# Lepocranus and Valalyllum gen. nov. (Orthoptera, Tetrigidae, Cladonotinae), endangered Malagasy dead-leaf-like grasshoppers 

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#### Abstract

Only two leaf-like pygmy grasshopper species and specimens are known from Madagascar: the Leatherback Pygmy Grasshopper (Lepocranus fuscus Devriese, 1991) —which has a relatively low median carina of the pronotum; and the Malagasy Litterhopper (Valalyllum folium gen. et. sp. nov.), herein described - which has a high median carina. Lepocranus fuscus is known from the rainforests around Tampolo, Manakambahiny, and Mahavelona (Foulpointe). The new taxon, Valalyllum folium gen. et. sp. nov. is known only from the Belanono forest. Both species inhabit northeastern Madagascar. The new species could be rare or not-easy-to-spot in the rainforest leaf litter, where it most probably lives. A new tribe, Valalyllini trib. nov., is described for the two mentioned genera because its members are different from the Caribbean leaf-like Choriphyllini Cadena-Castañeda \& Silva, 2019, from the African leaf-like Xerophyllini Günther, 1979, and from the Asian leaf-like Cladonotini Bolívar, 1887. A tabular key to the tribes of Cladonotinae with leaf-like representatives is provided, together with photographs of type specimens of both species belonging to the newly described tribe. The holotype of the new species belongs to the Muséum national d'Histoire naturelle Orthoptera collection, Paris.


## Keywords

Gondwana, identification traits, mimicry, new genus, new species, new tribe, phylogenetic position, rainforest, taxonomy, Valalyllini

## Introduction

Leaf mimicry in animals has evolved independently a number of times (Skejo et al. 2019). From the leaf-like satanic leaf-tailed gecko (Uroplatus phantasticus Boulenger, 1888) to the Indian oakleaf (Kallima inachus Boisduval, 1836), this curious morphology certainly survived as an adaptation for avoiding diurnal predators. Anyone who has visited a tropical rainforest will know how unlikely it is to spot a leaf-like critter in diverse leaf litter. Among the pygmy grasshoppers (Tetrigidae), there are several leaf-like species, mostly belonging to the subfamily Cladonotinae (Tumbrinck 2014). Katydids are well-known for their various leaf-like forms (Nickle and Castner 1995) as well.

Until now, only a single leaf-like species of Tetrigidae was known to inhabit Madagascar-the Leatherback Pygmy Grasshopper, Lepocranus fuscus (Devriese 1991) (Fig. 1). This species has been known from several individuals, of which the only digitalized one is a male holotype from the Tampolo forest, slightly northwest of Toamasina (Tamatave). The species has also been reported from the rainforests around Manakamahiny and Mahavelona (Foulpointe) (Danielczak et al. 2017). With this paper, we add one more species to the diverse Malagasy fauna, Valalyllum folium gen. et sp. nov. (Fig. 2). Our description is based on a single male from the Belanono rainforest, in the northernmost rainforests of Madagascar. The aim of this study is to describe V. folium sp. nov., compare it to L. fuscus, provide identification traits for the Malagasy leaf-like pygmy grasshoppers, and discuss their position in the Tetrigidae tree of life by describing a new tribe, Valalyllini trib. nov. (Fig. 3).

## Materials and methods

Museum acronyms used were MNCN for Museo Nacional de Ciencias Naturales (Madrid) and MNHN for the Muséum national d'Histoire naturelle. Taxonomy follows the Orthoptera species file (Cigliano et al. 2022) and nomenclature follows the International Code of the Zoological Nomenclature. Morphology and measurements follow Devriese (1991) and Tumbrinck's (2014) capital work on the subfamily Cladonotinae; we have added a new important character, whether the anterior pronotal tip comes before the face or not. Measurements were taken in ImageJ (version 1.8.0_172) after calibration with millimetre paper. For comparison of the new tribe with already described taxa, we have consulted Devriese (1999) monograph on Xerophyllini Günther, 1979, and the Orthoptera species file (Cigliano et al. 2022) digital catalogue of museum specimens for Choriphyllini Cadena-Castañeda \& Silva, 2019 and Cladonotini Bolívar, 1887. The holotype male of Lepocranus fuscus is deposited in MNCN and was examined and digitalized by the authors. The holotype male of Valalyllum folium gen. et. sp. nov. is deposited in MNHN, Paris and was examined and digitalized by the authors. Both Lepocranus fuscus and Valalyllum folium gen. et. sp. nov. holotypes are now digitalized and high-resolution photographs are uploaded to the Orthoptera species file database (Cigliano et al. 2022).

## Results

Class Insecta<br>Order Orthoptera<br>Suborder Caelifera<br>Infraorder Acrididea<br>Superfamily Tetrigoidea Rambur, 1838

Family Tetrigidae Rambur, 1838
Diversity in Madagascar. Altogether, 76 Tetrigidae species (including the new one) are known to inhabit Madagascar, of which 73 (= 96\%) can be found only in Madagascar and nowhere else in the world (Cigliano et al. 2022). These species are assigned to altogether 29 genera, of which 24 are endemic to the island. Hitherto, only a single leaf-like species was known from the island, Lepocranus fuscus. This study adds one more species with this interesting cryptic morphology, Valalyllum folium gen. et sp. nov.

## Subfamily Cladonotinae

Composition and distribution. This diverse and polyphyletic family includes altogether 314 species assigned to 74 genera (Cigliano et al. 2022). Most genera require revision and are not adequately classified in the Tetrigidae system (Zhang et al. 2020). Within the subfamily Cladonotinae, several monophyletic tribes exist, such as Caribbean Choriphyllini (Choriphyllum Serville, 1838 and Phyllotettix Hancock, 1902), African Xerophyllini (including leaf-like Acmophyllum Karsch, 1890, Seyidotettix Rehn, 1938, Trypophyllum Karsch, 1890, Xerophyllum Fairmaire, 1846), and Asian Cladonotini (Hymenotes Westwood, 1837, Holoarcus Hancock, 1909, Dolatettix Hancock, 1907).

Diversity in Madagascar. Altogether, 18 Tetrigidae species of Madagascar are assigned to the subfamily Cladonotinae: two members of Valalyllini trib. nov. (L. fuscus and V. folium); 13 species of the genus Thymochares, which is currently not assigned to any of the tribes, and it is questionable whether it represents a genus of Cladonotinae; Microthymochares pullus, an endemic genus and species with the same taxonomic problem as Thymochares; Epitettix spheniscus also with the same problematic assignment; and Xerophyllini and its member Morphopoides madagascariensis (Cigliano et al. 2022).

Tribe Valalyllini trib. nov. [Leaf-like Cladonotinae of Madagascar] http://zoobank.org/49EABF65-9F2F-482B-B545-B1A997AE9957
Figs 1-3; Table 1
Cladonotini (partim): Tumbrinck et al. (2020): 336 (tentatively assigned to Cladonotinae: Cladonotini).

Type genus. Valalyllum gen. nov., herein described (see below), type species V. folium sp. nov., also described herein.

Composition and distribution. Two monotypic genera, Lepocranus (including only L. fuscus) and Valalyllum gen. nov. (including V. folium sp. nov.). Lepocranus fuscus is endemic to the rainforests east of Mahavelona (Foulpointe) (Danielczak et al. 2017), while V. folium sp. nov. is endemic to northern Malagasy rainforests around Sambava and Andapa. Lepocranus folium sp. nov. lives 350 km more northerly than L. fuscus.

Descriptive diagnosis. Members of Valalyllini (Lepocranus, Valalyllum gen. nov.) are very similar to and thus most likely closely related to Asian Cladonotini (genera Misythus Stål, 1877, Hymenotes Westwood, 1837, Holoarcus Hancock, 1909) and Caribbean Choriphyllini (Choriphyllum Serville, 1838 and Phyllotettix Hancock, 1902). Head, pronotum and legs in these three tribes show remarkable similarity and are regarded as homologous. Superficially, the leaf-like morphology of Valalyllini trib. nov. also resembles that of leaf-like Xerophyllini, but detailed comparisons reveal no homologous parts.

Antenna shorter than hind femur; vertex very wide; vertex slightly and obliquely elevated above the compound eyes; vertex without horns; upper margin of the antennal grove in the level of the lower margin of a compound eye; pronotal tip of the pronotum projected above the head, but not before the eyes or before the face; transverse pronotal veins weak, almost unrecognizable; pronotal tip obliquely bilobate in dorsal view; legs smooth, only with weak (small) undulations and triangular projections, not toothed or sawed. Found only in Madagascar.

Table 1 shows a comparison between leaf-like Caribbean Choriphyllini (Choriphyllum Serville, 1838 and Phyllotettix Hancock, 1902), Asian Cladonotini (Hymenotes Westwood, 1837, Misythus Stål, 1877, Holoarcus Hancock, 1909 and Dolatettix Hancock, 1907), and African Xerophyllini (Xerophyllum Fairmaire, 1846, Trypophyllum Karsch, 1890, Acmophyllum Karsch, 1890). The leaf-like Tetrigidae were visually compared by Skejo et al. (2019). For comparison and elucidation of the mentioned characters, the reader is urged to refer to the abovementioned publication.

Table I. Tabular key to the four Cladonotinae tribes with leaf-like representatives (Valalyllini trib. nov. from Madagascar, Choriphyllini from the Caribbean, Cladonotini from SE Asia, and Xerophyllini from Africa).

|  | Valalyllini trib. nov. | Choriphyllini | Cladonotini | Xerophyllini |
| :---: | :---: | :---: | :---: | :---: |
| Distribution | Madagascar | Caribbean | Philippines, Papua | Tropical Africa |
| Antenna | short | long or short | short | short |
| Vertex horns | absent | absent | absent | present |
| Upper margin of the antennal groove | on the level of the lower margin of a compound eye | on the level of the lower margin of a compound eye | on the level of the lower margin of a compound eye | below the lower margin of a compound eye |
| Anterior tip of the pronotum | not falling in front of the face | strongly projected in front of the face | projected in front of the eyes | variable, not falling in front of the face (Trypophyllum) or projected in front of the face (Xerophyllum) |
| Transverse veins on the pronotum | weak | strong | very strong | weak |
| Posterior tip of the pronotum | obliquely bilobate | pointed | variable (pointed in Misythus, bilobate in Cladonotus) | pointed |
| Legs texture | smooth, with small protrusions | smooth | toothed or spiky | toothed or spiky |

## Genus Lepocranus Devriese, 1991

Lepocranus: Devriese 1991

Type species. Lepocranus fuscus, by original designation and by the original monotypy.
Composition and distribution. A single species only (L. fuscus), endemic to the Malagasy rainforests east of Mahavelona (Foulpointe), such as Tampolo, where the holotype is from, and Manakambahiny (reported in Danielczak et al. 2017).

Original etymology. Complex noun, male in gender, composed of Latinized Ancient Greek "lepos" ( $\lambda$ ह̇ $\pi \circ \varsigma)$, meaning bark, and "chranos" ( $\chi \dot{\alpha} \nu \circ \varsigma)$, meaning helmet (Devriese 1991).

Diagnosis. Same as for the species, see below.

## Lepocranus fuscus Devriese, 1991

Fig. 1
Lepocranus fuscus: Devriese 1991
Material examined. Holotype. Madagascar • $1 \widehat{J}^{\text {T}}$; "Foret de Tampolo, Madagascar"; May. 1932; A. Seyrig leg.; MNCN 7230.

Genus and species diagnosis. Small (< 10 mm long), apterous, leaf-like, cryptic species endemic to northern Madagascar. Antennae are short and filiform, composed of 15 antennomeres. The upper margins of the antennal groove are at the lower margin of a compound eye. Frontal costa bifurcates between the eyes into rounded facial carinae (parallel in Valalyllum gen. nov.), between which there is a wide scutellum, as wide as a compound eye. Vertex obliquely projected above the eyes in frontal view. Vertex about 3 times wider than a compound eye. In the frontal view, a compound eye is rounded (ovoid in Valalyllum gen. nov.). Pronotum is leaf-like, 2.4 times as long as high (1.75 times in Valalyllum gen. nov.) because of the compressed and elevated median carina. In the dorsal view, the median carina of the pronotum is sulcate. Median carina of the pronotum is straight in the dorsal view (undulated in Valalyllum gen. nov.). Pronotum dips above the head in dorsal view, then smoothly curves upwards and gradually descends towards the posterior apex at an obtuse angle ( $150^{\circ}$ slope, already reported in Devriese 1991) (posterior slope is abrupt in Valalyllum gen. nov. and forms a right-angle). Posterior slope of the median carina is weakly undulated (much more in Valalyllum gen. nov.). The posterior apex of the pronotum is bilobated in the dorsal view. Pronotum does not cover the whole abdomen, the last segments are not covered, and the subgenital plate is visible (fully covered in Valalyllum gen. nov.). Dorsal margin of hind femora bearing strong projections (lappets). Genicular and antegenicular teeth are large and blunt (small and angular in Valalyllum gen. n). Mid tibia is stout. The top margins of the mid and the fore femora undulated but lack strong tubercules. Pulvilli of the hind tarsi are rounded.

Original etymology. Latin adjective in male gender, "fuscus, -a, -um" meaning brown (Devriese 1991).


Figure I. Lepocranus fuscus Devriese, 1991 A head detail in frontal view B habitus in right lateral view $\mathbf{C}$ habitus in frontal view $\mathbf{D}$ habitus in light lateral view $\mathbf{E}$ habitus in dorsal view $\mathbf{F}$ labels. Photo: J. Skejo and MNCN Madrid.

Vernacular name. Leatherback Pygmy Grasshopper (Danielczak et al. 2017).
Measurements (male holotype). Body lengtb: (from the tip of the head to the tip of the subgenital plate) 8.6 mm (cited 9.8 mm in Devriese 1999, from the tip of the pronotum to the tip of the subgenital plate). Pronotum length: 8.1 mm (cited 8.2 mm in Devriese 1991). Pronotum maximum beight: 3.3 mm . Pronotum width between the lateral lobes: 3.8 mm . Pronotum width between the shoulders: 2.5 mm . Eye width: 0.4 mm . Vertex width: 1.2 mm . Fore femur length: 2.2 mm . Fore femur width: 0.5 mm . Mid femur length: 2.5 mm . Mid femur width: 0.8 mm . Hind femur
length: 5.3 mm (cited 5.3 mm in Devriese 1991). Hind femur width: 2.1 mm . Hind femur length/width ratio: 2.5.

Locus typicus. Madagascar, Tampolo.
IUCN Red List Assessment. The Leatherback Pygmy Grasshopper was listed as an endangered species on the IUCN Red List because (1) the minimal geographic range it inhabits (the extent of occurrence is only about $3000 \mathrm{~km}^{2}$ ), (2) the population seems to be fragmented, and (3) the decline in both the number of mature individuals and the size and quality of the range area due to inferred severe deforestation (Danielczak et al. 2017).

## Genus Valalyllum gen. nov.

http://zoobank.org/40A42840-9E96-4ED6-8D4A-2872D80E2E70

Type species. Valalyllum folium gen. et sp. nov., here described, by original monotypy.
Composition and distribution. A single species only ( $V$. folium), endemic to Madagascar (Belanono).

Etymology. Noun of neuter gender. From Malagasy "valala" or "vahalala"- grasshopper and Latinized Ancient Greek "phyllum" - leaf

Diagnosis and description. Same as for the species, see below.

## Valalyllum folium sp. nov.

http://zoobank.org/0DA809A6-3F8A-48D2-8DAB-2E135114FC73
Fig. 2
Material examined. Holotype. Madagascar - 1 §; East Madagascar, Belanono, "30 km SW de Sambava, sur la route d'Andapa" [along the road to Andapa]; Valdon and Peyrieras leg.; MNHN.

Diagnosis. Large ( $>11 \mathrm{~mm}$ long), apterous, leaf-like, rectangular, and cryptic species endemic to northern Madagascar. Antennae are short and filiform, composed of 15 antennomeres. Upper margins of the antennal groove in the level of the lower margin of a compound eye. Frontal costa bifurcates between the eyes into parallel facial carinae (more rounded in Lepocranus), between which there is a wide scutellum, as wide as a compound eye. Vertex obliquely projected above the eyes in the frontal view. Vertex about 3 times wider than a compound eye (measured at its widest part as seen in the frontal view). In the frontal view, a compound eye is ovoid (rounded in Lepocra$n u s)$. Pronotum is rectangular and 1.75 times as long as high (2.4 times in Lepocranus) because of the compressed and elevated median carina. In the dorsal view, the median carina of the pronotum is sulcate. Median carina of the pronotum is undulated in a sinusoid fashion in the dorsal view (straight in Lepocraus). Pronotum dips above the head in dorsal view, then smoothly curves upwards and sharply descends towards the posterior apex. The posterior slope of the pronotum is abrupt (gradual in Lepocranus), forming a right-angle (obtuse angle in Lepocranus), and undulated. The posterior apex of the pronotum is bilobated in the dorsal view. Pronotum covers the whole abdomen


Figure 2. Valalyllum folium gen. et sp. nov. from Madagascar, male holotype, deposited in MNHN Paris. A habitus in left lateral view $\mathbf{B}$ head in frontal view $\mathbf{C}$ habitus in frontal view $\mathbf{D}$ habitus in dorsal view. Photo J. Skejo and MNHN Paris.
(last segments not covered in Lepocranus). Dorsal margin of the hind femora bears three small projections (lappets). Genicular and antegenicular teeth are small and angular. Mid tibia is stout. Top margins of the mid and the fore femora lack tubercules. Pulvilli of the hind tarsi are rounded.

Etymology. The specific epitheton is a noun in apposition, from Latin "folium, $-i$, $n$." leaf, because of the species' leaf-like morphology.

Proposed vernacular name. Malagasy Litterhopper
Description. Holotype (male). General appearance. Valalyllum folium gen. et sp. nov. is a large (> 11 mm ); smooth; rectangular; cryptic; dead-leaf-mimicking species with fine leaf-like venation on the elevated part of pronotum; uniformly brown except for the yellowish tarsi of all legs, as well as pale-yellowish hind tibiae.

Head. Antenna (Fig. 2B, C) short, filiform, composed of 15 antennomeres. The first segment is the largest scapus, second is a barrel-like pedicel, both circular in crosssection, while the remaining 13 antennomeres make up the flagellum. Segments $3^{\text {rd }}$ to $6^{\text {th }}$ are basal segments of the flagellum and they are robust, about two times as long as wide. Segments $7^{\text {th }}$ to $10^{\text {th }}$ are the central segments, and they are elongated, from four to five times as long as wide. Segments $11^{\text {th }}$ to $13^{\text {th }}$ are the subapical segments, shorter and bulkier than the mid segments. Last two antennomeres, i.e., $14^{\text {th }}$ and $15^{\text {th }}$ are reduced apical antennomeres. In the frontal view (Fig. 2B, C), the vertex is 3 times wider than the width of the compound eye ( 1.2 mm wide vertex, 0.4 mm wide compound eye); the vertex tip is above the top margin of the compound eyes, forming a smooth convex bulge. Frontal costa is smooth before the bifurcation, without visible projections or teeth. Bifurcation of the frontal costa is situated just below the line connecting the mid portion of the compound eyes. Height of the compound eye is greater than the height of the scutellum. Compound eye is ovoid. Width of the compound eye is the same as the width of the scutellum. Scutellum in its widest part and at the level of antennal groove is significantly wider than the antennal groove. Facial carinae are visible, and run straight after the bifurcation. Facial carinae are however, slightly sub-parallel, so the scutellum is slightly widened at the level of the median ocellus. Facial carinae are compressed and elevated but smooth. Paired ocelli are situated below the line connecting the mid portion of the compound eyes, but above the lower margin of compound eyes. Top margin of the antennal groove is placed at the level of the bottom margin of the compound eyes. In the lateral view (Fig. 2A), both vertex and face are visible around an ovoid compound eye. Fastigium protrudes beyond the furthest margin of the compound eyes for a half of the compound eye width. Occipital area is visible and narrow. In the dorsal view (Fig. 2D), the vertex is 3 times wider than the width of the compound eye. However, in this view, the vertex is also visibly very short and reaches to about a third of the compound eye length, i.e., it is not projected, but indrawn. Lateral and transverse carinae are slightly visible and elevated. Medial carina not visible due to pronotal occlusion.

Pronotum In the frontal view (Fig. 2C), the pronotum is visible above the head as a large, compressed and elevated and undulated projection with sulcated (ditched) ridge. In lateral view (Fig. 2A), the pronotum has the appearance of a dead leaf, because of the strongly compressed and elevated median carina of the pronotum. Prozonal and extralateral carinae are invisible. The pronotum is long ( 13.4 mm ) and high ( 7.6 mm ), giving a rectangular shape to the insect. Tip of the median carina of the pronotum has striped pale-dark-pale-dark coloration. Anterior margin is projected above the head as an oblique projection and then goes in circular fashion towards the dorsal portion, where it becomes the highest in the level of the mid coxae. After its highest portion, the dorsal margin of the median carina slowly decreases in height, and then in the level of the subgenital plate it abruptly falls in almost rectangular angle. The whole distal portion of the pronotum after this abrupt slope is finely toothed and undulated. Ventral sinus of the pronotum is large and visible, while tegminal sinus lacks because of the absence of wings. Infrascapular area, covered by the hind femora, is wide, smooth and


Figure 3. The diversity and the distribution of the tribe Valalyllini trib. nov. (Valalyllum folium gen. et sp. nov. in the north, and Lepocranus fuscus in the south), the Malagasy dead-leaf-like Cladonotinae. Both species are endemic to small areas and are likely endangered because of deforestation. Both species most probably inhabit rainforest leaf litter.
convex. The compressed dorsum of pronotum is finely granulated and intercepted with many fine veins (carinulae), giving an insect even more credible dead-leaf mimicry. Paranota are triangular, with truncate-oblique apex. In the dorsal view (Fig. 2D), the median carina has a sulcated ridge and forms a sinusoid undulation from the head caudad. The pronotum is finely granulated. Prozonal and extralateral carinae are absent,
not visible. No pronotal projections visible. Shoulders (humeral angles) oblique, not projected outwards. Posterior apex of the pronotum is widely bilobate.

Legs. Fore legs (Fig. 2A, C, D). The fore femur is stouter than the mid one, length/ width ratio is about 3.1. The dorsal margin of the fore femur has a continuous carina without tubercles. Ventral margin with the same femur has one tubercle close to the femur's distal end. The fore tibia is finely serrated and rectangular in cross-section. Distal segment of the fore tarsus is much longer than the proximal one. Mid legs (Fig. 2A, D). Dorsal and ventral margins of the mid femur bear continuous carinae, but ventral carina also has three small tubercles. Length/width ratio of the mid femur is 3.7. The mid tibia is finely tuberculated and rectangular in cross-section. Distal segment of the fore tarsus is much longer than the proximal one. Hind legs (Fig. 2A, D). The dorsal margin of the hind femur bears three large, but relatively blunt teeth. Ventral margin with continuous carina. Length/width ratio of the hind femur is 2.7. The hind tibia bears numerous strong teeth, but otherwise has a very smooth surface. First segment of the tarsus bears three strongly protruding, smooth and rounded pulvilli. First tarsal segment is much longer than the third.

Measurements (male holotype). Body length (from the tip of head to the tip of the subgenital plate) 11.3 mm . Pronotum length 13.4 mm . Pronotum maximum height 7.6 mm . Pronotum width between lateral lobes 5.2 mm . Pronotum width between the shoulders 3.5 mm . Eye width 0.4 mm . Vertex width 1.2 mm . Fore femur length 2.5 mm . Fore femur width 0.8 mm . Mid femur length 3.3 mm . Mid femur width 0.9 mm . Hind femur length 7.5 mm . Hind femur width 2.8 mm . Hind femur length/width ratio 2.7.

Locus typicus. Madagascar, Sava region, Belanono (rainforest between Sambava and Adapa).

Proposed IUCN Red List Assessment. Similar to the Leatherback Pygmy Grasshopper (Lepocranus fuscus) (Danielczak et al. 2017), the Malagasy Litterhopper (Valalyllum folium gen. et sp. nov.) should be immediately listed as an endangered species in the IUCN Red List because (1) of a very small geographic range it inhabits (known from a single locality, (2) the population might be fragmented and (3) the decline in both the number of mature individuals and the size and quality of the range because of expected severe deforestation.

## Discussion

Despite its relatively small area, Madagascar is one of the richest biodiversity hotspots on Earth, boasting a high number of endemic taxa from the species level and above. This fact characterizes Madagascar as a high-priority area for conservation (Myers et al. 2000). Even at the level of Tetrigidae we find unbelievable diversity, with the new genus being the $24^{\text {th }}$ endemic genus of the total number of 29 (Cigliano et al. 2022). Tragically, Madagascar has a long tradition of deforestation. Only $15 \%$ of the island remains forested, with the northern rainforest being the healthiest (Ganzhorn et al. 2001; Harper et al. 2007;

Vieilledent et al. 2018). The deforestation continues despite efforts to stop it (Tabor et al. 2017). Undescribed species can hardly be protected (Liu et al. 2022) and our models hinge on abundant and reliable data (Tedesco et al. 2014; Régnier et al. 2015). The nomenclatural acts we perform reflect our endeavor to assist in preservation of biodiversity.

The two members of the Valalyllini trib. nov. are similar but can be readily separated by several characters, namely the size and the shape of the eyes, the size of the leaf-like crest, and the general size of the body. Although these features can be variable individually, L. fuscus as a whole appears as a neotenic form of $V$. folium gen. et sp. nov. and is thus likely closely related to it but morphologically consistently separate. A remarkable similarity between the leaf-like tribes Valalyllini trib. nov. (Madagascar), Cladonotini (Asia), and Choriophyllini (Caribbean) is evident, so it is reasonable to hypothesize that those characters represent homologies and reflect the common ancestry on the ancient continent of Gondwana, which is a pattern that has been observed in numerous other taxa (Turner 2004; Gray et al. 2009; Reguero and Goin 2021). Cladonotinae are polyphyletic; their taxonomy is still far from resolved (Zhang et al. 2020). There is a strong possibility that more tribes could be defined within the subfamily and that some of the defined ones could contain genera that better fit elsewhere. The broad picture we present could lead to the solution, but there is more work to be done.

Both Lepocranus fuscus and Valalyllum folium gen. et sp. nov. are known only from a small area. A single digitalized specimen of L. fuscus is not exceptionally well preserved due to the presence of mould (Devriese 1991) and the new species is known only from a single specimen. Clearly, this situation is not ideal as the identity of both species remains incomplete. Lepocranus fuscus is already considered endangered (Danielczak et al. 2017), and could become extinct before we know anything about it. The same could be true for $V$. folium sp. nov. Considering the previously discussed points, a clear distinction between two Malagasy dead-leaf-mimic genera is presented. It is certainly going to be updated in the light of new findings as it is definitely incomplete, but it serves as a valuable starting point for future research.

## Conclusions

A new dead-leaf-like genus and species of Malagasy Tetrigidae, Valalyllum folium gen. et sp. nov. is described and compared to Lepocranus fuscus, the only species of the genus Lepocranus. The two species are similar, but clearly separable by the general size, the shape of the pronotal crest and the shape of the eyes. Valalyllum folium gen. et sp. nov. and Lepocranus fuscus are assigned to the newly described tribe Valalyllini trib. nov. as the only Malagasy tetrigids with leaf-like pronotal crests. The comparison of the present tribe with others of similar morphology has revealed several likely homologies, which imply common ancestry. As the taxonomy of Cladonotinae is not resolved yet, new groups within the subfamily are certainly going to be defined, which will help in elucidating their evolutionary relationships.

Lepocranus fuscus is already considered endangered according to the IUCN Red List and, following the same pattern, we propose that the new species should be considered endangered as well.

Both species from the newly described tribe are defined only by a single specimen each, which is a fact that obscures the variability of the species. However, considering the rampant deforestation of Madagascar and the fact that the species can be differentiated by intraspecifically invariable characters homologous with other leaf-like tribes, we find it vital to describe the new, likely endangered, species and thus assist the efforts to classify and protect the rich Malagasy biodiversity.

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# A new systematic arrangement for the blister beetle genus Eurymeloe (Meloini, Meloidae, Coleoptera) with the description of a new species from Spain 

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#### Abstract

The taxonomic status and subgeneric arrangement of the genus Eurymeloe have been debated for decades. In this work, the internal taxonomy of Eurymeloe is redefined by recognising three subgenera: Eurymeloe for the former Eurymeloe brevicollis species group, Coelomeloe for Eurymeloe tuccia, and Bolognaia Ruiz, García-París, Sánchez-Vialas \& Recuero, subgen. nov., to accommodate the species of the formerly recognised Eurymeloe rugosus species group. Additionally, a new species of the newly described subgenus Bolognaia is described from the Iberian Peninsula based on molecular and morphological traits. The new species, Eurymeloe (Bolognaia) orobates sp. nov., can be distinguished from all other species of Eurymeloe by the following combination of morphological traits: dispersed brownish setae over the body that are arranged in small tufts on the abdominal terga; a small, very transverse pronotum that presents a unique macrosculpture; a deeply and densely punctured integument of the head and pronotum; and the very rugose elytra. The characters displayed by $E$. orobates suggest that the species groups that were previously defined and recognised for Eurymeloe, and that are now integrated within the newly erected subgenus Bolognaia, are non-monophyletic.


## Keywords

Bolognaia subgen. nov., Coelomeloe, Iberian Peninsula, new species, phylogeny, systematics

## Introduction

The taxonomy of Eurymeloe Reitter, 1911, originally described as a subgenus of Meloe Linnaeus, 1758, has been controversial due to both its differential use at the genus (Selander 1985; Sánchez-Vialas et al. 2021) or the sub-genus level (see Bologna 1988, 1991, 2008, 2020a; Bologna et al. 1989; Bologna and Pinto 2001; García-París et al. 2010) and its relationship with the monospecific Coelomeloe Reitter, 1911, which has been considered a synonym of Eurymeloe (Selander 1985; Bologna 2020b; SánchezVialas et al. 2021) or as a separate but closely related subgenus (Bologna 1988, 1991; Bologna et al. 1989; Bologna and Pinto 2001; Di Giulio et al. 2013). Based on morphology, Eurymeloe was defined and recognised as two clearly diagnosable species groups, "E. brevicollis" and "E. rugosus" [Bologna, 1988; as Meloe (Eurymeloe)]. However, according to recent molecular phylogenetic analyses (Sánchez-Vialas et al. 2021), Coelomeloe (a former subgenus of Meloe) and the species of the "E. brevicollis species group" sensu Bologna (1988) constitute a monophyletic assemblage that is the sister clade of the "E. rugosus species group" (sensu Bologna 1988). These analyses have highlighted that, although the generic taxonomic status of Eurymeloe is well supported by molecular and morphological data (Selander 1985; Sánchez-Vialas et al. 2021), its internal taxonomic structure remains in need of reassessment.

The Eurymeloe rugosus species group was comprehensively revised by Bologna (1988), who established two main subgroups based on several morphological traits. The first one, subgroup A or the "E. rugosus subgroup", was formed by species presenting black (or partially brownish) pilosity over the body, an opaque or shiny black integument, deep punctures on the head and pronotum, and rugose elytra. The species forming the second one, subgroup B or the " $E$. murinus subgroup", were characterised by having a general brownish yellow pilosity, greyish black or brownish black (exceptionally pale brown) integument with a matte or satin appearance, shallow punctures on the surface of the head and pronotum, and elytra that are less rugose than those of the first subgroup. In addition to these two subgroups, Ruiz et al. [2010; as Meloe (Eurymeloe)] proposed a third one, the "E. saharensis subgroup", which includes Eurymeloe saharensis (Chobaut, 1898) (widely distributed from the Canary Islands to the Arabian Peninsula) and a closely related species, E. vignai (Bologna, 1990). Compared to the first two subgroups, this last one presents distinctive morphological characters such as a slender appearance, long antennae and legs (especially tarsi), silky black integument, pilosity consisting of short reddish setae, thick but sparse punctures on the head and pronotum, and a very shallow elytral rugosity. To date, ca. 20 Palaearctic species have been included in the E. rugosus species group (Bologna 1988; Ruiz and García-París 2009, 2015; Ruiz et al. 2010). However, very few studies have examined the phylogenetic relationships among them.

Recently, Sánchez-Vialas et al. (2021) showed that the clade corresponding to the $E$. rugosus species group is comprised of at least three main lineages: one represented by Eu rymeloe fernandezi (Pardo Alcaide, 1951) with the other two corresponding generally to Bologna's (1988) subgroups A (E. rugosus subgroup) and B (E. murinus subgroup). How-
ever, as pointed by Ruiz and García-París (2009), the morphological traits defining these groups require further study as the specific composition of each is unclear. Moreover, although Sánchez-Vialas et al. (2021) included several representatives of Bologna's (1988) two subgroups, their study lacked specimens of E. rugosus (Marsham, 1802), i.e., the primary species characterising subgroup A, as well as those of the $M$. saharensis subgroup.

We recently collected specimens representing a new distinctive species of the E. rugosus species group from central Spain. The new species, which can be easily diagnosed by both conspicuous morphological characters and molecular data, is characterised by a combination of morphological traits present in either one or the other of Bologna's (1988) subgroups, suggesting that these subgroups are not monophyletic and that some of the characters used to distinguish them (e.g., body setae colouration) are not diagnostic.

In this study, we (1) describe the new species of Eurymeloe and analyse its phylogenetic relationships by using the Meloini molecular framework of Sánchez-Vialas et al. (2021), together with other published (Rulik et al. 2017; Ohnishi et al. 2021) and unpublished data, including sequences of $E$. rugosus; (2) describe a new subgenus within Eurymeloe, Bolognaia subgen. nov., to accommodate the species of the $E$. rugosus species group (sensu Bologna 1988, 1991); (3) redefine, in a more restricted sense, the subgenus Eurymeloe; and (4) discuss the internal taxonomic structures within the new subgenus.

## Materials and methods

We studied the external morphology of a total of 326 specimens belonging to 17 species of Eurymeloe. All specimens are listed in either Ruiz and García-París $(2009,2015)$ or Sánchez-Vialas et al. (2021), and housed in collections at the Museo Nacional de Ciencias Naturales (MNCN-CSIC), Madrid, Spain; the Museu de Zoologia, Barcelona, Spain (MZB); and the Natural History Museum, London, England (NHMUK); or in the personal collections of M. A. Bologna, University "Roma Tre", Rome, Italy (MAB); and J. L. Ruiz, Ceuta, Spain (JLR). A specimen from Biel (Zaragoza) recorded as Meloe rugosus (currently Eurymeloe rugosus, see Sánchez-Vialas et al. 2021) by Recalde et al. (2002) and Pérez-Moreno et al. (2003) and housed in the entomological collection of the Sociedad Entomológica Aragonesa (SEA) [Maynar-Duplá Collection], Zaragoza, Spain, was also studied. In addition, we examined specimens of Eurymeloe gomari (Ruiz and García-París 2009) from Morocco, E. ganglbaueri (Apfelbeck, 1907) from Sardinia (Italy), and a new species of Eurymeloe from central Spain. Comparisons with the remaining species included in Bologna's (1988) E. rugosus species group (i.e., those distributed in the Middle East and the eastern Mediterranean) were made using diagnostic morphological traits extracted from the literature (mainly Kaszab 1958, 1983; Bologna 1988, 1991; Ruiz and García-París 2009, 2015; Ruiz et al. 2010). The geographic distributions of the studied species were also extracted from the literature, mostly from studies by Bologna (1988, 1991, 2008, 2020a).

The description of the new species of Eurymeloe is based on a total of five specimens (one male, dried preserved, and four females, ethanol-preserved), all belonging to
the type series. These specimens were collected from the mountains of central Spain, at Puerto de la Quesera (Province of Guadalajara, Autonomous Community of CastillaLa Mancha). The type series is held at MNCN-CSIC.

For the morphological study, dry-mounted specimens were examined under a stereomicroscope. The male specimen was rehydrated in water before the extraction of the genital structures, which was subsequently mounted on a piece of cardboard using dimethylhydantoin formaldehyde (DMHF) resin and pinned adjacent to the specimen. Measurements were taken using a micrometre that was coupled to one of the microscope eyepieces. Digital images of live, dry-mounted specimens, and of male and female genital structures, were taken with a reflex camera (Canon 77D) fitted with a macro-lens (Sigma 105 mm F2.8) and two external flashes. We used the terminology suggested by Selander (1966) to describe the various parts of the male genitalia.

For the molecular analyses, we used sequences of Eurymeloe available from GenBank and those of four new specimens that we had collected and preserved in ethanol (now housed at the MNCN-CSIC), including one from Morocco corresponding to Eurymeloe gomari, one from Sardinia (Italy) corresponding to E. ganglbaueri, and two from central Spain corresponding to the new species, Eurymeloe sp. nov. From GenBank, we downloaded the sequences of 31 specimens belonging to 13 species of Eurymeloe (Rulik et al. 2017; Ohnishi et al. 2021; Sánchez-Vialas et al. 2021) (Table 1). A total of 31 specimens of 16 species from ten genera of Meloidae was used as the outgroup (Sánchez-Vialas et al. 2021) (Table 1). Tissue sampling, DNA extraction, sequencing, and alignments were performed as described by Sánchez-Vialas et al. (2021).

The molecular data set consisted of two mitochondrial (COI and 16S rRNA) and two nuclear ( Wg and 18 S rRNA ) gene fragments from 66 specimens (including the four new ones). All sequences were compiled and revised using Sequencer v. 4.9 and aligned with MAFFT (Katoh and Toh 2008). Final alignments were visually inspected with Mesquite v. 3.04 (Maddison and Maddison 2019). Phylogenetic analyses using a Bayesian inference approach, as implemented in MrBayes v. 3.2.3 (Ronquist et al. 2012), were performed on a combined data set consisting of 2917 bp from the four mitochondrial and nuclear sequences (COI, 16S, Wg, 18S) (Table 1). Analyses, which started with a randomly generated tree, consisted of four Metropolis-coupled Markov chains Monte Carlo (one cold, three heated) and two simultaneous runs of $10 \times 10^{6}$ generations each, sampling every 1000 generations. We discarded the first $25 \%$ of the obtained trees as burn-in and generated the consensus tree in MrBayes. Posterior clade probabilities were used to assess nodal support.

To delimit species, we adopted the evolutionary species concept (Wiley 1978; Wiley and Mayden 2000) in which a species is considered "a single lineage of ancestral descendant populations of organisms which maintains its identity from other such lineages and which has its own evolutionary tendencies and historical fate" (Wiley 1978: 18). This concept combines implications derived from the phylogenetic species concept, such as reciprocal monophyly, with additional subjective properties (e.g., phenetic distinguishability and reproductive isolation, among other lines of evidence) that can be used to assess the historical fate of lineages (Ruiz and García-París 2015; Sánchez-Vialas et al. 2020).

Table I. Specimen identity, collection locality, voucher number, and GenBank accession numbers of the new and previously published sequences analysed in this work.

| Taxon | Locality | Voucher number | GenBank \# CoxI | $\begin{gathered} \hline \text { GenBank \# } \\ 16 S \\ \hline \end{gathered}$ | $\begin{gathered} \text { GenBank } \\ \# W g \end{gathered}$ | $\begin{gathered} \hline \text { GenBank \# } \\ 18 \mathrm{~S} \\ \hline \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Eurymeloe apivorus | Morocco: Fès-Meknès: Ifrane, 2 km South of Cedro Gouran, Middle Atlas | mel 81009 | MW158218 | MW158046 | MW157964 | MW158119 |
| Eurymeloe apivorus | Morocco: Fès-Meknès: Ifrane, 2 km South of Cedro Gouran, Middle Atlas | mel 81013 | MW158220 | MW158048 | MW157966 | MW158121 |
| Eurymeloe apivorus | Morocco: Fès-Meknès: Ifrane, 2 km South of Cedro Gouran, Middle Atlas | mel 81054 | MW158219 | MW158047 | MW157965 | MW158120 |
| Eurymeloe brevicollis | Spain: Cantabria: Brañavieja, Pico Tres Mares | mel 04107 | MW158305 | MW158088 | MW157987 | MW158142 |
| Eurymeloe brevicollis | Andorra: Arinsal | mel 07092 | MW158306 | MW158089 | MW157988 | MW158143 |
| Eurymeloe corvinus | Japan: Niigata, Sado, Kitaushima | 9060 | LC583106.1 |  |  |  |
| Eurymeloe corvinus | Japan: Saga, Karatsu, Hirose | 11881 | LC583105.1 |  |  |  |
| Eurymeloe corvinus | Japan: Nagano, Iriyamabe | 10521 | LC583104.1 |  |  |  |
| Eurymeloe fernandezi | Spain: Islas Canarias: La Palma, Los Sauces | mel 07045 | MW158266 | MW158068 | MW157972 | MW158127 |
| Eurymeloe fernandezi | Spain: Islas Canarias: La Palma, Los Sauces | mel 07048 | MW158267 | MW158069 | MW157973 | MW158128 |
| Eurymeloe ganglbaueri | Italy: Lazio: Tarquinia | mel 81064 | MW158268 | MW158070 | MW157974 | MW158129 |
| Eurymeloe ganglbaueri | Italy: Lazio: Tarquinia | mel 81065 | MW158269 | MW158071 | MW157975 | MW158130 |
| Eurymeloe ganglbaueri | Italy: Sardinia: 4 km North-West of Orgosolo | ASV19011 |  | OM918705 |  | OM925566 |
| Eurymeloe glazunovi | Romania: Dobruja: Istria | mel 07001 | MW158265 | MW158067 | MW157971 | MW158126 |
| Eurymeloe gomari | Morocco: 9 km South-West of Moulay Abdeselam | ASV18019 | OM936883 | OM918704 |  | OM925565 |
| Eurymeloe gomari | Morocco: Tangier-Tetouan: Chaouen, Talassemtane National Park | mel 81063 | MW158275 | MW158076 | MW157981 | MW158136 |
| Eurymeloe ibericus | Spain: Ávila: Villanueva del Campillo, Puerto de Villatoro | mel 06039 | MW158307 | MW158090 | MW157989 | MW158144 |
| Eurymeloe ibericus | Spain: Ávila: Hoyos del Espino, Plataforma de Gredos | mel 81039 | MW158308 | MW158091 | MW157990 | MW158145 |
| Eurymeloe mediterraneus | Spain: Cádiz: Puerto Real | mel 04255 | MW158221 | MW158049 | MW157967 | MW158122 |
| Eurymeloe mediterraneus | Morocco: Moulay Abdelsalam | mel 07010 | MW158222 | MW158050 | MW157968 | MW158123 |
| Eurymeloe mediterraneus | Spain: Cuenca: Saelices | mel 07147 | MW158224 | MW158052 | MW157970 | MW158125 |
| Eurymeloe mediterraneus | Morocco: Tetouan: Agnan, Sierra del Haus | mel 81066 | MW158223 | MW158051 | MW157969 | MW158124 |
| Eurymeloe murinus | Morocco: Marrakesh-Safi: 2 km North of Aguelmouse, Tizi n'Tichka, High Atlas | mel 81018 | MW158270 | MW158072 | MW157976 | MW158131 |
| Eurymeloe murinus | Spain: Madrid: Colmenar Viejo | mel 81053 | MW158271 |  | MW157977 | MW158132 |
| Eurymeloe nanus | Spain: Madrid: 7 km South of Tielmes | mel 01028 | MW158273 | MW158074 | MW157979 | MW158134 |
| Eurymeloe nanus | Spain: Madrid: Tielmes | mel 81042 | MW158274 | MW158075 | MW157980 | MW158135 |
| Eurymeloe nanus | Spain: Toledo: Villacañas, Sierra del Romeral | mel 05001 | MW158272 | MW158073 | MW157978 | MW158133 |
| Eurymeloe orobates sp. nov. | Spain: Guadalajara: Puerto de la Quesera | ASV18002 | OM936884 |  | OM925567 |  |
| Eurymeloe orobates sp. nov. | Spain: Guadalajara: Puerto de la Quesera | ASV18003 | OM936885 |  | OM925568 |  |
| Eurymeloe rugosus | Germany: Saxony-Anhalt | $\begin{aligned} & \text { ZFMK- } \\ & \text { TIS-2003300 } \end{aligned}$ | KU918912.1 |  |  |  |
| Eurymeloe tuccia | Spain: Menorca: 2 km South of Binimella | mel 06034 | MW158276 | MW158077 | MW157982 | MW158137 |


| Taxon | Locality | Voucher number | GenBank \# CoxI | $\begin{gathered} \text { GenBank \# } \\ 16 \mathrm{~S} \end{gathered}$ | $\begin{gathered} \text { GenBank } \\ \text { \# Wg } \\ \hline \end{gathered}$ | $\begin{gathered} \text { GenBank \# } \\ 18 \mathrm{~S} \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Eurymeloe tuccia | Spain: Islas canarias: La Palma: Don Pedro | mel 07058 | MW158277 | MW158078 | MW157983 | MW158138 |
| Eurymeloe tuccia | Spain: Almería: La Mela | mel 81001 | MW158278 | MW158079 | MW157984 | MW158139 |
| Eurymeloe tuccia | Portugal: Algarve: Sagres, Praia do Martinhal | mel 81002 | MW158279 | MW158080 | MW157985 | MW158140 |
| Eurymeloe tuccia | Morocco: Larache: Lixus | mel 81006 | MW158280 | MW158081 | MW157986 | MW158141 |
| Lampromeloe aff. variegatus | Morocco: Marrakesh-Safi: 5.5 km <br> North-East of Aguelmouse, Tizi n’Tichka, High Atlas | mel 81010 | MW158202 | MW158033 | MW157953 | MW158108 |
| Lampromeloe cavensis | Morocco: Casablanca-Settat: Ouled Bahmad | mel 06011 | MW158201 | MW158032 | MW157952 | MW158107 |
| Lampromeloe variegatus | Spain: Salamanca: 5 km West of Palencia de Negrilla | mel 05015 | MW158203 | MW158034 | MW157954 | MW158109 |
| Lampromeloe variegatus | Hungary: Komárom-Esztergom: Vèrtesszölös | mel 81068 | MW158204 | MW158035 | MW157955 | MW158110 |
| Meloe (Anchomeloe) autumnalis | Spain: Guadalajara: Villanueva de la Torre | mel 04246 | MW158189 | MW158025 | MW157949 | MW158104 |
| Meloe (Anchomeloe) autumnalis | Spain: Zaragoza: El Frago | mel 10070a | MW158191 | MW158027 | MW157951 | MW158106 |
| Meloe (Anchomeloe) autumnalis | Morocco: Tangier-Tetouan: Chaouen, Djebel Tissouka | mel 81071 | MW158190 | MW158026 | MW157950 | MW158105 |
| Meloe (Meloe) proscarabaeus | Spain: Menorca: 3.5 km SouthWest of Fornells | mel 06026 | MW158148 | MW157993 | MW157939 | MW158094 |
| Meloe (Meloe) proscarabaeus | Morocco: Souss-Massa: 2 km South of Chafarni | mel 81007 | MW158150 | MW157995 | MW157941 | MW158096 |
| Meloe (Meloe) proscarabaeus | Hungary: Tolna: Bátaapáti | mel 06004 | MW158151 | MW157996 | MW157942 | MW158097 |
| Meloe (Meloe) proscarabaeus | Italy: Tuscany: Alberese | mel 81082 | MW158149 | MW157994 | MW157940 | MW158095 |
| Meloe (Meloe) tropicus | Guatemala: El Quiché: 9 km North-East of Santa Cruz Quiché | mel 81075 | MW158188 | MW158024 | MW157948 | MW158103 |
| Meloe (Meloe) cf. violaceus | Hungary: Tolna: Mőcsény | mel 07033 | MW158186 | MW158022 | MW157946 | MW158101 |
| Meloe (Meloe) cf. violaceus | Hungary: Vas: Csákánydoroszló | mel 07036 | MW158187 | MW158023 | MW157947 | MW158102 |
| Meloe (Meloe) violaceus | Spain: Ávila: Hoyos del Espino | mel 05024 | MW158183 | MW158019 | MW157943 | MW158098 |
| Meloe (Meloe) violaceus | Spain: León: Correcillas: Pico Polvareda | mel 81051 | MW158185 | MW158021 | MW157945 | MW158100 |
| Meloe (Meloe) violaceus | Spain: León: Correcillas: Pico Polvareda | mel 81052 | MW158184 | MW158020 | MW157944 | MW158099 |
| Meloegonius cicatricosus | Hungary: Pest: Tatárszentzgyörgy | mel 06002 | MW158208 | MW158038 | MW157959 | MW158114 |
| Meloegonius cicatricosus | Hungary: Komárom-Esztergom: Vèrtesszölös | mel 81069 | MW158209 | MW158039 | MW157960 | MW158115 |
| Mesomeloe coelatus | Morocco: Guelmine-Smara: Reg Labyad | Mcoelatus_ labyad | MW805179 |  |  |  |
| Mesomeloe coelatus | Morocco: Guelmine-Smara: Jbel Ouarkziz | Mcoelatus_ ouarkziz | MW805180 |  |  |  |
| Mesomeloe ottomerkli | Qatar: 1.8 km West of Al Marrawnah | mel Qatar1 | HG003653 | MW158044 | MW157962 | MW158117 |
| Mesomeloe ottomerkli | Qatar: 1.8 km West of Al Marrawnah | mel Qatar2 | HG003654 | MW158045 | MW157963 | MW158118 |
| Physomeloe corallifer | Spain: Madrid: 5 km South-East of Agustín de Guadalix, 618 m as | mel 09051 | MW158210 | MW158040 | MW157961 | MW158116 |
| Taphromeloe erythrocnemus | Morocco: 2.5 km North-West of Douar Azerzou | ASV18007 | MW158309 | MW158092 | MW157991 | MW158146 |
| Treiodous gracilicornis | Guatemala: El Quiché: 3.4 km North of Uspantán | mel 81077 | MW158205 | MW158036 | MW157956 | MW158111 |
| Treiodous gracilicornis | Mexico: Guerrero: 5 km West of Carrizal de Bravo | mel 874 | MW158206 |  | MW157957 | MW158112 |


| Taxon | Locality | Voucher <br> number | GenBank \# <br> CoxI | GenBank \# <br> $\mathbf{1 6 S}$ | GenBank <br> \# Wg | GenBank \# <br> 18S |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Treiodous laevis | Mexico: Guanajuato: Dolores | mel 08094 | MW158207 | MW158037 | MW157958 | MW158113 |
| Lytta vessicatoria | Hidalgo | Spain: Ourense: A Acea | mel 05073 | MW158147 | MW157992 |  |
| Phodaga alticeps | Mexico: Baja California Norte: <br> Ejido Luchadores del Desierto, <br> Northwest of Laguna Salada | KRN14 | MK024506 | MK024642 | MW158093 |  |
|  |  |  |  | MK024601 |  |  |
| Cordylospasta fulleri | USA: California: 4.8 km North- <br> East of Big Pine | KRN23 | MK024478 | MK024619 |  | MK024572 |
|  |  |  |  |  |  |  |

## Results

## Phylogenetic relationships within Eurymeloe

Our Bayesian phylogenetic tree, which has a topology similar to the one presented by Sánchez-Vialas et al. (2021), recovered three major lineages within the genus Eurymeloe (Fig. 1). The first lineage is represented by the type species of the genus Eurymeloe, E. brevicollis (Panzer, 1793), plus the species E. ibericus (Reitter, 1895) and E. corvinus (Marseul, 1877), which together form a clade with E. tuccia (Rossi, 1790), the type species of the subgenus Coelomeloe. These two clades together form the sister group of the clade comprising the remaining species of Eurymeloe that were previously included in the $E$. rugosus species group (sensu Bologna 1988).

