

# Unveiling the identity of *Diaurora* Cockerell, 1903 (Bivalvia, Unionidae): morphology, molecular phylogenetics, and the description of a new species

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## Abstract

The monotypic freshwater mussel genus *Diaurora* Cockerell, 1903 has long been enigmatic due to its rarity and morphological confusion with *Acuticosta*. In this study, we comprehensively redescribed *Diaurora aurea* (Heude, 1883) through a detailed analysis of shell morphology and molecular phylogenetics of recently collected specimens. Moreover, a new species, *Diaurora laeve* **sp. nov.**, was identified from the Fuyishui River, a tributary of the Zishui River in Shaoyang County, Shaoyang City, Hunan Province, China. Molecular phylogenetic analyses showed that *D. aurea* and *D. laeve* **sp. nov.** were reciprocally monophyletic and formed a clade as sister to *Schistodesmus*. Our study underscores the necessity of further exploring the diversity of freshwater mussels in understudied small tributaries throughout China.

**Key words:** Changjiang River, freshwater mussel, morphology, phylogenetic analysis, taxonomy



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## Introduction

Freshwater mussels, an important component of freshwater ecosystems (Liu et al. 1979; Vaughn and Hoellein 2018; Lopes-Lima et al. 2021), represent a diverse group of bivalve mollusks that are often identified by their unique shell morphology (Lopes-Lima et al. 2017; Chernyshev et al. 2020). However, reliance on morphological characteristics for taxonomy has resulted in some confusion within the group, as some species may share similar morphological characteristics and exhibit phenotypic plasticity (Huang et al. 2018b; Wu et al. 2020; Wu et al. 2022). One such example is *Diaurora* Cockerell, 1903, a monotypic genus of small-sized freshwater mussels that are endemic to the middle and lower reaches of the Changjiang River, also known as the Yangtze River (Heude 1883; Simpson 1900; Tchang and Li 1965; Tchang et al. 1965; Wu et al. 2000; Shu et al. 2009; He and Zhuang 2013; Guo 2022).

Initially, Simpson (1900) described the genus as a subgenus of *Parreysia* Conrad, 1853, based on the original description of *Unio aureus* Heude, 1883, and later renamed as *Diaurora* by Cockerell (1903). *Unio aureus* was originally collected

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and described by Heude (1883) based on the holotype USNM 472411 (designated by monotype) from Ning-kouo hien in China. However, only a few additional specimens have been reported sporadically since its original description, rendering this species little-known and enigmatic for over a century. Due to the limited availability of specimens for study and the potential confusion arising from its easily confused morphology with *Acuticosta* Simpson, 1900, the validity and systematic status of *Diaurora* have remained controversial for an extended period. It has been treated as a separate genus (Haas 1924), a subgenus of *Parreysia* Conrad, 1853 (Simpson 1900), or even a synonym of *Acuticosta* (Wu et al. 1999).

To address these taxonomic uncertainties, we analyzed the validity and systematic status of *Diaurora* using a combination of shell morphology and molecular phylogeny. Furthermore, we redescribed the enigmatic species *Diaurora aurorea* and described a new species, *Diaurora laeve* sp. nov., from the Zishui River of Hunan Province, China. Our study sheds light on the distribution, ecology, and protection of this distinct genus, and highlights the possibility of small Chinese tributaries harboring unique freshwater mussels.

## Material and methods

### Specimen collection and identification

Specimens were collected by hand net from the tributaries of the Changjiang River and the Jiulongjiang River from 2003–2022. These specimens were fixed in 95% ethanol, and some specimens were dissected for observation of the internal structure. Measurements were taken point-to-point with digital calipers recorded to the nearest 0.1 mm.

### Molecular phylogenetic analyses

Genomic DNA was extracted from somatic tissues preserved in 95% ethanol using a TIANamp Marine Animals DNA Kit (Tiangen Biotech, China). The quality and concentration of the DNA were checked on 1% agarose gel electrophoresis and NanoDrop 2000 (Thermo Scientific, USA). F-type mitochondrial COI sequences, which are considered the barcode marker, were amplified using primers LC022me2 (GGTCAACAAAYCATAARGATATTGG) and HCO700dy2 (TCAGGGTGACCAAAAAAYCA) (Walker et al. 2007). Polymerase chain reaction (PCR) amplifications of COI were performed in a final 25 µL volume mixture containing 1 µL of template DNA, 1 µL of each pair of primers, 12.5 µL of Green Taq Mix (Vazyme, China), and 9.5 µL ddH<sub>2</sub>O. Thermal cycling began with one cycle at 98 °C for 10 s, followed by 35 cycles of denaturation at 94 °C for 1 min, 50 °C for 1 min, and 72 °C for 1 min, with a final extension step at 72 °C for 7 min. The PCR products were purified and sequenced using an ABI 3730XL analyzer by Sangon Biotech (China). The accession numbers of all newly obtained sequences are given in Table 1.

Sequences were aligned using MEGA v. 6.0 (Tamura et al. 2013) and checked by eye. The genetic distance, based on the uncorrected *p*-distance model, was calculated using MEGA v. 6.0. Phylogenetic relationships were reconstructed using maximum likelihood (ML) and Bayesian inference (BI). Thirty-six reliable unionid sequences from GenBank were used for phylogenetic analysis.

**Table 1.** GenBank accession numbers of sequences used in this study. \* indicates the sequences newly obtained in this study.

