

New species in old mountains: integrative taxonomy reveals ten new species and extensive short-range endemism in *Nesticus* spiders (Araneae, Nesticidae) from the southern Appalachian Mountains

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Abstract

This revision is based on sampling efforts over the past three decades in the southern Appalachian Mountains which have provided *Nesticus* (Araneae, Nesticidae) collections of approximately 2100 adult specimens from more than 475 unique collecting events. Using a “morphology first” framework we examined recently collected specimens plus museum material to formulate morphology-based species hypotheses for putative new taxa (discovery phase). Using sequence capture of nuclear ultraconserved elements (UCEs) we analyzed 801 nuclear loci to validate new (and prior) morphology-based species hypotheses (validation phase) and reconstructed a robust backbone phylogeny including all described and new species. Sanger sequencing and UCE-bycatch were also used to gather mitochondrial data for more than 240 specimens. Based on our integrative taxonomic framework ten new *Nesticus* species are herein described, including *N. binfordae* sp. nov., *N. bondi* sp. nov., *N. canei* sp. nov., *N. cherokeensis* sp. nov., *N. dellingeri* sp. nov., *N. dykemanae* sp. nov., *N. jeminisae* sp. nov., *N. lowderi* sp. nov., *N. roanensis* sp. nov., and *N. templetoni* sp. nov. Previously unknown males are also described for *N. bishopi* Gertsch, 1984, *N. crosbyi* Gertsch, 1984, and *N. silvanus* Gertsch, 1984, as well as the previously unknown female for *N. mimus* Gertsch, 1984. Based on combined evidence *N. cooperi* Gertsch, 1984 is placed in synonymy with *N. reclusus* Gertsch, 1984. Overall, the montane radiation of Appalachian *Nesticus* reveals a general lack of species sympatry and compelling biogeographic patterns. Several regional *Nesticus* taxa are rare, microendemic habitat specialists that deserve conservation attention and detailed future monitoring as conservation sentinels.

Keywords

Cryophilic, invertebrate conservation, mitochondrial introgression, montane speciation, short range endemism, ultraconserved elements

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Introduction

Systematists and evolutionary biologists have long been interested in mountains. Mountains function as habitat islands, serve as refugia in the face of climatic variation, and generate ecological gradients (Wollenberg et al. 2008; Starrett et al. 2018; Rahbek et al. 2019; Perrigo et al. 2020). These combinations of isolation and selective forces act as engines for the origin and persistence of species diversity. In North America, the several physiographic provinces that together comprise the southern Appalachian Mountains represent an ancient and biodiverse region (Stein et al. 2000; Niemiller and Zigler 2013). A combination of climatic variability and long-term habitat availability, in concert with high topographic complexity, has promoted species diversification. For example, endemic radiations of upland arthropod taxa are found in millipedes (Marek and Bond 2006, 2009; Marek 2010; Means et al. 2021; Hennen et al. 2022),

harvestmen (Thomas and Hedin 2008; Hedin and Thomas 2010; Hedin and McCormack 2017; Derkarabetian et al. 2022), spiders (Hendrixson and Bond 2005; Keith and Hedin 2012; Hedin et al. 2015; Newton et al. 2020), and beetles (Sokolov et al. 2004; Caterino and Langton-Myers 2019).

The spider genus *Nesticus* Thorell, 1869 (family Nesticidae) is taxonomically diverse in southern Appalachia, with 28 described species distributed over a geographic area extending from southern West Virginia to central Alabama (Gertsch 1984; Coyle and McGarity 1992; Hedin 1997a; Hedin and Dellinger 2005; Zigler and Milne 2022). Appalachian *Nesticus* are habitat specialists with apparently strict physiological constraints that limit these spiders to dark, cool and moist microhabitats (Fig. 1A–C). Suitable microhabitats include limestone caves (at lower elevations), higher-elevation fissure caves, void spaces in north-facing rock fields, and deep north-facing litter. Appalachian *Nesticus* must be specifically targeted for collecting and are uncommon in general collections. Recent updates to the revisionary work of Gertsch (1984) have focused on cave-dwelling, highly troglomorphic *Nesticus* from the Appalachian Valley and Ridge and Cumberland Plateau geologic provinces. This fauna includes several species known only from single localities (Hedin and Dellinger 2005; Carver et al. 2016; Zigler and Milne 2022), making these taxa susceptible to population decline and perhaps extinction.

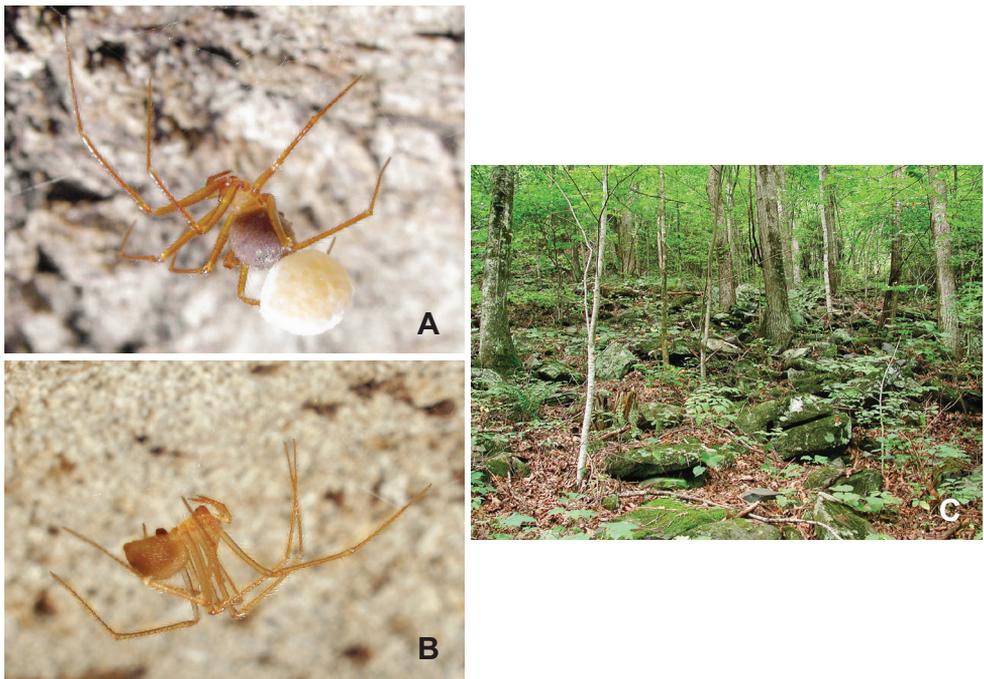


Figure 1. **A** ♀ *Nesticus stupkai* with egg sac, Tennessee, Sevier Co., Wear Cove **B** ♀ *N. barri*, Alabama, Jackson Co., Horseshull Cave **C** montane boulderfield habitat, North Carolina, Macon Co., S of Wayah Bald, MCH 02_169.

This revision focuses more specifically, but not exclusively, on montane Appalachian *Nesticus*, particularly taxa from the mountains of northern Georgia, western North Carolina, northeastern Tennessee, and southwestern Virginia. Both Gertsch (1984) and Coyle and McGarity (1992) described *Nesticus* species from this region but based taxonomic conclusions on small sample sizes often from widely separated geographic locations. For previously described taxa, much denser sampling is needed to fully understand geographic distributions, patterns of geographic variation, and species interactions at geographic boundaries. In addition, because of habitat specificity and high regional topographic complexity, denser geographic sampling is expected to result in the discovery of new montane microendemic species.

In his 1984 revision of North American nesticid spiders, Gertsch (1984) argued that his treatment of Appalachian *Nesticus* was preliminary. In particular, Gertsch predicted that additional geographic and specimen sampling would likely increase regional species diversity and knowledge of geographic distributions, and possibly alter species limits hypotheses. Here we combine morphological data from more than 2100 specimens, original ultraconserved element (UCE) DNA sequence data for 95 specimens, and Sanger / UCE-bycatch mitochondrial data for 241 specimens. Most of these specimens are derived from *Nesticus*-devoted collecting efforts from the past 25 years. We take a “morphology first” taxonomic approach, formulating species hypotheses based on study of male and female genitalia, then independently test these hypotheses using genomic-scale nuclear data bolstered by mitochondrial evidence. Overall, this research reveals a remarkable radiation of short-range endemic (*sensu* Harvey et al. 2011), mostly parapatric *Nesticus* species, including several new species which are rare in suitable habitats and deserve conservation attention.

Materials and methods

Specimen and geographic sampling

We acknowledge that the land upon which we searched for and collected specimens is the traditional and ancestral territories of the Calicuas, Cheraw, Chickasaw, Eno, Kaskaskia, Keyauwee, Lumbee, Manahoac, Miccosukee, Monacan, Moneton, Mvskoke (Muscogee), Myaamia, Occaneechi, Osage, Pee Dee, Saponi, S’atsoyaha (Yuchi), Shakori, Shawandasse Tula (Shawanwaki / Shawnee), Sissipahaw, Skaruhreh / Tuscarora, Sugaree, Tsalaguwetiya (Cherokee), Waxhaw, Yesan (Tutelo), and Yī İsuwā (Catawba) peoples. Most specimens used in this revision were obtained from collections made during the past 25 years by the authors, many collaborators, and prior students of the first author (see Acknowledgements). The types of all previously described taxa were loaned from the American Museum of Natural History (AMNH). Members of field expeditions searched appropriate microhabitats for spiders and collected specimens by hand or using an aspirator. Most spiders were preserved in the field in either 80% or 100% EtOH for subsequent molecular analysis. Molecular samples were later stored in a -80 °C freezer.

Geographic location data were taken in the field using a global positioning system (GPS) device, and later verified/adjusted using ACME Mapper (<https://mapper.acme.com/>). Map figures were generated by importing CVS files into the USGS Survey Map Viewer (<https://maps.usgs.gov/map/>) then adjusting terrain overlay and zoom levels.

We identified immature specimens using the following guidelines: 1) If immatures were collected in association with adults from the same geographic location and in the same microhabitats, these specimens were attributed to the same species, reflecting a very low probability of syntopy (three locations of > 450 unique collecting events; Suppl. material 1). We define syntopy as finding two or more species at the same geographic location (same collecting event), even though we cannot claim that spiders were found in identical microhabitats. 2) If only immatures were collected from a previously published location (generally caves), these specimens were attributed to the known species from this location. 3) Immature spiders from new locations without associated adults were not identified to species (see Suppl. material 2).

Morphology-based species discovery

Some authors have divided species delimitation into a two-step process (Carstens et al. 2013), including a discovery phase (formulating species hypotheses) and a validation phase (formally testing these hypotheses using typically independent evidence). Of course, the validation phase can also be used to test previously formulated hypotheses, i.e., species described by prior authors. We used patterns of morphological variation to formulate putative new species hypotheses. Our general approach was to rely most on patterns of male genitalic variation for a priori species delimitation (as is almost universally applied in araneomorph spider taxonomy, see Bond et al. 2022), assuming that the complex structures of the male nesticid palp best reflect species divergence.

One caveat to our morphology-first approach is that we expect some morphological variation within species, and the distinction between geographic variation vs. species-level divergence is not obvious using only a qualitative approach (e.g., vs. conducting morphometrics and statistical analyses). We expected morphological variation within species because the habitats occupied by these spiders are naturally fragmented (e.g., populations found in caves, isolated mountain ranges, talus fields within mountain ranges, etc.), and the spiders themselves are dispersal-limited. For example, Hedin (1997a) used mitochondrial DNA sequence data to show that gene flow is highly constrained in both cave- and surface-dwelling taxa of the *tennesseensis* group. The combination of natural fragmentation plus dispersal-limitation provides ample opportunity for the evolution of morphological geographic variation. This expected pattern is revealed in the specimens examined here, where genitalic variation is sometimes observed within a single population (e.g., within the confines of a single cave), and variation across populations is common. Yaginuma (1977) discussed similar patterns of challenging intraspecific variation in Japanese nesticids.

This revision shows that female epigynal morphology is generally (but not always) more conserved within groups of closely related taxa. This impacted our revisionary

research because adult males were not always available from all collecting locations. In these cases, our species assignments for female-only locations were less confident, and sometimes relied more heavily on geographic and/or genetic (a posteriori) evidence.

UCE data collection and analysis

To formally test or validate morphological hypotheses using independent character evidence we gathered phylogenomic-scale UCE data and supplemented this nuclear perspective with mitochondrial data. Original UCE data were gathered for 95 specimens representing all but one previously described Appalachian *Nesticus* species and all putative new species (Suppl. material 3). Original data were combined with previously published UCE data for two specimens (see Suppl. material 3). We paid particular attention to genetic sampling from the type localities (or locations nearby) for previously described taxa, and more extensive geographic sampling was conducted for taxa with larger geographic distributions. UCE outgroup data were generated for *Nesticus* species from California and Mexico, and *Nesticella mogera* (Yaginuma, 1972) from Japan. In recent analyses of Ribera and Dimitrov (2023), Appalachian taxa were recovered as monophyletic and sister to taxa from Japan (*Cyclocarcina floronoidea* Kishida, 1942) and South Korea (*Nesticus kyongkeomsanensis* Namkung, 2002); these authors however did not include *Nesticus* taxa from Mexico in their analysis.

Genomic DNA was extracted from leg tissues using the DNeasy Kit (Qiagen). At least 200 ng was sent to RAPID genomics for UCE library prep (Suppl. material 3), where UCES were captured using the spider-specific probe set (Kulkarni et al. 2020). Libraries were sequenced using HiSeq 4000 paired-end 150 bp reads. TRIMMOMATIC v. 0.39 (Bolger et al. 2014) was used to trim adapters and low quality base calls using the following commands: PE ILLUMINACLIP:\$adaptersfasta:2:30:10:2:keepBothReads LEADING:5 TRAILING:15 SLIDINGWINDOW:4:15 MINLEN:40. SPADES v. 3.15.2 (Prjibelski et al. 2020) was then used for assembling clean reads, using the commands: spades.py --sc --careful --cov-cutoff auto. Assembled contigs were imported into the UCE pipeline PHYLUCE 1.6.7 (Faircloth 2016), where the merged arachnid and spider probesets (see Maddison et al. 2020) were matched to contigs using default (80, 80) match values. Sequence alignments were conducted in PHYLUCE using MAFFT (Katoh and Standley 2013) and trimmed using GBLOCKS (Castresana 2000). Matrices with at least 70% occupancy were imported into Geneious Prime 2021.1.1, where alignments were spot-checked.

Using individual UCE loci alignments ($n = 801$) as separate partitions, optimal models were selected using the merging strategy as described in Lanfear et al. (2012), implemented in IQ-TREE 2 (Nguyen et al. 2015). IQ-TREE 2 was also used to reconstruct concatenated maximum likelihood trees and calculate gene (gCF) and site (sCF) concordance factors. For every node of a reference tree, gCF is the percentage of “decisive” gene trees containing that node while sCF is the percentage of decisive sites (in an alignment) supporting a node (Lanfear 2018; Minh et al. 2018).

A species tree was also estimated under a multispecies coalescent model using ASTRAL v. 5.7.8 (Mirarab et al. 2014; Mirarab and Warnow 2015; Rabiee et al. 2019). Input gene trees were estimated using IQ-TREE 2 and treated as unrooted. Internal branch lengths were estimated in coalescent units, with branch support measured as both quartet scores (Sayyari and Mirarab 2016) and local posterior probability values (a function of number of loci and quartet frequencies; Sayyari and Mirarab 2016).

Mitochondrial data collection and analysis

We generated mitochondrial sequences for 218 specimens using standard polymerase chain reaction (PCR) combined with Sanger sequencing. These Sanger data were collected prior to the UCE data (Suppl. material 3). Because there are very few instances of species sympatry as noted above, immature specimens were sometimes used for mitochondrial analysis, but in almost all cases immatures were associated with a sample of adult specimens from the same collecting event. PCR experiments targeted an approximately 1050 bp fragment of the mitochondrial cytochrome c oxidase subunit I (COI) gene region, using C1-J-1510/C1-N-2776 primers and amplification parameters as in Hedin and McCormack (2017). PCR amplification products were sequenced in both directions and sequence contigs were assembled and edited using Sequencher v. 4.2.2.

For specimens for which we had UCE data but no Sanger mitochondrial data, we used BLAST searches in Geneious to recover COI mitochondrial “by-catch” from UCE contigs. As proof of concept, we also included specimens for which we already had Sanger data to confirm the accuracy of the by-catch method and captured mitochondrial data (Suppl. material 3). In all cases by-catch and Sanger sequences from the same specimens were identical (see Results).

Combined Sanger and UCE by-catch mitochondrial data were manually aligned in Geneious. Phylogenetic analysis of the COI matrix was conducted using maximum likelihood searches implemented in IQ-TREE 2. The matrix was partitioned by codon position, with a best-fitting partition scheme (following Lanfear et al. 2012) found by possibly merging partitions (command: `-s -p -m MFP+MERGE`, with ultrafast bootstrap `-B 1000`).

Integrative species delimitation

We defined species under an integrative species delimitation framework as follows: “single populations or sets of populations that share diagnostic male palpal morphologies and that are supported by nuclear phylogenomic monophyly”. This definition includes some necessary caveats. First, our nuclear sample is representative but obviously not exhaustive. We did not generate UCE data for all available sample locations and as such could not formally validate the placement of all specimens / populations. Second, as it was not possible to apply the monophyly criterion for putative species including only a single UCE sample (typically species known only from one location), we here considered long branch lengths, either in a concatenation or coalescent unit framework (long internal branch lengths for the latter). Third, given only qualitative assessments

of variation, the concept of “diagnostic male palpal morphologies” is necessarily subjective, and we allowed for some intraspecific variation (for reasons argued above). The distinction between species level morphological variation vs. geographic variation within a species is not always obvious a priori, and we thus allowed the molecular results (nuclear data in particular) to help guide these decisions.

