RESEARCH ARTICLE



Revision of Diplocirrus Haase, 1915, including Bradiella Rullier, 1965, and Diversibranchius Buzhinskaja, 1993 (Polychaeta, Flabelligeridae)

Sergio I. Salazar-Vallejo^{1,†}, Galina Buzhinskaja^{2,‡}

l Depto. Ecología Acuática, ECOSUR, Unidad Chetumal, México 2 Zoological Institute, Russian Academy of Sciences, Saint-Petersburg, Russia

turn:lsid:zoobank.org:author:7D3C2D7E-5D08-45F7-A1E4-84A1563EA14F urn:lsid:zoobank.org:author:743B5217-0701-409A-8804-AF75B92729BD

Corresponding author: Sergio I. Salazar-Vallejo (savs551216@hotmail.com)

Academic editor: Robert Blakemore Received 10 December 2010 Accepted 5 May 2011 Published	1 15 June 2011
urn:lsid:zoobank.org:pub:7BC0CD4B-19D2-453E-866D-6A7B4B76E568	

Citation: Salazar-Vallejo SI, Buzhinskaja G (2011) Revision of *Diplocirrus* Haase, 1915, including *Bradiella* Rullier, 1965, and *Diversibranchius* Buzhinskaja, 1993 (Polychaeta, Flabelligeridae). ZooKeys 106: 1–45. doi: 10.3897/zooKeys.106.795

Abstract

Diplocirrus Haase, 1915, includes flabelligerids having cylindrical to club-shaped bodies, with cirriform papillae, multiarticulate chaetae in both parapodial rami, 8 branchial filaments of two types (thick and rarely lamellate, or cirriform), gonopodial lobes in chaetigers 5 or 6, or multiple gonopores along some anterior chaetigers. Bradiella Rullier, 1965, has included only the type species: B. branchiata Rullier, 1965, described from Eastern Australia. The original description has been overlooked and it lacked enough details on branchial and chaetal features. Diversibranchius Buzhinskaja, 1993, with D. nicolaji Buzhinskaja, 1994, as the type species, was introduced for a similar species from the Japan Sea. These two monotypic genera share the same morphologic features with Diplocirrus, and are herein regarded as its junior synonyms. As herein redefined, Diplocirrus includes, besides its type species, D. glaucus (Malmgren, 1867) from Scandinavia : D. branchiatus (Rullier, 1965), comb. n. from Queensland, Australia, D. capensis Day, 1961 from South Africa, D. erythroporus Gallardo, 1968 from Vietnam, D. hirsutus (Hansen, 1882) from Arctic and subarctic regions, D. incognitus Darbyshire & Mackie, 2009 from South Africa, D. kudenovi sp. n. from off Western Mexico, D. longisetosus (von Marenzeller, 1890) restricted to the Bering Sea, D. micans Fauchald, 1972 from deep water off Oregon and Western Mexico, D. nicolaji (Buzhinskaja, 1994), comb. n. from the Japan Sea, D. normani (McIntosh, 1908), comb. n. from Scandinavia, D. octobranchus (Hartman, 1965), comb. n. from off New England, and D. stopbowitzi Darbyshire & Mackie, 2009 from the Irish Sea.

Copyright Sergia I. Salazar-Valleja, Galina Buzhinskaja. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Keywords

Multiarticulated capillaries, body papillae, soft-bottoms

Introduction

The delineation of flabelligerid genera has been problematic since Grube (1877a); especially because the eversible anterior end, carrying the branchiae and palps, is rarely exposed. Branchial and chaetal features were employed to propose most genera, but their delineation was not clear-cut in most instances, especially because the branchiae are rarely everted. Thus, Diplocirrus was proposed by Haase (1915:194) following some earlier indications by von Marenzeller (1889:130), and by de Saint-Joseph (1898:366). These two authors have commented on the need to separate the species placed in *Sty*larioides delle Chiaje, 1831 by using the branchial arrangement. The species transferred to Diplocirrus have four pairs of cirriform, heteromorphic branchiae: the four distal filaments are shorter and thicker, whereas the proximal two pairs include thinner, longer filaments. The branchial filaments in the distal or posterior row differ from those found on the proximal or anterior row; they are basally prismatic due to the fact that when specimens are alive, they are closely packed making a branchial wall. Handling specimens often causes the branchial filaments to separate from the others, such that their lateral connections are not noticed. Further, although the posterior row filaments are thicker than the proximal row filaments, they are dehiscent. The proximal filaments are cirriform separated as two lateral pairs, but are completely free from each other, such that in living or preserved specimens, they look loose and and are deshiscent as well.

It is noteworthy that *D. capensis* Day, 1961 was described as having all branchiae of the same size and width, neurochaetae distally falcate, and without a cephalic cage. This combination of characters made Day expand the generic diagnosis with some hesitation (Day 1961:510). The whole body was later illustrated (Day 1967:665, Fig. 32.4e), and the emended diagnosis was confirmed. On the other hand, Hartman (1965:178) described *Ilyphagus octobranchus* and made some comments on its affinity with *D. capensis*; later, Day (1973:106–107) repeated her observations and because of their proximity, regarded *Ilyphagus* Chamberlin, 1919 as a junior synonym to *Diplocirrus*. As stated elsewhere (Salazar-Vallejo et al. 2008:204), this synonymy cannot be supported because of, among other things, the striking differences in body shape, cephalic cage development, and type of neurochaetae. Further, Darbyshire & Mackie (2009:96), after studying the type material, have found that it has the *Diplocirrus*, typical branchial pattern, and they compiled a table with the morphological characters for most species in the genus.

Chamberlin (1919b:397) introduced *Saphobranchia* for *Stylarioides longisetosus* von Marenzeller, 1890; however, he overlooked the revision by Haase (1915) who had established *Diplocirrus*, including this species into his generic definition. Thus, *Saphobranchia* is a junior synonym of *Diplocirrus*.

Further, eight genera in the polychaete family Flabelligeridae de Saint-Joseph, 1894 have been regarded as monotypic: *Bradabyssa* Hartman, 1967, *Bradiella* Rullier, 1965, *Coppingeria* Haswell, 1892, *Diversibranchius* Buzhinskaja, 1993, *Flabelliderma* Hartman, 1969, *Pantoithrix* Chamberlin, 1919, *Poeobius* Heath, 1930, and *Therochaetella* Hartman, 1967. The proposal of some of these genera may be explained by the lack of a revisionary work that clarifies the generic delimitations in the family. For example, *Flabelliderma* has been redefined recently and it is no longer a monotypic genus (Salazar-Vallejo 2007). *Coppingeria* has been merged into *Stylarioides* delle Chiaje, 1831, as indicated elsewhere (Salazar-Vallejo 2011a), and *Therochaetella* has been regarded as a junior synonym of *Trophoniella* Caullery, 1944 (Salazar-Vallejo 2011b).

On the other hand, *Bradiella* has been only known by its type species, *B. branchiata* Rullier, 1965, which was described from Moreton Bay, Queensland, Australia. Spies (1975) studied some specimens from the type locality, but they were identified as *Diplocirrus* cf. *capensis* Day, 1961. This was an unfortunate decision because *B. branchiata* was overlooked by posterior scientists working in the same area. Specimens from a similar species were found in the Sea of Japan by Buzhinskaja (1994); she documented several interesting morphological features, and concluded they were different enough from *D.* cf. *capensis*. Thus, she proposed a new genus: *Diversibranchius*, because of the strikingly different branchial filaments.

With this contribution, we revise *Diplocirrus* and regard *Bradiella* and *Diversibranchius* as junior synonyms based on review of type, topotype, and additional materials. *Diplocirrus* is amended and now contains 13 species that live on soft bottoms in sublittoral depths throughout the world.

Materials and methods

The relative size of notochaetae and their articulation pattern are based on median chaetigers, about chaetiger 10. As in other contributions in this series, specimens were photographed using available equipments; specimens were often temporarily stained with an over-saturated alcoholic methyl green solution. When available, head are depicted in frontal views once branchiae and palps are removed. Plates were prepared by combining several photographs by hand or by using HeliconFocus. Type and non-type materials belong to the following institutions.

Museum acronyms

AM	Australian Museum, Sydney.
BMNH	The Natural History Museum, London.
CAS	California Academy of Sciences, San Francisco.
ECOSUR	Colección de Referencia, El Colegio de la Frontera Sur, Chetumal.
IRFA	Institut de Recherche Fondamental et Appliquée, Université Catholique
	de l'Ouest, Angers, France.

LACM-AHF	Natural History Museum of Los Angeles County, Allan Hancock Foun-
	dation Polychaete Collection.
MNHN	Museum National d'Histoire Naturelle, Paris.
NTM	Museum and Art Gallery of the Northern Territory, Darwin, Australia.
QM	Queensland Museum, South Brisbane, Australia.
SAM	South Australian Museum, Adelaide (GR: Greg Rouse pers. coll.).
SMF	Forschungsinstitut und Naturmuseum Senckenberg, Frankfurt.
USNM	National Museum of Natural History, Smithsonian Institution, Wash-
	ington.
ZIRAS	Zoological Institute, Russian Academy of Sciences, Sankt Peterburg.
ZMUB	Zoologisk Museum, Univesiteet i Bergen, Bergen.

Results

Morphology

Body shape and color. The body is clavate, subcylindrical, with the anterior few chaetigers often swollen and longer than other segments. Although most species are pale or alternatively take the sediment pigmentation on their body wall, at least along the first few chaetigers, Scandinavian species have been separated by using their overall pigmentation. Thus, *D. hirsutus* has been regarded as reddish, at least along few anterior chaetigers, whereas *D. glaucus* and *D. normani* are grayish. These differences might be due to the sediment particles, and thus be variable depending on the sediment quality rather than a diagnostic feature.

Cephalic cage. The first chaetiger is poorly developed in *Diplocirrus* species. However, the relative size of the cephalic cage as well as the number of chaetae per bundle can be used to separate similar species.

Sediment cover. The body of the members of the *Diplocirrus* species is variously covered by sediment particles. Fine sediment particles may be adhered to each papilla, whereas larger particles are often trapped between papillae; they are rarely forming a sediment crust. Becausee body papillae are fragile, brushing off the excess of sediment might also remove the papillae, such that particle removal should be done carefully.

Body papillae. The relative shape and size of the body papillae has been used to separate similar species. The papillae can be short, being about 3–5 times longer than wide and giving a velvety appearance, or they can be long, being about 8–10 times longer than wide and giving a hirsute outlook. Further, their relative size in relation to notochaetae has been included in the key and descriptions below as an additional means to separate similar species.

Prostomium. The prostomium includes a short lobe carrying two pairs of eyes and a posterior projection, the caruncle, which tapers posteriorly, or is distally expanded. To observe this feature, branchial filaments must be removed and to decide if the caruncle is posteriorly expanded, an exploration throughout its length is needed.

5

Branchiae. Branchial filaments are made in two different types and can be separated in two series in relation to the prostomium. The proximal series is the anterior row and the distal one is the posterior row. The posterior row includes prismatic or cuneiform filaments, whereas the anterior row is made of cirriform filaments. The posterior row includes four filaments laterally fused to each other, forming a branchial wall that was illustrated by Haase (1915:29, 197, Fig. 5). This wall is formed because each branchia has two lateral sockets keeping them together, and making their separation difficult. Once separated, each filament is more or less triangular in cross section, but there are two basic modifications; filaments have ciliary bands in most species, whereas in a few of them, filaments are convoluted, with transverse ridges along their surface. Further, because in the latter species the dorsal side is often projected with a long wing, whereas the ventral side might be widened by the presence of multiple blades or lamellae, each filament has a prismatic or cuneiform appearance. These lamellae might be restricted to the proximal or cirriform branchial filaments, even in those species lacking the complex features seen in some posterior branchial filaments. Because they are variable within species, the shape of the filaments and the presence of ciliary bands are not useful for distinguishing genera. These blades are made by either a single series of convoluted filaments, or by a series of transverse filaments arranged as twin blades, but marginally independent of the following blade. Further, these blades can extend over different regions along the back of each branchial filament. The anterior row includes four filaments too, but they are separated in two lateral pairs. Each filament is cirriform, thinner, usually provided with a series of transverse ciliated ridges and, if provided with filamentous blades, hence lamellate, they are more or less restricted to the basal region. As in other flabelligerids carrying two series of branchial filaments as in Pherusa, there are some basal branchial knobs between the posterior branchial filaments. They resemble some short, rounded reinforcements present in sabellids or serpulids, and their relative development, whenever evident, might be useful to separate similar species.

Chaetae. All chaetae in *Diplocirrus* are multiarticulated with notochaetae thinner than neurochaetae. The multiarticulated notochaetae provide useful diagnostic features by their relative size, in relation to body width, or by the relative size of articles along the chaetae. Thus, articles are regarded as short if they are wider than long, medium-sized if they are as long as wide, and long if they are longer than wide. This variation in the articulation pattern is also present in neurochaetae and because it is a conservative feature, is often used to separate similar species.

Gonopodial lobes and gonopores. Adult members of some *Diplocirrus* species carry two projected lobes in chaetigers 4 or 5 which were regarded as neprhidial papillae since Haase (1915). However, nephridial lobes are restricted to the branchial plate, the projected, segmental lobes have a reproductive role and are consequently regarded as gonopodial. Their position in a given chaetiger, as well as their relative color and shape can be used to separate similar species. Three species lack gonopodial lobes, but have several pairs of ventral, rounded, reddish or dark orange structures of unknown function; pending a histological confirmation, they are herein regarded as gonopodial.

These multiple paired structures have been described for *D. erytrhoporus* Gallardo, 1968, and *D. glaucus orientalis* Gibbs, 1971, which is regarded as a junior synonym for the former. Because some species of *Diplocirrus* lack gonopodial lobes, and because they might be present only during reproduction, their presence or absence could not be employed as a generic diagnostic feature, and the multiple gonopores would be in the same condition.

Systematics

Class Polychaeta Grube, 1850 Order Flabelligerida Pettibone, 1982 Family Flabelligeridae de Saint-Joseph, 1894

Diplocirrus Haase, 1915

http://species-id.net/wiki/Diplocirrus

Diplocirrus Haase 1915:194; Day 1967:664–666; Fauchald 1977:116; Darbyshire & Mackie 2009:93.
Saphobranchia Chamberlin 1919:397; proposed for Stylarioides longisetosa von Marenzeller, 1890.
Bradiella Rullier 1965:188.

Diversibranchius Buzhinskaja 1994:231.

Type species. Trophonia glauca Malmgren, 1867, by original designation.

Diagnosis. Body clavate or subcylindrical, often anteriorly swollen. Cephalic cage variably developed. Body papillae abundant, short giving a velvety appearance, or very long, giving a hirsute outlook, sometimes adhering sediment particles. All chaetae multiarticulated capillaries; neurochaetae thicker, sometimes falcate. Branchiae sessile, 4 pairs, distal branchiae thicker, often shorter, proximal branchiae thinner, often longer, sometimes basally lamellate. Gonopodial papillae present in chaetiger 4 or 5, or a series of paired ventrolateral gonopores along some anterior chaetigers.

Remarks. Haase (1915) proposed *Diplocirrus* for those species formerly included in *Stylarioides* having two different sizes of branchiae, and multiarticulated capillaries only. Some species currently included in the genus had been previously described in either *Trophonia* or *Stylarioides*. However, as an independent genus, it differs by having two different sizes of branchiae, and all chaetae are multiarticulated capillaries.

Webster and Benedict (1887:730) proposed Zorus, with Zorus sarsi as the type and only species. They indicated that it had a body anteriorly swollen, becoming thinner posteriorly, only with capillary chaetae, and stated that branchiae and palps arise from an eversible stalk but gave no details on the size relationship of branchiae. Hartman (1961:122) regarded Zorus as a junior synonym of *Piromis* Kinberg, 1867. However, because of the body form and chaetal features, it rather resembles *Diplocirrus*, because *Piromis* has few papillae arranged in longitudinal rows and sometimes bifid neurohooks, which were not found in *Zorus*. Webster (1879:46) had already stated the differences among capillary chaetae and ventral hooks when he described another flabelligerid; so, there is no room for any such confusion. The only illustration provided by Webster and Benedict (1887, Pl. 5, Fig. 67), shows a cross section of a middle segment with very long chaetae, and long papillae. These features resemble *D. hirsutus* (Hansen, 1879), which is comm in the Bay of Fundi (Appy et al. 1980:32). However, because there is no type material, the generic definition did not include a size relationship of branchial filaments, and the description and illustration lack critical information, *Zorus sarsi* has been regarded as indeterminable (Salazar-Vallejo et al. 2008).

The record by Langerhans (1881:102–103, Figs. 14a–d) of *Brada inhabilis* apparently belongs to *Diplocirrus*, but the illustrations and characters are not clear enough to assign it to any species. The record of *D. longisetosus* by Rullier (1964:1094) off Cameroon belongs in *Pycnoderma* Grube, 1877b, likewise *D. erythroporus* Gallardo, 1968, includes *D. glaucus orientalis* Gibbs, 1971 (:181, no figures; orange globular papillae below each neuropodium in chaetigers 4–14(16)), and might also include the Indian Ocean record of *D. glaucus* by Fauvel (1932:186–187, Fauvel 1953:353, Fig. 184a–d).

As stated above, *D. capensis* Day (1961:509, Fig. 9a–f, South Africa), was described as lacking cephalic cage, with branchiae of a single kind, and with distally hooked neurochaetae (against generic diagnosis, *cf* Fauvel, Støp-Bowitz). The same different group might include *Diplocirrus* sp A Milligan (1984). The records of the former, originally described from Southern Africa (Day 1967:666, Figs. 32.4e–j; Fauchald 1972:4120), for North Carolina (Day 1973:105–107), and the Gulf of Mexico (Milligan 1984:47.9–11, Figs. 47.5–6), require confirmation to define if they belong to the same species. As stated above, Darbyshire and Mackie (2009) have clarified the branchial features for *D. capensis*, whereas the other records remain unsolved.

These differences prompted Day (1961:510), to propose a misfortunate redefinition of *Diplocirrus*, because the branchial features have been employed to establish it by Haase (1915:26, 194). Especially because the posterior row of branchiae are not just thicker than the anterior row filaments; rather, they tend to be closely packed, with each filament laterally fused forming a branchial wall. Further, the cirriform thinner branchiae are contractile, and if they were observed completely relaxed by Day, he might have had the impression that they were of about the same thickness. Later, Day (1973:106–107) modified the generic definition of *Diplocirrus* concluding that it would also include *Ilyphagus* Chamberlin, 1919. However, as stated above, this second emendation is problematic as well, because of marked differences in neurochaetae, because in the species of *Ilyphagus* neurochaetae are aristate neurospines whose handle is made of fused or anchylosed, short articles, and a hyaline fragile tip, whereas in *Diplocirrus* there are only multiarticulate, often falcate, neurochaetae.

Bradiella Rullier (1965:190) was compared with *Diplocirrus* and *Brada* Stimpson, 1854. The branchial features were incompletely described (see below); it was regarded as different from these two genera because of the branchiae, and because it lacks gonopodial lobes. The potential differences between *Bradiella* and *Diplocirrus* would

be that in Bradiella there are no gonopodial lobes in chaetigers 4-5, but gonopodial lobes have not been recorded in some Diplocirrus species at all. Further, the surface of branchial filaments is very complex in *Bradiella*, because it is provided with lamellate complex filaments, in contrast to the cirriform or tapering filaments which might barely have some ciliated bands, but some *Diplocirrus* species have a complex lamellar structure along the branchial filaments bases. Spies (1975) studied specimens from the type locality, Moreton Bay, Queensland, Australia, but overlooking the paper by Rullier (1965), identified them as Diplocirrus cf. capensis Day, 1961. He noticed that the branchiae include eight filaments, not just two as stated by Rullier, with four cirriform and four lamellate filaments. Spies (1975, Pl. 6, Fig. 18) illustrated a (lateral) dorsal spoon-like branchia provided with a flat lateral lobe, and a series of independent branchial blades. Thus, because there are variations in the presence of gonopodial lobes and in the development of lamellar structures in branchial filaments, the only difference to separate the Bradiella-like species would be the presence of paired ventrolateral pores. However, because there is no other major difference in chaetal types, Diplocirrus and Bradiella are regarded as synonyms.

Because of the rediscovery of these peculiar branchial features, Buzhinskaja (1994) established *Diversibranchius*. However, she overlooked Rullier (1965) as well, and compared his specimens with *Diplocirrus*, stressing its resemblance with *D*. cf. *capensis*. She found that branchiae were of two types, cirriform and prismatic, or cuneiform, provided with foliose projections, and illustrated that both have convoluted branchial lamellae giving the impression of a series of independent blades, as was illustrated by Spies (1975). *Bradiella* and *Diversibranchius* Buzhinskaja, 1994 resemble each other by having two different types of branchiae, short to long body papillae, and multi-articulated neurohooks. These two genera are herein regarded as junior synonyms to *Diplocirrus*, such that the type species are redescribed, and transferred and newly combined into *Diplocirrus*.

As herein redefined, *Diplocirrus* includes, besides the type species from Scandinavia, *D. branchiatus* (Rullier, 1965) comb. n. from Queensland, Australia, *D. capensis* Day, 1961 from South Africa, *D. erythroporus* Gallardo, 1968 from Vietnam, *D. hirsutus* (Hansen, 1882) from Arctic and subarctic regions, *D. incognitus* Darbyshire & Mackie, 2009 from South Africa, *D. kudenovi* sp. n. from off Western Mexico, *D. longisetosus* (von Marenzeller, 1890) restricted to the Bering Sea, *D. micans* Fauchald, 1972 from deep water off Oregon and Western Mexico, *D. nicolaji* (Buzhinskaja, 1994) comb. n. from the Japan Sea, *D. normani* (McIntosh, 1908) comb. n. reinst., from Scandinavia, *D. octobranchus* (Hartman, 1965) from off New England, and *D. stopbowitzi* Dabryshire & Mackie, 2009, from the Irish Sea.

Two of these species (*D. incognitus* and *D. stopbowitzi*), have been recently described and only their diagnosis and illustrations are included. On the other hand, three other currently undescribed species are informally characterized but not all have been included in the key because the quality of the materials; one is from Morocco, another one from off Sri Lanka, and the other from Antarctica. The species can be separated using several morphological features as stated below.