Species	Locality	Accession number	References
Unioninae Rafinesque, 1820			
<i>Diaurora aurea</i> (Heude, 1883) 1	Ji'an, Jiangxi, China	<a href="#">OQ829360*</a>	This study
<i>Diaurora aurea</i> (Heude, 1883) 2	Ji'an, Jiangxi, China	<a href="#">OQ829361*</a>	
<i>Diaurora aurea</i> (Heude, 1883) 3	Ji'an, Jiangxi, China	<a href="#">OQ829362*</a>	
<i>Diaurora aurea</i> (Heude, 1883) 4	Fuzhou, Jiangxi, China	<a href="#">OQ829366*</a>	
<i>Diaurora laeve</i> Chen, Dai, Huang & Wu, sp. nov. 1	Shaoyang, Hunan, China	<a href="#">OQ829363*</a>	
<i>Diaurora laeve</i> Chen, Dai, Huang & Wu, sp. nov. 2	Shaoyang, Hunan, China	<a href="#">OQ829364*</a>	
<i>Diaurora laeve</i> Chen, Dai, Huang & Wu, sp. nov. 3	Shaoyang, Hunan, China	<a href="#">OQ829365*</a>	
<i>Acuticosta chinensis</i> (Lea, 1868)	Jiangxi, China	<a href="#">MG462921</a>	Huang et al. 2018a
<i>Schistodesmus lampreyanus</i> (Baird & Adams, 1867)	Jiangxi, China	<a href="#">MG463037</a>	
<i>Schistodesmus spinosus</i> (Simpson, 1900)	Jiangxi, China	<a href="#">MG463046</a>	
<i>Schistodesmus</i> sp.	Hunan, China	<a href="#">MG463043</a>	
<i>Unio pictorum</i> (Linnaeus, 1758)	Europe	<a href="#">KC429109</a>	Sharma et al. 2013
<i>Unio crassus</i> Philipsson, 1788	Poland	<a href="#">KY290446</a>	Burzynski et al. 2017
<i>Tchangsinia piscicula</i> (Heude, 1874)	Jiangxi, China	<a href="#">MG462977</a>	Huang et al. 2018a
<i>Cuneopsis celtiformis</i> (Heude, 1874)	Jiangxi, China	<a href="#">MG462964</a>	
<i>Cuneopsis heudei</i> (Heude, 1874)	Jiangxi, China	<a href="#">MG462970</a>	
<i>Aculamprotula fibrosa</i> (Heude, 1877)	Jiangxi, China	<a href="#">MG462909</a>	
<i>Nodularia douglasiae</i> (Gray, 1833)	China	<a href="#">KX822653</a>	Lopes-Lima et al. 2017
<i>Nodularia breviconcha</i> Lee, Kim, Bogan & Kondo, 2020	South Korea	<a href="#">MT020662</a>	Lopes-Lima et al. 2020
<i>Inversiunio yanagawensis</i> (Kondo, 1982)	Japan	<a href="#">MT020654</a>	
<i>Inversiunio reinianus</i> (Kobelt, 1879)	Japan	<a href="#">MT020657</a>	
<i>Pseudocuneopsis sichuanensis</i> Huang, Dai, Chen & Wu, 2022	Sichuan, China	<a href="#">MZ540966</a>	Wu et al. 2022
<i>Pseudocuneopsis capitata</i> (Heude, 1874)	Anhui, China	<a href="#">NC042469</a>	Wu et al. 2019
<i>Alasmidonta marginata</i> Say, 1818	USA	<a href="#">AF156502</a>	Graf and O'Foighil 2020
<i>Lasmigona compressa</i> (Lea, 1829)	USA	<a href="#">AF156503</a>	
<i>Anodonta anatina</i> (Linnaeus, 1758)	Russia	<a href="#">KX822632</a>	Lopes-Lima et al. 2017
<i>Pseudanodonta complanata</i> (Rossmässler, 1835)	Ukraine	<a href="#">KX822661</a>	
<i>Lanceolaria gladiola</i> (Heude, 1877)	Jiangxi, China	<a href="#">KY067441</a>	Unpublished
<i>Lanceolaria oxyrhyncha</i> (Martens, 1861)	Japan	<a href="#">MT020648</a>	Lopes-Lima et al. 2020
<i>Cristaria plicata</i> (Leach, 1814)	Jiangxi, China	<a href="#">MG462956</a>	Huang et al. 2018a
<i>Lepidodesma languilati</i> (Heude, 1874)	Jiangxi, China	<a href="#">MG463015</a>	
<i>Sinanodonta woodiana</i> (Lea, 1834)	China	<a href="#">KX822668</a>	Lopes-Lima et al. 2017
<i>Beringiana beringiana</i> (Middendorff, 1851)	Japan	<a href="#">MT020557</a>	Lopes-Lima et al. 2020
<i>Pletholophus tenuis</i> (Gray, 1833)	Vietnam	<a href="#">KX822658</a>	Lopes-Lima et al. 2017
<i>Anemina arcaeformis</i> (Heude, 1877)	Jiangxi, China	<a href="#">MG462936</a>	Huang et al. 2018a
<i>Amuranodonta kijaensis</i> Moskvicheva, 1973	Russia	<a href="#">MK574204</a>	Bolotov et al. 2020
Parreysiinae Henderson, 1935			
<i>Coelatura aegyptiaca</i> (Cailliaud, 1823)	Egypt	<a href="#">KJ081162</a>	Graf et al. 2014
<i>Indonaia andersoniana</i> (Nevill, 1877)	Myanmar	<a href="#">MF352275</a>	Bolotov et al. 2017
<i>Parresysia nagpoorensis</i> (Lea, 1860)	India	<a href="#">JQ861229</a>	Unpublished
Gonideinae Ortmann, 1916			
<i>Pronodularia japonensis</i> (Lea, 1859)	Japan	<a href="#">LC505454</a>	Fukata and Iigo 2020
<i>Lamprotula leaii</i> (Gray, 1833)	Jiangxi, China	<a href="#">MG462996</a>	Huang et al. 2018a

