

Annotated catalogue of the Tachinidae (Insecta, Diptera) of Chile

By

James E. O'Hara, D. Monty Wood, Christian R. González

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Annotated catalogue of the Tachinidae (Insecta, Diptera) of Chile

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Abstract

The Tachinidae (Diptera) of Chile are catalogued and information is given on distributions, name-bearing types, synonyms, nomenclatural issues, and pertinent literature. The history of tachinid collectors in Chile and authors who have contributed to the systematic knowledge of Chilean tachinids is extensively reviewed. The classification has been updated and 122 genera and 264 species are recognised in Chile. There is a significant amount of endemism with 28 genera and 100 species known only from Chile. There are also 113 species with distributions shared only between Chile and Argentina, particularly in the southern portions of these countries comprising Patagonia.

The catalogue is based on examination of the original descriptions of all nominal species and all other references known to us containing relevant taxonomic and distributional information, for a total of approximately 450 references. Many of the name-bearing types and other Chilean specimens housed in collections were examined. Taxa are arranged hierarchically and alphabetically under the categories of subfamily, tribe, genus, subgenus (where recognised), and species. Nomenclatural information is provided for genus-group and species-group names, including lists of synonyms (mostly restricted to Neotropical taxa) and name-bearing type data. Species distributions are recorded by country within the New World and by larger geographical divisions in the Old World. Additional information is given in the form of notes and references under valid names at the level of tribe, genus, and species. Two genera are newly recorded from Chile: *Chaetoepalpus* Vimmer & Soukup, 1940 (Tachinini) (also newly recorded from Argentina) and *Patelloa* Townsend, 1916 (Goniini). Four species are newly recorded from Chile or

† Deceased

other countries: *Lypha ornata* Aldrich, 1934 (Chile); *Chaetoepalpus coquilletti* Vimmer & Soukup, 1940 (Argentina and Chile); *Phytomyptera evanescens* (Cortés, 1967) (Argentina); and *Xanthobasis unicolor* Aldrich, 1934 (Chile). Eight species previously recorded from Chile are deemed to have been misidentified or misrecorded from Chile (known distributions in parentheses): *Archytas incertus* (Macquart, 1851) (Argentina, Brazil, Paraguay, Uruguay); *Archytas seminiger* (Wiedemann, 1830) (Brazil, Colombia); *Gonia crassicornis* (Fabricius, 1794) (Brazil, Peru, Venezuela, Middle America, West Indies, Nearctic); *Lespesia andina* (Bigot, 1888) (Cuba); *Lespesia archippivora* (Riley, 1871) (widespread Nearctic and most of Neotropical); *Neothilla ignobilis* (van der Wulp, 1890) (Mexico, United States); *Siphona* (*Siphona*) *geniculata* (De Geer, 1776) (Palearctic, Nearctic [introduced]); and *Winthemia quadripustulata* (Fabricius, 1794) (Palearctic, Nearctic, Oriental]. As First Reviser we fix *Paratheresia rufiventris* Townsend, 1929 as the senior homonym and *Sarcoprosena rufiventris* Townsend, 1929 as the junior homonym when the two are placed together in *Billaea* Robineau-Desvoidy, 1830; and we fix *Mayophorinia angusta* Townsend, 1927 as the senior homonym and *Metarrhinomyia angusta* Townsend, 1927 as the junior homonym when the two are placed together in *Myiopharus* Brauer & Bergenstamm, 1889. New replacement names are proposed for eight preoccupied names of Neotropical species (country of type locality in parentheses): *Billaea rufescens* O'Hara & Wood for *Sarcoprosena rufiventris* Townsend, 1929, preoccupied in the genus *Billaea* Robineau-Desvoidy, 1830 by *Paratheresia rufiventris* Townsend, 1929 (Peru), **nom. nov.**; *Billaea triquetrus* O'Hara & Wood for *Sarcoprosena triangulifera* Townsend, 1927, preoccupied in the genus *Billaea* Robineau-Desvoidy, 1830 by *Dexia triangulifera* Zetterstedt, 1844 (Peru), **nom. nov.**; *Eucelatoria nudioculata* O'Hara & Wood for *Eucelatorioidea nigripalpis* Thompson, 1968, preoccupied in the genus *Eucelatoria* Townsend, 1909 by *Chetolyga nigripalpis* Bigot, 1889 (Trinidad), **nom. nov.**; *Eucelatoria oblonga* O'Hara & Wood for *Urodexodes elongatum* Cortés & Campos, 1974, preoccupied in the genus *Eucelatoria* Townsend, 1909 by *Exorista elongata* van der Wulp, 1890 (Chile), **nom. nov.**; *Lespesia thompsoni* O'Hara & Wood for *Sturmiopsoidea obscura* Thompson, 1966, preoccupied in the genus *Lespesia* Robineau-Desvoidy, 1863 by *Eurigaster obscurus* Bigot, 1857 (Cuba), **nom. nov.**; *Myiopharus charapensis* O'Hara & Wood for *Metarrhinomyia angusta* Townsend, 1927, preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Mayophorinia angusta* Townsend, 1927 (Peru), **nom. nov.**; *Myiopharus incognitus* O'Hara & Wood for *Stenochaeta claripalpis* Thompson, 1968, preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Neoxynopsoidea claripalpis* Thompson, 1968 (Trinidad), **nom. nov.**; and *Myiopharus rufopalpus* O'Hara & Wood for *Paralipse palpalis* Townsend, 1929, preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Myioxynops palpalis* Townsend, 1927 (Peru), **nom. nov.** New type species fixations are made under the provisions of Article 70.3.2 of the ICZN Code for three genus-group names: *Parafabricia* Brauer & Bergenstamm, 1894 (synonym of *Archytas* Jaennicke, 1867), type species newly fixed as *Parafabricia perplexa* Townsend, 1931; *Tachinodes* Brauer & Bergenstamm, 1889 (synonym of *Archytas* Jaennicke, 1867), type species newly fixed as *Jurinia metallica* Robineau-Desvoidy, 1830; and *Willistonina* Brauer & Bergenstamm, 1889 (synonym of *Belvosia* Robineau-Desvoidy, 1830), type species newly fixed as *Willistonina aldrichi* Townsend, 1931. Lectotypes are designated for the following four nominal species, all described or possibly described from Chile: *Echinomyia pygmaea* Macquart, 1851 (a valid name in the genus *Peleteria* Robineau-Desvoidy, 1830); *Gonia chilensis* Macquart, 1844 (a junior synonym of *Gonia pallens* Wiedemann, 1830); *Masicera auriceps* Macquart, 1844 (a valid name in the genus *Lespesia* Robineau-Desvoidy, 1863); and *Prosopochaeta nitidiventris* Macquart, 1851 (a valid name in the genus *Prosopochaeta* Macquart, 1851). The following 27 new or revived combinations are proposed (distributions in parentheses): *Blepharipeza andina* Bigot, 1888 is moved to *Lespesia* Robineau-Desvoidy, 1863 as *L. andina*, *nomen dubium* (Cuba), **comb. nov.**; *Camposodes evanescens* Cortés, 1967 is moved to *Phytomyptera* Rondani, 1845 as *P. evanescens* (Argentina, Chile), **comb. nov.**; *Ectophasiopsis ypiranga*

Dios & Nihei, 2017 is moved to *Trichopoda* Berthold, 1827 and assigned to subgenus *Galactomyia* Townsend, 1908 as *T. (G.) ypiranga* (Argentina, Brazil), **comb. nov.**; *Embiomyia australis* Aldrich, 1934 is moved to *Steleoneura* Stein, 1924 as *S. australis* (Argentina, Chile), **comb. nov.**; *Eurigaster modestus* Bigot, 1857 is moved to *Lespesia* as *L. modesta* (Cuba), **comb. nov.**; *Eurigaster obscurus* Bigot, 1857 is moved to *Lespesia* as *L. obscura* (Cuba), **comb. nov.**; *Macropatelloa tanumeana* Townsend, 1931 is moved to *Patelloa* Townsend, 1916 as *P. tanumeana* (Argentina, Chile), **comb. nov.**; *Masicera insignis* van der Wulp, 1882 is moved to *Drino* Robineau-Desvoidy, 1863 as *D. insignis* (Argentina, Chile), **comb. nov.**; *Parasetigena hichinsi* Cortés, 1967 is moved to *Chetogena* Rondani, 1856 as *C. hichinsi* (Chile), **comb. nov.**; *Parasetigena porteri* Brèthes, 1920 and junior synonym *Stomatotachina splendida* Townsend, 1931 are moved to *Chetogena* as *C. porteri* (Chile), both **comb. nov.**; *Phorocera calyptrata* Aldrich, 1934 is moved to *Admontia* Brauer & Bergenstamm, 1889 as *A. calyptrata* (Argentina, Chile), **comb. nov.**; *Poliops auratus* Campos, 1953 is moved to *Admontia* Brauer & Bergenstamm, 1889 as *A. aurata* (Chile), **comb. nov.**; *Poliops striatus* Aldrich, 1934 is moved to *Admontia* as *A. striata* (Argentina, Chile), **comb. nov.**; *Ruiziella frontosa* Cortés, 1951 is moved to *Chaetoepalpus* Vimmer & Soukup, 1940 and placed in synonymy with *C. coquilleti* Vimmer & Soukup, 1940 (Argentina, Chile, Peru), **comb. nov.**; *Ruiziella luctuosa* Cortés, 1951 is moved to *Chaetoepalpus* as *C. luctuosus* (Argentina, Chile), **comb. nov.**; *Sarcoprosena luteola* Cortés & Campos, 1974 is moved to *Billaea* Robineau-Desvoidy, 1830 as *B. luteola* (Chile), **comb. nov.**; *Sarcoprosena rufiventris* Townsend, 1929 is moved to *Billaea* where it is a junior secondary homonym and is renamed *B. rufescens* O'Hara & Wood (Peru), **comb. nov.**; *Sarcoprosena triangulifera* Townsend, 1927 is moved to *Billaea* where it is a junior secondary homonym and is renamed *B. triquetrus* O'Hara & Wood (Peru), **comb. nov.**; *Saundersia aurea* Giglio-Tos, 1893 is moved to "Unplaced species of Tachinini" (Mexico), **comb. nov.**; *Schistostephana aurifrons* Townsend, 1919 is moved to *Billaea* as *B. aurifrons* (Peru), **comb. nov.**; *Siphoactia charapensis* Townsend, 1927 is moved to *Clausicella* Rondani, 1856 as *C. charapensis* (Peru), **comb. nov.**; *Siphoactia peregrina* Cortés & Campos, 1971 is moved to *Clausicella* as *C. peregrina* (Chile), **comb. nov.**; *Sturmia festiva* Cortés, 1944 is moved to *Drino* as *D. festiva* (Argentina, Chile), **comb. nov.**; *Sturmiopsoidea obscura* Thompson, 1966 is moved to *Lespesia* Robineau-Desvoidy, 1863, where it is a junior secondary homonym and is renamed *L. thompsoni* O'Hara & Wood (Trinidad), **comb. nov.**; *Trichopoda arcuata* Bigot, 1876 is returned to *Trichopoda* from *Ectophasiopsis* Townsend, 1915 and assigned to subgenus *Galactomyia* (Argentina, Chile), **comb. revived**; and *Trichopoda gradata* Wiedemann, 1830 is returned to *Trichopoda* from *Ectophasiopsis* and assigned to subgenus *Galactomyia* (Argentina, Brazil, Uruguay), **comb. revived**. New or revived generic and specific synonymies are proposed for the following 14 names: *Camposodes* Cortés, 1967 with *Phytomyptera* Rondani, 1845, **syn. nov.**; *Ectophasiopsis* Townsend, 1915 with *Trichopoda* Berthold, 1827, subgenus *Galactomyia* Townsend, 1908, **syn. nov.**; *Embiomyia* Aldrich, 1934 with *Steleoneura* Stein, 1924, **syn. nov.**; *Fabricia andicola* Bigot, 1888 with *Peleteria robusta* (Wiedemann, 1830), **syn. revived**; *Macropatelloa* Townsend, 1931 with *Patelloa* Townsend, 1916, **syn. nov.**; *Peleteria inca* Curran, 1925 with *Peleteria robusta* (Wiedemann, 1830), **syn. revived**; *Poliops* Aldrich, 1934 with *Admontia* Brauer & Bergenstamm, 1889, **syn. nov.**; *Ruiziella* Cortés, 1951 with *Chaetoepalpus* Vimmer & Soukup, 1940, **syn. nov.**; *Ruiziella frontosa* Cortés, 1951 with *Chaetoepalpus coquilleti* Vimmer & Soukup, 1940, **syn. nov.**; *Sarcoprosena* Townsend, 1927 with *Billaea* Robineau-Desvoidy, 1830, **syn. nov.**; *Schistostephana* Townsend, 1919 with *Billaea*, **syn. nov.**; *Siphoactia* Townsend, 1927 with *Clausicella* Rondani, 1856, **syn. nov.**; *Stomatotachina* Townsend, 1931 with *Chetogena* Rondani, 1856, **syn. nov.**; and *Sturmiopsoidea* Thompson, 1966 with *Lespesia* Robineau-Desvoidy, 1863, **syn. nov.**

Keywords

Argentina, Chile, Neotropical Region, Oestroidea, parasitoids, Patagonia

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“The joke goes in Chile, that God the Creator, after seven days of hard labor, setting of the world out of chaos, was indeed tired and weary, when Angels approached Him with concern to tell that there still were huge assortment of deserts, oceans, mountains, lakes, forests, volcanoes, rocks, glaciers, islands and the rest, that they didn’t know where to place. The sensible answer from the Almighty was: well, throw it away in any remote corner still available! And this, gentlemen, was the way in which Chile was supposedly built.”

– Campos (1975: 7)

Introduction

Chile is a long slender country nestled between the Pacific Ocean and the Andes and stretching for more than 4000 kilometres from Peru to the southern tip of South

America. The Atacama Desert in the north gradually transitions to the fertile Central Valley that runs through the middle of the country for over 600 kilometres. This is the agricultural heartland of Chile and is noted for its Mediterranean climate and large variety of produce that is exported to countries around the world. Farther to the south are the ecoregions of the Valdivian temperate rainforest and Magellanic subpolar rainforest. The former has an especially diverse fauna and flora with a high percentage of endemics whereas the harsher and less hospitable conditions of the latter have limited its biodiversity. The Andes Mountains stretch along the eastern edge of Chile and faunistically separate it from the rest of South America except in the more southern reaches of the continent (i.e., Patagonia). The Tachinidae fauna of Chile has not been catalogued for fifty years, since Guimarães (1971) published on the family in *A catalogue of the Diptera of the Americas south of the United States*. That work was of necessity built upon the taxonomic contributions of predecessors and was heavily influenced by the most prolific describer of New World Tachinidae, C.H.T. Townsend. The culmination of Townsend's life's work was a twelve-volume series entitled *Manual of Myiology* (Townsend 1934–1942) that laid out an idiosyncratic classification for tachinids and related groups and provided keys to, and descriptions of, all of the world's genera. The *Manual of Myiology* and the many other publications of Townsend set forth a taxonomic scheme in which many genera were monotypic and the arrangement of genera into tribes and families followed ideas about relationships that have since been largely abandoned. An alternative to such restricted genera was proposed for a portion of the South American fauna in Aldrich's (1934) treatment of Tachinidae in *Diptera of Patagonia and South Chile*, but that work did not arrange genera into higher categories. Younger taxonomists who came along near the end of Townsend's era, like R. Cortés in Chile and E. Blanchard in Argentina, were obliged to interpret their faunas within the context of *Manual of Myiology*. As a result, as noted by Wood and Zumbado (2010: 1355): "Subsequent authors faced with this multiplicity of generic names have had little choice, when their specimens did not fit the existing narrow definitions, but to describe yet more genera".

Our main objective here has been to catalogue the described taxa of Chile rather than revise the classification, but we have done as much of the latter as seemed appropriate given our level of understanding of the fauna. We have updated the higher categories of tribes and subfamilies (following Sabrosky 1999 for priority of family-group names), proposed new synonymies and combinations, and replaced some preoccupied names with new ones. Our placements of taxa have been aided by recent advances in our understanding of tachinid phylogeny and especially by the morphological study of Cerretti et al. (2014) and molecular study of Stireman et al. (2019). Our own DNA barcoding of Chilean tachinid specimens in the Canadian National Collection of Insects has also helped with the placement of certain taxa. Certain groups are too problematic to be easily reclassified here (e.g., the Polideini) and we have maintained the current classification of these until more thorough revisions can be undertaken.

The tachinids of Chile, as catalogued below, consist of 122 genera and 264 species, with 28 genera and 100 species which are endemic to Chile according to presently

known distributions. Another 113 species are uniquely shared between Chile and Argentina, particularly in the southern portions of these countries. There is still a significant portion of the tachinid fauna of Chile that is undescribed and it is our hope that this catalogue will benefit those who pursue systematic studies of the fauna in the future.

A historical perspective on the Tachinidae of Chile

The earliest accounts of entomological pursuits in Chile were traced back to the 1500s by Cortés and Herrera (1989) in their review of the history of entomology in Chile. It was not until the 1800s that any organised progress was made. As the fledgling country struggled towards independence from Spanish rule, an ambitious plan was developed for a comprehensive account of virtually all aspects of Chilean natural and political history, including entomology. The person chosen to lead this grand endeavour was a young French naturalist who had emigrated to Chile just a year or so earlier and was not yet fluent in Spanish. His name was Claude [or Claudio] Gay (1800–1873) and he was commissioned in 1830 to research and write what would later be entitled *Historia física y política de Chile según documentos adquiridos en esta república durante doce años de residencia en ella y publicada bajo los auspicios del supremo gobierno* [“Physical and political history of Chile according to documents acquired in this republic during twelve years of residence in it and published under the auspices of the supreme government”].

Preparing the *Historia* consumed much of the next 40+ years of Gay’s life. He travelled extensively throughout Chile gathering information and making collections of the fauna and flora before returning to France in 1842. There he commenced the writing of the *Historia* subject by subject in a long series of volumes. When finally completed the *Historia* filled ca. 30 volumes and was published over the course of 28 years, from 1844 to 1871. The cumulative effect of this extraordinary work was to define the physical characteristics of Chile and thus provide a foundation for decision-making for years to come. Its coverage of life forms was thought to be so complete that Cortés and Herrera (1989: 303) remarked: “Hasta bien entrado el siglo XX se acostumbraba a decir en Chile ‘si no está en Gay (por plantas, insectos y animales) es nuevo’” [“Until well into the 20th Century it was customary to say in Chile ‘if it is not in Gay (for plants, insects and animals) it is new’.”].

Eight volumes of the *Historia* were devoted to zoology and a section in the seventh dealt with the Diptera. Gay had enlisted the aid of French zoologist C. Émile Blanchard (1819–1900) to prepare this section. No keys were included but species were arranged in a hierarchical classification of genera, tribes and families and each category was accompanied by a diagnosis and description (Blanchard 1854). Tachinids were grouped with the “Muscianos” and arranged into three tribes, but at this early stage in Chilean entomology there were few tachinid species known from the country. Blanchard recorded only seven, consisting of six described by French dipterist Pierre-Justin-Marie Macquart (1778–1855) and one described by German dipterist Christian Rudolph Wilhelm Wiedemann (1770–1840). The last species, treated as

Scotiptera melaleuca (Wiedemann) and now considered a synonym of *Scotiptera venatoria* (Fabricius), was described from Brazil and was questionably recorded from Chile; it is no longer recognised as a Chilean species.

By the time of Blanchard (1854), specimens of Chilean Tachinidae were trickling back to Europe and making their way into institutional and private collections. The first of these to be described were collected by Captain Philip Parker King during the voyage of the English naval vessels HMS *Adventure* and HMS *Beagle* to the “Straits of Magellan” at the southern tip of South America in 1825–1830. The specimens were from “Cape Gregory” [Cabo San Gregorio] and “Port Famine” [Puerto del Hambre] in the Magallanes province of Chile. King gave the specimens to the British Museum (Natural History) in London (now the Natural History Museum, NHMUK) where they were later examined and described by English entomologist Francis Walker (1809–1874) in a paper devoted to King’s insects (Walker 1836). Walker was a general entomologist who described insects of all kinds with no special talent for taxonomy or descriptions. He described five tachinid species from King’s material and placed them all in the genus *Tachina*. Subsequent study of the specimens led to their reassignment to two subfamilies and three tribes, with one of Walker’s species having been described twice under different names (see catalogue below for further details).

Walker (1849) later described another Chilean species, this one simply from “Chili” and collected by Hugh Cuming (1791–1865), an Englishman who settled in Chile and became a prosperous businessman and amateur naturalist. Walker also assigned this species to *Tachina* but it was later synonymised with an earlier Macquart name and is now placed in the common genus *Archytas*.

None of the species described by Walker was included by Blanchard in the *Historia*. We have been unable to determine the reason for this but it was likely an intentional act given the care with which the insect chapters were prepared and the presumed availability of the Walker publications at the time.

Species described from Chile by Macquart (1844, 1851) and listed in the *Historia* by Blanchard (1854) were based on specimens in the Muséum National d’Histoire Naturelle (the “Paris Museum”, MNHN) received from French collectors residing in or visiting Chile during the first half of the 19th Century. These collectors were frequently prominent and adventurous men better known for their non-entomological achievements. Macquart gave credit to them by naming the collector at the end of each species description. They were among the first to collect Tachinidae in Chile and are listed here in recognition of the important role they played in making these insects available to others who could describe them (collectors arranged by year of birth):

- 1) “Du Brésil ou du Chili. M. Gaudichand.” Charles Gaudichand-Beaupré (1789–1854), naturalist. Two species, as *Masicera auriceps* [= *Lespesia auriceps*] and *Gonia virescens*; neither included in the *Historia* and both to this day of uncertain provenance.
- 2) “Du Chili; de la Conception. M. Dumont-Durville.” Jules Dumont d’Urville (1790–1842), second in command on the French frigate *La Coquille*, which reached Concepción in January 1823. Macquart named *Trichoprosopus durvillei* in his honour.

- 3) “Du Chili. M. Gay.” Claude [or Claudio] Gay (1800–1873), author of *Historia física y política de Chile...* (see above for further details about Gay). Four species, as *Jurinia scutellata* [= *Archytas scutellatus*], *Gonia chilensis* [= *Gonia pallens* Wiedemann], *Prosopochaeta nitidiventris* and *Hyalomyia chilensis* [= *Phasia chilensis*].
- 4) “De la Patagonie. M. d’Orbigny.” Alcide Charles Victor Dessalines d’Orbigny (1802–1857), naturalist. One species, *Gonia lineata*; not included in the *Historia* presumably because of the uncertain provenance; current distribution as Argentina, Chile and Peru. This collector should not be confused with younger brother Charles Henry Dessalines d’Orbigny (1806–1876), author of *Dictionnaire Universel d’Histoire Naturelle*.
- 5) “Du Chili. M. Pissis.” Pierre Joseph Aimé Pissis (1812–1889), geographer. One species, as *Echinomyia pygmaea* [= *Peleteria pygmaea*]. Monte Pissis in Argentina, one of the highest mountains in South America, was named in his honour.

This practice of collecting insects in Chile and sending them back to Europe to be preserved in collections and described by specialists continued throughout the second half of the 19th Century. Tachinids seem not to have been the most popular of insects to send back to Europe during this time judging from the few that fell into the hands of the leading dipterists. The dipterists involved in describing them are reviewed below.

French dipterist Jacques-Marie-Frangille Bigot (1818–1893) described his first Chilean tachinid in 1857. The description was based on material received from Philibert Germain (1827–1913), a French entomologist who had emigrated to Chile in 1850. Germain was a man of considerable talents; over the course of the next 50 years he held important positions in Chilean entomology and collected throughout the country (see review of Germain in Cortés and Herrera 1989). Bigot described an additional 12 Chilean tachinids in six papers between 1876 and 1888. These species were all from “Chili” and caught by unnamed collectors except for two described in Bigot’s (1888a) section on Diptera in the multi-volume report *Mission scientifique du Cap Horn, 1882–1883*. These tachinids and other natural history specimens were collected by naval physician Paul Daniel Jules Hyades (1847–1919) in the vicinity of “Orange Bay” [Isla Hoste, Bahía Orange] during the voyage to Cape Horn of the French frigate *La Romanche*. Bigot (1888a: 26) dedicated the genus *Hyadesimyia* to Hyades.

French dipterist André-Jean-Baptiste Robineau-Desvoidy (1799–1857) achieved notoriety in part because of his prodigious output: nearly 600 generic names and over 3000 specific names (Evenhuis et al. 2010). These names were mostly of European schizophoran Diptera and a sizable portion of them later became junior synonyms. He named just one Chilean tachinid, *Jurinia andana* [= *Archytas scutellatus* (Macquart)], from “Chili” (Robineau-Desvoidy 1863a). The collector was not given and the type(s) is lost (Cortés 1963).

The eminent Italian dipterist Camillo Rondani (1808–1879) described five species from Chile based on material collected by Rudolph [or Rodulfo] Amandus Philippi (1808–1904). Philippi was born in Germany and received his higher education in Berlin, where coincidentally he was taught “Physische Geographie” by the famed naturalist and South American explorer Alexander von Humboldt (1769–1859) (Kabat and Coan 2017). Philippi emigrated to Chile in 1851 after revolutions in Germany forced

him to leave for his personal safety. He settled first in Valdivia in southern Chile but moved to Santiago in 1853 when the President of Chile appointed him director of the Museo de Historia Natural at the Universidad de Chile. He continued as director when the museum was relocated to another part of Santiago and renamed Museo Nacional de Historia Natural. He relinquished the position in 1897 at the age of 88. Philippi was well respected within the scientific community during his tenure as director and became one of the most influential voices for the natural sciences in Chile. He was also a prolific author of taxonomic papers on a wide variety of organisms throughout the animal kingdom (Cortés and Herrera 1989; Kabat and Coan 2017). His *Aufzählung der chilenischen Dipteren*, published in 1865, stands out as his most impressive contribution to dipterology with descriptions of 424 species of Diptera, but no Tachinidae (Camousseight 2005).

Three of the five species described by Rondani (1863) based on Philippi material are still recognised as valid. One of them was named *Spathipalpus philippii* in honour of Philippi and was based on material from Valdivia. Philippi had a farm there and much of the material described in his *Aufzählung der chilenischen Dipteren* was collected from that area.

German amateur entomologist Johann Friedrich Jaennicke (1831–1907) published a significant paper entitled *Neue exotische Dipteren* in 1867 that included 18 new species of Tachinidae from Indonesia, Ethiopia, Cuba, Mexico, Panama, Venezuela and Chile. The single Chilean species was described as *Demoticus ratzeburgii* [= *Deopalpus pruinosus* (Rondani)] and was presumably named in honour of Julius Theodor Christian Ratzeburg (1801–1871), a prominent German professor famous for his pioneering work in forest entomology. The type(s) of *D. ratzeburgii* was from “Chile”; the collector “Bayrhofer” is unknown to us. Jaennicke (1867) was also responsible for describing the common New World genus *Archytas*.

The circumnavigation of the world by the Austrian frigate SMS *Novara* in 1857–1859 was the largest naval expedition ever undertaken by Austria. It was primarily a voyage of scientific discovery and its bountiful harvest of natural history specimens would greatly increase the holdings and global status of the natural history collection in Vienna. The principal zoologist responsible for collecting insects during the *Novara* voyage was Georg Ritter von Frauenfeld (1807–1873), a curator at the natural history museum in Vienna. The task of describing the expedition's Diptera went to Ignatz Rudolph Schiner (1813–1873 [first initials often as J.R. on his publications]), a talented Viennese dipterist well-known for authoring the Diptera section of *Fauna Austriaca* (published in parts between 1860 and 1864). Schiner (1868) described 37 species of Tachinidae from the *Novara* expedition but only three were specifically from Chile; another 15 were simply from “Süd-Amerika”. Twenty years after these *Novara* tachinids were described the vast natural history collection in Vienna was moved into the new Naturhistorisches Museum Wien (or “Vienna Museum”, NHMW) along the *Ringstrasse*, where it could be displayed in regal splendour and the scientific staff could more easily study its specimens.

The *Novara* was preceded in its circumnavigation of the world by the Swedish frigate HSwMS *Eugenie* during the years 1851 to 1853. A huge number of insects was

collected during the expedition and various Swedish experts published on the newly discovered species. The Diptera were described by Carl Gustaf Thomson (1824–1899) and comprised over 300 new species (Thomson 1869). Only one was a tachinid, *Degeeria antarctica* [= *Admontia antarctica*]. The locality was given as “Patagonia” but was probably “Port Famine” [Puerto del Hambre] according to the review of the *Eugenie* localities by Persson (1971: 168). The collector was likely the ship’s physician and zoologist, Johan Gustaf Hjalmar Kinberg (1820–1908). Although the voyage of the *Eugenie* preceded that of the *Novara*, the Diptera species described by Schiner (1868) have priority over those of Thomson (1869).

Dutch dipterist Frederik Maurits van der Wulp (1818–1900) published extensively on world Diptera, with an emphasis on Tachinidae. He described close to 500 tachinid species, including over 400 from Mexico and mostly published in the remarkable *Biologia Centrali-Americana* (e.g., van der Wulp 1890a–e). Three tachinids were described from Chile in van der Wulp (1882) and were based on specimens in the “Leyden Museum” (Leiden, RMNH) received from or collected by entomologist and beetle specialist Carl August Dohrn (1806–1892). Dohrn held the position of president of the Entomological Society of Stettin (Entomologischer Verein zu Stettin) for over 40 years. [Stettin, now Szczecin, has been part of Poland since the end of World War II.]

The first catalogue of Chilean Diptera after Blanchard’s (1854) treatment in Gay’s *Historia* was published in 1888 by Edwyn Charles Reed (1841–1910). Reed, an English naturalist, emigrated to Chile in 1869 and held various positions in entomology and natural history within the country, including director of the Museo de Historia Natural de Concepción at the time of his death (Cortés and Herrera 1989; Etcheverry 1993; Edmundson 2009). The catalogue was a minimalistic listing of 716 species of Diptera. Twelve species of Tachinidae are listed in the main body of the text and they comprise the species in the *Historia*, the three Schiner species mentioned above, and two species originally described from Brazil by Wiedemann (details above). The Chilean species of Bigot were originally overlooked but were added at the end of the list prior to *Anales de la Universidad de Chile* going to press. The Chilean species described by Rondani (1863), Robineau-Desvoidy (1863a) and Jaennicke (1867), along with the species described from the southern tip of Chile not in the *Historia* (to which can be added the single species described by Thomson in 1869), were not included in Reed’s (1888) catalogue. Aldrich (1928b) named the tachinid genus *Reedia* after Reed and then renamed it *Edwynia* (Aldrich 1930) when *Reedia* Aldrich was discovered to be a junior homonym of *Reedia* Ashmead, 1904.

Austrian zoologist Friedrich Moritz Brauer (1832–1904) was employed at the Naturhistorisches Museum Wien (NHMW) as a curator for many years (of Mollusca and then Insecta) and then as director. He engaged the assistance of Vienna dipterist Julius Edler von Bergenstamm (1837–1896) to prepare a monumental work entitled *Vorarbeiten zu einer Monographie der Muscaria schizometopa (exclusive Anthomyidae)*, published in four parts between 1889 and 1894 (see O’Hara (2013a) for more about these authors, their monographs, and the NHMW). Brauer and Bergenstamm (1891) and Brauer (1898) each described one tachinid species from Chile. The first was col-

lected by Philippi (in 1870 according to a label quoted by Aldrich (1925: 459)) and the other came from the Bigot collection (collector unknown). Both names are now junior synonyms of other names.

By the end of the 19th Century there were ca. 40 species of Tachinidae described from Chile and a few more described from elsewhere and later recorded from Chile. The specimens upon which they were based had been collected over many years in different parts of the country by a variety of collectors, but all had something in common: they were described by European dipterists who had never been to Chile. The collectors were usually adventurous naturalists and the describers were prominent figures in European scientific institutions. Both of these groups were essential to the early knowledge that was being generated on Chilean Tachinidae.

One of the sons of Edwyn Charles Reed (see above), Carlos Samuel Reed (1888–1949), pursued a career in zoology and was most active in entomology and ornithology (Etcheverry 1993). He became an unintentional author of the tachinid name *Tachina porteri* when he published on the biology of this undescribed species (Reed 1907). He wrote that his father had set aside specimens of it in the Museo de Historia Natural de Concepción under the “nombre MS. de [manuscript name of] *Tachina Porteri*, Reed”. By publishing this name and providing a partial description of the species, Reed made the name nomenclaturally available. The senior Reed had chosen the name to honour Carlos Emilio Porter (1867–1942), a director of the Museo de Historia Natural de Valparaíso. *Tachina porteri* Reed, 1907 was the first tachinid described from Chile by a Chilean-born author.

A more formal but brief description of *Tachina porteri* Reed was given by Brèthes (1910) with name and authorship as “*Exorista porteri* (Reed) Brèthes”. Specimens of this species have not been located in recent times and Cortés and Hichins (1969: 90) treated it as *incertae sedis*. Jean [or Juan] Brèthes (1871–1928) was a French-born entomologist and professor in Argentina. From 1902 until his death, Brèthes was the curator in charge of the entomological collection in the Museo Argentino de Ciencias Naturales “Bernardino Rivadavia” in Buenos Aires (MACN). He described more than 1100 species of insects, mostly from Argentina, Chile, and other South American countries (Dallas 1928; Mulieri et al. 2013; Rossi Belgrano and Rossi Belgrano 2018). Approximately 20 species belonged to the Tachinidae including five from Chile. Brèthes’ name-bearing types in MACN were discussed in Mulieri et al. (2013).

The American-born dipterist Charles Henry Tyler Townsend (1863–1944) began his taxonomic studies of Tachinidae in North America in the 1890s with short papers of a regional nature and concluded his career decades later with his huge *Manual of Myiology in Twelve Parts*, 1934–1942. During the course of his career he described ca. 1500 genera and nearly 1600 species (Arnaud 1958; Evenhuis et al. 2015), most belonging to the Tachinidae. In his *Manual of Myiology* he gave an overview of world Tachinidae and other related flies with keys to, and diagnoses of, all tribes and genera. Townsend was a well-disciplined taxonomist with an intimate knowledge of the literature and a remarkable ability to recognise species, but his legacy has been tarnished by his propensity for monotypic genera and adoption of a higher classification that now

appears to be overly artificial (O'Hara 2013b). He has left behind so many genera, especially in South America, that modern systematists are still struggling to determine how best to reduce and reorganise them into more natural and manageable tribes.

Townsend was an avid and adventurous collector who caught many of the New World specimens that he described as new species and genera. He spent much of the latter half of his life in South America, collecting and describing tachinids and ultimately preparing his *Manual of Myiology*. He lived off and on in Peru before permanently settling in Itaquaquecetuba in the São Paulo province of Brazil in 1929 on a property he had purchased about ten years earlier (Hansen and Toma 2004; Evenhuis et al. 2015). Only once did Townsend visit Chile, in 1927 on a journey that took him all the way from Punta Arenas in the south to the bordering country of Peru in the north (Cortés 1944a) where he was currently employed. That trip resulted in the description of four tachinid species and one new genus from Chile (Townsend 1928b). Townsend described a total of 16 species from Chile between 1915 and 1931, ten of which are currently valid. Besides by himself, the type specimens of his new species were collected by Anastasio Piri6n (1888–1959; six species including patronyms *Pirionimyia paradoxa* and *Dolichocryptera pirioni*), E.C. Reed (see above; two species), Paul [or Pablo] Herbst (1861–1927; two species) and persons unknown (two species). The three known collectors were discussed or mentioned by Cortés and Herrera (1989). Twenty-nine species described by Townsend are currently recognised as valid in Chile and they comprise the aforementioned ten species described from Chile, 15 from Peru, two from Argentina, and one each from Bolivia and Brazil. Twenty-seven Townsend genera are recognised in Chile and 123 of his generic names are listed as junior generic synonyms.

There is no account of Townsend's journey through Chile in 1927 but we know from the dates of collection of his Chilean specimens that he was in Punta Arenas on February 5th and Valparaíso on February 15th. He likely travelled southward through Argentinian Patagonia and northward through Chilean Patagonia. Either known or unknown to him, he had just missed a major Diptera expedition to Patagonia by barely a month. The results of that expedition would lead to a profound advance in tachinid taxonomy in both Chile and Argentina.

The Patagonian expedition was conceived by English dipterist, Frederick Wallace Edwards (1888–1940) and had the backing of his employer, the British Museum (Natural History) (NHMUK). His partner in the expedition was American dipterist Raymond Corbett Shannon (1894–1945), who had worked as an assistant at the Bureau of Entomology, United States Department of Agriculture, in Washington but was currently working in Argentina for the Argentine Government on insects of public health concern (McAtee and Wade 1951). Their plan was to collect a broad range of Diptera in the southern temperate forest of Chile and Argentina at a latitude of ca. 41°S, corresponding to northern Patagonia. The small party of Edwards, Shannon and their wives started their collecting expedition near the mouth of the Río Negro in Argentina in late October, 1926 and finished at Concepción in Chile on 27 December (Edwards 1929). A total of 40,000 insects was collected including 30,000 Diptera.

The Edwards-Shannon expedition was part of a grander plan. The Diptera were sent to specialists to serve as a foundation for family treatments in a monographic series called *Diptera of Patagonia and South Chile*. Authors were expected to incorporate previously known species from Patagonia into their treatments and describe the new species they discovered. When they were done, the Edwards material went back to NHMUK and the Shannon material was later donated to the Smithsonian Institution (USNM).

John Merton Aldrich (1866–1934), a well-respected American dipterist at USNM, assumed responsibility for working up the Tachinidae for *Diptera of Patagonia and South Chile*. He was renowned for his catalogue of North American Diptera (Aldrich 1905) and his revisionary work on a number of Diptera families including Tachinidae. His monograph on Patagonian Tachinidae (Aldrich 1934) was an impressive achievement and the first comprehensive revision of a regional tachinid fauna in Chile. It included keys to genera and species, descriptions or diagnoses of genera and species, synonymy, illustrations and notes, but no arrangement into tribes and subfamilies. The number of genera and species was summarised in the introduction: “There are found to be 138 species and 2 varieties, of which 90 species and the 2 varieties are described as new; the total number of genera is 70, of which 28 are new” (Aldrich 1934: 1). Of the 90 new species that Aldrich described from Chile and/or Argentina in that work, 78 are recognised as valid in our catalogue below; another eleven species from earlier Aldrich papers (particularly Aldrich 1928b) are also recognised from Chile. Among these are species named in honour of Edwards (*Callotroxis edwardsi* and *Lypha edwardsi*), Shannon (*Metopomuscopteryx* [= *Alexogloblinia*] *shannoni*), Piri6n (discussed above; *Piriona fasciculata* and *Myiopharus piri6ni*) and English naturalist Charles Darwin (1809–1882) (*Pelycops darwini*). The holotype of the last was collected by Darwin from “Port Famine” [Puerto del Hambre] during the second voyage of HMS *Beagle*, likely in February 1834—almost exactly 100 years before Aldrich’s monograph was published on 24 March 1934. Sadly, Aldrich passed away unexpectedly two months later on 27 May 1934. The new genera of Aldrich (1934) were included in Townsend’s *Manual of Myiology*. Guimar6es (1971) named the genus *Aldrichiopa* after Aldrich and Cort6s (1944e) named a species after him, as *Cylindromyia aldrichi*.

Canadian-born Charles Howard Curran (1894–1972) was hired in 1922 as the first dipterist at the Canadian National Collection of Insects (CNC) in Ottawa but moved to New York for a position at the American Museum of Natural History (AMNH) in 1928. He rose to prominence as a general dipterist with many taxonomic revisions to his credit but is best remembered for his masterful coverage of flies in *The Families and Genera of North American Diptera* (Curran 1934). Only a few of the tachinid genera and species described by Curran are relevant to the Chilean fauna. Three of his generic names (proposed for species in Jamaica and Panama) and three of his species names (two named for Chilean specimens and one for Peruvian specimens) are junior synonyms. The only valid species is *Archytas peruanus*, described from Peru and since recorded from Chile.

A contemporary of Curran was American entomologist Henry Jonathan Reinhard (1892–1976). Reinhard spent most of his career in College Station, first as a general

entomologist at the Texas Agricultural Experiment Station and then as a professor at Texas A&M University (Burke 1977). In addition to his professional responsibilities he had an unwavering passion for tachinids and sarcophagids and published frequently on them throughout his adult life. He was instrumental in describing the rich but little-known fauna of his part of Texas but also wrote generic revisions and described species from more distant parts of the New World. Reinhard, like Curran, was not significantly involved with Chilean tachinids. Several of his generic revisions are helpful from a Chilean perspective (e.g., *Pseudochaeta*, *Winthemia*, *Leucostoma*) and one genus that he described from the United States, *Clastoneuriopsis*, has a species in Chile. Two of Reinhard's generic names (proposed for species in United States and Chile) and two of his species names (both based on Chilean specimens) are junior synonyms. One species described from Argentina, *Winthemia singularis*, is valid and has been recorded from Chile. Reinhard's personal collection was purchased in 1968 by the CNC and his name-bearing types located there are listed in Cooper and O'Hara (1996).

Everardo Eels Blanchard (1899–1971) was born in Buenos Aires and received training in entomology at the University of Maine in United States (Pirán 1972). After returning to Argentina he embarked on a long and illustrious career that encompassed nearly all aspects of entomology. He worked for the Ministerio de Agricultura and in time became director of the Instituto de Sanidad Vegetal. Among other pursuits and responsibilities, Blanchard found time to describe Argentine insects across a broad range of families and orders, with a general focus on those of agricultural importance. The Tachinidae ranked high among the insects he studied and his new taxa comprised ca. 50 genera and 125 species, all described from Argentina. Nearly all of the species names continue to be recognised as valid and six species have been recorded from Chile. Blanchard, like Townsend, had a restricted view of tachinid genera and only half of the ones he described are still considered valid. Three of his genera are recognised in Chile and another 18 generic names are listed in the catalogue below as junior synonyms. Cortés (1973b) wrote an obituary for Blanchard and later (Cortés 1979) named a Chilean species after him (*Ateloglutus blanchardi*). Blanchard has received two other patronyms in Tachinidae, one by Guimarães (1971: 44, replacement name *Pel-eteria blanchardi*) and another by Toma and Guimarães (2002: 45, Ecuadorian species *Leschenaultia blanchardi*). Blanchard's name-bearing types in MACN were discussed in Mulieri et al. (2013).

We come next to the central figure in Chilean tachinidology, Raúl Eduardo Cortés (1915–2001). Cortés was born in the coastal city of Coquimbo in northern Chile. He was educated at the Universidad de Chile in Santiago and spent a couple of years at Harvard University in the United States. His study of tachinid taxonomy began early while he was an entomologist in the Sección Zoología Agrícola of the Departamento de Sanidad Vegetal in the Ministerio de Agricultura, and professor at the Universidad Católica de Chile (both in Santiago). His first taxonomic papers on Tachinidae appeared in 1944, coincidentally the same year as the death of the patriarch of tachinidology at the time, C.H.T. Townsend. Cortés (1944a) published a short biography of Townsend outlining his major achievements. That same year, Cortés (1944b) pub-

lished a concise review of the history of tachinid studies in Chile that provides a good companion to our treatment of this subject here.

A milestone for dipterology in Chile was reached with the 1946 publication of *Catálogo de los dípteros de Chile* under the leadership of Carlos Stuardo [Ortíz] (1895–1962). Just a year before, Cortés (1945e) had dedicated one of his first tachinid genera, *Stuardomyia*, to his senior colleague. The tachinid section of the catalogue was prepared by the young Cortés (1946). Chilean Tachinidae were still relatively little known in the 1940s. Early European authors had described a spattering of species that they had generally assigned to new genera or Old World genera that were familiar to them. Aldrich brought some order to these early names but his emphasis was on the fauna of southern (Patagonian) Chile and he provided no higher classification beyond genus. Townsend then rearranged the entire Tachinidae of the world in his remarkable but idiosyncratic *Manual of Myiology*. Cortés (1946) had to make a choice of whom to follow and he sided with Aldrich:

“Al preparar esta lista de los Tachinidae de Chile, el autor ha querido conservar esencialmente el criterio sistemático con que el Dr. J. M. Aldrich tratara las especies patagónicas en Diptera of Patagonia and South Chile (7, 1:1–170, 1934). Se han introducido, sin embargo, las modificaciones que obvia y naturalmente había que hacer, especialmente en aquellos géneros y grupos que el autor ha podido estudiar con más abundante y representativo material.

Al adoptar este criterio—a pesar de que según Townsend prácticamente ninguna de nuestras especies fué correctamente ubicada por el Dr. Aldrich—el autor ha preferido continuar la línea sistemática por la cual el estudio de nuestros Tachinidae hasta ahora se ha guiado. Un cambio del criterio conservador de Aldrich al concepto extremadamente radical de Townsend, seguramente traería más confusión que beneficios para el estudio futuro de esta familia.”

[In preparing this list of the Tachinidae of Chile, the author wanted to essentially preserve the systematic criterion with which Dr. J. M. Aldrich treated the Patagonian species in *Diptera of Patagonia and South Chile* (7, 1: 1–170, 1934). However, the modifications that obviously and naturally had to be made have been introduced, especially in those genera and groups that the author has been able to study with more abundant and representative material.

By adopting this criterion—although according to Townsend practically none of our species was correctly placed by Dr. Aldrich—the author has preferred to continue the systematic line by which the study of our Tachinidae has so far been guided. A shift from the conservative approach of Aldrich to the extremely radical concept of Townsend would surely bring more confusion than benefits for the future study of this family.] (Cortés 1946: 172.)

Cortés (1946) arranged the known Tachinidae of Chile into 72 genera and 121 species (plus an additional 13 names listed as “Species *incertae sedis*”) and, following the style of the catalogue as a whole, provided no higher classification. The compilation of this catalogue and the earlier review of the history of Chilean Tachinidae studies provided Cortés with a solid understanding of the state of tachinid taxonomy in

Chile at this early point in his life-long dedication to the family. Cortés published ca. 50 papers on mostly Chilean tachinids over a span of 50+ years (González 2001) and during that time described 30 genera (24 currently valid) and 73 species (70 currently valid) of Tachinidae.

Most papers published by Cortés were straightforward taxonomic treatments with descriptions of species and/or genera, keys if appropriate, and notes about types and synonymy. Tachinid taxonomy in Chile advanced incrementally in this fashion for a number of years until the fauna as a whole was reviewed in a comprehensive monograph by Cortés and Hichins (1969) entitled *Distribución Geográfica y Huéspedes Conocidos de los Taquínidos de Chile* [Geographic Distribution and Known Hosts of Chilean Tachinids]. It was the summation of 25 years of collection-building on the part of the senior author and his examination of other collections within and outside (Cortés 1963) the country. The northernmost desert regions of Chile were excluded because only one species was known from there at the time. A careful compilation of distributions and hosts was given for 135 species and another 21 species were listed that were essentially known only from their type specimens and type localities. A further ten names were listed as *incertae sedis* following Cortés (1946). No new taxa were described. The monograph concluded with an interpretative discussion of the biogeographic patterns of tachinid distributions in Chile.

The arid northern portion of Chile was treated a couple of years later when Cortés and Campos (1971) published their findings on *Taquínidos de Tarapacá y Antofagasta*. This monograph recorded 33 genera and 53 species in these regions and described two genera and 15 species as new. A key to genera and a biogeographic discussion were included. One new species was named *Trichophoropsis* [= *Andicesa*] *sabroskyi* in recognition of Curtis Williams Sabrosky (1910–1997), a dipterist with the United States Department of Agriculture (at USNM) who had kindly provided them with information about types and answered their other questions for more than 15 years.

An addendum to Cortés and Campos (1971) was published by Cortés and Campos (1974) and added seven genera to the previous list (including one new genus) and nine species (including four new). A revised key to the genera of these northern regions was given. One new genus was named *Caltagironea* and dedicated to Leopoldo Enrique Caltagirone (1927–present) of the University of California Berkeley. Caltagirone (1953) had studied the life history and biological control potential of *Incamiya chilensis*. Caltagirone (1966) named a new species, *Opsophagus cortesi*, in honour of Cortés.

A second addendum to *Taquínidos de Tarapacá y Antofagasta* by Cortés and Hichins (1979) added new data on a number of previously reported species and also recorded two genera as new to the regions (one a new record for Chile). A third and last addendum was published by Cortés (1984). It brought the number of genera known from the regions to 47 and number of species to 70. One new genus and two new species were described and a new key to genera was included. In 2007, the Government of Chile divided the region of Tarapacá *sensu* Cortés and Hichins (1969, 1979) and Cortés and Campos (1971) into two regions, Arica & Parinacota and Tarapacá.

Chilean entomologist Nelson Hichins published on tachinids twice with Cortés (see above) and once in a sole-authored paper about a survey he conducted near Maipú,

just west of Santiago (Hichins 1969). The specimens collected during that survey were identified by Cortés and 68 species were recorded. A few years earlier, Louis Marnef had described the new genus and species *Lafuentemyia yanezi* based on specimens collected east of Valparaíso by Hichins (Marnef 1965). Cortés (1967b) dedicated the species *Parasetigena* [= *Chetogena*] *hichinsi* to Hichins, who had collected the type series.

Luciano Elliot Campos (1927–1989) was an agricultural entomologist and later dean of the Facultad de Ciencias Agronómicas, Universidad de Chile, Santiago. He coauthored twice with Cortés in the 1970s (see above) but he had also published a paper on tachinids 20 years earlier (Campos 1953) that contained notes on species and the description of a new species, *Poliops auratus*. Cortés (1967a) named the genus *Camposodes* in Campos' honour. An informative and entertaining oral presentation by Campos entitled *Insects – Men and Environment in Chile* was reproduced in *Revista Chilena de Entomología* (Campos 1975).

Two American dipterists who ran Malaise traps throughout Chile in the 1960s and made their material available to specialists were honoured with generic patronyms by Cortés. *Irwinia* Cortés (1967a) [= *Phytomyptera*] was named for Michael Edward Irwin (1940–present) and *Schlingermiya* Cortés (1967b) was named for Evert Irving Schlinger (1928–2004).

An intriguing suggestion was made by Cortés (1983) that the “Trichoprosopini” (the members of which are now included in the Megaprosopini, Tachininae) are the sister group to the New Zealand “Occisorini” [= Proscissionini] based on shared morphological features and host associations. This would represent the first indication of a transantarctic relationship in the Tachinidae. More substantial morphological evidence has not been forthcoming to support this hypothesis and recent molecular evidence contradicts it (Stireman et al. 2019), as does the inferred timeline for tachinid diversification (Cerretti et al. 2017). Although this transantarctic relationship has not been substantiated, the formal transfer of the “Trichoprosopini” from Dexiinae (Guimarães 1971, as “Proseninae”) to Tachininae by Cortés (1986) has been accepted and followed by later authors.

This last mentioned work of Cortés (1986) was another significant regional study reminiscent of *Taquínidos de Tarapacá y Antofagasta* (Cortés and Campos 1971). This one dealt with the southern regions under the title *Taquínidos de Aysén (XI Region) y Magallanes (XII Region) Chile*. It contributed new information about the tachinids of these regions and was based in large part on material accumulated or examined since the monograph of Cortés and Hichins (1969). The fauna comprised 51 genera and 71 species, including four new genera and eight new species.

The size of the Chilean tachinid fauna was cited as 125 genera and 250 species in a short paper about “non-generic characters in Chilean tachinid flies” (Cortés 1989). That same year, Cortés and Herrera (1989) published a detailed history of key figures in Chilean entomology since earliest times and some of the people are discussed above because of their involvement, large or small, with tachinids.

One of the last papers published by Cortés was a review of Chilean Tachinidae and was co-authored by a young graduate student, Christian Raúl González (1963–present)

(Cortés and González 1989). This student is now, more than 30 years later, a co-author on this catalogue of Chilean Tachinidae. González (2001) wrote a tribute to “Professor Raúl E. Cortés Peña” after his death that year. Cortés has been honoured with the following (and possibly more) patronyms: *Opsophagus* [= *Cyrtophloeoba*] *cortesi* Caltagirone (1966; Chile), *Raulcortesia* Artigas & Papavero (1991; Chile, Asilidae), *Leschenaultia cortesi* Toma & Guimarães (2002; Venezuela), *Chaetocnephalia cortesi* González in González & Vergés (2004; Chile) and *Dasyuromyia cortesi* Gramajo (2011; Argentina).

William Robin Thompson (1887–1972) was a Canadian entomologist with a strong background in biological control. He was appointed director of the Farnham House Laboratory of the Imperial Institute of Entomology in England in 1928 and continued to head the Institute as it went through changes in organisation, name, and headquarters. By the time Thompson retired in 1958, it was known as the Commonwealth Institute of Biological Control and headquartered in Ottawa. Thompson then took up a new vocation and spent the next ten years (throughout his 70s) working on the eight volumes of *The Tachinids of Trinidad* (Thompson 1961–1968). The series was unique in incorporating features of larval tachinids for many species and was well illustrated thanks to the artistic talents of his wife Mary.

Thompson’s higher classification of Tachinidae was influenced more by the great contemporary masters of European Tachinidae, Louis-Paul Mesnil (1904–1986) and Benno Wilhelm Herting (1923–2004), than by Townsend’s *Manual of Myiology*. Nevertheless, he tended towards restricted genera in *The Tachinids of Trinidad* and described 41 from the island. Only 18 are still valid and not all of them have been re-evaluated. Most of the synonymy was proposed in Wood’s (1985) conspectus of the Blondeliini and as a result seven of Thompson’s generic names are listed in the catalogue below as synonyms of *Eucelatoria* and *Myiopharus*.

The Brazilian José Henrique Guimarães (1937–2008) was educated at agricultural schools in Rio de Janeiro before beginning the study of Tachinidae as an intern at the Laboratório de Zoologia Médica e Parasitologia da Escola Nacional de Veterinária and the Instituto Oswaldo Cruz, in Rio (Lamas et al. 2009). A series of five revisionary papers on *Archytas* resulted from these studies (Guimarães 1960, 1961a, b, 1963a, b) in addition to several papers on other tachinine tachinids. In 1963 he was hired as a biologist in São Paulo at what is now the Museu de Zoologia, Universidade de São Paulo (MZSP). A scholarship permitted Guimarães to spend 1967 at the Smithsonian Institution (USNM) to work on a much-needed catalogue of Neotropical Tachinidae. Not only were the resources of the Smithsonian available to Guimarães at this time, in particular the collection (including the majority of Townsend’s types) and library, but the taxonomic and nomenclatural advice of resident USDA dipterist Curtis Sabrosky (details above) was indispensable. The catalogue was completed over the next several years and published in the series *Catalogue of the Diptera of the Americas South of the United States* (Guimarães 1971). Guimarães left Washington for California to start a revision of North American *Winthemia* at the University of California Riverside for a Master’s degree. With that completed he returned to Brazil in 1970 to resume his former position and to begin study of the Mesembrinellidae for

a Ph.D. degree, completed in 1973. Guimarães remained at the Museu de Zoologia until 1985 and then moved on to other positions before retiring in 1993 (Lamas et al. 2009). His sole-authored papers on Tachinidae ended in 1983 but he assisted with other tachinid papers up to 2002. Guimarães did not describe any Chilean Tachinidae but his revisions helped people like Cortés sort out the Chilean species of the groups that were revised.

Guimarães' most influential and enduring contribution to tachinidology was his 1971 tachinid catalogue. Sabrosky and Arnaud (1965) had recently published on the Tachinidae in *A Catalog of the Diptera of America North of Mexico* and a companion catalogue for the Tachinidae of the southern portion of the Americas was sorely needed. This was not an easy task because the number of species was much greater and the existing classification (due in large measure to Townsend) was more bewildering. His result was a progressive catalogue, to a point. As Guimarães (1971: 3) conceded, the "catalogue arrangement leaves much to be desired". This is true but the result must be viewed in the context of the task at hand, which was a compilation of data on a difficult fauna of close to 3000 species and ca. 950 genera. After 50 years this catalogue is still the chief resource for anyone studying Neotropical Tachinidae.

The Peruvian entomologist Luis A. Valencia (1945–present) has also contributed to Chilean Tachinidae. He co-authored with Cortés on a partial revision of the genus *Ateloglutus* Aldrich (Cortés and Valencia 1972), described the new genus and species *Velardemyia ica* (Valencia 1972a) and described a new species of *Winthemia* [*W. roblesi*, since synonymised with *W. singularis* Reinhard] (Valencia 1972b).

Argentine entomologist María Cecilia Gramajo (1973–present) has published several papers on the Tachinidae of Argentina. In a preliminary list of Tachinidae of Patagonian Argentina, Gramajo (1998) recorded 80 species of which ca. 20 were originally described from Chile and were newly recorded from Argentina. A paper on *Dasyuromyia* (Gramajo 2011) has a key to species that includes all but one of the Chilean species.

Xuekui Sun (1963–present) emigrated from China to Canada and completed a Ph.D. thesis on *Phasia* under the supervision of dipterist Stephen Archer Marshall (1954–present) at the University of Guelph, Ontario. The *Phasia* revision treated the species of the world except for the Neotropical ones. Seven generic names based on New World species were newly synonymised with *Phasia* in Sun and Marshall (2003) and are listed in the catalogue below.

The Brazilian tachinid specialist Silvio Shigueo Nihei (1976–present) at the Universidade de São Paulo has published primarily on the Brazilian fauna. A few of his publications are relevant to the Chilean fauna or to the list of generic synonyms in the catalogue below (Nihei 2015, 2016; Nihei and Dios 2016). A former student of Nihei's, Brazilian Rodrigo de Vilhena Perez Dios (1987–present), published a revision of *Ectophasiopsis* that includes the single species known from Chile (Dios and Nihei 2017).

A prominent figure among Chilean entomologists was Luis Enrique Peña (1921–1995), a great explorer and professional collector with a vast knowledge of the country and its fauna and flora, particularly its insects. He worked for a short time at the Universidad de Chile in Santiago and elsewhere, but his true calling was travelling

throughout Chile and neighbouring countries in search of insects, often assisting specialists from abroad who valued his expertise in the field. More than 400 species bear his name and he described more than 100 species of Tenebrionidae (Coleoptera). He did not name any tachinids but he collected many of them and his specimens are in Chilean collections, CNC and elsewhere. He collected the type series of both *Caltagironea vera* (Cortés and Campos 1974) and *Enchomyia penai* [= *Comops ruficornis*] (Cortés 1967b) and the latter species was named in his honour.

The authors of this present catalogue have been involved with the Chilean Tachinidae to a greater or lesser degree as reviewed below.