Key to species of Diplocirrus Haase, 1915

1	Body papillae abundant
_	Body papillae scarce, long, tunic looks bare
2 (1)	Body papillae short, giving a velvety outlook
_	Body papillae long, giving a hirsute outlook11
3 (2)	Body without sand particles4
_	Body with sand particles9
4 (3)	Ventrolateral gonopores present in some anterior chaetigers5
-	Ventrolateral gonopores absent7
5 (4)	First chaetiger with long chaetae, about half as long as body width; caruncle
	posteriorly expanded D. erythroporus Gallardo, 1968
-	Anterior end with short chaetae, about 1/3–1/5 as long as body width; carun-
	cle posteriorly tapering
6 (5)	Median chaetigers with neurochaetae tapering, 22-25 articles, and tip deli-
	cately falcate; cirriform branchiae with basal ¼–1/5 with lamella
	<i>D. branchiatus</i> (Rullier, 1965), comb. n.
-	Median chaetigers with neurochaetae barely tapering, 8-11 articles, and tip
	markedly falcate; cirriform branchiae with basal 1/3–1/2 with lamella
	<i>D. nicolaji</i> (Buzhinskaja, 1994), comb. n.
7 (4)	Papillae digitate, longer than wide, often swollen basally; median chaetigers
	with 5–6 notochaetae and 4–5 neurochaetae
_	Papillae hemispherical, about as long as wide
8 (7)	Median chaetigers with 5-6 neurochaetae, smaller than notochaetae, with
	articles 2.0–2.5 times longer than wide <i>D. kudenovi</i> sp. n.
_	Median chaetigers with 2–3 neurochaetae, about as long as notochaetae, with
	articles 7–8 times longer than wide
O(2)	D. stopbowitzi Darbyshire & Mackie, 2009
9 (3)	Anterior chaetigers swollen, much wider than following ones; sediment par-
	ticles scattered
_	Anterior chaetigers barely wider than following ones; sediment grains abun-
10(0)	dant, forming a thin crust
10 (9)	Lateral papiliae 1/3–1/10 as long as longest notocnaetae; median chaetigers no-
	tochaetae with basal articles poorly definedD. guarcus (Maingren, 180/)
_	Lateral papiliae up to 1/5 as long as longest hotochaetae; median chaetigers
	notochaetae with medium-sized articles basany
11(2)	Body without and particles
11 (2)	Body with out said particles, median chaetigers with 7, 8 notochaetee per hun
_	dle: neurochaetae with long articles distally
12(11)	Median chaetigers with notochaetae as long as hody diameter; popullae very
12 (11)	long single chaetiger 1 with 4-5 notochaetae (body often reddich)
	D hirestus (Hanson 1882)

-	Median chaetigers with notochaetae longer than body diameter13
13 (12)	Median neurochaetae with distal articles barely longer than wide14
_	Median neurochaetae with most articles 2-4 times longer than wide; no go-
	nopodial lobes
14 (13)	Gonopodial lobes dark (papillae core and tip blackish); body papillae thick,
	digitate (body often grayish)
	D. normani (McIntosh, 1908), comb. n., reinst.
_	Gonopodial lobes pale; body papillae thin, filiform (body often pale)
	D. longisetosus (von Marenzeller, 1890)
15 (11)	Sand particles restricted to the bases of papillae; neurochaetae with anchy-
	losed region about one-fifth of chaetal length
	D. octobranchus (Hartman, 1965)
_	Sand particles fixed along the papillae; neurochaetae with anchylosed region
	1/2–1/3 of chaetal length

Diplocirrus glaucus (Malmgren, 1867)

http://species-id.net/wiki/Diplocirrus_glaucus Fig 1

Trophonia glauca Malmgren 1867:192, Pl. 14, Fig. 78; McIntosh 1915:96–98, Pl. 96, Fig. 2, Pl. 104, Fig. 9 (syn.; simult. Haase 1915; his references stop in 1914).
Diplocirrus glaucus: Haase 1915:195–197, Textfigs. 3–5 (comb. n.); Fauvel 1927:120–121, Figs. 43a–d; Rioja-LoBianco 1931:98–100, Pl. 30; Støp-Bowitz 1948a:25–28, Fig. 6a–c; Hartmann-Schröder 1971:374–376, Fig. 132; Hartmann-Schröder 1996:416–417, Fig. 202; Jirkov & Philippova 2001:358–359, Figs. 1–3; Darbyshire & Mackie 2009:97, Table 1.

Stylarioides flabellata: Fauvel 1946:401 (non Sars, 1871).

Type material. Norway. Probably lost.

Additional material. Norway. One specimen (MNHN-A183), broken in two, without posterior end, anterior end exposed, appendices lost, Solsvick, no further data. Many specimens , Hardangerfjorden (60°10'00"N, 06°00'00"E) separated as follows: 14 anterior fragments (LACM-AHF 2620), Stat. Z20, 7 Jun. 1957, 25–16 m (up to 36 chaetigers, all with multiarticulated neurohooks; in posterior chaetigers with over 10 long articles). Two posteriorly incomplete specimens (LACM-AHF 2622), Stat. Z21, 88–78 m, 7 Jun. 1957. A mature female and a posterior fragment (LACM-AHF 2624), Stat. Z35, 98–104 m, 22 Sep. 1958 (oocytes about 125 μm). Anterior fragment, Stat. Z67, 102 m, 18 Oct. 1958. Seven specimens (LACM-AHF 2683), apparently fixed in alcohol, Stat. Z71, 102–78 m, 20 Oct. 1958 (used for details of branchiae; up to 27 chaetigers, all with multiarticulated neurohooks with articles medium-sized). Two specimens (LACM-AHF 2627), Stat. 121, 66–87 m, 15 Nov. 1958 (used for description). Faroe Islands. One specimen (MNHN-A183), anterior fragment, diges-



Figure 1. *Diplocirrus glaucus* (Malmgren, 1867). Non-type specimens (LACM-AHF 2683), Norway **A** anterior end, dorsal view, head exposed **B** same, close-up of branchiae showing longitudinal striae **C** another specimen (221), head exposed, anterior end, lateral view **D** same, cirriform branchiae with basal ridges **E** same, head, frontal view, branchiae and pals removed (BS: branchial scars, DL: dorsal lip, LL: lateral lip, NL: nephridial lobes, PS: palp scar), another specimen (LACM-AHF 2627) **F** chaetiger 24, basal, medial and distal notochaetal regions **G** same, basal, medial and distal neurochaetal regions.

tive system mostly expulsed from the body, most chaetae broken, RV Pourquoi-Pas? Expedition, off Klaksvik (62°13'26"N, 06°34'43"W), 8–15 m, 30 Jul. 1929. Sweden. Many specimens, Tjarno (58.52°N, 11.10°E) and surroundings, Apr. 2002, L. H. Harris, coll., including: One specimen (LACM-AHF 2684) complete, light dark (24 mm long, 2 mm wide, cephalic cage 1,8 mm long, 44 chaetigers; gonad lobes in chaetigers 5 and 6). One anterior fragment (LACM-AHF 2685) with anterior end exposed (used to describe the palp bases and lips). Russia. One specimen (ECOSUR), White Sea, 60 m, 28 Jun. 1998, A. Filippova, coll. (7 mm long, 0.8 mm wide, cephalic cage 2 mm long, 21 chaetigers; papillae short, capitate). Denmark. Four specimens (USNM-332), damaged, donated by C. Lütken, id. by M. Pettibone (Most chaetae broken; slide with median and posterior chaetiger, median one is only the chaetae. Zero to one chaetae in cephalic cage. Notochaetae very thin, neurochaetae thicker, tips falcate). Germany. Two specimens (USNM-175143), North Sea, German Bight, Senckenberg Stat. 24ku, 49.2 m, 12 Aug. 1990, M. Boggemann id.

Description. Largest specimens (LACM-AHF 2627) pale (some specimens Stat. Z71 with rusty pigmentation in chaetigers 1–3), posteriorly incomplete. Body soft, whitish (Fig. 1A, C), cylindrical, anteriorly swollen, posteriorly tapered; 17–20 mm long, 2–3 mm wide, cephalic cage 2 mm long, 23–27 chaetigers. Tunic with a thin

layer of fine sediment grains, papillated. Papillae short, capitate or club-shaped, arranged in 10–12 irregular rows per segment, longer in chaetal lobes, even longer in posterior chaetigers.

Cephalic hood exposed in one specimen (Fig. 1C, LACM-AHF, Stat. 221), almost transparent, smooth. Prostomium low cone (LACM-AHF LH2-514); eyes not seen. Caruncle poorly developed, not reaching the posterior margin of branchial plate, lateral ridges low, median keel not projected (Fig 1E). Palps long, thick; palp keels rounded, reduced. Lateral lips larger, thick, dorsal lip smaller, rhomboid, ventral lip reduced, rounded. Branchiae (LACM-AHF, Stat. Z71) of two different types; posterior branchiae thicker, prismatic, laterally fused to adjacent filaments (Fig. 1B), arranged in a continuous line; anterior branchiae cirriform, slightly longer than posterior branchiae, arranged as two lateral pairs, some with a basal thickening or reinforcement, occupying about 1/6–1/7 of branchial length (Fig. 1D). Palps longer than anterior branchiae. Nephridial lobes, two pairs, placed between posterior and anterior branchiae, each short, rounded (taking methyl-green stain).

Cephalic cage chaetae as long as, or slightly longer than body width. Only notochaetae of chaetiger 1 involved in the cephalic cage, chaetae directed dorsally. Chaetae arranged in a short transverse line; 2–3 notochaetae per ramus. Anterior dorsal margin of first chaetiger papillated, papillae similar to those along the body. Chaetiger 1 short, chaetigers 2–3 longer. Post-cephalic cage chaetigers not elongated, progressively widening to chaetigers 7–8, and then tapering posteriorly. Neurohooks start in chaetiger 1. Gonopodial lobes not seen (other specimens with low, blackish, rounded spots in chaetigers 5–6).

Parapodia reduced, chaetae emerge from the body wall. Parapodia lateral; median neuropodia ventrolateral. Notopodia (Fig. 1F) and neuropodia with slightly elongated papillae in chaetal lobes. Median notochaetae arranged in a short transverse line, as long as about 1/3 body width, 7–8 per bundle; all notochaetae multiarticulated capillaries, articles medium-sized basally, slightly long medially and distally. All neurochaetae multiarticulated hooks with short articles basally, becoming longer medially, tip falcate, smooth (Fig. 1G); median neurochaetae arranged in a transverse line, 4–5 per bundle.

Posterior end tapering, blunt (LACM-AHF-LH-2-522); pygidium with anus dorsoterminal, without anal cirri. A mature female with oocytes, each about 125 µm.

Remarks. *Diplocirrus glaucus* (Malmgren, 1867) is closely allied to *D. incognitus* Darbyshire & Mackie, 2009 because both have swollen anterior chaetigers and some sediment particles scattered over the body. They differ in the relative size of lateral papillae and notochaetal articulation; thus, *D. glaucus* has smaller papillae (up to one-fifth notochaetal length), and poorly defined basal articles in notochaetae, whereas *D. incognitus* has longer papillae (up to one-third notochaetal length) and medium-sized basal articles in notochaetae.

The original description (Malmgren 1867) indicated that the color was variable from bluish-gray to greenish or pale, but the number of chaetae in chaetiger 1 was stated as about 3, which has been used to separate it from similar species. The species was originally described from Bahusiae (Malmgren 1867:192), corresponding with the current Bohuslan (58.88° N, 10.51° E), where the Tjarno Marine Biological Station is, and where some of the specimens used for this description were collected.

Distribution. Northeastern Atlantic Ocean, Russian Northwestern Antarctic seas, in shallow water.

Diplocirrus branchiatus (Rullier, 1965), comb. n.

http://species-id.net/wiki/Diplocirrus_branchiatus Fig 2

Bradiella branchiata Rullier 1965:188–190, Fig. 7; Darbyshire & Mackie 2009:97, Table 1.

Diplocirrus cf. capensis Spies 1975:187, 189, 190, Pl. 3, Fig. 7, Pl. 6, Fig. 18.

Type material. Australia. Holotype of *Bradiella branchiata* Rullier, 1965 (AM-W3793), Moreton Bay (27°15'00"S, 153°15'00"E), Brisbane, Queensland, 1.2 km SW of M3 red beacon, coll. Party, 10 Nov. 1961. Two permanent slides (IRFA-W40, -W40'); W40 has three chaetal lobes and a small piece of skin; W40' has a branchial blade.

Additional material. Australia. One anterior fragment (NTM-18913), anterior end exposed, appendages lost, Stat. A16a (12°11.7'S, 136°41.3'E), Melville Bay, 2.7 m, 7 Jul. 1991, Marine Ecology Unit, coll. (14 mm long, 2 mm wide, chaetiger 1 chaetae 0.5 mm long, 17 chaetigers, gonopores in chaetigers 3–12). One complete specimen (QM-G10334), Southwest Rocks, 0.8 km south of Peel Island (27.3° S, 153.21° E), Moreton Bay, Queensland, 6.4 m, shell, grit and sand, Sep. 1970, W. Stephenson, coll. (id. R. B. Spies; dorsally dissected, some parapodia removed, damaged, 13.5 mm long, 3 mm wide, chaetiger 1 chaetae 1 mm long, 21 chaetigers, gonopores pale, in chaetigers 3-8). Anterior fragment (QM-G10379), 1.6 km SE off Southwest Rocks, Peel Island (27.3° S, 153.21° E), Moreton Bay, Queensland, 4–7 m, mud, Mar. 1970, S. Cook, coll. (id. R. B. Spies; dorsally dissected, some parapodia removed, damaged, 38 mm long, 4 mm wide, chaetiger 1 chaetae 1.2 mm long, 18 chaetigers, gonopores reddish, in chaetigers 3–16). Anterior fragment (SAM-GR-201), under Edithburgh Jetty (35°05.172' S, 137°44.825' E), Victoria, South Australia, 5 m, in sediment, 1 Mar. 2004, G. Rouse, coll. (it is 12 mm long, 2.5 mm wide, chaetiger 1 chaetae 0.5 mm long, 15 chaetigers, gonopores pale, in chaetigers 3–6).

Description. Holotype brown yellowish (other specimens pale, dirty orange or rusty). Body cylindrical, tapering posteriorly (Fig. 2A), contorted, with a previous dorsal longitudinal dissection, and other smaller ones to remove chaetigers 5 and 18; 53 mm long, 6 mm wide, cephalic cage 1.3 mm long, 37 chaetigers. Tunic with abundant papillae, long, cirriform, slightly capitate, with a thin layer of fine sediment particles, forming a thick base, arranged in over 20 irregular bands per segment.

Cephalic hood exposed, with smaller sparse papillae, as long as the following 3–4 chaetigers (swollen in holotype, annulated in QM-10334); cephalic hood mar-



Figure 2. *Diplocirrus branchiatus* (Rullier, 1965), comb. n. Holotype (AM-W3793) **A** entire, ventral view **B** same, head, lateral view (DB: dorsal branchiae) **C** same, frontal view (LL: lateral lip, PS: palp scar) **D** same, right anterior chaetigers 3–6, ventral view **E** same, chaetiger 5 showing broken neurochaetae (Ne) and ventral pore (VP) **F** same, median chaetiger, notopodium (insert: notochaetal distal region) **G** same, median chaetiger, neurochaeta (inserts: anterior neurochaetal tips) **H** non-type specimen (SAM-GR-201), anterior end, head exposed, lateral view **I** same, proximal, cirriform branchiae showing basal ridges **J** same, distal, complex branchiae (BS: branchial sockets, DL: dorsal lamella).

gin smooth. Anterior end not everted, observed through the already done dissection. Prostomium elevated, eyes and caruncle not seen because it is bent and covered by the lateral lips (Fig. 2B, C, in SAM-GR201 prostomium flat lobe, no eyes). Palps lost (in SAM-GR201 palps thick, as long as branchiae); palp lobes reduced (thick, rounded in SAM-GR201, and two lateral well-developed lobes. Caruncle projected dorsally to the base of branchiae, lateral ridges elevated, posteriorly separated, laterally expanded. Dorsal lip projected anteriorly; lateral lips thicker; ventral lip reduced. Nephridial lobes in branchial plate not seen).

Holotype with branchial plate damaged. Posterior branchiae compressed, lateral filaments lost, median filament bent towards the mouth, lamellate; cirriform branchiae

lost, two lateral scars per side, placed below a dorsal crest. Slide IRFA-W40' shows a branchial blade made of fused branchial filaments. Another specimen (SAM-GR201), with head slightly exposed (Fig. 2H), branchiae complete of two different types. Posterior row with four prismatic, thicker, lamellate branchiae, tips bare (Fig. 2J); lateral branchiae smaller (one in regeneration), each with dorsal keel reduced, with longitudinal bands, dorsal surface laterally expanded with a thin axis, provided with two rounded lateral lobes; median branchiae larger, dorsal keel large, foliose, markedly corrugated. Distal branchiae with ventral side with a blade made of fused branchial filaments, convoluted, looking like a series of successive blades, but actually made by a single convoluted blade. Anterior branchial row with four thin, cirriform filaments, shorter than palps, arranged in two lateral pairs, each filament with a convoluted laterally. Branchial basal lobes between median and lateral branchiae (dorsal), and outside the lateral ones (lateral); dorsal lobes small, rounded, lateral lobes rounded, larger).

Cephalic cage chaetae slightly longer than following ones. First chaetiger displaced dorsally, with multiarticulated capillaries. Notochaetae in a short transverse tuft, with 6–7 multiarticulated capillaries. Anterior dorsal margin of first chaetiger papillated, as following segments. Anterior chaetigers without longer papillae, chaetiger 1 shorter than following ones, chaetal lobes lateral, very close to each other. First 10 chaetigers without marked segmentation between them; following chaetigers shorter, better defined. Ventral gonopores in chaetigers 3–12, orange-red, low papillae (Fig. 2D, E).

Parapodia poorly developed; chaetae emerge from the body wall. Notopodia and neuropodia with papillae as long as other papillae. Noto- and neuropodia close to each other. Median neuropodia lateral, very close to notopodia.

Chaetal transition from first chaetiger to body chaetae abrupt; notochaetae of chaetigers 2–3 large multiarticulated hooks, distal article hooked, entire. All other notopodia with multiarticulated capillaries. Median notochaetae arranged in a longitudinal line. Notochaetae of chaetigers 1 and beyond the third, multiarticulated capillaries; by chaetiger 11, as long as half body width, 10–11 per bundle (6–7 in smaller specimen), each with long articles throughout the chaeta (Fig. 2F). Neurochaetae multiarticulated hooks from chaetiger 1, arranged in a short J-shaped pattern, 4–5 per bundle, each with long articles of about the same length, tips falcate (Fig. 2G), with a hood-like membrane.

Posterior end invaginated in holotype; other specimens with truncated rounded lobe; notochaetae directed posteriorly; without anal cirri.

Variation. Pigmentation varies from pale orange to dark yellowish, or to dirty pink with gonopodial pores reddish or pale. Further, there are two main variations related to body size: papillae are longer in larger specimens, and gonopores become more pigmented, and are probably present along more segments as body enlarges.

Remarks. *Diplocirrus branchiatus* (Rullier, 1965) comb. n. is very similar to *D. nicolaji* (Buzhinskaja, 1994), comb. n. because both species have bodies without sediment particles, ventrolateral gonopores along several anterior chaetigers, short chaetae in the first chaetiger, and their caruncle tapers posteriorly. They differ in the relative

development of neurochaetae and of the extent of the lamellate area in their cirriform branchiae; thus, in *D. branchiatus* median chaetigers have neurochaetae with about 23 articles, tapering to a delicately falcate tip, and the lamellate region might be up to one-fifth of the branchial length, whereas in *D. nicolaji*, neurochaetae are barely tapering, having about 10 articles, their tips are markedly falcate, and the lamellate region might extend up to one-third of branchial length.

Diplocirrus branchiatus (Rullier, 1965) has been known only through the original description. Spies (1975) studied some specimens from the type locality (herein re-examined); they fit the original description but the anterior end was previously removed. Rullier's description is fairly complete, though the presence of multiarticulated hooks in notopodia 2–3 was overlooked, as well as the presence of the gonopores. The anterior end has a symmetrical pattern and the original description does not provide complete details about branchiae; however, the drawings show that there were two larger lamellate branchiae (his figure 7C), and that there were smaller lateral branchiae (his figure 7D), but there are no details on cirriform branchiae; they might have been lost during dissection. As originally shown by Rullier (1965), and confirmed by the observation of one permanent slide, branchial blades include a series of parallel filaments; however, they are not arranged as successive, independent blades but rather as a continuous, convoluted, branchial blade. So far, this special type of branchial pattern is only known for a few species in Diplocirrus. Further, Rullier illustrated that neurohooks are distally tapering (his figure 7G), but he described them as (p. 190) "plus courtes et recourbées à leur extrémité" (shorter and distally curved), which is the correct description. Spies (1975) made some observations and his drawings are slightly inaccurate in several features: the caruncle does not taper posteriorly, and does not reach the posterior margin of the branchial plate, interbranchial lobes were not illustrated, and the lateral palp lobes were not seen.

Distribution. Originally described from Eastern Australia, *D. branchiatus* is present from Northeastern Australia to Southern Australia, in shallow water sediments. The data in the same publication by Rullier (1965), indicate that the type specimen was found in muddy bottoms in shallow depths.

Diplocirrus capensis Day, 1961

http://species-id.net/wiki/Diplocirrus_capensis

Diplocirrus capensis Day 1961:509, Fig. 9a–f; Day 1967:666, Figs. 32.4e–j; Day 1973:105–107; Milligan 1984:47.9–11, Figs. 47.5–6; Darbyshire & Mackie 2009:96–98, Table 1 (redescr.).

Type material. The specimens are housed in the South African Museum, Cape Town, but were not made available. Reexamined by Darbyshire & Mackie (2009).

Additional material. Madagascar. One specimen (SMF-15355), anterior fragment, damaged, Stat. 11 bis, 47 m, 3 Apr. 1970, R. Plante, coll. (6 mm long, 1 mm wide, cephalic cage chaetae 0.3 mm, 15 chaetigers; gonopores in chaetigers 5–12). Two fragments (SMF-15374), Nosy Iranja, Stat. 4, Benne, 17 Sep. 1966, R. Plante, coll. **Northwestern Atlantic Ocean.** 18 specimens (USNM-51039), damaged, 12 anterior previously dissected or with some parapodia previously removed, and 6 median fragments, off North Carolina, BST 51X (34°20'N, 75°55'W), 165 m, sandy mud, J.H. Day, coll. (larger anterior fragments 6.0–10.5 mm long, 1 mm wide, cephalic cage 0.8–1.0 mm long, 16–28 chaetigers; gonopores not seen).

Description. (modified from Day 1961, 1967 and combined with data from Darbyshire and Mackie 2009. Data from North Carolina specimens in parenthesis, if they differ): Body muddy brown (golden), anteriorly swollen with segmental lines indistinct, tapering posteriorly with better defined segments. Holotype an anterior fragment, 12 mm long, 2 mm wide, no cephalic cage, 18 chaetigers. Tunic papillated; each papillae short, 8-shaped to long, clavate, basally swollen (lateral papillae longer, cirriform).

Cephalic hood not exposed. Prostomium with four small, black eyes. Palps thick, as long as branchiae. Caruncle projected dorsally, not reaching the posterior margin of branchial plate. Lips corrugated, fused. Nephridial lobes in branchial plate not seen. Branchiae very dark, of two types. Posterior row with four wedge-shaped filaments; anterior row branchiae cirriform, separated in two lateral pairs by the caruncle. Interbranchial lobes not seen. Lamellate region difficult to evaluate.

Chaetigers 1–2 with 2–3 fine notochaetae and 4–6 shorter multiarticulated neurochaetae. Anterior dorsal margin of first chaetiger papillated, as following segments; no other modification. Anterior chaetigers without longer papillae, chaetiger 1 shorter than following ones, chaetal lobes lateral, very close to each other. First 10 chaetigers without marked segmentation; posterior segments better defined. Gonopodial lobes not seen.

Parapodia poorly developed; chaetae emerge from the body wall. Notopodia and neuropodia with papillae longer than other body ones. Noto- and neuropodia close to each other. Median neuropodia lateral, very close to notopodia.

Median notochaetae arranged in a longitudinal line, as long as body width, 10–12 (4–6) per bundle, each with short rings basally, long medially and distally. Neurochaetae multiarticulated hooks from chaetiger 1, arranged in a short J-pattern, 6–8 (3–4) per bundle, each with articles of about the same length, tip falcate.

Posterior end unknown.

Remarks. *Diplocirrus capensis* Day, 1961 is closely related to *D. kudenovi* sp. n. and *D. stopbowitzi* Darbyshire & Mackie, 2009 because their bodies do not incorporate sand particles, and by lacking ventrolateral gonopores. However, these two latter species are provided with hemispherical papillae whereas in *D. capensis* papillae are elongate, often basally swollen, but never hemispherical.

The records of *D. capensis* by Day (1973:105–107), and Milligan (1984:47.9–11, Figs. 47.5–6) differ from the typical South African form because they have different body color, cephalic cage, larger lateral papillae, and by the relative numbers of chaetae. They might represent a different species but their description as new species must wait for better specimens. There is a similar, apparently undescribed species in the Mediterranean Sea, which has been recorded as *D. glaucus* by Fauvel (1937:34, *non*

Malmgren, 1867). The materials are damaged (MNHN-406), many chaetae broken, anterior regions smashed or without exposed head, and were collected off Alexandria, Egypt. Better specimens would help clarify its affinities with *D. capensis*.

