RESEARCH ARTICLE



# An updated list of butterflies (Lepidoptera, Papilionoidea) of two Guatemalan seasonally dry forests

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#### Abstract

Guatemala has a great diversity of butterflies, although there have been few intensive surveys on Lepidoptera in the country so far. We present an updated list of 218 species in 149 genera, 19 subfamilies, and six families of butterflies sampled at two seasonally dry forests in the Salamá and Motagua valleys in central and eastern Guatemala, by integrating new data from field surveys conducted in 2014–2021 into our previously published data (Yoshimoto et al. 2018, 2019), with *Amblyscirtes elissa elissa* Godman, 1900, *Repens florus* (Godman, 1900), and *Niconiades nikko* Hayward, 1948 (Hesperiidae: Hesperiinae) as new country records. We collected a hairstreak species, *Chalybs hassan* (Stoll, 1790) (Lycaenidae: Theclinae), at the Motagua Valley site, representing the second record for Guatemala since the early 20<sup>th</sup> century, after we rediscovered it at the Salamá Valley site in 2011 and 2012 (Yoshimoto and Salinas-Gutiérrez 2015). Nymphalidae and Hesperiidae had larger numbers of species than the other four families at both sites. In Pieridae and Nymphalidae, species composition was similar between the sites, whereas in Lycaenidae, Riodinidae, and Papilionidae it differed more greatly between the sites. These results confirm the relatively high lepidopteran diversity of Guatemalan dry forests, noteworthy for the small areas that comprise the study sites, and represent marked similarities and differences in butterfly fauna and phenology within these forests.

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#### **Keywords**

Annotated list, dissimilarity, Hesperiidae, inventory, Mesoamerica, Neotropics, seasonality

## Introduction

Neotropical seasonally dry forests are rich in flora and fauna (Pennington et al. 2006; Dirzo et al. 2011), although their ecosystems have been deteriorating because of various anthropogenic disturbances, such as deforestation due to agricultural expansion (e.g., Chazdon et al. 2011). Dry forests in Guatemala also harbor high lepidopteran diversity as well; our previous studies documented more than 150 and 100 butterfly species at the two small forest reserves in central and eastern Guatemala, respectively (Yoshimoto et al. 2018, 2019). These species lists, however, are still incomplete, and obviously, more species remain to be sampled at these sites. Moreover, we detected marked seasonal patterns in butterfly species richness and several conspicuous differences in the lepidopteran fauna between the two sites (Yoshimoto et al. 2019). Thus, it was apparent that additional field surveys were needed to make quantitative between-site comparisons of species composition, in order to enhance our understanding of butterfly fauna and phenology of these forests.

In Guatemala, approximately 400 species of Hesperiidae and nearly 700 species of the remaining families of Papilionoidea have been reported (Austin et al. 1998; Barrios et al. 2006; Salinas-Gutiérrez et al. 2009, 2012; Salinas-Gutiérrez 2013). Despite such high lepidopteran diversity, Guatemala's butterfly fauna has been studied less intensively compared to neighboring countries; for example, in Mexico, exhaustive species lists for the whole country and for several states have been published (de la Maza et al. 1989, 1991; Luis-Martínez et al. 2011, 2016; Llorente-Bousquets et al. 2014), whereas Guatemala has had little research on Lepidoptera and few published inventories since the 20<sup>th</sup> century (but see Austin et al. 1996 and Yoshimoto et al. 2021). Continued field surveys in various parts of Guatemala are thus important to fill a gap in our knowledge of the Neotropical butterfly fauna, which will in turn contribute to biodiversity conservation in the country.

Here, we present an updated and integrated list of papilionoid species (including Hesperiidae; van Nieukerken et al. 2011) for the same dry forest sites where we conducted our previous studies (Yoshimoto et al. 2018, 2019), by adding the new data from subsequent field surveys performed in 2014–2021, correcting identification errors, and modifying some of the species names based on taxonomic changes. Additionally, we examine between-site differences in butterfly fauna by comparing species composition at the family level, and identify seasonal patterns at the species level.

#### Materials and methods

This study was conducted at the Los Cerritos Municipal Park (hereafter, Los Cerritos; Fig. 1a) in the Salamá Valley (a subwatershed of the Chixoy region) of Baja Verapaz

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Department in central Guatemala (15°05'N, 90°18'W, 960–1160 m a.s.l., 69 ha), and at the Heloderma Natural Reserve (hereafter, Heloderma Reserve; Fig. 1b) in the Motagua Valley of Zacapa Department in eastern Guatemala (14°53'N, 89°47'W, 510– 790 m a.s.l., 58 ha). The rainfall patterns are similar between the two areas, in which the rainy season usually begins in late May and ends in October; these six months were accordingly defined as the rainy season and the remaining months (November-April) as the dry season. This climatic trait fits the definition of seasonally dry tropical forests (4–6 months with rainfall being < 100 mm; Dirzo et al. 2011); see fig. 1 in Yoshimoto et al. (2018) and fig. 3 in Yoshimoto et al. (2019) for detailed precipitation information of each area.

The vegetation of both regions is characterized by an abundance of various aculeate plants such as cacti (Cactaceae). The most dominant species is a columnar cactus *Stenocereus pruinosus* (Otto) Buxb., with *Pilosocereus leucocephalus* (Poselg.) Byles & G. D. Rowley and *Pereskia lychnidiflora* DC., also being abundant at both sites. On the other hand, there exist some marked differences in flora and in forest landscape. Heloderma Reserve has a dense forest with many arboreal species such as *Bucida macrostachya* Standl. (Combretaceae), *Lysiloma divaricatum* (Jacq.) J. F. Macbr., *Leucaena collinsii* Britton & Rose (both Mimosaceae), and *Bursera excelsa* (Kunth) Engl. (Burseraceae), all of which can grow taller than the columnar cactus (Ariano-Sánchez and Salazar 2015; D. Ariano-Sánchez, pers. comm.; Fig. 1b). By contrast, none of these species have been reported from Los Cerritos (M. R. Álvarez, pers. comm.), where there are fewer high arboreal species and abundant shrubs and herbaceous plants (thus commonly called a spiny bush or scrub), thereby the columnar cactus being prominent in its forest landscape (Fig. 1a).

Field surveys were conducted on 21 days from July 2014 to August 2021 at Los Cerritos, and on 19 days from October 2017 to November 2021 at Heloderma Reserve. We collected adult butterflies with an insect net or photographed them in the daytime (09:00-17:00) at each site and in neighboring areas (a small garden at the foot of Los Cerritos and on a farm road adjacent to Heloderma Reserve). The individuals collected were mounted as voucher specimens and were deposited at the Colección de Artrópodos, Laboratorio de Entomología Sistemática, Universidad del Valle de Guatemala. All the individuals collected or photographed were identified to species or subspecies according to Warren et al. (2017). We did not include data for specimens that were not identified to species, except for Calephelis spp. (Riodinidae) and Bolla sp. (Hesperiidae: Pyrginae); see the footnotes of the Appendix 1 for the rationales for the inclusion of these data. We added all these data to our previous data (Yoshimoto et al. 2018, 2019), corrected identification errors, and modified scientific names of some of the species based on taxonomic changes, in order to compile an updated and integrated species list of the two sites. Note that the sampling methods were partially different in the previous surveys; only netting was done at Los Cerritos from January 2011 to November 2012, whereas at Heloderma Reserve between February 2016 and March 2017, data were obtained through netting, photographing, and observation; see Yoshimoto et al. (2018, 2019) for detailed information on the sampling methods for each site.



Figure 1. a Forest landscape of Los Cerritos Municipal Park and b Heloderma Natural Reserve.