While mitochondrial evidence was sometimes useful in *Nesticus* species delimitation, we relied most heavily on nuclear gene tree patterns. Conspicuously, mitochondrial data sometimes failed to recover well-supported nuclear lineages, including some well-supported species, likely because of high mitochondrial divergences (see Results). Evidence for mitochondrial introgression and/or deep coalescence of mitochondrial lineages is also apparent within the Appalachian *Nesticus* fauna. Because of the incongruence sometimes observed between hypothesized species (supported by morphology and nuclear data) vs. clades recovered on mitochondrial gene trees, we put less emphasis on the mitochondrial evidence for validating morphological species limits. However, because of larger sample sizes (more geographic populations sampled) and generally higher rates of molecular evolution, the mitochondrial data did provide useful phylogeographic information, and were sometimes used to place some populations for which we only collected female specimens.

Geography also played a secondary role in species delimitation because almost all *Nesticus* species are found in allopatry and typically occupy spatially contiguous geographical distributions that reflect landscape features (e.g., isolated mountain ranges).

Taxonomy

Standard terminology used to describe male and female genitalic morphology follows Coyle and McGarity (1992) and Hedin and Dellinger (2005), as illustrated in Fig. 2A–G. For the medial processes of the paracymbium (Fig. 2D), unless all three are present simultaneously, exact positional homology over distant taxa is uncertain because the relative placement of these processes does appear to evolve (move) across taxa. For example, the definition of dorsomedial vs. distomedial depends upon the relative placement of these two when both are present; if one is lacking then our inference of deeper homology (across species groups) should be viewed as necessarily uncertain. The same argument applies to paradistal vs. dorsal paracymbial processes in some cases. If both processes are not present in a species (or species group), we have observed that the relative positioning of these processes can vary among taxa, challenging our inference of deeper (among species group) positional homology. We emphasize that these issues of positional homology do not impact our species diagnoses, as most diagnoses are restricted to comparisons among relatively closely related taxa in the same species group (as defined below).

With regards to epigynal morphology, the internal fertilization and copulatory ducts are difficult to visualize in *Nesticus* without examination under a compound microscope. Because our drawings and digital images generally do not reveal these details our verbal descriptions similarly emphasize more readily visualized aspects of epigynal morphology.

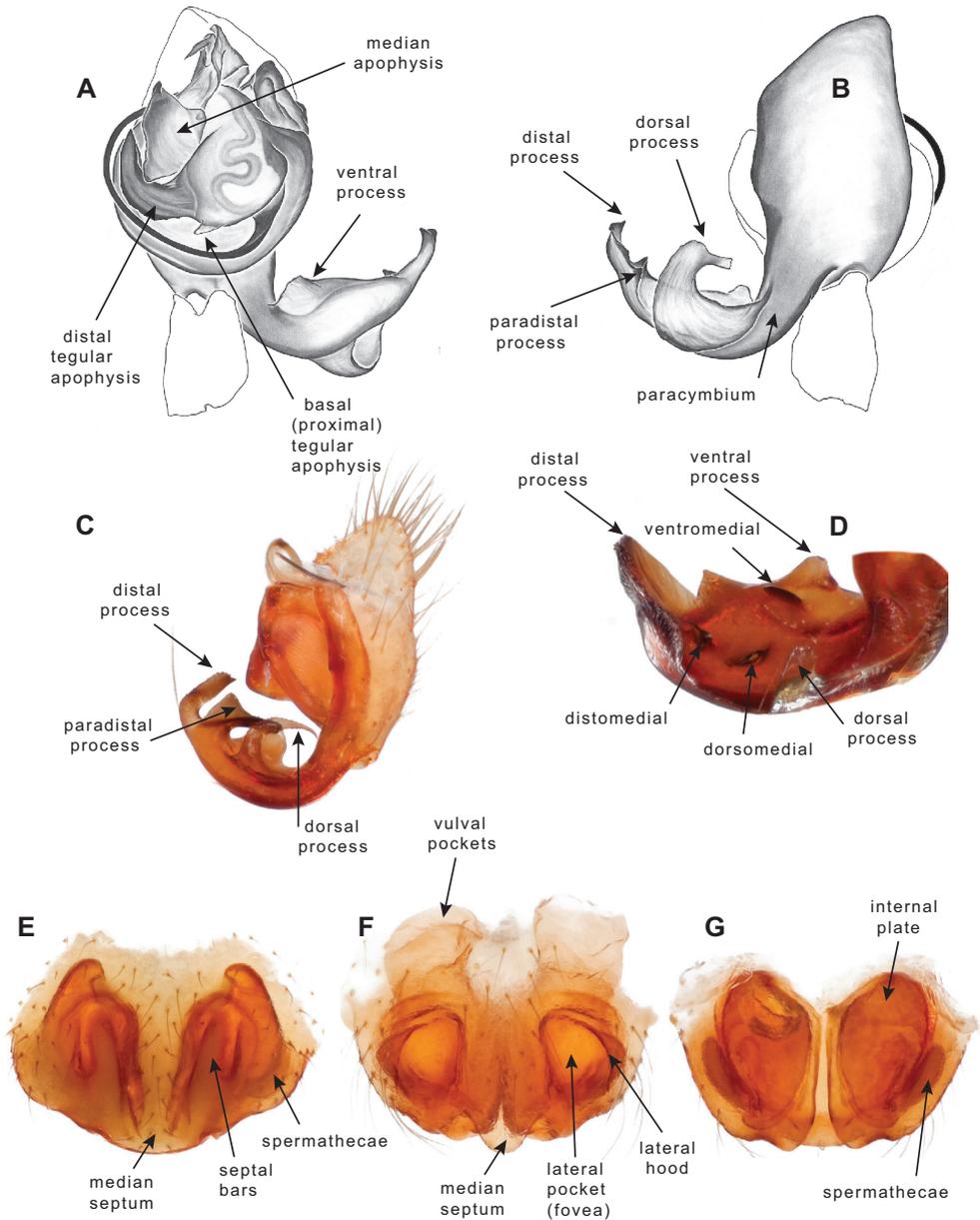


Figure 2. Graphical overview of terminology used to describe genitalic morphology. Tennessee, Overton Co., Obe Lee Cave, *Nesticus stygius*, ♂ MCH specimen #1882 palp ventral (**A**), dorsal (**B**) **C** North Carolina, Haywood Co., near Steestachee Bald overlook, *N. silvanus*, MCH specimen #1145, ♂ palp dorsal **D** North Carolina, Rutherford Co., Moonshiner's Cave, *N. brimleyi*, MCH 99_014, ♂ paracymbium medial **E** North Carolina, Clay Co., Big Tuni Creek, MCH 02_171, epigynum ventral **F** North Carolina, Henderson Co., W of Bat Cave, MCH 07_134, epigynum ventral **G** North Carolina, Buncombe Co., SW of Cane River Gap, MCH 01_167, epigynum dorsal.

Because adult body size, leg lengths, and carapace / abdominal color patterns were found to be variable both within and among populations of the same species (see examples below), we generally do not comment upon this variation in the species descriptions below. Instead, intraspecific variation in male and female genitalia (if present) is emphasized.

Holotype and paratype specimens have been deposited at the Bohart Museum of Entomology (**BME**) at UC Davis. All other specimens referenced with San Diego State University (**SDSU**) or M Hedin (**MCH**) numbers are currently housed in the San Diego State University Terrestrial Arthropod Collection (**SDSU_TAC**).

The following character abbreviations are used in our species descriptions: **BL** = body length (CL plus length of abdomen measured in dorsal view); **CL** = carapace length (from posterior edge to front edge of clypeus, measured at midline); **CW** = maximum carapace width. Lengths of leg I segments are measured in retrolateral view as a straight-line distance from opposite articulation points on the dorsal surface of each segment, reported in Species Descriptions as: Total (fm, pt, ti, mt, ta). All appendage measurements were recorded from the left appendage, unless noted otherwise, and are reported in mm. Measurements at SDSU were taken with an Olympus SZX12 stereomicroscope with 10× ocular lenses fitted with an eyepiece micrometer. All measurements were performed at 2× magnification. Measurements at the University of Indianapolis were taken using a Leica M165C stereomicroscope with an attached DMC2900 camera and calibrated annotation tools within the Leica Application Suite X software (Leica Microsystems, LAS Suite X, v. 3.0.12.21488).

Ink drawings of male and female genitalia were made by Nadine Dupérré. A digital camera attached to a stereomicroscope was used to capture images, which were then enlarged and printed. A tracing of this printed image was detailed and shadowed with repeated reference to the specimen under a microscope. Epigyna were removed and cleared with lactic acid prior to illustration. The left palp of male spiders was illustrated in all cases.

Specimens were digitally imaged at SDSU using a Visionary Digital BK plus system including a Canon 40D digital camera and Infinity Optics Long Distance Microscope. Individual images were combined into a composite image using Zerene Stacker v. 1.04 software; this composite image was then edited using Adobe Photoshop. Epigyna were dissected from specimens using fine forceps, immersed for 2–5 min in BioQuip specimen clearing fluid on a depression slide, then imaged directly in this fluid on the slides. Other images were taken with specimens immersed in filtered 70% EtOH, using KY jelly to secure samples.

Results and discussion

Specimen and geographic sampling

The total morphological sample considered is summarized in Suppl. material 1. This included more than 2100 adult specimens from ~ 480 unique collecting events. We also examined type specimens for all previously described taxa housed at the AMNH.

Morphology-based species discovery

Based on the examination of male and female morphology we hypothesized the following new species *a priori* (discovery phase): two undescribed species in the *tennesseensis* group, two undescribed species in the *nasicus* group, two undescribed species in the *barrowsi* group, and three undescribed species in the *reclusus* group (see group definitions below). We also questioned the species-level status of taxa for two species pairs in the *reclusus* group (*Nesticus stupkai* vs. *N. bishopi*; *N. reclusus* vs. *N. cooperi*). We also noted novel patterns of morphological variation within several described species, which could technically represent new species, but *a priori* treated this as intraspecific variation.

For species delimitation of *Nesticus jemisiae* sp. nov. we did not follow the morphology first framework. Instead, a female of uncertain affinity was first included in a UCE experiment and a long phylogenetic branch was discovered. We subsequently requested that colleagues collect additional specimens from the type locality, and upon inspection, both males and females proved to be morphologically unique.

UCE data and results

We gathered original UCE data for 95 specimens, supplemented with previously published data for two specimens (Suppl. material 3). Specimens from the type locality (or near type locality) were sampled for 27 of 28 previously described species (Suppl. material 3). Raw read data have been submitted to the Sequence Read Archive (BioProject ID PRJNA912717). After processing in PHYLUCE and GENEIOUS data were available for 801 nuclear loci with a total concatenated length of 863,026 base pairs. Input alignments, analysis log files, and output tree files have been included as Suppl. material 4.

Concatenated maximum likelihood and coalescent-based ASTRAL analyses, with very different analytical assumptions, largely agree on overall tree structure (Figs 3, 4). Rooting with *Nesticella mogera* recovers an Appalachian clade sister to a *Nesticus* species from Mexico, both sister to *Nesticus silvestrii* Fage, 1929 from California. We here define seven primary regional species groups within the Appalachian radiation (Figs 3, 4), all strongly supported on both concatenated and ASTRAL trees (Figs 3, 4) and typically diagnosed by morphological synapomorphies. These morphological synapomorphies are discussed in the Taxonomy section below. The seven primary species groups are as follows: *archeri* group, sister to all other Appalachian lineages; the latter clade including the *tennesseensis* group (first recognized by Gertsch 1984) sister to the *nasicus* group (hypothesized by Coyle and McGarity 1992); remaining taxa in a clade including the *barrowsi*, *barri*, *carteri*, and *reclusus* groups.

Mitochondrial data and results

COI sequences were generated for 241 total specimens (Suppl. material 3). For five specimens we gathered duplicate Sanger and UCE by-catch data (always 100% identical), and data for 18 specimens were derived from UCE by-catch only (Suppl. material 3). All mitochondrial sequences have been deposited to GenBank

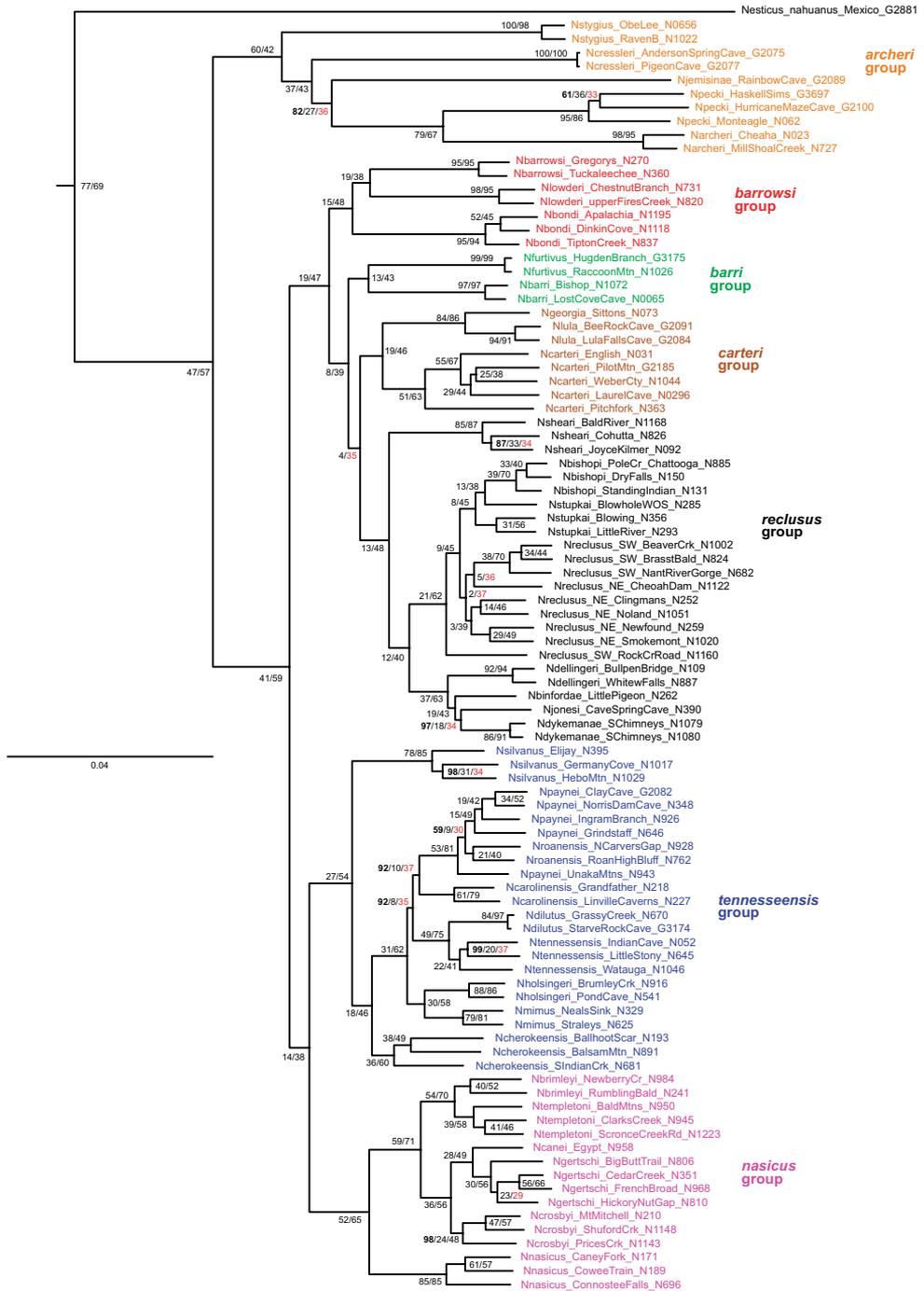


Figure 3. UCE concatenated maximum likelihood tree. Distant outgroups removed (for graphical purposes), specimen numbers correspond to those in Suppl. material 3 (with detailed location provided in Suppl. material 1). Node numbers correspond to bootstrap (bold text) / gCF / sCF. Only bootstrap values below 100 shown, all others 100. gCF and sCF values rounded to nearest integer; sCF values below 38 highlighted with red text.