Distribution. The distribution for the nominal form is apparently restricted to the Cape province, South Africa, in 11 m; it is questionably recorded from North Carolina, 165 m depth.

Diplocirrus erythroporus Gallardo, 1968

http://species-id.net/wiki/Diplocirrus_erythroporus Fig 3

- Diplocirrus erythroporus Gallardo 1968:108, Pl. 49, Figs. 7–10; Darbyshire & Mackie 2009:97, Table 1.
- *Diplocirrus glaucus*: Fauvel 1932:186–187; Fauvel 1953:353, Fig. 184a–d (*non* Haase, 1915).

Diplocirrus glaucus orientalis Gibbs 1971:181, no figs.

Type material. Viet Nam. Holotype (LACM-AHF 306), off Hon Mot Island (12°10'34"N, 109°16'11"E), R.V. Mao Tien, Naga Expedition Stat. 113, 22 m, 10 Feb. 1960.

Additional material. Viet Nam. Two specimens (LACM-AHF 2606), Western side of Hon Lon Island (12°12'49"N, 109°14'22"E), R.V. Mao Tien, Naga Expedition Stat. 323, 14 m, 4 Apr. 1960. Australia. Two specimens (NTM-18920), one complete, the other without anterior end, Stat. DW69A (12°32.28'S, 130°46.66'E), Darwin Harbor, Australia, 3 m, 17 Mar. 1994, Marine Ecology Unit, coll. (complete: 34 mm long, 3 mm wide, cephalic cage 1.8 mm long, 64 chaetigers, gonopores in chaetigers 4–14). Yellow Sea. Anterior fragment (ZISP-10854), plus few chaetigers, Yellow Sea, R.V. Venus, no station data, Chzhan coll.; B. Wu id. as *Brada longicirrata* sp. n. It was 11 mm long, 2 mm wide, cephalic cage 1.5 mm long, 23 chaetigers; four large erect papillae on chaetigers 1–2, one per ramus (resembling a cirrus on each chaetal bundle and hence the name); dorsal ones rise behind the first chaetiger notochaetae whereas the ventral ones stem halfway between the neurochaetae of chaetigers 1 and 2; nephridial pores without pigmentation, in chaetigers 4–12.

Description. Holotype an anterior fragment, soft, pale, with dispersed dark brown spots (Fig. 3A). Body cylindrical, anteriorly swollen, posteriorly tapered; 19 mm long, 2.8 mm wide (by chaetiger 7), cephalic cage 0.9 mm long, 33 chaetigers. Tunic papillated, with fine sediment particles.

Cephalic hood exposed, paler than following segments, almost transparent, with smaller papillae; anterior margin papillated, papillae sparse (anterior end dissected in another specimen, LACM-AHF 2606). Prostomium low cone (Fig. 3C); eyes not seen. Caruncle not seen. Palps pale; palp keels reduced. Branchiae of two types, distal row with filaments thick, cirriform; proximal branchiae in two lateral groups, filaments cirriform, thinner, with a thin distal part. Branchiae shorter than palps. Nephridial lobes rounded, low, brownish.



Figure 3. *Diplocirrus erythrophorus* Gallardo, 1968. Holotype (LACM-AHF 11144) **A** dorsal view **B** same, anterior end, ventral view, showing the ventrolateral pores **C** non-type specimen (LACM-AHF 11147), head, frontal view, palps and branchiae removed **D** same, left parapodium, chaetiger 16 **E** same, close-up of notochaetae **F** same, neurochaetal tips **G** same, chaetiger 21, right parapodium.

Cephalic cage chaetae as long as 1/3 body width. Only chaetiger 1 involved in the cephalic cage, slightly displaced dorsally. Chaetae arranged in a short lateral line; 3–4 chaetae per ramus. Anterior dorsal margin of first chaetiger papillated, papillae similar to those along the body but with one pair of stiff, long notopodial papillae; posterior chaetigers without long papillae but slightly longer papillae restricted to chaetal lobes.

Chaetigers 1–3 of about the same length (NTM-18920 with chaetiger 2 very thin, chaetiger 3 much longer, almost without papillae). Post-cephalic cage chaetigers not elongated, but progressively widening reaching the widest dimension by chaetiger 7, and then posteriorly reduced. Chaetal transition from cephalic cage to body chaetae gradual; neurohooks start by chaetiger 10. No gonopodial lobes; orange-reddish, disk-shaped gonopores in chaetigers 4–12 (Fig. 3B); in larger specimens along chaetigers 4–14.

Parapodia reduced, chaetae emerge from the body wall (Fig. 3D, G). Parapodia lateral; median neuropodia ventrolateral. Notopodia and neuropodia with slightly longer papillae in chaetal lobes. Median notochaetae arranged in a tuft, oblique to body axis. Median notochaetae as long as ¼ body width, about 9 per bundle; all notochaetae multiarticulated capillaries, articles very short basally, longer medially, becoming medium-sized distally (Fig. 3E). Neurochaetae multiarticulated capillaries resembling notochaetae in chaetigers 1–9; from chaetiger 10, neurochaetae thicker, multiarticulated hooks with short articles basally, becoming long medially, distal article longest, falcate, smooth (Fig. 3F). Median neurochaetae arranged in a transverse line, 4–5 per bundle.

Posterior end missing in holotype; non-type specimen (NTM-18920) with posterior end tapering to a blunt cone; pygidium with anus terminal, no anal cirri.

Remarks. *Diplocirrus erythroporus* Gallardo, 1968 resembles *D. branchiatus* (Rullier, 1965), comb. n. and *D. nicolaji* (Buzhinskaja, 1994), comb. n. because they all have ventrolateral gonopores along some anterior chaetigers. However, these two latter species have very short chaetae in their first chaetiger, whereas *D. erythroporus* has long chaetae. Additionaly, the caruncle of *D. erythroporus* is posteriorly expanded unlike that of *D. branchiatus* and *D. nicolaji*.

The original description (Gallardo 1968) was brief. It indicated that there were six tentacles (branchiae), four larger and two smaller ones, and there were no details on the extent of the cephalic cage. Thus, a redescription was required in order to separate this species from other similar ones in the Indo-Pacific regions. The two additional specimens were one maculated with rounded dark brown spots (11 mm long, 2.5 mm wide, cephalic cage 0.9 mm long, 22 chaetigers, gonopores in chaetigers 4–13), which was dissected to study the anterior end, and another without dark spots (14 mm long, 2.8 mm wide, cephalic cage 1.0 mm long, 23 chaetigers, nephridial pores in chaetigers 4–12; it is a mature female). *Diplocirrus glaucus orientalis* Gibbs, 1971 was described without illustrations; it has orange globular papillae below each neuropodium in chaetigers 4–14(16). This could include the record of *D. glaucus* by Fauvel (1932:186–187, Fauvel 1953:353, Fig. 184a–d). It is being regarded as a junior synonym of *D. erytrhoporus*.

Distribution. Vietnam, Solomon Islands, Northeastern Australia, in shallow depths (up to 24 m depth).

Diplocirrus hirsutus (Hansen, 1878)

http://species-id.net/wiki/Diplocirrus_hirsutus Fig 4

Trophonia hirsuta Hansen 1878:9–10, Pl. 7, Figs. 1–4; Hansen 1882:38, Pl. 7, Figs. 5–8.

Stylarioides hirsutus: von Marenzeller 1889:129–130 (comb. n.); Ditlevsen 1911:426, Pl. 29, Fig. 11, Pl. 31, Figs. 23, 24.

Diplocirrus hirsutus: Haase 1915:198–200 (comb. n.); Støp-Bowitz 1948a:28–30, Fig. 7, Støp-Bowitz 1948b:37–38, map; Wesenberg-Lund 1950:35; Fauchald 1972:412; Jirkov & Philippova 2001:359, Figs. 1–7; Darbyshire & Mackie 2009:97, Table 1.

Type material. Norway. Syntypes (ZMUB-2287), four anterior fragments and a dissected anterior end, two previously dissected, NMH Expedition, Stat. 18 (62°44'N, 01°48'E), and Stat. 31 (63°10'N, 05°00'E) (syntypes yellowish, incomplete, 4–5 no-



Figure 4. *Diplocirrus hirsutus* (Hansen, 1878). Syntypes (ZMUB-2287) **A** anterior fragments, dorsal view **B** larger syntype, anterior end, dorsal view **C** complete, non-type specimen (ZMUB-27459) **D** same, anterior end, oblique ventral view **E** same, head, frontal view, branchiae and palps removed **F** another syntype, chaetiger 8, right notopodial chaetae **G** same, neurochaetae (inserts: neurochaetal tips).

tochaetae in chaetiger 1; 8–9 notochaetae in median chaetigers; 10 transversal rows of papillae in chaetiger 10; gonopodial lobes not visible).

Additional material. Norway. Two specimens (ZMUB-25216), Norkse Nordhavs. Expedition, Stat. 262 (no data) (two anterior fragments, dried out). Five specimens (ZMUB-27459), NMH (N. Nordhosk Expedition, Stat. 326 (no data), Hansen, coll. (complete 19–22 mm long, 2.5–2.7 mm wide, cephalic cage 2.0–2.5 mm long, 29–37 chaetigers; gonopodial lobes in chaetigers 5–6 in two specimens).

Description. Larger syntype pale, soft, yellowish (Fig. 4A, B). Body club-shaped, swollen anteriorly, progressively narrowing to chaetiger 12, then cylindrical to the end of the fragment (and body; Fig. 4C)); 10 mm long, 3.5 mm wide, cephalic cage chaetae 2 mm long, 20 chaetigers. Tunic papillated, fine sediment particles on papillae basis only (other specimens with sediment cover towards the tip). Papillae long, abundant, capitate, with basal sediment making a rounded lobe (Fig 4B), about 10 transverse rows in chaetiger 10, much longer dorsally, longest about 2/3 as long as notochaetae.

Anterior end observed in a previously dissected specimen and in non-type specimen. Cephalic hood short, smooth, margin smooth. Prostomium low cone, grayish, eyes barely pigmented (Fig. 4E), difficult to be seen in syntype or non-types. Caruncle not observed in syntype, weakly defined in non-types. Palps thick, longer than the only available cirriform branchia; palp bases rounded, projected. Lateral lips projected, thick, well-developed, dorsal and ventral lips reduced. Branchiae mostly lost, scars remain; posterior row with thicker scars, anterior row with a single cirriform branchiae without basal blades (all cirriform, posterior ones slightly thicker, smooth). Nephridial lobes rounded, elevated, separating anterior and posterior branchial rows (taking methyl green stain deeply).

Cephalic cage chaetae shorter than body width. Chaetiger 1 involved in the cephalic cage, chaetae arranged in short dorsolateral lines, with 4–5 noto- and 9–10 neurochaetae per bundle. Anterior dorsal margin of first chaetiger papillated; anterior chaetigers with papillae longer than those present in following chaetigers. Chaetigers 1–3 progressively longer. No chaetal transition from cephalic cage chaetae to body chaetae; all neurochaetae multiarticulate falcigers but first chaetigers with shorter articles. Gonopodial lobes not seen in syntypes (oval, bare, pale areas in chaetigers 5–6 in non-types; Fig. 4D).

Parapodia lateral, poorly developed, chaetae emerge from the body wall; median neuropodia ventrolateral. Notopodia without conical lobes. Noto- and neuropodia distant to each other.

Median notochaetae arranged in a longitudinal, transverse, short line; all notochaetae multiarticulated capillaries (Fig. 4F), medium-sized articles basally, longer medial- and distally; 8–9 notochaetae per bundle in median chaetigers (up to 14 in non-types), about as long as body width. All neurochaetae multiarticulated hooks, markedly tapering subdistally (Fig. 4F); basal articles short, ill-defined, longer medialand distally, but diminishing in size towards the tip, 5–6 per bundle.

Posterior end, observed in non-type specimens, truncate (Fig. 4C); pygidium with anus terminal, without anal cirri.

Remarks. Diplocirrus hirsutus (Hansen, 1878) resembles D. longisetosus (von Marenzeller, 1890) and D. normani (McIntosh, 1908), comb. n. because they have bodies provided with long papillae but without sand particles. Their main difference lies in the relative length of notochaetae in median chaetigers, because the latter two species have notochaetae markedly longer than body width, whereas in D. hirsutus they are about as long as body width.

Haase (1915:199) noticed the cinnamon-red color for specimens of this species. The available specimens show a concentration of the pigment towards the anterior end, making a thin crust surrounding papillae and chaetae. Thus, it is not the basic color of the organism but rather some adsorbed minerals on these structures and, whenever this pigmentation is present, chaetae are darker, which indicates that the minerals are either ingested and later used for chaetal formation, or adsorbed to chaetae as well as over the tunic. This pigmentation should rely on the minerals available in the sediments, and therefore should not be used as a diagnostic feature.

Distribution. Originally described from Norway, it ranges in Arctic and Subarctic environments in shallow water. The Antarctic records by Hartmann-Schröder and Rosenfeldt (1989:71–72; 1991:74–75) are questionable.

Diplocirrus incognitus Darbyshire & Mackie, 2009

http://species-id.net/wiki/Diplocirrus_incognitus Fig. 5

Diplocirrus incognitus Darbyshire & Mackie 2009:99-102, Figs. 3B, 4, Table 1.

Diagnosis. Body anteriorly swollen (Fig. 5A). Papillae abundant, short, giving a velvety oultlook, with scattered sediment particles (Fig. 5B, C). Lateral papillae 1/3 as long as longest notochaetae. Median notochaetae with long articles (Fig. 5D). Neurochaetae with long articles, tips barely curved (Fig. 5E).

Remarks. As stated above, *Diplocirrus incognitus* Darbyshire & Mackie, 2009 resembles *D. glaucus* (Malmgren, 1867), because both have bodies anteriorly swollen and few sediment particles spread over the body. They differ in the relative size of lateral papillae and on the notochaetal basis articulation; thus, in *D. incognitus* papillae are longer (up to one-third notochaetal length), and notochaetal bases have mediumsized articles, whereas in *D. glaucus* papillae are smaller (up to one-fifth notochaetal length) and notochaetal bases have poorly-defined articles.

Distribution. South Africa, offshore, in muddy bottoms of about 100 m depth.

Diplocirrus kudenovi sp. n.

urn:lsid:zoobank.org:act:1EED4521-10A1-4EA5-87AE-FDCCF9E389A7 http://species-id.net/wiki/Diplocirrus_kudenovi Fig. 6

Type material. Eastern Pacific Ocean. Holotype (LACM-AHF 2594) and 14 paratypes (LACM-AHF 2595), Southern Bay, Isla Cedros, Baja California, Mexico, RV Velero IV, Stat. 2026 (20°05'00"N, 115°19'45"W), 16 fathoms, mud and sand, 19 Apr. 1951 (best paratypes: 8.0–22.5 mm long, 1–2 mm wide, cephalic cage 1.0–1.5 mm long, 24–49 chaetigers).

Additional material. Gulf of California. One anterior fragment (LACM-AHF 2596), damaged, off southeastern tip of Isla Angel de la Guarda, Baja California, Mexico, Stat. P-71–59 (29°20.0'N, 113°11.2'W), 40 fathoms (7 mm long, 1.5 mm wide, cephalic cage 1.5 mm long, 19 chaetigers).

Description. Holotype (LACM-AHF2594), without posterior end, soft, whitish (Fig. 6A). Body club-shaped, anteriorly swollen, progressively narrowing to chaetiger 15, then cylindrical, tapering to the end of the fragment; 19 mm long, 2 mm wide, cephalic cage 1.5 mm long, 47 chaetigers. Tunic papillated, fine sediment particles on papillae basis only. Papillae short, abundant, capitate, with basal sediment making a rounded lobe, about 13–15 irregular rows in anterior chaetigers (about 10 rows in median chaetigers), slightly longer dorsally and in posterior chaetigers; in median chaetigers papillae as long as 1/5–1/6 notochaetal length.



Figure 5. *Diplocirrus incognitus* Darbyshire & Mackie, 2009. Holotype (BMNH 1961.19.694) **A** dorsal view **B** same, anterior end, dorsal view **C** posterior end, dorsal view **D** median chaetiger, notopodium (insert: notochaetal tip) **E** same, neurochaetae (insert: neurochaetal tip).

Anterior end completely exposed, slightly damaged (Fig. 6B). Cephalic hood short, smooth, margin smooth. Prostomium low, pale, eyes not seen. Caruncle poorly developed, lateral ridges low, median keel reduced, not continued to the posterior margin of the branchial plate (Fig. 6C). Palps lost in holotype (pale in one paratype), palp bases rounded. Lateral lips well developed, dorsal lip reduced, ventral lip rounded. Branchiae mostly lost, branchial scars on branchial plate, arranged in two rows, posterior row with 4 thicker branchial scars, anterior row discontinuous, two narrower branchial scars, one long cirriform branchia left. Nephridial lobes rounded, separating posterior and anterior branchiae.

Cephalic cage chaetae shorter than body width. Chaetiger 1 involved in the cephalic cage, slightly displaced dorsally; chaetae arranged in short dorsolateral lines, with 2 noto- and 4 (–6) neurochaetae. Anterior dorsal margin of first chaetiger papillated; anterior chaetigers without especially long papillae. Chaetigers 1–3 progressively larger. No chaetal transition from cephalic cage to body chaetae, all neurochaetae similar, but first chaetiger with shorter articles. Gonopodial lobes present in chaetiger 5 (or 5 and 6 in some paratypes), a transverse papillae-free area.



Figure 6. *Diplocirrus kudenovi* sp. n. Holotype (LACM-AHF 2594) **A** lateral view **B** same, anterior end, lateral view **C** same, head, frontal view **D** another specimen, posterior end, dorsal view **E** same, chaetiger 26, right parapodium **F** same, notochaetae **G** same, neurochaetae (insert: neurochaetal tip).

Parapodia lateral, poorly-developed, chaetae emerge from the body wall (Fig. 6E); median neuropodia ventrolateral. Notopodia without conical chaetal lobes. Noto- and neuropodia distant to each other.

Median notochaetae arranged in a longitudinal, short line; all notochaetae multiarticulated capillaries, short articles basally, long medially and distally (Fig. 6F). About 6–8 chaetae per bundle, 1/2–2/3 as long as body width. All neurochaetae multiarticulated hooks, feebly-defined short articles basally, medial- and distally with long articles, distally falcate (Fig. 6G); neurohooks arranged in a transverse line, with 5–6 per bundle.

Posterior end (observed in a paratype) tapering to a rounded lobe (Fig. 6D); pygidium with anus terminodorsal, without anal cirri.

Etymology. This species is named after Jerry D. Kudenov, who has studied several polychaete families on a world-wide basis, and especially for his series of publications on the polychaetes from the Gulf of California, which have been very useful for many researchers working in the region, including one of us (SISV). The epithet is a noun in the genitive case.

Type locality. Southern Bay, Isla Cedros, Baja California, México, in mud-sand bottoms, at 16 fathoms depth.

Remarks. *Diplocirrus kudenovi* sp. n. is very similar to *D. stopbowitzi* Darbyshire & Mackie, 2009, because both have bodies without sand particles, with papillae hemispherical, and by lacking ventrolateral gonopores. They differ in chaetal features, especially regarding neurochaetae; thus, in *D. kudenovi*, median chaetigers have 5–6 neurochaetae and each has articles about twice as long as wide, whereas in *D. stopbowitzi*, there are 2–3 neurochaetae and each has longer articles, being about seven times longer than wide.

Distribution. Western Mexico, in both sides of the Baja California Peninsula, in subtidal waters.

Diplocirrus longisetosus (von Marenzeller, 1890), restricted

http://species-id.net/wiki/Diplocirrus_longisetosus Fig. 7

Stylarioides longisetosus von Marenzeller 1890:5 Fig. 3, von Marenzeller 1892:426–427. Diplocirrus longisetosus: Haase 1915:200–202, Textfigs. 6–7 (partim); Ushakov 1955:307(1965:285), Fig. 114G, H; Darbyshire & Mackie 2009:97, Table 1.

Type material. Gulf of Alaska. Neotype (CAS-27933), and paraneotypes (CAS), off Pitt Point, Alaska, Stat. 1546 (71°19.5'N, 152°58.0'W), 55 m, sandy silt, 11 Aug. 1977, R.E. Ruff, coll. and id. (paraneotypes 10–14 mm long, 1 mm wide, cephalic cage 2.5–3.0 mm long, 25–31 chaetigers).

Additional material. Bering Sea. Two anterior fragments (ZIRAS-27133), Providence Bay, Stat. 74 (no specific data), 18 m, mud, P. Uschakov, coll. (10.0/10.5 mm long, 2.0/2.5 mm wide, cephalic cage chaetae 3.0/2.5 mm long, 18/16 chaetigers; gonopodial lobes in chaetiger 5).

Description. Neotype complete (CAS-27933), pale yellowish. Body club-shaped, anteriorly swollen, progressively narrowing to chaetiger 13, then cylindrical, tapering to the posterior end (Fig. 7A); 12 mm long, 1.5 mm wide, cephalic cage 2.5 mm long, 33 chaetigers. Tunic papillated, detached in several portions, with fine sediment particles. Papillae pale, cirriform, sparse, about 5–6 transverse rows in median chaetigers, slightly longer dorsally; in median chaetigers about 1/5 as long as notochaetae.

Anterior end modifications observed in a paraneotype. Cephalic tube short, smooth, margin apparently smooth. Prostomium low, pale, eyes black, small. Caruncle not seen. Palps pale, thick, deeply furrowed, as long as branchiae; palp keels reduced. Lips damaged by dissection. Branchiae thick, cirriform, sessile on branchial plate; posterior branchiae thicker, anterior branchiae cirriform, two thinner filaments per lateral group. Nephridial lobes very thin, long, placed below the posterior row central filaments.

Cephalic cage chaetae 1/5 as long as body length, or 2/3 as long as body width. Chaetigers 1–2 involved in the cephalic cage; chaetae arranged in short, dorsolateral lines, 5(–8) noto- and 5 neurochaetae per bundle. Anterior dorsal margin of first chaetiger papillated; anterior chaetigers without especially long papillae. Chaetigers 1–3 progressively larger. Post-cephalic cage chaetigers not elongated. No chaetal transition from cephalic cage to body chaetae, all neurochaetae similar. Gonopodial lobes present in chaetiger 5, low, round, pale lobes, covered by small papillae, difficult to be seen even after methyl green staining (Fig. 7B).



Figure 7. *Diplocirrus longisetosus* (von Marenzeller, 1890) restricted. Neotype (CAS-27933) **A** dorsal view **B** same, anterior end, ventral view **C** paraneotype, chaetiger 25, right parapodium **D** same, basal, medial and distal notochaetal regions **E** same, basal, medial and distal neurochaetal regions.

Parapodia lateral, poorly-developed, chaetae emerge from the body wall (Fig. 7C); median neuropodia ventrolateral. Notopodia 1–2 with low, conical, chaetal lobes directed forward, remaining parapodia without conical lobes. Neuropodia 1–4 with low, conical chaetal lobes. Noto- and neuropodia distant to each other.

Median notochaetae arranged in a transverse horizontal C-shaped pattern; all notochaetae multiarticulated capillaries, short articles basally, medium-sized medially, long distally (Fig. 7D). About 11(-13) chaetae per bundle, twice as long as body width. All neurochaetae multiarticulated capillaries, very short articles basally, well-defined, medium-sized medially, long distally (Fig. 7E); tips straight; arranged in a transverse line, 8–9 per bundle.

Posterior end tapering to a rounded lobe; pygidium with anus terminal, without anal cirri.

Neotype locality. Off Pitt Point, Alaska, 55 m, sandy silt.

