

# Systematics of the Phyllognathopodidae (Copepoda, Harpacticoida): re-examination of *Phyllognathopus viguieri* (Maupas, 1892) and *Parbatocamptus jochenmartensi* Dumont and Maas, 1988, proposal of a new genus for *Phyllognathopus bassoti* Rouch, 1972, and description of a new species of *Phyllognathopus*

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## Abstract

The family Phyllognathopodidae (Crustacea, Copepoda, Harpacticoida) is heavily affected by the floating taxonomic status of the type-genus *Phyllognathopus*. A revision of the different character states displayed by members of the family is presented, and new phylogenetically informative characters are described, enlarging the analysis to the remaining genera of the family, *Parbatocamptus* and *Allophyllognathopus*. *Phyllognathopus viguieri* (Maupas, 1892) and *Parbatocamptus jochenmartensi* Dumont and Maas, 1988 are redescribed in detail, and *Phyllognathopus inexpectatus* sp. n. is described from ground water in Italy. The new genus *Neophyllognathopus* is established to accommodate *Phyllognathopus bassoti* Rouch, 1972, originally collected from Long Island (Papua - New Guinea), and subsequently recorded also from the Bantayan Island (Philippines), and from the Indian subcontinent. The new genus is presently monotypic and is easily defined by the unique construction and morphology of leg 5 in both male and female, of male leg 6, and by the peculiar ornamentation of male third and fourth urosomites. Biogeographical and ecological considerations are presented for members of the family.

**Keywords**

Harpacticoida, Phyllognathopodidae, taxonomy, *Phyllognathopus*, *Parbatocamptus*, *Neophyllognathopus bassoti* comb. n.

**Introduction**

The harpacticoid family Phyllognathopodidae exhibits a low diversity, currently containing 13 species according to Boxshall and Halsey (2004), accommodated in three genera: *Phyllognathopus* Mrázek, 1893, *Allophyllognathopus* Kiefer, 1967 and *Parbatocamptus* Dumont and Maas, 1988. Members of the family are predominantly recorded from semi-terrestrial and freshwater habitats (i.e. phytotelmata, leaf litter, epibenthic habitats in streams and springs, hyporheic habitats, ground water) (Reid 2001), and findings in anchialine caves or brackish ground water (Huys et al. 1996, pers. comm.; Bruno and Cottarelli 1999) represent mere exceptions.

The family is undoubtedly monophyletic, being instantly recognizable by the unique phyllopodial lamelliform maxilliped, in conjunction with the first pedigerous somite not being incorporated into the cephalosome. Despite the low diversification in the family, the taxonomic history of the type-genus *Phyllognathopus* has been and still is highly controversial (Glatzel and Königshoff 2005). The root cause for this state of affairs lies in three questionable assumptions surrounding the taxonomy of *Phyllognathopus viguieri* (Maupas, 1892) and which were employed early in delimiting species within the genus: 1) its cosmopolitanism; 2) its ecological plasticity, enabling itself to colonize virtually any kind of habitat, from the truly aquatic to semi-terrestrial; and 3) as a reflection of the latter, its morphological variability. The taxonomic confusion surrounding the type-species of *Phyllognathopus* is also reflected in the entire systematics of the family (Boxshall and Halsey 2004, Wells 2007), preventing any reconstruction of the phylogenetic relationships among members of this family.

Recent discoveries of new phyllognathopodid representatives in Italian ground water, together with the re-examination of different species and populations coming from several localities world-wide, allowed a re-analysis of key-taxa within the family, the description of a new stygobiotic species of *Phyllognathopus* and the establishment of a new taxonomic rank for *Phyllognathopus bassoti* Rouch, 1972, assigned herein to the new genus *Neophyllognathopus*.

**Material and methods**

Specimens were collected from hyporheic habitats by using the Bou-Rouch method (Bou and Rouch 1967) and filtering through a 60 µm mesh net. Epibenthic samples from springs were taken by washing sediments with a hand net or by using a drift net positioned at the major outlets of the sampled springs. Specimens were preserved in 7%

formalin solution and dissected in polyvinyl lactophenol. Drawings were made using camera lucida on a Leitz Laborlux phase contrast microscope. Some details gained from scanning electron microscopy (SEM) are added to line drawings. For SEM, 10 females and 8 males of *Phyllognathopus viguieri*, and 4 females and 2 males of *Phyllognathopus bassoti* were dehydrated in a graded ethanol series, critical point dried in a Balzers Union CPD 020 apparatus and coated with gold in a Balzers Union SCD 040 sputter. Observations were made with a Philips SEM XL30 CP scanning electron microscope.

Additional material, preserved on slides, was loaned by the Natural History Museum (London), the Museum National d'Histoire Naturelle (Paris), the National Museum of Natural History, Smithsonian Museum, Washington, D.C. (U.S.A.), the Senckenberg Museum (Germany). The descriptive terminology of Huys and Boxshall (1991) is adopted. Abbreviations used in the text and figures are: P1-P6, first to sixth thoracopods; exp., exopod; enp., endopod; exp (enp) -1 (-2, -3) to denote the proximal (middle, distal) segment of a ramus.

## Results

### Order HARPACTICOIDA Sars, 1903

### Family PHYLLOGNATHOPODIDAE Gurney, 1932

### Genus *Phyllognathopus* Mrázek, 1893

<http://species-id.net/wiki/Phyllognathopus>

**Emended diagnosis.** Phyllognathopodidae. Habitus slender, with no clear demarcation between prosome and urosome. Integumental dorsal window on cephalosome not confirmed for all members of the genus. Integument with surface pits, moderately sclerotized. Cephalosome rounded; rostrum clearly articulated to cephalosome. First pedigerous somite free. P5-bearing somite with large paired pores laterodorsally. Anal operculum plain or ornamented by fine spinules or extruded in strong spinular processes. Sexual dimorphism in antennule, P5, P6, urosomal segmentation and ornamentation. Female first and second abdominal somites fused forming genital double-somite. Anal somite with paired sensilla on dorsal side. Male urosome consisting of 6 segments. Caudal rami sub-quadrate, or longer than wide, with incomplete setal pattern (6 setae). Dorsal seta inserted on distal third of caudal ramus. Setae III and V variable in morphology among species. Antennule: 8-segmented in female, basically 10-segmented in male, although a suture line marking original segmentation between former segments 10 and 11 may be still discernible in some species; geniculation between segments 7 and 8; segment 9 discrete. Long tube-pores on segments 1 and 2 in both sexes. Antenna: armature of the second endopodal segment consisting of 10 elements. Exopod 1-segmented, with 3 lateral and 2 apical setae. Mandible: mandibular palp biramous, basis unarmed; exopod with 1 apical and 1 inner setae; endopod with 1 inner, 1 subapical and 2 apical setae. Armature of maxillule and maxilla as in *Phyllognathopus viguieri*. Maxilliped: phyl-

lopodial, lamelliform, 1-segmented. Trace of ancestral 2-segmented condition marked by the presence of outer incision; armature consisting of 10 elements.

P1-P3 with praecoxa and 3-segmented exopods and endopods. P4 small-sized, praecoxa missing, with 3- or 2- or 1-segmented exopod and 2- or 1-segmented endopod. Female P5 free, with clear articulation to P5-bearing somite; right and left legs distinct; intercoxal sclerite absent; baseoendopod and exopod coalescent, feeble incision marking original segmentation between them; endopodal lobe not pronounced, bearing 2 apical setae. Exopodal lobe fully incorporated into baseoendopod, not pronounced; basipodal outer seta present. Female P6 present, right and left legs represented by small chitinous lamellar plates, each leg bearing 1 normal seta or a stout spine with rounded tip. Male P5 free, with clear articulation to P5-bearing somite; right and left legs coalescent, intercoxal sclerite absent. Exopod discrete, but sometimes incorporated to basis. Endopod 1-segmented, normally conformed, cylindrical, bearing 1 leaf-like seta, alternatively transformed in a curved and stout element bearing 1 bipinnate seta, inserted on posterior surface of the endopod, close to its articulation to basis. Male P6 present; symmetrical, right and left legs coalescent along their medial margin, forming a continuous lamellar plate; each leg bearing 2 inner spines of different length and 1 outer seta.

### *Phyllognathopus viguieri* (Maupas, 1892)

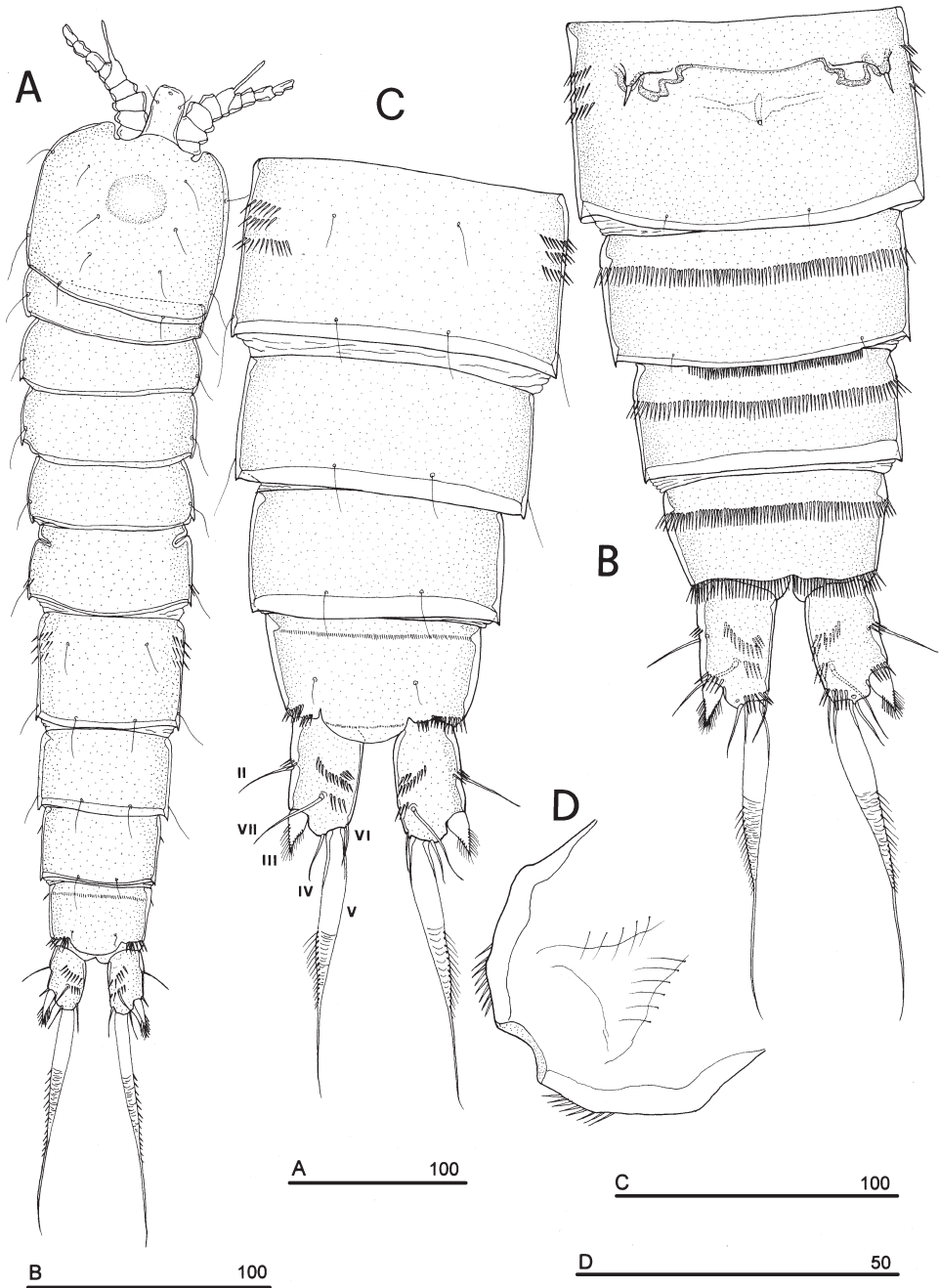
[http://species-id.net/wiki/Phyllognathopus\\_viguieri](http://species-id.net/wiki/Phyllognathopus_viguieri)

Figs 1–9

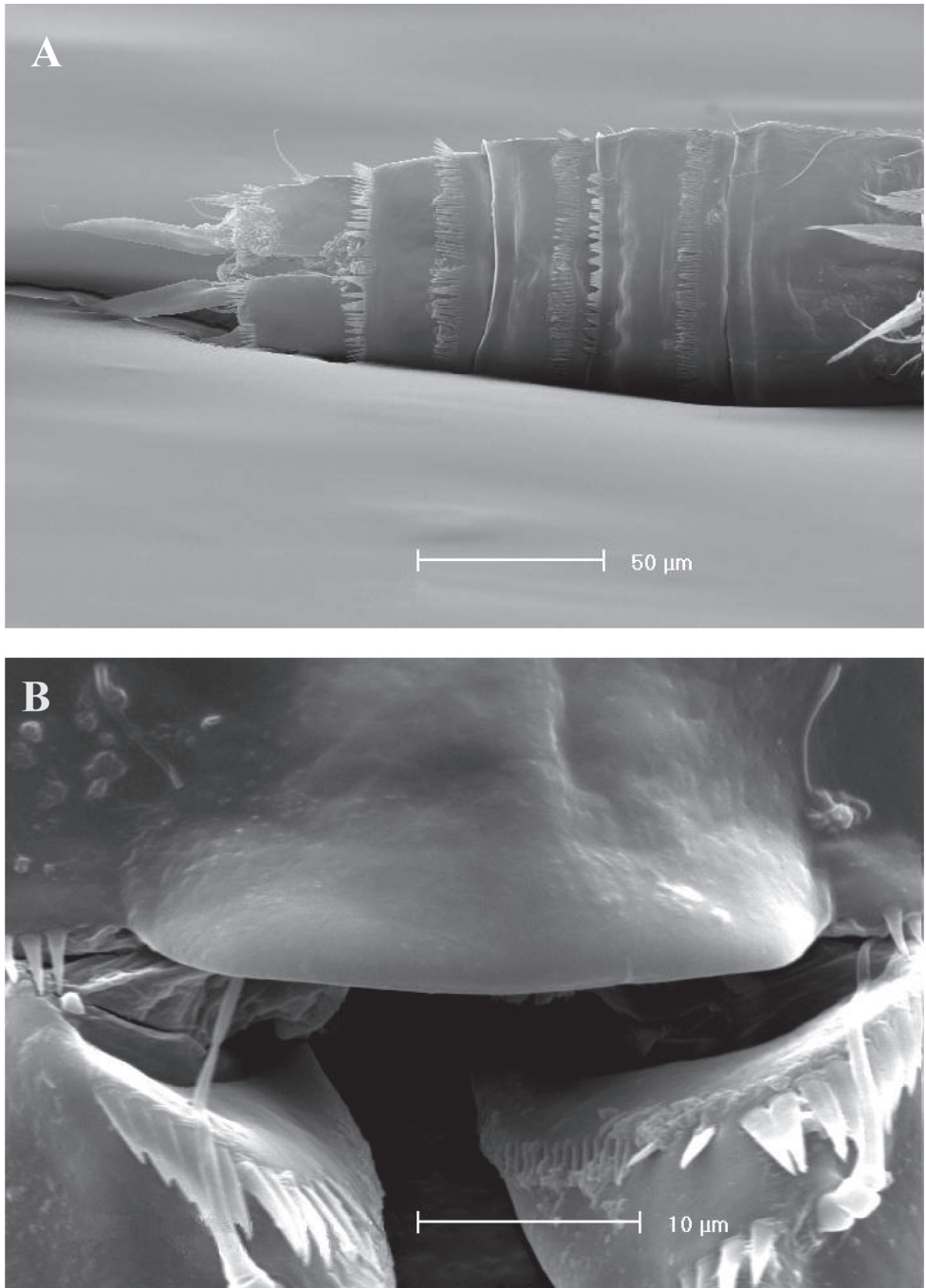
**Material examined.** 11 ♀♀ and 2 ♂♂, completely dissected and mounted in polyvinyl lactophenol, S. Anna D'Alfaedo, Progno di Valpantena (Verona, Italy), hyporheic habitat, 25.06.2002, E. Gattone coll.; 1 ♀, karstic spring in the hydrogeological basin of Rio Biondo, Progno di Valpantena (Verona, Italy), karstic habitat, 7. 07. 2003, B. Fiasca coll.; 3 ♀♀, Lake Bracciano (Latium, Italy), interstitial habitat, 27.05.02, V. Cottarelli coll.; 10 ♀♀ and 5 ♂♂, Avisio floodplain (Trento, northern Italy), hyporheic habitat, 30.05.2006, T. Di Lorenzo coll.; 3 ♀♀ and 1 ♂, Oignin stream (French Jura Mountains), hyporheic habitat, 30.07.2002, M.-J. Dole-Olivier coll.; 2 ♀♀ and 1 ♂, Ariège floodplain (France), P. Dumas coll.; 1 ♀, Lac Léman (France), slide code MNHN - Cp922, Paris; 2 ♂♂, S. Pierre, (France), slide code MNHN - Cp456, Paris; 3 ♂♂ and 2 ♀♀, Ruhr floodplain (Germany), T. Glatzel coll.; 6 ♀♀ and 1 ♂, R. Krishna, India, Y. Ranga Reddy coll.; 1 ♀, in pitcher of *Nepenthes mirabilis* (Hong Kong), B. Coker det., slide code 1982.329, Natural History Museum, London; 2 ♀♀ deposited at the Smithsonian Institution, Washington D.C., code USMN 251806, 204501; 1 ♂, code USMN 204500; 1 ♂ (juvenile), code USMN 204501.

*Phyllognathopus* cf. *viguieri* A. 2 ♂♂, 2 ♀♀, Madagascar, vial code, MNHN - Cp910, Paris, B. Dussart coll..

*Phyllognathopus* cf. *viguieri* B. 1 ♀, slide code 66/52, freshwater well, Mindoro Island, Philippines, 17.8.1992, V. Cottarelli coll.



**Figure 1.** *Phyllognathopus viguieri* (Maupas, 1892) (♀). **A** habitus, dorsal view **B** abdomen, ventral view **C** abdomen, dorsal view **D** labrum (scale bars in  $\mu\text{m}$ ).



**Figure 2.** SEM micrographs of *Phyllognathopus viguieri* (Maupas, 1892) (♀). **A** ventral surface of urosome (first urosomite omitted) **B** anal operculum.

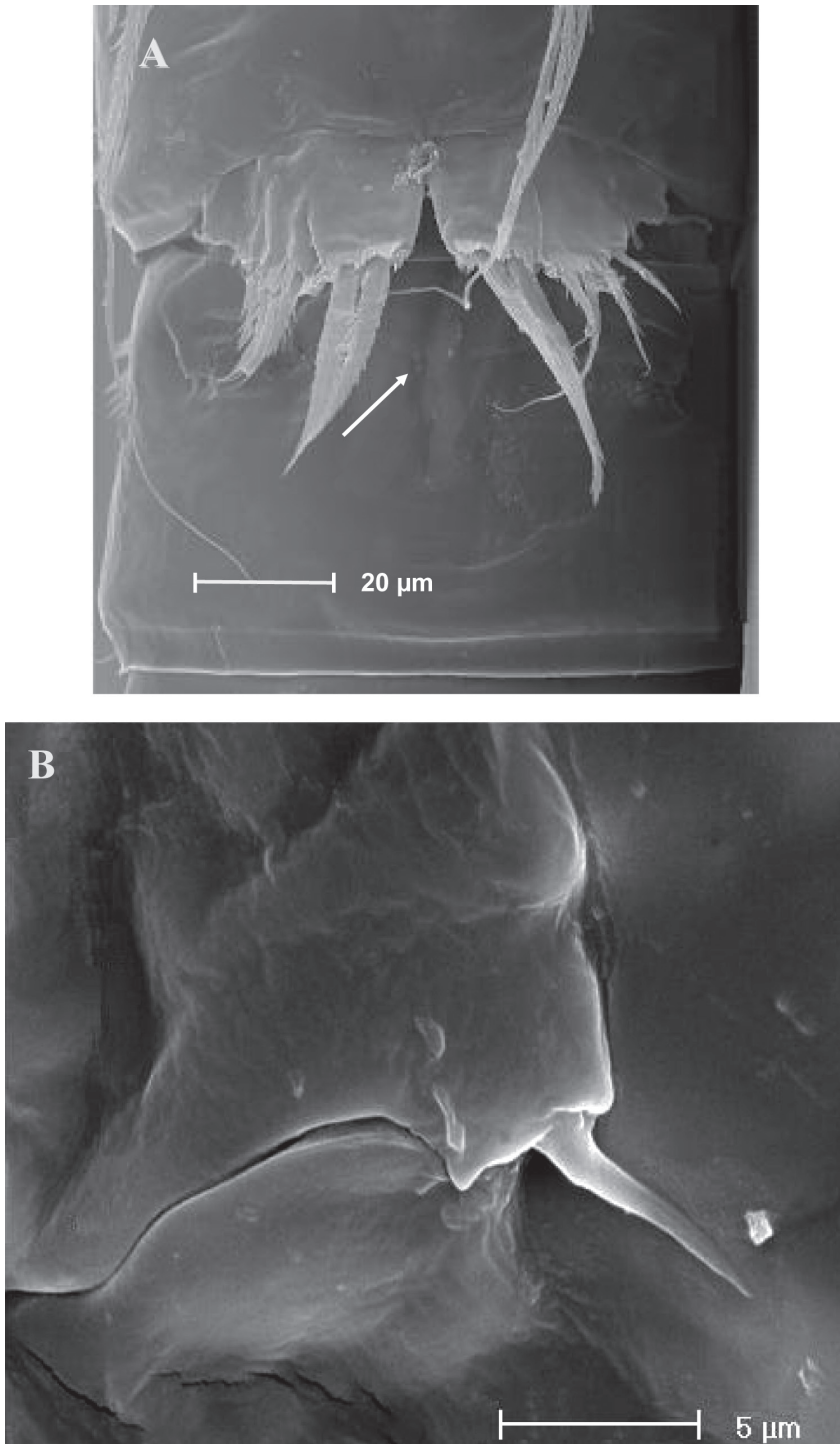
*Phyllognathopus viguieri* ?. 10 ♀♀ and 5 ♂♂, Andhra Loyola College Campus, Vijaya-Woda, Andhra Pradesh, India, Y. Ranga Reddy coll..

*Phyllognathopus viguieri menzeli*. 1 ♀, vial code USNM 150192, labelled: Pacific Ocean, Mariana Islands, Guam, 19 November 1970, Watkins R.L. coll. (remaining material in the vial: 17 ♀♀).

*Phyllognathopus viguieri menzeli* 2. ♂♂, vial code USNM 150193, labelled: Pacific Ocean, Mariana Islands, Guam, 1 April 1971, Belk and Watkins R.L. coll. (remaining material in the vial: 8 specimens, of which several copepodids).

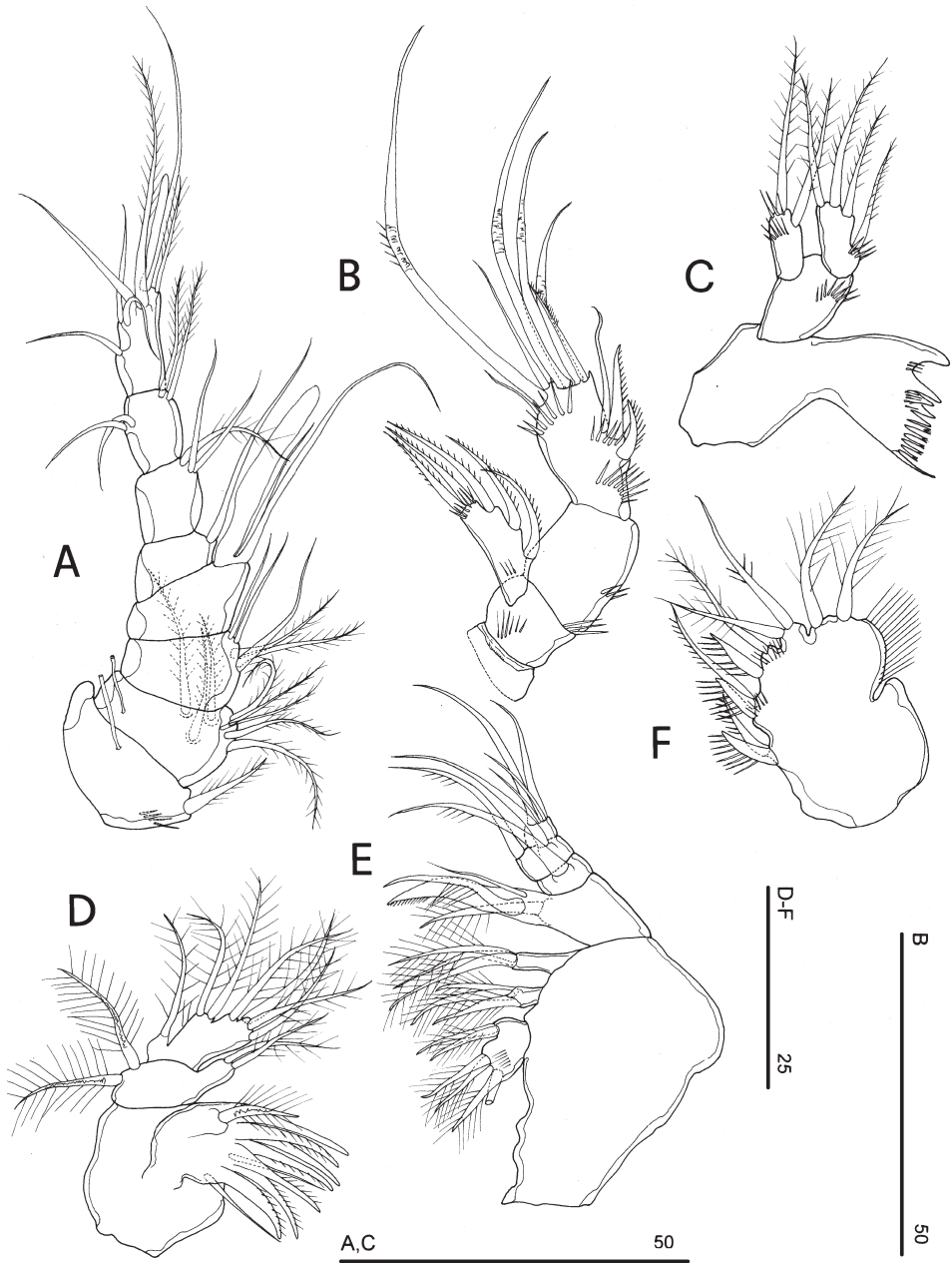
**Supplementary description.** FEMALE. Body length, measured from tip of rostrum to posterior margin of caudal rami, from 400 to 600  $\mu\text{m}$  (mean = 439  $\mu\text{m}$ ;  $n = 27$ ). Habitus slender, no clear demarcation between prosome and urosome. Integument with surface pits, moderately sclerotized as in Fig. 1A. Cephalosome sub-quadrangle, with a dorsal rounded protuberance, hardly observable, plausibly referable to a dorsal integumental window. Setule pattern as in Fig. 1A. Rostrum elongate, subrectangular in shape, clearly articulated to the cephalosome; two dorsal sensilla laterally inserted on its distal third, and one pore apically. Cephalosome and both thoracic and abdominal somites with cuticular ornamentation apparently represented by reduced number of paired sensilla (Fig. 1A). First pedigerous somite free. Hyaline frills of cephalosome, somites bearing P1-P4 and urosome dorsally smooth. Urosomites with smooth hyaline frill ventrally, except third urosomite (Figs 1B, 2A). Last three urosomites with spinular fringe on proximal third ventrally; anal somite with distal continuous spinule row. Anal somite with paired sensilla on dorsal side only (Fig. 1C), and two short spinule rows close to the anal operculum. Anal operculum rounded, only slightly protruding beyond insertion line of caudal rami (Figs 1C, 2B). P5-bearing somite with large paired pores laterodorsally and paired spinule rows laterally inserted on distal third of somite (Fig. 1A). Genital double-somite with three lateral spinule rows. Female genital field located between first and second third of genital double-somite. Genital apparatus apparently simplified; copulatory pore located halfway of genital double-somite (Figs 1B, 3A). Seminal receptacles located laterally and condensed close to the lamellar P6 (Fig. 3B).

Caudal rami rectangular, parallel, distinctly longer than wide (length/width ratio about 1.7), with incomplete setal pattern (6 setae) (Fig. 1B–C). Anterolateral accessory seta (I) absent, anterolateral seta (II) smooth, inserted at half of caudal ramus; posterolateral seta (III) inserted on distal third of ramus, transformed in a large and stout spiniform element. Outer terminal seta (IV) very short, thin, and naked, shorter than caudal ramus, inner terminal seta (V) unipinnate and relatively short, without articulation at base, with very enlarged proximal part tapering in a subtle tip; terminal accessory seta (VI) slightly shorter than outer terminal seta, thin and naked; dorsal seta (VII) inserted on distal third of caudal ramus, about as long as caudal ramus. Three spinule rows inserted dorsolaterally and two spinule rows inserted at distal margin of caudal ramus ventrally. Two pores located close to the insertion of setae II and IV, ventrally.

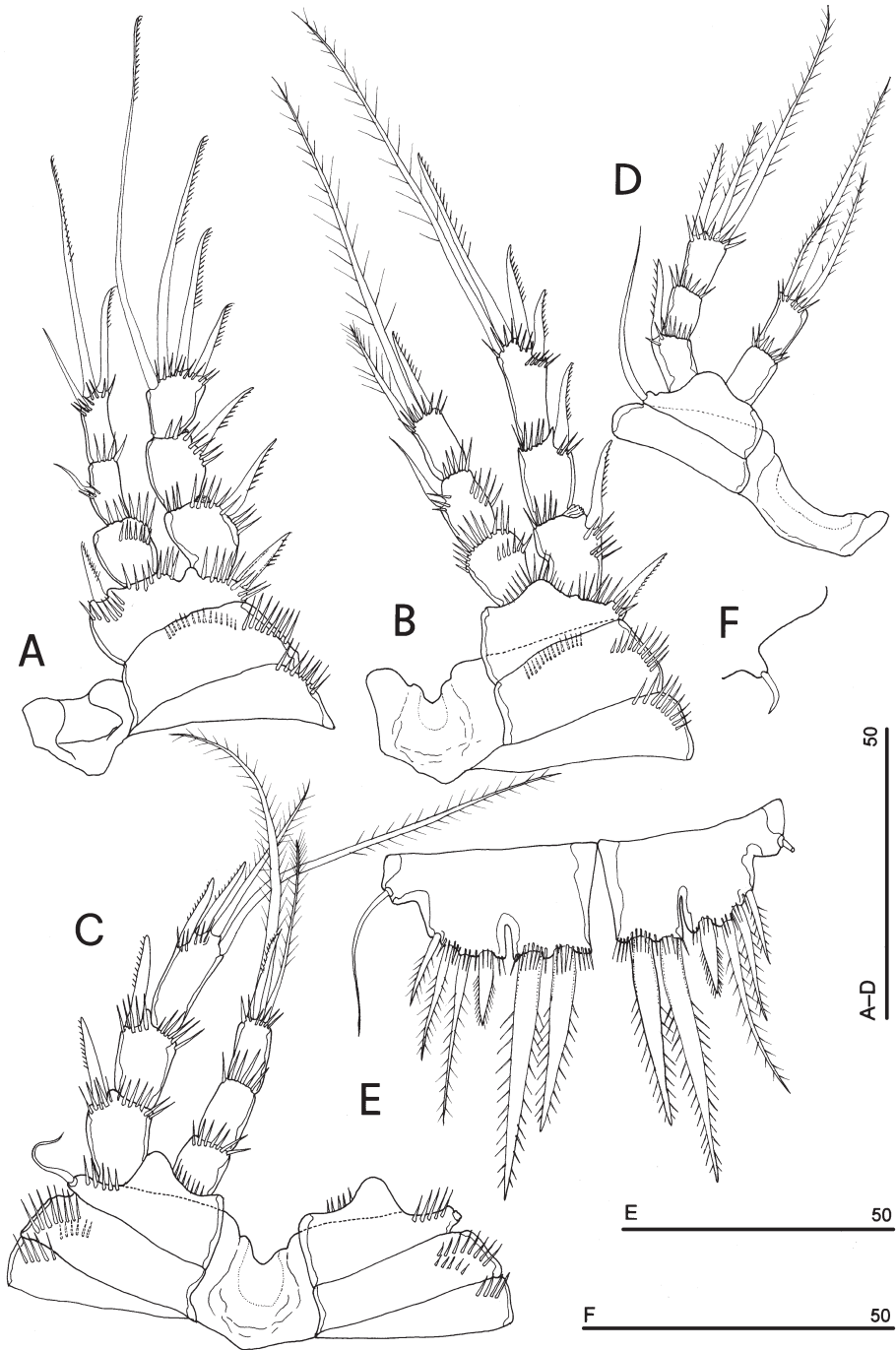


**Figure 3.** SEM micrographs of *Phyllognathopus viguieri* (Maupas, 1892) (♀). **A** P5 and genital double-somite (copulatory pore arrowed) **B** P6.

