

Systematic revision of *Stegodera* Martens, 1876 (Gastropoda, Stylommatophora, Camaenidae), with description of a new genus

Zhe-Yu Chen^{1,2}, Zhi-Tong Lyu³, Min Wu²

1 College of food science and engineering, Wuhan Polytechnic University, Wuhan 430023, China **2** School of Life Sciences, Nanjing University, Nanjing 210023, China **3** The Museum of Biology, School of Life Sciences, Sun Yat-sen University, Guangzhou 510275, China

Corresponding author: Min Wu (minwu1969@aliyun.com)

Academic editor: Frank Köhler | Received 7 May 2021 | Accepted 8 August 2021 | Published 1 September 2021

<http://zoobank.org/650D7F4C-990E-4CBF-88E3-F0CD38EEC377>

Citation: Chen Z-Y, Lyu Z-T, Wu M (2021) Systematic revision of *Stegodera* Martens, 1876 (Gastropoda, Stylommatophora, Camaenidae), with description of a new genus. ZooKeys 1059: 1–21. <https://doi.org/10.3897/zookeys.1059.68385>

Abstract

The monotypic genus *Stegodera* Martens, 1876 is systematically revised based on anatomical and morphological examination of freshly collected specimens. A new species from southern Hunan, which resembles *Stegodera angusticollis*, is confirmed to represent a new genus evidenced by comparative shell morphology and anatomy as well as by molecular phylogenetic analyses. The new genus might be more closely related to *Stegodera* and *Nesiohelix* Kuroda & Emura, but differs anatomically from the latter two genera by the absence of a dart apparatus.

Keywords

16S rRNA, Bradybaeninae, Camaeninae, China, *COI*, new genus, new species

Introduction

Stegodera Martens, 1876 is a monotypic camaenid genus endemic to southern Hubei and northern Jiangxi, China (Zilch 1960; Richardson 1983). Since the original description (Martens 1875a), only a few additional specimens of this species have been reported (Heude 1882; Chen and Gao 1987; Qian et al. 2008; Qian and Zhou 2014). The typical shell character states of *Stegodera* are “shell sinistral, disk-shaped, with low spire and

open, deep umbilicus; solid, opaque, brown. Inner whorls slowly increasing, regular; latter half of the last whorl distorted, straightened, covering the preceding whorl above. Aperture very oblique, crescentic, toothless; peristome reflexed; throat very much contracted” (Pilsbry 1895), while the anatomy of this genus kept unknown. Recently, we received several fresh specimens of *Stegodera angusticollis* (Martens, 1875) from southeastern Hubei. In another recent field investigation in southern Hunan, Dr Lu Qiu found two land snail specimens belonging to a species, which is conchologically similar to *Stegodera angusticollis*, but has been found to differ from it in anatomical characteristics.

Materials and methods

A piece of foot tissue was cut from the living animal and preserved in 99.7% ethanol for molecular analysis. Then the animal was relaxed by drowning in water before being transferred to 70% ethanol for fixation, which was replaced with ethanol of the same concentration after three days. Photographs of the shell and reproductive system were taken using a Canon camera with Macro lens, and then modified in Adobe Photoshop CC 2018. The shells were measured with digital vernier calipers to the nearest 0.1 mm. Measurement abbreviations:

$S_{D_{maj}}$ shell major diameter;
 $S_{D_{min}}$ shell minor diameter;
 S_H Shell height.

Whorls were counted as described by Kerney and Cameron (1979). Directions used in descriptions: proximal, towards the genital atrium; distal, away from the genital atrium.

Molecular phylogenetics

Muscle tissue was obtained from eleven species in this study (Table 1), including *Stegodera angusticollis* and the single paratype of *Pseudostegodera qiului* gen. et sp. nov. Genomic DNA was extracted by using Tiangen DNA Extraction Kit (for SYS samples. Abbreviations see below) and TIANamp Marine Animals DNA Kit (for HBUMM samples). Two mitochondrial genes, partial 16S ribosomal RNA gene (16S) and partial cytochrome *C* oxidase 1 gene (CO1), were amplified. Primers used for 16S were 16SA/16SB (Palumbi et al. 1991), and for CO1 were LCO1490/HCO2198 (Folmer et al. 1994). PCR amplifications were performed in a 20 μ l (for SYS samples) / 50 μ l (for HBUMM samples) reaction volume with the cycling conditions of an initial denaturing step at 94° C for 2 min, 35 cycles of denaturing at 94° C for 30 s, annealing at 58° C (for 16S)/ 50° C (for CO1) for 30 s and extending at 72° C for 30 s, and final extending step of 72° C for 10 min. PCR amplicons were inspected on a 1% agarose gel for quality and fragment size, then were purified, and sequenced on an automated sequencer.

For phylogenetic analysis, sequences from seven camaenid species and one out-group species were obtained from GenBank and incorporated into our dataset

Table 1. Vouchers and the GenBank accession numbers of the species for phylogenetic study (*, from NCBI).

Species	Subfamily	16S/CO1	Museum voucher	Voucher inf.
<i>Pseudostegoderia qiului</i> gen. et sp. nov.	Camaeninae	MW810083/ MW810790	SYS m001017	Paratype of the species; see text
<i>Stegoderia angusticollis</i>	Camaeninae	MW810079/ MW810793	SYS m001016	See text
<i>Amphidromus inversus</i>	Camaeninae	AB112400*/ FJ472655*	/	/
<i>Camaena cicatricosa</i>	Camaeninae	KU586474*/ KU061276*	/	/
<i>Camaena poyuensis</i>	Camaeninae	KU586468*/ KU061273*	/	/
<i>Satsuma guandi</i>	Camaeninae	MW804648/ MW810791 MW804647/ MW810792	HBUMM08239a1 HBUMM08239a2	Guangdong, Shaoguan, coll. Di Yu
<i>Exiligada gregoriana</i>	Camaeninae	JX393672*/ JX393761*	/	/
<i>Falspleuroxia overlanderensis</i>	Camaeninae	KU519178*/ KU519261*	/	/
<i>Tatemelon musgum</i>	Camaeninae	KU519194*/ KU519277*	/	/
<i>Sinumelon vagente</i>	Camaeninae	KP965282*/ KP965358*	/	/
<i>Acusta ravidia</i>	Bradybaeninae	MW800197/ MW810782	HBUMM06616a	Sichuan, Jiuzhaigou, coll. Min Wu
<i>Bradybaena qixiaensis</i>	Bradybaeninae	MW810081/ MW810783	HBUMM06900–1/ HBUMM06900–2	Jiangsu, Nanjing, coll. Min Wu
<i>Cathaica fasciola</i>	Bradybaeninae	MW800200/ MW810784	HBUMM06477–1/ HBUMM06477–2	Jiangsu, Zhenjiang, coll. Min Wu
<i>Coccolypta liui</i>	Bradybaeninae	MK680922*/ MK680001*	/	/
<i>Coccolypta pinchoniana</i>	Bradybaeninae	MK680923*/ MK680002*	/	/
<i>Dolicheulota formosensis</i>	Bradybaeninae	KR338956*/ KR338956*	/	/
<i>Euhadra dixonii</i>	Bradybaeninae	AF098711*/ AB916773*	/	/
<i>Laeocathaica polytyla</i>	Bradybaeninae	MW810082/ MW810787	HBUMM06726a1	Sichuan, Jiuzhaigou, coll. Min Wu
<i>Laeocathaica distinguenda</i>	Bradybaeninae	MW810084/ MW810785 MW810085/ MW810786	HBUMM06491a1 HBUMM06491a2	Gansu, Wenxian, coll. Min Wu
<i>Nesiobelx moreletiana</i>	Bradybaeninae	MW810080/ MW810788	HBUMM06796	Zhejiang, Hangzhou, coll. Min Wu
<i>Pseudobuliminus piligerus</i>	Bradybaeninae	MW800362/ MW810789	HBUMM06527a1	Gansu, Wenxian, Coll. Min Wu
<i>Cornu aspersum</i> (out-group)	Helicidae	KU586459*/ KU586502*	/	/

(Table 1). *Cornu aspersum* is used as the out-group of the studied Camaenidae, following the previous studies (Wang, Xiao et al. 2014; Wang, Yang et al. 2014; Huang et al. 2015) considering that the branch including Helicidae is a close group of Bradybaenidae + Camaenidae (Asian and Australasian) + Polygyridae (Wade et al. 2007; Razkin et al. 2015). DNA sequences of the two genes were aligned respectively by the Clustal W algorithm with default parameters (Thompson et al. 1997) in MEGA 7.0.26 (Kumar et al. 2015). The substitution saturation assessment for CO1 genes was done using DAMBE 7.0 (Xia 2018) employing the methods introduced by Xia et al. (2003) and Xia and Lemey (2009). The poorly aligned positions and divergent regions of the alignment of 16S were eliminated using Gblocks 0.91b (Castresana 2000), a concatenated matrix of 20 (including outgroup, Table 1)×703 bp was used for the subsequent analyses. Model selection was performed by “Models” in MEGA 7.0.26 (Kumar et al. 2015). The data set was analyzed using Bayesian Inference (BI) in MrBayes 3.2.4 (Ronquist et al. 2012) and the Maximum Likelihood analysis in raxmlGUI 2.0 beta (Edler et al. 2019) (Fig. 1). In Bayesian Inference analysis, three independent runs were conducted, each of which was performed for 10,000,000 generations and sampled every 1000 generations with the first 25% samples discarded as burn-in. Convergence of the Markov Chain Monte Carlo simulations was assessed using Tracer v1.7 (Rambaut et al. 2018), verifying that all ESS values exceeded 200. In addition, we repeated these analyses for a dataset containing a larger number of species (Table 1) (Fig. 1B).

