

Twenty-four new species of *Aleiodes* Wesmael from the eastern Andes of Ecuador with associated biological information (Hymenoptera, Braconidae, Rogadinae)

Eduardo Mitio Shimbori^{1,†}, Scott Richard Shaw^{2,‡}

1 Universidade Federal de São Carlos, Departamento de Ecologia e Biologia Evolutiva, Rod. Washington Luís, km 235, CEP 13565-905, São Carlos, SP, Brazil **2** University of Wyoming, Department of Ecosystem Science and Management (3354), 1000 E. University Avenue, Laramie, WY 82071 USA

[†] <http://zoobank.org/3A171087-3715-4AA2-8731-DB3BEC7DECDA>

[‡] <http://zoobank.org/B4DCD509-07A6-45E1-AF25-296430A49EED>

Corresponding author: Eduardo Mitio Shimbori (shimbori@gmail.com)

Academic editor: K. van Achterberg | Received 1 Mart 2014 | Accepted 8 April 2014 | Published 28 April 2014

<http://zoobank.org/0EC88104-E98F-4E99-9397-DB767D38050E>

Citation: Shimbori EM, Shaw SR (2014) Twenty-four new species of *Aleiodes* Wesmael from the eastern Andes of Ecuador with associated biological information (Hymenoptera, Braconidae, Rogadinae). ZooKeys 405: 1–81. doi: 10.3897/zookeys.405.7402

Abstract

Aleiodes Wesmael is the most diverse roga-dine genus worldwide, with specialized koinobiont endoparasitic development in Lepidoptera caterpillars resulting in mummification of the host remains. This paper focuses on describing new *Aleiodes* species from the Yanayacu Biological Station, with special interest in those with biological information. We describe 24 new species (*Aleiodes albidactyl* sp. n., *Aleiodes albigena* sp. n., *Aleiodes albiviria* sp. n., *Aleiodes bimaculatus* sp. n., *Aleiodes cacuangoi* sp. n., *Aleiodes colberti* sp. n., *Aleiodes delicatus* sp. n., *Aleiodes dyeri* sp. n., *Aleiodes elleni* sp. n., *Aleiodes falloni* sp. n., *Aleiodes frosti* sp. n., *Aleiodes kingmani* sp. n., *Aleiodes longikeros* sp. n., *Aleiodes luteosicarius* sp. n., *Aleiodes marilynnae* sp. n., *Aleiodes mirandae* sp. n., *Aleiodes napo* sp. n., *Aleiodes nubicola* sp. n., *Aleiodes onyx* sp. n., *Aleiodes shakirae* sp. n., *Aleiodes stewarti* sp. n., *Aleiodes townsendi* sp. n., *Aleiodes tzantza* sp. n., and *Aleiodes yanayacu* sp. n.) from Napo Province in Ecuador, 16 of which were reared from host caterpillars. With these results 89 species of Neotropical *Aleiodes* are now known, with 41 of them having host records. The most commonly reared species were in the *circumscriptus/gastritor* species-group, and mostly associated with Geometridae hosts (six of ten species). Three species of *seriatus* species-group, in contrast, were each reared from a different family. One of these species (i.e. *A. frosti* sp. n.), reared from Notodontidae, cuts a posterior radial opening in the mummy for emergence, a unique behavior in *Aleiodes*, recorded here for

the first time. *A. luteosicarius* **sp. n.** is the first described species from Ecuador in the *pallidator* species-group. Differing from previously described *pallidator* species, which attack only Lymantriinae larvae, *A. luteosicarius* **sp. n.** attacks several species of Arctiinae larvae, being both subfamilies within Erebidae with densely setose caterpillars. We also describe new species of the *gressitti* and *pulchripes* species-groups.

Keywords

Taxonomy, biology, rearing, caterpillar, Napo Province, Yanayacu

Introduction

Aleiodes is the most common and speciose rogaeline braconid genus worldwide. Species richness estimations have changed dramatically in last decade. In the Taxapad catalog (Yu et al. 2012) 431 described species are recorded, but Butcher et al. (2012), in a single work published after that catalog, described 179 new species from Thailand. Those authors estimated the *Aleiodes* fauna of Thailand alone to include more than 400 species, pushing the diversity of *Aleiodes* to a much higher level than previously estimated, especially for the tropical fauna, previously considered not as rich as the Holarctic (S. Shaw 2006). Quicke (2012) discussed evidence suggesting a greatly unknown tropical Ichneumonidae fauna, where the lack of studies on small body sized groups and highly speciose genera are the main reasons for the underestimation. For Neotropical *Aleiodes*, Delfin–Gonzalez and Wharton (2002) estimated about 200 undescribed species, but in light of these recent works, this number is also likely to be much higher (in addition, for example, there are at least 160 undescribed species from Brazil. EMS, previously unpublished data). Prior to this study, there are 65 described species of *Aleiodes* in Neotropical region, 25 of them with host records (Fortier 2009, Townsend and Shaw 2009, Shimbori and Penteadó–Dias 2011).

The first division of *Aleiodes* into species-groups (S. Shaw et al. 1997) accounts for fifteen different groups. After phylogenetic analyses three additional groups were proposed (Fortier and Shaw 1999). Townsend and Shaw (2009) found the species of the closely-related *gastritor* and *circumscriptus* groups in Ecuador to be difficult to separate, and recommended treating these as one single group for Neotropical fauna. Although the existing species-group system presents some limitations when applied to Neotropical fauna (Townsend and Shaw 2009), it provides a working framework to investigate this diverse group and to improve our taxonomic knowledge. Most species-groups are represented in Neotropical Region, except for the *compressor*, *praetor*, *procerus*, *rugulosus*, *ufei* and *unipunctator* groups (S. Shaw et al. 1997, S. Shaw et al. 1998a, Marsh and S. Shaw 1998, S. Shaw et al. 1998b, Marsh and S. Shaw 1999, Marsh and S. Shaw 2001, Marsh and S. Shaw 2003, S. Shaw et al. 2006, S. Shaw et al. 2013), being the *pallidator* species-group represented only by undescribed species in Neotropical Region. Based on undescribed material from Neotropical collections, the *seriatus* species-group is the most diverse group in this region, followed by *circumscriptus*/*gastritor* species-group.

Rogadinae is a cosmopolitan subfamily of koinobiont endoparasitoids of Lepidoptera (M. Shaw 1983, M. Shaw and Huddleston 1991, M. Shaw 1994, S. Shaw 1997), currently divided in five tribes: Stiriopini, Clinocentrini, Yeliconinae, Rogadini and Aleiodini (= *Aleiodes*+*Heterogamus*) (van Achterberg 1995, Zaldívar-Riverón et al. 2008), the later containing *Aleiodes*, the most speciose genus with about 612 described species of the current 1,141 named Rogadinae species (Yu et al. 2012, Butcher et al. 2012, S. Shaw et al. 2013). All Rogadinae induce the hardening of the host larva before pupation, producing the so-called “mummy,” with the mummification of the host larva been considered the only biological synapomorphy of this subfamily (van Achterberg 1995). *Aleiodes* is known to attack almost exclusively exposed-feeding macrolepidopterans, especially the superfamilies Noctuoidea and Geometroidea, and to a lesser extent, Sphingoidea and Papilionoidea (S. Shaw et al. 1997, S. Shaw 2006). A few exceptions include some exposed-feeding microlepidopterans (e.g. Zygaenidae) and, in rare cases, such as *Aleiodes compressor* (Herrich-Schäffer, 1838), macrolepidopterans with semi-concealed habit (M. Shaw 2002). *Aleiodes* commonly attacks second and third instar caterpillars, but first instar records are known, though less successful (M. Shaw 1983). The host is mummified in later instars, and within the hardened skin of the host the parasitoid pupates and eventually emerges through a hole on posterior region of the mummy.

The present work presents descriptions of 24 *Aleiodes* new species, 16 of which with information on biology and photographs of the mummified host larvae. These comprise mostly exposed-feeding host records, with the parasitoids attacking hosts living in small trees and bushes, rather than herbaceous plants near ground level, or higher in the canopy.

Methods

Specimens for this study were collected during the *Caterpillars and Parasitoids of the Eastern Andes of Ecuador* project (Miller and Dyer 2009), and reared at the Yanayacu Biological Station and Center for Creative Studies (YBS). The YBS is situated on the northeastern slope of the Andes in Napo Province, Ecuador, the watershed streams at the YBS flow to the Amazon basin through the Río Napo, a major tributary of the Amazon River (S. Shaw 2012). The reserve comprises both primary- and secondary-growth montane forests (Miller and Dyer 2009). The plant community at Yanayacu is very diverse and complex, comprising species from at least 76 families (Rab Green et al. 2011).

Specimens were sampled using varied methods including yellow pan trap, Malaise traps situated on the ground and suspended in the canopy, Maxi net, hand collected with aspirators at a light sheet, ultraviolet light trap (= U.V. light trap or black light trap), and during daylight off vegetation with net or vials. Most of the specimens were reared from caterpillars (Greeney 2007), from 2007 to 2013. Caterpillars were sampled by walking through various habitats using two different methods: by hand collecting after inspecting herbs, shrubs and trees up to a height of approximately 2.5 m; or beating

plants over a white cloth square of 1×1 m size. Caterpillars were collected in clear plastic bags with their food plant, assigned identification codes, and transported to the rearing shed at YBS. Reared parasitoid specimens are associated with the voucher number of the respective host larva (e.g. YY-00000). Caterpillars and host plants were identified and recorded. Rearing took place in plastic bags in an open-air shelter with ambient temperatures and natural day length. Frass and decaying plant material were removed every other day and new plant material was provided as necessary. While cleaning out the bags, the caterpillars were inspected to note the date of caterpillar pupation or date of parasitoid pupation. Parasitoid pupae were inspected daily for emergence. All emerging adult parasitoids were kept with the original code given to the caterpillar to preserve host data. The parasitoids were preserved in alcohol and transferred to the University of Wyoming where they were dried and point mounted for identification.

Type specimens are deposited at University of Wyoming Insect Museum, University of Wyoming, Laramie, Wyoming, U.S.A. (UWIM). Additional specimens from Canadian National Collection, Ottawa, Canada (CNC) were added whenever appropriate to the scope of this work.

All the species proposed here are satisfactorily distinguished by morphological features. Nevertheless, for some of species, a comparison of ribosomal DNA sequences in gene COI by Donald Quicke was helpful in the definition of species limits.

Because of the supporting grant funding, this work focused on species sampled at the YBS. Some paratypes outside the boundaries of this refuge were included, but only if at least one type specimen is from YBS. All specimens included in this project are from the eastern face of the Andes Mountains, in Napo Province. The altitude of sampling sites ranged from 2,000 to 2,800 meters. In previous work on *Aleiodes* from Napo Province by Townsend and Shaw (2009) the altitudinal range was from 1,383 to 2,837m. Most of additional material is also from Napo province, with altitude ranging from 1,700 to 2,000 meters; only two type specimens are from lowland Manabí Province, at 400 meters elevation.

For recognition of subfamily Rogadinae see van Achterberg (1993) and Sharkey (1997). The definition of *Aleiodes* follows that of van Achterberg (1991) and S. Shaw (1993). Species-group definitions follow S. Shaw et al. (1997), Fortier and S. Shaw (1999) and Townsend and Shaw (2009). Terminology follows Sharkey and Wharton (1997) and S. Shaw et al. (1997). Microsculpture terminology follows that of Harris (1979). Wing vein terminology follows the system adopted by Sharkey and Wharton (1997).

We present descriptions for 24 new species from Northeastern Andes in Ecuador. Along with the description of the new species, we provide summarized taxonomic information on the nine previously described species from the same region (Townsend and Shaw 2009), and also a set of additional characters, not discussed in the original description. New biological information for these species is presented whenever available. Images were captured with a 3MP Leica video camera and a Leica M205C stereomicroscope running Leica Application Suite (LAS) software, and focus-stacked using the same software. Some minor adjustments in images and plate preparation were

performed in Adobe Photoshop version CS6. Pictures of caterpillars and host mummies before parasitoid emergence were taken, at the rearing site, by Wilmer Rosendo Simbaña, José Arturo Simbaña and Luis Alberto Salagaje.

Key to species of *Aleiodes* from eastern Andes of Ecuador

- 1 Occipital and epicnemial carina entirely absent (Figure 70) **2**
- Occipital carina present, at least laterally (Figures 46, 85); epicnemial carina present **3**
- 2(1) Head mostly black and granulate, rugose below antenna (Figure 2), pronotum and propleuron black; malar space 1/2 eye height (Figure 2) ***capillosus* Townsend, 2009**
- Head mostly white to light yellow and smooth, pronotum and propleuron yellow (Figures 69, 70); malar space longer, nearly as long as eye height (Figure 3) ***marilynae* sp. n.**
- 3(2) Apex of hind tibia with a row of flat setae along inner margin forming a distinct fringe (Figures 4, 20) **4**
- Apex of hind tibia without a row of flat setae along inner margin, setae may be present but not flattened and not forming a distinct fringe **12**
- 4(3) Occipital carina weak or absent at vertex (Figure 19) **5**
- Occipital carina complete and well-defined at vertex (Figures 35, 43) **10**
- 5(4) Mesopleuron yellow, sometimes with antero-dorsal corner brown to black (Figures 54); propodeum mostly uniformly colored, without strongly contrasting colors; fore wing vein 1CUa shorter than or equal to length of vein 1CUB (Figure 21) **6**
- Mesopleuron black with a posterior white thumb-like marking (Figure 4); propodeum black anteriorly with white spot posteriorly (Figure 6); vein 1CUa 1.5× longer than vein 1CUB (Figure 5) ***albidactyl* sp. n.**
- 6(5) Ocell–ocular distance about equal to diameter of lateral ocellus (Figure 19); metasomal tergum 1 at most 1.4× longer than its apical width; species with known biology with normal emergence behavior, cutting a postero-dorsal exit hole in the mummified host caterpillar (Figures 40, 57) **7**
- Ocelli larger, ocell–ocular distance 1/2 diameter of lateral ocellus; metasomal tergum 1 unusually long and narrow, more than 2× longer than its apical width (Figure 47); species with a unique emergence behavior, cutting the whole posterior tip away from the mummified host caterpillar (Figure 51) ***frosti* sp. n.**
- 7(6) Metasomal terga mostly black to dark brown (Figures 38, 54), sometimes lighter posteriorly or with light spots medially; flagellum dark brown to black; wings hyaline or weakly infusate **8**
- Metasoma entirely orangish yellow; flagellum black with white band medially (Figure 18); wing membrane tinged honey yellow ***albiviria* sp. n.**

- 8(7) Pronotum and mesonotum honey yellow; hind coxa bicolored, black apically and white basally (Figures 38, 54)..... **9**
- Pronotum and mesonotum black except light brown square postero-medially on mesoscutum, hind coxa light brown ***greeneyi* Townsend, 2009**
- 9(8) Propodeum and metapleuron black (Figure 54); fore wing vein 1M strongly curved basally (as in Figure 120); hind wing vein 1M more than 2.0× longer than vein r-m, vein m-cu absent ***longikeros* sp. n.**
- Propodeum and metapleuron pale yellow (Figure 38); fore wing vein 1M weakly curved (as in Figure 21); hind wing vein 1M shorter than vein r-m, vein m-cu present and distinctly antefurcal to r-m (as in Figure 83) ***dyeri* sp. n.**
- 10(4) Mesonotum dark brown (Figure 34); ocelli extremely small, ocell–ocular distance 2× ocellus width (Figure 35)..... ***delicatus* sp. n.**
- Mesonotum orange (Figures 41, 82); ocelli larger, ocell–ocular roughly as long as ocellus width (Figure 43) **11**
- 11(10) Apical 2/3 of hind coxa, metanotum, propodeum, metapleuron dorsally and mesopleuron on antero-dorsal corner black (Figure 41); fore wing second submarginal cell long and rectangular, veins 2RS and 3RS forming a right angle, and vein r less than half length of 3RSa; wings hyaline; frons with lateral ridges (Figure 43) ***elleni* sp. n.**
- Hind coxa, propodeum, metanotum, metapleuron, and mesopleuron off-white (Figure 82); second submarginal cell shorter and trapezoidal (Figure 83), angle between veins 2RS and 3RS obtuse, vein r 0.85× vein 3RSa; wings moderately infusate; lateral ridges on frons absent..... ***nebulosus* Townsend, 2009**
- 12(3) Ocelli small, ocell–ocular distance longer than width of lateral ocellus (Figures 14, 26, 89, 118)..... **13**
- Ocelli moderate-sized to large, ocell–ocular distance equal to or shorter than width of lateral ocellus (Figures 32, 46, 59) **23**
- 13(12) First and second metasomal terga with median carina present (Figures 23, 87); ovipositor sheaths at most 2/3 length of hind basitarsus **14**
- First and second metasomal terga with median carina absent (Figure 10); ovipositor about 2× length of hind basitarsus ***albiterminus* Townsend, 2009**
- 14(13) Malar space about as long as width of mandible base (Figure 11); head mostly black to dark brown, except for a crescent moon-shaped brown mark vertex, contrasting to thorax mostly yellow (Figure 12) .. ***arbitrium* Townsend, 2009**
- Malar space at least 1.25× width of mandible base (Figure 13); head and thorax coloration not as above: head and thorax mostly with same color, or head contrasting lighter than thorax (Figures 14, 22, 26)..... **15**
- 15(14) Occipital carina weak or interrupted at vertex (Figure 15) **16**
- Occipital carina complete and well-defined at vertex (Figures 26, 85)..... **20**
- 16(15) Mesopleuron with central disc lacking setae, smooth and shining (Figures 74, 77) **17**
- Mesopleuron with central disc setose and mostly granulate (Figures 16, 95) ... **19**

- 17(16) Tergite 2 mostly black with white markings, hind coxa black (Figures 74, 77, 78); hind wing vein M+CU distinctly shorter than 1M (Figure 76)..... **18**
- Tergite 2 entirely whitish, hind coxa yellowish (Figures 114, 116); hind wing vein M+CU about as long as 1M..... ***yanayacu* sp. n.**
- 18(17) Head, pronotum, propleuron and scutellum orangish yellow (Figure 74), except ocellar triangle black; tergite 1 entirely white; tergite 2 with median carina complete; mesoscutum with mid-posterior depressed area finely rugulose ***mirandae* sp. n.**
- Head and thorax black, except reddish brown mark on temples, just behind eyes; tergite 1 white with large black medial spot (Figure 77); tergite 2 with median carina incomplete, not reaching end of segment (Figure 78); mesoscutum not depressed postero-medially, entirely granular (Figure 79) ***napo* sp. n.**
- 19(16) Head orange, except for black ocellar triangle, contrasting with mostly black body (Figure 92); mesopleuron entirely black (Figure 95); metasoma stout, wider than propodeum, tergum 1 shorter than its apical width (Figure 94); ovipositor sheaths shorter than 1/2 length of hind basitarsus; hind wing vein m-cu distinct ***onyx* sp. n.**
- Head mostly yellowish brown with large black semicircular spot on occiput, vertex and ocellar triangle also black (Figures 14, 15); mesopleuron with ventral 1/2 yellowish brown, dorsally black (Figure 16); metasoma narrower than propodeum, tergum 1 about 1.2× longer than apical width (Figure 17); ovipositor sheaths longer than 1/2 length of hind basitarsus; hind wing vein m-cu absent (Figure 21) or at most weakly indicated by infumate pigmentation..... ***atripileatus* Townsend, 2009**
- 20(15) Central disc of mesopleuron smooth and shining (Figure 108); wings black; tarsal claws strongly pectinate (Figure 126); metasomal tergite 2 coarsely longitudinally costate (Figure 107) ***stilpnos* Townsend, 2009**
- Central disc of mesopleuron with various types of surface micro-sculpturing, not smooth and shining (Figure 28); wings clear or weakly infusate; tarsal claws simple or weakly pectinate basally; metasomal tergite 2 with various weaker micro-sculpturing, not coarsely longitudinally costate (Figures 23, 27, 87, 88)..... **21**
- 21(20) Pronotum and mesonotum mostly black (Figure 28); hind coxa surface granulate..... **22**
- Pronotum and mesonotum yellowish brown (Figure 22); hind coxa rugose dorsally ***bimaculatus* sp. n.**
- 22(21) Mesoscutum with square orangish brown mark postero-medially (Figure 89); head mostly dark brown to black with crescent moon-shaped honey brown area bordering eyes at temples (Figure 84); metasoma mostly dark brown, light markings when present most frequent apically beyond tergite 4 (Figure 87), rarely reaching apex of tergite 1 in males (Figure 88)..... ***nubicola* sp. n.**
- Mesoscutum entirely black (Figure 26); head color variable, mostly yellowish with black occiput and vertex, and dark brown frons and face medially (Figures 25,

- 26); metasoma black with small basal spot on tergite 1 and finger-shaped mid-basal spot on tergite 2 (Figure 27) **cacuangoi sp. n.**
- 23(12) Mesosoma and metasoma mostly honey brown–honey yellow (Figures 45, 58, 104), reddish brown (Figure 31), sometimes with dark marks on mesoscutum (Figure 105) **24**
- Propodeum and most of metasomal terga black to dark brown (Figures 7, 98, 111), or sometimes first tergite white (Figure 52, 109); hind coxa sometimes bicolored black and white (Figures 7, 98, 109) **27**
- 24(23) Hind wing vein 2-1A present (Figure 119); diameter of lateral ocellus at least 2× ocell–ocular distance (Figure 59) **25**
- Hind wing vein 2-1A absent; diameter of lateral ocellus roughly as long as ocell–ocular distance (Figure 46) **falloni sp. n.**
- 25(24) Hind wing marginal cell gradually widening toward apex, vein RS well pigmented; dorsal carina on first tergite meeting subbasally, enclosing a basal polished triangular area (Figures 36, 106) **26**
- Hind wing marginal cell narrowest point at middle, vein RS faint, difficult to trace (Figure 119); dorsal carina on first tergite meeting basally enclosing a semicircular basal area (Figure 37) **luteosicarius sp. n.**
- 26(25) Antenna dark brown to brown proximally, lightening gradually toward pale brown apex (Figure 104); wings uniformly weakly infusate; ocelli large, about 2× ocell–ocular distance (Figure 105)..... **stewarti sp. n.**
- Antenna black with white median annulus (Figure 31); wings weakly infusate with dark band just below stigma (Figure 31); ocelli extremely large, about 8× ocell–ocular distance (Figure 32)..... **colberti sp. n.**
- 27(23) Malar space moderately wide, at least slightly longer than width of mandibular base; median carina present on propodeum **28**
- Malar space short, length 0.7× width of mandibular base (Figure 1); median carina absent on propodeum..... **achlydis Townsend, 2009**
- 28(27) Mesoscutum and scutellum honey yellow, hind coxa bicolored black and white (Figures 7, 98, 109)..... **29**
- Mesoscutum and scutellum mostly black (Figures 52, 111); hind coxa one color, either black (Figure 52), yellow, or white (Figure 111) **31**
- 29(28) Head honey yellow, ocellar triangle dark brown (Figure 99); fore wing vein 1M strongly curved basally (Figure 120) **30**
- Head dark brown, gena white (Figure 7); fore wing vein 1M almost straight or weakly and evenly curved (as in Figure 119) **albigena sp. n.**
- 30(29) First metasomal tergite about 2× longer than its apical width, dark brown to black (Figure 100); hind coxa basally white and apically black (Figures 98, 100)..... **shakirae sp. n.**
- First metasomal tergite about as long as apical width, white with small black spot mid-apically; hind coxa black basally and apically white (Figure 109) ...
..... **townsendi sp. n.**

- 31(28) First metasomal tergite white, contrasting to remainder mostly dark brown metasoma (Figure 103); pronotum mostly black; mesopleuron dark brown at least on anterior corner (Figure 52).....32
- First metasomal tergite dark brown; pronotum white with brown mark dorsally; mesopleuron entirely yellow (Figure 111) *tzantza* sp. n.
- 32(31) Mesopleuron, metapleuron, hind coxa and propodeum medially and anteriorly smooth; mesopleuron except for anterior corner and hind coxa orangish.....
- *speciosus* Townsend, 2009
- Mesopleuron, metapleuron, hind coxa and propodeum granulate; mesopleuron and hind coxa black (Figure 52) *kingmani* sp. n.

Systematics

Aleiodes aclydis Townsend, 2009

http://species-id.net/wiki/Aleiodes_aclydis

Figure 1

Diagnosis. Body length 6.1 mm; antenna with 44 segments; head with vertex black, occiput light orangish brown; ocelli large, ocell–ocular distance less than width of lateral ocellus; occipital carina interrupted at vertex; mesosoma mostly light orangish brown, except propodeum black; wings slightly darkened; mesopleuron granulate; apex of hind tibia without comb of modified setae; propodeum without median propodeal carina; metasomal terga entirely black; metasomal tergum 3 costate on anterior 2/3, with median carina along with this sculpturing; ovipositor short, about 0.25× length of hind basitarsus.

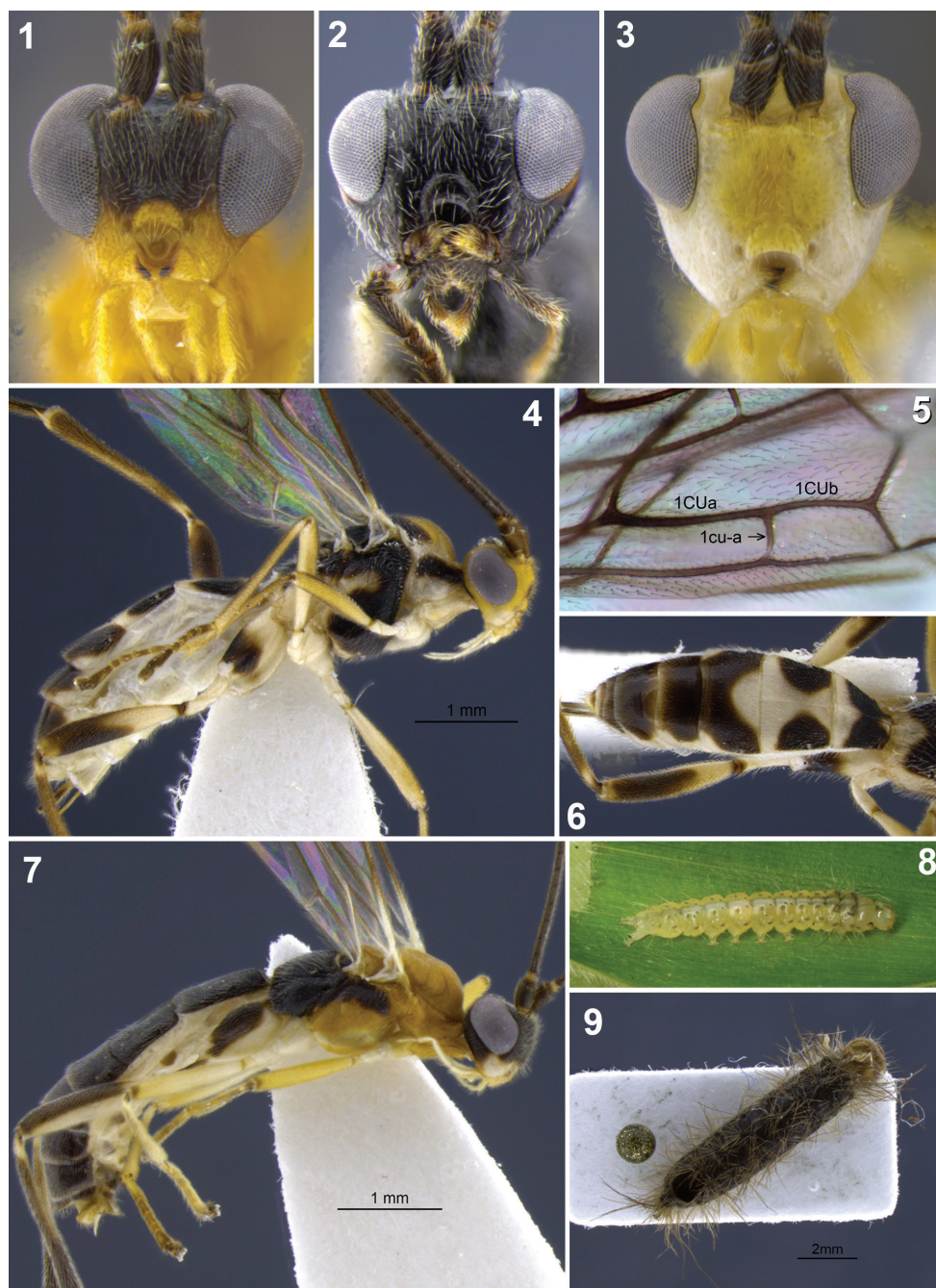
Additional characters. Last flagellomere with “bottle nipple”-like tip; mesoscutum with carina only in front of scutellar sulcus; scutellar sulcus with complete median carina plus two pairs of weak and incomplete lateral carina; fore wing vein 1M only slightly curved at base; hind wing vein 2-1A absent, vein m-cu present and well pigmented, antefurcal to r-m in left wing and interstitial in the right wing; ovipositor sheaths about as long as hind tarsomere II, 0.6× hind basitarsus.

Type material examined. (UWIM)

Biology. *Aleiodes aclydis* has been reared from an unidentified Geometridae found on *Ocotea* sp. (Lauraceae).

Distribution. Known only from the type locality, Isla de Las Palmas, Napo province, ECUADOR, at 1,883 meters elevation.

Discussion. From the newly described species, *Aleiodes aclydis* is very similar to *A. albigena* sp. n. in color features, including a mostly blackish head with a lighter gena. However, in *aclydis* the gena is yellow instead of the white gena of *albigena* sp. n. Both species belong to *circumscriptus/gastritor* species-group but *aclydis* is the unique species with a malar space as short as 0.7 times the basal width of mandi-



Figures 1–9. 1–3 head, frontal. 1 *Aleiodes aclydis* Townsend 2 *Aleiodes capillosus* Townsend; *Aleiodes marilynnae* sp. n. 4–6 *Aleiodes albidactyl* sp. n. 4 habitus 5 fore wing, detail of 1st subdiscal and subbasal cells 6 metasoma, dorsal 7–9 *Aleiodes albigena* sp. n. 7 habitus 8 host larva, nr. *Desmotricha* (Erebidae) 9 host mummy after parasitoid emergence.

bles. No additional *A. aclydis* specimens were found since Townsend and Shaw's (2009) work. The dorso-medially elevated area on mesopleuron is well-demarcated posteriorly. In the original description the sulcus demarcating this region is called the "sternaulus" (Townsend and Shaw 2009). We now consider that this sulcus is not a true sternaulus, as defined by Sharkey and Wharton 1997, and neither is it the precoxal sulcus, as defined by other authors (van Achterberg 1991, Wharton 2006). Additionally, the absence of sternaulus is a common feature in all species treated in this work. Therefore, this term is avoided in the descriptions of the new species. The same matter is found in the descriptions of *Aleiodes atripileatus* Townsend and *Aleiodes nebulosus* Townsend.

***Aleiodes albidactyl* sp. n.**

<http://zoobank.org/4969BD1E-3D57-4EC5-B5A0-F6B097D0148C>

http://species-id.net/wiki/Aleiodes_albidactyl

Figures 4–6

Description of holotype. Female (holotype). Body length 5.2 mm; antenna length 6.6 mm; fore wing length 4.9 mm.

Color. Distinctive black and white color pattern. Head mostly pale yellow with small dark brown spot just above clypeus; occiput laterally and stemmaticum dark brown. Antenna dark brown, scape and pedicel brown. Mesosoma mostly black; mesoscutum mostly pale yellow with postero-lateral borders dark brown; metapleuron, propodeum posteriorly based triangular area, thumb shaped area posteriorly on mesopleuron, ventral half of pronotum, and propleuron white. Metasoma mostly black dorsally and completely white ventrally; T1 apical 1/3 and small mid-basal spot white; T2 mostly white with two round antero-lateral spots; T3 with white semicircular basal spot; lateral borders of remainder terga white; ovipositor sheaths basal 1/5 white, remainder dark brown. Fore and mid legs pale light brown. Hind legs brown; basal half of femur and coxa, trochanter and trochantellus white. Wings very weakly infusate, veins dark brown.

Head. Antenna 50 antennomeres, apical flagellomere with "bottle-nipple"-shaped apex; malar space slightly longer than basal width of mandible, 0.4× eye height; in dorsal view eyes 2.75× longer than temples; occipital carina incomplete, directed toward vertex, getting close to lateral ocelli, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus not swollen; ocell-ocular distance as long as diameter of lateral ocellus; maxillary palp not swollen; head surface sculpture shining granulate, occiput smooth and shining; vertex with few wrinkles; higher face with few transverse rugosity; frons smooth with pairs of wrinkles concentric to each toruli, frons excavated with excavation bordered by a "W-shaped" carina.

Mesosoma. Sculpturing granulate; pronotum smooth latero-ventrally, latero-dorsally foveate; mesopleuron with antero-dorsal corner rugose, mid-longitudinally

depressed area foveate; propodeum coarsely shining granular with complete mid-longitudinal carina; notauli very shallow anteriorly, virtually absent; posterior margin of mesoscutum with complete carina; scutellar sulcus with five carina, relatively strong but irregular and not reaching anterior margin of sulcus.

Wings. Fore wing: stigma $3.4\times$ longer than high; vein r about as long as vein 2RS, $1.5\times$ longer than vein RS+Mb, and $0.8\times$ vein m-cu; vein 3RSa $0.43\times$ vein 3RSb, and $0.9\times$ vein 2M; vein 1CUa $3.3\times$ vein 1cu-a; vein 1CUB $0.7\times$ vein 1CUa; vein 1M slightly and evenly curved. Hind wing: vein m-cu only very weakly indicated; vein M+CU $1.8\times$ longer than vein 1M; 1M slightly shorter than r-m; RS smoothly curved at middle; vein 1M straight, almost unpigmented; vein 2-1A absent.

Legs. Hind tibia with comb of modified setae; tarsal claws simple, with a comb of relatively long thin setae basally; hind basitarsus $3.6\times$ longer than inner apical spur on hind tibia.

Metasoma. T1, T2 and basal 2/5 of T3 granulose-striate, longitudinal carina present along this sculpturing; remainder visible terga granular coriaceous; ovipositor sheaths about as long as hind tarsomere II; ovipositor sheaths parallel sided with lanceolate apex; T1 about as long as its apical width.

Male unknown.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, August 17, 2006, A. Townsend.

Type-specimen: Holotype female, point mounted. Top label: "ECUADOR: Napo Prov. / Yanayacu Biological Station / 17-Aug-2006 Macucoloma trail / Yellow pan 2163m. A. Townsend" (UWIM).

Discussion. This species belongs to *seriatus* species-group. It differs from all other species of this group by the white posterior thumb like marking on the black mesopleuron, and the long vein 1CUa on fore wing, about $1.5\times$ longer than 1CUB and $3.5\times$ vein 1cu-a. This species is most closely related to *dyeri* sp. n., but the color patterns of both species are quite distinct (see comments for *dyeri* sp. n.).

Etymology. From the Latin roots meaning "white finger", a reference to the white thumb like mark on mesopleuron.

***Aleiodes albigena* sp. n.**

<http://zoobank.org/E47103FE-3B7C-4AF0-8646-C67E62D7E968>

http://species-id.net/wiki/Aleiodes_albigena

Figures 7–9

Description of holotype. Male (holotype). Body length 5.2 mm; antenna length 7.0 mm; fore wing length 5.0 mm.

Color. Mostly black. Head and antenna dark brown; gena, mandibles and palp white, mandible tips dark brown. Pronotum, propleuron, mesoscutum, most of mesopleuron and scutellum honey yellow; metanotum, propodeum, metapleuron and dor-

sal 1/4 of mesopleuron, including the border with metapleuron, black; mesopleuron with whitish longitudinal stripe. Legs: all coxa, trochanter and trochantellus, mid femur and tibia, and most hind femur white; fore femur yellowish; fore tibia and tarsi and mid tarsi light brown; hind tibia and tarsi dark brown to black, but tibia with white basal band and fourth and fifth tarsi lighter; hind coxa, trochanter, trochantellus and femur black dorsally, except basal 1/5 of femur; tip of fore and mid femur and mid tibia with infuscate stains. Metasoma dark brown to black dorsally, laterally and ventrally white, but some lateral spots and the last sternites dark brown.

Head. 46 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with short pointed apex; malar space moderate, length 1.4× basal width of mandible, and 0.4× eye height; in dorsal view eye height about 3× temples; occipital carina complete, ventrally touching hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus not swollen; ocell-ocular distance 0.6× diameter of lateral ocellus; maxillary palp not swollen. Face somewhat bulging, frons barely excavated and without lateral ridges. Head surface sculpturing, including frons, granulate, face coarsely granulate, occiput smooth and shining.

Mesosoma. Sculpturing mostly granulate; pronotum with few laterally running wrinkles laterally; mesopleuron rugose on dorso-anterior corner; propodeum rugose-granulate, with complete mid-longitudinal carina; notauli very shallowly indicated anteriorly, posteriorly disappearing in a depressed rugose area; posterior margin of mesoscutum with very short carina, just anterior to scutellar sulcus; scutellar sulcus with five short carina, not reaching anterior margin of sulcus.

Wings. Fore wing: stigma 5× longer than high; vein r 0.5× vein 2RS, about 0.64× vein RS+Mb, and 0.56× as long as vein m-cu; vein 3RSa about 0.4 times vein 3RSb, and as long as vein 2M; vein 1CUa 2.7× vein 1cu-a; vein 1Cub about 2× vein 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu indicated as short nebulous vein interstitial to vein r-m; vein M+CU as long as vein 1M; vein 1M 1.5× vein r-m; RS smoothly curved at middle; vein 2-1A present as a short stub.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, with a comb of thin bristles medially; hind basitarsus 2.5× length of inner apical spur of hind tibia.

Metasoma. T1–T3 granular-striated; remainder terga granular; mid longitudinal carina complete from T1 throughout T2; T1 length 1.3× its apical width.

Female. Unknown.

Mummy. Length 9.0 mm, body reddish brown, head yellowish, setae pale brown, covered with setae, thorax compact and wrinkled, glue hole located ventrally on the thorax, exit hole irregular, located postero-dorsally, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, S00°35.9', W77°53.4', 2163 m, cloud forest, May 14, 2010

Type-specimen: Holotype female and mummy, point mounted. Top label: "ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA – NSF-BSI-07-17458 / (hand written) April 2010 / 47082", back (hand written): "14-MAY-2010"; bottom label (hand written): "coll. 8 April 2010 / YY road, Chusquea / Arctiidae, inst. 2 / Beat 650 / pup. 23 April / em. 14 May 2010" (UWIM).

Biology. Reared from a species near *Desmotricha* Hampson (Erebidae) larvae (voucher number YY-47082), feeding on *Chusquea scandens* (Poaceae). Parasitoid took three weeks to emerge after host mummification. This is the only described *circumscriptus/gastritor*-group species in Neotropical region known to attack Arctiinae hosts, producing a densely setose mummy.

Discussion. *Aleiodes albigena* sp. n. belongs to *circumscriptus/gastritor* species-group. Some diagnostic characters are the very shallow notauli present only anteriorly; scutellar sulcus without bisecting carina; occipital carina complete dorsally; face bulging; and frons surface granular, not excavated, without a lateral carina. This species is similar to *townsendi* sp. n. and *shakirae* sp. n., but it can be distinguished from both by the mostly black head with contrasting whitish gena, yellowish in *townsendi* sp. n. and *shakirae* sp. n., and the almost straight vein 1M on fore wing, strongly curved in *townsendi* sp. n. and *shakirae* sp. n. *A. albigena* sp. n. resembles *A. arbitrium* in color pattern but differs in the shorter ocell–ocular distance relative to lateral ocelli diameter.

Etymology. From the Latin roots meaning “white cheeks,” named in reference to the contrasting white gena, as compared with the black head of this species.

Aleiodes albiterminus Townsend, 2009

http://species-id.net/wiki/Aleiodes_albiterminus

Figure 10

Diagnosis. Body length 4.4 mm; antenna with 33 segments; head color mostly black, with light orangish brown mark below eye; malar space 1.25× basal width of mandible, ocelli small, ocell–ocular distance 1.3× width of lateral ocellus; occipital carina interrupted at vertex; mesopleuron granulate; apex of hind tibia without comb of flattened setae; propodeum mostly granulate, with median propodeal carina present but incomplete; metasomal tergum 3 black basally, apically with irregular “half heart-shaped” off-white spot; ovipositor sheaths long, 1.3× length of hind basitarsus.

Additional characters. Last flagellomere lanceolate; mesoscutum with complete carina on posterior margin though not well defined; scutellar sulcus with a strong complete median carina and some irregular carina laterally; fore wing vein 1M slightly and evenly curved; hind wing vein 2-1A present as a very short stub, vein m-cu present and distinctly postfurcal to vein r-m; ovipositor sheaths 1.3× longer than hind basitarsus.

Type material examined. (UWIM)

Biology. *Aleiodes albiterminus* has been reared from an unidentified Geometridae larva on *Alnus acuminata* (Betulaceae).

Distribution. Known only from the type locality, Rio Chalpi Grande, Napo province, ECUADOR, at 2,837 meters elevation.

Discussion. For standardization reasons we provide the measurement of the ovipositor sheaths instead of the ovipositor itself. The ovipositor and the sheaths length in *A. albiterminus* are unusually long for *Aleiodes* species. This is the only species from Ecuadorian Northeastern Andes with the ovipositor sheaths longer than

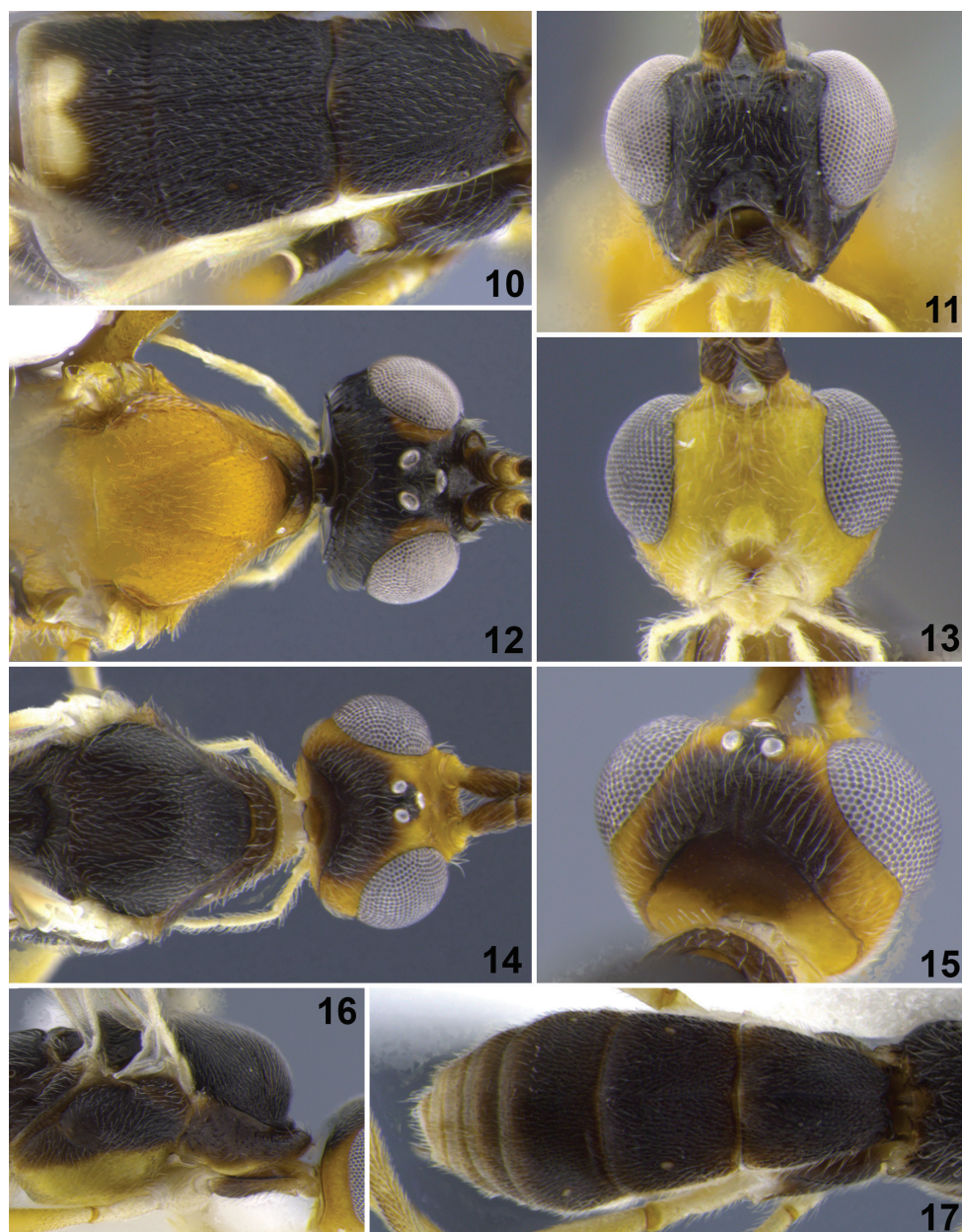


Figure 10–17. 10 *Aleiodes albiterminus* Townsend, metasoma, dorsal. 11, 12 *Aleiodes arbitrium* Townsend 11 head frontal 12 head and mesoscutum, dorsal 13–17 *Aleiodes atripileatus* Townsend 13 head, frontal 14 head and mesoscutum, dorsal 15 head, occiput and vertex 16 thorax, lateral 17 metasoma, dorsal.

its hind basitarsus. *A. albiterminus* can be distinguished also by the absence of a median carina on the first to third metasomal terga, and the distinctive off-white markings at the apex of metasomal tergum 3.

***Aleiodes albiviria* sp. n.**

<http://zoobank.org/099A4E94-7013-4B08-930B-1DB134C7EB3E>

http://species-id.net/wiki/Aleiodes_albiviria

Figures 18–21

Description of holotype. Female (holotype). Body length 7.1 mm; antenna length 9.0 mm; fore wing length 5.8 mm.

Color. Mostly yellowish “bronze” (orangish yellow). Ocellar triangle brown. Antenna black; scape yellowish ventrally; flagellum with white band medially from flagellomeres 18–19th to 29–30th. Brown regions of mesosoma: mesoscutum lateral borders, notauli region and depressed posterior area; lunules; metanotum; propodeum. Dorsal mesopleuron and metasoma ventrally pale yellow. Wings tinged honey yellow, veins honey brown.

Head. Antenna with 57 antennomeres, mid flagellomere roughly 2.0× longer than wide, apical flagellomere with small pointed apex; malar space moderate, about 1.25× longer than basal width of mandible, 0.33× eye height; in dorsal view eyes 3× longer than temples; occipital carina incomplete, directed toward vertex, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus not swollen; ocelli moderate, ocell–ocular distance slightly shorter than diameter of lateral ocellus; maxillary palp not swollen; head surface sculpture finely shining granulate, occiput smooth and shining; higher face with a small longitudinal ridge and transverse rugosity directed to it; frons excavated, excavation bordered by a weak “W-shaped” carina.

Mesosoma. Sculpturing shining granulate; pronotum foveate laterally; mesopleuron with antero-dorsal corner rugose; propodeum granular with some longitudinal diverging wrinkles laterally, mid-longitudinal carina present only on anterior 1/3; notauli with few crenulae and shallow anteriorly, meeting on depressed rugose area posteriorly; posterior margin of mesoscutum with complete carina; scutellar sulcus with median carina plus two pairs of complete lateral carina and one irregular.

Wings. Fore wing: stigma 3.8× longer than high; vein r as long as vein 2RS, 1.2× longer than vein RS+Mb, and 0.78× vein m-cu; vein 3RSa 0.45× vein 3RSb, and 0.9× vein 2M; vein 1CUa 2.7× vein 1cu-a; vein 1CUB about as long as vein 1CUa; vein 1M weakly curved basally. Hind wing: vein m-cu present, antefurcal to r-m; vein M+CU about 1.6× 1M; vein 1M almost as long as r-m; vein RS gradually opening from wing margin; vein 1M straight, dark brown, well pigmented; vein 2-1A absent.

Legs. Hind tibia with comb of modified setae; tarsal claw simple, with a comb of relatively long thin setae basally; hind basitarsus 3.5× longer than inner apical spur of hind tibia.

Metasoma. T1, T2 and basal 2/5 of T3 costate, longitudinal carina present along this sculpturing; remainder visible terga smooth; ovipositor sheaths short and lanceolate, about as long as hind tarsomere IV (half length of tarsomere II); T1 1.4× longer than its apical width.



Figures 18–20. *Aleiodes albiviria* sp. n. **18** habitus **19** head and mesonotum, dorsal **20** hind tibia inner side with comb of modified setae at apex.

Male unknown.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, April 1–8, 2007, J. Simbaña col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 1-8 April 2007, J. Simbaña /Macucoloma trail, Malaise trap / CAPEA - NSF-BSI-07-17458, S.R. Shaw”; bottom label: “SRS - 00043” (UWIM).

Discussion. This species is assigned to the *seriatus* species-group, where it most resembles *Aleiodes greeneyi* Townsend because of the dorsally incomplete occipital carina. *Aleiodes albiviria* sp. n. differs from other New World species of this species group by the mostly honey brown body with dark brown notauli and mid-posterior mesoscutum, and the white middle band on blackish antenna. *A. albiviria* sp. n. also resembles the Brazilian *Aleiodes scriptus* (Enderlein, 1920), by the costate sculpturing on metasomal tergite 1, which is rugose–costate in all other Neotropical species in *seriatus* species-group, but differs from *scriptus* by the shape of hind wing vein RS (parallel to wing margin basally and bent downward apically, as opposed to sinuate at middle in *scriptus*).

Etymology. From the Latin *albus*, meaning “white,” and *viria* meaning “bracelet,” a reference to the white band on the antenna.

***Aleiodes arbitrium* Townsend, 2009**

http://species-id.net/wiki/Aleiodes_arbitrium

Figures 11, 12

Diagnosis. Body length 5.7 mm; antenna with 43 segments; head mostly black; malar space slightly wider than basal width of mandible; ocelli small, ocell–ocular distance slightly wider than width of lateral ocellus; occipital carina at least shortly interrupted at vertex; mesosoma mostly light yellowish brown, except propodeum black; wings clear; mesopleuron granulate; apex of hind tibia without comb of flattened setae; propodeum rugulose basally, granulate apically, with median propodeal carina present; metasomal terga mostly black, except terga 2 and 3 often each with an oval-shaped off white marking; metasomal terga 1–3 costate, median carina complete to end of tergum 2; ovipositor sheaths slightly shorter than half of hind basitarsus.

Additional characters. Last flagellomere with short pointed tip; mesoscutum with carina on posterior margin only in front of scutellar sulcus; scutellar sulcus with a median carina barely complete plus two or three more or less weak and incomplete carina laterally; fore wing vein 1M slightly and evenly curved; hind wing vein 2-1A present as a very short stub, vein m-cu position varying from distinctly antefurcal to interstitial to vein r-m; ovipositor sheaths 0.45 to 0.49 times longer than hind basitarsus.

Morphological variation. All specimens examined agree well with original description of *Aleiodes arbitrium*, however the occipital carina in all specimens is at least shortly interrupted at vertex, and not complete but weak at vertex as described originally. The two spots on terga vary in size. In some females it is darker, not so contrasting, while in others it is larger, with the T2 spot reaching apical margin of tergite. In some males the spots tends to meet and form one single irregular elongate spot. In all specimens the gena is black–dark brown but the borders with mandibles are light brown.

Material. Type material examined. (UWIM)

Non-type material: 14 females and 9 males, ECUADOR, Napo, Yanayacu Biological Station, Napo Province, S00°35.9', W77°53.4', 2,163 m. 7 females and 6 males reared from Geometridae: ♂, YY-29185; ♀, YY-36318; ♂, YY-39980; ♂, YY-39990; ♀, YY-47187; ♀, YY-47708; ♀, YY-48161; ♂, YY-49147; ♂, YY-53729; ♂, YY-57356; ♀, YY-62371; ♀, YY-70773; ♀, YY-71345. 5 females collected by Malaise trap; 2 females and 3 males collected by yellow pan trap. (UWIM)

Biological additions. All new *Aleiodes arbitrium* Townsend rearings were from caterpillars feeding on polypod ferns: *Diplazium costale* (Dryopteridaceae) or *Dennstaedtia cornuta* (Dennstaedtiaceae). Most caterpillars were identified as Geometridae. Caterpillar pictures and mummy morphology corroborate family level identification, and two different species of *Psaliodes* Guenée (Geometridae), and one species of Pyralidae as hosts of *A. arbitrium*.

The holotype and eight non-type specimens were reared from *Psaliodes* sp. larva feeding on *D. costale* or *D. cornuta*. The two paratypes and four other non-type specimens were reared from *Psaliodes castanea* (Warren) feeding on *D. costale*. A single non-type specimen

was reared from unknown Pyralidae larva on *D. cornuta*. One paratype collected from Urticaceae did not feed, so may have simply wandered there prior to mummification.

Distribution. Known only from the Yanayacu Biological Station, Napo province, ECUADOR, at 2,163 meters elevation.

Discussion. After publication of Townsend and Shaw (2009) paper, 11 new specimens were reared and ten collected by traps, all from Yanayacu Biological Station, Napo Province, S00°35.9', W77°53.4', 2163. This additional reared material provided good biological information at least on feeding preferences of the host caterpillars. It is likely that *A. arbitrium* attacks mostly Geometridae species feeding on polypod ferns. In the *circumscriptus/gastritor* group, this species resembles *A. albigena* sp. n. in most color patterns; however, the smaller ocelli of *A. arbitrium* and the honey brown marks on vertex, absent in *A. albigena*, distinguish these two species. Other species with small ocelli and interrupted occipital carina, as *A. onyx* sp. n. and *A. atripileatus*, have more extensively dark bodies than *arbitrium*.

***Aleiodes atripileatus* Townsend, 2009**

http://species-id.net/wiki/Aleiodes_atripileatus

Figures 13–17, 21

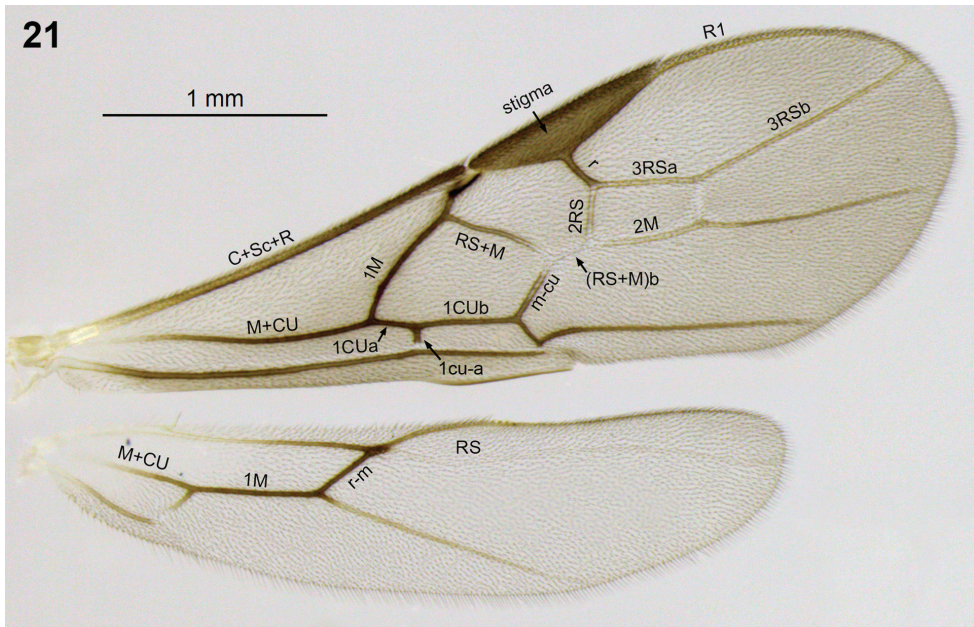
Diagnosis. Body length 4.1–4.8 mm; antenna with 36–39 segments; head color black dorsally, light orangish brown ventrally; malar space 1.5× basal width of mandible, ocelli small, ocell–ocular distance 1.7× width of lateral ocellus; occipital carina interrupted at vertex; mesopleuron granulate; apex of hind tibia without comb of flattened setae; propodeum mostly rugulose-granulate, with complete median carina; metasomal terga 1–4 entirely dark brown to black; ovipositor sheaths about 2/3 of the hind basitarsus length.

Additional characters. Last flagellomere with short “bottle-nipple”-like tip; mesoscutum posterior margin with carina only in front of scutellar sulcus; scutellar sulcus with a median plus two or three pairs lateral carina well defined and almost complete; fore wing vein 1M only slightly and evenly curved; hind wing vein 2-1A absent, vein m-cu absent; ovipositor sheaths about 2/3 length of hind basitarsus, one specimen in type series have 2.5/3, and another 1.8/3 proportions for this character.

Material. Type material examined. (UWIM)

Non-type material: 3 females and 2 undetermined sex (?), ECUADOR, Napo, Yanayacu Biological Station, Napo Province, S00°35.9', W77°53.4', 2163 m; voucher numbers: ♀, YY-28538: prob. Noctuidae (*Hypena* sp2) on *Phenax rugosus* (Urticaceae); ♀, YY-29079: *Hypena* sp2 (Noctuidae) on *Boehmeria ulmifolia* (Urticaceae); ♀, YY-37062: Noctuidae on *Phenax rugosus* (Urticaceae); ?, YY-58626: Noctuidae on *Phenax rugosus* (Urticaceae); ?, YY-63838: Noctuidae on *Phenax rugosus* (Urticaceae). (UWIM).

Biology. *Aleiodes atripileatus* has been reared from a species of *Hypena* Schrank (Noctuidae) caterpillars feeding on Urticaceae, including *Phenax rugosus*, *Boehmeria*



Figures 21. *Aleiodes atripileatus* Townsend wings.

bullata, *Miriocarpa* sp., and three other unidentified urticaceous plants. Other noctuids on the same host plants might be utilized as hosts.

Distribution. Known only from the YBS, Napo province, ECUADOR, at 2,163 meters elevation.

Discussion. *Aleiodes atripileatus* is one of the most commonly reared *Aleiodes* species in YBS. The species belongs to the *circumscriptus/gastritor* species-group. It is very similar to *A. nubicola* sp. n. and *A. cacuangoi* sp. n., differing from these two species in having the occipital carina distinctly interrupted at vertex. This character is shared with *A. onyx* sp. n., from which it differs by having a black vertex, a thinner metasoma, and the vein m-cu of hind wing absent. Different than originally described, the true sternaulus is absent.

***Aleiodes bimaculatus* sp. n.**

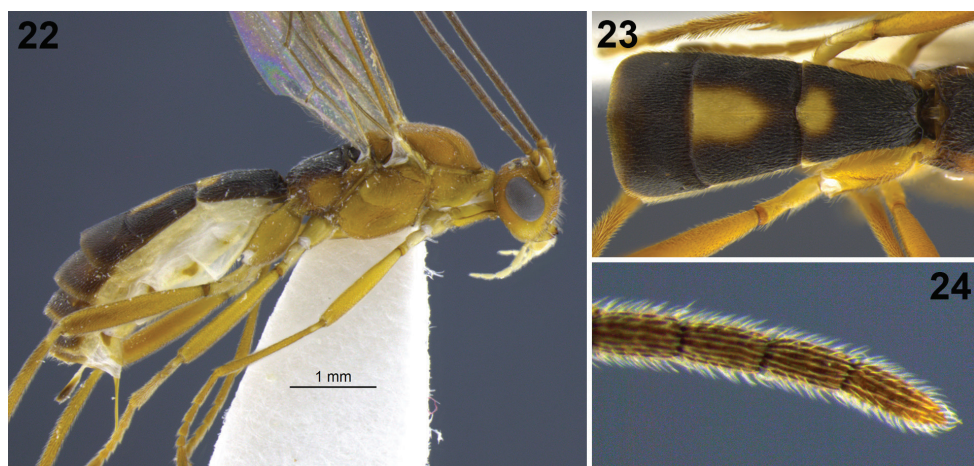
<http://zoobank.org/65E1286E-44D3-48DF-A403-F895A9D504A1>

http://species-id.net/wiki/Aleiodes_bimaculatus

Figures 22–24

Description of holotype. Female (holotype). Body length 5.6 mm; antenna length 6.4 mm; fore wing 5.4 mm.

Color. Mostly honey brown. Antenna brown, scape and pedicel honey brown as head; cheeks and palp light yellow; ocellar triangle brown; fore and mid coxa, and all



Figures 22–24. *Aleiodes bimaculatus* sp. n. **22** habitus **23** metasomal terga 1–3, dorsal **24** apical flagellomeres.

trochanter and trochantellus whitish; metanotum and propodeum dark brown; metasoma dark brown dorsally with mid-apical pale yellow spots on T1 and T2; ovipositor sheaths brown. Wings hyaline with brown veins and stigma, parastigma contrasting darker–black.

