RESEARCH ARTICLE



A new species of *Procamallanus* Baylis, 1923 (Nematoda, Camallanidae) from *Astronotus* ocellatus (Agassiz, 1831) (Perciformes, Cichlidae) in Brazil

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Abstract

A new species of *Procamallanus* Baylis, 1923 was found as a parasite of the fish *Astronotus ocellatus* (Agassiz, 1831) from a lake in the Jardim Botânico Bosque Rodrigues Alves, Belém, Brazil. *Procamallanus spiculastriatus* **sp. n.** has a smooth buccal capsule and a well-developed basal ring that is armed with four sclerotized tooth-like structures. The male of the new species is similar to the two species that are known from Brazilian fish, *P. peraccuratus* Pinto, Fábio, Noronha & Rolas, 1976, and *P. annipetterae* Kohn & Fernandes, 1988, by the absence of the gubernaculum. It differs from these two by the morphology of the buccal capsule, the number are arrangement of the caudal papillae in males, the size and morphology of the spicules and the shape of the tail of both sexes. *Procamallanus spiculastriatus* **sp. n.** is the third species discovered in fish from Brazil. This finding extends the geographical distribution of the genus into the Brazilian Amazon.

Keywords

Amazon, fish, helminth, nematode

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Introduction

The genus *Astronotus* is comprised of two species, *A. crassipinnis* (Heckel, 1840) and *A. ocellatus* (Agassiz, 1831) (Perciformes: Cichlidae) (Kullander 2003; Froese and Pauly 2017). *Astronotus ocellatus* (known as Acaraú-açu; Apaiari; Oscar) is popular among aquarists, but it is not as popular for aquaculture because of its slow growth rate. Early attempts at cultivation were encouraged by the government in the early 1970s, but they were not successful they only succeeded in the introduction of the species into almost all of Brazil (Fontenele and Nepomuceno 1983). Because it is an introduced species, there has been little interest on studies of its parasites; to date, only three studies (Azevedo et al. 2007; Malta 1984; Neves et al. 2013) have included this species of fish.

Nematodes of the genus *Procamallanus* Baylis, 1923 (Camallanida, Procamallaninae) are predominately parasites of freshwater fish that are distributed over several zoogeographical regions (Moravec 1998; Hoffman 1999). Members of the genus are easily recognized by the presence of a buccal capsule that is strongly sclerotized, but without ridges (Baylis 1923; Moravec and Scholz 1991; Moravec and Thatcher 1997). Despite the importance of introduced species of fish as mechanisms of co-introduction of parasites into native populations (Bautista-Hernández et al. 2014), helminths of most of introduced species have not been studied. As part of an ongoing study of the helminths of vertebrates of eastern Brazil, samples of *A. ocellatus* were collected and necropsied. Procamallanin nematodes were found as parasites of these fish, but they could not be assigned to a known species; therefore, the new species is described herein.

Materials and methods

Forty specimens of A. ocellatus were collected from Iara Lake, at the Jardim Botânico Bosque Rodrigues Alves (1°25'49"S, 48°27'22"W), located in an urban area of the city of Belém, state of Pará, eastern Brazilian Amazon. Fish were collected during March to July 2015 with the aid of a casting net. Fish were transported alive to the Laboratório de Biologia Celular e Helmintologia "Profa. Dra. Reinalda Marisa Lanfredi", Instituto de Ciências Biológicas, Universidade Federal do Pará-UFPA, for necropsy. Nematodes were collected, washed in phosphate-buffered saline, fixed in AFA solution (93 parts 70% ethanol, 5 parts formalin, and 2 parts glacial acetic acid) (Pritchard and Kruse 1982), and stored in 70% ethanol. For light microscopy, helminths were cleared in Amman's lactophenol solution (Giese et al. 2009) and examined under an Olympus BX41 microscope with a drawing tube. Measurements are given in micrometers unless otherwise noted and are presented as the range (minimum and maximum values) followed by the mean in parentheses. For scanning electron microscopy (SEM), helminths were washed in phosphate-buffered saline with a pH of 7.0 (Sodium Phosphate Monobasic 3.12 g, Sodium Phosphate Dibasic 2.83 g, and 17 g Sodium Chloride in 200 ml of distilled water), post-fixed in 1% osmium tetroxide, dehydrated to the critical point using CO₂, coated with gold+palladium, and studied using a scanning

