# Morphology and phylogenetic analysis of five deepsea golden gorgonians (Cnidaria, Octocorallia, Chrysogorgiidae) in the Western Pacific Ocean, with the description of a new species 

Yu Xu ${ }^{1,4^{*}}$, Zifeng Zhan ${ }^{1,2,3^{*}}$, Kuidong $\mathrm{Xu}^{1,2,3,4}$<br>I Laboratory of Marine Organism Taxonomy and Phylogeny, Institute of Oceanology, Chinese Academy of Sciences, Qingdao 266071, China 2 Laboratory for Marine Biology and Biotechnology, Pilot National Laboratory for Marine Science and Technology (Qingdao), Qingdao 266237, China 3 Center for Ocean Mega-Science, Chinese Academy of Sciences, Qingdao 266071, China 4 University of Chinese Academy of Sciences, Beijing 100049, China<br>Corresponding author: Kuidong Xu (kxu@qdio.ac.cn)

Academic editor: B.W.Hoeksema | Received 10 April 2020 | Accepted 20 September 2020 | Published 9 November 2020
http://zoobank.org/8957A11D-0098-4990-B073-A1999A8AFBC6
Citation: Xu Y, Zhan Z, Xu K (2020) Morphology and phylogenetic analysis of five deep-sea golden gorgonians (Cnidaria, Octocorallia, Chrysogorgiidae) in the Western Pacific Ocean, with the description of a new species. ZooKeys 989: 1-37. https://doi.org/10.3897/zookeys.989.53104


#### Abstract

Explorations of seamounts in the Western Pacific Ocean and South China Sea resulted in collecting 18 specimens of golden gorgonians. Based on the morphology and the genetic analysis of mtMutS, they are described as one new species, Chrysogorgia carolinensis sp. nov., and four known species, including Chrysogorgia dendritica Xu, Zhan \& Xu, 2020, Metallogorgia melanotrichos (Wright \& Studer, 1889), Metallogorgia macrospina Kükenthal, 1919, and Pseudochrysogorgia bellona Pante \& France, 2010. Chrysogorgia carolinensis belongs to the Chrysogorgia "group A, Spiculosae" with rods or spindles distributed in the pol-yp-body wall and tentacles, and differs from all of its congeners except $C$. dendritica by the $1 / 3 \mathrm{~L}$ branching sequence and amoeba-shaped sclerites at the basal polyp body. The mtMutS sequence of $C$. carolinensis $\mathbf{s p}$. nov. has six deletion mutations compared to those of its congeners, supporting the establishment of the new species. Although no genetic variability was observed between the closely related species C. dendritica and C. abludo Pante \& Watling, 2012, the former is different from the latter by the apparently irregular sclerites in the polyp body wall. The two specimens of Metallogorgia melanotrichos match well with the original description except for relatively larger polyps, while the $M$. macrospina specimens have slightly


[^0]smaller polyps than the holotype. The juvenile of Metallogorgia has an obvious morphological difference with the adults in the colony shape and branches, but they can be unified by the same polyps and sclerites as well as mitochondrial MutS sequences. Thus, the generic diagnosis of Metallogorgia is slightly extended to include the morphology of juveniles. The morphology of Pseudochrysogorgia bellona Pante \& France, 2010, as a new record for the South China Sea, matches well with that of the original description. In the phylogenetic trees, the Chrysogorgia species are separated into two clades, and while Metallogorgia and Pseudochrysogorgia formed a sister clade.

## Keywords

Anthozoa, Chrysogorgia dendritica, Chrysogorgia carolinensis, Metallogorgia, Pseudochrysogorgia, seamount

## Introduction

Chrysogorgiids are found in all major oceans from Iceland to Antarctica, and they are conspicuous members of deep-water benthic assemblages (Pante et al. 2012). The family Chrysogorgiidae Verrill, 1883 currently contains 14 genera, with 12 genera having been analyzed by molecular phylogenetic methods with and six of them forming a monophyletic clade (called the Monophyletic Chrysogorgiidae Clade, or MCC), including Chrysogorgia Duchassaing \& Michelotti, 1864, Pseudochrysogorgia Pante \& France, 2010, Iridogorgia Verrill, 1883, Rhodaniridogorgia Watling, 2007, Radicipes Stearns, 1883 and Metallogorgia Versluys, 1902 (Watling et al. 2011; Pante et al. 2012).

The genus Chrysogorgia Duchassaing \& Michelotti, 1864 contains 75 species and is distributed worldwide (Watling et al. 2011; Cairns 2018; Xu et al. 2020). It is characterized by four branching forms of the colony, including a single ascending spiral (clockwise or counterclockwise, bottlebrush-shaped colony), a single fan (planar colony), two or more fans emerging from a short main stem (bi- or multi-flabellate colony) and a bush of branches perched on top of a long straight stem (tree-shaped colony) (Pante and Watling 2012; Cordeiro et al. 2015; Xu et al. 2020). Based on the shapes of rods or scales in the polyp-body wall and tentacles, four groups have been recognized for the separation of Chrysogorgia species, including "group A, Spiculosae" with 42 species, "group B, Squamosae aberrantes" with 13 species, "group C, Squamosae typicae" with 19 species and "group D, Spiculosae aberrantes" with only one species (Versluys 1902; Cairns 2001; Cordeiro, Castro and Pérez 2015; Xu et al. 2019).

The genus Metallogorgia Versluys, 1902 is distinguished from Chrysogorgia by its distinctive monopodial stem with branchlets forming a sympodium pattern in strong branches, sclerites with no distinction in coenenchyme or polyps, and coenenchyme thin with few sclerites (Versluys 1902). The establishment of Metallogorgia was well supported by a phylogenetic analysis (Pante et al. 2012). At present, there are four species in this genus: M. melanotrichos (Wright \& Studer, 1889), M. macrospina Kükenthal, 1919, M. splendens (Verrill, 1883) and M. tenuis Pasternak, 1981 (Verrill 1883; Wright and Studer 1889; Kükenthal 1919; Pasternak 1981; Watling et al. 2011). The genus

Pseudochrysogorgia Pante \& France, 2010 only contains one species P. bellona Pante \& France, 2010, and has a close relationship with Metallogorgia (Pante and France 2010).

During an investigation of the seamount benthic diversity in the Western Pacific Ocean and the South China Sea, we obtained 18 specimens of golden gorgonians. Based on morphological and phylogenetic analyses, they were described as one new species Chrysogorgia carolinensis sp. nov., and four known species, including Chrysogorgia dendritica Xu, Zhan \& Xu, 2020, Metallogorgia melanotrichos (Wright \& Studer, 1889), Metallogorgia macrospina Kükenthal, 1919 and Pseudochrysogorgia bellona Pante \& France, 2010. Their phylogenetic positions are also discussed.

## Materials and methods

Specimen collection and morphological examination
One specimen of Chrysogorgia Duchassaing \& Michelotti, 1864 was collected from a seamount (tentatively named as M2) near the Mariana Trench by the ROV (remotely operated vehicle) FaXian (Discovery) in the tropical Western Pacific during the cruises of the R/V KeXue (Science) in 2016. One specimen of Pseudochrysogorgia Pante \& France, 2010 was initially collected from the Ganquan Plateau in the South China Sea in 2018, unfortunately only a few frozen fragments of this specimen and a picture were obtained. Ten specimens of Chrysogorgia and six specimens of Metallogorgia Versluys, 1902 were obtained from the seamounts (tentatively named as M5-M8) located on the Caroline Ridge during the cruises of the R/V KeXue (Science) in 2019. The Chrysogorgia and Metallogorgia specimens were photographed in situ before sampling, and photographed on board and then stored in $75 \%$ ethanol after collection. Small branches were detached and stored at $-80^{\circ} \mathrm{C}$ for molecular study.

The morphological terminology follows Bayer et al. (1983). The general morphology and anatomy were examined by using a stereo dissecting microscope. The sclerites of the polyps and branches were isolated by digestion of the tissues in sodium hypochlorite, and then were washed with deionized water repeatedly. Polyps and sclerites were air-dried and mounted on carbon double adhesive tape and coated for scanning electron microscopy (SEM) observation to investigate their structure. SEM scans were obtained and the optimum magnification was chosen for each kind of sclerite by using TM3030Plus SEM.

The type and voucher specimens have been deposited in the Marine Biological Museum of Chinese Academy of Sciences (MBMCAS) at Qingdao, China.

## DNA extraction and sequencing

Total genomic DNA was extracted from the polyps of each specimen using the TIANamp Marine Animal DNA Kit (Tiangen Bio. Co., Beijing, China) following the
manufacturer's instructions. PCR amplification for the mitochondrial genomic region 5'-end of the DNA mismatch repair protein - mutS - homolog (mtMutS) was conducted using primers AnthoCorMSH (5'-AGGAGAATTATTCTAAGTATGG-3'; Herrera et al. 2010) and Mut-3458R (5’-TSGAGCAAAAGCCACTCC-3’; Sánchez et al. 2003). PCR reactions were performed as described by Xu et al. (2019). PCR purification and sequencing were performed by TsingKe Biological Technology (TsingKe Biotech, Beijing, China).

## Genetic distance and phylogenetic analyses

The mtMutS gene in octocorals was selected for molecular identification and phylogenetic analyses. All the available mtMutS sequences of Chrysogorgia, Metallogorgia, Pseudochrysogorgia and the out-group species from related chrysogorgiid and plexaurid genera were downloaded from GenBank, and those from duplicate isolates or without associated publications or not identified to species level were omitted from the molecular analyses. To correct possible mistakes, all the selected sequences were visually inspected, and translated to amino acids (AA) to insure all the AA sequences did not include stop codons and suspicious substitutions. The nucleotide and AA sequences were aligned using MAFFT v. 7 (Katoh and Standley 2013) with the G-INS-i algorithm. With the guidance of the AA alignment, the nucleotide alignment was refined using BioEdit v7.0.5 (Hall 1999), and only the nucleotide alignment was used in the subsequent analyses. Genetic distances, calculated as uncorrected "p" distances within each species and among species, were estimated using MEGA 6.0 (Tamura et al. 2013).

For the phylogenetic analyses, only one known sequence was randomly selected from the conspecific sequences without genetic divergence (see Table 4). The TPM1uf+G evolutionary model was the best-fitted model for mtMutS, selected by AIC as implemented in jModeltest2 (Darriba et al. 2012). Maximum likelihood (ML) analysis was carried out using PhyML-3.1 (Guindon et al. 2010). For the ML bootstraps, we consider values $<70 \%$ as low, $70-94 \%$ as moderate and $\geq 95 \%$ as high following Hillis and Bull (1993). Node support came from a majority-rule consensus tree of 1000 bootstrap replicates. Bayesian inference (BI) analysis was carried out using MrBayes v3.2.3 (Ronquist and Huelsenbeck 2003) on CIPRES Science Gateway. Posterior probability was estimated using four chains running 10,000,000 generations sampling every 1000 generations. The first $25 \%$ of sampled trees were considered burn-in trees. Convergence was assessed by checking the standard deviation of partition frequencies (<0.01), the potential scale reduction factor (ca. 1.00), and the plots of $\log$ likelihood values (no obvious trend was observed over time). For the Bayesian posterior probabilities, we consider values $<0.95$ as low and $\geq 0.95$ as high following Alfaro et al. (2003). The museum voucher specimen and GenBank accession numbers of the mtMutS sequences were listed next to the species names in the phylogenetic trees (Figure 21).

## Results

## Class Anthozoa Ehrenberg, 1834

## Subclass Octocorallia Haeckel, 1866

Order Alcyonacea Lamouroux, 1812
Suborder Calcaxonia Grasshoff, 1999
Family Chrysogorgiidae Verrill, 1883

## Genus Chrysogorgia Duchassaing \& Michelotti, 1864

Diagnosis (based on Xu et al. 2020). Colony branching usually sympodial, occasionally monopodial, arising from a single ascending spiral (clockwise or counterclockwise, bottlebrush-shaped colony), a fan (planar colony), two fans emerging from a short main stem (biflabellate colony), or an unbranched main stem forming a tree-shaped colony. Axis with a metallic shine, dark to golden in color. Branch subdivided dichotomously or pinnately. Most polyps relatively large to the size of the branches they sit on, few in number and well separated from one another. Sclerites in the form of spindles, rods, scales, and rare plates with little ornamentation.

Type species. Chrysogorgia desbonni Duchassaing \& Michelotti, 1864, by monotypy.

Distribution. Worldwide in a depth range of 10-4492 m (Watling et al. 2011).

Chrysogorgia dendritica Xu, Zhan \& Xu, 2020
Figures 1-4; Table 1

Chrysogorgia dendritica Xu, Zhan \& Xu, 2020: 6-8, figs 2, 3.
Type locality. Kocebu Guyot in the Magellan Seamount chain, 1821 m depth.
Voucher specimens. MBM286353, station FX-Dive 71 ( $11^{\circ} 20.83^{\prime} \mathrm{N}$, $139^{\circ} 15.87^{\prime} \mathrm{E}$ ), a seamount (tentatively named as M2) near the Mariana Trench, depth $1375 \mathrm{~m}, 28$ March 2016. MBM286442, station FX-Dive $211\left(10^{\circ} 02.97^{\prime} \mathrm{N}\right.$, $140^{\circ} 10.48^{\prime} \mathrm{E}$ ), a seamount (tentatively named as M5) located on the Caroline Ridge, depth $1475 \mathrm{~m}, 29$ May 2019. MBM286443, station FX-Dive 211 ( $10^{\circ} 03.27^{\prime} \mathrm{N}$, $140^{\circ} 10.70^{\prime} \mathrm{E}$ ), a seamount (M5) located on the Caroline Ridge, depth $1387 \mathrm{~m}, 29$ May 2019. MBM286444, station FX-Dive 227 ( $10^{\circ} 37.92^{\prime} \mathrm{N}, 140^{\circ} 05.62^{\prime} \mathrm{E}$ ), a seamount (tentatively named as M8) located on the Caroline Ridge, depth $1702 \mathrm{~m}, 15$ June 2019. GenBank accession number: MT269888.

Extended diagnosis. Chrysogorgia "group A, Spiculosae" with 1/3L Branching sequence and a monopodial or a little zigzagging stem. Juvenile with a bottlebrush-like colony, while adult usually having a tree-shaped colony. Branches nearly perpendicular to stem, subdivided dichotomously. Polyps with a long neck and an expanded base.


Figure I. External morphology, polyps and sclerites of Chrysogorgia dendritica MBM286353 A, B the tree-shaped colony in situ and after collection (likely an adult), with a broken stem and a branching part $\mathbf{C}$ a single polyp under a light microscope $\mathbf{D}$ long polyp neck under SEM E sclerites in tentacles $\mathbf{F}$ sclerites of the polyp neck $\mathbf{G}$ sclerites in coenenchyme $\mathbf{H}$ sclerites at the basal polyp body. Scale bars: $10 \mathrm{~cm}(\mathbf{B})$; $1 \mathrm{~mm}(\mathbf{C}) ; 200 \mu \mathrm{~m}(\mathbf{D}) ; 50 \mu \mathrm{~m}(\mathbf{E}), 100 \mu \mathrm{~m}(\mathbf{F}, \mathbf{G}, \mathbf{H})$.

Rods and rare scales in tentacles, longitudinally arranged. Rods/spindles and elongate scales in polyp neck longitudinally arranged, coarse with many warts on surface. Scales and rare plates at the basal polyp body irregularly and alternately arranged, irregular and often amoeba-shaped. Scales in coenenchyme sparse and elongate, usually lobed with irregular edges.

Description. For morphological measurements, see Table 1.


Figure 2. External morphology, polyps and sclerites of Chrysogorgia dendritica MBM286442 A, B the bottlebrush-like colony in situ and after collection (likely a juvenile) $\mathbf{C}$ a single polyp under a light microscope $\mathbf{D}$ a single polyp under SEM E tentacular part under SEM $\mathbf{F}$ coenenchyme under SEM $\mathbf{G}$ sclerites in tentacles $\mathbf{H}$ sclerites extending to pinnules $\mathbf{I}$ sclerites of the polyp neck $\mathbf{J}$ sclerites in coenenchyme $\mathbf{K}$ sclerites at the basal polyp body. Scale bars: $10 \mathrm{~cm}(\mathbf{B}) ; 1 \mathrm{~mm}(\mathbf{C}, \mathbf{D}) ; 300 \mu \mathrm{~m}(\mathbf{E}, \mathbf{F}) ; 200 \mu \mathrm{~m}$ (G, I-K at the same scale), $100 \mu \mathrm{~m}(\mathbf{H})$.

Distribution. Kocebu Guyot, 1821 m (Xu et al. 2020); a seamount adjacent to the Mariana Trench and seamounts on the Caroline Ridge, 1375-1702 m depth.

Remarks. The four specimens match the holotype of Chrysogorgia dendritica Xu , Zhan $\& \mathrm{Xu}, 2020$ in having a monopodial stem and the same sclerite form, for example, rods and rare scales in tentacles, rods/spindles and elongate scales in polyp neck, irregular scales at the basal polyp body, and elongate scales in coenenchyme. Moreover, their mtMutS gene sequences are identical (see the genetic analysis below). Thus, we identified the four specimens as C. dendritica. The sclerites in the four


Figure 3. External morphology, polyps and sclerites of Chrysogorgia dendritica MBM286443 A, B the tree-shaped colony in situ and after collection (likely an adult) $\mathbf{C}$ a single polyp under SEM D sclerites in tentacles $\mathbf{E}$ sclerites at the basal polyp body $\mathbf{F}$ sclerites of the polyp neck $\mathbf{G}$ sclerites in coenenchyme. Scale bars: $10 \mathrm{~cm}(\mathbf{B}) ; 1 \mathrm{~mm}(\mathbf{C}) ; 300 \mu \mathrm{~m}$ (D-G at the same scale).
voucher specimens and the holotype showed some differences: (1) rod-like scales with an obvious medial contraction often present in MBM286353 and MBM286442, while rare in the holotype, MBM286443 and MBM286444 (Figures 1A, 2H vs. Figures 3D, 4D); (2) large cured spindles are rarely present in the polyp neck of the holotype, MBM286443 and MBM286444, while absent in the other two specimens (Figures 3F, 4F vs. Figures 1F, 2I); (3) scales at the basal polyp body of the holotype and MBM286444 are more irregular and amoeba-shaped than the other specimens (Figure 4 E vs. Figures $1 \mathrm{H}, 2 \mathrm{~K}, 3 \mathrm{E}$ ); and (4) scales in coenenchyme are more elongate in MBM286442 than the other specimens (Figure 2J vs. Figures 1G, 3G, 4G).


Figure 4. External morphology, polyps and sclerites of Chrysogorgia dendritica MBM286444 A, B the colony in situ and after collection (likely an intermediate state) $\mathbf{C}$ a single polyp under SEM D sclerites in tentacles $\mathbf{E}$ sclerites of the polyp neck $\mathbf{F}$ sclerites at the basal polyp body $\mathbf{G}$ sclerites in coenenchyme. Scale bars: $20 \mathrm{~cm}(\mathbf{B}) ; 2 \mathrm{~mm}(\mathbf{C}) ; 300 \mu \mathrm{~m}$ (D-G at the same scale).

However, these differences are minor and not constant, and we thus treate as the conspecific variation.

The four specimens of $C$. dendritica showed a series of growth stages, from bottle-brush-like colony (juvenile) to tree-shaped colony (adult). Considering the diameter size of the stem base and scars on the stem, the specimen MBM286442 is likely a
juvenile with a narrow stem and without scars, while the other specimens have wider stems and some old scars of the past branches (Table 1). The juvenile has a bottlebrushlike colony (Figure 2B), while the adult has a long monopodial stem with branches occurring on the top, forming a tree-shaped colony (Figures 1B, 3B). Compared to the changing colony shapes, the sclerite forms showed small variation in the growth stages and can be used as a main character to identify the species.

## Chrysogorgia carolinensis sp. nov.

http://zoobank.org/A37B30BA-6396-4679-9D39-1FFB51FCCD4D
Figures 5, 6; Table 1
Type material. Holotype. MBM286494, station FX-Dive 226 ( $10^{\circ} 38.18^{\prime} \mathrm{N}$, $140^{\circ} 04.08^{\prime} \mathrm{E}$ ), a seamount (tentatively named as M8) located on the Caroline Ridge, depth 1832 m, 14 June 2019. GenBank accession number: MT269889.

Paratypes. MBM286493, station FX-Dive 224 ( $10^{\circ} 37.63^{\prime} \mathrm{N}, 140^{\circ} 05.45^{\prime} \mathrm{E}$ ), depth $1509 \mathrm{~m}, 12$ June 2019. MBM286495, station FX-Dive 227 ( $10^{\circ} 37.92^{\prime} \mathrm{N}$, $140^{\circ} 05.62^{\prime} \mathrm{E}$ ), depth 1709 m. MBM286496, station FX-Dive 227 ( $10^{\circ} 37.92^{\prime} \mathrm{N}$, $140^{\circ} 05.62^{\prime} \mathrm{E}$ ), depth $1706 \mathrm{~m}, 15$ June 2019. MBM286497, station FX-Dive 227 ( $10^{\circ} 37.90^{\prime} \mathrm{N}, 140^{\circ} 05.62^{\prime} \mathrm{E}$ ), depth $1695 \mathrm{~m}, 15$ June 2019. MBM286498, station FXDive 227 ( $10^{\circ} 37.68^{\prime} \mathrm{N}, 140^{\circ} 05.48^{\prime} \mathrm{E}$ ), depth $1537 \mathrm{~m}, 15$ June 2019. MBM286499, station FX-Dive $227\left(10^{\circ} 37.60^{\prime} \mathrm{N}, 140^{\circ} 05.43^{\prime} \mathrm{E}\right)$, depth $1506 \mathrm{~m}, 15$ June 2019. They were all collected from a seamount (tentatively named as M8) located on the Caroline Ridge.

Diagnosis. Chrysogorgia "group A, Spiculosae" with $1 / 3 \mathrm{~L}$ branching sequence. Branches subdivided dichotomously, up to sixth order. Polyps only present in the end of terminal branchlets. Polyps large with pitcher shape, up to 8 mm long. Rods and spindles slender and coarse with many warts on surface in the back and base of tentacles. Scales amoeba-shaped, branched toward to any directions, irregularly and alternately arranged at basal polyp body. Scales rare, transversally arranged in coenenchyme.

Description. Specimen of holotype ca. 31 cm long and 14 cm wide excluding the holdfast (Figure 5C). Colony bottlebrush-shaped, with branching sequence 1/3L. Stem golden with metallic luster, ca. 1 mm in diameter at base. Branches subdivided dichotomously, up to sixth order, with distance between adjacent branches $6-12 \mathrm{~mm}$ long and orthostiche interval $19-35 \mathrm{~mm}$ long in measurements from all specimens. Branches up to 6 cm with the first branch internodes $7-15 \mathrm{~mm}$ long and terminal branchlets up to 2 cm long. Polyps only present in the end of the terminal branchlets. Polyps large with pitcher shape, some of them contracted and narrow at the base of tentacles, average 5 mm long with the terminal one up to 8 mm long, and $1-3 \mathrm{~mm}$ wide (Figure 5B, D, F). No polyps in axis internodes.

Rods and spindles slender and coarse with many warts on surface, some of them branched, rarely with one end a little flat, longitudinally arranged in the back of tentacles and usually forming eight distinct columns, and transversally or longitudinally arranged in the base of tentacles, measuring 107-814 $\times 10-78 \mu \mathrm{~m}$ (length $\times$ width,
Table I. Morphological comparisons between Chrysogorgia carolinensis sp. nov., Chrysogorgia dendritica Xu, Zhan \& Xu, 2020 and Chrysogorgia abludo Pante \&
Watling, 2012, including detailed morphological measuring data of five specimens of $C$. dendritica.

| Characters/species | Chrysogorgia dendritica |  |  |  |  | Chrysogorgia carolinensis $\mathbf{s p}$. nov.holotype | Chrysogorgia abludo |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Specimen | holotype | MBM286353 | MBM286442 | MBM286443 | MBM286444 |  | holotype | paratype |
| Group type | A |  |  |  |  | A | A |  |
| Branching sequence | 1/3L |  |  |  |  | 1/3L | 1/3, 1/4L, irregular |  |
| Axis | monopodial, or a little zigzagging |  |  |  |  | sympodial | monopodial |  |
| Colony shape | tree-shaped | tree-shaped | bottlebrush-like | tree-shaped | a little tree-shaped | bottlebrush-shaped | bottlebrushshaped | tree-shaped |
| Colony height (cm) | 57 | 85.5 | 55 | 50 | 110 | 31 | 16 | 50 |
| Basal stem width (mm) | 2 | 7 | 1.2 | 3 | 3 | 1 | No data | 2.2 |
| Interbranch distance (mm) | 16-22 | 18 | 5-22 | 11-15 | 12-24 | 6-12 | 4.3-6.8 | 7.5-15.0 |
| Orthostiche interval (mm) | 50-55 | 50-55 | 37-47 | 36-47 | 44-61 | 19-35 | No data | No data |
| First branch internode (mm) | 15-20 | 14-22 | 11-20 | 9-23 | 20-30 | 7-15 | 6.1-11.0 | 16 |
| Polyps on internodes | 1-5 | 2-3 | 1-3 | 1-4 | 1-4 | 0 | 1-2 | No data |
| Polyps on terminal branchlets | 1-6 | 1-4 | 1-4 | 1-8 | 1-8 | 1 | 1-3 | 1-6 |
| Polyps height (mm) | 3 | 3-5 | 1.5-2.0 | 2-3 | 3-4 | 3-8, average 5 | 0.8-2.2 | 0.8-2.2 |
| Sclerites in coenenchyme ( $\mu \mathrm{m}$ ) | flat elongate scales often with lobed edges | 82-200 $\times 10-96$ | $139-420 \times 16-112$ | $92-266 \times 15-57$ | $53-379 \times 10-34$ | elongate scales occasionally with lobed edges | small rugged scales with less lobed edges |  |
| Sclerites in body wall ( $\mu \mathrm{m}$ ) | scales, rods and spindles | 68-190 $\times 7-58$ at basal body; 171-516 $\times$ $20-55$ in neck | $\begin{array}{\|c\|} \hline 100-306 \times 20-72 \text { at } \\ \text { basal body; } 121-353 \times \\ 12-56 \text { in neck } \\ \hline \end{array}$ | $80-229 \times 13-82$ at basal body; 207-614 $\times$ $14-76$ in neck | $\begin{array}{\|c\|} \hline 39-236 \times 8-128 \text { at } \\ \text { basal body; } 137-590 \times \\ 12-98 \text { in neck } \\ \hline \end{array}$ | scales, rods and spindles | scales and rods |  |
| Sclerites in tentacles ( $\mu \mathrm{m}$ ) | scales and rods | 74-135 + 4-32 | 105-365 × 6-45 | 75-429 $\times 10-43$ | 93-275 $\times 24$-93 | rods and spindles | rods |  |
| Distribution | Kocebu Guyot | an unnamed seamount adjacent to the Mariana Trench | unnamed seamounts on the Caroline Ridge |  |  | an unnamed seamount on the Caroline Ridge | North Atlantic |  |
| References | Xu et al. 2020 | Present study |  |  |  | Present study | Pante and Watling 2012 |  |



Figure 5. External morphology and polyps of Chrysogorgia carolinensis sp. nov. A the holotype in situ $\mathbf{B}$ close-up branches in situ $\mathbf{C}$ the holotype after collection $\mathbf{D}$ two polyps under a light microscope $\mathbf{E}$ tentacles under a light microscope $\mathbf{F}$ a single polyp under SEM $\mathbf{G}$ upper part of a polyp under SEM $\mathbf{H}$ basal polyp body under SEM I coenenchyme under SEM. Scale bars: $10 \mathrm{~cm}(\mathbf{B}) ; 2 \mathrm{~mm}(\mathbf{D}) ; 1 \mathrm{~mm}(\mathbf{E}-\mathbf{G})$; $300 \mu \mathrm{~m}(\mathbf{H}) ; 100 \mu \mathrm{~m}(\mathbf{I})$.
the same below, Figures 5E, G, 6A). Rare sclerites extend into pinnules, but mostly pinnules free of sclerites. Scales amoeba-shaped, branched in any directions, irregularly and alternately arranged at the basal polyp body, and measuring 161-483 $\times$ 14-170 $\mu \mathrm{m}$ (Figures 5H, 6B). Scales rare and elongate, some of them lobed with irregular edges, transversally arranged in coenenchyme, and measuring 139-221 $\times 29-67 \mu \mathrm{~m}$ (Figures 5I, 6C).

Type locality. A seamount (tentatively named as M8) located on the Caroline Ridge with a depth range of $1506-1832 \mathrm{~m}$.

Etymology. Named after the type locality, the Caroline Ridge, where the species was discovered.

Distribution and habitat. Found only from a seamount located on the Caroline Ridge. Colony attached to a rocky substrate (Figure 5A).

Remarks. Chrysogorgia carolinensis sp. nov. belongs to the "group A, Spiculosae" with an unusual branching sequence of $1 / 3 \mathrm{~L}$ and bottlebrush-shaped colony, which is similar to C. midas Cairns, 2018 and C. abludo Pante \& Watling, 2012. However, the new species differs distinctly from these species by the presence of amoeba-shaped scales, which branch toward to any directions at the basal polyp body (vs. absence in


Figure 6. Sclerites of Chrysogorgia carolinensis sp. nov. A sclerites in the back and at the base of tentacles B sclerites at the basal polyp body $\mathbf{C}$ sclerites in coenenchyme. Scale bar: $300 \mu \mathrm{~m}$ (all images at the same scale).
both species). Chrysogorgia carolinensis sp. nov. is also similar to C. dendritica by 1/3L branching sequence and the conspicuously amoeba-shaped sclerites at the basal polyp body. However, the new species is easily separated by the bottlebrush-shaped colony (vs. tree-shaped), absence of polyps on internodes (vs. presence) and larger polyps up to 8 mm long (vs. no more than 5 mm ) (Table 1).

Genus Metallogorgia Versluys, 1902
Diagnosis (based on Versluys 1902; Nutting 1908; Kükenthal 1919). Main stem monopodial with a few branches occurring on the side. Axis round and solid with a smooth surface and strong metallic luster. Branches irregular, subdivided dichotomously with branchlets forming a sympodium. The coenenchyme usually thin with a few sclerites or not well differentiated this layer.

Type species. Metallogorgia melanotrichos (Wright \& Studer, 1889)

Distribution. Atlantic Ocean, Pacific Ocean and Indian Ocean, in a depth range of 567-2311 m (Watling et al. 2011, and the present study).

## Metallogorgia melanotrichos (Wright \& Studer, 1889)

Figures 7-10; Tables 2, 3

Dasygorgia melanotrichos Wright \& Studer, 1889: 15, pl. IV, fig. 3, pl. V, fig. 5.
Metallogorgia melanotrichos: Versluys, 1902: 87.
Metallogorgia melanotrichos: Nutting, 1908: 593-594, pl. LI, fig. 5.
Metallogorgia melanotrichos: Kükenthal, 1919: 503.
Metallogorgia melanotrichos: Pasternak, 1981: 51.

Type locality. Ascension Island in the South Atlantic Ocean, 778 m depth (Wright and Studer 1889).


Figure 7. External morphology and polyps of Metallogorgia melanotrichos MBM286485 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}$ the colony after collection $\mathbf{C}$ a branch under a light microscope $\mathbf{D}, \mathbf{E}$ a single polyp under a light microscope $\mathbf{F}, \mathbf{G}$ a single polyp under SEM $\mathbf{H}$ head of one polyp under SEM. Scale bars: $20 \mathrm{~cm}(\mathbf{B}), 2 \mathrm{~mm}(\mathbf{C})$, and $1 \mathrm{~mm}(\mathbf{E}-\mathbf{H})$.


Figure 8. Sclerites of Metallogorgia melanotrichos MBM286485 A rods in tentacles B rods and scales in the polyp-body wall C scales in coenenchyme. Scale bar: $200 \mu \mathrm{~m}$ (all images at the same scale).


Figure 9. External morphology and polyps of Metallogorgia melanotrichos MBM286486 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}, \mathbf{C}$ the colony after collection $\mathbf{D}$ a branch under a light microscope E, $\mathbf{F}$ a single polyp under a light microscope $\mathbf{G}$ three polyps under SEM $\mathbf{H}$ head of one polyp under SEM $\mathbf{I}$ coenenchyme under SEM. Scale bars: 10 cm (B, C), 2 mm (D) and $500 \mu \mathrm{~m}$ ( $\mathbf{E}-\mathbf{I} ; \mathbf{H}, \mathbf{I}$ at the same scale).


Figure IO. Sclerites of Metallogorgia melanotrichos MBM286486 A rods in tentacles B rods and scales in the polyp-body wall $\mathbf{C}$ scales in coenenchyme. Scale bar: $300 \mu \mathrm{~m}$ (all images at the same scale).

Voucher specimens. MBM286485, station FX-Dive 222 ( $10^{\circ} 04.73^{\prime} \mathrm{N}$, $140^{\circ} 09.45^{\prime} \mathrm{E}$ ), depth $1839 \mathrm{~m}, 10$ June 2019; MBM286486, station FX-Dive 227 ( $10^{\circ} 37.92^{\prime} \mathrm{N}, 140^{\circ} 05.62^{\prime} \mathrm{E}$ ), depth $1706 \mathrm{~m}, 15$ June 2019. They were collected from two seamounts (tentatively named as M5 and M8, respectively) located on the Caroline Ridge in the West Pacific Ocean.

Diagnosis (extended on the basis of Versluys 1902; Kükenthal 1919; Mosher and Watling 2009). In adults, main stem monopodial with a few large branches occurring on the distal end. Branching angle between these large branches usually obtuse. In each large branch, strong branchlets subdivided dichotomously and forming a sympodium pattern. In juveniles, main stem monopodial and gracile with branches
producing on the lateral of the trunk randomly and subdivided dichotomously in multiple planes. Polyps conical or cylindrical, absent on the stem of adults but present in juveniles. Sclerites elongated and often crossed, arranged densely in polyps and relatively sparsely in coenenchyme. Rods longitudinally arranged in tentacles, covered with sparse fine warts. Rods longitudinally arranged on upper part of polyp body, and scales partially crosswise or transversely arranged on bottom, with nearly smooth surface. Scales transversely arranged in coenenchyme, usually with rounded ends and occasionally irregular edges. Nematozooids absent.

Description. For morphological measurements, see Table 2.
Distribution. Central Indo-Pacific Ocean (Wright and Studer 1889; Versluys 1902), Western and Central Pacific (Nutting 1908; Pasternak 1981; present study), Atlantic Ocean, 183-2265 m depth (Kükenthal 1919; Watling et al. 2011; Pante et al. 2012).

Remarks. Metallogorgia melanotrichos (Wright \& Studer, 1889) is characterized by its completely monopodial stem, with the branches in adults occurring on the distal end, and scales in both body wall and coenenchyme (Table 3). Furthermore, M. melanotrichos has a much more extensive distribution than its congeners, while other species have a relatively limited distribution. Our specimens match well with the original description in the sclerites, but possess a relatively larger polyp (most 2 mm , up to

Table 2. The morphological measuring data of the specimens of Metallogorgia. "-" means nonexistent or meaningless data.

| Characters/ Specimens | MBM286485 | MBM286486 | MBM286484 | MBM286487 | MBM286488 | MBM286489 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species | M. melanotrichos | M. melanotrichos | M. macrospina | M. macrospina | M. macrospina | M. macrospina |
| Colony height (cm) | 76 | 35 | 56 | 51 | 69 | 48 |
| Basal stem width (mm) | 2.7 | 1.5 | 1.0 | 2.5 | 3.0 | 2.5 |
| Branching part height (cm) | 9 | 8 | - | 12 | 11 | 7 |
| Branching part width (cm) | 18 | 18 | 6 | 26 | 20 | 14 |
| Branch maximal length (cm) | 10 | 12 | 4 | 16 | 13 | 12 |
| Branches | 2 | 2 | 22 | 14 | 7 | 6 |
| Interbranch distance (mm) | - | - | 9-25 | 6-13 | 13-18 | 5-18 |
| Internode length (mm) | 4-12 | 3-11 | 3-7 | 3-8 | 3-10 | 3-11 |
| First internode length (mm) | 13-24 | 12-18 | 3-8 | 5-8 | 6-8 | 6-11 |
| Polyp height (mm) | 1.0-2.5 | 1.0-4.0, most 2.0 | 1.0-2.5, most 1.0 | average 1.0 | 1.0-1.5 | 1.0-1.5 |
| Polyps in internode | 1, rarely 2 | 1 , rarely 2 | 1, rarely 2 | 1, rarely 2 | 1, rarely 2 | 1, rarely 2 |
| Polyps in terminal branchlets | 1-3 | 1-4 | 1-4 | 1-4 | 1-5 | 1-2 |
| Inter-polyp distance (mm) | 1-6 | 1-4 | 2-5 | 1-3 | 0-5 | 0-4 |
| Sclerites measured in tentacles ( $\mu \mathrm{m}$ ) | $129-433 \times 17-55$ | $125-467 \times 21-125$ | $96-450 \times 10-85$ | $44-542 \times 6-117$ | $120-492 \times 12-98$ | $115-460 \times 14-78$ |
| Sclerites measured in body wall ( $\mu \mathrm{m}$ ) | $172-571 \times 14-93$ | $128-429 \times 25-125$ | $96-450 \times 10-85$ | $44-542 \times 6-117$ | $137-473 \times 17-93$ | $57-417 \times 11-66$ |
| Sclerites measured in coenenchyme ( $\mu \mathrm{m}$ ) | 96-378 $\times 18-66$ | $148-379 \times 31-113$ | $112-456 \times 10-62$ | $175-394 \times 17-75$ | $78-328 \times 11-51$ | $70-372 \times 14-50$ |
| Figures | 7,8 | 9, 10 | 11, 12 | 13, 14 | 15,16 | 17, 18 |

Table 3. The comparisons of the species (adults) in the genus Metallogorgia Versluys, 1902 and Pseudochrysogorgia bellona Pante \& France, 2010.

| Characters/ Species | M. macrospina | M. melanotrichos | M. splendens* | M. tenuis | P. bellona |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Branching part | sympodial | monopodial | unknown | sympodial $^{*}$ | monopodial |
| Branchlets forming a <br> sympodium | yes | yes | unknown | No $^{*}$ | No |
| Interbranch distance <br> (mm) | $5-18$ | - | unknown | $10-18^{*}$ | $9.9-17.2$ |
| Polyp height (mm) | $1-1.5$ | $1.5-2.0$ | 1.5 | $1.5-2.0$ | spindles |

*, putative species or questionable regarded by Watling et al. 2011.

4 mm vs. 1.75 mm ), and longer scales in coenenchyme (up to $379 \mu \mathrm{~m}$ vs. $225 \mu \mathrm{~m}$ ) (Kükenthal 1919, Table 2). The branches of the specimen MBM286486 on one side is irregular and tend to form a fake spiral with branches subdivided dichotomously in multiple planes, while it is regular and forms a planar layer that nearly perpendicular to the trunk in the specimen MBM286485.

## Metallogorgia macrospina Kükenthal, 1919

Figures 11-18; Tables 2, 3
Metallogorgia macrospina Kükenthal, 1919: 504-505, figs 227-229, Taf.XXX, Fig. 6.
Type locality. $0^{\circ} 58.2^{\prime} \mathrm{S}$, $90^{\circ} 43.2^{\prime} \mathrm{E}$, West Sumatra, 1280 m depth (Kükenthal 1919).
Voucher specimens. MBM286484, station FX-Dive $210\left(10^{\circ} 04.68^{\prime} \mathrm{N}\right.$, $140^{\circ} 12.07^{\prime} \mathrm{E}$ ), depth $911 \mathrm{~m}, 28$ May 2019. MBM286487, station FX-Dive 215 ( $10^{\circ} 04.97^{\prime} \mathrm{N}, 140^{\circ} 10.75^{\prime} \mathrm{E}$ ), depth $986 \mathrm{~m}, 2$ June 2019. MBM286488, station FXDive 215 ( $10^{\circ} 04.82^{\prime} \mathrm{N}, 140^{\circ} 10.90^{\prime} \mathrm{E}$ ), depth $902 \mathrm{~m}, 2$ June 2019. MBM286489, station FX-Dive 223 ( $10^{\circ} 04.63^{\prime} \mathrm{N}, 140^{\circ} 15.12^{\prime} \mathrm{E}$ ), depth $1072 \mathrm{~m}, 11$ June 2019. They were collected from three seamounts (tentatively named as M5, M7 and M8) located on the Caroline Ridge.

Diagnosis (extended on the basis of Kükenthal 1919). In adults, main stem monopodial with branches forming a similar spiral on the top. Strong branches subdivided dichotomously, with branchlets forming a sympodium pattern in each plane. In juveniles, main stem monopodial and thin/gracile with branches occurring on the lateral of the trunk randomly and subdivided dichotomously in mul-


Figure II. External morphology and polyps of juvenile Metallogorgia macrospina MBM286484 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}$ close-up of branches in situ $\mathbf{C}$ the colony after collection $\mathbf{D}$ a branch under a light microscope $\mathbf{E}, \mathbf{F}$ a single polyp under a light microscope $\mathbf{G}$ three polyps under SEM H polyp-body wall under SEM I coenenchyme under SEM. Scale bars: $10 \mathrm{~cm}(\mathbf{C}), 5 \mathrm{~mm}$ (D), $1 \mathrm{~mm}(\mathbf{G})$, and $500 \mu \mathrm{~m}(\mathbf{E}, \mathbf{F}, \mathbf{H}, \mathbf{I} ; \mathbf{H}, \mathbf{I}$ at the same scale).
tiple planes. Polyp cylindrical, some of them with a slightly expanded base, absent on stem of adults but present in juveniles. Sclerites relatively coarse with many small warts on surface, cross-shaped occasionally. Rods relatively regular, longitu-


Figure 12. Sclerites of juvenile Metallogorgia macrospina MBM286484 A some small rods in polyps B larger rods in polyps $\mathbf{C}$ rods and elongated scales in coenenchyme. Scale bars: $50 \mu \mathrm{~m}(\mathbf{A}), 300 \mu \mathrm{~m}$ (B, C at the same scale).
dinally arranged in tentacles and the upper part of the polyp body, and partially crosswise or transversely arranged on the body bottom. Scales and rods elongated, usually coarse with serrated edges, transversely arranged in coenenchyme. Nematozooids not present.

Description. In juvenile (specimen MBM286484), colony slightly bottlebrushshaped, with branches occurring on the lateral side of the stem randomly. Main stem monopodial and gracile. Branches subdivided dichotomously in multiple planes. Polyps cylindrical, occasionally present on the top of the stem. The branch coenenchyme


Figure 13. External morphology and polyps of Metallogorgia macrospina MBM286487 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}$ close-up of branches in situ $\mathbf{C}$ the colony after collection $\mathbf{D}$ a branch under a light microscope $\mathbf{E}, \mathbf{F}$ a single polyp under a light microscope $\mathbf{G}$ three polyps under SEM H head of a polyp under SEM I coenenchyme under SEM. Scale bars: $10 \mathrm{~cm}(\mathbf{C}), 2 \mathrm{~mm}(\mathbf{D})$, and $500 \mu \mathrm{~m}(\mathbf{E}-\mathbf{I} ; \mathbf{H}, \mathbf{I}$ at the same scale).
well differentiated with a layer of sclerites. In the adults (specimen MBM286487286489), colony like a tree shape with branches forming a similar spiral on the top of the stem. Main stem monopodial and strong. Branches subdivided dichotomously with branchlets forming a sympodium pattern in each plane. Polyps cylindrical, some


Figure 14. Sclerites of Metallogorgia macrospina MBM286487 A some small rods in polyps B larger rods in polyps C rods and elongated scales in coenenchyme. Scale bars: $100 \mu \mathrm{~m}(\mathbf{A})$, and $300 \mu \mathrm{~m}$ (B, C at the same scale).
of them with a slightly expanded base, absent from the stem. Branch coenenchyme usually not well differentiated.

Sclerites with same forms and arrangement in juveniles and adults, both relatively coarse with many small warts on surface, cross-shaped occasionally. Rods relatively regular, longitudinally arranged in tentacles and the upper part of the polyp body, and partially crosswise or transversely arranged on the body bottom. Scales and rods elongated, usually coarse with serrated edges, transversely arranged in coenenchyme. For detailed morphological measurements, see Table 2.

Distribution. West Sumatra (Kükenthal 1919); the unnamed seamounts on the Caroline Ridge in the Western Pacific (present study); Southwest Pacific (Pante et al. 2012), 720-1280 m depth.

Remarks. According to Kükenthal (1919), the sclerites in the M. macrospina polyps contain rods and spindles, and those in coenenchyme are slender rods, some of them flat or irregular. The sclerites in our specimens match well with the original description as well as pictures. Therefore, we identify our specimen as $M$. macrospina.


Figure I5. External morphology and polyps of Metallogorgia macrospina MBM286488 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}$ close-up of branches in situ $\mathbf{C}$ the colony after collection $\mathbf{D}$ a branch under a light microscope $\mathbf{E}, \mathbf{F}$ a single polyp under a light microscope $\mathbf{G}$ three polyps under SEM H head of one polyp under SEM I coenenchyme under SEM. Scale bars: $10 \mathrm{~cm}(\mathbf{C}) ; 2 \mathrm{~mm}(\mathbf{D})$, and $500 \mu \mathrm{~m}(\mathbf{E}-\mathbf{I} ; \mathbf{H}, \mathbf{I}$ at the same scale).


Figure 16. Sclerites of Metallogorgia macrospina MBM286488 A rods in tentacles B rods in the polypbody wall C rods and elongated scales in coenenchyme. Scale bar: $200 \mu \mathrm{~m}$ (all images at the same scale).

Metallogorgia macrospina is similar to $M$. melanotrichos by the branchlets forming a sympodium pattern in the large branches. In the original description, Kükenthal (1919) pointed out that M. macrospina differed from M. melanotrichos by its more densely arranged branches, larger polyps, longer sclerites in coenenchyme and different color. However, the polyps of M. macrospina in our specimens are generally smaller than those of $M$. melanotrichos (Table 2), and there are no conspicuous differences in color (almost brown to black) among these colonies. Therefore, based on the morphological features of our specimens, M. macrospina can be separated from M. melanotrichos by its sympodial branching part forming a spiral on the colony top (vs. monopodial), only rods in the polyp-body wall (vs. rods and scales), and rods and scales in coenenchyme (vs. only scales).

The juvenile of Metallogorgia macrospina (specimen MBM286484) has significant differences in the branching pattern from the adult specimens MBM286487, MBM286488 and MBM286489 in morphology. It differs also by having a slightly bottlebrush-shaped colony (vs. similarly tree-shaped colony), gracile and flexible stem (vs. hard and strong), monopodial branching part (vs. sympodial), branchlets


Figure 17. External morphology and polyps of Metallogorgia macrospina MBM286489 A the colony in situ. Laser dots spaced at 33 cm used for scale $\mathbf{B}$ the colony after collection $\mathbf{C}$ a branch under a light microscope $\mathbf{D}$ a terminal branchlet under a light microscope $\mathbf{E}, \mathbf{F}$ a single polyp under a light microscope $\mathbf{G}$ three polyps under SEM $\mathbf{H}$ head of one polyp under SEM I coenenchyme under SEM. Scale bars: 10 $\mathrm{cm}(\mathbf{B}), 2 \mathrm{~mm}(\mathbf{C})$, and $500 \mu \mathrm{~m}$ (D-I; H, I at the same scale).


Figure 18. Sclerites of Metallogorgia macrospina MBM286489 A rods in tentacles B rods in the polypbody wall $\mathbf{C}$ rods and elongated scales in coenenchyme. Scale bar: $300 \mu \mathrm{~m}$ (all at the same scale).
in multiple planes (vs. forming a sympodium in one plane), and well differentiated coenenchyme with more sclerites (vs. not well differentiated and with relatively sparse sclerites). However, the same sclerites in polyps and coenenchyme, and particularly the $\mathrm{mtMut} S$ gene data analyzed below indicate these specimens belong to the same species.

Genus Pseudochrysogorgia Pante \& France, 2010
Diagnosis (based on Pante and France 2010). Main stem monopodial or slightly zigzagging with branches subdivided dichotomously in multiple planes, forming a bot-tlebrush-shaped colony. Most polyps leaning distad and neck narrower than head. Sclerites slightly ornamented, in the form of plates, scales and rods. When polyp not leaning distad, sclerites arranged obliquely on the polyp body. When polyps leaning distad, sclerites mostly arranged longitudinally (parallel to branch) on the polyp body, placed
obliquely in the area of neck, and arranged longitudinally on head and along the back of tentacles. Scales and plates in branch coenenchyme mostly parallel to main branch axis.

Type species. Pseudochrysogorgia bellona Pante \& France, 2010.
Distribution. Southwest Pacific (Coral Sea and northeast of New Zealand), 8001462 m depth (Pante and France 2010, Pante et al. 2012).

## Pseudochrysogorgia bellona Pante \& France, 2010

Figures 19, 20; Table 3
Pseudochrysogorgia bellona Pante \& France, 2010: 595-612.
Type locality. Bellona Plateau, New Caledonia, 800-923 m depth (Pante and France 2010).

Voucher specimen. MBM286490, the Ganquan Plateau in the South China Sea, 586-910 m.

Description. No whole colony was obtained, thus the description was based on a picture (Figure 19A) and a few frozen fragments. Colony ca. 50 cm long and 11 cm wide with dendritic holdfast ca. 12 cm long. Main stem more zigzagging. Branching sequences unknown. Branches subdivided dichotomously, up to sixth order with distance between adjacent branches $10-25 \mathrm{~mm}$. Polyps cylindrical, or with an expanded body base and obviously narrow neck, $1.0-2.5 \mathrm{~mm}$ long and $0.5-2.5 \mathrm{~mm}$ wide at bases, with tentacles up to 1 mm long (Figure 19B-G).

Rods with many small warts on surface, usually with two round ends and relatively less ornamentation, longitudinally arranged along the back of tentacles, and measuring $142-598 \times 11-84 \mu \mathrm{~m}$ (Figures 19H, 20A). Sclerites in the polyp-body wall including rods, usually with one sharp end, and scales, slightly ornamented, some of them thick and elongated, and measuring $150-620 \times 31-127 \mu \mathrm{~m}$. When the polyp not leaning distad, rods and elongated scales arranged obliquely in the polyp-body wall. While polyps leaning distad, scales mostly arranged longitudinally (parallel to the branch) in upper of body, rods and scales placed obliquely in neck region, and arranged longitudinally on head and the base of tentacles (Figures 19I, 20B). Scales slightly ornamented, mostly parallel to main branch axis in coenenchyme, and measuring $115-397 \times 16-$ $150 \mu \mathrm{~m}$ (Figures 19J, 20C).

Distribution. Bellona Plateau, Coral Sea and Otara Seamount, at southern tip of the Kermadec Ridge (Pante and France 2010, Pante et al. 2012); and South China Sea (present study), 586-1462 m depth.

Remarks. Pseudochrysogorgia bellona Pante \& France, 2010 resembles the species of Chrysogorgia Duchassaing \& Michelotti, 1864 by having dichotomously subdivided branches arising from the main stem in a spiraling fashion and forming a bottlebrush-shaped colony. However, it differs in the monopodial or more zigzagging stem (vs. almost sympodial, except Chrysogorgia abludo Pante \& Watling, 2012 and C. dendritica Xu, Zhan $\& \mathrm{Xu}, 2020$ ) and obviously different polyps (Xu et al. 2020).


Figure 19. External morphology and polyps of Pseudochrysogorgia bellona $\mathbf{A}$ the colony after collection B-E a single polyp under a light microscope $\mathbf{F}, \mathbf{G}$ a single polyp under SEM $\mathbf{H}$ head of one polyp under SEM I basal polyp body under SEM J coenenchyme under SEM. Scale bars: $10 \mathrm{~cm}(\mathbf{A}), 1 \mathrm{~mm}(\mathbf{B}-\mathbf{G})$, and $500 \mu \mathrm{~m}(\mathbf{H}-\mathbf{J})$.

Furthermore, P. bellona is similar to species of Metallogorgia Versluys, 1902 in its monopodial colony and has a close genetic distance as well. However, P. bellona can be separated by its polyps usually with obviously narrow neck (vs. none), dendritic


Figure 20. Sclerites of Pseudochrysogorgia bellona $\mathbf{A}$ rods in tentacles $\mathbf{B}$ rods and scales in the polyp-body wall C scales in coenenchyme. Scale bar: $300 \mu \mathrm{~m}$ (all images at the same scale).
holdfast (vs. discoid) and sclerites including plates, scales and rods with more ornamentation (vs. almost rods and scales with little ornamentation) (Pante and France 2010). In our specimen, the rods in tentacles are more regular usually with two round ends and less ornamentation. The elongated thick scales are more abundant in coenenchyme, and plates are scarcer than the type colony. We suggest that these differences are population-dependent.


Figure 2I. Maximum likelihood (ML) tree inferred from the mtMutS sequences of Chrysogorgia, Metallogorgia, Pseudochrysogorgia and related species. Numbers at the nodes represent the ML bootstrap values and the posterior probability values of Bayesian analysis (BI). Newly sequenced species are in bold. Support $\leq 50 \% / 0.50$ is shown as '--'. Alignment flatfile showed the mtMutS deletion for Chrysogorgia carolinensis sp. nov. with "-", using C. averata KC788265 as a reference.

## Molecular sequences, genetic distances, and phylogenetic analyses

The conspecific sequences for each newly sampled species were identical, and only the holotype sequence was deposited in GenBank and analysed here. The accession number and the length are as follows: MT269889, 686 bp for Chrysogorgia carolinensis sp. nov., MT269888, 693 bp for C. dendritica, MT050468, 691 bp for M. macrospina; MT050469, 695 bp for M. melanotrichos, MT050470, 690 bp for P. bellona. The alignment dataset comprised 629 nucleotide positions. Compared to the other Chrysogorgia species, C. carolinensis sp. nov. has six-nucleotide deletion in the mtMust sequence (Figure 21). There was no intraspecific variability, while the interspecific dis-
tances ranged from zero to $2.42 \%$ (Table 4). The genetic distances between the new species $C$. carolinensis sp. nov. and its congeners were in range of $0-2.42 \%$, and those between $C$. dendritica and its congeners were in the range of $0-1.63 \%$. To date, only three species are available for the genetic distance and phylogenetic analysis for Metallogorgia and Pseudochrysogorgia. The mtMutS genetic distances between M. macrospina and $M$. melanotrichos was $0.16 \%$, while there was no intraspecific variation within the two species and P. bellona (Table 4). The genetic distances between Metallogorgia and Pseudochrysogorgia were $1.94 \%$ and $2.10 \%$.

The ML and BI phylogenetic trees are identical in topology, and thus only the former with the both support values is shown (Figure 21). In both trees, Metallogorgia, Pseudochrysogorgia, Iridogorgia, Radicipes and Chrysogorgia formed a "monophyletic Chrysogorgiidae clade" (MCC) sensu Pante et al. (2012) with moderate to high support (ML 86\%; BI 0.95). The Chrysogorgia species were separated into two main clades (Clade I and II) with high support values, which is consistent with Xu et al. (2019, 2020). Clade I includes the sister species C. binata, C. cf. stellata and C. chryseis, and Clade II contains all the rest species. The species C. dendritica and C. abludo formed a sister subclade, followed by C. fragilis. Chrysogorgia carolinensis sp. nov. formed a sister subclade with the cluster C. ramificans $+C$. monticola, the group $C$. dendritica + C. abludo + C. fragilis and the rest species within Clade II. Metallogorgia melanotrichos and M. macrospina formed sister clade, followed by P. bellona with full node support (Figure 21).