Head. Antenna 44 segmented, apical flagellomere lanceolate, without pointed tip; malar space about $1.3\times$ longer than mandible basal width, and $0.6\times$ eye height; occipital carina complete; oral space moderate and circular, maximum width about equal to basal width of mandible; clypeus swollen; ocelli small, ocell–ocular distance $1.4\times$ diameter of lateral ocellus; in dorsal view temples almost as long as eye height; head sculpturing mostly granular, face coarsely granular to rugose, occiput smooth.

Mesosoma. Sculpturing mostly granulate; pronotum covered with wrinkles; mesopleuron mostly rugose otherwise granulate, wrinkles stronger on sternaulus area; metapleuron with rugosity posteriorly; propodeum rugose, with complete mid-longitudinal carina; notauli deep and mostly smooth anteriorly, with two or three crenulae, meeting on rugose area posteriorly; posterior margin of mesoscutum bordered by short carina just anterior to scutellar sulcus; scutellar sulcus with median carina plus one pair of lateral carina.

Wings. Fore wing: stigma $3.5\times$ longer than high; vein r $0.65\times$ vein 2RS, $1.2\times$ vein RS+Mb, and $0.7\times$ as long as vein m-cu; vein 3RSa about $0.5\times$ vein 3RSb, and as long as vein 2M; vein 1CUa $1.5\times$ vein 1cu-a; vein 1CUB $3.0\times$ vein 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu indicated as short not tubular vein interstitial to vein r-m; vein M+CU $1.4\times$ longer than vein 1M; vein 1M $1.2\times$ vein r-m; RS smoothly curved at middle; vein M unpigmented; vein 2-1A present as a short stub.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, with a comb of relatively long thin setae basally; hind coxa rugose dorsally; hind basitarsus $3.5\times$ longer than inner apical spur on hind tibia.

Metasoma. T1 and T2 rugose costate with granulate background; T3 longitudinally striate basally; remainder terga granulate coriaceous; mid-longitudinal carina complete from T1 throughout T2; ovipositor sheaths about as long as hind tarsomere II, T1 1.2× longer than its apical width.

Paratype variation. Body length 5.0–6.0 mm; antennomeres 39–46; occipital carina is only very shortly interrupted in some paratypes, but never curved toward vertex; vein 2-1A of hind wing varying from short to absent. The paratypes from outside the YBS are distinctly smaller (body length 5.0–5.3 mm) than the type specimens from the YBS (5.4–6.0 mm), with fewer segments on antenna (39–40 *vs.* 43–46). The metasoma in this specimens is lighter than the holotype and paratypes from YBS: the apical terga beyond T3 are mostly honey yellow, and the spots on T1 and T2 are frequently larger, forming one large spot covering apical T1 and all T2 medially. We consider the specimens from Baeza as a geographical variant within *bimaculatus* sp. n. Since all but one females were collected at once and shows virtually none variation, the variation could be just an artifact. Further samplings could both confirm this hypothesis with some intermediate forms or support an alternative hypothesis (e.g. speciation process). The metasoma in one of the females from Manabí is almost entirely honey brown, the light spots are not contrasting but still visible.

Male. Body length 4.7–5.0 mm. Antennomeres 48–42. Considerable color pattern variation in males: antenna dark brown, pedicel brown, scape honey brown, face brown, hind coxa whitish, all tibia and tarsi darker, ocellar triangle black, metanotum dark brown as propodeum, T2 pale yellow spot varying from smaller than in female to covering most of the tergite, T3 also with pale yellow spot. The metasoma is narrower, T1 up to 1.4× longer than its apical width; eyes in dorsal view 1.55× longer than temple; occipital carina weak dorsally, barely interrupted at vertex. The males from Baeza follow the same pattern of the females, with the apical metasomal terga honey brown instead of dark brown, however the body length in these males is not distinctly shorter than the specimens from YBS.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, January 1–8, 2008, J. Simbaña col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 1-8 January 2007, J. Simbaña / Macucoloma trail, Malaise trap / NSF-BSI-07-17458, S.R. Shaw”; bottom label: “SRS - 00037”. (UWIM)

Paratypes. 11 females and 2 males (UWIM), same locality as holotype, different date and/or method: 1♀, September 5, 2005, malaise trap (Pumayacu ridge); 1♀, March 1–6, 2006, hand collected at light sheet, G. Gentry col; 2♀, same data as holotype; 1♀, no date/method; 2♀, June 1–8, 2007, malaise trap; 1♀, June 10 – July 10, 2010, canopy malaise on bamboo, S.R. Shaw col; 1♀, December 3–10, 2007, malaise trap; 1♀, October 3–10, 2007, malaise trap; 1♀, August 3–10, 2007, yellow pan; 2♂ May 14, 2011, black light, N. Zikani col.; 2♂ May 10–20, 2011, gate pan trap, M. Bryant col. ECUADOR, 12 females and 7 males (CNC): 9♀ and 1♂, Napo, Baeza,

2000m, February 1979, Mason; 1♀ and 3♂, Napo, 5km South Baeza, 1700m, February 9, 1983, Masner & Sharkey; 3♂, Napo, Baeza, 1900m, February 9, 1983; 2♀ Manabi, Montecristi, 400m, February 6, 1983, Masner & Sharkey.

Discussion. *Aleiodes bimaculatus* sp. n. belongs to *circumscriptus/gastritor* species group. It resembles *nubicola* sp. n. and *cacuangoi* sp. n. because of the complete occipital carina and the small ocelli. *A. bimaculatus* sp. n. differs from these species by the honey brown body color, with propodeum and metasoma dark brown and two pale yellow spots on T1 and T2, while *nubicola* sp. n. and *cacuangoi* sp. n. are mostly black. The stigma is yellowish in *bimaculatus* sp. n., but brownish in *cacuangoi* sp. n. and light brown to whitish in *nubicola* sp. n., and the body larger, 5.0–6.0 mm length while *cacuangoi* sp. n. and *nubicola* sp. n. are shorter, with maximum body length of 4.6 mm.

Etymology. From the Latin roots *bi*=two and *macula*=stain, refers to the two distinctive yellow spots on the dark brown metasomal terga of this species.

***Aleiodes cacuangoi* sp. n.**

<http://zoobank.org/3A2C8D97-3E7C-4EF9-A86C-57C095F39946>

http://species-id.net/wiki/Aleiodes_cacuangoi

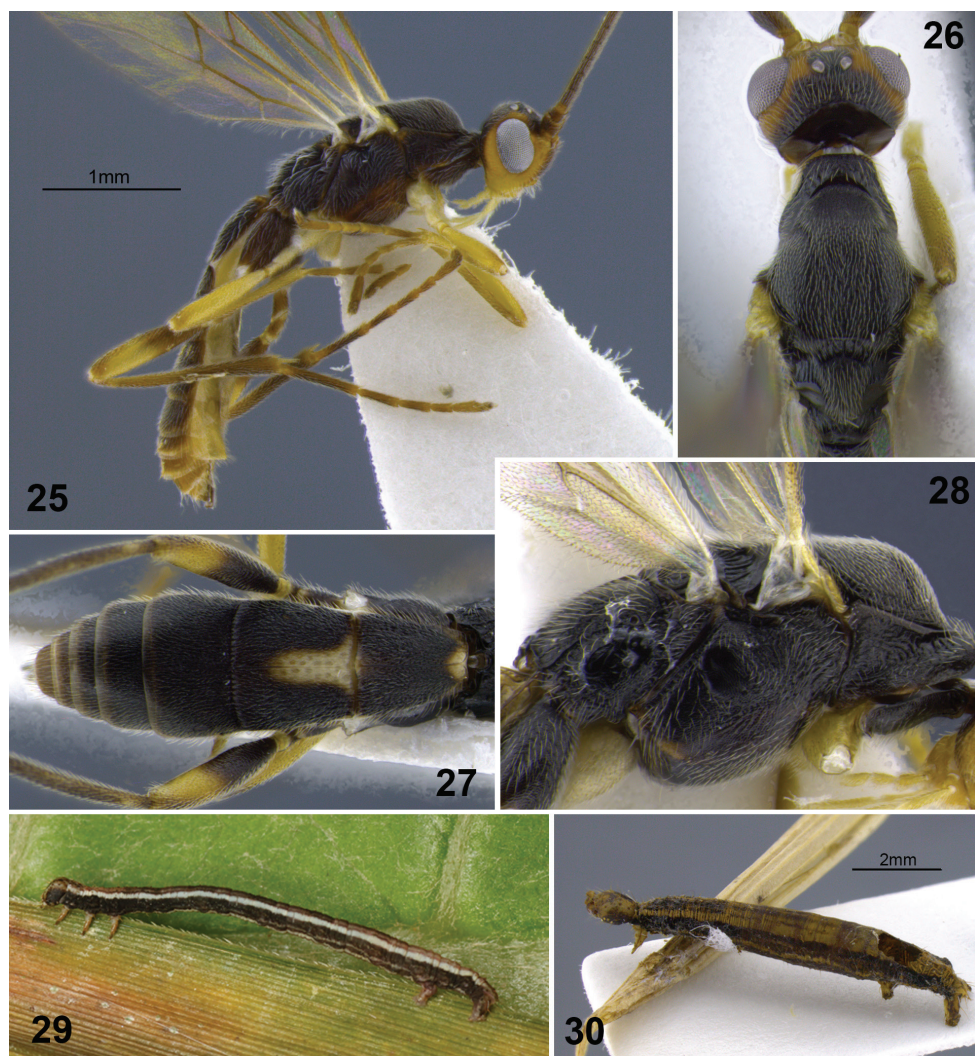
Figures 25–30

Description of holotype. Female (holotype). Body length: 4.77 mm; antenna length: 5.5 mm; fore wing length: 4.8 mm

Color. Mostly black. Face, gena, eyes orbits, and apical half of hind coxa yellowish brown; mandibles, gena at border with mandibles and palp whitish, mandibles tips (tooth) brown; fore and mid legs yellowish darkening distally; tarsi dark brown, hind tibial spurs yellowish; ventral metasoma light yellow; metasomal terga 1 and 2 with mid basal whitish marks, spot on tergite 1 small, on tergite 2 extending over basal half. Ovipositor sheaths dark brown on apical half, basal half whitish.

Head. Antenna comprising 43 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with short pointed apex; malar space moderate, length 1.6× basal width of mandible, and approximately 0.55× eye height; occipital carina complete and well defined, bordered by a more or less deep sulcus on temples and vertex, reaching hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus not swollen; ocelli small, ocell–ocular distance about 1.5× diameter of lateral ocellus; temples about 1.8× eye height in dorsal view; maxillary palp not swollen; head surface sculpturing mostly granulate, vertex more coarsely granulate, occiput very weakly shining coriaceous, apparently smooth.

Mesosoma. Sculpturing mostly granulate; pronotum rugose laterally, foveate dorsally; mesopleuron rugose anteriorly, with elevated area centrally smooth and shining, and some wrinkles centrally, otherwise coriaceous; propodeum rugose posteriorly, with long mid-longitudinal carina on anterior 2/3; notauli deep and renulated anteriorly, meeting a coarsely punctate area posteriorly; scutellar sulcus with well defined median carina and two pairs of lateral irregular carina.



Figures 25–30. *Aleiodes cacuangoi* sp. n. **25** habitus **26** head and mesonotum, dorsal **27** metasoma, dorsal **28** mesosoma, lateral **29** host larva (Geometridae) **30** host mummy after parasitoid emergence.

Wings. Fore wing: stigma $5\times$ longer than high; vein r as long as veins 2RS and RS+Mb, and $0.7\times$ as long as vein m-cu; vein 3RSa about $0.4\times$ vein 3RSb, about as long as 2M; vein 1CUa $1.7\times$ vein 1cu-a; vein 1CUB about $2.9\times$ 1CUa; vein 1M evenly curved. Hind wing: m-cu absent; vein M+CU $1.36\times$ 1M; vein r-m $1.5\times$ 1M; vein RS smoothly curved at middle; vein 2-1A absent.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, not pectinate, with a comb of relatively long thin setae basally; hind basitarsus about 3.4 times the length of inner apical spur on hind tibia.

Metasoma. T1 and T2 granulate and longitudinally striated; remainder terga granulate; mid longitudinal carina complete from T1 throughout T3; petiole length about $1.1 \times$ its apical width; ovipositor sheaths about as long as hind tarsomere II.

Paratype variation. Body length 4.2–4.6 mm; antennomeres 42–45; malar space about 1.4 times basal width of mandible; malar space/eye height = 0.53–0.60; ocell–ocular distance/diameter lateral ocellus ~ 1.4 –1.5; temple/eye height = 1.75–2.00; fore wing veins: $r/2RS \sim 0.75$ –1, $r/m-cu \sim 0.6$ –0.7, $r/RS+Mb = 0.75$ –1.00, $1CUb/1CUa = 2.4$ –2.9; hind wing veins: $1M/r-m \sim 1.40$ –1.55, $M+CU/1M = 1.15$ –1.40, in most paratype $m-cu$ weak but present, its position varying from just antefurcal to interstitial (frequently in the same individual); hind basitarsus/apical inner spur on hind tibia = 3.2–3.4; tergite 1 length/apical width = 1.1–1.2; ovipositor sheaths/hind tarsomere II ~ 0.8 –1.1. Color pattern: some paratypes have brown face medially, and/or a white stripe on mesopleuron; occipital carina somewhat irregular at vertex in few specimens; one or two pairs of lateral carina on scutellar sulcus, more or less irregular.

Male. Similar to females but body slightly smaller, antenna shorter, with 40–41 segments, metasoma slightly thinner, and ocelli relatively larger: ocell–ocular distance $1.25 \times$ diameter of lateral ocellus. In one specimen the occipital carina is shortly interrupted but the occiput and vertex limits are easily distinguishable. One male paratype have considerably darker head and metasoma: head brown to dark brown, gena at borders with mandibles pale yellow, middle face darker, clypeus and most of temples honey brown, white spots on metasoma reduced, on tergite 2 yellowish and covering less than half its length.

Mummy. Length 6.7–8.0 mm in males and 7.3–10.0 mm in females, mottled gray and yellowish, with lateral bands more or less defined mottled dark brown and brown, thorax compact and wrinkled, exit hole irregular, located postero-dorsally between anal and abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Beat 10A, YY-58947, S00°35.9', W77°53.4', 2163 m, cloud forest, September 5, 2011.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: "ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA – NSF-BSI-07-17458 / (hand written) July 2011 / 58947; back (hand written): "em. 5 Sept 2011". (UWIM)

Paratypes (UWIM): 12 females and 8 males, same data as holotype, except different dates. *Females*: August 15, 2007, YY-22975; August 20, 2009 YY-40348; December 21, 2009, YY-43751; April 22, 2010, YY-46509; May 4, 2010, YY-46887; May 12, 2010, YY-47603; July 8, 2010, YY-48837; August 13, 2010, YY-49757; August 25, 2010, YY-50103; November 12, 2010, YY-52466; June 14, 2011, YY-56911; June 16, 2011, YY-56954. *Males*: December 21, 2009, YY-43737; April 1, 2010, YY-45833; April 19, 2010, YY-46665; May 17, 2010, YY-47596; August 7, 2010, YY-49712; August 9, 2010, YY-49716; August 27, 2010, YY-50307; June 14, 2011, YY-56894.

Biology. All specimens were reared from the same Geometridae host caterpillar morphospecies (“línea blanca en la espalda chusquea”) feeding on bamboo, *Chusquea scandens* (Poaceae). Consistent morphology of mummies and rearing data of caterpillars support a single host species for this parasitoid, most frequently sampled between July and August. Most host caterpillars collected were in the 3rd larval instar, but one was in 4th and one in 2nd instar. Time span between host mummification and adult emergence varying in days from: 19–35 for females, and 16–26 for males.

Discussion. This species belongs to *gastritor/circumscriptus* species-group. It is similar to *A. atripileatus*, differing from it by having the occipital carina complete (interrupted at vertex in *atripileatus*), smooth area on mesopleuron (granulate–coriaceous in *atripileatus*), 43 antenomeres (maximum 39 in *atripileatus*), longitudinal carina complete on T1–T3 (incomplete in *atripileatus*), and one basal light spot on T1 and one on T2 (absent in *atripileatus*). This species is also very similar to *A. nubicola* sp. n. The most evident character to distinguish these species being the color patterns on meso- and metasoma. *A. cacuangoi* sp. n. have a mostly black metasoma dorsally, with one small to tiny basal white spot on T1 and a finger like mid-basal white spot on T2, the size of the spot on T2 varies from 1/3 to 2/3 of the tergite length. In *nubicola* sp. n. the metasoma varies from entirely black to black with apical whitish spots, these spots are larger on apical terga and extends to T4 in females, but one male paratype has the spots extending from the apical terga throughout apex of T1. There are not apical spots in none of the terga in *cacuangoi* sp. n., while in *nubicola* sp. n. there is not basal white spot on T1. The mesosoma in *cacuangoi* sp. n. is entirely black except for some reddish brown–brown stripe on mesopleuron, which is distinctly lighter close to mid coxal insertion. *A. nubicola* sp. n. have similar color pattern but with a postero-median square on mesoscutum and scutellum medially orangish. In *cacuangoi* sp. n. the head is mostly yellowish with occiput and vertex medially always black, the face varies from pale yellow to dark brown, and temples and gena are pale yellow to brownish orange. In *nubicola* sp. n. the head is mostly dark brown with a crescent moon-shaped honey yellow area on temples, bordering eyes, and the color of gena is variable. Mummies of *atripileatus*, *cacuangoi* sp. n., and *nubicola* sp. n. are very distinctive: *cacuangoi* sp. n. mummies are mottled gray and yellowish with more or less defined dark brown lateral stripes, *atripileatus* mummies are black with extended anal prolegs and head yellow, and *nubicola* sp. n. mummies are entirely brown and gradually narrowing anteriorly.

Etymology. The species is named in honor to Dolores Cacuaño, for her pioneering, outstanding brave efforts for the indigenous rights in Ecuador.

Aleiodes capillosus Townsend, 2009

http://species-id.net/wiki/Aleiodes_capillosus

Figure 2

Diagnosis. Body length 5.6 mm; antenna with 42 segments; head color black, with small brown spot at gena; malar space 1.8× basal width of mandible, ocelli small, ocell–

ocular distance 2.0× width of lateral ocellus; occipital carina completely absent; mesopleuron mostly smooth, epicnemial carina absent; apex of hind tibia without comb of flattened setae; propodeum smooth, without median carina; metasomal tergite 1 white with a black oval spot medially, remainder terga mostly black, laterally white; ovipositor sheaths about 2/3 of the hind basitarsus length.

Additional characters. Last flagellomere lanceolate; mesoscutum with carina on posterior margin varying from present only in front of scutellar sulcus to completely absent; scutellar sulcus with one to five incomplete carina; fore wing vein 1M slightly to moderately curved at base; hind wing vein 2-1A present and relatively long, vein m-cu present and pigmented but never tubular, interstitial to vein r-m; ovipositor sheaths about 2/3 as long as hind basitarsus.

Morphological variation. The non-type material fits well in original description, but most of the mummies are considerably lighter.

Material. Type material examined. (UWIM)

Non-type material: 2 females and 2 males, ECUADOR, Napo, Yanayacu Biological Station, Napo Province, S00°35.9', W77°53.4', 2163 m; voucher numbers: ♀, YY-53697: Geometridae on *Piper* sp1 (Piperaceae) [probably collected as mummy]; ♂, YY-40671: Geometridae on *Alnus acuminata*; ♀, YY-40675: Geometridae on *Alnus acuminata*; ♂, YY-40655: Geometridae on *Alnus acuminata*.

Biology. *Aleiodes capillosus* has been reared from unidentified Geometridae caterpillars on *Diplazium vesiculosum* (Dryopteridaceae). The non-type specimens were also reared from Geometridae, but on *Alnus acuminata* (Betulaceae) and *Piper* sp. (Piperaceae). Feeding on these plant species by the geometrid caterpillars is not confirmed.

Distribution. Known only from the type locality, YBS, Napo province, ECUADOR, at 2,163 meters elevation.

Discussion. *Aleiodes capillosus* is very similar to *A. marilynae* sp. n. (for a discussion on distinguishing characters see discussion section under *A. marilynae* sp. n.). These are the only two species in the *gressitti* species-group from Neotropical Region. As well as the morphological similarities in the adult parasitoids, the host mummies of *A. capillosus* and *A. marilynae* are very similar. Both produce a relatively swollen mummy with a strongly shrunken thorax, where it is attached to a leaf or branch in a distinct angle.

Aleiodes colberti sp. n.

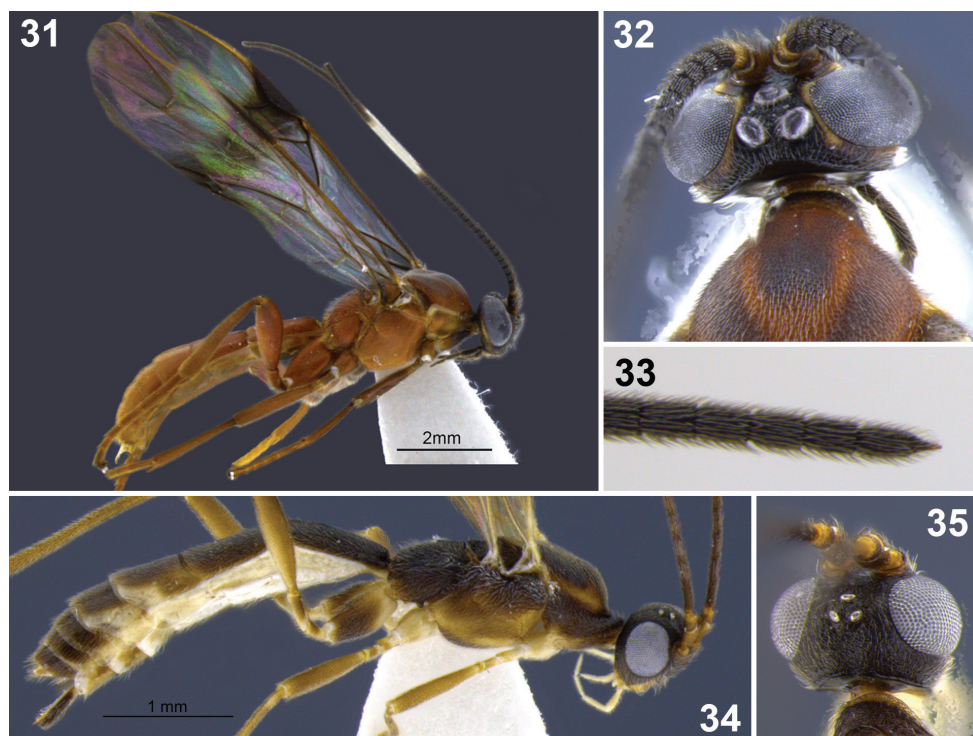
<http://zoobank.org/62EFAC20-EA55-4035-A4F9-5063A52DE15F>

http://species-id.net/wiki/Aleiodes_colberti

Figures 31–33, 36, 121

Description of the holotype. Male (holotype). Body length 8.8 mm; antenna length 8.5 mm; fore wing length 8.3 mm.

Color. Mostly reddish brown. Head mostly dark brown to black; yellowish stripe around eyes on frons, temple and gena; palp dark brown. Antenna black with white middle band between flagellomeres 20–29 (+/- 1). Mesosoma mostly reddish brown;

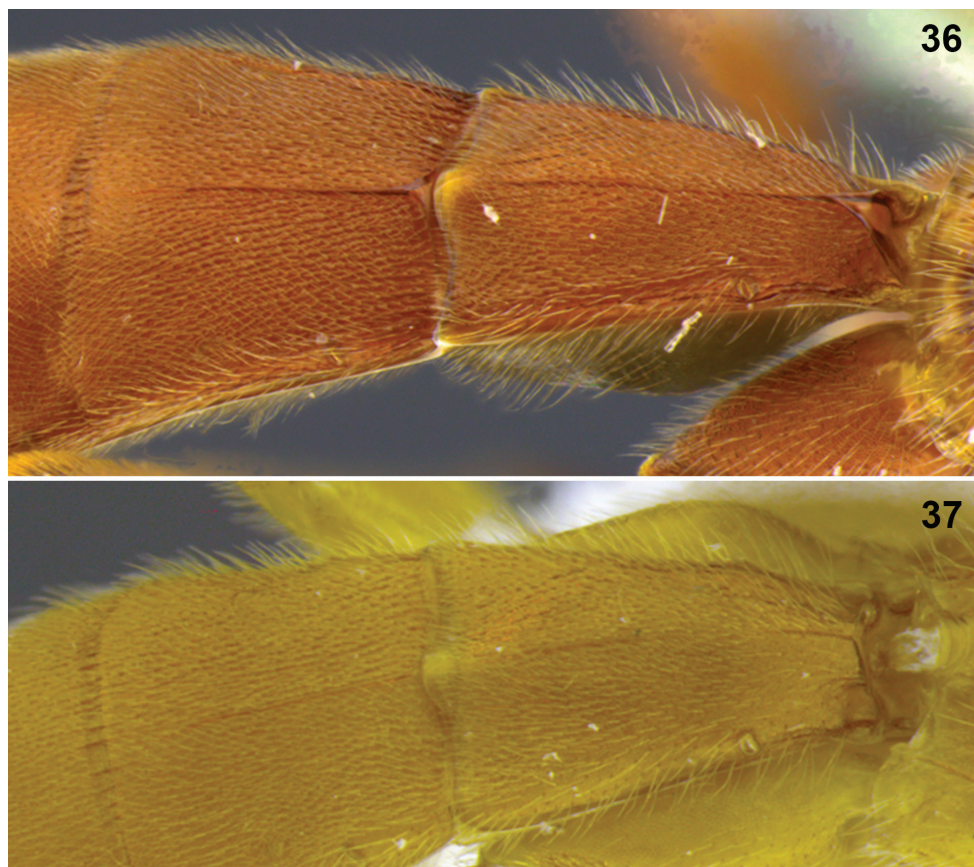


Figures 31–35. *Aleiodes colberti* sp. n. **31** habitus **32** head and mesoscutum, dorsal **33** apical flagellomeres **34, 35** *Aleiodes delicatus* sp. n. **34** habitus **35** head dorsal.

pronotum brown dorsally, laterally irregularly yellowish brown; propleuron brown, lighter posteriorly; mesoscutum infuscate except on notauli region; scutellum brown medially and postero-laterally. Wings weakly infuscate with dark band just bellow stigma; veins brown and stigma dark brown; tegula brown.

Head. 54 antennomeres, flagellomeres as long as wide, except apical 1/3 and basal 1/6, slightly longer than wide, apical flagellomere with distinctly pointed apex; malar space very short, 0.5× basal width of mandible and 1/6 of eye height; temple very narrow, in dorsal view eyes 5.5× longer than temples; occipital carina complete though irregular dorsally, ventrally barely reaching hypostomal carina; oral space small and circular, diameter about equal to basal width of mandible; clypeus weakly swollen; ocelli extremely large, ocell–ocular distance 1/8 diameter of lateral ocellus; face, gena and temples transverse rugose–costate, with mid-longitudinal ridge just bellow toruli, face granulose medially, clypeus granular–rugose, frons smooth and excavated, bordered by “W-shaped” carina, vertex coarsely granulate.

Mesosoma. Sculpturing mostly opaque granular; pronotum with median scrobiculate line; epicnemial carina incomplete dorsally; propodeum laterally with few wrinkles, mid-longitudinal carina on anterior 1/2, posterior half with irregular wrinkles; metapleuron rugose posteriorly; notauli virtually absent, only indicated anteriorly and meeting rugose



Figures 36–37. Metasomal terga 1–2. **36** *Aleiodes colberti* sp. n. **37** *Aleiodes luteosicarius* sp. n.

depressed area posteriorly; posterior margin of mesoscutum bordered by short carina just in front of scutellar sulcus; scutellar sulcus with five complete and well defined carina.

Wings. Fore wing: stigma $4\times$ longer than high; vein r about $0.7\times$ vein $2RS$, as long as vein $RS+Mb$, and $0.7\times$ vein $m-cu$; vein $3RSa$ $0.64\times$ vein $3RSb$, and $0.85\times$ vein $2M$; vein $1CUa$ $1.33\times$ vein $1cu-a$; vein $1CUB$ $2.25\times$ vein $1CUa$; vein $1M$ moderately curved at basal half. Hind wing: vein $m-cu$ absent; vein $M+CU$ slightly shorter $1M$; $1M$ about $2\times$ length of $r-m$; vein RS well pigmented, almost straight, gradually diverging from wing margin; vein M dark brown, well pigmented; vein $2-1A$ present and relatively long.

Metasoma. $T1$ and $T2$ granular-striate, longitudinal carina complete on $T1$ and $T2$; remainder visible terga granular; $T4-T6$ with small circular median dorsal pits, these terga densely setose except on median line, pubescence around the bare line directed to it; $T1$ length $2\times$ apical width.

Legs. Tarsal claws pectinate, with 6–7 thick bristles, and distinct gap between apical claw and basal pectination; hind basitarsus $1.7\times$ longer than inner apical spur of hind tibia.

Female unknown.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, March 2–5, 2006, S.R. Shaw col.

Type-specimen: Holotype male, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 2-5 March 2006, S.R. Shaw / Ex. Yellow pan trap”. (UWIM)

Discussion. *Aleiodes colberti* sp. n. belongs to the *pulchripes* species-group. This species can be distinguished from other New World species of this group by the infusate band on the forewing just below the pterostigma, and the white middle band on antenna. Despite these characters, it is similar to *Aleiodes earinos* Shaw, 1997, differing from which in its mostly black head and some dark stains on the mesoscutum and scutellum, as compared with the unicolored body in *earinos*. The pattern of wing veins resembles *Aleidoes arizonensis* Marsh & Shaw, 1997, mainly by the very long second submarginal cell. The infuscation below the stigma is also present in the Cuban and Costa Rican species *Aleiodes pedalis* Cresson, 1869, but *pedalis* also has distinct infumation apically, not present in *colberti* sp. n., as well as the apical hind tibia black, as opposed to reddish in *colberti* sp. n. In the key to New World *pulchripes* species (S. Shaw et al. 1997), *colberti* does not run easily to any of the described species. Considering the body color it will be forcibly run to *Aleiodes notozophus* Marsh & Shaw, 1997, but differs from that species by the large gap between apical claw and basal pectination, which is absent in *notozophus*. The male of *colberti* sp. n. has small setose pits on terga 4–6 (4–7 in *notozophus*). Disregarding the presence of the pits it will run to *Aleiodes vaughani* Muesebeck, 1960, but the ocelli are larger in the new species.

Etymology. This species is named after Stephen Tyrone Colbert, an American comedian, political satirist, writer, actor, and host of *The Colbert Report*.

Aleiodes delicatus sp. n.

<http://zoobank.org/2C718F94-CE3D-4120-BD76-BD3F2FE1713D>

http://species-id.net/wiki/Aleiodes_delicatus

Figures 34, 35

Description of holotype. Female (holotype). Body length 5.4 mm; antenna length 7.3 mm; fore wing length 4.5 mm.

Color. Mostly dark brown; legs pale brown but fore and mid coxae and trochanter whitish, hind coxa mostly dark brown, whitish on basal 1/4; all palp whitish; scape and pedicel, notauli and ventral border of mesoscutum, and mandibles yellowish; scutellum brown mid-anteriorly; propleuron pale brown, anteriorly darker; mesopleuron pale brown laterally, ventrally whitish with a roughly defined inverted “heart-shaped” honey brown infuscation; metasomal terga 3 and 4 slightly lighter than remainder metasoma and with whitish lateral borders; metasoma ventrally white except for last two sternites mostly light brown. Wing veins brown, but basally whitish, stigma pale brown, tegula white.

Head. Antenna with 45 segments, flagellomeres roughly $2.0\times$ as long as wide, apical flagellomere with small pointed apex; malar space wide, about $1.6\times$ basal width of mandible, $0.6\times$ eye height; in dorsal view eyes $2.4\times$ longer than temples; occipital carina complete dorsally, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width about as long as basal width of mandible; clypeus not swollen; ocelli small, ocell–ocular distance $2\times$ diameter of lateral ocellus; maxillary palp not swollen; head surface sculpture coarse granulate, occiput smooth and shining, frons not excavated, higher face with well defined mid-longitudinal ridge.

Mesosoma. Sculpturing mostly coarse granulate; pronotum foveate; propodeum rugulose with granular background, mid-longitudinal carina present on anterior $1/3$; notauli very shallow and narrow, with few crenulae anteriorly; posteriorly meeting on almost flat and weakly rugose area; posterior margin of mesoscutum bordered by carina; scutellar sulcus long, with three strong but incomplete carina.

Wings. Fore wing: stigma $4.5\times$ longer than high; vein r about $1.3\times$ vein 2RS, $1.4\times$ vein RS+Mb, and $1.0\times$ vein m-cu; second submarginal cell small and trapezoidal, vein 3RSa $0.28\times$ vein 3RSb, and $0.77\times$ vein 2M; vein 1CUa $1.6\times$ vein 1cu-a; vein 1Cub $2.7\times$ 1CUa; vein 1M weakly curved on basal half. Hind wing: vein m-cu absent; vein M+CU $0.85\times$ 1M; 1M about $2.7\times$ length of r-m; vein RS sinuate at middle, then only slightly diverging from wing margin; vein M dark brown, well pigmented; vein 2-1A absent.

Legs. Hind tibia with comb of modified setae; tarsal claw simple, not pectinate; inner apical spur on hind tibia extremely short, hind basitarsus $5\times$ longer than inner spur.

Metasoma. T1, T2 and basal $2/5$ of T3 finely rugose–striate with coarse granulate background, longitudinal carina present along T1 and T2; apical $3/5$ of T3 coarse granulate; remainder visible terga granular coriaceous; ovipositor sheaths as long as hind tarsomere II, and $0.47\times$ length hind basitarsus; T1 long and narrow, $1.6\times$ longer than its apical width.

Male. Unknown

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, August 15, 2006, A. Townsend col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Prov. / Yanayacu Biological Station / 15-Aug-2006 Macucoloma trail / Maxinet 2163m. A. Townsend”. (UWIM)

Discussion. *Aleiodes delicatus* sp. n. belongs to *seriatus* species-group. It is very similar to *Aleiodes pectoralis* (Ashmead, 1894) (= *sanctivicentensis* Shenefelt, 1975), from St. Vincent, and *A. akidnus* Marsh & Shaw, 1998, from Florida – USA, by the very small ocelli and the wing venation, remarkably the short second submarginal cell of the forewing, and also the longitudinal carina incomplete on propodeum. *A. delicatus* sp. n. is mostly dark brown with some lighter regions on mesoscutum and mesopleuron (legs also lighter and metasoma ventrally white), being the color pattern very similar to *pectoralis*, while *akidnus* is entirely honey yellow with black stemmaticum; however, the gena of *delicatus* sp. n. is dark brown, compared to light yellowish in *pectoralis*. Though short, the second submarginal cell is not almost square as in *akidnus* and *pectoralis*. The vein r is $0.75\times$

vein 3RSa in *delicatus* sp. n., but in *akidnus* and *pectoralis* r is slightly longer than 3RSa. In *delicatus* sp. n. the longitudinal carina is absent on metasomal tergite 3, but it is complete in *akidnus* and present on basal half in *pectoralis*, and the first metasomal tergite is 1.6× longer than apical width but roughly as long as wide in *akidnus* and about 1.3× in *pectoralis*.

Etymology. From the Latin, meaning delicate.

***Aleiodes dyeri* sp. n.**

<http://zoobank.org/6319072D-5E3A-4963-B0E2-135DBA18F0E4>

http://species-id.net/wiki/Aleiodes_dyeri

Figures 38–40, 122

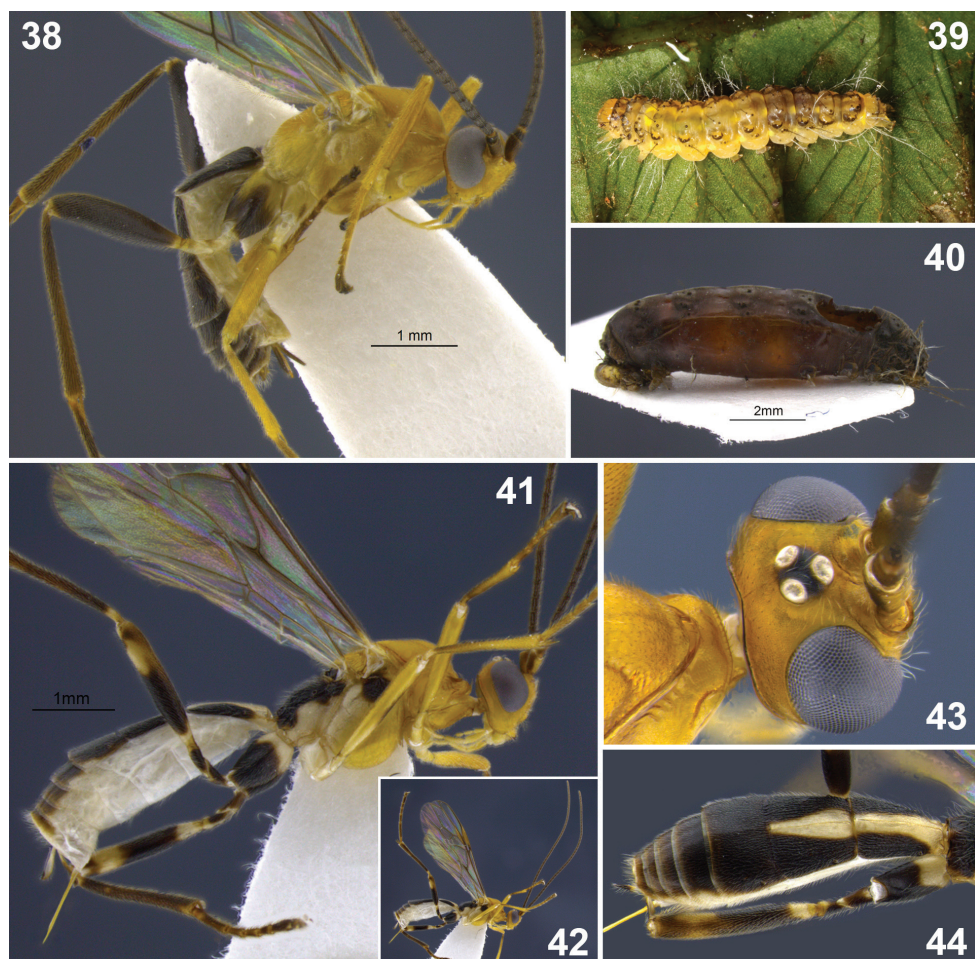
Description of holotype. Female (holotype). Body length 6.1 mm; antenna length 6+ mm; fore wing length 5.5 mm.

Color. Head honey yellow, ocellar triangle black; antenna dark brown, scape slightly lighter. Mesosoma honey yellow, some lighter parts on metapleuron and dorsal mesopleuron. Fore leg honey yellow, tarsi slightly darker but 5th tarsomere brown. Mid leg with same pattern of fore leg, but all tarsi brown; coxa, trochanter and trochantellus whitish, coxa with dark lateral stains. Hind leg: coxa mostly dark brown, basal third pale yellow; trochanter and trochantellus white with infuscate stains dorsally; femur black except for narrow basal whitish band; tibia and tarsi dark brown, tibial spurs honey brown. Metasoma black dorsally, ventrally white. Wings hyaline basally, becoming weakly infuscate apically; veins dark brown.

Head. Antenna with 53 antennomeres, flagellomeres roughly 2.0× as long as wide; malar space short, just slightly longer than basal width of mandible, and 0.3× eye height; eyes large, in lateral view temple very narrow, in dorsal view eyes 4.7× longer than temples; occipital carina incomplete, not meeting dorsally and curving toward lateral ocelli, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus slightly swollen; ocell–ocular distance about 0.7× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing shining granulate, occiput smooth and shining, frons also smooth with small weak concentric wrinkles; higher face with a small longitudinal ridge and transverse rugosity directed to it; frons excavated with excavation bordered by a weak “W-shaped” carina.

Mesosoma. Sculpturing shining granulate; pronotum foveate; propodeum coarsely shining granular with complete mid-longitudinal carina; notauli shallow and crenulate anteriorly, posteriorly meeting on depressed rugose area; posterior margin of mesoscutum bordered with complete carina; scutellar sulcus with median carina plus three pairs of almost complete lateral carina.

Wings. Fore wing: stigma 3.5× longer than high; vein r 0.75× vein 2RS, slightly longer than vein RS+Mb, and 0.7× vein m-cu; vein 3RSa 0.55× vein 3RSb, and 0.9× vein 2M; vein 1CUa 3× vein 1cu-a; vein 1CUb almost as long as vein 1CUa; vein



Figures 38–44. *Aleiodes dyeri* sp. n. **38** habitus **39** host larva, *Holophaea* sp. (Erebidae) **40** host mummy after parasitoid emergence **41–44** *Aleiodes elleni* sp. n. **41** , habitus **42** whole specimen habitus **43** head dorsal **44** metasoma, dorsal.

1M evenly slightly curved. Hind wing: vein m-cu distinct, pigmented and apparently tubular, distinctly antefurcal; M+CU $1.5\times$ 1M; vein 1M short, $0.85\times$ vein r-m; RS almost parallel to wing margin on basal 1/3 then slightly sinuate; vein M dark brown, well pigmented; vein 2-1A absent.

Legs. Hind tibia with comb of modified setae; tarsal claw pectinate, bristles relatively long and tightly arranged, with a short gap between pectination and claw base; hind basitarsus $3.4\times$ longer than inner apical spur of hind tibia.

Metasoma. T1, T2 and basal 2/5 of T3 rugose costate, longitudinal carina present along this sculpturing; remainder visible terga granular coriaceous; ovipositor sheaths about as long as tarsomere II; T1 length $1.36\times$ its apical width.

Male unknown.

Mummy. Length 8.5 mm, reddish brown (similar to a dipteran Brachycera puparium), most setae fell apart, exit hole irregular, located postero-dorsally.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-53568, S00°35.9', W77°53.4', 2163 m, cloud forest, February 15, 2011.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4', 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) Dec. 2010 / YY-53568; back (hand written): “15-Feb-2011”. (UWIM)

Biology. Reared from a species of *Holophaea* Druce (Erebidae) caterpillar (YY-53568), feeding on *Diplazium costale* var. *robustum* (Dryopteridaceae). Time span, from host mummification until adult emergence, about 5 weeks.

Discussion. *Aleiodes dyeri* sp. n. belongs to the *seriatus* species-group, where it resembles *A. greeneyi*, because of the dorsally incomplete occipital carina. It can be distinguished from *greeneyi* by the honey yellow mesosoma (dorsally black in *greeneyi*), the fore wing vein r 1.5× longer than RS+Mb (1.0× in *greeneyi*), and the hind wing vein r-m longer than 1M (shorter in *greeneyi*). *A. dyeri* sp. n. is similar to *longikeros* sp. n. in color patterns. These two species differ in the sculpturing of mesopleuron, being entirely granular in *dyeri* sp. n. but with a smooth elevated area in *longikeros* sp. n. The hind wing vein 1M is shorter than r-m in *dyeri* sp. n., as opposed to being 2.4× longer in *longikeros* sp. n., and the shape of fore wing vein 1M is weakly sinuate in *dyeri* sp. n., as compared with strongly curved in *longikeros* sp. n. Within the Nearctic species, *dyeri* sp. n. is more similar to *Aleiodes preclarus* Marsh & Shaw, 1998, from which it differs in the entire yellowish head but ocellar triangle black (several black spots in *preclarus*), wing veins mostly dark brown except fore wing veins M+CU and 1A, and hind wing veins 1M and M+CU proximally yellowish (pterostigma and fore wing vein C+SC+R with yellow spots in *preclarus*), and frons smooth (porcate in *preclarus*).

Comments. The antenna tips of the type specimen have a withered aspect, which makes impossible to measure the exact length of the antenna or describe the shape of the apical flagellomere.

Etymology. This species is named after Dr. Lee Dyer, of the University of Nevada (Reno), the lead investigator of the *Caterpillars and Parasitoids of the Eastern Andes of Ecuador* (CAPEA) project.

Aleiodes elleni sp. n.

<http://zoobank.org/B69D7595-D459-4032-B4BA-FC7CD67DFE48>

http://species-id.net/wiki/Aleiodes_elleni

Figures 41–44

Description of holotype. Female (holotype). Body length 6.1 mm; antenna length 7.1 mm; fore wing length 5.5 mm.

Color. Head yellow, ocellar triangle dark brown; antenna dark brown–black; scape, pedicel and first flagellomere proximally honey brown; scape darker dorsally; mesosoma yellow,

anterior corner of mesopleuron, metanotum propodeum and metapleuron dorsally dark brown–black, mesopleuron light yellow–whitish postero-dorsally; fore and mid leg coxa, trochanter and trochantellus whitish, darkening toward apex, 5th tarsomere honey brown, but tibia darker basally; mid trochanter, trochantellus and femur with infusate marking on inner side; hind coxa basal 1/3 white, apical 2/3 black; hind trochanter dark brown; hind trochantellus black with ventral white stripe; hind femur ventrally from base to apex with alternating bands: about 1/6 white, 1/2 dark brown, 1/4 pale yellow and 1/12 brown; dorsally only the basal white band well defined; tibia dark brown with basal pale yellow band; tibial spurs honey brown; tarsi brown. Metasomal terga black; median white stripe throughout length of terga 1 and 2; ovipositor sheaths dark brown apically, base whitish. Wings weakly infusate; stigma brown; most veins dark brown; vein C+SC+R darker (extreme base whitish) connecting to a black parastigma with light brown central spot.

Head. Antenna with 45 segments, flagellomeres roughly 2.0× as long as wide, apical flagellomere with short “bottle-nipple”-shaped tip; malar space moderate, about 1.3× basal width of mandible, 0.4× eye height; in dorsal view eyes 2.5× longer than temples; occipital carina complete dorsally, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus slightly swollen; ocell–ocular distance about as long as diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing shining granulate, occiput smooth and shining, frons also smooth with weak concentric wrinkles; frons excavated, excavation bordered by a weak “W-shaped” carina, but stronger laterally.

Mesosoma. Sculpturing shining granulate; pronotum foveate; propodeum rugulose with granular background and complete mid-longitudinal carina. Meso- and metapleuron surface rugose on dorsal dark brown areas; notauli shallow and crenulate anteriorly, posteriorly meeting on depressed rugose area; posterior margin of mesoscutum bordered with complete carina; scutellar sulcus with complete median carina plus two pairs of poorly defined lateral carina.

Wings. Fore wing: stigma 3.4× longer than high; vein r about 0.8× vein 2RS, 1.2× vein RS+Mb, and 0.7× vein m-cu; vein 3RSa 0.48× vein 3RSb, and 0.9× vein 2M; vein 1CUa 3.2× vein 1cu-a; vein 1CUB about as long as vein 1CUa; vein 1M slightly curved on basal half. Hind wing: m-cu is distinct, apparently tubular, distinctly antefurcal; vein M+CU about 2× longer than vein 1M; 1M about 0.8× length of r-m; vein RS almost straight, slightly diverging from wing margin; vein M dark brown, well pigmented; vein 2-1A present as a very short stub.

Legs. Hind tibia with comb of modified setae; tarsal claw simple, not pectinate; hind basitarsus 3× longer than inner apical spur on hind tibia.

Metasoma. T1, T2 and basal 2/5 of T3 granular–striate, longitudinal carina present along this sculpturing; remainder visible terga granular coriaceous; ovipositor sheaths about 1/2 length of hind basitarsus; T1 1.4× longer than its apical width.

Male. Antenna with 42–43 segments; metasoma narrower, T1 about 1.5–1.6× longer than its apical width; ocell–ocular distance shorter, 0.7× diameter of lateral ocellus; stigma narrower, 4.2× longer than high; one male has the white median stripe on metasoma interrupted on posterior half of tergite 2.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, November 5–12, 2007, J. Simbaña col.

Type-specimen: Holotype female. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 5-12 Nov. 2007, J. Simbaña / Macucoloma trail, Malaise trap / NSF-BSI-07-17458, S.R. Shaw”; second label: “SRS-00034”. (UWIM)

Paratypes, 2 males (UWIM), same locality as holotype, different collection methods and dates: 1♂, June 18, 2010, yellow pan trap, S.R. Shaw col.; 1♂, May 12, 2011, U.V. light, N. Zitani col.

Discussion. This species belongs to the *seriatus* species-group. *Aleiodes elleni* sp. n. is the only newly described species in this group with a strong, complete occipital carina on vertex. This character is present in two other Neotropical species: *A. scriptus* (from Brazil) and *A. nebulosus* (from Ecuador) from which *elleni* sp. n. differs by having the hind wing vein RS straight. *A. elleni* sp. n. also differs from *scriptus* by having the sculpturing of metasoma rugose–costate, as compared with widely costate in *scriptus*. It differs from *nebulosus* mostly in color patterns: pronotum yellow (white in *nebulosus*), apical 2/3 of hind coxa, propodeum, metanotum, metapleuron dorsally and mesopleuron on antero-dorsal corner black (white in *nebulosus*), and white medial marking on metasoma extending throughout tegite 1 and 2 (only anteriorly on tergite 1 in *nebulosus*). *A. elleni* sp. n. also differs from *nebulosus* in having the 2nd submarginal cell in fore wing long and rectangular (short and trapezoidal in *nebulosus*), 2RS and 3RSa forming a right angle (obtuse in *nebulosus*), and vein r less than half length of vein 3RSa (0.85× in *nebulosus*), wings hyaline (wings moderately infusate in *nebulosus*), and frons with lateral ridges (absent in *nebulosus*). The straight vein RS of the hind wing is shared with two other species of this species group: *A. frosti* sp. n. and the Nearctic species *Aleiodes femoratus* Cresson 1869; however, *elleni* sp. n. differs from *frosti* sp. n. by its smaller ocelli and relatively shorter petiole, and from both by the mostly black and medially whitish metasoma (yellowish in *frosti* sp. n. and *femoratus*). It also differs from *femoratus* by the smooth frons, as compared with porcate frontal sculpture in *femoratus*.

Etymology. This species is named after the American actress, comedian, and television host Ellen Lee DeGeneres.

Aleiodes falloni sp. n.

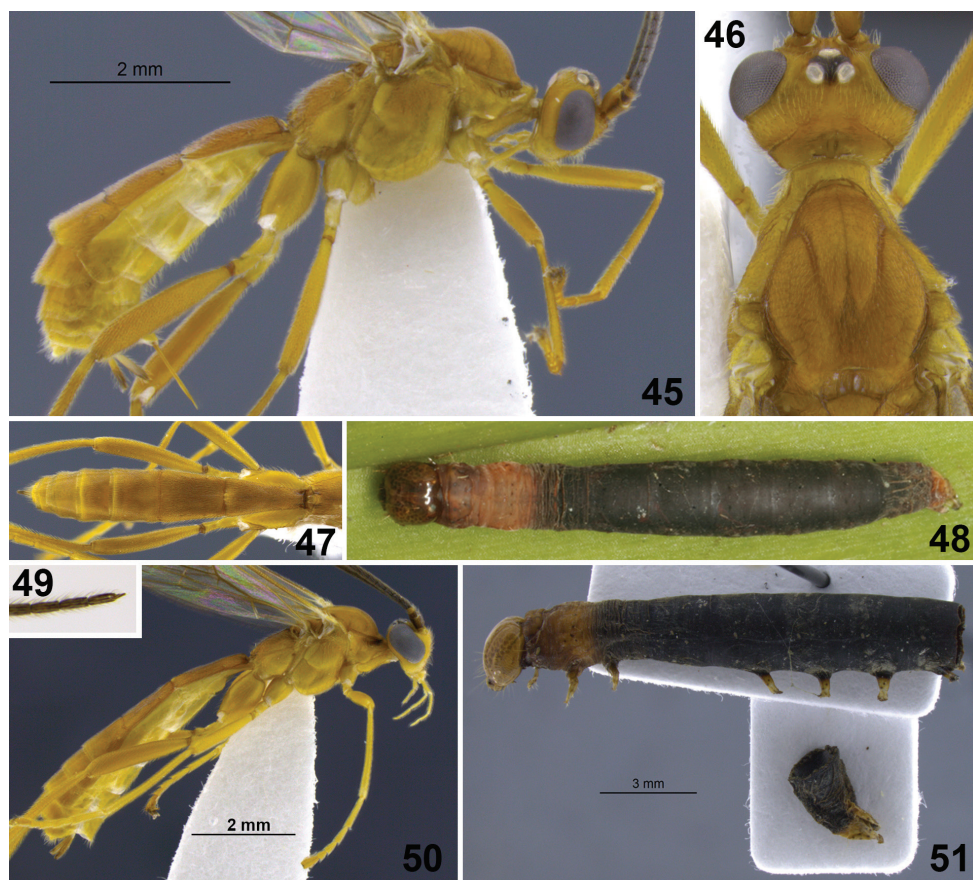
<http://zoobank.org/63DE928D-1CA0-4B80-B9B7-FDC81B5D4AC3>

http://species-id.net/wiki/Aleiodes_falloni

Figures 45–46, 123

Description of holotype. Female (holotype). Body length 6.9 mm; antenna length 7.6 mm; fore wing length 6.0 mm.

Color. Body gold–honey brown to orangish brown. Flagellum dark brown, scape and pedicel laterally brown; ocellar triangle black; tarsal claws brown; metasoma ven-



Figures 45–46. *Aleiodes falloni* sp. n. **45** habitus **46** head and mesonotum, dorsal **47–51** *Aleiodes frosti* sp. n. **47** metasoma, dorsal **48** host mummy, *Scotiuropsis* Hering sp. (Notodontidae), before parasitoid emergence **49** apical flagellomeres **50** habitus **51** host mummy after parasitoid emergence

trally, mandibles, fore and mid coxa light yellowish. Wings weakly infusate; veins dark brown but C+SC+R honey brown; stigma and vein R1 pale yellow–pale honey brown.

Head. Antenna with 50 antennomeres, flagellomeres roughly $2.0\times$ as long as wide, apical flagellomere with short pointed apex; malar space as long as basal width of mandible, and $0.35\times$ eye height; in dorsal view eye height $2.2\times$ temple; occipital carina incomplete dorsally (but not curved toward vertex), otherwise complete but not touching hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus bulging; ocell–ocular distance $0.86\times$ diameter of lateral ocellus; maxillary palp not swollen. Head surface sculpturing finely granulate, higher face with small longitudinal ridge and transverse rugosity directed to it, occiput smooth and shining.

Mesosoma. Sculpturing mostly granulate; pronotum foveate laterally; mesopleuron rugose on anterior corner; propodeum rugose–granulate, posteriorly with diverging and

some transverse wrinkles and weakly rugose laterally, with mid-longitudinal carina present on anterior 2/3; notauli present anteriorly, wide and shallow, posteriorly disappears in a depressed area with striations running antero-laterally from mid-posterior region; posterior margin of mesoscutum with short carina, just anterior to scutellar sulcus; scutellar sulcus with median carina plus two pairs of well defined lateral carina.

Wings. Fore wing: stigma 3× longer than high; vein r 0.55× vein 2RS, as long as vein RS+Mb, and 0.5× as long as vein m-cu; vein 3RSa about 0.5 times vein 3RSb, and as long as vein 2M; vein 1CUa 2× vein 1cu-a; vein 1CUb 1.8× vein 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu indicated as short pigmented vein antefurcal to vein r-m (in this species and others, the vein m-cu is very short but is also indicated by a slight bent on vein M, where these veins meet); vein M+CU about 1.4× 1M; vein 1M 1.4× vein r-m; vein RS smoothly curved at middle; vein M straight; vein 2-1A absent.

Legs. Hind tibia without comb of modified setae; tarsal claw pectinate at base, with a distinct gap between apical claw and basal pectination, pecten with 5–6 bristles; hind basitarsus 3× longer than inner apical spur on hind tibia; few rugositie dorso-laterally on outer side of hind coxa.

Metasoma. T1–T2 striated; remainder terga coriaceous; mid longitudinal carina complete from T1 throughout T2; ovipositor sheaths slightly shorter than hind tarsomere II, parallel sided and sharpened apically, setae longer pre-apically, longest setae about 1.5× longer than maximum width of sheaths; T1 slightly longer than apical width.

Variation. Antenna with 49–53 segments; some specimens with weak striation on basal 1/5 of metasomal tergite 3; scutellar sulcus with 3 to 5 well defined carina; one specimen with hind wing vein m-cu interstitial.

Male unknown.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, May 1–8, 2007, J. Simbaña col.

Type-specimen: Holotype female. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 1-8 May 2007, J. Simbaña / Macucoloma trail, Malaise trap / NSF-BSI-07-17458, S.R. Shaw”; second label: “SRS-00028”. (UWIM)

Paratypes. 1♀, same as holotype; 9♀s, same data as holotype but different dates: 5♀s, May 1–10, 2010; 1♀, June 3–13, 2009; 1♀, April 1–8, 2007; 2♀ February 1–8, 2007. (UWIM)

Discussion. This species belongs to *circumscriptus/gastritor* species group. It differs from the described species in this group by its mostly honey brown body color. *A. falloni* sp. n. resembles *Aleiodes speciosus* Townsend, from which it can be distinguished by the entire honey brown body color (mostly black–dark brown dorsally with first tergite white in *speciosus*) and the hind wing vein 2-1A absent (present in *speciosus*). It is very similar to *A. luteosicarius* sp. n., which belongs to the *pallidator* species-group, especially in color pattern (for distinguishing features see discussion at *luteosicarius* sp. n. section). Morphological distinction between *A. falloni* sp. n. and *A. luteosicarius* sp. n. is difficult due to their general resemblance. Separation of specimens in two different entities was supported

by comparison of ribosomal COI sequences, resulting in two groups with considerably different genetic information.

Etymology. This species is named after James Thomas Fallon, known as Jimmy Fallon, an American television host, comedian, actor, singer, musician and producer.

***Aleiodes frosti* sp. n.**

<http://zoobank.org/41C9635C-F5EF-4949-8822-01B80BEE93D0>

http://species-id.net/wiki/Aleiodes_frosti

Figures 47–51

Description of holotype. Female (holotype). Body length 8.1 mm; antenna length 10.7 mm; fore wing length 7.6 mm.

Body color. Yellowish to honey yellow, except for the ocellar triangle and antenna dark brown. Wings tinged yellowish; veins honey brown, parastigma blackish with central yellowish spot.

Head. Antenna 65 antennomeres, antenna 1.3× longer than body, flagellomeres roughly 2× longer than wide, apical flagellomere with long and narrow “bottle-nipple”-shaped apex; malar space short, about as long as basal width of mandible, and about 0.33× eye height; in dorsal view eyes 5× longer than temples; occipital carina incomplete dorsally, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus large, not swollen; ocelli large, ocell–ocular distance about 0.4× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpture finely shining granulate, occiput smooth and shining; higher face with a small longitudinal ridge and transverse rugosity directed to it; frons polished and excavated, without lateral ridges.

Mesosoma. Sculpturing shining granulate; pronotum with few wrinkles posteriorly and dorso-laterally; mesopleuron with small area at antero-dorsal corner rugose, and some wrinkles dorsally; propodeum longitudinally rugose posteriorly, mid-longitudinal carina almost complete; notauli with few crenulae and shallow anteriorly, meeting on depressed rugose area posteriorly; posterior margin of mesoscutum with complete carina; scutellar sulcus shallow and smooth except for the median carina.

Wings. Fore wing: stigma 5.3× longer than high; vein r 0.7× vein 2RS, 0.7× longer than vein RS+Mb, and 0.5× vein m-cu; vein 3RSa 0.5× vein 3RSb, and as long as vein 2M; vein 1CUa 1.7× vein 1cu-a; vein 1CUB 2.4× vein 1CUa; vein 1M weakly curved at its basal portion; RS+M straight. Hind wing: m-cu present, short and weakly pigmented, interstitial or just antefurcal to r-m; M+CU 1.25× vein 1M; 1M as long as r-m; RS mostly straight and gradually opening from wing margin, slightly bent downward at mid length; vein M straight, dark brown, well pigmented; vein 2-1A present as a short stub.

Legs. Hind tibia with comb of modified setae; tarsal claw simple, not pectinate; hind basitarsus 3× longer than inner apical spur of hind tibia.

Metasoma. T1, T2 and basal 2/3 of T3 rugose–striate, longitudinal carina complete on T1 and almost complete on T2, but not reaching posterior margin; ovipositor

sheaths parallel sided and truncate, about as long as hind tarsomere III; metasoma unusually long and narrow, T1 2.2× longer than its apical width.

Variation. Body length about 8.5 mm; antennomeres = 63; ocelli larger, ocell–ocular distance 0.25× diameter of lateral ocellus; hind wing vein 2-1A absent to short.