We used Mesquite v. 3.61 (Maddison and Maddison 2019) to reconstruct the evolutionary history of five reproductive characters, which are usually emphasized in camaenid taxonomy, and mapped these characters on the Bayesian phylogeny under Maximum Likelihood-based criterion.

Depository abbreviations

HBUMM	Mollusk collection of Museum of Hebei University, Baoding, China;
IZCAS	Institute of Zoology, Chinese Academy of Sciences (Beijing, China);
MYNU	Mianyang Normal University (Mianyang, China);
OYKC	Private collection of Kai-Chen Ouyang (Kunming, China);
QL	Private collection of Lu Qiu (Luzhou, China);
SMF	Senckenberg Forschungsinstitut und Naturmuseum (Frankfurt am Main, Germany);
SYS	Sun Yat-sen University (Guangzhou, China);
ZMB	Museum für Naturkunde (Berlin, Germany).

Anatomical abbreviations

AS	accessory sac;
At	atrium;
BC	bursa copulatrix;
BCD	bursa copulatrix duct;
DS	dart sac;
DSC	dart sac chamber;
Ep	epiphallus;
Fl	flagellum;
FO	free oviduct;
MG	mucous glands;
MGP	papilla distally leading to mucous glands on inner wall of accessory sac;
P	penis;
PC	penial caecum;
PR	penial retractor muscle;
PS	penis sheath;
Va	vagina; V
VD	vas deferens.

Results

Phylogeny

The substitution saturation assessment for CO1 sequences suggested that the first and second codon positions were relatively conserved but that the third codon po-

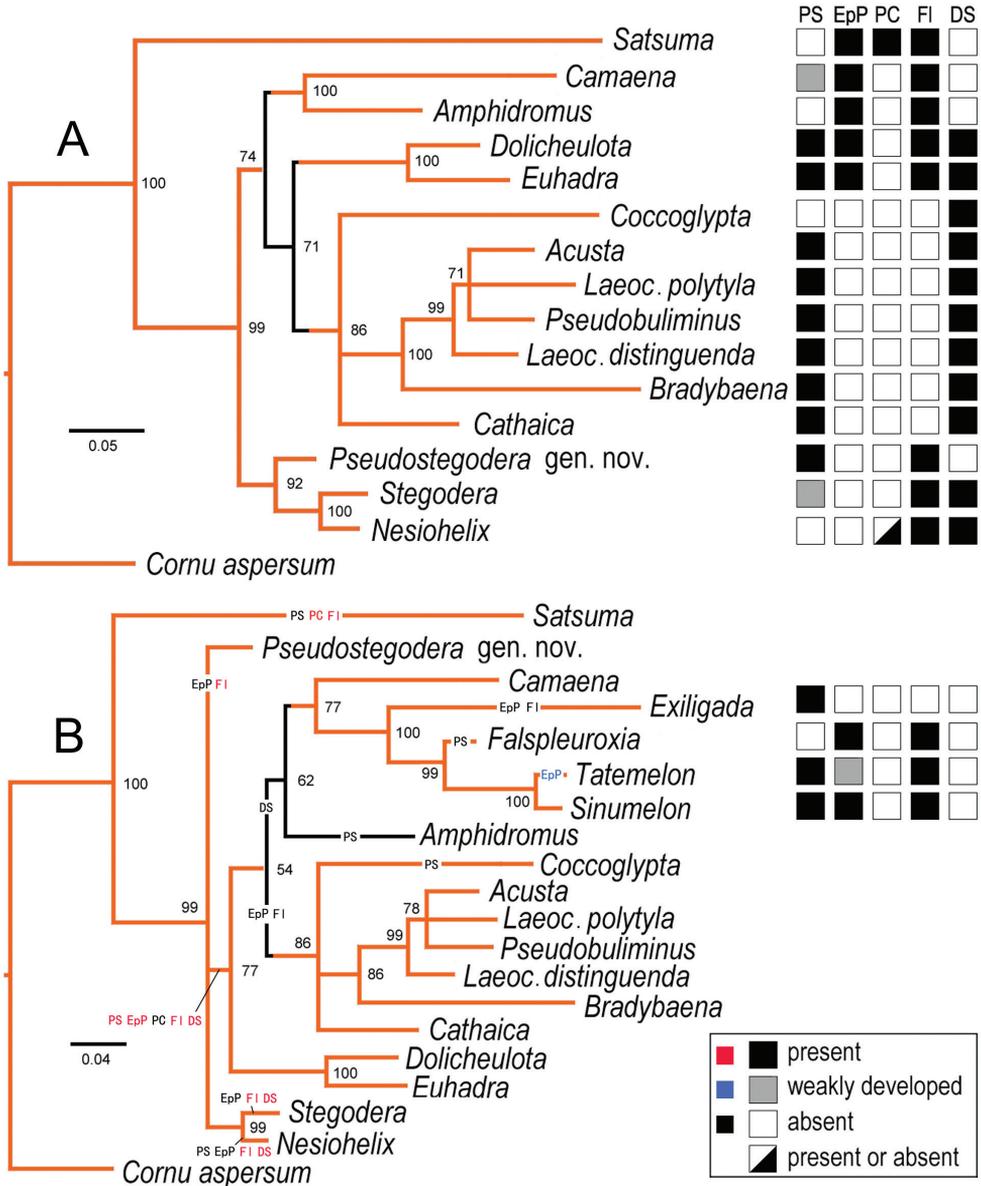


Figure 1. Bayesian Inferences of camaenids (representatives of genera in Table 1) based on partial mitochondrial 16S and CO1 sequences. Numbers near nodes indicate Bayesian posterior probabilities. The squares on the right of each taxon name indicate its character states. Abbreviations: PS: penial sheath; EpP: epiphallallic papilla; PC: penial caecum; FI: flagellum; DS: dart sac apparatus **A** phylogram without Australian camaenids **B** the results of ancestral states (tagged on branch in red, blue, or black) reconstruction mapping on the phylogram with some Australian camaenid genera added. Orange part: the topologies supported by the results respectively by using both Bayesian Inference and Maximum Likelihood methods. Character states are from Schileyko (2003) and Wu (2019). Scale bars for substitutions per site.

sitions revealed sequence saturation and are therefore not suitable for phylogenetic inference (for both symmetrical and asymmetrical trees, $Iss > Iss.c$, $ps < 0.001$). Our final molecular dataset contained nineteen sequences of partial 16S and CO1 genes. After eliminating poorly aligned positions and divergent regions of the alignment of 16S using Gblocks 0.91b (Castresana 2000), a concatenated matrix of 20 (including outgroup, Table 1)×703 bp was used for the subsequent analyses. The models “T92 + G” and “GTR + G + I” were chosen as the best nucleotide substitution models for 16S (lnL = -1877.8, BIC = 4100.7) and CO1 (lnL = -1399.1, BIC = 3216.9), respectively. The model of the combined dataset is “TN93 + G + I” (lnL=-3387.6, BIC = 7192.6).

The phylograms produced by both Maximum Likelihood Inference and Bayesian Inference based on partial 16S + partial CO1 sequences are topologically identical in major branches (Fig. 1). All trees reveal a primary division (posterior probability PP = 1) between penial caecum-bearing *Satsuma* A. Adams, 1868 (Camaeninae) and the remaining studied groups, a mixture of so-called camaenine and bradybaenine genera that usually have no penial caecum. The monophyly of Bradybaeninae is not supported because neither *Nesiohelix* Kuroda & Emura, 1943 nor *Dolicheulota* Pilsbry, 1901, and *Euhadra* Pilsbry, 1890, all well-known bradybaenine genera, are not included in the clade where most bradybaenine genera stay. The monophyly of Camaeninae, which is morphologically characterized by the absence of dart sac apparatus is also not supported when considering that *Satsuma* is situated most basally on the phylograms and meanwhile *Camaena* Albers, 1850 and its sister group *Amphidromus* Albers, 1850 are deeply nested in the clade that comprises most bradybaenine genera. *Stegoderma* E. Martens, 1876 is found to be the sister group of *Nesiohelix* with strong nodal support (PP = 1). In one phylogram (Fig. 1A), *Pseudostegoderma* gen. nov. is a sister group of clade *Stegoderma* + *Nesiohelix* and in the other, it is in an unresolved trichotomy with these two taxa.

As suggested by the analyses of character evolution, the ancestral character states among most studied camaenids, except *Satsuma*, *Pseudostegoderma* gen. nov., *Stegoderma*, and *Nesiohelix*, are: penial sheath, epiphallic papilla, flagellum and dart sac apparatus present, penial caecum absent (Fig. 1B). The penial sheath has been lost in all the studied taxa at least for five times. The epiphallic papilla has been lost at least for four times but regained once in *Tatemelon*. Amongst the ingroup, the dart sac was acquired for three times but lost in *Camaena*, *Amphidromus* and the Australian camaenids. The flagellum has been lost twice, once in *Exiligada* Iredale, 1939, and once in the clade including *Coccolypta* Pilsbry, 1895, *Acusta* Martens, 1860, *Laeocathaica* Moellendorff, 1899, *Pseudobuliminus* Gredler, 1887, *Bradybaena* Beck, 1837, and *Cathaica* Moellendorff, 1884, which are all bradybaenine genera mainly distributed in the mainland of China.