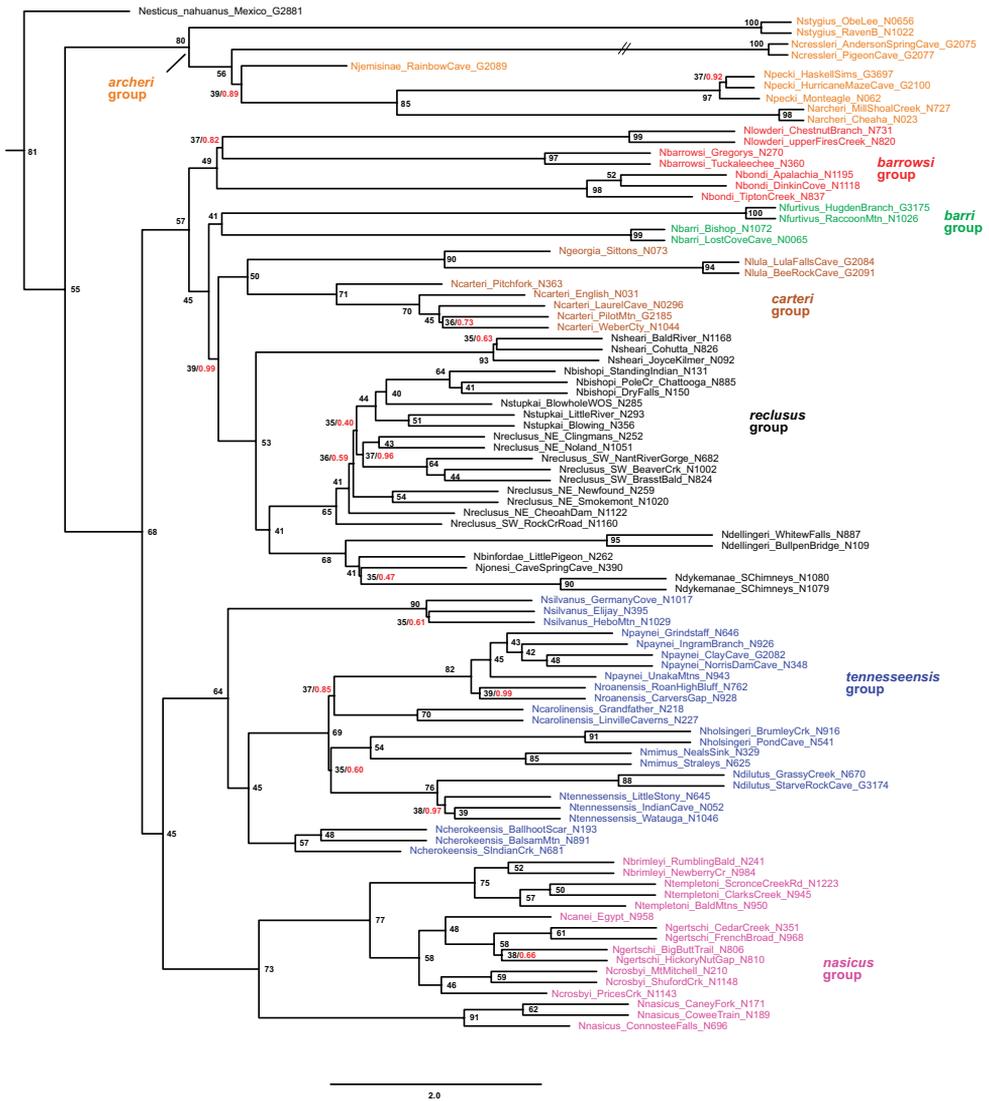


Figure 4. UCE ASTRAL species tree. Distant outgroups removed (for graphical purposes), specimen numbers correspond to those in Suppl. material 3 (with detailed location provided in Suppl. material 1). Included are quartet support values (rounded to nearest integer), and local posterior probability values (if less than 1.0, red text). Branch lengths in coalescent units for internal branches only, terminal branch lengths arbitrary. Internal branch to *Nesticus cressleri* truncated for graphical purposes.

(OQ094967–OQ095207). Input alignments, analysis log files, and output tree files have been included as Suppl. material 4.

Mitochondrial outgroup relationships are as recovered in the UCE data, and the Appalachian fauna is recovered as monophyletic (Fig. 6). The *archeri* group is recovered and identical to nuclear data in internal species relationships resolution. The *tennesseensis* group is also recovered. Beyond this, mitochondrial data failed to recover the remaining

groups. Several otherwise well-supported species are also not recovered on the mitochondrial tree (e.g., *N. carteri*). Multiple mitochondrial sequences are available for most species, and all exhibit high intraspecific mitochondrial divergences, where essentially all sample locations are genetically unique. Additional mitochondrial tree details are discussed in the Taxonomy section below for each species or species group.

Integrative species delimitation

For the nine new species discovered by examination of patterns of male palpal morphology (see above), six are strongly supported (validated) by nuclear gene tree monophyly and associated support metrics (ML bootstrap, concordance factors, ASTRAL quartet scores, and ASTRAL local posterior probabilities; see Fig. 5). One of the nine newly discovered species, *N. roanensis* sp. nov., was weakly supported by phylogenomic evidence and not supported by mitochondrial evidence but was retained as valid (see taxonomic section for this species). Two of the nine species hypotheses could not be formally validated using the monophyly criterion as only a single UCE sample was included.

The nuclear phylogenomic data also mostly strongly supported previously described species. We note that none of these prior hypotheses have ever been tested (validated) using independent data, as is true for almost all described spider species (see Bond et al. 2022). The previously described species pairs (*Nesticus reclusus* vs. *N. cooperi*; *N. stupkai* vs. *N. bishopi*) proved to represent difficult species delimitation scenarios. These challenging cases revealed phylogenetically discordant patterns (e.g., morphological groups conflicting with gene tree clades, mitonuclear discordance, genetic divergence without coincident morphological change) and are discussed more fully in the relevant Taxonomy sections below.

Patterns of genitalic variation across populations within what we considered as single species are summarized in the Taxonomy section for each species. These patterns of variation are based on much larger sample sizes than previously considered, so most are newly reported here.

Niche conservatism and conservation

Below we briefly describe patterns of niche conservatism in Appalachian *Nesticus*, and how this might have impacted the evolution of sympatry, geographic distributions, and species endemism. Future manuscripts will more comprehensively address these issues.

Contrasting with patterns of intraspecific genetic divergence, the Appalachian *Nesticus* fauna is rather conserved in somatic morphology and ecology. This is certainly true for montane species, all of which occur in similar microhabitats and are very similar in their general habitus. Most ecological and somatic morphological evolution is associated with the evolution of cave-dwelling (troglomorphy), involving characters such as body size, leg length, eye reduction, and pigment reduction. We hypothesize that this general niche conservatism has strongly impacted the evolution of sympatry and syntopy. Either because of ecological similarity or reproductive interference (e.g., Ribeiro and Spielman 1986; Goldberg and Lande 2007), syntopy is extremely rare in

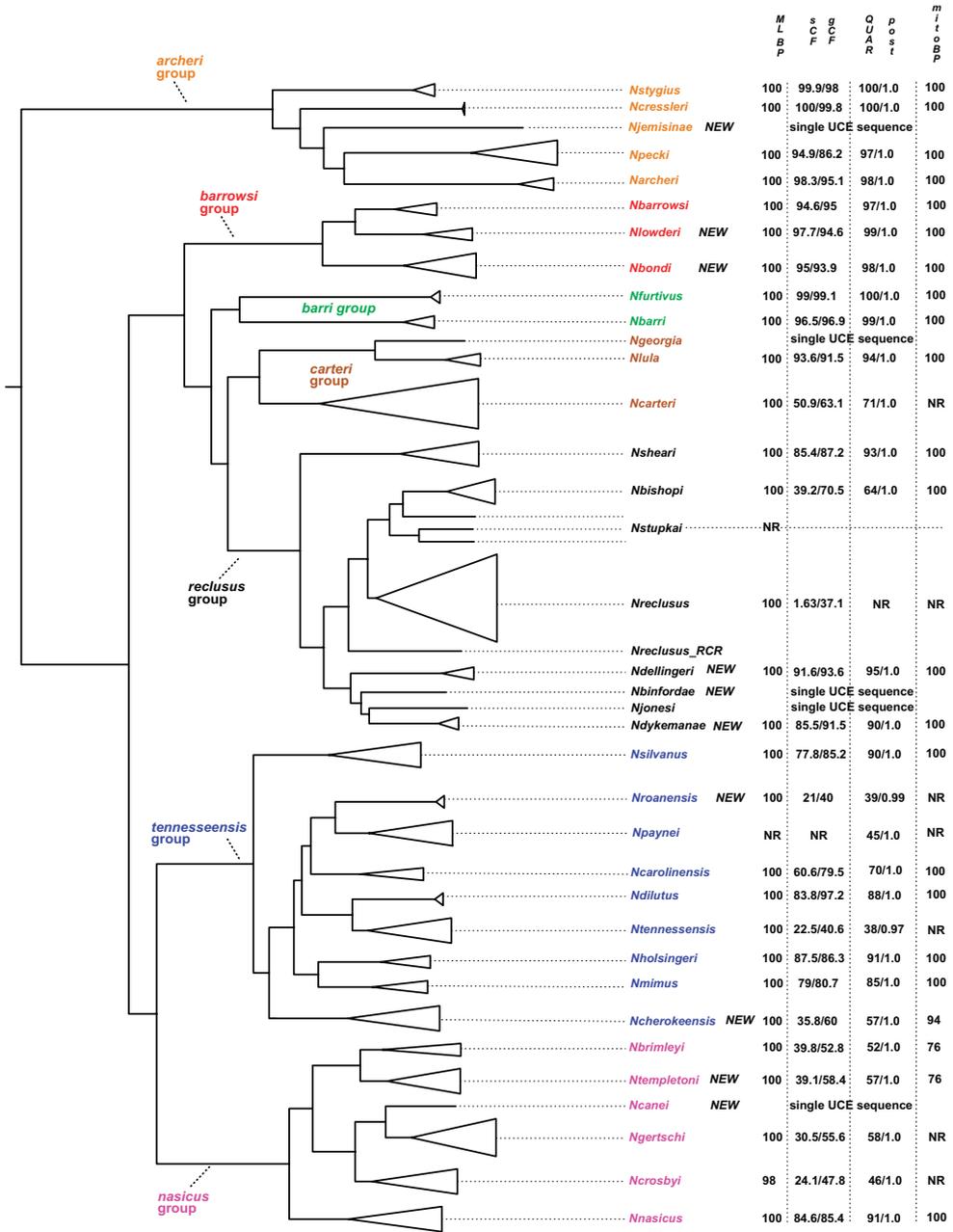


Figure 5. Summary species tree (from UCE topologies), with summary statistics for species support. Branch lengths are arbitrary. New species highlighted. NR = not recovered as clade.

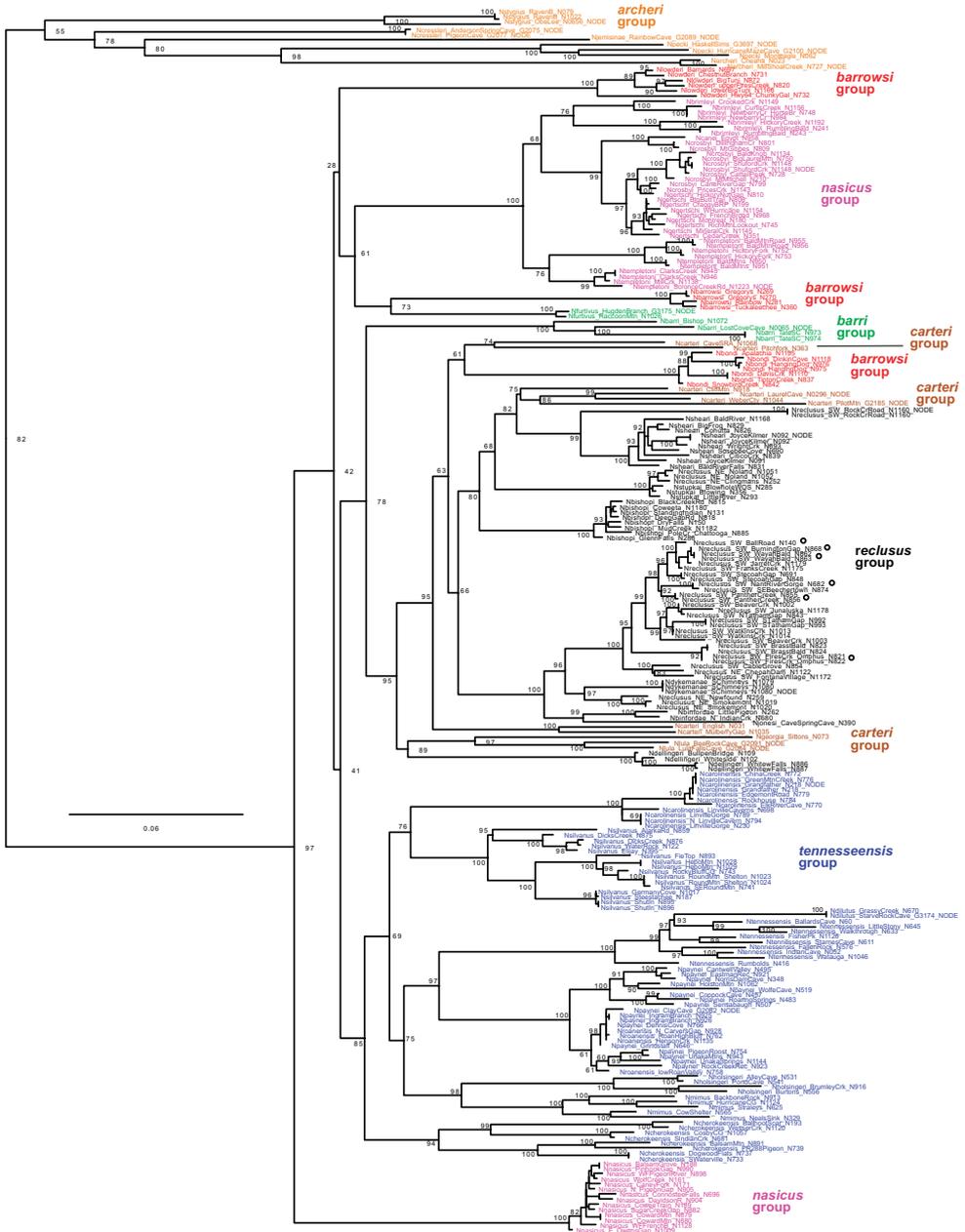


Figure 6. COI IQT gene tree. Distant outgroups trimmed. Specimen numbers correspond to those in Suppl. material 3 (with detailed location provided in Suppl. material 1). Sequences from UCE capture denoted with “NODE”. Populations with a “*Nesticus cooperi*-like” male paracymbium denoted by small circles. Not all tip bootstrap values shown (see Suppl. material 4 for .tree file with all bootstrap values).

the Appalachian radiation. Of 480 unique collecting events, we found members of two different species in syntopy on only three occasions, each time involving relatively distant phylogenetic relatives. These cases are more fully discussed in the Taxonomy section below.

We hypothesize that niche conservatism, and perhaps interactions with competing *Nesticus* species over evolutionary time, has impacted the evolution of endemism in the group. Many species have very small geographic distributions, including three species (*Nesticus canei* sp. nov., *N. jemisinae* sp. nov., *N. jonesi*) known only from single locations, and many others are known only from a handful of geographically adjacent locations. Many of these microendemic species also appear to be naturally rare (at low abundance). These taxa deserve conservation attention and continued conservation monitoring to ensure their long-term persistence.

The Appalachian *Nesticus* radiation has many parallels with the spider genus *Troglohyphantes* Joseph, 1882 (Linyphiidae) from southern Europe, including many taxa in a small area, many microendemic species, habitat and physiological specializations, rare sympatry, and conservation relevance (Isaia et al. 2017). Researchers have begun to understand how climate change will impact the evolution of both cave and montane-restricted species in *Troglohyphantes* (Mammola et al. 2018, 2019), and have developed conservation profiles for several at-risk species (Milano et al. 2022), developing this taxon into a model genus for invertebrate conservation. With the phylogenomic, geographic, and morphological taxonomic framework provided in this revision we argue that *Nesticus* might similarly be developed into a model taxon to understand the impacts of climate (and other abiotic and biotic) change on the specialized cave- and montane-restricted animal fauna of the Appalachian region.

Taxonomy

The taxonomy presented below is structured to follow phylogenomic results, including species groups (treating the *archeri* group first) and order of presentation of species within groups (treating early diverging species first). We do not provide a key to Appalachian species, but rather rely upon the combination of diagnostic morphological features and mostly allopatric geographic distributions to identify specimens. Fig. 7 illustrates the geographic distribution of type localities for all known 37 Appalachian species. Qualitative assessments of conservation status, based on data amassed during the past 25 years, are also emphasized below.

Family Nesticidae Simon, 1894

Genus *Nesticus* Thorell, 1869

archeri group, including:

Nesticus stygius Gertsch, 1984

Nesticus cressleri Zigler & Milne, 2022

Nesticus jemisiae sp. nov.

Nesticus archeri Gertsch, 1984

Nesticus pecki Hedin & Dellinger, 2005

Both UCE and mitochondrial data recover this clade and identical species relationships within this clade (Figs 3, 4, 6). Two eyeless taxa (*Nesticus stygius*, *N. cressleri*) form a paraphyletic grade with respect to three eyed taxa. Species in the group are each morphologically distinctive, subtended by long internal phylogenetic branches with high support values for all metrics and minimal apparent gene tree conflict as measured by concordance factor values (Figs 3–6). Overall, species delimitation within this group is straightforward, likely reflecting a relatively more ancient history (and extinction of intervening lineages) within the group.

Males within this species group possess palps with a forked tegular apophysis, including a larger, darkened distal tegular apophysis that extends underneath the median apophysis and a pointed basal tegular apophysis (Figs 2A, 9A, 11A). Females possess distinctive skinny or banana-shaped spermathecae that extend anteriorly near the outer edges of the epigynum (Figs 10A–C, 11C).