Remarks. As currently restricted, *Diplocirrus longisetosus* (von Marenzeller, 1890), closely resembles *D. micans* Fauchald, 1972 and *D. normani* (McIntosh, 1908), comb. n. These species have notochaetae longer than the body width, and long papillae with-

out sand particles, although *D. micans* separates from the other two species by having neurochaetae with long articles, and because it lacks gonopodial lobes. Then, *D. longisetosus* and *D. normani* differ especially in the relative body color, papillae and gonopodial lobes, and on the relative resolution of neurochaetal basal articles. In *D. longisetosus*, papillae and gonopodial lobes are pale, and basal neurochaetal articles are well-defined, whereas in *D. normani*, the body is grayish, and papillae and gonopodial lobes are darker or blackish, whereas neurochaetal basal articles are poorly-defined.

Further, *D. longisetosus* was described from Providence Bay, Russia, in the Bering Sea, with a single anterior fragment. Haase (1915:200) studied the supposed holotype (which is now lost), an additional specimen sent him by von Marenzeller, probably coming from Spitzbergen, Norway, and an additional broken specimen. This combination resulted in a mixture of morphological features and the species has been recorded from several localities in the Arctic Ocean as well as in the Northern Atlantic and Northern Pacific. Consequently, a redescription and proposal of a neotype is needed to clarify if there is more than one species. Støp-Bowitz (1948:32) noticed the nephridial lobes in the branchial plate, but he regarded them as accessory branchiae.

After the International Code of Zoological Nomenclature (1999, Art. 75), a neotype is being designated because there is no name-bearing type specimen, and because of the confusion between the above two species requires a designation to objectively define *D. longisetosus*. Consequently, in order to satisfy the qualifying conditions (Art. 75.3), it must be stated that this designation will clarify the taxonomic status, a description and illustrations have been presented to ensure the recognition of the species. Further, collection managers in several German museums were contacted in order to find the type material for this species, but none exists. On the other hand, the neotype fits the characteristics originally noticed in the species, it was found in a locality with ecological conditions similar to the ones prevailing in the original type locality, and has been deposited in the California Academy of Sciences.

Distribution. Originally described from Providence Bay (64°30'N, 173°30'W), Russia, these specimens come from Northern Alaska, about 1,200 km away, but despite the distance between them, these localities share the same environmental conditions, and the incomplete topotype specimens have most of the same morphological features.

Diplocirrus micans Fauchald, 1972

http://species-id.net/wiki/Diplocirrus_micans Fig. 8

Diplocirrus micans Fauchald 1972:218–219, Pl. 44, Figs. a–e; Darbyshire & Mackie 2009:97, Table 1.

Type material. Eastern Pacific Ocean. Holotype (LACM-AHF992), off Natividad Island, Baja California, RV Velero IV, Stat. 7229 (27°54'25"N, 115°40'00"W), 957–942 fathoms, 31 Dec. 1960.



Figure 8. *Diplocirrus micans* Fauchald, 1972. Holotype (LACM-AHF-993) **A** ventral view **B** same, anterior end, ventral view **C** non-type specimen (LACM-AHF-13754), anterior end, dorsal view **D** another non-type specimen (LACM-AHF-13755), chaetiger 14, right parapodium **E** same, basal, medial and distal notochaetal regions **F** same, basal and distal neurochaetal regions.

Additional material. Eastern Pacific Ocean. Several fragments (LACM-AHF 2615), off Natividad Island, Baja California, RV Velero IV, Stat. 7231 (from 27°24'00"N, 115°12'15"W, to 27°23'17"N, 115°13'45"W), 1355–1312 fathoms, green mud, 1 Jan. 1961. Median fragment (LACM-AHF 2612), off Natividad Island, Baja California, RV Velero IV, Stat. 7249 (27°36'25"N, 115°56'25"W), 2050–2027 fathoms, red clay and rock, 4 Jan. 1961. Two specimens (LACM-AHF 2611), 44 miles, 192 degrees N from Cabo Corrientes Lighthouse, RV Velero IV, Stat. 13754-70 (19°41"15"N, 105°53'00" W), 1220 fathoms, Campbell grab, 18 Jan 1970 (25–30 mm long, 1.2–1.5 mm wide, cephalic cage 1.5–3.0 mm long; chaetiger 1 with 2–3 noto- and 5–6 neurochaetae per bundle, 39 chaetigers; female with oocytes 125 µm). An anterior fragment (LACM-AHF 2611a), 35.3 miles 205 degrees T (T=true

north) from Cabo Corrientes Lighthouse, RV Velero IV, Stat. 13755-70 (19°51'30"N, 105°58'00"W), 1400 fathoms, Campbell grab, 18 Jan 1970 (7 mm long, 1.5 mm wide, cephalic cage chaetae 7 mm long; chaetiger 1 with 4–5 noto- and 5–6 neuro-chaetae).

Description. Holotype pale, damaged, without posterior end (in regeneration?), several parapodia removed, many chaetae broken. Body slightly swollen anteriorly, tapering posteriorly (Fig. 8A); 11 mm long, 1 mm wide, cephalic cage 1 mm long, 26 chaetigers. Tunic papillated, with abundant, fine sediment particles adhered. Papillae short, abundant (most eroded), cylindrical, longer in first chaetiger and in chaetal lobes, less than 1/3 chaetal length (very long in LACM-AHF 2615, as long as half notochaetal length).

Anterior end not exposed; not dissected to avoid further damage. Cephalic cage chaetae as long as body width. Chaetigers 1–2 involved in the cephalic cage; chaetae arranged in short, lateral lines, 2 chaetae per ramus. Anterior dorsal margin of first chaetiger papillated. Anterior chaetigers without long papillae. Chaetigers 1–3 progressively larger; notopodia with suprachaetal conical lobes. Post-cephalic cage chaetigers not elongated. Chaetal transition from cephalic cage to body chaetae abrupt; multi-articulared neurochaetae start in chaetiger 3. Gonopodial lobes not seen (Fig. 8B).

Parapodia porly-developed, chaetae emerge from the body wall (Fig. 8D). Parapodia lateral; median neuropodia ventrolateral. Noto- and neuropodia low, rounded lobes, very close to each other. All notochaetae multiarticulated capillaries, articles short basally, become long medially and distally (Fig. 8E). Median notochaetae arranged in a short, transverse line, holotype with 2–3 per bundle (other specimens with 8–9 chaetae per bundle), twice as long as body width. Neurochaetae multiarticulated capillaries in chaetigers 1–2; multiarticulated, thicker neurospines start in chaetiger 3, arranged in a transverse line, 4 per bundle (up to 8 in larger fragments Stat. 7231). Neurochaetae with short articles basally, become long medially, slightly decreasing their length distally; tips slightly falcate (Fig. 8F).

Posterior end unknown.

Remarks. *Diplocirrus micans* Fauchald, 1972 resembles other species with abundant papillae and long chaetae such as *D. longisetosus* (von Marenzeller, 1890), and *D. normani* (McIntosh, 1908), comb. n. However, *D. micans* separates from the two other species because its neurochaetae have long articles, and there are no gonopodial lobes, whereas the two other species have distal articles barely longer than wide, and gonopodial lobes.

The record by Fauchald and Hancock (1981:36) was based on a single, damaged specimen collected off Oregon, United States. The specimen (LACM-AHF 2616) resembles *D. micans* but it is brittle, apparently it has dried out in the past, so the conical lobes in first few chaetigers cannot be confirmed. However, this specimen has many more chaetae per bundle, especially in the anterior end, and articles are much longer than in *D. micans*, so it may be a different species, but the specimen is in poor shape and more specimens are required to describe it.

Distribution. Western Mexico, in deep water (1900–2800 m depth).

Diplocirrus nicolaji (Buzhinskaja, 1994), comb. n.

http://species-id.net/wiki/Diplocirrus_nicolaji Fig. 9

Diversibranchius nicolaji Buzhinskaya 1994:231, Figs. 2–7; Darbyshire & Mackie 2009:97, Table 1.

Flabelligeridae from Japan: Rouse & Pleijel 2001, Plate 11, Fig. f.

Type material. Northwestern Sea of Japan. Holotype (ZIRAS-48504), Vostok Bay (42°30'N, 133°00'E), Peter the Great Bay, Russia, 7 m, muddy sand, 26 Oct. 1989, G. Buzhinskaja, coll. Several paratypes (ZIRAS-48506), five anterior fragments (four with anterior end exposed, variously damaged), and several median fragments, Vostok Bay (42°50'N, 132°45'E), Peter the Great Bay, Russia, 7 m, muddy sand, 26 Oct. 1989, sample 2, G. Buzhinskaja & S. Kiyashko, coll. (anterior fragments 5.5–12.0 mm long, 0.7–2.0 mm wide, 12–24 chaetigers, chaetiger 1 notochaetae 0.3–0.6 mm, 10–22 transversal rows of papillae, gonopodial pores in chaetigers 3–7(–8, 9, 14 one each). Five paratypes (ZIRAS-48507), Vostok Bay (42°50'N, 132°45'E), Peter the Great Bay, 3m, muddy sand, 21 Sep. 1989, G. Buzhinskaja, coll. (6–9 mm long, 0.6–1.0 mm wide, 10–18 chaetigers, chaetiger 1 notochaetae 0.4–0.5 mm, 12–20 transversal rows of papillae, gonopodial pores in chaetigers 3–9(–10 in 2 paratypes, –11 in one; gut sediment particles heterogeneous, up to 0.6 mm long).

Additional material. Northwestern Pacific Ocean. Northwestern Sea of Japan, Peter the Great Bay, Russia. One specimen (ZIRAS-2/48505), Vostok Bay (42°50'N, 132°45'E), 7 m, muddy sand, 26 Oct. 1989, G. Buzhinskaja, coll. Five specimens (ZI-RAS-3/48506), one beheaded, Vostok Bay (42°50'N, 132°45'E), 7m, muddy sand, 26 Oct.1989, G.Buzhinskaja & S.Kiyashko, coll. Five specimens (ZIRAS-4/48507), Vostok Bay (42°50'N, 132°45'E), 3m, muddy sand, 21 Sept.1989, G.Buzhinskaja, coll. One specimen (ZIRAS-5/48508), beheaded, Posyet Bay (42°30'N, 131°00'E), 3 m, muddy sand, among *Zostera asiatica*, diving, sample from 0.25 m², 10 Mar. 1966, A.N. Golikov, coll. Three anterior fragments (ZIRAS-6/48509), beheaded, Tikhaya Bay, Posyet Bay (42°30'N, 131°00'E), 3m, muddy sand, among *Patiria pectinifera* and *Chaetopterus*, 3 Mar. 1966, diving, sample from 0.1 m², A.N. Golikov, coll. One specimen (ZIRAS-7/48510), Tikhaya Bay, Posyet Bay (42°30'N, 131°00'E), 4–5 m, muddy sand, among *Patiria pectinifera* and *Chaetopterus*, 21 Apr. 1965, diving, sample from 0.3 m², L. Chislenko, coll.

Description. Holotype (ZISP-48504) orange yellow, slightly macerated, without posterior end. Body anteriorly swollen, posteriorly tapering; 19 mm long, 1.9 mm wide, no cephalic cage (chaetiger 1 notochaetae 0.3 mm), 30 chaetigers. Tunic densely covered by papillae (Fig. 9A, C, E); papillae short, most 8-shaped, others digitate, with fine sediment adhered to their base, about 12 rows per segment.

Cephalic hood exposed, as long as the following 4 chaetigers, with small, sparse papillae, cephalic hood margin smooth. Prostomium low, eyes not seen. Palps thick, slightly longer than branchiae; palp lobes reduced, rounded. Other features from paratypes. Caruncle projected dorsally to the base of posterior branchiae, tapering, lateral



Figure 9. *Diplocirrus nicolaji* (Buzhinskaja, 1994), comb. n. Holotype (ZIRAS-48504) **A** anterior end, oblique lateral view, body in ventral view **B** same, head, frontal view, palps and one posterior branchia removed **C** same, anterior chaetigers, ventral view (arrows indicate ventral pores) **D** paratype (ZI-RAS-48506), posterior end, ventral view (arrows indicate ventral pores) **E** same, posterior end **F** same, chaetiger 13, right parapodium **G** Same, basal and distal notochaetal regions **H** same, basal and distal neurochaetal regions (insert: neurochaetal tip).

lobes elevated, posteriorly fused. Dorsal lip projected, lateral lips thicker, ventral lip reduced. Nephridial lobes in branchial plate not seen (Fig. 9B).

Branchiae of two different types (Fig. 9A, B). Posterior row with four prismatic, thicker, lamellate branchiae, lamella reaching the tips; lateral branchiae of the same size, with dorsal keel rounded, reduced, with longitudinal bands and laterally expanded dorsal surface, with a thin axis, branchial lateral margins with two rounded, sucker-like sockets; median branchiae with dorsal keel as those present in lateral branchiae, not foliose, corrugated. All posterior branchiae with a series of successive transverse blades on their ventral side; in median branchiae, all laterally fused making a single convoluted blade; in lateral branchiae the transverse blades laterally free. Anterior row with four thin, cirriform branchiae, shorter than palps, arranged in two lateral pairs, each filament with a convoluted lamella along its basal third, and successive ciliary bands medial- and distally. Interbranchial lobes small, between median and lateral branchiae (dorsal), and outside the lateral ones (lateral); dorsal lobes small, rounded, lateral lobes rounded, slightly larger).

First chaetiger displaced dorsally, notochaetae slightly longer than following ones. Notochaetae arranged in a short, oblique line with 2 multiarticulated hooks. Anterior dorsal margin of first chaetiger papillated, as following segments; no other modification. Anterior chaetigers without longer papillae, chaetiger 1 shorter than following ones, chaetal lobes lateral, very close to each other. Chaetigers 5–10 swollen, without

marked segmentation between them; therafter segments better defined. Gonopores orange-red, in chaetigers 2–12 (Fig. 9C, D).

Parapodia poorly developed; chaetae emerge from the body wall (Fig. 9F). Notopodia and neuropodia with papillae as long as the others. Noto- and neuropodia close to each other. Notochaetae multiarticulated capillaries, all articles long (Fig. 9G). Median notochaetae arranged in a longitudinal line, with 4 per bundle in holotype (11 per bundle in larger; 6–7 in smaller specimens), longest about as long as one-third body width. Median neuropodia lateral, very close to notopodia. Neurochaetae multiarticulated hooks from chaetiger 1 (Fig. 9G), arranged in a short longitudinal line (J-pattern in other specimens), 3–4 per bundle (6–8 in other specimens), each with long articles of about the same length, distal article falcate, finely transversely divided, not articulated, with a hood-like membrane.

Posterior end missing in holotype (probably invaginated); a posterior fragment (ZISP-48507, Fig. 9E) tapering to a rounded lobe; pygidium with anus dorsoterminal, dark, muscular, without anal cirri.

Variation. Living specimens dark-orange, gills green. The paratypes were orangeyellow to orange-brown, with 29–31 chaetigers.

Remarks. Diplocirrus nicolaji (Buzhinskaja, 1994), comb. n. is closely allied to *D. branchiatus* (Rullier, 1965) because the bodies of these species lack sediment particles, have ventrolateral gonopores in some anterior chaetigers, reduced chaetae in the first chaetiger, and their caruncles taper posteriorly. Their main differences rely on the relative neurochaetal development in median chaetigers, and on the area covered by lamellae in the cirriform branchiae; thus, *D. nicolaji* has barely tapering neurochaetae, with some 10 articles of about the same length, tips markedly falcate, and their cirriform branchiae has a lamellate region extending up to one-third of the branchial length, whereas in *D. branchiatus*, the neurochaetae are tapering, provided with about 23 articles, decreasing in size distally, tips delicately falcate, and the lamellate region along cirriform branchiae might reach one-fifth of the branchial length.

Distribution. Originally described from Vostok Bay, Peter the Great Bays, Northwestern Sea of Japan, in shallow water soft bottoms (3–7 m).

Diplocirrus normani (McIntosh, 1908), comb. n.

http://species-id.net/wiki/Diplocirrus_normani Fig. 10

Stylarioides normani McIntosh 1908:542–543, Pl. 12, Figs. 3, 8.
Stylarioides longisetosus von Marenzeller 1892:426–427 (non von Marenzeller, 1890).
Diplocirrus longisetosus: Haase 1915:200–202, Textfigs. 6–7 (partim); Støp-Bowitz 1948a:30–33, Fig. 8; Støp-Bowitz 1948b:38–39, map (non von Marenzeller, 1890).

Type material. Barents Sea. Holotype of *Stylarioides normani* (BMNH-1921.5.1.2646), Finmark, Northern Norway, Stat. 49, 1890, C. Norman, coll. (anterior fragment,



Figure 10. *Diplocirrus normani* (McIntosh, 1908), comb. n., reinst. Non-type specimen (ECOSUR): **A** complete, dorsal view **B** same, anterior end, dorsal view **C** same, anterior end, ventral view **D** same, chaetiger 13, right parapodium **E** same, basal to distal notochaetal regions **F** same, basal to distal neurochaetal regions.

dried-out, 7.5 mm long, 1.5 mm wide, cephalic cage 3 mm long, 14 chaetigers; right chaetiger 10 previously removed).

Additional material. Barents Sea. Two specimens (ECOSUR), White Sea, Russia, 5 Aug. 1999, A. Filippova, coll. (complete specimen used for redescription; anterior fragment 6 mm long, 1.5 mm wide, cephalic cage 2 mm long, 14 chaetigers). One specimen (ECOSUR), White Sea, Russia, 15 m, Jul. 1999, A. Filippova, coll. (anterior fragment 3 mm long, 1 mm wide, cephalic cage 2.3 mm long, 11 chaetigers). Two specimens (ECOSUR) complete, slightly damaged, Kandalalsha Bay, White Sea, Russia, 40 m, mud, 1 Aug. 2004, A. Zhadan, coll. (specimen with anterior end exposed used for description; 5.0–5.5 mm long, 0.8–1.0 mm wide, cephalic cage 1.0–1.3 mm long, 20–21 chaetigers; exposed anterior end 0.7 mm long). Northwestern Atlantic Ocean. Several specimens (USNM-48491), Cape Cod Bay, Massachusetts, Stat. 1424, 35.1–33.6 m, 19 Nov. 1968, C.D. Long. Coll. Id.

Description. Non-type specimen (ECOSUR) complete, yellowish gray. Body club-shaped, anteriorly swollen, slightly narrowing to chaetiger 13, then apparently regenerating the posterior, cylindrical region, tapering to posterior end (Fig. 10A); 12 mm long, 1.5 mm wide, cephalic cage 3 mm long, 25 chaetigers. Tunic papillated, with fine sediment particles. Papillae eroded, core and tips black, cirriform, sparse,

fragile, about 7–8 transverse rows in median chaetigers, becoming longer dorsally (Fig. 10B); in median chaetigers about 1/5–1/6 as long as notochaetae.

Cephalic tube long, smooth, margin apparently smooth. Prostomium low, eyes not seen. Caruncle not seen. One palp remaining, thick, longer than remaining branchiae, longitudinal furrow shallow; palp keels reduced. Dorsal and ventral lips reduced, lateral lips thicker. Branchiae cirriform, most lost, sessile on branchial plate, arranged in two concentric rows, distal row continuous with 4 thicker filaments bases, proximal row discontinuous, filaments probably thinner, lower filaments bases smaller. Nephridial lobes very thin, long, placed below the posterior row lateral filaments.

Cephalic cage chaetae ¹/₄ as long as body length, or twice as long as body width. Chaetigers 1–2 involved in the cephalic cage; chaetae arranged in short dorsolateral lines, 6–8 noto- and 4–6 neurochaetae per bundle. Anterior dorsal margin of first chaetiger papillated, black; anterior chaetigers without especially long papillae. Chaetigers 1–3 of about the same length. Post-cephalic cage chaetigers not elongated. No chaetal transition from cephalic cage to body chaetae, all neurochaetae similar. Gonopodial lobes present in chaetiger 5, low, rounded, black, covered by small papillae (Fig. 10C).

Parapodia lateral, poorly developed, chaetae emerge from the body wall (Fig. 10D); median neuropodia ventrolateral. Notopodia 2–3 with very low conical lobes directed forward, remaining notopodia less prominent. Neuropodia 2–5 with low conical chaetal lobes. Noto- and neuropodia distant to each other.

Median notochaetae arranged in a transverse horizontal C-shaped pattern; all notochaetae multiarticulated capillaries, short articles basally and distally, long medially (Fig. 10E). About 10 (-12) chaetae per bundle, at least twice as long as body width. All neurochaetae multiarticulated capillaries, short, poorly-defined articles along basal half or 2/3 chaetal length, better-defined, medium-sized and then long articles along the rest of chaetae (Fig. 10F), tips straight, arranged in a transverse line, 9–10 per bundle.

Posterior end tapering to a rounded lobe; pygidium with anus terminal, blackish, without anal cirri.

Remarks. *Diplocirrus normani* (McIntosh, 1908), comb. n. was regarded as a junior synonym of *D. longisetosus* (von Marenzeller, 1890) by Haase (1915:200) because they are very similar. As stated above, they also resemble *D. micans* Fauchald, 1972, though the latter separates from the other two species because it lacks gonopodial lobes and its neurochaetae have long articles. Thus, once *D. longisetosus* has been restricted, these species differ regarding coloration of body, papillae and gonopodial lobes, and because of the relative resolution of neurochaetal basal articles. Thus, in *D. normani,* although the body is grayish, papillae and gonopodial lobes are darker or blackish, and neurochaetal basal articles are poorly-defined, whereas in *D. longisetosus*, on the contrary, the papillae and gonopodial lobes are pale, and the basal articles of neurochaetae are well-defined.

Distribution. Originally described from Finmark, Northern Norway, Barents Sea. It ranges along Northeastern and Northwestern Atlantic areas, in shallow water.

Diplocirrus octobranchus (Hartman, 1965), comb. n.

http://species-id.net/wiki/Diplocirrus_octobranchus Fig. 11

- *Ilyphagus octobranchus* Hartman 1965:178–179, Pl. 39; Hartman & Fauchald 1971:120–121.
- *Diplocirrus octobranchus*: Day 1973:107 (informal comb. n.); Darbyshire & Mackie 2009:97, Table 1.

Type material. Eastern Atlantic Ocean. Holotype (LACM-AHF 540) and 19 paratypes (LACM-AHF 541), off New England, United States, RV Atlantis Stat. Slope 3 (39°58'24"N, 70°41'18"W), 300 m, 28 Aug. 1962, H. Sanders, coll. (two complete paratypes 7–16 mm long, 0.8–1.0 mm wide, cephalic cage 0.8–2.0 mm long, 24–42 chaetigers; gonopodial papillae not visible; smaller paratypes with relatively more sand particles over their bodies; broken mature female with oocytes about 120 μ m).

Additional material. North Carolina. One specimen (USNM-54938), Eastward Stat. 6269 (34°16.5'N, 75°44'W), 500–520 m, 11 Nov. 1966, G. Rowe, coll. One specimen (USNM-54932), Eastward Stat. 6241 (33°13.6' N, 76°13.4' W), small biological trawl, 1000–1020 m, 9 Nov. 1966, G. Rowe coll.

Description. Holotype an anterior fragment, brownish. Body anteriorly swollen, posteriorly tapered (Fig. 11A); 8.5 mm long, 1 mm wide (widest by chaetigers 5–6, 2 mm), cephalic cage 2 mm long, 17 chaetigers. Tunic papillated, sediment particles mostly fine, adherent on papillae bases, and few larger sand grains, especially dorsally (Fig. 11B); smaller specimens with more sand particles on the body. Papillae of varying lengths, longer dorsally and on chaetal lobes, may be as long as chaetae, shorter in the rest of the body, 4–5 rows per chaetiger.