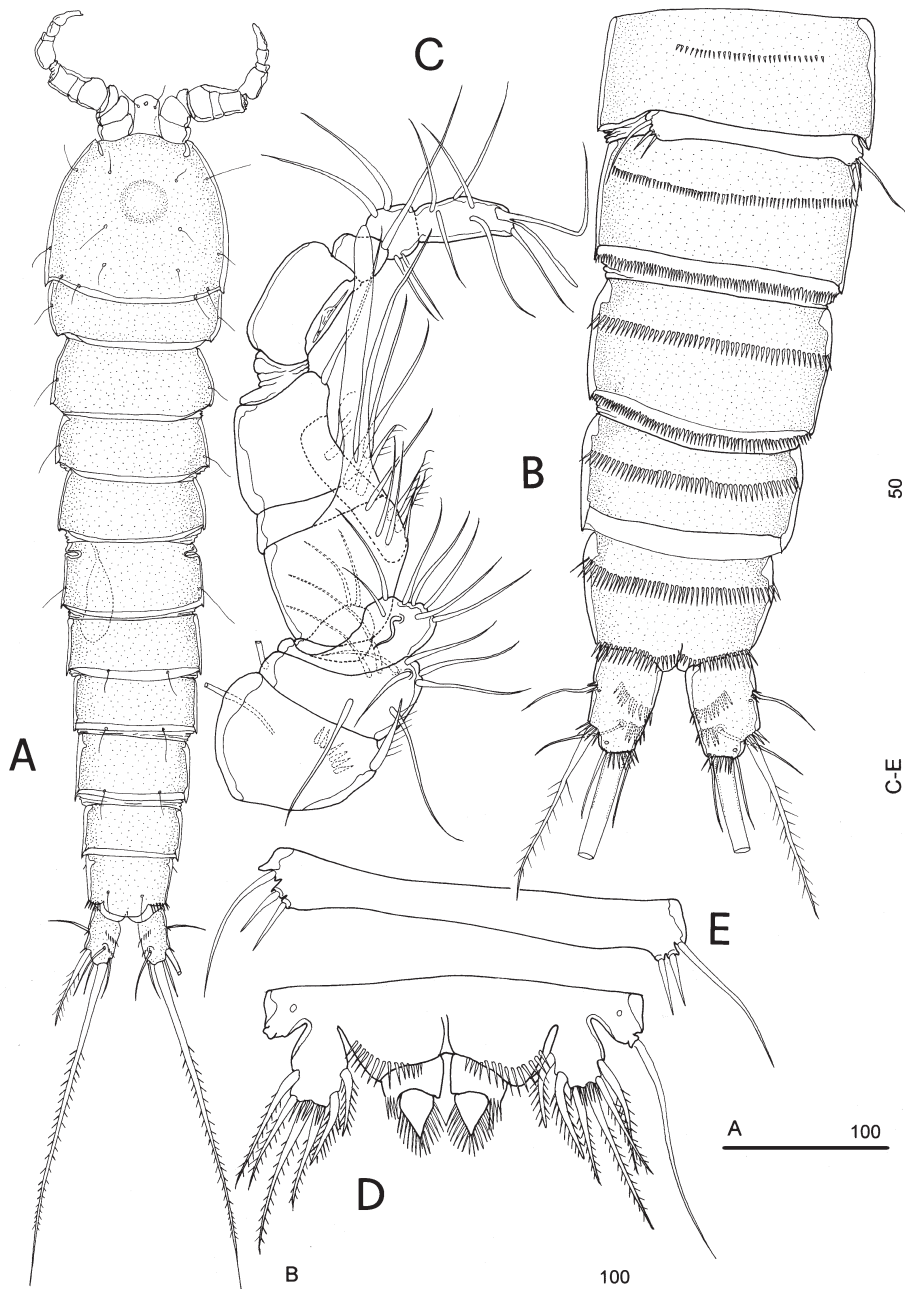




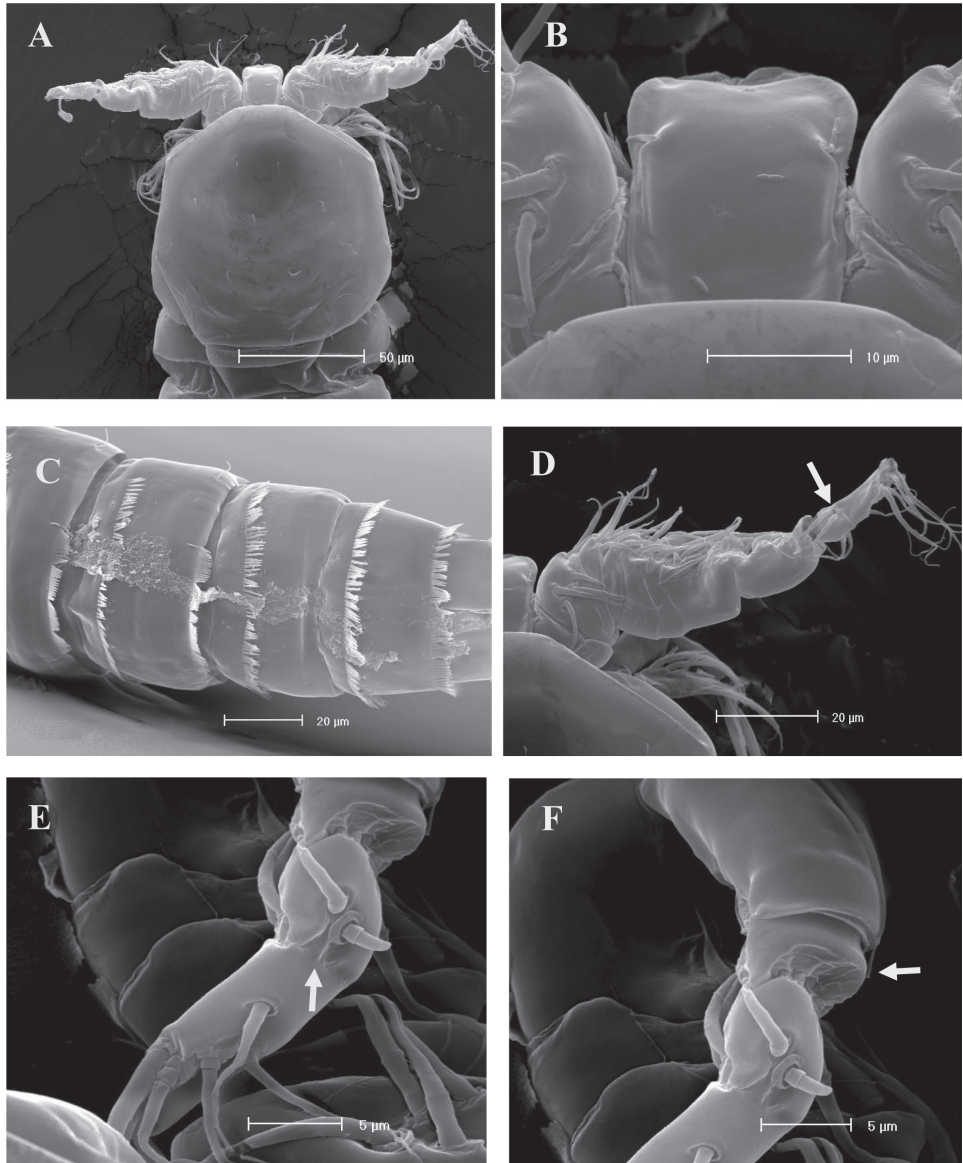
**Figure 4.** *Phyllognathopus viguieri* (Maupas, 1892) (♀). **A** antennule **B** antenna **C** mandible **D** maxillule **E** maxilla **F** maxilliped (scale bars in  $\mu\text{m}$ ).



**Figure 5.** *Phyllognathopus viguieri* (Maupas, 1892) (♀). **A** P1 **B** P2 **C** P3 **D** P4 **E** P5 **F** P6 (scale bars in  $\mu\text{m}$ ).

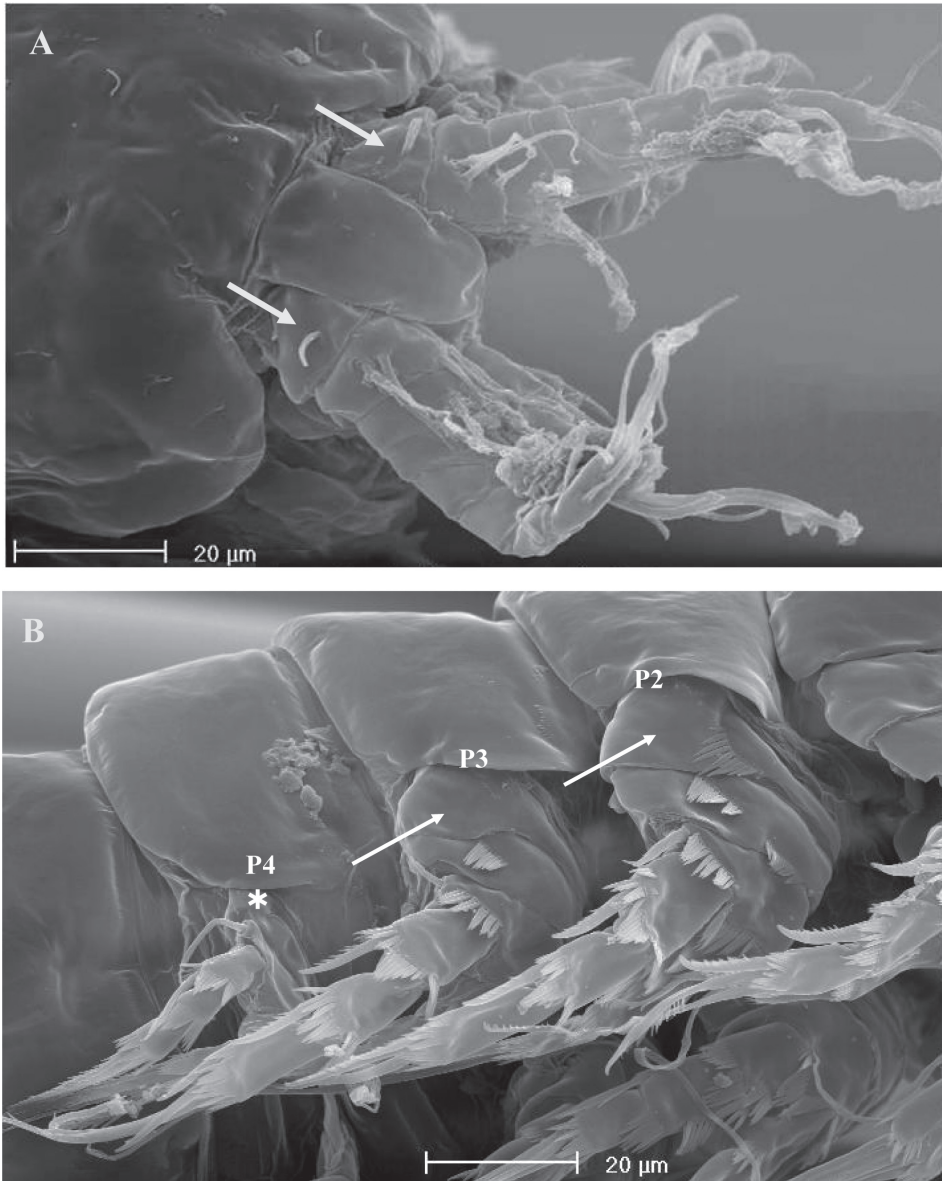


**Figure 6.** *Phyllognathopus viguieri* (Maupas, 1892) (♂). **A** habitus, dorsal view **B** abdomen, ventral view (first urosomite omitted) **C** antennule **D** P5 **E** P6 (scale bars in  $\mu\text{m}$ ).



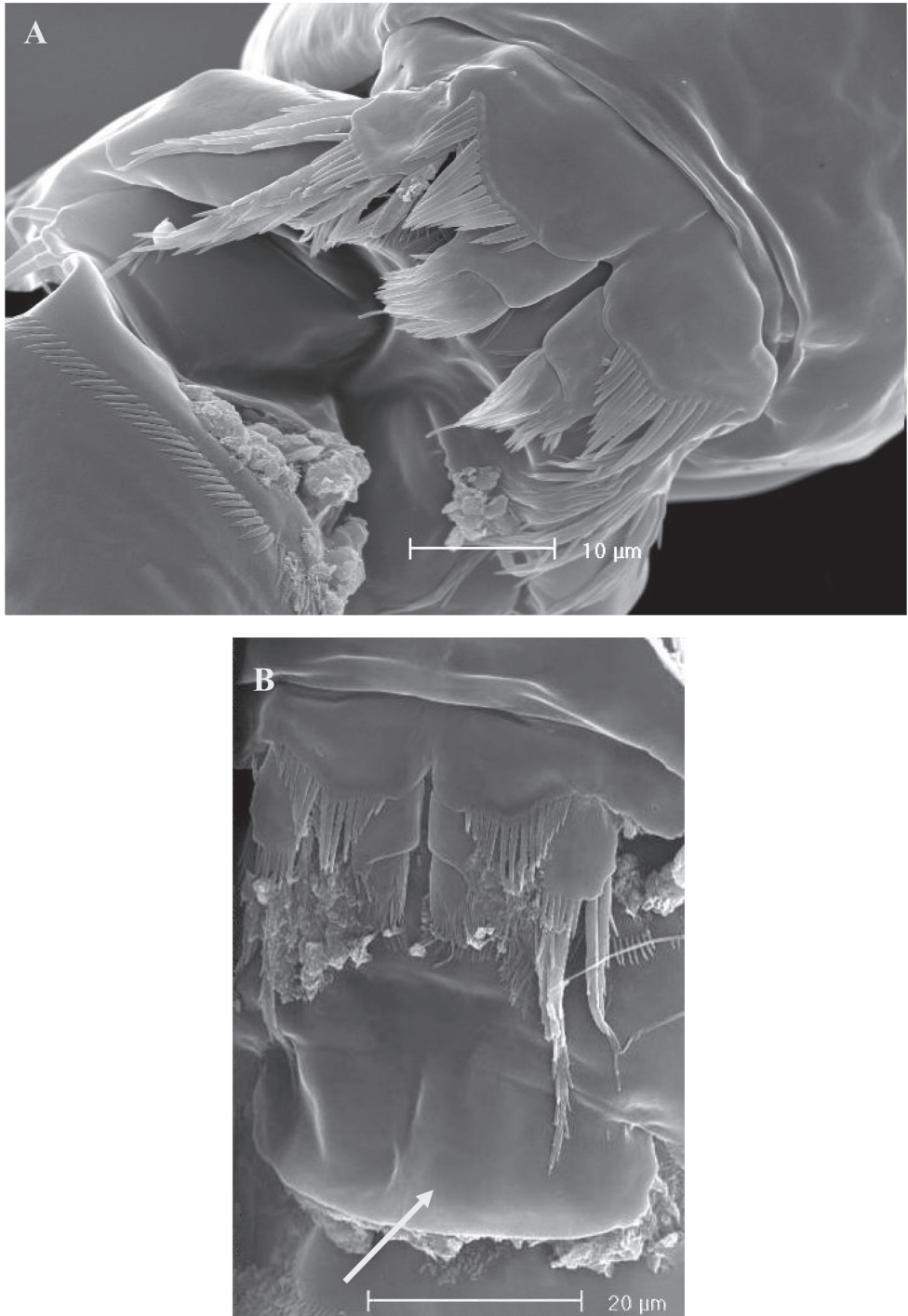
**Figure 7.** SEM micrographs of *Phyllognathopus viguieri* (Maupas, 1892) (♂). **A** cephalosome, rostrum and antennule **B** rostrum (detail) **C** ornamentation of third to six urosomites, ventral surface **D** antennule, segments 10 and 11 discrete on posterior surface (boundary line arrowed) **E** antennule, detail of segments 10 and 11 fused on anterior surface (suture line arrowed) **F** antennule, segment 9 arrowed.

Antennule (Fig. 4A): short, 8-segmented. Segment 1 with 1 spinule row. Both segments 1 and 2 bearing long and flaccid tube-pores. Armature formula: 1-[1], 2-[8], 3-[5], 4-[1 + (1+ae)], 5-[1], 6-[3], 7-[4], 8-[6 + (1+ae)]. Aesthetasc on segment 4 large, reaching about the proximal part of the penultimate antennulary segment.



**Figure 8.** SEM micrographs of *Phyllognathopus viguieri* (Maupas, 1892) (♂). **A** antennule, tube-pores on the first antennular segment arrowed **B** P2-P4 (P2-P3 praecoxa arrowed; asterisk indicates the P4 praecoxa missing).

Antenna (Fig. 4B): coxa unarmed; basis with 1 transverse spinule row on surface, a spinule row inserted on inner margin; exopod 1-segmented, well-defined at base, with surface spinule row, bearing 3 lateral and 2 apical setae; free endopod 2-segmented; both segments robust, of about the same length; segment 1 with inner spinule row; segment 2 with two inner spinule rows; armature consisting of 2 inner spines and 1



**Figure 9.** SEM micrographs of *Phyllognathopus viguieri* (Maupas, 1892) (♂). **A** P5 **B** P6 (arrowed).

seta, 1 unipinnate apical spine, 4 geniculate setae, 1 apical slender seta and 1 subapical slender seta; a row of spinules at outer corner.

Labrum (Fig. 1D): trapezoidal, with two spinule rows on free distal margin. Paired rows of hair-like elements on medioventral surface.

Mandible (Fig. 4C): coxal gnathobase elongate, cutting edge with 2 large and coarse teeth, three smaller teeth and row of fringed teeth; naked seta at dorsal corner. Mandibular palp biramous, basis with inner spinule row, exopod with 1 apical and 1 inner bipinnate setae; endopod with 1 inner, 1 subapical and 2 apical bipinnate setae.

Maxillule (Fig. 4D): well developed arthrite incorporated into praecoxa, with 7 strong curved spines inserted on free distal margin and 1 short seta inserted on a sort of surface peduncle and 2 anterior surface setae. Coxal epipodite represented by 2 setae; coxo-endite with 2 plumose setae. Exopod and endopod incorporated into basis, with a total of 7 plumose setae.

Maxilla (Fig. 4E): syncoxa with 3 endites. Proximal endite with 6 setae; medial and distal endites, each with 3 plumose setae. Allobasis drawn out into a strong claw, distally spinulose, accompanied by 2 robust and 1 thin setae; endopod 3-segmented; segment 1 with 1 robust curved seta; segment 2 with 2 robust curved setae; segment 3 with 2 robust curved and 2 slender setae.

Maxilliped (Fig. 4F): phyllopodial, lamelliform, and 1-segmented. Trace of ancestral 2-segmented condition marked by the presence of outer incision, representing original segmentation boundary between former segments 1 and 2. Armature consisting of 10 elements, of which 5 bipinnate setae in apical position, 1 unipinnate seta inserted along inner margin together with 4 strong unipinnate stout spines. No trace of incision along inner margin.

P1-P3 with 3-segmented exopods and endopods. P4 with 3-segmented exopod and 2-segmented endopod. Intercoxal sclerites: boundary between intercoxa and basis not well defined at posterior surface of P2-P4 (Fig. 5B-D). P1-P3 praecoxa well developed, with outer spinule row. P4 praecoxa absent.

P1 (Fig. 5A): praecoxa and coxa with outer spinule row on anterior surface; one posterior row of thin spinules inserted close to coxo-basis boundary. Basis with 1 outer spiniform seta and 1 inner spine, with spinule rows along outer margin, between exopod and endopod and at the insertion of inner spine, respectively. Exopod about as long as endopod; exp-1 and -2 with 1 outer unipinnate spine; exp-3 with 2 unipinnate spines in apical position, and 1 apical and 1 subapical inner setae. Endopod: enp-1 unarmed, about as long as enp-2 and enp-3, wider than enp-2 and enp-3. Enp-2 cylindrical, with short inner seta inserted at the middle of segment. Enp-3 with 1 inner spine, 1 apical seta and 1 curved apical spine. Ornamentation as in Fig. 5A.

P2 (Fig. 5B): ornamentation of praecoxa and coxa as in P1. Basis with 1 outer spine, with spinule rows along outer margin, and between exopod and endopod. Exopod slightly longer than endopod; exopodal segments of about the same length; exp-1 and -2 with 1 outer unipinnate spine; exp-3 with 2 outer unipinnate spines, 1 apical unipinnate seta and 1 subapical long inner seta. Endopod: enp-1 unarmed; enp-2 with

1 naked inner seta; enp-3 with 1 spine and 1 long bipinnate seta in apical position, and 1 short bipinnate subapical seta. Ornamentation as in Fig. 5B.

P3 (Fig. 5C): ornamentation of praecoxa and coxa as in P2. Basis with short outer seta and spinule rows along outer margin and at the insertion of the endopod. Exopod distinctly longer than endopod. Exp-1 and -2 with 1 unipinnate outer spine; exp-3 with 2 unipinnate outer spines, 1 bipinnate apical seta and 1 long bipinnate subapical seta. Endopod: enp-1 and -2 unarmed; enp-3 with 1 spine and 2 bipinnate setae in apical position. Ornamentation as in Fig. 5C.

P4 (Fig. 5D): reduced in size, praecoxa absent, coxa and basis without ornamentation; basis with long outer naked seta; exopod and endopod about as long as half of remaining legs; the exopod only slightly longer than endopod. Exp-1 with 1 unipinnate outer spine; exp-2 unarmed; exp-3 with 1 bipinnate outer spine and 2 apical setae of different length. Endopod: enp-1 unarmed; enp-2 with 3 apical setae. Ornamentation as in Fig. 5D.

P5 (Figs 3A, 5E): free, with clear articulation to P5-bearing somite; right and left legs distinct; baseoendopod and exopod coalescent, incision marked original segmentation between them still present; basipodal outer seta present, exopodal armature consisting of 3 bipinnate setae and 1 stout spine: all elements in apical position; baseoendopod armed with 2 robust bipinnate setae, the outermost the longest.

P6 (Figs 3B, 5F): rudimentary, consisting of paired small chitinous lamellar plates not coalescent along medial margin, partially covering seminal receptacles. Armature consisting of 1 short naked spine with rounded tip on each leg.

**Male.** Body length, measured from tip of rostrum to posterior margin of caudal rami, from 370 to 541  $\mu\text{m}$ , with mean of 424  $\mu\text{m}$  based on 8 individuals. Sexual dimorphism in antennule, abdominal segmentation, P5, P6 and caudal setae morphology. Habitus, cephalosome (Figs 6A, 7A–B), sensilla and pore patterns as in female. Integument with surface pits. Urosome as in Figs 6B, 7C. Caudal rami with 6 setae (Fig. 6B). Anterolateral seta (II) as in female, posterolateral seta (III) setiform, not transformed (length seta/length caudal ramus: about 2) and bipinnate. Outer terminal seta (IV) as in female, inner terminal seta (V) not transformed, plumose and long, not articulated at base; terminal accessory seta (VI) and dorsal seta (VII) as in female (Fig. 6). Ornamentation and pore patterns as in female. Anal operculum as in female.

Antennule (Figs 6C, 7D–F): elongate, basically 10-segmented, last segment still showing a surface suture line only on anterior surface, indicating an incipient 11-segmented condition. Segment 1 with 1 ventral spinule row and 1 tube-pore (Fig. 8A). Segment 2 with tube-pore. Segment 4 represented by small U-shaped sclerite. Segment 6 the largest, sclerotized. Segment 8 elongate and transformed, moderately sclerotized, segment 9 distinct, not incorporated into segment 8, segment 10 derived by incomplete fusion between former segments 10 and 11 (Fig. 7D–F). Armature formula: 1-[1], 2-[9], 3-[8], 4-[2], 5-[7+(1 + ae)], 6-[2], 7-[2], 8-[0], 9-[1], 10-[10 + (1 + ae)]. Aesthetasc on segment 5 very large. Segment 8 with medial pointed protrusions as in Fig. 6C.



P1-P4 as in female; for morphological details of P1-P4 see Fig. 8B. P5 (Figs 6D, 9A): free, with clear articulation to P5-bearing somite; right and left legs coalescent; exopod clearly discernible but incorporated to basis: no trace of articulation between them observable, bearing 2 inner, 2 apical and 2 outer bipinnate setae; endopod discrete, distinctly 1-segmented, bearing 1 large leaf-like transformed seta and a spinule row along its free outer margin. Basipodal outer seta slender and naked, one pore near its insertion.

P6 (Figs 6E, 9B): right and left legs coalescent, forming a single linear lamellar plate, with no trace of incision between right and left P6; armature consisting of 2 spines and 1 outer seta.

***Phyllognathopus inexpectatus* Galassi & De Laurentiis, sp. n.**

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[http://species-id.net/wiki/Phyllognathopus\\_inexpectatus](http://species-id.net/wiki/Phyllognathopus_inexpectatus)

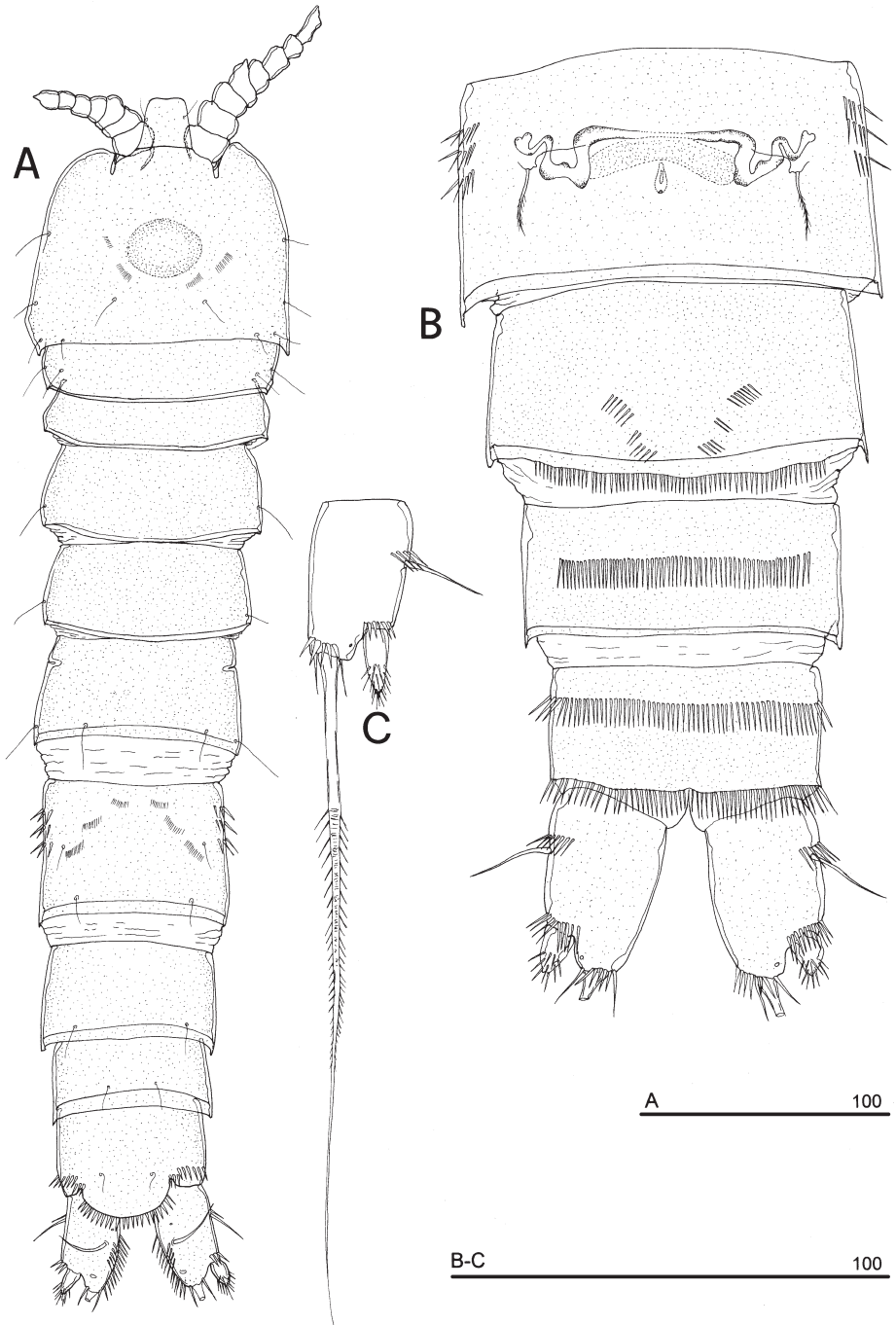
Figs 10–13

*Synonymy.* *Phyllognathopus* sp. (in Di Lorenzo et al. 2005).

**Type material.** ♀ holotype completely dissected and mounted in glycerine, deposited at the Natural History Museum, London (UK); January 2003, D. Cipriani coll.; 3 ♀♀ paratypes completely dissected and mounted in lactophenol; May 2003; January 2004. D. Cipriani coll..

**Type locality.** Mazzoccolo karstic spring (Latium, central Italy), coordinates: 41°15'17"N 13°27'08"E; Western Aurunci Mountains; 20 m a.s.l.; water temperature 13.5 ± 0.3 °C; pH 7.5 ± 0.1; O<sub>2</sub> 9.1 ± 0.9 mg/L (n = 11).

**Description.** FEMALE. Total body length, measured from tip of rostrum to posterior margin of caudal rami, 474 µm (holotype), 468 µm (paratypes mean value; n = 3). Body depigmented and eyeless. Habitus slender, no clear demarcation between prosome and urosome. Integument with surface pits, moderately sclerotized. Cephalosome subquadrate, with a dorsal rounded protuberance, hardly observable, referable to the dorsal integumental window (Fig. 10A). Couples of setule rows located on surface of cephalic shield. Rostrum elongate, subrectangular in shape, clearly articulated to the cephalosome; two dorsal sensilla laterally inserted on distal third, and one pore apically. Cephalosome and both thoracic and abdominal somites with cuticular ornamentation represented by reduced number of paired sensilla (Fig. 10A). First pedigerous somite free. Hyaline frills of cephalosome, somites bearing P1-P4 and urosome dorsally smooth. P5-bearing somite with large paired pores laterodorsally. Genital double-somite with three lateral spinule rows and three pairs of setule rows inserted dorsally. Female genital field located at the middle of genital double-somite. Genital apparatus simplified, copulatory pore located at half of the genital double-somite. Seminal receptacles laterally located and condensed close to the lamellar sixth legs.



**Figure 10.** *Phyllognathopus inexpectatus* sp. n. (♀). **A** habitus, dorsal view **B** abdomen, ventral view **C** caudal ramus, ventral view (scale bars in  $\mu\text{m}$ ).

Urosomites with smooth hyaline frill ventrally, except third urosomite (Fig. 10B). Last two urosomites with spinular fringe on proximal third; anal somite with distal continuous spinule row.

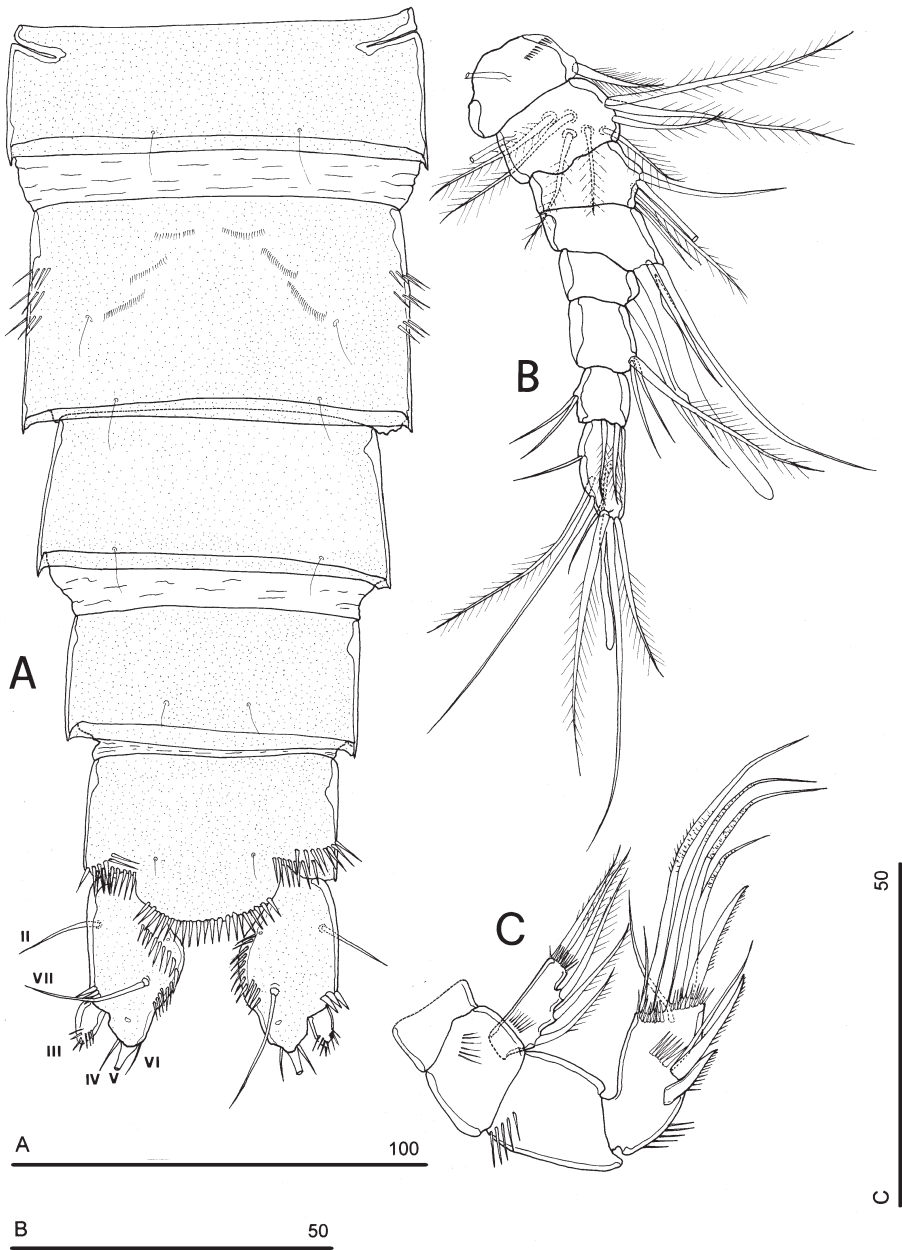
Anal somite with paired sensilla on dorsal surface (Fig. 11A), and two short spinule rows close to the anal operculum. Anal operculum rounded, protruding beyond insertion line of caudal rami and armed with strong spinules on free distal margin (Fig. 11A). Caudal rami rectangular with strongly expanded inner margin, slightly divergent, distinctly longer than wide (length/width ratio: about 1.5), with incomplete setal pattern (6 setae) (Figs 10C, 11A); anterolateral accessory seta (I) absent, anterolateral seta (II) smooth, inserted at proximal third of caudal ramus; posterolateral seta (III) inserted on distal third of ramus, transformed in a short and stout spiniform seta, with tuft of spinules apically. Outer terminal seta (IV) very short, thin, and naked, without articulation at base (Fig. 10C), distinctly shorter than caudal ramus; inner terminal seta (V) not transformed, very long, without articulation at base; terminal accessory seta (VI) as long as outer terminal seta, thin and naked; dorsal seta (VII) inserted at half of caudal ramus, about as long as caudal ramus or slightly shorter. A continuous spinule row along inner margin of caudal ramus and three spinule rows inserted close to the anterolateral seta (Figs 10B, 11A), at the basis of the posterolateral seta and at distal margin of ramus ventrally, respectively. Two pores are located dorsally on each caudal ramus, and one pore ventrally.