With respect to the internal taxonomic structure of Eurymeloe (sensu SánchezVialas et al. 2021), it consists of three main clades, all morphologically diagnosable, with the following subgeneric status: (1) Coelomeloe; (2) Eurymeloe sensu stricto (s. str.), which includes the species in Bologna's (1988) E. brevicollis species group, and (3) the lineage including all species within the $E$. rugosus species group (Fig. 1). As the namebearing type species of Eurymeloe (E. brevicollis) lies within the E. brevicollis species group, Eurymeloe (s. str.), the clade formed by the E. rugosus species group requires a new subgeneric name (as there is no name available for this group). We propose and describe herein a new subgenus of Eurymeloe to accommodate the species of Bologna's M. rugosus species group (sensu Bologna 1988): Bolognaia subgen. nov. We also redefine the subgenus Eurymeloe.

Bolognaia is comprised of three main lineages: one represented by E. fernandezi, an endemic of the Canary Islands, which resolved as the sister taxon to the other two lineages formed, respectively, by $E$. mediterraneus (Müller, 1925) and its closely related species and by E. murinus (Brandt and Erichson 1832) and its related species, including the new species of Eurymeloe from central Spain. The sequences of the new samples from Spain resolved as a distinctive lineage that is the sister taxon to the clade formed by $E$. murinus and $E$. ganglbaueri. This new lineage is not morphologically congruent with any other described species of Eurymeloe (Fig. 2).

Lastly, our molecular phylogenetic analysis confirmed the species identification of the other two newly sequenced specimens as E. gomari and E. ganglbaueri, and their relationship with the other specimens. As a result, the northern limit of


Figure I. Bayesian phylogeny of the genus Eurymeloe based on the concatenated matrix of mitochondrial and nuclear genes (COI, 16S, Wg, 18S). The three subgeneric taxonomic categories are indicated as follows: Eurymeloe (in green), Coelomeloe (in violet) and Bolognaia (in blue). Numbers on the nodes represent the posterior probabilities of the clades. Letters in parentheses (A-J) correspond to the species portrayed in Fig. 2. Phylogeny in lower left shows the outgroups (black) and the lineage of Eurymeloe (orange). Additional information for each specimen is provided in Table 1.
the distribution range of the Moroccan endemic E. gomari, which was previously known only from its type locality, has been expanded by approximately 37 km to the northwest along the Rif Mountains. The new specimen of E. ganglbaueri from Sardinia resolved, as expected, within the same clade as the conspecific Italian continental samples.

## Redefinition of the genus Eurymeloe

Given the new systematic arrangement proposed here for Eurymeloe as three subgenera (Eurymeloe, Coelomeloe, and Bolognaia subgen. nov.), we consider it necessary to present a revised diagnosis of Eurymeloe based on the morphology of adult specimens.


Figure 2. Habitus of some living adult specimens of the current subgenera of Eurymeloe $\mathbf{A}$ subgenus Eurymeloe, represented by a specimen of Eurymeloe (Eurymeloe) ibericus from Puebla de la Sierra, northern Madrid, Spain B subgenus Coelomeloe, represented by a specimen of Eurymeloe (Coelomeloe) tuccia found 3 km east of Celín, Sierra de Gádor, Almería, Spain C-I subgenus Bolognaia C Eurymeloe (Bolognaia) affinis from the surroundings of Bab Taza, Rif Mountains, Morocco D E. (Bolognaia) mediterraneus (type species of the subgenus Bolognaia) from Puebla de la Sierra, northern Madrid, Spain E E. (Bolognaia) glazunovi from Istria, Romania F E. (Bolognaia) gomari from the surroundings of Bab Taza, Rif Mountains, Morocco G E. (Bolognaia) nanus from Alcázar de San Juan, Ciudad Real, Spain H E. (Bolognaia) fernandezi from Los Sauces, La Palma, Canary Islands I Eurymeloe orobates sp. nov., from Puerto de la Quesera, Guadalajara, Spain J Eurymeloe (Bolognaia) murinus from Tizi n`Tichka, High Atlas, Morocco. Photographs: A-C, F, G, I (ASV) E, H, J (MGP) D (J. Aznar González de Rueda).

## Genus Eurymeloe Reitter, 1911

Type species. Meloe brevicollis Panzer, 1793, by subsequent designation of Pinto and Selander (1970).

Description (adult). Size small to medium ( $6-36 \mathrm{~mm}$ ), with diverse appearance, ranging from very robust to comparatively slender. Body integument colour variable, black, dull grey or dark brown (exceptionally sandy brown) to moderately metallic blue, opaque, bright, silky or sometimes with an oily shininess (Fig. 2A). Body pubescence short, sometimes quite distinct (Bolognaia) or very short, recumbent, often almost imperceptible (subgenus Eurymeloe) or absent dorsally (Coelomeloe; Fig. 2B), variable in colour, from yellowish to reddish brown and black. Head rounded, sides straight to arched, convergent to eyes. Eyes small or medium, usually subreniform, variably protruding, without longitudinal depressions behind them. Antennae unmodified in males, moniliform or submoniliform, robust or slender, short to moderate length, not reaching posterior margin of pronotum (Eurymeloe, Coelomeloe) or often reaching or even surpassing it (Bolognaia). Antennomeres subcylindrical or subconical, relatively robust or slender, highly variable width/length ratio, V to VII neither enlarged nor geniculated. Clypeus transverse, approximately twice as wide as long. Frontoclypeal suture angulated. Labrum wide, fore margin broadly emarginate. Maxillary and labial palpomeres unmodified. Mandibles robust, regularly and strongly curved on the outer margins. Pronotum from flat (Coelomeloe) to moderately convex (Eurymeloe, Bolognaia), subrectangular (Coelomeloe, Eurymeloe), subhexagonal or trapezoidal (Bolognaia), transverse or very transverse, usually equal to or more than $1.5 \times$ as wide as long (exceptionally, $1.2-1.3 \times$ as wide as long), with sides subparallel or converging backward; posterior margin usually broadly emarginated, with base not incised in the middle. Head and pronotum punctation from fine and scattered, sometimes almost absent (Eurymeloe), to somewhat deep and dense (Bolognaia), even very broad, dense, and deep, foveate in appearance (Coelomeloe). Hind margin of mesonotum straight or weakly arcuate. Metanotum short and barely visible, covered by the base of the elytra. Mesosternum short and wide, lacking scutum. Mesepisterna meet or not at the midline of the body. Elytra short and dehiscent, imbricate basally, not completely covering abdomen, smooth to densely coriaceous, rugose (Eurymeloe, Bolognaia) or with a surface densely foveate (Coelomeloe). Hind wings absent. Legs normal, unmodified in male, robust or more or less slender, pilose. Tibiae with two spurs at apex; outer spur of metatibiae widened and obliquely truncate, spoon-shaped. Tarsomeres with or without hair pads or dense setose pubescence on the inferior sides. Tarsal claws smooth, with distinct lower blades. Abdomen large, inflated, hypertrophied. Abdominal terga with medium or small highly sclerotised central plates. Last abdominal ventrite broadly emarginated at hind margin in males. Male genitalia: gonoforceps evenly sclerotised with gonostyli from moderately short to elongate, with distal regions more or less wide, usually digitiform (in lateral view), rounded at apex; gonocoxal plate broadly widened at the middle (in dorsal view); aedeagus robust (Eurymeloe, Coelomeloe) or relatively slender (Bolognaia), usually shorter than or approximately equal in length
to the gonoforceps (in some species of Bolognaia, sometimes a little longer than the gonoforceps), with two dorsal hooks and one endophallic hook.

Larva. The morphological traits of previously known first instar larvae of Eurymeloe (triungulines), including Coelomeloe, and the descriptions of the triungulines of several additional species of Eurymeloe, have been synthesised and studied by Di Giulio et al. (2013, 2014). We herein refer to these works for larval morphological traits.

Taxonomic remarks. The adult instar of species of the genus Eurymeloe is morphologically diverse, and the three subgenera can be recognised based on this diversity. Adults of the subgenera Eurymeloe, Coelomeloe, and the newly described Bolognaia are distinguishable particularly by the shape of the antennae and the pronotum, the macrosculpture of the pronotum, body integument and pilosity, and the punctation of the head, pronotum, and elytra, among other traits.

## Description of the subgenus

## Bolognaia Ruiz, García-París, Sánchez-Vialas \& Recuero, subgen. nov. https://zoobank.org/6B062E01-EF30-47F3-8ED2-42394FE7D532

Type species. Meloe mediterraneus Müller, 1925, by present designation.
Description (adult). Size small or medium to large ( $8-36 \mathrm{~mm}$ ). Body integument black, dull grey or dark brown, occasionally sandy brown [E. pallidicolor (Martínez de la Escalera, 1909)], with an opaque, silky or bright appearance, never bluish or metallic (Fig. 2C-J). Body pubescence quite distinct, black, yellowish, whitish or golden, short or very short. Head rounded, temples usually forming a regular arc, except in $E$. murinus (Brandt and Erichson 1832) and E. affinis (Lucas, 1847), which have strongly enlarged temples; occiput usually weakly concave. Medium-sized eyes [smaller in E. affinis and E. apivorus (Reitter, 1895)], subreniform, moderately protruding, without a longitudinal depression behind them. Antennae moniliform, normally slender, not thickened towards the apex; long or medium in length, usually reaching the posterior margin of the pronotum or exceeding it; unmodified in males, straight. Antennomeres IV-IX subcylindrical, always longer than wide. Clypeus transverse, approximately twice as wide as long. Labrum wide, fore margin broadly emarginate. Mandibles robust, regularly curved along the outer margins. Pronotum slightly convex, transverse, mainly subhexagonal or subtrapezoidal, wider than long, usually $1.4-2.1 \times$ as wide as long [in E. fernandezi, 1.2-1.3 $\times$ as wide as long], sides not parallel, converging backward, posterior margin broadly emarginated, posterior corners rounded. Pronotum surface variable, with or without a depressed area or groove in the middle, frequently with two depressed or smooth areas, diffuse, on both sides of the disc. Head and pronotum punctation fine to coarse, of variable density, always with pubescence. Posterior margin of mesonotum straight or weakly arcuate. Mesepisterna usually not meeting at the midline of the body. Elytra short and dehiscent, smooth to strongly rough, usually rugulose. Legs normal, usually slender, pubescent. Tarsomeres without hair pads on the inferior side, though some species [e.g., E. nanus (Lucas, 1847), E. baudueri (Grenier,
1863), E. gomari] have fairly dense setose pubescence that appears as small and short brushes. Last abdominal ventrite broadly and deeply emarginated at the hind margin in males. Male genitalia: Gonostyli usually elongate, distal regions narrow with their apices acuminate or digitiform in lateral view; gonocoxal plate long, usually narrow and slightly widened at the middle in dorsal view; aedeagus usually elongate, equally long as or longer than gonoforceps.

Etymology. The name Bolognaia, formed by the noun "Bologna" plus the Italian suffix "-aia" derived from the Latin "-aria" (used, in this case, to form a word meaning an animal associated with the specified noun Bologna), is in honour of Marco A. Bologna, a distinguished Italian entomologist specialising in the Meloidae and a friend who, among other excellent works, was able to clarify, for the first time, the complex taxonomy of the small-sized species of Eurymeloe related to E. rugosus for which the new subgenus is here erected.

Taxonomic remarks. We selected M. mediterraneus as the type species of Bolognaia because it is a morphologically well-characterised species, with low morphological or genetic intraspecific geographic differentiation (Bologna 1988, 1991; SánchezVialas et al. 2021), and without nomenclatural or identity problems associated to synonyms (García-París et al. 2010; Bologna 2020a). Bolognaia, a monophyletic subgenus, largely corresponds to the E. rugosus species group of Eurymeloe defined by Bologna (1988, 1991). It includes species whose adults are characterised mainly by the following traits: small or medium body size; black, dull grey, or dark brown body colour; a distinctive black or pale-coloured (yellowish, whitish, or golden) pilosity; elongated and subcylindrical antennomeres that are longer than wide; and generally marked punctation and rugosity.

Based on molecular data (this work) and adult morphology (see Reitter 1895; Martínez de la Escalera 1909; Pliginskij 1910; Pardo Alcaide 1951; Kaszab 1958, 1983; Bologna 1988, 1991, 1994a, 1994b; Ruiz and García-París 2009, 2015; García-París and Ruiz 2011; Di Giulio et al. 2013), we include within the subgenus Bolognaia the following species: Eurymeloe (Bolognaia) affinis (Lucas, 1847), E. (B.) apivorus (Reitter, 1895), E. (B.) apenninicus (Bologna, 1988), E. (B.) baamarani (Ruiz and GarcíaParís 2015), E. (B.) baudii (Leoni, 1907), E. (B.) baudueri (Grenier, 1863), E. (B.) fernandezi, E. (B.) flavicomus (Wollaston, 1854), E. (B.) ganglbaueri, E. (B.) glazunovi (Pliginskij, 1910), E. (B.) gomari, E. (B.) kandaharicus (Kaszab, 1958), E. (B.) mediterraneus, $E$. (B.) murinus, E. (B.) nanus, E. (B.) omanicus (Kaszab, 1983), E. (B.) pallidicolor, and $E$. (B.) rugosus.

According to our molecular analyses (Fig. 1), three sublineages can be recognised within Bolognaia: two generally corresponding to Bologna's (1988) subgroups A and B, defined by presenting, respectively, an entirely black body pilosity (in our analyses, E. mediterraneus, E. apivorus, and E. glazunovi) or a pale-coloured (whitish, yellowish, reddish yellow, or golden) pilosity over the entire body or parts of it [in our analyses, E. ganglbaueri, E. murinus, E. nanus, E. gomari, E. rugosus, and Eurymeloe orobates sp. nov. from central Spain]. Notably, based on our molecular analyses, E. rugosus, which was included in Bologna's (1988) subgroup A, appears to be genetically more related to
the species included in his subgroup B. In this regard, following a detailed examination of some specimens belonging to $E$. rugosus, we observed that several have inconspicuous brownish and reddish to yellowish setae (but not tufts) on their abdominal tergites, similar to those observed on the morphologically related species E. apenninicus (JLR, pers. obs.). In fact, Escherich $(1890)$ and Bologna $(1988,1991)$ pointed out that some specimens of $E$. rugosus show yellowish brown setae on the last abdominal tergites; these correspond to the named var. abdominalis (Escherich 1890) (which has even been confused with E. ganglbaueri, see Bologna 1988: 247). Likewise, E. ganglbaueri, which presents a golden-yellow pilosity on a part of the body, resolved as genetically more related to species in subgroup B. The third sublineage diverged from its sister group, the A and B sublineages, during the Middle Miocene (Sánchez-Vialas et al. 2021). This sublineage is composed of only one species, E. fernandezi, an endemic of the Canary Islands that is morphologically singular and isolated within the subgenus (Pardo Alcaide 1951; Bologna 1988, 1991, 1994a; Ruiz and García-París 2015).

For practical purposes, but also supported by our analyses and some morphological traits (mainly, pilosity colour, integument aspect, pronotum punctation, and elytra rugosity; see Bologna 1988; Ruiz and García-París 2009, 2015), we redefine the specific composition of the subgroups established by Bologna (1988) within Bolognaia (defined as the E. rugosus species group by Bologna 1988) as follows:
(1) group A or E. mediterraneus group (now renamed), characterised mainly by a dark body pilosity (black or dark brown) and a black body integument that is usually glossy, semi-glossy, or silky in appearance [exceptionally, it is matte as in $E$. (B.) baamarani]. This group integrates the following species: $E$. (B.) affinis, E. (B.) apivorus, E. (B.) baamarani, E. (B.) baudii, E. (B.) glazunovi, and E. (B.) mediterraneus. In some specimens of $E$. (B.) mediterraneus, particularly those from Sardinia (Bologna 1988, 1991), the pilosity of the temples, pronotum and, sometimes, abdomen, is brown. The unstudied E. (B.) affinis setosus Escherich, 1890 from Algeria, which differs from the typical form of the species by the presence of isolated yellowish setae along the abdominal tergites, among other traits (e.g., smaller size, constant frontal furrow, and different elytral sculpture) (Escherich 1890; Bologna 1988, 1991; Di Giulio et al. 2013), possibly constitutes a distinct species, as mentioned by Di Giulio et al. (2013) and previously suggested by Escherich (1890) and Peyerimhoff (in Cros 1934: 90).
(2) group B or E. murinus group, characterised mainly by a pale-coloured (reddish, golden, brownish, yellowish, or whitish) body pilosity, either over the entire body or parts of it, and a body integument that is usually greyish, greyish black, or dark brown and opaque; exceptionally, it is glossy black as in $E .(B$.$) apenninicus and$ $E$. (B.) rugosus. This group comprises the following species: $E$. (B.) apenninicus, $E$. (B.) baudueri, E. (B.) flavicomus, E. (B.) ganglbaueri, E. (B.) gomari, E. (B.) kandaharicus, $E$. (B.) murinus, E. (B.) nanus, E. (B.) omanicus, E. (B.) pallidicolor, and $E$. (B.) rugosus. Within this group, $E$. (B.) apenninicus and $E$. (B.) rugosus can be clearly differentiated from the others by having a glossy black body integument and dark reddish brown
(sometimes almost black) body setation, with scattered and sparse yellowish brown short setae on the abdominal tergites that are often barely noticeable.
(3) group C, composed by only E. (B.) fernandezi, well characterised morphologically within Bolognaia (see Pardo Alcaide 1951; Ruiz and García-París 2015).

Bologna (1988, 1990) integrated M. saharensis (= M. otini Peyerimhoff, 1949, M. marianii Kaszab, 1983; see Ruiz et al. 2010; Bologna 2020a) and M. vignai in the E. rugosus species group. Ruiz et al. (2010) considered the closely related E. saharensis and E. vignai as morphologically isolated and proposed a new group for them. As neither $E$. saharensis nor E. vignai have been studied at the molecular level, we have tentatively ascribed them to Bolognaia as Eurymeloe (Bolognaia) saharensis (Chobaut, 1898) and E. (B). vignai (Bologna, 1990).

Sánchez-Vialas et al. (2021) considered the six Asian species that Bologna (1988) included in the E. rugosus species group as belonging to Eurymeloe. These species are Eurymeloe heptapotamicus (Pliginski, 1910), E. primaeveris (Kaszab, 1958), E. punjabensis (Kaszab, 1958), E. schmidi (Kaszab, 1978), E. scutellatus (Reitter, 1895), and E. subsetosus (Reitter, 1895). However, the available information on these taxa is currently insufficient to assign them to the subgenus Bolognaia; therefore, they require further study at both the morphological and the molecular levels.

## Redefinition of the subgenus

## Eurymeloe Reitter, 1911

Type species. Meloe brevicollis Panzer, 1793 (by subsequent designation of Pinto and Selander, 1970).

Description (adult). Size small or medium ( $6-30 \mathrm{~mm}$ ), usually robust in appearance. Body integument colour black to moderately metallic blue, bright, silky, or with an oily shininess (Fig. 2I). Body pubescence very short, recumbent, or absent on the head and pronotum. Head rounded, sides almost straight, convergent to the eyes. Eyes small, subreniform, weakly protruding, and without longitudinal depression behind them. Antennae submoniliform, robust, short or medium in length, usually not reaching the posterior margin of the pronotum, smoothly thickened towards the apex in some species (e.g., E. brevicollis); in males, unmodified. Antennomeres subcylindrical or subconical, V to VII (in some species IV to IX, e.g., E. brevicollis), wider than long or, at most, as wide as long. Clypeus transverse, approximately twice as wide as long. Labrum wide, fore margin broadly emarginate. Mandibles robust, often curved along the outer margin. Pronotum slightly or moderately convex, very transverse, usually more than $1.7 \times$ wider than long, sides not parallel and obtusely rounded, posterior margin broadly emarginated, posterior corners rounded. Pronotum surface slightly variable, moderately convex, usually with a weak, diffuse, median longitudinal groove. Head and pronotum punctation from fine and scattered, sometimes almost absent, to deep and dense, with or without ( $E$. brevicollis) very
short pubescence. Hind margin of mesonotum straight or weakly arcuate. Elytra short and dehiscent, smooth to densely coriaceous or rugose. Legs normal, robust, pilose. Tarsomeres without hair pads or dense setose pubescence on the inferior side. Last abdominal ventrite broadly emarginated in males. Male genitalia: Gonostyli moderately short, distal regions wide, usually digitiform in lateral view, rounded at apex; gonocoxal plate broadly widened at the middle in dorsal view; aedeagus robust, relatively shorter than the gonoforceps or, at most, similar in length.

Taxonomic remarks. According to the present definition of the subgenus Eurymeloe, it is correlated with the E. brevicollis species group defined by Bologna (1988). It comprises a heterogeneous group of species characterised mainly by the following features in adults: small or medium in size, with a robust appearance; metallic blue or black body colour; reduced pilosity that is very scarce and short, often almost absent; wide antennomeres with V-VII usually wider than long; and variable head and pronotum punctation and elytral rugosity (see Bologna 1988, 1991).

Bologna (1988) tentatively included 22 species in the E. brevicollis species group. However, as this author pointed out, most of these species are very poorly known and, in some cases, the only morphological information on them is from the original description. As a result, the internal taxonomy of Eurymeloe s. str. is very complex and unclear (Bologna 1988).

On the basis of the molecular and morphological data (Reitter 1895, 1911; Escherich 1896; Martínez de la Escalera 1914; Peyerimhoff 1926; Bologna 1988, 1991, 1994a, 1994b; García-París et al. 2010; Di Giulio et al. 2013; this study), we ascribe to Eurymeloe s. str. the following species: Eurymeloe (Eurymeloe) algiricus (Escherich, 1890) (or E. brevicollis algiricus, see Bologna 2008), E. (E.) austrinus (Wollaston, 1854), E. (E.) brevicollis, $E$. (E.) corvinus (possibly co-specific with the previous species according to Di Giulio et al. 2013), E. (E.) crosi (Peyerimhoff, 1926), E. (E.) curticornis (Martínez de la Escalera, 1914) (or E. brevicollis curticornis, see Bologna 2008, 2020a), E. (E.) ibericus, and E. (E.) lederi (Reitter, 1895). The taxonomic positions of E. luctuosus (Brandt \& Erichson, 1832) (related to E. crosi) and E. scabriusculus (Brandt \& Erichson, 1832) (morphologically similar to E. baudii and E. glazunovi, both now included in Bolognaia) are still uncertain (Bologna 1988, 1991), and their assignment to Eurymeloe s. str. requires further studies.

Another 13 species [from Palaearctic Asia, except E. aleuticus (Borchmann, 1942), from the Aleutian Islands] were provisionally assigned by Bologna (1988) to the E. brevicollis species group: Eurymeloe aleuticus, E. curticollis (Kraatz, 1882), E. escherichi (Reitter, 1889), E. frontalis (Reitter, 1905), E. gaberti (Reitter, 1907), E. laevipennis (Brandt \& Erichson, 1832), E. lobicollis (Fairmaire, 1891), E. mandli (Borchmann, 1942), E. mathiesseni (Reitter, 1905), E. primulus (Semenow, 1903), E. servulus (Bates, 1879), E. transversicollis (Fairmaire, 1891), and E. zolotarevi (Pliginskij, 1914). As in the previous case, additional molecular and morphological studies are required to determine the subgeneric assignment of these species.

Regarding other species of Eurymeloe, Shapovalov (2012) described Meloe (Eurymeloe) sarmaticus Shapovalov, 2012 from Russia and Central Kazakhstan and
considered it closely related to the Russian-Kazakh E. aeneus (Tauscher, 1812). The last species, together with E. pusio (Wellman, 1910) and E. asperatus (Tan, 1981), were considered incertae sedis by Bologna (1988). However, recently, Bologna (2020a) integrated them into Eurymeloe (at the subgenus level), although he still considers $E$. aeneus a doubtful ascription. We did not examine material of these species; therefore, we cannot add new information on their current taxonomic placement.

## Key to the subgenera of Eurymeloe

$1 \quad$ Body entirely black and opaque. Body pubescence absent dorsally. Pronotum flat, subrectangular, transverse, depressed in middle of the base, with sides straight, parallel. Punctation of the head and pronotum very broad, dense, subcontiguous (less dense in Sicilian and southern Italian populations) and deep, clearly foveate in appearance (Fig. 2B). Elytral surface smooth, with punctation usually broad, dense and foveate (reduced and barely visible in Sicilian and southern Italian populations). Size medium to large ( $14-31 \mathrm{~mm}$ )

Coelomeloe

- Body black, dull grey or dark brown (exceptionally sandy brown) to moderately metallic blue, bright, silky or more seldom opaque in appearance, sometimes with an oily shininess. Body pubescence quite distinct, or very short, recumbent, often almost imperceptible. Pronotum slightly to moderately convex, wider than long, with sides not parallel, more or less converging backward, and posterior angles usually broadly rounded. Head and pronotum punctation from fine and scattered, sometimes almost absent, to somewhat deep and dense, but never foveolate (Fig. 2A, C-J). Elytral surface smooth to densely coriaceous, subrugose or rugose, not foveolate. Size small to large ( $6-36 \mathrm{~mm}$ ), but usually small to medium ( $6-22 \mathrm{~mm}$ ) 2
2 Body colour black to moderately metallic blue, bright or silky. Overall appearance robust. Body pubescence very short, recumbent, almost imperceptible or even absent on the head and pronotum. Antennae compact, robust, sometimes smoothly thickened towards the apex, short or medium in length, not reaching the posterior margin of the pronotum. Antennomeres subcylindrical or subconical, V to VII (in some species IV to IX) wider than long or, at most, as wide as long.
- Body integument black, dull grey or dark brown, exceptionally sandy brown, with an opaque, silky or bright appearance, never bluish or metallic. Body pubescence quite distinct. Overall appearance more graceful, sometimes moderately robust. Antennae normally slender, not thickened towards the apex, long or medium in length, usually reaching the posterior margin of the pronotum or exceeding it. Antennomeres IV to IX subcylindrical, always longer than wide.

Bolognaia

## Description of a new species of Eurymeloe from the Iberian Peninsula

Our molecular results revealed a distinctive lineage of Eurymeloe nested within the clade comprising E. rugosus, E. murinus, and E. ganglbaueri. This lineage, morphologically distinguishable from all its congeneric species, represents a new species that we herein describe.

## Eurymeloe (Bolognaia) orobates sp. nov.

https://zoobank.org/509BD098-E303-406E-AAE4-052059AC1865

Holotype. adult male (Fig. 3), labelled: "Puerto de la Quesera, Guadalajara, Spain, $41^{\circ} 12^{\prime} 32.2^{\prime \prime N}, 3^{\circ} 24^{\prime} 44.2^{\prime \prime} \mathrm{W}, 1738 \mathrm{~m}, 15-X I-2015$, F. Gutiérrez-Pérez et C. Cano leg." [white label, printed]; "Holotypus Meloe (Bolognaia) orobates Sánchez-Vialas, Ruiz, Recuero, Gutiérrez-Pérez \& García-París des. 2022" [white label, printed]; Holotipo [red label, printed]; MNCN_Ent 324740 [greyish label, printed]. Dissected and mounted genitalia (Fig. 4A-D). Dry-preserved, held at MNCN-CSIC.

Paratypes. four adult females, labelled: two females: "Puerto de la Quesera, Guadalajara, Spain $41^{\circ} 11^{\prime} 30.11 " \mathrm{~N}, 3^{\circ} 24^{\prime} 27.55^{\prime \prime} \mathrm{W}, 1625 \mathrm{~m}, 22-\mathrm{V}-2016, \mathrm{M}$. García París, A. Fernández Liger, A. Corral Lou leg. [white label, printed]; ASV 18002 and ASV 18003, respectively [white label, handwritten]; MNCN_Ent 325407 and $\mathrm{MNCN}_{-}$ Ent 325408, respectively [white label, printed]. One adult female (Fig. 5): "Puerto de la Quesera, Guadalajara, Spain, $41^{\circ} 11^{\prime} 30.11^{\prime \prime N}, 3^{\circ} 24^{\prime} 27.55^{\prime \prime} \mathrm{W}, 1625 \mathrm{~m}, 8$-XII-2018, A. Sánchez-Vialas leg." [white label, printed]; MNCN_Ent 325409 [white label, printed]. One adult female: "Puerto de la Quesera, Guadalajara, Spain, $41^{\circ} 12^{\prime} 58.10 " \mathrm{~N}$, $3^{\circ} 25^{\prime} 14.37$ "W, $1712 \mathrm{~m}, 28-X I I-2021$, A. Sánchez-Vialas leg." [white label, printed]; MNCN_Ent 325410 [white label, printed]. -All paratypes labelled: "Paratypus, Meloe (Bolognaia) orobates Sánchez-Vialas, Ruiz, Recuero, Gutiérrez-Pérez \& García-París des. 2022" [white labels, printed]. All paratypes are preserved in ethanol (except for the female gonostyli of the specimen MNCN_Ent 325409 [Fig. 4E], which was dissected, mounted on a piece of cardboard using DMHF, and preserved dry, bearing the following labels: "Puerto de la Quesera, Guadalajara, Spain, $41^{\circ} 11^{\prime} 30.11^{\prime N} \mathrm{~N}, 33^{\circ} 24^{\prime} 27.55^{\prime W} \mathrm{~W}$, 1625 m, 8-XII-2018, A. Sánchez-Vialas leg." [white label, printed]; "Paratypus, Meloe (Bolognaia) orobates Sánchez-Vialas, Ruiz, Recuero, Gutiérrez-Pérez \& García-París des. 2022" [white label, printed]; Paratipo [red label, printed]), held at MNCN-CSIC.

Description of the holotype. Total length (frons to apex of the tergite VIII): 11.05 mm . Length from the frons to the posterior margin of elytra: 6.55 mm . Maximum width (located slightly anterior to the apex of the elytra): 6.81 mm . Body relatively robust, with slender appendages (Fig. 3). Voluminous abdomen. Coloration black all over body and appendages, except tibial spines and tarsal claws, which are brownish. Integument finely microreticulated, silky or semi-glossy in appearance. Setation decumbent, reddish brown, fairly dark, sometimes almost black ventrally and on


Figure 3. Holotype of Eurymeloe orobates sp. nov. MNCN 324740 A dorsal view of the head and pronotum B frontal view of the head $\mathbf{C}$ and $\mathbf{D}$ dorsal and dorsolateral views. Scale bars: 2 mm . Photographs: ASV.
the legs, and very short on the head, pronotum and elytra, longer on the abdominal tergites, legs, pygidium and ventral region, and arranged in relatively conspicuous tufts or single reddish yellow setae on the sclerotised plates of the abdominal tergites.

Head voluminous, broadly rounded and clearly wider than the pronotum, weakly truncated on the posterior margin of the temples, with integument black, silky


Figure 4. Genitalia of the male holotype MNCN 324740 (A-D) and of the female paratype MNCN 325409 (E) A-C ventral, dorsal and lateral views of the gonoforceps D lateral view of the aedeagus; scale bar: $1 \mathrm{~mm} \mathbf{E}$ female gonostyli; scale bar: 0.2 mm . Photographs: ASV.
in appearance, finely microreticulated, and without longitudinal depressions behind the eyes (Fig. 3A, B). Maximum width in frontal view (at the level of the temples): 2.83 mm ; minimum distance between the inner edges of the eyes: 1.83 mm ; distance between the clypeus-frontal suture and the vertex (in frontal view): 1.81 mm . Temples wide and regularly rounded (Fig. 3B). Frons almost flat, with a weak and short longitudinal groove from the clypeus-frontal suture to the vertex, that is slightly deeper from the level of the eyes to the vertex; surface adjacent to the antennal insertions slightly elevated and with a weak and diffuse depression attached to the raised areas (Fig. 3B). Head punctation dense, consisting of rounded, markedly deep, closely positioned and subconfluent punctures, slightly larger in diameter in the frontal region and smaller
towards the vertex, almost uniformly distributed, except on a narrow longitudinal mid-band on the frons, which is almost smooth (Fig. 3A, B). Head setation inconspicuous, short, decumbent, dark reddish brown, distributed according to the pattern of punctures in which it is inserted. Eyes medium-sized, subreniform and protruding, with upper lobes larger than the lower ones, barely notched at the level of the antennal insertions; clypeus-frontal suture deeply marked, weakly arcuate (Fig. 3B). Clypeus flat, transverse, subtrapezoidal, $2.1 \times$ wider than long, with a brownish membranous anterior border; punctures medium-sized, separated by between 0.5 and $1 \times$ their diameter, with the highest density on the sides; long setae homogeneously distributed, following the puncture pattern, directed forward, longest on the sides (Fig. 3B). La-brum-clypeus suture almost straight. Labrum transverse, $2.5 \times$ wider than long, deeply emarginated in the middle, forming two clear lobes; punctures similar to those on the clypeus; setae longer on the lobes, following the punctation pattern, oriented forward and curved (Fig. 3B). Mandibles relatively robust, curved along the outer margins and notched in the distal region, glabrous at the apex, and scarcely pilose at the base. Maxillary and labial palps unmodified. Maxillary palps with palpomere I very short, subcylindrical ( 0.09 mm long, 0.1 mm wide); II longer, troncoconical, weakly curved in the proximal half ( 0.44 mm long, 0.21 mm wide); III troncoconical, shorter and wider than II ( 0.38 mm long, 0.23 mm wide); IV sub-trapezoidal, widened distally, broadly rounded at the apex and dorsoventrally flattened, with a narrow excavation along the distal margin ( 0.54 mm long, 0.3 mm wide); setae scattered and moderately long on palpomeres II and III, shorter and more scarce on palpomere IV. Labial palps short, not visible dorsally, with palpomere I subcylindrical, very short ( 0.11 mm long, 0.09 mm wide); II troncoconical ( 0.22 mm long, 0.15 mm wide); III similar in shape to the last maxillary palpomere (IV); setae as on maxillary palps.

Antennae with 11 antennomeres, moniliform, slender and long, surpassing the base of the pronotum when extended backward (Fig. 3A). Antennomeres not modified, subcylindrical or subconical, I-VIII black, semi-glossy, IX-XI dark brown, opaque but becoming reddish brown in XI. Antennomere I widened apically, subconical, $\sim 1.92 \times$ longer than wide ( 0.48 mm long, 0.25 mm wide); II short, subglobose, slightly wider than long ( 0.81 mm long, 0.82 mm wide); III-X subcylindrical, similar to each other, between 1.84 and $2.22 \times$ longer than wide (III: 0.49 mm long, 0.22 mm wide; IV: 0.5 mm long, 0.24 mm wide; V: 0.48 mm long, 0.26 mm wide; VI: 0.46 mm long, 0.25 mm wide; VII: 0.49 mm long, 0.24 mm wide; VIII: 0.48 mm long, 0.23 mm wide; IX: 0.47 mm long, 0.21 mm wide; $\mathrm{X}: 0.48 \mathrm{~mm}$ long, 0.23 mm wide); XI is the longest, $\sim$ $3.71 \times$ longer than wide ( 0.78 mm long, 0.21 mm wide), subfusiform, with a blunt tip. Pilosity of antennomeres I-V comprised of short black setae, most decumbent though a few semi-erected, longer on segments I-III; antennomere VI with a mixture of short reddish brown and black setae; and antennomeres VII-XI with very short yellowish red setae, almost imperceptible.

Pronotum black, silky in appearance (Fig. 3A), small, sub-hexagonal, transverse, $1.59 \times$ wider than long; length in the middle: 1.37 mm ; maximum width (at the level of the lateral angles): 2.18 mm ; lateral margins weakly converge backwards in the
posterior two thirds and strongly converge forward in the anterior third, with the lateral angles well marked and rounded; fore margin almost straight; posterior margin or base broadly emarginated, with a thin flange. Dorsal surface of the pronotum clearly convex, gently sloping forward from the mid-region and steeply sloping back, with a slight and narrow depressed longitudinal-middle area with ambiguous boundaries (without a marked longitudinal midline or groove), such that two raised areas are observed on both sides of the central depression with two shallow and small rounded depressions observed anterior to the raised areas. Pronotal punctation relatively dense and unevenly distributed, consisting of relatively large, circular and deep punctures, subcontiguous, similar to those of the vertex but with a slightly larger diameter (Fig. 3A); the highest density is in the elevated areas on both sides of the midline, and the lowest densities are in the first quarter (just behind the fore margin), the depressed midband, and the central area of the base; integument surface with several fine, small and semi-wavy wrinkles between the punctures, located mainly on the sides, where they are arranged transversely and longitudinally in the middle depression. Pronotal setation inconspicuous, made up of short, curved dark reddish brown setae, mostly applied against the pronotal surface, distributed according to the pattern of punctures in which they are inserted; anterior margin, adjacent to the neck, with somewhat longer, semierect setae. Mesonotum mostly covered by the pronotum, showing, in dorsal view, only its posterior margin, which is weakly arcuate and with dense setation, consisting of setae longer than those of the pronotum, almost straight and lying backwards. Metanotum smooth, almost completely covered by the elytra. Prosternum narrow, very slightly extended posteriorly, broadly rounded at the central tip. Mesosternum relatively narrow and very transverse (width: 1.82 mm ; length in the middle: 0.69 mm ), with a small triangular prolongation backwards, ending in a rounded tip that extends to the level of the fore third of the mesocoxae; surface with long transverse wrinkles and dispersed punctures, similar to those of the vertex, and short setae. Metasternum subtrapezoidal, wide, covered by the mesocoxae, deep and closely notched in the middle of the posterior margin.

Elytra relatively short (length: 4.05 mm ), strongly dehiscent and weakly convex, imbricated basally (the right over the left), divergent backwards and reaching the middle area of the fourth tergite, covering the first tergite, almost completely covering the second, and covering the lateral areas of the third (of which, only the central plate is clearly visible), and lateral basal portions on both sides of the fourth; elytral surface strongly rugose, corrugated, with marked wavy foveoles (Fig. 3C); punctation small, fine, shallow and scattered, confused with the roughness of the foveoles; integument with very dispersed and isolated setae, similar to those of the pronotum, although somewhat shorter.

Abdomen black, voluminous (Fig. 3C); maximum width, at level of the fourth tergite: 6.78 mm . Tergites semi-matte in appearance, with very weak and indistinct foveoles scattered on its surface; central sclerotised plates of the tergites elliptical, with a semi-glossy aspect and an integument surface that is slightly rough, with fine wrinkles, arranged transversely and concentrically. Dorsal setation decumbent, consisting of isolated and scattered short, reddish brown setae on the semi-matte sides of the tergites,
and longer yellowish red (some almost golden) setae on the central plates, denser and forming inconspicuous tufts on the posterior margin of tergites II-IV. Ventrites silky in appearance, with dense punctation, made up of small, subcontiguous and slightly marked punctures that give them a finely vermiculated appearance; with short and decumbent dark brown, almost black, setae, homogeneously distributed; last ventrite clearly emarginated at the apex, with longer yellowish setae.

Legs relatively slender (Fig. 3C, D); surface with punctation fine and shallow, very dense in the tibiae and scarcer in the femurs, covered by relatively dense setation, consisting of short, dark brown (sometimes almost black) lying setae, denser on the tibiae. Length (in mm) of pro-, meso-, and metafemur as follows: 2.32, 2.6, and 3.1. Length (in mm ) of pro-, meso-, and metatibia as follows: 2.25, 2.24 and 2.55. Length (in mm ) of pro-, meso-, and metatarsus (and respective tarsomeres) as follow (claws excluded): 2.41 (I: 0.65 ; II: 0.4; III: 0.36; IV: 0.33; V: 0.67), 3.13 (I: 0.98; II: 0.53; III: 0.46 ; IV: 0.43 ; V: 0.73 ) and 3.56 (I: 1.36; II: 0.73 ; III: 0.61 ; IV: 0.86 ). Tarsi slender, clearly longer than the respective tibiae, with tarsomeres subcylindrical, slightly enlarged distally. Tarsomeres showing, on their ventral side, a small brush of very short, hirsute black setae, quite reduced in the last ones (V, V, IV). Pro- and mesotibiae with two similar distal spurs, short, narrow and straight; metatibial spurs dissimilar: outer spur spoon-shaped, inner spur similar to those of the fore- and mesotibiae but a little wider at the base and weakly curved at the apex. Coxae dense and finely punctate, with dense and short setation. Claws smooth, curved, with the lower lobe narrower than the upper one but equal in length.

Male genitalia with gonoforceps dark brown, hairless, moderately elongated, slender in dorsal, ventral, and lateral views (Fig. 4A-C). Gonostyli relatively long, $4.4 \times$ longer than wide in lateral view ( 1.33 mm long, 0.3 mm wide in lateral view), no excavated or depressed areas laterally in the distal regions; distal portion of each gonostylus separated dorsally by a fusiform longitudinal notch that extends to approximately the middle of the structure (Fig. 4A, B); distal lobes narrow and rounded at the apexes in lateral view (Fig. 4C). Gonocoxal plate relatively narrow and long, $1.38 \times$ longer than wide in dorsal view ( 1.36 mm long, 0.98 mm wide in ventral view), with the greatest width roughly in the middle of the plate, markedly emarginated at its distal margin (in ventral view) (Fig. 4A); surface almost flat. Aedeagus slender and narrow in lateral view ( 1.96 mm long, 0.2 mm wide in lateral view) flattened, narrowly rounded at the apex with two dorsal hooks that are similar in shape, although the distal hook is somewhat larger than the proximal one (Fig. 4D); endophallic hook small, located close to the apex and barely visible.

Variability. Female similar to the male (Fig. 5) but with the last abdominal ventrite rounded and not emarginated in its posterior margin, and with relatively shorter antennae. Morphological measurements of the studied female specimens (paratypes): total length (frons to apex of tergite VIII): $10-14 \mathrm{~mm}$ (mean = $12 \mathrm{~mm} ; n=4$ ); body length (frons to posterior border of elytra): $6.5-9.5 \mathrm{~mm}$ (mean $=8.3 ; \mathrm{n}=4$ ); body maximum width (between the elytral external borders): $6-8.1 \mathrm{~mm}$ (mean $=7.4$; $n=4$ ); pronotum length: $1.6-1.8 \mathrm{~mm}$ (mean $=1.7 ; n=4$ ); pronotum maximum


Figure 5. A-C habitus of a living female paratype (MNCN 325409) of Eurymeloe orobates sp. nov. D detail of the dorsal surface of the abdominal tergite I and the elytra. Note the brownish tufts. Photographs: ASV.
width: $2.23-2.74 \mathrm{~mm}$ (mean $=2.55 \mathrm{~mm}$; $n=4$ ); head maximum width: 2.74-3.43 mm (mean $=3.17 \mathrm{~mm} ; n=4$ ); elytra length: $4-5.5 \mathrm{~mm}$ (mean $=5 \mathrm{~mm} ; n=4$ ). Marked variability in the density of the pilose tufts on the dorsal side of the abdomen was observed: the studied females present lighter yellowish brown pilosity than that of the male, with more numerous and denser tufts located on the small, rounded depressed areas of the integument, homogeneously distributed, giving it an irregular appearance. Female gonostyli as in Fig. 4E.

Diagnosis and morphological comparisons. Eurymeloe (B.) orobates is characterised morphologically, with respect to all the other species of the subgenus Bolognaia, by the following combination of diagnostic traits: (1) body size small or medium (total length: $10-14 \mathrm{~mm}$ ); (2) body integument entirely black, semi-glossy in appearance; (3) setation of the head, pronotum and elytra, short and decumbent, reddish brown, moderately dark, sometimes very dark (almost black) ventrally and on the legs; (4) setation of the central plates of the abdominal tergites yellowish red (some almost golden), longer and forming inconspicuous tufts; (5) antennae slender and long, surpassing
the base of the pronotum when extended backwards; (6) head broadly rounded, with a weak and relatively short longitudinal median groove; (7) pronotum small, very transverse (more than $1.5 \times$ wider than long), sub-hexagonal; (8) pronotal surface showing a weak and narrow depressed longitudinal-middle area, but without a marked groove; (9) punctation of the head and pronotum dense, forming rounded and markedly deep punctures; (10) elytral surface strongly rugose, corrugated, with marked foveoles; and (11) male genitalia with long gonostyli with no excavated or depressed areas in the distal regions and a narrow and long gonocoxal plate.