The site-level estimated species richness was calculated by using the Chao II index (Chao et al. 2005; Gotelli and Colwell 2011), after pooling the data across observation dates for each month for each site. The total estimated species richness was similarly obtained after pooling these data across both sites. The Jaccard dissimilarity index was used to quantify the between-site similarity in species composition; this index was calculated for all data and for each of the six families (Papilionidae, Pieridae, Lycaenidae, Riodinidae, Nymphalidae, and Hesperiidae). All the analyses were performed using R 4.1.2. (R Development Core Team 2021) with the package Vegan (Oksanen et al. 2020).

# Results

By integrating our previous data (Yoshimoto et al. 2018, 2019), a total of 218 species (including one unidentified taxon and 107 subspecies) in 149 genera from 19 subfamilies of six families were recorded at the two sites (Appendix 1). Hesperiidae was the richest family (71 species), followed by Nymphalidae, Lycaenidae, Pieridae, Riodinidae, and Papilionidae (66, 36, 20, 16, and 9 species, respectively). Los Cerritos had 166 species in 117 genera, and Heloderma Reserve had 139 species in 107 genera (Appendix 1), 16 and 41 species of which had been newly recorded in the subsequent surveys, respectively (Fig. 2). The estimated species richness (mean $\pm$ SE) of each site based on the Chao II index is 216.27 $\pm$ 16.35 and 187.35 $\pm$ 17.74, respectively, indicating that approximately 76.8% and 74.2% of the species inhabiting each site were sampled. The total estimated species richness for both sites is 272.80 $\pm$ 17.85 (79.9%).

We detected identification errors for 20 individuals (identified as 11 species in our previous studies) and determined them to represent 13 species in this study; ten individuals from Los Cerritos and ten from Heloderma Reserve have been determined to number eight and six different species, respectively, with one species, *Cissia themis*, shared between sites (Table 1). Additionally, Yoshimoto et al. (2019) incorrectly listed *Piruna* (Hesperiidae: Heteropterinae) in the subfamily Hesperiinae.

The following three skipper species (Hesperiidae: Hesperiinae) were recorded for the first time in Guatemala:

• *Amblyscirtes elissa elissa* Godman, 1900. Reserva Heloderma, Cabañas, Zacapa, GUATEMALA. Three specimens: 30-08-2016, J442; 26-09-2016, J478; 01-06-2018, J769. Collected by Jiichiro Yoshimoto. Identified by Andrew D. Warren. Note that the two individuals (J442 and J478) were misidentified as *Piruna* sp.1 in Yoshimoto et al. (2019), as shown in Table 1. The specimens were deposited in the Colección de Artrópodos, Laboratorio de Entomología Sistemática, Universidad del Valle de Guatemala, and are being cataloged (Fig. 3c). Distribution: Southwestern Mexico (Warren et al. 2017).

• *Repens florus* (Godman, 1900). Reserva Heloderma, Cabañas, Zacapa, GUATEMALA. One specimen: 23-10-2018, J800. Collected by Jiichiro Yoshimoto. Identified by Andrew D. Warren. The specimen was deposited as above and is being cataloged (Fig. 3d). Distribution: Eastern and Western Mexico, Belize, and Nicaragua (Warren et al. 2017).

• *Niconiades nikko* Hayward, 1948. Los Cerritos, Salamá, Baja Verapaz, GUA-TEMALA. One specimen: 16-11-2020, J1024. Collected and identified by Jiichiro Yoshimoto. The specimen was deposited as above and is being cataloged (Fig. 3e). Distribution: Eastern Mexico to Ecuador, Southern Brazil, and Paraguay (Warren et al. 2017).

Eighty-six species were shared between Los Cerritos and Heloderma Reserve (Table 2), which amounts to 51.8% and 61.9% of the species sampled at each site (the Jaccard dissimilarity index is 0.606). At both sites, species richness of Nymphalidae



Figure 2. Six of the species that were newly recorded in the present study at Los Cerritos or Heloderma Reserve **a**, **b** *Heraclides rumiko* Shiraiwa & Grishin, 2014 (Papilionidae) **c** *Archaeoprepona demophon centralis* (Fruhstorfer, 1905) **d** *Caligo telamonius memnon* (C. Felder & R. Felder, 1867) (both Nymphalidae) **e** *Abaeis nicippe* (Cramer, 1779) (Pieridae) **f** *Leptotes cassius cassidula* (Boisduval, 1870) (Lycaenidae) **g** *Piruna aea* (Dyar, 1912) (Hesperiidae) **a–d, g** Heloderma Reserve **e, f** Los Cerritos. Note that *P. aea* had already been collected and identified to genus (*Piruna* sp.1) by Yoshimoto et al. (2019).

**Table 1.** Butterfly species that were sampled at Los Cerritos and Heloderma Reserve (abbreviated as LC and HR, respectively) and were misidentified in Yoshimoto et al. (2018, 2019). Corrected species names are shown in bold.

Family	Species		Sampling month, year,
	Correct identification	Previous identification	and site
Papilionidae	<i>Heraclides rumiko</i> Shiraiwa & Grishin, 2014	Heraclides cresphontes (Cramer, 1777) <sup>A</sup>	Oct 2016 HR*
Pieridae	Abaeis nicippe (Cramer, 1779)	Pyrisitia proterpia (Fabricius, 1775) <sup>A</sup>	Jul 2016 HR
Lycaenidae	Strymon megarus (Godart, [1824])	Strymon melinus franki W. D. Field, 1938 <sup>A</sup>	Oct 2016 HR
Nymphalidae	Anthanassa tulcis (H. Bates, 1864)	Anthanassa dracaena phlegias (Godman, 1901) <sup>B</sup>	May 2011 LC
Nymphalidae	Chlosyne erodyle erodyle (H. Bates, 1864)	Chlosyne lacinia lacinia (Geyer, 1837) <sup>B</sup>	Oct 2011 LC, Jul 2012 LC
Nymphalidae	Chlosyne rosita rosita A. Hall, 1924	Chlosyne lacinia lacinia (Geyer, 1837) <sup>B</sup>	Sep 2011 LC
Nymphalidae	Cissia similis (A. Butler, 1867)	<i>Cissia pompilia</i> (C. Felder & R. Felder, 1867) <sup>B</sup>	May 2012 LC, Jun 2012 LC
Nymphalidae	Cissia themis (A. Butler, 1867)	<i>Cissia pompilia</i> (C. Felder & R. Felder, 1867) <sup>A, B</sup>	Aug 2011 LC, Aug 2016 HR**, Oct 2016 HR
Hesperiidae	<i>Urbanus viterboana</i> (Ehrmann, 1907)	Urbanus proteus proteus (Linnaeus, 1758) <sup>B</sup>	Nov 2011 LC
Hesperiidae	Heliopetes macaira macaira (Reakirt, [1867])	Heliopyrgus domicella domicella (Erichson, [1849]) <sup>B</sup>	Jul 2012 LC
Hesperiidae	Amblyscirtes elissa elissa Godman, 1900	Piruna sp.1 <sup>A</sup>	Aug 2016 HR, Sep 2016 HR
Hesperiidae	Copaeodes aurantiaca (Hewitson, 1868)	Ancyloxypha arene (W. H. Edwads, 1871) <sup>B</sup>	Mar 2011 LC
Hesperiidae	Cymaenes trebius (Mabille, 1891)	<i>Cymaenes tripunctus theogenis</i> (Capronnier, 1874) <sup>A</sup>	Sep 2016 HR**

<sup>A</sup> Listed in Yoshimoto et al. (2019).

<sup>B</sup> Listed in Yoshimoto et al. (2018).

\*Specimen not collected (recorded only by photographing; see Fig. 2a, b for its images).

\*\*Two individuals collected.

and Hesperiidae was greater than that of the other four families, although family-level species richness differed greatly between the sites (Table 2; Fig. 4). In particular, the proportion of Lycaenidae was much higher at Los Cerritos (18.7%) than at Heloderma Reserve (8.6%), which was mainly due to differences in the subfamily Theclinae (26 and 7 species, respectively: Appendix 1).

