

Research Article

A new species of *Achalinus* Peters, 1869 (Squamata, Xenodermidae) from Hunan Province, China

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Abstract

A new species, Achalinus hunanensis **sp. nov.**, is described from middle and western Hunan Province based on the results of molecular systematics and morphological characters. It diverges from known congeners by a significant genetic divergence (*p*-distance 3.2%-16.9% based on CO1 mitochondrial gene), and it can be distinguished from all known congeners by the following morphological characters: (1) all dorsal scales strongly keeled, 23 rows throughout the body, the outmost one strongly keeled and enlarged; (2) tail relatively short, TaL/TL 0.221 ~ 0.225; (3) maxillary teeth 23; (4) the suture between internasals 2 × as long as that between prefrontals; (5) loreal one, subrectangular, LorH/LorL 0.62 ~ 0.70; (6) supralabials 6, the 4th and 5th touch the eye; (7) the two anterior temporals in contact with eye; (8) ventrals 163–165, subcaudals 69–72, not paired. This raises the number of known species of *Achalinus* to 24.

Key words: Achalinus hunanensis sp. nov., morphology, phylogeny, divergence time, taxonomy

Introduction

The odd-scaled snake genus *Achalinus* Peters, 1869, which is widely distributed in northern Vietnam, China, and Japan, is a group of small to medium-sized, nocturnal, fossorial, and non-venomous snakes (Zhao et al. 1998; Zhao 2006). It is the most diverse genus of six known genera of the family Xenodermidae: 23 species in *Achalinus*; two species in *Fimbrios* Smith, 1921; two species in *Parafimbrios* Teynié, David, Lottier, Le, Vidal & Nguyen, 2015; one species in *Paratenodermus* Deepak, Lalronunga, Lalhmingliani, Das, Narayanan, Das & Gower, 2021; two species in *Stoliczkia* Jerdon, 1870; one species in *Xenodermus* Reinhardt, 1836 (Ha et al. 2022; Yang et al. 2022; Zhang et al. 2023). Among *Achalinus* species, more than half (14/23) have been described since 2019 (Wang et al. 2019; Ziegler et al. 2019; Li et al. 2020; Luu et al. 2022; Yang et al. 2022; Zhang et al. 2022; David et al. 2022; Jang et al. 2022; Jang et al. 2022; Jang et al. 2020; Hou et al. 2021; Huang et al. 2021; Li et al. 2021; Ha et al. 2022; Yang et al. 2022; Zhang et al. 2022; Jang et al. 2023). Besides, previous studies revealed that *A. ater* Bourret, 1937, *A. formosanus*



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Boulenger, 1908, *A. huangjietangi* Huang, Peng & Huang, 2021, *A. niger* Maki, 1931, *A. rufescens* Boulenger, 1888, and *A. spinalis* Peters, 1869 required further studied, due to their distinct morphology variation, deep intraspecific divergence, or non-monophyletic relationship (Zhao et al. 1998; Miller et al. 2020; Huang et al. 2021; Zhang et al. 2023). However, because of poor sampling, morphological and molecular information of *Achalinus* were lacking, leading to unresolved taxonomic problems and poorly known distribution patterns. Therefore, the diversity of this genus is underestimated and further research is required.

During our field work, two specimens were collected from Hunan Province, China (Fig. 1). They could be assigned to *Achalinus* by body small, slender, and cylindrical; whole body scales strongly keeled, lanceolate-shaped, and metallic luster; and preocular and postocular absent, loreal and temporals contacting the eyes directly. However, further morphological and molecular analyses revealed that these specimens comprise a separate taxon from other known species. Thus, we described them as a new species herein.

Materials and methods

Molecular phylogenetic analyses

Two specimens of the genus *Achalinus* were collected in Hunan Province, China: CIB 119039 was collected in Huaihua City, and CIB 119040 was collected in Ningxiang County, and they were deposited in Chengdu Institute of Biology (CIB) of Chinese Academy of Sciences (CAS). Genomic DNA were extracted from preserved muscle tissues of them using QIAamp DNA Mini Kit (QIAGEN, Changsheng Biotechnology Co. Ltd). A fragment of the mitochondrial cytochrome c oxidase subunit 1 (CO1) was amplified using the primer pairs dglco and dghco (Meyer et al. 2005). The polymerase chain reaction (PCR) was performed in 25 μ L of reactant with the following cycling conditions: 95 °C for 4 min; 35 cycles of denaturing at 95 °C for 30 s, annealing at 48 °C for 30 s, and extending at 72 °C for 60 s; and a final extending step of 72 °C for 10 min (Wang et al. 2019). PCR products were sequenced by Beijing Qingke New Industry Biotechnology Co., Ltd.

For phylogenetic analysis, 25 sequences (Table 1) were selected, among which 23 sequences (NO. 3–25) were obtained from National Center for Biotechnology Information (NCBI) including 22 sequences from 19 species of *Achalinus* and three sequences of *Fimbrios klossi* Smith, 1921, *Parafimbrios lao* Teynié, David, Lottier, Le, Vidal & Nguyen, 2015 and *Xenodermus javanicus* Reinhardt, 1836, which were used as outgroups (Luu et al. 2020; Ha et al. 2022).

CO1 sequences (681 bp) were input in MEGA11 (Tamura et al. 2021) and aligned by MUSCLE (Edgar 2004), and then the uncorrected pairwise distances (*p*-distance) were calculated in MEGA11. IQ-TREE 1.6.12 was used to conduct the maximum likelihood (ML) analysis (Nguyen et al. 2015) under the best-fit model TN+F+I+G4 selected by Modelfinder according to BIC (Kalyaanamoorthy et al. 2017). Ultrafast Bootstrap Approximation (UFB) node support as assessed by using 5000 ultrafast bootstrap replicates and the UFB \geq 95 was considered significantly supported (Hoang et al. 2018). The single branch tests were made using SH-like approximate likelihood ratio test (SH-aLRT) via 1000 replicates, and the SH \geq 80 was also considered supported well (Stephane et al. 2010). For Bayesian inference (BI), the best-fitting model HKY+I+G was selected by jModelTest 2.1.10



Figure 1. Distribution of *Achalinus hunanensis* sp. nov. and its sister taxon *A. ningshanensis*. Blue pentacle: the type locality of *A. ningshanensis*: Xunyangba, Ningshan County, Shaanxi Province, China. Red triangle: the type locality of *A. hunanensis* sp. nov. (CIB 119039): Hecheng District, Huaihua City, Hunan Province, China. Red circle: *A. hunanensis* sp. nov. (CIB 119040): Wazizhai, Ningxiang County, Changsha City, Hunan Province, China.

identified via BIC (Darriba et al. 2012) on CIPRES (Miller et al. 2010). The Bayesian inference analysis was conducted using MrBayes v. 3.2.1 (Ronquist et al. 2012) under HKY+I+G this model, and four chains run was calculated for 10 million generations, sampled every 1000 with the first 25% of samples discarded as burn-in, resulting in a potential scale reduction factor (PSRF) of \leq 0.005. Bayesian posterior probabilities (BI) \geq 0.95 were considered supported well.