Species	Locality	Accession number	References
Ambleminae Rafinesque, 1820			
<i>Lampsilis siliquoidea</i> (Barnes, 1823)	USA	<a href="#">MH560773</a>	Unpublished
<i>Quadrula quadrula</i> (Rafinesque, 1820)	USA	<a href="#">HM230409</a>	
Margaritiferidae Henderson, 1929			
<i>Margaritifera dahurica</i> (Middendorff, 1850)	Russia	<a href="#">KJ161516</a>	Bolotov et al. 2015
<i>Gibbosula rochechouartii</i> (Heude, 1875)	Jiangxi, China	<a href="#">MG463022</a>	Huang et al. 2018a

*Margaritifera dahurica* (Middendorff, 1850) and *Gibbosula rochechouartii* (Heude, 1875) were used as the outgroup for rooting the tree. ML analyses were performed in IQ-TREE v. 1.6.12 (Minh et al. 2013) with 1000 reiterations. The most appropriate model of sequence evolution (TIM3+I+G) was selected under PartitonFinder2 v. 1.1 (Robert et al. 2017). Bayesian inference (BI) was conducted in MrBayes v. 3.2.6 (Ronquist et al. 2012). The most appropriate model of sequence evolution (GTR+I+G) was selected under ModelFinder (Subha et al. 2017). Four simultaneous runs with four independent Markov Chain Monte Carlo (MCMC) were implemented for 10 million generations, and trees were sampled every 1000 generations with a burn-in of 25%. The convergence was checked with the average standard deviation of split frequencies <0.01 and the potential scale reduction factor (PSRF) ~1.

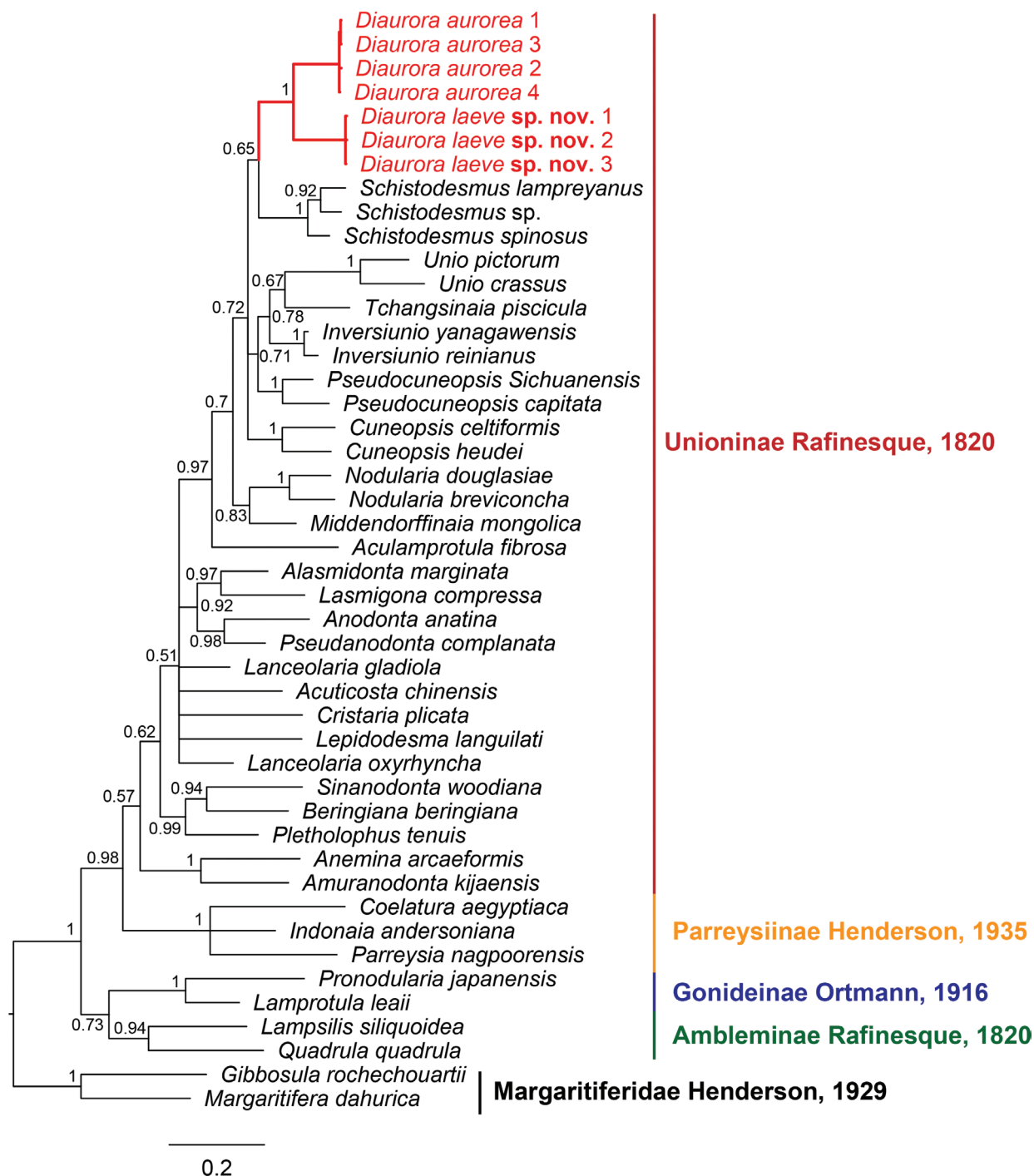
### Institutional abbreviations

<b>NCU</b>	Nanchang University (Nanchang, Jiangxi, China)
<b>USNM</b>	United States National Museum of Nature History (Washington, DC, USA)
<b>NZMC</b>	National Zoological Museum of China (Beijing, China)
<b>SMF</b>	Senckenberg Forschungsinstitut und Naturmuseum (Frankfurt am Main, Germany)
<b>YHC</b>	Collection of Mr Hao Yang (Xiamen, Fujian, China)

