

Molecular identification and larval morphology of spionid polychaetes (Annelida, Spionidae) from northeastern Japan

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Academic editor: G. Rouse | Received 21 May 2020 | Accepted 29 December 2020 | Published 4 February 2021

<http://zoobank.org/F6BD9213-9DB7-4564-AA00-3C61B2F43B2D>

Citation: Abe H, Sato-Okoshi W (2021) Molecular identification and larval morphology of spionid polychaetes (Annelida, Spionidae) from northeastern Japan. ZooKeys 1015: 1–86. <https://doi.org/10.3897/zookeys.1015.54387>

Abstract

Planktonic larvae of spionid polychaetes are among the most common and abundant group in coastal meroplankton worldwide. The present study reports the morphology of spionid larvae collected mainly from coastal waters of northeastern Japan that were identified by the comparison of adult and larval 18S and 16S rRNA gene sequences. The molecular analysis effectively discriminated the species. Adult sequences of 48 species from 14 genera (*Aonides* Claparède, 1864; *Boccardia* Carazzi, 1893; *Boccardiella* Blake & Kudenov, 1978; *Dipolydora* Verrill, 1881; *Laonice* Malmgren, 1867; *Malacoceros* Quatrefages, 1843; *Paraprionospio* Caullery, 1914; *Polydora* Bosc, 1802; *Prionospio* Malmgren, 1867; *Pseudopolydora* Czerniavsky, 1881; *Rhynchospio* Hartman, 1936; *Scolelepis* Blainville, 1828; *Spio* Fabricius, 1785; *Spiophanes* Grube, 1860) and larval sequences of 41 species from 14 genera (*Aonides*; *Boccardia*; *Boccardiella*; *Dipolydora*; *Laonice*; *Paraprionospio*; *Poecilochaetus* Claparède in Ehlers, 1875; *Polydora*; *Prionospio*; *Pseudopolydora*; *Rhynchospio*; *Scolelepis*; *Spio*; *Spiophanes*) of spionid polychaetes were obtained; sequences of 27 of these species matched between adults and larvae. Morphology of the larvae was generally species-specific, and larvae from the same genus mostly shared morphological features, with some exceptions. Color and number of eyes, overall body shape, and type and arrangement of pigmentation are the most obvious differences between genera or species. The morphological information on spionid larvae provided in this study contributes to species or genus level larval identification of this taxon in the studied area. Identification keys to genera and species of planktonic spionid larvae in northeastern Japan are provided. The preliminary results of the molecular phylogeny of the family Spionidae using 18S and 16S rRNA gene regions are also provided.

Keywords

Larval identification, meroplankton, molecular identification, phylogeny, planktonic larvae, 16S rRNA, 18S rRNA

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Introduction

Many marine invertebrates including polychaetes pass through a planktonic larval phase during their early life history. As such, planktonic larvae derived from the benthic polychaetes are one of the most numerous and diverse groups of coastal zooplankton (Thorson 1946; Hansen 1999; Omel'yanenko and Kulikova 2002; Blake 2017). Polychaetes are often represented in coastal benthic fauna with high species richness all over the world, and members of this group play a large role in the functioning and food webs of marine ecosystems (Aller 1982; Laffaille et al. 2005; Tomiyama et al. 2005, 2007). However, the field study of larval ecology has been restricted because of difficulties in larval identification, which is largely caused by the radical morphological differences between larval and adult stages, lack of diagnostic key characters of larvae, and lack of information on larval forms of many species as is also the case for other marine invertebrates (Branscomb and Vedder 1982; Shanks 1986; Levin 1990). Extensive efforts to describe planktonic polychaete larvae from coastal waters have been performed for species from European and American waters (Blake 2017, and the references cited therein). However, most of these studies are very limited regarding other areas, although Carrasco (1976) and Wu et al. (1978) described polychaete larvae of many species from Chilean and Chinese waters, respectively. In Japan, the larval development and morphology of some polychaete species have been studied (e.g., Izuka 1912: Nereididae Blainville, 1818; Okada 1930: Syllidae Grube, 1850; Okuda 1946: 9 families; Imajima 1959: Spionidae Grube, 1850; Choe 1960: Eunicidae Berthold, 1827; Yamaji 1966: 16 families; Tokioka 1970: Amphinomidae Lamarck, 1818; Imai 1975, 1982: Eunicidae; Miura and Kajihara 1981: Serpulidae Rafinesque, 1815; Yokoyama 1981, 1996: Spionidae; Sasaki and Brown 1983: Saccocirridae Bobretzky, 1872; Yokouchi 1985, 1988: 27 families; Sato and Tsuchiya 1991: Nereididae; Nishi and Yamasu 1992a, 1992b, 1992c, 1992d: Serpulidae; Yokouchi and Yokouchi 1997: 23 families; Koya et al. 2003: Nereididae; Tosuji and Sato 2006: Nereididae; Kondoh et al. 2017: Spionidae; Kan et al. 2020: Nereididae), and some field ecological investigations of polychaete larvae have been conducted (Yokouchi 1984, 1991; Yokoyama 1990, 1995; Abe et al. 2011, 2014; Kan et al. 2020).

Spionidae is one of the largest taxa of polychaete annelids and currently comprises more than 500 nominal species belonging to approximately 38 genera (Radashevsky 2012; Read and Fauchald 2020, excluding *Poecilochaetus* Claparède in Ehlers 1875 and *Trochochaeta* Levensen, 1883). Planktonic spionid larvae are often the most common and abundant group in the coastal meroplankton (Anger et al. 1986; Levin 1986; Abe et al. 2011, 2014) because of their high abundance and species richness in coastal zones, high reproductive capacity, and relatively long planktonic stage (Blake 1996; Blake and Arnofsky 1999). As they are often seasonally dominant in coastal zooplankton communities, spionid larvae can play a major role in planktonic trophic dynamics (Martin et al. 1996; Pedersen et al. 2010). They are also reported to constitute a large portion of ballast water species (Carlton and Geller 1993; Carlton 1996). Spionidae includes species that adult inhabit a wide range of substrates and some are symbionts of other invertebrates (Martin and Britayev 1998, 2018; Sato-Okoshi 1999, 2000; Abe et al. 2019b). Among these, symbionts polydorids (i.e., from the *Polydora* complex or tribe Polydorini, see Radashevsky 2012) are well known as harmful pests in molluscan aquaculture because of their shell boring activities (Blake and Evans 1972; Handley and Bergquist 1997; Simon et al. 2006; Simon and Sato-Okoshi 2015). Understanding larval dynamics and dispersal is important to prevent the settlement of pest spionid species on the shells of aquaculture mollusks (Sato-Okoshi et al. 1990; Simon 2015; David et al. 2016). The host/substrate selectivity and settlement mechanism of spionid larvae during their developmental process are also interesting aspects of larval biology. Although various morphological characteristics of larvae including body shape, pigment patterns (placement and number), ciliary organization, and to some extent, chaetae can be generally used to identify the planktonic larvae of spionid species (Blake and Arnofsky 1999), species-level identification is still difficult because of the lack of information on the larval forms of many species.

The link between larval and adult form has been traditionally achieved by labor-intensive culturing approaches either through rearing larvae collected from plankton or by spawning adults in the laboratory (Shanks 2001). In recent years, ecological studies on the diversity and distribution of marine planktonic larvae are increasingly depending on molecular methods for accurate taxonomic identification to species level (Andre et al. 1999; Hosoi et al. 2004; Pradillon et al. 2007; Phillips et al. 2008; Heimeier et al. 2010). For future metabarcoding studies, establishment of a comprehensive DNA barcoding library is very useful for rapid identification of planktonic larvae. Meanwhile, the use of molecular methods for identifying planktonic larvae in extensive field surveys handling large numbers of collected samples still requires extensive cost. Since the direct microscopic observation, which allows prompt identification of larvae at low cost, remains the popular technique for distinguishing planktonic larvae, information on larval morphology would be useful for such studies.

The aim of the present study is identification of spionid larvae that dominantly appear among the planktonic polychaete larvae from northeastern Japan (Abe et al. 2011, 2014) by comparing adult and larval gene sequences. The 18S rRNA and 16S

rRNA genes was herein used as a marker for species-level discrimination in Spionidae. Moreover, we report the results of preliminary phylogenetic analysis using these genetic regions and describe the morphologies of spionid larvae with photomicrographs of living specimens.

Materials and methods

Sample collection and morphological observation

Planktonic larvae of spionid polychaetes were collected mainly from a coastal station in Onagawa Bay (38°26'15"N, 141°27'42"E; depth: 22 m), but also from Gobu-ura (38°24'01"N, 141°27'59"E), Sasuhama (38°24'22"N, 141°22'08"E), Sendai Port (38°16'22"N, 141°00'01"E), and Gamo Lagoon (38°15'18"N, 141°00'48"E) in Miyagi Prefecture, northeastern Japan, and Tomiura (35°02'20"N, 139°49'16"E) in Boso Peninsula and Habu Port (34°41'09"N, 139°26'16"E) in Izu-Oshima Island in eastern Japan (Table 1, Fig. 1). Plankton samples were collected in Onagawa Bay once a month, from April 2011 to August 2012, by vertical hauls from the bottom to the surface using a NORPAC net (Motoda 1957) with a mesh size of 110 μm . In the other areas, the plankton samples were collected in 2011–2016 by using a simple plankton net with a mesh size of 100 μm and a mouth diameter of 30 cm. Morphological characteristics of live spionid larvae were observed under stereomicroscopes (Leica, WILD MZ8; Olympus, SZX 16), and light photomicrographs were taken by using digital cameras (Nikon E950, E4500; Olympus DP25, DP73; Sony α 6000) attached to the microscope. The larvae were anesthetized with magnesium chloride solution when necessary before the photography. Background, brightness, and contrast of the obtained images were adjusted using GNU Image Manipulation Program (GIMP) 2.10.6 (www.gimp.org). The terms trochophore, metatrochophore, and nectochaeta were defined as larvae with prototroch, clear signs of segmentation, and functional parapodia, respectively according to Rouse (2006).

Adult spionid polychaetes were collected from coastal waters in Shinminato (45°12'27"N, 141°08'09"E) and Numaura (45°06'54.0"N, 141°17'10.0"E) in Rishiri Island, Onagawa Bay, Sasuhama, Matsushima Bay (38°19'54"N, 141°08'44"E), Gamo Lagoon, Ninzaki (37°12'14"N, 136°55'07"E) and Kashima (37°05'13"N, 136°55'35"E) in Nanao Bay, Iwaki (36°55'14"N, 140°51'31"E), Moroiso Bay (35°09'27"N, 139°36'43"E), Ena Bay (35°08'46"N, 139°39'57"E), Tomiura, Akinohama (34°47'12"N, 139°24'32"E) in Izu-Oshima Island, Ishigaki Island (24°24'01"N, 124°08'30"E), and from a 103-m depth (by dredging) in Sagami Bay (35°05'N, 139°37'E) in Japan in 2011–2018 (Table 1, Fig. 1). Specimens were fixed in 70% or 99% ethanol. These fixed specimens of adult spionids were observed under stereomicroscopes (Leica, WILD MZ8; Olympus, SZX 16) and a biological light microscope (Nikon, Eclipse80i) and identified based on their mor-

Classification	Type locality		Sampling locality		Accession no. (length: bp)	
					18S	16S
	Adult Larvae	Adult Larvae	Adult Larvae	Adult Larvae	Adult Larvae	Adult Larvae
<i>Scolecipis cf. kudenovii</i> Hartmann-Schröder, 1981	Australia	Sasuhama	Ena Bay	LC545880 (1819)	–	LC595710 (505)
<i>Scolecipis planata</i> Imajima, 1992	Japan	–	Nanao Bay, Maotukawa-ura	LC545881 (1816)	LC595711 (501)	–
<i>Scolecipis tesana</i> Foster, 1971	USA	–	Lagoon	LC545882 (1821)	LC595712 (501)	–
<i>Scolecipis</i> sp. 1	–	Onagawa Bay	Onagawa Bay	LC545883 (1819)	LC595713 (505)	LC595714 (505)
<i>Scolecipis</i> sp. 2	–	Onagawa Bay	Onagawa Bay	LC545885 (1820)	–	LC595715 (505)
<i>Spiophanes</i> Grube, 1860	Greenland Sea	–	Onagawa Bay	LC545886 (1750)	LC595716 (500)	–
<i>Spiophanes</i> aff. <i>kroyeri</i> Grube, 1860	Russia	–	Onagawa Bay	LC545887 (1750)	–	LC595717 (504)
<i>Spiophanes uschakovii</i> Zachs, 1933	Russia	–	Onagawa Bay, Sasuhama	LC545888 (1750)	LC595718 (504)	LC595719 (504)
<i>Spiophanes</i> aff. <i>uschakovii</i> Zachs, 1933	Russia	–	Sasuhama	LC545889 (1750)	LC595719 (504)	–
<i>Spiophanes wigleyi</i> Pettibone, 1962	Georges Bank	–	Sagami Bay	LC545890 (1749)	LC595720 (513)	–
Spioninae Söderström, 1920						
<i>Boccardia</i> Canazzi, 1893						
<i>Boccardia proboscidea</i> Hartman, 1940	USA	Sasuhama	Sasuhama	LC107607 (1768) [†]	LC545891 (1768)	LC595721 (472)
<i>Boccardia pseudonatrix</i> Day, 1961	South Africa	Tomiuira	Tomiuira	LC545892 (1745)	LC545893 (1745)	LC595723 (466)
<i>Boccardia</i> sp. 1	–	–	Onagawa Bay	–	LC545894 (1705)	LC595725 (472)
<i>Boccardia</i> sp. 2	–	–	Onagawa Bay, Sasuhama, Sendai Port	–	LC545895 (1705)	LC595726 (472)
<i>Boccardiella</i> Blake & Kudenov, 1978						
<i>Boccardiella hamata</i> (Webster, 1879)	USA	Sasuhama, Gamo Lagoon	Onagawa Bay, Gobi-ura, Sasuhama	LC107608 (1772) [†]	LC545896 (1772)	LC595727 (472)
<i>Dipolydora</i> Verrill, 1881						
<i>Dipolydora armata</i> (Langenhans, 1880)	Madeira	Akinohama	–	LC545897 (1772)	–	LC595729 (473)
<i>Dipolydora bidentata</i> (Zachs, 1933)	Russia	Sasuhama	Onagawa Bay	LC107609 (1770) [†]	LC545898 (1770)	LC595730 (475)
<i>Dipolydora</i> cf. <i>commensalis</i> (Andrews, 1891)	USA	–	Sasuhama	–	LC545899 (1769)	LC595732 (474)
<i>Dipolydora giardi</i> (Mesnil, 1893)	France	Onagawa Bay	Onagawa Bay	LC545900 (1770)	LC545901 (1766)	LC595733 (474)
<i>Dipolydora</i> cf. <i>socialis</i> (Schmarda, 1861)	Chile	Onagawa Bay, Sasuhama	Onagawa Bay	LC545902 (1770)	LC545903 (1770)	LC595735 (475)
<i>Dipolydora</i> sp.	–	–	Onagawa Bay	–	LC545904 (1770)	LC595737 (476)
<i>Polydora</i> Bosc, 1802						
<i>Polydora atra</i> Sato-Okoshi, 1998	Japan	Hiroshima Bay	–	AB705409 (1771) [‡]	–	LC500931 (473) [‡]
<i>Polydora brevipalpis</i> Zachs, 1933	Russia	Mutsu Bay, Onagawa Bay	Onagawa Bay, Sasuhama	AB705407 (1771) [‡]	LC545905 (1766)	LC595738 (474)
<i>Polydora calcareo</i> (Templeton, 1836)	UK	Kitaibaraki	–	AB705403 (1771) [‡]	–	LC595740 (475)
<i>Polydora cornuta</i> Bosc, 1802	USA	Sasuhama, Gamo Lagoon	Gamo Lagoon	LC541483 (1742) [‡]	LC545906 (1770)	LC541484 (470) [‡]

Classification	Type locality	Sampling locality		Accession no. (length: bp)	
		Adult Larvae		16S	
		Adult Larvae	Adult Larvae	Adult Larvae	Adult Larvae
<i>Polydora cf. glycymeriza</i> Radashkevsky, 1993	Russia	–	Onagawa Bay, Sendai Port	LC545907 (1771)	– LC595742 (472)
<i>Polydora hoplara</i> Claparède, 1868	Italy	Kitabaraki	Onagawa Bay, Gobu-ura	LC545908 (1769)	LC101870 (475) [†] LC595743 (475)
<i>Polydora neocarea</i> Williams & Radashkevsky, 1999	USA	Hiroshima Bay	–	–	LC595744 (471)
<i>Polydora onagawaensis</i> Teramoto, Sato-Okoshi, Abe, Nishitani & Endo, 2013	Japan	Onagawa Bay	Onagawa Bay	LC545909 (1771) ^d	LC595745 (473) LC595746 (473)
<i>Polydora cf. spongiola</i> Berkeley & Berkeley, 1950	Canada	Moroiso Bay	Sasubama	LC545910 (1771)	LC595747 (475) LC595748 (475)
<i>Polydora websteri</i> Hartman in Loosanoff & Engle, 1943	USA	Nakatsu tidal flats	–	–	LC595749 (468)
<i>Polydora</i> sp. 1	–	Sasubama	Onagawa Bay, Sasubama	LC545912 (1771)	LC595750 (476) LC595751 (476)
<i>Polydora</i> sp. 2	–	–	Sasubama, Gamo Lagoon	LC545914 (1771)	– LC595752 (4702)
<i>Polydora</i> sp. 3	–	–	Onagawa Bay, Sasubama	LC545915 (1771)	– LC595753 (473)
<i>Pseudopolydora</i> Czerniawsky, 1881					
<i>Pseudopolydora</i> aff. <i>achaeta</i> Radashkevsky & Hsieh, 2000	Taiwan	Onagawa Bay	Onagawa Bay	LC545916 (1773) [†]	LC595754 (468) LC595755 (468)
<i>Pseudopolydora cf. kempfi</i> (Southern, 1921)	India	Gamo Lagoon	Gamo Lagoon	LC019990 (1772) [†]	LC595756 (471) LC595757 (471)
<i>Pseudopolydora paucibranchiata</i> (Okuda, 1937)	Japan	Mangoku-ura Inlet	Onagawa Bay	LC019991 (1784) [†]	LC595758 (455) LC595759 (455)
<i>Pseudopolydora cf. reticulata</i> Radashkevsky & Hsieh, 2000	Taiwan	Gamo Lagoon	Onagawa Bay, Gamo Lagoon, Sendai Port	LC019988 (1775) [†]	LC595760 (470) LC595761 (470)
<i>Pseudopolydora tsukaki</i> Simon, Sato-Okoshi & Abe, 2017	Japan	Habu Port	Tomuira, Habu Port	AB973929 (1713) [†]	LC107857 (475) [†] LC595762 (425)
<i>Pseudopolydora uchiumi</i> Simon, Sato-Okoshi & Abe, 2017	Japan	Uranoouchi Bay	–	–	LC107855 (474) [†]
<i>Spio</i> Fabricius, 1785					
<i>Spio</i> sp. 1	–	–	Sasubama	LC545921 (1781)	– LC595763 (471)
<i>Spio</i> sp. 2	–	Rishiri Island	Onagawa Bay	LC545922 (1762)	LC595764 (467) LC595765 (467)
	–	Sasubama, Matsushima Bay	Onagawa Bay, Sasubama	LC545924 (1760)	LC595766 (462) LC595767 (462)

[†]: Sato-Okoshi and Abe (2012); [‡]: Sato-Okoshi and Abe (2013); [§]: Sato-Okoshi et al. (2017); [¶]: Teramoto et al. (2013); ^{||}: Abe et al. (2016); ^{††}: Simon et al. (2019); ^{†††}: Abe & Sato-Okoshi (2020).

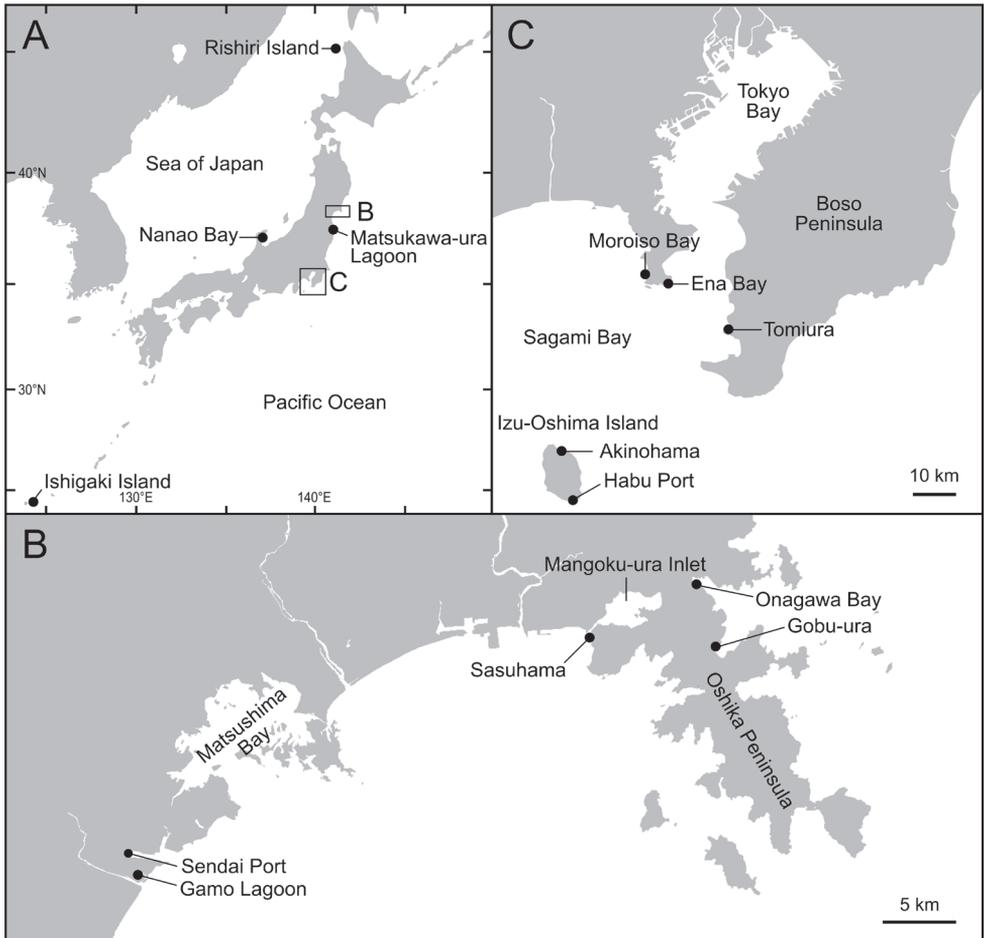


Figure 1. Maps showing sampling locations of the present study.

phological characteristics. Following Blake (2006) and Radashevsky et al. (2018), the genera *Poecilochaetus* and *Trochochaeta* were considered as belonging to the family Spionidae.

DNA analysis and larval identification

Adults and larvae of one or more individuals, respectively, were subjected to DNA analysis. In order to clarify the development links between the different stages, we analyzed the DNA of as many larvae of different stages as possible. Except for *Laonice* sp. 2 (Fig. 4E), all larvae pictured in this paper have been identified by DNA analysis. All individuals were washed by several transfers in sterile filtered (pore size 0.2 μm) seawater and distilled water to remove as much extraneous matter as possible before

DNA extraction. Genomic DNA was extracted from live or ethanol-preserved larval (from the whole body) and adult (from palp or a small piece of tissue) spionid specimens by grinding and heating at 95 °C for 20 min in 50 µl TE buffer (pH 8.0) with 10% Chelex 100 (Bio-Rad; Richlen and Barber 2005). Undiluted or 10-fold diluted extracted DNA in TE buffer was used as template for polymerase chain reaction (PCR) depending on the DNA concentration. Partial sequences of nuclear 18S rRNA gene were amplified by PCR according to the methods described by Sato-Okoshi and Abe (2012, 2013) and Teramoto et al. (2013) using the following primer pairs (Nishitani et al. 2012): 18S-1F1 (AACCTGGTTKATCCTGCCAG) and 18S-1R632 (ACTACGAGCTTTTTAAACYGCARC), 18S-2F576 (GGTAATTCAGCTCYAATRG) and 18S-2R1209 (AAGTTTYCCCGTGTTGARTC), and 18S-3F1129 (GCTGAAACTTAAAGRAATTGACGG) and 18S-R1772 (TCACCTACGGAAACCTTGTTACG). Partial sequences of mitochondrial 16S rRNA gene were amplified by PCR according to the methods described by Abe et al. (2019a) using the 16Sar (CGCCTGTTTATCAAAAACAT) and 16Sbr (CCGGTCTGAACTCAGATCACGT) primer pair (Palumbi et al. 1991). The PCR products were purified using ExoSAP-IT (Affymetrix, Cleveland, OH, USA) and sequenced by Eurofins Genomics (Tokyo, Japan). The forward and reverse complementary sequences and contigs were assembled using GeneStudio ver. 2.2.0.0 (GeneStudio, Inc. Suwanee, GA, USA). Larval and adult gene sequences obtained in the present study (Table 1) were aligned using the MAFFT online service ver. 7 with the L-INS-i algorithm (Katoh et al. 2017) with (Fig. 2) and without (Fig. 3) the sequences of other spionid species available in the DNA Data Bank of Japan (DDBJ), the European Nucleotide Archive (ENA), or GenBank databases (Table 2). The 18S and 16S ribosomal RNA gene sequences of *Sabella pavonina* Savigny, 1822 (DDBJ/EMBL/GenBank ID: U67144 and AY340482) and *Laonome* sp. (KP793139 and KP793138) obtained from DDBJ/ENA/GenBank were used as outgroup taxa. Ambiguously aligned regions of 2 alignments were eliminated by employing Gblocks (Talavera and Castresana 2007) implemented in PhyloSuite v.1.2.2 (Zhang et al. 2020) with the following relaxed settings: minimum number of sequences for a conserved/flank position: half the number of sequences + 1, maximum number of contiguous non-conserved positions: 10, minimum length of a block: 5, and with half of the allowed gap positions. The final lengths of the alignments were 1738 (18S) and 447 (16S) bp for the multiple sequence alignment (MSA) without DDBJ/ENA/GenBank sequences and 1644 (18S) and 410 (16S) bp for MSA with DDBJ/ENA/GenBank sequences. Phylogenetic trees were constructed based on the concatenated sequences of 18S and 16S rRNA gene region by maximum likelihood (ML) analyses performed using IQ-TREE (Nguyen et al. 2015) implemented in PhyloSuite under Edge-linked partition model. The TIM2e+I+G4 and TIM2+F+I+G4 models were selected for the 18S and 16S rRNA gene region, respectively as the best substitution model by ModelFinder (Kalyanamoorthy et al. 2017) as implemented in IQ-TREE under the Bayesian information criterion (BIC). The robustness of the ML trees was evaluated by the Shimodaira–Hasegawa–like approximate likelihood-ratio test (SH-

Table 2. Terminal taxa whose sequences were obtained from DDBJ/EMBL/GenBank and herein used in the phylogenetic analyses. Type and collection localities, accession numbers, sequence lengths, and references are shown.

	Classification		Type locality	Collection locality	Accession number (Length: bp)		Reference
					18S	16S	
Spironiidae							
Neriniinae							
Neriniinae	Söderström, 1920						
<i>Anoidella</i>	López-Jamar, 1989		Gulf of Cadiz, Spain	Great Mercor Seamount, NE Atlantic	KF434504 (483)	KF434508 (443)	Meißner et al. (2014)
<i>Anoidella</i> cf. <i>dryi</i>	Maciolek in López-Jamar, 1989						
<i>Anoides</i>	Claparède, 1864						
<i>Anoides</i>	<i>oxycephala</i> (Sars, 1862)		Norway	France	MG913226 (1699)	MG878895 (337)	Radashkevsky et al. (unpubl.)
<i>Anoides</i>	<i>selvagensis</i> Brito, Núñez & Riera, 2006		Savage Islands, Portugal	Irving Seamount, NE Atlantic	KF434507 (516)	–	Meißner et al. (2014)
<i>Aurospio</i>	Maciolek, 1981						
<i>Aurospio</i>	<i>albranchiata</i> Maciolek, 1981		Argentine Basin, SW Atlantic	Kaplan, Pacific Mn nodule province	EU340091 (1797)	EU340087 (484)	Mincus et al. (2009)
<i>Aurospio</i>	<i>foodhancsia</i> Mincus, Dyal, Paterson, Smith & Glover, 2009		Bellingshausen Sea, Antarctica	West Antarctic Peninsula shelf	EU340097 (1765)	EU340078 (552)	Mincus et al. (2009)
<i>Aurospio</i>	sp. Q		–	India	–	KF459948 (443)	Periasamy et al. (unpubl.)
<i>Aurospio</i>	sp. R		–	Ross Sea	–	KF713473 (397)	Gallego et al. (2014)
<i>Aurospio</i>	sp. S		–	Eastern Vema Fracture Zone	–	MN441726 (409)	Guggolz et al. (2020)
<i>Aurospio</i>	sp. T		–	Clanton Clipperton Fracture Zone	–	MN441512 (411)	Guggolz et al. (2020)
<i>Dispio</i>	Hartman, 1951						
<i>Dispio</i>	<i>remanei</i> Friedrich, 1956		Pacific Ocean, Central America	Brazil	KU9000474 (671)	–	Rebello & Schertini (unpubl.)
<i>Glandulospio</i>	Meißner, Bick, Guggolz & Göting, 2014						
<i>Glandulospio</i>	<i>oreste</i> Meißner, Bick, Guggolz & Göting, 2014		Little Mercor Seamount, NE Atlantic	Little Mercor Seamount, NE Atlantic	KF434505 (402)	KF434511 (446)	Meißner et al. (2014)
<i>Laonice</i>	Malmgren, 1867						
<i>Laonice</i>	cf. <i>antarctica</i> Hartman, 1953						
<i>Laonice</i>	<i>cirrata</i> (M. Sars, 1851)		Rio Grande do Sul	Antarctic	KM998754 (1744)	KX867280 (373)	Brasier et al. (2016)
<i>Laonice</i>	<i>norgensis</i> Sikorski, 2003		Norway	Russia	KF434506 (514)	–	Radashkevsky et al. (unpubl.)
<i>Laonice</i>	cf. <i>vietzei</i> López, 2011		Norwegian Sea, North Atlantic	Little Mercor Seamount	–	KF434512 (454)	Meißner et al. (2014)
<i>Laonice</i>	<i>weddellia</i> Hartman, 1978		Bellingshausen Sea, West Antarctica	Antarctic	–	KX867288 (368)	Brasier et al. (2016)
<i>Lapnice</i>	sp. VR-2006		Weddell Sea	Antarctic	–	KX867313 (379)	Brasier et al. (2016)
<i>Laonice</i>	sp. SLM-2008		–	Bohuslän, Sweden	DQ779655 (1705)	DQ779619 (342)	Roussier et al. (2007)
<i>Laonice</i>	sp. A		–	California borderland basins, USA	EU340089 (1784)	EU340088 (546)	Mincus et al. (2009)
<i>Laonice</i>	sp. B		–	Eastern Vema Fracture Zone	MK507647 (1017)	MK507653 (469)	Guggolz et al. (2019)
<i>Laonice</i>	sp. C		–	Eastern Vema Fracture Zone	MK507651 (1017)	MK507657 (470)	Guggolz et al. (2019)
<i>Laonice</i>	sp. D		–	Western Vema–Fracture Zone	MK507650 (1017)	MK507658 (450)	Guggolz et al. (2019)
<i>Laonice</i>	sp. E		–	Western Vema–Fracture Zone	MK507638 (1017)	MK507723 (475)	Guggolz et al. (2019)
<i>Laonice</i>	sp. E		–	Western Vema–Fracture Zone	MK507644 (966)	MK507706 (473)	Guggolz et al. (2019)

