

# Confirmation of the valid specific status of *Dolichovespula kuami* Kim & Yoon, 1996 (Hymenoptera, Vespidae) based on molecular and morphological evidence

Chang-Jun Kim<sup>1</sup>, Jiang-Li Tan<sup>2</sup>, Jeong Kyu Kim<sup>3</sup>, Moon Bo Choi<sup>4,5</sup>

<sup>1</sup> Division of Gardens and Education, Korea National Arboretum, Pocheon, 11186, Republic of Korea

<sup>2</sup> Key Laboratory for Animal Conservation / Key Laboratory of Resource Biology and Biotechnology in Western China, College of Life Sciences, Northwest University, Xi'an, Shaanxi 710069, China

<sup>3</sup> Department of Bio Environment Health, Dongnam Health University, Suwon, 16328, Republic of Korea

<sup>4</sup> Institute of Plant Medicine, Kyungpook National University, Daegu, 41566, Republic of Korea

<sup>5</sup> Department of R&D, Wild Beei, Chilgok, 39864, Republic of Korea

Corresponding authors: Moon Bo Choi ([kosinchoi@hanmail.net](mailto:kosinchoi@hanmail.net)); Jeong Kyu Kim ([hymjkk@dongnam.ac.kr](mailto:hymjkk@dongnam.ac.kr))

## Abstract

The taxonomic validity of *Dolichovespula kuami*, especially in relation to *D. flora*, has been the subject of a long-term debate. Herein, the valid specific status of the former was supported through an integrated analysis of morphological characters and DNA barcodes. The pronotal rugae and male genitalia of the two species are different, and partial mitochondrial genes (cytochrome oxidase subunit I, COI) indicate that they form significantly distinct lineages. The hitherto unknown male of *D. kuami* is described for the first time, and a brief discussion of the *D. maculata* species group is provided.

**Key words:** Description, DNA barcoding, *Dolichovespula flora*, male, mt-COI, taxonomy, Vespidae, wasp



Academic editor: Andreas Köhler

Received: 28 July 2023

Accepted: 15 February 2024

Published: 22 March 2024

ZooBank: <https://zoobank.org/0F3402DB-DD2D-4E62-9CB4-2C1E396025DB>

**Citation:** Kim C-J, Tan J-L, Kim JK, Choi MB (2024) Confirmation of the valid specific status of *Dolichovespula kuami* Kim & Yoon, 1996 (Hymenoptera, Vespidae) based on molecular and morphological evidence. ZooKeys 1196: 111–119. <https://doi.org/10.3897/zookeys.1196.110224>

**Copyright:** © Chang-Jun Kim et al.

This is an open access article distributed under terms of the Creative Commons Attribution License ([Attribution 4.0 International – CC BY 4.0](https://creativecommons.org/licenses/by/4.0/)).

## Introduction

To date, 19 species in the genus *Dolichovespula* Rohwer, 1916 (Hymenoptera: Vespidae) have been described from the Palearctic, Nearctic and Oriental regions (Archer 2012; Tan et al. 2014; Daglio 2020; Wang et al. 2022). Among these, the *Dolichovespula maculata* group includes a total of four species (*D. flora* Archer, 1987, *D. maculata* (Linnaeus, 1763), *D. media* (Retzius, 1783) and *D. kuami* Kim & Yoon, 1996), the members of which are characterized by the presence of pronotal striae (or furrows), the structure of the male aedeagus, and a strongly notched seventh gastral sternum (Archer 2012). Within this group, the taxonomic validity of *D. kuami* with respect to *D. flora* remains a matter of contention (Archer 1999, 2006, 2012). Whereas Tan et al. (2014) considered these to be conspecific taxa, Kim (2011) has presented evidence to indicate that *D. kuami* is a valid discrete species. This discrepancy can be attributed, at least in part, to a lack of information regarding male characteristics and differences in interpreting the related morphological variations.

In 2014, however, Tan et al. described the males of *D. flora* after discovering their nests. Recently, two of the authors of this study (JJK and MBC) identified two young and mature nests of *D. kuami* in Korea and obtained specimens of all relevant castes in 2018 and 2019, respectively.

In an attempt to resolve the longstanding debate surrounding the taxonomic validity of *D. kuami* and *D. flora*, in this study, we performed further morphological comparisons and DNA barcode analyses (using the partial mt-COI gene). We also thoroughly re-examined the orientation of the pronotal rugae and the structure of the male genitalia, and compared DNA barcodes of the two species to assess their genetic limits. A description of the previously unrecorded male of *D. kuami* is also provided.