Donald Montgomery ["Monty"] Wood (1933–2020) completed a Ph.D. degree on black flies (Simuliidae) at McMaster University in Hamilton, Ontario, in 1963 and was hired at CNC the following year to work on black flies and other families. Wood had been interested in tachinids for several years and the family came to dominate his research time and collecting activities. He soon realised that the New World classifications of Sabrosky and Arnaud (1965) and Guimarães (1971) were still overly influenced by Townsend's *Manual of Myiology* and saw much promise in the restructuring of European Tachinidae that was underway by Mesnil and Herting (see above). The result was a great pruning of generic names in his *Taxonomic Conspectus of the Blondeliini of North and Central America and the West Indies* (Wood 1985) and tachinid chapter in *Manual of Nearctic Diptera* (Wood 1987; synonymy reviewed in O'Hara and Wood 1998). Together, these two works proposed close to 400 new generic synonyms for New World genera with species north of continental South America. Included among these were quite a few generic names that are listed as synonyms in the catalogue below. A later chapter on Tachinidae and key to genera in *Manual of Central American Diptera* (Wood and Zumbado 2010) only provides a small measure of assistance with the Chilean genera because a significant number of the genera are not included. There are a few generic names mentioned as synonyms for the first time in Wood and Zumbado (2010) and also the occasional inflation of species numbers that reflect unstated generic synonymies.

Wood retired in 1986 and was an honorary research associate with CNC until his death a few months before this manuscript was completed. He continued to collect and build his knowledge of New World Tachinidae after retirement and was working towards a revised generic classification of the entire fauna until the final months of his life. He collected extensively throughout the Neotropics and built a large and significantly curated private collection. He augmented his own collecting efforts with specimens purchased from professional collectors Fritz Plaumann (Brazil) and Luis Peña (Chile). Wood and wife Grace collected in Chile and Argentina with Luis Peña in late 1993 to early 1994 and returned to Ottawa with a broad assortment of beautifully-mounted tachinids that were mostly sorted and identified before being donated to the CNC during the past few years. These tachinids from Peña and M. and G. Wood were helpful in understanding the Chilean fauna during the preparation of this paper.

James Edward O'Hara (1952–present) first became interested in Tachinidae as a summer student at CNC in 1977 while pursuing a B.Sc. degree at nearby Carleton University. There he was influenced by Monty Wood to undertake a revision of the

North American species of *Siphona* for a Master's degree, which was completed at the University of Alberta, Edmonton, under the supervision of coleopterist George E. Ball in 1981. The *Siphona* revision included a new generic synonym (*Phantasiosiphona*) listed in the catalogue below and a cautionary note that the European species *Siphona geniculata* is likely misidentified from Chile and elsewhere in South America (O'Hara 1983). The tribe Siphonini was revised for a Ph.D. degree, also under the supervision of Ball and completed in 1987 (O'Hara 1989). In that revision, Chilean species formerly in *Actia* were moved to *Ceromya* and subgenera of *Siphona* were recognised. O'Hara was hired into his present position with CNC in 1989. A later revision of North American Polideini and a reinterpretation of the tribe is followed here (O'Hara 2002). Specimens collected in Chile by O'Hara in late 2015 (Stireman et al. 2016) helped with the preparation of this paper. The most recent version of the *Preliminary Checklist of the Tachinidae of the World* (O'Hara et al. 2020) includes the current names and distributions of Chilean Tachinidae.

Christian Raúl González (1963–present) (also see above) became interested in Tachinidae as a student at the Universidad Metropolitana de Ciencias de la Educación (UMCE) in Santiago in 1986. There, he was influenced by Raúl Cortés to prepare a *List of Tachinidae from Chile* for an undergraduate thesis, which was completed under the supervision of Cortés in 1988. González then worked on Tabanidae for a Master's degree at the Universidad Metropolitana under the supervision of Sixto Coscarón. In 1989, González was hired into his present position at the same university. That same year, Cortés and González (1989) published a review of the Voriini of Chile, recognising nine genera and 16 species including a new genus with one new species (*Nothovoria praestans*). Other single-authored contributions on the Chilean Tachinidae consisted of reviews of the genus *Ateloglutus* (González 1989) and former tribe Cuphocerini (now Polideini and Tachinini) (González 1992a), and a survey of the tachinids of Reserva Nacional de Río Clarillo near Santiago (González 1992b). Collaborations resulted in reviews of the Chilean species of *Incamiya* (González and Henry 1992) and the Chilean Goniini (González and Vergés 2004). The single new species described in the latter was named *Chaetocnephalia cortesi* in honour of González's mentor.

Materials and methods

Format

This catalogue is arranged and formatted in a similar manner to the Tachinidae of the Afrotropical Region by O'Hara and Cerretti (2016). The sections here under Format are similar to the same sections in that work but are repeated here as a convenient guide and have been modified to apply to the Chilean Tachinidae. Any changes in format or interpretation of nomenclatural matters as compared to O'Hara and Cerretti (2016) are noted.

General

This catalogue cites all the species of Chile in their valid and original combinations, provides details about the name-bearing types of all nominal species, and gives known distributions. It is based on the examination of virtually all of the approximately 450 publications listed in the References.

Valid names are arranged hierarchically and alphabetically according to the categories of subfamily, tribe, genus, subgenus, and species. Synonyms are given for valid names of genera, subgenera, and species, and are listed chronologically. Synonymic lists comprise taxa described from south of the United States, synonyms that have been used as valid names in the literature on Chilean Tachinidae, and (where known and listed last) misidentifications and incorrect spellings.

Each genus-group name is listed with the following information: genus name in italics and capital letters (and additionally in bold if valid, unless misidentified from Chile; e.g., *Neoethilla* Cerretti et al.), author, year (with letter if applicable), page, note in parentheses if applicable (e.g., junior homonym or proposed as subgenus), type species with author and date, form of type fixation, and country (or region, such as Europe, if country unknown) of the type locality of the type species in square brackets (the last not given for all generic names in O'Hara and Cerretti 2016). Each type species is cited in its original binomen (Recommendation 67B of the *Code*, ICZN 1999), and if that name is a synonym then it is followed by the valid name of the species in parentheses. We have invoked Article 70.3.2 of the *Code* (ICZN 1999) to fix the intended species as the type species for generic names that were based on misidentified type species. This maintains the concepts of these generic names as currently accepted and in prevailing usage. The genera so affected are listed below under "Summary of new taxonomic and nomenclatural changes".

Type species were fixed by original designation, monotypy, subsequent designation, or in a few instances subsequent monotypy, except for type species newly fixed here for nominal genera based on misidentified type species. Fixation by original designation requires an explicit designation of a type species (Article 68.2 of the *Code*, ICZN 1999), so a new genus "proposed for" or "erected for" a single species has its type species fixed by monotypy. A new genus proposed before 1931 for a single species and accompanied by the expression "gen. n., sp. n." or an equivalent also has its type species fixed by monotypy (Article 68.2.1). If, on the other hand, the new genus is proposed for more than one new species and the expression "gen. n., sp. n." or an equivalent is applied to only one of the new species, then that species is fixed as type species by original designation (Article 68.2.1).

Species are listed by valid name followed by the available name(s) associated with it; i.e., the available name of the valid name plus synonyms. The valid name is represented by the valid specific epithet in bold and italics (in italics only if questionably recorded or misidentified from Chile; e.g., *Archytas incertus* (Macquart)) followed by the author, date (no letter suffix), and known distribution. Author and date are enclosed in parentheses if the species has moved from its original genus. The distribution is given first

for the Neotropical Region and then for other regions as explained under “Geographic divisions” and “Distributional data”. Each available name is given in italics in its original combination and spelling followed by author, year (with letter suffix if applicable to match a publication listed in the References), page, and a note in parentheses if applicable (e.g., junior homonym or subsequent spelling). A questionable synonym is preceded by a question mark (e.g., “? *Spathipalpus flavifrons* Rondani”). Given next is name-bearing type information consisting of status (holotype, lectotype, neotype, or syntypes), sex (of single type, or number and sex of syntypes), type depository (in parentheses), and type locality. If a neotype or lectotype was designated then a citation is given to the designation. Additional information may be given in parentheses with the type depository to cite the number and sex of syntypes existing in a collection if that number is different from the information given in the original description, or if the original description did not provide details about the type series; also, a reference may be cited wherein information can be found about the name-bearing type.

A subsequent spelling of a generic or specific name can be an incorrect subsequent spelling (which is not an available name) or an unjustified emendation (which is an available name with its own author and date). Incorrect subsequent spellings are cited where known to us but others surely exist. An unjustified emendation is cited with an author and date following O'Hara and Cerretti (2016); a name only was given in O'Hara and Wood (2004) and O'Hara et al. (2009) except in rare instances.

Notes and/or references are often given after genus and species entries. Notes provide explanations of some sort; e.g., priority of names, composition of type series, justification for a new combination or new name. References have a standardised format consisting of a source followed by the information provided therein; e.g., first records from countries (as explained under “Distributional data”), redescriptions, keys, figures, type notes. These references attempt to trace the history of name usage and synonymy but do not cite every occurrence of a name in species lists (unless it is a first record from a country).

The following abbreviations are used:

- Code** *International Code of Zoological Nomenclature*, specifically the fourth edition published by the International Commission on Zoological Nomenclature in 1999; cited as ICZN 1999.
- ICZN** International Commission on Zoological Nomenclature.
- JEOH** James E. O'Hara.
- DMW** D. Monty Wood.
- CRG** Christian R. González.

Name-bearing types

We follow the same method developed by O'Hara et al. (2009) and followed by O'Hara and Cerretti (2016) for citing name-bearing type information for species described without a holotype designation in the original publication or without a subsequent lectotype or neotype designation. Details are provided about name-bearing types based on the content of the original descriptions and are not biased by existing type material

in collections (that information being given in parentheses with the type depository). Our format for citing published data on name-bearing types other than a designated holotype, lectotype or neotype is as follows:

Type(s), male: One or more males. This citation is used for a species described from the male sex without indication of whether a single male (i.e., a holotype) or more than one male (i.e., syntypes) composed the type series.

Type(s), female: One or more females. See “Type(s), male”.

Type(s), unspecified sex: One or more specimens with no indication of sex.

Syntypes, [number] male[s] and [number] female[s] (e.g., “Syntypes, 3 males and 2 females”): Species described from an indicated number of males and females.

Syntypes, males and females: Species described from both sexes but the number of each sex was not given. A number in front of “males” with no number in front of “females” refers to the total number of males and females.

Syntypes, males: Species described from more than one male but without indication of the number of males.

Syntypes, females: Species described from more than one female but without indication of the number of females.

Syntypes, unspecified number and sex: Species described from more than one specimen but without indication of sex or number of specimens.

Avoidance of assumption of holotype

In following the foregoing format we have complied with Recommendation 73F of the *Code* (ICZN 1999), “Avoidance of assumption of holotype”, which states: “Where no holotype or syntype was fixed for a nominal species-group taxon established before 2000, and when it is possible that the nominal species-group taxon was based on more than one specimen, an author should proceed as though syntypes may exist and, where appropriate, should designate a lectotype rather than assume a holotype (see also Article 74.6)”. See O’Hara et al. (2009: 9–10) for a further discussion of this issue.

By following Recommendation 73F of the *Code*, assumed holotypes take on the status of syntypes. The recommendation favours “where appropriate” the designation of lectotypes. We have combined the spirit of Recommendation 73F and the provisions of Article 74.5 of the *Code* (ICZN 1999) to recognise certain published statements (as discussed in the next section) about assumed holotypes as lectotype fixations. This follows O’Hara et al. (2009) and O’Hara and Cerretti (2016) and is in our opinion the best way to reconcile assumed holotypes with the modern rules of nomenclature, while also giving credit of lectotype fixations to the authors who assumed holotypes.

Lectotypifications

There are two types of lectotypification in zoological nomenclature, explicit and implicit. In the former, a single syntype in a type series is designated as lectotype; in the latter, there is some form of statement that can be construed as the selection of a single name-bearing type. We follow O’Hara et al. (2009) in using the term “lectotype

designation” for an explicit lectotypification and “lectotype fixation” for an implicit lectotypification. There is good reason to distinguish between the two because implicit lectotypifications are open to some interpretation, especially with respect to Article 74.5 of the *Code* (ICZN 1999: 82–83) that deals in part (see also Article 74.6) with lectotype designations before 2000:

“In a lectotype designation made before 2000, either the term ‘lectotype’, or an exact translation or equivalent expression (e.g. ‘the type’), must have been used or the author must have unambiguously selected a particular syntype to act as the unique name-bearing type of the taxon. When the original work reveals that the taxon had been based on more than one specimen, a subsequent use of the term ‘holotype’ does not constitute a valid lectotype designation unless the author, when wrongly using that term, explicitly indicated that he or she was selecting from the type series that particular specimen to serve as the name-bearing type”.

What constitutes a valid lectotypification (or lectotype fixation in our terminology) in the foregoing is largely dependent on how one interprets the passage about an author explicitly indicating “that he or she was selecting from the type series that particular specimen to serve as the name-bearing type”. At one end of the spectrum is the mere mention of a “holotype” or “type” by a subsequent author when the original type series clearly consisted of two or more syntypes. This statement does not constitute a lectotype fixation because the “holotype” is not distinguishable from other syntypes. At the other end of the spectrum is the mention of a “holotype” or “type” with accompanying details about its labelling, features, damage, etc. that clearly distinguishes that specimen from other syntypes; or perhaps there is only one type specimen in a collection and it is an “assumed holotype” (see section above) for a species described from an unspecified number of specimens. We considered these latter statements about a single type to qualify as lectotype fixations under Article 74.5 because they contain an explicit indication that an author accepted the cited “holotype” or “type” as the name-bearing type and restricted the term to a single recognisable specimen in a collection.

O'Hara and Cerretti (2016) recognised lectotype fixations in Townsend's *Manual of Myiology* (Parts I–XII, 1934–1942), reversing the practice of O'Hara et al. (2009). We follow the former authors in recognising lectotype fixations in *Manual of Myiology* if there is a strong possibility of the lectotype being recognised in the stated collection. See O'Hara and Cerretti (2016: 11–12) for a more detailed discussion of this subject.

Type localities

Type localities are cited first by country and then by location within the country from larger to smaller geographic area or place. Spellings of geographic areas and places follow *The Times Comprehensive Atlas of the World* (Times Books 2007), if found in that work. Modern names and spellings are given where these have been determined. Country and higher administrative subdivisions (i.e., regions and provinces of Chile,

provinces of Argentina, states of Brazil, regions of Peru, etc.) are given only in their modern equivalents. For locality names that have changed since they were first published, the modern spelling is given first followed by the original spelling in square brackets and quotes; e.g., Puerto del Hambre [as “Port Famine”]. Elevations are cited in metres (m) or feet (ft) as given by the author. Coordinates given in an original publication are cited in parentheses after the type locality; e.g., Chile, Arica y Parinacota, Parinacota, Putre, 3530 m (18°12’S, 69°35’W). Coordinates are included for many type localities that we had difficulty locating and these are given in square brackets after the locality to distinguish them from coordinates provided by an author; e.g., Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9’S, 71°18’W]. Criteria for citing type localities in Sweden are explained in O’Hara et al. (2009: 11). The few localities we could not find are given in quotes; e.g., Trinidad, “Legerville Mt.”. A variety of resources were used to locate type localities not found in *The Times Comprehensive Atlas of the World* including other atlases, maps, literature, and Internet searches (often for the locality and/or collector).

Type localities in Chile are preceded by region and province (e.g., Valparaíso, Marga Marga). Those in other countries are preceded by province (Argentina, Ecuador, etc.), state (Brazil, Mexico, etc.) or department (Peru, Uruguay, etc.), or by no higher administrative division (e.g., countries of the West Indies).

Collections housing name-bearing types

The location of the name-bearing type (holotype, lectotype, neotype, or syntypes) is cited for each nominal species, where known. The collections housing these name-bearing types are listed below with the abbreviations used in the text. We largely accepted as accurate the statements about the deposition of name-bearing types given in the original literature unless we had reason to doubt the information given (e.g., types known to have been relocated or presumed lost).

The abbreviations of collections cited in this work are as follows:

AMNH	American Museum of Natural History, New York, USA.
CAS	California Academy of Sciences, San Francisco, California, USA.
CNC	Canadian National Collection of Insects, Arachnids and Nematodes, Agriculture and Agri-Food Canada, Ottawa, Canada.
CUIC	Cornell University Insect Collection, Department of Entomology, Cornell University, Ithaca, New York, USA.
EEAM	Estación Experimental Agronómica, Universidad de Chile, Maipú, Santiago, Chile. Cited as CEA in publications of R. Cortés.
EESC	Estación Experimental San Camilo [formerly Estación Experimental Agrícola], Ica, Peru.
INLA	INIA Subestación Experimental Control Biológico La Cruz, La Cruz, Chile. Cited as CENE [Estación Nacional de Entomología de La Cruz] in Cortés (1967b), Cortés and Hichins (1969) and Cortés and Campos (1971).

INTA	Instituto Nacional de Tecnología Agropecuaria, Castelar, Argentina.
MACN	Museo Argentino de Ciencias Naturales “Bernardino Rivadavia”, Buenos Aires, Argentina. Cited as MAHN in Cortés (1963).
MCZ	Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, USA.
MEUC	Museo Entomológico Luis Peña del Departamento de Sanidad Vegetal de la Facultad de Ciencias Agronómicas, Universidad de Chile, La Pintana, Santiago, Chile. Cited as DSV [Departamento de Sanidad Vegetal] in Cortés (1945c–e) and as CFA [Colección de la Facultad de Agronomía] in later publications of R. Cortés.
MLPA	Museo de La Plata, Universidad Nacional de La Plata, La Plata, Argentina.
MNHN	Muséum National d'Histoire Naturelle, Paris, France.
MNNC	Museo Nacional de Historia Natural, Santiago, Chile. Cited as CNI [Colección Nacional de Insectos, Ministerio de Agricultura, Santiago] in Cortés (1951b) and Campos (1953).
MZSP	Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.
MZUF	Museo Zoologico “La Specola”, Firenze [Florence], Italy.
MZUT	Museo e Istituto di Zoologia Sistemica dell'Università di Torino, Turin, Italy.
NHMUK	Natural History Museum, London, United Kingdom. Frequently cited as BMNH [British Museum (Natural History)] in previous publications.
NHMW	Naturhistorisches Museum Wien, Wien [Vienna], Austria.
NHRS	Naturhistoriska Riksmuseet [Swedish Museum of Natural History], Stockholm, Sweden.
NMPC	National Museum, Natural History Museum, Prague, Czech Republic.
RBINS	Royal Belgian Institute of Natural Sciences, Bruxelles [Brussels], Belgium. Frequently cited as IRSNB [Institut Royal des Sciences Naturelles de Belgique] in former publications.
RMNH	Naturalis Biodiversity Center, Leiden, Netherlands [formerly Nationaal Natuurhistorisch Museum and before that Rijksmuseum van Natuurlijke Historie]. The Zoölogisch Museum of the University of Amsterdam [ZMAN] has closed and its collections were merged with those of RMNH.
SDEI	Senckenberg Deutsches Entomologisches Institut, Leibniz-Zentrums für Agrarlandschaftsforschung, Müncheberg, Germany.
SEMC	Snow Entomological Museum Collection, KU Biodiversity Institute, University of Kansas, Lawrence, Kansas, USA.
SENASA	Laboratorio de Sanidad Vegetal, Servicio Nacional de Sanidad Agraria, Lima, Peru.
SMF	Forschungsinstitut und Naturmuseum Senckenberg, Frankfurt am Main, Germany.
UMCE	Universidad Metropolitana de Ciencias de la Educación, Santiago, Chile.
USNM	National Museum of Natural History [formerly United States National Museum], Smithsonian Institution, Washington, USA.

UVVC Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile. Cited as “Instituto de Biología, Universidad de Chile, Valparaíso” in Marnef (1965) and as CDCV [Colección del Departamento de Ciencias, Universidad de Chile, Valparaíso] in Cortés and Hichins (1969).

Imaging of specimens

Habitus images of *Billaea* Robineau-Desvoidy name-bearing types (Fig. 3a–d) were taken by DMW during a visit to USNM in 2011. The equipment used was not recorded and specimens were not measured.

Specimens shown in Figs 4–6 belong to CNC and were imaged using a Canon EOS 70D Digital SLR camera body mounted on a Kaiser RS1 copy stand. A Canon EF 100 mm f/2.8 macro lens was used to image the medium to large specimens and a Canon MP-E 65 mm 1–5× macro lens was used for small specimens. Helicon Remote software was used to tether the camera to a computer to capture images remotely and control lens functions (shutter, shutter speed, aperture, and focus). Lighting was provided by a ring light comprising 80 LEDs with a specimen holder in the middle. A dome cover with a reflective white coating was placed over the ring light and a hole in the centre of the dome permitted images to be taken of the specimen within. A series of images were captured using the Helicon Remote software paired with a Stackshot Macro Rail hardware package by Cognisys. The series of images was stacked using Zerene Stacker and final images were prepared in Adobe Photoshop Creative Cloud 2018. Specimen measurements given in the captions to Figs 4–6 refer to body length from the pedicel of the antenna to the tip of the abdomen, excluding setae.

Geographic divisions

The known distribution of each tachinid species recorded from Chile is given next to the valid name in the following order: Neotropical Region, Nearctic Region, Palearctic Region, Afrotropical Region, Oriental Region, and Australasian and Oceanian regions. These regions are delimited and mapped in O’Hara et al. (2020: 8–26). Each region is subdivided according to the scheme explained below, with the Neotropical Region subdivided more finely than the other regions. Geographical names of countries, political divisions within countries, places, and topographical features follow *The Times Comprehensive Atlas of the World* (Times Books 2007), if given therein. The abbreviations and names given below are those used for the distributions given in the catalogue.

Neotropical Region (Figs 1, 2)

Greater Antilles (part of the West Indies).

Bahamas; Cayman Islands (United Kingdom Overseas Territory); Cuba; Dominican Republic; Haiti; Jamaica; Puerto Rico; Turks & Caicos (United Kingdom Overseas Territory).

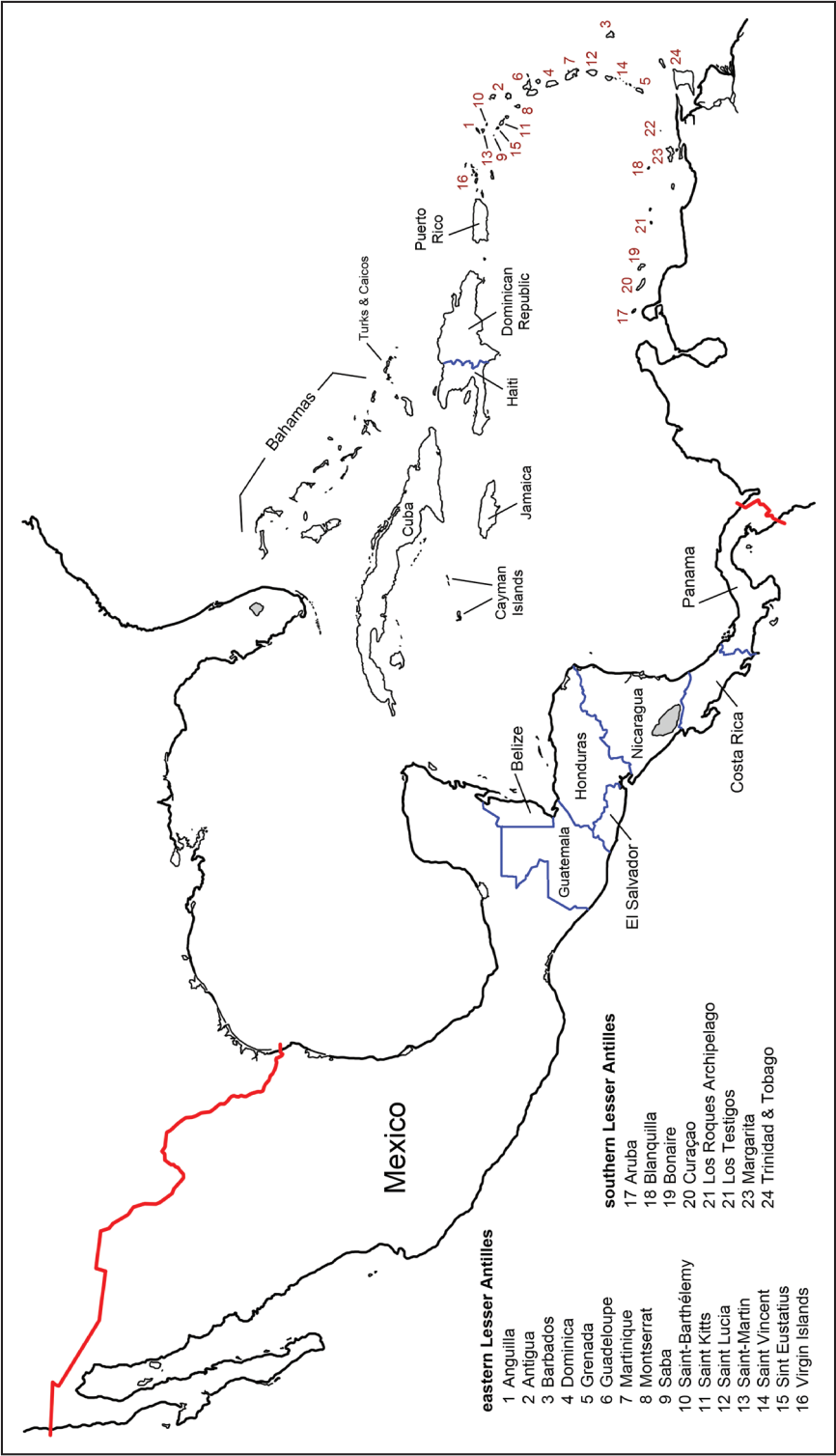


Figure 1. Countries and major islands of the Middle American portion of the Neotropical Region.



Figure 2. Countries and major islands of the South American portion of the Neotropical Region.

eastern Lesser Antilles (Leeward and Windward islands in the Lesser Antilles of the West Indies).

Anguilla (United Kingdom Overseas Territory); Antigua [Antigua & Barbuda] (including Redonda); Barbados; Dominica; Grenada; Guadeloupe (including Marie-

Galante, La Désirade, Îles des Saintes) (France); Martinique (France); Montserrat (United Kingdom Overseas Territory); Saba (Netherlands); Saint-Barthélemy (France); Saint Kitts [Saint Kitts and Nevis]; Saint Lucia; Saint-Martin (comprising Saint Martin [France] and Sint Maarten [Netherlands]); Saint Vincent [Saint Vincent and The Grenadines]; Sint Eustatius (Netherlands); Virgin Islands (including the United States islands of Saint Thomas, Saint John, and Saint Croix, and the British Virgin Islands of Tortola, Virgin Gorda, Anegada, and Jost Van Dyke).

southern Lesser Antilles (islands north of the Venezuelan coast in the Lesser Antilles of the West Indies).

Aruba (Netherlands); Blanquilla (Venezuela); Bonaire (Netherlands); Curaçao (Netherlands); Los Roques Archipelago (Venezuela); Los Testigos (Venezuela); Margarita (including smaller neighbouring islands, principally La Tortuga, Coche, and Cubagua; all comprising Nueva Esparta state, Venezuela); Trinidad & Tobago.

Middle America (mainland Middle America).

Belize; Costa Rica; El Salvador; Guatemala; Honduras; Mexico; Nicaragua; Panama.

South America. [Cited as South America when more detailed distributional data is not available.]

Argentina; Bolivia; Brazil; Chile (excluding Juan Fernández Islands); Colombia; Ecuador (excluding Galápagos Islands); Falkland Islands (disputed United Kingdom Overseas Territory); French Guiana (France); Juan Fernández Islands (Chile); Galápagos Islands (Ecuador); Guyana; Paraguay; Peru; South Georgia (including the South Sandwich Islands; disputed United Kingdom Overseas Territory); Suriname; Uruguay; Venezuela.

Nearctic Region

The limits of the Nearctic Region follow O'Hara et al. (2020: 8, 18 [map 1]) and include the following subdivisions: Bermuda (United Kingdom Overseas Territory); Canada; Greenland (Denmark); United States [United States of America, as "USA" for type localities; Hawaii as part of Australasian and Oceanian regions].

Palearctic Region

See O'Hara et al. (2020: 11, 21 [map 4], 22 [map 5]) for the countries included in the broader subdivisions recognised here: Central Asia; China (Palearctic part, *sensu* O'Hara et al. 2020); Europe; Japan (excluding Ryukyu Islands); Kazakhstan; Korean Peninsula; Middle East; Mongolia; North Africa; Russia; Transcaucasia.

Afrotropical Region

This region is subdivided by country, as explained and mapped in O'Hara and Cerretti (2016: 15, 16 [fig. 1]) and O'Hara et al. (2020: 13, 23 [map 6]).

Oriental Region

The Oriental Region is bounded on the north by the Palaearctic Region (O'Hara et al. 2020) and on the south by Weber's Line (Evenhuis 1989: 31). See O'Hara et al. (2020: 15, 22 [map 5], 24 [map 7], 25 [map 8]) for a list of countries/subdivisions and maps.

Australasian and Oceanian regions

The Australasian and Oceanian regions are bounded on the north by Weber's Line (Evenhuis 1989: 31). See O'Hara et al. (2020: 16, 26 [map 9]) for a list of countries/subdivisions and maps.

Sample distribution

A species recorded from all regions and subdivisions recognised here would be cited with the following distribution:

Neotropical: Greater Antilles (Bahamas, Cayman Islands, Cuba, Dominican Republic, Haiti, Jamaica, Puerto Rico, Turks & Caicos), eastern Lesser Antilles (Anguilla, Antigua, Barbados, Dominica, Grenada, Guadeloupe, Martinique, Montserrat, Saba, Saint-Barthélemy, Saint Kitts, Saint Lucia, Saint-Martin, Saint Vincent, Sint Eustatius, Virgin Islands), southern Lesser Antilles (Aruba, Blanquilla, Bonaire, Curaçao, Los Roques Archipelago, Los Testigos, Margarita, Trinidad & Tobago), Middle America (Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama), South America (Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Falkland Islands, French Guiana, Juan Fernández Islands, Galápagos Islands, Guyana, Paraguay, Peru, South Georgia, Suriname, Uruguay, Venezuela). Nearctic: Bermuda, Canada, Greenland, United States. Palaearctic: Central Asia, China [Pal.], Europe, Japan, Kazakhstan, Korean Peninsula, Middle East, Mongolia, North Africa, Russia, Transcaucasia. Oriental: Andaman & Nicobar Islands, Bangladesh, Brunei, Bhutan, Cambodia, China [Orien.], Christmas & Cocos Islands, India, Indonesia [Orien.], Japan [Ryukyu Islands], Laos, Malaysia, Maldives etc., Myanmar, Nepal, Pakistan, Philippines, Singapore, Sri Lanka, Taiwan, Thailand, Vietnam. Australasian & Oceanian: Australia, Hawaii, Indonesia [Aust.], Melanesia, Micronesia, New Zealand, Papua New Guinea, Polynesia.

Distributional data

Distributions within the Neotropical Region

Distributions are cited at the country level within the Neotropical Region (as listed in Geographic Divisions section) for each species based on published records and our examination of specimens in CNC and UMCE. The principal sources for published

records were Aldrich (1934) and the publications of Cortés (and co-authors, 1944–1992, mainly Chile) and Blanchard (1935–1966, mainly Argentina). The CNC has more than 3000 Chilean specimens but few have been reported in the literature. These mostly originated from the following sources: nearly 2000 collected by the esteemed Chilean entomologist Luis E. Peña [Guzmán] from 1959–1980 and 1994–1995 (privately sold to DMW over a period of years and recently donated to CNC); ca. 1000 collected by DMW and wife Grace between December 1993 and February 1994 (recently donated to CNC and including ca. 150 specimens from Argentina); and over 200 collected by JEOH in December 2015 (see Stireman et al. 2016).

A reference is cited, if known, for the first record of a species from countries different from the one(s) from which the species was described. Subsequent records from the same country are not generally given unless significant in some way. The first record is considered the most important because it is sometimes the source for later records even if it was based on a misidentification.

Classification

Summary of new taxonomic and nomenclatural changes

Genera newly recorded from Chile

Two genera are newly recorded from Chile (one also newly recorded from Argentina).

Chaetoepalpus Vimmer & Soukup, 1940 (based on new records of *Chaetoepalpus coquilleti* Vimmer & Soukup, 1940). New records from Argentina and Chile.

Patelloa Townsend, 1916 (based on new synonymy of *Macropatelloa* Townsend, 1931 with *Patelloa*). New record from Chile.

Species newly recorded from Chile

The following species are newly recorded from Chile or other countries.

Lypha ornata Aldrich, 1934. New record from Chile.

Chaetoepalpus coquilleti Vimmer & Soukup, 1940. New records from Argentina and Chile.

Phytomyptera evanescens (Cortés, 1967). New record from Argentina.

Xanthobasis unicolor Aldrich, 1934. New record from Chile.

Species misidentified or misrecorded from Chile

Species newly recognised as misidentified or misrecorded from Chile are listed here. The reasons for not recognising them from Chile are given under each name in the catalogue.

- Archytas incertus* (Macquart, 1851).—Not Chile [Argentina, Brazil, Paraguay, Uruguay].
Archytas seminiger (Wiedemann, 1830).—Not Chile [Brazil, Colombia].
Gonia crassicornis (Fabricius, 1794).—Not Chile [Brazil, Peru, Venezuela; also Middle America, West Indies and Nearctic].
Lespesia andina (Bigot, 1888), *nomen dubium*.—Not Chile [Cuba].
Lespesia archippivora (Riley, 1871).—Not Chile [widespread throughout the Nearctic Region and most of Middle and South America].
Neoethilla ignobilis (van der Wulp, 1890).—Not Chile [Mexico; United States].
Siphona (*Siphona*) *geniculata* (De Geer, 1776).—Not Chile [Nearctic (introduced), Palearctic].
Winthemia quadripustulata (Fabricius, 1794).—Not Chile [Palearctic; also Nearctic and Oriental].

First Reviser actions

Billaea Robineau-Desvoidy, 1830

Paratheresia rufiventris Townsend, 1929 and *Sarcoprosena rufiventris* Townsend, 1929 are secondary homonyms when placed together in *Billaea*. As the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999), we fix *Paratheresia rufiventris* as the senior homonym.

Myiopharus Brauer & Bergenstamm, 1889

Mayophorinia angusta Townsend, 1927 and *Metarrhinomyia angusta* Townsend, 1927 are secondary homonyms when placed together in *Myiopharus*. As the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999), we fix *Mayophorinia angusta* as the senior homonym.

New replacement names

Eight new names are proposed for preoccupied names that came to our attention during the preparation of this catalogue and belong to genera recorded from Chile. The preoccupied names do not concern Chilean species except for one but are renamed to avoid the confusion of having two Neotropical species with the same name in the same genus. The etymology of each new name is given in the catalogue. The country of the type locality of each preoccupied species name is given at the end of each entry.

Billaea rufescens O'Hara & Wood is proposed as a *nomen novum* for *Sarcoprosena rufiventris* Townsend, 1929, a name preoccupied in the genus *Billaea* Robineau-Desvoidy, 1830 by *Paratheresia rufiventris* Townsend, 1929 [Peru]. **Nom. nov.**

Billaea triquetrus O'Hara & Wood is proposed as a *nomen novum* for *Sarcoprosena triangulifera* Townsend, 1927, a name preoccupied in the genus *Billaea* Robineau-Desvoidy, 1830 by *Dexia triangulifera* Zetterstedt, 1844 [Peru]. **Nom. nov.**

Eucelatoria nudioculata O'Hara & Wood is proposed as a *nomen novum* for *Eucelatorioidea nigripalpis* Thompson, 1968, a name preoccupied in the genus *Eucelatoria* Townsend, 1909 by *Chetolyga nigripalpis* Bigot, 1889 [Trinidad]. **Nom. nov.**

Eucelatoria oblonga O'Hara & Wood is proposed as a *nomen novum* for *Urodexodes elongatum* Cortés & Campos, 1974, a name preoccupied in the genus *Eucelatoria* Townsend, 1909 by *Exorista elongata* van der Wulp, 1890 [Chile]. **Nom. nov.**

Lespesia thompsoni O'Hara & Wood is proposed as a *nomen novum* for *Sturmiopsoidea obscura* Thompson, 1966, a name preoccupied in the genus *Lespesia* Robineau-Desvoidy, 1863 by *Eurigaster obscurus* Bigot, 1857 [Cuba]. **Nom. nov.**

Myiopharus charapensis O'Hara & Wood is proposed as a *nomen novum* for *Metarrhinomyia angusta* Townsend, 1927, a name preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Mayophorinia angusta* Townsend, 1927 [Peru]. **Nom. nov.**

Myiopharus incognitus O'Hara & Wood is proposed as a *nomen novum* for *Stenochaeta claripalpis* Thompson, 1968, a name preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Neoxynopsoidea claripalpis* Thompson, 1968 [Trinidad]. **Nom. nov.**

Myiopharus rufopalpus O'Hara & Wood is proposed as a *nomen novum* for *Paralispe palpalis* Townsend, 1929, a name preoccupied in the genus *Myiopharus* Brauer & Bergenstamm, 1889 by *Myioxynops palpalis* Townsend, 1927 [Peru]. **Nom. nov.**

New type species fixations

Article 70.3.2 of the *Code* (ICZN 1999) allows the type species of a nominal genus to be fixed as the species intended by the original author if the type species designated by that author was misidentified. We have invoked Article 70.3.2 for the three instances of misidentified type species in this catalogue that had not been dealt with previously (e.g., O'Hara and Wood 2004) to preserve the current concepts of the genera involved. Type species are fixed for the following nominal genera (see catalogue for further details).

Parafabricia Brauer & Bergenstamm, 1894: 612 [also 1895: 76]. Type species newly fixed as *Parafabricia perplexa* Townsend, 1931. Synonym of *Archytas* Jaennicke, 1867.

Tachinodes Brauer & Bergenstamm, 1889: 133 [also 1889: 65]. Type species newly fixed as *Jurinia metallica* Robineau-Desvoidy, 1830. Synonym of *Archytas* Jaennicke, 1867.

Willistonina Brauer & Bergenstamm, 1889: 97 [also 1890: 29]. Type species newly fixed as *Willistonina aldrichi* Townsend, 1931. Synonym of *Belvosia* Robineau-Desvoidy, 1830.

Lectotype designations

Lectotypes are designated for four nominal species (see Lectotype Designations section).

Echinomyia pygmaea Macquart, 1851. This is a valid name in the genus *Peleteria* Robineau-Desvoidy, 1830, as *Peleteria pygmaea* (Macquart).

Gonia chilensis Macquart, 1844. This is a junior synonym in the genus *Gonia* Meigen, 1803. The valid name of the species is *Gonia pallens* Wiedemann, 1830.

Masicera auriceps Macquart, 1844. This is a valid name in the genus *Lespesia* Robineau-Desvoidy, 1863, as *Lespesia auriceps* (Macquart).

Proso-pochoeta nitidiventris Macquart, 1851. This is a valid name in the genus *Proso-pochoeta* Macquart, 1851.

New and revived combinations

New and revived combinations proposed in this work are listed below. These are based on the study of type material, authoritatively identified specimens, and/or descriptions and figures in the literature, mostly by DMW.

Blepharipeza andina Bigot, 1888 is moved from an unplaced species in “Sturmiini” or Tachinidae to *Lespesia* Robineau-Desvoidy, 1863 as a *nomen dubium*. Distribution: Cuba (not Chile as published). **Comb. nov.**

Camposodes evanescens Cortés, 1967 is moved from its original placement in *Camposodes* Cortés, 1967 to *Phytomyptera* Rondani, 1845. Distribution: Argentina, Chile. **Comb. nov.**

Ectophasiopsis ypiranga Dios & Nihei, 2017 is moved from its original placement in *Ectophasiopsis* Townsend, 1915 to *Trichopoda* Berthold, 1827 (and assigned to subgenus *Galactomyia* Townsend, 1908). Distribution: Argentina, Brazil. **Comb. nov.**

Embiomyia australis Aldrich, 1934 (type species of *Embiomyia* Aldrich, 1934) is moved from its original placement in *Embiomyia* to *Steleoneura* Stein, 1924 (with *Embiomyia* in synonymy). Distribution: Argentina, Chile. **Comb. nov.**

Eurigaster modestus Bigot, 1857 is moved from its position in unplaced species of Exoristinae (as “Goniinae”) by Guimarães (1971: 215) to *Lespesia* Robineau-Desvoidy, 1863. Distribution: Cuba. **Comb. nov.**

Eurigaster obscurus Bigot, 1857 is moved from its position in unplaced species of Exoristinae (as “Goniinae”) by Guimarães (1971: 215) to *Lespesia* Robineau-Desvoidy, 1863. Distribution: Cuba. **Comb. nov.**

Macropatelloa tanumeana Townsend, 1931 (type species of *Macropatelloa* Townsend, 1931) is moved from its original placement in *Macropatelloa* to *Patelloa* Townsend, 1916 (with *Macropatelloa* in synonymy). Distribution: Argentina, Chile. **Comb. nov.**

Masicera insignis van der Wulp, 1882 is moved from its placement in *Sturmia* Robineau-Desvoidy, 1830 by previous authors (e.g., Cortés and Hichins 1969: 59; Guimarães 1971: 192; Henry 1987: 200) to *Drino* Robineau-Desvoidy, 1863. Distribution: Argentina, Chile. **Comb. nov.**

Parasetigena hichinsi Cortés, 1967 is moved from its original placement in *Parasetigena* Brauer & Bergenstamm, 1891 to *Chetogena* Rondani, 1856. Distribution: Chile. **Comb. nov.**

Parasetigena porteri Brèthes, 1920 is moved from its placement in *Stomatotachina* Townsend, 1931 by previous authors (e.g., Guimarães 1971: 160; Mulieri et al. 2013: 169) to *Chetogena* Rondani, 1856. Distribution: Chile. **Comb. nov.**

Phorocera calyptrata Aldrich, 1934 is moved from uncertain placements by Guimarães (1971: 152, unplaced in Blondeliini), Henry (1987: 206, in *Phorocera* Robineau-Desvoidy, 1830 but genus unplaced in Tachinidae) and González (1992b: 183, in *Phorocera* but genus unplaced in Exoristinae [as “Goniinae”]) to *Admontia* Brauer & Bergenstamm, 1889. Distribution: Argentina, Chile. **Comb. nov.**

Poliops auratus Campos, 1953 is moved from its original placement in *Poliops* Aldrich, 1934 to *Admontia* Brauer & Bergenstamm, 1889. Distribution: Chile. **Comb. nov.**

Poliops striatus Aldrich, 1934 is moved from its original placement in *Poliops* Aldrich, 1934 to *Admontia* Brauer & Bergenstamm, 1889. Distribution: Argentina, Chile. **Comb. nov.**

Ruiziella frontosa Cortés, 1951 is moved from its original placement in *Ruiziella* Cortés, 1951 to *Chaetoepalpus* Vimmer & Soukup, 1940, where it is placed in synonymy with *C. coquilleti* Vimmer & Soukup, 1940, **syn. nov.** Distribution of *C. coquilleti*: Argentina, Chile, Peru. **Comb. nov.**

Ruiziella luctuosa Cortés, 1951 is moved from its original placement in *Ruiziella* Cortés, 1951 to *Chaetoepalpus* Vimmer & Soukup, 1940. Distribution: Argentina, Chile. **Comb. nov.**

Sarcoprosena luteola Cortés & Campos, 1974 is moved from its original placement in *Sarcoprosena* Townsend, 1927 to *Billaea* Robineau-Desvoidy, 1830. Distribution: Chile. **Comb. nov.**

Sarcoprosena rufiventris Townsend, 1929 is moved from its original placement in *Sarcoprosena* Townsend, 1927 to *Billaea* Robineau-Desvoidy, 1830. **Comb. nov.** The name *S. rufiventris* is a junior secondary homonym of *Paratheresia rufiventris* Townsend, 1929 when placed in *Billaea* and is renamed herein as *Billaea rufescens* O'Hara & Wood, **nom. nov.** Distribution: Peru.

Sarcoprosena triangulifera Townsend, 1927 (type species of *Sarcoprosena* Townsend, 1927) is moved from its original placement in *Sarcoprosena* to *Billaea* Robineau-Desvoidy, 1830 (with *Sarcoprosena* in synonymy). **Comb. nov.** The name *S. triangulifera* is a junior secondary homonym of *Dexia triangulifera* Zetterstedt, 1844 when placed in *Billaea* and is renamed herein as *Billaea triquetrus* O'Hara & Wood, **nom. nov.** Distribution: Peru.

Saundersia aurea Giglio-Tos, 1893 is moved from its placement in *Epalpus* Rondani, 1850 by Guimarães (1971: 64) to “Unplaced species of Tachinini”. Distribution: Mexico. **Comb. nov.**

Schistostephana aurifrons Townsend, 1919 (type species of *Schistostephana* Townsend, 1919) is moved from its original placement in *Schistostephana* to *Billaea* Robineau-Desvoidy, 1830 (with *Schistostephana* in synonymy). Distribution: Peru. **Comb. nov.**

Siphoactia charapensis Townsend, 1927 (type species of *Siphoactia* Townsend, 1927) is moved from its original placement in *Siphoactia* to *Clausicella* Rondani, 1856 (with *Siphoactia* in synonymy). Distribution: Peru. **Comb. nov.**

Siphoactia peregrina Cortés & Campos, 1971 is moved from its original placement in *Siphoactia* Townsend, 1927 to *Clausicella* Rondani, 1856. Distribution: Chile. **Comb. nov.**

Stomatotachina splendida Townsend, 1931 (type species of *Stomatotachina* Townsend, 1931) is moved from its original placement in *Stomatotachina* to *Chetogena* Rondani, 1856 (with *Stomatotachina* in synonymy). *Stomatotachina splendida* continues to be treated as a junior subjective synonym of *Parasetigena porteri* Brèthes, 1920 (see above). **Comb. nov.**

Sturmia festiva Cortés, 1944 is moved from its original placement in *Sturmia* Robineau-Desvoidy, 1830 to *Drino* Robineau-Desvoidy, 1863. Distribution: Argentina, Chile. **Comb. nov.**

Sturmiopsoidea obscura Thompson, 1966 (type species of *Sturmiopsoidea* Thompson, 1966) is moved from its original placement in *Sturmiopsoidea* to *Lespesia* Robineau-Desvoidy, 1863 (with *Sturmiopsoidea* in synonymy). **Comb. nov.** The name *S. obscura* is a junior secondary homonym of *Eurigaster obscurus* Bigot, 1857 when placed in *Lespesia* and is renamed herein as *Lespesia thompsoni* O'Hara & Wood, **nom. nov.** Distribution: Trinidad.

Trichopoda arcuata Bigot, 1876 is returned to *Trichopoda* Berthold, 1827 (and assigned to subgenus *Galactomyia* Townsend, 1908) from its placement in *Ectophasiopsis* Townsend, 1915 by previous authors (e.g., Aldrich 1934: 12; Guimarães 1971: 12; Dios and Nihei 2017: 5). Distribution: Argentina, Chile. **Comb. revived.**

Trichopoda gradata Wiedemann, 1830 is returned to *Trichopoda* Berthold, 1827 (and assigned to subgenus *Galactomyia* Townsend, 1908) from its placement in *Ectophasiopsis* Townsend, 1915 by Dios and Nihei (2017: 10). Distribution: Argentina, Brazil, Uruguay. **Comb. revived.**

New and revived synonymies

New and revived generic and specific synonymies are proposed for the names below. As with the new and revived combinations listed above, they result from the study of type material, authoritatively identified specimens, and/or descriptions and figures in the literature, mostly by DMW.

Camposodes Cortés, 1967, formerly treated as a genus (e.g., Guimarães 1971: 166; Cortés 1984: 378), is synonymised with *Phytomyptera* Rondani, 1845. **Syn. nov.**

Ectophasiopsis Townsend, 1915, formerly treated as a genus (e.g., Aldrich 1934: 11; Guimarães 1971: 12; Dios and Nihei 2017: 4), is synonymised with *Trichopoda* Berthold, 1827, subgenus *Galactomyia* Townsend, 1908. **Syn. nov.**

Embiomyia Aldrich, 1934, formerly treated as a genus (e.g., Cortés and Hichins 1969: 32; Guimarães 1971: 85), is synonymised with *Steleoneura* Stein, 1924. **Syn. nov.**

Fabricia andicola Bigot, 1888, treated as a junior synonym of *Peleteria filipalpis* (Rondani, 1863) by Guimarães (1971: 44), is returned to synonymy with *Peleteria robusta* (Wiedemann, 1830) as proposed earlier by Guimarães (1962: 484). **Syn. revived.**

Macropatelloa Townsend, 1931, formerly treated as a genus (e.g., Cortés 1986: 144, 158), González (1992b: 179), is synonymised with *Patelloa* Townsend, 1916. **Syn. nov.**

- Peleteria inca* Curran, 1925, treated as a junior synonym of *Peleteria filipalpis* (Rondani, 1863) by Guimarães (1971: 44), is returned to synonymy with *Peleteria robusta* (Wiedemann, 1830) as proposed earlier by Guimarães (1962: 484). **Syn. revived.**
- Poliops* Aldrich, 1934, formerly treated as a genus (e.g., Guimarães 1971: 169; Cortés 1979: 81), is synonymised with *Admontia* Brauer & Bergenstamm, 1889. **Syn. nov.**
- Ruiziella* Cortés, 1951, formerly treated as a genus (e.g., Guimarães 1971: 45; Cortés and Campos 1974: 116), is synonymised with *Chaetoepalpus* Vimmer & Soukup, 1940. **Syn. nov.**
- Ruiziella frontosa* Cortés, 1951, formerly treated as a valid species of *Ruiziella* (e.g., Guimarães 1971: 45), is synonymised with *Chaetoepalpus coquilleti* Vimmer & Soukup, 1940 in the genus *Chaetoepalpus* Vimmer & Soukup, 1940. **Comb. nov., syn. nov.**
- Sarcoprosena* Townsend, 1927, formerly treated as a genus (e.g., Guimarães 1971: 38; Cortés 1984: 381), is synonymised with *Billaea* Robineau-Desvoidy, 1830. **Syn. nov.**
- Schistostephana* Townsend, 1919, formerly treated as a genus (e.g., Guimarães 1971: 38), is synonymised with *Billaea* Robineau-Desvoidy, 1830. **Syn. nov.**
- Siphoactia* Townsend, 1927, formerly treated as a genus (e.g., Guimarães 1971: 170; Cortés 1984: 381), is synonymised with *Clausicella* Rondani, 1856. **Syn. nov.**
- Stomatotachina* Townsend, 1931, formerly treated as a genus (e.g., Guimarães 1971: 160; Mulieri et al. 2013: 169; Nihei 2015: 1), is synonymised with *Chetogena* Rondani, 1856. **Syn. nov.**
- Sturmiopsoidea* Thompson, 1966, formerly treated as a genus (e.g., Guimarães 1971: 192), is synonymised with *Lespesia* Robineau-Desvoidy, 1863. **Syn. nov.**

Catalogue

Subfamily DEXIINAE

Tribe DEXIINI

Genus *BILLAEA* Robineau-Desvoidy, 1830

- THERESIA* Robineau-Desvoidy, 1830: 325. Type species: *Theresia tandrec* Robineau-Desvoidy, 1830 (= *Musca rutilans* Fabricius, 1781), by monotypy [United States].
- BILLAEA*** Robineau-Desvoidy, 1830: 328. Type species: *Billaea grisea* Robineau-Desvoidy, 1830 (= *Dexia pectinata* Meigen, 1826), by monotypy [France].
- EUTHERESIA* Townsend, 1911: 149. *Nomen nudum* (named for “Coquillett’s *Theresia analis*”, itself a *nomen nudum*).
- EUTHERESIA* Townsend, 1912a: 117. Type species: *Eutheresia monohammi* Townsend, 1912, by monotypy [United States].
- PARATHERESIA* Townsend, 1915c: 65. Type species: *Paratheresia signifera* Townsend, 1915 (= *Sarcophaga claripalpis* van der Wulp, 1895), by original designation [Peru].
- SCHISTOSTEPHANA* Townsend, 1919b: 551. Type species: *Schistostephana aurifrons* Townsend, 1919, by original designation [Peru]. **Syn. nov.**

SARCOPROSENA Townsend, 1927a: 228. Type species: *Sarcoprosena triangulifera* Townsend, 1927 (junior secondary homonym of *Dexia triangulifera* Zetterstedt, 1844; = *Billaea triquetrus* O'Hara & Wood, **nom. nov.**, see below), by original designation [Peru]. **Syn. nov.**

BATHYTHERESIA Townsend, 1928a: 146. Type species: *Bathytheresia bassleri* Townsend, 1928 (= *Sarcophaga claripalpis* van der Wulp, 1895), by original designation [Peru].

PARABILLAEA Blanchard, 1937: 44. Type species: *Parabillaea rhynchophorae* Blanchard, 1937, by original designation [Argentina].

PARABILAEA. Incorrect subsequent spelling of *Parabillaea* Blanchard, 1937 (Guimarães 1977b: 269).

Notes: The concept of *Billaea* Robineau-Desvoidy adopted here is similar to that of *Theresia* Robineau-Desvoidy *sensu* Aldrich (1934: 106) for the Patagonian fauna and *Billaea sensu* Wood (1987: 1248) for the Nearctic fauna, but is expanded geographically to include variation within the lineage throughout the Neotropics. The restricted generic concepts of Townsend (1936b: 142), and to a lesser degree Guimarães (1971: 36, 1977b), were based on what we believe to be morphological differences within a larger clade that constitutes our concept of *Billaea*. This genus is characterised in part by a plumose arista, bare eye and parafacial, face with not more than a small carina dividing antennae, bare prosternum, haired propisternum, and often three black vittae on the scutum resembling the common pattern seen in *Sarcophaga* Meigen, 1826 (Sarcophagidae). The species formerly assigned to *Sarcoprosena* differ from the other species in having a narrower parafacial and gena.

The relative priority of *Billaea* Robineau-Desvoidy, 1830 and *Theresia* Robineau-Desvoidy, 1830, when the two are treated as synonyms, was established by Wood (1987: 1248), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999).

References: Coquillett (1910: 614), type species of *Theresia*; Aldrich (1934: 5, 106), in key to Patagonian genera (as *Theresia*), synonymy of *Eutheresia*, *Paratheresia* and *Schistostephana* with *Theresia*, taxonomic notes; Townsend (1936b: 142), diagnosis of adults and immatures of Theresiini and key to genera (including *Bathytheresia*, *Billaea*, *Eutheresia*, *Paratheresia*, *Sarcoprosena*, *Schistostephana* and *Theresia*); Townsend (1938: 391, 392, 397, 402, 403, 404, 405), redescrptions of *Bathytheresia*, *Billaea*, *Eutheresia*, *Paratheresia*, *Sarcoprosena*, *Schistostephana* and *Theresia*; van Emden (1949: 507), synonymy of *Bathytheresia* and *Parabillaea* with *Paratheresia*; Guimarães (1971: 37, 38), *Paratheresia*, *Schistostephana*, *Sarcoprosena* and *Theresia* recognised as valid genera; Cortés and Campos (1974: 115), *Sarcoprosena* in key to tachinid genera of Tarapacá and Antofagasta regions; Guimarães (1977b), revision of *Paratheresia*; Cortés (1984: 381), *Sarcoprosena* in key to tachinid genera of Tarapacá and Antofagasta regions; Wood (1987: 1248), synonymy of *Eutheresia*, *Paratheresia* and *Theresia* with *Billaea*; O'Hara and Wood (1998: 756, 759), review of synonymy of Wood (1987).

aurifrons (Townsend, 1919).—Not Chile [Peru]. **Comb. nov.**

Schistostephana aurifrons Townsend, 1919b: 552. Holotype male (USNM, examined by DMW, Fig. 3a). Type locality: Peru, Cajamarca, Río Charapi [as “Rio Charape”, ca. 5°25'S, 78°59'W], 4500 ft.

References: Aldrich (1934: 107), as species of *Theresia*; Townsend (1936b: 147, 1938: 404), as species of *Schistostephana*; Guimarães (1971: 38), as species of *Schistostephana*.

erecta (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Theresia erecta Aldrich, 1934: 107. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Peulla.

References: Guimarães (1971: 37), as sole species of *Theresia* in America south of United States; Gramajo (1998: 93), first record from Argentina.

luteola (Cortés & Campos, 1974).—Neotropical: South America (Chile). **Comb. nov.**

Sarcoprosena luteola Cortés & Campos, 1974: 122. Holotype female (MEUC). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, km 41/42, Mollepampa [ca. 18°24'S, 70°2'W].

rufescens O'Hara & Wood, **nom. nov.**—Not Chile [Peru].

Sarcoprosena rufiventris Townsend, 1929: 367 (junior secondary homonym of *Paratheresia rufiventris* Townsend, 1929, by First Reviser action below). Syntypes, 1 male and 1 female (USNM, examined by DMW, Fig. 3b). Type locality: Peru, Río Ushpayacu, 1300 ft. **Comb. nov.**

Billaea rufescens O'Hara & Wood, **nom. nov.** for *Sarcoprosena rufiventris* Townsend, 1929.

Note: *Paratheresia rufiventris* Townsend, 1929 and *Sarcoprosena rufiventris* Townsend, 1929, both from Peru, were described in the same publication on the same page (Townsend 1929: 367) and are secondary homonyms when placed together in *Billaea*. As the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999), we hereby fix *Paratheresia rufiventris* as the senior homonym. We propose the new name *Billaea rufescens* to replace the name of the junior homonym *Sarcoprosena rufiventris*. The same type material applies to the new name. The specific epithet *rufescens* is formed in part from *rufus*, Latin for red, alluding to the underlying reddish tinge of the abdomen that likely suggested the original name *rufiventris*.

Reference: Guimarães (1971: 37), as *Sarcoprosena rufiventris*.

triquetrus O'Hara & Wood, **nom. nov.**—Not Chile [Peru].

Sarcoprosena triangulifera Townsend, 1927a: 356 (junior secondary homonym of *Dexia triangulifera* Zetterstedt, 1844). Holotype male (USNM, examined by DMW, Fig. 3c, d). Type locality: Peru, Arequipa, Yahuar Mayo. **Comb. nov.**

Billaea triquetrus O'Hara & Wood, **nom. nov.** for *Sarcoprosena triangulifera* Townsend, 1927.

Note: *Sarcoprosena triangulifera* Townsend, 1927, when moved to *Billaea*, is a junior secondary homonym of *Dexia triangulifera* Zetterstedt, 1844, the valid name of a *Billaea* species in the Palaearctic Region (O'Hara et al. 2009: 28). We hereby propose the new name *Billaea triquetrus* to replace the name of the junior homonym *Sarcoprosena triangulifera*. The same type material applies to the new name. The specific epithet *triquetrus* is Latin for three-cornered or triangular, referring to the triangular markings on the abdomen and particularly tergite 3, which presumably inspired Townsend's name *triangulifera*.

Reference: Guimarães (1971: 37), as *Sarcoprosena triangulifera*.

Genus *CALLOTROXIS* Aldrich, 1929

CALLOTROXIS Aldrich, 1929b: 7. Type species: *Callotroxis edwardsi* Aldrich, 1929, by original designation [Chile].