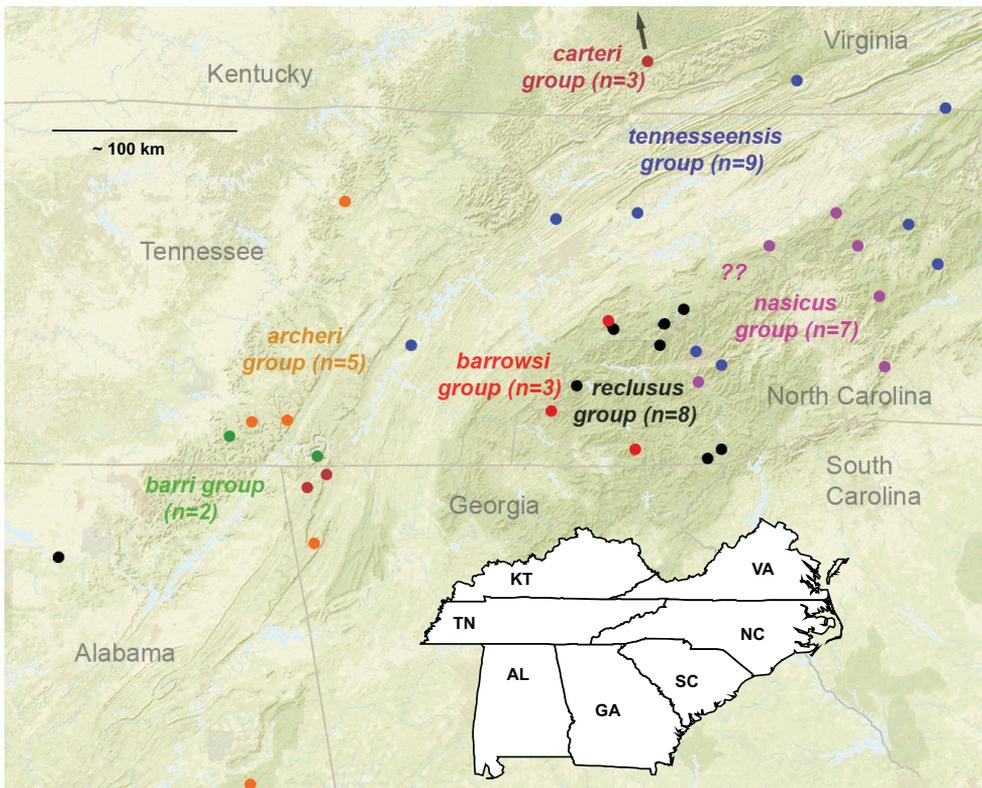


Figure 7. Map of all type localities. Type locality for *Nesticus secretus* is geographically uncertain (see text), designated with a question mark. Inset map shows full map for US states of interest. Scale bar: approximately 100 kilometers.

Species in this species group are distributed in caves on the Cumberland Plateau, and southwards to suitable surface microhabitats at Talladega Mountain in east-central Alabama (Fig. 8). Almost all species appear to have relictual distributions, most species are microendemic, including *Nesticus jemisinae* sp. nov. known only from a single geographic location (Fig. 8). Because of both rarity and microendemism, additional undiscovered populations and species representing this species group are likely. For example, collecting efforts in disjunct “sky island” ridges to the north of Talladega Mountain in east-central Alabama (e.g., Choccolocco Mountain, Weisner Mountain, Bogan Mountain) will likely uncover undescribed *archeri* group species.

Nesticus stygius Gertsch, 1984

Fig. 2A, B

Nesticus stygius Gertsch, 1984: 36, figs 170–172; Hedin and Dellinger 2005: 8, figs 11, 12.

Material examined. New collections from type locality: USA – Tennessee, Overton Co.

• ♂, 3♀; Obe Lee Cave, N of Monterey; 11 Oct. 1993; M. Hedin, C. Phillips leg.; **Non type material:** – Overton Co. • ♂, 1 imm; East Water Supply Cave, TOV15; 13 Jul. 2013; M.L. Niemiller, K.D. Kendall leg.; MLN 13–036; • 7♀; Raven Bluff Cave, NW of Allons; 26 Sep. 1992; M. Hedin, S. O’Kane leg.; • 2♂, ♀; Raven Bluff Cave; 1 Oct. 1991; M. Hedin, K. Crandall, A. Gerber leg.; • ♀; Raven Bluff Cave; 28 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_101; • 4♀, 1 imm; Raven Bluff Cave, TOV28; 3 Sep. 2017; M.L. Niemiller, N. Mann leg.; MLN 17–008.6; • ♀, 1 imm; Webb Cave, TOV39; 1 Oct. 2017; N.S. Gladstone, E.T. Carter, L. Hayter leg.; NSG 17–TOV39.17.

Diagnosis. Morphological diagnosis as in Hedin and Dellinger (2005).

Distribution. This highly troglomorphic taxon is only known from a few caves on the western margin of the Cumberland Plateau in Overton County, north-central Tennessee (Fig. 8; Hedin and Dellinger 2005: fig. 1).

Nesticus cressleri Zigler & Milne, 2022

Nesticus cressleri Zigler & Milne, 2022: 293, figs 1B, D, 4, 5, 7.

Material examined. Type material: USA – Georgia, Walker Co. • ♂, ♀, 1 imm; Anderson Spring Cave (GWK46); 11 Jun. 2014; K.S. Zigler, L. Carver, W.T. Coleman leg.; KSZ 13–159; **Non type material:** – Walker Co. • ♀, 1 imm; Pigeon Cave (GWK57); 3 Aug. 2013; L. Carver, A. Cressler, K.S. Zigler leg.; KSZ 13–184.

Diagnosis. Morphological diagnosis as in Zigler and Milne (2022).

Distribution and natural history. This troglomorphic taxon is known only from three geographically adjacent caves on Pigeon Mountain in Walker County, Georgia (Fig. 8; Zigler and Milne 2022: fig. 7).

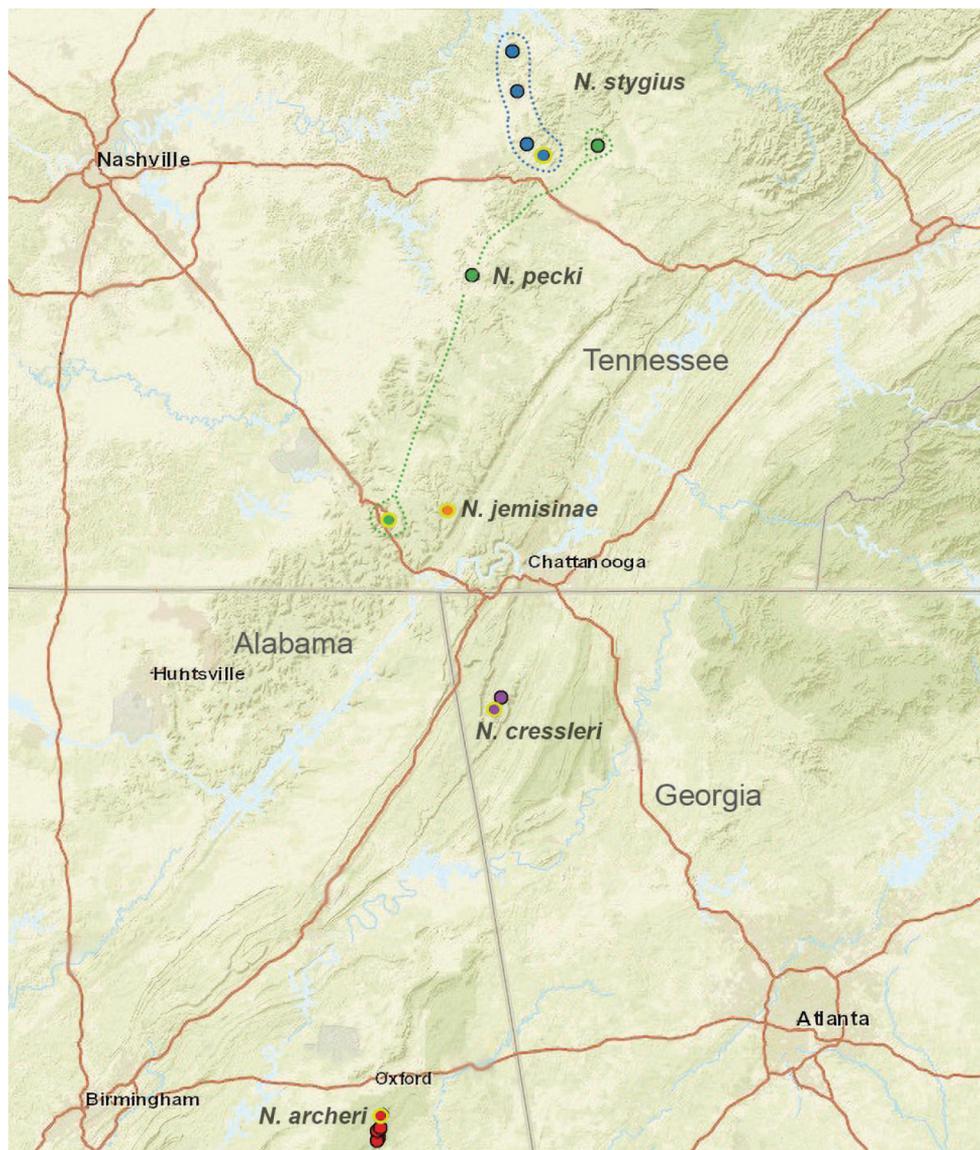


Figure 8. Distribution of the *archeri* group, including *Nesticus archeri*, *N. pecki*, *N. jemininae*, *N. cressleri*, and *N. stygius*. State boundaries and major cities shown for geographic context. Dashed lines circumscribe the distributions of *N. pecki* and *N. stygius*.

***Nesticus jemininae* sp. nov.**

<https://zoobank.org/EC278B87-636E-41F5-AA3E-59599C83AB18>

Fig. 9A–D

Material examined. Type material: *Holotype*: USA – Tennessee, Marion Co.

• ♂ holotype; Rainbow Cave (TMN20); 20 Oct. 2021; K.S. Zigler leg.; SDSU_

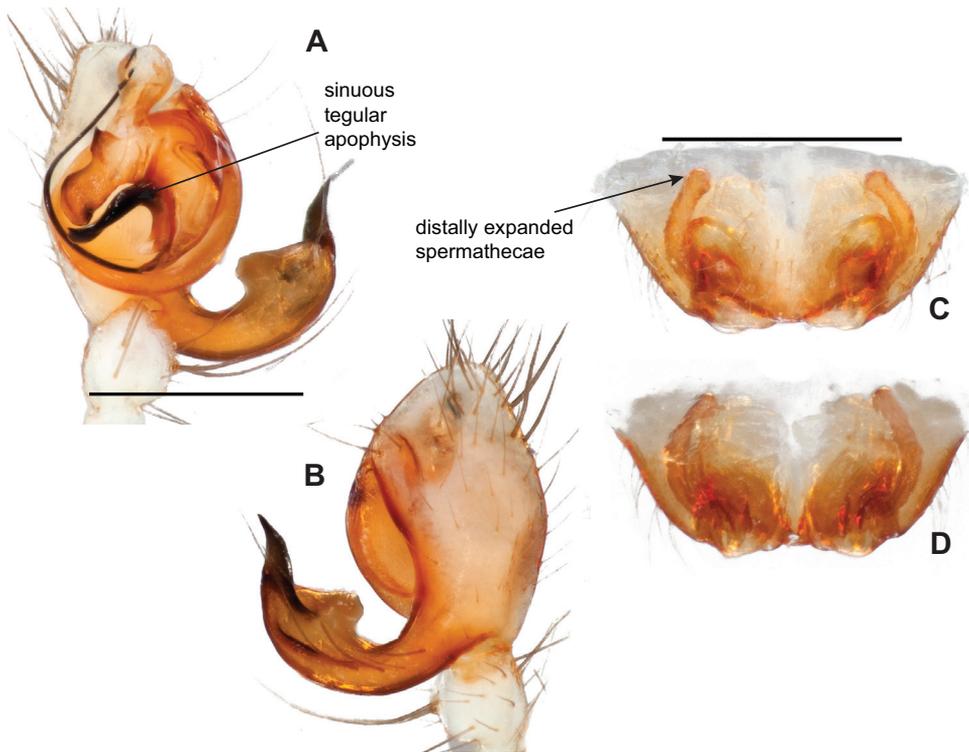


Figure 9. *Nesticus jemisinae* sp. nov. Tennessee, Marion Co., Rainbow Cave, ♂ holotype, SDSU_TAC000662, ventral (A), dorsal (B). Rainbow Cave, ♀ paratype, SDSU_TAC000663, ventral (C), dorsal (D). Scale bar: 0.5 mm.

TAC000662; **Paratypes:** • ♀ paratype; data as for holotype; SDSU_TAC000663; • ♂, 2♀ paratypes; data as for holotype; SDSU_TAC000664; **Non type material:** – **Marion Co.** • ♀; Rainbow Cave (TMN20); 10 Nov. 2013; K.S. Zigler leg.; KSZ 14–248. • 5 imm; Rainbow Cave (TMN20); 20 Oct. 2021; K.S. Zigler leg.

Diagnosis. Easily distinguished from other members of the *archeri* group. Distinctly small-bodied *Nesticus jemisinae* possesses well-developed eyes, different from the eyeless *N. cressleri* and *N. stygius*. Males possess a relatively simple paracymbium, contrasting with the multiple apophyses of the complex paracymbium of *N. archeri* (Fig. 10A) and *N. pecki* (Hedin and Dellinger 2005: figs 17, 18). The distinctive tegular apophysis is dark and sinuous, extending under the median apophysis. Female *N. jemisinae* may also be distinguished from the latter species by the epigynum. The epigynum of *N. archeri* is subtriangular with large anterior fovea and a narrow median septum (Fig. 10B, C), *N. pecki* possesses a broad posteriorly-broadening median septum (Fig. 11A–C), and *N. jemisinae* possesses a posteriorly-pointed median septum with the spermathecae expanded into small bulbs distally (Fig. 9C, D).

Description of ♂ holotype (SDSU_TAC000662; Fig. 9A, B). Carapace and appendages are a dusky yellow. Abdomen mottled gray with darker patches between lighter

parts. Eyes ringed with black and equally well-developed except for AME, which are significantly reduced. Carapace 1.18 long, 1.09 wide. Total body length 2.43. Leg I total length 9.50 (2.74, 0.46, 2.73, 2.56, 1.01), leg I / CW ratio 8.72, leg formula 1423. Paracymbium of palp relatively simple with a large proximally directed ventral process and a dark sinuous distal process. Dorsal process of paracymbium largely reduced to a shallow pocket on the distal edge. Palp tegular apophysis dark, long, narrow, pointed, and extends under median apophysis. Median apophysis elongated towards base of palp and angled proximally at tip.

♂ **Variation.** No significant genitalic variation was noted in the material examined.

Description of ♀ paratype (SDSU_TAC000663; Fig. 9C, D). Color of carapace, appendages, and abdomen as in male. Eyes as in male. CL 1.20, CW 0.97. Total body length 2.61. Leg I total length 7.96 (2.30, 0.44, 2.29, 1.98, 0.95), leg I / CW ratio 8.21, leg formula 1423. Epigynum width approximately half the width of the abdomen. Median septum pointed posteriorly and flanked by fovea along posterior margin. Internal foveal pockets visible from ventral inspection without dissection extending anteriorly angled outwards. Thin, elongate spermathecae curve slightly on outside margins of epigynum, extending anteriorly beyond foveal pockets, expanded into small bulbs distally.

♀ **Variation.** No significant genitalic variation was noted in the material examined.

Distribution and natural history. Known only from Rainbow Cave, located near Pocket Creek, a tributary to the Little Sequatchie River (Fig. 8). This cave is approximately 200 meters in length; spiders were collected ~ 50 meters from the cave entrance on the cave ceiling and walls, in total darkness.

Etymology. The specific name is a matronym in honor of N. K. Jemisin whose 'Broken Earth' book series features a subterranean colony, including scientists who study caves.

Remarks. *Nesticus jemisinae* sp. nov. is a relictual, single-site endemic whose morphology is quite distinct from that of other members of the species group. This species is nested within a diverged nuclear and mitochondrial subclade of the *archeri* group, sister to *N. pecki* and *N. archeri* (Figs 3, 4, 6).

Nesticus archeri Gertsch, 1984

Fig. 10A–C

Nesticus archeri Gertsch, 1984: 32, figs 115–117, 129–131.

Material examined. Type material: Holotype: USA – Alabama • 1♂; Mt. Cheaha, Cheaha State Park; 21 Apr. 1947; A.F. Archer leg; AMNH. **Non type material:** USA – Alabama, **Clay Co.** • 2♂, 3♀; Cheaha State Park, 0.5 mi N Hernandez Peak, along Pinhoti Trail; 33.4645°N, -85.8113°W; 26 Mar. 1995; M. Hedin, B. Dellinger leg.; • 2♂, 2♀; Cheaha State Park, N side of McDill Point; 33.4547°N, -85.8205°W; 26 Mar. 1995; M. Hedin, B. Dellinger leg.; • 2♂, 4♀; Cheaha State Park, Pinhoti Trail, 0.5 mi. S Hernandez Peak; 33.4537°N, -85.8144°W; 26 Mar. 1995; M. Hedin, B. Dellinger leg.; • ♂, 3♀; Talladega Mountain, Cheaha Wilderness, Nubbin Creek Trail,

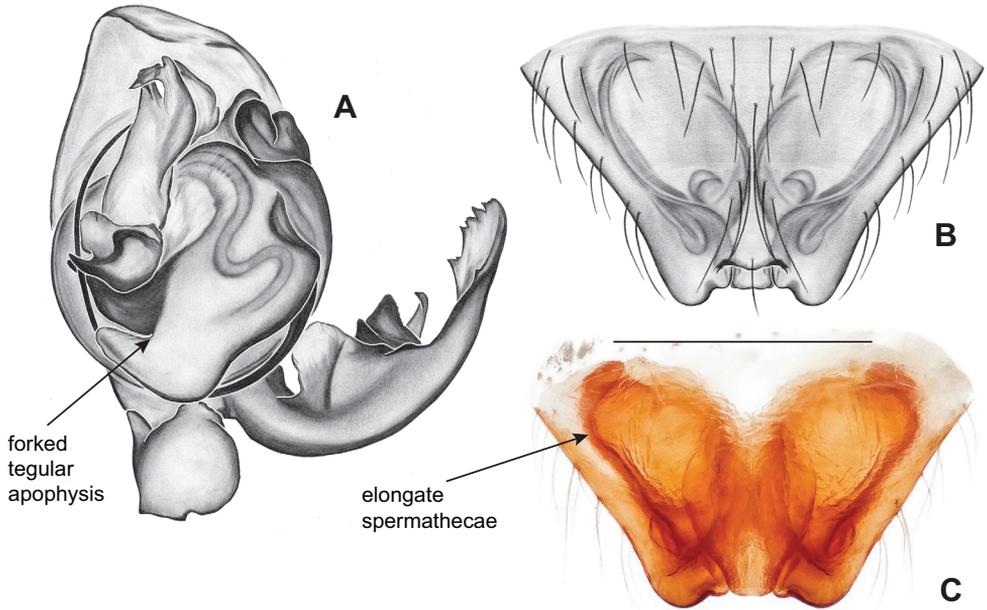


Figure 10. *Nesticus archeri* **A** Alabama, Clay Co., vicinity Mill Shoal Creek, ♂ specimen MCH #2132, ventral palp **B** Alabama, Clay Co., vicinity Mill Shoal Creek, ♀ specimen MCH #2129, epigynum, ventral view **C** dorsal view. Scale bar: 0.5 mm.

vicinity Mill Shoal Creek; 33.4269°N, -85.8177°W; 26 Mar. 1995; M. Hedin, B. Dellinger leg.; • ♂, 3♀; Talladega Mountain, Talladega National Forest, near headwaters of Cave Creek; 33.4344°N, -85.8128°W; 26 Mar. 1995; M. Hedin, B. Dellinger leg.; – **Cleburne Co.** • 1 imm; Cheaha State Park, just north of Bald Rock; 33.4966°N, -85.8075°W; 27 Sep. 1991; M. Hedin, K. Crandall leg.; • 4♂, 13♀; Cheaha State Park, just north of Bald Rock; 33.4966°N, -85.8075°W; 24 Sep. 1992; M. Hedin, S. O’Kane leg.; • 2♂, 2♀; Cheaha State Park, vicinity Bald Rock saddle, 0.5 mi NE Bald Rock; 33.4985°N, -85.8028°W; 25 Mar. 1995; M. Hedin, B. Dellinger leg.