## Materials and methods

### Morphological terminology

The terminology used in this study follows that described by Archer (1987, 1999).

### Illustrations

The images were captured using a Leica DFC 495 camera mounted on a Leica M205A stereozoom microscope (Leica Microsystems, Solms, Germany) and acquired by using LAS v.4.1.0 (Leica Microsystems, Switzerland). In addition, to observe the pronotal rugae of *D. kuami* and *D. flora*, scanning electron microscopy (SEM) images of specimens selected from each species were obtained. Subsequently, the head, metasoma, wings and legs were removed and only the mesosoma was retained. The separated mesosomes were immersed in 10% NaOCl to remove excess tissue. The detached mesosoma was washed with distilled water using a soft brush to remove the remaining tissues. After cleaning, the samples were dried and coated with gold ion particles for SEM observation (Hitachi SU8220 & SU8230, Tokyo, Japan). Image plates were prepared using Adobe Photoshop CS6 (Adobe Systems Inc., San Jose, CA, USA).

### DNA extraction and amplification

Genomic DNA was extracted from the hind legs of dried or 100% alcohol-preserved specimens using the DNeasy Blood and Tissue Kit (Qiagen) after incubating for 24 h at 56 °C in lysis buffer and Proteinase K, as per the manufacturer's instructions. A total of 26 specimens of *D. kuami*, eight specimens of *D. media*, and two specimens of *D. flora* were sequenced. The cytochrome c oxidase I barcoding region was amplified using the primer pair LepF1 and LepR1 (Hebert et al. 2004). PCR was performed using Solg™ 2X Taq PCR Pre-Mix (Solgent). We prepared 30 µL of reaction mixture containing 15 µL of PCR Pre-Mix, 11 µL of nuclease-free dH<sub>2</sub>O, 2 µL DNA template, and 1 µL of 10 pmol of each primer. A typical PCR program started with 4 min of initial denaturation at 94 °C, followed by 40 cycles of 30 s at 94 °C, 45 s of annealing at 45 °C, and 45 s of elongation at 72 °C, ending with a 6-min period of final elongation at 72 °C.

## Molecular phylogenetic analyses

Sequence reads were edited and assembled using Geneious 11 (Biomatters, Auckland, New Zealand). The optimal model (GTR+I+G) for each partition was selected using the Akaike information criterion in jModelTest (Posada 2008). The tree was constructed using the maximum-likelihood method with RAxML v.8.1.2 (Stamatakis 2014), with 1000 bootstrap replications. Successful sequences were submitted to the NCBI GenBank (accession numbers presented in Table 1).

In addition, for phylogenetic analysis, other *Dolichovespula* species and outgroups, excluding the *D. maculata* group (*D. flora*, *D. maculata*, *D. media* and *D. kuami*), included were as follows: KU874880 (*Dolichovespula norvegica*), KM568773 (*D. alpicola*), MG374965 (*D. norvegicoides*), KJ147234 (*D. saxonica*), KJ147233 (*D. pacifica*), KJ147230 (*D. arenaria*), KM567260 (*D. adulterina*), KJ147235 (*D. sylvestris*), KJ147244 (*Vespa crabro*) and LT977378 (*Formica fusca*) (Table 1). The latter two species, from Vespidae and Formicidae, respectively, were included to test the monophyly of the family and root the tree, respectively.

## Specimens

Thirty-five specimens were used in this study to review the taxonomic positions of the focus species, *D. kuami* (25) from the Korea National Arboretum (Pocheon, Republic of Korea) and *D. flora* (2) from Northwest University (Xi'an, China), and *D. media* (8).