## Results

### Phylogenetic relationships

A total of 43 mitochondrial COI sequences from 38 unionoid species were utilized in the phylogenetic analyses (Table. 1). Phylogenetic analyses revealed ML and BI trees with largely consistent topologies (Figs 1, 2). Our results showed that *Diaurora* occupied a distinct position in the subfamily Unioninae and was the sister group of *Schistodesmus* Simpson, 1900 instead of *Acuticosta*. However, this sister relationship between *Diaurora* and *Schistodesmus* was not well-supported (bootstrap support value = 70 and Bayesian inference posterior probability support value = 0.65). Furthermore, *Diaurora* can be divided into two separate clades: one consisted of *Diaurora aurea*, and the other represents a yet-to-be-described species with distinct morphological characters. The genetic distance (uncorrected *p*-distance) between the two clades in *Diaurora* was 10.4%. Indeed, we acknowledged that deep relationships in our phylogenetic trees based on the COI dataset were not well resolved but should not overturn the main conclusion drawn here. It is necessary to further reveal the exact phylogenetic relationships using more data such as mitochondrial genomes in the future.



**Figure 1.** Bayesian inference (BI) tree inferred from COI gene sequences. Posterior probabilities are shown on the left of nodes on the tree if greater than 50%. Red font indicates the species from this study.

## Systematics

### Family Unionidae Rafinesque, 1820

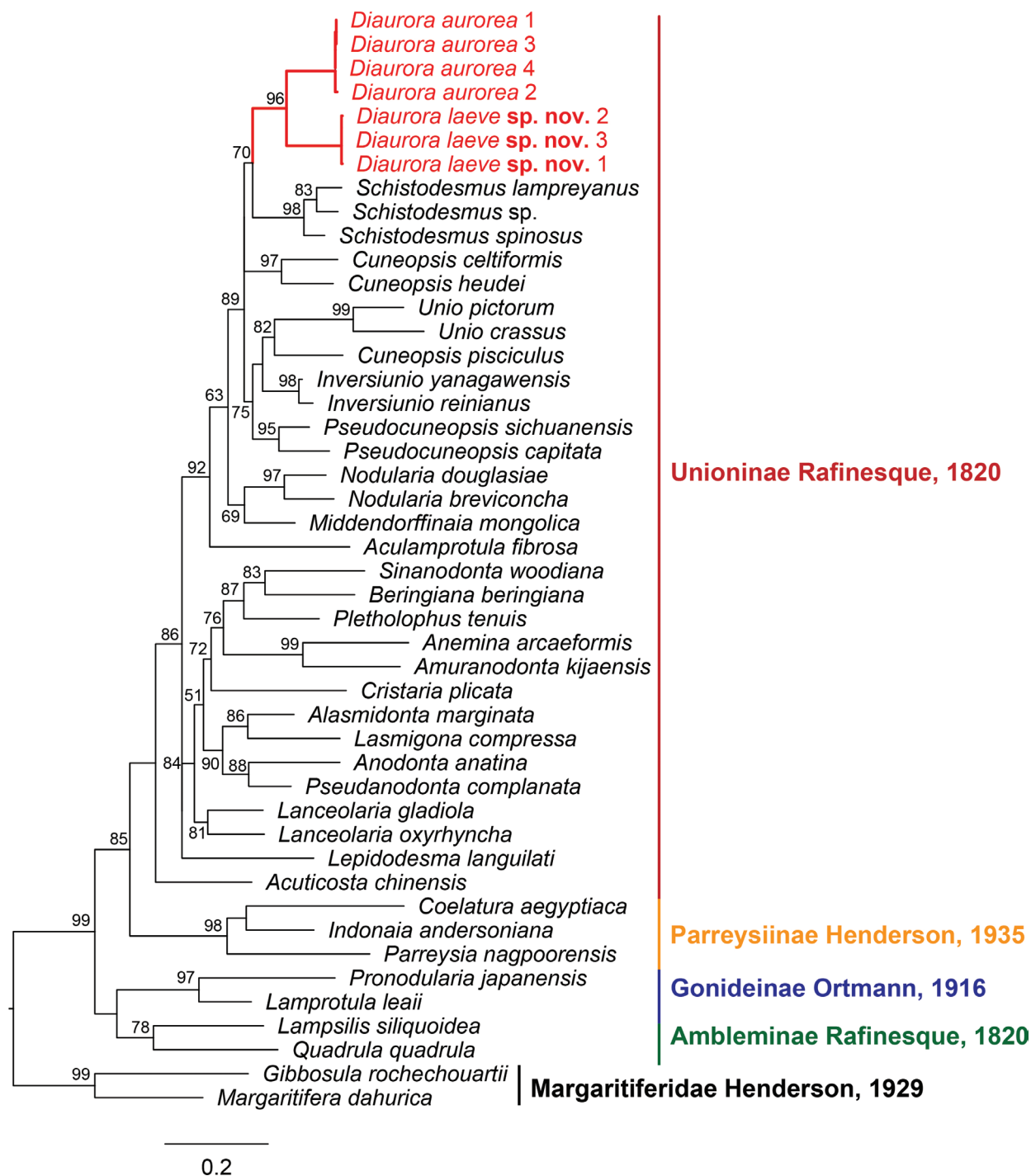
#### Subfamily Unioninae Rafinesque, 1820

#### Genus *Diaurora* Cockerell, 1903

Figs 3A, B, 4, 5

**Type species.** *Unio aureus* Heude, 1883, by original designation.





**Figure 2.** Maximum likelihood (ML) tree inferred from COI gene sequences. Bootstrap supports are shown on the left of nodes on the tree if greater than 50%. Red font indicates the species from this study.