Male. Body length about 8.5 mm; antenna with 63 segments. Virtually identical to female, but ocell–ocular distance 0.3× diameter of lateral ocellus.

Mummy. Length 14.0 mm, black, thorax brown, head honey brown mottled brown, tubular in shape, thorax compact and wrinkled, exit mode unique within *Aleiodes*: the parasitoid cuts a radial opening at posterior side of the mummy, just behind the hind abdominal prolegs, releasing a “lid” with the anal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-50211, Beat C-16, S00°35.9', W77°53.4', 2163 m, cloud forest, September 5, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) July 2010 / YY-50211; back (hand written): “5-Sep-2010”. (UWIM)

Paratypes, 1 female and 1 male (UWIM), same data as holotype, except: 1♀, 23 May 2008, Yanayacu Road, YY-31409, colección por golpeo / ex. Poaceae *Chusquea scandens* – wasp emerged 02 July 2008; 1♂ YY-46923, beat 638, ex. host plant: Poaceae *Chusquea scandens* parasitoid emerged 10 May 2010.

Biology. Host plant *Chusquea scandens* (Poaceae); host Lepidoptera: *Scoturopsis* Hering sp. (Notodontidae); time span from pupation to emergence: about 5 weeks for females, unknown for the male. The parasitoid cuts a radial opening at posterior side of the mummy releasing a “lid”, comprising the anal apex of mummified caterpillar, before emergence. The mummy exit mode of this species is unique for the genus, since all previously known mummies produced by *Aleiodes* species had a posterior hole cut for emergence (Zaldívar–Riverón et al. 2008).

Discussion. *Aleiodes frosti* sp. n. belongs to the *seriatus* species-group. This species resembles *Aleiodes nigricosta* (Enderlein 1920) because of its entirely yellowish to honey yellow body, black stemmaticum and brown antenna, but differs in the honey brown fore wing vein C+SC+R, black in *nigricosta*. *A. frosti* sp. n. also differs in the extension of median longitudinal carina on metasoma, which is incomplete on tergite 2, but extends to half of tergite 3 in *nigricosta*, and the exceptionally elongate metasoma. *A. frosti* sp. n. is also similar to *elleni* sp. n. by the nearly straight hind wing vein RS, enclosing a marginal cell gradually widening toward wing apex, but it can be readily distinguish by the interrupted occipital carina on vertex, compared to the complete occipital carina of *elleni* sp. n. The diameter of lateral ocelli, 3–4× longer than ocell–ocular distance, is also a diagnostic character shared only with one Neotropical species, *Aleiodes nigribasis* (Enderlein 1920); however, most of the already mentioned diagnostic features for *frosti* sp. n. (e. g. shape of metasoma and hind wing vein RS, and color pattern) are also useful to distinguish it from *nigribasis*. Within the Yanayacu

species in the *seriatus* group it is similar to *greeneyi* because of the incomplete occipital carina at vertex. It differs from *greeneyi* by the entire yellowish body and its unusual long and narrow metasoma.

Etymology. The species is named after the American poet Robert Frost (1874 – 1963), author of the poem “The Road Not Taken.” This species name is also a reference to that poem, and to the unusual emergence mode of this species, recorded here for the first time. The following quotation extracted from this poem summarizes its idea: “Two roads diverged in a wood, and I – I took the one less traveled by,” Robert Frost, 1920. This *Aleiodes* species takes a “road not taken” by other species, to its adulthood, by emerging in a different and unique way.

***Aleiodes greeneyi* Townsend, 2009**

http://species-id.net/wiki/Aleiodes_greeneyi

Diagnosis. Body length 4.9 mm; antenna with 42 segments; head color honey brown with black ocellar triangle; malar space $1.25\times$ basal width of mandible; ocell–ocular distance about as long as width of lateral ocellus; occipital carina interrupted at vertex; mesopleuron granulate; apex of hind tibia with comb of flattened setae; propodeum strongly punctate with several longitudinally oriented rugae, longitudinal carina complete; metasomal terga 1–2 dark brown, terga 3–4 brown medially and dark brown laterally, remainder terga light brown; ovipositor sheaths about $0.6\times$ hind basitarsus length.

Additional characters. Last flagellomere with “bottle-nipple”-like tip; mesoscutum with carina at posterior margin complete although not well defined; scutellar sulcus with complete median carina plus one pair of slightly weaker lateral carina; fore wing vein 1M moderately curved at base; hind wing vein 2-1A indicated as a very short stub, vein m-cu present, interstitial to vein r-m; ovipositor sheaths about as long as hind tarsomere II, $0.6\times$ hind basitarsus.

Type material examined. (UWIM)

Biology. *Aleiodes greeneyi* has been reared from a Geometridae caterpillar feeding on *Evodanthus funifer* (Cyclanthaceae).

Distribution. Known only from the type locality, YBS, Napo province, ECUADOR.

Discussion. *Aleiodes greeneyi* is known only by the holotype. It belongs to the *seriatus* species-group. The incomplete occipital carina at vertex and the moderate ocelli size are similar to those presented in *A. longikeros* sp. n. and *A. dyeri* sp. n.; however, the wing vein pattern is somewhat intermediate between these two species. *A. greeneyi* differs from *longikeros* sp. n. and *dyeri* sp. n. in having the pronotum and mesoscutum mostly black (yellow in *longikeros* sp. n. and *dyeri* sp. n.), and the hind coxa light brown (bicolored black and white in *longikeros* sp. n. and *dyeri* sp. n.). *A. greeneyi* was reared from geometrid larva. Within the *seriatus*-group species from Ecuador, *A. nebulosus* and *longikeros* sp. n. were also reared from Geometridae hosts.

***Aleiodes kingmani* sp. n.**

<http://zoobank.org/4A47BA21-4AE9-4074-8F1A-B67F5EEB8EE2>

http://species-id.net/wiki/Aleiodes_kingmani

Figures 52, 53

Description of holotype. Male. Body length 4.7 mm; antenna length 5.5 mm; fore wing length 4.3 mm.

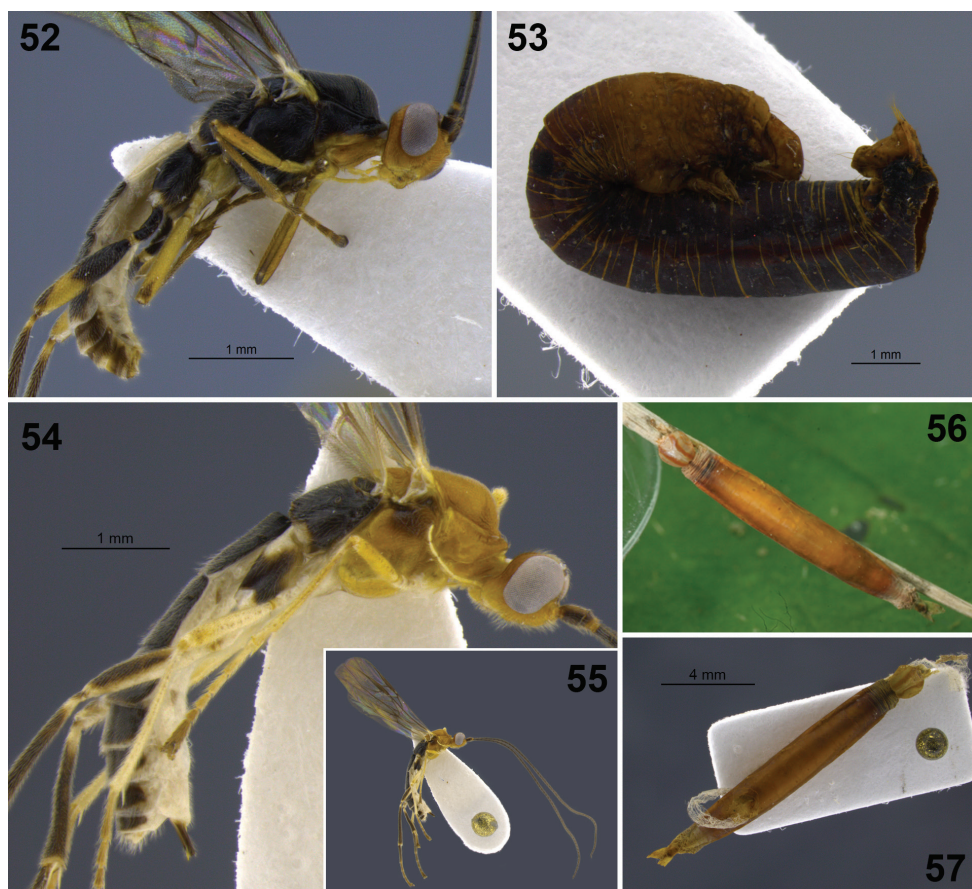
Color. Mostly black. Head honey yellow, ocellar triangle black; antenna dark brown except apical border of pedicel honey yellow. Mesosoma almost entirely black; propleuron and ventral quarter of pronotum, mesoscutum mid-posteriorly and scutellum medially yellowish to honey yellow; posterior border of propodeum white. Fore and mid legs with whitish coxa, darkening apically to honey yellow apical femur and tibia, and brown tarsi; hind leg coxa, trochanter and trochantellus black, but apical border of trochanter and trochantellus, and a small ventral spot on trochantellus white; femur black on basal 2/5 and dorso-apically, otherwise yellowish; tibia and tarsi brown, subbasal whitish small band on tibia. Metasoma black dorsally except for the white T1; apical borders of T4–T7 whitish; basal T5–T7 brownish. Wings moderately infusate; veins dark brown.

Head. 40 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with short pointed apex; malar space moderate, length 1.3× basal width of mandible, and approximately 1/3 eye height; in dorsal view eye 2.6× temples; occipital carina incomplete dorsally, curving toward lateral ocelli, well defined laterally but not meeting hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus slightly swollen; ocellus moderate, ocell–ocular distance short, about half diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing shining granulate, occiput smooth and shining; frons excavated with short lateral ridges.

Mesosoma. Sculpturing finely granulate; pronotum with some wrinkles laterally; mesopleuron coriaceous on central elevated area and some irregular latero-ventral parts; propodeum with mid-longitudinal carina incomplete, granulate postero-laterally with irregular wrinkles, and triangular rugose area mid-anteriorly with diverging wrinkles; notauli well defined anteriorly, narrow and crenulate, meeting a depressed rugose area posteriorly; posterior margin of mesoscutum bordered by complete carina; scutellar sulcus shallow, with median carina plus two pairs of poorly defined lateral carina.

Wings. Fore wing: stigma 4× longer than high; vein r 0.8× vein 2RS, 0.9× vein RS+Mb, and 0.7× as long as vein m-cu; vein 3RSa about 0.4 times vein 3RSb, and 0.8× vein 2M; vein 1CUa 2.7× vein 1cu-a; vein 1CUB slightly shorter than vein 1CUa; vein 1M moderately curved at basal portion. Hind wing: m-cu absent; vein M+CU 1.3× vein 1M; vein 1M 1.8× vein r-m; RS smoothly diverging from margin beyond middle; vein M dark brown, well pigmented; vein 2-1A absent.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, with a comb of thin bristles medially; hind tibial spurs relatively long, hind basitarsus 2.4× longer than inner apical spur.



Figures 52, 53. *Aleiodes kingmani* sp. n. **52** habitus **53** host mummy after parasitoid emergence
54–57 *Aleiodes longikeros* sp. n. **54** habitus **55** whole specimen habitus **56** host mummy before parasitoid emergence **57** host mummy after parasitoid emergence.

Metasoma. T1, T2 and basal 3/4 of T3 rugose costate, longitudinal carina present along this sculpturing; remainder T3 and T4 granular; remainder visible terga weakly shining coriaceous; T1 about as long as its apical width.

Female unknown.

Mummy. Length 9.5 mm, abdomen dark reddish brown, thorax, head and anal prolegs light brown, mummy bent (curled) ventrally, “J-shaped”, thorax inflated, exit hole barely round, located postero-dorsally at apex of mummy, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-57074, S00°35.9', W77°53.4', 2163 m, cloud forest, June 20, 2011.

Type-specimen: Holotype male and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4', 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) May 2011 / YY-57074; back (hand written): “em. 20-June-2011”. (UWIM)

Biology. Reared from mummified Geometridae caterpillar on *Chusquea scandens*. The host was collected as mummy.

Discussion. This species belongs to *circumscriptus/gastritor* species-group. It produces an unusual curled mummy, “J-shaped”, and emerge through a large and almost round exit hole at the posterior end. The white first tergite of *A. kingmani* sp. n., contrasting to the mostly black metasoma, is a useful diagnostic character shared only with *speciosus* and *townsendi* sp. n. *A. kingmani* sp. n. differs from *townsendi* sp. n. by the mostly black mesosoma (mostly yellowish in *townsendi* sp. n.), and the entirely black hind coxa (bicolored black and white in *townsendi* sp. n.). *A. kingmani* sp. n. differs from *speciosus* in having mesopleuron black (mostly yellow in *speciosus*) and entirely granulate (mostly smooth in *speciosus*); black region on head restricted to ocellar triangle (covering most of vertex and occiput dorsally in *speciosus*); hind coxa, trochanter and trochantellus black (yellow in *speciosus*); and occipital carina not meeting the hypostomal carina.

Etymology. This species is named after Eduardo Kingman (Loja, February 23, 1913 – Quito, November 27, 1997), one of the greatest Ecuadorian artists, who dedicated his art to portray the indigenous people of Ecuador.

***Aleiodes longikeros* sp. n.**

<http://zoobank.org/28CEB0BB-417E-4E91-A5FF-D232646AEEB9>

http://species-id.net/wiki/Aleiodes_longikeros

Figures 54–57

Description of holotype. Female (holotype). Body length 7.3 mm; antenna length 12.0 mm; fore wing length 7.0 mm

Color. Head honey yellow, ocellar triangle brown; antenna dark brown, scape and pedicel honey brown with darker outer side. Mesosoma honey yellow, dorso-anterior corner of mesopleuron infusate; metapleuron and propodeum black, posterior 1/5 of metapleuron white. Fore and mid leg whitish, tarsi darkening gradually, 5th tarsomeres brown. Hind leg: coxa mostly dark brown, basal third white; trochanter and trochantellus white; femur white with dorso-subapical dark brown stain; tibia and tarsi dark brown except for sub-basal white band on tibia which is larger ventrally; tibial spurs white. Metasoma black dorsally, with white spots mid-apically on T1 and medially from T2 to T4; ventrally white. Wings weakly hyaline; veins dark brown.

Head. Antenna with 60 antennomeres, about 1.6× as long as body, flagellomeres narrower than in other species, most flagellomeres 2.5× longer than wide, apical flagellomere with long “bottle-nipple”-shaped apex; malar space 1.1× longer than basal width of mandible, 0.4× longer than eye height; in dorsal view eyes 3× longer than temples; occipital carina barely incomplete, almost meeting dorsally and directed toward vertex, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width 0.8× basal width of mandible; clypeus not swollen; ocellular distance about as long as diameter of lateral ocellus; maxillary palp not swollen;

head surface sculpturing shining granulate, occiput smooth and shining; higher face with a small longitudinal ridge and transverse rugosity directed to it; frons only shallowly excavated.

Mesosoma. Sculpturing shining granulate; pronotum smooth laterally, dorsally granulate and foveate; mesopleuron with central elevated area smooth, antero-dorsal corner rugose; propodeum coarsely shining granular with complete mid-longitudinal carina; notauli very shallow anteriorly, virtually absent. Mesoscutum with some transverse wrinkles on anterior region of notauli, mid-posterior depressed area with few longitudinal wrinkles; posterior margin of mesoscutum with complete carina; scutellar sulcus with median carina plus two pairs of complete lateral carina.

Wings. Fore wing: stigma $5.7\times$ longer than high; vein r as long as vein 2RS, slightly longer than vein RS+Mb, and $0.75\times$ vein m-cu; vein 3RSa $0.55\times$ vein 3RSb, and as long as vein 2M; vein 1CUa $1.8\times$ vein 1cu-a; vein 1CUB $2\times$ vein 1CUa; vein 1M strongly curved in its basal portion. Hind wing: m-cu absent; vein M+CU $0.8\times$ vein 1M; 1M about $2.4\times$ longer than r-m; vein RS smoothly curved at middle; vein M straight dark brown, well pigmented; vein 2-1A present.

Legs. Hind tibia with comb of modified setae at apex; tarsal claw simple, not pectinate, with a comb of relatively long thin setae basally. Hind tibial spurs relatively short, hind basitarsus $3\times$ longer than inner spur.

Metasoma. T1, T2 and basal 2/5 of T3 granulose striate, longitudinal carina present along this sculpturing; remainder visible terga granular coriaceous; ovipositor sheaths about $0.85\times$ hind tarsomere II; apex of ovipositor sheaths truncate; T1 length $1.4\times$ its apical width, strongly widening posteriorly, its apical width about $2\times$ basal width.

Male unknown

Mummy. Length 12.5 mm, light reddish brown, head and prolegs light yellow, head with two longitudinal brownish stripes, thorax compact and wrinkled, posterior apex withered, glue hole located ventrally on the thorax, exit hole irregular, located postero-dorsally, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-57074, S00°35.9', W77°53.4', 2163 m, cloud forest, April 12, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: "ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4', 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) March 2010 / 46444"; back (hand written): "em. 12 April 2010". (UWIM)

Biology. Reared from mummified Geometridae caterpillar (YY-46444), on *Chusquea scandens* (Poaceae). The host was collected as mummy.

Discussion. *Aleiodes longikeros* sp. n. belongs to the *seriatus* species-group. This species has the longest antenna of any of these 24 newly described species, even though this is a medium sized species. Its antenna is about $1.6\times$ longer than body, as compared with at most $1.3\times$ in other species. The fore wing vein 1M strongly curved basally is also seen in *townsendi* sp. n. and *shakirae* sp. n., both belonging to *circumscriptus/gastritor* species-group, therefore both distinct from *longikeros* sp. n. by the absence of apical comb of flattened setae on hind tibia. *A. longikeros* sp. n. is similar to *dyeri* sp. n. (distinguishing

features are discussed on diagnosis section on *dyeri* sp. n.). It also resembles *greeneyi* because of the dorsally incomplete occipital carina, from which it can be distinguished by the honey yellow mesosoma (dorsally black in *greeneyi*), and white mid-apical spots on metasomal terga 1–4 (mostly black to dark brown in *greeneyi*); mesopleuron smooth on dorsal elevated area (granulate in *greeneyi*). *A. longikeros* sp. n. can be distinguished from all New World species by the following combination of characters: fore wing second submarginal cell long and narrow, vein 2RS 0.35× longer than vein 2M; the long antenna; and strongly curved vein 1M on fore wing. In the key to Nearctic species of the *seriatus* group (Marsh and Shaw 1998), *longikeros* sp. n. runs to *preclarus*. The new species differs from *preclarus* by the above mentioned character combination, and also by the entirely yellowish head and mesosoma, with several dark spots in *preclarus*, and the smooth frons, as compared with porcate frontal sculpturing in *preclarus*.

Etymology. From the Latin, meaning “long horned,” being a reference to the unusually long antenna of this species.

***Aleiodes luteosicarius* sp. n.**

<http://zoobank.org/E5AD2981-1E53-4CBD-9BFF-DFB446513ECD>

http://species-id.net/wiki/Aleiodes_luteosicarius

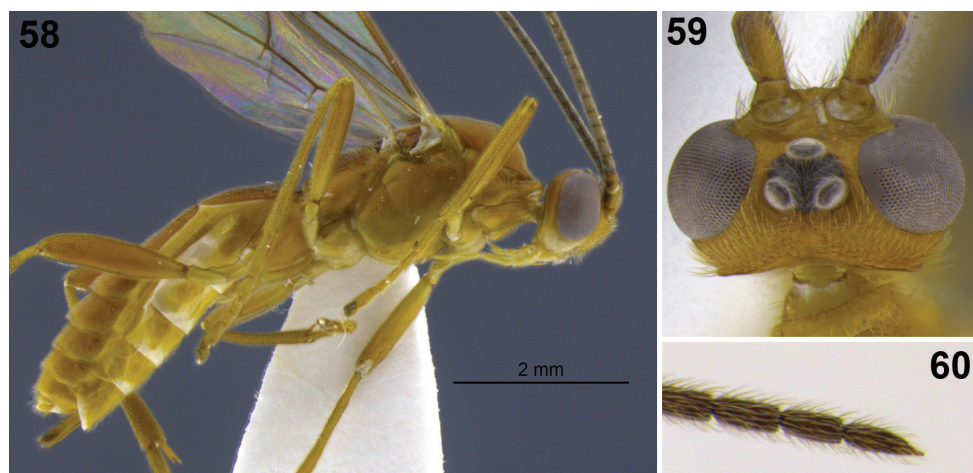
Figures 58–60, 61–68, 119, 124

Description of holotype. Female (holotype). Body length 6.9 mm; antenna length 7.6 mm; fore wing length 6.0 mm.

Color. Entire body honey brown to bronze, notum slightly darker; ocellar triangle black; antenna dark brown; wings hyaline; veins brown except C+SC+R, parastigma centrally, stigma, and R1 yellow; ovipositor mostly with same color of body, only weakly darkening apically.

Head. Antenna with 48 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with “bottle-nipple”-shaped apex; malar space as long as basal width of mandible, and 0.3× eye height; in dorsal view eye height 2.7× temple; occipital carina incomplete dorsally but not curved toward vertex, otherwise complete but not touching hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus not swollen; ocell–ocular distance 0.9× diameter of lateral ocellus; maxillary palp not swollen. Head surface sculpturing finely granulate, higher face with small longitudinal ridge and transverse rugosity directed to it, vertex coarsely granulate with some transverse wrinkles, occiput smooth and shining.

Mesosoma. Sculpturing mostly granulate; pronotum foveate laterally; mesopleuron rugose on anterior corner; propodeum mostly coarsely granulate with few longitudinal wrinkles posteriorly, laterally weakly rugose, with mid-longitudinal carina present on anterior 2/3; notauli present anteriorly, wide and shallow, posteriorly disappears in a depressed longitudinally rugose–striate area; posterior margin of mesoscutum with short carina just in front of scutellar sulcus; scutellar sulcus with strong median carina plus two pairs of strong but incomplete lateral carina.



Figures 58–60. *Aleiodes luteosicarius* sp. n. **58** habitus **59** head, dorsal **60** apical flagellomeres.

Wings. Fore wing: stigma about 4× longer than high; vein r 0.7× vein 2RS, about as long as vein RS+Mb, and 0.5× as long as vein m-cu; vein 3RSa about 0.5× vein 3RSb, and as long as vein 2M; vein 1CUa 2.5× vein 1cu-a; vein 1Cub 1.8× vein 1CUa; vein 1M virtually straight, only very slightly curved at basal half. Hind wing: m-cu indicated as short pigmented vein just postfurcal to vein r-m; vein M+CU about 1.3× 1M; vein r-m as long as vein 1M; vein RS faint, smoothly curved at middle; vein M straight; vein 2-1A present.

Legs. Hind tibia without comb of modified setae; tarsal claw pectinate with distinct gap between apical claw and bristles; hind basitarsus 3× longer than inner apical spur on hind tibia; few rugositie dorso-laterally on outer side of hind coxa.

Metasoma. T1–T2 and basal half of T3 striated; remainder terga coriaceous; mid longitudinal carina complete from T1 throughout T2; ovipositor sheaths slightly shorter than hind tarsomere II, relatively thick and somewhat rounded at tip, uniformly and densely covered with relative short regular sized pubescence, but dorso-basally bare, setae length no longer than width of sheaths; T1 slightly longer than apical width.

Paratype variation. Body length 6.7–7.4 mm; antenna 48–50 segments; 3 to 5 carina on scutellar sulcus; about half of paratypes have a lighter body color, otherwise very similar to holotype.

Male. Body length 5.7 mm; 45 antennomeres; body color a little darker; ocelli larger, ocell–ocular distance 0.6× diameter of lateral ocellus; hind wing vein 1M relatively shorter, and vein m-cu just postfurcal to r-m.

Mummy. All mummies densely setose, setae mostly yellowish to light brown contrasting with dark brown body, head varying from dark brown to yellowish brown; morphology of mummies variable according to different host species, exit hole irregular, postero-dorsal.

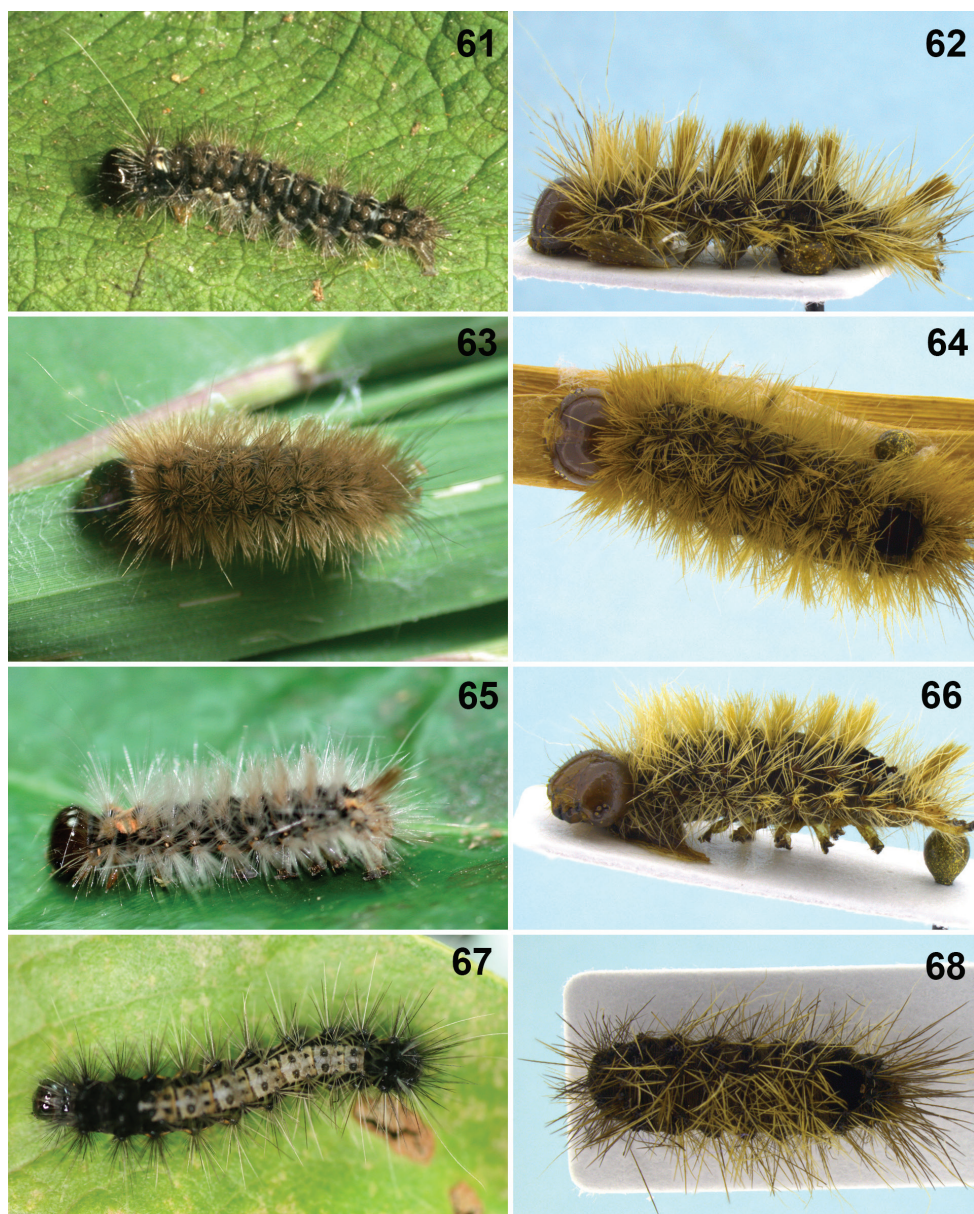
Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-47385, S00°35.9', W77°53.4', 2163 m, cloud forest, May 26, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) Apr. 2010 / YY-47385”; back (hand written): “26-May-2010”. (UWIM)

Paratypes, 9 females and 1 male (UWIM). 8♀ and 1♂, same data as holotype, except: 1♀, October 27, 2009, reared from *Amastus* nr. *hyalina* (Erebidae), YY-42218, feeding on *C. scandens*; 3 ♀ reared from *Pelochyta gandolfii* (Erebidae) feeding on *C. scandens*: April 27, 2009, YY-37911, March 16, 2007, YY-20534, and March 9, 2011, YY-54645; 1♀, December 2, 2009, YY-43108, reared from *Desmotricha imitata* (Erebidae) on *Barnadesia parviflora* (Asteraceae); 1♀ April 25, 2013, YY-73553, reared from *P. gandolfii*, on unknown host plant; 1♀ September 24, 2013, YY-78795, reared from Erebidae; 1♀ October 3, 2013, reared from *A. nr. hyalina*; 1♂, May 21, 2011, hand collected at daylight by aspirator (Ridge trail), H. Aguirre col. 1♀, ECUADOR, Napo Province, Cuyuja, Chalpi Grande, 2800m, March 3, 2006, hand collected at light sheet – Yanayacu exp. 2006, D.L. Wagner col.

Biology. Parasitoids on several Arctiinae (Erebidae) species: *Amastus coccinator* Schaus (including the holotype), *Pelochyta gandolfii* Schaus and *Amastus* nr. *hyalina* Dognin, collected on *Chusquea scandens* (Poaceae), and *Desmotricha imitata* Druce feeding on *Barnadesia parviflora* (Asteraceae). Nevertheless, *A. coccinator* and *P. gandolfii* have no previous rearing records feeding on *C. scandens*, which is the most common plant in the sampling sites. Since Arctiinae caterpillars are highly active, it is possible that they were just wandering over this plant; therefore, feeding on *C. scandens* for these two species needs confirmation. Time span from host mummification to adult emergence was about one month.

Discussion. *Aleiodes luteosicarius* sp. n. is the only species herein described which belongs to *pallidator* species-group. It is very similar to *falloni* sp. n., despite *falloni* sp. n. belongs to *circumscriptus/gastritor* species-group. *A. luteosicarius* sp. n. differs from *falloni* sp. n. in the following characters: ocell–ocular distance $0.3\times$ diameter of lateral ocellus (about $0.9\times$ in *falloni* sp. n.); at least half of tergite 3 striated, as in tergite 2 (tergite 3 mostly smooth–coriaceous); m-cu of hind wing slightly postfurcal to almost interstitial (antefurcal in females of *falloni* sp. n.); clypeus not swollen (swollen in *falloni* sp. n.); ovipositor sheaths relatively thick and somewhat rounded at tip, uniformly and densely covered with relative short regular sized pubescence, but dorso-basally bare, setae length no longer than width of sheaths (ovipositor sheaths sharpening at apex, setae concentrated apically, and not regular sized, longest setae about $1.5\times$ longer than width of sheaths in *falloni* sp. n.); fore wing as long as body length (distinctly shorter in *falloni* sp. n.); stigma narrower ($4\times$ longer than high in *luteosicarius* sp. n. vs. $3\times$ in *falloni* sp. n.); hind wing vein r-m as long as vein 1M (distinctly shorter in *falloni* sp. n.); depressed area on mesoscutum longitudinally rugose–striate (striations running antero-laterally from mid-posterior region); propodeum mostly coarsely granulate with few longitudinal wrinkles posteriorly, laterally weakly rugose (distinct pattern of rugosity on propodeum in *falloni* sp. n.); occipital carina strong and abruptly interrupted dorsally (weaker and gradually disappearing in *falloni* sp. n.); vertex coarsely



Figures 61–68. Arctiinae hosts of *Aleiodes luteosicarius* sp. n. **61** *Amastus coccinator* Schaus larva **62** host mummy (*A. coccinator*) after parasitoid emergence **63** *Pelochitas gandolfi* Schaus larva **64** host mummy (*P. gandolfi*) after parasitoid emergence **65** *Amastus* nr. *hyalina* Dognin larva **66** host mummy (*A. nr. hyalina*) after parasitoid emergence **67** *Desmotricha imitata* Druce larva **68** host mummy (*D. imitata*) after parasitoid emergence.

granulate with some transverse wrinkles (finely granulate in *falloni* sp. n.). In the key to species of *pallidator* species-group from North America (S. Shaw et al. 2013) it runs to *Aleiodes pallidator* (Thunberg, 1822). The new species closely resembles *pallidator*,

differing from it by the following: propodeum surface is entire rugose (laterally coriaceous in *pallidator*) and the longitudinal carina is present on anterior 3/4 (complete in *pallidator*); vein 1CUa about 2× length of 1cu-a (just slightly longer in *pallidator*); basal cell of hind wing very broad, vein r-m as long as vein 1M (narrower in *pallidator*); metasomal tergite 1 granular–rugose and tergite 2 striate–rugose (costate in *pallidator*); parasitoid on Arctiinae (Erebidae) (*pallidator* attacks Lymantriidae).

Comments. All previous known species of the *pallidator* species-group are parasitoids on Lymantriinae caterpillars (S. Shaw et al. 2013, as Lymantriidae). This is the first record of a species in this group attacking Arctiinae, and also the first species of the group with known host from Neotropical region. The status of subfamily for these groups is relatively recent, and both belong to Erebidae. Species within the *pallidator* group has been consistently reared from the setose mummified caterpillars of Lymantriinae in Japan, Europe and North America. The host associations reported here broaden the known host range for this species-group, but also denotes its ecological preferences for attacking densely setose caterpillars.

Etymology. From the Latin meaning “yellow killer,” referring to the main color of this parasitoid.

***Aleiodes marilynae* sp. n.**

<http://zoobank.org/6AB9A206-B015-42CC-9632-31E33DFF6DCD>

http://species-id.net/wiki/Aleiodes_marilynae

Figures 69–73

Description of holotype. Female (holotype). Body length 6.6 mm; antenna length 7.5 mm; fore wing length 6.6 mm.

Color. Head whitish, except for the ocellar triangle black, and the light yellow face and palp. Mesosoma black but pronotum and propleuron light yellow, and small posterior spot on metapleuron and posterior 1/5 of propodeum whitish; fore legs coxa, trochanter, trochantellus and mostly femur yellow; fore femur with dorso-apical brown stain, reminder fore legs brown; mid legs dark brown, but trochanter and trochantellus white; hind legs black. Metasomal tergite 1 with a black oval spot medially spanning from just behind basal carina to posterior edges of the tergite, remainder of tergite 1 white; tergite 2 black medially, dark region almost quadrate in shape with lateral borders convex at 2/3 posterior, white laterally; tergite 3 mostly white, with basal almost semi-circular black mark; reminder terga white apically, basally black, more or less concealed by the preceding tergite; metasoma ventrally white with a pair of latero-basal spots on each sternite, the spots larger on the second sternite; ovipositor sheaths black, ovipositor yellow.

Head. 50 antennomeres; most flagellomeres roughly 2× longer than wide, apical flagellomere with small pointed apex; malar space wide, about 2.5× times basal width of mandible and almost as long as eye height; temple wide, in dorsal view about as long as eye; occipital carina absent; oral space small and circular, diameter about equal



Figures 69–73. *Aleiodes marilynæ* sp. n. **69** habitus **70** head and mesoscutum, dorsal **71** metasoma, dorsal **72** host larva (Geometridae) **73** host mummy after parasitoid emergence.

to basal width of mandible; clypeus weakly swollen; ocelli very small, ocell–ocular distance $2.2\times$ diameter of lateral ocellus; head polished (smooth and shining).

Mesosoma. Sculpturing polished (smooth and shining); pronotum shining granular dorsally, otherwise shining coriaceous; mesopleuron sparsely punctate; notauli very weakly indicated only anteriorly; posterior margin of mesoscutum without carina, smoothly depressing into scutellar sulcus; scutellar sulcus with five incomplete carina posteriorly; mesopleuron central region bare; epicnemial carina absent; longitudinal carina on propodeum absent.

Wings. Fore wing: stigma $3.8\times$ longer than high; vein r $0.75\times$ length of $2RS$, $0.75\times$ length of $m-cu$, and as long as vein $RS+Mb$; vein $3RSa$ $0.44\times$ vein $3RSb$, and as long as vein $2M$; vein $1CUa$ about $2\times$ vein $1cu-a$; $1Cub$ $2\times$ length of $1CUa$; vein $1M$ almost straight. Hind wing: vein RS curved at middle, marginal cell narrowest point at middle; vein $1M$ about $2\times$ longer than vein $r-m$; vein $M+CU$ $0.9\times$ vein $1M$; vein $m-cu$ absent; vein $2-1A$ present.

Legs. Tarsal claws simple, not pectinate; hind basitarsus $3\times$ longer than inner apical spur of hind tibia; hind coxa smooth.

Metasoma. $T1-T3$ smooth and shining with punctuations on setal “pores”; remainder terga coriaceous; longitudinal carina barely indicated on $T1$ and absent on

T2; petiole broad, 0.8× longer than its apical width; ovipositor sheaths about as long as hind tarsomere II.

Male unknown.

Mummy. Length 14.9 mm, mostly pale brown with some dark spots, dorsally at middle of abdomen with faint “X-shape” mark and four withered expansions on each tip of the mark, thorax very strongly compact and abdomen angled upward, very similar to *A. capillosus* mummies, glue hole ventrally on thorax, exit hole postero-dorsal, between abdominal and anal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-44198, S00°35.9', W77°53.4', 2163 m, cloud forest, February 5, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) Dec. 2009 / 44198”; back (hand written): “em. 5 Feb 2010”. (UWIM).

Biology. Host plant: *Acalypha macrostachya* (Euphorbiaceae). Host Lepidoptera: Geometridae – common name: “cachos X blanca en la espalda”. The parasitoid adult emerged one month after host mummification.

Discussion. This is the fourth species of the *gressitti* species-group described. Of the former three, two are from New World, *Aleiodes lissos* Marsh & Shaw, 2003 and *Aleiodes capillosus* Townsend, and one from Campbell Island in the South Pacific: *A. gressitti* Muesebeck, 1964. This new species most resembles the Neotropical *capillosus* because of the absence of occipital and epicnemial carina, and also in the mostly black mesosoma and infusate wings. It differs from *capillosus* in the whitish head, except for the ocellar triangle being black and the face light yellow, pronotum and propleuron, mid trochanter and trochantellus white, and fore leg coxa, trochanter, trochantellus and mostly femur yellow (while all these parts are black in *capillosus*). The malar space is very large, being about as long as the eye height (at most 1/2 in other *gressitti*-group species). The mummies produced by *marilynae* sp. n. and *capillosus* are very similar in the extremely contracted thorax and relatively swollen abdomen, and also in the characteristic acute angle at which the mummy is attached to the substrate.

Etymology. This species is named in honor of Marilyn Rieden Shaw, wife of the co-author, Scott R. Shaw, in gratitude for her support for his entomological studies over many years.

Aleiodes mirandae sp. n.

<http://zoobank.org/80CF0827-3BF3-473E-94D4-4E35D0200C8A>

http://species-id.net/wiki/Aleiodes_mirandae

Figures 74–76

Description of holotype. Female (holotype). Body length 5.4 mm; antenna length 6.3 mm; fore wing length 4.8 mm.

Color. Mostly black. Head orangish yellow, including mandibles and palp, but mandibles tip brown; ocellar triangle black; antenna dark brown except apical border of pedicel honey yellow. Mesosoma almost entirely black; propleuron, pronotum, and scutellum medially honey yellow; posterior border of propodeum white. Fore leg yellowish; outer apical 1/3 of femur infusate; basal half of tibia and tarsi brown. Mid leg with same pattern of fore leg but ground color whitish. Hind leg black; light yellow markings on apical border of trochanter and trochantellus, small ventral spot on trochantellus, apical half of femur latero-ventrally, and small sub-basal band on tibia. Metasoma black dorsally, bordered white; T1, triangular mid-basal area on T2 and apical borders of T3–T7 white; ventrally white but infusate medially; ovipositor sheaths basal 1/3 whitish, apical 2/3 black. Wings weakly infusate; veins dark brown.

Head. Antenna with 46 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with very short pointed tip; malar space wide, about 1.8× basal width of mandible, and 0.6× eye height; in dorsal view eye 1.4× temple; occipital carina incomplete, close but not meeting dorsally and curving toward lateral ocelli, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus slightly swollen; ocelli small, ocell–ocular distance about 1.8× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing granulate, occiput smooth and shining; frons excavated with short lateral ridges.

Mesosoma. Sculpturing finely granulate; pronotum foveate; mesopleuron central disc mostly smooth and bare, posteriorly and ventrally coriaceous, antero-dorsal corner rugose; propodeum coarsely granular with complete mid-longitudinal carina; notauli well defined and crenulate anteriorly, barely defined but traceable posteriorly, meeting a depressed rugose area; posterior margin of mesoscutum bordered with complete carina; scutellar sulcus with median carina plus two pairs of incomplete lateral carina.

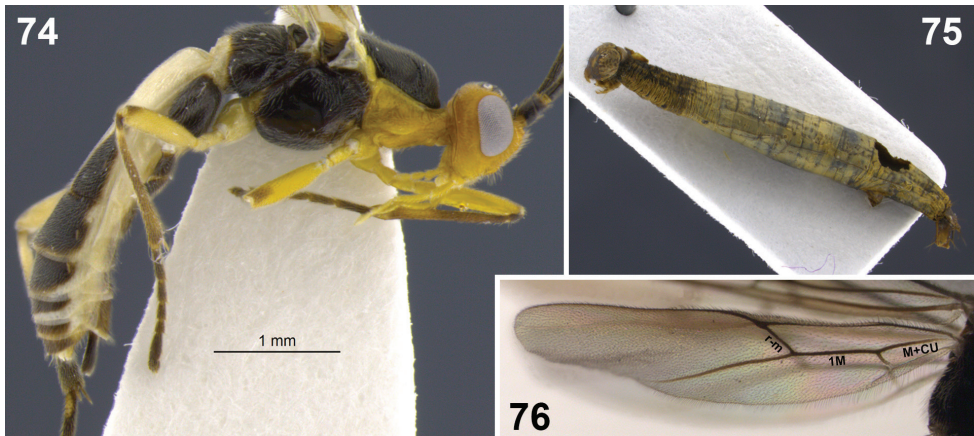
Wings. Fore wing: stigma about 4× longer than high; vein r 0.8× vein 2RS, as long as vein RS+Mb, and 0.75× as long as vein m-cu; vein 3RSa about 0.5× vein 3RSb, and 0.9× vein 2M; vein 1CUa 2.5× vein 1cu-a; vein 1CUB 2× vein 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu absent; M+CU 0.8× 1M; vein 1M almost 3× vein r-m; vein RS smoothly curved at middle; vein M dark brown, well pigmented; vein 2-1A present, short.

Legs. Hind tibia without apical comb of modified setae; tarsal claw simple, not pectinate, with a comb of relatively long thin setae basally; hind tibial spurs relatively short, about 1/4 basitarsus length.

Metasoma. T1, T2 and basal 3/4 of T3 rugose costate, longitudinal carina present along this sculpturing; remainder T3 and T4 granular; remainder visible terga weakly shining coriaceous; ovipositor sheaths 0.7× length of hind basitarsus; T1 length slightly longer than its apical width.

Variation. 45–46 antennomeres; scutellar sulcus with 5 or 7 carina; white markings larger: almost all apical 1/2 on hind femur, basal 1/2 of hind tibia, basal spot on inner hind basitarsus, and throughout T2 length medially (widening toward base) reaching T3 basally as tiny central mark.

Male unknown.



Figures 74–76. *Aleiodes mirandae* sp. n. **74** habitus **75** host mummy after parasitoid emergence **76** hind wing.

Mummy. Length 10.0 mm, entire mottled pale yellow and gray, thorax compact and wrinkled, anal prolegs extended posteriorly, exit hole irregular, located postero-dorsally, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, S00°35.9', W77°53.4', 2163 m, cloud forest, 16 May 2011, M. Bryant col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / M. Bryant, 10–20 May 2011 / NSF-DEB-10-20751 / (hand written) blacklight / 16 May 2011”. (UWIM).

Paratypes, 3 females (UWIM): same locality as holotype, different collecting dates and methods: 1♀, March 15, 2010, reared, YY-45457; 1♀, February 7, 2010, reared YY-53818; 1♀, February 9, 2010, reared, YY-53961.

Biology. Reared from a Geometridae species, common name “palito café chusquea”, feeding on *Chusquea scandens* (Poaceae). Morphology of mummies and rearing data corroborate a single host species. Three weeks elapsed between mummification until adult emergence.

Discussion. *Aleiodes mirandae* sp. n. belongs to *circumscriptus/gastritor* species-group. It is similar to *A. napo* sp. n. in the very small ocelli (ocell–ocular distance about 2× diameter of lateral ocelli), and also the mostly smooth mesopleuron. It differs from *napo* sp. n. in the rugose depressed mid-posterior area on mesoscutum, flat and granular in *napo* sp. n., the head, except black ocellar triangle, pronotum, propleuron and scutellum orangish yellow, all black in *napo* sp. n. except for small reddish marking on temples. The color pattern is very similar to *kingmani* sp. n., but additionally to already mentioned diagnostic characters *mirandae* sp. n. have hind wing vein M+CU shorter than 1M, while in *kingmani* sp. n. 1M is more than 2× longer than r-m. The host species “palito café chusquea” (Geometridae) is the same species attacked by *Aleiodes nubicola* sp. n. and *Aleiodes shakirae* sp. n.

Etymology. This species is named after Miranda Bryant, collector of the holotype.

***Aleiodes napo* sp. n.**

<http://zoobank.org/6779D464-D253-4A4D-B873-B955737CD320>

http://species-id.net/wiki/Aleiodes_napo

Figures 77–81

Description of holotype. Female. Body length 5.0 mm; antenna length 6.3 mm; fore wing length 5.3 mm.

Color. Mostly black. Head with a small brown stain on temples, just behind eyes; palp, fore and mid coxa, trochanter and trochantellus pale yellowish. Metasomal tergite 1 white, with a black oval spot medially spanning from just behind basal carina to posterior edges of the tergite; remainder terga mostly black, white laterally; metasoma ventrally white with a pair of latero-basal spots on each sternite, the spots larger on the second sternite; ovipositor sheaths black, ovipositor yellow.

Head. 47 antennomeres; most flagellomeres about $1.5\times$ longer than wide, apical flagellomere with small pointed tip; malar space moderate, about $1.5\times$ times basal width of mandible, and $0.5\times$ eye height; temple wide, in dorsal view slightly shorter than eye; occipital carina absent dorsally, running toward vertex, ventrally almost meeting hypostomal carina; oral space small and circular, diameter about equal to basal width of mandible; clypeus with distinct line separating dorsal and ventral surfaces, dorsally coriaceous, ventrally convex and smooth; ocelli very small, ocell–ocular distance about $2.5\times$ diameter of lateral ocellus; head, including frons, granulate.

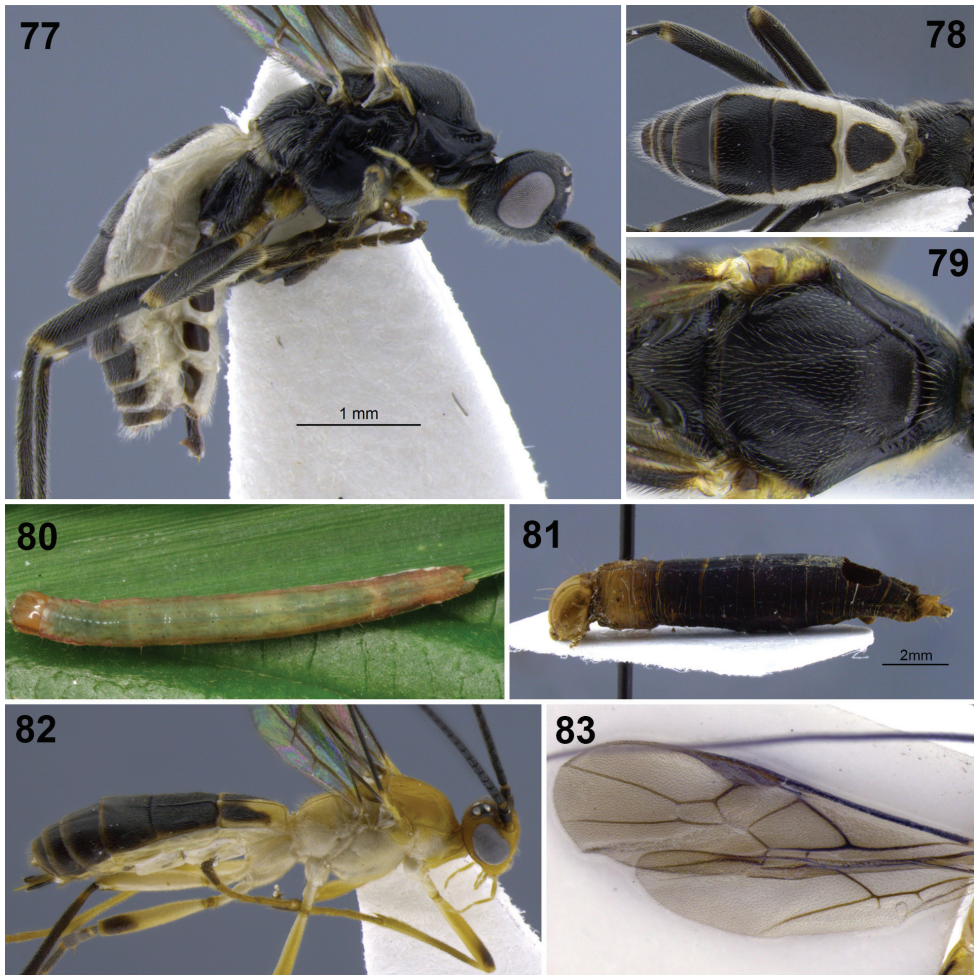
Mesosoma. Mesoscutum and scutellum granular coriaceous; pronotum dorsally and laterally on anterior half granular with scrobiculate sulcus, remainder lateral pronotum smooth except for the posterior corner coarsely rugose; propleuron granular–coriaceous; mesopleuron mostly smooth centrally to finely coriaceous, antero-dorsal corner weakly rugose–punctate; metapleuron and propodeum coarsely granular; longitudinal carina on propodeum almost complete; notauli present only anteriorly, narrow and crenulate, mid-posterior area of mesoscutum not depressed and granulate; entire posterior margin of mesoscutum bordered by carina; scutellar sulcus with median carina plus two pairs of lateral carina.

Wings. Fore wing: stigma $4.4\times$ longer than high; vein r $0.65\times$ length of 2RS, $0.7\times$ length of m-cu, and $0.85\times$ vein RS+Mb; vein 3RSa $0.3\times$ vein 3RSb, and $0.93\times$ vein 2M; vein 1CUa $1.4\times$ vein 1cu-a; 1CUB $2.8\times$ length of 1CUa; vein 1M almost straight. Hind wing: vein RS curved at middle, marginal cell narrowest point at middle; vein 1M $2.2\times$ longer than vein r-m; vein M+CU $0.68\times$ vein 1M; vein m-cu absent; vein 2-1A present.

Legs. Apex of hind tibia without comb of modified setae; tarsal claws simple, not pectinate; hind basitarsus about $3\times$ inner apical spur of hind tibia; hind coxa granulate basally.

Metasoma. T1 striate–rugose; T2 striated on basal $2/3$; apical $1/3$ of T2 and basal half of T3 coriaceous; longitudinal carina present along with the striated sculpturing on T1 and T2; T1 $0.87\times$ longer than its apical width; ovipositor sheaths about as long as hind tarsomere II.

Male unknown.



Figures 77–81. *Aleiodes napo* sp. n. **77** habitus **78** metasoma, dorsal **79** mesonotum, dorsal **80** host larva (Noctuidae) **81** host mummy after parasitoid emergence **82, 83** *Aleiodes nebulosus* Townsend **82** habitus **83** wings.

Mummy. Length 12.0 mm, head honey yellow, thorax pale brown, abdomen dark reddish brown, almost tubular in shape, exit hole located postero dorsally.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-48553, S00°35.9', W77°53.4', 2163 m, cloud forest, July 14, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: "ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA – NSF-BSI-07-17458 / REARED / 2010 (hand written) May 48553"; back (hand written): "14-Jul-2010". (UWIM)

Biology. Reared from a Noctuidae caterpillar (common name "raya roja a los lados chusquea") collected on *Chusquea scandens* (Poaceae). The parasitoid took six weeks from host mummification until emergence.

Discussion. This species belongs to *circumscriptus/gastritor* species-group. The color pattern of *Aleiodes napo* sp. n. is similar to those of *capillosus*; however, it does not belong to the *gressitti* species group because of the sculpturing on metasomal tergite 3, which is granular coriaceous on basal half. *A. napo* sp. n. differs from *capillosus* also in the presence of both occipital and epicnemial carina, though the former is incomplete dorsally, the presence of a complete longitudinal carina on metasomal terga 1 and 2, body sculpturing mostly granular–coriaceous, and longitudinal carina on propodeum almost complete (in *capillosus* all the above mentioned carina are absent, and the body sculpturing mostly smooth). Within the *circumscriptus/gastritor* group, *A. napo* sp. n. is similar to *mirandae* sp. n. in having a smooth central disc of mesopleuron and hind wing vein M+CU shorter than 1M, but differs from it in the almost entirely black head and thorax, mostly orangish yellow in *mirandae* sp. n., and the posterior central region of mesoscutum flat and granular, depressed and rugose in *mirandae* sp. n.

Etymology. This species is named after the indigenous inhabitants of the eastern Ecuador, for whom the Province (locality of the type) is also named: the Napo Runas.

Aleiodes nebulosus Townsend, 2009

http://species-id.net/wiki/Aleiodes_nebulosus

Figures 82, 83

Diagnosis. Body length 6.1 mm; antenna with 47 segments; head color honey brown with black ocellar triangle; malar space 1.2× basal width of mandible; ocell–ocular distance 1.25× width of lateral ocellus; occipital carina complete; mesopleuron granulate; apex of hind tibia with comb of flattened setae; propodeum granulate anteriorly and rugose posteriorly, longitudinal carina complete; metasomal terga mostly dark brown–black, tergite 1 with antero-median off-white marking; ovipositor sheaths length about half of hind basitarsus length.

Additional characters. Last flagellomere with short pointed tip; mesoscutum with complete and well defined carina bordering posterior margin; scutellar sulcus with seven carina; fore wing vein 1M slightly curved at base; hind wing vein 2-1A absent, vein m-cu present, distinctly antefurcal to vein r-m; ovipositor sheaths about as long as hind tarsomere II, 0.5× hind basitarsus. Different than originally described, the sternaulus is absent.

Type material examined. (UWIM)

Biology. *Aleiodes nebulosus* has been reared from unidentified Noctuidae caterpillar feeding on *Acalypha platyphylla* (Euphorbiaceae). One hyperparasitoid, *Mesochorus* sp. (Ichneumonidae), was reared from a similar mummy.

Distribution. Known only from the type locality, Isla de Las Palmas, Napo province, ECUADOR, at 1,885 meters elevation.

Discussion. *Aleiodes nebulosus* is known only by the holotype. In the *seriatus* species-group, *A. nebulosus* is similar to *A. elleni* sp. n. in having the occipital carina complete at vertex. These two species are also similar in color pattern because of the most yellowish

head and mesoscutum, and the mostly dark brown to black metasoma with a whitish mark antero-medially. Distinguishing features between *nebulosus* and *elleni* sp. n. are presented in the discussion section for *elleni* sp. n.

***Aleiodes nubicola* sp. n.**

<http://zoobank.org/198B4299-54FC-41C7-A661-CEE1AABFCB89>

http://species-id.net/wiki/Aleiodes_nubicola

Figures 84–91

Description of holotype. Female (holotype). Body length 3.8 mm; antenna length 4.9 mm; fore wing length 3.6 mm.

Color. Mostly dark brown to black. Head: gena and clypeus honey yellow; mandibles and palp whitish, but teeth brown. Mesosoma: lighter stripe, brown to reddish brown, laterally on mesopleuron; mesoscutum and scutellum mid apically yellowish; metapleuron just posteriorly whitish. Legs mostly yellowish but all tibia and tarsi brownish, hind coxa, trochanter and trochantellus dark brown to black, 1/3 basal of hind femur brown. Metasoma: ventrally, central apical T4 and remainder apical visible terga whitish to light yellow; ovipositor sheaths dark brown. Wings hyaline, veins dark brown, but some apical veins, stigma and base of fore wing M+CU and 1A light brown to almost colorless.

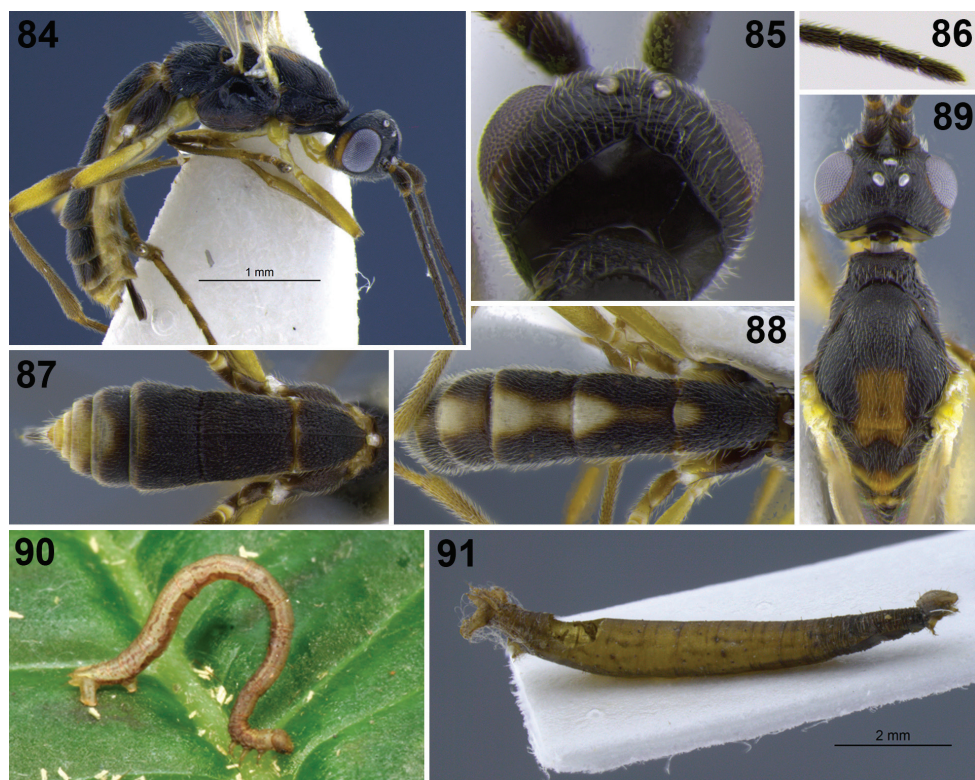
Head. Antenna with 38 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere lanceolate, without pointed apex; malar space moderate, length 1.65× basal width of mandible, and about 0.65× eye height; in dorsal view sub-rectangular (temples not receding), eye height 2× temples; occipital carina complete and well defined, reaching hypostomal carina; oral space small and circular, maximum width about equal to basal width of mandible; clypeus not swollen; ocelli small, ocell–ocular distance 1.6× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing finely granulate, but occiput smooth and shining.

Mesosoma. Sculpturing mostly granulate; pronotum with some wrinkles; mesopleuron anterior corner rugose; propodeum more coarsely granulate, with long mid-longitudinal carina on basal 2/3; notauli deep and crenulate anteriorly, posteriorly meeting on depressed area; posterior margin of mesoscutum with short carina, just anterior to scutellar sulcus; scutellar sulcus with incomplete median carina, otherwise smooth.

Wings. Fore wing: stigma about 5× longer than high; vein r 0.65× vein 2RS, 0.8× vein RS+Mb, and 0.7× as long as vein m-cu; vein 3RSa about 0.3 times vein 3RSb, about 0.8× vein 2M; vein 1CUa about 2.5 times vein 1cu-a; vein 1Cub about 2.3 times 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu indicated as short pigmented not tubular vein just postfurcal to vein r-m; vein M+CU as long as vein 1M; vein 1M 1.8× vein r-m; vein RS smoothly curved at middle; vein 2-1A present as a very short stub.

Legs. Hind tibia without apical comb of modified setae; tarsal claw simple, not pectinate; apical spurs on hind tibia small, hind basitarsus 4× longer than inner spur.

Metasoma. T1–T3 rugose–striated with granulate background; remainder terga granulate; mid longitudinal carina complete from T1 throughout; metasoma outline



Figures 84–91. *Aleiodes nubicola* sp. n. **84** habitus **85** head occiput and vertex **86** apical flagellomeres **87** female holotype metasoma, dorsal **88** male paratype metasoma, dorsal **89** head and mesonotum, dorsal **90** host larva (Geometridae) **91** host mummy after parasitoid emergence.

narrow, petiole relatively small, T1 1.2× longer than its apical width, ovipositor sheaths as long as hind tarsomere II.