## Results

### Comparison of the pronotal rugae and carina of *D. kuami* and *D. flora*

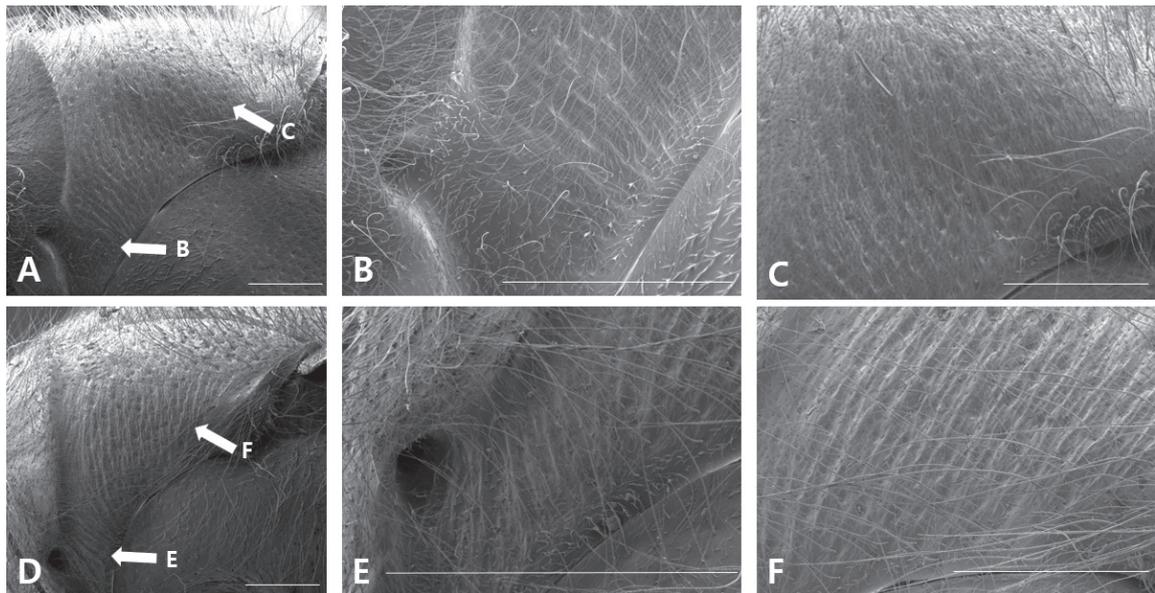
The pronotum rugae of *D. kuami* were generally very dull and faint (Fig. 1A), whereas those of *D. flora* were relatively more distinct (Fig. 1D). We found that *D. kuami* has faint longitudinal rugae on the pronotal lateral face next to the pronotal pit (Fig. 1B), and the remaining posterior area has fine rugae running vertically (Fig. 1C). On the other hand, *D. flora* has distinct longitudinal rugae (Fig. 1E) that run downward, except in the upper pronotal area (Fig. 1F). Thus, there is a clear difference between the pronotal rugae of these two species. Additionally, the pronotum carina was sharper in *D. kuami* than in *D. flora*.