electron microscope (VEGA 3 LMU/TESCAN) in the Laboratório de Histologia e Embriologia Animal - Instituto da Saúde e Produção Animal - Universidade Federal Rural da Amazônia – UFRA, campus Belém, state of Pará, Brazil. Type material was deposited in the Coleção de Invertebrados of the Museu Paraense Emílio Goeldi (MPEG), Belém, Pará, Brazil. Other material examined included specimens of Procamallanus peraccuratus Pinto, Fábio, Noronha & Rolas, 1976, Coleção Helmintologica do Instituto Oswaldo Cruz (CHIOC): females-16.747A-C, 16.759A-C, 31.082A, 31.083A-C; males-16.753B-D, 16.757A-B, 16.769A-C, 16.773B, 31.083A-B); Spirocamallanus inopinatus Travassos, Artigas & Pereira, 1928, CHIOC 31.315A-B, CHIOC 31.323A-B, CHIOC 31.324, CHIOC 31.325A-B, CHIOC 31.326A-B, CHIOC 31.327, CHIOC 31.328 and CHIOC 31.329; S. rarus Travassos, Artigas & Pereira, 1928, CHIOC 31.027A-B, CHIOC 31.328A-C; S. pexatus Pinto, Fabio, Noronha & Rolas, 1974, CHIOC 31.086A-D, CHIOC 31.087, CHIOC 31.088A-B, CHIOC 31.089A-B and 32.504A-B; S. paraensis Pinto, Fabio, Noronha & Rolas, 1976, CHIOC 31.342A-E; and S. saofranciscensis Moreira, Oliveira & Costa, 1994, CHIOC 37.857, CHIOC 37.858 and CHIOC 38.042.

Systematics

Family Camallanidae Railliet & Henry, 1915 Subfamily Camallaninae Railliet & Henry, 1915 Genus *Procamallanus* Baylis, 1923

Procamallanus spiculastriatus sp. n. http://zoobank.org/AD83ABF3-9B63-455C-8AC6-3B3795CD6156 Figures 1, 2, 3

Material. Type specimens. Holotype male (MPEG 195), allotype female (MPEG 196), and four paratypes (MPEG 197; MPEG 198; MPEG 199; MPEG 200) were deposited in the Coleção de Invertebrados of the Museu Paraense Emílio Goeldi (MPEG), Belém, Pará, Brazil.

Type host. *Astronotus ocellatus* (Agassiz) (Perciformes: Cichlidae). Average length = 24.7±2.6 cm; average weight = 331.8±96.3 g.

Type locality. Iará Lake, Jardim Botânico Bosque Rodrigues Alves (1°25'49"S, 48°27'22"W), Belém, Pará, Amazon Biome, Brazil.

Site in host. Mid-intestine.

Host-parasite data. Prevalence 55% (22 infected, 40 examined); Mean intensity = 14.8; Mean abundance = 8.5; Range = 1–59.

Etymology. The species name refers to the unique morphology of the spicules, which membranous alae that are supported by rays, giving them a striated appearance.

Description. [Based on 10 males, 11 females, 20 eggs (from allotype female), and 20 intrauterine larvae (from allotype female)] Medium-sized nematodes, red while

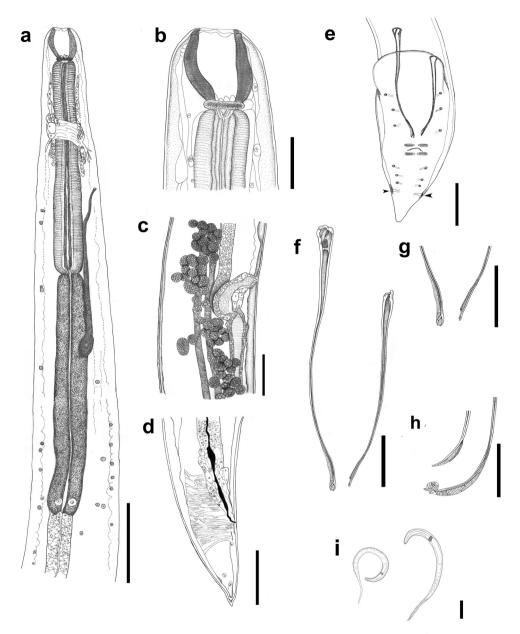


Figure 1. *Procamallanus spiculastriatus* sp. n., line drawings **a** adult male, anterior part of the body, lateral view **b** buccal capsule, lateral view **c** vulvar opening and vagina **d** tail of a female worm, lateral view **e** posterior end of a male worm and phasmids (arrowheads), ventral view **f** spicules, ventral view **g** distal part of spicules, ventral view **h** distal end of spicules, lateral view **i** larvae. Scale bars: 150 μ m (**a**); 40 μ m (**b**); 100 μ m (**c**, **d**); 20 μ m (**e**); **f**, 50 μ m; 50 μ m (**g**, **h**, **i**).

