insecta: Hemiptera: Coccidae). Journal of Natural History 26: 103-164. doi: 10.1080/00222939200770061
Reyne A (1963) Scale insects from Thailand with description of a Filippia n. sp. Beaufortia. Amsterdam 10: 29-39.
Takahashi R (1929) Observations on the Coccidae of Formosa, Part I. The Institute Taihoku, Formosa 40: 1-82.
Tang FT (1991) The Coccidae of China. Shanxi United Universities Press, Taiyuan, China, 377pp. [In Chinese]
Tao CCC (1999) List of Coccoidea (Homoptera) of China. Taiwan Agricultural Research Institute 78: 1-176.
Wu SA (1999) A new species of the genus Scythia (Homoptera: Coccidae) from North China. Entomotaxonomia 21 (2): 115-117.
Young PL (1982) General classification of scale insects in China. Shanghai Science \& Technology Press, Shanghai, China, 425 pp. [In Chinese]

# Dental morphology of the Lesser Bamboo Rat, Cannomys badius (Rodentia, Spalacidae) 

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#### Abstract

Cannomys and Rhizomys are the sole living genera of the tribe Rhizomyini (Rhizomyinae, Spalacidae, Rodentia), known in the fossil record since the Late Miocene. The dental morphology of fossil Rhizomyini has been described in detail but until recently such descriptions were unavailable for extant species. A detailed account of the morphology and dental wear pattern of the cheek teeth of $C$. badius is provided here based on the examination of museum specimens. Three stages of wear are recognized. Cannomys shares with Rhizomys the synapomorphy of having a mesolophid that is a long continuation of the protoconid on the first lower molar. There are significant differences between these taxa, such as the much smaller size of the cheek teeth and the trilophodont dental pattern of the M2, M3, and m2 in Cannomys.


## Keywords

Rhizomyinae, Rhizomyini, Cannomys, dental wear pattern

## Introduction

The subfamily Rhizomyinae (Spalacidae, Rodentia) is known in the fossil record since the Oligocene. It is represented by three modern genera: the Asian bamboo rats Rhizomys (a trispecific genus) and Cannomys (monospecific) and the African mole rats Tachyoryctes. The latter genus is considered bispecific by some authors (Missone 1971, Rahm 1980, Bucher 1982, Corbet and Hill 1991, Beolchini and Corti 2004), but this
may be an underestimation of true diversity (Allen 1939, Ellerman 1941, Musser and Carleton 1993, 2005). The Asian bamboo rats belong to the tribe Rhizomyini, whereas Tachyoryctes is the only extant representative of the Tachyoryctini (Flynn 2009; LópezAntoñanzas et al. in press). The lesser bamboo rat Cannomys badius ranges from eastern Nepal, through northeast India, Bhutan, southeastern Bangladesh, Myanmar, south China, northwest Vietnam, Thailand and Cambodia (Musser and Carleton 2005). It is known to live in a wide variety of habitats, from bamboo forest to cultivated land and other modified areas (IUCN 2010).

Cannomys badius was originally named as Rhizomys badius by Hodgson (1841; p. 60 ) on the basis of a male specimen obtained "some miles north of the great valley" of Nepal. The type specimen is BMNH 1843.1.12.61 (skin and skull) (P. Jenkins, pers. comm. 2012). Thomas (1915a) created the new genus Cannomys for which he took Rhizomys badius as type, but in which he also included two other species: C. castaneus (Blyth 1843) and C. minor (Gray 1842). Thomas (1915b) later described another species of Cannomys, C. pater, and split C. castaneus into two subspecies (C. castaneus castaneus and C. castaneus plumbescens). Later, Gyldenstolpe (1916) described a new subspecies of Cannomys minor (C. minor lönnbergi). All these taxa are currently regarded as synonyms of C. badius (Musser and Carleton 2005).

The dental morphology of all living Rhizomyinae other than Cannomys badius, has been described in detail recently (Tachyoryctes; López-Antoñanzas 2012) or are in the process thereof (Rhizomys). The aim of the present work is to provide an in-depth description of the dental morphology of Cannomys.

## Material and methods

All the specimens of Cannomys badius housed in the Laboratoire de ZoologieMammifères et Oiseaux of the Muséum national d'Histoire naturelle, Paris, France (MNHN) and in the Institut für Systematische Zoologie-Zoologische Sammlung of the Museum für Naturkunde, Berlin, Germany (ZMB) have been examined. MNHN individuals come from Thailand and Nepal, whereas those at ZMB are from Myanmar.

First, second, and third lower molars are designated as $\mathrm{m} 1, \mathrm{~m} 2$, and m 3 , respectively, and first, second, and third upper molars as M1, M2, and M3, respectively. The terminology used in the tooth descriptions follows the rodent dental terminology of Flynn (1982) with some adjustments (see Fig. 1).

## Dental morphology and wear pattern

Incisors of Cannomys badius are strongly proodont, flattened anteriorly, lack major ornamentation and have the enamel pigmented orange.

The upper molars of Cannomys badius show unilateral hypsodonty, with crowns higher lingually than labially. M1 has four roots, the anterolabial one being the


Figure I. Dental terminology used in this paper. Cannomys badius. a Left upper cheek teeth $\mathbf{b}$ Right lower cheek teeth. Scale bar equals 5 mm .
most developed. Its occlusal outline is square. In early wear (e.g., MNHN C.G. 2000-271; Fig. 2a), it has four transverse lophs (anteroloph, protoloph, mesoloph, and metaloph-posteroloph) and all labial and lingual sinuses are open. In later wear (e.g., ZMB 44769 and MNHN C.G. 2000-761; Fig. 2b-c), the number of lophs is reduced to three as the anteroloph and protoloph combine and join at the margin of the tooth, isolating two transversely elongated enamel lakes, whereas the lingual sinus remains open. The latter, narrow and short, is directed toward the middle enamel lake. The connection between the anterior and posterior parts of the tooth persists through wear.

The occlusal outline of M2 is square, with its posterior side more reduced than the anterior one. This tooth is much shorter than M1. In early wear (e.g., MNHN C.G. 1860-382; Fig. 2a), it has three transverse lophs (anteroloph-protoloph, mesoloph, and metaloph-posteroloph). The sinus is directed toward the anterosinus. All reentrants remain open. After moderate wear (e.g., ZMB 44769, Fig. 2b), the sinus becomes narrower and the anterosinus and posterosinus are closed-off, isolating two enamel lakes. Late in wear (e.g., MNHN C.G. 2000-761; Fig. 2c), the morphology


Figure 2. Dental wear pattern in Cannomys badius. a-c Upper molars: a Stage of wear 1, juvenile individual, left maxilla with M1-M3 in occlusal view (MNHN C.G. 1860-382) b Stage of wear 2, left maxilla with M1-M3 (ZMB 44769) c Stage of wear 3, left maxilla with M1-M3 in occlusal view (MNHN C.G. 2000-761). d-f Lower molars: d Stage of wear 1, juvenile individual, right hemimandible with m1-m3 in occlusal view (MNHN C.G. 1860-382), e Stage of wear 2, right hemimandible with m1-m3 in occlusal (ZMB 44768) f Stage of wear 3, left hemimandible with m1-m3 in occlusal (reversed) (MNHN C.G. 2000761). Scale bar equals 5 mm .
of M2 is quite similar to that in the previous wear stage. However, the posterior part of the tooth becomes more reduced and, even though the lingual sinus remains open, it turns out to be more anterolabially directed due to the labial displacement of the hypocone. The connection between the anterior and posterior parts of the tooth persists through wear.

Cannomys has a reduced M3. In early wear (e.g., MNHN C.G. 1860-382; Fig. 2a), it is morphologically similar to M2 but with its posterior part smaller due to the more labial position of the hypocone. This tooth is trilophodont, with anterolophprotoloph, mesoloph, and metaloph-posteroloph. After moderate wear (e.g., ZMB 44769; Fig. 2b), the anteroloph-protoloph is nearly connected to the mesoloph and the metaloph-posteroloph joins the mesoloph, isolating a labial circular enamel lake. The hypocone is much more labially displaced and the sinus much more anterolabi-
ally oriented. Therefore, the posterior part of the tooth becomes much reduced. In late wear (e.g., MNHN C.G. 2000-761; Fig. 2c), the first and second lophs combine and the anterior enamel lake disappears. At this stage of wear only a labial enamel lake persists and the hypocone is located on the posterior margin of the tooth.