### Comparison of the external genitalic features of male *D. kuami* and *D. flora*

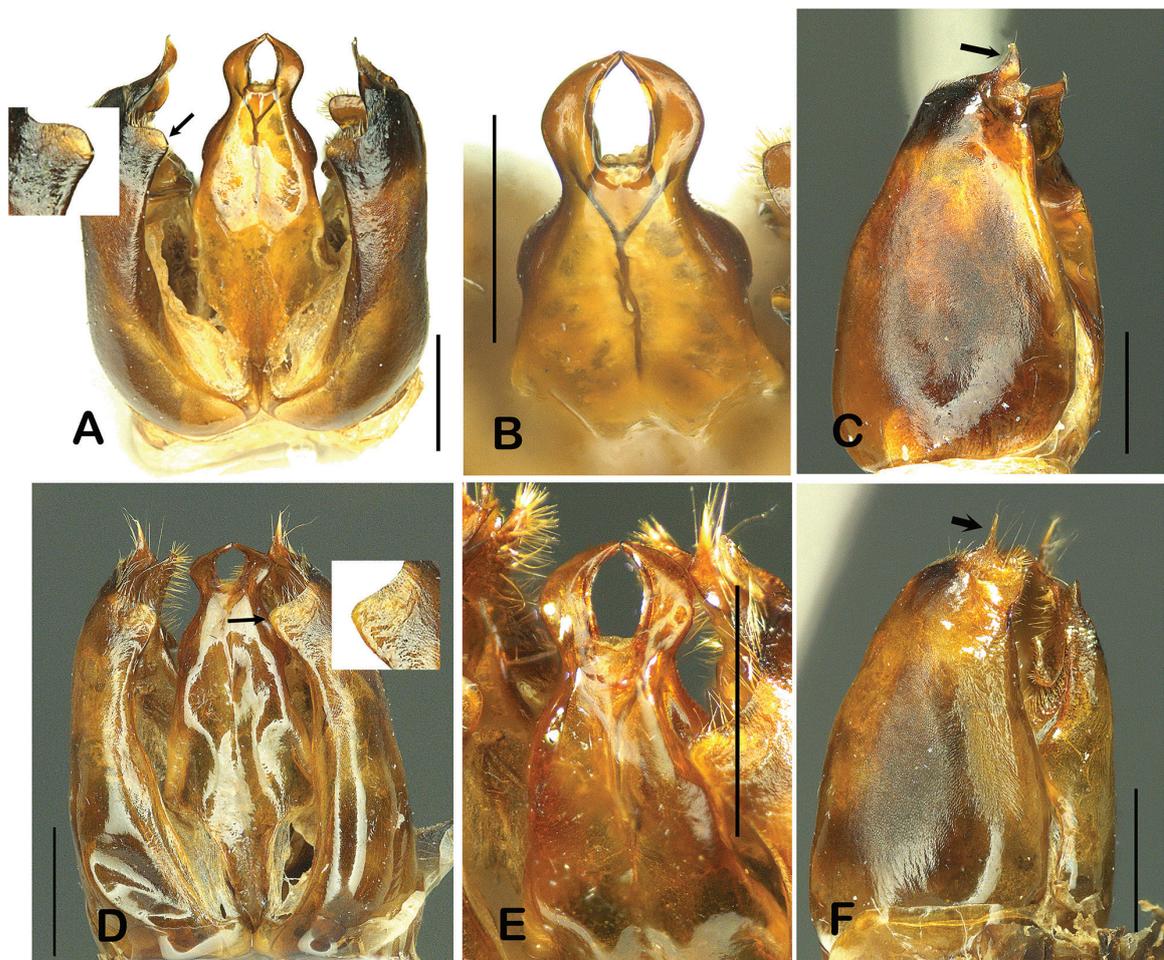
Genitalia (Fig. 2A–F). The external features of the genitalia of *D. kuami* and *D. flora* are very similar (Fig. 2A, D; also refer to Tan et al. 2014 for *D. flora*). However, *D. kuami* had a triangular parameral spine (Fig. 2C, arrow), whereas that of *D. flora* is short and slender (Fig. 2F, arrow). In addition, *D. kuami* has a truncated dorsal terminal process (Fig. 2A, arrow), whereas it is somewhat edged or shortly rounded in *D. flora* (Fig. 2D, arrow).