## Discussion

Both the morphology and molecular phylogenetic analysis support the assignment of the new species to the genus Chrysogorgia Duchassaing \& Michelotti, 1864. The barcoding analysis of mtMutS is considered as the first step in molecular identification of octocorals (McFadden et al. 2011; Pante et al. 2012). However, the mtMutS genetic distances within Chrysogorgia are relatively low, and there is no barcoding gap (intraspecific zero vs. interspecific $0-2.42 \%$ ) for species identification, as shown by Xu et al. (2020). Thus, the discrimination of the closely related species was mainly relied on other molecular evidence and morphological characters. Alternatively, single mutations on mtMutS can be used to separate Chrysogorgia species (Pante and Watling 2012; Pante et al. 2015; Xu et al. 2020). The mtMutS sequence of C. carolinensis sp. nov. has six deletion mutations compared to those of its congeners (Figure 20), supporting the establishment of the new species. Furthermore, the closely related species C. carolinensis sp. nov., C. artospira and C. pinnata are easily separated by the $1 / 3 \mathrm{~L}$ branching sequence (vs. 2/5L and flabellate, respectively) and amoeba-shaped scales at the basal polyp body (vs. regular scales and rods, respectively) (Cairns 2007; Pante and Watling 2012). No genetic variability was observed between C. dendritica and the closely related species C. abludo. However, the former is morphologically distinctly different from the latter by the presence of irregular sclerites in the polyp-body wall
Table 4. The uncorrected pairwise distances at mtMutS between Chrysogorgia, Metallogorgia and Pseudochrysogorgia species/populations.

|  | Species/populations | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | C. abludo GQ180139, JN227999 | - |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 2 | C. dendritica MN510469, MT269888 | 0 | - |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 3 | C. fragilis MN510470 | 0.16\% | 0.16\% | - |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 4 | C. gracilis MN510472 | 0.97\% | 0.97\% | 1.13\% | - |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 5 | C. carolinensis sp. nov. MT269889 | 0.81\% | 0.81\% | 0.65\% | 0.33\% | - |  |  |  |  |  |  |  |  |  |  |  |  |
| 6 | C. artospira GQ180132-GQ180135, GQ353317 | 0.81\% | 0.81\% | 0.65\% | 0.48\% | 0 | 0 |  |  |  |  |  |  |  |  |  |  |  |
| 7 | C. pinnata JN227988 | 0.81\% | 0.81\% | 0.65\% | 0.48\% | 0 | 0 | - |  |  |  |  |  |  |  |  |  |  |
| 8 | C. tricaulis JN227998, JN227990, JN227991, GQ180123-GQ180131, EU268056 | 0.97\% | 0.97\% | 0.81\% | 0.65\% | 0.16\% | 0.16\% | 0.16\% | 0 |  |  |  |  |  |  |  |  |  |
| 9 | C. averata KC788265, GQ180136 | 1.13\% | 1.13\% | 0.97\% | 0.81\% | 0.33\% | 0.32\% | 0.32\% | 0.48\% | 0 |  |  |  |  |  |  |  |  |
| 10 | C. monticola JN227989 | 1.45\% | 1.45\% | 1.29\% | 1.13\% | 0.65\% | 0.65\% | 0.65\% | 0.81\% | 0.97\% | - |  |  |  |  |  |  |  |
| 11 | C. ramificans MK431863 | 1.45\% | 1.45\% | 1.29\% | 1.13\% | 0.65\% | 0.65\% | 0.65\% | 0.81\% | 0.97\% | 0.32\% |  |  |  |  |  |  |  |
| 12 | C. chryseis JN227992, DQ297421 | 2.42\% | 2.42\% | 2.26\% | 2.10\% | 1.63\% | 1.62\% | 1.62\% | 1.78\% | 1.94\% | 2.26\% | 2.26\% | - |  |  |  |  |  |
| 13 | C. binata MK431862 | 2.42\% | 2.42\% | 2.26\% | 2.10\% | 1.63\% | 1.62\% | 1.62\% | 1.78\% | 1.94\% | 2.26\% | 2.26\% | 0.48\% | - |  |  |  |  |
| 14 | C. cf. stellata JN227920 | 2.26\% | 2.26\% | 2.10\% | 1.94\% | 1.46\% | 1.45\% | 1.45\% | 1.62\% | 1.78\% | 2.10\% | 2.10\% | 0.32\% | 0.16\% | - |  |  |  |
| 15 | Metallogorgia macrospina MT050468, JN228001, JN227906 | 5.33\% | 5.33\% | 5.17\% | 4.85\% | 4.55\% | 4.52\% | 4.52\% | 4.68\% | 4.52\% | 5.17\% | 5.17\% | 5.17\% | 5.17\% | 5.01\% | 0 |  |  |
| 16 | M. melanotrichos MT050469, GQ868333, GQ868339, GQ868340, GQ180146GQ180155, GQ180156-GQ180158, GQ180162, GQ180163, GQ353314, EU268057, DQ297423 | 5.49\% | 5.49\% | 5.33\% | 5.01\% | 4.72\% | 4.68\% | 4.68\% | 4.85\% | 4.68\% | 5.33\% | 5.33\% | 5.33\% | 5.33\% | 5.17\% | 0.16\% | 0 |  |
| 17 | Pseudochrysogorgia bellona MT050470, GQ868331, GQ868332 | 4.04\% | 4.04\% | 3.88\% | 3.55\%. | 3.25\% | 3.23\% | 3.23\% | 3.39\% | 3.55\% | 3.88\% | 3.88\% | 4.52\% | 4.52\% | 4.36\% | 1.94\% | 2.10\% | 0 |

(often branched and amoeba-shaped with more lobed edges vs. relatively regular with less lobed edges; Pante and Watling 2012; Xu et al. 2020).

Colonial branching pattern was regarded as one of the diagnostic characters to distinguish chrysogorgiid octocorals (Pante et al. 2012). However, it should be carefully applied to the identification of Metallogorgia species, since polymorphism of the colony shape is known for $M$. melanotrichos during its life cycle (Mosher and Watling 2009). In the present study, one juvenile and three adults of $M$. macrospina Kükenthal, 1919 were collected and identified. There are also obvious morphological differences between the juvenile and the adults in the colony shape and branch pattern (see before, Figures 11C, 15C). Nevertheless, like the case of $M$. melanotrichos, their conspecific assignment can be determined by the same polyps and sclerites, and the MutS gene sequences (Figures 11-18, 21). On the other hand, the M. melanotrichos and M. macrospina juveniles rather than the adults, are most similar to the Pseudochrysogorgia in having lateral branches subdivided dichotomously and not forming a sympodium, and a slightly bottlebrush-shaped colony, mirroring their sister relationship showed by the phylogenetic trees (Figures 11C, 21). Seemingly, the colony branch morphogenesis may be useful to infer the phylogenetic relationship for chrysogorgiids, and the colonial branching pattern may be an apomorphy for chrysogorgiids.

To include the juvenile morphology, we slightly extend the diagnosis of the genus Metallogorgia on the basis of Versluys (1902) and Kükenthal (1919): Colonies with distinctly monopodial stem giving rise to a few lateral branches. Branches arising distally in adults, while arising on the lateral of stem randomly in juveniles. Branch subdivided dichotomously, with branchlets either forming a sympodium in one plane in adults, or in multiple planes in juvenile. Branching part monopodial forming an approximate planar layer or a slight bottlebrush shape, or sympodial forming a spiral or tree shape. Axis round, hard and strong with a smooth surface and an extremely pronounced metallic luster. Polyps cylindrical without obvious neck, relatively little to the size of branches they sit on and well separated from one another, covered with abundant and dense sclerites. Coenenchyme thin, most with a few sclerites or not differentiated well into this layer. Sclerites in the main form of rods, spindles and scales, with little ornamentation.

Furthermore, based on the present morphological descriptions, we provide a preliminary key to full grow grown specimens of the genus Metallogorgia Versluys, 1902.
1 Only scales present in coenenchyme ..... 2

- Rods present in coenenchyme ..... 3
2 Rods and spindles present in polyp body wall M. tenuis- Rods and scales present in polyp body wall .......................M. melanotrichos3 Plates present in tentacles and only rods in coenenchyme

$\qquad$
M. splendens - Plates absent in tentacles, rods and scales present in coenenchyme.

## Acknowledgements

This work was supported by the Science \& Technology Basic Resources Investigation Program of China (2017FY100804), the Strategic Priority Research Program of the Chinese Academy of Sciences (XDB42000000, XDA19060401), the National Natural Science Foundation of China (41930533, 41876178) and the Senior User Project of RV KEXUE. We thank the assistance of the crew of R/V KeXue and ROV FaXian for sample collection, and Dr. Dong Dong for providing the Pseudochrysogorgia specimen. We also appreciate the editor Bert W. Hoeksema for his editorial work and two reviewers for their constructive comments on an early version of the manuscript.

## References

Alfaro ME, Zoller S, Lutzoni F (2003) Bayes or Bootstrap? A simulation study comparing the performance of Bayesian Markov chain Monte Carlo sampling and bootstrapping in assessing phylogenetic confidence. Molecular Biology and Evolution 20(2): 255-266. https://doi. org $/ 10.1093 / \mathrm{molbev} / \mathrm{msg} 028$
Bayer FM, Grasshoff M, Verseveldt J (1983) Illustrated Trilingual Glossary of Morphological and Anatomical Terms Applied to Octocorallia. E.J. Brill/Dr. W. Backhuys, Leiden, 75 pp.
Cairns SD (2001) Studies on western Atlantic Octocorallia (Gorgonacea: Chrysogorgiidae). Part 1: The genus Chrysogorgia Duchassaing \& Michelotti, 1864. Proceedings of the Biological Society of Washington 114(3): 746-787. https://doi.org/10.2988/0006-324X(200 7) 120 [243:SOWAOG]2.0.CO;2

Cairns SD (2007) Calcaxonian Octocorals (Cnidaria: Anthozoa) from the Eastern Pacific seamounts. Proceedings of the California Academy of Sciences 58(25): 511-541.
Cairns SD (2018) Deep-Water Octocorals (Cnidaria, Anthozoa) from the Galápagos and Cocos Islands. Part 1: Suborder Calcaxonia. ZooKeys 729: 1-46. https://doi.org/10.3897/ zookeys.729.21779.figure10
Cordeiro RTS, Castro CB, Pérez CD (2015) Deep-water octocorals (Cnidaria: Octocorallia) from Brazil: family Chrysogorgiidae Verrill, 1883. Zootaxa 4058(1): 81-100. https://doi. org/10.11646/zootaxa.4058.1.4
Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: more models, new heuristics and parallel computing. Nature Methods 9(772): 1-1. https://doi.org/10.1038/ nmeth. 2109
Deichmann E (1936) The Alcyonaria of the western part of the Atlantic Ocean. Memoirs of the Museum of Comparative Zoology, Harvard 53: 1-317. https://doi.org/10.5962/bhl. title. 49348
Duchassaing P, Michelotti J (1864) Supplément au mémoire sur les coralliaires des Antilles. Memorie della Reale Accademia delle Scienze di Torino 2(23): 97-206. https://doi. org/10.5962/bhl.title. 105196

Ehrenberg CG (1834) Beiträge zur physiologischen Kenntniss der Corallenthiere im allgemeinen, und besonders des rothen Meeres, nebst einem Versuche zur physiologischen Systematik derselben. Abhandlungen der Königlichen Akademie der Wissenschaften, Berlin 1: 225-380.
Grasshoff M (1999) The shallow-water gorgonians of New Caledonia and adjacent islands (Coelenterata, Octocorallia). Senckenbergiana Biologica 78: 1-121.
Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Systematic Biology 59: 307-321. https://doi.org/10.1093/sysbio/syq010
Haeckel E (1866) Generelle morphologie der Organismen (Vol. 2). Verlag von Georg Reimer, Berlin, 652 pp.
Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95-98.
Herrera S, Baco A, Sánchez JA (2010) Molecular systematics of the bubblegum coral genera (Paragorgiidae, Octocorallia) and description of a new deep-sea species. Molecular Phylogenetics and Evolution 55(1): 123-135. https://doi.org/10.1016/j.ympev.2009.12.007
Hillis DM, Bull JJ (1993) An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Systematic Biology 42(2): 182-192. https://doi. org/10.1093/sysbio/42.2.182
Katoh K, Standley DM (2013) MAFFT Multiple Sequence Alignment Software version 7: improvements in performance and usability. Molecular Biology and Evolution 30: 772-780. https://doi.org/10.1093/molbev/mst010
Kükenthal W (1919) Gorgonaria. Wissenschaftliche Ergebnisse der Deutschen Tiefsee- Expedition auf dem Dampfer"Valdivia"1891-1899 13(2): 1-946.
Lamouroux JVF (1812) Extrait d'un mémoire sur la classification des polypiers coralligènes non entièrement piérreux. Nouveau Bulletin des Sciences, Société Philomathique de Paris 3(63): 181-188.
Mosher CV, Watling L (2009) Partners for life: a brittle star and its octocoral host. Marine Ecology Progress Series 397: 81-88. https://doi.org/10.3354/meps08113
Nutting CC (1908) Descriptions of the Alcyonaria collected by the U.S. Bureau of Fisheries steamer Albatross in the vicinity of the Hawaiian Islands in 1902. Proceedings of the United States National Museum 34: 543-601. [pls 41-51.] https://doi.org/10.5479/ si.00963801.34-1624.543
Pante E, France SC (2010) Pseudochrysogorgia bellona n. gen., sp. nov.: a new genus and species of chrysogorgiid octocoral (Coelenterata, Anthozoa) from the Coral Sea. Zoosystema 32(4): 595-612. https://doi.org/10.5252/z2010n4a4
Pante E, Watling L (2012) Chrysogorgia from the New England and Corner Seamounts: Atlan-tic-Pacific connections. Journal of the Marine Biological Association of the United Kingdom 92(5): 911-927. https://doi.org/10.1017/S0025315411001354
Pante E, France SC, Couloux A, Cruaud C, McFadden CS, Samadi S, Watling L (2012) DeepSea Origin and In-Situ Diversification of Chrysogorgiid Octocorals. PLoS ONE 7(6): e38357. https://doi.org/10.1371/journal.pone. 0038357

Pante E, Abdelkrim J, Viricel A, Gey D, France SC, Boisselier MC, Samadi S (2015) Use of rad sequencing for delimiting species. Heredity 114: 450-459. https://doi.org/10.1038/ hdy.2014.105
Pasternak FA (1981) Alcyonacea and Gorgonacea. In: Benthos of the Submarine Mountains of the Marcus-Necker and adjacent Pacific Regions. Academy of Sciences of the USSR, P.P. Shirshov Institute of Oceanology, Moscow: 40-56.
Ronquist FR, Huelsenbeck JP (2003) Mrbayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572-1574. https://doi.org/10.1093/bioinformatics/btg180
Sánchez JA, Lasker HR, Taylor DJ (2003) Phylogenetic analyses among octocorals (Cnidaria): mitochondrial and nuclear DNA sequences (lsu-rRNA, 16S and ssu-rRNA, 18S) support two convergent clades of branching gorgonians. Molecular Biology and Evolution 29: 31-42. https://doi.org/10.1016/S1055-7903(03)00090-3
Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. Molecular Biology and Evolution 30(12): 2725-2729. https://doi.org/10.1093/molbev/mst197
Verrill AE (1883) Report on the Anthozoa, and on some additional species dredged by the "Blake" in 1877-1879, and by the U.S. Fish Commission steamer "Fish Hawk" in 188082. Bulletin of the Museum of Comparative Zoology at Harvard College 11: 1-72.

Versluys J (1902) Die Gorgoniden der Siboga-Expedition I. Die Chrysogorgiidae. Siboga-Expeditie 13: 1-120.
Watling L, France SC, Pante E, Simpson A (2011) Chapter Two-Biology of Deep-Water Octocorals. Advances in Marine Biology 60(3): 41-122. https://doi.org/10.1016/B978-0-12-385529-9.00002-0
Wright EP, Studer T (1889) Report on the Alcyonaria collected by H.M.S. Challenger during the years 1873-1876. Report on the Scientific Results of H.M.S. Challenger during the years 1873-76. Zoology 31(64): 1-314.
Xu Y, Li Y, Zhan ZF, Xu KD (2019) Morphology and phylogenetic analysis of two new deep-sea species of Chrysogorgia (Cnidaria, Octocorallia, Chrysogorgiidae) from Kocebu Guyot (Magellan seamounts) in the Pacific Ocean. Zookeys 881: 91-107. https://doi. org/10.3897/zookeys.881.34759
Xu Y, Zhan ZF, Xu KD (2020) Morphology and molecular phylogeny of three new deep-sea species of Chrysogorgia (Cnidaria, Octocorallia) from seamounts in the tropical Western Pacific Ocean. PeerJ 8: e8832. https://doi.org/10.7717/peerj. 8832

# A new species of Pseudosinella Schäffer, 1897 (Collembola, Entomobryidae) from Altamira Caves (Cantabria, Spain) 

Enrique Baquero', Rafael Jordana', Lucía Labrada ${ }^{2,3}$, Carlos G. Luque ${ }^{2}$<br>I University of Navarra, Faculty of Sciences, Department of Environmental Biology, University Campus, 31080 Pamplona, Spain 2 Impress Group Consulting S.L., P.O. Box 879, 39080 Santander, Cantabria, Spain 3 Coastal Interpretive Centre, Santander City Council, Barrio Monte-La Maruca, 56D, 39012 Santander, Cantabria, Spain

Corresponding author: Enrique Baquero (ebaquero@unav.es)

Academic editor:L. Deharveng | Received 24 March 2020 | Accepted 11 September 2020 | Published 9 November 2020
http://zoobank.org/4C91B9F4-A43E-4FF0-97E0-E97B6DC6DA9D
Citation: Baquero E, Jordana R, Labrada L, Luque CG (2020) A new species of Pseudosinella Schäffer, 1897 (Collembola, Entomobryidae) from Altamira Caves (Cantabria, Spain). ZooKeys 989: 39-54. https://doi.org/10.3897/ zookeys.989.52361


#### Abstract

This paper describes Pseudosinella altamirensis sp. nov. from the Altamira Caves, municipal district of Santillana del Mar (Cantabria, Spain), and five other caves near the coast of Cantabria (northern Spain). Its taxonomic position is discussed and differences and similarities among morphologically and geographically close species are highlighted. The new species can be identified by its five eyes, the basal and small inner paired teeth on the claw, and the length of the uncrenulated part of the distal dens.


## Keywords

cave dwelling fauna, chaetotaxy, northern Spain, Pseudosinella altamirensis sp. nov., taxonomy

## Introduction

During sampling work to increase the knowledge of the Collembolan cave fauna of the Cantabrian Mountains, we have captured a considerable number of undescribed species of Collembola. Some of these, mainly Pseudosinella Schäffer, 1897, are new species, and it is necessary to describe them. In this paper, a new species of the genus Pseudosinella

[^1]is described, found in some limestone caves within the municipal districts of Miengo, Santillana del Mar, Reocín, and Cabuérniga (Fig. 1). A complete review of specimens collected in Altamira and other nearby caves (see Luque and Labrada 2016 for details) has shown that this new species was erroneously attributed to Pseudosinella superoculata Gisin \& Gama, 1969 by Luque and Labrada. Dorsal macrochaetotaxy and other morphological characters have been used here for species identification. Christiansen et al. (1983) produced a catalogue of world Pseudosinella species and established a code for dorsal macrochaetotaxy characters. Christiansen et al. (1990) designed a computer-assisted identification key (Delta key) of the species of Pseudosinella with more characters. This electronic key is now available on the Web and regularly updated (Jordana et al. 2020). The new species was easily detected using this electronic key by the combination of chaetotaxy formula and other characters used in Pseudosinella species description.

## Materials and methods

## Abbreviations used in the description and figures:

```
Abd abdominal tergite
accp accessorial p-sensilla
Ant antennal segment
a.s.l. above sea level
Mc macrochaeta(e)
mes mesochaeta(e)
mic microchaeta(e)
MZNA Museum of Zoology, University of Navarra
pse pseudopore
Th thoracic tergite
```


## Terminology

Dorsal head and body chaetotaxy follows Gisin (1965, 1967a, b), Szeptycki (1979), and Soto-Adames (2010). Equivalence between the notation proposed by Gisin and the AMS system sensu Soto-Adames 2010 is given in Table 1.

The characters defined by Christiansen et al. (1990) for Pseudosinella and those used by Christiansen (2013) and Jordana et al. (2020) in the Delta key have been used for identification. Some characters proposed by Mateos (2008) and Winkler and Mateos (2018) have also been considered.

## Study area

The Saja River catchment is in the central sector of the Cantabrian mountain range and flows to the Cantabrian Sea after following a course of 67 km in which it descends approximately 1700 m in altitude. It follows a practically rectilinear south-north course


Figure I. Location of the studied area. A Cantabrian Mountains (northern Spain) B UTM grid map with 10 km squares of the central region of the Cantabrian Mountain to show the outcrops of the rock systems (in grey) which contain limestone and geographical situation of the study area with the distribution of the cave-dwelling Pseudosinella altamirensis sp. nov.: (1) Cudón cave; (2) La Venta del Cuco cave; (3) Altamira and Stalactites caves; (4) Peña Caranceja cave; (5) Cobezo cave. Source: Spanish Geological Institute and Cartographic Service of the Cantabrian Government, scale 1:50 000.
in its middle and upper sections, and in its lower one it flows in a west-east direction, which changes to a south-north direction in its final reach at the mouth of the San Martín de la Arena estuary between the municipalities of Miengo and Suances (Fig. 1). This course coincides with the main fracturing and folding lines which characterise the area. In the two municipalities which compose the middle course (Ruente and Cabuérniga), a dismantling of Cretaceous sandstones and clays has taken place, promoting the rising to the surface of Jurassic carbonate materials (Fig. 1). Among the few cavities known over these Jurassic limestones are the caves of Poyo and Cobezo (also the caves located in the Saja-Besaya Natural Park) and the flooded Fuentona de Ruente
cave. The first Creaceous limestone outcrops are in the largest municipalities (Cabezón de la Sal, Reocín, Santillana, and Miengo), which are located along the middle-lower course of the Saja River. In the geologic context, this coastal area is distinguished by the abundance of lower to upper Cretaceous carbonates, which favours karst development. Among the abundant explored cavities in these limestone rocks are two (the caves of Peña Caranceja and La Venta del Cuco) that harbor the new species described here. Cudón cave, which has prehistoric paintings, is near the town of the same name at the opening of the Saja River that forms the San Martín de la Arena estuary; Cudón cave is approximately 9 km from Altamira Cave (Fig. 1).

The Altamira Cave is in the central region of Cantabria (northern Spain), in one of the limestone hills that surround the small valley in Santillana village. On the whole, the geological and structural characterisation of Altamira Cave ( 270 m in length) indicates the evolution of a karstic complex from the first prehistoric human occupation until present. It is one of the many caves in the upper vadose area of the tabular polygenic karstic system that developed on Cretaceous calcarenite limestones (Sánchez et al. 2007). This area is known as Santa Olaja, although it is also known by the name of Planes (Santillana del Mar), where the medieval chronicles place the site of the "Sant[a] Illana" monastery (Luque and Labrada 2016). The small plain that extends over the Altamira Cave has an elevated position ( 159 m a.s.l.) below the Mount Santa Olaja hill range ( 168 m a.s.l.), and is protected between the mounts of Vispieres ( 226 m a.s.l.) and Cildad ( 287 m a.s.l.). This area separates the Saja River catchment from the Santillana physiographic basin ( 66 m a.s.l.). Altamira Cave, lying 4 km from the sea, is little more than 2 km from the nearby Saja River. It is situated on a topographical high point ( 152 m a.s.l.) and is only $3-22 \mathrm{~m}$ (averaging 8 m ) below the surface. Having a length of 270 m , the cave features a main passage 2-12 m high, and 6-20 m wide (Fig. 2). It was discovered in 1868; exploration began in 1875 , but it was not until 1879 when the first paintings, one of the first to be cataloged as Palaeolithic, were discovered by Marcelino Sanz de Sautuola (Lasheras 2009). Regarding the cave biology, Ignacio Bolívar was the first entomologist to explore the Altamira Cave in July 1883 (Luque and Labrada 2014, 2016). Since then, entomological visits to the cave have been rather sporadic. For example, in the early $20^{\text {th }}$ century, Charles Alluaud (28 June 1913) and Cándido Bolívar (28 August 1915) explored this cave (Luque and Labrada 2014, 2016). On 7 May 1924, the cave and its surrounding area was protected by a resolution of the Government of Spain. Since 1985, the cave and its paintings have been included in the UNESCO list of World Heritage Sites (SC-85/Conf. 008/9 1985).

Furthermore, a small cave with stalactites was discovered in the summer of 1928 when stone was being quarried out for the construction of a road to Altamira Cave. It is purely of geological interest, with beautiful stalactites and stalagmites, as no cave art has been found within. In October 1935, Hans Jürgen Stammer was the first entomologist that explored the Stalactites Cave (Luque and Labrada 2016). This cave also hosts the new species described here.


Figure 2. Geographical location of Altamira Caves (Cantabria, Spain). A location and ground plan of the Altamira Caves and its museum B location of the Pseudosinella altamirensis sp. nov. sampling sites along Altamira Cave (August 2000): (1) areas where soil contained mouldy rat or bat faeces; (2) areas where soil contained a rat cadaver (Rattus rattus); (3) areas where soil contained a bat cadaver (Rhinolophus hipossideros); (4) areas where walls had insect cadavers with visible fungal; (5) areas where water drips or flows towards the cave (hygropetric habitats); (6) areas where water had flooded the surface with small rim stone dams or gours; (7) areas with high clay soil content in the water; (8) areas of continuous monitoring of radon. Plan of the caves with its location courtesy of the National Museum and Research Centre of Altamira.

## Specimen collection and measurement

The specimens were obtained by direct capture. Additionally, baited pitfall traps were used in the parts of the cave that were considered favourable for the presence of fauna (Fig. 2) (see Luque and Labrada 2016 for details). The specimens were captured using a manual aspirator and then preserved in vials containing 70\% ethyl alcohol. Each vial was labelled with the following data: collection site, coordinates, date of capture, name of the organisation, and person involved in the capture. The specimens were mounted in Hoyer medium, and optical observations were made under an Olympus BX51-TF microscope with a multiviewing system and phase contrast, and an Olympus BX50-F4 microscope with differential interference contrast (DIC). For the measurements, a U-DA drawing attachment UIS (Universal Infinity System) and a scale calibrated with a slide by Graticules Ltd ( 1 mm divided in 100 parts) was used. For electron microscopy, three specimens were fixed with $4 \%(\mathrm{v} / \mathrm{v})$ glutaraldehyde in 0.1 M cacodylate buffer ( pH 7.3 ) for 48 h , then stored for 24 h in a 0.25 M sucrose buffer containing 0.1 M cacodylate, and dehydrated using an ethanol series followed by critical-point drying in $\mathrm{CO}_{2}$, mounted on aluminium SEM stubs, and coated in Argon atmosphere with 16 nm of gold in an Emitech K550 sputter-coater. SEM observations and photographs were made with a Zeiss DSM940A.

## Taxonomy

Class Collembola Lubbock, 1873
Order Entomobryomorpha Börner, 1913
Family Entomobryidae Schäffer, 1896
Subfamilia Lepidocyrtinae Wahlgren, 1906
Genus Pseudosinella Schäffer, 1897

## Pseudosinella altamirensis sp. nov.

http://zoobank.org/2f1163c5-c7d2-43e7-93a7-b7d30c2f11d0
Figs 3-7
Type material. Holotype: Spain - $q$; Cantabria, municipal district of Santillana del Mar, Altamira Cave, Sala de Polícromos (Polychromes Room), National Museum and Research Centre of Altamira; $43^{\circ} 22.61^{\prime} \mathrm{N}, 4^{\circ} 7.1^{\prime} \mathrm{W} ; 148 \mathrm{~m}$ a.s.l.; 29 Aug. 2000; C. Glez.-Luque leg.; slide labelled "MZNA-Altamira 6d-01". Paratypes: • 49 specimens on ethyl alcohol and 3 specimens mounted on SEM stubs; Cueva de las Estalactitas (Stalactites cave); $43^{\circ} 22.64^{\prime} \mathrm{N}, 4^{\circ} 7.21^{\prime} \mathrm{W} ; 148 \mathrm{~m}$ a.s.l.; 29 Aug. 2000; C. Glez.Luque leg.; slides MZNA-Altamira 6d-02 to 05 - 3 specimens on slides and 25 in ethyl alcohol; Polychromes Room; $43^{\circ} 22.61^{\prime} \mathrm{N}, 4^{\circ} 7.18^{\prime} \mathrm{W} ; 148 \mathrm{~m}$ a.s.l.; 24 Mar. 2008; Cesáreo Saiz leg.; slides MZNA-Altamira01-01 and MZNA-Altamira01-02; deposited at the Museum of Zoology, University of Navarra, Pamplona, Spain (MZNA).


Figure 3. Pseudosinella altamirensis sp. nov. Habitus. Scale bar: 0.5 mm .

Other material. Spain - Cantabria - 2 specimens on slides and 14 on ethyl alcohol; La Venta del Cuco cave, Ubiarco, Santillana del Mar; $43^{\circ} 24.28^{\prime} \mathrm{N}, 4^{\circ} 6.35^{\prime} \mathrm{W}$ W; 145 m a.s.l.; 13 Sep. 1995; C. Glez.-Luque leg.; slides MZNA-Luque Coll. 13d-01 and 02 - 1 specimen on slide and 1 in ethyl alcohol; Cudón cave, Cudón, Miengo; $43^{\circ} 24.94^{\prime} \mathrm{N}$, $4^{\circ} 0.74^{\prime} \mathrm{W} ; 22 \mathrm{~m}$ a.s.l.; 14 Sep. 1995; C. Glez.-Luque leg.; slide MZNA-Luque Coll. 36d - 1 specimen on slide and 3 in ethyl alcohol; Peña Caranceja or La Peñona cave, Barcenaciones, Reocín; $43^{\circ} 20.33^{\prime} \mathrm{N}, 4^{\circ} 9.45^{\prime} \mathrm{W} ; 125 \mathrm{~m}$ a.s.l.; 7 Oct. 2000; C. Glez.-Luque leg.; slide MZNA-Luque Coll. $3 \mathrm{~d} \cdot 2$ specimens on slide and three in ethyl alcohol; Cobezo, Cobeján or Perro cave, Viaña, Cabuérniga; $43^{\circ} 11.61^{\prime} \mathrm{N}, 4^{\circ} 16.52^{\prime} \mathrm{W} ; 360 \mathrm{~m}$ a.s.l.; 15 Jun. 1996; C. Glez.-Luque leg.; slide MZNA-Luque Coll. 7d.

Diagnosis. $5+5$ ocelli. Antennae moderately long. Ant III sense organ with two paddle- or leaf-shaped and two additional sensilla. Area not ringed of dentes nearly five times the length of mucro. Claw elongate, with two paired basal teeth; tenent hair acuminate. Reduced formula: $\mathrm{R}_{0} \mathrm{R}_{1} \mathrm{R}_{2} 000 / 00 / 0101+2 / \mathrm{s},-\mathrm{aBq}_{1} \mathrm{q}_{2}, \mathrm{M}_{1} \mathrm{~m}_{2} \mathrm{R}^{*} \mathrm{el}_{1} \mathrm{l}_{2}(* 1 / 3$ to $2 / 3$ of M ; sometimes $\mathrm{M}_{1}$ smooth and $\mathrm{L}_{2}$ ciliated, and usually asymmetrically).

Description. Habitus (Figs 3, 4A). Body length up to 3.1 mm (holotype: 2.3 mm ). Colour: blue pigment laterally on body from Th II to Abd IV, but extended to dorsal part in Th II, coxae I-III, first third of femur III, laterally on head and vertex and Ant I-III. Abd IV paler. Eyes and a spot on central vertex strongly pigmented.

Head. With five eyes (ABDEF or ABCDF; C and F smaller, almost imperceptible in some specimens). Intraocular chaetae $\mathrm{p}, \mathrm{r}$, and $s$ present. Only $\mathrm{A}_{0}, \mathrm{~A}_{2}, \mathrm{~A}_{3}, \mathrm{An}_{1}, \mathrm{An}_{2}$, $\mathrm{An}_{3 \mathrm{a}}$, and $\mathrm{An}_{3}$ as Mc (Fig. 7C). Ratio antenna/cephalic diagonal 1.64-2.16. Antennal


Figure 4. Pseudosinella altamirensis sp. nov. A habitus B distal part of tibiotarsus III and claw complex $\mathbf{C}$ trochanteral organ. Scale bars: $0.5 \mathrm{~mm}(\mathbf{A}) ; 0.02 \mathrm{~mm}(\mathbf{B}, \mathbf{C})$.
segments I/II/III/IV ratios 1/2-2.7/2.3-2.7/3.5-4.1. AntIV proximal area in Fig. 5A; apical vesicle absent (Fig. 7A); in this segment there are up to four special leaf-shaped sensilla on the distal third, two of them aligned, and at least five other different types of sensilla (some described by Beruete et al. 2002): smooth and cylindrical, some with short fringes, clearly different from the normal chaetae (Fig. 5B); smooth, shorter and narrowed in its distal third (Fig. 5C); and leaf-shaped, similar to the sensorial chaetae 's' of sensory organ of Ant III (Fig. 5D). Ant III sense organ with a peculiar configuration: two paddle-shaped sensilla (individually encased in a pit and more or less one above the other), another two similar ones but in a dorso-internal position, and two small, rounded, spiny guard sensilla on both sides of the first one (Fig. 7B). Apical region of the Ant II-III with pseudopore in internal-ventral position, far from chaetae line. Antennae without scales. Prelabral chaetae ciliated; labral row a, m, and p all smooth (distal row (a) on papillae). Formula of the labial base $\mathrm{M}_{1} \mathrm{~m}_{2} \mathrm{Rel}_{1} \mathrm{l}_{2} ; \mathrm{M}_{1}$ sometimes smooth, asymmetrical in some specimens; R ciliated, $1 / 3$ to $1 / 2$ length of the neighbouring smooth Mc $m_{2} ; l_{1}$ occasionally ciliated; the remaining chaetae smooth, but with minute fringes or ciliation seen only in SEM (Fig. 5E). Bifurcate maxillary palp with three smooth sublobal chaetae. Labial papilla (l.p.) E with finger-shaped process just reaching the base of apical appendage. Maxilla in Fig. 5F.

Body. Th II without Mc; pseudopore of this tergite in Fig. 6A. Th III without Mc. Abd II: chaetae a, $\mathrm{q}_{1}$ and $\mathrm{q}_{2}$ as ciliated mic, chaeta B as broad ciliated Mc (Fig. 7D). Abd III chaetotaxy shown in Fig. 7E. Accessory chaeta 's' in the anterior trichobothrial


Figure 5. Pseudosinella altamirensis sp. nov. A proximal half of Ant IV B detail of Ant IV showing normal chaetae and different types of sensilla: leaf shaped, smooth, with small cilia $\mathbf{C}$ detail at high magnification of the small sensilla narrowed at tip found on Ant II-IV D two leaf-shaped sensilla on Ant IV E labium and labral palps $\mathbf{F}$ maxillae. Scale bars: $0.02 \mathrm{~mm}(\mathbf{A}, \mathbf{E}) ; 0.007 \mathrm{~mm}(\mathbf{B}) ; 0.002 \mathrm{~mm}(\mathbf{C}) ; 0.006 \mathrm{~mm}(\mathbf{D}) ; 0.009 \mathrm{~mm}(\mathbf{F})$.


Figure 6. Pseudosinella altamirensis sp. nov. A Th II pseudopore B claw and empodium appendage of leg III $\mathbf{C}$ distal par of dens and mucro $\mathbf{D}$ tip of dens and mucro, not visible by the covering scales of terminal dens $\mathbf{E}$ mucro partially covered by scales and chaetae $\mathbf{F}$ mucro and mucronal spine. Scale bars: $0.004 \mathrm{~mm}(\mathbf{A}, \mathbf{F}) ; 0.02 \mathrm{~mm}(\mathbf{B}, \mathbf{D}) ; 0.04 \mathrm{~mm}(\mathbf{C}) ; 0.007 \mathrm{~mm}(\mathbf{E})$.


Figure 7. Pseudosinella altamirensis sp. nov. A antennae, tip of Ant IV B antennae, Ant III sensory organ C head, dorsal chaetotaxy; Abdomen dorsal chaetotaxy: D abd II E abd III F abd IV G abd V. White dots, Mc (size of the socket proportional to reality); black dots, mic. Scale bars: $0.01 \mathrm{~mm}(\mathbf{A}) ; 0.02 \mathrm{~mm}$ (B); $0.05 \mathrm{~mm}(\mathbf{C}-\mathbf{E}) ; 0.1 \mathrm{~mm}(\mathbf{F}, \mathbf{G})$.
complex of Abd IV present. Medial chaeta $\mathrm{B}_{5}$ below the level of the trichobothrium $\mathrm{T}_{4}$. Pseudopore between $B_{5}$ and $B_{6}$. Legs scales only on coxae. Trochanteral organ with ca 30 chaetae (Fig. 4C). Remaining chaetae clearly visible on all legs, acuminate and largest on leg III. Differentiated supraempodial inner chaeta on hind tibiotarsus well differentiated
and acuminate. Dorsal tibiotarsal tenent hairs acuminate, 0.3 times the length of inner margin of claw. Claw with only three internal teeth: dental plate occupying $35 \%$ of the basal internal edge, with the basal paired teeth of different sizes (posterior one larger and slightly more distal than anterior); unpaired tooth well developed, approximately $40 \%$ from base; lateral tooth, anterior, less frequently posterior, present in some specimens on leg I and in basal positions; dorsal tooth basal, observed only in one specimen at SEM. Empodium appendage acuminate, basally swollen, externally smooth, with a minute tooth in some specimens (Figs 4B, 6B). Retinaculum with $4+4$ teeth and one ciliated chaeta. Ventral tube without scales; lateral flap with a maximum of eight smooth and six ciliated chaetae. Manubrium and dens with scales only ventrally (anteriorly); two internal and $7-12$ external chaetae related to two distal pseudopores of manubrial plate; area not ringed of dentes nearly five times the length of mucro; mucro with distal tooth longer than the anteapical; basal spine reaching the tip of distal tooth (Fig. 6C-F). Chaetotaxy from head to Abd V illustrated in Fig. 7C-G.

Biology. This species is always found over organic matter accumulation. In Cudón cave, it has been found over the residue of rotten and wet wood; in the other caves it was found over old, mouldy rat and bat faeces and generally in insect cadavers with visible fungi. Although this species reaches to the dark zone of the caves near very wet areas, it has been found in deep zones on walls, roofs, and soils where the environmental humidity is near the saturation point (Fig. 2). The species appears to be restricted to the karst systems associated with the Saja River and within the municipal districts of Miengo, Santillana del Mar, Reocín, and Cabuérniga (Luque and Labrada 2016) (Fig. 1).

Etymology. The name is toponymical and refers to the type locality, the Altamira Cave, one of the most important Palaeolithic art sites in Europe.

## Discussion

According to the dorsal macrochaetotaxy (R000/00/0101+2), the presence of chaeta ' $s$ ' in the anterior trichobothrial complex of the Abd IV, the Abd II chaetotaxy $\left(-\mathrm{aBq}_{1} \mathrm{q}_{2}\right)$, and the formula of the labial base $\left(\mathrm{M}_{1} \mathrm{~m}_{2} \operatorname{Rel}_{1} \mathrm{l}_{2} ; \mathrm{M}_{1}\right.$ or $\left.\mathrm{m}_{1} ; \mathrm{L}_{1} \mathrm{ol} \mathrm{l}_{1}\right)$, this species is similar to Pseudosinella goughi Gisin \& Gama, 1972, P. suboculata Bonet, 1931, and P. superoculata (with more constant labial base formula: $\mathrm{M}_{1} \mathrm{~m}_{2} \mathrm{Rel}_{1} \mathrm{l}_{2}$ except in the case of $P$. gough i with $\mathrm{M}_{1}$ or $\mathrm{m}_{1}$ ). Nevertheless, these three species have six eyes (A, B, C, D, E, and F). Pseudosinella suboculata can have five eyes, but its dorsal tibiotarsal tenent hair is clavate and its claw is clearly different, with longer paired teeth, impaired tooth distal (more than $60 \%$ from base of inner claw), and the empodium appendage not basally swollen. Pseudosinella superoculata has the paired teeth of the inner claw at the same level, claw approximately $30 \%$ longer than in the new species, manubrial plate with three chaetae (two in one specimen) internal to pseudopore, and sensorial chaetae 's' of sensory organ of Ant III rod-like (after Gisin and Gama 1969 and observed also in some specimens from the Cantabria and Navarra caves). Pseudosinella goughi exceptionally has up to seven eyes and only two internal teeth on claw. In addition, the
Table I. Equivalence table of Gisin $(1965,1967 \mathrm{a}, 1967 \mathrm{~b})$ notation formulae for chaetotaxy to the modern notation of Soto-Adames 2010 (AMS) as presented in the figures.

|  | Dorsal Mc formula |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  | Additional sensilla |  |  |  | Abd II formula |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Head |  |  |  |  |  | Th |  |  |  |  |  |  |  | bd |  |  |  |  |  |  |  |  |  |
| Character from Gisin | $\mathrm{R}_{0}$ | $\mathrm{R}_{1}$ | $\mathrm{R}_{2}$ | S | T | P | / | Th II | Th III | 1 | Abd | A2 | A3 | Abd IV | + | Abd IV | I | s | Character from Gisin | p | a | b | $\mathrm{q}_{1}$ | $\mathrm{q}_{2}$ |
| Possible status | 0-1 | 0-1 | 0-1 | 0-1 | 0-1 | 0-1 |  | 0-2 | 0-2 |  | 0 | 0-2 | 0 | 0-1 |  | 0-3 |  | 0-1 | Possible status | --p | a-A | b-B | $\mathrm{q}_{1}-\mathrm{Q}_{1}$ | $\mathrm{q}_{2}-\mathrm{Q}_{2}$ |
| Actual status | mic- <br> Mc | mic- <br> Mc | mic- <br> Mc | mic- <br> Mc | mic- <br> Mc | mic- <br> Mc |  |  | mic-M | cr | numb |  |  | $\begin{aligned} & \text { mic- } \\ & \mathrm{Mc} \end{aligned}$ |  | micMc <br> number |  | presenceabsence | Meaning | 0 -mic | $\begin{aligned} & \text { mic- } \\ & \mathrm{Mc} \end{aligned}$ | mic- <br> Mc | $\begin{aligned} & \text { mic- } \\ & \mathrm{Mc} \end{aligned}$ | $\begin{aligned} & \text { mic- } \\ & \mathrm{Mc} \end{aligned}$ |
| Chaeta AMS system notation | $\mathrm{A}_{0}$ | $\mathrm{A}_{1}$ | $\mathrm{A}_{2}$ | M | $\begin{gathered} \mathrm{S}_{2} \text { or } \\ \mathrm{S}_{4} \end{gathered}$ | $\mathrm{Pa}_{5}$ |  | m or p | peries |  |  | p se |  | $\mathrm{C}_{1}$ |  | row B |  |  | Chaeta AMS system notation | $\mathrm{a}_{2 p}$ | $\mathrm{a}_{2}$ | $\mathrm{m}_{3}$ | $\mathrm{m}_{3 \mathrm{c}}$ | $\mathrm{P}_{4}$ |


| Species | C1 | C2 | C3 | C4 | C5 | C6 | C7 | C8 | C9 | C10 | C11 | C12 | C13 | C14 | C15 | C16 | C17 | C18 | C19 | C20 | C21 | C22 | D |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| P. alpina | 3* | $1^{*}$ | 2 | 2 | 4 | 2 | 2 | 2 | $1^{*}$ | 1 | 3 | $1^{*}$ | 2 | 3* | 3 | 5-7* | 1 | 2.4* | U | 32-36* | 1.7-2.0 | $3^{*}$ | 9 |
| P. astronomica | U | $1 *$ | 4 | 2 | 4 | 2 | 2 | 2 | $1^{*}$ | 1 | 3 | $1^{*}$ | 1 | $3^{*}$ | U | U | 1 | 1.85* | U | $60^{*}$ | $1.4 *$ | U | 7 |
| P. christiani | $1^{*}$ | $0^{*}$ | 2-4 | 2 | 4 | 2 | 2 | 2 | $1^{*}$ | 1 | 3 | $1 *$ | 1 | 3* | U | U | 1 | 1.5* | U | 35* | 1.5* | 1 | 8 |
| P. goughi | $1^{*}$ | 6-7* | 2-4 | 2 | 4 | 2 | 2 | 2 | 2 | 1 | $2^{*}$ | 2 | 1 | 3* | U | U | 1 | 1.9* | 25-30 | - | 1.7-1.8* | U | 6 |
| P. mucronata | $1^{*}$ | 5-6 | 2 | 2 | 4 | 2 | 2 | 2 | $1^{*}$ | 1 | 3 | $1^{*}$ | 1 | $2^{*}$ | U | U | 1 | $2^{*}$ | U | $60^{*}$ | 1.8-2* | U | 7 |
| P. suboculata | 2 | 5-6 | 4 | 2 | 4 | 2 | 2 | 2 | 2 | $2^{*}$ | 3 | 2 | 1-2 | 2* | U | U | 3* | 2.5* | 35 | $70^{*}$ | 1.7-1.8* | 2 | 6 |
| P. superoculata | $1^{*}$ | $6^{*}$ | 4 | 2 | 4 | 2 | 2 | 2 | 2 | 1 | 3 | $1^{*}$ | 1-2 | $3^{*}$ | 3* | 9-10 | 1 | 2.3* | 30 | 45-50 | 2.1-2.3 | 2 | 6 |
| P. thibaudi | U | $2^{*}$ | 4 | 2 | 4 | 2 | 2 | $4^{*}$ | $1^{*}$ | 1 | 3 | 1-2 | 1 | U | 2 | 2-3* | $2^{*}$ | 1.3* | U | $70^{*}$ | 1.3-1.4* | U | 8 |
| P. vandeli relicta | $1^{*}$ | $0^{*}$ | 4 | 2 | 4 | 2 | 2 | 2 | $1^{*}$ | 1 | 3 | $1^{*}$ | 2 | 3* | U | U | 1 | U | U | 30-37 | 1.5* | 2 | 6 |
| P. altamirensis sp. nov. | 2 | 5 | 2-4 | 2 | 3-4 | 2 | 2-4 | 2 | 2 | 1 | 3 | 2 | 1-2 | 1 | 2 | 7-12 | 1 | 3.1 | 35 | 40 | 2.16 | 2 | - |

Legend. C1: apical organ of third antennal segment: (1) peg or rod-like, (2) expanded. C2: number of eyes per side. C3: M1 < ventral labial chaeta > shape: (2) smooth Mc, (3) ciliated mic or mes, (4) ciliated Mc. C4: M2 < ventral labial chaeta > shape: (2) smooth Mc. C5: R < ventral labial chaeta > shape: (3) ciliated mic or mes, (4) ciliated Mc. C6: E < ventral labial chaeta > shape: (2) smooth Mc. C7: L1 < ventral labial chaeta > shape: (2) smooth Mc, (4) ciliated Mc. C8: L2 < ventral labial chaeta > shape: (2) smooth Mc. C9: Abd IV supplementary seta s: (1) absent, (2) present. C10: tenent hair shape: (1) acuminate, (2) clavate. C11: claw total teeth number. C12: claw wing tooth: (1) absent, (2) present. C13: empodium appendage wing tooth: (1) smooth, (2) minute. C14: empodium appendage shape: (1) acuminate, (2) truncate, (3) basally swollen. C15: inner chaeta on manubrial plate, number. C16: outer chaeta on manubrial plate, number. C17: habitat: (1) cave, (2) surface, (3) both cave and surface. C18: maximum length, in mm. C19: distance distal paired claw tooth from base/total claw, \%. C20: distance distal unpaired claw tooth from base/total claw, \%. C21: Antennae/head ratio. C22: differentiated inner chaeta on hind tibiotarsus: (1) unclear or absent, (2) clear, acuminate, (3) clear, truncate or clavate. D: accumulated differences. "*", difference with $P$. altamirensis sp. nov. "U", unknown. "-", not applicable.
four special leaf-shaped sensilla present on Ant IV in the new species separate it from its most similar species. With the same dorsal macrochaetotaxy, but without chaeta 's' on Abd II, there are six species: P. alpina Gisin, 1950 (one eye); P. astronomica Gisin \& Gama, 1970 (one eye); P. christiani Stomp, 1986 (without eyes); P. mucronata Gouze \& Deharveng, 1987 (five or six); P. thibaudi Stomp, 1977 (two eyes); and P. vandeli relicta Gisin, 1964 (no eyes). The differences between these nine species and the new species are shown in Table 1.

## Acknowledgements

The specimens referred to in this paper were collected at the end of August 2000 during several biospeleological expeditions authorised by the National Museum and Research Centre of Altamira (Cantabria, Spain). We especially thank, unfortunately posthumously, its director, José Antonio Lasheras (1956-2016), for his invaluable collaboration throughout the whole period of research, and we dedicate this work to his memory. At the end of August 2000, studies were conducted with the assistance and support of Dr Lucía Labrada, who was employed as a member of the research staff of the University of Columbia (New York, USA). Some specimens were kindly donated by Dr Cesáreo Saiz Jiménez (Institute of Natural Resources and Agrobiology-CSIC, Sevilla, Spain). We thank the reviewers (Eduardo Mateos and an anonymous reviewer), and Louis Dehaveng (Editor) for their careful reading of our manuscript and their many insightful comments and suggestions, which have allowed us to produce a more robust manuscript.

## References

Beruete E, Baquero E, Jordana R (2002) New species of Pseudosinella (Collembola: Entomobryidae) from karst caves on the Basque bio-speleologic district. Annales de la Société Entomologique de France (Nouvelle série) 38(4): 385-398. https://doi.org/10.1080/00379271.2002.10697351
Bonet F (1931) Estudios sobre Colémbolos cavernícolas con especial referencia a los de la fauna española. Memorias de la Sociedad Española de Historia Natural, Tomo XIV, Memoria 4a: 231-403.
Börner C (1913) Die Familien der Collembolen. Zoologischer Anzeiger 41: 315-322.
Christiansen K, Bellinger P, Gama MM da (1990) Computer Assisted Identification of Specimens of Pseudosinella (Collembola: Entomobryidae). Revue d'Écologie et de Biologie du Sol 27(2): 231-246.
Christiansen K, Gama MM da, Bellinger P (1983) A catalogue of the species of the genus Pseudosinella. Ciencia Biologica Ecology and Systematics 5: 13-31.
Christiansen K (2013) NaviKey 2.3 for Collembola dataset. http://www.math.grin.edu/ -twitchew/ coll/navikey.html [Accessed on: 2013-5-29]
Gisin H (1950) Quelques collemboles cavernicoles d'Italie du nord. Bollettino della Società Entomologica Italiana 80(9-10): 93-95.

Gisin H (1964) Collemboles d’Europe. VII. Revue Suisse de Zoologie 71: 649-678. https:// doi.org/10.5962/bhl.part. 75615
Gisin H (1965) Nouvelles notes taxonomiques sur les Lepidocyrtus. Revue d'Écologie et de Biologie du Sol 2(4): 519-524.
Gisin H (1967a) Deux Lepidocyrtus nouveaux pour l'Espagne (Collembola). Eos, Revista Española de Entomología 42: 393-395.
Gisin H (1967b) Espèces nouvelles et lignées évolutives de Pseudosinella endogés (Collembola). Memórias e Estudos do Museu Zoologico da Universidade de Coimbra 301: 1-25.
Gisin H, Gama MM da (1969) Espèces nouvelles de Pseudosinella cavernicoles (Insecta, Collembola). Revue suisse de Zoologie 76(6): 143-181. https://doi.org/10.5962/bhl.part. 146030
Gisin H, Gama MM da (1970) Pseudosinella cavernicoles de France (Insecta: Collembola). Revue suisse de Zoologie 77(1): 161-188. https://doi.org/10.5962/bhl.part. 75885
Gisin H, Gama MM da (1972) Pseudosinella cavernicoles d'Espagne (lnsecta: Collembola). Revue suisse de Zoologie 79(1): 261-278. https://doi.org/10.5962/bhl.part. 97127
Gouze A, Deharveng L (1987) Deux espèces nouvelles de Pseudosinella cavernicoles du Var (Coll.). Bulletin de la Société Entomologique de France 91(3-4): 65-68.
Jeannel R, Racovitza EG (1914) Biospeologica XXXIII: énumération des grouttes visitées 1911-1913 ( $5^{e}$ série). Archives de Zoologie Expérimentale et Générale 53: 325-547.
Jordana R, Baquero E, Ariño AH (2020) Continuously updated: Collembola DELTA database. University of Navarra. http://www.unav.es/unzyec/collembola/ [Accessed on: 2018-2-28]
Lashera JA (2009) The cave of Altamira: 22000 years of history. Adoranten Journal of Scandinavian Society for Prehistoric Art 2009: 5-33. https://www.rockartscandinavia.com/articles.php?article_id=20 [Accessed on: 2020-3-16]
Lubbock J (1873) Monograph of the Collembola and Thysanura. Ray Society, London, 276 pp. https://doi.org/10.5962/bhl.title. 11583
Luque CG, Labrada L (2014) Historia de la espeleología en Cantabria (1861-1936). Contribución de la sección de Santander de la Real Sociedad Española de Historia Natural a la institucionalización de la Espeleología Científica en España a principios del siglo XX. Boletín de la Real Sociedad Española de Historia Natural, Sección Biológica 108: 27-51.
Luque CG, Labrada L (2016) La fauna subterránea de las cuevas de Altamira (España). Consideraciones para la conservación del arte rupestre clasificado Patrimonio Mundial. Boletín de la Real Sociedad Española de Historia Natural, Sección Biológica 110: 93-120.
Mateos E (2008) The European Lepidocyrtus Bourlet, 1839 (Collembola: Entomobryidae). Zootaxa 1769: 35-59. https://doi.org/10.11646/zootaxa.1769.1.2
Sánchez MA, Foyo A, Tomillo C, Iriarte E (2007) Geological risk assessment of the area surrounding Altamira Cave: A proposed natural risk index and safety factor for protection of prehistoric caves. Engineering Geology 94: 180-200. https://doi.org/10.1016/j. enggeo.2007.08.004
SC-85/Conf. 008/9 (1985) Comité du Patrimoine Mondial, Neuvième Session (Paris, França, 2 a 6 dez. 1985). Relatório do relator. dez. 1985, 20 pp. [anexos, 10 pp .] http://whc.unesco.org/en/documents/895 [Accessed on: 2020-3-16]
Schäffer C (1896) Die Collembolen der Umgebung von Hamburg und benachbarter Gebiete. Mitteilungen aus dem Naturhistorischen Museum Hamburg 13: 149-216.
Schäffer C (1897) Apterygoten. Hamburger Magalhaensische Sammelreise Apterygoten 8: 1-48.

Soto-Adames FN (2010) Two new species and descriptive notes for five Pseudosinella species (Hexapoda: Collembola: Entomobryidae) from West Virginian (USA) Caves. Zootaxa 2331: 1-34.
Stomp N (1977) Nouvelle contribution a l'etude des Pseudosinella endoges d'Europe (Collemboles, Entomobryidae). Revue d'Écologie et de Biologie du Sol 14(1): 231-239.
Stomp N (1986) Le groupe de Pseudosinella vandeli (Insectes, Collemboles) dans les Alpes Orientales. International Seminar on Apterygota 85-91.
Szeptycki A (1979) Morpho-systematic studies of Collembola. IV. Chaetotaxy of the Entomobryidae and its phylogenetical significance. Polska Akademia Nauk, Zakład Zoologii Systematycznej i Doświadczalnej, Państwowe Wydawnictwo Naukowe, Warszawa and Kraków, 218 pp.
Wahlgren E (1906) Apterygoten aus Ägypten und dem Sudan nebst Bemerkungen zur Verbreitung und Systematik der Collembolen. Results of the Swedish Zoological Expedition to Egypt and the White Nile 1901 under the Direction of L.A. Jägerskiöld, Uppsala, 1-72.
Winkler D, Mateos E (2018) New species of Pseudosinella Schäffer, 1897 (Collembola, Entomobryidae) from Hungary. Zootaxa 4382(2): 347-366. https://doi.org/10.11646/zootaxa.4382.2.7

# New species and records of Cloeodes Traver, 1938 (Ephemeroptera, Baetidae) from Costa Rica 

Oscar Vásquez-Bolaños', Fabián Sibaja-Araya', Meyer Guevara-Mora'<br>I Laboratorio de Entomología (LEUNA), Escuela de Ciencias Biológicas, Universidad Nacional, Heredia, Costa Rica<br>Corresponding author: Oscar Vásquez-Bolaños (oscarv06@gmail.com)

Academic editor: E. Dominguez | Received 17 April 2020 | Accepted 3 September 2020 | Published 9 November 2020
http://zoobank.org/828AC238-8375-4893-81DD-3DB345429ECD
Citation: Vásquez-Bolaños O, Sibaja-Araya F, Guevara-Mora M (2020) New species and records of Cloeodes Traver, 1938 (Ephemeroptera, Baetidae) from Costa Rica. ZooKeys 989: 55-72. https://doi.org/10.3897/zookeys. 989.53018


#### Abstract

The nymph of Clooodes danta sp. nov. is described from male and female nymphs collected from highland streams in the Caribbean Slope of the Costa Rica Central Volcanic Mountain Range. Adults are unknown. In addition, C. excogitatus and C. redactus are recorded for the first time in the country. Cloeodes danta sp. nov. can be differentiated from all described species by the predominantly brownish coloration on females and a similar coloration on males but with segments VII-IX light yellow and light brown, with no conspicuous marks or patterns; abundant scale-bases throughout most parts of the body; hindwings pads absent; the presence of three spines in the corners of the posterior margin of sternum III, and the posterior margin of tergum III with 28-30 spines on each side of the middle line (spine with a base width up to $0.5 \times$ spine length).


## Keywords

diversity, freshwater, mayflies, Neotropics, taxonomy

## Introduction

The genus Cloeodes (Ephemeroptera, Baetidae) was erected by Traver (1938) for a group of species described from Puerto Rico with hind wings absent. Later, the genus was reviewed by Waltz and McCafferty (1987) and established for other species distributed across the Neotropics, Southern Nearctic, Afrotropics, and Oriental region. More recently, through phylogenetic analysis the genus has been depicted as exclusive for the New World within a complex including Bungona spp., Crassolus spp. and Potamocleoen spp. (Salles et al. 2016). Currently, the number of Cloeodes species has doubled in the last ten years with 26 species described for the Neotropics (Salles et al. 2016; Kluge 2017).

Despite the increase in the number of papers related to Cloeodes and the report of 142 mayfly species in Central America and the Caribbean (Cornejo and Flowers 2015; Salles et al. 2016), there are only two Cloeodes species formally reported from the Central American region: C. excogitatus (Waltz \& McCafferty, 1987) and C. redactus (Waltz \& McCafferty, 1987), registered from Guatemala and Honduras, respectively (McCafferty and Lugo-Ortiz 1996; McCafferty et al. 2004). In Costa Rica, the presence of the genus has been recorded for lowland and midland streams with scarce human intervention and good water quality (McCafferty et al. 1992; Flowers and De la Rosa 2010), but no species have been formally reported until now.

During an ongoing study of the richness and taxonomy of the family Baetidae in Costa Rica, nymphs of a new Cloeodes species and nymphs of C. excogitatus and $C$. redactus were collected in low order streams. This work provides a complete description of this new species and diagnostic characters for distinguishing C. excogitatus and C. redactus, in order to improve the taxonomic knowledge of the genus for future natural history or ecological studies of mayflies, which have crucial knowledge gaps in the tropics (Balian et al. 2008; Ramírez and GutiérrezFonseca 2014).

## Material and methods

Nymphs of these species were collected using a hand net to scrape large stone substrates from different streams in Costa Rica (Fig. 1). Line drawings for the new species description were made from original pictures taken with an AmScope 1803 digital camera adapted to a Premiere (MRP-161) microscope and stereomicroscope Premiere SMZ-05. The type material examined is preserved in $80 \%$ ethanol and Euparal permanent slides, and it has been deposited in Museo de Nacional de Costa Rica, San José Province; Laboratorio de Entomología (LEUNA), Escuela de Ciencias Biológicas, Universidad Nacional, Heredia; Museo de Zoología, Universidad de Costa Rica; and Purdue University Entomological Research Collection (PERC), West Lafayette, Indiana, USA.


Figure I. Geographic distribution in Costa Rica of Cloeodes danta sp. nov. (black circle), Cloeodes excogitatus (black triangle) and Cloeodes redactus (black square).