The lower molars are lower crowned than the upper molars. As for the lower jaw bone, the mandibular foramen is located well caudal to the posterior margin of m3 (a little dorsal to m3, at the level of the tip of the coronoid process), whereas the mental foramen is situated rostrally to the anterior border of ml (approximately on the midline of the dentary).

The occlusal outline of ml is triangular, with its anterior part much narrower than its posterior. In early wear (e.g., MNHN C.G. 1860-382; Fig. 2d), it shows a pentalophodont dental pattern with anterolophid, metalophid, mesolophid, hypolophid, and posterolophid. The metalophid joins lingually the anterolophid and labially the protoconid, isolating a small and oval anterior enamel lake. The mesolophid is a long continuation of the protoconid. The anterior part of the tooth is isolated from the rest of the crown by a long sinusid. The latter results from the junction of the sinusid (or labial reentrant) with the mesosinusid (or middle labial reentrant). All lophids join the lingual margin of the tooth, isolating four transversely elongated enamel lakes. The m1 shows two open labial sinusids (protosinusid and sinusid). After moderate wear (e.g., ZMB 44768; Fig. 2e), all reentrants are closed-off, the protosinusid disappears, and the anterior part of the tooth is deprived of any enamel lake. However, two enamel lakes persist posteriorly. In late wear (e.g., MNHN C.G. 2000-761; Fig. 2f), the occlusal surface is completely flat and generally devoid of enamel lakes although a tiny and circular posterolabially located enamel lake may persist. The enamel has disappeared from the anterior border of the tooth, but it persists posteriorly.

The m 2 has four roots, the posterior ones being the most developed. Its occlusal outline is square and it is anteroposteriorly compressed. In early wear (e.g., MNHN C.G. 1860-382, Fig. 2d); this tooth has three lophs (anterolophid-metalophid, mesolophid, and hypolophid-posterolophid). The mesolophid is short and joins labially the anterior lophid (through the protoconid) and lingually the posterior one. The sole lingual reentrant is closed-off, isolating a labial enamel lake, which is elongated. The narrow and posterolingually directed sinusid is open. After moderate wear (e.g., ZMB 44768, Fig. 2 e ); the morphology of the tooth is similar to that of the preceding stage. In late wear (e.g., MNHN C.G. 2000-761; Fig. 2f), the sinusid is closed off and the tooth shows both a labial and a lingual enamel lake. The anterior side of the tooth loses the enamel.

In early wear (e.g., MNHN C.G. 1860-382; Fig. 2d), m3 has three lophids (an-terolophid-metalophid, mesolophid, and hypolophid-posterolophid). The first lophid joins the second one through the protoconid. The posterior lophid is isolated from the rest of the crown by a long reentrant. At this stage of wear, all reentrants are open. After moderate wear (e.g., ZMB 44768, Fig. 2e), the lingual reentrants are closed-off. The tooth has an elongate anterior enamel lake and a long sinusid. In late wear (e.g., MNHN C.G. 2000-761; Fig. 2f), the lingual reentrant is closed-off, isolating two enamel lakes, and the anterior enamel lake persists.

## Conclusion

The examination of the cheek teeth in various specimens of Cannomys badius has allowed determining the changes undergone by the dental pattern during wear. Three fundamental stages of wear have been recognized.

As postulated by Flynn (1990), modern and extinct rhizomyines are united by dental features that include lophodonty, high crowned teeth, a strong wear gradient decreasing from first to last molar, junction between the metalophid and the anterolophid or the protoconid on the lower molars, and isolation of enamel lakes. In addition, Cannomys shares with Rhizomys, the other extant representative of the tribe Rhizomyini, the synapomorphy of having on the m 1 a mesolophid that is a long continuation of the protoconid. However, Cannomys badius is much smaller than all the extant species of Rhizomys and exhibits the particularity of a trilophodont dental pattern on m2, M2, and M3, whereas it is tetralophodont in Rhizomys.

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## References

Allen GM (1939) A checklist of African mammals. Bulletin of the Museum of Comparative Zoology at Harvard College 83: 1-763.
Beolchini F, Corti M (2004) The taxonomy of the genus Tachyoryctes: a geometric morphometric approach. Italian Journal of Zoology 71: 35-43. doi: 10.1080/11250000409356548
Blyth E (1843) Mr. Blyth's monthly Report for December Meeting, 1842, with Addenda subsequently appended. Journal of the Asiatic Society of Bengal 12: 925-1011.
Bucher JE (1982) Family Rhizomyidae. In: Honacki J, Kinmam KE, Koeppl JW (Eds) Mammals Species of the World, First Edition. Association of Systematics Collections, Lawrence, 477-478.

Corbet GB, Hill JE (1991) A World List of Mammalian Species, Third edition. Oxford University Press, Oxford.
Ellerman JR (1941) The families and genera of living rodents. Volume II. Family Muridae. British Museum (Natural History), London.
Flynn LJ (1982) Systematic revision of Siwalik Rhizomyidae (Rodentia). Geobios 15: 327-389. doi: 10.1016/S0016-6995(82)80084-3
Flynn LJ (1990) The natural history of rhizomyid rodents. In: Nevo E, Reig OA (Eds) Evolution of subterranean Mammals at the organismal and molecular Levels. A.R. Liss, New York, 155-183.
Flynn LJ (2009) The antiquity of Rhizomys and independent acquisition of fossorial traits in subterranean muroids. Bulletin of the American Museum of Natural History 331: 128-156. doi: 10.1206/582-4.1
Gyldenstolpe N (1916) Zoological Results of the Swedish Zoological Expeditions to Siam 1911-1912 \& 1914-1915. Mammals II. Kungliga Svenska Vetenskapsakademiens Handlingar 57: 1-59. doi: 10.5962/bhl.title. 57007
Gray JE (1842) Descriptions of some new Genera and fifty unrecorded Species of Mammalia. Annals and Magazine of Natural History 10: 255-267. doi: 10.1080/03745484209445232
Hodgson BH (1841) New species of Rhizomys discovered in Nepal. Calcutta Journal of Natural History 2: 60-62.
IUCN (2010) IUCN red list of threatened species. Version 2010.1. Available at www.iucnredlist.org
López-Antoñanzas R (2012) Dental morphology and wear pattern in Tachyoryctes (Spalacidae, Rodentia). Mammalia 76: 309-314. doi: 10.1515/mammalia-2012-0018
López-Antoñanzas R, Flynn LJ, Knoll F (in press) A comprehensive phylogeny of extinct and extant Rhizomyinae (Rodentia): evidence for multiple intercontinental dispersals. Cladistics.
Misonne X (1971) Order Rodentia. In: Meester J, Setzer HW (Eds) The Mammals of Africa: an identification manual. Smithsonian Institution Press, Washington, 1-39. doi: 10.5479/ si.03629236.298.1
Musser GG, Carleton MD (1993) Family Muridae. In: Wilson DE, Reeder DM (Eds) Mammal Species of the World, Second Edition. The Johns Hopkins University Press, Maryland, 501-755.
Musser GG, Carleton MD (2005) Superfamily Muroidea. In: Wilson DE, Reeder DM (Eds) Mammal Species of the World, Third Edition. The Johns Hopkins University Press, Baltimore, 894-1531.
Rahm U (1980) Die Afrikanische Wurzelratte Tachyoryctes. A. Ziemsen, Lutherstadt Wittenberg.
Thomas O (1915a) Notes on the Asiatic Bamboo-Rats (Rhizomys, etc.). Annals and Magazine of Natural History 16: 56-61.
Thomas O (1915b) Further notes on Asiatic Bamboo-Rats. Annals and Magazine of Natural History 16: 313-317. doi: 10.1080/00222931508693719

# Taxonomic notes on the Macrocheilus Hope (Coleoptera, Carabidae, Helluonini) from Oriental Region, with description of one new species from the Philippines 

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#### Abstract

Taxonomic notes, together with illustrated characters, on the oriental species of the genus Macrocheilus Hope, 1838 (Carabidae, Helluonini) are provided. One new species, Macrocheilus dewvie sp. n. is described from the Philippines. A key to all species of Macrocheilus in the Oriental Region is also provided.