Classification	Type locality	Collection locality	Accession number (Length:bp)		Reference
			18S	16S	
<i>Laonice</i> sp. F	–	Vema Transform Fault	MK507623 (1017)	MK507718 (473)	Guggolz et al. (2019)
<i>Laonice</i> sp. G	–	Puerto Rico Trench	MK507624 (1017)	MK507708 (474)	Guggolz et al. (2019)
<i>Laonice</i> sp. H	–	Puerto Rico Trench	MK507617 (1017)	MK507720 (474)	Guggolz et al. (2019)
<i>Malacoceros</i> Quatrefores, 1843					
<i>Malacoceros fuliginosus</i> (Claparède, 1868)	Italy	St. Efflaur, France	AV525632 (1765)	–	Struck and Purschke (2005)
<i>Malacoceros indicus</i> (Fauvel, 1928)	Gulf of Mannar	Helgoland, Germany	–	EF431961 (417)	Blank and Bastrop (2009)
<i>Malacoceros Mesnil</i> , 1896		Lizard Island, Australia	KT636512 (454)	KP636511 (391)	Meißner and Görting (2015)
<i>Marenzelleria arctica</i> (Chamberlin, 1920)	Beaufort Sea	Kara Sea, Russia	KJ546264 (1775)	KJ546306 (343)	Radaevskiy et al. (2014)
<i>Marenzelleria bastropi</i> Bick, 2005	North Carolina, USA	USA	EF446959 (468), EF446967 (577)	EF431963 (419)	Blank and Bastrop (2009)
<i>Marenzelleria neglecta</i> Sikorski & Bick, 2004	Germany	Baltic Sea	EF446955 (470), EF446963 (578)	DQ309248 (419)	Bastrop and Blank (2006), Blank and Bastrop (2009)
<i>Marenzelleria viridis</i> (Verrill, 1873)	New Jersey, USA	Barlow's Landing, MA, USA	EU418860 (1810)	–	Struck et al. (2008)
<i>Marenzelleria wirens</i> Augener, 1913	Franz Josef Land, Russia	Ringsjøbing Fjord	EF446957 (472), EF446965 (579)	DQ309252 (419) EF431980 (417)	Bastrop and Blank (2006) Blank and Bastrop (2009)
<i>Panprionospio</i> Caullery, 1914					
<i>Panprionospio condifolia</i> Yokoyama, 2007	Wakasa Bay, Japan	Eastern Arabian Sea, India	KT900309 (1655)	–	Rengaiyan and Ingole (2018)
<i>Panprionospio cristata</i> Zhou, Yokoyama & Li, 2008	East China Sea, China	India	KY704338 (520)	–	Vijapur et al. (unpubl.)
<i>Panprionospio patiens</i> Yokoyama, 2007	Osaka Bay, Japan	India	KT900307 (1684)	KY704331 (519)	Rengaiyan and Ingole (2018), Vijapur et al. (unpubl.)
<i>Panprionospio</i> sp. EPK-2019	–	–	MN069511 (588)	–	Kiskaddon et al. (unpubl.)
<i>Pocillochaetus</i> Claparède in Ehlers, 1875	English Channel	Arcachon, France	AV569652 (1833)	AV569680 (463)	Bleidorn et al. (2005)
<i>Pocillochaetus serpens</i> Allen, 1904	–	Banyuls, France	DQ779667 (1710)	DQ779630 (344)	Roussel et al. (2007)
<i>Pocillochaetus</i> sp. VR-2006	–	Clarion–Clipperton Fracture Zone	–	MK971106 (419)	Bonifacio et al. (2020)
<i>Pocillochaetus</i> sp. 18 PB					
<i>Pocillochaetus</i> sp. 18 PB					
<i>Prionospio</i> Malmgren, 1867					
<i>Prionospio dahlia</i> Day, 1961	South Africa	Southern New England, MA, USA	EU418859 (1823)	–	Struck et al. (2008)
<i>Prionospio</i> sp. A	–	Clarion Clipperton Fracture Zone	–	MN441557 (416)	Guggolz et al. (2020)
<i>Prionospio</i> sp. B	–	Eastern Vema Fracture Zone	MN447146 (846)	MN441645 (331)	Guggolz et al. (2020)
<i>Prionospio</i> sp. C (as <i>Prionospio</i> sp. 29 PB)	–	Clarion–Clipperton Fracture Zone	MK971148 (1677)	MK971035 (422)	Bonifacio et al. (2020)
<i>Prionospio</i> sp. D	–	Eastern Vema Fracture Zone	MN447192 (842)	MN441641 (409)	Guggolz et al. (2020)
<i>Prionospio</i> sp. E (as <i>Prionospio ehlerti</i>)	–	GROZEX	EU340095 (1812)	EU340081 (549)	Mincek et al. (2009)
<i>Prionospio</i> sp. F	–	Clarion Clipperton Fracture Zone	–	MN441542 (405)	Guggolz et al. (2020)
<i>Prionospio</i> sp. G	–	Eastern Vema Fracture Zone	MN447188 (844)	MN441564 (397)	Guggolz et al. (2020)
<i>Prionospio</i> sp. H	–	Clarion Clipperton Fracture Zone/ eastern Vema Fracture Zone	MN447158 (844)	MN441554 (411)	Guggolz et al. (2020)

Classification	Type locality	Collection locality	Accession number (Length:bp)		Reference
			18S	16S	
<i>Prionospio</i> sp. I	–	Puerto Rico Trench	MN447157 (844)	MN441749 (409)	Guggolz et al. (2020)
<i>Prionospio</i> sp. K	–	Clarion Clipperton Fracture Zone	–	MN441555 (413)	Guggolz et al. (2020)
<i>Prionospio</i> sp. L	–	Western Vema Fracture Zone	MN447168 (844)	MN441745 (408)	Guggolz et al. (2020)
<i>Prionospio</i> sp. M	–	Eastern Vema Fracture Zone	MN447160 (844)	MN441561 (411)	Guggolz et al. (2020)
<i>Prionospio</i> sp. N	–	Eastern Vema Fracture Zone	MN447180 (844)	MN441604 (342)	Guggolz et al. (2020)
<i>Prionospio</i> sp. O	–	Eastern Vema Fracture Zone	MN447159 (844)	MN441748 (342)	Guggolz et al. (2020)
<i>Prionospio</i> sp. P	–	Eastern Vema Fracture Zone	MN447173 (844)	MN441753 (339)	Guggolz et al. (2020)
<i>Prionospio</i> sp. KJO-2005	–	Monterey Bay, CA, USA	DQ209226 (1703)	–	Osborn et al. (2007)
<i>Pygospio</i> Claparède, 1863					
<i>Pygospio elegans</i> Claparède, 1863	Normandy, France	Russia	KJ747074 (1719)	KJ747084 (468)	Radashevsky et al. (2016b)
<i>Pygospio</i> sp. 1 (as <i>Pygospio</i> sp. 2583)	–	Russia	KP940584 (1709)	KP940582 (306)	Radashevsky et al. (2016b)
<i>Pygospio</i> sp. 2 (as <i>Pygospio</i> sp. VVP-2014)	–	USA	KJ747077 (1756)	KJ747087 (306)	Radashevsky et al. (2016b)
<i>Rhynchospio</i> Hartman, 1936					
<i>Rhynchospio arenicola</i> Hartman, 1936	CA, USA	USA	KJ546286 (1737)	KJ546318 (341)	Radashevsky et al. (2014)
<i>Rhynchospio</i> aff. <i>asiatica</i> sensu Radashevsky et al. (2014)	–	South Korea	KJ546296 (1731)	KJ546345 (492)	Radashevsky et al. (2014)
<i>Rhynchospio daruini</i> Radashevsky, 2015 (as <i>Rhynchospio</i> sp. 44)	Australia	Australia	KP986493 (1789)	KP986492 (316)	Radashevsky et al. (2016a)
<i>Rhynchospio</i> cf. <i>filiosa</i> Imaizumi, 1991 (as <i>Rhynchospio filiosa</i>)	Japan	USA	KP986489 (1765)	KP986488 (450)	Radashevsky et al. (2016a)
<i>Rhynchospio glutata</i> (Ehlers, 1897)	Strait of Magellan, Chile	Argentina	KJ546281 (1747)	KJ546332 (341)	Radashevsky et al. (2014)
<i>Rhynchospio mizansis</i> Simon, Williams & Henninger, 2018	South Africa	South Africa	MF625258 (1662)	MF625254 (290)	Simon et al. (2019b)
<i>Rhynchospio nharungwi</i> Radashevsky, 2007	Vietnam	Vietnam	KJ546299 (1717)	KJ546343 (499)	Radashevsky et al. (2014)
<i>Scolecopsis</i> Blainville, 1828					
<i>Scolecopsis acuta</i> (Treadwell, 1914)	San Diego, USA	Brazil	KU900479 (683)	–	Rebello & Schertini (unpubl.)
<i>Scolecopsis bonnieri</i> Mesnil, 1896	English Channel	Helgoland, Germany	EU084878 (1711)	–	Vorsepneva et al. (2008)
<i>Scolecopsis chilensis</i> (Hartmann-Schröder, 1962)	Chile	Brazil	KU900475 (689)	–	Rebello & Schertini (unpubl.)
<i>Scolecopsis daphninos</i> Zhou, Ji & Li, 2009	China	China	–	GU362676 (461)	Zhou et al. (2010)
<i>Scolecopsis elaninae</i> Blake, 1983	Ross Sea	Antarctica	KF713431 (333)	KF713470 (398)	Gallego et al. (2014)
<i>Scolecopsis godbodyi</i> (Jones, 1962)	Jamaica	Brazil	KU900477 (441)	–	Rebello & Schertini (unpubl.)
<i>Scolecopsis kudnowi</i> Hartmann-Schröder, 1981	Australia	Lizard Island, Australia	KP636517 (464)	–	Meißner and Görting (2015)
<i>Scolecopsis laoncola</i> (Tzetlin, 1985) (as <i>Acetacalanyx laoncola</i>)	White Sea, Russia	White Sea, Russia	EF569206 (1323)	–	Vorsepneva et al. (2008)
<i>Scolecopsis squamata</i> (Müller, 1806)	Denmark	Sylt, Germany	AF448164 (1848)	–	Bleidorn et al. (2003)
<i>Scolecopsis</i> sp. sco206	–	Eastern Arabian Sea, India	KT900310 (1759)	–	Rengaiyan and Ingole (2018)
<i>Scolecopsis</i> sp. sco207	–	Eastern Arabian Sea, India	KT900311 (1759)	–	Rengaiyan and Ingole (2018)
<i>Spiophanes</i> Grube, 1860					
<i>Spiophanes berkeleyorum</i> Pettibone, 1962	Vancouver Island, Canada	California, USA	MN186816 (1724)	–	Radashevsky et al. (2020a)
<i>Spiophanes bombyx</i> (Claparède, 1870)	Gulf of Naples, Italy	Adriatic Sea, Italy	–	MG878899 (484)	Radashevsky et al. (2020a)
<i>Spiophanes</i> cf. <i>convexus</i> Delgado-Blas, Diaz-Diaz & Viéitez, 2019	Ria de Vigo, Spain	Brittany, France	MG913229 (1742)	MG878902 (505)	Radashevsky et al. (2020a)

Classification	Type locality	Collection locality	Accession number (Length:bp)		Reference
			18S	16S	
<i>Spiophanes duplex</i> (Chamberlin, 1919) (as <i>Spiophanes berkeleyorum</i> isolate 205-48-2)	California, USA	California, USA	MN186817 (1682)	–	Radashevsky et al. (2020a)
<i>Spiophanes hakatensis</i> Radashevsky & Pankova in Radashevsky et al. 2020	British Columbia, Canada	California, USA	MG913241 (1746)	MG878914 (369)	Radashevsky et al. (2020a)
<i>Spiophanes cf. kroeyeri</i> Grube, 1860	Greenland Sea, NW Atlantic	Barents Sea, Norway	MG913238 (1738)	MG878907 (340)	Radashevsky et al. (2020a)
<i>Spiophanes aff. kroeyeri</i> Grube, 1860 (as <i>Spiophanes kroeyeri</i>)	Greenland Sea, NW Atlantic		EU340094 (1769)	EU340080 (544)	Mincek et al. (2009)
<i>Spiophanes norrtii</i> Meißner & Blank, 2009	Mexico	USA	GQ202716 (535)	–	Meißner and Blank (2009)
<i>Spiophanes psinnus</i> Meißner & Huchings, 2003	New South Wales, Australia	Australia	GQ202721 (534)	–	Meißner and Blank (2009)
<i>Spiophanes soederstromi</i> Hartman, 1953	off Rio Grande do Sul, Brazil	Paraná, Brazil	MG913232 (1735)	MG878905 (340)	Radashevsky et al. (2020a)
<i>Spiophanes uschakovii</i> Zachs, 1933	northern Sea of Japan, Russia	Russia	KM998760 (1747)	MG878915 (342)	Radashevsky et al. (2020a)
<i>Spiophanes viriosus</i> Meißner & Huchings, 2003	Australia	Lizard Island, Australia	KP636519 (451)	–	Meißner and Görting (2015)
<i>Spiophanes</i> sp. A	–	East China Sea, South Korea	MG913244 (1732)	MG878920 (417)	Radashevsky et al. (2020a)
<i>Spiophanes</i> sp. RG-2014	–	Anarctica	KF713435 (318)	KF713474 (372)	Gallego et al. (2014)
<i>Sreblospio</i> Webster, 1879	New Jersey, USA	Netherlands	KC686673 (411)	–	van Pel-Heerschap (unpubl.)
<i>Sreblospio</i> sp.	–	India	KY704336 (578)	KY704328 (523)	Vijapur et al. (unpubl.)
<i>Trochochaeta</i> Levinsen, 1884	–	–	–	–	–
<i>Trochochaeta multisetosa</i> (Örsted, 1844)	Danmark	Askerøfjord, Sweden/North Sea, Norway	MN296517 (1728)	MN193552 (341)	Radashevsky et al. (2020a)
Spioninae	–	–	–	–	–
<i>Boccardia</i> Canazzi, 1893	–	–	–	–	–
<i>Boccardia penata</i> (Chlebovitsch, 1959)	Kurile Islands	Sea of Japan, Russia	–	MH493047 (473)	Radashevsky et al. (2019)
<i>Boccardia polybranchia</i> (Haswell, 1885)	New South Wales, Australia	South Africa	KY677891 (1714)	–	Williams et al. (2017)
<i>Boccardia proboscidea</i> Hartman, 1940	CA, USA	CA, USA	KJ546254 (1763)	MH493027 (435)	Radashevsky et al. (2014, 2019)
<i>Boccardia pseudonatrix</i> Day, 1961	Kynsna Estuary, South Africa	South Africa	KY677895 (1719)	–	Williams et al. (2017)
<i>Boccardella</i> Blake & Kudenov, 1978	USA	Incheon, South Korea	MT482710 (1741)	–	Lee et al. (2020)
<i>Boccardella hamata</i> (Webster, 1879)	–	–	–	–	–
<i>Dipolydora</i> Verrill, 1881	–	–	–	–	–
<i>Dipolydora bidentata</i> (Zachs, 1933)	northern Sea of Japan, Russia	Peter the Great Bay, Russia	JX228065 (900)	JX228103 (475)	Radashevsky and Pankova (2013)
<i>Dipolydora capensis</i> (Day, 1955)	South Africa	South Africa	KY677896 (1714)	–	Williams et al. (2017)
<i>Dipolydora candallii</i> (E. Berkeley, 1927)	British Columbia, Canada	South Africa	KY677897 (1714)	–	Williams et al. (2017)
<i>Dipolydora carunculata</i> (Radashevsky, 1993)	Vostok Bay, Russia	Peter the Great Bay, Sea of Japan	JX228073 (900)	JX228113 (475)	Radashevsky and Pankova (2013)
<i>Dipolydora quadrilobata</i> (Jacobi, 1883)	Kiel Canal, Germany	Peter the Great Bay, Sea of Japan	JN048711 (942)	JN048698 (475)	Radashevsky and Pankova (2013)
		Russia	–	MH493041 (309)	Radashevsky et al. (unpubl.)

Classification	Type locality	Collection locality	Accession number (Length:bp)		Reference
			18S	16S	
<i>Dipolydora cf. socialis</i> (Schmarda, 1861)	Chile	South Africa	KY677899 (1715)	–	Williams et al. (2017)
<i>Microspio Mesnil</i> , 1896					
<i>Microspio granulata</i> Blake & Kudenov, 1978	Australia	Lizard Island, Australia	KP636515 (457)	KP636514 (362)	Meißner and Görting (2015)
<i>Polydora</i> Bosc, 1802					
<i>Polydora brevipalpa</i> Zschs, 1933	northern Sea of Japan	China	KP231289 (1725)	–	Ye et al. (2019)
<i>Polydora cornuta</i> Bosc, 1802	South Carolina	Netherlands	KC686637 (421)	–	van Pel-Heerschap (unpubl.)
<i>Polydora neocataca</i> Williams & Radashkevsky, 1999 (as <i>Polydora husuehl</i>)	Rhode Island	China	KF562242 (1792)	KF562235 (511)	Ye et al. (2019)
<i>Polydora linguisensis</i> Ye, Tang, Wu, Su, Wang, Yu & Wang, 2015	China	China	KF562240 (1791)	KF562233 (462)	Ye et al. (2015)
<i>Polydora cf. nuchalis</i> Woodwick, 1953	California, USA	South Africa	KY677903 (1715)	–	Williams et al. (2017)
<i>Polydora triglandata</i> Radashkevsky & Hsieh, 2000	Taiwan	Taiwan	JN048718 (941)	JN048705 (475)	Radashkevsky and Pankova (2013)
<i>Polydora cf. websteri</i> Hartman in Loosanoff & Engle, 1943	Milford Harbor, USA	South Africa	KY677904 (1716)	–	Williams et al. (2017)
<i>Polydrella Augener</i> , 1914					
<i>Polydrella dauyidoffi</i> Radashkevsky, 1996	South China Sea	Nha Trang Bay, Vietnam	–	MG460900 (308)	Radashkevsky et al. (2020b)
<i>Pseudopolydora Czerniavsky</i> , 1881					
<i>Pseudopolydora achata</i> Radashkevsky & Hsieh, 2000	Taiwan	Erhjen River, Taiwan, Taiwan	–	MG460903 (304)	Radashkevsky et al. (2020b)
<i>Pseudopolydora basarginensis</i> (Zschs, 1933)	northern Sea of Japan	Vostok Bay, Sea of Japan, Russia	–	MG460894 (306)	Radashkevsky et al. (2020b)
<i>Pseudopolydora dayii</i> Simon, 2009	South Africa	South Africa	KY677907 (1716)	–	Williams et al. (2017)
<i>Pseudopolydora diopatra</i> Hsieh, 1992	Taiwan	Hsinchu, Taiwan	–	MG460906 (308)	Radashkevsky et al. (2020b)
<i>Pseudopolydora kempii japonica</i> Inajima & Abe, 2017	South Africa	South Africa	AB973933 (1713)	LC107863 (471)	Simon et al. (2019)
<i>Pseudopolydora eriysii</i> Simon, Sato-Okoshi & Abe, 2017	Japan	Vostok Bay, Sea of Japan, Russia	–	MG460897 (306)	Radashkevsky et al. (2020b)
<i>Pseudopolydora paucibranchiata</i> (Okuda, 1937)	Japan	Gulf of Naples, Italy	–	MG460937 (455)	Radashkevsky et al. (2020b)
<i>Pseudopolydora pulchra</i> (Carazzi, 1893)	Gulf of Naples, Mediterranean	Bay of Morlaix, Brittany, France	–	MG460932 (471)	Radashkevsky et al. (2020b)
<i>Pseudopolydora uphonia</i> Simon, Sato-Okoshi & Abe, 2017	South Africa	South Africa	LC107848 (1711)	LC107866 (472)	Simon et al. (2019)
<i>Pseudopolydora vexillosa</i> Radashkevsky & Hsieh, 2000 (as <i>Pseudopolydora</i> sp. B)	Taiwan	Mung Is., Nha Trang Bay, Vietnam	–	MG460890 (295)	Radashkevsky et al. (2020b)
<i>Pseudopolydora</i> sp. A	–	Northern Territory, Australia	–	MG460921 (296)	Radashkevsky et al. (2020b)
<i>Pseudopolydora</i> sp. B (as <i>Pseudopolydora</i> sp. C)	–	Arabian Gulf, Kuwait	–	MG460957 (295)	Radashkevsky et al. (2020b)
<i>Pseudopolydora</i> sp. C (as <i>Pseudopolydora</i> sp. D)	–	Arabian Gulf, Kuwait	–	MG460941 (309)	Radashkevsky et al. (2020b)
<i>Pseudopolydora</i> sp. D (as <i>Pseudopolydora</i> sp. E)	–	Raunefjord, North Sea, Norway	–	MG460960 (305)	Radashkevsky et al. (2020b)
<i>Pseudopolydora</i> sp. Sodwana 32-4	–	South Africa	LC107849 (1724)	LC107867 (473)	Simon et al. (2019)
<i>Spio</i> Fabricius, 1785					
<i>Spio armati</i> Meißner, Bick & Bastrop, 2011 (as <i>Spio</i> sp. LK-2011-2)	Baltic Sea	Baltic Sea	FR823434 (1765)	FR823439 (453)	Meißner et al. (2011)
<i>Spio blakeri</i> Maciolek, 1990	Botany Bay, New South Wales, Australia	Lizard Island, Australia	KP636507 (458)	KP636502 (348)	Meißner and Görting (2015)
<i>Spio filicornis</i> (O. F. Müller, 1776)	Itulifarsutuk, Greenland	Itulifarsutuk, Greenland	FR823431 (1765)	FR823436 (454)	Meißner et al. (2011)
<i>Spio symphyta</i> Meißner, Bick & Bastrop, 2011 (as <i>Spio</i> sp. LK-2011-1)	North Sea	North Sea	FR823433 (1766)	FR823438 (453)	Meißner et al. (2011)

Classification	Type locality	Collection locality	Accession number (Length; bp)		Reference
			18S	16S	
<i>Spio</i> sp. 2573	–	Koni Peninsula, Sea of Okhotsk, Russia	KT200135 (1688)	KT200126 (310)	Radashovsky et al. (2016b)
Outgroup					
Sabellidae Latreille, 1825					
<i>Laonome</i> Malmgren, 1866					
<i>Laonome</i> sp.	–	Pärnu Bay, Baltic Sea	KP793139 (1813)	KP793138 (450)	Kotta et al. (2015)
<i>Sabella</i> Linnaeus, 1767					
<i>Sabella pavonina</i> Savigny, 1822	Plymouth	–/Brittany, France	U67144 (1726)	AY340482 (476)	Nador & Grant (unpubl.), Rousset et al. (2007)

aLRT) with 5,000 replicates (Guindon et al. 2010), approximate Bayes (aBayes) test (Anisimova et al. 2011), and ultrafast bootstraps (UFBoot) with 5000 replicates (Hoang et al. 2018). SH-aLRT \geq 80%, aBayes \geq 0.95, and UFBoot \geq 95% were defined as robust statistical support. All the sequences newly generated in this study were deposited in the DDBJ/ENA/GenBank nucleotide sequence database under accession numbers LC545853 to LC545925 and LC595683 to LC595767 (Table 1). Part of the sequences used in the present study was reported in the previous studies (see Table 1). The planktonic spionid larvae were identified by comparing larval and adult sequences obtained in the present study and/or by larval morphology.

Results

The 18S and 16S rRNA gene analyses of larval and adult spionids

Nuclear 18S and mitochondrial 16S rRNA gene sequences of adult spionid polychaetes were successfully obtained from 48 species belonging to 14 genera (Table 1).

In the phylogenetic analysis using only the sequences obtained in the present study (i.e., without DBJ/ENA/GenBank sequences), species from the genera *Paraprionospio*, *Pseudopolydora*, *Scolecopsis*, *Spio*, and *Spiophanes* were recovered as monophyletic groups with robust statistical supports (i.e., SH-aLRT \geq 80%, aBayes \geq 0.95, and UFBoot \geq 95%, Fig. 2). Species belonging to the genus *Boccardia* except for *B. pseudonatrix* and those from the genus *Dipolydora* except for *D. armata* and *D. cf. commensalis* were recovered as monophyletic groups with robust statistical supports. The tribe Polydorini and subfamily Spioninae were also recovered as monophyletic although UFBoot of monophyly of the subfamily Spioninae was with low support (\leq 95%).

In the phylogenetic analysis with the sequences obtained in the present study and from DDBJ/ENA/GenBank databases, species belonging to the genera *Poecilochaetus*, *Laonice*, *Marenzelleria*, *Pseudopolydora*, *Pygospio*, *Rhynchospio*, *Scolecopsis*, *Spio* + *Microspio*, and *Spiophanes*, were recovered as monophyletic groups with robust statistical supports (Fig. 3). Species belonging to the genus *Polydora* were recovered as a monophyletic group but with low UFBoot support. Tribe Polydorini + *Pygospio*, that plus *Glandulospio*, and *Spiophanes* + *Trochochaeta* were also recovered as monophyletic groups with robust statistical supports. The genera *Poecilochaetus* and *Trochochaeta*, which were previously considered as belonging to the family Poecilochaetidae and Trochochaetidae, respectively, were recovered as ingroup taxa of the family Spionidae with robust statistical supports (Fig. 3).

In total, 41 species belonging to 14 genera of planktonic spionid larvae were identified (Table 1; Fig. 2), 27 of which were identified by the 100% or nearly 100% match between the sequences obtained from adult and larvae (Fig. 2). The other 14 species of spionid larvae were identified to species or genus level based on their phylogenetic position and/or larval morphology. Tentative larval diagnosis for each genus and larval identification keys to species of each genus based on the morphol-

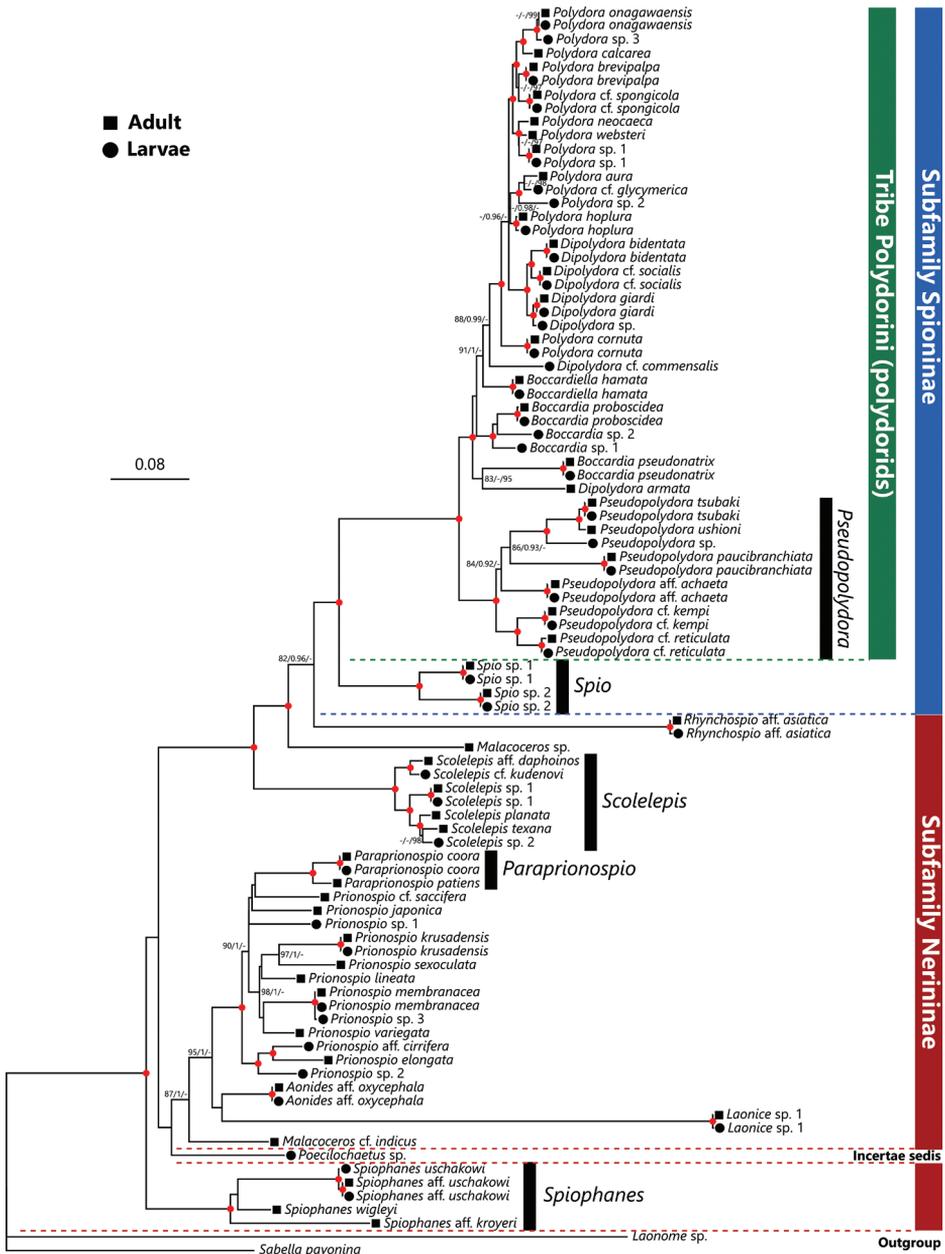


Figure 2. Maximum Likelihood tree inferred from nuclear 18S and mitochondrial 16S rRNA gene sequences of spionids obtained from Japan in the present and previous studies (provided in Table 1). The gene sequences of adult and larval spionid polychaetes are indicated by solid squares and circles in front of each species name, respectively. SH-aLRT/approximate Bayes support/ultrafast bootstrap support values of $\geq 80\%$, ≥ 0.95 , $\geq 95\%$, respectively are given beside the respective nodes. Nodes with red circles indicate triple high support values of SH-aLRT ≥ 80 , approximate Bayes support ≥ 0.95 , and ultrafast bootstrap support ≥ 95 . The scale bar represents the number of substitutions per site. Sequences of *Laonome* sp. and *Sabella pavonina* Savigny, 1822 obtained from DDBJ/EMBL/GenBank were used for outgroup rooting.

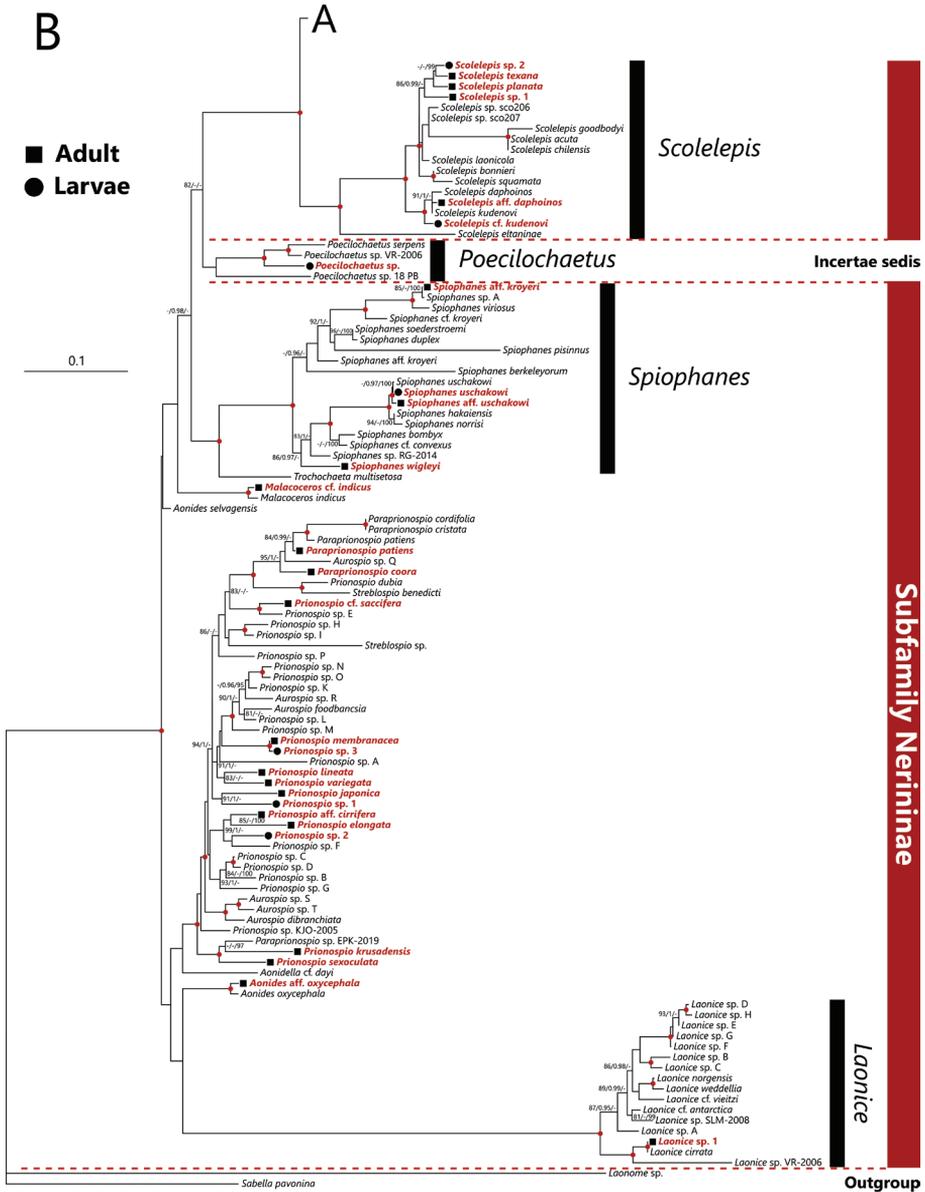


Figure 3. Continued.