Antennule (Fig. 11B): short, 8-segmented. Segment 1 with ventral spinule row. Both segments 1 and 2 bearing long and flaccid tube-pores. Armature formula: 1-[1], 2-[8], 3-[5], 4-[1 + (1 + ae)], 5-[1], 6-[3], 7-[4], 8-[6 + (1 + ae)]. Aesthetasc on segment 4 very large, reaching about the last antennular segment.

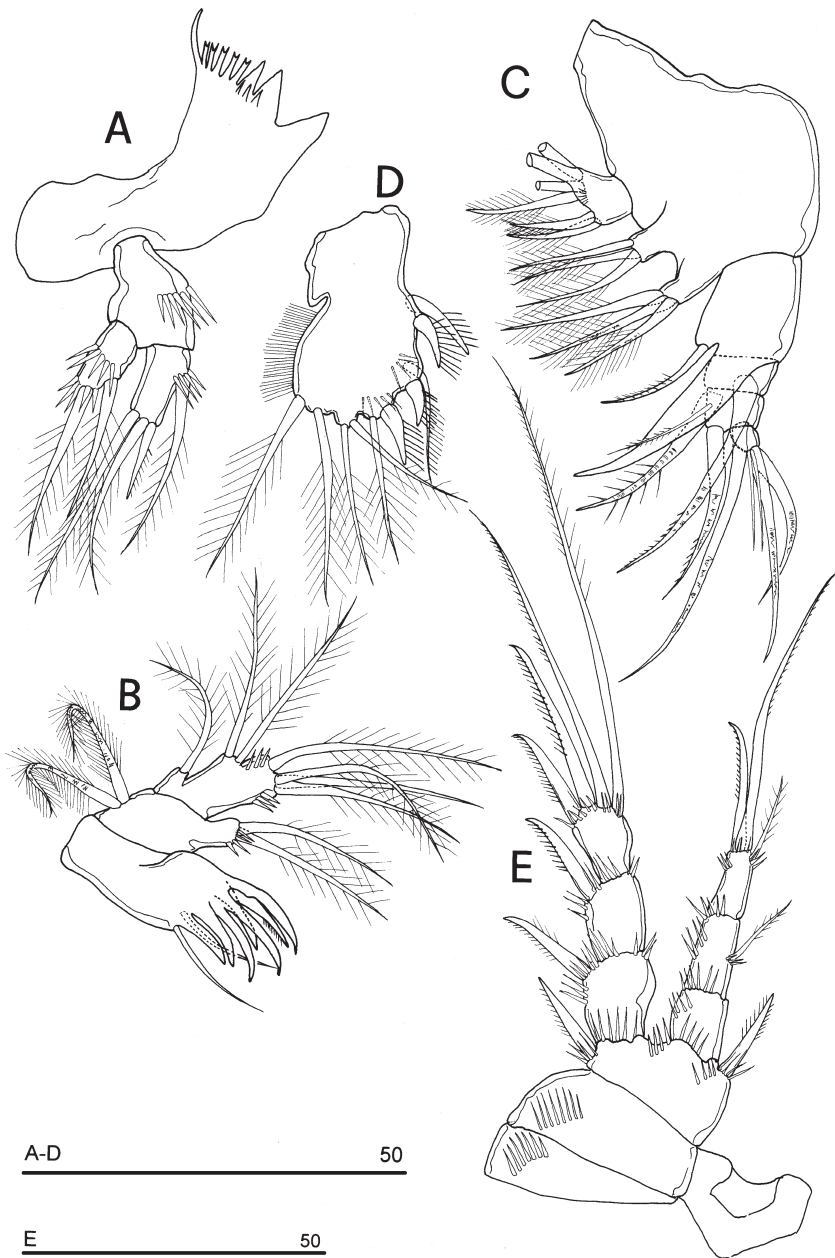
Antenna (Fig. 11C): coxa unarmed; basis with 1 transverse spinule row on surface; exopod 1-segmented, well-defined at base, with spinule row on surface, bearing 3 lateral unipinnate and 2 apical bipinnate setae; free endopod 2-segmented; both segments robust, of about the same length; segment 1 with inner spinule row; segment 2 with one inner and one surface spinule rows; armature consisting of 2 inner spines and 1 thin seta, 1 apical unipinnate spine, 4 geniculate setae, and 1 apical and 1 surface slender setae; two rows of spinules at outer corner and in subapical position on free distal margin, respectively.

Mandible (Fig. 12A): coxal gnathobase elongate, cutting edge with 3 large and coarse teeth, 5 smaller fringed teeth; naked seta at dorsal corner. Mandibular palp biramous, basis with inner strong spinule row, exopod with 1 apical and 1 inner bipinnate setae; endopod with 1 inner bipinnate, and 1 spiniform and 2 bipinnate apical setae. Ornamentation as in Fig. 12A.

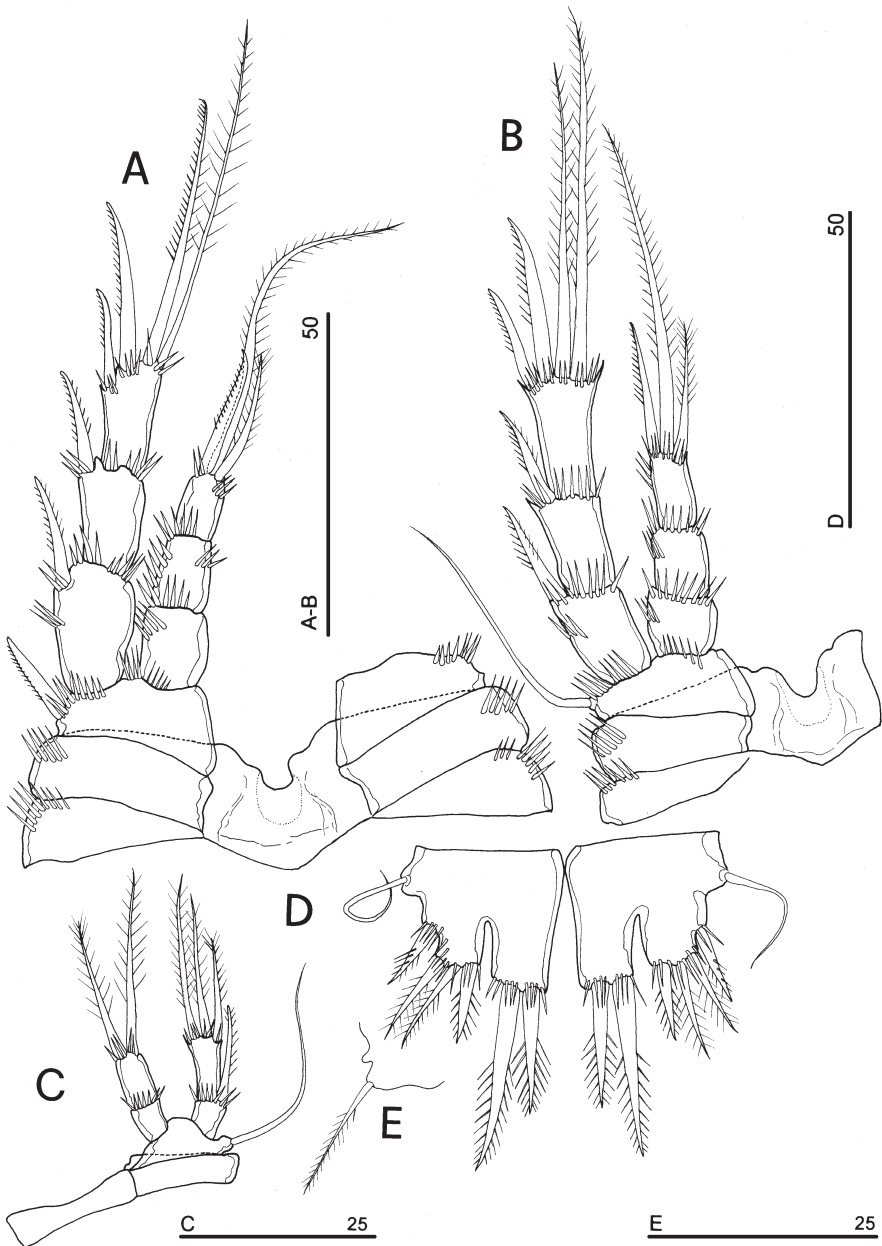
Maxillule (Fig. 12B): well developed arthrite incorporated into praecoxa, with 7 strong curved spines inserted on free distal margin, and 2 anterior surface setae. Proximal surface bipinnate seta inserted on tubercle absent (vs. present in *Phyllognathopus viguieri*, see Fig. 4D). Coxal epipodite represented by 2 setae; coxo-endite with 2 plumose setae. Exopod and endopod incorporated into basis, bearing 7 plumose setae.



**Figure 11.** *Phyllognathopus inexpectatus* sp. n. (♀) **A** urosome, dorsal view **B** antennule **C** antenna (scale bars in  $\mu\text{m}$ ).



**Figure 12.** *Phyllognathopus inexpectatus* sp. n. (♀). **A** mandible **B** maxillule **C** maxilla **D** maxilliped **E** P1 (scale bars in μm).



**Figure 13.** *Phyllognathopus inexpectatus* sp. n. (♀). **A** P2 **B** P3 **C** P4 **D** P5 **E** P6 (scale bars in  $\mu\text{m}$ ).

Maxilla (Fig. 12C): syncoxa with 3 endites. Proximal endite free, with 6 setae; medial and distal endites incorporated to syncoxa, each with 3 plumose setae, inserted as in Fig. 12C. Allobasis drawn out into a strong claw apparently smooth, accompanied by 1 robust and 2 thin setae; endopod 3-segmented; segment 1 with 1 robust curved seta; segment 2 with 2 robust curved setae; segment 3 with 2 robust curved and 2 slender setae.

Maxilliped (Fig. 12D): phyllopodial, lamelliform, 1-segmented, and slender than in *P. viguieri*. Trace of ancestral 2-segmented condition marked by the presence of outer incision, probably representing original segmentation boundary between segments 1 and 2. Armature consisting of 10 elements, of which 5 bipinnate setae in apical position, two of which with independent insertion, 1 unipinnate seta inserted along inner margin together with 4 strong unipinnate spines. No trace of incision along inner margin.

P1-P3 with 3-segmented exopods and endopods. P4 with 2-segmented exopod and endopod. Intercoxal sclerites: boundary between intercoxa and basis not well defined at posterior surface in P2-P4. P1-P3 praecoxa well developed, with 1 outer spinule row. P4 praecoxa absent.

P1 (Fig. 12E): praecoxa and coxa with outer spinule row on anterior surface. Basis with 1 outer spiniform seta and 1 inner spine; with spinule rows along outer margin, between exopod and endopod and at the insertion of inner spine, respectively. Exopod slightly longer than endopod: exp-1 and -2 with 1 outer unipinnate spine; exp-3 with 2 outer unipinnate spines and 2 setae, respectively inserted apically and subapically. Endopod: enp-1 unarmed, about as long as enp-2 and enp-3. Enp-2 cylindrical, with 1 inner short seta. Enp-3 with 1 inner bipinnate seta, 1 long unipinnate curved seta and 1 spiniform curved seta in apical position. Ornamentation as in Fig. 12E.

P2 (Fig. 13A): praecoxa and coxa as in P1; basis with 1 outer spiniform seta, with spinule rows along outer margin, and between exopod and endopod. Exopod distinctly longer than endopod; exopodal segments of about the same length; exp-1 and -2 with 1 outer bipinnate spine; exp-3 with 2 outer unipinnate spines, 1 apical unipinnate seta and 1 subapical long bipinnate seta. Endopod: enp-1 and -2 unarmed; enp-3 with 1 spine and 1 long bipinnate seta in apical position, 1 subapical short bipinnate seta. Ornamentation as in Fig. 13A.

P3 (Fig. 13B): ornamentation of praecoxa and coxa as in P1-P2. Basis with long outer seta and spinule rows along outer margin and at the insertion of the endopod. Exopod distinctly longer than endopod. Exp-1 and -2 with 1 outer bipinnate spine; exp-3 with 2 outer unipinnate spines, and 2 apical long bipinnate setae. Endopod: enp-1 and -2 unarmed; enp-3 with 1 unipinnate spine and 1 long bipinnate seta in apical position, 1 subapical bipinnate seta. Ornamentation as in Fig. 13B.

P4 (Fig. 13C): small sized, if compared to P1-P3; praecoxa absent, coxa and basis without ornamentation; basis with outer long and naked seta; exopod as long as endopod. Exp-1 with 1 outer long unipinnate spine; exp-2 with 1 outer long spine and 2

apical setae. Endopod: enp-1 unarmed; enp-2 with 2 apical plumose setae. Ornamentation as in Fig. 13C.

P5 (Fig. 13D): free, with clear articulation to P5-bearing somite; right and left legs distinct; baseoendopod and exopod coalescent, incision marked original segmentation still present; basipodal outer seta present, exopodal armature consisting of 1 outer spine, 2 apical short setae, of about the same length, and 1 apical spine; baseoendopod armed with 2 robust bipinnate setae, the outer the longest.

P6 (Fig. 13E): rudimentary, consisting of small paired chitinous lamellar plates not coalescent along medial margin, partially covering the genital field. Armature consisting of 1 long and slender bipinnate seta on each side.

Male unknown.

**Etymology.** The specific name derives from the Latin adjective *inexpectatus* which means “unexpected”, alluding to the surprising geographical location of the species, being the taxonomically related *Phyllognathopus* distributed in the Southern Hemisphere, and to the ecological finding of this species, which was collected from a large karstic aquifer in Central Italy, whereas all the other members of the genus are epigeal.

**Ecology.** At present knowledge the species is to be considered a stygobiotic species, collected from a karstic aquifer of the Western Aurunci Mountains (Latium) (Di Lorenzo et al. 2005). Although this aquifer is intensively fissured and karstified, with diffuse landforms of sinkholes and a discharge which is strictly linked to rainy events, stygobiont species were only sporadically present and represented by few individuals, due to the absence of a surface hydrological network, a landscape feature which is typical for coastal Mediterranean areas.

## Genus *Parbatocamptus* Dumont and Maas, 1988

### *Parbatocamptus jochenmartensi* Dumont and Maas, 1988

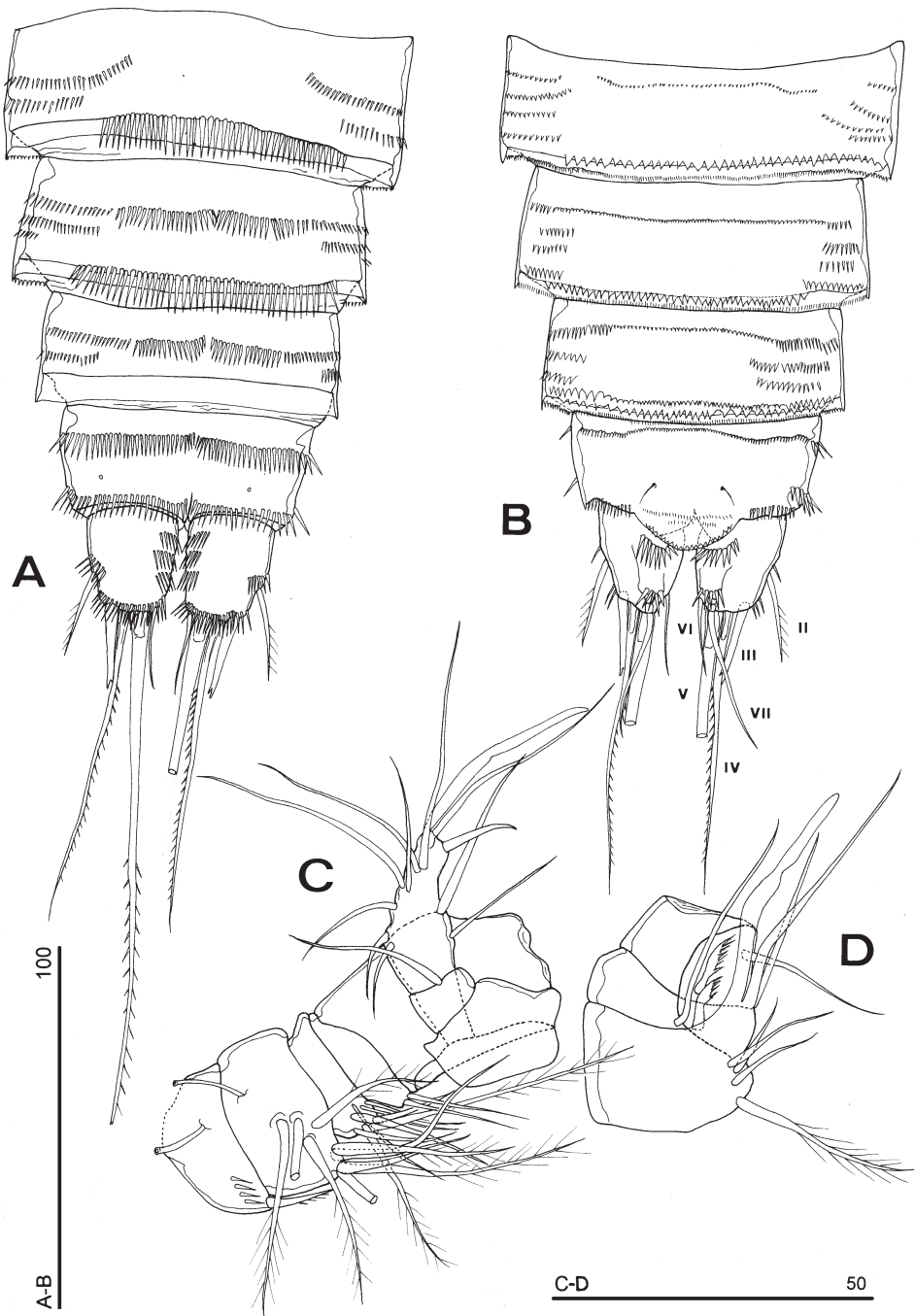
[http://species-id.net/wiki/Parbatocamptus\\_jochenmartensi](http://species-id.net/wiki/Parbatocamptus_jochenmartensi)

Figs 14–16

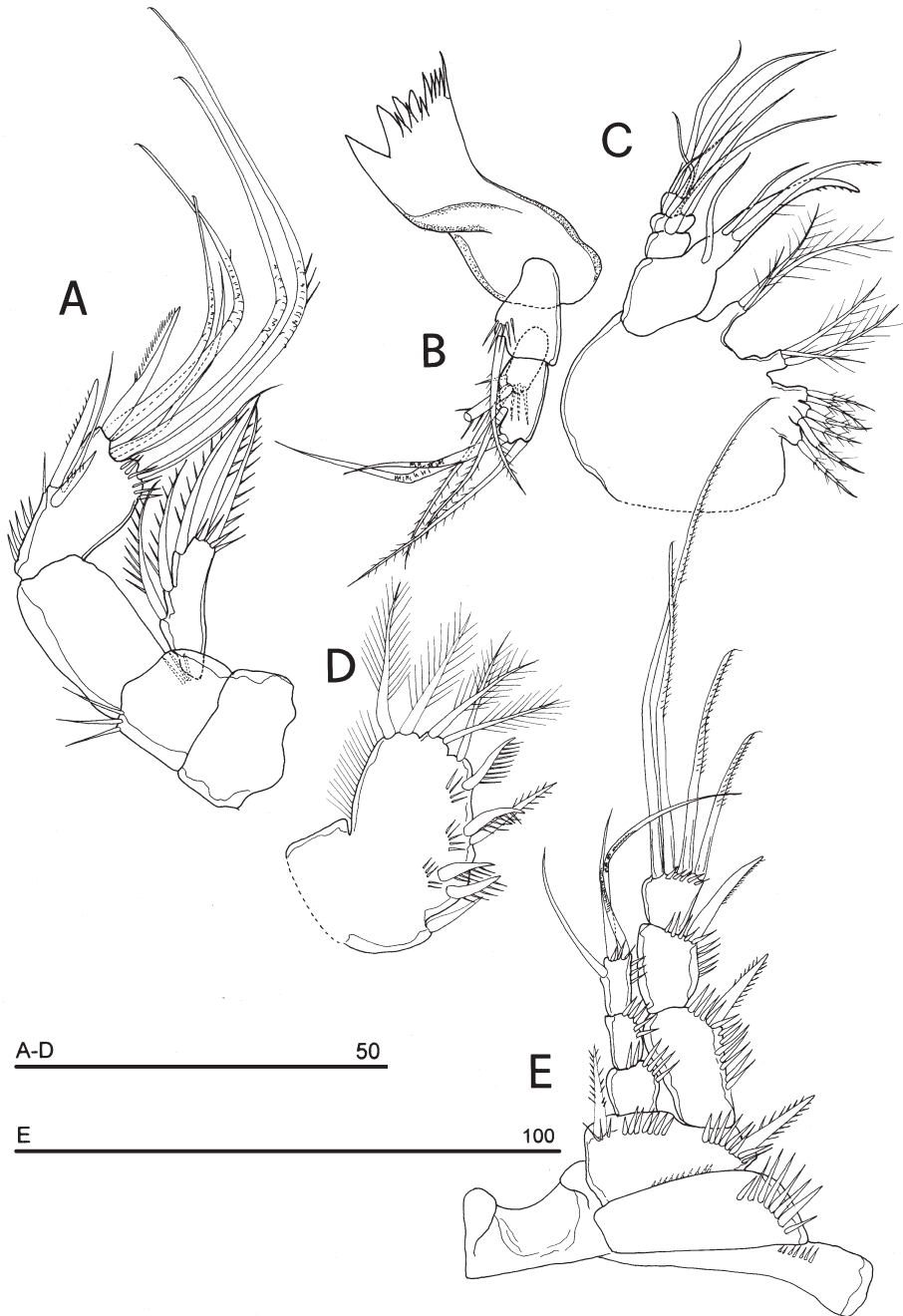
**Material examined.** ♂ holotype, completely dissected and mounted on slide labeled SMF 14657, deposited at the Senckenberg Museum (Germany).

**Emended diagnosis of the genus *Parbatocamptus* Dumont and Maas, 1988.** Phyllognathopodidae. Body flattened. First pedigerous somite free. Hyaline frills of abdominal somites ventrally smooth. Anal operculum prominent, subdistally crenulated. Caudal ramus subquadrate in ventral view, tapering on free distal margin in dorsal view, with 6 setae; anterolateral accessory seta (I) absent; dorsal seta inserted on spinulose protuberance closely located to free distal margin. Male antennule 10-segmented, with geniculation between segments 7 and 8; segment 9 distinct, as usual in the family; tube-pores on segments 1 and 2; aesthetascs on segments 5 and 10. Antenna: exopod 1-segmented, well-defined at base, bearing 3 lateral and 2 apical unipinnate setae; free endopod 2-segmented; both segments robust, of about the same length. Mandibular





**Figure 14.** *Parbatocamptus jochenmartensi* Dumont and Maas, 1988 (♂). **A** abdomen, ventral view **B** abdomen, dorsal view **C** antennule **D** antennule, detail of segments 5-7 (scale bars in  $\mu\text{m}$ ).



**Figure 15.** *Parbatocamptus jochenmartensi* Dumont and Maas, 1988 (♂). **A** antenna **B** mandible **C** maxilla **D** maxilliped **E** P1 (scale bars in  $\mu\text{m}$ ).

palp biramous; basis with inner long bipinnate seta; exopod with 2 setae; endopod with 2 apical geniculate setae; 1 inner and 1 subapical bipinnate setae. Maxillary syncoxa with 3 endites. Proximal endite quadrilobate, with 6 apical setae; medial and distal endites, each with 3 setae. Allobasis drawn out into a strong claw, accompanied by 1 curved and 2 normal setae; endopod 3-segmented; segment 1 with 1 robust curved seta; segment 2 with 2 robust curved setae; segment 3 with 2 robust curved and 2 slender setae. Maxilliped phyllopodial, lamelliform, 1-segmented. Clear trace of ancestral 2-segmented condition well discernible and marked by outer and inner incisions; armature consisting of 11 elements: 1 strong spine inserted on inner corner of former proximal segment; 4 spines and 6 setae along free distal margin of former second segment, two of which inserted on independent little knob. P1-P2 with 3-segmented exopods and endopods. P3-P4 with 3-segmented exopods and 2-segmented endopods. P1-P4 praecoxa present. P1 exp-1 long, about as long as exp-2 and -3 together. P1-P4 endopods distinctly shorter than exopods, not overreaching distal margin of exp-2. P2 enp-2 transformed: outer margin produced into a comb-like structure; P2 enp-3 with 1 apical strong curved spine and 1 apical seta. P5 with 2-segmented exopod; endopod incorporated to basis, forming a baseoendopod; suture line still observable on posterior surface; rudimentary intercoxal sclerite still discernible. P6 symmetrical, consisting of a well developed, deeply incised lamella, marking original division between left and right legs. Armature consisting of 1 outer seta and 2 inner short spines of different length.

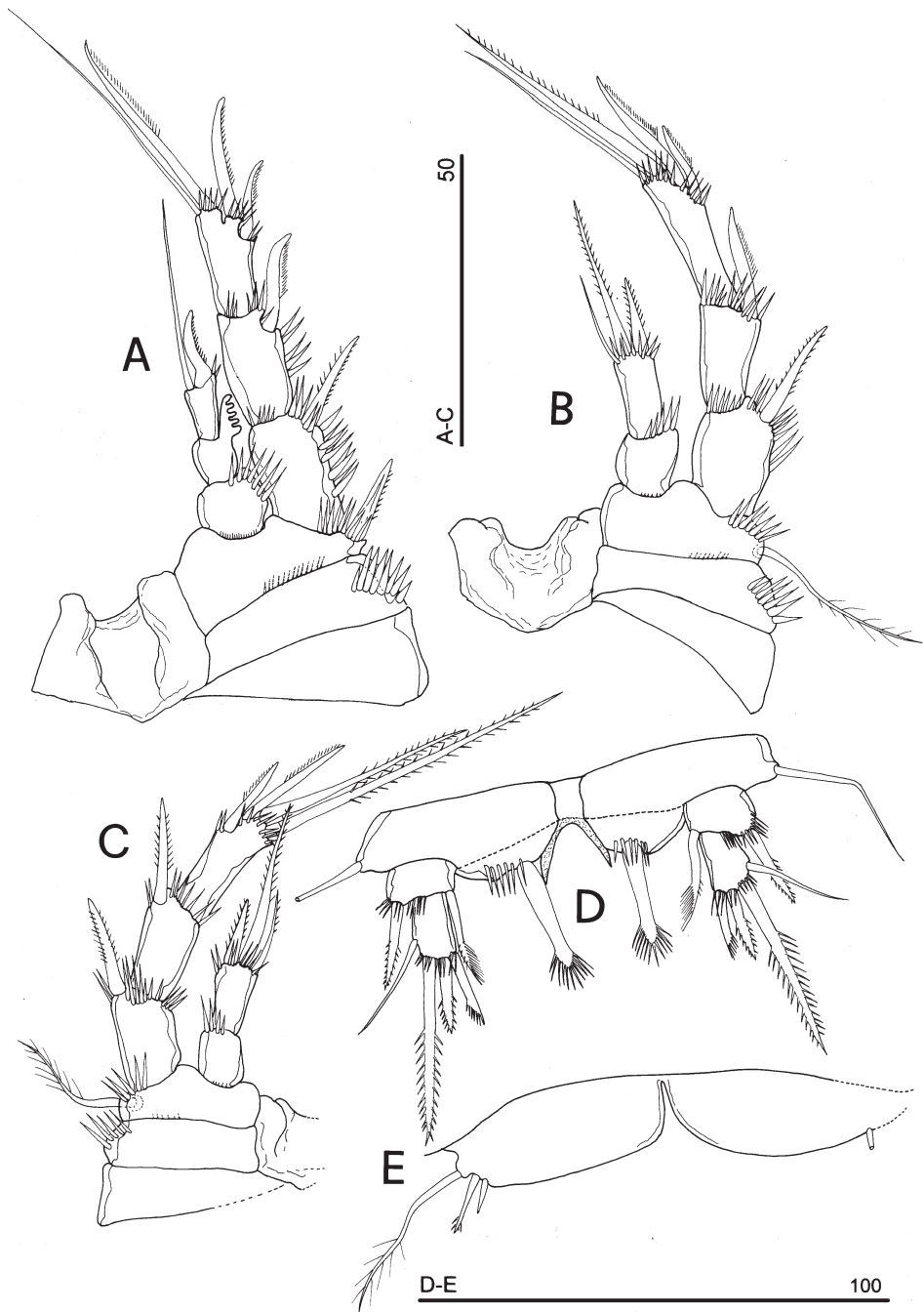
Female unknown.

**Supplementary description of the holotype.** The description deals with major morphological details, omitted or overlooked in the original description, and with improvements of observational errors.

Integumental pitting not detectable on the dissected holotype; integument well sclerotized.

Urosomites 3–5 with smooth hyaline frills ventrally (Fig. 14A); third and fourth urosomites with spinular fringe closely located to hyaline frill; fifth urosomite with surface spinule rows ventrally, anal somite with distal continuous spinule row. Other ornamentation as in Fig. 14A. Urosomites 3–5 with crenulated hyaline frills dorsally. Indented surface rows are observable on third, fourth and fifth urosomites dorsally (Fig. 14B). Anal somite with row of fine spinules on proximal third; spinule row at the insertion of each caudal ramus; paired dorsal sensilla. Anal operculum slightly protruding the insertion line of caudal rami, subdistally crenulated (Fig. 14B).

Caudal rami ventrally subquadrate, parallel, longer than wide (length/width ratio: about 1.4), with incomplete setal pattern (6 setae) (Fig. 14A–B). Anterolateral accessory seta (I) absent, anterolateral seta (II) unipinnate, inserted at second third of caudal ramus; posterolateral seta (III) inserted in subdistal position, transformed in spiniform bifid seta. Outer terminal seta (IV) well developed, unipinnate, with articulation at base; inner terminal seta (V) bipinnate and long; terminal accessory seta (VI) thin and naked, about as long as posterolateral seta; dorsal seta (VII) inserted on a spinulose knob, close to free distal margin of caudal ramus, distinctly longer than caudal ramus.



**Figure 16.** *Parbatocamptus jochenmartensi* Dumont and Maas, 1988 (♂). **A** P2 **B** P3 **C** P4 **D** P5 **E** P6 (scale bars in  $\mu\text{m}$ ).

A spinule row on proximal third of each caudal ramus dorsally; three spinule rows inserted ventrolaterally and one spinule row at distal margin of caudal ramus, ventrally.

Antennule (Fig. 14C): 10-segmented. Segment 1 with spinule row and tube-pore. Segment 2 with tube-pore. Segment 4 represented by small U-shaped sclerite. Segment 5 large, sclerotized (Fig. 14D). Segment 8 very large and transformed, moderately sclerotized, segment 9 short, discrete, segment 10 derived by complete fusion of former segments 10 and 11. Armature formula: 1-[1], 2-[9], 3-[8], 4-[2], 5-[5?(1 + ae)], 6-[2], 7-[2], 8-[0], 9-[1], 10-[10 + (1 + ae)].

Antenna (Fig. 15A): coxa unarmed; basis with 1 transverse spinule row on surface, a spinule row inserted on apical inner margin; exopod 1-segmented, well-defined at base, bearing 3 lateral and 2 apical unipinnate setae; free endopod 2-segmented; both segments robust, of about the same length; segment 1 naked; segment 2 with one inner spinule row, armature consisting of 2 inner spines and 1 seta, 1 apical unipinnate spine, 4 geniculate setae, 1 apical slender seta and 1 subapical tiny seta; a row of spinules at outer corner.

Mandible (Fig. 15B): coxal gnathobase elongate, cutting edge with 2 large and coarse teeth, three smaller teeth and row of tiny teeth; naked seta at dorsal corner. Mandibular palp biramous, basis with inner long bipinnate seta and spinule row, exopod with 2 bipinnate setae; endopod with 2 apical geniculate setae, 1 inner and 1 subapical bipinnate setae.

Maxillule not observable.

Maxilla (Fig. 15C): syncoxa with 3 endites fully incorporated to syncoxa. Proximal endite quadrilobate, with 6 setae; the first two distal lobes each bearing 2 setae; the proximal ones, each with 1 plumose seta; medial and distal endites, each with 3 bipinnate setae. Allobasis drawn out into a strong claw, distally spinulose, accompanied by 1 robust and curved seta and 2 naked setae, respectively; endopod 3-segmented; segment 1 with 1 robust curved seta; segment 2 with 2 robust curved setae; segment 3 with 2 curved and 2 slender setae.

Maxilliped (Fig. 15D): phyllopodial, lamelliform, 1-segmented. Clear trace of ancestral 2-segmented condition marked by the presence of both outer and inner incisions. Armature consisting of 11 elements: 1 strong spine inserted at inner corner of former segment 1; 4 strong unipinnate spines along inner margin, 1 bipinnate seta inserted along inner margin and 5 bipinnate setae in apical position.

P1-P2 with 3-segmented exopods and endopods. P3-P4 with 3-segmented exopods and 2-segmented endopods. P1-P4 praecoxa well developed. P1 (Fig. 15E): praecoxa and coxa with outer spinule row on anterior surface; one posterior row of thin spinules inserted on coxo-basis boundary. Basis with 1 outer spiniform seta and 1 inner spine, with spinule rows along outer margin, between exopod and endopod and at the insertion of inner spine, respectively. Endopod distinctly shorter than exopod, reaching about distal third of exp-2; exp-1 long, about as long as exp-2 and -3 together; exp-1 and -2 with 1 outer spine; exp-3 with 2 outer curved, unipinnate spines, and 1 apical and 1 subapical geniculate setae. Endopod: enp-1 unarmed, about as long as enp-2 and enp-3, wider than enp-2 and enp-3. Enp-2 cylindrical, unarmed. Enp-3 with 1 inner long seta, and 2 apical geniculate setae of different length. Ornamentation as in Fig. 15E.