We also estimated divergence time by BEAST v. 2.6.7 using CO1 sequences dataset (Bouckaert et al. 2019). Three calibration constraints were used: the divergence time between *Achalinus* and other Xenodermatidae: 38.6 Mya; the divergence time between *Xenodermus* and *Fimbrios* + *Parafimbrios*: 29.67

NO.	Species	Locality	Voucher	CO1 GenBank No.	References
1	A. hunanensis sp. nov.	Huaihua, Hunan, China	CIB 119039	OQ848425	This study
2	A. hunanensis sp. nov.	Ningxiang, Hunan, China	CIB 119040	OQ848426	This study
3	A. ningshanensis	Ningshan, Shaanxi, China	ANU 20220006	ON548422	Yang et al. 2022
4	A. ningshanensis	Ningshan, Shaanxi, China	ANU 20220007	ON548423	Yang et al. 2022
5	A. ater	Huaping Nature Reserve, Guangxi, China	SYS r00852	MN380334	Wang et al. 2019
6	A. dehuaensis	Dehua, Fujian, China	YBU 13013	MZ442662	Li et al. 2021
7	A. emilyae	Hoanh Bo, Quang Ninh, Vietnam	IEBR 4465	MK330857	Ziegler et al. 2019
8	A. formosanus	Taiwan, China	RN2002	KU529452	Unpublished
9	A. huangjietangi	Huangshan, Anhui, China	HSR18030	MT380191	Huang et al. 2021
10	A. juliani	Ha Lang, Cao Bang, Vietnam	IEBR A.2018.8	MK330854	Ziegler et al. 2019
11	A. meiguensis	Mianyang, Sichuan, China	GP835	MZ442641	Li et al. 2021
12	A. niger	Taiwan, China	RN0667	KU529433	Unpublished
13	A. panzhihuaensis	Yanbian, Sichuan, China	KIZ 040189	MW664862	Hou et al. 2021
14	A. pingbianensis	Honghe, Yunnan, China	YBU 18273	MT365521	Li et al. 2020
15	A. rufescens	Hongkong, China	SYS r001866	MN380339	Wang et al. 2019
16	A. spinalis	Badagong Mountains, Hunan, China	SYS r001327	MN380340	Wang et al. 2019
17	A. timi	Thuan Chau, Son La, Vietnam	IEBR A.2018.10	MK330856	Ziegler et al. 2019
18	A. tranganensis	Ninh Binh, Vietnam	VNUF R.2018.21	MW023086	Luu et al. 2020
19	A. vanhoensis	Van Ho, Son La, Vietnam	VNUF R.2019.13	ON677935	Ha et al. 2022
20	A. yangdatongi	Wenshan Nature Reserve, Yunnan, China	KIZ 034327	MW664865	Hou et al. 2021
21	A. yunkaiensis	Dawuling Forestry Station, Guangdong, China	SYS r001443	MN380329	Wang et al. 2019
22	A. zugorum	Bac Me, Ha Giang, Vietnam	IEBR 4698	MT502775	Miller et al. 2020
23	Fimbrios klossi	Quang Ngai, Vietnam	IEBR 3275	KP410744	Teynié et al. 2015
24	Parafimbrios lao	Louangphabang, Laos	MNHN 2013.1002	KP410746	Teynié et al. 2015
25	Xenodermus javanicus	Sumatera Barat, Indonesia:	_	KP410747	Teynié et al. 2015

Table 1. Localities, voucher information, GenBank numbers, and references for all samples used in this study.

Mya; and the divergence time between *Fimbrios* and *Parafimbrios*: 17.66 Mya (Kumar et al. 2022). Two independent searches of 20 million generations were conducted, sampling every 1000 iterations with 25% of the initial samples discarded as burn-in. Tracer v. 1.7.2 was used to evaluate estimate ESS for all parameters (Rambaut et al. 2018).

Morphological examination

Morphological data of known species of *Achalinus* were obtained from the two newly collected specimens, examination of museum specimens (*A. ater:* n = 1; *A. rufescens:* n = 1; *A. yunkaiensis:* n = 1) (Appendix 1) and many key references (Boulenger 1888, 1893, 1896; Denburgh 1912; Bourret 1935, 1937; Hu and Zhao 1966; Hu et al. 1973; Koshikawa 1982; Zong and Ma 1983; Ota and Toyama 1989; Zhao et al. 1998; Zhao 2006; Wang et al. 2019; Ziegler et al. 2019; Li et al. 2020; Luu et al. 2020; Miller et al. 2020; Yu et al. 2020; Hou et al. 2021; Huang et al. 2021; Li et al. 2021; Chen et al. 2022; Ha et al. 2022; Xu et al. 2023; Yang et al. 2022; Zhang et al. 2023).

Morphological descriptions followed Zhao (2006) and Yang et al. (2022). Three characters were measured to the nearest 1 mm by Deli Stainless Ruler (No.

8460): snout-vent length (SVL), tail length (TaL) and total length (TL); the other characters were measured to the nearest 0.01 mm by Deli Digital Vernier Caliper (DL91150): head length (HL), head width (HW), eye horizontal diameter (ED), loreal height (LorH), loreal length (LorL), length of the suture between internasals (LSBI), length of the suture between prefrontals (LSBP). We counted the following ratios: TaL/TL: tail length/total length, LorH/LorL: loreal height/loreal length, LSBI/LSBP: length of the suture between internasals/length of the suture between prefrontals, HL/HW: ratio head length/head width. We also directly compared the length of the sutures between internasals and prefrontals (LSBI vs. LSBP).