**Diagnosis.** Shell small size, symmetric, flat, sub-glossy, triangular-ovate to reniform, orangish to brownish, with broken blackish-green rays. Zigzag sculpture present. Posterior ridge absent.

**Remarks.** *Aurora* Simpson, 1900 was originally described as a subgenus of *Parreysia* based on shell morphology. Simpson (1900) indicated uncertainty regarding the taxonomic placement of *Unio aureus* and whether it merited a generic rank. As a result, he provisionally designated it as a new subgenus. Cockerell (1903) renamed *Aurora* as *Diaurora* due to the former being a junior

homonym of *Aurora* Ragonot, 1887 (Insecta: Lepidoptera: Pyralidae) and *Aurora* Sollas, 1888 (Porifera: Demospongiae: Ancorinidae). Haas (1924) elevated *Diaurora* to an independent genus, but this viewpoint was not widely accepted. Wu et al. (1999) recombined *Parreysia aurea* as *Acuticosta aurea* based on shell morphology and marsupium type. While *Diaurora* shares some similarities in shell morphology with *Acuticosta*, it can be differentiated from *Acuticosta* by its distinct shell shape and the absence of a posterior ridge (Fig. 3).

***Diaurora aurea* (Heude, 1883)**

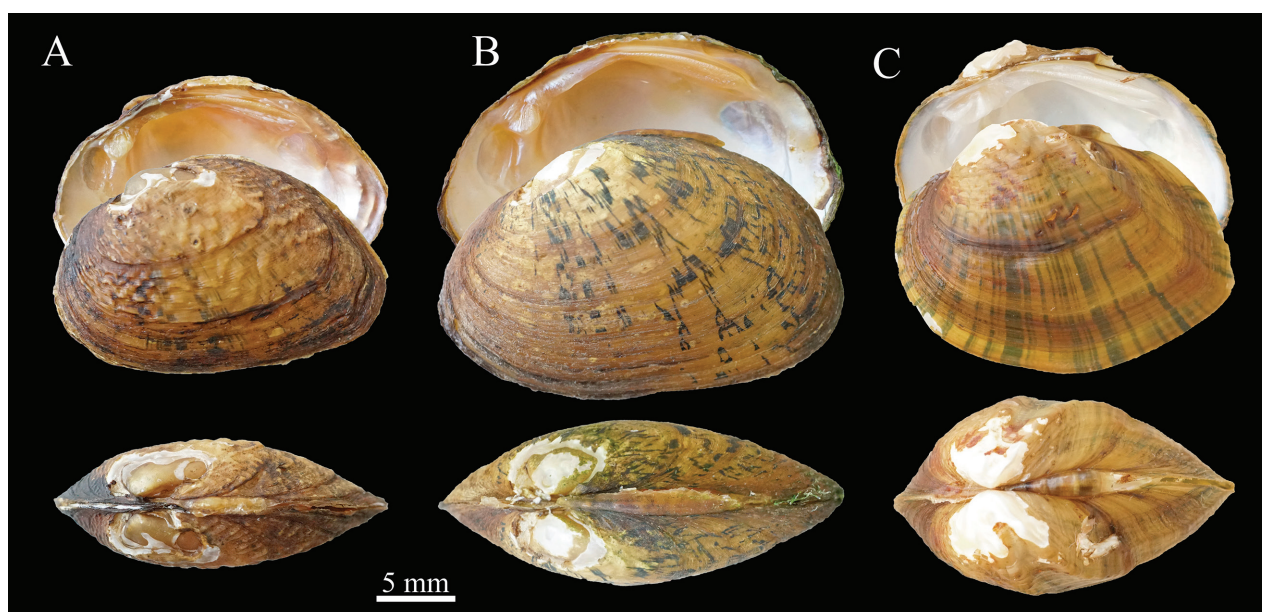
Figs 3A, 4A, 5A

**Museum material. Holotype:** USNM472411, Ning-kouo hien [宁国县], Kien-té hien [建德县], CHINA. NZMC FM00732, Yuanjiang River [沅江], Hunan Province [湖南省], CHINA. SMF24000a–b, Yütanchiao [玉潭桥], Hunan [湖南], CHINA, 28°15'7"N, 112°33'45"E.

**New material.** 22\_NCU\_XPWU\_DA01–02, Fuhe River [抚河], Fuzhou City [抚州市], Jiangxi Province [江西省], CHINA, 27°53'57"N, 116°34'03"E, collected by Zhong-Guang Chen, Yu-Ting Dai, Chen-Chen Jia & Ying-Ying Zhang in September 2022. 22\_NCU\_XPWU\_DA03–15, Ganjiang River [赣江], Jian City [吉安市], Jiangxi Province [江西省], CHINA, 27°12'39"N, 115°09'44"E, collected by Zhong-Guang Chen & Zheng-Jie Lou in November 2022. YHC0028001, Jiulong River [九龙江], Fujian Province [福建省], CHINA, collected by Hao Yang in 2003.