Identification key to genera of the spionid larvae in northeastern Japan

- 1 Two pairs of red or dark red eyes present; distinct black pigmentation or melanophore (branching black chromatophores) absent; gastrotrochs from chaetiger II or III onwards in all following chaetigers (except *Rhynchospio*: not in all following chaetigers) [subfamily Nerininae] 2
- Three pairs of black eyes present (most lateral two pairs of eyes often double-eyes: see Hacker 1896; Hannerz 1956); distinct black pigmentation present;

- gastrotrochs from chaetiger III, V, or VII onwards not in all following chaetigers [subfamily Spioninae] **8**
- Two pairs of dark red eyes present; small black pigmentation present laterally between parapodia on every chaetigers; gastrotrochs from chaetiger I onwards in all following chaetigers..... **Genus *Poecilochaetus***
- 2 Lateral parts of peristomium well developed and distinctly demarcated from prostomium **3**
- Lateral parts of peristomium less developed and less demarcated from prostomium..... **5**
- 3 Prostomium not pointed anteriorly, more or less stumpy; lateral parts of peristomium not clearly demarcated **4**
- Prostomium pointed anteriorly and tip of prostomium terminates in a tapered tip; lateral parts of peristomium clearly demarcated as large peristomial umbrella **Genus *Scolelepis***
- 4 Parapodia well differentiated; long larval chaetae only in notopodia..... **Genus *Rhynchospio***
- Parapodia less differentiated; serrated larval chaetae occur in both noto- and neuropodia **Genus *Laonice***
- 5 Slender, moderately long in overall shape; body not transparent; some pigmentation of various colors on pharynx, proctodaeum, prostomium, peristomium, pygidium, and/or various locations of the body in late larvae **6**
- Slender, fairly long in overall shape with numerous chaetigers; body nearly transparent; pigmentation almost completely absent except red or green pigmentation on pharynx and/or pygidium **7**
- 6 Body not rich in yolk; larval chaetae on first chaetiger medium length; pharynx not colored in black; prostomium rounded anteriorly **Genus *Spiophanes***
- Body rich in yolk; larval chaetae on first chaetiger fairly long especially in early larvae; pharynx colored in black; prostomium rectangular anteriorly (prostomium extended and tapered anteriorly in juvenile) **Genus *Aonides***
- 7 Prostomium anteriorly rounded; lateral parts of peristomium relatively demarcated from prostomium; quite large and long larvae with well-developed branchiae in late larvae..... **Genus *Paraprionospio***
- Prostomium anteriorly rounded; lateral parts of peristomium less demarcated from prostomium; long and slender larvae with no or less developed branchiae..... **Genus *Prionospio***
- 8 Overall body shape long and slender **9**
- Overall body shape thick/slender and fusiform **11**
- 9 Modified chaetae in chaetiger V present in late larvae; larval chaetae on first chaetiger medium length **10**
- Modified chaetae in chaetiger V absent; larval chaetae on first chaetiger fairly long **Genus *Spio***
- 10 Pairs of large branching dorsal melanophores present..... **Genus *Polydora***
- Pairs of large branching dorsal melanophores absent..... **Genus *Dipolydora***

- 11 Lateral prostomium expansion moderate (except for *Boccardia* sp. 2); mid-dorsal melanophores arranged in a single row (except *B. pseudonatrix*); vestibule or pharynx pigmented with black or brown..... **Genus *Boccardia***
- Lateral prostomium expand greatly; arrangement of dorsal melanophores not arranged in a single row; vestibule or pharynx not pigmented with black or brown **12**
- 12 A mid-dorsal branching melanophore on first chaetiger absent; more than two pairs of dorsal black pigmentation spots/bands on each chaetiger..... **Genus *Boccardiella***
- A mid-dorsal branching melanophore on first chaetiger present (except for *Pseudopolydora* cf. *kempi*: mid-dorsal melanophore on first chaetiger usually absent); one or two pairs of dorsal melanophores on each chaetiger **Genus *Pseudopolydora***

Description of larval morphology

Family SPIONIDAE Grube, 1850

Subfamily NERININAE Söderström, 1920

Genus *Aonides* Claparède, 1864

Larval diagnosis. The overall shape slender. Prostomium rounded or rectangular anteriorly. The lateral parts of the peristomium more or less demarcated from prostomium. Two pairs of red eyes present. Melanophore absent, some brown or dark pigmentation may be present in pharynx and pygidium. Larval chaetae coarsely or slightly serrated. Larval chaetae in first chaetiger very long, extend beyond pygidium in late trochophore and early nectochaete stages. Nototrochs develop in late larval stages. Gastrotrochs occur in all chaetigers from chaetiger II onwards. Two pairs of pygidial cirri develop in late larval stage. The body of early larvae covered by egg envelope, yellowish opaque appearance with abundant yolk. Two parallel rows of encircling vesicles of egg envelope present in pretrochophore and trochophore stages. Holopelagic lecithotrophic development unique among spionids (Hannerz 1956; Blake and Arnofsky 1999, as *Dispio uncinata* Hartman, 1951: see Radashevsky et al. 2011; Blake 2006, as *D. uncinata*).

Aonides aff. *oxycephala* (Sars, 1862)

Fig. 4A–C

Larval morphology. Remnants of egg envelope apparent in early trochophore (Fig. 4A). In ten-chaetiger larvae, egg envelope becomes incorporated into larval cuticle, two pairs of red eyes arranged in an approximately straight line (Fig. 4B). Larval chaetae on first chaetiger long especially in early larvae (Fig. 4A, B). Late larvae long and slender in shape (Fig. 4C). Prostomium rectangular anteriorly in larval stages, considerably ex-

tended and tapered in juvenile stage. Lateral parts of peristomium moderately demarcated from prostomium. Black pigment in pharynx. Pigmentation absent except in the eyes and pharynx. Pygidium acquires two pairs of dorsal cirri in late larvae (Fig. 4C).

Remarks. Adult individuals of this species were collected from muddy bottom sediment at 22 m depth in Onagawa Bay in January 2011 and 2012 using a Smith-McIntyre grab sampler. Adult morphology agrees with the description of *A. oxycephala* by Imajima (1989). *Aonides oxycephala* originally described from Norway has been reported worldwide and is considered cosmopolitan. However, these reports may comprise a series of similar or sibling species, as pointed out by Radashevsky (2015). The gene sequences obtained in the present study were 100% match in 18S rRNA but 8.6% (29/337 bp) different in 16S rRNA from that of *A. oxycephala* from France (MG913226 and MG878895). Therefore, the species collected in the present study was referred to *A. aff. oxycephala*. The larvae and adults were confirmed to match (18S: 1753/1753, 16S: 447/448 bp) using molecular data (Fig. 2).

Planktonic larvae were found in Onagawa Bay from October to December. In early larval stages, the larvae of this species are similar to those of *Laonice* sp. (Fig. 4D); but larval chaetae are longer, and the body is yolkier and opaquer in this species. The larval morphology of *A. aff. oxycephala* is similar to that of *A. oxycephala* described by Hannerz (1956). However, the peristomium of the former species is more developed and demarcated from the prostomium compared to the latter. Black pigmentation of the pharynx in late larval stages was not reported by Hannerz (1956).

Genus *Laonice* Malmgren, 1867

Larval diagnosis. Overall shape short, thick, and fusiform. Prostomium stumpy, rectangular, notched anteriorly. Lateral parts of peristomium clearly demarcated from prostomium. The short palps attached to outer end of lateral parts of peristomium. Two pairs of red eyes present. Melanophores and pigmentation absent except eyes. Nototrochs absent. Gastrotrochs occur in all chaetigers from chaetiger III onwards. Well-developed serrated larval chaetae occur both in noto- and neuropodia, notochoetae characteristically introverted toward medial line of dorsal side. Early larvae covered by egg envelope (Hannerz 1956; Plate and Husemann 1994).

Laonice sp. 1

Fig. 4D

Larval morphology. Remnants of egg envelope apparent in early trochophore (Fig. 4D). Two pairs of red eyes located in approximately a straight line, lateralmost pair larger in early larvae. Parapodia weakly differentiated; serrated larval chaetae introverted toward medial line of dorsal side. The body opaque yellowish with abundant yolk internally. Pigmentation absent except in the eyes.

Remarks. Adult individuals of this species were collected from bottom sediments at 22 m depth in Onagawa Bay in December 2011 using a Smith-McIntyre grab sampler. To date, two *Laonice* species, *L. cirrata* (Sars, 1851) and *L. japonica* (Moore, 1907) have been recorded from Japan. Sikorski (2002) indicated that *L. cirrata*, a previously presumed widespread species, is probably limited to Norway and adjacent regions. This was supported by a molecular study that suggested previously unrecognized diversity within this species (Bogantes et al. 2018). *Laonice japonica*, originally described as *Spionides japonicus* from Japan and later considered as synonymous with *L. cirrata* (e.g., Söderström 1920; Berkeley and Berkeley 1936; Okuda 1937; Imajima and Hartman 1964; Foster 1971), was reexamined and considered a valid species by Maciolek (2000) and Sikorski (2011). However, even after that, since *L. cirrata* has been recorded from Japan (e.g., Imajima 2006, 2009, 2011), the validity of these records is ambiguous and might represent different species. In addition to these two species, unidentified *Laonice* sp. was also reported from Japan by Imajima (1990c) but it is unclear whether the species is identical to the species reported here. Although the 18S rRNA gene sequences obtained in the present study match (1731/1731 bp, except for gaps) with *Laonice cirrata* sequences from Russia in DDBJ/EMBL/GenBank (KM998754), because taxonomic knowledge on this genus in Japan is still limited and the 18S rRNA gene is relatively conservative, this species was referred to *Laonice* sp. The larvae and adults were confirmed to match (18S: 1754/1754, 16S: 500/504 bp) using molecular data (Fig. 3).

Larvae of this species were rare in the planktonic community found in Onagawa Bay in September 2011 and October 2012. Although two parallel rows of encircling vesicles of egg envelope, similar to those of *Aonides* pretrochophore and trochophore stages, were reported in oocytes of *Laonice* species (Radashevsky and Lana 2009), this characteristic was not observed in early larval stages with egg envelope in the present study (Fig. 4D). Blake (2006) described the larval development of *Laonice* sp. from California; however, the identification of these larvae is doubtful because they seem to lack serrated larval notochaetae introverted toward the medial line of the dorsal side, which are characteristic of *Laonice* larvae.

***Laonice* sp. 2**

Fig. 4E

Larval morphology. Late larvae thick and stumpy in shape (Fig. 4E). Prostomium stumpy, somewhat notched at tip. Lateral parts of peristomium well demarcated from prostomium. In late larvae, two pairs of red eyes are arranged in a trapezoidal shape, the medial pair bigger and situated anteriorly. Short palps developed in late larvae, attached to outer end of lateral parts of peristomium. Parapodia weakly differentiated; serrated larval chaetae in both noto- and neuropodia; notochaetae characteristically introverted toward medial line of dorsal side. Gut dark green in color internally. Pigmentation absent except in the eyes.

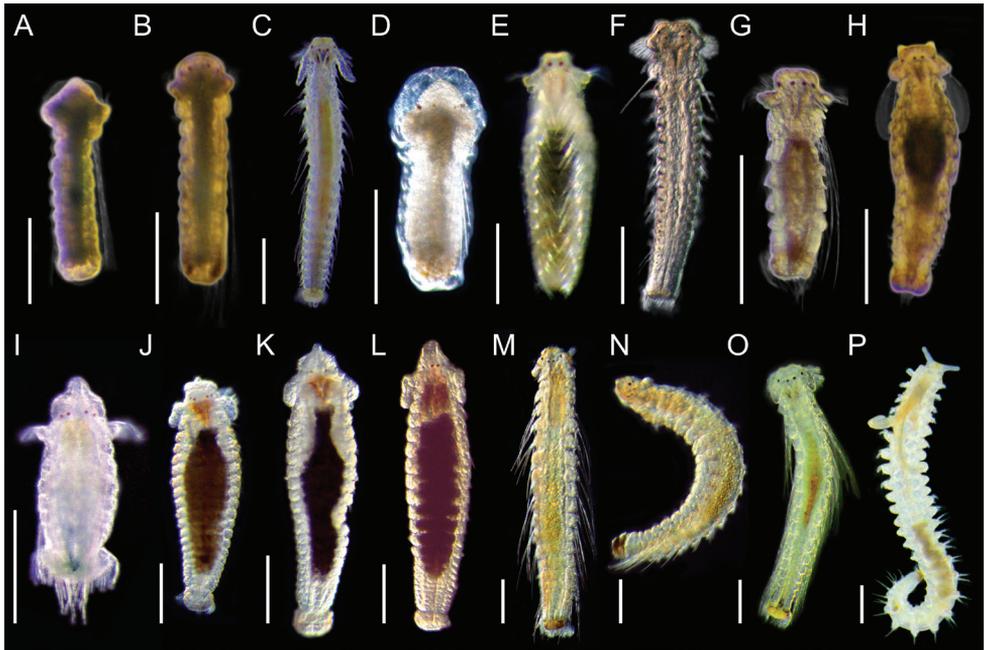


Figure 4. Light micrographs showing morphologies of living spionid larvae of *Aonides*, *Laonice*, *Rhyncospio*, *Scolelepis*, and *Spiophanes* **A–C** *Aonides* cf. *oxycephala*, dorsal view of early planktonic (**A**), 8-chaetiger (**B**), and 18-chaetiger larvae (**C**) **D** *Laonice* sp. 1, dorsal view of early planktonic larva **E** *Laonice* sp. 2, dorsal view of 12-chaetiger larva **F** *Poecilochaetus* sp., dorsal view of 17-chaetiger larva **G**, **H** *Rhyncospio* aff. *asiatica*, dorsal view of 6-chaetiger (**G**) and 12-chaetiger larvae (**H**) **I** *Scolelepis* cf. *kudenovi*, dorsal view of 7-chaetiger larva **J**, **K** *Scolelepis* sp. 1, dorsal view of 17-chaetiger (**J**) and 19-chaetiger larvae (**K**) **L** *Scolelepis* sp. 2, dorsal view of 20-chaetiger larva **M**, **N** *Spiophanes uschakowi*, dorsal (**M**) and lateral view (**N**) of 18-chaetiger larvae **O**, **P** *Spiophanes* aff. *uschakowi*, dorsal view of 16-chaetiger larva (**O**) and 27-chaetiger juvenile (**P**). Scale bars: 300 μ m.

Remarks. Adult individuals of this species were not collected in the present study. Only one individual of larva of this species were collected in Habu Port in June 2016. Even though the 18S and 16S rRNA gene sequences were not obtained, the larvae were identified as belonging to *Laonice* because the larval morphology of this species agrees with that of *L. cirrata* described by Hannerz (1956) and *L. cf. cirrata* described by Plate and Husemann (1994). However, the hooded hooks in neuropodia described by Hannerz (1956) were not observed in the specimens of the present study (nor in those reported by Plate and Husemann [1994]); this may be because the larvae collected here were less developed and the hooks were reported only from chaetiger XIV onwards.

Genus *Paraprionospio* Caullery, 1914

Larval diagnosis. Overall shape long and slender, large in size (> 4 mm) and number of chaetigers (> 35 chaetigers) at metamorphosis. Prostomium rounded. Lateral parts

of peristomium moderately demarcated from prostomium. Two pairs of red or dark red eyes present. Pigmentation absent except eyes and some reddish pigmentation on pygidium. Nototrochs absent. Gastrotrochs occur in all chaetigers from chaetiger II onwards. Branchiae well developed and elongated in late larvae. Long larval chaetae may be absent in chaetiger II (Berkeley and Berkeley 1961, as *Prionospio*; Carrasco 1976; Yokoyama 1981, 1996; Blake and Arnofsky 1999).

***Paraprionospio coora* Wilson, 1990**

Fig. 5A, B

Larval morphology. Long and thin in shape, quite large and long body with numerous chaetigers. Prostomium anteriorly rounded, lateral lips elevated from the ventrolateral side of prostomium (Fig. 5A). Late larvae acquire caruncle extending posteriorly from posterior part of prostomium (Fig. 5B). Peristomium fuses with the first larval segment at late larval stage (Fig. 5B). First pair of branchiae well developed, branchial pinnation still absent. Two pairs of red eyes arranged in somewhat trapezoidal shape, lateral pair kidney-shaped, situated anteriorly. Posterior part of pygidium pigmented reddish brown, anal cirri develop in late larvae.

Remarks. Adult individuals of this species were collected from muddy bottom sediments at 22 m depth in Onagawa Bay in December 2011 by using a Smith-McIntyre grab sampler. Adult morphology agrees with the description of *P. coora* by Yokoyama (2007), and therefore this species was referred to *P. coora*. The larvae and adults were confirmed to match (18S: 1754/1754, 16S: 500/500 bp) using molecular data (Fig. 2).

Only three planktonic larvae of this species were found in Onagawa Bay in November 2011 and Sasuhama in January 2013. The morphological characteristics and size of these larvae are similar to those in previous descriptions of the species from the same genus (Yokoyama 1981, 1996). However, the larvae of *P. coora* lack red pigmentation on the dorsolateral side of the lateral lips, which characterizes the larvae of *Paraprionospio patiens* Yokoyama, 2007 (Yokoyama 1981; as *P. pinnata*: see Yokoyama 2007). Additionally, lamellae of the first pair of branchiae in *P. coora* are less developed in late larvae with more than 30 chaetigers (Fig. 5B) compared with the larvae of *P. patiens* (Yokoyama 1981) and *Paraprionospio cordifolia* Yokoyama, 2007 (Yokoyama 1996, as *Paraprionospio* sp. form B: see Yokoyama 2007). Yokoyama (1981) suggested that the larvae of *Paraprionospio* are the largest in size and number of chaetigers at metamorphosis among the spionid larvae. However, late larvae of *Poecilochaetus* exceeding 5 mm (Magalhães et al. 2015) and with more than 40 chaetigers are often reported (Hannerz 1956; Plate and Husemann 1994).

Genus *Poecilochaetus* Claparède in Ehlers, 1875

Larval diagnosis. Overall shape long and slender, large in size (> 5 mm) and number of chaetigers (> 30 chaetigers) at metamorphosis. Body transparent, characterized by

total absence of pigmentation except pairs of small pigment spot between parapodia or ventro-lateral side of each chaetiger. Two pairs of red or dark red eyes present. Gastrotrochs from chaetiger I onwards in all following chaetigers, gastrotrochs in first and second chaetigers represented by solitary lateral patches of cilia and complete gastrotrochs occur from third chaetiger onwards. Nototrochs absent. Larvae prior to ca. 30–40 chaetiger stages remain in metatrochophore stage, characterized by absence of functional parapodia for swimming and presence of well-developed proto-, telo-, and gastrotrochs for swimming. Metatrochophore have broadened trapezoidal prostomium with tactile cilia in anterior part, broad and low caruncle, provisional larval chaetae, pygidium without anal cirri. Larval stage after metatrochophore stage (often called nectosoma) characterized by reduced trochs, the presence of functional parapodia, and rapid serpentine swimming behavior. In nectosoma stage, caruncle, nuchal lobes, a pair of palps, parapodia, cirriform or digitiform dorsal and ventral postchaetal lobes, and two pairs of anal cirri on pygidium develop gradually (Thorson 1946; Hannerz 1956; Berkeley and Berkeley 1961; Reddy and Mohan 1982; Plate and Husemann 1994; Magalhães et al. 2015).

***Poecilochaetus* sp.**

Fig. 4F

Larval morphology. Overall shape long and slender. Two pairs of dark red eyes present. Metatrochophore larvae with 17 chaetigers have broadened trapezoidal prostomium with tactile cilia in anterior part, broad and low caruncle, provisional larval chaetae, and well developed prototrochs (Fig. 4F). Body of metatrochophore larvae transparent, characterized by small pigment spot between parapodia from chaetiger II onwards. Pygidium without anal cirri. Incomplete anterior fragment of nectosoma larvae characterized by extremely long body, rounded prostomium, broad and low caruncle, reduced prototrochs, parapodia with digitiform dorsal postchaetal lobes, and rapid serpentine swimming behavior. The body of nectosoma larvae transparent and small pigment spot laterally on each side of chaetigers. Occipital antenna and pair of palps not observed.

Remarks. Adult individuals of this species were not collected in the present study. Even though the 18S and 16S rRNA gene sequences obtained from larvae in the present study did not match any of the *Poecilochaetus* sequences from DDBJ/EMBL/GenBank, this species was referred to *Poecilochaetus* sp. because specimens formed a monophyletic clade with the other *Poecilochaetus* species with robust statistical support (Fig. 3).

Planktonic larvae of this species were collected in Onagawa Bay in August 2010 and January 2013. The larval morphology of this species is similar to that of *Poecilochaetus serpens* Allen, 1904 described by Hannerz (1956) and Plate and Husemann (1994). However, the former species differs from the latter by not having yellow chromatophores on the “head” and pygidium as described by Plate and Husemann (1994). The

***Prionospio membranacea* Imajima, 1990**

Fig. 5E–G

Larval morphology. Long and thin in shape. Prostomium rounded anteriorly, lateral parts of peristomium not especially well demarcated. Two pairs of red or dark red eyes arranged somewhat in trapezoidal shape, lateral pair kidney-shaped and situated anteriorly. Small caruncle develop in late larvae, extends posteriorly from posterior part of prostomium (Fig. 5G). In late larvae branchial anlagen occur from chaetiger II, pygidium acquires anal cirri (Fig. 5G). Palps not yet developed in 15- and 20-chaetiger larva (Fig. 5E, F) but developed in 24-chaetiger larva (Fig. 5G). Pigmentation absent except eyes.

Remarks. Adult individuals of this species were collected from muddy bottom sediments at 22 m depth in Onagawa Bay in December 2011 by using a Smith-McIntyre grab sampler. The species was referred to *P. membranacea* as the adult morphology agrees with the descriptions of this species by Imajima (1990b). The larvae and adults were confirmed to match (18S: 1752/1752 and 1747/1750 except for gaps, 16S: 502/505 bp) using molecular data (Fig. 2). Planktonic larvae of this species were found in Onagawa Bay during August to October.

***Prionospio* spp. 1 and 2**

Fig. 5H, I

Larval morphology. Long and thin in shape. Prostomium rounded anteriorly, lateral parts of the peristomium not especially well demarcated. Two pairs of red or dark red eyes arranged somewhat in trapezoidal shape, lateral pair kidney-shaped and situated anteriorly. Small caruncle develop in late larvae, extends posteriorly from posterior part of the prostomium. In *Prionospio* sp. 2, branchial anlagen occur from chaetiger II, pygidium acquires anal cirri (Fig. 5I). Pigmentation absent except eyes in *Prionospio* sp. 1. Gut pigmented in orange in *Prionospio* sp. 2. Palps developed in 19-chaetiger larva of *Prionospio* sp. 2.

Remarks. Two unidentified species of planktonic larvae of the genus *Prionospio* other than *P. krusadensis* and *P. membranacea* were collected from Onagawa Bay. The adult individuals of these species were not collected in the present study. Even though the 18S and 16S rRNA gene sequences obtained from these larvae in the present study did not match any *Prionospio* sequences obtained in the present study nor with those registered in DDBJ/EMBL/GenBank, these species were referred to *Prionospio* sp. 1 and 2 as the larvae were similar to the other *Prionospio* species in their morphology and gene sequences (Figs 2, 3). The larvae of *Prionospio* sp. 1 and 2 are similar to each other and to that of *P. membranacea*, and it is difficult to distinguish among them based only on their morphology.



Figure 5. Light micrographs showing the morphologies of living spionid larvae of genera *Paraprionospio* and *Prionospio* **A, B** *Paraprionospio coora*, lateral view of 25-chaetiger (**A**) and 33-chaetiger larvae (**B**) **C, D** *Prionospio krusadensis*, lateral view of 17-chaetiger larvae **E–G** *Prionospio membranacea*, lateral view of 15-chaetiger (**E**), 20-chaetiger (**F**), and 24-chaetiger larvae (**G**) **H**, *Prionospio* sp. 1, lateral view of 11-chaetiger larva **I** *Prionospio* sp. 2, lateral view of 19-chaetiger larva. Scale bars: 300 μ m.

Genus *Rhynchospio* Hartman, 1936

Larval diagnosis. Overall body shape short and thick. Prostomium broad and straight or slightly notched anteriorly. Lateral parts of peristomium clearly demarcated from prostomium. Palps attached to outer end of lateral parts of peristomium. Two pairs of red or black eyes present. Faint yellow pigment may be present in anterior part of prostomium and posterior part of pygidium. In late larvae, pair of prominent anterolateral processes on prostomium developed. Melanophore absent, black or yellowish pigmentation occur in some species. Nototrochs weakly developed, occur in all chaetigers except first chaetiger. Gastrotrachs occur regularly in every other chaetiger from chaetiger III onwards (Carrasco 1976; Radashevsky 2007).

Rhynchospio aff. *asiatica* sensu Radashevsky et al., 2014

Figs 4G, H, 6

Larval morphology. Overall body shape short and thick in relation to length. Prostomium broad, stumpy, somewhat notched anteriorly. Peristomium well developed, forming wide collar on sides of prostomium. Two pairs of red or dark red eyes arranged

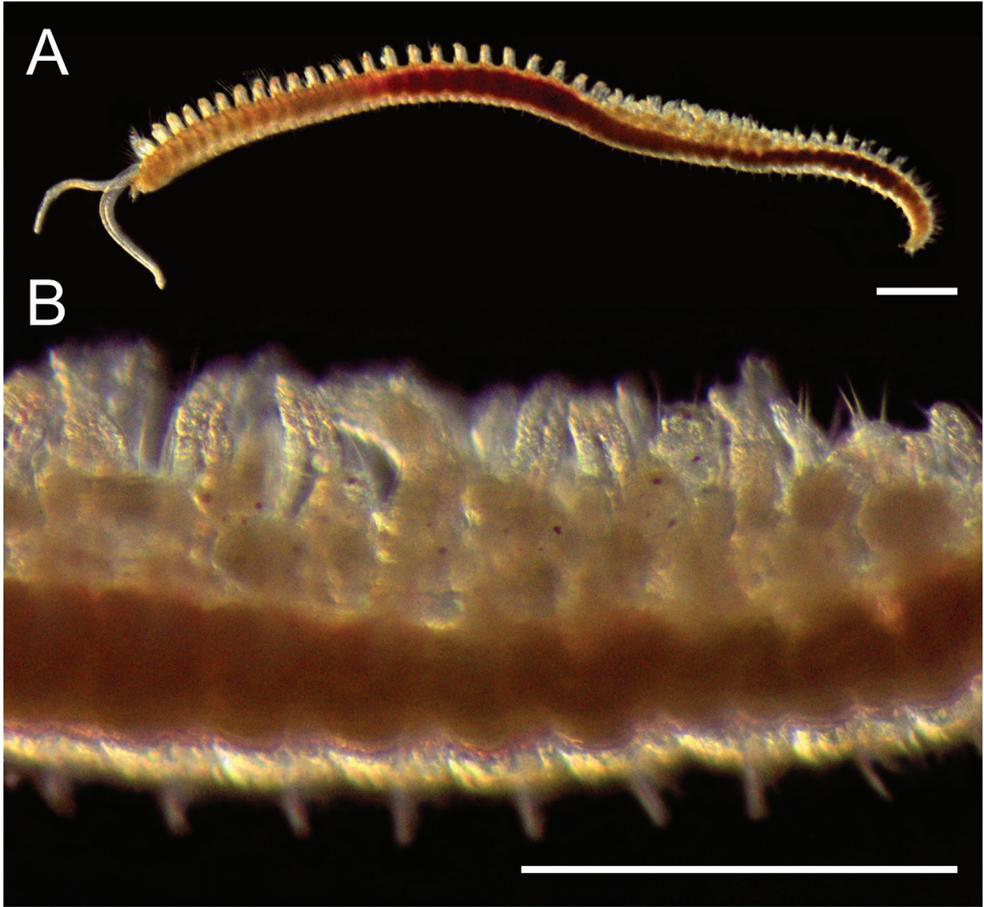


Figure 6. Light micrographs showing dorsal brooding of *Rhynchospio* aff. *asiatica*. Hermaphroditic individual broods their larvae between dorsal branchiae on posterior chaetigers. Scale bars: 1 mm.

in straight line, lateral pair in kidney-shape. Parapodia strongly differentiated in late larvae. Larval chaetae occur only in notopodia. Pygidium large and round, acquires dorsal cirri in late larvae. Late larvae have two antero-lateral processes on prostomium (Fig. 4H). Pigmentation usually absent, some individuals have brownish pigmentation on peristomium, dorsum, and/or pygidium, and/or two medial black pigmentation ventrally on approximately chaetiger VI and anterior margin of the pygidium.

Remarks. Adult individuals of this species were collected from intertidal and shallow subtidal sandy or muddy bottom sediments in Gamo Lagoon in January 2011 and Sasuhama in September 2011 by using a hand-scoop. To date, three *Rhynchospio* species, *R. foliosa* Imajima, 1991, *R. gutaea* (Ehlers, 1987), and *R. tuberculata* Imajima, 1991, have been recorded from Japan (Imajima 1991a). However, extensive morphological and molecular studies revealed the absence of records of *R. glutaea* from the

northern Pacific Ocean (Radashevsky 2007; Radashevsky et al. 2014, 2016a). Radashevsky et al. (2014) also referred to *R. arenicola* Hartman, 1936, *R. asiatica* Chlebovich, 1959, *R. aff. asiatica*, and *R. glutaea* as members of the *R. glutaea* complex because they resembled each other so closely. Adult morphology and 18S and 16S rRNA gene sequences of *Rhynchospio* specimens obtained in the present study agree (18S: 1716/1716, 16S: 486/492 bp) with those of *R. aff. asiatica* (Fig. 3) from South Korea (KJ546296) reported by Radashevsky et al. (2014); therefore, this species was referred to *R. aff. asiatica* sensu Radashevsky et al. (2014). The larvae and adults were confirmed to match (18S: 1783/1783, 16S: 471/477 bp) using molecular data (Fig. 2).

This species is recorded from Japan for the first time in the present study. The brooding of larvae beneath dorsal branchiae in this species was observed in September 2011 (Fig. 6). The larvae adhere to their parents and are enclosed by branchiae present on the posterior chaetigers (26th–39th chaetigers). The larvae are retained on the parents' dorsum even when the parent individuals leave their tube, unless the parent is disturbed. Larvae seemed to be released at around the 3-chaetiger stage; the fact that planktonic larvae with more than three chaetigers were commonly collected from plankton supports this observation. Similar dorsal larval brooding was reported in other *Rhynchospio* species (Levin 1982; Radashevsky 2007; Radashevsky et al. 2014) and in *Streblospio benedicti* Webster, 1879 (Levin 1982, 1984).

Planktonic larvae were found in Onagawa Bay, Gamo Lagoon, and Sasuhama in almost every season of the study period, but few were found in winter season (November to March). Larval morphology of this species resembles that of *R. glutaea* and *R. nhatrangii* Radashevsky, 2007 described by Carrasco (1976) and Radashevsky (2007), respectively. The overall larval morphology of *Rhynchospio* species is quite similar to that of the genus *Malacoceros* described in Hannerz (1956, as *Scolecopsis*), but it differs in the latter having three pairs of black eyes, the most lateral pairs with double-eyes.

Genus *Scolecopsis* Blainville, 1828

Larval diagnosis. Overall shape thick and fusiform. Prostomium pointed anteriorly, terminates in retractile, muscular tip. Lateral parts of peristomium clearly demarcated from prostomium, forming large peristomial umbrella. Short palps on lateral-most parts of peristomium. Two pairs of red eyes present. Melanophore absent, black, brown, orange, red, or green pigmentation patches often present in body surface, pharynx, gut, and/or proctodaeum. Nototrochs present or absent. Gastrotrochs occur in all chaetigers from chaetiger II or III onwards. Pygidium large, inflated, and surrounded by thick telotroch (Okuda 1946, as *Spio filicornis*; Hartman 1941, as *Nerinides*; Thorson 1946, as *Nerine* in part see Hannerz 1956; Hannerz 1956, as *Nerine* and *Nerinides*; Imajima 1959, as *Nerinides*; Dean and Hatfield 1963, as *Nerinides*; Carrasco 1976, as *Nerine* and *Nerinides*; Plate and Husemann 1994; Scheltema et al. 1997; Blake and Arnofsky 1999; Blake 2006).

