RESEARCH ARTICLE



New records of fish parasitic isopods of the gill-attaching genus *Mothocya* Costa, in Hope, 1851 from the Virgin Islands, Caribbean, with description of a new species

Kerry A. Hadfield¹, Paul C. Sikkel², Nico J. Smit¹

I Water Research Group (Ecology), Unit for Environmental Sciences and Management, Potchefstroom Campus, North West University, Private Bag X6001, Potchefstroom, 2520, South Africa **2** Department of Biological Sciences, Arkansas State University, P.O. Box 599, State University, AR, 72467, USA

Corresponding author: Kerry A. Hadfield (kerry.malherbe@nwu.ac.za)

Academic editor: Niel Bruce Received 11 June 2014 Accepted 13 August 2014 Published 10 September 2014
http://zoobank.org/6E790FDC-7C33-47E1-94A6-56C056334C48

Citation: Hadfield KA, Sikkel PC, Smit NJ (2014) New records of fish parasitic isopods of the gill-attaching genus *Mothocya* Costa, in Hope, 1851 from the Virgin Islands, Caribbean, with description of a new species. ZooKeys 439: 109–125. doi: 10.3897/zookeys.439.8093

Abstract

Two species of *Mothocya* Costa, in Hope, 1851 are reported from the Virgin Islands. *Mothocya xeno-branchia* Bruce, 1986 was collected from St. John Island from the gills of the Atlantic needlefish, *Strongy-lura marina*, which is a new locality record and also confirms a previously uncertain host identity. *Mothocya bertlucy* **sp. n.** is described from St. Thomas, St John and Guana Islands, from the gills of the redlip blenny, *Ophioblennius macclurei*, the first record of a blenny as host for any *Mothocya*. The distinguishing characters of *Mothocya bertlucy* **sp. n.** include its small size (< 9 mm) and eyes, the slender pleotelson with a narrowly rounded caudomedial point, extended uropod peduncle and uropods which do not extend past the pleotelson posterior margin, and the narrow pleon which is only slightly overlapped by pereonite 7.

Keywords

Cymothoidae, Mothocya, gill chamber, fish parasite, Caribbean Sea, St. Thomas, St. John, Guana Island

Introduction

Cymothoid isopods are one of the most recognisable groups of isopods to fisherman and anglers (Smit et al. 2014). These large (> 6 mm) aquatic parasites are commonly found on the external surface, inside the buccal cavity, or in the branchial cavity of their fish host. Cymothoids removed from the gills are often asymmetrical in body shape, twisted slightly due to the shape of the gill arches and operculum in the branchial cavity (Kensley and Schotte 1989).

In some cases, these parasites cause gill and branchial filament damage (Kroger and Guthrie 1972, Colorni et al. 1997). Williams and Williams (1978) commented on the discolouration and considerable erosion of the gill filaments and opercular flap in some fish they studied. Rokicki (1982) noted atrophy of the gill filaments which automatically affects the fish's development; and Colorni et al. (1997) reported on deformed and calcified gill rakers as well as gill filaments which were dystrophic and fused together with total obliteration of both primary and secondary lamellae.

One of these gill-attaching cymothoid genera is *Mothocya* Costa, in Hope, 1851. Historically the systematics and biology of this genus had not been considered problematic, but Bruce (1986) showed that *Irona* Schioedte & Meinert, 1884 and *Mothocya* were synonymous and that many of the species were misidentified, which had led to the misrepresentation of their hosts and distributions. Bruce (1986) comprehensively reviewed *Mothocya* and corrected many of these errors, revising seven species and describing 18 new species. Since then, another four species have been described (WoRMS 2014), making a total of 29 valid *Mothocya* species in the world (Smit et al. 2014).

There are six known species of *Mothocya* in the Caribbean Sea. These are *M. argenosa* Bruce, 1986 (Bermuda; Florida and Georgia, USA; Cuba; and the British Virgin Islands); *M. bermudensis* Bruce, 1986 (Bermuda; Haiti; Saint Barthélemy, Leeward Islands); *M. bohlkeorum* Williams & Williams, 1982 (Florida, USA; Bahamas; Saint Eustatius, Leeward Islands; and Puerto Rico); *M. nana* (Schioedte & Meinert, 1884) (Florida, Georgia and Maryland USA; Saint Barthélemy, Leeward Islands; and Panama), *M. omidaptria* Bruce, 1986 (Brazil and West Indies), and *M. xenobranchia* Bruce, 1986 (Florida, USA; and Venezuela). To date there are no known species recorded from the US Virgin Islands and only one species known from the British Virgin Islands (*M. argenosa*). The new species described here increases the number of species known from the Caribbean to seven.

Methods

Collections were made from the Virgin Islands, specifically St. John, and St. Thomas, US Virgin Islands, and Guana Island, British Virgin Islands, in the Caribbean Sea during 2013 as part of a study on blood parasites of Caribbean reef fishes. Atlantic needlefish (*Strongylura marina*) were collected near the surface at night by snorkelers using hand nets, while redlip blennies (*Ophioblennius macclurei*) were collected by hand nets during

the day from reef habitat in shallow bays by snorkelers or divers. Isopods were removed from the gills of their infected hosts using forceps, preserved in 70% ethanol, and processed according to techniques described in Hadfield et al. (2010, 2011). Species descriptions were prepared in DELTA (Descriptive Language for Taxonomy, see Coleman et al. 2010) using a general Cymothoidae character set (as in Hadfield et al. 2013, 2014). Ratios and measurements were rounded off to one decimal place and were made using maximum values of the specific measured article. Classification follows Brandt and Poore (2003). Host nomenclature and distribution are from FishBase (Froese and Pauly 2014).