Cephalic hood tube long, made of two rings, basal one shorter, both smooth; cephalic hood margin smooth. Prostomium low, eyes not seen (Fig. 11C). Caruncle low, wide. Palps lost (pale, laterally corrugated, 1.5 times longer than branchiae in one paratype); palp keels rounded, elevated. Lateral lips thick, projected outwards, rounded. Ventral lip reduced. Dorsal lip projected as a triangular lobe. Branchiae cirriform of two different widths; posterior row with thicker filaments, rectangular, with a middorsal black band, branchial bases continuous, anterior row with branchiae thinner, cirriform, separated in two lateral pairs. Branchiae of about the same length; size relationships with palps unknown. Nephridial lobes in branchial plate low, whitish.

Cephalic cage chaetae as long as widest body section. Only chaetiger 1 involved in the cephalic cage; chaetae arranged in a short, dorsolateral line with 4(-5) notoand 2(-8 in paratypes) neurochaetae. Anterior dorsal margin of first chaetiger papillated. Chaetigers 1–3 progressively larger. Post-cephalic cage chaetigers not elongated. Chaetal transition from cephalic cage to body chaetae abrupt, thicker neurospines present from chaetiger 2. Gonopodial lobes not seen.

Parapodia poorly-developed, chaetae emerge from the body wall (Fig. 11D). Parapodia lateral; median neuropodia ventrolateral. Noto- and neuropodia without pro-


Figure 11. *Diplocirrus octobranchus* (Hartman, 1965), comb. n. **A** holotype (LACM-AHF 540), dorsal view **B** same, anterior end, dorsal view **C** paratype (LACM-AHF 541), head, frontal view, palps and branchiae removed **D** same, chaetiger 18 **E** same, basal, medial and distal notochaetal regions **F** same, basal, medial and distal neurochaetal regions.

jected chaetal lobes. Papillae abundant, 2–4 larger ones in chaetal lobes. Noto- and neuropodia close to each other.

Median notochaetae arranged in a short transverse line, chaetae directed dorsally. All notochaetae multiarticulated capillaries. Median notochaetae 1.5–2.0 times as long as body width, 7 per bundle, articles short basally, feebly defined, become medium-sized medially, long distally (Fig. 11E). Neurochaetae multiarticulated capillaries in chaetiger 1; thicker multiarticulated neurospines from chaetiger 2, two (–5 in paratypes) per ramus, become thinner in the tapered median and posterior region, being 5 per ramus, arranged in a transverse line. Neurochaetae with feebly-defined short, basal articles, become very long medially, and decrease progressively to the straight tip (Fig. 11F). Posterior end observed in one complete paratype, tapering to a swollen pygidium, with anus dorsoterminal, without anal cirri. One paratype is a damaged female, oocytes $100-150 \mu m$.

Remarks. *Diplocirrus octobranchus* (Hartman, 1965), comb. n., is closely allied to an undescribed species from Antarctica, and both differ from other species with long papillae because they have sand particles over the body. These two species differ in the extent of sediment particles along the papillae and on the relative length of the neurochaetal anchylosed region. Thus, in *D. octobranchus* sediment particles are restricted to the base of papillae, and their neurochaetae have an anchylosed region of about onefifth of the chaetal length, whereas in the Antarctic undescribed species, the sediment particles spread along the papillae, and the anchylosed region might be about half or one-third of the chaetal length.

Diplocirrus octobranchus is a typical member of the genus because its branchiae are of two different widths. It does not belong in *Ilyphagus* because it has multiarticulated neurospines, with long articles in the medial and distal regions, and short articles only basally, whereas in *Ilyphagus* neurochaetae are aristate spines with very short articles basal- and medially, and distally hyaline. Further, the cephalic cage chaetae in *Ilyphagus* are clearly dorsal whereas in *Diplocirrus* they are lateral, or dorsolateral at most. After Hartman amended *Ilyphagus* (Hartman 1965:177), the correct placement for her new species as a member of *Diplocirrus* was indirectly stated by comparing it to *D. glaucus*, the type species for the genus (Hartman 1965:179). This made Day (1973:106) suggest the informal, new combination, which is herein confirmed after the examination of the type material and of the redefinition of *Diplocirrus*.

Distribution. Apparently discontinuous; off New England in 300–1000 m, and off northeastern South America in 770–805 m.

Diplocirrus stopbowitzi Darbyshire & Mackie, 2009

http://species-id.net/wiki/Diplocirrus_stopbowitzi Fig. 12

Diplocirrus stopbowitzi Darbyshire & Mackie 2009:93–96, Figs. 1–3A, Table 1.

Material examined. One specimen, broken in three pieces, Stat. BSA 449, 12 mm long, 0.8 mm wide, 25 chaetigers, A. Ravara, coll. (no further data available).

Diagnosis. Body slightly swollen anteriorly (Fig. 12A). Papillae abundant, short, giving a velvety oultlook, without sediment particles (Fig. 12B). Median chaetigers with 5–6 notochaetae and 2–3 neurochaetae; posterior chaetigers with three notochaetae and two neurochaetae. Notochaetae with long articles throughout the chaeta (Fig. 12C). Neurochaetae with long articles, being 7–8 times longer than wide, tips falcate (Fig. 12D).



Figure 12. *Diplocirrus stopbowitzi* Darbyshire & Mackie, 2009. Non-type specimens **A** complete, lateral view (photo by Teresa Darbyshire) **B** another specimen, chaetiger 20, left parapodium **C** same, notochaetal basal regions **D** same, basal, medial and distal neurochaetal regions.

Remarks. As stated above, *Diplocirrus stopbowitzi* Darbyshire & Mackie, 2009 resembles *D. kudenovi* sp. n. because in both species the body has hemispherical papillae, but lacks sand particles or ventrolateral gonopores. They especially differ regarding some neurochaetal features in median chaetigers such as their number and the relative length of articles; thus, *D. stopbowitzi* has 2–3 neurochaetae, each with long articles being about seven times longer than wide, whereas *D. kudenovi* has 5–6 neurochaetae and each has shorter articles, each being twice as long as wide.

Distribution. Southern Irish Sea, offshore, in gravel or gravelly-sand bottoms, 38–112 m depth.

Diplocirrus sp. n. Antarctica

Material examined. One specimen (USNM 46405), without posterior region, RV Staten Islands, Stat. 9-63 (64°48'S, 63°30'W), Port Lockroy, off Wiencke Island, Anvers Island, 31 fathoms, dredged at anchorage, mud bottom, 26 Jun. 1963, W.L. Schmitt, coll. (5.5 mm long, 0.7 mm wide, cephalic cage 0.9 mm long, 20 chaetigers). One slide with three segments (USNM 56470).

Remarks. This undescribed species is closely allied to *D. octobranchus* because both have sediment particles on the body and 7–8 notochaetae per bundle in median chaetigers. They differ because in *Diplocirrus* sp Antarctica, the sediment particles are adhered in the body wall and in the whole papillae, whereas in *D. octobranchus*, sediment particles are restricted to the base of the papillae leaving bare both the body wall and the papillae. Another important difference is the extension of the anchylosed articles; thus, the anchylosed portion is one-half or at least one-third of notochaetal length in the Antarctic species, whereas it is only one-fifth or less of neurochaetal length in *D. octobranchus*.

Diplocirrus sp. n. Morocco

Stylarioides scutigeroides: Fauvel 1936:77 (partim, non Augener, 1918).

Material examined. Morocco. Two anterior fragments (MNHN-361), most chaetae broken, RV Vanneau, Stat. 6 (31°42'N, 09°43'W), 22 m, 1 Jul. 1923, R.P. Dollfus & J. Liouville, coll. (5.0–5.5 mm long, 1.5 mm wide, cephalic cage 1.5 mm long, 12 chaetigers; anterior end dissected, it has the typical *Diplocirrus* pattern; i.e. 8 branchial filaments with the posterior ones thicker).

Remarks. This species differs from other species with short papillae because *D*. sp. Morocco has very short lateral papillae and the body wall has a thin layer of sediment grains. However, there are no more specimens available from the same expedition.

Distribution. Only known from off Cape Guir, Morocco, in 22 m depth.

Diplocirrus sp. n. Sri Lanka

Material examined. Three specimens (MNHN-unnumb.), off SW Sri-Lanka, RV Marion Dufresne, SAFARI II Cruise, Stat. 2 (05°37'N, 78°24'E), 3660 m, Jul. 1981.

Description. Three anterior fragments variously damaged. Body cylindrical, tapering posteriorly; 2.5–3.5 mm long, 0.7–1.2 mm wide, cephalic cage (broken) 1 mm long, 8–10 chaetigers. Tunic thin, without foreign particles, with 4 longitudinal rows of elongate papillae.

Cephalic hood not exposed. Anterior end not dissected to avoid further damage. Cephalic cage chaetae about as long as body width. Chaetiger 1 involved in the cephalic cage; chaetae in short ventrolateral lines, 1–2 noto- and 2–3 neurochaetae per ramus. Anterior dorsal margin of first chaetiger papillated, papillae elongate, clavate. Anterior chaetigers without especially long papillae. Chaetigers 1–3 of about the same length. Chaetal transition from cephalic cage to body chaetae abrupt; thicker neurochaetae start in chaetiger 2. Gonopodial lobes not seen.

Parapodia poorly-developed, chaetae emerge from the body wall. Parapodia lateral; median neuropodia ventrolateral. Noto- and neuropodia close to each other, each with 2–3 longer clavate papillae. Median notochaetae arranged in a tuft, most broken; all notochaetae multiarticulated capillaries, articles long; in median chaetigers 2–3 per bundle, as long as 2/3 body width. Neurochaetae multiarticulated capillaries in chaetiger 1; thicker articulated neurospines from chaetiger 2, with articles short basally, medial- and distally long, 2–3 per bundle.

Posterior end unknown.

Remarks. With the available specimens and as indicated in the key above, this species differs from all other species in the genus because it has a rather smooth body. Better specimens would clarify its affinities and allow a description.

Distribution. Only known from the type locality, off Sri-Lanka, in 3660 m depth.

Acknowledgments

This contribution could not be possible without the warm support from many colleagues, curators and collections managers of several museums or research centers. In alphabetical order by name they were Anna Filippova and Anna Zhadan (Moscow University), Ascensao Ravara (Univ. Aveiro, Portugal), Emma Sherlock and Miranda Lowe (BMNH), Chris Glasby (NTM), Dieter Fiege (SMF), Fredrik Pleijel (then in MNHN), Kristian Fauchald and Goeff Keel (USNM), Leslie Harris, Kathy Omura and Kirk Fitzhugh (LAMNH), Mal Bryant and Rob Adlard (QM), Marie-Noelle Helleout and Tarik Meziane (MNHN), Norma Emilia González (ECOSUR), Pat Hutchings and Penny Barents (AM), Patrick Gillet (IRFA), Robert van Syoc (CAS), Teresa Darbyshire (Wales Museum), Tore Hoisater and Jon Anders Kongsrud (ZMUB). Most of this research was made during a sabbatical leave from Ecosur and after a research visit in Sankt-Peterburg, Russia. Eunice Salazar, Joanna Zanol, Rosario Reyes, and Mario Londoño helped by scanning or typing some German documents, to be translated for this work. Many librarians were very helpful: Martha Rose, David Steer, and Richard Greene, all from USNM, and Julia Dunaeva (ZIRAS), searched and found important publications in Washington and in Moscow. The careful reading by Karen Osborn and the editor resulted in a significant improvement for this final version. Funding was provided by Ecosur and Conacyt (61609). Additional funds came from the U.S.A. Department of the Interior, Fulbright Commission, and from the Université Catholique de l'Ouest, Angers, France.

References

- Appy TD, Linkletter LE, Dadswell MJ (1980) Annelida: Polychaeta. In: A Guide to the Marine Flora and Fauna of the Bay of Fundi. Canada, Fisheries and Marine Service, Technical Report 920: vi + 124 pp.
- Buzhinskaja GN (1994(1993)) *Diversibranchius nicolaji* gen. et sp. n. from the Sea of Japan with unique branchial structure (Polychaeta: Flabelligeridae). Zoosystematica Rossica 2: 229–231.
- Chamberlin RV (1919) The Annelida Polychaeta of the *Albatross* Tropical Pacific Expedition, 1891–1905. Memoirs of the Museum of Comparative Zoology, Harvard College 48: 1–514.
- Darbyshire T, Mackie ASY (2009) Two new species of *Diplocirrus* (Polychaeta: Flabelligeridae) from the southern Irish Sea and South Africa. Zoosymposia 2: 91–103.
- Day JH (1961) The Polychaeta Fauna of Sout Africa, 6. Sedentary species dredged off Cape coast with a few new records from the shore. Zoological Journal of the Linnean Society, London 44: 463–560. doi:10.1111/j.1096-3642.1961.tb01623.x
- Day JH (1967) A Monograph on the Polychaeta of Southern Africa. British Museum (Natural History) Publications 656: 38 + 878 pp.
- Day JH (1973) New Polychaeta from Beaufort, with a key to all species recorded from North Carolina. National Oceanographic and Atmospheric Administration, Technical Report of the National Marine Fisheries Service, Circular 375: 1–153.
- de Saint-Joseph A (1894) Les annélides polychètes des côtes de Dinard, troisième partie (Nephtydiens – Serpuliens). Annales des Sciences Naturelles, Zoologie, 7ème série 17: 1–395.
- de Saint-Joseph A (1898) Les annélides polychètes des côtes de France (Manche et Océan). Annales des Sciences Naturelles, Zoologie, 8ème série, 5: 209–464.
- delle Chiaje S (1831(1829)) Memoria sulla Storia e Notomia degli Animali senza Vertebre del Regno di Napoli: Memoria III + IV. Societa' Tipographica, Napoli, Volume 4: 117–214.
- Ditlevsen H (1911) Annelids from the Danmark Expedition. Danmark-Ekspeditionen til Grønlands nordøstkyst 1906–1908 5: 410–432.
- Fauchald K (1972) Benthic polychaetous annelids from deep waters off Western Mexico and adjacent areas in the Eastern Pacific Ocean. Allan Hancock Monographs in Marine Biology 7: 1–575.
- Fauchald K (1977) The polychaete worms: Definitions and keys to the orders, families and genera. Natural History Museum of Los Angeles County, Science Series 28: 1–190.
- Fauchald K, Hancock DR (1981) Deep-water polychaetes from a transect off Central Oregon. Allan Hancock Foundation Monographs 11: 1–73.
- Fauvel P (1927) Polychètes sédentaires & addenda aux polychètes errantes. Faune de France 16: 1–494.
- Fauvel P (1932) Annelida Polychaeta of the Indian Museum, Calcutta. Memoirs of the Indian Museum, Calcutta 12: 1–262.
- Fauvel P (1937) Les fonds de pêche près d'Alexandrie, 11. Annélides polychètes. Direction des Recherches des Pêcheries, Le Caire, Notes & Mémoires 19: 1–60.
- Fauvel P (1946) Annélides polychètes des croisières du *Pourquoi-Pas?* Bulletin du Museum d'Histoire Naturelle, Paris, 2ème série 18: 397–403.
- Fauvel P (1953) The Fauna of India including Pakistan, Ceylon, Burma and Malaya. Annelida Polychaeta. Indian Press, Allahabad, 12 + 507.

- Gallardo VA (1968(1967)) Polychaeta from the Bay of Nha Trang, South Viet Nam. Scientific Results of Marine Investigations of the South China Sea and the Gulf of Thailand, 1959–1961. Scripps Institution of Oceanography NAGA Report 4: 35–279.
- Gibbs PE (1971) The polychaete fauna of the Solomon Island. Bulletin of the British Museum (Natural History), Zoology 21: 99–211.
- Grube AE (1850) Die Familien der Anneliden. Archiv für Naturgeschichte, Berlin 16: 249–364.
- Grube AE (1877a) Mittheilungen über die Familie der Chlorhaeminen. Jahres-Bericht der Schlesischen Gesellschaft für Vaterländische Cultur 45: 60–73.
- Grube AE (1877b) Annelidenausbeute von S.M.S. Gazelle. Monatsbericht der Königlich Preussischen Akademie der Wissenschafen zu Berlin 1877: 507–554.
- Haase P (1915) Boreale und arktisch Chloraemiden. Wissenschaftliche Meeresuntersuchungen der Kommission zur Wissenschaftlichen Untersuchung der Deutschen Meere, Neue Folge, Kiel 17: 169–228.
- Hansen A (1879(1878)) Annelider fra den norske Nordhavsexpedition i 1876. Nyt Magazin for Naturvidenskaberne 24: 1–17.
- Hansen A (1880) Annelider fra den norske Nordhavsexpedition i 1878. Nyt Magazin for Naturvidenskaberne 25: 224–234.
- Hansen A (1882) The Norwegian North-Atlantic Expedition 1876–1878, VII. Zoology, Annelida, Christiania, 54 pp. [bilingual publication, English and Norwegian]
- Hartman O (1961) Polychaetous annelids from California. Allan Hancock Pacific Expeditions 25: 1–226.
- Hartman O (1965) Deep-water benthic polychaetous annelids off New England to Bermuda and other North Atlantic areas. Allan Hancock Occasional Papers 28: 1–378.
- Hartman O, Fauchald K (1971) Deep-water benthic polychaetous annelids off New England to Bermuda and other North Atlantic areas, 2. Allan Hancock Monographs in Marine Biology 6: 1–327.
- Hartmann-Schröder G (1971) Annelida, Borstenwürmer, Polychaeta. Die Tierwelt Deutschlands 58: 1–594.
- Hartmann-Schröder G (1996) Annelida, Borstenwürmer, Polychaeta. Die Tierwelt Deutschlands 58, 2nd ed., Fischer, Jena, 645 pp.
- Hartmann-Schröder G, Rosenfeldt P (1989) Die Polychaeten der "Polarstern"-Reise ANT III/2 in die Antarktis 1984, 2. Cirratulidae bis Serpulidae. Mitteilungen aus dem Hamburgischen Zoologischen Museum und Institut 86: 65–106.
- ICZN (International Commission on Zoological Nomenclature) (1999) International Code of Zoological Nomenclature, 4th ed. International Trust for Zoological Nomenclature (The Natural History Museum), London, 306 pp (www.iczn.org/iczn/index.jsp).
- Jirkov IA, Filippova AV (2001) Flabelligeridae Saint-Joseph, 1894; pp 348–363 In: Polikhety Severnogo Ledovitogo Okeana. I.A. Jirkov. Yanus-K, Moskva, 632 pp.
- Langerhans P (1881) Die Wurmfauna von Madeira, 3. Zeitschrift für wissenschaftliche Zoologie 34: 87–143.
- Malmgren AJ (1867) Annulata Polychaeta: Spetsbergiae, Groenlandiae, Islandiae et Scandinaviae. Hactenus Cognita. Kongelige Vetenskaps-Akademiens Förhandlingar 1867(4): 127–235, Pl. 2–15.

- McIntosh WC (1908) Notes from the Gatty Marine Laboratory, St. Andrews, 30: 1. On the stranding of an adult female *Mesoplodon bidens*, Sowerby, at St. Andrews; 2. On an abnormal plaice with a precaudal fin-frill on the left side; 3. On *Orthogoriscus mola*, Bl.; 4. On the British Sphaerodoridae, Chloraemidae, and Chaetopteridae; 5. On the same families dredged in the 'Porcupine' expeditions of 1869 and 1870; 6. On the foregoing families dredged in the Gulf of St. Lawrence, Canada, by Dr. Whiteaves; 7. On the same families dredged in Norwegian waters and in Finmark by Canon Norman. Annals and Magazine of Natural History, series 8, 2: 524–545.
- McIntosh WC (1915) A Monograph of the British Annelids, 3. Polychaeta: Opheliidae to Ammocharidae. Ray Society, London, 368 pp.
- Milligan MR (1984) Family Flabelligeridae Saint Joseph, 1894; pp 47.1–47.20 In: Uebelacker JM, Johnson PG (Eds) Taxonomic Guide to the Polychaetes of the Northern Gulf of Mexico. Barry A. Vittor, Mobile, 7 vols.
- Pettibone MH (1982) Annelida. In: Parker SP (Ed) Synopsis and Classification of Living Organisms. McGraw Hill, New York, 1–43
- Rioja-LoBianco E (1931) Estudio de los poliquetos de la península Ibérica. Memorias de la Academia de Ciencias Físicas, Exactas y Naturales, Madrid 2: 1–471.
- Rouse GW, Pleijel F (2001) Polychaetes. Oxford University Press, Oxford, 354 pp.
- Rullier F (1964) Contribution à la faune des annélides polychaètes du Cameroun. Bulletin de l'Institut Français d'Afrique Noire, A, 36(4): 1071–1102.
- Rullier F (1965) Contribution à la faune des annélides polychètes de l'Australie. University of Queensland Papers, Department of Zoology 2(9): 163–201.
- Salazar-Vallejo SI (2007) Revision of *Flabelliderma* Hartman, 1969 (Polychaeta: Flabelligeridae). Journal of Natural History 41: 2037–2061. doi:10.1080/00222930701536443
- Salazar-Vallejo SI (in press) Revision of *Stylarioides* delle Chiaje, 1831 (Polychaeta: Flabelligeridae). Italian Journal of Zoology.
- Salazar-Vallejo SI (submitted) Revision of *Trophoniella* Caullery, 1944 (Polychaeta: Flabelligeridae). Zoosystema.
- Salazar-Vallejo SI, Carrera-Parra LF, Fauchald K (2008) Phylogenetic affinities of the Flabelligeridae (Annelida, Polychaeta). Journal of Systematic Zoology and Evolutionary Research 46: 203–215. doi:10.1111/j.1439-0469.2008.00464.x
- Spies RB (1975) Structure and function of the head in flabelligerid polychaetes. Journal of Morphology 147: 187–208. doi:10.1002/jmor.1051470206
- Støp-Bowitz C (1948a) Les Flabelligériens Norvégiens. Bergens Museums Arbok 1946(2): 1–59.
- Støp-Bowitz C (1948b) Sur les polychètes arctiques, des famillies des glicériens, des ophéliens, des scalibregmiens et des flabelligériens. Tromsø Museums Årshefter 66(2): 1–58.
- Uschakov PV (1955) Polychaeta of the Far Eastern Seas of the USSR. Israel Program for Scientific Translations, Jerusalem, (transl. 1965), 26 + 419.
- von Marenzeller E (1889) Beiträge zur Fauna Spitzbergens. Spitzbergische Anneliden. Archiv für Naturgeschichte, Berlin 55: 127–132.
- von Marenzeller E (1890) Annulaten des Beringmeeres. Annalen des Kaiserliche Naturhistorische Hofmusem, Wien 5: 1–18.

- von Marenzeller E (1892) Zoologische Ergebnisse der im Jahre 1889 auf Kosten der Breme Geographischen Gesellschaft von Dr. Willy Kükenthal und Dr. Alfred Walter ausgeführten Expedition nach Ostspitzbergen. Zoologische Jahrbücher, Abteilung für Systematik, Ökolobie und Geographie der Tiere 1891: 397–434.
- Webster HE (1879) On the Annelida Chaetopoda of the Virginian Coast. Transactions of the Albany Institute 9: 202–269.
- Webster HE, Benedict JE (1887) The Annelida Chaetopoda from Eastport, Maine. Annual Report of the Commissioner of Fish and Fisheries 1885: 707–758.
- Wesenberg-Lund E (1950) The Polychaeta of West Greenland with special reference to the Fauna of Nordre Strømfjord, Kvane-, and Bredefjord. Meddelelser om Grønland udgivne af Kommissionen for Videnskabelige undersøgelser I Grønland 151(2): 1–171.