Systematics

Family Camaenidae Pilsbry, 1895

Stegoderma Martens, 1876

盖螺属

Helix (*Stegodera*) Martens in Pfeiffer 1876: 150; Schmacker and Boettger 1894: 173.
Stegodera. – Pilsbry 1905: 64, Yen 1939: 126; Zilch 1960: 610; Schileyko 2003: 1512.
Steganodera Kobelt, 1879: 236 (incorrect subsequent spelling or unjustified emendation); Schileyko 2003: 1512 (syn pro *Stegodera* Martens, 1876).
Plectopylis (*Stegodera*). – Pilsbry 1894: 147.
Planispira (*Stegodera*). – Thiele 1931: 681.

Type species. *Helix angusticollis* Martens, 1875, by original designation.

Diagnosis. Shell sinistral. Apical whorls with dense fine ribs that gradually becoming granules. The last $\frac{1}{4}$ body whorl compressed, apically covering the contacted penultimate whorl. Peristome expanded and slightly reflexed. Head wart absent. Each side of mantle edge with a leaf-shaped appendage. Penis sheath weakly present. Penis externally simple. Epiphallic papilla absent. Flagellum present. Dart sac apparatus present. Accessory sac well developed. Mucous glands with numerous gland tubes. Membranous sac surrounding terminal genitalia absent. Poly-layered structure in dart sac and/or accessory sac absent.

Remarks. This genus was considered as a subgenus of *Helix* Linnaeus, 1758 or *Plectopylis* Benson, 1860 for some time (Martens 1876; Schmacker and Boettger 1894; Pilsbry 1894). Pilsbry (1905) formally established its independent status. *Stegodera* and *Traumatophora* Ancey, 1887 were considered to be closely related, and *Traumatophora* was once treated as a subgenus of *Stegodera* (Pilsbry 1905; Gude 1920). Pilsbry (1905) believed that the two shallow grooves in the throat area of *Stegodera* are likely to be homologous to the dentition in *Traumatophora*.

In light of the genital system, *Stegodera* shares with *Nesiohelix* the most important character states like the presences of flagellum, numerous tubes of mucous glands and the papilla distally leading to mucous glands on the inner wall of the accessory sac, and the absences of the epiphallic papilla, poly-layered structure in dart sac apparatus and membranous sac surrounding terminal genitalia (Wu 2019). The close relationship between them in genitalia is supported by the phylogenetic result of this work (Fig. 1A, B).

***Stegodera angusticollis* (Martens, 1875)**

狭缘盖螺

Figures 1, 2, 3A, 4A–D, 5A–C, 6A, B, 8A–C

Helix angusticollis Martens, 1875a: 2. Martens 1875b: 185; Pfeiffer 1875: 449, 1876: 149, pl. 134, figs 7–10; Gredler 1882: 175; Heude 1882: 36, pl. 15, fig. 8; Möllendorff 1884: 387.

Helix (*Stegodera*) *angusticollis*. – Martens, 1876 (in Pfeiffer 1870–1876): 149–150; Pilsbry 1890: 7, pl. 1, figs 15–17.

Plectopylis (*Stegodera*) *angusticollis*. – Pilsbry 1894: 147.

Planispira (Stegodera) angusticollis. – Thiele 1931: 681.

Stegodera angusticollis. – Gude 1902: 4; Kobelt 1905: 78; Pilsbry 1905: 64, 66, pl. 2, figs 1–3; Gude 1920: 60; Yen 1939: 126, pl. 13, fig. 9; Zilch 1960: fig. 2138; Richardson 1983: 291; Schileyko 2003: 1512, fig. 1948; Chen and Gao 1987: 107, fig. 137; Qian et al. 2008: 289, fig. 169; Qian and Zhou 2014: 120, figure in text.

Museum material examined. •ZMB. Moll. 31044, syntype, Poyangsee, China, slg. v. Richthofen; ZMB. •ZMB. Moll. 3710/1 China, slg. Paetel; •SMF27113/3, China: Prov. Hupei, slg. O. v. Moellendorff; •SMF27114/2, China: Wü-chang-fú, slg. K. Hashagen (ex. Schmacker); •SMF42579/3, China: Húbei (Hupei), slg. Ehrmann; •IZ-CAS TM108733/1, Kieou-Kiang; •IZCAS TM158903-158920/18, Ou-tchang h[ien], ex. Musée Heude; •IZCAS TM159272-159397/126, Kieou-Kiang, ex. Musée Heude.

New material examined. •One shell of HBUMM08435 (dissected), Guanyin Cave [观音洞], Taizi Town [太子镇], Yangxin County [阳新县], Huangshi City [黄石市], Hubei Province, China, 30°0'3.816"N, 115°11'24.428"E, 140 m a.s.l., 2020-X-15, leg. Xiao-Long Wang & Zi-Hao Shen ($S_{D_{maj}}$ = 26.7 mm, $S_{D_{min}}$ = 21.3 mm, S_H = 11.3 mm). •Three shells of OYKC, one shell of QL, Lushui Lake [陆水湖], Chibi City [赤壁市], Xianning City [咸宁市], Hubei Province, China, 29°40'12"N, 113°58'35"E, 2020-X-03, leg. Di Yu & Kai-Chen Ouyang.

Type locality. Poyang-See (= Poyang Lake [鄱阳湖], Jiangxi).

Measurements of new material. $S_{D_{maj}}$ = 26.7–30.6 mm, $S_{D_{min}}$ = 21.3–23.7 mm, S_H = 11.3–12.2 mm (n = 6).

Diagnosis. Body whorl completely covering partial penultimate whorl. Penial sheath weakly present. Epiphallic papilla absent. Dart sac apparatus and flagellum present. Mucous glands with numerous tubes.

Redescription. Shell (Fig. 3A). Sinistral, large, solid, rather flat, five whorls, in chestnut and darker near aperture. Suture impressed. Protoconch $1\frac{1}{4}$ whorls. After $\sim 3\frac{1}{2}$ whorls, growth lines broken into regularly arranged tubercles. Body whorl compressed from the last $\frac{1}{4}$ to the last $\frac{1}{8}$ whorls, completely covering corresponding part of penultimate whorl including the suture. After the last $\frac{1}{8}$ whorls, body whorl becoming as broad as normal again. On the above compressed region, one shorter ventral depression and one longer apical depression present. Aperture semilunar, slightly descending. Peristome chestnut, thickened, expanded, and slightly reflexed. Umbilicus moderately broad; approximately $\frac{1}{5}$ of shell major diameter. Protoconch visible through umbilicus.

General anatomy (Fig. 4A–D). Head wart absent. On internal body wall of head region between ommatophore insertions with tiny pits rather than glands (Fig. 4C). Each side of mantle edge with a leaf-shaped appendage (Fig. 4A, B). Body reddish brown, central dorsa with light longitudinal stripes. Sole dirty white. Jaw arcuate; with 12 more or less projecting ribs (Fig. 4D).

Genitalia (Figs 5A–C, 6A, B). Penis sheath present but thin and very short. Penis medially slightly thickened, moderately long, ~ 8.8 mm, externally simple. Inside penis with several thin longitudinal pilasters. Epiphallus slightly longer than penis. A

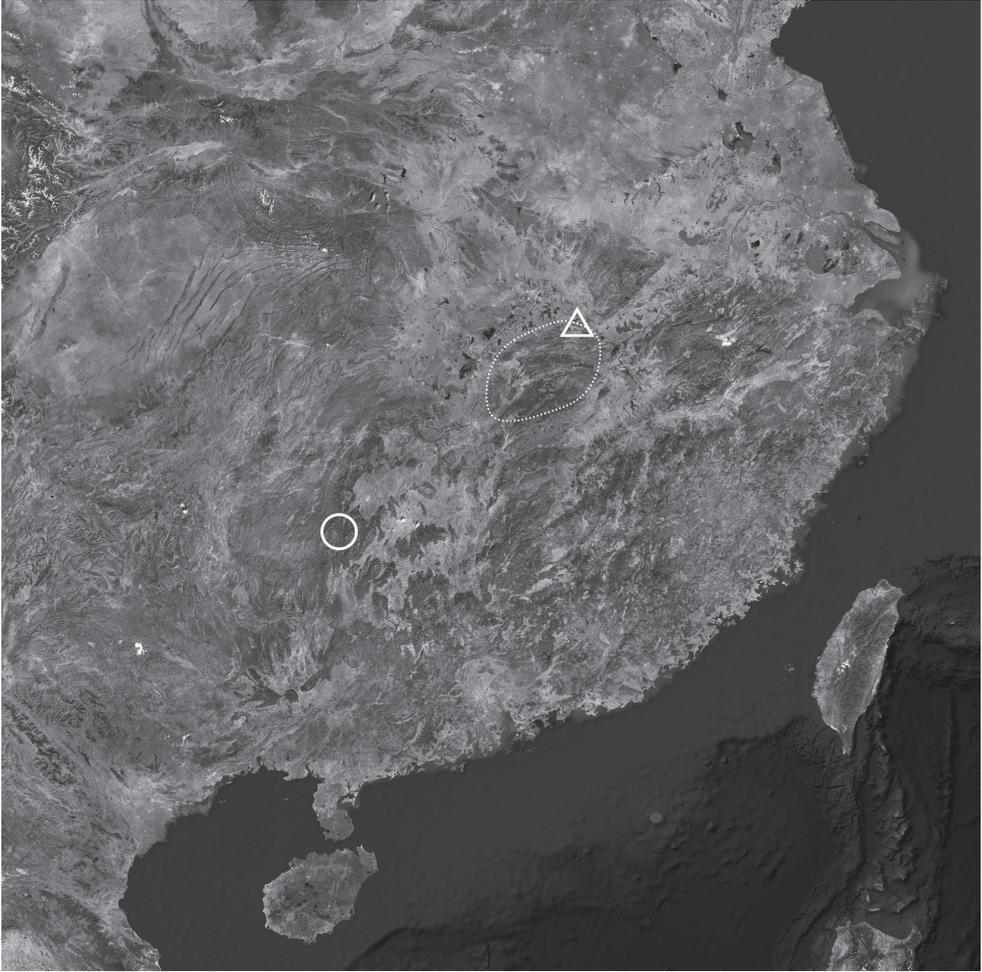


Figure 2. Distribution of *Stegoderia* and *Pseudostegoderia qiului* gen. et sp. nov. Triangle: new locality of *Stegoderia angusticollis* (Martens, 1875); dashed range: area of *Stegoderia angusticollis*; circle: type locality of *Pseudostegoderia qiului* gen. et sp. nov.