Identification key to species of the larvae belonging to the genus *Scolelepis* in northeastern Japan

- 1 Pharynx and pygidium colored green; pygidium very broad and horseshoe-shaped *Scolelepis cf. kudenovi*
- Pharynx pigmented orange and gut pigmented brown; Pygidium broad and spherical shaped **2**
- 2 Prostomium sharply tapered anteriorly; gut diverticula not strongly segmented *Scolelepis sp. 1*
- Prostomium bluntly tapered anteriorly; gut diverticula strongly segmented....
..... *Scolelepis sp. 2*

Scolelepis cf. kudenovi Hartmann-Schröder, 1981

Fig. 4I

Larval morphology. Thick and fusiform in shape. Prostomium pointed anteriorly, terminates in retractile, muscular tip. Lateral parts of peristomium clearly demarcated from prostomium, forming peristomial umbrella. Peristomial umbrella carrying well-developed prototroch. Two pairs of red eyes arranged in somewhat trapezoidal shape, medial pair situated anteriorly. Greenish pigment in pharynx and proctodaeum. Pygidium very broad and horseshoe-shaped.

Remarks. Only three individuals of early larvae of this species were collected from Sasuhama in January 2012. The 18S rRNA gene sequences obtained in the present study for these specimens match (464/464 bp) that of *S. kudenovi* from Lizard Island, Australia (KP636517: Meißner and Götting 2015). Since the species identification is unreliable because of the short reference sequence, this species was referred to *S. cf. kudenovi*.

The sequence of an adult individual, which collected from the surf zone of the sandy beach in Rishiri Island and previously identifies as *Scolelepis kudenovi* (Abe et al. 2019c) as the morphology agrees with the descriptions of *S. kudenovi* by Imajima (1992) and Meißner and Götting (2015), 100% matched with that of the larvae of *Scolelepis cf. kudenovi* in 18S rRNA gene (1819/1819 bp) but largely differed in 16S rRNA gene (462/505 bp). Because the 16S rRNA gene of the adult individual was rather closer to *S. daphoinos* (430/455 bp) from China (GU362676, Zhou et al. 2010), it is referred to *S. aff. daphoinos* in the present study (Table 1, Figs 2, 3).

Scolelepis sp. 1

Fig. 4J, K

Larval morphology. Thick and fusiform in overall shape. Prostomium pointed anteriorly as a small process, tip of prostomium terminates in retractile, muscular tip. Lateral

parts of peristomium clearly demarcated from prostomium, forming large peristomial umbrella. Peristomial umbrella carrying well-developed prototroch. Short palps developed in late larvae, attached on lateral-most parts of peristomium. Two pairs of red eyes arranged in an approximately straight line. Pharynx pigmented orange and the gut pigmented brown. Pygidium broad and spherical.

Remarks. Adult individuals of this species were collected from muddy sediments at 22 m depth in Onagawa Bay in December 2011 by using a Smith-McIntyre grab sampler. These adults were morphologically identified as *Scoelelepis*, but they were not identified to species level as these specimens were all incomplete and in poor condition. As the 18S and 16S rRNA gene sequences obtained in the present study did not match any available *Scoelelepis* sequences (Figs 2, 3), this species was referred to *Scoelelepis* sp. 1. Planktonic larvae of this species were collected in Onagawa Bay in October during the study period. The larvae and adults were confirmed to match using molecular data (Fig. 3).

Scoelelepis sp. 2

Fig. 4L

Larval morphology. Thick and fusiform in overall shape. Prostomium bluntly pointed anteriorly, terminates in retractile, muscular tip. Lateral parts of peristomium clearly demarcated from prostomium, forming large peristomial umbrella. Peristomial umbrella carrying well-developed prototroch. Short palps developed in late larvae, attached on lateral-most parts of peristomium. Two pairs of red eyes arranged in an approximately straight line. Pharynx widely pigmented orange and the gut pigmented brown. Pygidium broad and spherical shaped.

Remarks. No adult individuals of this species were collected in the present study. Even though the 18S and 16S rRNA gene sequences obtained from larvae in the present study did not match any available *Scoelelepis* sequences, this species was referred to *Scoelelepis* sp. 2 as the larvae constitute a monophyletic clade with the other *Scoelelepis* species with robust statistical support (Figs 2, 3).

Planktonic larvae of this species were found in Onagawa Bay in September. The larvae of this species are quite similar to those of *Scoelelepis* sp. 1; however, the prostomium is more broadly pointed anteriorly and the gut diverticula are more strongly segmented in the former species.

Genus *Spiophanes* Grube, 1860

Larval diagnosis. Overall shape slender. Prostomium small or broad, rounded or slightly notched anteriorly. Lateral parts of the peristomium slightly or moderately demarcated from the prostomium, palps on lateral-most parts of peristomium. Two pairs of red eyes present. In late larvae, a pair of prominent or small antero-lateral processes on prosto-

mium are often developed. Melanophore absent, some pigmentation patches of various colors are present on pharynx, proctodaeum, prostomium, peristomium, pygidium, and/or various locations of the body in late larvae. Nototrochs occur in all chaetigers from chaetigers II–IV onwards. Gastrotrochs occur in all chaetigers from chaetiger II onwards (Thorson 1946; Hannerz 1956; Carrasco 1976; Plate and Husemann 1994; Blake 2006).

Identification key to species of the larvae belonging to the genus *Spiophanes* in northeastern Japan

- 1 Prostomium small; pharynx and proctodaeum colored in brown; yellow or yellow-brown pigments on prostomium, peristomium, and pygidium; a pair of small lateral processes on the prostomium developed in late larvae; small red pigment spots present on lateral part of posterior chaetigers.....
*Spiophanes uschakowi*
- Prostomium broad; pharynx and proctodaeum colored in black; a pair of prominent lateral processes on the prostomium developed in late larvae; small red pigment spots absent.....*Spiophanes aff. uschakowi*

Spiophanes uschakowi Zachs, 1933

Fig. 4M, N

Larval morphology. Overall shape slender. Prostomium small and rounded anteriorly. Two pairs of red or dark red eyes present, lateral pair situated anteriorly. Late larvae bear very small antero-lateral processes on prostomium. Lateral parts of peristomium slightly demarcated from prostomium, palps attached on lateral-most parts of peristomium. Nototrochs occur from chaetiger IV onwards (Fig. 4N). Yellow pigments on the prostomium and pygidium, intense yellow-brown pigment on peristomium, inside of pharynx, and pygidium. Small red pigment spots present on lateral part of body (Fig. 4N). Black pigment in pharynx and proctodaeum absent.

Remarks. No adult individuals of this species were collected in the present study. However, gene sequences obtained from larvae of this species were almost identical (18S: 1732/1732, 16S: 341/342 bp) to that of *S. uschakowi* (KM998760 and MG878915) from Russia (Radashevsky et al. 2020a); therefore, this species was referred to *S. uschakowi*. Imajima (1991b) recorded four *Spiophanes* species from Japan: *S. kroeyeri* Grube, 1860 (as *S. kroeyeri*); *S. japonicum* Imajima, 1991; *S. bombyx* (Claparède, 1870); and *S. urceolata* Imajima, 1991. Then, Meißner and Hutchings (2003) synonymized *S. urceolata* with *S. wigleyi* Pettibone, 1962. Additionally, the specimens from Japan formerly identified as *S. bombyx* were morphologically reexamined and identified as *S. cf. uschakowi* by Meißner and Blank (2009). In the present study, the presence of *S. uschakowi* in Japan was further supported by molecular analysis.

Only a few larvae of this species were collected in Onagawa Bay in November 2011. The overall larval morphology of this species somewhat resembles that of *S. kroeyeri* de-

scribed by Hannerz (1956) in the following aspects: prostomium is relatively small and anteriorly rounded, the peristomium is not quite sharply demarcated from the prostomium, nototrochs occur from chaetiger IV onwards, and the brown pigmentation is present on the pygidium and inside the pharynx. However, *S. kroyeri* lacks small red pigment spots on the lateral part of the body and lateral processes on the prostomium even in 22-chaetiger larvae.

***Spiophanes* aff. *uschakowi* Zachs, 1933**

Fig. 4O, P

Larval morphology. Overall shape slender. Prostomium broad and slightly notched anteriorly. In late larvae, a pair of prominent antero-lateral processes on prostomium developed. Two pairs of red or dark red eyes present, lateral ones situated somewhat anteriorly. Lateral parts of peristomium moderately demarcated from prostomium, palps on lateral-most parts of peristomium. Nototrochs occur from chaetiger II onwards. Pharynx and proctodaeum black in color internally. Pygidium acquires dorsal cirri in late larvae. Some brownish, yellowish, or greenish pigmentation occurred on various locations of body in late larvae.

Remarks. Adult individuals of this species were collected from muddy bottom sediments at 22 m depth in Onagawa Bay in April and May 2012 by using a Smith-McIntyre grab sampler and from bottom sediments of the shallow subtidal zone in Sasuhama in February 2012. Adult morphology agrees with the description of *S. cf. uschakowi* by Meißner and Blank (2009) as well as with that of *S. bombyx* by Imaijima (1991b). However, the 18S and 16S rRNA gene sequences of this species did not match those of *S. uschakowi* obtained from DDBJ/EMBL/GenBank (KM998760): there was a 0.29% (5/1750 bp) and 0.88% (3/342) difference, respectively between these two species. Therefore, this species was referred to *S. aff. uschakowi*. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

Planktonic larvae of this species were collected in Onagawa Bay in November 2011 and in Sasuhama in February 2012. The larval morphology of *S. aff. uschakowi* is different from that of *S. uschakowi* in the following aspects: prostomium of the former is broad and slightly notched anteriorly, whereas that of the latter is relatively small and anteriorly rounded; the peristomium of the former is well demarcated from the prostomium, but that of the latter is relatively less demarcated; nototrochs of the former occur from chaetiger II onwards, whereas those of the latter occur from chaetiger IV onwards; pigmentation inside the pharynx is black in the former but brown in the latter; and black pigmentation in the proctodaeum is present in the former but absent in the latter. Black pigmentation in the pharynx and proctodaeum were also reported in the larvae of *S. bombyx* (Hannerz 1956), *S. cf. bombyx* (Blake 2006), and *S. duplex* (Chamberlin, 1919) (Blake 2006). However, the illustrations of *S. cf. bombyx* provided by Blake (2006: fig. 13.10C, D) are seemingly more similar to larvae of *Rhynchospio* than to those of *Spiophanes*.

Subfamily SPIONINAE Söderström, 1920

Genus *Boccardia* Carazzi, 1893

Larval diagnosis. Overall shape thick or slender and fusiform. Prostomium small or broad and rounded anteriorly. Three pairs of black eyes present, most lateral often double-eyes. Dorsal pigment pattern consists of single row of branching melanophores in most species, some species lack distinct dorsal melanophore. Lateral pigments present or absent. Ventral pigments absent. Nototrochs occur in all chaetigers except first two chaetigers. Gastrotrochs occur in irregular pattern. Modified chaetae develop in chaetiger V in late larvae (Söderström 1920, as *Polydora natrix*; Hartman 1941; Carrasco 1976; Woodwick 1977; Blake and Kudenov 1981; Duchêne 1984, 1989; Guérin 1991; Gibson 1997; Blake and Arnofsky 1999; Gibson and Smith 2004; Blake 2006; Kamel et al. 2010; Oyarzun and Brante 2015; Blake 2017).

Identification key to species of the larvae belonging to the genus *Boccardia* in northeastern Japan

- 1 Distinct dorsal melanophore absent; faint yellow coloration present on all over body *Boccardia pseudonatrix*
- Mid-dorsal melanophores arranged in a single row **2**
- 2 Dorso-lateral spots of black pigment absent; overall body shape thick and fusiform; pharynx pigmented with black *Boccardia* sp. **2**
- Dorso-lateral spots of black pigment present; overall body shape slender and fusiform; black pigment at pharynx present or absent **3**
- 3 A prominent row of mid-dorsal melanophores from chaetiger III; dorso-lateral spots of black pigment present on chaetigers VII and VIII; black pigment in pharynx absent *Boccardia proboscidea*
- A prominent row of mid-dorsal melanophores from chaetiger IV; dorso-lateral spots of black pigment present from chaetiger V onwards; black pigment in pharynx present *Boccardia* sp. **1**

Boccardia proboscidea Hartman, 1940

Fig. 7A, B

Larval morphology. Slender and fusiform in overall shape, widest in middle of body. Prostomium rounded and slightly notched anteriorly. Three pairs of eyes present, most median pair rounded, lateral pairs double-eyes. Body entirely faint green in color. A prominent row of dorsal melanophores occurs medially from chaetiger III, lateral black pigment spots present on chaetigers VII and VIII in late larvae (Fig. 7B). Pygidium has dorsal gap, pigmented with weak dark color. Internally, vestibule light brown, gut either yellow or brown. Gastrotrochs on chaetigers V and VII.

Remarks. Adults of this species were non-boring and collected from mud deposits in crevices of shells of living *Crassostrea gigas* (Thunberg, 1793) (recently assigned to *Magallana*: see Backeljau 2018) oysters in Sasuhama in May 2011 and February 2016. Adult morphology agrees with the description of *B. proboscidea* by Sato-Okoshi (2000). The 18S and 16S rRNA gene sequences obtained in the present study match (18S: 1748/1748, 16S: 435/435 bp) that of *B. proboscidea* from USA (KJ546254) reported by Radashevsky et al. (2014) (Fig. 3). Therefore, this species was referred to *B. proboscidea*. The larvae and adults were confirmed to match (18S: 1768/1768, 16S: 472/472 bp) using molecular data (Fig. 2).

Planktonic larvae of this species were rare, and only one 15-chaetiger larva (Fig. 7B) was collected in Sasuhama in May 2011. Another 9-chaetiger larvae, which accidentally hatched from its egg capsule in an adult tube during the process of extraction of the adult specimens (Fig. 7A), was also collected on the same date. *Boccardia proboscidea* has been reported to have poecilogonous development (Gibson 1997; Oyarzun et al. 2011). However, Sato-Okoshi (2000) reported that Japanese populations only show lecithotrophic development, with no (or a very short) planktonic stage after hatching. The larval morphology of this species agrees with the description of that of *B. proboscidea* documented in Hartman (1941), Woodwick (1977), Blake and Kudenov (1981), Gibson (1997), Gibson and Smith (2004), Kamel et al. (2010), and Oyarzun and Brante (2015). The dorsal pigment pattern of these larvae resembles that of the larvae of *B. tricuspa* (Hartman, 1939) described by Carrasco (1976, as *B. proboscidea*; *vide* Blake and Kudenov 1978), *B. natrix* (Söderström, 1920) described by Söderström (1920, as *Polydora natrix*), and *B. columbiana* Berkeley, 1927 described by Blake and Arnofsky (1999) and Blake (2006) in having a single row of mid-dorsal melanophores. However, the dorsal pigment pattern of the larvae of *B. tricuspa* differs from that of *B. proboscidea* in having branching dorsal melanophores on chaetiger I and in lacking small black lateral pigment spots on chaetigers VII and VIII. The larvae of *B. natrix* also lack small black lateral pigment spots on chaetigers VII and VIII. *Boccardia columbiana* has extensively branching mid-dorsal melanophores from chaetiger II onward, whereas mid-dorsal melanophores are less branching and start from chaetiger III in *B. proboscidea*.

***Boccardia pseudonatrix* Day, 1961**

Fig. 7C

Larval morphology. Slender and fusiform in overall body shape. Prostomium rounded with a slight anterior notch. Three pairs of eyes present, most median pair rounded and lateral pairs double-eyes. Body entirely faint yellow. Dorsal melanophore absent, slight black pigmentation present (Fig. 7C, left). Pygidium with dorsal gap, pigmented with yellow. Internally, the vestibule and pharynx brown or black, gut green in color. Gastrotrochs on chaetigers V and VII.

Remarks. Adults of this species were non-boring and collected from mud deposits in crevices of shells of living *C. gigas* oysters in Tomiura. Adult morphology (see Abe

et al. 2019b) agrees with the descriptions of *B. pseudonatrix* from South Africa (Day 1967; Simon et al. 2010) and Australia (Sato-Okoshi et al. 2008, as *B. knoxi*; see Walker 2013). The 18S rRNA gene sequences obtained in the present study completely match (1714/1714 bp) that of *B. pseudonatrix* from South Africa (KY677895) reported by Williams et al. (2017) (Fig. 3). Therefore, this species was referred to *B. pseudonatrix*.

Boccardia pseudonatrix has been reported to have adelphophagic larvae with a short or absent planktonic phase (Sato-Okoshi et al. 2008; Simon 2015). The larvae herein reported accidentally hatched from egg capsules in an adult tube during the process of extraction of the adult specimens.

***Boccardia* sp. 1**

Fig. 7D–F

Larval morphology. Slender and slightly fusiform in overall shape, widest in anterior part of body. Prostomium rounded anteriorly. Three pairs of black eyes present, most median pair rounded and lateral pairs double-eyes. Body entirely yellowish in color. A prominent row of dorsal melanophores occurs medially from chaetiger IV, lateral black pigment spots present from chaetiger V onwards (Fig. 7E). Pygidium with dorsal gap, pigmented with black color. Vestibule black, gut orange in color internally. Larval chaetae on first chaetiger long especially in early larvae (Fig. 7D). Gastrotrachs on chaetigers III, V, VII, X, and XIII.

Remarks. No adult individuals of this species were collected in the present study. The 18S and 16S rRNA gene sequences obtained from the larvae did not match any available *Boccardia* sequences, but this species is very similar to the other *Boccardia* species in larval morphology and gene sequences (Figs 2, 3). Therefore, this species was referred to *Boccardia* sp. 1.

Planktonic larvae of this species were collected from Onagawa Bay in April, May, November, and December 2011, and January, February, March, and May 2012. The overall body shape of these larvae is slender and slightly fusiform, similar to those of *B. proboscidea*. However, other larval morphological characteristics differ between these two species: overall body color is faint yellow in the former species and faint green in the latter; the larval chaetae are longer in the former species than in the latter, especially in early larvae (Fig. 7D); lateral black spots are present on chaetiger V onwards in the former species but only on chaetigers VII and VIII in the latter species.

***Boccardia* sp. 2**

Fig. 7G

Larval morphology. Thick and fusiform in overall shape, widest at middle part of body. Prostomium extensively broad and anteriorly rounded. Three pairs of black eyes present, most median pair rounded and lateral pairs double-eyes. Body entirely faint



Figure 7. Light micrographs showing the morphologies of living spionid larvae of genera *Boccardia*, *Boccardiella*, and *Dipolydora* **A, B** *Boccardia proboscidea*, dorsal view of accidentally hatched 9-chaetiger (**A**) and 15-chaetiger larvae (**B**) **C** *Boccardia pseudonatrix*, dorsal (left) and ventral (right) view of accidentally hatched 10-chaetiger larvae **D–F** *Boccardia* sp. 1, dorsal view of 9-chaetiger (**D**) and dorsal (**E**) and ventral view (**F**) of 17-chaetiger larvae **G** *Boccardia* sp. 2, dorsal view of 15-chaetiger larva **H, I** *Boccardiella hamata*, dorsal view of 16-chaetiger (**H**) and 18-chaetiger larvae (**I**) **J, K** *Dipolydora bidentata*, dorsal view of 13-chaetiger (**J**) and 18-chaetiger larvae (**K**) **L** *Dipolydora* cf. *commensalis*, dorsal view of 21-chaetiger larva **M, N** *Dipolydora giardi*, dorsal view of 19-chaetiger (**M**) and lateral view of 21-chaetiger larvae (**N**) **O** *Dipolydora* cf. *socialis*, dorsal view of 18-chaetiger larva **P** *Dipolydora* sp., dorsal view of 7-chaetiger larva. Scale bars: 300 μ m.

green in color in late larvae. A prominent row of dorsal ramified melanophores occurs medially from chaetiger IV onwards, lateral black pigment spots absent. Pygidium with dorsal gap, pigmented with weak dark color. Internally, vestibule black, gut orange in color. Gastrotrichs on chaetigers III, V, VII, X, and XIII.

Remarks. No adult individuals of this species were collected in the present study. The 18S and 16S rRNA gene sequences herein obtained from the larvae did not match any of the available *Boccardia* sequences, but this species is similar to the other *Boccardia* species in larval morphology and gene sequences (Figs 2, 3); therefore, this species was referred to *Boccardia* sp. 2.

Planktonic larvae of this species were collected from Onagawa Bay in December 2010 and November and December 2011, from Sasuhama in January 2013, and from Sendai Port in December 2010. The larval morphology of this species differs from that of other *Boccardia* larvae in having a thick and fusiform body shape.

Genus *Boccardiella* Blake & Kudenov, 1978

Larval diagnosis. Overall shape thick and fusiform. Prostomium extensively broad and rounded anteriorly. Three pairs of black eyes present, most lateral pairs usually double-eyes. More than two pairs of dorsal melanophores from chaetiger III onwards. Lateral and ventral pigments present. Nototrochs occur in all chaetigers except first two. Gastrotrochs occur in irregular pattern. Modified chaetae develop in chaetiger V in late larvae (Rullier 1960, as *Polydora redeki*; Dean and Blake 1966, as *Boccardia*).

Boccardiella hamata (Webster, 1879)

Fig. 7H, I

Larval morphology. Thick and fusiform in overall shape, widest at middle part of body. Prostomium broad and anteriorly rounded, usually dusky brown anteriorly. Three pairs of black eyes present, most median pair rounded, lateral pairs usually double-eyes, occasionally divided into respective eyes. Black pigmentation usually presents ventrally on each lateral lip, occasionally absent. Dorsal pigmentation basically consists of a pair of medial bands, lateral branching melanophores, and small pigment patch at the base of notopodia in each chaetiger from chaetiger III onwards (Fig. 7H). These melanophores undergo expansion and contraction, sometimes coalescing to cover almost the whole of the dorsal surface as ramified pigmentation (Fig. 7I). Four transverse lines of black pigmentation sometimes fused as a single transverse band in chaetiger I. One or two pairs of lateral black pigmentation on chaetiger II. Two rows of band-shaped ventral pigmentation usually located on posterior edges of some chaetigers posterior to second chaetiger. A pair of black pigment patches on pygidium. Gastrotrochs on chaetigers III, V, VII, X, and XIII.

Remarks. Adults of this species were non-boring and collected from mud deposits in crevices of shells of living *C. gigas* oysters in Sasuhama in May 2011 and February 2016. Adult morphology agrees with the description of *B. hamata* by Sato-Okoshi (2000). Therefore, this species was referred to *B. hamata*. The larvae and adults were confirmed to match (18S: 1772/1772, 16S: 480/481 bp) using molecular data (Fig. 2).

Planktonic larvae of this species were frequently collected from Onagawa Bay, Gobu-ura, and Sasuhama in July and August. The larval morphology of this species agrees with that of *B. hamata* described by Dean and Blake (1966, as *Boccardia*).

Genus *Dipolydora* Verrill, 1879

Larval diagnosis. Overall shape slender or slightly fusiform. Prostomium small rounded anteriorly. Three pairs of black eyes present, most lateral pairs often double-eyes. Ramified melanophore between central and lateral pairs of eyes usually absent, but present in some species (e.g., *D. cf. commensalis*). Dorsal pigment pattern consists of two rows of band or spot shaped melanophores or a transverse row of small melanophores at each chaetiger in most species, while some species have single row of branching mid-

dorsal melanophores (e.g., *D. cf. commensalis*) or completely lack melanophores (e.g., *D. armata*). Lateral and ventral pigments are present or absent. Nototrochs occur in all chaetigers except the first two chaetigers. Gastrotrochs occur in irregular pattern. Modified chaetae develop in chaetiger V in late larvae (Andrews 1891, as *Polydora*; Hannerz 1956, as *Polydora*; Hatfield 1965, as *Polydora*; Blake 1969, as *Polydora*; Carrasco 1976, as *Polydora*; Day and Blake 1979, as *Polydora*; Radashevsky 1989, as *Polydora*; Plate and Husemann 1994, as *Polydora*; Lewis 1998; Blake 2006; Blake 2017).

Identification key to species of the larvae belonging to the genus *Dipolydora* in northeastern Japan

- 1 Mid-dorsal single row of distinct melanophores present
..... *Dipolydora cf. commensalis*
- Arrangement of dorsal melanophore otherwise **2**
- 2 Black pigmentation on lateral peristomium present; a pair of band-shaped ventral black pigment present; notopodial lobes tipped with orange pigment in late larvae..... *Dipolydora cf. socialis*
- Black pigmentation on lateral peristomium absent; ventral black pigment absent; notopodial lobes not tipped with orange pigment..... **3**
- 3 Some patchy black pigment between head and first chaetiger present
..... *Dipolydora bidentata*
- Black pigment between head and first chaetiger absent **4**
- 4 Two pairs of dorsal black pigment spots present; yellow-brown pigment on anterior margin of prostomium absent..... *Dipolydora giardi*
- A pair of dorsal black pigment spots present; weak yellow-brown pigment on anterior margin of prostomium present *Dipolydora sp.*

Dipolydora bidentata (Zachs, 1933)

Fig. 7J, K

Larval morphology. Overall shape elongated. Prostomium and pygidium small. Three pairs of black eyes present, most lateral pairs double-eyes. Black pigmentation patches on lateral peristomium absent. Some patchy black pigment occurs between head and first chaetiger. Two dorsal black bands begin on chaetiger II and continue to posterior end. Dorso-lateral pigment extend posteriorly along lateral side found on most chaetigers. Some black or brown pigment may occur on pygidium. Ventral pigment absent. Gastrotrochs on chaetigers V, VII, X, XIII, and XV.

Remarks. Adults of this species are shell-borers and were collected from shells of wild *C. gigas* oysters in Sasuhama in July 2012. Adult morphology agrees with the description of *D. bidentata* by Sato-Okoshi (1999). The 18S and 16S rRNA gene sequences obtained in the present study match (18S: 900/900, 16S: 473/475 bp) that of *D. bidentata* from Russia (JX228065) reported by Radashevsky and Pankova (2013) (Fig. 3). Planktonic larvae of this species were collected from Onagawa Bay in Novem-

ber 2011 and from Sasuhamama in February 2012. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

***Dipolydora cf. commensalis* (Andrews, 1891)**

Fig. 7L

Larval morphology. Overall shape elongated and slender. Prostomium small but wider than body and rounded anteriorly. Three pairs of eyes present, most lateral pairs double-eyes of kidney-shaped appearance. Ramified melanophores present around eyes. Black pigmentation on lateral peristomium absent. Median row of ramified melanophores from chaetiger I onwards. Lateral and ventral pigments absent. A central black pigment spot and a pair of dark brown pigments on pygidium. Pygidium has a dorsal notch and lacks appendages. Gastrotrochs on chaetigers III, V, VII, X, XIII, XV, XVII, XIX, XXI, and XXIII. Modified chaetae develop in chaetiger V in late larvae.

Remarks. No adults of this species were collected in the present study. The 18S and 16S rRNA gene sequences obtained from the larvae of this species neither match nor constitute a monophyletic clade with any of the other available spionid sequences (Figs 2, 3). However, this species was tentatively identified as *D. cf. commensalis* based on its larval morphology, as it includes the characteristic dorsal pigment pattern of larvae of *D. commensalis* as described by Andrews (1891), Hatfield (1965), Blake (1969), and Radashevsky (1989) (described as *Polydora commensalis* by all of these authors). The combination of a slender body and a single dorsal median row of distinct melanophores from chaetiger I to the end of the body is distinctive among spionid larvae and has not been reported for any other spionid species. Currently, there are no records of *D. commensalis* from Japan; however, the presence of this species in Japan is expected as it has been reported from the Asian continental coast of the Sea of Japan and the Kurile Islands (Radashevsky 1993). This species is an obligate symbiont of hermit crabs (Blake 1996; Williams and McDermott 1997), but little effort was devoted to collecting hermit crab shells in the present study.

Notably, the results of the phylogenetic analysis in the present study showed that *D. cf. commensalis* deviates from the monophyletic clade constituted by many other *Dipolydora* species. This result supports the suggestion by Blake (1971) that *D. commensalis* may represent a distinct genus as its morphology deviates widely from other species of the genus *Polydora* and *Dipolydora*.

Only three individuals of planktonic larvae of this species were collected in Sasuhamama in January 2013. A small patch of lateral black pigments on the anterior margin of each chaetiger in late larvae was described in Hatfield (1965) and Blake (1969). However, these pigments were not observed in the present study, as in Andrews (1891) and Radashevsky (1989). Although Hatfield (1965) and Blake (1966) noted the high similarity between the larval morphologies of *D. commensalis* and *Polydora hermaphroditica* Hannerz, 1956, the adult morphologies of these two species were reported to be completely different (Bhaud 1966). The dorsal pigment pattern of *P. hermaphroditica* larvae reported by Hannerz (1956) rather resembles those of *Polydora glycymerica* and

Polydora cf. *glymymerica* larvae reported by Radashevsky (1989) and in the present study, respectively.

Dipolydora giardi (Mesnil, 1896)

Fig. 7M, N

Larval morphology. Overall body shape elongated and slender. Prostomium small and rounded anteriorly. Three pairs of black eyes arranged in transverse row, most lateral pairs double-eyes. Black pigmentation on lateral peristomium absent. Two pairs of dorsal black spots begin on chaetiger III onwards and continue to posterior end, sometime medial pair in first 2–4 chaetigers band-shaped. Small medial spot of black pigment on posterior margin of each chaetiger usually from chaetigers III, rarely from V or VI, in late larvae. Two small spots of black pigmentation occur lateral to the medial black pigmentation from approximately chaetiger VI or VII. A small black pigment spot, not visible dorsally, present on antero-lateral edges from chaetiger II onwards. Black pigment occurs on pygidium. Rust-colored pigment occurs in pharynx. Ventral pigment absent. Some metamorphosing larvae reduce pigmentation over the entire body and present whitish appearance with eyes fused and appears as one pair (Fig. 7N). Gastrotrochs on chaetigers III, V, VII, X, XIII, and XV.

Remarks. Adults of this species are shell-borers and were collected from shells of cultured *Mizuhopecten yessoensis* (Jay, 1857) (formerly as *Patinopecten yessoensis*) scallops suspended in Onagawa Bay in December 2010. Adult morphology agrees with the description of *D. giardi* by Sato-Okoshi (1999). Therefore, this species was referred to *D. giardi*. The larvae and adults were confirmed to match using molecular data (Fig. 3).

The 18S rRNA gene sequences of this species are very similar to that of *D. capensis* 1PE from South Africa (KY677896) reported by Williams et al. (2017), but there is a slight difference between their sequences (0.12% difference: 2/1714 bp). It is unclear whether this difference indicates that these two are the same or different species because two different 18S rRNA gene sequences have been reported from South Africa and are currently under the same species name (*D. capensis*) (Table 2, Fig. 3). No gene sequences of *D. cf. giardi* previously recorded from South Africa (Simon 2011) are available.

Planktonic larvae of this species were collected from Onagawa Bay in December 2010, June, July, October, November, and December 2011, and December 2012, and from Sasuhama in January 2013. The larval morphology of this species was previously described from California by Day and Blake (1979, as *Polydora giardi*). The morphology and dorsal pigment pattern of late larvae described by these authors resembles that reported here, but there are slight differences: two golden pigment spots present on either side of chaetiger I in the former description but absent in the latter; two small lateral melanophores present on chaetigers I and II in the former description but absent in the latter; a medial black pigmentation beginning from chaetiger II onwards in the former description but from chaetiger III onwards in the latter description; and two small spots of black pigmentation lateral to the medial black pigmentation starting from chaetiger III in the former description but from more posteriorly in the latter

description. These differences between specimens from Japan and California may indicate that they are different species, or that intraspecific variation occurs in larval dorsal pigmentation. Day and Blake (1979) pointed out differences in reproductive traits between the Californian and French populations and suggested the existence of two different species. Therefore, more than one species may be included under the name of *D. giardi*, which currently is reported with a worldwide distribution (Radashevsky and Petersen 2005).

***Dipolydora cf. socialis* (Schmarda, 1861)**

Fig. 7O

Larval morphology. Late larvae usually thick and slightly fusiform in shape, although not as much as the larvae of *Boccardia* sp. 2 (Fig. 7G), *Boccardiella hamata* (Fig. 7H, I), and *Pseudopolydora* species (Fig. 9A–I). Anterior margin of prostomium has yellow-brown pigment. Three pairs of black eyes arranged in transverse row, most lateral pairs double-eyes. Band of black pigment on each lateral part of the peristomium. First dorsal black melanophores occur as paired bands on chaetiger III and continue through to chaetiger V. From chaetiger VI, two pairs of dorsal black spots or bands occur and continue to posterior end of body. From chaetiger IV or V and continuing posteriorly, clusters of small black pigmented cells present in transverse row on dorsal posterior half of chaetigers. Lateral pigment found on late larvae on chaetiger II. Each notopodial lobe tipped with orange pigment, small patch of black pigment at the base of notopodial lobes. Ventral pigment consists of paired bars on posterior border of chaetigers, commencing with chaetiger II. Some black or brown pigment may occur on pygidium. Gastrotrochs occur on chaetigers III, V, VII, X, XIII, XV and XVII.