## Results

## Cloeodes danta Vásquez-Bolaños, Sibaja-Araya \& Guevara-Mora, sp. nov.

 http://zoobank.org/F2F0FC62-E8EE-4E79-93F6-48F711DAC45CFigures 2-6

Material examined. Holotype: mature $\widehat{0}$ nymph slide-mounted in Euparal, Costa Rica, Heredia Province, Central Volcanic Mountain Range, Cerro Chompipe zone, Refugio de Vida Silvestre Cerro Dantas, Río Las Vueltas, $10^{\circ} 5^{\prime} 38.24^{\prime \prime N}$, $84^{\circ} 3^{\prime} 42.57^{\prime \prime} \mathrm{W}$, 2000 m above sea level, VI/09/2017, Sibaja-Araya F. and Acuña F. (colls), deposited in Museo de Nacional de Costa Rica, San José Province. Paratypes: 5 mature nymphs same data as holotype, preserved in $95 \%$ alcohol (mouthparts, legs, gills, terga and paraprocts in micro-vial) deposited at Laboratorio de Entomología (LEUNA), Escuela de Ciencias Biológicas, Universidad Nacional, Heredia ( $1 \delta^{\lambda}, 1$ ) ; Museo de Zoología, Universidad de Costa Rica (1q) and PERC, West Lafayette, Indiana, USA (1才, 1q).

Additional material. Nine nymphs, Quebrada Grande, Refugio de Vida Silvestre Cerro Dantas, $10^{\circ} 5^{\prime} 38.24^{\prime \prime N}$, $84^{\circ} 3^{\prime} 42.57 " \mathrm{~W}$, about 1 km from the Río Las Vueltas at 2054 m asl, where type material was collected, Heredia Province, Costa Rica V/13/2018, F. Sibaja, D. Romero, M. Guevara, D. Segura, O. Vásquez (colls), deposited at Laboratorio de Entomología (LEUNA) of the Escuela de Ciencias Biológicas, Universidad Nacional.

Diagnosis. Mature nymph. 1) Brownish body coloration, without conspicuous marks or patterns (Fig. 2), 2) abundant scale-bases throughout most parts of the body (Figs 5b, 6h, 6i, 6k), 3) Absence of hind wing pads, 4) Tibia with two parallel lines of fine long hairs (Fig. 5b), 5) posterior margin of tergum III with 28-30 spines on each side of the middle line (Fig. 6h), 6) sterna with 26-28 variable size spines on each side of the middle line and three thick, spine-like teeth in each corner of the posterior margin of sternum III (Fig. 6i, 6j), 7) paraproct with about 13-15 spines (Fig. 7k).

Description. (based on last instar male and female nymphs; adults unknown).
Size (Mature nymphs): Body length: 5.3-6 mm males, $5.5-6.2 \mathrm{~mm}$ females; antennae $1.5-1.7 \mathrm{~mm}$; cerci $2.4-2.7 \mathrm{~mm}$; terminal filament $2.2-2.5 \mathrm{~mm}$.

Body coloration: Brownish in general (Fig. 2A, 2B), the head light brownish, with clearer area from central ocelli to the border of clypeus and between antennal and labrum bases; ocelli black, with two tiny white symmetrical dots on each side and clear brownish coloration, darker toward ocellus; eyes black, turbinated portion of compound eyes brownish. Fore wing pads brownish, foreleg brown with slightly lighter areas. Abdomen with even brownish coloration on females and males with predominantly brownish color but with segments VII-IX light yellow and light brown, both with no patterns of marks or spots, and upper corners of abdominal segments darkened; terga I-VIII with visible black posterior sigilla in the middle of every segment, terga VII-IX lighter with upper corners darkened; sterna pale yellowish-brown.

Head (Fig. 3A): Capsule longer than wide. Antennae subequal to $1.3 \times$ length of head, with scape subequal to $1.3 \times$ times length of pedicel (Fig. 3B). Intra-antennal extension of frons prolongs to ocelli.

Labrum (Fig. 3C). Subrectangular, broader than long, anterolateral margins rounded; dorsally with anterior margin with about 20 small, double frayed setae. Lateral margin with eight apically frayed setae; arc of anterodorsal setae with four simple setae; intermediate seta tiny; and several small scattered simple setae near posterior margin. Ventrally with small curved fine setae near anterior margin, and seven small simple setae near lateral margin. Anterior margin slightly asymmetric, with the left side of the cleft not extended the same length as the right side (Fig 3D).

Left mandible (Fig. 4A). Incisors with seven denticles, middle one reduced and others similar in size; prostheca robust, apically with three lobes and four elongated projections. Row of 5 or 6 minute spine-like setae between prostheca and molar region, only visible at high magnification (40x). Ventral surface of molar region with tuft of spines next to thumb as part of molar structure (Fig. 4B).


Figure 2. Clooodes danta sp. nov. Body coloration $\mathbf{A}$ male $\mathbf{B}$ female.


Figure 3. Cloeodes danta sp. nov. A head color pattern and intra-antennal extension detail B scape and pedicel C labrum (left d. v., right v. v.) D detail of labrum asymmetry on anterior margin.

Right mandible (Fig. 4C). Incisors with seven denticles, middle one reduced and others similar in size; prostheca with broad base, bifid, inner projection longer, and both parts frayed; 3-4 tiny, simple setae between prostheca and molar region only visible at high magnification $(40 \times)$; tuft of spines next to molar region.


Figure 4. Cloeodes danta sp. nov. A left mandible $\mathbf{B}$ detail of molar region, left mandible $\mathbf{C}$ right mandible $\mathbf{D}$ hypopharynx (left d. v., right v. v.) $\mathbf{E}$ maxilla.


Figure 5. Cloeodes danta sp. nov. A labium (left d. v., right v. v.) B foreleg $\mathbf{C}$ forefemur apex detail $\mathbf{D}$ gill III $\mathbf{E}$ gill III apex detail.

Hypopharynx (Fig. 4D). Lingua rounded with no apical lobes, slightly longer and broader than superlinguae, both apically covered with short fine hairs on dorsal and ventral surface.

Maxillae (Fig. 4E). Palpi slightly shorter or as long as galea-lacinia, two segmented; segment I slender in mid part; segment II $1.4 \times$ length of segment I; both segments with several simple short setae. Crown of galea-lacinia without thick distal dentisetae, with numerous setae on the inner-dorsal and inner ventral rows, and longer and slender towards the biting edge; medial region of galea-lacinia with one short seta and 5 or 6 long, simple setae.

Labium (Fig. 5A). Glossa and paraglossa similar in length, basally broad and apically narrow; with the base of the glossa reaching more than half of the paraglossa; glossae dorsally with 7-8 setae next to the inner margin and some scattered setae, ventrally with 12 or 13 setae on the inner margin and 9 or 10 on the outer margin, increasing in length apically in both cases. Paraglossae curve inwards, apex subtriangular, dorsally with 7 or 8 setae on the inner margin and 17 or 18 setae on outer margin. Palpi with segment I broad and $0.8 \times$ length of segment II and III combined, numerous micropores and simple, tiny setae on both dorsal and ventral surfaces; segment II with row of 6 or 7 setae on the dorsal surface; segment III suboval, dorsally with several short simple setae on apex and 20-25 simple setae of different size on the ventral surface, and the inner margin subequal to the outer margin.

Thorax: Hindwing pads absent; foreleg with abundant scale-bases and scattered micropores (Fig. 5B). Femur length about $4 \times$ maximum with; dorsal edge with 10 or 11 short, simple setae, ventrally bare, apex rounded with no evident projections, and two concave and apically rounded setae. Tibia with two lines of fine hairs arranged parallel to each other along the tibia, subtending bristle elongated and rounded (Fig. 5B, C). Tarsi dorsal edge bare, ventrally with 10 or 11 minute spines, increasing in size distally; tarsal claw length $0.5 \times$ length of tarsi. Mid and hind legs are similar to the foreleg.

Abdomen: Gills (Fig. 6A-G). Present on segments I-VII, colorless, oval, margins with curved fine setae (Fig. 6D, 6E); light-brown tracheae extending from main trunk, branching to margins. Gill I smaller than segment II; gill IV length about $1.3 \times$ length of segment III; gills on segment VII not extending beyond apex of segment X (Fig. 2A, B).

Terga. Tergum I with no spines on posterior margin; tergum III with 28-30 spines on posterior margin of each side of the middle line, spine bases almost same width as height (Fig. 6J).

Sterna (Fig. 6I). Posterior margin of sternum III with group of 26-28 variable size spines on each side of middle line, spines bases width about one-third of height; group of fine hairs arranged in slightly curved line on each side of sterna and three thick spine-like teeth on each corner of posterior margin of sternum III (Fig. 6H).

Paraprocts (Fig. 6K). Lateral margin with 13-15 spines, different in size and arranged in irregular line, with some overlapping; dorsal surface with numerous micropores and scale-bases; posterolateral extension with about 11 or 12 marginal spines, and several scale-bases and micropores.

Adults. Unknown.
Etymology. The name of this species honors the Danta (Tapirus bairdii) (Mammalia: Tapiridae), a common species in the Cerro Chompipe cloud forest zone, whose three-toed back feet resemble the sternal spine-like teeth described as a diagnostic character of C. danta sp. nov.


Figure 6. Cloeodes danta sp. nov. A gill I B gill II C gill III D gill IV E gill V F gill VI G gill VII Habdominal tergum III I abdominal sternum III J detail of sternum spine-like teeth $\mathbf{K}$ paraproct.

Distribution. Costa Rican, Caribbean slope basin, first order streams, above 2000 m asl.

Biology. Habitat preferences in C. danta sp. nov. were observed in relation to elevation. Individuals were found at 2054 m asl in cold waters of two cloud forest streams (Fig. 7); other non-Andean species have been reported between 159-1800 m asl (Nieto and Richard 2008; Gutiérrez and Reinoso-Flórez 2010; Massariol and Salles 2011; Massariol et al. 2013; De Paul et al. 2013; Gutiérrez and Dias 2015). Also, the new species was collected in a pristine forest region, which may be similar to other Cloeodes species in Brazil that seem to prefer remnant forest areas (Salles et al. 2015). In addition, C. danta sp. nov. is common in riffles, on igneous boulders that are covered with periphyton; this is typical for this genus, part of the grazer functional feeding group (Baptista et al. 2006).

## Cloeodes excogitatus (Waltz \& McCafferty, 1987)

Cloeodes excogitatus Waltz \& McCafferty, 1987: 200; McCafferty and Lugo-Ortiz 1996: 23; Wiersema and Baumgardner 2000: 61; McCafferty et al. 2004: 207.

Material examined. Paratype, Clooodes excogitatus, R.W. Koss and R. Baumann, 1 ठ / Collected 12/V/1968, Arizona, Oak Creek Canyon, slide-mounted in Euparal (Abs. Alc.) by R.B. Waltz VI/1983, det. by Waltz and MacCafferty, 1984. (Paratype),


Figure 7. Habitat of Cloeodes danta sp. nov. at Río Las Vueltas.

PERC $(0,012,327)$. Cloeodes excogitatus, DE Baumgardner, 1 male/ 1 female / Collected 21/V/2004, Arizona, Geenlee CO. San Francisco R. at FS Road, 212, ca 1 mi . N. Clifton. $33^{\circ} 04^{\prime} 30^{\prime \prime} \mathrm{N}, 109^{\circ} 18^{\prime} 04^{\prime \prime} \mathrm{W}$, Elev. 3700 ft , (DB04-21), det. M. Meyer 2005. 10 mature nymphs, Río Torito, La Fuente, Santa Teresita, Turrialba, Cartago, Costa Rica, $9^{\circ} 59^{\prime} 13.04^{\prime \prime N}, 83^{\circ} 40^{\prime} 44.95^{\prime \prime} \mathrm{W}, 1063 \mathrm{~m}$ asl, II/28/2017. Romero D. and Siba-ja-Araya F, (colls). Material in 95\% alcohol, deposited at Laboratorio de Entomología (LEUNA), Escuela de Ciencias Biológicas, Universidad Nacional, Heredia, Costa Rica.

Distribution. Guatemala, Mexico, USA (Waltz and McCafferty 1987; McCafferty and Lugo-Ortiz 1996; Wiersema and Baumgardner 2000; McCafferty et al. 2004), and Costa Rica.

## Cloeodes redactus (Waltz \& McCafferty, 1987)

Cloeodes redactus Waltz \& McCafferty, 1987: 204; McCafferty and Lugo-Ortiz 1996: 23; Nieto and Emmerich 2011: 58; Massariol et al. 2013: 11; Forero-Céspedes et al. 2016: 462; Kluge 2017: 104.

Material examined. Five mature nymphs, Río Claro tributary, Río Claro, Golfito, Puntarenas, Costa Rica. $8^{\circ} 41^{\prime} 13.05^{\prime \prime N}$, $83^{\circ} 02^{\prime} 48.78^{\prime \prime W}$ W, 79 m asl, XII/25/2018. Coll. Siba-ja-Araya F. (coll.). Material in 95\% alcohol deposited at Laboratorio de Entomología, Escuela de Ciencias Biológicas (LEUNA), Universidad Nacional, Heredia, Costa Rica.

Distribution. Colombia, Honduras, Peru (Waltz and McCafferty 1987; McCafferty and Lugo-Ortiz 1996; Massariol et al. 2013; Forero-Céspedes et al. 2016; Kluge 2017), and Costa Rica.

## Discussion

Cloeodes danta sp. nov. shares morphological affinities with the following species: C. caraibensis Hofmann et al. 1999, C. excogitatus, C. redactus, and C. maculipes Traver, 1938, including, absence of hind wing pads, segment III of labial palp rounded, maxillary palp shorter than galea-lacinia, and the general shape of other structures like labrum, labium and the apex of the femur that were recognized by Hofmann et al. (1999) and Nieto and Richard (2008). Based on the material and literature reviewed, we consider that Central American and Caribbean species can be separated in two groups according to the depth of the central cleft on the right mandible incisives: in C. excogitatus and C. redactus the cleft breaks down below the level of mandible first inner incisor apex (Waltz and McCafferty 1987: fig. 27; Kluge 2017: fig. 66), while in C. danta sp. nov., C. caraibensis and C. maculipes the cleft breaks down at the level of the first inner incisor apex (Waltz and McCafferty 1987: figs 3, 4 ; Hofmann et al. 1999: fig. 37; Fig. 4C).

In regard to C. caraibensis from Lesser Antilles, these species mentioned above have setae on segment III of the labial palp as long as the setae on the glossa and paraglossa
(Waltz and McCafferty 1987: fig. 6; Hofmann et al. 1999: fig. 41), while in C. danta these setae are shorter (Fig. 5A). Also, labrum intermediate setae are well developed in C. maculipes (Waltz and McCafferty 1987: fig. 2), while in C. danta sp. nov. they are minute (Fig. 3C); in regard to C. caraibensis this character is absent, and this species also shares some diagnostic characters with C. danta sp. nov. like scale-bases present on most of the body, shape of III segment of labial palp and spines on the corner of the posterior margins of the sterna (Hofmann et al. 1999: figs 36, 41, 50); however, $C$. danta sp. nov. can be differentiated by no spots or color pattern on the abdomen, right mandible prostheca with two well-developed branches, glossa apically not lobulated, distal portion of gills rounded, minute spines on the terga and sterna margins, and the posterior margin of the paraproct with no bifurcated spines (Figs $2 \mathrm{~A}, 2 \mathrm{~B}, 4 \mathrm{C}, 4 \mathrm{D}$, 6A-6G, 6K).

Furthermore, C. danta sp. nov. shares similar features with C. excogitatus, such as the number of spines in the terga III, the shape segment III of labial and the abundance of setae on it (20-25), the number of spines in the paraproct, and body size (Waltz and McCafferty 1987, but C. danta sp. nov. can be identified by the different abdominal color pattern, abundant scale-bases throughout most parts of the body, the maxillar palp is equal in length to the galea-lacinia and the absence of a narrow intra-antennal extension. Also, the new species resembles $C$. redactus in the length of the maxillary palp being about as long as the galea-lacinia, the number of spines in the paraproct and the lack of any projections on the apex of the femur (Waltz and MacCafferty 1987; Kluge 2017); however it could be distinguished by the shape and number of spines in terga III, the absence of the colorless spots in some terga, a larger body size, the presence of an intermediate setae on the labrum, and the presence of thick spine-like teeth in the inferior corner of sterna III.

In order to improve the identification of Cloeodes species in the Central American region, we provide a key to distinguish C. danta sp. nov., C. excogitatus and C. redactus. This will be a useful tool for future aquatic research in the region, which has been increasing over the last 20 years due to development of water quality monitoring using aquatic insects (Alonso-Eguía et al. 2014; Carrie et al. 2017; Castillo 2018; DávilaRecinos et al. 2019; Pérez 2019).

## Key to mature nymphs of Cloeodes species in Central America

1 Abdomen with brownish uniform color pattern, without spots or distinctive marks; body covered with scale-bases; intermediate setae minute; cleft on right mandible incisive breaks down at level of first inner incisor apex and thick spines like teeth present in sternal III corners (Figs 2A, 2B, 4C, 6I, 6J); highlands ( 2054 m asl)
C. danta sp. nov

- Abdomen with marks or spotted color pattern (Figs 8A, 9A); body not covered with scale-bases cleft on right mandible incisives breaks down below level of mandible first inner incisor apex; intermediate setae absent; no spines on sternal corners of sterna III (Waltz and McCafferty 1987; Kluge 2017); mid or lowlands.

2 Abdominal coloration on terga I-III and V-VII with 3 pale spots (the middle spot being smaller); intra-antennal extension narrow, labrum arc of anterodorsal setae with 5 simple setae; segment III of labial palp ventrally with 20-25 setae; maxillary palp shorter than galea-lacinia; $30-35$ spines on tergum III (Fig. 8B, C; Waltz and McCafferty 1987: figs 25, 36, 40); midlands ( 1063 m asl)
C. excogitatus

Abdominal coloration with terga I-III and V-VII with dark brown transverse stripes on posterior margins; intra-antennal extension not narrowed; labrum arc of anterodorsal setae with 3 simple setae; segment III of the labial palp with 30-35 setae; maxillary palp about same length as galea-lacinia; 15-20 spines on tergum III (Fig. 9B, C; Kluge, 2017: figs 6, 12, 14, 58, 59); lowlands ( 79 m asl)
C. redactus


Figure 8. Cloeodes excogitatus: A abdomen coloration B intra-anternal extension (scale:1000 pixels : $2 \mathrm{~mm}) \mathbf{C}$ spines on tergum III.


Figure 9. Clooodes redactus: $\mathbf{A}$ abdomen coloration $\mathbf{B}$ intra-anternal extension $\mathbf{C}$ spines on tergum III.

## Acknowledgments

We thank our team at the Laboratorio de Entomología (LEUNA) (Carlos Esquivel, David Romero, and Francisco Bravo) for their help both in the field and in the lab with their companionship. We also thank the Escuela de Ciencias Biológicas from Universidad Nacional for their financial and logistical support. We are also grateful with Luke Jacobus (Division of Science, Indiana University Purdue University Columbus for the comments in the early draft, and Sudeep Chandra (Aquatic Ecosystems Laboratory University of Nevada Reno) for facilitating a space to examine part of the material.

This work is part of the research project "Estudio preliminar de la ecología y diversidad del orden Ephemeroptera en Costa Rica", SIA 0230-13, and Laboratorio de Entomología (LEUNA), SIA 0484-18, Escuela de Ciencias Biológicas

## References

Alonso-Eguía P, Mora JM, Campbell B, Springer M (2014) Diversidad, conservación y uso de los macroinvertebrados dulceacuícolas de México, Centroamérica, Colombia, Cuba y Puerto Rico. Instituto Mexicano de Tecnología del Agua. http://repositorio.imta.mx/handle/20.500.12013/1684
Balian EV, Segers H, Lévêque C, Martens K (2008) The freshwater animal diversity assessment: an overview of the results. Hydrobiologia 595(1): 627-637. https://doi.org/10.1007/978-1-4020-8259-7_6
Baptista DF, Buss DF, Dias LG, Nessimian JL, Da Silva ER, Neto DM, Andrade LR (2006) Functional feeding groups of Brazilian Ephemeroptera nymphs: ultrastructure of mouthparts. Annales de Limnologie-International Journal of Limnology 42(2): 87-96. https:// doi.org/10.1051/limn/2006013
Carrie R, Dobson M, Barlow J (2017) Challenges using extrapolated family-level macroinvertebrate metrics in moderately disturbed tropical streams: a case-study from Belize. Hydrobiologia 794(1): 257-271. https://doi.org/10.1007/s10750-017-3100-z
Cornejo A, Flowers W (2015) Orden Ephemeroptera (Insecta) en Panamá: Listado de especies, distribución de géneros y comparación con la riqueza taxonómica regional. Puente Biológico 7: 37-67. http://pluseconomia.unachi.ac.pa/index.php/puentebiologico/article/ view/396
Castillo FS (2018) Evaluación de la calidad ambiental del Humedal Refugio de Vida Silvestre Sistema Lagunar de Tísma, Masaya, Nicaragua. Thesis. Universidad Nacional Agraria. Managua, Nicaragua. http://repositorio.una.edu.ni/id/eprint/3682
Dávila-Recinos G, Ortiz JR, Reyes-Morales F (2019) Efecto del microhábitat sobre la abundancia y riqueza específica de los macroinvertebrados bentónicos en dos ríos tropicales de montaña, Guatemala. Ciencia, Tecnología y Salud 6(1): 7-21.
De Paul MA, Nieto C, De Roman LE (2013) Cloeodes incus (Waltz and McCafferty) (Ephemeroptera: Baetidae): primer registro de Argentina, características hidrológicas y ambientales. Revista de la Sociedad Entomológica Argentina 72(3-4): 227-230.https://www.redalyc. org/pdf/3220/322030024012.pdf
Flowers RW, De la Rosa C (2010) Ephemeroptera. Revista de Biología Tropical 58: 63-93. https://www.scielo.sa.cr/pdf/rbt/v58s4/a04v58s4.pdf
Forero-Céspedes AM, Gutiérrez C, Reinoso-Flórez G (2016) Composición y estructura de la familia Baetidae (Insecta: Ephemeroptera) en una cuenca andina colombiana. Hidrobiológica 26(3): 459-474. https://doi.org/10.24275/uam/izt/dcbs/hidro/2016v26n3/Forero
Gutiérrez C, Reinoso-Flórez G (2010) Géneros de ninfas del orden Ephemeroptera (Insecta) del departamento del Tolima, Colombia: listado preliminar. Biota Colombiana 11(1-2): 23-32. http://revistas.humboldt.org.co/index.php/biota/article/view/229/227

Gutiérrez Y, Dias LG (2015) Ephemeroptera (Insecta) de Caldas-Colombia, claves taxonómicas para los géneros y notas sobre su distribución. Papéis Avulsos de Zoologia 55(2): 13-46. https://doi.org/10.1590/0031-1049.2015.55.02
Hofmann C, Sartori M, Thomas A (1999) Les Ephéméroptères (Ephemeroptera) de la Guadeloupe (petites Antilles françaises). Mémoires de la Société Vaudoise des Sciences Naturelles 20(1): 20-70.
Kluge NJ (2017) Contribution to the knowledge of Cloeodes Traver 1938 (Ephemeroptera, Baetidae). Zootaxa 4319(1): 91-127. https://doi.org/10.11646/zootaxa.4319.1.5
Massariol FC, Salles FF (2011) Two new species of Cloeodes Traver (Ephemeroptera: Baetidae) from Espírito Santo, Southeastern Brazil. Zootaxa 3058: 1-21. https://doi.org/10.11646/ zootaxa.3058.1.1
Massariol FC, Lima RC, Pinheiro UDS, Quieroz LL, Oliveira LG, Salles FF, Takiya D (2013) Two-winged Cloeodes in Brazil: New species, stage descrip-tion, and key to South American species. Journal of Insect Science 13(60): 1-20. https://doi.org/10.1673/031.013.6001
McCafferty WP, Baumgardner DE, Guenther JL (2004) The Ephemeroptera of Central America. Part 1: Guatemala. Transactions of the American Entomological Society 130: 201-219. http://www.ephemeroptera-galactica.com/pubs/pub_m/pubmccaffertyw2004p201.pdf
McCafferty WP, Flowers RW, Waltz RD (1992) The biogeography of Mesoamerican mayflies. Insects of Panama and Mesoamerica, D. Quintero y A. Aiello (Eds) Oxford University Press, Oxford 173-193. http://www.ephemeroptera-galactica.com/pubs/pub_m/pubmccaffertyw1992p173.pdf
McCafferty WP, Lugo-Ortiz CR (1996) Los efemerópteros (Ephemeroptera) de América Central. Revista Nicaragüense de Entomología 35: 19-28. http://www.insecta.bio.spbu.ru/z/ pdf/McCaffertyLugo-Ortiz1996p19.pdf
Nieto C, Emmerich DE (2011) Three new species of the genus Cloeodes Traver (Ephemeroptera: Baetidae) from Uruguay. Zootaxa 2996: 57-65. https://doi.org/10.11646/ zootaxa.2996.1.4
Nieto C, Richard B (2008) The genus Cloeodes (Ephemeroptera: Baetidae) in Argentina with new generic synonymy and new species. Zootaxa 1727: 1-21. https://doi.org/10.11646/ zootaxa.1727.1.1
Pérez AM (2019) Informe sobre la evaluación de la calidad del agua de la quebrada Zapatero del municipio de La Plata Huila, mediante la aplicación de los métodos BMWP/COL y ASPT correlacionados con parámetros fisicoquímicos. Thesis. Universidad Nacional Abierta y a Distancia. La Plata Huila, Honduras. https://repository.unad.edu.co/bitstream/handle/10596/28229/adperezlo.pdf?sequence=1
Ramirez A, Gutiérrez-Fonseca PE (2014) Studies on Latin American freshwater macroinvertebrates: recent advances and future directions. Revista de Biología Tropical 62: 9-20. https:// doi.org/10.15517/rbt.v62i0.15775
Salles FF, Batista JD, Cabette HRS (2004) Baetidae (Insecta: Ephemeroptera) de Nova Xavantina, Mato Grosso, Brasil: novos registros e descrição de uma nova espécie de Cloeodes Traver. Biota Neotropica 4(2): 1-8. https://doi.org/10.1590/S0085-56262011005000048
Salles FF, Massariol FC, Angeli KB, Lima MM, Gattolliat JL, Sartori MS (2015) Revealing the diversity of Cloeodes Traver, 1938 (Ephemeroptera: Baetidae) in the Neotropics: descrip-
tion of eleven new species from Brazilian mountain ranges. Zootaxa 4020(1): 1-50. http:// dx.doi.org/10.11646/zootaxa.4020.1.1

Salles FF, Gattolliat JL, Sartori M (2016) Phylogenetic analyses of Cloeodes Traver and related genera (Ephemeroptera: Baetidae). Systematic Entomology 41(1): 93-111. https://doi. org/10.1111/syen. 12144
Traver JR (1938) Mayflies of Puerto Rico. The Journal of Agriculture of the University of Puerto Rico 22: 5-42.
Waltz RD, McCafferty WP (1987) Revision of the genus Cloeodes Traver (Ephemeroptera: Baetidae). Annals of the Entomological Society of America 80 (2): 191-207. https://doi. org/10.1093/aesa/80.2.191
Wiersema NA, Baumgardner DE (2000) Distribution and taxonomic contributions to the Ephemeroptera fauna of Mexico and Central America. Entomological news 111(1): 6066. http://www.insecta.bio.spbu.ru/z/pdf/WiersemaBaumgardner2000.pdf

# A third species of the rarely collected Oriental hoverfly genus Furcantenna Cheng, 2008 (Diptera, Syrphidae, Microdontinae) 

Menno Reemer ${ }^{1}$<br>I Naturalis Biodiversity Center, P.O. Box 9517, 2300, RA Leiden, the Netherlands<br>Corresponding author: Menno Reemer (menno.reemer@naturalis.nl)

Academic editor: K. Jordaens \| Received 30 July 2020 | Accepted 21 September 2020 \| Published 9 November 2020
http://zoobank.org/1 B6E00CB-D9D7-4D76-A446-163BCBCC744F
Citation: Reemer M (2020) A third species of the rarely collected Oriental hoverfly genus Furcantenna Cheng, 2008 (Diptera, Syrphidae, Microdontinae). ZooKeys 989: 73-78. https://doi.org/10.3897/zookeys.989.57087


#### Abstract

Furcantenna malayana sp. nov. is described from Peninsular Malaysia, based on a single female collected in 1962. The other two known species of this genus are also known from single specimens, from Southeastern China and Nepal. A key to the species is given, and the taxonomy and distribution of the genus are discussed.


## Keywords

Identification key, Malaysia, morphology, new species, taxonomy

## Introduction

Few species of Syrphidae are known in which the basal antennal flagellomere (the apical segment of the antenna, which carries the arista) is furcate. In the subfamily Eristalinae a bifurcate basal antennal flagellomere is only found in males of the genus Cacoceria Hull, 1936. All other instances of this remarkable character among Syrphi-
dae are restricted to the subfamily Microdontinae. In this subfamily, the character is known in the New World genera Carreramyia Doesburg, 1966, Masarygus Brèthes, 1909, Schizoceratomyia Carrera, Lopes \& Lane, 1947 and an unnamed genus; in the Australian genus Cervicorniphora Hull, 1945; and in the Oriental genus Furcantenna Cheng, 2008 (Reemer and Ståhls 2013a).

The genus Furcantenna was described based on a single male specimen of F. yangi Cheng, 2008, collected in 1982 in the Guangxi Province in the Oriental part of China (Cheng and Thompson 2008). The second species, F. nepalensis Reemer, 2013, was described based on a single male specimen collected in 1967 in Nepal (Reemer and Ståhls 2013a). Ongoing taxonomic research in various entomological collections by the author revealed a third specimen of the genus, belonging to a third species, collected in 1962 in Peninsular Malaysia. This species is described here, and a key to distinguish it from the other two Furcantenna species is given.

## Material and methods

The studied specimen is deposited in the collection of the California Academy of Sciences (CAS) in San Francisco, USA. Photos of the specimen were made through a Zeiss Stereo Discovery v12 microscope, and processed (focus stacking) by Axiovision (version SE64 4.9) software. The map was made in Adobe Illustrator, based on locality information in Cheng and Thompson (2008), Reemer and Ståhls (2013a) and the present article (the symbols for the localities were placed by approximation, as available locality information is imprecise). Measurements of the studied specimen were taken using an ocular micrometer in a Wild M3B stereo microscope. Body length was measured from the anterior part of head (excluding antenna) to the apex of the abdomen. Morphological terminology follows Cumming and Wood (2017). For the type specimen, label data are given in quotation marks ("...") and line breaks on the label are indicated with a slash (/).

## Results

## Generic diagnosis

The genus Furcantenna is characterized by the combination of the following characters: wing vein $R_{4+5}$ without posterior appendix; wing vein $M_{1}$ straight, perpendicular to wing vein $\mathrm{R}_{4+5}$; abdomen broadly oval; antenna inserted below dorsal eye margin; basal antennal flagellomere much longer than scape, longer than distance between antennal socket and anterior oral margin; scutellum apicomedially sulcate; metasternum developed.

The pilosity of the katepisternum is variable: bare in F. yangi (Cheng and Thompson 2008), pilose in the other two species. The pilosity of the metasternum is also variable: bare in F. malayana sp. nov., pilose in the other two species.

## Key to the species of Furcantenna

1 Face and vertex black. Wing in apical half dark, at least anteriorly. Metasternum pilose. Katepimeron pilose or bare

- Face medially and vertex partly yellow (Figs 1-4). Wing in apical half whitish (Fig. 7). Metasternum bare. Katepimeron pilose ...... F. malayana Reemer, sp. nov.
2 Body colour brownish without violet shine. Katepimeron pilose. Scutellum slightly sulcate................................................................. F. nepalensis Reemer
- Body colour blackish with violet shine. Katepimeron bare. Scutellum deeply sulcate $\qquad$ .F. yangi Cheng


## Furcantenna malayana sp. nov.

 http://zoobank.org/DED6DB6F-ADDA-4027-A0CB-295956C9CC47 Figures 1-8Material. Holotype: Malaysia. • 1 , holotype of Furcantenna malayana sp. nov.; SE of Ipoh; alt. $50 \mathrm{~m} ; 10$ Oct. 1962; E.S. Ross \& D.Q. Cavagnaro leg.(CAS).

Label 1: "MALAYA: 3 mi. / SE. Ipoh / 50m VII-10-62"; label 2: "Collectors: / E.S. Ross / D.Q. Cavagnaro".

Diagnosis. Furcantenna malayana is the only known species of the genus with a partly yellow face and vertex, and a bare metasternum. The whitish apical half of the wing is also characteristic.

Description. Adult female. Body length: 9 mm (Fig. 1).
Head. Face occupying slightly more than $1 / 2$ of head width in frontal view; yellow medially, dark brown laterally; silvery white pilose. Oral cavity with lateral margins not produced. Frons medially yellow, bare; laterally dark brown, golden yellow pilose. Vertex swollen; yellow, except brown around ocellar triangle; yellow pilose. Occiput dark brown; yellow pilose. Eye bare, except for a few scattered, very short pile, only visible under high magnification (Fig. 4). Antennal socket about $1.5 \times$ as high as wide. Antenna brown; ratio of scape:basal flagellomere approx. 1:2.1; basal flagellomere swollen at apical $1 / 4$, appearing club-shaped, with apex approx. twice as wide as base. Arista yellowish, about 3/4 as long as basal flagellomere (Fig. 3).

Thorax. Postpronotum yellowish brown; yellow pilose. Mesoscutum brown, quite pale along margins and more blackish in middle; golden yellow pilose in anterolateral corners and along posterior margin, leaving a black pilose area in the shape of an upside-down letter T in between (Fig. 5). Postalar callus yellowish brown; yellow pilose. Scutellum without calcars, with shallow apicomedian sulcus; pale brown; black pilose (Fig. 6). Pleuron brown. Anterior and posterior part of anepisternum divided by a weak sulcus; almost entirely yellow pilose, except bare on narrow strip along dividing sulcus and on small ventral part. Anepimeron on anterior part with mixed yellow and black pile, other parts bare. Katatergum and anatergum long and short microtrichose, respectively. Katepisternum dorsally whitish pilose, ventrally bare. Katepimeron white pilose. Metasternum bare (except for dark microtrichia). Mediotergite dark brown;


Figures I-8. Furcantenna malayana sp. nov.,female, holotype $\mathbf{I}$ habitus dorsal $\mathbf{2}$ habitus lateral $\mathbf{3}$ head frontal $\mathbf{4}$ head lateral $\mathbf{5}$ thorax dorsal $\mathbf{6}$ scutellum dorsal $\mathbf{7}$ wing $\mathbf{8}$ abdomen dorsal.


Figure 9. Known localities of Furcantenna species.
shining, except narrowly microtrichose anteriorly and on rudimentary subscutellum. Calypter grey. Halter pale yellow.

Wing. Brown in basal half, whitish in apical half (Fig. 7). Microtrichose, except bare on posterobasal $1 / 2$ of cell br.

Legs. Pale brown, except hind femur and tibia darker brown; black pilose, except ventral surface of tarsi yellow pilose. Hind tibia strongly swollen, widest in the middle, with dorsal pile much longer than on other parts of the legs (Fig. 2). Coxae and trochanters brown; black pilose.

Abdomen. Pale brown (Fig. 8). Tergites short pilose: tergite 1 yellow pilose; tergite 2 black pilose except yellow pilose along anterior margin; tergite 3 black pilose; tergites 4 and 5 yellow pilose. All sternites yellow pilose.

Etymology. The specific epithet is an adjective which refers to Malaysia, the country where the species was collected.

Distribution. Only known from Peninsular Malaysia (Fig. 9).

## Discussion

The description of Furcantenna malayana is based on the first known female of the genus. Even though the basal antennal flagellomere is not furcate, as it is in the males of the other two described species of Furcantenna, the specimen is considered to belong to
this genus. The combination of the following characters supports this placement: wing vein $R_{4+5}$ without posterior appendix; wing vein $M_{1}$ straight, perpendicular to wing vein $\mathrm{R}_{4+5}$; abdomen broadly oval; basal antennal flagellomere longer than distance between antennal socket and anterior oral margin; scutellum apicomedially sulcate; katepisternum pilose. Also, in most Neotropical counterparts of Furcantenna (species of Carreramyia, Masarygus and Schizoceratomyia) the furcate antenna is found only in males, with the exception of Schizoceratomyia malleri (Curran, 1947) and Masarygus carrerai Papavero, 1962 (the female of the Australian genus Cervicorniphora is unknown).

Furcantenna was placed in a clade with the Neotropical genera Carreramyia, Masarygus, and Schizoceratomyia in a phylogenetic analysis based on morphological characters (Reemer and Ståhls 2013b). If these genera are indeed closely related, this would be a case of a trans-Pacific distribution pattern as defined by Cranston (2005). Similar examples among Microdontinae are found in the subgenus Chymophila Macquart, 1834 of Microdon Meigen, 1803, and in the genus Paramicrodon de Meijere, 1913 (Reemer 2012).

Apparently the genus Furcantenna is quite widespread in the Oriental region, even though the species are very rarely collected. Whether their elusiveness is a result of very restricted ranges, occurrence in very low densities, or a lifestyle that makes them difficult to notice, remains a matter of speculation.

## Acknowledgements

I would like to thank Jeroen van Steenis for bringing the specimen of Furcantenna malayana sp. nov. under my attention. Jere Schweikert (CAS) is thanked for arranging the loan of the specimen.

## References

Cheng X-Y, Thompson FC (2008) A generic conspectus of the Microdontinae (Diptera: Syrphidae) with the description of two new genera from Africa and China. Zootaxa 1879: 21-48. https://doi.org/10.11646/zootaxa.1879.1.3
Cranston P (2005) Biogeographic patterns in the evolution of Diptera. In: Yeates DK, Wiegmann BM (Eds) The Evolutionary Biology of Flies. Columbia University Press, New York, 274-311.
Cumming JM, Wood DM (2017) 3. Adult morphology and terminology. In: Kirk-Spriggs AH, Sinclair B (Eds) Manual of Afrotropical Diptera (Vol. 1). Introductory chapters and keys to Diptera families. Suricata 4. South African National Biodiversity Institute, Pretoria, 89-134.
Reemer M (2012) Speculations on the historical biogeography of Microdontinae (Diptera: Syrphidae). In: Reemer M (Ed.) Unravelling a hotchpotch: phylogeny and classification of the Microdontinae (Diptera: Syrphidae). PhD thesis, Leiden University, 351-357.
Reemer M, Ståhls G (2013a) Generic revision and species classification of the Microdontinae (Diptera, Syrphidae). ZooKeys 288: 1-213.
Reemer M, Ståhls G (2013b) Phylogenetic relationships of Microdontinae (Diptera: Syrphidae) based on molecular and morphological characters. Systematic Entomology 38: 661-688.

# A taxonomic revision of the whitefish of lakes Brienz and Thun, Switzerland, with descriptions of four new species (Teleostei, Coregonidae) 

Oliver M. Selz ${ }^{1,2}$, Carmela J. Dönz ${ }^{1,2}$, Pascal Vonlanthen ${ }^{1,3}$, Ole Seehausen ${ }^{1,2}$<br>I Department of Fish Ecology and Evolution, Centre for Ecology, Evolution \& Biogeochemistry, Eawag: Swiss Federal Institute of Aquatic Science and Technology, 6047 Kastanienbaum, Switzerland $\mathbf{2}$ Aquatic Ecology and Evolution, Institute of Ecology and Evolution, University of Bern, 3012 Bern, Switzerland $\mathbf{3}$ Aquabios GmbH, Les Fermes 57, 1792 Cordast, Switzerland

Corresponding author: Oliver M. Selz (oliver.selz@eawag.ch)

Academic editor: M. E. Bichuette \| Received 3 January 2019 | Accepted 7 August 2019 | Published 9 November 2020
http://zoobank.org/F78F6D87-9DDB-4CD9-8E4C-60E4883A59B6
Citation: Selz OM, Dönz CJ, Vonlanthen P, Seehausen O (2020) A taxonomic revision of the whitefish of lakes Brienz and Thun, Switzerland, with descriptions of four new species (Teleostei, Coregonidae). ZooKeys 989: 79-162. https:// doi.org/10.3897/zookeys. 989.32822


#### Abstract

The alpha taxonomy of the endemic whitefish of lakes Brienz and Thun, Switzerland, is revised. We evaluate the status of seven known species: Coregonus steinmanni sp. nov., Coregonus profundus sp. nov. and Coregonus acrinasus sp. nov. are endemic to Lake Thun; Coregonus brienzii sp. nov. is endemic to Lake Brienz; and C. alpinus, C. albellus, and C. fatioi from lakes Brienz and Thun are redescribed. One of these species, C. alpinus, is revised, since the lectotype for this species is incongruent with the species description given by Kottelat (1997) and Kottelat and Freyhof (2007). The name C. alpinus is thus retained for the lectotype designated by Kottelat (1997) and a new description of this taxon provided. For the species otherwise described by Kottelat (1997) and Kottelat and Freyhof (2007) as C. alpinus the new name C. profundus is designated. Coregonus acrinasus is genetically partially of allochthonous origin, closely related to the radiation of Lake Constance, and we therefore compare it to all recognized species of Lake Constance, C. wartmanni, C. macrophthalmus, C. arenicolus, and C. gutturosus.


## Keywords

adaptive radiation, Coregonus, ecological speciation, taxonomy, whitefish

## Introduction

The European whitefish (Coregonus spp.) provide prime examples of postglacial adaptive radiations, with several lakes in the boreal, subarctic and prealpine climate zones harbouring multiple, often closely related and endemic species. Up to six species can occur in single lakes of the pre-alpine region (Hudson et al. 2016; Dönz et al. 2018), and perhaps even more in the largest lakes of north-east Europe (Bernatchez 2004; Hudson et al. 2007; Kottelat and Freyhof 2007). Many of these radiations diversified after the most recent retreat of the ice shields 10 '000 to $15^{\prime} 000$ years ago (Bernatchez 2004; Østbye et al. 2005; Hudson et al. 2011). Diversification was by a combination of geographically sympatric and allopatric speciation in boreal and subarctic lakes (Østbye et al. 2005; Præbel et al. 2013) and mainly by geographically sympatric speciation from an ancestral hybrid population in pre-alpine lakes (Hudson et al. 2011). Multiple axes of divergence appear to structure whitefish radiations in Europe that repeatedly and independently evolved ecologically similar sets of species ("ecomorphs" sensu William 1972) which exhibit parallel patterns of divergence in traits related to foraging (i.e., gill raker number, benthic vs. limnetic feeding ecology), physiology (i.e., growth rate, depth partitioning during feeding and breeding) and reproductive ecology (i.e., spawning season and spawning habitat varying along lake depth and along the benthic-pelagic axis) (Fatio 1890; Steinmann 1950; Østbye et al. 2005; Vonlanthen et al. 2009, 2012; Harrod et al. 2010; Lundsgaard-Hansen et al. 2013; Hudson et al. 2016; Dönz et al. 2018; Öhlund et al. 2020). The two common combinations of traits among species in European whitefish radiations are large, fast growing, sparsely gill-rakered, benthivorous fish spawning in shallow water versus small sized, slow growing, densely rakered, zooplanktivorous fish spawning in deep water, but other combinations of some of these traits can also be found in some species (Steinmann 1950; Vonlanthen et al. 2012; Hudson et al. 2016). The number of gill rakers on the first gill arch have shown to be heritable (Rogers and Bernatchez 2007; Roesch et al. 2013). Variation in this trait relates to variation in the relative efficiency of feeding on zooplankton and benthic prey items (Lundsgaard-Hansen et al. 2013; Roesch et al. 2013). Interspecific differences in body shape (Lundsgaard-Hansen et al. 2013) as well as in growth rates resulting in different body size at a given age have both been shown to be heritable too (Rogers and Bernatchez 2007; Lundsgaard-Hansen et al. 2013). A large number of molecular genetic tests of reproductive isolation among sympatric whitefish species in various Swiss lakes have confirmed that sympatric forms are generally genetically clearly differentiated species (Douglas and Brunner 2002; Douglas et al. 2003; Hudson et al. 2011; Vonlanthen et al. 2012; Hudson et al. 2016; Dönz et al. 2018; Feulner and Seehausen 2018; Jacobs et al. 2018; De-Kayne et al. unpublished).

Here, we revise the whitefish species of lakes Brienz and Thun, Switzerland. Whitefish can be found in the large pre-alpine lakes of France, Germany, Austria, and Switzerland, which historically harboured approximately 50 different species native to approximately 30 lakes in three major river drainages: the Rhine, the Danube, and the Rhone (Svärdson 1957; Kottelat and Freyhof 2007; Winkler et al. 2011; Vonlanthen
et al. 2012). Part of this species diversity has been lost during increased lake eutrophication in the last century (Vonlanthen et al. 2012). Phylogeographic studies have shown that the European C. lavaretus species complex, which diverged from the North American C. clupeaformis species complex at least 500,000 years ago, comprises of two divergent mitochondrial lineages (Bernatchez and Dodson 1991, 1994; Douglas et al. 2003; Østbye et al. 2005; Kottelat and Freyhof 2007; Hudson et al. 2011; Winkler et al. 2011). The two lineages overlap in their geographical distribution, whereby the "northern lineage" (mitochondrial N clade) is predominantly found in Scandinavia and the Baltic Sea region while the "central European lineage" (C clade) has a higher frequency of occurrence in the pre-alpine and North Sea region, hence more westerly (Hudson et al. 2011). The entire pre-alpine whitefish radiation is a monophyletic radiation as revealed by genomic AFLP-markers and whole-genome resequencing data when compared to the closest relatives from northern Germany and Scandinavia (Hudson et al. 2011; De-Kayne et al. unpublished). The occurrence of both central and north mitochondrial haplotype lineages within the pre-alpine radiation, today (Hudson et al. 2011) as well as already 5000 years ago (Alonso et al. 2017), and the frequent occurrence of both lineages within species of the radiation suggest that the entire pre-alpine radiation is of hybrid origin (Hudson et al. 2011). The ancient carriers of the two divergent mitochondrial lineages probably correspond to two glacial refugial lineages that came into secondary contact and hybridized before the hybrid population spread across much of western Europe and diversified into the modern species flocks. As the Alpine ice shields retreated, this hybrid population would have colonized the pre-alpine lakes and radiated within each of the larger lakes into several endemic species (Hudson et al. 2011). As a result, both haplotypes are shared among many of the approx. 25 contemporary endemic species that are native to 17 Swiss lakes (Steinmann 1950; Kottelat and Freyhof 2007; Hudson et al. 2011; Vonlanthen et al. 2012). Only one century ago Swiss lakes harboured approximately 35 endemic species of whitefish, but one third of this original diversity has been lost in the middle of the $20^{\text {th }}$ century due to a combination of speciation reversal through hybridization and demographic declines, both driven by loss of habitat for foraging and spawning and possibly of selective regimes, associated with anthropogenic eutrophication of lakes (Vonlanthen et al. 2012; Hudson et al. 2013; Alexander et al. 2017).

In this paper we compile and review morphological, genetic and ecological data for seven species of whitefish from the connected lakes Thun and Brienz, three of which are found in both lakes. Three of the species were previously described as C. alpinus Fatio, 1885, C. albellus Fatio, 1890, and C. fatioi Kottelat, 1997. We describe four new species that are endemic to one of the two lakes. Three of them are endemic to Lake Thun, C. steinmanni, C. profundus and C. acrinasus. One is endemic to Lake Brienz, C. brienzii.

One of the previously described species, C. alpinus was designated a lectotype by Kottelat (1997) for which the species description (biology and morphology) in Kottelat (1997) and Kottelat and Freyhof (2007) is incongruent and not that of this species. We show this by tracing back Fatio's description of C. alpinus in his compendium on

Swiss fauna (1890) and comparing it to the lectotype of C. alpinus and to contemporary samples of this species.

One of the newly described species, C. acrinasus, shows ancestry contributions from whitefish of Lake Constance, besides its Lake Thun ancestry (Hudson et al. 2011, 2016; Dönz et al. 2018). We therefore do not only compare it to the five other species from Lake Thun but also to the four described species from Lake Constance, namely C. wartmanni Bloch, 1784, C. macrophthalmus Nüsslin, 1882, C. arenicolus Kottelat, 1997, and the extinct C. gutturosus Gmelin, 1818.

We studied the type material designated by Kottelat (1997) in his systematic revision of the nomenclature of European freshwater fishes for the three described species from lakes Thun and Brienz, C. alpinus, C. albellus, and C. fatioi and the four described species from Lake Constance, C. wartmanni, C. macrophthalmus, C. arenicolus, and the extinct C. gutturosus. Altogether, we compared 240 of our own contemporary samples from lakes Thun and Brienz to these type series.

## Materials and methods

## Study lakes and fish collection

Type material of all currently valid species (based on Kottelat's (1997) systematic revision of the nomenclature of Swiss whitefish) was inspected in the collections of the Natural History Museum of Geneva and Bern (MHNG and NMBE, respecitively), Switzerland and in the Steinmann collection of Eawag, Switzerland, that has recently been transferred together with the Seehausen-Eawag collection to the Natural History Museum of Bern (NMBE), Switzerland. All contemporary specimens are part of the Seehausen-Eawag collection. In some cases, more than one fish is stored in the same jar and thus we provide next to the NMBE number in brackets the individual labels of each fish with Eawag followed by the individual number.

The different whitefish species in this study derive from different lakes, namely Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$, surface area $48 \mathrm{~km}^{2}$, max depth 217 m ), Lake Brienz $\left(46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}\right.$, surface area $30 \mathrm{~km}^{2}$, max depth 261 m ), Lake Biel ( $47^{\circ} 5^{\prime} \mathrm{N}, 7^{\circ} 10^{\prime} \mathrm{E}$, surface area $39.3 \mathrm{~km}^{2}$, max depth 74 m ) and Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$, surface area and max depth of Upper Lake Constance $473 \mathrm{~km}^{2}$ and 251 m and of lower Lake Constance $63 \mathrm{~km}^{2}$ and 46 m depth). Lakes Thun and Brienz are among the deepest and most oligotrophic lakes of the northern pre-alpine region. Lake Constance was historically also among the most deep and oligotrophic lakes of the northern pre-alpine region but is today a mesotrophic lake (Vonlanthen and Périat 2013; Vonlanthen et al. 2015; Alexander et al. 2016). Lakes Thun and Brienz are connected through a short stretch of river (the Bödeli Aare) forming a super-lake system, and used to be part of a much larger postglacial lake, Lake Wendel, before high bed rock load from the river Lütschine separated the basin into the current two lake basins several thousand years ago (Steinmann 1950; Ammann et al. 1991; Hantke and Sc-
heidegger 2007). The species flock of the two lakes, except for $C$. acrinasus, forms a monophyletic group based on independent multilocus microsatellite, large AFLP and whole genome datasets (Douglas et al. 2003; Hudson et al. 2011; Hudson et al. 2016; De-Kayne et al. unpublished). Furthermore, within the super-lake system, the populations of each species from the two sister lakes, Thun and Brienz, are more closely related to each other than the different species from the same lake (i.e., species monophyly), suggesting that the origin of the radiation predates separation of Lake Wendel into lakes Brienz and Thun (Hudson et al. 2011; Hudson et al. 2016; Dönz et al. 2018).

Contemporary samples of whole specimens from lakes Thun and Brienz were collected in the course of many projects of the Seehausen research group (Eawag and the University of Bern). Contemporary material (whole specimens and tissue samples) used here was collected in the years 2005, 2011, 2013, 2014 and 2015 in lakes Thun and Brienz, and in 2016 in Lake Biel. Some of the fish were obtained from commercial fisheries catches. Additionally, in lakes Thun and Brienz fishing was done with monofilament bottom- and pelagic gill nets of various mesh size ranging from 5 to 60 mm , and across many depth ranges in the limnetic and benthic habitats of the lakes (details on net fishing protocols can be found in Alexander et al. (2015)). The fish come from three different sampling methods: targeted fishing on known spawning grounds of the different species at the respective spawning season and water depth (Hudson et al. 2011; Hudson 2011; Vonlanthen et al. 2012), targeted fishing each at one spawning site in lakes Thun and Brienz along a depth gradient four times during the whole spawning season of all species (this study), and habitat-stratified fishing of the whole lake during the summer months (Vonlanthen and Périat 2013; Vonlanthen et al. 2015; Dönz et al. 2018). Additionally, individuals of some species were retrieved from local fisherwomen or fishermen. The sampling locations of all contemporary specimens are plotted on a map in the Suppl. material 1: Figure S10.

## Sample processing

Sampling details for the fish collected in the years 2005, 2011, 2013, and 2014 can be found in the corresponding publications (Bittner 2009; Hudson 2011; Vonlanthen et al. 2012; Dönz et al. 2018). For the fish collected in the year 2015 and 2016 the procedure was as follows: upon capture, fish were anaesthetised and subsequently euthanised with appropriate concentrations of MS222 solutions. Muscle tissue and scales below the dorsal fin, as well as a part of the pectoral fin on the right side of the body, were taken for genetic and isotopic analysis and to determine the age of each fish. The left side of each fish was photographed in two ways: once in water in a custom-made photo cuvette and once on a flat surface with the fins spread. Fish were then fixed in $4 \%$ formalin solution for at least 1 month and afterwards transferred through a series of ethanol of increasing concentration ( $30 \%, 50 \%$ ) to the final concentration of $70 \%$ for storage. Permits for collecting fish in the lakes were issued by the canton of Bern.

In the field the fish were identified to species level as good as possible. Sex, fresh mass (to the nearest 0.1 g ), ripeness ( $4=$ not ripe; $5=$ partially ripe, i.e., slow flow of egg sand sperm when stripped; $6=$ ripe, i.e. eggs and sperm flow easily when stripped) and the presence of tubercles on the scales (modified from Kekäläinen et al. 2015: $0=$ not present; $1=$ small to medium-sized tubercles; $2=$ large tubercles) were noted in some but not all of the field campaigns. Fish which were not ripe, and thus where the sex could not be determined externally, were examined internally by opening the abdominal cavity and inspected for the presence of testis or ovaries.

The age of the specimens that were used in this study was determined in the lab by counting the annual growth rings of four scales under a confocal microscope following Lehtonen and Nylund (1995). If the ages differed between the four scales and three out of four scales did not correspond to the same age, further scales were measured to acquire the same age in $75 \%$ of the scales.

## Morphological and meristic characters

Morphological measurements and counts on the old type material $(\mathrm{N}=31)$ and on contemporary specimens $(\mathrm{N}=340)$ were taken of 25 body, 19 head, and 4 gill characters with adigital calliper to the nearest 0.1 mm . Twelve meristic characters were counted. The measurements and counts were taken on the left body side of the fish, unless a specific character was missing or deformed, in which case that character was measured or counted on the right side of the fish. The mean of two measurements were taken for each character, whereby the difference between two measurements had to be less than $5 \%$. If agreement was less good, the distance was measured again two times. The average inaccuracy between two measurements taken over all morphological characters was $1.4 \%$. Not all measurements could be taken for several specimens since characters where damaged or absent, and we thus sometimes report incomplete character lists for certain specimens. This results in varying sample sizes for each character. All characters for which we had missing values were not retained in the multivariate ratio analyses (see below). The number of characters used for each analysis is explicitly mentioned in the results section. Most of the morphological and meristic characters follow Hubbs and Lagler (1964). However, we also included some additional characters and refined the measurement of some characters found in Hubbs and Lagler (1964). A brief description of each character can be found in Table 1 for the morphological characters and in Table 2 for the meristic characters. Furthermore, illustrations in Suppl. material 1: Figures S1, S2 depict the measurements of the morphological characters. For all morphological characters the mean and for all meristic characters the mode are reported together with the standard deviation and the range for each species, lake population, and sex. Some times if the sample sizes were too small, no mode could be calculated and thus, we report "na". For the four newly described species, the holotype is included in the range. Both sexes are included for the full range of each character of each species from both lakes Thun and Brienz.