alive and white after fixation. Cuticle with fine transverse striations. Oral opening circular, surrounded by three concentric circles with four papillae each, inner circle with six small pores at base proximal to oral opening, pair of small lateral amphids present

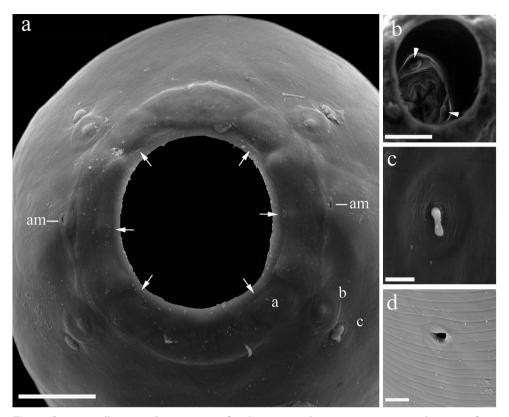


Figure 2. *Procamallanus spiculastriatus* sp. n., female, scanning electron microscopy. **a** oral opening, frontal view, three circles of cephalic papillae (a, b, c), amphid (am, arrowhead), pore-like structures (arrows) **b** oral opening, view of buccal capsule, two teeth are visible at the base of the basal ring (arrows) **c** deirid **d** excretory pore. Scale bars: 10 μ m (**a**, **b**); 2 μ m (**c**); 5 μ m (**d**).

(Figure 2a). Buccal capsule, orange-brown, barrel-shaped with a well-developed basal ring armed with four sclerotized tooth-like structures (Figs 1a, b, 2b). Inner surface of capsule smooth (Figure 1a, b) without ridges. Muscular esophagus somewhat shorter than glandular esophagus. Deirids minute, simple with rounded tip, situated between the buccal capsule and nerve ring (Figure 2c).

Males (based on holotype and 9 paratypes): body 8–11 (9) mm long; maximum width at esophageal/intestinal junction 105–147 (130). Buccal capsule including basal ring 57–74 (65) long and 32–39 (36) wide, basal ring 5–8 (6.5) long, 22–29 (26) wide. Maximum length/width ratio of buccal capsule 1:0.55. Deirids, nerve-ring and excretory pore at 91–119 (104), 156–188 (171) and 248–292 (263), respectively, from anterior extremity. Muscular portion of esophagus 316–395 (353) long and 42–53 (48) wide; glandular portion of esophagus 421–558 (470) long and 42–63 (51) wide. Muscular/glandular esophagus length ratio 1:1.3. Length of entire esophagus and buccal capsule 9–12% of body length. Posterior end of body ventrally curved, provided with wide caudal alae bearing six pairs of pedunculated papillae: three precloacal pairs and three postcloacal pairs (Figs 1e, 3a). Two pairs of adcloacal papillae. (Figs 1e, 3a).

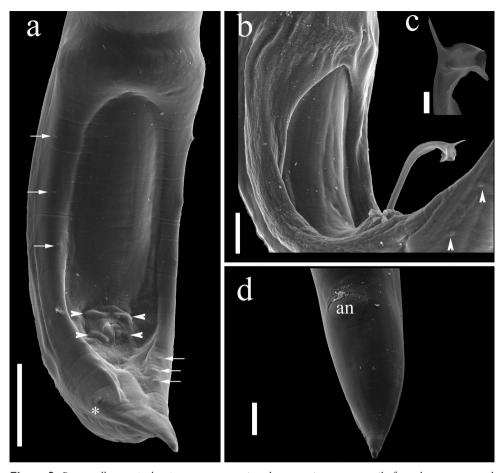


Figure 3. *Procamallanus spiculastriatus* sp. n., scanning electron microscopy. **a** tail of a male worm, ventral view, three preanal pairs (arrows) and three postanal pairs (arrows), four adcloacal papillae (arrowheads) and a lateral phasmid (*) are visible **b** tail of a male worm, lateral view with the spicule partially extroverted, and two pairs of sessile papillae are located along lateral margin (arrowhead); inset **c** detail of the tip of a spicule **d** tail of a female worm, ventral view, anus (an). Scale bars: 25 μm (**a**); 50 μm (**b**); 5 μm (**c**); 50 μm (**d**).