## Keywords

Coleoptera, Carabidae, Helluonini, Macrocheilus, Oriental Region, new species

## Introduction

The genus Macrocheilus Hope, 1838 is composed of Oriental, Palaearctic and Ethiopian species (Csiki 1932, Reichardt 1974). In total, fifty-six species of the genus are reported, among them, thirty-five species in the Ethiopian Region, nineteen in Oriental Region, and only two in Palaearctic Region (Lorenz 2005, Hůrka 2003, Zhao and Tian 2010).

For the Oriental Macrocheilus, Heller (1900) gave a table of ten species. But of them, two (dorsalis Klug and scapularis Klug) are actually African species, one (distactus Wiedemann) belongs to the genus Creagris, and one (tripustulatus Fairmaire) is a species of Pheropsophus. Andrewes (1920) dealt with ten Oriental species of the genus. Jedlička (1963) treated the East Asian Macrocheilus species and mentioned only five species. Park et al. (2006) listed six species from Vietnam. Zhao and Tian (2010) described seven new species and gave a key to Chinese species.

The aim of this paper is to provide taxonomic notes on all Oriental species of Macrocheilus by means of checking most of the type materials and a few other materials. As a result of the study, the examined materials are preserved in the Museum National d'Histoire Naturelle, Paris, France (MNHN). One new species of Macrocheilus from the Philippines is described. In addition, a distribution map of the genus in the Oriental Region is provided.

## Materials and methods

Materials for this study were dry mounted specimens. Dissection of specimens was done following the standard technique described by Lindroth (1974). Observations and measurements were made under stereo microscope (Leica, MZ125) and ocular microscope (Wild M5).

Abbreviations for the collections in which the type materials are deposited as follows:

| LMN | Leiden Museum, the Netherlands |
| :--- | :--- |
| MDSG | Museum Dresden, Saxony, Germany |
| MGI | Museum Genoa, Italy |
| MNHN | Museum National d'Histoire Naturelle, Paris, France |
| NHML | Natural History Museum, London, the U.K. |
| NMP | National Museum Prague, Czech Republic |
| SCAU | South China Agricultural University, Guangzhou, China |
| SNSD | Staatliches Naturhist. Sammlungen Museum für Tierkund, Dresden, |
| Germany |  |
| ZMUC | Zoological Museum University of Copenhagen, Copenhagen, Denmark |

## Taxonomic treatment

Genus Macrocheilus Hope, 1838
http://species-id.net/wiki/Macrocheilus
Macrocheilus Hope, 1838: 166. Type species: Macrocheilus bensoni Hope, 1838. Jeannel, 1949: 1041. Zhao \& Tian, 2010:4.
Acanthogenius Reiche, 1842: 334. Type species: Helluo impictus Wiedemann, 1823.

Generic diagnosis. Medium sized (length from 8.0 to 17.0 mm ), elongate, whole body coarsely punctuate and pubescent, except for labrum and middle region of ventral side of head. Head with two supraorbital setae on each side; ligula fairly wide, deeply impressed beneath at sides of base; mentum deeply emarginated, with a long and slender tooth; palpi varied in form; labrum extraordinarily or well developed; mandibles dorsally covered by labrum or not covered; antennae stout and flat, densely pubescent from antennomere 5. Pronotum truncate-cordiform or quadrate, basal margin more or less produced backwards in middle; lateral margin with two setae, one just before middle, and the other at hind angle. Elytra with deep striae and setiferous pores; intervals slightly or rather convex, interval 8 usually wide. Metepisterna elongate and very narrow in all species. Tarsomere 4 emarginate. Wings fully developed. No externally visible sexual dimorphic characters present. Median lobe of aedeagus varied in form. Parameres of aedeagus quite similar, elongate, the left one larger than the right one.

Differences with other genera of Helluonini in the Oriental Region. Macrocheilus spp., with larger body size, simple $4^{\text {th }}$ tarsomeres, and non-securiform labial palpomere are distinctly different from Colfax spp. (smaller body size, securiform labial palpomere) and Creagris spp. (bilobed $4^{\text {th }}$ tarsomere; smaller body size). Functional wings and the occurrence of a long spiniform median tooth of mentum of Macrocheilus distinguish them from Omphra spp. (which are brachypterous and have a short median tooth of the mentum).

Generic range. Oriental Region (China, Vietnam, Laos, Cambodia, Myanmar, India, Sri Lanka, the Philippines, Malaysia and Indonesia), Palaearctic Region (Pakistan, Syria, Asia Minor), Ethiopian Region (Ethiopia, Tanzania, Uganda, Senegal, Guinea, Guinea-Bissau, Sierra Leone, Togo, Nigeria, Central Africa, Cameroon, Gabon, Congo, Zimbabwe, Angola, South Africa, Madagascar).

## Key to species of Oriental Macrocheilus

1 Elytra without spots.................................................................................... 2

- Elytra spotted ............................................................................................ 3

2 Head and pronotum red; ligula with apical outer angles rounded; apex of lateral lobes of mentum fairly acute (Fig. 9); labrum with apical margin not sinuate (Fig. 1). Length 6.5 mm M. bicolor Andrewes

- Whole body piceous; ligula with apical outer angles rectangular; apex of lateral lobes of mentum rounded (Fig. 10); labrum with apical margin pluris-

3 Maxillary palpomere 4 not flattened and dilated, lateral lobes of mentum rounded on outer margin, mandibles not exposed, lateral margin of labrum rounded, front margin of clypeus not or slightly emarginate in middle, elytron with one or two spots.4
- Maxillary palpomere 4 strongly flattened and dilated, lateral lobes of mentum sinuate behind middle or on anterior one-third, mandibles exposed, lateral margin of labrum distinctly sinuate on anterior one-third, front margin of clypeus deeply and widely emarginate in middle, elytron with one spot.... 16
4 Elytron with one spot ..... 5
Elytron with two spots ..... 10
5 Labrum with anterior setae distinctly on upper surface margin ..... 6
Labrum with anterior setae close to or on the apical margin ..... 8
Labrum with apex narrow (Fig. 3); mentum with median tooth stout andwide, strongly sinuate at middle of lateral margins (Fig. 11). Length 25.0mmM. immanis Andrewes
Labrum with apex wide; mentum with median tooth slender, almost straight at sides ..... 77 Elytral spot transverse and almost rectangular (Fig. 46); median tooth ofmentum with four or five setae (Fig. 12). Length 16.0 mm .M. niger Andrewes
Length 14.5 mm M. asteriscus (White)
Elytral spot rounded, maxillary palpomere 4 roundly elongate; anterior setae of labrum close to apical margin. Length $12.5-14.0 \mathrm{~mm}$M. vitalisi Andrewes
Elytral spot oblong, maxillary palpomere 4 short and stout (Fig. 21); anterior setae of labrum along or on apical margin ..... 9Elytral spot covering intervals 3-7, anterior seta of labrum on apical margin
(Fig. 5). Length 13.0 mm ......................................M. binotatus AndrewesElytral spot covering intervals 2-7, anterior seta of labrum along apical mar-gin. Length 10.0 mm .M. macromaculatus Louwerens
10
Labrum with anterior setae on apical margin; ligula thickened at apex. ..... 11
Labrum with anterior setae beneath apex; ligula thin at apex. ..... 13
Labrum with apex wide; mentum with median tooth sinuate near apex onlateral margin, lateral lobes obtuse at apex. Length $15.5-17.0 \mathrm{~mm}$
M. gigas Zhao \& Tian
Labrum with apex pointed; mentum with median tooth not sinuate on lat- eral margin, lateral lobes sharp at apex ..... 12
12
Maxillary palpomere 4 strongly dilated ..... 15
14 Head and pronotum reddish brown (Fig. 53); labrum relatively long; prono-tum elongate; tibiae testaceous. Length 8.4 mm..... M. chaudoiri Andrewes
-tionaly convex on anterior portion which results apex can not be seen; maxil-lary palpomere 4 strongly and roundly dilated on anterior half portion.
$\qquad$M. parvimaculatus Zhao \& Tian
Body length $12.0-12.5 \mathrm{~mm}$; elytral spots larger (Fig. 52); labrum normallyconvex, apex visible (Fig. 6); maxillary palpomere 4 slightly dilated (Fig. 22)
black. Length 8.0 mmM. nigrotibialis Heller