Scalation features and their abbreviations are as follows: loreals (Loreal), supralabials (SPL), infralabials (IFL), the number of chin shield pairs (Chins), the number of infralabials touch the first pair of chin shields (IFL-1st Chin), supraoculars (SPO), temporals (TEM), the number of anterior temporals touch the eye (aTEM-Eye) (those head bilateral scale counts were given as left/right), pre-ventral scales (PrV), ventral scales (VEN), subcaudal (SC), entire or divided of the anal (Anal), dorsal scale rows (DSR) (counted at one-head-length behind the head, at midbody, at one-head-length before the anal). We also counted the number of maxillary teeth (MT) under the microscope.

Results

Molecular systematics

All Achalinus samples cluster in a monophyletic group with high supporting values (SH 100/ UFB 100/ BI 1), and they can be divided into six clades, although the relationships among these clades are still unresolved (Fig. 2). Clade A contains A. dehuaensis Li, Wu, Xu. Zhu, Ren, Guo & Dong, 2021; clade B contains A. emilyae Ziegler, Nguyen, Pham, Nguyen, Pham, Van Schingen, Nguyen & Le, 2019, A. rufescens and A. tranganensis Luu, Ziegler, Ha, Lo, Hoang, Ngo, Le, Tran & Nguyen, 2020; clade C A. zugorum Miller, Davis, Luong, Do, Pham, Ziegler, Lee, De Queiroz, Reynolds & Nguyen, 2020, A. huangjietangi, A. panzhihuaensis Hou, Wang, Guo, Chen, Yuan & Che, 2021, A. meiguensis Hu & Zhao, 1966, A. spinalis, A yunkaiensis Wang, Li & Wang, 2019, A. niger and A. formosanus; Clade D includes A. timi Ziegler, Nguyen, Pham, Nguyen, Pham, Van Schingen, Nguyen & Le, 2019 and A. vanhoensis Van Ha, Ziegler, Sy, Le, Nguyen & Luu, 2022, and Clade E contains A. pingbianensis Li, Yu, Wu, Liao, Tang, Liu & Guo, 2020, respectively. Clade F consists five species with a significantly high nodal support (SH 100/ UFB 100/ BI 1; divergence time: 1.76 Mya, 95% highest posterior density interval (HPD): 2.473 ~ 0.988 Mya), of which, the two specimens newly collected in this work are firstly clustered together as a lineage with well support (SH 88/UFB 96/BI 0.98; divergence time: 0.21 Mya, 95% HPD: 0.534 ~ 0.003 Mya), and then clustered with A. ningshanensis Yang, Huang, Jiang, Burbrink, Gong, Yu, Zhang, Huang & Huang, 2022 (SH 98/UFB 100/BI 1; divergence time: 0.48 Mya, 95% HPD: 0.914 ~ 0.120 Mya), forming a sister group of A. yangdatongi Hou, Wang, Guo, Chen, Yuan & Che, 2021 diverged at 0.83 Mya $(95\% \text{ HPD: } 1.418 \sim 0.279 \text{ Mya})$, and then they form a sister group of the lineage which contains the A. ater and A. juliani Ziegler, Nguyen, Pham, Nguyen, Pham, Van Schingen, Nguyen & Le, 2019 (divergence time: 1.27 Mya, 95% HPD: 1.930 ~ 0.602 Mya) (Fig. 3).



Figure 2. Phylogenetic tree of the genus *Achalinus* inferred from CO1 gene fragment (681 bp) using Maximum Likelihood. The tree nodes present the supporting values: SH-like approximate likelihood ratio test, Ultrafast Bootstrap Approximation and Bayesian posterior probabilities, respectively (SH, %/UFB, %/BI) (the ones lower than 50 are displayed as "-"). *Achalinus ningshanensis* is noted in blue and *A. hunanensis* sp. nov. is noted in red.





The genetic distances range from 5.0% (*A. timi* and *A. vanhoensis*) to 18.1% (*A. dehuaensis* and *A. meiguensis*) among the known *Achalinus* species studied in this work (Table 2), of which, within the Clade F, the genetic distances among the four known species range from 5.8% (*A. ningshanensis* and *A. yangdatongi*) to 9.6% (*A. juliani* and *A. ningshanensis*), while the lineage composed by those two newly collected specimens range from 3.2% to 8.8% divergent from congeners.

The results above indicate that the Hunan samples are close to the species *A. ningshanensis* but consist independent evolution lineage.

Morphological systematics

The two specimens of the genus *Achalinus* newly collected from Hunan Province can be easily distinguished from all other known congeners (Tables 3, 4). By having 23-23-23 dorsal scale rows, the two specimens can be distinguished from these species including *A. formosanus* formosanus Boulenger, 1908 (vs. 29-27-25), *A. f. chigirai* Ota & Toyama, 1989 (vs. (25-27)-(25-27)-25), *A. meiguensis* (vs. (21-23)-(19-21)-(19-21)), *A. niger* (vs. 25-25-23), *A. panzhihuaensis* (vs. 23-23-19), *A. timi* (vs. 25-25-23), *A. tranganensis* (vs. 25-23-23), *A. vanhoensis* (vs. 25-23-23), *A. zugorum* (vs. 25-23-23). By having loreal separated from prefrontal, they are different from *A. jinggangensis* and *A. pingbianensis* (vs. loreal fused with prefrontal). By having LSBI vs. LSBP > 1, they differ from *A. dabieshanensis* Zhang, Liu, Huang, Hu, Yu, Sun, Zhang, Wen & Zhang, 2023 (vs. < 1),