RESEARCH ARTICLE



Monomorium dryhimi sp. n., a new ant species (Hymenoptera, Formicidae) of the M. monomorium group from Saudi Arabia, with a key to the Arabian Monomorium monomorium-group

Abdulrahman S. Aldawood[†], Mostafa R. Sharaf[‡]

Plant Protection Department, College of Food and Agriculture Sciences, King Saud University, Riyadh 11451, P. O. Box 2460, Saudi Arabia

turn:lsid:zoobank.org:author:477070A0-365F-4374-A48D-1C62F6BC15D1 urn:lsid:zoobank.org:author:E2A42091-0680-4A5F-A28A-2AA4D2111BF3

Corresponding author: Mostafa R. Sharaf (antsharaf@yahoo.com)

Academic editor: Donat Age	ti Received 15 April 2	2011 Accepted 2	9 April 2011	Published 15 June 2011
urn:	id:zoobank.org:pub:917A65	55B-F4CB-490C-84CI	E-C6E106E8FC81	D

Citation: Aldawood AS, Sharaf MR (2011) *Monomorium dryhimi* sp. n., a new ant species (Hymenoptera, Formicidae) of the *M. monomorium* group from Saudi Arabia, with a key to the Arabian *Monomorium monomorium*-group. ZooKeys 106: 47–54. doi: 10.3897/zooKeys.106.1390

Abstract

A new ant species, *Monomorium dryhimi*, is described based on workers from a single colony collected in Al Bahah, Asir Province, Saudi Arabia. This is the fourth species of the *Monomorium monomorium*-group collected from Arabian Peninsula, and appears to be closely related to *Monomorium holothir* Bolton, 1987, from Kenya. It can be distinguished by the following characters: head in profile with a weakly convex dorsal surface and a clearly convex ventral surface; eyes of moderate size with maximum diameter EL 0.19–0.25 × HW and with 6 ommatidia in the longest row; body colour yellow to light brownish yellow. In some individuals, head and gaster slightly but conspicuously darker than rest of body. Second halves of first and second gastral tergites with two characteristic brownish transverse bands. An identification key to the workers of the Arabian species of the *Monomorium monomorium*-group is presented. Scanning electron micrographs are given to illustrate the new species.

Keywords

Myrmicinae, Arabia, new species, alpha taxonomy, Palaearctic region, Asir Province, endemism

Introduction

The ant genus *Monomorium* was established by Mayr (1855) for the newly described species *M. minutum* Mayr (which was given the new name *Monomorium monomorium* Bolton (1987:287)). This genus includes more than 300 species and subspecies (Bolton 1995; Bolton et al. 2007) found in all zoogeographic regions with most species occurring in the Old World tropics and temperate regions (Brown 2000). Taxonomic revisions of the Australian and Malagasy *Monomorium* fauna were carried out by Heterick (2001, 2006) respectively. The Afrotropical *Monomorium* fauna was comprehensively revised by Bolton (1987). Two new South American species of *Monomorium* were described by Fernández (2007) and notes on the genus were presented. The *Monomorium* fauna of Arabian Peninsula was reviewed and listed giving 53 species for the region (Collingwoood and Agosti 1996). Most species of *Monomorium* nest in rotten wood, under stones, or directly in the soil.

Members of the genus *Monomorium* can be distinguished by the following characters: monomorphic to polymorphic; antennae 10–12 segmented (most frequently 12), usually with a conspicuous 3-segmented club; mandibles with 3–5 teeth, median clypeal seta conspicuous; median portion of clypeus raised, the raised section longitudinally bicarinate; the carinae usually distinct; metanotal groove present, commonly impressed; propodeal dorsum usually unarmed and rounding into the declivity; propodeal spiracle usually circular, located at about the midlength of the sclerite (Bolton 1987).

The *Monomorium monomorium*-group (Bolton 1987) can be distinguished by the following characters: Monomorphic; mandibles unsculptured; the masticatory margin usually with 4 teeth; palp formula predominantly 2,2; cephalic dorsum unsculptured and glossy smooth except for scattered hair-pits; eyes always present, size small to large (0.15–0.38×HW), with 4 or more ommatidia in the longest row; head always longer than broad (CI 72–89); metanotal groove moderately to strongly impressed, with distinct cross-ribs; propodeal dorsum rounding into declivity, not angulate or dentate; petiole, postpetiole and gaster usually unsculptured.

The first treatment of the Arabian *Monomorium* fauna was Collingwood's (1985) study of the genus in Saudi Arabia where 20 species were recorded, of which a single species was of the *monomorium*-group, *M. clavicorne* Andre, 1881. The second and more comprehensive contribution was that of Collingwood and Agosti (1996) for the *Monomorium* in the Arabian Peninsula. In that study, 53 species were recorded, 17 of which were described from Saudi Arabia including two species belonging to the *Monomorium monomorium*-group, *M. montanum* and *M. qarahe*. Since that time the contributions to the Arabian *Monomorium* fauna were descriptions of only two new species, *M. nimihil* Collingwood from Socotra Archipelago (Collingwood et al. 2004) and *M. moathi* Sharaf & Collingwood from Yemen (Aldawood et al. 2010).

In the present paper a new species of the genus *Monomorium*, *M. dryhimi* is described from Saudi Arabia based on worker caste. A key to the four Arabian species of the *Monomorium monomorium*-group is presented.

Materials and methods

All measurements are in millimeters and follow the standard measurements (Bolton 1987).

- **TL** Total Length; the outstretched length of the ant from the mandibular apex to the gastral apex.
- **HW** Head Width; the maximum width of the head behind eyes in full-face view.
- HL Head Length; the maximum length of the head, excluding the mandibles.
- **CI** Cephalic Index (HW × 100/HL).
- **SL** Scape Length, excluding basal neck.
- **SI** Scape Index (SL × 100/HW).
- **EL** Eye Length; the maximum diameter of the eye.
- **ML** Mesosoma Length; the length of the mesosoma in lateral view, from the point at which the pronotum meets the cervical shield to the posterior base of the propodeal lobes or teeth.
- **PRW** Pronotal width, maximum width in dorsal view.
- **PL** Petiole Length; the maximum length measured in dorsal view, from the anterior margin to the posterior margin.
- **PW** Petiole Width; maximum width measured in dorsal view.
- **PPL** Postpetiole Length; maximum length measured in dorsal view.
- **PPW** Postpetiole Width; maximum width measured in dorsal view.

Images taken under the scanning electron microscope ((SEM) JSM-6380 LA) were used to record morphological details of the new species (Figs 1–7).

Results

Monomorium dryhimi Aldawood & Sharaf, sp. n.

urn:lsid:zoobank.org:act:4C171A6D-B1F5-4D4D-BBAD-CE2250167E5B http://species-id.net/wiki/Monomorium_dryhimi Figs 1–7

Holotype worker. TL1.84, HL 0.48, HW 0.34, SL 0.31, ML 0.46, EL 0.08, PRW 0.22, PL 0.14, PW 0.11, PPL 0.08, PPW 0.11, SI 91, CI 71.

Paratypes. TL 1.42–1.84, HL 0.42–0.49, HW 0.32–0.36, SL 0.26–0.32, ML 0.39–0.46, EL 0.07–0.08, PRW 0.19–0.24, PL 0.09–0.14, PW 0.08–0.11, PPL 0.05–0.09, PPW 0.09–0.12, SI 74–91, CI 73–83.(N=13).

Holotype worker. SAUDI ARABIA, Al Bahah province, Amadan forest, Al Mandaq governorate, 20°12'N, 41°13'E, 1881 m.a.s.l. 19.V.2010 (*M. R. Sharaf & A. S. Aldawood Leg.*); King Saud Museum of Arthropods (KSMA), College of Food and Agriculture Sciences, King Saud University, Riyadh, Kingdom of Saudi Arabia.



Figures 1–7. *Monomorium dryhimi* sp.n., paratype worker **1**, **3** Head in full-face view **2** head in profile **4** body in profile **5** propodeum **6** propodeal spiracle **7** petiole and postpetiole.

Paratypes. 27 workers, same locality and data as holotype; 1 deposited in the Muséum d'Histoire Naturelle, Geneva, Switzerland (Dr Bernhard Merz); 1 in Naturhistorisches Museum, Basel, Switzerland (Mrs. Isabelle Zürcher-Pfander); 1 in California Academy of Science (Dr Brian Fisher); 1 in World Museum Liverpool, Liverpool, U.K (Dr Guy Knight), 1 in The Natural History Museum, London (Mr. Barry Bolton); 15 workers, SAUDI ARABIA, Elqamh park, Belgershi, Al Bahah, 17.V.2010 (*M. R. Sharaf & A. S. Aldawood Leg.*) These paratypes are in the King Saud Museum of Arthropods, King Saud University, Riyadh, Saudi Arabia.

Worker. Head distinctly much longer than broad with weakly convex sides and straight or feebly concave posterior margin (Fig. 1). Underside of head with several long hairs but not forming a psammophore (Fig. 2). Head in profile with a weakly convex dorsal surface and a distinctly convex ventral surface (Fig. 2). Clypeal carinae sharply developed and distinctly elevated, divergent anteriorly and reaching the anterior margin at a pair of short low triangular projecting angles (Fig. 3). The median portion of anterior clypeal margin clearly concave. Eyes oval and of moderate size (EL $0.19-0.25 \times HW$) with 6 ommatidia in the longest row (Figs 2, 3). With head in profile, eyes consist of a peripheral ring of ommatidia encircling two rows of ommatidia within the ring (Figs 2, 3). In lateral view, the maximum diameter of the eyes clearly greater than the distant between the anteriormost point of the eyes and the nearest point of the mandibular articulation. Frontal lobes farther apart. Antennal scapes, when laid straight back, fail to reach posterior margin (Fig. 1). Mesosoma in profile with a flat promesonotal dorsum, which slopes posteriorly to a well developed metanotal groove (Fig. 4). Metanotal crossribs relatively long and distinct (Fig. 5). Propodeal spiracles small and pinhole-like (Fig. 6). Propodeal dorsum evenly sloping, the posterior third more strongly sloping than the anterior two-thirds (Fig. 5). Petiole node high and narrowly subconical, narrowly rounded above (Fig. 7). Petiole peduncle short and stout with a distinct ventral process. Postpetiole node smaller, lower, and more broadly rounded than petiole. Petiole and postpetiole each with three pairs of long backward directed hairs. Body pilosity abundant, shorter on head dorsum. Anterior pronotal margin with two pairs of hairs, middle part of pronotum with a single pair, promesonotum with 3-4 pairs of hairs. Dorsum and declivity of propodeum each with one pair of hairs. Overall yellow to light brownish yellow. In many individuals head and gaster are slightly but conspicuously, darker than rest of body. Second halves of first and second gastral tergites with characteristic brownish transverse bands. Body smooth and shining.

Etymology. This species is named in honor of Prof. Yousif N. Aldryhim, economic entomologist, Department of Plant Protection, College of Food and Agriculture Sciences, King Saud University, Kingdom of Saudi Arabia.

Discussion

This new species is a member of the *Monomorium monomorium* group with closest resemblance to *Monomorium holothir* Bolton, 1987, which was described from Kenya.

Both species sharing the following characters: clypeal carinae sharply developed and distinctly elevated; head sides behind eyes weakly convex; posterior margin feebly concave; in lateral view the maximum diameter of eyes clearly greater than the distance between anteriormost point of the eye and the nearest point of the mandibular articulation; body colour yellow to light brownish yellow; relatively similar body dimensions e.g. HL, HW, SI, and CI.

Monomoroium dryhimi can be easily separated from M. holothir by the following characters: eyes relatively small, their maximum diameter EL 0.19–0.25 × HW and with 6 ommatidia in the longest row, while in holothir eyes larger, their maximum diameter EL 0.30 × HW and with 8–9 ommatidia in the longest row. In M. dryhimi, the median portion of anterior clypeal margin is clearly concave, whereas it is transverse to feebly concave in holothir. Moreover, in M. dryhimi head in profile with a weakly convex dorsal surface and a clearly convex ventral surface, whereas in M. holothir, head in profile dorsoventrally flattened. Furthermore, the promesonotum in M. dryhimi has 3–4 pairs of hairs whereas in holothir the promesonotum have 8 pairs of hairs.

Biology of Monomorium dryhimi

The type locality is a forest called Amadan, Al Mandaq governorate, Al Bahah province, Kingdom of Saudi Arabia, with much wild vegetation including *Erica arborea* L, *Juniperus procera* Hochst. Ex Endle.and *Acacia gerrardii* Benth (Fig. 8). *Monomorium dryhimi* type was taken from a nest under a stone on hard-packed soil which contained tens of workers and was found in relatively elevated area of a valley which is high enough to avoid direct impacts of flooding. No additional nests were found despite extensive surveys. In addition, we were not able to collect foraging workers near the nest. It appears that members of the *Monomorium monomorium* group may prefer inhabiting areas of high elevations. All the four Arabian species were found inhabiting elevated localities with more than 1800 m asl., except *M.clavicorne* which was also recorded from both relatively lower elevated areas including Riyadh and Al Qatif, in Central and Eastern regions of Saudi Arabia, respectively, and also from a much elevated area, Fayfa, Asir province (Collingwood 1985). Apparently this species is endemic to the chain of Asir Mountains which extends to Yemen.

Key to the Arabian species of the Monomorium monomorium-group

1	Antennae with 11 segments; terminal funicular segment broadly swollen
_	Antennae with 12 segments; terminal funicular segment enlarged, not
	swollen



Figure 8. Type locality, Amadan forest, Al Mandaq governorate, Al Bahah province, Kingdom of Saudi Arabia.

2	Head, in full-face view, with long hairs surrounding posterior margin and
	head sides forming a fringe; metanotal groove shallowqarabe
-	Head, in full-face view, without a fringe of long hairs; metanotal groove sharp
	and distinct
3	Larger yellow species; TL 1.70-2.30, HW 0.40; metanotal groove sharp but
	too small to break the dorsal outline; pronotum with a single pair of curved
	hairsmontanum
_	Smaller yellowish to light brownish yellow species, first and second gastral
	tergites with light brownish bands; TL 1.42-1.84; HW 0.32-0.36; metano-
	tal groove sharp and distinctly breaks the dorsal outline; anterior pronotal
	margin with two pairs of hairs, middle part of pronotum with a single pair
	<i>dryhimi</i> sp. n.

Acknowledgements

The authors thank Plant Protection Department and King Saud University for funding this work. The authors are grateful to Mr. Barry Bolton for valuable comments, Prof. Loutfy El-Juhany for identifying the plants in the type locality, Dr Magdi El Hawagry and Dr Ehab Moazz for assistance in the field, and Mr. Hassan Badri for technical assistance. The senior author thanks his wife Mrs. Amal El Saadany, Dr Hathal Alzhafer, Prof. Hoda Farid, Dr Mahmoud El Dera'a and Mr. Ismail Farid for thier continuous encouragements.

References

- Aldawood AS, Sharaf MR, Collingwood CA (2010) *Monomorium moathi* sp. n., a new ant species from Yemen related to the *salomonis*-group. Egyptian Academic Journal of Biological Sciences 3 (2): 37–42.
- Bolton B (1987) A review of the *Solenopsis* genus group and revision of Afrotropical *Monomorium*. Bulletin of the British Museum (Natural History) Entomology. 54: 263–452.
- Bolton B (1995) A new general catalogue of the ants of the World. Harvard University Press, Cambridge, Massachusetts, 504 pp.
- Bolton B, Alpert G, Ward PS, Naskrecki P (2007) Bolton's Catalogue of the Ants of the World: 1758–2005. Compact Disc Edition, Harvard University Press.
- Brown WL Jr (2000) Diversity of ants. In: Agosti D, Majer JD, Alonso LE, Schultz TR (Eds) Ants. Standard methods for measuring and monitoring biodiversity. Biological diversity Hand Book Series. Smithsonian Institution Press, Washington, D.C., 45–79.
- Collingwood CA (1985) Hymenoptera: Fam. Formicidae of Saudi Arabia. Fauna of Saudi Arabia 7: 230–301.
- Collingwood CA, Agosti D (1996) Formicidae of Saudi Arabia (part 2). Fauna of Saudi Arabia 15: 300–385.
- Collingwood CA, Pohl H, Güsten R, Wranik W, Van Harten A (2004) The ants of the Socotra Archipelago. Fauna of Arabia 20: 473–495.
- Fernández F (2007) Two new species of South American Monomorium Mayr with taxonomic notes on the genus (pp. 128–145). In: Snelling RR, Fisher BL, Ward PS (Eds) Advances in ant systematics: homage to E.O. Wilson – 50 years of contributions. Memoirs of the American Entomological Institute 80: 1–690.
- Heterick BE (2001) Revision of the Australian ants of the genus Monomorium. Invertebrate Taxonomy 15: 353–459. doi:10.1071/IT00003
- Heterick BE (2006) A revision of the Malagasy ants belonging to genus *Monomorium* Mayr, 1855. Proceedings of the California Academy of Sciences 57: 69–202.
- Mayr G (1855) Formicina austriaca. Beschreibung der bisher im osterreichischen Kaiserstaate aufgefunden Ameisen nebst Hinzufugung jener in Deutschland, in der Schweiz und in Italien vorkommenden Arten. Verhandlungen des zoologisch-botanischen Vereins in Wien 5: 273–478.

RESEARCH ARTICLE



New myrmecomorphous longhorned beetles from Haiti and the Dominican Republic with a key to Anaglyptini and Tillomorphini of Hispaniola (Coleoptera, Cerambycidae, Cerambycinae)

Steven W. Lingafelter

Systematic Entomology Laboratory, Plant Sciences Institute, Agriculture Research Service, U.S. Department of Agriculture, National Museum of Natural History, Washington, D.C. 20013-7012, U.S.A.

urn:lsid:zoobank.org:author:8CB85FB5-19CF-4357-81E3-FE9BFC9B04AA

Corresponding author: Steven W. Lingafelter (elaphidion@gmail.com)

Academic	editor:	Terry	Erwin		Received 2 May	2011		Accepted 20	May	2011		Published	15	June 2	2011
			urn:lsi	d:zo	obank.org:pub:68E0)A7A9-8	8A.	95-410D-8528	R-E9EF	6 <i>B7A4</i>	061	,			

Citation: Lingafelter SW (2011) New myrmecomorphous longhorned beetles from Haiti and the Dominican Republic with a key to Anaglyptini and Tillomorphini of Hispaniola (Coleoptera, Cerambycidae, Cerambycinae). ZooKeys 106: 55–75. doi: 10.3897/zookeys.106.1470

Abstract

First records of the tribes Anaglyptini and Tillomorphini (Coleoptera: Cerambycidae: Cerambycinae) are documented for Hispaniola. A new genus of a highly myrmecomorphic longhorned beetle (*Licracantha* **gen. n.**) is described and illustrated based on one species (*Licracantha formicaria* **sp. n.**) and provisionally assigned to Tillomorphini. Three other new species of ant mimic longhorned beetles are described and illustrated: *Calliclytus macoris* **sp. n.** (Tillomorphini), *Tilloclytus baoruco* **sp. n.**, and *Tilloclytus neiba* **sp. n.** (Anaglyptini). An identification key and distribution map to all known Hispaniolan species of these two tribes are presented.

Keywords

Island, endemic, mimicry, myrmecophily, West Indies, taxonomy

Introduction

Hispaniola is among the most critical biodiversity regions in the world and is part of the Caribbean Islands Hotspot (Conservation International and McGinley 2008). The entomological riches there were first discovered and described by Palisot de Beauvois

Copyright S.W. Lingafelter. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. (1805–1821). Despite periods of intense beetle collecting in the late 19th, early 20th, and early 21st centuries, many species of Hispaniolan Cerambycidae still remain undescribed. In the last 5 years, 20 new species representing about 12% of the known fauna have been described, demonstrating how poor our prior knowledge was (Lingafelter 2008, 2010; Lingafelter and Micheli 2009; Lingafelter and Nearns 2007; Lingafelter and Woodley 2007).

Batesian mimicry has been well documented in longhorned beetles and has been summarized in Shelford (1902) and Linsley (1959). The genera in the tribe Tillormorphini Pascoe [and some in Anaglyptini Lacordaire] are considered as group mimics of ants with generalized structural modifications (Linsley 1959). Morphological adaptations that suggest an antlike facies include a constriction of the elytra around the middle, accentuated with a light colored fascia or ivory colored callus (representing a petiole) and an inflated pronotum (representing a large ant head when combined with the smaller, actual head) (McIver and Stonedahl 1993).

Studies such as Fisher (1932), Micheli (2003), Chalumeau and Touroult (2005), and Micheli (2010) have documented the species diversity in the West Indies for Anaglyptini and Tillomorphini, nearly all of which display varying degrees of myrmecomorphy. Several species have been documented as collected with ants in Puerto Rico and Hispaniola, suggesting that their mimicry of, and association with ants may provide a selective protection (Micheli 2003; Micheli 2010; and herein).

Prior to this work, the tribes Anaglyptini and Tillomorphini were unknown for Hispaniola (Perez-Gelabert 2008; Monné and Bezark 2010) [note that the genus *Hormathus* Gahan was previously placed in Tillomorphini but transferred to Ibidionini by Lingafelter and Nearns 2007]. The authorship of Tillomorphini and Anaglyptini was corrected in Bousquet, et al. (2009): Tillomorphini was proposed by both Pascoe and Lacordaire in 1869, but Pascoe takes precedence since he published first; Anaglyptini was proposed by Lacordaire and the publication date was determined to be 1868 despite the 1869 date imprint on the title page.

In addition to the nomenclatural confusion, these tribes are very difficult to differentiate from each other (and Clytini) because they were never clearly defined and the currently recognized diversity in each tribe has escalated beyond their original, meager definitions. For example, the type genus of Anaglyptini, *Anaglyptus* Mulsant, has subsequently been considered a clytine (Gressitt 1951; Löbl and Smetana 2010), while some authors retained it as a separate tribe based on the presence of a mesal spine on antennomere 3 and lack of carinae or grooves on the frons (Bíly and Mehl, 1989). Adding to the confusion, the type species of Tillomorphini, *Tillomorpha lineoligera* Blanchard, has very coarsely faceted eyes and lacks a transverse ivory-like ridge or fascia on the elytra, unlike most of the species currently placed in that tribe (Lingafelter and Nearns 2007). Despite this, Pascoe (1869) used finely faceted eyes as the main feature to place Tillomorphini in the "second section" of Cerambycidae. Aurivillius (1912) listed *Tilloclytus* Bates and *Tillomorpha* Blanchard next to each other in the tribe Tillomorphini. However, Linsley (1964) placed *Tilloclytus* in the tribe Anaglyptini based on some characters he ascribed to Anaglyptini such as: head large; pronotum constricted at base; prosternum with intercoxal process narrow; and elytra gibbose at base and lacking transverse, ivory-like ridges. Zayas (1975) returned the six Cuban species of *Tilloclytus* to Tillomorphini (along with *Calliclytus* Fisher and *Pentanodes* Shaeffer), perhaps correctly so, but without explanation. The most current catalog of the Cerambycidae of the Neotropical Region (Monné 2005) followed Linsley (1964) and returned *Tilloclytus* to Anaglyptini, leaving all other West Indian tillomorphine genera mentioned by Chalumeau and Touroult (2005), Micheli (2010), and Zayas (1975) in the Tillomorphini. Thus, currently in the West Indies there is only one genus (*Tilloclytus*) in Anaglyptini and six genera (*Arawakia* Villiers, *Bonfilsia* Villiers, *Calliclytus* Fisher, *Gourbeyrella* Lane, *Lamproclytus* Fisher, and *Pentanodes* Schaeffer) in Tillomorphini (Monné and Bezark, 2010).

A spectacularly myrmecomorphic, monotypic new genus and new species (*Licra-cantha formicaria*) is described and provisionally placed in the tribe Tillomorphini. A new species of *Calliclytus* (*C. macoris*, sp. n.) is also described in this tribe. Two other species of Hispaniolan ant mimic Cerambycidae (*Tilloclytus baoruco*, sp. n. and *Tilloclytus neiba*, sp. n.) are described in the tribe Anaglyptini. Species in *Tilloclytus* and *Lamproclytus* are known to have significant intraspecific variation in color, and Micheli and Hovore (2003) observed this when recognizing several synonymies of Puerto Rican species described by Fisher (1932, 1935). While most of the new species described herein are based on one or a few specimens each, they possess structural features (in addition to color patterns) that are unique to each, further demonstrating their taxonomic status.