Family-level species composition also differed between the sites, and the magnitude of this difference varied among the six families (Table 2). The dissimilarity indices for Riodinidae, Lycaenidae, and Papilionidae were considerably larger, indicating that species composition differed more greatly between the sites in these families. Pieridae and Nymphalidae, by contrast, had smaller indices with many shared species, demonstrating that their species composition was relatively similar between the sites.

Ninety-three species (42.7%) occurred in both dry and rainy seasons, whereas 103 (47.2%) appeared only in the rainy season and 22 species (10.1%) only in the



**Figure 3.** One species of hairstreak (Lycaenidae: Theclinae) **a** *Chalybs hassan* (Stoll, 1790), one species of skipperling (Hesperiidae: Heteropterinae) **b** *Piruna aea* (Dyar, 1912), and three species of grass-skippers (Hesperiidae: Hesperiinae) **c** *Amblyscirtes elissa elissa* Godman, 1900 **d** *Repens florus* (Godman, 1900), and **e** *Niconiades nikko* Hayward, 1948. The three grass-skipper species were newly recorded for Guatemala. Dorsal and ventral views, respectively, are shown at the left and right in each photograph.

**Table 2.** Species richness for six families at Los Cerritos and Heloderma Reserve, and comparisons of species composition at the family level between the sites, based on the number of shared species and the Jaccard dissimilarity index.

Family	Total No. species		No. shared species	Jaccard index
	Los Cerritos	Heloderma Reserve		
Papilionidae	7	4	2	0.778
Pieridae	16	17	13	0.350
Lycaenidae	31	12	7	0.806
Riodinidae	10	9	3	0.813
Nymphalidae	57	46	37	0.439
Hesperiidae	45	51*	24	0.662*

\*The data for *Bolla* sp. were included in the species count but excluded from the Jaccard index analysis (see the footnote 7 of the Appendix 1 for its rationale).

dry season. The most frequently recorded species was *Eurema daira eugenia* (Wallengren, 1860) (Pieridae: Coliadinae), which was collected or observed throughout the year (Appendix 1). The second most frequently recorded species (in 11 months) were *Kricogonia lyside* (Godart, 1819) (Coliadinae) and *Hamadryas glauconome glauconome* (H. Bates, 1864) (Nymphalidae: Biblidinae), followed by *Pyrisitia proterpia* (Fabricius, 1775) (Coliadinae: in ten months), *Phoebis sennae marcellina* (Cramer, 1777) (Coliadinae), *Mestra amymone* (Ménétriés, 1857) (Biblidinae), and *Urbanus dorantes dorantes* (Stoll, 1790) (Hesperiidae: Eudaminae: all in nine months).



Figure 4. Proportion of species richness at the family level at Los Cerritos and Heloderma Reserve.

# Discussion

A total of 218 species were recorded at the two dry forest sites during our 10-year field surveys, which confirms the relatively high lepidopteran diversity of Guatemalan seasonally dry forests for the small areas that comprise the study sites (<70 ha each). The estimated species richness suggests that nearly a quarter of the species inhabiting each site have yet to been recorded. The number of the additional species yielded in the subsequent surveys was more than twice greater at Heloderma Reserve than at Los Cerritos. The proportion of newly recorded species was much higher in Lycaenidae and Riodinidae; seven lycaenid species were added to the list for Los Cerritos, and seven lycaenid and four riodinid species were added to that of Heloderma Reserve, which nearly doubled the species richness of each family at this site (six lycaenid and five riodinid species in Yoshimoto et al. 2019). Among these species, the record of Chalybs hassan (Stoll, 1790) at Heloderma Reserve is highly important (Fig. 3a), as this species had not been reported for more than 100 years in Guatemala before we collected four individuals at Los Cerritos in 2011 and 2012 (Yoshimoto and Salinas-Gutiérrez 2015). These results highlight the importance of continuing butterfly surveys at both sites to create more exhaustive inventories, especially on small and taxonomically difficult taxa such as Lycaenidae and Riodinidae. Moreover, it is important to conduct research in other dry regions (e.g., the Nentón Valley in

northwestern Guatemala) and to make quantitative among-site comparisons of species richness and composition as well. All these studies will contribute to a comprehensive understanding of Neotropical butterfly fauna and distribution, and would serve as a scientific baseline for biodiversity conservation in Guatemalan dry regions.

More than half of the species sampled at each site were shared between the sites, suggesting that species composition is partially and moderately similar between Los Cerritos and Heloderma Reserve. Importantly, between-site similarity greatly differed among the six families. Higher similarity in Pieridae (especially in Coliadinae) would likely be associated with the distribution and abundance of their host plants, considering that coliadine larvae mostly feed on fabaceous plants such as *Senna* (e.g., DeVries 1987) and that these plants appear to be abundant at both sites.

In Lycaenidae and Riodinidae, species composition largely differed between the sites; in particular, Theclinae had considerable differences in species richness and composition (Appendix 1). In addition, most of these thecline species tended to be highly seasonal, as 25 out of 30 species were sampled only in the rainy season. In contrast to their marked seasonal pattern, *Strymon megarus* (Godart, [1824]) and *S. rufofusca* (Hewitson, 1877) occurred frequently also in the dry period at Heloderma Reserve; three and five individuals of each species were collected in both December and January at this site (Appendix 1). It should also be mentioned that *Hechtia guatemalensis* Mez (Bromeliaceae), one of the dominant bromeliad species at Heloderma Reserve (Fig. 1b), may be a possible foodplant for *S. megarus* at this site, as the larvae of this species are known to feed on bromeliads (Robbins 2010). Examination of abundance and distribution of host- and nectar-plants, as well as of larval and adult feeding behavior in relation to their phenology, would be an initial step to elucidate the bionomics of these species. Such surveys may also identify factors underlying the regional similarity and dissimilarity in the butterfly fauna.

We recorded *Amblyscirtes elissa elissa* Godman, 1900, *Repens florus* (Godman, 1900), and *Niconiades nikko* Hayward, 1948 (Hesperiidae: Hesperiinae) for the first time in Guatemala (Fig. 3c, d, e). Austin et al. (1998) listed *A. e. elissa* and *N. nikko* as species with a potential distribution in Guatemala. *Repens florus* could have been included in this category as well, as it is known to be distributed in the adjacent countries (Mexico, Belize, and Nicaragua; Warren et al. 2017). These results indicate that there still exists a gap in our knowledge of geographic distribution of Neotropical skipper species, again emphasizing the importance of more intensive research in Guatemala to bridge this gap.

Four individuals of *Piruna aea* (Dyar, 1912) (two in the previous survey and two in the subsequent one: Figs 2g, 3b) were collected at Heloderma Reserve. This is an interesting result, since most species in this genus are distributed in humid areas at higher elevation (1000–2700 m; Warren and González-Cota 1998). As Yoshimoto et al. (2019) pointed out, the wing pattern of these individuals is somewhat different from Mexican *P. a. aea* (Dyar, 1912), implying that *Piruna cingo sombra* Evans, 1955, described from Guatemala and currently considered a synonym of *P. a. aea*, may be a valid subspecies-level taxon. At present, this is difficult to determine, as very few specimens of this species have been sampled in Guatemala (Barrios et al. 2006).