Two pairs of dorso-lateral sessile papillae, between cloaca and tip of tail present (Figure3b). Caudal alae anteriorly interconnected, forming a pseudosucker, and not reaching tip of tail; a pair of phasmids located immediately posterior to the 6th pair of pedunculated papillae (Figs 1e, 3a). Spicules elongate and ventrally curved, with slightly sclerotized core; distal end of spicules with membranous alae supported by sclerotized rays (Figs 1e–h, 3b, c). Spicules with terminal bifurcation, similar in form, bifurcation of right spicule always larger than that of left spicule. Spicules dissimilar in length; left spicule shorter, 229–284 (247) long and right spicule 312–355 (332) long (Figure 1e–h). Gubernaculum absent. Length of tail 156–205 (184).

Females with larvae (based on 4 specimens): body 17–20 mm (18 mm) long; maximum width at esophageal/intestinal junction 160–173 (167). Buccal capsule including

basal ring 78–83 (81) long and 47–52 (50) wide, basal ring 12–13 (12) long, 32–43 (36) wide. Maximum length/width ratio of buccal capsule 1:0.62. Deirids, nerve-ring, and excretory pore at 147–167 (156), 220–233 (225), and 330–397 (371), respectively, from the anterior extremity (Figure 1c). Muscular portion of esophagus 413–453 (433) long and 53–60 (58) wide; glandular portion of esophagus 600–693 (633) long and 67–80 (70) wide. Muscular/glandular esophagus length ratio 1:1.5. Length of entire esophagus and buccal capsule representing 6–7% of body length. Vulva situated at, 8–11 (9) mm from anterior end, at about 50% of body length; vulval lips not elevated. Muscular vagina directed posteriorly (Figure 1c); uterus filled with larvae 210–280 (245±22 long) (Figure 1i) and eggs 17–29 (24) long by 14–26 (22) wide. Tail conical, 190–220 (208) long, without cuticular projections (Figs 1d, 3d).

Females with eggs (based on 7 specimens): body 11–13 mm (12 mm) long; maximum width at esophageal/intestinal junction 100–133 (124). Buccal capsule including basal ring 80–87 (82) long and 50–60 (53) wide, basal ring 10–12 (10) long, 33–38 (35) wide. Maximum length/width ratio of buccal capsule 1:0.64. Deirids, nerve-ring, and excretory pore at 127–163 (144), 183–227 (208), and 283–357 (308), respectively, from the anterior extremity. Muscular portion of esophagus 387–460 (427) long and 47–60 (52) wide; glandular portion of esophagus 447–527 (489) long and 60–67 (61) wide. Muscular/glandular esophagus length ratio 1:1.5. Length of entire esophagus and buccal capsule representing 8–9% of body length. Vulva situated at 6–7 mm (6 mm) from anterior end, at 52% of body length; vulval lips not elevated. Muscular vagina directed posteriorly; uterus filled with eggs 28–32 (29) long by 25–30 (26) wide. Tail conical, 140–180 (166) long, without cuticular projections.

Discussion

The family Camallanidae was established for species with a prominent, sclerotized buccal capsule (Railliet and Henry 1915). Yeh (1960) divided the family into two subfamilies, Camallaninae Railliet & Henry, 1915, for species with the buccal capsule divided into two halves, and Procamallaninae Yeh, 1960, for those with a single, cup-like buccal capsule. The new species with its cup-like buccal capsule composed of two lateral halves is identified as a member of Procamallaninae. Six genera have been assigned to Procamallaninae: Procamallanus, Spirocamallanus Olsen, 1952, Malayocamallanus Jothy & Fernando, 1970, Punctocamallanus Moravec & Scholz, 1991, Spirocamallanoides Moravec & Sey, 1988 and Denticamallanus Moravec & Thatcher, 1997 (Baylis 1923; Olsen 1952; Jothy and Fernando 1970; Moravec and Scholz 1991; Moravec and Thatcher 1997; Rigby and Rigby 2014). The genus Procamallanus consists of approximately 40 known species distributed worldwide, but only two species have been reported from Brazil, *P. peraccuratus* and *P. annipetterae* Kohn & Fernandes, 1988, both from southern Brazil (Pinto et al. 1976; Petter and Dlouhy 1985; Kohn and Fernandes 1988). The new species is assigned to Procamallanus because its cuplike buccal capsule with smooth walls (without striations); according to Moravec and

Thatcher (1997) the main characteristic of this genus is the presence of a smooth buccal capsule in both sexes.