**Diagnosis.** Shell triangular-ovate; posterior margin obliquely arc-shaped. Periostracum with straight broken blackish-green rays. Zigzag sculpture presented on all over the shell surface.

**Description.** Shell (Figs 3A, 4A). Shell small size, symmetric, solid, moderately thick, sub-glossy, triangular-ovate. Anterior margin oval, inflated; dorsal margin curved downwards; ventral margin slightly rounded or nearly straight; posterior



**Figure 3.** Specimens of *Diaurora* and *Acuticosta chinensis* **A** *Diaurora aurea*, 22\_NCU\_XPWU\_DA03 **B** *Diaurora laeve* sp. nov., holotype: 22\_NCU\_XPWU\_DL01 **C** *Acuticosta chinensis*, 22\_NCU\_XPWU\_AC01.

margin obliquely arc-shaped. Umbo inflated, above hinge line, located at 1/3 of the dorsal margin, and often eroded. Periostracum orangish to brownish, with straight broken blackish-green rays and thin growth lines. Growth lines arranged in irregular concentric circles. Zigzag sculpture presented on all over the shell surface but weakening from umbo to edge. Hinge short. Ligament short and strong. Mantle muscle scars obvious. Anterior adductor muscle scars oval, deep, smooth in junior but rough in adult; posterior adductor muscle scars long oval, smooth. Left valve with two pseudocardinal teeth, equal height, anterior tooth small and flat, posterior tooth thick and pyramidal; anterior pseudocardinal tooth of the right valve well developed, posterior pseudocardinal tooth reduced, connected to lateral teeth. Lateral teeth of both valves long and thick. Nacre white or light orangish.

Length 32.0–41.3 mm, height 20.5–26.0 mm, width 12.4–17.2 mm.

**Soft anatomy** (Fig. 5A). Mantle off-white to light-brownish, aperture margins brown, flap margin with blackish to brown papillae. Gills light-brownish, inner gills slightly longer and wider than outer gills. Labial palps yellowish to brown (fade to grayish white in alcohol), distally pointed and irregularly fan-shaped in appearance. Visceral mass creamy white, foot orange (fade to grayish white in alcohol).

**Vernacular name.** 金黄蚌 (Pinyin: jin huang bang).

**Distribution and ecology.** Scattered in the middle and lower Changjiang River basin and the upper Jiulongjiang River, including Anhui Province, Jiangxi Province, Hunan Province and Fujian Province (Fig. 6). It mainly co-occurs with *Aculamprotula polysticta* (Heude, 1877), *Acuticosta chinensis* (Lea, 1868), *Cuneopsis heudei* (Heude, 1874), *Nodularia douglasiae* (Gray, 1833), *Lamprotula caveata* (Heude, 1877), *Lanceolaria triformis* (Heude, 1877) and *Ptychorhynchus pfisteri* (Heude, 1874) in the sandy substrates of tributaries of large rivers (Fig. 7). Besides, *D. aurea* was reported in Vietnam by Dang et al. (1980), but the specimens were not available for examination (Do et al. 2018). This record is highly dubious due to its considerable distance from the known distribution range of *D. aurea* as documented in this study. It is possible that this record was a misidentification of *Acuticosta* species.

**Remarks.** The rarity of *D. aurea* has been seriously underestimated for a long time due to the common misidentification of museum material. Upon examination of museum specimens, it has been discovered that specimen NZMC10542 in the National Zoological Museum of China and specimens 15\_NCU\_XPWU\_AA01–11 in Nanchang University were actually *Acuticosta chinensis*. Since its original description, only a handful of additional specimens of *D. aurea* have been correctly identified. It has always been shrouded in mystery as there are hardly any photographs of specimens other than the type specimen. The new specimens examined in this study indicate that it is far more widespread than previously recorded. However, the habitats of *D. aurea* are severely fragmented and the population size is very small. Therefore, it requires more attention and protection in the future.

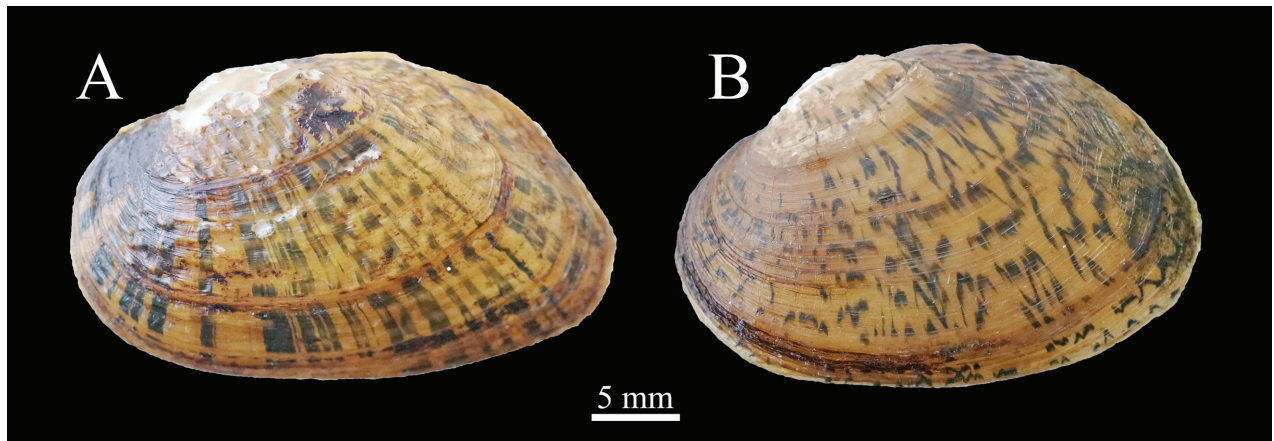
***Diaurora laeve* Chen, Dai, Huang & Wu, sp. nov.**