Abbreviations. AMNH – American Museum of Natural History, New York, NY, USA; TL – total length; USNM – National Museum of Natural History, Smithsonian Institution, Washington, DC, USA; W – width.

Taxonomy

Family Cymothoidae Leach, 1814

Genus Mothocya Costa, in Hope, 1851

Mothocya Costa, in Hope, 1851: 48. – Trilles 1968: 168. – Monod 1971: 174. – Bruce 1986: 1092–1095. – Trilles 1994: 197.

Irona Schioedte & Meinert, 1884: 381. – Stebbing 1905: 27. – Richardson 1905: 265. – Hale 1926: 218. – Monod 1971: 174. – Kussakin 1979: 307. – Trilles 1994: 166.

Diagnosis. Body not vaulted, widest at pereonite 5, usually twisted to one side. Cephalon with rostrum folded back, anterior margin rounded. Antennae widely separated, antennula longer and more stout than antenna. Eyes distinct. Maxilliped article 3 with 3–5 recurved robust setae; without oostegite lobe. Maxilla mesial lobe partly fused to lateral lobe. Maxillula simple. Pereonite 1 anterolateral angles slightly extended around cephalon. Pleon subequal to pereon. Pleonite 1 partly concealed by pereonite 7. Coxae 5–7 dorsally visible, projecting posteriorly past respective somite; large, and rounded, reniform. Brood pouch formed from coxae 2–4 and 6. Pereopods without carina, never enlarged or with protrusions. Pleopods simple, without setae. Pleopods 3–5 with lamellar proximomedial lobe, frequently with peduncle lobe. Uropod peduncle without retinaculae, exopod longer than endopod.

Type species. *Mothocya epimerica* Costa, in Hope, 1851; by subsequent designation (Bruce 1986). Costa, in Hope (1851) described three species, *M. contracta* Costa, in Hope, 1851, *M. detecta* Costa, in Hope, 1851 and *M. epimerica* of which only *M. epimerica* is recognised as a valid species.

Remarks. Female *Mothocya* are often twisted to one side due to the confines of the gill chamber. *Mothocya* can be identified by the asymmetrical body shape, antennula longer than the antenna, a maxilliped with an oostegite lobe and the brood pouch from coxae 2–4 and 6. Males are smaller and not twisted, with appendix masculina on pleopod 2.

A detailed diagnosis of *Mothocya* was given by Bruce (1986), including female and male characters as well as additional characters for the genus. The current diagnosis is a shortened and updated version with more information on the main defining characters such as the body, pleopod and uropod morphology. These important characteristics are very useful in species identifications, as is the host species with some *Mothocya* species being host species or host genus specific.

Bruce (1986) synonymised *Irona* with *Mothocya*, with many of the *Irona* species actually being junior synonyms for *Mothocya* species. The validity of the genus *Irona* was considered uncertain for many years (Monod 1923, 1971, Trilles 1968) after Schioedte and Meinert (1884) described it as well as redescribing *Mothocya* in the same paper. Bruce (1986) described 18 new species of *Mothocya* in his review, nine of which had synonymies from earlier misidentifications. Many species appear very similar in general appearance, with the antennulae, antennae, mouthparts and pereopods uniform across species and thus not very informative at species level (Bruce 1986).

When looking at individual characters, *Mothocya* can be distinguished from other gill-inhabiting genera. *Elthusa* Schioedte & Meinert, 1884 is similar to *Mothocya* and can be distinguished by the antennula being shorter than the antenna (longer in *Mothocya*), maxilliped article 3 is slender with setae (robust and without setae in *Mothocya*), and the pereopod dactyli are relatively short whereas they are long and robust in *Mothocya* (Bruce 1990). *Ichthyoxenus* Herklots, 1870, differs from *Mothocya* with the antennula being shorter than the antenna, having a strongly ovate and vaulted body, as well as a narrow pleon and short and rounded coxae.

Mothocya occurs in all oceans and is predominantly tropical and subtropical in its distribution. Currently 29 species names are valid (*Mothocya contracta* Costa, in Hope, 1851 designated as *nomen dubium*), with four species described since Bruce's (1986) monograph.

Mothocya xenobranchia Bruce, 1986

Figs 1-2

Mothocya xenobranchia Bruce, 1986: 1116–1119, figs. 13–14. – Trilles 1994: 203. – Bunkley-Williams et al. 1998: 29. – Bunkley-Williams et al. 2006: 178. – Schotte et al. 2009: 983.

Material examined. \bigcirc (15.0 mm TL; 10.0 mm W), \bigcirc (9.0 mm TL; 4.0 mm W) collected from Lameshur Bay, 18°18'59"N, 64°43'25"W, St. John Island, US Virgin Islands, from the gills of the Atlantic needlefish (34 mm TL), *Strongylura marina*, 18 May 2013, coll. Nico J. Smit (AMNH_IZC 00197448).