P2 (Fig. 16A): praecoxa unornamented, coxa as in P1 and P2. Basis with 1 outer spiniform seta, with spinule rows along outer margin. Exopod distinctly longer than endopod; endopod reaching about the proximal half of exopodal segment 2; exopodal segments of about the same length; exp-1 and -2 with 1 outer spine; exp-3 with 2 outer unipinnate spines, 1 apical fringed seta and 1 subapical long and slender seta. Endopod: enp-1 unarmed; enp 2 transformed, with outer strong comb-like process; enp-3 with 1 transformed spine and 1 long slender and naked seta in apical position. Ornamentation as in Fig. 16A.

P3 (Fig. 16B): praecoxa unornamented; ornamentation of coxa as in P1 and P2. Basis with long outer plumose seta and spinule rows along outer margin and at the insertion of the endopod. Exopod distinctly longer than endopod; endopod reaching about half of exp-2. Exp-1 and -2 with 1 outer spine; exp-3 with 2 outer fringed spines, 1 apical unipinnate seta and 1 subapical long and naked seta. Endopod: enp-1 unarmed; enp-2 with 2 apical spinulose setae and 1 subapical thin and naked seta. Ornamentation as in Fig. 16B.

P4 (Fig. 16C): slightly smaller than the other swimming legs, praecoxa present, unornamented; coxa with outer spinule row, one posterior row of thin spinules inserted on coxo-basis boundary; basis with outer plumose seta, and spinule rows along outer margin and at the insertion of endopod; exopod distinctly longer than endopod. Exp-1 and -2 with 1 outer bipinnate spine; exp-3 with 2 outer unipinnate spines and 2 apical bipinnate setae of different length. Endopod: enp-1 unarmed; enp-2 with 1 spine and 1 seta in apical position, and 1 subapical short unipinnate seta. Ornamentation as in Fig. 16C.

P5 (Fig. 16D): free, with clear articulation to P5-bearing somite; right and left legs distinct, trace of intercoxal sclerite present but hardly observable, with coxo-basis protrusions; exopod discrete, 2-segmented, segment 1 with 1 outer spine and 1 inner seta distally fringed; segment 2 with 4 elements: 1 outer slender and naked seta, 1 apical long spine, 1 medial short spine and 1 short seta distally crested; endopod incorporated to basis forming a baseoendopod, trace of original segmentation still recognizable on posterior surface (Fig. 16D); rudimentary endopod 1-segmented, bearing 1 strong spiniform element, crested on its distal margin. Basipodal outer seta slender and naked.

P6 (Fig. 16E): well developed, symmetrical, right and left legs distinct, deep medial incision marking boundary between legs; armature consisting of 1 outer long, bipinnate seta and two inner short spines, the innermost the shortest.

***Neophyllognathopus Galassi & De Laurentiis, gen. n.***

urn:lsid:zoobank.org:act:D06A6D6C-129B-4142-8C3C-B220A4E7CFAD

<http://species-id.net/wiki/Neophyllognathopus>

**Diagnosis.** Phyllognathopodidae. Habitus slightly dorsoventrally flattened with no clear demarcation between prosome and urosome. Integumental dorsal window on cephalosome not confirmed. Integument without surface pits, moderately sclerotized. Cepha-

losome rounded; rostrum elongate, clearly articulated to cephalosome. Cephalosome and both thoracic and abdominal somites with cuticular ornamentation represented by dorsal sensilla. First pedigerous somite free. Hyaline frills of cephalosome, somites bearing P1-P4 plain both dorsally and ventrally. P5-bearing somite with large paired pores laterodorsally. Sexual dimorphism in antennule, P5, P6, urosomal segmentation and ornamentation, and morphology of anal operculum. Female first and second abdominal somites fused forming the genital double-somite. Female urosomal segments with plain hyaline frills ventrally. Female genital apparatus simplified; copulatory pore located at the end of the proximal third of the genital double-somite. Seminal receptacles laterally located and condensed close to the lamellar sixth legs. Male urosome with different arrangement of hyaline frill ornamentations: urosome consisting of 6 segments, second urosomite with indented hyaline frill, third and fourth urosomites with deep ventral sockets; socket on third urosomite plicate, with smooth free distal margin, and 2 setules laterally inserted close to the socket opening; socket on fourth urosomite with free distal margin ornamented by strong and long spinules, covering the opening; fifth urosomite with indented hyaline frill. Anal somite with paired sensilla on dorsal side. Anal operculum protruding free distal margin of anal somite and extruded in strong spinular processes. Sexual dimorphism in the number of spinular processes of anal operculum (3 in females vs. 4 in males; and, in general, anal operculum in male more armed than in female). Caudal rami sub-quadrate, with incomplete setal pattern (6 setae). Dorsal seta inserted on distal third of caudal ramus. Antennule: 8-segmented in female, basically 9-segmented in male; geniculation between segments 7 and 8; penultimate and last segments, each with suture line marking original segmentation between former segments 8 and 9, and 10 and 11, respectively. Long tube-pores on segments 1 and 2 in both sexes. Antenna: armature of the second endopodal segment as in *Phyllognathopus* and *Parbatocamptus*, consisting of 10 elements. Exopod 1-segmented, with 3 lateral and 2 apical setae. Mandible: mandibular palp biramous, basis with inner spinule row, exopod with 1 apical and 1 inner setae; endopod with 1 inner, 1 subapical and 2 apical setae. Armature of maxillule and maxilla as in *Phyllognathopus*. Maxilliped: phyllopodial, lamelliform, 1-segmented. Clear trace of ancestral 2-segmented condition marked by the presence of outer and inner incisions as in *Parbatocamptus*. Armature consisting of 11 elements: 1 strong spine inserted at inner corner of former segment 1; 4 spines and 1 spiniform short seta inserted along inner margin, 5 bipinnate setae in apical position, armature topology basically referable to that of *Parbatocamptus*.

P1-P3 with 3-segmented exopods and endopods. P4 with 2-segmented exopod and endopod. P1-P3 praecoxa well developed. P1 exopod and endopod of about the same length; P2-P3 endopods shorter than exopods, reaching about tip of exp-2. P4 small - sized, praecoxa missing. Female P5: free, with clear articulation to P5-bearing somite; right and left legs distinct; baseoendopod and exopod coalescent, deep incision marking original segmentation between them; endopodal lobe well developed, elongate, longer than exopodal lobe, rectangular in shape, bearing 1 long pinnate seta, subdistally inserted, close to outer margin and a spinule row apically inserted. Exopodal lobe wide, fully incorporated into baseoendopod; exopodal armature consisting of 4

elements, the outermost bipinnate seta inserted in subdistal position, and three apical elements: 2 spinulose and 1 short setae; basipodal outer seta present. Female P6 rudimentary, each leg defined by a small cuticular lateral plate bearing a short, naked seta with rounded tip. Male P5: free, with clear articulation to P5-bearing somite; right and left legs separate, intercoxal sclerite rudimentary, but still discernible. Basis of each leg expanded, endopod strongly transformed, consisting of a sclerotized and strong protrusion articulated to basis. Endopodal seta bipinnate, inserted on posterior surface of the endopod, close to its articulation to basis. Exopod distinct, clearly articulated to basis, wide and short, rectangular in shape, representing most part of the free distal margin of each leg; exopodal armature consisting of 6 elements, the innermost spiniform seta curved inward. Male P6: right and left legs distinct but closely adjacent to each other along their medial margin, and symmetrical; each leg consisting of a well developed lamellar plate, with spinule row on the anterior surface; armature consisting of 2 inner spines of different length and 1 outer seta.

**Type species by monotypy.** *Phyllognathopus bassoti* Rouch, 1972 = *Neophyllognathopus bassoti* (Rouch, 1972), comb. n.

**Etymology.** The genus name is derived from the type genus *Phyllognathopus* and the Latinised Greek prefix *νέος* which means “new”, referring to the new position of *Phyllognathopus bassoti* in the systematics of the family Phyllognathopodidae.

***Neophyllognathopus bassoti* (Rouch, 1972), comb. n.**

[http://species-id.net/wiki/Neophyllognathopus\\_bassoti](http://species-id.net/wiki/Neophyllognathopus_bassoti)

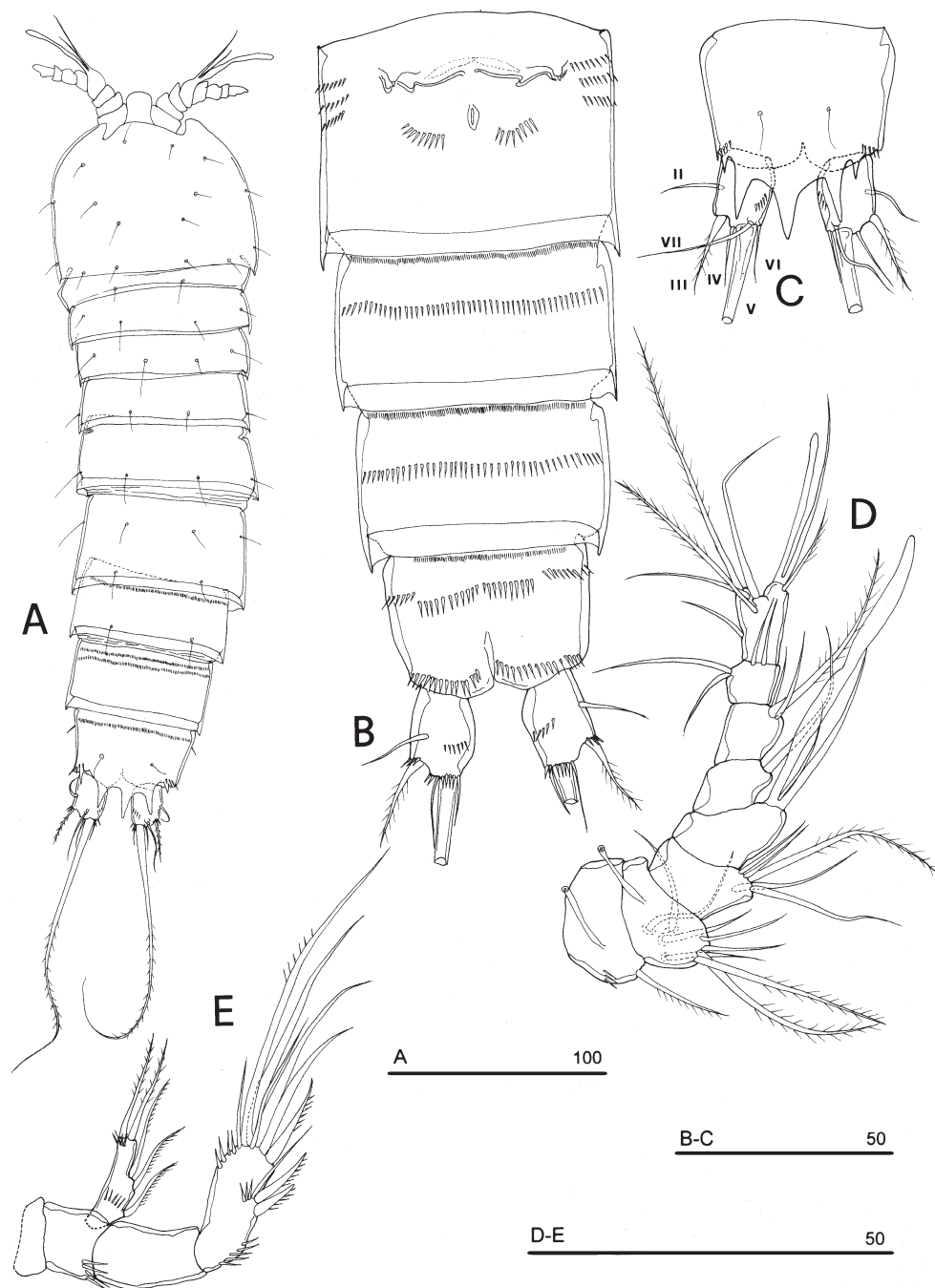
Figs 17–25

**Neotype designation.** Female neotype completely dissected and mounted in polyvinyl lactophenol, deposited at the Natural History Museum, London (reg. No. NHM.2008. neotype). Other material: 5 ♀♀ and 3 ♂♂ mounted on slides, 5 ♀♀ and 5 ♂♂ processed for SEM; India, 7 January 1999, Y. Ranga Reddy coll.; 1 ♀, slide code 66/49, 1 ♀, slide code 66/53, 1 ♂, slide code 66/55, Santa Fe, Bantayan island, Pooc, Philippines, V. Cottarelli coll. (see Bruno and Cottarelli 1999, for locality details).

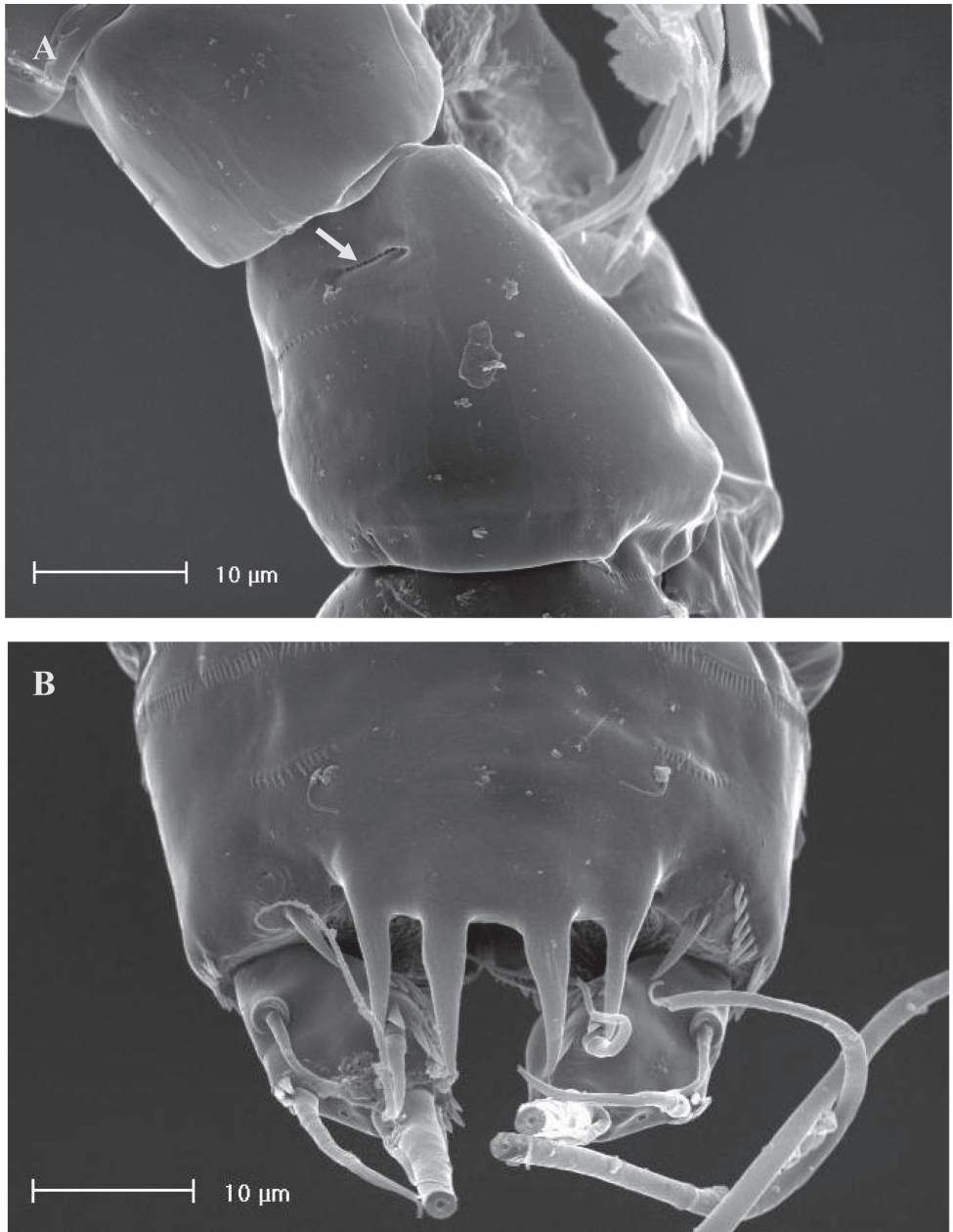
*Neophyllognathopus bassoti* is proposed herein as new combination for *Phyllognathopus bassoti* assigned by Rouch (1972) in the original description to the genus *Phyllognathopus*. According to ICZN (2000), a neotype may be designated when *no name-bearing type specimen (i.e. holotype, lectotype, syntype or prior neotype) is believed to be extant and an author considers that a name-bearing type is necessary to define the nominal taxon objectively* (Article 75.1). Article 75.3 asks also for *qualifying conditions* for the establishment of a neotype; among them: *.... a statement that it is designated with the express purpose of clarifying the taxonomic status or the type locality of a nominal taxon* (Article 75.3.1), and *the author’s reasons for believing the name-bearing type specimen(s) ... to be lost or destroyed, and the steps that have been taken to trace it or them* (Article 73.3.4).

The specimens on which Rouch (1972) based the original description of *P. bassoti* no longer exist, and most part of the Rouch’s collection has been lost (Rouch, in litt.).



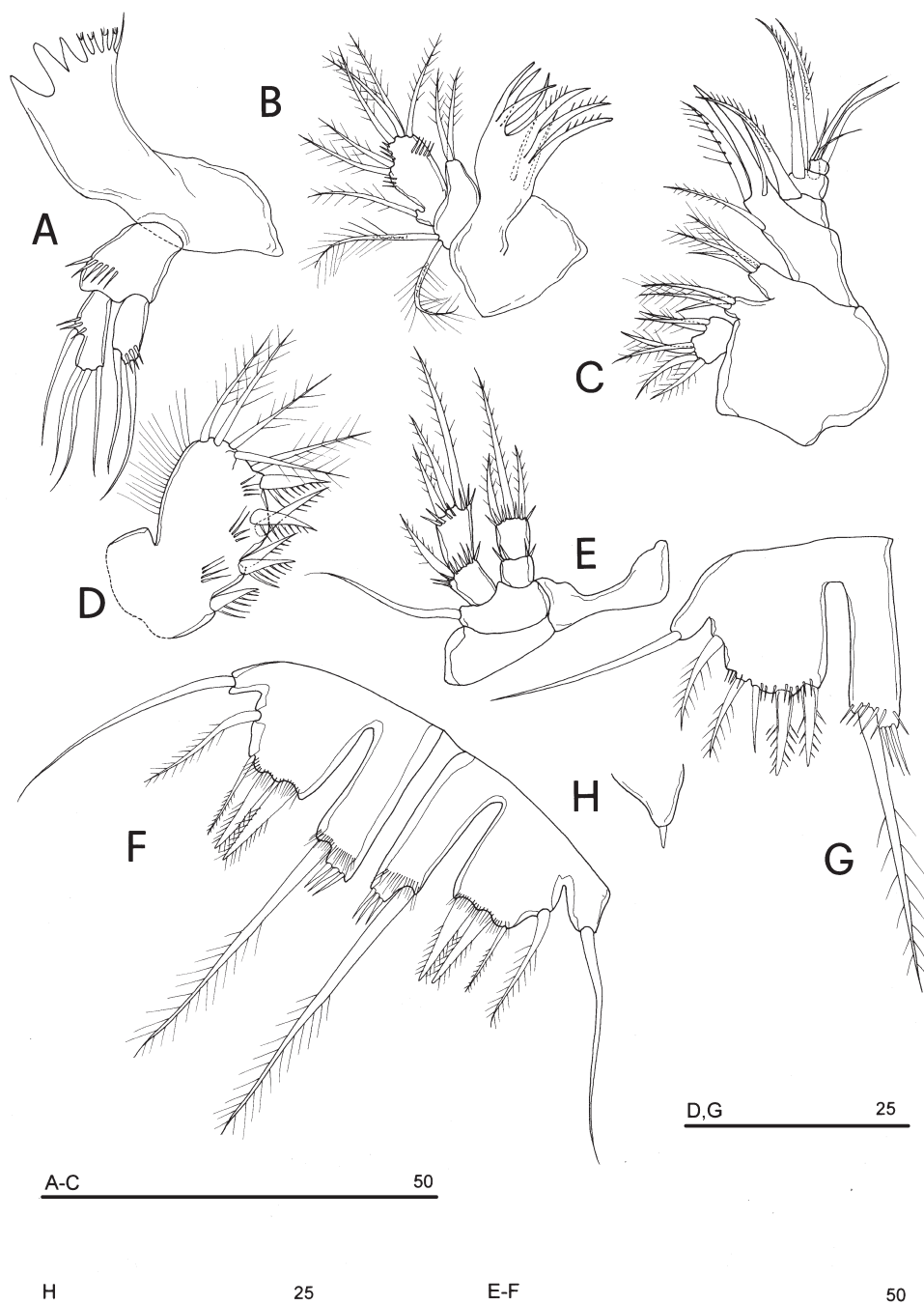


**Figure 17.** *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♀). **A** habitus, dorsal view **B** abdomen, ventral view **C** anal somite and operculum **D** antennule **E** antenna (scale bars in  $\mu\text{m}$ ).

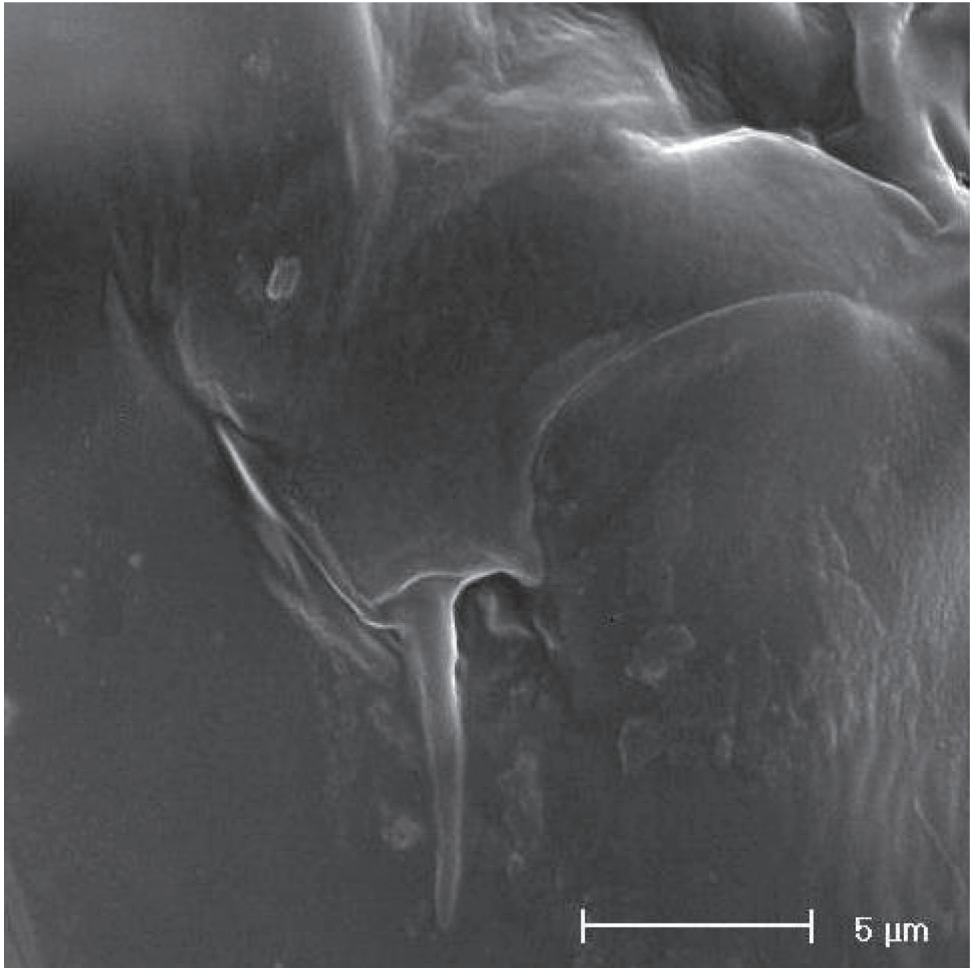


**Figure 18.** SEM micrographs of *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂). **A** lateral large pore (arrowed) on P5-bearing somite **B** anal somite and operculum.

Consequently, the ICZN (2000) recommendation 75A cannot be met, because no extant paratypes or paralectotypes, nor topotypic specimens are available, in order to select among them a neotype. The need to clarify the taxonomic status of this species, which is ranked herein to a new genus, imposed to follow another formal procedure, which, if



**Figure 19.** *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♀) **A** mandible **B** maxillule **C** maxilla **D** maxilliped **E** P4 **F** P5 **G** P5 (anomaly) **H** P6 (scale bars in  $\mu\text{m}$ ).

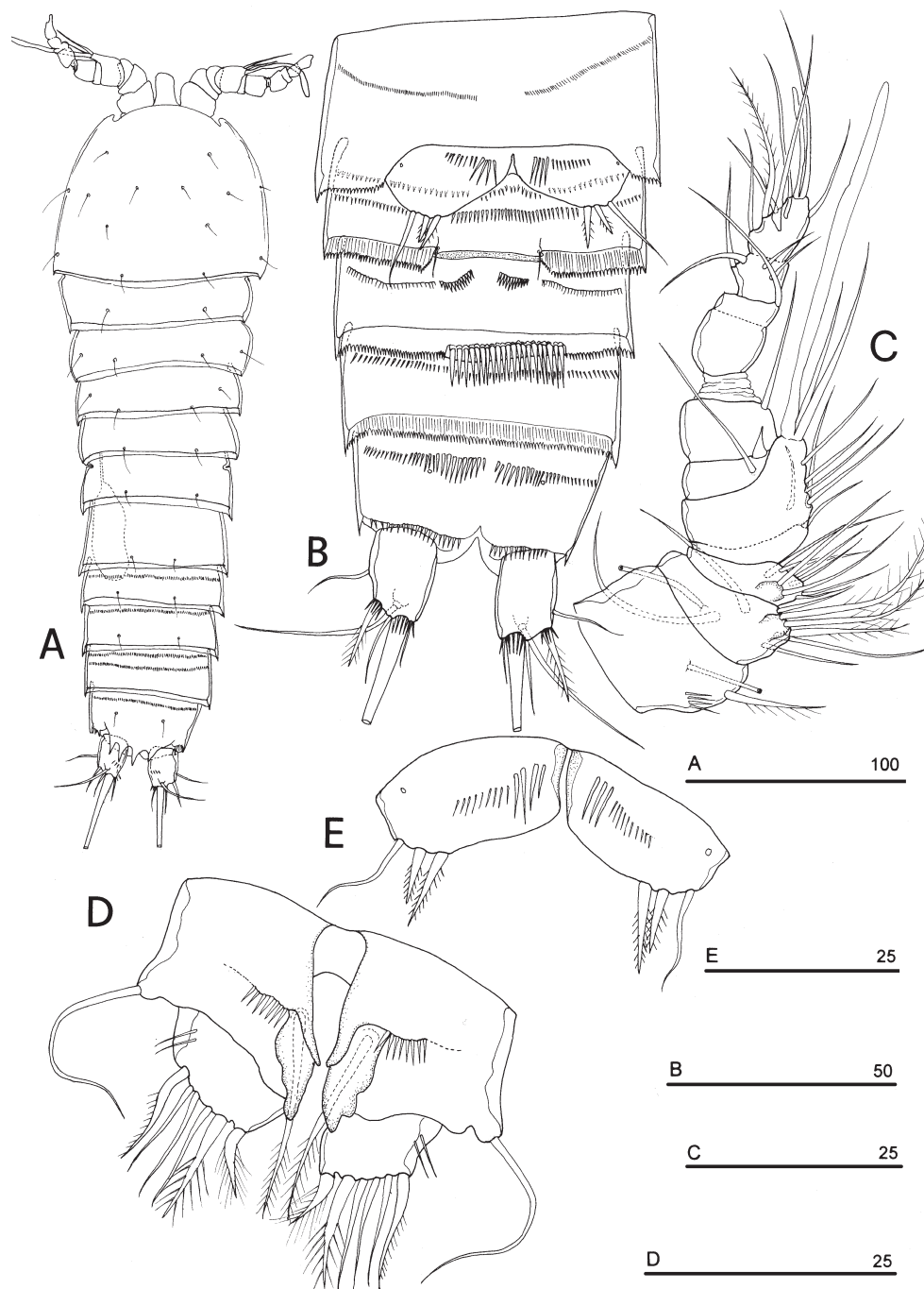


**Figure 20.** SEM micrograph of *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♀): P6.

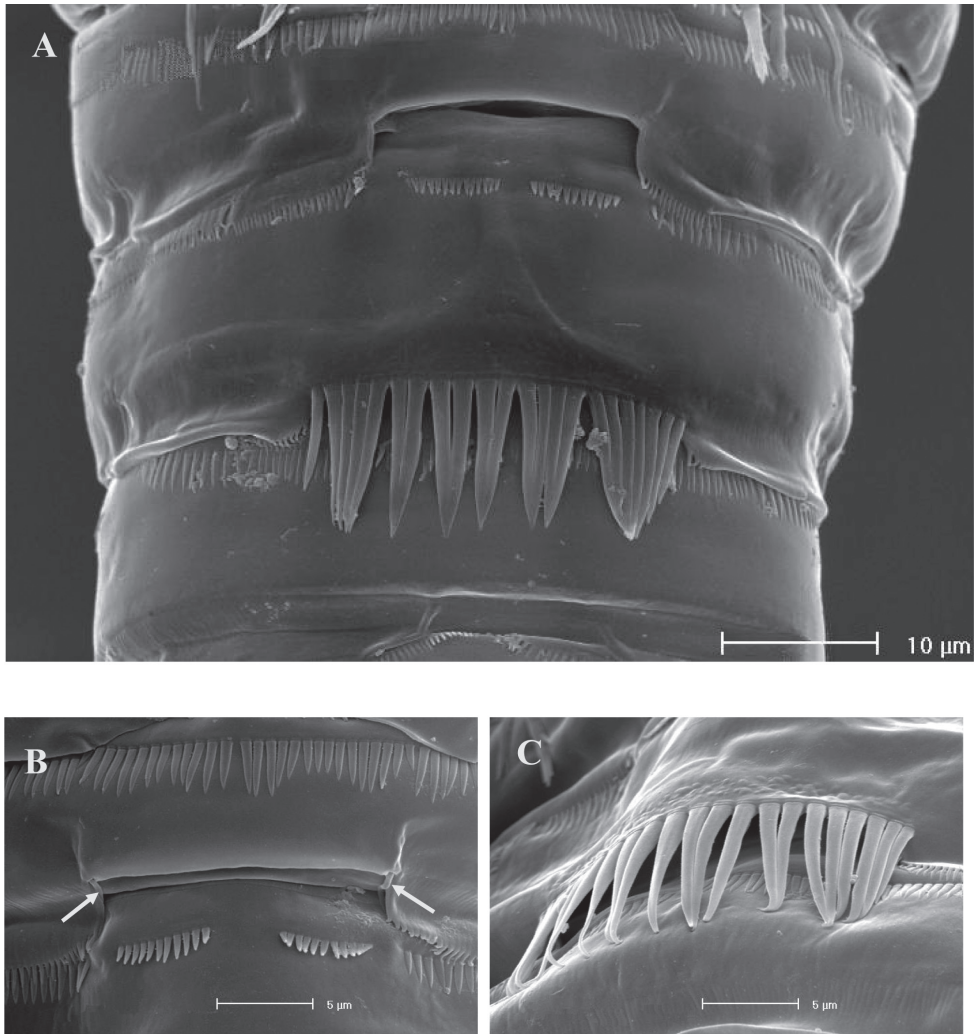
not completely fulfills the ICZN rules (Article 75.3.6), is accepted by the Code (Article 76.3): *the place of origin of the neotype becomes the type locality of the nominal species-group taxon, despite any previously published statement of the type locality*. Following these arguments, a consistent population from India has been selected to establish the new genus, together with additional material from Indonesia. It is relevant to observe that other researchers have given consensus (Bruno and Cottarelli 1999, Karanovic and Ranga Reddy 2004) on the attribution of both populations to the species *Phyllognathopus bassoti* described by Rouch (1972) from the Lake Wisdom (New Guinea).

**Neotype locality.** India, Andhra Pradesh, town of Guntur, Brindavan Gardens, domestic water reservoir filled by a freshwater bore well; coordinates: approx. 16°18'N 80°29'E (see Karanovic and Ranga Reddy 2004, for more details).