Paratype variation. Body length 3.8–4.6 mm; antenna with 38–40 antennomeres; head in most specimens mostly dark brown, except most of gena and temples just behind eyes honey yellow, but the gena is entirely dark brown in two type specimens; the clypeus color is also variable, in most specimens it is contrasting honey yellow, but in two specimens the clypeus has the same color of face; scutellum color varies from entire black to yellow on apical half, scutellar sulcus varies from yellow to black; light lateral stripes on mesopleuron varies from brown, reddish brown to yellow, in one specimen the stripes are connected by ventral yellowish stripe; most paratypes with metasomal terga 4 and 5 mostly blackish; position of hind wing vein m-cu varies from just postfurcal to just antefurcal; hind basitarsus 4–5× longer than inner apical spur on hind tibia.

Male. Essentially as in female, but eyes slightly smaller; antenna with 35 and 38 segments; metasomal terga entire dark brown in one specimen, the other with whitish central markings throughout all metasomal terga from apical tergite 1.

Mummy. Length 8.5–10.8 mm, entire brown, mummy with elongate aspect and thin skin, widening gradually from neck to posterior exit hole, thorax wrinkled, mummy attached to the substrate by silk posteriorly at prolegs region, exit hole irregular, located postero-dorsally anterior to abdominal prolegs, but in two specimens the hole is located postero-ventrally.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-47035, S00°35.9', W77°53.4', 2163 m, cloud forest, May 7, 2010.

Type-specimen: Holotype female and mummy, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) em. 7 May 2010 / YY-47035”; back (hand written): “Abr-2010 / 7-May-2010”. (UWIM)

Paratypes, 5 females and 3 males (UWIM), same data as holotype, different dates: 1♀ March 17, 2010, YY-45382; 2♀ May 4, 2010, YY-47069 and YY-46993; 1♀, October 8, 2010, YY-50796; 1♀, November 12, 2010, YY-52470; 1♂ March 18, 2010, YY-45448; 1♂ April 21, 2010, YY-46669; 1♂ April 22, 2010, YY-46568. 1♀, ECUADOR, Napo, Baeza, 2000m, Feb. '79 Mason. (CNC)

Biology. All specimens reared on the same Geometridae host caterpillar species (common name “palito café chusquea”) feeding on *Chusquea scandens* (Poaceae), collected from February to April. Consistent morphology of mummies and caterpillars support a single host species for this parasitoid. There is a variation in dorso-ventral orientation of the exit hole as observed in other species by M. Shaw (1983). All host caterpillars were collected during 2nd (all males and one female) or 3rd (only females) larval instars. Time span between host mummification and adult emergence varied mostly from two to three weeks, but one female took almost two months to emerge.

Discussion. *Aleiodes nubicola* sp. n. is similar to *A. cacuangoi* sp. n. and *A. atripileatus* (see diagnosis of *A. cacuangoi* for differences). This species also resembles *A. arbitrium* in the size of ovipositor and the mostly dark brown head. It differs from *arbitrium* by the longer malar space, about 1.6× longer than mandible width at base (about 1.0× in *arbitrium*), the mostly black pronotum and mesonotum (mostly brownish orange in *arbitrium*), and the position of light marks on head bordering eyes on temples (same marks on vertex in *arbitrium*). The host species “palito café chusquea” (Geometridae) is the same species attacked by *Aleiodes mirandae* sp. n. and *Aleiodes shakirae* sp. n.

Etymology. From Latin, means “cloud inhabiting”, a reference for the cloud forest habitat.

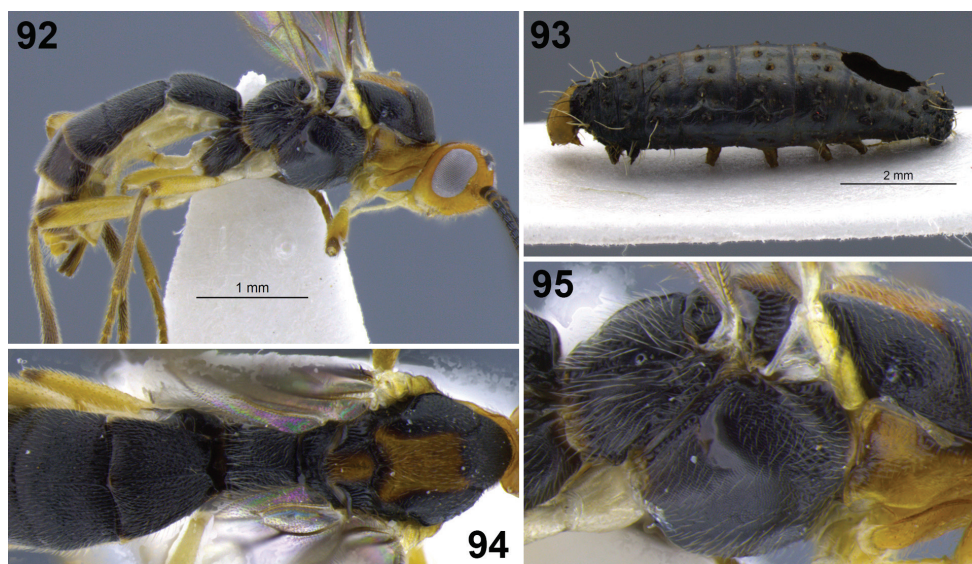
Aleiodes onyx sp. n.

<http://zoobank.org/6BBAEAE0E-F0BA-4CFF-B472-0FED7C7656DF>

http://species-id.net/wiki/Aleiodes_onyx

Figures 92–95

Description of holotype. Female (holotype). Body length 4.2 mm; antenna length 4.5 mm; fore wing length 4.1 mm.



Figures 92–95. *Aleiodes onyx* sp. n. **92** habitus **93** host mummy after parasitoid emergence **94** mesopleuron **95** mesosoma and metasomal terga 1–2, dorsal.

Color. Mostly black. Head orangish except black ocellar triangle; cheeks, palp and mandibles whitish; mesocutum with postero-median yellowish square mark; scutellum lighter; fore coxa and femur honey yellow; mid coxa, and all trochanter and trochantellus whitish; most of mid and hind femur whitish; all femur with small brown spot apical–dorsally; all tibia and tarsi brown; hind coxa black; hind tibial spurs and base of basitarsi honey brown; tip of metasoma behind T5 brown; ovipositor sheaths dark brown on apical half, basal half whitish.

Head. Antenna with 33 antennomeres, flagellomeres roughly $2.0\times$ as long as wide, apical flagellomere with small pointed tip; malar space moderate, length about $1.5\times$ basal width of mandible, and half eye height; in dorsal view eye height $2\times$ temples; occipital carina interrupted at vertex, ventrally reaching hypostomal carina; oral space small and circular, maximum width equal to basal width of mandible; clypeus not swollen; ocelli moderate, ocell–ocular distance as long as diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing finely granulate, but occiput smooth and shining.

Mesosoma. Sculpturing mostly granulate; pronotum with some wrinkles; mesopleuron with anterior corner rugose; propodeum more coarsely granulate, with long mid-longitudinal carina on basal $2/3$; notauli deep and crenulate anteriorly, meeting on depressed area posteriorly; posterior margin of mesoscutum with short carina, just anterior to scutellar sulcus; scutellar sulcus with median carina plus two pairs of poorly defined lateral carina.

Wings. Fore wing: stigma $5.5\times$ longer than high; vein r $0.75\times$ vein 2RS, $1.25\times$ vein RS+Mb, and $0.7\times$ as long as vein m-cu; vein 3RSa about $0.3\times$ vein 3RSb, and $0.8\times$ vein 2M; vein 1CUa $2.4\times$ vein 1cu-a; vein 1CUb $2\times$ 1CUa; vein 1M evenly slightly

curved. Hind wing: m-cu indicated as short pigmented not tubular vein just postfurcal to vein r-m; vein M+CU about as long as vein 1M; vein 1M 1.4× vein r-m; RS smoothly curved at middle; vein 2-1A present.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, not pectinate, with a comb of relatively long thin setae basally; hind basitarsus 3× longer than inner apical spur on hind tibia.

Metasoma. T1–T3 rugose–striated with granulate background; remainder terga granulate; mid longitudinal carina complete from T1 throughout T3; T1 0.8× longer than its apical width; ovipositor sheaths as long as hind tarsomere II.

Paratype variation. None observed, virtually identical to holotype.

Male unknown.

Mummy. Length about 6.5 mm, body entire graphite metallic black color, head orangish yellow, mummy aspect robust, body with one row of setal sockets on each segment and sparse setae except dorsally, wrinkles on thorax; exit hole irregular, located postero-dorsally.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-57102, S00°35.9', W77°53.4', 2163 m, cloud forest, July 11, 2011.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / CAPEA - NSF-BSI-07-17458 / (hand written) May 2011 / 57102”; back (hand written): “11-Ago-2011”. (UWIM)

Paratype, female: same data as holotype, except: July 8, 2011, YY-57101. (UWIM)

Biology. All type specimens were reared from the same Zygaenidae caterpillar species (“espalda tomate rubiacea”) feeding on *Notopleura plagiantha* (Rubiaceae). Parasitoid emerged about five weeks after host mummification. Five caterpillars were collected together from the same plant, suggesting gregarious feeding behavior by the host caterpillars; four of them were parasitized, but adult parasitoids emerged from only two.

Discussion. This species belongs to *circumscriptus*/gastritor species group. *A. onyx* sp. n. is similar to *A. atripileatus*; however, it can be distinguished by color patterns: head entirely orangish except ocellar triangle black (occiput mostly black in *atripileatus*), propleuron and ventral 1/4 pronotum honey brown (dark brown–black in *atripileatus*), mesopleuron whole black (ventral 1/2 honey brown in *atripileatus*), mesoscutum postero-central region honey yellow (whole dark-brown–black in *atripileatus*); as well as sculpturing features: mesopleuron central elevated area smooth (granulate in *atripileatus*), propodeum extensively rugose (granulate in *atripileatus*); and antenna with fewer flagellomeres: 31 in *onyx* sp. n. vs. 34 or more in *A. atripileatus*. Other diagnostic characters for *onyx* sp. n. are the occipital carina interrupted on vertex, the very short ovipositor, about as long as hind 3rd tarsomere, metasoma stout, T1 about 0.9× as long as its apical width and slightly wider than mesosoma. The host mummy is similar to that of *atripileatus*, but the anal prolegs are not posteriorly extended in mummies made by *onyx* sp. n. The base color of the mummy is a metallic graphite-like tone, as opposed to opaque black in *A. atripileatus* mummies. *A. onyx* sp. n. is the first *Aleiodes* species known from Ecuador to be reared from Zygaenidae caterpillars.

Etymology. From the Greek, the word *onyx* means “nail”. It is the name of a rock, used in adornments since ancient times, with several colors, being the black ones the most appreciated. The name is a reference for the main black color of this parasitoid mummy, which resembles the color of the black onyx rocks.

***Aleiodes shakirae* sp. n.**

<http://zoobank.org/DD5650C8-3A93-4088-88A2-300A07CE2557>

http://species-id.net/wiki/Aleiodes_shakirae

Figures 96–100, 120

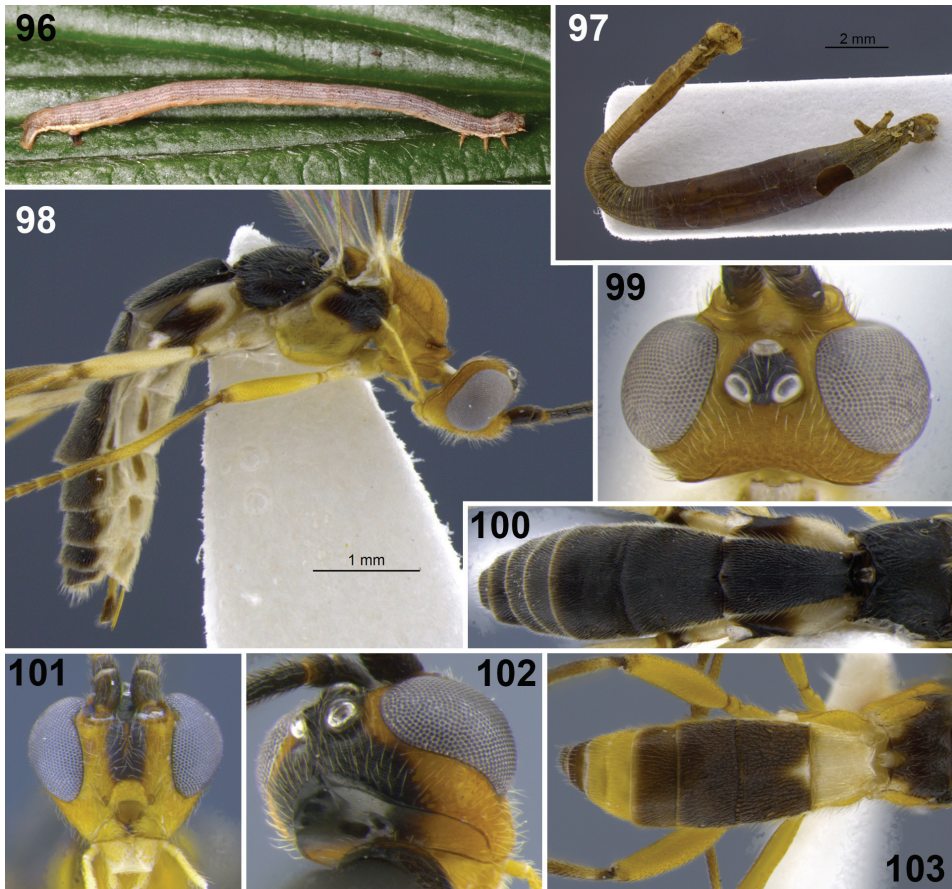
Description of holotype. Female (holotype). Body length 6.0 mm; antenna length 7.4 mm; fore wing length 5.7 mm

Color. Head yellow, ocellar triangle dark brown; antenna dark brown–black, scapus lighter dorsally; mesosoma yellow, anterior corner of mesopleuron, metanotum and part of lunules, propodeum and metapleuron dark brown–black, mesopleuron light yellow–whitish at broader with metapleuron but blackish in immediate border; fore and mid legs whitish at coxa, trochanter and trochantellus, darkening toward apex from light yellow to honey brown; hind leg: 1/2 basal coxa white, 1/2 apical black, trochanter white, trochantellus black with ventral white stripe, femur and tibia whitish with infuscate stains mid laterally and at apex, tibial spurs and telotarsi honey brown, remaining tarsi brown. Metasomal terga black, but T4–T7 white laterally; metasoma ventrally white; ovipositor sheaths dark brown on apical half, basal half whitish. Wings hyaline; stigma brown with central area lighter; most veins dark brown; vein C+SC+R black (extreme base whitish) connecting to a black parastigma with whitish central spot.

Head. Antenna with 47 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with long “bottle-nipple”-shaped apex; malar space as long as basal width of mandible, and approximately 1/4 eye height; in dorsal view eyes 3.4× temples; occipital carina incomplete dorsally, curving toward lateral ocelli, laterally complete and meeting hypostomal carina; oral space small and circular, maximum width slightly equal to basal width of mandible; clypeus not swollen; ocellus moderate, ocell–ocular distance short, about 0.45× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing finely granulate, occiput smooth and shining; higher face with some wrinkles just below toruli.

Mesosoma. Sculpturing finely granulate; propodeum with mid-longitudinal carina complete; notauli weak, shallow and smooth; posterior margin of mesoscutum with complete carina; scutellar sulcus with median carina plus two pairs of lateral carina.

Wings. Fore wing: stigma 4.5× longer than high; vein r 1.27× vein 2RS, 1.2× vein RS+Mb, and as long as vein m-cu; vein 3RSa about 0.44× vein 3RSb, and 0.8× vein 2M; vein 1CUa 1.6× vein 1cu-a; vein 1Cub 2.2× vein 1CUa; vein 1M strongly curved at basal portion. Hind wing: m-cu virtually absent; vein M+CU 1.4× vein 1M; vein 1M 1.2× vein r-m; RS smoothly curved at middle; vein M dark brown, well pigmented; vein 2-1A absent.



Figures 96–100. *Aleiodes shakirae* sp. n. **96** host larva (Geometridae) **97** host mummy after parasitoid emergence **98** habitus **99** head, dorsal **100** metasoma, dorsal. **Figures 101–103.** *Aleiodes speciosus* Townsend. **101** head frontal **102** head, occiput and vertex **103** metasoma, dorsal.

Legs. Hind tibia without comb of modified setae; tarsal claws pectinate basally, with several very short and tight bristles, longer and sparser apically, wide gap between apical bristles and claw; hind basitarsus 3× longer than inner apical spur on hind tibia.

Metasoma. T1–T3 granular–rugose; remainder terga granular; mid longitudinal carina complete from T1 throughout T3; ovipositor sheaths about as long as hind tarsomere II; T1 long and narrow, T1 length 1.7× its apical width.

Paratypes variation. Body length 4.5–6.2 mm; antenna with 47–50 segments; legs color somewhat darker in few paratypes, and/or mid trochantellus laterally infuscate; scutellar sulcus with 3 to 5 carina; other color and proportions with only minimal variation.

Male. Antenna with 44 or 46 segments; ocelli larger, ocell–cular distance 1/3 diameter of lateral ocellus; tergite 1 narrower, about 2× longer than apical width.

Mummy. Length 14.0–18.7 mm, dark reddish brown medially, light brown anteriorly and posteriorly, thorax narrow and wrinkled, mummy withered posteriorly be-

hind exit hole, general long and narrow aspect, curved down- or sideward “V-shaped” to almost straight, exit hole irregular, located postero-dorsally, anterior to prolegs, but one specimen with ventral exit hole.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, canopy Malaise trap on bamboo, S00°35.9', W77°53.4', 2163 m, cloud forest, July 10–16, 2010, S.R: Shaw col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 10-16 June 2010, Scott R. Shaw / canopy Malaise trap, bamboo / NSF-BSI-07-17458 expedition”. (UWIM)

Paratypes 5 females and 6 males (UWIM), same data as holotype, except: 1♀, June 1–8, 2007, Malaise trap, SRS-00047; 3♀ and 6 males, reared from Geometridae larvae on *Chusquea scandens* (Poaceae): 1♂ June 2, 2006, YY-14220; 1♂ March 25, 2008, YY-29593; 1♂ August 26, 2009, YY-40478; 1♀ January 30, 2010, YY-44297; 1♂ July 29, 2010, YY-48878; 1♂ September 9, 2010, YY-50943; 1♂ November 16, 2010, YY-52497; 1♀, January 7, 2011, YY-52852; 1♀ March 10, 2011, YY-54224; 1♀ September 3, 2013, YY-78769.

Biology. Reared from Geometridae caterpillar feeding on *Chusquea scandens* (Poaceae). Most host caterpillars were commonly named “palito café chusquea.” Considering associated caterpillar pictures, rearing information, and similar mummy morphology, it is likely that all the hosts were conspecific. Host caterpillars were collected in 2nd and 3rd instars. Time span, from pupation until adult emergence, varied from 2.5 up to 6 weeks. As the mummy dries, the middle section bends and dries in different ways, resulting in different but distinctive bent mummies.

Discussion. *Aleiodes shakirae* sp. n. belongs to *circumscriptus/gastritor* species group. The “V-shaped” mummies from Geometridae and some color features resemble *A. townsendi* sp. n.; however, *A. shakirae* sp. n. differs from *townsendi* sp. n. in the ocellular distance, which is about half the diameter of the lateral ocellus, but is nearly the same length as the lateral ocellus in *townsendi* sp. n. In *shakirae* sp. n. the metapleuron and metasoma are dorsally black (in *townsendi* sp. n. the metapleuron and T1 are mostly white with black markings), the hind coxa basally white and apically black (colors inverted in *townsendi* sp. n.), and hind tibia and tarsi are lighter in *shakirae* sp. n., the hind tibial spurs are yellowish to honey yellow (as compared with black in *townsendi*). The sculpturing of metasomal terga 1–3 in *shakirae* sp. n. is granular–rugose (rugose–striate in *townsendi* sp. n.), the metasoma is much slender in *A. shakirae* sp. n., petiole is 1.7–2.0 times longer than apical width (as compared with 1.2 times in *townsendi* sp. n.), and the hind wing m-cu is absent in *shakirae* sp. n. (present as a short pigmented stub in *townsendi* sp. n.). The host species “palito café chusquea” (Geometridae) is the same species attacked by *Aleiodes nubicola* sp. n. and *Aleiodes mirandae* sp. n.

Etymology. This species is named after the famous Colombian singer Shakira. Since parasitism by this species causes the host caterpillar to bend and twist its abdomen in various ways, and Shakira is also famous for her belly-dancing, the name seems particularly appropriate for this species.

***Aleiodes speciosus* Townsend, 2009**

http://species-id.net/wiki/Aleiodes_speciosus

Figures 101–103

Diagnosis. Body length 4.9 mm; antenna with 42 segments; head color honey brown with black mark covering most of occiput, frons and vertex, and face medially; malar space $1.3\times$ basal width of mandible; ocellus large, ocell–ocular distance $0.6\times$ width of lateral ocellus; occipital carina shortly interrupted at vertex; mesopleuron mostly smooth, rugose on anter-dorsal corner; apex of hind tibia without comb of flattened setae; propodeum granulate anteriorly and rugose posteriorly, longitudinal carina complete; metasomal tergite 1 white contrasting with most dark-brown–brown remainder terga, except terga 5–6 yellow.

Additional characters. Last flagellomere with short pointed tip; mesoscutum with carina at posterior margin almost complete but weakly defined; scutellar sulcus shallow, with incomplete median carina; wings moderately infusate; fore wing vein 1M moderately curved at base; hind wing vein 2-1A present, vein m-cu absent.

Type material examined. (UWIM)

Biology. *Aleiodes speciosus* has been reared from unidentified Lepidoptera caterpillar on *Miconia* sp. (Melastomataceae).

Distribution. Known only from the type locality, Camino a Loreto, Napo province, ECUADOR, at 1,383 meters elevation.

Discussion. *Aleiodes speciosus* is known only by the male holotype. This species belongs to the *circumscriptus/gastritor* species-group. *A. speciosus* is similar to *kingmani* sp. n. mainly by color pattern, but also in having relatively large ocelli. These two species have a white metasomal tergite 1, contrasting with the remainder dark brown terga; however the mesosoma laterally and ventrally, and hind coxa of *speciosus* is mostly yellowish, compared with being almost entirely black in *kingmani* sp. n. The mummy produced by *speciosus*, although decapitated, is noticeably distinct from the ventrally bent “J-shaped” mummy of *kingmani* sp. n.

***Aleiodes stewarti* sp. n.**

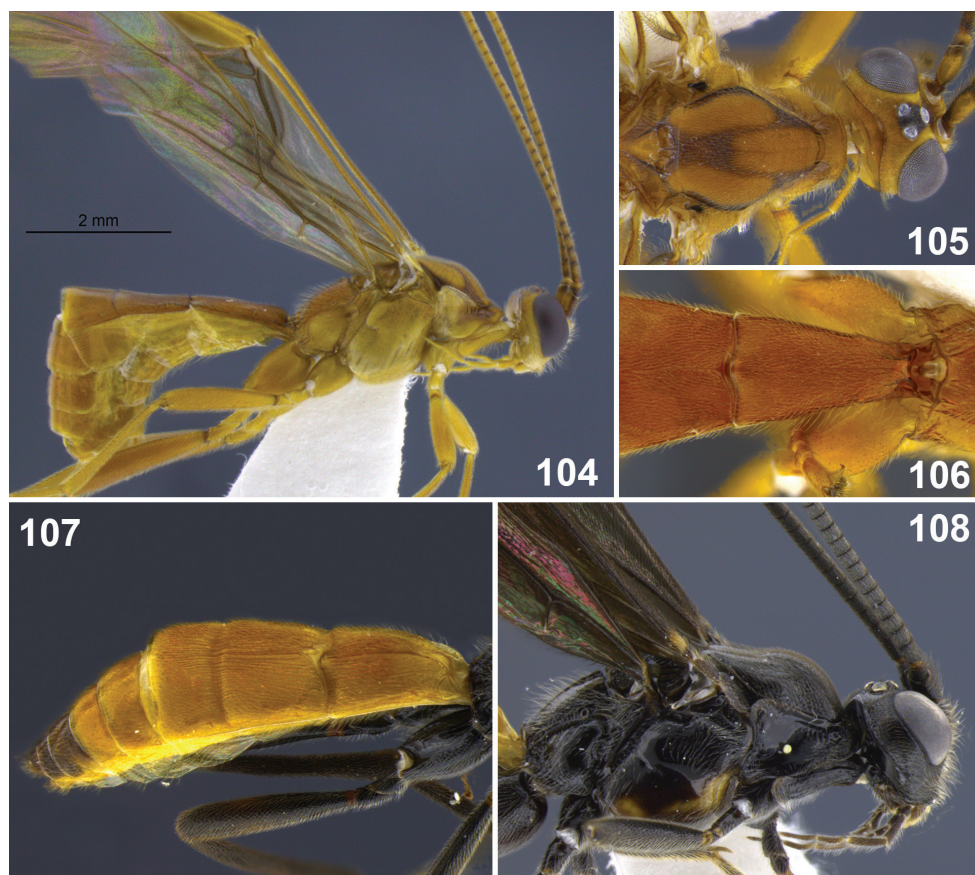
<http://zoobank.org/FE5FEE90-69ED-466B-88EA-7E9D52E58ADB>

http://species-id.net/wiki/Aleiodes_stewarti

Figures 104–106, 125

Description of holotype. Female (holotype). Body length 9.1 mm; antenna length 9.4 mm; fore wing length 7.4 mm.

Color. Entire body yellowish brown to honey brown, darker dorsally; antenna basally dark brown, lightening gradually toward apex, apical $1/3$ pale brown, scape light brown dorsally; face pale yellow, ocellar triangle black; lateral borders of mesoscutum, notauli and posterior depressed area brown; ovipositor sheaths mostly dark brown,



Figures 104–106. *Aleiodes stewarti* sp. n. **104** habitus **105** head and mesonotum, dorsal **106** metasomal tergite 1 **107, 108** *Aleiodes stilpnos* Townsend. **107**, metasoma **108** head and mesosoma, lateral.

basally whitish; wings slightly brown infuscate; veins dark brown except C+SC+R, stigma and R1 honey yellow.

Head. Antenna with 61 segments; flagellomeres about as long as wide, except apical 1/3 and basal 1/6 slightly longer than wide, apical flagellomere with small pointed apex; malar space as long as basal width of mandible, and $0.3\times$ eye height; temple narrow, in dorsal view about eyes $5\times$ longer than temples; occipital carina complete, reaching hypostomal carina; oral space small and circular, diameter about equal to basal width of mandible; clypeus weakly swollen; ocelli moderate, ocell–ocular distance about $1/2$ diameter of lateral ocellus; face and gena rugose–costate, with mid-longitudinal ridge just below toruli, frons smooth and excavated, bordered by weak “W-shaped” carina; temples and vertex granulate.

Mesosoma. Sculpturing mostly granular; pronotum with median scrobiculate line; mesopleuron mostly shining granular–coriaceous, antero-dorsal corner rugose, central elevated area sharply defined and smooth, epicnemial carina complete; propodeum on

posterior 1/3 smooth with longitudinal wrinkles, mid-longitudinal carina on anterior 2/3; metapleuron rugose posteriorly; notauli shallow and crenulate anteriorly, meeting rugose depressed area posteriorly; posterior margin of mesoscutum with carina interrupted laterally; scutellar sulcus with long median carina plus two pairs of incomplete lateral carina.

Wings. Fore wing: stigma $3.7\times$ longer than high; vein r $0.55\times$ length of 2RS, $0.45\times$ length of m-cu, and $0.7\times$ vein RS+Mb; vein 3RSa $0.46\times$ vein 3RSb, and $0.88\times$ vein 2M; vein 1CUa about $2\times$ vein 1cu-a; 1Cub $1.6\times$ length of 1CUa; vein 1M moderately curved at basal half. Hind wing: marginal cell widening toward apex, vein RS smoothly curved downward on base and well pigmented throughout; vein M+CU $1.3\times$ longer than 1M; vein 1M about $1.4\times$ longer than r-m; vein m-cu short, pigmented and non-tubular; vein 2-1A present and relatively long.

Legs. Tarsal claws strongly pectinate, with several relatively short bristles extending over the base of apical claw; basitarsus $3\times$ longer than inner apical spur of hind tibia.

Metasoma. T1 and T2 striate, longitudinal carina complete on T1, incomplete on posterior $\frac{1}{2}$ of T2; T3 with weak striation on anterior corners; remainder visible terga smooth; petiole long, very narrow basally, T1 $1.7\times$ longer than apical width; ovipositor sheaths about as long as hind tarsomere III.

Paratype variation. Essentially as holotype but antennomeres 60–64, scutellar sulcus with one or two pairs of lateral carina more or less defined and incomplete.

Male unknown.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, Macucoloma trail, S00°35.9', W77°53.4', 2163 m, cloud forest, January 1–8, 2007, J. Simbaña col.

Type-specimen: Holotype female, point mounted. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4', 2163m / 1-8 January 2007, J. Simbaña / Macucoloma trail, Malaise trap / NSF-BSI-07-17458, S.R. Shaw”. (UWIM).

Paratypes. 2♀, same data as holotype; 3♀, same data as holotype, except: 1♀, black light, May 15, 2011, S.R. Shaw col; 1♀, June–December 2011, canopy malaise trap (*Chusquea*); 1♀, September 5, 2005, malaise trap (Pumayacu ridge). (UWIM)

Discussion. *Aleiodes stewarti* sp. n. belongs to the *pulchripes* species-group. In the key to New World species (S. Shaw et al. 1997), this species will run to *Aleiodes rossi* Marsh & Shaw, 1997. *A. stewarti* sp. n. can be distinguished from *rossi* by its mostly bronze color with a distinct color pattern on mesoscutum: notauli+posterior depressed area brown, all wing veins brown and antenna lightening apically (in *rossi* the body is entirely light yellow with all tarsi brown and apex of hind tibia black, the antenna is entirely brown with scape and pedicel yellowish). The bristles in tarsal claws in *stewarti* sp. n. are more numerous than in *rossi* and shortening apically, very similar to those on *Aleiodes cazieri* Marsh & Shaw, 1997 and *Aleiodes vaughani* Muesebeck 1960. The first tergite of *stewarti* sp. n. is about $1.7\times$ longer than its apical width, distinctly more slender than the previous described species in *pulchripes*-group, in which this proportion is around $1.0\times$, but in *colberti* sp. n. it is $1.5\times$. *A. stewarti* sp. n. and *colberti* sp. n.

are the only two species in *pulchripes* species-group found so far from Yanayacu. *A. stewarti* sp. n. differs from *colberti* sp. n. in the antenna dark brown basally, gradually lightening toward pale brown apex (black with mid white band in *colberti* sp. n.), wings uniformly weakly infuscate (with dark band below stigma in *colberti* sp. n.), ocelli about 2× ocell–ocular distance (about 8× in *colberti* sp. n.); body mostly honey brown (reddish brown in *colberti* sp. n.), tarsal claw pectination with several short bristles extending to base of claw (in *colberti* sp. n. the pectination have less and larger bristles, and a distinct gap with claw base).

Comments. Since *stewarti* sp. n. is described based on several females and *colberti* sp. n. is described based on one male, and considering the geographical distribution and sexual dimorphism in the group, there is a possibility of these two species are one single species with very extreme sexual dimorphism. However, we do not think that it is likely because none of the known species in *pulchripes* species-group, having both males and females described, exhibit anything close to such extreme variation, which compels us to maintain these two entities as distinct species.

Etymology. This species is named after Jon Stewart (John Stuart Leibowitz), an American comedian, political satirist, writer, director, actor, and television host of *The Daily Show*.

Aleiodes stilpnos Townsend, 2009

http://species-id.net/wiki/Aleiodes_stilpnos

Figures 107, 108, 126

Diagnosis. Body length 9.0 mm; antenna with 65 segments; head and mesosoma color mostly black, contrasting with yellowish orange metasoma; malar space 1.3× basal width of mandible; ocellus moderate, ocell–ocular distance slightly longer than width of lateral ocellus; occipital carina complete; mesopleuron smooth; apex of hind tibia without comb of flattened setae; propodeum mostly rugose, longitudinal carina complete.

Additional characters. Last flagellomere with “bottle-nipple”-shaped tip; mesoscutum with carina at posterior margin present only in front of scutellar sulcus; scutellar sulcus with three strong and short carina; fore wing vein 1M slightly curved at base; hind wing vein 2-1A present and relatively long, vein m-cu absent.

Type material examined. (UWIM)

Biology. *Aleiodes stilpnos* has been reared from unidentified Noctuidae caterpillar on *Polygonum punctatum* (Polygonaceae).

Distribution. Known only from the type locality, YBS, Napo province, ECUADOR.

Discussion. *Aleiodes stilpnos* is the only species treated in this work in the *albitibia* species-group. The *albitibia*-group is relatively rarely collected in the Neotropical region, with only two described – *A. stilpnos* from Ecuador and *Aleiodes fuscipennis* (Szépligeti, 1904) from Peru, Venezuela (Torres and Briceño 2005) and Chile (deposited at UWIM), and at least two undescribed species from Costa Rica. *A. stilpnos*

differs from other species in Ecuador by the following characters: head, mesosoma and legs mostly black contrasting with light orange metasoma, and wings infusate; mesopleuron central disc smooth and bare; tarsal claws strongly pectinate; and costate sculpturing on metasomal terga 1–3.

***Aleiodes townsendi* sp. n.**

<http://zoobank.org/DE46E183-E9B7-4C43-9DCA-CCAEC880B559>

http://species-id.net/wiki/Aleiodes_townsendi

Figures 109, 110

Description of holotype. Female (holotype). Body length 5.7 mm; antenna length 6.7 mm; fore wing length 5.1 mm.

Color. Head yellow, ocellar triangle dark brown; antenna dark brown–black, scapus lighter dorsally, extreme base of scapus and first flagellomere yellow; mesosoma yellow, anterior corner of mesopleuron, lunules, metanotum, propodeum and dorsal 1/4 of metapleuron dark brown, remainder metapleuron and mesopleuron, at border with metapleuron, white; latero-ventral mesopleuron with slightly lighter stripe; fore leg yellow, telotarsus brown; mid coxa, trochanter, trochantellus and femur basally white, remainder mid leg yellow, darkening toward apex, telotarsi and tibial spurs brown; hind leg: coxa black on 1/2 basal and white on 1/2 apical, trochanter and trochantellus black, but apical border of trochanter, and trochantellus ventral-apical 1/3 plus a small ventral spot white; femur black on basal 2/5 and at extreme apex, otherwise white; tibia and tarsi black, small white subbasal band on tibia. Metasoma white with black dorsal triangle beginning on central apex of T1 and covering most dorsal portion of subsequent terga; ovipositor sheaths dark brown on apical half, basal half whitish. Wings weakly infusate; most veins and stigma dark brown; vein C+SC+R black (extreme base whitish) connecting to a black parastigma with whitish central spot.

Head. Antenna with 46 antennomeres, flagellomeres roughly 2.0× as long as wide, apical flagellomere with “bottle-nipple”-shaped apex; malar space as long as basal width of mandible, and 1/3 eye height; in dorsal view eye height 2.8× temple; occipital carina incomplete dorsally, curving toward lateral ocelli, laterally complete and meeting hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus not swollen; ocellus moderate, ocell–ocular distance 0.8× diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing finely granulate, occiput smooth and shining; higher face with some transverse wrinkles just below toruli.

Mesosoma. Sculpturing finely granulate; propodeum more coarsely granulate, with mid-longitudinal carina complete and some diverging wrinkles posteriorly; notauli weak, shallow and smooth; posterior margin of mesoscutum with complete carina; scutellar sulcus with median carina plus one pair of weak lateral carina.

Wings. Fore wing: stigma 4.5× longer than high; vein r 1.4× vein 2RS, 1.5× vein RS+Mb, and about as long as vein m-cu; vein 3RSa about 0.5× vein 3RSb, and 0.85×



Figures 109–113. 109, 110 *Aleiodes townsendi* sp. n. 109 habitus 110 host mummy after parasitoid emergence. 111–113 *Aleiodes tzantza* sp. n. 111 habitus 112 host mummy 113 scutellar sulcus, arrows indicating carina at posterior margin of mesoscutum.

vein 2M; vein 1CUa $1.7\times$ vein 1cu-a; vein 1CUB $1.7\times$ vein 1CUa; vein 1M strongly curved at basal portion. Hind wing: m-cu indicated as short pigmented not tubular vein interstitial to vein r-m; vein M+CU $1.3\times$ vein 1M; vein 1M $1.4\times$ vein r-m; RS smoothly curved at middle; vein M dark brown, well pigmented; vein 2-1A absent.

Legs. Hind tibia without comb of modified setae; tarsal claw pectinate basally, with several very short and tight bristles, longer and more sparse apically, wide gap between apical claw and basal pectination; hind basitarsus about $3\times$ longer than inner apical spur on hind tibia.

Metasoma. T1–T3 rugose–striate; remainder terga coriaceous; mid longitudinal carina complete from T1 throughout T3; ovipositor sheaths about as long as hind tarsomere II; T1 length $1.2\times$ its apical width.

Paratype variation. Antenna broken, otherwise essentially as holotype.

Male unknown.

Mummy. Length 17.0 mm, entire mummy mottled with gray and brownish, thorax narrow and wrinkled, mummy withered posteriorly behind exit hole, mummy as-

pect long and narrow, curved upward “V-shaped”, exit hole irregular, located postero-dorsally, anterior to prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-44074, S00°35.9', W77°53.4', 2163 m, cloud forest, January 23, 2010.

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Province / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / 23 December 2010, Yanayacu Road / YY-44074, ex. Geometridae”. (UWIM)

Paratype, female, same data as holotype, except: August 3, 2005, Malaise trap, A. Townsend col. (UWIM)

Biology. Reared from a Geometridae caterpillar (no common name) on *Dendrophobium lloense* (Asteraceae). Time span from host mummification until adult emergence was 20 days.

Discussion. *Aleiodes townsendi* sp. n. belongs to the *circumscriptus/gastritor* species group. It is similar to *A. shakirae* sp. n. because of the presence of a strongly curved vein 1M in the fore wing, some similar color patterns, and the elongate and curved “V-shaped” mummy. It differs from *shakirae* sp. n. by having the metasomal tergite 1 mostly white with a small mid-apical black spot and about as long as apical width, entirely black or dark brown and about 2× longer than apical width in *shakirae* sp. n. The metapleuron in *townsendi* sp. n. is bicolored, black and white, while entirely black in *shakirae* sp. n. *A. townsendi* sp. n. hind coxa is black basally and apically white, compared with the inverse color pattern in *shakirae* sp. n., and the wings are moderately infuscate, while hyaline in *shakirae* sp. n. (additional features are cited in discussion section for *A. shakirae* sp. n.). The ocelli in *A. townsendi* sp. n. are relatively small, but its large eyes, almost 3× longer than temple in dorsal view, make the ocell–ocular distance shorter, the width of lateral ocellus being roughly equal to ocell–ocular distance.

Etymology. This species is named after Andrew Townsend, collector of one of the type specimens, for his contributions to the knowledge of the Ecuadorian Braconidae fauna.

***Aleiodes tzantza* sp. n.**

<http://zoobank.org/9351F3F9-1AE2-4940-BC7B-CF592B3DBD2A>

http://species-id.net/wiki/Aleiodes_tzantza

Figures 111–113

Description of holotype. Female (holotype). Body length 5.0 mm; antenna length 5.8 mm; fore wing length 5.0 mm.

Color. Dorsally dark brown to black, except head; laterally and ventrally mostly yellowish to whitish. Head honey yellow; mandibles, cheeks, and palp whitish, but teeth brown; black stain dorsally, from ocellar triangle through occiput mid-dorsally. Mesosoma black dorsally; pronotum and propleuron whitish, but mid-dorsally pronotum dark brown; mesopleuron honey yellow; metapleuron black. Legs mostly

yellowish, darkening toward apex, from whitish coxa to light brown, tarsi and dark brown claws. Metasoma dark brown dorsally, apex lightening behind T3, ventrally whitish; ovipositor sheaths dark brown. Wings weakly infuscate, veins brown.

Head. Antenna with 42 segments, flagellomeres roughly $2.0\times$ as long as wide, apical flagellomere with short “bottle-nipple”-shaped apex; malar space moderate, length $1.3\times$ basal width of mandible, and $0.4\times$ eye height; in dorsal view eye height $2.6\times$ temple; occipital carina incomplete dorsally, otherwise well defined, reaching hypostomal carina; oral space small and circular, maximum width slightly smaller than basal width of mandible; clypeus not swollen; ocell–ocular distance $0.6\times$ diameter of lateral ocellus; maxillary palp not swollen. Head surface sculpturing finely granulate, vertex sculpturing coarser, face with irregular transverse rugosity concentrated near toruli, occiput smooth and shining.

Mesosoma. Sculpturing mostly finely granulate; propodeum rugose–granulate, with complete mid-longitudinal carina; notauli anteriorly, narrow, posteriorly indicated by carina and meeting in front of scutellar sulcus; mesiscutum mid-posterior area depressed and granular, posterior margin of mesoscutum bordered with complete carina; scutellar sulcus with short median carina.

Wings. Fore wing: stigma $3.8\times$ longer than high; vein r $0.9\times$ vein 2RS, as long as vein RS+Mb, and $0.7\times$ as long as vein m-cu; vein 3RSa about $0.45\times$ vein 3RSb, and as long as vein 2M; vein 1CUa $2\times$ vein 1cu-a; vein 1Cub $2.2\times$ vein 1CUa; vein 1M evenly slightly curved. Hind wing: m-cu indicated as short fold, interstitial to r-m; M+CU $1.3\times$ 1M; vein 1M $1.45\times$ vein r-m; RS smoothly curved at middle; vein 2-1A present as a short stub.

Legs. Hind tibia without comb of modified setae; tarsal claw simple, with a comb of thin bristles medially; hind basitarsus about $4\times$ longer than inner apical spur on hind tibia.

Metasoma. T1–T3 rugose–striated with granulate background; remainder terga granulate; mid longitudinal carina complete from T1 throughout T2, absent on T3; ovipositor sheaths about as long as hind tarsomere II; T1 $1.2\times$ longer than its apical width.

Paratype variation. Very similar to holotype, but with 41 antennomeres.

Male unknown.

Mummy. Length 9.0–10.0 mm, abdomen black, head, thorax, legs, and prolegs light pale brown, thorax compact and wrinkled, anal prolegs largely extended posteriorly, glue hole located ventrally on the thorax, exit hole irregular, located posterodorsally, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, YY-48320, S00°35.9', W77°53.4', 2163 m, cloud forest, June 5, 2010

Type-specimen: Holotype female and mummy, point mounted separately. Top label: “ECUADOR: Napo Prov. / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / REARED / (hand written) 48320”; back (hand written): “May 2010 / 5-May-2010”. (UWIM)

Paratype, female, same locality as holotype, August 14, 2005, reared YY-5189. (UWIM).

Biology. The holotype was reared from a mummified larva collected on *Paliourea ulloana* (Rubiaceae). The sampling date is listed in the database as May 12, 2010, and the adult emergence date is listed as May 5, 2010, so clearly one of these dates must be incorrect. Since this caterpillar record is nested within a large group of other caterpillar records also collected on May 12, 2010, it seems most likely that the emergence date was recorded incorrectly. Since the pupation date is assigned as May 18, 2010, this also corroborates that the emergence date could not possibly have been May 5, 2010. It seems most likely that the emergence month was recorded incorrectly and perhaps the actual emergence date was June 5, 2010. The holotype host is probably a Noctuidae due to similarity with the paratype mummy, which is possibly conspecific, identified as a Noctuidae. The host plant for the paratype is unknown, and it is not possible to determine if the holotype host caterpillar fed or not on *P. ulloana*.

Discussion. This species belongs to *circumscriptus/gastritor* species group. *Aleiodes tzantza* sp. n. is similar to *A. atripileatus*, from which it can be distinguished by the larger ocelli: ocell–ocular distance about half diameter of lateral ocelli (1.5 to 1.7× in *A. atripileatus*), and the color of mesopleuron entirely yellow (dark brown at least dorsally in *A. atripileatus*). *A. tzantza* sp. n. mummies are very similar to those of *atripileatus*, but the projecting anal prolegs are much longer in this species, and the body is distinctly longer. Its short ocell–ocular distance is similar to *A. speciosus*, from which it differs by having the mesopleuron surface granular (mostly smooth with anterior quarter rugose in *speciosus*) and entirely honey yellow (anterior 1/4 black in *speciosus*), pronotum whitish laterally (mostly dark brown in *speciosus*), and metasomal terga all dark brown (first tergite white in *speciosus*).

Etymology. “Tzantza” is the Shuar word for the ritual of reducing heads by a mummification process used by the Shuar, a people native from the current Ecuadorian Amazon territory, resulting in a shrunken mummy as the ones produced by the *Aleiodes* species.

***Aleiodes yanayacu* sp. n.**

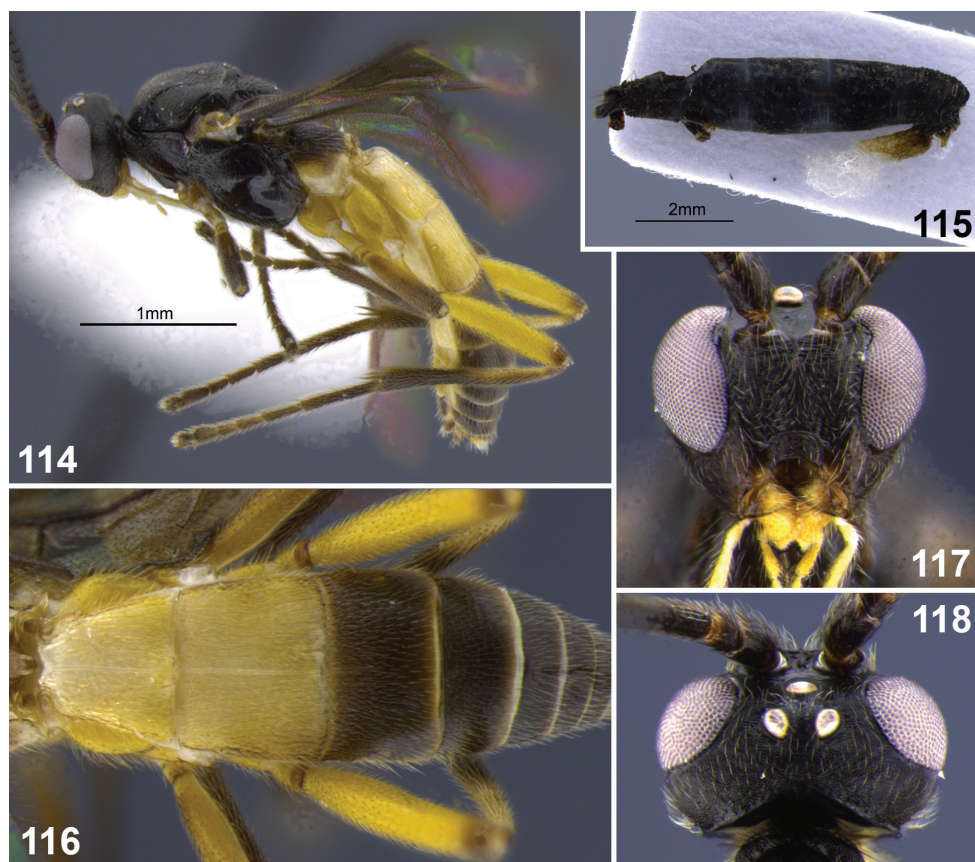
<http://zoobank.org/FEA654AE-3EF8-444B-839B-CE6B241EBED9>

http://species-id.net/wiki/Aleiodes_yanayacu

Figures 114–118

Description of holotype. Male (holotype). Body length 4.4 mm; antenna length 5.5 mm; fore wing length 4.5 mm.

Color. Mostly black. Propodeum with mid-apical white mark; metapleuron mostly whitish, but brown dorsally. Metasoma with T1 and T2 pale light yellow, reminder terga dark brown. Fore legs mostly dark brown, trochanter whitish with brown mark on inner side; mid legs mostly dark brown, coxa, trochanter and trochantellus yellowish, femur mostly light brown; hind legs yellowish basally, apex of femur with brown

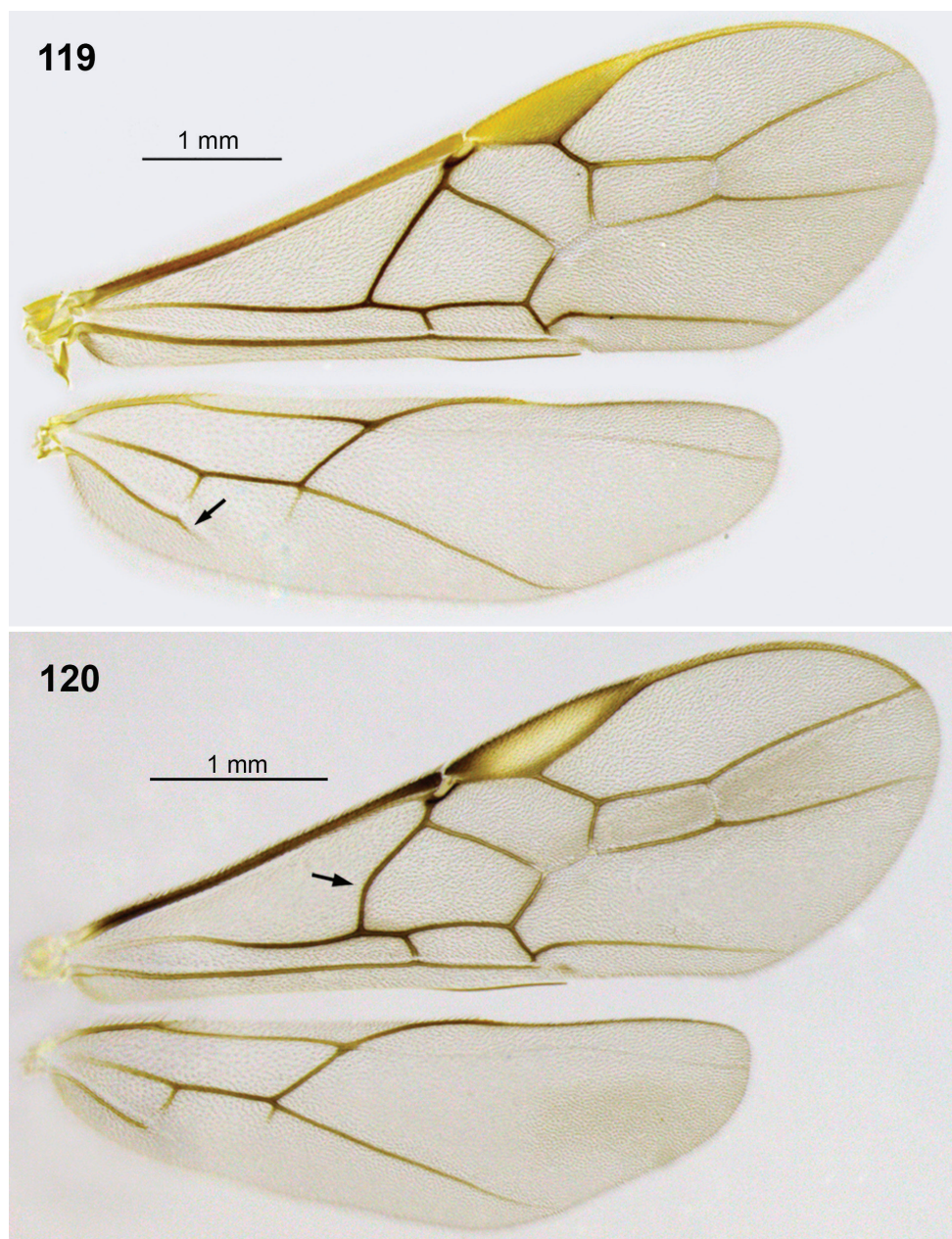


Figures 114–118. *Aleiodes yanayacu* sp. n. 114 habitus 115 host mummy after parasitoid emergence 116 metasoma, dorsal 117 head, frontal 118 head, dorsal.

stain dorsally, tibia and tarsi dark brown except for whitish stain covering basal half of inner side of tibia. Wings infusate with dark brown veins and stigma.

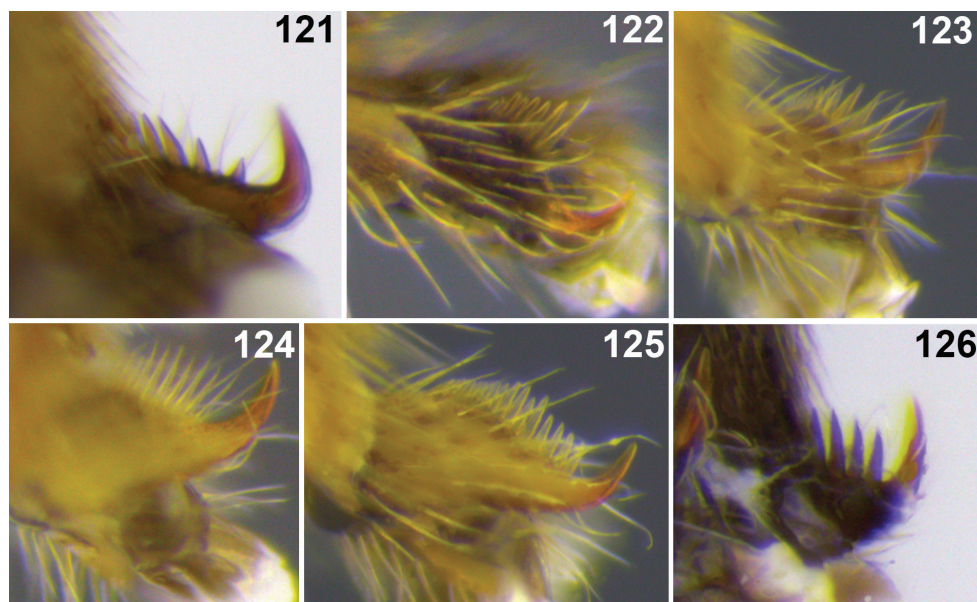
Head. Antenna with 42 antennomeres, flagellomeres roughly $2.0\times$ as long as wide, apical flagellomere with pointed tip; malar space about as long as basal width of mandible, and $0.4\times$ eye height; in dorsal view eye $1.4\times$ temple; occipital carina incomplete, interrupted at vertex, well defined laterally and meeting hypostomal carina; oral space small and circular, maximum width roughly equal to basal width of mandible; clypeus slightly swollen; ocell–ocular distance about $1.3\times$ diameter of lateral ocellus; maxillary palp not swollen; head surface sculpturing granulate, occiput smooth and shining; frons slightly excavated, shining coriaceous, sculpturing concentrically arranged; face with some transverse rugae medially, and a short but well defined mid-longitudinal carina higher on face, carina extending dorsally between toruli.

Mesosoma. Sculpturing finely granulate; pronotum foveate; mesopleuron central disc smooth and bare, antero-dorsal corner rugose; metapleuron smooth; propodeum coarsely granular with complete mid-longitudinal carina and some longitudinal rugosity posteriorly;



Figures 119–120. Wings. **119** *Aleiodes luteosicarius* sp. n., arrow = vein 2-1A, hind wing, fore wing **120** *Aleiodes shakirae* sp. n., arrow = vein 1M.

notauli well defined and crenulate anteriorly, barely defined but traceable posteriorly, meeting a rugose area; posterior margin of mesoscutum bordered with complete carina; scutellar sulcus with median carina plus two pairs of incomplete lateral carina.



Figures 121–126. Tarsal claws. **121** *Aleiodes colberti* sp. n. **122** *Aleiodes dyeri* sp. n. **123** *Aleiodes falloni* sp. n. **124** *Aleiodes luteosicarius* sp. n. **125** *Aleiodes stewarti* sp. n. **126** *Aleiodes stilpnos* Townsend.

Wings. Fore wing: stigma about $3.3\times$ longer than high; second submarginal cell relatively large; vein r $0.87\times$ vein 2RS, $1.4\times$ as long as vein RS+Mb, and $0.8\times$ as long as vein m-cu; vein 3RSa about $0.5\times$ vein 3RSb, and $0.9\times$ vein 2M; vein 1CUa $2.8\times$ vein 1cu-a; vein 1CUB about as long as vein 1CUa; vein 1M slightly curved at base. Hind wing: m-cu absent; M+CU as long as 1M; vein 1M $1.7\times$ vein r-m; vein RS smoothly curved at middle; vein M well pigmented; vein 2-1A present, short.

Legs. Hind tibia without apical comb of modified setae; tarsal claw simple, not pectinate, with a comb of relatively long thin setae basally; hind tibial spurs relatively short, about $1/4$ basitarsus length. Hind coxa smooth.

Metasoma. T1, T2 and basal $1/2$ of T3 costate, costa widely spaced over underlying granulation, sculpturing weaker at T3, longitudinal carina present along this sculpturing; remainder T3 and apical terga weakly shining coriaceous; T1 length about as long as its apical width.

Female unknown.

Mummy. Length 7.5 mm, head missing, black, exit hole irregular, located postero-dorsally, posterior to hind abdominal prolegs.

Type material. Type-locality: ECUADOR, Napo Province, Yanayacu Biological Station, S00°35.9', W77°53.4', 2163 m, cloud forest, 18 July 2013, reared from caterpillar: YY-75311.

Type-specimen: Holotype male, point mounted. Top label: "ECUADOR: Prov. Napo / Yanayacu Biological Station / S00°35.9', W77°53.4' 2163m / REARED: Jun 2013 / (hand written) 75311. Back: "18-Jul-2013". (UWIM)

Biology. Reared from a Geometridae species, feeding on *Phenax rugosus* (Urticaceae). Three weeks elapsed from mummification until adult emergence.

Discussion. *Aleiodes yanayacu* sp. n. belongs to *circumscriptus/gastritor* species-group. This species is similar to *A. mirandae* sp. n. and *A. napo* sp. n. in the mostly smooth mesopleuron, and the occipital carina, interrupted at vertex. *A. yanayacu* sp. n. differs from both species in having a slightly larger ocellus, the ocell–ocular distance is about $1.3\times$ the diameter of lateral ocellus, while in *mirandae* sp. n. and *napo* sp. n. it is about $2.0\times$. The hind wing vein M+CU is about as long as vein 1M in *yanayacu* sp. n., but distinctly shorter in *mirandae* sp. n. and *napo* sp. n. The sculpturing of metasomal terga 1 and 2 is also distinctive in *yanayacu* sp. n., with widely spaced costa, as compared with the finely rugose–costate sculpturing in *mirandae* sp. n. and *napo* sp. n. The metasomal tegite 2 is entirely withish in *yanayacu* sp. n. (black and white in *mirandae* sp. n. and *napo* sp. n.) and the hind coxa yellowish (black *mirandae* sp. n. and *napo* sp. n.).

Etymology. The species is named after the sampling and rearing location, the Yanayacu Biological Station.

Acknowledgments

Harold Greeney and Andrew Townsend are thanked for providing local arrangements and hospitality while at the Yanayacu Biological Station. This research was supported by National Science Foundation grants BSI -03-46729, BSI -07-17458, DEB-07-17034, DEB-10-20751 (Caterpillars and parasitoids in the Eastern Andes of Ecuador, CAPEA), and NSF Research Experience for Undergraduates (REU) supplemental grants DEB-08-23094, DEB-09-13110, and DEB-10-26103. Any opinions, findings, and conclusions expressed are those of the authors and do not necessarily reflect the views of the National Science Foundation. Travel to Ecuador was also supported, in part, by grants from U.W. International Programs, U.W. Environment and Natural Resources Program, Wyoming NASA Space Consortium, and U.W. Global Perspectives Program. Additional financial support was provided to EMS by the Science Without Borders program (213182/2013-0) of CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico), FAPESP (Fundação de Amparo à Pesquisa do Estado de São Paulo), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior), and INCT-HYMPAR SUDESTE (Instituto Nacional de Ciência e Tecnologia dos Hymenoptera Parasitoides da Região Sudeste Brasileira). Thanks to Lee Dyer and Suzane Rab Green for identification of host caterpillars, Donald Quicke for providing molecular data and helpful comments to determine some species limits, and Angélica Penteadó–Dias for all the support given and the unequalled contribution to the Neotropical fauna.