**Table 1.** Specimens, vouchers and GenBank accession information of *Dolichovespula* species included in the molecular phylogenetic analysis.

Species	Specimens	Vouchers	GenBank Accession #
1. <i>Dolichovespula kuami</i> Kim & Yoon, 1996	Korea: Gyeonggi-do, Pocheon-si	20Ves0603	<a href="#">OR029465</a>
	Korea: Gyeonggi-do, Pocheon-si	20Ves0604	<a href="#">OR029466</a>
	Korea: Gyeonggi-do, Yangpyeong-si	20Ves0605	<a href="#">OR029467</a>
	Korea: Gangwon-do, Hwacheon-gun	20Ves0606	<a href="#">OR029468</a>
	Korea: Gangwon-do, Hwacheon-gun	20Ves0607	<a href="#">OR029469</a>
	Korea: Gyeonggi-do, Cheorwon-gun	20Ves0608	<a href="#">OR029470</a>
	Korea: Gyeonggi-do, Pocheon-si	20Ves0609	<a href="#">OR029471</a>
	Korea: Gangwon-do, Hwacheon-gun	20Ves0610	<a href="#">OR029472</a>
	Korea: Gyeonggi-do, Namyangju-si	20Ves0611	<a href="#">OR029473</a>
	Korea: Gyeonggi-do, Yeosu-gun	20Ves0613	<a href="#">OR029474</a>
	Korea: Gyeonggi-do, Namyangju-si	20Ves0614	<a href="#">OR029475</a>
	Korea: Gyeonggi-do, Yangju-si	20Ves0615	<a href="#">OR029476</a>
	Korea: Gangwon-do, Hwacheon-gun	20Ves0616	<a href="#">OR029477</a>
	Korea: Gyeonggi-do, Yangpyeong-si	20Ves0617	<a href="#">OR029478</a>
	Korea: Gyeonggi-do, Yangpyeong-si	20Ves0619	<a href="#">OR029479</a>
	Korea: Gyeonggi-do, Pocheon-si	20Ves0621	<a href="#">OR029480</a>
	Korea: Gyeonggi-do, Pocheon-si	20Ves0622	<a href="#">OR029481</a>
	Korea: Gyeonggi-do, Paju-si	20Ves0623	<a href="#">OR029482</a>
	Korea: Gyeonggi-do, Yangpyeong-si	20Ves0624	<a href="#">OR029483</a>
	Korea: Gangwon-do, Yanggu-gun	20Ves0627	<a href="#">OR029484</a>
	Korea: Gyeonggi-do, Yeosu-gun	20Ves0628	<a href="#">OR029485</a>
	Korea: Gangwon-do, Yanggu-gun	20Ves0630	<a href="#">OR029486</a>
	Korea: Gyeonggi-do, Pocheon-si	20Ves0631	<a href="#">OR029487</a>
Korea: Gyeonggi-do, Pocheon-si	20Ves0632	<a href="#">OR029488</a>	
Korea: Gyeonggi-do, Pocheon-si	20Ves0633	<a href="#">OR029489</a>	
Korea: Gyeonggi-do, Pocheon-si	20Ves0635	<a href="#">OR029490</a>	
2. <i>D. media</i> (Retzius, 1783)	Korea: Gangwon-do, Yanggu-gun	20Ves0594	<a href="#">OR029457</a>
	Korea: Gangwon-do, Goseong-gun	20Ves0595	<a href="#">OR029458</a>
	Korea: Gangwon-do, Yanggu-gun	20Ves0596	<a href="#">OR029459</a>
	Korea: Gangwon-do, Goseong-gun	20Ves0597	<a href="#">OR029460</a>
	Korea: Gangwon-do, Hwacheon-gun	20Ves0598	<a href="#">OR029461</a>
	Korea: Gangwon-do, Yanggu-gun	20Ves0599	<a href="#">OR029462</a>
	Korea: Gangwon-do, Yanggu-gun	20Ves0600	<a href="#">OR029463</a>
	Korea: Gangwon-do, Goseong-gun	20Ves0601	<a href="#">OR029464</a>
3. <i>D. flora</i> Archer, 1987	China: Shaanxi, Huangbaiyuan	20Ves0643	<a href="#">OR029491</a>
	China: Shaanxi, Huaxian	China004	<a href="#">OR029492</a>
4. <i>D. maculata</i> (Linnaeus, 1763)	GenBank search	GenBank	<a href="#">KU874876</a>
	GenBank search	GenBank	<a href="#">KJ147231</a>
5. <i>D. norvegica</i> (Fabricius, 1781)	GenBank search	GenBank	<a href="#">KU874880</a>
6. <i>D. alpicola</i> (Wagner, 1978)	GenBank search	GenBank	<a href="#">KM568773</a>
7. <i>D. norvegicoides</i> (Sladen, 1918)	GenBank search	GenBank	<a href="#">MG374965</a>
8. <i>D. saxonica</i> (Fabricius, 1793)	GenBank search	GenBank	<a href="#">KJ147234</a>
9. <i>D. pacifica</i> (Birula, 1930)	GenBank search	GenBank	<a href="#">KJ147233</a>
10. <i>D. arenaria</i> (Fabricius, 1775)	GenBank search	GenBank	<a href="#">KJ147230</a>
11. <i>D. adulterina</i> (Buysson, 1905)	GenBank search	GenBank	<a href="#">KM567260</a>
12. <i>D. sylvestris</i> (Scopoli, 1763)	GenBank search	GenBank	<a href="#">KJ147235</a>
13. <i>Vespa crabro</i> Linnaeus, 1758	GenBank search	GenBank	<a href="#">KJ147244</a>
14. <i>Formica fusca</i> Linnaeus, 1758	GenBank search	GenBank	<a href="#">LT977378</a>