Remarks. Adults of this species were non-boring and collected from muddy bottom sediment at 22 m depth in Onagawa Bay in December 2010 by using a Smith-McIntyre grab sampler and from bottom sediments of shallow subtidal zone in Sasuhama in April 2013. Adult morphology agrees with the description of *D. socialis* by Sato-Okoshi (2000). The 18S rRNA gene sequence obtained in the present study showed a 0.35% (6/1715 bp) difference with that of *D. cf. socialis* from South Africa (KY677899) reported by Williams et al. (2017), which may indicate that these two are different species. The 18S rRNA gene sequence obtained in the present study rather closer to that of *D. carunculata* (940/942 bp match) reported by Radashevsky and Pankova (2013), but the 16S gene sequence showed a 2.3% (11/475 bp) difference with that of *D. carunculata*. As described above, since the taxonomic status of the species reported here is uncertain, we tentatively referred to it as *D. cf. socialis*.

Planktonic larvae of this species were collected from Onagawa Bay in November 2010 and 2011, and in October 2012. The larvae and adults were confirmed to match (18S: 1770/1770, 16S: 473/475 bp) using molecular data (Fig. 2). The larval morphology of this species agrees with that of *D. socialis* described as *Polydora socialis* by Blake (1969) and Carrasco (1976).

***Dipolydora* sp.**

Fig. 7P

Larval morphology. Overall body shape slender. Prostomium small and rounded anteriorly. Anterior margin of prostomium has weak yellow-brown pigment. Three pairs of black eyes present in transverse row, most lateral pairs double-eyes. Black pigmentation on lateral peristomium absent. A pair of dorsal black spots present on chaetiger III onwards. A small medial spot of black pigment on posterior margin of chaetiger III. Some black pigment occurs on pygidium. Ventral pigment absent. Gastrotrochs occur on chaetigers III and V.

Remarks. No adult individuals of this species were collected in the present study. The 18S and 16S rRNA gene sequences obtained from larvae in the present study did not match any available *Dipolydora* sequences. As the larvae specimens formed a monophyletic clade with the other *Dipolydora* species (excluding *D. armata*, *D. capensis* 1GG, *D. cf. commensalis*, and *D. quadrilobata*) with robust statistical supports (Figs 2, 3), this species was referred to *Dipolydora* sp.

Genus *Polydora* Bosc, 1802

Larval diagnosis. Overall shape slender or slightly fusiform. Prostomium broad or small and rounded anteriorly. Three pairs of black eyes present, most lateral pairs often double-eyes. Some species have ramified melanophore between central and lateral pairs of eyes. Dorsal pigmentation usually consists of two rows of bands, spots, or branching melanophores in most species, while some species have a single row of mid-dorsal melanophores (e.g., *Polydora* cf. *glycymerica*). Lateral and ventral pigments present or absent. Nototrochs occur in all chaetigers except first two. Gastrotrochs occur in irregular pattern. Modified chaetae develop on chaetiger V in late larvae (Wilson 1928; Thorson 1946; Hannerz 1956; Hopkins 1958; Woodwick 1960; Blake 1969; Carrasco 1976; Radashevsky 1986, 1988, 1989; 1994, 2005; Plate and Husemann 1994; Sato-Okoshi 1994; Williams 2001; Radashevsky and Cárdenas 2004; Blake 2006; Radashevsky et al. 2006; Zhang et al. 2009; Gao et al. 2011; David et al. 2014; Barros et al. 2017; Blake 2017; Radashevsky and Migotto 2017; Ye et al. 2017).

Identification key to species of the larvae belonging to the genus *Polydora* in northeastern Japan

- | | | |
|---|--|---|
| 1 | Mid-dorsal single row of branching melanophores present..... | |
| | | <i>Polydora</i> cf. <i>glycymerica</i> |
| – | Mid-dorsal single row of branching melanophores absent..... | 2 |
| 2 | Vestibule and pharynx with black pigmentation..... | <i>Polydora brevipalpa</i> |
| – | Vestibule and pharynx not pigmented with black..... | 3 |
| 3 | Dorsal melanophores on each chaetiger faint..... | <i>Polydora</i> sp. 2 |
| – | Dorsal melanophores on each chaetiger distinct..... | 4 |

- 4 Black or brown pigmentation on lateral part of peristomium present..... 5
 – Black or brown pigmentation on lateral part of peristomium absent..... 8
 5 Distinct ventral pigment spot (yellow-green, brown, or black) present.....
 *Polydora cornuta*
 – Distinct ventral pigment spot absent..... 6
 6 Black pigmentation on lateral part of lateral peristomium present.....
 *Polydora sp. 3*
 – Brown pigmentation on lateral part of lateral peristomium present..... 7
 7 Two rows of dorsal melanophores from chaetigers III–VI or VII band-shaped,
 followed by large branching melanophores in posterior chaetiger.....
 *Polydora onagawaensis*
 – Two rows of dorsal melanophores on anterior chaetiger dot-like or short
 band-shaped, followed by dot-like not branching melanophores in posterior
 chaetiger *Polydora sp. 1*
 8 Two rows of dorsal melanophores mostly band-shaped with some of them
 slightly branching *Polydora cf. spongiicola*
 – Two rows of dorsal melanophores band-shaped in anterior chaetigers, fol-
 lowed by branching melanophores in posterior chaetiger 9
 9 Two rows of dorsal melanophores from chaetigers III–VII band-shaped, fol-
 lowed by pairs of large branching melanophores *Polydora hoplura*
 – Two rows of faint dorsal melanophores from chaetigers II–V or VII band-
 shaped, followed by pairs of branching melanophores in posterior chaetigers
 or whole of dorsal surface covered by finely ramified black pigmentation.....
 *Polydora sp. 2*

Polydora brevipalpa Zachs, 1933

Fig. 8A, B

Larval morphology. Overall shape slender and slightly fusiform. Prostomium broad and rounded anteriorly. Three pairs of black eyes present, innermost pair rounded, lateral pairs double-eyes, ramified melanophore between innermost and lateral two pairs of eyes usually present. Black pigment on lateral peristomium absent. Dorsal pigmentation consists of two rows of melanophores from chaetiger III. Dorsal melanophores undergo expansion and contraction, may expand to branching melanophores or ramified appearance or covered almost whole of dorsal surface by very finely ramified black pigments (Fig. 8B), or they contract to dot-like pigmentation patches (Fig. 8A). Lateral and ventral pigments absent. Vestibule and pharynx pigmented with black, gut pigmented with orange color. Modified chaetae develop on chaetiger V in late larvae. Gastrotrochs occur on chaetigers III, V, VII, X, XIII, XV, and XVII.

Remarks. Adults of this species are boring and were collected from shells of cultured *M. yessoensis* scallops suspended in Onagawa Bay in February 2011 and Mutsu Bay in October 2011. This species was identified as *P. brevipalpa* as adult morphology

agrees with the descriptions by Sato-Okoshi (1999) and Sato-Okoshi and Abe (2012). The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

Planktonic larvae of this species were collected from Onagawa Bay in April, May, and July 2011 and from Sasuhama in April 2011. The pair of large ramified or dot-like melanophores from chaetiger III distinguishes larvae of this species from those of other *Polydora* species. Blake (2017) reported similar dorsal pigment patterns in the larvae of *Polydora spongicola* Berkeley & Berkeley, 1950. However, the larvae of *P. spongicola* in Blake (2017) differ from those of *P. brevipalpa* in having dorsal melanophores from chaetiger II instead of chaetiger III, dark green colored intestine instead of orange, and non-pigmented pharynx instead of pigmented with black. Reproduction and life history of this species was reported in Sato-Okoshi et al. (1990) and Sato-Okoshi (1994) (both as *P. variegata*).

***Polydora cornuta* Bosc, 1802**

Fig. 8C, D

Larval morphology. Overall shape slender. Prostomium broad and rounded anteriorly. Three pairs of black eyes present, median pair rounded, most lateral pairs double-eyes, ramified melanophores between first median and the second lateral pair of eyes usually present. In late larval stage, anterior part of prostomium and lateral lips of peristomium pigmented yellow or brown. Small spots of black pigments occur on lateral parts of peristomium. Dorsal pigmentation consists of two rows of melanophores from chaetiger III with those of anterior four chaetigers band-shaped and then replaced by rounded or ramified melanophores from chaetiger VII onwards. Three rows of small faint dorsal spots of brown pigment present on posterior edge from chaetigers III or IV onwards in late larvae. Lateral pigment on chaetigers II, III, and often VI–XI extensive compared to that on other chaetigers. Large yellow or brown chromatophores occur ventrally from chaetigers V or VI onwards, usually three chromatophores arranged in transverse line except on gastrotroch-bearing chaetigers where single midventral chromatophores present. Black pigment spots occur on ventral side of body (Fig. 8D) and mid-dorsal part on pygidium (Fig. 8C). Gastrotrochs occur on chaetigers III, V, VII, X, XIII, XV and XVII.

Remarks. Adults of this species were non-boring and collected from mud deposits in crevices of shells of living *C. gigas* oysters in Sasuhama in June 2011 and from intertidal bottom sediment in Gamo Lagoon in August 2012. This species was identified as *P. cornuta* as adult morphology agrees with the description by Sato-Okoshi (2000) and Radashevsky (2005). The larvae and adults were confirmed to match (18S: 1770/1770, 16S: 468/470 bp) using molecular data (Fig. 2).

Rice et al. (2008) suggested that at least three sibling species may be involved in North America under the name of *P. cornuta* by differences of mitochondrial COI sequences between California, Florida, and Maine populations. Takata et al. (2011) reported that the *P. cornuta* from Fukuyama in the Seto Inland Sea, western Japan is

genetically close with the California/New Zealand lineage. It is unclear to which lineage the eastern Japan populations belong. The 18S rRNA gene sequence obtained in the present study showed a 1.9% (5/421 bp) difference with that of *P. cornuta* from Netherlands (KC686637).

Planktonic larvae of this species were collected from Gamo Lagoon in August 2012. The larval morphology of this species generally agrees with the descriptions of *P. cornuta* by Hannerz (1956, as *P. ligni*), Blake (1969, as *P. ligni*), Plate and Husemann (1994, as *P. ligni*), and Radashevsky (2005). Peristomial melanophores, which were reported by Hannerz (1956) and Blake (1969) but not by Radashevsky (2005), and middorsal vesiculate melanophores, which were reported by Radashevsky (2005) but not described by Hannerz (1956) and Blake (1969), were both present in specimens of the present study. Ventral pigmentation pattern was consistent with the description by Blake (1969) and Radashevsky (2005) instead of Hannerz's (1956) description. The larval dorsal pigmentation pattern, similar to that of *P. cornuta*, is typically found in many other *Polydora* species. This species can, however, be distinguished by the characteristic ventral yellow pigmentation pattern as the yellow pigment on the ventral side of the other *Polydora* species is diffusely scattered and does not appear regularly arranged when present (Radashevsky 2005).

***Polydora* cf. *glycymerica* Radashevsky, 1993**

Fig. 8E

Larval morphology. Overall shape elongated and slender. Prostomium small and rounded anteriorly. Three pairs of black eyes present, most lateral pairs double-eyes. Ramified melanophores between middle and lateral pair of eyes absent. Pigmentation on lateral peristomium absent. Two rows of ramified melanophores on chaetigers III–VI, and a median row of ramified melanophores from chaetiger VII onwards. Lateral and ventral pigments absent. A pair of black pigments occur on pygidium. Pygidium has a dorsal notch and lacks appendages. Gastrotrochs absent in 25-chaetiger larvae, probably already lost. Modified chaetae develop on chaetiger V.

Remarks. No adult individuals of this species were collected in the present study. The 18S and 16S rRNA gene sequences obtained from larvae in the present study did not match any of the available *Polydora* sequences. However, as the larvae formed a robustly supported clade with other *Polydora* species (Figs 2, 3), this species was referred to as the genus *Polydora*. Furthermore, the larval morphology including the characteristic dorsal pigment pattern of this larvae matches that of the larvae of *P. glycymerica* described by Radashevsky (1989). Therefore, this larva was tentatively identified as *P. cf. glycymerica*. However, there were slight differences between the present specimens and Radashevsky (1989) description: the two rows of ramified melanophores continued until chaetiger VI in the present description, whereas it continues to chaetigers VII–X according to Radashevsky (1989); ramified melanophores between the middle and lateral pair of eyes are present in the former description while absent in the latter; larvae of



Figure 8. Light micrographs showing the morphologies of living spionid larvae of the genus *Polydora* **A, B** *Polydora brevipalpa*, dorsal view of 15-chaetiger (**A**) and 17-chaetiger larvae (**B**) **C, D** *Polydora cornuta*, dorsolateral view of 11-chaetiger larva (**C**) and ventral view of 17-chaetiger larva (**D**) **E** *Polydora* cf. *glycymerica*, dorsal view of 25-chaetiger larva **F** *Polydora hoplura*, dorsal view of 15-chaetiger larva **G–I** *Polydora onagawaensis*, dorsal view of 10-chaetiger (**G**) and 18-chaetiger larvae (**H**), and lateral view of 16-chaetiger larva (**I**) **J** *Polydora* cf. *spongicola*, dorsal view of 17-chaetiger larva **K, L** *Polydora* sp. 1, dorsal view of 7-chaetiger (**K**) and 16-chaetiger larvae (**L**) **M** *Polydora* sp. 2, dorsal view of 23-chaetiger larva **N–P** *Polydora* sp. 3, dorsal view of 13-chaetiger (**N**) and dorsal (**O**) and lateral view (**P**) of 18-chaetiger larvae. Scale bars: 300 μ m.

P. cf. glycymerica collected in the present study were 25-chaetigers with > 2.0 mm long (Fig. 8E), whereas the largest larva observed by Radashevsky (1989) was a 20-chaetiger specimen 1.8 mm long. Further studies should test whether these differences are attributable to individual or developmental variabilities or interspecific differences.

The dorsal median single row of ramified melanophores is distinct in the larvae of the genus *Polydora*. The larvae of *Polydora hermaphroditica* also have a dorsal median row of ramified melanophores such as that of the larvae of *P. cf. glycymerica* and *P. glycymerica* (Hannerz 1956; Plate and Husemann 1994). However, the first species differs from the other two by the absence of two rows of ramified melanophores on anterior chaetigers.

Only one individual of planktonic larva of *P. cf. glycymerica* was collected in Onagawa Bay in October 2011. *Polydora glycymerica* was previously recorded as a shell-borer of *Macridiscus aequilatera* (G. B. Sowerby I, 1825) from Oarai, Japan (Sato-Okoshi 1999).

***Polydora hoplura* Claparède, 1868**

Fig. 8F

Larval morphology. Overall body shape slender or somewhat fusiform. Prostomium broad and rounded anteriorly. Three pairs of black eyes present, most lateral pairs double-eyes. Ramified melanophores between first and second innermost pair of eyes absent. Black pigmentation patches on lateral peristomium absent. Dorsal pigmentation consists of two rows of melanophores from chaetiger III with those of first five pairs band-shaped and then replaced by ramified melanophores in posterior chaetigers. Lateral pigments found on late larvae on chaetigers II–IV. Dorsolateral pigments at base of the parapodia start from chaetiger VII. A pair of black pigment occur on pygidium. Ventral pigment absent. Modified chaetae develop in chaetiger V in late larvae. Gastrotrochs occur on chaetigers III, V, VII, X, XIII, and XV.

Remarks. This species is a shell-borer, and adult specimens were collected from the turban snail *Omphalius rusticus* (Gmelin, 1791) in Gobu-ura and Onagawa Bay. This species was identified as *P. hoplura* as its adult morphology agrees with descriptions by Sato-Okoshi and Abe (2012, as *P. uncinata*) and Sato-Okoshi et al. (2017). The larvae and adults were confirmed to match (18S: 1769/1769, 16S: 464/475 bp) using molecular data (Fig. 2).

Only late larvae were found in July in Onagawa Bay. The larval morphology of this species agrees with descriptions by Wilson (1928) and Radashevsky and Migotto (2017). This species has adelphophagic and lecithotrophic larval development, in which larvae feed on nurse eggs in brood capsules, hatch at a very late stage, and have only a short pelagic life (Wilson 1928; Read 1975; Sato-Okoshi et al. 2008, as *P. uncinata*; Radashevsky and Migotto 2017). The poecilogenous development of this species with planktotrophic and adelphophagic planktonic larvae was reported by David et al. (2014), David and Simon (2014), and Simon (2015).

***Polydora onagawaensis* Teramoto, Sato-Okoshi, Abe, Nishitani & Endo, 2013**

Fig. 8G–I

Larval morphology. Overall body shape slender. Prostomium slightly broad and rounded anteriorly. Three pairs of black eyes present; median pair of eyes rounded, most lateral pairs double-eyes. Ramified melanophore between middle and lateral pair of eyes usually present (Fig. 8G, H). Weak brown pigmentation located on lateral parts of peristomium, behind prototroch, occasionally much paler or absent. Dorsal pigmentation consists of two rows of melanophores from chaetiger III with those of first IV–VI band-shaped and subsequently replaced by ramified melanophores. These melanophores undergo expansion and contraction. Lateral pigment found on chaetigers II and III in late larvae (Fig. 3G). Dorsolateral pigment at base of most parapodia, often appears to coalesce with dorsal pigment bands on anterior part of body (Fig. 8H). Pygidium has a dorsal notch and lacks appendages; a pair of black pigment patches occur on pygidium. Ventral brown pigment may be present on posterior part of late larvae

(Fig. 8I). Telotroch well developed. Gastrotrochs on chaetigers III, V, VII, X, XIII, and XV; those of chaetigers III and V lost in late larvae (Fig. 8I). In late larvae, modified chaetae develop in chaetiger V.

Remarks. This species is a shell-borer, and adult individuals were collected from shells of the wild turban snail *O. rusticus*, cultured scallop *M. yessoensis*, and wild and cultured *C. gigas* oysters in Onagawa Bay and Sasuhama, northeastern Japan. This species was identified as *P. onagawaensis* as adult morphology agrees with the description by Teramoto et al. (2013). The larvae and adults were confirmed to match (18S: 1771/1771, 16S: 472/473 bp) using molecular data (Fig. 2).

Planktonic larvae of this species were abundant from November to June in Onagawa Bay during the study period. The larval morphology of this species is similar to that of *Polydora* sp. 3 (see below). However, the former species has weak brown pigmentation on the lateral parts of the peristomium, whereas the latter species has large patches of black pigment on this region.

Polydora cf. *spongicola* Berkley & Berkeley, 1950

Fig. 8J

Larval morphology. Overall body shape slender and slightly fusiform. Prostomium broad and rounded anteriorly. Three pairs of black eyes present; median eyes rounded, most lateral pairs double-eyes. Ramified melanophores between middle and lateral pair of eyes absent. Black pigment on lateral peristomium absent. Dorsal pigmentation consists of two rows of band-shaped melanophores from chaetiger II. These melanophores undergo expansion and contraction, expand to ramified melanophores or contract to non-ramified band-shaped melanophores. Lateral and ventral pigments absent. In late larvae modified chaetae develop in chaetiger V. Gastrotrochs on chaetigers III, V, VII, X, XIII, and XV.

Remarks. Adults of this species were collected from mud tubes constructed on the sponge *Mycale* sp. in Morois Bay, Misaki Peninsula (Table 1). The morphology of its modified spines in chaetiger V and the sponge-associated ecology of adults match the description of *P. spongicola* by Radashevsky (1993). However, this species was referred to *P. cf. spongicola* because the adult specimens were in poor condition, which hindered their morphology examination. The larvae and adults were confirmed to match (18S: 1770/1771, 16S: 474/475 bp) using molecular data (Fig. 2).

Only one planktonic larva of this species was collected in Sasuhama in January 2013. The larval morphology of *P. cf. spongicola* closely resembles that of *P. spongicola* described by Radashevsky (1988, as *Polydora uschakovi* Buzhinskaja, 1971) from Russia. *Polydora uschakovi* originally described from Russia was synonymized with *P. spongicola* (type locality: Canada) by Radashevsky (1993). Later, Blake (2017) described the larvae of *P. spongicola* from California and doubted this synonymization because, despite the similarities between the larvae from Russian and California, there are several morphological differences including the nature of the major spines of chaetiger V and the distribution of nototrochs and gastrotrichs. However, the larval dorsal pigment

pattern of *P. spongicola* described by Blake (2017) greatly differs from those of *P. cf. spongicola* in the present study and of *P. spongicola* in Radashevsky (1988) but resembles that of *P. brevipalpa* in the present study. Conspecificity between *P. uschakovi* and *P. spongicola* should be verified in future studies.

***Polydora* sp. 1**

Fig. 8K, L

Larval morphology. Overall body shape slender. Prostomium broad and rounded anteriorly. Three pairs of black eyes present; median eyes rounded and lateral pairs double-eyes. Ramified melanophore between innermost and next to innermost pairs of eyes absent. Weak brown pigmentation on lateral parts of peristomium present or absent. Dorsal pigmentation consists of two rows of melanophores from chaetiger III, those of first five pairs band-shaped and remaining pairs dot-like in late larvae (Fig. 8L). These melanophores all dot-like in early larvae (Fig. 8K). Lateral pigment found on chaetigers II, IX, X, and XI in late larvae. Dorsolateral pigment at base of parapodia on posterior chaetigers. A pair of black and brown pigment patches occur on pygidium. Ventral brown pigment present on posterior part of late larvae. Pygidium has a dorsal notch and lacks appendages. Telotroch well developed. In late larvae, modified chaetae develop in 5th chaetiger.

Remarks. Adults of this species are shell-borer and were collected from the shell of the turban snail *O. rusticus* in Sasuhama. The adults of this species have characteristic conspicuous black bars in their palps and are morphologically similar to *Polydora neocaeca* Williams & Radashevsky, 1999. *Polydora haswelli* previously recorded in Japan (Sato-Okoshi and Abe 2013) was reexamined as *P. neocaeca* by comparing morphology and molecular sequences with the specimens from near the type locality (Malan et al. 2020). As the 18S and 16S rRNA gene sequences of *Polydora* sp. 1 and *P. neocaeca* showed differences (18S: 8/1771, 16S: 40/476 bp), the specimens collected in the present study were referred to a different species. Only two individuals of planktonic larvae of this species were collected in Onagawa Bay in April and July 2011. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

***Polydora* sp. 2**

Fig. 8M

Larval morphology. Overall body shape slender. Prostomium broad and rounded anteriorly. Three pairs of black eyes present, most lateral pairs double-eyes. Ramified melanophores between first and second innermost pairs of eyes absent. Pigmentation on lateral peristomium weak brown or absent. Dorsal pigmentation consists of two rows of melanophores from chaetiger II, with those of first 4–6 chaetigers being band-shaped and then replaced by ramified melanophores in posterior chaetigers. Dorsal pigments faint, undergo expansion and contraction, expand to cover almost whole of

dorsal surface as finely ramified black pigmentation (Fig. 8M) or contract to band-shaped or dot-like black pigments without ramification. Faint lateral pigment found on late larvae on chaetigers VII onwards. Ventral pigments absent. A pair of brown pigments occur on the pygidium. Pygidium has a dorsal notch and lacks appendages. Gastrotrochs on chaetigers III, V, VII, X, XIII, XV, XVII, and XIX. Modified chaetae develop in chaetiger V in late larvae.

Remarks. No benthic adult stages were collected in the present study. These larvae formed a robustly supported monophyletic clade with other *Polydora* species (Figs 2, 3). Nevertheless, this species was identified as a member of *Polydora*. As the 18S and 16S rRNA gene sequences obtained from the larvae did not match any other available *Polydora* sequences, this species was referred to *Polydora* sp. 2.

Only two individuals of planktonic larvae of this species were collected from Sasuhama and Gamo Lagoon in January 2013. The faint dorsal pigmentation of the larvae of this species is unique among the members of *Polydora* collected in the present study.

Polydora sp. 3

Fig. 8N–P

Larval morphology. Overall body shape slender. Prostomium broad and rounded anteriorly. Three pairs of black eyes present; median pair of eyes rounded, most lateral pairs double-eyes, ramified melanophore between innermost and next to innermost pairs of eyes present. Large patches of black pigment located on lateral part of peristomium, behind prototroch. Dorsal pigment pattern consists of two rows of melanophores from chaetiger III with those of first four or five chaetigers being band-shaped and then replaced by ramified branching melanophores (Fig. 8O). These melanophores undergo expansion and contraction. Lateral pigment found on chaetigers II–IV, resumes again from chaetiger VII in late larvae (Fig. 8P). A pair of black pigment patches occur on pygidium. Ventral brown and black pigment present on posterior part in late larvae ready to metamorphose. Pygidium has a dorsal notch and lacks appendages. Telotroch well developed. Gastrotrochs on chaetigers III, V, VII, IX, X, XIII, XV, and XVII, lost on chaetigers III and V in late larvae (Fig. 8P). In late larvae, modified chaetae develop in chaetiger V.

Remarks. No benthic adult stages were collected in the present study. The 18S rRNA gene sequences obtained from the larvae did not match any available *Polydora* sequences. As the larvae formed a robustly supported monophyletic clade with other *Polydora* species (Figs 2, 3), this species was referred to *Polydora* sp. 3.

Planktonic larvae of this species were collected from December to June in Onagawa Bay every year during the study period. Planktonic larvae of this species were previously reported to be abundant in Onagawa Bay in the winter season from December to March (Abe et al. 2014, as *Polydora* sp.). Large patches of black pigment on the lateral peristomium are the main characteristic of this species and differentiate it from the other species of the genus observed in the present study, even at early planktonic stages (Fig. 8N).

Genus *Pseudopolydora* Czerniavsky, 1881

Larval diagnosis. Overall body shape thick and fusiform. Prostomium broad and rounded or gently notched anteriorly. Three pairs of black eyes present, most lateral often double-eyes. Mid-dorsal melanophore on the first chaetiger present in many species, absent in some species. Dorsal pigmentation consists of one or two pairs of branching melanophores (except *P. rosebelae*: mid-dorsal single row of melanophores present). Lateral and ventral pigments present or absent. Nototrochs occur in all chaetigers except first two chaetigers. Gastrotrochs occur in irregular pattern. Modified chaetae in chaetiger V and ventral hooded hooks from chaetiger VIII onwards develop in late larvae (Hannerz 1956, as *Polydora*; Rullier 1963, as *Polydora*; Rasmussen 1973; Blake and Woodwick 1975; Srikrishnadhas and Ramamoorthi 1977; Wu and Chen 1980; Radashevsky 1983, 1985; Plate and Husemann 1994, as *Polydora*; Hsieh 1994; Blake 2006; Radashevsky and Migotto 2009; Kondoh et al. 2017).

Identification key to species of the larvae belonging to the genus *Pseudopolydora* in northeastern Japan

- 1 A pair of dorsal melanophores on each chaetigers..... **2**
- Two pairs of dorsal melanophores on each chaetigers..... **4**
- 2 A pair of dorsal melanophores lack ramification; three pairs of black eyes are arranging more or less a straight line ***Pseudopolydora paucibranchiata***
- A pair of dorsal melanophores greatly ramified; lateral and anterior pairs of eyes link each other and form dumbbell-shaped eyes **3**
- 3 Ramification of dorsal melanophores covering most of dorsal side; a conspicuous large black pigment on pygidium ***Pseudopolydora tsubaki***
- Ramification of dorsal melanophores not covering most of dorsal side; a conspicuous black pigment spot on pygidium ***Pseudopolydora* sp.**
- 4 A central pair of dorsal black pigment “tilted wheels” shaped in anterior chaetigers; a weak mid-dorsal pigment present from chaetiger VI ***Pseudopolydora* aff. *achaeta***
- A central pair of dorsal melanophore dot-like or ramified; mid-dorsal melanophores absent except the first chaetiger **5**
- 5 Distinct ramified mid-dorsal melanophore present on first chaetiger..... ***Pseudopolydora* cf. *reticulata***
- Mid-dorsal melanophore on first chaetiger absent or not distinct and not ramified ***Pseudopolydora* cf. *kempi***

***Pseudopolydora* aff. *achaeta* Radashevsky & Hsieh, 2000**

Fig. 9A, B

Larval morphology. Overall body shape fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium gently notched an-

teriorly. Three pairs of black eyes present in more or less a straight line, most lateral pairs double eyes. Mid-dorsal melanophore on first chaetiger present. Dorsal pigmentation consists of two pairs of lateral and central rows of melanophores. Lateral ones dot-like, beginning on chaetiger II. Central ones shaped like “tilted-wheels” (inverted v-shape) begin on chaetiger III. A central pair of dorsal pigment patches gradually become dot-like on posterior chaetiger. Weak mid-dorsal pigments occur from chaetiger VI. Two medial black pigmentation areas occasionally present ventrally, on approximately chaetiger VI and anterior margin of pygidium. Anterior and posterior margin of prostomium have considerable brown pigment. Black pigment spots occur on sides of prostomium and peristomium. Pygidium has a central black pigment spot. Gastrotrachs on chaetiger III, V, VII, and XII in 13-chaetiger larvae.

Remarks. Adult individuals of this species were collected from muddy bottom sediments at 22 m depth in Onagawa Bay in December 2010 and September and December 2011 by using a Smith-McIntyre or Ekman-Birge grab sampler. Adult morphology agrees with the descriptions of *P. achaeta* by Radashevsky and Hsieh (2000) and Abe et al. (2016). However, the 16S rRNA gene sequence obtained in the present study showed a 11.5% (35/304 bp) difference with that of *P. achaeta* from Taiwan (country of type locality), which indicate that these two are different species. Therefore, this species is referred to *P. aff. achaeta*. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

Planktonic larvae of this species with more than 3-chaetiger stages were abundant in Onagawa Bay during July to November (Abe et al. 2014). A dorsal pigmentation area shaped like “tilted wheels” is a unique characteristic of this species among the known *Pseudopolydora* larvae.

Pseudopolydora cf. kemp (Southern, 1921)

Fig. 9C

Larval morphology. Overall body shape fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium gently notched anteriorly. Three pairs of black eyes present in more or less a straight line, most lateral pairs double-eyes. Mid-dorsal melanophore on first chaetiger usually absent (Fig. 9C), small non-ramified melanophore present in some individuals. Dorsal pigment consists of four rows of lateral and central pairs of pigment spots. Lateral and central pigments usually begin from chaetigers II and III, respectively. There pigment spots undergo expansion and contraction. Ventral pigment begins on chaetiger III, consists of paired bars on posterior border of each chaetiger. Anterior and posterior margin of prostomium have considerable brown pigment. Black pigment spots occur on sides of peristomium. Pygidium has black central spot. Gastrotrachs on chaetigers V and VII in 13-chaetiger larvae.

Remarks. Adult individuals of this species were collected from muddy sediment in Gamo Lagoon in January, May, and December 2011, and April 2013. Adult morphology agrees with the description of *P. cf. kemp* by Abe et al. (2016). Therefore, these

individuals were referred to *P. cf. kempfi*. The 16S rRNA gene sequence obtained in the present study showed a 99.7% (305/306 bp) similarity with that of *P. kempfi japonica* Imajima & Hartman, 1964 from Russia (MG460897) reported by Radashevsky et al. (2020b), indicating these two are same species. It will need to be clarified whether *P. kempfi* (type locality India) and subspecies *P. kempfi japonica* (type locality Japan) are the same species. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

Planktonic larvae of this species larger than 12-chaetiger stages were collected from Gamo Lagoon in August 2012. The larval morphology of this species observed in the present study agrees with the descriptions of *P. kempfi* by Blake and Woodwick (1975) and of *P. cf. kempfi* by Kondoh et al. (2017). These species have adelphophagic and lecithotrophic larval development, in which larvae feed on nurse eggs in brood capsules, hatch at a very late stage, and have a short pelagic life (Blake and Woodwick 1975; Kondoh et al. 2017). Reproduction and larval development of these species under the name of *P. kempfi* and *P. kempfi japonica* were also described by Srikrishnadhas and Ramamoorthi (1977), Myohara (1979), and Radashevsky (1985). However, the larvae of species in these descriptions resemble those of *Pseudopolydora cf. reticulata* Radashevsky & Hsieh, 2000 described by Kondoh et al. (2017) and of the present study in having planktotrophic development without nurse eggs and distinct dorsal melanophores including a middorsal melanophore on the first chaetiger. The taxonomy of *P. kempfi* is unclear because its original description is quite brief, and the current location of type specimen is unknown (Radashevsky and Hsieh 2000). Therefore, studies resolving the taxonomy of *P. kempfi* are necessary.