The species most similar to E. orobates are E. rugosus and E. apenninicus (both belonging to the group B or $E$. murinus group). Both species present dark (black or dark brown) pilosity all over the body and, on the abdominal tergites, some inconspicuous (usually barely perceptible) yellowish brown or yellow setae, but not tufts. In addition to the colour pattern of the body setation, $E$. rugosus and $E$. apenninicus differ from $E$. orobates by the shape of their pronotum, which is longer, less transverse, and flatter (less convex), and has a strong median longitudinal groove (absent in E. orobates). The punctation of the head and pronotum are also markedly larger, deeper, and denser in $E$. rugosus and $E$. apenninicus (see Bologna 1988, 1991). In E. orobates, the antennae are slenderer and longer.

Within group B ( $E$. murinus group), in which $E$. (B.) orobates is integrated, the new species can be readily distinguished from E. (B.) baudueri (southern France, Iberian Peninsula, and northern Morocco), E. (B.) flavicomus (Canary Islands), E. (B.) ganglbaueri (mainland Italy, Sardinia, Corsica, Sicily, Greece, Albania, Bulgaria, Bosnia and Herzegovina, Montenegro, Turkey, Syria, Spain, and southern France), E. (B.) gomari (northern Morocco), E. (B.) kandaharicus (Iran and Afghanistan), E. (B.) murinus (Iberian Peninsula, Sicily, Sardinia, Corse, Crete, Maghreb, and Libya), E. (B.) nanus (Iberian Peninsula, North Africa, and Middle East), E. (B.) omanicus (eastern Arabian peninsula), and $E$. (B.) pallidicolor (western Morocco). For instance, in contrast to $E$. (B.) orobates, all these species present, among other specific traits, a body integument that is dull grey or dark brown, occasionally reddish brown or, rarely, sandy brown (E. pallidicolor) or almost black (E. ganglbaueri). In addition, the body integument is generally opaque or matte in appearance or, at most, silky (but never glossy or semi-glossy as in $E$. orobates). The setation of these species is also quite distinct from that of E. orobates: it is yellowish, whitish, or golden all over the body and usually longer, and on the abdominal tergites, the tufts of setae, when present, are highly conspicuous (see Fig. 2). Moreover, the punctation of the head and pronotum in these species is clearly finer and shallower, and the elytral sculpture is distinctly smoother (not corrugated) and without marked foveoles, except in E. ganglbaueri; however, in this last species, the foveoles are clearly more attenuated than in E. orobates (see Kaszab 1958, 1983; Bologna 1988, 1991; Ruiz and García-París 2009, 2015).

Eurymeloe (B.) orobates differs from the species of group A ( $E$. mediterraneus group, composed of, at least, E. affinis from the Maghreb and Libya; E. apivorus and E. baamarani, which are restricted to Morocco; E. baudii from the Italian Peninsula, Sicily, and Croatia; E. glazunovi from Eastern Europe and Central Asia; and E. mediterraneus, which is widely distributed throughout Europe, the Mediterranean basin, the Canary

Islands, and the Middle East) by presenting reddish brown body setation and abdominal tergites with small tufts of reddish yellow setae, among other diagnostic characters (see above). By contrast, the body pilosity, including on the abdominal tergites, of species of the E. mediterraneus group is black (see Bologna 1988, 1991; Ruiz and GarcíaParís 2015). The Sardinian specimens of $E$. mediterraneus with brown setae can clearly be distinguished from $E$. orobates by the shape of the pronotum: in the first species, it is subrectangular and has subparallel sides; in the second, it is markedly transverse and subhexagonal and has sides that converge backwards.

The only species in group C is $E$. (B.) fernandezi (endemic to the Canary Islands). In comparison with $E$. (B.) orobates, this species presents, among other distinctive characters, an entirely black body setation; a clearly longer, not transverse pronotum with sinuous margins; an integument surface with wrinkles and parallel ridges that form eddies; and an elytral sculpture consisting of a fine zig-zag roughness (see Pardo Alcaide 1951; Ruiz and García-París 2015). Lastly, the two species tentatively assigned to Bolognaia, E. (B.) saharensis (widely distributed throughout North Africa, the Canary Islands, the Iberian Peninsula, Israel, and Saudi Arabia) and $E$. (B.) vignai (only known from Djibouti), are phenetically very different to $E$. (B.) orobates: their body setation is entirely reddish and longer, without reddish yellow setae forming tufts on the abdominal tergites; a subsquare, not transverse pronotum; relatively fine, shallow, and scattered punctation of the head and pronotum; very long legs; elytra with a soft sculpture, without foveoles or marked roughness; and highly distinctive male genitalia (Bologna 1988; Ruiz et al. 2010).

Distribution and notes on natural history. Eurymeloe orobates is only known from a single locality, Puerto de la Quesera (in the province of Guadalajara, Spain) in the Iberian Peninsula (Fig. 6). This site, which is at an elevation of 1738 m above sea level (a.s.l.), is within the supra-Mediterranean bioclimatic level (see Rivas-Martínez 1987; Rivas-Martínez et al. 2002). Specifically, Puerto de la Quesera is in the Sierra de Ayllón, at the eastern edge of the Sistema Central mountain range. This region is characterised predominantly by micaceous schist, slate and quartzite soils (Rivas-Martínez et al. 1990; Vera 2004). Vegetation cover around Puerto de la Quesera consists of, at lower altitudes (below 1500 m a.s.l), deciduous oak forests of Quercus pyrenaica Willd. and, at higher altitudes (1500-1700 m a.s.l.), formations of Fagus sylvatica L. Above the deciduous tree cover level, there are shrubs such as Erica arborea L., Juniperus communis L., and Arctostaphyllos uva-ursi L., whereas grasslands dominate at altitudes over 1800 m a.s.l (Ibáñez et al. 1982). Hostile climatic conditions including low temperatures, late spring frosts, and strong winds characterise the high-altitude areas (Ibánez et al. 1982). Furthermore, this region has been strongly altered by human activities (e.g., deforestation and overgrazing), particularly by the establishment of terraced pinewood plantations of Pinus sylvestris L. (Fig. 6A) (Gil-García et al. 1995). In this region, adult specimens of $E$. orobates have been found actively wandering, under stones, and on tree barks, between November and May, usually in open areas or at the boundaries of the terraced plantations of P. sylvestris (authors, pers. obs.). Biological aspects of the new species remain unknown; however, we expect them to be similar to the ones described for other species of the E. murinus group (Bologna 1988, 1991).


Figure 6. A Puerto de la Quesera, Guadalajara, Spain. Type locality of Eurymeloe orobates sp. nov. B adult female of Eurymeloe orobates sp. nov. in situ (paratype MNCN 325407). Photographs A (ASV) B (MGP).

Etymology. The specific epithet orobates is derived from the Greek word "oros", meaning mountain, and "bates", meaning walker. This name alludes to the mountainous environment where the specimens of the new species were found, sometimes, wandering on mountain pastures and trails (Fig. 6B).

## Discussion

In light of previous morphological data and recent phylogenetic analyses (SánchezVialas et al. 2021; this study), we have updated the internal taxonomy of the genus Eurymeloe. In order to reflect the morphologically distinguishable, main monophyletic units within Eurymeloe, and to maintain the validity of the widely used name Coelomeloe, we have deemed it necessary to consider that each of the three main molecular lineages represents an independent subgenus: Eurymeloe, Coelomeloe, and Bolognaia subgen. nov.

Morphological traits of larvae have been traditionally considered relevant in the systematics of the group, sometimes even more informative than adult characters for phylogenetic studies (Bologna and Pinto 2001). In fact, traits of the first instar larva (triungulin) have been studied for most of the genera and subgenera of Meloini (Bologna 1988, 1991; Selander 1989; Bologna et al. 1989, 1990; Bologna and Pinto 1992, 1998; Pinto and Bologna 1993; Bologna and Aloisi 1994; Di Giulio et al. 2002; Di Giulio et al. 2013, 2014). However, not having a resolved internal taxonomy for Eurymeloe confuses explanations of the evolutionary history of some of these traits. For instance, a particular morphological trait related to the shape of the abdominal spiracle I that is shared between the first instar larvae of Coelomeloe and Eurymeloe sensu stricto [the brevicollis group of Meloe (Eurymeloe) sensu Bologna 1988] was previously suggested to be the result of parallel biological adaptation (Di Giulio et al. 2013). However, considering our results and consistent with those shown by Sánchez-Vialas et al. (2021), this trait can be better explained as a synapomorphic character state for these sister subgenera.

Some conspicuous adult traits are also shared between the subgenera Eurymeloe and Coelomeloe, including antennae that are submoniliform, robust, short or medium in length, and which do not usually not reach the posterior margin of the pronotum; antennomeres V to VII that are wider than long or, at most, as wide as long; and very short or not [e.g., E. (E.) brevicollis, E. (E.) ibericus, and E. (C.) tuccia] body pubescence. These character states differ from those of the subgenus Bolognaia, which usually presents antennae that are moniliform, normally slender, long or medium in length, and which usually reach or exceed the posterior margin of the pronotum; antennomeres IV-IX that are subcylindrical, always longer than wide; and distinctive short or very short (black, yellowish, whitish, or golden) body pubescence. Therefore, the close relationship between Eurymeloe s. str. and Coelomeloe is supported by both our molecular analysis $(\mathrm{BPP}=0.9)$ and morphology.

Our results confirm that $E$. (B.) rugosus, a species previously assigned to Bologna's (1988) subgroup A ( $E$. rugosus subgroup) on the basis of morphology, should instead be included in subgroup B (M. murinus subgroup). The morphology of E. rugosus, which presents a completely black coloration without noticeable brownish pilosity, led Bologna (1988) to separate it from subgroup B. Although both subgroups are now integrated within the subgenus Bolognaia, they are not monophyletic groups since, according to our molecular phylogeny, the morphological traits used to diagnose them are homoplastic. As a result, the assignation of some species to each of these groups has
been controverted. For example, ambiguous morphological traits in E. ganglbaueri (see Ruiz \& García-París 2009) has blurred its systematic allocation, as it was included in subgroup A based on morphology sensu Bologna (1988) but ascribed to that author's subgroup B based on molecular data (Sánchez-Vialas et al. 2021). The newly discovered species, Eurymeloe ( $B$.) orobates sp. nov., notably presents a pattern of pilosity that is intermediate between $E$. (B.) rugosus and $E$. (B.) murinus- $E$. (B.) ganglbaueri, but a body integument that is more similar to $E$. (B.) rugosus.

With the addition of the new species, eight species of the subgenus Bolognaia are known from the Iberian Peninsula: E. (B.) baudueri, E. (B.) ganglbaueri, E. (B.) mediterraneus, $E$. (B.) murinus, $E$. (B.) nanus, $E$. (B.) orobates, $E$. (B.) rugosus, and $E$. (B.) saharensis (García-París et al. 2010; Bologna, 2020a; this study). The existence of a new, morphologically distinctive species of Meloidae, which was found in an apparently well surveyed area of central Spain (Puerto de la Quesera, in the province boundaries between Madrid and Guadalajara), suggests that an undefined portion of the diversity of Bolognaia and other secretive species of Eurymeloe still awaits discovery.

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# Comparative mitogenomics of the genus Motacilla (Aves, Passeriformes) and its phylogenetic implications 

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#### Abstract

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#### Abstract

The genus Motacilla belongs to Motacillidae (Passeriformes), where mitochondrial features are poorly understood and phylogeny is controversial. Whole mitochondrial genome (mitogenome) data and large taxon sampling are considered to be ideal strategies to obtain this information. We generated four complete mitogenomes of M. flava, M. cinerea, M. alba and Dendronanthus indicus, and made comparative analyses of Motacilla species combined with mitogenome data from GenBank, and then reconstructed phylogenetic trees based on 37 mitochondrial genes. The mitogenomes of four mitogenome sequences exhibited the same gene order observed in most Passeriformes species. Comparative analyses were performed among all six sampled Motacilla mitogenomes. The complete mitogenomes showed A-skew and C-skew. Most protein-coding genes (PCGs) start with an ATG codon and terminate with a TAA codon. The secondary structures of RNAs were similar among Motacilla and Dendronanthus. All tRNAs except for $\operatorname{trnS}$ (agy) could be folded into classic clover-leaf structures. Three domains and several conserved boxes were detected. Phylogenetic analysis of 90 mitogenomes of Passeriformes using maximum likelihood (ML) and Bayesian inference (BI) revealed that Motacilla was a monophyletic group. Among Motacilla species, M. flava and M. tschutschensis showed closer relationships, and $M$. cinerea was located in a basal position within Motacilla. These data provide important information for better understanding the mitogenomic characteristics and phylogeny of Motacilla.


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## Keywords

Comparative analysis, mitogenome, phylogeny

## Introduction

In most animals, the mitochondrial genome (mitogenome) contains 13 protein-coding genes (PCGs), two rRNA genes (rRNAs), 22 tRNA genes (tRNAs), and one noncoding region (the control region, CR) (Wolstenholme 1992; Boore 1999). Mitochondrial sequences are commonly used for inferring phylogeny (Hassanin et al. 2005), and the mitogenome has been used as an effective marker for exploring the phylogenies of some avian taxa (Li et al. 2016a; Mackiewicz et al. 2019; Cai et al. 2019).

Passeriformes comprises 6533 currently described species (Gill et al. 2020). The genus Motacilla belongs to Motacillidae (Passeriformes) and contains 12 species (Alström et al. 2003; del Hoyo et al. 2004), which show striking plumage pattern variation (Harris et al. 2018). Motacilla flava Linnaeus, 1758 is a small, insectivorous oscine (Ödeen and Björklund 2003) and is closely related to M. alba Linnaeus, 1758, distributed in the Palearctic (Dong and Zhang 2011). Some mitochondrial fragments, such as nad2 and CR of M. alba (Li et al. 2016b), have been used to study the phylogeography and population history of Motacilla. Additionally, some mitochondrial genes, such as nad2 (Suppl. material 1: Fig. S1A; Dong et al. 2016) and cytb (Suppl. material 1: Fig. S1B; Zhang et al. 2016), have been used to study the phylogenetic relationships of Motacilla. However, the phylogenetic position of some Motacilla species is still controversial. For example, M. alba has been reported to form a sister group with M. madaraspatensis Gmelin, 1789 (Suppl. material 1: Fig. S1A, Dong et al. 2016), but it has also been grouped with M. cinerea Tunstall, 1771 (Suppl. material 1: Fig. S1B; Zhang et al. 2016). In addition, phylogenetic results reconstructed from genome-wide SNPs (Suppl. material 1: Fig. S1C, Harris et al. 2018) have some incongruence with those based on mitochondrial genes or mitogenomes (Suppl. material 1: Fig. S1A, B, D; Dong et al. 2016; Zhang et al. 2016; Gao et al. 2019).

An increasing number of avian mitogenome sequences are being generated with high-throughput sequencing technology (Morinha et al. 2016; Yang et al. 2018), facilitating the identification of mitogenomic characteristics such as gene order and base composition through the comparison of mitogenomes. However, the limited Motacilla mitogenomic sequences available from the GenBank database restricts the exploration of mitogenome features in this genus. For example, recent studies of Motacilla (Dong et al. 2016; Zhang et al. 2016; Harris et al. 2018; Gao et al. 2019) have focused on the phylogenetic relationships within this genus but have not conducted further comparative analyses among mitogenomes. In the present study, we obtained complete mitogenome sequences of M. flava, M. cinerea, M. alba, and Dendronanthus indicus Gmelin, 1789, performed comparative analyses and generated phylogenies (Subspecies differentiation was not discussed here). The new mitogenome data not only may help us understand the mitogenomic characteristics of Motacilla but also provide a basis for exploring phylogenetic relationships.

## Methods

## Specimen collection

Muscle samples were collected from the following species: M. flava (from China, Shaanxi Province, Hongjiannao in 2013); M. cinerea (from China, Shaanxi Province, Feng County in 2017); and M. alba and D. indicus (from China, Shaanxi Province, Lantian in 2018). All specimens of muscle samples were preserved in 100\% ethanol and stored at $-20^{\circ} \mathrm{C}$ at the Shaanxi Institute of Zoology, Shaanxi Province, China.

## Mitogenome sequencing, assembly and annotation

The mitogenome of M. flava was sequenced by Genesky Biotechnologies Inc., Shanghai, China, using the Illumina HiSeq 2000 platform, while those of M. cinerea, M. alba and $D$. indicus were sequenced at Biomarker Technologies Inc., Beijing, China, using the Illumina Xten platform and a 150 bp paired-end strategy. Genomic DNA was extracted using a DNeasy kit and fragmented using ultrasonic methods to prepare a small-inserted-fragment library. The library data were obtained via Bridge PCR and Illumina paired-end sequencing.

There were $15,149,744$ paired-end raw reads of $M$. Alava, of which 47,390 reads were used for mitogenome assembly, with average coverage of 417.1X. There were $20,702,440$ paired-end raw reads of $M$. cinerea, with clean data 6.92 G . A total of 261,229 reads were used for mitogenome assembly, with average coverage of 2256.2X. There were $7,868,047$ raw reads in $M$. alba, with $7,860,296$ reads with clean data, and $8,430,436$ raw reads of $D$. indicus, with $8,420,710$ reads with clean data.

The raw data from $M$. flava, $M$. cinerea and $M$. alba were quality trimmed with CLC Genomics Workbench 9.5.2 (CLC bio, Aarhus, Denmark) using the default parameters. Mitogenome assembly was performed in MITOBIM 1.8 (Hahn et al. 2013), with M. alba (GenBank: NC029229) as a reference. The mitogenomic sequences of $D$. indicus were assembled using MitoZ 2.4 (Meng et al. 2019). Mitochondrial PCGs were identified using Geneious 11.1.3 (Kearse et al. 2012) by searching for open reading frames and employing the M. alba mitogenome (GenBank: NC029229) as a reference. Most tRNAs were identified using tRNAscan-SE 1.21 (Lowe and Eddy 1997), with secondary structures used as references. The remaining tRNAs, rRNAs and CRs were identified by comparison with other Motacilla species. Each mitochondrial gene was confirmed by alignment with the corresponding homologous genes from other Motacilla species available in GenBank. The secondary structures of $\operatorname{rrnS}$ and $\operatorname{rrnL}$ were generated using the mitogenomic rRNAs of Remiz consobrinus as a reference (Gao et al. 2013).

## Comparative analysis and phylogenetic reconstruction

The six mitogenomes ( $M$. flava, M. cinerea and $M$. alba mitogenomes from collected specimens combined with $M$. tschutschensis, M. alba and M. cinerea genomes from GenBank) were used for comparative analysis. A mitogenome of M. lugens
(KU246035/NC_029703) was excluded because this has been shown to represent a chimera (Sangster and Luksenburg 2021). The nucleotide compositions of the mitogenomes and different datasets were calculated using Geneious 11.1.3 (Kearse et al. 2012). Nucleotide bias was calculated using the formulas AT-skew $=(\mathrm{A}-\mathrm{T}) /(\mathrm{A}+\mathrm{T})$ and GC-skew $=(\mathrm{G}-\mathrm{C}) /(\mathrm{G}+\mathrm{C})$ (Perna and Kocher 1995). Relative synonymous codon usage (RSCU) was calculated with MEGA 11 (Tamura et al. 2021).

A total of 90 mitogenomes of Passeriformes were used to reconstruct phylogenetic relationships; the included mitogenomes came from 12 taxonomic families with Aethopyga gouldiae (Nectariniidae) used as an outgroup (Suppl. material 7: Table S1). Each mitochondrial gene was aligned individually using MUSCLE in MEGA 11 (Tamura et al. 2021), starting with the alignment of PCGs to amino acid sequences. One mitogenomic dataset (mtDNA) was used for phylogenetic analysis, which included the nucleotide sequences of 13 PCGs, two rRNAs and 22 tRNAs, with a length of $15,722 \mathrm{bp}$. The best models of GTR $+\mathrm{F}+\mathrm{R} 5$ for maximum likelihood (ML) analysis and GTR $+\mathrm{F}+\mathrm{I}+\mathrm{G} 4$ for Bayesian inference (BI) analysis were assessed in ModelFinder (Kalyaanamoorthy et al. 2017) using the Bayesian information criterion (BIC) in PhyloSuite 1.2.1 (Zhang et al. 2020). Phylogenetic relationships were analyzed using ML phylogenies with IQ-TREE 1.6.8 (Nguyen et al. 2015) with 1000 bootstrap replicates. The BI phylogeny was analysed with MrBayes 3.2.7 (Ronquist et al. 2012). Two independent runs with four simultaneous Markov chains were run for $5,000,000$ generations and were sampled every 100 generations. The first $25 \%$ of generations were discarded as burn-in. The effective sample size (ESS) values were estimated in Tracer 1.7 (Rambaut et al. 2018), with ESS values > 200 .

## Results and discussion

## Mitogenomic structure and organization

The obtained complete mitogenomes of M. flava, M. cinerea, M. alba and D. indicus ranged from $16,831 \mathrm{bp}$ to $16,870 \mathrm{bp}$ in length and each contained 37 genes and a noncoding region (CR) (Fig. 1). The complete mitogenome sequences were submitted to GenBank (MW929088-MW929091). Four gene arrangements have been identified among the Passeriformes mitogenomes sequenced to date (Caparroz et al. 2018; Mackiewicz et al. 2019). The gene order cytb-trnT-trnP-nad6-trnE-CR-trnF-rrnS is found in the mitogenomes of three Motacilla species and D. indicus, which is consistent with the order observed in most Passeriformes species (Mackiewicz et al. 2019). The major strand (J-strand) encodes 12 PCGs and two rRNAs as well as $\operatorname{trnF}$, $\operatorname{trnV}$, $\operatorname{trnL}($ uur $), \operatorname{trnI}, \operatorname{trn} \mathrm{M}, \operatorname{trnW}, \operatorname{trnD}, \operatorname{trnK}, \operatorname{trnG}$, trnR, the HSL cluster [trnH, trnS(agy), trnL(cun)] and $\operatorname{trnT}$ (Fig. 1). The lengths of the intergenic spacers range from $1-23 \mathrm{bp}$ in the three Motacilla mitogenomes and $1-18 \mathrm{bp}$ in $D$. indicus, with the longest intergenic spacer being located between the trnP and nad6 genes.


Figure I. Gene map of four newly sequenced mitogenomes. Notes: tRNAs are abbreviated with a single letter; mitochondrial genes encoded by the J - and N -strands, indicated in bold, are located outside and inside of the circle, respectively.

## Comparative analysis of Motacilla mitogenomes

The gene orders and nucleotide compositions of the six sampled Motacilla mitogenomes were generally similar. For instance, the $\mathrm{A}+\mathrm{T}$ content ranges from $53.5 \%$ to $53.9 \%$, which was slightly higher than the $\mathrm{G}+\mathrm{C}$ contents. All mitogenomes showed a tendency toward A-skew and obvious C-skew (Suppl. material 8: Table S2), which was similar to findings in other birds (Kan et al. 2010; Eberhard and Wright 2016; Li et al. 2016a).

## Protein-coding genes

The A+T contents of the 13 PCGs excluding stop codons ranged from $52.4 \%$ to $52.9 \%$ in sampled Motacilla mitogenomes (Suppl. material 8: Table S2). The highest


Figure 2. RSCU analysis of the PCGs of six mitogenomes from the genus Motacilla. Note a M. alba b M. cinerea c M. flava d M. tschutschensis e M. alba (MN356232) f M. cinerea (NC_027933).
$A+T$ content was found at the second codon position in all Motacilla mitogenomes. Obvious T-skew was recovered at the second codon position, while the most significant A-skew was found at the third codon position. The three codon positions showed different degrees of C-skew, which was most obvious at the third codon position.

The first and last codons of the PCGs of Motacilla were compared (Suppl. material 9: Table S3). Twelve of the 13 PCGs started with an ATG codon, while nad3 started with ATT. The start codons were conserved in the six mitogenomes. The termination codons of the PCGs included TAA, TAG, AGG, AGA, TA and T, which are conserved among PCGs with the exception of $\operatorname{cox} 2, \operatorname{cox} 3$ and nad 2 . The incomplete T termination codons found in cox 3 and nad 4 have also been reported in other avian mitogenomes (Ma et al. 2014; Li et al. 2015; Eberhard and Wright 2016).

RSCU analysis indicated that among all PCGs, codons including A or C at the third position were frequently overused relative to other synonymous codons (Fig. 2). The codon usage among Motacilla species was found to be conserved, with CUA (L), CGA (R) and UCC(S) representing the most frequently used codons.

## RNA genes

Similar to other avian mitogenomes, rrnS was found to be located between trnF and trnV, and rrnL was located between $\operatorname{trnV}$ and $\operatorname{trnL}(u u r)$. The length of rrnS was 975 bp
in M. alba (MN356232) and 973 bp in the other Motacilla mitogenomes, while the length of rrnL was 1595 bp in all Motacilla species. The A+T content was slightly greater than the $\mathrm{G}+\mathrm{C}$ content in the rRNA genes, ranging from $52.2 \%$ to $52.3 \%$ in rrnS and $55.2 \%$ to $55.4 \%$ in rrnL, and both rRNA genes exhibited A-skew and Cskew (Suppl. material 8: Table S2).

The rrnS included three domains and 47 helices in M. flava (Suppl. material 2: Fig. S2), while rrnL included six domains and 60 helices (Suppl. material 3: Fig. S3). Most of the identified sequences and secondary structures were conserved compared with those of other Motacilla rRNAs. In addition, most of the stems of the rRNA secondary structures were similar to those found in other Passeriformes mitogenomes. For example, stems 21 and 47 of rrnS and 15 and 40 of rrnL were consistent with those found in $R$. consobrinus (Gao et al. 2013).

A total of eight $\mathrm{tRNAs}(\operatorname{trn} \mathrm{Q}, \operatorname{trnA}, \operatorname{trnN}, \operatorname{trnC}, \operatorname{trn} \mathrm{P}, \operatorname{trn} S(\mathrm{ucn}), \operatorname{trnP}$ and $\operatorname{trnE})$ were located on the N -strand, while the remaining 14 tRNAs were located on the Jstrand (Fig. 1). The lengths of the 22 tRNAs in each Motacilla species ranged from 66 to 75 bp . The A+T content ranged from $58.3 \%$ to $58.6 \%$ in the tRNAs, which exhibited A-skew and G-skew (Suppl. material 8: Table S2).

Twenty-one of the 22 tRNAs of $M$. flava were folded into a clover-leaf-like secondary structure, with the exception of $\operatorname{trnS}$ (agy), lacking a dihydrouridine ( DHU ) stem (Suppl. material 4: Fig. S4), which is considered to be a typical feature of metazoan mitogenomes (Wolstenholme 1992). Comparisons among Motacilla tRNAs showed that the most conserved tRNAs were $\operatorname{trnL}(\mathrm{UUR}), \operatorname{trnM}, \operatorname{trnW}, \operatorname{trnA}, \operatorname{trnC}, \operatorname{trnH}$, $\operatorname{trnL}(\mathrm{CUN}), \operatorname{trnT}, \operatorname{trnP}$ and $\operatorname{trnE}$ (Suppl. material 4: Fig. S4), which contained the same nucleotides. Some mismatched base pairs found in Motacilla were similar to those observed in some other Passeriformes species (Pyrgilauda ruficollis, Ma et al. 2014; R. consobrinus, Gao et al. 2013), such as the C-C pair located in the acceptor stem of $\operatorname{trnL}$ (uur) and the anticodon stem of $\operatorname{trnG}, \mathrm{A}-\mathrm{A}$ in the $\mathrm{T} \varphi \mathrm{C}$ stem of $\operatorname{trnD}$, and $\mathrm{U}-\mathrm{U}$ in the anticodon stem of trnG.

## Control region

The CR was located between the trnE and trnF genes and were $1243-1250 \mathrm{bp}$ in length. The average $\mathrm{A}+\mathrm{T}$ content was $56.2 \%$ among all sampled Motacilla mitogenomes, which was slightly higher than that of G+C. The CRs showed a tendency toward T-skew and C-skew (Suppl. material 8: Table S2), with C-skew being more obvious. This C-skew was consistent with findings in other reported avian CRs (e.g., Huang et al. 2017).

The CR regulates the replication of the H strand and the transcription of all mitochondrial genes (Clayton 1992) and can be divided into three domains: extended termination-associated sequence (ETAS) domain I, central conserved domain II, and conserved sequence block (CSB) domain III (Sbisà et al. 1997; Randi and Lucchini 1998; Ruokonen and Kvist 2002). Among the three domains of the CR, domain I showed slight A-skew and obvious C-skew, domain II showed a tendency
toward T-skew and C-skew, and domain III exhibited A-skew and a highly significant C-skew (Suppl. material 8: Table S2).

The proportions of variable sites among the three domains were $3.6 \%, 2.4 \%$ and $9.0 \%$, respectively. Thus, most variation was found in domain III, similar to the findings of previous studies (Ruokonen and Kvist 2002; Huang et al. 2017). A poly-C sequence was found near the $5^{\prime}$ end of CR domain I in $M$. Alava, with a sequence of ССССССССССТТСССССССС, and this sequence was relatively conserved in the sampled mitogenome CRs (Suppl. material 5: Fig. S5). Within the M. flava CR sequence, boxes F, E, D, C, B and a bird similarity box in domain II were identified. The F, E, D and C boxes were similar to those found in other avian mitogenomes (Suppl. material 5: Fig. S5; Huang et al. 2017). Among these boxes, the F-box, bird similarity box and B-box were fully conserved among sampled mitogenomic sequences. Domain III contained CSB1, whose sequence was similar to that found in other birds (Huang et al. 2017). However, it was difficult to identify sequences corresponding to $\mathrm{O}_{\mathrm{H}}$, CSB2, CSB3 and bidirectional LSP/HSP promoters found in other birds (Li et al. 2015), which might play important roles in mitogenome replication. Furthermore, tandem repeat sequences in CRs are found in many avian mitogenomes (Yang et al. 2018). However, none of the sampled Motacilla CRs contained tandem repeats.

## Phylogenetic analysis

The ML and BI phylogenetic trees were reconstructed using the mtDNA dataset, showing consistent topological results among Motacillidae (Fig. 3 and Suppl. material 6: Fig. S6). The analyses supported the monophyly of Motacillidae with $100 \%$ bootstrap support and posterior probabilities of 1.0 . Among the three sampled genera among Motacillidae (Anthus, Dendronanthus and Motacilla), Anthus was sister to D. indicus and Motacilla. The monophyly of Motacilla was also recovered, with $D$. indicus forming a sister group with Motacilla.

Within Motacilla, the following phylogenetic relationships were recovered: $(((M$. flava + M. tschutschensis $)+$ M. alba $)+$ M. cinerea), similar to previous studies (Suppl. material 1: Fig. S1A; Dong et al. 2016; Suppl. material 1: Fig. S1D; Gao et al. 2019). Motacilla cinerea was in the basal position within Motacilla, and M. flava showed a closer relationship with $M$. tschutschensis. However, M. alba showed a closer relationship with M. cinerea (Suppl. material 1: Fig. S1B; Zhang et al. 2016), while M. cinerea presented a closer phylogenetic relationship with M. flava (Suppl. material 1: Fig. S1C; Harris et al. 2018). These differences might be due to the different data types, dataset sizes and sampling strategies involved. For example, the phylogenetic tree topologies obtained from the complete mitogenome are not identical to those resulting from individual mitochondrial genes in some avian taxa (Campillo et al. 2019). In addition, the phylogenetic relationships recovered from nuclear segment datasets are inconsistent with those recovered from mitogenomes in some aves (Li et al. 2016a; Campillo et al. 2019). Therefore, our results indicate that further studies are needed to address the phylogenetic relationships within Motacilla by adding more sampling and some nuclear data.


Figure 3. Phylogenetic results based on the maximum likelihood method using the mtDNA dataset.

## Conclusions

The complete mitogenomes of Motacilla flava, M. cinerea, M. alba and Dendronanthus indicus were sequenced and were shown to present the typical genome organization and gene order found in other Passeriformes mitogenomes. We focused on comparative analyses of the six mitogenomes to identify the mitogenomic characteristics of the genus Motacilla, such as the base composition, codon usage and RNA secondary structures. The complete mitogenomes showed a tendency toward Askew and C-skew. Most PCGs start with typical ATG codons and terminated with TAA codons. All tRNAs could be folded into classic clover-leaf structures except for trnS(agy), which lacked a DHU arm. In addition, 90 mitogenomes of Passeriformes were used to build the tree of phylogenetic relationships. The phylogenetic tree supported the monophyly of Motacillidae. Within Motacilla, the phylogenetic topology of $(((M$. flava $+M$. tschutschensis $)+M$. alba $)+M$. cinerea $)$ was recovered.

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## Supplementary material I

## Figure S1

Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Phylogenetic hypotheses of previous studies. a: Dong et al. (2016); b: Zhang et al. (2016); c: Harris et al. (2018); d: Gao et al. (2019).
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl1

## Supplementary material 2

## Figure S2

Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Secondary structures of $\operatorname{rrnS}$ of M. flava. Note: differences within the six mitogenomes from the genus Motacilla are indicated by filled grey circles.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl2

## Supplementary material 3

Figure S3
Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Secondary structures of rrnL of M. flava. Note: differences within the six mitogenomes from the genus Motacilla are indicated by filled grey circles.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl3

## Supplementary material 4

Figure S4
Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Secondary structures of tRNAs of M. flava. Note: differences within the six mitogenomes from the genus Motacilla are indicated by filled grey circles.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl4

## Supplementary material 5

Figure S5
Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Control region structures of M. flava. Note: differences within the six mitogenomes from the genus Motacilla are indicated with filled circles.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl5

## Supplementary material 6

Figure S6
Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Image.
Explanation note: Maximum likelihood and Bayesian inference phylogenetic results based on the mtDNA dataset, corresponding to Fig. 3. Note: a: ML tree, b: BI tree. Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1109.81125.suppl6

## Supplementary material 7

## Table S1

Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye Data type: Doc file.
Explanation note: Mitogenome sequences employed for reconstructing phylogenetic trees.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl7

## Supplementary material 8

## Table S2

Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Doc file.
Explanation note: Nucleotide composition and bias of six mitogenomes of the genus Motacilla. Notes: Stop codons of protein-coding genes were excluded; AT-skew=[A$\mathrm{T}] /[\mathrm{A}+\mathrm{T}]$, GC-skew=[G-C]/[G+C]. The sequenced mitogenome species in this study are shown in the bold format.
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Link: https://doi.org/10.3897/zookeys.1109.81125.suppl8

## Supplementary material 9

## Table S3

Authors: Chao Yang, Xiaojuan Du, Yuxin Liu, Hao Yuan, Qingxiong Wang, Xiang Hou, Huisheng Gong, Yan Wang, Yuan Huang, Xuejuan Li, Haiyan Ye
Data type: Doc file.
Explanation note: Initial and terminal codons of protein-coding genes in the six mitogenomes from the genus Motacilla. Notes: The sequenced mitogenome species in this study are shown in the bold format.
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# Two new species and a new genus of ray spiders (Araneae, Theridiosomatidae) from the Ryukyu Islands, southwest Japan, with notes on their natural history 

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#### Abstract

This paper provides descriptions of two new theridiosomatid species, Theridiosoma nigrivirgatum sp. nov. and Sennin tanikawai gen. nov., sp. nov. from the Ryukyu Islands, southwest Japan, with photographs and illustrations of both sexes. Sennin gen. nov. is a troglophilic genus composed of two species, S. tanikawai sp. nov. (Iriomote Island, Japan) and S. coddingtoni (Zhu, Zhang \& Chen, 2001), comb. nov. (southern China). Zoma dibaiyin Miller, Griswold \& Yin, 2009, which recently joined the Japanese fauna, was morphologically reexamined based on specimens from the Ryukyus, and taxonomic features of Zoma males were reassessed. A distributional map of theridiosomatid spiders in the Ryukyus is also provided, including T. dissimulatum Suzuki, Serita \& Hiramatsu, 2020, and T. alboannulatum Suzuki, Serita \& Hiramatsu, 2020 with their habitat types, web morphology, and web-building behavior in detail.


## Keywords

Araneoidea, embolic apophysis, limestone cave, Iriomote Island, new combination, Okinawa Island, orb web, taxonomy

## Introduction

The family Theridiosomatidae Simon, 1881 (Araneae: Araneoidea) is composed of smallsized (body length, ca. $0.5-3 \mathrm{~mm}$ ) spiders that prefer dark and humid environments such as forest floors, mountainous streams, and caves (Coddington 1986a). The family is characterized by a sternal pit organ (Wunderlich 1980) that has a pair of pits on the anterior margin of the sternum, except in Chthonos Coddington, 1986 (Coddington 1986a), large globular palp of males, a pair of spermathecae fused or in contact with each other in females except in Coddingtonia Miller, Griswold \& Yin, 2009 (Miller et al. 2009), and elongated trichobothria on the dorsum of the third and fourth legs (Coddington 1986a). All genera except Chthonos are known as web-builders (Coddington 1986a), and several genera build conventional orb webs (e.g., Baalzebub Coddington, 1986, Epeirotypus O. Pickard-Cambridge, 1894, and Naatlo Coddington, 1986), while others construct orb webs with radial anastomosis (e.g., Theridiosoma O. Pickard-Cambridge, 1879) or deformed webs, called sparse networks (e.g., Ogulnius O. Pickard-Cambridge, 1882, Wendilgarda Keyserling, 1886). Webs of some genera are characterized by a tension line, a non-sticky isolated radius that stretches from the center of the web and attaches to the substrates (e.g., Epeirotypus, Naatlo, and Theridiosoma). Spiders of these genera drag the tension line with forelegs and hold it into coiled conditions while holding the web with hindlegs. Therefore, the web acquires a distorted conical shape (Shinkai and Shinkai 1985; Coddington 1986a). When flying insects approach the web, the tension line is promptly released, and the launched web captures the prey. This latch-mediated spring actuation results in an ultrafast web shooting motion (Alexander and Bhamla 2020).

Currently, 19 genera and 133 species of Theridiosomatidae are recorded mainly in tropical and subtropical regions worldwide (World Spider Catalog 2022). After the revision by Coddington (1986a), new genera and new species were described mostly from China and Southeast Asia (Miller et al. 2009; Chen 2010; Wunderlich 2011; Zhao and Li 2012; Labarque and Griswold 2014; Zhao and Li 2014; Prete et al. 2018). Twelve genera, namely Baalzebub, Chthonopes Wunderlich, 2011, Coddingtonia, Epeirotypus, Karstia Chen, 2010, Menglunia Zhao \& Li, 2012, Ogulnius, Sinoalaria Zhao \& Li, 2014, Tagalogonia Labarque \& Griswold, 2014, Theridiosoma, Wendilgarda, and Zoma Saaristo, 1996, were recorded from East to Southeast Asia. These genera are distributed in both neotropics and Asia, with several exceptions such as Karstia and Menglunia, which are endemic to China, and Tagalogonia, endemic to the Philippines (World Spider Catalog 2022).

The Ryukyu Islands, comprising hundreds of continental islands located between Kyushu and Taiwan, were formed by a complicated geological history of several land bridge connections with the Chinese continent. Consequently, the fauna constitutes continental components and consists of many endemic species that have been derived from the continental ancestry (e.g., Ota 1998, 2000). Spider fauna of the Ryukyu Islands has been surveyed by many arachnologists (e.g., Shimojana 1977), but small-sized spiders such as Theridiosomatidae have only recently been examined (e.g., Shinkai and Hiramatsu 2000). Our recent surveys on the islands revealed two new species of Theridiosoma from the Ryukyus, and two theridiosomatid species as new mem-
bers of the spider fauna in the Ryukyus: Theridiosoma dissimulatum Suzuki, Serita \& Hiramatsu, 2020, T. alboannulatum Suzuki, Serita \& Hiramatsu, 2020, Wendilgarda ruficeps Suzuki, 2019 and Zoma dibaiyin Miller, Griswold \& Yin, 2009 (Ono and Ogata 2018; Suzuki et al. 2020; Suzuki and Matsushima 2021; Suzuki and Serita 2021).

During our survey in the Ryukyu Islands conducted between 2020 and 2022, several unidentified specimens of theridiosomatid spiders were further discovered from secondary forests, grasslands, and bushes in Okinawa, Kume and Aka Islands, and limestone caves on Iriomote Island. Based on morphological examination, we concluded that these specimens belong to two new species. One species was determined to be an undescribed Theridiosoma species. We confirmed that the second undescribed species from caves on Iriomote Island possesses unique characteristics that do not correspond to the taxonomic characteristics of known theridiosomatid genera. We were also aware that Karstia coddingtoni (Zhu, Zhang \& Chen, 2001), known from Southern China, shares common features with the undescribed species in Iriomote Island. Here, we suggest the establishment of a new genus named Sennin gen. nov., that comprises of these two species. Furthermore, several specimens of the Chinese species Zoma dibaiyin Miller, Griswold \& Yin, 2009, which was recently recognized in the Japanese fauna (Ono and Ogata 2018; Suzuki and Serita 2021), were morphologically reexamined, and taxonomic characteristics of Zoma males were reassessed. We provide descriptions of these three theridiosomatid species, including two new species with illustrations and photographs of both sexes. Furthermore, geographical distributional data including other theridiosomatids in the Ryukyus are also provided with a comparison of their natural history, especially habitat types, web morphology, and web-building behavior.

## Materials and methods

The specimens were preserved in $80 \%(\mathrm{v} / \mathrm{v})$ ethanol solution. Morphological features of the specimens were observed, and photographs were taken using a stereoscopic microscope (Nikon AZ100M, Japan). Photographed images were stacked using microscope imaging software (Nikon NIS-Elements D 4.20.00 64-bit, Japan). Photographs of $Z$. dibaiyin were taken using a digital camera (Nikon CF Plan X20 objective lens + Olympus M. Zuiko 75-300 mm attached to Olympus OM-D E-M1) and stacked using imaging software (Zerene Stucker; Zerene Systems, Washington, USA). The vulvae were treated with Proteinase K before being photographed. Measurements of the legs are given in the following format: femur + patella + tibia + metatarsus + tarsus $=$ total, in millimeters. The formula of macrosetae on the legs is as follows: $\mathbf{d}$, dorsal; $\mathbf{p}$, prolateral; $\mathbf{r}$, retrolateral. All specimens used in this study were deposited in the collection of the Department of Zoology, National Museum of Nature and Science (NSMT; curator: Ken-ichi Okumura), Tsukuba, Japan. Specimens without registration numbers in the 'material examined' section were deposited in the personal collection of YS.

Observations of Sennin tanikawai sp. nov. were conducted in Yutsun-do cave and Ôtomi-daiichi-do cave on Iriomote Island in August 1998, and April and June 2021.

The measurements of webs and visual observations of web-building behavior were conducted using a 6 V search light. The web size was measured for the horizontal and vertical diameters of the capture area. The number of sticky spirals was counted along a radius located at an angle of $45^{\circ}$ in the upper right sector of the orb. Web-building behavior was observed in the adult females. Webs were photographed and behaviors on the webs were recorded as movie in the spiders' natural habitat using a digital camera (Laowa 50 mm Ultra Macro + Olympus OM-D-E-M1; Canon DS6041 + Canon Macro Lens EF 100 mm ).

Abbreviations of morphological terminology are in accordance with Coddington (1986a), Zhu et al. (2001), Miller et al. (2009), Chen (2010), and Zhao and Li (2012). MAL, MAW, PCP, and RCP are defined herein for the first time; see below for all abbreviations for morphology:

| AL | abdomen length; | FD | fertilization duct; |
| :--- | :--- | :--- | :--- |
| ALE | anterior lateral eye; | MA | median apophysis; |
| AME | anterior median eye; | MAL | length of dorsal protrusion on me- |
| AW | abdomen width; |  | dian apophysis; |
| C | conductor; | MAW | width of dorsal protrusion on me- |
| CA | cymbial apophysis; |  | dian apophysis; |
| CaL | carapace length; | PC | paracymbium; |
| CAW | cymbial apophysis width; | PCP | posterior conductor projection; |
| CaW | carapace width; | PLE | posterior lateral eye; |
| CB | copulatory bursae; | PME | posterior median eye; |
| CD | copulatory duct; | PTL | palpal tibia length; |
| CL | cymbial lamella; | RCP | retrolateral conductor projection; |
| E | embolus; | S | spermatheca; |
| EA | embolic apophysis; | ST | subtegulum; |
| ED | embolic division; | T | tegulum; |
| ES | epigynal scape; | VW | vulva width. |
| ESL | epigynal scape length; |  |  |

Abbreviations for web architecture:

| HL | hub loop; | RD | radii; | TS | temporary spiral. |
| :--- | :--- | :--- | :--- | :--- | :--- |
| OH | open hub; | SS | sticky spirals; |  |  |
| RA | radial anastomosis; | TL | tension line; |  |  |

Key to the theridiosomatid species in the Ryukyu Islands
$\qquad$
_ Female ........................................................................................................ 6
2 Cymbium with a long dorsal cymbial apophysis, embolic apophyses entirely covered with conductor.

- Cymbium lacking cymbial apophysis.......................................................... 3

3 One embolic apophysis exposed from conductor ..................Zoma dibaiyin

- Two embolic apophyses exposed from conductor........................................ 4

4 Conductor with a projection............................ Theridiosoma dissimulatum

- Conductor lacking a projection. 5
5 Two paralleled embolic apophyses of same length
Theridiosoma alboannulatum
- One embolic apophysis longer than the other. $\qquad$
Theridiosoma nigrivirgatum sp. nov.
6 Epigyne with a scape on posterior margin Sennin tanikawai sp. nov.
- Epigyne lacking a scape 7
7 Epigyne with a sclerotized median pit ...................................Zoma dibaiyin
- Epigyne lacking a sclerotized median pit..................................................... 8

8 Epigyne with an invagination on posterior margin...................................... 9

- Epigyne lacking an invagination on posterior margin.

Theridiosoma alboannulatum
9 Heart-shaped invagination with a pair of spurs Theridiosoma dissimulatum

## Taxonomy

## Family Theridiosomatidae Simon, 1881

## Genus Theridiosoma O. Pickard-Cambridge, 1879

Type species. Theridiosoma gemmosum (L. Koch, 1877), from Nuremberg, West Germany (not examined).

Remarks. Males of Theridiosoma species can be distinguished from other theridiosomatid genera by the morphology of the embolic division of the male palp: short and tubular embolus with embolic apophyses fragmented into several long bristle-like parts (Coddington 1986a: figs 131, 133). Embolic apophysis varies in number and shape among species and is regarded as an important taxonomic character (e.g., Zhao and Li 2012; Suzuki et al. 2020). Median apophysis is less sclerotized, curved and attenuates distally (Coddington 1986a: figs 132, 133), which is less useful for distinguishing species. A sclerotized projection ('conductor projection') is present on prolateral side of conductor in some species, while absent in others (Coddington 1986a; Suzuki et al. 2020). Distal margin of conductor beneath embolic apophyses ('posterior margin of embolic division' in Suzuki et al. 2020) is generally sclerotized and the shape is useful as a taxonomic character (e.g., Suzuki et al. 2020: figs 7E, 8E, 10E, 11E). Tegular surface beneath conductor is generally sclerotized with many folds (referred as 'ventral side of tegulum beneath posterior edge of embolic division’ in Suzuki et al. 2020), of which shape and surface texture vary among species (Zhao and Li 2012; Suzuki et al. 2020).