The new species can be distinguished from all known members of the genus outside of Brazil in having tooth-like structures on the basal ring of the buccal capsule. In addition to the above characteristic, it differs from *P. annulatus* Yamaguti, 1955 (Indonesia), *P. elatensis* Fusco & Overstreet, 1979 (Israel), *P. laeviconchus* Wedl, 1861 (Egypt), *P. planoratus* Kulkarni, 1935 (India) and *P. pseudolaeviconchus* Moravec & Van As, 2015 (Botswana) by the absence of a sclerotized gubernaculum, present in the other five species (Wedl 1861; Kulkarni 1935; Yamaguti 1955; Fusco and Overstreet 1979; Moravec and Van As 2015). *P. spiculastriatus* can be distinguished from *P. pacificus* Moravec, Justine, Würtz, Taraschewski & Sasal, 2006 (New Caledonia) also not found in Brazil, by the absence of the small processes (mucrons) (*sensu* Moravec et al. 2006) on the tip of the tail.

Two species of *Procamallanus* have been found in Brazil: *P. peraccuratus* Pinto, Fábio, Noronha & Rolas, 1976, from *Geophagus brasiliensis* (Quoy & Gaimard) and *Australoheros facetus* (Jenyns) (both Cichlidae) in the State of Espirito Santo (Southern Region of Brazil) and *P. annipetterae* Kohn & Fernandes, 1988 (= *P. petterae* Kohn & Fernandes, 1988), from *Corydoras paleatus* (Jenyns) in the Iguaçu River, State of Paraná (south of Brazil) (Pinto et al. 1976; Kohn and Fernandes 1988). Pinto et al. (1976) suggested that the characteristics of the buccal capsule and the morphological and morphometric data of each taxon should be considered for the differentiation of species, a view shared by Moravec (1998).

Procamallanus spiculastriatus sp. n. has tooth-like structures on the basal ring of the buccal capsule similar to these in *P. annipetterae* although the new species has four distinct tooth-like structures, whereas these are six in *P. annipetterae* as described by Petter (1990; see also her fig. 5A) in addition the letter species is distinct. Kohn and Fernandes (1988) provide no info on the number of cephalic papillae in females; only in the holotype male *P. spiculastriatus* sp. n. can be distinguished from *P. annipetterae* by the tail shape (conical in female of the new species vs. pointed in both sexes with a marked long and narrow posterior part); number of caudal papillae (three precloacal, two adcloacal, and five postcloacal vs. four precloacal and four postcloacal), shape of oral opening (circular in *P. spiculastriatus* vs. oval in *P. annipetterae*) and morphometric parameters such as spicule length (smaller spicule 229–284 µm, larger spicule 312–355 µm vs. 150–160 µm and 180–210 µm, respectively) and length of the tail (184 µm in males and 208 µm in females vs. 336 µm in males and 281 µm in females), comparisons made based on the description of Kohn and Fernandes (1988).

Procamallanus spiculastriatus sp. n. resembles *P. peraccuratus* in the morphology of the buccal capsule, oral opening circular and presence of caudal alae of males, but differs by the presence of four internal sclerotized tooth-like structures on the basal ring, the presence of two postcloacal dorsal papillae, and the presence of spicules with alate distal end supported by sclerotized rays of *P. spiculastriatus* and those characters are absent in *P. peraccuratus* (Moravec et al. 1993; Moravec 1998). Despite sharing hosts from the same family (Cichlidae), the species differ with respect to species of host and

the biomes where they are found: *P. spiculastriatus* sp. n. is a parasite of *A. ocellatus* (biome Amazonia), whereas *P. peraccuratus* is a parasite of *G. brasiliensis* and *Au. facetus* (biome Brazilian Atlantic Forest). This finding extends the geographical distribution of the genus into the Brazilian Amazon. Additional morphometric comparisons between *P. spiculastriatus* sp. n. and the two species found in Brazil are presented in Table 1.