Diagnosis. The only *Nesticus* species with a surface-dwelling habitus (small-bodied, darkly pigmented, well-developed eyes) in the region, and the only known *Nesticus* from Talladega Mountain. Male palp with a forked tegular apophysis, distal (highly sclerotized) fork lying behind pointed basal part of median apophysis in ventral view (Fig. 10A.). Paracymbium also distinctive, with prominent, sclerotized paradistal process. Very distinctive truncate, heart-shaped epigynum with a narrow median septum, large lateral pockets, and elongate spermathecae that hug the outer edges of the epigynal pockets, curving inwards anteriorly (Fig. 10B, C).

Variation. No significant male or female genitalic variation was noted in the material examined.

Distribution and natural history. All known records are from high elevation habitats (most above 600 m, but as low as 430 m) on Talladega Mountain, east-central Alabama (Fig. 8). This species was previously known only from type material

collected at an unspecified location within Cheaha State Park, at the northern end of Talladega Mountain. New collections suggest that *Nesticus archeri* can be found at several places on Talladega Mountain, spanning an approximately 10² kilometer area from near Bald Rock (northern side of Cheaha Mountain) in the north to Mill Shoal Creek in the south.

This species has been collected in dark, cool, relatively moist near-surface habitats. For example, field notes from 1992 collections north of Bald Rock indicate that spiders were collected from “below bluffs on a steep hillside”, in “talus with a heavy leaf litter cover”, where spiders were “most abundant under large rocks close to the surface”. This situation compares favorably with the original “heavy talus of ravine” collections made by A.F. Archer, and the 1995 Hedin and Dellinger collections, most of which were made in north-facing talus, although at least one collection was from southwest-facing talus. This species is perhaps not as uncommon as previously believed and (in the late 1990s) was found consistently in suitable microhabitats.

Remarks. We view Gertsch’s drawing of the male conductor (fig. 129) as inaccurate (compare to Fig. 10A).

Nesticus pecki Hedin & Dellinger, 2005

Fig. 11A–C

Nesticus pecki Hedin & Dellinger, 2005: 14, figs 17–20.

Material examined. Type material: Holotype: USA – Tennessee, Marion Co. • ♂ holotype; Monteagle Saltpeter Cave, ~ 6.4 km SE of Monteagle; 26 Sep. 1992; M. Hedin, J. Hedin, S. O’Kane leg.; MCH1624; • ♀ paratype; data as for holotype; MCH1625; • 2♀; Monteagle Saltpeter Cave; 29 Sep. 1991; M. Hedin, K. Crandall, A. Gerber leg.; MCH1012, MCH1013; **Non type material:** – Fentress Co. • ♀; Hurricane Maze Cave (TFE331); 31 Aug. 2013; M.L. Niemiller, G. Moni, K. Bobo, A. Crabtree, B. Reeves, K. Pasternak leg.; MLN 13–063; – White Co. • 8♀; Haskell Sims Cave; 18 Jan. 2014; M.L. Niemiller, E.T. Carter, G. Moni, C. Sutherland leg.; MLN 14–004.

Diagnosis. Morphological diagnosis as summarized in Hedin and Dellinger (2005), spiders small-bodied with well-developed eyes, males with a thickened and chisel-like regular apophysis, females with a posteriorly-broadened median septum (Fig. 11A–C).

Variation. The Hurricane Maze Cave specimen is similar to specimens from the type locality, possessing a short and wide epigynum with a posteriorly flaring median septum and banana-shaped spermathecae with narrow bases, lying just lateral to fovea but inside of the sclerotized epigynal outline (Fig. 11A, B). Haskell Sims Cave specimens are fairly different, the posterior edge of the median septum rounded instead of flared, and with internal plates close to touching (Fig. 11C). Given this relatively divergent female morphology it would be useful to attempt to collect adult males from this northern disjunct cave location (Fig. 8).

Distribution and natural history. Previously known only from the type locality in southeastern Tennessee (Hedin and Dellinger 2005, fig. 1). We report here new

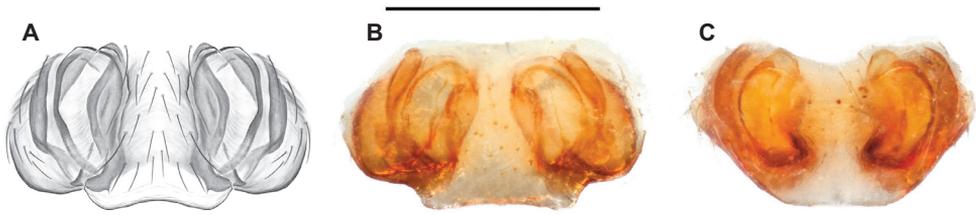


Figure 11. *Nesticus pecki* epigynum variation, ventral views **A** Tennessee, Marion Co., Monteagle Saltpeter Cave, specimen #1625 **B** Tennessee, Fentress Co., Hurricane Maze Cave, MLN 13–063 (SDSU_G2100) **C** Tennessee, White Co., Haskell Sims Cave, MLN 14–004 (SDSU_G3697). Scale bar: 0.5 mm.

important northern records from Haskell Sims and Hurricane Maze Caves, extending the geographic distribution of this species significantly northwards (Fig. 8, Suppl. material 1). More collecting on the Cumberland Plateau will likely result in additional new distributional records, although the species appears to be naturally rare. Hedin and Dellinger (2005) and Carver et al. (2016) reported on the rarity of this species at the type locality.

***tennesseensis* group, including:**

Nesticus silvanus Gertsch, 1984

Nesticus cherokeensis sp. nov.

Nesticus holsingeri Gertsch, 1984

Nesticus mimus Gertsch, 1984

Nesticus tennesseensis (Petrunkevitch, 1925)

Nesticus dilutus Gertsch, 1984

Nesticus carolinensis (Bishop, 1950)

Nesticus paynei Gertsch, 1984

Nesticus roanensis sp. nov.

This species group is recovered as monophyletic with both nuclear (Figs 3, 4) and mitochondrial data (Fig. 6), but lacks strong bootstrap support in the latter. Species relationships within this group are consistent and well supported for concatenated and coalescent UCE analyses (Figs 3, 4), and our presentation below follows this (accepted) phylogenomic structuring. Mitochondrial relationships among taxa within the group do not reflect UCE results, with many relatively low bootstrap support values (below 90) along the phylogenetic backbone; following arguments made in the Materials and methods we defer here to the nuclear phylogenomic results.

Male and female genital morphology suggests common ancestry for this complex of nine species, also defined as a species group by Gertsch (originally not including *Nesticus cherokeensis* sp. nov. or *N. roanensis* sp. nov.). Males of this species group include palps characterized by a translucent dorsal process of the paracymbium, projecting anteriorly then medially, with a thin medial projection with fine anterior serrations (Fig. 12A–G). Viewed dorsally, the dorsal process lies above a rectangular, dark paradistal process which itself lies above a narrower distal process with fine serrations

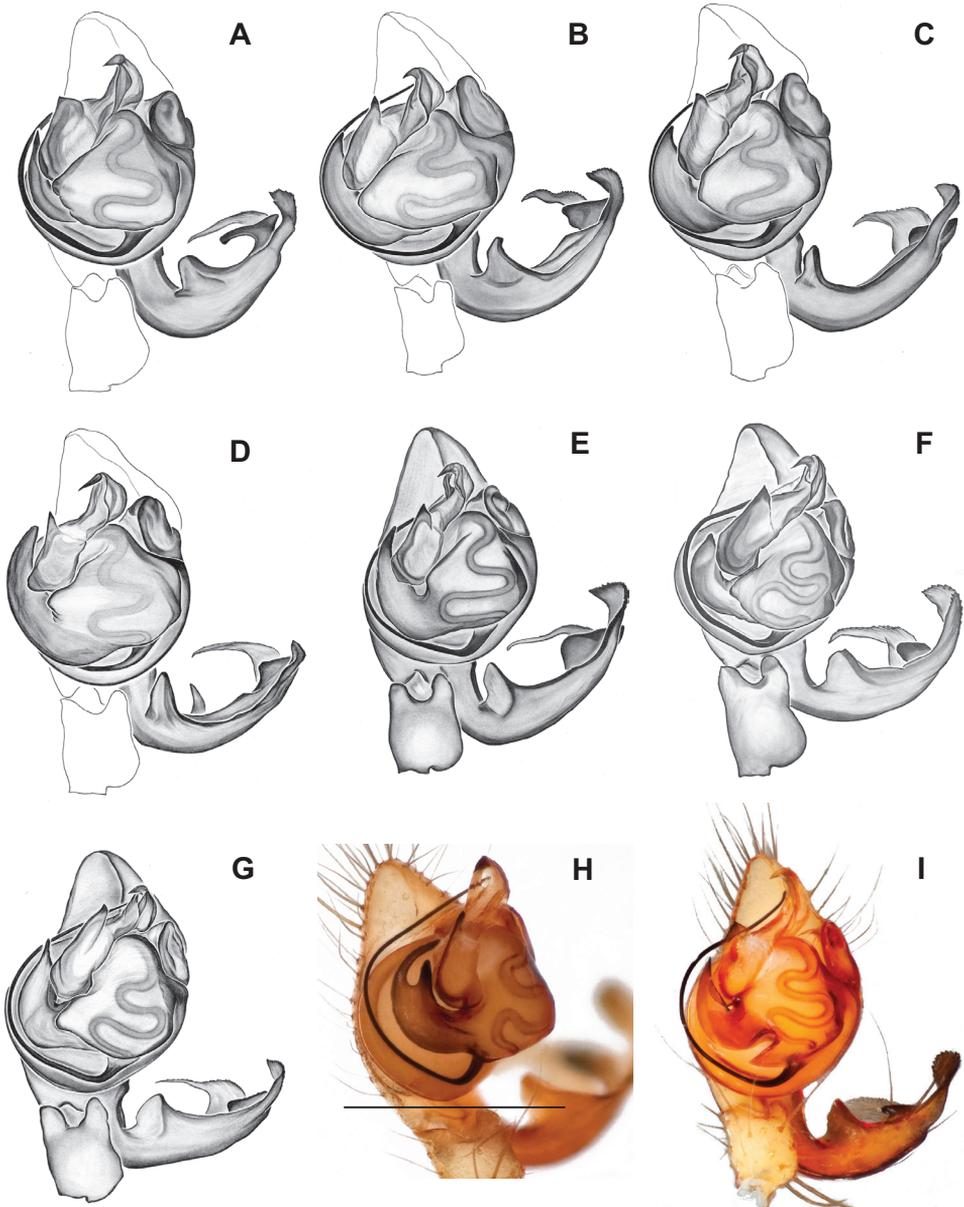


Figure 12. Comparative ♂ palps of *tennesseensis* group **A** *Nesticus silvanus* **B** *N. cherokeensis* **C** *N. holsingeri* **D** *N. mimus* **E** *N. tennesseensis* **F** *N. dilutus* **G** *N. carolinensis* **H** *N. paynei*, and **I** *N. roanensis*. All views are ventral. See subsequent figures for specimen locations and voucher details.

along the anteroventral edge (e.g., Fig. 12F). Palps viewed ventrally include a shoe-shaped tegular apophysis, projecting distally behind the rectangular median apophysis (Fig. 12A–I). Noticeable variation among species is found in the shape of the base of the tegular apophysis (where the tegular apophysis projects from tegulum), the shape

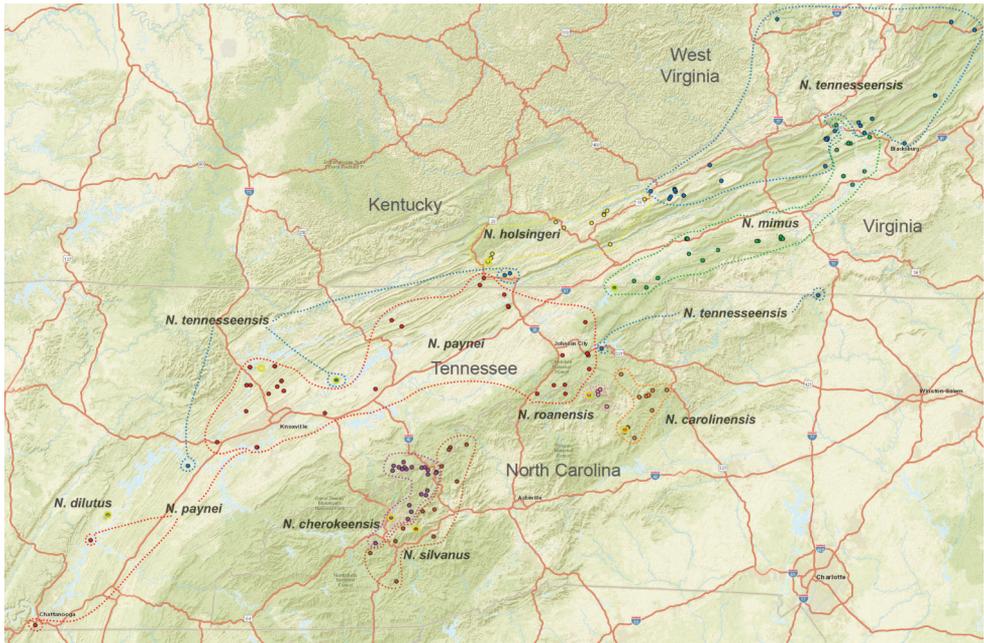


Figure 13. Distribution of *tennesseensis* group species. Type localities designated with yellow circles. State boundaries and major cities shown for geographic context. Dashed lines circumscribe known species distributions.

of the tegular apophysis itself, and the shape of the various paracymbial processes. Epigynal morphology is relatively conserved across species, with variation in the length and width of the epigynum and variation in spermathecal shape in certain taxa.

The *tennesseensis* group includes a combination of mostly cave-dwelling species distributed in the Appalachian Valley and Ridge (*Nesticus dilutus*, *N. holsingeri*, *N. mimus*), surface-dwelling species found entirely in the montane southern Blue Ridge (*N. carolinensis*, *N. cherokeensis*, *N. roanensis*, *N. silvanus*), and species found both in caves of the Valley and Ridge and mountains of the Blue Ridge (*N. tennesseensis*, *N. paynei*; see Fig. 13). The montane species *N. silvanus* and *N. cherokeensis* sp. nov. are successively sister to the remaining members of this species group, suggesting a south to north and mountain to cave biogeographic directionality (Fig. 13).

Nesticus silvanus Gertsch, 1984

Figs 14A–D, 15A–E

Nesticus silvanus Gertsch, 1984: 27, figs 141–143.

Material examined. Type material: Holotype: USA – North Carolina, Jackson–Haywood Co. • ♀ holotype; Water Rock Knob summit, elev. 1918 m, 30 Oct. 1969, W.

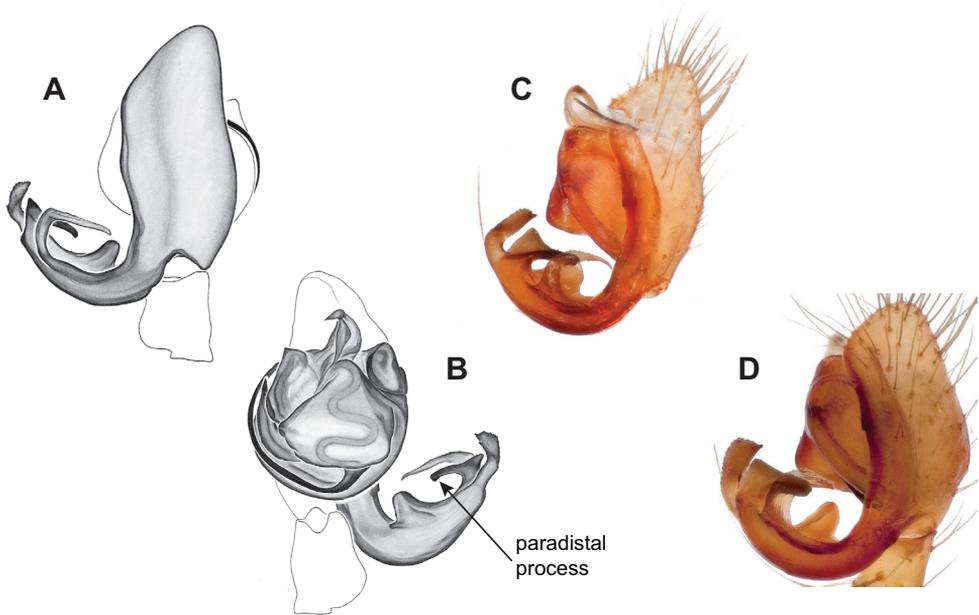


Figure 14. *Nesticus silvanus* ♂ palps. North Carolina, Jackson Co., vicinity of Water Rock Knob, MCH specimen #1080, dorsal (**A**), ventral (**B**) **C** North Carolina, Haywood Co., Blue Ridge Parkway, near Steestachee Bald overlook, MCH specimen #1145, dorsal **D** North Carolina, Madison Co., W of Rocky Bluff campground, MCH 01_139, dorsal. Scale bar: 0.5 mm.