**Description based on the designed neotype.** FEMALE NEOTYPE. Body length, measured from tip of rostrum to posterior margin of caudal rami, 348 μm. Habitus slight-

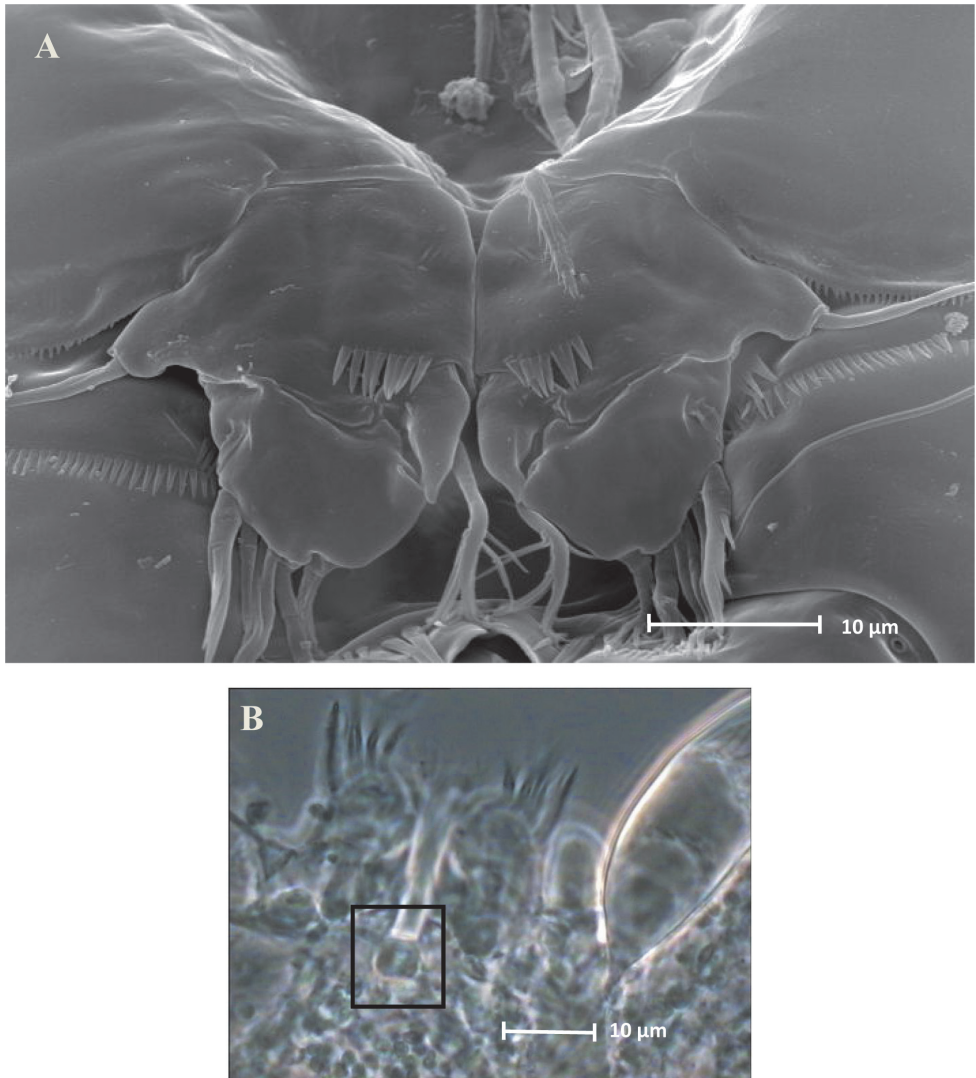


**Figure 21.** *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂). **A** habitus, dorsal view **B** abdomen, ventral view **C** antennule **D** P5 **E** P6 (scale bars in  $\mu\text{m}$ ).



**Figure 22.** SEM micrographs of *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂). **A** general view of the urosomal sockets **B** plicate socket on third urosomite (paired lateral setules arrowed) **C** socket on fourth urosomite covered by strong spinules.

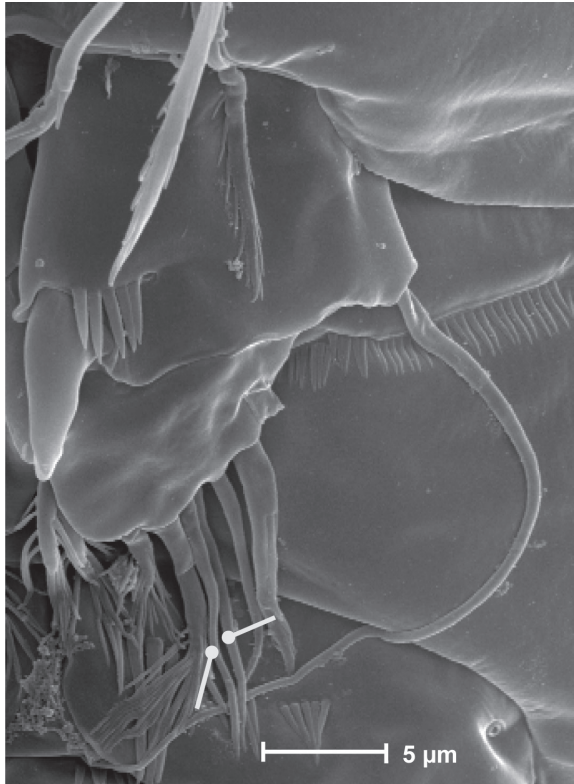
ly dorsoventrally flattened (Fig. 17A), with no clear demarcation between prosome and urosome. Body depigmented and eyeless. Integumental dorsal window on cephalosome not confirmed. First pedigerous somite free. Integument without surface pits, moderately sclerotized. Cephalosome rounded; rostrum elongate, clearly articulated to cephalosome. Hyaline frills of cephalosome, somites bearing P1-P4 and urosome plain both dorsally and ventrally (Fig. 17 A, B). Cephalosome and both thoracic and abdominal somites (except fourth urosomite) with cuticular ornamentation represented by dorsal sensilla. P5-bearing somite with lateral paired and large pores (Fig. 18A). Female genital field located between first and second third of genital double-somite. Genital apparatus simplified; copulatory



**Figure 23.** *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂). **A** SEM micrograph of P5 **B** contrast phase micrograph of P5 rudimentary intercoxa (framed).

pore located at half of genital double-somite. Seminal receptacles laterally located and condensed close to the lamellar sixth legs. Three spinular processes on free distal margin of anal operculum (Figs 17A–C). Caudal rami sub-quadrangle, with incomplete setal pattern (6 setae). Dorsal seta inserted close to free distal margin of caudal ramus (Fig. 17C).

Antennule (Fig. 17D): consisting of 8 segments, segments 1 and 2 with long tube-pores. Armature formula: 1-[1], 2-[8], 3-[5], 4-[1 + (1 + ae)], 5-[1], 6-[3], 7-[4], 8-[6 + (1 + ae)]. Aesthetasc on segment 4 very large and long, well overreaching the last antennular segment.



**Figure 24.** SEM micrograph of *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂) P5, detail (white lines showing setae III and IV, which are closely adherent to each other and hardly discernible as distinct under contrast phase microscope).

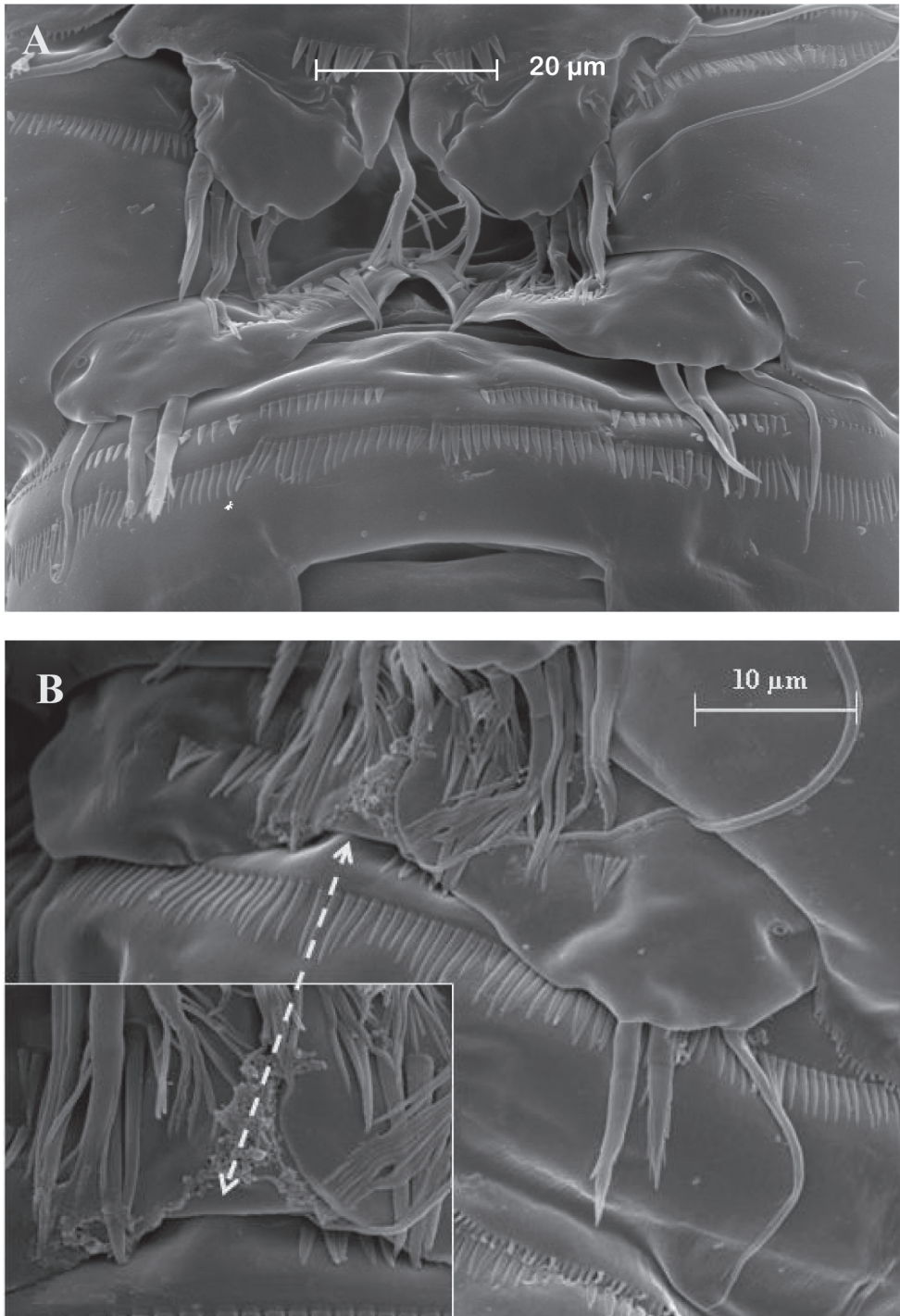
Antenna (Fig. 17E): exopod and armature of the second endopodal segment as in *Phyllognathopus* and *Parbatocamptus*. Exopod 1-segmented, with 3 lateral and 2 apical setae.

Mandible (Fig. 19A): mandibular palp biramous, basis with inner spinule row, exopod with 1 apical and 1 inner setae; endopod with 1 inner, 1 subapical and 2 apical setae. Armature of maxillule (Fig. 19B) and maxilla (Fig. 19C) as in *Phyllognathopus*.

Maxilliped (Fig. 19D): phyllopodial, lamelliform, 1-segmented. Clear trace of ancestral 2-segmented condition marked by the presence of both outer and inner incisions. Armature consisting of 11 elements: 1 strong spine inserted at inner corner of former segment 1; 4 spines and 1 spiniform short seta inserted along inner margin, 5 bipinnate setae in apical position, armature topology basically referable to that of *Parbatocamptus*.

P1-P3 with 3-segmented exopods and endopods. P4 with 2-segmented exopod and endopod. P1-P3 praecoxa well developed. P4 praecoxa absent (Fig. 19E). P1 exopod of about the same length of endopod. P2-P3 exopods longer than endopods, endopod not overreaching exp-2, fitting the original description (Rouch





**Figure 25.** SEM micrographs of *Neophyllognathopus bassoti* (Rouch, 1972), comb. n. (♂). **A** P5 and P6 **B** detail of P6 showing an interconnecting lamella (rudimentary intercoxa?) between right and left legs arrowed.

1972) and the subsequent ones (Bruno and Cottarelli 1999, Karanovic and Ranga Reddy 2004).

P5 (Fig. 19F, G): free, with clear articulation to P5-bearing somite; right and left legs separate; baseoendopod and exopod coalescent, deep incision marking original segmentation between them; endopodal lobe well developed, elongate, rectangular in shape, longer than exopod, bearing 1 long pinnate seta, subdistally inserted, close to outer margin, and 2 spinule rows, the proximal one composed by tiny elements, the distal one of long spinules; exopodal lobe well discernible, with armature consisting of 4 (rarely 5 elements, observed in only one female) elements, the outermost seta inserted in subdistal outer position, the remaining ones in apical position; the outer apical seta slender and bipinnate, the remaining two spiniform. Basipodal outer seta present.

P6 (Figs 19H, 20): rudimentary, consisting of small paired chitinous lamellar plates not coalescent along medial margin, partially covering seminal receptacles. Armature consisting of 1 short smooth spine with rounded tip on each side.

**Male.** No marked sexual dimorphism in body size. Body length, measured from tip of rostrum to posterior margin of caudal rami, 335  $\mu\text{m}$ . Rostrum and ornamentation of cephalosome as in female (Fig. 21A). Male urosome consisting of 6 segments (Fig. 21A), third and fourth urosomites with deep ventral sockets (Figs 21B, 22A); socket on third urosomite plicate, with smooth free distal margins, and 2 setules laterally inserted close to the socket opening (Fig. 22B); socket on fourth urosomite with ornamented anterior margin, armed by strong spinules covering the opening (Fig. 22C). Anal somite with paired sensilla on dorsal side. Anal operculum protruding free distal margin of anal somite and extruded in 4 strong spinular processes, rarely 5 (in general anal operculum in males more armed than in females) (Fig. 18B). Antennule (Fig. 21C): basically 9-segmented, geniculation between segments 7 and 8; penultimate and last segments, each with suture line marking original segmentation between former segments 8 and 9, and 10 and 11, respectively. Long tube-pores on segments 1 and 2. Armature formula: 1-[1], 2-[8], 3-[8], 4-[1], 5-[7+(1 + ae)], 6-[2], 7-[1], 8-[1], 9-[10 + (1 + ae)].

P5 (Figs 21D, 23A): free, with clear articulation to P5-bearing somite; right and left legs separate, intercoxal sclerite rudimentary but still discernible (Figs 21D, 23B). Basis of each leg well developed, representing most part of each leg; endopod rudimentary, consisting of a sclerotized and strong process articulated to basis. Row of surface spinules inserted near articulation between endopod and basis. Endopodal seta bipinnate, inserted on posterior surface of the endopod, close to its articulation to basis. Exopod distinct, clear articulated to the basis, wide and short, with unusual topology, being placed at the inner free distal margin of basis; exopodal armature consisting of 6 elements, all of which in apical position. Inner spinulose seta short and distinctly curved inward, the remaining setae of about the same length, 2 of which (the second and the fourth, beginning from the inner margin of the exopod) are respectively bipinnate and unipinnate; the remaining 3 smooth and slender, frequently closely adherent to each other and not easily discernible as distinct (Fig. 24). Male P6 (Figs 21E, 25A–B): right and left legs distinct, closely adherent along inner margin, and symmetrical, each leg consisting of a well developed lamellar plate, with some spinule rows on the anterior surface. A membranous lamella

is observable between right and left P6 (rudimentary intercoxa?) (Fig. 25B); armature consisting of 2 inner spines of different length and 1 outer naked seta.

## Discussion

According to Dussart and Defaye's (1990) world catalogue of freshwater harpacticoids, eleven species of *Phyllognathopus* are formally accepted as valid. However, two species, *P. labicauda* Por, 1964 and *P. medius* Por, 1964 must be discounted, since they belong to the genus *Phyllopodopsyllus* T. Scott, 1906 (Tetragonicipitidae), and were erroneously assigned to the genus *Phyllognathopus*. Moreover, *Phyllognathopus coecus* (Maupas, 1892) is still considered a valid species by some authors (Borutzky 1964, Dámian-Georgescu 1970, Dussart and Defaye 1990), whereas it is in reality a junior synonym of *P. viguieri*, as already pointed out by Lang (1948). Dussart and Defaye (1990) also ranked *Phyllognathopus coecus* var. *brevisetosus* (Daday, 1901) and *P. fodinatus* (Zieglmayer, 1923) as taxa *incertae sedis*, and we have followed their decision since neither Daday (1901) nor Zieglmayer (1923), strongly criticized by Chappuis (1924), provided the detail required for a correct identity of the taxa. Dussart and Defaye (1990) did not list *Phyllognathopus* sp., reported and partially figured by Dussart (1984) from New Caledonia. According to the author, this species possesses several peculiar morphological characters of high taxonomic significance. Similarly omitted was the species reported from Madras (India) and described by Krishnaswamy (1957) under the name *P. viguieri*, which in our opinion should be transferred to a new species, characterized by the combination of several features, not least the presence of 2-segmented P4 exopods. Unfortunately Krishnaswamy's type-material no longer exists (Ranga Reddy, pers. comm.), so we admittedly based our conclusions on newly collected material from India (although not from the type-locality). Conflicting opinions on the validity of the individual species of *Phyllognathopus* instigated serious taxonomic confusion. Most workers attempted to solve the problem by synonymising quite orthodoxically the largest number of species and subspecies possible. On the other hand, because of the low standard of published descriptions and the alleged variability observed among (sometimes sympatric) populations, it is not surprising that this taxonomic practice became popular. Most of the supposed morphological variability was found in the morphology and armature of caudal rami and male P5. In different botanical gardens in Great Britain, harbouring different tropical plants, several "morphological types" were observed and discussed by Gurney (1932). In this paper, the author concluded that these forms are to be considered phenotypic variations of the same species *P. viguieri*. Unfortunately, the drawings for each population, although of good quality by contemporary standards, are incomplete for the remaining morphological characters, thus preventing us from attributing a more accurate taxonomic status to the different populations. Moreover, it is not unlikely that they may belong to different species imported from different places together with their host-plants. Chappuis (1940) already reached an analogous conclusion. He observed that the different populations

described by Gurney (1932) originated from different botanical gardens in Kew, Oxford and Edinburgh, suggesting that the populations containing males with different P5 baseoendopodal armature and females with different caudal ramus morphology and setation may have been imported together with the tropical plants with which they were associated. For this reason, it is conceivable that they are not native for the country where they were collected. Phytotelmata are the more common habitats for *Phyllognathopus* species. For example, the type-species was originally described from decaying banana trees in Algeria (Maupas 1892). One year later, Mrázek (1893) described *P. paludosus*, but subsequently both Hartwig (1896) and Scourfield (1906) considered it a junior synonym of *P. viguieri*. In particular, Scourfield (1906) argued that Mrázek (1893) based its description of *P. paludosus* on copepodids. Later on, Chappuis (1916) published a comparative table, illustrating major differences between *P. viguieri* (= *Viguiarella coeca*) and *P. paludosus*. However, some of the differences listed are doubtful. For instance, in *P. viguieri* the antenna is 4-segmented instead of 3-segmented in *P. paludosus*. In our opinion this difference is based on an observational error, i.e. the failure to recognize the boundary between the small basis and the proximal endopodal segment. This explains why the exopod is either figured (correctly) at the basis-endopod boundary or (erroneously) halfway of the outer margin of what appears to be an allobasis. Other characters, such as the relative length of caudal rami (more than twice longer than wide in *P. paludosus*, vs. 1.5 longer than wide in *P. viguieri*), the shape of the inner terminal caudal seta V (transformed in *P. viguieri*, vs. normal in *P. paludosus*), the anal operculum (smooth in *P. viguieri*, vs. armed with fine spinules in *P. paludosus*), the inner protrusion (= transformed endopod) of male P5 baseoendopod with seta in *P. paludosus*, vs. 1-segmented cylindrical endopod bearing a transformed seta in *P. viguieri*, seem to support the validity of *P. paludosus*, which is considered a valid species also by Borutzky (1964), Damian-Georgescu (1968), and Barclay (1969). More recently, Chang and Yoon (2007) redescribed both *P. viguieri* and *P. paludosus* from South Korea, and the populations they assigned to the above species show minor differences in relation to the available descriptions. In particular, the Korean *P. viguieri* possesses anal operculum with free distal margin smooth or “with several minute projections” (Chang and Yoon 2007: 60), whose nature remains doubtful, since this minor ornamentation is not homologous to the spinules of the free distal margin of the anal operculum of several phyllognathopodids; the male P6 is figured and described as a small protrusion, bearing 3 elements, without any mention to the presence/absence of a continuous lamellar plate connecting, or not, right and left P6; again, this condition should require confirmation, because it has never been reported in other descriptions of *P. viguieri*. Moreover, some discrepancies are also observable between drawings and text descriptions. In particular, the maxilliped of *P. viguieri* is figured with 10 elements, vs. described with 9; the male P5 endopod is figured as distinctly 1-segmented, with an additional element at the insertion of the free endopod with basis (Chang and Yoon 2007: figure 2E) but described as *partly fused with exopod with 1 protuberance bearing about 10 spinules or setules around distal margin* (Chang and Yoon 2007: 60). In the same occasion, the authors described and figured *P. paludosus*. The male P5 does not

fit previous descriptions in the morphology and construction of the endopod, and the male P6 is partly figured and described as small protrusion. Unfortunately, all the available descriptions of *P. paludosus* are incomplete, preventing any clear statement and critical assessment of the diagnostic features of this species.

Chappuis (1938) assigned one population from the River Ondo, close to Lake Kibuga (Zaire) to *P. viguieri* without describing or figuring any specimens. This species, which had already been recorded from tropical Africa (Lake Tanganyika) (Gurney 1928) was subsequently reported by Chappuis (1956) from a cave in La Réunion. Again, no text description or figures were provided except for the observation that the female caudal rami were at least twice longer than wide and for the remaining characters both sexes in the population fitted the diagnosis of *P. viguieri*. Barclay (1969) described *P. volcanicus* from New Zealand, which resembles *P. paludosus* in most aspects, especially in the morphology and armature of the caudal rami, and differs from the latter only in the relative length of the exopodal setae of the female P5. In *P. paludosus* (as well as in *P. viguieri*) the longest seta is the third, whereas in *P. volcanicus* it is the fourth.

Chappuis (1928) described a new subspecies from bromeliads on the Island Sumatra (Chappuis 1928, 1931), named *Phyllognathopus viguieri menzeli* (Chappuis, 1928) [as *Viguiarella coeca menzeli* Chappuis, 1928], on the basis of the armature and morphology of the male P5 baseoendopod, bearing a well developed inner protrusion, representing the former endopod, plus 1 normal pinnate seta, probably corresponding to the *Viguiarella* sp. "Salakform" described by Menzel (1926) from Buitenzorg in Java (Indonesia). Subsequently, Chappuis (1931) supplemented the original description with some details on the morphology and armature of the male P2 distal endopodal segment, which bears a long apical transformed spine, giving more robust support for the validity and possibly specific status of this taxon. *P. viguieri menzeli* has also been reported by Watkins and Belk (1975) from phytotelmata in Guam. Their identification was supposedly based on the close resemblance of the male P5 with that originally described by Chappuis (1928) but direct comparison of the respective drawings failed to reveal such close similarity. In the original description the baseoendopodal seta is normally built and setiform, whereas in the material from Guam it is drawn as a large and stout element, not dissimilar in ornamentation from the inner protrusion (transformed endopod), suggesting that they are not homologous. Our examination of the specimens from Guam, on which Watkins and Belk (1975) based their assignment, revealed a quite different situation: 1) male P5 endopod identical to that of *P. viguieri* (1-segmented endopod bearing a large and short leaf-like transformed seta); 2) male P5 exopod wider than in *P. viguieri* but with identical armature; 3) no trace of transformed seta on male P2 enp-3; 4) anal operculum armed with spinules. These observations definitively confirm that the specimens from Guam cannot be assigned to *P. viguieri menzeli*, and they more likely represent a new species, closely related to the nominotypical species *P. viguieri*. Jakubisiak (1929) described *Viguiarella coeca parvula* (= *Phyllognathopus viguieri parvulus* (Jakubisiak, 1929)) from mosses in Poznam (Poland). This subspecies differs from the nominotypical species by the smaller body size (288  $\mu\text{m}$ ), and the different P5 morphology in both sexes.

Unfortunately, this subspecies was insufficiently described and not figured at all in the original description, although some drawings were provided in a subsequent publication (Jakubisiak 1931). In this paper, the author recognized some similarity with the P5 of *P. fodinatus*, but the description and figures of this species by Ziegelmeier (1923) are vague and erroneous in several aspects. For instance, Ziegelmeier's (1923) fig. 7 represents the male P5 and not the mandible as cited in the legend. This male P5 shows the outer basal seta arising from the exopod, and the baseoendopods being coalescent, displaying no discernible trace of armature, apart from some tiny setules along the free distal margin. Interpreting the swimming leg setation pattern is a most intractable issue because it is impossible to distinguish between ornamentation and armature elements. Lang (1948), being unable to resolve the taxonomic confusion, synonymised several species and subspecies with *Phyllognathopus viguieri*, namely *P. coecus* (Chappuis, 1916), *P. coecus menzeli* (Chappuis, 1928), *P. coecus parvulus* (Jakubisiak, 1929), *P. paludosus*, *P. coecus brevisetosus* (Daday, 1901), *P. fodinatus*, and *P. chappuisi* Delachaux, 1924, claiming the high morphological variability of this species as a reflection of its ecological plasticity.

Chappuis (1940) (not cited by Lang 1948), in his description of *P. insularis* Chappuis, 1940 from mosses collected in the subantarctic Marion Island (Southern Indian Ocean), lent support to the taxonomic validity of *P. chappuisi*, originally described by Delachaux (1924) from a similar habitat in Surinam (South America). Delachaux figured only the antennules, antennae and P5 (all based on a single male provided by Chappuis). Chappuis (1924) himself pointed out the 2-segmented condition of both rami of the P4 and figured this appendage and the P5 in a subsequent paper (Chappuis 1940). With the discovery of this clear-cut character, the systematics of the genus *Phyllognathopus* started to be viewed in a different light. Some characters were considered to have a more robust taxonomic significance than others, but for most morphological features such significance remained obscure. Chappuis (1940) for the first time recognized the importance of P4 exopodal segmentation in assessing the taxonomic position and status of *Phyllognathopus* populations. Both *P. chappuisi* and *P. insularis* share a 2-segmented P4 exopod, together with the widespread 2-segmented endopod, with the distal segment bearing the 3 basic elements. They differ in the arrangement and armature of the male P5 baseoendopod, the inner protrusion (transformed endopod) being accompanied by 1 normal seta in *P. insularis*, but being completely absent in *P. chappuisi*. The male P4 shows an identical setation pattern in both species, differing otherwise only in the relative length of the armature elements, and in the length of the distal exopodal segment (being slightly longer in *P. insularis*). The synonymy proposed by Karanovic and Ranga Reddy (2004) between these species is not strongly supported by evidence and the observed differences were probably underestimated, following Lang (1948)'s practice. Božić (1965, 1966) started to adopt a biological approach to the taxonomy of *Phyllognathopus* by demonstrating that superficially similar forms did not interbreed and consequently individual species boundaries are much narrower than traditionally believed, necessitating the re-instatement of some taxa previously considered as mere "forms". He also stressed the necessity to re-validate the taxonomic

significance of some morphological characters. On this basis, he described *P. camptoides* Božic, 1965 collected from dead wood on a forest floor near a pond in Gabon, and *P. paracamptoides* Božic, 1968 from mosses in La Réunion (Božic 1968). In regard to *P. camptoides*, Karanovic and Reddy (2004) considered this species junior synonym of *P. chappuisi*, basing their conclusion on the description of *P. cf. camptoides* given by Defaye and Heymer (1996) for a phyllognathopodid population collected from the soil cover of a shaded forest in Irangi (Zaire), and arguing that these authors “found enough variability to synonymise these three species” (Karanovic and Ranga Reddy 2004: 131) (i.e. *P. chappuisi*, *P. insularis* and *P. camptoides*). Actually, *P. cf. camptoides* differs from the species described by Božic (1965) by the setation pattern of P5 in both sexes. In the Zaire population the female P5 exopod is armed with 4 setae, vs. 3 in *P. camptoides*; the male P5 exopod bears 5 setae, vs. 6 in *P. camptoides*, and the basis bears the endopodal protrusion accompanied by 1 seta, vs. the same is absent in *P. camptoides*. On the basis of these differences, Defaye and Heymer’s (1996) material could in our opinion be assigned to a different species.

Recently, cross-breeding experiments carried out by Glatzel and Königshoff (2005) strengthened Božic’s results, encouraging a deep re-visitation of the entire putative

**Table 1.** Genera, species and subspecies presently recognized in the family Phyllognathopodidae.

<b>Genus <i>Phyllognathopus</i> Mrázek, 1893</b>	
	<i>P. viguieri</i> (Maupas, 1892)
	<i>P. viguieri menzeli</i> (Chappuis, 1928)
	<i>P. paludosus</i> Mrázek, 1893
	<i>P. insularis</i> Chappuis, 1940
	<i>P. chappuisi</i> Delachaux, 1924
	<i>P. camptoides</i> Božic, 1965
	<i>P. paracamptoides</i> Božic, 1968
	<i>P. volcanicus</i> Barclay, 1969
	<i>P. inexpectatus</i> <b>sp. n.</b>
<b>Species and subspecies inquirendae</b>	
	<i>P. coecus brevisetosus</i> (Daday, 1901)
	<i>P. fodinatus</i> (Zieglmayer, 1923)
	<i>P. viguieri parvulus</i> (Jakubisiak, 1929)
	<i>P. sp.</i> ( <i>sensu</i> Dussart 1984)
	<i>P. sp.</i> ( <i>sensu</i> Krishnaswamy 1957)
	<i>P. cf. camptoides</i> ( <i>sensu</i> Defaye and Heymer 1996)
<b>Genus <i>Allophyllognathopus</i> Kiefer, 1967</b>	
	<i>A. brasiliensis</i> Kiefer, 1967 (type-species by monotypy)
<b>Genus <i>Parbatocamptus</i> Dumont and Maas, 1988</b>	
	<i>P. jochenmartensi</i> Dumont and Maas, 1988 (type-species by monotypy)
<b><i>Neophyllognathopus</i> gen. n.</b>	
	<i>N. bassoti</i> (Rouch, 1972), <b>comb. n.</b> (type-species by monotypy)

range of *P. viguieri*, arguing also the potential relevance of differences eventually observed on microcharacters.

A taxonomic dilemma was raised with the description of *Phyllognathopus bassoti*. The most striking morphological features of *P. bassoti*, as reported in the original description given by Rouch (1972), were: 1) the possession of a 2-segmented P4 exopod in both male and female, 2) anal operculum with 3–4 large, long and strong spinules not articulated to free distal margin of anal operculum, and 3) the peculiar morphology and armature of P5 in both sexes. The morphology of P5 is undoubtedly the most distinctive trait of this species, which can easily be distinguished from all other known species of *Phyllognathopus*, *Allophyllognathopus* and *Parbatocamptus*. *Phyllognathopus bassoti* was also recorded from two different wells on Bantayan Island (Philippines) by Bruno and Cottarelli (1999). These populations show some differences with respect to the original description given by Rouch (1972). In particular the female P5 differs slightly in the shape of the inner lobe of the basipodite, which is shorter with one more seta in our specimens (Bruno and Cottarelli 1999: 525). Moreover, the male P5 differs in the armature of both exopod and baseoendopod: in the original description from Long Island the exopod bears 5 (as also described and figured by Karanovic and Ranga Reddy 2004 on Indian populations), vs. 6 setae in the Philippine material; the baseoendopod bears a short pinnate seta apparently inserted on the tip of the small endopodal protrusion, incorporated into the baseoendopod in the material from Long Island, vs. in the Philippine specimens it bears two different elements, one long pinnate seta, inserted proximal to the strong inner protrusion. Our re-examination of both Philippine and Indian populations (the latter also by using SEM) revealed a constant exopodal armature consisting of 6 elements, where the innermost spiniform seta is always strong, short and curved inward, and the three outermost setae are naked and slender, two of which closely adherent and superimposed to each other, making difficult their identity (see Fig. 24).

It is therefore not unlikely that differences observed in the relative development and armature of the P5 endopodal lobe are related to different perception of morphological details, because of intrinsic difficulties in observing the mutual position of the inner protrusion (which is a modified endopod) and the relative seta, whose surface insertion is hardly discernible under optical microscopy. Both populations of *P. bassoti* recently described from India (Karanovic and Ranga Reddy 2004) show minor differences in respect to the original description of the species given by Rouch (1972) and more consistent differences in respect to the populations described by Bruno and Cottarelli (1999) from Philippines. We consider the differences observed among populations as reflection of intraspecific variability, and the derived features shared by all the populations strong enough to rank this species to the new genus *Neophyllognathopus*. The other phyllognathopodid genera *Allophyllognathopus* and *Parbatocamptus* are monotypic. *Allophyllognathopus brasiliensis* Kiefer, 1967 is known from the upstream sector (“caatingas”) of the Rio Negro (Brazil) (Kiefer 1967), and *Parbatocamptus jochenmartensi* has been recorded from high-altitude leaf litter in Nepal (Dumont and Maas 1988) (Table 1).



**Review of morphological characters in the family Phyllognathopodidae.** Comparisons were based almost exclusively on material examined directly and only sporadically on existing descriptions. This course of action was necessary because in most descriptions many morphological details are missing and the drawings are usually so deficient that any comparisons made may be potentially misleading.

**Male antennules.** Male antennules are primarily 11-segmented, but, at present knowledge, this segmental pattern is not showed by any member of the family, and the derived condition of 10 segments is the most widespread in the family. An 8-segmented male antennule has been reported in *P. viguieri* (cf. Gurney 1932) and *Parbatocamptus jochenmartensi* (cf. Dumont and Maas 1988), and a 9-segmented antennule in *Neophyllognathopus bassoti* comb. n. (see Bruno and Cottarelli 1999, Karanovic and Ranga Reddy 2004). Glatzel and Königshoff (2005) reported a 10-segmented male antennule for *P. jochenmartensi*, probably resulted by counting segments 10 and 11 as distinct; whereas the male antennule of *A. brasiliensis* is reported by the same authors as 9-segmented, vs. 7-segmented in the original description (Kiefer 1967). These different segmental patterns partly reflect the level of fusion between the penultimate and last segments, which are usually fused together, but at least in some species the boundary between them is still discernible on posterior surface. Our examination of several species and different populations revealed the presence of an additional segment distal to segment 8. In most phyllognathopodids, this short segment is discrete, and was overlooked in past descriptions. The setation patterns on individual segments are virtually impossible decipher on the basis of published illustrations due to omission of setation elements. Our comparative analysis suggests that there is a relatively common, conservative setation pattern among populations and species, although the total number of setae counted per each segment may change on the basis of the different fusion patterns observed in different species.