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### References

- Ariano-Sánchez D, Salazar G (2015) Spatial ecology of the endangered Guatemalan Beaded Lizard *Heloderma charlesbogerti* (Sauria: Helodermatidae), in a tropical dry forest of the Motagua Valley, Guatemala. Mesoamerican Herpetology 2(1): 64–74. https://library.iucnisg.org/documents/2015/Ariano-Sanchez\_2015\_Mesoamerican\_Herpetology.pdf
- Austin GT, Haddad NM, Méndez C, Sisk TD, Murphy DD, Launer AE, Ehrlich PR (1996) Annotated checklist of the butterflies of the Tikal National Park area of Guatemala. Tropical Lepidoptera 7(1): 21–37. https://journals.flvc.org/troplep/article/view/90056
- Austin GT, Méndez C, Launer AE (1998) A preliminary checklist of Guatemala butterflies: Hesperiidae (Lepidoptera: Hesperioidea). Tropical Lepidoptera 9(Supplement 2): 8–19. https://journals.flvc.org/troplep/article/view/90138
- Barrios MV, Méndez CA, Austin GT (2006). Las Hesperiidae (Lepidoptera: Hesperioidea) de Guatemala. In: Cano EB (Ed.) Biodiversidad de Guatemala Vol. I. Universidad del Valle de Guatemala, Guatemala, 431–439.
- Brower AVZ (2006) Problems with DNA barcodes for species delimitation: 'ten species' of Astraptes fulgerator reassessed (Lepidoptera: Hesperiidae). Systematics and Biodiversity 4(2): 127–132. https://doi.org/10.1017/S147720000500191X
- Brower AVZ (2010) Alleviating the taxonomic impediment of DNA barcoding and setting a bad precedent: names for ten species of 'Astraptes fulgerator' (Lepidoptera: Hesperiidae: Eudaminae) with DNA-based diagnoses. Systematics and Biodiversity 8(4): 485–491. https://doi.org/10.1080/14772000.2010.534512
- Chao A, Chazdon RL, Colwell RK, Shen TJ (2005) A new statistical approach for assessing similarity of species composition with incidence and abundance data. Ecology Letters 8(2): 148–159. https://doi.org/10.1111/j.1461-0248.2004.00707.x
- Chazdon RL, Harvey CA, Martínez-Ramos M, Balvanera P, Stoner KE, Schondube JE, Avila Cabadilla LD, Flores-Hidalgo AM (2011) Seasonally dry tropical forest biodiversity and conservation value in agricultural landscapes of Mesoamerica. In: Dirzo R, Young HS, Mooney HA, Ceballos G (Eds) Seasonally dry tropical forests: ecology and conservation. Island Press, Washington DC, 195–219.

- Cong Q, Grishin NV (2014) A new *Hermeuptychia* (Lepidoptera, Nymphalidae, Satyrinae) is sympatric and synchronic with *H. sosybius* in southeast US coastal plains, while another new *Hermeuptychia* species not *hermes* inhabits south Texas and northeast Mexico. ZooKeys 379: 43–91. https://doi.org/10.3897/zookeys.379.6394
- de la Maza RG, de la Maza J, White-López A (1989) La fauna de mariposas de México. Parte I. Papilionoidea (Lepidoptera: Rhopalocera). Revista de la Sociedad Mexicana de Lepidopterología 12: 39–98.
- de la Maza J, White-López A, de la Maza RG (1991) La fauna de mariposas de México. Parte II. Hesperioidea (Lepidoptera: Rhopalocera). Revista de la Sociedad Mexicana de Lepidopterología 14: 3–44.
- DeVries PJ (1987) The Butterflies of Costa Rica and their Natural History. Papilionidae, Pieridae and Nymphalidae. Princeton University Press, Princeton, 327 pp.
- Dirzo R, Young HS, Mooney HA, Ceballos G (2011) Seasonally dry tropical forests: ecology and conservation. Island Press, Washington DC, 392 pp. https://doi.org/10.5822/978-1-61091-021-7
- Gotelli NJ, Colwell RK (2011) Estimating species richness. In: Magurran AE, McGill BJ (Eds) Biological diversity: frontiers in measurement and assessment. Oxford University Press, Oxford, 39–54.
- Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W (2004) Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly *Astraptes fulgerator*. Proceedings of the National Academy of Sciences of the United States of America 101(41): 14812–14817. https://doi.org/10.1073/pnas.0406166101
- Llorente-Bousquets J, Vargas-Fernández I, Luis-Martínez A, Trujano-Ortega M, Hernández-Mejía BC, Warren AD (2014) Biodiversidad de Lepidoptera en México. Revista Mexicana de Biodiversidad 85(Supplement 1): 353–371. https://doi.org/10.7550/rmb.31830
- Luis-Martínez MA, Salinas-Gutiérrez JL, Llorente-Bousquets J (2011) Papilionoidea y Hesperioidea (Lepidoptera: Rhopalocera). In: Álvarez F (Ed.) Chiapas: estudios sobre su diversidad biológica. Instituto de Biología, UNAM, México D.F., 363–391.
- Luis-Martínez A, Hernández-Mejía B, Trujano-Ortega M, Warren A, Salinas-Gutiérrez J, Ávalos-Hernández O, Vargas-Fernández I, Llorente-Bousquets J (2016) Avances faunísticos en los Papilionoidea sensu lato (Insecta: Lepidoptera) de Oaxaca, México. Southwestern Entomologist 41(1): 171–224. https://doi.org/10.3958/059.041.0119
- Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, Minchin PR, O'Hara RB, Solymos P, MH Stevens H, Szoecs E, Wagner H, Barbour M, Bedward M, Bolker B, Borcard D, Carvalho G, Chirico M, De Caceres M, Durand S, Evangelista HBA, FitzJohn R, Friendly M, Furneaux B, Hannigan G, Hill MO, Lahti L, McGlinn D, Ouellette M-H, Cunha ER, Smith T, Stier A, Ter Braak CJF, Weedon J (2020) Vegan: Community Ecology Package. R package version 2.5-7. https://CRAN.R-project.org/ package=vegan
- Pennington T, Lewis G, Ratter J (2006) Neotropical savannas and seasonally dry forests: plant diversity, biogeography, and conservation. CRC Press, Boca Raton, Florida, 484 pp. https://doi.org/10.1201/9781420004496
- R Development Core Team (2021) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna.

- Robbins RK (2010) The "upside down" systematics of hairstreak butterflies (Lycaenidae) that eat pineapple and other Bromeliaceae. Studies on Neotropical Fauna and Environment 45(1): 21–37. https://doi.org/10.1080/01650521003751712
- Salinas-Gutiérrez JL (2013) Registros nuevos y aclaratorios de ninfálidos (Papilionoidea: Nymphalidae) para Guatemala. Acta Zoológica Mexicana 29(2): 431–436. http://www.scielo. org.mx/scielo.php?script=sci\_arttext&pid=S0065-7372013000200015
- Salinas-Gutiérrez JL, Méndez C, Barrios M, Pozo C, Llorente-Bousquets J (2009) Hacia una síntesis de los Papilionoidea (Insecta: Lepidoptera) de Guatemala con una breve reseña histórica. Caldasia 31: 407–440. http://www.scielo.org.co/scielo.php?script=sci\_arttext& pid=S0366-52322009000200013
- Salinas-Gutiérrez JL, Llorente-Bousquets J, Méndez C, Barrios M, Pozo C (2012) Introducción a los Papilionoidea (Papilionidae, Pieridae, Lycaenidae, Riodinidae y Nymphalidae) de Guatemala. In: Cano EB, Schuster JC (Eds) Biodiversidad de Guatemala. Vol. II. Universidad del Valle de Guatemala, Guatemala, 155–173.
- Trujano-Ortega M, Callaghan CJ, Arellano-Covarrubias A, Luis-Martínez A, Avalos-Hernández O, Llorente-Bousquets J (2021) Geographical distribution of *Emesis* Fabricius (Lepidoptera: Riodinidae) in Mexico: Updated checklist and temporal patterns. Zootaxa 4964(3): 401–442. https://doi.org/10.11646/zootaxa.4964.3.1
- van Nieukerken EJ, Kaila L, Kitching IJ, Kristensen NP, Lees DC, Minet J, Mitter C, et al. (2011) Order Lepidoptera Linnaeus, 1758. In: Zhang ZQ (Ed.) Animal biodiversity: an outline of higher-level classification and survey of taxonomic richness. Zootaxa 3148: 212– 221. https://doi.org/10.11646/zootaxa.3148.1.41
- Warren AD, González-Cota L (1998) Notes on the genus *Piruna* in western Mexico, with description of a new species (Lepidoptera: Hesperiidae). Tropical Lepidoptera 9(Supplement 2): 1–7. https://journals.flvc.org/troplep/article/view/90137
- Warren AD, Davis KJ, Stangeland EM, Pelham JP, Willmott KR, Grishin NV (2017) Illustrated Lists of American Butterflies (North and South America). [21–XI–2017] http://www.butterfliesofamerica.com/
- Yoshimoto J, Salinas-Gutiérrez JL (2015) First record of *Atlides gaumeri* and notes on *Chalybs hassan* in Guatemala. Southwestern Entomologist 40(3): 497–502. https://doi. org/10.3958/059.040.0307
- Yoshimoto J, Salinas-Gutiérrez JL, Barrios M (2018) Annotated list of butterflies (Lepidoptera: Papilionoidea) of a Guatemalan dry forest, with two first records for Guatemala. Tropical Lepidoptera Research 28(1): 1–8. https://doi.org/10.5281/zenodo.1248159
- Yoshimoto J, Salinas-Gutiérrez JL, Barrios M (2019) Butterfly fauna and phenology in a dry forest of the Motagua Valley, Guatemala. Journal of the Lepidopterists Society 73(3): 191– 202. https://doi.org/10.18473/lepi.73i3.a8
- Yoshimoto J, Barrios M, Salinas-Gutiérrez JL, Warren AD (2021) Fauna y fenología de mariposas diurnas (Lepidoptera: Papilionoidea) de un bosque secundario en el área urbana de Guatemala. Revista Mexicana de Biodiversidad 92(0): e923469. https://doi.org/10.22201/ ib.20078706e.2021.92.3469
- Zhang J, Shen J, Cong Q, Grishin NV (2019) Genomic analysis of the tribe Emesidini (Lepidoptera: Riodinidae). Zootaxa 4668(4): zootaxa.4668.4.2. https://doi.org/10.11646/ zootaxa.4668.4.2