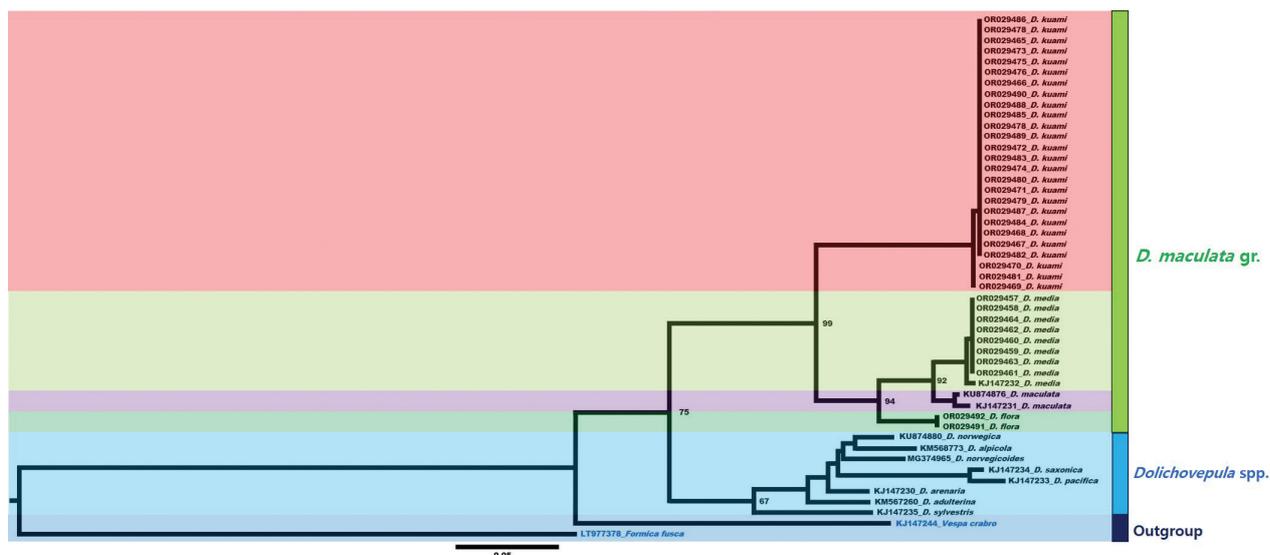
**Figure 1.** Comparison between the pronotal rugae and carinas of *D. kuami* and *D. flora*. Pronotum of *D. kuami* (A) and *D. flora* (D); rugae and carinas in the pronotal lateral part to the pronotal pit of *D. kuami* (B) and *D. flora* (E); rugae on the pronotum of *D. kuami* (C) and *D. flora* (F). Arrows in figures A, D indicate the enlarged parts in B, C, E, F. Scale bars: 0.5 mm.



**Figure 2.** Genitalia of *Dolichovespula kuami* (A–C) and *D. flora* (D–F) A genitalic capsules, in dorsal view (truncated dorsal terminal process, arrow) B aedeagus C gonostipes and triangular parameral spine (arrow) D genitalic capsules, in dorsal view (shortly rounded terminal process, arrow) E aedeagal tip F gonostipes and slender parameral spine (arrow). Scale bars: 1 mm.

### DNA barcoding

Phylogenetically, *D. maculata* clearly clustered with other *Dolichovespula* spp. (Fig. 3). In the *D. maculata* group, *D. flora* was more closely related to *D. maculata* and *D. media*, whereas *D. kuami* clustered as a sister species to the clade that included these three species (Fig. 3). Thus, the two species *D. flora* and *D. kuami* stat. rev. were clearly separated into well-supported clusters, and the genetic distance between them was relatively high (average DNA barcode distance: 0.0996), suggesting that they represent two biological species. These results were further supported by those of the morphological examination.



**Figure 3.** A maximum-likelihood phylogenetic tree of the successfully DNA barcoded *Dolichovespula* specimens. The numbers above the branches indicate bootstrap proportions.