Females of the genus can be distinguished from related genera (Baalzebub, Epilineutes and Wendilgarda) by having relatively sclerotized, robust copulatory ducts running
from the bursa to the spermathecae (Coddington 1986a: figs 145, 152). Surface of epigynal plate is smooth and its posterior margin generally lacks scape-like structures. Shape of posterior margin of epigynal plate varies among Theridiosoma species: rounded or almost straight in some species, while having a pair of small, sclerotized processes (named as 'spurs' in Coddington 1986a) or a small slit-like invagination in others (Coddington 1986a; Miller et al. 2009; Suzuki et al. 2020). Zoma females possess a similar genitalia except in having a sclerotized median pit on the surface of the epigynal plate and lacking any processes nor invaginations on the posterior margin of the epigynal plate in known species (Saaristo 1996; Miller et al. 2009; Zhao and Li 2012; Ballarin et al. 2021).

## Theridiosoma nigrivirgatum sp. nov.

https://zoobank.org/C1CB60C1-B782-4E34-9FAB-F44D3B12B4A0
[New Japanese name: Jyabara-karakara-gumo]
Figs 1-3, 11, 12C, 13A-C, 15A
Type material. Holotype: Japan, Okinawa Is. (Okinawa Prefecture): đ (NSMT-Ar 21717), Urasoe City, Nakama, Urasoe-daikoen Park ( $26^{\circ} 1^{\prime} 50.2^{\prime \prime} \mathrm{N}, 127^{\circ} 43^{\prime} 49.8^{\prime \prime} \mathrm{E}$, alt. 112 m ), 8 Mar. 2021, Y. Suzuki leg. Paratypes: 2 , same data as the holotype; $1 \delta^{\AA}$ 1 Q (NSMT-Ar 21718), Urasoe City, Nakama, Urasoe-daikoen Park ( $26^{\circ} 14^{\prime} 59.2^{\prime \prime} \mathrm{N}$, $127^{\circ} 43^{\prime} 54.6^{\prime \prime} \mathrm{E}$, alt. 64 m ), 16 Apr. 2021, Y. Suzuki leg.; 1 ठ 1 q (NSMT-Ar 21719), Nakagami District, Nishihara Town, Senbaru ( $26^{\circ} 15^{\prime} 01.8^{\prime \prime N}$, $127^{\circ} 45^{\prime} 57.8^{\prime \prime E}$ E, alt. 104 m), 25 Apr. 2021, Y. Suzuki leg.

Other material examined. Japan, Okinawa Is. (Okinawa Prefecture): 10 q, Naha City, Shuri-sueyoshi Town, Sueyoshi-koen Park ( $26^{\circ} 13^{\prime} 45.0^{\prime \prime} \mathrm{N}, 127^{\circ} 42^{\prime} 49.8^{\prime \prime} \mathrm{E}$, alt. 49 m), [7 Mar. 2021 (1 Q ), 8 Mar. 2021 (9 q)], Y. Suzuki leg.; 5 Q, Nakagami District, Nishihara Town, Tanabaru, Tanabaru Gusuku ( $26^{\circ} 14^{\prime} 44.3^{\prime \prime} \mathrm{N}, 127^{\circ} 45^{\prime} 16.4^{\prime \prime} \mathrm{E}$, alt. 141 m), 8 Apr. 2021, Y. Suzuki leg.; $2 \circlearrowleft^{\lambda}$, Kunigami District, Kunigami Village, Yona ( $26^{\circ} 44^{\prime} 35.2^{\prime \prime} \mathrm{N}, 128^{\circ} 14^{\prime} 55.1^{\prime \prime} \mathrm{E}$, alt. 195 m ), 19 Sep. 2021, Y. Suzuki leg.; 1 q, Ôgusuku ( $26^{\circ} 17^{\prime} 09.5^{\prime \prime N}$, $127^{\circ} 48^{\prime} 13.1^{\prime \prime} \mathrm{E}$, alt. 136 m ), Nakagami Distirct, Kitanakagusuku Village, R. Serita leg. Kume Is. (Okinawa Prefecture): 1 q 2 juv., Shimajiri District, Kumejima Town, Jyanado ( $26^{\circ} 20^{\prime} 46.2^{\prime \prime N}, 126^{\circ} 47^{\prime} 52.0^{\prime \prime} \mathrm{E}$, alt. 16 m ), 10 Sep. 2021, Y. Suzuki leg. Aka Is. (Okinawa Prefecture): 1 q, Shimajiri District, Zamami Village, Aka, streamside at dim forest ( $26^{\circ} 11^{\prime} 47.11^{\prime \prime} \mathrm{N}, 127^{\circ} 16^{\prime} 57.09^{\prime \prime} \mathrm{E}$, alt. 65 m), 16 Mar. 2022, Y. Suzuki leg.

Etymology. The specific name is a Latin adjective derived from the black striped pattern on the dorsal abdomen of the new species.

Diagnosis. Males of the new species resemble T. alboannulatum Suzuki, Serita \& Hiramatsu, 2020 in having two parallel embolic apophyses exposed from conductor and lacking a conductor projection on the male palp. They can be distinguished by the presence of one embolic apophysis longer than another and the shape of the sclerotized distal margin of conductor beneath embolic apophyses: a ridge separates the two triangular surfaces and sharply cornered at the terminal of the ridge in T. nigrivirgatum sp. nov. (Figs 2C, 3C), while a ridge is lacking in


Figure I. Theridiosoma nigrivirgatum sp. nov., male holotype (NSMT-Ar 21717 A-D) and female paratype (NSMT-Ar 21718 E-H) A, E habitus, dorsal view B, $\mathbf{F}$ habitus, lateral view $\mathbf{C}, \mathbf{G}$ habitus, ventral view $\mathbf{D}, \mathbf{H}$ abdomen, posterior view. Scale bars: 0.5 mm .
T. alboannulatum (Suzuki et al. 2020: fig. 12E, F). Females of the new species resemble those of T. diwang Miller, Griswold \& Yin, 2009 in having a small and narrow slit on the posterior margin of the epigynal plate, but can be distinguished by the shape of the vulva: genital plate is bell-shaped and longer than wide; spermathecae are positioned at the anterior part of the vulva in T. nigrivirgatum sp. nov. (Figs 2G, 3G), while the vulva is wider than long, copulatory ducts extend anteriorly, and the position of spermatheca is lower than the anterior margin of the copulatory ducts in T. diwang (Miller et al. 2009: fig. 3G). Both sexes can be distinguished from congeners by their abdominal color and patterns: a dark marking on the anterior dorsum, two pairs of dark markings on the dorsolateral side, and dark striped markings on the posterior dorsum (Fig. 1).

Description. Male (holotype, NSMT-Ar 21717). Measurements. Body 1.02 long. Carapace 0.45 long, 0.46 wide, and 0.36 high. Eye size and interdistances, AME 0.054 , ALE 0.047, PME 0.050, PLE 0.042, AME-AME 0.022, AME-ALE 0.017, PME-PME 0.012 , PLE-PLE 0.030 . Leg length: $\operatorname{leg} \mathrm{I} 0.47+0.17+0.31+0.29+0.20=1.44$; leg II $0.38+0.15+0.26+0.24+0.18=1.21 ; \operatorname{leg}$ III $0.23+0.13+0.14+0.18+0.13=0.81$; leg IV $0.30+0.13+0.20+0.20+0.15=0.98$. Abdomen 0.58 long, 0.60 wide, 0.80 high.


Figure 2. Theridiosoma nigrivirgatum sp. nov., male holotype genitalia (NSMT-Ar 21717 A-D) and female paratype genitalia (NSMT-Ar 21718 E-G) A retrolateral view $\mathbf{B}$ ventral view $\mathbf{C}$ posterior-ventral view $\mathbf{D}$ prolateral view $\mathbf{E}$ ventral view $\mathbf{F}$ ventral view $\mathbf{G}$ dorsal view. Abbreviations: $\mathbf{C}$ conductor CD copulatory ducts $\mathbf{E}$ embolus EA embolic apophysis FD fertilization ducts MA median apophysis PC paracymbium $\mathbf{S}$ spermatheca $\mathbf{S T}$ subtegulum $\mathbf{T}$ tegulum. Scale bars: 0.1 mm .

Carapace oval, wider than long (CaL/CaW 0.98). Chelicerae with three teeth on promargin. Abdomen oval and wider than long (AL/AW 0.97).

Coloration and markings (Fig. 1A-D). Carapace, chelicerae, and legs dark yellowish brown (turning to yellowish brown in ethanol). Cephalic groove stained with dark spots. Anterolateral margin of carapace dark grey. Mouthparts dark yellowish brown. Sternum pale yellowish brown with black lateral margins. Eyes on the dark bases. Legs yellowish brown with femora pale and lacking annulations. Abdomen pale yellowish brown with a dark greyish marking on anterior dorsum, two pairs of dark greyish spots on dorsolateral sides, and dark-colored longitudinal stripes on posterior dorsum. Spinnerets and ventral side of abdomen dark grey.

Palp (Figs 2A-D, 3A-D). Palpal patella with a strong retrolateral macroseta. Paracymbium hook-like with a blunt tip. Tegulum bulbous. Embolic division covered with a semitransparent conductor and composed of several apophyses. Conductor lacking conductor projection. Two long and parallel bristle-like embolic apophyses exposed from the conductor. Posterior margin of the embolic division strongly sclerotized


Figure 3. Theridiosoma nigrivirgatum sp. nov., male holotype genitalia (NSMT-Ar 21717 A-D) and female paratype genitalia (NSMT-Ar 21718 E-G) A retrolateral view B ventral view C posterior-ventral view $\mathbf{D}$ prolateral view $\mathbf{E}$ ventral view $\mathbf{F}$ ventral view $\mathbf{G}$ dorsal view. Abbreviations: C conductor $\mathbf{C D}$ copulatory ducts $\mathbf{E}$ embolus $\mathbf{E A}$ embolic apophysis $\mathbf{F D}$ fertilization ducts MA median apophysis PC paracymbium $\mathbf{S}$ spermatheca $\mathbf{S T}$ subtegulum $\mathbf{T}$ tegulum. White and black arrows indicate a ridge that separate the two triangular surfaces and a sharply cornered terminal of the ridge, respectively. Scale bars: 0.1 mm .
with angular corners, the middle one pointed, the retrolateral one blunt, a ridge separates two triangular surfaces: one is covered by embolic division and the other is not (Fig. 3C). Tegular surface beneath conductor weakly sclerotized with denticles. Median apophysis narrower toward the pointed tip.

Female (paratype: NSMT-Ar 21718). Measurements. Body 1.31 long. Carapace 0.51 long, 0.50 wide, 0.41 high. Eye size and interdistances: AME 0.057, ALE 0.058, PME 0.060, PLE 0.053, AME-AME 0.019, AME-ALE 0.032, PME-PME 0.009, PLE-PLE 0.043. Leg length: leg I $0.61+0.20+0.29+0.26+0.19=1.55$; leg II $0.40+0.16+0.24+0.19+0.16=1.15$; leg III $0.24+0.15+0.14+0.17+0.13=0.83$; leg IV $0.40+0.17+0.23+0.20+0.14=1.97$. Abdomen 0.86 long, 0.87 wide, 0.90 high.

Carapace oval and almost as long as wide ( $\mathrm{CaL} / \mathrm{CaW} 1.02$ ). Chelicerae with three teeth on promargin. Abdomen oval and as long as wide (CaL/CaW 0.99).

Coloration and markings (Fig. 1E-H). Carapace and chelicerae pale yellowish brown. Lateral margin of carapace dark grey. Eyes on the dark bases. Eyes, mouthparts, sternum, and legs as in male. Abdomen yellowish brown with dark greyish markings similar to the male, and dark orange markings on the dorsum and sides.

Genitalia (Figs 2E-G, 3E-G). Epigyne a wide plate with a short and narrow slit in the middle of the posterior margin. Vulva. Copulatory ducts moderately complicated. Spermathecae rounded triangular and juxtaposed. Fertilization ducts with curved tips.

Variations. The color and patterns of the abdomen vary: male specimens collected from Northern Okinawa lack longitudinal stripes on the posterior dorsum of the abdomen.

Taxonomic justification. Theridiosoma nigrivirgatum sp. nov. can safely be assigned to the genus according to the male palpal morphology: embolus short and tubular, and embolus apophyses fragmented into several long bristle-like parts.

Remarks. The males and females are considered to be the same species because of the similarity of body color and patterns and their sympatric occurrences. Although this species sympatrically occurred with T. dissimulatum on southern Okinawa Island (Fig. 11), no other undescribed candidates were collected.

Distribution. Japan (Okinawa, Kume and Aka Islands; Fig. 11).
Habitat. The new species inhabits forest floors of secondary forests, bushes, and grasslands. The species is frequently collected from an open environment covered by Poaceae grasses, where T. dissimulatum is never found (Fig. 12C). The habitat of this species resembles that of T. alboannulatum (Fig. 12D).

Web morphology. This species weaves a concave orb web with radial anastomosis and a tension line connected to substrates (Fig. 13A-C). A spider drags a tension line with a strong force so that the web is deformed to a conical shape. The web is similar to that of the congeners.

Egg sac. pale whitish brown and spherical with a long horizontal line and a short stalk (Fig. 15A).

Theridiosoma dissimulatum Suzuki, Serita \& Hiramatsu, 2020
Figs 11, 12A, 15B

Theridiosoma dissimulatum Suzuki, Serita \& Hiramatsu, 2020: 137, figs 1E-H, 3D-F, $5 \mathrm{C}, \mathrm{D}, 8 \mathrm{~A}-\mathrm{J}, 9 \mathrm{~A}-\mathrm{P}, 13 \mathrm{E}-\mathrm{H}$ (holotype male and paratypes from Amami Island, Japan; not examined).

Material examined. Japan, Amami Is. (Kagoshima Prefecture): 1 , Amami City, Nase-uragami Town ( $28^{\circ} 23^{\prime} 55.6^{\prime \prime N}$, $129^{\circ} 32^{\prime} 27.5^{\prime \prime} \mathrm{E}$, alt. 139 m ), 1 Jul. 2021, Y. Suzuki leg.; 4 Q, Amami City, Sumiyo Town, Nishinakama, Santaro-toge Pass ( $28^{\circ} 15^{\prime} 48.7^{\prime \prime N}$ N, $129^{\circ} 25^{\prime} 09.0^{\prime \prime} \mathrm{E}$, alt. 141 m ), 6 May 2021, Y. Suzuki leg.; 1 q, Ôshima District, Yamato Village, Ôganeku, Materiya-no-taki Waterfall ( $28^{\circ} 19^{\prime} 04.4^{\prime \prime N}, 129^{\circ} 21^{\prime} 08.2^{\prime \prime} \mathrm{E}$, alt. 176 m), 4 Jul. 2021, Y. Suzuki leg. Okinoerabu Is. (Kagoshima Prefecture): 2 $\delta^{\top} 4$, Ôshima District, China Town, Tokudoki ( $27^{\circ} 21^{\prime} 34.5^{\prime \prime N}$, $128^{\circ} 33^{\prime} 02.8^{\prime \prime} \mathrm{E}$, alt. 134 m), 8 Dec. 2021, Y. Suzuki leg. Okinawa Is. (Okinawa Prefecture): 2 § 3 , Naha City, Shuri-sueyoshi Town, Sueyoshi-koen Park ( $26^{\circ} 13^{\prime} 39.6^{\prime \prime} \mathrm{N}, 127^{\circ} 42^{\prime} 55.3^{\prime \prime} \mathrm{E}$, alt. 25 m), 7 Mar. 2021, Y. Suzuki leg.; 2 , Kunigami District, Ogimi Village, Nerome ( $26^{\circ} 40^{\prime} 49.7^{\prime \prime N}, 128^{\circ} 08^{\prime} 01.1^{\prime \prime} \mathrm{E}$, alt. 128 m ), 14 Apr. 2021, Y. Suzuki leg.; 1 \&, Kunigami District, Ôgimi Village, Ôgimi ( $26^{\circ} 40^{\prime} 57.9^{\prime \prime N}, 128^{\circ} 08^{\prime} 21.6^{\prime \prime} \mathrm{E}$, alt. $311 \mathrm{~m}), 15$ May 2021, Y. Suzuki leg. Iriomote Is. (Okinawa Prefecture): 1 § 3 Q, Yaeyama District, Taketomi Town, Haiminaka, Ôtomi-rindo Path $\left(24^{\circ} 17^{\prime} 52.6^{\prime \prime N}\right.$, $123^{\circ} 52^{\prime} 47.3^{\prime \prime} \mathrm{E}$, alt. 18 m ), 30 Apr. 2021, Y. Suzuki leg.; $1 \circlearrowleft^{\lambda} 3$. Ôtomi-daiichi-do Cave ( $24^{\circ} 17^{\prime} 31.0^{\prime \prime N}$, $123^{\circ} 52^{\prime} 45.7^{\prime \prime} \mathrm{E}$, alt. 30 m ), 1 May 2021, Y. Suzuki leg.

Remarks. This species can easily be distinguished from T. nigrivirgatum sp. nov. by the presence of a conductor projection on the male palp and a heart-shaped invagination with a pair of spurs on the posterior margin of the female epigynal plate (Suzuki et al. 2020). Refer to the description in Suzuki et al. (2020) for further morphological information.

Distribution. Japan (Amami, Okinoerabu, Okinawa, Ishigaki, and Iriomote Islands; Fig. 11).

Habitat. This species was collected from dim moist forests, especially from locations beside streams (Fig. 12A).

Web morphology. Theridiosoma dissimulatum weaves a concave orb web and drags a tension line with the forelegs.

Egg sac. pale reddish brown and spherical with a long horizontal line and a short stalk (Fig. 15B).

## Theridiosoma alboannulatum Suzuki, Serita \& Hiramatsu, 2020

Figs 12D, 13D-E, 15C

Theridiosoma alboannulatum Suzuki, Serita \& Hiramatsu, 2020: 149, figs 2I-L, 4G-I, 6E-F, 12A-J, 13P (holotype male and paratypes from Iriomote Island, Japan; not examined).

Material examined. Japan, Kurima Is. (Okinawa Prefecture): 2 §, Miyakojima City, Shimojikuruma ( $24^{\circ} 43^{\prime} 29.2^{\prime \prime} \mathrm{N}, 125^{\circ} 15^{\prime} 09.2^{\prime \prime} \mathrm{E}$, alt. 41 m ),, 17 Nov. 2021, Y. Suzuki leg. Miyako Is. (Okinawa Prefecture): $3 \widehat{\delta}^{\wedge} 3$, Miyakojima City, Hiraranishihara, grassland at roadside ( $24^{\circ} 49^{\prime} 53.5^{\prime \prime} \mathrm{N}, 125^{\circ} 18^{\prime} 55.0^{\prime \prime} \mathrm{E}$, alt. 46 m ), 24 Apr. 2022, Y. Suzuki leg. Kuroshima Is. (Okinawa Prefecture): $2 \Uparrow 1 q$, Yaeyama District, Taketomi

Town, edge of coastal forest besides Hokei beach ( $24^{\circ} 14^{\prime} 26.2^{\prime \prime} \mathrm{N}, 123^{\circ} 59^{\prime} 32.7^{\prime \prime} \mathrm{E}$, alt. 0 m ), 2 Nov. 2021, Y. Suzuki leg. Yonaguni Is. (Okinawa Prefecture): 1 q, Yaeyama District, Yonaguni Town, Yonaguni ( $24^{\circ} 27^{\prime} 58.8^{\prime \prime} \mathrm{N}, 123^{\circ} 01^{\prime} 20.1^{\prime \prime} \mathrm{E}$, alt. 49 m ), 12 Oct. 2021, Y. Suzuki leg.; 2 § 2 , Yaeyama District, Yonaguni Town, Sonai Village ( $24^{\circ} 28^{\prime} 10.5^{\prime \prime N}$, $123^{\circ} 00^{\prime} 31.7^{\prime \prime} \mathrm{E}$, alt. 19 m ), 12 Oct. 2021, Y. Suzuki leg.; 1 § 1 Q, Yaeyama District, Yonaguni Town, wetland beside secondary forest ( $24^{\circ} 27^{\prime} 16.3^{\prime \prime} \mathrm{N}$, $122^{\circ} 59^{\prime} 24.3^{\prime \prime} \mathrm{E}$, alt. 38m), 14 Oct. 2021, Y. Suzuki leg.

Note. See diagnosis section for comparison with T. nigrivirgatum sp. nov.
Habitat. This species inhabits grasslands, bushes, and secondary forests. Spiders were collected from the basal parts of grasses.

Web morphology. The spider weaves a concave web between the grasses (Fig. 13D, E).
Egg sac. similar to that of T. nigrivirgatum sp. nov. (Fig. 15C).
Distribution. Japan (Miyako, Kurima, Iriomote, Kuroshima, and Yonaguni Islands; Fig. 11).

## Genus Zoma Saaristo, 1996

Type species. Zoma zoma Saaristo, 1996, from Seychelles (not examined).
Composition. Zoma zoma Saaristo, 1996, Z. dibaiyin Miller, Griswold \& Yin, 2009, Z. fascia Zhao \& Li, 2012, Z. taiwanica (Zhan, Zhu \& Tso, 2006).

Remarks. Females of the genus can be distinguished by the flat and bluntly triangular genital plate with a sclerotized median pit and a pair of smaller, generally less recognizable, lateral pits (Fig. 4J; see also Saaristo 1996). Males of the type species Z. zoma have not yet been described. Therefore, the taxonomic characteristics of Zoma males are poorly defined. Males of three Zoma species, Z. dibaiyin, Z. fascia, and Z. taiwanica have relatively simpler palps with a filiform embolic apophysis emerging beneath from the conductor, while two or more apophyses in Theridiosoma (Fig. 3B, C vs. Fig. 5A, B). Zoma species have wider and straight median apophysis, while curved and sharp tip in Theridiosoma (Fig. 3B-D vs. Fig. 5A-C). Zoma species can be distinguished from congeners by the presence of a transverse whitish silver band on the dorsum abdomen (Saaristo 1996; Miller et al. 2009).

## Zoma dibaiyin Miller, Griswold \& Yin, 2009

Figs 4-5, 11, 12B, 13F-H
Zoma dibaiyin Miller, Griswold \& Yin, 2009: 27, figs 10A-F, 11A-B, 13A-D (holotype male and paratypes from China; not examined); Ono and Ogata 2018: 120, 504, 505; Suzuki and Serita 2021a: 231, figs 1-3.

Material examined. Japan, Amami Is. (Kagoshima Prefecture): $1 \AA^{\lambda} 1$ (NSMTAr. 21720, 21721), Ôshima District, Setouchi Town, Katsuura ( $28^{\circ} 12^{\prime} 33.7^{\prime \prime N}$,
$129^{\circ} 19^{\prime} 54.0^{\prime \prime} \mathrm{E}$, alt. 354 m ), 4 Jul. 2021, Y. Suzuki leg.; 1 § 1 q, Amurogama ( $28^{\circ} 13^{\prime} 15.4^{\prime \prime N}, 129^{\circ} 18^{\prime} 59.3^{\prime \prime E}$, alt. 111 m ), 4 Jul. 2021, Y. Suzuki leg.; 1 §, Amami City, Sumiyo Town, Nishinakama, Santarou-toge Pass ( $28^{\circ} 15^{\prime} 48.7^{\prime \prime} \mathrm{N}, 129^{\circ} 25^{\prime} 09.0^{\prime \prime} \mathrm{E}$, alt. 141 m ), 6 May 2021, Y. Suzuki leg.; Okinawa Is. (Okinawa Prefecture): 1 ठ 1 Q, Kunigami District, Ôgimi Village, Nerome ( $26^{\circ} 40^{\prime} 49.7^{\prime \prime N}$, $128^{\circ} 08^{\prime} 01.1^{\prime \prime} \mathrm{E}$, alt. 128 m), 14 Apr. 2021, Y. Suzuki leg.; Kume Is. (Okinawa Prefecture): 1 § 1 ㅇ, Shimajiri District, Kumejima Town, Uezu, Mt. Daruma-yama ( $26^{\circ} 21^{\prime} 42.9^{\prime \prime N}$, $126^{\circ} 45^{\prime} 34.9^{\prime \prime} \mathrm{E}$, alt. 149 m ), 10 Sep. 2021, Y. Suzuki leg.

Diagnosis. Males of this species can be distinguished from congeners by the embolic apophysis with a curved tip running along the sclerotized surface of the ventral tegulum, and females by a nearly transverse posterior margin of the epigynal plate (more convex in Z. zoma and rounded in Z. taiwanica) and lower position of the spermathecae (higher in Z. taiwanica) (Miller et al. 2009; Ballarin et al. 2021).

Description. Male (NSMT-Ar 21720). Measurements. Body 1.62 long. Carapace 0.74 long, 0.68 wide, 0.60 high. Eye size and interdistances: AME 0.090, ALE 0.076, PME 0.093, PLE 0.065, AME-AME 0.015, AME-ALE 0.032, PME-PME 0.008, PLE-PLE 0.070. Leg length: leg I $0.80+0.23+0.57+0.47+0.28=2.35$; leg II $0.50+0.21+0.46+0.31+0.28=1.76$; leg III $0.40+0.19+0.23+0.24+0.24=1.30$; leg IV $0.44+0.14+0.34+0.27+0.27=1.46$. Abdomen 0.88 long, 0.95 wide, 1.01 high.

Carapace oval, longer than wide ( $\mathrm{CaL} / \mathrm{CaW} 1.34$ ). Chelicerae with three teeth on promargin. Abdomen oval, wider than long (AL/AW 0.93).

Coloration and markings (Fig. 4A, B). Carapace, chelicerae, maxillae, labium, sternum, and legs yellowish brown. Eyes on the dark bases. Cephalic groove stained with darkspots. Legs lacking annulation. Abdomen dark brown encircled dorsolaterally with a whitish silver band.

Palp (Figs 4C-H, 5). Paracymbium with sharp tip. Tegulum bulbous. Median apophysis weakly sclerotized, wider than long. Embolic division branched into a few bristle-like apophyses, embolus short and tubular, and a long and filiform embolic apophysis emerging beneath from a translucent conductor. Tip of embolic apophysis curved and running along sclerotized surface of the tegulum beneath the conductor. Conductor having two projections: posterior conductor projection strongly sclerotized and triangular with a blunt tip; retrolateral conductor projection strongly sclerotized and weakly curved anteriorly with a triangular posterior tip. Posterior margin of conductor with a sharp tip.

Female (NSMT-Ar 21721). Measurements. Body 2.04 long. Carapace 0.82 long, 0.76 wide, 0.67 high. Eye size and interdistances: AME 0.089, ALE 0.082 , PME 084, PLE 0.076, AME-AME 0.023, AME-ALE 0.034, PME-PME 0.009, PLE-PLE 0.076. Leg length: leg I $0.66+0.32+0.42+0.33+0.22=1.95$; leg II $0.64+0.30+0.4$ $1+0.33+0.21=1.89$; leg III $0.36+0.18+0.20+0.25+0.18=1.17$; leg IV $0.53+0.21+0.34+0.27+0.22=1.57$. Abdomen 1.21 long, 1.32 wide, 1.16 high.

Carapace, mouthparts, and abdomen as in male (CaL/CaW 1.08; AL/AW 0.92).
Coloration and markings (Fig. 4I) similar to male.
Genitalia (Fig. 4J, K). Epigynal plate flat and wider than long with a sclerotized posterior margin and a median pit. Spermathecae touching each other, copulatory


Figure 4. Zoma dibaiyin Miller, Griswold \& Yin, 2009, male habitus and genitalia (NSMT-Ar 21720 $\mathbf{A}-\mathbf{H}$ ) and female habitus and genitalia (NSMT-Ar 21721 I-K) A habitus, dorsal view $\mathbf{B}$ habitus, lateral view $\mathbf{C}$ palp, retrolateral view $\mathbf{D}$ palp, ventral view $\mathbf{E}$ palp, posterior-ventral view $\mathbf{F}$ palp, prolateral view G embolic division, prolateral view $\mathbf{H}$ embolic division, posterior-ventral view I habitus, dorsal view J epigyne, ventral view $\mathbf{K}$ vulva, dorsal view. Abbreviations: $\mathbf{C}$ conductor $\mathbf{C D}$ copulatory ducts $\mathbf{E}$ embolus EA embolic apophysis FD fertilization ducts MA median apophysis PC paracymbium PCP posterior conductor projection RCP retrolateral conductor projection $\mathbf{S}$ spermatheca $\mathbf{S T}$ subtegulum $\mathbf{T}$ tegulum. Arrows in E, F, H indicate a cornered margin of posterior membrane of conductor. Arrow in $\mathbf{J}$ indicates a sclerotized median pit. Scale bars: $1.0 \mathrm{~mm}(\mathbf{A}, \mathbf{B}, \mathbf{I}) ; 0.1 \mathrm{~mm}(\mathbf{C}-\mathbf{H}, \mathbf{J}, \mathbf{K})$.


Figure 5. Zoma dibaiyin Miller, Griswold \& Yin, 2009, male genitalia (NSMT-Ar 21720) A ventral view $\mathbf{B}$ posterior-ventral view $\mathbf{C}$ prolateral view $\mathbf{D}$ conductor and embolic division, retrolateral view $\mathbf{E}$ conductor and embolic division, posterior-ventral view. Abbreviations: $\mathbf{C}$ conductor $\mathbf{C D}$ copulatory ducts $\mathbf{E}$ embolus EA embolic apophysis MA median apophysis PC paracymbium PCP posterior conductor projection RCP retrolateral conductor projection ST subtegulum $\mathbf{T}$ tegulum. Arrows in $\mathbf{B}-\mathbf{E}$ indicate a cornered margin of posterior membrane of conductor. Scale bars: 0.1 mm .
ducts wide at their openings, and the course of the ducts simple. See Miller et al. (2009) for further details.

Remarks. A strongly sclerotized triangular projection with a rounded tip (Figs 4E, 5B) and a cornered margin of the posterior membrane of the conductor (Figs 4E, 5B, arrowed) were visible in the ventro-posterior view of the male palp. The triangular projection does not seem to be homologous to conductor projection in Theridiosoma, as the former protrudes from the posterior margin of the conductor, while the latter was positioned on the surface of the conductor. Herein, we define it as posterior conductor projection (PCP).

The embolus on the male palp was not determined in $Z$. dibaiyin and Z. fascia (Miller et al. 2009; Zhao and Li 2012). In Z. taiwanica, the embolus is described as a 'short and tubular structure' but lacks explanations in illustrations (Zhang et al. 2006). Ballarin et al. (2021) determined the embolus as a strongly sclerotized, thin, stick-like apophysis located on the retrolateral side of the embolic division (Ballarin et al. 2021). Considering the shape of Theridiosoma's embolus, which is short, tubular, and hidden under conductor (see Coddington 1986a: fig. 131), we suppose the 'embolus' of Zoma species determined in Ballarin et al. (2021) is not an embolus. The true embolus of Z. dibaiyin is found beneath basal part of embolic apophyses (Fig. 5D). Hereafter we defined the sclerotized stick-like structure as a retrolateral conductor projection (RCP), and distinguishable from an embolus. The shape of RCP is useful as a taxonomic character for differentiating species within the genus Zoma: S-shaped with pointed tip in Z. fascia, claviform with rounded tip in Z. taiwanica (Zhao and Li 2012: figs 28, 30; Zhang et al. 2006: figs 4-6; Ballarin et al. 2021: fig. 5B, C), and weakly curved anteriorly with a triangular posterior tip in $Z$. dibaiyin (Figs 4D, 5D).

Distribution. China (Yunnan), Japan (Honshu to the Ryukyu Islands; Fig. 11).
Habitat. This species inhabits the forest floor and streamside in dim and wet forests (Fig. 12B).

Web morphology. This species weaves a concave orb web with radial anastomosis above the ground (Fig. 13F-H). The orb web is almost horizontal. A tension line stretched from the center of the web and attached to substrates such as rocks and dead leaves. The mesh of sticky spirals tends to be fine (occasionally, the number of sticky lines is $>30$ ). For details of the web morphology, see Hiramatsu (2021).

Egg sac. light brown with a distinct circular suture at the upper end. The sac was suspended from a long horizontal line with a short stalk (see Hiramatsu 2021).

## Genus Sennin gen. nov.

https://zoobank.org/AAA86579-ABA5-4385-B0D7-68D7676BD871
[New Japanese name: Hora-ana-karakara-gumo-zoku]
Type species. Sennin tanikawai sp. nov.
Etymology. The generic name Sennin is noun in apposition, masculine, and derived from the Japanese word meaning mountain hermits, a person who acquires a spiritual power after living a secluded life deep in the mountains. Iriomote Island, where the new species inhabits, is famous for a man called Sennin, who was self-sufficient, lived in the coastal caves, and single-handedly built a wooden hut.

Diagnosis. This genus can be distinguished from other theridiosomatid genera by the following characteristics: a large, oblong cymbial outgrowth (cymbial apophysis) protruding from the basal and dorsal part of cymbium of male palp (Figs 7A-C, 9 A-C); an embolic division with three elongated bristle-like embolic apophyses with the longest one coiled (Figs 7F-J, 9F-J); the anterior margin of the epigynal plate with
a pair of sclerotized, triangular extensions protruding anteriorly from the anterolateral side (Figs 8A, D; 10A, D; arrowed; Zhu et al. 2001: fig. 4; Chen 2010: figs 19, 20); the vulva with long copulatory ducts coiling at the lateral side of the spermatheca (Figs 8C, D, 10C, D).

Composition. Sennin tanikawai sp. nov., S. coddingtoni (Zhu, Zhang \& Chen, 2001), comb. nov.

Remarks. This genus is related to Baalzebub Coddington, 1986, based on the shape of the median apophysis on the male palp, the embolic apophyses that are not exposed from the conductor, and the general morphology of the epigyne. The elongated and oblong dorsal cymbial apophysis, one of the most conspicuous characters of Sennin gen. nov. (Figs 7A-C, 9A-C), differentiates the new genus from Baalzebub. Although some species of Baalzebub have a small protrusion on the retrolateral-dorsal side of basal part of cymbium (e.g., paracymbium in B. acutum; Prete et al. 2016: figs 2D, 3C; named 'Höcker' (= lump) in B. brauni; Wunderlich 1976: figs 17, 18), it is not as prominent as that of Sennin gen. nov. The embolic apophyses of Baalzebub are short, blunt, and spatulate, but those of Sennin are longer, bristle-like, and strongly curved or coiled (Figs 7F-J, 9F-J). As for the female genitalia of species of Baalzebub, the epigynal plate is upside-down triangular with sclerotized central epigynal pit, the spermathecae elliptical, and longer laterally with connate tips, and the course of copulatory ducts is simple (Coddington 1986a). Sennin gen. nov. has similar spermathecae, but the course of the copulatory duct is more complex, with a coiled trajectory at the basal side of the spermathecae (Figs 8C, D, 10C, D).

Sennin coddingtoni comb. nov. was formerly placed in Karstia Chen, 2010, but it shares conspicuous characteristics with $S$. tanikawai sp. nov. and can clearly be differentiated from K. upperyangtzica Chen, 2010, the type species of the genus. Therefore, we transferred it from Karstia to Sennin gen. nov. Karstia upperyangtzica and K. cordata Dou \& Li (2012) females have an upside-down triangular epigynal plate with a sclerotized epigynal pit, and a simple course of copulatory ducts; males have cymbial apophysis as a very small protrusion, and embolic division with short, spatulate embolic apophyses (Chen 2010; Dou and Lin 2012; Zhang and Wang 2017). Based on these morphological characteristics, it is difficult to differentiate K. upperyangtzica and K. cordata from Baalzebub; therefore, taxonomic revision of Karstia is needed. In this study, we defer revision of Karstia, which may require direct examination of the type specimens and further molecular analysis.

As mentioned above, taxonomic relationship between Sennin gen. nov. and its potentially closest-related genera (Baalzebub and probably Karstia) is not yet well defined. This also indicated that the establishment of Sennin gen. nov. could render these related genera polyphyletic. To revise taxonomic status of these taxa in terms of monophyly, further integrative phylogenetic approach covering large number of species and genera is required.

According to the morphology and a potential close-relatedness to Baalzebub, Sennin gen. nov. is here suggested to be assigned to the subfamily Theridiosomatinae.

## Sennin tanikawai sp. nov.

https://zoobank.org/2FB64512-C697-4195-AE82-62C4563508DD
[New Japanese name: Iriomote-hora-ana-karakara-gumo]
Figs 6-10, 11, 12E, 14A-G, 15D-E
Type material. Holotype: Japan, Iriomote Is. (Okinawa Prefecture): $\begin{gathered} \\ \text { (NSMT-Ar }\end{gathered}$ 21722), Yaeyama District, Taketomi Town, Haiminaka, Ôtomi-Daini-Do Cave, 31 Mar. 1985, A. Tanikawa leg. Paratypes: 2 \& (NSMT-Ar 21723), 27 Mar. 1995, A. Tanikawa leg.; 9 ô (NSMT-Ar 21724-21725), 31 Mar. 1985, A. Tanikawa leg.; 1 ㅇ (NSMT-Ar 21726), 1 Aug. 1970, Y. Shirota leg.; 2 \& (NSMT-Ar 21727), 27 Oct. 1977, N. Tsurusaki leg.; above paratypes are collected at same locality as the holotype; $1 \widehat{o}^{\lambda}$ (NSMT-Ar 21728), a small opening of Ôtomi-Daini-Do Cave ( $24^{\circ} 17^{\prime} 09.4$ "N, $123^{\circ} 53^{\prime} 24.9^{\prime \prime} \mathrm{E}$, alt. 10 m ), 24 Jun. 2021, Y. Suzuki leg.

Additional material examined. Japan, Iriomote Is. (Okinawa Prefecture): 1 q, Yaeyama District, Taketomi Town, Haemi, Ôtomi-daiichi-do Caves ( $24^{\circ} 17^{\prime} 31.0^{\prime \prime} \mathrm{N}$, $123^{\circ} 52^{\prime} 45.7^{\prime \prime}$ E, alt. 30 m ), 3 May 2021, Y. Suzuki leg.; 1 个, Yaeyama District, Taketomi Town, Takana, Yutsun-Do Caves, a small cave on coastal cliff ( $24^{\circ} 23^{\prime} 08.90^{\prime \prime} \mathrm{N}$, $123^{\circ} 53^{\prime} 27.89^{\prime \prime}$ E, alt. 10 m ), 21 Mar. 2019, Y. Suzuki leg.; 1 \&, Takana, Yutsun-Do Caves, a large cave opening on coastal cliff ( $24^{\circ} 23^{\prime} 05.88^{\prime \prime} \mathrm{N}, 123^{\circ} 53^{\prime} 25.00^{\prime \prime} \mathrm{E}$, alt. 7 m), 28 Mar. 2008, T. Hiramatsu leg.; 6 § 7 \& , Takana, Yutsun-Do Caves, a large cave opening on coastal cliff ( $24^{\circ} 23^{\prime} 05.88^{\prime \prime} \mathrm{N}, 123^{\circ} 53^{\prime} 25.00$ " E , alt. 7 m ), 1 May 2021, Y. Suzuki leg.; 2 \& , Takana, Yutsun-Do Caves, cavities of rocks on coastal cliff ( $24^{\circ} 23^{\prime} 04.96^{\prime \prime N}$, $123^{\circ} 53^{\prime} 23.82^{\prime \prime} \mathrm{E}$, alt. 10 m ), 22 Jun. 2021, Y. Suzuki leg.; 3 ㅇ, Takana, Yutsun-Do Caves, a cave beside Shirahama-haemi-sen road ( $24^{\circ} 23^{\prime} 06.5^{\prime \prime} \mathrm{N}$, $123^{\circ} 53^{\prime} 31.2^{\prime \prime} \mathrm{E}$, alt. 27 m ), 24 Jun. 2021, Y. Suzuki leg.; $1 \delta^{\lambda} 1$ q, Haemi, limestone rocky walls in a secondary forest ( $24^{\circ} 16^{\prime} 08.48^{\prime \prime} \mathrm{N}, 123^{\circ} 52^{\prime} 01.52^{\prime \prime} \mathrm{E}$, alt. 16 m ), 24 Jun. 2021, Y. Suzuki leg.

Etymology. The specific name is patronym dedicated to Dr. Akio Tanikawa, a Japanese arachnologist who has contributed remarkably to the elucidation of the spider fauna in Iriomote Island and offered us many specimens including type specimens.

Diagnosis. Males of this species can be distinguished from the allied Sennin coddingtoni comb. nov. by the following characteristics: cymbial apophysis is wider in relation to palpal tibia length while it is almost the same length as $S$. coddingtoni comb. nov. $($ CAW/PTL $=2.41$ in S. tanikawai sp. nov., also see Fig. 9A, B; 1.00 in S. coddingtoni comb. nov.; also see Zhu et al. 2001: fig. 7); median apophysis of S. tanikawai sp. nov. is longer and narrower dorsally (MAL/MAW 1.63; Fig. 9E) compared to that of the latter (MAL/MAW 0.85, based on Chen 2010: fig. 24); the less-sclerotized distal part of median apophysis is lanceolate with pointed tip on ventral terminal in S. tanikawai sp. nov. (arrows in Figs 7E, 9E), while that of S. coddingtoni comb. nov. is falcate (Chen 2010: fig. 24). Females of S. tanikawai sp. nov. can be distinguished from S. coddingtoni comb. nov. by the following characteristics: longer epigynal scape (ESL/VW 0.46 in S. tanikawai sp. nov., Fig. 10; 0.16 in S. coddingtoni comb. nov., based on Chen 2010: fig. 19); tip of spermatheca is strongly curved anteriorly in


Figure 6. Sennin tanikawai sp. nov., male holotype (NSMT-Ar 21722 A-D) and female paratype (NS-MT-Ar $21723 \mathbf{E - H} \mathbf{A}, \mathbf{E}$ habitus, dorsal view $\mathbf{B}, \mathbf{F}$ habitus, lateral view $\mathbf{C}, \mathbf{G}$ eye region, dorsal view D, $\mathbf{H}$ chelicera, anterior view. Scale bars: $0.5 \mathrm{~mm}(\mathbf{A}, \mathbf{B}, \mathbf{E}, \mathbf{F}) ; 0.1 \mathrm{~mm}(\mathbf{C}, \mathbf{G}, \mathbf{D}, \mathbf{H})$.
S. tanikawai sp. nov., whereas it is almost straight in S. coddingtoni comb. nov.; the course of copulatory ducts: ducts from both sides juxtaposed at the middle of vulva ventral to spermathecae and continue posteriorly straight toward epigynal scape, then make a right-angle turn and apart laterally (arrows in Figs 8C, 10C), while in S. coddingtoni comb. nov. the ducts apart to each other ventrally to the spermatheca and curved laterally (Zhu et al. 2001: fig. 5).

Description. Male (NSMT-Ar 21722). Measurements. Body 2.30 long. Carapace 1.07 long, 1.10 wide, 0.72 high. Eye size and interdistances: AME 0.09, ALE 0.09, PME 0.10, PLE 0.08, AME-AME 0.02, AME-ALE 0.03, PME-PME 0.03, PLE-PLE 0.08 , Leg length: leg I $1.62+0.53+1.30+1.08+0.50=5.03$; leg II $1.30+0.49$ $+1.03+0.91+0.49=4.22$; leg III $0.70+0.39+0.56+0.63+0.38=2.66$; leg IV $0.93+0.40+0.73+0.69+0.38=3.14$. Abdomen 1.32 long, 1.44 wide, 1.61 high.


Figure 7. Sennin tanikawai sp. nov., male holotype genitalia (NSMT-Ar 21722) A-D male palp F-J embolic division $\mathbf{A}$ retrolateral view $\mathbf{B}$ prolateral view $\mathbf{C}$ dorsal view $\mathbf{D}$ ventral view $\mathbf{E}$ median apophysis, ventral view $\mathbf{F}$ posterior-ventral view $\mathbf{G}$ ventral view $\mathbf{H}$ anterior-dorsal view I posterior-dorsal view $\mathbf{J}$ prolateral view. Abbreviations: $\mathbf{C}$ conductor $\mathbf{C A}$ cymbial apophysis $\mathbf{C L}$ cymbial lamella $\mathbf{E}$ embolus EA embolic apophysis ED embolic division EM embolus MA median apophysis PC paracymbium ST subtegulum $\mathbf{T}$ tegulum. Arrow in $\mathbf{E}$ indicates the tip of less-sclerotized region of median apophysis. Scale bars: 0.1 mm .


Figure 8. Sennin tanikawai sp. nov., female paratype genitalia (NSMT-Ar 21723) A epigyne, ventral view $\mathbf{B}$ epigyne, lateral view $\mathbf{C}$ vulva, dorsal view $\mathbf{D}$ vulva, anterior view. Abbreviations: $\mathbf{C B}$ copulatory bursae $\mathbf{C D}$ copulatory ducts ES epigynal scape FD fertilization ducts $\mathbf{S}$ spermatheca. Arrows in $\mathbf{A}, \mathbf{D}$ indicate a pair of sclerotized extensions on the anterior margin of epigynal plate. Arrow in $\mathbf{C}$ indicates a pair of copulatory ducts juxtaposed. Scale bars: 0.1 mm .

Carapace oval, wider than long (CaL/CaW 0.97). Chelicerae with six teeth on promargin with the largest one positioned close to the fang base, no teeth on retromargin (Fig. 6D). Anterior eye row recurved, posterior eye row straight. Cymbial apophysis of palp 0.297 long, 0.111 wide. Macrosetae: leg I: femur r1-p1, patella d1, tibia d2-r1-p1; leg II: femur r1, patella d1, tibia d2-r1; leg III: patella d1, tibia d1; leg IV: patella d1, tibia d1. Abdomen oval, wider than long (AL/AW 0.92). Abdomen covered with long and thin setae.

Coloration and markings (Fig. 6). Carapace, mouthparts, sternum, and legs dark yellowish brown (turning to yellowish brown in ethanol). Eyes on dark bases. Legs lacking annulation. Abdomen pale yellowish grey, dorsum of abdomen with two pairs of sigilla.


Figure 9. Sennin tanikawai sp. nov., male holotype genitalia (NSMT-Ar 21722) A-D male palp F-J embolic division $\mathbf{A}$ retrolateral view $\mathbf{B}$ prolateral view $\mathbf{C}$ dorsal view $\mathbf{D}$ ventral view $\mathbf{E}$ median apophysis, ventral view $\mathbf{F}$ posterior-ventral view $\mathbf{G}$ ventral view $\mathbf{H}$ anterior-dorsal view $\mathbf{I}$ posterior-dorsal view $\mathbf{J}$ prolateral view. Abbreviations: $\mathbf{C}$ conductor $\mathbf{C A}$ cymbial apophysis $\mathbf{C L}$ cymbial lamella $\mathbf{E}$ embolus $\mathbf{E A}$ embolic apophysis ED embolic division EM embolus MA median apophysis PC paracymbium ST subtegulum $\mathbf{T}$ tegulum. Arrow in $\mathbf{E}$ indicates the tip of less-sclerotized region of median apophysis. Scale bars: 0.1 mm .


Figure 10. Sennin tanikawai sp. nov., female paratype (NSMT-Ar 21723) A epigyne, ventral view B epigyne, lateral view $\mathbf{C}$ vulva, dorsal view $\mathbf{D}$ vulva, anterior view $\mathbf{E}$ course of copulatory duct. Abbreviations: CB copulatory bursae CD copulatory ducts ES epigynal scape ESL epigynal scape length FD fertilization ducts $\mathbf{S}$ spermatheca $\mathbf{V W}$ vulva width. Arrows in $\mathbf{A}, \mathbf{D}$ indicate a pair of sclerotized extensions on the anterior margin of epigynal plate. Arrow in $\mathbf{C}$ indicates the pair of copulatory ducts juxtaposed. Scale bars: 0.1 mm .