References

- Butcher BA, Smith MA, Sharkey MJ, Quicke DLJ (2012) A turbo–taxonomic study of Thai *Aleiodes* (*Aleiodes*) and *Aleiodes* (*Arcaleiodes*) (Hymenoptera: Braconidae: Rogadinae) based largely on COI barcoded specimens, with rapid descriptions of 179 new species. *Zootaxa* 3457: 1–232.
- Delfin–Gonzalez H, Wharton RA (2002) Distribution of species and species-groups of *Aleiodes* (Hymenoptera: Braconidae) in Mexico. *Folia Entomologica Mexicana* 41: 215–227.
- Fortier JC, Shaw SR (1999) Cladistics of the *Aleiodes* Lineage of the Subfamily Rogadinae (Hymenoptera: Braconidae). *Journal of Hymenoptera Research* 8: 204–237.
- Fortier JC (2009) A Revision of the *Tetrasphaeropyx* Ashmead Lineage of the Genus *Aleiodes* Wesmael (Hymenoptera: Braconidae: Rogadinae). *Zootaxa* 2256: 1–126.
- Harris RA (1979) A glossary of surface sculpturing. *Occasional Papers in Entomology of the California Department of Food and Agriculture* 28: 1–31.
- Marsh PM, Shaw SR (1998) Revision of North American *Aleiodes* (Part 3): the *seriatus* species-group. *Proceedings of the Entomological Society of Washington* 100: 395–408.
- Marsh PM, Shaw SR (1999) Revision of North American *Aleiodes* (Part 5): the *melanopterus* species-group. *Journal of Hymenoptera Research* 8: 98–108.
- Marsh PM, Shaw SR (2001) Revision of North American *Aleiodes* (Part 6): the *gasterator* and *unipunctator* species-group. *Proceedings of the Entomological Society of Washington* 103: 291–307.
- Marsh PM, Shaw SR (2003) Revision of North American *Aleiodes* (Part 7): the *compressor*, *ufei*, *gressitti* and *procerus* species-groups. *Proceedings of the Entomological Society of Washington* 105: 698–707.
- Miller JS, Dyer L (2009) Special feature: diversity of insect – plant interactions in the eastern Andes of Ecuador. *Journal of Insect Science* 9 (26). <http://insectscience.org/9.26>, doi: 10.1673/031.009.2601
- Quicke DLJ (2012) We Know Too Little about Parasitoid Wasp Distributions to Draw Any Conclusions about Latitudinal Trends in Species Richness, Body Size and Biology. *PLoS ONE* 7(2): e32101. doi: 10.1371/journal.pone.0032101
- Rab Green SB, Gentry GL, Greeney HF, Dyer LA (2011) Ecology, natural history, and larval descriptions of Arctiinae (Lepidoptera: Noctuiodea: Erebiidae) from a cloud forest in the eastern Andes of Ecuador. *Annals of the Entomological Society of America* 104: 1135–1148. doi: 10.1603/AN10165
- Sharkey MJ (1997) Key to New World subfamilies of the Braconidae. In: Wharton RA, Marsh PM, Sharkey MJ (Eds) *Manual of the New World Genera of the Family Braconidae* (Hymenoptera). International Society of Hymenopterists Special Publication, 39–63.
- Sharkey MJ, Wharton RA (1997) Morphology and terminology. In: Wharton RA, Marsh PM, Sharkey MJ (Eds) *Manual of the New World Genera of the Family Braconidae* (Hymenoptera). International Society of Hymenopterists Special Publication, 39–63.
- Shaw MR (1983) On[e] evolution of endoparasitism: the biology of some genera of Rogadinae (Braconidae). *Contributions of the American Entomological Institute* 20: 307–328.

- Shaw MR, Huddleston T (1991) Classification and biology of braconid wasps (Hymenoptera: Braconidae). Handbooks for the identification of British insects 1991 (7): 1–126.
- Shaw MR (1994) Parasitoid host ranges. In: Hawkins BA, Sheehan W (Eds) Parasitoid Community Ecology, Oxford, University Press, 111–144.
- Shaw MR (2002) Host ranges of *Aleiodes* species (Hymenoptera: Braconidae), and an evolutionary hypothesis. In: Melika G, Thuróczy C (Eds) Parasitic wasps: evolution, systematics, biodiversity and biological control. Agroiinform, Budapest, 322–327.
- Shaw SR (1993) Systematic status of *Eucystomastax* Brues and characterization of the Neotropical species (Hymenoptera: Braconidae: Rogadinae). Journal of Hymenoptera Research 2: 1–11.
- Shaw SR (1997) Subfamily Rogadinae. In: Wharton RA, Marsh PM, Sharkey MJ (Eds) Manual of the New World Genera of the Family Braconidae (Hymenoptera) 403–412. International Society of Hymenopterists Special Publication.
- Shaw SR (2006) *Aleiodes* wasps of eastern forests: a guide to parasitoids and associated mummified caterpillars. Forest Health Technology Enterprise Team, Technology Transfer Series, FHTET-2006-08, November 2006, 121 pp.
- Shaw SR (2012) A new genus and new species of the tribe Dinocampini from Napo Province in Ecuador (Hymenoptera: Braconidae: Euphorinae). International Journal of Tropical Insect Science 32: 101–107. doi: 10.1017/S1742758412000112
- Shaw SR, Marsh PM, Fortier JC (1997) Revision of North American *Aleiodes* (Part 1): the *pulchripes* species-group in the New World. Journal of Hymenoptera Research 6: 10–35.
- Shaw SR, Marsh PM, Fortier JC (1998a) Revision of North American *Aleiodes* (Part 2): the *apicalis* species-group in the New World. Journal of Hymenoptera Research 7: 62–73.
- Shaw SR, Marsh PM, Fortier JC (1998b) Revision of North American *Aleiodes* (Part 4): the *albitibia* and *praetor* species-groups in the New World. Proceedings of the Entomological Society of Washington 100: 553–565.
- Shaw SR, Marsh PM, Fortier JC (2006) Revision of North American *Aleiodes* (Part 8): the *coxalis* species-group. Zootaxa 1314: 1–30.
- Shaw SR, Marsh PM, Talluto MA (2013) Revision of North American *Aleiodes* (Part 9): the *pallidator* (Thunberg) species-group with description of two new species (Hymenoptera: Braconidae, Rogadinae). Zootaxa 3608(3): 204–214. doi: 10.11646/zootaxa.3608.3.4
- Shimbori EM, Pentead-Dias AM (2011) Taxonomic contribution to the *Aleiodes melanopterus* (Erichson) species-group (Hymenoptera, Braconidae, Rogadinae) from Brazil. ZooKeys 142: 15–25. doi: 10.3897/zookeys.142.1705
- Torres DN, Briceño RA (2005) Riqueza, abundancia y distribución de la subfamilia Rogadinae (Hymenoptera: Braconidae) en tres ecosistemas naturales de Venezuela. Entomotropica 20(3): 205–211.
- Townsend AC, Shaw SR (2009) Nine new species of *Aleiodes* Wesmael reared at Yanayacu Biological Station (Hymenoptera: Braconidae: Rogadinae) in eastern Ecuador. Journal of Insect Science 9(37): 1–22. <http://insectscience.org/9.37>, doi: 10.1673/031.009.3701
- van Achterberg C (1991) Revision of the genera of the Afrotropical and W. Palearctic Rogadinae Foerster (Hymenoptera: Braconidae). Zoologische Verhandlungen 273: 1–102.

- van Achterberg C (1995) Generic revision of the subfamily Betylobraconinae (Hymenoptera: Braconidae) and other groups with modified fore tarsus. *Zoologische Verhandelingen* 298: 1–242.
- van Achterberg C (1993) Illustrated key to the subfamilies of the Braconidae (Hymenoptera: Ichneumonoidea). *Zoologische Verhandelingen* 283: 1–189.
- Wharton RA (2006) The species of *Sternalopius* Fischer (Hymenoptera: Braconidae: Opiinae) and the Braconiae *Sternaulus*. *Journal of Hymenoptera Research* 15(2): 317–347.
- Yu D, van Achterberg K, Horstmann K (2012) *World Ichneumonoidea 2012: Taxonomy, Biology, Morphology and Distribution*. CD. Taxapad, Vancouver, Canada.
- Zaldívar-Riverón A, Shaw MR, Sáez AG, Mori M, Belokoblylskij SA, Shaw SR, Quicke DLJ (2008) Evolution of the parasitic wasp subfamily Rogadinae (Braconidae): phylogeny and evolution of lepidopteran host ranges and mummy characteristics. *BMC Evolutionary Biology* 8 (329): 1–20. doi: 10.1186/1471-2148-8-329

Revision of the genus *Paridea* Baly, 1886 from Taiwan (Coleoptera, Chrysomelidae, Galerucinae)

Chi-Feng Lee^{1,†}, Jan Bezděk^{2,‡}

1 Applied Zoology Division, Taiwan Agricultural Research Institute, 189 Chung-Cheng Road, Wufeng, Taichung 413, Taiwan **2** Department of Zoology, Mendel University, Zemědělská 1, 613 00 Brno, Czech Republic

† <http://zoobank.org/9F5D86AB-3DCE-4DE8-A079-75B2073BAB01>

‡ <http://zoobank.org/668F3A35-3E6E-40F3-9F06-356EEB50E45F>

Corresponding author: Chi-Feng Lee (chifeng@tari.gov.tw)

Academic editor: R. Beenen | Received 9 March 2014 | Accepted 12 April 2014 | Published 28 April 2014

<http://zoobank.org/1880D172-9007-46BE-B6A5-82EBAEDF0F9A>

Citation: Lee C-F, Bezděk J (2014) Revision of the genus *Paridea* Baly, 1886 from Taiwan (Coleoptera, Chrysomelidae, Galerucinae). ZooKeys 405: 83–125. doi: 10.3897/zookeys.405.7458

Abstract

The Taiwanese species of the genus *Paridea* Baly, 1886, are revised. Two new species, *Paridea* (*Semacia*) *houjyai* sp. n. and *P.* (*S.*) *kaoi* sp. n. are described. Both were confused previously with *P.* (*S.*) *sexmaculata* (Laboissière, 1930) and *P.* (*S.*) *angulicollis* (Motschulsky, 1854) respectively. *Paridea* (*P.*) *sauteri* (Chûjô, 1935) and *P.* (*P.*) *taiwana* (Chûjô, 1935) are removed from synonymy with *P.* (*P.*) *sinensis* Laboissière, 1930. The synonymy of *Paraulaca flavipennis* Chûjô, 1935 with *Paridea* (*Paridea*) *testacea* Gressitt & Kimoto, 1963 is supported. *Paridea* (*Semacia*) *nigrimarginata* Yang, 1991 is regarded as a junior synonym of *P.* (*S.*) *angulicollis* and excluded from the Taiwan fauna. Lectotypes are designated for *Paraulaca costata* Chûjô, 1935, *P. flavipennis* Chûjô, 1935, *P. taiwana* Chûjô, 1935, *Semacia nipponensis* Laboissière, 1930, and *Paridea sinensis* Laboissière, 1930.

Keywords

Paridea, *Semacia*, Taiwan, taxonomic revision

Introduction

The genus *Paridea* Baly, 1886, comprising 83 species (Bezděk, unpublished data), is one of the most species-rich genera of Oriental Galerucinae. The genus occurs in Nepal, Bhutan, India, Pakistan, China, Korea, Japan and Southeast Asia. *Paridea apicalis* (Jacoby, 1886), from New Guinea, evidently is not congeneric and is not counted. As in most large Oriental genera of Galerucinae, a comprehensive revision based on the study of primary types has not been performed. Recently, only two larger papers were devoted to *Paridea*: the Chinese species were revised by Yang (1991) and Medvedev and Samoderzhnikov (1997) provided a key to species from the Himalayas and adjacent regions.

Paridea Baly, 1886 is similar to other genera of the subtribe Aulacophorina, particularly to *Agetocera* Hope, 1831, *Aulacophora* Chevrolat, 1836, *Pseudocophora* Jacoby, 1884 and *Paragetocera* Laboissière, 1929. All these genera share unmarginated anterior pronotal margins, the presence of a transverse depression on the pronotum, and all feed on leaves of Cucurbitaceae (cf. Kimoto 1989b, Jolivet and Hawkeswood 1995). *Paridea* is separated easily from other genera by the appendiculate tarsal claws (bifid tarsal claws in others).

Laboissière (1930) described the first *Paridea* species from Taiwan as *Semacia sexmaculata* Laboissière. Chûjô (1935) added five species to the Taiwan fauna as *Paraulaca costata* Chûjô, *Paraulaca cyanipennis* Chûjô, *Paraulaca flavipennis* Chûjô, *Paraulaca sauteri* Chûjô, and *Paraulaca taiwana* Chûjô. Chûjô (1938) later listed a newly recorded species, *Paraulaca angulicollis* (Motschulsky, 1854). Kimoto (1966) synonymized *Paraulaca taiwana* Chûjô, 1935 with *Paraulaca sauteri* Chûjô, 1935; *Paraulaca flavipennis* Chûjô, 1935 with *Paridea testacea* Gressitt & Kimoto, 1963. *Paridea testacea* remains valid because *Paraulaca flavipennis* Chûjô, 1935 is a secondary homonym of *P. flavipennis* (Laboissière, 1930). Kimoto (1974) regarded *Paraulaca sauteri* Chûjô, 1935 as a junior synonym of *Paridea sinensis* Laboissière, 1930. Yang (1991) studied the Chinese species of *Paridea* and described a new species, *Paridea nigrimarginata* from Mt. Takao. Beenen (2010) added this species to Taiwan fauna since he assumed the type locality is in Taiwan. Thus, in total seven species have been described or recorded from Taiwan.

Taxonomic confusion and misidentification often occurs in this genus because some members have variable color patterns and some others have sexually dimorphic characters. Thus, robust sample sizes are required for revising this genus. The Taiwan Chrysomelid Research Team (TCRT) was founded in 2005 and is composed of 10 members. All of them are amateurs interested in making an inventory of all Chrysomelid species in Taiwan. Specimens of the genus have been extensively collected and studied, and host plants recorded. Diagnostic characters were assessed and the status of all species was evaluated based on a large series of specimens.

Materials and methods

More than 1000 specimens have been examined. Most of them (> 90%) were collected either by TCRT or belonged to the historical collection at the Taiwan Agricultural Research Institute (TARI).

To prepare drawings of the adult reproductive systems, the abdomens of adults were separated and boiled in a 10% KOH solution, cleared in distilled water, and then mounted on microscope slides in glycerin for observation. Specimens were examined and drawings were made using a Leica M165 stereomicroscope. Microscope slides were examined and illustrated using a Nikon ECLIPSE 50i microscope. Body parts were then stored in glycerin tubes with the dry mounted specimens.

Host plants are recorded by observing adult feeding behavior in the field. Plants were identified by Chih-Kai Yang.

Specimens examined are deposited at the following institutes and museums.

BMNH	The Natural History Museum, London, UK [Michael Geiser];
BPBM	Bernice P. Bishop Museum, Hawaii, USA [Shepherd Myers];
CAS	California Academy of Sciences, California, USA [David H. Kavanaugh];
FKCC	František Kantner collection, České Budějovice, Czech Republic;
HNHM	Hungarian Natural History Museum, Budapest, Hungary [Otto Merkl];
ISNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium [Pol Limbourg];
IZAS	Institute of Zoology, Academia Sinica, Beijing, China [YongYing Ruan];
JBCB	Jan Bezděk collection, Brno, Czech Republic;
KMNH	Kitakyushu Museum of Natural History, Kitakyushu, Japan [Kyoichiro Ueda];
MNHN	Museum National d'Histoire naturelle, Paris, France [Antoine Mantille-ri];
NMNS	National Museum of Natural Science, Taichung, Taiwan [Ming-Luen Jeng];
SDEI	Senckenberg Deutsches Entomologisches Institut, Müncheberg, Germany [Stephan Blank];
TARI	Taiwan Agricultural Research Institute, Taichung, Taiwan;
ZMUH	Zoologisches Institut und Zoologisches Museum, Universität von Hamburg, Hamburg, Germany [Hossein Rajaei and Kai Schuette];

Exact label data are cited for all type specimens of the described species; a double slash (//) divides the data on different labels and a single slash (/) divides the data in different rows. Other comments and remarks are in square brackets: [p] – preceding data are printed, [h] – preceding data are handwritten, [w] – white label, [y] – yellow label, [o] – orange label, [g] – green label, and [r] – red label.

Taxonomy

Classification of subgenera of *Paridea*

Five generic or subgeneric names (*Paraulaca* Baly, 1888, *Aeropa* Weise, 1889, *Semacia* Fairmaire, 1889, *Semacianella* Laboissière, 1930, and *Carapaula* Chûjô, 1962) were erected based on various sexually dimorphic characters of *Paridea*. Kimoto (1989b) and Medvedev and Samoderzhnikov (1997) used *Paridea* (s. str.), *Paraulaca* and *Semacia* as valid subgenera, while Yang (1991) considered only *Paridea* and *Semacia* as monophyletic after reviewing Chinese species. This arrangement was followed also by Beenen (2010) (with *Paraulaca*, *Carapaula* and *Semacianella* as synonyms of *Paridea* (s. str.) and *Aeropa* as synonym of *Semacia*) and is used also in our study but some characters are modified in the key

Key to species of the genus *Paridea* from Taiwan

- 1 Pygidium of both sexes projecting beyond elytral apices; apex of eighth abdominal tergite in male modified into two processes, apical margin without setae **2 (subgenus *Semacia*)**
- Pygidium of both sexes covered by elytra; apex of eighth abdominal tergite not modified, apical margin with setae..... **4 (subgenus *Paridea*)**
- 2 Female pygidium deeply emarginated (Fig. 4); elytron of male with one pre-median cavity near lateral margin (Fig. 3); without black spot on postscutellar common area in female (Fig. 4) ***P. (S.) houjayi* sp. n.**
- Female pygidium entire; elytron of male without a cavity near lateral margin but with postscutellar area depressed (Figs 6, 28, 31); flat and with black spot in female (Figs 9, 32, 33) **3**
- 3 Elytron with longitudinal black stripe along lateral margin from base to middle (Fig. 8) ***P. (S.) kaoi* sp. n.**
- Elytron without black stripe along lateral margin, but with one subbasal black spot near lateral margin (Fig. 30), sometimes extending inwards (Figs 31, 33)..... ***P. (S.) sexmaculata* (Laboissière)**
- 4 Elytron at least partly metallic blue or black **5**
- Elytron yellowish brown, sometimes with two pairs of black spots, in some individuals black spots extending to entire elytron except apex **6**
- 5 Vertex black; pronotum with one black spot at middle near basal margin; first tarsomeres of front and middle legs in male strongly swollen (Fig. 34); entire elytra metallic blue (Figs 34, 36) ***P. (P.) costata* (Chûjô)**
- Head and pronotum yellowish brown; first tarsomeres in male not modified (Fig. 52); lateral margin of elytron from base to apical 1/3 yellowish brown (Fig. 54)..... ***P. (P.) cyanipennis* (Chûjô)**
- 6 Femora and tibia yellowish brown, but outer margins black (Figs 74–79)..... ***P. (P.) taiwana* (Chûjô)**

- Femora yellowish; tibiae blackish brown or black 7
- 7 Elytra yellowish brown (Fig. 80) ***P. (P.) testacea* Gressitt & Kimoto**
- Elytra black, but apices yellowish brown (Fig. 55) ... ***P. (P.) sauteri* (Chûjô)**

***Paridea (Semacia) houjayi* sp. n.**

<http://zoobank.org/6D9F8E80-A589-489A-A9A0-E0129A38E5E8>

http://species-id.net/wiki/Paridea_houjayi

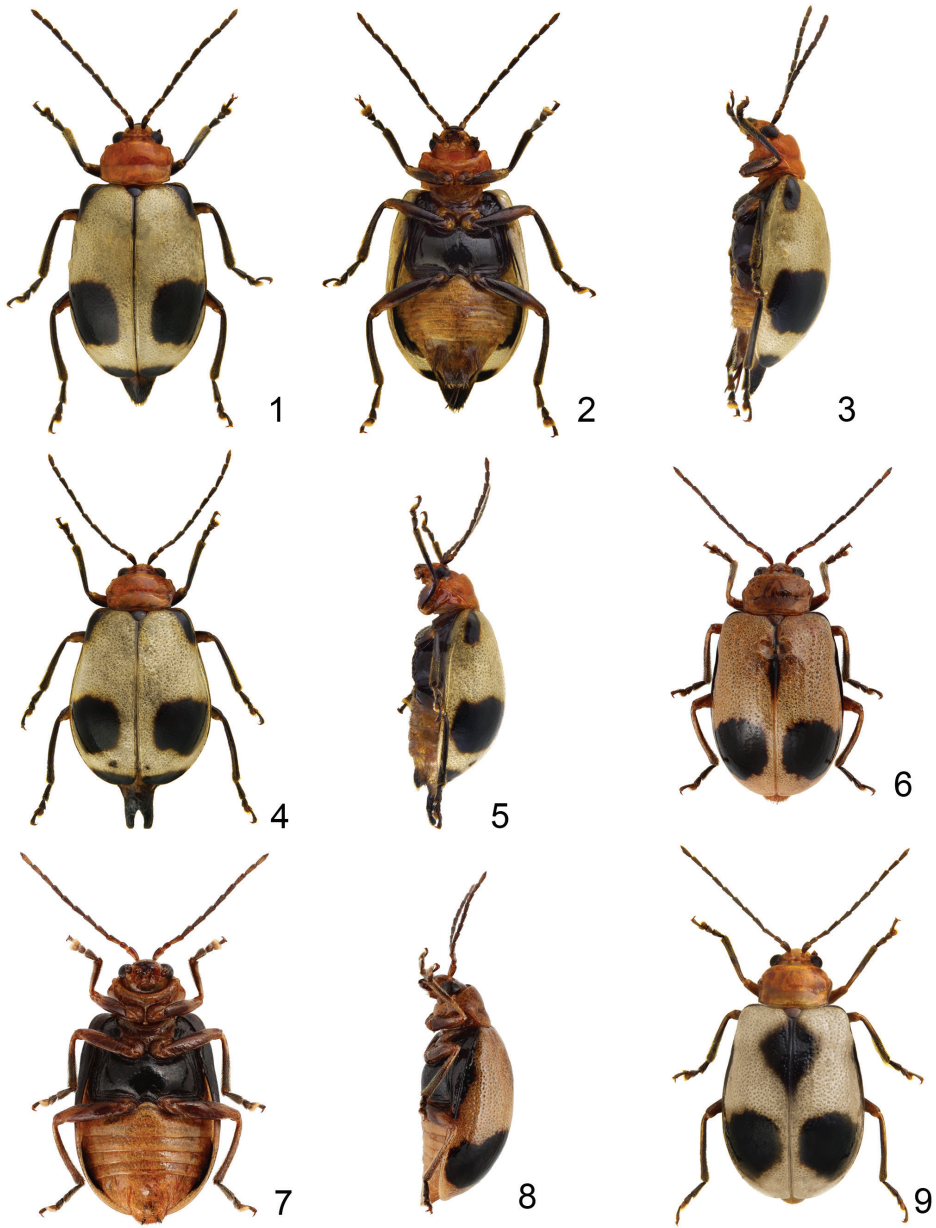
Paridea (Paraulaca) sexmaculata: Kimoto & Takizawa, 1997: 180 (misidentification)

Type locality. Taiwan, Ilan, Ssuchi, 24°29'N, 121°25'E, 700 m, broad-leaf forest.

Type material (n= 16). Holotype male (TARI), mounted on card, labeled: “Taiwan: Ilan (12458) [p] / Ssuchi (四季) [p] / 01.VIII.2009, leg. M.-H. Tsou [p, w] // **Holotypus** [p] / *Paridea (Semacia)* ♂ [p] / *houjayi* n. sp. [p] / des. C.-F. Lee, 2014 [p, r]”. Paratypes: 4♀♀, same data as holotype, but with “12459–12462” (TARI); 1♀: “Taiwan: Hsinchu (8723) [p] / Lupi (魯壁) [p] / 12.III.2009, leg. H. Lee [p, w]” (TARI); 1♂: “Taiwan: Ilan (4098) [p] / Fushan Chihwu Yuan [= Botanical Park] [p] / 01.IV.2008, leg. M.-H. Tsou [p, w]” (TARI); 1♂: “Hatonosawa [p] [= Chiuchihtse] / Mt. Taiheizan [p] [= Taipingshan] / 23.vii.1940 [p] / FORMOSA [p] / Col. M. CHUJO [p, w]” (TARI); 1♀: “Taiwan: Ilan (4712) [p] / Mingchi (明池) [p] / 27.IV.2008, leg. M.-H. Tsou [p, w]” (TARI); 1♂: “N. TAIWAN: Taipingshan [p] / 1950m. Ilan Hsien [p] / 26–28.VII.1983 [p] / L. Y. Chou [p, o]” (TARI); 1♂: “Taiwan: Taipei (4772) [p] / Fushan [p] / 30.IV.2008, leg. H.-J. Chen [p, w]” (TARI); 1♀: “Taiwan: Taipei (18945) [p] / Fushan (福山) [p] / 01.IV.2011, leg. S.-F. Yu [p, w]” (TARI); 1♂: “Taiwan: Taipei (989) [p] / Wulai [p] / 15.III.2007, leg. H.-J. Chen [p, w]” (TARI); 1♂: “Taiwan: Taipei [p] / Wulai [p] / 05.III.2009, leg. Y.-L. Lin [p, w]” (TARI); 1♀: “Taiwan: Taoyuan (3980) [p] / Hsuanyuan (萱源) [p] / 16.III.2008, leg. M.-H. Tsou [p, w]” (TARI); 1♂: “Taiwan: Taoyuan (4691) [p] / Tungyanshan (東眼山) [p] / 25.IV.2008, leg. H. Lee [p, w]” (TARI); 1♂: “TAIWAN, Ilan county, [p] / Mingchyh Forest [p] / Recreation Area, 1200 m, [p, w] // swept from vegetation, [p] / 5.IV.2002, [p] / leg. Gy. Fábíán & O. Merkl [p, w]” (HNHM). Each paratype has a type label: “**Paratypus** [p] / *Paridea (Semacia)* ♂ [or ♀] [p] / *houjayi* n. sp. [p] / des. C.-F. Lee, 2014 [p, pink label]”.

Diagnosis. This new species is recognized easily by the elytra of males possessing a lateral cavity near each lateral margin and the pygidium of each female with deeply emarginated apex.

Males. Length 6.2–6.3 mm, width 3.6–3.7 mm. General color (Figs 1–3) yellowish brown; antenna dark brown; scutellum black; elytra pale yellow, with one small, longitudinal black spot at humerus, one large, rounded black spot at postermedian area, apex black; mesepimeron and metathoracic ventrites black; outer margins of femora and tibiae black; tarsi dark brown. Elytron with one premedian cavity near lateral margin; three tufts of long hairs at anterior area and one tubercle at posterior



Figures 1–9. *Paridea* species. **1** *P. (S.) houjayi* sp. n., male, dorsal view **2** ditto, ventral view **3** ditto, lateral view **4** *P. (S.) houjayi* sp. n., female, dorsal view **5** ditto, lateral view **6** *P. (S.) kaoi* sp. n., male, dorsal view **7** ditto, ventral view **8** ditto, lateral view **9** *P. (S.) kaoi* sp. n., female, dorsal view.

of depression; postscular common area flat and impunctate. Eighth abdominal tergite (Fig. 12) strongly sclerotized, base extremely slender, with one pair of extremely long processes, each process flattened and widened near apex and outer margin of widened

area serrate. Pygidium projecting beyond elytral apices, apex shallowly emarginate. Penis (Figs 10–11) strongly asymmetric, dorsum with longitudinal groove at right side; almost straight from lateral view; apex forming angular process and directed ventrally; endophallic sclerites with two sclerites, one extremely elongate, about 0.75 times as long as penis, other curved and apically pointed.

Females. Length 6.2–6.9 mm, width 3.5–3.7 mm. Color (Figs 4–5) similar to male but elytra without excavation. Apical margin of last abdominal ventrite (Fig. 14) with deep notch at middle, between one pair of shallow processes. Pygidium projecting beyond elytral apices, deeply emarginate and forming bilobed process. Gonocoxae (Fig. 13) slender, apex of each gonocoxa with eight to nine setae from apical 1/4 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 15) weakly sclerotized; apex wide, apical margin truncate, surface with extremely dense long setae along apical margin, spiculum short. Spermathecal receptaculum (Fig. 16) strongly swollen; pump much longer than receptaculum, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Etymology. The new species name honors Mr. Hou-Jay Chen, who discovered this interesting new species.

Distribution. Taiwan. This new species occurs in northern Taiwan (Fig. 17).

Host plants. Cucurbitaceae: *Gynostemma pentaphyllum* (Thunb.) Makino; *Thladiantha nudiflora* Hemsl. ex Forbes & Hemsl.; Compositae: *Aster lasiocladus* Hayata.

Notes. Males of this new species were misidentified as *Paridea* (S.) *sexmaculata* by Kimoto and Takizawa (1997).

***Paridea* (*Semacia*) *kaoi* sp. n.**

<http://zoobank.org/9FB73A3A-10B1-4603-98D0-D2D8523F37AC>

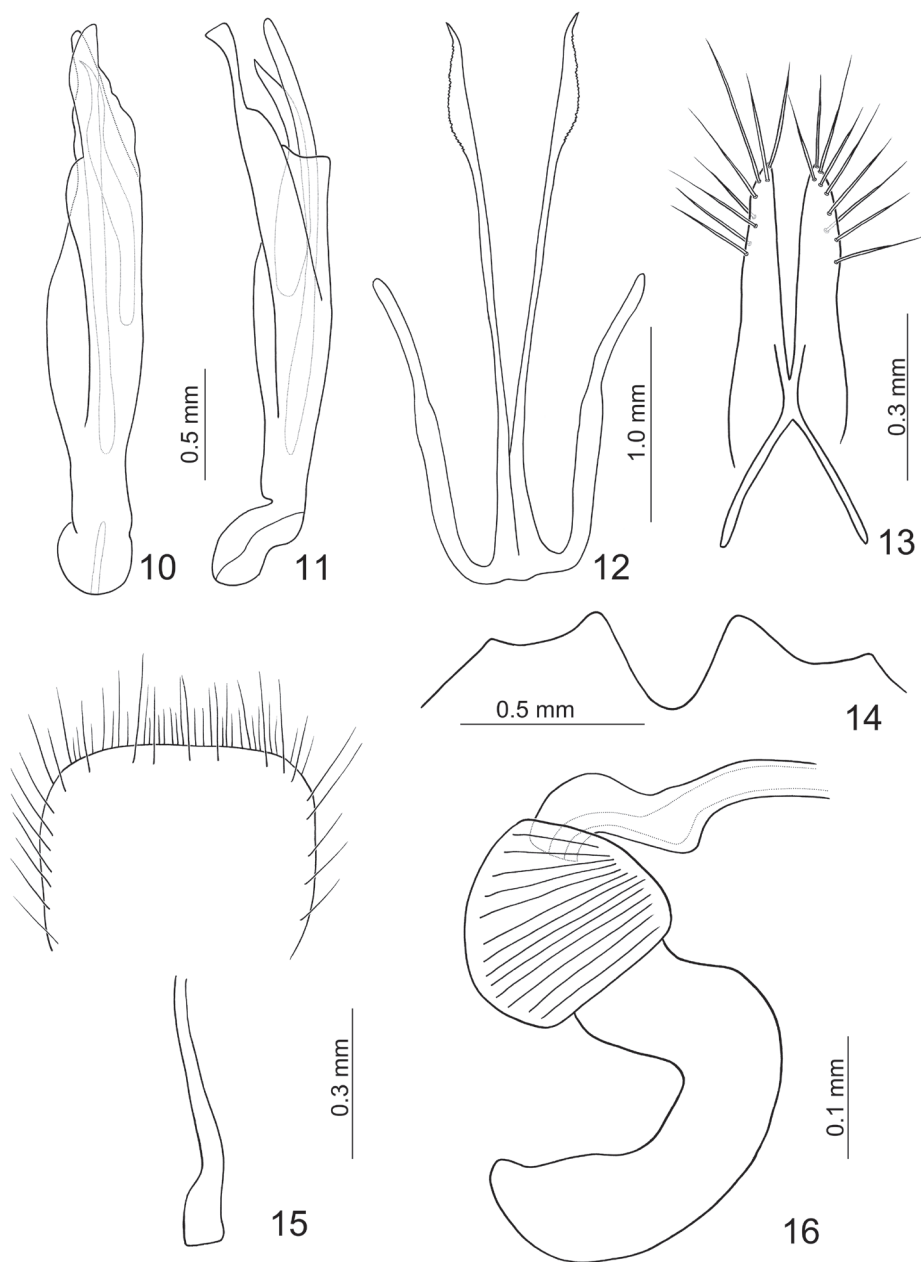
http://species-id.net/wiki/Paridea_kaoi

Paraulaca angulicollis: Chûjô 1938: 138 (misidentification)

Paraulaca (*Paraulaca*) *angulicollis*: Chûjô 1962: 195 (redescription); Chûjô 1963: 395.

Type locality. Taiwan, Taipei, Wulai, 24°51'N, 121°33'E, 150 m, broad-leaf forest.

Type material (n= 27). Holotype male (TARI), mounted on card, labeled: Taiwan: Taipei (130) [p] / Wulai [p] / 05.IX.2006, leg. H.-T. Cheng [p, w] // **Holotypus** [p] / *Paridea* (*Semacia*) ♂ [p] / *kaoi* n. sp. [p] / des. C.-F. Lee, 2014 [p, r]". Paratypes: 1♀: "Taihorinsho [h] / Formosa [p] / Sauter [p] IX [h] __ 07 09 [p, w] // *Paraulaca* [h] / *angulicollis* [h] / MOTSCHULSKY [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04198 [p, g]" (SDEI); 1♀: "LB [h] 973 [h, w, circular label] / Taiwan: Taipei (973) [p] / Wulai [p] / 15.III.2007, leg. H.-J. Chen [p,w]" (TARI); 3♂♂: "Taiwan: Taipei, [p] / Wulai, 30.III.2007 [p] / leg. C.-F. Lee [p, w]" (TARI); 1♀: "Taiwan: Taipei (10866) [p] / Wulai [p] / 22.V.2009, leg. H.-J. Chen [p,w]" (TARI); 1♀: "Taiwan: Taipei (11678) [p] / Wulai [p] / 26.VI.2009, leg. H.-J. Chen [p, w]" (TARI); 1♂: "TAIWAN: Wulai [p] / nr. Taipei, 300- [p] / 500m, 23.IX.1957



Figures 10–16. *Paridea (Semacia) houjayi* sp. n. **10** Penis, dorsal view **11** Penis, lateral view **12** Eighth abdominal tergite **13** Gonocoxae **14** Fifth abdominal ventrite **15** Eighth abdominal sternite **16** Spermatheca.

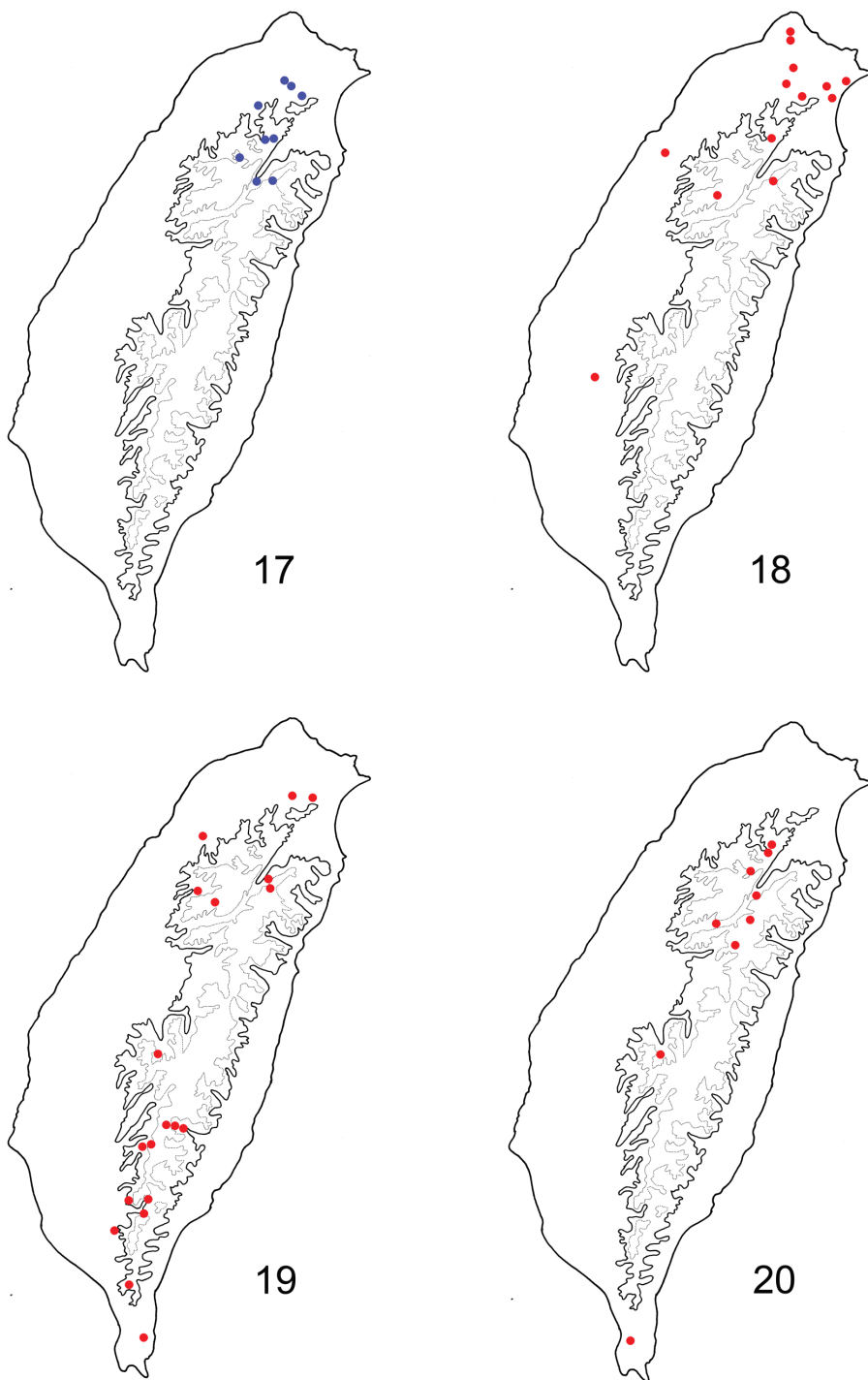
[p, w] // K.S. Lin [p] / Collector [p] / BISHOP“ (BPBM); 1♂: “TAIWAN (NE): [p] / Taiheizan [= Taipingshan], 1500m [p] / 6 [h]. VI/1934 [p] / L. Gressitt [p, w]“ (BPBM); 1♂: “Taiwan: Ilan (5258) [p] / Fushan Chiwuyuan [p] [= Botanical Park] /

08.V.2008, leg. M.-H. Tsou [p,w]" (TARI); 1♂: "Taiwan: Ilan [p] / Fushan (福山) 植物園 [p] / 13.IV.2011, leg. C.-F. Lee [p,w]" (TARI); 1♂: "Taiwan: Ilan (4711) [p] / Mingchi (明池) [p] / 27.IV.2008, leg. M.-H. Tsou [p, w]" (TARI); 1♀: "Taiwan: Ilan [p] / Yingtzuling (鶯仔嶺) [p] / 15.IV.2012, leg. Y.-L. Lin [p,w]" (TARI); 1♂: "Taiwan: Miaoli (4776) [p] / Hohsinglinchang (和興林場) [p] / 01.V.2008, leg. Y.-C. Lin [p,w]" (TARI); 1♂: "Taiwan: Taichung (3193) [p] / Anmashan (鞍馬山) [p] / 22.IX.2007, leg. M.-H. Tsou [p,w]" (TARI); 1♂: "TAIWAN: [p] / Kwantzing, [p] / Tainan Hsien, 250m [p] / 6–7.IV.1965 [p, w] // C. Yoshimoto [p, w]" (BPBM); 2♂♂, 2♀♀: "Taiwan: Taipei (5934–5937) [p] / Chutzuhu (竹子湖) [p] / 15.VI.2008, leg. S.-F. Yu [p, w]" (TARI); 1♂: "Taiwan: Taipei (5661) [p] / Maokung (貓空) [p] / 29.V.2008, leg. S.-F. Yu [p, w]" (TARI); 1♂: "Taiwan: Taipei [p] / Pinglin (坪林) [p] / 28.VII.2007, leg. Y.-L. Lin [p, w]" (TARI); 1♀: "TAIWAN: [p] / Taipei & vicinity [p] / IX.1964 [p,w] // T. C. Maa [p] / Collector [p,w] // *Paridea* [h] / *angulicollis* [h] / 鑒定者: [p] Motschulsky [h, w]" (BPBM); 2♂♂, 2♀♀: "Taiwan: Taipei (916–919) [p] / Yangmingshan (陽明山) [p] / 10.III.2007, leg. M.-H. Tsou [p, w]" (TARI); 1♀: "TAIWAN, Ilan county, [p] / Mingchyh Forest [p] / Recreation Area, 1200 m, [p, w] // swept from vegetation, [p] / 5.IV.2002, [p] / leg. Gy. Fábíán & O. Merkl [p, w]" (HNHM); 1♂: "TAIWAN bor. or. 2.vi.2008 [p] / Yilan County, 50 m [p] / cca 20 km N of Yilan City [p] / N 24°49,25'; E 121°44,39' [p] / leg. F. & L. Kantner [p, w]" (FKCC). Each paratype has a type label: "**Paratypus** [p] / *Paridea* (*Semacia*) ♂ [or ♀] [p] / *kaoi* n. sp. [p] / des. C.-F. Lee, 2014 [p, pink label]"

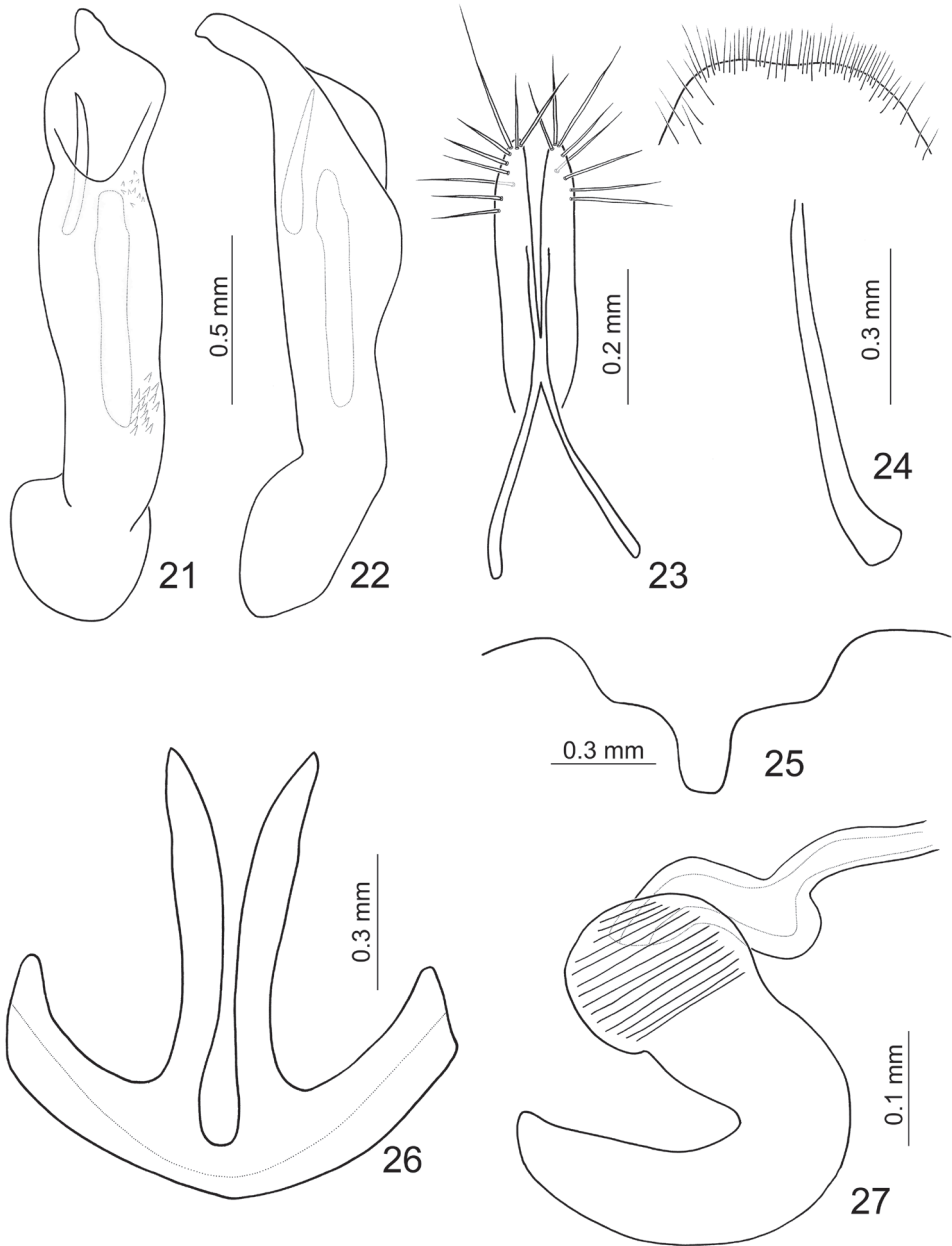
Diagnosis. This new species is similar to *P. angulicollis* but differs by the separation between basal black lateral margins and subapical spots on the elytra (Fig. 8) (connected in *P. angulicollis* (Fig. 98)), the wider apical processes of the eighth abdominal tergite in males (Fig. 26) (more slender in *P. angulicollis* (Fig. 107)), the wider penis in lateral view, and the deep notch on apical margin of the fifth abdominal ventrite in females (Fig. 25) (one pair of rounded processes in *P. angulicollis* (Fig. 106)).

Males. Length 5.0–5.3 mm, width 2.7–3.1 mm. Head and prothorax yellowish brown (Figs 6–8), labrum black, antenna brown; scutellum pale yellow; elytra pale yellow, postscutellar common area depressed; with one extremely slender black stripe along suture behind excavation, sometimes reduced; with one pair of large black spots subapically, lateral margin and epipleuron black, abbreviated at middle and separated from subapical black spots; meso- and metathoracic ventrites black; legs dark brown, apex of femur and base of tibia paler; abdomen yellow. Eighth abdominal tergite (Fig. 26) strongly sclerotized, transverse and slender, with one pair of slender and curved processes. Pygidium slightly projecting beyond elytral apices. Penis (Figs 21–22) strongly asymmetric, moderately narrowed at apical 1/6; apex narrow, tubular, and small; straight from lateral view; endophallic sclerites with one pointed sclerite, one elongate sclerite, an anterior cluster of small setae, and a posterior cluster of large setae.

Females. Length 5.7–6.1 mm, width 3.3–3.5 mm. Color (Fig. 9) similar to male; elytra without excavation but with black spot instead. Apical margin of last abdominal ventrite (Fig. 25) with shallow emargination at middle, margin of emargination truncate with deep notch at middle. Pygidium slightly projecting beyond elytral apices.



Figures 17–20. Distribution map of *Paridea* species, solid line: 1000 m, broken line: 2000 m. **17** *P. (S.) houjayi* sp. n. **18** *P. (S.) kaoi* sp. n. **19** *P. (S.) sexmaculata* **20** *P. (P.) costata*.



Figures 21–27. *Paridea* (*Semacia*) *kaoi* sp. n. **21** Penis, dorsal view **22** Penis, lateral view **23** Gonocoxae **24** Eighth abdominal sternite **25** Fifth abdominal ventrite **26** Eighth abdominal tergite **27** Spermatheca.

Gonocoxae (Fig. 23) slender, apex of each gonocoxa with eight or nine setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 24) weakly sclerotized; apex narrow, apical margin a little emarginate at middle,

surface with dense long setae along apical margin, spiculum short. Spermathecal receptaculum (Fig. 27) swollen; pump much longer than receptaculum, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Etymology. The name is dedicated to Mr. Shu-Jung Kao who financially supported the Taiwan Chrysomelid Research Team.

Distribution. Taiwan. This species occurs in central and northern Taiwan (Fig. 18).

Host plant. Cucurbitaceae: *Gynostemma pentaphyllum* (Thunb.) Makino.

Notes. This species was misidentified as *Paridea* (*Semacia*) *angulicollis* by Chûjô (1938, 1962, 1963).

***Paridea* (*Semacia*) *sexmaculata* (Laboissière, 1930)**

http://species-id.net/wiki/Paridea_sexmaculata

Semacia sexmaculata Laboissière, 1930: 336; Chûjô 1935: 168.

Paraulaca (*Paraulaca*) *semaculata*: Chûjô 1962: 194; Chûjô 1965: 97.

Paridea (*Paraulaca*) *sexmaculata*: Kimoto 1966: 30; Kimoto 1969: 33; Kimoto 1989: 250.

Paridea (*Semacia*) *sexmaculata*: Yang 1991: 268.

Paridea (*Paridea*) *sexmaculata*: Beenen 2010: 468.

Paraulaca taiwana Chûjô, 1935: 167 (part).

Type locality. Taiwan, Tainan.

Type material. Holotype female (ZMUH), pinned, labeled: “Tainan [h] / Formose [h, w] // TYPE [red letters, p] / ♀ [h, w] // *Semacia* [h] / *sexmaculata* [h] / m [h] / V. Laboissière – Dét. [p, w] // Le Moul’t Vend. [p] / via Reinbek [p] / Eing Nr. 1, 1957 [p, w]”. It was indicated as male in the original description but marked as female in the examined specimen.

Additional material examined (n= 36). **CHINA: Hunan:** 1♀, Yungshun, 6.VIII.1988, leg. S.-U. Wang (TARI); **TAIWAN: Hsinchu:** 1♂, Shinchiku (= Hsinchu), 1–30.VII.1918, leg. J. Sonan & K. Miyake (BMNH); 1♂, Wuchihshan, 27.III.2008, leg. S.-F. Yu (TARI); **Hualien:** 2♂♂, 1♀, YuShan N. P., 8.VI.2008, leg. F. & L. Kantner (FKCC, JBCB), 1♀, same locality, 7.VI.2008 (FKCC); **Ilan:** 1♂, Fushan Botanical Garden, 8–11.IV.2002, leg. O. Merkl (HNHM); 1♀, Taipingshan, 12.VI.2007, leg. Y.-C. Chang (TARI); 1♀, Tulishan, 10.III.2007, leg. H.-H. Li (TARI); **Kaoshiung:** 1♀, Erchituan, 1.V.2009, leg. U. Ong (TARI); 1♂, Tona logging trail, 3.II.2013, leg. B.-X. Guo (TARI); 1♂, Wukungshan, 23.I.2009, leg. M.-H. Tsou (TARI); **Nantou:** 1♀, Tungpu, 28.IV–2.V.1981, leg. T. Lin & C. J. Lee; 1♀, 19–23.VII.1982, leg. L. Y. Chou & T. Lin (TARI); **Pingtung:** 2♂♂, Machia, 11.III.2013, leg. Y.-T. Chung (TARI); 1♂, same locality, 17.III.2013, leg. W.-C. Liao (TARI); 1♀, Nanjenshan, 1.III.2010, M.-L. Jeng (TARI); 1♀, Peitawushan, 18.VI.2012, leg. J.-C. Chen (TARI); 1♂, Tahanshan, 18.VII.2007, leg. C.-F. Lee (TARI); 1♀, same locality, 27.VIII.2009, leg. J.-C. Chen (TARI); 1♀, same locality, 6.VI.2012, leg. C.-F. Lee (TARI); 1♂, same locality, 19.VII.2012, leg. C.-F. Lee (TARI); 1♂, same locality, 26.III.2013, leg. C.-

F. Lee (TARI); **Taichung**: 1♀, Anmashan, 7.VI.2010, leg. C.-F. Lee (TARI); 1♂, Wushihkeng, 19.III.2008, leg. C.-F. Lee (TARI); **Taipei**: 1♂, Fushan, 12.III.2009, leg. H.-J. Chen (TARI); 1♀, Tunghou, 27.VI.2007, leg. M.-H. Tsou (TARI); 1♀, Wulai, 12.III.2009, leg. S.-F. Yu (TARI); **Taitung**: 1♀, Hsiangyang, 14.VIII.2012, leg. C.-F. Lee (TARI); 1♂, Liyuan, 19.VI.2013, leg. C.-F. Lee; 1♀, Motien, 23.VI.2010, leg. S.-F. Yu (TARI); 1♂, same locality, 19.VI.2011, leg. C.-F. Lee (TARI).

Diagnosis. *Paridea* (*Semacia*) *sexmaculata* is similar to *P. (S.) kaoi* sp. n. and *P. (S.) angulicollis* with the depression on postscutellar common area of males but flat and replaced with a black spot in females. It differs by the presence of a black spot near the lateral margin instead of black stripe along lateral margin in *P. (S.) kaoi* sp. n. and *P. (S.) angulicollis*, the well sclerotized and extremely slender penis (Fig. 37) and processes on eighth abdominal tergite of males (Fig. 42), and the presence of two acute processes on the fifth abdominal ventrite in females (Fig. 41).

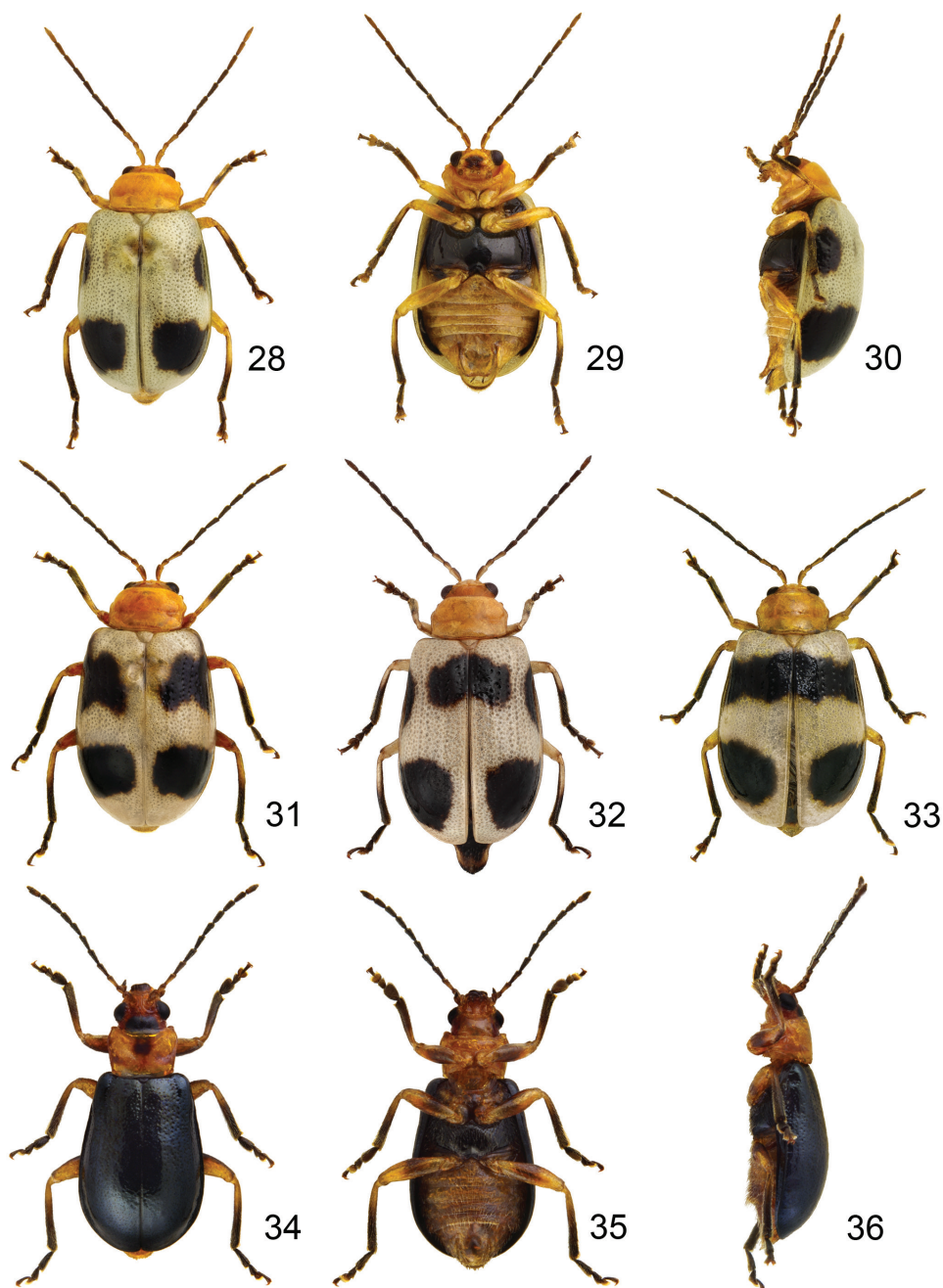
Males. Length 5.1–6.0 mm, width 2.9–3.3 mm. Head and prothorax yellowish brown (Figs 28–30), labrum black, antenna blackish brown; scutellum pale yellow; elytra pale yellow, postscutellar common area depressed; with one pair of black spots near lateral margin at same level with excavation, spots extending inwards in some individuals (Fig. 31; one pair of large black spots subapically, lateral margin and epipleuron yellow; meso- and metathoracic ventrites black; legs yellowish brown, apical half of tibia, and tarsi black; abdomen yellow. Eighth abdominal tergite (Fig. 42) strongly sclerotized, transverse and slender, with one pair of extremely slender and curved processes. Pygidium slightly projecting beyond elytral apices. Penis (Figs 37–38) very slightly asymmetric, weakly narrowed at apical 1/6; apex narrow, tubular, and extremely long; moderately curved from lateral view; endophallic sclerites with one elongate sclerite and one layer of small setae.

Females. Length 5.3–6.2 mm, width 3.1–3.4 mm. Similar to male; elytra without excavation but with transverse black spot instead (Fig. 32), sometimes connected with lateral black spots (Fig. 33). Apical margin of last abdominal ventrite (Fig. 41) with one pair of small, wide, and acute processes at middle, with shallow notch between processes. Pygidium slightly projecting beyond elytral apices. Gonocoxae (Fig. 39) slender, apex of each gonocoxa with eight setae from apical 1/4 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 40) weakly sclerotized; apex wide, apical margin slightly concave at middle, surface with dense long setae along apical margin, spiculum short. Spermathecal receptaculum (Fig. 43) slightly swollen; pump much longer than receptaculum, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

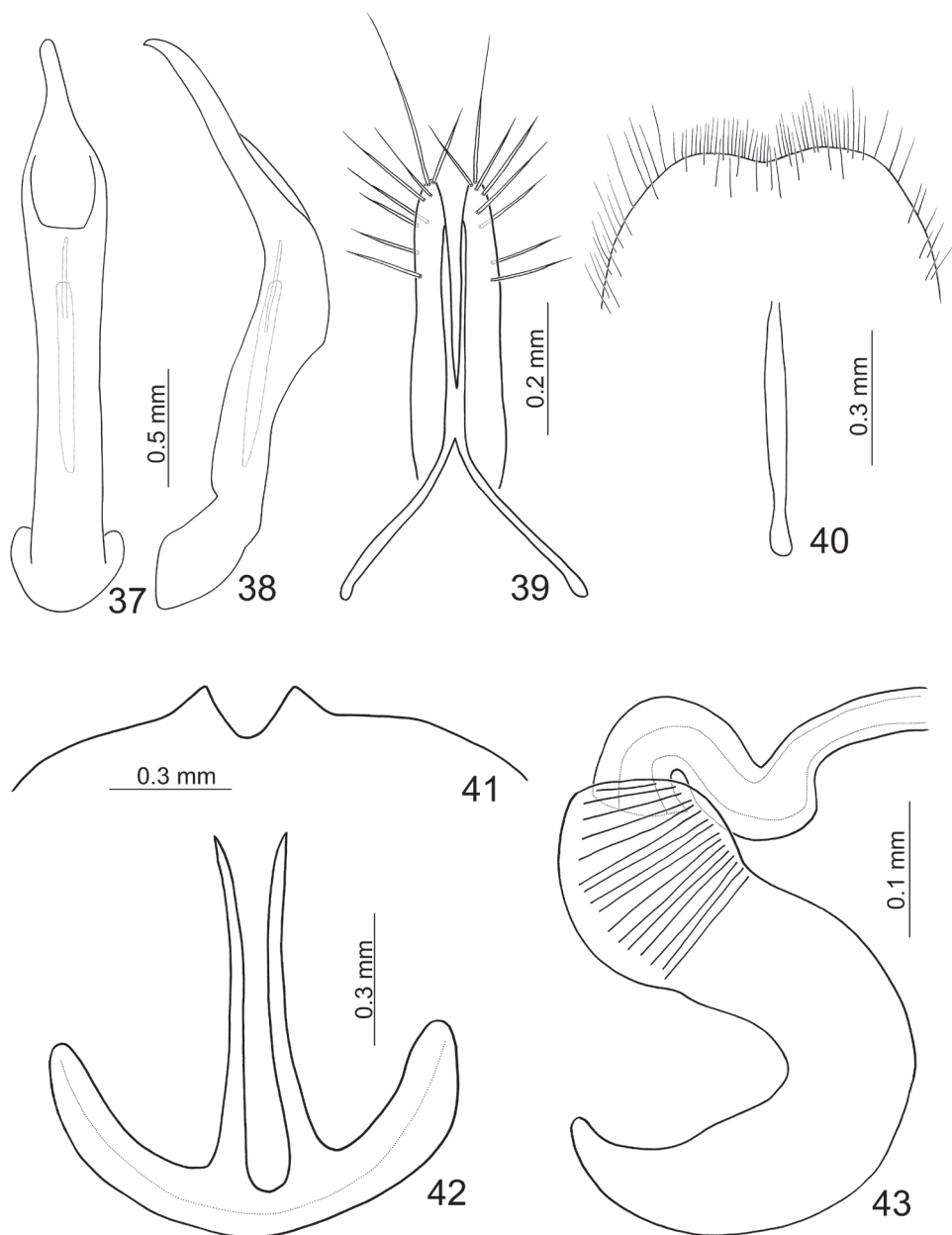
Distribution. Taiwan, China (Hunan). It is widespread in Taiwan (Fig. 19). Yang (1991) indicated that this species was also found in Hebei, Beijing, Jiangsu, Shanghai, Zhejiang, Fujian, and Hainan provinces of China without voucher specimens. These records require confirmation.