Table I. Morphological characters, their acronyms and a brief description of each character.

| Morphological characters | Acronym | Description |
| :---: | :---: | :---: |
| Body |  |  |
| Pelvic fin base | PelvFB | Length between insertions of fin |
| Pelvic fin "spine" length | PelvFS | Length from upper insertion point of fin to tip of spine; the spine is actually an elongated scale structure |
| Pelvic fin length | PelvF | Length from upper insertion point of fin to tip of longest branched ray |
| Pectoral fin base | PecFB | Length between insertions of fin |
| Pectoral fin 1 length | PecF1 | Length from upper insertion point of fin to tip of unbranched ray |
| Pectoral fin 2 length | PecF2 | Length from upper insertion point of fin to tip of longest branched ray |
| Dorsal fin base | DFB | Length between insertions of fin |
| Length of anterior part of dorsal fin erected | DFAe | Length from anterior insertion point of fin to tip of longest unbranched ray, when fin is fully erected |
| Length of anterior part of dorsal fin depressed | DFAd | Length from anterior insertion point of fin to tip of longest unbranched ray, when fin is depressed |
| Length of posterior part of dorsal fin erected | DFPe | Length from posterior insertion point of fin to tip of most posterior branched ray, when fin is erected |
| Anal fin base | AFB | Length between insertions of fin |
| Length of anterior part of the anal fin | AFAe | Length from anterior insertion point of fin to tip of longest branched ray, when fin is fully erected |
| Adipose fin base | AdFB | Length between insertions of fin |
| Caudal fin length | CF | Length from the middle of hypural plate of the caudal fin (internally this is the expanded bones at the end of the backbone that support the caudal fin, externally where the lateral line scales end) to the tip of the longest unbranched ray either being on the dorsal or ventral part of the caudal fin |
| Caudal peduncle depth | CD | Vertical distance between dorsal and ventral margins of the caudal peduncle at its narrowest part |
| Caudal peduncle length | CL | Length from posterior insertion point of anal fin to the middle of the hypural plate of the caudal fin |
| Length from anterior part of adipose fin to caudal fin base | PAdC | Length from anterior insertion point of adipose fin to the middle of the hypural plate of the caudal fin |
| Dorsal head length | DHL | Length from tip of snout to most posterior part of the frontal head bone |
| Prepelvic length | PreP | Length from tip of snout to anterior insertion point of pelvic fin |
| Preanal length | PreA | Length from tip of snout to anterior insertion point of anal fin |
| Standard length | SL | Length from tip of snout to the middle of the hypural plate of the caudal fin |
| Total length | TL | Length from tip of snout to the tip of longest unbranched ray either being on the dorsal or ventral part of the caudal fin |
| Predorsal length | PreD | Length from tip of snout to anterior insertion point of dorsal fin |
| Body depth | BD | Vertical distance between dorsal and ventral margins of body from anterior insertion point of dorsal fin to anterior insertion of pelvic fin: not necessarily the greatest body depth |
| Postdorsal length | PostD | Length from posterior insertion point of dorsal fin to middle of hypural plate of the caudal fin |
| Head |  |  |
| Eye diameter | ED | Horizontal distance across the midline of the eye from the anterior to the posterior margin of the soft eye tissue |
| Eye cavity | EC | Horizontal distance across the midline of the eye from the anterior margin of the eye socket to the posterior margin of the eye cavity |
| Eye height | EH | Vertical distance across the midline of the eye from the dorsal margin of the eye cavity to the ventral margin of the eye cavity |
| Eye socket | ES | Horizontal distance from the anterior margin of the eye socket to the most anterior point of the the posterior margin of the eye socket |
| Postorbital length | PostO | Length from posterior margin of the eye to the most posterior point of the operculum |
| Head length | HL | Length from the tip of snout to most posterior point of the operculum margin |
| Head depth | HD | The transverse distance between margins at the widest point of the head. |
| Head width | HW | Distance between the posterior margins of the left and right operculum |
| Mouth width | MW | The transverse distance between margins of the upper and lower jaw |
| Upper jaw length | UJ | Length from the tip of the snout to most posterior point of the upper jaw |
| Lower jaw length | LJ | Length from the most anterior point of the lower jaw to the lower jaw insertion |
| Lower jaw width | LJW | Length between the anterior left and right side of the lower jaw |
| Uperr jaw width | UJW | Length between the posterior left and right point of the upper jaw |
| Length of maxilla | M | Length from the most anterior point of the maxilla to the most posterior point of the maxilla |


| Morphological characters | Acronym | Description |
| :---: | :---: | :---: |
| Snout length | SN | Length from tip of snout to anterior margin of the eye |
| Snouth depth | SD | Vertical distance from the upper to the lower margin of the rostral plate |
| Snouth width | SW | Horizontal distance from the left to the right margin of the rostral plate |
| Interorbital width | IOW | Distance between the anterior margin of the left and right eye cavity |
| Internarial width | INW | Distance between the right and left nostrils |
| Gill |  |  |
| Upper arch length | UA | Length of the first hypobranchial (upper arch) from the most anterior point to the joint of the hypo- and ceratobranchial where the middle raker emerges |
| Lower arch length | LA | Length of the first ceratobranchial (lower arch) from the most anterior point to the joint of the hypo- and ceratobranchial where the middle raker emerges |
| Middle gill raker length | MGR | Length of the gill raker directly at the joint of the the upper and lower first arch, from the insertion of the gill raker to the tip of the gill raker |
| Longest gill raker length | LGR | Length of the longest gill raker either on the upper and lower first arch, from the insertion of the gill raker to the tip of the gill raker |

Table 2. Meristic characters, their acronyms and a brief description of each character.

| Mersitic characters | Acronym | Description |
| :--- | :---: | :---: | :---: |
| Pelvic fin rays | PelvFR | Number of unbranched and branched rays |
| Pectoral fin rays | PecFR | Number of unbranched and branched rays |
| Dorsal fin rays | AFR | Number of unbranched and branched rays; the posteriormost dorsal rays are often borne from <br> a single pterygiophore (the bones on which the rays articulate), in such a case the two rays are <br> acounted as 2 rays, rudimentary unbranched rays in front of the fin are counted |
| Anal fin rays | Number of unbranched and branched rays; the posteriormost anal rays are often borne from <br> a single pterygiophore (the bones on which the rays articulate), in such a case the two rays are <br> acounted as 2 rays, rudimentary unbranched rays in front of the fin are counted |  |
| Lateral line scales | LS | Scales bearing the lateral-line column canal from the head to the end of the hybpural plate of the <br> caudal peduncle |
| Predorsal scales | TDS | "Dorsal scales starting from the posterior end of the head to the anterior insertion of the dorsal fin |

## Analysis of morphological data

The average sizes of fish from each species differ between lakes enough that for certain species such as for $C$. albellus the average size and the maximum size of adult fish of the population in Lake Brienz do not overlap with the average size and the minimum size of adult fish of the population in Lake Thun (Suppl. material 1: Figures S4-S6). The lakes differ naturally in several abiotic factors (max lake depth, bathymetric slope, average lake temperature, water turbidity; see Alexander et al. 2015) that may be related to the different growth rates of conspecific populations of several species and thus different size-at-age between the lakes (Kirchhofer 1995; Müller et al. 2007). Comparisons with multivariate statistical methods (PCA, LDA see below) are difficult in such cases, when size differences between populations or species are large and there is little to no overlap (Baur et al. 2014). We thus performed separate multivariate ratio analysis (see
below for details) on the species from lakes Thun and Brienz. In the Lake Thun dataset the partially allochthonous species $C$. acrinasus was not included in the comparison between the types of the previously described species of C. alpinus, C. albellus, and C. fatioi (type locality Lake Thun) and the contemporary specimens, because the introduction of whitefish from Lake Constance, from which C. acrinasus shares genetic ancestry contributions, postdates the collection year of the types. Furthermore, in Lake Brienz we divided the data into two subsets to avoid allometry issues; a subset containing individuals smaller than 163.5 mm SL and one subset containing individuals larger than 163.5 mm SL. This threshold was chosen to retain several small individuals of the three larger whitefish species, C. alpinus, $C$. brienzii, and $C$. fatioi for the analysis with all individuals of the small whitefish species C. albellus. All four species of Lake Brienz are represented in both subsets albeit unequally distributed.

Multivariate ratio analysis is a method that performs principal component analysis (PCA) and linear discriminant analysis (LDA) on morphological ratios (Baur and Leuenberger 2011; Baur et al. 2014). Analysis of morphological ratios are especially well suited in a taxonomic context (László et al. 2013). A scree plot was used to identify the number of PC-axes that should be retained and plotted. In most cases the first two PC axes were retained, and, in a few cases, the third PC axis was also retained. We thus use the first three axes to visualize shape variation between the species. The eigenvalues of the PC-axes and the loadings of each trait can be found in the Suppl. material 1: Tables S1-S4. We further plotted the scores of each PC-axis against isosize to investigate the contribution of allometry to individual shape PC-axes. Isosize is an isometric size axis defined as the geometric mean of all characters used in the PCA. We report the linear regression coefficient $R$ as a metric of the contribution of allometry to each PC-axes (Baur and Leuenberger 2011) (Suppl. material 1: Tables S1-S4). If the relationship between size and shape is strong, then such PC-axes are not informative to distinguish species based on shape itself.

For the development of a species identification key we used LDA analysis on all characters together and on subsets of only head or only body characters for all contemporary specimens from lakes Thun and Brienz separately to calculate the first two ratios of characters that best separate each of the species in each lake. This method also allows to estimate the extent of shape change with size (i.e., the contribution of allometry to these ratios) which is given as $\delta$ and describes how good shape discriminates in comparison to size (see Baur and Leuenberger 2011: Page 818, formula 14). In several pairwise species comparisons, we had more variables than individuals which will not allow to calculate the best LDA ratios. In such cases we used a subset of the variables to match the number of individuals. The variables that were retained in this subset were chosen such that possibly informative characters in each pairwise comparison were kept. All the comparisons with a subset of characters are marked in the table and the respective characters that were excluded are listed (Tables 10, 11). Due to large size differences between the species of Lake Brienz the LDA ratios were calculated with three different datasets; once each with individuals larger or smaller than 163.5 mm SL and once with the full-size ranges of all species. Ratios marked in the table with an asterisk $\left(^{*}\right.$ ) have very little or no
overlap with other species and were thus used in the identification key and the species diagnoses. All analyses were performed in RStudio v1.0.143 (R Studio Team 2015).

## Genetics

Genetic analysis of ten microsatellite loci were used for the Bayesian clustering algorithm program STRUCTURE (Pritchard et al. 2000) to assign all contemporary specimens of lakes Thun and Brienz to the different whitefish species present in either lake. DNA was extracted from fin tissue using Chelex and Proteinase-K following the manufacturer's standard protocol. All individuals were genotyped at ten microsatellite loci that were combined into two multiplex sets: $\mathrm{CoCl} 49, \mathrm{CoCl68}, \mathrm{CoCl6}, \mathrm{C} 2-157$, $\mathrm{CoCl61}, \mathrm{CoCl} 45$ and BWF-2, CoCl4, CoCl18, CoCl10 (Patton et al. 1997; Turgeon et al. 1999; Rogers et al. 2004). DNA fragments were resolved on an automated DNA sequencer (ABI 3130xl) and genotypes were determined with the software Gene Mapper (ver. 4.0) with the same scoring-panel as in Dönz et al. (2018). Individuals that had missing data at more than two loci were excluded from further analysis.

From the targeted spawning fisheries (each at one spawning site in lakes Thun and Brienz along a depth gradient) a total of 663 individuals from Lake Thun had complete genotypes, ten individuals had one missing locus, and four individuals had two missing loci. A total of 284 individuals from Lake Brienz had complete genotypes, eighteen individuals had one missing locus, and four individuals had two missing loci. These individuals were assigned to the different species using the program STRUCTURE with reference populations of each species deriving from the study by Dönz et al. (2018). A detailed description of the assignment procedure can be found in Dönz et al. (2018). In brief, Dönz et al. (2018) had a dataset comprising 2388 fish from both lakes with the same set of ten microsatellite loci and the same scoring-panel as in our study. To find the most likely number of genetic clusters (K), they conducted a hierarchical cluster analysis (Coulon et al. 2008; Roy et al. 2015) using the individual-based Bayesian clustering algorithm implemented in STRUCTURE (Pritchard et al. 2000). They determined the most likely K for the full dataset of 2388 individuals, then the most likely K within each of the data subsets suggested by the previous analysis, and so forth until all subsets supported a value of $\mathrm{K}=1$. To determine correspondence of genetic clusters to known species, they assessed how individuals from targeted samplings of known species were distributed among the clusters. They then chose the 50 individuals with highest assignment likelihood to the corresponding clusters at each previous step in the hierarchical analysis and designated them as a reference panel for the six clusters. Afterwards this method can be used to obtain individual genetic assignment proportions to the six clusters inferred in the hierarchical analysis.

We used the reference panel from Dönz et al. (2018) as reference populations and assigned all the individuals from our data set to the six species clusters with the function PopFlag in STRUCTURE. Subsets of 50 individuals out of the 973 genotyped individuals (from the depth gradient data set) were run in separate assignment runs to
avoid issues with unequal sample sizes. For each of these analyses, we performed 10 replicates of $\mathrm{K}=6$ with $200^{\prime} 000$ burn-in steps and $200^{\prime} 000 \mathrm{MCMC}$ steps using the admixture and correlated allele frequency model. We used Structure Harvester to generate input files for CLUMPP (ver. 1.1.2, Jakobsson and Rosenberg 2007), which we used to generate consensus percentages of assignment proportions from the 10 structure runs. We first retained all individuals with assignment probabilities higher than $70 \%$ to one cluster and chose among these individual's specimens for the taxonomic work (Dönz et al. 2018). This resulted in a total of 244 out of $677(36 \%)$ individuals for Lake Thun and a total of 147 out of 296 ( $50 \%$ ) individuals for Lake Brienz. We aimed at obtaining for each lake a total of ca. 20 specimens of each species for the taxonomic work. In cases where this number was not reached with specimens that had assignment probabilities higher than $70 \%$, we supplemented the data set with individuals with lower assignment probabilities and checked if they cluster in morphospace with the respective specimens with higher assignment probabilities. A few additional specimens were taken from previous sampling campaigns, which were assigned by Dönz et al. (2018). We also took into consideration - next to the genetic species assignment - information regarding catch date and depth (reflecting spawning season and habitat). The percentage of assignment proportions for each contemporary specimen can be found together with all the other data underpinning the analyses reported in this paper in the online Dryad Data Repository (http://doi.org/10.5061/dryad.pd2tq5g).

Newer genomic findings by De-Kayne et al. (unpublished) suggest that whitefish from Lake Brienz, that have previously been assigned based on genetic analysis (see above; Dönz et al. 2018) to C. steinmanni and have also been selected based on these assignments for our taxonomic work, actually comprise of an endemic species in Lake Brienz, C. brienzii. The assignment probability to C. steinmanni of specimens of C. brienzii, which is based on the assignment method by Dönz et al. (2018), are for the sake of completeness still reported in the online dataset.

## Results

## Summary

The principal component analyses (shapePCA) on the morphological characters show that the type specimens of all previously described species C. alpinus, C. albellus, and C. fatioi group in morphospace within the ranges or adjacent to the ranges of the respective contemporary specimens of these species in Lake Thun (Figure 1a, b; Suppl. material 1: Figure S11a, b). The types of each of the three previously described species further mostly overlap within the ranges of each of the contemporary specimens of the three species from Lake Thun (Tables 3-5). We thus use the contemporary specimens of the previously described species C. alpinus, C. albellus, and C. fatioi, together with the holo- and paratypes of the newly described species $C$. steinmanni, C. brienzii, C. profundus, and C. acrinasus to delineate the species. The shape PCA on


Figure I. Principal Component Analysis showing that the types of the previously described species C. alpinus, C. albellus and C. fatioi (type locality: Lake Thun) lie within or adjacent to the ranges of the contemporary species of Lake Thun A, B shape PCA of the first vs. the second or third PC-axes explain together $70.05 \%$ of the variation in shape and are based on a subset (Suppl. material 1: Table S1) of 30 out of a total of 48 measured linear morphological characters (Table 1), since the type material lacked certain characters. Name-bearing types of the formerly described species are highlighted with enlarged symbols in the plots. The proportion of variance explained by each shape PC is given in brackets in the axis legend. PC-loadings and amount of shape variation explained by size are reported in Suppl. material 1: Table S1.


Figure 2. Principal Component Analysis showing the morphospace of the contemporary whitefish species C. acrinasus, C. alpinus, C. steinmanni, C. albellus, C. profundus and C. fatioi from Lake Thun A, $\mathbf{B}$ shape PCA of the first vs. the second or third PC-axes explain together $56.5 \%$ of the variation in shape and are based on all 48 measured linear morphological characters (Table 1). The proportion of variance explained by each shape PC is given in brackets and the PC-loadings and amount of shape variation explained by size in Suppl. material 1: Table S2.
the contemporary specimens of Lake Thun shows three main clusters, one containing the species C. alpinus and C. steinmanni, one containing C. albellus and C. fatioi, and a third cluster with C. profundus, while C. acrinasus lies intermediate between these three


Figure 3. Principal Component Analysis showing the morphospace of the contemporary whitefish species C. alpinus, C. brienzii, C. albellus and C. fatioi from Lake Brienz (A-D) A, B shape PCA of the first vs. the second or third PC-axes explain together 53-60.7\% of the variation in shape and are based on all 48 measured linear morphological characters (Table 1), with a dataset containing specimens once smaller (A, B) and once larger ( $\mathbf{C}, \mathbf{D}$ ) than 163.5 mm standard length (SL) to avoid allometry issues. The proportion of variance explained by each shape PC is given in brackets and the PC-loadings and amount of shape variation explained by size in Suppl. material 1: Table S3 ( $<163.5 \mathrm{~mm}$ ) and Suppl. material 1: Table S4 (> 163.5 mm ).
clusters (Figure 2a, b). The shape PCA on the contemporary specimens of $C$. alpinus, C. albellus, C. fatioi, and C. brienzii of Lake Brienz reveals three clusters, one cluster containing C. alpinus, one containing C. fatioi and C. brienzii, and the third cluster containing C. albellus (Figure 3a-d). Some species cluster together in morphospace: In Lake Thun C. steinmanni groups with C. alpinus and C. fatioi with C. albellus and in Lake Brienz C. brienzii groups with C. fatioi. The species can be delineated further by several morphological and meristic characters as well as by morphological ratios extracted from linear discriminant analyses (Tables 3-11). Coregonus acrinasus can also be distinguished from all four described Lake Constance whitefish species C. wartmanni, C.macrophthalmus, C. arenicolus, and C. gutturosus (Tables 9, 12).

## Species descriptions

## Coregonus albellus Fatio, 1890

## Figure 4

Coregonus exiguus albellus: Fatio 1890
Coregonus "Brienzlig": Surbeck 1917; Steinmann 1950, Rufli 1978, 1979; Kirchhofer and Tschumi 1986; Kirchhofer 1995; Bittner et al. 2010
Coregonus "Brienzlig", "Winterbrienzlig": Kirchhofer 1990; Kirchhofer 1995
Coregonus "Small type": Maurer and Guthruf 2005; Müller et al. 2007
Coregonus sp. "winter spawning": Kottelat and Freyhof 2007
Coregonus "Sommerbrienzlig", "BRI2": Douglas et al. 1999; Douglas and Brunner 2002
Coregonus "Sommerbrienzlig", "THU5": Douglas et al. 2003
Coregonus "Sommerbrienzlig", "Winterbrienzlig", "THU4", "THU5": Douglas and Brunner 2002
Coregonus sp. "Brienzlig": Vonlanthen and Périat 2013
Coregonus "Kropfer": Heuscher 1901 (see also synonymy of C. profundus)
Coregonus lavaretus natio arurensis, oekot. nanus: Steinmann 1950
Coregonus "Zwergalbock": Steinmann 1950

Material examined. Lectotype. MHNG-816.022, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}$, $7^{\circ} 46^{\prime} \mathrm{E}$ ), 165 mm SL, sex unknown.

Non-types. NMBE-1077186-1077202, NMBE-1077221-1077237, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=34,177-271 \mathrm{~mm} \mathrm{SL}$; NMBE-1059754; 1059768; 1059791; 1059801; 1059814, NMBE-1077129-1077131, NMBE-10773181077341, Switzerland, Lake Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ), $\mathrm{N}=32,101-164 \mathrm{~mm}$ SL.

Diagnosis. Coregonus albellus is a very small whitefish species in Lake Brienz and a small whitefish species in Lake Thun with weak pigmentation of all fins and body; the colouration on the flanks above the lateral line of specimens from Lake Thun are pale rose to brown and from Lake Brienz pale brown to light green; no or few small pigmented dots on the edge of the scales along the flank for specimens from Lake Thun and specimens from Lake Brienz sometimes have rather large pigmented dots more or less in a row on the upper dorsum; elongate slender body; large eye with a thin and roundish eye socket; tip of snout fleshy and roundish; many and long gill rakers.

Differential diagnosis. No single character was sufficient to distinguish C. albellus against all the five other species from Lake Thun and the species is diagnosed by a combination of characters. Based on ratios for the subset of whitefish from Lake Brienz smaller than $163.5 \mathrm{~mm}, C$. albellus can be distinguished from the other three species from Lake Brienz by a smaller "postdorsal length / eye height" ratio (PostD/EH: 5.47-6.93 vs. 7.5-8.9). Also, when taking the full-size range (100-290 mm) of all species from Lake Brienz C. albellus can be distinguished from the three other species by a smaller "predorsal length / eye height" ratio (PreD/EH: 6.1-7.58 vs. 8.12-10.5) (Table 11).


Figure 4. Coregonus albellus, lakes Thun and Brienz, Switzerland A lectotype, MHNG-816.022, Lake Thun, 165 mm SL, sex unknown B non-type, Eawag-123825, Lake Thun, 221 mm SL, male $\mathbf{C}$ nontype, NMBE-1077320, Lake Brienz, 115.5 mm SL, male. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.

## Coregonus albellus-Coregonus alpinus

The specimens from lakes Thun and Brienz of C. albellus differ from those of C. alpinus of both lakes in having a higher number of gill rakers (UGR\#: 9-17, mode $=13$ vs. $8-11$, mode $=10$; LGR\#: $20-29$, mode $=25$ vs. $15-23$, mode $=19$; total GR: $32-44$, mode $=38$ vs. $25-34$, mode $=29$ ), a longer longest gill raker $(14.1-21.8 \%$ HL, mean $=17.7$ vs. $10-15.2 \% \mathrm{HL}$, mean = 11.9), a deeper adipose fin (4.5-9.2\% SL, mean $=6.5$ vs. $3.4-5.5 \% \mathrm{HL}$, mean $=4.4)$, a longer lower jaw $(38.4-49.2 \% \mathrm{HL}$, mean $=43.6$ vs. $33.8-41.4 \% \mathrm{HL}$, mean $=38.4)$ and a thinner eye socket $(2-4.9 \%$ HL, mean $=3.4$ vs. $3.3-6.3 \% \mathrm{HL}$, mean $=5$ ).
Table 3. Morphological and meristic data of C. albellus Fatio, 1890 from lakes Thun and Brienz, MHNG-816.022 lectotype from Lake Thun; non-type material
$\mathrm{N}=34$ from Lake Thun and $\mathrm{N}=32$ from Lake Brienz.

| Morphological/ characters | $\begin{array}{\|l\|} \hline \text { C. albellus } \\ \hline \text { Lectotype } \end{array}$ | Both lakes |  | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-types both sexes |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  | $N$-total $=66$ |  | $N$-total $=34$ |  | $N$-females $=21$ |  | $N$-males $=13$ |  | $N$-total $=32$ |  | N-females $=19$ |  | N -males $=13$ |  |
|  |  | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\text { Mean } \pm$ StDev | Range |
| SL (mm) | 165.0 | $173.2 \pm 51.9$ | (101-271) | $221.4 \pm 16.4$ | (177-271) | $224.4 \pm 14.9$ | (205-271) | $216.6 \pm 18.1$ | (177-254) | $122.1 \pm 11.0$ | (101-164) | $122.9 \pm 13.5$ | (101-164) | $120.9 \pm 6.4$ | (108-129) |


| $(3.2-5.6)$ | $4.0 \pm 0.5$ | $(3.2-5.6)$ | $3.6 \pm 0.2$ | $(3.3-4.0)$ |
| :--- | :--- | :--- | :--- | :--- |


| $(5.5-7.8)$ |
| :---: |
| $(15.3-18.1)$ |
| $(2.9-3.6)$ |

 $(18.1-21.2)$
$(10.7-13.2)$ (17.3-20.8)




 (19.6-21.8) $(16.1-19.7)$
$(48.4-54.0)$ $(74.0-78.7)$
$(46-51.3)$ (16.9-22.6)

 | $122.1 \pm 2.3$ | $(119.6-130)$ | $121.7 \pm 2.2$ | $(118.2-125.5)$ |
| :--- | :--- | :--- | :--- |




| Morphological/ characters | $\begin{array}{\|l\|} \hline \text { C. albellus } \\ \hline \text { Lectotype } \\ \hline \end{array}$ | Both lakesNon-types both sexesN-total $=\mathbf{6 6}$ |  | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  |  |  | $N$-total $=34$ |  | $N$-females $=21$ |  | $N$-males $=13$ |  | N -total $=32$ |  | N -females $=19$ |  | N-males $=13$ |  |
|  |  | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ StDev | Range | Mean $\pm$ <br> StDev | Range | $\text { Mean } \pm$ StDev | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ StDev | Range |
| EC | 27 | $29.9 \pm 3.7$ | (23.1-36.8) | $26.6 \pm 1.4$ | (23.1-29.2) | $26.9 \pm 1.4$ | (24-29.2) | $26.2 \pm 1.4$ | (23.1-28.1) | $33.3 \pm 1.6$ | (30.4-36.8) | $33.2 \pm 1.9$ | (30.4-36.8) | $33.5 \pm 1.2$ | (31.5-36.2) |
| EH | na | $26.1 \pm 2.9$ | (20.8-30.6) | $23.5 \pm 1.3$ | (20.8-26.2) | $23.7 \pm 1.4$ | (20.8-26.2) | $23.4 \pm 1$ | (21.4-25.1) | $28.7 \pm 1.1$ | (26.5-30.6) | $28.7 \pm 1.2$ | (26.5-30.6) | $28.9 \pm 0.9$ | (27.1-30.4) |
| ES | na | $3.4 \pm 0.7$ | (2-4.9) | $3.2 \pm 0.8$ | (2.0-4.9) | $3.1 \pm 0.8$ | (2.0-4.5) | $3.3 \pm 1.0$ | (2.1-4.9) | $3.5 \pm 0.5$ | (2.5-4.8) | $3.5 \pm 0.5$ | (2.9-4.8) | $3.5 \pm 0.5$ | (2.5-4.3) |
| PostO | 49.8 | $50.2 \pm 2.5$ | (45.5-55.6) | $52 \pm 1.5$ | (48.5-55.6) | $51.8 \pm 1.6$ | (48.5-55.6) | $52.3 \pm 1.4$ | (50.1-55.1) | $48.2 \pm 1.8$ | (45.5-53.1) | $48.3 \pm 2$ | (45.5-53.1) | $48.2 \pm 1.4$ | (45.7-50.5) |
| HD | 75.5 | $68.3 \pm 4.5$ | (59.2-80.3) | $71.8 \pm 2.9$ | (66.9-80.3) | $71.5 \pm 3.2$ | (66.9-80.3) | $72.2 \pm 2.4$ | (67.1-75.8) | $64.6 \pm 2.3$ | (59.2-69) | $64.4 \pm 2.4$ | (59.2-68.8) | $64.8 \pm 2.3$ | (61.3-69) |
| MW | na | $10.4 \pm 0.8$ | (8.8-12.2) | $10.3 \pm 0.7$ | (8.8-12) | $10 \pm 0.6$ | (8.8-11.2) | $10.8 \pm 0.7$ | (9.4-12) | $10.5 \pm 0.8$ | (8.8-12.2) | $10.5 \pm 0.8$ | (8.9-12.2) | $10.5 \pm 0.8$ | (8.8-11.7) |
| UJ | na | $31.7 \pm 1.5$ | (28.6-34.9) | $31.2 \pm 1.6$ | (28.8-34.7) | $30.9 \pm 1.5$ | (28.8-33.9) | $31.8 \pm 1.7$ | (29-34.7) | $32.1 \pm 1.3$ | (28.6-34.9) | $31.8 \pm 1.1$ | (28.6-33.4) | $32.6 \pm 1.3$ | (30.3-34.9) |
| LJ | 45.5 | $43.6 \pm 2.8$ | (38.4-49.2) | $41.4 \pm 1.6$ | (38.4-44.6) | $41.2 \pm 1.7$ | (38.6-44.4) | $41.6 \pm 1.6$ | (38.4-44.6) | $45.9 \pm 1.6$ | (41.9-49.2) | $45.6 \pm 1.7$ | (41.9-48.5) | $46.4 \pm 1.5$ | (43.9-49.2) |
| M | na | $23.5 \pm 1.7$ | (20.1-26.9) | $22.4 \pm 1.4$ | (20.1-26.8) | $22.1 \pm 1.4$ | (20.1-26.8) | $22.8 \pm 1.3$ | (21.5-26.4) | $24.7 \pm 1.1$ | (22.6-26.9) | $24.4 \pm 1.1$ | (22.6-26.5) | $25.1 \pm 1.2$ | (23.4-26.9) |
| SD | na | $8.8 \pm 1.6$ | (5.6-13.1) | $9.6 \pm 1.6$ | (6-13.1) | $9.2 \pm 1.4$ | (6-12.7) | $10.3 \pm 1.7$ | (7.4-13.1) | $8 \pm 1$ | (5.6-10.4) | $8.2 \pm 1.1$ | (5.6-10.4) | $7.7 \pm 0.6$ | (6.4-8.6) |
| SW | na | $17.9 \pm 1.5$ | (13.5-21.1) | $17.4 \pm 1.7$ | (13.5-20) | $16.9 \pm 1.5$ | (13.5-19.1) | $18.1 \pm 1.6$ | (14.3-20) | $18.4 \pm 1.2$ | (15.4-21.1) | $18.4 \pm 1.3$ | (15.4-20.6) | $18.4 \pm 1.2$ | (16.3-21.1) |
| HW | 48.1 | $48.5 \pm 4.3$ | (37.5-56.1) | $50.7 \pm 3.3$ | (43.5-56.1) | $50.6 \pm 3.2$ | (45.1-56.1) | $50.7 \pm 3.6$ | (43.5-55.1) | $46.2 \pm 4.1$ | (37.5-54.7) | $45.7 \pm 3.7$ | (37.5-51.7) | $46.9 \pm 4.5$ | (41.3-54.7) |
| IOW | 25.8 | $26.9 \pm 2.2$ | (22.7-33.6) | $28.4 \pm 2$ | (24.4-33.6) | $28.4 \pm 2$ | (24.8-33.6) | $28.4 \pm 2.1$ | (24.4-31.9) | $25.4 \pm 1.2$ | (22.7-27.5) | $25.4 \pm 1.3$ | (23.1-27.5) | $25.5 \pm 1.2$ | (22.7-27.4) |
| INW | na | $11.5 \pm 1.3$ | (7.8-15.4) | $11.9 \pm 1.3$ | (9.9-15.4) | $11.8 \pm 1.4$ | (9.9-15.4) | $12.1 \pm 1.3$ | (9.9-15) | $11 \pm 1.2$ | (7.8-13.7) | $11.1 \pm 1.1$ | (9-13.7) | $11 \pm 1.3$ | (7.8-13) |
| LJW | 12.2 | $11.7 \pm 2.4$ | (7.5-17.5) | $13.3 \pm 2.2$ | (8.6-17.5) | $13.1 \pm 2$ | (9-16.5) | $13.5 \pm 2.4$ | (8.6-17.5) | $10 \pm 1.3$ | (7.5-12.3) | $9.9 \pm 1.2$ | (8.4-12.3) | $10 \pm 1.5$ | (7.5-12.2) |
| UJW | na | $24.5 \pm 2.2$ | (19.1-28.9) | $25.4 \pm 1.9$ | (20.9-28.7) | $24.9 \pm 1.9$ | (20.9-28.7) | $26.2 \pm 1.8$ | (22.7-28.4) | $23.5 \pm 2$ | (19.1-28.9) | $23.4 \pm 2.1$ | (19.1-28.9) | $23.6 \pm 1.9$ | (19.8-27.4) |
| MGR | 16.1 | $16 \pm 1.5$ | (11.7-19.4) | $15.6 \pm 1.7$ | (11.7-18.3) | $15.7 \pm 1.6$ | (11.7-18.3) | $15.5 \pm 1.8$ | (12-18.2) | $16.5 \pm 1.2$ | (13.7-19.4) | $16.4 \pm 1.3$ | (13.7-19.3) | $16.6 \pm 1.2$ | (15.1-19.4) |
| LGR | 17 | $17.7 \pm 1.6$ | (14.1-21.8) | $17.2 \pm 1.5$ | (14.1-20.3) | $17.3 \pm 1.6$ | (14.1-20.3) | $17 \pm 1.4$ | (14.7-18.9) | $18.2 \pm 1.5$ | (14.9-21.8) | $18 \pm 1.6$ | (14.9-21.8) | $18.6 \pm 1.2$ | (16.8-21.5) |
| UA | 20 | $19.5 \pm 1.4$ | (14.8-22.6) | $19.2 \pm 1.4$ | (14.8-22.6) | $19.1 \pm 1.6$ | (14.8-22.6) | $19.5 \pm 1.2$ | (17.5-21.3) | $19.7 \pm 1.4$ | (17.2-22.1) | $19.5 \pm 1.5$ | (17.2-22.1) | $20 \pm 1.1$ | (17.8-21.3) |
| LA | 37.8 | $36.4 \pm 2.7$ | (29.6-42.2) | $35.4 \pm 1.7$ | (32.3-39.6) | $35.4 \pm 1.9$ | (32.3-39.6) | $35.4 \pm 1.4$ | (32.8-37.5) | $37.5 \pm 3.1$ | (29.6-42.2) | $36.8 \pm 3.6$ | (29.6-42.2) | $38.7 \pm 1.9$ | (35.3-42) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 11 | 10 | (9-12) | 10 | (9-11) | 10 | (9-11) | 10 | (9-11) | 10 | (10-12) | 10 | (10-11) | 10 | (10-12) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 16 | 16 | (14-17) | 16 | (14-17) | 16 | (14-17) | 16 | (15-17) | 16 | (15-17) | 16 | (15-17) | 16 | (15-17) |
| DF unbranched | 3 | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) |
| DF branched | 11 | 10 | (9-12) | 10 | (9-12) | 10 | (9-12) | 10 | (9-12) | 10 | (9-12) | 10 | (9-12) | 10 | (9-11) |


| Meristic characters | C. albellus <br> Lectotype | Both lakesNon-types both sexes$N$-total $=66$ |  | Lake Thun Non-types |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | $N$-total $=32$ |  | Non-typesN-females $=19$ |  | $N-m a l e s=13$ |  |
|  |  |  |  | $N$-total $=34$ | $N$-females $=21$ |  | $N$-males $=13$ |  |
|  |  | Mode | Range |  |  | Mode | Range |  |  | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range |
| AF unbranched | 3 | 3 | (2-4) | 3 | (2-4) |  |  | 3 | (2-4) | 3 | (2-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) |
| $\begin{aligned} & \hline \text { AF } \\ & \text { branched } \end{aligned}$ | 14 | 12 | (10-14) | 12 | (10-13) | 12 | (10-13) | 12 | (11-13) | 12 | (10-14) | 12 | (11-14) | 12 | (10-14) |
| LS | 75 | 79 | (70-86) | 79 | (73-86) | 79 | (73-83) | 77 | (76-86) | 76 | (70-84) | 77 | (70-80) | 78 | (70-84) |
| PDS | na | 31 | (26-37) | 34 | (29-37) | 34 | (29-37) | 34 | (30-37) | 31 | (26-35) | 32 | (30-35) | 31 | (26-34) |
| TDS | 9 | 9 | (7-10) | 9 | (8-10) | 9 | (8-10) | 9 | (9-10) | 8 | (7-9) | 8 | (7-9) | 8 | (7-9) |
| TAS | 7 | 8 | (6-9) | 8 | (7-9) | 8 | (7-9) | 8 | (8-9) | 7 | (6-8) | 7 | (6-8) | 8 | (6-8) |
| TPS | 8 | 9 | (7-9) | 9 | (8-9) | 9 | (8-9) | 9 | (8-9) | 8 | (7-9) | 8 | (7-9) | 8 | (7-8) |
| UGR | 11 | 13 | (9-17) | 13 | (9-17) | 13 | (9-15) | 14 | (11-17) | 13 | (11-16) | 13 | (11-16) | 13 | (12-14) |
| LGR | 23 | 25 | (20-29) | 24 | (20-28) | 24 | (20-28) | 26 | (22-27) | 25 | (22-29) | 25 | (22-29) | 27 | (24-28) |
| total GR | 34 | 38 | (32-44) | 38 | (32-44) | 37 | (32-42) | 38 | (33-44) | 40 | (35-42) | 38 | (35-42) | 40 | (37-42) |

In Lake Brienz C. albellus further differs from C. alpinus by having translucent pelvic and anal fins compared to the moderately to strongly pigmented pelvic and anal fins of C. alpinus, a longer pectoral fin (Pectoral fin 1 length: 15.7-22.6\% SL, mean $=$ 18 vs. $13.9-17.9 \%$ SL, mean $=16.3$; Pectoral fin 2 length: 16.9-23.8\% SL, mean $=$ 19.4 vs. $14.4-17.7 \%$ SL, mean $=16.9$ ), a longer distance from the anal fin to the hypural plate of the caudal peduncle (17.7-24.2\% SL, mean $=20.7$ vs. $15.3-19.5 \%$ SL, mean $=17.6)$, a longer head $(16.1-23.1 \% \mathrm{SL}$, mean $=17.9$ vs. $14-16.3 \% \mathrm{SL}$, mean $=$ 15.4 ), a larger eye and eye cavity (eye diameter: $26.1-32 \% \mathrm{HL}$, mean $=29.3$ vs. $21.8-$ $27.2 \% \mathrm{HL}$, mean $=24.3$; eye height: $26.5-30.6 \% \mathrm{HL}$, mean $=28.7$ vs. $22.4-27.1 \%$ HL, mean $=23.9$; eye cavity: $30.4-36.8 \% \mathrm{HL}$, mean $=33.3$ vs. $26.4-31.5 \% \mathrm{HL}$, mean $=29$ ), and a longer upper jaw ( $28.6-34.9 \% \mathrm{HL}$, mean $=32.1$ vs. $25.4-29.1 \%$ HL, mean $=26.8$ ). Finally, C. albellus smaller than 163.5 mm SL can be distinguished from C. alpinus by a smaller "preanal length / lower jaw" ratio (PreA/LJ: 6.33-7.44 vs. 9.24-9.97) and a larger "pectoral fin 2 length / length of the depressed anterior part of the dorsal fin" ratio (PecF2/DFAd: $0.81-1.06$ vs. $0.78-0.8$ ). With the full size range of Lake Brienz specimens, C. albellus can be distinguished from C. alpinus by the smaller "predorsal length / lower jaw" ratio (PreD/LJ: 3.99-4.68 vs. 5.6-6.81), "erected anterior part of the dorsal fin / upper jaw" ratio (DFAe/UJ: 2.14-2.79 vs. 3.25-4.1), "head depth / upper jaw" (HD/UJ: 1.87-2.2 vs. 2.38-2.78) and a larger "lower jaw / interorbital width" ratio (LJ/IOW: 1.53-1.99 vs. 1.33-1.57). (Tables 3, 4, 11).

In Lake Thun C. albellus can further be distinguished from C. alpinus by having a less deep caudal peduncle ( $6.4-7.9 \% \mathrm{SL}$, mean $=7.1$ vs. $7.6-8.9 \%$ SL, mean $=8.2$ ) and a longer upper jaw ( $28.8-34.7 \% \mathrm{HL}$, mean $=31.2$ vs. $24.3-30.1 \% \mathrm{HL}$, mean $=$ 27.7). Based on pigmentation of the fins $C$. albellus can be distinguished from $C$. alpinus from Lake Thun by having translucent to weakly pigmented fins compared to strongly pigmented fins, respectively. In Lake Thun C. albellus can further be distinguished from C. alpinus by the smaller "caudal peduncle depth / upper jaw length" ratio (CD/UJ: $0.96-1.29$ vs. $1.36-1.65$ ) and "caudal peduncle depth / dorsal head length" ratio (CD/DHL: $0.44-0.54$ vs. $0.54-0.62$ ) (Tables 3, 4, 10).

## Coregonus albellus-Coregonus fatioi

In Lake Brienz C. albellus can be distinguished from C. fatioi by having a larger head $(16.1-23 \%$ SL, mean $=17.9$, vs. $14.5-16.8 \%$ SL, mean $=15.7)$, a larger eye and eye cavity (eye diameter: $26.1-32 \% \mathrm{HL}$, mean $=29.4$ vs. $21.2-27.6 \% \mathrm{HL}$, mean $=$ 24.8; eye cavity: $30.4-36.8 \% \mathrm{HL}$, mean $=33.3$ vs. $25.3-33 \% \mathrm{HL}$, mean $=29$; eye height: $26.5-30.6 \% \mathrm{HL}$, mean $=28.7$ vs. $22.1-26.3 \% \mathrm{HL}$, mean $=24.4$ ), a longer maxilla ( $22.6-26.9 \% \mathrm{HL}$, mean $=24.7$ vs. $18.7-24.2 \% \mathrm{HL}$, mean $=21.7$ ) and longer gill rakers (middle gill raker: 13.7-19.4\% HL, mean $=16.5$ vs. $10.5-15 \% \mathrm{HL}$, mean = 13.2; longest gill raker: $14.9-21.8 \% \mathrm{HL}$, mean $=18.2$ vs. $12.3-16.4 \% \mathrm{HL}$, mean $=$ 14.3). Based on ratios C. albellus smaller than 163.5 mm SL can be distinguished from C. fatioi by a larger "pectoral fin 2 length / preanal length" ratio (PecF2/PreA: 0.220.28 vs. $0.2-0.22$ ), "upper jaw length / eye socket width" ratio (UJ/ES: 6.81-12.42 vs. 4.51-6.15) and "eye socket width / head length" ratio (ES/HL: 0.27-0.31 vs. $0.23-$ 0.27). With the full-size range of Lake Brienz specimens (100-290 mm), C. albellus
can be distinguished from C. fatioi by a smaller "prepelvic length / eye height" ratio (PreP/EH: 6.56-7.98 vs. 8.94-11.43) (Tables 3, 5, 11).

In Lake Thun C. albellus can be distinguished from C. fatioi by its live colouration above the lateral line on the dorsum ranging from a pale rose to a pale brown compared to a light to dark green colouration in C. fatioi. C. albellus can further be differentiated from $C$. fatioi by having no or few small pigmented dots on the edge of the scales or on the boundary of two scales on the flank and dorsum compared to moderate or many dots on the flanks and dorsum in C. fatioi.

## Coregonus albellus-Coregonus brienzii

Coregonus albellus from Lake Brienz differs from C. brienzii by having a longer longest gill raker ( $14.9-21.8 \% \mathrm{HL}$, mean $=18.2$ vs. $12.1-16.8 \% \mathrm{HL}$, mean $=14.7$ ), a longer maxilla ( $22.6-26.9 \% \mathrm{HL}$, mean $=24.7$ vs. $15.4-24 \% \mathrm{HL}$, mean $=21$ ), anterior a longer dorsal fin (anterior dorsal fin erected: 17.3-24.7\% SL, mean = 19.7 vs. 15.5-19.8\% SL, mean = 17.9; anterior dorsal fin depressed: 18.3-26.6\% SL, mean $=20.6$ vs. $15.3-20.8 \%$ SL, mean $=18.6)$, a longer head $(16.1-23.1 \%$ SL, mean $=$ 17.9 vs. $14.6-16.8 \%$ SL, mean $=15.6$ ) and a larger eye and eye cavity (eye diameter: $26.1-32 \% \mathrm{HL}$, mean $=29.3$ vs. $23.1-28.3 \% \mathrm{HL}$, mean $=25.3$; eye height: $26.5-$ $30.6 \% \mathrm{HL}$, mean $=28.7$ vs. $22-27.2 \% \mathrm{HL}$, mean $=24.4$; eye cavity: $30.4-36.8 \%$ HL, mean $=33.3$ vs. $25.6-32.9 \%$ HL, mean $=29$ ). Based on ratios $C$. albellus smaller than 163.5 mm SL can be distinguished from C. brienzii by a larger "maxilla length / eye socket width" ratio (M/ES: 5.35-9.76 vs. 3.31-4.37), "pectoral fin 2 length / predorsal length" ratio (PecF2/PreD: 0.36-0.45 vs. 0.29-0.32), "lower jaw length / eye socket width" ratio (LJ/ES: 9.62-17.28 vs. 6.01-6.49) and a smaller "predorsal length / lower jaw length" ratio (PreD/LJ: 3.99-4.68 vs. 5.05-5.57). With the full size range (100-290 mm) of Lake Brienz specimens, C. albellus can be distinguished from C. brienzii by a larger "eye height / head length" ratio (EH/HL: 0.27-0.31 vs. $0.22-0.27$ ) and a smaller "predorsal length / eye height" ratio (PreD/EH: 6.1-7.58 vs. 8.12-10.32) (Tables 3, 7, 11).

## Coregonus albellus-Coregonus steinmanni

Coregonus albellus from Lake Thun can be distinguished from C. steinmanni by having a longer longest gill raker ( $14.1-20.3 \% \mathrm{HL}$, mean $=17.2$ vs. $10-14.4 \% \mathrm{HL}$, mean $=12.1$ ), a longer maxilla ( $20.1-26.8 \% \mathrm{HL}$, mean $=22.4$ vs. $18.1-21.8 \% \mathrm{HL}$, mean $=19.7)$, a less deep caudal peduncle $(6.4-7.9 \% \mathrm{SL}$, mean $=7.1$ vs. $7.5-8.6 \%$ SL, mean $=8.0)$ and a deeper adipose fin $(4.5-7.7 \% \mathrm{SL}$, mean $=5.8$ vs. $3.7-5.4 \%$ HL, mean $=4.5$ ). Based on ratios C. albellus can be distinguished from C. steinmanni by a smaller "caudal peduncle depth / upper jaw length" ratio (CD/UJ: 0.96-1.29 vs. 1.36-1.55) (Tables 3, 6, 10).

## Coregonus albellus-Coregonus profundus

Coregonus albellus from Lake Thun differs from C. profundus by having more and longer gill rakers (upper arch gill raker number: $9-17$, mode $=13$ vs. $5-10$, mode $=9$; lower arch gill raker number: $20-28$, mode $=24$ vs. $10-18$, mode $=14$; total number of gill rakers: $32-44$, mode $=38$ vs. $15-27$, mode $=21$; middle gill raker length: 11.7$18.3 \% \mathrm{HL}$, mean $=15.6$ vs. $7.6-11.7 \% \mathrm{HL}$, mean $=9.2$; longest gill raker length:
$14.1-20.3 \% \mathrm{HL}$, mean $=17.2$ vs. $7.8-12.4 \% \mathrm{HL}$, mean $=10.1)$. Based on ratios C. albellus can be distinguished from C. profundus by a larger "caudal peduncle length / eye cavity length" ratio (CL/EC: 1.97-2.87 vs. 1.56-2.09) (Tables 3, 8, 10).

## Coregonus albellus-Coregonus acrinasus

Coregonus albellus from Thun can be distinguished from C. acrinasus by having a deeper adipose fin ( $4.5-7.7 \% \mathrm{SL}$, mean $=5.8$ vs. $3.7-6.2 \% \mathrm{SL}$, mean $=4.7$ ), a thinner eye socket (ES: $2-4.9 \% \mathrm{HL}$, mean $=3.2$ vs. $3.2-6.4 \% \mathrm{HL}$, mean $=4.7$ ) and a longer longest gill raker ( $14.1-20.3 \% \mathrm{HL}$, mean $=17.2$ vs. $11.4-16.9 \% \mathrm{HL}$, mean $=14.5)($ Tables 3,9$)$.

Description. General appearance is shown in Figure 4. Morphological and meristic characters of both sexes can be found in Table 3 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Tables 10, 11. The description is valid for both sexes and both lakes; differences between the populations of lakes Thun and Brienz are mentioned.

Shape: Body elongate, slender. Greatest body depth anterior of the dorsalfin. Ventral profile and dorsal profile similar and slightly arched. Dorsal and ventral profile from tip of snout to interorbital mostly straight and then slightly convex to dorsal and pelvic fin origin respectively. Head long. Snout often $40-50^{\circ}$ angle to the body axis anterior of the eye, such that the profile from the tip of the snout to the vertical projection where the anterior part of the eye crosses the dorsal profile is straight and afterwards slightly convex. Mouth (i.e., width of upper and lower jaw) wide, long and often terminal and only rarely slightly sub-terminal. Snout mostly wider than deep, not strongly pronounced, since the tip of the snout is often fleshy and roundish. Large eye, which is more pronounced in specimens from Lake Brienz. Individuals from both lakes have a thin and roundish eye-socket from the middle to the outer margins. Pectoral fin long and moderately tapered. Anterior unbranched ray of the erected dorsal fin range from almost vertically straight to an approx. $70-80^{\circ}$ angle to body axis and only bent slightly posteriorly at the end of the ray. Caudal peduncle narrow and elongated with caudal fin forked and sometimes moderately to strongly asymmetrical with either the ventral or dorsal part being longer. Unbranched ray of anal fin straight and rarely bent posteriorly at the end of the ray. Anal fin longest anteriorly and progressively shortening posteriorly with the outer margin of the anal fin slightly concave.

Meristics: Many and long gill rakers.
Colour: Pigmentation of fins and body overall weak in live specimens. In specimens from Lake Thun the pectoral fin is translucent, sometimes yellowish with faint pigmentation at the median to distal parts of the fin. Pelvic fin is translucent and only weakly to moderately pigmented. Dorsal, adipose, anal and caudal fins are moderately pigmented. In specimens from Lake Brienz all fins are translucent, with the dorsal, anal and caudal fins sometimes showing some very faint pigmentation. In both lakes fish have a silvery appearance along the flanks and dorsally above the lateral line the silvery appearance changes to a pale rose colouration (e.g., RGB $(247,187,175)$ ) and then to a pale brown (e.g., RGB $(230,202,110)$ ). In specimens from Lake Thun the flanks very rarely have few pigmented small dots on the scales. Distribution of dots are bound
to the scale patterning (i.e., at the edge of the scales or at the boundary point of two scales. In specimens from Lake Brienz the upper dorsum ranges from pale brown (e.g., RGB (230, 202, 110)) to a light green colouration (e.g., RGB $(136,245,205)$ ) and sometimes has pigmented dots more or less in a row on the upper dorsum that are rather large ("cheetah look") (Suppl. material 1: Figure S7). Distribution of the dots not restricted to the scale patterning (i.e., at the edge of the scales or at the boundary point of two scales), as can be found for the species of C. alpinus, C. steinmanni, C. brienzii and $C$. fatioi. For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. Dorsal part of head of specimens of Lake Brienz is weakly pigmented, whereas that of specimens from Lake Thun is moderately pigmented. Snout around the nostrils is weakly (Lake Brienz) to moderately (Lake Thun) pigmented with a gap of little pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. Operculum and pre-operculum are silvery with one black dot on the lower margin of the pre-operculum. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. The silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB (239, $210,40)$ ), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus albellus is found in the lakes Thun ( $46^{\circ} 40{ }^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ) and Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ) that are connected by the short river Bödeli Aare at Interlaken. It is believed to have been endemic to these lakes yet,individual fish have been caught in Lake Biel $\left(47^{\circ} 05^{\prime} \mathrm{N}, 7^{\circ} 10^{\prime} \mathrm{E}\right)$ in recent years (since 2005), after it was artificially connected with Lake Thun through the river Aare during the Jura water correction project dating back to 1868-1878. Individuals of C. albellus were first identified by local fishermen and fisherwomen, which reported that they had caught small, ripe fish during the summer months (Bittner 2009). The native whitefish species of Lake Biel only spawn in the winter months (Fatio 1885; Steinmann 1950; Rufli 1978). Genetic analysis has shown that these summer-ripe individuals belong to the species C. albellus (Bittner 2009). We show for two ripe specimens caught in summer in Lake Biel, genetically assigned based on the assignment method of Dönz et al. (2018) to C. albellus with $84 \%$ and $94 \%$ probability, that they can also be assigned to C. albellus based on their morphology (gill raker number, morphological characters) (Suppl. material 1: Figure S9). The species may have established an independent population in Lake Biel, since ripe fish have now been caught for several years in reasonable numbers during the usual spawning period known for this species from Lake Thun (Bittner 2009; Vonlanthen and Périat 2018). Coregonus albellus feeds predominantly on zooplankton (stomach content for Lake Brienz: Maurer and Guthruf 2005; Müller et al. 2007; isotopic signature for both lakes: Selz 2008; Hudson 2011; Ingram et al. 2012) and has a slow growth rate (Kirchhofer 1995; Müller et al. 2007; Bittner et al. unpublished). The gill raker number and length of C. albellus (many and long gill rakers) also suggests that, based on the functional properties of the number of gill rakers (experimentally tested with specimens of this species and other whitefish species from lakes Thun and Lucerne) (Lundsgaard-Hansen et al. 2013; Roesch
et al. 2013), that C. albellus feeds predominantly on zooplankton. Habitat-stratified random sampling of lakes Thun (mid-October 2013: Vonlanthen et al. 2015) and Brienz (mid-September 2011: Vonlanthen et al. 2013) show for a snapshot of a few months in summer, that C. albellus in Lake Thun occupies the moderately shallow to the deepest benthic waters (approx. 30-217 m; N=29) and the moderately shallow to moderately deep pelagic waters (approx. $10-70 \mathrm{~m} ; N=44$ ) (Dönz et al. 2018). In Lake Brienz C.albellus occupies the very shallow (few meters) to the deepest waters $(260 \mathrm{~m})$ of the benthic habitat $(N=78)$ and the very shallow to the deeper waters of the pelagic habitat (few meters down to approx. 60 m and exceptionally down to 130 $\mathrm{m} ; N=47$ ) (Dönz et al. 2018). It is to note that the habitat-stratified random sampling data for both lakes only covers a short period of time (one month in late summer) and it is thus not clear how the species are distributed spatially through the rest of the year. Furthermore, the habitat-stratified random sampling in both lakes did not distinguish between ripe and unripe specimens, and thus in the case of C. albellus the distribution pattern along the depth in the benthic zone is biased by the spawning aggregation of this species since the sampling period in both lakes coincides with the main spawning season of this species. Most of the whitefish that were phenotypically assigned as C. albellus and that were caught in deeper waters during habitat stratified sampling of lakes Brienz and Thun were ripe (PV pers. obs.). In Lake Thun C. albellus phenotypically resembles $C$. fatioi and to some extent $C$. profundus. Interestingly, Steinmann (1950) already mentioned for Lake Thun that C. albellus (Steinmann, 1950: Coregonus lavaretus L. nat. arurenis, oekot. nanus; common name: "Zwergalbock" or "Brienzlig") resembles morphologically C. fatioi (Steinmann, 1950: Coregonus lavaretus L. nat. arurenis, oekot. pelagicus; common name: "Schwebalbock" or "Albock"). The average size (total length) at 3 years of age for specimens in this study is $258 \pm 13 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=9$ ) and $152 \pm 8 \mathrm{~mm}(\mathrm{~N}=14)$ for lakes Thun and Brienz, respectively (Suppl. material 1: Figures S4-S6). In Lake Brienz the size of 3-year old specimens of $C$. albellus is considerably smaller than that of the other three whitefish species (C. alpinus, C. brienzii, C. fatioi), whereas in Lake Thun it is similar to that of C. profundus and C. fatioi (Suppl. material 1: Figure S6) and smaller than that of C. alpinus, C. steinmanni, and C. acrinasus. Coregonus albellus has a long spawning season with two peaks. The main spawning peak is in late summer/early autumn from August to October (Locally known as "Sommer-Brienzlig") and the second peak is in early to late winter from December to March (locally known as "Winter-Brienzlig") (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). Spawning depth varies with spawning season and can range from approx. 30 m to max. lake depth at 217 m in Lake Thun and approx. 50 m to max. lake depth at 261 m in Lake Brienz (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). The spawning season and depth of C. albellus partially overlaps with that of C. steinmanni, C. fatioi, and C. profundus in Lake Thun and with that of C. brienzii and C. fatioi in Lake Brienz.

Common names. Brienzlig, Brienzling; often the time of the year the fish is caught on the spawning grounds is added to the name and shows that this species has a very wide spawning season: Sommer-Brienzlig (for summer) or Winter-Brienzlig (for win-
ter). This species was historically known by local fishermen and fisherwomen as white whitefish (German: "Weissfelchen", but also Albele and Albuli). The common name for this species today is Brienzling which has an ending that is known as a diminutive suffix.

## Coregonus alpinus Fatio, 1885

## Figure 5

Coregonus "Albock": Rufli 1978, 1979; Kirchhofer and Tschumi 1986; Kirchhofer 1995 (see also synonymy of C. steinmanni and C. acrinasus)
Coregonus balleus: Fatio 1885
Coregonus "Balchen", "THU2": Douglas et al. 1999, 2003; Douglas and Brunner 2002 (see also synonymy of C. steinmanni)
Coregonus "Balchen": Heuscher 1901; Surbeck 1917 (see also synonymy of C. steinmanni)
Coregonus "Felchen": Kirchhofer 1990; Kirchhofer 1995 (see also synonymy of C. fatioi and C. brienzii)
Coregonus "Large type": Maurer and Guthruf 2005; Müller et al. 2007 (see also synonymy of C. fatioi and C. brienzii)
Coregonus lavaretus natio arurensis, oekot. litoralis: Steinmann 1950
Coregonus lavaretus natio arurensis, oekot. primigenius: Steinmann 1950 (see also synonymy of C. fatioi and C. steinmanni)
Coregonus schinzii alpinus: Fatio 1885
Coregonus schinzii helveticus: Fatio 1890
Coregonus schinzii helveticus var. Thunensis: Fatio 1890
Coregonus sp. "Balchen": Hudson et al. 2011, 2013, 2016; Ingram et al. 2012; Vonlanthen et al. 2012, 2015; Lundsgaard-Hansen et al. 2013; Roesch et al. 2013; Vonlanthen and Périat 2013 (see also synonymy of C. steinmanni and C. brienzii)
Coregonus sp. "Balchen 1": Dönz et al. 2018
Coregonus "Albock", "Uferalbock": Steinmann 1950 (see also synonymy of C. steinmanni and C. fatioi)

Material examined. Lectotype. MHNG-717.045, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}$, $7^{\circ} 46{ }^{\prime} \mathrm{E}$ ), 283 mm SL, sex unknown.

Non-types. NMBE-1077241-1077261, Switzerland, Lake Thun (4640'N, $7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=21,210-364 \mathrm{~mm}$ SL; NMBE-1059817; 1059821; 1077134, NMBE-10771101077115 , Switzerland, Lake Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ), $\mathrm{N}=9,147-290 \mathrm{~mm}$ SL.

Diagnosis. Coregonus alpinus is a large whitefish with strong pigmentation of all fins and the body; greenish blue colour on the flanks above the lateral line; moderate to many pigmented small dots on the scales along the flank and the dorsum; deep bodied; truncated blunt snout; short head; sub-terminal mouth; small eye with a thick and triangular-shaped eye socket; short and stout caudal peduncle; few and short gill rakers.

Differential diagnosis. Differential diagnosis against C. albellus is given under that species account. The total number of gill rakers of 25 to 34 with mode-values
of 28,29 , and 30 distinguishes $C$. alpinus from all other six whitefish species of lakes Thun and Brienz, by either having more gill rakers than the species C. profundus (total GR: $15-27$, mode $=21$ ) or fewer gill rakers than $C$. fatioi (total GR: 32-43, mode $=$ 38), C. albellus (32-44, mode $=38$ ), C. steinmanni $(30-35$, mode $=31)$, C. brienzii $(32-39$, mode $=37)$ and C. acrinasus $(30-40$, mode $=36)$ (Suppl. material 1: Table S6). The contemporary gill raker range is congruent with the historical gill raker range (23-27) given in Fatio (1890).

For specimens in Lake Brienz smaller than 163.5 mm SL C. alpinus can be distinguished from the other three whitefish species by a larger "length of the depressed anterior part of the dorsal fin / lower jaw length" ratio (DFAd/LJ: 2.57-2.58 vs. 1.6-2.1). For fish larger than 163.5 mm SL, C. alpinus can be distinguished from C. brienzii and C. fatioi by a larger "length of the erected anterior part of the dorsal fin / upper jaw length" ratio (DFAe/UJ: 3.28-4.1 vs. 2.58-3.19). With the full-size range (100290 m ) of Lake Brienz specimens, C. alpinus can be distinguished from the other three whitefish species by a larger "length of the erected anterior part of the dorsal fin / upper jaw length" ratio (DFAe/UJ: 3.25-4.1 vs. 2.14-3.19) (Table 11).