Shear leg; AMNH; **New collections from type locality:** USA – North Carolina, Jackson Co. • 2♂, 7♀; vicinity of Water Rock Knob, off Blue Ridge Parkway; 35.4597°N, -83.1417°W; 9 Aug. 1992; M. Hedin leg; **Non type material:** – **Haywood Co.** • 2♂, 8♀; Blue Ridge Parkway, Mile 438, near Steestachee Bald overlook; 35.4263°N, -83.0388°W; 13 Aug. 1992; M. Hedin leg.; • 2♀; Cold Springs Creek, NE of I-40; 35.7585°N, -82.9938°W; 19 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_137; • ♀, 1 imm; Fie Top Road, along Fie Creek; 35.5451°N, -83.1045°W; 3 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_187; • 3♀; Germany Cove Road, vicinity Hemphill Creek; 35.5543°N, -83.036°W; 25 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_087; • 2♀, 4 imm; NW Hebo Mountain, Hwy 209; 35.6869°N, -82.9065°W; 25 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_086; – **Jackson Co.** • 6♀; Dicks Creek, near Dicks Creek Church, N of Dillsboro; 35.4056°N, -83.2586°W; 31 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_173; • 11♀, 1 imm; Soco Creek, up Shut-in Creek road; 35.4653°N, -83.2148°W; 3 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_188; – **Macon Co.** • 2♀; Falls branch of Elijay Creek, 2 mi. E Elijay; 35.2135°N, -83.2535°W; 11 Aug. 1992; M. Hedin leg.; – **Madison Co.** • ♂, 12♀; Hwy 209, W Rocky Bluff campground at Long Mountain Branch; 35.8599°N, -82.8502°W; 19 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_139; – **Swain Co.** • 14♀, 4 imm; Alarka Road, N

Deep Gap church; 35.3482°N, -83.4064°W; 28 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_168; – **Tennessee, Cocke Co.** • 4♀; south of Round Mountain, Shelton Branch, Hwy 107; 35.835°N, -82.9519°W; 27 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_095; • 5♀; southeast of Round Mountain, W Rattlesnake Gap; 35.8472°N, -82.9443°W; 19 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_138.

Diagnosis. Morphologically very similar to geographically parapatric *Nesticus cherokeensis* (Fig. 13). Males of *N. silvanus* can be distinguished from *N. cherokeensis* by the shape of the paradistal paracymbial process, which possesses a well-sclerotized ventral edge and a long prolateral extension (Fig. 14A–D). Also, the *N. silvanus* paracymbium lacks a basal projection of the dorsal process (Fig. 14A, C, D) as (sometimes) present in *N. cherokeensis* (Fig. 16B, F). Epigyna of *N. silvanus* are very similar to those of *N. cherokeensis* but (when viewed dorsally) possess epigynal plates with well separated medial margins, while in *N. cherokeensis* these plate margins are long, parallel, and touching (but see Fig. 15E vs. Fig. 17B). Additionally, *N. silvanus* epigyna possess anteriorly elongated epigynal pockets, lateral lobes that are shorter than the median septum, and relatively short spermathecae that lie perpendicular to the median septum, separating them from remaining members of the *tennesseensis* group.

Description of previously undescribed ♂ from type locality (MCH specimen #1080). Carapace light cream colored, gray pigmentation behind ocular area leading to midline and around edges. Legs pale yellow / cream. Abdomen with paired faint gray blotches on a light gray background. All eyes approximately equal in size, except for AMEs, ~ 1/3 width of ALEs. Eyes with rings of dark pigment. CL 1.39, CW 1.16, abdomen length 1.89, total body length 3.28. Leg I total length 8.66 (2.41, 0.54, 2.48, 2.25, 0.98), leg formula 1423, leg I / CW ratio 7.5. Paracymbium possesses a hook-shaped paradistal process with a well-sclerotized ventral edge and a long prolaterally-directed extension. Paracymbial dorsal process transparent and concave. Distal paracymbial process directed anteriorly, rounded, with a serrate edge. Ventral paracymbial process triangular with a blunted anterior edge. Median apophysis oval with a sharp anterior edge. Tegular process elongate, narrowing distally, and directed anteriorly. Nose-like bulge at the base of the tegular apophysis. Distal tip of conductor bent and directed prolaterally.

Variation. Minimal palpal variation was observed for males from three sample locations, the dorsal paracymbial process in a single Rocky Bluff male being slightly wider and shorter (Fig. 14A–D). Female genitalic variation across sample locations was minimal (Fig. 15A–E).

Distribution and natural history. Originally recorded from three locations (Gertsch 1984), now known to be relatively broadly distributed in appropriate surface microhabitats, including high-elevation habitats above 1900 m (e.g., Water Rock Knob, Steestachee Bald, etc.). This species is closely parapatric with *Nesticus cherokeensis* directly to the west, with an almost parallel geographic distribution (Fig. 13).

Strong phylogeographic structuring is observed in the mitochondrial data with a well-supported subclade found east of the Pigeon River (FieTop, Hebo Mtn, Rocky

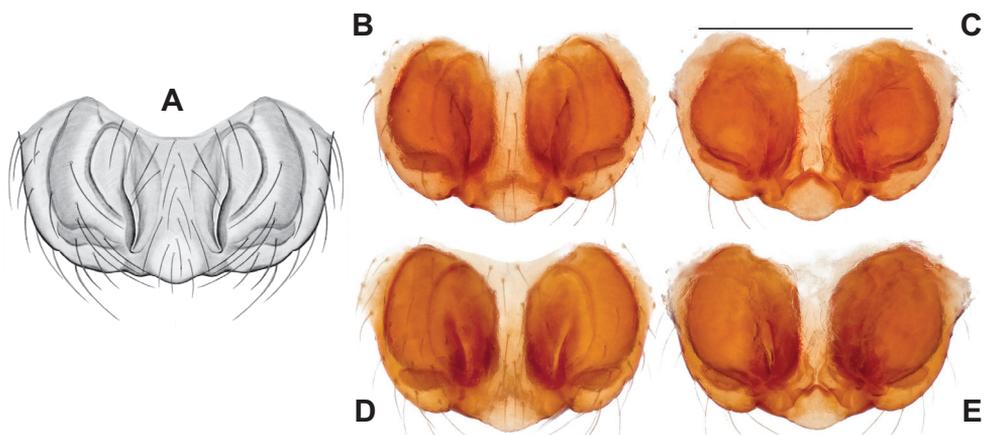


Figure 15. *Nesticus silvanus* epigynal variation **A** North Carolina, Jackson Co., vicinity of Water Rock Knob, MCH specimen #1083, ventral. North Carolina, Macon Co., Falls branch of Elizay Creek, MCH specimen #1115, ventral (**B**), dorsal (**C**) North Carolina, Madison Co., W of Rocky Bluff campground, MCH 01_139, ventral (**D**), dorsal (**E**). Scale bar: 0.5 mm.

Bluff, etc.; Fig. 6), suggesting a possible role for riverine barriers in phylogeographic structuring, and further suggesting a southwest to northeast biogeographic directionality.

As an example of natural history, one male and 12 females were collected from rocky void spaces in a moist, rocky ravine near Rocky Bluff campground (MCH 01_139) during a 30-minute devoted *Nesticus* search.

Remarks. This species is strongly supported as sister to remaining members of the *tennesseensis* group based on UCE evidence (Figs 3, 4).

***Nesticus cherokeensis* sp. nov.**

<https://zoobank.org/6057E5AC-B191-4964-B829-A347E2B74B0D>

Figs 16A–F, 17A–H

Material examined. Type material: *Holotype*: USA – North Carolina, Swain Co. • ♂; Blue Ridge Parkway, below Ballhoot Scar overlook near Ravensford; 35.5167°N, -83.2837°W; 9 Aug. 1992; M. Hedin leg.; (MCH specimen #1177). ***Paratypes*:** – North Carolina, Swain Co. • ♂, 10♀; Blue Ridge Parkway, below Ballhoot Scar overlook near Ravensford; 35.5167°N, -83.2837°W; 9 Aug. 1992; M. Hedin leg.; (MCH specimens #1176, #1178–1187). **Non type material:** – North Carolina, Haywood Co. • 2♂, 3♀; Dogwood Flats Creek, W Longarm Mountain; 35.7201°N, -83.0731°W; 18 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_135; • ♂, 12♀; Flat Branch Creek of Mt Sterling Creek, south of Waterville; 35.7407°N, -83.0741°W; 18 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_134; • 2♀; Flat Branch Rd, SE Mt. Sterling, along Laurel Creek; 35.7526°N, -83.0895°W; 12 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_046; • ♂, 3♀; FR 288 above Pigeon River; 35.726°N, -83.0265°W;

18 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_136; • ♀; FR 288 along Pigeon River, 0.4 mi. SW of I-40; 35.7308°N, -83.025°W; 27 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_094; • ♀, 4 imm; Great Smoky Mountains NP, Big Creek, 100 yards up Baxter Creek trail from picnic area; 35.7506°N, -83.1088°W; 17 Oct. 1994; F. Coyle leg.; • 3♀, 6 imm; Great Smoky Mountains NP, Boogerman Trail 0.5 mi from Northern end, N extension Den Ridge; 35.6225°N, -83.0847°W; 11 Sep. 1994; F. Coyle, J. Miller leg.; • ♀; Great Smoky Mountains NP, Cataloochee area, Sag Branch, 1.5 mi from N end Caldwell Fork Trail; 35.6435°N, -83.0766°W; 10 Sep. 1994; F. Coyle, J. Miller leg.; • ♂; Great Smoky Mountains NP, Cataloochee, 150 meters S mouth Palmer Branch at Caldwell Fork; 35.6251°N, -83.1121°W; 4 Jun. 1996; F. Coyle, Edwards, Stiles, Wright leg.; – **North Carolina, Jackson Co.** • ♂, ♀; Blue Ridge Parkway, Mile 460, near Bunches Bald Tunnel; 35.5092°N, -83.1883°W; 9 Aug. 1992; M. Hedin leg.; – **North Carolina, Swain Co.** • ♂, ♀; Great Smoky Mountains NP, 0.25 mi. NW Hintooga Overlook/Picnic Area on Hintooga Round Bottom Road; 35.5748°N, -83.1805°W; 3 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_186; • ♂, 4♀, 1 imm; Great Smoky Mountains NP, road to Balsam Mountain, N Black Camp Gap; 35.5437°N, -83.1679°W; 3 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_185; • ♂, ♀; Wesser Creek, Dills Road, S of Whittier; 35.3953°N, -83.3746°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_118; – **Tennessee, Cocke Co.** • ♂, 5♀; Great Smoky Mountains NP, above Cosby CG on Snake Den Ridge trail; 35.7432°N, -83.2218°W; 1 Aug. 1995; F. Coyle, Carbiener leg.; • ♀; Great Smoky Mountains NP, Cosby Ranger Station along Cosby Creek, behind ATBI residence house; 35.7779°N, -83.2135°W; 28 Jul. 2000; M. Hedin, J. Cokendolpher leg.; MCH 00_138; • ♂; Great Smoky Mountains NP, Maddron Bald trail to Albright Grove; 35.7608°N, -83.271°W; 3 Aug. 2000; M. Hedin, W. Reeves leg.; MCH 00_149; • ♂; Great Smoky Mountains NP, N side Gabes Creek at Gabes Mountain trail; 35.7523°N, -83.2419°W; 1 Aug. 1995; F. Coyle, Williams, Carbiener leg.; • 2♀; Great Smoky Mountains NP, near Cosby campground, below group camp parking area; 35.7533°N, -83.2066°W; 27 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_097; • ♀; Great Smoky Mountains NP, S side Indian Camp Creek on Maddron Bald Trail; 35.7378°N, -83.2777°W; 16 Apr. 1994; M. Hedin, B. Dellinger leg.; • 3♂, 4♀; Great Smoky Mountains NP, trail from Cosby to Low Gap; 35.7453°N, -83.197°W; 1 Aug. 2000; M. Hedin leg.; MCH 00_145; • 2♀; Great Smoky Mountains NP, trail from Low Gap to Mt. Cammerer; 35.754°N, -83.1658°W; 1 Aug. 2000; M. Hedin leg.; MCH 00_146.

Diagnosis. As discussed above, this species is morphologically most similar to geographically adjacent *Nesticus silvanus*. Males have a fan-shaped paradistal paracymbial process (Fig. 16A–F) that lacks the elongate retrolateral extension and well-sclerotized ventral edge found in *N. silvanus*. Epigyna are very similar to that of *N. silvanus*, but when viewed dorsally possess adjacent medial plate margins that are parallel to each other, unlike the indistinct or pointed margins in *N. silvanus* (but see Fig. 15E. vs. Fig. 17B).

Description of ♂ holotype (MCH specimen #1177). Carapace cream colored, gray pigmentation behind ocular area leading to midline and around edges. Legs pale

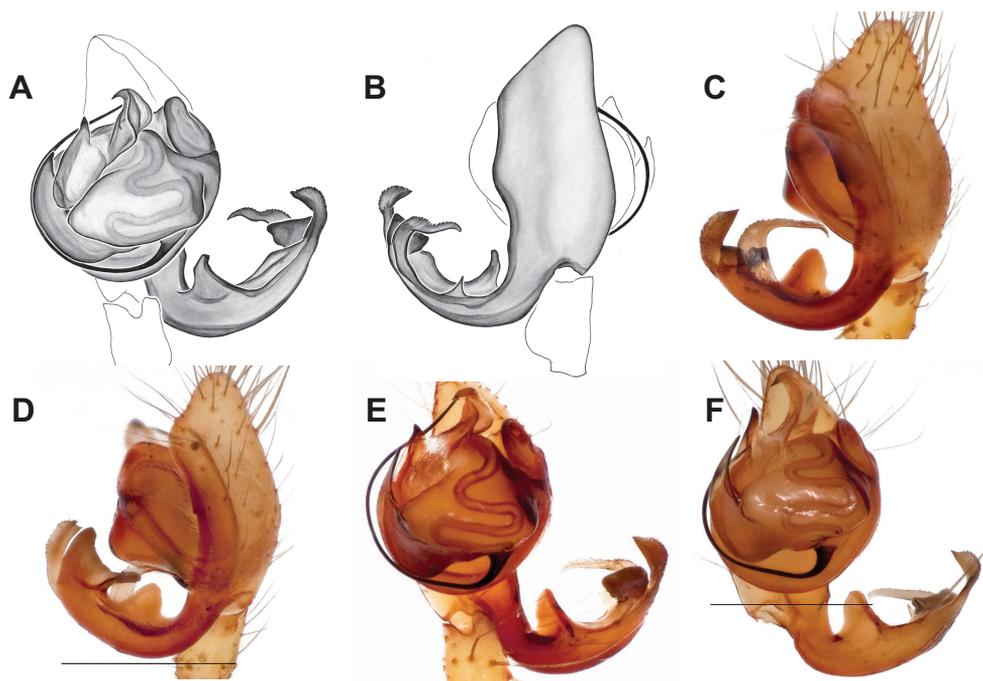


Figure 16. *Nesticus cherokeensis* sp. nov. ♂ palps. North Carolina, Swain Co., Blue Ridge Parkway, below Ballhoo Scar overlook, MCH specimen #1177, ventral (**A**), dorsal (**B**) **C** North Carolina, Haywood Co., S of Waterville, MCH 01_134, dorsal **D** North Carolina, Jackson Co., Blue Ridge Parkway, near Bunches Bald Tunnel, MCH specimen #1089, dorsal **E** North Carolina, Haywood Co., FR 288 above Pigeon River, MCH 01_136, ventral **F** North Carolina, Swain Co., road to Balsam Mountain, MCH 02_185, ventral. Scale bar: 0.5 mm.

yellow / cream. Abdomen with paired gray blotches on a light gray background. All eyes approximately equal in size, except for AMEs, ~ 1/2 width of ALEs. Eyes ringed with dark pigment. CL 1.45, CW 1.27, abdomen length 1.77, total body length 3.22. Leg I total length 10.55 (2.96, 0.54, 3.18, 2.8, 1.07), leg formula 1423, leg I / CW ratio 8.3. Paracymbium with a triangular ventral process with a sclerotized retrolateral edge, a dorsal process with an expanded serrate, distal portion, a heavily sclerotized triangular paradistal process, and a transparent, elongated, prolaterally directed dorsal process with a small triangular basal extension. Median apophysis a narrow oval with anteriorly directed edge coming to a point. Tegular process thick and sharp-tipped distally. Nose-like bulge at the base of the tegular apophysis. Distal tip of conductor bent and directed prolaterally.

♂ **Variation.** Males from different sample locations vary in the presence / absence of a small basal projection of the dorsal process (Fig. 16B, C).

Description of ♀ paratype (MCH specimen #1181). Carapace cream colored, gray pigmentation behind ocular area leading to midline and around edges. Legs pale yellow / cream. Abdomen with paired dark gray blotches on a pale cream background.