References: Aldrich (1934: 4, 80), in key to Patagonian genera, taxonomic notes; Townsend (1936b: 131), diagnosis of adults and immatures of Prosenini and key to genera (including *Callotroxis*); Townsend (1938: 321), redescription.

edwardsi Aldrich, 1929.—Neotropical: South America (Chile).

Callotroxis edwardsi Aldrich, 1929b: 8. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Reference: Aldrich (1934: 81), redescription, first description of female.

Genus *DASYUROMYIA* Bigot, 1885

DASYUROMYIA Bigot, 1885a: 237. *Nomen nudum*.

DASYUROMYIA Bigot, 1885c: liv [also 1885c: liv, *Bull. Soc. Ent. France*]. Type species: *Dasyuromyia penicillata* Bigot, 1885 (= *Tachina inornata* Walker, 1836), by monotypy [Chile].

SELENOMYIA Brauer & Bergenstamm, 1891: 361 [also 1891: 57]. Type species: *Selenomyia brevicornis* Brauer & Bergenstamm, 1891 (= *Hyadesimyia sarcophagidea* Bigot, 1888), by monotypy [Chile].

MESEMBRIOPHYTO Townsend, 1916c: 301. Type species: *Mesembriophyto magellana* Townsend, 1916 (= *Tachina inornata* Walker, 1836), by original designation [Chile].

References: Aldrich (1934: 6, 156), in key to Patagonian genera, synonymy of *Mesembriophyto* and *Selenomyia* with *Dasyuromyia*, redescription, key to six Patagonian species; Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Dasyuromyia*); Townsend (1939c: 32), redescription of *Dasyuromyia*; Dugdale (1969: 624), figure of terminal segments of first instar larva of *Dasyuromyia* sp.; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; Gramajo (2011: 175), key to species of Patagonian Argentina including all species listed below except for *Dasyuromyia nervosa* (known only from Chile).

aperta Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Dasyuromyia aperta Aldrich, 1934: 161. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Reference: Cortés and Hichins (1969: 29), first record from Chile.

inornata (Walker, 1836).—Neotropical: South America (Argentina, Chile).

Tachina inornata Walker, 1836: 349. Lectotype male (NHMUK), by fixation of Austen (1907: 329) (examination of male “type” from Cape Gregory in NHMUK is regarded as a lectotype fixation). Type locality: Chile, Magallanes

y de la Antártica Chilena, Magallanes, Cabo San Gregorio [as “Cape Gregory”, ca. 52°39'S, 70°13'W].

Dasyuromyia penicillata Bigot, 1885c: lv [also 1885c: lv, *Bull. Soc. Ent. France*]. Lectotype male (NHMUK), by fixation of Townsend (1931a: 93) (examination of “male Ht” from Chile in NHMUK [as “Newmarket”] is regarded as a lectotype fixation). Type locality: Chile.

Mesembriophyto magellana Townsend, 1916e: 301. Holotype female (USNM). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Punta Arenas [as “Sandy Point”].

References: Aldrich (1934: 157), synonymy, redescription, figures of male abdomen, first record from Argentina (as “Southern Patagonia”, which is interpreted here as Argentina based on the travels of the collector of the specimen, paleontologist Barnum Brown); Cortés (1963: 244), notes on name-bearing type of *Tachina inornata* in NHMUK.

nervosa (Walker, 1836).—Neotropical: South America (Chile).

Tachina nervosa Walker, 1836: 349. Lectotype male (NHMUK), by fixation of Austen (1907: 329) (examination of male “type” from Port Famine in NHMUK is regarded as a lectotype fixation). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Puerto del Hambre [as “Port Famine”].

Reference: Cortés (1963: 245), notes on name-bearing type in NHMUK.

nigriceps Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Dasyuromyia nigriceps Aldrich, 1934: 162. Holotype female (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangué.

References: Cortés (1963: 244), notes on holotype in NHMUK; Gramajo (2011: 174), first record from Argentina.

sarcophagidea (Bigot, 1888).—Neotropical: South America (Argentina, Chile).

Hyadesimyia sarcophagidea Bigot, 1888a: 28. Syntypes, 6 males and females (MNHN, see note). Type locality: Chile, Magallanes y de la Antártica Chilena, Antártica Chilena, Isla Hoste, Bahía Orange area [ca. 55°31'S, 68°6'W].

Selenomyia brevicornis Brauer & Bergenstamm, 1891: 361 [also 1891: 57] (as “*S. brevicornis* Phil.”). Lectotype male (NHMW), by fixation of Townsend (1939c: 32) (mention of “Male Ht” from Chile in NHMW is regarded as a lectotype fixation). Type locality: Chile.

Notes: Bigot (1888a: 28) described *Hyadesimyia sarcophagidea* from six specimens of both sexes from the Bahía Orange area of Magallanes, Chile. The MNHN database records ten possible syntypes in the Macquart collection with numbers MNHN-ED-ED10217 to MNHN-ED-ED10226.

Brauer and Bergenstamm (1891: 361) described *Selenomyia brevicornis* from one or more males collected from Chile by Philippi. Aldrich (1925: 459) partially redescribed one male borrowed from NHMW and labelled “Philippi, Chili, 1870”. This specimen is undoubtedly a name-bearing type of *S. brevicornis* and is assumed to be the “Ht male” of Townsend (1939c: 32).

Reference: Aldrich (1934: 159), synonymy, redescription, first record from Argentina.

sternalis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Dasyuromyia sternalis Aldrich, 1934: 160. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Reference: Cortés (1979: 79), first record from Argentina.

tarsalis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Dasyuromyia tarsalis Aldrich, 1934: 160. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Gutiérrez.

References: Cortés (1963: 245), notes on holotype in NHMUK; Cortés (1986: 146), first record from Chile.

Genus *HYADESIMYIA* Bigot, 1888

HYADESIMYIA Bigot, 1888a: 26. Type species: *Hyadesimyia clausa* Bigot, 1888, by subsequent designation of Bigot (1891: cxxxvi) [Chile].

References: Coquillett (1910: 553), type species (given as “*Hyadesimyia clausa* Bigot, the first species, by present designation”); Aldrich (1934: 4, 84), in key to Patagonian genera, taxonomic notes; Townsend (1936b: 112), diagnosis of adults and immatures of Aulacephalini and key to genera (including *Hyadesimyia*); Townsend (1938: 259), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

clausa Bigot, 1888.—Neotropical: South America (Argentina, Chile).

Hyadesimyia clausa Bigot, 1888a: 27. Holotype male (MNHN, number MNHN-ED-ED10216). Type locality: Chile, Magallanes y de la Antártica Chilena, Antártica Chilena, Isla Hoste, Bahía Orange area [ca. 55°31'S, 68°6'W].

References: Aldrich (1934: 85), redescription, first record from Argentina; Cortés (1973a: 101), partial redescription, head and wing figures.

Genus *HYOSOMA* Aldrich, 1934

HYOSOMA Aldrich, 1934: 139. Type species: *Hyosoma limbisquama* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Hyosoma*); Townsend (1939c: 41), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

limbisquama Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Hyosoma limbisquama Aldrich, 1934: 140. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Nahuel Huapí, Puerto Blest.

Note: *Hyosoma limbisquama* was recorded from both Argentina and Chile in the original description.

Reference: Cortés (1963: 244), notes on type series in NHMUK.

Genus *MORPHODEXIA* Townsend, 1931

MORPHODEXIA Townsend, 1931c: 342. Type species: *Morphodexia microphthalmoidea* Townsend, 1931 (= *Camarona barrosi* Brèthes, 1920), by original designation [Chile].

References: Aldrich (1934: 6, 146), in key to Patagonian genera, redescription, key to five Patagonian species; Townsend (1936b: 116), diagnosis of adults and immatures of Dexillini and key to genera (including *Morphodexia*); Townsend (1938: 288), redescription; Cortés (1986: 142), in key to tachinid genera of Aysén and Magallanes regions.

barrosi (Brèthes, 1920).—Neotropical: South America (Argentina, Chile).

Camarona barrosi Brèthes, 1920a: 42. Lectotype male (MACN), by fixation of Cortés (1963: 250–251) (examination of male “tipo” from Río Blanco in MACN is regarded as a lectotype fixation). Type locality: Chile, Los Andes, Río Blanco.

Morphodexia microphthalmoides Townsend, 1931c: 343. Holotype female (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

References: Aldrich (1934: 147), redescription, head figure (as *Morphodexia microphthalmoides*); Verbeke (1962: 97), description of male terminalia; Cortés (1963: 250), notes and partial redescription of name-bearing type of *Camarona barrosi* in MACN; Cortés (1967b: 13), synonymy of *Morphodexia microphthalmoides* with *Camarona barrosi*; Cortés (1979: 80), first record from Argentina; Mulieri et al. (2013: 160), notes on name-bearing type (as syntype) of *C. barrosi* in MACN.

clausa Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Morphodexia clausa Aldrich, 1934: 149. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Nahuel Huapí, Puerto Blest.

Reference: Cortés and Hichins (1969: 43), first record from Chile.

facialis (Aldrich, 1928).—Neotropical: South America (Argentina, Chile).

Selenomyia facialis Aldrich, 1928b: 23. Holotype female (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

References: Aldrich (1934: 150), redescription, first description of male; Gramajo (1998: 93), first record from Argentina.

nigra Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Morphodexia nigra Aldrich, 1934: 149. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Chiloé, Ancud.

Reference: Cortés (1979: 80), first record from Argentina.

palpalis Aldrich, 1934.—Neotropical: South America (Chile).

Morphodexia palpalis Aldrich, 1934: 150. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

subaenea Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Morphodexia nigra subaenea Aldrich, 1934: 149. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Gutiérrez.

References: Cortés and Hichins (1969: 44), first record from Chile (as *Morphodexia nigra* var. *subaenea*); Guimarães (1971: 24), raised from subspecies to *Morphodexia subaenea*.

Genus *MYIODEXIA* Cortés & Campos, 1971

MYIODEXIA Cortés & Campos, 1971: 36. Type species: *Myiodexia deserticola* Cortés & Campos, 1971, by original designation [Chile].

References: Cortés and Campos (1971: 21, 1974: 112) and Cortés (1984: 378), in keys to tachinid genera of Tarapacá and Antofagasta regions.

deserticola Cortés & Campos, 1971.—Neotropical: South America (Chile).

Myiodexia deserticola Cortés & Campos, 1971: 38. Holotype male (EEAM). Type locality: Chile, Tarapacá, Tamarugal, 15 km south of Pozo Almonte, Junoy, 1200 m (20°18'S, 69°48'W) (coordinates and elevation given on p. 11).

Genus *NOTODYTES* Aldrich, 1934

NOTODYTES Aldrich, 1934: 163. Type species: *Notodytes variabilis* Aldrich, 1934, by original designation [Argentina].

References: Aldrich (1934: 163), key to species; Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Notodytes*); Townsend (1939c: 51), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

aurea Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Notodytes aurea Aldrich, 1934: 165. Holotype female (USNM). Type locality: Chile, Los Lagos, Llanquihue, Ensenada.

References: Cortés (1973a: 100), taxonomic notes; Cortés (1979: 80), first record from Argentina.

major Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Notodytes major Aldrich, 1934: 165. Syntypes, 14 females (NHMUK, USNM). Type locality: Chile, Araucanía, Malleco, Angol.

References: Cortés (1963: 245), notes on two syntypes in NHMUK; Cortés (1979: 80), first record from Argentina.

variabilis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Notodytes variabilis Aldrich, 1934: 164. Holotype male (NHMUK). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

Note: *Notodytes variabilis* was recorded from both Argentina and Chile in the original description. References: Cortés (1963: 245), notes on type series in NHMUK; Dugdale (1969: 624), figure of first instar larva; Cortés (1973a: 100), taxonomic notes.

Genus *OLIGOOESTRUS* Townsend, 1932

OLIGOOESTRUS Townsend, 1932c: 1. Type species: *Oligooestrus oestroides* Townsend, 1932, by original designation [Argentina].

OLIGOESTRUS. Incorrect subsequent spelling of *Oligooestrus* Townsend, 1932 (Guimarães 1971: 216, 302).

Note: *Oligooestrus oestroides*, the single known species of *Oligooestrus*, is an unusual-looking tachinid with a round yellow head in frontal view, thorax and abdomen dark, and body length of ca. 5 mm. It has a suite of distinctive features (see descriptions and figures in Townsend 1932: 2 and Aldrich 1934: 6) including tiny antenna, arista micropubescent, vibrissae closely approximated and very high (closer to antenna than to oral cavity), vein M_1 petiolate and ending in wing margin far from wing tip, and one katepisternal seta. Townsend (1932: 1) regarded the species as “the most important oestromuscoid discovery of the twentieth century from the taxonomic point of view”, a form he believed supported his view that “the oestriform tachinids or Tachino-Oestridae of Villeneuve (Aulacephalini, Ormiini, Trixodini, Trixini, Palpostomatini, Paratrixini, Glaurocarini, and Myiotrixini) all belong in the same family with the Oestrini”. Townsend assigned *Oligooestrus* to a narrowly interpreted Oestrini within the Oestridae (see also Townsend 1936b: 108). *Oligooestrus* was later listed as an unplaced genus of Tachinidae in Guimarães (1971: 216). It was recorded from Chile for the first time in Stireman et al. (2016: 30) as “Dexiinae ... ?Dufouriini: *Oligooestrus* ?*oestroides* Townsend”. The identity of that specimen as *O. oestroides* has since been confirmed and a second specimen of *O. oestroides* collected on the same trip and identified later bears the following data: Chile, Araucanía Region, road R-955, south of Punta Negra, 1090 m, 38°34.96'S, 71°26.35'W, 15.xii.2015, J.E. O'Hara (CNC487480). The recent molecular phylogeny of Tachinidae places *Oligooestrus* in the tribe Dexiini of the Dexiinae (Stireman et al. 2019: 9 [fig. 4], 30) and we follow this placement.

References: Aldrich (1934: 2, 6), in key to Patagonian genera, taxonomic notes; Townsend (1936b: 108), diagnosis of adults and immatures of Oestrini and key to genera (including *Oligooestrus*); Townsend (1938: 252), redescription; Guimarães (1971: 216), listed as an unplaced genus of Tachinidae.

oestroides Townsend, 1932.—Neotropical: South America (Argentina, Chile). **New record from Chile.** (Fig. 4a)

Oligooestrus oestroides Townsend, 1932c: 4. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Nahuel Huapí, San Carlos de Bariloche [as “Bariloche”].

Note: *Oligooestrus oestroides* was tentatively recorded from Chile (Parque Nacional Nahuelbuta, Araucanía) in a trip report by Stireman et al. (2016: 30). It is here confirmed from Chile based on a male specimen in CNC collected by JEOH during the same trip but identified later. The specimen bears the following data: Araucanía Region, road R-955, south of Punta Negra, 1090 m, 38°34.96'S, 71°26.35'W, 15.xii.2015, J.E. O'Hara (CNC487480).

Reference: Aldrich (1934: 7), redescription, taxonomic notes, head and wing figures.

Genus *PELYCOPS* Aldrich, 1934

PELYCOPS Aldrich, 1934: 168. Type species: *Pelycops darwini* Aldrich, 1934, by original designation [Chile].

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Pelycops*); Townsend (1939c: 56), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

darwini Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Pelycops darwini Aldrich, 1934: 169. Holotype female (NHMUK). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Puerto del Hambre [as “Port Famine”].

Notes: *Pelycops darwini* was recorded from both Argentina and Chile in the original description. There is variation in the amount of orange setation on the abdomen and there could be more than one species.

References: Verbeke (1962: 98, pl. V fig. 6), description and figure of male terminalia; Cortés (1963: 245), notes on holotype in NHMUK; Cortés (1986: 147), taxonomic notes; Stireman et al. (2016: 37), habitus images of *P.* “nr. *darwini*”.

Genus *PIRIONIMYIA* Townsend, 1931

PIRIONIMYIA Townsend, 1931c: 343. Type species: *Pirionimyia paradoxa* Townsend, 1931, by original designation [Chile].

PIRIONOMYIA. Incorrect subsequent spelling of *Pirionimyia* Townsend, 1931 (Aldrich 1934: 5, 105).

References: Aldrich (1934: 5, 105), in key to Patagonian genera, taxonomic notes (as “*Pirionomyia*”); Townsend (1936b: 131), diagnosis of adults and immatures of Prosenini and key to genera (including *Pirionimyia*); Townsend (1938: 361), redescription.

paradoxa Townsend, 1931.—Neotropical: South America (Chile).

Pirionimyia paradoxa Townsend, 1931c: 344. Holotype female (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

Reference: Aldrich (1934: 105), redescription, head figure.

Genus *PSECACERA* Bigot, 1880

PSECACERA Bigot, 1880: 69 [also 1880: liii]. Type species: *Psecacera chiliensis* Bigot, 1880, by monotypy [Chile].

TRIXODOPSIS Townsend, 1933: 527. Type species: *Trixodopsis facialis* Townsend, 1933, by monotypy (not by original designation as cited by Evenhuis et al. 2015: 270) [Chile].

References: Aldrich (1934: 6, 151), in key to Patagonian genera, synonymy, taxonomic notes, key to six species; Townsend (1936b: 121), diagnosis of *Trichoprosopini* and key to genera (including *Psecacera*); Townsend (1938: 299), redescription of *Psecacera*; Cortés (1986: 141, 142), synonymy of *Trixodopsis* with *Psecacera*, in key to tachinid genera of Aysén and Magallanes regions.

atriventris Aldrich, 1934.—Neotropical: South America (Chile).

Psecacera atriventris Aldrich, 1934: 154. Holotype male (USNM). Type locality: Chile, Los Lagos, Llanquihue, Ensenada.

chiliensis Bigot, 1880.—Neotropical: South America (Argentina, Chile).

Psecacera chiliensis Bigot, 1880: 70 [also 1880: liii]. Syntypes, 2 specimens of unspecified sex [2 males, examined by DMW] (NHMUK). Type locality: Chile.

Selenomyia plena Aldrich, 1928b: 23. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

chiliensis. Incorrect subsequent spelling of *chiliensis* Bigot, 1880 (Stireman et al. 2016: 29, 30).

Note: Aldrich (1928b: 23) described *Selenomyia plena* from two males and two females collected from four localities in Argentina and Chile. Aldrich cited a male “Type” in USNM with “Cat. No. 41383” but did not give the type locality. The holotype in USNM, examined by DMW, is from Angol in Chile.

References: Brauer (1898: 494), taxonomic notes on *Psecacera chiliensis*; Aldrich (1934: 152), synonymy, redescription, figure of male surstylus; Verbeke (1962: 96), description of male terminalia.

facialis (Townsend, 1933).—Neotropical: South America (Chile).

Trixodopsis facialis Townsend, 1933: 527 (named for *Psecacera chiliensis* of Townsend, 1931a, not Bigot, 1880). Holotype male (NHMW). Type locality: Chile.

Psecacera chiliensis of Townsend (1931a: 98), not Bigot, 1880. Misidentification (Townsend 1933: 527).

latiforceps Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Psecacera latiforceps Aldrich, 1934: 155. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Reference: Henry (1987: 194), first record from Chile.

robusta Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Psecacera robusta Aldrich, 1934: 154. Syntypes, 2 males (NHMUK, USNM). Type locality: Chile, Biobío, Concepción, San Rosendo.

Note: Aldrich (1934: 154) described *Psecacera robusta* from two male “cotypes” (i.e., syntypes) but did not name the type depository. There appears to be three specimens labelled as types in collections, one in NHMUK and two in USNM (all examined by DMW).

Reference: Gramajo (1998: 93), first record from Argentina.

tibialis Aldrich, 1934.—Neotropical: South America (Chile).

Psecacera tibialis Aldrich, 1934: 154. Holotype male (USNM). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangue.

virens (Aldrich, 1928).—Neotropical: South America (Chile).

Selenomyia virens Aldrich, 1928b: 22. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales, Chile, near Santiago”, ca. 33°9'S, 71°18'W].

Genus *SETOLESTES* Aldrich, 1934

SETOLESTES Aldrich, 1934: 142. Type species: *Setolestes genalis* Aldrich, 1934, by original designation [Chile].

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Setolestes*); Townsend (1939c: 65), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

genalis Aldrich, 1934.—Neotropical: South America (Chile).

Setolestes genalis Aldrich, 1934: 142. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Chiloé, Castro.

References: Cortés (1963: 244), notes on holotype in NHMUK; Cortés (1973a: 100), taxonomic notes.

Tribe DUFOURIINI

Genus *GONZALEZODORIA* Cortés, 1967

GONZALEZODORIA Cortés, 1967b: 18. Type species: *Gonzalezodoria gonioides* Cortés, 1967, by original designation [Chile].

Notes: *Gonzalezodoria gonioides* is a small dark tachinid with a globous abdomen that Cortés (1967b: 18) noted would run to Dexiinae in the keys of Mesnil (1939), or to the Prosenidae in the family key of Townsend (1936b). In Aldrich's (1934) key to Tachinidae of Patagonia and South Chile it runs to *Myiophasia* Brauer & Bergenstamm, 1891 (now a synonym of *Gnadochaeta* Macquart, 1851) but “con el cual no tiene ningun parecido” [“with which it has no resemblance”] according to Cortés (1967b: 20). Guimarães (1971: 26) did not agree and placed *Gonzalezodoria* in the Myiophasiini along with a dozen species of *Myiophasia* and five monotypic genera. We have examined five females of *G. gonioides* in CNC with the following data: Chile, Los Lagos, Volcán Osorno, La Picada, 600 m, 1980, L. Peña (CNC1546958, CNC1546966–CNC1546969). They clearly belong to the “Oestrophasiini” *sensu* Guimarães (1977a), differing from the genera treated therein in having setae on the parafacial that are continuous with the orbitals on the fronto-orbital plate (as shown in figs 1–2 in Cortés 1967b: 21) and lacking banding on the wing. Guimarães (1977a: 216) remarked that the male terminalia of Oestrophasiini “are very similar to those of the Old World Dufouriini ... but it will be many years before the interrelationships of this difficult group ... becomes clear”. The recent molecular phylogeny of Tachinidae of Stireman et al. (2019: 9 [fig. 4]) places *Oestrophasia* in the Dufouriini and *Gonzalezodoria* belongs there as well.

The male of *G. gonioides* has not been described but a single male in CNC with the following data might be the male of this species: Santiago Metro. Region, Mirador de Los Tres Valles, 1820 m, 7.xii.2015, J.E. O'Hara [CNC487604]. This male is a good match morphologically with the aforementioned females but the colouration is different. The females have a yellow head and thorax and a black abdomen; the male has a yellow head except for black underlying the continuous orbital and parafacial setae, black thorax except for yellow scutellum, and yellow abdomen except for black syntergite 1+2, median vitta and bands posteriorly on tergites 3–5.

gonioides Cortés, 1967.—Neotropical: South America (Chile). (Fig. 4b)

Gonzalezodoria gonioides Cortés, 1967b: 19. Holotype female (EEAM). Type locality: Chile, Coquimbo, Limarí, 15 km southwest of Pachingo, near Parque Nacional Bosque Fray Jorge, 110–250 m.

Tribe EUTRIXINI

The South American genus *Xanthobasis* appears to belong to one of the most basal lineages of the Dexiinae according to the recent molecular phylogeny of the Tachinidae of Stireman et al. (2019: 9). In the same clade is the small North American genus *Eutrixia* Coquillett, 1897. The extant tachinids of early lineages like this one are often difficult to classify morphologically and molecular evidence is helping with their phylogenetic, and hence taxonomic, placements. Historically, Guimarães (1971: 110) followed Townsend (1936c: 49, 1939c: 163) in placing *Xanthobasis* in the tribe Ebeiniini, subfamily Dexiinae (“Dexiidae” *sensu* Townsend). Sabrosky and Arnaud (1965: 981) followed Townsend (1936b: 115, 1938: 258) in placing *Eutrixia* in the tribe Aulacephalini, with the former authors assigning the tribe to subfamily Proseninae (“Dexiinae of authors”) in contrast to Townsend’s placement in the family Oestridae. O’Hara and Wood (2004: 45) transferred *Eutrixia* to the Palpostomatini as the first New World member of the tribe. They were influenced by such similarities as a weak genal dilation, weakly developed postscutellum, lower calypter strongly diverging from scutellum, and shared parasitism of scarab beetles. These are traits of *Xanthobasis* as well. However, the molecular phylogenetic evidence of Stireman et al. (2019: 9) suggests that the Palpostomatini are an Old World lineage and the name Eutrixini therefore applies to *Xanthobasis* and allies (*Eutrixia*, *Isidotus* Reinhard, 1962, and genera named below in the *Xanthobasis* note).

Genus *XANTHOBASIS* Aldrich, 1934

XANTHOBASIS Aldrich, 1934: 110. Type species: *Xanthobasis angustifrons* Aldrich, 1934, by original designation [Argentina].

PROXANTHOBASIS Blanchard, 1966b: 219. Type species: *Proxanthobasis rufipes* Blanchard, 1966, by original designation [Argentina].

Note: Cortés (1973a: 101) synonymised *Proxanthobasis* with *Xanthobasis* and Cortés (1986: 147) later commented that two additional monotypic genera described from Argentina, *Neoxanthobasis* Blanchard, 1966 and *Paraxanthobasis* Blanchard, 1966, may also be synonyms.

We did not examine specimens of these last two nominal genera and cannot comment on the merits of this proposition but we do include all three nominal genera in the Eutrixini based on their presumed close relationship.

References: Aldrich (1934: 110), key to three Patagonian species; Townsend (1936c: 47), diagnosis of Ebeniini and key to genera (including *Xanthobasis*); Townsend (1939c: 163), redescription of *Xanthobasis*; Cortés (1973a: 101), synonymy of *Proxanthobasis* with *Xanthobasis*; Cortés (1986: 143, 147), in key to tachinid genera of Aysén and Magallanes regions, comments on likely additional generic synonymy.

angustifrons Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Xanthobasis angustifrons Aldrich, 1934: 111. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Note: *Xanthobasis angustifrons* was recorded from both Argentina and Chile in the original description.

References: Blanchard (1966b: 225), taxonomic notes; Cortés (1973a: 101), taxonomic notes.

rufescens (Blanchard, 1966).—Neotropical: South America (Argentina, Chile). (Fig. 4c)

Proxanthobasis rufescens Blanchard, 1966b: 222. Holotype male (not located). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

Reference: Cortés (1973a: 101), moved to *Xanthobasis*, first record from Chile.

unicolor Aldrich, 1934.—Neotropical: South America (Argentina, Chile). **New record from Chile.**

Xanthobasis unicolor Aldrich, 1934: 112. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Note: *Xanthobasis unicolor* is recorded from Chile for the first time based on material from Rofuco [in Los Ríos Region, Valdivia Province] in MZSP identified by R. Cortés (ex. unpublished notes of Cortés in UMCE examined by CRG).

References: Blanchard (1966b: 218), redescription and wing figure, assigned to *Paraxanthobasis* Blanchard, 1966; Guimarães (1971: 110), as *Paraxanthobasis unicolor*; Cortés (1986: 147), as *Xanthobasis unicolor*.

Tribe VORIINI

Genus *ACTINOPLAGIA* Blanchard, 1940

ACTINOPLAGIA Blanchard, 1940: 234. Type species: *Actinoplusia koehleri* Blanchard, 1940, by original designation [Argentina].

Reference: Cortés (1967b: 12), key to separate *Actinoplusia* Blanchard and *Chaetodemoticus* Brauer & Bergenstamm.

koehleri Blanchard, 1940.—Neotropical: South America (Argentina, Chile, Uruguay).

Actinoplusia koehleri Blanchard, 1940: 234. Holotype male (MACN). Type locality: Argentina, Buenos Aires, Arrecifes.

References: Parker et al. (1951 [pages unknown], also 1953: 53, 66), first record from Uruguay; Parker (1953: 62), figures of first instar larva and puparium; Blanchard (1963: 170), redescription, head figures; Cortés (1967b: 12), first record from Chile; Mulieri et al. (2013: 165), notes on holotype in MACN.

Genus *ALDRICHIOPA* Guimarães, 1971

APHELOGASTER Aldrich, 1934: 22 (junior homonym of *Aphelogaster* Kolbe, 1897).

Type species: *Aphelogaster coracella* Aldrich, 1934, by original designation [Argentina].

ALDRICHIOPA Guimarães, 1971: 165 (*nomen novum* for *Aphelogaster* Aldrich, 1934).

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Aphelogaster*); Townsend (1940a: 192), redescription of *Aphelogaster*.

coracella (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Aphelogaster coracella Aldrich, 1934: 23. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Reference: Townsend (1940a: 192), first record from Chile.

Genus *ALEXOGLOBLINIA* Cortés, 1945

ALEXOGLOBLINIA Cortés, 1945b: 256. Type species: *Metopomuscopteryx shannoni* Aldrich, 1934, by original designation [Argentina].

shannoni (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Metopomuscopteryx shannoni Aldrich, 1934: 46. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Nahuel Huapi.

Reference: Stireman et al. (2016: 34), first record from Chile (Araucanía Region, Parque Nacional Conguillío, Laguna Conguillío), single female in Stireman collection.

Genus *ALPINOPLAGIA* Townsend, 1931

ALPINOPLAGIA Townsend, 1931d: 475. Type species: *Alpinoplagia boliviana* Townsend, 1931, by original designation [Bolivia].

References: Townsend (1936b: 232), diagnosis of adults and immatures of Voriini and key to genera (including *Alpinoplagia*); Townsend (1939a: 373), redescription; Cortés and Campos (1971: 23, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés and González (1989: 116), in key to genera of Chilean Voriini.

boliviana Townsend, 1931.—Neotropical: South America (Bolivia, Chile).

Alpinoplagia boliviana Townsend, 1931d: 476. Holotype female (NHMW). Type locality: Bolivia, La Paz, Cerro Sillutincara [as “Cuesta de Cillutincara”, ca. 16°17'S, 67°53'W], 11,000 ft.

Reference: Cortés and Campos (1971: 46), first record from Chile, redescription, head figure.

Genus *ATELOGLUTUS* Aldrich, 1934

References: Cortés and Valencia (1972: 66), key to the two subgenera of *Ateloglutus* and the three species of new subgenus *Proteloglutus*; Cortés (1984: 378), in key to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; González (1989), review, key to Chilean species; Cortés and González (1989: 116), in key to genera of Chilean Voriini.

Subgenus *ATELOGLUTUS* Aldrich, 1934

***ATELOGLUTUS* Aldrich, 1934: 24.** Type species: *Ateloglutus ruficornis* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Ateloglutus*); Townsend (1940a: 192), redescription of *Ateloglutus*.

***blanchardi* Cortés, 1979.**—Neotropical: South America (Argentina, Chile).

Ateloglutus (Ateloglutus) blanchardi Cortés, 1979: 77. Holotype female (MLPA). Type locality: Argentina, Santa Cruz, Caleta Olivia, 5 km northwest of Piedrabuena, 130 m.

Reference: González (1989: 226, 227), first description of male, head figure, first record from Chile.

***lanfrancoi* Cortés, 1986.**—Neotropical: South America (Chile).

Ateloglutus (Ateloglutus) lanfrancoi Cortés, 1986: 147. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Última Esperanza, Sierra de Los Baguales, 600 m [ca. 50°47'S, 72°24'W].

***ruficornis* Aldrich, 1934.**—Neotropical: South America (Argentina, Chile).

Ateloglutus ruficornis Aldrich, 1934: 25. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Nahuel Huapi.

Note: *Ateloglutus ruficornis* was recorded from both Argentina and Chile in the original description.

Subgenus *PROTELOGLUTUS* Cortés & Valencia, 1972

***PROTELOGLUTUS* Cortés & Valencia, 1972: 66.** Type species: *Phorichaeta chilensis* Brèthes, 1920, by original designation [Chile].

***chilensis* (Brèthes, 1920).**—Neotropical: South America (Argentina, Chile).

Phorichaeta chilensis Brèthes, 1920a: 42. Type(s), unspecified sex (1 female in MACN, Mulieri et al. 2013: 162). Type locality: Chile, Los Andes, Río Blanco.

References: Aldrich (1934: 25), redescription, first record from Argentina; Cortés (1963: 251), notes on a female with data of name-bearing type in MACN (with no mention of “type” and hence not a lectotype fixation); Cortés and Valencia (1972: 66), in key, taxonomic notes; Mulieri et al. (2013: 162), notes on syntype in MACN.

***nitens* Aldrich, 1934.**—Neotropical: South America (Argentina, Chile).

Ateloglutus nitens Aldrich, 1934: 26. Holotype female (NHMUK). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí.

References: Cortés and Valencia (1972: 66), in key, first record from Chile; Cortés (1979: 77), taxonomic notes.

velardei Cortés & Valencia, 1972.—Neotropical: South America (Argentina, Chile, Peru).

Ateloglutus (*Proteloglutus*) *velardei* Cortés & Valencia, 1972: 67. Holotype male (EESC). Type locality: Peru, Ica, Hacienda Paraya.

Note: *Ateloglutus velardei* was recorded from both Peru and Chile in the original description.

Reference: González (1989: 226, 227), wing figure, first record from Argentina.

Genus **CHAETODEMOTICUS** Brauer & Bergenstamm, 1891

CHAETODEMOTICUS Brauer & Bergenstamm, 1891: 385 [also 1891: 81]. Type species: *Demoticus chilensis* Schiner, 1868, by monotypy [Chile].

References: Townsend (1936b: 218), diagnosis of adults and immatures of Germariini and key to genera (including *Chaetodemoticus*); Townsend (1939a: 319), redescription; Cortés (1967b: 12), key to separate *Chaetodemoticus* and *Actinoplusia* Blanchard; Cortés and Campos (1971: 25, 1974: 115) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions.

chilensis (Schiner, 1868).—Neotropical: South America (Chile).

Demoticus chilensis Schiner, 1868: 324. Holotype male [not female as published, Aldrich 1927b: 5] (NHMW). Type locality: Chile.

References: Aldrich (1927b: 5), redescription of holotype; Cortés (1945c: 24), redescription; Cortés and Campos (1971: 56, 61), notes, head figure.

Genus **CHILOCLISTA** Townsend, 1931

CHILOCLISTA Townsend, 1931c: 334. Type species: *Chiloclista bicolor* Townsend, 1931, by original designation [Chile].

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Chiloclista*); Townsend (1939c: 30), redescription.

bicolor Townsend, 1931.—Neotropical: South America (Chile).

Chiloclista bicolor Townsend, 1931c: 334. Holotype male (USNM). Type locality: Chile, O'Higgins, Cardenal Caro, Tanumé [ca. 34°13'S, 71°55'W].

Reference: Stireman et al. (2016: 37), habitus images.

Genus **CORACOMYIA** Aldrich, 1934

CORACOMYIA Aldrich, 1934: 21. Type species: *Coracomyia crassicornis* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Coracomyia*); Townsend (1940a: 206), redescription; Cortés (1986: 143),

in key to tachinid genera of Aysén and Magallanes regions; Cortés and González (1989: 116), in key to genera of Chilean Voriini.

crassicornis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Coracomyia crassicornis Aldrich, 1934: 22. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Nahuel Huapí.

Reference: Cortés (1976: 8), partial redescription including first description of female, first record from Chile.

woodi Cortés, 1976.—Neotropical: South America (Chile).

Coracomyia woodi Cortés, 1976: 8. Holotype male (MEUC). Type locality: Chile, Los Lagos, Osorno, Parque Nacional Puyehue, Paso Cardenal Antonio Samoré [as “Paso Puyehue”], 1200 m [ca. 40°42’S, 71°57’W].

Genus **CYRTOPHLOEBA** Rondani, 1856

CYRTOPHLOEBA Rondani, 1856: 207. Type species: *Tachina ruricola* Meigen, 1824, by original designation [Europe].

EUCYRTOPHLOEBA Townsend, 1916: 316. Type species: *Eucyrtophloeoba rhois* Townsend, 1916, by original designation [Mexico].

OPSOPHAGUS Aldrich, 1926a: 15. Type species: *Opsophagus ornatus* Aldrich, 1926, by original designation [Peru].

CYRTHOPHLEBA. Incorrect subsequent spelling of *Cyrtophloeoba* Rondani, 1856 (Rondani 1857: 13) (see O’Hara et al. 2011: 68).

CYRTOPHLEBA. Incorrect original spelling of *Cyrtophloeoba* Rondani, 1856 (Rondani 1856: 68) (see O’Hara et al. 2011: 69).

References: Coquillett (1910: 530), type species of *Cyrtophloeoba* (as “*Cyrtophleba*”); Aldrich (1934: 3, 32), in key to Patagonian genera, taxonomic notes (as *Opsophagus*); Townsend (1936b: 232), diagnosis of adults and immatures of Voriini and key to genera (including *Cyrtophloeoba* [as “*Cyrtophleba*”], *Eucyrtophloeoba* and *Opsophagus*); Townsend (1939a: 378, 379, 393), redescrptions of *Cyrtophloeoba* (as “*Cyrtophleba*”), *Eucyrtophloeoba* and *Opsophagus*; Caltagirone (1966: 63), key to Neotropical species (as *Opsophagus*); Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions (as *Opsophagus*); Cortés and González (1989: 116), in key to genera of Chilean Voriini (as *Opsophagus*); Wood and Zumbado (2010: 1402), synonymy of *Opsophagus* with *Cyrtophloeoba* (as “*Cyrtophleba*”).

cortesi (Caltagirone, 1966).—Neotropical: South America (Argentina, Chile).

Opsophagus cortesi Caltagirone, 1966: 64. Holotype male (INLA). Type locality: Chile, Maule, Talca, Gualleco.

References: Cortés (1967b: 12), taxonomic notes; Gramajo (1998: 95), first record from Argentina.

nigripalpis (Aldrich, 1926).—Neotropical: South America (Argentina, Chile, Ecuador).

Opsophagus nigripalpis Aldrich, 1926a: 16. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9’S, 71°18’W].

References: Aldrich (1934: 33), redescription, taxonomic notes, first record from Argentina; Cortés (1979: 80), taxonomic notes; Cortés (1980: 106), first record from Ecuador.

Genus *DISCHOTRICHIA* Cortés, 1944

DISCHOTRICHIA Cortés, 1944f: 54. Type species: *Dischotrichia caelibata* Cortés, 1944, by original designation [Chile].

Reference: Cortés (1975: 36), in key to related genera.

caelibata Cortés, 1944.—Neotropical: South America (Chile).

Dischotrichia caelibata Cortés, 1944f: 56. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga.

Reference: Campos (1953: 25), first description of female.

Genus *GANOPLEURON* Aldrich, 1934

GANOPLEURON Aldrich, 1934: 118. Type species: *Ganopleuron divergens* Aldrich, 1934, by original designation [Chile].

divergens Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Ganopleuron divergens Aldrich, 1934: 119. Holotype female (NHMUK). Type locality: Chile, Los Lagos, Chiloé, Castro.

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Ganopleuron*); Townsend (1939c: 39), redescription; Cortés (1963: 243), notes on holotype in NHMUK; Cortés (1973a: 98), first description of male; Gramajo (1998: 94), first record from Argentina.

Genus *LAFUENTEMYIA* Marnef, 1965

LAFUENTEMYIA Marnef, 1965: 243. Type species: *Lafuentemyia yanezi* Marnef, 1965, by original designation [Chile].

Reference: Cortés (1975: 36), in key to genera with a modified hind femur in male.

yanezi Marnef, 1965.—Neotropical: South America (Chile).

Lafuentemyia yanezi Marnef, 1965: 246. Holotype male (UVVC). Type locality: Chile, Valparaíso, Valparaíso, Reserva Nacional Lago Peñuelas.

Reference: Cortés (1975: 36), taxonomic notes.

Genus *MYIOCHAETA* Cortés, 1967

MYIOCHAETA Cortés, 1967b: 24. Type species: *Myiochaeta marnefi* Cortés, 1967, by original designation [Chile].

Reference: Cortés and González (1989: 116), in key to genera of Chilean Voriini.

marnefi Cortés, 1967.—Neotropical: South America (Chile).

Myiochaeta marnefi Cortés, 1967b: 25. Holotype male (EEAM). Type locality: Chile, Metropolitana de Santiago, Santiago, Maipú, Rinconada, Quebrada de La Plata, 510 m.

Genus **NEOCHAETOPLAGIA** Blanchard, 1963

NEOCHAETOPLAGIA Blanchard, 1963: 173. Type species: *Neochaetoplagia pastranai* Blanchard, 1963, by original designation [Argentina].

Reference: Cortés and González (1989: 116), in key to genera of Chilean Voriini.

pastranai Blanchard, 1963.—Neotropical: South America (Argentina, Chile).

Neochaetoplagia pastranai Blanchard, 1963: 173. Holotype male (presumed lost, Mulieri et al. 2013: 168). Type locality: Argentina, Buenos Aires, Buenos Aires [as “Capital Federal”].

References: Cortés and González (1989: 118), first record from Chile; Mulieri et al. (2013: 168), notes on type series.

Genus **NOTHOVORIA** Cortés & González, 1989

NOTHOVORIA Cortés & González, 1989: 120. Type species: *Nothovoria praestans* Cortés & González, 1989, by original designation [Chile].

Reference: Cortés and González (1989: 116), in key to genera of Chilean Voriini.

praestans Cortés & González, 1989.—Neotropical: South America (Chile).

Nothovoria praestans Cortés & González, 1989: 120. Holotype female (UMCE). Type locality: Chile, Tarapacá, Iquique, 40 km from Iquique, Pampa del Tamarugal, Estación Refresco, 1200 m.

Genus **PHAEODEMA** Aldrich, 1934

PHAEODEMA Aldrich, 1934: 145. Type species: *Phaeodema mystacina* Aldrich, 1934, by original designation [Chile].

References: Townsend (1936c: 47), diagnosis of Ebeiniini and key to genera (including *Phaeodema*); Townsend (1939c: 160), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

mystacina Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Phaeodema mystacina Aldrich, 1934: 145. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Puerto Montt.

Reference: Gramajo (1998: 94), first record from Argentina.

Genus *PIRIONA* Aldrich, 1928

PIRIONA Aldrich, 1928b: 24. Type species: *Piriona fasciculata* Aldrich, 1928, by original designation [Chile].

References: Aldrich (1934: 3, 44), in key to Patagonian genera, taxonomic notes; Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Piriona*); Townsend (1939c: 58), redescription; Cortés (1975: 36), in key to genera with a modified hind femur in male.

fasciculata Aldrich, 1928.—Neotropical: South America (Argentina, Chile).

Piriona fasciculata Aldrich, 1928b: 24. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga.

Note: Aldrich (1928b: 25) described *Piriona fasciculata* from five males and one female collected from three localities in Argentina and Chile. Aldrich cited a male “Type” in USNM with “Cat. No. 41385” but did not give the type locality. The holotype in USNM was examined by DMW and is from Marga Marga Province in Chile (see also Aldrich 1934: 45). References: Aldrich (1934: 45), redescription; Cortés (1963: 243), notes on specimens in NHMUK; Cortés (1975: 36), taxonomic notes.

Genus *PROSOPOCHAETA* Macquart, 1851

PROSOPOCHAETA Macquart, 1851: 183 [also 1851: 210] (as “*Prosopochaeta*”, see note). Type species: *Prosopochaeta nitidiventris* Macquart, 1851, by original designation [Chile].

PUNACLISTA Townsend, 1915e: 406. Type species: *Punaclista setosa* Townsend, 1915, by original designation [Peru].

PROSOPOCHOETA. Incorrect original spelling of *Prosopochaeta* Macquart, 1851 (Macquart 1851: 183, see note).

Notes: The name *Prosopochaeta* Macquart, 1851 was originally published as *Prosopochaeta* but subsequent authors (e.g., Aldrich 1934; Cortés and Hichins 1969; Guimarães 1971; González 1992b) used the spelling *Prosopochaeta*. This changed spelling would normally be considered an incorrect subsequent spelling but because it is in prevailing usage and is attributed to Macquart (1851), it is deemed to be the correct original spelling (Article 33.3.1 of the Code, ICZN 1999).

Macquart (1851: 184 [also 1851: 211]) noted about his new genus *Prosopochaeta* (as *Prosopochaeta*), “Le type de ce genre est du Chili” [“The type of this genus is from Chile”]. This statement is accepted as a type species designation for *Prosopochaeta* of the single included species, *Prosopochaeta nitidiventris* Macquart, from Chile.

References: Aldrich (1934: 5, 115), in key to Patagonian genera, synonymy of *Punaclista* with *Prosopochaeta*, redescription; Townsend (1936b: 121), diagnosis of Trichoprosopini and key to genera (including *Prosopochaeta*); Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Punaclista*); Townsend (1938: 299), redescription of *Prosopochaeta*; Townsend (1939c: 62), redescription of *Punaclista*; Parker (1953: 66), figures of puparium of *Prosopochaeta* sp.; Cortés and Campos (1971: 45), taxonomic notes

including reinstating synonymy of *Punaclista* with *Prosopochaeta*; Cortés and Campos (1971: 26, 1974: 116) and Cortés (1984: 381), in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 142), in key to tachinid genera of Aysén and Magallanes regions.

anomala Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Prosopochaeta anomala Aldrich, 1934: 118. Holotype male (USNM). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí, Jones Estancia.

Note: *Prosopochaeta anomala* was recorded from both Argentina and Chile in the original description.

caliginosa Cortés & Campos, 1971.—Neotropical: South America (Argentina, Chile).

Prosopochaeta caliginosa Cortés & Campos, 1971: 43. Holotype male (EEAM). Type locality: Chile, Antofagasta, Antofagasta, north of Quebrada de Paposo, 200–1000 m (25°03'S, 70°25'W) (coordinates and elevation given on p. 12).

Reference: Cortés (1979: 81), first record from Argentina.

nitidiventris Macquart, 1851.—Neotropical: South America (Argentina, Chile).

Prosopochaeta nitidiventris Macquart, 1851: 184 [also 1851: 211]. Lectotype male (MNHN), by designation herein (see Lectotype Designations section). Type locality: Chile (“Coquimbo, etc.” according to Blanchard 1854: 424).

Notes: *Punaclista setosa* Townsend, 1915 from Peru was treated as a synonym of *Prosopochaeta nitidiventris* Macquart by Aldrich (1934: 116) but is currently recognised as a valid species of *Prosopochaeta* (e.g., Cortés and Campos 1971: 46; Guimarães 1971: 100).

References: Aldrich (1934: 116), redescription, head figure, first record from Argentina; Verbeke (1962: 103), description of male terminalia.

Genus *TRICHODISCHIA* Bigot, 1885

TRICHODISCHIA Bigot, 1885a: 237. *Nomen nudum*.

TRICHODISCHIA Bigot, 1885b: xlv [also 1885b: xlv, *Bull. Soc. Ent. France*]. Type species: *Trichodischia soror* Bigot, 1885, by subsequent designation of Townsend (1916b: 9) [Argentina].

TRICHORAEA Cortés, 1975: 37. Type species: *Trichodischia caerulea* Bigot, 1885, by original designation [Argentina].

TRICODISCHIA. Incorrect subsequent spelling of *Trichodischia* Bigot, 1885 (Henry 1987: 194).

Note: The two Bigot species *Trichodischia caerulea* and *T. soror* were treated as generically different by Cortés (1975: 37) and the former was assigned to new genus *Trichoraea* Cortés. Subsequent authors have continued to treat the original combination *Trichodischia caerulea* as valid (e.g., Guimarães 1977c: 76; Henry 1987: 195; Molina-Ochoa et al. 2003: 269) and this classification is followed here, but without further study.

References: Townsend (1936c: 13), diagnosis of adults and immatures of Macquartiini and key to genera (including *Trichodischia*); Townsend (1939c: 68), redescription of *Trichodischia*; Parker (1953: 66), figures of first instar larva and puparium of *Trichodischia* sp., from Argentina; Cortés (1969: 98), key to separate the two species; Cortés (1975: 36), *Trichodischia* and *Trichoraea* in key to genera with a modified hind femur in male.

caerulea Bigot, 1885.—Neotropical: South America (Argentina, Chile, Uruguay).

Trichodischia caerulea Bigot, 1885b: xlv [also 1885b: xlv, *Bull. Soc. Ent. France*].

Syntypes, 3 specimens as “♀?” (2 males and 1 female in NHMUK, examined by DMW). Type locality: Argentina, Buenos Aires, Buenos Aires.

caerulai. Incorrect subsequent spelling of *caerulea* Bigot, 1885 (Blanchard 1963: 184).

caerulia. Incorrect subsequent spelling of *caerulea* Bigot, 1885 (Blanchard 1963: 185).

coerulea. Incorrect subsequent spelling of *caerulea* Bigot, 1885 (Brauer 1899: 498).

References: Cortés (1969: 98), separation from *Trichodischia soror*, first record from Chile; Cortés (1975: 36, 37), first record from Uruguay (distribution given for *Trichoraea* in key, a monotypic genus based on *T. caerulea*), taxonomic notes.

soror Bigot, 1885.—Neotropical: South America (Argentina, Brazil, Chile, Uruguay).

Trichodischia soror Bigot, 1885b: xlvi [also 1885b: xlvi, *Bull. Soc. Ent. France*]. Holotype male (NHMUK). Type locality: Argentina, Buenos Aires, Buenos Aires.

Trichodischia caerulea of Cortés (1944f: 51, 1946: 175) and Blanchard (1963: 184), not Bigot, 1885. Misidentification (Cortés 1969: 97, 1975: 37).

References: Cortés (1944f: 54), first records from Chile and Uruguay, misidentified as *Trichodischia caerulea*; Cortés (1969: 98), separation from *T. caerulea*; Cortés (1975: 37), taxonomic notes; Cortés (1980: 105), first record from Brazil.

Genus **VELARDEMYIA** Valencia, 1972

VELARDEMYIA Valencia, 1972a: 364. Type species: *Velardemyia ica* Valencia, 1972, by original designation [Peru].

Reference: Cortés and González (1989: 116), in key to genera of Chilean Voriini.

ica Valencia, 1972.—Neotropical: South America (Chile, Peru).

Velardemyia ica Valencia, 1972a: 364. Holotype male (SENASA, Lozada et al. 2005: 460). Type locality: Peru, Ica, Arrabales.

Reference: Cortés and González (1989: 122), first record from Chile.

Genus **VORIA** Robineau-Desvoidy, 1830

VORIA Robineau-Desvoidy, 1830: 195. Type species: *Voria latifrons* Robineau-Desvoidy, 1830 (= *Tachina ruralis* Fallén, 1810), monotypy [France].

PLAGIA Meigen, 1838: 201. Type species: *Tachina verticalis* Meigen, 1824 (= *Tachina ruralis* Fallén, 1810), by subsequent designation of Rondani (1856: 69) [Europe].

XENOPLAGIA Townsend, 1914a: 13. Type species: *Xenoplagia setosa* Townsend, 1914, by original designation [Peru].

ITAVORIA Townsend, 1931d: 474. Type species: *Itavoria aurescens* Townsend, 1931, by original designation [Brazil].

References: Coquillett (1910: 591, 619), type species of *Plagia* and *Voria* (with *Plagia* [and others] in synonymy with *Voria*); Townsend (1936b: 232), diagnosis of adults and immatures of Voriini and key to genera (including *Itavoria*, *Voria* and *Xenoplagia*); Townsend (1936c: 280), *Plagia* as synonym of *Voria*; Townsend (1939a: 385, 402, 403), redescrptions

of *Itavoria*, *Voria* (with *Plagia* in synonymy) and *Xenoplagia*; Mesnil (1974: 1261), redescription (with *Plagia* in synonymy), taxonomic notes; Cortés and Campos (1974: 113) and Cortés (1984: 379), *Voria* in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés and González (1989: 116), *Voria* in key to genera of Chilean Voriini; Fleming et al. (2017: 7), synonymy of *Itavoria* and *Xenoplagia* with *Voria* [also the synonymy with *Voria* of the Afrotropical and Oriental genus *Hystriovoria* Townsend, 1928 and its synonyms *Afro- voria* Curran, 1938 and *Anavoria* Mesnil, 1953, but the present authors follow O'Hara and Cerretti (2016: 56) in recognising *Hystriovoria* as a valid genus].

ruralis (Fallén, 1810).—Neotropical: southern Lesser Antilles (Trinidad & Tobago), Middle America (Mexico, Nicaragua), South America (Argentina, Brazil, Chile, Colombia, Peru, Uruguay, Venezuela). Nearctic: Canada, United States. Palaearctic: Central Asia, China [Pal.], Europe, Japan, Korean Peninsula, Middle East, Mongolia, Russia, Transcaucasia. Afrotropical: Kenya to South Africa, Yemen. Oriental: China (Yunnan), India, Nepal, Pakistan, Taiwan. Australasian & Oceanian: Australia, Papua New Guinea.

Tachina ruralis Fallén, 1810: 265. Lectotype male (NHRS), by designation of Crosskey (1973: 163). Type locality: Sweden, Skåne, Äsperöd [as “Esperöd”].

Plagia americana van der Wulp, 1890c: 102. Syntypes, males and females (NHMUK). Type localities: Mexico, Veracruz (Orizaba), Guerrero (Venta del Zopilote [ca. 17°46'N, 99°32'W], 2800 ft; Xocomanatlán [as “Xucumanatlan”, ca. 17°34'N, 99°37'W], 7000 ft; Omiltemi [as “Omilteme”, ca. 17°33'N, 99°41'W], 8000 ft), and Tabasco (Teapa).

Plagia mexicana Giglio-Tos, 1893: 5. Type(s), female (MZUT). Type locality: Mexico.

Voria brasiliiana Townsend, 1929: 380. Syntypes, “many males and females” (USNM). Type locality: Brazil, São Paulo, Itaquaquecetuba.

Voria ayerzai Blanchard, 1937: 47 (as “*Voria ayerzai*, (Brethes)”). *Nomen nudum*.

Voria ayerzai Blanchard, 1943c: 157 (as “*Plagia ayerzai*, Brèthes in lit.”). Syntypes, 3 males and females (MLPA). Type locality: Argentina, Buenos Aires [province or city].

Notes: The mention of a “Ht” for *Tachina ruralis* from Sweden in NHRS by Townsend (1939a: 402) is not accepted as a lectotype fixation because the specimen in question is not distinguishable from the other specimens in the type series.

Voria ruralis as here interpreted is almost certainly a species complex (see also Fleming et al. 2017). This complex in the New World may not include the true *Voria ruralis* described from the Palaearctic Region by Fallén (1810). Some of the names listed here in synonymy may represent distinct species.

References: Aldrich (1926a: 14), synonymy of *Plagia americana* with *Tachina ruralis*; Cortés (1944d: 142), *Voria brasiliiana* and *Voria ayerzai* as possible synonyms of *Tachina ruralis*, first record from Chile; Parker (1953: 64), figures of first instar larva and puparium; Thompson (1961: 28), synonymy including *Plagia mexicana* as a questionable synonym of *Tachina ruralis*, redescription, first record from Trinidad; Blanchard (1963: 169), taxonomic notes, record from Argentina; Guimarães (1971: 93), synonymy; Nihei (2016: 911), first record from Colombia.

Unplaced genus of Dexiinae

Genus *SCHLINGERMYIA* Cortés, 1967

SCHLINGERMYIA Cortés, 1967b: 20. Type species: *Schlingermiya venusta* Cortés, 1967, by original designation [Chile].

Note: This genus was not placed beyond subfamily Dexiinae by the original author, Cortés (1967: 20). Guimarães (1971: 100) listed *Schlingermiya* under subfamily Dexiinae, tribe Macquartiini, following Townsend's (1936c: 13) concept of the tribe. Most of the Neotropical genera then assigned to the Macquartiini are currently placed in the dexiine tribes Dexiini and Voriini.

venusta Cortés, 1967.—Neotropical: South America (Chile).

Schlingermiya venusta Cortés, 1967b: 22. Holotype male (EEAM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Los Perales”, ca. 33°9'S, 71°18'W].

Subfamily EXORISTINAE

Tribe ACEMYINI

Genus *CERACIA* Rondani, 1865

CERACIA Rondani, 1865: 221. Type species: *Ceracia mucronifera* Rondani, 1865, by monotypy [Italy].

MYOTHYRIA van der Wulp, 1890a: 44, in key [1890e: 208, description]. Type species: *Myothyria majorina* van der Wulp, 1890, by subsequent designation of Coquillett (1910: 573) (see O'Hara and Cerretti 2016: 63) [Mexico].

ACEMYIOPSIS Townsend, 1915e: 433. Type species: *Acemyiopsis punensis* Townsend, 1915, by original designation [Peru].

CLYTHOPSIS Townsend, 1927a: 276. Type species: *Clythopsis confundens* Townsend, 1927 (= *Myobia brachyptera* Thomson, 1869), by original designation [Brazil].

References: Coquillett (1910: 573), type species of *Myothyria*; Aldrich (1934: 5, 136), in key to Patagonian genera, synonymy of *Acemyiopsis*, *Clythopsis* and *Myothyria* with *Ceracia*, taxonomic notes; Townsend (1936c: 71, 270, 273, 278), diagnosis of adults and immatures of Acemyini and key to genera (including *Ceracia*), synonymy; Townsend (1939c: 255), redescription of *Ceracia* (with *Acemyiopsis*, *Clythopsis* and *Myothyria* in synonymy).

dentata (Coquillett, 1895).—Neotropical: Middle America (Mexico), South America (Chile). Nearctic: Canada, United States.

Acemyia dentata Coquillett, 1895a: 311. Syntypes, 4 females (2 females in USNM [one with abdomen missing] and 2 females in MCZ). Type localities: USA, Florida (Georgetown), Alabama (Mobile), and California (Los Angeles County).

Reference: Aldrich (1934: 137), redescription, first record from Chile.

subandina Blanchard, 1943.—Neotropical: South America (Argentina, ?Chile).

Ceracia subandina Blanchard, 1943b: 19. Holotype male (INTA, Patitucci et al. 2015: 567). Type locality: Argentina, Río Negro, Comallo.

Note: Guimarães (1971: 123) recorded *Ceracia subandina* from three places in Chile (“Angol, Concepción, Santiago”) but we have not found any of these records in the Chilean literature and suspect that they were listed for *C. subandina* in error.

Tribe BLONDELIINI

References: Sabrosky (1981: 3), key to genera of Blondeliini in which females possess an abdominal keel and sharp, curved piercer; Wood (1985), revision of the Blondeliini of North and Central America and the West Indies.

Genus *ADMONTIA* Brauer & Bergenstamm, 1889

GRAVENHORSTIA Robineau-Desvoidy, 1863a: 924 (junior homonym of *Gravenhorstia* Boie, 1836). Type species: *Gravenhorstia longicornis* Robineau-Desvoidy, 1863 (= *Tachina grandicornis* Zetterstedt, 1849), by original designation [France].

ADMONTIA Brauer & Bergenstamm, 1889: 104 [also 1890: 36]. Type species: *Admontia podomyia* Brauer & Bergenstamm, 1889, by monotypy [Austria, Germany, Italy, Poland, Germany and Czech Republic].