## Coregonus alpinus-Coregonus fatioi

The specimens from lakes Thun and Brienz of C. alpinus can be distinguished from those of $C$. fatioi by having a shorter under jaw ( $24.3-30.1 \%$ HL, mean $=27.4$ vs. $27.6-34.1 \% \mathrm{HL}$, mean $=30$ ), and a shorter longest gill raker $(10-15.2 \% \mathrm{HL}$, mean $=$ 11.9 vs. $12.3-22.6$, mean $=15.6$ ).

In Lake Brienz C. alpinus can be distinguished from C. fatioi by having a shorter caudal peduncle ( $11.3-13.9 \%$ SL, mean $=12.5$ vs. $13.1-16.1 \%$ SL, mean $=14.2$ ) and a shorter and narrower lower jaw (lower jaw length: 33.8-39.4\% HL, mean = 38.2 vs. $37.6-48.4 \% \mathrm{HL}$, mean $=42.6$; lower jaw width: $7.3-10.6 \% \mathrm{HL}$, mean $=$ 8.8vs. $8.6-13.3 \% \mathrm{HL}$, mean $=11.6$ ). For fish from Lake Brienz larger than 163.5 mm SL, C. alpinus can be distinguished based on ratios from C. fatioi by having a larger "length of the erected anterior part of the anal fin / upper jaw length" ratio (AFAe/ UJ: 1.96-2.5 vs. 1.66-1.96) and a larger "head length / upper jaw length" ratio (HL/ UJ: 3.55-3.93 vs. 3.13-3.55). With the full size range ( $100-290 \mathrm{~mm}$ ) of Lake Brienz specimens, C. alpinus can be distinguished from C. fatioi by having a larger "length of the erected anterior part of the dorsal fin / upper jaw length" ratio (DFAe/UJ: 3.25-4.1 vs. 2.14-3.19) (Table 11).

In Lake Thun C. alpinus can be further distinguished from C. fatioi by having a shorter postdorsal length (38.3-43.9\% SL, mean $=42.7$ vs. $41.6-50.7 \% \mathrm{SL}$, mean $=$ 44.9) and a thicker eye socket ( $3.4-6.3 \% \mathrm{HL}$, mean $=5.1$ vs. $1.7-5.9 \% \mathrm{HL}$, mean = 3.6). Based on ratios C. alpinus can be distinguished from C. fatioi by having a larger "caudal peduncle depth / postdorsal length" ratio (CD/PostD: 0.17-0.21 vs. 0.14-0.17) (Tables 4, 5, 10).

## Coregonus alpinus-Coregonus brienzii

C. alpinus from Lake Brienz can be differentiated from C. brienzii by having a shorter caudal peduncle ( $11.3-13.9 \% \mathrm{SL}$, mean $=12.5$ vs. $12.2-15.8 \% \mathrm{SL}$, mean $=$ 13.8), a shorter upper and lower jaw (upper jaw: $25.4-29.1 \%$ HL, mean $=26.8$ vs. $27.1-32 \% \mathrm{HL}$, mean $=29.5$; lower jaw: $33.8-39.4 \% \mathrm{HL}$, mean $=38.2$ vs. $40.5-45.7 \%$
Table 4. Morphological and meristic data of C. alpinus Fatio 1885 from lakes Thun and Brienz, MHNG-717.045 lectotype from Lake Thun; non-type material
$\mathrm{N}=21$ from Lake Thun and $\mathrm{N}=9$ from Lake Brienz.

| Morphological characters | $\begin{array}{\|l\|} \hline \text { C. alpinus } \\ \hline \text { Lectotype } \\ \hline \end{array}$ | Both lakes |  |  |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-types both sexes |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  |  |  | $N$-total $=21$ |  | $N$-females $=12$ |  | $N$-males $=9$ |  | N-total $=9$ |  | N-females = 6 |  | N -males $=3$ |  |
|  |  | Mean $\pm$ StDev | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ <br> StDev | Range | Mean $\pm$ StDev | Range | Mean $\pm$ StDev | Range | Mean $\pm$ StDev | Range | $\begin{aligned} & \text { Mean } \pm \\ & \text { StDev } \end{aligned}$ | Range |
| SL (mm) | 283.0 | $266.1 \pm 56.4$ | (147-364) | $288.3 \pm 45.1$ | (210-364) | $299.6 \pm 37.3$ | (267-364) | $273.3 \pm 52.2$ | (210-352) | $214.3 \pm 46.0$ | (147-290) | $240.1 \pm 27.9$ | (213-290) | $162.7 \pm 23.3$ | (147-190) | Percentage of standard length


| $(4.0-4.8)$ | $3.7 \pm 0.3$ | $(3.1-4.0)$ | $3.9 \pm 0.1$ | $(3.7-4.0)$ | $3.5 \pm 0.5$ | $(3.1-4.0)$ |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | | $(6.0-7.6)$ | $6.5 \pm 0.6$ | $(5.9-7.1)$ |
| :---: | :---: | :---: |
| $(14-18.1)$ | $16.8 \pm 1.4$ | $(15.2-18)$ |











| $(42.7-53.0)$ | $50.1 \pm 3.8$ | $(42.7-53.0)$ | $51.4 \pm 0.7$ | $(50.6-51.9)$ |
| :--- | :---: | :---: | :---: | :---: |
| $(76.1-81.8)$ | $78.2 \pm 2.0$ | $(76.1-81.8)$ | $76.8 \pm 0.3$ | $(76.5-77.0)$ |




 Percentage of head length


| Morpho-logicalcharacters | $\begin{array}{\|l\|} \hline \text { C. alpinus } \\ \hline \text { Lectotype } \\ \hline \end{array}$ | Both lakes |  | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-types both sexes |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  | $N$-total $=30$ |  | $N$-total $=21$ |  | $N$-females $=12$ |  | $N$-males $=9$ |  | N-total $=9$ |  | N-females $=\mathbf{6}$ |  | $N$-males $=3$ |  |
|  |  | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range |
| EH | 23.7 | $22.2 \pm 1.8$ | (19.1-27.1) | $21.4 \pm 1.4$ | (19.1-23.6) | $21.1 \pm 1.2$ | (19.1-23) | $21.9 \pm 1.6$ | (19.6-23.6) | $23.9 \pm 1.5$ | (22.4-27.1) | $23.3 \pm 0.9$ | (22.4-24.6) | $25.1 \pm 2$ | (23.2-27.1) |
| ES | 5.7 | $5.0 \pm 0.8$ | (3.3-6.3) | $5.1 \pm 0.8$ | (3.4-6.3) | $5.0 \pm 0.7$ | (3.7-6.0) | $5.2 \pm 0.9$ | (3.4-6.3) | $4.7 \pm 0.8$ | (3.3-5.8) | $4.6 \pm 0.9$ | (3.3-5.5) | $4.9 \pm 0.8$ | (4.3-5.8) |
| PostO | 51.2 | $52.4 \pm 1.5$ | (48.9-55.4) | $52.6 \pm 1.2$ | (50.2-55.4) | $52.5 \pm 1.3$ | (50.8-55.4) | $52.8 \pm 1.2$ | (50.2-54.4) | $51.7 \pm 1.9$ | (48.9-54.8) | $52.2 \pm 1.5$ | (50.7-54.8) | $50.8 \pm 2.6$ | (48.9-53.8) |
| HD | 71.7 | $71 \pm 3.7$ | (65.5-79.6) | $71.6 \pm 4$ | (65.6-79.6) | $71.5 \pm 4.6$ | (65.6-79.6) | $71.8 \pm 3.4$ | (67.9-76.7) | $69.4 \pm 2.5$ | (65.5-73.2) | $69.7 \pm 2.7$ | (65.5-73.2) | $68.7 \pm 2.3$ | (66.3-70.8) |
| MW | 8.9 | $9.4 \pm 0.5$ | (8.4-10.4) | $9.4 \pm 0.5$ | (8.4-10.4) | $9.3 \pm 0.6$ | (8.4-10.4) | $9.4 \pm 0.6$ | (8.8-10.2) | $9.5 \pm 0.5$ | (8.7-10.2) | $9.6 \pm 0.5$ | (9.2-10.2) | $9.3 \pm 0.5$ | (8.7-9.7) |
| UJ | 27.8 | $27.4 \pm 1.5$ | (24.3-30.1) | $27.7 \pm 1.5$ | (24.3-30.1) | $27.2 \pm 1.5$ | (24.3-29.4) | $28.4 \pm 1.3$ | (26.5-30.1) | $26.8 \pm 1.3$ | (25.4-29.1) | $26.5 \pm 1$ | (25.6-28.2) | $27.3 \pm 1.8$ | (25.4-29.1) |
| LJ | 38.2 | $38.4 \pm 1.7$ | (33.8-41.4) | $38.6 \pm 1.7$ | (36.6-41.4) | $38.5 \pm 1.8$ | (36.6-41.4) | $38.7 \pm 1.6$ | (37-41.2) | $38.2 \pm 1.8$ | (33.8-39.4) | $38 \pm 2.1$ | (33.8-39.2) | $38.5 \pm 1.3$ | (36.9-39.4) |
| M | 22.5 | $20 \pm 1.2$ | (16.6-22.5) | $20 \pm 1.1$ | (17.7-22.1) | $19.7 \pm 0.9$ | (18.4-21) | $20.3 \pm 1.3$ | (17.7-22.1) | $20 \pm 1.6$ | (16.6-22.5) | $19.6 \pm 1.6$ | (16.6-21.1) | $20.8 \pm 1.5$ | (19.5-22.5) |
| SD | 9.9 | $10.1 \pm 1.4$ | (7.2-12.9) | $10.5 \pm 1.3$ | (8.5-12.9) | $10.5 \pm 1.4$ | (8.5-12.9) | $10.6 \pm 1.2$ | (8.7-12.2) | $9.2 \pm 1.1$ | (7.2-10.5) | $9.2 \pm 0.8$ | (7.9-10) | $9.3 \pm 1.9$ | (7.2-10.5) |
| SW | 13.8 | $15.6 \pm 1.1$ | (13.7-17.6) | $15.6 \pm 1.2$ | (13.7-17.6) | $15.7 \pm 1.1$ | (14.1-17.6) | $15.5 \pm 1.4$ | (13.7-17.2) | $15.7 \pm 0.9$ | (14.6-17.6) | $15.6 \pm 0.8$ | (14.6-16.7) | $16 \pm 1.3$ | (15.3-17.6) |
| HW | 47.1 | $50 \pm 4.5$ | (39.2-59.5) | $51.3 \pm 4.1$ | (44.2-59.5) | $51.5 \pm 5$ | (44.2-59.5) | $51 \pm 2.8$ | (46.3-55.9) | $46.9 \pm 4$ | (39.2-52.3) | $47.4 \pm 5$ | (39.2-52.3) | $45.8 \pm 0.4$ | (45.4-46.1) |
| IOW | 24.5 | $27.7 \pm 2.2$ | (22.4-32.5) | $28.3 \pm 2.3$ | (22.4-32.5) | $28 \pm 2.4$ | (22.4-32.4) | $28.7 \pm 2.3$ | (24.9-32.5) | $26.5 \pm 1.2$ | (24.9-28) | $26.5 \pm 1.4$ | (24.9-28) | $26.6 \pm 1$ | (25.5-27.4) |
| INW | 10.7 | $11.7 \pm 1$ | (9.5-14.1) | $12.1 \pm 1$ | (10.5-14.1) | $12 \pm 1.1$ | (10.5-14.1) | $12.1 \pm 0.9$ | (10.7-14) | $11 \pm 0.7$ | (9.5-11.9) | $10.9 \pm 0.9$ | (9.5-11.9) | $11.2 \pm 0.4$ | (10.9-11.6) |
| LJW | 10.3 | $11.1 \pm 2.2$ | (7.3-15.7) | $12.1 \pm 1.7$ | (10.1-15.7) | $11.9 \pm 1.8$ | (10.1-15.4) | $12.4 \pm 1.6$ | (10.5-15.7) | $8.8 \pm 1.2$ | (7.3-10.6) | $9 \pm 1.2$ | (7.5-10.6) | $8.5 \pm 1.4$ | (7.3-10.1) |
| UJW | 19.8 | $23.2 \pm 2.2$ | (18.4-27.2) | $23.5 \pm 2.2$ | (19.9-27.2) | $23 \pm 2.5$ | (19.9-27.2) | $24.1 \pm 1.6$ | (21.9-26.5) | $22.8 \pm 2.2$ | (18.4-25.6) | $23.7 \pm 1.6$ | (21.4-25.6) | $20.9 \pm 2.2$ | (18.4-22.8) |
| MGR | 10.6 | $10.9 \pm 1.4$ | (8.3-15.2) | $11.3 \pm 1.4$ | (9.3-15.2) | $11.3 \pm 1.4$ | (9.3-15.2) | $11.3 \pm 1.4$ | (9.5-13.2) | $9.8 \pm 1$ | (8.3-11.2) | $9.9 \pm 1.1$ | (8.3-11.2) | $9.7 \pm 0.8$ | (8.7-10.2) |
| LGR | 11.5 | $11.9 \pm 1.2$ | (10-15.2) | $12.3 \pm 1.1$ | (10.6-15.2) | $12.2 \pm 1.2$ | (10.6-15.2) | $12.5 \pm 1$ | (11.3-14.1) | $10.8 \pm 0.7$ | (10-12.3) | $10.9 \pm 0.8$ | (10-12.3) | $10.6 \pm 0.6$ | (10-11.2) |
| UA | na | $18.5 \pm 1.4$ | (15.6-21.5) | $18.4 \pm 1.3$ | (15.6-20.7) | $18.6 \pm 1.5$ | (15.6-20.7) | $18.2 \pm 1.1$ | (16.1-19.7) | $18.7 \pm 1.7$ | (16.4-21.5) | $18.2 \pm 1.4$ | (16.4-20) | $19.8 \pm 1.7$ | (18-21.5) |
| LA | 35 | $33.9 \pm 2.1$ | (28.6-38.8) | $33.5 \pm 1.8$ | (28.6-36.3) | $34.1 \pm 1.5$ | (30.9-36.3) | $32.7 \pm 1.9$ | (28.6-35) | $34.7 \pm 2.7$ | (30.4-38.8) | $34.5 \pm 2.7$ | (30.4-38.2) | $35.1 \pm 3.2$ | (32.8-38.8) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 11 | 11 | (10-11) | 11 | (10-11) | 11 | (10-11) | 11 | (10-11) | 10 | (10-11) | 10 | (10-11) | 11 | (10-11) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 16 | 15 | (14-17) | 15 | (14-17) | 15 | (14-17) | 16 | (14-16) | 15 | (15-17) | 15 | (15-16) | na | (15-17) |
| DF unbranched | 3 | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (4-4) |
| DF <br> branched | 11 | 11 | (10-13) | 11 | (10-13) | 11 | (10-13) | 11 | (10-12) | 11 | (10-11) | 11 | (10-11) | 11 | (11-11) |
| AF <br> unbranched | 3 | 3 | (3-4) | 3 | (3-4) | 3 | (3-3) | 3 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) |


| Meristic characters | C. alpinus Lectotype |  | kes | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-types both sexes |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  | N -total $=30$ |  | $N$-total $=21$ |  | $N$-females $=12$ |  | $N$-males $=9$ |  | $N$-total $=9$ |  | $N$-females $=6$ |  | N -males $=3$ |  |
|  |  | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range |
| AF branched | 12 | 12 | (10-14) | 12 | (10-14) | 12 | (10-14) | 12 | (10-13) | 11 | (11-13) | 12 | (11-13) | 11 | (11-11) |
| LS | 82 | 84 | (77-93) | 81 | (77-93) | 84 | (77-93) | 80 | (78-84) | 84 | (80-88) | 86 | (80-88) | na | (81-84) |
| PDS | 39 | 36 | (32-42) | 36 | (32-42) | 36 | (32-42) | 33 | (33-38) | 33 | (32-42) | na | (33-42) | na | (32-37) |
| TDS | 10 | 10 | (8-11) | 10 | (9-11) | 10 | (9-11) | 10 | (9-11) | 10 | (8-10) | 10 | (8-10) | 9 | (9-10) |
| TAS | 8 | 8 | (7-9) | 8 | (8-9) | 8 | (8-9) | 8 | (8-9) | 8 | (7-8) | 8 | (7-8) | 8 | (7-8) |
| TPS | 8 | 8 | (7-9) | 9 | (8-9) | 9 | (8-9) | 8 | (8-9) | 8 | (7-8) | 8 | (8-8) | 8 | (7-8) |
| UGR | 10 | 10 | (8-11) | 10 | (8-11) | 10 | (9-11) | 11 | (8-11) | 9 | (9-11) | 9 | (9-11) | na | (9-11) |
| LGR | 18 | 19 | (15-23) | 19 | (16-23) | 19 | (16-21) | 20 | (17-23) | 18 | (15-21) | 17 | (17-21) | na | (15-19) |
| total GR | 28 | 29 | (25-34) | 30 | (25-34) | 28 | (26-32) | 30 | (25-34) | 28 | (26-30) | 29 | (27-30) | 28 | (26-28) |

HL, mean $=42.2$ ), a narrower snout $(14.6-17.6 \% \mathrm{HL}$, mean $=15.7$ vs. $15.7-20.2 \%$ HL, mean $=17.8$ ), a narrower lower jaw ( $7.3-10.6 \% \mathrm{HL}$, mean $=8.8$ vs. $10.1-14.1 \%$ HL, mean $=11.5$ ) and shorter gill rakers (middle gill raker length: $8.3-11.2 \% \mathrm{HL}$, mean $=9.8$ vs. $10.9-15.1 \% \mathrm{HL}$, mean = 13.5; longest gill raker length: $10-12.3 \%$ HL , mean $=10.8$ vs. $12.1-16.8 \% \mathrm{HL}$, mean $=14.7$ ). For fish larger than 163.5 mm SL, C. alpinus from Lake Brienz can be distinguished based on ratios from C. brienzii by having a larger "caudal peduncle depth / snout width" ratio (CD/SW: 2.25-2.64 vs. 1.82-2.04), "length of the erected anterior part of the dorsal fin / length from the adipose fin to the caudal fin base" ratio (DFAe/PAdC: $1.11-1.32$ vs. $0.96-1.16$ ) and by having a smaller "lower jaw width / upper jaw width" ratio (LJW/UJW: 0.33-0.44 vs. $0.45-0.55)$. With the full size range ( $100-290 \mathrm{~mm}$ ) of Lake Brienz specimens, C. alpinus can be distinguished from C. brienzii by having a larger "length of the depressed anterior part of the dorsal fin / lower jaw width" ratio (DFAd/LJW: 9.84-14.82 vs. 6.05-8.91), "dorsal head length / lower jaw length" ratio (DHL/LJ: 1.84-2.22 vs. 1.63-1.82), "head depth / lower jaw width" ratio (HD/LJW: 6.72-9.39 vs. 5.236.66), "head length / lower jaw length" ratio (HL/LJ: 2.54-2.96 vs. 2.19-2.47) and a smaller "length of the pectoral fin 2 / length of the depressed anterior part of the dorsal fin" ratio (PecF2/DFAd: $0.74-0.85$ vs. $0.85-1.03$ ) (Tables 4, 7, 11).

## Coregonus alpinus-Coregonus profundus

Coregonus alpinus from Thun differs from C. profundus by having shorter pectoral fins (pectoral fin 1 length: 13.6-18.7\% SL, mean $=16.2$ vs. $16.6-21 \%$ SL, mean $=$ 18.4; pectoral fin 2 length: $15.3-19.7 \% \mathrm{HL}$, mean $=17$ vs. $17.7-23.2 \% \mathrm{SL}$, mean $=$ 20.2), a deeper caudal peduncle ( $7.6-8.9 \%$ SL, mean $=8.2$ vs. $6.5-7.9 \%$ SL, mean $=7.3$ ), a shorter head ( $12.6-15.6 \% \mathrm{SL}$, mean $=14.2$ vs. $15.5-18.4 \% \mathrm{SL}$, mean $=$ 16.4) and longer gill rakers (middle gill raker length: 9.3-15.2\% HL, mean $=11.3$ vs. $7.6-11.7 \% \mathrm{HL}$, mean $=9.2$; longest gill raker length: $10.6-15.2 \% \mathrm{HL}$, mean $=$ 12.3 vs. $7.8-12.4 \% \mathrm{HL}$, mean $=10.1$ ). Based on ratios C. alpinus can be distinguished from C. profundus by having a larger "caudal peduncle depth / dorsal head length" ratio (CD/DHL: $0.54-0.62$ vs. $0.4-0.49)$ (Tables 4, 8, 10).

## Coregonus alpinus-Coregonus acrinasus

Coregonus alpinus can further be differentiated from C. acrinasus by having a shorter lower jaw ( $36.6-41.4 \% \mathrm{HL}$, mean $=38.6$ vs. $38.6-47 \% \mathrm{HL}$, mean $=40.9$ ). Based on ratios $C$. alpinus can be distinguished from $C$. acrinasus by having a larger "caudal fin length / maxilla length" ratio (CF/M: 5.55-6.55 vs. 4.4-5.57) (Tables 4, 9, 10).

Description. General appearance is shown in Figure 5. Morphological and meristic characters of both sexes can be found in Table 4 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Tables 10, 11. The description is valid for both sexes and both lakes; differences between the populations of lakes Thun and Brienz are mentioned.

Shape: Generally deep bodied with greatest body depth anterior of the dorsal fin. Dorsal profile strongly arched compared to ventral profile such that the dorsal profile from the tip of snout to the anterior origin of dorsal fin is moderate to strongly convex. Ventral profile slightly arched such that almost straight or slightly convex from the


Figure 5. Coregonus alpinus, lakes Thun and Brienz, Switzerland A lectotype, MHNG-717.045, Lake Thun, 283 mm SL, sex unknown B close-up of head of lectotype MHNG-717.045 C non-type, NMBE1077246, Lake Thun, 251.5 mm SL, male, freshly caught specimen $\mathbf{D}$ non-type, NMBE-1077115, Lake Brienz, 253 mm SL, female, frozen and defrosted specimen. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.
interorbital area to the pelvic fin origin. Head short. Mouth thin (i.e., width of upper and lower jaw), short and sub-terminal. Rostral plate pronounced and almost equally wide as deep resulting in an almost square shape. Tip of the snout often blunt. Small eye, which is less pronounced in specimens from Lake Brienz. Eye-socket thick and triangular (i.e., sickle-shaped). Pectoral fin moderately tapered. Dorsal fin long with the anterior unbranched ray of the erected dorsal fin approx. $60-70^{\circ}$ angle to body axis and only slightly bent posteriorly at the end of the ray. Caudal peduncle stout and short with the caudal fin forked and sometimess lightly asymmetrical with either the ventral or dorsal part being longer. Unbranched ray of anal fin slightly bent posteriorly. Anal fin longest anteriorly and progressively shortening posteriorly with the outer margin of the anal fin mostly straight and only rarely slightly concave.

Meristics: Few short gill rakers, which are shorter for specimens from Lake Brienz.
Colour: Pigmentation of fins and body over all strong in live specimens. In specimens from Lake Thun the pectoral fin is moderately to strongly pigmented. Dorsal, adipose, pelvic, anal and caudal fins are strongly pigmented. In specimens from Lake Brienz all fins are less pigmented. The pectoral fin is sometimes yellowish and ranges from translucent to moderately pigmented at the median to distal parts of the fin. Dorsal, adipose, pelvic, anal, and caudal fins are moderately pigmented. In both lakes fish have a silvery appearance along the flanks with few to many pigmented small dots on the scales along the flank and the dorsum (as can be found for the species of C. fatioi, C. steinmanni, C. brienzii). The distribution of the dots is bound to the scale patterning such that the dots are found at the edge of the scales or at the boundary point of two scales. Dorsally above the lateral line the silvery appearance changes to a light (e.g., $\operatorname{RGB}(135,236,179))$ or darker greenish blue colour (e.g., RGB $(7,168,125)$ ). The dorsal part of the head of specimens of Lake Brienz is moderately pigmented, whereas that of specimens from Lake Thun is strongly pigmented. The snout around the nostrils is moderately (Lake Brienz) to strongly (Lake Thun) pigmented. Specimens in Lake Brienz have a gap of very weak pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. The pre-operculum and operculum are silvery with one black dot on the lower margin of the pre-operculum. In some specimens of Lake Thun, the pre-operculum and operculum has some pigmented dots, similar to those found on the scales and extending also to the dorsal part of the head. For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. The silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB (239, 210, 40)), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus alpinus is found in the lakes Thun $\left(46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}\right)$ and Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ) that are connected through the river Aare at Interlaken. Coregonus alpinus feeds predominantly on benthic prey and parts
of the year on zooplankton (stomach content for Lake Brienz: Maurer and Guthruf 2005; Müller et al. 2007; isotopic signature for both lakes: Selz 2008; Hudson 2011; Ingram et al. 2012) and has a rapid growth rate (Lake Brienz: Müller et al. 2007; both lakes: Kirchhofer 1995; Bittner et al. unpublished). It has to be noted that the stomach content and isotopic work did not distinguish between all species in lakes Thun or Brienz and thus in some cases lumped different species together into few groups. The stomach content work by Maurer and Guthruf (2005) and Müller et al. (2007) differentiated between "small-type" and "large-type" whitefish based on cohort-specific threshold values for length-at-age. Based on morphology and ecology Kirchhofer (1995) differentiated in Lake Thun between "Albock" (comprising most likely of C. alpinus, C. steinmanni and C. acrinasus), "Brienzlig" (comprising most likely of C. albellus and C. fatioi) and "Kropfer" (C. profundus) and in Lake Brienz between "Felchen" (comprising most likely of C. alpinus, C. fatioi and C. brienzii) and "Brienzlig" and "Winter-Brienzlig" (comprising of summer- and winter-spawning specimens of C. albellus). Finally, Selz (2008), Hudson (2011) and Ingram et al. (2012) did not yet differentiate between C. alpinus and C. steinmanni in Lake Thun, which were most likely both grouped under C. "Balchen". The gill raker number and length of C. alpinus (few and short gill rakers) suggests, based on the functional properties of the number of gill rakers experimentally tested with specimens of this species and other whitefish species from lakes Thun and Lucerne (Lundsgaard-Hansen et al. 2013; Roesch et al. 2013), that C. alpinus feeds more on benthic prey and less on zooplankton. However, this assumption needs to be verified with stomach content analysis that distinguish between the different species within a lake. The relative species abundances in the pelagic and benthic habitat from a habitat-stratified random sampling of Lake Thun (mid-October 2013: Vonlanthen et al. 2015) and Brienz (midSeptember 2011: Vonlanthen et al. 2013) shows, that C. alpinus can only be found in shallow water in the benthic habitat (first $15 \mathrm{~m} ; N=1$ each for lakes Thun and Brienz) and is completely absent from the pelagic habitat in Lake Thun, while in Lake Brienz it can be also found in the very shallow waters (approx. first $5 \mathrm{~m} ; N=2$ ) of the pelagic habitat (Dönz et al. 2018). It is noteworthy that the habitat-stratified random sampling data for both lakes only covers a short period of time (one month in late summer) and it is thus not clear how the species are distributed spatially throughout the rest of the year. In Lake Thun C. alpinus resembles phenotypically C. steinmanni and to some extent $C$. acrinasus. The average size (total length) at 3 years of age for specimens in this study is $321 \pm 20 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=8$ ) and 273 $+14 \mathrm{~mm}(\mathrm{~N}=4)$ for lakes Thun and Brienz respectively (Suppl. material 1: Figures S4-S6). The average size at 3 years of age for the specimens of C. alpinus from Lake Thun from this study are similar to those for the years 1969-1970 (333.8 $\pm \mathrm{mm}, N=$ 13) and 2004-2005 ( $342.8 \pm 21 \mathrm{~mm}, N=14$ ) (Bittner et al. unpublished; Vonlanthen et al. unpublished). In Lake Thun the size at 3 years of age of C. alpinus is similar to that of $C$. steinmanni, larger than that of $C$. acrinasus and considerably larger than that of C. albellus, C. fatioi and C. profundus (Suppl. material 1: Figure S6). In Lake Brienz the size at 3 years of age of C. alpinus is larger than that of C. fatioi and C. brienzii and
considerably larger than that of C. albellus (Suppl. material 1: Figure S6). However, the size-at-age comparisons should be treated with some caution since the sample size for C. alpinus is rather small. Coregonus alpinus has a short spawning season in late December and spawns mostly in very shallow water ( $1-2 \mathrm{~m}$ ) and to a lesser extent down to 10 m and very seldom down to 30 m or more (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). Coregonus alpinus spawns earlier in Lake Brienz than in Lake Thun (Fatio 1890; Dönz et al. 2018). The spawning season and depth of C. alpinus overlaps largely with that of C. acrinasus, C. fatioi and C. steinmanni in Lake Thun and with that of C. fatioi and C. brienzii in Lake Brienz.

Kottelat (1997) has designated a lectotype as C. alpinus which is incongruent with his description of the species (with the common name "Kropfer": Kottelat (1997) and Kottelat and Freyhof (2007)). Fatio $(1885,1890)$ was unaware of this species (the "Kropfer") as it is not considered in his compendium of the Swiss fauna (Fatio 1890) nor in his earlier work on the Swiss whitefish (Fatio 1885). The lectotype designated by Kottelat (1997) to C. alpinus clearly and correctly resembles the description given by Fatio (Fatio 1885, 1890) for the species-group Coregonus schinzii alpinus (Fatio, 1885) and later Coregonus schinzii helveticus (Fatio, 1890), known then and today by its local name as "Balchen". Fatio (1890) describes the "Balchen"-type whitefish as a relatively large whitefish,with few and short gillrakers, a short and stout head with a thick and squared snout, mouth inferior and often subterminal especially for specimens from Lake Thun, a more or less small eye, caudal peduncle short and stout, long pectoral fin, all fins more or less strongly pigmented, colour of live specimens generally olive or grey-olive with greenish or blueish reflections on the back, head more or less strongly pigmented, black pigmented dots more or less abundant on the dorsum, and on the margins of the scales, spawning season in winter (November-December) and spawning depth rather shallow at the shoreline over boulders or stones (Fatio 1885: Page 663, Tables 1, 2; Fatio 1890: Pages 222-234). This description is very clearly and accurately that of a "Balchen", and very different from "Kropfer". We thus identify C. alpinus as the species from lakes Thun and Brienz known under the common name "Balchen". The description of C. alpinus in Kottelat (1997) and Kottelat and Freyhof (2007) and the photograph in Kottelat and Freyhof (2007) does not describe and depict "Balchen" but C. profundus, the species from Lake Thun known by the common name "Kropfer".

Common name. Balchen.

## Coregonus fatioi, Kottelat, 1997 <br> Figure 6

Coregonus "Albock": Heuscher 1901
Coregonus "Albock", "BRI1": Douglas et al. 1999, 2003; Douglas and Brunner 2002 (see also synonymy of C. acrinasus)
Coregonus "Felchen": Kirchhofer 1990; Kirchhofer 1995 (see also synonymy of C. alpinus and C. brienzii)
Table 5. Morphological and meristic data of $C$. fatioi Kottelat, 1997 from lakes Thun and Brienz, MHNG-809.059 lectotype from Lake Thun; non-type material $\mathrm{N}=30$ from Lake Thun and $\mathrm{N}=30$ from Lake Brienz.

| Morphological characters | C. fatioiLectotype | Both lakes |  | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-types both sexes |  | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  | Ntotal $=60$ |  | Ntotal $=30$ |  | Nfemales $=17$ |  | Nmales $=13$ |  | Ntotal $=30$ |  | Nfemales = 12 |  | Nmales $=18$ |  |
|  |  | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ StDev | Range | Mean $\pm$ StDev | Range |
| SL (mm) | 154.5 | $207.5 \pm 35.2$ | (132-288) | $230.2 \pm 21.2$ | (191-288) | $226.6 \pm 14.3$ | (191-245) | $234.9 \pm 27.8$ | (202-288) | $184.8 \pm 31.7$ | (132-244) | $195.9 \pm 29.7$ | (141-244) | $177.3 \pm 31.6$ | (132-225) |


|  |
| :---: |
|  |
| $(3.2-4.9-7.4)$ |
| $(14.6-17.4)$ |
| $(2.7-3.4)$ |
| $(14.1-18.7)$ |
| $(15.3-19.7)$ |
| $(16.5-20.0)$ |
| $(17.1-21.1)$ |
| $(4.3-7.0)$ |
| $(10.6-13.3)$ |
| $(10.2-12.7)$ |
| $(4.0-7.7)$ |
| $(22.7-26.4)$ |

$\frac{\left(\mathrm{I} 9 \mathrm{I}-\mathrm{I}^{\circ} \mathrm{E} \mathrm{I}\right)}{(8-6 \cdot 9)}$


$(45.9-48)$
$(22.1-24.9)$



 :c|c ล $\widetilde{2}$
$\underset{\sim}{2}$

$\underset{\sim}{i}$ | $\approx$ |
| :---: |
|  |
| $\substack{1 \\ \\ \\ \hline}$ |


| Morphological characters | $\begin{array}{\|c\|} \hline \text { C. fatioi } \\ \hline \text { Lectotype } \\ \hline \end{array}$ | Both lakes |  | Lake Thun |  |  |  |  |  | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Non-type | both sexes | Non-types |  |  |  |  |  | Non-types |  |  |  |  |  |
|  |  | Ntotal $=60$ |  | Ntotal $=30$ |  | Nfemales $=17$ |  | Nmales $=13$ |  | Ntotal $=30$ |  | Nfemales $=12$ |  | Nmales $=18$ |  |
|  |  | Mean $\pm$ StDev | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ StDev | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | Mean $\pm$ StDev | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range | $\begin{gathered} \text { Mean } \pm \\ \text { StDev } \end{gathered}$ | Range |
| PostO | 49.8 | $51.3 \pm 1.8$ | (46.8-54.8) | $52.2 \pm 1.4$ | (48.7-54.8) | $52 \pm 1.6$ | (48.7-54.1) | $52.5 \pm 1.1$ | (51.4-54.8) | $50.5 \pm 1.7$ | (46.8-54) | $50.7 \pm 1$ | (49.8-53) | $50.3 \pm 2.1$ | (46.8-54) |
| HD | 68.3 | $69.6 \pm 3.1$ | (63.6-78.6) | $70.7 \pm 3.3$ | (65.5-78.6) | $70.6 \pm 3.7$ | (65.5-78.6) | $70.8 \pm 2.7$ | (66.3-74.8) | $68.5 \pm 2.6$ | (63.6-73.2) | $68.8 \pm 2.8$ | (65.7-73.2) | $68.3 \pm 2.5$ | (63.6-72.2) |
| MW | 11.5 | $10.1 \pm 0.8$ | (8.2-12.1) | $10 \pm 0.9$ | (8.2-12.1) | $10.1 \pm 0.9$ | (8.5-12.1) | $9.8 \pm 0.9$ | (8.2-11) | $10.1 \pm 0.8$ | (8.5-11.4) | $10 \pm 0.9$ | (8.5-11.4) | $10.2 \pm 0.8$ | (8.6-11.4) |
| UJ | 30.5 | $30 \pm 1.4$ | (27.6-34.1) | $30.5 \pm 1.5$ | (28-34.1) | $30.8 \pm 1.6$ | (28-34.1) | $30.2 \pm 1.3$ | (28.2-33.1) | $29.5 \pm 1.3$ | (27.6-32) | $29.4 \pm 1.2$ | (28.1-31.2) | $29.6 \pm 1.3$ | (27.6-32) |
| LJ | 38.4 | $41.6 \pm 2.6$ | (36.9-48.4) | $40.7 \pm 2.1$ | (36.9-46.1) | $41.3 \pm 2.1$ | (37.4-46.1) | $39.8 \pm 1.8$ | (36.9-42.3) | $42.6 \pm 2.6$ | (37.6-48.4) | $43.1 \pm 2.3$ | (40.6-47.8) | $42.2 \pm 2.9$ | (37.6-48.4) |
| M | 26.6 | $21.8 \pm 1.3$ | (18.5-25.6) | $21.8 \pm 1.6$ | (18.5-25.6) | $22.1 \pm 1.4$ | (19.8-25.6) | $21.5 \pm 1.7$ | (18.5-25.1) | $21.7 \pm 1$ | (18.7-24.2) | $22 \pm 0.8$ | (21-23.8) | $21.5 \pm 1.1$ | (18.7-24.2) |
| SD | 8.1 | $9.3 \pm 1.2$ | (6.7-12.4) | $9.5 \pm 1.2$ | (6.7-12.4) | $9 \pm 1$ | (6.7-10.5) | $10.1 \pm 1.1$ | (8.7-12.4) | $9 \pm 1.1$ | (6.7-10.9) | $9.4 \pm 1.2$ | (6.9-10.9) | $8.8 \pm 1$ | (6.7-10.4) |
| SW | 15 | $17.7 \pm 1.3$ | (14.7-20.4) | $17.5 \pm 1.3$ | (14.7-20.4) | $17.6 \pm 1.5$ | (14.7-20.4) | $17.5 \pm 1.1$ | (16-19.6) | $17.8 \pm 1.3$ | (14.7-19.7) | $17.4 \pm 1.6$ | (14.7-19.7) | $18 \pm 1.1$ | (15.8-19.7) |
| HW | 45.2 | $49.8 \pm 3.1$ | (42.3-57.2) | $51.1 \pm 3$ | (45.8-56.6) | $51.5 \pm 3.4$ | (45.8-56.6) | $50.6 \pm 2.3$ | (47.8-54.1) | $48.5 \pm 2.7$ | (42.3-57.2) | $48.8 \pm 3.5$ | (42.3-57.2) | $48.3 \pm 2$ | (44.8-52.8) |
| IOW | 24.5 | $27 \pm 1.5$ | (22.8-31.5) | $27.7 \pm 1.6$ | (23.6-31.5) | $27.2 \pm 1.4$ | (23.6-29.7) | $28.3 \pm 1.6$ | (25.4-31.5) | $26.4 \pm 1.1$ | (22.8-28.8) | $26.6 \pm 0.9$ | (25.4-28.8) | $26.2 \pm 1.3$ | (22.8-28.5) |
| INW | 10.9 | $11.5 \pm 1.1$ | (9.2-13.5) | $11.7 \pm 1$ | (9.8-13.5) | $11.6 \pm 1.1$ | (9.8-13.3) | $11.8 \pm 0.9$ | (10.6-13.5) | $11.4 \pm 1.2$ | (9.2-13.5) | $11.2 \pm 1$ | (10.1-13.3) | $11.5 \pm 1.3$ | (9.2-13.5) |
| LJW | 14.7 | $12 \pm 1.9$ | (7.9-16) | $12.4 \pm 2.3$ | (7.9-16) | $12.5 \pm 2.2$ | (7.9-15.8) | $12.4 \pm 2.5$ | (7.9-16) | $11.6 \pm 1.2$ | (8.6-13.3) | $11.4 \pm 1.3$ | (8.7-13.3) | $11.8 \pm 1.2$ | (8.6-13.2) |
| UJW | 23.7 | $24 \pm 1.8$ | (20.3-30.3) | $24.7 \pm 1.8$ | (21-30.3) | $25.2 \pm 2$ | (21-30.3) | $24.1 \pm 1.3$ | (22.1-26.8) | $23.4 \pm 1.5$ | (20.3-26.5) | $23.1 \pm 1.7$ | (20.3-26.5) | $23.5 \pm 1.5$ | (21-26) |
| MGR | 14.3 | $14.5 \pm 2$ | (10.5-21.3) | $15.8 \pm 1.9$ | (12.5-21.3) | $16.3 \pm 1.8$ | (13.9-21.3) | $15 \pm 1.8$ | (12.5-19.6) | $13.2 \pm 1.2$ | (10.5-15) | $13 \pm 1$ | (11.5-14.4) | $13.3 \pm 1.4$ | (10.5-15) |
| LGR | 14.9 | $15.6 \pm 2.1$ | (12.3-22.6) | $16.9 \pm 2$ | (12.8-22.6) | $17.5 \pm 1.9$ | (15.4-22.6) | $16.1 \pm 1.9$ | (12.8-19.6) | $14.3 \pm 1.1$ | (12.3-16.4) | $14.3 \pm 0.9$ | (12.7-15.5) | $14.3 \pm 1.2$ | (12.3-16.4) |
| UA | 19 | $18.6 \pm 1.5$ | (15.7-22.6) | $19.1 \pm 1.6$ | (16.1-22.6) | $18.6 \pm 1.4$ | (16.1-21.2) | $19.7 \pm 1.7$ | (17.3-22.6) | $18.2 \pm 1.1$ | (15.7-20.1) | $18 \pm 0.8$ | (16.7-19.4) | $18.4 \pm 1.3$ | (15.7-20.1) |
| LA | 35.3 | $35.8 \pm 1.8$ | (32.5-41.3) | $35.9 \pm 1.7$ | (32.5-41.3) | $36.1 \pm 1.4$ | (34-38.6) | $35.7 \pm 2.1$ | (32.5-41.3) | $35.7 \pm 1.9$ | (32.9-39.8) | $35.7 \pm 1.2$ | (33.4-37.3) | $35.8 \pm 2.3$ | (32.9-39.8) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 11 | 10 | (9-11) | 10 | (9-11) | 10 | (9-11) | 10 | (9-11) | 10 | (9-11) | 10 | (10-11) | 10 | (9-11) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 16 | 16 | (14-17) | 16 | (14-17) | 16 | (14-17) | 16 | (15-17) | 16 | (14-17) | 16 | (14-17) | 16 | (14-17) |
| DF unbranched | 4 | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) |
| DF branched | 10 | 10 | (10-13) | 10 | (10-11) | 10 | (10-11) | 10 | (10-11) | 11 | (10-13) | 11 | (10-12) | 11 | (10-13) |
| AF unbranched | 3 | 3 | (2-5) | 3 | (2-4) | 3 | (2-4) | 2 | (2-4) | 4 | (3-5) | 3 | (3-5) | 4 | (3-4) |
| AF branched | 12 | 12 | (10-14) | 12 | (10-14) | 12 | (11-14) | 12 | (10-14) | 12 | (11-13) | 12 | (11-13) | 12 | (11-13) |
| LS | 82 | 86 | (78-93) | 86 | (78-93) | 85 | (78-86) | 86 | (78-93) | 86 | (79-92) | 86 | (81-92) | 86 | (79-91) |
| PDS | 32 | 36 | (30-44) | 32 | (30-44) | 36 | (30-40) | 38 | (31-44) | 34 | (30-40) | 34 | (30-38) | 34 | (33-40) |
| TDS | 10 | 9 | (8-11) | 9 | (8-11) | 9 | (9-10) | 10 | (8-11) | 10 | (9-10) | 10 | (9-10) | 10 | (9-10) |
| TAS | 7 | 8 | (7-10) | 8 | (7-9) | 8 | (7-9) | 8 | (7-9) | 8 | (7-9) | 8 | (8-9) | 8 | (7-9) |
| TPS | 8 | 9 | (7-10) | 9 | (7-9) | 9 | (8-9) | 9 | (7-9) | 9 | (8-9) | 9 | (8-9) | 9 | (8-9) |
| UGR | 11 | 12 | (10-16) | 14 | (10-16) | 14 | (12-16) | 14 | (10-15) | 12 | (11-15) | 12 | (11-14) | 12 | (11-15) |
| LGR | 22 | 24 | (19-27) | 24 | (22-27) | 24 | (22-27) | 24 | (22-26) | 22 | (19-27) | 22 | (22-26) | 24 | (19-27) |
| total GR | 33 | 38 | (32-43) | 38 | (32-43) | 38 | (34-43) | 38 | (32-40) | 35 | (32-40) | 37 | (33-38) | 39 | (32-40) |

Coregonus "Large type": Maurer and Guthruf 2005; Müller et al. 2007 (see also synonymy of C. alpinus and C. brienzii)
Coregonus lavaretus wartmanni natio fatioi: Berg 1932
Coregonus lavaretus natio arurensis, oekot. pelagicus: Steinmann 1950 (see also synonymy of C. steinmanni)
Coregonus lavaretus natio arurensis, oekot. primigenius: Steinmann 1950 (see also synonymy of $C$. steinmanni and $C$. alpinus)
Coregonus "Bodenalbock", "Albock", "Schwebalbock", "Wanderalbock": Steinmann 1950 (see also synonymy of C. alpinus, C. steinmanni)
Coregonus sp. "Felchen": Hudson et al. 2011, 2013, 2016; Ingram et al. 2012
Coregonus sp. "Tiefenalbock": Vonlanthen et al. 2015
Coregonus wartmanni alpinus: Fatio 1890

Material examined. Lectotype. MHNG-809.059, Switzerland, Lake Thun (4640'N, $7^{\circ} 46^{\prime} \mathrm{E}$ ), 154.5 mm SL, sex unknown.

Non-types. NMBE-1077133, NMBE-1077180-1077185, NMBE-10771351077157, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=30,191-288 \mathrm{~mm}$ SL; NMBE-1077342, NMBE-1077291-1077317, NMBE-1077266, NMBE-1077267, Switzerland, Lake Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ), $\mathrm{N}=30,132-244 \mathrm{~mm}$ SL.

Diagnosis. Coregonus fatioi is a medium-sized whitefish with weak pigmentation of all fins and body; light to dark green colour on the flanks above the lateral line; moderate to many pigmented small dots on the scales along the flank and the dorsum; slender, elongated and slightly torpedo-like body; long head; tip of snout is fleshy and roundish; small eye with a thin and triangular to roundish eye socket for individuals from Lake Thun and a thick and triangular shaped eye socket for individuals from Lake Brienz; many and long gill rakers.

Differential diagnosis. Differential diagnoses against C. albellus and C. alpinus are given under those species' accounts.

## Coregonus fatioi-Coregonus brienzii

In Brienz C. fatioi can be differentiated from C. brienzii by being deeper bodied $(22.1-26.2 \%$ SL, mean $=23.9$ vs. $19.6-25.1 \%$ SL, mean $=22.6)$ and having a smaller eye (eye depth: $21.2-27.6 \% \mathrm{HL}$, mean $=24.8$ vs. $23.1-28.3 \% \mathrm{SL}$, mean $=25.3)(\mathrm{Ta}-$ bles 5, 7, 11).

## Coregonus fatioi-Coregonus steinmanni

The specimens of C. fatioi from Lake Thun differ from those of C. steinmanni by having longer gill rakers (middle gill raker length: $12.5-21.3 \% \mathrm{HL}$, mean $=15.8$ vs. 9.1-14.3\% HL, mean = 11.5; longest gill raker length: $12.8-22.6 \% \mathrm{HL}$, mean $=16.9$ vs. $10-14.4 \% \mathrm{HL}$, mean $=12.1$ ), a longer and wider underjaw (under jaw length: $28-34.1 \% \mathrm{HL}$, mean $=30.5$ vs. $25.2-30 \% \mathrm{HL}$, mean $=27.3$; under jaw width: $21-$ $30.3 \%$ HL, mean $=24.7$ vs. $19.3-25 \%$ HL, mean $=23$ ). Based on ratios C. fatioi can be differentiated from C. steinmanni by having a smaller "caudal peduncle depth / upper jaw length" ratio (CD/UJ: 1.02-1.34 vs. 1.36-1.55) and "caudal peduncle depth / postdorsal length" ratio (CD/PostD: 0.14-0.17 vs. 0.17-0.20 (Tables 5, 6, 10).


Figure 6. Coregonus fatioi, lakes Thun and Brienz, Switzerland A, B lectotype, MHNG-809.059, Lake Thun, 154.5 mm SL, sex unknown, left and right side of the specimen $\mathbf{C}$ non-type, NMBE-1077139, Lake Thun, 240 mm SL, male, freshly caught specimen D non-type, NMBE-1077317, Lake Brienz, 202 mm SL, male, frozen and defrosted specimen. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.

## Coregonus fatioi-Coregonus profundus

Coregonus fatioi from Lake Thun can be distinguished from C. profundus by having more and longer gill rakers (upper arch gill raker number: $10-16$, mode $=14$ vs. $5-10$, mode $=9$; lower arch gill raker number: $22-27$, mode $=24$ vs. $10-18$, mode $=$ 14; total gill raker number: $32-43$, mode $=38$ vs. $15-27$, mode $=21$; middle gill raker length: $12.5-21.3 \% \mathrm{HL}$, mean $=15.8$ vs. $7.6-11.7 \% \mathrm{HL}$, mean $=9.2$; longest gill raker length: $12.8-22.6 \% \mathrm{HL}$, mean $=16.9$ vs. $7.8-12.4 \% \mathrm{HL}$, mean $=10.1$ ), shorter pectoral fin (pectoral fin 1 length: 13.3-18.9\% SL, mean $=16.5$ vs. $16.6-21 \%$ SL, mean $=18.4$; pectoral fin 2 length: $13.8-20.6 \%$ SL, mean $=17.7$ vs. $17.7-23.2 \%$ SL, mean $=20.2$ ), a shorter head $(13.6-16.2 \%$ SL, mean $=14.8$ vs. $15.5-18.4 \%$ SL, mean $=16.4)$, a longer postdorsal length ( $41.6-50.7 \% \mathrm{SL}$, mean $=44.9$ vs. $38.9-44.5 \%$ SL, mean $=42.5)$, and a longer upper jaw $(28-34.1 \% \mathrm{HL}$, mean $=30.5$ vs. $26.4-30.6 \%$ HL, mean $=28.7)($ Tables 5, 8, Suppl. material 1: Table S6).

## Coregonus fatioi-Coregonus acrinasus

Coregonus fatioi can be distinguished from C. acrinasus by having a longer postdorsal length ( $41.6-50.7 \%$ SL, mean $=44.9$ vs. $40.3-45.6 \%$ SL, mean $=43$ ) and longer gill rakers (middle gill raker length: $12.5-21.3 \% \mathrm{HL}$, mean $=15.8$ vs. $9.1-16.6 \% \mathrm{HL}$, mean $=13.4$; longest gill raker length: $12.8-22.6 \% \mathrm{HL}$, mean $=16.9$ vs. $11.4-16.9 \%$ HL, mean $=14.5)($ Tables 5, 9).

Description. General appearance is shown in Figure 6. Morphological and meristic characters of both sexes can be found in Table 5 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Tables 10, 11. The description is valid for both sexes and both lakes; differences between the populations of lakes Thun and Brienz are mentioned.

Shape: Elongated. Slender bodied with greatest body depth anterior of the dorsal fin resulting in a slightly torpedo-like form. Dorsal and ventral profile similar and slightly arched. Dorsal and ventral profile from tip of snout to interorbital area mostly straight and then slightly convex to dorsal and pelvic fin origin respectively. Head long. Very rarely does the snout have an approx. $40-50^{\circ}$ angle to the body axis anterior of the eye, such that the profile from the tip of the snout to the vertical projection where the anterior part of the eye crosses the dorsal profile is straight and afterwards slightly convex. Mouth thick (i.e., width of upper and lower jaw), long and often terminal and only rarely slightly sub-terminal. Snout mostly wider than deep, not strongly pronounced, since the tip of the snout is often fleshy and roundish. Specimens from Lake Thun have a thin, roundish and rarely triangular shaped eye-socket, whereas specimens from Lake Brienz have an eye-socket that is thick and triangular (i.e., sickle-shaped). Pectoral fin moderately tapered. Anterior unbranched ray of the erected dorsal fin ranges from almost vertically straight to an approx. $60-80^{\circ}$ angle to body axis and only bent slightly posteriorly at the end of the ray. Caudal peduncle narrow and elongated with caudal fin forked in specimens from both lakes and sometimes moderately asymmetrical (mostly the ventral part is longer) in specimens from Lake Thun but very rarely in specimens from Lake Brienz. Unbranched ray of anal fin straight and rarely bent posteriorly at the end of the ray. Anal fin longest anteriorly and progres-
sively shortening posteriorly with the outer margin of the anal fin ranging from being straight to slightly concave.

Meristics: Specimens of Lake Thun have many and long gill rakers, whereas specimens from Lake Brienz have a bit less and moderately long gill rakers.

Colour: Pigmentation of fins and body overall weak in live specimens. In specimens from Lake Thun the pectoral fin is translucent, sometimes yellowish with faint pigmentation at the median to distal parts of the fin. In Thun the pelvic fin ranges from completely translucent to moderately pigmented and the dorsal, adipose, anal and caudal fins are moderately pigmented. Specimens from Lake Brienz have a fully translucent pectoral fin that sometimes has a faint pigmentation on the unbranched ray. Pelvic and anal fins range from fully transparent to moderately pigmented and dorsal, adipose and caudal fins are moderately pigmented. In both lakes fish have a silvery appearance along the flanks. Specimens from both lakes sometimes have many pigmented small dots on the scales along the flank and the dorsum, which is rare in specimens from Lake Thun and common in specimens from Lake Brienz. Distribution of the dots is bound to the scale patterning such that the dots are found at the edge of the scales or at the boundary point of two scales (as can be found for the species of C. alpinus, C. brienzii and C. steinmanni). Colouration on the dorsum above the lateral line of specimens from Lake Thun ranges from a light green colouration (e.g., $\operatorname{RGB}(136,245,205)$ ) to an olive-green colouration (e.g., RGB (176, 192, 125)), where the former is more common. In specimens from Lake Brienz the upper dorsum is light greenish in colouration (e.g., RGB (136, 245, 205)). For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. The dorsal part of the head of specimens of Lake Brienz is weakly pigmented, whereas that of specimens from Lake Thun is moderately pigmented. The snout around the nostrils is weakly (Lake Brienz) to moderately (Lake Thun) pigmented with a gap of very weak (Brienz) to moderate (Thun) pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. Operculum and pre-operculum are silvery with one black dot on the lower margin of the pre-operculum. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. Silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB $(239,210,40)$ ), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus fatioi is found in the lakes Thun $\left(46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}\right)$ and Brienz $\left(46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}\right)$ that are connected through the river Aare at Interlaken. Based on isotopic signatures C. fatioi feeds predominantly on zooplankton (Selz 2008; Hudson 2011; Ingram et al. 2012). Stomach content analyses of specimens of C. fatioi from Lake Brienz suggest that C. fatioi feeds on a mix of zooplankton and benthic prey (Maurer and Guthruf 2005; Müller et al. 2007). Coregonus fatioi has a moderately fast growth rate (Both lakes: Kirchhofer 1995; Bittner et al. unpublished; Lake Brienz: Müller et al. 2007). It has to be noted that the work by Kirchhofer (1995), Maurer and Guthruf (2005) and Müller et al. (2007) did not
distinguish between all species in lakes Thun or Brienz and thus lumped different species together into few groups. Maurer and Guthruf (2005) and Müller et al. (2007) differentiated between "small-type" and "large-type" whitefish based on cohort-specific threshold values for length-at-age. Based on morphology and ecology Kirchhofer (1995) differentiated in Lake Thun between "Albock" (comprising most likely of C. alpinus, C. steinmanni and C. acrinasus), "Brienzlig" (comprising most likely of C. albellus and C. fatioi) and "Kropfer" (C. profundus) and in Lake Brienz between "Felchen" (comprising most likely of C. alpinus, C. fatioi and C. brienzii) and "Brienzlig" and "Winter-Brienzlig" (comprising of summer- and winter-spawning specimens of C. albellus). The gill raker number and length of C. fatioi (many and long gill rakers) suggests, based on the functional properties of the number of gill rakers (Lundsgaard-Hansen et al. 2013; Roesch et al. 2013), that C. fatioi feeds more on zooplankton and less on benthic prey. However, this assumption needs to be verified with stomach content analyses that distinguish between the different species within a lake. Habitat stratified random sampling of Lake Thun (mid-October 2013; Vonlanthen et al. 2015) and Brienz (mid-September 2011; Vonlanthen et al. 2013) shows, that $C$. fatioi occupies the moderately shallow (Brienz: approx. $1-48 \mathrm{~m}, N=9$; Thun: approx. $25-140 \mathrm{~m}, N=4)$ to the deepest waters of the benthic habitat in both lakes (down to 217 m and 261 m in lakes Thun and Brienz, respectively) (Dönz et al. 2018). In the pelagic habitat C. fatioi aggregates in lakes Thun and Brienz in moderate water depths (Brienz: approx. $1-100 \mathrm{~m}, N=10$; Thun: approx. $10-40 \mathrm{~m}, N=9$ ) (Dönz et al. 2018). Note that the habitat-stratified random sampling data for both lakes only covers a short period of time (one month in late summer) and it is thus not clear how the species are distributed spatially through the rest of the year. Furthermore, the habitat-stratified random sampling in the both lakes did not distinguish between ripe and unripe specimens, and thus in the case of $C$. fatioi the distribution pattern along the depth in the benthic zone is biased by the spawning aggregation of this species since the sampling period in both lakes coincides partially with the spawning season of this species. Coregonus fatioi resembles phenotypically C. albellus and to some extent C. profundus. Interestingly, Steinmann (1950) already mentioned for Lake Thun that the ecotype "Coregonus lavaretus L. nat. arurenis, oekot. nanus" (most likely C. albellus) should be grouped based on its ecology closely to the ecotype "Coregonus lavaretus L. nat. arurenis, oekot. pelagicus" (most likely C. fatioi). Steinmann mentions the German name "Schwebalbock" for the ecotype "pelagicus", which means verbally translated the "floating whitefish" and mentions that the "nanus" ecotype seems to be a small species with similar ecological properties. For the large whitefish species in Lake Thun, Steinmann (1950) defined one central ecotype, the "primigenius" ecotoype, which he places - based on the size - with two other ecotypes namely the "litoralis" ecotoype (most likely C. alpinus) and the "pelagicus" ecotype (most likely C. fatioi). Besides referring to a "primigenius" ecotype, Steinmann (1950) also refers to a "primigenius"-group, which most likely comprises of the "pelagicus" and "litoralis" ecotypes. A further indication of this is that he also mentions that yet another ecotype, namely the "profundus" ecotype, can be directly deduced from the
"primigenius" ecotype. Steinmann (1950) further mentions that specimens, which he places in the "primigenius"-group, used to migrate before the construction of water gates (see below) upstream from Lake Thun into the river Aare, which connects Lake Thun with Lake Brienz. Steinmann (1950) mentions that these fish belong to the "primigenius"-group, but did not specify if the migrating population constituted of individuals of the "litoralis" or the "pelagicus" ecotype or both. This migrating population was referred to as "Wanderalbock" (i.e. migrating whitefish) in German and historically migrated from Lake Thun into Lake Brienz during the spawning season, before migration became impossible due to the construction of water gates in 1856 (Fatio 1890; Dönz et al. 2018). Fatio (1890) mentioned that a large part of the population of C. fatioi "disappeared" at the beginning of the spawning season in late August and was caught by fishermen in the river Aare downstream (near the city of Thun or Bern) or upstream (near the city of Interlaken) of Lake Thun before and after the construction of the water gate. We compared six whitefish specimens from the museum collections of the MHNG and NMBE, which had no species designation but where it was mentioned that they were caught in the river Aare near the city of Bern (in the years 1881 and 1895), Thun (in the year 1950) and Interlaken (in the year 1945), to the contemporary specimens of Lake Thun including the type specimens of C. albellus, C. fatioi and C. alpinus. All the specimens were caught after the construction of the water gate, when free movement between the lakes was already constrained. All six specimens from the river Aare group in morphospace within the range or adjacent to the range of the contemporary specimens of $C$. fatioi including the type specimen (Suppl. material 1: Figure S11a-c), suggesting that the historically migrating population of whitefish from Lake Thun most likely belonged to the species C. fatioi. Bittner (2009) sampled and genotyped individuals of a population of whitefish spawning in the river Aare near Interlaken. Dönz et al. (2018) re-analysed those individuals and was able to assign 4 individuals with high assignment probability ( $>70 \%$ ) to several different contemporary species of Lake Thun, namely C. alpinus (individual assignment probability of THL15N18 $=86 \%$ ), C. acrinasus (ind. assign. prob. of THL15N07 and THL15N23 $=77 \%$ and $80 \%$, respectively) and C. fatioi (ind. assign. prob. of THL15NfS1124 = 92\%). This suggest either that historically more species than just C. fatioi migrated to the river Aare for spawning and were missed both by Fatio (1890) and Steinmann (1950) and are thus not represented in our PCA morphospace of Aare river whitefish (Suppl. material 1: Figure S11a-c). Or the historical migratory population consisted - as has been suggested by Fatio (1890) and Steinmann (1950) - of individuals of C. fatioi. The average size (total length) at 3 years of age for specimens in this study is $266 \pm 15 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=14)$ and $244 \pm 14 \mathrm{~mm}(\mathrm{~N}=16)$ for lakes Thun and Brienz respectively (Suppl. material 1: Figures S4-S6). In Lake Brienz the size of 3-year-old specimens of C. fatioi is considerably larger than that of C. albellus and similar to that of $C$. alpinus and $C$. brienzii, whereas in Lake Thun it is similar to that of C. profundus and C. albellus and smaller than that of C. alpinus, C. steinmanni, and C. acrinasus (Suppl. material 1: Figure S6). Coregonus fatioi has a long spawning season with two peaks. One spawn-
ing peak is in late summer/early autumn from August to October, which seems more common in Lake Thun than Lake Brienz, and the second peak is in early to late winter from December to March (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). Spawning depth varies with spawning season and can range from approx. 40 m down to the max. depth of 210 m and 261 m in lakes Thun and Brienz, respectively (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). Occasionally C. fatioi can be found spawning shallower (up to 10 m ), but generally it spawns in deeper waters. The spawning season and depth of C. fatioi partially overlaps with that of C. steinmanni, C. albellus, C. acrinasus, and C. profundus in Lake Thun and with that of C. albellus and C. brienzii in Lake Brienz.