# Appendix I

**Table A1.** Butterfly species observed in 2011–2021 at two dry forests in Guatemala: Los Cerritos Municipal Park and Heloderma Natural Reserve, based on our previous studies (Yoshimoto et al. 2018, 2019) and on subsequent field surveys (July 2014 to August 2021 at Los Cerritos and October 2017 to November 2021 at Heloderma Reserve). Species and months in bold indicate the data newly obtained in the subsequent surveys. Year information is also shown with sampling months, when necessary. Nomenclature follows Warren et al. (2017).

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies		
Papili	onidae		
Papili	oninae		
1	Neographium epidaus epidaus (E. Doubleday, 1846) <sup>PH, A</sup>	Apr, May, <b>Jun</b> , <b>Jul</b> , <b>Aug</b> , <b>Nov</b>	_
2	Neographium philolaus philolaus (Boisduval, 1836) <sup>PH, A</sup>	Jun, Sep	Mar, Apr, May, Jun
3	Battus polydamas polydamas (Linnaeus, 1758) <sup>PH, A</sup>	Mar, Jul, <b>Aug</b> , Sep, <b>Dec</b>	_
4	Parides photinus (E. Doubleday, 1844)	_	Sep
5	Heraclides erostratus erostratus (Westwood, 1847) A, Y	May, Oct	_
6	Heraclides thoas autocles (Rothschild & Jordan, 1906) $^{\rm PH,A}$	Feb, <b>Mar</b> , Apr, <b>Jun</b> , <b>Aug</b> , <b>Nov</b>	-
7	Heraclides ornythion ornythion (Boisduval, 1836) PH	_	May, Jun
8	Heraclides rumiko Shiraiwa & Grishin, 2014 MI, PH	Jul	Oct'16 <sup>MI</sup> , Dec
9	Papilio polyxenes asterius Stoll, 1782 PH	Mar, Apr, May, <b>Nov</b>	_
Pierid	ae		
Colia	dinae		
10	Kricogonia lyside (Godart, 1819) <sup>PH, A</sup>	Mar	Jan, Feb, Mar, Apr, May, Jun, Jul, Aug, Sep, Nov, Dec
11	Eurema daira eugenia (Wallengren, 1860) <sup>PH, A, Y</sup>	Jan, Feb, Aug, Nov	Jan, Mar, Apr, May, Jun, Jul, Aug, Sep, Oct, Dec
12	Eurema boisduvaliana (C. Felder & R. Felder, 1865) PH, A	Feb, Oct, Nov	Jun, Jul, Aug, Sep, Oct, Nov
13	Abaeis nicippe (Cramer, 1779) MI, PH	Nov	Jul'16 <sup>MI</sup>
14	<i>Pyrisitia proterpia</i> (Fabricius, 1775) <sup>PH, A</sup>	May, Jun, Jul, Oct, <b>Dec</b>	Feb, Apr May, Jun, Jul, Aug, Sep, Oct, Nov
15	Pyrisitia dina westwoodi (Boisduval, 1836) PH, A	-	Jan, Feb, Jun, Oct, Nov, Dec
16	<i>Pyrisitia nise nelphe</i> (R. Felder, 1869) $PH, A, Y$	Jun, Jul, Aug, Nov	Jun, Jul, Aug, Sep, Oct, <b>Nov</b> , Dec
17	Zerene cesonia cesonia (Stoll, 1790) PH, Y	Jun, Aug	Jun, <b>Jul</b>
18	Anteos maerula (Fabricius, 1775) PH.A	Jun, Sep, Nov	<b>May</b> , Jun, Jul, Aug, Sep, Oct
19	Anteos clorinde (Godart, [1824]) PH, A	Apr, Jun	Jun, Aug
20	Phoebis sennae marcellina (Cramer, 1777) <sup>PH, A</sup>	Feb, <b>Mar</b> , Apr, Jun, Jul, Nov	Mar, May, <b>Jun</b> , Jul, Aug, Sep
21	Phoebis philea philea (Linnaeus, 1763) <sup>A, Y</sup>	May	Jul
22	Phoebis argante ssp. <sup>A</sup>	May	Jul
23	Aphrissa statira statira (Cramer, 1777) <sup>A</sup>	Oct	_
Pierin	ae		
24	Hesperocharis crocea crocea H. Bates, 1866	Mar, Aug	-
25	Ascia monuste monuste (Linnaeus, 1764) PH, A, Y	Feb, Jun	Jun