The six genera currently assigned to Procamallaninae were reduced to subgenera of *Procamallanus* by Moravec (1998). However, Moravec (1998) did not provide any arguments why the former genera should be demoted to a lower level. Rigby and Rigby (2014) noted the overlap in the diagnostic characteristics of the buccal capsules of these taxa. Two molecular phylogenetic studies (Černotíková et al. 2011; Choud-

Caracteres	Procamallanus spiculastriatus n. sp.				P. peraccuratus		P. annipetterae	
	Holotype	Allotype	Male	Female	Male	Female	Male	Female
Length (mm)	10.76	17	8-11	17-20	9.42-9.75	12.78-22.34	9.69	21.8
Width	147.36	166	105-147	160-173	150-170	210-400	500	720
Buccal capsule (L)a	64.93	80	57–74	78-83	72-87	87-113	131	180
Buccal capsule (W)	38.96	51	32–39	47-52	49	52–66	123	187
Mouth opening	Circular		Circular		Circular		-	
Teeth	Present	Present	Present	Present	Absent	Absent	Present	Present
Deirids	107.79	150	91–119	147-167	_	-	_	-
Nerve ring	168.83	226.66	156-188	220-233	220	230-240	298	326
Excretory pore	280.51	330	248–292	330-397	_	260-330	_	-
Muscular esophagus (L)	352.63	420	316-395	413-453	410-440	560-660	625	644
Glandular esophagus (L)	557.89	600	421-558	600–693	450-520	580-660	868	887
Ratio L/Oc and esophagus	9%	6.5%	9–12%	6–7%	10.32% ^b	7.57% ^b	16.76% ^b	7.8% ^b
Vulva (mm)	-	8	-	8-11	_	6.7-10.90	_	-
Preanal papillae (pairs)	3	-	3	_	3	-	2	-
Additional papillae (pairs)	2	_	2	_	2	-	2	-
Postanal papillae (pairs)	3 + 2DL ^a	-	3 + 2DL ^a	-	4	-	1	-
Spicule large	342.17	-	312-355	-	270-290	-	210	-
Spicule small	230.8	-	229–284	-	180-200	-	160	-
Caudal alae	Present	_	Present	_	Present	-	Absent	-
Tail	166.23	190	156-205	190-220	140	220-310	336	281
Host	Astronotus ocellatus				Geophagus brasiliensis and Australoheros facetus		Corydoras paleatus	
Site	Intestine				Intestine		Intestine	
Locality	Belém, Pará, Brazil				Vale do Rio Itaúnas, Espírito Santo, Brazil		Rio Iguaçu, Paraná, Brazil	
Biome	Amazonia				Atlantic Forest		Atlantic Forest	
Author	In this study				Pinto et al. (1976)		Kohn and Fernandes (1988)	

Table 1. Comparison of morphometric characteristics of the known South American species of *Procamallanus* with those of *Procamallanus spiculastriatus* n. sp. Except as noted for individual characteristics, all data for *P. peraccuratus*, and *P. annipetterae* were taken from the original descriptions.

^aAbbreviations: L = length; W = width, e = esophagus, DL = dorsolateral papillae; Ratio L/Oc and esophagus = Length of entire oesophagus and buccal capsule representing of body length. ^bDerived or calculated from the original publication of the species description. hury and Nadler 2016), further revealed that *Procamallanus* and *Spirocamallanus* are paraphyletic. We find that phylogenetic relationships within Procamallaninae have not been evaluated in an objective analysis using both morphological and molecular data. There is no evidence that such different taxa should be assigned to the same genus-level group (i.e., that they should be recognized as sub-genera rather than distinct genera, which is an arbitrary decision). Therefore, based on the current knowledge of the studied group we follow the concept accepting the full generic status of all recognized genus-group names within Procamallaninae. Even in the absence of a rigorous test of the monophyly of the genera, all but Procamallanus have contain species with consistent features; all known species of Procamallanus have smooth-walled buccal capsules without tooth-like structures except for *P. spiculastriatus* sp. n. and *P. annipetterae*, both of which have tooth-like structures on the basal ring. This suggests that these two species might represent a distinct genus. Currently, we do not feel confident in describing a new genus without the support of a formal phylogenetic analysis. Among the camallanid species parasitizing A. ocellatus, only Spirocamallanus inopinatus from northern Brazil (Moravec 1998; Thatcher 2006) and Camallanus sp. from Midwestern Brazil have been reported (Kohn et al. 1985; Vicente et al. 1985).

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References

Agrawal V (1966) On a new nematode, *Procamallanus muelleri* n. sp. from the stomach of a freshwater fish, *Heteropneustes fossilis*. Proceedings of the Helminthological Society of Washington 33: 204–208.