TRICHOPAREIA Brauer & Bergenstamm, 1889: 103 [also 1890: 35]. Type species: *Tachina seria* Meigen, 1824, by monotypy [Germany].

AUSTROSTAUROCHAETA Townsend, 1931d: 476. Type species: *Degeeria antarctica* Thomson, 1869, by original designation [probably Chile].

POLIOPS Aldrich, 1934: 94. Type species: *Poliops striatus* Aldrich, 1934, by original designation [Argentina]. **Syn. nov.**

Notes: The relative priority of *Admontia* Brauer & Bergenstamm, 1889 and *Trichopareia* Brauer & Bergenstamm, 1889, when the two are treated as synonyms, was established by Strobl (1910: 137), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999).

The new synonymy of *Poliops* with *Admontia* is explained below under *Admontia striata*. References: Coquillett (1910: 503), type species of *Admontia* (as synonym of *Hyperecteina* Schiner, 1861); Aldrich (1934: 4, 94, 95), *Admontia* and *Poliops* in key to Patagonian genera, synonymy including *Trichopareia* and *Austrotaurochaeta* with *Admontia*, taxonomic notes, key to six Patagonian species of *Admontia*; Townsend (1936c: 129, 154), diagnosis of adults and immatures of Actiini and key to genera (including *Austrotaurochaeta* and *Poliops*), diagnosis of adults and immatures of Trichopareiini and key to genera (including *Admontia* and *Trichopareia*); Townsend (1940a: 193, 254, 301, 304), redescrptions of *Admontia*, *Austrotaurochaeta*, *Poliops* and *Trichopareia*; Cortés (1973a: 100), separation of *Admontia* and *Austrotaurochaeta* from *Notomanes* Aldrich; Wood (1985: 14, 17), in key to the Blondeliini of North and Central America and the West Indies, synonymy including *Austrotaurochaeta* with *Admontia*, diagnosis, taxonomic notes; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions.

antarctica (Thomson, 1869).—Neotropical: South America (Argentina, Chile).

Degeeria antarctica Thomson, 1869: 527. Lectotype male (NHRS), by fixation of Townsend (1931b: 183) (examination of “Male Ht” from Patagonia in NHRS is regarded as a lectotype fixation). Type locality: “Patagonia” (most likely Chile, Magallanes y de la Antártica Chilena, Magallanes, Puerto del Hambre [frequently as “Port Famine”], based on localities where insects were collected during the voyage of the Swedish frigate *Eugenie*, including the lectotype of *Degeeria antarctica*; see Persson 1971: 168).

References: Aldrich (1934: 97), redescription, first record from Argentina; Cortés (1973a: 97), taxonomic notes.

aurata (Campos, 1953).—Neotropical: South America (Chile). **Comb. nov.**

Poliops auratus Campos, 1953: 27. Holotype male (MNNC). Type locality: Chile, Biobío, Concepción, Tomé.

Note: *Poliops auratus* is assumed to have the same generic features as the type species of *Poliops*, *P. striatus*, and for this reason is transferred to *Admontia*. The transfer of *P. striatus* to *Admontia* is discussed below under *A. striata*.

calyptrata (Aldrich, 1934).—Neotropical: South America (Argentina, Chile). **Comb. nov.** (Fig. 4d)

Phorocera calyptrata Aldrich, 1934: 73. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Correntoso.

Note: *Phorocera calyptrata* Aldrich has the *Admontia* features of a setose facial ridge, haired parafacial and tiny fore claws in the female but the eye has scattered long hairs rather than the usual bare condition. We interpret the species as an aberrant member of the *Admontia* lineage and move it here to *Admontia* from its prior indefinite placements in Tachinidae (see References below).

References: Aldrich (1934: 69), in key to Patagonian species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*); Cortés (1945d: 158), in key to Chilean species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*) and *Parasetigena* Brauer & Bergenstamm, 1891, known from Argentina and not Chile; Guimarães (1971: 152), as unplaced species of Blondeliini; Henry (1987: 206), first record from Chile (in *Phorocera* but genus unplaced in Tachinidae); González (1992b: 183), record from Chile, as *Phorocera calyptrata* (*sensu* previous authors).

communis Aldrich, 1934. —Neotropical: South America (Argentina, Chile).

Admontia communis Aldrich, 1934: 99. Holotype, unspecified sex (NHMUK).

Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí.

Admontia communis albescens Aldrich, 1934: 100. Syntypes, 5 males (NHMUK).

Type localities: Argentina, Río Negro, eastern end of Lago Nahuel Huapí and San Carlos de Bariloche [as “Bariloche”].

Notes: *Admontia communis* was recorded from both Argentina and Chile in the original description. Cortés and Hichins (1969: 16) gave the depository for the holotype of *A. communis* as USNM, in error.

debilis Aldrich, 1934. —Neotropical: South America (Argentina, Chile).

Admontia debilis Aldrich, 1934: 102. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangué.

finisterrae Cortés, 1986.—Neotropical: South America (Chile).

Admontia finisterrae Cortés, 1986: 155. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Antártica Chilena, Islas Hermite, Isla Deceit, Caleta Toledo.

flavibasis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Admontia flavibasis Aldrich, 1934: 103. Holotype female (USNM). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Reference: González (1992b: 178), first record from Chile.

pictiventris Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Admontia pictiventris Aldrich, 1934: 100. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Peulla.

Reference: Gramajo (1998: 96), first record from Argentina.

striata (Aldrich, 1934).—Neotropical: South America (Argentina, Chile). **Comb. nov.**

Poliops striatus Aldrich, 1934: 94. Holotype male (NHMUK). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí.

Notes: *Poliops striatus* was recorded from both Argentina and Chile in the original description.

Aldrich (1934: 94) described *Poliops* as a monotypic genus based on the new species *P. striatus*, noting: "Very similar to *Admontia*, but costal spine about as long as distance between auxiliary and first vein on costa, and third vein at base with a single setula of about the same length". A male homotype of *P. striatus* in CNC that DMW compared to the holotype in NHMUK possesses the usual features of the Blondeliini and diagnostic characters of *Admontia* (Wood 1985: 18), including bare eye, setose facial ridge, and a few hairs on the upper parafacial below lowest frontal seta. A single setula at the base of wing vein R_{4+5} is not unique in *Admontia* to the two species originally described in *Poliops* and was even given in the original description as a characteristic of *A. finisterrae* Cortés, 1986.

Genus *EUCELATORIA* Townsend, 1909

EUCELATORIA Townsend, 1909: 249. Type species: *Tachina armigera* Coquillett, 1889, by monotypy [United States].

SPATHIMYIA Townsend, 1912b: 318. Type species: *Spathimyia ferox* Townsend, 1912, by original designation [Peru].

XIPHOMYIA Townsend, 1917: 125. Type species: *Xiphomyia gladiatrix* Townsend, 1917, by original designation [Panama].

URODEXODES Townsend, 1919b: 572. Type species: *Urodexodes charapensis* Townsend, 1919, by original designation [Peru].

MACHAIROMASICERA Townsend, 1919b: 577. Type species: *Machairomasicera carinata* Townsend, 1919, by original designation [Ecuador].

LIXINIA Curran, 1926: 108. Type species: *Lixinia jamaicensis* Curran, 1926, by original designation [Jamaica].

TINALYDELLA Townsend, 1927a: 265. Type species: *Tinalydella tinensis* Townsend, 1927, by original designation [Peru].

- OROPHOROCERA* Townsend, 1927a: 267. Type species: *Orophorocera ocellaris* Townsend, 1927, by original designation [Peru].
- HYPOMYOTHYRIA* Townsend, 1927a: 276. Type species: *Hypomyothyria hypodermica* Townsend, 1927, by original designation [Brazil].
- EUCELATORIOPSIS* Townsend, 1927a: 276. Type species: *Eucelatoriopsis teffeensis* Townsend, 1927, by original designation [Brazil].
- HELIOLYDELLA* Townsend, 1927a: 277. Type species: *Heliolydella aurata* Townsend, 1927, by original designation [Brazil].
- TACHINOPHYTOPSIS* Townsend, 1927a: 277. Type species: *Tachinophytopsis carinata* Townsend, 1927 (junior secondary homonym of *Machairomasicera carinata* Townsend, 1919; = *Eucelatoria paracarinata* Nihei & Dios, 2016), by original designation [Brazil].
- HEMILYDELLA* Townsend, 1927a: 278. Type species: *Hemilydella fasciata* Townsend, 1927, by original designation [Peru].
- LYDELLOHOUGHIA* Townsend, 1927a: 280. Type species: *Lydellohoughia nana* Townsend, 1927, by original designation [Brazil].
- EUPTILODEGEERIA* Townsend, 1931d: 465. Type species: *Hypostena obumbrata* van der Wulp, 1890, by original designation [Mexico].
- COROZALIA* Curran, 1934: 465. Type species: *Corozalia longula* Curran, 1934, by original designation [Panama].
- CELATORIOPSIS* Blanchard, 1963: 228. Type species: *Celatoriopsis eucelatorioides* Blanchard, 1963, by original designation [Argentina].
- EUCELATORIOIDEA* Thompson, 1968: 176. Type species: *Eucelatorioidea nigripalpis* Thompson, 1968 (junior secondary homonym of *Chetolyga nigripalpis* Bigot, 1889; = *Eucelatoria nudiculata* O'Hara & Wood, **nom. nov.**, see below), by original designation [Trinidad & Tobago].
- DEXODIMYIA* Thompson, 1968: 181. Type species: *Dexodimyia discalis* Thompson, 1968, by original designation [Trinidad & Tobago].
- PSEUDOCELATORIA* Thompson, 1968: 190. Type species: *Pseudocelatoria robusta* Thompson, 1968, by original designation [Trinidad & Tobago].
- HELIODEXODES* Thompson, 1968: 197. Type species: *Heliodexodes argenteus* Thompson, 1968, by original designation [Trinidad & Tobago].
- DEXODIOPSIS* Thompson, 1968: 202. Type species: *Dexodiopsis aurea* Thompson, 1968, by original designation [Trinidad & Tobago].

Notes: The relative priority of *Eucelatorioidea* Thompson, 1968, *Dexodimyia* Thompson, 1968, *Pseudocelatoria* Thompson, 1968, *Heliodexodes* Thompson, 1968 and *Dexodiopsis* Thompson, 1968, when the five are treated as synonyms, has not been established and is not of concern while all are junior synonyms of *Eucelatoria* Townsend, 1909 (as proposed by Wood 1985: 40).

References: Townsend (1936c: 86, 237), diagnosis of adults and immatures of Compsilurini and key to genera (including *Eucelatoria*, *Eucelatoriopsis*, *Euptilodegeeria*, *Heliolydella*, *Hemilydella*, *Hypomyothyria*, *Machairomasicera*, *Orophorocera*, *Spathimyia*, *Tachinophytopsis*, *Tinilydella* and *Urodexodes*), diagnosis of adults and immatures of Trypherini and key to genera (including *Corozalia*, *Lixinia*, *Lydellohoughia* and *Xiphomyia*); Townsend (1940a: 48, 50, 53,

54, 55, 56, 63, 76, 94, 97, 98, 100), redescrptions of *Eucelatoria*, *Eucelatoriopsis*, *Euptilodegeeria*, *Heliolydella*, *Hemilydella*, *Hypomyothyria*, *Machairomasicera*, *Orophorocera*, *Spathimyia*, *Tachinophytopsis*, *Tinalydella* and *Urodexodes*; Townsend (1941: 258, 279, 281, 327), redescrptions of *Corozalia*, *Lixinia*, *Lydellohoughia* and *Xiphomyia*; Thompson (1968: 174, 176), revision of *Eucelatoria* species of Trinidad, as nine genera (five new) collectively termed the “Trinidad compsilurines”; Cortés and Campos (1971: 26, 1974: 115, 116), *Hemilydella* and *Eucelatoria* in keys to tachinid genera of Tarapacá and Antofagasta regions; Sabrosky (1981: 3), in key to genera of Blondeliini in which females possess an abdominal keel and sharp, curved piercer (key also including the following generic names later synonymised with *Eucelatoria*: *Eucelatoriopsis*, *Heliodexodes*, *Heliolydella*, *Hemilydella*, *Lydellohoughia*, *Machairomasicera*, *Spathimyia*, *Tinalydella*, *Urodexodes* and *Xiphomyia*), synonymy of *Celatoriopsis* with *Eucelatoria*; Cortés (1984: 381, 382), *Eucelatoria*, *Hemilydella* and *Urodexodes* in key to tachinid genera of Tarapacá and Antofagasta regions; Wood (1985: 13, 40), in key to the Blondeliini of North and Central America and the West Indies, new synonymy of all generic names listed above with *Eucelatoria* (with the exception of the previously synonymised *Celatoriopsis*), diagnosis, taxonomic notes.

australis Townsend, 1911.—Neotropical: eastern Lesser Antilles (Saint Vincent), southern Lesser Antilles (Trinidad & Tobago), South America (Brazil, Chile, Peru).

Eucelatoria australis Townsend, 1911: 140, based on female reproductive system [1912b: 315, adult description]. Lectotype female (USNM), by fixation of Townsend (1912b: 316) (description of female “Type” [dissection TD 4025] from Piura in USNM is regarded as a lectotype fixation). Type locality: Peru, Piura.

Note: Aldrich (1927a: 19) synonymised *Compsilura oppugnator* Walton, 1914 from Puerto Rico with *Eucelatoria australis* Townsend and this synonymy was followed by Guimaraes (1971: 133). Sabrosky (1981) recognised *Eucelatoria oppugnator* as valid and Wood (1985: 44) did also but with the note: “[? = *australis* (c/f Aldrich 1927a: 19)]”.

References: Aldrich (1927a: 19), first record from St. Vincent; Sauer (1946: 21), first record from Brazil; Thompson (1968: 200), redescription, first record from Trinidad; Cortés and Campos (1971: 81), first record from Chile; Cortés (1984: 386), taxonomic notes; Vergara de Sánchez (1987: 10), redescription, figures of male and female terminalia.

digitata Sabrosky, 1981.—Neotropical: South America (Chile, Peru).

Eucelatoria digitata Sabrosky, 1981: 11. Holotype male (USNM). Type locality: Peru, Lima, San Diego.

Note: *Eucelatoria digitata* was recorded from both Chile and Peru in the original description.

References: Cortés (1984: 386), taxonomic notes; Vergara de Sánchez (1987: 10, 11), redescription, figures of male and female terminalia.

fasciata (Townsend, 1927).—Neotropical: southern Lesser Antilles (Trinidad & Tobago), South America (Chile, Peru).

Hemilydella fasciata Townsend, 1927a: 315. Holotype male (USNM). Type locality: Peru, Piura, Río Macará, La Tina, on border with Ecuador, 1370 ft.

nudioculata O’Hara & Wood, **nom. nov.**—Not Chile [Trinidad].

Eucelatorioidea nigripalpis Thompson, 1968: 177 (junior secondary homonym of *Chetolyga nigripalpis* Bigot, 1889). Holotype female (CNC). Type locality: Trinidad.

Eucelatoria nudioculata O'Hara & Wood, **nom. nov.** for *Eucelatorioidea nigripalpis* Thompson, 1968.

Note: *Eucelatorioidea nigripalpis* Thompson, 1968 from Trinidad, the type species of *Eucelatorioidea* Thompson, 1968, became a junior secondary homonym of *Chetolyga nigripalpis* Bigot, 1889 from Mexico when transferred to *Eucelatoria* Townsend, 1909 by Wood (1985: 44). Both names were treated as valid in that work and the junior homonym was not renamed “pending a revision of the genus” (Wood 1985: 44). This situation has continued to the present and both names are listed as valid in the most recent version of the checklist of world Tachinidae (O'Hara et al. 2020: 225). In the interests of nomenclatural stability, we hereby propose the new name *Eucelatoria nudioculata* to replace the preoccupied name *Eucelatorioidea nigripalpis* Thompson. The same type material applies to the new name. The specific epithet *nudioculata* refers to the bare eye that was noted by Thompson (1968: 176) as a characteristic of his new genus *Eucelatorioidea*, for which *E. nigripalpis* was designated type species.

oblonga O'Hara & Wood, **nom. nov.**—Neotropical: South America (Chile).

Urodexodes elongatum Cortés & Campos, 1974: 124 (junior secondary homonym of *Exorista elongata* van der Wulp, 1890). Holotype male (MEUC). Type locality: Chile, Arica y Parinacota, Parinacota, Belén, 3500 m.

Eucelatoria oblonga O'Hara & Wood, **nom. nov.** for *Urodexodes elongatum* Cortés & Campos, 1974.

Note: *Urodexodes elongatum* Cortés & Campos, 1974, from Chile, is a junior secondary homonym of *Exorista elongata* van der Wulp, 1890, the valid name of a Costa Rican species of *Eucelatoria* (Wood 1985: 43). The two species names are listed as valid in the most recent version of the checklist of world Tachinidae (O'Hara et al. 2020: 224). In the interests of nomenclatural stability, we hereby propose the new name *Eucelatoria oblonga* to replace the preoccupied name *Urodexodes elongatum* Cortés & Campos. The same type material applies to the new name. The specific epithet *oblonga*, Latin for longer than broad, refers to the elongated appearance of the species.

parkeri (Sabrosky, 1952).—Neotropical: South America (Argentina, Brazil, Chile, Uruguay).

Eucelatoriopsis parkeri Sabrosky, 1952: 325. Holotype male (USNM). Type locality: Uruguay, Montevideo, Montevideo.

References: Cortés (1967b: 12), first record from Chile; Guimarães (1977c: 35), first record from Brazil; Boldt et al. (1991: 843), first record from Argentina.

Genus *EUHALIDAYA* Walton, 1914

EUHALIDAYA Walton, 1914: 130. Type species: *Euhallidaya severinii* Walton, 1914 (= *Biomyia genalis* Coquillett, 1897), by original designation [United States].

OOMEIGENIA Townsend, 1915c: 434 (as “*Oömeigenia*”). Type species: *Oomeigenia chosica* Townsend, 1915, by original designation [Peru].

CLYTHOXYNOPS Townsend, 1927a: 272. Type species: *Clythoxynops orbitalis* Townsend, 1927, by original designation [Brazil].

BACULOCAPTUS Cortés, 1968a: 106. Type species: *Baculocaptus valparadisi* Cortés, 1968, by original designation [Chile].

EUHALLIDAYA. Incorrect original spelling of *Euhallidaya* Walton, 1914 (Walton 1914: 130) (see note).

CLITHOXYNOPS. Incorrect original spelling of *Clythoxynops* Townsend, 1927 (Townsend 1927a: 272).

Notes: The genus name *Euhallidaya* Walton was originally proposed as *Euhallidaya* but subsequent authors (e.g., Curran 1934: 460) changed the spelling to *Euhallidaya*. This changed spelling would normally be considered an incorrect subsequent spelling but because it is in prevailing usage and is attributed to Walton (1914), it is deemed to be the correct original spelling (Article 33.3.1 of the *Code*, ICZN 1999).

There are two original spellings of *Clythoxynops* in Townsend (1927a): *Clithoxynops* (p. 272) and *Clythoxynops* (p. 299). The correct original spelling was selected as *Clythoxynops* by Townsend (1927b, see entry for “page 272, line 17 [from] top” in the unpaginated errata of Townsend 1927a), as the First Reviser (Article 24.2.3 of the *Code*, ICZN 1999).

References: Townsend (1936c: 71, 237), diagnosis of adults and immatures of Acemyini and key to genera (including *Euhallidaya*), diagnosis of adults and immatures of Trypherini and key to genera (including *Clythoxynops* and *Oomeigenia*); Townsend (1939c: 259), redescription of *Euhallidaya*; Townsend (1941: 257, 296), redescrptions of *Clythoxynops* and *Oomeigenia*; Cortés (1976: 3), difference between *Clythoxynops* and *Baculocaptus*; Wood (1985: 14, 45), in key to the Blondeliini of North and Central America and the West Indies, synonymy of *Baculocaptus*, *Clythoxynops* and *Oomeigenia* with *Euhallidaya*, diagnosis, taxonomic notes.

valparadisi (Cortés, 1968).—Neotropical: South America (Chile).

Baculocaptus valparadisi Cortés, 1968a: 108. Holotype male (EEAM). Type locality: Chile, Valparaíso, Valparaíso, Viña del Mar.

Genus *INCAMYIA* Townsend, 1912

INCAMYIA Townsend, 1912b: 317. Type species: *Incamyia cuzcensis* Townsend, 1912, by original designation [Peru].

SPHALLOGLANDULUS Townsend, 1915e: 438. Type species: *Sphalloglandulus unicus* Townsend, 1915, by original designation [Peru].

PROPHRYNOPSIS Townsend, 1927a: 273. Type species: *Prophrynopsis peruviana* Townsend, 1927, by original designation [Peru].

References: Aldrich (1928b: 14), synonymy of *Sphalloglandulus* with *Incamyia*, key to four species; Aldrich (1934: 4, 65), in key to Patagonian genera, *Sphalloglandulus* in synonymy, taxonomic notes, key to three Patagonian species; Townsend (1936c: 86, 190, 282), diagnosis of adults and immatures of Compsilurini and key to genera (including *Incamyia*), diagnosis of adults and immatures of Sturmiini and key to genera (including *Prophrynopsis*), synonymy; Townsend (1940a: 560), redescription of *Incamyia* (with *Sphalloglandulus* in synonymy); Townsend (1941: 124), redescription of *Prophrynopsis*; Parker (1953: 68), figures of

first instar larva and puparium; Cortés (1968b: 18), key to four species; Cortés and Campos (1971: 25, 87), in key to tachinid genera of Tarapacá and Antofagasta regions, key to six species of these provinces; Guimarães (1971: 136), synonymy of *Prophrynopsis* with *Incamyia*; Cortés and Campos (1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions; Sabrosky (1981: 3), in key to genera of Blondeliini in which females possess an abdominal keel and sharp, curved piercer; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Henry (1992: 36), key to the ten Chilean species.

charlini Cortés, 1968.—Neotropical: South America (Chile).

Incamyia charlini Cortés, 1968b: 19. Holotype male (EEAM). Type locality: Chile, Metropolitana de Santiago, Santiago, Maipú, Rinconada.

Reference: Cortés (1986: 158), taxonomic notes.

chilensis Aldrich, 1928.—Neotropical: South America (Argentina, Chile, Uruguay).

Incamyia chilensis Aldrich, 1928b: 16. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

References: Aldrich (1934: 66), redescription, first record from Argentina; Parker et al. (1951 [pages unknown], also 1953: 55, 58, 68), first record from Uruguay; Cortés (1952: 109), first record from Juan Fernández Islands with note on the possible subspecific status of the island population; Caltagirone (1953: 90, 92), description and figure of first instar larva; Blanchard (1963: 203), redescription, wing figure; Molina-Ochoa et al. (2003: 259), record from Brazil attributed to Guimarães (1977c) but *Incamyia chilensis* not recorded from Brazil in that work.

cinerea Cortés & Campos, 1971.—Neotropical: South America (Chile).

Incamyia cinerea Cortés & Campos, 1971: 88. Holotype male (EEAM). Type locality: Chile, Tarapacá, Tamarugal, Mamiña, 2600 m (20°06'S, 69°16'W) (coordinates and elevation given on p. 11).

cuzcensis Townsend, 1912.—Neotropical: South America (Chile, Peru).

Incamyia cuzcensis Townsend, 1912b: 317. Holotype female (USNM). Type locality: Peru, Cusco [region or city, as “Cuzco”].

cuzcoensis. Incorrect subsequent spelling of *cuzcensis* Townsend, 1912 (Vimmer and Soukup 1940b: 361).

References: Aldrich (1928b: 14), synonymy of *Sphalloglandulus unicus* Townsend, 1915 from Peru with *Incamyia cuzcensis*, but this synonymy overlooked or not followed by later authors (except Aldrich 1934: 65); Cortés and Campos (1971: 89), first record from Chile.

nuda Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Incamyia nuda Aldrich, 1934: 66. Syntypes, 6 males (USNM). Type locality: Argentina, Río Negro, Lago Nahuel Huapi, San Carlos de Bariloche [as “Bariloche”].

Reference: González and Henry (1992: 36), first record from Chile.

perezi Cortés & Campos, 1971.—Neotropical: South America (Chile).

Incamyia perezi Cortés & Campos, 1971: 89. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Parinacota, Putre, 3530 m (18°12'S, 69°35'W) (coordinates and elevation given on p. 11).

picta Cortés, 1976.—Neotropical: South America (Chile).

Incamyia picta Cortés, 1976: 5. Holotype male (MEUC). Type locality: Chile, Coquimbo, Elqui, Baños El Toro, 3300–4000 m [ca. 29°50'S, 70°1'W].

sandovali Cortés & Campos, 1971.—Neotropical: South America (Chile).

Incamyia sandovali Cortés & Campos, 1971: 90. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Parinacota, Putre, 3530 m (18°12'S, 69°35'W) (coordinates and elevation given on p. 11).

spinicosta Aldrich, 1928.—Neotropical: South America (Argentina, Chile).

Incamyia spinicosta Aldrich, 1928b: 15. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

Reference: Aldrich (1934: 67), first description of female, first record from Argentina.

striata Aldrich, 1928.—Neotropical: South America (Chile, Peru).

Incamyia striata Aldrich, 1928b: 16. Holotype male (USNM). Type locality: Peru, Junín, La Oroya.

Reference: Cortés and Campos (1971: 91), first record from Chile.

Genus *MYIOPHARUS* Brauer & Bergenstamm, 1889

MYIOPHARUS Brauer & Bergenstamm, 1889: 161 [also 1890: 93]. Type species:

Myiopharus metopia Brauer & Bergenstamm, 1889, by monotypy [Mexico].

DIDYMA van der Wulp, 1890a: 43, in key [1890e: 156, description]. Type species:

Didyma albomicans van der Wulp, 1890, by subsequent designation of Townsend in Williston (1908: 379, as “*albomicans*”) [Mexico].

PARALISPE Brauer & Bergenstamm, 1891: 337 [also 1891: 33]. Type species: *Paralipse brasiliensis* Brauer & Bergenstamm, 1891, by monotypy [Brazil].

PARADORIA Brauer & Bergenstamm, 1891: 339 [also 1891: 35]. Type species: *Paradoria nigra* Brauer & Bergenstamm, 1891, by monotypy [Venezuela].

MESOCHAETA Brauer & Bergenstamm, 1891: 341 [also 1891: 37]. Type species: *Didyma commixta* van der Wulp, 1890 (= *Phorocera barbata* Bigot, 1889; *commixta* cited as “*connexa*” by Brauer & Bergenstamm 1891: 341, in error), by monotypy [Mexico].

METADORIA Brauer & Bergenstamm, 1893: 29 [also 1893: 117]. Type species:

Metadoria mexicana Brauer & Bergenstamm, 1893 (= *Phorocera barbata* Bigot, 1889), by monotypy [Mexico].

HEMIARGYRA Townsend, 1908: 88. Type species: *Hemiargyra nigra* Townsend, 1908

(junior secondary homonym of *Paradoria nigra* Brauer & Bergenstamm, 1891; = *Phorocera nigrita* van der Wulp, 1890), by original designation [Costa Rica].

MUSCINOTHELAIIRA Townsend, 1916e: 310. Type species: *Muscinothelaira lutzi*

Townsend, 1916, by original designation [Brazil].

AUSTROLYDELLA Townsend, 1919b: 573. Type species: *Austrolydella assimilis*

Townsend, 1919, by original designation [Peru].

GYMNODORIA Townsend, 1927a: 260. Type species: *Gymnodoria capitata* Townsend,

1927, by original designation [Peru].

EUHEMIARGYRA Townsend, 1927a: 260. Type species: *Euhemiargyra parva*

Townsend, 1927, by original designation [Brazil].

- HEMIARGYROPSIS* Townsend, 1927a: 260. Type species: *Hemiargyropsis frontalis* Townsend, 1927, by original designation [Peru].
- DACTYLODIDYMA* Townsend, 1927a: 260. Type species: *Dactylodidyma dubia* Townsend, 1927, by original designation [Brazil].
- THELYPHAENOPSIS* Townsend, 1927a: 262. Type species: *Thelyphaenopsis atra* Townsend, 1927, by original designation [Brazil].
- BOLODORIA* Townsend, 1927a: 262. Type species: *Bolodoria yahuarmaryana* Townsend, 1927, by original designation [Peru].
- DIDYMOPS* Townsend, 1927a: 262 (junior homonym of *Didymops* Rambur, 1842 and *Didymops* Szilády, 1922). Type species: *Didymops yahuarmaryensis* Townsend, 1927, by original designation [Peru].
- MAYOPHORINIA* Townsend, 1927a: 263. Type species: *Mayophorinia angusta* Townsend, 1927, by original designation (see note) [Peru].
- ARGYRODORIA* Townsend, 1927a: 265. Type species: *Argyrodoria hemiargyroides* Townsend, 1927, by original designation [Brazil].
- NEARGYROPHYLAX* Townsend, 1927a: 265. Type species: *Neargyrophylax argentescens* Townsend, 1927, by original designation [Brazil].
- HEMIARGYROPHYLAX* Townsend, 1927a: 265. Type species: *Hemiargyrophylax punctilucis* Townsend, 1927, by original designation [Peru].
- OXYNOPSIS* Townsend, 1927a: 270. Type species: *Oxynopsis brasiliensis* Townsend, 1927, by original designation [Brazil].
- MYIOXYNOPS* Townsend, 1927a: 278. Type species: *Myioxynops palpalis* Townsend, 1927, by original designation [Peru].
- HYPOPHORINIA* Townsend, 1927a: 279. Type species: *Hypophorinia hyphena* Townsend, 1927, by original designation [Brazil].
- METARRHINOMYIA* Townsend, 1927a: 279. Type species: *Metarrhinomyia angusta* Townsend, 1927 (junior secondary homonym of *Mayophorinia angusta* Townsend, 1927; = *Myiopharus charapensis* O'Hara & Wood, **nom. nov.**, see below), by original designation (see note) [Peru].
- MELANODORIA* Townsend, 1927a: 280. Type species: *Melanodoria nigrisquamis* Townsend, 1927, by original designation [Peru].
- NEOXYNOPS* Townsend, 1934b: 403. Type species: *Neoxynops nana* Townsend, 1934, by original designation [Brazil].
- OXYNOPSALIA* Curran, 1934: 467. Type species: *Oxynopsalia nitida* Curran, 1934, by original designation [Panama].
- ANOXYNOPSELLA* Townsend, 1935: 226. Type species: *Anoxynopsella argentescens* Townsend, 1935 (junior secondary homonym of *Neargyrophylax argentescens* Townsend, 1927; = *Myiopharus argentata* Nihei & Dios, 2016), by original designation [Brazil].
- NEOXYNOPSOIDEA* Thompson, 1968: 149. Type species: *Neoxynopsoidea claripalpis* Thompson, 1968, by original designation [Trinidad & Tobago].
- STENOCHAETA* Thompson, 1968: 159. Type species: *Stenochaeta claripalpis* Thompson, 1968 (junior secondary homonym of *Neoxynopsoidea claripalpis* Thompson,

1968; = *Myiopharus incognitus* O'Hara & Wood, **nom. nov.**, see below), by original designation [Trinidad & Tobago].

NEOARGYROPHYLAX. Incorrect subsequent spelling of *Neargyrophylax* Townsend, 1927 (Guimarães 1971: 142, 297; Toma and Nihei 2006: 242, 249).

References: Coquillett (1910: 533, 550, 568, 572), type species of *Didyma*, *Hemiargyra*, *Metadoria* and *Myiopharus* (with *Hemiargyra* in synonymy with *Metadoria*); Aldrich (1924: 216), synonymy of *Hemiargyra* with *Myiopharus*; Aldrich (1934: 4, 62), in key to Patagonian genera, synonymy (*Hemiargyra* with *Myiopharus*), taxonomic notes; Townsend (1936c: 78, 82, 86, 204, 237, 273, 275), diagnosis of adults and immatures of Anacamptomyiini and key to genera (including *Gymnodoria*), diagnosis of adults and immatures of Elodiini and key to genera (including *Metarrhinomyia*), diagnosis of adults and immatures of Compsilurini and key to genera (including *Anoxynopsella*), diagnosis of adults and immatures of Carceliini and key to genera (including *Myioxynops*), diagnosis of adults and immatures of Trypherini and key to genera (including *Argyrodoria*, *Austrolydella*, *Bolodoria*, *Didyma*, *Didymops*, *Hemiargyra*, *Hemiargyrochylax*, *Hemiargyropsis*, *Hypophorinia*, *Mayophorinia*, *Melanodoria*, *Mesochaeta*, *Metadoria*, *Muscinothelaira*, *Myiopharus*, *Neargyrophylax*, *Neoxynops*, *Oxynopsalia*, *Oxynopsis*, *Paradoria*, *Paralispe* and *Thelyphaenopsis*), *Dactylodidyma* and *Euhemiargyra* as synonyms of *Paradoria*; Townsend (1940a: 9, 21, 32), redescrptions of *Gymnodoria*, *Metarrhinomyia* and *Anoxynopsella*; Townsend (1941: 154, 239–322), redescrptions of *Myioxynops* and the aforementioned genera of Trypherini (with *Dactylodidyma* and *Euhemiargyra* in synonymy with *Paradoria*); Guimarães (1971: 141), *Hemiargyra* in synonymy with *Myiopharus* (following Aldrich 1924, 1934, not Townsend 1941); Wood (1985: 13, 14, 15, 60), in key to the Blondeliini of North and Central America and the West Indies, synonymy (including many of the names above as new generic synonyms), diagnosis, taxonomic notes.

charapensis O'Hara & Wood, **nom. nov.**—Not Chile [Peru].

Metarrhinomyia angusta Townsend, 1927a: 329 (junior secondary homonym of *Mayophorinia angusta* Townsend, 1927, by First Reviser action below). Holotype female (USNM). Type locality: Peru, Cajamarca, Río Charapi [as “Rio Charape”, ca. 5°25'S, 78°59'W].

Myiopharus charapensis O'Hara & Wood, **nom. nov.** for *Metarrhinomyia angusta* Townsend, 1927.

Note: *Mayophorinia angusta* Townsend, 1927 (type species of *Mayophorinia*) and *Metarrhinomyia angusta* Townsend, 1927 (type species of *Metarrhinomyia*), both from Peru, were described in the same publication (Townsend 1927a: 326, 329) and became secondary homonyms when the generic names were synonymised with *Myiopharus* by Wood (1985: 61, 62). The two species names are listed as valid in the most recent version of the checklist of world Tachinidae (O'Hara et al. 2020: 257). As the First Reviser (Article 24.2.2 of the Code, ICZN 1999), we hereby fix *Mayophorinia angusta* as the senior homonym. In the interests of nomenclatural stability, we propose the new name *Myiopharus charapensis* to replace the name of the junior homonym *Metarrhinomyia angusta*. The same type material applies to the new name. The specific epithet *charapensis* is based on the type locality of Río Charapi.

incognitus O'Hara & Wood, **nom. nov.**—Not Chile [Trinidad].

Stenochaeta claripalpis Thompson, 1968: 159 (junior secondary homonym of *Neoxynopsoidea claripalpis* Thompson 1968). Holotype male (CNC). Type locality: Trinidad, “Legerville Mt.” [not located].

Myiopharus incognitus O'Hara & Wood, **nom. nov.** for *Stenochaeta claripalpis* Thompson, 1968.

Note: *Neoxynopsoidea* and *Stenochaeta* were described from Trinidad in the same work by Thompson (1968), along with their type species *Neoxynopsoidea claripalpis* and *Stenochaeta claripalpis*. The names of the two type species became secondary homonyms when the generic names were transferred to *Myiopharus* Brauer & Bergenstamm by Wood (1985: 62) and both were listed under “Included species” of *Myiopharus* (p. 64). The relative priority of *Neoxynopsoidea claripalpis* and *Stenochaeta claripalpis*, when both are placed in *Myiopharus*, was established by Wood (1985: 64) as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999) when he noted under the latter: “Although a secondary homonym of *claripalpis* (Thompson) 1968: 149, this species is not renamed, pending a revision of the genus”. This situation has continued to the present and both names are listed as valid in the most recent version of the checklist of world Tachinidae (O'Hara et al. 2020: 258). In the interests of nomenclatural stability, we hereby propose the new name *Myiopharus incognitus* to replace the name of the junior homonym *Stenochaeta claripalpis*. The same type material applies to the new name. The specific epithet *incognitus* was inspired by the type locality of “Legerville Mt.” that we have been unable to locate.

pirioni Aldrich, 1934.—Neotropical: South America (Chile).

Myiopharus pirioni Aldrich, 1934: 64. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

Note: Marshall et al. (2008) observed a female of *Myiopharus pirioni* Aldrich feeding on the regurgitate of a leaf beetle larva (*Procalus* Clark, Chrysomelidae).

rufopalpus O'Hara & Wood, **nom. nov.**—Not Chile [Brazil].

Paralispe palpalis Townsend, 1929: 376 (junior secondary homonym of *Myioxynops palpalis* Townsend, 1927). Holotype female (USNM). Type locality: Brazil, São Paulo, Itaquaquecetuba.

Myiopharus rufopalpus O'Hara & Wood, **nom. nov.** for *Paralispe palpalis* Townsend, 1929.

Note: *Myioxynops palpalis* Townsend, 1927 (type species of *Myioxynops* Townsend, 1927) from Peru and *Paralispe palpalis* Townsend, 1929 from Brazil became secondary homonyms when the genera to which they belonged, *Myioxynops* Townsend, 1927 and *Paralispe* Brauer & Bergenstamm, 1891, were synonymised with *Myiopharus* Brauer & Bergenstamm, 1889 by Wood (1985: 60, 62). The two species names are listed as valid in the most recent version of the checklist of world Tachinidae (O'Hara et al. 2020: 260). In the interests of nomenclatural stability, we hereby propose the new name *Myiopharus rufopalpus* to replace the name of the junior homonym *Paralispe palpalis*. The same type material applies to the new name. The specific epithet *rufopalpus* refers to the colour of the palpus, described by Townsend (1929: 376) as “light rufous on swollen portion”.

subaeneus Aldrich, 1934.—Neotropical: South America (Chile).

Myiopharus subaeneus Aldrich, 1934: 63. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

Genus **NOTOMANES** Aldrich, 1934

NOTOMANES Aldrich, 1934: 93. Type species: *Tachina maura* Walker, 1836 (= *Tachina basalis* Walker, 1836), by original designation [Chile].

Reference: Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions.

basalis (Walker, 1836).—Neotropical: South America (Chile).

Tachina basalis Walker, 1836: 351. Lectotype female (NHMUK), by fixation of Aldrich (1934: 94) (examination of “type female” from Port Famine in NHMUK is regarded as a lectotype fixation). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Puerto del Hambre [as “Port Famine”].

Tachina maura Walker, 1836: 352. Lectotype male (NHMUK), by fixation of Aldrich (1934: 94) (examination of “type male” from Port Famine in NHMUK is regarded as a lectotype fixation). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Puerto del Hambre [as “Port Famine”].

Note: The relative priority of *Tachina basalis* Walker, 1836 and *Tachina maura* Walker, 1836, when the two are treated as synonyms, was established by Austen (1907: 330), as the First Reviser (Article 24.2.2 of the Code, ICZN 1999). Aldrich (1934: 93), Cortés (1963: 243, 1986: 158) and Cortés and Hichins (1969: 66) treated *maura* as the valid name but Guimaráes (1971: 99) gave priority to *basalis* based on the First Reviser action of Austen (1907). Cortés (1973a: 99) argued that *maura* should have priority because it was chosen over *basalis* as the type species of *Notomanes* Aldrich, 1934 and was based on a male type in better condition than the female type of *basalis*, but the nomenclatural action of the First Reviser cannot be set aside on these grounds. Cortés (1986: 158) continued use of the name *Notomanes maura*. References: Aldrich (1934: 93), taxonomic notes; Cortés (1963: 243), notes on name-bearing types of *Tachina basalis* and *Tachina maura* in NHMUK; Cortés (1973a: 99), comparison of recent Chilean specimens with the redescription of Aldrich (1934: 93, as *Notomanes maura*).

Genus **PHASMOPHAGA** Townsend, 1909

PHASMOPHAGA Townsend, 1909: 243. Type species: *Phasmophaga antennalis* Townsend, 1909, by original designation [United States].

PHASMOVORA Cortés, 1968a: 102. Type species: *Phasmovora phasmophagae* Cortés, 1968, by original designation [Chile].

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Phasmophaga*); Townsend (1940a: 246), redescription of *Phasmophaga*; Cortés (1976: 4), difference between *Gilvella* Mesnil, 1960 (a synonym of *Anisia* van der Wulp, 1890) and *Phasmovora*; Wood (1985: 14, 72), in key to the Blondeliini of North and Central America and the West Indies, synonymy of *Phasmovora* with *Phasmophaga*, diagnosis, taxonomic notes.

phasmophagae (Cortés, 1968).—Neotropical: South America (Chile).

Phasmovora phasmophagae Cortés, 1968a: 105. Holotype male (EEAM). Type locality: Chile, Maule, Curicó, Cajón del Río Claro, 15 km east of Los Queñes, 900 m.

Genus **STELEONEURA** Stein, 1924

STELEONEURA Stein, 1924: 151. Type species: *Steleoneura czernyi* Stein, 1924, by monotypy [Spain].

EMBIOMYIA Aldrich, 1934: 29. Type species: *Embiomyia australis* Aldrich, 1934, by original designation [Argentina]. **Syn. nov.**

Notes: *Steleoneura* was most recently characterised by Wood (1985: 80) and was included in the keys to tachinid genera of America north of Mexico (Wood 1987: 110) and Central America (Wood and Zumbado 2010: 1380). A similarity between *Steleoneura* and Chilean *Embiomyia* (and South African genus *Pararondania* Villeneuve, 1916) was noted by Wood (1985: 81) based on the shared possession of “medially separated antennae, elongate pedicel, short first flagellomere and bulbous-based arista, long straight prosternal setae, absence of lateral scutellar bristles, 2 postpronotal bristles, and vein M ending in R_{4+5} ”. Wood and Zumbado (2010: 1412) further noted that *Embiomyia* “may be congeneric with *Steleoneura*”. These previous authors were reluctant to synonymise *Embiomyia* with *Steleoneura* because they had not seen a female of the single known species, *E. australis*. The female abdomen in *Steleoneura* is distinctive: “globular, ovipositor telescopic, extending ventrally from apex of abdomen (Fig. 50)” (Wood 1985: 81). Aldrich (1934: 31) had simply described the shape of the female abdomen as “bluntly pointed”. We have examined three females of *E. australis* in CNC collected in Chile by JEOH in 2015 (and unidentified until after the trip report published by Stireman et al. 2016) and they possess the same peculiar ventrally-directed ovipositor that characterises *Steleoneura* species. Based on this finding and the other similarities between *Embiomyia* and *Steleoneura* noted above [but not “medially separated antennae”], we here synonymise the two generic names. *Steleoneura* is a genus with unusual features for a blondeline and it could be misplaced here.

References: (*Steleoneura* Stein, 1924 is inexplicably missing from Townsend’s comprehensive *Manual of Myiology*); Townsend (1936c: 106), diagnosis of adults and immatures of Phoroceratini and key to genera (including *Embiomyia*); Townsend (1940a: 121), redescription of *Embiomyia*.

australis (Aldrich, 1934).—Neotropical: South America (Argentina, Chile). **Comb. nov.** (Fig. 4e)

Embiomyia australis Aldrich, 1934: 30. Holotype male (NHMUK). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapi.

Note: *Embiomyia australis* was recorded from both Argentina and Chile in the original description.

References: Wood (1985: 81, 86) and Wood and Zumbado (2010: 1412), taxonomic notes.

Tribe ERYCIINI

Genus *CARCELIA* Robineau-Desvoidy, 1830

CARCELIA Robineau-Desvoidy, 1830: 176. Type species: *Carcelia bombylans* Robineau-Desvoidy, 1830, by subsequent designation of Coquillett (1910: 518) (see Evenhuis et al. 2010: 52) [France].

References: Coquillett (1910: 518), type species (given as “*bombylans* ... by designation of Desvoidy ... vol. 1, 1863, p. 220”); Townsend (1936c: 204), diagnosis of adults and immatures of Carceliini and key to genera (including *Carcelia*); Townsend (1941: 143), redescription of *Carcelia*; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions.

halliana Cortés, 1945.—Neotropical: South America (Argentina, Chile).

Carcelia halliana Cortés, 1945c: 27. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Note: This species has not been assigned to a *Carcelia* subgenus.

Reference: Gramajo (1998: 97), first record from Argentina.

Genus *DRINO* Robineau-Desvoidy, 1863

DRINO Robineau-Desvoidy, 1863a: 250. Type species: *Drino volucris* Robineau-Desvoidy, 1863 (= *Tachina lota* Meigen, 1824), by original designation [France].

Note: The more notable diagnostic features of *Drino* within the Erycini are the short or absent ocellar setae, bare parafacial, facial ridge bare above lowest third, postpronotal setae more or less in line, four katapisternal setae, single setula at the base of wing vein R_{4+5} , and row of even and closely spaced anterodorsal setae on the hind tibia (Wood 1987: 1215; Wood and Zumbado 2010: 1366). The two species moved here to *Drino* share these characteristics.

References: Aldrich (1934: 5, 138), in key to Patagonian genera, taxonomic notes (as *Sturmia* Robineau-Desvoidy, 1830); Cortés (1944g: 161), key to Chilean species (as *Sturmia*); Thompson (1966: 391, 393), key to Trinidad species (as *Drino*), taxonomic notes; Cortés and Campos (1974: 115) and Cortés (1984: 381), in keys to tachinid genera of Tarapacá and Antofagasta regions (as *Sturmia*); ICZN (2012: 242), ruling to conserve current usage of generic names *Sturmia* and *Drino*.

festiva (Cortés, 1944).—Neotropical: South America (Argentina, Chile). **Comb. nov.**

Sturmia festiva Cortés, 1944g: 163. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales, prov. de Valparaíso”, ca. 33°9'S, 71°18'W].

Note: *Sturmia festiva* was treated as a species of *Sturmia* Robineau-Desvoidy, 1830 by previous authors (e.g., Guimaraes 1971: 192; Cortés and Hichins 1979: 115; Liljesthöm 1980: 135; Henry 1987: 200). It is moved here to *Drino* Robineau-Desvoidy and resembles the widespread *Drino rhoeo* (Walker, 1849) in possessing a bright yellow abdominal tergite 5 that contrasts with the gray pruinose colour of the previous segments. *Drino rhoeo*, if correctly

identified throughout its range, occurs from Canada (O'Hara and Wood 2004: 120) to Argentina (Blanchard and De Santis 1975: 34), including Costa Rica (Smith et al. 2007: 4968). References: Campos (1953: 25), first description of female; Cortés and Hichins (1979: 115), redescription of female; Liljesthröm (1980: 135), first record from Argentina.

insignis (van der Wulp, 1882).—Neotropical: South America (Argentina, Chile).

Comb. nov.

Masicera insignis van der Wulp, 1882: 85. Syntypes, 1 male and 1 unknown (abdomen missing at time of description) (RMNH). Type locality: Chile.

Note: *Masicera insignis* was treated as a species of *Sturmia* Robineau-Desvoidy, 1830 by previous authors (e.g., Cortés and Hichins 1969: 59; Guimarães 1971: 192; Henry 1987: 200) but is moved here to *Drino* Robineau-Desvoidy (see note under genus for generic characters).

Reference: Cortés (1944g: 166), redescription, first record from Argentina.

piceiventris (Walker, 1836).—Neotropical: South America (Chile).

Tachina piceiventris Walker, 1836: 350. Lectotype female (NHMUK), by fixation of Aldrich (1934: 139) (examination of female “type” in NHMUK is regarded as a lectotype fixation). Type locality: not given; somewhere along the South American coast “from St. Paul’s [São Paulo, see Thompson (1974: 2)] in Brazil to Valparaíso [Chile]” according to Curtis (1836: 315), here interpreted as Chile based on the known distribution of the species.

Note: *Tachina piceiventris* was treated as a species of *Sturmia* Robineau-Desvoidy, 1830 by most previous authors (e.g., Aldrich 1934: 139; Cortés 1944g: 161; Cortés 1963: 242; Cortés and Hichins 1969: 59; Henry 1987: 200) but was moved to *Drino* Robineau-Desvoidy by Guimarães (1971: 189).

References: Aldrich (1934: 139), redescription; Cortés (1944g: 161), redescription; Campos (1953: 24), partial redescription; Cortés (1963: 242), notes on name-bearing type in NHMUK.

Genus **LESPESIA** Robineau-Desvoidy, 1863

LESPESIA Robineau-Desvoidy, 1863a: 567. Type species: *Achaetoneura anisotae* Webber, 1930, by designation under the Plenary Powers of ICZN (1983: 97) [United States].

ACHAETONEURA Brauer & Bergenstamm, 1891: 334 [also 1891: 30]. Type species: *Achaetoneura hesperus* Brauer & Bergenstamm, 1891 (= *Masicera frenchii* Williston, 1889), by subsequent designation of Townsend (1908: 88) [North America].

PARAFRONTINA Brauer & Bergenstamm, 1893: 27 [also 1893: 115]. Type species: *Parafrontina apicalis* Brauer & Bergenstamm, 1893 (= *Tachina archippivora* Riley, 1871), by monotypy [United States].

ZYGOFRONTINA Townsend, 1915e: 427. Type species: *Zygofrontina capitis* Townsend, 1915 (= *Tachina archippivora* Riley, 1871), by original designation [Peru].

MASICEROPSIS Townsend, 1916c: 178. Type species: *Masicera pauciseta* Coquillett, 1897 (= *Tachina archippivora* Riley, 1871), by original designation [United States].

YPOPHAEMYIA Townsend, 1916d: 75. Type species: *Ypophaeomyia malacosomae* Townsend, 1916 (= *Tachina archippivora* Riley, 1871), by original designation [United States].

- EUPARAFRONTINA* Brèthes, 1917: 17. Type species: *Euparafrontina martinezi* Brèthes, 1917, by monotypy [Peru].
- PROPHRYNO* Townsend, 1927a: 262. Type species: *Prophryno aurulans* Townsend, 1927 (= *Tachina lata* Wiedemann, 1830), by original designation [Brazil].
- ACHAETONEUOPSIS* Townsend, 1927a: 272. Type species: *Achaetoneuopsis affinis* Townsend, 1927, by original designation [Brazil].
- MYIOSTURMIA* Townsend, 1927a: 272. Type species: *Myiosturmia mixta* Townsend, 1927, by original designation [Brazil].
- ZYGOFRONTINOPSIS* Blanchard, 1959: 173. Type species: *Zygofrontinopsis williamsoni* Blanchard, 1959, by original designation [Argentina].
- STURMIOPSOIDEA* Thompson, 1966: 359. Type species: *Sturmiopsoidea obscura* Thompson, 1966 (junior secondary homonym of *Eurigaster obscurus* Bigot, 1857; = *Lespesia thompsoni* O'Hara & Wood, **nom. nov.**, see below), by monotypy [Trinidad & Tobago]. **Syn. nov.**
- ACHATONEURA*. Incorrect subsequent spelling of *Achaetoneura* Brauer & Bergengstamm, 1891 (Townsend 1927a: 268 [not p. 230 as cited by Evenhuis et al. 2015: 38], subsequently corrected to *Achaetoneura* in Townsend 1927b, see entry for "page 268, line 7 [from] bottom" in the unpaginated errata of Townsend 1927a).
- ZYGOFRONTINOPSIS*. Incorrect subsequent spelling of *Zygofrontinopsis* Blanchard, 1959 (Guimarães 1983: 14, etc.; Toma 2010: 166; Nihei 2016: 929).

Note: The new synonymy of *Sturmiopsoidea* with *Lespesia* is explained below under *Lespesia thompsoni* O'Hara & Wood, which is a new replacement name for type species *Sturmiopsoidea obscura* Thompson, 1966 (a junior homonym of *Eurigaster obscurus* Bigot, 1857 when the two names are placed together in *Lespesia*).

References: Coquillett (1910: 502, 545), type species of *Achaetoneura* and *Parafrontina* (both as synonyms of *Frontina* Meigen, 1838); Webber (1930: 1), synonymy of *Masiceropsis*, *Parafrontina* and *Ypophamyia* with *Achaetoneura*, revision of North American species; Aldrich (1934: 4, 91), in key to Patagonian genera, synonymy of *Achaetoneuopsis*, *Euparafrontina* and *Zygofrontina* with *Achaetoneura*, taxonomic notes; Townsend (1936c: 190, 218, 227, 277), diagnosis of adults and immatures of *Sturmiini* and key to genera (including *Myiosturmia*), diagnosis of adults and immatures of *Lydellini* and key to genera (including *Masiceropsis*); diagnosis of adults and immatures of *Phrynoini* and key to genera (including *Achaetoneura*, *Achaetoneuopsis*, *Euparafrontina*, *Parafrontina*, *Prophryno*, *Ypophamyia* and *Zygofrontina*), *Lespesia* as synonym of *Istocheta* Rondani, 1859; Townsend (1941: 109, 192, 203–227), redescrptions of *Myiosturmia*, *Masiceropsis*, and the aforementioned genera of *Phrynoini*; Mesnil (1950: 109), synonymy of *Achaetoneura* and *Prophryno* with *Lespesia*; Beneway (1963), revision of North American species, synonymy following non-Townsend authors; Thompson (1966: 371), revision of three Trinidad species, taxonomic notes; Cortés and Campos (1971: 25, 1974: 115) and Cortés (1984: 381), in keys to tachinid genera of Tarapacá and Antofagasta regions; Sabrosky (1980: 65), revised key to Nearctic species; Guimarães (1983: 12), synonymy of *Myiosturmia* and *Zygofrontinopsis* with *Lespesia*, revision of Brazilian species; Toma (2010), revision of Venezuelan species.

archippivora (Riley, 1871).—Not Chile [widespread throughout the Nearctic Region and most of Middle and South America].

Tachina archippivora Riley, 1871: 150 (name not authored by Williston as cited by some early authors).

Note: *Lespesia archippivora* was recorded from Chile by Molina-Ochoa et al. (2003: 262), citing Etcheverry (1957) as the source. There is no mention of *L. archippivora* in Etcheverry (1957) and the Chilean record of this species in Molina-Ochoa et al. (2003) is assumed to be in error.

auriceps (Macquart, 1844).—Neotropical: South America. Distribution not known beyond the imprecise type locality of Brazil or Chile.

Masicera auriceps Macquart, 1844: 59 [also 1844: 216]. Lectotype male (MNHN), by designation herein (see Lectotype Designations section). Type locality: Brazil or Chile.

References: Guimarães (1971: 209), as recognised species of *Lespesia*; Guimarães (1983: 23), as unrecognised species of *Lespesia*.

modesta (Bigot, 1857).—Not Chile [Cuba]. **Comb. nov.**

Eurigaster modestus Bigot, 1857b: 341. Type(s), unspecified sex (2 syntypes in MNHN, see note). Type locality: Cuba.

Note: The online MNHN database records two syntypes in the Guérin-Meneville in Macquart collection for *Eurigaster modestus*. One is a female based on the presence of proclinate orbital setae on the head (number MNHN-ED-ED10017) and the other is of undetermined sex (number MNHN-ED-ED10018, head missing). Bigot (1857b: 341) did not specify the sex of the name-bearing type but female is suggested by his comment under *Eurigaster obscurus* that this might be the male of *Eurigaster modestus*. The syntypes of *E. modestus* were examined by DMW and determined to belong to *Lespesia*.

Reference: Guimarães (1971: 215), as unplaced species of Exoristinae (as “Goniinae”).

obscura (Bigot, 1857).—Not Chile [Cuba]. **Comb. nov.**

Eurigaster obscurus Bigot, 1857b: 341. Type(s), male (1 male in MNHN, see note). Type locality: Cuba.

Note: The online MNHN database records a male holotype in the Guérin-Meneville in Macquart collection for *Eurigaster obscurus* (number MNHN-ED-ED10015) based on a holotype determination label that Paul Arnaud, Jr. attached to the specimen in 1972. However, Bigot did not restrict the name-bearing type to a single specimen and the “holotype” in MNHN is technically a syntype [see Recommendation 73F of the *Code* (ICZN 1999), “Avoidance of assumption of holotype”]. This specimen was examined by DMW and determined to be a species of *Lespesia* (see characters of the genus under *L. thompsoni*) This new combination is recorded here because the name is currently valid and a senior secondary homonym of *Sturmiopsoidea obscura* Thompson, 1966.

Reference: Guimarães (1971: 215), as unplaced species of Exoristinae (as “Goniinae”).

thompsoni O'Hara & Wood, **nom. nov.**—Not Chile [Trinidad].

Sturmiopsoidea obscura Thompson, 1966: 359 (junior secondary homonym of *Eurigaster obscurus* Bigot, 1857). Holotype male (CNC). Type locality: Trinidad, North Coast Road [as “American Road”], “Mauvan Hill” [not located]. **Comb. nov.**

Lespesia thompsoni O'Hara & Wood, **nom. nov.** for *Sturmiopsoidea obscura* Thompson, 1966.

Notes: *Sturmiopsoidea obscura* Thompson, 1966, from Trinidad, is a junior secondary homonym of *Eurigaster obscurus* Bigot, 1857, the valid name of a Cuban species that we transfer above to *Lespesia*. In the interests of nomenclatural stability, we hereby propose the new name *Lespesia thompsoni* to replace the name of the junior homonym *Sturmiopsoidea obscura*. The same type material applies to the new name. The specific epithet *thompsoni* is based on the surname of the describer of *S. obscura*, W.R. Thompson.

We have examined the holotype of *S. obscura* in CNC and it has the usual characteristics of Erycini and *Lespesia*, and runs to *Lespesia* in the keys of Wood (1987: 1211) and Wood and Zumbado (2010: 1363). Among the more diagnostic features of *Lespesia* are a setose facial ridge and four katepisternal setae. The eye can be haired or bare (haired in *S. obscura*) and this character splits *Lespesia* into two exit points in both of the aforementioned keys. Thompson (1966: 355) restricted *Lespesia* to species with a bare eye and an otherwise similar species (*obscura*) with a haired eye was assigned to new genus *Sturmiopsoidea*.

Reference: Guimaráes (1971: 192), as *Sturmiopsoidea obscura*.

Nomen dubium of *LESPESIA* Robineau-Desvoidy, 1863

andina (Bigot, 1888).—Not Chile [Cuba]. **Comb. nov.**

Blepharipeza andina Bigot, 1888b: 90. Holotype male (NHMUK). Type locality: Cuba (as “Chili” in error, see note).

Note: The holotype of *Blepharipeza andina* in NHMUK was examined by DMW. It is a male (published as “♂?”) of *Lespesia*, possibly near *L. aletiae* (Riley, 1879). The label indicates that it is from Cuba, not Chile as published and as subsequently interpreted. It is moved here to *Lespesia* as a *nomen dubium* from various uncertain placements (see references below).