Our re-examination of the segmental patterns of the male antennule revealed that *P. viguieri* possesses a basically 10-segmented antennule, where the penultimate and last segments are still distinct only on posterior surface (giving an incipient 11-segmented antennule) and fused on frontal surface. *Parbatocamptus jochenmartensi* shows a 10-segmented antennule, whereas *Neophyllognathopus bassoti* comb. n. displays a 9-segmented antennule, because both segments 8 and 9, and 10 and 11 respectively, are not distinct, probable reflection of heterochrony.

Novel structures were discovered on the antennules in both sexes; in particular, a truncated tubular extension, probably ending in a distal opening, is discernible on the first and second antennular segments in both males and females. In many harpacticoid families, these segments commonly possess (tube-) pores, as in, for example, the Neobryadyidae (Huys 1987), Cyliandropsyllidae (Huys and Conroy-Dalton 1993), Leptastacidae (Huys and Todaro 1997) and Ambunguipedidae (Huys 1990), and it is conceivable that the hyaline structures in phyllognathopodids represent tubular extensions of these pores. Similarly, flaccid structures can also be found on other appendages such as the antenna in the genera of the Leptopontiidae where it is expressed as a backwardly directed tubular "seta" on the allobasis (Huys and Conroy-Dalton 1996)

and almost certainly represents the external tubular extension of the persisting antennary gland (R. Huys, pers. comm.). An analogous aesthetasc-like structure is found on the maxilla in the Asterocheridae (Siphonostomatoida) where it forms a hyaline extension around the syncoxal exit of the maxillary gland (Huys and Boxshall 1991: Fig. 2.9.21D). Given their transparent nature, it is not surprising that antennular tube-pores have not been documented before in phyllognathopodid descriptions. Our analysis confirmed their presence in all *Phyllognathopus* species examined, as well as in *P. jochenmartensi* and *N. bassoti* comb. n., suggesting that this character may well be an autapomorphy for the family. The functional significance of the tubular extensions is as yet unknown but their presence and identical position in adults of both sexes appears to rule out a possible role in mate location or guarding.

**Antenna.** The antenna of *P. viguieri* has been reported as 4-segmented, vs. 3 segmented in *P. paludosus*, but this different segmentation pattern stems from failure to identify the segment boundary between the basis and the proximal endopod segment, and consequently the correct position of the exopod. In the original description (Mrázek 1893) of *P. paludosus*, the exopod is positioned on what appears to be an allobasis. Chappuis (1916) uncritically accepted this character as a diagnostic difference between *P. viguieri* and *P. paludosus*. The same observational error was made by Dámian-Georgescu (1970) and Borutzky (1964) in their redescrptions of *P. paludosus*. Our re-examination revealed that the antenna is invariably 4-segmented, comprising a coxa, a basis bearing the 1-segmented exopod, and a 2-segmented endopod.

**Mandible.** The basic structure of the mandible and the setation of the mandibular palp in the type-genus *Phyllognathopus* appear to be identical in all observed populations, the only exception being *N. bassoti* comb. n., which, according to Rouch (1972), bears one short seta on the basis. In the Philippine and Indian populations of this species, as redescrbed and figured by Bruno and Cottarelli (1999) and Karanovic and Ranga Reddy (2004) respectively, as well as in all species of *Phyllognathopus* we have examined, this seta is absent. As a matter of fact, the presence of a true seta is doubtful and requires confirmation as it could have been confused with one of the spinules forming the surface row that is always present in the same position (but it was not figured by Rouch 1972). On this regard, Karanovic and Ranga Reddy (2004) referred to a spinule element in the Indian populations of *N. bassoti* comb. n.

Re-examination of the male holotype of *P. jochenmartensi* revealed the presence of a well developed pinnate seta on the basis of the mandibular palp (not figured nor described in the original description), accompanied also by the typical spinule row. This observation suggests that the basis is armed in primitive phyllognathopodids. Interestingly, *P. jochenmartensi* also shows two transformed, prehensile setae on the endopod of the mandibular palp, a character already reported by Dumont and Maas (1988), which is a clear autapomorphy of the genus.

**Maxillule and maxilla.** The structure of the maxilla and maxillule is almost identical in all specimens observed, although comparison of the setation patterns with previously described species is hampered by inconsistencies and deficiencies contained in most descriptions. As regards to the maxilla, number of setae and their topology

are identical in all the species examined. In *P. jochenmartensi* the proximal endite is incorporated into the syncoxa and is composed by four lobes vs. the same endite is articulated to the syncoxa and the lobes are only hardly discernible in other members of the family. The maxillule is more conservative in both morphology and setation. The only exception is represented by the maxillule of *P. viguieri*, where the praecoxal arthrite bears 10 elements, whereas in all the other phyllognathopodids it bears 9 elements, being the proximal surface seta (inserted on a small knob) missing.

**Maxilliped.** The phyllopodial maxilliped is bilobed, with the basal part (syncoxa) fully incorporated in the compound distal part (basis and endopod fused). With regard to armature, a direct analysis of material among all the populations analysed, revealed two different setation patterns, accompanied by a different degree of fusion between the former syncoxa and the baseoendopod. In all the examined populations of *Phyllognathopus* the maxilliped bears 10 elements, and only a rudimentary incision between syncoxa and baseoendopod is still discernible. On the contrary, the primitive distinction between syncoxa and baseoendopod is still retained in *P. jochenmartensi* and *N. bassoti* comb. n. with both inner and outer incisions marking original segmentation; moreover, an additional element (a robust stout spine) is inserted along inner side of the boundary syncoxa-baseoendopod, suggesting this condition as primitive within the family.

**Integumental windows patterns.** The dorsal integumental window (“nuchal organ”) on the cephalosome has been reported only for *N. bassoti* comb. n. by Bruno and Cottarelli (1999). Our SEM observations failed to confirm its presence in *P. viguieri*, as well as in *N. bassoti* comb. n. SEM analysis revealed only a rounded globose structure in *P. viguieri* with the same topology of the nuchal organ; no solution of continuity of the cuticle is observable in all the specimens analysed.

**Swimming legs P1-P4.** Female P1-P3 are relatively identical in both morphology and armature in virtually all species and populations of *Phyllognathopus*. The only remarkable difference is observable in members of the family characterized by a 3-segmented P4 exopod. In particular, in *P. viguieri* and related species, the P4 exp-2 lacks the outer spine in all the examined populations, whereas it is present in *P. jochenmartensi* and absent in *A. brasiliensis*. Another exception refers to a population of *P. viguieri* collected from Lake Léman (Switzerland) by Dussart (1966, 1967) in which the female P3 exp-3 has been reported with 3 elements only, whereas in all other descriptions it consistently shows 4 elements (2 outer spines and 2 apical setae). This difference was first highlighted by Van de Velde (1974) in her description of *P. viguieri* from Lochristi (Belgium). Contrary to Dussart (1966), our re-examination of material from Lake Léman revealed the presence of 4 elements on the distal segment of P3 exopod.

The major differences between members of the family are found in the segmentation of the P4 exopod which can show three different patterns: 1) 3-segmented; 2) distinctly 2-segmented; or 3) 1-segmented with (*Phyllognathopus paracamptoides*) or without (*Phyllognathopus* sp. *sensu* Dussart 1984) a surface suture marking the original boundary between proximal and distal segments. In *A. brasiliensis* the swimming leg segmentation pattern is identical to that of *P. viguieri*. According to Dumont

and Maas (1988), *P. jochenmartensi* has 3-segmented P1-P4 exopods, 3-segmented P1 endopod, 2-segmented P2-P3 endopods and 3-segmented P4 endopod. Our re-examination of the male holotype revealed that the legs had been mixed up in the original description, as already supposed by Karanovic and Ranga Reddy (2004). In reality, the P2 has a 3-segmented endopod, whereas P3 and P4 display 2-segmented endopods.

Some variation is also expressed in the segmentation of the P4 endopod (in general, it is 2-segmented, but it was described and figured as 1-segmented in *Phyllognathopus* sp. by Dussart (1984).

By analysing different *Phyllognathopus* populations and species, as well as *N. bassoti* comb. n., the P4 praecoxa is always absent in both males and females, a character never described or reported for the family, whereas all the remaining swimming legs possess it. The absence of praecoxa is always accompanied by a noticeable reduction in the size of P4, irrespective of the segmentation pattern of this leg. Although the absence of the P4 praecoxa may be a potential synapomorphy of the *Phyllognathopus*-*Neophyllognathopus* lineage, its presence/absence in *Allophyllognathopus* requires confirmation before any phylogenetic inference can be drawn. *Parbatocamptus* still possesses a P4 praecoxa. Interestingly the P4 is large, and the exp-2 still bears 1 outer spine, suggesting that *P. jochenmartensi* possesses the most primitive P4 in the family. Members of the genus *Phyllognathopus* do not display sexual dimorphism in the segmentation of swimming legs. However, some authors have documented transformations of particular setae, especially on the male P2 endopod such as in *P. viguieri* by Gurney (1932: fig. 362) and in *P. viguieri menzeli* by Chappuis (1931: Figs 151–152). Unfortunately, we failed to trace the material on which the above authors based their descriptions, but the direct analysis of the specimens from Guam assigned to *P. viguieri menzeli* by Watkins and Belt (1975) revealed the absence of any kind of transformation of setae in P2 enp-3, as well as in any other of the remaining legs. The potential presence of sexual dimorphism in morphology and/or armature of P3 in *A. brasiliensis* and of P2 in *P. jochenmartensi* remains a hypothesis, since the female is unknown for both genera.

**Fifth legs.** The fifth pair of legs is sexually dimorphic in both structure and armature, and sometimes differs among species in the morphology of the exopod, in the degree of fusion between exopod and baseoendopod, in the number of exopodal and baseoendopodal setae, or alternatively, in their relative length. The female of *P. viguieri* shows an exopodal lobe with 4 apical elements, and a baseoendopod with 2 pinnate setae. The exopod is incorporated into the baseoendopod but the right and left legs are distinctly separate. *Phyllognathopus inexpectatus* sp. n. possesses a female P5 quite similar to that of *P. viguieri*, differing in the length of the exopodal setae, all shorter than in *P. viguieri* and, more importantly, in the topology of the outermost exopodal seta which is inserted in a clear outer subapical position. Only a few *Phyllognathopus* species show differences in the number of armature elements (e.g. *P. camptoides*). *N. bassoti* comb. n. differs from any phyllognathopodid species in the unique morphology of female P5. The female P5 baseoendopod and exopod are coalescent, deep incision marking original segmentation between them; the endopodal lobe is well developed,

elongate, rectangular in shape, longer than exopod, bearing 1 long pinnate seta, subdistally inserted, close to outer margin; exopodal lobe well discernible; exopodal armature consisting of 4 elements, the outermost seta inserted in subdistal outer position, the remaining ones in apical position.

The male P5 exhibits much more variation within the family, especially in the structure and armature of the baseoendopod. The most primitive condition is observed in *P. viguieri*, which exhibits a discrete 1-segmented endopod, on which the endopodal seta is terminally inserted. In some *Phyllognathopus* species, as well as in *A. brasiliensis* and *N. bassoti* comb. n., the endopod is secondarily transformed in a strong spinular process, sometimes described as incorporated into the basis, but more frequently articulating with it. In *P. jochenmartensi* the endopod is partially fused to basis forming a baseoendopod, although the suture line marking original segmentation is still discernible. In most phyllognathopodid species the endopod bears 1 element, but it was also reported without ornamentation. Despite its variation, no significance has been attributed to this character, in so far that populations displaying different setation patterns have been assigned to the same species.

The male P5 exopod shows different construction and armature among members of the family. The most primitive condition is showed by *P. jochenmartensi*, which retains a 2-segmented exopod; the exopod is distinctly 1-segmented in *N. bassoti* comb. n., and in *A. brasiliensis*, and appears as incorporated to the basis in *Phyllognathopus* (see also Glatzel and Königshoff 2005: 145).

Direct examination of the Philippine material assigned to *P. bassoti* by Bruno and Cottarelli (1999) (now *N. bassoti* comb. n.) and of another morphologically close population of this species from India (Karanovic and Ranga Reddy 2004) revealed new informative characters associated with the male P5. Males of both populations, and presumably also the type-series of *P. bassoti*, share the presence of an intercoxal sclerite joining the fifth legs which is completely absent in the female P5. Following this discovery, several populations assigned to *P. viguieri* were re-examined, as well as all available species with 2-segmented P4 exopods. Unfortunately the type-material of *P. camptoides* and *P. paracamptoides* no longer exists and the slide material of *P. cf. paracamptoides* deposited in the NMHN (Paris) is in a bad condition. Nevertheless, we had the opportunity to examine two new species with 2-segmented P4 exopods, *P. inexpectatus* sp. n. from Italian ground water and another population from India, which is currently being analysed. By comparing several populations and species, we observed that whereas most species of *Phyllognathopus* show a weakly defined, medial sclerotisation, fully incorporated into the baseoendopod, only the three known populations of *N. bassoti* comb. n. show a well defined intercoxal sclerite (also figured but not described by Bruno and Cottarelli 1999). *Parbatocamptus jochenmartensi* also seems to possess a P5 intercoxal sclerite, which is less easy to discern.

**Sixth legs.** The sixth pair of legs has only sporadically been described and solely in males. It is bilaterally symmetrical and bears 3 elements on either side: a long outer seta, presumably representing the original outer basal seta, and 2 spinulose inner setae. The male sixth pair of legs shows three different morphologies: 1) in most species it

appears as a hyaline linear and continuous lamella, lacking any trace of the primitive paired state showing distinct right and left legs (as in *P. viguieri*); 2) in a second species-group, the lamella appears medially incised as in some taxa currently under study, and in *Parbatocamptus jochenmartensi*, and 3) in *Neophyllognathopus bassoti* comb. n. the sixth legs are deeply incised forming a more complex structure, where also an interconnecting lamella has clearly been observed (rudimentary intercoxa?).

The P6 has never been described nor observed in females, frequently reported as absent (cf. Karanovic and Ranga Reddy 2004) and it was probably assumed that it was absent in phyllognathopodids. Our comparative study revealed the presence of the P6 in the female, bearing one seta only, which is usually short, robust and naked, and, less frequently, represented by a slender pinnate seta, being either very long or short. We conclude that the P6 is sexually dimorphic, being differently constructed in different species, or species-groups.

**Ornamentation of urosome.** The ornamentation of the urosome deserves special attention. By comparing several populations of *P. viguieri sensu lato*, several other *Phyllognathopus* species, *N. bassoti* comb. n., and *P. jochenmartensi*, clearly distinct ornamentation patterns could be distinguished, as well as previously unnoticed enigmatic structures located on both dorsal and ventral sides of the male urosome. The more complex structures were observed in *N. bassoti* using SEM (Fig. 22A–C). A similar ornamentation is presumably present in *P. camptoides*, as figured by Božić (1966: 37, and Fig. 2).

Both male and female urosome show large dorso-lateral pores located on the P5-bearing somite. They are referable to the pores observed in several Ameiridae (Galassi et al. 1999) and Canthocamptidae (Galassi, *pers. obs.*).

**Caudal rami.** Caudal rami are frequently sexually dimorphic and generally considered polymorphic in the females of *P. viguieri*. Variation in caudal rami morphology and setation pattern has been reported for several harpacticoids (Schminke 1991). Since different morphs of *Phyllognathopus* not infrequently co-occur in the same geographical area, some authors (e.g. Gurney 1932, Lang 1948) considered them as ecophenotypes of the widespread *P. viguieri*. Since Gurney's (1932) material came from different sites and, more importantly, different botanical gardens harbouring different imported tropical plants, it is highly conceivable that these different populations belong to different species and that caudal rami polymorphism is much more limited than previously assumed, as also observed by Königshoff and Glatzel (2008) in reared populations of *P. viguieri sensu lato*. On the other hand, the same authors stressed that the morphology of the posterolateral and inner terminal setae of female caudal rami is *per se* a weak diagnostic character, since species with the same setal morphology do not interbreed.

## Conclusion

The re-examination of type-material and/or topotypes of different phyllognathopodid species revealed a systematic scenario more complicate than supposed. The type species

of the genus *Phyllognathopus*, *P. viguieri*, was redescribed in detail, analysing several populations coming from different localities world-wide. Some morphological characters within the genus *Phyllognathopus* revealed taxonomic significance, giving ground for the description of *P. inexpectatus* sp. n. from ground water in Italy. The discovery of new informative phylogenetic characters led also to the proposal of a new genus for *P. bassoti*, namely *Neophyllognathopus bassoti* comb. n.

Re-examination of different populations and species of the genus *Phyllognathopus* led to the conclusion that the mouthparts show virtually no variation in structure and setation, and the few differences observed are usually autapomorphies of individual species (with the exception of the structure and setation of the maxilliped and the absence of a seta on mandibular basis which is a synapomorphy of a wider group of species/genera). The most robust discriminating features between species-groups are the segmentation of the P4 exopod, the general morphology of legs 5 and 6 in both sexes, and the morphology and ornamentation of male urosome. The different segmental patterns of P4 exopods observed among species of *Phyllognathopus* allow the identification of three morphological groups: 1) species with 3-segmented exopod (here defined *P. viguieri*-group, including *P. viguieri*, *P. viguieri menzeli*, *P. viguieri parvulus*, *P. viguieri brevisetosus*, *P. paludosus*, *P. volcanicus*); 2) species with 2-segmented exopod (here defined *P. chappuisi*-group, including *P. chappuisi*, *P. insularis*, *P. camptooides*, *P. cf. camptooides sensu* Defaye and Heymer (1996), *P. viguieri sensu* Krishnaswamy (1957), *Phyllognathopus* sp. (Galassi and Fiasca, under study), and *P. inexpectatus* sp. n.); 3) species with 1-segmented exopod (here defined *P. paracamptooides*-group, including *P. paracamptooides* and *Phyllognathopus* sp. *sensu* Dussart (1984)). After an in-depth re-examination of different species and populations we refrain from attributing any phylogenetic validity to these groups. The reason for this decision is twofold: 1) the more derived groups (*chappuisi*- and *paracamptooides*- groups) show only one derived character state in comparison to the other ones; 2) this difference exclusively pertains to the reduction in the number of exopodal segments of P4, probably resulting from heterochrony, such as post-displacement. Moreover, this character appears to be evolutionary labile since, for instance, in the *paracamptooides*-group, the boundary-line between the first and the second segment is still partly expressed, reinforcing the hypothesis that the development of P4 is post-displaced relative to that of the other legs (P5 excluded, development of which appears to be decoupled from that of the swimming legs, as also noticed in the Parastenocarididae by Galassi and De Laurentiis 2004). The pedomorphic origin of the fourth leg is reflected in its small size (compared to the dimension of other swimming legs), the absence of the outer spine on the exp-2 in all members of the family showing a 3-segmented P4 exopod, the absence of the praecoxa (vs. present in P1-P3), and the strong tendency towards a reduction of the segmental pattern in both exopod and endopod. Only *Parbatocamptus* retains a relatively large P4, a well developed P4 praecoxa, and the outer spine on P4 exp-2. Despite its unstable ontogeny, being the only variable appendage within the genus *Phyllognathopus*, specimens with 3-segmented exopods have never been found to co-occur in the same "population" with specimens displaying 2-segmented exopods, suggesting

that this character may be a useful discriminant at least at species level. Reductions in swimming leg segmentation should be employed with caution when inferring phylogenetic relationships between species or species-groups, particularly when no other evolutionary novelties accompany such derived character states. Evolution in copepods frequently entails character losses or fusions between segments. Nevertheless, it is not unlikely that such reductions may have occurred independently more than once in the evolutionary history of a family or genus. For example, endopodal segmentation can be highly variable in certain harpacticoid lineages, and it would be unwise to automatically attribute excessive phylogenetic significance to groups of species sharing a derived segmentation pattern (e.g. endopod 2-segmented, 1-segmented or absent at all, vs. the 3-segmented primitive condition) without having other synapomorphies in common. Identical endopodal segmentation patterns could potentially be homoplastic, either as the result of convergence by habitat selection, or of parallelism, due to the fact that the morphological character states in question share a common ontogenetic basis. Consequently, the evolutionary instability of the endopodal segmentation should be considered with great caution in assessing the common ancestry of derived taxa within a given lineage. This situation has already been observed in some harpacticoid families, such as the Ameiridae (Galassi et al. 1999, Galassi 2001, Galassi et al. 2009), where differences in endopodal segmentation originated as result of intrageneric evolution (Lee and Huys 2002).

Exopodal segmentation patterns of swimming legs are generally more conservative than those of endopods, and the explanatory power of derived states may be potentially higher in resolving phylogenetic issues. Within the Harpacticoida, members of the same genus very rarely exhibit different segmental patterns in the exopods of P2-P4. For example, in the canthocamptid genus *Hypocamptus* Chappuis, 1929, *H. brehmi* (Van Douwe, 1922) shows P3 and P4 with 3-segmented exopods, whereas *H. paradoxus* (Kreis, 1926) has 2-segmented P3 and P4. Similarly, the laophontid genus *Laophontina* Norman & T. Scott, 1905 contains species with 2-segmented (*L. acantha* Noodt, 1955; *L. noodti* Kunz, 1983) and 3-segmented (*L. dubia* Norman & T. Scott, 1905; *L. posidoniae* Fiers, 1986) P4 exopods. In another laophontid genus, *Robustunguis* Fiers, 1992, the type species *R. unguulatus* Fiers, 1992 possesses 3-segmented P2-P4 exopods but in its congener *R. minor* Fiers, 1992 these rami are only 2-segmented. Unfortunately, our analysis failed to reveal any congruence between exopodal segmentation patterns and other derived character states of phylogenetic significance, and consequently we were unable to delimit any “natural groups” within the genus *Phyllognathopus*. The *P. chappuisi*-“lineage” almost certainly evolved within the *P. vignieri*-group, although its phylogenetic position remains unresolved. Species belonging to this “lineage” all have the 2-segmented P4 exopod, which may have evolved independently and several times in the evolutionary history of the genus *Phyllognathopus*. However, since these species do not share any other derived character states, the monophyly of the *chappuisi*-“lineage” remains questionable as it may include the species of the *paracamptoides*-group and therefore be paraphyletic. On the other hand, if *Neophyllognathopus bassoti* comb. n. may theoretically enter into the “*chappuisi*-lineage” for



the possession of a 2-segmented P4 exopod, this species is defined by a combination of unique apomorphies, most of which discovered in the present study: 1) P4 with 2-segmented exopods; 2) unique morphology of P5 in both sexes; 3) peculiar arrangement of the ornamentation of the hyaline frills on the male urosome ventrally; 6) anal operculum bearing 3–4 long and stout spinules, not articulated to the operculum. It also shows several plesiomorphic character states, such as the presence of an intercoxal sclerite between male fifth legs, the bilobed, separate male sixth legs (with rudimentary intercoxa?), the phyllopodial maxilliped with clear trace of articulation between syncoxa and basis-endopod, and an additional spiniform seta on the same maxilliped. Within the Phyllognathopodidae, the new genus *Neophyllognathopus* shows feeble relationships with the genus *Parbatocamptus*, which plausibly is the most primitive genus within the family, showing the most plesiomorphic state of male P5, with 2-segmented exopod, trace of endopod together with the presence of a rudimentary intercoxal sclerite; a deeply incised and well sclerotized male P6, the basis of the mandibular palp bearing one long bipinnate seta, the phyllopodial maxilliped with 11 elements and clear trace of the primitive articulation between syncoxa and basis-endopod, and the presence of P4 praecoxa and the outer spine on P4 exp-2, always absent in all members of the family showing a 3-segmented P4 exopod. *Neophyllognathopus* gen. n. shares with *Parbatocamptus* the identical construction and armature of the maxilliped, and the presence of a rudimentary intercoxa in male P5. Such similarities are however symplesiomorphic and more detailed information about *Parbatocamptus* is required (for instance, its female is unknown) before such a relationship can be corroborated. Pending the arrival of new data (e.g. molecular analysis, Glatzel, *in litt.*), it seems justifiable to maintain the *P. chappuisi*-group and the *P. paracamptoides*-group in the genus *Phyllognathopus*, considering the differences in P4 exopodal segmentation as intrageneric variation, and to assign generic rank to *Phyllognathopus bassoti* by creating the new genus *Neophyllognathopus*.

Among members of the *chappuisi*-group, all defined by 2-segmented exopods and endopods, *P. inexpectatus* sp. n. is easily distinguishable by a P4 enp-2 with only 2 apical elements (vs. 3 in all members of this group), a spinulose free distal margin of anal operculum (vs. ciliate in *P. insularis* and armed with strong spinular processes in *P. camptoides*), caudal rami with posterolateral seta III transformed and subapical (vs. apical and not transformed in *P. insularis*). *P. camptoides*, as originally described by Božic (1965) shows only 3 elements on the exopodal lobe of female P5 vs. the widespread condition of 4 elements in all the remaining members of the *chappuisi*-group. The urosome of *P. camptoides* has been figured with spinulose hyaline frills ventrally (somewhat resembling the ornamentation of *Neophyllognathopus* gen. n., and, to lesser extent, *Parbatocamptus*), vs. the same are plain in the new species. The descriptions of *P. insularis* by both Delachaux (1924) and Chappuis (1940) are so generic that any conclusion is inadequate, apart from the armature of the P4 enp-2, described and figured with 3 elements. The *Phyllognathopus viguieri* described and figured by Krishnaswamy (1957), which, as already mentioned, is in need to be transferred to a different species, enters the *chappuisi*-group, being instantly recognizable as different species by the presence of

3 elements on P4 enp-2 and caudal inner terminal seta not transformed. A spinulose anal operculum is shared by this species and *P. inexpectatus* sp. n. The missing male of *P. inexpectatus* prevents us from further considerations on the interspecific relationships with apparently closely related species.

*P. inexpectatus* sp. n. is defined by the combination of the following morphological characters: maxillule with syncoxal proximal surface seta absent (vs. present in *P. viguieri*); P2 enp-2 without inner seta (vs. present in *P. viguieri*); 2-segmented P4 exopod (vs. 3-segmented in *P. viguieri*); 2 apical elements on P4 enp-2 (vs. 3 elements in *P. viguieri*); female P5 exopod with 3 apical and one distinctly subapical outer seta (vs. 4 apical setae in *P. viguieri*); female P6 with long bipinnate and slender seta (vs. short, naked and with rounded tip in *P. viguieri*); anal operculum with spinules (vs. smooth in *P. viguieri*); inner terminal seta normally shaped (vs. short and with proximal part enlarged in *P. viguieri*).

**Ecology and biogeography.** Phyllognathopodidae occur in both temperate and tropical areas, and at different altitudes, with high preference for phytotelmata, leaf litter, moist soils, pitcher plants, man-made and altered habitats. More rarely they occur in mosses (Reid 2001) and in abandoned coalmines. They invaded also genuine freshwater habitats, as they are frequently found in epibenthic layers of sediments in ponds, streams and lakes, in hyporheic habitats, as well as in phreatic and karstic groundwater systems. Their potential for dispersal seems to be very high, by both active and passive dispersion mechanisms. The most demonstrable example was reported by Rouch (1972) who collected *Neophyllognathopus bassoti* comb. n. from a small sandy island in Wisdom Lake only 20 months after its formation in the lake. The Karaman-Chappuis (Delamare Debutteville 1954) method used to take this sample prevents us from assessing more accurately the real ecology of the species. An ecological confusion comes also from Karanovic and Ranga Reddy (2004) which recorded this species from India and considered the species stygobiont at Kandukur, and stygophyle at Guntur, on the basis of the different habitats from which both populations have been collected.

At present, it is difficult to speculate about the plesiotypic habitat of the family, but it is not unlikely that epigeal, semi-terrestrial habitats represent the ancestral and still preferred environment for the family in temperate and, especially, in tropical areas. Circumstantial evidence supporting this hypothesis is provided by the high likelihood of discovering phyllognathopodids in these habitats world-wide, where they also appear to have their highest abundance and species diversity. How they can survive dehydration during low-water periods is unknown. Resting stages have never been found, and dormancy has not been documented until now.

From a biogeographical point of view, the cosmopolitanism of *P. viguieri* has unjustly been overemphasized as discussed also by Glatzel and Königshoff (2005). It is now obvious, however, that under this name several cryptic species are hiding, sometimes only recognizable on the basis of differences in microcharacters (morphology and ornamentation of anal operculum, ornamentation of urosomites, ornamentation and armature of both female and male P5 and P6). More precise information for discriminating different species will undoubtedly become available with the arrival of

molecular data analysis. Among true freshwater populations presently attributed to *P. viguieri*, some differences have been observed, the taxonomic significance of which is still debatable. For this reason, it seems more adequate to discuss the distribution of the *P. viguieri*-group, as defined above in its restricted sense: it is cosmopolitan in distribution, and utilizes different habitats, occurring more frequently in phytotelmata. The *chappuisi*-group consists of epigeal forms, predominantly distributed in tropical areas of the Southern Hemisphere, the only exceptions being *P. inexpectatus* sp. n., which is the first and only member of this group described from the Holarctic Region as a whole. *P. inexpectatus* may be classified as a stygobiont, as it was collected from a karstic aquifer. The colonization of ground water by the putative ancestor may have occurred before or during the Quaternary glaciation, when most of the epigeal elements disappeared, and only some populations survived in refuge habitats, like ground water. The widespread presence of the *P. viguieri*-group in the Northern Hemisphere is probably linked to post-glacial recolonizations, by both active and passive mechanisms.

The new genus *Neophyllognathopus* shows a disjunct geographical distribution in the Oriental and Australasian regions and it is thus far predominantly restricted to ground water *sensu lato* (subsurface freshwater habitats: this is the case of *N. bassoti* from Long Island (Papua, New Guinea) and from Philippines and India). Some doubts there are also for *Allophyllognathopus brasiliensis*, collected from the “caatingas” located in arid areas in Brazil. These areas are characterized by xeric vegetation and their hydrological regimes are regulated by intermittent rivers, which exist only during the rainy season.

The geographical distribution of the four defined groups is not of great assistance in corroborating or refuting phylogenetic affinity between or within groups. Members of the same group do not show a common track of distribution. The *P. viguieri*-group is cosmopolitan, and, at present, any speculation about the centre of origin of the group is premature. Members of the *chappuisi*-group are distributed in both Northern and Southern Hemispheres, predominantly in tropical areas with the exception of the geographically disjunct location of *P. inexpectatus*, which is recorded from temperate Europe. Two alternative hypotheses may be proposed: 1) the *chappuisi*-group was widespread in the past, and descended directly from a *P. viguieri*-like ancestral stock, but disappeared from the plesiotypic surface habitats in the Northern Hemisphere as a consequence of the drastic climatic changes linked to the Quaternary glaciations, and survived as relict species in refuge environments (e.g. ground water); 2) members of the *chappuisi*-group may have originated independently in different geographical areas from different surface ancestors closely related to the *P. viguieri*-group. In the first scenario, a common origin is hypothesized for the *chappuisi*-group, which could be considered monophyletic within the *P. viguieri*-group; in the second one, the “lineage” should be considered polyphyletic. The new genus *Neophyllognathopus* established for the *P. bassoti*-lineage seems to be the only one for which a monophyletic origin may be reasonably inferred. Its distribution is thus far limited to tropical India, the Philippines and New Guinea, in both the Oriental and Australasian Regions, a very problematic area from a biogeographical point of view (Lomolino et al. 2006). There is some ground to suppose an ancient origin for this group, and

a recent colonization from India-Philippines to Long Island by dispersal events. The ecological preferences of the species refer to groundwater habitats *sensu lato*, especially the ecotonal boundary between surface and subsurface environments (e.g. hyporheic and subsurface alluvial habitats). Against this ecological background, it is not surprising that *Neophyllognathopus* shows some relationships with the genus *Parbatocamptus*, collected from leaf-litter in Nepal.

## Keys to genera of Phyllognathopodidae

### Males

- 1 P5 with 1-segmented exopod and endopod, the latter cylindrical or transformed in a spiniform element..... **3**
- 2 P5 with 2-segmented exopod; endopod incorporated to basis forming a baseoendopod; endopod boundary marked by rudimentary suture; rudimentary intercoxa present; P6 represented by a deeply incised hyaline lamella.....  
..... *Parbatocamptus*
- 3 P3 endopod not transformed and with setae and/or spines normally conformed..... *Phyllognathopus*
- 4 P3 endopod transformed, with aesthetasc-like elements on enp-3.....  
..... *Allophyllognathopus*
- 5 P5 with intercoxa, exopod discrete, very large and translated at the inner margin of the basis; right and left P6 distinctly separated; third and fourth urosomites with deep ventral sockets..... *Neophyllognathopus* gen. n.

### Females\*

- 1 P5 baseoendopod and exopod coalescent; incision marking original articulation between basis and endopod little pronounced; baseoendopod bearing two elements..... *Phyllognathopus*
- 2 P5 baseoendopod and exopod coalescent; deep incision marking original articulation between basis and endopod; endopod bearing one element.....  
..... *Neophyllognathopus* gen. n.

\*female unknown for *Parbatocamptus* and *Allophyllognathopus*.

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# Key to the Tribes and Genera of Deltocephaline Leafhoppers (Auchenorrhyncha, Hemiptera, Cicadellidae) of Pakistan

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## Abstract

A key with accompanying figures is provided for the 14 tribes and 35 genera of Deltocephalinae (Cicadellidae) from Pakistan.

## Keywords

Deltocephalinae, taxonomy, morphology

## Introduction

Deltocephalinae is the largest and most diverse subfamily of Cicadellidae with 6200 described species placed in over 850 genera (McKamey in press), in 36 tribes (Zahniser and Dietrich 2010). Members of the subfamily are also important vectors of plant diseases (Weintraub and Beanland 2006) and account for 117 of the 151 cicadellid vector species listed by Nielson (1968).

Early leafhopper taxonomic work in Pakistan (from 1960 onwards) focused on Typhlocybinae. Work on Deltocephalinae was scattered in various publications (Ahmed 1986, Ahmed and Aziz 1988, Ahmed and Rao 1986, Ahmed et al. 1988, Ara and Ahmed 1988, Fatima et al. 1995, 1997, 1998, Mahmood 1979, 1980, Mahmood and Aziz 1979, Mahmood and Meher 1973, Mahmood et al. 1972), but a thorough review

of this literature was given by Khatri and Webb (2010). These authors also provided a checklist, new combinations, new species, new synonymy and new record and a key to the 14 tribes.