17 Color brown; labrum with only three pairs of labral setae, apex pointed rounded; mandibles obtuse at apices; median tooth of mentum with lateral margin not sinuate. Length 10.8 mm $\qquad$ M. fuscipennis Zhao \& Tian

- Color black; labrum with four pairs of labral setae, apex widely rounded; mandibles sharp at apices; median tooth of mentum with lateral margin sinuate. 18
18 Labrum with additional setae located between the anterior and intermediate setae (Fig. 8 in Zhao \& Tian, 2010); median tooth of mentum with lateral margin sinuate in middle; mandibles less sharp at apices. Length 12.3-12.5 mm.
M. solidipalpis Zhao \& Tian
- Labrum with additional setae located before the anterior setae (Fig. 9 in Zhao \& Tian, 2010); median tooth of mentum with lateral margin sinuate on anterior one-third; mandibles sharper at apices. Length 11.0 mm
M. cheni Zhao \& Tian

Labrum with anterior and intermediate setae distance from each other, apex pointed rounded (Fig. 10 in Zhao \& Tian, 2010); mandibles sharp at apices; median tooth of mentum sharp at apex; pronotum almost quadrate. Length 12.1 mm
M. quadratus Zhao \& Tian

- Labrum with anterior and intermediate setae closed each other, apex widely rounded (Fig. 11 in Zhao \& Tian, 2010); mandibles obtuse at apices; median tooth of mentum strongly obtuse at apex; pronotum nearly cordiform. Length 11.7 mm $\qquad$ M. sinuatilabris Zhao \& Tian


## Macrocheilus bicolor Andrewes, 1920

http://species-id.net/wiki/Macrocheilus_bicolor
Figs 1, 9, 17, 37, 43
Macrocheilus bicolor Andrewes, 1920: 503; Andrewes, 1930: 206; Csiki, 1932: 1573; Lorenz, 2005: 512. Type locality: India (Bombay: Belgaum), deposited in NHML.

Diagnosis. Length $6.3-6.5 \mathrm{~mm}$, width 2.5 mm . Head and prothorax red; elytra black or sometimes bluish black. Labrum (Fig. 1) semicircular in front, shortly depressed towards base, front pair of setae small and closely placed along the front margin, intermediate one at a distance from margin; ligula rectangular, with a wide and deep median impression, a pair of setae at a distance form apex depressed at base; apex truncate, outer apical angles rounded; mentum (Fig. 9) glabrous at base,


Figures I-8. Labrum and clypeus of Macrocheilus (dorsal view): I M. bicolor (holotype) $\mathbf{2}$ M. impictus (male) $\mathbf{3}$ M. immanis (holotype) $\mathbf{4} M$. niger (holotype) $\mathbf{5}$ M. binotatus (female) $\mathbf{6} M$. tripustulatus (holotype) $\mathbf{7}$ M. nigrotibialis (holotype) $\mathbf{8}$ M. dewvie (holotype). Scale bar: 0.5 mm .
both tooth and lobes elongate, slender and sharp at apex; tooth almost as long as lobes, two pairs of setae on base; lobes sinuate at a distance from apex along outer margin; maxillary palpi (Fig. 17) not dilated. Elytra without spots.

Female genitalia. Gonocoxite short, stout, inner margin arcuate and not sinuate, apex short and sharp (Fig. 37).


Figures 9-24. Mentum and Left maxillary palpi of Macrocheilus (ventral view): 9, 17 M. bicolor (holotype) I0, 18 M . impictus (male) II, 19 M . immanis (holotype) I2, 20 M . niger (holotype) I3, $\mathbf{2 I}$ M. binotatus (female) 14, 22 M . tripustulatus (holotype) I5, 23 M . nigrotibialis (holotype) 16, $\mathbf{2 4} \mathrm{M}$. denvie (holotype). Scale bar: 0.5 mm .

Remarks. This species is allied to the next species, M. impictus (Wiedemann). Both differ from other species in the absence of elytral spot.

Materials examined. Holotype: 1 female, labeled "Kaoeqaoe, 740, Type, Macrochilus bicolor Type Andr., H. E. Andrewes det.; H. E. Andrewes Coll. B. M. 1945-97.;

Macrochilus bicolor Andrewes, 1920", deposited in NHML. 1 ex, sex unknown (the abdomen missing), labeled "Bangalore, Chikkangalur, Taboourel, 1900; Macrocheilus bicolor Andrewes, H. E. Andrewes det.", deposited in MNHN.

Distribution. India.

## Macrocheilus impictus (Wiedemann, 1823)

http://species-id.net/wiki/Macrocheilus_impictus
Figs 2, 10, 18, 25, 26, 38, 44
Helluo impictus Wiedemann, 1823: 49. Dejean, 1825: 287; Reiche, 1842: 335; Heller, 1900: 3; Andrewes, 1920: 503; 1921: 168; Csiki, 1932: 1574; Lorenz, 2005: 512. Type locality: India, deposited in ZMUC.

Diagnosis. Length $14.5-15.0 \mathrm{~mm}$, width $5.0-5.5 \mathrm{~mm}$. Black. Labrum (Fig. 2) with front margin rounded and bisinuate, three pairs of setae close to margin, and front two pairs on sinuated area; mandibles slightly obtuse at apex; mentum (Fig. 10) irregularly setose in basal half, both tooth and lobes elongate and sharp at apex, tooth slightly shorter than lobes; maxillary palpomere 4 (Fig. 18) cylindrical and gradually dilated toward apex, rounded and obliquely truncate at apex. Elytra without spots.

Male genitalia. Median lobe dilated on dorsal side in middle partion, sinuate on ventral side; apical lamella elongate, not sinuate near apex, rounded at apex (Figs 25-26).

Female genitalia. Gonocoxite slender, five setae on dorsal surface, sharp at apex (Fig. 38).
Remarks. Similar to $M$. bicolor without elytral spots, distinctly differs by larger size, body black, sinuate front margin of labrum and plurisetose mental tooth.

Materials examined. 1 male, labeled "Indes Orientales, $\mathrm{M}^{\text {ts }}$ Kodeicanel, J. Castets 1886"; 1 male, "India, Bangalore. P.S. Nathan, 1936"; 1 male, "Punjab Baddia (Indes Angl.); G. Babault Avril 1914"; 1 male, "S. India, Medungadu, P.S. Nathan. 1936"; 3 males, "Java", "Ex-Musaeo Chaudoir from Coll. Dejean"; 1 female, "Java", "ExMusaeo Mniszech from Coll. Dejean". All deposited in MNHN.

Distribution. India (East India) and Indonesia (Java).

## Macrocheilus immanis Andrewes, 1920

http://species-id.net/wiki/Macrocheilus_immanis
Figs 3, 11, 19, 27, 28, 45
Macrocheilus immanis Andrewes, 1920: 498. Csiki, 1932: 1574; Jedlička, 1963: 469; Lorenz, 2005: 512. Type locality: Myanmar (Taung-ngu), deposited in NHML.