very thin membranous sac wrapping distal half of penis with distal end connecting basal penial retractor muscle (Fig. 6B). Flagellum cylindrical, tapering. Vas deferens thin throughout. Dart sac apparatus distally inserting on vagina. Dart sac small. Dart not observed. Membranous sac surrounding terminal genitalia absent. Accessory sac relatively large. Tubes of mucous glands more than twenty, neatly inserting in a single row on distal and same side of dart sac on accessory sac (Figs 5C, 6B). From proximal to distal accessory sac, mucous gland tubes increased gradually in length (Figs 5C, 6B). Papilla distally leading to mucous glands on inner wall of accessory sac integrated into one long thick spongy pilaster on the side of mucous gland tube insertion (Fig. 6A). Bursa copulatrix ball-shaped.

Ecology. This species was observed under litter layer, on nearby rocks and crevices.

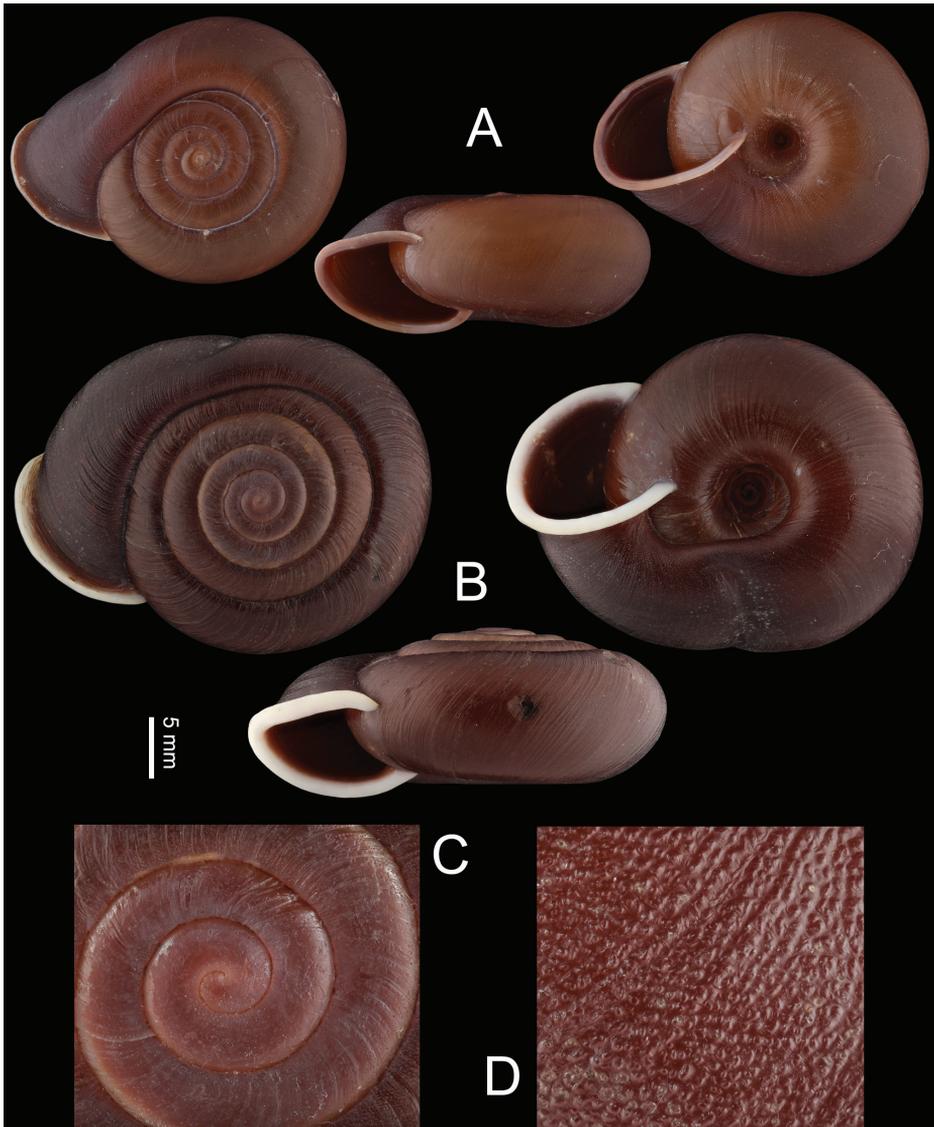


Figure 3. Shells **A** *Stegodera angusticollis* Martens, 1876, HBUMM08435 **B–D** *Pseudostegodera qiului* gen. et sp. nov., holotype IZCAS TM206978 **C** p rotoconch, magnified **D** shell surface, magnified. Scale bar: 5 mm (**A, B**).

Distribution. Central China: Hubei, Jiangxi (Fig. 2).

Remarks. The peristome of *Stegodera angusticollis* is in chestnut in fresh specimens, but in most museum specimens this color gets faint to white. In *Stegodera angusticollis*, the compressed part of body whorl usually completely cover the penultimate whorl, while in *Pseudostegodera qiului* gen. et sp. nov. the corresponding part

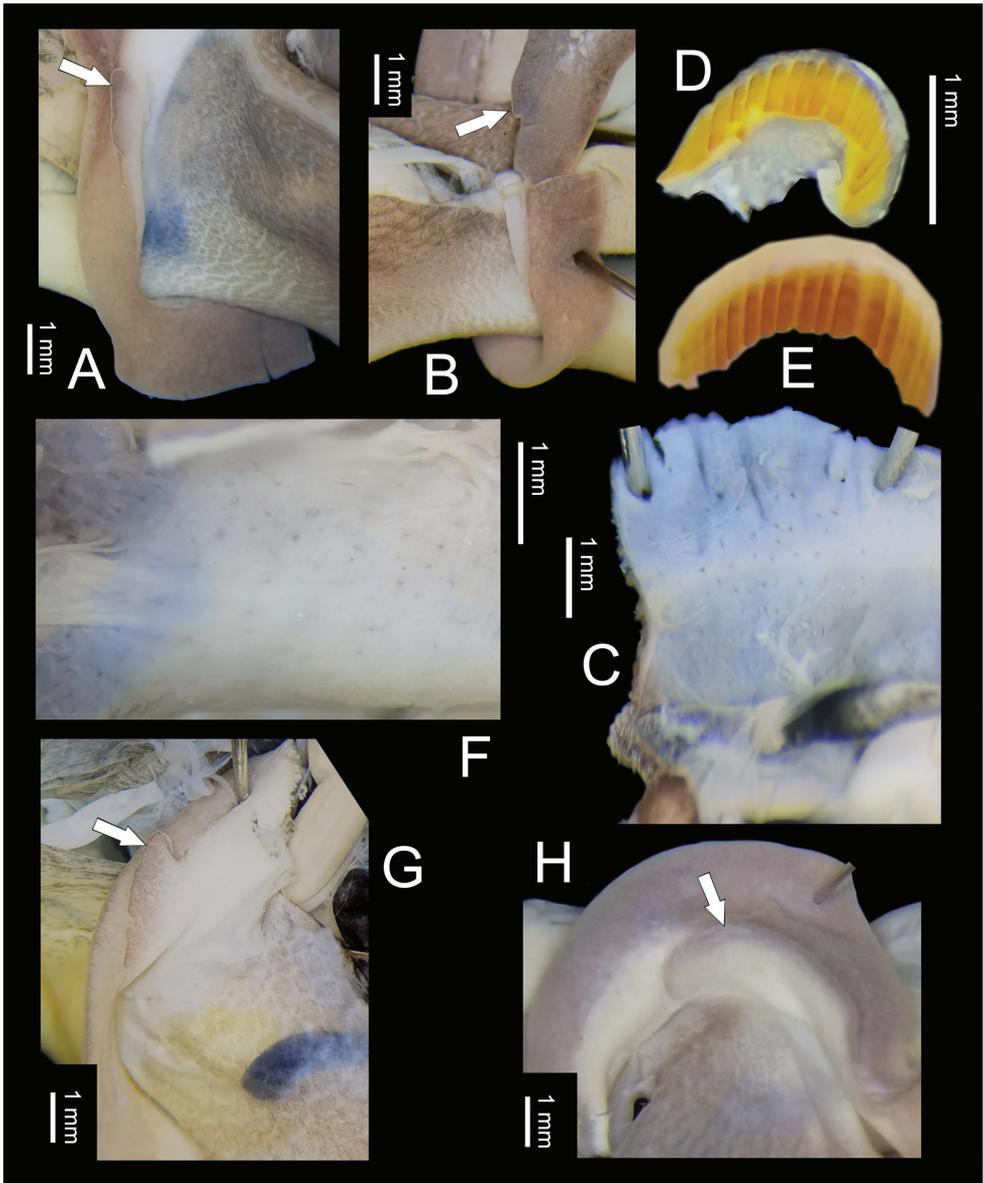


Figure 4. General anatomy **A–D** *Stegodera angusticollis* (Martens, 1875), HBUMM08435 **A** lobe (arrowed) on the left side of mantle edge **B** lobe (arrowed) on the right side of mantle edge **C** internal surface of head region **D** jaw **E–H** *Pseudostegodera qiului* gen. et sp. nov., holotype IZCAS TM206978 **E** jaw **F** internal surface of head region **G** lobe (arrowed) on the left side of mantle edge **H** lobe (arrowed) on the right side of mantle edge.

is not so compressed and the compressed part of body whorl can hardly cover the penultimate whorl.