In the present paper we revise the above tribal key to include Mukariini, following its recent discovery in Pakistan (Khatri and Webb 2011) and its inclusion in the sub-family by Zahniser and Dietrich (2010), together with Drabescini (Paraboloponina) and Penthimiini, also included by Zahniser and Dietrich (2010). Drabescini is represented in Pakistan by *Dryadomorpha pallida* Kirkaldy (1906: 336), recorded from Pakistan as *Rhombopsis viridis* Pruthi see (Webb 1981) and Penthimiini, represented by *Neodartus acocephaloides* Melichar (1903: 163), from Hafizabad and *Penthimia compacta* Walker (1851: 842) from Murree Hills (National Pusa Collection, IARI, New Delhi, pers. com. Dr. C.A. Viraktamath).

An additional genus and species, omitted by Khatri and Webb (2010), is also included: *Gurawa minorcephala* Pruthi (1930: 29) (described from Pakistan: Murree Hills) and following Zahniser (2008) is placed in Chiasmini. We here follow Khatri and Webb's (2010) broad concept of Athysanini to include *Osbornellus* Ball and *Scaphoideus* Uhler which were left unplaced to tribe by Zahniser and Dietrich (2010).

One other species, *Hengchunia pakistanica* Asche & Webb 1994 (from India) was erroneously recorded from Pakistan.

In total 35 known genera are included in the key. Figures are also provided and additional figures can be found in Khatri and Webb (2010).

### Key to Dectocephalinae tribes and genera from Pakistan

Note: The characters given in the key will separate the Pakistan genera but not necessarily the tribes on a wider distribution. As the genera in the following key are grouped by tribe and as some of these are based only on the male genitalia, several genera in the latter part of the key are based only on the male.

- 1 Anterior margin of head with transverse striations or carinae (Figs 1j, l).....2
- Anterior margin of head smooth or shagreen .....7
- 2 Clypellus elongate, extending beyond margin of genae, tapered to apex and slightly bent subapically under head..... (**Grypotini**).....3
- Clypellus short not extending beyond margin of genae.....4
- 3 Vertex of head medially longer than next to eyes; ocelli equidistant between eye and median line of head; 5–6 mm in length.....  
..... **Sobipona Ghauri & Viraktamath**
- Vertex of head of uniform length; ocelli placed slightly closer to eye than to median line; less than 5 mm in length ..... **Pinopona Viraktamath & Sohi**
- 4 Antennae very long, extending beyond midlength of body, situated at upper corner of eyes (Fig. 1k).....  
..... **Drabescini (Paraboloponina) Dryadomorpha Kirkaldy**

- Antennae short, not reaching midlength of body, situated below upper corner of eyes..... **5**
- 5 Ocelli on foremargin of head bound both dorsally and ventrally by at least one carina (fig. 1j)..... **6**
- Ocelli located before foremargin of head, with carinae only ventrally.....  
..... **Penthimiini** *Penthimia* Distant
- 6 Head depressed anteriorly (Fig. 1j); forewing venation reticulate; aedeagus with one shaft ..... **Penthimiini** *Neodartus* **Melichar**
- Head not depressed anteriorly, forewing venation not reticulate; aedeagus with two shafts (Fig. 3b) ..... **Mukariini** (*Mukaria* Distant)
- 7 Robust species; vertex distinctly broader than long and only slightly longer medially than next to eyes (Fig. 1b); forewing appendix extending to outer apical cell (Fig. 1r); subgenital plates fused to each other and to valve (Fig. 2m); style apical process expanded apically; aedeagus fused to connective (Fig. 3e) ..... **Goniagnathini** (*Goniagnathus* Fieber)
- Without the above combination of characters..... **8**
- 8 Genae of face broad (Fig. 1h), visible in dorsal view (Fig. 1a) .....  
..... **Scaphytopiini**..... **9**
- Genae of face narrow, not visible dorsally..... **11**
- 9 Forewing obliquely truncate at apex; green species with red longitudinal stripes on head, thorax and forewings (Fig. 1e) ..... **Varta** Distant
- Forewing rounded at apex (Fig. 1o); colour not as above..... **10**
- 10 Pronotum with lateral carina..... **Grammacephalus** Haupt
- Pronotum without lateral carina ..... **Masiripius** Dlabola
- 11 Face with laterofrontal sutures directed mediad of and terminating distad of corresponding ocelli (Fig. 1g). Head, particularly in female, somewhat spatulate. Ovipositor with second valvulae, lacking teeth (Fig. 3l) ..... **Hecalini**..... **12**
- Without the above combination of characters..... **13**
- 12 Green species, male pygofer without caudal marginal stout setae (Fig. 2d) ....  
..... **Hecalus** Stål
- Brown species; male pygofer with caudal marginal stout setae.....  
..... **Glossocratus** Fieber
- 13 Vertex narrow basally (Fig. 1f). Male pygofer elongate, without a membranous laterobasal slit, with a lateroposterior triangular process (Fig. 2f); valve long; subgenital plate short; connective with stem two pronged apically, arms parallel (Fig. 2o); aedeagal shaft whip-like (Fig. 3d). Ovipositor with second valvulae lacking teeth (Fig. 3m) ..... **Stenometsopiini** (*Stirellus* Osborn & Ball)
- Without the above combination of characters..... **14**
- 14 Forewing brachypterous or if macropterous then appendix (when present) extending to fourth (outer) apical cell (Fig. 1m). Aedeagal shaft hinged (Fig. 3a) or not hinged; connective with arms looped (Fig. 2n). Ovipositor with sculpture granular, not extending to dorsal margin (Fig. 3n) .....  
..... **Chiasmini**..... **15**

–	Forewing appendix extending to second apical cell (Fig. 1n). Aedeagal shaft not hinged; connective with arms looped or divergent. Ovipositor with sculpture reticulate, extending to dorsal margin .....	<b>19</b>
15	Forewing with two subapical cells (Figs 1m) .....	<b><i>Aconurella</i> Ribaut</b>
–	Forewing with three subapical cells (Figs 1n, o) .....	<b>16</b>
16	Head margin depressed in lateral view .....	<b>17</b>
–	Head margin rounded in lateral view .....	<b>18</b>
17	Forewing lacking appendix. Aedeagal shaft not hinged at base with atrium; compressed in apical region with serrated margin .....	<b><i>Gurawa</i> Distant</b>
–	Forewing when well developed, with appendix. Aedeagal shaft hinged at base with atrium (Fig.3a), shaft cylindrical and lacking serration .....	<b><i>Chiasmus</i> Mulsant &amp; Rey</b>
18	Predominantly green species .....	<b><i>Nephotettix</i> Matsumura</b>
–	Predominantly pale brown species.....	<b><i>Exitianus</i> Ball</b>
19	Forewing with two subapical cells (Fig. 1q). Connective with arms divergent.....	<b>Macrostelini.....20</b>
–	Forewing with three subapical cells (Fig. 1n, o), sometimes outer subapical cell subdivided (Fig. 1s), or if two subapical cells connective with arms convergent apically .....	<b>22</b>
20	Vertex of head short, of uniform length, more than 4 times wider than long (Fig. 1c) .....	<b><i>Balclutha</i> Kirkaldy</b>
–	Vertex longer medially than next to eyes, twice or less as wide as medial length .....	<b>21</b>
21	Head and thorax golden yellow, vertex with two round dark brown spots (Fig. 1d). Male pygofer with long, slender hook-like process; without marginal comb-like serrations (Fig. 2a).....	<b><i>Cicadulina</i> China</b>
–	Head and thorax not as above. Male pygofer without above process; with comb-like serrations on caudal margin (Fig. 2b) .....	<b><i>Macrosteles</i> Fieber</b>
22	Aedeagus with two shafts. ....	<b>Opsiini.....23</b>
–	Aedeagus with one shaft.....	<b>25</b>
23	Aedeagal shafts fused in basal half, then divergent describing a circle (Figs 3c) .....	<b><i>Nealiturus (Circulifer)</i> Distant</b>
–	Aedeagal shafts separate at base (Figs 3g, 3h) .....	<b>24</b>
24	Aedeagal shaft with ventral pair of processes (Figs 3g, 3j) .....	<b><i>Opsius</i> Fieber</b>
–	Aedeagal shaft without ventral pair of processes (Fig. 3h)....	<b><i>Orosius</i> Distant</b>
25	Connective with arms divergent (Fig. 2k ). ....	<b>Athysanini.....26</b>
–	Connective with arms at most only divergent basally, convergent apically ...	<b>32</b>
26	Vertex acutely pointed, disc depressed; forewing venation reticulate (Fig. 1s) ...	<b><i>Platymetopius</i> Burmeister</b>
–	Vertex and forewings not as above.....	<b>27</b>
27	Three to four cross veins from outer apical cell reaching costal margin (Fig. 1n).....	<b>28</b>
–	At most two cross veins from outer apical cell reaching costal margin .....	<b>29</b>

- 28 Male connective with paraphyses (Fig. 2g) ..... **Scaphoideus** Uhler  
 – Male connective without paraphyses (Fig. 2k)..... **Bampurius** Dlabola  
 29 Male subgenital plates with mesal sclerotized process (Fig. 2h).....  
 ..... **Neolimnus** Linnavuori  
 – Male subgenital plates without mesal sclerotized process..... **30**  
 30 Aedeagal shaft without basal processes, apophysis of style long, subequal to  
 length of aedeagal shaft (Fig. 2e) ..... **Pseudosubhimalus** Ghauri  
 – Aedeagal shaft with basal pair of dorsal (Fig. 3i) or ventral (Fig. 3k) processes,  
 apophysis of style shorter than aedeagal shaft ..... **31**  
 31 Aedeagal shaft with basal processes arising on dorsal surface of shaft (Fig. 3i)...  
 ..... **Monobazus** Distant  
 – Aedeagal shaft with basal process arising on ventro-lateral surface of shaft  
 (Fig. 3k)..... **Osbornellus** Ball  
 32 Connective fused to aedeagus (Fig. 3f). ..... **Deltocephalini**..... **33**  
 – Connective not fused to aedeagus (Fig. 2i)..... **Paralimnini**..... **34**  
 33 Vertex with transverse black stripe; pygofer with dorsal marginal appendage...  
 ..... **Paramesodes** Ishihara  
 – Vertex without transverse black stripe; pygofer without appendage (Fig. 2c) ...  
 ..... **Maiestas** Distant  
 34 Male subgenital plates short (Fig. 2i); mesal arm of style longer than outer  
 arm (Fig. 2j)..... **Psammotettix** Haupt  
 – Male subgenital plate longer; mesal arm of style shorter than outer arm (Fig.  
 2l)..... **Jilinga** Ghauri

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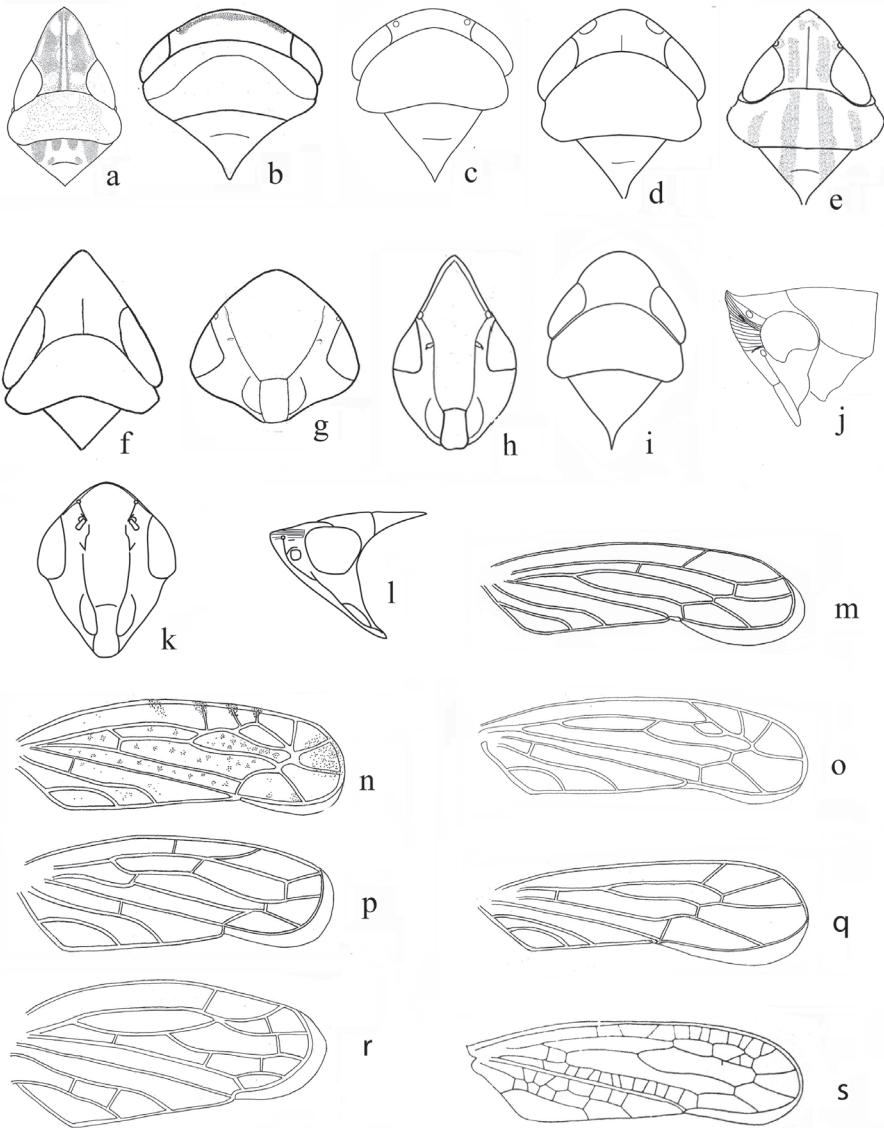
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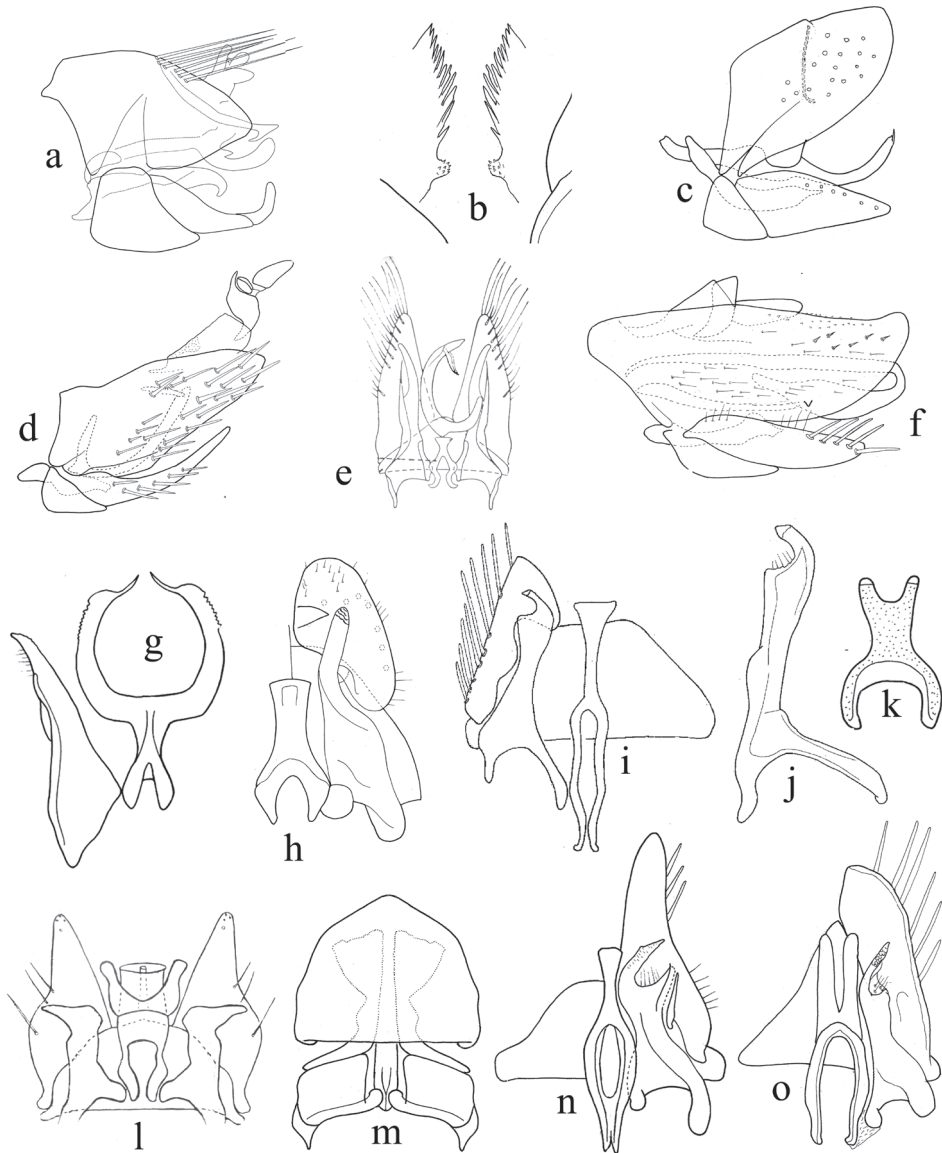
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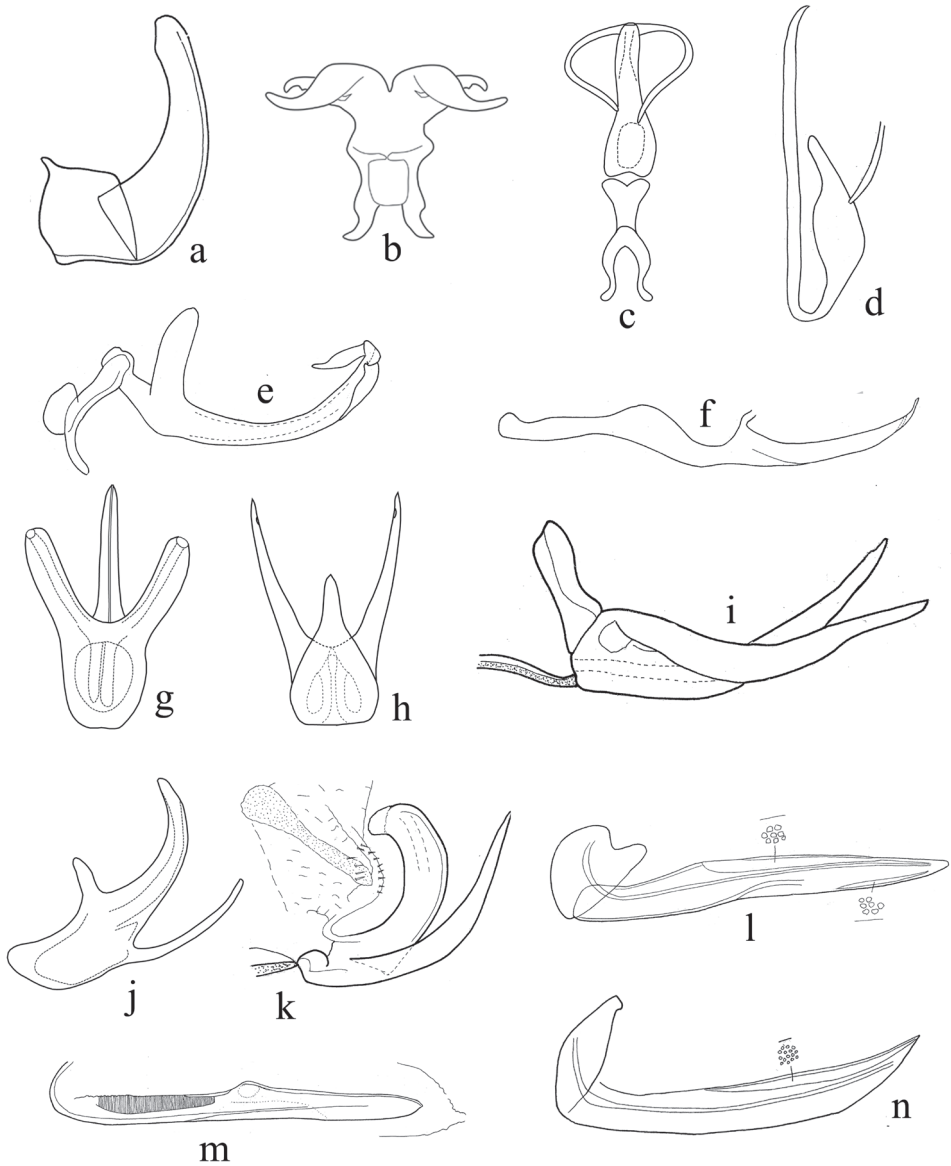
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**Figure 1.** a–f, i (head and thorax dorsal view); g, h, k (face); j, l (head and thorax lateral view); m–s (right forewing) **A** *Grammacephalus indicus* Viraktamath & Murthy **B** *Goniagnathus (T.) quadripinnatus* Dash & Viraktamath **C** *Balclutha incisa* (Matsumura, 1902) **D** *Cicadulina bipunctata* Ghauri **E** *Varta rubrofasciata* Distant **F** *Stirellus lahorensis* (Distant) **G** *Hecalus sindhensis* (Ahmed & Aziz) **H** *Grammacephalus indicus* Viraktamath & Murthy **I** *Mukaria splendida* Distant **J** *Neodartus acocephaloides* Melichar **K, L** *Dryadomorpha pallida* Kirkaldy **M** *Aconurella prolixa* (Lethierry) **N** *Bampurius pakistanicus* Khatri & Webb **O** *Grammacephalus indicus* Viraktamath & Murthy **P** *Chiasmus* sp. **Q** *Macrosteles indrina* (Pruthi) **R** *Goniagnathus (T.) quadripinnatus* Dash & Viraktamath **S** *Platymetopius* sp.



**Figure 2.** a–o male genitalia. a, c, d, f (genital capsule); b (pygofer, caudal view); e (genital capsule, internal view); g (connective and style); h, i, n, o (valve, style and connective, dorsal view). j (style); k (connective); l (valve, styles, subgenital plates, connective and base of aedeagus); m (fused subgenital plates and valve (setae omitted), styles and base of connective). **A** *Cicadulina bipunctata* Ghauri **B** *Macrosteles indrina* (Pruthi) **C** *Maiestas pruthii* (Metcalf) **D** *Hecalus sindhensis* (Ahmed & Aziz) **E** *Pseudosubhimalus bicolor* Pruthi **F** *Stirellus lahorensis* (Distant) **G** *Scaphoideus harlani* Kitbamroong & Freytag **H** *Neolimnus quadricornis* Khatri & Webb **I, J** *Psammotettix emarginata* Singh **K** *Bampurius pakisticus* Khatri & Webb **L** *Jilinga gopii* (Pruthi) **M** *Goniagnathus (T.) quadripinnatus* Dash & Viraktamath **N** *Aconurella prolixa* (Lethierry) **O** *Stirellus lahorensis* (Distant).



**Figure 3.** a–n male and female genitalia; a, d, g–j (aedeagus); b–c, e–f (aedeagus and connective); k aedeagus, apex of connective and pygofer; l, n (first valvulae); m (second valvulae). **A** *Aconurella prolixa* (Lethierry) **B** *Mukaria splendida* Distant **C** *Neolaliturus (Circulifer) tenellus* (Baker) **D** *Stirellus lahorensis* (Distant, 1918) **E** *Goniagnathus (T.) quadripinnatus* Dash & Viraktamath, 2001 **F** *Maiestas tareni* (Dash & Viraktamath) **G** *Opsius versicolor* (Distant) **H** *Orosius albicinctus* Distant **I** *Monobazus dissimilis* (Distant) **J** *Opsius versicolor* (Distant) **K** *Osbornellus (Mavromoustaca) macchiaie* (Lindberg) **L** *Hecalus sindhensis* (Ahmad & Aziz) **M** *Stirellus lahorensis* (Distant) **N** *Aconurella prolixa* (Lethierry).

# Two new species of *Pterostichus* Bonelli subgenus *Pseudoferonina* Ball (Coleoptera, Carabidae, Pterostichini) from the mountains of central Idaho, U.S.A.

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## Abstract

Two new species of *Pterostichus* Bonelli subgenus *Pseudoferonina* Ball, are described from the mountains of central Idaho: *Pterostichus bousqueti* Bergdahl [type locality = small tributaries of South Fork of Payette River watershed, ca. 1170 m (3840 ft), 44.0675°/-115.6822°, near Lowman, Salmon River Mountains, Boise County, Idaho, U.S.A.] and *Pterostichus lolo* Bergdahl [type locality = Cottonwood/Orogrande Creek, ca. 870 m (2850 ft), 46.5528°/-115.5522°, North Fork of Clearwater River watershed, Clearwater Mountains, near Bungalow, Clearwater County, Idaho, U.S.A.]. Males of *P. bousqueti* and *P. lolo* are easily distinguished from each other and the seven previously described *Pseudoferonina* species by the form of the median lobe of the aedeagus, and from most individuals of the other species of *Pseudoferonina* in Idaho by features of pronotal shape and macrosculpture. Both species appear to be obligate ripicolous hygrophiles, restricted in distribution primarily to the margins of small montane streams in forested areas. Widespread intensive stream surveys for *Pseudoferonina* over many years indicate the geographic ranges of both species are highly localized, and additional undescribed species may occur in Idaho.

## Keywords

Coleoptera, Carabidae, *Pterostichus*, *Pseudoferonina*, *Melvilleus*, North America, Pacific Northwest, Idaho, Salmon River Mountains, Clearwater Mountains, flightlessness, endemism, montane streams

## Introduction

*Pterostichus* Bonelli (1810) is one of the largest genera of carabid beetles in North American north of Mexico, including about 200 described species and subspecies, classified in about 20 species groups and at least as many recognized subgenera (Bousquet and Laroche 1993; Bousquet 1999). They represent almost 8% of this region's carabid fauna (ca. 2640 species and recognized subspecies). The *Pterostichus* subgenus *Pseudoferonina* Ball (1965) (type species = *Pterostichus lanei* Van Dyke 1925) is one of three *Pterostichus* subgenera narrowly restricted (endemic) to the Pacific Northwest (Washington, Oregon, Idaho and British Columbia), and the most diverse (Bergdahl 1995). Adults of all *Pseudoferonina* species are strict ripicolous hygrophiles whose habitat is the margin of small, forested headwater streams in mountainous terrain. They can be found from near sea level to subalpine elevations, but especially mid-montane zones where low (0–2) order streams (Strahler 1957) are common. None of the species are known to be alpine.

The carabid fauna of the Pacific Northwest [ca. 705 recognized species and subspecies (Bousquet and Laroche 1993, Bergdahl 1995)] is reasonably well known. Most of the species were described many decades ago (mean date of publication = ca. 1865), and any remaining undiscovered species are probably very localized and difficult to find (e.g., Kavanaugh and Labonte 2006; Labonte 2006), or cryptic sibling species. *Pseudoferonina* is one of the least understood and most fascinating groups of closely related carabids in the Pacific Northwest. Over the last 26 years, 6 of the last 13 new carabid species to be described from the Pacific Northwest are members of subgenus *Pseudoferonina*.

*Pseudoferonina* includes six previously described species that appear to form a distinct group: *P. lanei* (Van Dyke 1925), *P. humidulus* (Van Dyke 1943), *P. shulli* (Hatch 1949), *P. campbelli* Bousquet (1985), *P. smetanai* Bousquet (1985), and *P. spathifer* Bousquet (1992) (Bousquet and Laroche 1993). The subgenus also includes *Pterostichus vexatus* Bousquet (1985), which both Hatch (1953) and Ball (1965) misidentified as *P. shulli*. Males of *P. vexatus* differ from those of other members of the subgenus in having: 1) a short median lobe, with apical lamella flattened dorso-ventrally (not laterally), 2) the gonopore terminal, not basal, on the internal sac, 3) the internal sac everted dorsally, not ventrally, and 4) lacking the semicircular indentation (flattening) of the last visible abdominal sternite that is distinct in males of all of the other species. Based on these features, Ball (1965) described a new subgenus for *P. vexatus* (i.e., his "*P. shulli*"), *Melvilleus* Ball, which is currently considered a subjective junior synonym of *Pseudoferonina* (Bousquet 1985, 1992, 1999, Bousquet and Laroche

1993). Among species of *Pseudoferonina* found in Idaho, *P. vexatus* has the broadest geographical and habitat ranges.

The purpose of this paper is to describe two new species of *Pseudoferonina* from the Clearwater and Salmon River mountains of central Idaho, including information about form and structure, geographical and habitat distributions, and way of life. Hatch (1953) provided a key to the carabids of the Pacific Northwest that includes *P. lanei* and *P. vexatus*. Lindroth (1966) provided excellent keys to most of the subgenera and species of *Pterostichus* in the Pacific Northwest, but he did not include *Pseudoferonina*. Bousquet (1985) provided a key for identification of adults of the seven *Pseudoferonina* species known to him and later modified that key (Bousquet 1992). In this paper we suggest modifications to his key to accommodate the two new species we describe.

## Materials and methods

All specimens of the new species described here were collected by the first author [JCB] in 1999, 2009 or 2010. These specimens were acquired by timed hand collecting along small streams in mountainous terrain (e.g., Dupuis and Friele 2006), turning over rocks and organic debris primarily with a handheld garden fork, and flooding small gravel bars (Darlington 1971) using a plastic gold pan. Specimens were preserved in the field in a mixture of 70% ethanol + 30% white vinegar. As soon as possible in the lab, specimens were washed in warm water and then stored in fresh aliquots of the same ethanol/vinegar mixture to await preparation. Stream names and the names of other geographic features were derived from 1:100,000 United States Geographic Survey topographic maps and United States Forest Service forest district road maps. Geographical coordinates and elevations for collecting sites were determined using Google Earth ([www.earth.google.com](http://www.earth.google.com)).

Specimens were examined using an 8–50x Nikon dissecting scope with a high-intensity light tube. The only measurement recorded was apparent body length (ABL), measured from the apex of the labrum to the apex of the longer elytron. Digital photographs of dorsal habitus and male genitalia were taken using an Auto-montage imaging system by Syncroscopy with a Leica M420 dissecting microscope.

Specimens examined in the course of this study are deposited in the following collections:

<b>CAS</b>	California Academy of Sciences, San Francisco, California, U.S.A.
<b>CMNH</b>	Carnegie Museum of Natural History, Pittsburgh, Pennsylvania, U.S.A.
<b>CNC</b>	Canadian National Collection, Ottawa, Ontario, Canada
<b>JCB</b>	James C. Bergdahl Collection, Spokane, WA, U.S.A.
<b>OSU</b>	Oregon State University Arthropod Collection, Corvallis, Oregon, U.S.A.
<b>UICM</b>	University of Idaho, Moscow, Idaho, U.S.A.
<b>WSU</b>	Washington State University, Pullman, WA, U.S.A.

## Taxonomy

### *Pterostichus (Pseudoferonina) bousqueti* Bergdahl, sp. n.

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[http://species-id.net/wiki/Pterostichus\\_\(Pseudoferonina\)\\_bousqueti](http://species-id.net/wiki/Pterostichus_(Pseudoferonina)_bousqueti)

Figs 1, 3A, 4A, 4C, 5, 6

**Type locality.** U.S.A., Idaho, Boise County, small tributaries of the South Fork of the Payette River near Lowman, ca. 1200 m (3940 ft) elevation, 44.0675°/-115.6822°.

**Type material.** Holotype: a male, deposited in CAS, labeled: “USA: Boise Co., Pine Flat Creek at South Fork Payette River Road, 6 km W of Lowman, 44°04'03"N at 115°40'56"W, T9N, R7E, S31, elev. 1160 m (3805 f), South Fork Payette River Basin, Boise National Forest, 05 May 1999, #26-1999, J. C. Bergdahl, coll.”/ “HOLOTYPE *Pterostichus bousqueti* Bergdahl designated 2011” [red label]. Paratypes: A total of 11, with 8 males and 1 female with the same data as the holotype, and 2 males labeled: “USA: Boise Co., Archie Creek near confluence with SF Payette R., 8 km E of Lowman, Boise Natn. Forest, 44°04'15"N at 115°31'06"W, T9N R8E S33, elev. 1240 m (4070 ft), South Fork Payette R. watershed, 06 May 1999, #33-1999, J. C. Bergdahl, coll.” The single female paratype is deposited in CAS, the male paratypes in CNC, CMNH, OSU, UICM, WSU, and JCB.

**Etymology.** The specific epithet, *bousqueti*, is a Latinized version (genitive case) of the surname of Dr. Yves Bousquet (Agriculture Canada, Ottawa, Ontario, Canada), in recognition of his outstanding contribution (Bousquet 1985) to our knowledge of *Pseudoferonina*, which brought order to the chaos in taxonomy of the species known at that time.

**Diagnosis.** A *Pterostichus* with the characteristics of members of subgenus *Pseudoferonina* (see Ball 1965, and Bousquet 1999); males easily distinguished from those of all other species of *Pseudoferonina* by the shape of the median lobe of the aedeagus, especially the apical lamella (apex), which is shaped (in left lateral view) like the blade of a hatchet or tomahawk (Figs 4A and 4C). Also, the shaft of the median lobe in lateral view is thicker in the middle than at either end (where it joins to the base or the apical lamella) and lacks the distinct mid-shaft sinuation (Bousquet 1985, 1992) seen in *P. shulli*, *P. spathifer*, the following new species (*P. lolo*), and, to a lesser degree, *P. smetanai*. The small size and thin body form of *P. bousqueti* adults are most similar to *P. campbelli* adults, which occur only west of the Cascade Range in Oregon.

**Description.** Size medium-small for subgenus, ABL males 8.5–9.5 mm, female 8.9 mm. Body form slightly slender for subgenus (Fig. 1).

**Color.** Dorsal body surface black and shiny, pronotal lateral beads piceous, legs, antennae and palpi piceous.

**Microsculpture.** Head with isodiametric meshes on frons; pronotum with faintly impressed transverse meshes on most of disc, meshes more isodiametric basolaterally; elytra with markedly transverse meshes, faintly iridescent in some areas.

