RESEARCH ARTICLE



A revision of the purse-web spider genus Calommata Lucas, 1837 (Araneae, Atypidae) in the Afrotropical Region

René Fourie^{1,3,†}, Charles R. Haddad^{1,‡}, Rudy Jocqué^{2,§}

l Department of Zoology & Entomology, University of the Free State, P.O. Box 339, Bloemfontein 9300, South Africa **2** Section of Invertebrates, Royal Museum for Central Africa, B-3080 Tervuren, Belgium **3** Quintiles International, 196 Nelson Mandela Drive, Brandwag, Bloemfontein 9301, South Africa

turn:lsid:zoobank.org:author:FEE7B91B-DEE2-4079-A3CA-66AC2C0902FD
turn:lsid:zoobank.org:author:417ED537-9B99-48BD-B2AB-CC27E762C850
urn:lsid:zoobank.org:author:CF15016C-8CD1-4C9D-9021-44CA7DC7A5D5

Corresponding author: Charles R. Haddad (haddadcr@ufs.ac.za)

Academic editor: Jeremy Miller | Received 19 November 2010 | Accepted 15 April 2011 | Published 4 May 2011

urn:lsid:zoobank.org:pub:3252AEA9-3D2A-42B3-B65E-6072F8E30D35

Citation: Fourie R, Haddad CR, Jocqué R (2011) A revision of the purse-web spider genus *Calommata* Lucas, 1837 (Araneae, Atypidae) in the Afrotropical Region. ZooKeys 95: 1–28. doi: 10.3897/zooKeys.95.745

Abstract

The purse-web spider genus *Calommata* Lucas, 1837 is revised in the Afrotropical Region. Following examination of the female type material, *C. transvaalica* Hewitt, 1916 is removed from synonymy with *C. simoni* Pocock, 1903 and revalidated. The females of both species are redescribed and their males described for the first time. While *C. simoni* is very widespread across tropical Africa, *C. transvaalica* is endemic to northern South Africa. Four new species are described, all known only from males: *C. megae* **sp. n.** (Zimbabwe), *C. meridionalis* **sp. n.** (South Africa), *C. namibica* **sp. n.** (Namibia) and *C. tibialis* **sp. n.** (Ivory Coast and Togo). Notes are presented on the biology of each species.

Keywords

conservation, endemic, Mygalomorpha, new species, revalidation

Introduction

The spider family Atypidae includes a small group of distinctive Mygalomorpha, commonly known as purse-web spiders. Presently it comprises three genera, *Atypus* Latreille, 1804, *Calommata* Lucas, 1837 and *Sphodros* Walckenaer, 1835, representing 43 species in Africa, Europe, Asia and North America (Platnick 2010). Morphologically, the family possesses many plesiomorphic features, yet they also have some derived features in both their morphology and technique of prey capture (Gertsch and Platnick 1980). Atypids are burrowing spiders that line their burrows with silk, with an upper section forming a tough web level with the ground, which is expanded and camouflaged as a trap for wandering arthropods (Jocqué and Dippenaar-Schoeman 2006).

Atypidae represents an ancient branch of the Mygalomorpha, with morphological and molecular phylogenetic analyses placing the family basally in the suborder, sister to the Antrodiaetidae in the clade Atypoidina = Atypoidea (Raven 1985, Goloboff 1993, Hedin and Bond 2006, Ayoub et al. 2007). Unfortunately, previous phylogenies have not included African *Calommata*, and thus their relationships to species from other regions and the biogeographical origins of *Calommata* remain totally unexplored.

The Atypidae, with a residual dorsal abdominal scutum in males, can be characterised by (Raven 1985): 1) the extremely elongate, curved inner portion of the maxillary lobes (Figs 10, 11), 2) the broad and obliquely truncated posterior median spinnerets (Figs 32, 35), 3) the rotated orientation of the maxillae (Figs 12–19), and 4) the teeth on the paired and unpaired claws of males and females that are raised on a common process, giving the appearance of a single multipectinate tooth (Raven 1985). However, the latter character is not applicable to the *Calommata* investigated here, neither in males (Figs 23, 27) nor females. Within the family, the three genera can be separated by palpal, spermathecal and sternal morphology, discussed in the diagnosis below.

While the taxonomy of the Atypidae of North America (Gertsch and Platnick 1980), Europe (Kraus and Baur 1974, Schwendinger 1990) and Asia (Schwendinger 1990, Oliger 1998, Tanikawa 2006, Zhu et al. 2006, Levy 2007) has been quite thoroughly studied, the African fauna has not yet been subjected to a modern revision, the last taxonomic paper of note having been published more than 40 years ago (Benoit 1967). Similarly, considerable information is available on the biology (e.g. Clark 1969, Kraus and Baur 1974, Gertsch and Platnick 1980, Coyle and Shear 1981, Anderson 1987, Edwards and Edwards 1990), microhabitat preferences (Řezáč et al. 2007, Řezáč 2009), dispersal (Coyle 1983, Coyle et al. 1985), population genetics (Pedersen and Loeschcke 2001) and karyology (Řezáč et al. 2006) of *Atypus* and *Sphodros*, while considerably less information is available on *Calommata*, particularly the African species, and relates mainly to habitat, burrow structure and prey capture behaviour (Van Dam and Roberts 1917, Blandin 1971, Charpentier 1995, Sato et al. 2007).

Until now, *Calommata* was thought to be represented in the Afrotropical Region by a single species described from Cameroon, *C. simoni* Pocock, 1903 (Benoit 1967, Gertsch and Platnick 1980, Dippenaar-Schoeman and Jocqué 1997, Dippenaar-Schoeman 2002, Jocqué and Dippenaar-Schoeman 2006). A second species, *C. transvaali*- *cus* Hewitt, 1916, described from Roodeplaat, South Africa, was synonymised with *C. simoni* by Benoit (1967). The remaining six species in the genus, *C. fulvipes* (Lucas, 1835), *C. obesa* Simon, 1886, *C. pichoni* Schenkel, 1963, *C. signata* Karsch, 1879, *C. sundaica* (Doleschall, 1859) and *C. truculenta* (Thorell, 1887), were all described from East Asia, although *C. sundaica* has recently been recorded from Israel (Levy 2007).

In this paper the *Calommata* of the Afrotropical Region are revised, *C. transvaalica* is removed from synonymy with *C. simoni*, and four new species are described. While the biology of *C. simoni* and *C. transvaalica* has been studied (Van Dam and Roberts 1917, Blandin 1971, Charpentier 1995), little is known about the habits of the new species, which is largely due to the general scarcity of these spiders. We suspect that the biological studies related to *C. simoni* do, in fact, refer to *C. tibialis* sp. n., based on available material from one study and contrasting habitat preferences of the two species. By resolving the taxonomy of the Afrotropical fauna, the representative species can be included in future phylogenetic analyses and conservation assessments.

Material and methods

All spiders were studied under an Olympus SZX10 stereomicroscope under 10× magnification. Measurements (body and legs) were taken with a measuring eyepiece and are given in millimetres (mm). The proportional indices used follow Levy (2007) and are the carapace index (length divided by width), and patella-tibia index (combined length of the patella and tibia segments of the first leg divided by the length of the carapace). Total length is given as the length from the front of the chelicerae to the tip of the abdomen, excluding the spinnerets. A range of total length measurements is provided for each species from the material examined. Where geographical locality data was not provided on specimen labels or was not available in collection databases, geographical co-ordinates were searched for in the Global Gazetteer Version 2.2 (http://www.fallingrain.com) or using Google Earth Version 5.0 (http://www.google. com/earth), and are indicated in square parenthesis.

Material of male specimens of *Calommata meridionalis* sp. n., *C. namibica* sp. n., *C. simoni* and *C. tibialis* sp. n. was prepared for Scanning Electron Microscopy (SEM). Specimens were transferred to absolute (100 %) ethanol and left overnight. After drying in hexamethyldisilazane (HMDS), the specimens were glued to rounded aluminium rivets using two-sided copper strips and then coated with gold for examination using a JEOL JSM-6480 scanning electron microscope at the MRAC. Digitised micrographs were taken.

Digital photographs of the dorsal habitus of a *C. simoni* female were taken with a Leica MZ16 and stacked using the LAS automontage software at the MRAC. Digital photographs of the dorsal habitus of the remaining species and a *C. simoni* male were taken using a Nikon Coolpix 8400 mounted on a Nikon SMZ800 stereomicroscope at the UFS. The extended focal range images were stacked using CombineZM software to increase depth of field (www.hadleyweb.pwp.blueyonder.co.uk.).

Abbreviations:

AL	abdomen length
ALS	anterior lateral spinneret(s)
AMNH	American Museum of Natural History (New York, U.S.A.)
AW	abdomen width
BMNH	British Natural History Museum (London, U.K.)
CL	carapace length
CW	carapace width
MRAC	Royal Museum for Central Africa (Tervuren, Belgium)
NCA	National Collection of Arachnida, Plant Protection Research Institute
	(Pretoria, South Africa)
NMBA	National Museum (Bloemfontein, South Africa)
NMSA	Natal Museum (Pietermaritzburg, South Africa)
PLS	posterior lateral spinneret(s)
PMS	posterior median spinneret(s)
UFS	University of the Free State (Bloemfontein, South Africa)
TL	total length
TMSA	Ditsong National Museum of Natural History (Pretoria, South Africa)
USNM	National Museum of Natural History (Washington D.C., U.S.A.)

Taxonomy

Family Atypidae Subfamily Atypinae

Genus Calommata Lucas, 1837

http://species-id.net/wiki/Calommata

Type species. Pachyloscelis fulvipes Lucas, 1835 from Java and Sumatra

Diagnosis. *Calommata* can be distinguished from *Atypus* and *Sphodros* by three main characteristics: 1) Spermathecal structure. In *Atypus*, there are two broad plates each bearing two or more small receptacula (Schwendinger 1990: figs 14–25), in *Sphodros* the four spermathecae are each highly coiled and without distinct receptacula (e.g. Gertsch and Platnick 1980: fig. 29), whereas in *Calommata* there are four spermathecae (Fig. 62), each bearing several closely packed terminal receptacula positioned in a cauliflower-like arrangement,). However, the latter definition was based only on the spermathecal structure of three of the seven *Calommata* species (Gertsch and Platnick 1980), and variation does indeed occur within the spermathecal structure of the genus. The female genitalia of *C. transvaalica* only bear one pair of oval spermathecae (Fig. 69). 2) Male palpal cymbium structure. In *Calommata* the palpal cymbium is short and truncate (Fig. 48), while in *Atypus* and *Sphodros* it is short and acuminate (Gertsch and Platnick 1980: figs 15, 25;

Raven 1985). 3) Labiosternal suture. In *Calommata* the labiosternal suture is positioned anteriorly on the sternum (Levy 2007: fig. 8), but is considered by Gertsch and Platnick (1980) to be absent and by Raven (1985) to have migrated posteriorly in *Atypus* and *Sphodros* (see Gertsch and Platnick 1980: figs 14, 22). Further morphological characters unique to *Calommata* (Gertsch and Platnick 1980) include the bipartite, longitudinal thoracic groove (fovea) (Figs 1–9); basal ledge on the outer surface of the fangs of both sexes, the posteriorly positioned ocular tubercle, enormously elongated endites and dorsally expanded chelicerae (Figs 10, 11); short leg I, particularly the femur, and flattened palpal tibia and tarsus of females (Figs 4, 7); and the greatly elongate tarsi of legs III and IV of males, which are clearly pseudosegmented (e.g. Figs 3, 5, 30).

Description. Medium to large sexually dimorphic spiders (Figs 1–9), females 18.60– 33.40 in length and males 5.10–9.35 in length. Carapace with an anterior, strongly elevated median ocular tubercle (Figs 10, 11), with a flattened posterior part traversed by a longitudinal thoracic furrow with a small deep pit in the middle (Figs 1–8). Three faint lines run from the ocular tubercle, converging at the fovea. Chelicerae massive, dorsally expanded with flattened sides, bearing sharp teeth usually in one (rarely two) rows, and a long arched fang with a slit-like distal opening and distinctive basal ledge along its outer margin (Figs 10–19, 20, 21, 26, 29). Cheliceral dentition of both sexes



Figures 1–8. Dorsal habitus of Afrotropical *Calommata* species 1 *C. megae* sp. n., male (Harare, Zimbabwe) 2 *C. meridionalis* sp. n., male (Erfenis Dam, South Africa) 3 *C. namibica* sp. n., male (Etosha, Namibia) 4 *C. simoni* Pocock, female (Galim, Cameroon) 5 same, male (Dja Reserve, Cameroon) 6 *C. tibialis* sp. n., male (Bassari–Sokodé, Togo) 7 *C. transvaalica* Hewitt, female (Blouberg, South Africa) 8 same, male (Groenkloof, South Africa). Scale bars: 5mm.



Figures 9–19. Somatic morphology of *C. tibialis* sp. n., male 9, 17 *C. simoni* Pocock, female 10, 15 and male 11, 16 *C. megae* sp. n., male 12 *C. meridionalis* sp. n., male 13 *C. namibica* sp. n., male 14 and *C. transvaalica* Hewitt, female 18 and male 19: 9 dorsal habitus 10, 11 lateral view of chelicera, endites and anterior of carapace 12–19 left chelicera, ventral view, indicating dentition. Scale bars: 5mm (9), 1mm (10–19).



Figures 20–31. Scanning electron micrographs of *Calommata meridionalis* sp. n. 20–25 *C. simoni* Pocock 26–28 and *C. tibialis* sp. n. 29–31 males 20, 29 chelicera in ventral view 21 tip of fang 22 right endite, ventral view 23, 27 tarsus and claw, leg I (note pseudosegmentation of tarsus) 24, 30 tarsus IV, lateral and ventral view (note pseudosegmentation of tarsus) 25, 28, 31 detail of ventral scopulate setae on tarsus IV 26 chelicera in prolateral view.

often variable between specimens and even between opposing chelicerae of a specimen. The general dentition pattern of each species and sex is indicated in Figs 12–19. Chelicerae of females have larger, more numerous teeth and an extensive field of tiny denticles retrolateral of the teeth row near the cheliceral base (Figs 15, 18). Chilum single, large, varying in shape, oval, rectangular or pentagonal (Figs 1–8). Endites strongly elongated and curved upwards on the prolateral side (Figs 10, 11, 22). Sternum with a distinctive labio-sternal suture anteriorly. Legs of females short and stout, leg formula of females 4231. Legs of males of normal size, with the tarsi pseudosegmented (Figs 23, 24, 30) and tarsi of legs III and IV distinctly longer (Figs 1–3, 5, 6, 8, 9); leg formula of males



Figures 32–40. Scanning electron micrographs of *Calommata simoni* Pocock 32–38 and *C. meridionalis* sp. n. 39, 40 male spinnerets 32 spinneret field, ventral view 33 ALS 34 detail of single ALS spigot 35 PMS 36, 39 detail of PMS spigots 37, 40 PLS 38 detail of PLS spigots.

usually 4321, rarely 4132. Tarsi with three claws, paired claws of males with single row of teeth and unpaired claw without teeth (Figs 23, 27), contra Raven (1985: 123). Tarsi ventrally scopulate, consisting of pointed setae (Figs 25, 31) or setae with a rounded tip (Fig. 28). Abdomen of male with small, distinctive anterior dorsal scutum, absent in females (Figs 1-8). Venter with three pairs of spinnerets, examined in detail in males (Figs 32-40) but not females; ALS small and finger-like, single-segmented, with a single short, stout distal fused spigot (Figs 32-34); PMS small and subtriangular, single-segmented, with several fused and articulated spigots in distal half of segment (Figs 32, 35, 36, 39); PLS large, elongate, three segmented, distal segment digitiform, with several fused and articulated spigots in distal half of second and entire ventral surface of third segment (Figs 32, 37, 38, 40). Anal tubercle widely separated from the spinnerets. Female palp short, with flattened tibia and tarsus (Fig. 7). Female epigyne forming broad, weakly sclerotised plate in ventral view; epigyne with two pairs of spermathecae, median pair subrectangular and rounded anteriorly, and lateral pair subtriangular (Fig. 62), or with single pair of transversely oval spermathecae (Fig. 69). Male palp with swollen tibia, bearing a retrolateral ventral row of several trichobothria, the bases of which are raised to one side; palpal tegulum with curved broad conductor with tooth on its dorsal surface, embolus straight, tapering towards tip (Figs 41–52).

Key to the Afrotropical Calommata

[females of *C. megae* sp. n., *C. meridionalis* sp. n., *C. namibica* sp. n. and *C. tibialis* sp. n. unknown]

1	Females2
_	Males
2	Chelicerae with a single row of teeth along promargin running from fang base close to cheliceral base (Fig. 15); epigyne with two pairs of spermathecae (Fig. 62) (tropical Africa)
_	Chelicerae with an additional row of 2–4 teeth along promargin placed close to fang base, prolateral of main teeth row (Fig. 18); epigyne with one pair of transversely oval spermathecae (Fig. 69) (South Africa) <i>C. transvaalica</i>
3	CL subequal to CW
_	Carapace clearly longer than wide, CL>1.15CW
4	Embolus and conductor in ventral view directed obliquely and retrolater- ally towards endites (Fig. 54); palpal tibia clearly longer than wide (Fig. 53), patella-tibia index ~1.3 (Zimbabwe)
_	Embolus and conductor in ventral view directed obliquely and retrolaterally to- wards distal end of chelicerae (Fig. 67); palpal tibia swollen, only slightly longer than wide (Fig. 66), patella-tibia index ~0.9 (Côte d'Ivoire and Togo)
F	
5	Embolus and conductor in ventral view directed transversely across palpal axis
_	Embolus and conductor in ventral view directed obliquely relative to palpal axis
6	Chelicerae with one or two very large teeth near fang base, remaining teeth distinctly smaller; several small denticles retrolateral of teeth row near base of chelicerae (Fig. 13); cymbium tip in ventral view broad and rounded (Fig. 57) (South Africa)
_	Chelicerae without one or two large teeth near fang base, teeth gradually de- creasing in size from fang base to cheliceral base; denticles absent near base of chelicerae (Fig. 19); cymbium tip in ventral view tapering to rounded point (Fig. 71) (South Africa)
7	Cheliceral teeth variable in size, with at least three large teeth in midsection of teeth row and several denticles retrolateral of teeth row (Fig. 16); conductor and embolus not projecting beyond cymbial margin in ventral view (Fig. 64); conductor with distinct tooth and second ridge resembling a tooth distally on its dorsal surface (Fig. 49) (tropical Africa)
_	Cheliceral teeth tiny, subequal, without denticles retrolateral of teeth row (Fig. 14); conductor and embolus projecting far beyond cymbial margin in ventral view (Fig. 60); conductor with single tooth distally on its dorsal surface (Fig. 46) (Namibia)



Figures 41–52. Scanning electron micrographs of *Calommata meridionalis* sp. n. 41–43, *C. namibica* 44–46, *C. simoni* Pocock 47–49 and *C. tibialis* sp. n. 50–52 males 41, 44, 47, 50 trichobothrial bases on palpal tarsus 42, 43, 46, 49, 52 distal end of conductor and embolus 45 palpal organ in prolateral ventral view 48, 51 palpal organ in prolateral view.

Calommata megae Fourie, Haddad & Jocqué, sp. n.

urn:lsid:zoobank.org:act:ED324676-F4FB-4F0D-816E-4670E7EE2577 http://species-id.net/wiki/Calommata_megae Figs 1, 12, 53–55

Type material. Holotype male. ZIMBABWE: Harare, Highlands [17°48'S, 31°05'E], 13.I.2003, on soil, by hand, M. Cumming (NCA 2004/362).

Other material examined. ZIMBABWE: 1imm.: same locality as holotype, 21.II.2000, in garden, M. Cumming (NCA 2004/1361).



Figures 53–55. Left palp of male *Calommata megae* sp. n. 53 prolateral view 54 ventral view 55 retrolateral view. Scale bars: 1mm.

Diagnosis. The male of this species is recognised by the conductor that narrows and makes a half twist before broadening distally, and the obliquely orientated embolus and conductor (Figs 53–55). This species shares with *C. tibialis* sp. n. the carapace that is subequal in length and width (longer than wide in the other four species).

Etymology. The specific epithet is a patronym in honour of the collector of the holotype, Meg Cumming, in recognition of her contributions to African arachnology, particularly in Zimbabwe.

Description. Male holotype. Measurements: CL 2.00, CW 2.05, AL 2.95, AW 1.80, TL 7.00. Length of leg segments, and total: I 2.40 + 0.90 + 1.70 + 1.90 + 1.70 = 8.60; II 2.15 + 0.95 + 1.40 + 2.20 + 2.15 = 8.85; III 1.80 + 0.95 + 1.00 + 2.15 + 3.55 = 9.45; 2.35 + 1.10 + 1.28 + 2.50 + 4.35 = 11.58. Carapace index 0.98; patella-tibia index 1.30.