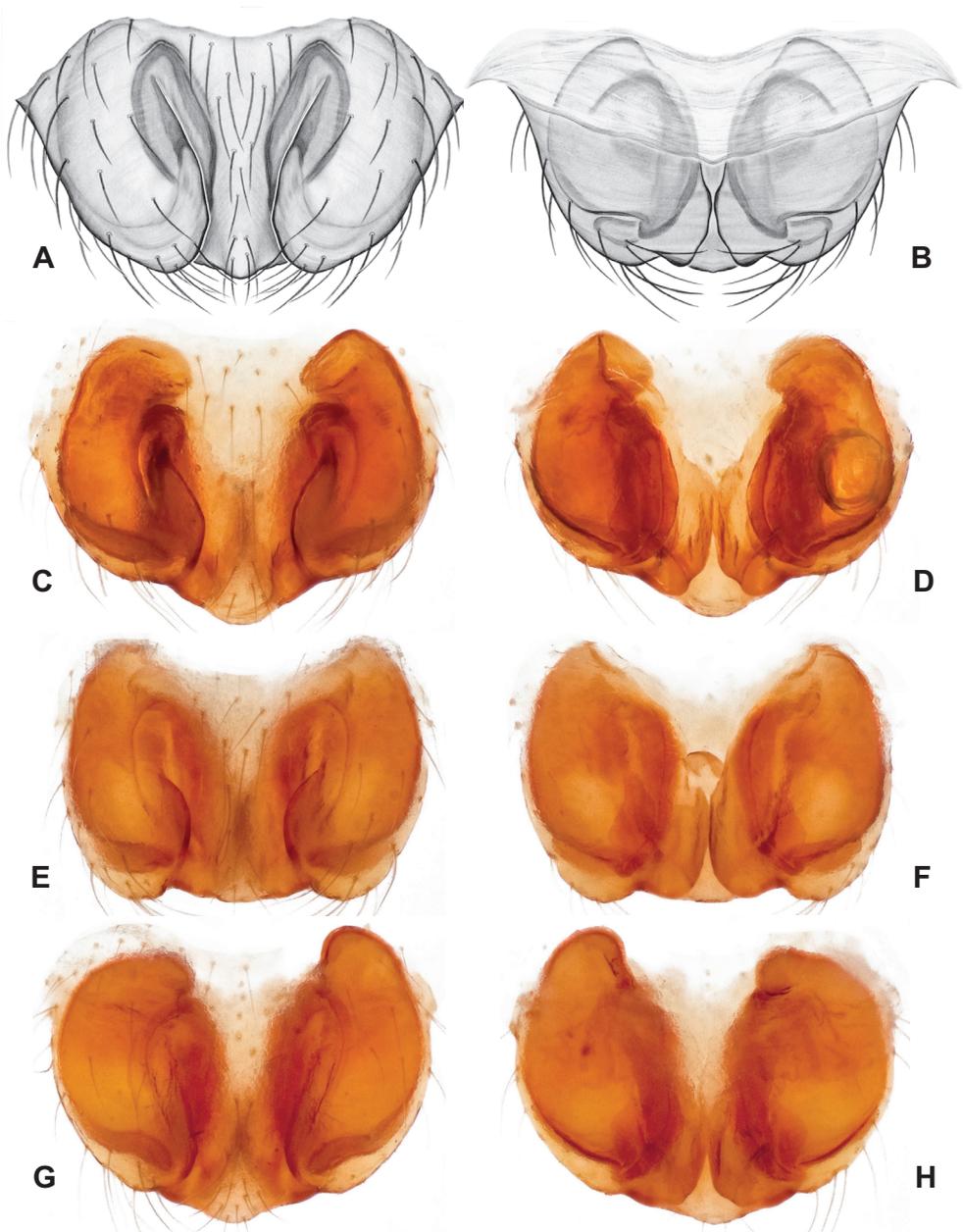


Figure 17. *Nesticus cherokeensis* sp. nov. epigynal variation. North Carolina, Swain Co., Blue Ridge Parkway, below Ballhoot Scar overlook, MCH specimen #1181, ventral (**A**), dorsal (**B**) North Carolina, Haywood Co., S of Waterville, MCH 01_134, ventral (**C**), dorsal (**D**) Tennessee, Cocke Co., S side of Indian Camp Creek, MCH specimen #1982, ventral (**E**), dorsal (**F**) North Carolina, Haywood Co., Cataloochee area, Sag Branch, ventral (**G**), dorsal (**H**). Scale bar: 0.5 mm.

Eyes approximately equal in size, except for AMEs, ~ 1/2 width of ALEs. Eyes with rings of dark pigment. CL 1.51, CW 1.28, abdomen length 2.08, total body length 3.59. Leg I total length 9.27 (2.71, 0.58, 2.68, 2.23, 1.07), leg formula 1423, leg I / CW ratio 7.2. Epigynum possesses oval-shaped lateral lobes that extend to the posterior end of the median septum. Spermathecae visible beneath posterior lateral lobes, short and angled slightly upwards from perpendicular to septum. Viewed dorsally, large internal lobes extend anteriorly and possess sclerotized rims. Medial margins parallel to each other and touching along the midline.

♀ **Variation.** Females from different sample locations vary in the symmetry of the interior epigynal plates (Fig. 17C, G).

Distribution and natural history. Found in rocky microhabitats from the rugged mountains of the eastern Great Smoky Mountains National Park, and adjacent eastern and southern locations (Fig. 13). The apparent gap at high elevations in this region (Fig. 13) likely reflects a lack of sampling in these less accessible high-elevation locations. Most collections include a modest number of specimens, suggesting a natural rarity for this taxon.

Along the Maddron Bald (along Indian Camp Creek) and the Low Gap to Mt. Cammerer trails (MCH 00_146) both *Nesticus cherokeensis* and *N. binfordae* sp. nov. were collected, indicating that these species are syntopic or nearly so at these locations. At both locations multiple collections were taken along an elevational transect and unfortunately lumped into a single collecting event. It is therefore not possible to discern if different species were collected at the exact same location (truly syntopic) or were closely parapatric along these elevational transects.

Etymology. The species epithet (*cherokeensis*) honors the larger Cherokee Nation whose ancestral homelands included the mountains of western North Carolina. *Nesticus cherokeensis* can also be found near The Qualla Boundary, home of the Eastern Band of Cherokee.

Remarks. This species is strongly supported as sister to remaining members of the *tennesseensis* group based on UCE evidence (Figs 3, 4). Mitochondrial structuring is very pronounced, with each sample location (or set of adjacent locations) genetically distinct (Fig. 6).

Nesticus holsingeri Gertsch, 1984

Fig. 18A–G

Nesticus holsingeri Gertsch, 1984: 25, figs 66–67, 91–93.

Material examined. Type material: Holotype: USA – Virginia, Scott Co. • ♂ holotype; Pond Cave; 5 Nov. 1966; J. Holsinger, S. Taylor leg; AMNH; **New collections from type locality:** USA – Scott Co. • 2♂, 7♀; Pond Cave, Rye Cove; 6 Oct. 1993;

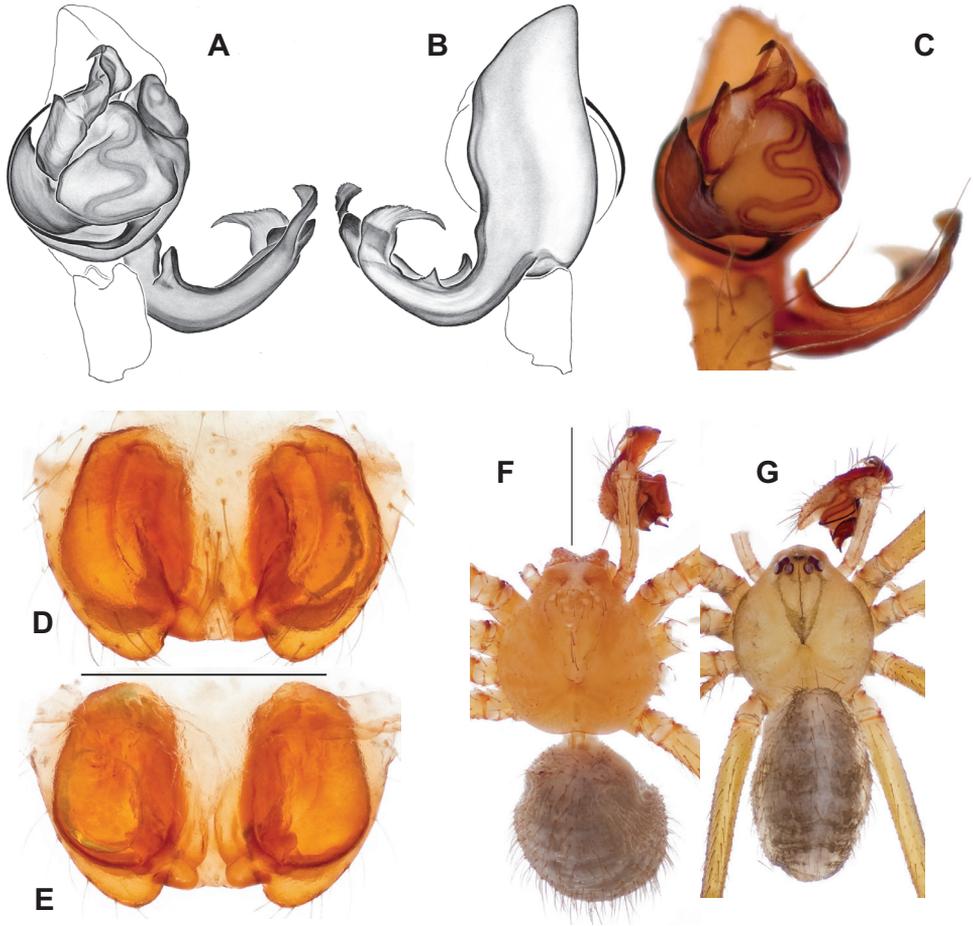


Figure 18. *Nesticus holsingeri*. Virginia, Scott Co., Pond Cave, MCH specimen #1780, ♂ palp, ventral (A), dorsal (B) C Virginia, Washington Co., Brumley Creek, MCH 04_026, ♂ palp, ventral. ♀ epigynum. Virginia, Washington Co., Brumley Creek, MCH 04_026, epigynum, ventral (D), dorsal (E). Scale bar: 0.5 mm. ♂ habitus images F Virginia, Washington Co., Brumley Creek, MCH 04_026 G Virginia, Scott Co., Pond Cave, MCH specimen #1780. Scale bar: 1 mm.

M. Hedin, C. Phillips leg; **Non type material:** – **Russell Co.** • ♂; Banners Corner Cave; 10 Apr. 2017; T. Malabad leg.; • ♂, 2♀; Bundys Cave No. 1, west of Lebanon, VA; 31 Jan. 2020; T. Malabad, R. Reynolds leg.; • ♂, ♀, 8 imm; Concrete Tank Cave; 24 Jul. 2017; T. Malabad leg.; • ♂, 3♀, 12 imm; Daugherty Cave, northeast of Lebanon, VA; 10 Apr. 2017; T. Malabad leg.; • 2♀; Daugherty Cave; 26 Jun. 2020; T. Malabad, A. Malabad leg.; • ♀; Ferrells Cave, northeast of Rosedale, VA; 26 Jun. 2020; T. Malabad, A. Malabad leg.; –**Scott Co.** • 4♂, 11♀; Alley Cave, E of Natural Tunnel SP; 19 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♂; Alley Cave; 6 Oct. 1993; M. Hedin, C. Phillips leg.; • 3♂, ♀; Alley Cave; 22 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_072; • 2♀; Cox Ram Pump Cave; 3 Aug. 2016;

T. Malabad leg.; – **Washington Co.** • 3♂, ♀; Brumley Creek, near Brumley Gap; 36.7933°N, -82.0229°W; 7 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_026; – **Wise Co.** • 4♂, 6♀; Burton's Cave, SW of St Paul; 8 Oct. 1993; M. Hedin, C. Phillips leg.

Diagnosis. Strongly supported by both mitochondrial and UCE data as sister to *Nesticus mimus*. The male tegular apophysis of *N. holsingeri* is shorter (nearly as wide as long) with a more pronounced narrowing tip (Fig. 18A, C) than in *N. mimus* (Fig. 19B, C). *Nesticus holsingeri* epigyna are more squarish and lack conspicuous elongate spermathecae (Fig. 18D, E), as compared to longer (anterior to posterior) *N. mimus* epigyna with conspicuous elongate spermathecae (Fig. 20A–H).

Variation. This species exhibits interesting variation in somatic morphology. Specimens from Pond Cave are pale and long-legged with reduced eye pigmentation, specimens from Burton's Cave and Alley Cave are pale and long-legged but with well-developed eyes, and specimens from the surface Brumley Creek population exhibit an “epigean” habitus, with dark abdomens, shorter legs, and well-developed eyes (Fig. 18F, G). Minimal genitalic variation was observed across sample locations.

Distribution and natural history. Known only from a small area in southwestern Virginia, in the upper Clinch River drainage basin (Fig. 13). All known records are from limestone caves, except for the Brumley Creek population where spiders were collected from under surface rocks in a moist, north-facing stream valley. This species has been collected in near syntopy with *Nesticus carteri* at Alley Cave, Virginia. At this site *N. carteri* is found under rocks in talus in a sink leading to the cave entrance, whereas *N. holsingeri* has only been collected from the dark zone of the cave.

Remarks. The variation in degree of troglomorphy within this single species suggests that this character suite (eye development, pigmentation, leg length, etc.) can evolve relatively rapidly, as seen in other cave spider taxa (e.g., Arnedo et al. 2007).

Nesticus mimus Gertsch, 1984

Figs 19A–C, 20A–H

Nesticus mimus Gertsch, 1984: 26, figs 64, 65.

Nesticus tennesseensis Gertsch, 1984: 24, 25 (in part).

Material examined. Type material: Holotype: USA – Virginia, Washington Co. • ♂ holotype; Shiloh School Cave, SE of Abingdon; 25 Nov. 1960; C.W. Greever leg; AMNH. **Non type material: USA – Tennessee, Johnson Co.** • ♂, 3♀; Backbone Rock Rec Area, 4 mi. S Damascus, off Hwy 133; 36.594°N, -81.8163°W; 7 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_025; **Tennessee, Sullivan Co.** • ♂, ♀; Potter's Cave, S Abingdon; 14 Jul. 1979; J.R. Holsinger et al. leg.; AMNH; – **Virginia, Giles Co.** • 2♀, 26 imm; Curve Saltpetre Cave; 14 May. 2019; T. Malabad leg.; • ♀; Harris Cave, S of Pearisburg; 29 Jun. 1974; J. Holsinger, L. Ferguson leg; AMNH; • ♀, 2 imm; Harris Cave; 14 Sep. 2018; T. Malabad leg.;

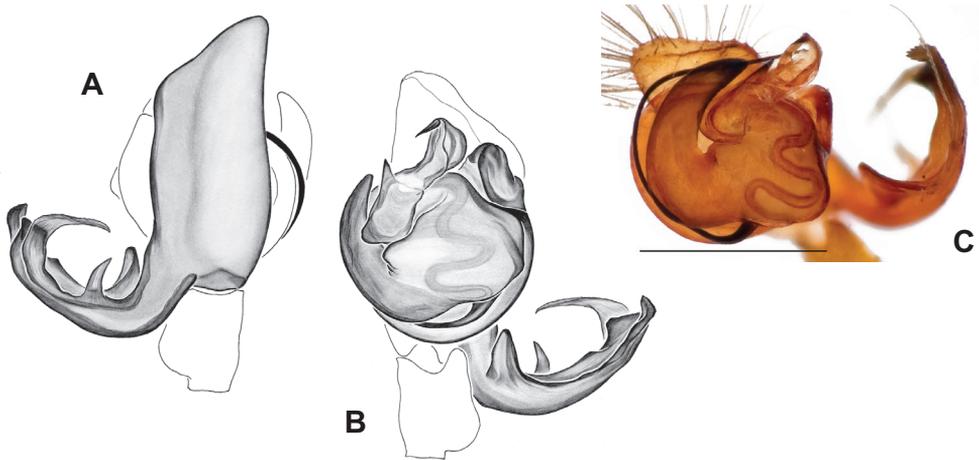


Figure 19. *Nesticus mimus* ♂ palps. Virginia, Giles Co., Straley's Cave #1, MCH specimen #1396, dorsal (A), ventral (B) C Tennessee, Johnson Co., Backbone Rock Recreational Area, MCH 04_025, ventral. Scale bar: 0.5 mm.