Ovigerous female. Body moderately twisted, 1.4 times as long as greatest width, strongly arched longitudinally, widest at pereonite 3, most narrow at pereonite 1, lateral margins slightly convex. Cephalon 0.7 times longer than wide, visible from dorsal view,

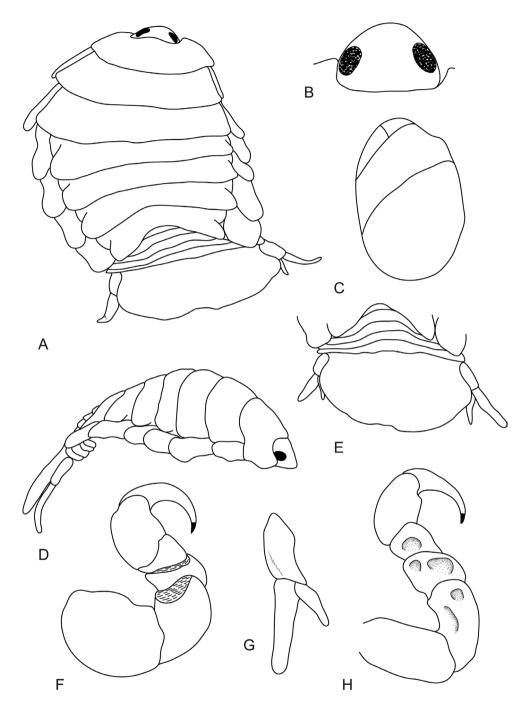


Figure 1. *Mothocya xenobranchia* Bruce, 1986 (15 mm) (AMNH_IZC 00197448): **A** dorsal view **B** dorsal view of cephalon **C** oostegites **D** lateral view **E** dorsal view of pleotelson **F** pereopod 1 **G** uropod **H** pereopod 7 showing indentations.

subtriangular. Eyes oval with distinct margins, 0.2 times width of cephalon, 0.4 times length of cephalon. Coxae 2–3 narrow; 4–7 large, rounded and produced, slightly produced past pereonite margin. Pereonites 1–4 increasing in length and width; 5–7 decreasing in length and width; becoming more progressively rounded posteriorly. Pleon with pleonite 1 largely concealed by pereonite 7; pleonites posterior margin smooth, mostly concave; pleonites 2–5 partially overlapped by pereonite 7; pleonite 5 posterior margin straight. Pleotelson 0.6 times as long as anterior width, dorsal surface smooth, anterolateral margin recessed, lateral margins widen slightly then curve inwards, posterior margin broadly truncate, without median point.

Antennula comprised of 7 articles; articles 1 and 2 distinct and articulated; article 2 0.8 times as long as article 1; article 3 as long as wide, 0.5 times as long as combined lengths of articles 1 and 2; last article terminating in 4–7 short simple setae. Antenna comprised of 7 articles; article 3 1.2 times as long as article 2, 2.1 times as long as wide; article 4 2.3 times as long as wide, 0.9 times as long as article 3; article 5 0.7 times as long as article 4, 1.7 times as long as wide; last article terminating in 6–7 short simple setae.

Molar process present, mandible palp without setae. Maxillula with 4 terminal robust setae. Maxilla lateral lobe with 2 recurved robust setae; mesial lobe with 2 large recurved robust setae. Maxilliped weakly segmented, palp article 2 with no simple setae, article 3 with 4 recurved robust setae and no simple setae.

Pereopod 1 basis 1.2 times as long as greatest width; ischium 0.9 times as long as basis; merus proximal margin with slight bulbous protrusion; carpus with straight proximal margin; propodus 1.3 times as long as wide; dactylus slender, 1.1 times as long as propodus, 2.3 times as long as basal width. Pereopod 7 basis 1.9 times as long as greatest width; ischium 0.9 as long as basis, without protrusions; merus proximal margin without bulbous protrusion, 0.5 as long as ischium, 0.9 times as long as wide; carpus 0.9 as long as ischium, without bulbous protrusion, 1.1 times as long as wide; propodus 0.8 as long as ischium, 1.7 times as long as wide; dactylus slender, 0.9 as long as propodus, 2.4 times as long as basal width. Pereopod 7 with small indentations on the inner side of the ischium, merus and carpus.

Pleopod 1 exopod as long as wide, lateral margin strongly convex, distally broadly rounded, mesial margin strongly convex; endopod 1.2 times as long as wide, lateral margin weakly convex, distally narrowly rounded, mesial margin straight, peduncle 0.7 times as wide as long. Pleopods 2–5 similar in structure to pleopod 1. Large medial lobes present and increasing in size from pleopods 1 to 5. Peduncle lobes increasing in size from pleopods 2 to 5.

Uropod longer than pleotelson; peduncle 0.7 times longer than exopod, lateral margin without setae; rami extending beyond pleotelson, marginal setae absent, apices broadly rounded. Endopod apically slightly pointed, 3.6 times as long as greatest width, lateral margin weakly convex, mesial margin weakly convex, terminating without setae. Exopod extending beyond endopod, 1.9 times longer than endopod, 3.8 times as long as greatest width, apically rounded, lateral margin straight, mesial margin straight, terminating without setae.