Carapace and chelicerae brown in colour (Fig. 1). Carapace flat and robust. Median ocular tubercle raised, darker in colour. Chelicerae with a single row of small teeth, increasing in size from fang base to base of chelicerae, with a few denticles near base of chelicerae (Fig. 12). Sternum and coxae light brown, remainder of legs dark brown, fading to light yellow-brown at tarsi. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules (thicker and shorter than bristles). Abdomen dark brown, with an irregular brown scutum present anteriorly (Fig. 1). Palpal cymbium short with rounded distal margin; embolus and conductor orientated obliquely, pointing retrolaterally towards base of chelicerae; conductor narrow, making a half twist before broadening distally, with single small curved tooth on its dorsal surface distally; embolus short and straight, slightly curved near tip (Figs 53–55).

Female. Unknown.

Distribution. Known only from the type locality (Fig. 73).

Biology. Poorly known. The holotype male was collected in mid-summer on the soil surface. The second instar juvenile specimen was captured while ballooning and landing on the porch of a house (M. Cumming, pers. comm.). Amongst atypids, ballooning has previously been recorded in both *Atypus* and *Sphodros* (Wiehle 1953 cited in Pedersen and Loeschcke 2001, Coyle 1983, Coyle et al. 1985).

Calommata meridionalis Fourie, Haddad & Jocqué, sp. n.

urn:lsid:zoobank.org:act:7E2F6F05-6995-4C3B-AC59-30A0CE451914 http://species-id.net/wiki/Calommata_meridionalis Figs 2, 13, 20–25, 39–43, 56–58

Type material. Holotype male. SOUTH AFRICA: *Free State Province*: Erfenis Dam Nature Reserve, 28°29.888'S, 26°48.488'E, 21.IX–22.X.2005, pitfalls, unburned site 1, C. Haddad, S. Otto & R. Poller (NCA 2009/3488).

Paratypes. SOUTH AFRICA: *Free State Province*: 33° : Bloemfontein, National Botanical Gardens, 29°03.006'S, 26°12.701'E, 27.X–16.XI.2009, pitfalls, grassland, C. Haddad (NMBA 13981); 73° : Same data, 16–21.XI.2009 (NMSA 22616); 13° : Same locality, 21.XI–7.XII.2009, pitfalls, grassland, C. Haddad & R. Fourie (MRAC 229029); 13° : Erfenis Dam Nature Reserve, 28°30.373'S, 26°48.437'E, 21.IX–22.X.2005, pitfalls, burned site 1, C. Haddad, S. Otto & R. Poller (NCA 2009/3663); 43° : Same locality, 28°30.134'S, 26°48.427'E, 22.X–22.XI.2005, pitfalls, burned site 2, C. Haddad, S. Otto & R. Poller (NCA 2007/3142); 13° : Same locality, 28°29.741'S, 26°48.065'E, 21.IX–22.X.2005, pitfalls, unburned site 3, C. Haddad, S. Otto & R. Poller (NCA 2009/3664); 13° : Oranjeville district, Vaal Dam, 26°59.523'S, 28°15.737'E, 1–29.X.2009, pitfalls, grassland, R. Fourie & A. Grobler (NCA 2009/3539).

Other material examined. SOUTH AFRICA: *Free State Province*: 1³: Erfenis Dam Nature Reserve, 28°30.134'S, 26°48.427'E, 21.IX–22.X.2005, pitfalls, burned site 2, C. Haddad, S. Otto & R. Poller (MRAC, prepared for SEM).

Diagnosis. The male of this species can be easily recognised from African congeners by the presence of one or two very large teeth at the fang base (Fig. 13), and the transversely orientated curved embolus with distally broadened conductor bearing a single tooth on its dorsal surface (Figs 42, 43, 57). The raised median ocular tubercle is broader than in the other species.

Etymology. This specific epithet is Latin for southern, referring to the distribution of the species, southernmost in the genus.

Description. Male holotype. Measurements: CL 1.60, CW 1.25, AL 3.15, AW 1.68, TL 6.35 (5.60–7.80). Length of leg segments, and total: I 1.40 + 0.55 + 1.00 + 1.15 + 1.25 = 5.35; II 1.30 + 0.55 + 0.85 + 1.24 + 1.45 = 5.39; III 1.05 + 0.64 + 0.64 + 1.20 + 2.40 = 5.93; IV 1.45 + 0.64 + 0.75 + 1.44 + 2.90 = 7.18. Carapace index 1.28; patella-tibia index 0.97.



Figures 56–58. Left palp of male *Calommata meridionalis* sp. n. 56 prolateral view 57 ventral view 58 retrolateral view. Scale bars: 1mm.

Carapace and chelicerae dark brown in colour (Fig. 2). Median ocular tubercle raised, darker in colour; median ocular tubercle broader than in other species. Chelicerae with single prolateral row of teeth, one very large tooth close to fang base, sometimes accompanied by second large tooth, remaining teeth distinctly smaller and subequal in size, with several denticles retrolateral of teeth row close to cheliceral base (Fig. 13). Sternum and coxae pale brown, remainder of legs brown, gradually fading to yellow at tarsi. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules. Abdomen grey-brown, with a round brown scutum anteriorly (Fig. 2). Palpal cymbium short with rounded distal margin; embolus and conductor orientated transversely across palpal axis, pointing retrolaterally; conductor broadened distally, with a single small tooth on its dorsal surface distally; embolus long, slightly curved in a S-form along its length (Figs 42, 43, 56–58).

Female. Unknown.

Distribution. Endemic to central and northern Free State Province, South Africa (Fig. 73).

Biology. The species was collected exclusively by pitfalls in spring and early summer (September to early December) in the Grassland Biome of South Africa. Specimens were only collected in dark vertic clay and loamy-clay soils and not from sites with sandy soils. Most of the specimens were collected from sites near to freshwater streams and dams. Despite exhaustive attempts to locate burrows in the vicinity of pitfall sites (Erfenis Dam Nature Reserve and Botanical Gardens) none could be found.

Calommata namibica Fourie, Haddad & Jocqué, sp. n. urn:lsid:zoobank.org:act:099D7F5A-BBF3-482E-B570-3D8053C88E93 http://species-id.net/wiki/Calommata_namibica Figs 3, 14, 44–46, 59–61

Type material. Holotype male and paratype male. NAMIBIA: Etosha National Park, Beisebvlakte, 18°32'S, 17°02'E, 10–14.XI.1996, A. Russell-Smith (MRAC 215409).

Other material examined. None.

Diagnosis. The male of this species can be recognised by the tiny cheliceral teeth in a single row, without retrolateral denticles (Fig. 14), and the obliquely orientated conductor and embolus projecting far beyond the retrolateral cymbial margin (Fig. 60).

Etymology. The specific epithet refers to the country of the type locality.

Description. Male holotype. Measurements: CL 1.80, CW 1.24, AL 2.75, AW 1.54, TL 6.30 (5.10–6.30). Length of leg segments, and total: I 1.21 + 0.50 + 0.83 + 1.09 + 1.35 = 4.98; II 1.40 + 0.51 + 0.62 + 1.13 + 1.40 = 5.06; III 1.05 + 0.49 + 0.47 + 1.16 + 1.95 = 5.12; IV 1.44 + 0.60 + 0.65 + 1.38 + 2.40 = 6.47. Carapace index 1.45; patella-tibia index 0.74.

Carapace and chelicerae dark brown (Fig. 3). Median ocular tubercle raised, narrow, darker in colour. Chelicerae with single prolateral row of tiny teeth, without denticles near cheliceral base (Fig. 14). Sternum and coxae light brown, femora dark brown; subsequent segments fading to light yellow at tarsi. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules. Abdomen dark grey, nearly black, with brown scutum present in the anterior half (Fig. 3). Palp with elongate cymbium, with tapering pointed distal margin; embolus and conductor orientated obliquely, pointing retrolaterally and distally, projecting far beyond retrolateral cymbial margin; conductor short, slightly broadened distally, with a very prominent, very long and slender tooth distally on its dorsal surface; embolus long, with slight bend in distal half (Figs 45, 46, 59–61).

Female. Unknown.

Distribution. Known only from the type locality (Fig. 73).

Biology. Poorly known. This species was collected in late spring in arid savanna.

Calommata simoni Pocock, 1903

http://species-id.net/wiki/Calommata_simoni Figs 4, 5, 10, 11, 15, 16, 26–28, 32–38, 47–49, 62–65

Calommata simoni Pocock, 1903: 259; Benoit, 1967: 283, figs 1–4; Kraus 1978: 245, fig. 13; Dippenaar-Schoeman and Jocqué 1997: 52, figs 40a–h; Dippenaar-Schoeman 2002: 23, figs 9, 10a–g; Jocqué and Dippenaar-Schoeman 2006: 82, figs 18a–h.

Type material. Lectotype female. CAMEROON: Efulen [02°46'N, 10°43'E], G.L. Bates (BMHN, examined).



Figures 59–61. Left palp of male *Calommata namibica* sp. n. **59** prolateral view **60** ventral view **61** retrolateral view. Scale bars: 1mm.

Other material examined. CAMEROON: 1 \bigcirc : Galim [07°06'N, 12°28'E], 15.VIII–19.VIII.1971, F. Puylaert (MRAC 143671); 3 \bigcirc : N of Dja Reserve, 03°41'N, 13°14'E, 8.III.2005, pitfalls, old secondary forest, I. Deblauwe (MRAC 220674); 1 \bigcirc : Same locality, 8.III.2005 (MRAC 219754); 6 \bigcirc : Same locality, 8.III.2005, pitfalls, riverine forest, I. Deblauwe (MRAC 220663); 1 \bigcirc : Same data, 6.V.2005 (MRAC 219754); 6 \bigcirc : Same locality, 8.III.2005, pitfalls, young secondary forest, I. Deblauwe (MRAC 220659). CONGO D.R.: 1 \bigcirc : Kisangani, Masako Forest, 00°35'N, 25°11'E, 25.II.2003, J.-L. Juakaly (MRAC 216031); 1 \bigcirc : Same locality, 11.III.2003, pitfalls, primary forest, J.-L. Juakaly (MRAC 214347); 1 \bigcirc : Same data, 11.III.2003 (MRAC 214354); 1 \bigcirc : Same data, 11.III.2003 (MRAC 214385); 1 \bigcirc : Same data, 11.III.2003 (



Figures 62–65. *Calommata simoni* Pocock female genitalia **62** and left palp of male **63–65: 62** dorsal view **63** prolateral view **64** ventral view **65** retrolateral view. Scale bars: 1mm.

Appouesso, Bossematié Forest, $06^{\circ}35'N$, $03^{\circ}28'W$, 19.IX.1994, pitfalls, rain forest, R. Jocqué & N. Séabé (MRAC 202481); 13° : Same locality, 12.III.1995, pitfalls, forest, R. Jocqué & Tanoh (MRAC 205382); 13° : Same data, 26.III.1995 (MRAC 205383); 13° : Mankono, Ranch de la Marahoué, $08^{\circ}27'N$, $06^{\circ}52'W$, III.1980, riverine forest, J. Everts (MRAC 172117); 13° : Same data (MRAC 172118); 13° : Same data (MRAC 172119); 23° : Same data (MRAC 172120). GUINÉE: 33° : F.C. de Ziama, $08^{\circ}24'N$, $09^{\circ}17'W$, 22.IV.1998, pitfalls, rain forest, D. Flomo (MRAC 216239); 23° : Same data, 5.V.1998 (MRAC 216248); 23° : Same data, 18.V.1998 (MRAC 216249); 13° : Same data, 14.VI.1998 (MRAC 216247); 13° : Same data, 14.VI.1998 (MRAC 216247); 13° : Same data, 31.III.1999 (MRAC 216245); 23° : Same data, 26.IV.1999 (MRAC 216243); 13° : Same data, 26.IV.1999 (MRAC 216244);

13: Same data, 9.V.1999 (MRAC 216242); 23: Same data, 22.V.1999 (MRAC 216240); 13: Same data, 17.VI.1999 (MRAC 216241); 13: Same data, 31.III.2000 (MRAC 216246). KENYA: 13: Kakamega Forest, 00°13'N, 34°54'E, 24.I.2002, pitfalls, D. Shilabira Smith (MRAC 228141); 13: Same data, 13.IV.2002 (MRAC 220536). LIBERIA: 1imm.: Mount Coffee, Bensonville [06°29'N, 10°38'W], II.1894, constructs a tube-like nest under a log, collector unknown (USNM). MALAWI: 13: Chisasira Forest, 25km S of Chintheche, 11°50'S, 33°13'E, 1.XII.1977, pitfalls, *Brachystegia* woodland, R. Jocqué (MRAC 169498); 13: Same data, 1.XII.1977 (MRAC 169499). TANZANIA: 1imm.: Bunduki, Uluguru Mountains, 07°02'S, 37°38'E, 2.V.1957, nest, forest ground in litter, P. Basilewsky & N. Leleup (MRAC 111792).

Diagnosis. The female of this species has the cheliceral teeth in a single row (Fig. 15), while an additional row is found in *C. transvaalica* (Fig. 18). The female genitalia have two pairs of small spermathecae (Fig. 62), while *C. transvaalica* only has a single pair of large transverse spermathecae (Fig. 69). The male of this species is recognised by the conductor ending broadly with a prominent tooth and sharp edge, appearing to be a second tooth, on its dorsal surface (Fig. 49). In *C. simoni* females the patella-tibia index is double that of *C. transvaalica*.

Redescription. Female (measurements provided for female lectotype from Efulen, colouration for female from Galim). Measurements: CL 7.70, CW 6.10, AL 13.10, AW 8.85, TL 27.80 (25.80–33.40). Length of leg segments, and total: I 3.95 + 1.62 + 1.75 + 2.10 + 1.38 = 10.80; 3.70 + 2.25 + 1.70 + 2.23 + 1.60 = 11.48; III 3.65 + 2.75 + 1.55 + 1.78 + 1.39 = 11.12; IV 3.90 + 2.90 + 2.10 + 2.15 + 1.50 = 12.55. Carapace index 1.26; patella-tibia index 0.44.

Robustly built with short legs, carapace faded to creamy brown (Fig. 4). Median ocular tubercle raised, narrow, sloping sharply at fovea (Fig. 10). Single median line running from anterior of eye area to approximately middle of chilum. Chelicerae pale orange brown, darker laterally; chelicerae with a single row of small and medium sized teeth along promargin running from fang base close to cheliceral base, with extensive denticle field retrolateral of teeth row near cheliceral base (Fig. 15). Endites strongly elongated prolaterally, strongly curved upwards (Fig. 10). Sternum and legs light yellowish brown. Legs short and stout, leg formula 4231; legs III and IV more robust than legs I and II; leg I without bristles or spinules; leg II with few spinules distally on tibiae, and dorsal and lateral spinules on metatarsi and tarsi; legs III and IV with spinules from patellae to tarsi (mainly dorsal and prolateral) and covered in bristles. Abdomen globose and pale grey, with indistinct median heart marking in anterior half (Fig. 4). Epigyne forming a broad, weakly sclerotised plate ventrally, in dorsal view with two pairs of spermathecae; median pair subrectangular, rounded anteriorly, lateral pair subtriangular (Fig. 62). Female palp short, tibiae and tarsi flattened.

Male from Cameroon. Measurements: CL 2.20, CW 1.90, AL 3.80, AW 2.32, TL 8.20 (5.60–9.35). Length of leg segments, and total: I 2.30 + 0.75 + 1.64 + 1.91 + 1.55 = 8.15; II 2.25 + 0.86 + 1.45 + 2.16 + 1.88 = 8.60; III 1.84 + 0.90 + 0.98 + 2.23 + 2.95 = 8.90; IV 2.50 + 1.05 + 1.43 + 2.60 + 3.90 = 11.48. Carapace index 1.16; patella-tibia index 1.09.

Carapace and chelicerae brown (Fig. 5). Median ocular tubercle raised, narrow, darker in colour (Fig. 11). Chelicerae with single prolateral row of teeth, largest teeth in midsection of teeth row interspersed with smaller teeth anteriorly and posteriorly, with several denticles retrolateral of teeth row close to cheliceral base (Fig. 16). Endites elongated prolaterally, curving upwards (Fig. 11). Sternum and coxae light yellowish brown, rest of leg segments brown, fading to light yellow at tarsi. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules. Abdomen grey brown, with elongate brown scutum present in anterior half (Fig. 5). Palp with short cymbium, with rounded distal margin; embolus and conductor orientated obliquely, pointing retrolaterally and distally, not projecting beyond retrolateral cymbial margin; conductor short, broadened distally, with a prominent elongate tooth and sharp edge opposite the tooth, appearing as a second tooth; embolus short and straight (Figs 48, 49, 63–65).

Remarks. Benoit's (1967: 286, figs 1–4) drawings of a male "allotype" of *C. simoni* correspond with the males we have studied. However, Pocock (1903: 259) never described the male of *C. simoni* nor listed any males in his material studied, and thus the specimen examined by Benoit could not possibly be an allotype. The loan request to BMNH also only yielded the female lectotype of *C. simoni*, and no allotype or paratypes as indicated by Benoit. Benoit indeed wrongly considered the specimen used to describe the unknown sex for the first time to be the allotype, even when that occurred separately from the original description. Comments on the revalidation of *C. transvaalica* are provided under remarks for that species below.

Charpentier (1995) studied the biology of *C. simoni* in Benin, but no specimens collected by him could be traced in any collection. He collected specimens at four localities: Ayou [06°43'N, 02°07'E], Ahota [06°39'N, 02°09'E], Sè [06°28'N, 01°49'E] and Toffo [06°50'N, 02°04'E].

Distribution. Widespread across tropical Africa in forests and savanna woodlands (Fig. 73).

Biology. The biology of "*C. simoni*" was studied by Blandin (1971) and Charpentier (1995) in Côte d'Ivoire and Benin, respectively (localities listed above). However, examination of the specimens reported on by Blandin indicates that they are, in fact, *C. tibialis* sp. n.. In considering the habitats of the available material of *C. simoni* and *C. tibialis* sp. n., it is evident that the two species are ecologically separated, the former occurring in forests and the latter in woodland savannah. As the material collected by Charpentier (1995) could not be traced, it is impossible to determine whether he studied the biology of *C. simoni* or *C. tibialis* sp. n.. However, his indication of the habitat types at the four localities he sampled (grassland, patches of subsistence agriculture, near rivers and open ground near palm forests) suggests that the material he studied is *C. tibialis* sp. n. and not *C. simoni*. Thus, we have included biological information from their two studies under *C. tibialis* sp. n..

Most of the specimens studied here from the MRAC collected in Guinée, Côte d'Ivoire, Kenya, Tanzania and Congo D.R. were collected in contrasting forest types across tropical Africa, indicating that *C. simoni* is tolerant and adaptable to a wide range of soil, vegetation and climatic variables.

Calommata tibialis Fourie, Haddad & Jocqué, sp. n.

urn:lsid:zoobank.org:act:E31586AD-B428-49F1-8F43-100356C90DCA http://species-id.net/wiki/Calommata_tibialis Figs 6, 9, 17, 29–31, 50–52, 66–68

Type material. Holotype male. TOGO: Between Bassari and Sokodé, 09°15'N, 00°47'E, V–VII.1984, pitfalls, wooded savanna, P. Douben (MRAC 169501).

Paratypes. 1⁽²⁾: together with holotype. CÔTE D'IVOIRE: 1⁽²⁾: Kossou, 06°57'N, 04°58'W, 28.IV.1975, pitfalls, wooded savanna, R. Jocqué (MRAC 169500).

Other material examined. CÔTE D'IVOIRE: 1 subadult \mathcal{Q} : Lamto, 06°12'N, 05°20'W, 6.III.1968, savanna with *Borassus aethiopum*, C. Girard (MRAC 232547); 1 \mathcal{O} : Same locality, 11.II.1974, dirt road near biological station, P. Blandin (MRAC 232548).

Diagnosis. The male of the species can be recognised by the carapace that is subequal in length and width (Fig. 6), the short, swollen palpal tibia, and the narrow conductor ending in a thick prominent tooth (Figs 51, 52).

Etymology. The specific epithet refers to the palpal tibia of the male, which is distinctly shorter and more swollen compared to that of other African congeners.

Description. Male holotype. Measurements: CL 2.13, CW 1.95, AL 3.45, AW 2.02, TL 6.65 (6.00–6.65). Length of leg segments, and total: I 1.98 + 0.65 + 1.35 + 1.71 + 1.63 = 7.32; II 1.80 + 0.75 + 1.10 + 1.60 + 1.80 = 7.05; III 1.49 + 0.80 + 0.75 + 1.70 + 2.45 = 7.19; IV 1.94 + 0.90 + 1.03 + 2.00 + 3.03 = 8.90. Carapace index 1.10; patella-tibia index 0.94.



Figures 66–68. Left palp of male *Calommata tibialis* sp. n. **66** prolateral view **67** ventral view **68** retrolateral view. Scale bars: 1mm.