References: Cortés (1946: 184), listed under “Species *incertae sedis*” at end of Tachinidae; Cortés and Hichins (1969: 90), listed under “Especies excluidas de la lista (*incertae sedis*)”; Guimaráes (1971: 193), listed as an unplaced species of Sturmiini.

Genus *RCORTESIA* Koçak & Kemal, 2010

HYPSONOMYIA Cortés, 1984: 382 (junior homonym of *Hypsomyia* McAlpine, 1965).

Type species: *Hypsomyia hispida* Cortés, 1983, by original designation [Chile].

RCORTESIA Koçak & Kemal, 2010: 159 (*nomen novum* for *Hypsomyia* Cortés, 1983).

Reference: Cortés (1984: 380), in key to tachinid genera of Tarapacá and Antofagasta regions.

hispida (Cortés, 1983).—Neotropical: South America (Chile).

Hypsomyia hispida Cortés, 1984: 383. Holotype male (MEUC). Type locality: Chile, Arica y Parinacota, Arica, Las Cuevas, Parque Nacional Lauca, 4800 m.

Genus *TELONOTOMYIA* Cortés, 1986

TELONOTOMYIA Cortés, 1986: 151. Type species: *Telonotomyia remota* Cortés, 1986, by original designation [Chile].

Reference: Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

remota Cortés, 1986.—Neotropical: South America (Chile).

Telonotomyia remota Cortés, 1986: 152. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Río Seco [as “Los Robles”, ca. 53°5'S, 70°53'W].

Unplaced species of Eryciini

The genus *Phorocera* Robineau-Desvoidy, 1830 *sensu* Aldrich (1934: 69) and Cortés (1945d: 158, 1950: 7) was cosmopolitan in distribution and consisted of many species. The main characters were given by Aldrich (1934: 69) as “hairy eyes, receding face, and bristly facial ridges”. None of the six Chilean species described in *Phorocera* by Aldrich and Cortés belong in the genus as defined by Wood (1972). We are only able to place one of these species to genus (*Phorocera calypttrata* Aldrich, 1934 is a species of *Admontia* Brauer & Bergenstamm) and the others are left unplaced to genus (two here in Eryciini, one in Goniini, and one in Winthemiini) or subfamily (one unplaced species of Tachinidae).

chilensis Cortés, 1950.—Neotropical: South America (Chile). (Fig. 5b)

Phorocera chilensis Cortés, 1950: 7. Holotype male (INLA). Type locality: Chile, Coquimbo, Elqui, Gualiguaica.

Note: Cortés and Hichins (1969: 27) placed *Phorocera chilensis* in the Old World genus *Clemelis* Robineau-Desvoidy, 1863 (Goniini) and this was followed by Henry (1987: 196) and González (1992b: 183). We follow Guimarães (1971: 214) in treating this species as unplaced in the Eryciini.

Reference: Cortés (1950: 10), in key to Chilean species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*).

elisae Cortés, 1945.—Neotropical: South America (Chile). (Fig. 5c)

Phorocera elisae Cortés, 1945d: 162. Holotype female (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Note: Guimarães (1971: 161) listed *Phorocera elisae* as an unrecognised species of Exoristini. We treat it as a recognised but unplaced species of Eryciini based on the examination of the holotype by DMW.

References: Cortés (1945d: 159), in key to Chilean species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*) and *Parasetigena* Brauer & Bergenstamm, 1891; Cortés (1950: 10), in key to Chilean species of *Phorocera* (*s. lato*); Cortés (1951a: 65), first description of male; Henry (1987: 206), in *Phorocera* but genus unplaced in Tachinidae.

Tribe ETHILLINI

Genus *NEOETHILLA* Cerretti, Wood & O'Hara, 2012

NEOETHILLA Cerretti, Wood & O'Hara, 2012: 28. Type species: *Exorista ignobilis* van der Wulp, 1890, by original designation [Mexico].

ignobilis (van der Wulp, 1890).—Not Chile [Mexico, United States].

Exorista ignobilis van der Wulp, 1890b: 71.

Note: *Exorista ignobilis* was assigned to *Winthemia* Robineau-Desvoidy by Reinhard (1931: 16) and stayed in this genus until recently recognised as the sole New World member of the Ethillini and placed in the new genus *Neoethilla* by Cerretti et al. (2012). Its distribution is thought to be limited to United States and Mexico (Cerretti et al. 2012) and reports of the species from South America in the following works are likely based on misidentifications of one or more *Winthemia* species: Reinhard (1931: 17, Argentina, Chile), Aldrich (1934: 44, Argentina, Chile), Cortés (1946: 175, Chile), Cortés (1948: 124, Chile), Cortés and Hichins (1969: 63, Chile), Henry (1987: 200, Chile), Coelho et al. (1989: 275, as "*ignobilis*", Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Peru, Venezuela), Vergara de Sánchez and Raven (1990: 100, Chile) and González (1992b: 180, 183, Chile). Cerretti et al. (2012) suspected that the species called *W. ignobilis* in Chile and Argentina might be *Winthemia reliqua* Cortés & Campos, 1971, which Coelho et al. (1989: 281) has treated as a synonym of *Winthemia trinitatis* Thompson, 1963.

Tribe EXORISTINI

Genus *CHETOGENA* Rondani, 1856

SALIA Robineau-Desvoidy, 1830: 108 (junior homonym of *Salia* Hübner, 1818).

Type species: *Salia echinura* Robineau-Desvoidy, 1830 (= *Tachina obliquata* Fallén, 1810), by subsequent designation of Robineau-Desvoidy (1863a: 553) [France].

CHETOGENA Rondani, 1856: 68. Type species: *Salia rondaniana* Villeneuve, 1931, by fixation of O'Hara and Wood (2004: 145) under Article 70.3.2 of the *Code* (ICZN 1999), misidentified as *Tachina gramma* Meigen, 1824 in the original designation by Rondani (1856) (see O'Hara et al. 2011: 54) [France].

SPOGGOSIA Rondani, 1859: 182. Type species: *Spoggosia oclusa* Rondani, 1859 (= *Tachina obliquata* Fallén, 1810), by monotypy [Italy and Malta].

STOMATOMYIA Brauer & Bergenstamm, 1889: 98 [also 1890: 30]. Type species: *Chetogena filipalpis* Rondani, 1859, by subsequent designation of Brauer (1893: 483) [Italy].

TETRAGRAPHIA Brauer & Bergenstamm, 1891: 351 [also 1891: 47]. Type species: *Tetragrapha tessellata* Brauer & Bergenstamm, 1891, by monotypy [Cuba].

EUPHOROCERA Townsend, 1892b: 112. Type species: *Euphorocera tachinomoides* Townsend, 1892, by original designation [United States].

PLAGIPROSPHERYSA Townsend, 1892b: 113. Type species: *Plagiprospherysa valida* Townsend, 1892 (= *Prospheysa parvipalpis* van der Wulp, 1890), by original designation [United States].

TACHINOPSIS Coquillett, 1897: 38, 120. Type species: *Tachinopsis mentalis* Coquillett, 1897 (= *Prospheysa parvipalpis* van der Wulp, 1890), by original designation [United States].

CHAETOGENA Bezzi & Stein, 1907: 315. Unjustified emendation of *Chetogena* Rondani, 1856 (see O'Hara et al. 2011: 54, 259).

PLAGIOTACHINA Townsend, 1927a: 261. Type species: *Plagiotachina peruviana* Townsend, 1927 (junior secondary homonym of *Euphorocera peruviana* Townsend, 1912; = *Euphorocera townsendi* Guimarães, 1971), by original designation [Peru].

STOMATOTACHINA Townsend, 1931d: 464. Type species: *Stomatotachina splendida* Townsend, 1931 (= *Parasetigena porteri* Brèthes, 1920), by original designation [Chile]. **Syn. nov.**

EPIPLAGIOPS Blanchard, 1943a: 450. Type species: *Epiplagiops littoralis* Blanchard, 1943 (junior secondary homonym of *Plagiops littoralis* Townsend, 1911; = *Plagiprospherysa floridensis* Townsend, 1892), by original designation [Argentina].

Note: *Parasetigena* Brauer & Bergenstamm, 1891 is an Old World genus with four species and a native distribution throughout the Palaearctic Region and northern portion of the Oriental Region (southern China). One species, *P. silvestris* (Robineau-Desvoidy, 1863), was successfully introduced into eastern North America for biological control purposes and has become established. The assignment of South American species to this Old World genus have been the result of a misunderstanding of the difference between *Parasetigena* and *Chetogena* Rondani. The latter currently has 71 species and is worldwide in distribution (O'Hara et al. 2020: 385). The two genera have the typical features of the Exoristini (principally prosternum haired, first postsutural supra-alar seta short and bend of vein M_1 right-angled) and a setose facial ridge, but in *Parasetigena* the setae on the facial ridge are weak and decumbent and in *Chetogena* they are strong and erect (see Wood 1987: 1209, cf. head figs 35 [*Chetogena tachinomoides* (Townsend, 1892)] and 36 [*Exorista larvarum* (Linnaeus, 1758), illustrative of *Parasetigena*] and pp. 1220–1221, key couplets 107–111). Based on this interpretation of *Chetogena*, we transfer two Chilean species originally described as *Parasetigena porteri* Brèthes (currently in *Stomatotachina* Townsend with type species of that genus in synonymy) and *Parasetigena hichinsi* Cortés to *Chetogena*.

References: Coquillett (1910: 522, 542, 589, 591, 602, 608, 611, 613), type species of *Chetogena*, *Euphorocera*, *Salia*, *Spoggosia* (all four as synonyms of *Phorocera* Robineau-Desvoidy, 1830), *Plagiprospherysa*, *Tachinopsis* and *Tetragrapha*; Aldrich (1926a: 14), synonymy of *Tachinopsis* with *Plagiprospherysa*; Aldrich (1934: 3, 4, 34, 69), in key to Patagonian genera, synonymy, taxonomic notes (as *Plagiprospherysa* and *Phorocera* [in part]); Townsend (1936c: 116, 123, 273, 281), diagnosis of adults and immatures of Exoristini and key to genera (including *Euphorocera*, *Plagiotachina*, *Spoggosia* and *Tetragrapha*), diagnosis of adults and immatures of Phoriniini and key to genera (including *Plagiprospherysa*, *Stomatomyia*, *Stomatotachina* and *Tachinopsis*), *Chetogena* as synonym of *Phorocera*, *Spoggosia* as valid name for *Salia*; Townsend (1940a: 160, 167, 170, 171, 182, 184, 185, 186), redescrptions of *Eu-*

phorocera, *Plagiotachina*, *Spoggosia* (with *Salia* in synonymy), *Tetragrapha*, *Plagiprospherysa*, *Stomatomyia*, *Stomatotachina* and *Tachinopsis*; Mesnil (1946: 42), synonymy of *Plagiotachina* with *Euphorocera*; Cortés and Campos (1971: 23, 24, 1974: 113, 114) and Cortés (1984: 379, 380), in keys to tachinid genera of Tarapacá and Antofagasta regions (as *Plagiprospherysa* and *Euphorocera*); Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions (as *Plagiprospherysa*); Wood (1987: 1221), synonymy of *Euphorocera*, *Spoggosia* and *Stomatomyia* with *Chetogena*; O'Hara and Wood (1998: 755, 759), review of synonymy of Wood (1987) (*Spoggosia* and *Stomatomyia* overlooked); Nihei (2015: 1, 2), synonymy of the monotypic genera *Epiplagiops* and *Tetragrapha* with *Chetogena*.

hichinsi (Cortés, 1967).—Neotropical: South America (Chile). **Comb. nov.** (Fig. 4f)
Parasetigena hichinsi Cortés, 1967b: 13. Holotype male (EEAM). Type locality: Chile, Metropolitana de Santiago, Santiago, Maipú, Rinconada.

Note: We examined specimens of *P. hichinsi* in CNC and have determined that it belongs to *Chetogena* according to the criteria given above in genus note.

References: Guimarães (1971: 159), as sole species of *Parasetigena* Brauer & Bergenstamm in America south of United States; Henry (1987: 199), in *Parasetigena*.

parvipalpis (van der Wulp, 1890).—Neotropical: Middle America (Mexico), South America (Argentina, Chile). Nearctic: Canada, United States.

Prospherysa parvipalpis van der Wulp, 1890d: 124. Syntypes, 3 males and 1 female (NHMUK). Type localities: Mexico, northern Sonora, Guerrero (Tepetlapa [ca. 18°3'N, 99°10'W], 3000 ft; Omiltemi [as "Omilteme", ca. 17°33'N, 99°41'W], 8000 ft), and Sinaloa (Villa Unión [as "Presidio", ca. 23°11'N, 106°13'W]).

Plagiprospherysa valida Townsend, 1892b: 113. Holotype male (SEMC, Byers et al. 1962: 176). Type locality: USA, New Mexico, Las Cruces.

Tachinopsis mentalis Coquillett, 1897: 120. Holotype male (USNM). Type locality: USA, Washington [state].

References: Aldrich (1934: 34), synonymy, redescription, taxonomic notes, first record from Argentina (as "Southern Patagonia", which is interpreted here as Argentina based on the travels of the collector, paleontologist Barnum Brown); Cortés and Hichins (1969: 53), first record from Chile.

peruviana (Townsend, 1912).—Neotropical: South America (Chile, Peru).

Euphorocera peruviana Townsend, 1912b: 303. Holotype female (USNM). Type locality: Peru, Piura, Piura.

Reference: Cortés and Campos (1971: 81), first record from Chile.

porteri (Brèthes, 1920).—Neotropical: South America (Chile). **Comb. nov.** (Fig. 5a)
Parasetigena porteri Brèthes, 1920b: 12. Lectotype, unspecified sex [female according to Mulieri et al. 2013: 169] (MACN), by fixation of Cortés (1963: 251) (examination of "Type" from Santiago in MACN is regarded as a lectotype fixation). Type locality: Chile, Metropolitana de Santiago, Santiago, Santiago.

Stomatotachina splendida Townsend, 1931d: 464. Holotype female (SDEI, Rohlfien and Ewald 1974: 144). Type locality: Chile, Biobío, Concepción, Concepción. **Comb. nov.**

Notes: We examined specimens of *P. porteri* in CNC, including a male identified by R. Cortés, and have determined that it belongs to *Chetogena* according to the criteria given above in genus note.

The type locality of *Parasetigena porteri* was given as “Santiago” and that of *Stomatotachina splendida* as “Concepcion”, both of which could be interpreted as either the city or province of those names. Cortés and Hichins (1969: 46) cited the cities as the type localities, as “Santiago (Santiago)” and “Concepción (Concepción)”, and we follow this interpretation. References: Cortés (1945d: 158, 159), in key to Chilean species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*) and *Parasetigena* Brauer & Bergenstamm, 1891, first description of male, synonymy of *Stomatotachina splendida* with *Parasetigena porteri*, synonymy of *Stomatotachina* with *Parasetigena*; Cortés (1963: 251), notes on name-bearing type of *P. porteri* in MACN; Guimarães (1971: 160), *Stomatotachina* revived as valid genus name for *P. porteri*; Mulieri et al. (2013: 169), notes on name-bearing type (as syntype) of *P. porteri* in MACN, as *Stomatotachina porteri*.

Tribe GONIINI

Some of the genera recognised as valid below are almost certainly synonymous with the widespread New World genus *Spallanzania* Robineau-Desvoidy, 1830. Wood and Zumbado (2010: 1412) noted: “*Spallanzania* has six North American species and ca. 20 nominal species (presently assigned to nearly as many genera) at high elevations in the Andes and at high latitudes in Patagonia”. We hesitate to formally propose any synonymy here because the diversity of morphological forms in the *Spallanzania* lineage is best left for a more detailed study before the limits of the genus are revised. Chilean genera to be considered in such a study include *Chaetocnephalia* Townsend, *Chaetocraniopsis* Townsend, *Coscaronia* Cortés and *Dolichocnephalia* Townsend.

Reference: González and Vergés (2004), revision of ten (of the 14) genera of Chilean Goniini (not included were *Belvosia* Robineau-Desvoidy, *Leschenaultia* Robineau-Desvoidy, *Patelloa* Townsend [formerly as *Macropatelloa* Townsend] and *Pseudochaeta* Coquillett).

Genus *ARAUCOGONIA* Cortés, 1976

ARAUCOGONIA Cortés, 1976: 10. Type species: *Araucogonia speciosa* Cortés, 1976, by original designation [Chile].

References: Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 42), in key to Chilean genera of Goniini, redescription.

speciosa Cortés, 1976.—Neotropical: South America (Chile).

Araucogonia speciosa Cortés, 1976: 11. Holotype male (MEUC). Type locality: Chile, Araucanía, Malleco, Pehuenco Chico, Marimenuco, 1000 m [ca. 38°43'S, 71°7'W].

Reference: González and Vergés (2004: 42), redescription.

Genus *ARAUCOSIMUS* Aldrich, 1934

ARAUCOSIMUS Aldrich, 1934: 88. Type species: *Araucosimus bullocki* Aldrich, 1934, by original designation [Chile].

References: Townsend (1936c: 169), diagnosis of adults and immatures of Goniini and key to genera (including *Araucosimus*); Townsend (1941: 10), redescription; Cortés and Campos (1971: 22, 1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions; González and Vergés (2004: 41, 43), in key to Chilean genera of Goniini, redescription, key to species.

bullocki Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Araucosimus bullocki Aldrich, 1934: 88. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

References: Cortés (1979: 76), taxonomic notes, first record from Argentina; González and Vergés (2004: 43), redescription.

orfilanus Cortés, 1979.—Neotropical: South America (Argentina, ?Chile).

Araucosimus orfilanus Cortés, 1979: 76. Holotype male (MLPA). Type locality: Argentina, Mendoza, Mendoza.

Note: Cortés (1979: 76) described *Araucosimus orfilanus* from Argentina but also identified a female from Chile (from El Melocotón near Santiago) as tentatively belonging to this species.

Reference: González and Vergés (2004: 44), redescription, tentatively recorded from Chile.

superbus Cortés, 1945.—Neotropical: South America (Chile).

Araucosimus superbus Cortés, 1945a: 122. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales, prov. de Valparaíso”, ca. 33°9'S, 71°18'W].

References: Campos (1953: 26), first description of female; González and Vergés (2004: 44), redescription.

Genus *BELVOSIA* Robineau-Desvoidy, 1830

BELVOSIA Robineau-Desvoidy, 1830: 103. Type species: *Belvosia bicincta* Robineau-Desvoidy, 1830, by monotypy [Unites States and West Indies].

LATREILLIA Robineau-Desvoidy, 1830: 104 (junior homonym of *Latreillia* Roux, 1830; priority established by ruling of ICZN 1964: 343, see Evenhuis et al. 2010: 96). Type species: *Musca bifasciata* Fabricius, 1775, by subsequent designation of Coquillett (1910: 558) (see Evenhuis et al. 2010: 96) [America, probably West Indies].

WILLISTONIA Brauer & Bergenstamm, 1889: 97 [also 1890: 29]. Type species: hereby fixed under Article 70.3.2 of the *Code* (ICZN 1999) as *Willistonina aldrichi* Townsend, 1931, misidentified as *Musca esuriens* Fabricius, 1805 in the fixation by monotypy of Brauer and Bergenstamm (1889) [Brazil].

LATREILLIMYIA Townsend, 1908: 105 (*nomen novum* for *Latreillia* Robineau-Desvoidy, 1830).

GONIOMIMA Townsend, 1908: 105. Type species: *Belvosia luteola* Coquillett, 1900, by monotypy [Puerto Rico].

TRIACHORA Townsend, 1908: 105. Type species: *Latreillia unifasciata* Robineau-Desvoidy, 1830, by monotypy [America, probably West Indies].

BELVOSIOMIMA Townsend, 1915e: 413. Type species: *Belvosiomima fosteri* Townsend, 1915, by original designation [Paraguay].

BELVOSIOPSIS Townsend, 1927a: 248. Type species: *Belvosiosis brasiliensis* Townsend, 1927 (= *Belvosia weyenberghiana* van der Wulp, 1883), by original designation [Brazil].

PSEUDOBELVOSIA Blanchard, 1954: 8. Type species: *Pseudobelvosia lugubris* Blanchard, 1954, by original designation [Argentina].

PARABELVOSIA Blanchard, 1954: 12. Type species: *Parabelvosia tibialis* Blanchard, 1954, by original designation [Argentina].

EUBELVOSIOPSIS Blanchard, 1954: 15. Type species: *Eubelvosiosis formosana* Blanchard, 1954, by original designation [Argentina].

NEOBELVOSIOPSIS Blanchard, 1954: 20. Type species: *Neobelvosiosis bosqi* Blanchard, 1954, by original designation [Argentina].

References: Williston (1893: 240), synonymy of *Latreillia* and *Williston* with *Belvosia*; Coquillett (1910: 513, 547, 558, 615, 619), type species of *Belvosia*, *Goniomima*, *Latreillia*, *Latreillimyia*, *Triachora* and *Williston* (with *Latreillia*, *Latreillimyia* and *Williston* in synonymy with *Belvosia*, and *Triachora* in synonymy with *Goniomima*); Aldrich (1928a: 1), synonymy of *Belvosiomima*, *Belvosiosis*, *Goniomima*, *Latreillimyia* and *Triachora* with *Belvosia*; Townsend (1936c: 180, 277), diagnosis of adults and immatures of Belvosiini and key to genera (including *Belvosia*, *Belvosiomima*, *Belvosiosis*, *Goniomima*, *Latreillimyia*, *Triachora* and *Williston*), *Latreillimyia* as valid name for *Latreillia*; Townsend (1941: 57, 58, 60, 66, 67, 74, 76), redescrptions of *Belvosia*, *Belvosiomima*, *Belvosiosis*, *Goniomima*, *Latreillimyia* (with *Latreillia* in synonymy), *Triachora* and *Williston*; Cortés and Campos (1971: 27, 1974: 116) and (1984: 382), *Triachora* in keys to tachinid genera of Tarapacá and Antofagasta regions; Guimarães (1971: 181), synonymy of *Eubelvosiosis*, *Neobelvosiosis*, *Parabelvosia* and *Pseudobelvosia* with *Belvosia*; Wood (1987: 1214), synonymy of *Triachora* with *Belvosia*; O'Hara and Wood (1998: 757, 759), review of synonymy of Wood (1987).

barbosai (Cortés & Campos, 1971).—Neotropical: South America (Chile).

Triachora barbosai Cortés & Campos, 1971: 98. Holotype female (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Codpa, 2109 m (18°50'S, 69°47'W) (coordinates and elevation given on p. 10; longitude given as "70°47'W", a location in the Pacific Ocean and likely an error for 69°47'W).

Reference: Cortés and Campos (1974: 123), first description of male.

Genus *CHAETOCNEPHALIA* Townsend, 1915

CHAETOCNEPHALIA Townsend, 1915d: 63. Type species: *Chaetocnephalia alpina* Townsend, 1915, by original designation [Peru].

References: Aldrich (1934: 4, 89), in key to Patagonian genera, taxonomic notes; Townsend (1936c: 169), diagnosis of adults and immatures of Goniini and key to genera (including *Chaetocnephalia*); Townsend (1941: 13), redescription; Cortés and Campos (1971: 22, 1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 45), in key to Chilean genera of Goniini, redescription, key to species.

americana (Schiner, 1868).—Neotropical: South America (Argentina, Chile).

Cnephalia americana Schiner, 1868: 327. Holotype female (NHMW). Type locality: Chile.

References: Aldrich (1927b: 31), redescription of holotype; Aldrich (1934: 89), redescription; Cortés (1979: 78), first record from Argentina; González and Vergés (2004: 46), redescription.

andina Cortés & Campos, 1971.—Neotropical: South America (Argentina, Bolivia, Chile).

Chaetocnephalia andina Cortés & Campos, 1971: 76. Holotype male (EEAM). Type locality: Chile, Antofagasta, El Loa, Ojo Hécar, 4500 m (23°11'S, 68°01'W) (coordinates and elevation given on p. 12, locality as “Ojo Hécar (Láscar”).

References: Cortés (1980: 107), first records from Argentina and Bolivia; González and Vergés (2004: 46), redescription.

cortesi González, 2004.—Neotropical: South America (Chile).

Chaetocnephalia cortesi González in González and Vergés, 2004: 47. Holotype male (UMCE). Type locality: Chile, Tarapacá, Tamarugal, Mamiña, 2800 m [ca. 20°4'S, 69°13'W].

innupta Cortés, 1945.—Neotropical: South America (Argentina, Chile).

Chaetocnephalia innupta Cortés, 1945a: 120. Holotype female (USNM). Type locality: Chile, Metropolitana de Santiago, Santiago, Las Condes.

References: Campos (1953: 26), first description of male; Gramajo (1998: 97), first record from Argentina; González and Vergés (2004: 48), redescription.

Genus *CHAETOCRANIOPSIS* Townsend, 1915

CHAETOCRANIOPSIS Townsend, 1915d: 68. Type species: *Chaetocraniopsis chilensis* Townsend, 1915, by original designation [Chile].

VALPOGONIA Townsend, 1928b: 163. Type species: *Valpogonia chilensis* Townsend, 1928 (junior secondary homonym of *Chaetocraniopsis chilensis* Townsend, 1915; = *Chaetocraniopsis argenteiceps* Aldrich, 1928), by original designation [Chile].

References: Aldrich (1928b: 19), taxonomic notes on *Chaetocraniopsis*, key to two species; Townsend (1936c: 169), diagnosis of adults and immatures of Goniini and key to genera (including *Chaetocraniopsis* and *Valpogonia*); Townsend (1941: 14, 51), redescriptions of *Chaetocraniopsis* and *Valpogonia*; Cortés (1945a: 116), synonymy of *Valpogonia* with *Chaetocraniopsis*, key to Chilean species; Cortés (1980: 107), description of a species from Argentina (*C. transandinum* Cortés); Cortés and Campos (1971: 22, 1974: 113) and Cortés (1984: 379),

in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 49), in key to Chilean genera of Goniini, redescription, key to species.

argenteiceps Aldrich, 1928.—Neotropical: South America (Argentina, Chile).

Chaetocraniopsis argenteiceps Aldrich, 1928b: 20. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

Valpogonia chilensis Townsend, 1928b: 163 (junior secondary homonym of *Chaetocraniopsis chilensis* Townsend, 1915). Holotype female (USNM). Type locality: Chile, Valparaíso, Valparaíso, “hills back of Valparaíso”.

Note: *Valpogonia chilensis* Townsend (published in “early 1928” according to Evenhuis et al. 2015: 352) has priority over *Chaetocraniopsis argenteiceps* Aldrich (published on 1 December 1928 according to the Table of Contents of the journal volume) when the two are treated as synonyms, but *Valpogonia chilensis* is a junior secondary homonym of *Chaetocraniopsis chilensis* Townsend, 1915 and thus invalid.

References: Cortés (1945a: 119), synonymy, taxonomic notes; Cortés (1980: 107), first record from Argentina; González and Vergés (2004: 50), redescription; Stireman et al. (2016: 38), habitus images.

chilensis Townsend, 1915.—Neotropical: South America (Chile).

Chaetocraniopsis chilensis Townsend, 1915d: 69. Holotype male (USNM). Type locality: Chile.

References: Aldrich (1928b: 20), taxonomic notes; Cortés (1945a: 117), taxonomic notes; González and Vergés (2004: 50), redescription.

obliteratus Cortés, 1945.—Neotropical: South America (Chile).

Chaetocraniopsis obliteratus Cortés, 1945a: 117. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales, prov. de Valparaíso”, ca. 33°9'S, 71°18'W].

Reference: González and Vergés (2004: 51), redescription.

similis (Townsend, 1928).—Neotropical: South America (Chile).

Valpogonia similis Townsend, 1928b: 163. Holotype female (USNM). Type locality: Chile, Valparaíso, Valparaíso, “hills back of Valparaíso”.

References: Cortés (1945a: 120), first description of male; González and Vergés (2004: 52), redescription.

Genus *COSCARONIA* Cortés, 1979

COSCARONIA Cortés, 1979: 77. Type species: *Coscaronia atrogonia* Cortés, 1979, by original designation [Argentina].

COSCARCONIA. Incorrect original spelling of *Coscaronia* Cortés, 1979 (Cortés 1979: 78, see note).

Note: There are two original spellings for *Coscaronia* in Cortés (1979): *Coscaronia* (pp. 77, 78) and *Coscarconia* (p. 78). There is clear evidence in the work itself that the spelling

Coscarconia is an inadvertent error because the genus-group name is dedicated to dipterist S. Coscarón. Therefore, the spelling *Coscaronia* is deemed to be the correct original spelling (Article 32.5.1 of the *Code*, ICZN 1999).

References: Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 52), in key to Chilean genera of Goniini, redescription.

antennalis Cortés, 1986.—Neotropical: South America (Chile).

Coscaronia antennalis Cortés, 1986: 157. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Tierra del Fuego, Isla Grande de Tierra del Fuego, Puerto Espora.

Reference: González and Vergés (2004: 53), redescription.

propinqua Cortés, 1979.—Neotropical: South America (Argentina, Chile).

Coscaronia propinqua Cortés, 1979: 78. Holotype male (MEUC). Type locality: Chile, Aysén, General Carrera, Chile Chico.

References: Gramajo (1998: 97), first record from Argentina; González and Vergés (2004: 53), redescription.

Genus *DOLICHOCNEPHALIA* Townsend, 1915

DOLICHOCNEPHALIA Townsend, 1915d: 64. Type species: *Dolichocnephalia puna* Townsend, 1915, by original designation [Peru].

References: Townsend (1936c: 169), diagnosis of adults and immatures of Goniini and key to genera (including *Dolichocnephalia*); Townsend (1941: 21), redescription; Cortés and Campos (1971: 21, 1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions; González and Vergés (2004: 41, 53), in key to Chilean genera of Goniini, redescription.

puna Townsend, 1915.—Neotropical: South America (Chile, Peru).

Dolichocnephalia puna Townsend, 1915d: 66. Holotype female (USNM). Type locality: Peru, Junín, La Oroya, Valle del Río Mantaro, higher than 12,000 ft.

References: Cortés and Campos (1971: 80), first record from Chile; González and Vergés (2004: 53), redescription.

Genus *ENCHOMYIA* Aldrich, 1934

ENCHOMYIA Aldrich, 1934: 42. Type species: *Gonia erythrocer*a Bigot, 1888, by original designation [Chile].

References: Townsend (1936c: 169), diagnosis of adults and immatures of Goniini and key to genera (including *Enchomyia*); Townsend (1941: 23), redescription; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 54), in key to Chilean genera of Goniini, redescription.

***erythrocer*a** (Bigot, 1888).—Neotropical: South America (Chile).

*Gonia erythrocer*a Bigot, 1888b: 86. Holotype female (NHMUK). Type locality: Chile.

References: Aldrich (1934: 42), diagnosis, taxonomic notes; Cortés (1951a: 60), first description of male; González and Vergés (2004: 54), redescription.

shewelli Cortés, 1976.—Neotropical: South America (Chile).

Enchomyia shewelli Cortés, 1976: 5. Holotype male (CNC). Type locality: Chile, Coquimbo, Elqui, Baños El Toro, 3300–4000 m [ca. 29°50'S, 70°1'W].

Reference: González and Vergés (2004: 54), redescription.

Genus *GONIA* Meigen, 1803

SALMACIA Meigen, 1800: 38. Meigen (1800) suppressed by ICZN (1963: 339).

GONIA Meigen, 1803: 280. Type species: *Gonia bimaculata* Wiedemann, 1819, by subsequent designation of Sabrosky and Arnaud (1965: 1075) [South Africa].

SALMACIA Meigen *in* Hendel, 1908: 65. First usage of *Salmacia* (*sensu* Meigen, 1800) as a valid name after Meigen, 1800; no type species designated originally or subsequently (see note).

PHOSOCOCEPHALOPS Townsend, 1927a: 237. Type species: *Phosococephalops fulvus* Townsend, 1927 (= *Gonia pallens* Wiedemann, 1830), by original designation [Brazil].

Note: The name *Salmacia* Meigen, 1800 became unavailable when the pamphlet of Meigen (1800) was suppressed by ICZN (1963: 339). *Salmacia* became available later when given by Meigen *in* Hendel (1908: 65), as explained in Evenhuis and Pape (2017: 51). This last work cited the type species of *Salmacia* Meigen *in* Hendel, 1908 as *Musca capitata* De Geer, 1776 by designation of Coquillett (1910: 602) but this is incorrect; Coquillett (1910) designated a type species for *Salmacia* Meigen, 1800 (at the time an available name) not *Salmacia* Meigen *in* Hendel, 1908.

References: Coquillett (1910: 547, 602), type species of *Gonia* and *Salmacia* Meigen, 1800 (with *Gonia* [and others] in synonymy with *Salmacia*); Townsend (1931b: 177), synonymy of *Phosococephalops* with *Gonia*; Aldrich (1934: 4, 86), in key to Patagonian genera, synonymy, taxonomic notes, key to two Patagonian species; Townsend (1936c: 169, 281), diagnosis of adults and immatures of Goniini and key to genera (including *Gonia* and *Phosococephalops*), *Salmacia* as synonym of *Gonia*; Townsend (1941: 31, 42), redescrptions of *Gonia* (with *Salmacia* in synonymy) and *Phosococephalops*; Cortés and Campos (1971: 84), in key to tachinid genera of Tarapacá and Antofagasta regions, key to separate several species of *Gonia*; Cortés and Campos (1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González and Vergés (2004: 41, 55), in key to Chilean genera of Goniini, redescription.

crassicornis (Fabricius, 1794).—Not Chile [Brazil, Peru, Venezuela; also Middle America, West Indies and Nearctic].

Musca crassicornis Fabricius, 1794: 328.

Note: *Gonia crassicornis* was recorded from Chile and Puerto Rico by Ashley (1979), citing Jones (1913) and Van Dine (1913) as sources. Neither of these last two papers record *G. crassicornis* from Chile and Ashley's (1979) record is assumed to be in error.

lineata Macquart, 1851.—Neotropical: South America (Argentina, Chile, Peru).

Gonia lineata Macquart, 1851: 151 [also 1851: 178]. Lectotype male (MNHN, see note), by fixation of Aldrich (1934: 87) (examination of “type” in MNHN is regarded as a lectotype fixation). Type locality: “Patagonie” (i.e., Argentina or Chile).

Gonia chiliensis of Blanchard (1854: 422), not Macquart, 1844. Misidentification (Aldrich 1934: 87).

Note: The online MNHN database records a male holotype in the Macquart collection for *Gonia lineata* (number MNHN-ED-ED8338) based on a holotype determination label that DMW attached to the specimen in 1982. Macquart did not restrict the name-bearing type to a single specimen and the lectotype fixation of Aldrich (1934: 87) is accepted here [see Recommendation 73F of the *Code* (ICZN 1999), “Avoidance of assumption of holotype”]. References: Aldrich (1934: 87), taxonomic notes, recorded from Argentina, Chile and Peru; Cortés (1963: 249), notes on name-bearing type in MNHN; Cortés (1979: 79), separation of *Gonia lineata* and *Gonia pallens*; González and Vergés (2004: 55), redescription.

pallens Wiedemann, 1830.—Neotropical: Greater Antilles (Cuba, Jamaica), eastern Lesser Antilles (Saint Vincent), Middle America (Mexico), South America (Argentina, Brazil, Chile, Ecuador, Paraguay, Peru).

Gonia pallens Wiedemann, 1830: 346. Lectotype, unspecified sex (NHMW), by fixation of Townsend (1931b: 177) (examination of “Ht” from Brazil in NHMW is regarded as a lectotype fixation). Type locality: Brazil.

Gonia chilensis Macquart, 1844: 50 [also 1844: 207]. Lectotype female (MNHN), by designation herein (see Lectotype Designations section). Type locality: Chile or Cuba.

Phosococephalops fulvus Townsend, 1927a: 347 (as “*fulva*” on p. 237). Lectotype female (USNM), by fixation of Townsend (1931b: 177) (examination of “female Ht” from São Paulo in USNM [as “Lima” but later changed to “Washington” in Townsend 1941: 42] is regarded as a lectotype fixation for the single female in the type series). Type locality: Brazil, São Paulo, Itaquaquecetuba.

Notes: Cortés (1963: 248) questioned whether *Gonia pallens* as interpreted here is a single species but we tentatively accept the synonymy and distribution above following Aldrich (1934: 87), Guimarães (1971: 176), González and Vergés (2004: 57) and others.

References: Van der Wulp (1888: 39), distribution as Brazil, Chile, Argentina, Mexico, Cuba and Jamaica; Williston (1896: 354), record from Saint Vincent, taxonomic notes; Townsend (1931b: 177), synonymy of *Phosococephalops fulvus* with *Gonia pallens*; Aldrich (1934: 87), synonymy, partial redescription, distribution as Argentina, Brazil, Chile, Ecuador, Paraguay and Peru; Blanchard (1963: 226), redescription, wing figure; Cortés (1963: 248), notes on type series in MNHN; Cortés (1979: 79), separation of *Gonia pallens* and *Gonia lineata*; González and Vergés (2004: 57), redescription.

Nomen dubium of *GONIA* Meigen, 1803

virescens Macquart, 1844.—Neotropical: South America. Distribution not known beyond the imprecise type locality of Brazil or Chile.

Gonia virescens Macquart, 1844: 50 [also 1844: 207]. Type(s), female (1 female in MNHN, see note). Type locality: Brazil or Chile.

Note: The online MNHN database records a holotype for *Gonia virescens* Robineau-Desvoidy, 1863 (number MNHN-ED-ED6825) in the Macquart collection. This record is in error; Robineau-Desvoidy (1863a: 741) did not describe a new species but instead cited the earlier Macquart species as “*Gonia virescens*: Macq. Coll. du Muséum” and assigned it to the genus *Reaumuria* Robineau-Desvoidy, 1830. He gave the provenance as Egypt, whereas Macquart had cited the species from “Brésil ou du Chili”. Perhaps Egypt was given in error and the “holotype” of “*Gonia virescens* Robineau-Desvoidy, 1863” is an overlooked type of *Gonia virescens* Macquart, 1844.

Reference: Guimarães (1971: 176), unrecognised species of *Gonia*.

Genus *LESCENAULTIA* Robineau-Desvoidy, 1830

LESCENAULTIA Robineau-Desvoidy, 1830: 324. Type species: *Leschenaultia cilipes* Robineau-Desvoidy, 1830, by subsequent designation of Townsend (1916b: 7) (see Evenhuis et al. 2010: 97) [Suriname].

BLEPHARIPEZA Macquart, 1844: 54 [also 1844: 211]. Type species: *Blepharipeza rufipalpis* Macquart, 1844 (= *Leschenaultia cilipes* Robineau-Desvoidy, 1830), by monotypy [Mexico].

ECHINOMASICERA Townsend, 1915e: 413. Type species: *Echinomasicera hystrix* Townsend, 1915, by original designation [Peru].

HARRISIOPSIS Townsend, 1927a: 247. Type species: *Harrisiopsis spinosa* Townsend, 1927 (= *Leschenaultia cilipes* Robineau-Desvoidy, 1830), by original designation [Brazil].

PARACHAETOPSIS Blanchard, 1959: 163. Type species: *Parachaetopsis proseni* Blanchard, 1959 (= *Blepharipeza bicolor* Macquart, 1846), by original designation [Argentina].

BLEPHRARIPEZA. Incorrect subsequent spelling of *Blepharipeza* Macquart, 1844 (Vimmer and Soukup 1940a: 217).

References: Coquillett (1910: 514), type species of *Blepharipeza*; Townsend (1931b: 175), synonymy of *Blepharipeza* and *Harrisiopsis* with *Leschenaultia*; Townsend (1936c: 186, 272, 276), diagnosis of adults and immatures of *Harrisiini* and key to genera (including *Echinomasicera* and *Leschenaultia*), synonymy; Townsend (1941: 77, 79), redescrptions of *Echinomasicera* and *Leschenaultia* (with *Blepharipeza* and *Harrisiopsis* in synonymy); Cortés (1984: 380), *Echinomasicera* in key to tachinid genera of Tarapacá and Antofagasta regions; Toma and Guimarães (2002), revision, synonymy including *Echinomasicera* and *Parachaetopsis* with *Leschenaultia*.

hystrix (Townsend, 1915).—Neotropical: South America (Chile, Peru).

Echinomasicera hystrix Townsend, 1915e: 413. Holotype male (USNM). Type locality: Peru, Lima, Matucana, ca. 8000 ft.

References: Cortés (1984: 386), first description of female, first record from Chile; Toma and Guimarães (2002: 38, 66), in key to *Leschenaultia* species, figures, diagnosis.

Genus *PATELLOA* Townsend, 1916

PATELLOA Townsend, 1916a: 619. Type species: *Phorocera leucaniae* Coquillett, 1897, by original designation [United States]. **New record from Chile.**

PATELLOAPSIS Townsend, 1927a: 263. Type species: *Patelloapsis similis* Townsend, 1927, by original designation [Brazil].

YAHUARPHRYNO Townsend, 1927a: 263. Type species: *Yahuarphryno patelloides* Townsend, 1927, by original designation [Peru].

MACROPATELLOA Townsend, 1931d: 472. Type species: *Macropatelloa tanumeana* Townsend, 1931, by original designation [Chile]. **Syn. nov.**

Note: *Patelloa* is currently known from 19 species that are widely distributed throughout the New World, including Argentina (three species) but not Chile (O'Hara et al. 2020: 491). *Macropatelloa tanumeana* is a common species in Chile and Argentina and is well-represented in CNC. It is recognised here as a typical species of *Patelloa* based on the following diagnostic features of the genus: prosternum haired, parafacial bare, facial ridge with row of strong setae, first postsutural supra-alar seta well-developed, and setae on postpronotum arranged in a triangle (Wood 1987: 1206, Wood and Zumbado 2010: 1361).

References: Townsend (1936c: 218, 237), diagnosis of adults and immatures of Lydellini and key to genera (including *Macropatelloa*), diagnosis of adults and immatures of Trypherini and key to genera (including *Patelloa*, *Patelloapsis* and *Yahuarphryno*); Townsend (1941: 191, 307, 327), redescrptions of *Macropatelloa*, *Patelloa*, *Patelloapsis* and *Yahuarphryno*; Sabrosky and Arnaud (1965: 1104), synonymy of *Patelloapsis* with *Patelloa*; Guimarães (1971: 211), synonymy of *Yahuarphryno* with *Patelloa*.

tanumeana (Townsend, 1931).—Neotropical: South America (Argentina, Chile).

Comb. nov. (Fig. 5d)

Macropatelloa tanumeana Townsend, 1931d: 472. Holotype female (USNM). Type locality: Chile, O'Higgins, Cardenal Caro, Tanumé [ca. 34°13'S, 71°55'W].

Note: Townsend (1931d: 472) described *Macropatelloa tanumeana* from two males and one female from "Tanumé and Talagante, Chile". The type locality of the female holotype was not given in the original description but was cited as Tanumé in Townsend (1941: 191).

References: Aldrich (1934: 69, 71), in key to Patagonian species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*), redescription; Cortés (1945d: 158), in key to Chilean species of *Phorocera* (*s. lato*) and *Parasetigena* Brauer and Bergenstamm, 1891; Cortés (1950: 10), in key to Chilean species of *Phorocera* (*s. lato*); Guimarães (1971: 161), as unrecognised species of Exoristini; Cortés (1979: 80), first record from Argentina (as *Macropatelloa tanumeana*); Cortés (1986: 144, 158), *Macropatelloa* in key to tachinid genera of Aysén and Magallanes regions, taxonomic notes on *M. tanumeana*; González (1992b: 179), survey data, as *M. tanumeana*.

Genus *PHILOCORUS* Cortés, 1976

PHILOCORUS Cortés, 1976: 12. Type species: *Philocorus montanum* Cortés, 1976, by original designation [Chile].

PHILOCHORUS. Incorrect subsequent spelling of *Philocorus* Cortés, 1976 (González and Vergés 2004: 41, 60).

References: Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions. González and Vergés (2004: 41, 60), in key to Chilean genera of Goniini, redescription.

montanum Cortés, 1976.—Neotropical: South America (Chile).

Philocorus montanum Cortés, 1976: 13. Holotype male (CNC). Type locality: Chile, Coquimbo, Elqui, Baños El Toro, 3300–4000 m [ca. 29°50'S, 70°1'W].

References: Cortés (1986: 158), first description of female; González and Vergés (2004: 61), redescription.

Genus *PROTOGONIOPS* Townsend, 1913

PROTOGONIA Townsend, 1912b: 347 (junior homonym of *Protogonia* Cope, 1881).

Type species: *Protogonia ocellaris* Townsend, 1912, by original designation [Peru].

PROTOGONIOPS Townsend, 1913a: 133 (*nomen novum* for *Protogonia* Townsend, 1912).

References: Townsend (1936c: 169, 280), diagnosis of adults and immatures of Goniini and key to genera (including *Protogoniops*), *Protogoniops* as valid name for *Protogonia*; Townsend (1941: 44), redescription of *Protogoniops* (with *Protogonia* in synonymy); Cortés and Campos (1974: 115) and Cortés (1984: 381), in keys to tachinid genera of Tarapacá and Antofagasta regions; González and Vergés (2004: 41, 61), in key to Chilean genera of Goniini, redescription.

ocellaris (Townsend, 1912).—Neotropical: South America (Chile, Peru).

Protogonia ocellaris Townsend, 1912b: 348. Holotype male (USNM). Type locality: Peru, western base of Cordillera Occidental, Río Suyu, ca. 1500 ft.

References: Cortés and Campos (1974: 122), first description of female, first record from Chile; González and Vergés (2004: 61), redescription.

Genus *PSEUDOCOAETA* Coquillett, 1895

References: Coquillett (1910: 596, 615), type species of *Pseudochaeta*; Townsend (1936c: 237, 274), diagnosis of adults and immatures of Trypherini and key to genera (including *Metopiops*, *Phaenopsis* and *Pseudochaeta*), *Dimasicera* as synonym of *Phaenopsis*; Townsend (1941: 287, 309, 313), redescrptions of *Metopiops*, *Phaenopsis* (with *Dimasicera* in synonymy) and *Pseudochaeta*; Reinhard (1946), revision of North American species of *Phaenopsis* and *Pseudochaeta*, key to all New World species; Thompson (1964: 98), synonymy of *Phaenopsis* with *Pseudochaeta*, revision of Trinidad species; Wood (1987: 1210), synonymy of *Metopiops* and *Phaenopsis* with *Pseudochaeta*; O'Hara and Wood (1998: 756, 766), review of synonymy of Wood (1987).

Subgenus *METOPIOPS* Townsend, 1912

METOPIOPS Townsend, 1912b: 338. Type species: *Metopiops mirabilis* Townsend, 1912, by original designation [Peru].

There are no Chilean species in this subgenus.

Subgenus *PHAENOPSIS* Townsend, 1912

PHAENOPSIS Townsend, 1912b: 362. Type species: *Phaenopsis arabella* Townsend, 1912, by original designation [Peru].

DIMASICERA Townsend, 1915c: 62. Type species: *Dimasicera nitida* Townsend, 1915 (= *Phaenopsis arabella* Townsend, 1912), by original designation [Peru].

References: Cortés and Campos (1971: 23, 1974: 114) and Cortés (1984: 380), *Phaenopsis* in keys to tachinid genera of Tarapacá and Antofagasta regions.

arabella (Townsend, 1912).—Neotropical: South America (Chile, Peru).

Phaenopsis arabella Townsend, 1912b: 363. Holotype male (USNM). Type locality: Peru, Piura, Valle del Río Chira, Sullana.

Dimasicera nitida Townsend, 1915c: 64. Holotype female (USNM). Type locality: Peru, Piura, Valle del Río Chira, near Sullana.

References: Townsend (1941: 310), synonymy of *Dimasicera nitida* with *Phaenopsis arabella*; Reinhard (1946: 111), notes on synonymy, in key to New World species of *Phaenopsis* and *Pseudochaeta*; Cortés and Campos (1971: 97), first record from Chile; Guimarães (1971: 163), earlier synonymy of *Dimasicera nitida* with *Phaenopsis arabella* apparently overlooked and both names listed as valid.

Subgenus *PSEUDOAETHA* Coquillett, 1895

PSEUDOAETHA Coquillett, 1895a: 309. Type species: *Pseudochaeta argentifrons* Coquillett, 1895, by original designation [United States].

Reference: Townsend (1908: 96), comparison with new genus *Trepophrys*.

There are no Chilean species in this subgenus.

Unplaced species of *Goniini*

leliae Cortés & Campos, 1971.—Neotropical: South America (Chile).

Lespesia leliae Cortés & Campos, 1971: 91. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, Rosario, 352 m (18°26'S, 70°06'W) (coordinates and elevation given on p. 11).

Note: See note under *A. robusta* for comments on the tribal placement of this species.

negrensis Aldrich, 1934.—Neotropical: South America (Argentina, Chile). (Fig. 5e)

Phorocera negrensis Aldrich, 1934: 72. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Notes: *Phorocera negrensis* was recorded from both Argentina and Chile in the original description.

Phorocera negrensis was listed as an unplaced species of Blondeliini by Guimarães (1971: 153) but it unquestionably belongs to the *Cyzenis* Robineau-Desvoidy, 1863–*Frontiniella* Townsend, 1918 clade of Goniini, a complex with additional but undescribed species in Chile. These species typically have a haired eye, setose facial ridge (on lower half or more), and lack a pair of apical scutellar setae (Wood and Zumbado 2010: 1363). DNA barcoding by JEOH suggests that *Chrysoexorista* Townsend, 1915 is also close to or part of this clade. The molecular phylogeny of Stireman et al. (2019: 13 [fig. 8]) did not include *Cyzenis* but found a close relationship between *P. negrensis* and *Frontiniella*. We could assign *Phorocera negrensis* to *Cyzenis* here based on external morphology but we are reluctant to place it to genus without a proper study of the undescribed species related to it.

References: Aldrich (1934: 69, 72), in key to Patagonian species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*); Cortés (1945d: 158), in key to Chilean species of *Phorocera* (*s. lato*) and *Parasetigena* Brauer & Bergenstamm, 1891; Cortés (1950: 10), in key to Chilean species of *Phorocera* (*s. lato*); Guimarães (1971: 153), as unplaced species of Blondeliini.

nimia Cortés & Campos, 1971.—Neotropical: South America (Chile).

Lespesia nimia Cortés & Campos, 1971: 95. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, km 57.

Note: See note under *A. robusta* for comments on the tribal placement of this species.

robusta Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Achaetoneura robusta Aldrich, 1934: 91. Holotype male (USNM). Type locality: Chile, Metropolitana de Santiago, Santiago, Cerro San Cristóbal.

Note: Guimarães (1971: 194) assigned *Achaetoneura robusta* Aldrich to “Unplaced Species of Sturmiini” (i.e., Goniini) without commenting on its placement. Later, Guimarães (1983: 23, 28) transferred two additional species to this category and commented that all three produce microtype eggs (i.e., belong to Goniini, not Eryciini), writing: “*Lespesia* deposits membranous eggs on the body of the host ... Chilean species recorded to this complex, viz. *L. robusta* Aldrich, *L. leliae* Cortés and *L. nimiae* Cortés, definitely do not belong to *Lespesia*, and their correct placement have not yet been established. Dissections of females of the three Chilean species show the presence of microtype eggs, the male and female genitalia being differently shaped”.

Reference: Gramajo (1998: 96), first record from Argentina (as *Achaetoneura robusta*).

Tribe WINTHEMIINI

Genus *WINTHEMIA* Robineau-Desvoidy, 1830

WINTHEMIA Robineau-Desvoidy, 1830: 173. Type species: *Musca quadripustulata* Fabricius, 1794, by subsequent designation of Desmarest in d'Orbigny (1849: 301) (see Evenhuis and Thompson 1990: 239) [Germany].

MICROTRICHODES Macquart, 1846: 288 [also 1846: 160]. Type species: *Microtrichodes analis* Macquart, 1846, by original designation [Brazil].

- MASIPODA* Brauer & Bergenstamm, 1889: 162 [also 1889: 94]. Type species: *Masipoda geminata* Brauer & Bergenstamm, 1889, by monotypy [Mexico].
- HEMIMASIPODA* Townsend, 1927a: 267. Type species: *Hemimasipoda brasiliensis* Townsend, 1927, by original designation [Brazil].
- OKEOPSIS* Townsend, 1927a: 267. Type species: *Okeopsis palpalis* Townsend, 1927, by original designation [Brazil].
- PROWINTHEMIA* Townsend, 1928a: 151. Type species: *Prowinthemia paraguayensis* Townsend, 1928 (= *Exorista tricolor* van der Wulp, 1890), by original designation [Paraguay].
- BICRUCIOSTURMIA* Townsend, 1932b: 106. Type species: *Bicruciosturmia bicrucis* Townsend, 1932, by original designation [Brazil].
- PROMASIPODA* Townsend, 1934b: 399. Type species: *Promasipoda pinguoides* Townsend, 1934, by original designation [Brazil].
- PRONEMORILLA* Townsend, 1935: 229. Type species: *Pronemorilla mima* Townsend, 1935 (junior secondary homonym of *Winthemia mima* Reinhard, 1931; = *Winthemia trinitatis* Thompson, 1963), by original designation [Brazil].
- WINTHEMIOPSIS* Blanchard, 1963: 212. Type species: *Winthemiopsis grioti* Blanchard, 1963, by original designation [Argentina].
- MICROTRICHOMODES*. Incorrect subsequent spelling of *Microtrichodes* Macquart, 1846 (Guimarães 1972: 42).
- WINTHEMYA*. Incorrect subsequent spelling of *Winthemia* Robineau-Desvoidy, 1830 (Robineau-Desvoidy 1863a: 206–216; Vimmer and Soukup 1940a: 207).
- WINTHEMYIA*. Incorrect subsequent spelling of *Winthemia* Robineau-Desvoidy, 1830 (e.g., Vimmer and Soukup 1940b: 370).
- WITHEMLIA*. Incorrect subsequent spelling of *Winthemia* Robineau-Desvoidy, 1830 (Etcheverry 1957: 187).

Notes: There is much confusion in the literature regarding the valid names of *Winthemia* species and their synonyms in the New World. For practical purposes the synonymy proposed by Coelho et al. (1989) is followed here.

Macquart (1846: 289 [also 1846: 161]) noted about his new genus *Microtrichodes*, “Le type de ce genre est du Brésil” [“The type of this genus is from Brazil”]. This statement is accepted as a type species designation for *Microtrichodes* of the single included species, *Microtrichodes analis* Macquart, from Brazil.

The species treated by many authors as *Winthemia ignobilis* (van der Wulp) was moved to the Ethillini by Cerretti et al. (2012: 34) and is treated here under the name *Neoethilla ignobilis* (not recorded from Chile; see explanation under that name).

References: Coquillett (1910: 565, 620), type species of *Masipoda* and *Winthemia* (with former in synonymy with latter; type species of *Winthemia* given as “*Musca quadripustulata* Fabricius ... by designation of Desvoidy ... vol. 1, 1863, p. 207”); Reinhard (1931), revision of New World species, synonymy of *Hemimasipoda*, *Masipoda* and *Microtrichodes* with *Winthemia*, key, descriptions; Aldrich (1934: 3, 43), in key to Patagonian genera, synonymy of *Prowinthemia* with *Winthemia*; Townsend (1936c: 190), diagnosis of adults and immatures of Sturmiini and key to genera (including *Bicruciosturmia*, *Hemimasipoda*, *Masipoda*, *Microtrichodes*, *Okeopsis*, *Promasipoda*, *Pronemorilla*, *Prowinthemia* and *Winthemia*); Townsend

(1941: 90–138), redescrptions of the aforementioned genera; Thompson (1963b: 960), revision of Trinidad species; Cortés and Campos (1971: 101), synonymy including *Pronemorilla* with *Winthemia*; Cortés and Campos (1971: 23, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions; Guimarães (1971: 196), synonymy of *Okeopsis*, *Promasipoda* and *Winthemiopsis* with *Winthemia*; Guimarães (1972), revision of species from north of Mexico; Coelho et al. (1989), synonymy of *Bicruciostrumia* with *Winthemia*, key and review of South American species.

quadripustulata (Fabricius, 1794).—Not Chile [Palearctic; also Nearctic and Oriental].
Musca quadripustulata Fabricius, 1794: 324.

Note: *Winthemia quadripustulata* was recorded from Chile by Molina-Ochoa et al. (2003: 262) based on an earlier record by Etcheverry (1957: 187, as “*Withemia*” *quadripustulata*). This is undoubtedly a misidentification; *W. quadripustulata* is widely distributed in the Palearctic, Nearctic and Oriental regions and though possibly a species complex is not reliably known from South America. It was not recognised from America south of United States by Guimarães (1971, 1972) or from South America by Coelho et al. (1989).

singularis Reinhard, 1931.—Neotropical: southern Lesser Antilles (Trinidad & Tobago), South America (Argentina, Brazil, ?Chile, Colombia, Ecuador, Paraguay, Peru, Venezuela).

Winthemia singularis Reinhard, 1931: 38. Holotype male (USNM). Type locality: Argentina, Tucumán [province or city].

Hemimasipoda alabamae Townsend, 1940b: 892. Lectotype male (MZSP), by fixation of Coelho et al. (1989: 280) (examination of “holótipo macho” in MZSP is regarded as a lectotype fixation). Type locality: Brazil, São Paulo, Ribeirão Preto.

Winthemia aureonigra Thompson, 1963b: 978. Holotype male (CNC). Type locality: Trinidad, Maracas Valley.

Winthemia roblesi Valencia, 1972b: 366. Holotype male (SENASA, Lozada et al. 2005: 460). Type locality: Peru, Ica, Huamaní.

Note: The only record of *Winthemia singularis* from Chile was given in a table in Molina-Ochoa et al. (2003: 262), as *Winthemia roblesi*. Due to the difficult nature of identifying *Winthemia* specimens, the synonymy of Coelho et al. (1989: 280) needs confirmation, as does the presence of *Winthemia singularis* in Chile. *Winthemia aureonigra* (holotype examined by DMW) is a particularly unlikely synonym.

References: Coelho et al. (1989: 275, 280), in key to South American species, synonymy of *Hemimasipoda alabamae*, *Winthemia aureonigra* and *Winthemia roblesi* with *Winthemia singularis*, distribution; Nihei (2016: 935), in catalogue of Tachinidae of Colombia.

trinitatis Thompson, 1963.—Neotropical: southern Lesser Antilles (Trinidad & Tobago), South America (Argentina, Bolivia, Brazil, ?Chile, Colombia, Paraguay, Peru, Venezuela).

Pronemorilla mima Townsend, 1935: 230 (junior secondary homonym of *Winthemia mima* Reinhard, 1931). Holotype female (MZSP). Type locality: Brazil, São Paulo, São Vicente.

Winthemia trinitatis Thompson, 1963b: 971. Holotype male (CNC). Type locality: Trinidad, Chaguanas.

Winthemia reliqua Cortés & Campos, 1971: 101 (*nomen novum* for *Pronemorilla mima* Townsend, 1935).

reliquia. Incorrect subsequent spelling of *reliqua* Cortés & Campos, 1971 (Valencia 1972b: 365, etc.).