Diagnosis. Length 24.7 mm , width 8.3 mm . Labrum (Fig. 3) elongate, three pairs of equidistant setae on upper surface near margin; ligula thickened, apex slightly


Figures 25-42. Aedeagus of Macrocheilus (left lateral and dorsal view) and Left gonopods of female genitalia (dorsal view): 25, 26, 38 M . impictus $\mathbf{2 7}, 28 \mathrm{M}$. immanis (holotype) 29, 30, $\mathbf{3 9} \mathrm{M}$. niger $\mathbf{3 I}$, 32, 41 M . tripustulatus $\mathbf{3 3}, \mathbf{3 4 , 4 2} \mathrm{M}$. nigrotibialis (holotype) $\mathbf{3 5 , 3 6} \mathrm{M}$. deuvie (holotype) $\mathbf{3 7} \mathrm{M}$. bicolor (holotype) 40 M . binotatus. Scale bar: 0.5 mm .
narrowed, slightly emarginate in middle of front margin, deeply depressed near apex to form a median channel, with five pairs of setae along sides, hollowed out above with a median carina; mentum (Fig. 11) with both tooth and lobes stout, tooth shorter than lobes, contracted after middle, irregularly setose on ventral surface on basal half; maxillary palpomere 4 (Fig. 19) dilated, densely setose; labial


Figures 43-5 I. Habiti of Macrocheilus (dorsal view): 43 M . bicolor (holotype) 44 M . impictus (male) 45 M . immanis (holotype) 46 M . niger (holotype) 47 M . asteriscus (male) 48 M . vitalisi (holotype) 49 M . binotatus (female) $\mathbf{5 0}$ M. gigas (holotype) 5 I M. parvimaculatus (holotype).
palpomere 4 dilated, palpomere 3 not dilated inwards. Elytal spots nearly square, large, near the middle, cover intervals 2-7.

Male genitalia. Median lobe of aedeagus strongly dilated and stout, strongly sinuate near paramere, straight towards apex, apical lamera elongate and narrowed at apex (Figs 27-28).

Remarks. This species is similar to $M$. niger and $M$. asteriscus in having anterior setae of labrum obviously on upper surface, but differs from the other two species by the large and almost square elytral spots and stout median tooth of mentum.

Materials examined. 1 male, the holotype, labeled "Toungoo; Type; Macrochilus immanis, Type, Andr., H. E. Andrewes det.; Macrochilus immanis Andrewes, 1920", deposited in NHML.

Distribution. Myanmar.

## Macrocheilus niger Andrewes, 1920

http://species-id.net/wiki/Macrocheilus_niger
Figs 4, 12, 20, 29, 30, 39, 46
Macrocheilus niger Andrewes, 1920: 499. Csiki, 1932: 1574; Lorenz, 2005: 512. Type locality: India (Nilgiri Hills), deposited in NHML.

Diagnosis. Length $16.0-16.3 \mathrm{~mm}$, width $6.0-6.2 \mathrm{~mm}$. Labrum (Fig. 4) convex, with a short furrow on each side of base, narrowed and pointed in front, with three pairs of setae away from the margin; ligula thickened, apex truncate towards sides, apical margin slightly emarginate, deeply depressed towards base, with a small median impression near apex; mentum (Fig. 12) glabrous at base; median tooth shorter than lobes, rather narrow and with apex rather pointed, two pairs of setae at basal area; lateral lobes obtuse at apex. Palpi (Fig. 20) not dilated. Elytral spots transverse, close to the middle of interval 5, covering intervals 3-7.

Male genitalia. Median lobe stout, apical lamella short and round at apex (Figs 29-30).

Female genitalia. Gonocoxite subapically dilated, three setae on dorsal surface, sharp at apex (Fig. 39).

Remarks. Similar to $M$. asteriscus, but differs with the tooth of mentum plurisetose and elytral spot not cruciform.

Materials examined. 1 female, the holotype, labeled "type, 11, H. L. Andrewes, Nilgiri Hills, H. E. Andrewes Coll., B.M. 1945-97.; Macrochilus niger Type-, Andr., H. E. Andrewes det.; Macrochilus niger Andrewes, 1920", deposited in NHML. 1 male and 1 female, "Dehra-Dun 1940-45, Kumaon Himalaya, Liesenfeldt leg.", deposited in MNHN.

Distribution. India (Nilgiri Hills, Malabar, Madras and Bombay) and Sri Lanka.

## Macrocheilus asteriscus White, 1844

http://species-id.net/wiki/Macrocheilus_asteriscus Fig. 47

Macrocheilus asteriscus White, 1844: 422; Bates, 1892: 389; Andrewes, 1919: 180; 1920: 500;1924: 470; 1930: 206; Csiki, 1932: 1573; Wu, 1937: 188; Jedlička, 1963: 470; Hůrka, 2003:407; Lorenz, 2005: 512; Zhao \& Tian, 2010: 6. Type locality: China (Hongkong), deposited in NHML.
Planetes crucifer Redtenbacher, 1867: 4. Type locality: China (Hongkong), deposited in NHML.

Remarks. M. asteriscus differs from other species in having cruciform elytral spots, the slender median tooth of mentum and the narrow apex of ligula.

Materials examined. 2 males, 1 female, "Hongkong" (MNHN); 2 males, "Hainan, Oct. 1979, Shaoming Zhuo leg." (SCAU); 2 males, "Guangdong: Zhanjiang, Jul. 1982" (SCAU); 1 male, 1 female, "Guangdong: Zhanjiang, May, 1983" (SCAU). 1 male, 1 female, "Annam, Phuc-Son, Nov. to Dec., H, Fruhstorfer", Central Vietnam (MNHN); 1 male, "Tonkin, P. Lemée, 1903-1906", North Vietnam (MNHN); 1 male, "Laos. Mouhot" and "Janson Acq. 1884" (MNHN); 2 males, "Java, Preanger", Indonesia (MNHN); 1 female, "Nilgherries" and "Ex. Musaeo H. W. Bates, 1892", India (MNHN).

Distribution. China, Vietnam, Laos, Myanmar, Indonesia and India.

## Macrocheilus vitalisi Andrewes, 1920

http://species-id.net/wiki/Macrocheilus_vitalisi
Fig. 48
Macrocheilus vitalisi Andrewes, 1920: 500; Andrewes, 1930: 208; Csiki, 1932: 1575; Wu, 1937:188; Jedlička, 1963: 470; Hůrka, 2003: 407; Lorenz, 2005: 512; Zhao \& Tian, 2010: 7.

Remarks. M. vitalisi is similar to M. binotatus from Sumatra, but as stated by Andrewes (1931), M. binotatus differs from M. vitalisi by "the dark colour, the elytral spot oblong and red; the upper surface generally is more coarsely and less densely punctuate, the genae are contracted more sharply to the neck, the sides of the prothorax are less sinuate behind, the elytral intervals more convex, with punctuation along the side more widely spaced and coarser". In addition, according to our examination, the anterior seta of the labrum is closer to the apical margin in $M$. vitalisi than in $M$. binotatus and the 4th maxillary palpomere is cylindrically dilated in M. vitalisi.

Materials examined. 1 female, the holotype, "China, Bowring 63.47*, 986 27/2/53" (NHML); 1 female, "Tonkin, Région de Hoa-Binh", "Muséum Paris, 1932, A. de Cooman" (MNHN).

Distribution. China, Laos, Vietnam, Borneo.

## Macrocheilus binotatus Andrewes, 1931

http://species-id.net/wiki/Macrocheilus_binotatus
Figs 5, 13, 21, 40, 49
Macrocheilus binotutats Andrewes, 1931: 68. Csiki, 1932: 1573; Lorenz, 2005: 512. Type locality: Indonesia (Sumatra), deposited in LMN.

Diagnosis. Length 14.0 mm , width 4.8 mm . Labrum (Fig. 5) with apex rounded and pointed, front setae on apical margin; mandibles covered by the labrum, sharp at apex; mentum (Fig. 13) glabrous at base, median tooth nearly as long as lobes, about five setae at basal half; maxillary palpomere 4 (Fig. 21) cylindrically dilated, labial palpomere 4 flat and dilated, labial palpomere 3 dilated inwards; ligula wide and rectangular, apical margin straight, with a wide median impression beneath apex, a seta on either side near apex. Elytral spots large and oblong, covering intervals $3-7$ in the middle.

Female genitalia. Gonocoxite elongate, three setae on dorsal margin, sparsely setose on ventral surface; apex sharp (Fig. 40).

Remarks. This species is similar to $M$. macromaculatus having elytral spots oblong; and it differs from $M$. macromaculatus in having the anterior setae of labrum on the apical margin.

Materials examined. 1 female, "Paggar Alam, Sumatra, J. Bouchaud", deposited in MNHN.