	1-2	3-4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
1–2 A. hunanensis sp. nov.	0.5																			
3−4 A. ningshanensis	3.2-3.3	0.7																		
5 A. ater	7.1-7.3	7.6-7.7																		
6 A. dehuaensis	15.1-15.2	16.3-16.5	16.5																	
7 A. emilyae	12.9-13.3	13.5-14.1	11.7	15.7																
8 A. formosanus	13.8-13.9	14.8-15.1	14.1	15.7	14.6															
9 A. huangjietangi	16.8-16.9	17.2	15.0	16.8	14.8	16.2														
10 A. juliani	8.7-8.8	9.1-9.6	7.1	15.2	11.5	13.4	14.8													
11 A. meiguensis	16.4	17.0	15.4	18.1	15.4	15.6	15.2	16.8												
12 A. niger	13.2-13.3	14.6	13.5	15.7	12.9	9.0	14.6	12.9	13.9											
13 A. panzhihuaensis	16.2	17.1–17.4	16.2	15.3	16.6	16.0	15.2	15.5	11.6	14.4										
14 A. pingbianensis	11.1-11.2	11.7-12.4	11.8	14.8	13.0	14.5	13.0	12.2	16.8	11.7	14.9									
15 A. rufescens	12.1-12.2	12.3-12.7	12.7	14.3	8.0	14.1	14.3	12.3	17.3	12.7	16.0	12.9								
16 A. spinalis	14.0-14.3	15.1-15.6	15.2	14.3	13.9	13.9	13.4	13.9	16.0	13.5	15.8	13.3	13.0							
17 A. timi	12.1	13.6	13.3	15.8	12.9	14.0	14.8	13.7	15.8	12.0	15.5	12.2	13.9	14.3						
18 A. tranganensis	13.7-14.2	14.3-15.2	12.7	14.2	10.6	17.3	13.7	12.3	16.4	14.9	16.4	13.3	11.5	14.6	13.5					
19 A. vanhoensis	11.3-11.7	12.1-12.4	13.1	15.8	12.3	14.1	14.8	13.5	15.6	12.6	15.5	10.8	13.8	12.9	5.0	13.3				
20 A. yangdatongi	5.1	5.8-5.9	6.2	14.0	12.8	14.4	14.6	7.3	17.1	13.7	15.5	11.3	11.5	14.2	13.1	12.8	11.3			
21 A. yunkaiensis	11.7-12.1	13.0-13.7	12.8	14.7	13.1	12.3	12.5	12.5	15.8	12.2	15.7	11.6	13.3	12.0	14.1	13.5	13.6	12.0		
22 A. zugorum	11.6-11.9	12.8	13.1	14.3	12.9	13.7	14.4	13.5	15.0	13.4	15.3	10.9	13.5	13.3	13.4	12.5	12.0	12.2	10.9	

Table 2. Uncorrected p-distances (%) among Achalinus species inferred from mitochondrial CO1 gene.

Vouchor Number	CIB 119039	CIB 119040
voucher Number	Holotype	Paratype
Sex	Male	Male
SVL	255	204
TaL	74	58
TL	329	262
TaL/TL	0.225	0.221
Loreal	1/1	1/1
LorH	1.03/1.04	0.91/0.93
LorL	1.54/1.58	1.46/1.49
LorH/LorL	0.70/0.66	0.62/0.62
LSBI	1.78	1.52
LSBP	0.88	0.76
LSBI/LSBP	2.02	2.00
LSBI vs. LSBP	> 1	>1
HL	7.91	6.33
HW	4.80	3.38
HL/HW	1.65	1.87
ED	1.42/1.41	1.37/1.36
MT	23	23
SPL	3-2-1/3-2-1	3-2-1/3-2-1
IFL	5/6	5/5
IFL-1 st Chin	3/4	3/3
SPO	1/1	1/1
ТЕМ	2+2+4/2+2+4	2+2+4/2+2+4
aTEM-Eye	2/2	2/2
PrV	2	2
VEN	163	165
SC	69	72
Anal	Entire	Entire
DSR	23-23-23	23-23-23

Table 3. Main morphological characters of Achalinus hunanensis sp. nov.

A. hainanus Huang, 1975 (vs. = 1), *A. huangjietangi* (vs. < 1), *A. spinalis* (vs. < 1), *A. werneri* Van Denburgh, 1912 (vs. = 1) and *A. yunkaiensis* (vs. = 1). By having more ventrals (163–165), they can be distinguished from *A. dehuaensis* (vs. 142–154), *A. emilyae* (157–161), and *A. rufescens* (vs. 132–156).

Within the clade F, the two newly collected specimens from Hunan can be identified from *A. ater* by having nostril in the anterior part of the nasal (vs. nostril in the posterior part of the nasal), loreal length ~ $1.5 \times$ than loreal height (vs. loreal length > 2 × than loreal height), and more subcaudals (69–72 vs. 47–70). They are different from *A. juliani* by having different dorsal scale rows (23-23-23 vs. 25-23-23), less maxillary teeth (23 vs. 28), and less subcaudals (69–72 vs. 77–91). They differ from *A. yangdatongi* by having relatively shorter tail length in males (0.221 ~ 0.225 vs. 0.261 ~ 0.262), more ventrals in males (163–165 vs. 155), and fewer subcaudals in males (69–72 vs. 76) fewer maxillary teeth (23 vs. 24–26). They also can be easily distinguished from its sister group *A. ningshanensis* by the following morphological characters: (1) the suture between the internasals 2 × as long as the suture between the prefrontals vs. the suture between the

able 4. Morphological characters of Achalinus obtained from specimens examined in this study and literatures (Boulenger 1888, 1893, 1896; Denburgh 1912; Bourret 1935,
:020; Luu et al. 2020; Miller et al. 2020; Yu et al. 2020; Hou et al. 2021; Huang et al. 2021; Li et al. 2021; Chen et al. 2022; Ha et al. 2022; Yang et al. 2022; Zhang et al. 2023; Xu
t al. 2023). Int. fus.: internasal fused to prefrontal; Pre fus.: prefrontal fused to loreal; PtO: postoculars.