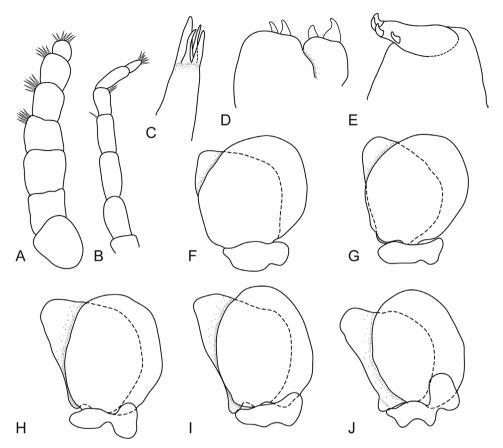


Figure 2. *Mothocya xenobranchia* Bruce, 1986 (15 mm) (AMNH_IZC 00197448): **A** antennula **B** antenna **C** tip of maxillula **D** tip of maxilla **E** tip of maxilliped article 3 **F** pleopod 1 **G** pleopod 2 **H** pleopod 3 **I** pleopod 4 **J** pleopod 5.

Type material. Holotype (16.2 mm TL) from the gill cavity of *Tylosurus crocodilis crocodilis* from Bahia Mochima, Venezuela (USNM 216274); Paratypes (USNM 216275–216278) (Bruce 1986; not examined).

Distribution. Off the coast of Florida, Florida Keys (USA); Cumaná, Venezuela (Bruce 1986, Bunkley-Williams et al. 1998, Schotte et al. 2009); and St. John Island, US Virgin Islands (present study).

Hosts. Known from the hound needlefish, *Tylosurus crocodilis crocodilis* (Péron & Lesueur, 1821) (Bruce 1986, Bunkley-Williams et al. 1998, Schotte et al. 2009) and *Strongylura marina* (Walbaum, 1792) (previously unconfirmed but verified in the present study). There is also another unconfirmed record from *S. notata notata* (Poey, 1860) in Florida (Bruce 1986).

Remarks. *Mothocya xenobranchia* is known from Belonidae fish hosts and distinguished by the broad body which is arched in lateral view, the invaginations on the inner portion of pereopod 7, antenna with seven articles, and the shape of the pleotelson which is tapered anteriorly, then widens before bluntly rounding off.

When comparing *M. xenobranchia* from the present study to the description given by Bruce (1986) there are a few minor differences but these are within the normal range of species variation. Variations include a different length to width ratio of the body and size of the eyes on the cephalon, more pronounced rostrum in the holotype, different number of setae on maxilla, but these characteristics given by Bruce (1986) are averages based on many specimens and can be variable depending on the specimen. In his remarks on the species, Bruce (1986) states the antenna can have seven or eight articles too and thus even this difference can be accounted for.

The other Caribbean species differ from *M. xenobranchia* in that *M. bermudensis* is smaller overall, with smaller eyes and less produced coxae; *Mothocya argenosa* has larger eyes, a larger and rounder pleotelson and smaller coxae; and *M. nana* has a narrower body shape and is not arched longitudinally. *Mothocya bohlkeorum* has a narrow strongly produced rostrum; antennula and antenna bases closer together; larger and rounder coxae; and less developed proximomedial and peduncle lobes on the pleopods. Lastly, *M. omidaptria* has much longer uropods, is not arched in lateral view, acute coxae on pereonite 7, and a narrowly produced rostrum. Furthermore, these species all have different hosts to *M. xenobranchia* and thus there is no overlap of this isopod species on its host species in the Caribbean.

This record of *M. xenobranchia* in the US Virgin Islands is a new locality record and also confirms the previously uncertain host record of *Strongylura marina* (Bruce 1986). The locality record conforms to the distribution of this species within the western Atlantic. Likewise, the host record is also from a Belonidae species and thus conforms to the host preference of this species.

Mothocya bertlucy sp. n.

http://zoobank.org/DC08E45E-5DDF-40D5-9310-B3AEA5C68265 Figs 3–7

Material examined. All material from the gills of the redlip blenny, Ophioblennius macclurei.

Holotype. Ovigerous \bigcirc (8.0 mm TL; 4.5 mm W), collected from Lameshur Bay, 18°18'59"N, 64°43'25"W, St. John Island, US Virgin Islands, July 2013, coll. L. Renoux & J. Sellers (AMNH_IZC 00197449).

Paratypes. \bigcirc dissected (7.0 mm TL; 3.5 mm W), three immature $\Diamond \Diamond$, one dissected (5.5–6.0 mm TL; 2.0–2.5 mm W), collected from Brewers Bay, 18°20'24"N, 64°58'44"W, St. Thomas Island, Caribbean Sea), 19 May 2013, coll. J. A. Barry & A. McCammon (AMNH_IZC 00197450). Ovigerous \bigcirc (9.0 mm TL; 5.0 mm W), collected from Lameshur Bay, 18°18'59"N, 64°43'25"W, St. John Island, US Virgin Islands, July 2013, coll, L. Renoux & J. Sellers (AMNH_IZC 00197451). Ovigerous \bigcirc (7.5 mm TL; 4.0 mm W), mature \Diamond (6.0 mm TL; 4.0 mm W), collected from Guana Island, 18°28'0"N, 64°33'59"W, British Virgin Islands, 07 July 2013, coll: R. Ditter & J. Barry (AMNH_IZC 00197452).