Host plant. Cucurbitaceae: *Gynostemma pentaphyllum* (Thunb.) Makino.

Notes. Two specimens of the type series of *Paraulaca taiwana* were misidentified. They are *P. (S.) sexmaculata* and labeled as follow: 1♂: “Shiigo [= Maopu, Wufeng



Figures 28–36. *Paridea* species. **28** *P. (S.) sexmaculata*, male, dorsal view **29** ditto, ventral view **30** ditto, lateral view **31** *P. (S.) sexmaculata*, male, color variation **32** *P. (S.) sexmaculata*, female, dorsal view **33** *P. (S.) sexmaculata*, female, color variation **34** *P. (P.) costata*, male, dorsal view **35** ditto, ventral view **36** ditto, ventral view.



Figures 37–43. *Paridea* (*Semacia*) *sexmaculata*. **37** Penis, dorsal view **38** Penis, lateral view **39** Gonocoxae **40** Eighth abdominal sternite **41** Fifth abdominal ventrite **42** Eighth abdominal tergite **43** Spermatheca.

township] Chikuto [= Chutung] [p] / SHINCHIKU [= Hsinchu county] [p] / 27–30. VI.1934 [p] / COL. M. CHUJO [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* Чûjô [h] / DET. M. CHUJO [p, g] // No. 1339 [p, w]” (TARI); 1♂: “Formosa [p] / Karenko, [= Hualien] – 19. [p] / VII 20-VIII 4. [p] / T. Okuni, [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* Чûjô [h] / DET. M. CHUJO [p, g] // 2185 [p, w]” (TARI).

***Paridea (Paridea) costata* (Chûjô, 1935)**

http://species-id.net/wiki/Paridea_costata

Paraulaca costata Chûjô, 1935: 164.

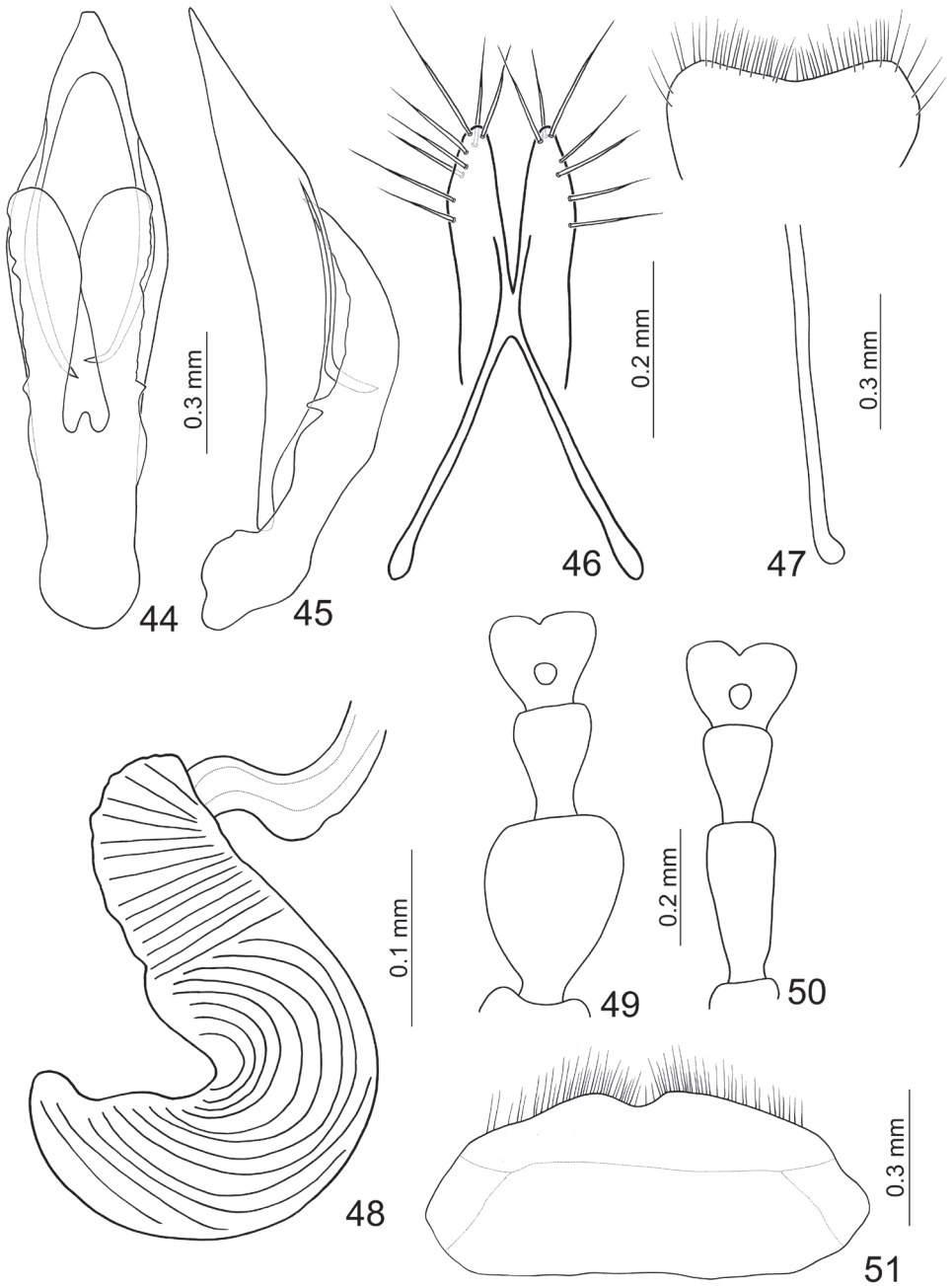
Paraulaca (Carapaula) costata: Chûjô 1962: 198 (redescription); Chûjô 1963: 395.

Paridea (Paridea) costata: Gressitt and Kimoto 1963: 512; Kimoto 1966: 29; Kimoto 1969: 33; Kimoto 1989a: 251; Kimoto 1991: 11; Takizawa et al. 1995: 12.

Type locality. Taiwan, Hualien.

Type material. Lectotype male (TARI), pinned, here designated to fix the concept of *Paraulaca costata* Chûjô and to ensure the universal and consistent interpretation of the same, labeled: “Formosa [p] / Karenko [= Hualien], -19 [p] / VII 20-VIII 4. T. Okuni, [p, w] / COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *costata* [h] / Чûjô [h] / DET. M. CHUJO [p, g] // 1491 [p, w] // **Lectotypus** [p] / *Paraulaca costata* ♂ [p] / Chûjô, 1935 [p] / des. C.-F. Lee, 2014 [p, r]”. Paralectotypes: 1♂ (TARI): “Formosa [p] / Y. Miwa [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *costata* [h] / Чûjô [h] / DET. M. CHUJO [p, g] // 2585 [p, w]”; 1♀ (TARI): “14/IV 1918 [h] / Bakuras [= Bakulasi, in Nantou] [h] / Col. I. Nitobe [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *costata* [h] / Чûjô [h] / DET. M. CHUJO [p, g] // 1490 [p, w]”; 1♀ (TARI): “Horisha [= Puli, in Nantou] [h] / Apr 2. 1919 [h, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *costata* [h] / Чûjô [h] / DET. M. CHUJO [p, g] // 2586 [p, w]”; 2♂♂ (SDEI): “Taihorin [= Talin, in Chiayi] [p] / Formosa [p] / H. Sauter, 1911 [p, w] // 7.VIII. or 7.VII. [p, w] // Syntypus [p, r] // *Paraulaca* [h] / *costata* [h] / Чûjô [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04199 and 04200 [p, g]”. Each paralectotype has a type label: “**Paralectotypus** [p] / *Paraulaca costata* ♂ [or ♀] [p] / Chûjô, 1935 [p] / des. C.-F. Lee, 2014 [p, pink label]”

Additional material examined (n= 29). **TAIWAN:** **Hualien:** 1♀, Taroko N. P., 3.vi.2008, F. & L. Kantner leg. (JBCB); **Hsinchu:** 1♀, Mamei, 4.V.2008, leg. S.-F. Yu (TARI); **Ilan:** 1♀, Suchi, 19.V.2010, leg. H.-J. Chen (TARI); **Nantou:** 3♂♂, 1♀, Sungkang, 2.VII.2008, leg. M.-H. Tsou (TARI); 2♂♂, 1♀, Tungpu, 28.IV.-2.V.1981, leg. T. Lin & C. J. Lee (TARI); 1♂, Wushe, 30.VIII–2.IX.1982, leg. L. Y. Chou & K. C. Chou (TARI); 1♂, same locality, 19–22.IV.1983, leg. K. C. Chou & S. P. Huang (TARI); **Pingtung:** 1♀, Suchunghsi, 8.V.2013, leg. Y.-T. Chung; **Tai-chung:** 2♂♂, 1♀, Kukuan, 16.VII.2007, leg. M.-H. Tsou (TARI); 1♀, Wuling, 30.



Figures 44–51. *Paridea* (*Paridea*) *costata*. **44** Penis, dorsal view **45** Penis, lateral view **46** Gonocoxae **47** Eighth abdominal sternite **48** Spermatheca **49** Tarsi of front leg, male **50** Tarsi of front leg, female **51** Eighth abdominal tergite.

VI.2008, leg. S.-F. Yu (TARI); **Taoyuan**: 1♀, Hsuanyuan, 16.III.2008, leg. S.-F. Yu (TARI); 6♂♂, 1♀, Tamanshan, 2.VIII.2008, leg. S.-F. Yu (TARI); 1♂, 5♀♀, same locality, 2.VIII.2008, leg. M.-H. Tsou (TARI).

Diagnosis. *Paridea* (*Paridea*) *costata* is recognized by the black vertex, presence of one black spot on the pronotum, the metallic blue elytra, and the swollen first tarsomeres of front and middle legs of males.

Males. Length 4.5–4.7 mm, width 2.2–2.4 mm. General color (Figs 34–36) yellowish brown; antenna blackish brown but three basal antennomeres paler; vertex with one big black spot; pronotum with a moderate black spot near center; scutellum and elytra bluish black; metathoracic ventrites black; tibia and tarsi dark brown; femora darkened except bases and apices. Elytra with one longitudinal costa arising from humerus, reduced in some individuals. Median lobe at fifth abdominal ventrite deeply depressed. Eighth abdominal tergite (Fig. 51) weakly sclerotized but apex well sclerotized, transverse and wide, apical margin emarginate at middle, with dense long setae along apical margin. First tarsomeres of pro- and mesotarsi swollen (Fig. 49). Penis (Figs 44–45) wide, apically tapering, apex truncate, slightly curved near based from lateral view; with one pair of elongate processes extending from near apex to middle, base curved upwards from lateral view; external process large and wide, lateral margin irregular from middle to apex, with a small process at middle of lateral margin; endophallus without visible sclerites.

Females. Length 4.7–4.9 mm, width 2.5 mm. Similar to male; apical margin of last abdominal ventrite smooth, not modified. Gonocoxae (Fig. 46) slender, apex of each gonocoxa with seven or eight setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 47) weakly sclerotized; apex wide, apical margin emarginate at middle, surface with dense long setae on apical margin, spiculum short. Spermatheca (Fig. 48) strongly sclerotized, disk with dense transverse grooves; receptaculum narrower than pump; pump short and wide, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Distribution. China (Gansu, Jiangxi, Sichuan, Zhejiang), Taiwan. It is uncommon but widespread in Taiwan (Fig. 20).

Host plants. Cucurbitaceae: *Thladiantha nudiflora* Hemsl. ex Forbes & Hemsl.

Paridea (*Paridea*) *cyanipennis* (Chûjô, 1935)

http://species-id.net/wiki/Paridea_cyanipennis

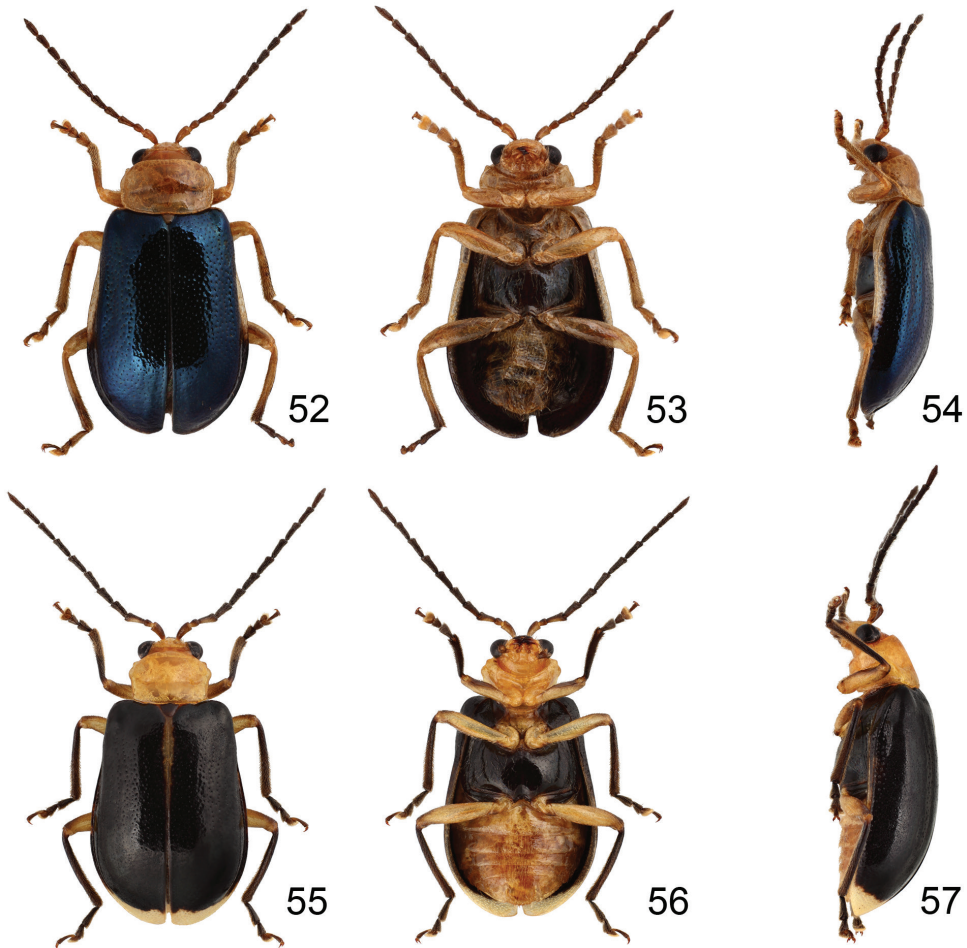
Paraulaca cyanipennis Chûjô, 1935: 164

Paraulaca (*Paraulaca*) *cyanipennis*: Chûjô 1962: 192 (redescription);

Paridea (*Paridea*) *cyanipennis*: Kimoto 1966: 30; Kimoto 1989a: 250; Kimoto 1991: 11.

Type locality. Taiwan, Pingtung, Henchun (= Koshun), 22°00'N, 120°44'E, 50 m, broad-leaf forest.

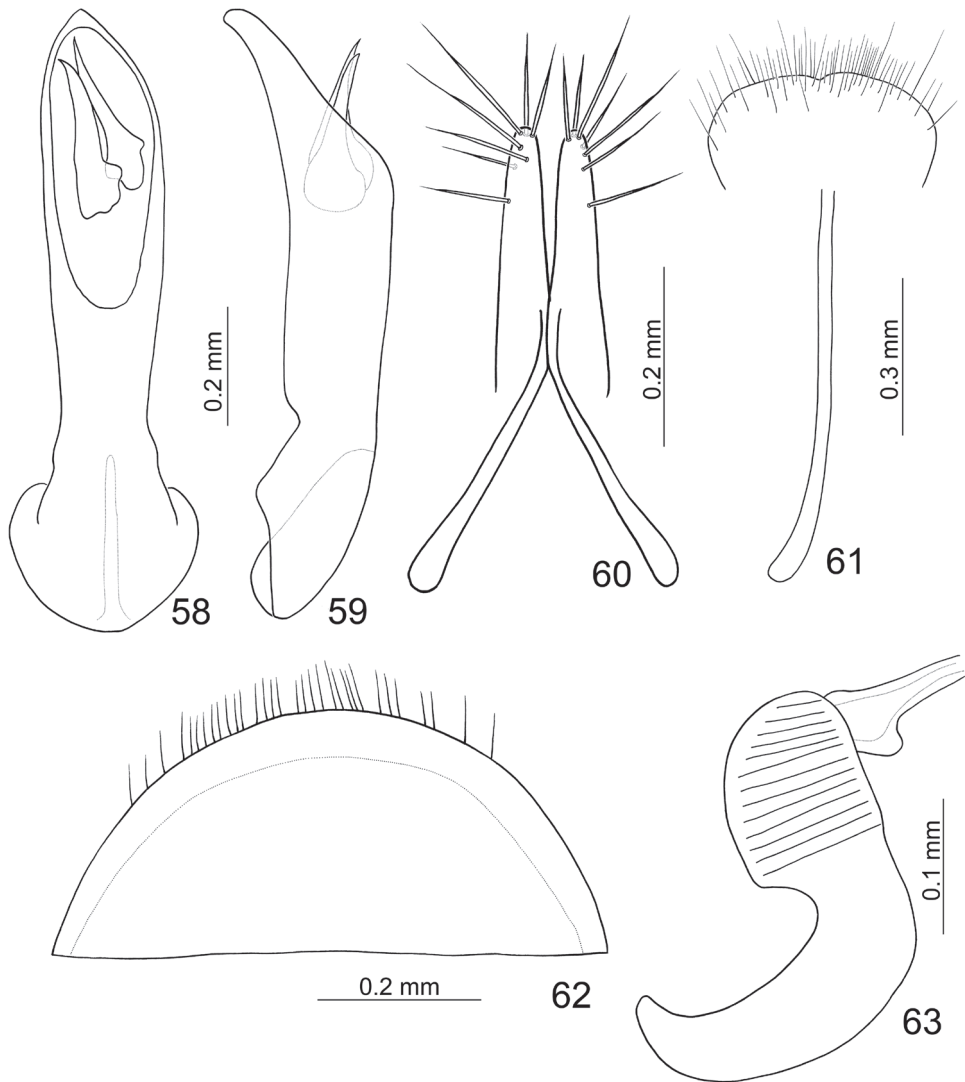
Type material. Holotype male (SDEI), mounted on card, labeled: “ Kankau (Koshun) [p] / Formosa [p] / H. Sauter VI. 1912 [p, w] // Holotype [h, red letters]



Figures 52–57. *Paridea* species. **52** *P. (P.) cyanipennis*, male, dorsal view **53** ditto, ventral view **54** ditto, ventral view **55** *P. (P.) sauteri*, male, dorsal view **56** ditto, ventral view **57** ditto, ventral view.

// *Paraulaca* [h] / *cyanipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04201 [p, g]”. Paratypes: 1♂ (SDEI), same as holotype but with “Paratype” and “Col. – 04204”; 1♂ (TARI), labeled: “ Kankau (Koshun) [p] / Formosa [p] / H. Sauter VII. 1912 [p, w] // Paratype [h, red letters] // *Paraulaca* [h] / *cyanipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // 2848 [p, w]”; 1♂, 2♀♀ (SDEI), labeled: “ Kankau (Koshun) [p] / Formosa [p] / H. Sauter IX. 1912 [p, w] // Paratype [red letters, h] // *Paraulaca* [h] / *cyanipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04202, 04205–6 [p, g]”; 1♂ (TARI), same as preceding but with “2847 [p, w]”; 1♀ (SDEI), same as preceding but with “Allotype” and “DEI Müncheberg [p] / Col – 04203 [p, g]”.

Additional material examined (n= 16). **TAIWAN:** **Hualien:** 2♀♀, Wenlan, 21.X-9.XII.2009, leg. W. T. Yang & K. W Huang (NMNS); **Kaoshiung:** 1♂, Chi-



Figures 58–63. *Paridea (Paridea) cyanipennis*. **58** Penis, dorsal view **59** Penis, lateral view **60** Gonocoxae **61** Eighth abdominal sternite **62** Eighth abdominal tergite **63** Spermatheca.

asien, 10–13.V.1981, leg. C. C. Chen & C. C. Pan (TARI); 1♂, 1♀, Meinung, 17.VII.2012, leg. J.-C. Chen (TARI); 3♂♂, 2♀♀, Tengchih, 26.IV.2010, leg. J.-C. Chen (TARI); **Pingtung**: 1♂, Checheng, 1.XII.2012, leg. J.-C. Cheng (TARI); 2♂♂, 2♀♀, Kueishan, 3.VIII.2012, leg. J.-C. Chen (TARI); 1♀, Lilungshan, 6.VI.2013, leg. J.-C. Chen (TARI).

Diagnosis. This species is similar to *Paridea (Paridea) costata* with the metallic blue elytra but differs in possessing a yellowish brown lateral margin of each elytron, the yellowish brown head and pronotum, and the unmodified first tarsomeres of males.

Males. Length 4.9 mm, width 2.4 mm. General color yellowish brown (Figs 52–54); antennomeres IV–XI darkened; elytra metallic blue, but basal margin, base to apical 1/3 of lateral margin yellowish brown; metathoracic ventrites metallic blue. Elytra with one longitudinal costa arising from humerus, reduced in some individuals. Eighth abdominal tergite (Fig. 62) weakly sclerotized but apex well sclerotized, transverse and wide, apical margin rounded, with dense long seta along apical margin. Penis (Figs 58–59) wide, apically tapering, apex slightly asymmetric and pointed, slightly curved near apex and apex narrowly rounded from lateral view; endophallus with two visible sclerites, apically curved and pointed, one smaller.

Females. Length 5.5–6.1 mm, width 3.0–3.2 mm. Similar to male; apical margin of last abdominal ventrite smooth, not modified. Gonocoxae (Fig. 60) slender, apex of each gonocoxa with seven setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 61) weakly sclerotized; apex wide, apical margin emarginate at middle, surface with longer setae near apical margin and shorter setae on apical margin, spiculum long. Spermatheca (Fig. 63) weakly sclerotized; receptaculum slightly swollen; pump short and wide, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Distribution. The species occurs in eastern and southern Taiwan (Fig. 64).

Host plant. Cucurbitaceae: *Momordica cochinchinensis* (Lour.) Spreng.

***Paridea (Paridea) sauteri* (Chûjô, 1935), stat. n.**

Paraulaca sauteri Chûjô, 1935: 166; Kimoto 1974: 26 (as synonym of *P. sinensis*)

Paraulaca (Paraulaca) sauteri: Chûjô 1962: 192 (redescription); Chûjô 1963: 395.

Paridea (Paraulaca) sauteri: Kimoto 1965a: 489.

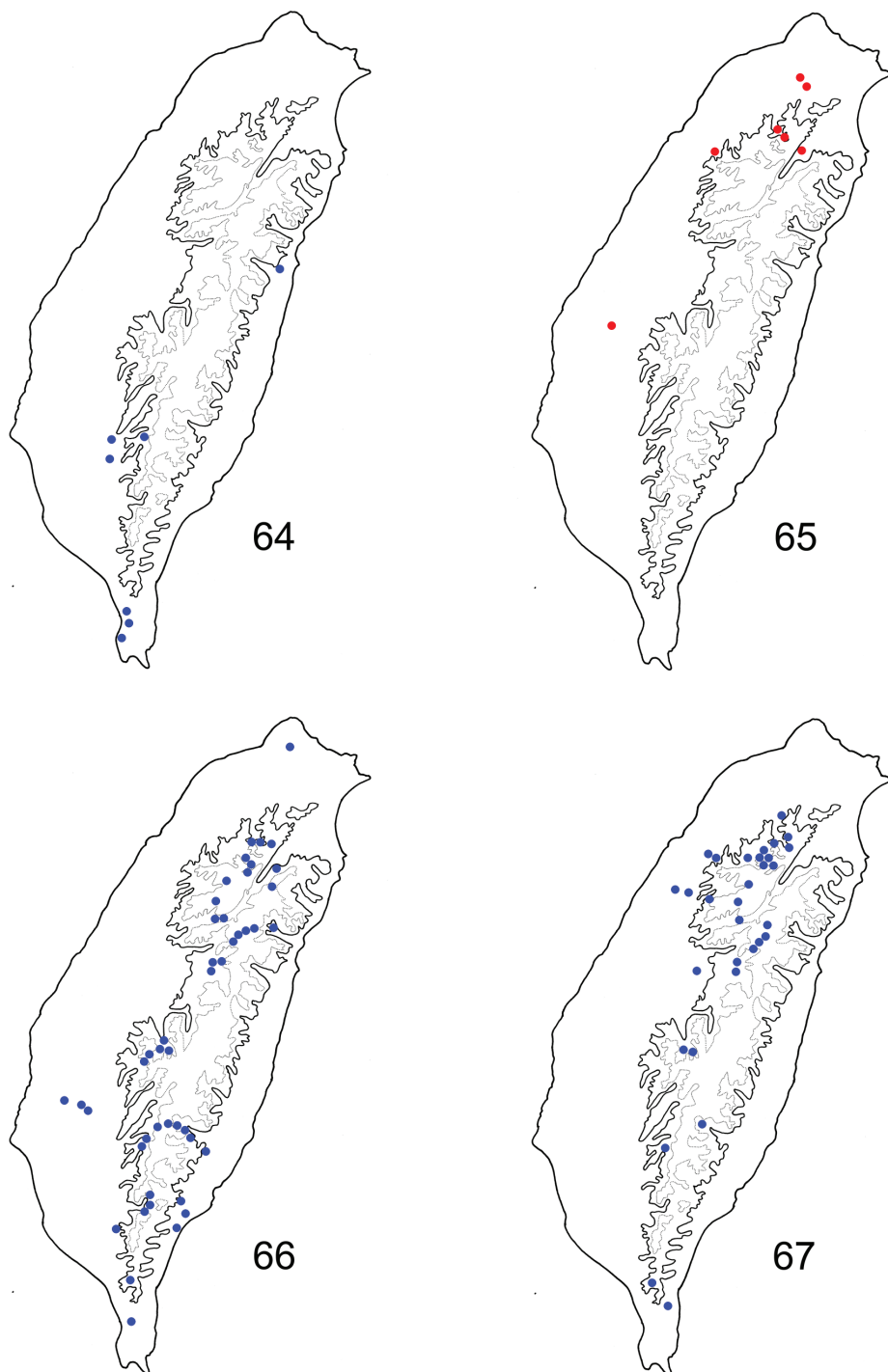
Paridea (Paridea) sauteri: Kimoto 1966: 29; Kimoto 1969: 35; Takizawa et al. 1995: 12.

Paridea (Paridea) sinensis: Kimoto 1986: 57; Kimoto 1989a: 251; Kimoto 1991: 11.

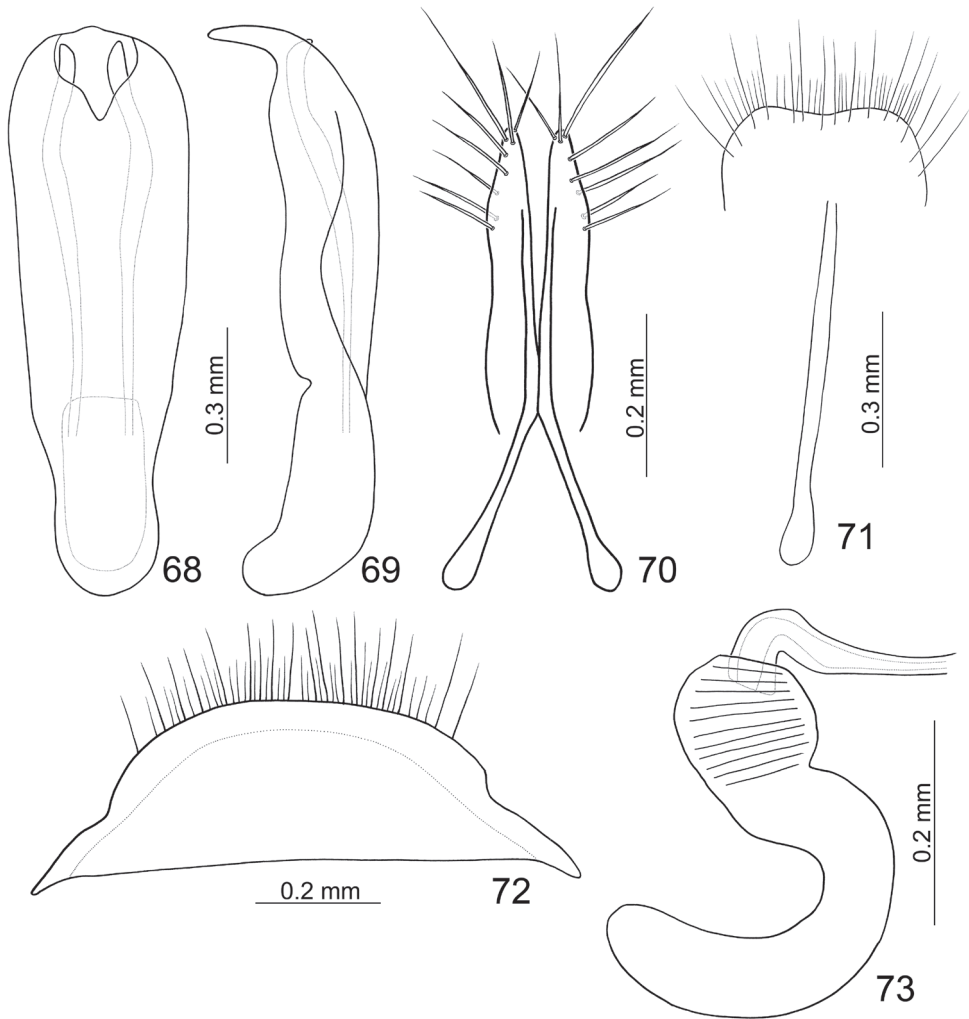
Type locality. Taiwan, Chiayi, Talin (= Taihorin), 23°35'N, 120°28'E, 50 m, broad-leaf forest.

Type material. Holotype ♂ (SDEI), labeled: “Taihorin [p] / Formosa [p] / H. Sauter, 1911 [p] // 7. VIII. [p] // Holotype [h, red letters] // *Paraulaca* [h] / *sauteri* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04207 [p, g]”. Paratypes: 1 ♀ (SDEI), same as holotype but with “Allotype” and “Col – 04210”; 2 ♀♀ (SDEI), labeled: Taihorin [p] / Formosa [p] / H. Sauter, 1911 [p] // 7. VII. [p] // Paratype [h, red letters] // *Paraulaca* [h] / *sauteri* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04208 & 04209 [p, g]”; 1 ♀ (TARI), same as preceding, but with “2846 [p, w]”; 1 ♂ (TARI), labeled: “Taihorin [h] / Formosa [p] 10. [h] / Sauter [p] IV. [h] ___ 07 09 // Paratype [h, red letters] // *Paraulaca* [h] / *sauteri* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // 2845 [p, w]”.

Additional material examined (n= 51). TAIWAN: Hsinchu: 13♂♂, 3♀♀, Wufeng, 14–16.VII.1982, leg. K. C. Chou & C. C. Pan (TARI); Ilan: 2♂♂, Songluo,



Figures 64–67. Distribution map of *Paridea* species, solid line: 1000 m, broken line: 2000 m. **64** *P. (P.) cyanipennis* **65** *P. (P.) sauteri* **66** *P. (P.) taiwana* **67** *P. (P.) testacea*.



Figures 68–73. *Paridea (Paridea) sauteri*. **68** Penis, dorsal view **69** Penis, lateral view **70** Gonocoxae **71** Eighth abdominal sternite **72** Eighth abdominal tergite **73** Spermatheca.

7.IV.2007, leg. M.-H. Tsou (TARI); **Taipei:** 2♂♂, Fushan, 5.IV.2007, leg. S.-F. Yu (TARI); 3♂♂, same locality, 2.III.2012, leg. M.-H. Tsou (TARI); 1♂, Shintien, 2.X.2010, leg. Y.-F. Hsu (TARI); 1♂, Wulai, 7.X.2006, leg. S.-F. Yu (TARI); 1♂, same locality, 30.III.2007, leg. C.-F. Lee (TARI); 1♀, same locality, 16.V.2007, leg. G. Martin & J. Quicke (BMNH); 2♂♂, same locality, 15.VI.2007, leg. C.-F. Lee (TARI); 8♂♂, same locality, 26.VI.2009, leg. C.-F. Lee (TARI); 2♂♂, same locality, 26.VII.2009, leg. H.-J. Chen (TARI); 7♂♂, same locality, 30.VIII.2009, leg. C.-F. Lee (TARI); 1♂, same locality, 1.III.2010, leg. C.-F. Lee (TARI); **Taoyuan:** 1♂, Hsiaowulai, 19.IV.2008, leg. S.-F. Yu (TARI); 1♂, Paling, 2.IX.2009, leg. H. Lee (TARI).

Diagnosis. *Paridea* (*Paridea*) *sauteri* is similar to some individuals of *P. (P.) taiwana* having yellow apices of the black elytra but differs by possessing yellow femora and black tibiae (black outer margins of femora and tibiae in *P. (P.) taiwana*).

Males. Length 4.8–5.5 mm, width 2.9–3.2 mm. General color (Figs 55–57) yellowish brown; antenna dark brown; apex of labrum darkened; elytra black but apex pale yellow; scutellum, meso- and metachoracic ventrites black; tibiae and tarsi blackish brown. Eighth abdominal tergite (Fig 72) weakly sclerotized, transverse and wide, apical margin truncate, with dense long seta along apical margin. Penis (Figs 68–69) wide, apex pointed, ventrally curved; almost straight from lateral view; endophallic sclerites composed of one pair of elongate and longitudinal sclerites, curved inwards near apex, apex projecting from opening.

Females. Length 5.5–5.7 mm, width 3.2–3.3 mm. Similar to male; apical margin of last abdominal ventrite smooth, not modified. Gonocoxae (Fig. 70) slender, apex of each gonocoxa with eight setae from apical 1/4 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 71) weakly sclerotized; apex wide, apical margin slightly emarginate at middle, surface with longer setae near apical margin, and shorter and denser setae on apical margin, spiculum long. Spermathecal receptaculum (Fig. 73) slightly swollen; pump extremely long, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Distribution. Endemic to Taiwan, and with a scattered distribution. The species seems to be allopatric with *P. (P.) taiwana* (Fig. 65). For example, it is common in Wulai, northern Taiwan. But no individuals of *P. (P.) taiwana* have been found in Wulai.

Host plants. Cucurbitaceae: *Thladiantha nudiflora* Hemsl. ex Forbes & Hemsl.

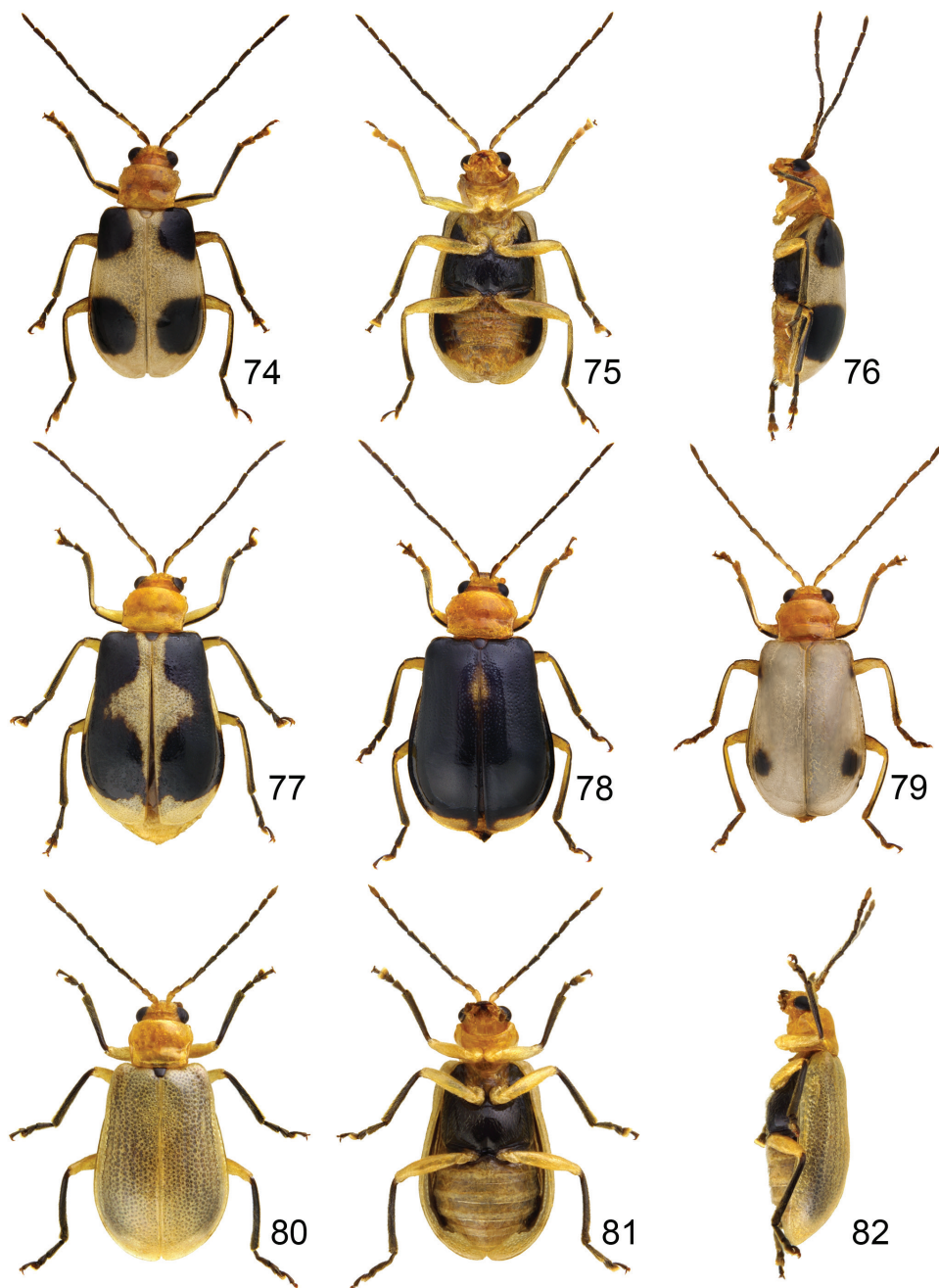
Paridea (*Paridea*) *taiwana* (Chûjô, 1935), stat. r.

Paraulaca taiwana Chûjô, 1935: 167; Kimoto 1966: 29 (as synonym of *P. sauteri*).

Paraulaca (*Carapaula*) *taiwana*: Chûjô 1962: 200 (redescription); Chûjô 1965: 98.

Type locality. Taiwan, Hualien.

Type material. Lectotype male (TARI), pinned, here designated to fix the concept of *Paraulaca taiwana* Chûjô and to ensure the universal and consistent interpretation of the same, labeled: “Formosa [p] / Karenko [= Hualien], -- 9. [p] / VII 20-VIII 4. [p] / T. Okuni, [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJÔ [h] / DET. M. CHUJO [p, g] // 2186 [p, w] // **Lectotypus** [p] / *Paraulaca taiwana* ♂ [p] / Chûjô, 1935 [p] / des. C.-F. Lee, 2014 [p, r]”. Paralectotypes: 3♀♀ (TARI), same as lectotype but with “756, 2188, and 2189”; 1♂ (TARI): “Arisan [= Alishan], [h] / 1912-X-10 [h] / Col. I. Nitobe [p, w] // *Paridea* n sp. [h] / Det. Shiraki [p, w] // *Aulacophora* ? [h] / *quadriplagiata* Baly [h] / Det. Shiraki [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJÔ [h] / DET. M. CHUJO [p, g] // 755 [p, w]”; 1♀ (TARI): “Arisan [= Alishan] [p] /



Figures 74–82. *Paridea* species. **74** *P. (P.) taiwana*, male, dorsal view **75** ditto, ventral view **76** ditto, lateral view **77** *P. (P.) taiwana*, black spots enlarged **78** *P. (P.) taiwana*, black elytra with yellow apices **79** *P. (P.) taiwana*, black spots reduced **80** *P. (P.) testacea*, female, dorsal view **81** ditto, ventral view **82** ditto, ventral view.

Formosa [p] / 25.X.1933 [p] / Col. M. CHUJO [p] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJô [h] / DET. M. CHUJO [p, g] // No. 1338 [p, w]”; 1♂ (TARI): “Rakuraku [in Hualien] [p] / 18.IV.1924 [p] / T. Shiraki [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJô [h] / DET. M. CHUJO [p, g] // 2191 [p, w]”; 1♀ (TARI): “Funkiko [= Fenchihu, in Chiayi] [h] / 16.II.1926 [h] / Col. J. Sonan [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJô [h] / DET. M. CHUJO [p, g] // 2190 [p, w]”; 1♀ (TARI): “Raisha [= Laiyi, in Pingtung] [p] / 30-VIII-1927 [p] / J. Sonan [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *taiwana* CHÛJô [h] / DET. M. CHUJO [p, g] // 2187 [p, w]”; 1♀ (SDEI): “Suisharyo [= Shuisheliao, in Chiayi] [p] / Formosa [p] / H. Sauter X.11 [p, w] // Syntypus [p, r] // *Paraulaca* [h] / *taiwana* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04211 [p, g]”; 1♂ (SDEI): “Taihorinsho [= Talin, in Chiayi] [p] / Formosa [p] / H. Sauter [p] X 09 [h, w] // Syntypus [p, r] // *Paraulaca* [h] / *taiwana* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04212 [p, g]”. Each paralectotype has a type label: “**Paralectotypus** [p] / *Paraulaca taiwana* ♂ [or ♀] [p] / Chûjô, 1935 [p] / des. C.-F. Lee, 2014 [p, pink label]”

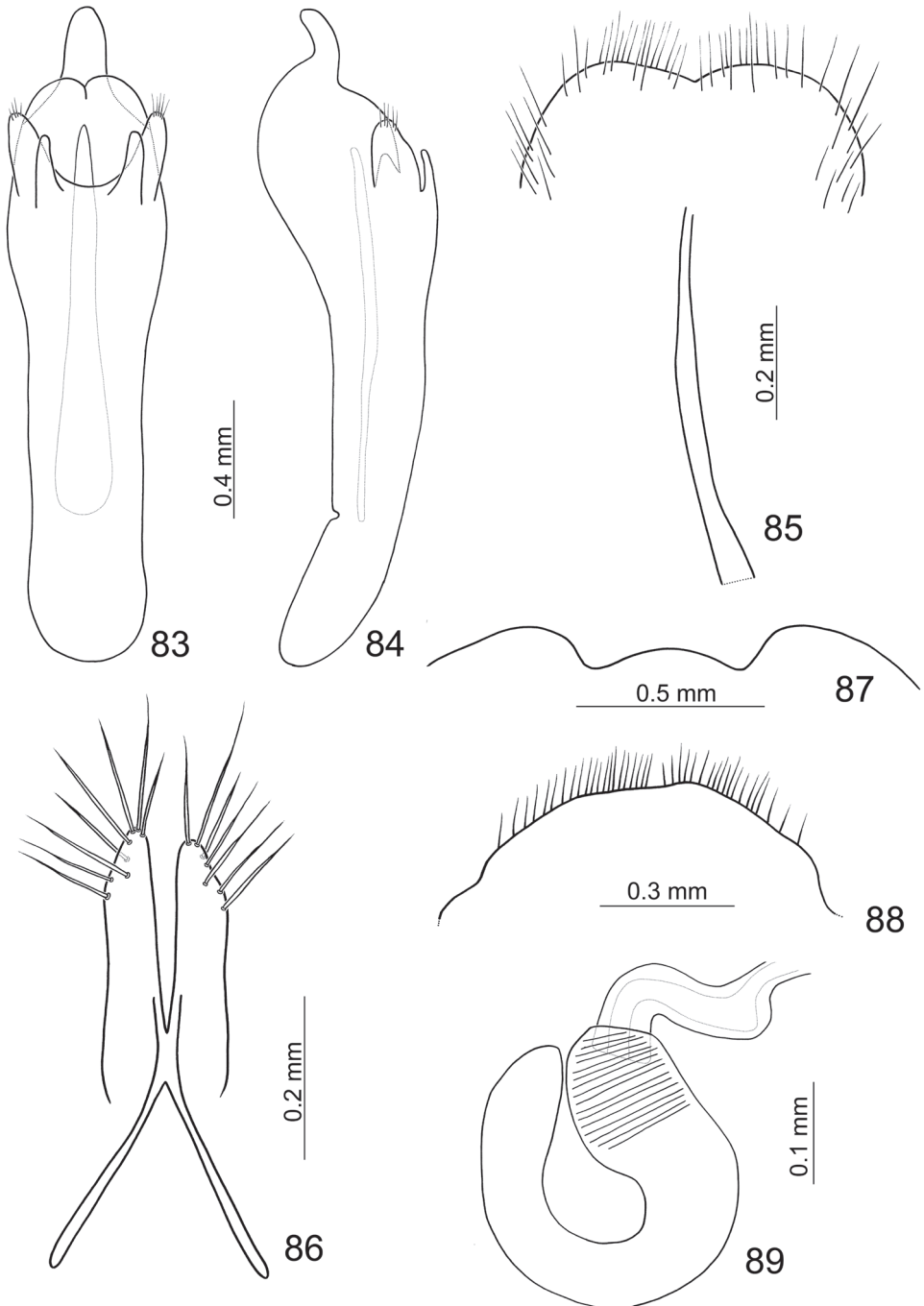
Additional material examined (n= 503). TAIWAN: Chiayi: 1♀, Arisan (= Alishan), 2–23.X.1918, leg. J. Sonan & M. Yoshino (BMHH); 2♀♀, Alishan, 5–9. VIII.1981, leg. L. Y. Chou and S. C. Lin; 1♀, same locality, 27.VI.2010, leg. U. Ong; 2♂♂, Laichitashan, 19.III.2009, leg. H. Lee; **Hsinchu:** 1♀, Litungshan, 3.V.2008, leg. Y.-L. Lin; 1♂, Mamei, 4.V.2008, leg. H.-F. Yu; 1♀, same locality, 18.V.2008, leg. M.-H. Tsou; 1♂, Shihlei, 6.III.2010, leg. Y.-L. Lin; 1♂, Talu logging trail, 29. IV.2008, leg. Y.-L. Lin; 1♀, same locality, 19.VI.2010, leg. Y.-L. Lin; 1♂, same locality, 17.III.2012, leg. Y.-L. Lin; **Hualien:** 1♂, Huitouwan, 10.VII.2007, leg. C.-F. Lee; 1♀, Tayuling, 9–16.VI.1980, leg. K. S. Lin and B. H. Chen; **Ilan:** 1♂, Mingchi, 20.IV.2007, leg. H.-H. Li; 1♀, Taipingshan, 12.VI.2007, leg. Y.-C. Chang; 1♂, Tuchang, 1.III.2007, leg. H.-H. Li; **Kaoshiung:** 1♂, Chungchihkuang, 16.IV.2012, leg. L.-P. Hsu; 2♂♂, 1♀, 10–13.X.2012, leg. L.-P. Hsu; 15♂♂, Erchituan, 1.III.2009, leg. U. Ong; 2♂♂, 1♀, same locality, 1.V.2009, leg. U. Ong; 3♂♂, 1♀, same locality, 21.VI.2010, leg. U. Ong; 1♂, same locality, 8.III.2013, leg. B.-X. Guo; 1♂, Southern C.-I. Hwy, 19.IX.2008, leg. L. Dembick (BMNH); 1♂, Shihshan logging trail, 23. III.2009, leg. H. Lee; 4♀♀, Tengchih, 2–5.VI.2008, leg. C.-F. Lee; 2♂♂, same locality, 12.III.2013, leg. Y.-T. Chung; 1♂, same locality, 19.III.2013, leg. Y.-T. Chung; 3♂♂, same locality, 31.III.2013, leg. W.-C. Liao; 2♂♂, same locality, 8.VI.2013, leg. W.-C. Liao; 1♀, same locality, 15.VI.2013, leg. B.-X. Guo; 2♂♂, same locality, 6. VII.2013, leg. W.-C. Liao; 2♂♂, 10.VIII.2013, leg. W.-C. Liao; 1♂, 1♀, Tona logging trail, 25.II.2013, leg. Y.-T. Chung; **Nantou:** 2♀♀, Hoshe, 22.VII.1982, leg. L. Y. Chou and T. Lin; 1♀, Hsitou, 12.IX.2009, leg. C.-F. Lee; 1♂, same locality, 10.V.2010, leg. Y.-T. Wang; 1♀, Lushan, 28.VI.1981, leg. K. S. Lin and W. S. Tang; 1♀, same locality, 6.VIII.2008, leg. H. Medel & M. V. L. Barclay (BMNH); 1♀, same locality, 8.VIII.2008, leg. H. Medel, U. Ong, M. V. L. Barclay & R. Ewers (BMNH); 1♀, Meifeng, 10.V.1979, leg. K. C. Chou; 3♂♂, 3♀♀, same locality, 20–

22.VI.1979, leg. K. S. Lin and B. H. Chen; 2♀♀, same locality, 29.VIII-10.IX.1979; 3♂♂, same locality, 2-12.X.1979; 1♂, 1♀, same locality, 24.X.1979, leg. K. C. Chou; 1♂, 1♀, same locality, 25.X-7.XI.1979; 1♀, same locality, 2-4.VI.1980, leg. L. Y. Chou & C. C. Chen; 3♀♀, same locality, 8.VI.1980, leg. K. S. Lin and B. H. Chen; 1♂, 3♀♀, same locality, 26.VIII.1980, leg. K. S. Lin and C. H. Wang; 1♂, 4♀♀, same locality, 5-9.X.1980, leg. C. C. Chen and C. C. Chien; 4♂♂, 7♀♀, same locality, 7-9.V.1981, leg. K. S. Lin and S. C. Lin; 2♀♀, same locality, 24-26.VI.1981, leg. K. S. Lin and W. S. Tang; 1♂, same locality, 28-29.VIII.1981, leg. L. Y. Chou and S. C. Lin; 1♀, same locality, 22.V.1982, leg. L. Y. Chou; 1♀, same locality, 15.VII.1982, leg. S. C. Lin and C. N. Lin; 1♂, same locality, 4-7.X.1982, leg. K. C. Chou; 1♂, 1♀, same locality, 8-11.V.1984, leg. K. C. Chou and C. C. Pan; 1♀, Mong Gwu, 14 km E of Puli, 20.IV.2002, leg. D. Anstine, G. Fabián & O. Merkl (JBCB); 1♀, Sungkang, 13-15.IX.1984, leg. K. S. Lin and S. C. Lin; 1♀, same locality, 3.VII.2008, leg. M.-H. Tsou; 1♀, Tsuifeng, 8.V.1981, leg. K. S. Lin and S. C. Lin; 1♀, same locality, 12-14.IX.1984, leg. K. S. Lin and S. C. Lin; 1♂, Tungpu, 20-22.VI.1980, leg. C. C. Chen; 6♂♂, 8♀♀, same locality, 25-29.IX.1980, leg. L. Y. Chou and T. Lin; 8♂♂, 9♀♀, same locality, 28.IV-2.V.1981, leg. T. Lin and C. J. Lee; 9♂♂, 6♀♀, same locality, 5-8.X.1981, leg. T. Lin and W. S. Tang; 9♂♂, 11♀♀, same locality, 18-23.XI.1981, leg. T. Lin and W. S. Tang; 9♂♂, 21♀♀, same locality, 19-23.VII.1982, leg. L. Y. Chou and T. Lin; 2♂♂, 4♀♀, same locality, 22-25.XI.1982, leg. K. C. Chou and S. P. Huang; 5♂♂, 6♀♀, same locality, 20-24.VI.1983, leg. K. C. Chou and C. Y. Wong; 16♂♂, 28♀♀, same locality, 23-27.VII.1984, leg. K. C. Chou and C. H. Yang; 2♂♂, same locality, XI.1985, leg. K. S. Lin; 1♂, 1♀, Salih sien-shi near Tungpu, 23.XI.2002, leg. L. Ronkay & O. Merkl (JBCB, HNHM); 2♂♂, 2♀♀, Wanfengtsun, 2.IV.2008, leg. W.-T. Liu; 1♀, same locality, 24.IV.2008, leg. W.-T. Liu; 1♂, same locality, 8.VII.2008, leg. W.-T. Liu; 2♂♂, Wushe, 19-23.VI.1979, leg. K. S. Lin and B. H. Chen; 1♀, same locality, 20-22.VI.1980, leg. C. C. Chen; 2♀♀, same locality, 8.X.1980, leg. C. C. Chen and C. C. Chien; 2♀♀, same locality, 6-11.V.1981, leg. K. S. Lin and S. C. Lin; 1♀, same locality, 23-28.VI.1981, leg. K. S. Lin and W. S. Tang; 1♀, same locality, 4.VIII.1981, leg. T. Lin and W. S. Tang; 1♂, same locality, 14.VII.1982, leg. S. C. Lin and C. N. Lin; 10♂♂, 4♀♀, same locality, 30.VIII-2.IX.1982, leg. L. Y. Chou and K. C. Chou; 4♂♂, 1♀, same locality, 7-8.X.1982, leg. K. C. Chou; 3♂♂, 3♀♀, same locality, 19-22.IV.1983, leg. K. C. Chou and S. P. Huang; 2♂♂, 2♀♀, same locality, 7.V.1984, leg. K. C. Chou and C. C. Pan; 2♂♂, 2♀♀, same locality, 17.VIII.1984, leg. K. S. Lin; 1♂, same locality, 11-15.IX.1984, leg. K. S. Lin; 6♂♂, 1♀, same locality, 23.III.2009, leg. U. Ong; 1♂, same locality, 21.VI.2009, leg. U. Ong; 1♂, 11♀♀, Kao-Leng Dyi, 18 km W of Wushe, 18-19.IV.2002, leg. D. Anstine, G. Fabián & O. Merkl (HNHM, 1♀ JBCB); **Pingtung**: 1♀, Lilungshan, 5.XI.2009, leg. M.-H. Tsou; 4♂♂, Machia, 11.III.2013, leg. Y.-T. Chung; 1♂, Peitawushan, 25.VI.2012, leg. J.-C. Chen; 2♂♂, 1♀, Tahanshan, 24.VI.2007, leg. C.-F. Lee; 4♂♂, 4♀♀, same locality, 18-20.VII.2007, leg. C.-F. Lee and M.-H. Tsou; 1♀, same locality, 25.V.2008, leg. C.-F. Lee; 1♂, 1♀, same locality, 4.VII.2008, leg. M.-H. Tsou; 2♀♀, 21.III.2009, leg. M.-H. Tsou; 1♀,

same locality, 5.IV.2009, leg. C.-F. Lee; 1♀, same locality, 18.V.2009, leg. M.-L. Jeng; 1♂, same locality, 28.VI.2009, leg. Y.-T. Chung; 1♂, same locality, 7.IX.2009, leg. U. Ong; 1♀, same locality, 3.XI.2009, leg. M.-H. Tsou; 1♂, 11.I.2010, leg. J.-C. Chen; 1♂, same locality, 28.VIII.2010, leg. Y.-L. Lin; 1♀, same locality, 5.VII.2011, leg. M.-H. Tsou; 1♂, same locality, 14.VIII.2011, leg. Y.-T. Wang; 2♂♂, same locality, 6.I.2012, leg. Y.-T. Chung; 1♀, same locality, 3.VI.2012, leg. W.-C. Liao; 1♀, same locality, 30.VII.2012, leg. Y.-T. Chung; 3♂♂, same locality, 20.X.2012, leg. W.-C. Liao; 1♀, same locality, 10.XI.2012, leg. W.-C. Liao; 1♂, same locality, 15.XII.2012, leg. W.-C. Liao; 1♂, same locality, 14.I.2013, leg. Y.-T. Chung; 3♂♂, same locality, 26.II.2013, leg. Y.-T. Chung; 1♀, same locality, 14.III.2013, leg. Y.-T. Chung; 8♂♂, 2♀♀, same locality, 26.III.2013, leg. C.-F. Lee; 1♂, 2♀♀, same locality, 3.IV.2013, leg. Y.-T. Chung; 2♀♀, same locality, 25.V.2013, leg. Y.-T. Chung; 1♀, same locality, 2.VII.2013, leg. Y.-T. Chung; 1♀, same locality, 3.VII.2013, leg. B.-X. Guo; 2♂♂, 1♀, same locality, 1.X.2013, leg. Y.-T. Chung; 6♂♂, 5♀♀, Wutai, 12.IV.2009, leg. U. Ong; 2♀♀, same locality 9–12.V.1009, leg. U. Ong; 1♂, same locality, 17.V.2009, leg. U. Ong; 1♂, same locality, 1.IV.2010, leg. U. Ong; **Tai-chung**: 2♂♂, Anmashan, 16.VII.2007, leg. M.-H. Tsou; 1♀, same locality, 22.IX.2007, leg. M.-H. Tsou; 2♀♀, same locality, 7.VI.2010, leg. C.-F. Lee; 2♂♂, Chiapaotai, 14–18.X.1980, leg. K. S. Lin and C. H. Wang; 2♀♀, Kukuan, 14–17.X.1980, leg. K. S. Lin and C. H. Wang; **Tainan**: 2♂♂, 1♀, Kantoushan, 14.III.2010, leg. M.-H. Tsou; 1♀, Meiling, 28.XII.2008, leg. U. Ong; 2♂♂, 5♀♀, same locality, 24.III.2011, leg. U. Ong; 1♀, same locality, 24.IV.2013, leg. B.-X. Guo; 1♂, same locality, 7.VI.2013, leg. Y.-T. Chung; 1♂, Pichien, 11.II.2009, leg. U. Ong; **Taipei**, 1♀, Chutzuhu, 26.V.1983, leg. K. C. Chou; 1♂, same locality, 29.IV.2007, leg. M.-H. Tsou; **Taitung**: 1♀, Chihpen, 17–18.II.1982, leg. L. Y. Chou and K. C. Chou; 2♂♂, 1♀, Guanshan, 31.X.2009, leg. P.-F. Wang; 7♂♂, Hsiangyang, 14.VIII.2012, leg. C.-F. Lee; 1♂, Lichia, 2.VI.2009, leg. U. Ong; 1♀, Litao, 23.VI.2010, leg. M.-H. Tsou; 1♀, same locality, 4.X.2010, leg. M.-H. Tsou; 1♀, Liyuan, 19.VI.2013, leg. C.-F. Lee; 5♀♀, Motien, 23–24.VI.2010, leg. M.-H. Tsou; 12♂♂, same locality, 5.X.2010, leg. C.-F. Lee; 1♂, same locality, 19.VI.2011, leg. C.-F. Lee; 4♂♂, Taimali, 20.III.2008, P.-F. Wang; 1♂, Wulu logging trail, 26.IX.2007, leg. J.-F. Tsai; **Taoyuan**: 1♂, Paling, 3–5.V.1983, leg. K. C. Chou and C. C. Pan; 2♂♂, 2♀♀, Sankuang, 17.X.2009, leg. Y.-L. Lin. If not otherwise stated, all specimens deposited in TARI.

Diagnosis. This species is similar to *Paridea* (*Paridea*) *sinensis* in general color pattern but differs in possessing extremely variable black spots on the elytra, the smaller external process on the penis and the abruptly widened apex of the penis in lateral view (the external processes are extremely long and the penis is slender in lateral view in *P. (P.) sinensis*) and the modified apical margin of the fifth abdominal ventrite of females (Fig. 87).