Species	TaL/TL	MT	Int fus.	Pre fus.	LorH/LorL	LSBI vs. LSBP	DSR	PtO	SPL	SPL-Eye	F	IFL-1 st Chin	TEM	aTEM-Eye	VEN	sc
A. ater	0.190 ~ 0.220	1	0	0	0.40	۲ ۲	(21-23)-(21-25)-(21-25)	0	9	4-5	5-6	1-3	2+2+3	2	160-170	47-70
A. dabieshanensis	0.168 ~ 0.223	ı	0	0	$0.73 \sim 0.83$	۲ ۷	23-23-23	0	9	4-5	£	1-3	2+2+3(4)	2	141-155	45-55
A. dehuaensis	0.206 ~ 0.286	31–33	0	0	1	~	23-23-23	0	9	4-5	5	1–3	2+2(3)+3(4)	1-2	142-154	63-81
A. emilyae	0.183 ~ 0.203	27-28	0	0	1	× 1	23-23-23	0	9	4-5	5	1-3	2+2+3	-	157-161	56-63
A. formosanus chigirai	0.317	14	0	-	I		(25-27)-(25-27)-25	0	9	4-5	5-6	I	2+2	2	161–167	96-97
A. f. formosanus	0.159	17	0	1(usually	-	"	29-27-25	0	9	4-5	6-7	I	2+2	-	158-184	61-83
A. hainanus	0.258 ~ 0.266	ı	0	0	I		23-23-23	0	9	4-5	2	1–3	1+2+3(4)	-	165-168	67-69
A. huangjietangi	0.152 ~ 0.232	ı	0	0	0.70 ~ 0.74	۲ ۷	23-23-23	0	9	4-5	5-6	1-3(4)	2+2+3(4)	2	157-170	40-67
A. hunanensis sp. nov.	0.221 ~ 0.225	23	0	0	0.62 ~ 0.70	~	23-23-23	0	9	4-5	5-6	1-3(4)	2+2+4	2	163-165	69-72
A. jinggangensis	0.174 ~ 0.217	ı	0	-	1	× 1	23-23-23	0	9	4-5	9	1-4	2(1)+2+3(4)	2	156-164	51-64
A. juliani	$0.224 \sim 0.268$	28	0	0	1	~	25-23-23	0	6(7)	4-5(5-6)	9	1-3(4)	2+2+4	2	163-179	77-91
A. meiguensis	0.142 ~ 0.238	17	-	0	I	I	(21-23)-(19-21)-(19-21)	-	9	4-5	9	1–3	2(3)+2(3)	-	146-173	39-60
A. niger	0.151 ~ 0.179	1	0	0	0.67	۲ ۲	25-25-23	0	9	4-5	9	1-3(4)	2+2(3)	2	169–185	52-72
A. ningshanensis	0.121 ~ 0.161	ı	0	0	$0.45 \sim 0.58$		23-23-23(21)	0	9	4-5	2	1-2(3)	2+2(3)+3(4)	1-2	159-174	41-46
A. panzhihuaensis	0.246	28	-	0	0.67	I	23-23-19	-	9	4-5	9	1–3	2+2+3	-	160	73
A. pingbianensis	0.243	ı	0	-	1		23-23-23	0	7	5-6	9	1-3	2+2+3	-	164	56
A. rufescens	0.191 ~ 0.276	23	0	0	0.80 ~ 1.00	~	23-(23-25)-23	0	9	4-5	2	1-3	2(1)+2+3(4)	1-2	132-156	58-82
A. spinalis	0.150 ~ 0.250	16-20	0	0	I	~	(23-25)-(23-25)-(23-25)	0	9	4-5	5-6	1–3	2+2(3)	1-2	138-175	48-67
A. timi	0.213	27	0	-	I	~	25-25-23	0	9	4-5	9	1–3	2+2+3	-	170	72
A. tranganensis	0.254(+)	29	0	0	I	~	25-23-23	0	9	4-5	9	1–3	2+2+3	2	171	73(+)
A. werneri	$0.250 \sim 0.300$	I	0	0	I		?-(21–23)-?	0	9	4-5	9	I	2+3(4)	I	157-191	67–98
A. yangdatongi	0.180 ~ 0.262	24–26	0	0	0.57	~	23-23-23	0	9	4-5	5-6	1–3	2+2/3+2/3	2	155-171	59-76
A. yunkaiensis	0.156 ~ 0.203	20-24	0	0	$0.49 \sim 0.64$	"	23-23-23	0	9	4-5	9	1-3(4)	2+2+3(4)	2	150-162	49–56
A. vanhoensis	0.264	32	0	-	I	× 1	25-23-23	0	6/7	4-5/5-6	9	1-4	2+2+3	7	176	84
A. zugorum	0.229	28	0	-	I	^	25-23-23	0	9	4-5	7	1–3	2+2+3	2	173	70

internasals subequal to the suture between the prefrontals; (2) relatively longer tail (TaL/TL: 0.221 ~ 0.225 vs. 0.121 ~ 0.161); (3) more subcaudals: 69–72 vs. 41–46; (4) two chin pairs vs. three chin pairs; (5) relatively narrow and long loreals (0.62 ~ 0.70 vs. 0.45 ~ 0.58) (more details are presented in Table 5).

Combined the results of molecular systematics and morphological characters above, the specimens newly collected in this work represent a new species, and we describe it herein.

 Table 5. Main morphological characters of Achalinus hunanensis sp. nov. and A. ning-shanensis.

Species	Achalinus hunanensis sp. nov.	A. ningshanensis
Sex	Males (<i>n</i> = 2)	Females $(n = 5)$
SVL	204-255	334-463
TaL	58-74	62-72
TL	262-329	398-527
TaL/TL	0.221 ~ 0.225	0.121 ~ 0.161
Loreal	1/1	1/1
LorH/LorL	0.62 ~ 0.70	0.45 ~ 0.58
LSBI/LSBP	2.00 ~ 2.02	0.95 ~ 1.11
LSBI vs. LSBP	>1	= 1
HL	6.33-7.91	11.17-13.72
HW	3.38-4.80	4.75-7.48
HL/HW	1.65-1.87	1.78-2.67
MT	23	_
SPL	3-2-1	3-2-1
IFL	5-6	5
Chins	2	3
IFL-1 st Chin	3-4	2-3
SPO	1	1
TEM	2+2+4	2+2+3/2+2+4/2+3+4
aTEM-Eye	2	1-2
VEN	163-165	159-174
sc	69-72	41-46
Anal	Entire	Entire
DSR	23-23-23	23-23-23 (rarely 21)
References	This study	Yang et al. 2022

Taxonomic account

Achalinus hunanensis sp. nov.

https://zoobank.org/05012A7E-84CE-4C6D-A915-51FCF365DFFE Figs 4, 5

Chresonymy. Achalinus ater: Shu et al. 2014; Shen et al. 2014; Gao et al. 2022.