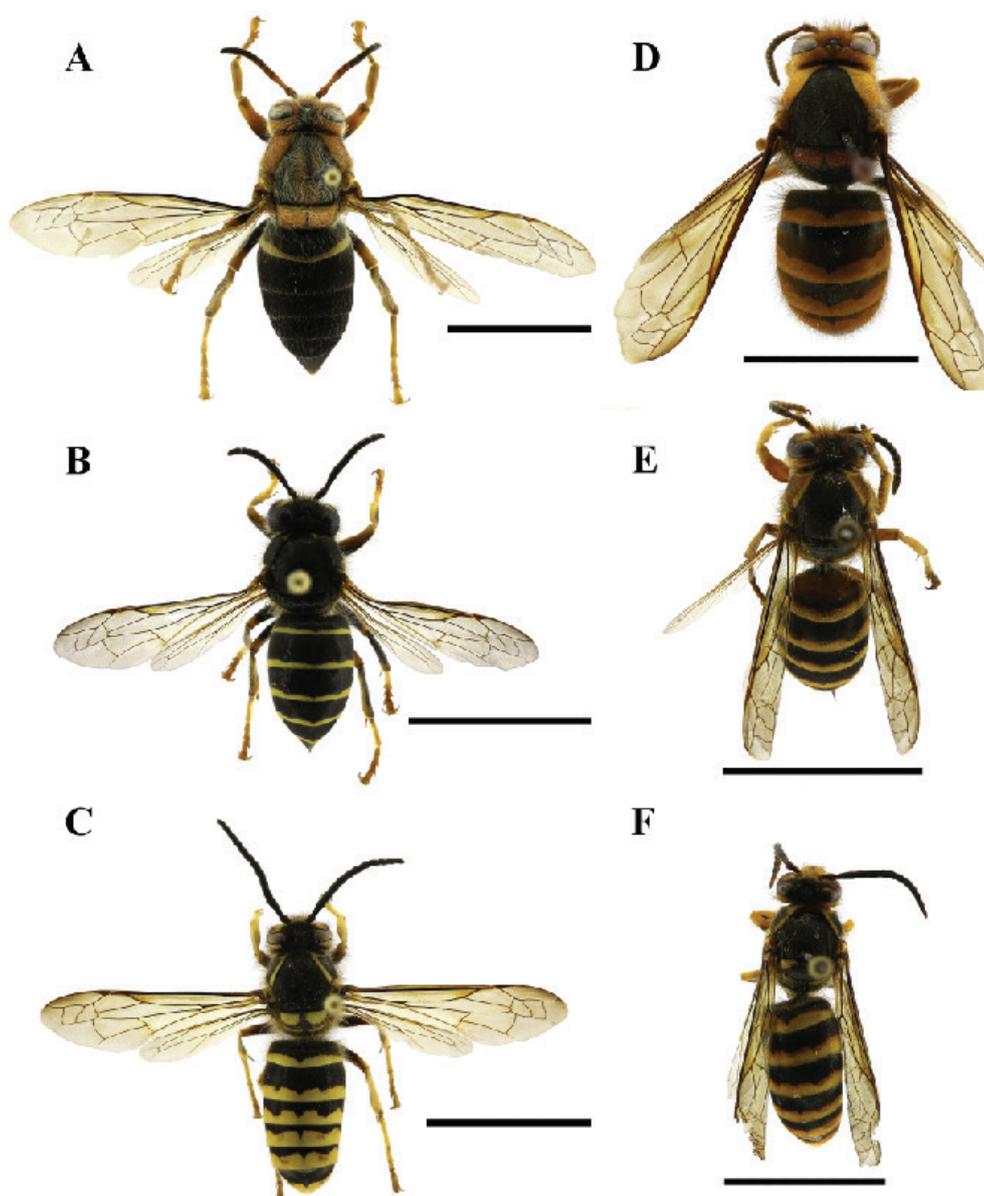
### Discussion

According to Archer (1999), *D. kuami* was treated as a conspecific to *D. flora* because the orientation of the rugae in the lateral part of the pronotum is the same, and the observed differences in body color are a type of variation. Although Kim (2011) re-described this aspect with additional specimens, Archer (2012) consistently insisted that these two species were conspecific. In fact, as shown by Kim (2011) (Fig. 2), the rugae of the pronotum were not clearly distinguished under light microscopy because they are the same color as the base color. Therefore, in this study, we attempted to obtain very clear rugae images using SEM and observed that the pronotal rugae of the two species were clearly different. Despite these morphological differences, differences in male genitalia or DNA sequences are most critically needed to provide evidence of the difference between these two species (Archer 1999; Tan et al. 2014).

*Dolichovespula kuami* and *D. flora* are uncommon species in Korea and China, respectively, and their nests and males have not been recorded for many years. Tan et al. (2014) collected males of *D. flora* and described their genitalia. We collected males for the first time in 2018, when the first nest of *D. kuami* was discovered. This discovery enabled us to compare the male genitalia of

the two species. In general, their external morphologies were relatively similar, but there were clear differences in the parameral spine and dorsal terminal processes. None of the *D. kuami* strains were conspecific to *D. flora* based on evidence of their pronotal rugae, color patterns (Fig. 4), genitalia and DNA sequences. These results support the conclusion of the long-term conspecific debate and the specific status of *D. kuami*.

Additionally, DNA barcoding is an excellent tool for accelerating species identification and complementing species delimitation (Mo et al. 2021; Zhang and Wenjun 2022; Jafari et al. 2023). In particular, COI barcode information from the genus *Polistes*, a related genus of Vespidae, provides insight into the phylogenetic relationships within the group (Schmid-Egger et al. 2017). Based on the results of this study, morphological evidence and DNA barcoding in Vespidae will provide critical evidence to resolve species delimitations in the future.



**Figure 4.** Comparison of the general habitus of *Dolichovespula kuami* and *D. flora* (**A, D** queen **B, E** worker **C, F** drone): *D. kuami* (**A–C**) *D. flora* (**D–F**). Scale bars: 1 cm.

## Additional information

### Conflict of interest

The authors have declared that no competing interests exist.

### Ethical statement

No ethical statement was reported.

### Funding

This study was supported by the Korea National Arboretum [project no. KNA1-1-20, 16-1].