<https://zoobank.org/06206EEC-D7DB-464E-AC35-56F9E9174000>

Figs 3B, 4B, 5B

**Type material. Holotype:** 22\_NCU\_XPWU\_DL01, Zishui River [资水], Shaoyang County [邵阳县], Shaoyang City [邵阳市], Hunan Province [湖南省], CHINA,





**Figure 4.** Comparison of the radial pattern on the shell surface in *Diaurora* **A** *Diaurora aurea*, 22\_NCU\_XPWU\_DA04 **B** *Diaurora laeve* sp. nov., 22\_NCU\_XPWU\_DL02.

26°59'27"N, 111°16'10"E, collected by Zhong-Guang Chen & Zheng-Jie Lou in November 2022; **Paratypes:** 22\_NCU\_XPWU\_DL02–100, other information same as holotype.

**Diagnosis.** Shell reniform. Periostracum with irregular broken blackish-green rays. Zigzag sculpture only presented in umbo area.

**Description.** Shell (Figs 3B, 4B). Shell small size, symmetric, solid, moderately thick, sub-glossy, reniform. Anterior margin oval, inflated; dorsal margin curved downwards and truncate; ventral margin slightly rounded or nearly straight; posterior margin oval. Umbo inflated, above hinge line, located at 1/3 of the dorsal margin, and often eroded. Periostracum orangish to brownish, with irregular broken blackish-green rays and thin growth lines. Growth lines arranged in irregular concentric circles. Zigzag sculpture only presented in umbo area. Hinge short. Ligament short and strong. Mantle muscle scars obvious. Anterior adductor muscle scars oval, deep, smooth in junior but rough in adult; posterior adductor muscle scars long oval, smooth. Left valve with two pseudocardinal teeth, equal height, anterior tooth small and flat, posterior tooth thick and pyramidal; anterior pseudocardinal tooth of the right valve well developed, posterior pseudocardinal tooth reduced, connected to lateral teeth. Lateral teeth of both valves long and thick. Nacre light orangish.

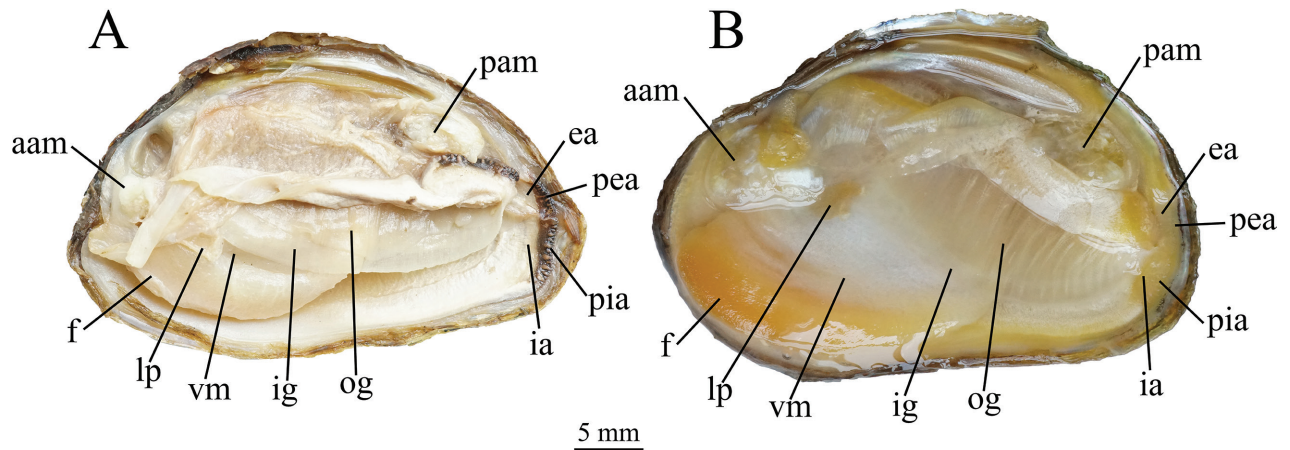
**Holotype:** length 41.8 mm, height 29.1 mm, width 17.0 mm; Paratypes: length 27.4–45.0 mm, height 17.6–30.8 mm, width 11.0–17.6 mm.

**Soft anatomy** (Fig. 5B). Mantle off-white to light-brownish, aperture margins brown, flap margin with yellowish papillae. Gills light-brownish, inner gills slightly longer and wider than outer gills. Labial palps yellowish to brown, distally pointed and irregularly fan-shaped in appearance. Visceral mass creamy white, foot orange.