Males. Length 5.1–5.9 mm, width 2.8–3.2 mm. General color (Figs 74–76) yellowish brown, two pairs of black spots on elytra, anterior one near humerus and other subapical, sizes of spots variable, sometimes spots enlarged and connected with each



Figures 83–89. *Paridea* (*Paridea*) *taiwana*. **83** Penis, dorsal view **84** Penis, lateral view **85** Eighth abdominal sternite **86** Gonocoxae **87** Fifth abdominal ventrite **88** Eighth abdominal tergite **89** Spermatheca.

other (Fig. 77), even whole elytra black except apices (Fig. 78), sometimes black spots reduced (Fig. 79); outer margins of femora and tibiae black; metathoracic ventrites black. Eighth abdominal tergite (Fig. 88) weakly sclerotized, transverse and wide, apical margin slightly emarginate at middle, with dense long seta along apical margin. Penis (Figs 83–84) wide; apex tubular, curved from lateral view; strongly widened near apex from lateral view; external process small, smaller than medial process; medial process wide apex with several setae.

Females. Length 4.8–7.1 mm, width 2.7–3.7 mm. Similar to male; apical margin of last abdominal ventrite emarginate at middle, slightly convex at emargination. Gonocoxae (Fig. 86) slender, apex of each gonocoxa with seven or eight setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 89) weakly sclerotized; apex wide, apical margin concave at middle, surface with dense long setae along apical margin, spiculum short. Spermathecal receptaculum (Fig. 15) slightly swollen; pump extremely long, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Distribution. Endemic to Taiwan. It is the most common and widespread species of the genus in Taiwan (Fig. 66).

Host plant. Cucurbitaceae: *Thladiantha nudiflora* Hemsl. ex Forbes & Hemsl.

Paridea (Paridea) testacea Gressitt & Kimoto, 1963

http://species-id.net/wiki/Paridea_testacea

Paridea (Paridea) testacea Gressitt & Kimoto, 1963: 515; Kimoto 1969: 33; Kimoto 1989a: 251; Kimoto 1991: 11.

Paraulaca flavipennis Chûjô, 1935: 165 (nec Laboissière 1930: 334); Chûjô 1938: 138;

Kimoto 1966: 29 (as synonym of *P. testacea*). **synonym confirmed**

Paraulaca (Paraulaca) flavipennis: Chûjô 1962: 193 (redescription); Chûjô 1965: 97.

Paridea (Paridea) formosana Yang, 1991: 272. (replacement name for *P. flavipennis*)

Type locality. China, Fujian, Shaowu, Tachulan.

Type material examined. *Paridea (Paridea) testacea*: Holotype female (BPBM), pinned, labeled: “Fukien, S. China [p] / Shawu, TaChu[p]Lan [h] / III-29-42 [h] T. C. Maa [p, w] // HOLOTYPE [p] ♂ [h] / *Paridea* [h] / (*Paridea*) [h] / *testacea* [h] / Gressitt & Kimoto [p, r] // *Paridea* (*P.*) [h] / *testacea* [h] / sp. ♂ [h] / holo [h] / Det. S. Kimoto [p, w]”. Gressitt and Kimoto (1963) indicated that the holotype is a male. Actually it is a female. Paratypes: 1♀: “Fukien, S. China [p] / Shaowu, Tachulan [p] / IV.11.[h]194[p]3[h] T. Maa [p, w] // T. C. Maa, collec- [p] / tor. L. Gressitt [p] / collection [p, w] // allo [h] // *Paridea* [h] / *testacea* [h] / ♀ G & K [h] / Gressitt & Kimoto det. 196[p]1[h, w] // ALLOTYPE [p] / *Paridea* [h] / *testacea* [h] / S. Kimoto [h] / J. L. Gressitt [p, r]; 1♀ (BPBM): “Fukien S. China [p] / ShaoWu TaChuFunG [h] / IV-24-[h],19[p]42[h] T.C.Maa [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt &

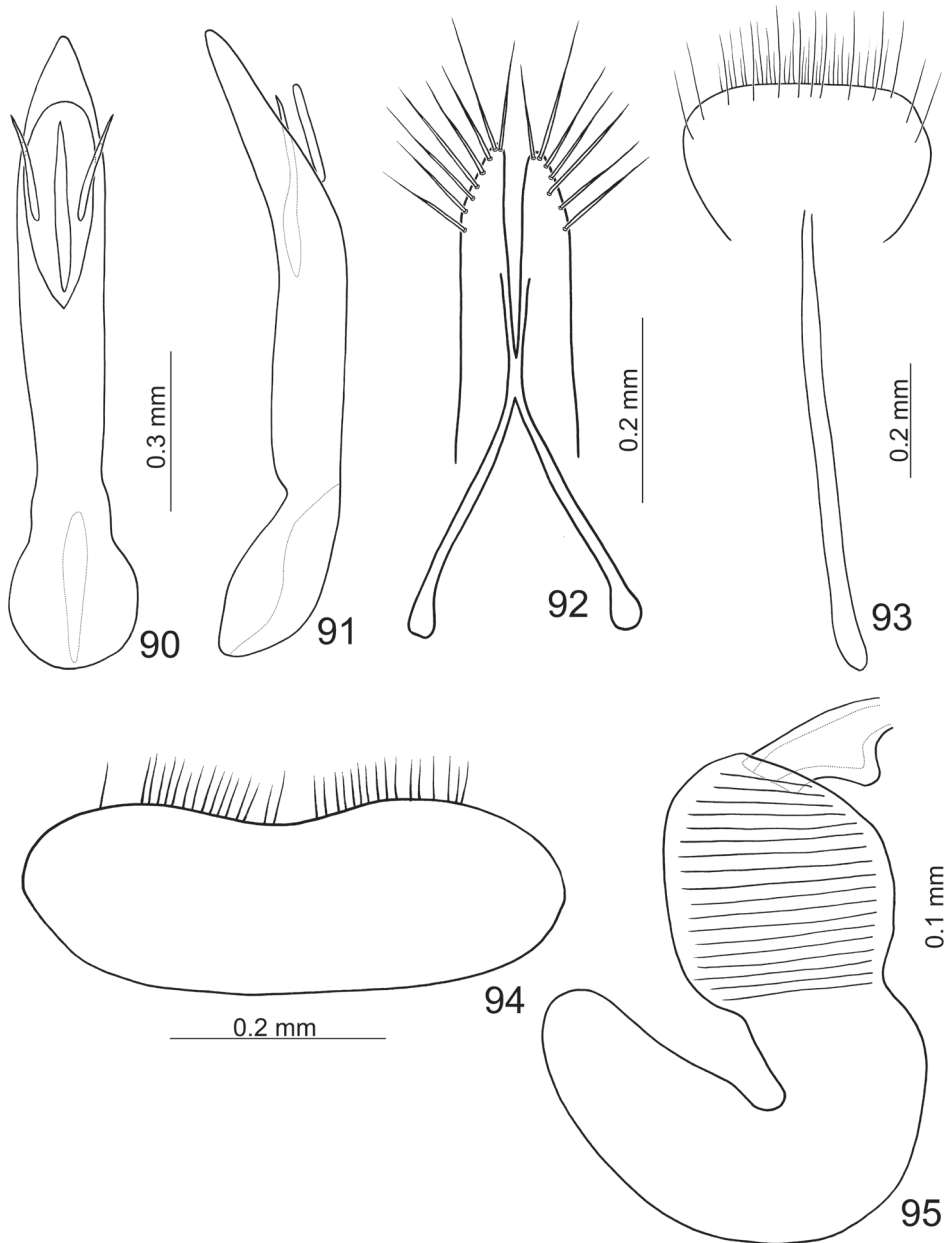
Kimoto det. 1961 [p, w]”; 1♀ (BPBM): “Fukien. S. China [p] / Shaowu, Tachulan [p] V.31[h].194[p]2[h] T. Maa [p, w] // T.C.Maa, collec- [p] tor. L.Gressitt [p] / collection [p] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 1961 [p, w]”; 1♀ (CAS): “FUKIEN, S. China [p] / Shaowu: Tachulan [p] / 1000 m. T. Maa [p, w] // Apr. 8, 1943 [h, w] // NO 6 [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 1961 [p, w]”; 1♀ (CAS): “FUKIEN, S. China [p] / Shaowu: Tachulan [p] / 1000 m. T. Maa [p, w] // Apr. 17, 1943 [h, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 1961 [p, w]”; 1♀ (BPBM): “FUKIEN, S. China [p] / Shaowu: Tachulan [p] / 1000 m. T. Maa [p, w] // APR. 27, 1943 [h, w] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 196 [p] 1 [h, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y]”; 1♀ (CAS): “FUKIEN, S. China [p] / Shaowu: Tachulan [p] / 1000 m. T. Maa [p, w] // V-3-1943 [h, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y]”; 1♀ (CAS): “Fukien, S. China [p] / Shaowu, TaChuFung May. 6–10. 1943 [p] T. C. Maa [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 1961 [p, w]”, although the locality of this type is different locality from that of holotype, it is still regarded as paratypes since collecting date was included in the original description and bearing the same type label; 1♀ (BPBM), same data as preceding; 1♀ (BPBM): “Fukien, S. China [p] / Shaowu, TaChuFung [p] / V [h] / 26-29[h]-43. T. C. Maa [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y]”; 1♀ [BPBM]: “FUKIEN, S. China [p] / Shaowu: Ta-ChuLan [p] / K.S. Lin [h] T. C. Maa [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / Gressitt & Kimoto det. 1961 [p, w]”; 1♀ (BPBM): “Fukien. S. China [p] / Shaowu, Tachulan [p] / VI.2[h].194[p]3[h] T. Maa [p, w] // T.C. Maa, collec- [p] / tor. L. Gressitt [p] / collection [p, w] // PARATYPE [p] / *Paridea* [h] / *testacea* [h] / Gressitt & Kimoto [p, y] // *Paridea* [h] / *testacea* [h] / G & K [h] / J.L.Gressitt det. [p, w]”.

Paraulaca flavipennis: Lectotype male (TARI), pinned, here designated to fix the concept of *Paraulaca flavipennis* Chûjô and to ensure the universal and consistent interpretation of the same, labeled: “Formosa [p] / Arisan [= Alishan, in Chiayi], 1918. [p] / X 2-23. [p] / J. Sonan. [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // 2177 [p, w] // **Lectotypus** [p] / *Paraulaca flavipennis* ♂ [p] / Chûjô, 1935 [p] / des. C.-F. Lee, 2014 [p, r]”. Paralectotypes: 1♀, same as lectotype but with “2178”; 1♂ (TARI): “Arisan [=Alishan, in Chiayi] [h] / 1912.X.10 [h] / Col. I. Nitobe [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // one kept [h] / (not paratype) [h, w]”; 1♂ (TARI): “Arisan [= Alishan, in Chiayi] [p] / FORMOSA [p] / 25.X.1933 [p] / Col. M. CHUJO [p] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJÔ [h] / DET. M. CHUJO [p, g] // No. 1345 [p, w]”; 1♂, 1♀ [TARI]: “Formosa [p] / Karenko [= Hualien], -19. [p] / VII 20-VIII 4. [p] / T. Okuni, [p] // COTYPE [p, circle label

with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2180 and 2181 [p, w]”; 1♀ (TARI): “Formosa [p] / Musha [= Wushe, in Nantou]. 1919 [p] / V 18 – VI 15 [p] / T. Okuni, [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2179 [p, w]”; 1♂, 1♀ (TARI): “Formosa [p] / Shinchiku [= Hsinchu], -18. [p, w] / VII 1-30. [p] / J. Sonan, [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2175 and 2176 [p, w]”; 1♂ (TARI): “Formosa [p] / Y. Miwa [p] // 西村 [= Hsitsun, in Maoli] [h] / 24.7.1929 [h, w] [on the back] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô (h) / DET. M. CHUJO [p, g] // 757 [p, w]”; 1♀ (TARI): “Formosa [p] / Y. Miwa [p] // Hsuangyung (in Taoyuan) [written in Japanese] [h] / 23.7.1929 [h, w] [on the back] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2173 [p, w]”; 1♂ (TARI): “Piyasan [written in Japanese] [in Taoyuan] [h] / VII. 1933 [h] / R. takahashi [written in Japanese] [h] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2172 [p, w]”; 1♀ (TARI): “Fukiko [= Fenchihu, in Chiayi] [p] / 29-IV-1931 [p] / Col. T. Shiraki [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2171 [p, w]”; 1♂ (TARI): “Jujiro [= Shihtzulu, in Chiayi] [p] / 26-IV-1931 [p] / Col. T. Shiraki [p, w] // COTYPE [p, circle label with yellow letters] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // 2170 [p, w]”; 1♂, 1♀ (SDEI): “Taihorin [= Talin, in Chiayi] [p] / Formosa [p] / H. Sauter, 1911 [p, w] // 7. VII [p, w] // Syntypus [p, r] // *Paraulaca* [h] / *flavipennis* [h] / CHÛJô [h] / DET. M. CHUJO [p, g] // DEI Müncheberg [p] / Col – 04213 and 04214 [p, g]”. Each paralectotype has a type label: “**Paralectotypus** [p] / *Paraulaca flavipennis* ♂ [or ♀] [p] / ChÛjô, 1935 [p] / des. C.-F. Lee, 2014 [p, pink label]”.

Specimens examined (n= 391). **CHINA: Fujian:** 1♂, Shaowu, Tachulan, 5.IV.1942, leg. T. C. Maa (BPBM); **TAIWAN: Hsinchu:** 1♀, Chienshih, 26.IX.2009, leg. H.-J. Chen (TARI); 1♀, Kuanwu, 30.IV.2010, leg. C.-F. Lee (TARI); 7♂♂, 1♀, same locality, 4.III.2010, leg. L.-H. Sun (TARI); 1♂, 1♀, Litungshan, 15.III.2009, leg. S.-F. Yu (TARI); 1♀, same locality, 6.VI.2010, leg. Y.-L. Lin (TARI); 1♂, same locality, 10.VII.2010, leg. Y.-L. Lin (TARI); 1♀, Lupi, 26.VII.2008, leg. M.-H. Tsou (TARI); 11♀♀, Mamei, 4.V.2008, leg. S.-F. Yu (TARI); 1♂, Peitelaman, 26.VI.2008, leg. S.-F. Yu (TARI); 1♂, Tahunshan, 8.IX.2009, leg. S.-F. Yu (TARI); 1♂, Wuchihshan, 27.III.2008, leg. H. Lee (TARI); 1♂, Wufeng, 17.III.2009, leg. S.-F. Yu (TARI); **Ilan:** 1♀, Taipingshan, 26–28.VII.1983, leg. L. Y. Chou (TARI); **Kaoshiung:** 2♂♂, Erhchituan, 21.VI.2010, leg. U. Ong (TARI); 3♀♀, same locality, 8.III.2013, leg. B.-X. Guo (TARI); **Miaoli:** 1♂, Kuantaoshan, 3.XI.2009, leg. S.-F. Yu (TARI); 1♂, same locality, 5.II.2012, leg. M.-H. Tsou (TARI); 1♂, Sanyi, 6.VII.2013, leg. Y.-T. Chung (TARI); **Nantou:** 1♂, Chingching, 5.III.2007, leg. H.-J. Chen (TARI); 2♂♂, 1♀, same locality, 27.VII.2013, leg. W.-C. Liao (TARI); 2♂♂, 3♀♀, Hoshe, 22.VII.1982, leg. L. Y. Chou & T. Lin (TARI); 1♂, 1♀, Hua-

kang, 14.IX.2010, leg. C.-F. Lee (TARI); 1♀, Nanshanchi, 11.VII.2007, leg. M.-H. Tsou (TARI); 1♀, Meifeng, 20–22.VI.1979, leg. K. S. Lin & B. H. Chen (TARI); 1♂, same locality, 29.VIII–10.IX.1979 (TARI); 1♂, same locality, 8–14.XI.1979 (TARI); 2♂♂, same locality, 2–12.X.1979 (TARI); 1♀, same locality, 24.X.1979, leg. K. C. Chou (TARI); 2♂♂, same locality, 25.X–7.XI.1979 (TARI); 1♂, 4♀♀, same locality, 5–9.X.1980, leg. C. C. Chen & C. C. Chien (TARI); 1♀, same locality, 7–9.V.1981, leg. K. S. Lin & S. C. Lin (TARI); 1♂, same locality, 28–29.VIII.1981, leg. L. Y. Chou & S. C. Lin (TARI); 1♂, 3♀♀, same locality, 7.XI.1981, leg. S. C. Lin & W. S. Tang (TARI); 1♂, same locality, 15.VII.1982, leg. S. C. Lin & C. N. Lin (TARI); 12♂♂, 9♀♀, same locality, 31.VIII–2.IX.1982, leg. L. Y. Chou & K. C. Chou (TARI); 1♂, 1♀, same locality, 4–7.X.1982, leg. K. C. Chou (TARI); 1♀, Rueiyen River Major Wildlife Habitat, 8.VIII.2008, leg. H. Medel & M. V. L. Barclay (BMNH); 3♂♂, 1♀, Sungkang, 13–15.IX.1984, leg. K. S. Lin & S. C. Lin (TARI); 1♂, 4.IV.2010, leg. Y.-T. Wang (TARI); 1♂, 4♀♀, Tsuifeng, 1–3.VIII.1981, leg. T. Lin & W. S. Tang (TARI); 1♂, same locality, 27.VIII.1981, leg. L. Y. Chou & S. C. Lin (TARI); 1♂, same locality, 1–3.IX.1982, leg. L. Y. Chou & K. C. Chou (TARI); 1♂, 4♀♀, Tungpu, 20–22.VI.1980, leg. C. C. Chen (TARI); 2♂♂, 23♀♀, same locality, 28.IV–2.V.1981, leg. T. Lin & C. J. Lee (TARI); 17♂♂, 10♀♀, same locality, 5–8.X.1981, leg. T. Lin & W. S. Tang (TARI); 10♂♂, 9♀♀, same locality, 18–23.XI.1981, leg. T. Lin & W. S. Tang (TARI); 27♂♂, 24♀♀, same locality, 19–23.VII.1982, leg. L. Y. Chou & T. Lin (TARI); 7♂♂, 2♀♀, same locality, 22–25.XI.1982, leg. K. C. Chou & S. P. Huang (TARI); 6♂♂, 5♀♀, same locality, 20–24.VI.1983, leg. K. C. Chou & C. Y. Wong (TARI); 1♂, 13♀♀, same locality, 16–20.IV.1984, leg. K. C. Chou & C. H. Yang (TARI); 27♂♂, 35♀♀, same locality, 23–27.VII.1984, leg. K. C. Chou & C. H. Yang (TARI); 1♂, Wanfengtsun, 2.IV.2008, leg. W.-T. Liu (TARI); 1♂, same locality, 13.IV.2010, leg. W.-T. Liu (TARI); 1♂, Wushe, 23–28.VI.1981, leg. K. S. Lin & W. S. Tang (TARI); 3♂♂, 6♀♀, 30.VIII–2.IX.1982, leg. L. Y. Chou & K. C. Chou (TARI); 1♂, 1♀, same locality, 7–8.X.1982, leg. K. C. Chou (TARI); 1♂, 2♀♀, same locality, 19–22.IV.1983, leg. K. C. Chou & S. P. Huang (TARI); 2♀♀, same locality, 7.V.1984, leg. K. C. Chou & S. P. Huang (TARI); 1♂, same locality, 4.VIII.1984, leg. K. S. Lin (TARI); 5♂♂, same locality, 17.VIII.1984, leg. K. C. Chou (TARI); 1♂, same locality, 11–15.IX.1984, leg. K. S. Lin (TARI); 4♂♂, 1♀, same locality, 21–23.III.2009, leg. U. Ong (TARI); **Pingtung**: 1♀, Shouka, 22.III.2009, leg. M.-H. Tsou (TARI); 1♀, Tahanshan, 18.VII.2007, leg. C.-F. Lee (TARI); **Taichung**: 2♂♂, 3♀♀, Kukuan, 16.VII.2007, leg. M.-H. Tsou (TARI); 2♂♂, 1♀, Lishan, 16.VIII.1984, leg. K. S. Lin & S. C. Lin (TARI); 4♀♀, Wushihkeng, 19.III.2008, leg. C.-F. Lee (TARI); 1♂, Yuantsuishan, 16.VII.2010, leg. J.-C. Chen (TARI); **Taipei**: 2♀♀, Guanyinshan, 14–21.IV.2002, leg. G. Fabián & Merkl O. (JBCB, HNHM); **Taitung**: 4♂♂, Motien, 5.X.2010, leg. C.-F. Lee (TARI); 1♂, 1♀, same locality, 23.V.2011, leg. C.-F. Lee (TARI); **Taoyuan**: 2♂♂, 3♀♀, Hsuanyuan, 16.III.2008, leg. M.-H. Tsou (TARI); 1♂, Sankuang, 17.X.2009, leg. Y.-L. Lin (TARI); 1♂, Tamanshan, 2.VIII.2008, leg. M.-H. Tsou (TARI); 1♀, Tungyangshan, 12.IV.2007, leg. H. Lee (TARI).



Figures 90–95. *Paridea (Paridea) testacea*. **90** Penis, dorsal view **91** Penis, lateral view **92** Gonocoxae **93** Eighth abdominal sternite **94** Eighth abdominal tergite **95** Spermatheca.

Diagnosis. The species is similar to a few individuals of *Paridea (Paridea) taiwana* having reduced black spots on the elytra but differs by the yellow femora and black tibiae (black outer margins of femora and tibiae in *P. (P.) taiwana*).

Males. Length 4.4–4.6 mm, width 2.4–2.5 mm. General color (Figs 80–82) pale yellow; antenna brown; scutellum, tibia, and tarsi blackish brown; mesepimeron and metathoracic ventrites black. Eighth abdominal tergite (Fig. 94) weakly sclerotized, transverse and wide, apical margin slightly emarginate at middle, with dense long seta along apical margin. Penis (Figs 90–91) slender, apically pointed; slightly curved at middle in lateral view; with one pair of elongated sclerites projecting beyond opening; endophallic sclerites composed of one slender sclerite.

Females. Length 4.6–5.3 mm, width 2.4–2.8 mm. Similar to male; apical margin of last abdominal ventrite smooth, not modified. Gonocoxae (Fig. 92) slender, apex of each gonocoxa with seven or eight setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base slender. Sternite VIII (Fig. 93) weakly sclerotized; apex wide, apical margin truncate, surface with dense long setae along apical margin, spiculum long. Spermathecal receptaculum (Fig. 95) slightly swollen; pump long, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Host plants. Cucurbitaceae: *Thladiantha nudiflora* Hemsl. ex Forbes & Hemsl.

Distribution. China (Fujian), China. This species is common and widespread in Taiwan (Fig. 67).

Notes. Taiwanese populations have a black scutellum which differs from the holotype *Paridea testacea* with a yellowish brown scutellum. Actually, most of types of *Paridea testacea* have the scutellum darkened. In addition, all of the studied types of *P. testacea* (including holotype and allotype) are females. One male was found from Maa's collection at the BBPM. Examination of the male confirms that both are conspecific.

Species excluded from Taiwan fauna

Paridea (*Semacia*) *angulicollis* (Motschulsky, 1854)

http://species-id.net/wiki/Paridea_angulicollis

Rhaphidopalpa angulicollis Motschulsky, 1854: 50.

Aulacophora angulicollis: Baly 1874: 186.

Paraulaca (*Aulacophora*) *angulicollis*: Baly 1888: 168.

Paraulaca (*Semacia*) *angulicollis*: Ogloblin 1936: 168.

Semacia (*Semacia*) *angulicollis*: Chûjô and Kimoto 1961: 168.

Paridea (*Paraulaca*) *angulicollis*: Gressitt and Kimoto 1963: 508; Kimoto 1965b: 376.

Paridea (*Paridea*) *angulicollis*: Yang 1991: 269.

Semacia nipponensis Laboissière, 1930: 355; Chûjô and Kimoto 1961: 168 (as synonym of *P. angulicollis*).

Paridea (*Semacia*) *nigrimarginata* Yang, 1991: 279. **syn. n.**

Type locality. China, Beijing.

Type material. *Rhaphidopalpa angulicollis*: Unavailable for study. The type specimens are not present in the Zoological Museum of Moscow State University (Med-

vedev 2006, 2014, personal communication) or in the Zoological Institute in Saint Petersburg (Moseyko 2014, personal communication).

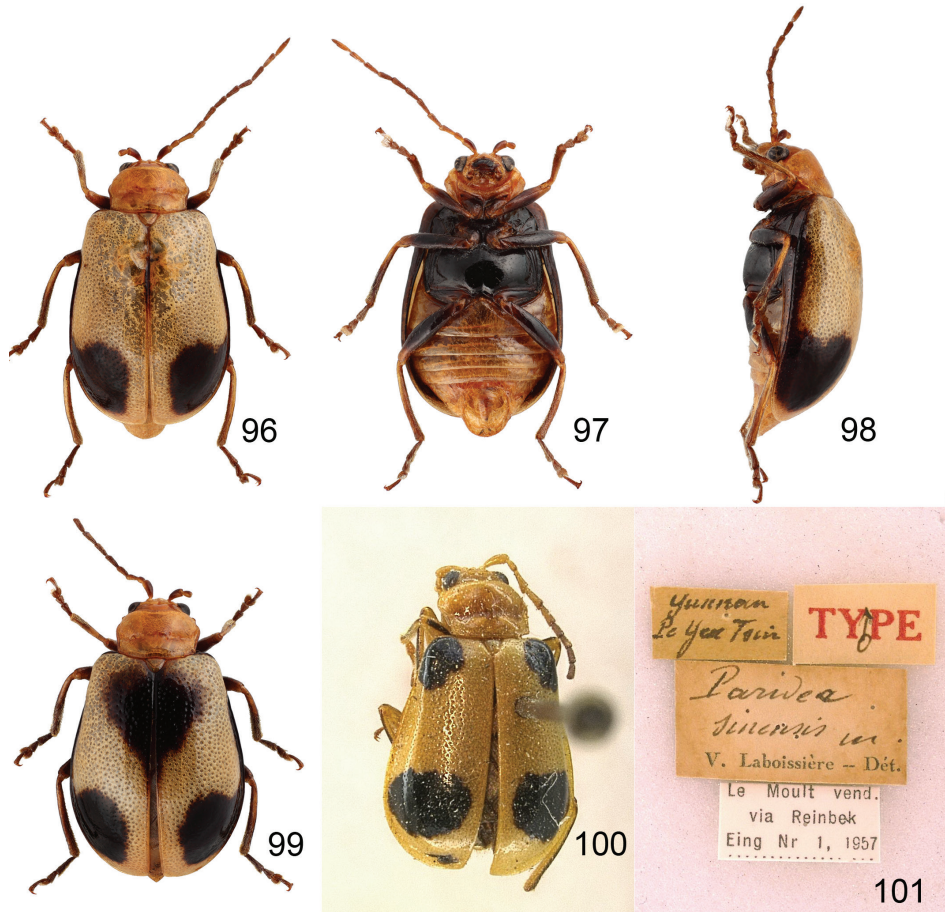
Semacia nipponensis: Lectotype male (MNHN), pinned, here designated to fix the concept of *Semacia nipponensis* and to ensure the universal and consistent interpretation of the same, labeled: “MUSEUM PARIS [p] / NIPPON MOYEN [p] / ENV. DE TOKIO [p] / ET ALPES DE NIKKO [p] / J. HARMAND 1901 [p, w] // *Semacia* [h] / *nipponensis* [h] / m. [h] / V. Laboissière – Dét. [p, w] // TYPE [red letters, p, w] ♂ [inserted between “Y” and “P”, h] // SYNTYPE [p] / *Semacia* [p] / *nipponensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] // MNHN [p] / EC4060 [p, w] // **Lectotypus** [p] / *Senacua nipponensis* ♂ [p] / Laboissière, 1930 [p] / des. C.-F. Lee, 2014 [p, r]”. Paralectotypes: 1♂ (MNHN): “MUSEUM PARIS [p] / NIPPON MOYEN [p] / ENV. DE TOKIO [p] / ET ALPES DE NIKKO [p] / J. HARMAND 1901 [p, w] // SYNTYPE [p] / *Semacia* [p] / *nipponensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] // MNHN [p] / EC4061 [p, w]”; 1♀ (ZMUH): “Tokio [h, w] // TYPE [red letters, p, w] ♀ [inserted between “Y” and “P”, h] // *Semacia* [h] / *nipponensis* m [h] / V. Laboissière – Dét. [p, w]”. Each paralectotype has a type label: “**Paralectotypus** [p] / *Semacia nipponensis* ♂ [or ♀] [p] / Laboissière, 1930 [p] / des. C.-F. Lee, 2014 [p, pink label]”

Paridea (Semacia) nigrimarginata: Holotype ♂ (IZAS), labeled: “Mt. Takao [p] / June 11 32 [blue letters, p, w] // HOLOTYPE [p, r] // *Paridea* (S.) [h] / *nigrimarginata* [h] / sp n [h] / 鑑定者 [p]: 楊.[h, w] [= det. Yang]”.

Additional material examined (n= 14). CHINA. Jiangxi: 1♂ (BPBM); **Hubei** (= Hupeh): 1♂, Lichuan, Lianghoken, 7.IX.1948, leg. Gressitt & Djou (BPBM); **Sichuan:** 2♂♂, Bayueshan, 21.IV.2013, leg. J. Y. Qiu & H. Xu (TARI); **JAPAN. Honshu:** 1♀ (KMNH), Aomori Pref., Towada, 20.VII.1980, leg. S. Kawauchi; 1♀ (KMNH), Kanagawa Pref., Tanzawa, Ooyama, 23.V.1966, leg. Y. Kusui; 1♂ (CAS), Tokyo, 27.IV.1930, leg. L. Gressitt; 1♀ (CAS), Tokyo Pref., Mt. Takao, 4.V.1930, leg. L. Gressitt; 1♀ (CAS), same locality, 14.VI.1959, leg. H. Toshimii; 1♀ (KMNH), Kyoto Pref., Minoo – Takayama, 8.V.1956, leg. K. Morimoto; **Kyushu:** 3♂♂ (KMNH), Fukuoka Pref., Mt. Sefuri, 10.VI.1956, leg. H. Kamiya; 1♂ (KMNH), Fukuoka Pref., Kurokidaira, 23.V.1979.

Diagnosis. See diagnosis of *Paridea (Semacia) kaoi* sp. n.

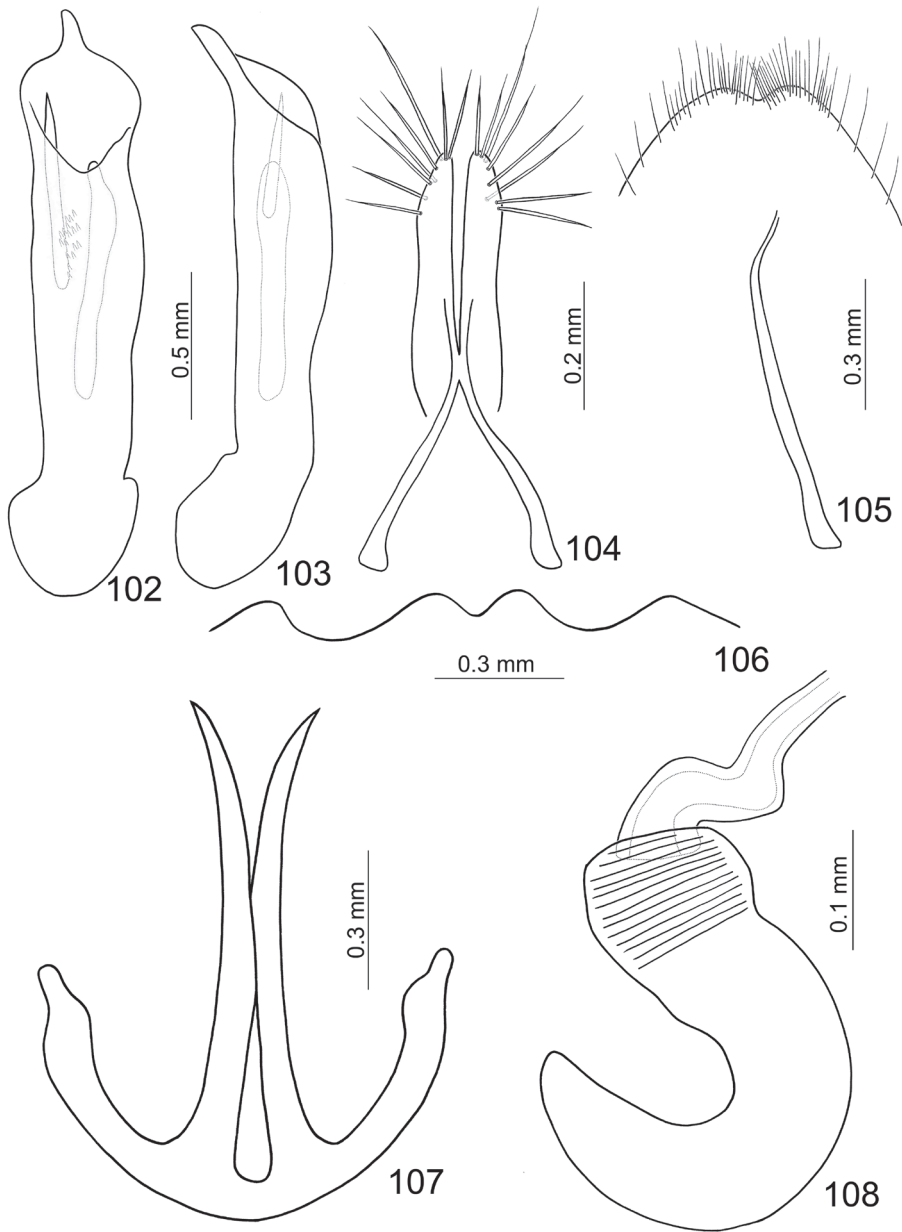
Males. Length 5.1–5.5 mm, width 3.1–3.3 mm. Head and prothorax yellowish brown (Figs 96–98), labrum black, antenna brown; scutellum pale yellow; elytra pale yellow, with deep excavation behind scutellum at suture; with one extremely slender black stripe along suture behind excavation, sometimes reduced; with one pair of large black spots subapically, lateral margin and epipleuron black, extending posterior and connected with subapical black spots; meso- and metathoracic ventrites black; legs dark brown, apex of femur and base of tibia paler; abdomen yellow. Eighth abdominal tergite (Fig. 107) strongly sclerotized, transverse and slender, with one pair of extremely slender and curved processes. Pygidium slightly projecting beyond elytral apices. Penis slightly asymmetric, slightly narrowed at apical 1/6; apex narrow, tubular, and small; straight from lateral view; endophallic sclerites with one pointed sclerite, one elongate sclerite, and a cluster of large setae.



Figures 96–101. *Paridea* species. **96** *P. (S.) angulicollis*, male, dorsal view **97** ditto, ventral view **98** ditto, lateral view **99** *P. (S.) angulicollis*, female, dorsal view **100** *P. (P.) sinensis*, lectotype, dorsal view **101** *P. (P.) sinensis*, lectotype, labels.

Females. Length 5.1–5.6 mm, width 3.0–3.4 mm. Similar to male (Fig. 99), elytra without excavation but black spot instead. Apical margin of last abdominal ventrite (Fig. 106) with one pair of small rounded processes at middle, slightly emarginate outside processes. Pygidium slightly projecting beyond elytral apices. Gonocoxae (Fig. 104) slender, apex of each gonocoxa with eight setae from apical 1/7 to apex; connection of gonocoxae extremely slender, base widened. Sternite VIII (Fig. 105) weakly sclerotized; apex narrow, apical margin emarginate at middle, surface with dense long setae along apical margin, spiculum short. Spermathecal receptaculum (Fig. 108) slightly swollen; pump much longer than receptaculum, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.,

Distribution. Japan, China (Jiangxi, Hubei, Sichuan). Yang (1991) indicated that this species was also found in Zhejiang, Fujian, Hainan, and Guansi provinces of



Figures 102–108. *Paridea (Semacia) angulicollis*. **102** Penis, dorsal view **103** Penis, lateral view **104** Gonocoxae **105** Eighth abdominal sternite **106** Fifth abdominal ventrite **107** Eighth abdominal tergite **108** Spermatheca.

China. These records are dubious since no voucher specimens were examined and Japanese populations were misidentified as *Paridea (Semacia) nigrimarginata* Yang, 1991 (see below).

Host plants. Cucurbitaceae: *Gynostemma pentaphyllum* (Thunb.) Makino, *Trichosanthes cucumeroides* (Ser.) Maxim. ex Fr. & Sav. (Chûjô and Kimoto 1961).

Notes. The position of the type locality of *Paridea* (*Semacia*) *nigrimarginata* was doubtful since information on the label is insufficient. Only “Mt. Takao” appears on the label and Beenen (2010) supposed that it was located in Taiwan since “Takao” is the Japanese name for Kaoshiung City, but Kaoshiung city it is not a mountain. Mt. Takao probably refers to a locality in Japan since a famous mountain (Mt. Takao-Yama, Takao-machi, Hachioji-shi, Tokyo Pref., Japan) exists there with similar names. Moreover, subsequent material (see specimens examined) came from this locality with additional information indicating the Japanese Mt. Takao.

***Paridea* (*Paridea*) *sinensis* Laboissière, 1930**

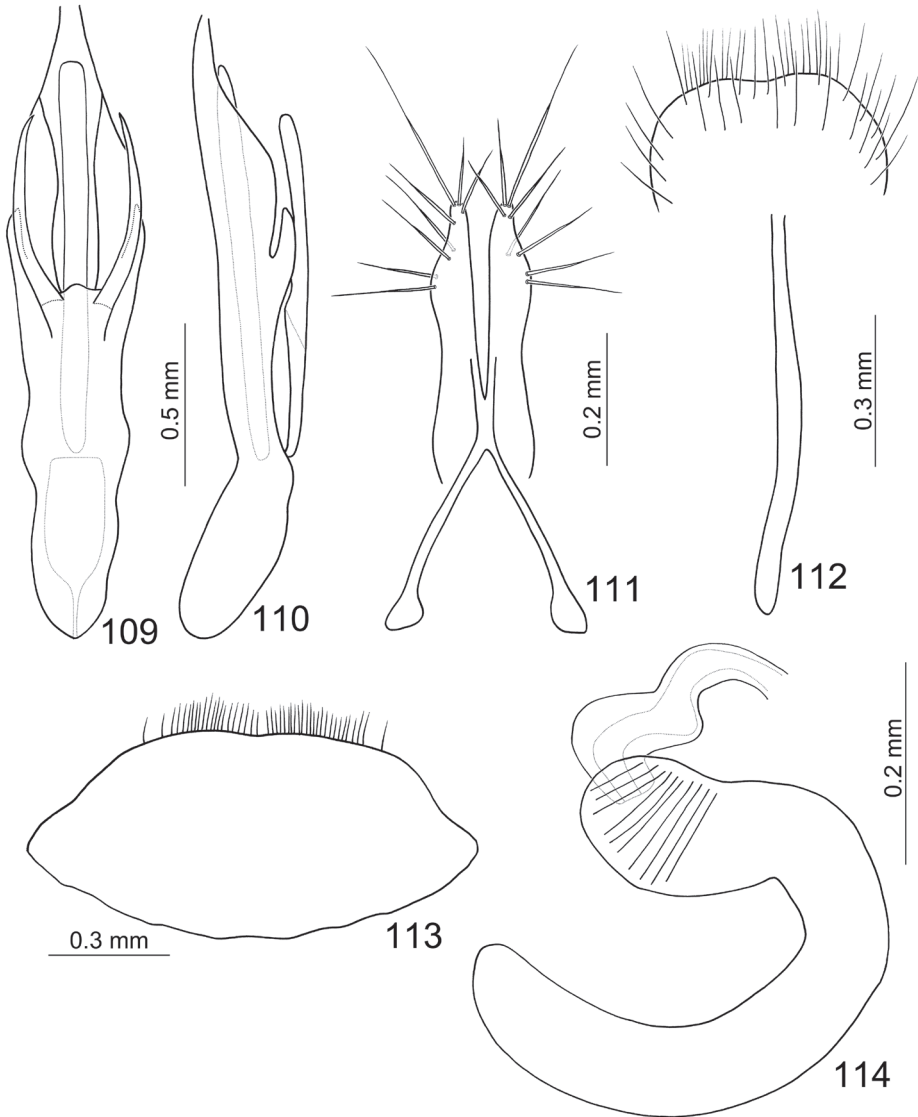
http://species-id.net/wiki/Paridea_sinensis

Paridea sinensis Laboissière, 1930: 342.

Paridea (*Paridea*) *sinensis*: Gressitt and Kimoto 1963: 514.

Type locality. China, Yunnan, PeYenTsin.

Type material. Lectotype male (Fig. 100) (ZMUH), pinned, here designated to fix the concept of *Paridea sinensis* and to ensure the universal and consistent interpretation of the same, labeled (Fig. 101): “Yunnan [h] / Pe Yen Tsin [h, w] // TYPE [red letters, p] ♂ [p] // *Paridea* [h] / *sinensis* [h] / m [h] / V. Laboissière – Dét. [p, w] // Le Moul't Vend. [p] / via Reinbek [p] / Eing Nr. 1, 1957 [p, w] // **Lectotypus** [p] / *Paridea sinensis* ♂ [p] / Laboissière, 1930 [p] / des. C.-F. Lee, 2014 [p, r]”. Paralectotypes: 1♀ (ZMUH): “Yunnan [h] / Pe Yen Tsin [h, w] // Le Moul't Vend. [p] / via Reinbek [p] / Eing Nr. 1, 1957 [p, w]”; 1♀ (ZMUH): “**PE YEN TSIN** [p] / **YUNNAN** [p] / Coll. de Touzalin [p, w] // Le Moul't Vend. [p] / via Reinbek [p] / Eing Nr. 1, 1957 [p, w]”; 1♀ (ISNB): “**PE YEN TSIN** [p] / **YUNNAN** [p] / Coll. de Touzalin [p, w], glued on larger card, labeled: “*Coll. R. I. Sc. N. B.* (p) / Chine [p, y] // *Paridea* [h] / *sinensis* [h] / m [h] / V. Laboissière – Dét. [p] / 1930 [vertical, h, w] // Para- [p] / type [p, o]”; 1♀ (MNHN): “MUSEUM PARIS [p] / YUNNAN [p] / S.-O 24 ° N [p] / PE. YEN. TSIN [p] / (MINES DE SEL) [p] / (PÈRE SIMÉON TEN) [p] / P. CUERRY 1924 [p, w] // NOVEMBRE [p, w] // SYNTYPE [p] / *Paridea* (*Paridea*) [p] / *sinensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] / MNHN [p] / EC4062 [p, w]”; 1♀ (MNHN): “MUSEUM PARIS [p] / KOUY-TCHÉOU [p] / RÉG. DE PIN-FA [p] / PÈRE CAVALERIE 1908 [p, w] // SYNTYPE [p] / *Paridea* (*Paridea*) [p] / *sinensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] / MNHN [p] / EC4063 [p, w]”; 1♀ (MNHN): “MUSEUM PARIS [p] / SE-TCHOUEN [p] / ENV DE TA-TSIEN-LOU [p] / MO-SY-MIEN [p] / Père AUBERT 1902 [p, w] // *Paridea* [h] / *sinensis* [h] / m [h] / V. Laboissière – Dét. [p, w] // COTYPE [red letters, p, w] // SYNTYPE [p] / *Paridea* (*Paridea*) [p] / *sinensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] / MNHN [p] / EC4064 [p, w]”; 2♀♀ (MNHN): “MUSEUM PARIS [p] / SE-TCHOUEN [p]



Figures 109–114. *Paridea (Paridea) sinensis*. **109** Penis, dorsal view **110** Penis, lateral view **111** Gonocoxae **112** Eighth abdominal sternite **113** Eighth abdominal tergite **114** Spermatheca.

/ ENV DE TA-TSIEN-LOU [p] / MO-SY-MIEN [p] / Père AUBERT 1902 [p, w] // SYNTYPE [p] / *Paridea (Paridea)* [p] / *sinensis* Laboissière, 1930 [p, w] // SYNTYPE [p, r] / MNHN [p] / EC4065 or 4066 [p, w]”. Each paralectotype has a type label: “**Paralectotypus** [p] / *Paridea sinensis* ♂ [or ♀] [p] / Laboissière, 1930 [p] / des. C.-F. Lee, 2014 [p, pink label]”

Additional material examined (n= 2). **CHINA: Sichuan:** 1♂ (TARI), Luding, Moxi, 20.VI.1983, leg. Y.-Q. Chen; 1♀ (TARI), same but with “leg. S.-Y. Wang.

Diagnosis. See diagnosis of *Paridea* (*Paridea*) *taiwana*.

Males. Length 6.5 mm, width 3.6 mm. General color (Fig. 100) yellowish brown, elytra with two pairs of black spots: one at humerus smaller, other subapical and larger; outer margins of femora and tibiae black; metathoracic ventrites black. Eighth abdominal tergite (Fig. 113) weakly sclerotized, transverse and wide, apical margin slightly emarginate at middle, with dense long seta along apical margin. Penis (Fig. 109–110) slender; external process extending anterior, much longer than medial process; medial process small; straight in lateral view.

Females. Length 6.8 mm, width 3.8 mm. Similar to male; apical margin of last abdominal ventrite smooth, not modified. Gonocoxae (Fig. 111) slender, apex of each gonocoxa with eight setae from apical 1/5 to apex; connection of gonocoxae extremely slender, base widened. Sternite VIII (Fig. 112) weakly sclerotized; apex narrow, apical margin slightly concave at middle, surface with longer setae near apical margin and shorter setae on apical margin, spiculum long. Spermathecal receptaculum (Fig. 114) slightly swollen; pump extremely long, strongly curved; spermathecal duct short, stout, shallowly projecting into receptaculum.

Distribution. China (Jiangxi, Hubei, Fujian, Sichuan, Yunnan, Guizhou). The distribution of *Paridea* (*Paridea*) *sinensis* should be reevaluated since specimens collected from Fujian identified by Gressitt & Kimoto are *Paridea* (*Paridea*) *fujiana* Yang, 1991.

Acknowledgements

We would like to thank all curators listed above for giving us the opportunity to study the material from their collections. We thank the Taiwan Chrysomelid Research Team for assistance in collecting material, including J.-C. Chen, H.-J. Chen, H.-T. Cheng, Y.-T. Chung, B.-X. Guo, H. Lee, W.-C. Liao, M.-H. Tsou, and S.-F. Yu. We especially thank T.-H. Lee and H.-T. Cheng for photos of dorsal and ventral habitus of each species. We also thank C.-K. Yang, graduate student at the department of Life Science, National Taiwan Normal University with expertise in plant taxonomy, for identification of host plants. This study was also supported by the Ministry of Science and Technology of Taiwan NSC 102-2313-B-055-002. We are grateful to Prof. Christopher Carlton (Louisiana State Arthropod Museum, USA) for reviewing the manuscript.

References

- Baly JS (1874) Catalogue of the Phytophagous Coleoptera of Japan, with descriptions of the species new to science. The Transactions of the Entomological Society of London 1874: 161–217.
- Baly JS (1886) Descriptions of a new genus and of some new species of Galerucinae, also diagnostic notes on some of the older described species of *Aulacophora*. The Journal of the Linnean Society of London (Zoology) 20: 1–27. doi: 10.1111/j.1096-3642.1886.tb01432.x

- Baly JS (1888) Descriptions of some genera and species of Galerucinae. The Journal of Linnean Society of London (Zoology) 20: 156–188. doi: 10.1111/j.1096-3642.1888.tb01444.x
- Beenen R (2010) Galerucinae. In: Löbl I, Smetana A (Ed) Catalogue of Palaearctic Coleoptera Vol. 6. Apollo Books, Stenstrup, 443–491.
- Chevrolat LAA (1836) [new taxa]. In: Dejean PFAM (Ed) Catalogue des coléoptères de la collection de M. le Comte Dejean. Deuxième édition. Livraison 5. Méquignon-Marvis Père et Fils, Paris, 361–442.
- Chûjô M (1935) H. Sauter's Formosa-Ausbeute: Subfamily Galerucinae (Coleoptera: Chrysomelidae). Arbeiten über Morphologische und Taxonomische Entomologie 2: 160–174.
- Chûjô M (1938) H. Sauter's Formosa-Collection: Subfamily Galerucinae. (Coleoptera: Chrysomelidae). Arbeiten über Morphologische und Taxonomische Entomologie 5: 135–152.
- Chûjô M (1962) A taxonomic study on the Chrysomelidae (Insecta: Coleoptera) from Formosa. Part IX. Subfamily Galerucinae. The Philippine Journal of Science 91: 1–239.
- Chûjô M (1963) Chrysomelid-beetles from Formosa, preserved in the Hungarian Natural History Museum, Budapest. Annales Historico-Naturales Musei Nationalis Hungarici 55: 379–402.
- Chûjô M (1965) Chrysomelid-beetles of Formosa (I). Special Bulletin of Lepidopterological Society of Japan 1: 88–104.
- Chûjô M, Kimoto S (1961) Systematic catalog of Japanese Chrysomelidae (Coleoptera). Pacific Insects 3: 117–202.
- Fairmaire L (1889) Coléoptères de l'intérieur de la Chine, 5^e partie. Annales de al Société Entomologique de France 6: 5–84.
- Gressitt JL, Kimoto S (1963) The Chrysomelidae (Coleopt.) of China and Korea, Part 2. Pacific Insects Monographs 1B: 301–1026.
- Hope FW (1831) Synopsis of new species of Nepaul insects in the collection of Major General Hardwicke. In: Gray JE (Ed) Zoological Miscellany. Vol. 1. Treuttel, Wurtz & Co., London, 21–32.
- Jolivet P, Hawkeswood TJ (1995) Host-plants of Chrysomelidae of the World. Backhuys Publishers, Leiden, 281 pp.
- Kimoto S (1965a) A list of specimens of Chrysomelidae from Taiwan preserved in the Naturhistorisches Museum / Wien (Insecta: Coleoptera). Annalen des Naturhistorischen Museums in Wien 68: 485–490.
- Kimoto S (1965b) The Chrysomelidae of Japan and the Ryukyu Islands. VII. Subfamily II. Journal of the Faculty of Agriculture, Kyushu University 13: 369–400.
- Kimoto S (1966) A list of the Chrysomelid specimens of Taiwan preserved in the Zoological Museum, Berlin. Esakia 5: 21–38.
- Kimoto S (1969) Notes on the Chrysomelidae from Taiwan II. Esakia 7: 1–68.
- Kimoto S (1974) Notes on the Chrysomelidae from Taiwan VII. Entomological Review of Japan 26: 21–26.
- Kimoto S (1986) The Chrysomelidae (Insecta: Coleoptera) collected by the Nagoya University Scientific Expedition to Taiwan in 1984. Kurume University Journal 35: 53–62.
- Kimoto S (1989a) The Taiwanese Chrysomelidae (Insecta: Coleoptera) collected by Dr. Kintaro Baba, on the occasion of his entomological survey in 1983 and 1986. Kurume University Journal 38: 237–272.

- Kimoto S (1989b) Chrysomelidae (Coleoptera) of Thailand, Cambodia, Laos and Vietnam IV. Galerucinae. *Esakia* 27: 1–241.
- Kimoto S (1991) The Taiwanese Chrysomelidae (Insecta: Coleoptera) collected by Dr. Kintaro Baba, on the occasion of his entomological survey in 1987, 1988 and 1989. *Kurume University Journal* 40: 1–27.
- Kimoto S, Takizawa H (1997) Leaf beetles (Chrysomelidae) of Taiwan. Tokai University Press, Tokyo, 581 pp.
- Laboissière V (1929) Observations sur les Galerucini asiatiques principalement du Tonkin et du Yunnan et descriptions de nouveaux genres et espèces. *Annales de la Société Entomologique de France* 96: 37–62.
- Laboissière V (1930) Observations sur les Galerucini asiatiques principalement du Tonkin et du Yunnan et descriptions de nouveaux genres et espèces. (2e partie). *Annales de la Société Entomologique de France* 99: 325–368.
- Medvedev LN (2006) To the knowledge of Chrysomelidae (Coleoptera) described by V. Motschulsky. *Russian Entomological Journal* 15: 409–417.
- Medvedev LN, Samoderzhenkov EV (1997) Revision of *Paridea* from the Himalaya and Himalaya and adjacent regions. *Russian Entomological Journal* 6: 57–65.
- Motschulsky V de (1854) Diagnoses de coléoptères nouveaux trouvés par M. M. Tatarinoff et Gaschkévitch aux environs de Pékin. *Études Entomologiques* 2: 44–51.
- Ogloblin DA (1936) Listoedy, Galerucinae. *Fauna SSSR. Nasekomye Zhestkokrylye* [Chrysomelidae, Galerucinae. Faune de l'URSS, Insectes Coléoptères], n. s. 8, 23(1). Moskva-Leningrad, Izdatel'stvo Akademii Nauk SSSR, 455 pp. [in Russian]
- Takizawa H, Nakamura S, Kojima K (1995) The Taiwanese chrysomelid beetles preserved in Hiwa Museum for Natural History (Coleoptera: Coleoptera). *Miscellaneous Reports of the Hiwa Museum for Natural History* 33: 1–16. [in Japanese]
- Yang X (1991) Study on the genus *Paridea* Baly from China (Chrysomelidae: Galerucinae). *Sinozoologia* 8: 267–295.

A new genus of nemonychid weevil from Burmese amber (Coleoptera, Curculionoidea)

Steven R. Davis^{1,†}, Michael S. Engel^{1,‡}

¹ Division of Entomology, Natural History Museum, and Department of Ecology & Evolutionary Biology, 1501 Crestline Drive – Suite 140, University of Kansas, Lawrence, Kansas 66045, USA

[†] <http://zoobank.org/A6D33B50-CDD6-460E-838E-AC7BF43888A2>

[‡] <http://zoobank.org/3714A7FF-E19E-495A-AAF9-98D2F597B757>

Corresponding author: Steven R. Davis (steved@ku.edu)

Academic editor: M. Alonso-Zarazaga | Received 22 October 2013 | Accepted 7 April 2014 | Published 28 April 2014

<http://zoobank.org/96C776FF-525D-48E4-BF07-455703DF6947>

Citation: Davis SR, Engel MS (2014) A new genus of nemonychid weevil from Burmese amber (Coleoptera, Curculionoidea). ZooKeys 405: 127–138. doi: 10.3897/zookeys.405.6475

Abstract

The first fossil nemonychid (Nemonychidae) in Burmese amber, belonging to the subfamily Rhinorhynchinae, is described and figured as *Burmonyx zigrasi* Davis & Engel, **gen. n.** and **sp. n.** While this specimen also comprises the first definitive record of the subfamily in the Asian continent, other compression fossils exist at least from the Yixian Formation of China and the Karatau site of Kazakhstan which may also deserve placement within this group. Although several important areas of the body are obscured by the shape and fragmented condition of the amber piece, a sufficient number of features are visible to consider adequate placement within Rhinorhynchinae, including the fairly strongly punctate elytral striae and appendiculate, nearly bifid pretarsal claws.

Keywords

Coleoptera, amber, Cretaceous, weevils, Nemonychidae, Mesozoic, taxonomy

Introduction

The origin and evolution of weevils (Curculionoidea) remains one of the more interesting and challenging areas of systematic research among the Coleoptera (Grimaldi and Engel 2005). The distinctive weevil rostrum has been implicated as a key innovation intimately tied to their considerable success in terms of species and ecological diversity (Anderson 1995). Although much remains to be illuminated regarding the larger patterns of weevil phylogeny, one thing is certain: among the extant families, the Nemomychidae are the most basal offshoot of the lineage (Kuschel 1995; Oberprieler et al. 2007). Often familiarly known as pine cone weevils owing to their association with diet of pollen from pines and related gymnosperms, nemomychids primitively retain non-geniculate (orthocerous) antennae, lack pronotal lateral carinae (except apparently in the nemomychid subfamily Eobelinae), and have the abdominal ventrites separate (rather than partially fused or entirely connate). Given this critical phylogenetic position, it is understandable why nemomychids play a large role in narratives regarding the early order of weevils (e.g., Legalov 2010b, 2012; Oberprieler et al. 2007).

Thorough accounts of the Mesozoic fossil record of Nemomychidae (and of the superfamily Curculionoidea) have been provided and reiterated several times (e.g., Kuschel 1983; Kuschel and Leschen 2011; Oberprieler and Oberprieler 2012; Riedel 2010), therefore it will not be repeated here. Extant members of the subfamily Rhinorhynchinae are distributed in the southwest Nearctic, Neotropical, and Australian regions (Kuschel and Leschen 2011). While there appear to be fossil representatives of this subfamily in various deposits in Asia (e.g., Karatau, Yixian), Asian fossil taxa have yet to be ascribed to this lineage. The description of the new genus and species herein represents the first nemomychid found in Burmese amber (Figs 1, 2), as well as the first record of the subfamily Rhinorhynchinae in Asia.

Material and methods

The amber piece containing the holotype was excavated from the strata in the northern state of Kachin in Myanmar as part of regular and ongoing mining operations and is from the collection of Mr. James S. Zigras, available for study through the American Museum of Natural History (AMNH), New York. The origin, age, and fauna of Burmese amber have been reviewed by Grimaldi et al. (2002), Ross et al. (2010), and Shi et al. (2012), the latter of which arrived at an age of approximately 99 Ma (providing a range close to the Aptian-Cenomanian boundary). Due to the presence of several other insect inclusions and the heavily fragmented state of the amber piece, further preparation beyond some initial polishing was not possible, leaving some aspects of the specimen unobservable or obscured. Due to the round surface of the piece, glycerin was applied to a small coverslip and placed on the area directly above the inclusion in order to acquire more satisfactory viewing and photography. Photomicrographs were obtained by combining a z-stack of around 25 images using the computer software CombineZ.

Illustrations were made through the aid of a drawing tube attached to an Olympus SZX9 stereomicroscope. The general familial classification of Curculionoidea follows that of Oberprieler et al. (2007).

Systematic paleontology

Family Nemonychidae Bedel

Subfamily Rhinorhynchinae Voss

Burmonyx Davis & Engel, gen. n.

<http://zoobank.org/C7493C8C-E242-4EC9-BD39-9BC4C4C579F9>

<http://species-id.net/wiki/Burmonyx>

Type species. *Burmonyx zigrasi* Davis & Engel, sp. n. Gender feminine.

Diagnosis. The new genus appears similar to several genera in Rhinorhynchinae, particularly to those of Rhinorhynchini, due to the fairly long, narrow rostrum (Figs 1, 3) and moderately wide pronotum. It appears to be differentiated based mainly on its elongate scutellary striole. The striole in extant Nemonychidae always is fairly short and not extending more than $0.25 \times$ the length of the elytra from the base. This genus, however, possesses a striole that extends approximately to the middle of the elytron. The relatively large stria punctures on the elytra may also be a defining feature of this genus, as the punctures typically are quite small in other members of Rhinorhynchinae.

Etymology. The genus name is a combination of the specimen's collection locality, Burma (former name of Myanmar), and the Greek *nyx*, meaning "night".

Burmonyx zigrasi Davis & Engel, sp. n.