Distribution. Indonesia (Sumatra).

## Macrocheilus macromaculatus Louwerens, 1949

http://species-id.net/wiki/Macrocheilus_macromaculatus
Macrocheilus macromaculatus Louwerens, 1949: 51. Lorenz, 2005: 512. Type Locality: Indonesia (Java), deposited in LMN.

Diagnosis. Length 10.0 mm , width 3.0 mm . Labrum large, semicircular anteriorly, with a flat depression on each sides, setae running along the sides in two small furrows; palpi short and stout, truncate at apex; mentum with a long, narrow, sharp median tooth, lateral lobes a little longer. Elytral spots oblong and covers intervals 2-7.

The above description is after Louwerens (1949).
Remarks. This species is similar to $M$. binotatus, but the former is smaller in size and has the sides of elytra gently rounded behind, in contrast to the larger size and more sharply rounded hind region of elytra in $M$. macromaculatus.

Distribution. Indonesia (Java).

## Macrocheilus gigas Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_gigas
Fig. 50
Macrocheilus gigas Zhao \& Tian, 2010: 8. Type Locality: China (Guangdong), deposited in SCAU.

Remarks. M. gigas is similar to M. parvimaculatus and M. tripustulatus in having anterior seta of the labrum on the apical margin and the ligula thickened at apex which distinguishes these three species from other species with two spots on each elytron. But M. gigas differs from the other two species by wide apex of labrum and having median tooth of mentum sinuate near apex on lateral margin.

Materials examined. 1 male, the holotype, Guangdong: Zhanjiang, Jun.1983, deposited in SCAU; 1 male, Guangdong: Zhanjiang, Oct.1982, Zhichang Tan leg., deposited in SCAU; 2 males, Guangdong: Zhanjiang, July 1982, deposited in SCAU and MNHN.

Distribution. China (Guangdong: Zhanjiang).

## Macrocheilus parvimaculatus Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_parvimaculatus
Fig. 51
Macrocheilus parvimaculatus Zhao \& Tian, 2010: 9. Type Locality: China (Guangxi), deposited in SCAU.

Remarks. This species is easily distinguished from other species of Macrocheilus by its strongly convex labrum, maxillary palpomere 4 dilated in apical half and with small spots on elytra.

Materials examined. 1 male, the holotype, Guangxi: Liuzhou: Luzhai, 26 May 1980, Shaozhou Ruan leg.; 1 male, same data as holotype. All specimens deposited in SCAU. Distribution. China (Guangxi).

## Macrocheilus tripustulatus (Dejean, 1825)

http://species-id.net/wiki/Macrocheilus_tripustulatus
Figs 6, 14, 22, 31, 32, 41, 52
Helluo tripustulatus Dejean, 1825: 286 (syn. in Reiche, 1842: 334). Reiche, 1842: 334; Chaudoir, 1872: 212; Heller, 1900: 3; Andrewes, 1919: 124; Andrewes, 1920: 501; Csiki, 1932: 1575; Lorenz, 2005: 512. Type locality: Indonesia (Java), deposited in MNHN.

Diagnosis. Length $12.0-12.5 \mathrm{~mm}$, width $4.0-4.3 \mathrm{~mm}$. Labrum (Fig. 6) strongly convex on anterior part, rounded and pointed in front, front seta on apical margin, intermediate one on upper surface close to margin; mandibles sharp at apex; mentum (Fig.
14) irregular and densely setose, both tooth and lobes stout, tooth shorter than lobes, densely setose at basal half; palpi slender, palpomere 4 (Fig. 22) not dilated, densely setose, rounded and truncate at apex, labial palpomere 3 not dilated, bisetose inside; ligula strongly thickened and dilated, deeply depressed at sides, apex rounded, slightly emarginate in middle with a seta on either side. Elytra with two pairs of spots, front spot rounded, covering intervals $3-7$ in the middle, hind spot close to apical inner angles, nearly rectangular, covering intervals $1-5$; apex roundly truncate.

Male genitalia. Median lobe dilated in the middle ventrally, apical lamella long, wide and round at apex (Figs 31-32).

Female genitalia: Gonocoxite slender, setose on ventral surface, three setae on dorsal side, apex sharp (Fig. 41).

Remarks. M. tripustulatus can be distinguished from $M$. parvimaculatus by the slender maxillary palpomere 4, greater convexity of labrum and larger elytral spots.

Materials examined. 1 female, the holotype, "tripustulatus Dejean (non Fabr.), Java, Coll. Dejean, 3-pustulatus Wiedemann (Helluo). Fabr. (Brachinus), Western, ExMusaeo Chaudoir"; 1 male, "Java merid., 1500, 1891, H. Fruhstorfer."; 1 male, " Java", "Ex-Musaeo Mniszech"; 1 female, "Carin Chebà, 900-1100 m, L. Fea, V Xii-88; Ex-Musaeo H.W.Bates 1892". All specimens deposited in MNHN.

Distribution. Myanmar and Indonesia (Java).

## Macrocheilus chaudoiri Andrewes, 1919

http://species-id.net/wiki/Macrocheilus_chaudoiri
Fig. 53
Macrocheilus chaudoiri Andrewes, 1919: 130; 1920: 502; 1924: 470; 1930: 207; Csiki, 1932: 1573; Jedlička, 1963: 470; Lorenz, 2005: 512; Zhao \& Tian, 2010: 11. Type Locality: India, deposited in MNHN.
Acanthogenius trimaculatus sensu Chaudoir, 1872: 171 (non Oliver). Type Locality: India (Decan), deposited in MNHN.
Macrocheilus ruficollis Heller, 1923: 296; Andrewes, 1926: 258. Type Locality: Philippines (Luzon), deposited in MDSG.

Remarks. M. chaudoiri, M. nigrotibialis, M. bensoni and M. deuvie are similar in having the anterior seta of the labrum beneath the apex and two pairs of elytral spots. This species is distinct from the other three species in having the head and pronotum reddish brown and maxillary palpomere 4 strongly dilated.

Materials examined. 1 female, the holotype, "Ex-Musaeo Chaudoir; Macrocheilus Chaudoiri Andr., H.S. Andrewes det.; 3-maculatus Chaud., Deccan, Coll. Jeakes" (MNHN); 1 male, "Maissour, Sakrabail, IX 1897" (MNHN); 1 female, "Museum Paris, Cochinchine, Harmand 1872" (MNHN); 1 female, "Java" and "Museum Paris, Lakhon, Harmand 1878" (MNHN); 1 female, "Insl. Phiip." and "Thorey" (MNHN).

Distribution. China (Macao), Cambodia, Laos, Vietnam, the Philippines, Indonesia (Sumatra) and India.


Figure 52-6I. 52 M . tripustulatus (holotype) 53 M . chaudoiri (holotype) 54 M . nigrotibialis (holotype) 55 M . bensoni (male) 56 M . deuvie (holotype) 57 M . fuscipennis (holotype) 58 M . solidipalpis (holotype) 59 M. cheni (holotype) 60 M. quadratus (holotype) 6I M. sinuatilabris (holotype). Scale bar: 5.0 mm .

## Macrocheilus nigrotibialis Heller, 1900

http://species-id.net/wiki/Macrocheilus_nigrotibialis
Figs 7, 15, 23, 33, 34, 42, 54
Macrochilus nigrotibialis Heller, 1900: 2. Andrewes, 1920: 497; Csiki, 1932: 1574; Lorenz, 2005: 512. Type locality: Indonesia (Sulawesi), deposited in MDSG.

Diagnosis. Length $8.0-10.1 \mathrm{~mm}$, width $3.0-3.5 \mathrm{~mm}$. Labrum (Fig. 7) convex, arcuate at apex, front setae beneath apex, intermediate one on apical margin, hind one close to middle margin; mandibles sharp at apex; mentum (Fig. 15) setose at base; median tooth triangular and shorter than lobes, with two pairs of setae at base; lobes obtuse at apex; maxillary palpomere 4 (Fig. 23) roundly dilated, labial palpomere 4 triangular in shape and dilated, 3 not dilated and bisetose on inner sides; ligula thickened, apical margin arcuate inwards, deeply depressed at sides, with a median impression close to apex, outer apical angles rounded, with a seta on either of median impression close to apex. Elytra with front spot large, just before middle, almost rounded, covering intervals $3-7$ and extended to small part of 2 and 8 ; hind spot on inner angles, more or less triangular, covering intervals $1-5$.