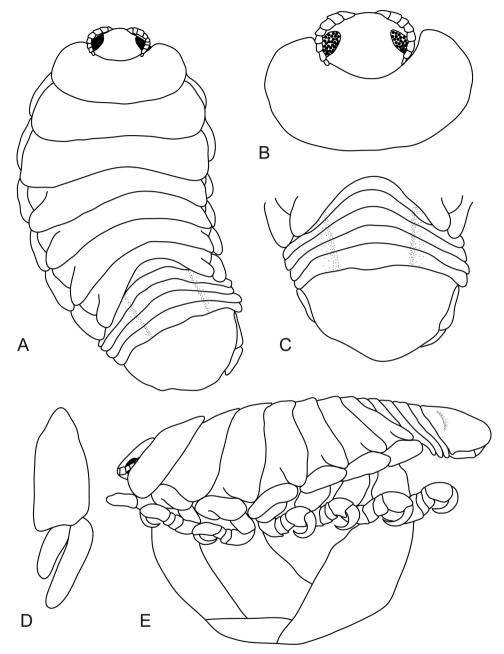


Figure 3. *Mothocya bertlucy* sp. n. ovigerous female holotype (7 mm) (AMNH_IZC 00197449): **A** dorsal view **B** anterodorsal view of pereonite 1 and cephalon **C** dorsal view of pleotelson **D** uropod **E** lateral view.

Ovigerous female holotype. Body oval and moderately twisted, 1.9 times as long as greatest width, widest at pereonite 3, most narrow at pereonite 1, lateral margins slightly convex. Cephalon 0.7 times longer than wide, visible from dorsal view. Eyes

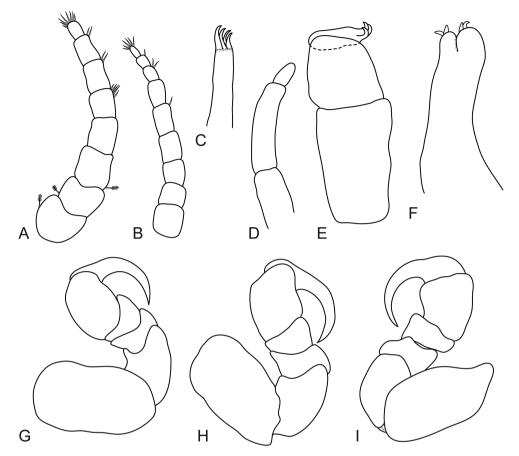


Figure 4. *Mothocya bertlucy* sp. n. female paratype (7 mm) (AMNH_IZC 00197450): **A** antennula **B** antenna **C** maxillula **D** molar process **E** maxilliped **F** maxilla **G** percopod 1 **H** percopod 2 **I** percopod 7.

oval with distinct margins, 0.2 times width of cephalon, 0.4 times length of cephalon. Pereonite 1 smooth, anterolateral angle rounded. Posterior margins of pereonites smooth and slightly curved laterally. Coxae narrow with rounded point, shorter or same length as pereonite. Pereonites 1–3 increasing in length and width; 4–7 decreasing in length and width, becoming progressively rounded posteriorly. Pleon with pleonite 1 largely concealed by pereonite 7, visible in dorsal view; pleonites posterior margin smooth, mostly concave; pleonite 2 partially overlapped by pereonite 7; pleonite 5 posterior margin slightly concave. Pleotelson 0.6 times as long as anterior width, dorsal surface smooth, lateral margins weakly concave, posterior margin converging to blunt caudomedial point.

Antennula comprised of 8 articles; articles 1 and 2 distinct and articulated with plumose setae; article 2 0.9 times as long as article 1; article 3 1.2 times as long as wide, 0.5 times as long as combined lengths of articles 1 and 2 with plumose seta; short simple

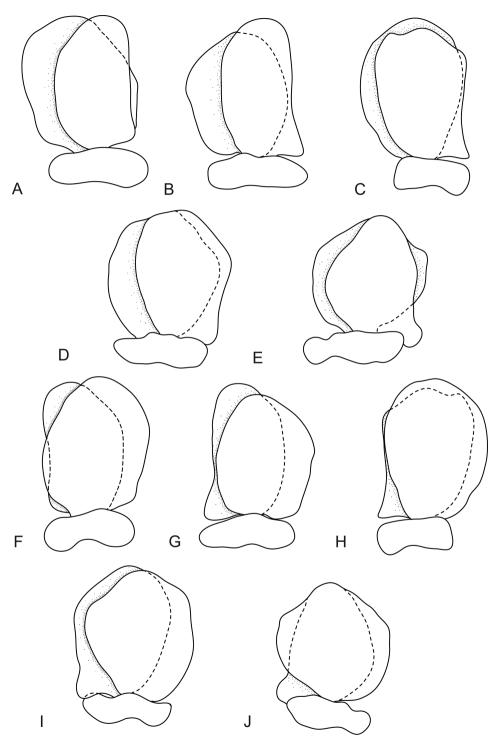


Figure 5. *Mothocya bertlucy* sp. n. female paratype (7 mm) (AMNH_IZC 00197450): **A–E** dorsal pleopod 1–5 respectively **F–J** ventral pleopod 1–5 respectively.

setae present on last four articles, last article terminating in 4–8 short simple setae. Antenna comprised of 9 articles; article 3 1.3 times as long as article 2, 1.3 times as long as wide; article 4 1.4 times as long as wide, 1.1 times as long as article 3; article 5 as long as article 4, 1.4 times as long as wide; short simple setae on last three articles, last article terminating in 6–7 short simple setae.