Palp (Figs 7, 9). Palpal patella with a strong dorsal macroseta. Paracymbium hooklike with a sharp tip. Cymbial lamella robust. Dorsal cymbial apophysis oblong, platelike with blunt tip, extending anterior-retrolaterally. Tegulum large, bulbous, and occupying a large part of the palpal organ. Embolic division is a complex of long bristle-like apophyses, entirely covered with translucent conductor, and none of the embolic apophyses are exposed. Embolus short, blunt, and covered with a membrane. Three embolic apophyses conspicuous, EA 1 thickest, protruding middle of embolic division, S-shaped and sharper distally; EA 2 longest among them, bristle-like, basal part swelled, forming a loop at the ventro-prolateral side, distal part along with EA 1; EA 3 thinnest, protruding from prolateral side of embolic division. Median apophysis with a deep groove dividing it into two parts, distal translucent, weakly sclerotized and sharper ventrally, basal triangular, strongly sclerotized with narrower dorsally and spatula-like at ventral tip, MAL 1.07, MAW 0.66.


Figure II. Distribution of theridiosomatid species in the Ryukyu Islands, Japan.

Female (paratype, NSMT-Ar 21723). Measurements. Body 2.37 long. Carapace 1.05 long, 1.06 wide, 0.65 high. Eye size and interdistances: AME 0.10, ALE 0.10, PME 0.12, PLE 0.09, AME-AME 0.02, AME-ALE 0.04, PME-PME 0.03, PLE-PLE 0.08. Leg length: leg I: $1.43+0.49+1.01+0.87+0.48=4.28$; leg II: $1.22+0.45$ $+0.85+0.70+0.40=3.62$; leg III: $0.77+0.40+0.52+0.56+0.37=2.62$; leg IV: $0.90+0.36+0.71+0.59+0.38=2.94$. Abdomen 1.58 long, 1.45 wide, 1.49 high.

Carapace oval, as long as wide (CaL/CaW 0.99). Chelicerae with five teeth on promargin with the largest one positioned close to fang base, no teeth on posterior margin (Fig. 6H). Anterior eye row recurved, posterior eye row straight. Macrosetae: leg I: femur p1, patella d1, tibia d2-r1-p1; leg II: patella d1, tibia d2-r1; leg III: patella d1, tibia d1; leg IV: patella d1, tibia d1. Abdomen as in male (AL/AW 1.09).

Coloration and markings (Fig. 6). As in male.
Genitalia (Figs 8, 10). Epigyne a wide plate with an epigynal scape protruding from the posterior margin, epigynal scape spoon-like, and convex ventrally. Anterior


Figure 12. Habitats of theridiosomatid species in Ryukyu Islands $\mathbf{A}$ streamside in dim forest at Iriomote Island $\mathbf{B}$ forest floor of secondary forest at Kume Island $\mathbf{C}$ grassland at Kume Island $\mathbf{D}$ grassland at Yonaguni Island $\mathbf{E}$ crevices on limestone rocky wall at Iriomote Island.
margin of epigynal plate with a pair of dark-colored, sclerotized extensions protruding anteriorly from anterolateral side. Vulva. Spermatheca elliptical, longer laterally juxtaposed at the tip. Copulatory bursae developed. Course of copulatory ducts complicated: originating from copulatory bursae at ventral side, touching each other along the mesial line of the vulva, running posterior-dorsally under spermathecae, bend at a right angle toward laterally, curving anterior-dorsally at lateral side of vulva, forming a coil at lateral side of spermathecae, and then connecting to spermathecae. Fertilization ducts running under copulatory ducts and tip dorsally.

Variations. There is a variation in the color of the abdomen: some individuals with dark grey abdomen, while others with pale yellowish grey abdomen. Course of embolic apophyses also varies among individuals: EA 2 tightly coiled with distal part along with EA 1 and EA 3 running below EA 1 in some individuals including the type specimens, while EA 2 loosely coiled with distal part apart from EA 1 and EA 2 running above EA 1.

Remarks. Males and females are considered the same species because no other candidates were sympatric.

Distribution. Japan (Iriomote Island; Fig. 11).
Habitat. The new species inhabits entrance or insides of limestone caves and crevices of limestone rocky walls (Fig. 12E). Spiders are found in high density at the entrance and twilight zones of humid caves, while sparsely deep inside the dark zone. Its general morphology (pigmented body, eight developed eyes, etc.) and habitat suggest that the species is troglophilic rather than obligate troglobite.

Web morphology. The newly reported species built a conventional orb web with an open hub and two hub loops (Fig. 14A-C). The web was almost vertical, and the tension line extended upward obliquely from the upper side of the hub to the surface of


Figure 13. Web of Theridiosoma and Zoma spiders at Ryukyu Islands, Japan A-C web of Theridiosoma nigrivirgatum sp. nov. D-E web of Theridiosoma alboannulatum $\mathbf{F - H}$ web of Zoma dibaiyin. Abbreviations: RA radial anastomosis SS sticky spirals TL tension line.
the rock (Fig. 14D). The angle of the trapline was approximately $60^{\circ}$ to the horizontal plane. The spider sat upward and held a trapline by both forelegs and grasped radii by legs III, and put legs IV on the hub (Fig. 14E). The web turned conical shape (Fig. 14D), but it seemed to be less distorted than that of Theridiosoma spp. The mean web diameter was $11.6 \times 10.1(\mathrm{~cm}$ vertical $\times$ horizontal $)(n=9)$, number of radii: $17 \pm 2.3(\mathrm{SD})$, and number of sticky spirals: $14.5 \pm 2.5$ (SD) $(n=8)$. As a result of observation of 209 webs in June 2021, it was found that some individuals do not make tension lines. The percentage of webs with a tension line was as follows: female adult, $77 \%$ and juvenile, $33 \%$ in Yutsun-do Cave; female adult, $67 \%$ and juvenile, $45 \%$ in Ôtomi-Daiichi-Do Cave. Juveniles were more likely to build ordinary webs lacking tension lines than adults at both sites. When a web is disturbed by wind, the spider immediately escapes from the web running along the tension line ( $n=22$, see Suppl. material 1). After the escape, some spiders try to hide themselves into limestone rock crevices.

Web-building behavior. $(\boldsymbol{n}=5$ 5). (1) Frames and radii were laid. (2) The spider returned to the hub and made a temporary spiral as a circle. (3) The spider pulled out a sticky line by using only the outer leg IV several times while touching the temporary spiral by the inner leg IV (in T. epeiroides Bösenberg \& Strand, 1906, it draws out a


Figure 14. Web structure and building behavior in Sennin tanikawai sp. nov. (A-G) and Theridiosoma epeiroides $(\mathbf{H}) \mathbf{A}$ orb web, frontal view $\mathbf{B}$ orb web, illustrated $\mathbf{C}$ open hub, frontal view $\mathbf{D}$ orb web, lateral view $\mathbf{E}$ spider holding tension line with forelegs $\mathbf{F}$ process of weaving sticky spirals by $S$. tanikawai sp. nov. $\mathbf{G}$ radial elongation behavior in $S$. tanikawai sp. nov. $\mathbf{H}$ web building processes of Theridiosoma epeiroides. Blue lines indicate elongated portion of radii. Abbreviations: HL hub loops $\mathbf{O H}$ open hub RD radii SS sticky spiral TL tension line TS temporary spiral.
sticky line using both legs IV alternately [Shinkai and Shinkai 1985]). (4) After drawing a sticky line, the spider walked to the frame along a radius holding it by the outer leg IV, shifted it inward, and then attached it to the radius (Fig. 14F; see Suppl. mate-
rial 2). (5) The spider turned to the hub by drawing a new sticky line by the outer leg IV and moved the next radius along the temporary spiral (Fig. 14F). (6) The spider repeated sequences (3) to (5) and laid sticky spirals from outside to inside. (7) After finishing laying the sticky spirals, the temporary spiral was removed. (8) The spider moved near the hub and bit off the radius. (9) After biting off the radius, it changed the direction and attached its spinnerets to the radius and drew out a radius. The elongated radius was attached to the hub (Suppl. material 3). Thus, all radii were elongated (Fig. 14G). (10) The spider returned to the hub, laid two hub loops, and bit out its center. It ingested the ball of threads using both forelegs and digested it. (11) The spider held a tension line, and the web formed a cone. It took approximately one hour to complete the web.

Egg sac. Spherical and dark brown. The size was approximately $3 \times 2$ (mm, height $\times$ width), which was suspended with a long vertical line on the roof of a cave (Fig. 15D, E). This vertical line (pendant line) ranged from 2.5 to 5.3 cm , and the mean was $4.1 \mathrm{~cm}(n=4)$. There was a single attachment point of the egg sac. The junction of the upper end of the egg sac and the lower end of the pendant line resembled a hatch, similar to a cap like structure in Theridiosoma. The lower end of the pendant line was thickened. The egg sac is cleaved at the joint. The egg sac of K. upperyangtzica resembles that of $S$. tanikawai sp. nov. but can be distinguished by the shape of 'cap' (thickened end of the pendant line): almost as long as wide in the former while clearly longer than wide in the latter (Chen 2010: fig. 29 vs. Fig. 15E).

## Sennin coddingtoni (Zhu, Zhang \& Chen, 2001), comb. nov.

Wendilgarda coddingtoni Zhu, Zhang \& Chen, 2001: 2, figs 1-7 (holotype female and paratypes from Liangxi Cave, Dongtang Village, Libo Country, Guizhou Prov., China; not examined).
Karstia coddingtoni: Chen 2010: 4, figs 15-28 (transferred from Wendilgarda).
Remarks. See diagnosis section in S. tanikawai sp. nov.
Distribution. China (Yunnan).

## Discussion

## Habitat and distribution

Although theridiosomatid species prefer dim and moist habitats, microhabitat preferences seem to differ among species. For example, among the Japanese Theridiosoma species, T. epeiroides prefers dim forests, while T. fulvum Suzuki, Serita \& Hiramatsu, 2020 and T. paludicola Suzuki, Serita \& Hiramatsu, 2020 mainly inhabit open and semi-aquatic environments such as wetlands, riverbeds, and pondside (Suzuki et al.


Figure 15. Egg sacs of theridiosomatid species $\mathbf{A}$ egg sac of Theridiosoma nigrivirgatum sp. nov. (cap opened) B egg sac of Theridiosoma dissimulatum $\mathbf{C}$ egg sac of Theridiosoma alboannulatum D-E egg sac of Sennin tanikawai sp. nov.
2020). In the Ryukyu Islands, both T. dissimulatum and Z. dibaiyin are predominantly collected from dim forests, whereas T. nigrivirgatum sp. nov. and T. alboannulatum were frequently found in open habitats such as grasslands (Fig. 12C, D), where the former two species are rarely found. Our survey revealed that T. nigrivirgatum sp. nov. is distributed on the Okinawa Islands, while T. alboannulatum is found on Miyako and Yaeyama Islands (Fig. 11), indicating that the distributional boundary of the two species can be found along the Tokara gap.

Sennin tanikawai sp. nov. showed habitat preferences for limestone caves. Although troglophilic theridiosomatid species have never been reported in other regions of Japan, the congener $S$. coddingtoni comb. nov. is also known to inhabit the insides of limestone caves (Zhu et al. 2001; Chen 2010). Troglophilic species are more common in the neotropical and Chinese genera, for example, Baalzebub in China and Central America, Alaria, Cuacuba, Karstia, and Sinoalaria (Coddington 1986a; Chen 2010; Zhao and Li 2012, 2014; Prete et al. 2018; Prete and Brescovit 2020). Congeners of Sennin gen. nov. are expected to be found in the region between southern China and Iriomote Island, especially Taiwan.

## Web architecture and construction behavior of S. tanikawai sp. nov.

Sennin tanikawai sp. nov. built a conventional orb with an open hub, resembling that of Meta (Araneae: Tetragnathidae). However, modification of the hub after the construction of sticky spirals, temporary spirals as circle, and elongation of radii clearly differentiate the new species from ordinary orb weavers (Tetragnathidae and Araneidae). Elongation of radii after spinning spirals is observed among tiny Araneoids of the families Anapidae, Symphytognathidae, and Mysmenidae (Shinkai and Shinkai 1985; Coddington 1986a,b; Eberhard 1987; Shinkai and Hiramatsu 2000), and is also seen in Theridiosoma epeiroides during the construction of radial anastomosis (as 'ray', Fig. 14H; also see Shinkai and Shinkai 1985). Sennin tanikawai sp. nov. elongates all the radii without anastomosing, and it finally bites the hub as a hole and adds two hub loops (Fig. 14G). A series of radial elongation and hub construction behaviors has never been described in other theridiosomatids. The use of legs during the spinning of sticky spirals also differs between S. tanikawai sp. nov. and T. epeiroides. Theridiosoma epeiroides reels a sticky line out using both forth legs alternately, but S. tanikawai sp. nov. pulls out it by only outer leg IV while touching a temporary spiral as circle by inner leg IV (Fig. 14F). The significance of this difference in behavior of both species in web building is uncertain because details of the behavior are largely unknown in other theridiosomatid spiders. Sennin tanikawai sp. nov. holds the radius away from its body with one leg IV after attaching a sticky line (Fig. 14F, also see Suppl. material 1). This behavior has also been reported in theridiosomatids (e.g., Theridiosoma, Epeirotypus, Ogulnius), and some anapids (Anapis, Anapisona) (Eberhard 1981). The function of this behavior is probably to avoid adhering sticky lines to the radius (Eberhard 1981).

Coddington (1986a) revised Theridiosomatidae mainly from neotropical and neosubtropical regions and discussed their natural histories, especially web morphology and web-building behaviors. The webs of Epeirotypus and Naatlo (Epeirotypinae) are typical orbs with tension lines, lack radial anastomosis, and hubs with two or more persistent hub loops (Table 1; also see Coddington, 1986a: figs 67, 69; Coddington 1986b: fig. 12.7), resembling the web of S. tanikawai sp. nov. Unlike S. tanikawai sp. nov., Epeirotypus species lacks radial elongation (Coddington 1986a). The web of Baalzebub also resembles that of S. tanikawai sp. nov. in appearance, but the former has a single hub loop without a tension line (Coddington 1986a: figs 165, 167), while the latter has two hub loops with a tension line. Baalzebub species adds a single hub loop after spinning sticky spirals, but the process of web building is unknown in detail. In appearance, the web of S. tanikawai sp. nov. is closer to those of Epeirotypus and Naatlo than that of Baalzebub. Based on the morphology of genitalia, S. tanikawai sp. nov. is closely related to Baalzebub. The subfamily Theridiosomatinae, to which Baalzebub belongs, is not closely related to Epeirotypinae in the cladistic analysis (Coddington 1986a). Therefore, the similarity in webs of two subfamilies might be the result of convergence. The multiple hub loops may contribute to the reinforcement of the central region of the web to defuse the tension by the 'slingshot' behavior. There are several differences in the webs and related behaviors of S. tanikawai sp. nov. and

Epeirotypinae: the upward running of the tension line (downward in Epeirotypinae), upward posture of the spider on the hub (dorsal side up position in Epeirotypinae), quick escape behavior along the tension line (not observed in Epeirotypinae), and a 'halfway' slingshot posture (the web is more strongly distorted in Epeirotypinae). These characteristics suggest that the principal function of the tension line in this species may be to escape from any predator, and the function of prey-capture may be secondary. The main predator of $S$. tanikawai sp. nov. is unknown. Bat (Chiroptera) is one of the major predators inhabiting caves, but they are less likely to forage for this species, as all cave-dwelling bats reported on Iriomote Island forage insects outside caves (Okinawa Prefectural Board of Education 1985). Plato is confirmed to have no tension line (Coddington 1986a; Eberhard 2020), while the presence or absence of tension lines has not been examined in most cave-dwelling theridiosomatids: Cuacuba, Karstia, and Sinoalaria (Chen 2010; Zhao and Li 2014; Prete et al. 2018). If these cave-living theridiosomatids also lack a tension line, it would be interesting to know whether or how they perform escape behavior.

Table I. Comparison of habitat and web morphology of theridiosomatid genera of which web morphology were described in published papers. Data source: a Coddington (1986a), ${ }^{\text {b }}$ Eberhard (1986), ${ }^{\text {c }}$ Eberhard (2020), ${ }^{\mathrm{d}}$ Chen (2010), ${ }^{\mathrm{e}}$ Shinkai and Shinkai (1985), ${ }^{\mathrm{f}}$ Hiramatsu (2021). + = present; -= absent, ? = unknown.

| Subfamily | Genus | Habitat | $\begin{aligned} & \text { Orb } \\ & \text { web } \end{aligned}$ | Web shape | Web angle | Tension line | Angle of tension line | Radial anasto-mosis | Open hub | $\begin{aligned} & \text { Hub } \\ & \text { loops } \end{aligned}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Platoninae | Chthonos ${ }^{\text {a }}$ | leaf litter ${ }^{1}$ | - | no web | - | - | - | - | - | - |
|  | Plato ${ }^{\text {a }}$ | caves, dark places | + | loose orb web | vertical or diagonal | - | ? | + | - | - |
| Epeirotypinae | Epeirotypus ${ }^{\text {a, b, c }}$ | shrubs, shaded wet forest | + | concave orb web | vertical or diagonal | + | almost horizontal or downward | - | + | $2-5$ |
|  | Naatlo ${ }^{\text {a }}$ | humid shaded forest | + | concave orb web | vertical or diagonal | + | ? | - | + | 2 |
| Ogulniinae | Ogulnius ${ }^{\text {a }}$, | wet, shaded forest | - | sparse network | - | - | - | - | - | - |
| Theridiosomatinae | Baalzebub ${ }^{\text {a }}$ | interior of hollow logs, under fallen tree, caves | + | ordinary orb web | vertical or diagonal | - | - | - | + | 1 |
|  | Epilineutes ${ }^{2}$ | over stream water | + | conventional orb web | vertical or diagonal | + (rare) | ? | + | - | - |
|  | Karstia ${ }^{\text {d }}$ | limestone caves | + | ? | vertical? | ? | ? | ? | ? | ? |
|  | Sennin gen. nov. | limestone caves | + | conventional orb web | vertical or diagonal | + | almost <br> upward, sometimes horizontal | - | + | 2 |
|  | Theridiosoma ${ }^{\text {a }}$ | wet, shaded forest, etc. | + | concave orb | vertical or diagonal | + | almost horizontal or downward | + | - | - |
|  | Wendilgarda | over stream water or ponds | - | Naruko web | - | - | - | - | - | - |
|  | Zoma ${ }^{\text {f }}$ | wet, shaded forest | + | concave orb | horizontal | + | vertical to the ground | + | - | - |

As the tension line is sporadic throughout theridiosomatids (Coddington 1986a), the origin and function of the tension line can vary (for example, Eberhard 2020: fig. 9.4.(d) (e)). Further morphological and molecular analyses of theridiosomatids, including Sennin gen. nov. species, are expected to elucidate the evolutionary process of the tension line.

## Acknowledgements

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## Supplementary material I

## Video S1

Authors: Yuya Suzuki, Takehisa Hiramatsu, Haruki Tatsuta
Data type: Video (mp4. file).
Explanation note: Sennin tanikawai sp. nov. escaping from the web via a tension line. Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1109.83807.suppl1

## Supplementary material 2

## Video S2

Authors: Yuya Suzuki, Takehisa Hiramatsu, Haruki Tatsuta
Data type: Video (mp4. file).
Explanation note: SS weaving behavior in Sennin tanikawai sp. nov.
Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1109.83807.suppl2

## Supplementary material 3

Video S3
Authors: Yuya Suzuki, Takehisa Hiramatsu, Haruki Tatsuta
Data type: Video (mp4. file).
Explanation note: Radial elongation behavior in Sennin tanikawai sp. nov.
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Link: https://doi.org/10.3897/zookeys.1109.83807.suppl3

# The complete mitochondrial genome of Meloe proscarabaeus (Coleoptera, Meloidae): genome descriptions and phylogenetic inferences 

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#### Abstract

Oil beetles are meloids, which are characterised for their cleptoparasitic habits in bee nests and oily fluid of cantharidin that causes blistering and swelling of the skin. The complete mitochondrial genome of Meloe proscarabaeus is determined using the next-generation sequencing technology and its genomic characteristics are described. The 15,653 -bp long genome is a circular molecule consisting of 13 protein-coding genes (PCG), 22 transport RNA, two ribosomal RNA, and a control region. The A + T bias of the mitochondrial genome is manifested in the complete sequence and the codon usage of protein-coding genes. The genetic distance within and between genera is calculated to confirm the taxonomic status of M. proscarabaeus. The phylogenetic relationships among 15 available meloid taxa are inferred by the maximum likelihood (ML) method based on 13 mitochondrial PCGs. The ML trees resulting from nucleotide and amino acid datasets recover both the monophyly of Meloe and Epicauta and the polyphyly comprising Hycleus and Mylabris. This study provides the first description of a mitochondrial genome belonging to the genus Meloe. The mitochondrial genome sequence and its characteristics are expected to be conducive to future studies on taxonomy, systematics, and molecular phylogenetics of the family Meloidae.


## Keywords

Genome feature, meloid, mitogenome, oil beetle, phylogenetic relationship

## Introduction

The oil beetle Meloe proscarabaeus Linnaeus, 1758 is a characteristic species of the meloid genus Meloe Linnaeus, 1758, which comprises about 155 species in 16 subgenera and is mainly distributed in the Holarctic region (Sánchez-Vialas et al. 2021; Pan and Bologna 2021). Oil beetles are well known for the oily fluid of hemolymph released from their leg joints, which contains the poison cantharidin that causes blistering and swelling of the skin (Muzzi et al. 2020; Du et al. 2021). Additionally, oil beetles are distinguished by their hypermetamorphic development and cleptoparasitic habits in bee nests (Saul-Gershenz and Millar 2006; Saul-Gershenz et al. 2018).

The taxonomy and phylogeny of the genus Meloe are based on morphological characteristics and molecular data (Di Giulio et al. 2002, 2014; Muzzi et al. 2020; Pan and Bologna 2021; Sánchez-Vialas et al. 2021). The genus is considered monophyletic, but more detailed molecular phylogenetic studies are necessary to solve the phylogenetic relationships within the genus. With the simple genetic structure, the high rate of evolution, and the advantage in acquiring methods, the mitochondrial genome has been used widely in many phylogenetic studies of animals (Avise 1994; Cameron 2014; Du et al. 2020). There are 15 meloid species in five genera that have had their complete mitochondrial genomes published in the GenBank database. Previous studies have utilised mitochondrial genome sequences to infer the phylogeny of Meloidae, but without data on the genus Meloe due to the absence of mitochondrial genomes for the genus (Du et al. 2017; Liu et al. 2020).

In this study, we determine the complete mitochondrial genome of $M$. proscarabaeus using next-generation sequencing, and we describe its genomic characteristics. Furthermore, we reconstruct the phylogenetic trees based on the nucleotide and amino acid sequences from all available mitochondrial genomes to analyse the phylogenetic relationships among the family Meloidae. The sequence and phylogenetic inferences of $M$. proscarabaeus mitochondrial genome will be a significant increase in furthering the study on coleopteran mitochondrial genome architecture and phylogenetics.

## Materials and methods

## Sample and genomic DNA extraction

The adults of Meloe proscarabaeus were collected from a hill slope in Dongsheng, Inner Mongolia, China ( $39^{\circ} 45.33^{\prime} \mathrm{N}, 110^{\circ} 01.83^{\prime} \mathrm{E}$ ). The fresh samples were immediately preserved in $100 \%$ ethanol and stored in a $-20^{\circ} \mathrm{C}$ refrigerator. We identified the specimens according to the morphological characters that descried by Pan and Bologna (2021). Total genomic DNA was extracted from a frozen adult using Tianamp Genomic DNA kit following the manufacturer's protocol. The quality of DNA was determined using $1 \%$ agarose gel electrophoresis.

## Next-generation sequencing and genomic assembly

The library was constructed using an Illumina TruSeq Library Preparation kit with an insert size of 250 bp and sequenced using the paired-end strategy on an Illumina HiSeq 2500 platform. A total of 4.4 Gb raw data was yielded with an average read length of 150 bp . The raw data were trimmed and filtered using fastp with parameters of phred quality $\geq 30$ and unqualified percent $<20$ to remove adapters and low-quality reads (Chen et al. 2018). Then the clean data were used to assemble the mitochondrial genome of $M$. proscarabaeus by MITObim v. 1.9.1 with the default settings (Hahn et al. 2013). The mitochondrial genome of Lytta caraganae (GenBank accession number NC_033339.1; Du et al. 2017) was employed as a reference sequence. The assembled sequence was aligned with other meloid mitochondrial genomes described by Du et al. (2017) to ensure the assembling quality.

## Gene annotation and sequence analysis

The complete mitochondrial genome of $M$. proscarabaeus was automatically annotated by the software MitoZ (Meng et al. 2019) and manually compared with other meloid mitochondrial genomes. Of these, PCGs were checked by the identification of open reading frames and aligning with mitochondrial PCGs of other meloids. The annotated mitochondrial genome was analysed its genome characteristics, including nucleotide composition, the composition of skewness, codon usages, and relative synonymous codon usage (RSCU), using MEGA6 (Tamura et al. 2013). All available mitochondrial genomes of 15 meloid species were used to calculate the genetic distances within and between genera in Meloidae. $P$-distances were calculated using MEGA6 with the bootstrap method of 1,000 replications for the variance estimation.

## Phylogenetic analysis

To infer the phylogenetic relationships among the family Meloidae, all available mitochondrial genomes of 15 meloid species, including the $M$. proscarabaeus, were employed to reconstruct the maximum-likelihood (ML) trees with the Tribolium castaneum (GenBank accession number NC_003081.2; Friedrich and Muqim 2003) as the outgroup. According to GenBank annotations, the nucleotide and amino acid sequences of 13 mitochondrial PCGs from each species were extracted and stop codons removed. Orderly combined sequences were aligned using MAFFT with the default settings (Katoh and Standley 2013) and the gaps and ambiguous sites removed to concatenate into consensus sequences including a $10,934 \mathrm{bp}$ nucleotide and a 3,633 amino acid dataset, respectively. IQ-Tree was employed to estimate the best-fit models of partitioning schemes (Nguyen et al. 2015). The nucleotide and the corresponding amino acid datasets were partitioned for 13 genes individually. The best-fitting models of partitioning schemes were selected with the greedy search algorithm, under the Bayesian information criterion. The ML trees of both datasets were also reconstructed using IQ-Tree with 1000 bootstraps to assess the node support.

## Results

## Genome structure and composition

The 5.4 Gb raw data was yielded by the next-generation sequencing with $36,048,728$ reads, and the $4.9 \mathrm{~Gb}(95.06 \%)$ clean data was obtained after filtering. The complete mitochondrial genome of $M$. proscarabaeus was assembled using 1,160,002 reads, and the average depth of coverage was assessed at 7,635.95 X. The mitochondrial genome was annotated and then submitted to the GenBank under the accession number OL840851. The complete mitochondrial genome of $M$. proscarabaeus is $15,653 \mathrm{bp}$ in length, which is of moderate length among mitochondrial genomes within Meloidae (Du et al. 2016, 2017; Zhou et al. 2021).

The complete mitochondrial genome of $M$. proscarabaeus is a circular DNA molecule consisting of 13 protein-coding genes, 22 transport RNAs, two ribosomal RNAs, and a control region (Fig. 1). The length of $r r n L$ and $r r n S$ were determined to be $1,329 \mathrm{bp}$ and 816 bp , respectively, and the control region was $1,013 \mathrm{bp}$ in length. The nucleotide base composition of the mitochondrial genome was $37.5 \%$ A, $31.6 \% \mathrm{~T}, 19.1 \% \mathrm{C}$, and $11.9 \% \mathrm{G}$. The total A + T content was $69.1 \%$, and the AT skew was 0.0854 .

## Protein-coding gene and codon usage

The total length of the 13 mitochondrial protein-coding genes in M. proscarabaeus is $11,127 \mathrm{bp}$, accounting for $71.10 \%$ of the total length of the genome, encoding 3,711 codons in total. All 13 protein-coding genes start using regular initiation codons, including ATT and ATG, and ATA (Table 1), which were commonly used as start codons in insect mitochondrial genomes. Most protein-coding genes terminated with conventional stop codons (such as TAA or TAG), except cox3, nad5, and nad4 stopped with T (Table 1).

The $\mathrm{A}+\mathrm{T}$ bias is also manifested in the codon usage of protein-coding genes (Fig. 2). Relatively synonymous codon usages, excluding stop codons, showed that the third position of synonymous codons always has more frequency with A or T than G or C. Additionally, the first three frequently used codons UUA (Leu2), UCU (Ser2), GUU (Val), and some other frequently used codons, including AUU (Ile), UUU (Phe) AAU (Asn), etc. are comprised of two or three A and/or T nucleotides.

## RNAs and control regions

All 22 transfer RNAs were annotated in the mitochondrial genome of M. proscarabaeus. Their length ranged from the shortest $\operatorname{trn} S$ with 59 bp to the longest trnK with 71 bp (Table 1). The maldistribution of transfer RNAs was also found in the mitochondrial genome. For example, there are two clusters comprising six transfer RNAs ( $\operatorname{trn} A-\operatorname{trn} R-\operatorname{trn} N-\operatorname{trn} S-\operatorname{trn} E-\operatorname{trn} F)$ between nad3 and nad5


Figure I. The circular structure of the mitochondrial genome of Meloe proscarabaeus. The inner circle represents GC content (the ratio of guanine to cytosine), the outer circle represents genetic characteristics, orange represents ribosomal RNA, red represents transport RNA, and green represents protein-coding regions. Genes in the inner circle (on the J chain) are transcribed clockwise, while those outside the circle (on the N chain) are transcribed counter-clockwise.
genes and three transfer RNAs ( $\operatorname{trn} W-\operatorname{trn} C-\operatorname{trn} Y$ ) between nad2 and coxl. The remaining RNAs are dispersed among other genes in a single or double way (Fig. 1). Two ribosomal RNAs mitochondrial genome of $M$. proscarabaeus were aligned with these of other meloid and assigned to the blanks between neighbouring genes. The $\operatorname{rrnL}$ gene was located between $\operatorname{trn} L$ and $\operatorname{trn} V$ with a length of $1,328 \mathrm{bp}$, the $r r n S$ was located between $\operatorname{tnrV}$ and the control region with a length of 815 bp (Table 1). The control region was located between $\operatorname{rrnS}$ and $\operatorname{trnI}$ with $1,015 \mathrm{bp}$ in length (Table 1; Fig. 1).

Table I. Annotation of the Meloe proscarabaeus mitogenome.

| Gene | Location | Inc | Size | Strand | Anticodon | Start codon | Stop codon |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| trnI | 1-66 |  | 66 | J | GAU |  |  |
| $t r n Q$ | 64-132 | -3 | 69 | N | UUG |  |  |
| trnM | 132-201 | -1 | 70 | J | CAU |  |  |
| nad2 | 220-1215 | 18 | 996 | J |  | ATT | TAA |
| trnW | 1214-1279 | -2 | 66 | J | UCA |  |  |
| $t r n C$ | 1279-1341 | -1 | 63 | N | GCA |  |  |
| $t r n Y$ | 1344-1408 | 2 | 65 | N | GUA |  |  |
| cox 1 | 1401-2948 | -8 | 1548 | J |  | ATT | TAA |
| trnL2 | 2944-3007 | -5 | 64 | J | UAA |  |  |
| cox2 | 3008-3695 | 0 | 688 | J |  | ATA | T* |
| $t r n K$ | 3696-3766 | 0 | 71 | J | CUU |  |  |
| $t r n D$ | 3767-3831 | 0 | 65 | J | GUC |  |  |
| atp 8 | 3832-3993 | 0 | 171 | J |  | ATT | TAA |
| atp 6 | 3984-4655 | -10 | 672 | J |  | ATG | TAA |
| cox3 | 4655-5437 | -1 | 783 | J |  | ATG | TAA |
| $t r n G$ | 5441-5503 | 3 | 63 | J | UCC |  |  |
| nad3 | 5504-5857 | 0 | 354 | J |  | ATA | TAG |
| $t r n A$ | 5856-5918 | -2 | 64 | J | UGC |  |  |
| $t r n R$ | 5919-5982 | 0 | 64 | J | UCG |  |  |
| $t r n N$ | 5983-6049 | 0 | 67 | J | GUU |  |  |
| $t r n S$ | 6050-6108 | 0 | 59 | J | UCU |  |  |
| $t r n E$ | 6109-6169 | 0 | 61 | J | UUC |  |  |
| $t r n F$ | 6168-6231 | -2 | 64 | N | GAA |  |  |
| nad5 | 6232-7942 | 0 | 1711 | N |  | ATT | T* |
| trnH | 7943-8006 | 0 | 64 | N | GUG |  |  |
| nad4 | 8007-9339 | 0 | 1333 | N |  | ATG | T* |
| nad 41 | 9333-9620 | -7 | 288 | N |  | ATG | TAA |
| $t r n T$ | 9623-9685 | 2 | 63 | J | UGU |  |  |
| $t r n P$ | 9686-9748 | 0 | 63 | N | UGG |  |  |
| nad6 | 9751-10242 | 2 | 492 | J |  | ATT | TAA |
| cob | 10242-11381 | -1 | 1140 | J |  | ATG | TAA |
| trnS2 | 11380-11447 | -2 | 68 | J | UGA |  |  |
| nad1 | 11465-12415 | 17 | 951 | N |  | ATT | TAG |
| $t r n L$ | 12416-12479 | 0 | 65 | N | UAG |  |  |
| $r m L$ | 12442-13769 | -38 | 1328 | N |  |  |  |
| $t r n V$ | 13757-13825 | -13 | 69 | N | UAC |  |  |
| rrnS | 13824-14638 | -2 | 815 | N |  |  |  |
| CR | 14639-15653 | 0 | 1015 | J |  |  |  |

Inc: intergenic nucleotides, negative values refer to overlapping nucleotides.
$\operatorname{trnL2} 2$ and $\operatorname{trnS2}$ refer to $\operatorname{trnL}$ (UAA) and $\operatorname{trnS}$ (UGA), respectively.
*TAA stop codon is completed by the addition of 3 ' A residues to the mRNA.

## Genetic distances

The genetic distances within and between genera were calculated using the nucleotide and amino acid data of 13 mitochondrial PCGs among 15 meloid taxa. The result from the nucleotide data showed that the $p$-distances within genera for Hycleus, Epicanta, and Mylabris are $0.167,0.173$, and 0.232 , respectively, with an average of 0.191 , while the $p$-distances between genera ranged from 0.234 to 0.281 with an average of 0.258 (Fig. 3). The result from the amino acid data showed that the $p$-distances within


Figure 2. Relative synonymous codon usage (RSCU) in the Meloe proscarabaeus mitochondrial genome. An average of 3,711 codons were analysed, excluding stop codons. Codon families are provided on the x -axis. Leu, Leu2, Ser, and Ser2 indicate trnL1 (CUN), trnL2 (UUR), trnS1 (AGN), and trnS2 (UCN), respectively.
genus of these three genera are $0.124,0.107$, and 0.163 , respectively, with the mean of 0.115 , and the $p$-distance between genera ranging from 0.173 to 0.226 with the mean of 0.187 (Fig. 3). The $p$-distances within genus are significantly lower than between genera from both datasets $(p<0.01)$. The $p$-distance between $M$. proscarabaeus and M. poggii was 0.116 and 0.064 from the nucleotide and the amino acid data, and far less than the corresponding distances between genera.

## Phylogenic relationship

The phylogenetic relationships within the family Meloidae were inferred by using maximum likelihood methods, based on the nucleotide and amino acid data from mitogenomes of 15 meloid taxa. The best-fit partitioning schemes and corresponding substitution models are shown in Table 2. Log likelihoods of consensus trees constructed from 1000 bootstrap trees are $-88,585.1106$ and $-32,785.8864$ for the nucleotide and amino acid data, respectively.

The ML trees resulting from both datasets strongly support the monophyly of Meloe and Epicauta, whereas the genera Hycleus and Mylabris are not monophyletic (Fig. 4). The monophyletic Meloe, including M. proscarabaeus and M. poggii, sisters with Lytta into a branch in both phylogenetic trees, but the branch clusters with Epicauta or Hycleus in the ML tree from nucleotide or amino acid data, respectively (Fig. 4).

Table 2. The best-fit schemes and evolutionary models for two datasets from mitochondrial genomes.

| Dataset | Subset | Best model | Partition names |
| :---: | :---: | :---: | :---: |
| nucleotide | 1 | TIM $+\mathrm{F}+\mathrm{I}+\mathrm{G} 4$ | nad2 |
|  | 2 | $\mathrm{GTR}+\mathrm{F}+\mathrm{I}+\mathrm{G} 4$ | cox 1 |
|  | 3 | $\mathrm{HKY}+\mathrm{F}+\mathrm{I}+\mathrm{G} 4$ | cox2, apt6, nad3, nad4, nad4l, nad6 |
|  | 4 | TPM3+F+G4 | apt8 |
|  | 5 | TIM2+F+I+G4 | cox3, cob |
|  | 6 | TPM3+F+I+G4 | nad5 |
|  | 7 | $\mathrm{K} 3 \mathrm{Pu}+\mathrm{F}+\mathrm{I}+\mathrm{G} 4$ | nad1 |
| Amino acid | 1 | mtMet+G4 | nad2, cox2, apt8, nad3, nad6 |
|  | 2 | mtZOA+G4 | cox 1 |
|  | 3 | mtVer+G4 | apt6 |
|  | 4 | $m \mathrm{mAM}+\mathrm{G} 4$ | cox3 |
|  | 5 | $m t I n v+I+G 4$ | nad5 |
|  | 6 | mtInv+G4 | nad4, nad4l |
|  | 7 | mtMAM $+\mathrm{I}+\mathrm{G} 4$ | cob |
|  | 8 | mtART+G4 | nad1 |



Figure 3. Genetic distance within and between genera. Each boxplot represents the $p$-distance based on the nucleotide and the amino acid datasets from 13 mitochondrial PCGs. The lower horizontal bar represents the smallest observation, the lower edge of the rectangle represents the 25 percentile, the central bar within the rectangle represents the median, the upper edge of the rectangle represents 75 percentile, and the upper horizontal bar represents the largest observation.

## Discussion

The first mitochondrial genome of an oil beetle Meloe proscarabaeus was sequenced and annotated in this study. The gene arrangement and orientation were the same as the common ancestor for the Insecta (Boore et al. 1998; Taanman 1999). The A + T content and AT skew of $M$. proscarabaeus is the moderate level within Meloidae, and the biased usage of A and T nucleotides is also exhibited in mitochondrial PCGs, because


Figure 4. Phylogenetic trees of 14 meloid species based on the nucleotide (a) and amino acid (b) dataset inferred from the maximum likelihood. The numbers abutting branches refer to the bootstrap supports. Tribolium castaneum (Tenebrionidae) was used to root the trees as an outgroup.
the $\mathrm{A}+\mathrm{T}$ bias is a common phenomenon in insect mitochondrial genomes ( Du et al. 2017). Incomplete stop codons commonly exist in mitochondrial genomes of insects (Du et al. 2016; 2017). It is usual that the single T was employed as the stop codon in many insect mitochondrial genomes, and the incomplete stop codon could be functional in polycistronic transcription cleavage and polyadenylation processes (Ojala et al. 1981). The control region was located between $r r n S$ and $t r n I$ with $1,015 \mathrm{bp}$ in length, which is similar to that of other meloid mitochondrial genomes ( Du et al. 2017). It is a noncoding region and a functional region that controls the replication and transcription of the mitochondrial genome and is the biggest region in which variations occurred to affect both sequence and the length of the entire mitochondrial genome (Andrews et al. 1999).

The genetic distance within and between genera was calculated to confirm the taxonomic status of $M$. proscarabaeus. The $p$-distances within genus are significantly lower than between genera from both datasets $(p<0.01)$, but the $p$-distance within Mylabris was a little higher than that between Hycleus and Mylabris (Fig. 3), which may be because of too a few Mylabris taxa and also discussed in some fishes and birds (Ma et al. 2020; Du et al. 2020). The $p$-distances resulting from the nucleotide and the amino acid data between $M$. proscarabaeus and M. poggii were both far less than the corresponding distances between genera. This indicates there exist a significant distinction in genetic distance within and between genera, and subsequently confirm the taxonomic status of M. proscarabaeus.

Phylogenetic analysis within Meloidae recover the monophyly of Meloe and Epicauta, and a polyphyly comprising Hycleus and Mylabris. Within the polyphyly, both trees based on the nucleotide and the amino acid datasets show that $M$. calida Pallas, 1782 clustered with $H$. chodschenticus Ballion, 1878 rather than M. aulica Menetries, 1832 (Fig. 4). The polyphyly of Mylabrini was also recovered by other studies utilising partial genes (mitochondrial 16 S and nuclear ITS2 sequences) and complete mitochondrial PCGs (Bologna et al. 2008; Du et al. 2017). It may be caused by the
high similarity between Hycleus and Mylabris and the inadequate number of taxa available for molecular phylogenetic studies. Previous studies also recovered the different topology (Bologna et al. 2008; Du et al. 2017), which might be limited by the lack of enough taxon sampling. To date, no phylogeny has definitively inferred the phylogenetic relationships within the families. In consideration of the diverse meloid species, the increasingly published information would help achieve more convincing conclusions for the phylogeny of the family.

## Conclusion

The mitochondrial genome of $M$. proscarabaeus was assembled and described. The genome descriptions provide an informative reference for mitochondrial genomes of Meloe beetles. The mitochondrial genome sequence and its characteristics would be beneficial for future studies on taxonomy, molecular phylogenetics, and systematics of meloid insects.

## Acknowledgements

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# Ten new species of genus Tachycines (Orthoptera, Rhaphidophoridae, Aemodogryllinae) from karst caves in Guizhou, China 

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#### Abstract

Ten new karst cave-dwelling rhaphidophorids species of the subgenus Gymnaeta of the genus Tachycines are described from Guizhou Province, southern China; i.e., Tachycines (Gymnaeta) zhongi sp. nov., Tachycines (Gymnaeta) jinniui sp. nov., Tachycines (Gymnaeta) shibenzhangi sp. nov., Tachycines (Gymnaeta) labaidensis sp. nov., Tachycines (Gymnaeta) pinglangus sp. nov., Tachycines (Gymnaeta) shanduensis sp. nov., Tachycines (Gymnaeta) buyii sp. nov., Tachycines (Gymnaeta) portae sp. nov., Tachycines (Gymnaeta) ziyunensis sp.nov., and Tachycines (Gymnaeta) jialiangensis sp. nov. All specimens were collected from Guizhou Plateau.


## Keywords

Aemodogryllini, caves, Guizhou, Gymnaeta, new species, rhaphidophorids

## Introduction

The subgenus Gymnaeta Adelung, 1902 belongs to the tribe Aemodogryllini that is comprised of 67 species predominantly distributed in China, with eight species extending southwards to Southeast Asia: six in Vietnam, one in Myanmar, and one

[^1]in the Philippines (Adelung 1902; Karny 1926, 1929, 1934a, b; Gorochov 1994, 1998, 2001, 2010, 2012; Zhang and Liu 2009; Qin et al. 2019; Cigliano et al. 2021). In addition to the surface species, several species of the subgenus Tachycines (Gymnaeta) inhabit cave habitats (Gorochov 2001; Gorochov et al. 2006; Jiao et al. 2008; Rampini et al. 2008; Feng et al. 2019, 2020; Qin et al. 2019; Zhou and Yang 2020; Zhu et al. 2020).

In recent years, the interest in cave organisms research has increased, and many cave beetles, spiders, cave crickets, ant-loving beetles, cave millipedes, and cave Gesneriaceae species have been reported (Figs 11-13; Gorochov et al. 2006; Lin and Li 2014; Yin et al. 2015; Tian et al. 2016, 2017, 2019; Song et al. 2017; Wang et al. 2017; Liu and Golovatch 2018; Deuve et al. 2020), but there are still many cave species remaining undiscovered, for example, the cave-dwelling rhaphidophorids covered in this study. Twenty-nine species of cave-dwelling rhaphidophorids have been reported in China. Guizhou province is the central area of karst distributions in southern China, where 18 cave-dwelling rhaphidophorids have been found (Gorochov et al. 2006; Rampini et al. 2008; Wen 2018; Feng et al. 2019, 2020; Qin et al. 2019; Zhou and Yang 2020; Zhu et al. 2020; Li et al. 2021; Zhu and Shi 2021). In this work, another ten new species are added to the Chinese fauna based on newly acquired material from Guizhou. This study further reveals the high degree of morphological similarity and cryptic diversity of species in the subgenus Gymnaeta, making it more challenging to delimitate these species using morphological characteristics. Furthermore, we agree with the views presented by Zhu et al. (2020) and confirm that Tachycines (Gymnaeta) aspes (Rampini \& Di Russo, 2008) is a valid species and not a synonym of Tachycines (Gymnaeta) proximus (Gorochov, Rampini \& Di Russo, 2006) according to the varying degrees of reduction of the fastigium vertices and eyes, the higher number of spines on the hind tibia, and the shape of male genitalia. Moreover, we consider that Eutachycines crenatus (Gorochov, Rampini \& Di Russo, 2006) should be transferred to the subgenus Gymnaeta, due to the following genitalic characteristics: median lobe that is shorter than lateral lobe and four lateral lobes that are not sclerotized.

## Materials and methods

All specimens in this article were collected in karst caves by hand, sometimes assisted with a swipe net, and preserved in $75 \%$ ethanol. Morphological characteristics were examined using an Olympus SZ61 stereomicroscope. The male genitalia was preserved in a solution of ethanol and glycerin. Photographs were taken by an Olympus DP22 digital camera and processed with Adobe Photoshop CS6. All specimens are deposited in the Institute of Karst Caves, Guizhou Normal University, Guizhou Province, China (IKCGZNU).

The morphological terms and classification follow Gorochov et al. (2006) and Qin et al. (2019).

## Taxonomy

## Genus Tachycines

## Subgenus Gymnaeta Adelung, 1902

Gymnaeta Adelung, 1902. Annuaire du Musée Zoologique de l'Académie Impériale des Sciences de St. Petersburg 7: 62; Kirby 1906. A Synonymic Catalogue of Orthoptera 2: 125.
Diestrammena (Gymnaeta): Jacobson, 1905[1902-1905]. In: Jacobson and Bianchi, Orthopteroid and Pseudoneuropteroid Insects of Russian Empire and adjacent countries, 329, 352, 434; Gorochov 1994. In: Gorochov and Kireichuk [Eds.], Proceedings of the Zoological Institute of the Russian Academy of Sciences, 257: 49; Otte 2000. Orthoptera Species File 8: 55; Jiao et al. 2008. Zootaxa 1917: 55; Zhang and Liu 2009. Zootaxa 2272: 21.
Tachycines (Gymnaeta): Karny, 1934. Konowia 13 (1-3): 218; Karny 1937. Genera Insectorum 206: 248; Storozhenko 1990. Entomologicheskoe Obozrenie 69(4): 845, 847; Qin et al. 2018. Zootaxa 4374(4): 452; Qin et al. 2019. Zootaxa 4560(2): 274; Feng et al. 2019. Zootaxa 4674(4): 492; Zhou and Yang 2020. ZooKeys 937: 21-29; Zhu et al. 2020. Zootaxa 4809(1): 72.

Type species. Gymnaeta berezovskii Adelung, by subsequent designation; authority: Kirby, W.F. 1906. A Synonymic Catalogue of Orthoptera (Orthoptera Saltatoria, Locustidae vel Acridiidae) 2: i-viii, 1-562.