Note: *Winthemia trinitatis* was recorded from Chile by Coelho et al. (1989: 275) but we are doubtful that this species, which was originally described from Trinidad, occurs there.

References: Coelho et al. (1989: 275, 281), in key to South American species, distribution, and synonymy of *Pronemorilla mima* Townsend, 1935 with *Winthemia trinitatis*; Nihei (2016: 935), in catalogue of Tachinidae of Colombia.

Unplaced species of Winthemiini

bullocki Aldrich, 1934.—Neotropical: South America (Argentina, Chile). (Fig. 5f)

Phorocera bullocki Aldrich, 1934: 70. Syntypes, 4 females (NHMUK, USNM, according to databases of these collections). Type localities: Chile, Araucanía (Malleco, Angol) and Metropolitana de Santiago (Santiago, Cerro San Cristóbal).

Note: *Phorocera bullocki* and two related but undescribed species are each represented in CNC by specimens from Chile and Argentina. A new genus in the Winthemiini may be warranted for these species. The katapimeron is haired as in other members of the tribe. The parafacial is bare (haired in *Winthemia* species) and facial ridge is setose.

References: Aldrich (1934: 69, 70), in key to Patagonian species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*), description; Cortés (1945d: 159), in key to Chilean species of *Phorocera* (*s. lato*) and *Parasetigena* Brauer & Bergenstamm, 1891; Cortés (1950: 10), in key to Chilean species of *Phorocera* (*s. lato*); Campos (1953: 25), first description of male (in *Phorocera*); Guimarães (1971), name missing from catalogue; Henry (1987: 206), first record from Argentina (in *Phorocera* but genus unplaced in Tachinidae).

Unplaced genus of Exoristinae

Genus **CALTAGIRONEA** Cortés & Campos, 1974

CALTAGIRONEA Cortés & Campos, 1974: 117. Type species: *Caltagironea vera* Cortés & Campos, 1974, by original designation [Chile].

Note: Cortés and Campos (1974: 117) placed their new genus *Caltagironea* in the “Sturmiini” *sensu* Crosskey (1973: 91), a group comprising genera that would later be assigned to the Eryciini or Goniini depending upon reproductive habit (with those producing microtype eggs being placed in the latter). Cortés and Campos (1974) recognised both the Goniini and Sturmiini and hence their sturmiines are generally eryciines in modern terminology. González and Vergés (2004) excluded *Caltagironea* from their revision of Chilean Goniini and although it likely belongs to the Eryciini we cannot rule out its placement elsewhere in the Tachinidae. Reference: Cortés (1984: 381), in key to tachinid genera of Tarapacá and Antofagasta regions.

scillina Cortés & Campos, 1974.—Neotropical: South America (Chile).

Caltagironea scillina Cortés & Campos, 1974: 120. Holotype male (MEUC). Type locality: Chile, Arica y Parinacota, Arica, Valle de Camarones, Taltape, 300–400 m.

vera Cortés & Campos, 1974.—Neotropical: South America (Chile).

Caltagironea vera Cortés & Campos, 1974: 119. Holotype male (MEUC). Type locality: Chile, Tarapacá, Tamarugal, south of (or road to) Chiapa, 3400–3800 m.

Subfamily PHASIINAE

Tribe CYLINDROMYIINI

Genus *CYLINDROMYIA* Meigen, 1803

References: Coquillett (1910: 529, 577), type species of *Cylindromyia* and *Ocyptera* (with latter in synonymy with former); Aldrich (1926: 2), revision of North American species, synonymy of *Apinocyptera* and *Odontocyptera* with *Cylindromyia*; Aldrich (1934: 2, 8), in key to Patagonian genera, synonymy of *Dolichocyptera*, *Glossidionophora* and *Melanocyptera* with *Cylindromyia*, taxonomic notes, key to three Chilean species; Townsend (1936b: 63), diagnosis of adults and immatures of *Cylindromyiini* and key to genera (including *Apinocyptera*, *Catocyptera*, *Cylindromyia*, *Dolichocyptera*, *Ecatocypterops*, *Glossidionophora*, *Melanocyptera*, *Ocypteryx* and *Odontocyptera*); Townsend (1938: 87–143), redescrptions of the aforementioned genera; Cortés (1944e), key and review of Chilean species; Guimarães (1971: 15), synonymy of *Ecatocypterops* with *Cylindromyia*; Guimarães (1976), revision of species from south of the United States, synonymy including *Catocyptera* with *Cylindromyia*, three subgenera recognised for New World *Cylindromyia*.

Subgenus *APINOCYPTERA* Townsend, 1915

APINOCYPTERA Townsend, 1915f: 94. Type species: *Apinocyptera signata* Townsend, 1915 (= *Ocyptera signatipennis* van der Wulp, 1892), by original designation [Guatemala].

ODONTOCYPTERA Townsend, 1915h: 233. Type species: *Odontocyptera nana* Townsend, 1915, by original designation [Mexico].

There are no Chilean species in this subgenus (Guimarães 1976: 24).

Subgenus *CYLINDROMYIA* Meigen, 1803

CYLINDROMYIA Meigen, 1803: 279. Type species: *Musca brassicaria* Fabricius, 1775, by monotypy [Europe].

OCYPTERA Latreille, 1804: 195. Type species: *Musca brassicaria* Fabricius, 1775, by subsequent designation of Curtis (1837: 629) [Europe].

GLOSSIDIONOPHORA Bigot, 1885a: 237. *Nomen nudum*.

GLOSSIDIONOPHORA Bigot, 1885c: lv [also 1885c: lv, *Bull. Soc. Ent. France*].

Type species: *Glossidionophora nigra* Bigot, 1885, by subsequent designation of Townsend (1916b: 7) [Argentina].

CATOCYPTERA Townsend, 1927a: 215. Type species: *Catocyptera brasiliiana* Townsend, 1927, by original designation [Brazil].

MELANOCYPTERA Townsend, 1927a: 215. Type species: *Melanocyptera carinata* Townsend, 1927, by original designation [Brazil].

DOLICHOCYPTERA Townsend, 1931c: 325. Type species: *Dolichocyptera pirioni* Townsend, 1931, by original designation [Chile].

OCYPTERYX Townsend, 1931c: 326. Type species: *Ocypteryx ochrescens* Townsend, 1931 (= *Ocyptera dorsalis* Wiedemann, 1830), by original designation [Paraguay].

ECATOCYPTEROPS Townsend, 1935: 217. Type species: *Ecatocypterops ater* Townsend, 1935 (junior secondary homonym of *Ocyptera atra* Röder, 1885; = *Melanocyptera carinata* Townsend, 1927), by original designation [Brazil].

aldrichi Cortés, 1944.—Neotropical: South America (Chile).

Cylindromyia aldrichi Cortés, 1944e: 178. Holotype male (USNM). Type locality: Chile, Metropolitana de Santiago, Santiago, Santiago.

Note: The type locality of *Cylindromyia aldrichi* was given as “Santiago” in Chile, which could be interpreted as either the city or province of that name. Cortés and Hichins (1969: 28) cited the former as the type locality (as “Santiago (Santiago)”) and we follow this interpretation.

Reference: Guimarães (1976: 8, 9), in key, taxonomic notes, figures.

apicalis (Bigot, 1878).—Neotropical: South America (Chile).

Ocyptera apicalis Bigot, 1878: 45. Lectotype male [original type(s) not female as published by Bigot] (NHMUK), by designation of Guimarães (1976: 10). Type locality: Chile.

References: Aldrich (1934: 10), redescription, taxonomic notes; Guimarães (1976: 8, 10), in key, redescription, taxonomic notes, figures.

nigra (Bigot, 1885).—Neotropical: South America (Argentina, Chile).

Glossidionophora nigra Bigot, 1885c: lv [also 1885c: lv, *Bull. Soc. Ent. France*]. Holotype female [not male as published, Guimarães 1976: 19] (NHMUK). Type locality: Argentina, Buenos Aires, Buenos Aires.

Glossidionophora cylindrica Brauer, 1899: 499. Holotype female [not male as published, Guimarães 1976: 19] (NHMUK). Type locality: Argentina, Buenos Aires, Buenos Aires.

Cylindromyia atricauda Aldrich, 1934: 10. Holotype female (NHMUK). Type locality: Chile, Valparaíso, San Felipe de Aconcagua, Llay-Llay [as “Llaillai”].

Note: The holotype of *Glossidionophora nigra* Bigot, 1885 is also the holotype of *Glossidionophora cylindrica* Brauer, 1899. Brauer described *Glossidionophora cylindrica* from a specimen in the Bigot collection labelled with that name but was unaware that Bigot had described *Glossidionophora nigra* from the same specimen a few years earlier (Guimarães 1976: 19–20). Reference: Guimarães (1976: 7, 19), in key, synonymy of *Cylindromyia atricauda* and *Glossidionophora cylindrica* with *Glossidionophora nigra*, redescription, taxonomic notes, figures, first record from Chile as *Cylindromyia nigra*.

pirioni (Townsend, 1931).—Neotropical: South America (Chile).

Dolichocyptera pirioni Townsend, 1931c: 326. Holotype female (USNM). Type locality: Chile, Metropolitana de Santiago, Santiago, Cerro San Cristóbal.

Reference: Guimarães (1976: 7, 22), in key, taxonomic notes.

porteri (Brèthes, 1925).—Neotropical: South America (Argentina, Chile).

Ocyptera porteri Brèthes, 1925: 208. Holotype female [not male as published, Mulieri et al. 2013: 169] (MACN). Type locality: Chile, Metropolitana de Santiago, Santiago, Las Condes.

References: Aldrich (1934: 9), redescription; Cortés (1963: 251), notes on name-bearing type in MACN, as male; Guimarães (1976: 8, 21), in key, redescription, taxonomic notes, figures; Gramajo (1998: 91), first record from Argentina; Mulieri et al. (2013: 169), notes on holotype in MACN.

Tribe GYMNOSOMATINI

Genus *GYMNOSOMA* Meigen, 1803

RHODOGYNE Meigen, 1800: 39. Meigen (1800) suppressed by ICZN (1963: 339).

GYMNOSOMA Meigen, 1803: 278. Type species: *Musca rotundata* Linnaeus, 1758 (as “*Musca rotundata* Fabr.”), by monotypy [Europe].

RHODOGYNE Meigen in Hendel, 1908: 66. Type species: *Musca rotundata* Linnaeus, 1758 (as “*M. rotundata* F.”), by monotypy (see Evenhuis and Pape 2017: 50) [Europe].

Note: The name *Rhodogyne* Meigen, 1800 became unavailable when the pamphlet of Meigen (1800) was suppressed by ICZN (1963: 339). *Rhodogyne* became available later when given by Meigen in Hendel (1908: 65), as explained in Evenhuis and Pape (2017: 50).

References: Coquillett (1910: 548, 600), type species of *Gymnosoma* and *Rhodogyne* Meigen, 1800 (with former in synonymy with latter); Townsend (1936b: 44), diagnosis of adults and immatures of Gymnosomatini and key to genera (including *Gymnosoma*); Townsend (1936c: 281), *Rhodogyne* as synonym of *Gymnosoma*; Townsend (1938: 7), redescription of *Gymnosoma* (with *Rhodogyne* in synonymy); Cortés and Campos (1971: 21, 1974: 112) and Cortés (1984: 378), in keys to tachinid genera of Tarapacá and Antofagasta regions.

neotropicale Cortés & Campos, 1971.—Neotropical: South America (Chile, Peru).

Gymnosoma neotropicale Cortés & Campos, 1971: 27. Holotype male (EEAM).

Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, km 23.

Reference: Vergara de Sánchez and Raven (1990: 94), first record from Peru.

Genus *TRICHOPODA* Berthold, 1827

References: Coquillett (1897: 47), key to species of America north of Mexico; Townsend (1897: 273), key to species in Vera Cruz; van der Wulp (1903: 433), revision of Central American species; Townsend (1908: 129), key to genera of Trichopodini; Coquillett (1910: 546, 593, 616), type species of *Galactomyia*, *Polistomyia* and *Trichopoda* (with first two in synonymy with last, *Trichopoda* as “*Trichiopoda* Latreille ... 1829”); Townsend (1936b: 47),

diagnosis of adults and immatures of Trichopodini and key to genera (including *Polistomyia*, “*Trichiopoda*” and *Trichopodopsis*); Townsend (1936c: 275), synonymy of *Galactomyia* with “*Trichiopoda*”; Townsend (1938: 25, 28, 30), redescrptions of *Polistomyia*, “*Trichiopoda*” (with *Galactomyia* in synonymy) and *Trichopodopsis*; Sabrosky (1950: 361, 366), key to genera of Trichopodini, synonymy of *Trichopodopsis* with *Trichopoda*, taxonomic notes; Blanchard (1966a: 62, 81), *Trichopodopsis* as valid genus, *Eutrichopodopsis* as new genus; Guimarães (1971: 7, 10), *Eutrichopodopsis* as valid genus, *Trichopodopsis* as synonym of *Trichopoda* subgenus *Galactomyia*; Liljesthröm (1992: 51, 56), revision of Argentinian species, synonymy of *Eutrichopodopsis* with *Trichopoda*; Dios and Nihei (2020), revision of Neotropical species.

Subgenus *GALACTOMYIA* Townsend, 1908

GALACTOMYIA Townsend, 1908: 135. Type species: *Trichopoda radiata* Loew, 1863 (= *Thereva lanipes* Fabricius, 1805), by subsequent designation of Coquillett (1910: 546) [United States].

TRICHOPODOPSIS Townsend, 1913b: 148, 313. Type species: *Musca pennipes* Fabricius, 1781, by subsequent monotypy of Anonymous (1913: 313) (see Evenhuis et al. 2015: 268) [North America].

ECTOPHASIOPSIS Townsend, 1915e: 439. Type species: *Ectophasiopsis chilensis* Townsend, 1915 (= *Trichopoda arcuata* Bigot, 1876), by original designation [Chile]. **Syn. nov.**

EUTRICHOPODOPSIS Blanchard, 1966a: 81. Type species: *Eutrichopodopsis funebris* Blanchard, 1966 (= *Musca pennipes* Fabricius, 1781), by original designation [Argentina].

ECTOPHASIOPS. Incorrect subsequent spelling of *Ectophasiopsis* Townsend, 1915 (Sabrosky 1950: 361).

TRICHOPODOSIS. Incorrect subsequent spelling of *Trichopodopsis* Townsend, 1913 (Mallea et al. 1977: 21, 23).

Note: *Ectophasiopsis* was recognised as a valid genus with a single species until recently revised by Dios and Nihei (2017), resulting in an increase in the number of species to three (with only the original species, *E. arcuata*, known from Chile). The species of *Ectophasiopsis*, *Trichopoda* (*sensu* Dios and Nihei 2020) and *Eutrichopoda* Townsend, 1908 (*sensu* Dios and Nihei 2016) all share a common habitus of black or black and yellow bodies and wings, and a row of long and distinctive “feather-like” setae on the hind tibia. A key to separate these three genera was given in Dios and Nihei (2017: 4) but the phylogenetic relationships among them have yet to be studied. Possibly all three taxa should be combined under one genus, for which the name *Trichopoda* would apply, but we have only explored the relationship between *Ectophasiopsis* and *Trichopoda*. Single legs of more than 50 CNC specimens of *Trichopoda* species from Canada, United States and Costa Rica and one Chilean specimen of *E. arcuata* (CNC487602, figs 30–31 in Stireman et al. 2016: 37) were sent to the Biodiversity Institute of Ontario (BIO) at the University of Guelph for DNA barcoding of the COI gene. A neighbor-joining tree clustered *E. arcuata* among ca. 40 samples of *T. pennipes* (Fabricius, 1781) and undetermined Costa Rican *Trichopoda*. These were all assigned to the same BIN (Barcode Index Number), BOLD:AAD9027, except for a few Costa Rican specimens. Sis-

ter to this clade was one consisting of three *Trichopoda indivisa* Townsend, 1897 and five *Trichopoda plumipes* (Fabricius, 1805), with each species in its own BIN. These last two species are members of the subgenus *Trichopoda*, whereas *T. pennipes* belongs to subgenus *Galactomyia*. We accept that the members of the BIN to which *E. arcuata* and *T. pennipes* belong are closely related, but we suspect from the morphological diversity within the sampled group that several or more species are involved and more sensitive molecular analyses may be needed to resolve them. For the present we restrict our taxonomic changes to the synonymy of *Ectophasiopsis* with *Trichopoda* and the assignment of *E. arcuata* and its allies *E. gradata* and *E. ypiranga* to subgenus *Galactomyia*.

References: Aldrich (1934: 2, 11), *Ectophasiopsis* in key to Patagonian genera, taxonomic notes; Townsend (1936b: 53), diagnosis of adults and immatures of Phasiini and key to genera (including *Ectophasiopsis*); Townsend (1938: 47), redescription of *Ectophasiopsis*; Sabrosky (1950: 361), *Ectophasiopsis* (as “*Ectophasiops*”) moved to Trichopodini; Dios and Nihei (2017), revision of *Ectophasiopsis*.

arcuata (Bigot, 1876).—Neotropical: South America (Argentina, Chile). Australasian & Oceanian: Polynesia (Easter Island, introduced). **Comb. revied.**

Trichopoda arcuata Bigot, 1876: 397. Lectotype male (NHMUK), by designation of Dios and Nihei (2017: 6). Type locality: Chile.

Ectophasiopsis chilensis Townsend, 1915e: 440. Holotype, unspecified sex [female, see note] (USNM). Type locality: Chile.

Note: Townsend (1915e: 440) described *Ectophasiopsis chilensis* from “one female and two males” and designated one of them as “Holotype.—Cat. No. 19460, U.S.N.M.”. This was a valid designation of a holotype even though Townsend did not specify which specimen it was. Townsend (1938: 47) later cited the holotype as the single female (as “Ht female”). The name-bearing type does not comprise all three of the original specimens as inferred by Dios and Nihei (2017: 5, as “Syntypes, two ♂♂ and one ♀”).

References: Aldrich (1934: 12), synonymy, redescription, taxonomic notes; Wolcott (1948: 471), introduced to Puerto Rico but not established; Verbeke (1962: 121, etc.), description of male terminalia; Cortés (1979: 79), first record from Argentina; Ripa et al. (1995: 432), introduced to Easter Island; Gramajo (1998: 92), cited as first record from Argentina but preceded by Cortés (1979); Stireman et al. (2016: 37), habitus images; Dios and Nihei (2017: 5, 7), in key, redescription, figures.

gradata Wiedemann, 1830.—Not Chile [Argentina, Brazil, Uruguay]. **Comb. revied.**

Trichopoda gradata Wiedemann, 1830: 275. Lectotype female (NHMW), by fixation of Dios and Nihei (2017: 10) (examination of “Holotype ♀” from Brazil in NHMW is regarded as a lectotype fixation). Type locality: Brazil.

Trichopodopsis incognita Blanchard, 1966a: 62. Holotype female (probably lost, Dios and Nihei 2017: 10). Type locality: Argentina, La Rioja [province].

Trichopodopsis argentinensis Blanchard, 1966a: 65. Holotype male (INTA). Type locality: Argentina, Córdoba [province].

Trichopodopsis christensenii Blanchard, 1966a: 78. Holotype male (INTA). Type locality: Argentina, Buenos Aires, José C. Paz.

Note: The relative priority of *Trichopodopsis incognita* Blanchard, 1966, *Trichopodopsis argentinensis* Blanchard, 1966 and *Trichopodopsis christenseni* Blanchard, 1966, when the three are treated as synonyms, was established by Liljesthrom (1992: 57), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999).

References: Liljesthrom (1992: 57), synonymy of *Trichopodopsis christenseni* and *T. incognita* with *T. argentinensis*, as species of *Trichopoda*; Dios and Nihei (2017: 10), synonymy of *T. argentinensis* and its two synonyms with *T. gradata*, reassigned to *Ectophasiopsis*, redescription, figures.

ypiranga (Dios & Nihei, 2017).—Not Chile [Argentina, Brazil]. **Comb. nov.**

Ectophasiopsis ypiranga Dios & Nihei, 2017: 18. Holotype male (FIOC). Type locality: Brazil, São Paulo, São Paulo, Ipiranga [as “Ypiranga”].

Note: Information about an intended paratype of *Ectophasiopsis ypiranga* was inadvertently removed from the manuscript of Dios and Nihei (2017) prior to publication (pers. comm., R. de V.P. Dios). It was the only specimen of *E. ypiranga* examined from Argentina and would have explained the inclusion of Argentina in the stated distribution and the dot on the distribution map. The specimen is a male in MNHN with the following data: “Museum Paris / Chaco de Santiago / del Estero / bords du rio Salado / la Palisa del Bracho / 25 kil. N. O. d’Icaño / E. R. Wagner 1909” (details courtesy of R. de V.P. Dios).

Subgenus *TRICHOPODA* Berthold, 1827

TRICHOPODA Berthold, 1827: 508 (as “*Trichopode*” (vernacular) by Latreille 1825: 498, name first latinised in Berthold’s German translation of Latreille (1825); see Sabrosky 1950: 366). Type species: *Thereva plumipes* Fabricius, 1805, by subsequent designation of Coquillett (1910: 616) [United States].

POLISTOMYIA Townsend, 1908: 132. Type species: *Trichopoda trifasciata* Loew, 1863 (= *Thereva plumipes* Fabricius, 1805), by original designation [United States].

THICHOPODA. Incorrect subsequent spelling of *Trichopoda* Berthold, 1827 (Guimarães 1971: 7).

TRICHIOPODA. Incorrect spelling of *Trichopoda* Berthold, 1827 (e.g., Latreille 1829: 512; Coquillett 1910: 616; Townsend 1913b: 147; see Sabrosky 1999: 313).

References: Townsend (1913b: 147), synonymy of *Polistomyia* with *Trichopoda* (as “*Trichiopoda*”), taxonomic notes; Townsend (1915g: 122), *Polistomyia* reinstated as valid genus, nomenclatural and taxonomic notes; Aldrich (1931: 3), synonymy of *Polistomyia* with “*Trichiopoda*”.

There are no Chilean species in this subgenus.

Tribe LEUCOSTOMATINI

Genus *LEUCOSTOMA* Meigen, 1803

LEUCOSTOMA Meigen, 1803: 279. Type species: *Ocyptera simplex* Fallén, 1815, by subsequent monotypy of Meigen (1824: 234) [Sweden].

PSALIDA Rondani, 1856: 76. Type species: *Psalida leucostoma* Rondani, 1856 (as “*Tachina Leucostoma* Mgn.”) (= *Ocyptera simplex* Fallén, 1815), by original designation (see O'Hara et al. 2011: 152) [Italy].

SIPHOPSALIDA Townsend, 1915e: 439. Type species: *Siphopsalida meridionalis* Townsend, 1915, by original designation [Peru].

CYCLODIONAEA Townsend, 1915h: 233. Type species: *Cyclodionaea acuminata* Townsend, 1915 (= *Musca aterrima* Villers, 1789), by original designation [United States].

PARADIONAEA Townsend, 1916a: 631. Type species: *Leucostoma atra* Townsend, 1891 (= *Ocyptera simplex* Fallén, 1815), by original designation [United States].

NEOPSALIDA Townsend, 1916a: 632. Type species: *Leucostoma neomexicana* Townsend, 1892 (= *Musca aterrima* Villers, 1789), by original designation [United States].

References: Coquillett (1910: 561, 595), type species of *Leucostoma* and *Psalida* (with latter in synonymy with former); Aldrich (1934: 3, 28), in key to Patagonian genera, synonymy, taxonomic notes; Townsend (1936b: 77), diagnosis of adults and immatures of Leucostomatini and key to genera (including *Cyclodionaea*, *Leucostoma*, *Neopsalida*, *Paradionaea* and *Siphopsalida*); Townsend (1936c: 280), *Psalida* as synonym of *Leucostoma*; Townsend (1938: 185, 189, 191, 192, 196), redescrptions of *Cyclodionaea*, *Leucostoma* (with *Psalida* in synonymy), *Neopsalida*, *Paradionaea* and *Siphopsalida*; Reinhard (1956), revision of New World species; Cortés and Campos (1971: 21, 1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions.

aterrimum (Villers, 1789).—Neotropical: Greater Antilles (Puerto Rico), Middle America (Mexico), South America (Argentina, Chile). Nearctic: Canada, United States. Palearctic: Europe. Australasian & Oceanian: Hawaii (immigrant).

Musca aterrima Villers, 1789: 548. Lectotype male (MNHN, see note), by fixation of Townsend (1932a: 33) (examination of “Male Ht” from Europe in MNHN is regarded as a lectotype fixation). Type locality: Europe.

Leucostoma neomexicana Townsend, 1892c: 169. Holotype male (SEMC, Byers et al. 1962: 175). Type locality: USA, New Mexico, Las Cruces.

Cyclodionaea acuminata Townsend, 1915h: 234. Holotype female (USNM). Type locality: USA, California, Santa Clara County.

Notes: The lectotype of *Musca aterrima* is not among the types currently listed in the online MNHN database but is assumed to be in the Muséum based on its study there by Townsend (1932a: 33).

Bezzi and Stein (1907: 327) listed both *L. aterrimum* and *L. simplex* as valid species in Europe and both species were recognised from the Americas in Reinhard's (1956) revision of *Leucostoma*. Subsequent catalogues in the Americas (Sabrosky and Arnaud 1965: 976; Guimarães 1971: 17; O'Hara and Wood 2004: 224) also recognised both species. However, Herting (1984: 174) treated *M. aterrima* as a questionable synonym of *L. simplex* and later Herting and Dely-Draskovits (1993: 420) removed the questionable status and listed the names as synonyms but continued to treat *L. simplex* as the valid name even though it is the junior synonym. Subsequent authors in Europe have recognised only *L. simplex* but the two names are in use

in the Americas for two broadly distributed species, both recorded here from Chile. Preliminary DNA barcoding by JEOH of seven specimens in CNC grouped four as *L. simplex* from Czech Republic (CNC DIPTERA 161997, CNC DIPTERA 161998) and New Mexico, USA (CNC DIPTERA 104203, CNC DIPTERA 104205) and three as *L. aterrimum* from Ontario, Canada (CNC602375) and Chile (CNC487609, CNC487610). What this signifies is not clear and further study is needed to determine if: 1) the two names are correctly applied in the Americas (and the names are thus not synonyms), 2) one or both names are misapplied in the Americas (and one or both of the species must be given a different available or new name), or 3) the names *L. aterrimum* and *L. simplex* are truly synonyms as currently treated in Europe and the valid name is *L. aterrimum* according to the *Code* (ICZN 1999).

References: Aldrich (1934: 29), synonymy, diagnosis, taxonomic notes, first records from Argentina and Chile; Reinhard (1956: 160), synonymy, redescription; Sabrosky and Arnaud (1965: 976), distribution including Mexico; Guimarães (1971: 17), distribution including Puerto Rico; Nishida (1992: 121), recorded from Hawaii as an immigrant (i.e., not purposely introduced).

simplex (Fallén, 1815).—Neotropical: South America (Argentina, Chile). Nearctic: Canada, United States. Palearctic: Central Asia, China [Pal.], Europe, Kazakhstan, Mongolia, Russia, Transcaucasia. Afrotropical: Cape Verde, Sierra Leone. Australasian & Oceanian: Australia, Hawaii (immigrant).

Ocyptera simplex Fallén, 1815: 240. Holotype female [not syntypes of both sexes as cited by Herting 1984: 174] (NHRS). Type locality: Sweden, Småland, Kalmar Län.

Psalida leucostoma Rondani, 1856: 76 (as “*Tachina Leucostoma* Mgn.”, see O’Hara et al. 2011: 152). Type(s), female (not located). Type locality: Italy.

Leucostoma atra Townsend, 1891: 380. Holotype male (SEMC, Byers et al. 1962: 174). Type locality: USA, Illinois, Carlinville.

Note: The identity of *Leucostoma simplex* is discussed above under *L. aterrimum*.

References: Aldrich (1934: 28), diagnosis, taxonomic notes, first record from Chile (as *Leucostoma atra*); Reinhard (1956: 159), synonymy of *Leucostoma atra* with *Ocyptera simplex*, redescription; Cortés and Hichins (1969: 40), Chilean records (as *Leucostoma ater*); Cortés and Campos (1971: 36), *Leucostoma simplex* accepted as the valid name with *Leucostoma atra* in synonymy; Cortés (1979: 80), first record from Argentina; Nishida (1992: 121), recorded from Hawaii as an immigrant (i.e., not purposely introduced).

Genus *PERIOSTOMA* Cortés, 1986

PERIOSTOMA Cortés, 1986: 145. Type species: *Periostoma flabellatum* Cortés, 1986, by original designation [Chile].

Reference: Cortés (1986: 142), in key to tachinid genera of Aysén and Magallanes regions.

flabellatum Cortés, 1986.—Neotropical: South America (Chile).

Periostoma flabellatum Cortés, 1986: 145. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Ultima Esperanza, Parque Nacional Torres del Paine, Laguna Amarga.

Tribe PHASIINI

Genus *PHASIA* Latreille, 1804

- PHASIA** Latreille, 1804: 195. Type species: *Conops subcoleoptratus* Linnaeus, 1767, by subsequent monotypy of Latreille (1805: 379); see rulings by ICZN (1970, 2006) [Sweden].
- ALOPHORA** Robineau-Desvoidy, 1830: 293. Type species: *Syrphus hemipterus* Fabricius, 1794, by subsequent designation of Robineau-Desvoidy (1863b: 226, as “*Thereva hemiptera* de Fabricius”) [United Kingdom].
- HYALOMYA** Robineau-Desvoidy, 1830: 298. Type species: *Phasia semicinerea* Meigen, 1824 (= *Phasia pusilla* Meigen, 1824), by subsequent designation of Westwood (1840: 140) [probably Germany].
- HYALOMYIA** Macquart, 1834: 69 [also 1834: 205]. Unjustified emendation of *Hyalomya* Robineau-Desvoidy, 1830 (see Evenhuis et al. 2010: 90).
- ALLOPHORA** Mik, 1894: 49. Unjustified emendation of *Alophora* Robineau-Desvoidy, 1830 (see Evenhuis et al. 2010: 36).
- PARAPHORANTHA** Townsend, 1915b: 20. Type species: *Alophora grandis* Coquillett, 1897, by original designation [United States].
- PHORANTHELLA** Townsend, 1915b: 23. *Nomen nudum* (by ruling of ICZN 1954: 311).
- ALOPHORELLOPSIS** Townsend, 1927a: 209. Type species: *Alophorellopsis capitata* Townsend, 1927, by original designation [Brazil].
- EPAULOPHASIA** Townsend, 1934a: 207. Type species: *Epaulophasia officialis* Townsend, 1934, by original designation [Brazil].
- HEYNEOPHASIA** Townsend, 1934a: 208. Type species: *Heyneophasia heynei* Townsend, 1934, by original designation [Costa Rica].
- XANTHOTRICHIVUS** Townsend, 1934a: 209. Type species: *Xanthotrichius xenos* Townsend, 1934, by original designation [Brazil].
- XIPHOPHASIA** Townsend, 1937a: 116. Type species: *Xiphophasia ushpayacua* Townsend, 1937, by monotypy [Peru].
- TRICHOPHASIA** Townsend, 1939b: 447 (junior homonym of *Trichophasia* Swainson, 1839). Type species: *Trichophasia transita* Townsend, 1939, by original designation [Brazil].
- PARAPHASIANA** Townsend, 1940b: 889. Type species: *Paraphasiana dysderci* Townsend, 1940 (junior secondary homonym of *Euphorantha dysderci* Townsend, 1938; = *Phasia aurodysderci* Nihei & Dios, 2016), by original designation [Brazil].
- ANDROEURYOPS** Beneway, 1961: 44. Type species: *Hyalomyia ecitonis* Townsend, 1897, by original designation [Mexico].

Note: Herting (1984: 168) designated *Conops subcoleoptrata* Linnaeus, 1767 as the type species of *Thereva* Fabricius, 1798, a junior homonym of *Thereva* Latreille, 1796 (Diptera, Therevidae). Later, an application to the International Commission on Zoological Nomenclature (Holston et al. 2003) resulted in the placement of the name *Thereva* Fabricius, 1798 on the Official Index of Rejected and Invalid Generic Names in Zoology (ICZN 2006).

References: Coquillett (1910: 505, 553, 587), type species of *Alophora*, *Hyalomyia* and *Phasia* (with first two in synonymy with *Phasia*); Aldrich (1934: 2, 13), in key to Patagonian genera, synonymy, taxonomic notes, key to four Patagonian species (as “*Hyalomyia*” Robineau-Desvoidy); Townsend (1936b: 53), diagnosis of adults and immatures of Phasiini and key to genera (including *Alophorellopsis*, *Epaulophasia*, *Heyneophasia*, *Hyalomyia*, *Paraphoranthia*, *Phasia*, *Phoranthella* and *Xanthotrichius*; Townsend (1936c: 271), *Alophora* as synonym of *Phasia*; Townsend (1938: 36, 50, 56, 57, 64, 65, 68, 73, 74), redescription of *Alophorellopsis*, *Epaulophasia*, *Heyneophasia*, *Hyalomyia*, *Paraphoranthia*, *Phasia* (with *Alophora* in synonymy), *Phoranthella*, *Xanthotrichius* and *Xiphophasia*; Sabrosky and Arnaud (1965: 968), synonymy of *Alophorellopsis* with *Hyalomyia* Robineau-Desvoidy; Herting (1974: 37, 1984: 170), synonymy of *Hyalomyia* with *Phasia* Latreille; Cortés (1986: 142), in key to tachinid genera of Aysén and Magallanes regions (as *Hyalomyia*); Wood (1987: 1258), synonymy including *Hyalomyia*, *Paraphoranthia* and *Phoranthella* with *Phasia*; O’Hara and Wood (1998: 756, 765), review of synonymy of Wood (1987); Sun and Marshall (2003: 19), synonymy of *Androeuryps*, *Epaulophasia*, *Heyneophasia*, *Paraphasiana*, *Trichophasia*, *Xanthotrichius* and *Xiphophasia* with *Phasia* Latreille.

chilensis (Macquart, 1851).—Neotropical: Middle America (Mexico), South America (Argentina, Brazil, Chile, Peru, Uruguay, Venezuela). Nearctic: United States.

Hyalomyia chilensis Macquart, 1851: 189 [also 1851: 216]. Lectotype male (MNHN, see note), by fixation of Aldrich (1934: 14) (examination of “type, a male” from Chile in MNHN is regarded as a lectotype fixation). Type locality: Chile.

Paraphoranthia peruviana Townsend, 1936a: 489. Syntypes, 2 males and 2 females (USNM). Type locality: Peru, La Libertad, Pacasmayo, Jequetepeque.

Paraphoranthia dimidiata Townsend, 1937b: 318. Syntypes, 1 male and 1 female (USNM). Type localities: Brazil, São Paulo, Tietê and Campinas.

Paraphoranthia pollinosa Brooks, 1945: 660. Holotype male (MCZ). Type locality: USA, Maryland, Chesapeake Beach.

Paraphoranthia auricaudata Brooks, 1945: 661. Holotype male (CNC). Type locality: USA, Oregon, Milton.

Note: The online MNHN database records a male holotype in the Macquart collection for *Hyalomyia chilensis* (number MNHN-ED-ED8382) based on a holotype determination label that DMW attached to the specimen in 1985. Macquart did not restrict the name-bearing type to a single specimen and the lectotype fixation of Aldrich (1934: 14) is accepted here [see Recommendation 73F of the *Code* (ICZN 1999), “Avoidance of assumption of holotype”].

References: Aldrich (1934: 14), redescription, taxonomic notes; Blanchard (1940: 224), first record from Argentina; Berry (1951: 339, 340), first record from Peru, figures of larval cephaloskeleton and puparium; Cortés (1963: 249), notes on name-bearing type of *Hyalomyia chilensis*; Sun and Marshall (2003: 158, 159, 165), in key, synonymy of *Paraphoranthia peruviana*, *Paraphoranthia dimidiata*, *Paraphoranthia pollinosa* and *Paraphoranthia auricaudata* with *Hyalomyia chilensis*, redescription, distribution.

curvipes (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Hyalomyia curvipes Aldrich, 1934: 16. Holotype, unspecified sex [male, examined by DMW] (NHMUK). Type locality: Chile, Metropolitana de Santiago, Santiago, Santiago.

Note: The type locality of *Hyalomyia curvipes* was given as “Santiago” in Chile, which could be interpreted as either the city or province of that name. Cortés and Hichins (1969: 37) cited the former as the type locality (as “Santiago (Santiago)”) and we follow this interpretation.

Reference: Cortés (1948: 122), first record from Argentina.

glauc (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Hyalomyia glauca Aldrich, 1934: 15. Holotype male (USNM). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

Reference: Cortés (1946: 173), first record from Chile.

metallica (Aldrich, 1934).—Neotropical: South America (Chile).

Hyalomyia metallica Aldrich, 1934: 15. Holotype female (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangué.

Tribe STRONGYGASTRINI

Genus **STRONGYGASTER** Macquart, 1834

STRONGYGASTER Macquart, 1834: 75 [also 1834: 211]. Type species: *Tachina globula* Meigen, 1824, by monotypy [Europe].

CLISTOMORPHA Townsend, 1892a: 79. Type species: *Clistomorpha hyalomoides* Townsend, 1892 (= *Hyalomyia triangulifera* Loew, 1863), by original designation [United States].

HYALOMYODES Townsend, 1893: 429. Type species: *Hyalomyodes weedii* Townsend, 1893 (= *Hyalomyia triangulifera* Loew, 1863), by monotypy [United States].

HYALOMYIODES. Incorrect subsequent spelling of *Hyalomyodes* Townsend, 1893 (Verbeke (1962: 118).

References: Coquillett (1910: 525, 553), type species of *Clistomorpha* (as synonym of *Elizeta* Rondani, 1856) and *Hyalomyodes*; Curran (1927: 297), key, synonymy of *Hyalomyodes* with *Clistomorpha*; Aldrich (1934: 2, 17), in key to Patagonian genera, taxonomic notes (as *Clistomorpha* with *Hyalomyodes* in synonymy); Townsend (1936b: 80), diagnosis of adults and immatures of Strongygastrini and key to genera (including *Clistomorpha*, *Hyalomyodes* and *Strongygaster*); Townsend (1938: 200, 201, 205), redescrptions of *Clistomorpha*, *Hyalomyodes* and *Strongygaster*; Brooks (1942: 140, 142), revisions of *Clistomorpha* and *Hyalomyodes*; Cortés (1986: 142), in key to tachinid genera of Aysén and Magallanes regions (as *Hyalomyodes*); Wood (1987: 1260), synonymy of *Clistomorpha* and *Hyalomyodes* with *Strongygaster*; O'Hara and Wood (1998: 754, 755, 766), review of synonymy of Wood (1987).

triangulifera (Loew, 1863).—Neotropical: Middle America (Mexico), South America (Argentina, Chile). Nearctic: Canada, United States.

Hyalomyia triangulifera Loew, 1863: 319. Type(s), female [not male as published] (2 females in MCZ). Type locality: USA, New York.

Clistomorpha hyalomoides Townsend, 1892a: 80. Holotype female [not male as published, Townsend 1938: 200 and verified by DMW] (SEMC, Byers et al. 1962: 173 [as male, in error]). Type locality: USA, New York, Ithaca.

Hyalomyodes weedii Townsend, 1893: 430. Lectotype male (SEMC, Byers et al. 1962: 174 [as “5 ♂? syntypes”]), by fixation of Townsend (1938: 201) (mention of “Ht male” from Hanover in SEMC is regarded as a lectotype fixation for the single male syntype from that locality). Type locality: USA, New Hampshire, Hanover.

triangulifer. Incorrect subsequent spelling of *triangulifera* Loew, 1863 (e.g., Cortés and Hichins 1969: 37; Guimarães 1971: 18; González 1992b: 179).

Note: The mention of a “Ht” for *Hyalomyia triangulifera* from New York in MCZ by Townsend (1938: 201) is not accepted as a lectotype fixation because the specimen in question is not distinguishable from the other specimen in the type series.

References: Aldrich (1934: 18), synonymy, redescription, first record from Chile; Verbeke (1962: 118, pl. XIV fig. 9), description and figure of male terminalia; Cortés (1979: 79), taxonomic notes, first record from Argentina; O’Hara and Wood (2004: 306), synonymy including *Clistomorpha hyalomoides* with *Hyalomyia triangulifera*.

Subfamily TACHININAE

Tribe ERNESTIINI

Genus *LINNAEMYA* Robineau-Desvoidy, 1830

References: Coquillett (1910: 515, 561, 565, 569), type species of *Bonnetia*, *Linnaemya*, *Marshamia* and *Micropalpis* (with *Marshamia* and *Micropalpis* in synonymy with *Bonnetia*); Townsend (1936b: 190, 197), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Marshamia*), diagnosis of adults and immatures of Linnaemyini and key to genera (including *Bonnetia*, *Gymnochaetopsis* and *Linnaemya*); Townsend (1936c: 106, 277), diagnosis of adults and immatures of Phoroceratini and key to genera (*Ophina* omitted from key), “*Micropalpus*” as synonym of *Bonnetia*; Townsend (1939a: 198, 221, 232, 238), redescrptions of *Marshamia*, *Bonnetia* (with “*Micropalpus*” in synonymy), *Gymnochaetopsis* and *Linnaemya*; Townsend (1940a: 140), redescription of *Ophina*.

Subgenus *LINNAEMYA* Robineau-Desvoidy, 1830

LINNAEMYA Robineau-Desvoidy, 1830: 52. Type species: *Linnaemya silvestris* Robineau-Desvoidy, 1830 (= *Tachina vulpina* Fallén, 1810), by subsequent designation of Robineau-Desvoidy (1863a: 131) (as *vulpina*, with *silvestris* in synonymy) [France].

BONNETIA Robineau-Desvoidy, 1830: 55. Type species: *Bonnetia oenanthis* Robineau-Desvoidy, 1830 (= *Tachina comta* Fallén, 1810), by subsequent designation of Townsend (1916b: 6) [France].

MARSHAMIA Robineau-Desvoidy, 1830: 57. Type species: *Marshamia analis* Robineau-Desvoidy, 1830 (junior secondary homonym of *Linnaemya analis* Robineau-Desvoidy, 1830; = *Tachina comta* Fallén, 1810), by subsequent designation of Townsend (1916b: 7) [United States].

MICROPALPIS Macquart, 1834: 180 [also 1834: 316]. Type species: *Tachina vulpina* Fallén, 1810, by subsequent designation of d'Orbigny (1846: 200, as "*Micropalpus*") (see Evenhuis and Thompson 1990: 237, as "*Micropalpus*") [Sweden].

LINNEMYIA Macquart, 1835: 81. Unjustified emendation of *Linnaemya* Robineau-Desvoidy, 1830 (see Evenhuis et al. 2010: 100).

LINNAEMYIA Aldrich, 1905: 451. Unjustified emendation of *Linnaemya* Robineau-Desvoidy, 1830 (see Evenhuis et al. 2010: 100).

MICROPALPUS. Incorrect subsequent spelling of *Micropalpis* Macquart, 1834 (Macquart 1835: 80).

References: Cortés and Campos (1971: 24, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions (as *Bonnetia* Robineau-Desvoidy).

comta (Fallén, 1810).—Neotropical: Middle America (Honduras, Mexico), South America (Chile, Peru). Nearctic: Canada, United States. Palearctic: Central Asia, China [Pal.], Europe, Kazakhstan, Korean Peninsula, Middle East, Russia, Transcaucasia. Oriental: China [Orien.], Taiwan. Misidentified from the Afrotropical Region (O'Hara and Cerretti 2016: 193–194).

Tachina comta Fallén, 1810: 277. Lectotype female (NHRS), by fixation of Townsend (1939a: 222) (mention of "Ht male" from Sweden in NHRS is regarded as a lectotype fixation of the single type specimen, a female, in NHRS; examined by JEOH). Type locality: Sweden.

Linnaemya distincta Robineau-Desvoidy, 1830: 54. Lectotype female (MNHN, see note), by fixation of O'Hara and Wood (2004: 242) (mention of "holotype female" from Philadelphia in MNHN is regarded as a lectotype fixation; examined by DMW). Type locality: USA, Pennsylvania, Philadelphia.

Linnaemya analis Robineau-Desvoidy, 1830: 54. Holotype, unspecified sex (MNHN or lost, see note). Type locality: France, Maine-et-Loire, Angers.

Marshamia analis Robineau-Desvoidy, 1830: 58 (junior secondary homonym of *Linnaemya analis* Robineau-Desvoidy, 1830; = *Micropalpus piceus* Macquart, 1835). Lectotype female (MNHN, see note), by fixation of Townsend (1939a: 198) (mention of "Ht male" from "Carolina" in MNHN is regarded as a lectotype fixation of the single type specimen, a female, in MNHN; examined by DMW). Type locality: USA, "Caroline" (i.e., North and South Carolina).

Marshamia nigripes Robineau-Desvoidy, 1830: 58. Lectotype female (MNHN, see note), by fixation of O'Hara and Wood (2004: 242) (mention of "holotype female" from "Carolina" in MNHN is regarded as a lectotype fixation; examined by DMW). Type locality: USA, "Caroline" (i.e., North and South Carolina).

Micropalpus piceus Macquart, 1835: 84 (*nomen novum* for *Marshamia analis* Robineau-Desvoidy, 1830, see note).

compta. Incorrect subsequent spelling of *comta* Fallén, 1810 (Meigen 1824: 262; numerous subsequent authors).

Notes: The relative priority of *Linnaemya analis* Robineau-Desvoidy, 1830 and *Marshamia analis* Robineau-Desvoidy, 1830, when both are placed in the same genus, was established by Macquart (1835: 84), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999). Macquart (1835: 84) gave the junior homonym, *Marshamia analis*, the new name *Micropalpus piceus* Macquart, 1835.

The single specimen of *Tachina comta* Fallén in NHRS (a female, examined by JEOH), was treated as the holotype by O'Hara and Wood (2004: 241).

The online MNHN database records a female holotype in the Macquart collection for *Linnaemya distincta* (number MNHN-ED-ED7203, mistakenly recorded as *Micropalpus distinctus*) based on a holotype determination label that DMW attached to the specimen in 1985. Robineau-Desvoidy did not restrict the name-bearing type to a single specimen and the lectotype fixation of O'Hara and Wood (2004: 242) is accepted here [see Recommendation 73F of the *Code* (ICZN 1999), "Avoidance of assumption of holotype"].

There is no record for *Linnaemya analis* Robineau-Desvoidy in the online MNHN database. There are also no records in the database for *Marshamia analis* Robineau-Desvoidy and *Marshamia nigripes* Robineau-Desvoidy, but the lectotypes of both names are assumed to be in the Muséum based on their examination there by DMW.

References: Giglio-Tos (1894: 482), first record from Mexico; Coquillett (1897: 87), synonymy of *Linnaemya distincta*, *Linnaemya analis*, *Marshamia analis*, *Marshamia nigripes* and *Micropalpus piceus* with *Tachina comta*; Cortés and Campos (1971: 59), head figure, first records from Chile and Peru; Cave (1992: 595), record from Honduras.

Subgenus *OPHINA* Robineau-Desvoidy, 1863

OPHINA Robineau-Desvoidy, 1863a: 298. Type species: *Ophina fulvipes* Robineau-Desvoidy, 1863 (= *Tachina picta* Meigen, 1824), by original designation [France].
GYMNOCHAETOPSIS Townsend, 1914a: 15. Type species: *Gymnochaetopsis analis* Townsend, 1914 (junior secondary homonym of *Linnaemya analis* Robineau-Desvoidy, 1830, see note), by original designation (see Evenhuis et al. 2015: 136) [Peru].

Note: The type species of *Gymnochaetopsis*, *G. analis* Townsend, 1914, is a junior secondary homonym of *Linnaemya analis* Robineau-Desvoidy, 1830, described from France. It is not renamed while *Linnaemya analis* is in synonymy with *Linnaemya comta* (Fallén, 1810).

References: Thompson (1963a: 433), redescription of *Gymnochaetopsis analis*, first record from Trinidad; Mesnil (1971: 1006, 1018), synonymy of *Gymnochaetopsis* with *Linnaemya* and treatment of the former as a subgenus; Shima (1986), assignment of *Gymnochaetopsis analis* to *Linnaemya* (*Ophina*).

There are no Chilean species in this subgenus.

Tribe GRAPHOGASTRINI

Reference: Andersen (1988: 48), treatment of the Graphogastrini, key to Palaearctic genera.

Genus *CLASTONEURA* Aldrich, 1934

CLASTONEURA Aldrich, 1934: 26. Type species: *Clastoneura brevicornis* Aldrich, 1934, by original designation [Argentina].

Note: Wood and Zumbado (2010: 1412) suggested in their note about *Steleoneura* Stein that “apparently, it [*Steleoneura*] is the sister genus of two Chilean genera, *Clastoneura* Aldrich and *Embiomyia* Aldrich; the latter may be congeneric with *Steleoneura*”. *Steleoneura* is a blondeline genus and we recognise it herein from Chile with *Embiomyia* in synonymy. We have examined males and females of *Clastoneura brevicornis* in CNC and believe it to be a graphogastrine and not a blondeline, as similarly interpreted by Guimarães (1971: 171) and Andersen (1988: 48–49, diagnosis and included genera of Graphogastrini).

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Clastoneura*); Townsend (1940a: 203), redescription.

brevicornis Aldrich, 1934.—Neotropical: South America (Argentina, Chile). (Fig. 6a)
Clastoneura brevicornis Aldrich, 1934: 27. Holotype male (NHMUK). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí.

Reference: Cortés (1967b: 11), taxonomic notes, first record from Chile.

Genus *CLASTONEURIOPSIS* Reinhard, 1939

CLASTONEURIOPSIS Reinhard, 1939: 68. Type species: *Clastoneuriopsis meralis* Reinhard, 1939, by original designation [United States].

References: Cortés (1986: 144, 156), in key to tachinid genera of Aysén and Magallanes regions, taxonomic affinities, diagnostic characters; Andersen (1988: 49), assigned to Graphogastrini.

magallanica Cortés, 1986.—Neotropical: South America (Chile).

Clastoneuriopsis magallanica Cortés, 1986: 156. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Última Esperanza, Sierra de Los Baguales, 600 m [ca. 50°47'S, 72°24'W].

Genus *PHYTOMYPTERA* Rondani, 1845

PHYTOMYPTERA Rondani, 1845: 32, 33. Type species: *Phytomyptera nitidiventris* Rondani, 1845 (= *Tachina nigrina* Meigen, 1824), by monotypy [Italy].

ELFIA Robineau-Desvoidy, 1849: 158. *Nomen nudum* (no description or included species).

ELFIA Robineau-Desvoidy, 1850: 190. Type species: *Actia cingulata* Robineau-Desvoidy, 1830, by subsequent designation of Robineau-Desvoidy (1863a: 672) [France].

LISPIDEA Coquillett, 1895b: 51. Type species: *Lispidea palpigera* Coquillett, 1895, by original designation [United States].

LISPIDEOSOMA Reinhard, 1943: 164. Type species: *Lispideosoma flavipes* Reinhard, 1943, by original designation [United States].

CAMPOSODES Cortés, 1967a: 4. Type species: *Camposodes evanescens* Cortés, 1967, by original designation [Chile]. **Syn. nov.**

IRWINIA Cortés, 1967a: 7. Type species: *Irwinia pollinosa* Cortés, 1967, by original designation [Chile].

LISPIDIA. Incorrect subsequent spelling of *Lispidea* Coquillett, 1895 (Vimmer and Soukup 1940a: 214).

Note: Cortés (1967a: 4, 7) described *Camposodes* and *Irwinia* as monotypic genera, each characterised by reduced and distinctive wing venation. They are, however, simply apomorphic forms of *Phytomyptera*, a genus in which the loss of wing veins is not unusual and has likely occurred independently in several lineages. Mesnil (1973: 1192) recognised this and synonymised *Irwinia* with *Phytomyptera*. We have examined a specimen of *C. evanescens* in CNC and it is also fundamentally *Phytomyptera*. The diagnostic features of *Phytomyptera* include a haired prosternum, a single setula at the base of wing vein R_{4+5} and lower proepimeral seta directed downward (Andersen 1988: 49; Wood and Zumbado 2010: 1372).

References: Coquillett (1910: 537, 562), type species of *Elfia* (as synonym of *Actia* Robineau-Desvoidy) and *Lispidea*; Aldrich (1934: 4, 5, 75), in key to Patagonian genera, synonymy, taxonomic notes, key to six Patagonian species (as *Lispidea*, in part); Townsend (1936c: 129, 274), diagnosis of adults and immatures of Actiini and key to genera (including *Lispidea* and *Phytomyptera*), *Elfia* as synonym of *Actia* Robineau-Desvoidy, 1830; Townsend (1940a: 232, 248), redescrptions of *Lispidea* and *Phytomyptera*; Sabrosky and Arnaud (1965: 1065), synonymy of *Lispidea* with *Elfia*; Cortés and Campos (1971: 20, 26, 1974: 112, 115, as *Camposodes* and *Lispidea*) and Cortés (1984: 378, 381, as *Camposodes* and *Elfia*), in keys to tachinid genera of Tarapacá and Antofagasta regions; Mesnil (1973: 1192), synonymy of *Irwinia* with *Phytomyptera*; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions (as *Elfia*); O'Hara (1985: 93), request for type species designation for *Actia* to separate the concepts of *Actia* and *Elfia*; ICZN (1987: 71), type species designation for *Actia*, thereby removing *Actia* as a senior synonym of *Elfia*; Wood (1987: 1220), synonymy of *Elfia*, *Lispidea* and *Lispideosoma* with *Phytomyptera*; Andersen (1988: 45, 49), in key to genera of Graphogastrini, synonymy, diagnosis; O'Hara and Wood (1998: 754, 755, 765), review of synonymy of Wood (1987).

atra (Aldrich, 1934).—Neotropical: South America (Chile).

Lispidea atra Aldrich, 1934: 78. Holotype female (USNM). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangué.

Reference: Cortés (1967b: 11), first description of male.

evanescens (Cortés, 1967).—Neotropical: South America (Argentina, Chile). **New record from Argentina. Comb. nov.**

Camposodes evanescens Cortés, 1967a: 4. Holotype male (EEAM). Type locality: Chile, Metropolitana de Santiago, Santiago, Maipú, Universidad de Chile, Estación Experimental Agronómica, Quebrada de La Plata, 510–550 m.

Note: The new combination for *Camposodes evanescens* is explained under the genus heading above. The new record from Argentina is based on four CNC specimens from three localities with the following data: [Santa Cruz], southeast of Lago Viedma, ca. 50°S, 72°W, 22.xii.1960, L. Peña (2 specimens); Jujuy, La Quiaca, 23.x.1968, 3500 m, L. Peña (1 specimen); Jujuy, 3 km north of Humahuaca, 3300 m, 22.x.1968, L. Peña (1 specimen).

frontalis (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Lispidea frontalis Aldrich, 1934: 80. Holotype female (NHMUK). Type locality: Argentina, Tierra del Fuego, Río Grande, Estancia Viamonte.

Reference: Cortés and Campos (1971: 97), first record from Chile.

interrupta (Aldrich, 1934).—Neotropical: South America (Chile).

Lispidea interrupta Aldrich, 1934: 79. Holotype female (USNM). Type locality: Chile, Los Lagos, Chiloé, Ancud.

Note: The suggestion by Andersen (1988: 46) that *Lispidea interrupta* “seems more likely belonging in Leskiini” was probably based on misidentified specimens.

pollinosa (Cortés, 1967).—Neotropical: South America (Chile).

Irwinia pollinosa Cortés, 1967a: 7. Holotype male (EEAM). Type locality: Chile, Coquimbo, Limarí, Pachingo, Parque Nacional Bosque Fray Jorge.

triangularis (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Lispidea triangularis Aldrich, 1934: 76. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Correntoso.

Reference: Cortés and Hichins (1969: 41), first record from Chile.

Genus *PLANOMYIA* Aldrich, 1934

PLANOMYIA Aldrich, 1934: 129. Type species: *Planomyia browni* Aldrich, 1934, by original designation [Chile].

PLANOMYIA. Incorrect subsequent spelling of *Planomyia* Aldrich, 1934 (Cortés 1967b: 11).

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Planomyia*); Townsend (1940a: 249), redescription; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; Andersen (1988: 49), in key to genera of Graphogastrini, characters given to separate this genus from the externally similar *Phytomyptera* Rondani.

browni Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Planomyia browni Aldrich, 1934: 129. Holotype female (NHMUK). Type locality: Chile, Biobío, Concepción, Concepción.

Note: The type locality of *Planomyia browni* was given as “Concepción” in Chile, which could be interpreted as either the city or province of that name. Cortés and Hichins (1969: 53) cited the former as the type locality (as “Concepción (Concepción)”) and we follow this interpretation. Seven paratypes of *P. browni* were collected from “So. Patagonia” by “B.

Brown" (Aldrich 1934: 130). The country of origin of these paratypes is interpreted here as Argentina based on the travels of the collector, paleontologist Barnum Brown.

Reference: Cortés (1967b: 11), first description of male.

vibrissata (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Lispidea vibrissata Aldrich, 1934: 78. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Correntoso.

References: Andersen (1988: 46), moved to *Planomyia*; González (1992b: 183), first record from Chile (in *Elfia* Robineau-Desvoidy).

Tribe LESKIINI

Genus *CLAUSICELLA* Rondani, 1856

CLAUSICELLA Rondani, 1856: 61. Type species: *Clausicella suturata* Rondani, 1856 (as "*Claus: Sutturata Mihi*"), by original designation (see O'Hara et al. 2011: 61) [Italy].

SIPHOAETIA Townsend, 1927a: 212. Type species: *Siphoactia charapensis* Townsend, 1927, by original designation [Peru]. **Syn. nov.**

Note: It is evident from the holotype of *Siphoactia charapensis* (examined by DMW) and head figure of *Siphoactia peregrina* (Cortés and Campos 1971: 68) that these two species are typical members of *Clausicella*, a genus not previously reported from South America. The genus can be recognised in part by the long proboscis and "membrane between lower facial margin and clypeus with pair of convex subtriangular sclerites" (Wood and Zumbado 2010: 1396). References: Coquillett (1910: 524), type species of *Clausicella*; Townsend (1936c: 129, 146), diagnosis of adults and immatures of Actiini and key to genera (including *Clausicella*), diagnosis of adults and immatures of Siphonini and key to genera (including *Siphoactia*; Townsend (1940a: 204, 290), redescription of *Clausicella* and *Siphoactia*; Cortés and Campos (1971: 25, 1974: 115) and Cortés (1984: 381), *Siphoactia* in keys to tachinid genera of Tarapacá and Antofagasta regions.

charapensis (Townsend, 1927).—Not Chile [Peru]. **Comb. nov.**

Siphoactia charapensis Townsend, 1927a: 357. Holotype female (USNM). Type locality: Peru, Cajamarca, Río Charapi [as "Rio Charape", ca. 5°25'S, 78°59'W].

peregrina (Cortés & Campos, 1971).—Neotropical: South America (Chile). **Comb. nov.**

Siphoactia peregrina Cortés and Campos, 1971: 67. Holotype female (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, km 31.