Carapace and chelicerae orange brown (Fig. 6). Chelicerae with single row of alternating small and medium sized teeth, gradually decreasing in size from fang base to cheliceral base, with several denticles retrolateral of teeth row near cheliceral base (Fig. 17). Eye area raised, narrow, darker in colour. Sternum and coxae yellow, remainder of legs orange, fading to pale yellow at tarsi. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules. Abdomen grey, with pale orange-brown scutum anteriorly (Fig. 6). Palp with short cymbium, with rounded distal margin; tibia shorter and broader than in the other five species, slightly longer than wide; embolus and conductor orientated obliquely, pointing retrolaterally and distally, not projecting beyond retrolateral cymbial margin; conductor narrow with a thick prominent tooth distally on its dorsal surface; embolus short and slightly curved (Figs 51, 52, 66–68).

Female. Unknown.

Remark. The specimens may possibly have faded over time in 70% ethanol, which can only be confirmed should fresh material become available. Although a subadult female is available it will not be described as the genitalic structure cannot be studied.

Distribution. Central Côte d'Ivoire and northern Togo (Fig. 73).

Biology. Present data indicates that *C. tibialis* sp. n. occurs in woodland savannah habitats and avoids forests, where *C. simoni* has been collected. This may indicate some degree of ecological separation between the species but requires further study. Taking this into account we consider the studies of Blandin (1971) and Charpentier (1995) to relate to *C. tibialis* sp. n. and not *C. simoni*, as indicated by them.

Charpentier (1995) located more than 50 nests of *C. tibialis* sp. n. at four localities in southern Benin with quite contrasting habitat structures, including grassland, patches of subsistence agriculture, in close proximity to rivers, and open ground near palm forests. He did not indicate the occurrence of the species in forests. In one of the habitats that he found *Calommata*, the soil was described as sandy, of poor quality and relatively acidic, and covered in 'grassland' vegetation, similar to the habitat characteristics described by Blandin (1971) for the Lamto area, from where *C. tibialis* sp. n. specimens are available.

The burrow of *C. tibialis* sp. n. slants obliquely downwards into the soil, and was estimated to be 25-30cm deep by Blandin (1971), while Charpentier (1995) indicated a maximum depth of 21cm in a female specimen, although generally shallower in other specimens (12–19cm). The top 1–2cm of the burrow is expanded to form a chamber covered by silk webbing that is camouflaged with soil, and the spider lies in wait hanging upside-down from the web for wandering prey (Charpentier 1995). Egg sacs are suspected to hatch in May; during incubation the female spins a silk veil at the base of the chamber that is suspected to firstly allow the spider access to the chamber to capture potential prey, and secondly hide the spider and its eggs from potential parasites once they have entered the chamber (Charpentier 1995).

Calommata transvaalica Hewitt, 1916

species-id.net/wiki/Calommata_transvaalica Figs 7, 8, 18, 19, 69–72

C. transvaalicus Hewitt, 1916: 180, fig. 3, pl. 26, fig. 11 revalidated *C. simoni* Benoit, 1967: 283 synonymy rejected

Type material. Female holotype. SOUTH AFRICA: *Gauteng Province*: Pretoria, Roodeplaat [25°38'S, 28°21'E], 3.IV.1915, G. van Dam (TMSA 2999 – examined).

Other material examined. SOUTH AFRICA: *Gauteng Province*: 13: Groenkloof Nature Reserve [25°47'S, 28°12'E], 21.I.2003, reptile trap, M. Forsythe (NCA 2004/750); 12: Pretoria district, Hatfield [25°45'S, 28°15'E], 25.IV.1915, G. van Dam (TMSA 4639); 133: Zwartkoppies Farm 364, Portion 2, ca. 21km E of Pretoria, 25°45'23.6"S, 28°24'47.7"E, 1347m a.s.l., 29.X.2010, open pitfall traps, I. Engelbrecht & GDARD Field Staff (TMSA 23875). *Limpopo Province*: 13: Blouberg Nature Reserve, 23°00.065'S, 29°03.855'E, 29.XI.2005, searching below the knee, *Philenoptera violaceae*, A. Dawood (NCA 2009/3665); 12: Soutpansberg district, Blouberg, Wilhan's Hohe [not traced], 28.VIII.1923, G. van Dam (TMSA 2772); 12: Same data, 29.VIII.1923 (TMSA 2773).

Diagnosis. The female of this species has an additional row of two to four large prolateral teeth close to the fang base in addition to the main row of teeth, which are larger and more strongly curved than in *C. simoni*. The epigyne comprises a single pair of transversely oval spermathecae, while *C. simoni* possesses two pairs of smaller spermathecae. The male of the species shares with *C. meridionalis* the transversely orientated embolus and conductor, but the conductor of *C. transvaalica* is clearly narrower at the tip and the embolus is straight and not slightly curved as in *C. meridionalis*. The male chelicerae of *C. transvaalica* also lack the one or two large teeth found near the fang base in *C. meridionalis*.

Redescription. Female from Blouberg Nature Reserve. Measurements: CL 6.72, CW 5.80, AL 13.70, AW 10.50, TL 25.20 (18.60–27.00). Length of leg segments, and total: I 1.85 + 0.60 + 0.74 + 0.85 + 0.60 = 4.64; II 1.60 + 1.05 + 0.69 + 0.90 + 0.72 = 4.96; III 1.45 + 1.28 + 0.60 + 0.80 + 0.55 = 4.68; IV 1.58 + 1.41 + 0.90 + 0.90 + 0.47 = 5.26. Carapace index 1.16; patella-tibia index 0.20.

Robustly built with short legs (Fig. 7), carapace pale creamy brown. Median ocular tubercle raised, narrow, sloping sharply at fovea. Single median line running from front of median ocular tubercle to middle of chilum. Chelicerae orange, darker laterally; chelicerae with a row of two to four large teeth close to fang base, prolateral of promarginal teeth row; teeth row comprising very large teeth curved at tips, interspersed with small teeth, with extensive denticle field retrolateral of teeth row near cheliceral base (Fig. 18). Endites strongly elongated and slender prolaterally, strongly curved upwards. Sternum and legs pale yellow-brown. Legs short and stout, leg formula 4231; legs III and IV more robust than legs I and II; leg I with three to five spines on patellae and



Figures 69–72. *Calommata transvaalica* Hewitt female genitalia 69 and left palp of male 70–72) 69 dorsal view 70 prolateral view 71 ventral view 72 retrolateral view. Scale bars: 1mm.

two on tibiae; leg II with few spinules on patellae and several spinules on tibiae and metatarsi; legs III and IV with spinules from patellae to tarsi (mainly dorsal and prolateral); legs II to IV covered in bristles. Abdomen globose and pale grey, with indistinct median heart marking anteriorly (Fig. 7). Epigyne forming a broad, weakly sclerotised plate ventrally, in dorsal view with single pair of large, transversely oval spermathecae (Fig. 69). Female palp short, tibiae and tarsi flattened.

Male from Groenkloof Nature Reserve. Measurements: CL 2.10, CW 1.85, AL 2.70, AW 1.70, TL 6.40 (6.20–6.40). Length of leg segments, and total: I 1.66 + 0.60 + 1.10 + 1.43 + 1.28 = 6.07; II 1.67 + 0.70 + 0.95 + 1.50 + 1.47 = 6.29; III 1.45 + 0.70 + 0.70 + 1.55 + 1.95 = 6.35; IV 1.85 + 0.78 + 1.05 + 1.85 + 1.90 = 7.43. Carapace index 1.14; patella-tibia index 0.81.

Carapace and chelicerae dark brown in colour (Fig. 8). Chelicerae with single row of large teeth, gradually decreasing in size from fang base to cheliceral base, without denticles near cheliceral base (Fig. 19). Carapace oval in shape. Median ocular tubercle raised, narrow, darker in colour. Sternum and coxae yellow-brown, femora, patellae and tibiae brown, metatarsi yellow-brown, tarsi yellow. Legs weakly covered with bristles; prolateral side of patellae, tibiae and metatarsi of legs II–IV covered with spinules. Abdomen dark

grey, nearly black, with dark orange-brown scutum anteriorly (Fig. 8). Palp with short cymbium, cymbium tip in ventral view tapering to rounded point; embolus and conductor orientated transversely to palpal axis, pointing retrolaterally, distal ends projecting beyond retrolateral cymbial margin; conductor short, slightly broadened distally, with single sharp, curved tooth on its dorsal surface; embolus long and straight (Figs 70–72).

Remarks. It is clear from the redescriptions of both sexes of *C. simoni*, and the redescription of the female and first description of the male of *C. transvaalica* provided here, that the two species have distinct differences in their somatic and genitalic morphology, most notably regarding their cheliceral dentition, number of spermathecae in the female epigyne and orientation and length of the male embolus and conductor. Consequently, we reject Benoit's (1967) synonymy of the two species and propose the revalidation of *C. transvaalica*. The specimens collected by Van Dam and Roberts (1917) between Villeria and Derdepoort near Pretoria could not be traced.

The abdomen of the female holotype of *C. transvaalica* is damaged and therefore the specimen was not redescribed. The holotype is the smallest of the known females of this species (18.60mm long). The colour of all available female specimens has likely faded over time in 70 % ethanol.

Distribution. Limpopo and Gauteng Provinces, South Africa (Fig. 73).

Biology. The biology of *C. transvaalica* was studied by Van Dam and Roberts (1917) at Roodeplaat near Pretoria, South Africa, in the days following heavy rainfall. They first detected a female (the holotype) by kicking up a tuft of grass that disclosed white webbing, which was followed downwards into the ground to locate the spider. They subsequently discovered additional nests on bare ground. They described the nests as slightly raised above the ground at the top, and then from the inner rim they were neatly rounded off, gradually sloping outwards and downwards to the level of the ground with the outer surface covered with earth that resembled the surroundings. The interior of the tube was lined with loose, highly adhesive silky webbing. They suggested that the adhesive webbing may afford the spider some protection against the intrusion of enemies. The nests were deep (22–25cm) and vertical for the greater part of their depth (Van Dam and Roberts 1917). Hewitt (1916) commented that *C. transvaalica* specimens had a very pronounced and objectionable odour and compared it to decomposing stable manure.

The recently collected series of males from Zwartkoppies Farm (TMSA 23875) was collected in open pitfalls without preservative from a site in open *Acacia karroo* woodland on red structured clay soils (30–45% clay in A horizon, Shortlands form). The site has a gentle slope and had recently been burned. The activity of the males appears to be related to heavy rainfall that had fallen two days prior to the collection of the males. It thus seems that males do not emerge on the night immediately following a heavy shower, but instead on the night thereafter (Ian Engelbrecht, pers. comm.).



Figure 73. Distribution of *Calommata* species in the Afrotropical Region: *C. megae* (open square), *C. meridionalis* (solid squares). *C. namibica* (solid triangle), *C. simoni* (solid circles), *C. tibialis* (open triangles) and *C. transvaalica* (open circles).

Discussion

Calommata is a small but widespread genus that is known from Africa, Israel and South East Asia. Two species are present in West Africa, one species in Central Africa and four species in southern Africa. The type of habitat seems to play a role in separating the species biogeographically. *Calommata simoni* is perhaps the most flexible in terms of habitat requirements, and occurs in forests, savannahs and grasslands in tropical Africa. In contrast, *C. tibialis* was only found in wooded savannahs in West Africa. In southern Africa, *C. megae*, *C. namibica* and *C. transvaalica* occur in the Savannah biome, but with contrasting vegetation structures and climatic variables. Lastly, *C. meridionalis* occurs only in the Grassland biome of central South Africa (Fig. 73).

Atypid spiders are widely regarded as being of conservation importance due to their generally specific environmental requirements, low rates of dispersal and general scarcity (Pedersen and Loeschcke 2001, Řezáč et al. 2007, Řezáč 2009). Several species are considered critically threatened or endangered and are included in various country or regional Red Data lists (e.g. Platen et al. 1996, Komposch and Steinberger 1999, Blick and Scheidler 2003, Farkač et al. 2005, Sato et al. 2007).

Calommata transvaalica (previously as C. simoni in South Africa) was until recently presumed nationally extinct in South Africa as the species had last been reported in the 1920's (Dippenaar-Schoeman 2002). It was subsequently rediscovered in 2003 (Groenkloof NR) and 2005 (Blouberg NR). This species was submitted for an initial Red List assessment in April 2008, but during the process of assessment, questions arose regarding the taxonomic status of this species and it was included in the Data Deficient category for taxonomic reasons (Engelbrecht 2008). As the genus has now been revised, the South African species can be resubmitted for Red Data listing to promote conservation of these spiders. In comparing the two South African species, populations of C. transvaalica are under severe threat in the south of its range due to rapid urbanisation and habitat loss in the Gauteng Province. The recent collection of 13 males from Zwartkoppies Farm near Pretoria suggests that healthy populations do still exist in natural habitats in the province. In contrast, C. meridionalis is experiencing considerably lower threat levels (the three known localities are from undisturbed grassland) and potential agricultural expansion, especially from cultivation agriculture, perhaps represents its greatest threat. If more attention can be paid to the apparent soil preferences of South African Calommata in future when conducting pitfall surveys it is likely that further populations could be located in suitable habitat. As such, South African Calommata may serve as ideal candidates of predictive modelling due to their restricted distributions, limited knowledge of their biology, and increasing threats to their survival (see Jiménez-Valverde and Lobo 2006, Jiménez-Valverde et al. 2007).

Acknowledgments

This study forms part of the M.Sc of the senior author at the University of the Free State and was funded through a National Research Foundation of South Africa (NRF) grant in the NRF Thuthuka program to the second author (TK2008050500003). The NRF also provided a student travel grant to the senior author through its SABI program to visit the MRAC (GUN 69525). The following curators of the collections are thanked for the loan of material that made this study possible: Norman Platnick (AMNH), Janet Beccaloni (BMNH), Ansie Dippenaar-Schoeman (NCA), Leon Lotz (NMBA), Audrey Ndaba (NMSA), Robin Lyle (TMSA) and Jonathan Coddington (USNM). Ansie Dippenaar-Schoeman is thanked for providing some rare taxonomic and biological literature, and she and Ian Engelbrecht are thanked for useful discussion on the conservation importance of the South African species. Meg Cumming kindly provided biological information on the C. megae specimens that she collected in Zimbabwe and Ian Engelbrecht on C. transvaalica specimens collected in South Africa. Field work by Ian Engelbrecht was funded by the Gauteng Dept. of Agriculture and Rural Development and Duncan MacFadyen of E. Oppenheimer & Son. Richard Gallon and Jeremy Miller are thanked for their constructive comments that helped improve the manuscript.

References

- Anderson JF (1987) Morphology and allometry of the purse-web of *Sphodros abboti* (Araneae, Atypidae): respiratory and energetic considerations. Journal of Arachnology 15: 141–150.
- Ayoub NA, Garb JE, Hedin M, Hayashi, CY (2007) Utility of the nuclear protein-coding gene, elongation factor-1 gamma (*EF-1γ*), for spider systematics, emphasizing family level relationships of tarantulas and their kin (Araneae: Mygalomorphae). Molecular Phylogenetics and Evolution 42: 394–409. http://dx.doi.org/10.1016/j.ympev.2006.07.018
- Benoit PLG (1967) Le genre *Calommata* Lucas en Afrique (Aran.-Orth.-Atypidae). Bulletin et Annales de la Société royal entomologique de Belgique 103: 283–288.
- Blandin P (1971) Découverte en Côte d'Ivoire de *Calommata simoni* Pocock [Aran.-Orth.-Atypidae]. Bulletin de l'Institut Fondamental Afrique Noire 33: 48–52.
- Blick T, Scheidler M (2003) Rote Liste gefährdeter Spinnen (Arachnida: Araneae) Bayerns. Schriftenreihe des Bayerischen Landesamtes für Umweltschutz 166: 308–321.
- Charpentier P (1995) New data on the African atypid spider *Calommata simoni* Pocock Orthognatha, Atypidae. Journal of the British Tarantula Society 10: 81–86.
- Clark DJ (1969) Notes on the biology of *Atypus affinis* Eichwald (Araneae Atypidae). Bulletin of the British Arachnological Society l: 36–39.
- Coyle FA (1983) Aerial dispersal by mygalomorph spiderlings (Araneae, Mygalomorphae). Journal of Arachnology 11: 283–286.
- Coyle EA, Greenstone MH, Hultsch, A-L, Morgan, CW (1985) Ballooning mygalomorphs: estimates of the masses of *Sphodros* and *Ummidia* ballooners (Araneae: Atypidae, Ctenizidae). Journal of Arachnology 13: 291–296.
- Coyle FA, Shear WA (1981) Observations on the natural history of *Sphodros abboti* and *Sphodros rufipes* (Araneae, Atypidae), with evidence for a contact sex pheromone. Journal of Arachnology 9: 317–326.
- Dippenaar-Schoeman AS (2002) Baboon and trapdoor spiders of southern Africa: an identification manual. Plant Protection Research Institute Handbook No. 13. Agricultural Research Council, Pretoria, 128 pp.
- Dippenaar-Schoeman AS, Jocqué R (1997) African spiders: an identification manual. Plant Protection Research Institute Handbook no. 9. Agricultural Research Institute, Pretoria, 392 pp.
- Edwards RL, Edwards, EH (1990) Observations on the natural history of a New England population of *Sphodros niger* (Araneae, Atypidae). Journal of Arachnology 18: 29–34.
- Engelbrecht I (2008) Red List Assessment: The baboon spiders *Harpactira hamiltoni* Pocock 1904 and *Brachionopus pretoriae* Purcell 1904 (Araneae: Theraphosidae) and the African Purse Web Spider *Calommata simoni* Pocock 1903 (Araneae: Atypidae). Red List assessment report. Gauteng Department of Agriculture, Conservation and Environment, Johannesburg, 4 pp.
- Farkač J, Král D, Škorpík M [eds.] (2005) Červený seznam ohrožených druhů České republiky. Bezobratlí. Agentura ochrany přírody a krajiny ČR, Praha, 760 pp.
- Gertsch WJ, Platnick, NI (1980) A revision of the American spiders of the family Atypidae (Araneae, Mygalomorphae). American Museum Novitates 2704: 1–39.

- Goloboff PA (1993) A reanalysis of mygalomorph spider families (Araneae). American Museum Novitates 3056: 1–32.
- Hedin M, Bond JE (2006) Molecular phylogenetics of the spider infraorder Mygalomorphae using nuclear rRNA genes (18S and 28S): conflict and agreement with the current system of classification. Molecular Phylogenetics and Evolution 41: 454–471. http://dx.doi. org/10.1016/j.ympev.2006.05.017
- Hewitt J (1916) Descriptions of new South African spiders. Annals of the Transvaal Museum 5: 180–213.
- Jiménez-Valverde A, García-Díez T, Bogaerts S (2007) First records of the endangered spider Macrothele calpeiana (Walckenaer, 1805) (Hexathelidae) in Portugal. Boletín Sociedad Entomológica Aragonesa 41: 445–446.
- Jiménez-Valverde A, Lobo JM (2006) Distribution determinants of endangered Iberian spider Macrothele calpeiana (Araneae, Hexathelidae). Environmental Entomology 35: 1491–1499. http://dx.doi.org/10.1603/0046-225X-35.6.1491
- Jocqué R, Dippenaar-Schoeman AS (2006) Spider Families of the World. Royal Museum for Central Africa, Tervuren, 336 pp.
- Komposch C, Steinberger, KH (1999) Rote Liste der Spinnen Kärntens (Arachnida: Araneae). Naturschutz in Kärnten 15: 567–618.
- Kraus O (1978) *Liphistius* and the evolution of spider genitalia. Symposia of the Zoological Society, London 42: 235–254.
- Kraus O, Baur H (1974) Die Atypidae der West-Paläarktis. Systematik, Verbreitung und Biologie (Arach.: Araneae). Abhandlungen aus der Naturwissenschaften Verein, Hamburg 17: 85–116.
- Levy G (2007) *Calommata* (Atypidae) and new spider species (Araneae) from Israel. Zootaxa 1551: 1–30.
- Oliger TI (1998) A new *Atypus* from the Russian Far East (Aranei: Atypidae). Arthropoda Selecta 7: 201–204.
- Pedersen AA, Loeschcke V (2001) Conservation genetics of peripheral populations of the mygalomorph spider *Atypus affinis* (Atypidae) in northern Europe. Molecular Ecology 10: 1133–1142. http://dx.doi.org/10.1046/j.1365-294X.2001.01266.x
- Platen R, Blick T, Sacher P, Malten A (1996) Rote Liste der webspinnen Deutschlands (Arachnida, Araneae). Arachnologische Mitteilungen 11: 5–31.
- Platnick NI (2010) The World Spider Catalog, Version 11.0. Family Atypidae. American Museum of Natural History. http://research.amnh.org/iz/spiders/catalog/ATYPIDAE.html [accessed 22.IX.2010, last updated 17.III.2010]
- Pocock RI (1903) Some new spiders from the Cameroons collected by Mr G. L. Bates. Annals and Magazine of Natural History 11: 258–264.
- Raven R (1985) The spider infraorder Mygalomorpha (Araneae): cladistics and systematics. Bulletin of the American Museum of Natural History 182: 1–180.
- Řezáč M (2009) Rozšíření a ochrana pavouků sklípkánků (Araneae: *Atypus* spp.) v České republice. Příroda 28: 3–43.