## Tachycines (Gymnaeta) zhongi sp. nov.

http://zoobank.org/705C74CA-5D8A-46E1-8223-FDA318E27D72
Figs 1A-D, 15
Specimens examined. Holotype, $1{ }^{\top}$, Daxiao Dong, Xinchang township, Liuzhi Special District, 900 m, 2019-VII-28, collected by Jinhua Zhong, Xulin Zhou, Lingzhi Ou, Guang Wang, Benzhang Shi, Juan Liao and Liangfeng An; paratypes, 5ठ, 2 ${ }^{\text {T, }}$ same collection data as for holotype.

Diagnosis. This new species is similar to T. (G.) caudatus (Gorochov et al., 2006) regarding the shape of the female subgenital plate, but the female subgenital plate of the new species has a small triangle on both sides, while the latter is without. Also similar to $T$. (G.) chenhui (Rampini \& Di Russo, 2008) regarding the shape of the male epiphallus, but the new species is smaller, with its body length not exceeding 13 mm , vertex conical tubercles extremely reduced, scarce (Fig. 1D), ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites less developed, forming smaller and shorter projections, hind tarsus keeled ventrally; $T$. (G.) chenhui has a larger body exceeding 13 mm , vertex conical tubercles of intermediate development, ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, forming larger and longer projections, hind tarsus with bristles ventrally.


Figure I. Tachycines (Gymnaeta) zhongi sp. nov., A male genitalia, dorsal view B female subgenital plate in ventral view $\mathbf{C}$ hind tarsus in lateral view $\mathbf{D}$ male live habitus dorsal view.

Description. Male. Body medium and small-sized (Fig. 1D). Eyes slightly reduced, ocelli absent; conical tubercles of vertex reduced. Legs elongate and slender; fore femur approx. 3.1-3.2 times longer than the pronotum, ventrally unarmed, the internal genicular lobe with single small spine, external genicular lobe with single elongate movable spur; ventral side of fore tibiae with one internal spur and two external spurs. Mid femur with an elongate movable spur on the inner and outer genicular lobes, ventrally unarmed; mid tibiae beneath with one internal spur and one external spur. Hind femur without ventral spine, internal genicular lobe with one small spine; hind tibiae dorsally on both sides with 23-25 spines, sparsely arranged. Supra-internal spur of hind tibiae not exceeding ventral apex of hind tarsus. Hind tarsus keeled ventrally and with one dorsal apical spine (Fig. 1C). Small and short ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, but distal ones obtuse and densely ciliated. Cerci extremely long. Male genitalia with H-shaped epiphallus, middle lobe and lateral sclerites of genitalia almost at the same level at the bottom (Fig. 1A).

Female. Other characteristics are similar to the male. Subgenital plate with three lobes, median lobe large and nearly triangular (Fig. 1B). Ovipositor is slightly longer than half the length of hind femur.

Coloration. Body uniformly yellowish brown.
Measurements (mm). Body $\begin{gathered}\text { © } \\ 11.2-11.6, ~\end{gathered} 10.8-12.1$; pronotum ${ }^{\lambda} 3.5, ~ Q 3.8$; fore femur $\delta^{\top} 11.1-11.5, \uparrow 10.8-12.3$; hind femur $\overparen{J}^{\lambda} 18.5-19.3,+18.4-20.0$, ovipositor 10.0-11.2.

Distribution of light zone. Weak light and dark light zones.
Cave adaptation type. Troglobite.
Etymology. The specific epithet refers to the person's last name who led us to collect the specimens.

## Tachycines (Gymnaeta) jinniui sp. nov.

http://zoobank.org/144C1AF3-1041-4D1C-A340-C9C6832F74CC
Figs 2A, 14, 15
Specimens examined. Holotype, 1 q, Jinniu Cave (Fig. 14), Libo County, 2017-X-23, collected by Xulin Zhou, Dongshan Xu, Weicheng Yang; paratype, 1q, same collection data as for holotype.

Diagnosis. The new species is very similar to Tachycines (Gymnaeta) trapezialis Zhou \& Yang, 2020 but differs from the latter by having slightly reduced eyes and the conical tubercles of the vertex intermediately reduce, the hind tibia dorsally on each side has 78-85 spines instead of 54-60 spines.

Description. Female. Body medium sized. Vertex conical tubercles slightly reduced, apex obtuse, ommateum black and well developed. Legs elongate and slender; fore femur approx. 2.5-2.7 times longer than the pronotum, ventrally unarmed, the internal genicular lobe with a small spine, external genicular lobe with one elongate mov-
able spur; ventral side of fore tibiae with one internal spur and two external spurs. Mid femur with an elongate movable spur on internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with two internal spurs and two external spurs. Hind femur without ventral spine, internal genicular lobe without spine; hind tibiae dorsally on both sides with 79-86 spines, arranged in groups. Supra-internal spur of hind tibiae not exceeding ventral apex of hind tarsus. Hind tarsus keeled ventrally, with one dorsal apical spine. Cerci long and slender. Ovipositor shorter than half length of hind femur.

Male. Unknown.
Coloration. Body color uniform, yellowish brown, eyes black.
Measurements (mm). Body $q$ 13.7-16.7; pronotum $q$ 5.1-5.3; fore femur $q$ 13.6-14.1; hind femur +25.2 -26.0; ovipositor 8.4-8.5.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglophile.
Etymology. The new species is named after the collection locality of the specimens (Jinniu cave).


Figure 2. Tachycines (Gymnaeta) jinniui sp. nov., female, subgenital plate in ventral view.

Tachycines (Gymnaeta) shibenzhangi sp. nov.
http://zoobank.org/84644FCC-1440-4FF1-B00E-3D1A70F9450A
Figs 3A-D, 15
Specimens examined. Holotype, $1 \delta^{\lambda}$, Xuehua Cave, Zhonghe Town, Sandu County, 2019-VII-28, collected by Xulin Zhou, Benchang Shi, Changzhen Zheng, Haixia Luo, Gui Liang, Hailian Lan, Panpan Ren and Juan Liao; paratypes, 16§, 15q, same data as the holotype.

Diagnosis. The characteristic of the male genitalia of the new species is distinct from that of other groups: the epiphallus of the male genitalia is semi-circular, and the lateral sclerites sub-elliptical. In addition, the conical tubercles of the vertex are absent, the ommateum are extremely degenerated, the mid tibiae ventrally without spur or spine, the ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, and the distal ones are obtuse and densely ciliated.

Description. Male. Body smaller than the average for the subgenus. Vertex conical tubercles absent, ommateum extremely degenerated, present by narrow stripes with several black facets (some individuals have no black facets and are completely blind). Legs elongate


Figure 3. Tachycines (Gymnaeta) shibenzhangi sp. nov., A male genitalia in dorsal view $\mathbf{B}$ male genitalia in ventral view $\mathbf{C}$ female live habitus in dorsal view $\mathbf{D}$ female subgenital plate in ventral view.
and slender, fore femur approx. 2.6-3.0 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe without spine; fore tibiae beneath with one external spur (sometimes with two external spurs), but without internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobes, ventrally unarmed; mid tibiae ventrally without internal or external spur. Hind femur without spines ventrally; hind tibiae dorsally with 11-18 inner spines and 13-18 outer spines, sparsely arranged. Supra-internal spur of hind tibiae not exceeding ventral apex of hind tarsus. Hind tarsus ventrally with bristles. Ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, but distal ones obtuse and densely ciliated. Epiphallus of male genitalia nearly semi-circular, lateral sclerites sub-elliptical (Fig. 3A, B).

Female. Appearance is similar to the male. Subgenital plate nearly triangular, apical area slightly blunt. Ovipositor is longer than half length of hind femur, dorsal margin smooth, and apical area of ventral margin denticulate.

Coloration. Body color uniform, pale yellow, abdomen slightly transparent and the internal organs are visible.

Measurements (mm). Body $\delta^{\top} 9.4-12.1$, q $11.2-12.5$; pronotum ${ }^{\top} 2.9-3.3$, ㅇ 3.0-3.2; fore femur $\circlearrowleft^{\lambda} 8.1-8.8$, $q 8.2-9.6$; hind femur ${ }^{\top} 13.6-14.6$, ㅇ 13.6-14.8; ovipositor 9.1-10.0.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglobite.
Etymology. The specific epithet refers to the name of the person who provided crucial help in collecting the specimens.

## Tachycines (Gymnaeta) lahaidensis sp. nov.

http://zoobank.org/365EE275-1199-4EC4-A0BA-0CD676277EEE
Figs 4A-D, 15
Specimens examined. Holotype, $1 \delta^{\lambda}$, Lahaide Dong, Pinglang Town, Duyun City, 2015-VII-24, collected by Qing Wen, Dongshan Xu, Yuanchan Yu, Yi Luo, Guang Zhang; paratypes, 6 ${ }^{3}, 8$, same data as the holotype.

Diagnosis. The new species is very similar to Tachycines (Gymnaeta) shibenzhangi sp. nov., as both species have an arc-shaped epiphallus. The difference is that the ventral surface of the hind tarsus keeled in the new species, but differs from the latter in that: lower notch of the epiphallus is rather small, hind tarsus keeled beneath, epiphallus of male nearly n-shaped; ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, apex mucronate without dense cilia.

Description. Male. Body medium sized. Vertex conical tubercles inconspicuous, eyes moderately reduced, approx. $1 / 2$ the size of the normal eye. Legs elongate, slender; fore femur approx. 2.1-2.5 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongated movable spur, internal knee lobe without spine; fore tibiae ventrally with two external spurs and one internal spur. Mid femur with an elongate movable spur on the internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with one external spur and one internal spur. Hind femur without


Figure 4. Tachycines (Gymnaeta) lahaidensis sp. nov., A male body in lateral view B male genitalia in dorsal view $\mathbf{C}$ hind tarsus in lateral view $\mathbf{D}$ female subgenital plate in ventral view.
spine ventrally; hind tibiae dorsally with 27-30 internal spines and 22-26 external spines, sparsely arranged. Supra-internal spur of hind tibiae not exceeding the ventral apex of hind tarsus. Hind tarsus keeled ventrally. Ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distally mucronate without cilia. Epiphallus of male genitalia nearly $n$-shaped, lateral sclerites distinctly long and narrow.

Female. Appearance is similar to the male. The subgenital plate is nearly triangular, its apical area slight obtuse (Fig. 4D). Ovipositor is longer than half length of the hind femur, dorsal margin smooth, apical area of ventral margin denticulate, bent slightly upwards.

Coloration. Body brown, ovipositor wheat.
Measurements (mm). Body đ $10.5-12.3$, $q$ 11.2-12.6; pronotum đ $4.0-4.8$, $q$
 18.1; ovipositor 10.0-11.0.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglobite.
Etymology. The specific epithet refers to the Lahaide cave.

## Tachycines (Gymnaeta) pinglangus sp. nov.

http://zoobank.org/C2D9C063-9A83-464F-8EE2-EDDD561C4745
Figs 5A-D, 15
Specimens examined. Holotype $1 \delta^{\lambda}$, Lagaobieran Dong, Pinglang Town, Duyun City, 2015-VII-25, collected by Qing Wen; paratypes, 11才, 16Q, 2015-VII-25, collected by Qing Wen, Dongshan Xu, Yi Luo, Yuanchan Yu, Guang Zhang.

Diagnosis. The new species is very similar to Tachycines (Gymnaeta) ferecaecus (Gorochov, Rampini \& Di Russo, 2006): both species have a nearly quadrate-shaped epiphallus, but the new species can be distinguished from the latter by the absence of an ommateum (without any black facets), only the base of the ommateum was faintly visible.

Description. Male. Body medium sized in the subgenus (Fig. 5A). Vertex conical tubercles almost absent, ommateum completely reduced (appears to be without any black facets, only ommateum base); Legs elongate and slender; fore femur approx. 2.8-3.2 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe without spine; fore tibiae beneath with one external spur (sometimes with two external spurs), but without internal spur. Mid femur with an elongate movable spur on the internal and external genicular lobes, ventrally unarmed; mid tibiae ventrally with one external spur and one internal spur. Hind femur without spines ventrally; hind tibiae dorsally with $12-15$ internal spines and 12 or 13 external spines, sparsely arranged. Supra-internal spur of hind tibiae shorter than the ventral apex of hind tarsus. Hind tarsus with bristles ventrally. Ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distally obtuse, and densely ciliated. Epiphallus of male genitalia nearly quadrate, median lobe of genitalia with a pair of wide apical lobules, but without distinct lateral sclerites (Fig. 5B).

Female. Appearance is similar to the male. The subgenital plate is nearly triangular, and the apical area slightly obtuse. Ovipositor is longer than half of the hind femur length, brown, dorsal margin smooth, apical area of ventral margin denticulate, bent slightly upwards.

Coloration. Body yellowish brown, ovipositor wheat.


Figure 5. Tachycines (Gymnaeta) pinglangus sp. nov., A male body in lateral view B male genitalia in dorsal view $\mathbf{C}$ female terminal in lateral view $\mathbf{D}$ female subgenital plate in ventral view.

Measurements (mm). Body ô $9.5-13.0$, $q 8.2-12.0$; pronotum § $3.2-3.8, q$ $3.2-3.4$; fore femur $\overbrace{}^{\Uparrow} 10.2-10.50$, $\uparrow 10.2-10.4$; hind femur ${ }^{\top} 13.2-14.7$, , $13.2-$ 14.6; ovipositor 8.0-10.4.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglobite.
Etymology. The specific epithet refers to the locality where the type specimens were collected.

## Tachycines (Gymnaeta) shanduensis sp. nov.

http://zoobank.org/ED670C87-E4D5-48C1-A93C-83949F4F8FC3
Figs 6A-D, 15

Specimens examined. Holotype $1 \delta^{\lambda}$, Shuilong Cave, Sandu County, 2019-VII-22, collected by Xulin Zhou, Benchang Shi, Changzhen Zheng, Gui Liang, Haixia Luo, Hailian Lan, Juan Liao; paratypes, $6 \widehat{\lambda}, 8 q$, same data as holotype.

Diagnosis. This species is rather similar to Tachycines (Gymnaeta) solida (Gorochov, Rampini \& Di Russso, 2006) and Tachycines (Gymnaeta) tongrenus Feng, Huang \& Luo, 2020, but the male epiphallus of the new species has a distal shallow notch clearly wider than the upper notch, the median process of the male genitalia is significantly longer than the lateral sclerites, hind tibiae dorsally on both sides with 34-46 spines, hind tarsus keeled beneath; however, in Tachycines (Gymnaeta) solida, the male epiphallus has the upper and lower notches almost the same size, hind tibiae dorsally on both sides with 62-69 spines; in Tachycines (Gymnaeta) tongrenus, the hind tibia dorsally with 48-49 inner spines and 54-56 outer spines, hind tarsus with bristles ventrally.

Description. Male. Body rather large for this subgenus. Vertex conical tubercles are well developed, bisected from the base; ommateum is black and well developed (Fig. 6A). Legs elongate and slender; fore femur approx. 1.9-2.1 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe with a small spine; fore tibiae beneath with two external spurs and one internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with one external spur and one internal spur. Hind femur without spines ventrally; hind tibiae dorsally with $34-43$ internal spines and 38-46 external spines, arranged in groups. Supra-internal spur of hind tibiae shorter than the dorsal apex of hind tarsus. Hind tarsus keeled ventrally, with one dorsal apical spine. Epiphallus of male genitalia nearly H-shaped, lateral sclerites distinctly long and narrow, upper notch rather smaller than lower notch.

Female. Appearance is similar to male (Fig. 6B). Subgenital plate with three lobes, median lobe large, triangular, and apical area sharp (Fig. 6C). Ovipositor is shorter than half of the hind femur length, dorsal margin smooth, apical area of ventral margin denticulate.

Coloration. Body dark brown, mixed with tawny stripes, hind femur with brown diagonal stripe.


Figure 6. Tachycines (Gymnaeta) shanduensis sp. nov., A male live habitus in dorsal view B female live habitus in dorsal view $\mathbf{C}$ female subgenital plate in ventral view $\mathbf{D}$ male genitalia in dorsal view.

 30.7; ovipositor 12.8-13.5.

Distribution of light zone. Light zone, weak light zone, and dark light zone.
Cave adaptation type. Troglophile.
Etymology. The name of the new species refers to the type locality.

## Tachycines (Gymnaeta) buyii sp. nov.

http://zoobank.org/935DF22A-FD7C-4C2C-80FF-04743039FDE2
Figs 7A-C, 15
Specimens examined. Holotype $1 \delta^{\lambda}$, Sanjiaoshan Cave, Ziyun County, 2019-X-2, collected by Xulin Zhou, Haixia Luo, Panpan Ren, Meizhen Deng and Suqin Zhao; paratypes, ${ }^{\top} 15, ~, ~ 18$, same data as holotype.

Diagnosis. This new species is rather similar to Tachycines (Gymnaeta) solida (Gorochov, Rampini \& Di Russo, 2006), but differs as follows: the new species epiphallus of male genitalia with upper notch smaller and shallower than lower notch, hind tarsus ventrally with bristles; in $T$. (G.) solida the epiphallus of the male genitalia with upper notch and lower notch almost the same size, hind tarsus keeled ventrally.

Description. Male. Body rather small for this subgenus. Vertex conical tubercles well-developed, bisected from the base; ommateum black and well developed. Legs elongate and slender; fore femur 2.0-2.1 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe without spine; fore tibiae beneath with two external spurs and one internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with one external spur and one internal spur. Hind femur without spines ventrally; hind tibiae dorsally with 35-44 internal spines and 36-46 external spines, arranged in groups. Supra-internal spur of hind tibiae not exceeding the ventral apex of hind tarsus. Hind tarsus ventrally with bristles. Epiphallus of male genitalia nearly H-shaped, lower notch rather deeper than upper notch (Fig. 7A, B).

Female. Appearance is similar to the male. Subgenital plate with three lobes, median lobe large and triangular. Ovipositor is slightly shorter than half of the hind femur length, dorsal margin smooth, apical area of ventral margin denticulate.

Coloration. Body brown, mixed with dark brown patches. Hind femur with brown stripe, and dark brown rings located at $2 / 3$ of the length.

Measurements (mm). Body © $10.5-11.3, ~ Q 10.8-11.5$; pronotum ${ }^{\top} 3.9-4.5$,
 ovipositor 7.2-8.6.

Distribution of light zone. Light zone, weak light zone, and dark light zone.
Cave adaptation type. Troglophiles.
Etymology. The specific epithet refers to the native BuYi people who have lived in southern Guizhou for generations.


Figure 7. Tachycines (Gymnaeta) buyii sp. nov., A male genitalia in dorsal view B male genitalia in ventral view $\mathbf{C}$ female subgenital plate in ventral view.

## Tachycines (Gymnaeta) portae sp. nov.

http://zoobank.org/6A1D6FF0-A4CC-4921-9E0B-0E6EDFC09E7E
Figs 8A-C, 15
Specimens examined. Holotype $1{ }^{\top}$, Niujingchongzi Dong Weining County, 2019-VII-17, collected by Xulin Zhou, Lingzhi Ou, Guang Wang, Rongxiang Su Benzhang Shi, Juan Liao and Liangfeng An. paratypes, 4 ${ }^{\top}, 2 q$, same data as holotype.

Diagnosis. The new species is most closely related to Tachycines (Gymnaeta) buyii sp. nov., but it can be distinguished from the latter by the structure of epiphallus, and hind tarsus keeled ventrally.

Description. Male. Body rather small for this subgenus. Vertex conical tubercles well developed, bisected from the base; ommateum black and well developed. Legs elongate and slender; fore femur 1.8-1.9 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe with a small spine; fore tibiae beneath with two external spurs and one internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with one external spur and one internal spur. Hind femur without spines ventrally; hind tibiae dorsally with 65-81 internal spines and 63-81 external spines, arranged in groups. Supra-internal spur of hind tibiae not exceeding the ventral apex of hind tarsus. Hind tarsus keeled ventrally. Epiphallus of male genitalia nearly door-shaped, lower notch rather deeper than upper notch (Fig. 8A, B).

Female. Appearance is similar to the male. Subgenital plate with three lobes, median lobe large triangular (Fig. 8C); ovipositor is slightly longer than half of the hind femur length, dorsal margin smooth, apical area of ventral margin denticulate.

Coloration. Body brown, mixed with brown patches; hind femur with brown stripe.
Measurements (mm). Body ${ }^{\top} 6.0-7.0$, $q 6.3-7.0$; pronotum ${ }^{\top} 4.0-4.5$, $\uparrow 3.8-$ 4.5 ; fore femur $\widehat{\top}^{\top} 6.8-8.0, q 6.5-7.5$; hind femur ${ }^{\top} 10.0-11.5$,, $10.5-11.5$; ovipositor 5.5-6.0.

Distribution of light zone. Light and weak light zone.
Cave adaptation type. Troglophile.
Etymology. The specific epithet refers to the shape of epiphallus, the Latin word porta meaning door.


Figure 8. Tachycines (Gymnaeta) portae sp. nov., A male genitalia in dorsal view $\mathbf{B}$ male genitalia in ventral view $\mathbf{C}$ female subgenital plate in ventral view.

Tachycines (Gymnaeta) ziyunensis sp. nov.
http://zoobank.org/53F85BED-D0B8-46F5-AC23-0DE66CF2C3F1
Figs 9A-E, 15
Specimens examined. Holotype $1{ }^{\AA}$, Sanjiaoshan cave, Ziyun County, 2019-X-2, collected by Xulin Zhou, Haixia Luo, Panpan Ren, Meizhen Deng and Suqin Zhao, paratypes $15 \delta^{\lambda}, 38$, same data as holotype.

Diagnosis. The new species is rather similar to Tachycines (Gymnaeta) shibenzhangi sp. nov., it can easily be distinguished by the eyes moderately reduced, ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distal mucronate without ciliated;
but the latter of eyes extremely reduce, ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distal obtuse and densely ciliated.

Description. Male. Body medium size (Fig. 9D, E). Vertex conical tubercles almost absent, ommateum moderately reduced. Legs elongate and slender; fore femur approx. 2.5-3.1 times longer than the pronotum, ventrally unarmed, external genicular lobe with one elongate movable spur, internal knee lobe with a small spine; fore tibiae beneath with one external spur and one internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobe, ventrally unarmed; mid tibiae beneath without internal and external spur. Hind femur without spines ventrally; hind tibiae dorsally with 9-15 internal spines and 9-13 external spines, sparsely arranged, supra-internal spur of hind tibiae not exceeding the ventral apex of hind tarsus. Hind tarsus ventrally with bristles (Fig.9C). ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distal mucronate without cilia. Epiphallus of male genitalia nearly semi-circular, lateral sclerites sub-elliptical; median process of male genitalia with semi-sclerotized lobules at apical part and divided into two lobes, significantly longer than lateral sclerites (Fig. 9A, B).

Female. Appearance is similar to the male. Subgenital plate with three lobes, median lobe large, triangular; ovipositor is slightly longer than half of the hind femur length.

Coloration. The body color is yellowish, face without dark brown stripes, uniformly pale yellow, ventral conical projections of abdominal sternites shiny white. Ovipositor is brownish yellow.

Measurements (mm). Body đ $11.4-12.8, q 12.2-13.2$, pronotum đ $3.5-4.3$, $q$ 3.9-4.3, fore femur ơ $11.0-12.1$, $+10.2-10.8$, hind femur ${ }^{\top} 17.9-19.2$, $+16.9-$ 17.9; ovipositor 8.9-10.3.


Figure 9. Tachycines (Gymnaeta) ziyunensis sp. nov., A male genitalia in dorsal view B male genitalia in ventral view $\mathbf{C}$ hind tarsus in lateral view $\mathbf{D}$ male live habitus in dorsal view $\mathbf{E}$ nymph of male, living habitus in dorsal view.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglobite.
Etymology. The name of the new species refers to the type locality.

## Tachycines (Gymnaeta) jialiangensis sp. nov.

http://zoobank.org/68C1DDDD-C8AD-432B-9BBD-85D4610A79A5
Figs 10A-C, 15

Specimens examined. Holotype $1 \delta^{\lambda}$, Lajilou Cave, Jialiang Town, Libo County, 2017-X-23, collected by Xulin Zhou, Dongshan Xu, Weicheng Yang; paratypes, 4 ${ }^{\lambda}$, 2 中, same data as holotype.

Diagnosis. This species is similar to T. (G.) ziyunensis sp. nov. but differs in that the hind tarsus ventrally bears bristles in $T$. (G.) ziyunensis sp. nov., and by the hind tibiae armed with 9-15 spines on both sides; however, the hind tarsus is keeled ventrally in $T$. (G.) jialiangensis sp. nov., and the hind tibiae are provided with 17-25 spines on both sides.

Description. Male. Body medium in size. Vertex conical tubercles almost absent, ommateum moderately reduced. Legs elongate and slender; fore femur 2.9-3.0 times longer than the pronotum, ventrally unarmed, external genicular lobe with single elongate movable spur, internal knee lobe without spine; fore tibiae beneath with two external spurs and one internal spur. Mid femur with an elongate movable spur on both internal and external genicular lobes, ventrally unarmed; mid tibiae beneath with an external spur and without internal spur. Hind femur without spines ventrally; hind tibiae dorsally with 18-25 internal spines and 17 or 18 external spines, sparsely arranged. Supra-internal spur of hind tibiae


Figure I0. Tachycines (Gymnaeta) jialiangensis sp. nov., A male genitalia in dorsal view $\mathbf{B}$ male genitalia in ventral view $\mathbf{C}$ hind tarsus in lateral view.
not exceeding ventral apex of hind tarsus. Hind tarsus keeled ventrally (Fig. 10C). ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites developed, distally mucronate without cilia. Epiphallus of male genitalia nearly semi-circular, lateral sclerites sub-elliptical, median process of male genitalia with semi-sclerotized lobules at apical part and divided into two lobes, significantly longer than lateral sclerites (Fig. 10A, B).

Female. Appearance is similar to the male. Subgenital plate with three lobes, median lobe large, triangular; ovipositor slightly shorter than half of the hind femur length, dorsal margin smooth, apical area of ventral margin denticulate.

Coloration. Body color uniform, pale yellow; face without dark brown stripes; ventral conical projections of $3^{\text {rd }}-8^{\text {th }}$ abdominal sternites shiny white.

Measurements (mm). Body § $11.8-12.0$, $\uparrow$ 11.2-13.0; pronotum § 3.5-3.6, Q 3.4-4.2; fore femur $\widehat{O}^{\top} 10.3-10.6$,, $10.4-13.2$; hind femur ${ }^{\top} 17.5-18.4$,, $17.1-$ 21.66, ovipositor 8.3-9.9.

Distribution of light zone. Dark light zone.
Cave adaptation type. Troglobite.
Etymology. The name of the new species refers to the type locality.

## Discussion

Environmental conditions typical of the habitat deep within caves include the complete absence of light, a stable and usually very high humidity (> 95\%), relatively low levels of available nutrients, and a nearly constant temperature (Taylor 2004). Cave organisms have been evolving in unusual and fascinating habitats, and the nature of these seems to be the loss of some structures (generally of eyes and pigment), such as in cavefish, cave-dwelling rhaphidophorids, and cave-adapted ground beetles. These organisms successfully navigate within such environments, find and capture food, identify and reproduce with conspecifics, and compete with one another for resources, all in the absence of visual cues. During the adaptive evolution in a cave environment, these cave-dwellers have evolved morphological, physiological, and behavioral modifications, many of which could promote their success lives in constant darkness (William et al. 2019).

There are two subfamilies of Rhaphidophoridae, Aemodogryllinae and Rhaphidophorinae, occurring in East Asia. Among them, species of the subgenus Gymnaeta are widely adapted to cave ecosystems, and belong to the Aemodogryllinae. The distribution of cave species of Gymnaeta is consistent with that of karst landforms, while the surface species are widely distributed in East Asia, ranging from forests to swamps, deserts, and the Tibet Plateau. According to our observation of many years, the surfacedwelling species of Gymnaeta usually demonstrate the following characteristics: a larger body size, darker body coloration, the ommateum and conical tubercles on the vertex well-developed, abdominal sternites without ventral projections, ventral spurs of the
fore tibiae and mid tibiae well developed, the dorsal spines of hind tibiae dark and arranged in dense clusters, and the hind femur muscles well-developed, and hence these species are physically agile and good jumpers. Surface-dwelling species of Gymnaeta are widely distributed throughout China.


Figure II. The ecdysial Tachycines (Gymnaeta) solida (Gorochov, Rampini \& Di Russo, 2006). A in cave Donggou Dong B, C in cave Yu Dong.


Figure 12. Other species found in the caves $\mathbf{A}$ Rhinolophus sp. (Chiroptera, Rhinolophidae) in cave Donggou Dong B a running earwig in cave Xuehua Dong C Glyphiulus sp. in cave Xuehua Dong D a Polydesmida in cave Da Dong E an Odorrana sp. in cave Wuming Dong near Banzhu town $\mathbf{F}$ an Epanerchodus millipede in cave Sanjiaoshan Dong $\mathbf{G}$ a Tachycines (Gymnaeta) sp. nymph feeding on rat droppings from Donggou Dong.


Figure 13. A Entrance of Ban Dong B entrance of Mawan Dong from Fuyan town C Entrance of Mawan Dong from Lengjiagou D entrance of Donggou Dong E Primulina eburnea in entrance of Ban Dong $\mathbf{F}$ two collectors in a small humid passageway in cave Sanjiaoshan Dong, showing the habitat where Tachycines (Gymnaeta) ziyunensis sp. nov. was collected.

In contrast, the cave-adapted species of the subgenus have the ommateum and conical tubercles of the vertex reduced, the body appears thinner and smaller, the legs appear more slender, the ventral spurs of fore tibiae and mid tibiae are reduced and sometimes absent, the dorsal spines of hind tibiae are not pigmented and are sparse, the hind femur muscles are not obvious, and the ventral projections of abdominal sternites are developed. The cave-dwelling Gymnaeta are not good at jumping, as they have less developed legs muscles, leading them to almost walking on the ground like cave beetles when in danger. They are mainly distributed in karst areas of south China. We think that these morphological changes have occurred through adaptive evolution to cave environments. In addition, we consider the presence or absence of cilia on the distal of abdominal sternites projections in cave-adapted species of the subgenus Gymnaeta an important taxonomic characteristic. While these species in Chinese karst caves may be morphologically similar to surface-dwelling species, we suggest they may be genetically distinct, potentially representing different subspecies or lineages and we expect our future work to expand on these theories.


Figure 14. Map of Cave Jinniu Dong, type locality of Tachycines (Gymnaeta) jinniui sp. nov. The arrowhead indicates where in the cave system the rhaphidophorids were found.


Figure 15. Distribution map for new species of subgenus Gymnaeta from Guizhou mentioned in this study.

## Conservation suggestions for China＇s karst cave habitats and species

Caves and their associated ecosystems（mostly karst）represent resources of great val－ ue．These values can be grouped into three general clusters：ecological－scientific，eco－ nomic，and cultural．Cave habitats and species are attracting increasing interest and concern among conservationists，cavers，and speleobiologists，and for good reason． Most troglobionts are highly restricted geographically and often are numerically rare， making them vulnerable to even relatively minor disturbances．As cave organisms are an essential part of biodiversity，it is hoped that we can raise public awareness and take adequate measures to protect karst cave ecosystems．

In China，we have a rich diversity of troglobionts，but the biodiversity of Karst caves in China is under serious threat．We should choose high value and high priority caves，which need emergency attention in relation to protection，manage－ ment，and conservation actions in the karst region of China．Such a fact does not exclude the need for the conservation of the other caves that should require atten－ tion，management，and／or conservation plans coordinated by the environmental supervisory agencies．

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# Revision of the water scavenger beetle genus Notionotus Spangler, 1972 in the Neotropical Region (Coleoptera, Hydrophilidae, Enochrinae) 

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#### Abstract

The Neotropical species of the water scavenger beetle genus Notionotus Spangler, 1972 are revised using an integrative taxonomic approach combining morphology with DNA sequence data from two genes. Support exists for four putative species groups into which 18 species are placed, including twelve that are described here as new: $N$. bicolor sp. nov. (Suriname), $N$. bifidus sp. nov. (Venezuela), $N$. brunbadius sp. nov. (Brazil), N. garciae sp. nov. (Brazil), N. giraldoi sp. nov. (Brazil), N. insignitus sp. nov. (Venezuela), N. juma sp. nov. (Brazil), N. parvus sp. nov. (Suriname), N. patamona sp. nov. (Guyana), N. peruensis sp. nov. (Peru), N. retusus sp. nov. (Guyana), and $N$. vatius sp. nov. (Brazil). Four new synonymies are created: N. shorti Queney syn. nov. is found to be conspecific with $N$. dilucidus Queney; N. edibethae García syn. nov., $N$. nucleus Perkins syn. nov., and $N$. perijanus García syn. nov. are found to be conspecific with $N$. tricarinatus Perkins. New records are provided for all previously described species except $N$. mexicanus Perkins. Within the Neotropical region, the range of the genus is greatly expanded and now known from as far south as Bolivia and the Brazilian state of Mato Grosso do Sul. While a few species are found in hygropetric habitats, most are associated with the margins of forested streams. Genitalia and habitus images are provided for nearly all species, as well as a key to the four species groups.


[^2]
#### Abstract

Resumen Las especies de escarabajos acuáticos detritívoros neotropicales del género Notionotus Spangler, 1972 son revisadas usando taxonomía integrativa, combinando morfología con datos de secuencias de ADN para dos genes. Se encontró soporte para cuatro grupos de especies conformados por 18 especies, incluyendo 12 que son aquí descritas como nuevas: $N$. bicolor sp. nov. (Suriname), $N$. bifidus sp. nov. (Venezuela), N. brunbadius sp. nov. (Brazil), N. garciae sp. nov. (Brazil), N. giraldoi sp. nov. (Brazil), N. insignitus sp. nov. (Venezuela), N. juma sp. nov. (Brazil), N. parvus sp. nov. (Suriname), N. patamona sp. nov. (Guyana), N. peruensis sp. nov. (Peru), N. retusus sp. nov. (Guyana), N. vatius sp. nov. (Brazil). Se sinonimizan cuatro especies: $N$. shorti Queney syn. nov. se considera conespecífico con $N$. dilucidus Queney; N. edibethae García syn. nov., N. nucleus Perkins syn. nov., y N. perijanus García syn. nov. son conespecíficos con $N$. tricarinatus Perkins. Nuevos registros son provistos para todas las especies previamente descritas excepto para $N$. mexicanus Perkins. Hasta el momento, en la región neotropical, el rango de distribución del género es ampliamente expandido, se conoce desde el Sur de Bolivia hasta el estado brasileño de Mato Grosso do Sul. Si bien algunas especies son encontradas en hábitats higropétricos, la mayoría de ellas están asociadas a las orillas de arroyos boscosos. Se proveen imágenes del hábito dorsal y genitalia para la mayoría de las especies, al igual que la clave para los cuatro grupos de especies.


## Keywords

aquatic insects, integrative taxonomy, new species, South America

## Introduction

The genus Notionotus Spangler, 1972 is a group of small to minute water scavenger beetles that occur in streams and seepages in the tropics of the New World and Southeast Asia. The original concept of Notionotus was based on two new species from the Venezuelan Andes (Spangler 1972). In the Neotropics, its range was soon expanded north to Mexico, with three species described by Perkins (1979) from Guatemala, Mexico and Panama, and subsequent additional species from Venezuela (García 2000). Most recently, three species were described from French Guiana and Guyana (Queney 2010) with additional new records being provided in a series of biotic surveys in Suriname and Guyana (Short and Kadosoe 2011; Short 2013; Short et al. 2017, 2018). Notionotus has also been reported from the Old World tropics, with seven species described from southeast India and Southeast Asia (Hebauer 2001, 2003) and southern China (Jia and Short 2011). Currently there are 17 described species, including ten from the Neotropics and seven from tropical Asia.

During the last two decades, extensive fieldwork for aquatic beetles in the Neotropics has resulted in the collection of many new specimens of Notionotus, including some that represent new species. We here use an integrative approach, combining adult morphological data with DNA sequences from two gene loci, to revise the Neotropical species of the genus. This revision provides a taxonomic foundation not only for future descriptive work on the genus, but also for evolutionary studies on the diversity and biogeography of the Neotropical region.

## Materials and methods

## Molecular methods

We amplified and sequenced the mitochondrial gene COI and the nuclear ribosomal gene 28 S for 56 specimens of Notionotus, including 55 specimens from the Neotropical region and one unidentified specimen from India which we used as an outgroup to root the tree. We sampled specimens from most localities for which we had appropriate material, including a broad geographic sampling of $N$. tricarinatus and $N$. dilucidus as these two species had large ranges with some observed variation in the aedeagus. All specimens were preserved as frozen tissue samples since collection with the exception of the specimen from Costa Rica (SLE2381) which was pinned. Molecular extraction and sequencing methods follow those of Kohlenberg and Short (2017). Resulting DNA sequences were assembled and edited in Geneious R 8.0.5 (Biomatters, http:// www.geneious.com/). All sequences are deposited in GenBank (see Table 1 for accession numbers). We used IQ-TREE 1.4.4 (Nguyen et al. 2015) to infer phylogenetic relationships. Each gene was placed in its own partition. The optimal models of substitution for each partition were selected using the Auto function in IQ-TREE 1.4.4. In order to assess nodal support, we performed 1000 ultrafast bootstrap replicates (Minh et al. 2013).

Table I. List of DNA voucher specimens and GenBank accession numbers used in this study. "N/A" indicates the gene fragment was not successfully amplified or sequenced.

| Taxon | Extraction | Locality | COI Accession | 28S Accession |
| :---: | :---: | :---: | :---: | :---: |
| N. bicolor | SLE1810 | Suriname: Sipaliwini: Kabalebo Nature Resort | ON239446 | ON243733 |
| N. bicolor | SLE2120 | Suriname: Sipaliwini: Kabalebo Nature Resort | ON23944 | /A |
| N. bifidus | SLE1113 | Venezuela: Amazonas: Tobogán de la Selva | ON239437 | ON243725 |
| N. bifidus | SLE2369 | Venezuela: Amazonas: Tobogán de la Selva | ON23943 | ON243726 |
| N. brunbadius | SLE1553 | Brazil: Amazonas: Ducke Reserve | ON239441 | ON243731 |
| N. brunbadius | SLE2102 | Brazil: Amazonas: Ducke Reserve | ON239442 | ON243732 |
| N. dilucidus | SLE0505 | Suriname: Brokopondo: Brownsberg Nature Park | ON239422 | N/A |
| N. dilucidus | SLE0506 | Suriname: Brokopondo: Brownsberg Nature Park | ON239420 | ON243688 |
| N. dilucidus | SLE1799 | Suriname: Sipaliwini: Kabalebo Nature Resort | ON239418 | ON243695 |
| N. dilucidus | SLE1811 | Suriname: Sipaliwini: Kabalebo Nature Resort | ON239419 | ON243696 |
| N. dilucidus | SLE2107 | Guyana: Region IX: Kusad Mountain | ON239411 | ON243702 |
| N. dilucidus | SLE2108 | Guyana: Region VI: Upper Berbice | ON239417 | ON243692 |
| N. dilucidus | SLE2113 | French Guiana: Crique Eau Chire | ON239406 | ON243693 |
| N. dilucidus | SLE2114 | Suriname: Sipaliwini: Sipaliwini Savanna Reserve | ON239413 | ON243703 |
| N. dilucidus | SLE2121 | Suriname: Brokopondo: Brownsberg Nature Park | ON239421 | ON243694 |
| N. dilucidus | SLE2330 | Brazil: Roraima: ca. 13 km NE of Caroebe | N/A | ON243704 |
| N. dilucidus | SLE2335 | Guyana: Region 9: Kusad Mountain | ON239412 | ON243705 |
| N. dilucidus | SLE2339 | French Guiana: St. Laurent du Maroni (ca. 15 km SW ) | ON239415 | ON243690 |
| N. dilucidus | SLE2366 | Venezuela: Amazonas: Tobogán de la Selva | ON239404 | ON243687 |
| N. dilucidus | SLE2368 | Guyana: Region 8: 7 km NW Chenapau | ON239409 | ON243699 |
| N. dilucidus | SLE2374 | Brazil: Roraima: Rio Cocal, near Tepequem | ON239408 | ON243698 |
| N. dilucidus | SLE2375 | Suriname: Sipaliwini: Wehepai | ON239416 | ON243706 |


| Taxon | Extraction | Locality | COI Accession | 28S Accession |
| :---: | :---: | :---: | :---: | :---: |
| N. dilucidus | SLE2377 | Brazil: Roraima: Serra do Tepequém | ON239410 | ON243700 |
| $N$. dilucidus | SLE2378 | Venezuela: Bolívar: Piedra de la Virgen | ON239405 | ON243701 |
| N. dilucidus | SLE2380 | Guyana: Region IX: Parabara | ON239414 | ON243707 |
| $N$. dilucidus | SLE2383 | French Guiana: Carbet ONF Grillon | ON239407 | ON243697 |
| N. dilucidus | SLE2389 | Suriname: Sipaliwini: Raleighvallen | ON239423 | ON243689 |
| N. dilucidus | SLE2394 | Suriname: Sipaliwini: Raleighvallen | N/A | ON243691 |
| N. garciae | SLE1900 | Brazil: Amazonas: Pres. Fig. (ca. 57 km E) | ON239440 | ON243730 |
| N. giraldoi | SLE2088 | Brazil: Rondonia: Vale do Cachoeiras | ON239428 | ON243718 |
| N. giraldoi | SLE2332 | Brazil: Rondonia: Ji-Parana ( 27 km SW ) | ON239427 | ON243719 |
| N. giraldoi | SLE2334 | Brazil: Rondonia: Ji-Parana ( 27 km SW ) | ON239429 | ON243720 |
| N. insignitus | SLE1115 | Venezuela: Bolívar: La Escalera | ON239443 | ON243734 |
| N. juma | SLE1269 | Brazil: Amazonas: Novo Airão | ON239435 | ON243727 |
| N. juma | SLE2100 | Brazil: Amazonas: Ducke Reserve | ON239436 | ON243728 |
| N. liparus | SLE2111 | Venezuela: Aragua: Henri Pittier National Park | ON239403 | ON243714 |
| N. liparus | SLE2123 | Venezuela: Barinas: Barinitas (ca. 13 km NW) | ON239401 | ON243715 |
| N. liparus | SLE2124 | Venezuela: Mérida: Santo Domingo (ca. 2 km SE ) | ON239400 | ON243716 |
| N. liparus | MSC1820 | Venezuela: Aragua: Henri Pittier National Park | ON239402 | KC992598 |
| N. lohezi | SLE2337 | French Guiana: Savane Roche Virginie | ON239444 | ON243737 |
| N. lohezi | SLE2387 | French Guiana: Savane Roche Virginie | ON239445 | ON243738 |
| N. parvus | SLE2388 | Suriname: Sipaliwini: Grensgeberte Moutains | ON239434 | ON243729 |
| N. retusus | SLE2110 | Guyana: Region 8: ca. 7 km NW Chenapau | N/A | ON243735 |
| N. retusus | SLE2372 | Guyana: Region 8: Ayanganna Airstrip | ON239439 | ON243736 |
| N. tricarinatus | SLE1112 | Venezuela: Zulia: Tukuko | ON239425 | ON243710 |
| N. tricarinatus | SLE2371 | Venezuela: Zulia: Tukuko | ON239426 | ON243711 |
| N. tricarinatus | SLE2381 | Venezuela: Aragua: Henri Pittier National Park | ON239424 | ON243712 |
| N. tricarinatus | SLE2391 | Venezuela: Portuguesa: Biscucuy | N/A | ON243713 |
| N. tricarinatus | SLE2392 | Venezuela: Portuguesa: Biscucuy | N/A | ON243708 |
| N. tricarinatus | SLE2397 | Costa Rica: Cartago: Tapanti National Park | N/A | ON243709 |
| N. vatius | SLE2104 | Brazil: Bahia: Cachoeira Domingos Lopes | ON239430 | ON243722 |
| N. vatius | SLE2324 | Brazil: Mato Grosso do Sul: Aquidauana (ca. 27 km S ) | ON239432 | ON243724 |
| N. vatius | SLE2327 | Brazil: Mato Grosso do Sul: Aquidauana (ca. 15 km E) | ON239433 | ON243723 |
| N. vatius | SLE2385 | Brazil: Bahia: Livramento de Nossa Senhora | ON239431 | ON243721 |
| $N$. sp. | SLE0092 | India: Tamil Nadu: Bodi Hills, Western Ghats | ON239399 | KC992599 |
| N. sp. | SLE2140 | Peru: Cusco: 1 km N. Quince Mil | ON239398 | ON243717 |

## Morphological methods

We examined more than 900 specimens for this work, including the holotypes of most previously described Neotropical species. The specimens were examined using Olympus SZ61 and SZX7 microscopes and preparation of the specimens and dissection of the genitalia were carried out following the methodology of Girón and Short (2017). There was a modification in the time of exposure to heat of the genitalia during the clearing process depending on the level of sclerotization of the structure, which we reduced the time from 60 min to $30-40 \mathrm{~min}$. Morphological terminology mainly followed Hansen (1991) and terminology adapted from Lawrence and Ślipiński (2013) regarding the use of meso- and metaventrite. Cleared genitalia photos were taken with Olympus BX51 microscope to $400 \times$ magnification (except for $N$. rosalesi genitalia to $100 \times$ ), $\sim 7-15$
photos were taken per genitalia and stacked using CombineZP software. We generated the species distribution maps using the software SimpleMappr (Shorthouse 2010). The data on the holotype labels are provided verbatim and cited in quotation marks.

## Depositories of examined material

CBDG Center for Biological Diversity, University of Guyana, Georgetown (G. Maharaj);
INPA Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil (M. Oliveira);
NMW Naturhistorisches Museum, Vienna, Austria (M. Jäch);
MALUZ Museo de Artrópodos de la Universidad del Zulia, Maracaibo, Venezuela (J. Camacho, M. García);
MNHN Muséum national d'Histoire naturelle, Paris, France;
MIZA Museo del Instituto de Zoología Agrícola, Maracay, Venezuela (L. Joly);
NZCS National Zoological Collection of Suriname, Paramaribo (P. Ouboter, V. Kadosoe);
SCC Private collection of Simon Clavier, Kourou, French Guiana;
SEMC Snow Entomological Collection, University of Kansas, Lawrence, KS (A. Short);
UMSP University of Minnesota Insect Collection, St. Paul, MN (R. Holzenthal, R.Thomson);

USNM U.S. National Museum of Natural History, Smithsonian Institution, Washington, DC (C. Micheli).

## Results

From our integrated approach of combining morphological data with DNA sequence data from two genes, we found support for 18 species in the Neotropical region, including twelve new species. At the same time, we found that two of the most widespread species have been described multiple times, resulting in four new synonymies. For the 14 species that we had molecular data, the maximum likelihood analysis (Fig. 1) recovered two well-supported species groups. We found morphological characters that can be used to diagnose these two clades, which we here define as the lohezi and liparus species groups. In addition, two species that were not included in our molecular analysis are assigned to their own monotypic species groups as they present unique character combinations that were not consistent with either of the other species groups.

Among the 14 species for which we had molecular data, the minimum uncorrected pairwise distances in COI between any two species was $5.0 \%$ (between N. giraldoi and $N$. vatius) with the exception of $N$. dilucidus, in which the two specimens from Venezuela (SLE2366 and SLE2378) were 6\% divergent from the remaining populations further to the east (Guyana, Brazil, Suriname, French Guiana). We sequenced one specimen from Peru that is known only from a single female, and we were unable
to associate it with confidence to any of the described species for which we did not have DNA; however, due to the importance of the male genitalia in identifications, we refrain from describing this species here.