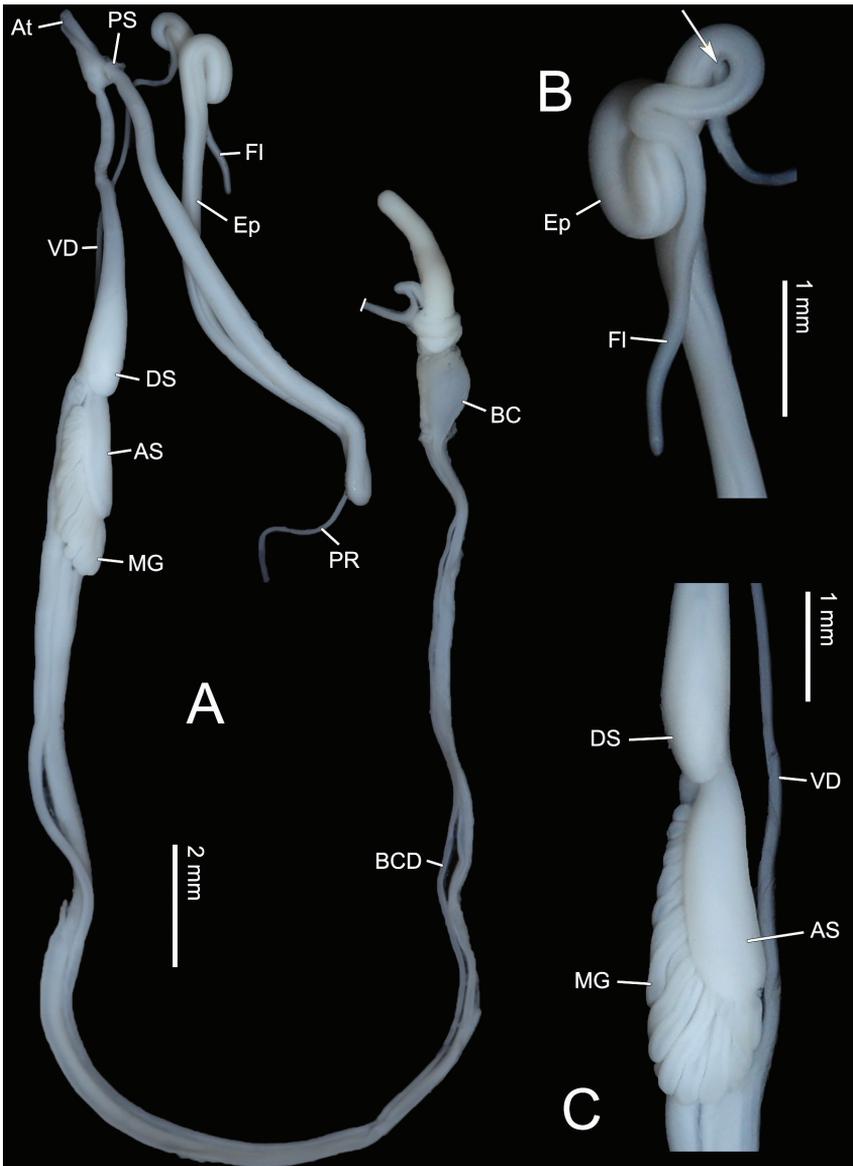


Figure 5. Genitalia of *Stegodera angusticollis* (Martens, 1875), HBUMM08435 **A** general view **B** male part, showing the vas deferens connecting epiphallus (arrowed) **C** female part, showing the dart sac, accessory sac, and mucous glands.

Some authors (Chen and Gao 1987; Qian et al. 2008; Qian and Zhou 2014) recorded this species in Jiangsu, Zhejiang and Anhui, but voucher specimens from these provinces were neither recorded therein nor being found in the mollusk collection deposited in IZCAS where some of them once worked. Therefore, the accuracy of these records needs to be further confirmed.

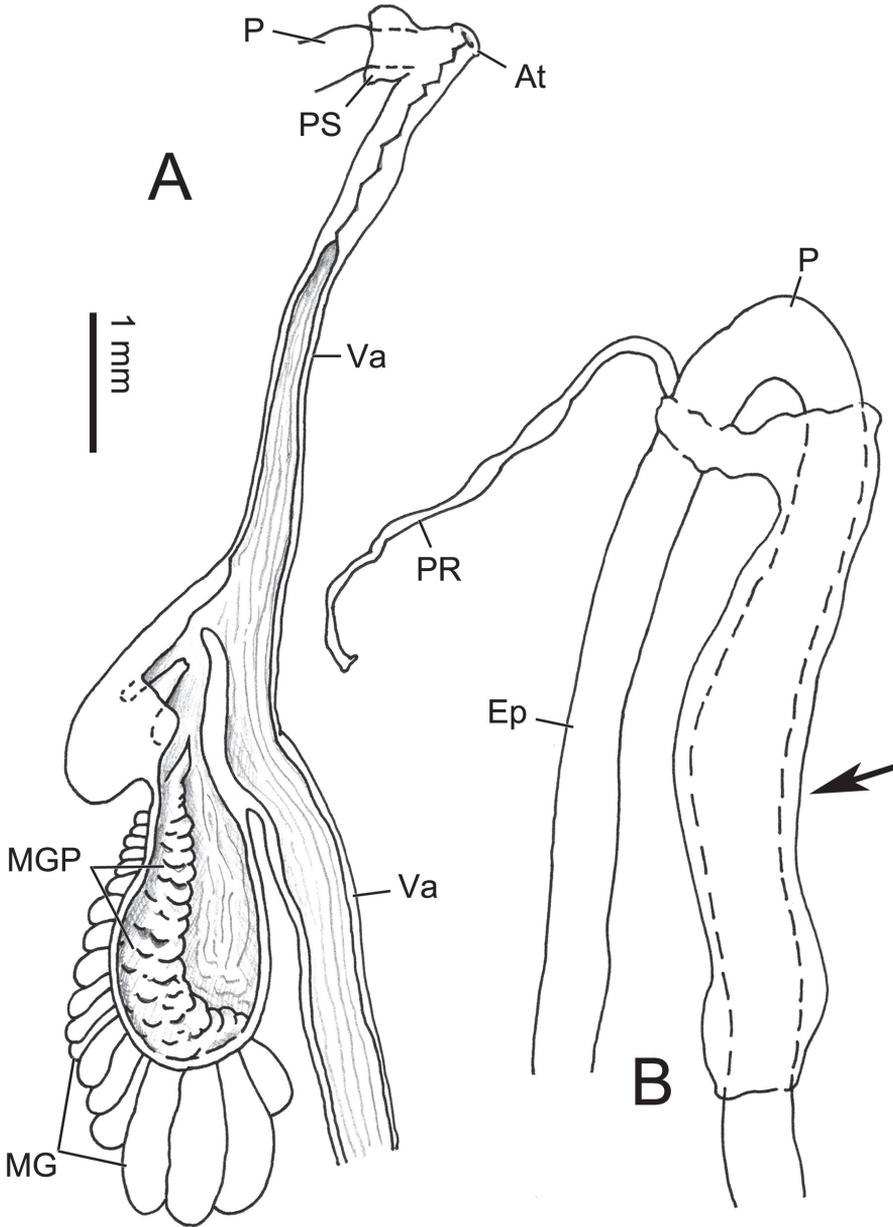


Figure 6. Genitalia of *Stegodera angusticollis* (Martens, 1875), HBUMM08435 **A** exposed dart sac apparatus **B** magnified partial penis and partial epiphallus. Arrow showing a very thin membranous sac wrapping distal half of penis with distal end connecting basal penial retractor muscle.

Pseudostegodera Wu & Chen, gen. nov.