Male genitalia. Median lobe slender, apical lamella round (Figs 33-34).
Female genitalia. Gonocoxite slender, arcuate, with three or four setae on dorsal side, apex slightly obtuse (Fig. 42).

Remarks. This species differs from following two species in having the tibiae black, maxillary palpomere 4 more dilated, the median lobe of male genitalia stouter and the apical lamella more rounded.

Materials examined. 1 male, the holotype, "typus!", "Drs. Sarasin N. Celebes Panot-Maimang", "12623", "Staatl. Museum fur Tierkunde. Dresden", deposited in SNSD. 1 female, labeled "Nord Borneo Mont Kina Balu 5-8, 1903, John Waterstradt; Macrochilus nigrotibialis Heller, det. Andrewes", deposited in MNHN.

Distribution. Indonesia (Sumatra, Malaysia and Sulawesi).

## Macrocheilus bensoni Hope, 1838

http://species-id.net/wiki/Macrocheilus_bensoni
Fig. 55
Macrocheilus bensoni Hope, 1838: 166; Chaudoir, 1872: 212; Bates, 1892: 389; Heller, 1900: 3; Andrewes, 1919: 176, 202; Hůrka, 2003: 407; Lorenz, 2005: 512: Zhao \& Tian, 2010: 10. Type locality: India, deposited in NHML.
Carabus trimaculatus Olivier, 1790: 347 (non Villers, 1789); Andrewes, 1919: 129, 176; 1920: 502; 1930, 208; Csiki, 1932: 1574; Wu, 1937: 188; Jedlička, 1963: 470. Type locality: China, deposited in NHML.

Helluo quadrimaculata Guérin-Méneville, 1840: 38; Chaudoir, 1872: 212. Type locality: India, deposited in MNHN.

Helluo tripustulata sensu Guérin-Méneville, 1843: 34 (non Dejean, 1825); Andrewes, 1923: 460. Type locality: China, deposited in MNHN.
Macrocheilus quadripustulatus Schmidt-Göbel, 1846: 65. Type locality: Myanmar, deposited in NMP.
Acanthogenius infuscatus Bates 1892, 389; Andrewes, 1920: 493. Type locality: Myanmar (Bhamo), deposited in MGI.

Remarks. This species can be distinguished from M. chaudoiri by the black head and pronotum and from $M$. nigrobibialis by the slender maxillary palpomere 4.

Materials examined. 1 male, "North China, 1884, Janson" (MNHN); 1 male, "North India, Coll. Benson, Ex-Musaeo H.W. Bates, 1892" (MNHN); 1 male, "Ind. Angl., Coimbatore Dt, Siruveni, VI. 1937" (MNHN); 1 female, "Ind. Angl., Mysore" (MNHN); 1 male, "Guangdong: Zhanjiang, Oct.1982, Zhichang Tan leg." (SCAU); 2 males, "Guangdong: Yingde, 27 Mar.2003, Danyang Zhao leg." (SCAU and MNHN); 1 male, Guangxi, Dec.1983" (SCAU); 1 male, "Yunnan: Jinghong, Jul. 1985" (SCAU); 1 female, "Hainan: Diaoluoshan, 26 Nov.1963" (SCAU).

Distribution. China (Fujian, Jiangxi, Guangdong, Guangxi, Guizhou, Yunnan, Hongkong, Hainan), Laos, Vietnam, Myanmar, India and Sri Lanka.

## Macrocheilus deuvie sp. n.

urn:lsid:zoobank.org:act:0181841F-FB7C-4A7D-9612-AB0BFEAD91F3
http://species-id.net/wiki/Macrocheilus_deuvie
Figs 8, 16, 24, 35, 36, 56

Description. Length 9.5 mm , width 3.5 mm .
Black; ligula, antennomeres 1-4, a spot on vertex, lateral margin of pronotum and legs reddish brown; sides of ligula, palpi, antennomeres 5-11 and elytral spots brown.

Head convex; neck short and punctate on dorsal surface; frontoclypeal sulcus faint, frontal foveae short and shallow; clypeus with apical margin truncate, two setae on each side of apical outer angels, a row of 6 setae along apical margin, irregularly setose basally on each side; labrum (Fig. 8) convex anteriorly, apical margin rounded, front setae beneath near apex, middle one just on apical margin, hind one close to middle margin; mandibles covered by labrum, sharp at apex; mentum (Fig. 16) irregularly setose and punctate at base, median tooth triangular and shorter than lobes, with a pair of setae at base, lobes obtuse at apex; maxillary palpi (Fig. 24) not dilated, labial palpomere 3 with two setae on inner side; ligula thickened, apex deeply and widely emarginated in middle.

Pronotum flat; widest before middle; faint median line, median and apical impression distinct, basal foveae deep; lateral margin round in front, strongly sinuate near base; hind angles nearly rectangular, with a small obtuse tooth and an emargination before tooth.

Elytra flat, striae with large, close punctures and setae; intervals convex, with two rows of regular punctures and setae, interval 8 wider than others and densely and irregularly punctate and setose; spots small, front spot rounded, just before middle, covering intervals 3-6, hind spot rhombic, on inner apical angles, covering intervals $1-5$.

Male genitalia. Median lobe dilated in middle on ventral margin; apical lamella elongated, rounded at apex (Figs 35-36).

Remarks. This species is very similar to M. bensoni, but differs by it's smaller size, curved labrum at anterior part, clypeus glabrous on middle, pronotum with lateral margin strongly sinuate near base, and male genitalia dilated on ventral margin.

Type material. 1 male, the holotype, "Philippines, Bohol. Ch. Semper", deposited in MNHN.

Etymology. This species is named in honor of Dr. Thierry Deuve (MNHN), a well known carabidologist.

Distribution. The Philippines.

## Macrocheilus fuscipennis Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_fuscipennis
Fig. 57
Macrocheilus fuscipennis Zhao \& Tian, 2010: 12. Type locality: China (Guangxi), deposited in SCAU.

Remarks. This species is similar to M. solidipalpis and M. cheni in having the larger elytral spots, but easily distinguishable from them by the labrum without additional setae, mandibles obtuse at apex, median tooth of mentum with lateral margin not sinuate and body brownish.

Materials examined. 1 male, the holotype, "Guangxi: Napo, 10 Oct.1970, by light trap".

Distribution. China (Guangxi: Napo).

## Macrocheilus solidipalpis Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_solidipalpis
Fig. 58
Macrocheilus solidipalpis Zhao \& Tian, 2010: 13. Type locality: China (Guangxi), deposited in SCAU.

Remarks. This species is similar to $M$. cheni but easily distinguishable from the latter by the presence of additional setae between the anterior and anterior setae, the median tooth of the mentum with sinuate lateral margins in middle (sinuate on api-
cal one-third in $M$. cheni), the median lobe larger, and the apical lamella long and narrowed towards apex.

Materials examined. 1 male, the holotype, "Guangxi: Dibei, Oct.1980, Xiuzhen Mao leg., by light trap".

Distribution. China (Guangxi: Dibei).

## Macrocheilus cheni Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_cheni
Fig. 59
Macrocheilus cheni Zhao \& Tian, 2010: 14. Type locality: China (Guangxi), deposited in SCAU.

Remarks. This species is similar to M. solidipalpis and the differences from the latter were mentioned above.

Materials examined. 1 male, the holotype,"Guangxi: Tengxian, Oct.1980". Distribution. China (Guangxi: Tengxian).

## Macrocheilus quadratus Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_quadratus
Fig. 60
Macrocheilus quadratus Zhao \& Tian, 2010: 16. Type locality: China (Guangxi), deposited in SCAU.

Remarks. This species is easily distinguished from other species by the shape of pronotum.
Materials examined. 1 male, the holotype, labeled "Guangxi: Cangwu, 1980, in paddy field".

Distribution. China (Guangxi: Cangwu).