Figure I. Maximum likelihood phylogeny of Notionotus spp. inferred from COI and 28 S sequence data. Extraction numbers are given next to each terminal name (see Table 1).

## List of species

## Notionotus liparus species group

1. Notionotus dilucidus Queney, 2010: Brazil (Roraima), French Guiana, Guyana, Suriname, Venezuela
Notionotus shorti Queney, 2010, syn. nov.
2. Notionotus giraldoi sp. nov.: Brazil (Rondonia)
3. Notionotus liparus Spangler, 1972: Venezuela
4. Notionotus mexicanus Perkins, 1979: Mexico
5. Notionotus tricarinatus Perkins, 1979: Costa Rica, Guatemala, Panama, Venezuela Notionotus edibethae García, 2000, syn. nov.
Notionotus nucleus Perkins, 1979, syn. nov.
Notionotus perijanus García, 2000, syn. nov.
6. Notionotus vatius sp. nov. :Brazil (Bahia, Mato Grosso do Sul)

## Notionotus lohezi species group

7. Notionotus bicolor sp. nov.: Suriname
8. Notionotus bifidus sp. nov.: Venezuela
9. Notionotus brunbadius sp. nov.: Brazil (Amazonas)
10. Notionotus garciae sp. nov.: Brazil (Amazonas)
11. Notionotus insignitus sp. nov.: Venezuela
12. Notionotus juma sp. nov.: Brazil (Amazonas)
13. Notionotus lohezi Queney, 2010: Guyana
14. Notionotus parvus sp. nov.: Suriname
15. Notionotus patamona sp. nov.: Guyana
16. Notionotus retusus sp. nov.: Guyana

Notionotus peruensis species group
17. Notionotus peruensis sp. nov.: Peru

## Notionotus rosalesi species group

18. Notionotus rosalesi Spangler, 1972: Trinidad, Venezuela

## Taxonomy

Genus Notionotus Spangler, 1972
Notionotus Spangler, 1972: 139.
Type species. Notionotus rosalesi Spangler, 1972: 141; by original designation.

Differential diagnosis for Neotropical species. Small to very small beetles, total body length $1.5-2.0 \mathrm{~mm}$. Color yellow, reddish brown, dark and pale brown to black. Body shape oval in dorsal view; moderately convex to convex in lateral view. Antennae with eight antennomeres. Maxillary palps short, nearly half the width of the head, second segment bending outwards, apical segment $\sim 2 \times$ as long as the penultimate segment (Fig. 4J; e.g., N. bifidus sp. nov.). Eyes reniform in dorsal view. Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Head, pronotum and elytra with ground and systematic punctures; systematic punctures of the head very sparse. The elytral ground punctation is more evident in some species (Fig. 2A; e.g., N. liparus) than in others (Fig. 4G; e.g., N. garciae sp. nov.); systematic punctures extremely reduce and sparse detectable for short seta, forming very sparse rows (Fig. 4B; e.g., $N$. patamona sp. nov.); elytra without sutural stria. Prosternum carinate medially, strongly raised, and projected anteromedially. Elevation of mesoventrite strongly raised forming an anteromedial carina consisting of one (Fig. 10A, B) or two longitudinal ridges and one transverse (Fig. 10C, D), extending between procoxae on the same plane as metaventrite. Metaventrite densely pubescent, slightly elevated, with elevation broad posteromedially and convex medially forming a glabrous patch drop-shaped; and two posterolateral glabrous patches in a half-circle shape. Pro- and mesofemora mostly covered with pubescence on basal three-quarters (Fig. 3B; e.g., N. tricarinatus); metafemora with pubescence, sometimes on basal three-quarters, or along basal three-quarters of the anterior margin with some setae on the posterior basal margin (Fig. 2H; e.g., N. rosalesi). Abdominal ventrites densely pubescent, with fifth ventrite bearing an apical emargination to shallowly truncate. Aedeagus trilobed, size and form variable.

Remarks. As this revision only treats the New World species, we did not comprehensively examine the Old World species to generate a global genus description. Therefore, the diagnosis above should be considered for Neotropical species only. The Old World species of Notionotus are generally similar to the New World species, but some species do differ in significant characters: for example, N. suturalis Hebaeur, 2003 has a sutural stria and antennae with 9 antennomeres.

Remarks of diagnostic features of Notionotus Spangler, 1972. Body shape and coloration. The degree of convexity between species is variable; some are moderately convex, others weakly convex. The general dorsal coloration of the body among species ranges from yellow (e.g., N. tricarinatus, Fig. 3A) to nearly black (e.g., N. liparus, Fig. 2A), however color alone is usually not sufficient on its own to definitively identify most Notionotus species. This is due both to the fact that some species share the same coloration, as well as some species have a slight variation in dorsal coloration. Species with unique (so far as currently known) color patterns include N. liparus (entirely dark brown to black, Fig. 2A), $N$. rosalesi (tricolored, Fig. 2G) and $N$. insignitus sp. nov. (with pale spot on the elytral disc Fig. 4D). The coloration of the head in some species is uniform, but in others is bicolorous (with typically the frons being darker than the clypeus).

Mesoventrite. In Notionotus the elevation of the mesoventrite is composed of two or three laminae: one transverse ridge and one longitudinal (Fig. 10A, B) or two transverse ridges and one longitudinal ridge (Fig. 10C, D), which generally converge and fuse


Figure 2. Habitus and labels of Notionotus spp.: N. liparus (holotype): A dorsal view $\mathbf{B}$ ventral view $\mathbf{C}$ labels; $N$. mexicanus (holotype): $\mathbf{D}$ dorsal view $\mathbf{E}$ ventral view $\mathbf{F}$ labels; $N$. rosalesi (holotype): $\mathbf{G}$ dorsal view $\mathbf{H}$ ventral view $\boldsymbol{I}$ labels.


Figure 3. Habitus and labels of Notionotus spp.: $N$. tricarinatus (holotype): A dorsal view $\mathbf{B}$ ventral view $\mathbf{C}$ labels; $N$. nucleus (holotype): $\mathbf{D}$ dorsal view $\mathbf{E}$ ventral view $\mathbf{F}$ labels; $N$. edibethae (holotype): $\mathbf{G}$ dorsal view $\mathbf{H}$ ventral view I labels.
medially; the shape of the longitudinal ridge shows high variation among species. In general, the transverse ridges are medially elevated and laterally concave. The apex of the transverse ridge can be nearly acute (Fig. 10B), or blunt (Fig. 10C), and lateral sides vary from very to slightly concave or straight. The longitudinal ridge varies, it can be completely sharp or sharp anteriorly and broadening posteriorly reaching the end of the elevation, but it can also be broad anteriorly and sharp posteriorly. The point where the two or three ridges merged can be rounded and obtuse or wide and blunt respectively.

Elytral punctation. The density of the ground punctation is typically sparse, and the degree of impression is variable between species within the genus. In some species, the ground punctation is very weakly impressed and may almost appear absent and low magnification (Fig. 4H; e.g., N. vatius sp. nov.); in other cases, it is more coarse and moderately impressed (Fig. 4E; e.g., N. juma sp. nov.).

Aedeagus. The shape of the aedeagus is the most important and often crucial feature to identify species of Notionotus. Most species in the Neotropics exhibit two different generalized aedeagal forms: in some species, the median lobe and basal piece have the same length, or the median lobe is slightly longer than the parameres (e.g., $N$. tricarinatus, Fig. 6A). However, some species present the median lobe and basal lobe that are shorter than the parameres (e.g., N. bicolor sp. nov. (Fig. 8A), N. retusus sp. nov. (Fig. 8E), N. parvus sp. nov. (Fig. 9E)). In terms of shape, some species present variation in the apex of the median lobe, this is usually rounded (e.g., N. patamona sp. nov., Fig. 8D), but it varies from acute (e.g., N. liparus, Fig. 7A), emarginated (e.g., N. juma, Fig. 8G), and bifurcated (e.g., N. bifidus sp. nov., Fig. 9D). Additionally, the width of the median lobe varies from very slender (e.g., N. giraldoi, Fig. 7D) to very wide (e.g., N. bifidus, Fig. 9D). Nevertheless, in most of the species, the median lobe is wide at the base and slightly narrowing towards the apex.

## Notionotus liparus species group

Diagnosis. The species of this group can be diagnosed by the following combination of characters: (1) the shape of the elevation mesoventrite, having one transverse ridge and one longitudinal ridge (Fig. 10A, B); (2) parameres nearly as long as the basal piece; the length of the median lobe and the length of the parameres is approximately subequal. (e.g., Notionotus liparus, Fig. 7A).

## Notionotus dilucidus Queney, 2010

Figs 5A-J, 10A, 15

Notionotus dilucidus Queney, 2010: 130.

Type material examined. Paratype (male): " "", "Notionotus/dilucidus n. sp./ PARATYPE/ P. QUENEY descr. 2010", "Guyane [= French Guiana]: Roura,/ Cacao, Chemin/ Molokoi, crique,/ 16-IX-2009/ leg. P. Queney" (SEMC). Paratypes (8 exs.): same data as the dissected paratype (8 exs., SEMC).

## Notionotus shorti Queney, 2010: 133. syn. nov.

Type material examined. Holotype (male): " $\overline{0}$ ", "Notionotus shorti/n. sp. HOLOTYPE/P. QUENEY descr.2010", "Guyana: Mazaruni-Potaro/District, Takutu Mountains,/ stream debris berlesed, 18-/XII-1983, leg. P.J. Spangler,/ W.E. Steiner \& M. Levine" (USNM). Paratypes (8 exs.): same data as the holotype (8 exs., SEMC).

Additional material examined (362 exs.). Brazil: Roraima State: Serra do Tepequém, $3^{\circ} 47.334^{\prime} \mathrm{N}, 61^{\circ} 42.570^{\prime} \mathrm{W}, 14.1 .2018$, leg. Short, Benetti, \& Santana, small forested stream, BR18-0114-02A (1 ex., SEMC, DNA voucher SLE2377); Caroebe, Rio Caroebe, ca. 13 km NE of Caroebe, $00^{\circ} 54.786^{\prime} \mathrm{N}, 59^{\circ} 34.397^{\prime} \mathrm{W}, 150 \mathrm{~m}, 17 . \mathrm{i} .2018$, leg. Short and Benetti, margins of sandy river, BR18-0117-04A (26 exs., INPA, SEMC, including DNA voucher SLE2330); Rio Jatapú nr. Usina de Jatapú, $00^{\circ} 50.939^{\prime} \mathrm{N}, 59^{\circ} 18.262^{\prime} \mathrm{W}, 145 \mathrm{~m}, 17 . \mathrm{i} .2018$, leg. A. Short, marginal pools of river, BR18-0117-02A (1 ex., SEMC); Amajari, Rio Cocal, $3^{\circ} 44.135^{\prime} \mathrm{N}, 61^{\circ} 43.542^{\prime} \mathrm{W}$, 237 m, 15.i.2018, leg. A. Short, clearwater creek, BR18-0115-01A (6 exs., SEMC, including DNA voucher SLE2374). French Guiana: St. Laurent du Maroni ca. 15 km SW, Crique des cascades, 5.34662, $-54.10539,17 \mathrm{~m}, 4 . \mathrm{iii} .2020$; leg. Short and Neff, stream margin, FG20-0304-02A (1 ex., SEMC); same data except leg. Short, leaf packs on rock, FG20-0304-02B ( 2 exs., SEMC, including DNA voucher SLE2339); same data except small rock pools in granite FG20-0304-02C (1 ex., SEMC); Carbet ONF Grillon, Piste Bélizon, Crique Grillon, 4.28219, -52.45163 , $65 \mathrm{~m}, 11 . \mathrm{iii} .2020$; leg. Short, rock pools by waterfall, FG20-0311-01C (2 exs., SEMC, including DNA voucher SLE2383); Crique à l'Est above Crique Eau Chire, 3.66383, -53.22193, $156 \mathrm{~m}, 10 . x i .2016$, leg. D. Post (1 ex., SEMC, DNA voucher SLE2113). Guyana: Region 6: Upper Berbice, ca. 1 km , W basecamp, $4^{\circ} 09.143$ 'N, $58^{\circ} 11.207^{\prime} \mathrm{W}, 170 \mathrm{~m}$, 22.ix.2014, leg. Short, Salisbury, La Cruz, margins of creek, GY14-0921-03H (6 exs., SEMC); Upper Berbice, Basecamp 1, creek next to basecamp (upstream), 4.154817, -58.178616, 24.ix.2014, leg. Short, creek margins, GY14-0924-01A (1 ex., SEMC, DNA voucher SLE2108); Upper Berbice, Basecamp 1, $4^{\circ} 09.289^{\prime} \mathrm{N}, 58^{\circ} 10.717^{\prime} \mathrm{W}$, 96 m, 22.ix.2014, leg. Short, Salisbury, La Cruz, GY14-0924-01A (2 ex., SEMC). Region 8: Upper Potaro Camp I (ca. 7 km NW Chenapau), $5^{\circ} 0.673^{\prime} \mathrm{N}, 59^{\circ} 38.358^{\prime} \mathrm{W}$, 500 m, 14.iii.2014, leg. Short, Salisbury, La Cruz, clear water creek rapids, GY14-0314-01A (7 exs., SEMC); same data except $5^{\circ} 0.660^{\prime} \mathrm{N}, 59^{\circ} 38.283^{\prime} \mathrm{W}, 484 \mathrm{~m}, 11$. iii.2014, leg. Short, Baca, Salisbury, La Cruz, clear water creek rapids, GY14-031104A (6 exs., SEMC, including DNA voucher SLE2368); same data except Potaro margin trail, $5^{\circ} 0.571^{\prime} \mathrm{N}, 59^{\circ} 38.202^{\prime} \mathrm{W}, 524 \mathrm{~m}, 11 . \mathrm{iii} .2014$, leg. Short and Baca, leaf packs in rocky stream, GY14-0311-05A (1 ex., SEMC). Region 9: Kusad, Mts., Mokoro Creek, $2^{\circ} 48.531^{\prime} \mathrm{N}, 59^{\circ} 51.900^{\prime} \mathrm{W}, 170 \mathrm{~m}, 27 . x .2013$, leg. Short, Isaacs and Salisbury, main seepage area, GY13-1027-03B (2 exs., SEMC); same data except basecamp, leg. A. Short and Washington, on wet rocks, GY13-1024-03C (4 exs., SEMC); same data except 24.x.2013, leg. Salisbury, small rock pool with detritus, GY13-102403A (1 ex., SEMC); Kusad, Mts., basecamp area, $2^{\circ} 48.588^{\prime} \mathrm{N}, 59^{\circ} 51.931^{\prime} \mathrm{W}, 194 \mathrm{~m}$, 23.x.2013, leg. A. Short, leaf packs in flow of creek, GY13-1023-02B (9 exs., CDBG,

SEMC, including DNA voucher SLE2335); Kusad, Mts., Taraara Wao, $2^{\circ} 47.417{ }^{\prime} \mathrm{N}$, $59^{\circ} 53.986^{\prime} \mathrm{W}, 113 \mathrm{~m}, 28 . x .2013$, leg. Short, Isaacs and Salisbury, margin and isolated side pools, GY13-1028-01A (5 exs., SEMC, including DNA voucher SLE2107); N. Parabara, creek by basecamp (Bototo wau creek), $2^{\circ} 10.908^{\prime} \mathrm{N}, 59^{\circ} 20.306^{\prime} \mathrm{W}$, leg. Short, 31.x.2013, stream margins, GY13-1031-01A (1 ex., SEMC, DNA voucher SLE2380). Region Mazaruni-Potaro: Takutu Mountains, $6^{\circ} 15^{\prime} \mathrm{N}, 59^{\circ} 5^{\prime} \mathrm{W}, 2-14$. xii.1983, leg. P.D. Perkins, stream ex. leaf packs and twigs (8 exs., SEMC) Suriname: Brokopondo District: Brownsberg Nature Park, near Capaci House, $4^{\circ} 56.934^{\prime}$ N, $55^{\circ} 10.825^{\prime} \mathrm{W}, 460 \mathrm{~m}, 17 . \mathrm{iii} .2017$, leg. A. Short, small stream, SR17-0317-01A (1 ex., SEMC); Leo Val/lrene Val return loop trail, 4.95069 'N, $-55.18599^{\prime} \mathrm{W}, 470 \mathrm{~m}, 18$. iii.2017, leg. Baca and Johnson, stream, SR17-0318-01B (1 ex., SEMC); Leo Val, $4^{\circ} 57^{\prime} 16.08^{\prime \prime N}$, $-55^{\circ} 11^{\prime} 26.82^{\prime \prime} \mathrm{W}, 317 \mathrm{~m}, 19 . i i i .2017$, leg. Short, leaf packs/detritus from behind waterfall, SR17-0319-01C ( 5 exs., SEMC); same data except SR17-0323-01C (6 exs., SEMC); same data except 23.iii.2017, leg. Baca, submerged woody debris, SR17-0323-01A (1 ex., SEMC); same data except side pools in creek, SR17-0323-01D (1 ex., SEMC); Witti Kreek, $4^{\circ} 55.674^{\prime} \mathrm{N}, 55^{\circ} 09.874$ 'W, $84 \mathrm{~m}, 21 . i i i .2017$, leg. Short and Baca, small side stream, SR17-0321-01D (2 exs., SEMC, including DNA voucher SLE2121); Brownsberg Nature Park, Mazaroni Val, $04^{\circ} 56.351^{\prime}$ N, $55^{\circ} 12.108^{\prime} \mathrm{W}, 394 \mathrm{~m}, 5 . v i i i .2012$, leg. Short, Maier, Mcintosh, forested waterfall and stream, SR12-0805-01A (22 exs., NZCS, SEMC); Brownsberg Nature Park, $04^{\circ} 57.268^{\prime} \mathrm{N}, 55^{\circ} 11.447^{\prime} \mathrm{W}, 317 \mathrm{~m}, 4$. viii.2012, leg. Short, Maier, Mcintosh, forested waterfall and stream, SR12-0804-02A (21 exs., SEMC); Brownsberg Nature Park, $04^{\circ} 56.871^{\prime} \mathrm{N}, 55^{\circ} 10.911^{\prime} \mathrm{W}, 462 \mathrm{~m}, 4$. viii.2012, leg. Short, Maier, McIntosh, SR12-0804-01A (2 exs., SEMC including SLE505 and SLE506). Sipaliwini District: Kabalebo Nature Resort: Bwkw rapids, $4.40041^{\circ} \mathrm{N}, 57.24658^{\circ} \mathrm{W}, 90 \mathrm{~m}, 9-10.9 \mathrm{iii} .2019$, leg. Short and class, large isolated muddy pool by river, SR19-0309-02A (1 ex., SEMC); Moi Moi creek, $4.42313^{\circ}$ N, $57.19198^{\circ} \mathrm{W}, 104 \mathrm{~m}, 10-14.1 i i .2019$, leg. Short and class, rock and detrital pools along creek, SR19-0310-01A (3 exs., SEMC); same data except leg. Short and Baca, small seeps, SR19-0310-01F (10 exs., SEMC); same data except leg. Short, margin of detrital pool in drying creekbed, SR19-0310-01G (5 exs., SEMC); same data except leg. Short, Baca and class, SR19-0310-01J (1 ex., SEMC); same data except leg. Baca, margin of stream pool with root mats, SR19-0310-01L (5 exs., SEMC); same data except leg. Short and class, SR19-0310-01M (10 exs., SEMC, including DNA voucher SLE1799); same data except Leg. S. Baca, upstream riparian habitats, SR19-0310-01N (2 exs., SEMC); Sand Crk, $4.38476^{\circ} \mathrm{N}$, $57.24636^{\circ} \mathrm{W}, 72 \mathrm{~m}, 13-15 . \mathrm{iii} .2019$, leg. Short and Baca, upper marginal pool along river, SR19-0313-01D (2 exs., SEMC); Charlie Falls, $4.38302^{\circ} \mathrm{N}, 57.21161^{\circ} \mathrm{W}$, $174 \mathrm{~m}, 11 . \mathrm{iii} .2019$, leg. Short and class, rock pools in creekbed, SR19-0311-01A (39 exs., NZCS, SEMC, including DNA voucher SLE1811); same data except leg. Short, seepage, SR19-0311-01B (7 exs., SEMC); Sipaliwini Savanna Nature Res. 4-Brothers Mts, $2^{\circ} 00.342^{\prime} \mathrm{N}, 55^{\circ} 58.149^{\prime} \mathrm{W}, 337 \mathrm{~m}, 31 . \mathrm{iii} .2017$, leg. Short and Baca, clear water stream sandy with detritus, SR17-0331-01B (31 exs., SEMC, including DNA Voucher SLE2114); same data except leg. Baca, small rocky creek, SR17-0331-01A (1 ex.,

SEMC); Raleighvallen Nature Reserve, Fungu Island, $4^{\circ} 43.459^{\prime} \mathrm{N}, 56^{\circ} 12.658^{\prime} \mathrm{W}$, $30 \mathrm{~m}, 14 . \mathrm{iii} .2016$, leg. A. Short, isolated river margin pools, rocky bottom, SR16-0314-01E (1 ex., SEMC); Coppename Rvr-Voltzberg trail, 15.iii.2016, leg. A. Short, small sandy stream, SR16-0315-02A (1 ex., SEMC); Lolopaise Area, $4^{\circ} 42.48^{\prime} \mathrm{N}$, $56^{\circ} 13.15908^{\prime} \mathrm{W}, 24 \mathrm{~m}, 18 . i i i .2016$, leg. Short et al., intermittent stream margins, flotation, SR16-0318-01D (1 ex., SEMC); Brownsberg Nature Park, $04^{\circ} 56.871^{\prime} \mathrm{N}$, $55^{\circ} 10.911^{\prime} \mathrm{W}, 462 \mathrm{~m}, 4 . v i i i .2012$, leg. Short, Maier, Mcintosh, forested stream with lots of detritus, SR12-0804-01A (8 exs., SEMC); Raleighfallen Nature Reserve, trail to Raleighfallen, $04^{\circ} 42.480^{\prime} \mathrm{N}, 56^{\circ} 13.159^{\prime} \mathrm{W}, 24 \mathrm{~m}, 27 . v i i .2012$, leg. Short, Mcintosh and Kadosoe, creek margins, SR12-0727-03A (22 exs., SEMC, including DNA voucher SLE2394); same data except leg. C. Mcintosh, detrital pools near creek in forest, SR12-0727-03D (1 ex., SEMC); Raleighvallen Nature Reserve Voltzberg Station, $04^{\circ} 40.910^{\prime} \mathrm{N}, 56^{\circ} 11.138^{\prime} \mathrm{W}, 78 \mathrm{~m}, 9 . v i i .2012$, leg. Short, Maier, McIntosh, and Kadosoe, stream margins, SR12-0729-02A (1 ex., SEMC, DNA voucher SLE2389); Raleighfallen Nature Reserve, Voltzberg trail, $04^{\circ} 40.910^{\prime} \mathrm{N}, 56^{\circ} 11.138^{\prime} \mathrm{W}, 78 \mathrm{~m}, 30$. vii.2012, leg. C. Maier and V. Kadosoe, margin of stream, SR12-0730-01A (1 ex., SEMC); Raleighfallen Nature Reserve, Fungu island, $04^{\circ} 43.459^{\prime} \mathrm{N}, 56^{\circ} 12.658^{\prime} \mathrm{W}$, 30 m, 1.viii.2012, leg. Short, Maier, Mcintosh and Kadosoe, small creek, SR12-080101B (2 exs., SEMC); Camp 2, on Sipaliwini District, CI-RAP Survey, $2^{\circ} 10.973^{\prime} \mathrm{N}$, $56^{\circ} 47.235^{\prime} \mathrm{W}, 210 \mathrm{~m}, 29-30 . v i i i .2010$, leg. Short and Kadosoe, Inselberg, SR10-0829-01A ( 2 exs., SEMC); Camp 1, Upper Palumeu, CI-RAP Survey, $2.47700^{\circ} \mathrm{N}$, $55.62941^{\circ} \mathrm{W}, 275 \mathrm{~m}, 11 . \mathrm{iii} .2012$, leg. Short and Kadosoe, around waterfall, SR12-0311-03A (2 exs., SEMC); Camp 3, Wehepai, $2^{\circ} 21.776^{\prime} \mathrm{N}, 56^{\circ} 41.861^{\prime} \mathrm{W}, 237 \mathrm{~m}$, 4-6.ix.2010, leg. Short and Kadosoe, sandy forest creek, SR10-0904-01A (36 exs., SEMC, including DNA voucher SLE2375). Venezuela: Bolívar State: Piedra de la Virgen, $6^{\circ} 5^{\prime} 14.1^{\prime \prime N}, 61^{\circ} 23^{\prime} 55.8^{\prime W} \mathrm{~W}, 400 \mathrm{~m}, 31 . v i i .2008$, leg. A. Short M. García, L. Joly, small forest stream, AS-08-056 (7 exs., SEMC, including DNA voucher SLE2378). Amazonas State: Tobogán de la Selva, $5^{\circ} 23.207^{\prime} \mathrm{N}, 67^{\circ} 36.922^{\prime} \mathrm{W}, 125 \mathrm{~m}$, 14.i.2009, leg. A. Short, clumps of wet leaves on rock, VZ09-0114-01D (6 exs., MIZA, SEMC, including DNA voucher SLE2366).

Differential diagnosis. The general coloration of Notionotus dilucidus is similar to $N$. giraldoi, N. mexicanus, and N. tricarinatus and these species are very difficult to differentiate with external characters alone. However, the shape of the aedeagus in $N$. dilucidus is quite distinct: the outer and inner margins of the parameres are sinuate, the apex of the parameres presents an indentation in which the depth can vary from very deep, slightly deep, or almost not distinguishable and pointing outwards (Fig. 5A-J).

Description. Size and form: Body length $1.6-1.8 \mathrm{~mm}$. Body form elongate oval, moderately convex in lateral view (e.g., Fig. 2D). Color and punctation: Dorsally yellow to pale brown, head mostly yellow and frons pale brown; pronotum paler than elytra, with two small black round spots along posterior margin (e.g., Fig. 2D). Ventrally brown; maxillary palps and antennae yellow (antennal club slightly darker), mouth parts and legs pale brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $1-2 \times$ their width); pronotum and elytra
with ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp, the point where the two ridges merged rounded and obtuse (Fig. 10A); elevation flat in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area, medial region patch drop-shaped; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters (e.g., Fig. 2E). Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 5A) basal piece nearly the same length of a paramere. Base of the parameres slightly narrower than the base of the median lobe; outer margins sinuate, tapering almost reaching the apex, inner margins sinuate, apex of the parameres slightly bending outwards, rounded in the outer margin with an indentation in the inner margin (the depth of the indentation varies, Fig. 5A-J). Median lobe nearly as long as the parameres or a bit longer, broad at the base and gradually narrowing to the apex, apex of the median lobe rounded.

Variation. There is significant variation in the apex of the parameres of the aedeagus. The paramere tip is weakly sclerotized, as evidenced by its very pale to white appearance in cleared specimens (e.g., Fig. 5A-J). This tip varies from nearly evenly rounded (e.g., Fig. 5D, G) to possessing some level of indentation on the inner margin (e.g., Fig. 5A, H). Although this variation seems in part to be real, it also seems to be partly caused by artificial distortions; for example, the weakly sclerotized apex may become indented or inflated depending on the state of the genitalia during preparation. This would explain why there is a large variation in form as well as why the tip appears to be exhibit subtly asymmetry between parameres (e.g., Fig. 5C, F).

Distribution. This species is widely distributed throughout the Guiana Shield region, from Tobogán de la Selva in Venezuela east to the coast of French Guiana (Fig. 15). Reported from Suriname in Short and Kadosoe (2011) and Short (2013; as N. shorti) and from Guyana as Notionotus sp. B in Short et al. (2018).

Life history. This species is the most common and abundant species of the genus in the Guiana Shield region and is frequently collected in leaf packs or along the margins of forested streams. They are typically found in streams that are lined with detritus and have rocky or sandy substrates. Although some specimens have been collected in seepage-like habitats or adjacent to seepages, this does not seem to be a primary or favored habitat for this species.

Remarks. Queney (2010) described N. dilucidus based on specimens from several closely situated localities in French Guiana, and $N$. shorti based on a long series of specimens from a single collecting event in Guyana. The primary feature used to separate the two taxa was the apex of the parameters, which was given as "parameres apically shortly curved outwards" in $N$. dilucidus and "parameres apically almost straight" in $N$. shorti. We examined material from the type series of both species and confirmed there
are notable differences in the paramere shape. If we only had access to the specimens that Queney (2010) examined and no other data, we also would have very likely concluded they were different species and described them as such. However, as we examined recently collected specimens from numerous other localities in the eastern Guiana Shield, we observed quite a bit of variation in the shape of the parameres that encompassed the forms present in $N$. dilucidus and $N$. shorti. In some cases, we noticed variation in this feature even among specimens from the same series. Additionally, the apex of the paramere even occasionally appears slightly asymmetrical in some dissections (e.g., Fig. 5F, J).

To help interpret these morphological observations, we sequenced 22 specimens that span the entire width of the Guiana Shield, including specimens from Venezuela, Brazil, Guyana, Suriname, and French Guiana. These specimens also represented a range of paramere form diversity. In general, we found little meaningful molecular divergence among these populations (Fig. 1) and these differences were not correlated to observed variations in the paramere apex. Therefore, we here consider N. shorti as a junior subjective synonym of $N$. dilucidus.

Among all sequenced specimens, the maximum pairwise divergence in the gene COI was $6.0 \%$. Although this is on the higher end of intraspecific divergence observed in hydrophilids, it is seen in some taxa (see Short and Girón 2018; Smith and Short 2020). Additionally, given that its geographic range spans 1500 kilometers, it is not surprising to see such divergences. The two Venezuelan populations (from Tobogán de Selva in Amazonas State and the Escalera region of Bolívar State) were the most genetically distinct and sister to the more eastern populations. Indeed, with these samples removed, the maximum intraspecific divergence among remaining specimens drops to just $3.8 \%$. However, we did not observe any significant morphological differences from these Venezuelan populations and therefore consider them to be part of a broader definition of $N$. dilucidus.

As both $N$. dilucidus and $N$. shorti were proposed in the same work (Queney 2010), we use our authority as first revisors (ICZN article 24.2.2) to give precedence to $N$. dilucidus as the valid name for this species.

## Notionotus giraldoi sp. nov.

http://zoobank.org/05F6195D-EB81-47FB-B7FD-2256F49D877F
Figs 4I, 7D, 13A, 14

Type material. Holotype (male): "BRAZIL: Rondonia: Novo [sic: Nova] Uniao/ $-10.91764^{\circ},-62.377^{\circ}, 359 \mathrm{~m} /$ Vale do Cachoeiras; 10.vii.2018/ leg. Short; Margin of rocky/ stream; BR18-0710-02B" (INPA). Paratypes (56 exs.): Brazil: Mato Grosso do Sul State: Rio Bento Gomes (Pantanal), Campo Allegre I, $15^{\circ} 45^{\prime} \mathrm{S}, 56^{\circ} 33^{\prime} \mathrm{W}$, 1993-1994, leg. E. Stuhr, spring-fed brook, (9 exs., NMW, SEMC). Rondonia State: Same data as holotype (13 exs., SEMC, including DNA Voucher SLE2334); same data except margin of rocky stream with waterfall (12 exs., SEMC); same data except small sandy-bottom stream margin BR18-0710-02A (20 exs., INPA, SEMC, including DNA voucher SLE2088); same data except flotation of marginal root mats, BR18-0711-01C (1 ex., SEMC); Ji-Parana ( 27 km SW ), Rio Urupa, rock pools along


Figure 4. Habitus of Notionotus spp.: A N. bicolor (paratype) B N. patamona (holotype) C N. brunbadius (holotype) D N. insignitus (paratype) E N. juma (paratype) F N. parvus (holotype) G N. garciae (paratype) H $N$. vatius (holotype) I $N$. giraldoi (paratype) J $N$. bifidus (paratype) K $N$. peruensis (holotype).
margins of river, $-11.03618,-62.14465,135 \mathrm{~m}$, leg. Short, $10 . v i i .2018$, rock pools along margins of river, BR18-0710-01A (1 ex., SEMC, DNA voucher SLE2332).

Differential diagnosis. The dorsal coloration, shape of the elevation of the mesoventrite, area of pubescence on the metafemur and degree of impression of the ground
punctation of Notionotus giraldoi are very similar to $N$. dilucidus, $N$. mexicanus, $N$. tricarinatus. It can be distinguished only by its aedeagus, including the unique shape of the parameres with a depression of the inner margin, as well as by the abrupt narrowing in the midlength of the median lobe (Fig. 7D).

Description. Size and form: Body length $1.7-1.9 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4I). Color and punctation: Dorsally yellow, head yellow; pronotum paler than elytra, with two small black round spots along posterior margin (Fig. 4I). Ventrally brown; maxillary palps, mouthparts, antennae (antennal club slightly darker) and legs yellow. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra with ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp, the point where the two ridges merged rounded and obtuse (e.g., Fig. 10A, B); elevation flat in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area, medial region patch drop-shaped; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 7D) with basal piece nearly the same length of a paramere. Base of the parameres slightly narrower than the base of the median lobe; outer margins sinuate, inner margins depressed in the midlength, depression extending to apex without reaching it; apex of parameres wide and blunt. Median lobe length almost equal to the parameres, wide at basal region, narrowing abruptly in the midlength, apical third slender, narrow, and rounded.

Etymology. L. M. González-Rodríguez names this species in honor of Juan José Giraldo Gutiérrez in gratitude for the encouragement and support in her career.

Distribution. Known from several localities in the Brazilian States of Rondonia and Mato Grosso do Sul (Fig. 14).

Life history. This species was collected along the margins of two adjacent streams, one with a sandy bottom and one with a rocky bottom (Fig. 13A). Specimens were collected by agitating the sand and detritus along the stream margin as well as washing root mats in tubs of water.

## Notionotus liparus Spangler, 1972

Figs 2A-C, 7A, 11A, B, 14
Notionotus liparus Spangler, 1972: 144.
Type material examined. Holotype (male): "VENEZUELA/ Bar., 24 Km. NW/ Bar-initas/II-23-1969/P.\&P. Spangler", "Collected with/ 207 Oocyclus", "HOLOTYPE/ Notionotus/liparus/P.J. Spangler" (USNM).


Figure 5. Aedeagi of Notionotus dilucidus A N. dilucidus (paratype) B $N$. shorti (paratype) C-J N. dilucidus $\mathbf{C}$ specimen from Roraima, Brazil $\mathbf{D}$ specimen from Guyana $\mathbf{E}$ specimen from French Guiana $\mathbf{F}$ specimen from Suriname $\mathbf{G}$ specimen from Roraima, Brazil $\mathbf{H}$ specimen from Bolívar State, Venezuela I specimen from Amazonas State, Venezuela $\mathbf{J}$ specimen from Suriname.

Additional material examined (139 exs.). Venezuela: Aragua State: Henri Pittier National Park, Río Curucuruma, $10^{\circ} 21.070^{\prime}$ N, $67^{\circ} 34.920^{\prime} \mathrm{W}, 11 . i .2006$, leg. A.E.Z. Short, waterfall/seep, AS-06-023 (75 exs., MIZA, SEMC, including DNA Voucher MSC1820); Henri Pittier National Park, Río Castańo Regresiva del Diablo,
$10.35669^{\circ}$ N, $67.60645^{\circ}$ W, 6.i.2009, leg. A.E.Z. Short, log in stream, VZ09-010601A (5 exs., SEMC); same data except seeps/wet rock, VZ09-0106-01B (19 exs., SEMC); Henri Pittier National Park, Ranch Grande, 10.i.2006, leg. Short, stream and seep at Toma, AS-06-021, (8 exs., SEMC), same data except 2.vii.2020, VZ10-070201A (1 ex., SEMC, DNA voucher SLE2111). Barinas State: 13 km NW Barinitas, $8^{\circ} 48.424^{\prime} \mathrm{N}, 70^{\circ} 31.139^{\prime} \mathrm{W}, 992 \mathrm{~m}, 24 . \mathrm{i} .2012$, leg. A. Short \& Gustafson, seepage by road, VZ12-0124-02A (22 exs., SEMC, including DNA voucher SLE2123); same data except small stream pool, VZ12-0124-02B (1 ex., SEMC). Mérida State: ca. 2.5 km S . La Azulita, $8^{\circ} 44.335^{\prime} \mathrm{N}, 70^{\circ} 37.131^{\prime} \mathrm{W}, 842 \mathrm{~m}, 28.1 .2012$, leg. G.T. Gustafson, stream pools, VZ12-0128-02B (1 ex., SEMC); ca. 12 km , SE of Santo Domingo, $8^{\circ} 51.933^{\prime} \mathrm{N}$, $70^{\circ} 37.131^{\prime} \mathrm{W}, 1682 \mathrm{~m}, 22 . \mathrm{i} .2012$, leg. Short $\&$ Arias, wall seep 1, VZ12-0122-03A (7 exs., SEMC, including DNA voucher SLE2124). Trujillo State: ca. 3 km NE Laguna Agua Negro, $9^{\circ} 19.371^{\prime} \mathrm{N}, 70^{\circ} 9.303^{\prime} \mathrm{W}, 1770 \mathrm{~m}, 21 . i .2009$, leg. Short, García \& Camacho, small mountain stream w/ detritus, VZ09-0121-03x (1 ex., SEMC).

Differential diagnosis. Notionotus liparus can be recognized by the distinct black coloration among the other dark (reddish brown) species such as $N$. brunbadius, N. parvus and $N$. retusus, also for being the only dark species in the liparus group (Fig. 2A). It can also be differentiated by sharply marked punctation of the pronotum and elytra. Moreover, the outer margin of the parameres is sinuate and the apical third is slim and tapered. It is the only species in the liparus group with the apex of the median lobe acute.

Description. Size and form: Body length $1.6-1.8 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 2A). Color and punctation: Dorsally black, with lateral margins of the pronotum and elytra reddish brown (Fig. 2A). Ventrally reddish brown, except for black abdominal ventrites; maxillary palps and antennae yellow (antennal club slightly darker) (Fig. 2B). Clypeus and labrum with dense, coarse, and moderately impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation dense, coarse, and moderately impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge narrowed anteriorly and broadening posteriorly, the point where the two ridges merged blunt (e.g., Fig. 10A, B); elevation concave in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of posterior margin, then apical half of the posterior margin with sparse setae (Fig. 2B). Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 7A) with basal piece $1.3 \times$ the length of a paramere. Base of the parameres slightly narrower than the base of the median lobe; outer margin sinuate, inner margin nearly straight, tapering along apical third, parameres thin and rounded at apex. Median lobe shorter than the parameres, broad at the base and gradually widening to the apex, with the apex acute.

Distribution. This species is widespread in the Mérida Andes and Coastal Mountains of Venezuela (Fig. 14). Originally described from localities in the Venezuelan states of Barinas and Mérida (Spangler 1972, it was later recorded from the state of Aragua (García 2000). Here, we report additional localities in these three states as well as report it from the state of Trujillo for the first time.

Life history. This species is found in rock seepage habitats and wet rocks adjacent to waterfalls (Fig. 11A, B). Occasionally it is found in the pools that form at the best of these habitats.

Remarks. We sequenced four specimens from three Venezuelan states across the range of this species (Aragua, Barinas, and Mérida). The sequences are nearly identical (Fig. 1), supporting the concept of a widespread species in the Mérida Andes and Coastal mountains.

Notionotus mexicanus Perkins, 1979
Figs 2D-F, 7B, 14

Notionotus mexicanus Perkins, 1979: 306.

Type material examined. Holotype (male): "MEXICO, Оахаса, 8 mi . E./Tapanatepec, tropical/stream with lg. boulders/3-VII-1974/ME\&PD Perkins", "Type No/76326/U S N M", "HOLOTYPE/Notionotus/mexicanus/P.D.Perkins" (USNM).

Differential diagnosis. Notionotus mexicanus is very similar morphologically to $N$. tricarinatus sharing characters such as body length, yellow dorsal coloration, pronotum and elytra with fine ground punctation, and elevation of the mesoventrite with a transversal and a longitudinal ridge. It can be distinguished by the shape of the aedeagus, specifically the shape of the parameres: inner margins straight and sinuate reaching the apex, parameres narrowing along apical third, and narrower than $N$. tricarinatus.

Description. Size and form: Body length 1.8 mm . Body form elongate oval, convex in lateral view (Fig. 2D). Color and punctation: Dorsally yellow, head mostly yellow, frons pale brown; pronotum with two small black round spots along posterior margin (Fig. 2D). Ventrally brown; maxillary palps, mouthparts, antennae, pro and meso legs yellow, meta legs pale brown (Fig. 2E). Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp, the point where the two ridges merged rounded and obtuse (e.g., Fig. 10A, B); elevation concave in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending
to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters (Fig. 2E). Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 7B) with basal piece $1.1 \times$ the length of a paramere. Base of the parameres broader than the base of the median lobe; outer margin slightly convex along basal two-thirds, then slightly sinuate apically, inner margins nearly straight along basal two-thirds and then sinuate apically; apex of parameres rounded. Median lobe shorter than the parameres, approximately triangular, with acute apex.

Distribution. Only known from the type locality in Mexico (Fig. 14).
Life history. The type series was collected "from plant debris which had become trapped between stones in a rapid stream" (Perkins 1979).

Notionotus tricarinatus Perkins, 1979
Figs 3A-I, 6A-I, 11C, D, 14

Notionotus tricarinatus Perkins, 1979: 309.

Type material examined. Holotype (male): "PANAMA, C.Z./Albrook Forest Site/ ground, 22-III-/1968, R.S. Hutton", Type No/76324/U S N M", "HOLOTYPE/Notionotus/tricarinatus/P.D. Perkins" (USNM).

## Notionotus edibethae García, 2000: 250. syn. nov.

Type material examined. Holotype (male): "VENEZUELA. Trujillo/Mcpo. Rafael Rangel/La Guaira, Quebrada la/amarilla, 530 m./24-VIII-1997/Det. M. García, 199", "Col:/M. García/J. Camacho/E. Gomez", "Holotipo đ̄/ Notionotus/edibethae/ Dcrip. M. García, 1997" (MALUZ). Paratypes (2 exs.): "VENEZUELA: Trujillo/ Mcpo. Rafael Rangel/ La Gira, Quebrada la/ amarilla, 530 m./ 24-VIII-1997/ Det. M. García, 199", "Col/ M. Garcia/ J. Camacho/", "Paratipo q/ Notionotus/ edibethae/ Dcrip. M. García, 1997" (1 ex. SEMC); "VENEZUELA, Trujillo,/ Mcpo. Rafael Rangel, La/ Gira, Qda. La Amarilla,/ 520 m 20-22 / V / 1995/ Trampa Interceptación/", "Colectores:/ J. Camacho/ M. García/", "Paratipo ठ / Notionotus/ edibethae/ Dcrip. M. García, 1997" (1 ex., SEMC). The labeled holotype is an undissected male with the aedeagus visible and still attached to the abdomen. We also examined a permanent genitalia slide that had been presumed to be from holotype and is labeled as this species.

## Notionotus nucleus Perkins, 1979: 308. syn. nov.

Type material examined. Holotype (male): "GUATE., Alta Verapaz/5 mi. W. La Tinta/small tropical brook/6-VI-1974, ME\&PD Perkins", "Type No/76325/U S N M", "HOLOTYPE/Notionotus/nucleus/P.D. Perkins" (USNM).


Figure 6. Aedeagi of Notionotus tricarinatus A $N$. tricarinatus (holotype) B $N$. nucleus (holotype) C $N$. perijanus (holotype) D N. edibethae (holotype) E specimen from Barinas State, Venezuela $\mathbf{F}$ specimen from Portuguesa State, Venezuela G specimen from Aragua State, Venezuela H specimen from Zulia State, Venezuela I specimen from Costa Rica.

Notionotus perijanus García, 2000: 252. syn. nov.

Type material examined. Holotype (male): [only the permanent slide mount of the aedeagus was examined] (MALUZ). As this species was described only from one male and one female specimen, we presume this slide is of the holotype specimen (Fig. 6C).

Additional material examined (173 exs.). Costa Rica: Cartago Province: Tapanti National Park, Building by Río Villegas, 29.v.2006, leg. A.E.Z. Short, HG-vapor light, AS-06-066 (5 exs., SEMC, INBio, including DNA voucher SLE2397). Panama: Panama province: Barrio Colorado, $9^{\circ} 11^{\prime} \mathrm{N}, 79^{\circ} 51^{\prime} \mathrm{W}, 40 \mathrm{~m}, 22-25-\mathrm{VI}-2000$, leg. S. Chatzimanolis, flight intercept trap, PAN1C00 022 (3 exs., SEMC); same data except PAN1C00 024 (5 exs., SEMC); same data except PAN1C00 025 (2 exs., SEMC); same data except PAN1C00 033 (1 ex., SEMC); same data except PAN1C00 0234 (1 ex., SEMC); same data except PAN1C00 014 (1 ex., SEMC); same data except 07-VI1994, leg. D. Banks ( 2 exs., SEMC); same data except 08-VIII-1994, leg. D. Banks (1 ex., SEMC), same data except 08-VIII-1994, leg. D. Banks (1 ex., SEMC); same data except 04-VIII-1994, leg. D. Banks (1 ex., SEMC); same data except 01-VIII-1994, leg. D. Banks (1 ex., SEMC); Old plantation Rd. 6.9 km S Gamboa, $09^{\circ} 05^{\prime} \mathrm{N}, 79^{\circ} 40^{\prime} \mathrm{W}, 80$ m, 04-07-VI-1995, leg. J, Ashe \& R. Brooks, \#137 flight intercept trap (1 ex., SEMC), same data except 07-22-VI-1995, \#266 flight intercept trap (1 ex., SEMC); Colón, Parque Nacional Soberanía, Pipeline Rd km 6.1, $09^{\circ} 07^{\prime} \mathrm{N}, 79^{\circ} 45^{\prime} \mathrm{W}, 40 \mathrm{~m}, 07-21-\mathrm{VI}-$ 1995, leg. J, Ashe \& R. Brooks, \#265 flight intercept trap (2 exs., SEMC); Colón, Escobal \& Piña Rds, 14 km N jct. 02-11-VI-1996, leg. J, Ashe \& R. Brooks flight intercept trap PAN1AB96 181B (1 ex., SEMC). Venezuela: Aragua State: Henri Pittier Natural Park Río Cumboto, $10.39376^{\circ}$ N, $67.79597^{\circ} \mathrm{W}, 130 \mathrm{~m}, 4 . \mathrm{i} .2009$, leg. Short, García \& Miller, river side pools, VZ09-0104-02B (18 exs., SEMC, including DNA voucher SLE2381); Río La Trilla $10.37319^{\circ} \mathrm{N}, 67.74250^{\circ} \mathrm{W}, 295 \mathrm{~m}$, leg. Short, García \& Miller, pools, VZ09-0104-01A (1 ex., SEMC). Barinas State: Río Santa Barbara, E. Santa Barbara. $7^{\circ} 50.028 \mathrm{~N}, 71^{\circ} 11.188 \mathrm{~W}, 177 \mathrm{~m}, 26 . \mathrm{i} .2012$, leg. Short, Arias, Gustafson, sandy sidepool in floodplain, VZ12-0126-01B, (1 ex., SEMC). Portuguesa State: Trib. of Río Guanare, S. Biscucuy, $9^{\circ} 14.457^{\prime} \mathrm{N}, 69^{\circ} 55.994^{\prime} \mathrm{W}, 370 \mathrm{~m}, 19 . \mathrm{i} .2009$, leg. Short, García \& Miller, gravel stream, VZ09-0119-03X (15 exs., SEMC, including DNA vouchers SLE2391 and SLE2392). Zulia State: Perijá Natural Park Tukuko: Río Manantial, $9^{\circ} 50.490^{\prime} \mathrm{N}, 72^{\circ} 49.310^{\prime} \mathrm{W}, 270 \mathrm{~m}, 29 . \mathrm{i} .2009$, leg. Short, García \& Camacho, gravel margin, VZ09-0129-01A (91 exs., MIZA, SEMC, including DNA vouchers SLE1112 and SLE2371); same data except 29.i.2009, leg. Short, García \& Miller, detrital pool, VZ09-0129-01B (1 ex., SEMC); same data except 22.ix.2007, leg. A.E.Z. Short, rock pools/margin, AS-07-020b (8 exs., SEMC); Toromo, $10^{\circ} 03.058^{\prime} \mathrm{N}, 72^{\circ} 49.974^{\prime} \mathrm{W}, 435$ m, 31.xii.2005, leg. A.E.Z. Short, small stream and seep, AS-06-001 (6 exs., SEMC); same data except 28.i.2009, detrital pool, VZ09-0128-01A (3 exs., SEMC).