- Řezáč M, Král J, Musilová J, Pekár S (2006) Unusual karyotype diversity in the European spiders of the genus *Atypus* (Araneae: Atypidae). Hereditas 143: 123–129. http://dx.doi. org/10.1111/j.2006.0018-0661.01949.x
- Řezáč M, Řezáčová V, Pekár S (2007) The distribution of purse-web Atypus spiders (Araneae: Mygalomorphae) in central Europe is constrained by microclimatic continentality and soil compactness. Journal of Biogeography 34: 1016–1027.http://dx.doi.org/10.1111/j.1365-2699.2006.01670.x
- Sato T, Wada T, Nakashima C, Tsurusaki N (2007) New records of *Calommata signata* (Araneae: Atypidae) from eastern part of Tottori Prefecture, Honshu, Japan. Natural History Research of San'in 3: 6–10.
- Schwendinger PJ (1990) A synopsis of the genus *Atypus* (Araneae, Atypidae). Zoologica Scripta 19: 353–366. http://dx.doi.org/10.1111/j.1463-6409.1990.tb00263.x
- Tanikawa A (2006) A new species of the spider genus *Atypus* (Araneae: Atypidae) from Amamiôshima, Japan. Acta Arachnologica 55: 25–27. http://dx.doi.org/10.2476/asjaa.55.25
- Van Dam G, Roberts A (1917) Notes on nests of some trapdoor spiders and the nest of *Calom-mata transvaalicus* Hewitt. Annals of the Transvaal Museum 5: 218–233.
- Zhu M-S, Zhang F, Song D, Qu P (2006) A revision of the genus *Atypus* in China (Araneae: Atypidae). Zootaxa 1118: 1–42.

RESEARCH ARTICLE



Revision of the stiletto fly genera Acupalpa Kröber and Pipinnipons Winterton (Diptera, Therevidae, Agapophytinae) using cybertaxonomic methods, with a key to Australasian genera

Shaun L. Winterton

California State Collection of Arthropods, California Department of Food & Agriculture, Sacramento, CA, USA

urn:lsid:zoobank.org:author:37F5AC48-EC3A-47ED-902B-2BD1467CCA72

Corresponding author: Shaun L. Winterton (wintertonshaun@gmail.com)

Academic editor: Martin Hauser | Received 3 September 2010 | Accepted 15 March 2011 | Published 4 May 2011

urn:lsid:zoobank.org:pub:3252AEA9-3D2A-42B3-B65E-6072F8E30D35

Citation: Winterton SL (2011) Revision of the stiletto fly genera *Acupalpa* Kröber and *Pipinnipons* Winterton (Diptera, Therevidae, Agapophytinae) using cybertaxonomic methods, with a key to Australasian genera. ZooKeys 95: 29–78. doi: 10.3897/zookeys.95.1461

Abstract

Australian stiletto flies of the sister-genera Acupalpa Kröber, 1912 and Pipinnipons Winterton, 2001 (Diptera: Therevidae: Agapophytinae) are revised. Twelve new species of Acupalpa are described, while Acupalpa imitans (White, 1915), comb. n. is transferred from Pipinnipons and Acupalpa albimanis (Kröber, 1914), comb. n. is transferred from Ectinorhynchus Macquart as a senior synonym of Acupalpa pollinosa Mann. The total number of species of Acupalpa is therefore increased to 19: A. albimanis (Kröber), comb. n., A. albitarsa Mann, A. boharti sp. n., A. divisa (Walker), A. dolichorhyncha sp. n., A. glossa sp. n., A. imitans (White), comb. n., A. irwini Winterton, A. melanophaeos sp. n., A. miaboolya sp. n., A. minuta sp. n., A. minutoides sp. n., A. notomelas sp. n., A. novayamarna sp. n., A. rostrata Kröber, A. semirufa Mann, A. westralica sp. n., A. yalgoo sp. n. and A. yanchep sp. n. Three new species of Pipinnipons are described, increasing the total number of species to five: P. chauncyvallis sp. n., P. fascipennis (Kröber), P. kampmeierae sp. n., P. kroeberi Winterton, and P. sphecoda sp. n. Pipinnipons and Acupalpa are rediagnosed in light of the new species presented herein and revised keys to species are included. A dichotomous key to genera of Australasian Therevidae is included. As an empirical example of cybertaxonomy, taxonomic descriptions were composed using a character matrix developed in Lucid Builder (in Structured Descriptive Data (SDD) format) to generate natural language descriptions supplemented by online specimen and image databases. Web resources are provided throughout the document including: a) links to high resolution colour images of all species on Morphbank, b) registration of authors, publications, taxon names and

Copyright Shaun L Winterton. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

other nomenclatural acts in Zoobank, with assignment of Life Science Identifiers (LSIDs) for each, c) links to Genbank accession records for DNA sequences, and d) assignment of LSIDs to specimen records with links to respective records in an online Therevidae specimen database.

Keywords

cybertaxonomy, LSID, character matrix, natural language description

Introduction

The stiletto fly subfamily Agapophytinae is comprised of 23 described genera restricted in the Australasia region, yet with three additional described genera endemic to Chile and Argentina (Winterton 2006). *Acupalpa* Kröber, 1912 and *Pipinnipons* Winterton, 2001 are agapophytine sister genera found exclusively in Australia. Along with *Agapophytus* Guérin, 1831, these genera form a relatively derived clade characterized by an antenna with an elongate cylindrical scape and flagellum (Winterton 2000; Winterton et al. 2001). *Agapophytus* is differentiated from these genera by an elongate scape typically longer than, or equal to, the length of the flagellum, while in *Acupalpa* and *Pipinnipons* the scape is shorter than the flagellum. *Acupalpa* contains seven previously described species: *A. albimanis* (Kröber, 1914), comb. n. (=*A. pollinosa* Mann, 1929, syn. n.), *A. albitarsa* Mann, 1929, *A. divisa* (Walker, 1850), *A. imitans* (White, 1915), comb. n., *A. irwini* Winterton, 2000, *A. rostrata* Kröber, 1912, and *A. semirufa* Mann, 1929. *Pipinnipons* includes two previously described species: *P. fascipennis* (Kröber, 1928) and *P. kroeberi* Winterton, 2001.

Twelve new species of Acupalpa are described herein: A. boharti sp. n., A. dolichorhyncha sp. n., A. glossa sp. n., A. melanophaeos sp. n., A. miaboolya sp. n., A. minuta sp. n., A. minutoides sp. n., A. notomelas sp. n., A. novayamarna sp. n., A. westralica sp. n., A. yalgoo sp. n., and A. yanchep sp. n. Many of these species are from Western Australia, indicating a rich diversity of this genus in the western region of the continent. Addition of these new species significantly broadens the concept of Acupalpa beyond the characters defining the genus in previous treatments (i.e. Mann 1929; Winterton 2000; Winterton et al. 2001), therefore the genus and constituent species is rediagnosed herein and a revised key to species presented. Acupalpa imitans (White, 1915), comb. n. is transferred from Pipinnipons based on the discovery of new material matching the original description, while A. albimanis (Kröber, 1914), comb. n. is transferred from Ectinorhynchus Macquart, 1850 as the latter is a senior subjective synonym of A. pollinosa. Three new species of Pipinnipons are described, increasing the total number of species to five: P. chauncyvallis sp. n., P. fascipennis, P. kampmeierae sp. n., P. kroeberi, and P. sphecoda sp. n. A key to Australasian stiletto fly genera is also included.

The modern taxonomic enterprise represents a gradual paradigm shift away from tedious traditional methods toward rapid, semi-automated ones (i.e. recently termed cybertaxonomy), with increased efficiency in data handling through the use of online databases for information such as label metadata, specimen images, name registration, semantic mark-up and natural language descriptions from character matrices (Winterton 2009). The concept itself is not new, with single zoological registries (e.g. Brown 1961) and rapid descriptive processes (e.g. Erwin and Johnson 2000) espoused previously, but the actual empirical use of informatics tools to enhance the taxonomic descriptive process (i.e. online databases) is only now becoming a reality. Several authors have embraced modern cybertaxonomic methods through the incorporation of such digital, web-based, resources in taxonomic descriptions (e.g. Pyle et al. 2008; Johnson et al. 2008; Deans and Kawada 2008; Miller et al. 2009; Winterton 2009; Penev et al. 2010; Blagoderov et al. 2010). The methods used by these authors, and this paper, are empirical examples of how digital tools can significantly speed the process of documenting biodiversity through rapid generation of natural language descriptions derived from matrix based character data in a standard-ized format suitable for multiple use (e.g. distributed morphological ontologies).

Materials and methods

Adult morphological terminology follows McAlpine (1981) as modified by Winterton et al. (1999a) and Winterton (2006). Hauser and Irwin (2003) provide a convincing argument for the use of the term pubescence (*sensu* Nichols 1989) instead of pruinescence (*sensu* Winterton et al. 1999a) to describe the various types of microtrichia covering the adult body and is used here throughout the text. The term velutum (Winterton et al. 1999a) is retained to describe a particular type of very dense, unidirectional and reflective (i.e. velvet-like) microtrichia typically found on the male abdomen (silver coloured) or on the femora of agapophytine therevids (variously coloured but usually dark). Genitalia were macerated in 10% KOH to remove soft tissue, then rinsed in distilled water and dilute glacial acetic acid, and dissected in 80% ethanol. Genitalia preparations were placed in glycerine in a genitalia vial mounted on the pin beneath the specimen.

Types are deposited in the following institutions and collections: Queensland Museum (Brisbane) (QM), Australian Museum (Sydney) (AMS), Australian National Insect Collection (Canberra) (ANIC), Senckenberg Deutsches Entomologisches Institut, Müncheberg, Germany (DEI), University of Queensland Insect Collection (Brisbane) (UQIC), University of California, Davis, Bohart Museum (UCDC), Western Australian Museum (Perth) (WAM), Michael E. Irwin private collection [to be ultimately housed in the California Academy of Sciences] (MEIC/CAS), Greg Daniels private collection [to be ultimately housed in the Australian Museum] (GDCB/AMS), Naturhistorisches Museum Wien, (NMW), Harvard University Museum of Comparative Zoology (MCZ), Oxford University Museum of Natural History (OUMNH), University of California, Riverside (UCR), Universität von Hamburg Zoologisches Institut und Zoologisches Museum (ZMUH). All types have been examined. Numbers quoted with individual specimens as MEI000000 are unique identifiers in the therevid database MANDALA and are attached to each specimen as a yellow or white label (Kampmeier & Irwin 2009). Links are provided in this document to Life Science Identifiers (LSIDs) to specimen records with links to respective records in an online MANADALA Therevidae specimen database and Discover Life (http://www.discoverlife.org). Note that some web browsers are not able to read and format RSS (Really Simple Syndication) feeds and/or XML without additional software extensions or plug-ins. Details of current issues with select web browsers and LSID resolvers can be found on the Biodiversity Information Standards (TDWG) LSID resolver website (http://lsid.tdwg.org/). Material examined lists were exported from MANDALA. Descriptions were constructed using Lucid Builder 3.5, using a matrix database of character states, which were then exported using a natural language function into XML and a text document. Links are provided to Genbank accession records for DNA sequences where available. Specimen images were taken using a digital camera with a series of images montaged using Helicon Focus (©HeliconSoft). Descriptions are aided by the provision of embedded URL links in the document to high-resolution digital images of all species in Morphbank. All nomenclatural acts are registered in Zoobank as per the recent proposed amendment to the International Code of Zoological Nomenclature for a universal register for animal names (Polaszek et al. 2005a,b; Pyle et al. 2008; ICZN 2008).

Key to genera of Australasian Therevidae

The following key to genera supersedes those by Winterton et al. (1999b) and Winterton et al. (2001) and includes all genera found throughout the region east of Wallace's Line, incorporating landmasses such as Australia, New Zealand, New Caledonia, Papua New Guinea, and eastern Indonesia. The enigmatic *Taenogera* genus group (sensu Winterton et al. 1999b) is herein included within an expanded concept of Agapophytinae (Winterton 2006). The subfamilies Xestomyzinae and Phycinae are absent from the Australasian region.

1 Femora with multiple vestiture types, often with appressed, scale-like setae; strong macrosetae usually present on femora; two spermathecae in female, ventral apodeme of parameral sheath not forked, usually narrow, dorsal apodeme of parameral sheath well developed, usually broad and hood-like...... Femora with only a single type of setae, often short and dark setae admixed with longer pale setae, never appressed or scale-like, macrosetae sometimes present; three spermathecae (rarely reduced to one or two); ventral apodeme of parameral sheath forked or anteriorly emarginate, never as single narrow apodeme; dorsal apodeme of parameral sheath usually forked or greatly reduced, rarely broad or hood-likesubfamily AGAPOPHYTINAE - 4 2 Male frons narrow so that eyes almost contiguous medially; usually a single row of postocular setae dorsomedially in male; mid coxal pile present; wing cell m₂ open or closed; femora with distinct appressed pile (Indonesia, Papua New Guinea) (relatively slender flies)...... Irwiniella Lyneborg Male frons usually wider than ocellar tubercle so that eyes widely separated; usually many rows of postocular setae dorsally in both sexes; mid coxal pile absent (rarely present in Anabarhynchus); wing cell m₃ open; femora with

multiple vestiture types, sometimes without distinct appressed pile (robust 3 Size variable, but never uniformly black; (Australia, Papua New Guinea, New Caledonia, Fiji, New Zealand) Anabarbynchus Macquart Large, black, robust flies; female sternite 8 with posterolateral slits (New Zea-4 Elongate strip of velutum (velvet pubescence) on ventromedial surface of hind femur present; patch of velutum on ventral surface of male gonocoxite often present (rarely reduced or absent); wing cell m₂ open or closed (Australia, Papua New Guinea, Indonesia).....5 Femora without of velutum patches; gonocoxites without velutum patch ventrally (rarely present); wing cell m₂ open (Australia, New Zealand, Papua New Guinea, Indonesia, New Caledonia)18 5 Wing cell m₂ open6 Wing cell m, closed9 6 Fore femur without velutum patch on ventral surface; hind femur with one (rarely more) subapical anteroventral seta; antennae usually longer than head, flagellum cylindrical; occiput often overlain with silver, gold and matte black pubescence (Australia)Evansomyia Mann Fore femur with velutum patch on ventral surface; hind femur without subapical setae; antennae usually shorter than head, flagellum conical; occiput Short, relatively small flies; male genitalia with ventral lobe of male gono-7 coxite not broad or enlarged; medial atrium (Winterton et al. 2001: fig 22) usually present (Australia) Parapsilocephala Kröber (part) Elongate, small to relatively large flies; ventral lobe of gonocoxite broad, enlarged; medial atrium absent8 8 Eyes relatively small; occiput concave, postocular ridges angled, not in same plane; relatively few postocular setae, arranged in a poorly defined single row; setae absent on posterior surface of mid coxa; postspiracular setae absent; mid femur without elongate velutum patch; distiphallus spinose apically (Aus-Eyes regular size; occiput concave or convex, postocular ridges straight or almost in same plane, usually with multiple rows of postocular setae dorsally, some males with only a single row; setae sometimes present on prosternum, and on posterior surface of mid coxa; postspiracular setae sometimes present; additional elongate velutum patch on posteroventral surface of mid femur often present; distiphallus without spines (Australia) Laxotela Winterton & Irwin 9 Flagellum elongate, narrow cylindrical in cross-section; scape also narrow elongate, usually more than 3× length of pedicel; antennae typically longer than head (rarely equal in length); antennae appear positioned on middle or upper region of head, rarely on lower frons; antennae usually not projecting anteroventrally to body axis10

-	Flagellum conical, turbinate or oval shaped, usually flattened laterally; scape
	length variable but never elongate and narrow cylindrical, usually less than
	3× pedicel length, sometimes bulbous; antennae shorter than head, although
	sometimes close to equal head length, antennae positioned very low on head
	and projecting anteroventrally to body axis
10	Flagellum shorter than or equal to scape length; scape usually longer than
	head (Australia, Papua New Guinea)Agapophytus Guérin
-	Flagellum longer than scape length; scape never longer than head11
11	Palpi spatulate; face narrow, not expansive or protruding (Australia)
-	Palpi narrow to acuminate, not broadened apically; face expansive and often
	protruding anteriorly (Australia)
12	Black individuals with silver-white velutum stripe along lower half of thorax
	and abdomen; male genitalia with articulated gonocoxal process absent or
	greatly reduced; ventral lobe greatly enlarged into blade-like structure (Aus-
	tralia)
_	Body colour and markings otherwise (silver velutum stripe on pleuron pre-
	sent in some genera); male genitalia with well-developed articulated gono-
	coxal process; ventral lobe not large and blade-like
13	Wing typically strongly banded: abdomen slender and narrow basally, diameter
	of thorax distinctly greater than base of abdomen: hind femur longer than fore
	and mid femora: male genitalia with medial atrium present or absent
_	Wing hvaline or variably infuscate but not handed: abdomen thicker basally
_	diameter similar to thoray slightly tapered posteriorly but not slender: femora
	annever similar to morax, signify tapered posteriorly but not siender, remora
1/1	Mala anandrium archad to partially conceal conceavitou medial atrium present
14	Male epandrium arched to partially conceal gonocoxites; mediai atrium pre-
	sent between gonocoxites; nypandrium with patch of strong posterioriy di-
	rectly setae (Australia)
_	Male epandrium not concealing gonocoxites; gonocoxites meeting medi-
	ally such that medial atrium is absent; hypandrium without patch of strong
	setae
15	Small species (usually <6.0 mm body length); scutellum often dorsally acumi-
	nate; frequently excellent ant mimics; male with single row of postocular se-
	tae (Australia, Papua New Guinea, Timor)
-	Relatively larger species (8.0–12.0 mm body length), usually larger than 6.0mm
	total body length; scutellum rounded, never dorsally acuminate; male with mul-
	tiple rows of postocular setae (Australia)Acraspisoides Hill & Winterton
16	Ventral lobe very long and narrow, length equal to gonostylus; plate or cup-
	like velutum patch on gonocoxites; frons flattened or rounded, without callus
	above antenna (Australia) Patanothrix Winterton
_	Ventral lobe shorter than gonostylus; velutum barely evident on gonocoxite,
	found mainly on atrium membrane; frons typically with glossy callus above an-
	tenna

17	Three spermathecae present; transverse velutum plaques absent on male abdo- men; gonocoxal apodemes and distiphallus short; ejaculatory apodemes strong-
	ly sclerotized but not enlarged (Australia) Parapsilocephala Kröber (part)
_	One spermatheca present; transverse velutum plaques often present on male abdomen; gonocoxal apodemes and distiphallus often greatly elongate; ejacula- tory apodemes greatly enlarged (Australia) <i>Bonjeania</i> Irwin & Lyneborg
18	Hind femur without subapical setae
_	Hind femur with one (rarely more) subapical anteroventral seta
19	Male gonocoxite with processes absent; male usually with a single row of postocular setae dorsally; medium sized flies; colouration and markings variable, scutum often yellow or tan ground colour with dark brown tessellate or spotted pattern (Australia)
_	Male gonocoxite with gonocoxal process present; male usually with multiple
	poorly defined rows of postocular setae dorsally; relatively small flies; grey and black colouration and markings (Australia)
20	Body usually large to medium sized, robust, glossy dark metallic blue or or-
	ange; abdomen abruptly tapered; small patch of postspiracular setae present on thorax; wing extensively black (sometimes hyaline basally) or orange in-
_	Body size variable, usually relatively slender, never glossy metallic blue or or-
	ange; abdomen elongate, evenly tapered; thoracic postspiracular setae absent;
	wing infuscation variable, usually banded or hyaline, never uniform orange or black
21	Scape short, setae on antennae and head relatively short; two pairs of scutellar setae; wing with uniform orange infuscation (Australia)
	Eupsilocephala Kröber
_	Scape elongate with numerous enlarged setae; single pair of scutellar setae; wing either with uniform black infuscation or hyaline basally (Australia)
	Iohnmannia Irwin & Ivnehorg
2.2	Male and female occiput convex, variously overlain with bronze, matte black.
	silver and gold pubescence: multiple rows of postocular setae in male: abdo-
	men of equal diameter along length: distiphallus broad, cylindrical: medium
	to large individuals (Australia)
_	Male occiput typically flat to concave, not distinctly convex, rarely overlain
	with bronze, matte black, silver and gold pubescence; usually single row of
	postocular setae in male; abdomen tapered; distiphallus usually narrow; size variable
23	Antennae shorter than or equal to head length; scape usually $< 2 \times$ pedicel
-	length, usually with only small setae on scape and frons
_	Antennae longer than head; scape > 3× pedicel length, often with strong setae
	on scape and frons
24	Occiput with multiple rows of postocular setae in both sexes; male with ar- ticulated gonocoxal process greatly reduced or absent; gonocoxite sometimes

	with large horn-like posterior process (Australia)
	Actenomeros Winterton & Irwin
_	Occiput with single row of postocular setae in male; male with articulated
	gonocoxal process well developed; gonocoxite without horn-like posterior
	process (Australia)
25	Occiput overlain with silver and matte black pubescence; male with single
	row of postocular setae; typically larger species with banded wings
_	Occiput overlain with grey pubescence; male often with multiple rows of
	postocular setae; smaller species with hyaline or slightly suffused wings (Aus-
	tralia) <i>Taenogerella</i> Winterton & Irwin
26	Male with medial atrium between gonocoxites, articulated gonocoxal process
	greatly reduced; velutum patch present ventrally on gonocoxites; colouration
	and markings often sexually dimorphic (Australia, New Zealand)
	<i>Ectinorhynchus</i> Macquart
_	Male without medial atrium between gonocoxites, articulated gonocoxal process
	well developed; velutum absent on gonocoxites; colouration and markings not
	sexually dimorphic (Australia, Papua New Guinea)Squamopygia Kröber

Acupalpa Kröber

urn:lsid:zoobank.org:act:68450BF4-0179-4194-BE8D-422966FC95C7 http://species-id.net/wiki/Acupalpa

Acupalpa Kröber 1912: 152; Kröber 1913: 18; Mann 1929: 23; Hardy 1939: 47 [as Acutipalpa]; Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 227 [revision]; Winterton et al. 2001: 197. Type species: Acupalpa rostrata Kröber 1912: 152.