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies	-	
26	Ganyra josephina josepha (Salvin & Godman, 1868) PH, A	_	Jan, Oct
27	<i>Leptophobia aripa elodia</i> (Boisduval, 1836) <sup>Y</sup>	Jan	-
28	Itaballia demophile centralis Joicey & Talbot, 1928	-	Jan
29	<i>Glutophrissa drusilla tenuis</i> (Lamas, 1981) <sup>A</sup>	-	Jun, Aug
Lycaeı	nidae		
Thecli	nae		
30	Evenus regalis (Cramer, 1775) <sup>A</sup>	Sep	-
31	Atlides gaumeri (Godman 1901)	Aug	-
32	Atlides carpasia (Hewitson, 1868) <sup>A</sup>	Aug	-
33	Rekoa zebina (Hewitson, 1869)	Jun, Sep	-
34	Rekoa stagira (Hewitson, 1867) <sup>A</sup>	Aug	-
35	Arawacus sito (Boisduval, 1836) A, Y	Aug	-
36	Arawacus jada (Hewitson, 1867) <sup>A</sup>	Jul	-
37	Kolana lyde (Godman & Salvin, 1887) <sup>A</sup>	Sep	-
38	Chlorostrymon simaethis sarita (Skinner, 1895) <sup>A</sup>	Nov	-
39	Cyanophrys herodotus (Fabricius, 1793) <sup>A</sup>	Aug	-
40	Cyanophrys miserabilis (Clench, 1946)	-	Oct
41	Electrostrymon hugon (Godart, [1824])	_	Jul
42	Kisutam syllis (Godman & Salvin, 1887) <sup>A</sup>	_	Oct
43	Calycopis clarina (Hewitson, 1874)	Jun	-
44	Calycopis isobeon (A. Butler & H. Druce, 1872)	Aug, Sep	_
45	Strymon melinus franki W. D. Field, 1938	Aug, Sep	_
46	Strymon rufofusca (Hewitson, 1877) PH	Jul, Nov	Jan, Aug, Oct, Nov, Dec
47	Strymon bebrycia (Hewitson, 1868) PH	<b>Jun</b> , Aug	_
48	Strymon yojoa (Reakirt, [1867]) A	Jul	_
49	Strymon cestri (Reakirt, [1867]) <sup>A</sup>	Aug	-
50	Strymon bazochii bazochii (Godart, [1824]) <sup>A</sup>	Jul	_
51	Strymon istapa istapa (Reakirt, [1867])	Aug, <b>Nov</b>	-
52	Strymon megarus (Godart, [1824]) MI, PH	-	Jan, Jul, Oct'16 <sup>MI</sup> , Dec
53	Strymon ziba (Hewitson, 1868)	Jul	-
54	Ministrymon azia (Hewitson, 1873) A	Jun	Jul
55	Ostrinotes keila (Hewitson, 1869) A, Y	Aug	-
56	Panthiades bitias (Cramer, 1777) <sup>A</sup>	Jun	-
57	Michaelus hecate (Godman & Salvin, 1887)	Sep	-
58	Erora gabina (Godman & Salvin, 1887)	May, Jun, Aug, Oct	-
59	Chalybs hassan (Stoll, 1790)	Aug, Sep, Nov	Jul
Polyor	nmatinae		
60	Celastrina echo gozora (Boisduval, 1870) <sup>Y</sup>	Nov	-
61	Leptotes cassius cassidula (Boisduval, 1870) PH, A, Y	Jun, Dec	Sep, Oct
62	Cupido comyntas texana (F. Chermock, 1945) PH, A, Y	Sep, Nov	Oct, Nov, Dec
63	Hemiargus ceraunus astenidas (Lucas, 1857) <sup>A</sup>	Mar, <b>Jul</b> , Nov	Feb, Jun, <b>Dec</b>
64	Hemiargus hanno hanno (Stoll, 1790) <sup>PH, A</sup>	-	Jul, Aug, Sep, Oct
65	Echinargus isola (Reakirt, [1867])	Feb, Dec	Jun, Dec
Riodi	nidae		
Riodi	ninae		
66	Rhetus arcius castigatus Stichel, 1909 <sup>A</sup>	Sep	-
67	Calephelis spp. PH, 1	Jan, May, Jul, Aug,	Jan, Jul, Aug, Sep, Oct,
		Oct, Nov, Dec	Nov, Dec
68	<i>Lasaia sula sula</i> Staudinger, 1888 <sup>PH</sup>	-	Jun, Oct
69	Lasaia maria maria Clench, 1972	-	Jun, Jul, Oct
70	<i>Melanis pixe pixe</i> (Boisduval, 1836) <sup>A</sup>	Feb, Sep, Nov, Dec	-

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies	_	
71	Anteros carausius carausius Westwood, 1851 <sup>A</sup>	Aug, Nov	Sep, Nov
72	Calydna sturnula (Geyer, 1837) PH	Aug, Sep, Oct	-
73	<i>Emesis mandana</i> furor A. Butler & H. Druce, 1872 <sup>A</sup>	Aug	_
74	<i>Emesis tenedia</i> C. Felder & R. Felder, 1861 <sup>A, Y, 2</sup>	Iul	_
75	Emesis luping luping Godman & Salvin, 1886 <sup>2</sup>	Oct	_
76	Curvie emesia (Hewitson 1867) PH, A, 3	_	Jun Oct
77	Thishe lucarias (Hewitson, [1853]) A	Jun Jul Oct Nov	
78	Inditha caucana (Stichel 1911)		Oct
79	Sumarris mucana (Henritson 1865) A	Mar Jup Jul	Jul
80	Huboshulla counies a Boisduval 1836	iviai, juii, jui	Jui Ion
80 81	Theophysica Zearippa Boisduvai, 1830	—	Jan Eeb Nov
01 N	helider	-	red, nov
Nymp			
Libytr		T 1	T A C
82	Libytheana carinenta mexicana Michener, 1943	Jul	Jun, Aug, Sep
Danai			<b>N</b> 0
83	Lycorea halia atergatis E. Doubleday [1847] * PR. A. T	-	May, Sep
84	Danaus eresimus montezuma Talbot, 1943 Pri, A	Aug, Nov, Dec	Jun, Aug, <b>Sep</b>
85	Danaus gilippus thersippus (H. Bates, 1863) <sup>A</sup>	-	Mar
86	<i>Mechanitis lysimnia utemaia</i> Reakirt, 1866 <sup>PH, A, Y</sup>	-	May, Aug, Oct
87	<i>Mechanitis polymnia lycidice</i> H. Bates, 1864 <sup>PH, A, Y</sup>	Sep	Sep, Oct
88	Dircenna klugii klugii (Geyer, 1837) <sup>Y</sup>	Sep, Oct, Nov	-
Helico	oniinae		
89	Agraulis vanillae incarnata (N. Riley, 1926) <sup>PH, A</sup>	Jun, Jul, <b>Nov</b>	Aug
90	Dione moneta poeyii A. Butler, 1873 <sup>Y</sup>	Jun, Nov	-
91	Dione juno huascuma (Reakirt, 1866) PH, A	Feb, Mar, Dec	Jul
92	Dryas iulia moderata (N. Riley, 1926) <sup>PH, A, Y</sup>	Aug	Aug, Sep, Oct
93	<i>Eueides isabella eva</i> (Fabricius, 1793) <sup>A</sup>	Nov	-
94	Heliconius charithonia vazquezae W. Comstock & F.	Jul	Jul, Oct
	Brown, 1950 <sup>A, Y</sup>		
95	<i>Euptoieta hegesia meridiania</i> Stichel, 1938 <sup>PH, A, Y</sup>	Jun, Jul, Sep	Jun, Jul
Limen	itidinae	· · · •	
96	Adelpha paroeca paroeca (H. Bates, 1864) <sup>Y</sup>	Oct, Nov	_
97	Adelpha iphicleola iphicleola (H. Bates, 1864) PH	Aug	Jun, Jul, Sep, Oct
98	Adelpha melanthe (H. Bates, 1864)	Aug, Sep	-
Biblid	inae	0, 1	
99	Biblis hyperia aganisa Boisduval, 1836 PH, A	Jul. Dec	Sep
100	Mestra amymone (Ménétriés, 1857) <sup>A</sup>	May, Jun, Jul	Mar. Jul. Aug. Sep. Oct.
100		inay, juii, jui	Nov. Dec
101	Catonephele mexicana Jenkins & R.G. Maza 1985 A	Sep. Oct. Nov	
102	Eurica manima (Stoll 1782) PH	Jup Aug	Mar Jun Jul Aug Oct
102	Lunicu monimu (Stoll, 1762)	Juli, Aug	Dec
103	Funica tatila tatila (Herrich Schäffer [1855]) A	Iun	Dee
105	Hama duna atlantia atlantia (H. Batas, 1964) PH	Jun	- Iun Iul Nov
104	Hamadryas attantis attantis (11. Dates, 1804)	Sep	Juli, Jul, Nov
105	<i>Hamaaryas jeorua jerentina</i> (Godart, [1824])	Apr, May, Jun, Jul,	red, <b>Jui</b> , Oct
106	Hamadmuse alausonome alausonome (H. Botos, 1864) PH	Inov, Dec	Ion Esh Man Mary Ion
100	11umuar jus gunconome gunconome (11. Dates, 1804)	Jan, <b>Jun</b> , Oct, 1 <b>100</b> , Dec	Jul Aug Son Oct
107	Hamadmide augtom along augtom along (U Datos 1064)	May Ing Inl	Jui, Aug, Sep, Oct
10/	н, а	iviay, Jun, Jui	Jui
109	Rollangurg sulphis sulphis (H Barros 1964) PH	Jul Aug Son	Mar Jun Jul Son Oat
100	Existence adverte adverte Howisson 10(1)	Jui, Aug, Sep	iviai, juii, jui, sep, Oct
109	Epipinie aurasia aurasia riewitson, 1801	Aug, Oct	-