Genus *EPICORONIMYIA* Blanchard, 1940

EPICORONIMYIA Blanchard, 1940: 245. Type species: *Epigrimya mundelli* Blanchard, 1935 (as "*Epigrimya mundelli*"), by original designation [Argentina].

mundelli (Blanchard, 1935).—Neotropical: South America (Argentina, Chile).

Epigrymia mundelli Blanchard, 1935: 8. Holotype male (not located). Type locality: Argentina, Santiago del Estero [province or city].

Reference: Cortés (1976: 4), additional characters, first record from Chile.

Genus ***ORAEOSOMA*** Cortés, 1976

ORAEOSOMA Cortés, 1976: 8. Type species: *Oraeosoma proboscideum* Cortés, 1976, by original designation [Chile].

proboscideum Cortés, 1976.—Neotropical: South America (Chile).

Oraeosoma proboscideum Cortés, 1976: 10. Holotype male (MEUC). Type locality: Chile, Metropolitana de Santiago, Santiago, Pudahuel.

Genus ***SPATHIPALPUS*** Rondani, 1863

SPATHIPALPUS Rondani, 1863: 20 [also 1864: 20]. Type species: *Spathipalpus philippii* Rondani, 1863, by subsequent designation of Brauer and Bergenstamm (1893: 44 [also 1893: 132], as “*Spatipalpus* Rdi. Type: *Philippi* Rdi.”) (see O'Hara et al. 2011: 166) [Chile].

MACROPALPUS Rondani, 1863: 20 [also 1864: 20]. *Nomen nudum* (proposed in synonymy [with *Spathipalpus* Rondani, 1863] and not made available by subsequent usage before 1961) (see O'Hara et al. 2011: 111).

References: Aldrich (1934: 3, 31), in key to Patagonian genera, taxonomic notes; Townsend (1936c: 62, 277), diagnosis of adults and immatures of Leskiini and key to genera (including *Spathipalpus*), *Spathipalpus* as valid name for *Macropalpus*; Townsend (1940a: 237), redescription of *Spathipalpus* (with *Macropalpus* in synonymy); Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

philippii Rondani, 1863.—Neotropical: South America (Argentina, Chile).

Spathipalpus philippii Rondani, 1863: 21 [also 1864: 21]. Lectotype female (probably MZUF or lost), by fixation of Townsend (1939c: 237) (mention of “Ht female” from Valdivia in “Naples or Genoa” is regarded as a lectotype fixation). Type locality: Chile, Los Ríos, Valdivia, Valdivia.

? *Spathipalpus flavifrons* Rondani, 1863: 21 [also 1864: 21]. Type(s), ?male [described as female but possibly male, Aldrich 1934: 32 and Townsend 1939c: 239] (probably MZUF or lost). Type locality: Chile, Los Ríos, Valdivia, Valdivia. *philipii*. Incorrect subsequent spelling of *philippii* Rondani, 1863 (Henry 1987: 204). *philippi*. Incorrect subsequent spelling of *philippii* Rondani, 1863 (Brauer and Bergenstamm 1893: 44 [also 1893: 132]).

Note: The relative priority of *Spathipalpus philippii* Rondani, 1863 and *Spathipalpus flavifrons* Rondani, 1863, when the two are treated as synonyms, was established by Aldrich (1934: 32), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999). The synonymy of

these two names was questioned by Townsend (1939c: 239), Cortés (1951a: 65) and Guimarães (1971: 118) and has not been conclusively established.

References: Aldrich (1934: 32), taxonomic notes, head figure, first record from Argentina; Cortés (1951a: 62), first description of male.

Tribe MEGAPROSOPINI

References: Cortés (1945e: 150), key to the three Chilean genera here assigned to the Megaprosopini, treated as genera allied to *Trichoprosopus* Macquart; Cortés (1983), "Trichoprosopini" proposed as the sister group of the New Zealand tribe Proscissionini (as Occisorini) based in particular on the study of *Trichoceronia* Cortés and *Trichoprosopus* Macquart.

Genus *STUARDOMYIA* Cortés, 1945

STUARDOMYIA Cortés, 1945e: 157. Type species: *Stuardomyia crassiseta* Cortés, 1945, by original designation [Chile].

crassiseta Cortés, 1945.—Neotropical: South America (Argentina, Chile).

Stuardomyia crassiseta Cortés, 1945e: 158. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Reference: Cortés (1980: 105), first record from Argentina.

Genus *TRICHOCERONIA* Cortés, 1945

TRICHOCERONIA Cortés, 1945e: 150. Type species: *Trichoceronia thermitana* Cortés, 1945, by original designation [Chile].

THRICHOCERONIA. Incorrect subsequent spelling of *Trichoceronia* Cortés, 1945 (González (1992b: 183).

Reference: Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

latifrons (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Trichoprosopus latifrons Aldrich, 1934: 20. Holotype female (AMNH). Type locality: "South Patagonia" (interpreted as Argentina by Cortés and Hichins 1969: 60).

References: Cortés (1945e: 150), moved to *Trichoceronia* and partial redescription of female holotype; Cortés and Hichins (1969: 60), first record from Chile.

thermitana Cortés, 1945.—Neotropical: South America (Chile).

Trichoceronia thermitana Cortés, 1945e: 151. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Curacautín, Termas de Río Blanco.

Genus *TRICHOPROSOPUS* Macquart, 1844

TRICHOPROSOPUS Macquart, 1844: 70 [also 1844: 227]. Type species: *Trichoprosopus durvillei* Macquart, 1844, by original designation [Chile].

THRICHOPROSOPUS. Incorrect subsequent spelling of *Trichoprosopus* Macquart, 1844 (González 1992b: 183).

TRICHOPROSOPA. Incorrect subsequent spelling of *Trichoprosopus* Macquart, 1844 (Cortés 1963: 249, with note “*erratum pro Trichoprosopus*”).

References: Aldrich (1934: 2, 19), in key to Patagonian genera, taxonomic notes; Townsend (1936b: 121), diagnosis of adults and immatures of Trichoprosopini and key to genera (including *Trichoprosopus*); Townsend (1938: 300), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; Stireman et al. (2016: 38), habitus images of *Trichoprosopus* sp.

durvillei Macquart, 1844.—Neotropical: South America (Chile).

Trichoprosopus durvillei Macquart, 1844: 71 [also 1844: 228]. Lectotype male (MNHN, see note), by fixation of Townsend (1931a: 97) (examination of “Male Ht” from “Chile” in MNHN is regarded as a lectotype fixation). Type locality: Chile, Biobío, Concepción, Concepción.

Note: The online MNHN database records a male holotype in the Macquart collection for *Trichoprosopus durvillei* (number MNHN-ED-ED8374) based on a holotype determination label that DMW attached to the specimen in 1982. Macquart did not restrict the name-bearing type to a single specimen and the lectotype fixation of Townsend (1931a: 97) is accepted here [see Recommendation 73F of the *Code* (ICZN 1999), “Avoidance of assumption of holotype”].

The type locality of *Trichoprosopus durvillei* was given as “Concepcion” in Chile, which could be interpreted as either the city or province of that name. Cortés and Hichins (1969: 61) cited the former as the type locality (as “Concepción (Concepción)”) and we follow this interpretation.

References: Aldrich (1934: 20), redescription, taxonomic notes; Cortés (1945e: 154), redescription; Cortés (1963: 249), notes on name-bearing type in MNHN.

Tribe NEMORAEINI

Genus *XANTHOPHYTO* Townsend, 1916

XANTHOPHYTO Townsend, 1916a: 627. Type species: *Nemoraea labis* Coquillett, 1895, by original designation [United States].

References: Aldrich (1934: 4, 82), in key to Patagonian genera, synonymy, taxonomic notes; Townsend (1936b: 203), diagnosis of adults and immatures of Ernestiini and key to genera (including *Xanthophyto*); Townsend (1939a: 269), redescription.

erythropyga (van der Wulp, 1882).—Neotropical: South America (Argentina, Chile).

Nemoraea erythropyga van der Wulp, 1882: 83. Holotype male (RMNH). Type locality: Chile.

References: Aldrich (1934: 83), redescription, first description of female; Cortés (1973a: 101), taxonomic notes; Gramajo (1998: 96), first record from Argentina.

Tribe POLIDEINI

The concept of the Polideini and the North American members of the tribe were revised by O'Hara (2002). The Neotropical Polideini are not well understood and require significant revision at the generic and specific levels to better classify the fauna along phylogenetic lines and to accommodate numerous new species. The traditional classification of the Polideini is followed here pending a revision of the tribe.

Genus *ANDICESA* Koçak & Kemal, 2010

TRICHOPHOROPSIS Townsend, 1914a: 11. *Nomen nudum* (see Evenhuis et al. 2015: 267).

TRICHOPHOROPSIS Townsend, 1914b: 42 (junior homonym of *Trichophoropsis* Bonaparte, 1854). Type species: *Trichophoropsis puna* Townsend, 1914, by original designation [Peru].

ANDICESA Koçak & Kemal, 2010: 158 (*nomen novum* for *Trichophoropsis* Townsend, 1914).

ANICESA. Incorrect subsequent spelling of *Andicesa* Koçak & Kemal, 2010 (Evenhuis et al. 2015: 267).

References: Townsend (1936b: 190), diagnosis of adults and immatures of Cuphocerotini and key to genera (including *Trichophoropsis*), Townsend (1939a: 217), redescription of *Trichophoropsis*; Cortés and Campos (1971: 24, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions (as *Trichophoropsis*); González (1992a: 55, 63), in key to Chilean genera of "Cuphocerini", diagnosis, two new species (as *Trichophoropsis*).

bicolor (González, 1992).—Neotropical: South America (Chile).

Trichophoropsis bicolor González, 1992a: 64. Holotype male (UMCE). Type locality: Chile, Antofagasta, Antofagasta, Geyser del Tatio.

coscaroni (González, 1992).—Neotropical: South America (Chile).

Trichophoropsis coscaroni González, 1992a: 65. Holotype male (UMCE). Type locality: Chile, Ñuble, Diguillín, Termas de Chillán.

nitens (Townsend, 1914).—Neotropical: South America (Chile, Peru).

Trichophoropsis nitens Townsend, 1914b: 44. Syntypes, 3 males (USNM). Type locality: Peru, Junín, La Oroya, 12,250 ft.

Reference: Cortés and Campos (1971: 70), first record from Chile.

sabroskyi (Cortés & Campos, 1971).—Neotropical: South America (Argentina, Chile).

Trichophoropsis sabroskyi Cortés & Campos, 1971: 71. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, Rosario, 352 m (18°26'S, 70°06'W) (coordinates and elevation given on p. 11).

Reference: Cortés (1980: 106), first record from Argentina.

Genus *COMOPS* Aldrich, 1934

COMOPS Aldrich, 1934: 40. Type species: *Comops ruficornis* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936b: 190), diagnosis of adults and immatures of Cuphocerotini and key to genera (including *Comops*), Townsend (1939a: 177), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 55, 58), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

ruficornis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Comops ruficornis Aldrich, 1934: 40. Holotype, unspecified sex [male, examined by DMW] (NHMUK). Type locality: Argentina, Río Negro, eastern end of Lago Nahuel Huapí.

Enchomyia penai Cortés, 1967b: 16 (as “*peñai*”). Holotype female (EEAM). Type locality: Chile, Coquimbo, Choapa, Illapel, Hacienda Illapel, 2500–2800 m.

References: Guimarães (1971: 75), synonymy of *Enchomyia penai* with *Comops ruficornis*, citing “R. Cortés, *in litt.*”; Cortés (1973a: 99), taxonomic notes.

Genus *DELOBLEPHARIS* Aldrich, 1934

DELOBLEPHARIS Aldrich, 1934: 74. Type species: *Deloblepharis nigra* Aldrich, 1934, by original designation [Chile].

References: Townsend (1936b: 218), diagnosis of adults and immatures of Germariini and key to genera (including *Deloblepharis*); Townsend (1939a: 325), redescription; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

nigra Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Deloblepharis nigra Aldrich, 1934: 74. Holotype female (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangué.

Reference: Gramajo (1998: 95), first record from Argentina.

Genus *DESANTISODES* Cortés, 1973

DESANTISODES Cortés, 1973a: 102. Type species: *Desantisodes concinnum* Cortés, 1973, by original designation [Chile].

concinnum Cortés, 1973.—Neotropical: South America (Argentina, Chile).

Desantisodes concinnum Cortés, 1973a: 103. Holotype female (MEUC). Type locality: Chile, Maule, Curicó, Río Vergara, 2000–2300 m.

Reference: Cortés (1976: 5), first description of male, first record from Argentina.

Genus *DOLICHOSTOMA* Townsend, 1912

DOLICHOSTOMA Townsend, 1912b: 325. Type species: *Dolichostoma alpina* Townsend, 1912, by original designation [Peru].

ERIGONOPSIS Townsend, 1912b: 326. Type species: *Erigonopsis arequipae* Townsend, 1912, by original designation [Peru].

EPIDOLICHOSTOMA Townsend, 1927a: 238. Type species: *Epidolichostoma andina* Townsend, 1927, by original designation [Peru].

Note: The relative priority of *Dolichostoma* Townsend, 1912 and *Erigonopsis* Townsend, 1912, when the two are treated as synonyms, was established by Aldrich (1934: 37), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999).

References: Aldrich (1934: 3, 37), in key to Patagonian genera, synonymy of *Erigonopsis* with *Dolichostoma*, taxonomic notes; Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Dolichostoma*, *Epidolichostoma* and *Erigonopsis*), Townsend (1939a: 184, 187, 188), redescrptions of *Dolichostoma*, *Epidolichostoma* and *Erigonopsis*; Cortés and Campos (1971: 24, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions; Guimarães (1971: 77), earliest synonymy we have found of *Epidolichostoma* with *Dolichostoma*; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 55, 58), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

arequipae (Townsend, 1912).—Neotropical: South America (Chile, Peru).

Erigonopsis arequipae Townsend, 1912b: 326. Holotype female (USNM). Type locality: Peru, Arequipa, Arequipa.

Reference: Cortés and Campos (1971: 61), first record from Chile.

nigricaudum (Blanchard, 1963).—Neotropical: South America (Argentina, Chile).

Erigonopsis nigricauda Blanchard, 1963: 178. Holotype male (MACN). Type locality: Argentina, Tucumán [province or city].

References: González (1992a: 59), head figure, first record from Chile; Mulieri et al. (2013: 167), notes on type series in MACN.

puntarenensis (Townsend, 1928).—Neotropical: South America (Argentina, Chile).

Erigonopsis puntarenensis Townsend, 1928b: 163. Holotype female (USNM). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Punta Arenas.

References: Aldrich (1934: 38), redescription, head figure, first record from Argentina; Cortés (1986: 149), taxonomic notes.

Genus *ERNESTIOPSIS* Townsend, 1931

ERNESTIOPSIS Townsend, 1931d: 454. Type species: *Ernestiopsis erigonopsidis* Townsend, 1931, by original designation [Chile].

References: Aldrich (1934: 51), in synonymy with *Lypha* Robineau-Desvoidy; Townsend (1936b: 203), diagnosis of adults and immatures of Ernestiini and key to genera (including *Ernestiopsis*); Townsend (1939a: 256), redescription.

erigonopsidis Townsend, 1931.—Neotropical: South America (Argentina, Chile).

Ernestiopsis erigonopsidis Townsend, 1931d: 454. Holotype male (USNM). Type locality: Chile, Valparaíso, Marga Marga, Bosque Los Perales [as “Perales”, ca. 33°9'S, 71°18'W].

erygonopsidis. Incorrect subsequent spelling of *erigonopsidis* Townsend, 1931 (Cortés and Hichins 1969: 41).

Note: *Ernestiopsis erigonopsidis* was treated in *Lypha* Robineau-Desvoidy by Aldrich (1934: 57), Cortés (1946: 176), Cortés and Hichins (1969: 41) and Cortés (1986: 150), and in *Ernestiopsis* by Guimaraes (1971: 83), Gramajo (1998: 95) and O'Hara (2002: 10).

References: Aldrich (1934: 57), redescription; Gramajo (1998: 95), first record from Argentina.

Genus **GANOPROCTUS** Aldrich, 1934

GANOPROCTUS Aldrich, 1934: 36. Type species: *Ganoproctus argentifer* Aldrich, 1934, by original designation [Argentina].

References: Aldrich (1934: 36), key to the two species; Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Ganoproctus*), Townsend (1939a: 193), redescription; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 55, 59), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

argentifer Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Ganoproctus argentifer Aldrich, 1934: 36. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Correntoso.

References: Cortés (1967b: 11), first record from Chile; Cortés (1973a: 98), first description of female.

longicornis Aldrich, 1934.—Neotropical: South America (Chile).

Ganoproctus longicornis Aldrich, 1934: 37. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Lonquimay, Reserva Nacional Alto Biobío [as “Alto Biobio”, ca. 38°36'S, 70°58'W].

Genus **LYGAEOMYIA** Aldrich, 1934

LYGAEOMYIA Aldrich, 1934: 143. Type species: *Lygaeomyia tristis* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936c: 129), diagnosis of adults and immatures of Actiini and key to genera (including *Lygaeomyia*); Townsend (1940a: 233), redescription.

tristis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lygaeomyia tristis Aldrich, 1934: 144. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Correntoso.

Note: *Lygaeomyia tristis* was recorded from both Argentina and Chile in the original description.

Genus *LYPHA* Robineau-Desvoidy, 1830

LYPHA Robineau-Desvoidy, 1830: 141. Type species: *Tachina dubia* Fallén, 1810, by subsequent designation of Robineau-Desvoidy (1863a: 196) [Sweden].

APOROMYA Rondani, 1859: 90. Type species: *Tachina dubia* Fallén, 1810, by original designation [Sweden].

LYPHE. Incorrect subsequent spelling of *Lypha* Robineau-Desvoidy, 1830 (Coquillett 1910: 563).

Note: Preliminary study of some of the species below suggests they do not belong to the same lineage as the *Lypha* of North America and need to be reclassified under one or more other genera.

References: Coquillett (1910: 509, 563), type species of *Aporomya* and *Lypha* (as “*Lyphe*”, with *Aporomya* in synonymy); Townsend (1936b: 216), diagnosis of adults and immatures of Lyphini and key to genera (including *Lypha*); Townsend (1936c: 271), *Aporomya* as synonym of *Lypha*; Townsend (1939a: 306), redescription of *Lypha* (with *Aporomya* in synonymy); Aldrich (1934: 4, 51), in key to Patagonian genera, synonymy, taxonomic notes, key to Patagonian species; Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

angolensis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha angolensis Aldrich, 1934: 58. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Reference: Gramajo (1998: 95), first record from Argentina.

chaetosa Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha chaetosa Aldrich, 1934: 59. Holotype male (USNM). Type locality: Argentina, Río Negro, Lago Nahuel Huapi.

corax Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha corax Aldrich, 1934: 61. Holotype male (NHMUK). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

edwardsi Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha edwardsi Aldrich, 1934: 53. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Correntoso.

Reference: Cortés and Hichins (1969: 41), first record from Chile.

longicornis Aldrich, 1934.—Neotropical: South America (Chile).

Lypha longicornis Aldrich, 1934: 62. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Casa Pangue.

orbitalis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha orbitalis Aldrich, 1934: 60. Holotype male (NHMUK). Type locality: Chile, Los Lagos, Llanquihue, Ensenada.

Reference: Gramajo (1998: 95), first record from Argentina.

ornata Aldrich, 1934.—Neotropical: South America (Argentina, Chile). **New record from Chile.**

Lypha ornata Aldrich, 1934: 56. Holotype female (NHMUK). Type locality: Argentina, Tierra del Fuego, Río Grande, Estancia Viamonte.

Note: *Lypha ornata* is newly recorded from Chile based on four CNC specimens from two localities in the Ultima Esperanza Province of the Magallanes y de la Antártica Chilena Region, with the following data: “Natales East of Mount Payne”, Laguna Amarga, 200 m, 14–20.xii.1960, L. Peña (CNC1546165–CNC1546167); and “110 km north of Pto Natales”, Laguna Amarga, 28.i.1994, M. Wood (CNC1546168).

triangulifera (Jacobs, 1900).—Neotropical: South America (Argentina, Chile).

Hystricia triangulifera Jacobs, 1900: 107. Holotype female (RBINS). Type locality: Argentina, Tierra del Fuego, Canal Beagle, Puerto Harberton.

References: Aldrich (1934: 54), redescription; Cortés (1968c: 142), first record from Chile, southernmost record of a tachinid in the Americas.

truncata Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Lypha truncata Aldrich, 1934: 55. Holotype male (NHMUK). Type locality: Argentina, Tierra del Fuego, Lago Yehuín [as “Lake Yuvin”].

Reference: Cortés (1986: 150), first record from Chile.

Genus *NOTODERUS* Cortés, 1986

NOTODERUS Cortés, 1986: 150. Type species: *Notoderus maculatus* Cortés, 1986, by original designation [Chile].

Reference: Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions.

maculatus Cortés, 1986.—Neotropical: South America (Chile).

Notoderus maculatus Cortés, 1986: 150. Holotype male (MEUC). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, northeast of Punta Arenas, Punta Delgada.

Genus *OLLACHERYPHE* Townsend, 1927

OLLACHERYPHE Townsend, 1927a: 256. Type species: *Ollacheryphe facialis* Townsend, 1927, by original designation [Peru].

AEGLOPS Aldrich, 1934: 47. Type species: *Aeglops aenea* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936b: 218), diagnosis of adults and immatures of Germariini and key to genera (including *Aeglops* and *Ollacheryphe*); Townsend (1939a: 310, 347), redescrptions of *Aeglops* and *Ollacheryphe*; Cortés (1945c: 30), synonymy of *Aeglops* with *Ollacheryphe*; Cortés (1984: 380), in key to tachinid genera of Tarapacá and Antofagasta regions; O'Hara (2002: 105), taxonomic notes.

aenea (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Aeglops aenea Aldrich, 1934: 47. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Correntoso.

Note: *Aeglops aenea* was recorded from both Argentina and Chile in the original description.

facialis Townsend, 1927.—Neotropical: South America (Argentina, Brazil, Chile, Peru).

Ollacheryphe facialis Townsend, 1927a: 339. Holotype male (USNM). Type locality: Peru, Puno, Ollachea.

fascialis. Incorrect subsequent spelling of *facialis* Townsend, 1927 (Cortés and Hichins 1979: 114).

References: Guimarães (1971: 88), first record from Argentina; Cortés and Hichins (1979: 114), first record from Chile; Cortés (1980: 107), first record from Brazil.

Genus *TELODYTES* Aldrich, 1934

TELODYTES Aldrich, 1934: 50. Type species: *Telodytes analis* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936b: 203), diagnosis of adults and immatures of Ernestiini and key to genera (including *Telodytes*); Townsend (1939a: 268), redescription.

analis Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Telodytes analis Aldrich, 1934: 50. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Gutiérrez.

Reference: Henry (1987: 205), first record from Chile.

Genus *XANTHOPELTA* Aldrich, 1934

XANTHOPELTA Aldrich, 1934: 48. Type species: *Xanthopelta scutellaris* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936b: 218), diagnosis of adults and immatures of Germariini and key to genera (including *Xanthopelta*); Townsend (1939a: 362), redescription.

scutellaris Aldrich, 1934.—Neotropical: South America (Argentina, Chile). (Fig. 6b)

Xanthopelta scutellaris Aldrich, 1934: 49. Holotype female (NHMUK). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

Note: *Xanthopelta scutellaris* was recorded from both Argentina and Chile in the original description.

Tribe SIPHONINI

Reference: O'Hara (1989), revision of the genera of the Siphonini.

Genus *CEROMYA* Robineau-Desvoidy, 1830

CEROMYA Robineau-Desvoidy, 1830: 86. Type species: *Ceromya testacea* Robineau-Desvoidy, 1830 (= *Tachina bicolor* Meigen, 1824), by subsequent designation of Coquillett (1910: 520) (see Evenhuis et al. 2010: 54) [France].

CEROMYIA Agassiz, 1846: 7. Unjustified emendation of *Ceromya* Robineau-Desvoidy, 1830 (see Evenhuis et al. 2010: 54).

ACTINACTIA Townsend, 1927a: 248. Type species: *Actinactia lutea* Townsend, 1927, by original designation [Brazil].

Note: O'Hara (1989: 63) in his "List of examined, undescribed, species included in *Ceromya sensu stricto*" listed two undescribed species from Chile as "*Ceromya* Chile sp. 1: One male and one female from Magellanes (CNC)" and "*Ceromya* Chile sp. 2: One male from Isla de Chiloe (CNC)". These species are still undescribed.

References: Coquillett (1910: 520), type species of *Ceromya* (as synonym of *Ceranthia* Robineau-Desvoidy, 1830); Aldrich (1934: 5, 131), in key to Patagonian genera, synonymy, key to four Patagonian species (as *Actia* Robineau-Desvoidy, 1830); Townsend (1936c: 129, 146), diagnosis of adults and immatures of Actiini and key to genera (including *Ceromya*), diagnosis of adults and immatures of Siphonini and key to genera (including *Actinactia*); Townsend (1940a: 200, 274), redescrptions of *Ceromya* and *Actinactia*; O'Hara (1989: 38, 52), in key to genera of the Siphonini, synonymy including *Actinactia* with *Ceromya*, redescription.

amblycera (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Actia amblycera Aldrich, 1934: 132. Holotype male (USNM). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as "Bariloche"].

References: Cortés (1967b: 10), first description of female, first record from Chile; O'Hara (1989: 60), moved to *Ceromya* Robineau-Desvoidy.

cornuta (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Actia cornuta Aldrich, 1934: 131. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Note: *Actia cornuta* was recorded from both Argentina and Chile in the original description. Reference: O'Hara (1989: 61), moved to *Ceromya* Robineau-Desvoidy.

Genus *SIPHONA* Meigen, 1803

References: Coquillett (1910: 528, 606), type species of *Crocuta* Meigen and *Siphona* (with latter in synonymy with former); Townsend (1936c: 146, 273), diagnosis of adults and immatures of Siphonini and key to genera (including *Phantasiosiphona*, *Siphona* and *Siphonopsis*), *Crocuta* as synonym of *Siphona*; Townsend (1940a: 286, 292, 294), redescrptions of *Phantasiosiphona*, *Siphona* (with *Crocuta* in synonymy) and *Siphonopsis*; O'Hara (1989: 39, 84), in key to genera of the Siphonini, recognition, key to subgenera.

Subgenus *JIMIMYIA* Evenhuis, Pont & Whitmore, 2015

SIPHONOPSIS Townsend, 1916a: 622 (junior homonym of *Siphonopsis* Agassiz, 1846). Type species: *Siphona plusiae* Coquillett, 1895, by original designation [United States].

JIMIMYIA Evenhuis, Pont & Whitmore, 2015: 249 (*nomen novum* for *Siphonopsis* Townsend, 1916).

Note: O'Hara (1989: 123) in his "List of examined, undescribed, species included in New World *Siphona* (*Siphonopsis*)" listed one undescribed species from Chile as "*S. (Siphonopsis)*

sp. N: One male from Ecuador, males and two females from Chile (CNC)". This species is still undescribed.

Reference: O'Hara (1989: 86, 120), in key to subgenera of *Siphona*, first treatment as subgenus of *Siphona*, redescription (as *Siphonopsis*).

brasiliensis (Townsend, 1929).—Neotropical: South America (Brazil, Chile).

Siphonopsis brasiliensis Townsend, 1929: 374. Lectotype female (USNM), by fixation of O'Hara (1989: 123) (examination of "Holotype female" from São Paulo in USNM is regarded as a lectotype fixation). Type locality: Brazil, São Paulo, Itaquaquecetuba.

Note: Records of *Siphona brasiliensis* from Chile (e.g., Cortés 1944d: 142; Cortés 1946: 180; Cortés 1948: 123; Cortés and Hichins 1969: 57; Guimarães 1977c: 74) are possibly based on misidentifications (O'Hara 1989: 122).

Reference: Cortés (1944d: 142), first record from Chile.

Subgenus *SIPHONA* Meigen, 1803

CROCUTA Meigen, 1800: 39. Meigen (1800) suppressed by ICZN (1963: 339).

SIPHONA Meigen, 1803: 281. Type species: *Musca geniculata* De Geer, 1776, by designation under the Plenary Powers of ICZN (1974: 157) [Sweden].

CROCUTA Bezzi, 1907: 414. First usage of *Crocuta* (*sensu* Meigen, 1800) as a valid name after Meigen, 1800; no type species designated originally or subsequently (see note).

PHANTASIOSIPHONA Townsend, 1915f: 93. Type species: *Phantasiosiphona tropica* Townsend, 1915, by original designation [Mexico].

Notes: The name *Crocuta* Meigen, 1800 became unavailable when the pamphlet of Meigen (1800) was suppressed by ICZN (1963: 339). *Crocuta* became available later when used by Bezzi (1907: 414), as explained in Evenhuis and Pape (2017: 30). This last work cited the type species of *Crocuta* Bezzi, 1907 as *Musca geniculata* De Geer, 1776 by designation of Coquillett (1910: 528) but this is incorrect; Coquillett (1910) designated a type species for *Crocuta* Meigen, 1800 (at the time an available name) not *Crocuta* Bezzi, 1907.

O'Hara (1989: 119) in his "List of examined, undescribed, species included in *Siphona* (*Siphona*)" listed one undescribed species from Chile as "*S. (Siphona)* nr. *tropica*: One female from Coquimbo, Chile (CNC)". This species is still undescribed. Stireman et al. (2016) recorded specimens of unidentified (and presumably undescribed) *Siphona* (*Siphona*) species from several sites in Chile (Fig. 6c). One specimen of *S. (Pseudosiphona)* Townsend, 1916) was also reported in Stireman et al. (2016: 27) but we have not reexamined it to confirm its subgenus placement; it is in the Stireman collection at Wright State University, Dayton, USA.

References: Aldrich (1934: 5, 108), in key to Patagonian genera, synonymy, taxonomic notes; Cortés and Campos (1971: 22, 1974: 113) and Cortés (1984: 379), in keys to tachinid genera of Tarapacá and Antofagasta regions; O'Hara (1983: 275), synonymy of *Phantasiosiphona* with *Siphona*; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; O'Hara (1989: 85, 108), in key to subgenera of *Siphona*, redescription; Stireman et al. (2016: 27, 28, 29, 33, 35), records of unidentified specimens of *S. (Pseudosiphona)* and *S. (Siphona)* species.

geniculata (De Geer, 1776).—Not Chile [Nearctic (introduced), Palearctic].

Musca geniculata De Geer, 1776: 38.

Note: *Siphona geniculata* is native to the Palearctic Region and was introduced into North America for biological control purposes (Wilkinson 1984). There are many records of *S. geniculata* from South America but we believe they are based on misidentifications, as suggested by O'Hara (1983: 299). Among the records are those of Aldrich (1934: 109), Cortés (1946: 180), Cortés and Hichins (1969: 57), Cortés and Campos (1971: 98), Cortés (1979: 81), Cortés (1986: 158 [as possible misidentification based on pers. comm. with JEOH]), Avalos (1989: 49 [also citing possibility of misidentification]), González (1992b: 179) and Gramajo (1998: 98). Guimarães (1971: 170) did not recognise *S. geniculata* from America south of United States. We have not seen *S. geniculata* among examined specimens of *S. (Siphona)* from the Neotropical Region. Recent DNA barcoding of the COI gene for two *S. (Siphona)* specimens collected by JEOH in Chile in 2015 (CNC497684, CNC497688) confirmed that they are not *S. geniculata*.

kuscheli (Cortés, 1952).—Neotropical: South America (Chile [Juan Fernández Islands]). Not known from mainland Chile.

Phantasiosiphona kuscheli Cortés, 1952: 110. Holotype male (MEUC). Type locality: Chile, Valparaíso, Valparaíso, Juan Fernández Islands, Isla Robinson Crusoe [as “Masatierra”], Cerro Alto, 600 m.

Reference: O'Hara (1983: 279), moved to *Siphona* Meigen.

Tribe TACHININI

Reference: Cortés (1951b: 250), key to Chilean genera of Tachinini with strong setae on the lower parafacial.

Genus *ACROCERONIA* Cortés, 1951

ACROCERONIA Cortés, 1951b: 251. Type species: *Acroceronia elquiensis* Cortés, 1951, by original designation [Chile].

Reference: González (1992a: 56, 57), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

elquiensis Cortés, 1951.—Neotropical: South America (Chile).

Acroceronia elquiensis Cortés, 1951b: 252. Holotype male (MNNC). Type locality: Chile, Coquimbo, Elqui, Gualliguaica, 600 m.

Reference: Cortés (1951b: 251), in key to Chilean genera of Tachinini with strong setae on the lower parafacial.

Genus *AGICUPHOCERA* Townsend, 1915

AGICUPHOCERA Townsend, 1915e: 430. Type species: *Agicuphocera nigra* Townsend, 1915, by original designation [Peru].

References: Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Agicuphocera*), Townsend (1939a: 169), redescription; Cortés

and Campos (1974: 116) and Cortés (1984: 382), in keys to tachinid genera of Tarapacá and Antofagasta regions; González (1992a: 56, 57), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

nigra Townsend, 1915.—Neotropical: South America (Chile, Peru).

Agicuphocera nigra Townsend, 1915e: 430. Holotype female (USNM). Type locality: Peru, Lima, Chosica, ca. 2800 ft.

Reference: Cortés and Campos (1974: 116), first record from Chile.

Genus *ALLELOMYIA* González, 1992

ALLELOMYIA González, 1992a: 56. Type species: *Allelomyia discalis* González, 1992, by original designation [Chile].

discalis González, 1992.—Neotropical: South America (Chile).

Allelomyia discalis González, 1992a: 56. Holotype male (UMCE). Type locality: Chile, Metropolitana de Santiago, Cordillera, Reserva Nacional de Río Clarillo.

Genus *ANDROSOMA* Cortés & Campos, 1971

ANDROSOMA Cortés & Campos, 1971: 50. Type species: *Androsoma perhirsutum* Cortés & Campos, 1971, by original designation [Chile].

References: Cortés and Campos (1971: 24, 1974: 114) and Cortés (1984: 380), in keys to tachinid genera of Tarapacá and Antofagasta regions.

perhirsutum Cortés & Campos, 1971.—Neotropical: South America (Chile).

Androsoma perhirsutum Cortés & Campos, 1971: 52. Holotype male (EEAM). Type locality: Chile, Antofagasta, El Loa, Ojo Hécar, 4500 m (23°11'S, 68°01'W) (coordinates given on p. 12, locality as “Ojo Hécar (Láscar)”).

Genus *ARCHYTAS* Jaennicke, 1867

Note: Subgenera of *Archytas* Jaennicke are not recognised here because the subgeneric placements of the Neotropical species require more study.

ARCHYTAS Jaennicke, 1867: 392 [also 1868: 84]. Type species: *Archytas bicolor* Jaennicke, 1867 (= *Tachina diaphana* Fabricius, 1805), by monotypy [Venezuela].

NEMOCHAETA van der Wulp, 1888: 38. Type species: *Nemochaeta dissimilis* van der Wulp, 1888, by monotypy [Costa Rica].

TACHINODES Brauer & Bergenstamm, 1889: 133 [also 1889: 65]. Type species: hereby fixed under Article 70.3.2 of the *Code* (ICZN 1999) as *Jurinia metallica* Robineau-Desvoidy, 1830, misidentified as *Musca hystrix* Fabricius, 1775 in the fixation by monotypy of Brauer and Bergenstamm (1889) [United States].

- PARAFABRICIA* Brauer & Bergenstamm, 1894: 612 [also 1895: 76] (as subgenus of *Archytas* Jaennicke, 1867). Type species: hereby fixed under Article 70.3.2 of the Code (ICZN 1999) as *Parafabricia perplexa* Townsend, 1931, misidentified as *Tachina bicolor* Wiedemann, 1830 in the subsequent designation of Coquillett (1910: 513) [Brazil].
- EUFABRICIA* Townsend, 1908: 111. Type species: *Eufabricia flavicans* Townsend, 1908 (= *Tachina diaphanus* Fabricius, 1805), by original designation [Brazil].
- PSEUDOARCHYTAS* Townsend, 1915a: 185. Type species: *Pseudoarchytas marmorata* Townsend, 1915, by original designation [Peru].
- NEOARCHYTAS* Townsend, 1915e: 430. Type species: *Neoarchytas inambarica* Townsend, 1915, by original designation [Peru].
- MAKASINOCERA* Townsend, 1915e: 431. Type species: *Makasinocera unguis* Townsend, 1915, by original designation [Peru].
- PSEUDOARCHYTOPSIS* Townsend, 1927a: 252. Type species: *Pseudoarchytopsis brasiliensis* Townsend, 1927 (= *Gonia incerta* Macquart, 1851), by original designation [Brazil].
- PROARCHYTAS* Townsend, 1931c: 351. Type species: *Tachina daemon* Wiedemann, 1830, by original designation [Brazil].
- MAKASINOCEROPS* Townsend, 1935: 219. Type species: *Makasinoceros fulviventrus* Townsend, 1935 (junior secondary homonym of *Jurinia fulviventrus* Robineau-Desvoidy, 1830; = *Archytas shannoni* Guimarães, 1960), by original designation [Brazil].
- ITARCHYTAS* Blanchard, 1940: 225. Type species: *Itarchytas pseudodaemon* Blanchard, 1940, by original designation [Argentina].
- ARCHYNEMOCHAETA* Blanchard, 1941: 345. Type species: *Archynemochaeta frenquellii* Blanchard, 1941, by original designation [Argentina].
- ARCHYTODEJEANIA* Blanchard, 1941: 348. Type species: *Archytodejeania bruchi* Blanchard, 1941, by original designation [Argentina].
- PROARCHYTOIDES* Blanchard, 1941: 365. Type species: *Proarchytoides giacomellii* Blanchard, 1941, by original designation [Argentina].

References: Brauer and Bergenstamm (1893: 58 [also 1893: 146], synonymy of *Nemochaeta* and *Tachinodes* with *Archytas*; Coquillett (1897: 141), synonymy including *Parafabricia* with *Archytas*; Coquillett (1910: 509, 574, 584, 611), type species of *Archytas*, *Nemochaeta*, *Parafabricia* and *Tachinodes* (with last three in synonymy with *Archytas*); Curran (1928b–e), revision of New World species, synonymy including *Makasinocera*, *Neoarchytas*, *Proarchytas* and *Pseudoarchytas* with *Archytas* (*Proarchytas* not mentioned by name but type species included in *Archytas*); Aldrich (1934: 5, 133), in key to Patagonian genera, synonymy including *Eufabricia* and *Pseudoarchytopsis* with *Archytas*, redescription; Townsend (1936b: 167, 174), diagnosis of adults and immatures of Tachinini and key to genera (including *Makasinocera*, *Makasinoceros*, *Nemochaeta*, *Neoarchytas*, *Pseudoarchytas* and *Pseudoarchytopsis*), diagnosis of adults and immatures of Dejeaniini and key to genera (including *Archytas*, *Parafabricia* and *Proarchytas*); Townsend (1936c: 275, 282), *Eufabricia* and *Tachinodes* as synonyms of *Archytas*; Townsend (1939a: 46–58, 70–97), redescrptions of the aforementioned genera of Tachinini and Dejeaniini (with *Eufabricia* and *Tachinodes* in synonymy with *Archytas*); Cortés (1944c), notes on

Chilean species; Sabrosky (1955), notes on *Archytas* species; Guimarães (1960, 1961a, 1961b, 1963b, 1963c), series of papers revising *Archytas* species, synonymy including *Makasinocerops* with *Archytas* (Guimarães 1960: 116, 122), synonymy including *Archynemochaeta*, *Archytodejeania*, *Itarchytas* and *Proarchytoides* with *Archytas* (Guimarães 1961b: 356); Thompson (1963a: 361), revision of Trinidad species; Cortés and Campos (1971: 27, 1974: 116) and Cortés (1984: 382), in keys to tachinid genera of Tarapacá and Antofagasta regions; Ravlin and Stehr (1984), revision of species from America north of Mexico.

incasanus Townsend, 1912.—Neotropical: South America (Bolivia, Chile, Peru).

Archytas incasana Townsend, 1912b: 331. Holotype female (USNM). Type locality: Peru, Piura, Piura.

incasanus. Incorrect subsequent spelling of *incasanus* Townsend, 1912 (Silva et al. 2008: 493, 496).

Note: The identity of *Archytas incasanus* is currently confused in the literature with that of *Archytas divisus* (Walker, 1853), a non-Chilean species. *Archytas divisus* was treated as a tentative synonym of *A. analis* (Fabricius, 1805) by Guimarães (1961b: 374) but was later recognised as a valid name by the same author (Guimarães 1971: 49), with *A. incasanus* in synonymy. As a result of this synonymy, *A. incasanus sensu* Guimarães (1961b: 370) equals *A. divisus sensu* Guimarães (1971). Some authors have continued to use the name *A. incasanus*, see references below and additionally Cortés and Campos (1974: 117) [Chile], Cortés (1984: 385) [Chile], Henry (1987: 200) [Chile], Silva et al. (2008: 496) [Brazil], Nihei (2016: 918) [not Colombia, “but is likely to occur in the country”] and Zetina et al. (2018: 31) [Mexico]. Other authors have used the name *A. divisus*; e.g., Terán (1974: 20) [Venezuela], Avalos (1989: 48) [Argentina] and Vergara de Sánchez and Raven (1990: 95) [Peru, det. Cortés]. There are undoubtedly misidentifications of both *A. incasanus* and *A. divisus* throughout the literature and the only countries that can be conclusively recorded for each is Peru for *A. incasanus* and Brazil for *A. divisus*, based on the type localities. Pending further study of these species we conservatively record the distribution of *A. incasanus* as Peru, Chile and Bolivia and for present purposes record *A. divisus* from the other countries from which *A. incasanus* or *A. divisus* has been reported, see aforementioned references plus Curran (1928e: 275) [Peru, Ecuador, Costa Rica, Mexico] and Thompson (1963a: 383) [Trinidad]. References: Etcheverry (1957: 186), first record from Chile; Cortés and Campos (1971: 55), first record from Bolivia, notes on Chilean specimens (as *A. incasanus*).

incertus (Macquart, 1851).—Not Chile [Argentina, Brazil, Paraguay, Uruguay].

Gonia incerta Macquart, 1851: 152 [also 1851: 179].

Note: *Archytas incertus* was recorded from Chile only once, in a paper on cutworm control in northern Chile by Caltagirone (1953: 88). Although this record was cited much later in the host-parasite catalogue of Guimarães (1977c: 20), *A. incertus* was not mentioned in the Chilean literature after Caltagirone (1953) (e.g., Cortés and Hichins 1969; Cortés and Campos 1971) and is deemed here to have been misidentified from Chile.

marmoratus (Townsend, 1915).—Neotropical: Greater Antilles (Cuba, Haiti, Jamaica, Puerto Rico), eastern Lesser Antilles (Barbados, Grenada, Guadeloupe, Montserrat, Virgin Islands), southern Lesser Antilles (Trinidad & Tobago), Middle

America (Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama), South America (Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Guyana, Peru, Suriname, Venezuela). Nearctic: United States.

Pseudoarchytas marmorata Townsend, 1915a: 186. Holotype female (USNM). Type locality: Peru, Lima, Chosica (3000 ft according to label data).

Echinomyia piliventris of Coquillett (1897: 142, as “*Archytas piliventris*”) and Curran (1928c: 222, as “*Archytas piliventris*”), not van der Wulp, 1883. Misidentification (Sabrosky 1955: 78) (see note).

Note: The distributions of *Archytas marmoratus* and *A. incertus* (Macquart) are confused in the literature, in part because the name *Echinomyia piliventris* van der Wulp, that is currently accepted as a synonym of *A. incertus*, had also been used for misidentified specimens of *A. marmoratus* (e.g., Coquillett 1897: 142; Curran 1928c: 222). Authors who followed the concept of Curran (1928c) for *Archytas piliventris* were using the name in the current sense of *A. marmoratus*. It is likely that the early Argentinian records of “*Archytas piliventris*” by Blanchard (1935: 12, 1937: 47) and of “*Pseudoarchytopsis piliventris*” by Blanchard (1941: 348, 1963: 165) apply to *A. marmoratus* because both “*Pseudoarchytopsis piliventris*” and “*Pseudoarchytas incerta*” were redescribed in the same work by Blanchard (1963). A closer study of Blanchard’s descriptions is needed to determine with certainty the identities of the species he redescribed.

References: Curran (1928c: 202, 222), in key, redescription (as “*Archytas piliventris*” with *Pseudoarchytas marmorata* in synonymy); Sabrosky (1955: 78), modern interpretation of *A. marmoratus*, distribution given as most of the countries listed here with the notable exceptions of Brazil and Argentina; Guimarães (1961a: 168), redescription, distribution including first record from Brazil; Thompson (1963a: 375), redescription; Ravlin and Stehr (1984: 18), redescription; Avalos (1989: 48), first record from Argentina using the name *Archytas marmoratus* (see note above); Maes (1999: 1605), distribution, references; Nihei (2016: 918), in catalogue of Tachinidae of Colombia.

nigriventris (van der Wulp, 1882).—Neotropical: South America (Argentina, Chile).

Jurinia nigriventris van der Wulp, 1882: 81 (junior primary homonym of *Jurinia nigriventris* Robineau-Desvoidy, 1863). Syntypes, 2 females (RMNH). Type localities: Chile and Argentina.

Note: *Jurinia nigriventris* van der Wulp, 1882 is a junior primary homonym of *Jurinia nigriventris* Robineau-Desvoidy, 1863, a valid name for a Mexican species of *Jurinia* Robineau-Desvoidy, 1830. Junior primary homonyms are permanently invalid according to Article 70.3.2 of the *Code* (ICZN 1999), but Article 23.9.5 states: “the author must not automatically replace the junior homonym; the case should be referred to the Commission for a ruling under the plenary power and meanwhile prevailing usage of both names is to be maintained [Art. 82]”. Cortes (1944c: 140) suggested that *J. nigriventris* van der Wulp might be a synonym of *Tachina infirma* Walker, 1849, a name later synonymised by Cortés (1963: 242) with *Jurinia scutellata* Macquart, 1844, treated here as *Archytas scutellatus*. In light of this possible synonymy and the instructions of Article 23.9.5, no action is taken at this time to replace the name of the junior homonym *Jurinia nigriventris* van der Wulp.

peruanus Curran, 1928.—Neotropical: South America (Bolivia, Chile, Peru).

Archytas peruanus Curran, 1928d: 249. Holotype male (USNM). Type locality: Peru, Junín, La Oroya.

References: Guimarães (1961b: 392), redescription, first record from Bolivia; Cortés and Campos (1971: 58), first record from Chile.

pilifrons (Schiner, 1868).—Neotropical: South America (Argentina, Chile).

Echinomyia pilifrons Schiner, 1868: 331. Holotype male (NHMW). Type locality: Chile.

Jurinia nudigaena Brauer, 1898: 500. Lectotype female (NHMUK), by designation of Sabrosky (1955: 83). Type locality: Chile (see note).

Archytas pollinosus Curran, 1928d: 251. Holotype male (SDEI, Rohlfien and Ewald 1974: 142). Type locality: Chile.

Jurinia scutellata of Aldrich (1934: 135), Cortés (1944c: 139), Cortés (1946: 182) and Guimarães (1961a: 170) (all as “*Archytas scutellatus*”), not Macquart, 1844. Misidentification (e.g., Cortés 1963: 247; Cortés and Hichins 1969: 17; Guimarães 1971: 51).

Note: Sabrosky (1955: 83) reported that Aldrich saw three syntypes of *Jurinia nudigaena* in the Bigot collection and had written in his notes: “2 are *Archytas piliventris* V.d.W., the other is ♀ of *Archytas pilifrons* Sch.”. Sabrosky (1955) designated the last as lectotype of *J. nudigaena* and synonymised *J. nudigaena* with *Echinomyia pilifrons*. The lectotype of *J. nudigaena* is from Chile and the two paralectotypes, both males of a second species [*Archytas incertus* (Macquart, 1851), as *Archytas piliventris* (van der Wulp, 1883) in notes of Aldrich] are from Montevideo in Uruguay (all three examined by DMW). Brauer (1898: 500) cited the type locality of *Jurinia nudigaena* as “Chili, Montevideo” but the latter locality applies to the two males of the second species, *Archytas incertus*. The true *Archytas pilifrons* has not been recorded from Uruguay.

References: Aldrich (1929a: 27), taxonomic notes on holotype of *Echinomyia pilifrons*; Aldrich (1934: 135), synonymy of *Archytas pollinosus* with “*Archytas scutellatus*” (misidentification), records from Argentina and Chile (as *A. scutellatus*); Cortés (1963: 247), notes on synonymy and misidentifications.

platonicus Cortés & Campos, 1971.—Neotropical: South America (Chile, Peru).

Archytas platonicus Cortés & Campos, 1971: 58. Holotype male (EEAM). Type locality: Chile, Arica y Parinacota, Arica, Valle de Lluta, km 57.

Reference: Vergara de Sánchez and Raven (1990: 95), first record from Peru.

scutellatus (Macquart, 1844).—Neotropical: South America (Chile).

Jurinia scutellata Macquart, 1844: 41 [also 1844: 198]. Lectotype female (MNHN, see note), by fixation of Aldrich (1934: 135) (examination of “type” in MNHN is regarded as a lectotype fixation). Type locality: Chile.

Tachina infirma Walker, 1849: 719. Lectotype male (NHMUK), by fixation of Cortés (1963: 242) (examination of “type” from Chile in NHMUK is regarded as a lectotype fixation). Type locality: Chile.

Jurinia andana Robineau-Desvoidy, 1863a: 657. Type(s), female (lost, Cortés 1963: 247). Type locality: Chile.

Echinomyia ignobilis Rondani, 1863: 15 [also 1864: 15]. Type(s), unspecified sex (lost, Cortés 1963: 247). Type locality: Chile.

Archytas chilensis Curran, 1928c: 222. Holotype male (USNM). Type locality: Chile, Valparaíso, Valparaíso, Valparaíso.

Note: Macquart (1844: 41) described *Jurinia scutellata* from an unspecified number of specimens from “Chili” and “Santa-Fe de Bogota, en Colombie”. The online MNHN database records a female lectotype from Chile in the Macquart collection for *Jurinia scutellata* (number MNHN-ED-ED8299) based on a lectotype determination label that DMW attached to the specimen in 1982. This specimen is presumed to be the same one examined earlier by Aldrich (1934: 135, as “type”) and Cortés (1963: 246, as “tipo”). The paralectotypes from Colombia are not listed in the MNHN database and have not been discussed by subsequent authors. They are here presumed to have been misidentified because there is no corroborating evidence that *Archytas scutellatus* occurs in Colombia and it was not listed from Colombia by Nihei (2016). References: Aldrich (1934: 135), redescription (as *Archytas chilensis*); Cortés (1963: 242, 246), notes on name-bearing types of *Tachina infirma* (in NHMUK), *Jurinia scutellata* (in MNHN) and *Archytas chilensis* (in USNM), synonymy of *Tachina infirma*, *Jurinia andana*, *Echinomyia ignobilis* and *Archytas chilensis* with *Jurinia scutellata*.

seminiger (Wiedemann, 1830).—Not Chile [Brazil, Colombia].

Tachina seminigra Wiedemann, 1830: 296.

Note: *Archytas seminiger* was described from Brazil and was later recorded from Chile and Colombia by Schiner (1868: 331). Reed (1888: 305) listed *A. seminiger* from Chile but this was likely based on Schiner's earlier record. *Archytas seminiger* was considered a doubtful species in Chile by Cortés (1944c: 140, 1946: 172) and it has not been reported from Chile since. Reports of *A. seminiger* from Mexico and/or Puerto Rico by such authors as Giglio-Tos (1894: 484), Aldrich (1905: 487), Curran (1928a: 117), Wolcott (1948: 483), Nihei (2016: 918) and Zetina et al. (2018: 32) are presumed to have been based on misidentifications. *Archytas seminiger* was redescribed by Guimarães (1963b: 155).

Genus *CHAETOEPALPUS* Vimmer & Soukup, 1940

CHAETOEPALPUS Vimmer, 1940: 101. *Nomen nudum* (proposed after 1930 without designation of type species; no included species).

CHAETOEPALPUS Vimmer & Soukup, 1940a: 218. Type species: *Chaetoepalpus coquilleti* Vimmer & Soukup, 1940, by monotypy [Peru]. **New record from Chile.**

RUIZIELLA Cortés, 1951b: 254. Type species: *Ruiziella frontosa* Cortés, 1951, by original designation [Chile]. **Syn. nov.**

CHAETOPALPUS. Incorrect subsequent spelling of *Chaetoepalpus* Vimmer & Soukup, 1940 (Vimmer and Soukup 1940b: 371; Guimarães 1971: 264).

Note: The new synonymy of *Ruiziella* with *Chaetoepalpus* is explained under *C. coquilleti*.

References: Cortés (1951b: 251), *Ruiziella* in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Cortés and Campos (1971: 26, 1974: 116) and Cortés (1984: 381), *Ruiziella* in keys to tachinid genera of Tarapacá and Antofagasta regions; Stireman et al. (2016: 38), habitus images of *Ruiziella* sp.

coquilleti Vimmer & Soukup, 1940.— Neotropical: South America (Argentina, Chile, Peru). **New records from Argentina and Chile.** (Fig. 6d)

Chaetoepalpus coquilleti Vimmer & Soukup, 1940a: 218. Type(s), unspecified sex (1 female in NMPC, examined by DMW). Type locality: Peru, Puno [region or city].

Ruiziella frontosa Cortés, 1951b: 255. Holotype male (MNNC). Type locality: Chile, Metropolitana de Santiago, Cordillera, Cerro Punta de Damas, 3200 m [ca. 33°31'S, 70°26'W]. **Syn. nov.**

Note: *Chaetoepalpus coquilleti* was described from an unspecified number of specimens of unspecified sex. One type specimen, a female, was examined in NMPC by DMW in 2005 and matched a conspecific CNC female bearing the following data: Argentina, Jujuy, Río Seco, 5 km south of Station Catalina, 3500 m, 25.x.1968, L. Peña. There are additional specimens in CNC collected by Peña from various localities at high elevations in Jujuy Province of Argentina. There is a series of specimens from Chile in CNC with the following data: Coquimbo, La Laguna, 130 km east of Vicuña, 3200 m, 20.i.1994, G. & M. Wood (including CNC_Diptera162434–CNC_Diptera162436). Images of the specimen from Argentina that was compared to the type of *C. coquilleti* in NMPC and a specimen from Vicuña, Chile, were compared with the types of *R. frontosa* and *R. luctuosa* in MNNC by CRG. The type of *R. frontosa* is a match for *C. coquilleti* (i.e., palpus shorter and postpedicel less broad at apex than in type of *R. luctuosa*), and on this basis *Ruiziella* is synonymised with *Chaetoepalpus* and *R. frontosa* is synonymised with *C. coquilleti*.

The specific epithet “*coquilleti*” is not a misspelling or printer’s error that could be “corrected” to “*coquillett*” to correspond with the proper spelling of the surname of dipterist Daniel W. Coquillett. Vimmer and Soukup (1940a) mentioned *coquilleti* only once in their paper (p. 218) but mentioned the dipterist’s name as well in the same paper, as “Coquillett” (p. 221), and thus the specific epithet was spelled as intended.

References: Guimarães (1971: 217), *C. coquilleti* listed as an unrecognised species of Tachinidae; Cortés (1979: 81), first record of *R. frontosa* from Argentina.

luctuosus (Cortés, 1951).—Neotropical: South America (Argentina, Chile). **Comb. nov.**

Ruiziella luctuosa Cortés, 1951b: 257. Holotype male (MNNC). Type locality: Chile, Ñuble, Diguillín, Termas de Chillán.

Reference: Cortés (1980: 106), first record from Argentina.

Genus **CHILOEPALPUS** Townsend, 1927

CHILOEPALPUS Townsend, 1927c: 281. Type species: *Chiloepalpus aurifacies* Townsend, 1927 (= *Jurinia callipyga* Bigot, 1857), by original designation [Chile].

EUHELIOPROSOPA Reinhard, 1964: 123. Type species: *Euhelioprosopa pactilis* Reinhard, 1964 (= *Cuphocera aurea* Aldrich, 1926), by original designation [Chile].

References: Aldrich (1934: 5, 122), in key to Patagonian genera, redescription; Townsend (1936b: 182), diagnosis of Juriniini and key to genera (including *Chiloepalpus*); Townsend (1939a: 108), redescription of *Chiloepalpus*; Cortés (1951b: 250), in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; Cortés (1992: 236), synonymy of *Euhelioprosopa* with *Chiloepalpus*.

aureus (Aldrich, 1926).—Neotropical: South America (Chile).

Cuphocera aurea Aldrich, 1926b: 25. Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Euhelioprosopa pactilis Reinhard, 1964: 124. Holotype male (CAS). Type locality: Chile, Coquimbo, Elqui, 50 km south of La Serena (misspelled as “LaSorena” in original description, see citation of label data in Arnaud 1979: 400).

References: Aldrich (1934: 124), redescription in *Chiloepalpus*, taxonomic notes; Cortés (1992: 236), synonymy of *Euhelioprosopa pactilis* with *Cuphocera aurea*.

callipygus (Bigot, 1857).—Neotropical: South America (Argentina, Chile).

Jurinia callipyga Bigot, 1857: 299. Lectotype female (NHMUK), by fixation of Townsend (1939a: 108) (mention of “Ht female” from Chile in NHMUK [as “Newmarket”] is regarded as a lectotype fixation). Type locality: Chile.

Chiloepalpus aurifacies Townsend, 1927c: 281. Holotype female (SDEI, Rohlfien & Ewald 1974: 133). Type locality: Chile, Biobío, Concepción, Concepción.

Note: The type locality of *Chiloepalpus aurifacies* was given as “Concepcion” in Chile, which could be interpreted as either the city or province of that name. Cortés & Hichins (1969: 26) cited the former as the type locality (as “Concepción (Concepción)”) and we follow this interpretation.

Reference: Aldrich (1934: 123), synonymy, redescription, first record from Argentina.

Genus *COMOPSIS* Cortés, 1986

COMOPSIS Cortés, 1986: 148. Type species: *Comopsis regale* Cortés, 1986, by original designation [Chile].

References: Cortés (1986: 143), in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 55, 58), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

regale Cortés, 1986.—Neotropical: South America (Chile).

Comopsis regale Cortés, 1986: 148. Holotype male (MEUC). Type locality: Chile, Aysén, General Carrera, 5.8 km west of Chile Chico.

Genus *DEOPALPUS* Townsend, 1908

DEOPALPUS Townsend, 1908: 110. Type species: *Deopalpus hirsutus* Townsend, 1908, by original designation [Mexico].