Diagnosis. Antennal scape shorter than or equal to flagellum; antenna elongate, cylindrical, total length slightly longer than or equal to head length; upper part of frons flat or slightly concave above antenna; face either protruding anteriorly below antennal base, or broadly rounded, expansive, short dark setae often present; parafacial setae absent; palpus apically narrow or acute, not spatulate; mouthparts length variable, frequently elongate and forward projecting (Fig. 3H); male postocular ridge with single row of macrosetae immediately laterad of ocellar tubercle, female with more than one row; wing infuscate, usually strongly banded; setae absent on wing vein R₃; cell m₂ closed; velutum patches on fore and hind femora; femora without macrosetae; single type of setal pile on femora, setae not appressed; prosternal furrow without setae; post spiracular pile absent; pleuron orange to black, overlain with sparse silver pubescence; mid coxa without setae on posterior surface; gonocoxites with velutum patch on ventral surface (Fig. 3B); articulated gonocoxal process present; hypandrium present; ventral apodeme of parameral sheath forked; dorsal apodeme of parameral sheath 'T'-shaped (Fig. 3F); three spermathecae in female; spermathecal sac present, sac simple or with smaller additional lobes basally, often with outer elongate lobes; spermathecal ducts joining common duct before bursa (Fig. 3G), female


Figure 1. *Acupalpa divisa* (Walker), female, Brisbane, Queensland. Body length= 7.0 mm. (Photo: Anthony O'Toole, University of Queensland).



Figure 2. *Acupalpa yanchep* sp. n., female, Yanchep, Western Australia. Body length= 9.0 mm. (Photo: S.L. Winterton).



Figure 3. *Acupalpa* spp.: **A** *A. notomelas* sp. n., epandrium dorsal **B** gonocoxites, ventral **C** *A. imitans* (White) comb. n., gonocoxite, lateral **D** same, aedeagus, lateral **E** *A. rostrata* Kröber aedeagus, lateral **F** *A. imitans*, gonocoxites, epandrium removed and aedeagus *in situ*, dorsal **G** *A. rostrata*, female spermathecal sac complex, dorsal **H** *A. melanophaeos* sp. n., female head, lateral **I** *A. rostrata*, male head, anterolateral. Abbreviations: **ag** accessory gland **c** cercus **d** distiphallus **da** dorsal apodeme of parameral sheath **ea** ejaculatory apodeme **f** furca **ga** gonocoxal apodeme **gp** (articulated) gonocoxal process **gs** gonostylus **gx** gonocoxite **h** hypandrium **gp** gonocoxal process (articulated) **lea** lateral ejaculatory apodeme **s** spermathecal sac duct **va** ventral apodeme of parameral sheath **v** velutum patch **vl** ventral lobe. Scale lines = 0.2 mm.

with A1 and A2 acanthophorite spines well developed; female sternite 8 emarginate along posterior margin.

Comments. Acupalpa is a genus with some distinctive wasp mimicking species (Figs 1–2), often strikingly coloured with black and orange. The male terminalia are relatively conserved throughout both Acupalpa and Pipinnipons, and species identification is more easily and reliably accomplished using external characters of either sex. Closely related to Pipinnipons and Agapophytus, Acupalpa can be distinguished by the elongate, cylindrical antennae, scape not longer than flagellum, face usually expansive and protruding, and palpi that are acuminate or narrowly cylindrical. The latter two characters specifically differentiate Acupalpa from Pipinnipons, as the face is always narrow and the palpi spatulate in Pipinnipons. Agapophytus is separated from Pipinnipons and Acupalpa by the length of the scape ranging from relatively equal length, to significantly longer than the flagellum.

Included species. A. albimanis (Kröber), comb. n., A. albitarsa Mann, A. boharti sp. n., A. divisa (Walker), A. dolichorhyncha sp. n., A. glossa sp. n., A. imitans (White), comb. n., A. irwini Winterton, A. melanophaeos sp. n., A. miaboolya sp. n., A. minuta sp. n., A. minutoides sp. n., A. notomelas sp. n., A. novayamarna sp. n., A. rostrata Kröber, A. semirufa Mann, A. westralica sp. n., A. yalgoo sp. n., A. yanchep sp. n.

Key to Acupalpa species

1	Abdomen ground colour completely brown to black (terminal segments
	sometimes orange in female) (e.g. Figs 5, 10, 16)2
_	Abdomen with any number of segments 1-3 completely or partially orange
	or yellow (e.g. Figs 7–9, 11–12)
2	Fore and mid coxae brown to black, overlain with silver-grey pubescence (e.g.
	Figs 5–6, 13, 18)
_	Fore and mid coxae orange to pale yellow (Figs 16, 20)
3	Lower pleuron orange; fore femur brown; base of mid and hind femora or-
	ange; foreleg second tarsomere pale (Fig. 16) A. notomelas sp. n.
_	Pleuron uniformly brown to black; fore and mid femora brown to black dor-
	sally, pale ventrally; hind femur brown to black; foreleg second tarsomere
	brown to black (female only) (Fig. 20) A. westralica sp. n.
4	Scape and pedicel yellow; fore and mid femora yellow (Fig. 18)
_	Scape and pedicel brown to black (rarely pale basally); fore and mid femora
	brown to black5
5	Very small sized species (< 3.5 mm total body length); pleuron with longitu-
	dinal stripe of silver velutum; flagellum grossly enlarged (> 3 time combined
	scape and pedicel length) (Fig. 14); two notopleural macrosetae
_	Size variable, but usually larger than 5.0 mm; pleuron colour and vestiture
	variable but never with longitudinal silver velutum stripe; flagellum length

	sub-equal to 0.5 times combined scape and pedicel length; always more than
	two notopleural macrosetae7
6	Coxal macrosetae pale; frons width slightly narrower than ocellar tubercle
	width; wing vein M ₁ and M ₂ originating separately from discal cell; fore tibia
	uniformly brown to black (Fig. 14) (male only)A. minuta sp. n.
_	Coxal macrosetae black; male frons slightly wider than ocellar tubercle width;
	wing vein M ₁ and M ₂ , fused basally and originating as a petiolate stem from dis-
	cal cell; fore tibia yellowish basally (Fig. 15) (male only) A. minutoides sp. n.
7	Fore tibia pale yellow to white, rarely brown distally; male abdomen without
	silver velutum; small species (Figs 6, 13)
_	Fore tibia dark, rarely pale basally; male abdomen with silver velutum; larger
	species (Figs 5, 22)9
8	Scutum overlain with mostly uniform brown-bronze pubescence; fore tibia
	uniform white-cream (Fig. 6)
_	Scutum overlain with grey and brown striped pubescence: fore tibia white-
	cream but darker basally (Fig. 13)
9	Basitarsi white to cream fore-basitarsus darker basally all tarsomeres 3–5
/	dark (Fig 4) <i>A albimanis</i> (Kröber) comb n
_	Basitarsi brown to black tarsomeres 2–5 white
10	Male froms at parrowest point slightly parrower than width of ocellar tubercle
10	(western Australia) (Figs 2, 22)
	(western Australia) (Figs 2, 22)
_	(costorn Australia) (Fig. 5)
11	(eastern Australia) (Fig.)
11	$\mathbf{F}_{\text{remove bases bases to black}} = \left[\mathbf{F}_{\text{remove bases bases bases bases}} + \mathbf{F}_{\text{remove bases}} + \mathbf{F}_{$
10	Femora brown to black (Figs / -0, 11, 21)
12	Coxae pale orange; pleuron dark with orange surfusion; male abdomen with-
	out silver velutum (Figs 12, 1/)13
-	Coxae dark (rarely dark orange), overlain with silver-grey pubescence; pleu-
	ron completely dark, overlain with silver pubescence; male abdomen with or
10	without silver velutum dorsally (Figs 9–10, 19)14
13	Pleuron mostly dark; scutal setae minute; genal setae dark; male frons width
	equal to width of anterior ocellus (Fig. 12)
_	Pleuron mostly orange; scutal setae medium length; genal setae pale; male frons
	width equal to width of ocellar tubercle (Fig. 17)A. novayamarna sp. n.
14	Tibia and tarsi mostly darkened apically; frons profile concave; male abdo-
	men with dense silver velutum dorsally (Fig. 19)
_	Tibia and tarsi without distinctive darkening apically; frons profile rounded;
	male abdomen without dense silver velutum dorsally15
15	Abdominal segments 2–3 bright orange, with orange suffusion of segments 1
	and 4; haltere knob brown (Fig. 9) A. glossa sp. n.
-	Abdominal segments 2-3 slight orange-tan laterally; haltere knob white
	(Fig. 10)
16	(Fig. 10)

_	Tibia yellow to cream basally, dark apically (Figs 11, 21)18
17	Mouthparts greatly elongate, much longer than head length; foreleg basitar-
	sus entirely white to cream (Fig. 8)A. dolichorbyncha sp. n.
_	Mouthparts approximately equal to head length; foreleg basitarsus darker ba-
	sally, pale distally (Fig. 7)
18	Antennal scape and face with short, dark setae; face flat, not protruding ante-
	riorly (male only) (Fig. 21)
_	Antennal scape and face with pale setae; face protruding anteriorly (Fig. 11)

Acupalpa albimanis (Kröber), comb. n.

urn:lsid:zoobank.org:act:911E11F8-66AF-41CF-9B32-8C4B4D744C6A http://species-id.net/wiki/Acupalpa_albimanis Fig. 4

Ectinorhynchus albimanus Kröber 1914: 31. - Irwin and Lyneborg 1989: 356; *nec.* Mann 1928: 156; 1933: 334.

Acupalpa pollinosa Mann 1929: 25; Hardy 1939: 47 [as Acutipalpa polinosa]; Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 235; Winterton et al. 2001: 210. syn. n.

Type material. *Ectinorhynchus albimanus* Kröber, 1914 - **Holotype** female 'N. Holl. [Neu Holland] 878 IV/ TYPE (ANIC29_003432) (NMW).

Acupalpa pollinosa Mann, 1929 - Holotype male, AUSTRALIA: Queensland: Brisbane, 18.ix.1914, H. Hacker (MEI029468) [D3283] (QM). Paratypes: AUSTRALIA: Queensland: 2 males, Brisbane, 24.ix.1914, 24.ix.1923, H. Hacker (MEI108792, 108793) (QM).

Diagnosis. Frons profile concave above antenna; antenna black; wing dark banded; legs black with basitarsus and second tarsomere white; abdomen black, overlain with silver velutum in male.

Redescription. Body length= 6.9–9.3 mm. *Head.* Frons wider than ocellar tubercle, profile transversely concave above antennae, pubescence as two silver patches along eye margin, vestiture as minute setae; frons surface texture as irregular longitudinal striations; face projecting anteriorly, vestiture with dark to pale setae; gena with pale setae; parafacial glabrous; mouthparts relatively short (approximately equal to head length); palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape brown to black, length much shorter than flagellum, with sparse black setae ventrally; flagellum black, base of flagellum with short, dark setae. *Thorax.* Scutum uniform grey-black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings dark banded infuscate; haltere knob white, stem dark brown; coxae and femora brown to black; tibia dark; tarsi dark, basal ³/₄ of fore-basitarsus and entire second



Figure 4. *Acupalpa albimanis* (Kröber), comb. n., (Holotype of *A. pollinosa*), male, anterolateral view [Morphbank: 576222]. Body length = 6.5 mm.

tarsomere cream to white. Scutal chaetotaxy (macrosetae pairs): *np* (notopleural), 4; *sa* (supra-alar), 1; *pa* (post-alar), 1; *dc* (dorsocentral), 2–3; *sc* (scutellar), 1. *Abdomen*. Black, covered with silver velutum dorsally on tergites (male only); terminalia pale.

Comments. *Ectinorhynchus albimanis* is herein transferred to *Acupalpa* with *A. pollinosa* becoming a junior synonym of *Acupalpa albimanis* comb. n. Mann (1928) redescribed *E. albimanis* based on a series of specimens, but clearly did not examine the type, as his redescription does not match that in Kröber (1914) nor reflect characteristics of the type. *Acupalpa albimanis* is morphological similar to *A. albitarsa* and *A. yanchep* sp. n. The colouration of abdomen and tarsi is diagnostic for this species.

Acupalpa albitarsa Mann

urn:lsid:zoobank.org:act:44480644-4D44-460F-9092-7E610A4767F0 Genbank Accession: AF150967 http://species-id.net/wiki/Acupalpa_albitarsa Fig. 5

Acupalpa albitarsa Mann 1929: 24; Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 230; Winterton et al. 2001: 210.

Type material. Holotype male, AUSTRALIA: Queensland: Brisbane, 24.ix.1914, H. Hacker [-27.465, 153.017] (MEI029448) [D3282] (QM). Paratypes: AUSTRAL-

IA: Queensland: 4 males, same data as holotype, (MEI108766, 108768, 108770, 108771) (QM).

Diagnosis. Frons profile rounded above antenna; antenna black, scape sometimes brown; wing irregularly banded; pleuron black; tarsi white with brown to black basitarsus; abdomen with sparse silver velutum on anterior segments (denser in male).

Redescription. Body length= 7.4–9.7 mm. *Head.* Frons wider than ocellar tubercle; profile rounded above antenna, pubescence sparse silver-grey; frontal vestiture as minute setae, texture verrucous; face shape broadly rounded, vestiture with dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts elongate and projecting anteriorly, or relatively short; palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base raised; antenna longer than head; scape brown or black, length approximately equal to flagellum, with sparse black setae; flagellum black, base of flagellum with short, dark setae. *Thorax.* Scutum light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse, silver-grey pubescence; wing markings irregularly banded; haltere knob white; coxae black; femora brown to black; tibia dark, lighter basally; tarsi white with basitarsi dark. Scutal chaetotaxy: *np*, 3–4; *sa*, 1; *pa*, 1; *dc*, 2–3; *sc*, 1. *Abdomen.* Entirely black, segments 5–8 sometimes orange dorsally; silver velutum dorsally on tergites (1–3), bronze medially; terminalia dark (male) or pale (female).

Comments. The distinctive tarsal colouration of *A. albitarsa* and the closely related *A. yanchep* sp. n., separates these species from all other *Acupalpa. Acupalpa albitarsa* is an eastern species while *A. yanchep* sp. n. is western. Females are difficult to distinguish but males differ in the shape of the frons and in general body shape and size. The white patterning of the scutum is less pronounced in this species.

Acupalpa boharti sp. n.

urn:lsid:zoobank.org:act:8F89524D-85C5-4332-86AC-D278AB724C1D http://species-id.net/wiki/Acupalpa_boharti Fig. 6

Type material. Holotype female, AUSTRALIA: Western Australia: Norseman, [-32.167°, 121.75°], 24.xi.1979, R. M. Bohart (MEI029500) (UCDC). Paratypes. AUSTRALIA: Western Australia: male, female, Norseman, [-32.167°, 121.75°], 24.xi.1979, R. M. Bohart (MEI029499, 029501) (UCDC).

Diagnosis. Body size relatively small; frons rounded above antenna; scutum glossy black with bronze pubescence; tibia yellow with dark apices, fore tibia white-cream; abdomen black, without velutum.

Description. Body length= 5.0–6.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded above antenna, glabrous; frontal vestiture as minute setae, texture smooth; face broadly rounded, glabrous; gena with pale setae; parafacial glabrous; mouthparts elongate, projecting anteriorly, or sometimes relatively short; palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base flat; frons



Figure 5. Acupalpa albitarsa Mann, male, anterolateral view [576246]. Body length = 7.0 mm.



Figure 6. Acupalpa boharti sp. n., female, lateral view [576266]. Body length = 5.0 mm.

roughly level with eye in profile; antenna longer than head; scape brown or black, length shorter than flagellum, with sparse black setae; flagellum black or brown, base of flagellum with short dark setae. *Thorax*. Scutum glossy black, overlain with sparse bronze pubescence; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse, silver-grey pubescence; wing markings weakly infuscate with pale band midway, hyaline ocellations basally; haltere knob orange-yellow; coxae black; femora brown to black; tibia yellow, apices dark on mid and hind tibia, fore tibia white-cream; tarsi black; mid and hind basitarsi pale basally. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 4–6; *sc*, 1. *Abdomen*. Black, silver velutum absent, terminalia dark.

Comments. *Acupalpa boharti* sp. n. is a small, dark species similar to *A. miaboolya* sp. n. This species is known only from the type series collected in southwestern Australia. The small body size and leg and body colouration are diagnostic for this species.

Etymology. This species is named in honour of the collector, R. M. Bohart.

Acupalpa divisa (Walker)

urn:lsid:zoobank.org:act:318DBAA4-B88A-4276-897B-643F78FA4AA5 Genbank Accession: AF150966 http://species-id.net/wiki/Acupalpa_divisa Fig. 7

Dimassus divisus Walker 1850: 3.
Ectinorrhynchus divisus (Walker) - Kröber 1913: 19.
Acupalpa semiflava Mann 1929: 28.
Acupalpa divisa (Walker) - Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 232; Winterton et al. 2001: 210.

Type material. *Dimassus divisus* Walker 1850 - **Holotype** female, AUSTRALIA (OUMNH).

Acupalpa semiflava Mann 1929 - Holotype female, AUSTRALIA: Queensland: Brisbane, 24.ix.1914, H. Hacker (MEI029471) (QM). Paratypes: Queensland: 3 females, Brisbane, 24.ix.1912, 14.x.1913, 10.x.1916, H. Hacker (MEI092529, 092532, 092534) (QM).

Additional material. AUSTRALIA: Queensland: male, Barakula State Forest, Hellhole Creek, Auburn Rd., 52062, 13.x.2004, Queensland Museum party, open forest, hand collected [-26.33°, 150.7°] (ANIC29_016460) (QM).

Diagnosis. Frons profile concave above antenna; antenna black; pleuron black; wing dark banded; femora and tibia black; abdomen black, segments 1–3 yellow at least laterally; abdominal velutum present in male.

Redescription. Body length= 6.5–8.0 mm. *Head.* Frons wider than ocellar tubercle, profile transversely concave above antennae, pubescence as silver patches along eye margin, frontal vestiture as minute setae, texture as irregular longitudinal striations; face produced anteriorly, vestiture with dark or pale setae; gena with pale setae; parafacial gla-



Figure 7. Acupalpa divisa (Walker), male, anterolateral view [576245]. Body length = 7.0 mm.

brous; mouthparts elongate, projecting anteriorly, or sometimes relatively short; palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base raised, antennal length approximately equal to head; scape black, length approximately equal to flagellum, with sparse black setae; flagellum black, base of flagellum with short dark setae. *Thorax.* Scutum uniform grey-black, sometimes with faint white stripes; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings dark banded infuscate; haltere knob white; coxae black; femora brown to black; tibia black; tarsi black; fore basitarsus white distally, 2nd tarsomere basally, remaining basitarsi yellowish. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 3–4; *sc*, 1. *Abdomen.* Segments 2–3 yellow, remaining segments black (male tergites 1-3 dark medially), silver velutum dorsally on tergites (male) or absent (female); terminalia dark.

Comments. The male of *A. divisa* has long been unknown, and herein described for the first time. Hardy (1939) proposed that this species was a synonym of *A. pollinosa*, but has been subsequently proved incorrect as corresponding sexes of both species are now known.

Acupalpa dolichorhyncha sp. n.

urn:lsid:zoobank.org:act:56FF95E1-D936-4DBA-8EBF-3A6E79B24116 http://species-id.net/wiki/Acupalpa_dolichorhyncha Fig. 8

Type material. Holotype male, AUSTRALIA: **Western Australia**: 11 km N Cataby, 29.x.1987, M. E. Irwin & E. I. Schlinger, sweeping *Leptospernum* flowers [-30.733°,



Figure 8. Acupalpa dolichorhyncha sp. n., male, anterolateral view [576248]. Body length = 8.0 mm.

115.533°] (MEI029507) (ANIC). **Paratype.** AUSTRALIA: **Western Australia:** female, same data as holotype (MEI029506) (ANIC).