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies		
110	Temenis laothoe hondurensis Fruhstorfer, 1907 A	_	Oct
111	<i>Dynamine dyonis</i> Geyer, 1837 <sup>A</sup>	Jul, Aug, Oct, Nov, <b>Dec</b>	-
112	Dynamine postverta mexicana R.F. d'Almeida, 1952 PH, A	Jun, Jul, Oct, <b>Nov</b>	Sep, Oct
113	Dynamine theseus (C. Felder & R. Felder, 1861) A	Aug, Sep, Oct	_
114	Diaethria astala astala (Guérin-Méneville, [1844]) <sup>A, Y</sup>	May, Jul, Oct, Nov	Oct
Cyrest	inae		
115	<i>Marpesia petreus</i> ssp. <sup>A, Y</sup>	Jul, Sep	Jun
Nymp	halinae		
116	Historis odius dious Lamas, 1995 PH, A	Jun, Jul, Aug, Sep	Jul
117	Smyrna blomfildia datis Fruhstorfer, 1908 PH, A, Y	Jul	May, Nov
118	Anartia fatima fatima (Fabricius, 1793) PH, A, Y	May	Jun, Sep, Oct
119	Siproeta epaphus epaphus (Latreille, [1813]) <sup>A, Y</sup>	Sep	Sep
120	Siproeta stelenes biplagiata (Fruhstorfer, 1907) PH, A, Y	Jul, Sep	Jun, Aug, Sep, Oct
121	Junonia evarete (Cramer, 1779) PH, A	Jun, Jul, Aug	Jan, Jul, Oct
122	Chlosyne janais janais (Drury, 1782) <sup>A</sup>	Jun, Aug	-
123	Chlosyne erodyle erodyle (H. Bates, 1864) MI	Jul'12 <sup>MI</sup> , <b>Jul'19</b> , Oct'11 <sup>MI</sup> , <b>Oct'17</b>	_
124	Chlosyne rosita rosita A. Hall, 1924 MI	Sep'11 <sup>MI</sup>	Jun, Jul, Sep
125	Chlosyne theona theona (Ménétriés, 1855) PH	Apr, Jun, Sep	Jun, Jul, Aug, Sep
126	Chlosyne lacinia lacinia (Geyer, 1837) <sup>PH, A, Y</sup>	Mar, <b>Jun</b> , <b>Jul</b> , Aug, Nov	Jun, Jul, Aug
127	Chlosyne melanarge (H. Bates, 1864) PH	-	Aug, Sep, Oct
128	Microtia elva horni Rebel, 1906 PH	Jun, Jul, Aug, Nov	Jun, Jul, Aug, Sep, Oct, Nov
129	Anthanassa tulcis (H. Bates, 1864) MI, A	May'11 <sup>MI</sup>	Jun, Sep, Dec
130	Anthanassa ptolyca ptolyca (H. Bates, 1864) <sup>Y</sup>	Aug	Dec
131	Tegosa guatemalena (H. Bates, 1864) <sup>A</sup>	Feb, Nov	-
Chara	xinae		
132	Zaretis ellops (Ménétriés, 1855) <sup>A</sup>	Jul, Sep, Nov	_
133	Anaea aidea (Guérin-Méneville, [1844]) PH, A	May, Nov	Jun, Aug, Oct, Nov
134	Fountainea glycerium glycerium (E. Doubleday, [1849]) PH	Aug, Sep, Oct, Nov	_
135	Archaeoprepona demophon centralis (Fruhstorfer, 1905) * PH,A	-	Jul
Satyri	nae		
136	Morpho helenor ssp.* A	_	Sep
137	<i>Caligo telamonius memnon</i> (C. Felder & R. Felder, 1867) * PH, A	_	Oct
138	Manataria hercyna maculata (Hopffer, 1874) A, Y	_	Jun
139	Cissia similis (A. Butler, 1867) MI, PH, A	May'12 <sup>MI</sup> , Jun'12 <sup>MI</sup> , Oct, Nov	Jan, Feb, Apr, May, Jun, Oct, Nov, Dec
140	Cissia themis (A. Butler, 1867) MI, PH	Jul, Aug'11 <sup>MI</sup>	Feb, <b>Jun</b> , Jul, Aug'16 <sup>MI</sup> , Oct'16 <sup>MI</sup> , Dec
141	Cyllopsis gemma freemani (D. Stallings & J. Turner, 1947)	Sep, Nov	_
142	Cyllopsis hedemanni hedemanni R. Felder, 1869 <sup>Y</sup>	Feb	_
143	Cyllopsis hilaria (Godman, 1901)	Sep, Nov	_
144	Cyllopsis pephredo (Godman, 1901) Y	Jun, Nov	_
145	Euptychia fetna A. Butler, 1870	Aug. Sep	_
146	Hermeuptychia hermes (Fabricius, 1775) <sup>A, Y, 4</sup>	Jul	Ian, Feb. Sep. Oct
147	Taygetis thamyra (Cramer, 1779) PH	Nov	Jun, Oct