Molar process present, mandible palp without setae. Maxillula with 4 terminal robust setae. Mxilla lateral lobe with 2 recurved robust setae; mesial lobe with 2 large recurved robust setae. Maxilliped comprised of 3 articles, palp article 2 without simple setae, article 3 with 3 recurved robust setae, and no simple setae.

Pereopods without robust or simple setae. Pereopod 1 basis 1.8 times as long as greatest width; ischium 0.6 times as long as basis; merus proximal margin without bulbous protrusion; carpus with straight proximal margin; propodus 1.4 times as long as wide; dactylus slender, 1.3 times as long as propodus, 2.6 times as long as basal width. Pereopod 2 propodus 1.3 as long as wide; dactylus 1.3 as long as propodus. Pereopod 7 basis 1.7 times as long as greatest width; ischium 0.7 as long as basis, without protrusions; merus proximal margin with slight bulbous protrusion, 0.4 as long as ischium, 0.6 times as long as wide; carpus 0.9 as long as ischium, 1.3 times as long as wide; dactylus slender, 1.7 as long as propodus 0.9 as long as ischium, 1.3 times as long as wide; dactylus slender, 1.7 as long as propodus, 2.7 times as long as basal width.

Pleopod 1 exopod 1.3 times as long as wide, lateral margin weakly convex, distally narrowly rounded, medial margin weakly oblique, mesial margin strongly convex; endopod 1.8 times as long as wide, lateral margin weakly convex, distally narrowly rounded, mesial margin straight, peduncle 0.4 times as wide as long. Pleopods 2–5 similar to pleopod 1. Proximomedial lobes present and increasing in size from pleopod 1 to 5. Peduncle lobes absent.

Uropod more than half the length of pleotelson, peduncle 1.2 times longer than rami, peduncle lateral margin without setae; rami not extending beyond pleotelson, marginal setae absent, apices broadly rounded. Endopod apically rounded, 2.8 times as long as greatest width, lateral margin straight, mesial margin straight, terminating without setae. Exopod extending beyond endopod, 1.7 times longer than endopod, 4.2 times as long as greatest width, apically rounded, lateral margin straight, mesial margin straight, terminating without setae.

Male. Males similar to females but smaller. Body more oval and not twisted, 2.1 times as long as wide. Maxilliped article three with three recurved robust setae. Maxilla with one recurved robust seta on the medial lobe and two on the lateral lobe. Penis set close together, medially united. Pleopod 2 appendix masculina basally swollen, 0.8 times as long as endopod, distally bluntly rounded. Pleotelson triangular converging to a sharp caudal point. Uropods extend past posterior margin of pleotelson and endopod is longer, exopod 1.5 times as long as endopod.

Size. Ovigerous females (7.0–9.0 mm TL; 3.5–5.0 mm W), non-ovigerous females (7.0 mm TL; 3.0 mm W); mature male (6.0 mm TL; 4.0 mm W), immature males (5.5–6.0 mm TL; 2.0–2.5 mm W).

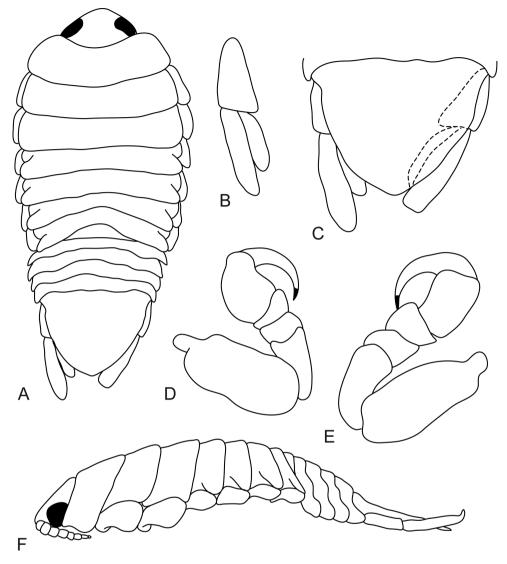


Figure 6. *Mothocya bertlucy* sp. n. male paratype (5.5 mm) (AMNH_IZC 00197450): A dorsal view **B** uropod **C** dorsal view of pleotelson **D** pereopod 1 **E** pereopod 7 **F** lateral view.

Etymology. This species is named in honour of Ernest H. ("Bert") Williams Jr. and Lucy Bunkley-Williams on the occasion of their retirement and in recognition of their contribution to Caribbean marine parasitology; noun in apposition.

Distribution. Known from St. John, St. Thomas, and Guana Islands, Caribbean Sea.

Hosts. Only known from the redlip blenny, Ophioblennius macclurei (Silvester, 1915).