Diagnosis. Mouthparts elongate; frons profile rounded above antenna; antenna black; scutum glossy black; pleuron black; wing dark banded; abdominal segments 1–3 orange, rest black; abdominal velutum absent.

Description. Body length= 7–10 mm. *Head.* Frons wider than ocellar tubercle (male), profile rounded above antenna, pubescence absent, glabrous, frontal vestiture glabrous or as minute setae, surface texture smooth; face shape broadly rounded, expansive, vestiture with dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts elongate, projecting anteriorly; palpus brown-black, narrowly cylindrical; occiput overlain with sparse, silver-grey pubescence; antennal base flat, frons roughly level with eye in profile; antennal length approximately equal to head; scape black with sparse black setae, length shorter than flagellum; flagellum black, base of flagellum with short dark setae. *Thorax.* Scutum glossy black, overlain with faint stripes of grey pubescence; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings faintly banded infuscate; haltere knob white; coxae black; femora brown to black; tibia brown or black; tarsi dark, basitarsi pale, dark distally, fore-basitarsus entirely white. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen.* Segments 1-3 orange, remaining segments black, silver velutum absent; terminalia dark.

Comments. Acupalpa dolichorhyncha sp. n. is a distinctive species with elongate mouthparts and orange-banded abdominal segments 1–3. This western species is morphologically similar to *A. melanophaeos* sp. n., also from Western Australia, and *A. glossa* sp. n. from Victoria.

Etymology. The specific epithet is derived from Gr. *dolichos*, long; *rhynchus*, snout, referring to the elongate mouthparts.

Acupalpa glossa sp. n.

urn:lsid:zoobank.org:act:60BC378C-1247-4CB5-A194-755C00BC5567 http://species-id.net/wiki/Acupalpa_glossa Fig. 9

Type material. Holotype male, AUSTRALIA: **Victoria:** 5 km S Rocket Lake, Murray-Sunset N.P., 34.39°S, 141.49°E, 25.xi.1992, swept, McEvey, Moulds, McAlpine (MEI165183) (AMS). **Paratypes.** AUSTRALIA: **Victoria:** male, 2 females, Murray-Sunset N.P., Millewa South Bore track, 20.7 km S Shearers Quarters, 17–23.xi.2002, C. Lambkin, D. Yeates, N. Starick, J. Recsei, 34°45'02"S, 141°03'56"E [Malaise trap] (MEI165184, 165185, 165186) (ANIC).

Diagnosis. Frons profile rounded above antenna; antenna black; scutum glossy black; pleuron orange to brown, darker posteriorly; wing banded infuscate; femora orange; tibia black; abdomen black, without velutum.

Description. Body length= 7.0–9.0 mm. *Head*. Frons wider than ocellar tubercle, profile rounded above antenna, glabrous, sometimes with silver patches of pubescence along eye margin, frontal vestiture as minute setae, surface texture smooth; face broad-



Figure 9. Acupalpa glossa sp. n., male, anterolateral view [576249]. Body length = 8.0 mm.

ly rounded, vestiture as dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts elongate, projecting anteriorly; palpus brown-black, narrowly cylindrical; occiput overlain with sparse, silver-grey pubescence; antennal base flat; antennal length approximately equal to head; scape brown, shorter than flagellum, with sparse black setae; flagellum black, base of flagellum with short dark setae. *Thorax*. Scutum black, overlain with grey pubescence; scutellum overlain with dense, matt-black pubescence; pleuron dark, overlain with sparse silver-grey pubescence, denser anteriorly and posteriorly, sparse around midway; wing banded infuscate; haltere knob white; coxae dark, overlain with dense silver pubescence; femora orange; tibia orange, darker distally; fore basitarsus white, 2nd tarsomere white basally, remaining basitarsi cream, darker distally. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen*. Segments 2–3 orange, rest black, intersegmental membranes white on segments 2–3, silver velutum absent; terminalia dark.

Comments. Acupalpa glossa sp. n. is similar to A. dolichorhyncha sp. n. in colour pattern and elongated mouthparts, but is easily differentiated based on leg colour. This species is known only from the type series collected in Victoria.

Etymology. The specific epithet is derived from Gr. *glossa*, tongue, referring to the elongate mouthparts.

Acupalpa imitans (White), comb. n.

urn:lsid:zoobank.org:act:749A106A-0357-4AAF-A88B-E2E46ECA7B9A http://species-id.net/wiki/Acupalpa_imitans Figs 3C–D, F, 10

Phycus imitans White 1915: 28.

Agapophytus imitans (White 1915: 28) - Mann 1929: 40; Irwin and Lyneborg 1989: 354 [catalogue].

Pipinnipons imitans (White 1915: 28) -Winterton et al. 2001: 211.

Acupalpa imitans (White 1915: 28), comb. n.

Type material. Type female, AUSTRALIA: **Tasmania:** Wedge Bay, 3.i.1914, G.H. Hardy [lost].

Neotype male, AUSTRALIA: **Queensland:** Indooroopilly, Long Pocket [-27.418°, 152.837°], 22.viii–7.ix.2007, S. L. Winterton, Malaise trap (MEI165187) (QM).

Other material examined. AUSTRALIA: **Queensland:** female, Brisbane Forest Park, Scrub Road, crossing at Enoggera Creek, [-27.428°, 152.843°], 200m, 10–14.xi.1995, malaise trap, M.E. Irwin. (MEI140857) (QM); male, Tambourine, [-27.88, 153.13], 12.vi.1925 ("Allotype" of Mann 1929) (MEI023602) (QM); female, Mount Tamborine, [-27.917°, 153.15°], 29.xi.1925, hand netted, H. Hacker (MEI108898) (QM).

Diagnosis. Frons profile rounded above antenna; antenna dark yellow; pleuron black; wing dark banded; femora orange to yellow; tibia yellow; abdomen dark, segments 2–3 red-brown laterally, without silver velutum.



Figure 10. Acupalpa imitans (White), male, anterolateral view [576250]. Body length = 7.0 mm.

Redescription. Body length= 6.3–7.0 mm. *Head*. Frons wider than ocellar tubercle (female) or narrower (male), profile rounded above antenna, pubescence as silver patches along eye margin, frontal vestiture glabrous, surface texture as irregular longitudinal striations or smooth; face as narrow strip below antennal base, vestiture glabrous; gena with pale setae; parafacial glabrous; mouthparts relatively short (approximately equal to head length); palpus brown-black, narrowly cylindrical; occiput glabrous, glossy black; antennal base flat, frons roughly level with eye in profile (or near so); antenna longer than head; scape dark yellow, length approximately equal to flagellum, scape with sparse black setae; flagellum dark yellow, base of flagellum with short dark setae. Thorax. Scutum black, overlain with grey pubescence, brown stripes of pubescence more expansive posteriorly; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings banded infuscate; haltere knob white; coxae black; femora orange or yellow; tibia yellow, apices sometimes dark; tarsi yellow, fore-basitarsus white. Scutal chaetotaxy: np 4-5; sa, 1; pa, 1; dc, 2; sc, 1. Abdomen. Dark, segments 2-3 red-brown medially, orange laterally, silver velutum absent (female) or small triangular patches on tergites 2–3 (male); terminalia pale.

Comments. The type of *Phycus imitans* was kept in the G.H. Hardy collection, which was moved from Brisbane to Katoomba, but is now apparently lost or destroyed by pests. While not stating whether he actually examined the type, Mann (1929) redescribed this species in *Agapophytus* based on an unjustified allotype designation and associated female from southeast Queensland. Winterton et al. (2001) transferred the

referred to by Mann (1929). The two specimens examined by Mann (1929) as putative *A. imitans* were located in the QM collection and no further material has been collected from Tasmania. Based on the discovery of new material from southeast Queensland more closely matching the original description by White (1915) than any other material examined (including the allotype designated by Mann (1929)), a male specimen (MEI165187) is herein designated as a Neotype to stabilise the concept of the species. This problematic species has characteristics that indicate a closer relationship to species of *Acupalpa* (particularly palpi shape) and is herein transferred from *Pipinnipons*.

Acupalpa irwini Winterton

urn:lsid:zoobank.org:act:6C3A49B8-6E77-4C55-B5D9-C77636280ECB http://species-id.net/wiki/Acupalpa_irwini Fig. 11

Acupalpa irwini Winterton, 2000: 232.

Type material. Holotype female, AUSTRALIA: **Western Australia:** 7.5 km WSW Lake Cronin, 32°23'S, 119°46'E, 19–26.ix.1978, T. F. Houston et al. (MEI029876) (WAM). **Paratypes.** AUSTRALIA: **Western Australia:** female, same data as holotype, (MEI029877) (WAM); male, 3 females, 53 km E Hyden nr. Emu Rock, 24–27.x.1985, R. W. Thorpe (MEI029502, 029503, 029504, 029505) (UCDC).



Figure 11. Acupalpa irwini Winterton, female, anterolateral view [576251]. Body length = 9.0 mm.

Diagnosis. Frons profile rounded above antenna; face projecting anteriorly; antenna black; scutum grey to black; pleuron black; wing dark banded; femora brown to black; abdomen black, segments 1–3 orange; abdominal velutum absent.

Redescription. Body length= 7.0–10.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded above antenna, pubescence sparse silver-grey, frontal vestiture small dark setae, surface texture verrucous; face projecting anteriorly, vestiture with dark or pale setae; gena with pale setae; parafacial glabrous or with pale setae; mouth-parts elongate, projecting anteriorly; palpus brown-black, narrowly cylindrical; occiput overlain with sparse, silver-grey pubescence; antennal base raised; antennal length approximately equal to head; scape black, length approximately equal to flagellum, scape with pale setae ventrally, shorter dark setae dorsally; flagellum black, base of flagellum with short dark setae. *Thorax*. Scutum light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with silver-grey pubescence; wing markings dark banded infuscate; haltere knob brown; coxae black; femora brown to black; tibia yellow-orange, darker distally; tarsi black, basitarsi pale, dark distally. Scutal chaetotaxy: np, 4; sa, 1; pa, 1; dc, 2–3; sc, 1. *Abdomen*. Segments 1–3 orange, remaining segments black, silver velutum absent; terminalia dark.

Comments. *Acupalpa irwini* is a relatively large species differentiated by the characteristic leg and abdomen colouration. This species is known only from the type series collected in Western Australia.

Acupalpa melanophaeos sp. n.

urn:lsid:zoobank.org:act:32E2D131-5108-46B5-B330-ABEAAC192948 http://species-id.net/wiki/Acupalpa_melanophaeos Figs 3H, 12

Type material. Holotype female, AUSTRALIA: **Western Australia:** Drummond Cove, Geraldton, 16.xi.1973, N. McFarland [-28.767°, 114.617°] (MEI029498) (ANIC). **Paratypes.** AUSTRALIA: **Western Australia:** female, same data as holo-type (MEI029496) (WAM); male, Bunbury, 3.i.1957, A. Snell [-33.317°, 115.633°] (MEI029509) (ANIC); female, Cape Le Grand Nat. Park [-33.96°, 122.12°], 12.i.1987, G. & A. Daniels (MEI029494) (GDCB/AMS).

Diagnosis. Frons profile rounded above antenna; scape yellow, flagellum dark; scutum grey to black; pleuron black (reddish posteriorly in female); wing banded infuscate; coxae orange; legs orange to yellow, tarsi dark distally and fore-basitarsus white; abdomen black, segments 1–3 orange to yellow; abdominal velutum absent.

Description. Body length= 7.1–9.6 mm. *Head.* Frons wider than ocellar tubercle (female) or narrower (male), profile rounded above antenna, pubescence absent, frontal vestiture glabrous or with minute setae, surface texture smooth; face shape broadly rounded, expansive, vestiture glabrous; gena with dark setae; parafacial glabrous; mouth-parts elongate, projecting anteriorly, or sometimes relatively short; palpus brown-black, narrowly cylindrical; occiput glabrous, glossy black; antennal base raised (male) or flat,



Figure 12. Acupalpa melanophaeos sp. n., female, anterolateral view [576252]. Body length = 9.0 mm.

frons roughly level with eye in profile (female); antenna longer than head; scape yellow, length much shorter than flagellum, scape with short, black setae; flagellum black, base of flagellum with short, dark setae. *Thorax.* Scutum light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron black (male) or darker anteriorly with dark orange posteroventrally (female), overlain with sparse silvergrey pubescence; wing markings banded infuscate; haltere knob brown; coxae orange; femora orange or dark yellow; tibia orange; tarsi yellow orange, dark distally, fore-basitarsus white. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 1–2; *sc*, 1. *Abdomen*. Segments 1-3 yellow or orange, remaining segments black, silver velutum absent; terminalia dark.

Comments. Acupalpa melanophaeos sp. n. is a relatively large species from Western Australia with distinctive leg and abdomen colouration. It is morphologically similar to *A. novayamarna* sp. n. and *A. notomelas* sp. n. The coxae are pale in this species along with a rounded face, rather than protruding anteriorly in similar species (e.g. *A. irwini*).

Etymology. The specific epithet is derived from the Gr. *melanos*, black; *phaeos*, light, shiny, referring to the scutum colouration.

Acupalpa miaboolya sp. n.

urn:lsid:zoobank.org:act:31A86965-E8AB-4D6C-9252-FE42BAC3269A http://species-id.net/wiki/Acupalpa_miaboolya Fig. 13

Type material. Holotype male, AUSTRALIA: **Western Australia:** 14.5 km N Carnarvon, Miaboolya Beach, [-24.76°, 113.65°], 4.x.1969, H. E. Evans, R. W. Mat-



Figure 13. Acupalpa miaboolya sp. n., male, anterolateral view [576253]. Body length = 5.0 mm.

thews. (MEI080305) (ANIC). **Paratype.** AUSTRALIA: **Western Australia:** female, same data as holotype (MEI080301) (MCZ).

Diagnosis. Frons profile rounded above antenna; antenna brown to black; scutum glossy black with pubescent stripes of grey and brown; pleuron black; wing faintly infuscate; femora and tibia dark, fore tibia pale distally; abdomen black, without silver velutum.

Description. Body length= 5.0–6.5 mm. *Head.* Frons wider than ocellar tubercle, profile rounded above antenna, pubescence sparse silver-grey, without setae, surface texture smooth; face broadly rounded, glabrous; gena with pale setae (female) or dark setae (male); parafacial glabrous; mouthparts relatively short (approximately equal to head length), or elongate and projecting anteriorly; palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base flat; frons roughly level with eye in profile; antennal length approximately equal to head; scape light brown to black, length approximately equal to flagellum, scape with sparse black setae; flagellum black, base of flagellum with short, dark setae. Thorax. Scutum black, overlain with stripes of grey and brown pubescence; scutellum overlain with sparse grey pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing largely hyaline, faint band midway (male) or infuscate with pale band midway, hyaline ocellations basally (female); haltere knob white; coxae black, overlain with silver pubescence; femora brown to black; tibia brown; fore tibia pale distally; tarsi black, mid and hind basitarsi pale basally. Scutal chaetotaxy: np, 3; sa, 1; pa, 1; dc, 3; sc, 1. Abdomen. Black, overlain with bronze pubescence, silver velutum absent; terminalia dark.

Comments. *Acupalpa miaboolya* sp. n. is a relatively small, dark species from Western Australia very similar to *A. boharti* sp. n. This species can be differentiated from the latter based on scutal pattern; the scutum has grey and brown stripes in *A. miaboolya* sp. n. while the scutum of *A. boharti* sp. n. has more uniform brown-bronze pubescence.

Etymology. This species is named after the region in which the specimens were collected, Miaboolya beach, on the north-central coast of Western Australia.

Acupalpa minuta sp. n.

urn:lsid:zoobank.org:act:C2B0BF78-A250-4F9E-B23C-44533147EFA5 http://species-id.net/wiki/Acupalpa_minuta Fig. 14

Type material. Holotype male, AUSTRALIA: **Western Australia:** Kalbarri, [-27.717, 114.167], 23.ix.1974, N. McFarland (MEI021410) (ANIC).

Diagnosis. Very small sized species; setae on coxae pale; flagellum greatly elongate; scape relatively short; frons equal to width of ocellar tubercle; tibia dark; two notopleural setae; abdomen dark, velutum absent.

Description. Body length= 3.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded, level with eye, pubescence sparse silver-grey; frontal vestiture glabrous, texture smooth; lower frons and face broadly rounded, expansive; face vestiture glabrous; gena with pale setae; parafacia overlain with silver pubescence; mouthparts elongate, project-ing anteriorly; palpus brown-black; occiput overlain with sparse, silver-grey pubescence; antennal base flat; antennal length longer than head; scape colour black, length much



Figure 14. Acupalpa minuta sp. n., male, lateral view [581505]. Body length = 3.0 mm.

shorter than flagellum, with sparse black setae; flagellum colour brown, base of flagellum without setae. *Thorax.* Scutum glossy black-brown with sparse grey pubescence and small brown setae; scutellum overlain with dense matt black pubescence; pleuron glossy black-brown with longitudinal stripe of silver velutum; wing markings dark banded infuscate; haltere knob orange-yellow; coxae brown, overlain with dense pubescence and pale setae; femora brown to black; tibia brown; tarsi brown. Scutal chaetotaxy (macrosetae pairs): *np*, 2; *sa*, 1; *pa*, 1; *dc*, 3, *sc*, 1. *Abdomen.* Colouration brown, tergites 2-4 with bronze pubescence, silver velutum absent; terminalia dark.

Comments. Acupalpa minuta sp. n. is closely related to A. minutoides sp. n., sharing characteristics such as very small size, two notopleural setae and an antenna with a short scape and a greatly elongate flagellum. It can be differentiated based on the colour of the setae on the coxae and by the width of the frons. This species is known only from a single male individual from Western Australia.

Etymology. The specific epithet is derived from the L. *minutus*, small, little, referring to the diminutive body size.

Acupalpa minutoides sp. n.

urn:lsid:zoobank.org:act:2429322A-907F-4330-8BF5-0F1C34FBFAD7 http://species-id.net/wiki/Acupalpa_minutoides Fig. 15

Type material. Holotype male, AUSTRALIA: Western Australia: Geraldton, Drummond's Cove, [-28.767, 114.617], 29.ix.1972, N. McFarland (MEI021412) (ANIC). Paratypes. Western Australia: male, Geraldton, Drummond's Cove, [-28.767, 114.617], 29.ix.1972, N. McFarland, on *Calandrinia* flowers (MEI029995) (CSCA); male, Geraldton, Drummond's Cove, [-28.767, 114.617], 18.ix.1972, N. McFarland, on *Calandrinia* flowers (MEI021411) (CSCA).

Diagnosis. Very small sized species; setae on coxae black; flagellum greatly elongate; scape relatively short; frons slightly wider than width of ocellar tubercle; wing veins M_1 and M_2 fused and petiolate basally from discal cell; tibia pale basally; two notopleural setae; abdomen dark, velutum absent.

Description. Body length= 3.0–4.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded, level with eye, pubescence sparse silver-grey; frontal vestiture glabrous with minute setae laterally, texture smooth; lower frons and face shape broadly rounded, expansive; face vestiture glabrous; gena with pale setae; parafacia overlain with silver pubescence; mouthparts elongate, projecting anteriorly; palpus brown-black; occiput overlain with sparse, silver-grey pubescence; antennal base flat; antennal length longer than head; scape black, length much shorter than flagellum, with sparse black setae; flagellum black or brown, base of flagellum without setae. *Thorax.* Scutum glossy black-brown with sparse grey pubescence and small brown setae; scutellum overlain with dense matt black pubescence; pleuron glossy black-brown with longitudinal stripe of silver velutum; wing markings dark banded infuscate; haltere knob orange-yellow; coxae brown, overlain with dense pubes-



Figure 15. Acupalpa minutoides sp. n., male, lateral view [581506]. Body length = 3.5 mm.

cence and dark setae; femora brown to black; tibia black, yellow-orange basally; tarsi brown. Scutal chaetotaxy (macrosetae pairs): *np*, 2, *sa*, 1, *pa*, 1, *dc*, 2, *sc*, 1. *Abdomen*. Colouration brown, tergites 2-4 with bronze pubescence, silver velutum absent; terminalia dark.

Comments. See comments under *A. minuta* sp. n. This species is known only from three male specimens from Western Australia.

Etymology. The specific epithet is derived from the L. *minutus*, small, little; *-oides*, like, referring to the similarity of this species to *A. minuta* sp. n.

Acupalpa notomelas sp. n.

urn:lsid:zoobank.org:act:68193274-CA84-43A8-A35A-6E819DE50B46 http://species-id.net/wiki/Acupalpa_notomelas Figs 3A–B, 16

Type material. Holotype male, AUSTRALIA: **Western Australia:** 22 km W Waroora Homestead [-23.483°, 113.8°], 25.x.1987, sand plain, M. E. Irwin (MEI029510) (ANIC). **Paratypes.** AUSTRALIA: **Western Australia:** female, Melaleuca Park, 38 km N Perth [-31.95°, 115.85°], 29.x.1987, M. E. Irwin, E. I. Schlinger (MEI029512) (ANIC); male, Yanchep National Park [-31.517°, 115.683°], 22–26.x.1985, truck trap, A. Dyce, W. Wirth (MEI029514) (ANIC).