<http://zoobank.org/15B9C6E0-166E-4D25-A142-5B8E12517468>

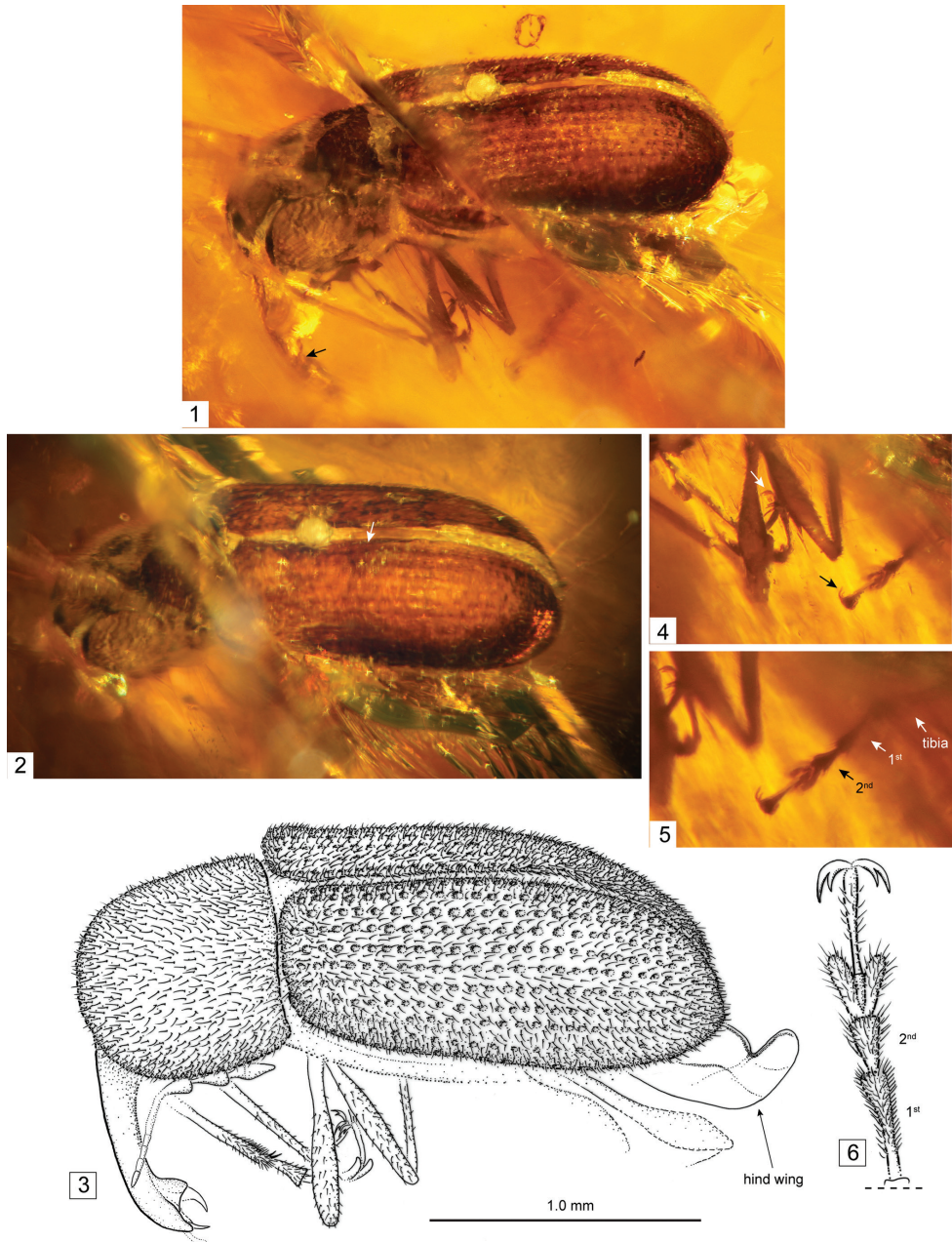
http://species-id.net/wiki/Burmonyx_zigrasi

Figs 1–7

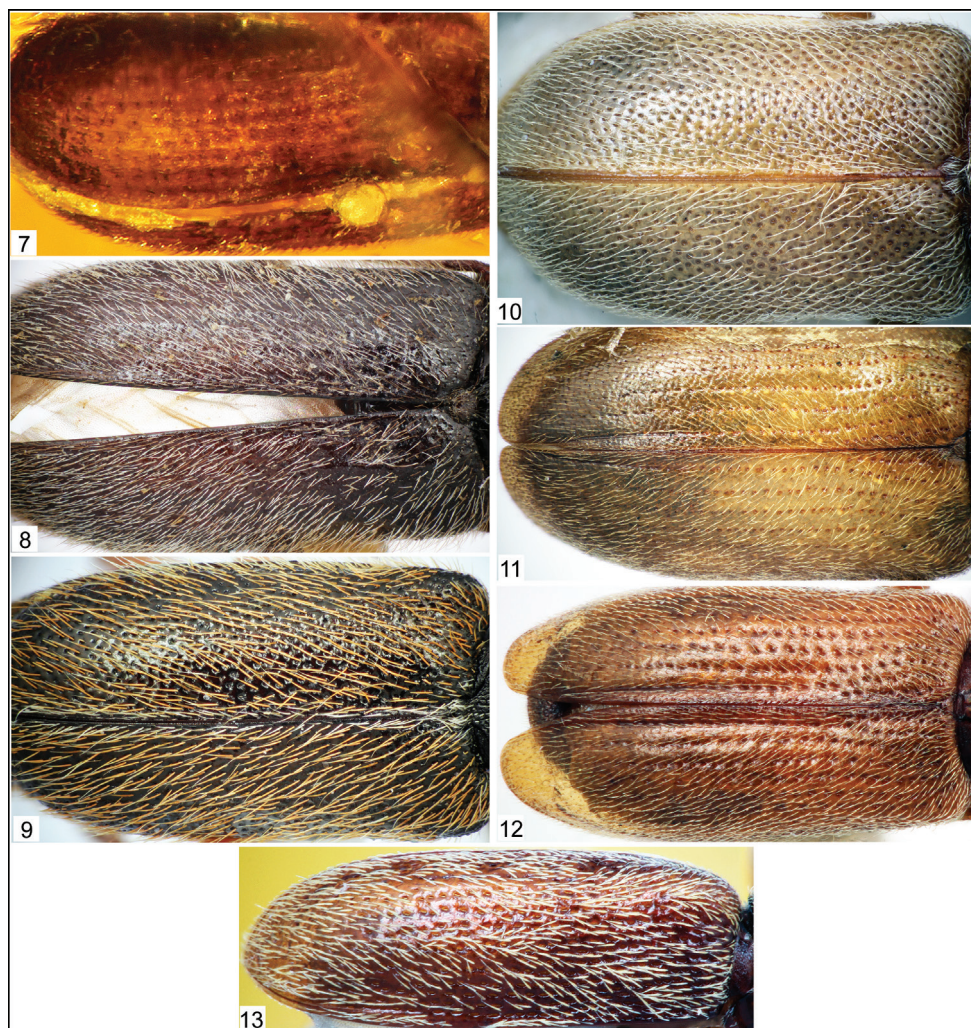
Holotype. JZC-Bu228, Myanmar: Kachin; Cretaceous: Early Cenomanian; in the private collection of Mr. James S. Zigras, available for study through the Division of Invertebrate Zoology, American Museum of Natural History (AMNH), New York, USA. Material was obtained from ongoing excavations in mines in the Hukawng Valley, northern Myanmar (Grimaldi et al. 2002). The raw material was brought back to Myitkyina where initial polishing and sorting was undertaken prior to final preparation of pieces in the AMNH.

Diagnosis. As for the genus (*vide supra*).

Description. Total body length (excluding rostrum): ca. 2.2 mm; maximal width (along middle of elytra): ca. 0.7 mm; elytral length: ca. 1.3 mm. Integument appearing light to dark brown (Figs 1, 2, 7). Scales absent, but dense covering of setae along at



Figures 1–6. Photomicrographs (**1**, **2**, **4**, **5**) and line drawings (**3**, **6**) of holotype of *Burmonyx zigrasi* Davis and Engel, sp. n. (JZC-Bu228). **1** Dorso-lateral view of specimen inclusion, arrow pointing to antennal scape **2** Slightly more dorsal view of specimen than in figure 1, arrow pointing to scutellary striae **3** Line drawing of specimen (scale bar only applies to this figure) **4** Legs, arrows pointing to appendiculate, nearly bifid pretarsal claws **5** Enlargement of legs, arrows pointing to metatibia and metatarsomeres **6** Illustration of metatarsus.



Figures 7–13. Photomicrographs of nemonychid elytra (dorsal aspect). **7** *Burmonyx zigrasi* Davis and Engel, sp. n. **8** *Nemonyx lepturoides* (Fabricius, 1801) **9** *Cimberis elongata* (LeConte, 1876) **10** *Doydirhynchus austriacus* (Olivier, 1807) **11** *Basiliorhinus araucariae* Kuschel, 1994 **12** *Nannomacer germaini* (Voss, 1952) **13** *Rhinorhynchus rufulus* (Broun, 1880).

least pronotum and elytra (lateral and ventral surfaces not clearly visible). Head and compound eyes not clearly visible (slightly pushed into and obscured by prothorax). Rostrum approximately as long as pronotum along middle (exact length ratio unclear due to obscured head), fairly slender, abruptly widening apically (Fig. 3). Mandibles large, falciform. Antennae orthocerous, inserted dorso-laterally at apical 1/4; clubs composed of 3 loose articles. Pronotum seemingly as wide or nearly as wide as elytral humeri; not constricted anteriorly at collar, slightly rugulose, bearing small,

dense, shallow punctures. Mesoscutellum not visible. Elytra with ten shallowly punctate striae (Figs 2, 3, 7); scutellary striae present, extending approximately to mid-length of elytra; interstices lacking punctures; elytral shoulders rounded. Abdomen with pygidium (tergite VII) concealed. Legs approximately equal in length, slender; femora slender; trochanters small, triangular; tibial spur formula 2-2-2; tarsomere 1 rather narrow, elongate (Fig. 6), approximately 2 × as long as tarsomere 2, 2 with rounded apico-lateral margins; 3 strongly bilobed (Figs 5, 6), lobes narrow; 4 short, slightly longer than 0.5 × length of tarsomere 3; 5 slender, approximately 2 × as long as tarsomere 2; pretarsal claws (ungues) divaricate strongly appendiculate, nearly bifid (Figs 4, 6).

Etymology. The specific epithet is dedicated to the collector, Mr. James S. Zigras, who permitted study of the material and has generously supported amber research.

Discussion

Presence of scutellary striae excludes a placement of *Burmonyx* in Caridae, Brentidae, and Curculionidae. However, as the dorsal area encompassing the elytral suture is somewhat difficult to observe, if the presence of striae represents a misinterpretation of complete striae, a superficial resemblance to the group Mesophyletinae Poinar (2006) may be assumed primarily in the shape of the tarsomeres and divaricate bifid pretarsal claws. Beyond these similarities, *Burmonyx* possesses orthocerous antennae (in which the scape is short and similar in length to subsequent antennomeres) and tibiae with smooth dorsal margins, sufficient information to exclude such a placement. The absence of a distinct pygidium, incompletely covered by the elytra and typically extending ventrad, and the conical second tarsomeres that do not envelope the base of the third, excludes it from Anthribidae and Attelabidae.

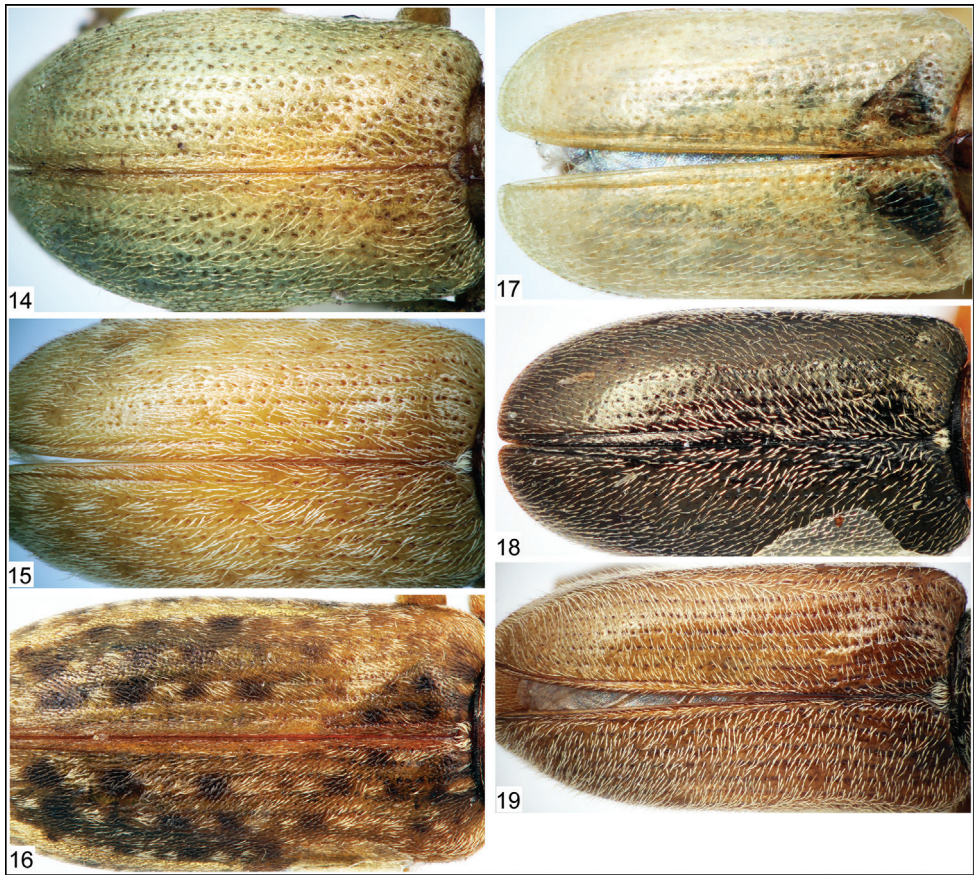
In Nemonychidae, as members of Nemonychinae do not possess distinctly punctate striae (Fig. 8), those of Cimberidinae lack striae (Figs 9–10), and both groups lack elytral striae, *Burmonyx* does not belong within those subfamilies and a possible placing within Rhinorhynchinae or Eobelinae remains. Eobelinae are a difficult assemblage of taxa to comprehend in the least, and it is yet rather unclear how these different groups relate to the extant nemonychid fauna. It is clear that many of the taxa, which appear to have simple, divaricate pretarsal claws and a rostrum emerging from the ventral part of the head capsule, also bear distinctly punctate elytral striae, and may also bear scutellary striae (Table 1; Figs 20–29). It also is interesting to note that there are some undescribed taxa from the Jurassic deposits of Karatau (Kazakhstan) and Daohugou (Inner Mongolia, China) that have a dense scattering of elytral punctures and lack elytral striae (appearing to also lack scutellary striae), similar to extant Cimberidinae. Nonetheless, *Burmonyx* can be excluded from Eobelinae based on its appendiculate, almost bifid pretarsal claws.

Kuschel (1995) and Kuschel and Leschen (2011) listed several synapomorphies for Rhinorhynchinae, of which only a few are visible in *Burmonyx*. In this new genus,

Table 1. Exemplar taxa sampled from the four recognized subfamilies of Nemonychidae (Alonso-Zaraza-ga and Lyal 1999; Kuschel and Leschen 2011), summarizing the layout of punctures on the elytra and indicating the presence of a scutellary striae.

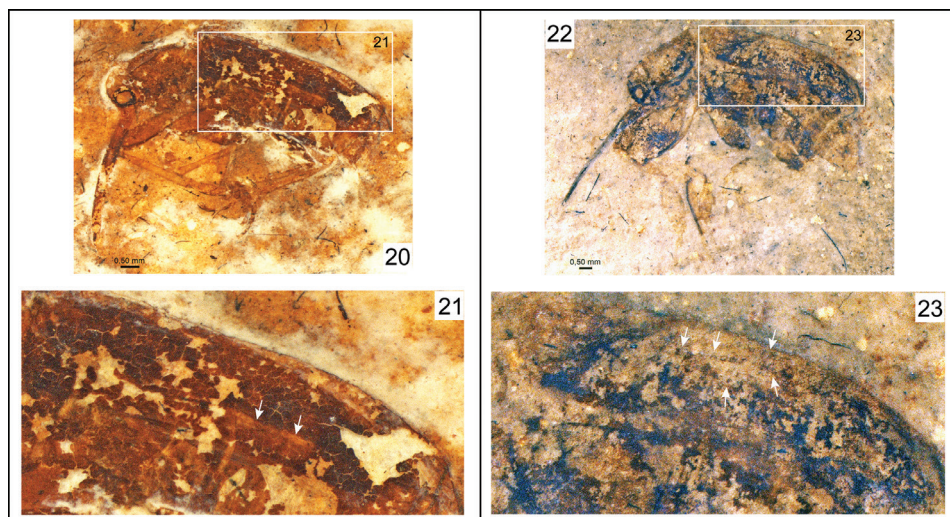
SUBFAMILY: Tribe	Species	Elytron puncture type
NEMONYCHINAE	<i>Nemonyx lepturoides</i> (Fabricius, 1801)	Punctures scattered, striae present, faint; striae absent
CIMBERIDINAE: Cimberidini	<i>Cimberis elongata</i> (LeConte, 1876)	Punctures scattered, striae absent; striae absent
Doydirhynchini	<i>Doydirhynchus austriacus</i> (Olivier, 1807)	Punctures scattered, striae absent; striae absent
RHINORHYNCHINAE: Rhinorhynchini	<i>Basiliorhinus araucariae</i> Kuschel, 1994	Punctures aligned into striae; striae present
	<i>Nannomacer germaini</i> (Voss, 1952)	Punctures aligned into striae; striae present
	<i>Rhinorhynchus rufulus</i> (Broun, 1880)	Punctures aligned into striae; striae present
	<i>Rhynchitomacer flavus</i> Voss, 1937	Punctures aligned into striae; striae present
Rhinorhynchini?	<i>Burmonyx zigrasi</i> Davis & Engel, sp. n.	Punctures aligned into striae; striae present
Mecomacerini	<i>Aragomacer leai</i> Kuschel, 1994	Punctures aligned into striae; striae present
	<i>Mecomacer scambus</i> Kuschel, 1954	Punctures aligned into striae; striae present
	<i>Brarus mystes</i> Kuschel, 1997	Punctures aligned into striae; striae present
	<i>Rhynchitomacerinus kuscheli</i> (Voss, 1952)	Punctures aligned into striae; striae present
	<i>Rhynchitoplesius eximius</i> (Voss, 1952)	Punctures aligned into striae; striae present
	<i>Eobelus longipes</i> Arnol'di, 1977	Punctures aligned into striae; striae present
EOBELINAE: Eobelini	<i>Archaeorhynchus paradoxopus</i> Arnol'di, 1977	Punctures aligned into striae; striae present
Oxycorynoidini	<i>Oxycorynoides similis</i> Arnol'di, 1977	Punctures aligned into striae; striae present
Brenthorhynchini	<i>Brenthorhynchoides mandibulatus</i> Gratshev & Zherikhin, 1996	Punctures aligned into striae; striae present
Distenorhynchini	<i>Distenorhynchus</i> spp.	Punctures aligned into striae; striae present

the elytral punctures are distinctly aligned into striae and the pretarsal claws are appendiculate, both characters specifying its placement to Rhinorhynchinae. Although the visible features of this specimen may also superficially appear to allow placement within Nemonychinae, the elytral striae in the latter subfamily are much less defined and the elytra contain a scattering of small punctures. In Rhinorhynchinae, all species have well-defined punctate elytral striae (Figs 11–19). According to its rather elongate, narrow rostrum, the fairly large striae punctures of the elytra, as well as the somewhat wide pronotum in relation to the width of the elytra, *Burmonyx* seems to be more closely related to taxa of Rhinorhynchini; however, the nearly bifid pretarsal claws also bear some likeness to those members of Mecomacerini. As the head also appears to be pushed into the pronotum in this specimen or at least concealed in dorso-lateral view by a small portion of the pronotum, it is possible that the compound eyes are not prominent in *Burmonyx*, though this statement is not verifiable due to the obscured head region. Alas, given that much of the head, a large portion of the rostrum and most detail of the mouthparts are obscured in this specimen (not to mention lack of visibility of any internal features), an informed tribal placement within Rhinorhynchinae seems rather implausible.

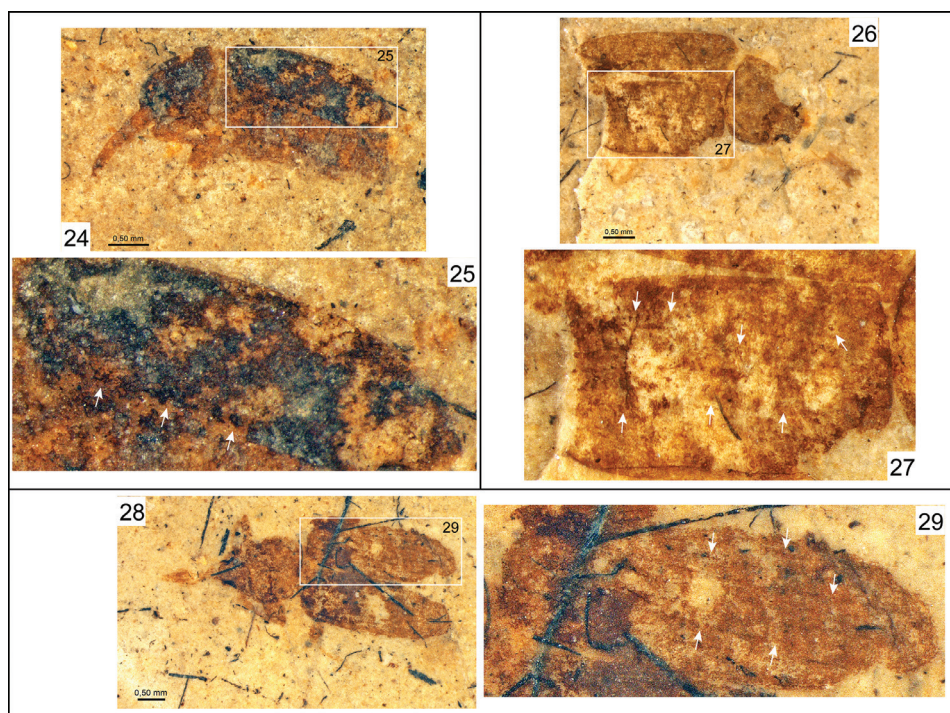


Figures 14–19. Photomicrographs of nemomychid elytra (dorsal aspect). **14** *Rhynchitomacer flavus* Voss, 1937 **15** *Aragomacer leai* Kuschel, 1994 **16** *Mecomacer scambus* Kuschel, 1954 **17** *Brarus mystes* Kuschel, 1997 **18** *Rhynchitomacerinus kuscheli* (Voss, 1952) **19** *Rhynchitoplesius eximius* (Voss, 1952).

The only nemomychids described from amber inclusions thus far include *Kuschelomacer kerneggeri* Riedel (2010), in Baltic amber, and *Libanorhinus succinus* Kuschel and Poinar (1993), in Lebanese amber. Although *Burmonyx* accounts for the first definitive fossil recorded in the subfamily Rhinorhynchinae (and possibly in Rhinorhynchini) from the Asian continent (certainly the first record in Burmese amber), Kuschel and Leschen (2011) transferred *Cratomacer* Zherikhin and Gratshev (2004) from its original placement in Rhinorhynchini (Rhinorhynchinae) to Mecomacerini, which includes two fossil species, *Cratomacer immersus* Zherikhin and Gratshev (2004) and *C. ephippiger* Zherikhin and Gratshev (2004). Fossils of Cimberidinae are now known from the Yixian Formation of China (Davis et al. 2013), and the Karatau site also appears to contain members of this subfamily (e.g., *Chinocimberis* Legalov 2009) and Rhinorhynchinae (personal observation), in addition to the well-known Eobelinae. This growing list also includes the recently described Late Jurassic compression fossil



Figures 20–23. Photomicrographs of fossil nemonychid taxa and their elytra. **20** *Eobelus longipes* Arnol'di, 1977 (holotype), PIN 2452-275 **21** Enlargement of outlined elytron in Figure 20 **22** *Archaeorrhynchus paradoxopus* Arnol'di, 1977 (holotype), PIN 2335-42 **23** Enlargement of outlined elytron in Figure 22.



Figures 24–29. Photomicrographs of fossil nemonychid taxa and their elytra. **24** *Oxycorynoides similis* Arnol'di, 1977 (holotype), PIN 2554-713 **25** Enlargement of outlined elytron in Figure 24 **26** *Brentborrhinoides mandibulatus* Gratshev and Zherikhin, 1996 (holotype), PIN 2239-1508 **27** Enlargement of outlined elytron in Figure 26 **28** *Distenorrhinus* sp., PIN 2239-1547 **29** Enlargement of outlined elytron in Figure 28.

Talbragarus averyi Oberprieler and Oberprieler (2012), which possibly belongs to either Rhinorhynchinae or Eobelinae. While Karatau certainly has yielded an impressive fauna of nemonychid fossils, this diversity remains a complicated assemblage and the taxa have been placed in several disparate groups based on characters of somewhat questionable interpretation and informativeness (e.g., Arnol'di 1977; Gratshev and Legalov 2009; Legalov 2010a, 2010b, 2012). It also appears that the Yixian Formation may contain fossils of Rhinorhynchinae, as represented by *Abrocar* Liu & Ren (2006); Davis et al. 2013). Although the studies thus far have made valorous strides in sorting this particular fossil fauna, more rigorous examinations of characters are required. Just as greater scrutiny is needed for more accurate placement of taxa, it is possible that in many cases, the level of preservation necessary for acute taxonomic identification (for example, beyond the subfamilial or tribal level) simply is absent and should be recognized, therefore further attempts to classify such taxa must be abandoned.

Acknowledgements

Much appreciation is given to James S. Zigras who provided the opportunity to study this specimen and has generally supported amber research at the AMNH. Partial support for this work was provided by the Byers Opportunity Fund, Division of Entomology, University of Kansas Natural History Museum (to SRD), and US National Science Foundation grants DEB-0542909 (to MSE) and DEB-1110590 (to MSE, P. Cartwright, and SRD). This is a contribution of the Division of Entomology, University of Kansas Natural History Museum.

References

- Alonso-Zarazaga MA, Lyal CHC (1999) A World Catalogue of Families and Genera of Curculionoidea (Insecta: Coleoptera) (Excepting Scolytidae and Platypodidae). Entomopraxis, Barcelona, Spain, 315 pp.
- Anderson RS (1995) An evolutionary perspective on diversity in Curculionoidea. *Memoirs of the Entomological Society of Washington* 14: 103–114.
- Arnol'di LV (1977) Rhynchophora. Family Eobelidae. *Trudy Paleontologicheskogo Instituta Akademii Nauk SSSR* 161: 142–176. [In Russian: English translation published as Arnol'di LV, Zherikhin VV, Nikritin LM, Ponomarenko AG. 1991. Mesozoic Coleoptera. Oxonian Press, New Delhi, India, xii+284 pp.]
- Broun T (1880) Manual of the New Zealand Coleoptera. Part I. Colonial Museum and Geological Survey Department, Wellington, New Zealand, xx+651 pp.
- Davis SR, Engel MS, Legalov A, Ren D (2013) Weevils of the Yixian Formation, China (Coleoptera: Curculionoidea): Phylogenetic considerations and comparison with other Mesozoic faunas. *Journal of Systematic Palaeontology* 11(3–4): 399–429.

- Fabricius JC (1801) *Systema eleutheratorum secundum ordines, genera, species: Adiectis synonymis, locis, observationibus, descriptionibus*. Tomus II. Bibliopoli Academici Novi, Kiliae [Kiel], Germany, 687 pp.
- Gratshev VG, Legalov AA (2009) New taxa of the family Nemonychidae (Coleoptera) from the Jurassic and Early Cretaceous. *Euroasian Entomological Journal* 8(4): 411–416. [In Russian]
- Gratshev VG, Zherikhin VV (1996) A revision of the nemonychid weevil subfamily Brenthorhinae (Insecta, Coleoptera: Nemonychidae). *Paleontological Journal* 29(4): 112–127.
- Grimaldi D, Engel MS (2005) *Evolution of the Insects*. Cambridge University Press, Cambridge, United Kingdom, xv+755 pp.
- Grimaldi DA, Engel MS, Nascimbene PC (2002) Fossiliferous Cretaceous amber from Myanmar (Burma): Its rediscovery, biotic diversity, and paleontological significance. *American Museum Novitates* 3361: 1–72. doi: 10.1206/0003-0082(2002)361<0001:FCAFMB>2.0.CO;2
- Kuschel G (1954) La familia Nemonychidae en la Región Neotropical (Aporte 15 de Coleoptera Curculionidae). *Revista Chilena de Historia Natural* 54(9): 97–126.
- Kuschel G (1983) Past and present of the relict family Nemonychidae (Coleoptera, Curculionoidea). *GeoJournal* 7(6): 499–504.
- Kuschel G (1994) Nemonychidae of Australia, New Guinea and New Caledonia. In: Zimmerman EC (Ed) *Australian Weevils: Volume I: Orthoceri, Anthribidae to Attelabidae: The primitive weevils*. CSIRO, Melbourne, Australia, 563–637. [total volume xxxii+741 pp.]
- Kuschel G (1995) A phylogenetic classification of Curculionoidea to families and subfamilies. *Memoirs of the Entomological Society of Washington* 14: 5–33.
- Kuschel G (1997) A new genus and species of Nemonychidae (Coleoptera) associated with *Araucaria angustifolia* in Brazil. *New Zealand Entomologist* 20(1): 15–22. doi: 10.1080/00779962.1997.9722664
- Kuschel G, Leschen RAB (2011) Phylogeny and taxonomy of Rhinorhynchinae (Coleoptera: Nemonychidae). *Invertebrate Systematics* 24(6): 573–615. doi: 10.1071/IS09027
- Kuschel G, Poinar Jr GO (1993) *Libanorhinus succinus* gen. & sp. n. (Coleoptera: Nemonychidae) from Lebanese amber. *Entomologica Scandinavica* 24(2): 143–146. doi: 10.1163/187631293X00253
- LeConte JL (1876) [Untitled sections]. In: LeConte JL, Horn GH. *The Rhynchophora of America North of Mexico*. *Proceedings of the American Philosophical Society* 15(96): 1–455.
- Legalov AA (2010a) Checklist of the Mesozoic Curculionoidea (Coleoptera) with description of new taxa. *Baltic Journal of Coleopterology* 10(1): 71–101.
- Legalov AA (2010b) Phylogeny of the family Nemonychidae (Coleoptera) with descriptions of new taxa. *Euroasian Entomological Journal* 9(3): 457–473.
- Legalov AA (2012) Fossil history of Mesozoic weevils (Coleoptera: Curculionoidea). *Insect Science* 19(6): 683–698. doi: 10.1111/j.1744-7917.2012.01508.x
- Liu M, Ren D (2006) First fossil Eccoptarthridae (Coleoptera: Curculionoidea) from the Mesozoic of China. *Zootaxa* 1176: 59–68.
- Oberprieler RG, Marvaldi AE, Anderson RS (2007) Weevils, weevils, weevils everywhere. *Zootaxa* 1668: 491–520.

- Oberprieler RG, Oberprieler SK (2012) *Talbragarus averyi* gen. et sp. n., the first Jurassic weevil from the Southern Hemisphere (Coleoptera: Curculionoidea: Nemonychidae). *Zootaxa* 3478: 256–266.
- Olivier AG (1807) Entomologie, ou Histoire Naturelle des Insectes, avec leurs caractères généraux et spécifiques, leur description, leur synonymie, et leur figure enluminée. Coléoptères. Tome Cinquieme. Desray, Paris, France, [i]+612 pp.
- Poinar G Jr (2006) *Mesophyletis calhouni* (Mesophyletinae), a new genus, species, and subfamily of early Cretaceous weevils (Coleoptera: Curculionoidea: Eccoptarthridae) in Burmese amber. *Proceedings of the Entomological Society of Washington* 108(4): 878–884.
- Riedel A (2010) A new tribe, genus and species of Nemonychidae from Baltic amber (Coleoptera: Curculionoidea: Nemonychidae: Cimberidinae). *Insect Systematics & Evolution* 41(1): 29–38. doi: 10.1163/139956009X12550095535792
- Ross AJ, Mellish C, York P, Crighton B (2010) Burmese amber. In: Penney D (Ed) *Biodiversity of Fossils in Amber from the Major World Deposits*. Siri Scientific Press; Manchester, UK, 208–235 [total volume 304 pp.]
- Shi G, Grimaldi DA, Harlow GE, Wang J, Wang J, Yang M, Lei W, Li Q, Li X (2012) Age constraint on Burmese amber based on U-Pb dating of zircons. *Cretaceous Research* 37: 155–163. doi: 10.1016/j.cretres.2012.03.014
- Voss E (1937) Über ostasiatische Curculioniden (Col., Curc.). *Senckenbergiana* 19(3–4): 226–282.
- Voss E (1951) [1952] Ueber einige Rhynchitinen der chilenischen Fauna (Coleoptera Curculionidae). (116. Beitrag zur Kenntnis der Curculioniden). *Revista Chilena de Entomología* 1: 175–185.
- Zherikhin VV, Gratshev VG (2004) Fossil curculionoid beetles (Coleoptera, Curculionoidea) from the Lower Cretaceous of northeastern Brazil. *Paleontological Journal* 38(5): 528–537.

Taxonomic study of the planthopper genus *Lacusa* Stål, 1862 (Hemiptera, Fulgoromorpha, Lophopidae)

Jichun Xing^{1,3,†}, Xiangsheng Chen^{1,2,3,‡}

1 Institute of Entomology, Guizhou University, Guiyang, Guizhou, P. R. China, 550025 **2** College of Animal Sciences, Guizhou University, Guiyang, Guizhou, P. R. China, 550025 **3** Special Key Laboratory for Development and Utilization of Insect Resources of Guizhou, Guiyang, Guizhou, P. R. China, 550025

† <http://zoobank.org/1CD0E0C8-D09A-41EE-B9C0-478889BED58A>

‡ <http://zoobank.org/D9953BEB-30E6-464A-86F2-F325EA2E4B7C>

Corresponding author: Xiangsheng Chen (chenxs3218@163.com)

Academic editor: M. Wilson | Received 8 January 2013 | Accepted 15 April 2014 | Published 28 April 2014

<http://zoobank.org/38646370-1FED-4DB6-8261-82774C65FCD3>

Citation: Xing J, Chen X (2014) Taxonomic study of the planthopper genus *Lacusa* Stål, 1862 (Hemiptera, Fulgoromorpha, Lophopidae). ZooKeys 405: 139–148. doi: 10.3897/zookeys.405.6981

Abstract

The planthopper genus *Lacusa* Stål, 1862 is reviewed. All species are illustrated and male genital characters are provided, and including two new species: *L. digitata* **sp. n.** and *L. producta* **sp. n.** from Yunnan Province, China. The species *L. orientalis* Liang, 2000 is removed from this genus based on the frons with median carina and fore femur and tibia flattened but not foliaceous. A key to species is also given. The type specimens of the new species are deposited in the Institute of Entomology, Guizhou University, Guiyang, China (GUGC).

Keywords

Auchenorrhyncha, morphology, taxonomy, distribution, new species

Introduction

The planthopper genus *Lacusa* was established by Stål (1862) for a single species *Elasmoscelis fuscofasciata* Stål, 1854 from India. Later, Atkinson (1886) placed *Cixius eminens* Walker, 1858 as a junior synonym of *Lacusa fuscofasciata* (Stål, 1854). Chou & Huang (see Chou et al. 1985) described a new species *Lacusa yunnanensis* from China. Liang

(1996) proposed *Lacusa yunnanensis* Chou & Huang, 1985 as a junior synonym of *Lacusa fuscofasciata* (Stål, 1854). Liang (2000) placed *Sarebasa* Distant, 1909 as a junior subjective synonym of *Lacusa*, 1862 and proposed the new combination *L. celebris* (Distant, 1909), and described a new species *L. orientalis* from Laos, Vietnam and China. Later, Soulier-Perkins (2001) reinstated *Sarebasa* as a separate genus based on the phylogenetic analysis in Lophopidae.

Here, we follow Soulier-Perkins (2001) in treating *Sarebasa* as a genus distinct from *Lacusa* based on the morphological characteristics for these two genera (see Table 1). The species *Lacusa orientalis* Liang, 2000 indicates that the original figures and description are not belong to the genus *Lacusa* based on the frons with median carina and fore femur and tibia flattened but not foliaceous, and it may be belong to the genera *Sarebasa* or *Acothrura*, and its placement is not treated further here. Consequently, this genus now contains only one species: *L. fuscofasciata*.

In this paper, two new species *Lacusa digitata* sp. n. and *Lacusa producta* sp. n. are described and illustrated from Yunnan Province, China. The type specimens of the new species and other materials examined are deposited in the Institute of Entomology, Guizhou University, Guiyang, China (GUGC). The genus *Lacusa* now contains 3 species, a key is given to separate all species.

Material and methods

Specimens were collected by sweeping net. Dry specimens were used for the description and illustration. External morphology was observed under a stereoscopic microscope and characters were measured with an ocular micrometer. Color pictures for adult habitus were obtained by KEYENCE VHX-1000 system. The genital segments of the examined specimens were macerated in 10% NaOH and drawn from preparations in glycerin jelly using a Leica MZ 12.5 stereomicroscope. Illustrations were scanned with Canon CanoScan LiDE 200 and imported into Adobe Photoshop CS8 for labeling and plate composition. Morphological terminology follows Liang (2000).

Taxonomy

Lacusa Stål

<http://species-id.net/wiki/Lacusa>

Lacusa Stål, 1862: 309; Atkinson 1886: 42; Distant 1906: 323; Muir 1930: 478; Chou et al. 1985: 125; Liang 2000: 283.

Type species. *Elasmoscelis fuscofasciata* Stål, 1854

Description. Body length (from apex of vertex to tip of forewings) less than 10 mm, size medium. Head short, approximately trapezoidal, ratio width of vertex from base to

length in middle line 1.6, narrower than pronotum; vertex broader than long, anterior margin straight in dorsal view and not produced anteriorly beyond proximal margin of eyes, lateral margins with carinate, with an obsolete median longitudinal carina. Frons with lateral carinae, and with sublateral carinae fused apically, and without median carina. Rostrum extending to meso-trochanter, with ratio subapical to apical segment 2.0. Pronotum slightly longer than vertex, anterior margin roundly produced, posterior margin approximately straight, tricarinate on disc, lateral areas curved down. Mesonotum broad, with tricarinate on disc. Forewings broadly round at apex, outer and inner margins nearly parallel, precostal area with many oblique transverse parallel crossveins; hindwings narrower than forewings. Legs moderately long, fore femora and tibiae foliaceous. Hind tibiae with 3 lateral spines and 3–4 rows with more than 70 small spines apically, apical spines of first hind tarsal segment separated by a pad of microsetae.

Head pale luteous, suffused with piceous brown. Eyes black brown. Frons brown and anteclypeus piceous brown. Antennae black brown. Rostrum yellowish brown. Forewings semiopaque, with three broad fuscous transverse band and irregularly piceous spots; hindwings semihyaline. Fore and middle legs blackish brown; hind legs luteous, pad yellowish. Abdomen piceous brown.

Male genitalia. Pygofer short, upper 1/3 very narrow in lateral view, without appendage. Anal tube longer than pygofer, apex forked in caudal view. Genital styles short to long, dorsolateral with a small hook or a finger-like process near posterior margin. Aedeagus with 2 dorsally directed, spinose processes, dorsally directed at medioventral margin; near base of aedeagus on dorsal side or mediolateral of aedeagus with 2 spinose processes or not; base of aedeagus on dorsal side with 1 spinose process or not, and each base laterally of aedeagus with 1 processes or not.

Diagnosis. The genus *Lacusa* resembles *Pitambara* Distant, 1906 and *Sarebasa* Distant, 1909 in having the vertex not produced anteriorly beyond proximal margin of eyes, apical spines of first hind tarsal segment separated by a pad of microsetae, and frontal disc not longitudinally deeply concave, but differs from these genera by the characters noted in Table 1.

Distribution. China, India, Nepal, Burma, Thailand.

Table 1. Differences among *Lacusa* with *Pitambara* and *Sarebasa*.

	<i>Lacusa</i>	<i>Pitambara</i>	<i>Sarebasa</i>
1.Body length (from apex of vertex to tip of forewings, male)	Less than 10mm	Less than 10mm	More than 10mm
2.Length of vertex	Broader than long	Longer than broad	Broader than long
3.Median carina on the frons	Absent	Present	Present even if rudimentary
4.Median anterior margin of pronotum	Regularly rounded	Anteriorly protuberant	Regularly rounded
5.Fore femur and tibia	Foliaceous	Flattened but not foliaceous	Relatively elongate, but not foliaceously dilated
6.Hind tibiae with lateral spines (number)	3	2	3

Key to species (male) of *Lacusa* from China

- 1 Aedeagus stout, its base with 1 spinose process on dorsal side and base laterally without process **2**
- Aedeagus long and curved in the middle, its base without spinose process on dorsal side and each base laterally with 1 process..... ***L. fuscofasciata***
- 2 Mediolateral of aedeagus with 2 long spinose processes.... ***L. producta* sp. n.**
- Near base of aedeagus on dorsal side with 2 short spinose processes.....
..... ***L. digitata* sp. n.**

***Lacusa fuscofasciata* (Stål, 1854)**

http://species-id.net/wiki/Lacusa_fuscofasciata

Figs 1–10

Elasmoscelis fuscofasciata Stål, 1854: 248.

Cixius eminens Walker, 1858: 42 (synonymized by Atkinson 1886: 42).

Lacusa fuscofasciata (Stål, 1854), comb. n. by Stål, 1862: 309; Atkinson 1886: 42; Distant 1906: 324, Fig. 159; Melichar 1915: 355; Liang 2000: 287, Figs 10, 18–22.

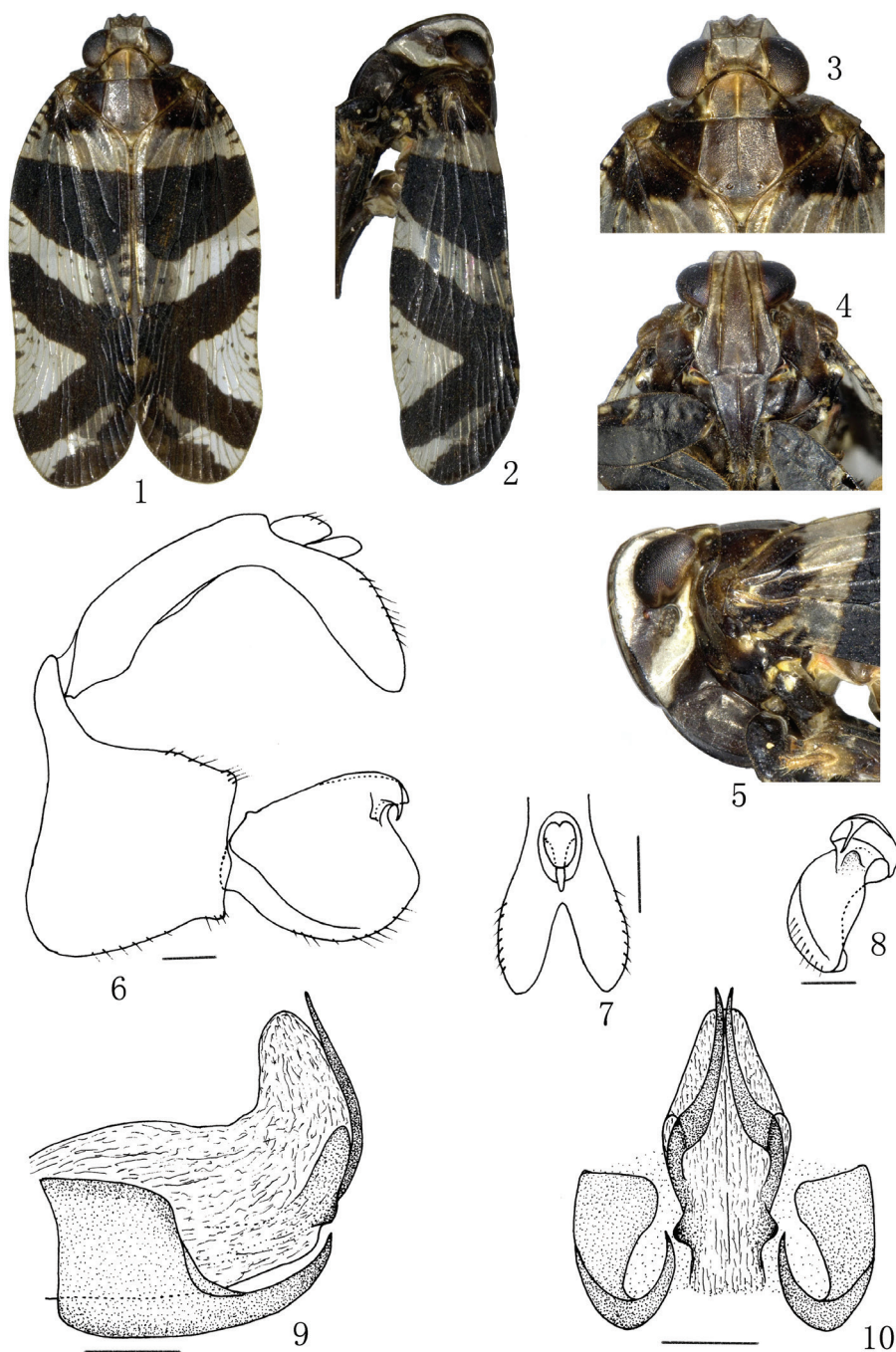
Lacusa yunnanensis Chou & Huang (in Chou et al.) 1985: 128, 137–138, Fig. 119 (synonymized by Liang 1996: 147).

Material examined. 1♂, China: **Guizhou** Province, Jiangkou County, Fanjingshan National Natural Reserve, 24 September 2011, coll. Weibin Zheng (GUGC); 26♂♂14♀♀, China: **Guizhou** Province, Wangmo County, 19 August 2012, coll. Zhimin Chang, Weibin Zheng, Jiankun Long and Weicheng Yang (GUGC); 4♂♂1♀, China: **Guizhou** Province, Ceheng County, 25 August 2012, coll. Zhimin Chang, Weibin Zheng and Jiankun Long (GUGC); 2♂♂, China: **Guizhou** Province, Luodian County, 16 May 2013, coll. Jiankun Long (GUGC); 6♂♂6♀♀, China: **Hainan** Province, Jianfengling National Natural Reserve, 9 April 2013, coll. Jiankun Long, Jichun Xing and Yubo Zhang (GUGC); 1♂, China: **Yunnan** Province, Ruili City, Nongdao, 15 July 2013, coll. Haiyan Sun (GUGC).

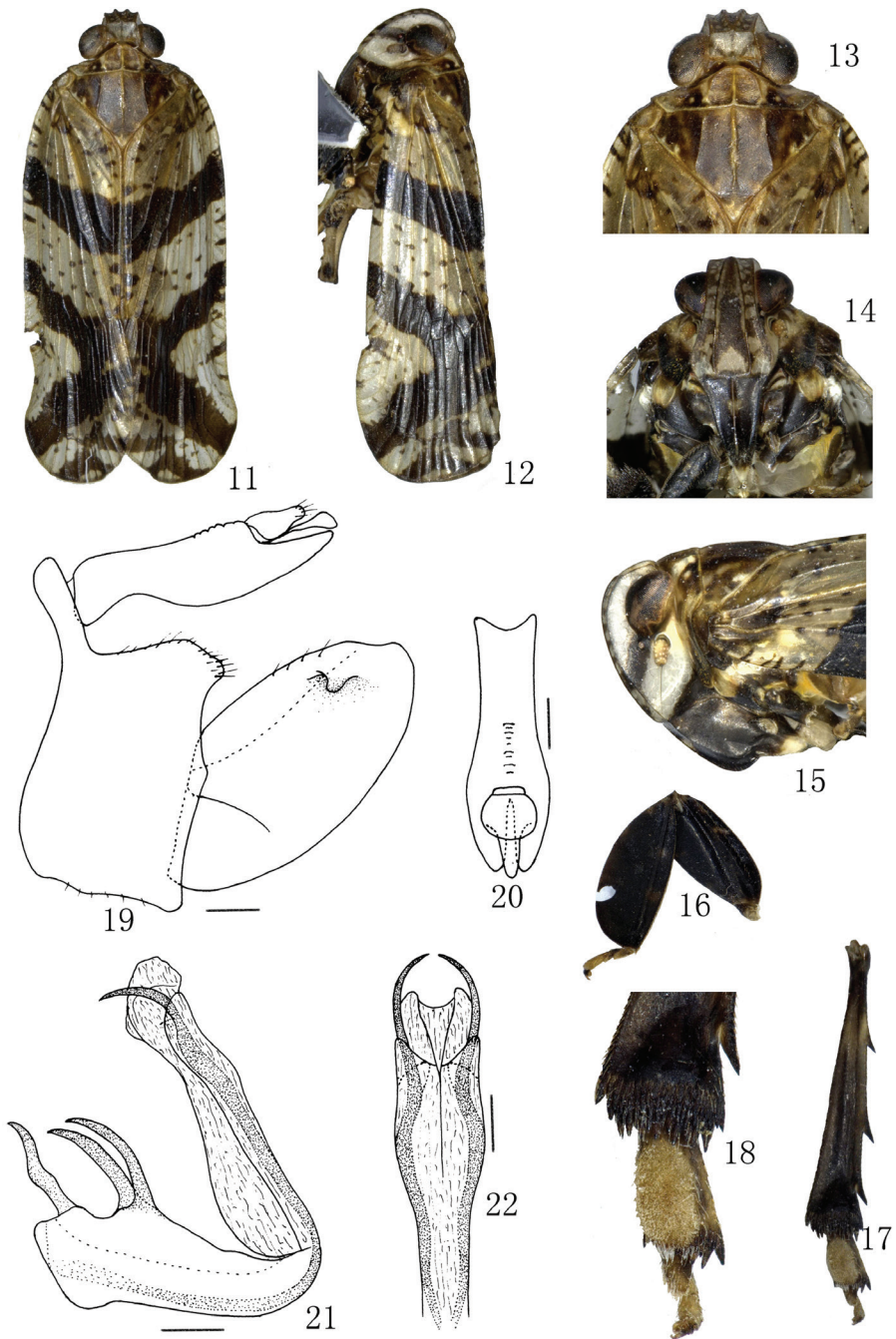
Distribution. China (Yunnan, Guizhou, Guangdong, Hainan), India, Nepal, Burma, Thailand.

Note. Chou & Huang (in Chou et al. 1985) described a new species *L. yunnanensis* from Menglun, Xishuangbanna, Yunnan Province, China, based on one female specimen, and they noted that *L. yunnanensis* was closely related to *L. fuscofasciata*, but that it could be distinguished from it by the fuscous transverse band near outer margin on forewing branched and the fuscous meso- and metathorax. Later, Liang (1996) proposed *L. yunnanensis* as a junior synonym of *L. fuscofasciata* based on examination of specimens from Yunnan, Guizhou, Guangdong and Hainan Provinces in China.

Body length (from apex of vertex to tip of forewings) of male specimen from Guizhou and Hainan Provinces, relatively small about 6.9 mm, but male specimen from Yunnan Province relatively larger about 8.0 mm.



Figures 1–10. *Lacusa fuscifasciata* (Stål, 1854). **1** Male habitus, dorsal view **2** Male habitus, lateral view **3** Male, head and thorax, dorsal view **4** Male, frons and clypeus **5** Male, head and thorax, lateral view **6** Male genitalia, lateral view **7** Apex of anal tube, caudal view **8** Left genital style, caudal view **9** Aedeagus, lateral view **10** Aedeagus, caudal view. Scale bars: **6–10** = 0.20 mm.



Figures 11–22. *Lacusa digitata* sp. n. **11** Male habitus, dorsal view **12** Male habitus, lateral view **13** Male, head and thorax, dorsal view **14** Male, frons and clypeus **15** Male, head and thorax, lateral view **16** Left fore femur and tibia, dorsal view **17** Left hind tibiae and lateral spines, and tarsomere, dorsal view **18** Apex of left hind tibiae and tarsomere, dorsal view **19** Male genitalia, lateral view **20** Anal tube, dorsal view **21** Aedeagus, lateral view **22** Aedeagus, caudal view. Scale bars: 19–22 = 0.20 mm.

***Lacusa digitata* Xing & Chen, sp. n.**

<http://zoobank.org/9CAB07EA-9508-458D-836B-A312AEC90CA1>

http://species-id.net/wiki/Lacusa_digitata

Figs 11–22

Description. Body length (from apex of vertex to tip of forewings): male 8.5–8.7 mm (n=2).

Color pattern of anterior dorsum and face as in Figs 11–15. Pronotum and mesonotum yellowish brown. Tegula yellowish. Ocelli red. Apical margin of forewings maculately piceous. External features as in generic description.

Male genitalia. Pygofer short, upper 1/3 very narrow in lateral view, dorsal margin strongly concave (Fig. 19). Anal tube in dorsal view with ratio length to maximum width 3.0 (Fig. 20). Genital styles long, apex approximately round in lateral view, dorsolateral with a finger-like process near posterior margin (Fig. 19). Aedeagus with 2 dorsally directed, spinose processes, dorsally directed at medioventral margin, and exceed the end of aedeagal shaft; near base of aedeagus on dorsal side of aedeagus with 2 short spinose processes; base of aedeagus on dorsal side with 1 short spinose process (Figs 21, 22).

Type material. Holotype: ♂, China: **Yunnan** Province, Ruili City, Mengla County, Moli, 5 June 2011, coll. Jiankun Long (GUGC); paratypes: 1 ♂, **Yunnan** Province, Lianghe County, Mengyang, 27 July 2013, coll. Zhihua Fan (GUGC).

Diagnosis. This species is similar to *Lacusa fuscofasciata* (Stål, 1854) but can be distinguished by the base of aedeagus on dorsal side with one spinose process, near base of aedeagus on dorsal side with two short spinose processes, the anal tube relatively short, and the genital styles much more long.

Etymology. The species name is derived from the Latin word “*digitata*”, indicating that the genital style dorsolateral with a finger-like process near posterior margin.

***Lacusa producta* Xing & Chen, sp. n.**

<http://zoobank.org/D770CD3E-2AC6-4C93-9A7D-AF6AEE307EA4>

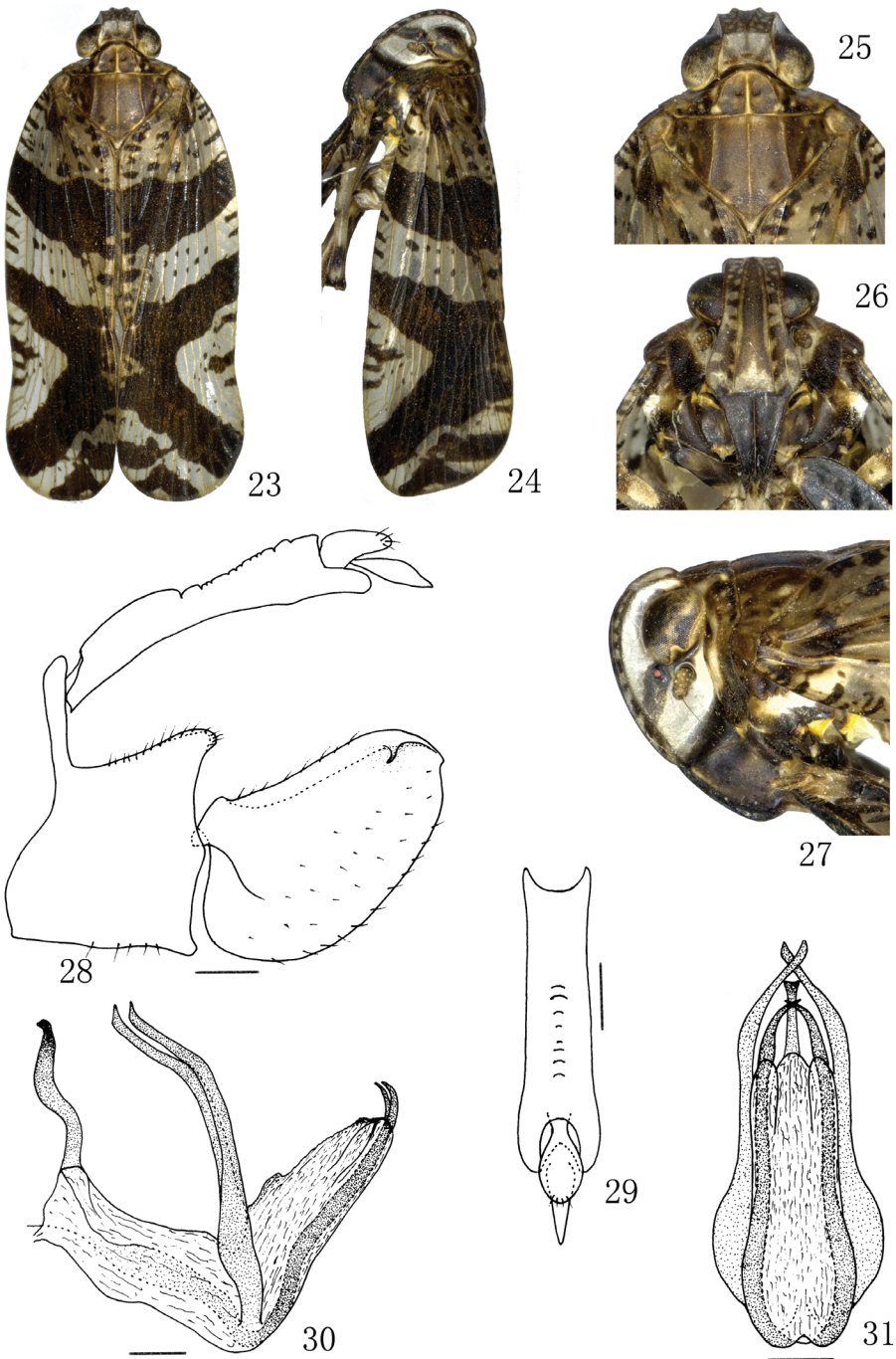
http://species-id.net/wiki/Lacusa_producta

Figs 23–31

Description. Body length (from apex of vertex to tip of forewings): male 9.2–9.3 mm (n=3).

Color pattern of anterior dorsum and face as in Figs 23–27. General appearance as in *Lacusa digitata* sp. n., but the body much larger.

Male genitalia. Pygofer short, upper 1/3 very narrow in lateral view, dorsal margin strongly concave, posterior margin angle produced near the middle; (Fig. 28). Anal tube in dorsal view with ratio length to maximum width 4.1 (Fig. 29). Genital styles long, apex approximately round in lateral view, dorsolateral with a small hook process near posterior margin (Fig. 28). Aedeagus with 2 dorsally directed, spinose processes, dorsally directed at medioventral margin, and exceed the end of aedeagal shaft; mediolateral of aedeagus with 2 long spinose processes; base of aedeagus on dorsal side with 1 spinose process (Figs 30, 31).



Figures 23–31. *Lacusa producta* sp. n. **23** Male habitus, dorsal view **24** Male habitus, lateral view **25** Male, head and thorax, dorsal view **26** Male, frons and clypeus **27** Male, head and thorax, lateral view **28** Male genitalia, lateral view **29** Anal tube, dorsal view **30** Aedeagus, lateral view **31** Aedeagus, caudal view. Scale bars: **28–31** = 0.20 mm.

Type material. Holotype: ♂, China: **Yunnan** Province, Xishuangbanna, Mengla County, Mohan, 25 July 2013, coll. Jichun Xing (GUGC); paratypes: 2♂♂, **Yunnan** Province, Xishuangbanna, Mengla County, Mohan, 25 July 2013, coll. Yuan Liu and Yangyang Liu (GUGC).

Diagnosis. This species is similar to *Lacusa digitata* sp. n. but can be distinguished by the mediolateral of aedeagus with two long spinose processes, genital style dorso-laterally with a small hook process near posterior margin, the anal tube relatively long.

Etymology. The species name is derived from the Latin word “*producta*”, indicating that the mediolateral of aedeagus with two long spinose processes.

Acknowledgements

We are grateful to all collectors for collecting specimens. We also thank Prof. Michael Wilson (Department of Biodiversity & Systematic Biology, National Museum of Wales, Cardiff, UK) and an anonymous referee for reading the manuscript and making some suggestions. This work was supported by the National Natural Science Foundation of China (No.31060290, 31093430) and the International Science and the Program for New Century Excellent Talents in University (NCET-07-0220).

References

- Atkinson ET (1886) Notes on Indian Rhynchota. No. 5. Journal and Proceedings of the Asiatic Society of Bengal 55: 12–83.
- Chou I, Lu JS, Huang J, Wang SZ (1985) *Economic insect fauna of China. Fasc. 36*. Homoptera: Fulgoroidea. Science Press, Beijing, China, 152 pp. [In Chinese with English summary]
- Distant WL (1906) The fauna of British India, including Ceylon and Burma. Rhynchota 3 (Heteroptera-Homoptera). Taylor & Francis, London, 503 pp.
- Distant WL (1909) Rhynchota malayana Part II. Records of the Indian Museum 3: 163–181.
- Liang AP (1996) Taxonomic changes in Chinese Lophopidae with a check list of Chinese species (Homoptera: Fulgoroidea). Pan-Pacific Entomologist 72: 145–151.
- Liang AP (2000) Oriental Lophopidae: new taxa and taxonomic changes (Insecta: Hemiptera: Fulgoroidea). Reichenbachia 33: 281–311.
- Melichar L (1915) Monographie der Lophopinen. Annales Historico Naturales Mus. Nationalis Hungarici 13: 337–384.
- Muir FAG (1930) On the classification of the Fulgoroidea. Annals and Magazine of Natural History (Ser.10) 6: 461–478.
- Soulie-Perkins A (1998) The Lophopidae (Hemiptera: Fulgoromorpha): description of three new genera and key to the genera of the family. European Journal of Entomology 95: 599–618.
- Soulie-Perkins A (2001) The phylogeny of the Lophopidae and the impact of sexual selection and coevolutionary sexual conflict. Cladistics 17: 56–78. doi: 10.1111/j.1096-0031.2001.tb00111.x

Stål C (1854) Nya Hemiptera. Ofversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar 11: 231–255.

Stål C (1862) Novae vel minus cognitae Homopterorum formae et species. Berliner Entomologische Zeitschrift 6: 303–315.