***Pseudopolydora paucibranchiata* (Okuda, 1937)**

Fig. 9D

Larval morphology. Overall body shape fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium gently notched anteriorly. Three pairs of black eyes present in more or less a straight line, most lateral pair comma-shaped. A mid-dorsal ramified melanophore on chaetiger I. A pair of melanophores present dorso-laterally from chaetigers II onwards. Black pigment spots occur on lateral surface of chaetiger II, on sides of peristomium, and pygidium. Two small medial black pigment spots occasionally present ventrally on approximately chaetiger VI and anterior margin of pygidium. Gut has yellow-green color due to ingested food. Gastrotrochs on chaetiger V, VII, and XI in 13-chaetiger larvae.

Remarks. Adult individuals were collected from muddy bottom sediment in the intertidal zone of Mangoku-ura Inlet in July 2014. Adult morphology agrees with the description of *Pseudopolydora paucibranchiata* by Okuda (1937, as *Polydora*). Therefore, these individuals were referred to this species. The larvae and adults were confirmed matching (18S: 1784/1784, 16S: 454/455 bp) using molecular data (Fig. 2).

The planktonic larvae of this species were reported to be common in Onagawa Bay during June to November (Abe et al. 2014). The larval morphology of this species observed in the present study agrees with the descriptions by Blake and Woodwick (1975), Ward (1977), Myohara (1980), Wu and Chen (1980), Radashevsky (1983), and Blake (2006). The dorsal pigment pattern of this species consists of one pair of melanophores, which agrees with that of the larvae of *Pseudopolydora vexillosa* Radashevsky & Hsieh, 2000 photographed by Mok et al. (2009) and Chandramouli et al. (2011, 2013), currently synonymized to *P. paucibranchiata* (Junqueira et al. 2009). The dorsal pigment pattern of these larvae is also similar to that of the larvae of *Pseudopolydora antennata* (Claparède, 1869) described by Hannerz (1956, as *Polydora antennata*), but the latter species has a more thickened body shape compared to the former.

***Pseudopolydora* cf. *reticulata* Radashevsky & Hsieh, 2000**

Fig. 9E, F

Larval morphology. Overall larval shape fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium slightly notched anteriorly. Three pairs of black eyes present in more or less a straight line, most lateral pairs double-eyes. Large patches of black pigment on lateral peristomium present. Mid-dorsal melanophore on chaetiger I usually present. Dorsal pigments undergo expansion and contraction, expanding to cover almost complete dorsal surface with finely ramified black pigment (Fig. 9F) or contract to dot-like black pigmentation without ramifications (Fig. 9E). Ventral pigment usually absent, consisting of paired bars on the posterior border on anterior chaetigers occasionally present. Black pigment on pygidium. Gastrotrochs on chaetigers V, VII, and XII in 17- and 18-chaetiger larvae, late larvae lose gastrotrochs on chaetigers V and/or XXII.

Remarks. Adult individuals of this species were collected from muddy sediment in Gamo Lagoon in April 2013 and Sasuhama in July and September in 2011. Adult morphology agrees with the description of *P. cf. reticulata* by Abe et al. (2016). Therefore, these individuals were referred to this species. The 16S rRNA gene sequence obtained in the present study showed a 99.4% (304/306 bp) similarity with that of *P. bassarginensis* (Zachs, 1933) from Russia (MG460894) reported by Radashevsky et al. (2020b), indicating these two are one species. Although the Japanese population shows intermediate morphological characteristics between *P. reticulata* (type locality Taiwan) and *P. bassarginensis* (type locality Russia), Abe et al. (2016) tentatively identified the Japanese population as *P. cf. reticulata* because the original description of *P. bassarginensis* is very brief and the status of the species remains unclear. The results of the present study indicate that the Japanese population likely belongs to *P. bassarginensis*, but whether the morphologically similar *P. reticulata* and *P. bassarginensis* are considered molecularly as the same or different species will need to be clarified. Planktonic larvae of *P. cf. reticulata* larger than the 3-chaetiger stage were collected from Gamo

Lagoon, Sasuhama, and Onagawa Bay mainly from July to September. The larvae and adults were confirmed to match (18S: 1775/1775, 16S: 468/470 bp) using molecular data (Fig. 2).

Pseudopolydora cf. *reticulata* and *P.* cf. *kempi* are very similar sister species; specimens from Japan once misidentified as *P.* cf. *kempi* or *P. kempi japonica* were distinguished based on their morphology and 18S and 28S rRNA gene sequences by Abe et al. (2016). The larvae of these two species are also quite similar, but the mid-dorsal melanophore on chaetiger I is usually present in *Ps.* cf. *reticulata* and absent in *P.* cf. *kempi*; moreover, the dorsal pigmentation is more distinct in the former species than in the latter. The two species also differ in reproduction and larval development: *P.* cf. *kempi* has lecithotrophic development with a short planktonic phase, whereas *P.* cf. *reticulata* has planktotrophic development with a long planktonic phase (Kondoh et al. 2017).

Pseudopolydora tsubaki Simon, Sato-Okoshi & Abe, 2017

Fig. 9G, H

Larval morphology. Overall larval shape fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium gently notched anteriorly. Three pairs of black eyes present, comprising one pair of rounded median eyes, one pair of large lateral eyes, and one pair of large anterior eyes. Lateral and anterior pairs of eyes link with each other and form dumbbell-shapes almost divided into two equal parts by a deep constriction. Mid-dorsal ramified melanophore present on chaetiger I in early larvae. Mid-dorsal melanophore on chaetiger I occasionally absent or expanded to finely ramified melanophore in late larvae. A paired of melanophores occur dorso-laterally from chaetiger II onwards, usually finely ramified in late larvae (Fig. 9H). Ramified melanophores cover almost entire ventral surface on chaetigers III–VII in 11-chaetiger larvae. Black pigment spots on sides of peristomium absent. Conspicuous large black pigment on pygidium. Gastrotrochs on chaetigers V and VII in 11-chaetiger larvae.

Remarks. Adult individuals were collected from mud deposits in crevices of shells of living *C. gigas* oysters in Habu Port, Izu-Oshima Island, and Tomiura, Boso Peninsula in April 2016. Adult morphology agrees with the description of *Pseudopolydora tsubaki* by Simon et al. (2019a). Therefore, these individuals were identified as *P. tsubaki*. The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

A small number of planktonic larvae of this species were collected in Habu Port and Tomiura in May and June 2016. The larvae of *P. tsubaki* are similar to those of *Pseudopolydora pulchra* (Carazzi, 1893) in having ramified melanophores covering the ventral side; however, these cover only the central part of the body in the former species, whereas those of latter species cover the ventral surface almost entirely (Hannerz 1956, as *Polydora pulchra*; Rullier 1963, as *Polydora pulchra*). The dorsal pigment pattern is also different in these two species: two pairs of melanophores are distinct in *P. pulchra*, whereas the melanophore pair is ambiguous in *P. tsubaki*.

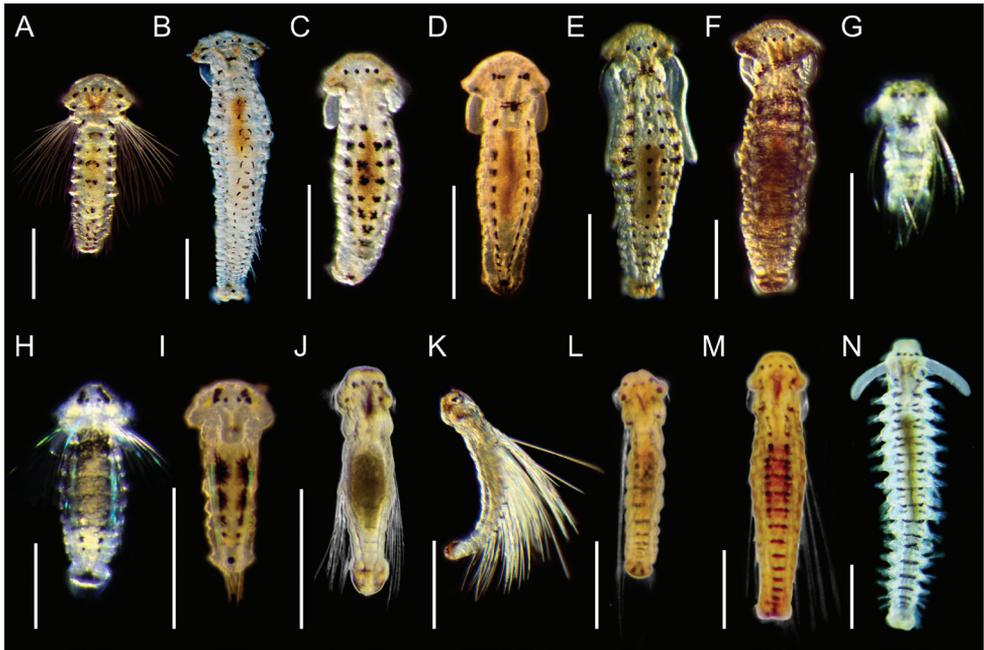


Figure 9. Light micrographs showing the morphologies of living spionid larvae of genera *Pseudopolydora* and *Spio* **A, B** *Pseudopolydora* aff. *achaeta*, dorsal view of 12-chaetiger (**A**) and 25-chaetiger larvae (**B**) **C** *Pseudopolydora* cf. *kempfi*, dorsolateral view of 12-chaetiger larva **D** *Pseudopolydora paucibranchiata*, dorsal view of 13-chaetiger larva **E, F** *Pseudopolydora* cf. *reticulata*, dorsal view of 17-chaetiger (**E**) and 16-chaetiger larvae (**F**) **G, H** *Pseudopolydora tsubaki*, dorsal view of 5-chaetiger (**G**) and 11-chaetiger larvae (**H**) **I** *Pseudopolydora* sp., dorsal view of 7-chaetiger larva **J, K** *Spio* sp. 1, dorsal view of 8-chaetiger (**J**) and lateral view of 12-chaetiger larvae (**K**) **L–N** *Spio* sp. 2, dorsal view of 10-chaetiger (**L**) and 17-chaetiger larvae (**M**), and 17-chaetiger metamorphosing larvae (**N**). Scale bars: 300 μ m.

Pseudopolydora sp.

Fig. 9I

Larval morphology. Overall larval shape slightly fusiform, head region enlarged due to broad prostomium and expanded lateral lips of vestibule. Prostomium rounded anteriorly. Three pairs of black eyes present, comprising one pair of rounded median eyes, one pair of large lateral eyes, and one pair of large anterior eyes. Lateral and anterior pairs of eyes link with each other and form a dumbbell-shape almost divided into two equal parts by a deep constriction. Small mid-dorsal melanophore present on chaetiger I. A distinct paired melanophore occurs dorso-laterally from chaetiger II onwards, ramified in anterior chaetigers. Black pigment spots on sides of peristomium absent. Dot-like black pigmentation on pygidium.

Remarks. No benthic adult stages were collected in the present study. The larvae formed a monophyletic clade with the other *Pseudopolydora* species with > 50% bootstrap support (Figs 2, 3). Therefore, this species was identified as a member of *Pseudopol-*

ydora. As the 18S rRNA gene sequences obtained from the larvae did not match any of the available *Pseudopolydora* sequences, this species is referred to *Pseudopolydora* sp.

Only one larva individual was collected from Sasuhama in August 2011. The dorsal pigment pattern of this larva somewhat resembles that of *P. paucibranchiata*; however, the mid-dorsal pigment of this species is weaker and its dorsolateral melanophores are more ramified than those of *P. paucibranchiata*. The eye arrangement of this larva resembles that of late *P. tsubaki* larvae: three pairs of black eyes are present, but not in a straight line.

Genus *Spio* Fabricius, 1785

Larval diagnosis. Overall body shape long, slender, and weakly or moderately fusiform. Prostomium small and rounded anteriorly. Lateral part of peristomium weakly demarcated from prostomium. Three pairs of black eyes present, most lateral often double-eyes. Dorsal pigmentation consists of transverse band-shaped or dot-like paired lateral melanophores. Some species lack black pigmentation. Ventral pigment usually absent. Dark-brown pigment may be present on pygidium. Nototrochs occur in all chaetigers except first one or two chaetigers, where nuchal organs develop. Gastrotrachs occur regularly in every other chaetiger from chaetiger III onwards. Larval chaetae on first chaetiger usually fairly long. Branchiae develop in late larvae, first on chaetiger II or III. One pair of anal cirri present on pygidium in late larvae (Thorson 1946, as spionid larva C, E, and F; Hannerz 1956; Wu et al. 1965; Simon 1963, 1967, 1968; Guérin 1972; Srikrishnadhas and Ramamoorthi 1981; Plate and Husemann 1994).

Identification key to species of the larvae belonging to the genus *Spio* in northeastern Japan

- 1 Two rows of black melanophore spots on each side of dorsum from chaetiger I onwards, linking by band-shaped medial black pigmentation from chaetiger IV or V ***Spio* sp. 2**
- Dorsal black melanophores not distinct; rows of faint transverse band-shaped black pigmentation present on dorsum from chaetiger IV onwards ***Spio* sp. 1**

***Spio* sp. 1**

Fig. 9J, K

Larval morphology. Overall larval shape slender and weakly fusiform. Larval chaetae on first chaetiger fairly long. Prostomium round anteriorly. Small patches of black pigment on peristomium ventrally. Three pairs of black eyes present, most lateral pairs double-eyes. Distinct black melanophore absent, rows of faint transverse band-shaped black pigmentation on dorsum from chaetiger IV onwards. Pharynx exhibits weak dark or brownish pigmentation. Gut yellow-green in color due to ingested food.

Remarks. Adult individuals of this species were collected from Rishiri Island, northern Japan, in July and August 2017. These specimens were previously identified as *S. arndti* Meißner, Bick & Bastrop, 2011 (Abe et al. 2019c) since adult morphology agreed. Although 18S rRNA gene sequence obtained in the present study 100% match with that of *S. arndti* (FR823434, 1761/1761 bp), because the 16S rRNA gene sequences were different (6.7%, 30/451 bp), the species reported here is referred to *Spio* sp. 1. The 16S rRNA gene sequence of *Spio* sp. 1 was rather more similar (96.1%, 298/310 bp) to that of *Spio* sp. 2573 from Russia (KT200126), but conspecificity of these two is unclear. A few planktonic larvae of this species were collected from Onagawa Bay only in May 2011. The larvae and adults were confirmed to match (18S: 1762/1762, 16S: 466/467 bp) using molecular data (Fig. 2).

The absence of distinct black melanophores in larvae of this species differentiates them from those of *Spio* sp. 2 (see below). Slight dorsal pigmentation was also reported in adelphophagic benthic larvae of *Spio setosa* Verrill, 1873 sensu Simon (1967, 1968), which were essentially unpigmented, and in those of *Spio multioculata* (Rioja, 1918) described by Hannerz (1956). However, the larval morphologies of these two species are different from that of *Spio* sp. 1 in lacking ventral black pigment on the peristomium (in both former species) and long larval chaetae on the first chaetiger (in *S. setosa*), and in having a relatively thickened body shape (in both species).

Spio sp. 2

Fig. 9L–N

Larval morphology. Overall larval shape elongated, slender, weakly fusiform. Larval chaetae on first chaetiger fairly long. Prostomium round anteriorly. Small patches of black pigment on lateral peristomium present ventrally. Three pairs of black eyes present, most lateral pairs double-eyes. Two rows of dot-like black melanophores on each side of dorsum from chaetiger I onwards, linking by band-shaped medial black pigmentation from chaetiger IV or V. Pharynx exhibits weak dark or brownish pigmentation. The larvae which are ready to metamorphose have branchiae from chaetiger II, pigment spot on palps, and a pygidium with four leaf-shaped anal cirri.

Remarks. Adult individuals were collected from muddy sand sediments of shallow water in Sasuhama in September 2011. These adults were morphologically identified as a *Spio* species, but they could not be identified to species level. *Spio* spp. 1 and 2 are distinguishable morphologically by the number of ventral epidermal glands. The 18S and 16S rRNA gene sequences obtained in the present study did not match any of the available *Spio* sequences (Figs 2, 3). The larvae and adults were confirmed to 100% match using molecular data (Fig. 2).

Planktonic larvae of this species were found in Sasuhama and Onagawa Bay from April to August during the study period. Larval morphology and pigmentation pattern of this species is similar to that of *Spio decorata* Bobretzky, 1870 described by Guérin 1972. However, the latter species was originally described from the Black Sea and has not been recorded in Japan.

Discussion

Larval identification based on the molecular data

The present study identified 41 species from 14 genera of planktonic spionid larvae by comparing adult and larval gene sequences and revealed high diversity of spionid larvae in neritic plankton communities (Table 1, Figs 2, 3). Planktonic spionid larvae of several species could not be identified to species level because of the lack of adult reference sequences or difficulties in adult identification. As the genetic information available for many marine invertebrate taxa including polychaetes is insufficient, the increase in gene sequence data based on accurate species identification and the establishment of a comprehensive database of adult reference sequences are essential for a more precise and efficient larval molecular identification. However, most of the larvae from the present study that did not have sequences that matched those of adults were identified to genus level based on their position within the phylogenetic tree; this was only possible because many of the spionid genera were recovered well or moderately supported monophyletic groups in our molecular phylogenetic analyses (Figs 2, 3). In contrast, the monophyly of some spionid taxa, particularly of the genera *Dipolydora*, *Malacoceros*, and *Prionospio* were ambiguous and not well supported in the phylogenetic tree recovered herein. It should be noted that *Malacoceros indicus* and *Malacoceros* cf. *indicus* were recovered as quite distant from *Malacoceros fuliginosus* and *Malacoceros* sp. (Fig. 3), potentially indicating the paraphyletic origins of these two clades. The results of the phylogenetic analyses also showed that the monophyly of subfamily Neriniinae is doubtful and more likely to be paraphyletic. Because intergeneric phylogenetic relationships were ambiguous due to the low statistical support of most of the higher internal nodes (Figs 2, 3), it was difficult to compare with the previous results of phylogenetic relationships among spionid genera provided by Sigvaldadóttir et al. (1997) and Blake and Arnofsky (1999). The results of our phylogenetic analyses reinforce the need for a more robust and comprehensive molecular phylogenetic study of this taxon to test the monophyly of each genus and subfamily and to shed light on the phylogenetic relationships among spionid genera.

In the present study, many spionid species were collected as planktonic larval stages. This emphasizes the effectiveness of field investigations of both larval and adult stages to assess the cryptic species diversity in benthic invertebrate fauna of coastal waters. The reference gene sequences used in the present study for adults covered most of the species belonging to the genera *Polydora* and *Pseudopolydora* hitherto recorded from Japan (Sato-Okoshi 1999, 2000; Sato-Okoshi and Abe 2012, 2013; Teramoto et al. 2013; Abe et al. 2016; Simon et al. 2019a). However, the sequences of some *Polydora* and *Pseudopolydora* larvae, namely *Polydora* sp. 2, *Polydora* sp. 3, and *Pseudopolydora* sp., did not match any adult reference sequences. This emphasizes the need for detailed taxonomic studies with a more comprehensive sampling of spionid adults to reveal the actual biological diversity of this taxon in Japan.

Morphology of spionid larvae

The family Spionidae can be divided into two subfamilies: 1) Spioninae Söderström, 1920, which includes the genera *Spio*, *Microspio* Mesnil, 1896, *Pygospio* Claparède, 1863, and genera of the tribe Polydorini; and 2) Nerininae Söderström, 1920, which includes almost all remaining spionid genera, except for *Atherospio* Mackie & Duff, 1986 and *Pygospioopsis* Blake, 1983 (Blake 2006), besides *Poecilochaetus* and *Trochochaeta*, which were recently placed within the family Spionidae (Radashevsky et al. 2018). The larvae of these two subfamilies were distinguished in the present study based on color and number of eyes, body pigmentation, shape of peristomium, and distribution of gastrotrachs, and by the following characteristics identified by Hannerz (1956): larvae of Spioninae have three pairs of black eyes (lateral eyes are often double eyes), distinct black pigmentation with melanophores, lateral parts of the peristomium not demarcated from prostomium, and gastrotrachs present from chaetiger III, V, or VII onwards, but absent in all of the succeeding chaetigers (Figs 7–9); larvae of Nerininae have two pairs of red or dark red eyes, lack distinct black pigmentation, lateral parts of the peristomium are well developed and often demarcated from prostomium, and gastrotrachs present from chaetiger II or III onwards and in all succeeding chaetigers (Figs 4, 5). Blake (1969) also discussed the presence of ventral ciliary patches in early larval stages as a common characteristic of subfamily Spioninae, but these cilia were not herein observed because they are lost in early larval stages.

Hannerz (1956) reported the following exceptions to the abovementioned typical larval morphologies: larvae of *Prionospio fallax* Söderström, 1920 (as *P. malmgreni*, see Blake and Arnofsky 1999) with two pairs of black eyes; larvae of *Malacoceros* (as *Scolecopsis*), which belongs to Nerininae, with intermediate characteristics between the two subfamilies, i.e., with three pairs of black eyes and gastrotrachs regularly distributed on every other chaetiger as in Spioninae larvae. However, Plate and Husemann (1994) reported that the larvae of *Malacoceros fuliginosus* (Claparède, 1868) have up to three pairs of red eyes in early stages and that eye color changes to black as larvae develop. Radashevsky and Migotto (2006) reported that the larvae of *Malacoceros* sp. have two pairs of red eyes. In the present study, the larvae of *Rhynchospio* have two pairs of dark red eyes (Fig. 4F, G), whereas their morphology resembled those of *Malacoceros* species described by Hannerz (1956). Radashevsky (2007) also reported that larvae of *Rhynchospio nhatrangi* have two pairs of red eyes. Besides the various reports on the number and color of eyes, the close relationships of *Malacoceros* and *Rhynchospio* to the subfamily Spioninae were indicated by larval morphology, and results of the phylogenetic analyses presented (Figs 2, 3) also provide some support for this hypothesis.

In the subfamily Spioninae, the most obvious larval differences between genera and species are the overall body shape and type and arrangement of pigmentation (Blake and Arnofsky 1999). The overall body shape of larvae of *Polydora*, *Dipolydora*, and *Spio* tended to be long and slender, whereas those of *Boccardiella*, *Boccardia*, and *Pseudopolydora* tended to be thick and fusiform (Figs 7–9), although

Boccardia proboscidea (Fig. 7A, B) and *Boccardia* sp. 1 (Fig. 7D–F) showed relatively slender body shapes. The lateral enlargement of the prostomium in Spioninae is variable: large in *Boccardiella* and *Pseudopolydora*, moderate in *Polydora* and *Boccardia*, and small in *Dipolydora* and *Spio*. Fairly long larval chaetae on the first chaetiger are highly characteristic of *Spio* within Spioninae.

The dorsal black pigmentation with melanophores is distinct in the subfamily Spioninae, and the pattern of rows of melanophores is generally diagnostic among Spioninae genera. The typical patterns of dorsal pigmentation rows in larvae are as follows: a pair of transverse bands of black pigment on some anterior chaetigers followed by a pair of large branching melanophores in *Polydora*; lack of large melanophores, but with a pair of medial spots or bands, a pair of lateral pigment patches, and mid-dorsal black pigment spot continuing posteriorly from the anterior chaetigers in *Dipolydora*; mid-dorsal melanophores arranged in a single row in *Boccardia*; medial and lateral pairs of spots or bands with black pigmentation and a small patch of pigment at the base of the notopodia present on almost all chaetigers in *Boccardiella*; a mid-dorsal melanophore on the first chaetiger, and one or two pairs of melanophores on each chaetiger in *Pseudopolydora*; a pair of black pigment spots and transverse black pigment bands linking them on each chaetiger in *Spio* (Figs 7–9). These typical dorsal pigment patterns were also reported in many previous studies (Hannerz 1956; Blake 1969, 2006; Blake and Arnofsky 1999; and references cited therein). However, unusual larval pigment patterns are often found in members of each of the aforementioned genera; therefore, these typical larval pigment patterns are not wholly consistent within each genus. For example, the single row of dorsal melanophores typical of *Boccardia* larvae was also observed in larvae of *Polydora* cf. *glycymerica* (Fig. 8E) and *Dipolydora* cf. *commensalis* (Fig. 7L), and have been reported in *Polydora glycymerica* (Radashevsky 1989), *Polydora hermaphroditica* (Hannerz 1956; Plate and Husemann 1994), *Dipolydora commensalis* (as *Polydora commensalis*: Andrews 1891; Hatfield 1965; Blake 1969; Radashevsky 1989), and *Pseudopolydora rosebelae* Radashevsky & Migotto, 2009. In contrast, the single row of dorsal melanophores is absent in *Boccardia chilensis* Blake & Woodwick, 1971 (Carrasco 1976; Blake and Kudenov 1981), *Boccardia pseudonatrix* (Fig. 7C), and *Boccardia semibranchiata* Guérin, 1990 (Guérin 1991). The larvae of *Pseudopolydora* cf. *kempi* lack a mid-dorsal melanophore on the first chaetiger, which is typical in *Pseudopolydora* larvae (Fig. 9C; Kondoh et al. 2017). The distinct dorsal black pigment is absent in *Spio setosa* Verrill, 1873 (Simon 1967, 1968) and *Spio* sp.1 (Fig. 9J, K).

Larvae of the following Spioninae genera were not collected in the present study: *Microspio*, *Pygospio*, and the polydorid genera *Amphipolydora* Blake, 1983, *Carazziella* Blake & Kudenov, 1978, *Polydorella* Augener, 1914, and *Tripolydora* Woodwick, 1964 (among them, *Microspio* and *Carazziella* have records from Japan by Okuda 1937, Sato-Okoshi 1998). Little is known about the larval morphology of the genera *Amphipolydora*, *Polydorella*, and *Tripolydora*. The larvae of *Microspio* resemble those of *Spio* in having a long and slender body shape and band-shaped dorsal black pigmentation (e.g., Hannerz 1956; Cazaux 1971). The larvae of *Carazziella* resemble those of the polydorid genus *Boccardia* in having a fusiform body shape and a single row of dorsal

melanophores (Carrasco 1976, as *Polydora citrona*; Blake and Arnofsky 1999; Blake 2006). The morphology of planktonic larval stages of *Pygospio elegans* as described in Hannerz (1956) resembles that of *Pseudopolydora* in having a thick and fusiform body shape, laterally enlarged prostomium, and mid-dorsal melanophore on the first chaetiger. Blake (1969) also noted the morphological similarity between the larvae of *Pseudopolydora* and *Pygospio elegans* and suggested the possibility that polydorids are closely related to *Pygospio* through *Pseudopolydora*. Subsequently, Blake and Woodwick (1975) reported the similarities of nurse egg feeding patterns between *Pseudopolydora kempfi* and *Pygospio elegans*, further strengthening the view of a close relationship between these two genera. This hypothesis is supported by the results of the phylogenetic analysis presented, showing that polydorids plus *Pygospio* form a monophyletic clade with robust statistical supports (Fig. 3).

In the subfamily Nerininae, as in Spioninae, the most obvious differences among genera are also regarding their overall body shapes. The lateral parts of the peristomium are conspicuous, well developed, and distinctly demarcated from the prostomium in larvae of *Laonice*, *Rhynchospio*, and *Scolecopsis*, but they are less pronounced in those of *Aonides*, *Paraprionospio*, *Prionospio*, and *Spiophanes*, as previously noted by Hannerz (1956). Larvae of the former group of genera (*Laonice*, *Rhynchospio*, *Scolecopsis*) also have a relatively wide body shape, whereas those of the latter group have a narrow body shape. Regarding the larvae of the former group, the prostomium is more or less stumpy and not pointed anteriorly in *Rhynchospio* and *Laonice*; however, *Scolecopsis* larvae have a unique body shape distinct from other spionid genera and their prostomium is pointed anteriorly, terminating in a tapered tip, and the lateral parts of the peristomium are demarcated and bearing a large peristomial umbrella. The larvae from the latter group (*Aonides*, *Paraprionospio*, *Prionospio*, *Spiophanes*), *Paraprionospio*, and *Prionospio* characteristically have extremely long and thin bodies with numerous chaetigers. In particular, larvae of *Paraprionospio* are extremely large in terms of body size and chaetiger number at metamorphosis among the spionid larvae (Yokoyama 1981). The larval morphology of the genus *Poecilochaetus* resembles that of *Paraprionospio* and *Prionospio*: larvae have extremely long and slender transparent bodies without distinct black pigmentation. However, the first differs from the other two in having small lateral pigment spots on each side of the chaetigers, a long metatrochophore stage (up to ca. 30–40 chaetiger stages), and a serpentine swimming behavior with developed parapodia bearing cirriform dorsal and ventral postchaetal lobes in the nectosoma stage. *Poecilochaetus* larvae are distinctive among spionid larvae in having gastrotrochs from chaetiger I onwards despite all other Nerininae larvae having gastrotrochs from chaetiger II or III onwards. In the present study, a pair of lateral processes on the prostomium developed in late larvae was found only in larvae of *Rhynchospio* and *Spiophanes*, it also previously described for *Malacoceros* larvae (Hannerz 1956). Fairly long and straight larval chaetae on the first chaetiger are highly characteristic of *Aonides* larvae, especially in the early stages (Fig. 4A, B); however, similar long and straight larval chaetae were also herein observed in larvae of *Spio* sp. 2 (Fig. 9L, M). Although larvae of the genera *Malacoceros*, *Marenzelleria* Mesnil, 1896, *Streblospio* Webster, 1879, and *Trochochaeta* (subfamily Nerininae) were not collected in the present study, larval morphologies of

these genera have been well described in previous studies. The larvae of *Malacoceros* resemble those of *Rhynchospio* (see above). The larvae of *Marenzelleria* (Bochert and Bick 1995) resemble those of *Laonice* in having the following characters: the remains of the egg membrane visible in early stages; in late larvae, lateral parts of the peristomium are conspicuous, well developed and distinctly demarcated from prostomium, and palps start developing laterally on the peristomium; the body is broader than the prototroch; both notopodial and neuropodial larval chaetae are present. However, the larvae of these two genera differ in the arrangement of nototrochs and gastrotrochs. The larval morphology of the genus *Streblospio*, which is included in the *Prionospio* complex (Dean 1965; Blake and Arnofsky 1999; Blake 2006) resembles that of *Prionospio*. The larval morphology of the genus *Trochochaeta* is distinctive among spionid larvae in having unusually long larval chaetae on the first chaetiger, very pronounced peristomial umbrella with two rows of robustly developed prototrochs, and total absence of nototrochs; larvae of this genus also present the typical morphological characteristics of Nerininae larvae, such as two pairs of red eyes, lack of distinct black pigmentation, and gastrotrochs from chaetiger II onwards on all succeeding chaetigers, although gastrotrochs of *Trochochaeta* larvae are weakly developed and those on chaetiger II are especially small and inconspicuous in late-stage larvae (Hannerz 1956, as *Disoma*; Blake and Arnofsky 1999; Blake 2006).

There is insufficient information on the larval morphology of the remaining genera of Nerininae. The larval development and morphology of *Dispio uncinata* Hartman, 1951 were described, and this species' close relationship with *Aonides* was suggested by Blake and Arnofsky (1999) and Blake (2006). However, Radashevsky et al. (2011) pointed out that the larvae described by these authors are most likely those of *Aonides californiensis* Rioja, 1947 rather than of a *Dispio* Hartman, 1951 species. Species-level identification of larvae from this subfamily is generally more difficult because of the lack of structured pigmentation, which is a useful characteristic for identifying species of Spioninae larvae. Especially in the genus *Prionospio*, larval morphology is quite simple and similar among species, which made it impossible to find morphological characters to distinguish between them in the present study.

Except for *Prionospio* spp., most of the planktonic spionid larvae collected in the present study have morphological characteristics that could be used to distinguish genera and species, and allowed morphological identification based on overall body shape and pigment patterns. The present paper provides identification keys to genera and species of planktonic spionid larvae from northeastern Japan; however, sufficient attention to developmental and/or intraspecific variation of larval morphological characteristics and the disappearance of pigments after fixation (only the black pigment usually remains after fixation) is required for accurate larval identification.