拟盖螺属

<http://zoobank.org/F0A6D222-8DB2-4B57-8910-17CAAE2B7FFA>

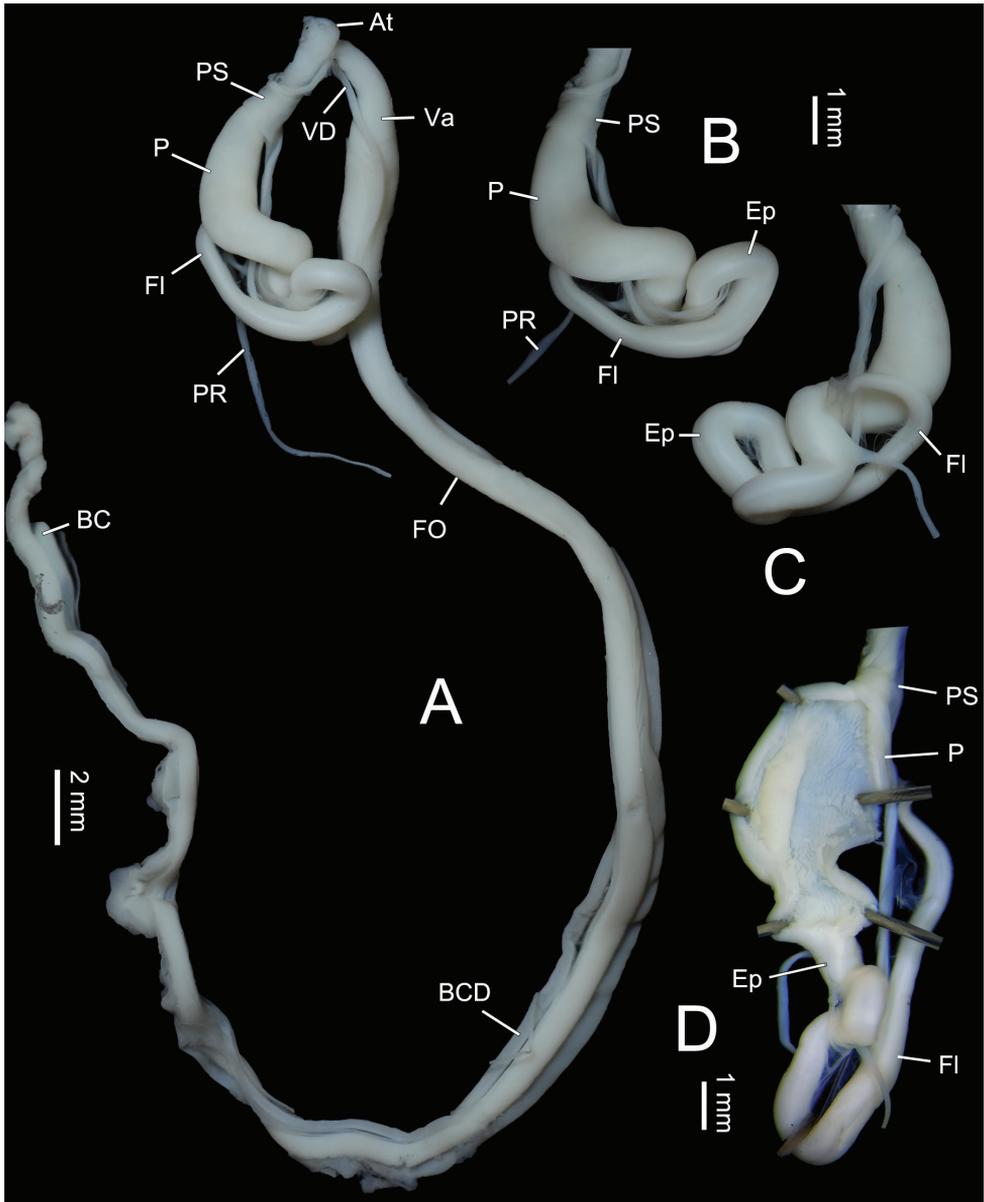


Figure 7. Genitalia of *Pseudostegodera qiului* gen. et sp. nov., holotype IZCAS TM206978 **A** general view **B, C** male part, two sides **D** exposed penis.

Type species. *Pseudostegodera qiului* gen. et sp. nov.

Diagnosis. Shell sinistral. Apical whorls with dense fine ribs gradually becoming granules. The last $\frac{1}{8}$ to $\frac{1}{4}$ body whorl compressed, only partly covering penultimate whorl. Umbilicus broad. Head wart absent. Each side of mantle edge with a leaf-shaped appendage. Penis sheath present. Penis externally simple. Epiphallic papilla absent. Flagellum present. Dart sac apparatus absent.

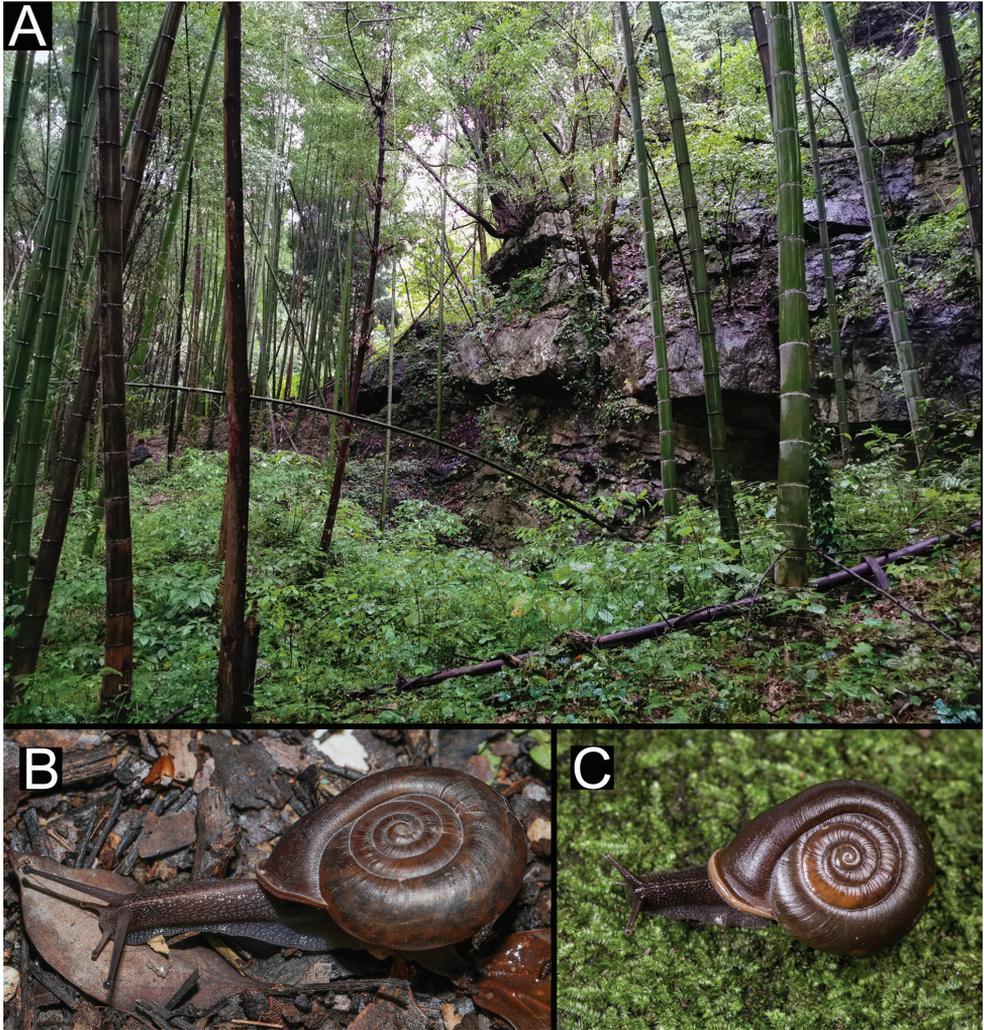


Figure 8. Habitat and living specimens of *Stegoderia angusticollis* (Martens, 1875) **A** habitat (photograph Kai-Chen Ouyang) **B** the living specimen of HBUMM08435 **C** a living specimen from Chibi City (photograph Lu Qiu).

Remarks. Conchologically the new genus can be distinguished from *Stegoderia* by the detached body whorl and penultimate whorl. The new genus is supported to be the closest relative of a clade containing *Stegoderia* + *Nesiohelix* as based on the Maximum Likelihood/Baysian Inference phylogeny shown in Fig. 1A. However, when the Australian camaenids were included in the analysis, the relationships between the new genus, *Stegoderia* + *Nesiohelix*, and all remaining taxa except *Satsuma* remained unresolved (Fig. 1B). The new genus is conchologically similar to *Stegoderia*, however, it differs in certain reproductive characters (see above). We consider that the genital anatomy is more informative than shell features in helicoid systematics (e.g., Criscione and Köhler 2013). In addition, the loss of the dart sac apparatus in

Bradybaeninae and/or Camaeninae is not a frequent event during the evolution of Camaenidae (Fig. 1B). Based on these two considerations, we consider it is reasonable to distinguish this new camaenid taxon as a monotypic genus represented by the new species described below.

***Pseudostegodera qiului* Chen, sp. nov.**

邱氏拟盖螺

<http://zoobank.org/E95104CE-D502-4E48-A256-F0727253745C>

(Figs 1, 2, 3B–D, 4E–H, 7A–D, 9A–C)

Type material. •*Holotype*, IZCAS TM206978, Zihuaping [紫花坪], core area of Shunhuang Mountain National Nature Reserve [舜皇山国家级自然保护区核心区], Huanglong Town [黄龙镇], Xinning County [新宁县], Shaoyang City [邵阳市], Hunan, China, 26°23'31"N, 110°0'25"E, 945 m a.s.l., 2020-VIII-23, leg. Lu Qiu. •*Paratype*, MYNU/1, same data as holotype.

Measurements. $S_{D_{maj}} = 31.0 - 34.0$ mm, $S_{D_{min}} = 25.0 - 26.4$ mm, $S_H = 12.0 - 13.7$ mm ($n = 2$).

Diagnosis. Body whorl incompletely covering partial penultimate whorl. Dart sac apparatus absent. Flagellum present.

Description. Shell (Fig. 3B–D). Sinistral, large, depressed, thick and solid, dark reddish-brown. Shell with $5\frac{1}{2}$ convex whorls. Suture impressed. Protoconch $1\frac{1}{4}$ – $1\frac{1}{2}$ whorls, with regularly arranged fine axial striae that may be invisible by weathering or erosion. Growth lines clear, broken into microscopic tubercles of irregular shape. The last $\frac{1}{8}$ to $\frac{1}{4}$ body whorl compressed, partly covering penultimate whorl. At $\frac{1}{4}$ whorl from the aperture, a spiral depression above periphery and a weak depression near umbilicus making a narrowing on body whorl. Whorl after the narrowing reverting to normal broadness. Aperture semilunar, slightly descending. Peristome white, strongly thickened, expanded, and slightly reflexed. Umbilicus broad, approximately $\frac{1}{3}$ of shell major diameter. Protoconch visible through umbilicus.