Diagnosis. Frons profile rounded above antenna; mouthparts elongate; antenna dark; scutum dark; pleuron orange ventrally; wing banded; legs dark yellow to orange [fore femur darker]; abdomen black without silver velutum.



Figure 16. Acupalpa notomelas sp. n., male, anterolateral view [576254]. Body length = 6.0 mm.

Description. Body length= 6.2–8.0 mm. *Head*. Frons wider than ocellar tubercle, profile rounded above antenna, glabrous or silver pubescent patches along eye margin [some sparse pubescence dorsally], frontal vestiture as small dark setae, surface texture smooth, face broadly rounded, vestiture as dark or pale setae; gena with pale setae or with dark setae (ventrally); parafacial glabrous; mouthparts elongate, projecting anteriorly; palpus brown-black, narrowly cylindrical; occiput overlain with sparse, silver-grey pubescence, antennal base flat; antennal length approximately equal to head; scape brown, length shorter than flagellum, scape with sparse black setae; flagellum dark, base of flagellum without setae. Thorax. Scutum light grey to glossy black, setal bases glossy black, overlain with faint stripes of grey pubescence; scutellum overlain with dense, matt-black pubescence; pleuron yellow-orange in lower 2/3, upper 1/3 concolourous with scutum, overlain with sparse silver-grey pubescence; wing markings banded infuscate; haltere knob white, dark basally; coxae yellow-orange; femora with darker fore femur, rest yellow-orange; tibia and tarsi dark yellow; fore-basitarsus white, darker basally, 2nd tarsomere white basally, remaining basitarsi yellow-brown. Scutal chaetotaxy: np, 4; sa, 1; pa, 1; dc, 2; sc, 1. Abdomen. black, silver velutum absent; terminalia pale.

Comments. Acupalpa notomelas sp. n. is similar to A. melanophaeos sp. n. and A. novayamarna sp. n., but can be differentiated based on abdomen colouration. The palpi are very slender and elongate.

Etymology. The specific epithet is derived from the Gr. *noto*, back; *melas* black, dark, referring to the scutal colouration.

Acupalpa novayamarna sp. n.

urn:lsid:zoobank.org:act:C0BEA372-6953-43E4-B957-0A521FD8D83B http://species-id.net/wiki/Acupalpa_novayamarna Fig. 17

Type material. Holotype male, AUSTRALIA: **Western Australia:** 25 km E New Yamarna Homestead, [-28.167°, 123.683°], 21.ix.1982, T. F. Houston, B. Hanich (WAM872128) (WAM).

Diagnosis. Frons profile rounded to slightly concave above antenna; mouthparts short; antenna dark; scutum dark; pleuron dark dorsally, orange ventrally; wing banded; legs orange, tarsi dark distally, fore-basitarsus white; abdomen orange, segments 1–2 black medially, velutum absent.

Description. Body length= 7.2 mm. *Head.* Frons wider than ocellar tubercle, profile rounded to slightly concave above antenna, pubescence absent or as silver patches along eye margin, frontal vestiture glabrous, surface texture smooth, face as narrow strip below antennal base, glabrous; gena with pale setae; parafacial overlain with silver pubescence; mouthparts relatively short (approximately equal to head length); palpus brown-black, narrowly cylindrical; occiput glabrous, glossy black; antennal base raised, antennal length approximately equal to head; scape orange-brown, length much shorter than flagellum, scape with sparse black setae; flagellum black, base of flagellum with short dark setae. *Thorax.* Scutum light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron orange, upper 1/3 concolourous with scutum, overlain with sparse silver-grey pubescence; wing markings banded infuscate, dark yellow basally;



Figure 17. Acupalpa novayamarna sp. n., male, anterolateral view [576255]. Body length = 7.0 mm.

haltere knob orange-yellow; coxae orange; femora orange; tibia orange; tarsi yellow orange, dark distally, fore-basitarsus white. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen*. Orange, segments 1–2 black medially, silver velutum absent; terminalia pale.

Comments. This species is very similar to *A. melanophaeos* sp. n., but can be differentiated by the pale genal setae, larger proportion of the pleuron being orange, and male frons slightly wider. Only the male is known for this western species.

Etymology. This species is named after the type locality of New Yamarna Homestead, Western Australia.

Acupalpa rostrata Kröber

urn:lsid:zoobank.org:act:4FFB52C9-C88E-426B-BB46-B2CF1EC04C9F http://species-id.net/wiki/Acupalpa_rostrata Figs 3E, G, I, 18

Acupalpa rostrata Kröber 1912: 152; Kröber 1913: 18; Mann 1929: 26; Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 235; Winterton et al. 2001: 210.

Type material. Holotype male, AUSTRALIA: **New South Wales**, Sydney (ZMUH) [destroyed]. **Neotype** male, AUSTRALIA: **New South Wales**: Kosciusko National Park, Round Mountain, Olgives Creek, [-35.682°, 149.533°] 1400m, 28.xii.1977, E. I. Schlinger. (MEI029931) (ANIC).

Diagnosis. Frons profile concave above antenna; scape and pedicel yellow-orange, flagellum black; scutum black, overlain with silver pubescence; pleuron black with silver pubescence; wing banded; femora yellow-orange [hind femur dark]; tibia yellow-orange; abdomen black, overlain with silver velutum in male.

Redescription. Body length= 7.0–8.0 mm. *Head.* Frons wider than ocellar tubercle, profile transversely concave above antennae, pubescence as silver patches along eye margin, frontal vestiture as numerous elongate setae, surface texture smooth; face projecting anteriorly, vestiture as dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts relatively short (approximately equal to head length); palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape and pedicel yellow-orange, length approximately equal to flagellum, with pale setae ventrally, shorter dark setae dorsally; flagellum black, base of flagellum with short dark setae. *Thorax.* Scutum uniform grey-black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with silver pubescence; wing markings banded infuscate; haltere knob white, dark basally; coxae black; femora yellow with hind femur dark; tibia yellow-orange, apices dark; tarsi yellow-orange, distal segments darker, basitarsus and second tarsomere on foreleg white. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen.* Black, silver velutum dorsally on tergites (male) or absent (female); terminalia pale.

Comments. The type of *A. rostrata* is apparently destroyed. Winterton (2000) redescribed this distinctive species, without designating a neotype as the species was still identifiable based on the original description alone. With the description of the new



Figure 18. Acupalpa rostrata Kröber, Neotype male, anterolateral view [576256]. Body length = 7.0 mm.

species herein a neotype is designated to stabilise the taxon and remove any possibility of confusion in the future. *Acupalpa rostrata* is differentiated from other *Acupalpa* species by the unique leg and antennal colouration.

Acupalpa semirufa Mann

urn:lsid:zoobank.org:act:CF947CED-BCB0-4A62-AFDF-433DDBF3E1D3 http://species-id.net/wiki/Acupalpa_semirufa Fig. 19

Acupalpa semirufa Mann 1929: 27; Irwin and Lyneborg 1989: 354 [catalogue]; Winterton 2000: 237; Winterton et al. 2001: 210.

Type material. Holotype male, AUSTRALIA: **New South Wales:** Blackheath, Hardy [not examined-location unknown]. **Paratypes.** AUSTRALIA: **Queensland:** 'Allotype' female, Bribie Island, 12.ix.1918, H. Hacker (MEI029439) (QM). **New South Wales:** 2 females, Sydney, Manly, 20.xi.1923 (MEI108786, 108787) (QM).

Diagnosis. Frons profile concave above antenna; antenna dark; scutum dark; pleuron black; wing dark banded; femora orange to yellow; tibia yellow, darker distally; abdomen black with segments 2–3 yellow-orange, overlain with silver velutum in male.

Redescription. Body length= 6.0–9.0 mm. *Head*. Frons wider than ocellar tubercle, profile transversely concave above antennae, pubescence as silver patches



Figure 19. Acupalpa semirufa Mann, male, anterolateral view [576257]. Body length = 7.0 mm.

along eye margin, sparse silver-grey dorsally, frontal vestiture as small dark setae, surface texture vertucous; face projecting anteriorly with dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts short (approximately equal to head length), or elongate, projecting anteriorly; palpus brown-black, acuminate; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape brown, length shorter than flagellum, scape with pale setae ventrally, shorter dark setae dorsally; flagellum black, base of flagellum with short dark setae. Thorax. Scutum uniform grey-black or light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings dark, banded infuscate; haltere knob white, dark basally; coxae black; femora orange or yellow; tibia yellow-orange, darker distally; basitarsi yellow-orange, rest black, fore-basitarsus white distally, 2nd tarsomere basally. Scutal chaetotaxy: np, 4; sa, 1; pa, 1; dc, 3; sc, 1. Abdomen. Segment 2 in male, or segments 2-3 in female orange with black medial patch, rest of segments black, silver velutum dorsally on tergites (male) or absent (female); terminalia pale.

Comments. Acupalpa semirufa is a common species in south-eastern Queensland and northern New South Wales. It is similar to *A. divisa* and *A. yalgoo* sp. n., but can be differentiated easily from the former by the orange leg colour (black in *A. divisa*) and from the latter by the projecting face (rounded in *A. yalgoo* sp. n.) and two wing bands (single band in *A. yalgoo* sp. n.).

Acupalpa westralica sp. n.

urn:lsid:zoobank.org:act:C2BCA147-940D-42D1-AB6C-0A106BF6709A http://species-id.net/wiki/Acupalpa_westralica Fig. 20

Type material. Holotype female, AUSTRALIA: **Western Australia:** Stirling Ranges N.P., Chester Pass Rd., Eucalyptus open woodland, 230m; C. Lambkin, J. Recsei, 3–15.xi.2003; Malaise, ANIC bulk sample 2191 [-34.433°, 118.076°] (MEI165188) (ANIC).

Diagnosis. Frons profile rounded above antenna; scape yellow-brown, flagellum black; scutum grey-black with dark and pale stripes; pleuron black; wing dark banded; femora yellow with extensive dark suffusion dorsally [hind femur dark]; tibia brown; abdomen black with brown pubescence, silver velutum absent.

Description. Body length= 8.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded above antenna, glabrous or with minute setae, surface texture smooth; face broadly rounded, expansive, with dark or pale setae; gena with pale setae; parafacial glabrous; mouthparts elongate, projecting anteriorly; palpus brown-black, narrowly cylindrical; occiput glabrous, glossy black; antennal base flat; frons roughly level with eye in profile; antennal length approximately equal to head; scape yellow-brown, shorter than flagellum, scape with sparse black setae; flagellum black, base of flagellum without setae. *Thorax.* Scutum uniform grey-black with diffuse brown and cream stripes; scutellum



Figure 20. Acupalpa westralica sp. n., female, anterolateral view [576258]. Body length = 8.0 mm.

overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silvergrey pubescence; wing markings dark banded infuscate; haltere knob white, dark basally; coxae yellow; hind femur dark, rest yellow with extensive dark suffusion dorsally; tibia and tarsi brown, fore-basitarsus white. Scutal chaetotaxy: *np*, 4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. Abdomen. Black with brown pubescence, silver velutum absent; terminalia dark.

Comments. *Acupalpa westralica* sp. n. is known only from the holotype female from southern Western Australia. This species superficially resembles *A. rostrata* in colouration, but the head shape suggests a close relationship to *A. notomelas* sp. n.

Etymology. The specific epithet is derived from the western distribution of this species.

Acupalpa yalgoo sp. n.

urn:lsid:zoobank.org:act:AA49E372-62DF-4607-9C4E-77CCE53D9CC4 http://species-id.net/wiki/Acupalpa_yalgoo Fig. 21

Type material. Holotype male, AUSTRALIA: Western Australia, 28 km W Yalgoo, [-28.35°, 116.683°], 2.ix.1981, G. A. Holloway (MEI029508) (AM). Paratype. AUS-TRALIA: Western Australia: male, Great Victoria Desert, Officer Basin, NE Streich Mound, 24–28.ix.1991, McMillan (MEI165193) (WAM).

Diagnosis. Frons profile rounded above antenna; antenna black; scutum grey to black; pleuron black; wing dark banded; femora black; tibia yellow in basal half; abdomen black, segments 1–3 orange, silver velutum absent.

Description. Body length= 9.0-10.0 mm. *Head.* Frons wider than ocellar tubercle, profile rounded above antenna, pubescence sparse silver-grey, frontal vestiture as small dark setae, surface texture as irregular longitudinal striations, face broadly rounded, expansive, with dark or pale setae; gena with pale setae; parafacial with short setae towards gena; mouthparts elongate; palpus brown-black, narrowly cylindrical; occiput overlain with sparse, silver-grey pubescence; antennal base flat; frons roughly level with eye in profile; antennal length shorter than head; scape black, length approximately equal to flagellum, scape with sparse black setae; flagellum black. *Thorax.* Scutum light grey to black, setal bases glossy black; scutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings dark banded [discal band broad], dark yellow basally; haltere knob orange-yellow; coxae black; femora brown to black; tibia yellow in basal 1/2, dark in distal 1/2; tarsi black, basitarsi pale, dark distally, rest of tarsomeres dark. Scutal chaetotaxy: *np*, 4–5; *sa*, 1; *pa*, 1; *dc*, 3; *sc*, 2. *Abdomen.* Segments 1–3 orange, remaining segments black, silver velutum absent; terminalia dark.

Comments. *Acupalpa yalgoo* sp. n. is a western species similar to *A. semirufa* and *A. glossa* sp. n., but differs from both in leg colouration and wing patterning.

Etymology. The specific epithet is derived from the type locality of this species, near the Western Australian township of Yalgoo.



Figure 21. Acupalpa yalgoo sp. n., male, anterolateral view [576259]. Body length = 8.0 mm.

Acupalpa yanchep sp. n.

urn:lsid:zoobank.org:act:A3312038-6598-4C6A-BBA4-09CFF0E002F0 http://species-id.net/wiki/Acupalpa_yanchep Figs 2, 22

Type material. Holotype male, AUSTRALIA: **Western Australia:** Yanchep N.P., Malaise trap, [-31.576°, 115.693°] 18–29.xii.1999; J. & A. Skevington, C. Lambkin, P. Bouchard (MEI165189) (ANIC). **Paratypes.** AUSTRALIA: **Western Australia:** male, female, same data as holotype (MEI165190, 165191) (ANIC); female, Yanchep [-31.525°, 115.626°], 21.xi.2008, fore-dune, S. L. Winterton & S. D. Gaimari (MEI165192) (QM).

Diagnosis. Frons profile rounded above antenna; antenna dark; scutum black with irregular brown and white pubescent markings; pleuron black; wing irregularly banded; legs dark, tibia pale basally; abdomen black [sometimes orange apically in female], silver velutum present in male.

Description. Body length= 6.0–10.0 mm. *Head.* Frons wider than ocellar tubercle (female) or narrower (male), profile rounded above antenna (male) or transversely concave above antennae (female), pubescence as silver patches along eye margin, frontal vestiture as small dark setae, surface texture as irregular longitudinal striations (female) or smooth (male); face broadly rounded with dark or pale setae; gena with pale setae; parafacial overlain with silver pubescence; mouthparts length variable, but usually rela-



Figure 22. Acupalpa yanchep sp. n., male, anterolateral view [576260]. Body length = 6.0 mm.

tively short; palpus brown-black, acuminate; occiput overlain with sparse, silver-grey pubescence; antennal base raised; antenna longer than head; scape brown, length approximately equal to flagellum, scape with sparse black setae; flagellum black, base of flagellum with short, dark setae. *Thorax.* Scutum dark, overlain with pubescence of irregular brown to grey markings with pale broken lines and spots, setal bases glossy black; scutellum overlain with grey and matte black pubescence; pleuron dark, overlain with silver-grey pubescence; wing markings irregularly banded to apparently fenestrate; haltere knob white; coxae black, overlain with silver pubescence; femora brown to black; tibia black, yellow-orange dorsal stripe in basal 1/2; tarsi white, basitarsi dark in basal 3/4. Scutal chaetotaxy: *np*, 3–5; *sa*, 1 [rarely 2–3]; *pa*, 1; *dc*, 2–3; *sc*, 1. *Abdomen.* Black (segments 6-8 orange in some females), silver velutum dorsally on tergites (male) or absent (female); terminalia pale.

Comments. Acupalpa yanchep sp. n. is morphologically similar to A. albitarsa, with females difficult to separate except for more pronounced white scutal patterning in many individuals. This species is found in western Australia while A. albitarsa is found in eastern and southern regions. There is a pronounced size difference in the sexes of A. yanchep sp. n. with males considerably smaller than females. The leg colouration and scutal patterning is distinctive for this species.

Etymology. This species is named after the type locality, the township of Yanchep, Western Australia.

Pipinnipons Winterton

urn:lsid:zoobank.org:act:157A683F-6C11-4309-A7F5-5B5C4C37E47C http://species-id.net/wiki/Pipinnipons

Diagnosis. Antenna elongate, cylindrical, total length slightly longer than or equal to head length; scape shorter than flagellum; frons flat, smooth; face as narrow strip below antenna, glabrous; palpus spatulate apically; mouthparts short; occiput with single row of postocular setae immediately laterad of ocellar tubercle in male, multiple rows in female; wing banded infuscate or hyaline; setae absent on wing vein R₁; cell m₃ closed; elongate velutum patches on fore and hind femora; femora without macrosetae; single type of setal pile on femora, setae not appressed; prosternal furrow without setae; mid coxa without setae on posterior surface; post spiracular pile absent; gonocoxites with velutum patch on ventral surface (Fig. 24); articulated gonocoxal process present; hypandrium present; ventral apodeme of parameral sheath forked; dorsal apodeme of parameral sheath 'T'-shaped; three spermathecae in female; spermathecal sac present, usually with two smaller, additional lobes and/or outer reticulated lobes along length; spermathecal ducts joining common duct before bursa; female with A1 and A2 acanthophorite spines well developed; female sternite 8 emarginate along posterior margin.

Comments. Pipinnipons is a distinctive genus of wasp mimicking therevids, often with metallic pubescence, yellow and black marking and banded wings (Fig. 23). It can be distinguished among related genera by the elongate, cylindrical antennae, scape not longer than flagellum, narrow face and palpi spatulate. The latter two characters specifically differentiate *Pipinnipons* from *Acupalpa*, as the face is broadly rounded, often produced, and the palpi are acuminate or narrowly cylindrical in Acupalpa. While the mouthparts are of variable length in Acupalpa (and often elongate and forward projecting), the mouthparts of *Pipinnipons* are always relatively short. As stated in the comments under Acupalpa, Agapophytus is separated from Pipinnipons and Acupalpa by the length of the scape ranging from relatively equal length, to significantly longer than the flagellum. The modified setae patch on abdominal tergite 2 mentioned by Winterton et al. (2001) as a characteristic of *Pipinnipons* is not present in all the new species described here, and is no longer considered diagnostic for the genus as it is also found sporadically in other, unrelated genera such as Neodialineura Mann, 1928 and Bonjeania Irwin & Lyneborg, 1989. The male terminalia are relatively conserved throughout the genus, and species identification is more easily done using external characters of both sexes. Pipinnipons is distributed along coastal eastern Australia from northern Queensland to Tasmania.

Included species. Pipinnipons chauncyvallis sp. n., P. fascipennis (Kröber), P. kampmeierae sp. n., P. kroeberi Winterton, P. sphecoda sp. n.

Pipinnipons Winterton, 2001: 205. Type species: *Pipinnipons kroeberi* Winterton, 2001: 206.

Key to Pipinnipons species:

1	Abdomen with bright yellow and black markings (Fig. 29)
	P. sphecoda sp. n.
_	Abdomen otherwise coloured and marked
2	Abdominal tergites 4–8 overlain with dense silver to gold velutum; legs and pleuron dark orange-maroon (Figs 23, 26) <i>P. fascipennis</i> (Kröber)
_	Abdominal tergites 4–8 without dense silver to gold velutum; leg colour vari- able, usually bright orange (Figs 27–28) to yellow with dark markings (Fig. 25); pleuron black
3	Legs and coxae brown and yellow; wings banded infuscate; abdomen entirely brown to black (Fig. 25) (Tasmania)
_	Legs and coxae orange; wings largely hyaline or very weakly banded infuscate; some abdominal segments orange (Queensland)
4	Hind coxa dark; abdomen mostly orange (segments 3–4 with black suffu- sion); intersegmental margin of segment 3 not lighter (Fig. 28)
	<i>P. kroeberi</i> Winterton
-	Hind coxa orange; abdominal segments 2-3 orange with black dorsum, rest
	glossy black; intersegmental margin of segment 3 white (Fig. 27)
	P. kampmeierae sp. n.

Pipinnipons chauncyvallis sp. n.

urn:lsid:zoobank.org:act:718CB653-E72C-4333-A6C0-C54249CFDAEE http://species-id.net/wiki/Pipinnipons_chauncyvallis Fig. 25

Type material. Holotype male, AUSTRALIA: **Tasmania:** Bagdad, Chauncyvale Wildlife Sanctuary [-42.614°, 147.256°], 18–19.xii.1998, D. Yeates, S. Winterton (ANIC29_021139) (ANIC). **Paratypes:** AUSTRALIA: **Tasmania:** 3 females, same data as holotype (ANIC29_021136, 021137, 021140) (ANIC).