- Azevedo RKD, Abdallah VD, Luque JL (2007) Ecologia da comunidade de metazoários parasitos do apaiarí Astronotus ocellatus (Cope, 1872) (Perciformes: Cichlidae) do rio Guandu, Estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 16: 15–20.
- Bautista-Hernández CE, Violante-González J, Monks S, Pulido-Flores G (2014) Helminth communities of *Xiphophorus malinche* (Pisces: Poeciliidae), endemic freshwater fish from the Pánuco River drainage, México. Revista Mexicana de Biodiversidad 85: 838–844. https://doi.org/10.7550/rmb.40560
- Baylis HA (1923) Report on a collection of parasitic nematodes, mainly from Egypt. Part III. Camallanidae, etc. with a note on *Probstmatria* and an Appendix on Acanthocephala. Parasitology 15: 24–38. https://doi.org/10.1017/S0031182000014475
- Černotíková E, Horák A, Moravec F (2011) Phylogenetic relationships of some spirurine nematodes (Nematoda: Chromadorea: Rhabditida: Spirurina) parasitic in fishes inferred from SSU rRNA gene sequences. Folia Parasitologica 58: 135–148. https://doi.org/10.14411/fp.2011.013
- Choudhury A, Nadler SA (2016) Phylogenetic relationships of Cucullanidae (Nematoda), with observations on Seuratoidea and the monophyly of *Cucullanus, Dichelyne* and *Truttaedac-nitis*. Journal of Parasitology 102: 87–93.https://doi.org/10.1645/15-806
- Fontenele O, Nepomuceno FH (1983) Exame dos resultados da introdução do *Astronotus ocellatus* (Agassiz, 1849), em açudes do nordeste do Brasil. Boletim Técnico do DNOCS 41: 85–99.
- Froese R, Pauly D (Eds) (2017) FishBase. Version (02/2017). www.fishbase.org [Version (02/2018), last visited 7 May 2018]
- Fusco AC, Overstreet RM (1979) Two camallanid nematodes from Red Sea fishes including *Procamallanus elatensis* sp. nov. from siganids. Journal of Natural History 13: 35–40. https://doi.org/10.1080/00222937900770041
- Giese EG, Santos JN, Lanfredi RM (2009) A new species of Camallanidae from Ageneiosus ucayalensis (Pisces: Siluriformes) from Pará state, Brazil. Journal of Parasitology 95: 407– 412. https://doi.org/10.1645/GE-1680.1
- Hoffman GL (1999) Parasites of North American Freshwater Fishes. Cornell University Press, New York, 539 pp.
- Jothy AA, Fernando CH (1970) A new camallanid nematode, *Malayocamallanus intermedius* gen. et. sp. nov., from a Malayan freshwater fish, *Fluta alba* (Zuiew) with a key to the genera of the subfamily Procamallaninae. Helminthologia 11: 87–91.
- Kohn A, Fernandes BMM (1988) Procamallanus annipetterae nom. nov. for Procamallanus petterae Kohn & Fernandes, 1988 preoccupied by Procamallanus (Procamallanus) petterae Moravec & Sey, 1988. Memórias do Instituto Oswaldo Cruz 83: 535. https://doi. org/10.1590/S0074-02761988000400025
- Kohn A, Fernandes BMM, Pipolo HV, Godoy MP (1988) Helmintos parasitos de peixes das Usinas Hidrelétricas Eletrosul (Brasil). II. Reservatórios de Salto Osório e de Salto Santiago. Bacia do Rio Iguaçu. Memórias do Instituto Oswaldo Cruz 83: 299–303. https:// doi.org/10.1590/S0074-02761988000300006
- Kohn A, Fernandes BMM, Macedo B, Abramson B (1985) Helminths parasites of freshwater fishes from Pirassununga, SP, Brazil. Memórias do Instituto Oswaldo Cruz 80: 327–336. https://doi.org/10.1590/S0074-02761985000300009