Methods

The material examined in this study was collected by Michael Ivie, Edmund Giesbert, Eugenio Nearns, Derek Sikes, Michael Thomas, Barry Valentine, and Norman Woodley. Holotypes are deposited in the Smithsonian Institution (USNM) and the Museum of Entomology at the Florida State Collection of Arthropods (FSCA). Holotype images in the USNM are available online in the Smithsonian Primary Type database (Lingafelter et al. 2011).

The species in this study are represented by one or a few specimens each. Many specimens are in imperfect condition; therefore, careful digital paintings were deemed preferable to show the beetles in natural, lifelike postures and to display the diagnostic characters. These paintings were produced by Taina Litwak (Systematic Entomology Laboratory, USDA [hereafter SEL]) using Photoshop in Adobe Creative Suite 4 on a G5 Macintosh with OS X.5.8.

Species determinations were aided by examination of material from many collections and type image websites. Those websites that were particularly useful, holding holotypes of related species, included: AMNH (2011) (which has the holotype of *Tilloclytus minutus* Fisher); MCZC (2011) (which has the holotype of *Tilloclytus rufipes* Fisher); and Lingafelter, et al. (2011) (which has the holotypes of *Tilloclytus* bruneri Fisher, *Tilloclytus cubae* Fisher, *Tilloclytus puertoricensis* Fisher, *Lamproclytus elegans* Fisher, *Lamproclytus oakleyi* Fisher, and *Calliclytus schwarzi* Fisher). The paper on the Cuban Cerambycidae of the Zayas collection by Nearns et al. (2006) was very useful since it provided photographs of the holotypes of *Tilloclytus elongatus* Zayas and *Tilloclytus pilosus* Zayas, leading to the discovery of a synonomy (Lingafelter and Nearns, in press).

Morphological terminology follows Lingafelter (1998). Measurements were made using Axiovision software on images taken with a Zeiss AxioCam HRc camera attached to a Zeiss Discovery V20 stereomicroscope with Sycop motorized zoom and focus control and a PlanApo S 1.0X objective.

Collection acronyms used in this study include:

ACMT	American Coleoptera Museum, San Antonio, Texas, U.S.A. (J. Wappes)
EFGC	Edmund F. Giesbert Collection (at FSCA), Gainesville, Florida, U.S.A. (M.
	Thomas, P. Skelley)
FSCA	Florida State Collection of Arthropods, Gainesville, Florida, U.S.A. (M.
	Thomas, P. Skelley)
USNM	National Museum of Natural History, Smithsonian Institution, Washing-
	ton, DC, U.S.A. (S. Lingafelter)
WIBF	West Indian Beetle Fauna Project, Bozeman, Montana, U.S.A. (M. Ivie)

Results and discussion

Licracantha Lingafelter gen. n.

urn:lsid:zoobank.org:act:923ACE0A-618A-47F6-A830-93E5F8F9C75B http://species-id.net/wiki/Licracantha Figs 1–2

Diagnosis. No other genus of Tillomorphini or Anaglyptini has the type of modified antenna, pronotum, and elytron as is present in *Licracantha*. The combination of the following character states is unique to *Licracantha*: antenna myrmecomorphic, 11-segmented, with elongate scape, antennomeres 3–5 with pronounced mesal spines, antennomeres 6–11 abruptly shortened, antennomeres 3–11 articulated in a potentially opposing direction from scape; pronotum highly and abruptly elevated at anterior four-fifths, with acute, suprascutellar projection posteriorly; eye finely faceted and as single lobe anteroventral to antennal insertion, elytra gibbose basally and apically with depression at oblique, unraised, ivory fascia; tibiae each with single, curved spine (most pronounced on meso- and metatibia).

Gourbeyrella, Tillomorpha, Bonfilsia, Arawakia, Pentanodes, and Tilloclytus each lack antennal spines. Further, Bonfilsia, Arawakia, Lamproclytus, and presumably, Calliclytus



Map 1. Distribution of ant-mimic longhorned beetles of tribes Tillomorphini and Anaglyptini in Hispaniola.

each have 10-segmented antennae. Calliclytus and Lamproclytus are further differentiated since each have upper eye lobes (along with the larger, lower lobe), a very short scape, and a pronotum that is not elevated anteriorly. A few species in the relatively large, heterogeneous genus *Tilloclytus* are most similar to *Licracantha* in having a moderately, anteriorly elevated pronotum, a single finely faceted eye lobe, and an elytron moderately gibbous at base and apex and with some type of pale transverse or oblique fascia near mid length. Tilloclytus minutus Fisher has the most similar antenna possessing an elongate scape and very short antennomeres 6-11; however, the myrmecomorphic modifications are not as extreme: the scape is shorter, extending only to the anterior third of the pronotum, the abrupt articulation allowing an opposing orientation of remaining antennomeres from 3–11 is not present, and antennomeres 3–5 are, at most, dentiform mesally. *Tilloclytus minutus* is further differentiated by having the elytron and pronotum uniformly, confluently alveolate-punctate, lacking a narrow, well defined pale elytral fascia, having the pronotum not abruptly elevated anteriorly, and in lacking a well developed gibbosity on the elytral base and apex. Tilloclytus bruneri Fisher is similar in having a posterior suprascutellar pronotal projection, but it is not as developed or acute as in Licracantha. Tilloclytus bruneri is further differentiated by having a longer, unmodified, unspined antenna and a pronotal elevation and elytral gibbosity that are less developed. It also has a glossy integument lacking micropunctation.

Type species. This genus is known only from *Licracantha formicaria* Lingafelter, described below.

Etymology. A latinized composite noun, female gender, derived from the Greek "Likros" meaning horn and the Greek "Akantha" meaning thorn. *Licracantha* refers to the pronounced spines on the antennae.

Remarks. This new genus is provisionally assigned to the tribe Tillomorphini. The definitions and boundaries of Anaglyptini and Tillomorphini are vague and trouble-

some, as discussed above, and each may contain a polyphyletic assemblage of taxa. A phylogenetic analysis of all the genera in these tribes is needed to develop a meaningful classification. Once those studies are completed, they may show that the genus *Tillocly-tus*, to which *Licracantha* shows some similarities, should be returned to Tillomorphini from Anaglyptini, in which case Anaglyptini would not be present in the Caribbean Region.

Licracantha formicaria Lingafelter sp. n. urn:lsid:zoobank.org:act:2C031E71-45D5-4FD7-9514-A28A8803D8DC http://species-id.net/wiki/Licracantha_formicaria Figs 1–2; Map 1

Diagnosis. The single known species is recognized by the modified myrmecomorphic antennae with strong mesal spines on antennomeres 3–5, the suprascutellar projection of the pronotum, the matte integument with micropunctation throughout the dorsal surface, the dense, white pubescence on the sides of the meso- and metasternum, the purple hue on the apical two-thirds of the elytron, and the glossy integument ventrally and on the elytral epipleuron.

Description. Male. 4.86 mm long; 1.37 mm wide at humeri. Color: Integument of head, pronotum, elytron, and legs mostly dark reddish brown with purple hue on apical two-thirds of elytron and legs; antenna dark reddish brown except for base of scape and antennomeres 3–5 which are lighter orange. Head: Matte, with micropunctation throughout and indistinct larger depressions; setae sparse, short, golden; gena short, produced anteriorly into acute tooth near base of mandible; frons short, broad, without evident frontal-genal ridge, anteclypeal sulcus, or interantennal groove or depression; eye with single small lobe present anteroventral to antennal tubercle; laterally as protuberant as pronotum; finely faceted; antennal tubercle weakly, gradually elevated; antenna 11-segmented, short, extending to middle of elytron; highly myrmecomorphic with elongate scape extending to nearly middle of pronotum; antennomere 2 short, angled, causing remaining antennomeres to be potentially articulated in opposing direction from scape; antennomeres 3-5 spinose apicomesally, lighter than remainder; antennomeres 6-11 abruptly shortened, shorter than scape; antennomeres 1-5 with sparse, elongate, golden setae; 6-11 with denser, appressed golden setae. Mandible yellow with piceous apex; terminal palpomeres broadly dilated. Pronotum: Matte, with uniform micropunctation throughout and interspersed, separate, shallow punctures; without calli or tubercles; distinctly longer than broad, 1.61 mm long, 1.04 mm wide (length/width = 1.55); anterior four-fifths abruptly elevated above constricted posterior fifth; base distinctly narrower than elytron; posteriorly produced into suprascutellar process at middle; sparsely pubescent with scattered, erect and appressed, short, golden setae, more dense at posterior margin of elevation. Prosternum: Densely, uniformly micropunctate, glabrous; prosternal process very narrow between procoxae; broadly expanded behind, closing procoxal cavities posteriorly; highly impressed



Figure 1. *Licracantha formicaria* sp. n., dorsal habitus. Digital painting by Taina Litwak.



Figure 2. Licracantha formicaria sp. n., lateral habitus. Digital painting by Taina Litwak.

anterior to procoxae; strongly downwardly curved anteriorly. Elytron: Matte, except for glossy, anterior epipleural region; uniformly micropunctate throughout; scattered, sparse, appressed, golden setae, mostly concentrated on basal third; dark reddish brown to piceous at basal third, separated from lighter purplish brown apical two thirds with oblique pale fascia that does not extend to suture; constricted and depressed near white fascia, gibbous at base and apex; elytral apex rounded to suture; 2.79 mm long, 0.67 mm wide (length/width = 4.16). Scutellum: Small, rounded posteriorly; short, golden setae present on middle. Legs: Femora short, stout, with strongly clavate apices on abruptly narrowed peduncles; metafemur not attaining elytral apex; tibiae straight, apically expanded, each with single, strong, curved tibial spine; sparsely pubescent with scattered golden setae. Venter: Mostly glossy, sparsely pubescent except on sides of posterior margin of metasternum and mesosternum which have dense, white pubescence, the former coinciding with white fascia of elytron; mesosternal intercoxal process about 3 times as broad as prosternal process, with small lateral projection into mesocoxa and middle notch receiving anterior projection of metasternum. Ventrite 1 most elongate; remaining ventrites successively shorter and elevated towards elytral apex; apex of fifth ventrite broadly rounded, without notch, sulcus, or other modification.

Etymology. The specific epithet, *formicaria*, is a Latin adjective referring to the very antlike facies.

Type material. Holotype, male: Dominican Republic, La Vega Prov., 4.7 km SE Jarabacoa, July 16, 1996, M. C. Thomas, collector (FSCA).

Remarks. Only a single specimen is known of this monotypic genus. It is similar to arboreal ants of the genus *Cephalotes* Latreille, and they may be the model for this myrmecomorphic longhorn. These distinctive ants with large heads and spines on the thorax that resemble the suprascutellar process of the beetle, are slow moving and frequently beaten from vegetation in the Dominican Republic where this specimen was collected (Lingafelter pers. obs). There are six species of these ants known from Hispaniola (Perez 2008).

Calliclytus macoris Lingafelter, sp. n.

urn:lsid:zoobank.org:act:CD9CE24D-72FB-4858-A16F-5D6DBDC55ACA http://species-id.net/wiki/Calliclytus_macoris Fig. 3; Map 1

Diagnosis. This species is very similar to the Cuban *Calliclytus schwarzi* Fisher with regard to proportions, shapes of anatomical structures, position of the antemedial, raised, ivory callus of the elytron, and hypothesized presence of only 10 antennomeres (the holotype of *C. schwarzi* is missing the terminal segment of both antennae; however, since the antennal proportions are similar to those of *C. macoris*, it presumably has only 10 antennomeres). *Calliclytus macoris* differs from *C. schwarzi* in having an alveolate-punctate pronotum (rugose in *C. schwarzi*), a diamond shaped, pale macula

at suture near elytral apex (elytral apex all black in *C. schwarzi*), and a densely pubescent scutellum (glabrous in *C. schwarzi*).

Calliclytus macoris is also similar to the Puerto Rican *Lamproclytus elegans* Fisher with regard to proportions and shapes of the major anatomical structures but differs in having the raised eburneous ridge of the elytron antemedially located (postmedially positioned in *L. elegans*), the diamond shaped, pale macula at the elytral apex (uniformly dark in *L. elegans*), and uniformly dark legs (femora pale at the base and dark at the apex in *L. elegans*).

Description. Female. 4.85 mm long; 1.26 mm wide at humeri. Color: Dorsal integument of head, pronotum, and legs dark reddish brown to black; elytral color dark reddish brown to black on periscutellar region and most of the apical two-thirds with the exception of orange, diamond shaped macula at apex and raised, transverse ivory callus antemedially; ventral color mostly dark reddish brown to black except for orange head. Head: Shiny, rugose-punctate throughout; moderately dense, long and short erect and suberect, yellow-translucent setae; frons and gena short, broad, without acute projection near base of mandible; with poorly defined frontal-genal ridge; without anteclypeal sulcus; without interantennal groove or depression; eye divided into two lobes connected by row of 5 facets, with large lobe anteroventral positioned to antennal tubercle and small lobe present posterodorsal to antennal tubercle; laterally not as protuberant as pronotum; finely faceted; antennal tubercle weakly elevated; antenna 10-segmented, without spines, short, extending to just beyond base of elytron; scape short, thickened at middle, extending just past anterior margin of pronotum; antennomere 2 short, but over half length of antennomere 3; antennomeres 2–5 successively increasing in length, 6–10 successively shorter, produced apicolaterally; scape through antennomere 5 pale orange; 6-10 piceous to black; antennomeres 1-5 with sparse, elongate, golden setae; 6-10 with denser, appressed golden setae along with sparse, erect setae. Mandible short, retracted, yellow with piceous apex; terminal palpomeres not broadly dilated. Pronotum: Semiglossy, with uniform alveolate punctation dorsally, becoming punctate at sides; without calli or tubercles; slightly longer than broad, 1.47 mm long, 1.27 mm wide (length/width = 1.16); evenly widened at middle; gradually rounded laterally and dorsally; base with constriction; distinctly narrower than elytra; sparsely but conspicuously pubescent with scattered, long, erect yellowish setae combined with shorter, appressed yellow setae. Prosternum: Glossy, with dense microsculpture and short, white, setae in front of procoxae; prosternal process narrow between procoxae; apex broadly expanded behind, closing procoxal cavities posteriorly. Elytron: Glossy; deeply, separately punctate throughout, becoming slightly more dense posteriorly; sparse but conspicous, erect, yellow setae throughout; dark reddish brown to black on periscutellar region and most of the apical two-thirds with exception of orange, diamond shaped macula at apex; transverse, raised, eburneous callus present, not extending to suture; weakly gibbous at periscutellar region only; elytral apex rounded to suture; 2.96 mm long, 0.62 mm wide (length/width = 4.77). Scutellum: Elongate, subtruncate at posterior apex; densely coated with appressed, short, yellowish setae. Legs: Femora short, stout, with strongly clavate apices on abruptly



Figure 3. Calliclytus macoris sp. n., dorsal habitus. Digital painting by Taina Litwak.

narrowed peduncles; metafemur not attaining elytral apex; tibiae straight, not expanded apically, each with two straight tibial spines; tibiae and femora sparsely but conspicuously pubescent with long, erect, white setae. **Venter**: Glossy; sparsely pubescent throughout with erect, long, white setae and dense, short, white setae on metasternum posterior and lateral margin, mesosternum, and prosternum; integument darker than most of dorsum; mesosternal intercoxal process about 2.2 times as broad as prosternal process, without lateral projection into mesocoxa. Ventrite 1 most elongate; remaining ventrites much shorter and subequal in length; apex of fifth ventrite broadly rounded, without notch, sulcus, or other modification.

Etymology. The specific epithet is based on the nearby Macorís River where this species was discovered by Edmund Giesbert.

Type material. Holotype, female: Dominican Republic, San Pedro de Macorís Prov., 12 km W San Pedro de Macorís, May 5–19, 1985, E. Giesbert, collector (EFGC in FSCA).

Remarks. The genera *Lamproclytus* and *Calliclytus* were not specifically compared to each other in Fisher's (1932) descriptions, despite their obvious similarities in nearly every feature. Careful phylogenetic work in Tillomorphini may suggest that synonymy of these genera is necessary, but that is beyond the scope of this work. Given that the position of the eburneous elytral ridge of *C. macoris* is closest to that of *C. schwarzi*, it is placed in that genus as opposed to *Lamproclytus*. Note that Monné (2005) and Monné and Bezark (2010) erroneously listed *Lamproclytus elegans* Fisher from the Dominican Republic, but that species does not occur in Hispaniola.

This new species is superficially similar to ants of the genus *Leptothorax* Mayr, which may serve as the mimicry model. In Puerto Rico, the similarly colored cerambycid, *Boricyrtinus nilseni* Micheli, was collected with *Leptothorax isabellae* (Wheeler) (Micheli 2003). There are seven species of these ants known from Hispaniola (Perez 2008).

Tilloclytus baoruco Lingafelter, sp. n.

urn:lsid:zoobank.org:act:8B4B5147-A091-4137-88FE-1D0EBC4CAC1E http://species-id.net/wiki/Tilloclytus_baoruco Fig. 4; Map 1

Diagnosis. This species is unique among West Indian *Tilloclytus* in having only 10 antennomeres. It is otherwise most similar to *Tilloclytus bruneri* Fisher from Cuba in that the antemedial pale band of short, appressed pubescence is incomplete, not reaching the suture, but they are easily differentiated by color: *T. baoruco* is mostly light brown to orange, while *T. bruneri* is darker bluish black.

Description. 3.34–4.67 mm long; 0.85–1.12 mm wide at humeri. **Color:** Dorsal integument of head, pronotum, elytra, antenna, and legs various shades of light brown to orange; head and pronotum lighter than remainder; elytral color interrupted by antemedial transverse, white, microstriate, unelevated fascia that does not reach suture; ventral color mostly light brown to orange except for sternites which are brown with very dark brown



Figure 4. *Tilloclytus baoruco* sp. n., dorsal habitus. Digital painting by Taina Litwak.

posterior margins. Head: Semi-matte, microsculptured but impunctate throughout; inconspicuous, sparse, translucent long and short erect and suberect setae; frons and gena short, broad, with short, acute projection near base of mandible; with incomplete frontalgenal ridge; without anteclypeal sulcus; without interantennal groove or depression; single eye lobe anteroventrally positioned to antennal tubercle; laterally nearly as protuberant as pronotum; finely faceted; antennal tubercle moderately elevated; antenna 10-segmented, without spines, short, extending to apical third of elytron; scape long, slender, extending beyond anterior fourth of pronotum; antennomere 2 short, but over one-third length of antennomere 3; antennomere 4 distinctly shorter than 3 and 5, 6-10 successively shorter, decreasing in length, produced apicolaterally; antennomeres dark brown with exception of scape which may have light brown base; sparse, elongate, suberect and appressed, white setae throughout. Mandible moderately produced, yellow with piceous apex; terminal palpomeres elongate, not broadly dilated in female; broadly dilated and securiform in male. Pronotum: Matte, with uniform ultra-microrugosity throughout, impunctate, without calli or tubercles; distinctly longer than broad, 1.15-1.24 mm long, 0.65-0.88 mm wide (length/width = 1.41 - 1.76); strongly constricted at basal fourth, elevated and widest anteriorly, base distinctly narrower than elytral base; distinct, rounded periscutellar projection at middle; sparsely but conspicuously pubescent with scattered, long, erect translucent to white setae. Prosternum: Glossy, impunctate, with sparse, elongate, white setae; prosternal process very narrow between procoxae; apex broadly expanded behind, closing procoxal cavities posteriorly. Elytron: Mostly glossy; impunctate (but with scattered, dark, subcuticular spots resembling punctures but not depressed on surface); microruguse at basal third, with unelevated antemedial, transverse, white, microstriate fascia not attaining suture; oblique, ultra-micropunctate region adjacent and posterior to white fascia; remainder of elytron to apex glossy; scattered, long, translucent setae sparsely distributed throughout; light brown throughout with exception of white fascia which is surrounded by darker brown on both sides, extreme base, and periscutellar regions which are darker brown; weakly gibbous at apex; elytral apex narrowly rounded to suture; 1.97-2.64 mm long, 0.40–0.55 mm wide (length/width = 4.80–4.93). Scutellum: Narrow, subtruncate at posterior apex; sparsely coated with appressed, short, yellowish setae. Legs: Femora short, stout, with strongly clavate apices on abruptly narrowed peduncles; metafemur not attaining elytral apex; tibiae straight, not expanded apically; meso- and metatibiae each with two asymmetrical, straight tibial spines; protibia with one; tibiae and femora sparsely but conspicuously pubescent with long, erect, white setae. Venter: Glossy; sparsely pubescent throughout with erect, long, white setae; dense, white, short, appressed setae present on posterior margin of metasternum to sides, corresponding with white macula of elytron, and along side of mesosternum; integument light brown, but darker on abdominal sternites; mesosternal intercoxal process narrow, but about twice as broad as prosternal process, with strong lateral projection into mesocoxa. Ventrite 1 most elongate; remaining ventrites much shorter and subequal in length; apex of fifth ventrite broadly rounded, without notch, sulcus, or other modification.

Etymology. The specific epithet, a noun in apposition, is based on the mountain range, Sierra de Baoruco, where the holotype was collected.

Type material. Holotype, female: Dominican Republic, Pedernales Prov., Parque Nacional Sierra de Baoruco, Las Abejas, 1150m, beating, E. H. Nearns and S. W. Lin-gafelter, June 18, 2005 (USNM). Paratypes: Haiti, Dept. Sud-Oueste, Parc National La Visite, ca. 1 km. S Roche Plat, May 22, 1984, M. C. Thomas, collector (FSCA, 1 male); Haiti, Dept. Sud-Oueste, Parc National La Visite, vicinity park headquarters, 1880 m, May 23, 1984, M. C. Thomas, collector (FSCA, 1 male); Haiti, Dept. Ouest, Furcy, July 9, 1956, B. and B. Valentine, collectors (USNM, 1 male, 1 female; ACMT, 2 females).

Remarks. This species is sexually dimorphic with respect to the terminal labial and maxillary palpi: in males, they are strongly dilated, nearly securiform; in females, weakly dilated and more elongate. One specimen was collected with an undetermined species of ant in the genus *Pheidole* Westwood.

Tilloclytus neiba Lingafelter, sp. n.

urn:lsid:zoobank.org:act:461DD063-89B4-46D3-812A-30C208D74C8B http://species-id.net/wiki/Tilloclytus_neiba Fig. 5; Map 1

Diagnosis. This species is similar to *Tilloclytus rufipes* Fisher from Cuba in proportions, color, and in having the antemedial elytral fascia extend completely to the suture, but in *T. neiba* the white elytral fascia is striate and without pubescence; in *T. rufipes*, the white fascia is a band of pubescence. *Tilloclytus neiba* also differs from *T. rufipes* in having very short, white pubescence covering the entire base of the elytron giving it a matte finish (in *T. rufipes*, this portion of the elytron is glossy and mostly free of appressed pubescence). From the Hispaniolan congener, *T. baoruco*, *T. neiba* differs most distinctly by having 11 antennomeres (10 in *T. baoruco*).