General anatomy (Fig. 4E–H). Eversible head wart absent. On internal body wall of head region between ommatophorous insertions with tiny pits rather than glands (Fig. 4F). Each side of mantle edge with a leaf-shaped appendage (Fig. 4G, H). Body reddish brown, central dorsa with light longitudinal stripes. Sole dirty white. Jaw arcuate, with ~ 14 more or less projecting ribs (Fig. 4E).

Genitalia (Fig. 7A–D). Penis sheath short but well developed. Penis somewhat swollen on proximal half, externally simple. In the middle of penis, internally with a single strong longitudinal pilaster almost as thick as epiphallus, ~ 2 mm long (Fig. 7D). Epiphallic papilla absent (Fig. 7D). Flagellum cylindrical, tapering. Vas deferens thin throughout, approximately as long as penis. Dart sac apparatus absent. Bursa copulatrix duct thickened basally. Bursa copulatrix duct longer than 50 mm. Bursa copulatrix rod-shaped.

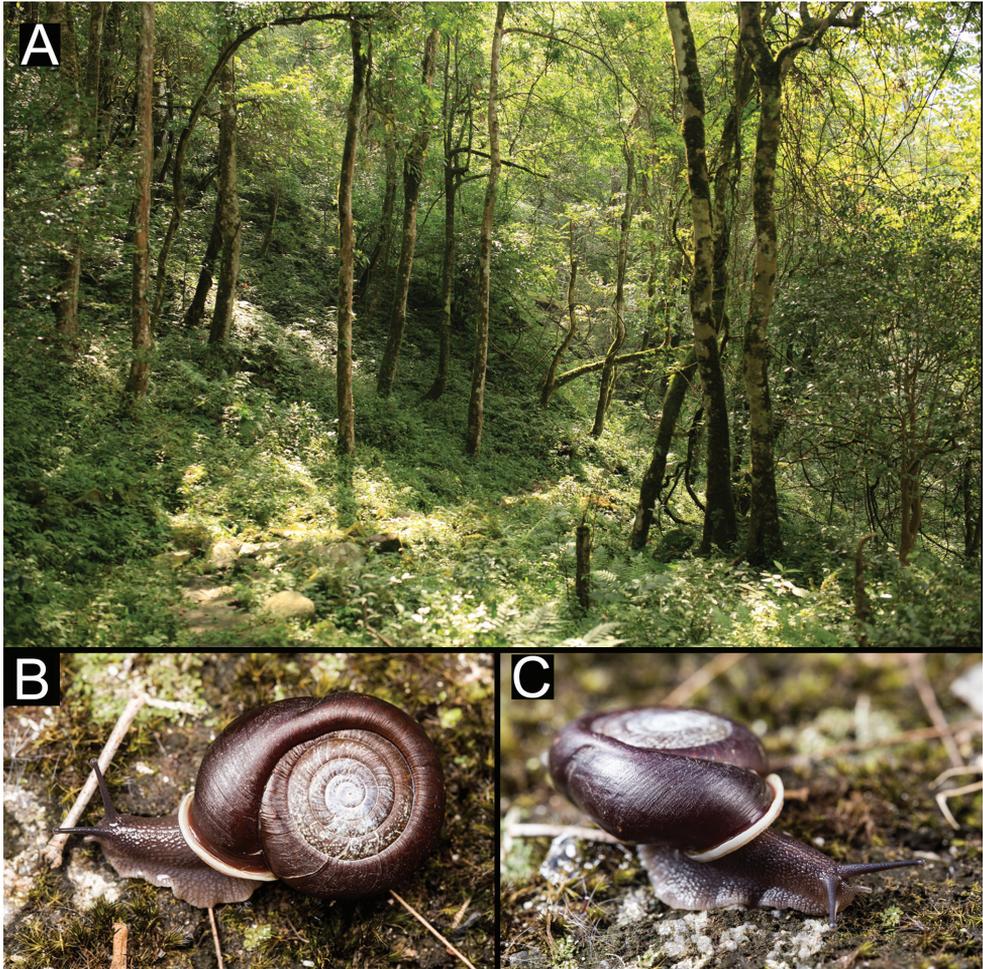


Figure 9. Habitat and living specimens of *Pseudostegodera qiului* gen. et sp. nov. **A** habitat **B, C** a living specimen. Photographs Lu Qiu.

Etymology. This new species is named after Dr Lu Qiu [邱鹭], who provided the specimens and field data.

Ecology. This species was found under rotten wood.

Distribution. Only known from the type locality (Fig. 2).

Remarks. See under the genus.

Discussion

The comparison of the phylograms (Fig. 1) suggests the phylogeny obtained in this paper is more or less robust when the Australian camaenids, which form a monophyly together with the Asian camaenids (as suggested by Scott 1996), are considered. The phy-

logenetic position of *Satsuma* agrees with that suggested by many authors (Chiba 1999: fig. 3; Wade et al. 2007: fig. 2). As indicated by some authors, *Satsuma* is the sister group of the common ancestor of Bradybaeninae and Camaeninae (Fig. 1; Wade et al. 2007: fig. 2), rather than a typical camaenine (sensu Azuma 1995) or bradybaenine (sensu Schileyko 2004). The monophyly made by *Stegodera*, *Nesiohelix* and the new genus is the sister group of all the remaining bradybaenines and camaenines involved in this study. It suggests that neither the Camaeninae including *Satsuma* (sensu Azuma 1995) nor the Bradybaeninae including *Nesiohelix* (sensu nearly all the authors) are monophyletic.

The current work also confirms that so-called important characters of genital system, including penial sheath, epiphallic papilla, penial caecum, flagellum, and dart sac apparatus, have homoplasiously evolved more than once in the studied terminal taxa and their ancestral nodes (Fig. 1B). In other words, widespread homoplasious changes in morphology (Hirano et al. 2014; Fig. 1) including dart sac apparatus based on which Bradybaeninae and Camaeninae are distinguished, rendering the basis of the establishment of Bradybaeninae particularly feeble, explaining that no agreement on the subdivision within Camaenidae was reached to date (Nordsieck 2002; Schileyko 2003, 2004; Wade et al. 2006, 2007; Gittenberger et al. 2012; Bouchet et al. 2017).

Acknowledgements

We are grateful to Sigrid Hof (SMF), Kaibaryer Meng (IZCAS), Christine Zorn and Le-Jia Zhang (ZMB) for searching and imaging the specimens of *Stegodera angusticollis* in their respective collections. Kai-Chen Ouyang (Kunming, China), Di Yu (Wuhan, China), Lu Qiu (MYNU, China), and Zi-Hao Shen (Wuhan, China) are appreciated for providing specimens and field data. We appreciate T. A. Pearce, F. Köhler, and an anonymous reviewer for their very helpful comments.

This work is supported by the National Natural Science Foundation of China (NSFC 31872196) and Guangdong Academy of Science (GDAS) Special Project of Science and Technology Development (No. 2020GDASYL-20200102021).

References

- Azuma M (1995) Colored Illustrations of the Land Snails of Japan. Hoikusha Publishing, 343 pp.
- Bouchet O, Rocroi JP, Hausdorf B, Kaim A, Kano Y, Nützel A, Parkhaev P, Schrödl M, Strong EE (2017) Revised classification, nomenclator and typification of gastropod and monoplacophoran families. *Malacologia* 61(1–2): 1–526. <https://doi.org/10.4002/040.061.0201>
- Castresana J (2000) Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* 17: 540–552. <https://doi.org/10.1093/oxfordjournals.molbev.a026334>