Material examined. *Holotype*. CIB 119039 (Collection No. 20130505001), subadult male (Fig. 4), collected in early May 2013, by Sheng-Chao Shi and Sun-Jun Xiang from Huangyan Village, Hecheng District, Huaihua City, Hunan Province, China (27°28'N, 110°02'E; ca. 880 m a.s.l.).



Figure 4. The holotype (CIB 119039, subadult male) of *Achalinus hunanensis* sp. nov. **A** dorsolateral view **B** ventral view **C** right side of middle body view **D** dorsal head view **E** left side of head view **F** right side of head view **G** ventral head view. Photographs by SCS.

Paratype. CIB 119040, subadult male (Fig. 5), collected on 16 June 2022, by Sheng-Qiang Liu from Wazizhai, Ningxiang County, Changsha City, Hunan Province, China (28°00'N, 111°53'E; ca. 1020 m a.s.l.).

Etymology. This new species is named after its known distribution range, which is endemic to Hunan Province. The Chinese name is suggested as "湖南脊蛇" (Hú Nán Jǐ Shé) and the English name "Hunan Odd-scale Snake" or "Hunan Burrowing Snake" is suggested.

Diagnosis. (1) 23 rows of dorsal scales throughout the body, all dorsal scales strongly keeled, and the outmost one strongly keeled and enlarged; (2) tail relatively short, TaL/TL 0.221 ~ 0.225; (3) maxillary teeth 23; (4) the suture between internasals 2 × as long as that between prefrontals; (5) loreal one, subrectangular, LorH/LorL 0.62 ~ 0.70; (6) supralabials six, the 4th and 5th touch the eye; (7) the two anterior temporals in contact with eye; (8) ventrals 163–165, subcaudals 69–72, not paired.

Description of holotype. A subadult male with a total length of 329 mm (SVL 255 mm and TaL 74 mm); tail relatively short, Tal/TL 0.225; body slender,





cylindrical; head length (HL) 7.91 mm, head width 4.80 mm, HL/HW 1.65, slightly distinct from neck; eye small, ED 1.42/1.41 mm; maxillary teeth 23, small, equally sized and curved. Rostral small, triangular, only the upper tip visible from above. Length of the suture between the internasals (LSBI 1.78 mm) ~ 2 × as long as length of the suture between the prefrontals (LSBP 0.88 mm). Nostril in the anterior part of the nasal. Loreal one, subrectangular, loreal height (LorH) 1.03/1.04 mm, loreal length (LorL) 0.70/0.66 mm, LorH/LorL 0.62 ~ 0.70. Supraocular one. Frontal one, pentagonal, pointed backwards, much shorter than the parietals. Parietals paired and elongated. No preoculars and postoculars. Temporals 2+2+4, the anterior two contact the eye, the lower anterior temporal much larger, the upper medium temporal much larger, the upper posterior temporal much larger and separated from the other side one by two small scales which contact the parietals. Supralabials 6, 4th and 5th contact the eye, the last one much elongated. One mental. Two chin shields, similar length. Infralabials 5/6, the first one contact with each other after the mental and before the 1st chin shields, $1^{st}-3^{rd}/1^{st}-4^{th}$ touch the 1^{st} chin shields.

Dorsal scales lanceolate and strongly keeled; 23 rows throughout the body; those of the outmost rows on both sides significantly enlarged and strongly keeled. Ventrals 163, with two preventrals; anal entire; subcaudals 69, not paired.

Coloration of holotype. In life, dorsum dark, slightly metallic, vent blackbrown, dark brown near the margin, grey in the margin. A yellowish brown patch on the head occipital. The head ventral anterior part dark brown and posterior part yellowish white (Shu et al. 2014). In preservation, dorsum brown, vent anterior part grey and posterior part light brown. Ventral side of tail brown. The head ventral anterior part brown and posterior part grey.

Variations. Main morphological characters were listed in Table 3. The other sample are very similar to the holotype except that: (1) more ventrals: CIB 119040: 165; (2) more subcaudals: CIB 119040: 72; (3) vent coloration: CIB 119040: dark brown throughout the vent.

Distribution and habits. Achalinus hunanensis sp. nov. is currently only known from Hunan Province, China: Hecheng District, Huaihua City and Ningxiang County, Changsha City (880–1020 m a.s.l.). The holotype was found at night, near a mountain stream (AT 24 °C, RH 80%) with shrubs under subtropical evergreen broadleaves forest. It was moving from leaf litter to the stream. Earthworms were found at the same place, which we speculated as its prey (Shu et al. 2014).

Discussion

Based on molecular evidence, the newly collected *Achalinus* specimens in this study are most closely to *A. ningshanensis* but a genetic differentiation (*p*-distance 3.2%) already exists between these two groups (Fig. 2, Table 2). The newly collected *Achalinus* specimens and its sister group *A. ningshanensis* have separate distribution ranges at south of Yangtze River and north of Yangtze River, respectively, isolated by Three Gorges (Fig. 1). In addition, their estimated divergence time was at 0.48 Mya (Fig. 3), which broadly coincides with the formation of Three Gorges of the Yangtze River (0.30 ~ 0.12 Mya) (Zhang et al. 2018), hindering their communication and driving allopatric speciation. Moreover, combining their distinct morphological differences (Table 5), we describe them as a new species. Currently, 24 *Achalinus* species are reported.

Achalinus ater was first recorded in Hunan Province only based one specimen (Shen et al. 2014; Shu et al. 2014; Gao et al. 2022). However, this study found that this record was a misidentification and this specimen was designed as the new species holotype. Therefore, *A. ater* recorded on Hunan reptile check list should be transferred as *A. hunanensis*.

Due to the secretive life history and morphological similarities, many cryptic species may be "hidden" within known widely distributed species, such as *A. spinalis, A. rufescens*, and *A. ater* (Wang et al. 2019; Yang et al. 2022), and the description of *A. hunanensis* sp. nov. indicated that further study is necessary to conduct by using different *Achalinus* species and geographic populations to revise the mystery snakes and reveal their evolutionary history.

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Additional information

Conflict of interest

No conflict of interest was declared.

Ethical statement

No ethical statement was reported.