Description. Male. 4.08–4.23 mm long; 1.01–1.25 mm wide at humeri. **Color:** Most of dorsal integument of head, pronotum, and elytra dark brown to golden brown; antenna, legs, mesosternum and sometimes metasternum, prosternum, and base of pronotum, light orange; elytral color interrupted by antemedial transverse, white, microstriate, unelevated fascia that reaches suture. **Head:** Semi-matte, microsculptured but impunctate throughout; covered with moderately dense mixture of short, semi-appressed and long, erect, translucent and golden setae; frons and gena short, broad, with short, acute projection near base of mandible; with incomplete frontal-genal ridge; without anteclypeal sulcus; without interantennal groove or depression; large, single eye lobe anteroventrally positioned to antennal tubercle; laterally as protuberant as pronotum; finely faceted; antennal tubercle moderately elevated; antenna 11-segmented, without spines, short, extending to apical third of elytron; scape long, slender, extending beyond anterior third of pronotum; antennomere 2 short, less than one-third length of antennomere 3; antennomere 4 distinctly shorter than 3 and 5, 6–10 successively shorter, decreasing in length, not produced apicolaterally; antennomeres orange to light (sometimes with 9–11



Figure 5. Tilloclytus neiba sp. n., dorsal habitus. Digital painting by Taina Litwak.

dark brown); sparse, elongate, suberect and appressed, yellow-translucent setae throughout. Mandible moderately produced, light brown with piceous apex; terminal palpomeres broadly dilated. Pronotum: Matte except for glossy posterior fifth and sides, with striate microsculpturing over most of disk; impunctate, without calli or tubercles; distinctly longer than broad, 1.23–1.30 mm long, 0.81–0.84 mm wide (length/width = 1.52–1.55); strongly constricted at basal fourth, elevated and widest anteriorly, base distinctly narrower than elytral base; without periscutellar projection at middle; moderately dense, appressed white to translucent pubescence, especially at anterior third and along posterior constriction, combined with scattered, sparse, long, erect white or translucent setae. Prosternum: Glossy, impunctate, with sparse, elongate, translucent setae; prosternal process narrow between procoxae; apex broadly expanded behind, closing procoxal cavities posteriorly; dark brown anteriorly to pale orange posteriorly near procoxae, or uniformly pale orange. Elytron: Mostly glossy and impunctate (but with scattered, dark, subcuticular spots resembling punctures but not depressed on surface); inconspicuously micropunctate at basal third with moderately dense patch of yellow-white and translucent, appressed setae combined with more sparse, long, erect setae; with unelevated antemedial, transverse, white, microstriate fascia attaining suture; apical two-thirds mostly covered by patch of moderately dense, short, yellow-white, appressed setae with interspersed long, erect setae; dark brown to light, golden brown throughout with exception of white fascia; weakly gibbous at apex; elytral apex rounded to suture; 2.52-2.56 mm long, 0.51-0.59 mm wide (length/width = 4.33-4.94). Scutellum: Broad, short, rounded at posterior apex; moderately coated with appressed, short, yellow-white setae. Legs: Femora short, stout, with strongly clavate apices on abruptly narrowed peduncles; metafemur not attaining elytral apex; tibiae straight, not expanded apically; meso- and metatibiae each with two asymmetrical, straight tibial spines; protibia with one broad, curved spine; tibiae and femora sparsely pubescent with long, erect, off-white setae. Venter: Glossy; sparsely pubescent with inconspicuous, erect, translucent setae; dense white, short, appressed setae present on posterior margin of metasternum to sides, corresponding with white macula of elytron, and along side of mesosternum; integument light brown to orange on mesosternum and sometimes metasternum; dark brown on sternites; mesosternal intercoxal process narrow, but about twice as broad as prosternal process, with strong lateral projection into mesocoxa. Ventrite 1 most elongate; remaining ventrites much shorter and subequal in length; apex of fifth ventrite broadly rounded, without notch, sulcus, or other modification.

Etymology. The specific epithet, a noun in apposition, is based on the mountain range, Sierra de Neiba, where the holotype was collected.

Type material. Holotype, male: Dominican Republic, San Juán Prov., Sierra de Neiba, trail to Sabana del Silencio, 10 km SSW of El Cercado, 1650-1700m, 18°39.935'N, 71°31.964'W, July 10-11, 2006, N. E. Woodley, collector, sweeping foliage (USNM). Paratype: Dominican Republic, La Vega Prov., Jarabacoa – El Rio Rd., 910 m, April 11, 1992, M. Ivie, D. Sikes, and W. Lanier, collectors (WIBF, 1 male).

Remarks. Only males are known. Although this species was not collected with ants, *Pheidole* could serve as the model given its similarity to *T. baoruco* which was collected with that genus of ant.

Key to Anaglyptini and Tillomorphini of Hispaniola

There are no worldwide keys to tribes or genera that include all Old and New World taxa of Anaglyptini and Tillomorphini. Linsley (1962) provided a key to the Cerambycinae tribes of North America, but it excluded most of the genera in these groups that occur in the Neotropical Region. The characters that he used to distinguish the tribes (Anaglyptini: "without transverse, ivory-like ridges"; Tillomorphini: "usually with transverse raised ivory-like ridges") are unsatisfactory (see Introduction). No keys exist to all the species in the West Indies. A key to the three genera of Tillomorphini from the Lesser Antilles is provided in Chalumeau and Touroult (2005). A key to all the Cuban species of Tillomorphini + Anaglyptini is provided by Zayas (1975). In Puerto Rico, only one species in each of the tribes Anaglyptini (*Tilloclytus minutus* Fisher) and Tillomorphini (*Lamproclytus elegans* Fisher) is known, so no keys to the species of those genera were necessary (Micheli 2010). The key below combines all the new species described herein and includes both tribes since there are no satisfactory characters to differentiate them.

1	Antenna with prominent mesal spines on antennomeres 3–5
_	Antenna without spines
2(1)	Antenna very short, extending only to extreme base of elytron; elytron with
	raised, ivory callus; pronotum uniformly alveolate-punctate
_	Antenna longer, reaching beyond middle of elytron; elytron with unraised,
	white fascia; pronotum without distinct punctures
3(2)	Antenna 10-segmented; elytral fascia not attaining suture
_	Antenna 11-segmented; elytral fascia attaining suture

Acknowledgments

I am grateful to the curators and caretakers of the collections above, especially Michael Ivie (WIBF), Michael Thomas (FSCA), Paul Skelley (FSCA), James Wappes (ACMT), and the late Edmund Giesbert (EFGC). For my expeditions to the Dominican Republic in 2005 and 2006 when some of the specimens were collected, I thank Kelvin Guerrero (Santo Domingo Este) for his aid in logistics and obtaining permits. I thank the Subsecretaria de Áreas Protegidas y Biodiversidad de la Secretaría de Estado de Medio Ambiente y Recursos Naturales, Dominican Republic for providing the collecting and export permits (2005 collecting permit #0876 and export permit #0403; 2006 collecting permit #01140 and export permit #0953). I thank Taina Litwak (SEL) for the beautiful digital paintings of the new species. I thank Norman Woodley (SEL)
and Eugenio Nearns (University of New Mexico) for their camaraderie on the expeditions and for collecting some of the specimens. I appreciate the consultation of Ted Schultz, Jeffrey Sosa-Calvo, and Christian Rabeling (Smithsonian Institution) who helped identify associated ants and suggest potential models based on the morphology of the cerambycids. I thank Jens Prena (SEL) for guidance on Greek and Latin name formation and Charyn Micheli (Smithsonian Institution), Robert Kula (SEL), Allen Norrbom (SEL), Julio Micheli, and James Wappes for their reviews of this paper. USDA is an equal opportunity provider and employer.

References

- American Museum of Natural History [AMNH] (2011) American Museum of Natural History Online Type Database. http://www.research.amnh.org/invertzoo/types_db/ [last accessed February 11, 2011]
- Aurivillius C (1912) Cerambycidae: Cerambycinae. In: Schenkling S (Ed) Coleopterorum Catalogus, auspicis et auxilio W. Junk, Berlin, Pars 39, 574 pp.
- Bousquet Y, Heffern DJ, Bouchard P, Nearns EH (2009) Catalogue of family-group names in Cerambycidae (Coleoptera). Zootaxa 2321: 1–80.
- Bíly S, Mehl O (1989) Longhorn Beetles (Coleoptera, Cerambycidae) of Fennoscandia and Denmark. Fauna Entomologica Scandinavika 22. Brill, New York, 203 pp.
- Chalumeau F, Touroult J (2005) Les Longicornes des Petites Antilles (Coleoptera, Cerambycidae) Taxonomie, Ethologie, Biogeographie. Pensoft Series Faunistica 51, 241 pp.
- Conservation International, McGinley M (2007) "Biological diversity in the Caribbean Islands." In: Encyclopedia of Earth. Cutler J. Cleveland (Ed) (Washington, D.C.: Environmental Information Coalition, National Council for Science and the Environment). http://www. eoearth.org/article/Biological_diversity_in_the_Caribbean_Islands [last revised: August 22, 2008; last accessed February 10, 2011].
- Fisher WS (1932) New West Indian cerambycid beetles. Proceedings of the United States National Museum 80(2922) Art. 22: 1–93.
- Fisher WS (1935) New cerambycid beetles from Puerto Rico. The Journal of Agriculture of the University of Puerto Rico 19(2): 51–63.
- Gressitt JL (1951) Longicorn beetles of China. Longicornia, Etudes et notes sur les longicornes, 2: Lechevalier, Paris, 667 pp + 22 plates.
- Lacordaire JT (1868) Histoire naturelle des insectes. Genera des Coléoptères ou exposé méthodique et critique de tous les genres proposés jusqu'ici dans cet ordre d'insectes. Tome huitième. Librairie Encyclopédique de Roret, Paris, 552 pp.
- Lacordaire JT (1869) Histoire naturelle des insectes. Genera des Coléoptères ou exposé méthodique et critique de tous les genres proposés jusqu'ici dans cet ordre d'insectes. Tome neuvième. Première partie. Librairie Encyclopédique de Roret, Paris, 409 pp.
- Lingafelter SW (1998) The genera of Elaphidiini Thomson (Coleoptera: Cerambycidae). Memoirs of the Washington Entomological Society No. 20: 1–118.

- Lingafelter SW (2008) Seven new species of Elaphidiini (Coleoptera: Cerambycidae) from the Dominican Republic with taxonomic notes, new country records, and a key to *Elaphidion* Audinet-Serville from Hispaniola. The Coleopterists Bulletin 62(3): 353–379. doi:10.1649/1110.1
- Lingafelter SW (2010) Methiini and Oemini (Coleoptera: Cerambycidae: Cerambycinae) of Hispaniola. The Coleopterists Bulletin 64(3): 265–269. doi:10.1649/0010-065X-64.3.265.14
- Lingafelter SW, Micheli CJ (2009) The genus *Leptostylopsis* of Hispaniola (Coleoptera, Cerambycidae, Acanthocinini). ZooKeys 17: 1–55. doi:10.3897/zookeys.17.217
- Lingafelter SW, Nearns EH (2007) Five new species of longhorned beetles (Coleoptera: Cerambycidae) from the Dominican Republic in genera *Ataxia* Haldeman, *Atimiola* Bates, *Drycothaea* Thomson, *Eburia* Lepeletier & Audinet-Serville, and *Hormathus* Gahan. The Coleopterists Bulletin 61(2): 177–191. doi:10.1649/0010-065X(2007)61[177:FNSOLB] 2.0.CO;2
- Lingafelter SW, Nearns EH (in press) New synonymy in Cuban Tilloclytus Bates (Coleoptera: Cerambycidae: Anaglyptini). The Coleopterists Bulletin 65(2).
- Lingafelter SW, Woodley NE (2007) A new species of *Derancistrus* Audinet-Serville (Coleoptera: Cerambycidae: Prioninae) from the Dominican Republic with notes on other species of Prioninae from Hispaniola. The Coleopterists Bulletin 61(2): 165–175. doi:10.1649/0010-065X(2007)61[165:ANSODA]2.0.CO;2
- Lingafelter SW, Monné MA, Nearns EH (2011) Online Image Database of Cerambycidae Primary Types of the Smithsonian Institution. Available from: http://www.elaphidion.com/ [last accessed on March 25, 2011]
- Linsley EG (1959) Mimetic form and coloration in the Cerambycidae (Coleoptera). Annals of the Entomological Society of America 52(2): 125–131.
- Linsley EG (1962) The Cerambycidae of North America. Part III. Taxonomy and classification of the subfamily Cerambycinae, Tribes Opsimini through Megaderini. University of California Publications in Entomology 20: 1–188.
- Linsley EG (1964) The Cerambycidae of North America. Part V. Taxonomy and classification of the subfamily Cerambycinae, Tribes Callichromini through Ancylocerini. University of California Publications in Entomology, 22: 1–197.
- Löbl I, Smetana A (2010) Catalogue of Palaearctic Coleoptera, Volume 6, Chrysomeloidea. Apollo Books, Stenstrup, Denmark, 924 pp.
- McIver JD, Stonedahl G (1993) Myrmecomorphy: Morphological and behavioral mimicry of ants. Annual Review of Entomology 38: 351–379. doi:10.1146/annurev. en.38.010193.002031
- Museum of Comparative Zoology, Harvard University [MCZC] (2011) Museum of Comparative Zoology, Harvard University, Database of Images of Primary Types. http://insects.oeb. harvard.edu/mcz/index.htm [last accessed 11 February 2011]
- Micheli JA (2003) New longhorn beetles from Puerto Rico (West Indies). The Coleopterists Bulletin, 57(2): 191–204. doi:10.1649/540
- Micheli JA (2010) Longicornios de Puerto Rico. Pensoft Publishers, Sofia, Bulgaria: 298 pp.

- Micheli JA, Hovore FT (2003) New synonymies and distributional records for Caribbean Cerambycidae (Coleoptera). The Coleopterists Bulletin 57(1): 1–4. doi:10.1649/0010-065X(2003)057[0001:NSADRF]2.0.CO;2
- Monné MA (2005) Catalogue of the Cerambycidae (Coleoptera) of the Neotropical region. Part I - Subfamily Cerambycinae. Zootaxa 946: 1–765.
- Monné MA, Bezark LG (2010) Checklist of the Cerambycidae, or longhorned beetles (Coleoptera) of the Western Hemisphere. Available online at: http://plant.cdfa.ca.gov/byciddb/ documents.html [last accessed February 11, 2011]
- Nearns EH, Branham MA, Bybee SM (2006) Cerambycidae (Coleoptera) types of the Fernando de Zayas Collection, Havana, Cuba. Zootaxa 1270: 1–17.
- Pascoe FP (1869) Longicornia Malayana; or, a descriptive catalogue of the species of the three longicorn families Lamiidae, Cerambycidae and Prionidae, collected by Mr. A. R. Wallace in the Malay Archipelago. Transactions of the Entomological Society of London (third series) 3(6): 497–710.
- Palisot de Beauvois AMFJ (1805-1821) Insectes recueillis en Afrique et en Amérique, dans les royaumes d'Oware et de Benin, a Saint-Domingue et dans les États-Unis, pendant les années 1786–1797; par A.-M.-F.-J Palisot de Beauvois. Paris, Impr. de Fain et compagnie.
- Perez-Gelabert DE (2008) Arthropods of Hispaniola (Dominican Republic and Haiti): A checklist and bibliography. Zootaxa 1831: 1–530.
- Shelford RWC (1902) Observations on some mimetic insects and spiders from Borneo and Singapore. Proceedings of the Zoological Society of London 2(1): 230–284.
- Zayas F (1975) Revisión de la familia Cerambycidae (Coleoptera, Phytophagoidea). Academia de Ciencias de Cuba, Instituto de Zoología, Havana, Cuba, 443 pp.

SHORT COMMUNICATION



Record of the invasive alien ladybird Harmonia axyridis (Coleoptera, Coccinellidae) from Kenya

Oldřich Nedvěd¹, Jiří Háva², Daniela Kulíková³

Faculty of Sciences, University of South Bohemia, and Institute of Entomology, Biology Center, Academy of Sciences of the Czech Republic, Branišovská 31, 370 05 České Budějovice, Czech Republic 2 Private Entomological Laboratory & Collection, Rýznerova 37, CZ-252 62 Únětice u Prahy, Praha-západ, Czech Republic
Podřipská 188, CZ-41185 Horní Beřkovice, Czech Republic

Corresponding author: Oldřich Nedvěd (nedved@prf.jcu.cz)

Academic editor: Michael Thomas | Received 11 March 2011 | Accepted 27 April 2011 | Published 15 June 2011

Citation: Nedvěd O, Háva J, Kulíková D (2011) Record of the invasive alien ladybird *Harmonia axyridis* (Coleoptera, Coccinellidae) from Kenya. ZooKeys 106: 77–81. doi: 10.3897/zooKeys.106.1242

Abstract

The biological control agent and alien invasive ladybird *Harmonia axyridis* (Pallas, 1773) was recorded for the first time in Kenya, and in equatorial Africa, in 2010.

Keywords

Multicolored Asian Ladybird, distribution, new record, Kenya, Afrotropical region, invasive predator

Introduction

The multicolored Asian lady beetle or harlequin ladybird *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae) is native to temperate (and mountain subtropical) Central and East Asia: China, Taiwan, Japan, Korea, Mongolia, Kazakhstan and eastern Russia (Kuznetsov 1992). It was introduced in many regions of the world as a biological control agent against aphids, and later became an invasive species, spreading 100–500

km each year. It is established in at least 37 countries in four continents (Brown et al. in press). In Africa, this species was intentionally introduced in two Mediterranean countries: Tunisia, where it did not survive, and Egypt (Ferran et al. 2000), where it established a limited population (Brown et al. in press). Conversely, it has invaded and established in South Africa (Stals and Prinsloo 2007) and neighbouring Lesotho (Stals 2010) although it was not intentionally introduced there.

This article reports the first record of this alien invasive ladybird beetle in Kenya.

Material examined

Kenya E, Coast province, Kikambala (3° 48.28'S; 39° 50.00'E; cca. 45 km N of Mombasa), 30.12.2010–8.1.2011, 2 \bigcirc lgt. + 20 exx. observ., Jiří Háva & Daniela Kulíková lgt., J. Háva coll. et det.

We observed the beetles on the plant *Ipomoea pescapre* (Convolvulaceae) on the sea coast (Fig. 1). All individuals belonged to the colour morph *succinea* (Hope), with 19 well-developed spots on the elytra and well-developed elytral ridges (Fig. 2). Like a previous record of *H. axyridis* in Uruguay (Nedvěd and Krejčík 2010), this finding was done by chance by a non-professional entomologist.



Figure 1. The host plant Ipomoea pescapre (Convolvulaceae) on the sea coast in Kikambala.



Figure 2. Female of *Harmonia axyridis* from Kikambala, colour morph *succinea*, with 19 spots and elytral ridge.

Discussion

Because Kikambala is a holiday centre, but not a port or transport node, we consider the occurrence of *H. axyridis* here to be the result of a wider and lasting invasion, rather than a singular incidental and ephemeral introduction with goods.

The observed colour morph *succinea* is the most common morph in the eastern part of its natural range (Blekhman et al. 2010) and in the invasive European population (Brown et al. 2008). The size of the spots suggests that the individuals recorded emerged from pupae at temperatures of around 25°C – the spots would be smaller or missing at higher temperatures (Michie et al. 2010).

High temperature may be limiting the continued spread of *H. axyridis*, at least at a local scale. The American (Acar et al. 2001) and European (Fois et al., unpublished) invasive populations do not survive temperatures above 33°C. However, the CLIMEX model that used known physiological limits of *H. axyridis* indicated that this species may tolerate most southern and eastern African countries, including Kenya (Poutsma et al. 2008). The coastal climate near Mombasa is rather hot (average annual temperature 26°C, Climate & Temperature 2011), while at higher elevations inland, mild temperatures (*e.g.* 18°C in the capital, Nairobi) are more favourable for *H. axyridis*.

Although there were several independent introductions of *H. axyridis* in Europe and North America, with different source populations from East Asia, there is a single main invasive population/strain in several continents (Lombaert et al. 2010). Thus in future the origin of the population in Kenya should be compared with known populations from both the native and invasive ranges, using molecular genetic methods (Blekhman et al. 2010, Thomas et al. 2010, Lawson Handley et al., in press) to determine if it is the same strain, or a different one that might have higher temperature requirements.

Conclusion

We consider that *H. axyridis* has established in Kenya, the first fully tropical country to be invaded, but that its further spread may be hampered by high temperature and low prey availability. In this region we suggest that *H. axyridis* may pose a low threat to biodiversity, such as the native ladybird beetles, which are mostly coccidophagous.

Acknowledgement

The study was supported by grant number QH82047 by the Ministry of Agriculture of the Czech Republic.

References

- Acar B, Smith BN, Hansen LD, Booth GM (2001) Calorespirometry to determine the effects of temperature on the metabolic efficiency of *Harmonia axyridis* (Col.: Coccinellidae) larvae. http://esa.confex.com/esa/2001/techprogram/paper_3292.htm [accessed 10 Feb 2011]
- Blekhman AV, Goryacheva II, Zakharov IA (2010) Differentiation of *Harmonia axyridis* Pall. according to polymorphic morphological traits and variability of the mitochondrial COI gene. Moscow University Biological Sciences Bulletin 65: 174–176. doi:10.3103/ S0096392510040140
- Brown PMJ, Adriaens T, Bathon H, Cuppen J, Goldarazena A, Hagg T, Kenis M, Klausnitzer BEM, Kovar I, Loomans AJM, Majerus MEN, Nedved O, Pedersen J, Rabitsch W, Roy HE, Ternois V, Zakharov IA, Roy DB (2008) *Harmonia axyridis* in Europe: spread and distribution of a non-native coccinellid. BioControl 53: 5–21. doi: 10.1007/s10526-007-9132-y
- Brown PMJ, Thomas CE, Lombaert E, Jeffries DL, Estoup A, Lawson Handley LJ (in press) The global spread of *Harmonia axyridis* (Coleoptera: Coccinellidae): distribution, dispersal and routes of invasion. BioControl.
- Climate & Temperature: http://www.climatetemp.info/kenya/mombasa.html [accessed 6 Mar 2011]
- Ferran A, El-Arnaouty SA, Beyssat-Arnaouty V, Galal H (2000) Introduction and release of the coccinellid *Harmonia axyridis* Pallas for controlling *Aphis craccivora* Koch on faba beans in Egypt. Egyptian Journal of Biological Pest Control 10: 129–136.

- Kuznetsov VN (1992) Family Coccinellidae ladybirds. In: Lera PA (Ed) Identification key to insects of Far East of Soviet Union III, 2. Nauka, 333–376. [In Russian]
- Lawson Handley LJ, Estoup A, Thomas C, Lombaert E, Facon B, Aebi A, Evans D, Roy HE (in press) Ecological genetics of invasive species. BioControl.
- Lombaert E, Guillemaud T, Cornuet JM, Malausa T, Facon B, Estoup A (2010) Bridgehead effect in the worldwide invasion of the biocontrol harlequin ladybird. Plos One 5: e9743. doi:10.1371/journal.pone.0009743
- Michie LJ, Mallard F, Majerus MEN, Jiggins FM (2010) Melanic through nature or nurture: genetic polymorphism and phenotypic plasticity in *Harmonia axyridis*. Journal of Evolutionary Biology 23: 1699–1707. doi: 10.1111/j.1420-9101.2010.02043.x
- Nedvěd O, Krejčík S (2010) Record of the ladybird *Harmonia axyridis* (Coleoptera: Coccinellidae) from Uruguay. Klapalekiana 46: 203–204.
- Poutsma J, Loomans AJM, Aukema B, Heijerman T (2008) Predicting the potential geographical distribution of the harlequin ladybird, *Harmonia axyridis*, using the CLIMEX model. BioControl 53:103–125. doi: 10.1007/s10526-007-9140-y
- Stals R (2010) The establishment and rapid spread of an alien invasive lady beetle: *Harmonia axy-ridis* (Coleoptera: Coccinellidae) in southern Africa, 2001-2009. IOBC WPRS Bulletin 58: 125–132. http://www.iobc-wprs.org/restricted_member/iobc-wprs_bulletin_2010_58.pdf
- Stals R, Prinsloo G (2007) Discovery of an alien invasive, predatory insect in South Africa: the multicoloured Asian ladybird beetle, *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae). South African Journal of Science 103: 123–126.
- Thomas C, Lombaert E, Ware RL, Estoup A, Lawson Handley LJ (2010) Investigating global invasion routes of the harlequin ladybird (*Harmonia axyridis*) using mtDNA. IOBC WPRS Bulletin 58:155–157. http://www.iobc-wprs.org/restricted_member/iobc-wprs_ bulletin_2010_58.pdf