- Chen DN, Gao JX (1987) Economic Fauna of China (Terrestrial Molluscs). Science Press, Beijing, 186 pp. [in Chinese]
- Chiba S (1999) Accelerated evolution of land snails *Mandarina* in the oceanic Bonin Islands: Evidence from mitochondrial DNA Sequences. *Evolution* 53(2): 461–471. <https://doi.org/10.1111/j.1558-5646.1999.tb03781.x>
- Criscione F, Köhler F (2013) Six new species of *Australocosmica* Köhler, 2011 from the Kimberley Islands, Western Australia (Mollusca: Pulmonata: Camaenidae). *Zootaxa* 3608(2): 101–115. <https://doi.org/10.11646/zootaxa.3608.2.1>
- Edler D, Klein J, Antonelli A, Silvestro D (2019) raxmlGUI 2.0 beta: a graphical interface and toolkit for phylogenetic analyses using RAxML. *bioRxiv*. <https://doi.org/10.1101/800912>
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3(5): 294–299.
- Gittenberger E, Hamann TD, Asami T (2012) Chiral speciation in terrestrial pulmonate snails. *PLoS ONE* 7(4): e34005. <https://doi.org/10.1371/journal.pone.0034005>
- Gredler V (1882) Uebersicht der Binnenschnecken von China. *Malakozoologische Blätter, Neue Folge* 5: 165–187.
- Gude GK (1902) A classified list of the helicoid land shells of Asia. *Proceedings of the Malacological Society of London* 9(1): 1–11.
- Gude GK (1920) The armature of land Mollusca. *Proceedings of the Malacological Society of London* 14(1): 52–73.
- Heude PM (1882) Notes sur les mollusques terrestres de la vallée du Fleuve Bleu. *Mémoires concernant l'histoire naturelle de l'Empire chinois* 1: 1–84. <https://doi.org/10.5962/bhl.title.50365>
- Hirano T, Kameda Y, Kimura K, Chiba S (2014) Substantial incongruence among the morphology, taxonomy, and molecular phylogeny of the land snails *Aegista*, *Landouria*, *Trishoplita*, and *Pseudobuliminus* (Pulmonata: Bradybaenidae) occurring in East Asia. *Molecular Phylogenetics and Evolution* 70: 171–181. <https://doi.org/10.1016/j.ympev.2013.09.020>
- Huang CW, Lin SM, Wu WL (2015) Mitochondrial genome sequences of landsnails *Aegista diversifamilia* and *Dolicheulota formosensis* (Gastropoda: Pulmonata: Stylommatophora). *Mitochondrial DNA, Early Online*: 1–3. <https://doi.org/10.3109/19401736.2015.1053070>
- Kerney MP, Cameron RAD (1979) *A Field Guide to the Land Snails of Britain and North-West Europe*. Collins, London, 288 pp. [24 pls.]
- Kobelt W (1879) *Illustriertes Conchylienbuch, vol. 2*. Bauer & Raspe, Nürnberg, 177–264. [pls 61–80]
- Kobelt W (1905 [1904–1906]) *Die Raublungenschnecken (Agnatha). Erste Abtheilung: Streptaxidae und Daudebardiidae*. In: Kobelt W (Ed.) *Systematisches Conchylien-Cabinet von Martini und Chemnitz. Neu herausgegeben und vervollständigt. Ersten Bandes, zwölfte Abtheilung (B). Erste Hälfte. (1) 12b*. Bauer & Raspe, Nürnberg, 1–96. [pls 42–59]
- Kumar S, Stecher G, Tamura K (2015) MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33: 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Maddison WP, Maddison DR (2019) *Mesquite: a modular system for evolutionary analysis. Version 3.61*. <http://www.mesquiteproject.org>

- Martens E von (1875a) Ostasiatische Land- und Süßwasserconchylien. Sitzungs-Berichte der Gesellschaft Naturforschender Freunde, Berlin 1875: 2–4.
- Martens E von (1875b) Binnen-Mollusken aus dem mittlern China. Malakozoologische Blätter 22: 185–188.
- Möllendorff OF (1884) Materialien zur Fauna von China. Jahrbücher der Deutschen Malakozoologischen Gesellschaft 11: 307–391.
- Palumbi S, Martin A, Romano S, McMillan WO, Stice L, Grabowski G (1991) The simple fool's guide to PCR. Department of Zoology, University of Hawaii, Honolulu.
- Pfeiffer L (1875 [1875–1876]) Monographia Heliceorum viventium. Sistens descriptiones systematicas et criticas omnium huius familiae fenerum et specierum hodie cognitarum. Volumen septimum (3): 321–480. F.A. Brockhaus, Lipsiae.
- Pfeiffer L (1870–1876) Novitates Conchologicae. Series prima. Mollusca extramarina. Descriptions et figures de coquilles extramarines nouvelles ou peu connues. Tome IV, 1–8, pls 109–111 [1870], 9–40, pls 112–117 [1871], 41–88, pls 118–125 [1872], 89–112, pls 126–130 [1874], 113–144, pls 131–134 [1875], 145–171, pls 135–137.
- Pilsbry HA (1890) [1890–1891] Manual of conchology, Structural and systematic, with illustrations of the species, Second series, Volume 6 (Helicidae, Vol. IV). Academy of Natural Science Philadelphia, PA, 366 pp. [pp.1–192 (1890)].
- Pilsbry HA (1895) [1893–1895] Manual of conchology, Structural and systematic, with illustrations of the species, Second series, Volume 9 (Helicidae, Vol. 7. Guide to the Study of Helices). Academy of Natural Science Philadelphia, PA, 253 pp. [pp. 161–336 (1895)]
- Pilsbry HA (1905) Notes on *Moellendorffia* and *Stegodera*. The Nautilus 19: 63–67.
- Qian ZX, Zhou WC [Ed.] (2014) Illustrated Handbook of Common Terrestrial Mollusks in China. Zhejiang People's Fine Arts Publishing House, Hangzhou, 228 pp. [in Chinese]
- Qian ZX (ed.) (2008) Farmland Molluscs from Zhejiang. Hangzhou Publishing House, Hangzhou, 425 pp. [in Chinese]
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA (2018) Posterior summarisation in Bayesian phylogenetics using Tracer 1.7. Systematic Biology 67: 901–904. <https://doi.org/10.1093/sysbio/syy032>
- Razkin O, Gómezmoliner BJ, Prieto CE, Martínezortí A, Arrébola JR, Muñoz B, Chuca LJ, Madeira MJ (2015) Molecular phylogeny of the western Palaearctic Helicoidea (Gastropoda, Stylommatophora). Molecular Phylogenetics and Evolution 83: 99–117. <https://doi.org/10.1016/j.ympev.2014.11.014>
- Richardson CL (1983) Bradybaenidae: Catalog of species. Tryonia 12: 1–253.
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Shileyko AA (2003) Treatise on recent terrestrial pulmonate molluscs, Part 11. Trigonochlamydidae, Papillodermidae, Vitrinidae, Limacidae, Bielziidae, Agriolimacidae, Boettgeriellidae, Camaenidae. Ruthenica, Supplement 2(11): 1467–1626. [figs 1896–2098]
- Shileyko AA (2004) Treatise on recent terrestrial pulmonate molluscs, Part 12. Bradybaenidae, Xanthonychidae, Epiphragmophoridae, Helminthoglyptidae, Elonidae, Sphincterochilidae, Cochlicellidae. Ruthenica, Supplement 2(12): 1627–1763. [figs 2099–2259]

- Schmacker B, Boettger O (1894) Description of some Chinese land-shells. Proceedings of the Malacological Society 1: 169–174. <https://doi.org/10.1093/oxfordjournals.mollus.a064109>
- Scott B (1996) Phylogenetic relationships of the Camaenidae (Pulmonata: Stylommatophora: Helicoidea). Journal of Molluscan Studies 62(1): 65–73. <https://doi.org/10.1093/mollus/62.1.65>
- Thiele J (1931) Handbuch der systematischen Weichtierkunde. Vol. 1, Teil 2: 377–778. Gustav Fischer, Jena.
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG (1997) The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. Nucleic Acids Research 25(24): 4876–4882. <https://doi.org/10.1093/nar/25.24.4876>
- Wade CM, Hudelot C, Davison A, Naggs F, Mordan PB (2007) Molecular phylogeny of the helicoid land snails (Pulmonata: Stylommatophora: Helicoidea), with special emphasis on the Camaenidae. Journal of Molluscan Studies 73: 411–415. <https://doi.org/10.1093/mollus/eym030>
- Wade CM, Mordan PB, Naggs F (2006) Evolutionary relationships among the Pulmonate land snails and slugs (Pulmonata, Stylommatophora). Biological Journal of the Linnean Society 87: 593–610. <https://doi.org/10.1111/j.1095-8312.2006.00596.x>
- Wang P, Xiao Q, Zhou WC, Hwang CC (2014) Revision of three camaenid and one bradybaenid species (Gastropoda, Stylommatophora) from China based on morphological and molecular data, with description of a new bradybaenid subspecies from Inner Mongolia, China. ZooKeys 372: 1–16. <https://doi.org/10.3897/zookeys.372.6581>
- Wang P, Yang HF, Zhou WC, Hwang CC, Zhang WH, Qian ZX (2014) The mitochondrial genome of the land snail *Camaena cicatricosa* (Müller, 1774) (Stylommatophora, Camaenidae): the first complete sequence in the family Camaenidae. ZooKeys 451: 33–48. <https://doi.org/10.3897/zookeys.451.8537>
- Wu M (2019) A taxonomic note on the helicoid land snail genus *Traumatophora* (Eupulmonata, Camaenidae). ZooKeys 835: 139–152. <https://doi.org/10.3897/zookeys.835.32697>
- Xia X (2018) DAMBE7: New and improved tools for data analysis in molecular biology and evolution. Molecular Biology and Evolution 35: 1550–1552. <https://doi.org/10.1093/molbev/msy073>
- Xia X, Lemey P (2009) Assessing substitution saturation with DAMBE. In: Lemey P, Salemi M, Vandamme A-M (Eds) The Phylogenetic Handbook: A Practical Approach to DNA and Protein Phylogeny. 2nd edn. Cambridge University Press, 615–630. <https://doi.org/10.1017/CBO9780511819049.022>
- Xia X, Xie Z, Salemi M, Chen L, Wang Y (2003) An index of substitution saturation and its application. Molecular Phylogenetics and Evolution 26: 1–7. [https://doi.org/10.1016/S1055-7903\(02\)00326-3](https://doi.org/10.1016/S1055-7903(02)00326-3)
- Yen TC (1939) Die chinesischen Land- und Süßwasser-Gastropoden des Natur-Museums Senckenberg. Abhandlungen der Senckenbergischen Naturforschenden Gesellschaft 444: 1–234. [16 pls.]
- Zilch A (1960 [1959–1960]) Teil 2, Euthyneura. In: Wenz W (Ed.) Gastropoda, Handbuch der Paläozoologie (ed. Schindewolf OH), Band 6, Gastropoda. Berlin: Gebrüder Borntraeger, 835 pp. [(4): 601–835 (1960)] [Abb. 2112–2515, I–XII]