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Author contributions

Shun Ma: Laboratory work, methodology, data analysis, validation, writing: orgination and draft, writing: review and editing; Sheng-Chao Shi: methodology, investigation and resources, writing: orgination and draft, writing: review and editing; Sun-Jun Xiang: investigation and resources, writing: review and editing; Fu Shu: investigation and resources, writing: review and editing; Jian-Ping Jiang: conceptualization, data curation, project administrition, resources, supervision, writing: review and editing.

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Data availability

All of the data that support the findings of this study are available in the main text.

References

- Bouckaert R, Vaughan TG, Barido-Sottani J, Duchêne S, Fourment M, Gavryushkina A, Heled J, Jones G, Kühnert D, De Maio N, Matschiner M, Mendes FK, Müller NF, Ogilvie HA, du Plessis L, Popinga A, Rambaut A, Rasmussen D, Siveroni I, Suchard MA, Wu CH, Xie D, Zhang C, Stadler T, Drummond AJ (2019) BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. PLoS Computational Biology 15(4): e1006650. https://doi.org/10.1371/journal.pcbi.1006650
- Boulenger GA (1888) Description of two new snakes from Hongkong, and note on the dentition of *Hydrophis viperina*. Annals & Magazine of Natural History, Series 6 7(2): 43–44. https://doi.org/10.1080/00222938809460874
- Boulenger GA (1893) Catalogue of the Snakes in the British Museum (Nature History). I. London (Taylor & Francis), 448 pp.
- Boulenger GA (1896) Catalogue of the Snakes in the British Museum (Natural History). III. London, 612 pp.
- Bourret R (1935) Notes herpétologiques sur l'Indochine française. VIII. Sur les *Achalinus* d'Indochine. Bulletin général de l'Instruction Publique, Hanoi, 14^e Année (1934–1935) (5 janvier 1935): 101–104. [Separate: 1–4]

- Bourret R (1937) Notes herpétologiques sur l'Indochine française. XV. Lézards et serpents reçus au Laboratoire des Sciences Naturelles de l'Université au cours de l'année 1937. Descriptions de deux espèces et de deux variétés nouvelles. Bulletin général de l'Instruction Publique, Hanoi, 17^e Année (1937–1938), (4 Décembre 1937), Annexe: 57–80.
- Chen CW, Zhang CW, Ding GH, Zhang BW, Wang YP (2022) A new snake record of *Achalinus huangjietangi* in Zhejiang Province, China. Yesheng Dongwu 43(4): 1149–1150.
- Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: More models, new heuristics and parallel computing. Nature Methods 9(8): 772. https://doi.org/10.1038/ nmeth.2109
- Denburgh JV (1912) Concerning certain species of reptiles and amphibians from China, Japan, the Loo Choo Islands, and Formosa. Proceedings of the California Academy of Sciences 3(10): 187–258.
- Edgar RC (2004) MUSCLE: Multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Research 32(5): 1792–1797. https://doi.org/10.1093/nar/gkh340
- Gao ZW, Qian TY, Jiang JP, Hou DJ, Deng XJ, Yang DD (2022) Species diversity and distribution of amphibians and reptiles in Hunan Province, China. Shengwu Duoyangxing 30(2): 21290. https://doi.org/10.17520/biods.2021290
- Ha NV, Ziegler T, Sy TD, Le MD, Nguyen TQ, Luu VQ (2022) A new species of the genus *Achalinus* (Squamata: Xenodermidae) from Son La Province, Vietnam. Zootaxa 5168(3): 375–387. https://doi.org/10.11646/zootaxa.5168.3.8
- Hoang DT, Chernomor O, Haeseler AV, Minh BQ, Vinh LS (2018) UFBoot2: Improving the ultrafast bootstrap approximation. Molecular Biology and Evolution 35(2): 518–522. https://doi.org/10.1093/molbev/msx281
- Hou SB, Wang K, Guo P, Chen JM, Yuan ZY, Che J (2021) Two new species and a new country record of the genus *Achalinus* (Reptilia: Squamata: Xenodermidae) from China. Zootaxa 4950(3): 528–546. https://doi.org/10.11646/zootaxa.4950.3.6
- Hu SQ, Zhao EM (1966) Three new species of reptiles from Szechwan. Acta Zootaxon Sinica 3(2): 158–164. [In Chinese with English abstract]
- Hu SQ, Zhao EM, Liu CZ (1973) A survey of amphibians and reptiles in Kweichow Province, including a herpetological analysis. Dong Wu Xue Bao 19(2): 149–181. [In Chinese with English abstract]
- Huang RY, Peng LF, Yu L, Huang TQ, Jiang K, Ding L, Chang JK, Yang DC, Xu YH, Huang S (2021) A new species of the genus *Achalinus* from Huangshan, Anhui, China (Squamata: Xenodermidae). Asian Herpetological Research 12(2): 178–187. https://doi. org/10.16373/j.cnki.ahr.200075
- Kalyaanamoorthy S, Minh BQ, Wong TKF, Haeseler AV, Jermiin LS (2017) ModelFinder: Fast model selection for accurate phylogenetic estimates. Nature Methods 14(6): 587–589. https://doi.org/10.1038/nmeth.4285
- Koshikawa A (1982) Three new species of reptiles from Hainan Island, Guangdong Province. Smithsonian Herpetological Information Service 53(53): 1–10. https://doi. org/10.5479/si.23317515.53.1
- Kumar S, Suleski M, Craig JE, Kasprowicz AE, Sanderford M, Li M, Stecher G, Hedges SB (2022) TimeTree 5: an expanded resource for species divergence times. Molecular Biology and Evolution 38(8): msac174. https://doi.org/10.1093/molbev/msac174
- Li K, Yu M, Wu YY, Liao LH, Tang K, Liu Q, Guo P (2020) A new species of the genus *Achalinus* (Squamata: Xenodermidae) from southeastern Yunnan Province, China. Zootaxa 4860(1): 116–128. https://doi.org/10.11646/zootaxa.4860.1.6