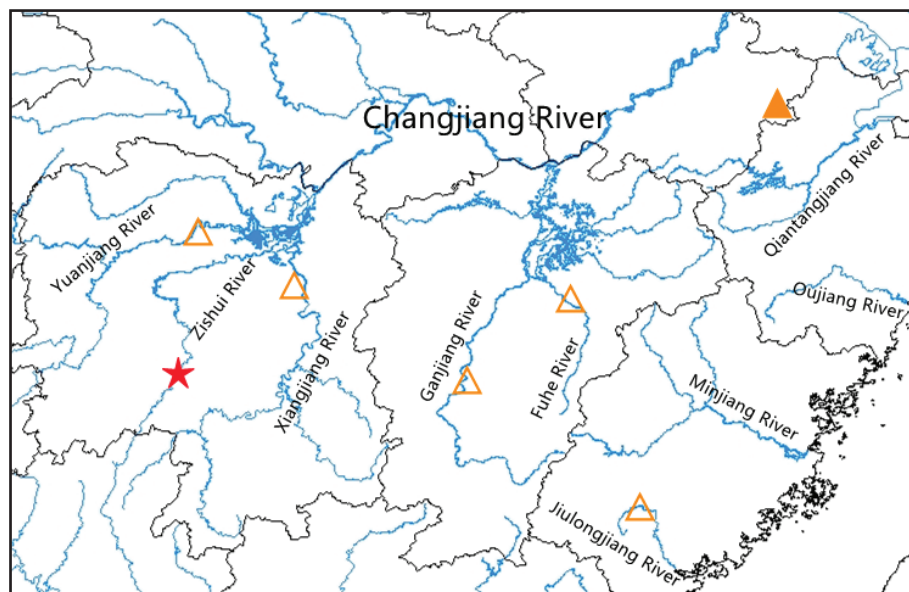
**Etymology.** The specific name *laeve* is made from the Latin *laeve* for smooth, an adjective, alluding to the smoother shell surface of this species.

**Vernacular name.** 平滑金黄蚌 (Pinyin: ping hua jin huang bang).

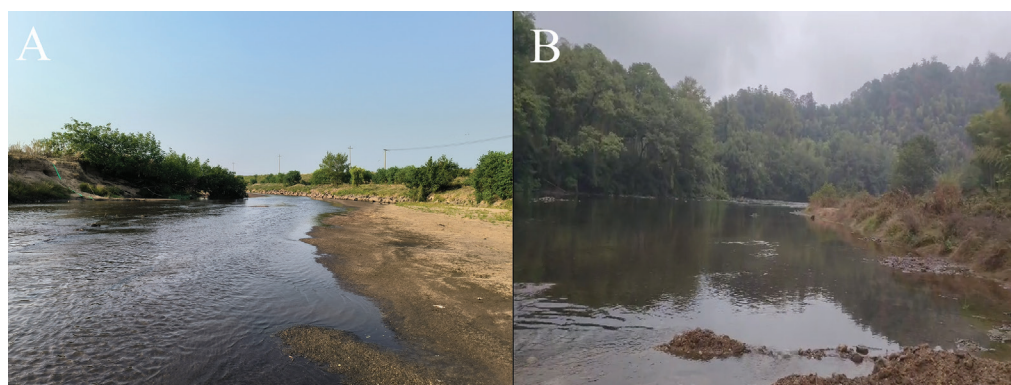
**Distribution and ecology.** *Diaurora laeve* sp. nov. is known from the type locality only (Fig. 6). It was found to occur in a pebbly substrate of the river together with *Nodularia douglasiae* and *Lanceolaria triformis* (Fig. 7). It is the dominant species in the habitat, accounting for 98% of the total density of freshwater mussels.



**Figure 5.** Soft anatomical characters of *Diaurora* **A** *Diaurora aurea*, 22\_NCU\_XPWU\_DA15 **B** *Diaurora laeve* sp. nov., 22\_NCU\_XPWU\_DL50. aam, anterior adductor muscle; pam, posterior adductor muscle; ea, excurrent aperture; ia, incurrent aperture; pia, papillae of the incurrent aperture; pea, papillae of the excurrent aperture; ig, inner gills; og, outer gills; m, mantle; lp, labial palps; vm, visceral mass; f, foot.



**Figure 6.** Distribution map of *Diaurora*. Star: type locality of *Diaurora laeve* sp. nov.; solid triangle: type locality of *Diaurora aurea*; hollow triangle: new locality of *Diaurora aurea*.



**Figure 7.** Habitat of *Diaurora*. **A** habitat of *Diaurora aurea*, Fuhe River, Fuzhou City, Jiangxi Province, China **B** habitat of *Diaurora laeve* sp. nov., Zishui River, Shaoyang City, Hunan Province, China.

**Remarks.** The placement of the new species in *Diaurora* is supported by both morphology and molecular phylogenetic analysis. *Diaurora laeve* sp. nov. can be easily distinguished from *D. aurea* by its reniform shell, smaller zigzag sculptured area, and irregular rays on the shell. The different habitat preferences of *Diaurora laeve* sp. nov. and *D. aurea* may have led to their differentiation. *Diaurora aurea* is commonly found in the middle reaches of sandy substrate tributaries, while *Diaurora laeve* sp. nov. prefers to inhabit the upper reaches of pebbly substrate tributaries. In recent years, with more in-depth investigations and field surveys, new freshwater mussel species such as *Inversidens rentianensis* Wu & Wu, 2021 and *Pseudocuneopsis sichuanensis* Huang, Dai, Chen & Wu, 2022 have been discovered in small tributaries of China (Wu et al. 2021; Wu et al. 2022). These findings suggest that the diversity of freshwater mussels in small tributaries of China remains to be fully explored, and future extensive exploration may lead to the discovery of other yet-to-be-described species.

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## Additional information

### Conflict of interest

The authors have declared that no competing interests exist.

### Ethical statement

No ethical statement was reported.

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### Author contributions

Zhong-Guang Chen, Yu-Ting Dai, Xiao-Chen Huang & Xiao-Ping Wu designed the study. Zhong-Guang Chen & Yu-Ting Dai collected the specimens. Zhong-Guang Chen & Yu-Ting Dai examined the specimens, performed morphological comparisons and conducted molecular analysis. All authors prepared the manuscript, read and approved the final version.

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### Data availability

All of the data that support the findings of this study are available in the main text.

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