## Macrocheilus sinuatilabris Zhao \& Tian, 2010

http://species-id.net/wiki/Macrocheilus_sinuatilabris
Fig. 61
Macrocheilus sinuatilabris Zhao \& Tian, 2010: 17. Type locality: China (Guangxi), deposited in SCAU.

Remarks. M. sinuatilabris is similar to M. quadratus in having three pairs of setae on the labrum and small elytral spots. However, it differs from M. quadratus in having
smaller elytral spots, the median tooth of the mentum obtuse at its apex, the rounded apex of the labrum, the close positioning of front and intermediate labral setae, and the slender median lobe of the aedeagus.

Materials examined. 1 male, the holotype, labeled "Guangxi: Fenghuangcheng, Sep.1981, by light trap".

Distribution. China (Guangxi: Fenghuangcheng).

## Distribution of Macrocheilus in the Oriental Region

The zoogeographical pattern of Macrocheilus is illustrated in Figure 62. Most species are distributed in limited small areas, but three are widespread, occurring in many countries: M. asteriscus in China, Vietnam, Laos, Myanmar, Indonesia and India; M. chaudoiri in China, Cambodia, Laos, Vietnam, the Philippines, Sumatra and India; and M. bensoni in China, Laos, Vietnam, Myanmar, India and Sri Lanka. At present the Macrocheilus faunas of Thailand and Nepal remain unknown.


Figure 62. Distribution map of Macrocheilus in the Oriental Region.

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## References

Andrewes HE (1919) On the types of Oriental Carabidae in the British Museum, and in the Hope Department of the Oxford University Museum. The transactions of the Entomological Society of London, 1919: 119-217.
Andrewes HE (1920) Papers on Oriental Carabidae.-V. The Annals and Magazine of Natural History, 9 (6): 493-506. doi: 10.1080/00222932008632476
Andrewes HE (1921) Notes on Synonymy and on some types of Oriental Carabidae in various foreign collections. The Transactions of the Entomological Society of London, 1921: 145-195.
Andrewes HE (1923) On the Oriental Carabidae of the "Reise Novara". The Transactions of the Entomological Society of London, 1923: 459-468.
Andrewes HE (1924) Part 2: Systematic list and description of a new species. In: Andrewes H.E. \& Scott H.: A list of Carabidae from Macao, South China, with a description of a new species and biological notes. The Annals and Magazine of Natural History, 9 (13): 466-472. doi: 10.1080/00222932408633058
Andrewes HE (1926) Papers on Oriental Carabidae.-XVI. The Annals and Magazine of Natural History, 9(17): 252-259. doi: 10.1080/00222932608633402
Andrewes HE (1930) Catalogue of Indian Insects (Part 18: Carabidae). Government of India Central Publication Branch, Calcutta, xxi+389 pp.
Andrewes HE (1931) Some keys to Sumatran Carabidae, together with descriptions of further new species. Zoologische Medemeelingen uitgegeven door s'Rijks Museum van Natuurlijke Historie te Leiden, 14: 54-78.
Bates HW (1892) Viaggio de Leonardo Fea in Birmania e regioni vicine. XLIV. List of the Carabidae. Annali del Museo Civico di Storia Naturale di Genova, 32: 267-428.
Chaudoir M de (1872) Descriptions d'espèces nouvelles de Carabiques de la tribu des Troncatipennes, et remarques synonymiques. Revue et Magasin de Zoologie pure et appliquée, (2) 23: 138-143, 168-172, 212-221.
Csiki E (1932) Carabidae: Harpalinae VII (Pars 124). In Junk W, Schenkling S (Eds) Coleopterorum Catalogus. Volumen III. Carabidae III. Berlin, 1279-1598.

Dejean PFMA (1825) Species général des Coléoptères de la collection de M. le comte Dejean. Tome premier. Chez Crevot, Libraire-Editeur, Paris, 463 pp.
Guérin-Méneville FE (1840) Coléoptères nouveaux du Plateau des Neelgherries dans les Indes Orientales, découvertes par M. Adolphe Delessert. Revue Zoologique, 37-42.
Guérin-Méneville FE (1843) Animaux articulés. In : Delessert A.: Souvenirs d'un voyage dans l'Inde exécuté de 1834 à 1839. Paris: Fortin, Masson \& Co, 33-98 + pl. 11-27.
Heller KM (1900) Neue Käfer von Celebes. IV. (Nr. 5). Abhandlungen und Berichte des Königlichen Zoologischen und Anthropologishe-Ethnographischen Museums zu Dresden, 9: 1-46.
Heller KM (1923) Some new Malayan Carabidae, especially Philippine. The Philippine Journal of Science, 23: 295-307.
Hope FW (1838) The coleopterist's manual, part the second, containing the predaceous land and water beetles of Linneus and Fabricius. London: Bohn, xvi+ 168, 3 pl., frontispiece.
Hůrka K (2003) Carabidae: Helluonini. In: Löbl I, Smetana A (Eds) Catalogue of Palaearctic Coleoptera. I: Archostemata-Myxophaga-Adephage. Apollo Books, Stenstrup, 407 pp.
Jeannel R (1949) Faune de l'Empire français. XI. Coléoptères Carabiques de la région Malgache (troisième partie). Paris: Librairie Larose, 767-1146.
Jedlička A (1963) Monographie der Truncatipennen aus Ostasien. Lebinnae - Odacanthinae - Brachyninae (Coleoptera, Carabidae). Entomologische Abhandlungen und Berichte aus dem Staatlchen Museum für Tierkunde in Dresden, 28: 269-578.
Lindroth CH (1974) Coleoptera Carabidae. Handbooks for the Identification of British Insects. London: Published by the Society and Sold at its Rooms, 148 pp.
Lorenz W (2005) Systematic List of Extant Ground Beetles of the World. Second Edition. Tutzing: Published by the author, 530 pp .
Louwerens CJ (1949) Some notes on the Carabidae, collected by Mr. P. H. van Doesburg in the Malay Archipelago with descriptions of new species. Tijdschrift voor Entomologie uitgegeven door de Nederlandsche Entomologische Vereeniging, 90: 45-53.
Park JK, Trac DH, Will K (2006) Carabidae from Vietnam. Journal of Asia-Pacific Entomology, 9(2): 85-105. doi: 10.1016/S1226-8615(08)60280-0
Olivier AG (1790) Encyclopédie méthodique, ou par ordre de matières; par une société de gens de lettres, de savans et d'artistes; précédée d'un vocabulaire universel, servant de table pour tout l'ouvrage, ornée des portraits de Mm . Diderot \& d'Alembert, premiers éditeurs de l'Encyclopédie. Histoire naturelle. Insectes. Tome cinquième. Livraison 41. Paris: C.J. Panckouche, 368 pp .
Reichardt H (1974) Monograph of the Neotropical Helluonini, with notes and discussions on Old World forms (Coleoptera: Carabidae). Studia Entomologia, 17: 211-302.
Redtenbacher L (1867) Zoologischer Theil. Zweiter Band. I. Abtheilung A. 1. Coleoptera. Reise der Österreichischen Fregatte Novara un die Erde in den Jahren 1857, 1858, 1859 unter der befehlen des Commodere B. von Wüllerstorf-Urbair. Karl Gerold's Sohn, Wien, 1-249.
Reiche L (1842) Recherches sur les Helluonides, ou révision du genre Helluo, Bonelli et Dejean. Annales de la Société Entomologique de France, 11[1842]: 323-344.
Schmidt-Göbel HM (1846) Faunula Coleopterorum Birmaniae, Adjectis Nonnulis Bengaliae Indigenis, Prague: Gottlieb Hasse Söhne, viii+ $94+3$ pls

White A (1844) Some new insects from China. The Annals and Magazine of Natural History, 14: 422-423. doi: 10.1080/037454809495213
Wu CF (1937) Catalogus Insectorum Sinensium. III. Peiping: The Fan Memorial Institute of Biology, 1312 pp.
Zhao DY, Tian MY (2010) A review of the genus Macrocheilus Hope (Coleoptera: Carabidae:
Helluonini) of China, with descriptions of seven new species. Polish Journal of Entomology, 29: 3-23.