- Li K, Wu YY, Xu RY, Zhu F, Ren JL, Guo P, Dong BJ (2021) A new species of the *Achalinus rufescens* complex (Xenodermidae: *Achalinus*) from Fujian Province, China. Zootaxa 5026(2): 239–254. https://doi.org/10.11646/zootaxa.5026.2.5
- Luu VQ, Ziegler T, Van HN, Van LO, Hoang TT, Ngo HT, Le MD, Tran DH, Nguyen TQ (2020) A new species of *Achalinus* (Squamata: Xenodermidae) from Trang An Landscape Complex, Ninh Binh Province, Vietnam. Zootaxa 4877(1): 174–184. https://doi. org/10.11646/zootaxa.4877.1.8
- Meyer CP, Geller JB, Paulay G (2005) Fine scale endemism on coral reefs: Archipelagic differentiation in turbinid gastropods. Evolution; International Journal of Organic Evolution 59(1): 113–125. https://doi.org/10.1111/j.0014-3820.2005.tb00899.x
- Miller MA, Pfeiffer W, Schwartz T (2010) Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: Proceedings of the Gateway Computing Environments Workshop (GCE), 14 Nov. 2010. New Orleans, LA, 1–8. https://doi.org/10.1109/GCE.2010.5676129
- Miller AH, Davis HR, Luong AM, Do QH, Pham CT, Ziegler T, Lee JL, Queiroz KD, Reynolds RG, Nguyen TQ (2020) Discovery of a new species of enigmatic odd-scaled snake (Serpentes: Xenodermidae: *Achalinus*) from Ha Giang Province, Vietnam. Copeia 108(4): 796–808. https://doi.org/10.1643/CH2020060
- Nguyen LT, Schmidt HA, Haeseler AV, Minh BQ (2015) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. Molecular Biology and Evolution 32(1): 268–274. https://doi.org/10.1093/molbev/msu300
- Ota H, Toyama M (1989) Taxonomic re-definition of *Achalinus formosanus* Boulenger, 1908 (Xenoderminae: Colubridae: Ophidia), with description of a new subspecies. Copeia 1989(3): 597–602. https://doi.org/10.2307/1445485
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA (2018) Posterior summarisation in Bayesian phylogenetics using Tracer 1.7. Systematic Biology 67(5): 901–904. https://doi.org/10.1093/sysbio/syy032
- Ronquist F, Teslenko M, Van DMP, Ayres DL, Darling A, Hohna S (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029
- Shen YH, Ye YY, Deng XJ (2014) Fauna Hunan: Reptilia. Hunan Science and Technology Press, Changsha, 220 pp. [In Chinese]
- Shu F, Shi SC, Tang ZJ, Liu ZX, Zhao DD, Xiang SJ, Deng XJ (2014) *Achalinus ater* was discovered in Hunan Province. Dongwuxue Zazhi 49(1): 113–115. [In Chinese]
- Stephane G, Jean-franc OD, Vincent L, Maria A, Wim H, And OG (2010) New algorithms and methods to estimate Maximum-likelihood phylogenies: Assessing the performance of PhyML 3.0. Systematic Biology 59(3): 307–321. https://doi.org/10.1093/ sysbio/syq010
- Tamura K, Stecher G, Kumar S (2021) MEGA11: Molecular evolutionary genetics analysis version 11. Molecular Biology and Evolution 38(7): 3022–3027. https://doi. org/10.1093/molbev/msab120
- Teynié A, David P, Lottier A, Le MD, Vidal N, Nguyen TQ (2015) A new genus and species of xenodermatid snake (Squamata: Caenophidia: Xenodermidae) from northern Lao People's Democratic Republic. Zootaxa 3926(4): 523–540. https://doi.org/10.11646/ zootaxa.3926.4.4
- Wang J, Li Y, Zeng ZC, Lyu ZT, Sung YH, Li YY, Lin CY, Wang YY (2019) A new species of the genus Achalinus from southwestern Guangdong Province, China (Squamata: Xenodermidae). Zootaxa 4674(4): 471–481. https://doi.org/10.11646/zootaxa.4674.4.6

- Xu RY, Li K, Zhang H, He H, Zhu F, Wu YY, Guo P (2023) Expanded description of *Achalinus yangdatongi* (Serpentes: Xenodermidae). Current Herpetology 42(1): 1–8. https://doi. org/10.5358/hsj.42.1
- Yang DC, Huang RY, Jiang K, Burbrink FT, Gong YN, Yu J, Zhang Y, Huang TQ, Huang S (2022) A new species of the genus *Achalinus* (Squamata: Xenodermidae) from Ningshan County, Shaanxi Province, China. Zootaxa 5190(1): 127–140. https://doi.org/10.11646/zootaxa.5190.1.5
- Yu M, Li K, Liu Q, Yang K, Wu YY, Guo P (2020) First Record of the *Achalinus yunkaiensis* from Maoershan National Nature Reserve, Guangxi, China. Chinese Journal of Zoology 55(6): 793–796. [In Chinese]
- Zhang XB, Liu Y, Wang SJ, Liu WM, Xue WX (2018) On the chronology of the Yellow Rivers and Yangtze Rivers. Mountain Research 36(5): 661–668. [In Chinese]
- Zhang C, Liu K, Huang RY, Hu TL, Yu L, Sun RL, Zhang YC, Wen J, Zhang BW (2023) A new species of the genus *Achalinus* (Squamata: Xenodermidae) from the Dabie Mountains, Anhui, China. Animals (Basel) 13(4): 708. https://doi.org/10.3390/ani13040708
- Zhao EM (2006) Snakes of China. I. Anhui Science and Technology Publishing House, Hefei, 372 pp. [In Chinese]
- Zhao EM, Huang MH, Zong Y (1998) Fauna Sinica: Reptilia. Vol. 3. Squamata, Serpentes. Science Press, Beijing, 522 pp. [In Chinese]
- Ziegler T, Nguyen TQ, Pham CT, Nguyen TT, Pham AV, Schingen VS, Nguyen TT, Le MD (2019) Three new species of the snake genus *Achalinus* from Vietnam (Squamata: Xenodermidae). Zootaxa 4590(2): 249–269. https://doi.org/10.11646/zootaxa.4590.2.3
- Zong Y, Ma J (1983) A new species of the genus *Achalinopsis* from Jiangxi and the restoration of this genus. Acta Herpetologica Sinica 2(2): 61–63. [In Chinese]

Appendix 1

Examined Achalinus specimens

- A. ater (n = 1): China, Guizhou Province, Xingyi County: CIB 63III5243.
- A. rufescens (n = 1): China, Hong Kong: CIB 119042.
- A. yunkaiensis (n = 1): China, Hunan Province, Xinning County: CIB 119041.