Acknowledgements

We would like to express our sincere appreciation to Yoshinari Endo and Goh Nishitani (Tohoku University) for valuable suggestions during the course of this study and

support with molecular analyses, respectively. We thank Captain Toyokazu Hiratsuka and the staff of Onagawa Field Science Center, Tohoku University and the members of Laboratory of Biological Oceanography, Tohoku University (Keijiro Hiwatashi, Katsuhisa Adachi, Genta Takeda, Yuta Yamakawa, Wataru Teramoto, Hiromasa Ohno, Akihiro Suzuki, Syuhei Uchiumi, Noritaka Ayakoji, Tomohiko Kondoh, Daiki Fujii, Jiro Endo, Chenchen Shen, and Yasushi Gomi) for their kind cooperation in sample collections in Onagawa Bay. We also thank Captain Hayashi and the crew of the 'Shinyo Maru' (Tokyo University of Marine Science and Technology) for valuable assistance in sample collections in Sagami Bay; Shinri Tomioka, Masahiko Sato (Rishiri Town Museum), Takao Suzuki (Michinoku Research Institute for Benthos), Gen Kanaya, Hajime Itoh (National Institute for Environmental Studies), Hisanori Kohtsuka (The University of Tokyo), Kenji Okoshi, Masanori Taru, Masahiro Suzuki, Motomi Imai (Toho University), Naoto Jimi (National Institute of Polar Research), Misuzu Aoki (Wetland International Japan), Tomoo Unagami (Center for Environmental Studies), Osamu Hoshino (Diving Service Chap), Genki Kobayashi (Kyoto University), and Masaatsu Tanaka (Keio University) for valuable assistance in the sample collections of adult spionids. We are grateful to Naoto Jimi, Genki Kobayashi, Shinri Tomioka, Jason D. Williams (Hofstra University), and Carol A. Simon (Stellenbosch University) for valuable comments on an early version of the manuscript and Vasily Radashevsky, Helena Wiklund, and Greg Rouse for helpful comments and suggestions in the peer review process. This study was partly supported by the research grant from Research Institute of Marine Invertebrates (No. 2011 IKU-1), Rishiri Research Project (2017), the Japanese Association for Marine Biology (JAMBIO) as a joint-research project (No. 27-56), JSPS KAKENHI (Grant Number: JP21580216, JP15K07540, JP18K05777, JP19K15899), and Environment Research and Technology Development Fund (Grant Number: JPMEERF20204R01) of the Environmental Restoration and Conservation Agency of Japan.

References

- Abe H, Kondoh T, Sato-Okoshi W (2016) First report of the morphology and rDNA sequences of two *Pseudopolydora* species (Annelida: Spionidae) from Japan. *Zoological Science* 33: 650–658. <https://doi.org/10.2108/zs160082>
- Abe H, Sato-Okoshi W, Endo Y (2011) Seasonal changes of planktonic polychaete larvae and chlorophyll *a* concentration in Onagawa Bay, northeastern Japan. *Italian Journal of Zoology* 78: 255–266. <https://doi.org/10.1080/11250003.2011.576037>
- Abe H, Sato-Okoshi W, Nishitani G, Endo Y (2014) Vertical distribution and migration of planktonic polychaete larvae in Onagawa Bay, north-eastern Japan. *Memoirs of Museum Victoria* 71: 1–9. <https://doi.org/10.24199/j.mmv.2014.71.01>
- Abe H, Tanaka M, Taru M, Abe S, Nishigaki A (2019a) Molecular evidence for the existence of five cryptic species within the Japanese species of *Marphysa* (Annelida: Eunicidae) known as “Iwa-mushi”. *Plankton & Benthos Research* 14: 303–314. <https://doi.org/10.3800/pbr.14.303>

- Abe H, Takeuchi T, Taru M, Sato-Okoshi W, Okoshi K (2019b) Habitat availability determines distribution patterns of spionid polychaetes (Annelida: Spionidae) around Tokyo Bay. *Marine Biodiversity Records* 12: 1–7. <https://doi.org/10.1186/s41200-019-0167-4>
- Abe H, Tomioka S, Kobayashi G, Itoh H (2019c) Spionidae (Annelida) from Rishiri Island, Northern Japan. *Rishiri Studies* 38: 15–27. [in Japanese with English abstract] <http://riishiri.sakura.ne.jp/Sites/RS/archive/382019/3803.pdf>
- Abe H, Sato-Okoshi W (2020) Novel symbiotic relationship between a spionid polychaete and *Lingula* (Brachiopoda: Lingulata: Lingulidae), with description of *Polydora lingulicola* sp. nov. (Annelida: Spionidae). *Zoosymposia* 19: 103–120. <https://doi.org/10.11646/zoosymposia.19.1.13>
- Aller RC (1982) The effects of macrobenthos on chemical properties of marine sediment and overlying water. In: McCall PL, Tevesz MJS (Eds) *Animal-Sediment Relations*. Plenum Press, New York, 53–102. https://doi.org/10.1007/978-1-4757-1317-6_2
- Andre C, Lindgarth M, Jonsson PR, Sundberg P (1999) Species identification of bivalve larvae using random amplified polymorphic DNA (RAPD): differentiation between *Cerastoderma edule* and *C. lamarcki*. *Journal of the Marine Biological Association of the United Kingdom* 79: 563–565. <https://doi.org/10.1017/S0025315498000691>
- Andrews EA (1891) A commensal annelid. *The American Naturalist* 25: 25–35. <https://doi.org/10.1086/275264>
- Anger K, Anger V, Hagmeier E (1986) Laboratory studies on larval growth of *Polydora ligni*, *Polydora ciliata*, and *Pygospio elegans* (Polychaeta, Spionidae). *Helgoländer Meeresuntersuchungen* 40: 377–395. <https://doi.org/10.1007/BF01983819>
- Anisimova M, Gil M, Dufayard JF, Dessimoz C, Gascuel O (2011) Survey of branch support methods demonstrates accuracy, power, and robustness of fast likelihood-based approximation schemes. *Systematic Biology* 60: 685–699. <https://doi.org/10.1093/sysbio/syr041>
- Backeljau T (2018) *Crassostrea gigas* or *Magallana gigas*: A community-based scientific response. *National Shellfisheries Association Quarterly Newsletter* 2018(1): 1–3. <https://www.shellfish.org/newsletter>
- Barros TL, Santos CSG, De Assis JE, de Souza JRB (2017) Morphology and larval development of *Polydora* cf. *websteri* (Polychaeta: Spionidae) in a tropical region of north-eastern Brazil. *Journal of Natural History* 51: 1169–1181. <https://doi.org/10.1080/00222933.2017.1316426>
- Bastrop R, Blank M (2006) Multiple invasions – a polychaete genus enters the Baltic Sea. *Biological Invasions* 8: 1195–1200. <https://doi.org/10.1007/s10530-005-6186-6>
- Berkeley E, Berkeley C (1936) Notes on Polychaeta from the coast of Western Canada. I. Spionidae. *Annals and Magazine of Natural History* 18: 468–477. <https://doi.org/10.1080/00222933608655215>
- Berkeley E, Berkeley C (1961) Notes on polychaeta from California to Peru. *Canadian Journal of Zoology* 39: 655–664. <https://doi.org/10.1139/z61-069>
- Bhaud M (1966) Les larves planctoniques d'Annélides Polychètes; leur intérêt pour une meilleure connaissance faunistique. *Vie et Milieu* 17: 435–437.
- Blake JA (1969) Reproduction and larval development of *Polydora* from northern New England. *Ophelia* 7: 1–63. <https://doi.org/10.1080/00785326.1969.10419288>

- Blake JA (1971) Revision of the genus *Polydora* from the east coast of North America (Polychaeta: Spionidae). *Smithsonian Contributions to Zoology* 75: 1–32. <https://doi.org/10.5479/si.00810282.75>
- Blake JA (1996) Family Spionidae Grube, 1850. In: Blake JA, Hilbig B, Scott PH (Eds) Taxonomic atlas of the benthic fauna of the Santa Maria Basin and Western Santa Barbara Channel (Vol. 6). Santa Barbara Museum of Natural History, Santa Barbara, 81–224.
- Blake JA (2006) Chapter 13. Spionida. In: Rouse G, Pleijel F (Eds) *Reproductive Biology and Phylogeny of Annelida*. Science Publishers, Enfield, 565–638. <https://doi.org/10.1201/9781482280159>
- Blake JA (2017) Larval development of Polychaeta from the northern California coast. Fourteen additional species together with seasonality of planktic larvae over a 5-year period. *Journal of the Marine Biological Association of the United Kingdom* 97: 1081–1133. <https://doi.org/10.1017/S0025315417000716>
- Blake JA, Arnofsky PL (1999) Reproduction and larval development of the spioniform Polychaeta with application to systematics and phylogeny. *Hydrobiologia* 402: 57–106. <https://doi.org/10.1023/A:1003784324125>
- Blake JA, Evans JW (1972) *Polydora* and related genera as borers in mollusk shells and other calcareous substrates (Polychaeta: Spionidae). *Veliger* 15: 235–249.
- Blake JA, Kudenov JD (1978) The Spionidae (Polychaeta) from southeastern Australia and adjacent areas with a revision of the genera. *Memoirs of the National Museum of Victoria* 39: 171–280. <https://doi.org/10.24199/j.mmv.1978.39.11>
- Blake JA, Kudenov JD (1981) Larval development, larval nutrition and growth for two *Boccardia* species (Polychaeta: Spionidae) from Victoria, Australia. *Marine Ecology Progress Series* 6: 175–182. <https://doi.org/10.3354/meps006175>
- Blake JA, Woodwick KH (1975) Reproduction and larval development of *Pseudopolydora paucibranchiata* (Okuda) and *Pseudopolydora kempfi* (Southern) (Polychaeta: Spionidae). *The Biological Bulletin* 149: 109–127. <https://doi.org/10.2307/1540483>
- Blank M, Bastrop R (2009) Phylogeny of the mud worm genus *Marenzelleria* (Polychaeta, Spionidae) inferred from mitochondrial DNA sequences. *Zoological Scripta* 38: 313–321. <https://doi.org/10.1111/j.1463-6409.2008.00370.x>
- Bleidorn C, Vogt L, Bartolomaeus T (2003) A contribution to sedentary polychaete phylogeny using 18S rRNA sequence data. *Journal of Zoological Systematics and Evolutionary Research* 41: 186–195. <https://doi.org/10.1046/j.1439-0469.2003.00212.x>
- Bleidorn C, Vogt L, Bartolomaeus T (2005) Molecular phylogeny of lugworms (Annelida, Arenicolidae) inferred from three genes. *Molecular Phylogenetics and Evolution* 34: 673–679. <https://doi.org/10.1016/j.ympev.2004.11.019>
- Bochert R, Bick A (1995) Reproduction and larval development of *Marenzelleria viridis* (Polychaeta: Spionidae). *Marine Biology* 123: 763–773. <https://doi.org/10.1007/BF00349119>
- Boeggemann M (2009) Polychaetes (Annelida) of the abyssal SE Atlantic. *Organisms Diversity & Evolution* 9: 251–428.
- Bogantes VE, Halanych KM, Meißner K (2018) Diversity and phylogenetic relationships of North Atlantic *Laonice* Malmgren, 1867 (Spionidae, Annelida) including the description of a novel species. *Marine Biodiversity* 48: 737–749. <https://doi.org/10.1007/s12526-018-0859-8>

- Bonifácio P, Martínez-Arbizu P, Menot L (2020) Alpha and beta diversity patterns of polychaete assemblages across the nodule province of the eastern Clarion-Clipperton Fracture Zone (equatorial Pacific). *Biogeosciences* 17: 865–886. <https://doi.org/10.5194/bg-17-865-2020>
- Branscomb ES, Vedder K (1982) A description of the naupliar stages of the barnacles *Balanus glandula* Darwin, *Balanus cariosus* Pallas, and *Balanus crenatus* Bruguiere (Cirripedia, Thoracica). *Crustaceana* 42: 83–95. <https://doi.org/10.1163/156854082X00722>
- Brasier MJ, Wiklund H, Neal L, Jeffreys R, Linse K, Ruhl H, Glover AG (2016) DNA barcoding uncovers cryptic diversity in 50% of deep-sea Antarctic polychaetes. *Royal Society Open Science* 3: 160432. <https://doi.org/10.1098/rsos.160432>
- Carlton JT (1996) Pattern, process, and prediction in marine invasion ecology. *Biological Conservation* 78: 97–106. [https://doi.org/10.1016/0006-3207\(96\)00020-1](https://doi.org/10.1016/0006-3207(96)00020-1)
- Carlton JT, Geller JB (1993) Ecological roulette: the global transport of nonindigenous marine organisms. *Science* 261: 78–82. <https://doi.org/10.1126/science.261.5117.78>
- Carrasco FD (1976) Larvas de la familia Spionidae (Polychaeta) en el plancton de la Bahía de Concepción, Chile. *Gayana Zoología* 38: 1–63. [in Spanish]
- Cazaux C (1971) Développement larvaire de *Microspio mecznikowianus*. In: Crisp DJ (Ed.) Fourth European Marine Biology Symposium. Cambridge University Press, Cambridge, 247–257. [in French with English summary]
- Chandramouli KH, Mok FS, Wang H, Qian PY (2011) Phosphoproteome analysis during larval development and metamorphosis in the spionid polychaete *Pseudopolydora vexillosa*. *BMC Developmental Biology* 11: 1–31. <https://doi.org/10.1186/1471-213X-11-31>
- Chandramouli KH, Sun J, Mok FS, Liu L, Qiu JW, Ravasi T, Qian PY (2013) Transcriptome and quantitative proteome analysis reveals molecular processes associated with larval metamorphosis in the polychaete *Pseudopolydora vexillosa*. *Journal of Proteome Research* 12: 1344–1358. <https://doi.org/10.1021/pr3010088>
- Choe S (1960) On the life history of the polychaete worm *Diopatra neapolitana* Delle Chiaje. *Bulletin of the Japanese Society of Scientific Fisheries* 26: 430–437. [in Japanese with English abstract] <https://doi.org/10.2331/suisan.26.430>
- Cowart DA, Pinheiro M, Mouchel O, Maguer M, Grall J, Miné J, Arnaud-Haond S (2015) Metabarcoding is powerful yet still blind: a comparative analysis of morphological and molecular surveys of seagrass communities. *PLoS ONE* 10: e0117562. <https://doi.org/10.1371/journal.pone.0117562>
- David AA, Simon CA (2014) The effect of temperature on larval development of two non-indigenous poecilognous polychaetes (Annelida: Spionidae) with implications for life history theory, establishment and range expansion. *Journal of Experimental Marine Biology and Ecology* 461: 20–30. <https://doi.org/10.1016/j.jembe.2014.07.012>
- David AA, Matthee CA, Simon CA (2014) Poecilognony in *Polydora hoplura* (Polychaeta: Spionidae) from commercially important molluscs in South Africa. *Marine Biology* 161: 887–898. <https://doi.org/10.1007/s00227-013-2388-0>
- David AA, Matthee CA, Loveday BR, Simon CA (2016) Predicting the dispersal potential of an invasive polychaete pest along a complex coastal biome. *Integrative and Comparative Biology* 56: 600–610. <https://doi.org/10.1093/icb/icw011>

- Day JH (1967) A Monograph on the Polychaeta of Southern Africa. Trustees of the British Museum (Natural History), London, 878 pp. <https://www.biodiversitylibrary.org/page/8725653>
- Day RL, Blake JA (1979) Reproduction and larval development of *Polydora giardi* Mesnil (Polychaeta, Spionidae). The Biological Bulletin 156: 20–30. <https://doi.org/10.2307/1541000>
- Dean D (1965) On the reproduction and larval development of *Streblospio benedicti* Webster. The Biological Bulletin 128: 67–76. <https://doi.org/10.2307/1539390>
- Dean D, Blake JA (1966) Life-history of *Boccardia hamata* (Webster) on the east and west coasts of North America. The Biological Bulletin. 130: 316–330. <https://doi.org/10.2307/1539739>
- Dean D, Hatfield PA (1963) Pelagic larvae of *Nerinides agilis* (Verrill). The Biological Bulletin 124: 163–169. <https://doi.org/10.2307/1539492>
- Duchêne JC (1984) Reproductive biology of *Boccardia polybranchia* (Carazzi) in Kerguelen (Subantarctic Province). Polar Biology 2: 251–257. <https://doi.org/10.1007/BF00263632>
- Duchêne JC (1989) Adelphopagie et biologie larvaire chez *Boccardia polybranchia* (Carazzi) (Annélide Polychète Spionidae) en province subantarctique. Vie et Milieu 39(3/4): 143–152.
- Foster NM (1971) Spionidae (Polychaeta) of the Gulf of Mexico and the Caribbean Sea. Studies on the Fauna of Curaçao and other Caribbean Islands 36: 1–183. <http://www.repositorio.naturalis.nl/record/506046>
- Gallego R, Lavery S, Sewell MA (2014) The meroplankton community of the oceanic Ross Sea during late summer. Antarctic Science 26: 345–360. <https://doi.org/10.1017/S0954102013000795>
- Gao Y, Zhang T, Yang H-S, Zhang X-F (2011) Studies on the embryo and larval development of *Polydora latispinosa*. Marine Sciences 35: 1–5. [in Chinese with English abstract] http://en.cnki.com.cn/Article_en/CJFDTotal-HYKX201110000.htm
- Gibson GD (1997) Variable development in the spionid *Boccardia proboscidea* (Polychaeta) is linked to nurse egg production and larval trophic mode. Invertebrate Biology 116: 213–226. <https://doi.org/10.2307/3226898>
- Gibson GD, Smith HL (2004) From embryos to juveniles: morphogenesis in the spionid *Boccardia proboscidea* (Polychaeta). Invertebrate Biology 123: 136–145. <https://doi.org/10.1111/j.1744-7410.2004.tb00149.x>
- Guérin JP (1972) Rapports taxonomiques et développement larvaire de *Spio decoratus* Bobretsky, 1871 (Annélide Polychète). Cahiers de Biologie Marine 13: 321–339. [in French with English summary]
- Guérin JP (1991) Cultivation of spionids (Annelida, Polychaeta). 3. Description of the larval development of *Boccardia semibranchiata*. Annales de l'Institut océanographique 67: 145–154. [in French with English summary]
- Guggolz T, Meißner K, Schwentner M, Brandt A (2019) Diversity and distribution of *Laonice* species (Annelida: Spionidae) in the tropical North Atlantic and Puerto Rico Trench. Scientific Reports 9: e9260. <https://doi.org/10.1038/s41598-019-45807-7>
- Guggolz T, Meißner K, Schwentner M, Dahlgren TG, Wiklund H, Bonifácio P, Brandt A (2020) High diversity and pan-oceanic distribution of deep-sea polychaetes: *Prionospio* and *Aurospio* (Annelida: Spionidae) in the Atlantic and Pacific Ocean. Organisms Diversity & Evolution 20: 171–187. <https://doi.org/10.1007/s13127-020-00430-7>

- Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Systematic Biology* 59: 307–321. <https://doi.org/10.1093/sysbio/syq010>
- Handley SJ, Bergquist PR (1997) Spionid polychaete infestations of intertidal Pacific oysters *Crassostrea gigas* (Thunberg), Mahurangi Harbor, northern New Zealand. *Aquaculture* 153: 191–205. [https://doi.org/10.1016/S0044-8486\(97\)00032-X](https://doi.org/10.1016/S0044-8486(97)00032-X)
- Hannerz L (1956) Larval development of the polychaete families Spionidae Sars, Disomidae Mensil, and Poecilochaetidae n. fam. in the Gullmar Fjord (Sweden). *Zoologiska bidrag från Uppsala* 31: 1–204.
- Hansen BW (1999) Cohort growth of planktotrophic polychaete larvae—are they food limited? *Marine Ecology Progress Series* 178: 109–119. <https://doi.org/10.3354/meps178109>
- Hartman O (1941) Some contributions to the biology and life history of Spionidae from California. *Allan Hancock Pacific Expeditions* 7: 289–324. [3 pls.]
- Hatfield PA (1965) *Polydora commensalis* Andrews—Larval development and observations on adults. *The Biological Bulletin* 128: 356–368. <https://doi.org/10.2307/1539899>
- Heimeier D, Lavery S, Sewell MA (2010) Using DNA barcoding and phylogenetics to identify Antarctic invertebrate larvae: Lessons from a large scale study. *Marine Genomics* 3: 165–177. <https://doi.org/10.1016/j.margen.2010.09.004>
- Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS (2018) UFBoot2: improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution* 35: 518–522. <https://doi.org/10.1093/molbev/msx281>
- Hopkins SH (1958) The planktonic larvae of *Polydora websteri* Hartman (Annelida, Polychaeta) and their settling on oysters. *Bulletin of Marine Science* 8: 268–277. <https://www.ingentaconnect.com/content/umrsmas/bullmar/1958/00000008/00000003/art00004>
- Hosoi M, Hosoi-Tanabe S, Sawada H, Ueno M, Toyohara H, Hayashi I (2004) Sequence and polymerase chain reaction-restriction fragment length polymorphism analysis of the large subunit rRNA gene of bivalve: Simple and widely applicable technique for multiple species identification of bivalve larva. *Fisheries Science* 70: 629–637. <https://doi.org/10.1111/j.1444-2906.2004.00850.x>
- Hsieh H-L (1994) Larval development and substrate preference at settlement in *Pseudopolydora diopatra* (Polychaeta: Spionidae). *Invertebrate Reproduction and Development* 25: 205–214. <https://doi.org/10.1080/07924259.1994.9672387>
- Imai T (1975) Iwamushi *Marphysa sanguinea* (MONTAGU) no Sanran to Syokikeitai nitsuite. [Spawning and early morphology of *Marphysa sanguinea* (Montagu)]. *Aquaculture Science* 23: 14–20. [in Japanese]
- Imai T (1982) The early development and breeding of *Marphysa sanguinea* (Montagu). *Benthos Research* 23: 36–41. [in Japanese] <https://doi.org/10.5179/benthos1970.1982.36>
- Imajima M (1959) A description of a new species of the Spionidae (Polychaeta), *Nerinides yamaguchii* n. sp., with notes on its development. *Journal of Hokkaido Gakugei University* 10: 155–165.
- Imajima M (1989) Spionidae (Annelida, Polychaeta) from Japan I. The genera *Aonides* and *Apoprionospio*. *Bulletin of the National Science Museum, Tokyo, Series A. Zoology* 15: 213–222.
- Imajima M (1990a) Spionidae (Annelida, Polychaeta) from Japan II. The genus *Prionospio* (*Aquilaspio*). *Bulletin of the National Science Museum, Tokyo, Series A. Zoology* 16: 1–13.

- Imajima M (1990b) Spionidae (Annelida, Polychaeta) from Japan IV. The genus *Prionospio* (*Prionospio*). Bulletin of the National Science Museum, Tokyo, Series A. Zoology 16: 105–140.
- Imajima M (1990c) Spionids (Annelida, Polychaeta) obtained by dredging from Oshima Strait and Yakiuchi Bay, Amami-Oshima of the Ryukyu Islands. Memoirs of the National Science Museum, Tokyo 23: 93–99.
- Imajima M (1991a) Spionidae (Annelida, Polychaeta) from Japan VI. The genera *Malacoceros* and *Rhynchospio*. Bulletin of the National Science Museum, Tokyo, Series A. Zoology 17: 5–17.
- Imajima M (1991b) Spionidae (Annelida, Polychaeta) from Japan VII. The genus *Spiophanes*. Bulletin of the National Science Museum, Tokyo, Series A. Zoology 17: 115–137.
- Imajima M (1992) Spionidae (Annelida, Polychaeta) from Japan. VIII. The genus *Scolelepis*. Bulletin of the National Science Museum, Tokyo, Series A. Zoology 18:1–34.
- Imajima M (2006) Polychaetous annelids from Sagami Bay and Sagami Sea, central Japan. Memoirs of the National Science Museum, Tokyo 40: 317–408.
- Imajima M (2009) Deep-sea benthic polychaetes off Pacific coast of the northern Honshu, Japan. National Museum of Nature and Science Monographs 39: 39–192.
- Imajima M (2011) Polychaetous annelids collected from Sagami Bay toward the Ogasawara Islands, Japan. Memoirs of the Museum of Nature and Science, Tokyo 47: 145–218.
- Imajima M, Hartman O (1964) The polychaetous annelids of Japan. Parts I, II. Allan Hancock Foundation Publications Occasional Paper 26: 1–452. [38 plates]
- Izuka A (1912) The errantiate Polychaeta of Japan. Journal of the College of Science, Imperial University, Tokyo 30: 1–262. [24 plates] <https://doi.org/10.15083/00037775>
- Junqueira AOR, Tavares MDS, Schaeffer-Novelli Y, Radashevsky VI, Cirelli JO, Julio LM, Romagnoli FC, Santos KC, Ferreira-Silva MAG (2009) Capítulo 6. Zoobentos. In: Lopes RM, Coradin L, Pombo VB, Cunha DR (Eds) Informe Sobre as Espécies Exóticas Invasoras Marinhas no Brasil. Ministério do Meio Ambiente, Brazil, 145–371.
- Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. Nature Methods 14: 587–589. <https://doi.org/10.1038/nmeth.4285>
- Kamel SJ, Oyarzun FX, Grosberg RK (2010) Reproductive biology, family conflict, and size of offspring in marine invertebrates. Integrative and Comparative Biology 50: 1–11. <https://doi.org/10.1093/icb/icq104>
- Kan K, Kuroki Y, Sato M, Tosuji H (2020) Larval recruitment process in the catadromous life history of *Hediste diadroma* (Nereididae, Annelida) in an estuary in Kagoshima Bay, Southern Japan. Plankton and Benthos Research 15: 30–43. <https://doi.org/10.3800/pbr.15.30>
- Katoh K, Rozewicki J, Yamada KD (2017) MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics 20(4): 1160–1166. <https://doi.org/10.1093/bib/bbx108>
- Kondoh T, Abe H, Sato-Okoshi W (2017) Reproduction and larval development of two sympatric *Pseudopolydora* species (Annelida: Spionidae) in Japan. Invertebrate Reproduction & Development 61: 172–181. <https://doi.org/10.1080/07924259.2017.1318095>
- Kotta J, Kotta I, Bick A, Bastrop R, Väinölä R (2015) Modelling habitat range and seasonality of a new, non-indigenous polychaete *Laonome* sp. (Sabellida, Sabellidae) in Pärnu Bay, the north-eastern Baltic Sea. Aquatic Invasions 10: 275–285. <https://doi.org/10.3391/ai.2015.10.3.03>

- Koya Y, Onchi R, Furuta Y, Yamauchi K (2003) Method for artificial fertilization and observation of the developmental process in Japanese palolo, *Tylorrhynchus heterochaetus* (Annelida: Polychaeta). Science Report of the Faculty of Education, Gifu University (Natural Science) 27: 85–94. [in Japanese with English abstract]
- Laffaille P, Petillon J, Parlier E, Valery L, Ysnel F, Radureau A, Feunteun E, Lefeuvre J-C (2005) Does the invasive plant *Elymus athericus* modify fish diet in tidal salt marshes? Estuarine, Coastal and Shelf Science 65: 739–746. <https://doi.org/10.1016/j.ecss.2005.07.023>
- Lee GH, Yoon SM, Min G-S (2020) DNA barcoding of *Boccardiella hamata* (Annelida: Polychaeta: Spionidae) in South Korea. Animal Systematics, Evolution and Diversity 36: 268–273. <https://doi.org/10.5635/ASED.2020.36.3.037>
- Levin LA (1982) The roles of life history, dispersal and interference competition in the population and community structure of a dense infaunal polychaete assemblage. PhD thesis, University of California, San Diego.
- Levin LA (1984) Multiple patterns of development in *Streblospio benedicti* Webster (Spionidae) from three coast of North America. The Biological Bulletin 166: 494–508. <https://doi.org/10.2307/1541157>
- Levin LA (1986) The influence of tides on larval availability in shallow waters overlying a mudflat. Bulletin of Marine Science 39: 224–233. <https://www.ingentaconnect.com/content/umrsmas/bullmar/1986/00000039/00000002/art00007#expand/collapse>
- Levin LA (1990) A review of methods for labelling and tracking marine invertebrate larvae. Ophelia 32: 115–144. <https://doi.org/10.1080/00785236.1990.10422028>
- Maciolek NJ (1985) A revision of the genus *Prionospio* Malmgren, with special emphasis on species from the Atlantic Ocean, and new records of species belonging to the genera *Apoprionospio* Foster and *Paraprionospio* Caullery (Polychaeta, Annelida, Spionidae). Zoological Journal of the Linnean Society, London 84: 325–383. <https://doi.org/10.1111/j.1096-3642.1985.tb01804.x>
- Magalhães WF, Bailey-Brock JH, Santos CSG (2015) A new species and two new records of *Poecilochaetus* (Polychaeta: Poecilochaetidae) from Hawaii. Journal of the Marine Biological Association of the United Kingdom 95: 91–100. <https://doi.org/10.1017/S002531541400109X>
- Malan A, Williams JD, Abe H, Sato-Okoshi W, Matthee CA, Simon CA (2020) Clarifying the cryptogenic species *Polydora neocaeca* (Annelida: Spionidae): a shell-boring invasive pest of molluscs from locations worldwide. Marine Biodiversity 50: 1–51. <https://doi.org/10.1007/s12526-020-01066-8>
- Martin D, Britayev TA (1998) Symbiotic polychaetes: review of known species. Oceanography and Marine Biology: An Annual Review 36: 217–340.
- Martin D, Britayev TA (2018) Symbiotic polychaetes revisited: an update of the known species and relationships (1998–2017). Oceanography and Marine Biology: An Annual Review 56: 371–448. <https://doi.org/10.1201/9780429454455-6>
- Martin D, Pinedo S, Sarda R (1996) Grazing by meroplanktonic polychaete larvae may help to control nanoplankton in the NW Mediterranean littoral: in situ experimental evidence. Marine Ecology Progress Series 143: 239–246. <https://doi.org/10.3354/meps143239>
- Maciolek NJ (2000) New species and records of *Aonidella*, *Laonice*, and *Spiophanes* (Polychaeta: Spionidae) from shelf and slope depths of the Western North Atlantic. Bulletin of

- Marine Science 67: 529–547. <https://www.ingentaconnect.com/content/umrsmas/bull-mar/2000/00000067/00000001/art00043>
- Meißner K, Blank M (2009) *Spiophanes norrisi* sp. nov. (Polychaeta: Spionidae) – a new species from the NE Pacific coast, separated from the *Spiophanes bombyx* complex based on both morphological and genetic studies. *Zootaxa* 2278: 1–25. <https://doi.org/10.11646/zootaxa.2278.1.1>
- Meißner K, Götting M (2015) Spionidae (Annelida: ‘Polychaeta’: Canalipalpata) from Lizard Island, Great Barrier Reef, Australia: the genera *Malacoceros*, *Scolecopsis*, *Spio*, *Microspio*, and *Spiophanes*. *Zootaxa* 4019: 378–413. <https://doi.org/10.11646/zootaxa.4019.1.15>
- Meißner K, Hutchings PA (2003) *Spiophanes* species (Polychaeta: Spionidae) from eastern Australia: with descriptions of new species, new records and an emended generic diagnosis. *Records of the Australian Museum* 55: 117–140. <https://doi.org/10.3853/j.0067-1975.55.2003.1379>
- Meißner K, Bick A, Guggolz T, Götting M (2014) Spionidae (Polychaeta: Canalipalpata: Spionida) from seamounts in the Northeast Atlantic. *Zootaxa* 3786: 201–245. <https://doi.org/10.11646/zootaxa.3786.3.1>
- Miralles L, Ardura A, Arias A, Borrell YJ, Clusa L, Dopico E, de Rojas AH, Lopez B, Munoz-Colmenero M, Roca A, Valiente AG, Zaiko A, Garcia-Vazquez E (2016) Barcodes of marine invertebrates from north Iberian ports: Native diversity and resistance to biological invasions. *Marine Pollution Bulletin* 112: 183–188. <https://doi.org/10.1016/j.marpolbul.2016.08.022>
- Mincks SL, Dyal PL, Paterson GLJ, Smith CR, Glover AG (2009) A new species of *Aurospio* (Polychaeta, Spionidae) from the Antarctic shelf, with analysis of its ecology, reproductive biology and evolutionary history. *Marine Ecology* 30: 181–197. <https://doi.org/10.1111/j.1439-0485.2008.00265.x>
- Minh BQ, Nguyen MA, von Haeseler A (2013) Ultrafast approximation for phylogenetic bootstrap. *Molecular Biology and Evolution* 30: 1188–1195. <https://doi.org/10.1093/molbev/mst024>
- Miura T, Kajihara T (1981) The development of a serpulid worm, *Hydroides ezoensis* (Annelida, Polychaeta). *Proceedings of the Japanese Society of Systematic Zoology* 20: 7–12. https://doi.org/10.19004/pjssz.20.0_7
- Mok FS, Thiyagarajan V, Qian PY (2009) Proteomic analysis during larval development and metamorphosis of the spionid polychaete *Pseudopolydora vexillosa*. *Proteome Science* 7: 1–44. <https://doi.org/10.1186/1477-5956-7-44>
- Motoda S (1957) North Pacific standard plankton net. *Inform Bulletin of Plankton Society of Japan* 4: 13–15. [in Japanese]
- Myohara M (1979) Reproduction and development of *Pseudopolydora kempji japonica* (Polychaeta: Spionidae), with special reference to the polar lobe formation. *Journal of the Faculty of Science, Hokkaido University. Series 6, Zoology* 21: 355–364. [figs 8, 9] <http://hdl.handle.net/2115/27642>
- Myohara M (1980) Reproduction and development of *Pseudopolydora paucibranchiata* (Polychaeta: Spionidae) under laboratory condition, with special regard to the polar lobe formation. *Journal of the Faculty of Science, Hokkaido University. Series 6, Zoology* 22: 145–155. [figs 1–3] <http://hdl.handle.net/2115/27657>

- Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ (2015) IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution* 32: 268–274. <https://doi.org/10.1093/molbev/msu300>
- Nishi E, Yamasu T (1992a) Brooding and development of *Rhodopsis pusilla* Bush (Serpulidae, Polychaeta). *Bulletin of the College of Science, University of the Ryukyus* 54: 93–100.
- Nishi E, Yamasu T (1992b) Brooding and development of a serpulid worm *Salmacina dysteri* (Huxley) (Sedentaria; Polychaeta). *Bulletin of the College of Science, University of the Ryukyus* 54: 107–121.
- Nishi E, Yamasu T (1992c) Brooding habit and larval development of a serpulid worm *Paraprotis dendrova* Uchida (Annelida, Polychaeta, Sedentaria). *Bulletin of the College of Science, University of the Ryukyus* 54: 83–92.
- Nishi E, Yamasu T (1992d) Observations on the reproductive behaviour and development of a common fouling spirorbid *Dexiospira foraminosa* Bush (Sedentaria, Polychaeta). *Bulletin of the College of Science, University of the Ryukyus* 54: 101–106.
- Okada YK (1930) A remark on the constitution of larval syllids. *Journal of the Marine Biological Association of the United Kingdom* 16: 479–487. <https://doi.org/10.1017/S002531540007288X>
- Okuda S (1937) Spioniform polychaetes from Japan. *Journal of the Faculty of Science Hokkaido Imperial University Series 6, Zoology* 5: 217–254. <http://hdl.handle.net/2115/27002>
- Okuda S (1946) Studies on the development of Annelida Polychaeta I. *Journal of the Faculty of Science, Hokkaido Imperial University. Series 6, Zoology* 9: 115–219. <http://hdl.handle.net/2115/27055>
- Omel'yanenko VA, Kulikova VA (2002) Composition, seasonal dynamics, and long-term fluctuations in the density of pelagic polychaetes in Amurskii Bay, Sea of Japan. *Russian Journal of Marine Biology* 28: 308–316. <https://doi.org/10.1023/A:1020903400805>
- Osborn KJ, Rouse GW, Goffredi SK, Robison BH (2007) Description and relationships of *Chaetopterus pugaporcinus*, an unusual pelagic polychaete (Annelida, Chaetopteridae). *The Biological Bulletin* 212: 40–54. <https://doi.org/10.2307/25066579>
- Oyarzun FX, Brante A (2015) A new case of poecilogony from South America and the implications of nurse eggs, capsule structure, and maternal brooding behavior on the development of different larval types. *The Biological Bulletin* 228: 85–97. <https://doi.org/10.1086/BBLv228n2p85>
- Oyarzun FX, Mahon AR, Swalla BJ, Halanych KM (2011) Phylogeography and reproductive variation of the poecilogonous polychaete *Boccardia proboscidea* (Annelida: Spionidae) along the West Coast of North America. *Evolution & Development* 13: 489–503. <https://doi.org/10.1111/j.1525-142X.2011.00506.x>
- Palumbi S, Martin A, Romano S, McMillan WO, Stice L, Grabowski G (1991) The simple fool's guide to PCR. Version 2.0. Department of Zoology and Kewalo Marine Laboratory, University of Hawaii, Honolulu, 47 pp.
- Pedersen TM, Almeda R, Fotel FL, Jakobsen HH, Mariani P, Hansen BW (2010) Larval growth in the dominant polychaete *Polydora ciliata* is food-limited in a eutrophic Danish estuary (Isefjord). *Marine Ecology Progress Series* 407: 99–110. <https://doi.org/10.3354/meps08563>
- Phillips NE, Wood AR, Hamilton JS (2008) Molecular species identification of morphologically similar mussel larvae reveals unexpected discrepancy between relative abundance of

- adults and settlers. *Journal of Experimental Marine Biology and Ecology* 362: 90–94. <https://doi.org/10.1016/j.jembe.2008.06.006>
- Plate S, Husemann E (1994) Identification guide to the planktonic polychaete larvae around the island of Helgoland (German Bight). *Helgoländer Meeresuntersuchungen* 48: 1–58. <https://doi.org/10.1007/BF02366201>
- Pradillon F, Schmidt A, Peplies J, Dubilier N (2007) Species identification of marine invertebrate early stages by whole-larvae *in situ* hybridisation of 18S ribosomal RNA. *Marine Ecology Progress Series* 333: 103–116. <https://doi.org/10.3354/meps333103>
- Radashevsky VI (1983) Reproduction and larval development of the polychaete *Pseudopolydora paucibranchiata* (Spionidae) in Peter the Great Bay of the Sea of Japan. *Soviet Journal of Marine Biology* 2: 38–46. [in Russian with English abstract]
- Radashevsky VI (1985) The larval development of the polychaete *Pseudopolydora kempji japonica* (Spionidae) in Peter the Great Bay of the Sea of Japan. *Soviet Journal of Marine Biology* 2: 39–46. [in Russian with English abstract]
- Radashevsky VI (1986) Reproduction and larval development of the polychaete *Polydora ciliata* in Peter the Great Bay of the Sea of Japan. *Biologiya Morya* 6: 36–42. [In Russian, with English abstract]
- Radashevsky VI (1988) Morphology, ecology, reproduction and larval development of *Polydora uschakovi* (Polychaeta, Spionidae) in the Peter the Great Bay of the Sea of Japan. *Zoologicheski Zhurnal* 67: 870–878. [in Russian with English abstract]
- Radashevsky VI (1989) Ecology, sex determination, reproduction and larval development of commensal polychaetes *Polydora commensalis* and *Polydora glycymerica* in the Japanese Sea. In: Sveshnicov VA (Ed.) *Symbiosis in marine animals*. Russian Academy of Sciences, Moscow, 137–164. [in Russian with English abstract]
- Radashevsky VI (1993) Revision of the genus *Polydora* and related genera from the North West Pacific (Polychaeta: Spionidae). *Publications of the Seto Marine Biological Laboratory* 36: 1–60. <https://doi.org/10.5134/176224>
- Radashevsky VI (2005) On adult and larval morphology of *Polydora cornuta* Bosc, 1802 (Annelida: Spionidae). *Zootaxa* 1064: 1–24. <https://doi.org/10.11646/zootaxa.1064.1.1>
- Radashevsky VI (2007) Morphology and biology of a new *Rhynchospio* species (Annelida: Spionidae) from the South China Sea, Vietnam, with the review of *Rhynchospio* taxa. *Journal of Natural History* 41: 985–997. <https://doi.org/10.1080/00222930701376717>
- Radashevsky VI (2012) Spionidae (Annelida) from shallow waters around the British Islands: an identification guide for the NMBAQC Scheme with an overview of spionid morphology and biology. *Zootaxa* 3152: 1–35. <https://doi.org/10.11646/zootaxa.3152.1.1>
- Radashevsky VI, Cárdenas CA (2004) Morphology and biology of *Polydora rickettsi* (Polychaeta: Spionidae) from Chile. *New Zealand Journal of Marine and Freshwater Research* 38: 243–254. <https://doi.org/10.1080/00288330.2004.9517234>
- Radashevsky VI, Lana PC (2009) *Laonice* (Annelida: Spionidae) from South and Central America. *Zoosymposia* 2: 265–295. <https://doi.org/10.11646/zoosymposia.2.1.19>
- Radashevsky VI, Lana PC, Nalesso RC (2006) Morphology and biology of *Polydora* species (Polychaeta: Spionidae) boring into oyster shells in South America, with the description of a new species. *Zootaxa* 1353: 1–37. <https://www.biotaxa.org/Zootaxa/article/view/zootaxa.1353.1.1>

- Radashevsky VI, Migotto AE (2006) Photoreceptive organs in larvae of Spionidae (Annelida) and Sipuncula. *Journal of Zoology* 268: 335–340. <https://doi.org/10.1111/j.1469-7998.2006.00092.x>
- Radashevsky VI, Migotto AE (2009) Morphology and biology of a new *Pseudopolydora* (Annelida: Spionidae) species from Brazil. *Journal of the Marine Biological Association of the United Kingdom* 89: 461–468. <https://doi.org/10.1017/S002531540800177X>
- Radashevsky VI, Pankova VV (2013) Shell-boring versus tube-dwelling: is the mode of life fixed or flexible? Two cases in spionid polychaetes (Annelida, Spionidae). *Marine Biology* 160: 1619–1624. <https://doi.org/10.1007/s00227-013-2214-8>
- Radashevsky VI, Petersen ME (2005) On the morphology and distribution of *Dipolydora giardi* and status of *D. trilobata* (Annelida: Spionidae). *Zootaxa* 1086: 25–36. <https://doi.org/10.11646/zootaxa.1086.1.2>
- Radashevsky VI, Migotto AE (2017) First report of the polychaete *Polydora hoplura* (Annelida: Spionidae) from North and South America and Asian Pacific. *Marine Biodiversity* 47: 859–868. <https://doi.org/10.1007/s12526-016-0515-0>
- Radashevsky VI, Díaz M, Bertrán C (2006) Morphology and biology of *Prionospio patagonica* (Annelida: Spionidae) from Chile. *Journal of the Marine Biological Association of the United Kingdom* 86: 61–69. <https://doi.org/10.1017/S0025315406012860>
- Radashevsky VI, Alexandrova YN, Yurchenko OV (2011) Spermiogenesis and spermatozoa ultrastructure of *Aonides oxycephala* (Annelida: Spionidae) from the Sea of Japan. *Invertebrate Reproduction & Development* 55: 168–174. <https://doi.org/10.1080/07924259.2011.553410>
- Radashevsky VI, Neretina TV, Pankova VV, Tzetlin AB, Choi J-W (2014) Molecular identity, morphology and taxonomy of the *Rhynchospio glutaea* complex with a key to *Rhynchospio* species (Annelida, Spionidae). *Systematics and Biodiversity* 12: 424–433. <https://doi.org/10.1080/14772000.2014.941039>
- Radashevsky VI, Malyar VV, Pankova VV, Nuzhdin S (2016a) Molecular analysis of six *Rhynchospio* species Hartman, 1936 species (Annelida: Spionidae) with comments on the evolution of brooding within the group. *Zootaxa* 4127: 579–590. <https://doi.org/10.11646/zootaxa.4127.3.10>
- Radashevsky VI, Pankova VV, Neretina TV, Stupnikova AN, Tzetlin AB (2016b) Molecular analysis of the *Pygospio elegans* group of species (Annelida: Spionidae). *Zootaxa* 4083: 239–250. <https://doi.org/10.11646/zootaxa.4083.2.4>
- Radashevsky VI, Rizzo AE, Peixoto AJM (2018) First record of *Trochochaeta japonica* (Annelida: Spionidae) in Brazil with identification key to species of the genus. *Zootaxa* 4462: 566–578. <https://doi.org/10.11646/zootaxa.4462.4.8>
- Radashevsky VI, Pankova VV, Malyar VV, Neretina TV, Wilson RS, Worsfold TM, Diez ME, Harris LH, Hourdez S, Labruno C, Houbin C, Kind B, Kuhlenkamp R, Nygren A, Bonifácio P, Bachelet G (2019) Molecular analysis and new records of the invasive polychaete *Boccardia proboscidea* (Annelida: Spionidae). *Mediterranean Marine Science* 20: 393–408. <https://doi.org/10.12681/mms.20363>
- Radashevsky VI, Pankova VV, Malyar VV, Neretina TV, Choi J-W, Yum S, Houbin C (2020a) Molecular analysis of *Spiophanes bombyx* complex (Annelida: Spionidae) with description of a new species. *PLoS ONE* 15: e0234238. <https://doi.org/10.1371/journal.pone.0234238>

- Radashevsky VI, Malyar VV, Pankova VV, Gambi MC, Giangrande A, Keppel E, Nygren A, Al-Kandari M, Carlton JT (2020b) Disentangling invasions in the sea: molecular analysis of a global polychaete species complex (Annelida: Spionidae: *Pseudopolydora paucibranchiata*). *Biological Invasions* 22: 3621–3644. <https://doi.org/10.1007/s10530-020-02346-x>
- Rasmussen E (1973) Systematics and ecology of the Isefjord marine fauna (Denmark). *Ophelia* 11: 1–495. <https://doi.org/10.1080/00785326.1973.10430115>
- Read G, Fauchald K (2020) World Polychaeta database. Spionidae Grube, 1850. World Register of Marine Species. <http://www.marinespecies.org/aphia.php?p=taxdetails&id=913> [Accessed on 5 May 2020]
- Reddy VP, Mohan PC (1982) On the occurrence of larval stages of *Poecilochaetus johnsoni* Hartman 1939 (Polychaeta: Trochochaetidae) in the plankton off Waltair Coast. *Mahasagar Bulletin of the National Institute of Oceanography* 15: 243–246.
- Rengaiyan P, Ingole B (2018) 18S rDNA sequencing data of benthic polychaetes from the Eastern Arabian Sea. Data in Brief 20: 1749–1752. <https://doi.org/10.1016/j.dib.2018.09.015>
- Rice SA, Karl S, Rice KA (2008) The *Polydora cornuta* complex (Annelida: Polychaeta) contains populations that are reproductively isolated and genetically distinct. *Invertebrate Biology* 127: 45–64. <https://doi.org/10.1111/j.1744-7410.2007.00104.x>
- Rouse GW (2006) Chapter 6 Annelid larval morphology. In: Rouse GW, Pleijel F (Eds) *Reproductive Biology and Phylogeny of Annelida*. Science Publishers, Enfield, 141–177.
- Rousset V, Rouse GW, Siddall ME, Tillier A, Pleijel F (2004) The phylogenetic position of Siboglinidae (Annelida), inferred from 18S rRNA, 28S rRNA, and morphological data. *Cladistics* 20: 518–533. <https://doi.org/10.1111/j.1096-0031.2004.00039.x>
- Rousset V, Pleijel F, Rouse GW, Erseus C, Siddall ME (2007) A molecular phylogeny of annelids. *Cladistics* 23: 41–63. <https://doi.org/10.1111/j.1096-0031.2006.00128.x>
- Rullier F (1963) Développement de *Polydora (Carazzia) antennata* Clap. var. *pulchra* Carazzi. *Cahiers de Biologie Marine* 4: 233–250.
- Sasaki S, Brown R (1983) Larval development of *Saccocirrus uchidae* from Hokkaido, Japan and *Saccocirrus krusadensis* from New South Wales, Australia (Archiannelida, Saccocirridae). *Annotationes Zoologicae Japonenses* 56: 299–314.
- Sato M, Tsuchiya M (1991) Two patterns of early development in nereidid polychaetes keying out to *Neanthes japonica*. *Ophelia supplement* 5: 371–382.
- Sato-Okoshi W (1994) Life history of the polychaete *Polydora variegata* that bores into the shells of scallops in Northern Japan. *Mémoires du Muséum national d'Histoire naturelle* 162: 549–558. <http://pascal-francis.inist.fr/vibad/index.php?action=getRecordDetail&idt=3360957>
- Sato-Okoshi W (1998) Three new species of polydorids (Polychaeta, Spionidae) from Japan. *Species Diversity* 3: 277–288. <https://doi.org/10.12782/specdiv.3.277>
- Sato-Okoshi W (1999) Polydorid species (Polychaeta: Spionidae) in Japan, with descriptions of morphology, ecology and burrow structure. 1. Boring species. *Journal of the Marine Biological Association of the United Kingdom* 79: 831–848. <https://doi.org/10.1017/S0025315498001003>
- Sato-Okoshi W (2000) Polydorid species (Polychaeta: Spionidae) in Japan, with descriptions of morphology, ecology and burrow structure. 2. Non-boring species. *Journal of the Marine Biological Association of the United Kingdom* 80: 443–456. <https://doi.org/10.1017/S0025315499002143>

- Sato-Okoshi W, Abe H (2012) Morphological and molecular sequence analysis of the harmful shell boring species of *Polydora* (Polychaeta: Spionidae) from Japan and Australia. *Aquaculture* 368–369: 40–47. <https://doi.org/10.1016/j.aquaculture.2012.08.046>
- Sato-Okoshi W, Abe H (2013) Morphology and molecular analysis of the 18S rRNA gene of oyster shell borers, *Polydora* species (Polychaeta: Spionidae), from Japan and Australia. *Journal of the Marine Biological Association of the United Kingdom* 93: 1279–1286. <https://doi.org/10.1017/S002531541200152X>
- Sato-Okoshi W, Sugawara Y, Nomura T (1990) Reproduction of the boring polychaete *Polydora variegata* inhabiting scallops in Abashiri Bay, North Japan. *Marine Biology* 104: 61–66. <https://doi.org/10.1007/BF01313158>
- Sato-Okoshi W, Okoshi K, Shaw J (2008) Polydorid species (Polychaeta: Spionidae) in southwestern Australian waters with special reference to *Polydora uncinata* and *Boccardia knoxi*. *Journal of the Marine Biological Association of the United Kingdom* 88: 491–501. <https://doi.org/10.1017/S0025315408000842>
- Sato-Okoshi W, Abe H, Nishitani G, Simon CA (2017) And then there was one: *Polydora uncinata* and *Polydora hoplura* (Annelida: Spionidae), the problematic polydorid pest species represent a single species. *Journal of the Marine Biological Association of the United Kingdom* 97: 1675–1684. <https://doi.org/10.1017/S002531541600093X>
- Scheltema RS, Blake JA, Williams IP (1997) Planktonic larvae of spionid and chaetopterid polychaetes from off the west coast of the Antarctic Peninsula. *Bulletin of Marine Science* 60: 396–404.
- Shanks AL (1986) Tidal periodicity in the daily settlement of intertidal barnacle larvae and an hypothesized mechanism for the cross-shelf transport of cyprids. *The Biological Bulletin* 170: 429–440. <https://doi.org/10.2307/1541852>
- Shanks AL (2001) *An Identification Guide to the Larval Marine Invertebrates of the Pacific Northwest*. Oregon State University Press, Corvallis, 314 pp. <http://hdl.handle.net/1794/6123>
- Sigvaldadóttir E, Mackie ASY (1993) *Prionospio steenstrupi*, *P. fallax* and *P. dubia* (Polychaeta, Spionidae): re-evaluation of identity and status. *Sarsia* 78: 203–219. <https://doi.org/10.1080/00364827.1993.10413535>
- Sigvaldadóttir E, Mackie ASY, Pleijel F (1997) Generic interrelationships within the Spionidae (Annelida: Polychaeta). *Zoological Journal of the Linnean Society* 119: 473–500. <https://doi.org/10.1111/j.1096-3642.1997.tb00144.x>
- Sikorski AV (2002) On distinguishing the morphologically close species, *Laonice cirrata* and *L. bahusiensis* (Polychaeta, Spionidae). *Zoologicheskij Zhurnal* 81: 406–419. [in Russian with English summary]
- Sikorski AV (2011) Review of *Laonice* (Spionidae, Annelida) with remarks on several species and a description of a new species from South Africa. *Italian Journal of Zoology* 78: 201–214. <https://doi.org/10.1080/11250003.2011.617218>
- Simon JL (1963) Reproduction and larval development of the spionid polychaete *Spio setosa*, Verrill. PhD thesis, University of New Hampshire, Durham.
- Simon JL (1967) Reproduction and larval development of *Spio setosa* (Spionidae: Polychaeta). *Bulletin of Marine Science* 17: 398–431. <https://www.ingentaconnect.com/content/umrsmas/bullmar/1967/00000017/00000002/art00015>

- Simon JL (1968) Occurrence of pelagic larvae in *Spio setosa* Verrill 1873 (Polychaeta: Spionidae). The Biological Bulletin 134: 503–515. <https://doi.org/10.2307/1539868>
- Simon CA (2011) *Polydora* and *Dipolydora* (Polychaeta: Spionidae) associated with molluscs on the south coast of South Africa, with descriptions of two new species. African Invertebrates 52: 39–50. <https://doi.org/10.5733/afin.052.0104>
- Simon CA (2015) Observations on the composition and larval developmental modes of polydorid pests of farmed oysters (*Crassostrea gigas*) and abalone (*Haliotis midae*) in South Africa. Invertebrate Reproduction & Development 59: 124–130. <https://doi.org/10.1080/07924259.2015.1044675>
- Simon CA, Sato-Okoshi W (2015) Polydorid polychaetes on farmed molluscs: distribution, spread and factors contributing to their success. Aquaculture Environment Interactions 7: 147–166. <https://doi.org/10.3354/aei00138>
- Simon CA, Ludford A, Wynne S (2006) Spionid polychaetes infesting cultured abalone *Haliotis midae* in South Africa. African Journal of Marine Science 28: 167–171. <https://doi.org/10.2989/18142320609504141>
- Simon CA, Worsfold TM, Lange L, Sterley J (2010) The genus *Boccardia* (Polychaeta: Spionidae) associated with mollusc shells on the south coast of South Africa. Journal of the Marine Biological Association of the United Kingdom 90: 585–598. <https://doi.org/10.1017/S0025315409990452>
- Simon CA, Sato-Okoshi W, Abe H (2019a) Hidden diversity within the cosmopolitan species *Pseudopolydora antennata* (Claparède, 1869) (Spionidae: Annelida). Marine Biodiversity 49: 25–42. <https://doi.org/10.1007/s12526-017-0751-y>
- Simon CA, Williams L-G, Henninger T (2019b) A new species of *Rhynchospio* (Annelida: Spionidae) in South Africa. Marine Biodiversity 49: 663–672. <https://doi.org/10.1007/s12526-017-0842-9>
- Söderström A (1920) Studien über die Polychäten familie Spionidae. PhD thesis, Almqvist & Wicksells, Uppsala, 286 pp. [1 plate]
- Srikrishnadhas B, Ramamoorthi K (1977) Development of *Pseudopolydora kempfi* (Southern, 1921) in the laboratory. Proceedings of the Symposium on Warm Water Zooplankton. Special Publication UNESCO/NIO, 671–677.
- Srikrishnadhas B, Ramamoorthi K (1981) Studies on the life-history of *Spio filicornis* (Muller 1776). Mahasagar – Bulletin of the National Institute of Oceanography 14: 303–307. <http://www.ijs.nio.org/index.php/msagar/article/view/2382>
- Struck TH, Purschke G (2005) The sister group relationship of Aeolosomatidae and Potamodrilidae (Annelida: “Polychaeta”) – a molecular phylogenetic approach based on 18S rDNA and cytochrome oxidase I. Zoologischer Anzeiger 243: 281–293. <https://doi.org/10.1016/j.jcz.2005.01.001>
- Struck TH, Schult N, Kusen T, Hickman E, Bleidorn C, McHugh D, Halanych KM (2007) Annelid phylogeny and the status of Sipuncula and Echiura. BMC Evolutionary Biology 7: 1–57. <https://doi.org/10.1186/1471-2148-7-57>
- Struck TH, Nesnidal MP, Purschke G, Halanych KM (2008) Detecting possibly saturated positions in 18S and 28S sequences and their influence on phylogenetic reconstruction of Annelida (Lophotrochozoa). Molecular Phylogenetics and Evolution 48: 628–645. <https://doi.org/10.1016/j.ympev.2008.05.015>

- Takata N, Noguchi D, Tanaka M, Awakihara H (2011) Genetic characteristics of *Polydora cornuta* complex (Spionidae; Polychaeta) inhabiting an eutrophic port of Fukuyama. *Eco-engineering* 23: 101–104. <https://doi.org/10.11450/seitaikogaku.23.101>
- Talavera G, Castresana J (2007) Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Systematic Biology* 56: 564–577. <https://doi.org/10.1080/10635150701472164>
- Teramoto W, Sato-Okoshi W, Abe H, Nishitani G, Endo Y (2013) Morphology, 18S rRNA gene sequence, and life history of a new *Polydora* species (Polychaeta: Spionidae) from northeastern Japan. *Aquatic Biology* 18: 31–45. <https://doi.org/10.3354/ab00485>
- Thorson G (1946) Reproduction and larval development of Danish marine bottom invertebrates. *Meddelelser fra Kommissionen for Danmarks Fiskeri-og Havundersøgelser, Series Plankton* 4: 1–523.
- Tokioka T (1970) Droplets from the plankton net. XXV. record of a Rostraria from Seto. *Publication of the Seto Marine Biological Laboratory* 18: 257–278. <https://doi.org/10.5134/175636>
- Tomiyama T, Katayama S, Omori M, Honda H (2005) Importance of feeding on regenerable parts of prey for juvenile stone flounder *Platichthys bicoloratus* in estuarine habitats. *Journal of Sea Research* 53: 297–308. <https://doi.org/10.1016/j.seares.2005.01.001>
- Tomiyama T, Omori M, Minami T (2007) Feeding and growth of juvenile stone flounder in estuaries: generality and the importance of sublethal tissue cropping of benthic invertebrates. *Marine Biology* 151: 365–376. <https://doi.org/10.1007/s00227-006-0491-1>
- Tosuji H, Sato M (2006) Salinity favorable for early development and gamete compatibility in two sympatric estuarine species of the genus *Hediste*. *Marine Biology* 148: 529–539. <https://doi.org/10.1007/s00227-005-0079-1>
- Vortsepneva E, Tzetlin A, Purschke G, Mugue N, Haß-Cordes E, Zhadan A (2008) The parasitic polychaete known as *Asetocalamyzas laonicola* (Calamyzidae) is in fact the dwarf male of the spionid *Scolecopsis laonicola* (comb. nov.). *Invertebrate Biology* 127: 403–416. <https://doi.org/10.1111/j.1744-7410.2008.00137.x>
- Walker LM (2013) A revision of the *Polydora*-complex (Annelida: Spionidae) fauna from Australia. PhD thesis. University of Queensland, Brisbane.
- Ward LA (1977) A Key to the larvae of the polychaetous annelids of the Hawaiian Islands. MOP Student Reports, University of Hawaii, Marine Option Program, Honolulu, 78 pp. <http://hdl.handle.net/10125/23518> [accessed on 7 Nov 2018]
- Williams JD (2001) Reproduction and larval development of *Polydora robi* (Polychaeta: Spionidae), an obligate commensal of hermit crabs from the Philippines. *Invertebrate Biology* 120: 237–247. <https://doi.org/10.1111/j.1744-7410.2001.tb00034.x>
- Williams JD, McDermott JJ (1997) Feeding behavior of *Dipolydora commensalis* (Polychaeta: Spionidae): particle capture, transport, and selection. *Invertebrate Biology* 116: 115–123. <https://doi.org/10.2307/3226975>
- Williams L-G, Karl S, Simon C (2017) Molecular identification of polydorid polychaetes (Annelida: Spionidae): Is there a quick way to identify pest and alien species? *African Zoology* 52: 105–117. <https://doi.org/10.1080/15627020.2017.1313131>
- Wilson DP (1928) The Larvæ of *Polydora ciliata* Johnston and *Polydora hoplura* Claparède. *Journal of the Marine Biological Association of the United Kingdom* 15: 567–603. <https://doi.org/10.1017/S0025315400009553>

- Woodwick KH (1977) Lecithotrophic larval development in *Boccardia proboscidea* Hartman. In: Reish DJ, Fauchald K (Eds) Essays on Polychaetous Annelids in Memory of Dr Olga Hartman. Allan Hancock Foundation, University of Southern California, Los Angeles, 347–371.
- Wu BL, Chen M (1980) Morphology, ecology, reproduction and larval development of *Pseudopolydora paucibranchia* (Okuda). *Acta Zoologica Sinica* 26: 356–364. [in Chinese with English abstract]
- Wu BL, Chen M, Sun RP (1965) On the occurrence of a spionid worm, *Spio martinensis* Mesnil in the Yellow Sea, with notes on its larval development. *Oceanologia et Limnologia Sinica* 7: 25–31. [in Chinese with English abstract] http://en.cnki.com.cn/Article_en/CJFDTotal-HYFZ196501001.htm
- Wu BL, Sun RP, Shen SP (1978) A preliminary report on the polychaete larvae in the plankton from the Zhongsha Islands, Guangdong Province, China. Report on the scientific results of marine biology of the Xisha Islands and Zhongsha Islands (South China Sea). South China Sea Institute of Oceanology, Academia Sinica, Guangzhou, 171–200. [in Chinese with English abstract]
- Yamaji I (1966) Illustrations of the marine plankton of Japan. Hoikusha Publishing Co. Ltd, Osaka, 369 pp. [in Japanese]
- Ye L, Cao C, Tang B, Yao T, Wang R, Wang J (2017) Morphological and molecular characterization of *Polydora websteri* (Annelida: Spionidae), with remarks on relationship of adult worms and larvae using mitochondrial COI gene as a molecular marker. *Pakistan Journal of Zoology* 49: 699–710. <https://doi.org/10.17582/journal.pjz/2017.49.2.699.710>
- Ye L, Tang B, Wu K, Su Y, Wang R, Yu Z, Wang J (2015) Mudworm *Polydora lingshuiensis* sp. n is a new species that inhabits both shell burrows and mudtubes. *Zootaxa* 3986: 88–100. <https://doi.org/10.11646/zootaxa.3986.1.4>
- Yokouchi K (1984) Surface distribution of polychaete larvae in Volcano Bay, southern Hokkaido, during the vernal phytoplankton blooms of 1982. *Bulletin of Plankton Society of Japan* 31: 113–122.
- Yokouchi K (1985) Reproduction and larval ecology of the sandworm *Neanthes virens* (Sars) from southern Hokkaido. *Bulletin of Plankton Society of Japan* 32: 1–13.
- Yokouchi K (1988) Hokkai-do Oshima-Hantou Usujiri Egan ni okeru Teisei Tamourui Huyuu-ki Yousei no Kisetsu-Bunpu ni kansuru Kenkyu [Study on the seasonal distribution of planktonic larvae of benthic polychaetes in coastal area of Usujiri, Oshima Peninsula, Hokkaido]. PhD thesis, Hokkaido University, Sapporo. [in Japanese]
- Yokouchi K (1991) Seasonal distribution and food habits of planktonic larvae of benthic polychaeta in Volcano Bay, southern Hokkaido, Japan. *Ophelia Supplement* 5: 401–410.
- Yokouchi K, Yokouchi M (1997) Zooplankton larvae phylum Annelida. In: Chihara M, Murano M (Eds) An illustrated guide to marine plankton in Japan. Tokai University Press, Tokyo, 1430–1438. [in Japanese]
- Yokoyama H (1981) Larval development of a spionid polychaete *Paraprionospio pinnata* (Ehlers). *Publication of the Seto Marine Biological Laboratory* 26: 157–170. <https://doi.org/10.5134/176017>
- Yokoyama H (1990) Life history and population structure of the spionid polychaete *Paraprionospio* sp. (form A). *Journal of Experimental Marine Biology and Ecology* 144: 125–143. [https://doi.org/10.1016/0022-0981\(90\)90024-7](https://doi.org/10.1016/0022-0981(90)90024-7)

- Yokoyama H (1995) Occurrence of *Paraprionospio* sp. (Form A) larvae (Polychaeta: Spionidae) in hypoxic water of an enclosed bay. *Estuarine Coastal and Shelf Science* 40: 9–19. [https://doi.org/10.1016/0272-7714\(95\)90009-8](https://doi.org/10.1016/0272-7714(95)90009-8)
- Yokoyama H (1996) Larvae of the spionid polychaete *Paraprionospio* sp. (form B) found in the plankton from Omura Bay. *Bulletin of National Research Institute of Aquaculture* 25: 17–22.
- Yokoyama H (2007) A revision of the genus *Paraprionospio* Caullery (Polychaeta: Spionidae). *Zoological Journal of the Linnean Society* 151: 253–284. <https://doi.org/10.1111/j.1096-3642.2007.00323.x>
- Zhang D, Gao F, Jakovlić I, Zou H, Zhang J, Li WX, Wang GT (2020) PhyloSuite: an integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Molecular Ecology Resources* 20: 348–355. <https://doi.org/10.1111/1755-0998.13096>
- Zhang S-L, Li X-D, Bi X-D (2009) Studies on the larval development of *Polydora ligni*. *Sichuan Journal of Zoology* 28: 801–809. [in Chinese with English abstract] http://en.cnki.com.cn/Article_en/CJFDTotal-SCDW200906002.htm
- Zhou H, Zhang Z, Chen H, Sun R, Wang H, Guo L, Pan H (2010) Integrating a DNA barcoding project with an ecological survey: a case study on temperate intertidal polychaete communities in Qingdao, China. *Chinese Journal of Oceanology and Limnology* 28: 899–910. <https://doi.org/10.1007/s00343-010-9131-1>