- Kulkarni RB (1935) A second species of *Procamallanus* Baylis, 1923, from India. Proceedings of the Indian Academy of Sciences 2: 29–32.
- Kullander SO (2003) Cichlidae (Cichlids). In: Reis RE, Kullander SO, Ferraris Jr CJ (Eds) Checklist of the freshwater fishes of South and Central America. EDIPUCRS, Porto Alegre.
- Malta JCdO (1984) Os peixes de um lago de várzea da Amazônia Central (Lago Janauacá, Rio Solimões) e suas relações com os crustáceos ectoparasitas (Branchiura: Argulidae). Acta Amazonica 14: 355–372. https://dx.doi.org/10.1590/1809-43921984143372
- Moravec F, Van As LL (2015) Procamallanus (Procamallanus) spp. (Nematoda: Camallanidae) in fishes of the Okavango river, Botswana, including the description of P. (P.) pseudolaeviconchus n. sp. parasitic in Clarias spp. (Clariidae) from Botswana and Egypt. Systematic Parasitology 90: 137–149. https://doi.org/10.1007/s11230-014-9541-0
- Moravec F, Justine JL, Würtz J, Taraschewski H, Sasal P (2006) A new species of *Procamallanus* (Nematoda: Camallanidae) from pacific eels (*Anguilla* spp.). Journal of Parasitology 92: 130–137. https://doi.org/10.1645/GE-3509.1
- Moravec F (1998) Nematodes of freshwater fishes of the Neotropical region. Academia, Prague, 464 pp.
- Moravec F, Thatcher VE (1997) Procamallanus (Denticamallanus subgen. n.) dentatus n. sp. (Nematoda: Camallanidae) from the characid fish, Bryconops alburnoides, in the Brazilian Amazon. Parasite 4: 239–243. https://doi.org/10.1051/parasite/1997043239
- Moravec F, Scholz T (1991) Observations on some nematodes parasitic in freshwater fishes in Laos. Folia Parasitologica 38: 163–178.
- Moravec F, Kohn A, Fernandes BMM (1993) Nematode parasites of fishes of the Paraná River, Brazil. Part 3. Camallanoidea and Dracunculoidea. Folia Parasitologica 40: 211–229
- Neves LR, Pereira FB, Tavares-Dias M, Luque JL (2013) seasonal influence on the parasite fauna of a wild population of *Astronotus ocellatus* (Perciformes: Cichlidae) from the Brazilian Amazon. Journal of Parasitology 99: 718–721. https://doi.org/10.1645/12-84.1
- Olsen LS (1952) Some nematodes parasitic in marine fishes. Publications of the Institute of Marine Science, University of Texas. 2: 175–215.
- Petter AJ (1990) Nématodes de poissons du Paraguay. VI. Description de deux nouvelles espèces du genre Spirocamallanus et compléments à la description de Procamallanus annipetterae Kohn & Fernandes, 1988. Revue Suisse de Zoologie 97: 327–338. https://doi. org/10.5962/bhl.part.82069
- Petter AJ, Dlouhy C (1985) Nématodes de poissons du Paraguay. III Camallanina. Description d'une espèce et d'une sous-espèce nouvelles de la famille des Guyanemidae. Revue Suisse de Zoologie 92: 165–175. https://doi.org/10.5962/bhl.part.81607
- Pinto RM, Fábio SPde, Noronha D, Rolas FJT (1976) Novas considerações morfológicas e sistemáticas sobre os *Procamallanus* brasileiros (Nematoda, Camallanoidea). Memórias do Instituto Oswald Cruz 74: 77–84. https://doi.org/10.1590/S0074-02761976000100008
- Pritchard MH, Kruse GOW (1982) The collection and preservation of animal parasites. University of Nebraska Press, Lincoln, Nebraska, 141 pp.
- Railliet A, Henry A (1915) Sur les nématodes du genre *Camallanus* Raill. et Henry, 1915 (*Cucullanus* auct., non Mueller, 1777). Bulletin de la Societe de Pathologie Exotique 8: 446–452.

- Rigby MC, Rigby E (2014) 7.20 Order Camallanida: Superfamilies Anguillicoloidea and Camallanoidea. In: Schmidt-Rhaesa A (Ed.) Handbook of Zoology: Gastrotricha, Cycloneuralia and Gnathifera: Vol. 2, Nematoda. Walter De Gruyter, Berlin, 637–659.
- Thatcher VE (2006) Amazon fish parasites. Pensoft Publishers, Sofia, 508 pp.
- Vicente JJ, de Oliveira Rodrigues H, Correa Gomes D (1985) Nematóides do Brasil. I.: Nematóides de peixes. Atas da Sociedade de Biologia do Rio de Janeiro 25: 1–79.
- Wedl K (1861) Zur Helminthenfauna Aegyptens. 2. Abtheilung, III. Nematoda. Sitzungsberichte der Kaiserlichen Akademie derWissenschaften, Mathematisch-Naturwissenschaftlichen Classe 44: 463–482.
- Yamaguti S (1955) Parasitic worms mainly from Celebes. Part 9. Nematodes of fishes. Acta Medicinae Okayama 9: 122–133.
- Yeh L-S (1960) On a reconstruction of the genus *Camallanus* Railliet & Henry, 1915. Journal of Helminthology 34: 117–124. https://doi.org/10.1017/S0022149X00020435