Etymology. The name given to this species by Fatio (1890) was preoccupied by another species described by Fatio (1885). Kottelat (1997) proposed C. fatioi as a replacement name. The specific epithet fatioi is the genitive of Fatio. It was named by Kottelat (1997) after the late researcher Viktor Fatio, a zoologist from Switzerland who wrote a standard reference work on the Swiss vertebrates entitled "Faune des Vertébrés de la Suisse Partie 1-3"and in which he also described part of the whitefish species diversity of Switzerland.

Common name. Tiefenalbock in Lake Thun and Felchen in Lake Brienz.

## Coregonus steinmanni Selz, Dönz, Vonlanthen \& Seehausen, sp. nov. http://zoobank.org/C03A9DA8-8492-4CBF-B87B-406D72594530 <br> Figure 7

Coregonus "Albock": Rufli 1978, 1979; Kirchhofer and Tschumi 1986; Kirchhofer 1995 (see also synonymy of C. alpinus and C. acrinasus)
Coregonus "Balchen": Heuscher 1901; Surbeck 1917 (see also synonymy of C. alpinus)
Coregonus"Balchen", "THU2": Douglas et al. 1999, 2003; Douglas and Brunner 2002 (see also synonymy of C. alpinus)
Coregonus lavaretus natio arurensis, oekot. primigenius: Steinmann 1950 (see also synonymy of C. fatioi and C. alpinus)
Coregonus sp. "Balchen": Hudson et al. 2011, 2013, 2016; Ingram et al. 2012; Vonlanthen et al. 2012, 2015; Vonlanthen and Périat 2013 (see also synonymy of C. alpinus and C. brienzii)
Coregonus sp. "Balchen 2": Dönz et al. 2018 (see also synonymy of C. brienzii)
Coregonus "Wanderalbock", "Bodenalbock", "Albock": Steinmann 1950 (see also synonymy of C. alpinus, C. fatioi, C. steinmanni and C. brienzii)

Material examined. Holotype. NMBE-1077219, Switzerland, Lake Thun (4640'N, $7^{\circ} 46$ E), 301 mm SL, female.

Paratypes. NMBE-1077132, NMBE-1077212-1077218, NMBE-1077220, NMBE-1077262-1077265, Switzerland, Lake Thun (4640'N, $7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=13$, 211-323 mm SL.

Table 6. Morphological and meristic data of C. steinmanni from Lake Thun, Switzerland, NMBE1077219, female, holotype from Lake Thun; paratypes $N=12$. For females and for both sexes the range and mean include the holotype.

| Morphological characters | $\begin{gathered} \hline \text { C. steinmanni } \\ \hline \text { Holotype } \end{gathered}$ | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=12$ |  | $N$-females $=3$ |  | $N$-males $=9$ |  |
|  |  | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range |
| SL (mm) | 301 | $275.3 \pm 29.4$ | (211-323) | $276.5 \pm 36.9$ | (234-301) | $274.9 \pm 29.2$ | (211-323) |
| Percentage of standard length |  |  |  |  |  |  |  |
| PelvFB | 4.0 | $4.4 \pm 0.3$ | (4.0-4.8) | $4.1 \pm 0.2$ | (4.0-4.3) | $4.5 \pm 0.3$ | (4.1-4.8) |
| PelvFS | 5.7 | $6.2 \pm 0.5$ | (5.3-6.9) | $6.2 \pm 0.4$ | (5.7-6.5) | $6.2 \pm 0.6$ | (5.3-6.9) |
| PelvF | 15.3 | $16.5 \pm 1.1$ | (14.6-18.3) | $16.1 \pm 1.1$ | (15.3-17.4) | $16.6 \pm 1.1$ | (14.6-18.3) |
| PecFB | 3.2 | $3.4 \pm 0.3$ | (3.1-3.8) | $3.2 \pm 0.2$ | (3.1-3.4) | $3.4 \pm 0.3$ | (3.1-3.8) |
| PecF1 | 14.7 | $16.2 \pm 1.3$ | (13.9-18.2) | $15.8 \pm 1.1$ | (14.7-16.9) | $16.4 \pm 1.4$ | (13.9-18.2) |
| PecF2 | 15.2 | $17 \pm 1.3$ | (15.2-19.1) | $16.2 \pm 1$ | (15.2-17.2) | $17.3 \pm 1.3$ | (15.5-19.1) |
| DFB | 11.4 | $12.6 \pm 0.8$ | (11.4-13.8) | $12.4 \pm 1.2$ | (11.4-13.7) | $12.6 \pm 0.7$ | (11.7-13.8) |
| DFAe | 16.2 | $18.8 \pm 1.7$ | (16.2-21.2) | $18.4 \pm 2.3$ | (16.2-20.9) | $19.0 \pm 1.6$ | (16.2-21.2) |
| DFAd | 17.8 | $20.1 \pm 1.6$ | (17.5-22.4) | $20.0 \pm 2.3$ | (17.8-22.4) | $20.2 \pm 1.4$ | (17.5-22.1) |
| DFPe | 4.8 | $4.8 \pm 0.7$ | (3.9-6.3) | $4.6 \pm 0.2$ | (4.4-4.8) | $4.9 \pm 0.7$ | (3.9-6.3) |
| AFB | 12.1 | $12.6 \pm 0.8$ | (11.5-14.2) | $12.6 \pm 0.5$ | (12.1-13.0) | $12.6 \pm 0.9$ | (11.5-14.2) |
| AFAe | 11.2 | $12.4 \pm 1.0$ | (10.8-13.7) | $12.3 \pm 1.3$ | (11.2-13.7) | $12.4 \pm 0.9$ | (10.8-13.5) |
| AdFB | 5 | $4.5 \pm 0.6$ | (3.7-5.4) | $4.4 \pm 0.5$ | (4.0-5.0) | $4.5 \pm 0.6$ | (3.7-5.4) |
| CF | 23.6 | $23.4 \pm 1.2$ | (22.2-25.9) | $23.7 \pm 1.2$ | (22.4-24.9) | $23.3 \pm 1.3$ | (22.2-25.9) |
| CD | 7.8 | $8 \pm 0.4$ | (7.5-8.6) | $7.9 \pm 0.2$ | (7.7-8.1) | $8 \pm 0.4$ | (7.5-8.6) |
| CL | 13 | $13.0 \pm 0.7$ | (11.4-14.0) | $13.5 \pm 0.5$ | (13.0-14.0) | $12.9 \pm 0.7$ | (11.4-13.9) |
| PAdC | 18.2 | $18 \pm 1$ | (16.4-19.6) | $17.8 \pm 0.8$ | (16.9-18.4) | $18 \pm 1.1$ | (16.4-19.6) |
| DHL | 13.6 | $14.0 \pm 0.7$ | (13.2-15.1) | $14.3 \pm 0.7$ | (13.6-14.9) | $13.9 \pm 0.7$ | (13.2-15.1) |
| PreP | 53.4 | $51.7 \pm 1.9$ | (48.6-54.3) | $52.8 \pm 1.2$ | (51.4-53.6) | $51.3 \pm 2.0$ | (48.6-54.3) |
| PreA | 78.1 | $77.5 \pm 0.9$ | (75.0-78.4) | $78.0 \pm 0.6$ | (77.4-78.4) | $77.3 \pm 0.9$ | (75.0-77.9) |
| PreD | 50 | $47.2 \pm 1.5$ | (44.5-50) | $48.2 \pm 1.7$ | (46.7-50) | $46.9 \pm 1.4$ | (44.5-49.7) |
| BD | 30 | $27.0 \pm 1.5$ | (24.6-30.0) | $28.0 \pm 1.8$ | (26.5-30.0) | $26.7 \pm 1.3$ | (24.6-28.7) |
| PostD | 43.3 | $43.3 \pm 1.2$ | (41.9-45.6) | $42.5 \pm 0.8$ | (41.9-43.3) | $43.6 \pm 1.2$ | (42-45.6) |
| TL | 120.1 | $119.6 \pm 2.3$ | (115.3-122.5) | $119.5 \pm 0.6$ | (118.8-120.1) | $119.6 \pm 2.7$ | (115.3-122.5) |
| HL (mm) | 58.7 | $55.3 \pm 4.9$ | (44.8-63.3) | $55.6 \pm 5.3$ | (49.4-58.7) | $55.2 \pm 5.1$ | (44.8-63.3) |
| Percentage of head length |  |  |  |  |  |  |  |
| SN | 22.2 | $23.2 \pm 1.7$ | (20.5-26.3) | $23.5 \pm 1.2$ | (22.2-24.6) | $23.1 \pm 1.9$ | (20.5-26.3) |
| ED | 22.2 | $22 \pm 1.1$ | (20.5-24.5) | $22.6 \pm 1.7$ | (21.1-24.5) | $21.8 \pm 0.8$ | (20.5-23) |
| EC | 25.5 | $26.2 \pm 1.2$ | (24.2-27.8) | $26.3 \pm 1.3$ | (25.5-27.8) | $26.2 \pm 1.2$ | (24.2-27.4) |
| EH | 22.5 | $21.6 \pm 1.1$ | (19.6-24.1) | $22.5 \pm 1.5$ | (21-24.1) | $21.3 \pm 0.8$ | (19.6-22) |
| ES | 5.1 | $4.8 \pm 0.6$ | (3.9-5.6) | $4.9 \pm 0.5$ | (4.3-5.2) | $4.8 \pm 0.7$ | (3.9-5.6) |
| PostO | 54.4 | $52.4 \pm 1.4$ | (50.3-54.4) | $53 \pm 1.4$ | (51.6-54.4) | $52.2 \pm 1.5$ | (50.3-54.3) |
| HD | 72.1 | $72.1 \pm 2.1$ | (68.9-76.3) | $72.8 \pm 0.9$ | (72.1-73.8) | $71.8 \pm 2.4$ | (68.9-76.3) |
| MW | 10.7 | $9.3 \pm 0.7$ | (8.3-10.7) | $9.7 \pm 0.8$ | (9.1-10.7) | $9.2 \pm 0.7$ | (8.3-10.6) |
| UJ | 27 | $27.3 \pm 1.4$ | (25.2-30) | $27.3 \pm 0.7$ | (26.9-28.1) | $27.3 \pm 1.6$ | (25.2-30) |
| LJ | 39.4 | $39 \pm 1.2$ | (36.6-40.4) | $39.7 \pm 0.3$ | (39.4-40) | $38.7 \pm 1.3$ | (36.6-40.4) |
| M | 19.7 | $19.7 \pm 1.2$ | (18.1-21.8) | $19.4 \pm 0.7$ | (18.6-19.9) | $19.8 \pm 1.3$ | (18.1-21.8) |
| SD | 10.4 | $10 \pm 1.7$ | (6.5-13.2) | $10.1 \pm 0.4$ | (9.7-10.4) | $10 \pm 2$ | (6.5-13.2) |
| SW | 15.8 | $16.7 \pm 1.1$ | (15.3-18.9) | $16 \pm 0.8$ | (15.3-17) | $16.9 \pm 1.1$ | (15.7-18.9) |
| HW | 53.1 | $51.6 \pm 3.1$ | (44.5-56.9) | $49 \pm 4.3$ | (44.5-53.1) | $52.4 \pm 2.2$ | (49.5-56.9) |
| IOW | 29.6 | $27.6 \pm 2.3$ | (23.8-31.2) | $27.9 \pm 2.2$ | (25.4-29.6) | $27.5 \pm 2.4$ | (23.8-31.2) |
| INW | 11.6 | $12.1 \pm 0.7$ | (11-13.2) | $11.7 \pm 0.1$ | (11.6-11.8) | $12.3 \pm 0.7$ | (11-13.2) |
| LJW | 14.3 | $11.9 \pm 1.4$ | (9.7-14.3) | $12 \pm 2.3$ | (9.7-14.3) | $11.9 \pm 1.1$ | (10.1-13.6) |
| UJW | 24.1 | $23 \pm 1.6$ | (19.3-25) | $21.6 \pm 2.4$ | (19.3-24.1) | $23.4 \pm 1.1$ | (21.2-25) |
| MGR | 11.3 | $11.5 \pm 1.7$ | (9.1-14.3) | $11.3 \pm 1.1$ | (10.2-12.4) | $11.5 \pm 1.9$ | (9.1-14.3) |


| Morphological characters | $\begin{gathered} \hline \text { C. steinmanni } \\ \hline \text { Holotype } \end{gathered}$ | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=12$ |  | $N$-females $=3$ |  | $N$-males $=9$ |  |
|  |  | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range |
| LGR | 11.7 | $12.1 \pm 1.5$ | (10-14.4) | $11.6 \pm 1.2$ | (10.4-12.9) | $12.3 \pm 1.6$ | (10-14.4) |
| UA | 19.6 | $18.6 \pm 0.6$ | (17.8-19.8) | $18.9 \pm 0.6$ | (18.4-19.6) | $18.6 \pm 0.6$ | (17.8-19.8) |
| LA | 34.7 | $34.3 \pm 1.2$ | (31.6-36.5) | $33.9 \pm 0.8$ | (33-34.7) | $34.4 \pm 1.3$ | (31.6-36.5) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 10 | 10 | (10-12) | na | (10-12) | 10 | (10-12) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 15 | 15 | (14-16) | na | (14-16) | 15 | (15-16) |
| DF unbranched | 4 | 4 | (3-4) | 4 | (3-4) | 4 | (3-4) |
| DF branched | 10 | 10 | (10-12) | 10 | (10-11) | 10 | (10-12) |
| AF unbranched | 3 | 3 | (3-3) | 3 | (3-3) | 3 | (3-3) |
| AF branched | 11 | 12 | (11-13) | 12 | (11-12) | 12 | (11-13) |
| LS | 78 | 78 | (78-87) | 78 | (78-80) | 85 | (78-87) |
| PDS | 40 | 36 | (32-40) | na | (32-40) | 35 | (33-40) |
| TDS | 9 | 10 | (8-10) | 9 | (9-10) | 10 | (8-10) |
| TAS | 8 | 8 | (8-9) | 8 | (8-8) | 8 | (8-9) |
| TPS | 8 | 9 | (8-9) | 8 | (8-8) | 9 | (8-9) |
| UGR | 10 | 11 | (10-12) | 11 | (10-11) | 12 | (10-12) |
| LGR | 20 | 20 | (19-23) | 20 | (20-21) | 21 | (19-23) |
| total GR | 30 | 31 | (30-35) | na | (30-32) | 31 | (30-35) |

Diagnosis. Coregonus steinmanni is a large whitefish with moderate pigmentation of all fins and body; light to dark greenish blue colour on the flanks above the lateral line; moderate to many pigmented small dots on the scales along the flank and the dorsum; deep bodied; stout caudal peduncle; short head; sub-terminal mouth; small eye with a thick and triangular shaped eye socket.

Differential diagnosis. Coregonus steinmanni occurs only in Lake Thun and we therefore compare the characters of this species specifically with the species of Lake Thun. Differential diagnoses against C. albellus, C. alpinus, and C. fatioi are given under those species' accounts.

## Coregonus steinmanni-Coregonus profundus

Coregonus steinmanni can be distinguished from C. profundus by having more and longer gill rakers (upper arch gill raker number: $10-12$, mode $=11$ vs. $5-10$, mode $=$ 9; lower arch gill raker number: $19-23$, mode $=20$ vs. $10-18$, mode $=14$; total gill raker number: $30-35$, mode $=31$ vs. $15-27$, mode $=21$; middle gill raker length: $9.1-14.3 \% \mathrm{HL}$, mean $=11.5$ vs. $7.6-11.7 \% \mathrm{HL}$, mean $=9.2$; longest gill raker length: $10-14.4 \% \mathrm{HL}$, mean $=12.1$ vs. $7.8-12.4 \% \mathrm{HL}$, mean $=10.1$ ), shorter pectoral fin (pectoral fin 1 length: 13.9-18.2\% SL, mean $=16.2$ vs. $16.6-21 \%$ SL, mean $=18.4$; pectoral fin 2 length: $15.2-19.1 \%$ SL, mean $=17$ vs. $17.7-23.2 \%$ SL, mean $=20.2$ ), a shorter head $(13.2-15.1 \% \mathrm{SL}$, mean $=14$ vs. $15.5-18.4 \% \mathrm{SL}$, mean $=16.4)$, a smaller eye cavity ( $24.2-27.8 \%$ HL, mean $=26.2$ vs. $26.2-32.1 \%$ HL, mean $=29.2$ ), a narrower underjaw (19.3-25, mean $=23 \%$ HL vs. $22.7-29.2 \% \mathrm{HL}$, mean $=26$ ), and a shorter prepelvic distance $(48.6-54.3 \%$ SL, mean $=51.7$ vs. $51.2-58.1 \%$ SL,


Figure 7. Coregonus steinmanni, Lake Thun, Switzerland A holotype, NMBE-1077219, Lake Thun, 301 mm SL, female, freshly caught specimen B, C NMBE-1077219, holotype, preserved specimen D paratype, NMBE-1077214, Lake Thun, 234 mm SL, female, freshly caught specimen. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.
mean $=54.2$ ). Based on ratios C. steinmanni can be differentiated from C. profundus by having a larger "caudal fin depth / dorsal head length" ratio ( $0.53-0.63$ vs. $0.4-0.49$ ) (Tables 6, 8, 10, Suppl. material 1: Table S6).

## Coregonus steinmanni-Coregonus acrinasus

Coregonus steinmanni differs from C. acrinasus by having a shorter maxilla (18.1$21.8 \% \mathrm{HL}$, mean $=19.7$ vs. $19.4-23.8 \% \mathrm{HL}$, mean $=21.8)($ Tables 6,8$)$ and can be differentiated based on ratios from C. acrinasus by having a larger "caudal peduncle depth / maxilla length" ratio (1.86-2.24 vs. 1.4-1.9) (Tables 6, 9, 10).

Description. General appearance is shown in Figure 7. Morphological and meristic characters of both sexes can be found in Table 6 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Table 10. The description is valid for both sexes.

Shape: Generally deep bodied with greatest body depth anterior of the dorsal fin. Dorsal profile strongly arched compared to ventral profile. Dorsal profile from the tip of snout to the anterior origin of dorsal fin moderate to strongly convex, whereas the ventral profile is slightly arched such that it is almost straight or slightly convex from the interorbital area to the pelvic fin origin. Mouth is rather thin (i.e., width of upper and lower jaw), short and sub-terminal. Snout is pronounced and almost equally wide as deep resulting in an almost square shape. Small eye. Eyesocket is thick and triangular (i.e., sickle-shaped). Pectoral fin moderately tapered. The anterior unbranched ray of the erected dorsal fin has an approx. $60^{\circ}$ angle to body axis and at the end of the ray it is bent posteriorly. Caudal peduncle is stout and short. Caudal fin forked and sometimes slightly asymmetrical with the dorsal part being longer. Un-branched ray of anal fin mostly straight and only sometimes slightly bent posteriorly. Anal fin longest anteriorly and progressively shortening posteriorly with the outermargin of the anal fin mostly slightly concave and only rarely straight.

Meristics: Few and short gill rakers.
Colour: Pigmentation of fins and body overall moderately strong in live specimens. Pectoral fin is moderately to strongly pigmented. Dorsal, adipose, pelvic, anal, and caudal fins are moderately to strongly pigmented. Silvery appearance along the flanks with moderate to many pigmented small dots on the scales. The dots are found along the flank and the dorsum. Distribution of the dots is bound to the scale patterning such that the dots are found at the edge of the scales or at the boundary point of two scales (as can be found for the species of C. fatioi, C. alpinus and C. brienzii). Dorsally above the lateral line the silvery appearance changes to a light (e.g., RGB $(135,236,179)$ ) or darker greenish blue colour (e.g., RGB $(7,168,125))$. Dorsal part of the head strongly pigmented. Snout around the nostrils strongly pigmented with a gap of moderate pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. Pre-operculum and operculum are silvery with one black dot on the lower margin of the pre-operculum. For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. Silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB $(239,210,40)$ ), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus steinmanni is found in Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ), which is connected to Lake Brienz through the river Aare at Interlaken. Based on isotopic signatures C. steinmanni feeds on a mix of benthic prey and zooplankton (Selz 2008; Hudson 2011; Ingram et al. 2012) and has a fast growth rate (Bittner 2009). It has to be noted that the work by Selz (2008), Hudson (2011) and Ingram et al. (2012) did not yet separate C. alpinus from C. steinmanni, which are phenotypically difficult to distinguish. Only recently has genetic work by Dönz and colleagues (2018) clearly resolved that these are two distinct species. Thus, the isotopic work by Selz (2008), Hudson (2011) and Ingram et al. (2012) most likely comprises of specimens of both species. The gill raker number of C. steinmanni (more gill rakers) and $C$. alpinus (fewer gill rakers) suggests - based on the functional properties of the number of gill rakers on feeding on different prey items (Lundsgaard-Hansen et al. 2013; Roesch et al. 2013) - that C. steinmanni feeds more on zooplankton and less on benthic prey than C. alpinus, but this assumption needs to be verified in the future with stomach content analyses. Interestingly, the relative species abundances in the pelagic and benthic habitat from a habitat stratified random sampling in Lake Thun (mid-October 2013: Vonlanthen et al. 2015) shows, that $C$. steinmanni is occupying the moderately deep waters of the benthic habitat $(76 \mathrm{~m} ; N=1)$ and the shallow waters of the pelagic habitat ( $8 \mathrm{~m} ; N=1$ ) (Dönz et al. 2018). Coregonus alpinus on the other hand can exclusively be found in shallow water in the benthic habitat (first 13 m ; $N=1$ ) and is completely absent from the pelagic habitat in Lake Thun (Dönz et al. 2018). It is to note that the habitat-stratified random sampling data only covers a short period of time (one month in late summer) and it is thus not clear how the species are distributed spatially through the rest of the year.

Coregonus steinmanni resembles phenotypically C. alpinus and to some extent C. acrinasus. The average size (total length) at 3 years of age for specimens in this study is $328 \pm 23 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=11$ ) (Suppl. material 1: Figures S4-S6). The average size at 3 years of age for the specimens of C. steinmanni from this study is similar to that for the years 2004-2005 ( $338.5 \pm 19 \mathrm{~mm}, N=8$ ) (Bittner et al. unpublished; Vonlanthen et al. unpublished). The size of 3 -year-old specimens of $C$. steinmanni is similar to that of C. alpinus, larger than that of C. acrinasus and considerably larger than that of C. albellus, C. fatioi and C. profundus (Suppl. material 1: Figure S6). Coregonus steinmanni has a short spawning season in late December and only rarely can be found spawning in late autumn (Suppl. material 1: Figure S3; Dönz et al. 2018). Coregonus steinmanni spawns mostly in moderately shallow waters of 10 m down to approx. 120 m (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). The spawning season and depth of C. steinmanni overlaps largely with that of C. acrinasus and C. alpinus and partially with that of C. fatioi. To a much lesser extent the spawning depth and time of $C$. steinmanni also overlaps with that of C. albellus and C. profundus.

Etymology. The specific epithet steinmanni is the genitive of Steinmann. We name this species after the high school teacher and researcher Paul Steinmann, a zoologist from Switzerland who wrote the most comprehensive compendium on Swiss whitefish to date and compiled throughout his lifetime a large collection of preserved specimens
of Swiss, but also European, fishes (Steinmann 1950). This collection and his work on the revision of Swiss whitefish together with work by Fatio (1890) has been essential to describe the whitefish diversity that was present in Switzerland just before or at the beginning of the strong anthropogenic-induced eutrophication of many Swiss lakes which was accompanied by population collapse, speciation reversals, and extinction of Swiss whitefish (Vonlanthen et al. 2012). For example, the only existing specimens of a now-extinct whitefish species, C. gutturosus Gmelin 1818, can only be found in the collection of Paul Steinmann.

Common name. None; this species was not recognized by local fishermen or fisherwomen as distinct from C. alpinus and was thus also called "Balchen". We suggest the German name "Steinmann's Balchen".

## C. brienzii Selz, Dönz, Vonlanthen \& Seehausen, sp. nov. <br> http://zoobank.org/C42663B8-4D34-4499-85D9-259AB7DA204B

## Figure 8

Coregonus "Felchen": Kirchhofer 1990; Kirchhofer 1995 (see also synonymy of C. alpinus and C. fatioi)
Coregonus "Large type": Maurer and Guthruf 2005; Müller et al. 2007 (see also synonymy of C. fatioi and C. alpinus)
Coregonus sp. "Balchen": Hudson et al. 2011, 2013, 2016; Ingram et al. 2012; Vonlanthen et al. 2012, 2015; Vonlanthen and Périat 2013 (see also synonymy of $C$. alpinus and C. steinmanni)
Coregonus sp. "Balchen 2": Dönz et al. 2018 (see also synonymy of C. steinmanni)

Material examined. Holotype. NMBE-1077126, Switzerland, Lake Brienz (4643'N, $7^{\circ} 57^{\prime} \mathrm{E}$ ), 223 mm SL, female.

Paratypes. NMBE-1077116-1077125, NMBE-1077127-1077128, Switzerland, Lake Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ), $\mathrm{N}=12,118-226 \mathrm{~mm}$ SL.

Diagnosis. Coregonus brienzii is a medium-sized whitefish with moderate pigmentation of all fins and body; light to dark greenish blue colour on the flanks above the lateral line; moderate to many pigmented small dots on the scales along the flank and the dorsum; deep bodied; stout caudal peduncle; short head; moderately large eye with a moderately thick and triangular shaped eye socket.

Differential diagnosis. Coregonus brienzii occurs only in Lake Brienz and we therefore compare the characters of this species specifically with the species of Lake Brienz. Differential diagnoses against C. albellus, C. alpinus, and C. fatioi are given under those species' accounts.

Description. General appearance is shown in Figure 8. Morphological and meristic characters of both sexes can be found in Table 7 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Table 11. The description is valid for both sexes.

Table 7. Morphological and meristic data of C. brienzii from Lake Brienz, Switzerland, NMBE1077126, female, holotype; paratypes $\mathrm{N}=12$. For females and for both sexes the range and the mean include the holotype.

| Morphological characters | $\begin{aligned} & \hline \text { C. brienzii } \\ & \hline \text { Holotype } \end{aligned}$ | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=13$ |  | $N$-females $=4$ |  | $N$-males $=9$ |  |
|  |  | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range |
| SL (mm) | 223.0 | $181.5 \pm 37.0$ | (118-226) | $187.8 \pm 47.4$ | (118-223) | $178.7 \pm 34.3$ | (129-226) |
| Percentage of standard length |  |  |  |  |  |  |  |
| PelvFB | 4.1 | $3.7 \pm 0.6$ | (2.8-4.8) | $3.6 \pm 0.6$ | (2.8-4.2) | $3.8 \pm 0.6$ | (2.9-4.8) |
| PelvFS | 6.1 | $6.1 \pm 0.8$ | (4.6-7.4) | $5.8 \pm 0.4$ | (5.1-6.1) | $6.3 \pm 0.9$ | (4.6-7.4) |
| PelvF | 15.2 | $15.6 \pm 1.1$ | (14-17.5) | $15 \pm 0.5$ | (14.6-15.7) | $15.9 \pm 1.2$ | (14-17.5) |
| PecFB | 3.1 | $3.1 \pm 0.2$ | (2.6-3.4) | $3.0 \pm 0.3$ | (2.6-3.2) | $3.1 \pm 0.2$ | (2.8-3.4) |
| PecF1 | 16.0 | $15.9 \pm 1.6$ | (13.9-20.1) | $15.4 \pm 0.7$ | (14.5-16.0) | $16.2 \pm 1.8$ | (13.9-20.1) |
| PecF2 | 17.0 | $16.8 \pm 1.6$ | (14-20.7) | $16.4 \pm 0.7$ | (15.5-17) | $17 \pm 1.9$ | (14-20.7) |
| DFB | 12.3 | $11.8 \pm 0.8$ | (10.4-12.9) | $11.7 \pm 0.4$ | (11.3-12.3) | $11.9 \pm 1.0$ | (10.4-12.9) |
| DFAe | 17.6 | $17.9 \pm 1.2$ | (15.5-19.8) | $17.7 \pm 0.7$ | (16.9-18.6) | $18.0 \pm 1.4$ | (15.5-19.8) |
| DFAd | 18.7 | $18.6 \pm 1.5$ | (15.3-20.8) | $18.3 \pm 0.4$ | (17.8-18.7) | $18.8 \pm 1.8$ | (15.3-20.8) |
| DFPe | 5.1 | $5.2 \pm 0.6$ | (4.2-6.5) | $5.0 \pm 0.2$ | (4.7-5.2) | $5.3 \pm 0.7$ | (4.2-6.5) |
| AFB | 13.7 | $12.4 \pm 0.9$ | (11.1-13.7) | $12.9 \pm 1.1$ | (11.4-13.7) | $12.2 \pm 0.7$ | (11.1-13.6) |
| AFAe | 11.1 | $11.2 \pm 1.0$ | (9.4-12.6) | $11.2 \pm 0.5$ | (10.5-11.6) | $11.2 \pm 1.2$ | (9.4-12.6) |
| AdFB | 5.1 | $5.5 \pm 0.8$ | (4.0-7.1) | $5.2 \pm 0.3$ | (5.0-5.7) | $5.6 \pm 0.9$ | (4.0-7.1) |
| CF | 23.8 | $24.1 \pm 1.1$ | (22.6-26.3) | $23.2 \pm 0.5$ | (22.6-23.8) | $24.5 \pm 1.1$ | (22.7-26.3) |
| CD | 7.3 | $7.3 \pm 0.3$ | (6.7-7.7) | $7.1 \pm 0.4$ | (6.7-7.5) | $7.4 \pm 0.2$ | (7.1-7.7) |
| CL | 13.9 | $13.8 \pm 1.0$ | (12.2-15.8) | $14.0 \pm 0.4$ | (13.7-14.6) | $13.7 \pm 1.1$ | (12.2-15.8) |
| PAdC | 18.8 | $19.1 \pm 0.7$ | (17.9-20.7) | $19 \pm 0.4$ | (18.6-19.4) | $19.1 \pm 0.9$ | (17.9-20.7) |
| DHL | 15.0 | $15.6 \pm 0.7$ | (14.6-16.8) | $15.4 \pm 0.6$ | (15.0-16.3) | $15.7 \pm 0.7$ | (14.6-16.8) |
| PreP | 48.6 | $51.1 \pm 1.7$ | (47.8-54.0) | $50.9 \pm 1.8$ | (48.6-52.8) | $51.2 \pm 1.8$ | (47.8-54.0) |
| PreA | 75.3 | $77.1 \pm 1.5$ | (74.3-79.5) | $76.2 \pm 1.7$ | (74.3-78.2) | $77.5 \pm 1.3$ | (75.4-79.5) |
| PreD | 46.2 | $47.5 \pm 1.7$ | (43.9-49.4) | $47.2 \pm 1.1$ | (46.2-48.2) | $47.6 \pm 2$ | (43.9-49.4) |
| BD | 24.6 | $22.6 \pm 1.7$ | (19.6-25.1) | $22.7 \pm 2.5$ | (20.5-25.1) | $22.6 \pm 1.5$ | (19.6-24.2) |
| PostD | 45.9 | $44.1 \pm 1.1$ | (42.4-45.9) | $44.6 \pm 1.3$ | (43-45.9) | $43.9 \pm 1.1$ | (42.4-45.5) |
| TL | 122.0 | $121.5 \pm 1.9$ | (117.8-124.4) | $121.2 \pm 2.5$ | (117.8-123.8) | $121.6 \pm 1.7$ | (119.2-124.4) |
| HL (mm) | 45.4 | $38.7 \pm 7.3$ | (26.7-47.4) | $39.2 \pm 8.5$ | (26.7-45.4) | $38.5 \pm 7.3$ | (28.3-47.4) |
| Percentage of head length |  |  |  |  |  |  |  |
| SN | 25.6 | $23.3 \pm 1.8$ | (20.5-26.3) | $23.6 \pm 2.1$ | (21.1-25.6) | $23.2 \pm 1.7$ | (20.5-26.3) |
| ED | 24.4 | $25.3 \pm 1.6$ | (23.1-28.3) | $25.2 \pm 1.6$ | (24.2-27.6) | $25.3 \pm 1.7$ | (23.1-28.3) |
| EC | 27.8 | $29 \pm 2.3$ | (25.6-32.9) | $28.8 \pm 3.1$ | (25.6-32.9) | $29.1 \pm 2.1$ | (26.5-32.7) |
| EH | 22.0 | $24.4 \pm 1.4$ | (22-27.2) | $23.9 \pm 1.7$ | (22-26.2) | $24.7 \pm 1.3$ | (23-27.2) |
| ES | 3.5 | $4.7 \pm 1.2$ | (3.3-7.2) | $4.8 \pm 1.4$ | (3.5-6.5) | $4.7 \pm 1.2$ | (3.3-7.2) |
| PostO | 50.9 | $50.7 \pm 1.1$ | (48.2-52.3) | $49.8 \pm 1.5$ | (48.2-51.1) | $51.1 \pm 0.7$ | (50.3-52.3) |
| HD | 75.2 | $68.5 \pm 3.3$ | (64.4-75.2) | $69.8 \pm 4.4$ | (65.2-75.2) | $67.9 \pm 2.8$ | (64.4-73.1) |
| MW | 9.7 | $9.9 \pm 0.9$ | (8.5-10.9) | $9.4 \pm 0.8$ | (8.5-10.3) | $10.1 \pm 0.8$ | (8.6-10.9) |
| UJ | 30.2 | $29.5 \pm 1.6$ | (27.1-32) | $29 \pm 1.8$ | (27.1-30.8) | $29.6 \pm 1.6$ | (27.3-32) |
| LJ | 42.9 | $42.2 \pm 1.5$ | (40.5-45.7) | $43.2 \pm 1.7$ | (42-45.7) | $41.8 \pm 1.2$ | (40.5-43.7) |
| M | 23.4 | $21 \pm 2.4$ | (15.4-24) | $21 \pm 3.9$ | (15.4-24) | $21.1 \pm 1.7$ | (18.3-23.8) |
| SD | 7.0 | $8.8 \pm 1.4$ | (6.4-11.6) | $7.4 \pm 0.9$ | (6.4-8.6) | $9.4 \pm 1.2$ | (8-11.6) |
| SW | 18.0 | $17.8 \pm 1.2$ | (15.7-20.2) | $17.6 \pm 0.6$ | (16.7-18) | $17.8 \pm 1.4$ | (15.7-20.2) |
| HW | 52.1 | $48.1 \pm 3.1$ | (44.1-52.4) | $48.5 \pm 4$ | (44.1-52.1) | $47.9 \pm 2.9$ | (44.1-52.4) |
| IOW | 28.4 | $26.2 \pm 1.9$ | (22.8-30.7) | $26.3 \pm 1.7$ | (25-28.4) | $26.1 \pm 2.1$ | (22.8-30.7) |
| INW | 9.7 | $11.1 \pm 0.8$ | (9.7-12.6) | $10.8 \pm 0.7$ | (9.7-11.2) | $11.3 \pm 0.9$ | (10-12.6) |
| LJW | 14.1 | $11.5 \pm 1.2$ | (10.1-14.1) | $11.9 \pm 2$ | (10.1-14.1) | $11.3 \pm 0.6$ | (10.5-12.4) |
| UJW | 25.9 | $23.4 \pm 1.6$ | (20.2-26.1) | $23.3 \pm 2.3$ | (20.2-25.9) | $23.5 \pm 1.3$ | (21.4-26.1) |
| MGR | 13.5 | $13.5 \pm 1.3$ | (10.9-15.1) | $13.7 \pm 1.6$ | (11.6-15.1) | $13.3 \pm 1.3$ | (10.9-14.9) |
| LGR | 13.9 | $14.7 \pm 1.6$ | (12.1-16.8) | $14.8 \pm 2.2$ | (12.1-16.8) | $14.7 \pm 1.4$ | (13-16.7) |
| UA | 20.4 | $18.5 \pm 1.7$ | (15.3-20.5) | $19.6 \pm 0.8$ | (18.5-20.4) | $18 \pm 1.7$ | (15.3-20.5) |
| LA | 40.4 | $35.5 \pm 2$ | (33-40.4) | $37.2 \pm 2.5$ | (35-40.4) | $34.8 \pm 1.4$ | (33-37.5) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 10 | 10 | (9-11) | 10 | (9-10) | 10 | (10-11) |


| Meristic characters | $\begin{array}{\|l} \hline \text { C. brienzii } \\ \hline \text { Holotype } \end{array}$ | Lake Brienz |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=13$ |  | $N$-females $=4$ |  | $N$-males $=9$ |  |
|  |  | Mode | Range | Mode | Range | Mode | Range |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 15 | 15 | (15-17) | 15 | (15-17) | 15 | (15-17) |
| DF unbranched | 4 | 4 | (3-4) | 4 | (4-4) | 4 | (3-4) |
| DF branched | 12 | 11 | (10-13) | 11 | (10-12) | 10 | (10-13) |
| AF unbranched | 4 | 4 | (3-4) | 4 | (4-4) | 4 | (3-4) |
| AF branched | 13 | 12 | (11-13) | 13 | (11-13) | 12 | (12-12) |
| LS | 89 | 86 | (80-91) | 89 | (80-91) | 86 | (80-88) |
| PDS | 36 | 35 | (32-40) | na | (34-37) | 32 | (32-40) |
| TDS | 9 | 9 | (7-10) | 9 | (7-9) | 9 | (8-10) |
| TAS | 8 | 8 | (7-8) | 8 | (7-8) | 8 | (7-8) |
| TPS | 8 | 8 | (8-9) | 8 | (8-8) | 8 | (8-9) |
| UGR | 14 | 14 | (11-14) | 13 | (13-14) | 12 | (11-14) |
| LGR | 25 | 24 | (20-25) | 24 | (24-25) | 23 | (20-25) |
| total GR | 39 | 37 | (32-39) | 37 | (37-39) | 32 | (32-38) |

Shape: Moderately deep bodied with greatest body depth anterior of the dorsal fin. Dorsal profile moderately arched compared to ventral profile. The dorsal profile from the tip of snout to the anterior origin of dorsal fin is moderately convex, whereas the ventral profile is slightly arched such that is almost straight or slightly convex from the interorbital area to the pelvic fin origin. In some specimens the ventral profile and dorsal profile are similar and only slightly arched. Head moderately short. Mouth is rather thin (i.e., width of upper and lower jaw), moderately short and terminal to sub-terminal. The snout can range from almost equally wide as deep to wider than deep, and is only moderately pronounced, since the tip of the snout can sometimes be fleshy and roundish. Moderately large eye. The eye-socket is thick and triangular (i.e., sickle-shaped). Pectoral fin moderately tapered. The anterior unbranched ray of the erected dorsal fin is almost vertically straight with an approx. $70-80^{\circ}$ angle to the body axis and is only bent slightly posteriorly at the end of the ray. Caudal peduncle is moderately stout and short. Caudal fin forked and sometimes slightly asymmetrical with the dorsal part being longer. Unbranched ray of anal fin mostly straight and only sometimes slightly bent posteriorly. Anal fin longest anteriorly and progressively shortening posteriorly with the outer margin of the anal fin mostly slightly concave and only rarely straight.

Meristics: Many gill rakers that are moderately long.
Colour: Pigmentation of fins and body overall moderate in live specimens. The pectoral fin is mostly translucent and only rarely moderately pigmented at the median to distal parts of the fin. The dorsal, adipose, pelvic, anal, and caudal fins are moderately pigmented. Silvery appearance along the flanks with moderate to many pigmented small dots on the scales. The dots are found along the flank and the dorsum. The distribution of the dots is bound to the scale patterning such that the dots are found at the edge of the scales or at the boundary point of two scales (as can be found for the species of C. alpinus and C. fatioi from both lakes and C. steinmanni from Lake Thun). Dorsally above the lateral line the silvery appearance changes to a light (e.g., RGB $(135,236,179))$ or darker greenish blue colour (e.g., RGB $(7,168,125))$. The dorsal part of the head is moderately pigmented. The snout around the nostrils is


Figure 8. Coregonus brienzii, Switzerland, Lake Brienz A holotype, NMBE-1077126, 223 mm SL, female, freshly caught specimen $\mathbf{B}, \mathbf{C}$ holotype, NMBE-1077126, preserved specimen $\mathbf{D}$ paratype, NMBE1077116, 210.5 mm SL, female.
moderately pigmented with a gap of very weak pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. The pre-operculum and operculum are silvery with one black dot on the lower margin of the pre-operculum. For a comparison
to the main colouration found in the other species see Suppl. material 1: Figure S8. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. The silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB (239, 210, 40)), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus brienzii is found in Lake Brienz ( $46^{\circ} 43^{\prime} \mathrm{N}, 7^{\circ} 57^{\prime} \mathrm{E}$ ) which is connected with Lake Thun through the river Aare at Interlaken. Our previous genetic work (Dönz et al. 2018) suggested that C. brienzii is the same species as C. steinmanni and that it together with the other three species, C. alpinus, C. fatioi, and C. albellus, is present in both lakes. All four species displayed the same genetic relationships in both lakes (i.e., the same hierarchical grouping into distinct genotypic clusters and similar extends of genetic divergence). However, recent analyses of whole-genome data (De-Kayne et al. unpublished) revealed, that specimens of C. steinmanni from Lake Thun do not group with those of C. brienzii, whereas those of the other three species from both lakes do cluster together. Instead the whole genome data suggests that C. steinmanni clusters closer to C. alpinus from Lake Thun - as has previously been shown with genetic data (Dönz et al. 2018) - and that C. brienzii clusters closer to C. fatioi from Lake Brienz. Interestingly, we also find morphological relationships to differ between the lakes; in Lake Thun C. steinmanni groups in morphospace with C. alpinus, whereas in Lake Brienz C. brienzii groups in morphospace with C. fatioi.

Coregonus brienzii most likely feeds on a mix of benthic prey and zooplankton (stomach content: Maurer and Guthruf 2005; Müller et al. 2007; isotopic signatures: Selz 2008; Hudson 2011) and has a moderatly fast growth rate (Müller et al. 2007). It has to be noted that the work by Kirchhofer (1995), Maurer and Guthruf (2005) and Müller et al. (2007) did not distinguish between all species in Lake Brienz and thus lumped different species together into few groups. Maurer and Guthruf (2005) and Müller et al. (2007) differentiated between "small-type" and "large-type" whitefish based on cohort-specific threshold values for length-at age. Based on morphology and ecology Kirchhofer (1995) differentiated in Lake Brienz between "Felchen" (comprising most likely of C. alpinus, C. fatioi and C. brienzii) and "Brienzlig" and "Winter-Brienzlig" (comprising of summer- and winter-spawning specimens of C. albellus). Also, the isotopic work by Selz (2008), and Hudson (2011) did not yet differentiate between C. fatioi and C. brienzii. The relative species abundances in the pelagic and benthic habitat from a habitat-stratified random sampling of Lake Brienz (mid-September 2011: Vonlanthen et al. 2013) shows, that C. brienzii is absent from the benthic habitat and is present in the moderately deep pelagic waters ( $30 \mathrm{~m} ; N=1$ ) ( $\mathrm{Dönz}$ et al. 2018). It is to note that the habitatstratified random sampling data only covers a short period of time (one month in late summer) and it is thus not clear how the species is distributed spatially through the rest of the year. Coregonus brienzii resembles phenotypically C. fatioi. The average size (total length) at 3 years of age for specimens in this study is $254+14 \mathrm{~mm}$ ( $\mathrm{N}=8$ ) (Suppl. material 1: Figures S5, S6). The size at 3 years of age of C. brienzii is similar to that of C. fatioi, slightly smaller than that of C. alpinus and consider-
ably larger than that of C. albellus (Suppl. material 1: Figure S6). Coregonus brienzii has a short spawning season in late December (Suppl. material 1: Figure S3; Dönz et al. 2018). Coregonus brienzii spawns mostly in moderately shallow waters of 10 m down to 60 m and rarely to 100 m (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). The spawning season and depth of C. brienzii overlaps largely with that C. fatioi.

Etymology. The specific epithet brienzii is the genitive of Brienz. We name this species after Lake Brienz, as it is the only endemic whitefish species known for Lake Brienz.

Common name. None. We suggest the German name «Brienzer Kleinbalchen»

## Coregonus profundus Selz, Dönz, Vonlanthen \& Seehausen, sp. nov. http://zoobank.org/6B17CFFD-08A3-4A6E-A4AA-CAE0678370FF

## Figure 9

Coregonus alpinus: Kottelat 1997; Kottelat and Freyhof 2007; Hudson et al. 2011, 2013, 2016; Ingram et al. 2012; Vonlanthen et al. 2012, 2015; Dönz et al. 2018 Coregonus lavaretus natio arurensis, oekot. profundus: Steinmann 1950
Coregonus "Tiefenalbock", "Kropfer": Steinmann 1950
Coregonus "Kropfer": Heuscher 1901
Coregonus "Kropfer": Rufli 1978, 1979; Kirchhofer and Tschumi 1986; Kirchhofer 1995; Bittner et al. 2010 (see also synonymy of C. albellus)
Coregonus"Kropfer", "THU3": Douglas et al. 1999, 2003; Douglas and Brunner 2002

Material examined. Holotype. NMBE-1077208, Switzerland, Lake Thun (4640'N, $7^{\circ} 46^{\prime} \mathrm{E}$ ), 194 mm SL, male.

Paratypes. NMBE-1077161-1077179, NMBE-1077203-1077207, NMBE-1077209-1077211, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=27,188-316 \mathrm{~mm} \mathrm{SL}$.

Diagnosis. Coregonus profundus is a small whitefish species with moderate pigmentation of all fins and the body; brown-orange colouration on the flanks above the lateral line; elongate slender body; long head; large eye with a thick and triangular shaped eye socket; tip of snout is fleshy and roundish; few (15-27) and short gill rakers.

Differential diagnosis. Coregonus profundus occurs only in Lake Thun and we therefore compare the characters of this species specifically with the species of Lake Thun. The differential diagnoses against C. albellus, C. alpinus, C. fatioi, and C. steinmanni are given under those species' accounts. The lower number of gill rakers of C. profundus (total gill raker number: $15-27$, mode $=21$ ) distinguishes this species from all other 5 whitefish species, C. albellus ( $32-44$, mode $=38$ ), C. alpinus (25-34, mode $=30)$, C. fatioi $(32-43$, mode $=38)$, C. steinmanni $(30-35$, mode $=31)$, and C. acrinasus (30-40, mode $=36$ ) (Suppl. material 1: Table S6).

## Coregonus profundus-Coregonus acrinasus

Coregonus profundus can be distinguished from C. acrinasus by having shorter gill rakers (middle gill raker length: $7.6-11.7 \% \mathrm{HL}$, mean $=9.2$ vs. $9.1-16.6 \% \mathrm{HL}$,

Table 8. Morphological and meristic data of C. profundus from Lake Thun, NMBE-1077208, male, holotype; paratypes $\mathrm{N}=27$. For ranges of males and for both sexes, the total range and mean include the holotype.

| Morphological characters | C. profundus Holotype | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=28$ |  | $N$-females $=6$ |  | $N$-males $=22$ |  |
|  |  | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range |
| SL (mm) | 194.0 | $223.3 \pm 26.7$ | (188-316) | $248.7 \pm 42.2$ | (188-316) | $216.3 \pm 16$ | (188-241) |
| Percentage of standard length |  |  |  |  |  |  |  |
| PelvFB | 4.4 | $4.2 \pm 0.3$ | (3.6-5.0) | $4.2 \pm 0.2$ | (4-4.5) | $4.2 \pm 0.4$ | (3.6-5) |
| PelvFS | 7.2 | $6.0 \pm 0.8$ | (4.0-7.2) | $5.7 \pm 1$ | (4-6.8) | $6.1 \pm 0.7$ | (4.8-7.2) |
| PelvF | 16.9 | $17.7 \pm 1.1$ | (15.1-19.6) | $17.3 \pm 0.9$ | (16.5-18.9) | $17.9 \pm 1.1$ | (15.1-19.6) |
| PecFB | 3.5 | $3.7 \pm 0.2$ | (3.2-4.3) | $3.6 \pm 0.2$ | (3.2-3.8) | $3.7 \pm 0.2$ | (3.4-4.3) |
| PecF1 | 16.9 | $18.4 \pm 1.1$ | (16.6-21.0) | $18.1 \pm 1.3$ | (16.6-19.8) | $18.5 \pm 1$ | (16.8-21) |
| PecF2 | 17.8 | $20.2 \pm 1.3$ | (17.7-23.2) | $19.9 \pm 1.5$ | (17.7-22.1) | $20.2 \pm 1.3$ | (17.8-23.2) |
| DFB | 12.6 | $12.5 \pm 0.9$ | (10.5-14.5) | $12.3 \pm 0.7$ | (11.3-13.4) | $12.5 \pm 1$ | (10.5-14.5) |
| DFAe | 18.7 | $19.5 \pm 1.4$ | (15.9-21.9) | $18.7 \pm 1.9$ | (15.9-21.6) | $19.7 \pm 1.2$ | (17-21.9) |
| DFAd | 20.6 | $20.7 \pm 1.3$ | (17.5-23.2) | $19.9 \pm 1.4$ | (17.5-21.5) | $20.9 \pm 1.2$ | (18.3-23.2) |
| DFPe | 5.1 | $5.0 \pm 0.5$ | (3.9-6.1) | $5.1 \pm 0.4$ | (4.5-5.6) | $5 \pm 0.6$ | (3.9-6.1) |
| AFB | 13.5 | $13.2 \pm 1.0$ | (10.8-15.3) | $13.4 \pm 0.8$ | (12.1-14.4) | $13.1 \pm 1.1$ | (10.8-15.3) |
| AFAe | 13.6 | $13.3 \pm 1.0$ | (10.9-14.7) | $12.8 \pm 1$ | (10.9-13.9) | $13.4 \pm 0.9$ | (11.3-14.7) |
| AdFB | 5.1 | $5.3 \pm 0.6$ | (3.8-6.3) | $5.3 \pm 0.4$ | (4.6-5.8) | $5.2 \pm 0.6$ | (3.8-6.3) |
| CF | 24.1 | $24.5 \pm 1.4$ | (21.8-27.8) | $24.3 \pm 2$ | (21.8-27.8) | $24.6 \pm 1.3$ | (22.2-27.8) |
| CD | 7.5 | $7.3 \pm 0.3$ | (6.5-7.9) | $7.5 \pm 0.2$ | (7.2-7.8) | $7.3 \pm 0.3$ | (6.5-7.9) |
| CL | 12.5 | $11.8 \pm 0.7$ | (10.2-13.0) | $12 \pm 0.8$ | (10.9-13) | $11.8 \pm 0.7$ | (10.2-13) |
| PAdC | 16.9 | $18.3 \pm 1.1$ | (15.8-20.1) | $18.5 \pm 0.9$ | (17.1-19.6) | $18.2 \pm 1.1$ | (15.8-20.1) |
| DHL | 16.4 | $16.4 \pm 0.6$ | (15.5-18.4) | $16.2 \pm 0.5$ | (15.5-16.7) | $16.5 \pm 0.6$ | (15.7-18.4) |
| PreP | 55.2 | $54.2 \pm 1.5$ | (51.2-58.1) | $53.3 \pm 1.2$ | (51.2-54.1) | $54.5 \pm 1.4$ | (52.1-58.1) |
| PreA | 79.2 | $78.4 \pm 1.4$ | (75.0-80.6) | $77.8 \pm 1.3$ | (75.8-79.4) | $78.6 \pm 1.4$ | (75-80.6) |
| PreD | 48.5 | $48.3 \pm 1.3$ | (45.8-51.1) | $47.8 \pm 1.8$ | (45.8-50) | $48.5 \pm 1.2$ | (46.9-51.1) |
| BD | 24.4 | $24.2 \pm 1.4$ | (22.1-27.6) | $25.4 \pm 1.3$ | (24-27.6) | $23.9 \pm 1.2$ | (22.1-26.6) |
| PostD | 40.6 | $42.5 \pm 1.5$ | (38.9-44.5) | $43.2 \pm 1.4$ | (41.3-44.5) | $42.3 \pm 1.5$ | (38.9-44.4) |
| TL | 122.2 | $121.3 \pm 1.7$ | (117.3-125.6) | $120.5 \pm 1.1$ | (118.9-121.8) | $121.5 \pm 1.8$ | (117.3-125.6) |
| HL (mm) | 41.2 | $48.9 \pm 5.5$ | (39.8-66.2) | $54.1 \pm 8.7$ | (39.8-66.2) | $47.4 \pm 3.2$ | (41.2-53.7) |
| Percentage of head length |  |  |  |  |  |  |  |
| SN | 23.6 | $23.5 \pm 0.8$ | (21.8-24.8) | $23.3 \pm 0.6$ | (22.5-24) | $23.6 \pm 0.8$ | (21.8-24.8) |
| ED | 23.3 | $23.8 \pm 1.4$ | (21.3-26.2) | $23.7 \pm 1.5$ | (21.9-25.7) | $23.8 \pm 1.4$ | (21.3-26.2) |
| EC | 30.9 | $29.2 \pm 1.4$ | (26.2-32.1) | $28.2 \pm 1.6$ | (26.2-31.1) | $29.5 \pm 1.3$ | (26.9-32.1) |
| EH | 24.5 | $23.6 \pm 0.9$ | (21.8-25.5) | $23.2 \pm 0.7$ | (21.9-23.9) | $23.7 \pm 0.9$ | (21.8-25.5) |
| ES | 5.7 | $4.6 \pm 0.8$ | (3.0-5.9) | $4.3 \pm 0.9$ | (3.5-5.9) | $4.7 \pm 0.7$ | (3-5.7) |
| PostO | 51.1 | $50.9 \pm 1.4$ | (48-54) | $52.2 \pm 1.8$ | (49.2-54) | $50.6 \pm 1$ | (48-52.1) |
| HD | 78.3 | $71.8 \pm 2.8$ | (65.9-78.3) | $73 \pm 2.1$ | (69.4-75.7) | $71.5 \pm 2.9$ | (65.9-78.3) |
| MW | 11.2 | $10 \pm 0.8$ | (8.5-11.7) | $10 \pm 0.5$ | (9.4-10.7) | $10 \pm 0.9$ | (8.5-11.7) |
| UJ | 29.1 | $28.7 \pm 1.2$ | (26.4-30.6) | $28.1 \pm 1.3$ | (26.4-30) | $28.9 \pm 1.1$ | (26.8-30.6) |
| LJ | 41.4 | $39.9 \pm 1.7$ | (37-43.6) | $39.1 \pm 1.4$ | (37-40.9) | $40.1 \pm 1.8$ | (37.2-43.6) |
| M | 24 | $20.7 \pm 1.2$ | (17.3-24) | $20.1 \pm 1.4$ | (17.3-21.2) | $20.8 \pm 1.1$ | (18.7-24) |
| SD | 10.1 | $10 \pm 0.8$ | (8.1-11.3) | $9.7 \pm 0.6$ | (8.8-10.7) | $10 \pm 0.8$ | (8.1-11.3) |
| SW | 17.6 | $15.8 \pm 1.3$ | (12.5-17.8) | $15.3 \pm 1.6$ | (13.7-17.3) | $16 \pm 1.1$ | (12.5-17.8) |
| HW | 57.3 | $52.4 \pm 3.3$ | (46.7-58.6) | $53.1 \pm 3.9$ | (46.7-58.6) | $52.2 \pm 3.1$ | (47.4-57.7) |
| IOW | 28.7 | $28.1 \pm 1.2$ | (26.1-30.3) | $28.9 \pm 1.4$ | (26.5-30.3) | $27.9 \pm 1.1$ | (26.1-29.5) |
| INW | 11.1 | $11.1 \pm 1$ | (8.2-13.3) | $11.7 \pm 1.1$ | (10.3-13.3) | $10.9 \pm 1$ | (8.2-12.5) |
| LJW | 9.3 | $11.7 \pm 2.2$ | (7.8-16.2) | $12.7 \pm 0.7$ | (11.4-13.6) | $11.5 \pm 2.4$ | (7.8-16.2) |
| UJW | 28.9 | $26 \pm 1.7$ | (22.7-29.2) | $25.2 \pm 1.5$ | (22.7-27.4) | $26.2 \pm 1.7$ | (22.8-29.2) |
| MGR | 10 | $9.2 \pm 1.1$ | (7.6-11.7) | $9.4 \pm 1.2$ | (7.6-10.9) | $9.2 \pm 1.1$ | (8-11.7) |
| LGR | 10.7 | $10.1 \pm 1.2$ | (7.8-12.4) | $10.5 \pm 1.6$ | (7.8-12.4) | $9.9 \pm 1.1$ | (8.1-12.3) |
| UA | 19.6 | $18 \pm 1.8$ | (15.5-21.8) | $18.7 \pm 2.4$ | (15.5-21.8) | $17.8 \pm 1.6$ | (15.5-21.2) |
| LA | 35.8 | $34.3 \pm 1.8$ | (30.3-37.7) | $35.1 \pm 2.1$ | (32.9-37.7) | $34.1 \pm 1.7$ | (30.3-36.6) |
| Meristic characters |  | Mode | Range |  |  |  |  |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 10 | 10 | (9-11) | 10 | (10-11) | 10 | (9-11) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |


| Meristic characters | $\begin{array}{\|c\|} \hline \text { C. profundus } \\ \hline \text { Holotype } \\ \hline \end{array}$ | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=28$ |  | $N$-females $=6$ |  | N -males $=22$ |  |
|  |  | Mode | Range | Mode | Range | Mode | Range |
| PecF branched | 16 | 16 | (13-17) | 16 | (16-16) | 16 | (13-17) |
| DF unbranched | 5 | 4 | (3-5) | 4 | (3-4) | 4 | (3-5) |
| DF branched | 10 | 10 | (9-12) | 11 | (10-12) | 10 | (9-11) |
| AF unbranched | 5 | 3 | (2-5) | 2 | (2-4) | 3 | (2-5) |
| AF branched | 11 | 12 | (11-14) | 12 | (12-14) | 12 | (11-13) |
| LS | 83 | 84 | (76-90) | 83 | (80-89) | 84 | (76-90) |
| PDS | 34 | 34 | (32-38) | 32 | (32-37) | 34 | (32-38) |
| TDS | 9 | 9 | (8-10) | 9 | (8-10) | 9 | (8-10) |
| TAS | 8 | 8 | (6-8) | 8 | (8-8) | 8 | (6-8) |
| TPS | 8 | 8 | (7-9) | 9 | (8-9) | 8 | (7-9) |
| UGR | 8 | 9 | (5-10) | 7 | (6-10) | 9 | (5-9) |
| LGR | 13 | 14 | (10-18) | 17 | (11-18) | 14 | (10-18) |
| total GR | 21 | 21 | (15-27) | na | (18-27) | 21 | (15-26) |

mean $=13.4$; longest gill raker length: $7.8-12.4 \% \mathrm{HL}$, mean $=10.1$ vs. $11.4-16.9 \%$ HL , mean $=14.5)$ and a longer head $(15.5-18.4 \% \mathrm{HL}$, mean $=16.4$ vs. $13.8-16.1 \%$ HL, mean $=15.2)($ Tables 8,9$)$.