Remarks. *Mothocya bertlucy* sp. n. can be identified by its unique host (redlip blenny), small size (like those reported from atherinids), relatively small eyes, the small

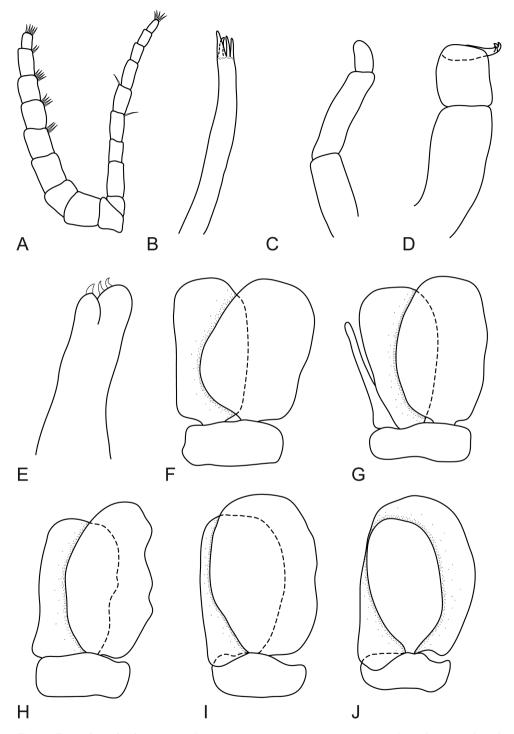


Figure 7. *Mothocya bertlucy* sp. n. male paratype (5.5 mm) (AMNH_IZC 00197450): **A** antennula and antenna **B** maxillula **C** molar process **D** maxilliped **E** maxilla **F–J** pleopod 1–5 respectively.

pleotelson with a narrowly rounded caudomedial point, large uropod peduncle with short rami, uropods which do not extend past the pleotelson posterior margin, and the narrow pleon which is only slightly overlapped by pereonite 7.

The species most similar to *Mothocya bertlucy* sp. n. is *M. rosea* Bruce, 1986 found on the Mexican and Californian coasts. In comparison to *M. bertlucy*, *M. rosea* has more produced proximomedial lobes on pleopods 3–5, larger eyes, broad truncate pleotelson, and four setae on the maxilliped article 3.

The three small *Mothocya* species from atherinids (*M. argenosa*; *M. epimerica*; and *M. waminda* Bruce, 1986) were all compared to the current species. *Mothocya argenosa* from the western Atlantic measures 5.6–9.8 mm, but has larger eyes, longer uropods, the pleotelson is more rounded and the posterolateral margins of pereonite 7 are acute. *Mothocya epimerica* from the Mediterranean has a more pointed rostrum, rounded pleotelson, larger eyes and four setae on the maxilliped. *Mothocya waminda* from the Indo-Pacific has an appendix masculina on pereopod 2 in the female and longer uropods.

Mothocya bertlucy sp. n. differs from all the other known Caribbean species in that *M. bohlkeorum* has much larger and more produced coxae and a larger truncate pleotelson; *M. nana* has a wider pleotelson, truncate rostrum and larger coxae; *M. bermudensis* has an antennula with only seven articles, large eyes and an arched body; and *M. omidaptria* has longer uropods extending past the pleotelson, a strongly produced rostrum and acute coxae as well as posterolateral angles of pereonite 7.

This is the first account of a *Mothocya* species from the US Virgin Islands and is also the first record on a blenny, which helps establish its status as a new species as Bruce (1986) commented that "host identity may be useful in making a *Mothocya* identification."

Acknowledgements

The financial assistance of the South African National Research Foundation (NRF project IFR2011040100022, N.J. Smit, PI and SFP12091012541, K.A. Hadfield, PI), the US National Science Foundation (NSF OCE-121615, P. Sikkel, PI), and the Falconwood Corporation towards this research is hereby acknowledged. Opinions expressed and conclusions arrived at are those of the authors and are not necessarily to be attributed to the funders. We thank J.A. Barry, R. Ditter, A. Mc Cammon, L. Renoux, J. Sellers, and J. Wagner for assistance with collection of fish specimens. We are also grateful to the staff of the MacLean Marine Science Center, Virgin Islands Environmental Resource Station, and Guana Island for logistic support. This is contribution number 116 from the University of the Virgin Islands Center for Marine and Environmental Studies. The authors would like to thank Dr Niel Bruce (Museum of Tropical Queensland, Australia) for valuable comments on the draft manuscript.