**Pronotum.** Fig. 3A. Almost as long as wide, lateral margins subparallel anterior to hind angles, slightly sinuate, hind angles rectangular or slightly acute, lateral margins (in lateral



view) bent ventrally near hind angles. Anterior and posterior transverse impressions present but faintly impressed, median line distinctly impressed, almost entire, but not quite extended to anterior and posterior margins in most specimens examined; anterior transverse impression faintly punctulate. Lateral margins finely beaded, posterior margin without margination, anterior margin minutely beaded in medial one-third. Basolateral fovea deep and broadly linear, slightly convergent toward midline anteriorly, not or only faintly punctulate.

*Elytra*. Intervals flat or only slightly convex, but less so than in any other *Pseudoferonina* species.

*Legs*. Male mesotibiae slightly curved apically.

*Abdomen*. Last visible sternite of male with broad, shallow medial indentation, anterior margin of depression not at all carinate, sternite without protuberances; last visible sternite of males with single pair of anal setae, female with two pairs.

*Male genitalia*. Median lobe of aedeagus as in Figs 4A and 4C. Apical one-third of median lobe (in ventral view) with lightly sclerotized oblique band extended longitudinally next to slight ridge (Fig. 4C); shaft of median lobe (in ventral view) swollen submedially on left, with shaft gradually tapered from swelling to the apical lamella, ventral margin not sinuate subapically (in lateral view); apical lamella average in length for *Pseudoferonina* species in Idaho, symmetrically hatchet-shaped (in left lateral view). Right paramere short and sharply pointed.

**Geographic distribution.** At this time, *P. bousqueti* is known only from Pine Flat Creek and Archie Creek near their confluences with the South Fork of the Payette River near Lowman, Boise County, Idaho (Fig. 5). The Payette River is a tributary of the Snake River, which is a major tributary of the Columbia River. To what extent the range of this species is narrowly restricted to this area is not known. Despite sampling many other creeks in this area, JCB has been unable to document any other *P. bousqueti* populations. Additional areas, if any, in which this species is most likely to occur include the Big Pine, Deadwood, Clear Creek and Middle Fork of the Payette River watersheds to the west of Lowman and east of the North Fork of the Payette River.

**Habitat.** The type locality along the South Fork of the Payette River near Lowman is in a steeply sided, sparsely pine-forested gorge (Fig. 6A) that has experienced intense forest fires. From 1908 to 2000, canopy fires burned more than 50% of the Boise National Forest in central Idaho. Large fires have a huge effect on vegetation, geomorphology, stream hydrographs (Pierce et al. 2004), and undoubtedly *P. bousqueti* habitat. Fire legacies of this extent on the landscape probably create severe barriers to dispersal of *Pseudoferonina* species. Areas at low to mid-elevations along the South Fork of the Payette River form a small, isolated outlier of the Hot Dry Canyons eco-region of the Idaho Batholith (McGrath et al. 2002). Most of this eco-region is associated with the main stem Salmon River, 145 km (90 miles) to the north. McGrath et al. (2002) describe this eco-region as: unglaciated, deep, precipitous canyons; annual precipitation = 305–889 mm (lowest in deep canyons); mean temperature: Jan. (min/max) = -8.9/0.6°C, July (min/max) = 8.8/31.7°C (warmer with increasing canyon depth). The vegetation is characterized by open ponderosa pine forest and sagebrush scrub on south facing-slopes, and ponderosa pine/Douglas fir forest on north-facing slopes.



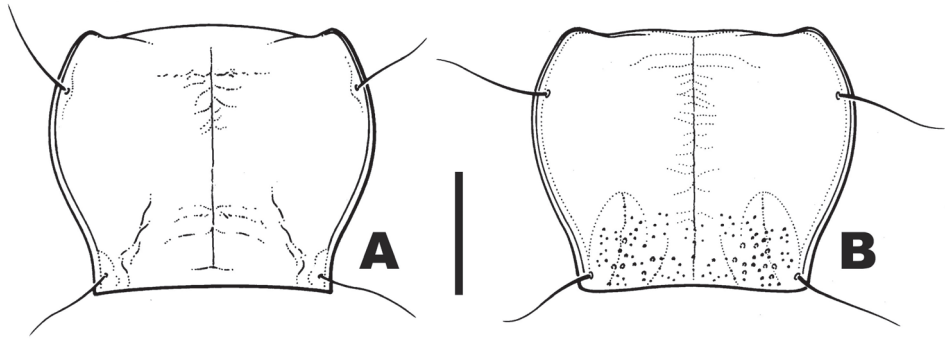
**Figure 1.** Holotype male, *Pterostichus (Pseudoferonina) bousqueti* Bergdahl, sp. n., dorsal habitus. Scale line = 1.0 mm. Automontage digital image by D. H. Kavanaugh.

Both Pine Flat Creek (Fig. 6B) and Archie Creek are small, perennial streams descending steep canyon slopes. Their lower sections run thru dry open ponderosa pine forests. At higher elevations on sun and wind-exposed south-facing slopes, the vegetation is primarily open scrub. Pine Flat Creek descends a steep scrubby hillside; pine forest is restricted to lower reaches near the Payette River. Archie Creek descends its watersheds at a gentler grade. Both of these creeks near their confluence with Payette River are only about 1.5 m bank-full-width, and are confined primarily to channels with stabilized vegetated banks. Consequently, there are very few gravel bars on which to search for beetles, even at low water. High water for these creeks most likely occurs



**Figure 2.** Holotype male, *Pterostichus* (*Pseudoferonina*) *lolo* Bergdahl, sp. n., dorsal habitus. Scale line = 1.0 mm. Automontage digital image by D. H. Kavanaugh.

March-May, during rapid melt of the snow at higher elevations on adjacent hillsides coincident with wet spring storms. Because the watersheds these creeks service are so small, large thunderstorms and cloudbursts, by themselves, also may cause significant peaks in runoff during summer and fall months after the snow pack has melted at high elevation. In exceptionally dry years, both of these creeks may actually dry up to the extent that even lower sections are without surface-flowing water. However, on 25



**Figure 3.** Pronotum, dorsal aspect **A** *Pterostichus (Pseudoferonina) bousqueti* Bergdahl, sp. n. **B** *Pterostichus (Pseudoferonina) lolo* Bergdahl, sp. n. Scale line = 1.0 mm. Line drawings by Go Sato.

September (2008) both creeks had nearly one meter of surface water stream width. Given their small watersheds, it is likely that major sections of both streams, especially high up where slopes are steep and the vegetation scrubby, dry up even in average years.

Other carabid species collected by JCB at or near the type locality include: forest habitat - *Psydus piceus* LeConte, *Pterostichus protractus* LeConte; streamside habitats - *Platynus brunneomarginatus* (Mannerheim), *Bembidion iridescens* LeConte, *Bembidion kuprianovi* Mannerheim, *Bembidion* sp., *Diplous aterrimus* (Dejean), *Nebria* sp., *Bradycellus californicus* (LeConte), *Bradycellus nigrinus* (Dejean), *Bradycellus nubifer* LeConte. The trout-stream beetle *Amphizoa insolens* LeConte (Coleoptera, Amphizoidae) also occurs along creeks in this area.

**Phenology and larval ecology.** JCB visited Pine Flat Creek and Archie Creek on two occasions: 05–06 May (1999) and 25 September (2008). In early May, he easily found adults of *P. bousqueti* at both sites, however in September collecting along these same stream sections on both creeks yielded no specimens. *Pseudoferonina* populations in hot dry regions at low elevations appear to aestivate or go into early hibernation, probably in August. Very few teneral specimens have been collected; however, males with soft, poorly chitinized median lobes are common in late summer and fall. This suggests that adults emerge from pupal chambers in late summer through early fall and overwinter as adults. Presumably they escape winter-spring floods by migrating out of the floodplain to higher positions along stream banks to hibernate in the fall before freeze-up, however they probably do not go very far. Once snow packs melt and daytime temperatures approach 10°C (late March and early April at lower elevations in Idaho), adults again become active on the surface. Given the fact that surface-active *Pseudoferonina* adults are so closely associated with the immediate vicinity of stream margins, the larvae are probably also narrowly restricted to these habitats and may spend a substantial amount of time subsurface at or very near the hyporheic zone.

**Dispersal power.** All known individuals are brachypterous (flightless) and restricted to the margins of small, isolated, forested streams, so dispersal power of *P. bousqueti* adults is expected to be extremely low. Like *P. shulli*, *P. spathifer* and *P. lolo* sp. n., the geographic range of *P. bousqueti* (Fig.5) appears to be highly restricted.

**Remarks.** Among Idaho's *Pseudoferonina* species, *P. bousqueti* appears to be a member of the *lanei* species-group (with *P. lanei*), as opposed to the *shulli* species-group (*P. shulli*, *P. spathifer* and *P. lolo* sp. n.). Idaho species in the *shulli* group have a distinct ventral sinuation (bulge and concavity) on the ventral surface of the median lobe of the aedeagus near the middle of the shaft (easily seen in lateral view). Species in the *lanei* group (and *P. vexatus*) do not have this feature. To what extent this feature, or its absence, is indicative of phylogenetic relationship is presently unknown.

Property ownership in the South Fork Payette River area in vicinity of the type locality is primarily the U.S. Forest Service (Boise National Forest), however according to the Boise National Forest map there are many small in-holdings along the river. Two of these in-holdings include both lower Pine Flat and lower Archie creeks, near where the type specimens were collected. Both of these creeks probably have experienced prospecting or mining activity. There is a small Forest Service campground and hot springs very near Pine Flat Creek on the north bank of the Payette River, but visitors seem to have little impact on the creek.

Because *P. bousqueti* is known from only two easily identifiable localities, we hope that collectors will exercise restraint when sampling at these sites, and instead focus on discovering new sites for this species in the immediate vicinity, such as in those watersheds listed above.

***Pterostichus* (*Pseudoferonina*) *lolo* Bergdahl, sp. n.**

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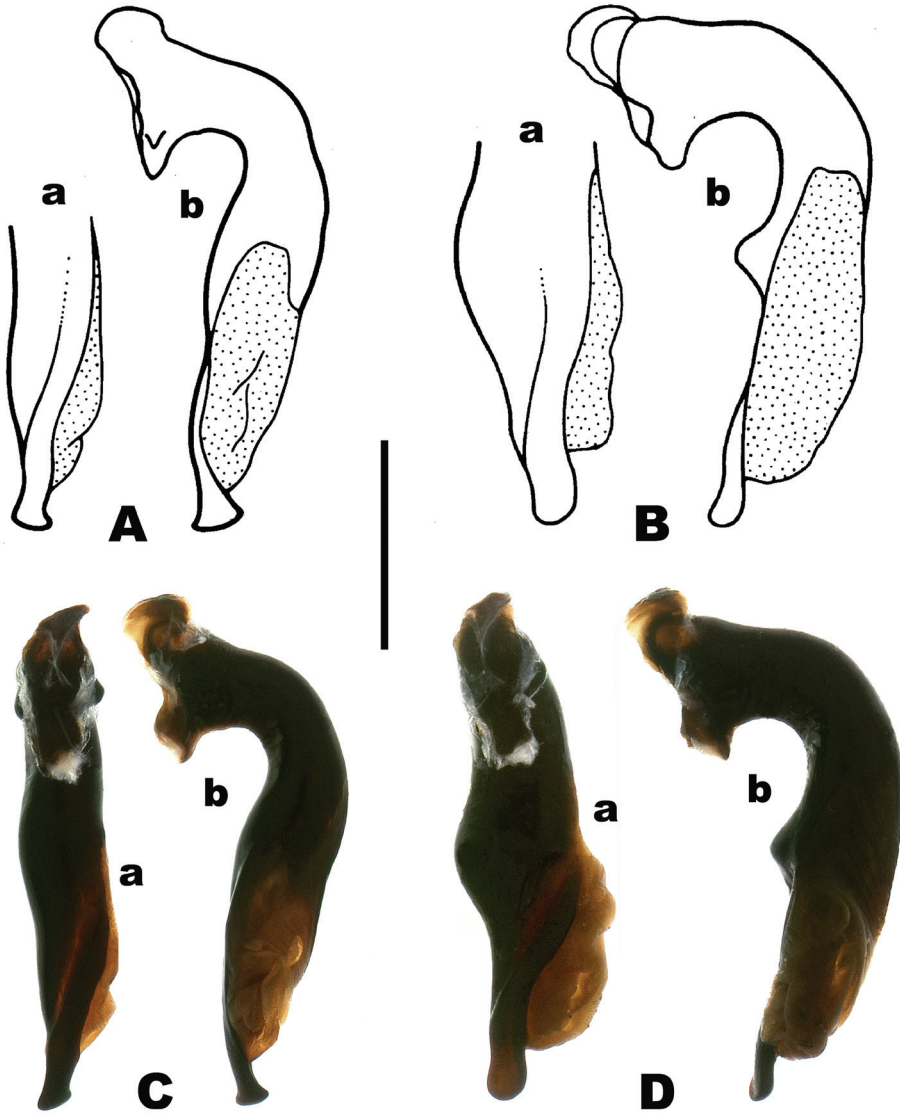
[http://species-id.net/wiki/Pterostichus\\_\(Pseudoferonina\)\\_lolo](http://species-id.net/wiki/Pterostichus_(Pseudoferonina)_lolo)

Figs 2, 3B, 4B, 4D, 5, 7

**Type locality.** U.S.A, Idaho, Clearwater County, Cottonwood Creek near the confluence of Orogrande Creek and the North Fork of the Clearwater River, ca. 870 m (2860 ft) elevation, 46.5528°/-115.5522°.

**Type material.** Holotype, a male, deposited in CAS, labeled: "USA: IDAHO, Clearwater Co.; Cottonwood Creek @ USFS Rd 250; Orogrande Ck. Rd.; NF Clearwater R. watershed; elev. ~870 m (2860 ft); 12.5 MI. ESE Headquarters, 3.5 MI. SW of Bungalow; 46°33'10"N, 115°33'08"W; T38N, R7E, S34; 17 July 2009; #73-2009; J. C. Bergdahl, coll."/ "HOLOTYPE *Pterostichus lolo* Bergdahl designated 2011" [red label]. Paratypes: A total of 22 paratypes from same locale as the holotype (7 males and 5 females from 17 July 2010; 2 males and 9 females from 03 July 2010) deposited in CAS, CMNH, CNC, JCB, OSU, UICM and WSU.

**Etymology.** The specific epithet, *lolo*, is a noun in apposition, and refers to the Lolo Trail, an ancient hunting and trade route developed by the Nez Perce tribe to cross the Bitterroot Mountains from their homelands on the Clearwater River near Kamiah, Idaho, to the buffalo herds of Montana. The beginning of this trail in its western foothills starts in the upper Lolo River watershed, near the type locality of *P. lolo*. The Lolo Trail is also the route the Lewis and Clark Expedition took over the Bitterroot Mountains heading west in September 1805 and east in June 1806.



**Figure 4.** Median lobe of male aedeagus **A–B** line drawings, a = apical region, ventral view; b = left lateral view **C–D** Automontage digital images, a = ventral view, b = left lateral view **A** *Pterostichus* (*Pseudoferonina*) *bousqueti* Bergdahl sp. n. **B** *P. lolo* Bergdahl sp. n. **C** *P. bousqueti* **D** *P. lolo*. Scale line = 1.0 mm. Line drawings by G. Sato; digital images by D. H. Kavanaugh.

**Diagnosis.** A *Pterostichus* with the characteristics of members of subgenus *Pseudoferonina* (see Ball 1965; and Bousquet 1999); males easily distinguished from those of all other species of *Pseudoferonina* by the shape of the median lobe of the aedeagus, especially the form of the mid-shaft sinuation and associated convexity (Figs 4B and 4D), which is markedly and abruptly convex, nearly tuberculate (in lateral view), and

the shape of the apical lamella (apex), which is broader in ventral view than in *P. shulli* males (see Bousquet 1992, Fig. 16) and narrower in lateral view than in *P. spathifer* males (see Bousquet 1992, Fig. 15).

**Description.** Size medium-large for subgenus, ABL males = 9.5–10.5 mm, females = 9.0–10.3 mm. Body form average for subgenus (Fig. 2).

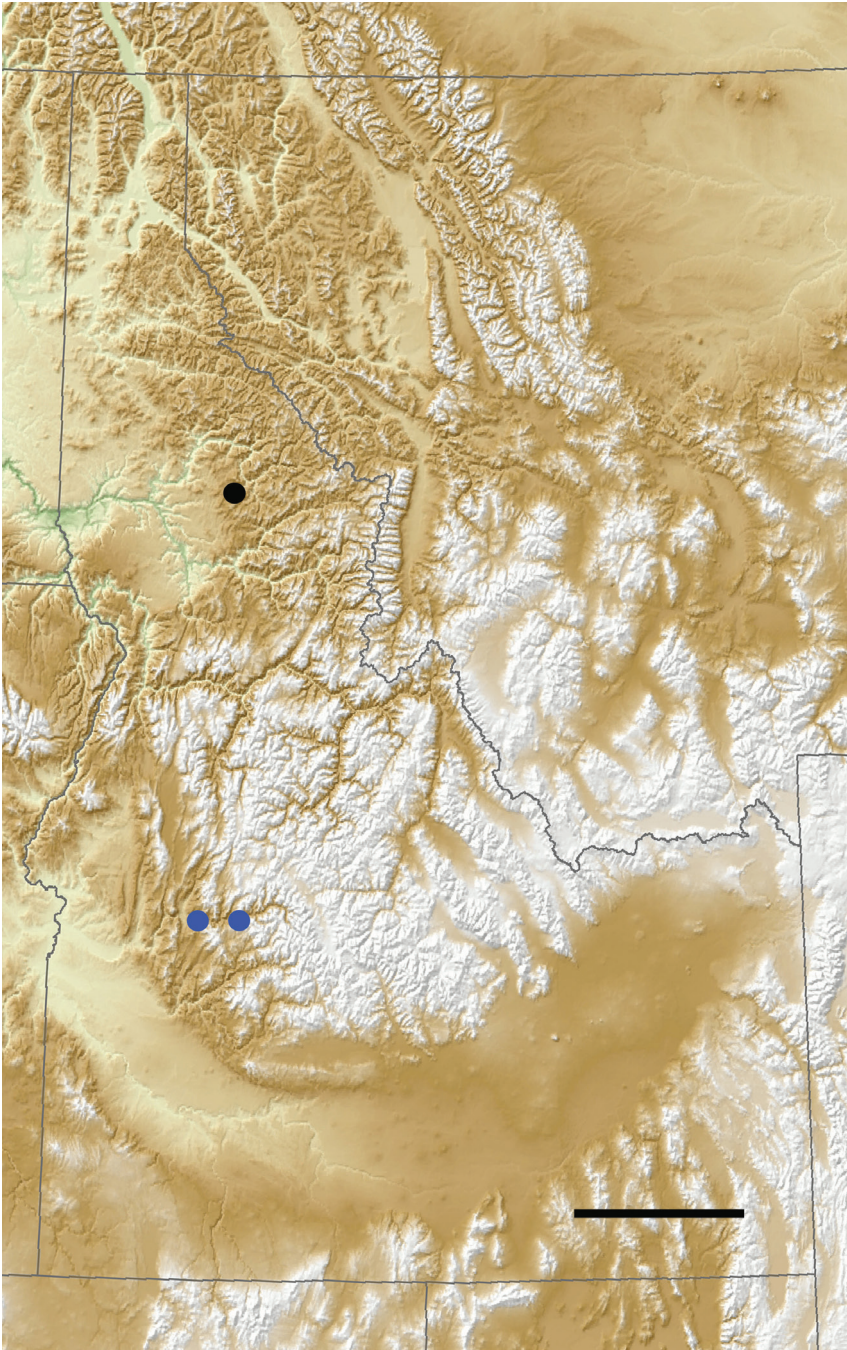
*Color.* Dorsal surface black and shiny, pronotal lateral beads piceous, antennae and palpi rufous or rufopiceous, femora piceous except distally rufopiceous, tibiae and tarsi rufopiceous or rufous.

*Microsculpture.* Head with faintly impressed isodiametric meshes on frons; pronotum with moderately impressed transverse meshes on most of disc, meshes more isodiametric posteriorly; elytra with moderately impressed transverse meshes and very slight iridescence. *Pronotum.* Fig. 3B. Almost as long as wide, widest point approximately at anterior one-third, lateral margins slightly sinuate in posterior half, almost parallel before hind angles, hind angles slightly obtuse to subrectangular, lateral margins (in lateral view) bent ventrally near hind angles, basal one-quarter of pronotum finely but distinctly punctate. Anterior and posterior transverse impressions faintly or not at all impressed, median longitudinal impression distinctly impressed, nearly extended to anterior and posterior margins in most specimens examined. Lateral margins finely beaded, posterior margin without margination, anterior margin minutely beaded in lateral thirds, without margination in medial one-third. Basolateral fovea deep and broadly linear, slightly convergent toward midline anteriorly, finely but distinctly punctate, areas between basal fovea and lateral margins convex, subcarinate.

*Elytra.* Intervals nearly flat, striae entire, moderately impressed, not or only very faintly punctulate. *Legs.* Male mesotibiae slightly curved apically. *Abdomen.* Last visible sternite of male with broad, shallow medial indentation, anterior margin of depression slightly carinate, sternite without protuberances; last visible sternite of males with single pair of anal setae, female with two pairs.

*Male genitalia.* Median lobe of aedeagus as in Figs 4B and 4D. Apical one-third of median lobe (in ventral view) with lightly sclerotized oblique band extended longitudinally next to slight ridge (Fig. 4Da); shaft of median lobe (in ventral view) markedly swollen submedially on left, with shaft abruptly tapered from swelling to the apical lamella, ventral margin distinctly sinuate subapically (in lateral view) with markedly abrupt, subtuberculate convexity basad of sinuation (Fig. 4Db); apical lamella average in length for *Pseudoferonina* species in Idaho, apex evenly rounded, very slightly swollen in ventral view (Fig. 4Da), narrow and very slightly reflexed ventrally at tip in lateral view (Fig. 4Db). Right paramere as illustrated by Bousquet (1992, Figs 15b and 16b) for *P. spathifer* and *B. shulli*.

**Geographic distribution.** Currently, *P. lolo* is known only from the type locality on Cottonwood Creek (at 870 m), which is a small, low-order tributary of Orogrande Creek, which is itself a tributary of the North Fork of the Clearwater River. In turn, the Clearwater is a tributary of the Snake River, which is a major tributary of the Columbia River. To what extent the range of this species is narrowly restricted to this area is not known. Despite sampling many other creeks in this area over the past decade, JCB has been unable to document any other *P. lolo* populations.



**Figure 5.** Toporelief map of Idaho illustrating the locations of known localities for *Pterostichus* (*Pseudoferonina*) *bousqueti* Bergdahl, sp. n. (blue dots) and *P. lolo* Bergdahl sp. n. (black dot). Scale line = 100 km.





**Figure 6.** Photographs of habitat for *Prerostichus* (*Pseudoferonina*) *bousqueti* Bergdahl, sp. n. **A** View north over South Fork Payette River gorge toward Deadwood Ridge above Lowman, Boise County, Idaho, 5 May 1999 **B** Pine Flat Creek, near Lowman, Boise County, Idaho, 25 September 2008. Photographs by J. C. Bergdahl.

**Habitat.** The type locality is in the Clearwater Mountains and Breaks section of the Northern Rockies eco-region (Nesser et al. 1997). This area has mountains of moderate relief with rounded landform, and steep breaklands. Elevations range between ca. 610 and 2070 m (2000 and 6800 ft.). The climate is generally cool, with a significant maritime influence. Summers are warm and dry, winters cool and moist. The soils are mantled by a thick layer of Quaternary volcanic ash and colluvium, underlain by Tertiary granitic rocks of the Idaho Batholith and Precambrian gneiss and schists (McGrath et al. 2002). Annual precipitation is ca. 889–2032 mm, with 40–50% falling as snow. Snow packs at higher elevations can become very deep and linger late into spring. Rain-on-snow events are common below 1370 m (4500 ft.); January min/max temperatures:  $-8.3/0.6^{\circ}\text{C}$ ; July min/max temperatures are  $7.2/27.2^{\circ}\text{C}$ .

These mountains have been largely unglaciated and support a highly dissected stream network of ancient narrow valleys and canyons. Drainage density (length of stream/area) is high. The natural vegetation is primarily a very species-rich mosaic of mesic conifer forest, including exceptional conifer species diversity (grand fir, Douglas fir, western red cedar, western hemlock, mountain hemlock, ponderosa pine, lodgepole pine, white pine, western larch, subalpine fir, Englemann spruce and yew). Ponderosa pine/Douglas fir and cedar/hemlock/pine forests occur at lower elevations, and spruce/fir or mountain hemlock forests at higher elevations. Some of the forest types in this area are the best examples of inland temperate rainforest in the Rocky Mountain region (Bergdahl 2008), including western red cedar trees estimated to be more than 3000 years old (Smith and Fischer 1997). The flora of this region has many unique coastal disjunct species and the area is often referred to as the Clearwater Refugium (Brunsfeld et al. 2001, Carstens et al. 2005, Brunsfeld and Sullivan 2006).

Like all other species of *Pseudoferonina*, *P. lolo* appears to be a habitat specialist, primarily on the wet margins of small, low (0–2) order forest streams descending hill-sides in mountainous country. These habitats are often isolated in headwater basins, or on breaks and canyon walls at lower elevations. These beetles are not aquatic, but they are strict hygrophiles. Surface-active adults are usually found within two meters of wet stream channels, primarily within one meter of the strandline and especially right at the water's edge. When disturbed during warm weather, they will often run into the water and crawl down into the gravel or float away.

Other carabid species collected by JCB at or in the vicinity of the Cottonwood Creek locality include: *Scaphinotus (Pseudonomaretus) regularis* (LeConte), *S. (P.) relictus* (Horn), *S. (P.) merckelli* (Horn), *Zacotus matthewsii* LeConte, *Trechus coloradense* Schaeffer, *Bembidion kuprianovi*, *B. iridescens*, *B. breve* (Motschulsky), *Pterostichus (Hypherpes) ecarinatus* Hatch, *P. (Leptoferonia) idahoae* Csiki, *P. (L.) beyeri* Van Dyke, *P. (Pseudoferonina) shulli* (Hatch) and *P. (P.) vexatus* Bousquet.

**Phenology and larval ecology.** Adults of *P. lolo* are probably active from the first warm days of spring after snowmelt to the first days of hard frosts in October. A single pair of adults was observed *in copulo*, on 17 July 2009 [JCB]. *Pterostichus lolo*, and all other *Pseudoferonina* species, are probably “spring breeders” (Bousquet 1986)—that is, they breed early in the active season, larval development takes place in late spring-sum-



**Figure 7.** Photographs of habitat for *Pterostichus* (*Pseudoferonina*) *lolo* Bergdahl, sp. n. Cottonwood Creek, near Bungalow, Clearwater County, Idaho, 3 July 2010. Photograph by J. C. Bergdahl.

mer, new adults emerge from pupal chambers in late summer-early fall, and individuals overwinter primarily as immature adults. Males collected in fall are usually immature, suggesting they may die after a single breeding season. These beetles probably vacate streambeds and floodplains in the fall so as to escape winter or spring freshets of high water. Flash floods from thunderstorms during the active season may be a major cause of mortality. The ecology of the larvae is poorly known because they are rarely seen on the surface. The extent to which larvae and adults of *Pseudoferonina* utilize the subterranean hyporheic zone may be substantial. As mentioned above, adults do not hesitate to crawl down into saturated streambed gravels to escape capture. The highest density of adults is often found along small creeks and reaches that have recently experienced small debris slides with rocks embedded in substantial amounts of wet organic mud. Presumably these muddy microhabitats also support the highest larval density and survival.

**Dispersal power.** All known individuals are brachypterous (flightless) and restricted to the margins of a small, isolated, forested stream, so dispersal power of *P. lolo* adults is expected to be extremely low. Like *P. shulli*, *P. spathifer* and *P. bousqueti* sp. n., the geographic range of *P. lolo* (Fig. 5) appears to be highly restricted.

**Remarks.** Based on the form of the median lobe of the male aedeagus, members of *P. lolo* appear to be most closely related to those of *P. shulli* and *P. spathifer*, (i.e., the *shulli* species-group). It is perhaps significant that both of these species, along with the more distantly related *P. vexatus*, occur in areas near the single known locality for *P. lolo*. The North Fork Clearwater River basin has the highest diversity of *Pseudoferonina* species.

Property ownership in the vicinity of type locality is primarily the U.S. Forest Service (Clearwater National Forest). Potlatch Corporation, Bennett Tree Farms, Inc. and the State of Idaho also have significant property ownership in this region. The primary land use impacts in this area are timber harvesting, roads, mining and off-road vehicle use. There is a large, poorly managed campground on U.S. Forest Service property along both sides of Cottonwood Creek at the type locality, with substantial off-road vehicle impacts.

Because *P. lolo* is known from only one, easily identifiable locality, we hope that collectors will exercise restraint when sampling at this site, and instead focus on discovering new sites for this species in the immediate vicinity.

### **Suggested modifications to Bousquet's key to adults of *Pseudoferonina* Ball**

Bousquet's key for identification of *Pseudoferonina* adults (Bousquet 1985, modified in 1992) works only for males, and is based mainly on characters of the median lobe of the aedeagus. Although a key that permits the identification of females is highly desirable, we do not yet have sufficient knowledge to construct such a key at this time.

Males of both *P. bousqueti* and *P. lolo* key to couplet "3" in Bousquet's 1985 key. In 1992, he expanded this couplet to accommodate the new species, *Pterostichus spathifer* Bousquet by adding a couplet "3'". We suggest the following additional modification to accommodate the two new species described here:

- 3 Median lobe of aedeagus strongly sinuate on ventral margin behind middle in left lateral view..... **3a**
- Median lobe of aedeagus not or very slightly sinuate on ventral margin behind middle in left lateral view..... **3c**
- 3a Median lobe of aedeagus with apical lamella wide, spoon-like, slightly distorted to right in left lateral view (see Bousquet, 1992, Fig. 15) ..... ***P. spathifer* Bousquet**
- Median lobe of aedeagus with apical lamella narrower, not spoon-like (see Bousquet, 1992, Fig. 15 and Figs 4B and 4D here)..... **3b**
- 3b Median lobe with a broad, evenly arcuate convexity on ventral margin at middle in left lateral view (see Bousquet, 1992, Fig. 16) .... ***P. shulli* (Hatch)**
- Median lobe with an abrupt, nearly tuberculate convexity on ventral margin at middle in left lateral view (Figs 4B and 4D) ..... ***P. lolo* Bergdahl sp. n.**
- 3c Specimen larger, ABL greater than 10.5 mm; apical lamella of male median lobe nearly parallel-sided, narrow in left lateral view (see Bousquet, 1985, Fig. 9b) ..... ***P. lanei* Van Dyke**
- Specimen smaller, ABL less than 9.6 mm; apical lamella of male median lobe expanded apically, symmetrically hatchet-shaped in left lateral aspect (Figs 4A and 4C) ..... ***P. bousqueti* Bergdahl sp. n.**

## Discussion

The carabid fauna of the Pacific Northwest (PNW) of North America is highly distinctive, with many endemic and taxonomically isolated taxa, some of which have close affinities (particularly at the generic level) with the southeastern United States or southeast Asia, especially China and Japan (Van Dyke 1940, Kavanaugh 1988). This fauna includes 93 known endemic species and subspecies (Bergdahl 1995). Some of these endemics are restricted to the northern parts of the region, which are assumed to have been fully or largely glaciated in Pleistocene time, as recently as 10,000 years ago (Kavanaugh 1988). There are few, if any, other areas in North America that are known to have been heavily glaciated and yet have such a distinctive fauna with a significant endemic component.

All nine described species in the *Pterostichus* subgenus *Pseudoferonina* are endemic to the PNW. No other group of closely related carabid species includes so many endemics restricted to the region. The large carabid genera *Nebria* Latreille and *Bembidion* Latreille have 13 and 9 species endemic to PNW respectively, but these species represent several subgenera (Bergdahl 1995). Therefore, *Pseudoferonina* may provide opportunities for unique insights into the evolutionary history of the insect fauna of the Pacific Northwest. *Pseudoferonina* includes two species from the Coast Ranges (*P. humidulus* from northwestern Oregon and southwestern Washington, and *P. campbelli* from northwestern Oregon), and two from the Cascade Mountains (*P. smetanai* from southern Washington and *P. campbelli* from northern Oregon). The other six species (*P. lanei*, *P. shulli*, *P. spathifer*, *P. lolo*, *P. bousqueti* and *P. vexatus*) are found in the interior on the west slope

of the Rockies, primarily in the Clearwater, Bitterroot and Salmon River mountains and their foothills in central Idaho. It is clear that the center of diversity of *Pseudoferonina* species is the mountains of central Idaho, especially the Clearwater River basin. This is one of the oldest and most extensive mountain regions in the Pacific Northwest. Given the number of newly described species of *Pseudoferonina* from Idaho and the difficulty of collecting there, it is likely that other species remain to be discovered.

The geographical ranges of all *Pseudoferonina* species remain incompletely known and/or circumscribed. The larvae of all species remain to be recognized and described. Perhaps most importantly, no sister group for *Pseudoferonina* has been identified and phylogenetic relationships among included species have not yet been analyzed. Without a robust hypothesis of relationship among the species, interpretation of the evolution, diversification and history of the group remains impossible. These gaps in our knowledge serve as obstacles to a better understanding of the group and prevent us from answering, at least for the present, interesting questions that should guide future research: What historical and ecological factors are responsible for the diversification of so many localized *Pseudoferonina* species? What is the significance of the disjunct distribution of *Pseudoferonina* in coastal versus inland forest regions? What can an understanding of the phylogenetic and biogeographic history of *Pseudoferonina* tell us about other unique elements of the Pacific Northwest's insect fauna and their conservation? Hopefully, future discoveries and analyses will help us to answer such questions.

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## Appendix I

Table of localities. (doi: 10.3897/zookeys.104.1272.app1) File format: Microsoft Office Excel.

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