Diagnosis. Wing banded; pleuron black; femora yellow [hind femur dark]; tibia yellow, darker apically]; abdomen black, segments 6–8 orange [female].

Description. Body length= 7.0–10.0 mm. *Head.* Frons wider than ocellar tubercle in female, equal in male, profile flat to rounded above antenna, pubescence absent, frontal vestiture as numerous elongate setae (longer in male), surface texture as irregular longitudinal striations or transverse striations; gena with pale setae; parafacial overlain with silver pubescence; palpus yellow-orange; occiput glabrous, glossy black; antennal base raised; antenna longer than head; scape yellow, length shorter than flagellum, with sparse black setae; flagellum dark yellow (darker basally), base of flagellum with short dark setae. *Thorax.* Scutum uniform grey-black with white pile and overlain with sparse grey pubescence; sutellum overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing markings banded infuscate; haltere

knob white, dark basally; coxae black; hind femur dark, rest yellow with dark patch; tibia yellow (apices sometimes darker); tarsi yellow orange, dark distally, fore-basitarsus white. Scutal chaetotaxy: *np*, 3–4; *sa*, 1; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen.* Black, segments 6–8 orange in female; terminalia pale.

Comments. *Pipinnipons chauncyvallis* sp. n. is known only from a small conservation area near Bagdad, Tasmania. This species differs from all other *Pipinnipons* by the body colouration and the numerous pale setae on the frons.

Etymology. This species is named after the type locality, Chauncyvale Wildlife Sanctuary, owned by the Chauncy family who established and maintain the sanctuary.

Pipinnipons fascipennis (Kröber)

urn:lsid:zoobank.org:act:336E5D1F-DAAF-4532-BFFB-1CAFD12670E8 Genbank Accession: AF150979 http://species-id.net/wiki/Pipinnipons_fascipennis Fig. 23, 24A–D, G, 26

Squamopygia fascipennis Kröber 1928: 36. *Pipinnipons fascipennis* (Kröber) - Winterton et al. 2001: 211.

Type material. Type male. AUSTRALIA: **Queensland:** Kuranda [-16.817°, 145.633°], Lichtwardt (MEI090896) (DEI).

Other material examined- AUSTRALIA: **Queensland:** male, Indooroopilly, Long Pocket, 22.viii–7.ix.2007, S. L. Winterton, Malaise trap (UQIC) (MEI165213).

Diagnosis. Wing dark banded; pleuron orange to maroon; legs orange to maroon, tarsi lighter; abdomen dark red, tergite 2–3 red-brown, gold-bronze velutum on segments 4–7.

Redescription. Body length= 7.0–9.0 mm. *Head.* Frons wider than ocellar tubercle (female) or narrower (male), profile rounded above antenna, pubescence matte black and bronze, surface texture smooth or striated; gena with pale setae; parafacial overlain with silver pubescence; palpus brown-black; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape orange to brown, shorter than flagellum, with sparse black setae; flagellum orange to brown, base of flagellum with short dark setae. *Thorax.* Scutum dark with irregular brown and grey pubescente; pleuron orange to maroon, overlain with sparse silver-grey pubescence; wing markings dark banded infuscate; haltere knob white; coxae, femora and tibia orange-maroon; tarsi lighter orange, foreleg with basitarsus dark, white apically, rest of foretarsi white with slightly darker apex. Scutal chaetotaxy: np, 4–5; sa, 1; pa, 1; dc, 1; sc, 1. *Abdomen.* Dark, tergites 2–3 red-brown laterally, gold-bronze velutum on tergites 4–7; terminalia pale.

Comments. The gold-bronze abdominal velutum covering in both sexes and the dark orange pleuron and leg colouration make *P. fascipennis* easily recognisable. This species is found in closed forest areas, including rainforest.



Figure 23. Pipinnipons fascipennis (Kröber), male. Body length = 6.0 mm. (Photo: S.L. Winterton).

Pipinnipons kampmeierae sp. n.

urn:lsid:zoobank.org:act:5F4F235B-49AE-431B-A79C-041BB73DE633 http://species-id.net/wiki/Pipinnipons_kampmeierae Fig. 27

Type material. Holotype male, AUSTRALIA: **Queensland:** Jimmy's Scrub State Forest, nr. Goomeri, 22.xi.1985, M. De Baar (MEI165194) (QM). **Paratype.** AUS-TRALIA: **Queensland:** male, Bribie Island, DPI Fisheries site, [-27.055°, 153.193°], 7.x.1997, S. L. Winterton, N. Power, D. White, heathland- *Acacia* regrowth, Malaise trap (MEI090764) (QM).

Diagnosis. Wing mostly hyaline; pleuron black; coxae, femora and tibia orange; abdomen black, segments 2–3 orange with dark medial patch.

Description. Body length= 6.0–7.0 mm. *Head.* Frons narrower than ocellar tubercle, profile rounded above antenna, surface texture smooth, glabrous; gena with pale setae; parafacial overlain with silver pubescence; palpus brown-black; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape orange-yellow, much shorter than flagellum, scape with sparse dark setae dorsally; flagellum orange-yellow, base of flagellum without setae. *Thorax.* Scutum glossy black, overlain with sparse yellow setae, grey pubescence laterally; scutellum overlain with sparse grey pubescence; pleuron black, overlain with sparse silver-grey pubescence; wing largely hyaline, faint band midway; haltere knob white; coxae, femora and tibia orange; tarsi white, fore-basitarsus dark, rest of basitarsi orange. Scutal chaetotaxy: *np*, 5; *sa*, 1–2; *pa*, 1; *dc*, 2; *sc*, 1. *Abdomen*. Segments 2–3 orange with black medial patch, remaining segments black; terminalia pale.

Comments. *Pipinnipons kampmeierae* sp. n. is very similar to *P. kroeberi*, but can be differentiated based on coxae and abdomen colouration. The wings are only weakly infuscate rather than strongly banded infuscate as in most other species of *Pipinnipons* and *Acupalpa*. The female is unknown for this species.

Etymology. This species is named in honour of Gail Kampmeier, in recognition of her excellent work on Therevidae bioinformatics.

Pipinnipons kroeberi Winterton

urn:lsid:zoobank.org:act:97517024-09B0-41E4-8E27-DA0D8A8155CF Genbank Accession: AF150980 http://species-id.net/wiki/Pipinnipons_kroeberi Figs 24E–F, 28

Pipinnipons kroeberi Winterton, 2001: 205.

Type material. Holotype male, AUSTRALIA: New South Wales: Warrumbungle N.P., Buckleys Creek, 1.7 km N Camp Blackman, 23.xii.1992 M. E. Irwin (MEI027580) (ANIC). Paratypes. AUSTRALIA: New South Wales: female, Warrumbungle N.P., Browns Creek, 2.5 km N Woolshed, 13.i.1994, M. E. Irwin (MEI039303) (ANIC). Queensland: male, female, Lake Broadwater, 25 km SW Dalby, on *Leptospermum flavescens* blossom, 18.x.1985, D. K. Yeates (MEI090894, 090895) (GDCB/AMS).

Diagnosis. Wing hyaline; pleuron black; femora and tibia orange to yellow; abdomen orange, segments 1–3 sometimes black medially.

Redescription. Body length= 8.0–9.5 mm. *Head.* Frons wider than ocellar tubercle (equal in male), profile rounded or transversely concave above antennae (female), pubescence as silver patches along eye margin, frons otherwise glabrous, surface texture as irregular longitudinal striations (female) or smooth (male); gena with dark setae; parafacial overlain with silver pubescence; palpus yellow-orange; occiput glabrous, glossy black; antennal base raised; antenna longer than head; scape yellow, length much shorter than flagellum, scape with sparse pale setae; flagellum yellow, base of flagellum with short, dark setae. *Thorax.* Scutum uniform grey-black; scutellum overlain with sparse, grey pubescence; pleuron black, overlain with sparse, silver-grey pubescence; wing hyaline, orange suffusion along costal margin; haltere knob orange-yellow; coxae orange-yellow (hind coxa dark); femora and tibia orange or yellow, fore tibia apex dark; tarsi yellow, fore-basitarsus dark basally, rest of foreleg tarsomeres white. Scutal chaetotaxy: *np.*, 4; *sa*, 1; *pa.*, 1; *dc.* 3; *sc.*, 1. *Abdomen.* Orange, segments 1–2 black medially; terminalia pale.

Comments. *Pipinnipons kroeberi* is similar to *P. kampmeierae* sp. n. and can be differentiated based on the body colouration. This species is found in southeastern Queensland and northeastern New South Wales.



Figure 24. *Pipinnipons* spp.: **A** *P. fascipennis* (Kröber), epandrium and tergite 8, dorsal **B** gonocoxites, ventral **C** same, lateral **D** aedeagus, lateral **E** *P. kroeberi* Winterton, aedeagus, lateral **F** gonocoxites, epandrium removed and aedeagus *in situ*, dorsal **G** *P. fascipennis*, male head, lateral **H** *P. kroeberi*, female internal genitalia showing spermathecal sac complex. Scale lines = 0.2 mm.


Figure 25. *Pipinnipons chauncyvallis* sp. n., male, lateral view [576261]. Body length = 7.0 mm.



Figure 26. Pipinnipons fascipennis (Kröber), male, anterolateral view [576262]. Body length = 7.0 mm.



Figure 27. *Pipinnipons kampmeierae* sp. n., male, lateral view [576263]. Body length = 6.0 mm.



Figure 28. *Pipinnipons kroeberi* Winterton, male, anterolateral view [576264]. Body length = 8.0 mm.

Pipinnipons sphecoda sp. n.

urn:lsid:zoobank.org:act:329B77AF-DF06-4FB5-9E0C-0EEB79F1638F http://species-id.net/wiki/Pipinnipons_sphecoda Fig. 29

Type material. Holotype female, AUSTRALIA: **Tasmania:** Claytons [-43.383°, 146.133°], Jan.1991, E. D. Edwards, E. S. Nielsen (MEI027583) (ANIC). **Paratypes**. AUSTRALIA: **Tasmania:** female, Lake St. Clair, [-42.067°, 146.167°], 25.i.1949, E. F. Riek (MEI027585) (ANIC); female, 10 km ENE Nunamara [-41.367°, 147.4°], 12.i.–7.ii.1983, malaise trap, I. D. Naumann, J. C. Cardale (MEI027586) (ANIC).

Diagnosis. Wing with yellow and black irregular banding; pleuron black; femora and tibia yellow, sometimes with dark suffusion midway along femur; abdomen black, bright yellow-orange markings on tergites 1–4 and 7–8.

Description. Body length= 10.0–14.0 mm. *Head.* Frons wider than ocellar tubercle, profile slightly transversely concave above antennae, pubescence absent, frontal vestiture as minute setae, surface texture as irregular longitudinal striations and transverse striations; gena with dark setae; parafacial overlain with silver pubescence; palpus yellow-orange; occiput glabrous, glossy black; antennal base raised; antennal length approximately equal to head; scape yellow, shorter than flagellum, scape with sparse black setae; flagellum yellow, base of flagellum with short dark setae. *Thorax.* Scutum glossy black, overlain with sparse yellow setae, sparse grey pubescence laterally; scutellum



Figure 29. Pipinnipons sphecoda sp. n., female, anterolateral view [576265]. Body length = 10.0 mm.

overlain with dense, matt-black pubescence; pleuron black, overlain with sparse silvergrey pubescence; wing with dark band midway, pale yellow suffusion basally and in discal area; haltere knob white to yellow; coxae black, overlain with silver pubescence; femora yellow-orange, sometimes with dark patch midway; tibia yellow-orange; tarsi yellow, fore leg with basitarsus and second tarsomere white, rest of tarsomeres dark. Scutal chaetotaxy: *np*, 4–5; *sa*, 1; *pa*, 2; *dc*, 1; *sc*, 1. *Abdomen.* Black, bright yelloworange markings on tergites 1–4 and 7–8; terminalia pale.

Comments. *Pipinnipons sphecoda* sp. n. is a relatively large, apparently wasp-mimicking species known only from female specimens collected from various sites in Tasmania. The dramatic colouration of species makes it quite unlike any other stiletto fly species.

Etymology. The species epithet is derived from the Gr. sphekodos, wasp-like.

Acknowledgements

Thank you to Gail Kampmeier for her valuable assistance with extracting information from the MANDALA therevid specimen database. Thank you to Dr Christine Lambkin (QM), Brian Hanich (WAM) and Dr Mike Irwin (Illinois Natural History Survey) for help with examining type material and loan of specimens. Thank you also to Christine Lambkin who recognised the synonymy of *Acupalpa pollinosa* with *Ectinorhynchus albimanus* and found the male of *A. divisa*, bringing both of these to my attention. Thank you to Donald Holbern (Atlas of Living Australia) and Garry Jolley-Rogers (CSIRO) for provision and help with setting up of LSIDs. Drs Donald Webb (Illinois Natural History Survey), Mark Metz, Stephen Gaimari, Martin Hauser and Christine Lambkin provided useful comments on the draft manuscript and interesting discussions regarding the future of the taxonomic paradigm. This research was supported by the National Science Foundation (NSF-DEB-0614213) and the Australian Biological Resource Study (ABRS-209-48). Statements and viewpoints expressed herein do not necessarily reflect the opinion of NSF or ABRS.

References

- Blagoderov V, Brake I, Georgiev T, Penev L, Roberts D, Rycroft S, Scott B, Agosti D, Catapano T, Smith V (2010) Streamlining taxonomic publications: a working example with Scratchpads and Zookeys. Zookeys 50: 17–28. http://dx.doi.org/10.3897/zookeys.50.539
- Brown WL (1961) An international taxonomic register: preliminary proposals. Systematic Zoology 10: 80–85. http://dx.doi.org/10.2307/2411725
- Deans AR, Kawada R (2008) Alobevania, a new genus of neotropical ensign wasps (Hymenoptera: Evaniidae), with three new species: integrating taxonomy with the World Wide Web. Zootaxa 1787: 28–44.
- Erwin TL, Johnson PJ (2000). Naming species, a new paradigm for crisis management in taxonomy: rapid journal validation of scientific names enhanced with more complete descrip-

tions on the internet. Coleopterists Bulletin 54: 269–278. doi:10.1649/0010-065X(2000)054[0269:NSANPF]2.0.CO;2

- Hardy GH (1939) Miscellaneous notes on Australian Diptera. V. Proceedings of the Linnaean Society of New South Wales 64: 34–50.
- Hauser M, Irwin ME (2003) The Nearctic Genus Ammonaios Irwin and Lyneborg 1981 (Diptera: Therevidae). Annals of the Entomological Society of America 96: 738-765. doi:10.1603/0013-8746(2003)096[0738:TNGAIA]2.0.CO;2
- International Commission on Zoological Nomenclature (2008) Proposed amendment of the International Code of Zoological Nomenclature to expand and refine methods of publication. Zootaxa 1908: 57–67.
- Irwin ME, Lyneborg L (1989) 39. Family Therevidae. In: Evenhuis NL (Ed) Catalog of Diptera of the Australasian and Oceanian regions. Bishop Museum Special Publication. Bishop Museum Press 86: 353–358.
- Johnson NF, Masner L, Musetti L, Van Noort S, Rajmohana K, Darling DC, Guidott A, Polaszek A (2008) Revision of world species of the genus *Heptascelio* Kieffer (Hymenoptera: Platygastroidea, Platygastridae). Zootaxa 1776: 1–51.
- Kampmeier GE, Irwin ME (2009) Meeting the interrelated challenges of tracking specimen, nomenclature, and literature data in Mandala. In: Pape T, Meier R, Bickel D (Eds) Diptera Diversity: Status, Challenges and Tools. Brill Academic Publishers, Leiden, 407–438.
- Kröber O (1912) Die Thereviden der indo-australischen Region. Entomologische Mitteilungen, 1, 116–125, 148–159, 183–189, 215–222, 242–256, 282–287.
- Kröber O (1913) Fam. Therevidae. Genera Insectorum, Bruxelles, 148: 1-69.
- Kröber O (1914) Beitrage zur Kenntnis der Thereviden und Omphraliden. Mitteilungen Naturhistorischen Museum, Hamburg 31: 29–74.
- Kröber O (1928) Neue Dipteren des Deutschen Entomolog. Museums in Dahlem. Entomologische Mitteilungen 17: 31–41.
- Mann JS (1928) Revisional notes on Australian Therevidae. Part 1. Australian Zoologist 5: 151–194.
- Mann JS (1929) Revisional notes on Australian Therevidae. Part 2. Australian Zoologist 6: 17–49.
- Mann JS (1933) Revisional notes on Australian Therevidae. Part 3. Australian Zoologist 7: 325–344.
- McAlpine JF (1981) Morphology and terminology-Adults. In: McAlpine JF, Peterson BV, Shewell GE, Teskey HJ, Vockeroth JR, Wood DM (Eds) Manual of Nearctic Diptera. Research Branch, Agriculture Canada Monograph 1: 9–63.
- Miller JA, Griswald CE, Yin CM (2009) The symphytognathoid spiders of the Gaoligongshan, Yunnan, China (Araneae, Araneoidea): Systematics and diversity of micro-orbweavers. Zookeys 11: 9–195. http://dx.doi.org/10.3897/zookeys.11.160
- Nichols SW (Ed) (1989) The Torre-Bueno glossary of entomology: revised edition. New York Entomological Society, 840 pp.
- Penev L, Agosti D, Georgiev T, Catapano T, Miller J, Blagoderov V, Roberts D, Smith VS, Brake I, Ryrcroft S, Scott B, Johnson NF, Morris RA, Sautter G, Chavan V, Robertson T, Remsen D, Stoev P, Parr C, Knapp S, Kress WJ, Thompson FC, Erwin T (2010) Semantic

tagging of and semantic enhancements to systematics papers: ZooKeys working examples. Zookeys 50: 1–16. http://dx.doi.org/10.3897/zookeys.50.538

- Polaszek A, Agosti D, Alonso-Zarazaga M, Beccaloni G, de Place Bjørn P, Bouchet P, Brothers DJ, Earl of Cranbrook, Evenhuis NL, Godfray HCJ, Johnson NF, Krell FT, Lipscomb D, Lyal CHC, Mace GM, Mawatari SF, Miller SE, Minelli A, Morris S, Ng PKL, Patterson DJ, Pyle RL, Robinson N, Rogo L, Taverne J, Thompson FC, van Tol J, Wheeler QD, Wilson EO (2005a) Commentary: A universal register for animal names. Nature 437: 477.
- Polaszek A, Alonso-Zarazaga M, Bouchet P, Brothers DJ, Evenhuis NL, Krell FT, Lyal CHC, Minelli A, Pyle RL, Robinson N, Thompson FC, van Tol J (2005b) ZooBank: the openaccess register for zoological taxonomy: technical discussion paper. Bulletin of Zoological Nomenclature 62: 210–220.
- Pyle RL, Earle JL, Greene BD (2008) Five new species of the damselfish genus *Chromis* (Perciformes: Labroidei: Pomacentridae) from deep coral reefs in the tropical western Pacific. Zootaxa 1671: 3–31.
- Walker F (1850) Diptera. Part I. In: Insecta Saundersiana: or characters of undescribed insects in the collection of William Wilson Saunders, Esq., F.R.S., F.L.S., &c. 1, John Van Voorst, London, 1–76.
- White A (1915) The Diptera-brachycera of Tasmania, Part ii, families Tabanidae and Therevidae. Proceedings Royal Society Tasmania, Hobart 1915: 1–59.
- Winterton SL, Irwin ME, Yeates DK (1999a) Systematics of Nanexila Winterton & Irwin, gen. n. (Diptera: Therevidae) from Australia. Invertebrate Taxonomy 13: 237–308. http:// dx.doi.org/10.1071/IT97029
- Winterton SL, Irwin ME, Yeates DK (1999b) Phylogenetic revision of the *Taenogera* Kröber genus-group (Diptera: Therevidae), with descriptions of two new genera. Australian Journal of Entomology 38: 274–290. http://dx.doi.org/10.1046/j.1440-6055.1999.00126.x
- Winterton SL (2000) Phylogenetic revision of Acupalpa Kröber (Diptera: Therevidae). Insect Systematics and Evolution 31: 225–240. http://dx.doi.org/10.1163/187631200X00417
- Winterton SL (2006) New species of *Eupsilocephala* Kröber from Australia (Diptera: Therevidae). Zootaxa 1372: 17–25.
- Winterton SL (2009) Revision of the stiletto fly genus *Neodialineura* Mann (Diptera: Therevidae): an empirical example of cybertaxonomy. Zootaxa 2157: 1–33.
- Winterton SL, Yang L, Wiegmann BM, Yeates DK (2001) Phylogenetic revision of Agapophytinae subf. n. (Diptera: Therevidae) based on molecular and morphological evidence. Systematic Entomology 26: 173–211. http://dx.doi.org/10.1046/j.1365-3113.2001.00142.x