SPANIPALPUS Townsend, 1908: 110. Type species: *Trichophora miscelli* Coquillett, 1897, by monotypy [United States].

PROCYANOPSIS Townsend, 1934a: 209. Type species: *Procyanopsis pictipennis* Townsend, 1934, by original designation [Brazil].

SPANIPALPIS. Incorrect subsequent spelling of *Spanipalpus* Townsend, 1908 (Coquillett 1910: 606).

Notes: The relative priority of *Deopalpus* Townsend, 1908 and *Spanipalpus* Townsend, 1908, when the two are treated as synonyms, was established by Sabrosky and Arnaud (1965: 1003), as the First Reviser (Article 24.2.2 of the *Code*, ICZN 1999).

References: Coquillett (1910: 531, 606) type species of *Deopalpus* (as synonym of *Cuphocera* Macquart) and *Spanipalpus* (as “*Spanipalpis*”); Aldrich (1934: 126), as *Cuphocera* Macquart, in part; Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Deopalpus*, *Procyanopsis* and *Spanipalpus*), Townsend (1939a: 183, 207, 212), redescrptions of *Deopalpus*, *Procyanopsis* and *Spanipalpus*; Cortés (1951b: 251), *Spanipalpus* in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Guimarães (1963a: 76), synonymy of *Procyanopsis* with *Deopalpus*; Sabrosky and Arnaud (1965: 1003), synonymy of *Spanipalpus* with *Deopalpus*; Cortés (1967b: 16), key to separate *Spanipalpus*, *Vibrissomyia* Townsend and *Epalpodes* Townsend; Cortés (1984: 382), *Spanipalpus* in key to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 144), *Spanipalpus* in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 56, 59), *Spanipalpus* in key to Chilean genera of “Cuphocerini”, key to species, diagnosis, notes, two new species.

australis (Townsend, 1928).—Neotropical: South America (Argentina, Chile).

Spanipalpus australis Townsend, 1928b: 164. Holotype female (USNM). Type locality: Chile, Magallanes y de la Antártica Chilena, Magallanes, Punta Arenas.

Helioprosopa finita Reinhard, 1964: 121. Holotype male (CAS). Type locality: Chile, Coquimbo, Elqui, 5 miles north of Laguna Dam, 8000 ft.

Note: The records of *Helioprosopa finita* from Mexico and Colombia given in Guimarães (1971: 78) were likely based on misidentifications; see also Nihei (2016: 922).

References: Aldrich (1934: 127), redescription; Reinhard (1934: 57), redescription; Cortés (1992: 236), synonymy of *Helioprosopa finita* with *Spanipalpus australis*; Gramajo (1998: 95), first record from Argentina.

conspiciendum (Cortés, 1976).—Neotropical: South America (Chile).

Spanipalpus conspicendum Cortés, 1976: 6. Holotype male (CNC). Type locality: Chile, Biobío, Arauco, Cordillera de Nahuelbuta, Cerro Pichinahuel [ca. 37°48'S, 73°2'W].

hiemalis (Cortés, 1983).—Neotropical: South America (Chile).

Spanipalpus hiemalis Cortés, 1984: 384. Holotype female (MEUC). Type locality: Chile, Tarapacá, Tamarugal, Zapahuira, ca. 2500 m.

ochricornis (Bigot, 1888).—Neotropical: South America (Chile).

Epalpus ochricornis Bigot, 1888b: 95. Holotype female (NHMUK). Type locality: Chile.

References: Brauer (1898: 503), taxonomic notes on *Epalpus ochricornis*; Aldrich (1930: 26), notes on type of *Epalpus ochricornis*; Guimarães (1971: 76), moved to *Deopalpus* based on “R. Cortés, *in litt.*”; Cortés (1976: 6), partial redescription (as *Spanipalpus ochricornis*).

picturatus (González, 1992).—Neotropical: South America (Chile).

Spanipalpus picturatus González, 1992a: 61. Holotype male (UMCE). Type locality: Chile, Tarapacá, Tamarugal, 1250 m (located at ca. 20°24'S, 69°56'W, as determined by CRG).

pulchriceps (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Cuphocera pulchriceps Aldrich, 1934: 128. Holotype male (NHMUK). Type locality: Argentina, Río Negro, San Carlos de Bariloche [as “Bariloche”].

Reference: Cortés and Hichins (1979: 27), first record from Chile.

rubidus (González, 1992).—Neotropical: South America (Chile).

Spanipalpus rubidus González, 1992a: 62. Holotype female (UMCE). Type locality: Chile, Magallanes y de la Antártica Chilena, Última Esperanza, Sierra de Los Baguales, 600 m [ca. 50°47'S, 72°24'W].

Nomina dubia of *DEOPALPUS* Townsend, 1908

pruinus (Rondani, 1863).—Neotropical: South America (Chile).

Ciphocera pruinosa Rondani, 1863: 16 [also 1864: 16]. Type(s), female (probably MZUF or lost). Type locality: Chile.

References: Guimarães (1971: 77), unrecognised species of *Deopalpus*; González (1992a: 61), not included in key to Chilean species of *Spanipalpus*.

ratzeburgii (Jaennicke, 1867).—Neotropical: South America (Chile).

Demoticus ratzeburgii Jaennicke, 1867: 386 [also 1868: 78]. Type(s), female (SMF). Type locality: Chile.

ratzeburgi. Incorrect subsequent spelling of *ratzeburgii* Jaennicke, 1867 (e.g., Cortés and Hichins 1969: 90; Guimarães 1971: 77, 315).

References: Guimarães (1971: 77), unrecognised species of *Deopalpus*; González (1992a: 61), not included in key to Chilean species of *Spanipalpus*.

Genus *EDWYNIA* Aldrich, 1930

REEDIA Aldrich, 1928b: 17 (junior homonym of *Reedia* Ashmead, 1904). Type species: *Reedia robusta* Aldrich, 1928, by original designation [Chile].

EDWYNIA Aldrich, 1930: 26 (*nomen novum* for *Reedia* Aldrich, 1928).

References: Aldrich (1934: 5, 125), in key to Patagonian genera, synonymy, taxonomic notes; Townsend (1936b: 182), diagnosis of adults and immatures of Juriniini and key to genera (including *Edwynia*); Townsend (1936c: 281), *Edwynia* as valid name for *Reedia*; Townsend (1939a: 116), redescription of *Edwynia* (with *Reedia* in synonymy); Cortés (1951b: 250), in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions.

robusta (Aldrich, 1928).—Neotropical: South America (Argentina, Chile).

Reedia robusta Aldrich, 1928b: 18. Holotype female (USNM). Type locality: Chile, Biobío, Concepción, Concepción.

Note: The type locality of *Reedia robusta* was given as “Concepcion” in Chile, which could be interpreted as either the city or province of that name. Cortés and Hichins (1969: 32) cited the former as the type locality (as “Concepción (Concepción)”) and we follow this interpretation. Reference: Aldrich (1934: 126), redescription, first record from Argentina.

Genus *EPALPODES* Townsend, 1912

EPALPODES Townsend, 1912b: 330. Type species: *Epalpodes equatorialis* Townsend, 1912, by original designation [Ecuador].

References: Townsend (1936b: 182), diagnosis of adults and immatures of Juriniini and key to genera (including *Epalpodes*); Townsend (1939a: 119), redescription; Cortés (1951b: 251), in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Cortés (1967b: 16), key to separate *Epalpodes*, *Vibrissomyia* Townsend and *Deopalpus* Townsend (as *Spanipalpus* Townsend); Cortés and Campos (1974: 116) and Cortés (1984: 382), in keys to tachinid genera of Tarapacá and Antofagasta regions; González (1992a: 56, 59), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

chillanensis Cortés, 1951.—Neotropical: South America (Argentina, Chile).

Epalpodes chillanensis Cortés, 1951b: 258. Holotype male (MNNC). Type locality: Chile, Ñuble, Diguillín, Termas de Chillán.

Reference: Liljesthrom (1980: 135), first record from Argentina.

malloi Cortés & Campos, 1971.—Neotropical: South America (Chile).

Epalpodes malloi Cortés & Campos, 1971: 62. Holotype male (EEAM). Type locality: Chile, Tarapacá, Tamarugal, Mamiña, 2700 m (20°06'S, 69°16'W) (coordinates given on p. 11).

Genus *EPALPUS* Rondani, 1850

EPALPUS Rondani, 1850: 168, 169. Type species: *Micropalpus rufipennis* Macquart, 1846, by subsequent designation of Coquillett (1910: 538) (see O'Hara et al. 2011: 81) [Colombia].

EUSIGNOSOMA Townsend, 1914b: 44. *Nomen nudum* (see Evenhuis et al. 2015: 122).

EUSIGNOSOMA Townsend, 1914c: 123. Type species: *Eusignosoma aureum* Townsend, 1914, by original designation [Peru].

ARGENTOEPALPUS Townsend, 1919a: 178. Type species: *Epalpus niveus* Townsend, 1914, by original designation [Peru].

Note: The name *Eusignosoma aureum* Townsend, 1914, type species of *Eusignosoma* Townsend, is a junior secondary homonym of *Saundersia aurea* Giglio-Tos, 1893 when the two names are placed together in the genus *Epalpus* Rondani, as in Guimarães (1971: 64). We have assessed the placement of *Saundersia aurea* and move it to “Unplaced species of Tachinini” herein. See under that heading for further details.

References: Coquillett (1910: 538), type species of *Epalpus*; Townsend (1936b: 182), diagnosis of adults and immatures of Juriniini and key to genera (including *Argentoepalpus*, *Epalpus* and *Eusignosoma*); Townsend (1939a: 105, 120, 128), redescrptions of *Argentoepalpus*, *Epalpus* and *Eusignosoma*; Sabrosky and Arnaud (1965: 1002), synonymy of *Argentoepalpus* with *Epalpus*; Guimarães (1971: 63), synonymy including *Eusignosoma* with *Epalpus*.

porteri Brèthes, 1918.—Neotropical: South America (Chile).

Epalpus porteri Brèthes, 1918: 50. Type(s), unspecified sex (1 female in MACN, Mulieri et al. 2013: 169). Type locality: Chile, Valparaíso, Petorca, La Ligua.

References: Cortés (1963: 250), notes on a specimen with data of name-bearing type in MACN (with no mention of “type” and hence not a lectotype fixation); Mulieri et al. (2013: 169), notes on syntype in MACN.

Genus *PELETERIA* Robineau-Desvoidy, 1830

Note: Subgenera of *Peleteria* Robineau-Desvoidy are not recognised here because the subgeneric placements of the Neotropical species require more study.

PELETERIA Robineau-Desvoidy, 1830: 39. Type species: *Peleteria abdominalis* Robineau-Desvoidy, 1830, by subsequent designation of Coquillett (1910: 586) (see Evenhuis et al. 2010: 129) [Italy].

CUPHOCERA Macquart, 1845: 267. Type species: *Micropalpus ruficornis* Macquart, 1835, by original designation [France].

PELETERIOPSIS Townsend, 1916a: 630. Type species: *Echinomyia flaviventris* van der Wulp, 1888, by original designation [Mexico].

APHRIOSPHYRIA Townsend, 1927a: 238. Type species: *Aphriosphyria communis* Townsend, 1927 (= *Tachina robusta* Wiedemann, 1830), by original designation [Brazil].

CUPHOCEROPSIS Townsend, 1935: 220. Type species: *Cuphoceropsis facialis* Townsend, 1935 (= *Echinomyia pygmaea* Macquart, 1851), by original designation [Brazil].

APHRIOSPHYRIOPSIS Blanchard, 1943c: 134. Type species: *Aphriosphyriopsis nemochaetoides* Blanchard, 1943, by original designation [Argentina].

CUPHOCEROMYIA Blanchard, 1943c: 136. Type species: *Cuphoceromyia aldrichi* Blanchard, 1943 (junior secondary homonym of *Peleteria aldrichi* Curran, 1925; = *Peleteria blanchardi* Guimarães, 1971), by original designation [Argentina].

PROSTEATOSOMA Blanchard, 1943c: 150. Type species: *Prosteatosoma lineata* Blanchard, 1943, by original designation [Argentina].

APHRYOSPHYRIA. Incorrect original spelling of *Aphriosphyria* Townsend, 1927 (Townsend 1927a: 287).

PELETIERIA. Incorrect subsequent spelling of *Peleteria* Robineau-Desvoidy, 1830 (Thompson 1963a: 341, 412).

Note: There are two original spellings of *Aphriosphyria* in Townsend (1927a): *Aphriosphyria* (p. 238) and *Aphryosphyria* (p. 287). The correct original spelling was selected as *Aphriosphyria* by Townsend (1927b, see entry for “page 287, line 7 [from] top” in the unpaginated errata of Townsend 1927a), as the First Reviser (Article 24.2.3 of the *Code*, ICZN 1999).

References: Coquillett (1910: 529, 586), type species of *Cuphocera* and *Peleteria*; Curran (1925), revision of New World species; Aldrich (1934: 5, 119, 126), in key to Patagonian genera, synonymy including *Aphriosphyria* and *Peleteriospis* with *Peleteria*, taxonomic notes (as both *Peleteria* and *Cuphocera*); Townsend (1936b: 167, 190), diagnosis of adults and

immatures of Tachinini and key to genera (including *Peleteria* and *Peleteriopsis*), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Aphriosphyria*, *Cuphocera* and *Cuphoceropsis*; Townsend (1939a: 54, 55, 172, 179, 180), redescription of *Peleteria*, *Peleteriopsis*, *Aphriosphyria*, *Cuphocera* and *Cuphoceropsis*; Cortés (1951b: 250), in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Guimarães (1962), revision of Brazilian species; Cortés and Campos (1971: 27, 1974: 116) and Cortés (1984: 382), in keys to tachinid genera of Tarapacá and Antofagasta regions; Guimarães (1971: 43), synonymy of *Aphriosphyriopsis*, *Cuphoceromyia*, *Cuphoceropsis* and *Prosteatosoma* with *Peleteria*.

filipalpis (Rondani, 1863).—Neotropical: South America (Argentina, Chile).

Echinomyia filipalpis Rondani, 1863: 15 [also 1864: 15]. Type(s), female (probably MZUF or lost). Type locality: Chile (likely the commune of Valdivia in southern Chile [in Los Ríos Region, Valdivia Province] according to Cortés and Campos 1971: 65).

Note: *Peleteria filipalpis* is restricted here to southern South America following Cortés and Campos (1971: 65).

References: Curran (1925: 257), redescription [but misidentified, see *Peleteria sordida* Aldrich under *P. pygmaea* (Macquart)]; Aldrich (1934: 120), synonymy, redescription, taxonomic notes, first records from Argentina; Cortés and Campos (1971: 65), distribution in Chile and Argentina, comments on type locality of *E. filipalpis*.

pygmaea (Macquart, 1851).—Neotropical: South America (Argentina, Brazil, Chile, Paraguay).

Echinomyia pygmaea Macquart, 1851: 143 [also 1851: 170]. Lectotype female (MNHN), by designation herein (see Lectotype Designations section). Type locality: Chile.

Peleteria sordida Aldrich, 1934: 122 (named for *Echinomyia filipalpis* of Curran, 1925, not Rondani, 1863). Holotype male (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Cuphoceropsis facialis Townsend, 1935: 220. Holotype female (USNM). Type locality: Brazil, Pernambuco, Tapéra.

Echinomyia filipalpis of Curran (1925: 257, as “*Peleteria filipalpis*”), not Rondani, 1863. Misidentification (Aldrich 1934: 122).

pygmea. Incorrect subsequent spelling of *pygmaea* Macquart, 1851 (Vergara de Sánchez and Raven 1990: 95).

References: Aldrich (1934: 122), first record from Argentina (as *Peleteria sordida*); Guimarães (1962: 488), redescription (as *Peleteria sordida*), distribution as Argentina, Brazil, Chile and Paraguay; Cortés (1963: 248), *Echinomyia pygmaea* moved to *Peleteria*, synonymy of *P. sordida* with *P. pygmaea*.

robusta (Wiedemann, 1830).—Neotropical: South America (Argentina, Brazil, Chile, Peru, Uruguay).

Tachina robusta Wiedemann, 1830: 290. Lectotype female (NHMW), by fixation of Aldrich (1929a: 28) (examination of female “type” from Montevideo in

NHMW is regarded as a lectotype fixation). Type locality: Uruguay, Montevideo, Montevideo.

Fabricia andicola Bigot, 1888b: 86. Holotype female (NHMUK). Type locality: Chile. **Syn. revived.**

Peleteria robusta marmorata Townsend, 1915a: 185. Syntypes, 8 males and 8 females (USNM). Type locality: Peru, Lima, Chosica, ca. 2700–3000 ft.

Peleteria inca Curran, 1925: 247. Holotype male (CUIC). Type locality: Peru, Lima, Matucana. **Syn. revived.**

Aphriosphyria communis Townsend, 1927a: 287 (genus as “*Aphryosphyria*”, see note above under generic synonyms). Syntypes, many males and females (USNM). Type locality: Brazil, São Paulo, Itaquaquecetuba.

Notes: The mention of a “female Ht” of *Aphriosphyria communis* by Townsend (1931b: 158) or “Ht female” by Townsend (1939a: 172) is not accepted as a lectotype fixation because the specimen in question is not distinguishable from the other females in the type series.

The distribution of *Peleteria robusta* is confused in the literature due to numerous misidentifications and is recognised here only from South America. Aldrich (1929a: 28) and Guimarães (1962: 484) treated *P. robusta* as widespread in the Americas with the northern portion of the distribution attributable to their treatment of *Peleteria texensis* Curran, 1925 as a junior synonym of *P. robusta*. Guimarães (1971: 45) later treated this synonymy as questionable and Richards (1973: 80) confirmed *P. texensis* as a separate species. *Peleteria texensis* was recorded from United States to Costa Rica by O'Hara and Wood (2004: 322). The South American distribution of *P. robusta* was given as Argentina, Brazil, Chile, Peru and Uruguay by Guimarães (1962: 488) and this distribution is followed here.

Fabricia andicola and *Peleteria inca* were treated as junior synonyms of *Peleteria robusta* by Guimarães (1962: 484) but were moved into synonymy with *Peleteria filipalpis* by Guimarães (1971: 44) based on their earlier placement there by Aldrich (1934: 120). However, Aldrich (1934) misinterpreted the type locality of *P. filipalpis* and misidentified the species from the Santiago area according to Cortés and Campos (1971: 65), thereby making the synonymy of Guimarães (1962: 484) more probable than that of Guimarães (1971: 44).

References: Brauer (1898: 495), taxonomic notes on *Fabricia andicola*; Aldrich (1929a: 28), synonymy (not entirely followed here, see note above), taxonomic notes; Parker (1953: 62), figures of egg, first instar larva and puparium; Guimarães (1962: 484), redescription (see note above); Blanchard (1963: 155), redescription (as *Aphriosphyria robusta*); Cortés and Campos (1971: 64), distribution of *P. robusta* vs. *P. filipalpis*.

Genus *PYRRHOTACHINA* Townsend, 1931

PYRRHOTACHINA Townsend, 1931d: 447. Type species: *Pyrrhotachina proboscidea* Townsend, 1931, by original designation [Argentina].

References: Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Pyrrhotachina*); Townsend (1939a: 209), redescription; Cortés (1984: 382), in key to tachinid genera of Tarapacá and Antofagasta regions; González (1992a: 55, 59), in key to Chilean genera of “Cuphocerini”, diagnosis.

proboscidea Townsend, 1931.—Neotropical: South America (Argentina, Chile).

Pyrrhotachina proboscidea Townsend, 1931d: 448. Holotype female (SDEI, Rohlfi and Ewald 1974: 142). Type locality: Argentina, Mendoza [province or city].

Reference: Cortés (1984: 387), taxonomic notes, first record from Chile.

Genus **SAUNDERSIOPS** Townsend, 1914

SAUNDERSIOPS Townsend, 1914d: 138. Type species: *Saundersiops confluens* Townsend, 1914, by original designation [Peru].

SIGNOEPALPUS Townsend, 1931d: 446. Type species: *Signoepalpus spinosus* Townsend, 1931, by original designation [Peru].

References: Townsend (1936b: 182), diagnosis of Juriniini and key to genera (including *Saundersiops* and *Signoepalpus*); Townsend (1939a: 152, 153), redescrptions of *Saundersiops* and *Signoepalpus*; Curran (1947: 94), revision, key to species, synonymy of *Signoepalpus* with *Saundersiops*; Cortés (1984: 382), in key to tachinid genera of Tarapacá and Antofagasta regions.

cruciatus Townsend, 1914.—Neotropical: South America (Chile, Peru).

Saundersiops cruciata Townsend, 1914d: 140. Holotype female (USNM). Type locality: Peru, Lima, Matucana, ca. 8000 ft.

Reference: Cortés and Hichins (1979: 114), first record from Chile.

Genus **STEATOSOMA** Aldrich, 1934

STEATOSOMA Aldrich, 1934: 112. Type species: *Steatosoma rufiventris* Aldrich, 1934, by original designation [Argentina].

References: Townsend (1936b: 167), diagnosis of adults and immatures of Tachinini and key to genera (including *Steatosoma*; Townsend (1939a: 63), redescription; Cortés (1951b: 251), in key to Chilean genera of Tachinini with strong setae on the lower parafacial. Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions. González (1992a: 56, 63), in key to Chilean genera of “Cuphocerini”, diagnosis, notes.

nigriventris Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Steatosoma nigriventris Aldrich, 1934: 115. Holotype male (NHMUK). Type locality: Argentina, Tierra del Fuego, Lago Yehuin [as “Lake Yuvín”].

nigripentris. Incorrect subsequent spelling of *nigriventris* Aldrich, 1934 (Cortés 1963: 246).

References: Cortés (1963: 246), notes on type series in NHMUK; Cortés (1973a: 98), taxonomic notes, first record from Chile.

rufiventris Aldrich, 1934.—Neotropical: South America (Argentina, Chile).

Steatosoma rufiventris Aldrich, 1934: 112. Holotype male (NHMUK). Type locality: Argentina, Tierra del Fuego, Río Grande, Estancia Viamonte.

Note: *Steatosoma rufiventris* was recorded from both Argentina and Chile in the original description.

Reference: Cortés (1963: 246), notes on holotype and paratype in NHMUK.

Genus *VIBRISSOMYIA* Townsend, 1912

VIBRISSOMYIA Townsend, 1912b: 327. Type species: *Vibrissomyia lineata* Townsend, 1912 (= *Epalpus lineolata* Bigot, 1888), by original designation [Peru].

VIBRISSOMYIA. Incorrect subsequent spelling of *Vibrissomyia* Townsend, 1912 (Cortés 1980: 106).

References: Townsend (1936b: 190), diagnosis of adults and immatures of Cuphoceratini and key to genera (including *Vibrissomyia*); Townsend (1939a: 218), redescription; Cortés (1951b: 251), in key to Chilean genera of Tachinini with strong setae on the lower parafacial; Cortés (1967b: 16), key to separate *Vibrissomyia*, *Epalpodes* Townsend, and *Deopalpus* Townsend (as *Spanipalpus* Townsend); Cortés and Campos (1971: 26, 1974: 116) and Cortés (1984: 382), in keys to tachinid genera of Tarapacá and Antofagasta regions; Cortés (1986: 144), in key to tachinid genera of Aysén and Magallanes regions; González (1992a: 56, 66), in key to Chilean genera of “Cuphocerini”, key to species, diagnosis, notes, one new species.

concinata González, 1992.—Neotropical: South America (Chile).

Vibrissomyia concinnata González, 1992a: 66. Holotype male (UMCE). Type locality: Chile, Ñuble, Diguillín, Termas de Chillán.

erythrostroma (Bigot, 1888).—Neotropical: South America (Chile).

Epalpus erythrostroma Bigot, 1888b: 95. Holotype female (NHMUK). Type locality: Chile.

Reference: Brauer (1898: 504), taxonomic notes.

lineolata (Bigot, 1888).—Neotropical: South America (Argentina, Chile, Peru).

Epalpus lineolatus Bigot, 1888b: 94. Holotype male (NHMUK). Type locality: Chile.

Vibrissomyia lineata Townsend, 1912b: 328. Holotype female (USNM). Type locality: Peru, Puno, high puna of Lake Titicaca Region, Tirapata, ca. 13,000 ft.

Vibrissomyia albilineata Blanchard, 1943c: 152. Syntypes, unspecified number and sex (MLPA). Type locality: Argentina, Santa Cruz, Parque Nacional Los Glaciares, Valle del Río Túnel [as “Valle Tunel, Santa Cruz”, ca. 49°23'S, 72°56'W].

References: Brauer (1898: 503), taxonomic notes on *Epalpus lineolatus*; Cortés (1945c: 29), notes, synonymy of *Vibrissomyia lineata* and *Vibrissomyia albilineata* with *Epalpus lineolatus*, first records from Argentina and Peru.

notata Cortés, 1967.—Neotropical: South America (Argentina, Chile).

Vibrissomyia notata Cortés, 1967b: 14. Holotype male (EEAM). Type locality: Chile, Metropolitana de Santiago, Cordillera, Cajón del Río Maipo, El Yeso, 2200–2500 m.

Reference: Cortés (1979: 81), first record from Argentina.

pullata Cortés, 1951.—Neotropical: South America (Chile).

Vibrissomyia pullata Cortés, 1951b: 260. Holotype male (MNNC). Type locality: Chile, Metropolitana de Santiago, Cordillera, El Canelo [ca. 33°35'S, 70°27'W].

Unplaced species of Tachinini

aurea Giglio-Tos, 1893.—Not Chile [Mexico]. **Comb. nov.**

Saundersia aurea Giglio-Tos, 1893: 3. Type(s), male (MZUT). Type locality: Mexico.

Note: Guimarães (1971: 64) listed *Saundersia aurea* as an unrecognised species of *Epalpus* Rondani. This species name is a senior secondary homonym of *Eusignosoma aureum* Townsend, 1914 (a Peruvian species) when the two names are placed together in *Epalpus*. Guimarães (1971: 64) was aware of this homonymy but chose not to rename the junior homonym “at present”. DMW examined a male in MZUT labelled as lectotype by P. Arnaud Jr., but unpublished, and called it a “*Trichoepalpus* with bristles on pfc [parafacial]”. This was a preliminary determination in a lineage that has yet to be satisfactorily delineated. We remove *Saundersia aurea* from *Epalpus* and leave it unplaced in Tachinini until it can be placed to genus with more confidence.

References: Giglio-Tos (1894: 492), redescription, number of specimens in original type series (2 males and 1 ?female) and type localities (Mexico and “Angang” [Mexico, Michoacán, Angangueoven]) given; Guimarães (1971: 64), as unrecognised species of *Epalpus* Rondani; Papavero and Ibáñez-Bernal (2001: 144), notes on type series and current name; Zetina et al. (2018: 33), erroneous listing of *Saundersia aurea* Giglio-Tos and *Eusignosoma aureum* Townsend as synonyms instead of homonyms with combined distribution of Mexico and Peru (the latter in error).

Unplaced tribe of Tachinidae

Tribe MYIOPHASIINI

The New World tribe Myiophasiini currently comprises nine genera and ca. 40 species, with most species in *Gnadochaeta* Macquart (O'Hara et al. 2020). Myiophasiines are mostly or entirely parasitoids of larval weevils (Curculionidae *s. lato*) (Guimarães 1977; Arnaud 1978) and have been assigned to the Dexiinae by some authors (e.g., Sabrosky and Arnaud 1965; Guimarães 1977) and the Tachininae by others (e.g., Mesnil 1966; Tschorsnig 1985; O'Hara and Wood 2004). Mesnil (1966: 882) treated the group as a basal lineage of Tachininae and the phylogenetic analyses of Cerretti et al. (2014) and Stireman et al. (2019) placed it as an early branch of the Tachinidae, basal to the four recognised subfamilies. It likely shares this basal position with a sister lineage, the Macquartini (Stireman et al. 2019). If this basal lineage is recognised as a subfamily then the name Macquartinae will apply (Sabrosky 1999) but we are hesitant to take this step without further confirmatory evidence. For the present we regard Myiophasiini as unplaced in the Tachinidae.

Genus *GNADOCHAETA* Macquart, 1851

GNADOCHAETA Macquart, 1851: 200 [also 1851: 227] (see note). Type species: *Gnadochaeta coerulea* Macquart, 1851, by original designation [Brazil].

ANGIORHINA Brauer & Bergenstamm, 1889: 163 [also 1890: 95]. Type species: *Tachina crudelis* Wiedemann, 1830, by monotypy [West Indies].

MYIOPHASIA Brauer & Bergenstamm, 1891: 362 [also 1891: 58]. Type species: *Tachina aenea* Wiedemann, 1830 (junior primary homonym of *Tachina aenea* Meigen, 1824; = *Myiophasia australis* Townsend, 1916), by monotypy [Uruguay].

PSEUDOCLISTA Brauer & Bergenstamm, 1893: 104 [also 1893: 192]. Type species: *Pseudoclista atra* Brauer & Bergenstamm, 1893, by original designation [Brazil].

ANGIORRHINA. Incorrect subsequent spelling of *Angiorhina* Brauer & Bergenstamm, 1889 (Guimarães 1971: 23, 253, 269).

GNADOCHOETA. Incorrect original spelling of *Gnadochaeta* Macquart, 1851 (Macquart 1851: 200, see note).

Notes: There are two original spellings for *Gnadochaeta* in Macquart (1851): *Gnadochoeta* in the text (pp. 200–201) and index (p. 291) and on plate 21, and *Gnadochaeta* in the figure explanation (p. 286). O'Hara and Wood (2004: 277) were unaware of the latter spelling in the original publication and treated it as an incorrect subsequent spelling in prevailing usage. As explained in Evenhuis et al. (2016: 66): “They [O'Hara and Wood 2004] used ICZN Code Art. 33.3.1 to treat *Gnadochaeta* as the original spelling in prevailing usage, but by doing so they implicitly acted as First Reviser in selecting *Gnadochaeta* as the correct original spelling”.

Macquart (1851: 201) noted about his new genus *Gnadochaeta*, “Le type est du Brésil” [“The type is from Brazil”]. This statement is accepted as a type species designation for *Gnadochaeta* of the single included species, *Gnadochaeta coerulea* Macquart, from Brazil.

References: Coquillett (1910: 506, 572), type species of *Angiorhina* and *Myiophasia*; Aldrich (1934: 6, 165), in key to Patagonian genera, synonymy, taxonomic notes (as *Myiophasia*); Townsend (1936b: 116, 121, 124), diagnosis of adults and immatures of Dexillini and key to genera (including *Angiorhina*); diagnosis of adults and immatures of Trichoprosopini and key to genera (including *Gnadochaeta*); diagnosis of adults and immatures of Myiophasiini and key to genera (including *Myiophasia* and *Pseudoclista*); Townsend (1938: 274, 297, 307, 309), redescrptions of *Angiorhina*, *Gnadochaeta*, *Myiophasia* and *Pseudoclista*.

antennalis (Aldrich, 1934).—Neotropical: South America (Argentina, Chile).

Myiophasia antennalis Aldrich, 1934: 167. Holotype male (NHMUK). Type locality: Argentina, Río Negro, Lago Nahuel Huapí, Puerto Blest.

Note: *Myiophasia antennalis* was recorded from both Argentina and Chile in the original description.

solitaria (Aldrich, 1934).—Neotropical: South America (Chile).

Myiophasia solitaria Aldrich, 1934: 168. Holotype female (USNM). Type locality: Chile, Araucanía, Malleco, Angol.

Unplaced genus of Tachinidae

Genus *MARNEFIA* Cortés, 1982

MARNEFIA Cortés, 1982: 142. Type species: *Marnefia mirifica* Cortés, 1982, by original designation [Chile].

mirifica Cortés, 1982.—Neotropical: South America (Chile). (Fig. 6f)

Marnefia mirifica Cortés, 1982: 143. Holotype (MEUC). Type locality: Chile, Valparaíso, Valparaíso, Viña del Mar, El Salto, Jardín Botánico Nacional.

Unplaced species of Tachinidae

casanuevai Cortés, 1945.—Neotropical: South America (Chile).

Phorocera casanuevai Cortés, 1945d: 160. Holotype male (MEUC). Type locality: Chile, Valparaíso, Marga Marga, Limache.

References: Cortés (1945d: 159), in key to Chilean species of *Phorocera* Robineau-Desvoidy, 1830 (*s. lato*) and *Parasetigena* Brauer & Bergenstamm; Cortés (1950: 10), in key to Chilean species of *Phorocera* (*s. lato*); Guimarães (1971: 161), listed as an unrecognised species of Exoristini.

porteri Reed, 1907.—Neotropical: South America (Chile).

Tachina porteri Reed, 1907: 1046. Syntypes, males and females (not located). Type locality: Chile, Biobío, Concepción [province].

Note: C.S. Reed (1907: 1046) referred to *Tachina porteri* as a manuscript name of E.C. Reed (his father, see history section) but gave descriptive details that made the name available from his paper. Brèthes (1910: 67) gave a more formal description of the species under the name “*Exorista porteri* (Reed) Brèthes”. The name was correctly attributed to C.S. Reed by Cortés and Hichins (1969: 90) but was attributed to Brèthes by Guimarães (1971: 215).

Reference: Guimarães (1971: 215), treated as *Exorista porteri* Brèthes, 1910 and listed as an unplaced species of Exoristinae (as “Goniinae”).

Nomina dubia of Tachinidae

albomaculata Robineau-Desvoidy, 1863.—Neotropical: South America (Chile).

Peleteria albomaculata Robineau-Desvoidy, 1863: 622 (as “*albo-maculata*, Macq.”). Type(s), female (MNHN, 1 specimen with number MNHN-ED-ED8298, see note). Type locality: Chile.

Note: Robineau-Desvoidy (1863: 622) noted that this species was listed as *Echinomya albomaculata* Macquart in the catalogue of the Muséum and was labelled as such in the collection. Macquart did not publish a description of it and hence the name *Peleteria albomaculata* dates

from Robineau-Desvoidy's (1863: 622) description of it. The single specimen in MNHN is coded with the name "*Echinomyia albomaculata* Macquart" and has not been recognised and photographed as the name-bearing type.

References: Cortés (1946: 185), listed under "Species *incertae sedis*" at end of Tachinidae as "*Peleteria albomaculata* Macquart [*nomen nudum*]; Robineau-Desvoidy ... 1863"; Cortés and Hichins (1969: 90), listed under "Especies excluidas de la lista (*incertae sedis*)" as "*Peleteria albomaculata* Macquart *nomen nudum*" followed by "*Peleteria albomaculata* Robineau-Desvoidy 1863"; Guimarães (1971: 215), listed as an unrecognised species of Tachinidae.

lateralis Robineau-Desvoidy, 1863.—Neotropical: South America (Chile).

Faurella lateralis Robineau-Desvoidy, 1863: 664. Type(s), female (no specimens listed in MNHN database). Type locality: Chile.

References: Cortés (1946: 185), listed under "Species *incertae sedis*" at end of Tachinidae; Cortés and Hichins (1969: 90), listed under "Especies excluidas de la lista (*incertae sedis*)"; Guimarães (1971: 216), listed as an unrecognised species of Tachinidae.

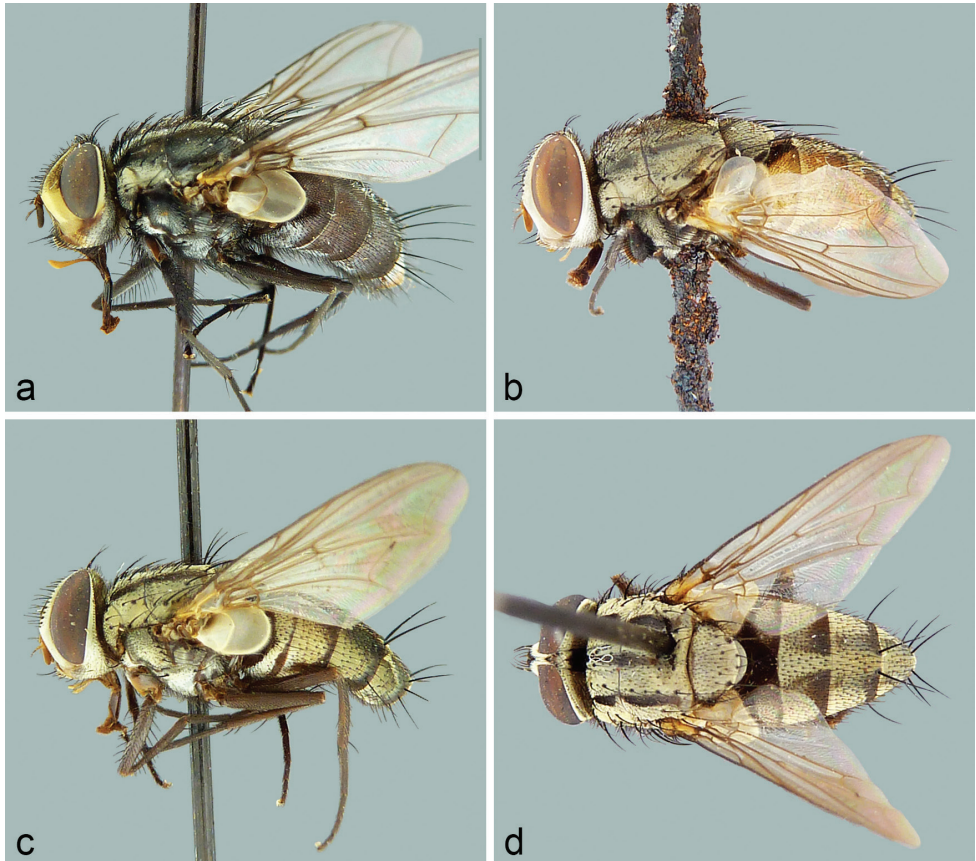


Figure 3. *Billaea* species (Dexiinae, Dexiini), habitus images **a** *B. aurifrons* (Townsend), comb. nov. ♂ (Peru) [holotype] **b** *B. rufescens* O'Hara & Wood, nom. nov., comb. nov. ♂ (Peru) [syntype] **c, d** *B. triquetrus* O'Hara & Wood, nom. nov., comb. nov. ♂ (Peru) [holotype].

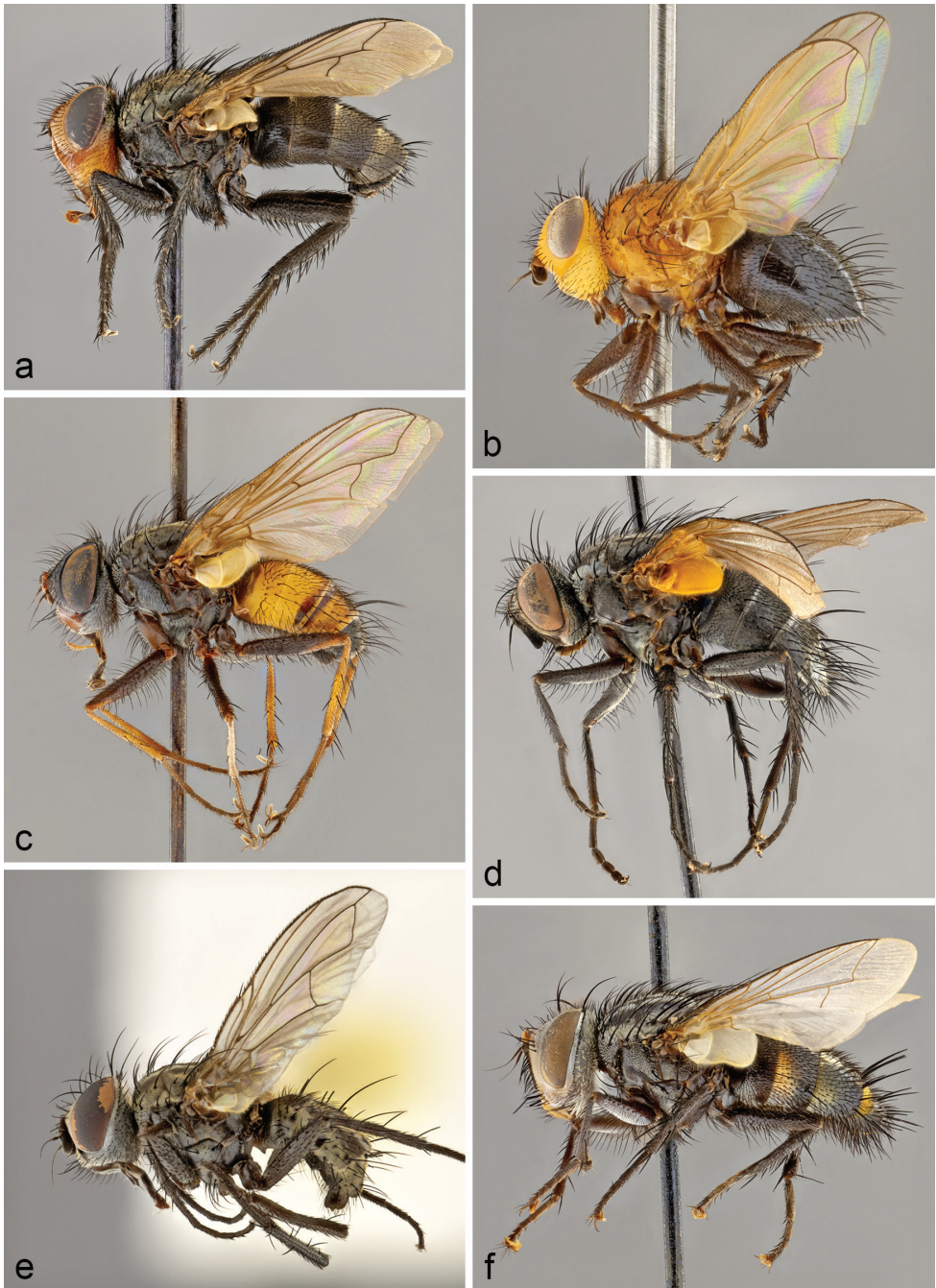


Figure 4. Habitus images **a** *Oligooestrus oestroideus* Townsend ♂ (Dexiinae, Dexiini) (Chile) [CNC487480], 6.0 mm **b** *Gonzalezodoria gonioides* Cortés ♀ (Dexiinae, Dufouriini) (Chile) [CNC1546958], 4.9 mm **c** *Xanthobasis rufescens* (Blanchard) ♂ (Dexiinae, Eutrixini) (Chile) [CNC1142102], 7.0 mm **d** *Admontia calyptata* (Aldrich) ♀ (Exoristinae, Blondeliini) (Chile) [CNC1143224], 7.5 mm **e** *Steleoneura australis* (Aldrich) ♀ (Exoristinae, Blondeliini) (Chile) [CNC487608], 2.5 mm **f** *Chetogena hichinsi* (Cortés) ♂ (Exoristinae, Exoristini) (Chile) [CNC1546959], 9.1 mm.

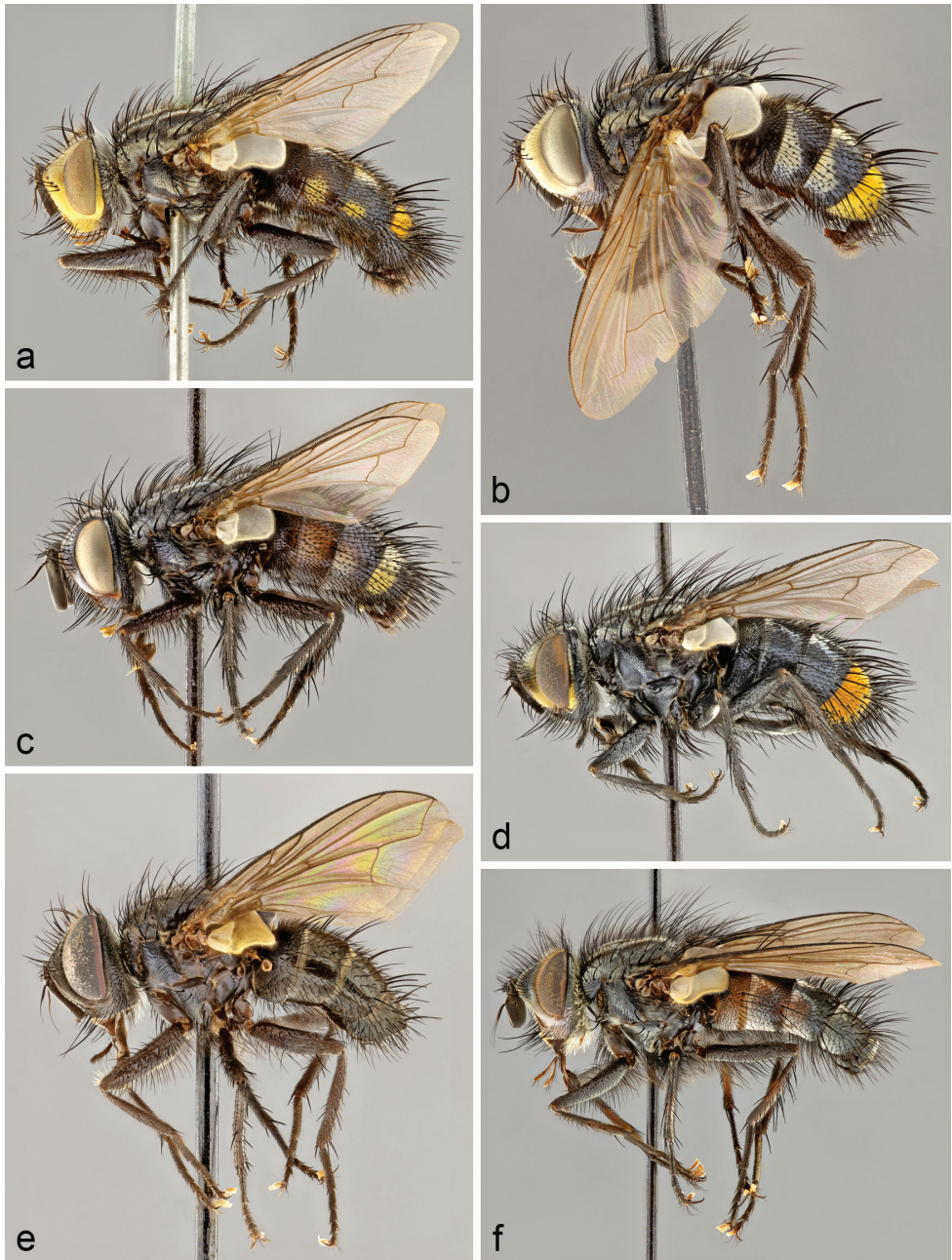


Figure 5. Habitus images **a** *Chetogena porteri* (Brèthes), comb. nov. ♂ (Exoristinae, Exoristini) (Chile) [CNC1546960], 11.0 mm **b** “*Phorocera chilensis* Cortés” ♂ (Exoristinae, unplaced species of Eryciini) (Chile) [CNC1546961], 6.7 mm **c** “*Phorocera elisae* Cortés” ♂ (Exoristinae, unplaced species of Eryciini) (Chile) [CNC1143104], 8.3 mm **d** *Patelloa tanumeana* (Townsend), comb. nov. ♂ (Exoristinae, Goniini) (Chile) [CNC487488], 10.9 mm **e** “*Phorocera negrensis* Aldrich” ♂ (Exoristinae, unplaced species of Goniini) (Argentina) [CNC1546962], 4.9 mm **f** “*Phorocera bullocki* Aldrich” ♂ (Exoristinae, unplaced species of Winthemiini) (Chile) [CNC1143251], 12.5 mm.

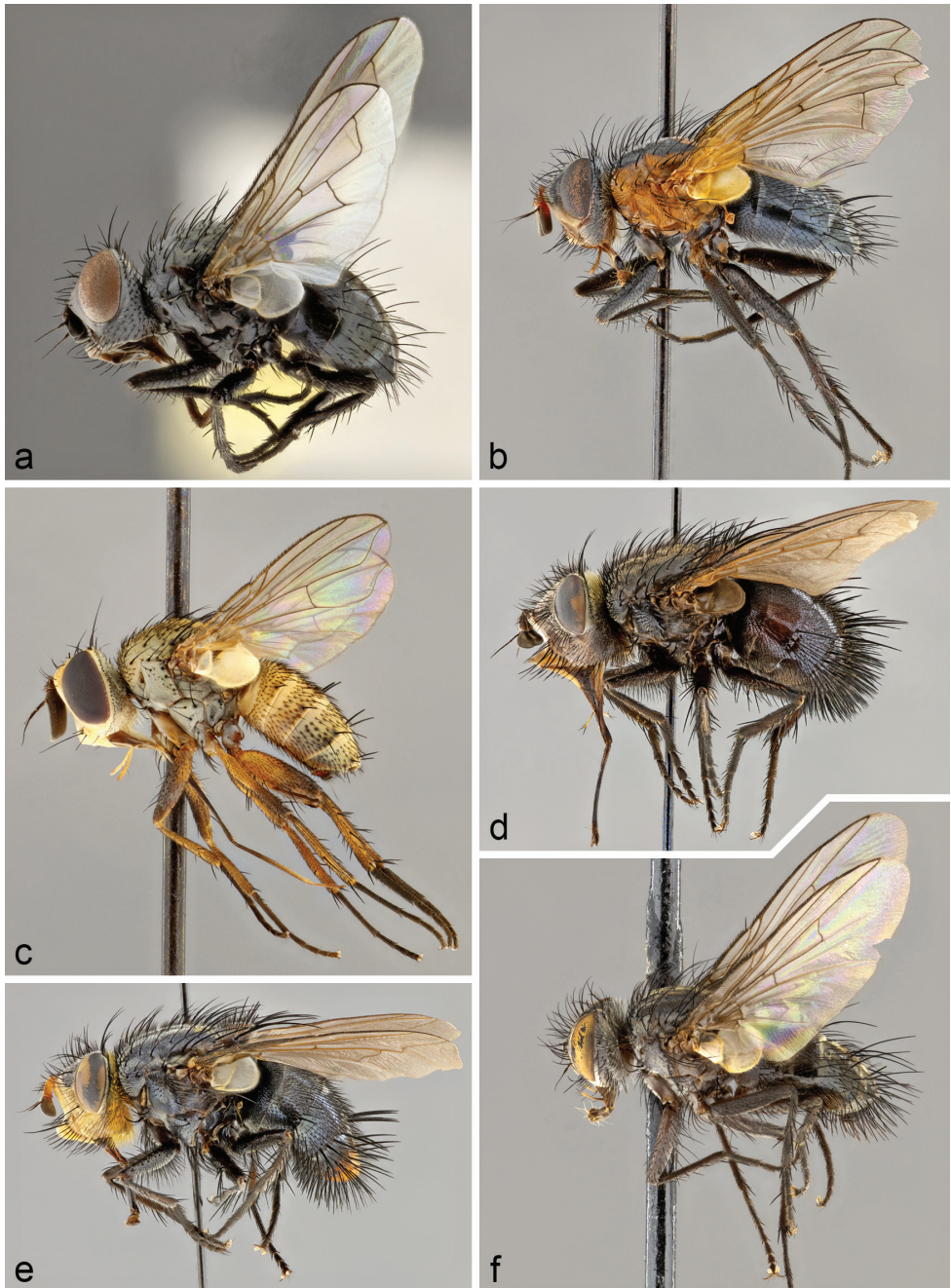


Figure 6. Habitus images **a** *Clastoneura brevicornis* Aldrich ♀ (Tachininae, Graphogastrini) (Chile) [CNC487612], 3.2 mm **b** *Xanthopelta scutellaris* Aldrich ♀ (Tachininae, Polideini) (Chile) [CNC487642], 7.6 mm **c** *Siphona* (*Siphona*) sp. ♂ (Tachininae, Siphonini) (Chile) [CNC497678], 4.0 mm **d** *Chaetoepalpus coquilleti* Vimmer & Soukup (Tachininae, Tachinini) ♀ (Chile) [CNC1546963], 12.4 mm **e** *Chiloepalpus callipygus* (Bigot) (Tachininae, Tachinini) ♀ (Argentina) [CNC1546964], 12.9 mm **f** *Marnefia mirifica* Cortés (Tachinidae, unplaced genus) ♂ (Chile) [CNC1546965], 4.2 mm.

Lectotype designations

In the interests of nomenclatural stability, DMW designates below four lectotypes for Macquart species based on his examination and labelling of the specimens in the 1980s. The lectotypes are housed in MNHN and their data and images can currently be accessed by searching the Diptera collection database at: https://science.mnhn.fr/institution/mnhn/collection/ed/item/search?lang=en_US.

***Echinomyia pygmaea* Macquart, 1851: 143 [also 1851: 170].**

Described from an unspecified number of females from “Chili” [Chile] collected by “M. Pissis” [Monsieur Pissis; i.e., Pierre Joseph Aimé Pissis, see history section] and deposited in “Muséum” [MNHN].

The online MNHN database records a female holotype in the Macquart collection for *Echinomyia pygmaea* (number MNHN-ED-ED8307) based on a holotype determination label that DMW attached to the specimen in 1980. However, Macquart did not restrict the name-bearing type to a single specimen and no lectotype fixation has been published subsequently. Cortés (1963: 248) examined a single female in MNHN, presumably the same specimen later examined by DMW, but did not explicitly refer to it as a name-bearing type and therefore did not fix it as lectotype.

In the interests of nomenclatural stability and to restrict the name to a single specimen, female syntype MNHN-ED-ED8307 in MNHN is hereby designated by DMW as lectotype of *Echinomyia pygmaea* Macquart, 1851.

The current combination for this species is *Peleteria pygmaea* (Macquart, 1851).

***Gonia chilensis* Macquart, 1844: 50 [also 1844: 207].**

Described from an unspecified number of females from “Chili” [Chile] collected by “M. Gay” [Monsieur Gay; i.e., Claude Gay, see history section] and deposited in “Muséum” [MNHN] and from Cuba collected by “M. de la Sagra” [Monsieur de la Sagra; i.e., Ramón de la Sagra] in “Muséum” [MNHN].

The online MNHN database records a female lectotype (from Chile, number MNHN-ED-ED8332) and three female paralectotypes (two from Chile [MNHN-ED-ED8333 and MNHN-ED-ED8334] and one from Cuba [MNHN-ED-ED8335]) for *Gonia chilensis* in the Macquart collection based on labels that DMW attached to these specimens in 1982 (the lectotype) and 1985 (the paralectotypes). However, the lectotype designation was not published.

In the interests of nomenclatural stability and to restrict the name to a single specimen, female syntype MNHN-ED-ED8332 from Chile in MNHN is hereby designated by DMW as lectotype of *Gonia chilensis* Macquart, 1844.

The current combination for this species is *Gonia pallens* Wiedemann, 1830.

***Masicera auriceps* Macquart, 1844: 59 [also 1844: 216].**

Described from an unspecified number of males from “Brésil ou du Chili” [Brazil or Chile] collected by “M. Gaudichand” [Monsieur Gaudichaud (as “Gaudichand”, typesetter error); i.e., Charles Gaudichaud-Beaupré, see history section] and deposited in “Muséum” [MNHN].

Guimarães (1983: 23) reported that the “type” of *Masicera auriceps* is “presumably lost” and treated the species as unrecognised. However, the online MNHN database records four male type specimens in the Macquart collection with numbers MNHN-ED-ED8355 to MNHN-ED-ED8358. The database has the first male as lectotype and the other three males as paralectotypes based on labels that DMW attached to these specimens in 1982 (the lectotype) and 1985 (the paralectotypes). However, the lectotype designation was not published. The database has Brazil as the country of origin but we have been unable to verify the restriction of the type locality to either of the two cited countries, Brazil or Chile.

In the interests of nomenclatural stability and to restrict the name to a single specimen, male syntype MNHN-ED-ED8355 in MNHN is hereby designated by DMW as lectotype of *Masicera auriceps* Macquart, 1844.

The current combination for this species is *Lespesia auriceps* (Macquart, 1844).

***Prosopochaeta nitidiventris* Macquart, 1851: 184 [also 1851: 211].**

Described from an unspecified number of males from “Chili” [Chile] collected by “M. Gay” [Monsieur Gay; i.e., Claude Gay, see history section] and deposited in “Muséum” [MNHN].

Aldrich (1934: 118) reported that the “types” of *Prosopochaeta nitidiventris* “apparently are lost”, but this was in error. Townsend (1938: 299) mentioned a “Ht male” for *P. nitidiventris* from Coquimbo in “Lille or lost” but this is not accepted as a lectotype fixation because the specimen in question is not distinguishable from the other males in the type series (in MNHN, not Lille). Cortés (1963: 249) reported examining five (type) specimens in poor condition in MNHN.

The online MNHN database records three male type specimens in the Macquart collection with numbers MNHN-ED-ED8367 to MNHN-ED-ED8369. The database has specimen MNHN-ED-ED8367 as lectotype based on a lectotype label that DMW attached to this specimen in 1982. The other two specimens were labelled as paralectotypes by DMW in 1985. However, the lectotype designation was not published.

In the interests of nomenclatural stability and to restrict the name to a single specimen, male syntype MNHN-ED-ED8367 in MNHN is hereby designated by DMW as lectotype of *Prosopochaeta nitidiventris* Macquart, 1851.

The current combination for this species is *Prosopochaeta nitidiventris* Macquart, 1851.

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A date of publication is given at the end of each citation in square brackets, if known. Dates that we have determined are accompanied by a brief explanation of where they were given, as follows: OS stands for “original source” and if not accompanied by further details then the date is given in the work itself; OS followed by details in parentheses indicates that a date is given somewhere else in the same volume,

perhaps in a wrapper of the issue, in the volume's table of contents, or on the last page of the issue or volume. Dates originating from other works (e.g., Evenhuis 1997) are credited to the sources we used. In some instances we cite the date an issue was received by the Neatby Library of the Canadian Agriculture Library, as indicated by a CAL date stamp. Some authors use "31 December+" to indicate that a date has not been determined beyond year but we use this only when we are citing it from another source (e.g., "[31 December+, Evenhuis et al. 2016: 250]") and give no date for works for which we could not find publication dates.

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Index

Listed here are the taxonomic names of the Tachinidae of Chile that appear in the catalogue, including valid names, synonyms, emendations, incorrect original and subsequent spellings, and *nomina nuda*. Type species, species mentioned in notes, and senior homonyms are not listed unless the species occurs in Chile. Taxon and author names are formatted as follows:

- 1) Names of subfamilies and tribes are given in capitals.
- 2) Valid generic and subgeneric names are given in bold with subgeneric names followed by “subg.”.
- 3) Valid species names are given in plain type.
- 4) Non-valid names (e.g., synonyms, *nomina nuda*, misidentifications, unjustified emendations) are given in italics.
- 5) Parentheses around an author’s name indicate that the present genus and species combination is not the original one.
- 6) Valid species-group names agree in gender with their valid generic names. Non-valid species names appear with their original endings as they do in the catalogue.

Author abbreviations: B. & B., Brauer & Bergenstamm; R.-D., Robineau-Desvoidy. Nomenclatural abbreviations: incorrect orig. spell., incorrect original spelling; incorrect sub. spell., incorrect subsequent spelling.

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