References

- Brandt A, Poore GCB (2003) Higher classification of the flabelliferan and related Isopoda based on a reappraisal of relationships. Invertebrate Systematics 17: 893–923. doi: 10.1071/ IS02032
- Bruce NL (1986) Revision of the isopod crustacean genus *Mothocya* Costa, in Hope, 1851 (Cymothoidae: Flabellifera), parasitic on marine fishes. Journal of Natural History 20(5): 1089–1192. doi: 10.1080/00222938600770781
- Bruce NL (1990) The genera Catoessa, Elthusa, Ichthyoxenus, Idusa, Livoneca and Norileca n. gen. (Isopoda, Cymothoidae), crustacean parasites of marine fishes, with descriptions of eastern Australian species. Records of the Australian Museum 42(3): 247–300. doi: 10.3853/j.0067-1975.42.1990.118
- Bunkley-Williams L, Williams EH Jr, Bashirullah AKM (1998) Some isopods associated with Venezuelan fishes. Caribbean Marine Studies 6: 27–30.
- Bunkley-Williams L, Williams EH Jr, Bashirullah AKM (2006) Isopods (Isopoda: Aegidae, Cymothoidae, Gnathiidae) associated with Venezuelan marine fishes (Elasmobranchii, Actinopterygii). Revista de Biologia Tropical 54(suppl. 3): 175–188.
- Coleman CO, Lowry JK, Macfarlane T (2010) DELTA for beginners. An introduction into the taxonomy software package DELTA. ZooKeys 45: 1–75. doi: 10.3897/zookeys.45.263
- Colorni A, Trilles J-P, Golani D (1997) *Livoneca* sp. (Flabellifera: Cymothoidae), an isopod parasite in the oral and branchial cavities of the Red Sea silverside *Atherinomorus lacunosus* (Perciformes, Atherinidae). Diseases of Aquatic Organisms 31: 65–71. doi: 10.3354/dao031065
- Costa A (1851) Caratteri di alcuni de'generi e specie nouve segnete nel presente catalogo. In: Hope FW (Ed) Catalogo dei crostacei Italiani e di molti altri de Mediterraneo, Napoli, 41–48.
- Froese R, Pauly D (2014) FishBase. World Wide Web electronic publication, version (04/2014). http://www.fishbase.org [accessed May 2014]
- Hadfield KA, Bruce NL, Smit NJ (2010) Redescription of the monotypic genus *Cinusa* Schioedte and Meinert, 1884 (Isopoda, Cymothoidae), a buccal-cavity isopod from South Africa. Zootaxa 2437: 51–68.
- Hadfield KA, Bruce NL, Smit NJ (2011) Cymothoa hermani sp. n. (Isopoda, Cymothoidae, Crustacea), a parasitic isopod, collected off the Zanzibar coast, Tanzania from the mouth of a parrotfish (Scaridae). Zootaxa 2876: 57–68.
- Hadfield KA, Bruce NL, Smit NJ (2013) Review of the fish-parasitic genus *Cymothoa* Fabricius, 1783 (Isopoda, Cymothoidae, Crustacea) from the south-western Indian Ocean, including a new species from South Africa. Zootaxa 3640: 152–176. doi: 10.11646/zootaxa.3640.2.2
- Hadfield KA, Bruce NL, Smit NJ (2014) Review of the fish parasitic genus *Ceratothoa* Dana, 1852 (Crustacea, Isopoda, Cymothoidae) from South Africa, including the description of two new species. Zookeys 400: 1–42. doi: 10.3897/zookeys.400.6878
- Hale HM (1926) Review of Australian isopods of the cymothoid group. Part II. Transactions of the Royal Society of South Australia 50: 201–234.
- Kensley B, Schotte M (1989) Guide to the Marine Isopod Crustaceans of the Caribbean. Smithsonian Institution Press, Washington, D.C. & London, 308 pp.

- Kroger RL, Guthrie JF (1972) Incidence of the parasitic isopod, Olencira praegustator, in juvenile Atlantic menhaden. Copeia 2: 370–374. doi: 10.2307/1442505
- Kussakin OG (1979) Marine and brackishwater likefooted Crustacea (Isopoda) from the cold and temperate waters of the Northern Hemisphere. Suborder Flabellifera. Opredeliteli po Faune SSSR, Izdavaemye Zoologicheskim Institutom Akademii Nauk SSSR. Izdatel'stvo Nauka, Leningrad, 1, 472 pp.
- Monod T (1923) Notes carcinologiques (parasites et commensaux). Bulletin de l'Institut Oceanographique de Monaco 427: 19–22.
- Monod T (1971) Sur quelques Crustacés de Tuléar. Tethys Supplément 1, 165-192.
- Richardson H (1905) A monograph on the isopods of North America. Bulletin of the United States National Museum 54: 1–727.
- Rokicki J (1982) Lironeca indica Edwards, 1840 (Crustacea, Isopoda) from Selar crumenophthalmus (Bloch). Wiadomości Parazytologiczne 28(1/2): 205–206.
- Schioedte JC, Meinert F (1884) Symbolæ ad monographium cymothoarum crustaceorum isopodum familiæ. IV. Cymothoidæ Trib. II. Cymothoinæ. Trib. III: Lironecinæ. Naturhistorisk Tidsskrift, Kjøbenhavn 14: 221–454.
- Schotte M, Markham JC, Wilson GDF (2009) Isopoda (Crustacea) of the Gulf of Mexico. In: Felder DL, Camp DK (Eds) Gulf of Mexico Origin, Waters, and Biota. Volume 1: Biodiversity. Texas A&M University Press, College Station, Texas, 973–986.
- Smit NJ, Bruce NL, Hadfield KA (2014) Global diversity of fish parasitic isopod crustaceans of the family Cymothoidae. International Journal for Parasitology: Parasites and Wildlife 3: 188–197.
- Stebbing TR (1905) Report on the Isopoda collected by Professor Herdman, at Ceylon, in 1902. Report to the Government of Ceylon on the Pearl Oyster Fisheries of the Gulf of Manaar, 1905, Supplementary Report 4(23): 1–64.
- Trilles J-P (1968) Recherches sur les Isopodes. Cymothoidae de cotes Francaises. I. Systematique et faunistique. II. Bionomie et parasitisme. Thèse de Doctorat es-sciences, Académie de Montpellier, Université des sciences et techniques du Languedoc, CNRS, AO 2305, 1–181.
- Trilles J-P (1994) Les Cymothoidae (Crustacea, Isopoda) du Monde. Podrome pour une faune. Studia Marina 21/22: 1–288. [for 1991]
- WoRMS Editorial Board (2014) World Register of Marine Species. http://www.marinespecies.org at VLIZ. [accessed 2014-07-29]
- Williams EH Jr, Williams LB (1978) Cymothoid isopods of some marine fishes from the northern Gulf of Mexico. Northeastern Gulf Sciences 2(2): 122–124.