Description. General appearance is shown in Figure 9. Morphological and meristic characters of both sexes can be found in Table 8 and Suppl. material 1: Table S6 and first- and second-best ratios for both sexes combined can be found in Table 10. The description is valid for both sexes.

Shape: Body elongate. Slender bodied with greatest body depth anterior of the dorsal fin. Dorsal and ventral profile similar and slightly arched. Dorsal and ventral profile from tip of snout to interorbital area mostly straight and then slightly convex to dorsal and pelvic fin origin respectively. Head long. Snout often $60^{\circ}$ angle to the body axis anterior of the eye, such that the profile from the tip of the snout to the vertical projection where the anterior part of the eye crosses the dorsal profile is straight and afterwards slightly convex. Mouth is wide (i.e., width of upper and lower jaw), rather short and mostly strongly sub-terminal and only rarely terminal. Snout is weakly pronounced, since the tip of the snout is often fleshy and roundish. Eye rather large with a large eye cavity and a thick and triangular eye-socket (i.e., sickle-shaped). Pectoral fin long and moderately tapered. Dorsal fin long with the anterior unbranched ray of the erected dorsal fin approx. $70-80^{\circ}$ angle to body axis and only slightly bent posteriorly at the end of the ray. Caudal peduncle narrow and short with caudal fin forked and sometimes moderately to strongly asymmetrical with either the ventral or dorsal part being longer. Unbranched ray of anal fin straight and rarely bent posteriorly at the end of the ray. Anal fin is longest anteriorly and progressively shortening posteriorly with the outer margin of the anal fin slightly concave and only rarely straight.

Meristics: Very few and very short gill rakers.
Colour: Pigmentation of fins and body is overall moderate in live specimens. Pectoral fin is translucent or yellowish in colouration with moderate pigmentation at the median to distal parts of the fin. Dorsal, adipose, pelvic, anal and caudal fins are moderately pigmented. Silvery appearance along the flanks and dorsally above the lateral line the silvery appearance changes to a pale brown-orange colouration (e.g., RGB


Figure 9. Coregonus profundus, Lake Thun, Switzerland A holotype, NMBE-1077208, Lake Thun, 194 mm SL, male, freshly caught specimen B, C holotype, NMBE-1077208, preserved specimen D paratype, NMBE-1077203, Lake Thun, 315.5 mm SL, male E paratype, NMBE-1077166, Lake Thun, 253.5 mm SL, female. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.
$(232,172,52))$ and very rarely the brown-orange colouration can have a hint of light greenish colour (e.g., RGB (136, 245, 205)). Sometimes the colouration above the lateral line is pale rose (e.g., $\operatorname{RGB}(247,187,175))$ and then towards the dorsum becomes a brown-orange. This transition from one colouration to another can also be observed in C. albellus. For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. Dorsal part of the head is moderately pigmented. Snout around the nostrils is moderately pigmented and rarely with a gap of less pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. The operculum and pre-operculum are silvery with one black dot on the lower margin of the pre-operculum. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. Silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB (239, 210, 40)), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus profundus is found in Lake Thun $\left(46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}\right.$ ). It is believed to have been endemic to this lake. Yet, based on matching genetic (microsatellite) and morphological (gill raker number, morphological characters) evidence one ripe specimen of $C$. profundus has been caught by a local fisherman, Stefan Dasen, in 2016 in Lake Biel ( $47^{\circ} 05^{\prime} \mathrm{N}, 7^{\circ} 10^{\prime} \mathrm{E}$ ) (Suppl. material 1: Figure S9). Lake Biel has been artificially connected with Lake Thun through the river Aare since the Jura water correction from 1868-1878, where the river Aare was artificially bypassed downstream from Lake Thun into Lake Biel. For another Lake Thun species, C.albellus, it had been known since at least 2004 that it can be found in Lake Biel (see details in the note on biology for C. albellus) (Bittner 2009; this study Suppl. material 1: Figure S9).

It is important to note that native whitefish species of Lake Biel were only known to spawn in the winter months (Fatio 1885; Steinmann 1950; Rufli 1978), whereas C.profundus as well as C.albellus spawn in late summer and winter. Our study reports the first record of C. profundus in Lake Biel. It is unclear though if C. profundus has established as a self-sustaining population in Lake Biel. So far, we only know of one ripe specimen of C. profundus from Lake Biel, whereas for C. albellus reasonable numbers of ripe specimen have been caught for several years in Lake Biel during what is the normal spawning period (late summer) of this species in lakes Thun and Brienz (Bittner 2009; 2016: Suppl. material 1: Figure S9). Based on isotopic signatures C. profundus feeds on benthic prey items (Selz 2008; Hudson 2011; Ingram et al. 2012) and has a slow growth rate (Bittner et al. unpublished). Interestingly specimens of C. profundus that have been caught on the spawning grounds of C. albellus were often in past-spawning condition and occasional stomach content analysis revealed that these fish had been heavily preying on whitefish eggs (Bittner 2009). Earlier stomach content analysis of C. profundus from the months of October and February of 1971 and 1972, respectively, showed that C. profundus mainly feed on chironomid larvae and occasionally on fish eggs (Rufli 1979). Even earlier stomach content analysis by Steinmann (1950) also show that they feed on chironomid larvae, but also on pisidium and other benthic invertebrates. Habitat-stratified random sampling of Lake Thun (mid-October 2013:

Vonlanthen et al. 2015) shows that C. profundus occupies mostly the moderately deep to the deepest waters in the benthic habitat (approx. $15-210 \mathrm{~m} ; N=16$ ) and rarely the moderately deep pelagic waters (approx. $15-45 \mathrm{~m} ; N=3$ )(Dönz et al. 2018). The habitat-stratified random sampling did not distinguish between ripe and unripe specimens, and thus in the case of $C$. profundus, the distribution pattern along the depth in the benthic zone is biased by the spawning aggregation of this species since the sampling period coincides partially with the spawning season of this species. Coregonus profundus phenotypically resembles superficially $C$. albellus. The average size (total length) at 3 years of age for specimens used in this study is $263 \pm 16 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=11$ ) (Suppl. material 1: Figures S4, S6). The size of 3-year-old specimens of C. profundus is similar to that of C. albellus and C. fatioi, but smaller than that of C. acrinasus and considerably smaller than that of C. alpinus and C. steinmanni (Suppl. material 1: Figure S6). Coregonus profundus has a moderately long spawning season from August to December with one major peak from late August to late September / early October (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). Spawning depth varies with spawning season and can range from approx. 30 m to 150 m (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). The spawning season and depth of C. profundus partially overlaps with that of C. steinmanni, C. fatioi, and C. albellus (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018).

Coregonus profundus is known by the common name "Kropfer" and has previously been described under the name C. alpinus (Kottelat (1997) and Kottelat and Freyhof (2007)). As we explain in detail under the species account of $C$. alpinus, the designated lectotype of C. alpinus is incongruent with the description of the species (with the common name "Kropfer": Kottelat (1997) and Kottelat and Freyhof (2007)). We have thus retained the name C. alpinus for the lectotype designated by Kottelat (1997) and provided a new description of this taxon. For the species otherwise described by Kottelat (1997) and Kottelat and Freyhof (2007) as C. alpinus (with the common name "Kropfer") we designated a new name, C. profundus.

Etymology. The adjective profundus means deep in Latin and is used for C. profundus to describe the species unique ecology of living and breeding in great depths in Lake Thun.

Common name. Kropfer.

## Coregonus acrinasus Selz, Dönz, Vonlanthen \& Seehausen, sp. nov. <br> http://zoobank.org/FEB8CAC5-E55D-4A8C-8E21-94E4DB0E77B2

## Figure 10

Coregonus "Albock": Kirchhofer 1995 (see also synonymy of C. alpinus and C. steinmanni)
Coregonus "Albock", "THU1": Douglas et al. 1999; Douglas and Brunner 2002; Douglas et al. 2003 (see also synonymy of C. fatioi)
Coregonus fatioi: Hudson et al. 2011, 2013, 2016; Ingram et al. 2012; Vonlanthen et al. 2012
Coregonus sp. "Albock": Doenz et al. 2018

Material examined. Holotype. NMBE-1077271, Switzerland, Lake Thun (4640'N, $7^{\circ} 46^{\prime} \mathrm{E}$ ), 239.5 mm SL, male.

Paratypes. NMBE-1077238-1077240, NMBE-1077268-1077270, NMBE-$1077272-1077290$, Switzerland, Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ), $\mathrm{N}=25,197-278 \mathrm{~mm}$ SL.

Diagnosis. Coregonus acrinasus is a medium-sized whitefish with moderate pigmentation of all fins and body; dark greenish blue colour on the flanks above the lateral line; moderate to many pigmented small dots on the scales; tip of the snout pointy; long head; small eye with a thick and triangular shaped eye socket; many and moderately long gill rakers.

Differential diagnosis. Coregonus acrinasus only occurs in Lake Thun and shows ancestry contributions from whitefish of Lake Constance, besides its Lake Thun ancestry. These derive from historically documented introductions of at least two whitefish species (C. wartmanni and C. macrophthalmus) into Lake Thun. Since, historically undocumented introductions of other whitefish from Lake Constance cannot be excluded and since there is no clear genetic assignment of C. wartmanni or C. macrophthalmus as likely source of the allochthonous introgression we compare the characters of this species with those of all whitefish species from Lake Constance and all other whitefish species from Lake Thun. The differential diagnoses against C. albellus, C. alpinus, C. fatioi, C. steinmanni and C. profundus are given under those species' accounts.

## Lake Constance comparison.

## Coregonus acrinasus-all four Lake Constance species

The wider underjaw of C. acrinasus $(9.2-14.3 \% \mathrm{HL}$, mean $=12.2)$ differentiates it from all other species from Lake Constance, C. gutturosus ( $6.8-9.9 \%$ HL, mean $=7.7)$, C. arenicolus $(7.8-8.5 \% \mathrm{HL}$, mean $=8.1)$, C. macrophthalmus $(6.4-8.8 \%$ HL, mean $=8$ ) and $C$. wartmanni $(8.1 \% \mathrm{HL})$ (Tables 9, 12).

## Coregonus acrinasus-Coregonus wartmanni

Coregonus acrinasus differs from C. wartmanni by having a larger eye and eye cavity (eye diameter: $21.6-25.5 \% \mathrm{HL}$, mean $=23.7$ vs. $18.9 \%$ HL; eye cavity: 26-29.6\% HL, mean = 27.7 vs. $23.9 \%$ HL; eye height: $21.7-24.8 \% \mathrm{HL}$, mean $=22.9$ vs. $19 \%$ HL) (Tables 9, 12).

## Coregonus acrinasus-Coregonus macrophthalmus

Coregonus acrinasus differs from C. macrophthalmus by having a wider head (43.9$56.2 \% \mathrm{HL}$, mean $=49.6$ vs. $39.3-43.3 \% \mathrm{HL}$, mean $=41.6)($ Tables 9,12$)$.

## Coregonus acrinasus-Coregonus gutturosus

Coregonus acrinasus differs from C. gutturosus by having more and longer gill rakers (upper arch gill raker number: $10-15$, mode $=13$ vs. $7-9$, mode $=7$; lower arch gill raker number: $20-26$, mode $=24$ vs. $9-12$, mode $=10$; total gill raker number: 30-40, mode $=36$ vs. $16-21$, mode $=19$; middle gill raker length: $9.1-16.6 \% \mathrm{HL}$, mean $=$ 13.4 vs. $4.1-8.7 \% \mathrm{HL}$, mean $=6.9$; longest gill raker length: $11.4-16.9$, mean $=14.5$ vs. $6.7-10.6 \% \mathrm{HL}$, mean $=8.2$ ), a longer lower jaw ( $38.6-47 \% \mathrm{HL}$, mean $=40.9$ vs. $34.3-39.1 \% \mathrm{HL}$, mean $=36.6)$ and a shorter head $(13.8-16.1 \% \mathrm{HL}$, mean $=15.2$ vs. 15.4-18.1\% HL, mean = 16.8) (Tables 9, 12, Suppl. material 1: Table S7).

## Coregonus acrinasus-Coregonus arenicolus

Coregonus acrinasus can be differentiated from C. arenicolus by having more and longer gill rakers (lower arch gill raker number: 20-26, mode $=24$ vs. 13-19;

Table 9. Morphological and meristic data of C. acrinasus from Lake Thun, NMBE-1077271, male, holotype; paratypes $\mathrm{N}=25$. For males and for both sexes, the range and mean include the holotype.

| Morphological characters | $\begin{array}{\|c\|} \hline \text { C. acrinasus } \\ \hline \text { Holotype } \\ \hline \end{array}$ | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=26$ |  | $N$-females $=4$ |  | $N$-males $=22$ |  |
|  |  | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range | Mean $\pm$ Stdev | Range |
| SL (mm) | 239.5 | $237.3 \pm 21.2$ | (197-278) | $235.5 \pm 26.5$ | (197-254) | $237.6 \pm 20.8$ | (197-278) |
| Percentage of standard length |  |  |  |  |  |  |  |
| PelvFB | 5.1 | $4.1 \pm 0.4$ | (3.5-5.1) | $3.7 \pm 0.4$ | (3.5-4.3) | $4.1 \pm 0.4$ | (3.5-5.1) |
| PelvFS | 7 | $6.2 \pm 0.7$ | (4.6-7.5) | $6.2 \pm 0.3$ | (5.7-6.4) | $6.2 \pm 0.7$ | (4.6-7.5) |
| PelvF | 17.4 | $16 \pm 0.9$ | (14.3-17.5) | $15.6 \pm 1.2$ | (14.6-17.2) | $16.1 \pm 0.8$ | (14.3-17.5) |
| PecFB | 3.6 | $3.4 \pm 0.2$ | (3.1-4.0) | $3.2 \pm 0.1$ | (3.1-3.4) | $3.5 \pm 0.2$ | (3.1-4) |
| PecF1 | 17.4 | $15.9 \pm 1.1$ | (13.8-18.2) | $15.6 \pm 1.8$ | (14.1-18.1) | $16 \pm 1$ | (13.8-18.2) |
| PecF2 | 18.6 | $16.9 \pm 1.3$ | (15-19.7) | $16.5 \pm 2.2$ | (15-19.7) | $17 \pm 1.1$ | (15-19) |
| DFB | 14.8 | $12.4 \pm 0.8$ | (11.2-14.8) | $12.3 \pm 0.8$ | (11.5-13.4) | $12.4 \pm 0.9$ | (11.2-14.8) |
| DFAe | 20.9 | $18.1 \pm 1.2$ | (15.7-20.9) | $17.8 \pm 1.5$ | (15.8-19.1) | $18.1 \pm 1.2$ | (15.7-20.9) |
| DFAd | 21.7 | $19.3 \pm 1.1$ | (17.0-21.7) | $19.1 \pm 1.1$ | (18.0-20.3) | $19.3 \pm 1.2$ | (17-21.7) |
| DFPe | 5.5 | $5.0 \pm 0.5$ | (4.0-6.1) | $4.9 \pm 0.5$ | (4.2-5.3) | $5 \pm 0.5$ | (4-6.1) |
| AFB | 13.6 | $12.6 \pm 0.6$ | (11.3-13.6) | $12.6 \pm 0.6$ | (11.9-13.3) | $12.6 \pm 0.6$ | (11.3-13.6) |
| AFAe | 13 | $11.6 \pm 0.8$ | (9.2-13.0) | $11.4 \pm 0.5$ | (11.0-12.2) | $11.7 \pm 0.9$ | (9.2-13) |
| AdFB | 4.5 | $4.7 \pm 0.7$ | (3.7-6.2) | $4.8 \pm 0.6$ | (4.2-5.6) | $4.7 \pm 0.7$ | (3.7-6.2) |
| CF | 24 | $23.3 \pm 0.9$ | (21.5-25.1) | $23.2 \pm 0.2$ | (23-23.4) | $23.4 \pm 1$ | (21.5-25.1) |
| CD | 7.5 | $7.6 \pm 0.4$ | (7.1-8.3) | $7.8 \pm 0.3$ | (7.6-8.2) | $7.6 \pm 0.4$ | (7.1-8.3) |
| CL | 11.7 | $12.8 \pm 0.6$ | (11.7-14.2) | $12.4 \pm 0.4$ | (11.9-12.8) | $12.9 \pm 0.6$ | (11.7-14.2) |
| PAdC | 15 | $18.1 \pm 1.2$ | (15-20.1) | $17.6 \pm 1$ | (16.2-18.4) | $18.2 \pm 1.2$ | (15-20.1) |
| DHL | 14.9 | $15.2 \pm 0.6$ | (13.8-16.1) | $14.9 \pm 0.9$ | (14.0-15.9) | $15.2 \pm 0.5$ | (13.8-16.1) |
| PreP | 50.3 | $52.6 \pm 1.6$ | (49.1-56.8) | $51.9 \pm 0.5$ | (51.4-52.4) | $52.7 \pm 1.8$ | (49.1-56.8) |
| PreA | 78.5 | $77.7 \pm 1.2$ | (75.3-80.3) | $77.1 \pm 0.5$ | (76.5-77.6) | $77.8 \pm 1.3$ | (75.3-80.3) |
| PreD | 45.4 | $47.5 \pm 1.4$ | (45-50.7) | $47.5 \pm 1.1$ | (46.3-48.6) | $47.5 \pm 1.4$ | (45-50.7) |
| BD | 25.6 | $24.7 \pm 1.6$ | (20.7-28.1) | $26.1 \pm 1.6$ | (24.4-28.1) | $24.4 \pm 1.5$ | (20.7-26.7) |
| PostD | 41.2 | $43 \pm 1.3$ | (40.3-45.6) | $42.2 \pm 1.6$ | (41-44.3) | $43.1 \pm 1.3$ | (40.3-45.6) |
| TL | 123.2 | $120.6 \pm 1.7$ | (116-123.2) | $119 \pm 2.5$ | (116-121.5) | $120.8 \pm 1.4$ | (118.2-123.2) |
| HL (mm) | 49 | $49.9 \pm 4$ | (41.5-58.4) | $48.5 \pm 4.7$ | (41.5-51.3) | $50.1 \pm 3.9$ | (41.5-58.4) |
| Percentage of head length |  |  |  |  |  |  |  |
| SN | 23.4 | $23.9 \pm 1.4$ | (20.5-27) | $22.6 \pm 1.8$ | (20.5-24.6) | $24.1 \pm 1.3$ | (21.6-27) |
| ED | 23.2 | $23.7 \pm 0.8$ | (21.6-25.5) | $23.8 \pm 0.8$ | (22.6-24.4) | $23.7 \pm 0.9$ | (21.6-25.5) |
| EC | 27.4 | $27.7 \pm 1$ | (26-29.6) | $28.6 \pm 1$ | (27.2-29.6) | $27.6 \pm 0.9$ | (26-28.8) |
| EH | 22.8 | $22.9 \pm 0.9$ | (21.7-24.8) | $23.6 \pm 1.1$ | (22.2-24.8) | $22.8 \pm 0.8$ | (21.7-24.5) |
| ES | 4.9 | $4.7 \pm 0.8$ | (3.2-6.4) | $5.6 \pm 0.9$ | (4.8-6.4) | $4.5 \pm 0.6$ | (3.2-6.1) |
| PostO | 51 | $50.9 \pm 1.5$ | (48.5-54.1) | $52 \pm 1.8$ | (49.8-54.1) | $50.7 \pm 1.4$ | (48.5-53) |
| HD | 69.8 | $69.1 \pm 2.4$ | (65.1-74.9) | $69.8 \pm 2$ | (67.8-72.5) | $68.9 \pm 2.5$ | (65.1-74.9) |
| MW | 9.7 | $9.8 \pm 0.7$ | (8.1-11.4) | $9.8 \pm 0.6$ | (8.8-10.3) | $9.8 \pm 0.8$ | (8.1-11.4) |
| UJ | 28.8 | $29.4 \pm 1.2$ | (26.7-30.9) | $30.1 \pm 0.8$ | (29.3-30.8) | $29.2 \pm 1.3$ | (26.7-30.9) |
| LJ | 40.5 | $40.9 \pm 1.7$ | (38.6-47) | $40.5 \pm 1.1$ | (39-41.5) | $41 \pm 1.8$ | (38.6-47) |
| M | 21.1 | $21.8 \pm 1$ | (19.4-23.8) | $21.9 \pm 0.9$ | (21.3-23.2) | $21.8 \pm 1$ | (19.4-23.8) |
| SD | 9.8 | $8.6 \pm 1.3$ | (6-11.3) | $9 \pm 1.1$ | (7.9-10.5) | $8.6 \pm 1.3$ | (6-11.3) |
| SW | 17.1 | $16 \pm 1.5$ | (13.1-18.1) | $16.2 \pm 1.3$ | (14.7-17.6) | $15.9 \pm 1.5$ | (13.1-18.1) |
| HW | 53.8 | $49.6 \pm 3.2$ | (43.9-56.2) | $51.1 \pm 2.4$ | (48.2-53.8) | $49.4 \pm 3.3$ | (43.9-56.2) |
| IOW | 25.8 | $27 \pm 2.1$ | (21.3-31.5) | $27.1 \pm 1.4$ | (25.1-28.1) | $27 \pm 2.3$ | (21.3-31.5) |
| INW | 10.8 | $11.7 \pm 1$ | (9.5-13.4) | $11.7 \pm 1.2$ | (10.5-13.3) | $11.7 \pm 1$ | (9.5-13.4) |
| LJW | 12.4 | $12.2 \pm 1.2$ | (9.2-14.3) | $12.5 \pm 1$ | (11-13.4) | $12.1 \pm 1.2$ | (9.2-14.3) |
| UJW | 24 | $22.8 \pm 2.1$ | (18.2-27.5) | $23.7 \pm 2.7$ | (20.4-26.3) | $22.6 \pm 2$ | (18.2-27.5) |
| MGR | 13.1 | $13.4 \pm 1.6$ | (9.1-16.6) | $14.4 \pm 1.9$ | (11.9-16.6) | $13.2 \pm 1.6$ | (9.1-15.1) |
| LGR | 15 | $14.5 \pm 1.4$ | (11.4-16.9) | $15.7 \pm 1.2$ | (14.4-16.9) | $14.3 \pm 1.3$ | (11.4-16.3) |
| UA | 19.7 | $18.1 \pm 1.6$ | (13.5-20.3) | $18.5 \pm 1.6$ | (16.2-19.8) | $18.1 \pm 1.6$ | (13.5-20.3) |
| LA | 36.7 | $34.9 \pm 1.7$ | (32.3-38.9) | $35.9 \pm 2.1$ | (34-38.9) | $34.8 \pm 1.7$ | (32.3-38.4) |
| Meristic characters |  | Mode | Range | Mode | Range | Mode | Range |
| PelvF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PelvF branched | 11 | 10 | (9-12) | 10 | (10-11) | 11 | (9-12) |
| PecF unbranched | 1 | 1 | (1-1) | 1 | (1-1) | 1 | (1-1) |
| PecF branched | 15 | 15 | (13-16) | 16 | (15-16) | 15 | (13-16) |


| Meristic characters | $\begin{gathered} \hline \text { C. acrinasus } \\ \hline \text { Holotype } \end{gathered}$ | Lake Thun |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Both sexes |  |  |  |  |  |
|  |  | $N$-total $=26$ |  | $N$-females $=4$ |  | $N$-males $=22$ |  |
|  |  | Mode | Range | Mode | Range | Mode | Range |
| DF unbranched | 3 | 4 | (3-4) | 3 | (3-4) | 4 | (3-4) |
| DF branched | 12 | 10 | (9-12) | 11 | (10-12) | 10 | (9-12) |
| AF unbranched | 3 | 3 | (2-4) | 3 | (3-3) | 3 | (2-4) |
| AF branched | 13 | 11 | (11-13) | 12 | (11-13) | 11 | (11-13) |
| LS | 80 | 84 | (79-88) | 85 | (84-85) | 80 | (79-88) |
| PDS | 34 | 34 | (33-42) | 34 | (34-41) | 35 | (33-42) |
| TDS | 10 | 10 | (9-10) | 10 | (9-10) | 10 | (9-10) |
| TAS | 8 | 8 | (8-9) | 8 | (8-8) | 8 | (8-9) |
| TPS | 9 | 8 | (8-9) | 9 | (8-9) | 8 | (8-9) |
| UGR | 13 | 13 | (10-15) | na | (10-15) | 13 | (10-14) |
| LGR | 20 | 24 | (20-26) | 24 | (21-24) | 24 | (20-26) |
| total GR | 33 | 36 | (30-40) | 35 | (34-36) | 36 | (30-40) |

total gill raker number: $30-40$, mode $=36$ vs. $22-31$; middle gill raker length: $9.1-16.6 \% \mathrm{HL}$, mean $=13.4$ vs. $9.8-10.6 \% \mathrm{HL}$, mean $=10.2$; longest gill raker length: $11.4-16.9$, mean $=14.5$ vs. $10.9-12 \% \mathrm{HL}$, mean $=11.5$ ), a larger eye (eye diameter: $21.6-25.5 \% \mathrm{HL}$, mean $=23.7$ vs. $17.3-19.6 \% \mathrm{HL}$, mean $=17.7$; eye cavity: $26-29.6 \% \mathrm{HL}$, mean $=27.7$ vs. $24.1-25.7 \% \mathrm{HL}$, mean $=25$; eye height: $21.7-24.8 \% \mathrm{HL}$, mean $=22.9$ vs. $18.8-20.8 \% \mathrm{HL}$, mean $=19.6$ ) and a shorter anal fin $(9.2-13 \%$ HL, mean $=11.6$ vs. $12.9-13.8 \% \mathrm{HL}$, mean $=13.3)($ Tables 9,12 , Suppl. material 1: Table S7).

Description. General appearance is shown in Figure 10. Morphological and meristic characters of both sexes can be found in Tables 9, 12, and Suppl. material 1: Tables S6, S7 and first- and second-best ratios for both sexes combined can be found in Table 10. The description is valid for both sexes.

Shape: Only slightly deep bodied with greatest body depth anterior of the dorsal fin. Dorsal and ventral profile equally arched such that both the dorsal profile from the tip of snout to the anterior origin of dorsal fin and the ventral profile from the interorbital area to the pelvic fin origin are moderately convex. Head long. Mouth (i.e., width of upper and lower jaw) is thick, moderately long and often sub-terminal and only rarely terminal. Rostral plate is mostly wider than deep, not strongly pronounced and the tip of the snout is often pointy in the sagittal plane. Eye-socket thick and triangular (i.e., sickle-shaped). Pectoral fin moderately tapered. Anterior unbranched ray of the erected dorsal fin has an approx. $40-60^{\circ}$ angle to body axis and from the middle to the end of the ray it is moderately bent posteriorly. Caudal peduncle stout and moderately long. Caudal fin forked and sometimes slightly asymmetrical with the ventral part being longer. Unbranched ray of anal fin mostly straight and only sometimes slightly bent posteriorly. Anal fin is longest anteriorly and progressively shortening posteriorly with the outer margin of the anal fin slightly concave.

Meristics: Many and moderately long gill rakers.
Colour: Pigmentation of fins and body overall moderately strong in live specimens. Pectoral fin is mostly transparent to moderately pigmented with a yellowish faint pigmentation and only very rarely strongly pigmented. Dorsal, adipose, pelvic, anal, and caudal fins are moderately to strongly pigmented. Fish have a silvery ap-
pearance along the flanks with moderate to many pigmented small dots on the scales. Dots along the flank and the dorsum. Distribution of the dots is bound to the scale patterning such that the dots are found at the edge of the scales or at the boundary point of two scales. Dorsally above the lateral line the silvery appearance changes to dark greenish blue colour (e.g., RGB $(7,168,125)$ ). The snout around the nostrils is strongly pigmented with a gap of very little pigmentation posteriorly of the nostrils up to the height of the middle of the eyes. Pre-operculum and operculum are silvery with one black dot on the lower margin of the pre- operculum. For a comparison to the main colouration found in the other species see Suppl. material 1: Figure S8. Preserved specimens are pale in colouration with similar pigmentation as described for live specimens. Silvery, translucent, not coloured or unpigmented parts of the body become brown-yellowish (e.g., RGB (239, 210, 40)), whereas the pigmented parts are conserved and the coloured parts (dorsally above the lateral line) become brownish (e.g., RGB (186, 140, 100)).

Distribution and notes on biology. Coregonus acrinasus is found in Lake Thun ( $46^{\circ} 40^{\prime} \mathrm{N}, 7^{\circ} 46^{\prime} \mathrm{E}$ ). Based on isotopic signatures C. acrinasus most likely feeds on a mix of benthic prey and zooplankton (Selz 2008; Hudson 2011; Ingram et al. 2012) and based on the size-at-age data C. acrinasus must have a rather fast growth rate (Suppl. material 1: Figures S4-S6). The gill raker number and length of C. acrinasus (many gill rakers and moderately long gill rakers) suggests, based on the functional properties of the number of gill rakers on feeding on different prey items (Lundsgaard-Hansen et al. 2013; Roesch et al. 2013), that C. acrinasus feeds more on zooplankton and less on benthic prey, but this assumption needs to be verified in the future with stomach content analyses. The relative species abundances in the pelagic and benthic habitat from a habitat-stratified random sampling of Lake Thun (mid-October 2013: Vonlanthen et al. 2015) also points to this.Coregonus acrinasus occupies only the shallow waters of the benthic habitat $(15 \mathrm{~m} ; N=1)$ and the moderately deep pelagic waters (approx. 10-35 m; N = 9) (Dönz et al. 2018). However, the habitat-stratified sampling needs to be treated with caution since it only shows a snapshot in time (one month) of the spatial distribution of this and the other species. Coregonus acrinasus phenotypically resembles to some extent C. alpinus and C. steinmanni. The average size (total length) at three years of age for specimens in this study of C. acrinasus is $304 \pm 21 \mathrm{~mm}$ (mean and standard deviation, $\mathrm{N}=9$ ) (Suppl. material 1: Figures S4, S6).The average size at 3 years of age for the specimens of $C$. acrinasus from this study is similar to that for the years 2004-2005 ( $322.8 \pm 18 \mathrm{~mm}, N=50$ ) (Bittner et al. unpublished; Vonlanthen et al. unpublished). The size of 3-year-old specimens of C. acrinasus is smaller to that of C. alpinus and C. steinmanni and considerably larger than that of C. albellus, C. fatioi, and C. profundus (Suppl. material 1: Figure S6). Coregonus acrinasus has a short spawning season in late December and very rarely have ripe individuals been caught in late autumn or winter (Suppl. material 1: Figure S3; Dönz et al. 2018). Coregonus acrinasus spawns mostly in moderately shallow waters of 10 m down to approx. 100 m (Suppl. material 1: Figure S3; Bittner 2009; Dönz et al. 2018). The spawning season and depth of C. acrinasus overlaps
largely with that of $C$. alpinus, $C$. steinmanni, and $C$. fatioi and to a much lesser extent with that of $C$. albellus and $C$. profundus.

Coregonus acrinasus appears to be a species of partially allochthonous origin, closely related to the radiation of Lake Constance with genetic contributions from Lake Thun. Indications of this situation were seen in several earlier genetic studies (Douglas and Brunner 2002; Douglas et al. 2003; Bittner 2009; Hudson et al. 2011) and this was clearly confirmed with large sample sizes recently (Hudson et al. 2016; Dönz et al. 2018). Historical records mention the stocking of alevins of the Lake Constance endemics $C$. wartmanni and $C$. macrophthalmus into Lake Thun. To fully understand the relationship of $C$. acrinasus to the Lake Constance species, we compared the morphology of $C$. acrinasus with that of all four described species of Lake Constance, C. wartmanni, C. macrophthalmus, C. arenicolus, and the now extinct $C$. gutturosus. Our data clearly reveal C. acrinasus as distinct from all Lake Constance species based on morphological characters. Historical records from Fatio (1890) in his book on Swiss fish (Fatio 1890: Page 123) and from Heuscher (1901) in his report on the biology of lakes Thun and Brienz (Heuscher 1901: Pages 69-70, 103) report several incidences of introductions of whitefish from other lakes. Evidence for additional introductions comes from historical records from a fisheries club that was responsible for the propagation of whitefish in lakes Thun and Brienz before stocking with allochthonous fish was forbidden in Lake Thun (nothing is stated regarding Lake Brienz) in 1946 by the local fisheries authorities (Douglas et al. 2003; Dönz et al. 2018). Since 1991 such introductions were banned in all of Switzerland through federal law (BGF 6 I b). These historical records reveal that in 1888, 1889, and 1934 in Lake Thun and 1892 in Lake Brienz between 20'000 and 750'000 (Lake Thun) and once $39^{\prime} 000$ (Lake Brienz) fry of either C. macrophthalmus (only Lake Thun) or C. wartmanni (both lakes) were stocked. Heuscher (1901: Page 70) further noted that the introductions of 1888, 1889, and 1892 were unsuccessful in both lakes, as fishermen did not catch adult fish of either of the Lake Constance species ever after those introductions. Steinmann (1950) in his monograph on Swiss whitefish diversity did not mention any species from Lake Constance to be present in Lake Thun or in Lake Brienz. Dönz et al. (2018) could recently show with genetic data from scales dating back to 1972 and earlier that C. acrinasus was completely absent in catches of that period. The first qualitative reports of this species in spawning fisheries catches are from around the year 2000 (Douglas et al. 1999; Bittner 2009), and our own genetic data from samples of more than 2000 whitefish from Lake Thun confirm the presence of the species. Based on a recent lake-wide quantitative survey in 2015 Dönz et al. (2018) showed that this species accounts for ca. $10 \%$ of all whitefish in Lake Thun in abundance when based on genetic assignments. Several independent multilocus microsatellite and AFLP data sets suggest that it has genetic contributions from the endemic Lake Thun species and cannot clearly be designated genetically to one of the Lake Constance species (Douglas and Brunner 2002; Douglas et al. 2003; Bittner 2009; Hudson et al. 2011; Hudson et al. 2016; Dönz et al. 2018). This suggests that some individuals of one or several of the introduced species from Lake


Figure 10. Coregonus acrinasus, Lake Thun, Switzerland A holotype, NMBE-1077271, Lake Thun, 239.5 mm SL, male, freshly caught specimen B, C holotype, NMBE-1077271, preserved specimen D paratype, NMBE-1077270, Lake Thun, 270 mm SL, male, freshly caught specimen $\mathbf{E}$ paratype, NMBE-1077279, Lake Thun, 234 mm SL, male, freshly caught specimen. The white scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.

Constance must have successfully reproduced in Lake Thun and hybridized with one or several of the local species.

Etymology. The name C. acrinasus is a combination of the ablative case of the Latin adjective acer resulting in acri, which means pointed and the noun nasus for nose. The name acrinasus refers to a phenotypic feature of this species, which often has a pointed snout when viewed in the sagittal plane.

Common name. Albock

## Lake Constance whitefish species

Coregonus gutturosus, Gmelin, 1818
Figure 11
Material examined. Non-types. NMBE-1076230 (Eawag-246), NMBE1076232 (Eawag-248-1), NMBE-1076233 (N = 6: Eawag-249-1, Eawag-249-2, Eawag-249-3, Eawag-249-4, Eawag-249-5, Eawag-249-6), NMBE-1076232 (N = 2: Eawag-248-2, Eawag-248-3), Switzerland, Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$ ), N $=10,169-292 \mathrm{~mm}$ SL.

Distribution and notes on biology. Coregonus gutturosus used to be endemic to Lake Constance but is now extinct.

Common name. Kilch

## Coregonus arenicolus, Kottelat, 1997

## Figure 11

Material examined. Holotype. NMBE-1076223 (Eawag-239-1), Switzerland, Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$ ), 296 mm SL, sex unknown.

Paratypes. NMBE-1076223 (N = 3: Eawag-239-2,Eawag-239-3, Eawag-239-4), Switzerland, Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$ ), $\mathrm{N}=3,289-314 \mathrm{~mm}$ SL.

Distribution and notes on biology. Coregonus arenicolus is found in the upper and lower basin of Lake Constance.

Common name. Sandfelchen.

## Coregonus macrophthalmus, Nüsslin, 1882

Figure 11
Material examined. Syntypes. MHNG-716.052, MHNG-716.051, MHNG816.02, MHNG-715.094 ( $\mathrm{N}=2$ : MHNG-715.094-1, MHNG-715.094-2), NMBE-1076211 (N = 2: Eawag-227-1, Eawag-227-2), Switzerland, Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$ ), $\mathrm{N}=7,193-235 \mathrm{~mm}$ SL.


Figure I I. Types of the Lake Constance species, Switzerland A Coregonus gutturosus, non-type, NMBE1076232 (Eawag-248-1), 250 mm , sex unknown, preserved specimen B Coregonus arenicolus, holotype, 296 mm, NMBE-1076223 (Eawag-239-1), sex unknown, preserved specimen C Coregonus macrophthalmus, syntype, MHNG-716.052, 215 mm , sex unknown, preserved specimen D Coregonus wartmanni, non-type, NMBE-1076206, 301 mm , female, preserved specimen. The white scale ( 1 cm ) below each fish acts as a reference for the actual size of the specimen.

Table 10. The first- and second-best ratios retrieved from the LDA ratio extractor of either head or body characters (see Table 1) alone or combined, used for pair-wise comparisons of all contemporary specimens from the six whitefish species of Lake Thun. For some comparisons only a subset of characters could be used (a-d); the respective characters that were excluded are listed at the end of the table. Only external characters were used for the LDA comparisons, since internal characters (gill raker and gill arch length) cannot be measured on live specimens, and are thus not informative to assign specimens to species in the field. Additionally, species were combined to find first- and second-best ratios that distinguish one species or a combination of species vs. all other species. For multi-species comparisons, only the comparisons that yielded distinguishing ratios are shown. $\delta$ is a measure of how good shape discriminates in comparison to size (i.e., the smaller $\delta$ the less allometry). Ratios marked with an asterisk * have very little (for the pairwise species comparisons not more than one specimen of one species overlaps with the other species) or no overlap and were thus eligible for use in the species key and the diagnoses.

| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\begin{gathered} \hline \delta \text { (Shape vs. size) } \\ \hline 0.1 \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| head + body | C. albellus vs. C. alpinus | 1: CD/UJ * | 0.96-1.29 | 1.36-1.65 | 18.09 |  |
|  |  | 2: AdFB/ES | 5.54-13.54 | 2.87-5.94 | 17.49 | 0.1 |
| body | C. albellus vs. C. alpinus | 1: CD/DHL * | 0.44-0.54 | 0.54-0.62 | 5.98 | 0.26 |
|  |  | 2: DFB/AdFB | 1.6-2.66 | 2.31-4.02 | 4.86 | 0.31 |
| head | C. albellus vs. C. alpinus | 1: UJ/ES | 6.48-16.01 | 4.52-7.93 | 6 | 0.22 |
|  |  | 2: HL/UJ | 2.88-3.47 | 3.33-4.11 | 5.26 | 0.25 |
| head + body | C. albellus vs. C. fatioi | 1: TL/EH | 21.93-27.57 | 21.91-29.46 | 3.07 | 0.08 |
|  |  | 2: PelvF/PecF1 | 0.87-1.03 | 0.87-1.07 | 2.89 | 0.08 |
| body | C. albellus vs. C. fatioi | 1: PecF1/TL | 0.12-0.16 | 0.11-0.15 | 1.4 | 0.19 |
|  |  | 2: DFAe/DFAd | 0.88-1 | 0.9-0.98 | 1.18 | 0.21 |
| head | C. albellus vs. C. fatioi | 1: EH/HL | 0.21-0.26 | 0.2-0.25 | 1.63 | 0.08 |
|  |  | 2: LJW/ES | 2.14-7.79 | 1.59-8.18 | 1.35 | 0.1 |
| head + body | C. albellus vs. C. steinmanni | 1: CD/UJ * | 0.96-1.29 | 1.36-1.55 | 13.8 | 0.12 |
|  |  | 2: AdFB/ES | 5.54-13.54 | 3.31-6.31 | 13 | 0.12 |
| head | C. albellus vs. C. steinmanni | 1: HL/UJ | 2.88-3.47 | 3.33-3.97 | 5.3 | 0.21 |
|  |  | 2: LJ/ES | 8.25-20.33 | 6.65-12.45 | 4.59 | 0.23 |
| head + body | C. albellus vs. C. profundus | 1: CL/EC * | 1.97-2.87 | 1.56-2.09 | 13.19 | 0.03 |
|  |  | 2: DHL/M | 2.77-3.53 | 3.21-3.79 | 12.79 | 0.03 |
| body | C. albellus vs. C. profundus | 1: CL/DHL | 0.75-1.04 | 0.61-0.82 | 4.43 | 0.06 |
|  |  | 2: CD/BD | 0.26-0.31 | 0.28-0.34 | 3.38 | 0.07 |
| head | C. albellus vs. C. profundus | 1: EC/UJ | 0.74-0.95 | 0.87-1.05 | 5.02 | 0.1 |
|  |  | 2: SW/ES | 3.22-9.67 | 2.31-5.26 | 4.05 | 0.12 |
| head + body | C. albellus vs. C. acrinasus | 1: AdFB/ES | 5.54-13.54 | 3.31-6.5 | 9.13 | 0.06 |
|  |  | 2: CD/UJW | 1.14-1.79 | 1.4-2 | 8.69 | 0.07 |
| body | C. albellus vs. C. acrinasus | 1: PecF1/CD | 2.13-2.76 | 1.8-2.39 | 4.5 | 0.11 |
|  |  | 2: DFB/AdFB | 1.6-2.66 | 1.98-3.45 | 3.88 | 0.13 |
| head | C. albellus vs. C. acrinasus | 1: UJW/ES | 5.27-13.65 | 3.22-7.96 | 4.19 | 0.14 |
|  |  | 2: ED/UJ | 0.66-0.84 | 0.74-0.9 | 3.51 | 0.16 |
| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (shape vs. Size) |
| head + body | C. alpinus vs. C. fatioi | 1: CD/PostD * | 0.17-0.21 | 0.14-0.17 | 22.73 | 0.07 |
|  |  | 2: DFAe/UJ | 3.14-3.93 | 2.43-3.41 | 22.33 | 0.07 |
| body | C. alpinus vs. C. fatioi | 1: CD/PostD * | 0.17-0.21 | 0.14-0.17 | 8.98 | 0.17 |
|  |  | 2: DFAe/DHL | 1.26-1.55 | 1.02-1.36 | 7.9 | 0.19 |
| head | C. alpinus vs. C. fatioi | 1: HD/UJ | 2.34-2.9 | 2.13-2.57 | 3.86 | 0.3 |
|  |  | 2: MW/ES | 1.47-3 | 1.82-6.16 | 3.15 | 0.34 |
| head + <br> body | C. alpinus vs. C. steinmanni (a) | ED/EC | 0.74-0.9 | 0.74-0.9 | 8.07 | 0.05 |
|  |  | CD/CL | 0.6-0.75 | 0.54-0.7 | 8.02 | 0.05 |
| body | C. alpinus vs. C. steinmanni | 1: DFAe/AFAe | 1.5-1.83 | 1.43-1.62 | 5.7 | 0.06 |
|  |  | 2: PelvFS/DFAe | 0.24-0.36 | 0.29-0.37 | 5.58 | 0.07 |
| head | C. alpinus vs. C. steinmanni | 1: EC/SW | 1.47-2.13 | 1.43-1.7 | 2.45 | 0.16 |
|  |  | 2: ED/EC | 0.74--0.9 | 0.79-0.9 | 2.2 | 0.18 |
| head + body | C. alpinus vs. C. profundus | 1: CD/DHL * | 0.54-0.62 | 0.4-0.49 | 19.86 | 0.07 |
|  |  | 2: PecF2/CF | 0.63-0.82 | 0.74-0.90 | 19.01 | 0.07 |


| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| body | C. alpinus vs. C. profundus | 1: CD/DHL * | 0.54-0.62 | 0.4-0.49 | 9.31 | 0.15 |
|  |  | 2: PecF2/CF | 0.63-0.82 | 0.74-0.90 | 7.32 | 0.19 |
| head | C. alpinus vs. C. profundus | 1: EH/PostD | 0.09-0.11 | 0.11-0.15 | 4.32 | 0.21 |
|  |  | 2: SD/UJW | 0.35-0.51 | 0.30-0.44 | 3.93 | 0.23 |
| head + <br> body | C. alpinus vs. C. acrinasus | 1: CD/LJ | 0.95-1.11 | 0.79-1 | 65.21 | 0.02 |
|  |  | 2: CF/M * | 5.55-6.55 | 4.4-5.57 | 65.13 | 0.02 |
| body | C. alpinus vs. C. acrinasus | 1: CD/DHL | 0.54-0.62 | 0.46-0.58 | 4.69 | 0.25 |
|  |  | 2: DFAe/DFPe | 3.39-4.72 | 2.84-4.54 | 3.91 | 0.29 |
| head | C. alpinus vs. C. acrinasus | 1: $\mathrm{PostO} / \mathrm{M}$ | 2.4-3 | 2.17-2.56 | 4.26 | 0.21 |
|  |  | 2: HD/MW | 6.57-8.7 | 6.02-8.87 | 3.65 | 0.24 |
| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (shape vs. Size) |
| head + body | C. fatioi vs. C. steinmanni (b) | 1: CD/UJ * | 1.02-1.34 | 1.36-1.55 | 33.96 | 0.04 |
|  |  | 2: PelvF/PAdC | 0.73-1 | 0.84-0.1 | 33.71 | 0.04 |
| body | C. fatioi vs. C. steinmanni | 1: CD/PostD * | 0.14-0.17 | 0.17-0.20 | 6.34 | 0.22 |
|  |  | 2: DHL/BD | 0.5-0.7 | 0.45-0.58 | 5.37 | 0.25 |
| head | C. fatioi vs. C. steinmanni | 1: HD/UJ | 2.13-2.57 | 2.42-2.83 | 4.41 | 0.23 |
|  |  | 2: HW/LJW | 3.17-6.12 | 3.72-5.1 | 3.25 | 0.29 |
| head + body | C. fatioi vs. C. profundus | 1: CL/EC | 1.84-2.98 | 1.56-2.09 | 10.03 | 0.02 |
|  |  | 2: DHL/UJ | 2.11-2.70 | 2.32-2.92 | 9.54 | 0.02 |
| body | C. fatioi vs. C. profundus | 1: CL/DHL | 0.76-1.04 | 0.61-0.82 | 4.44 | <0.01 |
|  |  | 2: DFPe/CD | 0.56-0.87 | 0.56.0.82 | 3.2 | <0.01 |
| head | C. fatioi vs. C. profundus | 1: EC/SW | 1.32-1.73 | 1.63-2.38 | 5.05 | 0.08 |
|  |  | 2: UJ/UJW | 1.04-1.50 | 1-1.29 | 4.28 | 0.09 |
| head + body | C. fatioi vs. C. acrinasus | 1: CD/PostD | 0.14-0.17 | 0.16-0.2 | 8.3 | 0.05 |
|  |  | 2: ED/SW | 1.08-1.5 | 1.3-1.79 | 8 | 0.05 |
| body | C. fatioi vs. C. acrinasus | 1: CD/PostD | 0.14-0.17 | 0.16-0.2 | 3.66 | 0.07 |
|  |  | 2: AFAe/DHL | 0.69-0.9 | 0.61-0.93 | 2.93 | 0.09 |
| head | C. fatioi vs. C. acrinasus | 1: ED/SW | 1.08-1.5 | 1.3-1.79 | 3.05 | 0.15 |
|  |  | 2: MW/ES | 1.82-6.16 | 1.4-3.02 | 2.45 | 0.18 |
| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (shape vs. Size) |
| head + <br> body | C. steinmanni vs. C. profundus (c) | 1: CD/DHL * | 0.53-0.63 | 0.4-0.49 | 23.9 | 0.05 |
|  |  | 2: CL/IOW | 2.05-2.69 | 1.69-2.28 | 23.3 | 0.05 |
| body | C. steinmanni vs. C. profundus | 1: CD/DHL * | 0.53-0.63 | 0.4-0.49 | 9.13 | 0.14 |
|  |  | 2: PecF2/DFAe | 0.76-0.96 | 0.82-1.21 | 7.44 | 0.17 |
| head | C. steinmanni vs. C. profundus | 1: SW/UJW | 0.65-0.80 | 0.54-0.69 | 5.9 | 0.12 |
|  |  | 2: EH/PostO | 0.36-0.47 | 0.41-0.52 | 5.37 | 0.13 |
| head + <br> body | C. steinmanni vs. C. acrinasus (d) | 1: CD/M * | 1.86-2.24 | 1.4-1.9 | 160.64 | <0.01 |
|  |  | 2: PostD/LJ | 4.96-5.9 | 4.65-5.43 | 160.6 | <0.01 |
| body | C. steinmanni vs. C. acrinasus | 1: CD/DHL | 0.53-0.63 | 0.46-0.58 | 4.46 | 0.23 |
|  |  | 2: PelvF/DHL | 1.08-1.26 | 0.95-1.16 | 3.83 | 0.26 |
| head | C. steinmanni vs. C. acrinasus | 1: ED/HD | 0.29-0.33 | 0.31-0.37 | 4.54 | 0.13 |
|  |  | 2: HL/M | 4.6-5.53 | 4.21-5.17 | 3.41 | 0.17 |
| Characters | Species comparison | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (shape vs. Size) |
| head + body | C. profundus vs. C. acrinasus | 1: PecF2/CD | 2.37-3.16 | 1.91-2.59 | 13.46 | 0.01 |
|  |  | 2: LJ/UJW | 1.34-1.86 | 1.54-2.27 | 13.12 | 0.01 |
| body | C. profundus vs. C. acrinasus | 1: PecF2/CD | 2.37-3.16 | 1.91-2.59 | 4.58 | 0.05 |
|  |  | 2: DHL/TL | 0.13-0.15 | 0.12-0.13 | 3.48 | 0.06 |
| head | C. profundus vs. C. acrinasus | 1: M/UJW | 0.69-0.94 | 0.8-1.22 | 4.45 | 0.02 |
|  |  | 2: EC/LJ | 0.66-0.81 | 0.58-0.72 | 3.88 | 0.02 |
| Characters | Mulitple species comparison | Best ratios | Range group 1 | Range group 2 | Standard distance | $\delta$ (shape vs. Size) |
| head + <br> body | C. alpinus + C. steinmanni vs. 4 other species | 1: CD/UJ * | 1.36-1.65 | 0.96-1.43 | 5.34 | 0.24 |

(a) PelvFS, PecF1, DFAd, DFAe, DFPe, TL, SL, EH, SD, SW, INW, IOW
(b) PelvFS, PecF 1, DFAd, TL
(c) PelvFS, PecF1, DFAd, TL,EH
(d) PelvFS, PecF1, DFAd, TL, EH, ES, EC

Table I I. The first- and second-best ratios retrieved from the LDA ratio extractor of either head or body characters (see Table 1) alone or combined, used for pair-wise comparisons of all contemporary specimens from the four whitefish species of Lake Brienz. For some species comparisons only a subset of characters could be used (a-l); the respective characters that were excluded are listed at the end of the table. Only external characters were used for the LDA comparisons, since internal characters (gill raker and gill arch length) cannot be measured on live specimens, and are thus not informative to assign specimens to species in the field. Due to large size differences between the species the LDA ratios were calculated with three different datasets; once each with individuals larger or smaller than 163.5 mm standard length and once with the full size ranges of all species. For the multi-species comparisons, only the comparisons that yielded distinguishing ratios are shown. $\delta$ is a measure of how good shape discriminates in comparison to size (i.e., the smaller the less allometry). Ratios marked with an asterisk * have very little (for the pairwise species comparisons not more than one specimen of one species overlaps with that of the other species) or no overlap and were thus eligible for use in the species key and the diagnoses.

| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| head + body | C. albellus vs. C. alpinus (a) | <163.5mm | 1: PreA/LJ * | 6.33-7.44 | 9.24-9.97 | 27.13 | 0.04 |
|  |  |  | 2: AFAe/M | 1.65-2.25 | 2.58-2.63 | 25.94 | 0.04 |
| body | C. albellus vs. C. alpinus | $<163.5 \mathrm{~mm}$ | 1: PecF2/DFAd* | 0.81-1.06 | 0.78-0.8 | 9.97 | 0.14 |
|  |  |  | 2: DHL/PreD | 0.34-0.42 | 0.32-0.34 | 9.4 | 0.15 |
| head | C. albellus vs. C. alpinus | $<163.5 \mathrm{~mm}$ | 1: HD/LJ | 1.30-1.55 | 1.77-1.92 | 15.43 | 0.02 |
|  |  |  | 2: IOW/UJW | 0.89-1.30 | 1.20-1.26 | 14.14 | 0.02 |
| head + <br> body | C. albellus vs. C. fatioi | $<163.5 \mathrm{~mm}$ | 1: PecF2/PreA* | 0.22-0.28 | 0.2-0.22 | 5.78 | 0.16 |
|  |  |  | 2: DHL/PreP | 0.31-0.38 | 0.30-0.32 | 4.49 | 0.2 |
| body | C. albellus vs. C. fatioi | $<163.5 \mathrm{~mm}$ | 1: PecF2/PreA* | 0.22-0.28 | 0.2-0.22 | 6.76 | 0.17 |
|  |  |  | 2: DHL/TL | 0.13-0.18 | 0.13-0.14 | 5.7 | 0.19 |
| head | C. albellus vs. C. fatioi | $<163.5 \mathrm{~mm}$ | 1: UJ/ES * | 6.81-12.42 | 4.51-6.15 | 8.63 | 0.12 |
|  |  |  | 2: EH/HL * | 0.27-0.31 | 0.23-0.27 | 7.3 | 0.14 |
| head + body | C. albellus vs. C. brienzii (b) | $<163.5 \mathrm{~mm}$ | 1: $\mathrm{PreD} / \mathrm{LJ} *$ | 3.99-4.68 | 5.05-5.57 | 47.9 | 0.01 |
|  |  |  | 2: M/ES* | 5.35-9.76 | 3.31-4.37 | 47.63 | 0.01 |
| body | C. albellus vs. C. brienzii | <163.5mm | 1: PecF2/PreD* | 0.36-0.45 | 0.29-0.32 | 15.95 | 0.06 |
|  |  |  | 2: DHL/TL | 0.13-0.18 | 0.13-0.14 | 9.91 | 0.05 |
| head | C. albellus vs. C. brienzii | $<163.5 \mathrm{~mm}$ | 1: LJ/ES * | 9.62-17.28 | 6.01-6.49 | 12.51 | 0.05 |
|  |  |  | 2: HL/UJ | 2.87-3.5 | 3.19-3.6 | 8.87 | 0.04 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + body | C. alpinus vs. C. fatioi <br> (b) | >163.5mm | 1: AFAe/UJ * | 1.96-2.5 | 1.66-1.96 | 26.08 | 0.04 |
|  |  |  | 2: CL/PreA | 0.14-0.18 | 0.17-0.21 | 26.46 | 0.04 |
| body | C. alpinus vs. C. fatioi | >163.5mm | 1: AFae/TL | 0.1-0.11 | 0.09-0.1 | 13.41 | 0.11 |
|  |  |  | 2: CL/PreA | 0.14-0.18 | 0.17-0.21 | 13.41 | 0.11 |
| head | C. alpinus vs. C. fatioi | >163.5mm | 1: HL/UJ * | 3.55-3.93 | 3.13-3.55 | 11.51 | 0.07 |
|  |  |  | 2: LJW/UJW | 0.33-0.44 | 0.38-0.55 | 11.02 | 0.07 |
| head + body | C. alpinus vs. C. brienzii (c) | >163.5mm | 1: CD/SW * | 2.25-2.64 | 1.82-2.04 | 34.25 | 0.02 |
|  |  |  | 2: LJW/UJW * | 0.33-0.44 | 0.45-0.55 | 33.91 | 0.02 |
| body | C. alpinus vs. C. brienzii (d) | $>163.5 \mathrm{~mm}$ | 1: DFAe/PAdC* | 1.11-1.32 | 0.96-1.16 | 18.53 | 0.07 |
|  |  |  | 2: CD/AFB | 0.61-0.68 | 0.52-0.62 | 18.31 | 0.07 |
| head | C. alpinus vs. C. brienzii (e) | $>163.5 \mathrm{~mm}$ | 1: LJW/UJW * | 0.33-0.44 | 0.45-0.55 | 7.44 | 0.08 |
|  |  |  | 2: PostO/UJ | 1.8-2.12 | 1.57-1.86 | 6.78 | 0.08 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + body | C. fatioi vs. C. brienzii (f) | >163.5mm | 1: CL/PAdC | 0.71-0.86 | 0.66-0.76 | 7.46 | 0.08 |
|  |  |  | 2: BD/LJ | 2.44-3.05 | 2.31-2.82 | 7.36 | 0.08 |
| body | C. fatioi vs. C. brienzii | $>163.5 \mathrm{~mm}$ | 1: CL/PAdC | 0.71-0.86 | 0.66-0.76 | 6.04 | 0.1 |
|  |  |  | 2: CF/BD | 0.87-1.13 | 0.93-1.12 | 5.92 | 0.1 |
| head | C. fatioi vs. C. brienzii | >163.5mm | 1: ED/M | 1.03-1.19 | 1.04-1.57 | 3.58 | 0.18 |
|  |  |  | 2: HW/UJW | 1.89-2.23 | 1.88-2.33 | 3.38 | 0.19 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + <br> body | C. alpinus vs. other 3 species | <163.5mm | 1: DFAd/LJ * | 2.57-2.58 | 1.6-2.1 | 23.47 | 0.03 |
|  |  |  | 2: AdFB/PAdC | 0.26-0.28 | 0.21-0.42 | 22.66 | 0.03 |


| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| head + <br> body | C. albellus vs. other 3 species | <163.5mm | 1: PostD/EH* | 5.47-6.93 | 7.5-8.9 | 48.36 | 0.02 |
|  |  |  | 2: UJW/ES | 4.88-9.3 | 3.41-5.31 | 48.13 | 0.02 |
| head + <br> body | C. alpinus vs. C. fatioi <br> + C. brienzii (g) | >163.5mm | 1: DFAe/UJ* | 3.28-4.1 | 2.58-3.19 | 24.71 | 0.05 |
|  |  |  | 2: CD/SW * | 2.25-2.64 | 1.76-2.27 | 24.37 | 0.05 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + <br> body | C. albellus vs. C. alpinus (h) | 100-290 | 1: PreD/LJ * | 3.99-4.68 | 5.6-6.81 | 22.86 | 0.13 |
|  |  |  | 2: DFAe/UJ * | 2.14-2.79 | 3.25-4.1 | 21.65 | 0.14 |
| head | C. albellus vs. C. alpinus | 100-290 | 1: HD/UJ * | 1.87-2.2 | 2.38-2.78 | 14.39 | 0.18 |
|  |  |  | 2: LJ/IOW * | 1.53-1.99 | 1.33-1.57 | 13.25 | 0.19 |
| head + <br> body | C. albellus vs. C. fatioi | 100-290 | 1: PreP/EH* | 6.56-7.98 | 8.94-11.43 | 15.95 | 0.13 |
|  |  |  | 2: CL/UJ | 1.44-2.02 | 1.93-2.72 | 15.09 | 0.14 |
| head + body | C. albellus vs. <br> C. brienzii (i) | 100-290 | 1: PreD/EH* | 6.1-7.58 | 8.12-10.32 | 50.86 | 0.04 |
|  |  |  | 2: CL/LJ | 0.99-1.45 | 1.38-1.65 | 50.6 | 0.04 |
| head | C. albellus vs. C. brienzii | 100-290 | 1: EH/HL * | 0.27-0.31 | 0.22-0.27 | 9.33 | 0.18 |
|  |  |  | 2: LJ/ES | 9.62-17.28 | 6.08-12.43 | 8.57 | 0.22 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + body | C. alpinus vs. C. brienzii ( $j$ ) | 100-290 | 1: DFAd/LJW * | 9.84-14.82 | 6.05-8.91 | 20.72 | 0.03 |
|  |  |  | 2: DHL/LJ * | 1.84-2.22 | 1.63-1.82 | 20.47 | 0.02 |
| body | C. alpinus vs. C. brienzii (k) | 100-290 | 1: PecF2/DFAd* | 0.74-0.85 | 0.85-1.03 | 87.52 | <0.01 |
|  |  |  | 2: CD/PostD | 0.17-0.2 | 0.15-0.18 | 87.48 | <0.01 |
| head | C. alpinus vs. C. brienzii | 100-290 | 1: HD/LJW* | 6.72-9.39 | 5.23-6.66 | 11.94 | 0.04 |
|  |  |  | 2: HL/LJ * | 2.54-2.96 | 2.19-2.47 | 11.61 | 0.04 |
| head + body | C. alpinus vs. C. fatioi (b) | 100-290 | 1: DFAe/UJ * | 3.25-4.1 | 2.45-3.17 | 18.98 | 0.03 |
|  |  |  | 2: PecF2/AFAe | 1.24-1.47 | 1.37-1.63 | 18.63 | 0.03 |
| body | C. alpinus vs. C. fatioi | 100-290 | 1: PecF2/DFAe | 0.77-0.89 | 0.87-1.02 | 9.25 | 0.08 |
|  |  |  | 2: AFAe/PostD | 0.27-0.32 | 0.22-0.29 | 8.71 | 0.08 |
| head | C. alpinus vs. C. fatioi | 100-290 | 1: LJW/UJW | 0.33-0.47 | 0.37-0.55 | 5.62 | 0.08 |
|  |  |  | 2: HL/UJ | 3.43-3.93 | 3.13-3.63 | 4.98 | 0.08 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + body | C. fatioi us. C. brienzii (l) | 100-290 | 1: AFB/BD | 0.45-0.67 | 0.44-0.58 | 28.19 | <0.01 |
|  |  |  | 2: $\mathrm{PreD} / \mathrm{M}$ | 8.87-14.85 | 9.13-11.41 | 28.16 | <0.01 |
| body | C. fatioi vs. C. brienzii | 100-290 | 1: AFB/BD | 0.45-0.67 | 0.44-0.58 | 2.76 | 0.05 |
|  |  |  | 2: PreP/PreA | 0.58-0.65 | 0.57-0.64 | 2.51 | 0.05 |
| head | C. fatioi vs. C. brienzii | 100-290 | 1: ED/M | 1.04-1.57 | 1.03-1.28 | 1.72 | 0.1 |
|  |  |  | 2: SN/MW | 2.09-2.63 | 1.78-2.87 | 1.52 | 0.1 |
| Characters | Species comparison | Size range | Best ratios | Range species 1 | Range species 2 | Standard distance | $\delta$ (Shape vs. size) |
| head + body | C.albellus vs. other 3 species | 100-290 | 1: PreD/EH * | 6.1-7.58 | 8.12-10.5 | 10.89 | 0.16 |
|  |  |  | 2: CL/UJ | 1.44-2.02 | 1.85-2.72 | 9.79 | 0.17 |
| head + <br> body | C.alpinus vs. other 3 species | 100-290 | 1: DFAe/UJ* | 3.25-4.1 | 2.14-3.19 | 9.59 | 0.11 |
|  |  |  | 2: LJW/UJW | 0.33-0.47 | 0.34-0.55 | 8.98 | 0.12 |

(a) PelvS, PecF1, DFAd, DFAe, DFPe, TL, SL, EH, SD, SW, INW, IOW
(b) PelvFS, PelvFB, PecFB, DFPe, TL, EH, ED, SD, IOW
(c) PelvFB, PelvFS, PelvF, PecF1, DFB, DFAe, DFPe, AFB, AFAe, AdFB, CF, PAdC, PreP, PreA, SL, TL, BD, PostD, DHL, ED, EH, ES, PostO, HD, MW, SN, SD, SW, IOW, INW
(d) PecFB, PelvFB, PelvF, PelvFS, PecF1, DFAe, DFPe, AFAe, AdFB, PreP, CF, TL, PostD
(e) ED, EH, HD, SD, SW, INW
(f) PelvFB, PelvFS, PelvF, PecFB, DFAe, DFAd, DFPe, CF, PreP, SL, TL, ED, EH, MW, SD, SW, IOW, INW, ES
(g) PelvFS, PecF1, PecFB, DFAd, DFPe, SL, TL, ED, EH, INW, CF
(h) PelvS, PecF1, DFAd, DFAe, DFPe, TL, SL, EH, SD, SW, INW, IOW
(i) PelvFS
(j) PelvFB, PelvFS, PelvF, PecF1,DFAe, DFPe, AFB, AFAe,CF, SL, TL, BD, ED, EH, ES, PostO, HD, MW, SN, SD, SW, IOW, INW
(k) PelvFS, PecF1, CF, SL, TL
(l) PelvS, TL, INW

Distribution and notes on biology. Coregonus macrophthalmus is found in Lake Constance, especially in the upper basin (Obersee). It is unclear if it also occurs in the lower basin (Untersee) of the lake.

Common name. Gangfisch.
Table 12. Morphological and meristic data of C. gutturosus Gmelin, 1818, C. arenicolus Kottelat, 1997, C. macrophthalmus Nüsslin, 1882 and C. wartmanni Bloch, 1784 from Lake Constance. Coregonus gutturosus Gmelin, 1818, non-types $\mathrm{N}=10$. Coregonus arenicolus Kottelat, 1997, holotype, NMBE-1076223 (Eawag-239-1),
sex unknown; paratypes $\mathrm{N}=3$. Coregonus macrophthalmus $\mathrm{Nüsslin}, 1882$, syntypes $\mathrm{N}=7$. C. wartmanni Bloch, 1784, non- type, NMBE-1076206, female.

| Morphological characters | C. gutturosus |  |  | C. arenicolus |  |  |  | C. macropbthalmus |  |  | $\begin{array}{\|l\|} \hline \text { C. wartmanni } \\ \hline \text { Non-type } \\ \hline \end{array}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Non-types ( $\mathrm{N}=10$ ) |  |  | Holotype | Paratypes (N=3) |  |  | Syntypes ( $\mathrm{N}=7$ ) |  |  |  |
|  | $N$-total | Mean $\pm$ StDev | Range |  | $N$-total | Mean $\pm$ StDev | Range | $N$-total | Mean $\pm$ StDev | Range |  |
| SL (mm) | 10 | $220.4 \pm 36.8$ | (169-292) | 296.0 | 3 | $301.3 \pm 12.5$ | (289-314) | 7 | $213.9 \pm 12.4$ | (193-235) | 301 |
| Percentage of standard length |  |  |  |  |  |  |  |  |  |  |  |
| PelvFB | 10 | $4.1 \pm 0.2$ | (3.7-4.4) | 3.9 | 3 | $4.4 \pm 0.3$ | (3.9-4.6) | 7 | $3.8 \pm 0.3$ | (3.3-4.2) | 3.8 |
| PelvFS | 10 | $6.1 \pm 0.4$ | (5.3-6.7) | 5.4 | 3 | $5.7 \pm 0.5$ | (5.2-6.1) | 7 | $5.7 \pm 0.7$ | (4.8-6.9) | 6.5 |
| PelvF | 10 | $17.1 \pm 1.2$ | (15.4-19.1) | 17.3 | 3 | $17.3 \pm 0.7$ | (16.8-18.1) | 7 | $16.5 \pm 0.9$ | (15.2-17.6) | 15.4 |
| PecFB | 10 | $3.4 \pm 0.3$ | (2.9-3.9) | 3.3 | 3 | $3.4 \pm 0.2$ | (3.2-3.5) | 7 | $3.2 \pm 0.4$ | (2.8-3.9) | 3 |
| PecF1 | 10 | $16.8 \pm 1.1$ | (14.8-18.9) | 14.4 | 3 | $16.8 \pm 0.5$ | (14.4-17.2) | 7 | $16.4 \pm 1.2$ | (15.1-18.1) | 16 |
| PecF2 | 10 | $18.2 \pm 1.5$ | (16.8-20.3) | 15.7 | 3 | $17.4 \pm 0.8$ | (15.7-18) | 7 | $17.1 \pm 1$ | (15.6-18.4) | 17 |
| DFB | 10 | $11.9 \pm 0.7$ | (10.7-12.8) | 12.2 | 3 | $12.2 \pm 1.1$ | (11.0-13.1) | 7 | $11.6 \pm 0.7$ | (10.8-12.4) | 11.2 |
| DFAe | 10 | $19.3 \pm 1.3$ | (17.6-21.6) | 18.9 | 3 | $19.2 \pm 1.2$ | (18.0-20.3) | 7 | $18.2 \pm 1.2$ | (16.6-19.6) | 16.6 |
| DFAd | 10 | $20.4 \pm 1.1$ | (19.0-22.2) | 20.2 | 3 | $20.5 \pm 1.3$ | (19.3-21.9) | 7 | $19.2 \pm 1.3$ | (17.2-20.5) | 18.2 |
| DFPe | 10 | $5.5 \pm 0.7$ | (4.8-7.0) | 5.2 | 3 | $5.5 \pm 0.2$ | (5.2-5.7) | 7 | $5.2 \pm 0.6$ | (4.4-5.9) | 4.6 |
| AFB | 10 | $12.4 \pm 0.8$ | (11.4-13.4) | 11.9 | 3 | $11.5 \pm 1.1$ | (10.7-12.7) | 7 | $12.3 \pm 1.3$ | (10.6-14.2) | 12.5 |
| AFAe | 10 | $12.3 \pm 1.0$ | (10.7-13.9) | 13.2 | 3 | $13.3 \pm 0.5$ | (12.9-13.8) | 5 | $12.1 \pm 1.3$ | (10.8-13.9) | 11.1 |
| AdFB | 10 | $5.6 \pm 0.4$ | (4.9-6.1) | 5.7 | 3 | $5.0 \pm 1.3$ | (3.7-6.2) | 7 | $5.3 \pm 0.3$ | (4.9-5.8) | 4 |
| CF | 9 | $23.2 \pm 1.9$ | (20.8-25.6) | na | 2 | $24 \pm 0.1$ | (24-24.1) | 3 | $22.6 \pm 1.2$ | (21.8-24) | 23.8 |
| CD | 10 | $7.4 \pm 0.4$ | (6.7-8.2) | 7.7 | 3 | $8.1 \pm 0.1$ | (7.7-8.2) | 7 | $7.4 \pm 0.4$ | (6.9-8) | 7.4 |
| CL | 10 | $12.9 \pm 0.8$ | (11.5-13.9) | 14.4 | 3 | $12.9 \pm 0.8$ | (12.0-14.4) | 7 | $13.8 \pm 1.3$ | (12.4-16.5) | 13.2 |
| PAdC | 10 | $18.5 \pm 0.6$ | (17.4-19.3) | 19.6 | 3 | $17.2 \pm 2.2$ | (14.6-19.6) | 7 | $18.9 \pm 1.1$ | (17.6-20.2) | 17.8 |
| DHL | 10 | $16.8 \pm 0.8$ | (15.4-18.1) | 15.1 | 3 | $15.1 \pm 0.2$ | (14.8-15.3) | 7 | $15.7 \pm 0.8$ | (14.4-16.5) | 14.5 |
| PreP | 10 | $52.7 \pm 1.4$ | (50.4-54.1) | 49.5 | 3 | $50.6 \pm 0.5$ | (49.5-51.0) | 7 | $51.7 \pm 1.7$ | (48.1-53.1) | 50.7 |
| PreA | 10 | $77.9 \pm 1.4$ | (76.0-80.4) | 75.0 | 3 | $79.2 \pm 0.9$ | (75.0-80.3) | 7 | $76.8 \pm 0.9$ | (75.7-78.3) | 77.4 |
| PreD | 10 | $48.4 \pm 0.9$ | (46.8-49.6) | 47.9 | 3 | $49.2 \pm 0.5$ | (47.9-49.6) | 7 | $47 \pm 1$ | (45.8-48.5) | 47.3 |
| BD | 10 | $25.9 \pm 1.9$ | (22.9-29.6) | 24.4 | 3 | $26.2 \pm 0.8$ | (24.4-27.1) | 7 | $23.5 \pm 1.9$ | (21.0-26.9) | 23.5 |
| PostD | 10 | $43 \pm 1.5$ | (40.6-45.4) | 44.8 | 3 | $42.2 \pm 1.7$ | (40.4-44.8) | 7 | $43 \pm 1.4$ | (41.6-45.7) | 44 |
| TL | 9 | $120.2 \pm 3$ | (115.1-124.3) | na | 2 | $121.5 \pm 2.9$ | (119.4-123.5) | 3 | $119.2 \pm 0.7$ | (118.9-120) | 120.6 |
| HL (mm) | 10 | $50.3 \pm 7.1$ | (41.6-62.4) | 61.8 | 3 | $63.7 \pm 3.9$ | (59.6-67.2) | 7 | $47.5 \pm 3.2$ | (42.6-51.3) | 64.7 |
| Percentage of head length |  |  |  |  |  |  |  |  |  |  |  |
| SN | 10 | $22.4 \pm 0.7$ | (21.1-23.1) | 23.4 | 3 | $23.4 \pm 1.6$ | (21.6-24.6) | 7 | $21.7 \pm 2.7$ | (18-25.6) | 24 |
| ED | 10 | $21.1 \pm 1.4$ | (19.4-23) | 19.6 | 3 | $17.7 \pm 0.4$ | (17.3-19.6) | 7 | $24.1 \pm 1.7$ | (21.3-26.1) | 18.9 |
| EC | 10 | $26.9 \pm 1.2$ | (25.4-29.3) | 25.7 | 3 | $25 \pm 0.8$ | (24.1-25.7) | 7 | $28.9 \pm 2$ | (25.4-30.8) | 23.9 |
| EH | 10 | $21.3 \pm 0.6$ | (20.5-22.6) | 20.8 | 3 | $19.6 \pm 0.9$ | (18.8-20.8) | 7 | $23.2 \pm 2.1$ | (19.5-25.6) | 19 |


| Morphological characters |  | C. gutturosus |  | C. arenicolus |  |  |  | C. macrophthalmus |  |  | $\begin{gathered} \hline \text { C. wartmanni } \\ \hline \text { Non-type } \\ \hline \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Non-types ( $\mathrm{N}=10$ ) |  |  | Holotype | Paratypes ( $\mathrm{N}=3$ ) |  |  | Syntypes (N=7) |  |  |  |
|  | $N$-total | Mean $\pm$ StDev | Range |  | $N$-total | Mean $\pm$ StDev | Range | $N$-total | Mean $\pm$ StDev | Range |  |
| ES | 10 | $4.8 \pm 0.7$ | (3.5-5.6) | 4.9 | 3 | $5.2 \pm 0.5$ | (4.6-5.5) | 7 | $3.9 \pm 0.8$ | (2.7-4.6) | 5.1 |
| PostO | 10 | $52.8 \pm 1$ | (51.5-54.4) | 55.7 | 3 | $54 \pm 1$ | (53-55.7) | 7 | $50.2 \pm 1.9$ | (48.5-53.2) | 53.4 |
| HD | 10 | $74.2 \pm 3.2$ | (69.9-80.6) | 68.2 | 3 | $72.6 \pm 3$ | (68.2-75) | 7 | $68.6 \pm 4.8$ | (61.6-76.3) | 67.6 |
| MW | 10 | $9.8 \pm 0.6$ | (9-11.2) | 10.4 | 3 | $10.5 \pm 0.5$ | (10-11) | 6 | $10.1 \pm 0.8$ | (8.7-11.1) | 10.6 |
| UJ | 10 | $26.8 \pm 1.2$ | (24.6-29) | 27.2 | 3 | $29.3 \pm 1$ | (27.2-30.1) | 7 | $30.3 \pm 2.3$ | (26.7-33.8) | 28.8 |
| LJ | 10 | $36.6 \pm 1.4$ | (34.3-39.1) | 37.8 | 3 | $38.7 \pm 0.6$ | (37.8-39.1) | 7 | $42.2 \pm 2$ | (40-44.4) | 43.5 |
| M | 10 | $18.9 \pm 1.3$ | (17.3-21.7) | 21.1 | 3 | $19.7 \pm 0.9$ | (18.6-21.1) | 5 | $23.1 \pm 1.9$ | (20.1-24.7) | 22 |
| SD | 10 | $10.2 \pm 0.8$ | (9.3-11.9) | 9.7 | 3 | $10.9 \pm 1.3$ | (9.7-12.3) | 7 | $7.4 \pm 1.2$ | (5.5-9.5) | 6.8 |
| SW | 10 | $15.1 \pm 1.6$ | (12.3-17.6) | 14.9 | 3 | $17.8 \pm 0.7$ | (14.9-18.5) | 7 | $15.6 \pm 1.2$ | (14.1-17.4) | 15 |
| HW | 10 | $56.1 \pm 4.3$ | (46.7-62.3) | 51.8 | 3 | $50.8 \pm 0.5$ | (50.5-51.8) | 7 | $41.6 \pm 1.5$ | (39.3-43.3) | 45.5 |
| IOW | 10 | $28.4 \pm 1.7$ | (26.2-31.6) | 29.6 | 3 | $29.7 \pm 1$ | (28.8-30.8) | 7 | $26.1 \pm 1.6$ | (23.8-28.9) | 24.2 |
| INW | 10 | $11.9 \pm 0.7$ | (10.7-12.7) | 12.0 | 3 | $13.7 \pm 0.1$ | (12-13.8) | 5 | $11.9 \pm 1.3$ | (10.7-14.1) | 12.7 |
| LJW | 10 | $7.7 \pm 1$ | (6.8-9.9) | 7.8 | 3 | $8.1 \pm 0.3$ | (7.8-8.5) | 5 | $7.8 \pm 1$ | (6.4-8.8) | 8.1 |
| UJW | 10 | $25.2 \pm 1.2$ | (23.1-26.8) | 24.9 | 3 | $26.4 \pm 0.8$ | (24.9-27.2) | 7 | $21.6 \pm 1.9$ | (18.6-24.6) | 22.7 |
| MGR | 9 | $6.9 \pm 1.3$ | (4.1-8.7) | 9.9 | 2 | $10.2 \pm 0.6$ | (9.8-10.6) | 4 | $12.5 \pm 1.4$ | (11.6-14.7) | 10.8 |
| LGR | 9 | $8.2 \pm 1.4$ | (6.7-10.6) | 10.9 | 2 | $11.5 \pm 0.6$ | (10.9-12) | 4 | $14.6 \pm 1.2$ | (13.3-16.1) | 11.3 |
| UA | na | na | na | na | na | na | na | na | na | na | na |
| LA | na | na | na | na | na | na | na | na | na | na | na |
| Meristic character | N -total | Mode | Range |  | N -total | Mode | Range | N -total | Mode | Range |  |
| PelvF nbranched | 10 | 1 | (1-1) | 1 | 3 | 1 | (1-1) | 7 | 1 | (1-1) | 1 |
| PelvF branched | 10 | 11 | (9-11) | 11 | 3 | 11 | (11-11) | 7 | 10 | (10-11) | 12 |
| PecF nbranched | 10 | 1 | (1-1) | 1 | 3 | 1 | (1-1) | 7 | 1 | (1-1) | 1 |
| PecF branched | 10 | 13 | (12-14) | 12 | 3 | 12 | (12-14) | 7 | 15 | (14-15) | 16 |
| DF unbranched | 10 | 4 | (3-4) | 4 | 3 | 4 | (4-4) | 7 | 4 | (4-4) | 4 |
| DF branched | 10 | 10 | (9-10) | 9 | 3 | 10 | (9-10) | 7 | 9 | (9-10) | 10 |
| AF unbranched | 10 | 3 | (3-4) | 3 | 3 | 3 | (3-3) | 7 | 3 | (3-4) | 4 |
| AF branched | 10 | 11 | (10-12) | 10 | 3 | 10 | (10-11) | 7 | 11 | (10-13) | 13 |
| LS | 10 | 78 | (76-82) | 82 | 3 | na | (82-90) | 7 | 80 | (73-80) | 84 |
| PDS | 10 | 33 | (31-35) | 36 | 3 | na | (36-44) | 7 | 32 | (32-36) | 34 |
| TDS | 10 | 10 | (9-10) | 10 | 3 | 10 | (10-11) | 7 | 9 | (9-10) | 10 |
| TAS | 10 | 8 | (7-9) | 8 | 3 | 9 | (8-9) | 7 | 7 | (7-9) | 8 |
| TPS | 10 | 8 | (8-9) | 8 | 3 | 9 | (8-9) | 7 | 7 | (7-9) | 8 |
| UGR | 9 | 7 | (7-9) | 9 | 2 | na | (9-12) | 4 | 14 | (12-14) | 11 |
| LGR | 9 | 10 | (9-12) | 13 | 2 | na | (13-19) | 4 | 24 | (22-24) | 23 |
| total GR | 9 | 19 | (16-21) | 22 | 2 | na | (22-31) | 4 | 36 | (36-38) | 34 |

## Coregonus wartmanni, Bloch, 1784

## Figure 11

Material examined. Non-type. NMBE-1076206, Switzerland, Lake Constance ( $47^{\circ} 38^{\prime} \mathrm{N}, 9^{\circ} 22^{\prime} \mathrm{E}$ ), 301 mm SL , sex female.

Distribution and notes on biology. Coregonus wartmanni is found in Lake Constance, especially in the upper basin (Obersee). It is unclear if it also occurs in the lower basin (Untersee).

Common name. Blaufelchen.

## Identification key to the species of lakes Thun and Brienz

## Lake Thun

1 Caudal peduncle depth / upper jaw length ratio is 1.36-1.65 and caudal peduncle depth / maxilla length ratio is 1.77-2.24

- Caudal peduncle depth / upper jaw length ratio is 0.96-1.43.................... 3

2 Total number of gill rakers 25-30 C. alpinus

- Total number of gill rakers 31-35....................................... C. steinmanni

3 Total number of gill rakers 15-27..........................................C. profundus

- Total number of gill rakers 30-44 4

4 Colouration above the lateral line on the dorsum from a pale rose colouration to a pale brown colouration; no or few small pigmented dots on the edge of the scales or on the boundary of two scales on the flank; no pigmented dots on the dorsum C. albellus

- $\quad$ Colouration above the lateral line on the dorsum from a light to dark green and rarely a light olive; moderate to many dots on the edge of the scales or on the boundary of two scales on the flank and/or the dorsum
- Angle to body axis of the erected dorsal fin approx. $40-60^{\circ}$...... C. acrinasus


## Lake Brienz

1 Total number of gill rakers 26-30 and erected dorsal fin length / upper jaw length ratio is $3.25-4.1$
C. alpinus

- $\quad$ Total number of gill rakers 32-42 and erected dorsal fin length / upper jaw length ratio is $2.14-3.19$


Figure 12. Illustrations of specimens of each species from Lake Thun. From top to bottom: Coregonus alpinus: nontype, NMBE-1077244, 343 mm , male; Coregonus steinmanni: paratype, NMBE-1077218, 289.5 mm , male; Coregonus acrinasus: paratype, NMBE-1077270, 270 mm , male; Coregonus fatioi: nontype, NMBE-1077138, 267 mm , male; Coregonus albellus: non-type, NMBE-1077188, 215 mm , male; Coregonus profundus: non-type, Eawag-123850, 195 mm , male. The black scale $(1 \mathrm{~cm})$ below each fish acts as a reference for the actual size of the specimen.


Figure 12. Continued.

## Discussion

Phylogeographic studies have shown that the pre-alpine whitefish are a monophyletic clade, most closely related to whitefish from northern Europe. The clade is of hybrid origin involving two glacial lineages that must have come into secondary contact several hundred thousand years after their separation. Independent events of intra-lacustrine speciation led to a series of adaptive radiations in each major lake system of the northern preAlps (Hudson et al. 2011). Up to six endemic species can be found in the most diverse of these adaptive radiations (Vonlanthen et al. 2012; Hudson et al. 2016; Dönz et al. 2018). Here we take an integrative taxonomic approach, combining genetic assignments (Dönz et al. 2018) with morphological and meristic traits and multivariate statistical methods to delineate species and revise the taxonomy of the whitefish radiation of lakes Thun and Brienz in the western Aare catchment of Switzerland. We distinguish and characterize seven whitefish species from these lakes. Three of them occur in both lakes and three may be unique to Lake Thun and one may be unique to Lake Brienz. The three species that occur in both lakes have been described more than 130 years ago. Two of these species, $C$. alpinus and C. albellus, were described by Fatio (1885 and 1890 respectively). The third species, C. fatioi, was renamed by Kottelat (1997), but first described by Fatio (1890). Kottelat (1997) proposed C. fatioi as a replacement name, since the name given by Fatio (1890) was preoccupied by another species described by Fatio (1885). Our own earlier research combining genetics and ecology had shown that a fourth species is present in both lakes (Dönz et al. 2018). However, recent whole-genome data (De-Kayne et al. unpublished) csuggest that this newly discovered species is genetically different between lakes. We thus describe this species here as Coregonus steinmanni sp. nov. for the specimens from Lake Thun and those from Lake Brienz we designate as C. brienzii. Our earlier research (Hudson et al. 2011; Vonlanthen et al. 2012; Dönz et al. 2018) had further revealed that Lake Thun harbours two additional undescribed species and we describe these here as Coregonus profundus sp. nov. and Coregonus acrinasus sp. nov. Consistent with previous work (Dönz et al. 2018), recent genomic analyses (whole-genome data: De-Kayne et al. unpublished) find that the three species, C. alpinus, C. fatioi and C. albellus, occurring in both lakes cluster by species, whereas $C$. steinmanni from Lake Thun and C. brienzii from Lake Brienz (formerly C. steinmanni from Lake Brienz; Dönz et al. 2018) are not each others closest relatives. Interestingly, we also find morphological relationships to differ between the lakes; in Lake Thun C. steinmanni groups in morphospace with C. alpinus, whereas in Lake Brienz C. brienzii groups in morphospace with C. fatioi.

Based on genetic, morphological and ecological data at least two species from the Lake Thun-Brienz radiation, namely C. albellus (since at least 2004: Bittner 2009; Vonlanthen and Périat 2018; this study) and C. profundus (since at least 2016: this study) have colonized Lake Biel. There are no indications and no historical records that the Bernese cantonal officials have translocated any whitefish from other lakes into Lake Biel. Importantly, Steinmann (1950) already mentions that fishermen reported that suddenly after the Jura water correction, whitefish that resembled C. albellus (common name Brienzlig), appeared in Lake Biel. It is hence possible, that colonization of Lake Biel happened in recent times through the river Aare, which became connected with

Lake Biel after the Jura water correction of 1868-1878. At least one of the species, $C$. albellus, has likely established a self-sustaining population in Lake Biel, since a reasonable number of ripe specimens of this species have been caught repeatedly over several years during the typical spawning period of this species (late summer: September-October; Bittner 2009; Vonlanthen and Périat 2018; Suppl. material 1: Figure S9). Today, Lake Biel harbours two native whitefish species, C. confusus, Fatio 1885 and C. palaea, Cuvier 1829 (Kottelat and Freyhof 2007) but it used to harbour a third species known by its local name as "Balch-Pfärrit" (Fatio 1885), which is extinct today (Vonlanthen et al. 2012). Fatio (1890: Page 192) mentions that the "Balch-Pfärrit" was intermediate in phenotype between C. confusus and C. palaea of Lake Biel and has been considered by the local fishermen as a natural hybrid between the latter two species. Yet, based on the overall phenotype and ecological characters (spawning season and depth) Fatio (1885, 1890) considered the "Balch-Pfärrit" as an independent albeit variable species. This species increased in abundance during the study period of Fatio, which coincided with the completion of the Jura water correction from 1868-1878 that by passed the river Aare from Lake Thun to Lake Biel. This led some fishermen to suggest, that the "BalchPfärrit" might have come from Lake Thun. Fatio dismissed this because these fish did not resemble the species known by then from Lake Thun, this being C. albellus, C. alpinus and C. fatioi. He rather suggested the rise in abundance of the "Balch-Pfärrit" may have been caused by the lake level reduction of Lake Biel following the Jura water correction. The Lake Biel and Lake Neuchatel species' C. confusus, C. palaea, and C. candidus form distinct monophyletic clades in population neighbour-joining trees and one genetic cluster in a structure analysis, based on microsatellite and genomic AFLP loci (Hudson et al. 2011, 2016). Based on this and on the fact that the historically reported three whitefish species of Lake Biel (C. confusus, C. palaea and the "Balch-Pfärrit") were all winter spawners (Fatio 1885, 1890; Steinmann 1950) and that the ripe whitefish, that were caught in recent years in late summer in Lake Biel, were assigned with high probability to whitefish species from Lake Thun (Bittner 2009; Suppl. material 1: Figure S9), suggests that the ripe specimens caught in late summer in Lake Biel are unlikely to be the extinct "Balch-Pfärrit". Instead, we suggest that two Lake Thun whitefish species, C. profundus and C. albellus, have colonized Lake Biel. Interestingly, Steinmann (1950) reports that he was able to examine two ripe whitefish in September 1944 from Lake Biel that he thought resembled, based on their morphology (e.g., gill raker number, eye size), very much C. albellus from lakes Thun and Brienz.

Lakes Thun and Brienz in the Bernes Highlands today harbour the most speciose pre-alpine whitefish radiation. These lakes have also suffered the least anthropogenic pressures of all the large pre-alpine lakes in Switzerland. Species delineation and description in such rich radiations require an integrative approach to taxonomy, combining morphology with population genetics and ecology and extensive contemporary and historical specimen collections. Such work is also much needed for conservationminded fisheries management because, as we have shown here and others before us (Douglas and Brunner 2002; Bittner 2009; Dönz et al. 2018), human-made changes to the connectivity of water bodies as well as deliberate introductions, are increasing the distribution ranges of species and cause previously isolated biota to mix.

## Acknowledgements

Many thanks to the commercial fisherwomen and fishermen of lakes Thun, Brienz, and Biel and the cantonal fisheries authorities of Bern for their help, hospitality, and sharing of knowledge about the whitefish species of all three lakes. Especially the fisherwomen and fishermen Edith Klopfenstein, Kurt Klopfenstein, Elsbeth Abegglen, Beat Abegglen and Stefan Dasen and the cantonal fisheries warden of Bern, Benjamin Gugger, have been essential in many aspects and especially so in the collection of specimens of the rare species from all three lakes. We would also like to thank the people who helped in the field (Marta Reyes, Pascal Reichlin, Felix Vögtli, Jana Jucker, Andreas Taverna, Dr. Marcel Häsler, Jonas Walker, Dr. Philine Feulner, Dr. Miguel Leal, Dr. Jaime Mauricio-Anayas, Hélène Hefti, Dr. Florian Moser, Andrin Krähenbühl, David Frei, Dr. Kotaro Kagawa). We are grateful to Dr. Rudolf Müller in introducing OMS into the methods of how to read and age scales. We thank Dr. Lukas Rüber and the Natural History Museum in Bern for the adoption and curation of the Seehausen/ Steinmann whitefish collection and the Natural History Museum in Geneva for granting us access to their collection. Many thanks to Verena Kälin (www.rubia-tinctorum. ch) for the brilliant scientific illustrations. Finally, thanks to Dr. David Bittner and the Eawag Fish Ecology \& Evolution group and University of Bern Aquatic Ecology group for helpful discussions. This work was funded through the BAFU (Swiss Federal Officefor the Environment) grant "A2310.0132 Wasser", financing the project "Felchenvielfalt der Schweizer Seen", which is an action plan to revise the Swiss whitefish taxonomy granted to OS. Additional funding derived from the Department of Fish Ecology and Evolution at EAWAG and the division Aquatic Ecology and Evolution at the University of Bern.

## References

Alexander TJ, Vonlanthen P, Seehausen O (2017) Does eutrophication-driven evolution change aquatic ecosystems? Philosophical Transactions of the Royal Society B 72(1712): 20160041. https://doi.org/10.1098/rstb.2016.0041

Alexander TJ, Vonlanthen P, Seehausen O, Périat G, Raymond JC, Degiorgi F (2016) Artenvielfalt und Zusammensetzung der Fischpopulation im Bodensee. Schlussbericht, 68 pp.
Alexander TJ, Vonlanthen P, Seehausen O, Périat G, Degiorgi F, Raymond JC, Seehausen O (2015) Evaluating gillnetting protocols to characterize lacustrine Fish communities. Fisheries Research 161: 320-329. https://doi.org/10.1016/j.fishres.2014.08.009
Alonso JDG, Häberle S, Hüster Plogmann H, Schibler J, Schlumbaum A (2017) Millennia Long Co-Existence of Two Major European Whitefish (Coregonus spp.) Lineages in Switzerland Inferred from Ancient Mitochondrial DNA. Diversity 9(3): 1-34. https://doi. org/10.3390/d9030034
Ammann B, Haeberli W, Wohlfarth B, Merki R, Presler J, Schälchli U, Kühne A (1991) Landschaftsentwicklung im Seeland seit der letzten Eiszeit - Modelle und Realität. Modelle der Geomorphologie - Beispiele aus der Schweiz, Freiburg, 73-100.

Baur H, Leuenberger C (2011) Analysis of ratios in multivariate morphometry. Systematic Biology 60: 813-825. https://doi.org/10.1093/sysbio/syr061
Baur H, Kranz-Baltensperger Y, Cruaud A, Rasplus J-V, Timokhov AV, Gokhman VE (2014) Morphometric analysis and taxonomic revision of Anisopteromalus Ruschka (Hymnoptera: Chalcidoidea: Pteromalidae) - an integrative approach. Systematic Entomology 39: 691-709. https://doi.org/10.1111/syen. 12081
Berg LS (1932) Übersicht der Verbreitung der Süsswasserfische Europas. Zoogeographica 1: 107-208.
Bernatchez L, Dodson JJ (1991) Phylogeographic structure in mitochondrial DNA of the lake whitefish (Coregonus clupeaformis) and its relation to Pleistocene glaciations. Evolution 45: 1016-1035. https://doi.org/10.1111/j.1558-5646.1991.tb04367.x
Bernatchez L, Dodson JJ (1994) Phylogenetic relationships among Palearctic and Nearctic whitefish (Coregonus sp.) populations as revealed by mitochondrial DNA variation. Canadian Journal of Fisheries and Aquatic Sciences 51: 240-251. https://doi.org/10.1139/f94-310
Bernatchez L (2004) Ecological Theory of Adaptive Radiation - An Empirical Assessment from 395 Coregonine Fishes (Salmoniformes). In: Hendry AP, Stearns SC (Eds) Evolution Illuminated. Oxford University Press, Oxford, 175-207.
Bittner D (2009) Gonad deformations in whitefish (Coregonus spp.) from Lake Thun, Switzerland - A population genetic and transcriptomic approach. PhD Thesis, University of Bern, Bern.

Bittner D, Excoffier L, Largiader CR (2010) Patterns of morphological changes and hybridization between sympatric whitefish morphs (Coregonus spp.) in a Swiss lake: a role for eutrophication? Molecular Ecology 19: 2152-2167. https://doi.org/10.1111/j.1365-294X.2010.04623.x
Coulon A, Fitzpatrick JW, Bowman R, Stith BM, Makarewich A, Stenzler LM, Lovette IJ (2008) Congruent population structure inferred from dispersal behaviour and intensive genetic surveys of the threatened Florida scrub-jay (Aphelocoma coerulescens). Molecular Ecology 17: 1685-1701. https://doi.org/10.1111/j.1365-294X.2008.03705.x
Dönz CJ, Bittner D, Vonlanthen P, Wagner CE, Seehausen O (2018) Rapid buildup of sympatric species diversity in Alpine whitefish. Ecology and Evolution 2018: 1-15. https://doi. org/10.1002/ece3.4375
Douglas MR, Brunner PC (2002) Biodiversity of central alpine Coregonus (Salmoniformes): impact of one-hundred years of management. Ecological Applications 12: 154-172. https://doi.org/10.1890/1051-0761(2002)012[0154:BOCACS]2.0.CO;2
Douglas MR, Brunner PC, Bernatchez L (2003) Doassemblages of Coregonus (Teleostei: Salmoniformes) in the Central Alpine region of Europe represent species flocks? Molecular Ecology 8: 589-603. https://doi.org/10.1046/j.1365-294x.1999.00581.x
Fatio V (1885) Les corégones de la Suisse (féras diverses) classification et conditions de frai. Recueil Zoologie Suisse 1, 2: 649-665. [22, 23 pls$]$
Fatio V (1890) Faune des Vertébrés de la Suisse (Vol. V, 2partie). In: Georg H (Ed.) Genf und Basel, 569 pp. https://doi.org/10.5962/bhl.title. 124573
Feulner GD, Seehausen O (2018) Genomic insights into the vulnerability of sympatric whitefish species flocks. Molecular Ecology 28: 615-629. https://doi.org/10.1111/mec. 14977
Hantke R, Scheidegger AE (2007) Zur Entstehungsgeschichte der Berner Oberländer Seen In: Jahrbuch 2007 des Uferschutzverband Thuner- und Brienzersee, Uferschutzverband Thuner- und Brienzersee, Thun, 11-34.

Harrod C, Mallela J, Kahilainen K (2010) Phenotype-environment correlations in a putative whitefish adaptive radiation. Journal of Animal Ecology 79: 1057-1068. https://doi. org/10.1111/j.1365-2656.2010.01702.x
Heuscher J (1901) Thuner- und Brienzer-See. Ihre Biologischen und Fischerei-Verhältnisse. Druck E. Zwingli, Pfäffikon, 104 pp.
Hubbs CL, Lagler KF (1964) Fishes of the Great Lakes Region. University of Michigan Press, Ann Arbor, 213 pp.
Hudson AG (2011) Population genomics and ecology of parallel adaptive radiations: the Alpine lake whitefish. PhD Thesis, University of Bern, Bern.
Hudson AG, Lundsgaard-Hansen B, Lucek K, Vonlanthen P, Seehausen O (2016) Managing cryptic biodiversity: fine- scale intralacustrine speciation along a benthic gradient in Alpine whitefish (Coregonus spp.). Evolutionary Applications 10: 251-266. https://doi.org/10.1111/eva. 12446
Hudson AG, Vonlanthen P, Bezault E, Seehausen O (2013) Genomic signatures of relaxed disruptive selection associated with speciation reversal in whitefish. BMC Evolutionary Biology 13: 1-108. https://doi.org/10.1186/1471-2148-13-108
Hudson AG, Vonlanthen P, Seehausen O (2011) Rapid parallel adaptive radiation from a single hybridogenic ancestral population. Proceedings of the Royal Society B: Biological Sciences 278: 58-66. https://doi.org/10.1098/rspb.2010.0925
Hudson AG, Vonlanthen P, Müller R, Seehausen O (2007) Review: The geography of speciation and adaptive radiation in coregonines. Advances in Limnology 60: 111-146.
Ingram T, Hudson AG, Vonlanthen P, Seehausen O (2012) Does water depth or diet divergence predict progress towards ecological speciation in whitefish radiations? Evolutionary Ecology Research 14: 487-502.
Jacobs A, Carruthers M, Eckmann R, Yohannes E, Adams CE, Behrmann-Godel J, Elmer KR (2018) Rapid niche expansion by selection on functional genomic variation after ecosystem recovery. Nature Ecology and Evolution. https://doi.org/10.1038/s41559-018-0742-9
Jakobsson M, Rosenberg NA (2007) CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. Bioinformatics 23: 1801-1806. https://doi.org/10.1093/bioinformatics/btm233
Kekäläinen J, Soler C, Veentaus S, Huuskonen H (2015) Male Investments in High Quality Sperm Improve Fertilization Success, but May Have Negative Impact on Offspring Fitness in Whitefish. PLoS ONE 10: e0137005. https://doi.org/10.1371/journal.pone. 0137005
Kirchhofer A (1990) Limnologische und Ichtyologische Untersuchungen im Brienzersee unter besonderer Berücksichtigung der Differenzierung der sympatrischen Felchenpopulationen. Dissertation, Phil.-nat. Fakultät der Universität Bern.
Kirchhofer A (1995) Growth characteristics of coregonid populations in three lakes with different trophic states and decreasing nutrient concentrations. Archiv für Hydrobiologie Special Issues Advances in Limnology 46: 61-70.
Kirchhofer A, Tschumi PA (1986) Age structure and growth of coregonid fish populations in Lake Thun. Archiv für Hydrobiologie - Beiheft Ergebnisse der Limnologie 22: 303-318.
Kottelat M (1997) European freshwater fishes. A heuristic checklist of the freshwater fishes of Europe (exclusive of former USSR), with an introduction for non-systematists and comments on nomenclature and conservation. Biologia, Bratislava, Section Zoology 52: 1-271. [Suppl. material 1]

Kottelat M, Freyhof J (2007) Handbook of European Freshwater Fishes. Kottelat, Cornol, and Freyhof, Berlin, 646 pp.
László Z, Baur H, Tóthmérész B (2013) Multivariate ratio analysis reveals Trigonoderus pedicellaris Thomson (Hymenoptera, Chalcidoidea, Pteromalidae) as a valid species. Systematic Entomology 38: 753-762. https://doi.org/10.1111/syen. 12026
Lehtonen H, Nylund V (1995) An atlas of fish scales. 2. Salmonid species found in European fresh waters. Samara Publishing Limited, Cardigan, 55 pp .
Lundsgaard-Hansen B, Matthews B, Vonlanthen P,Taverna A, Seehausen O (2013) Adaptive plasticity and genetic divergence in feeding efficiency during parallel adaptive radiation of whitefish (Coregonus spp.). Journal of Evolutionary Biology 26: 483-498. https://doi. org/10.1111/jeb. 12063
Maurer V, Guthruf K (2005) Brienzerseefelchen - Magenanalysen - Schlussbericht. Bericht i.A. Fischereiinspektorat des Kantons Bern. HYDRA: 1-18.

Müller R, Breitenstein M, Bia MM, Rellstab C, Kirchhofer A (2007) Bottom-up control of whitefish populations in ultra- oligotrophic Lake Brienz. Aquatic Sciences 69: 271-288. https://doi.org/10.1007/s00027-007-0874-5
Öhlund G, Bodin M, Nilsson KA, Öhlund S-O, Mobley KB, Hudson AG, Peedu M, Brännström A, Bartels P, Præbel K, Hein CL, Johansson P, Englund G (2020) Ecological speciation in European whitefish is driven by a large-gaped predator. bioRxiv. Evolution Letters.https://doi.org/10.1002/evl3.167
Østbye K, Naesje TF, Bernatchez L, Sandlund OT, Hindar K (2005) Morphological divergence and origin of sympatric populations of European whitefish (Coregonus lavaretus L.) in Lake Femund, Norway. Journal of Evolutionary Biology 18: 683-702. https://doi.org/10.1111/ j.1420-9101.2004.00844.x

Patton JC, Gallaway BJ, Fechhelm RG, Cronin MA (1997) Genetic variation of microsatellite and mitochondrial DNA markers in broad whitefish (Coregonus nasus) in the Colville and Sagavanirktok rivers in northern Alaska. Canadian Journal of Fisheries and Aquatic Science 54: 1548-1556. https://doi.org/10.1139/f97-062
Præbel K, Knudsen R, Siwertsson A, Karhunen M, Kahilainen KK, Ovaskainen O, Østbye K, Peruzzi S, Fevolden S-E, Amundsen P-A (2013) Ecological speciation in postglacial European whitefish: rapid adaptive radiations into the littoral, pelagic, and profundal lake habitats. Ecology and Evolution 3: 4970-4986. https://doi.org/10.1002/ece3.867
Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. Genetics 155: 945-959.
R Studio Team (2015) RStudio: Integrated Development for R. RStudio, Inc., Boston.
Roesch C, Lundsgaard-Hansen B, Vonlanthen P, Taverna A, Seehausen O (2013) Experimental evidence for trait utility of gill raker number in adaptive radiation of a north temperate fish. Journal of Evolutionary Biology 26: 1578-1587. https://doi.org/10.1111/jeb. 12166 Rogers SM, Marchand MH, Bernatchez L (2004) Isolation, characterization and cross-salmonid amplification of 31 microsatellite loci in the lake whitefish (Coregonus clupeaformis, Mitchill). Molecular Ecology Notes 4: 89-92. https://doi.org/10.1046/j.1471-8286.2003.00578.x
Rogers SM, Bernatchez L (2007) The genetic architecture of ecological speciation and the association with signatures of selection in natural lake whitefish (Coregonus sp. Salmonidae) species pairs. Molecular Biology and Evolution 24: 1423-1438. https://doi.org/10.1093/molbev/msm066

Roy D, Lucek K, Walter RP, Seehausen O (2015) Hybrid 'superswarm' leads to rapid divergence and establishment of populations during a biological invasion. Molecular Ecology 24: 5394-5411. https://doi.org/10.1111/mec. 13405
Rufli H (1978) Die heutigen sympatrischen Felchenpopulationen (Coregonus spp.) des Thunerund Bielersees und ihre Morphologie. Schweizerische Zeitung für Hydrologie 40: 7-31. https://doi.org/10.1007/BF02502369
Rufli H(1979) Ernährung und Wachstumder Felchen populationen (Coregonus spp.) des Thuner-und Bielersees. Schweizerische Zeitschrift für Hydrologie 41: 64-93. https://doi. org/10.1007/BF02551760
Selz OM (2008) Building a link between dN and dC stable isotopes signatures and the ecological phenotypic forms of Whitefish (Coregonus laveratus (L.)) in 4 Swiss Lakes. B.Sc. Thesis: University of Bern, Bern.
Steinmann P (1950) Monographie der schweizerischen Koregonen. Beitrag zum Problem der Entstehung neuer Arten. Spezieller Teil. Schweizerische Zeitung für Hydrologie 12: 340491. https://doi.org/10.1007/BF02489885

Surbeck G (1917) Der Brienzlig (Coregonus exiguus Klunz. var. albellus Fatio). Neue Beobachtungen, Fang- und Zuchtversuche. Schweizerisches Departement des Innern. Inspektion für Forstwesen, Jagd und Fischerei. Buchdruckerei HNERI KUNZ, Pfäffikon.
Svärdson G (1957) The Coregonid Problem. VI. The Palaearctic species and their Intergrades. Annual Report Drottningholm 38: 267-356.
Turgeon J, Estoup A, Bernatchez L (1999) Species flock in the North American Great Lakes: Molecular ecology of Lake Nipigon Ciscoes (Teleostei: Coregonidae: Coregonus). Evolution 53: 1857-1871. https://doi.org/10.2307/2640446
Vonlanthen P, Périat G (2018) Standardisierte Befischung Bielersee Resultate der Erhebungen vom September 2017. Aquabios GmbH. Auftraggeber: Amt für Landwirtschaft und Natur, Fischereiinspektorat, Kanton Bern, 49 pp.
Vonlanthen P, Périat G (2013) Artenvielfalt und Zusammensetzung der Fischpopulation im Brienzersee. ProjetLac, Eawag, Kastanienbaum, 44 pp. [in Zusammenarbeit mit: Seehausen O, Dönz C, Rieder J, Brodersen J, Degiorgi F, Guillard J, Colon M]
Vonlanthen P, Périat G, Dönz C, Hellmann J, Alexander TJ, Seehausen O (2015) Artenvielfalt und Zusammensetzung der Fischpopulation im Thunersee. Projet Lac, Eawag, Kastanienbaum, 41 pp .
Vonlanthen P, Bittner D, Hudson A, Young KA, Müller R, Lundsgaard-Hansen B, Roy D, Di Piazza S, Largiader CR, Seehausen O (2012) Eutrophication causes speciation reversal in whitefish adaptive radiations. Nature 482: 357-363. https://doi.org/10.1038/ nature 10824
Vonlanthen P, Roy D, Hudson AG, Largiadèr CR, Bittner D, Seehausen O (2009) Divergence along a steep ecological gradient in lake whitefish (Coregonus sp.). Journal of Evolutionary Biology 22: 498-514. https://doi.org/10.1111/j.1420-9101.2008.01670.x
William EE (1972) The Origin of Faunas. Evolution of Lizard Congeners in a Complex Island Fauna: A Trial Analysis. In: Dobzhansky T, Hecht MK, Steere WC (Eds) Evolutionary Biology. Springer, New York. https://doi.org/10.1007/978-1-4684-9063-3_3
Winkler KA, Pamminger-Lahnsteiner B,Wanzenböck J, Weiss S (2011) Hybridizationand restricted gene flow between native and introduced stocks of Alpine whitefish (Coregonus sp.)
across multiple environments Molecular Ecology 20: 456-472. https://doi.org/10.1111/ j.1365-294X.2010.04961.x

## Supplementary material I

## Tables S1-S8, Figures S1-S11

Authors: Oliver M. Selz, Carmela J. Dönz, Pascal Vonlanthen, Ole Seehausen
Data type: measurements
Explanation note: Table S1. PC-loadings of all characters (values $>0.1$ or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R-squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all contemporary specimens of the formerly described species C. alpinus, C. albellus, and C. fatioi and their types and the newly described species C. steinmanni and C. profundus from Lake Thun. Table S2. PC-loadings of all characters (values > 0.1 or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R -squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all contemporary specimens of the six species C. alpinus, C. steinmanni, C. fatioi, C. albellus, C. acrinasus and C. profundus from Lake Thun. Table S3. PC-loadings of all characters (values $>0.1$ or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R-squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all contemporary specimens of the four species C. alpinus, C. fatioi, and C. albellus and the newly described species C. brienzii from Lake Brienz. Only specimens smaller than 163.5 mm were used for the analysis to overcome allometry issues. Table S4. PC-loadings of all characters (values $>0.1$ or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R-squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all specimens of the three species $C$. alpinus, C. fatioi, and C. albellus and the newly described species C. brienzii from Lake Brienz. Table S5. PC-loadings of all characters (values $>0.1$ or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R -squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all whitefish species from Lake Thun and the three specimens of whitefish from Lake Biel that were assigned by genotype to C. albellus or C. profundus. Table S6. Frequency of occurrence of meristic values in the six whitefish species from Lake Thun and the four whitefish species from Lake Brienz. Table S7. Frequency of occurrence of the meristic values in the four whitefish species from Lake Constance, Switzerland, C. gutturosus Gmelin, 1818, C. arenicolus Kottelat, 1997, C. macrophthalmus Nüsslin, 1882, C. wartmanni Bloch, 1784, and the partially allochthonous whitefish species C. acrinasus from Lake Thun, Switzerland, that has ancestry contributions from Lake Constance whitefish. Table S8. PC-loadings of all characters (values $>0.1$ or $<-0.1$ are highlighted in bold), eigenvalues, proportion of explained variance, and R-squared from a linear regression of shape vs. isosize of the retained first three PC-axes from a PCA with all contemporary pecimens of the formerly described species C. alpinus, C. albellus, and
C. fatioi, their types, the newly described species C. steinmanni and C. profundus from Lake Thun and the museum specimens with the river Aare as type-locality. Figure S1. Illustration of the morphological body character measurements (see Table 1 in materials and methods for a detailed description of each character). Figure S2. Illustration of the morphological head character measurements (see Table 1 in materials and methods for a detailed description of each character). Figure S3. SSpawning distribution of the different whitefish species at one spawning site in Lake Thun (Faulensee: 46.673725 / 7.707944 ) and Lake Brienz (Iseltwald: 46.712079 / 7.961261 ) during the complete spawning season of the whitefish species from late summer to late winter. Figure S4. Size (TL=total length in mm ) at age of the species $C$. alpinus (A), C. albellus (B) C. steinmanni (C), C. fatioi (D), C. acrinasus (E), and C. profundus (F) from Lake Thun. Figure S5. Size (TL = total length in mm ) at age of the species C. alpinus (A), C. albellus (B), C. brienzii (C), and C. fatioi (D) from Lake Brienz. Figure S6. Size (TL = total length in mm ) of 3-year-old fish of the species C. acrinasus (red), C. alpinus (blue), C. steinmanni (light blue), C. albellus (green), C. profundus (orange), C. fatioi (violet) and C. brienzii (black) from lakes Thun (A) and Brienz (B). Figure S7. The "cheetah look" of some specimens of C. albellus from Lake Brienz with rather large pigmented dots arranged more or less in rows on the upper dorsum. Figure S8. The three main colouration types found on the dorsum and the dorsal part of the flanks above the lateral line in the whitefish species of Lakes Thun and Brienz. Figure S9. Scatterplot of the first three shape PC axes and the total number of gill rakers for the contemporary whitefish species from Lake Thun and the three contemporary specimens of whitefish from Lake Biel that were caught on the 9th of October 2016 and were genetically assigned (see Materials and methods for details on the genetic assignments) to C. albellus (specimens NMBE-1077160 with $84 \%$ and NMBE-1077159 with $94 \%$ genetic assignment) or C. profundus (specimen NMBE-1077158 with $96 \%$ genetic assignment). Figure S10. Map of Lake Thun, Brienz, and Biel and the locations where all contemporary specimens of the six species of Lake Thun (C. alpinus, C. steinmanni, C. fatioi, C. albellus, C. acrinasus, and C. profundus) and the four species of Lake Brienz (C. alpinus, C. brienzii, C. fatioi, C. albellus) were caught as well as the locations of the contemporary specimens of the 2 whitefish species from Lakes Thun and Brienz that were caught in Lake Biel. Figure S11. Principal Component Analysis showing that the whitefish specimens caught in the river Aare at the end of the 19th century and midtwentieth century (locality: either near the city of Bern (specimens: NMBE-1013589 and NMBE-1013603), below the outflow of Lake Thun near the city of Thun (specimens: Eawag-363-1 and Eawag-363-2) or in the Bödeli-Aare between the lakes Thun and Brienz near the city of Interlaken (specimens: Eawag-373-1 and Eawag-373-2) lie within the range or adjacent to the range of the contemporary specimens of C. fatioi. Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.989.32822.suppl1


[^0]:    * Contributed equally as the first authors.

[^1]:    Copyright Enrique Baquero et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