### Author contributions

Conceptualization, MB Choi and JK Kim; data curation, CJ Kim, MB Choi and JL Tan; formal analysis, CJ Kim and MB Choi; funding acquisition, CJ Kim; investigation, MB Choi, JK Kim and JL Tan; methodology, CJ Kim and JK Kim; project administration, CJ Kim and MB Choi; supervision, MB Choi and JK Kim; writing—original draft, MB Choi, JK Kim and CJ Kim; writing—review and editing, JK Kim and JL Tan. All authors have read and agreed to the published version of the manuscript.

### Author ORCIDs

Chang-Jun Kim  <https://orcid.org/0000-0002-5823-8703>

Jeong Kyu Kim  <https://orcid.org/0000-0003-2175-5798>

### Data availability

All of the data that support the findings of this study are available in the main text.

## References

- Archer ME (1987) Three new species of *Dolichovespula* (Hym., Vespidae) from China. *Entomologist's Monthly Magazine* 123: 27–31.
- Archer ME (1999) Taxonomy and world distribution of the Euro-Asian species of *Dolichovespula* (Hym., Vespidae). *Entomologist's Monthly Magazine* 135: 153–160.
- Archer ME (2006) Taxonomy, distribution and nesting biology of species of the genus *Dolichovespula* (Hymenoptera, Vespidae). *Entomological Science* 9(3): 281–293. <https://doi.org/10.1111/j.1479-8298.2006.00174.x>
- Archer ME (2012) Vespine wasp of the world behavior, ecology, taxonomy of the Vespinae. Siri Scientific Press. Manchester, UK.
- Daglio A (2020) Yellowjackets of the World. LAP LAMBERT Academic Publishing. Mauritius.
- Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W (2004) Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly *Astraptes fulgerator*. *Proceedings of the National Academy of Sciences of the United States of America* 101(41): 14812–14817. <https://doi.org/10.1073/pnas.0406166101>
- Jafari S, Müller B, Rulik B, Rduch V, Peters RS (2023) Another crack in the Dark Taxa wall: A custom DNA barcoding protocol for the species-rich and common Eurytomidae (Hymenoptera, Chalcidoidea). *Biodiversity Data Journal* 11: e101998. <https://doi.org/10.3897/BDJ.11.e101998>
- Kim JK (2011) *Dolichovespula kuami* (Vespidae, Hymenoptera): Taxonomic complement with newly found specimens. *Sociobiology* 57: 11–18.

- Kim JK, Yoon IB (1996) A new species of *Dolichovespula* (Insecta: Hymenoptera: Vespidae) from Korea. *Korean Journal of Systematic Zoology* 12: 199–202.
- Mo W-h, Chen H-y, Pang H, Liu J-x (2021) DNA barcoding for molecular identification of the genus *Oxyscelio* (Hymenoptera, Scelionidae) from southern China, with descriptions of five new species. In: Lahey Z, Talamas E (Eds) *Advances in the Systematics of Platygastridae III*. *Journal of Hymenoptera Research* 87: 613–633. <https://doi.org/10.3897/jhr.87.7191>
- Posada D (2008) jModelTest: phylogenetic model averaging. *Molecular Biology & Evolution* 25(7): 1253–1256. <https://doi.org/10.1093/molbev/msn083>
- Schmid-Egger C, van Achterberg K, Neumeyer R, Morinière J, Schmidt S (2017) Revision of the West Palaearctic *Polistes* Latreille, with the descriptions of two species – an integrative approach using morphology and DNA barcodes (Hymenoptera, Vespidae). *ZooKeys* 713: 53–112. <https://doi.org/10.3897/zookeys.713.11335>
- Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30(9): 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Tan JL, Chen X, van Achterberg C (2014) Description of the male *Dolichovespula flora* Archer (Hymenoptera: Vespidae). *Entomotaxonomia* 36(1): 75–80. <https://doi.org/10.3897/zookeys.391.6606>
- Wang H, Wen Q, Wang T, Ran F, Wang M, Fan X, Wei S, Li Z, Tan J (2022) Next-Generation Sequencing of Four Mitochondrial Genomes of *Dolichovespula* (Hymenoptera: Vespidae) with a Phylogenetic Analysis and Divergence Time Estimation of Vespidae. *Animals (Basel)* 12(21): 3004. <https://doi.org/10.3390/ani12213004>
- Zhang H, Wenjun B (2022) Exploring Large-Scale Patterns of Genetic Variation in the COI Gene among Insecta: Implications for DNA Barcoding and Threshold-Based Species Delimitation Studies. *Insects* 13(5): 425. <https://doi.org/10.3390/insects13050425>