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies		
Hespe	riidae		
Eudan	ninae		
148	Phocides polybius lilea (Reakirt, [1867]) A	Nov	-
149	Phocides urania urania (Westwood, 1852)	Aug	-
150	Proteides mercurius mercurius (Fabricius, 1787) PH, A	Jun	Jun, Sep
151	<i>Epargyreus exadeus cruza</i> Evans, 1952 <sup>A, Y</sup>	Feb, Mar, Apr, Jun, <b>Jul</b>	Aug
152	Polygonus leo arizonensis (Skinner, 1911)	Jul	Jun, Jul, Aug, Sep, Oct
153	Chioides albofasciatus (Hewitson, 1867) <sup>A</sup>	Jun	_
154	Chioides zilpa (A. Butler, 1872) <sup>A</sup>	Jan, Mar	-
155	Typhedanus undulatus (Hewitson, 1867) <sup>A</sup>	Feb, Mar, May	-
156	Typhedanus ampyx (Godman & Salvin, 1893) <sup>A</sup>	-	Oct
157	Polythrix asine (Hewitson, 1867) PH, A, 5	_	Jan, Dec
158	Polythrix octomaculata (Sepp, [1844]) A	_	May
159	Cephise aelius (Plötz, 1880)	_	Oct
160	Codatractus alcaeus alcaeus (Hewitson, 1867)	Mar, May	_
161	Codatractus melon (Godman & Salvin, 1893)	_ ,	Jun, Jul
162	Urbanus viterboana (Ehrmann, 1907) <sup>MI, A, Y</sup>	Sep, Nov'11 <sup>MI</sup>	Sep, Oct
163	Urbanus esmeraldus (A. Butler, 1877) A, Y	Aug. Sep	Iun
164	Urbanus dorantes dorantes (Stoll, 1790) <sup>PH, A</sup>	May. Jul. Aug. Dec	Apr. Jun. Jul. Aug. Sep.
		,, <b>j</b> , <b>g</b> ,	Oct, Nov
165	Urbanus procne (Plötz, 1881) PH, A, Y	May, Jul, Nov	_
166	Urbanus doryssus doryssus (Swainson, 1831) <sup>A</sup>	Jul	-
167	Astraptes fulgerator azul (Reakirt, [1867]) A, Y, 6	Oct, Nov	_
168	Astraptes alector hopfferi (Plötz, 1881) A	Sep	Jan
169	Astraptes anaphus annetta Evans, 1952 PH, A, Y	Jun, Oct	Jul
170	Achalarus toxeus (Plötz, 1882) <sup>A</sup>	_	Mar, Apr, Oct
171	Achalarus albociliatus albociliatus (Mabille, 1877) <sup>A</sup>	Feb, Mar	Oct, Nov
172	Cabares potrillo potrillo (Lucas, 1857) A, Y	_	Jun, Jul, Aug, Oct, Nov
173	Cogia cajeta eluina Godman & Salvin, 1894	May, <b>Jun</b>	_
Pyrgin	lae	, <b>,</b> , <b>,</b> ,	
174	Mysoria affinis (Herrich-Schäffer, 1869)	-	Oct
175	Celaenorrhinus fritzgaertneri (Bailey, 1880) PH	Feb, Aug	Mar, <b>Jun</b>
176	Noctuana stator (Godman, 1899) PH, A, Y	Feb, Mar, May, Jul, Sep	_
177	Bolla evippe (Godman & Salvin, 1896)	Mar	-
(177)	Bolla sp. <sup>7</sup>	_	Oct 7
178	Staphylus ascalaphus (Staudinger, 1876) Y	May	Jan, Sep
179	Staphylus azteca (Scudder, 1872)	_	Feb, Aug, Nov
180	Gorgythion vox Evans, 1953 A, Y	_	Jun, Jul, Aug, Sep, Oct
181	Mylon salvia Evans, 1953	Nov	-
182	Mylon pelopidas (Fabricius, 1793)	_	May, Jun
183	Grais stigmaticus stigmaticus (Mabille, 1883) PH	_	Jan, Jun, Jul
184	<i>Timochares trifasciata trifasciata</i> (Hewitson, 1868) <sup>A</sup>	_	Jan
185	Chiomara georgina georgina (Reakirt, 1868)	Jan, <b>Jul</b> , Sep, <b>Nov</b>	Aug
186	Erynnis funeralis (Scudder & Burgess, 1870)	Aug	Jun
187	Eantis tamenund (W. H. Edwards, 1871) PH	Feb, Jul, Aug, Nov,	_
		Dec	
188	Atarnes sallei (C. Felder & R. Felder, 1867) A	Jul	_
189	Carrhenes fuscescens fuscescens (Mabille, 1891)	_	Jun
190	Antigonus erosus (Hübner, [1812]) PH, A	Mar, Oct	Feb, Mar, <b>Jun</b> , Aug, Oct
191	Antigonus corrosus Mabille, 1878 A, Y	-	Sep

Family		Months when observed	
	Subfamily	Los Cerritos	Heloderma Reserve
	Species and subspecies		
192	Zopyrion sandace Godman & Salvin, 1896	Mar, Aug	Jan, Apr, May, Jun, Sep,
			Dec
193	Pyrgus oileus (Linnaeus, 1767) <sup>PH, A, Y</sup>	Apr	Jul, Aug, Sep, Oct, Dec
194	Pyrgus orcus (Stoll, 1780)	-	Jun
195	Heliopyrgus domicella domicella (Erichson, [1849])	Sep	Sep, Oct, Nov
196	Heliopetes laviana laviana (Hewitson, 1868) <sup>A</sup>	_	Feb
197	Heliopetes macaira macaira (Reakirt, [1867]) <sup>MI, A</sup>	Jul'12 <sup>MI</sup> , <b>Nov</b>	Oct
198	Heliopetes alana (Reakirt, 1868) A, Y	Jul	-
Heter	opterinae		
199	Piruna aea (Dyar, 1912) PH	-	Jun <sup>8</sup> , <b>Jul</b> , Sep <sup>8</sup> , <b>Oct</b>
Hespe	riinae		
200	Perichares adela (Hewitson, 1867) <sup>A</sup>	Aug, Sep	Oct
201	Copaeodes aurantiaca (Hewitson, 1868) MI	Mar'11 <sup>MI</sup>	-
202	Panoquina lucas (Fabricius, 1793) A		Jan
203	Zenis jebus hemizona (Dyar, 1918)	Jan	-
204	Synapte shiva Evans, 1955	Aug	Jun, Dec
205	Synapte syraces (Godman, 1901)	-	Sep
206	Callimormus saturnus (Herrich-Schäffer, 1869) A	-	Oct
207	Amblyscirtes elissa elissa Godman, 1900 NR, MI	-	Jun, Aug'16 <sup>MI</sup> , Sep'16 <sup>MI</sup>
208	Amblyscirtes tolteca tolteca Scudder, 1872 <sup>A</sup>	<b>Jun</b> , Jul	May
209	Methionopsis ina (Plötz, 1882) <sup>A</sup>	-	Jan, Oct, Nov
210	Repens florus (Godman, 1900) NR	-	Oct
211	Cymaenes trebius (Mabille, 1891) MI, A	Aug	Sep'16 <sup>MI</sup>
212	Lerema liris Evans, 1955	-	Jul, Sep
213	Niconiades nikko Hayward, 1948 <sup>NR</sup>	Nov	-
214	Vettius fantasos (Cramer, 1780) A, Y	Aug, Sep, Oct	Jan, Oct
215	<i>Hylephila phyleus phyleus</i> (Drury, 1773) <sup>A</sup>	Aug	-
216	Polites vibex praeceps (Scudder, 1872) A	Mar, Apr	-
217	<b>Pompeius pompeius (Latreille, [1824])</b> A, Y	-	Jun
218	Atrytonopsis ovinia (Hewitson, 1866) PH	Jan, Feb, Mar, Apr, <b>Oct</b>	Oct, Dec

PH Species photographed.

<sup>A</sup> Species reported at Tikal in northern Guatemala by Austin et al. (1996).

<sup>Y</sup> Species reported at Parque Cayalá in Guatemala City by Yoshimoto et al. (2021).

\* Specimens not collected (recorded only by direct observation or photographs).

<sup>MI</sup> Species misidentified in Yoshimoto et al. (2018 or 2019); their previous identification results are shown in Table 1.

<sup>1</sup> Listed at the genus level because of the difficulty of species-level identification derived from the confused taxonomic state of this genus.

<sup>2</sup> These two species, together with *Emesis tegula* and *E. toltec*, can be treated as a species complex. These taxa require further study, according to Trujano-Ortega et al. (2021).

<sup>3</sup> The genus name, treated as *Emesis* in Yoshimoto et al. (2019), was modified according to Zhang et al. (2019).

<sup>4</sup> The name "*hermes*" is correctly applied to a South American species, according to Cong and Grishin (2014). Thus, the individuals collected might include multiple species, none of which are true *hermes*.

<sup>5</sup> One sample which was photographed at Heloderma Reserve in October 2016 and was identified as this species in Yoshimoto et al. (2019) was excluded, as we found that it might have been of another species of this genus, which is unable to de determined because of the lack of a specimen.

<sup>6</sup> This is a species complex, which includes several species in Costa Rica (Hebert et al. 2004; Brower, 2006, 2010). Thus, the individuals collected might also be of multiple species.

<sup>7</sup> These data were included in the species count and Chao II analyses for Heloderma Reserve, since at this site none of the identified species of this genus had been recorded. In contrast, these data have been excluded from the Chao II analysis for all data (pooled across the sites) and from the Jaccard index analyses, as this individual might be of *Bolla evippe* (Godman & Salvin, 1896), which is unable to be examined because of the heavily damaged specimen of *Bolla* sp. <sup>8</sup> Identified to genus (*Piruna* sp.1) and incorrectly listed as Hesperiinae in Yoshimoto et al. (2019).

NR New record for Guatemala.