Differential diagnosis. See differential diagnosis for Notionotus mexicanus.
Description. Size and form: Body length $1.6-1.9 \mathrm{~mm}$. Body form elongate oval, strongly convex in lateral view (Fig. 3A). Color and punctation: Dorsally yellow, head mostly pale brown or yellow, frons brown or dark brown; pronotum paler than elytra, with two small black round spots along posterior margin (Fig. 3A, D, G). Ventrally brown; maxillary palps, mouthparts, antennae yellow (antennal club slightly darker), legs pale brown (Fig. 3B, E, H). Clypeus and labrum with dense, fine, and weakly ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures
separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp and broadening posteriorly almost to the end, the point where the two ridges merged rounded and obtuse (e.g., Fig. 10A, B); elevation concave in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters (Fig. 3B, E, H). Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 6A-I) basal piece $1.2 \times$ the length of a paramere. Parameres broad, base wider than the base of the median lobe, outer and inner margins convex, pinched at the apex, broad and rounded apex. Length of the median lobe can vary (median lobe as long as the parameres (Fig. 6F, I), slightly shorter (Fig. 6A, B) or longer than the parameres (Fig. 6H), median lobe with triangular shape, wide at base and gradually tapering to apical third, with rounded apex.

Distribution. Known from Guatemala, Costa Rica, Panama, and Venezuela (Fig. 14).
Life history. This species is found along the margins and in leaf packs of streams in the mountains and foothills of the Northern Andes and Central America. It prefers gravelly or rocky streams, especially in the foothills where it may sometimes be abundant (Fig. 11C, D).

Remarks. We examined more than 155 specimens from a dozen localities of this species from Guatemala to several chains of the Venezuelan Andes. Although there are subtle variations in the apex of the aedeagal parameres, this variation is relatively small and not correlated to geography, other morphological characters, or molecular data. These subtle variations in paramere shape likely explain why this species has been described four times, once each from Guatemala and Panama (Perkins 1979) and twice from Venezuela (García 2000). Three of these four species were described from single collecting events. However, with significantly more material available to us for this study from a range of additional localities, it is apparent these differences in paramere shape are more easily considered as intraspecific variation in a widespread, common species than as indicative of species boundaries. This hypothesis is also supported by available DNA evidence: we sequenced specimens from Costa Rica, the Serranía de Perijá, the Mérida Andes, as well as the Coastal mountains of Venezuela. All specimens form a clade (Fig. 1) with a maximum pairwise divergence in COI of $3.9 \%$ (although we only had COI data from specimens from several Venezuelan localities). However, $28 S$ sequence data from specimens from Venezuela and Costa Rica are identical, further supporting the concept of a single, widespread species. In addition, specimens throughout its range were found in very similar habitats: leaf packs or detrital margins of streams with a gravel or rocky substrate

As both $N$. nucleus and $N$. tricarinatus were proposed in the same work (Perkins 1979), we use our authority as first revisors (ICZN article 24.2.2) to give precedence to $N$. tricarinatus as the valid name for this species.

## Notionotus vatius sp. nov.

http://zoobank.org/CD665789-B94F-4C19-AAE1-83B7ACCE9413
Figs 4H, 7C, 13B, 14
Type material. Holotype (male): "BRAZIL: Mato Grosso do Sul/ -20.72281 ${ }^{\circ}$ $-55.69127^{\circ} ; 225 \mathrm{~m} / A q u i d a u a n a(c .27 \mathrm{~km} \mathrm{~S}$ ) on/MS-174; leg. Hamada \& team;/27. vi.2018; seepage \& debris nr./stream margin; BR18-0627-01E" (INPA). Paratypes (38 exs.): Brazil: Mato Grosso do Sul State: Same data as holotype (19 exs., INPA, SEMC, including DNA Voucher 2324); Aquidauana on plateau (ca. 15 km E), -20.4509, -55.6218, $380 \mathrm{~m}, 22 . v i .2018$, leg. Hamada \& team, detritus and washing roots at margin on rock, BR18-0622-03D (6 exs., SEMC, including DNA voucher SLE2327); Corumbá (ca. 27 km SE ) by mountains, -19.28382 , $-57.57506,146 \mathrm{~m}$, 24.vi.2018, leg. Hamada \& team, stream margin and leaf packs, BR18-0624-01A (1 ex., SEMC); Rio Bento Gomes (Pantanal), Campo Allegre I, $15^{\circ} 45^{\prime} \mathrm{S}, 56^{\circ} 33^{\prime} \mathrm{W}$, 1993-1994, leg. E. Stuhr, spring-fed brook, (10 exs., NMW, SEMC). Bahia: Morro do Chapéu, Cachoeira Domingos Lopes, -11.55965, -40.90635, 675 m, 24.ii.2018, leg. Benetti \& team, blackwater river and waterfall, BR18-0224-02A (1 ex., SEMC,


Figure 7. Aedeagi of Notionotus liparus species group A $N$. liparus (non-type specimen) B $N$. mexicanus (holotype) C $N$. vatius (holotype) D N. giraldoi (holotype).

DNA Voucher SLE2104); Livramento de Nossa Senhora, NE on BR-148, -13.6212, -41.81908, 536 m, 27.ii.2018, leg. Benetti $\&$ team, stream margins, BR18-0227-01A (1 ex., SEMC, DNA Voucher SLE2385).

Differential diagnosis. Among the species of liparus group, Notionotus vatius can be recognized by the brown dorsal coloration, and quite unique color pattern of the head frons and medial region of the clypeus dark brown, lateral side of the clypeus pale brown. In addition, the shape of aedeagus, especially the apex of the parameres is broad, blunt, and pointing slightly outwards, and the gonopore has rounded shape and it is situated at midlength of median lobe.

Description. Size and form: Body length 1.7-2.3 mm. Body form elongate oval, strongly convex in lateral view (Fig. 4H). Color and punctation: Dorsally brown, head mostly brown, frons and medial region of the clypeus dark brown, lateral side of the clypeus pale brown; pronotum pale brown with two small black round spots along posterior margin, elytra dark brown (Fig. 4H). Ventrally dark brown; maxilla, maxillary palps, antennae (antennal club slightly darker) yellow, legs pale brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp, the point where the two ridges merged rounded and obtuse (e.g., Fig. 10B); elevation flat in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area, medial region patch drop-shaped; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 7C) with basal piece nearly the same length as a paramere. Base of the parameres slightly narrower than the base of the median lobe; outer and inner margins sinuate; apex of parameres wide and blunt, pointing outwards. Median lobe of the same length as the parameres, wide at basal region, narrowing in the midlength, then widening along at apical third, apex rounded; gonopore with rounded shape and situated at midlength of median lobe.

Etymology. The name is derived from the Latin word vatius meaning bent outwards, after the form of the parameres slightly pointing outwards of the aedeagus.

Distribution. This species is known from several localities in Bahia and Mato Grosso do Sul States in Brazil (Fig. 14).

Life history. This species was collected on seepages and along the margins of rocky streams (Fig. 13B).

Remarks. Although the known localities of this species are widely dispersed in Brazil, they are from similar habitats at similar elevations on the Brazilian Shield. Moreover, the specimens from Bahia and Mato Grosso do Sul states are less than 3\% divergent in uncorrected pairwise distances in COI.

## Notionotus lohezi species group

Diagnosis. The lohezi species group can be distinguishable by the presence of three ridges in the elevation of the mesoventrite, two transverse ridge and one longitudinal (Fig. 10C, D); the basal piece is shorter than the parameres, the length of the median lobe is shorter than the parameres.

## Notionotus bicolor sp. nov.

http://zoobank.org/7954CB9D-66E9-492F-865E-90BA4991ADF3
Figs 4A, 8A, 10D, 12D, 15
Type material examined. Holotype (male): "SURINAME: Sipaliwini District/ $4.42313^{\circ}$ N, $57.19198^{\circ}$ W, $104 \mathrm{~m} /$ Kabalebo Nature Resort/ Moi Moi Creek; 10-14. iii.2019/ leg Short \& Baca small seeps/ SR19-0310-01F" (NZCS). Paratypes (88 exs.): Suriname: Sipaliwini District: Same data as holotype (8 exs., SEMC); Kabalebo Nature Resort: Charlie Falls, $4.38302^{\circ} \mathrm{N}, 57.21161^{\circ} \mathrm{W}, 174 \mathrm{~m}, 11 . \mathrm{iii} .2019$, leg. Short, Baca and class, rocks pools in creekbed, SR19-0311-01A (55 exs., NZCS, SEMC, including DNA vouchers SLE1810 and SLE2120); same data except leg. Short, Seepage, SR19-0311-01B (14 exs., SEMC); same data except leg. Baca, side pools in gravel in creekbed, SR19-0311-01E (2 exs., SEMC); Kabalebo Nature Resort Moi Moi creek, $4.42313^{\circ}$ N, $57.19198^{\circ} \mathrm{W}, 104 \mathrm{~m}, 10-14 . i i i .2019$ leg. Short, Baca and class, margin of seepage, SR19-0310-01E ( 2 exs., SEMC); same data except leg. Baca, margin of stream pool with root mats, SR19-0310-01L (1 ex., SEMC); same data except leg. Short and class, small trickle on rocks w/detritus, SR19-0310-01M (4 exs., SEMC); CSNR: Tafelberg Summit, near Caiman Creek Camp, $3^{\circ} 53.942^{\prime} \mathrm{N}, 56^{\circ} .10 .849^{\prime} \mathrm{W}$, 733 m, 19.viii.2013, leg. Short and Bloom, margins and leaf packs in Caiman Creek, SR13-0819-05D (2 exs., SEMC).

Differential diagnosis. Notionotus bicolor shares the bicolorous dorsal coloration that can also be observed in $N$. garciae, $N$. patamona, and $N$. lohezi. It can be recognized by the shape of the aedeagus, the median lobe is much shorter than in $N$. insignitus (Fig. 8B) and N. patamona (Fig. 8D), and slightly longer than in N. lohezi (Fig. $8 \mathrm{~F})$. The margins of the median lobe are sinuate with a widening in the middle length, this differs in the other species where the margins are slightly straight. Moreover, the apex is slightly narrow and rounded, this differs in $N$. insignitus with acute apex, $N$. patamona with rounded and wide apex and $N$. lohezi with blunt apex.

Description. Size and form: Body length $1.7-2 \mathrm{~mm}$. Body form elongate oval, moderately convex in lateral view (Fig. 4A). Color and punctation: Dorsally bicolor, head brown, frons dark brown; pronotum yellow with two small black round spots along posterior margin; elytra dark brown, elytra margins paler (Fig. 4A). Ventrally brown; maxillary palps, mouthparts, and antennae yellow (antennal club slightly darker), pro legs yellow, meso and meta legs pale brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their


Figure 8. Aedeagi of Notionotus lohezi species group A N. bicolor (holotype) B N. insignitus (holotype) C $N$. brunbadius (holotype) D N. patamona (holotype) E N. retusus (holotype) F N. lohezi (paratype) G $N$. juma (holotype) H $N$. garciae (holotype).
width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3-4 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides
concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (Fig. 10D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and sparse pubescence along the entire basal posterior margin. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8A) with basal piece $0.7 \times$ the length of a paramere. Base of the parameres slightly narrower than the base of the median lobe; outer margin convex, inner margins along basal two-thirds slightly convex, apical third straight; apex of parameres slightly acute, pointing inwards. Median lobe shorter than the parameres, wide at basal region, narrowing in the midlength, then sinuate, apex slightly acute; gonopore situated at the apex of median lobe.

Etymology. The name derived from the Latin words $b i$ meaning two and color meaning hue, referring to the dorsal coloration of the species. This species has yellow coloration in the pronotum and black in the elytra.

Distribution. Known from two localities in central Suriname: Kabalebo and the summit of Tafelberg Tepui (Fig. 15).

Life history. At Kabalebo, this species was collected in several streams, including both along the margin, rock pools with detritus in the creek bed, and in seepage habitats (Fig. 12D). At Tafelberg, the species was collected along a rocky creek margin with detritus.

## Notionotus bifidus sp. nov.

http://zoobank.org/AEAADA10-0A40-4AB9-B48E-FC27A126792B
Figs 4J, 9D, 12A, 14

Type material. Holotype (male): "VENEZUELA: Amazonas State/ 5²33.207'N, $67^{\circ} 36.922^{\prime}$ W, $125 \mathrm{~m} /$ Tobogan de la Selva; 8.viii.2008/leg. A. Short, M. García, L. Joly/ AS-08-080b; old "tobogancito"/on seepage area w/detritus" (MIZA). Paratypes (54 exs.): Venezuela: Amazonas State: Same data as holotype (18 exs., SEMC, including DNA voucher SLE1113); same data except 14.i.2009, leg. A. Short, clumps of wet leaves on rock, VZ09-0114-01D (15 exs., SEMC, including DNA voucher SLE2369); same date except 14.i.2009, leg. K. Miller, detrital rock pools, VZ09-011401E (1 ex., SEMC); same data except 14.i.2009, leg. Short \& Miller partly shaded wet rock w/algae, VZ09-0114-01G (20 exs., MIZA, SEMC).

Differential diagnosis. Notionotus bifidus can be separated from all other species of lohezi group by being the only species in the group that present uniformly dorsal yellow coloration (Fig. 4J), the rectangular shape of the median lobe and its bifurcation at the apex, and the abrupt tapering of the parameres along apical third (Fig. 9D).

Description. Size and form: Body length $1.5-1.7 \mathrm{~mm}$. Body form elongate oval, moderately convex in lateral view (Fig. 4J). Color and punctation: Dorsally yellow,


Figure 9. Aedeagi of Notionotus spp. A-C N. rosalesi A drawing from Spangler (1972) B holotype $\mathbf{C}$ specimen from Trinidad D $N$. bifidus $\mathbf{E} N$. parvus $\mathbf{F} N$. peruensis.
head mostly dark brown, frons and medial region of the clypeus dark brown, lateral sides of clypeus yellow; pronotum yellow with two small dark brown round spots along posterior margin; elytra yellow (Fig. 4J). Ventrally dark brown; maxillary palps, mouthparts, and antennae yellow. Pro legs yellow, meso and meta legs pale brown.

Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with broad glabrous patch on the medial and


Figure 10. Mesoventral process of Notionotus spp. A $N$. dilucidus B $N$. tricarinatus $\mathbf{C} N$. insignitus D N. bicolor. Red marks pointing to transverse ridge in A-D blue marks pointing to second transverse ridge in C, D.


Figure II. Habitat of Notionotus spp. in the Andean region of Venezuela A Venezuela, Barinas State, seepage habitat of $N$. liparus (collecting event VZ12-0124-02A) B Venezuela: Aragua, Henri Pittier National Park, seepage habitat of $N$. liparus, (collecting event AS-06-023) C Venezuela: Zulia, Rio Tukuko, habitat of $N$. tricarinatus (collecting event VZ09-0129-01A) D Venezuela, Portuguesa, tributary of the Rio Guanare, habitat of $N$. tricarinatus (collecting event VZ09-0119-03X).
posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of posterior margin, then apical half of posterior margin with sparse setae. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 9D) with basal piece $0.6 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; outer margins straight along basal two-thirds, then narrowing abruptly along apical third, inner margins convex along basal twothirds and then apically slightly concave; apex of parameres rounded. Median lobe shorter than the parameres, approximately rectangular, narrow along apical fifth, apex bifurcated; gonopore drop-shaped and situated at apical fourth of median lobe.

Etymology. The specific name comes from the Latin word bifidus meaning split into two parts, after the form of the median lobe "bifurcated apically" of the aedeagus.

Distribution. Known only from the type locality in Venezuela (Fig. 14).
Life history. This species was collected in seepage habitats that were covered with algae and detritus (Fig. 12A).

## Notionotus brunbadius sp. nov.

http://zoobank.org/23615726-EDE5-40AD-B408-8ED24AC30EDC
Figs 4C, 8C, 12C, 15
Type material. Holotype (male): "BRAZIL: Amazonas, Manaus/-2.93079, -59.97514, $75 \mathrm{~m} /$ Ducke Reserve/ leg. Short \& team; stream margin/ \& assoc. backwater swampy area/ 9-10.vi.2018; BR18-0609-03A", "DNA VOUCHER/ Extraction \#/ SLE-1553" (INPA). Paratypes (4 exs.): Brazil: Amazonas State: Same data as holotype (2 exs., SEMC); same data except Igarape Barro Branco, muddy pools in swampy area adjacent to stream, BR18-0609-02B (1 ex., SEMC, DNA voucher SLE2102); same data except by unnamed stream, water in palm fronds, BR18-0609-03B (1 ex., SEMC).

Differential diagnosis. This species has a particular coloration pattern, dark reddish brown, which makes it easily distinguishable among the other species of the lohezi group. The shape of the parameres is slightly similar to $N$. lohezi (Fig. 8F) but the apex is less acute and bend much less inward. It can be separated by the triangular shape of the median lobe and acute apex (Fig. 8C).

Description. Size and form: Body length $1.7-1.9 \mathrm{~mm}$. Body form elongate oval, moderately convex in lateral view (Fig. 4C). Color and punctation: Dorsally dark reddish brown, lateral margins of clypeus, pronotum and elytra yellow brown (Fig. 4C). Ventrally reddish brown, except for black abdominal ventrites; maxillary palps, mouth parts and antennae yellow (antennal club slightly darker). Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed, and sparser than on head (punctures separated by 3-4 $\times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along three-quarters of the anterior basal margin and sparse pubescence along the posterior basal margin. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8C) with basal piece $0.7 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; outer margins slightly convex, inner margins straight; apex of parameres acute and pointing outwards. Median lobe shorter than the parameres, approximately triangular, wide at the basal region, then slightly narrowing to the apex, apex acute, gonopore oval in shape and situated at the apex of the median lobe.

Etymology. The name is a combination of two Latin words brun meaning dark and badius meaning reddish brown, highlighting the distinguishable dark reddish brown color of this species.

Distribution. Known only from the type locality near Manaus, Brazil (Fig. 15).


Figure 12. Habitat of Notionotus spp. A Venezuela: Amazonas State, type locality and habitat of $N$. bifidus (collecting event AS-08-080b) B Venezuela: Bolívar State: Type locality and habitat of $N$. insignitus (collecting event AS-08-058) C Guyana: Region 9: type locality and habitat of $N$. patamona (collecting event GY13-0318-01C) D Suriname: Kabalebo, habitat and type locality for $N$. bicolor (collecting event SR19-0310-01F).

Life history. The only known specimens were collected along the margin of a stream (Fig. 12C).

## Notionotus garciae sp. nov.

http://zoobank.org/10EFC8E2-7C02-4F44-994B-54B3ABB388C8
Figs 4G, 8H, 13D, 15
Type material. Holotype (male): "BRAZIL: Amazonas: Manaus/ -2.93079, $-59.97514,75 \mathrm{~m} /$ Ducke Reserve, stream nr./ transect trail; leg. Short \& team/ stream margins \& leaf packs/ 9.vi.2018; BR18-0609-01A" (INPA). Paratypes (8 exs.): BrazIL: Amazonas State: Same data as holotype ( 3 exs., SEMC); same data except 9-10. vi.2018, stream margin and associate backwater swampy area, BR18-0609-03A ( 1 ex., SEMC); same data except Igarape Barro Branco, 6.vi.2018, shallow pools, BR18-0606-02D (1 ex., SEMC); same data except Igarape Barro Branco, margin of stream, 9.vi.2018, BR18-0609-02A (2 exs., SEMC); Presidente Figueiredo (ca. 57 km E) on AM-240, -1.98826, -59.51618, 80 m , airport stream, 19.vi.2018, leg. Short, margin of creek, (1 ex., SEMC, DNA voucher SLE1900).

Differential diagnosis. Notionotus garciae is very similar to $N . j u m a$ in the bicolor dorsal coloration, the shape of the elevation of the mesoventrite and the pubescent area of the metafemora. It can be distinguished by the punctation of the pronotum, the elytra shallowly marked (in $N$. juma is moderately impressed, Fig. 8 G ), and the distinctive rounded and broad apex of the median lobe (Fig. 8H).


Figure I 3. Habitat of Notionotus spp. A type locality and habitat of N. giraldoi (collecting event BR18-0710-02A) B type locality and habitat of $N$. vatius (collecting event BR18-0627-01A) C type locality and habitat of N. brunbadius (collecting event BR18-0609-03A) D type locality and habitat of N. garciae (collecting event BR18-0609-01A) E type locality and habitat of $N$. juma (collecting event BR18-0609-02B) F type locality and habitat of $N$. retusus (collecting event GY14-0311-02A).

Description. Size and form: Body length $1.7-1.8 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4G). Color and punctation: Dorsally bicolor, head mostly brown, frons dark brown, posteromedial margin of the clypeus pale brown and lateral sides and anterior margin yellow; pronotum yellow with two small black round spots along posterior margin; elytra brown, lateral and posterior side of the elytra yellow (Fig. 4G). Ventrally dark brown; maxillary palps, mouthparts, and antennae yellow. Pro and meso legs yellow, meta legs pale brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation weakly impressed, fine, and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and sparse pubescence along the apical posterior margin. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8 H ) with basal piece $0.4 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; outer margins nearly convex along basal two-thirds, then curved inwards along apex, inner margins nearly straight; apex of parameres acute. Median lobe shorter than the parameres, wide at basal region, narrowing in the midlength and then slightly broadening to apex, apex rounded and wide; gonopore ovate in shape and situated at apical fourth of median lobe.

Etymology. This species is named after Andrea Lorena García Hernández curator at the Colección de Insectos de la Universidad del Quindío (CIUQ) in recognition to her passion and contribution of the knowledge of the insects, specially hydrophilids in Colombia.

Distribution. This species is known from several collecting events at the Ducke Reserve near Manaus, Brazil (Fig. 15).

Life history. This species was collected along the margins of small streams in dense forest (Fig. 13D).

## Notionotus insignitus sp. nov.

http://zoobank.org/9B5D9358-AC4A-49D1-B9F6-44490E2FAA7C
Figs 4D, 8B, 10C, 12B, 15
Type material. Holotype (male): "VENEZUELA: Bolívar State/ 6²'10.5"N, $61^{\circ} 23^{\prime} 57.8^{\prime \prime W}$ W, $630 \mathrm{~m} /$ Along La Escalera; 31.vii.2008/ leg. A. Short, M. García, L. Joly/ AS-08-058; rocky stream" (MIZA). Paratypes (14 exs.): Venezuela: Bolívar State: same data as holotype ( 14 exs., SEMC, including DNA voucher SLE1115).

Differential diagnosis. See differential diagnosis for Notionotus bicolor. In addition, $N$. insignitus has a particular and unique color pattern in the elytra among the other congeners. The elytra are mostly dark brown, the posterior margins yellow and in the middle region has a characteristic yellow spot.

Description. Size and form: Body length $1.6-1.8 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4D). Color and punctation: Dorsally bicolor, head mostly brown, frons and middle region of the clypeus dark brown, lateral sides of the clypeus pale brown; pronotum yellow with two small black round spots along posterior margin; elytra dark brown, with a yellow spot on the anterior third of the elytra, lateral and posterior margins of the elytra yellow (Fig. 4D). Ventrally dark brown; maxillary palps, mouthparts, and antennae yellow (antennal club slightly darker). Pro and meso legs yellow, meta legs pale brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (Fig. 10C); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of the posterior margin, then anterior half of posterior margin with sparse setae. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8B) with basal piece $0.6 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; outer margin straight along basal two-thirds, then apically slightly convex, inner margins convex along basal two-thirds and then apically straight; basal two-thirds of the parameres broad then apical third narrower; apex of parameres rounded and pointing inwards. Median lobe shorter than the parameres, approximately triangular, with acute apex; gonopore triangular and situated at apex of median lobe.

Etymology. The specific name comes from the Latin word insignitus meaning marked and refers to the distinctive yellow spot in the elytra of this species.

Distribution. Known only from the type locality in southeastern Venezuela (Fig. 15).
Life history. The only known series of this species was collected along the margin of a forested rocky stream (Fig. 12B).

## Notionotus juma sp. nov.

http://zoobank.org/DA88A467-D7C8-4AE1-A8E2-4033D8AF3364
Figs 4E, 8G, 13E, 15

Type material. Holotype (male): "Brazil: Amazonas: Manaus/ -2.93079, -59.97514, 75 m/ Ducke Reserve, Igarape Barro/ Branco; Short \& team; muddy/ pools in swampy


Figure 14. Distribution map of Notionotus spp.
area by stream/ 9.vi.2018; BR18-0609-02B" (INPA). Paratypes (16 exs.): Brazil: Amazonas State: Same data as holotype ( 13 exs., SEMC); same data except stream margins, 6.vi.2018, BR18-0606-02B (2 exs., SEMC, including DNA voucher SLE2100); Novo Airão Municipio, $-2.68396,-60.93840$, leg. Benetti, 9 .vi.2017, densely vegetated margin of blackwater creek, BR17-0609-04A (1 ex., SEMC, DNA voucher SLE1269).

Differential diagnosis. See differential diagnosis for Notionotus garciae. In addition, the aedeagus of $N$. juma has an emargination in the apex of the medium lobe, being a particular feature among the species of the lohezi group.

Description. Size and form: Body length $1.6-1.9 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4E). Color and punctation: Dorsally bicolor, head pale brown, pronotum yellow with two small black round spots along posterior margin; elytra dark brown (Fig. 4E). Ventrally brown; maxillary palps, mouthparts, and antennae yellow (antennal club slightly darker). Pro and meso legs yellow, meta legs pale brown. Clypeus and labrum with dense, coarse, and moderately impressed ground punctation (punctures separated by $2 \times$ their width); elytra and pronotum ground punctation coarse, moderately impressed and less dense than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8G) with basal piece nearly the same length as a paramere. Base of the parameres as wide as the base of the median lobe; outer margins convex, then curved inwards along apex, inner margins convex along basal two-thirds, and concave at apical third; apex of parameres rounded. Median lobe shorter than the parameres, wide at basal region, slightly narrowing in the midlength, apex wide and emarginate medially; gonopore with an oval shape and situated at apex of the median lobe.

Etymology. This species is named after the Juma, an indigenous tribe located in the Açuâ River, in the southern part of the state of Amazonas-Brazil.

Distribution. Known only from the Ducke Reserve near Manaus, Brazil (Fig. 15).
Life history. Specimens were collected in two habitats at the same forest reserve: in detrital pools in an area of shallowly flooded forest with detrital and mud substrate (Fig. 13E), and along the margins of a small stream.

## Notionotus lohezi Queney, 2010

Figs 8F, 15
Notionotus lohezi Queney, 2010: 135.
Type material examined. Paratype (male): " ${ }^{\lambda}$ ", "Notionotus lohezil n. sp. PARATYPE/ P. QUENEY descr. 2010", "Guyane: Régina,/ Patawa, crique en/ forêt, 170 m ,/ 13-IX-2009,/ leg. P. Queney" (SEMC). Paratypes (5 exs.): same data as the paratype dissected (5 exs., SEMC).


Figure 15. Distribution map of Notionotus spp.

Additional material examined ( 2 exs.). French Guiana: Savane Roche Virginie, near RN 2, 4.1883, -52.13982, 64 m, Crique Chauve-souris, leg. Short, 10.iii.2020, solid granite substrate, detritus along margins on granite, FG20-0310-01A (2 exs., SEMC, including DNA vouchers SLE2337 and 2387).

Differential diagnosis. See differential diagnosis for Notionotus bicolor.
Description. Size and form: Body length $1.6-1.8 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (e.g., Fig. 4A). Color and punctation: Dorsally bicolor, head bicolor, frons dark brown, clypeus yellow; pronotum yellow with two small black round spots along posterior margin; elytra dark brown, elytra margins paler (e.g., Fig. 4A). Ventrally dark brown; maxillary palps, mouthparts, and antennae yellow (antennal club slightly darker), legs brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of the posterior margin, then apical half of the posterior margin with sparse setae. Abdomen: abdominal ventrites very densely pubescent. Aedeagus (Fig. 8F) with basal piece $0.8 \times$ the length of the paramere. Base of the parameres slightly narrower than the base of the median lobe, inner margins straight, then convex reaching the apex, outer margins convex, and rounded apex pointing inwards. Median lobe shorter than the parameres, approximately rectangular with apex blunt; gonopore situated at the apex of the median lobe.

Distribution. This species is only known from a few localities in French Guiana (Fig. 15).

Life history. This species was collected in rocky streams with detritus along margins.

## Notionotus parvus sp. nov.

http://zoobank.org/08140DE3-8B15-42F9-ADA9-3A1F952C1D3D
Figs 4F, 9E, 15

Notionotus sp. 2 in Short, 2013: 88.

Type material. Holotype (male): "SURINAME: Sipaliwini District/ N 2.97731, W $55.38500^{\circ}$, $200 \mathrm{~m} /$ Camp 4 (low), Kasikasima; sandy/ stream on trail to METS camp/ 20.iii.2012; SR12-0320-02A/ leg. A. Short; 2012 CI-RAP Survey" (NZCS). Paratypes (2 exs.): SURINAME: same data as holotype ( 2 exs., SEMC, including DNA voucher SLE2388).

Differential diagnosis. Notionotus parvus can be recognized by the pale reddish yellow color in the head and pronotum and reddish brown in the elytra (Fig. 4F). Furthermore, the aedeagus is quite unique, parameres tubular in shape with lanceolate appendages in the outer margin which are shorter than the length of the parameres, and gonopore with crown-shape located in the apical region of the median lobe (Fig. 9E).

Description. Size and form: Body length $1.7-1.9 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4F). Color and punctation: Dorsally reddish brown, head and pronotum pale reddish yellow; pronotum with two small black round spots along posterior margin; elytra dark reddish brown (Fig. 4F). Ventrally reddish brown; maxillary palps and mouthparts light yellow reddish, and antennae yellow (antennal club slightly darker). Pro and meso legs yellow, meta legs pale reddish brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation slightly convex in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and sparse pubescence along the apical posterior margin. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 9E) with basal piece $0.5 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; parameres tubular shape, outer margins sinuate and covered by lanceolate appendages that cover three-quarters of the margin; inner margins sinuate, apex of the parameres rounded. Median lobe shorter than the parameres, wide at basal region, narrowing apically, apex rounded and wide; gonopore crown-shaped and situated at apex of median lobe.

Etymology. The species name is derived from the Latin word parvus meaning little or small in reference to the small aedeagus size of this species.

Distribution. Only known from the type locality in southern Suriname (Fig. 15).
Life history. This species was collected along the margins of a small, sandy-bottomed stream.

## Notionotus patamona sp. nov.

http://zoobank.org/B8AC3A70-8620-44E5-8665-709DCF4307AA
Figs 4B, 8D, 12C, 15
Type material. Holotype (male): "GUYANA: Region XIII [!sic: Region 8]/ $5^{\circ} 18.264^{\prime} \mathrm{N}, 59^{\circ} 50.257^{\prime} \mathrm{W} ; 687 \mathrm{~m} /$ Ayanganna Airstrip; trail from air-/ strip to Ayanganna; seepage area; over rocks in forest flowing into/ stream; leg. A. Short; 18.iii.2014/

GY14-0318-01C" (CBDG). Paratype (12 exs.): Guyana: Region 8: Same data as holotype (8 exs., SEMC); Upper Potaro Camp I (ca. 7 km NW Chenapau), Potaro margin trail, $5^{\circ} 0.660^{\prime} \mathrm{N}, 59^{\circ} 38.283^{\prime} \mathrm{W}, 484 \mathrm{~m}, 11 . \mathrm{iii} .2014$, leg. Short, Baca, Salisbury and La Cruz, wet detritus in sandy area, GY14-0311-04A (1 ex., SEMC); top of falls on Potaro River, $5^{\circ} 0.730^{\prime}$ N, $59^{\circ} 38.965^{\prime}$ W, $585 \mathrm{~m}, 12 . \mathrm{iii} .2014$, leg. Short, Salisbury and La Cruz, seeps with roots and algae, GY14-0312-01B (1 ex., SEMC); stream near camp, $5^{\circ} 0.673^{\prime} \mathrm{N}, 59^{\circ} 38.358^{\prime} \mathrm{W}, 500 \mathrm{~m}, 14 . i i i .2014$, leg. Short, Salisbury and La Cruz, gravel/sandy stream w/ detritus, GY14-0314-01A (1 ex., SEMC); Kaieteur Natural Park, trail by guest house, $5^{\circ} 10.514^{\prime} \mathrm{N}, 59^{\circ} 28.970^{\prime} \mathrm{W}, 440 \mathrm{~m}, 21 . \mathrm{iii} .2014$, leg. Short, Salisbury and La Cruz, forest pools, GY14-0321-01B (1 ex., SEMC).

Differential diagnosis. See differential diagnosis for Notionotus bicolor.
Description. Size and form: Body length $1.6-1.8 \mathrm{~mm}$. Body form elongate oval, convex in lateral view (Fig. 4B). Color and punctation: Dorsally bicolor, head brown, frons dark brown, clypeus pale brown; pronotum yellow with two small black round spots along posterior margin; elytra dark brown, elytra margins paler (Fig. 4B). Ventrally brown; maxillary palps, mouthparts, antennae, and legs yellow. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of the posterior margin, then apical half of the posterior margin with sparse setae. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 4B) with basal piece $0.7 \times$ the length of a paramere. Base of the parameres narrower than the base of the median lobe; outer margins straight along basal two-thirds, then apically slightly convex, inner margins straight along basal two-thirds and then convex and tapering apically; apex of parameres rounded and pointing inwards. Median lobe shorter than the parameres, approximately triangular, gradually narrowing from the base, broad and rounded apex; gonopore oval-shaped and situated at apex of median lobe.

Etymology. This species is named after the Patamona, an indigenous tribe located in the mountainous region from which this species is known.

Distribution. Known from several closely situated localities in western Guyana (Fig. 15).

Life history. This species was collected at several a variety of stream-associated habitats, including along the margins of detritus and sandy-based streams, as well as in rock seepage habitats adjacent to streams (Fig. 12C).

## Notionotus retusus sp. nov.

http://zoobank.org/366A532F-DAAF-4521-9E0B-CD6AACA17266
Figs 8E, 13F, 15
Type material. Holotype (male): "GUYANA: Region XIII [!sic: Region 8]/ $5^{\circ} 0.730^{\prime} \mathrm{N}$, $59^{\circ} 38.965^{\prime} \mathrm{W}, 585 \mathrm{~m} /$ Upper Potaro Camp I (c. $7 \mathrm{~km} / \mathrm{NW}$ Chenapau), Ridge Trial/ leg. Short, Baca and Salisbury/ 11.iii.2014; GY14-0311-02A", "DNA VOUCHER/ Extraction \#/ SLE-2110" (CBDG). Paratype (1 ex.): Guyana: Region 8: Ayanganna Airstrip, $5^{\circ} 18.264^{\prime} \mathrm{N}, 59^{\circ} 50.257^{\prime} \mathrm{W}$; 687 m , trail from airstrip to Ayanganna, leg. A. Short, 18.iii.2014, seepage area over rocks in forest flowing into stream, GY14-031801C (1 ex., SEMC, DNA voucher 2372)

Differential diagnosis. The external characters of Notionotus retusus and N. lohezi are quite similar. The only way they can be separated is by the features of the aedeagus. In this species, the apex of the parameres is wide and rounded (acute in $N$. lohezi, Fig. 8 F ), the median lobe is much longer than in $N$. lohezi and the gonopore is elongated and oval (nearly rectangular in $N$. lohezi).

Description. Size and form: Body length 1.8 mm . Body form elongate oval, moderately convex in lateral view. Color and punctation: Dorsally bicolor, head mostly brown, frons brown, clypeus pale brown; pronotum yellow with two small black round spots along posterior margin; elytra dark brown. Ventrally brown; maxillary palps, mouthparts, antennae (antennal club slightly darker), legs brown. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed, and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, acute and pointing anteriorly. Elevation of mesoventrite with two transversal ridges, elevated medially, lateral sides concave; longitudinal ridge broad anteriorly and sharp posteriorly, the point where the three ridges merged wide and blunt (e.g., Fig. 10C, D); elevation flat in ventral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal one-quarter of posterior margin, then apical half of the posterior margin with sparse setae. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 8E) with basal piece $0.7 \times$ the length of a paramere. Base of the parameres nearly the same as the base of the median lobe; outer margins convex, inner margins nearly straight; apex of parameres broad, rounded and slightly pointing inwards. Median lobe shorter than the parameres, wide at basal region, narrowing in basal third, margins straight and apex rounded and nearly notched; gonopore with an oval shape and covering approximately two-thirds of the length of the median lobe.

Etymology. The specific name comes from the Latin word retusus meaning rounded and notched, after the form of the apex of the median lobe of the aedeagus.

Distribution. This species is only known from the type locality in western Guyana (Fig. 15).

Life history. This species was collected in detrital-filled pools in a shallow ravine with dense forest cover (Fig. 13F). Although the pools might not be considered a stream, the pools were part of a drainage network.

## Notionotus rosalesi species group

Diagnosis. The species of this group can be recognized by the unique dorsal coloration, being almost yellow with a wide brown band in the third anterior of the elytra (Fig. 2G), the elevation of the mesoventrite with one transversal and longitudinal ridge (e.g., Fig. 10B), the shape of the genitalia is quite distinct, being the only one within all Notionotus species of the Neotropical region with the apical third of the parameres membranous (Fig. 9A-C).

## Notionotus rosalesi Spangler, 1972

Figs 2G-I, 9A-C, 14

Notionotus rosalesi Spangler, 1972: 141

Type material examined. Holotype (male): "VENEZUELA/Arag., 10 Km S./Rancho Grande/II-14-1969/P.\&P. Spangler", "TypeNo/71950/U S N M", "HOLOTYPE/ Notionotus/rosalesi/P.J.Spangler" (USNM).

Additional material examined. Trinidad: Guanapo State: 4.1 km up Guanapo Valley, trib of Guanapo River, 460 ft , 11-VII-2005 (1 ex., SEMC); Verdant vale, Arima River, $10^{\circ} 42^{\prime} \mathrm{N}, 61^{\circ} 18^{\prime} \mathrm{W}, 570 \mathrm{ft}, 9-\mathrm{VII}-2005$ (1 ex., SEMC). Venezuela: Aragua: Rancho Grande Biol. Stn. 1150 m, $10^{\circ} 21^{\prime} \mathrm{N}, 67^{\circ} 41^{\prime} \mathrm{W}, 25-28$ II 1995, S. Marshall, yellow pan trap (1 ex., SEMC).

Differential diagnosis. Notionotus rosalesi can be distinguished by the wide brown band in the anterior third of the elytra, as well as, the unique shape of the genitalia, having many accessories at the base of the parameres, the apex of the parameres membranous and lanceolate.

Description. Size and form: Body length $1.8-1.9 \mathrm{~mm}$. Body form elongate oval, moderately convex in lateral view (Fig. 2G). Color and punctation: Dorsally bicolor, head mostly brown, frons brown, medially region of the clypeus pale brown with lateral margins yellow; pronotum yellow with two small black round spots along posterior margin; elytra yellow except by a brown wideband on the anterior third of the elytra (Fig. 2G). Ventrally brown; maxillary palps, mouthparts, antennae, and legs yellow (antennal club slightly darker) (Fig. 2H). Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $5 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing
setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge narrowed anteriorly and broadening posteriorly, the point where the two ridges merged acute (e.g., Fig. 10B); elevation concave in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area; anterior margin extending to mesoventrite elevation. Metafemora with dense hydrofuge pubescence along basal three-quarters of the anterior margin and along basal half of the posterior margin. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 9A-C) basal piece $0.4 \times$ the length of a paramere; broad parameres, base of the parameres much wider than the base of the median lobe, base of the parameres with two accessories with ovate shape, outer margins of parameres strongly sinuate, inner margins slightly sinuate, with membranous acuminate apex, bending outwards; median lobe shorter than the parameres, approximately triangular, with apex rounded.

Distribution. Originally described from the Venezuelan states of Aragua and Barinas (Spangler 1972), it was later reported from the states of Trujillo and Falcón (García 2000). Here we report it for the first time from Trinidad (Fig. 14).

Life history. Although specific habitat information is limited, all specimens were collected in association with streams. Spangler (1972) characterized this species as a hygropetric specialist, although not all specimens known at that time were from seepages (the others were from a stream pool that "was in the bedrock and the bottom was covered with rotting leaves").

Remarks. The genitalia of the holotype appears to have some modest fungal growth on the median lobe (Fig. 9B), the circular "halo" at the tip of the median lobe appears to be unnatural and is not part of the original structure.

## Notionotus peruensis species group

Diagnosis. The species of peruensis group can be diagnosed by the dorsal coloration completely yellow, the elevation of the mesoventrite with one transverse and one longitudinal (e.g., Fig. 10B), and by the shape of the genitalia (Fig. 9F).

## Notionotus peruensis sp. nov.

http://zoobank.org/4E05420D-577E-4B12-8B2B-783B0C0BB101
Figs 4K, 9F, 14

Type material examined. Holotype (male): "PERU: Dept. Madre de/ Dios: Pantiacolla Lodge, / Alto Madre de Dios R./ $12^{\circ} 39.3^{\prime} \mathrm{S}, 71^{\circ} 13.9^{\prime} \mathrm{W} 420 \mathrm{~m} / 14-19-\mathrm{XI}-2007 \mathrm{D}$. Brzoska/ ex. flight intercept trap/ PER1B07 004" (SEMC).

Differential diagnosis. Notionotus peruensis can be distinguished by the particular shape of the aedeagus, being nearly rectangular in the basal half and abruptly narrow in the apical half (Fig. 9F).

Description. Size and form: Body length 1.6 mm . Body form elongate oval, convex in lateral view (Fig. 4K). Color and punctation: Dorsally yellow, head mostly yellow and frons pale brown; pronotum with two small black round spots along posterior margin (Fig. 4K). Ventrally brown; maxillary palps, mouthparts, antennae (antennal club slightly darker) and legs yellow. Clypeus and labrum with dense, fine, and weakly impressed ground punctation (punctures separated by $2 \times$ their width); pronotum and elytra ground punctation fine, weakly impressed and sparser than on head (punctures separated by $3 \times$ their width). Head: Clypeus and labrum shallowly emarginate anteromedially, lateral margins of the labrum bearing setae. Thorax: Prosternum carinate medially, strongly raised, pointing anteriorly and acute. Elevation of mesoventrite with one transversal ridge, elevated medially, lateral sides concave; longitudinal ridge sharp, the point where the two ridges merged acute (e.g., Fig. 10B); elevation flat in lateral view; mesoventrite with triangular shape in ventral view. Metaventrite convex in the median region, pubescent with narrow glabrous patch on the medial and posterolateral area, medial region patch drop-shaped; anterior margin extending to mesoventrite elevation. Metafemora densely covered with hydrofuge pubescence on basal three-quarters. Abdomen: Abdominal ventrites very densely pubescent. Aedeagus (Fig. 9F) with basal piece $0.7 \times$ the length of a paramere. Base of the parameres wider than the base of the median lobe; outer margins straight along basal two-thirds, then apically sinuate, inner margins straight along basal two-thirds and then convex and tapering apically; apex of parameres rounded and pointing outwards. Median lobe much shorter than the parameres, basal half rectangular, apical half narrowing abruptly, apex rounded.

Etymology. The species is named after Peru, the country where it was collected, as well as for being the first species described for the genus in this country.

Distribution. Known only from the type locality in Peru (Fig. 14).
Life history. The single specimen was collected at a flight intercept trap; nothing is known about its habitat.

## Notionotus spp.

Fig. 14
Material examined (2 exs.). Bolivia: Santa Cruz: Amboro National Park, Guarda Parque Mataracu, 21-27.xi.2004, malaise trap, leg. Robertson, García, \& Vidaurre (1 female, UMSP). Peru: Cusco: Quispicanchi Province, streams 1 km N Quince Mil, $13^{\circ} 13.335^{\prime} \mathrm{S}, 70^{\circ} 46.035^{\prime} \mathrm{W}, 730 \mathrm{~m}, ~ 9 . i .2020$, leg. Baca., Slow rivulets \& pools w/ saturated detritus next to stream, PE20-0109-02A (1 female, SEMC, DNA voucher SLE2140).

Remarks. We examined two single female specimens from unique localities that we refrain from identifying or describing, due to the lack of male genitalia for comparison. The specimen from Peru likely represents an undescribed species, as suggested by its distant position in our DNA tree (Fig. 1). The single female from Bolivia is the first and only known record of the genus from that country.

## Key to the species groups of Notionotus of the Neotropical Region

Although it is fairly straightforward to key specimens (especially males) to species group, identification to species within each group almost always requires examination of the aedeagus. For this reason, as well as the fact that there are no doubt many yet-to-be-described species, particularly in the southern Amazon region, we do not include a species key here.

1 Elevation of the mesoventrite composed by two ridges (one transverse and one longitudinal) (e.g., Fig. 10A, B) 2

- Elevation of the mesoventrite composed by three ridges (two transverse and one longitudinal) (e.g., Fig. 10C, D)........................... lohezi species group
2 Length of the parameres nearly the same as the length of the basal piece. Length of the median lobe almost the same as the parameres (e.g., N. liparus, Fig. 7A)
liparus species group
- Length of the parameres longer than the basal piece. Length of the median lobe almost the same as the parameres3

3 Outer margin of the parameres convex and tapering reaching the apex, rounded apex. Median lobe rectangular along basal half then tapering abruptly (e.g., N. peruensis sp. nov., Fig. 9F)......................peruensis species group

- Outer margin of the parameres sinuate, apex of the parameres lanceolate. Median lobe wide and approximately triangular (e.g., N. rosalesi, Fig. 9F) ....
rosalesi species group


## Discussion

Since Notionotus was first described as a hygropetric genus from the Andean Region of Venezuela fifty years ago (Spangler 1972), our knowledge of the genus has substantially expanded. Within the Neotropical region, this present work expands the distribution throughout much of tropical South America, including the first records from Peru, Bolivia, and Brazil. We also find that while the originally described species do appear to prefer hygropetric habitats (especially $N$. liparus), most species are not found in seepages but prefer leaf packs and margins of forested streams. The vast majority of specimens have been collected at sites under 1000 meters in elevation, though N. liparus has been found as high as 1770 m . Although the highest density of known specimen records remains in the northern quarter of South America, particularly Venezuela and the Guiana Shield region, this is almost certainly an artifact of collecting effort, as aquatic beetles are in general less well known in the central and southern Amazon region. While many new species undoubtedly remain to be described, we also found that some species have quite expansive ranges (e.g., N. dilucidus, $N$. tricarinatus, N. vatius) and care should be taken to put even seemingly geographically distant new collections in context with existing species. The integration of DNA sequence data proved invaluable in helping to establish species limits in these widespread taxa.

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