• 3♀, 7 imm; Spruce Run Mountain Cave; 14 Nov. 2018; T. Malabad leg.; • 2 imm; Straley's Cave No. 1; 6 Sep. 1958; T.C. Barr leg; AMNH; • 2♂, 3♀; Co. Straley's Cave No. 1 off Eggleston Road, S Pembroke; 17 Sep. 1992; M. Hedin, S. O'Kane leg.; • ♂, 7♀; Straley's Cave No. 1 off Eggleston Road, S Pembroke; 10 Oct. 1993; M. Hedin, C. Phillips leg.; • ♀; Straleys Cave No. 2; 12 Sep. 2018; T. Malabad leg.; – **Pulaski Co.** • 4♀, 31 imm; Maze Cave; 25 Sep. 2018; T. Malabad leg.; • 2♀; Mebane Saltpetre Cave; 3 Oct. 2018; T. Malabad leg.; – **Radford Co.** • ♀, 3 imm; Adams Cave; 11 Apr. 2019; T. Malabad leg.; – **Smyth Co.** • 3♂, 6♀; Cow Shelter Cave, SE of Sugar Grove; 6 Oct. 1993; M. Hedin, C. Phillips leg.; • ♂, ♀; McMullin Cave, southwest of Marion, VA; 11 Jul. 2019; T. Malabad leg.; • ♀; McMullin Cave; 10 Sep. 2019; T. Malabad, K. Kosič Ficco leg.; • 2♀; McMullin Cave; 22 Oct. 2019; T. Malabad, K. Kosič Ficco leg.; • 9♂, 20♀; McMullin Cave; 2 Mar. 2020; T. Malabad, K. Kosič Ficco leg.; • ♂, 3♀; McMullin Cave; 23 Jun. 2020; T. Malabad, A. Malabad leg.; • 3♀; Mt. Rogers National Rec Area, 0.5 mi S of Hurricane CG on NF 84; 36.7186°N, -81.4911°W; 11 Aug. 2007; M. Hedin, R. Keith leg.; MCH 07_087; • ♀, 3 imm; Rowland Creek Cave; 10 Aug. 2018; T. Malabad leg.; • 8♂, 7♀, 1 imm; Whitetop Laurel Creek, Hwy 58, E of Damascus; 36.637°N, -81.75°W; 31 May. 2016; M. Hedin, S. Derkarabetian, J. Starrett, D. Proud leg.; MCH 16_034; – **Washington Co.** • 7♂, 9♀; Neal's Sinks, 1.5 mi S Alvarado, Sweet Hollow Road; 18 Sep. 1992; M. Hedin, S. O'Kane leg.; – **Wythe Co.** • ♂, 4♀, 12 imm; Canyon Cave No. 1; 21 Sep. 2018; T. Malabad leg.; • ♂, ♀, 1 imm; Deep Spring Cave; 21 Sep. 2018; W. Orndorff leg.; • 6 imm; Deep Spring Cave; 7 Jul. 2018; T. Malabad leg.; • 2♂, 2♀, 14 imm; Mockleys Cave; 20 Sep. 2018; T. Malabad leg.; • ♂, ♀, 4 imm; Sinking Spring Cave No. 1; 21 Sep. 2018; T. Malabad leg.; • ♀, 7 imm; Sinking Spring Cave No. 2; 21 Sep. 2018; T. Malabad leg.; • 2♀, 6 imm; Speedwell Cave No. 1; 20 Sep. 2018; T. Malabad leg.

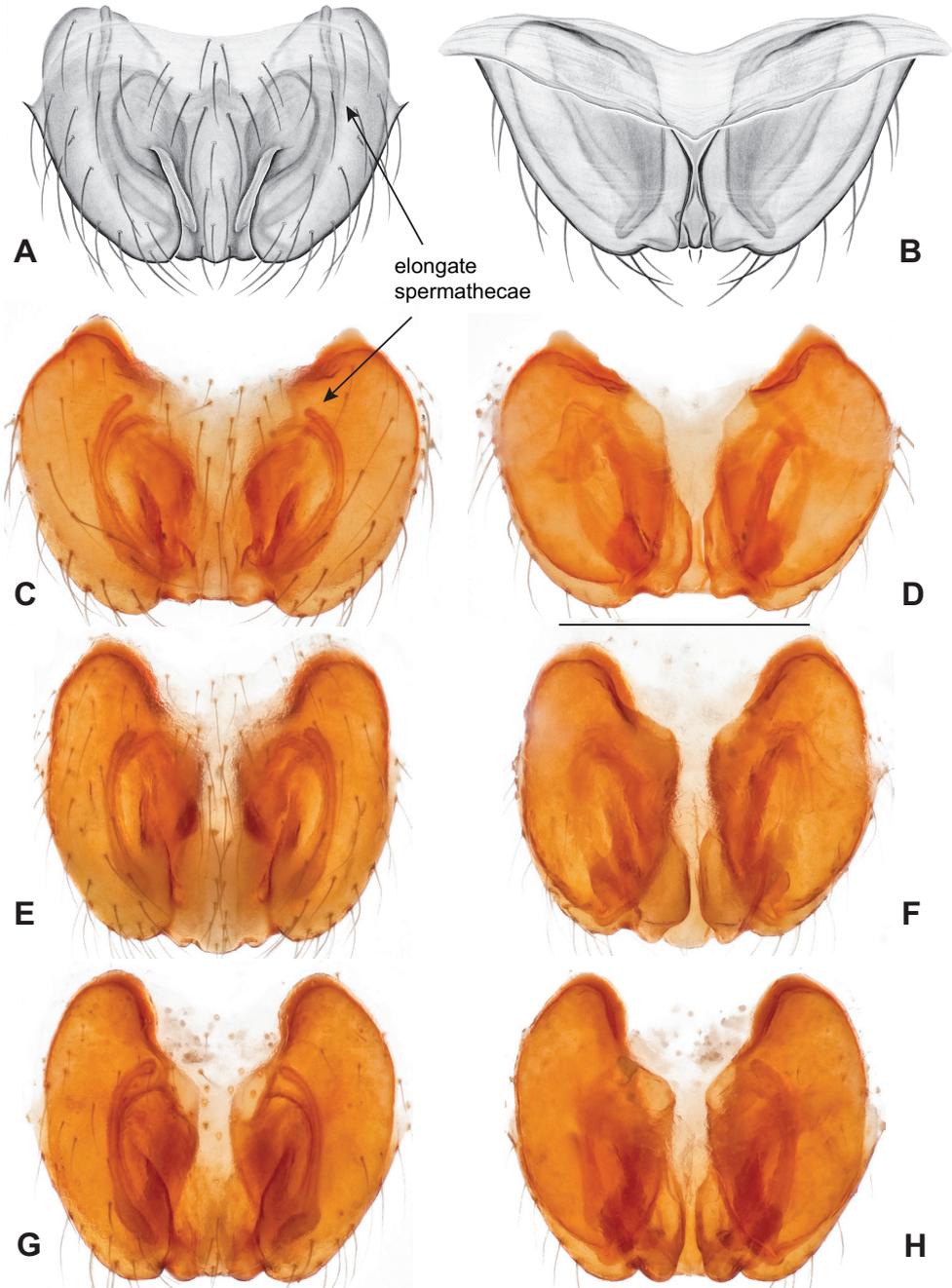


Figure 20. *Nesticus mimus* epigynal variation. Virginia, Giles Co., Straley's Cave #1, MCH specimen #1399, ventral (A), dorsal (B). Virginia, Washington Co., Neal's Sinks, MCH specimen #1430, ventral (C), dorsal (D). Virginia, Smyth Co., near Hurricane CG, MCH 07_087, ventral (E), dorsal (F). Tennessee, Johnson Co., Backbone Rock Recreational Area, MCH 04_025, ventral (G), dorsal (H). Scale bar: 0.5 mm.

Diagnosis. Strongly supported by both mitochondrial and UCE data as sister to *Nesticus holsingeri*. In *N. mimus* the basal dorsal process of the paracymbium is longer (Fig. 19A, B) and the tegular apophysis is relatively long and narrow, almost reaching the pronounced acute distal process of the median apophysis (Fig. 19B, C). Females possess a relatively elongate, narrow epigynum that is distinctive in the species group in possessing long, thin spermathecae that viewed ventrally bow upwards around the outer edge of the lateral pockets (Fig. 20A–H).

Description of previously undescribed ♀ (MCH specimen #1398). Carapace light cream colored, faint gray pigmentation behind ocular area leading to midline. Legs pale yellow / cream. Abdomen concolorous light cream. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.37, CW 1.19, abdomen length 1.89, total body length 3.26. Leg I total length 8.68 (2.36, 0.58, 2.5, 2.22, 1.02), leg formula 1423, leg I / CW ratio 7.3. Epigynum relatively elongate and narrow. Lateral lobes well-defined with internal edges that extend posteriorly to the end of median septum or slightly further. Spermathecae extremely long and curved around fovea to anterior edge of epigynum. Large internal lobes (viewed dorsally) extend anteriorly with sclerotized rims. Medial margins parallel to each other but not touching along midline.

Variation. No noteworthy variation in male or female genitalia was found across sample locations.

Distribution and natural history. Known only from a small area of the Appalachian Valley and Ridge in southwestern Virginia and adjacent eastern Tennessee (Fig. 13). Found in both caves and moist, dark near-surface microhabitats. As an example of the near-surface natural history, eight males and seven females were collected along White-top Laurel Creek (MCH 16_034) from rock piles in a rich, rocky streamside forest.

Remarks. Gertsch (1984) incorrectly identified specimens from the following locations as *Nesticus tennesseensis*: Straley's Cave No. 1 (2 immatures), Harris Cave (1 ♀), and Potter's Cave (1 ♂, 1 ♀) – specimens from these populations belong to *N. mimus*. Gertsch (1984) also provisionally identified specimens from two montane locations as *N. mimus*: a single female from Table Rock Mountain (Burke County, NC), and a male specimen from Grandfather Mountain (cited as Watauga County, NC but label reads Avery County, NC). We contend that specimens from both locations correspond to *N. carolinensis* (see further comments below).

Specimens from Hedin (1997b) referred to as *Nesticus* “nov sp 1” (from Neal's Sinks, Straley's Cave No. 1, and Cow Shelter Cave) are actually *N. mimus*, and those referred to as *N. mimus* (Grandfather Mtn, Linville Gorge) are actually *N. carolinensis*.

***Nesticus tennesseensis* (Petrunkevitch, 1925)**

Figs 21A–C, 22A–G

Ivesia tennesseensis Petrunkevitch, 1925: 321, pl. 20, figs 4, 7, 10.

Yvesella tennesseensis: Arndt 1928: 84.

Ivesia tennesseensis: Bishop 1950: 10, figs 5–8.

Nesticus tennesseensis: Gertsch 1984: 23, figs 58–63, 82–84.

Material examined. New collections from type locality: – Tennessee, Grainger Co. • 2♂, 2♀; Indian Cave, E of Blaine; 25 Sep. 1991; M. Hedin, K. Crandall leg.; • 3♂, 12♀; Indian Cave, E of Blaine; 21 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♂, ♀, 5 imm; Indian Cave, TGA4; 22 Feb. 2014; M.L. Niemiller, A.S. Engel, S. Engel, A. Paterson leg.; MLN 14–010.7; **Non type material: USA – North Carolina, Surry Co.** • ♀, 1 imm; vic Fisher Peak lookout tower, SE of Blue Ridge Parkway; 36.559°N, -80.8276°W; 12 Aug. 2007; M. Hedin, R. Keith leg.; MCH 07_091; – **Tennessee, Carter Co.** • 3♀; off Hwy 167/321, near Watauga Lake, along Little Stony Creek; 36.309°N, -82.0732°W; 23 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_078; – **Tennessee, Roane Co.** • 5♂, ♀, 11 imm; Berry Cave, TRN3; 28 Jun. 2014; M.L. Niemiller, C.D.R. Stephen, A.S. Engel, et al. leg.; MLN 14–034.1; – **Virginia, Alleghany Co.** • ♂; Island Ford Cave, west of Low Moor, VA; 12 Jun. 2020; T. Malabad, P. Tegelman Malabad, K. Malabad, A. Malabad leg.; • 6♂, 7♀; Rumbold’s Cave, near Callaghan; 16 Sep. 1992; M. Hedin, S. O’Kane leg.; – **Virginia, Craig Co.** • 2♂, 4♀; Walkthrough Cave, sw of Newcastle; 10 Oct. 1993; M. Hedin, C. Phillips leg.; – **Virginia, Giles Co.** • 6♂, 7♀; Ballard’s Cave, just S Pearisburg; 9 Oct. 1993; M. Hedin, C. Phillips leg.; • 2♂, 4♀; Dead Doe Cave; 1 Jul. 2019; T. Malabad leg.; • ♀; Doe Mountain Cave; 16 Aug. 2019; T. Malabad leg.; • ♂; Hodges Cave, 3.5 miles southwest of Pearisburg, VA; 26 Nov. 2019; T. Malabad, K. Kosič Ficco leg.; • ♀; Little Stony Creek, NE of Pembroke; 37.3565°N, -80.5921°W; 10 Oct. 1993; M. Hedin, C. Phillips leg.; • ♂; Mountain Lake Biological Station, Jefferson Nat Forest; 37.3755°N, -80.5158°W; 11 Jul. 2013; C Richart leg.; • 4♂, 10♀; Starne’s Cave, SW Pearisburg, Wilburn Valley; 10 Oct. 1993; M. Hedin, C. Phillips leg.; • ♂5 imm; Starnes Cave; 3 Jun. 2019; T. Malabad leg.; • ♂, 4♀, 15 imm; Yer Cave; 9 Aug. 2019; T. Malabad leg.; – **Virginia, Montgomery Co.** • 4♀, 1 imm; Hancock Blowhole Cave; 14 Dec. 2014; E. Koertge leg.; – **Virginia, Scott Co.** • ♀, 6 imm; Coley Cave #2; 15 Sep. 2015; W. Orndorff leg.; • ♀; Herrons Echo Hall; 4 Aug. 2016; T. Malabad leg.; – **Virginia, Tazewell Co.** • 6♂, 11♀; Cassell’s Farm Cave, Burkes Garden; 9 Oct. 1993; M. Hedin, C. Phillips leg.; • 2♀, 2 imm; Cauliflower Cave; 24 Oct. 2018; T. Malabad leg.; • 2 imm; Corkscrew Cave; 4 Mar. 2017; K. Kosič Ficco leg.; • ♀, 4 imm; Corkscrew Cave; 27 Oct. 2018; T. Malabad leg.; • 4♂, 8♀; Fallen Rock Cave, Ward Cove, S Maiden Spring; 9 Oct. 1993; M. Hedin, C. Phillips leg.; • ♂; Gillespie Water Cave, southwest of Liberty, VA; 9 Sep. 2019; T. Malabad, K. Kosič Ficco, A. Futrell leg.; • ♂, 4♀, 9 imm; Glenwood Church Cave; 24 Oct. 2018; T. Malabad leg.; • ♀, 5 imm; Gulley Cave; 22 Jul. 2019; T. Malabad leg.; • 2♀, 3 imm; Little River Cave; 29 Nov. 2018; T. Malabad leg.; • 3♀, 4 imm; Lost Mill Cave 1; 22 Jul. 2019; T. Malabad leg.; • 2♀, 2 imm; Lost Mill Cave 2; 22 Jul. 2019; T. Malabad leg.; • 2♀; Stompbottom Cave, southeast of Claypool Hill, VA; 5 May. 2021; T. Malabad, K. Kosič Ficco, M. Ficco leg.; • ♂, 3♀; Stonley Cave; 10 Jan. 2019; T. Malabad leg.; – **West Virginia, Raleigh Co.** • ♂, 5♀; Grandview State Park, New River Gorge; 37.8321°N, -81.0614°W; 15 Sep. 1992; M. Hedin, S. O’Kane leg.

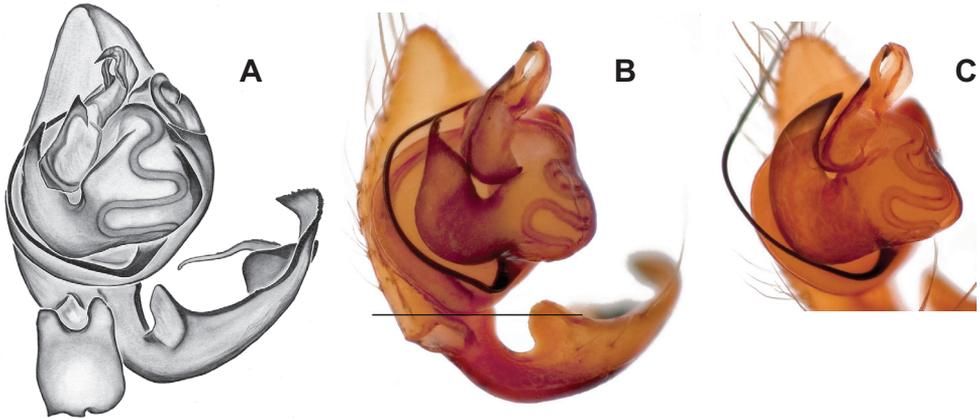


Figure 21. *Nesticus tennesseensis* ♂ palps, ventral view **A** West Virginia, Raleigh Co., Grandview SP, MCH specimen #1360 **B** Virginia, Tazewell Co., Fallen Rock Cave, MCH specimen #1808 **C** Fallen Rock Cave, MCH specimen #1807. Scale bar: 0.5 mm.

Diagnosis. *Nesticus tennesseensis* and *N. dilutus* are morphologically similar sister species. Male differences are noted in the Diagnosis of *N. dilutus* below. Male *N. tennesseensis* may be differentiated from other members of this species group by the combination of palps with a paracymbium with a wide, broad ventral process, rectangular paradistal paracymbial process, a rectangular median apophysis with an anteriorly-pointed sclerotized edge, and a narrow, singularly-pointed tegular apophysis that extends to ~ half the length of the median apophysis (Fig. 21A–C). Females may be differentiated from other members of this species group by an overall rounded epigynum with short, somewhat globular spermathecae that extend perpendicular to the median septum (Fig. 22A–G). Viewed dorsally, circular pockets lie above extended parallel separated medial margins that diverge posteriorly.

Variation. The shape of the tegular apophysis varies slightly across sample locations. Specimens from most locations (similar to type material from Indian Cave) possess a tegular apophysis with a broad, L-shaped base and an acute tip (see Gertsch, 1984: fig. 58), whereas other specimens have a narrower base with a gradually tapering tip (Fig. 21A, C). Different specimens from the Fallen Rock Cave population exhibit both conditions (Fig. 21B, C). We observed only minor variation in epigyna (Fig. 22A–G), even for geographically disjunct southeastern Surry and Carter County populations (Figs 13, 22C–F).

Distribution and natural history. Known from both limestone caves (both shallow and deeper situations) and dark, cool, relatively moist near-surface habitats (e.g., rock piles, shallow cliff caves). Most known populations are from caves in the upper-central Appalachian Valley and Ridge, with a few peripheral montane surface populations (e.g., Surry County, NC; Carter County, TN; Raleigh County, WV). The southeastern Surry and Carter County populations appear disjunct, separated from the remainder of the species' range by regions occupied by other taxa in the species group (Fig. 13).

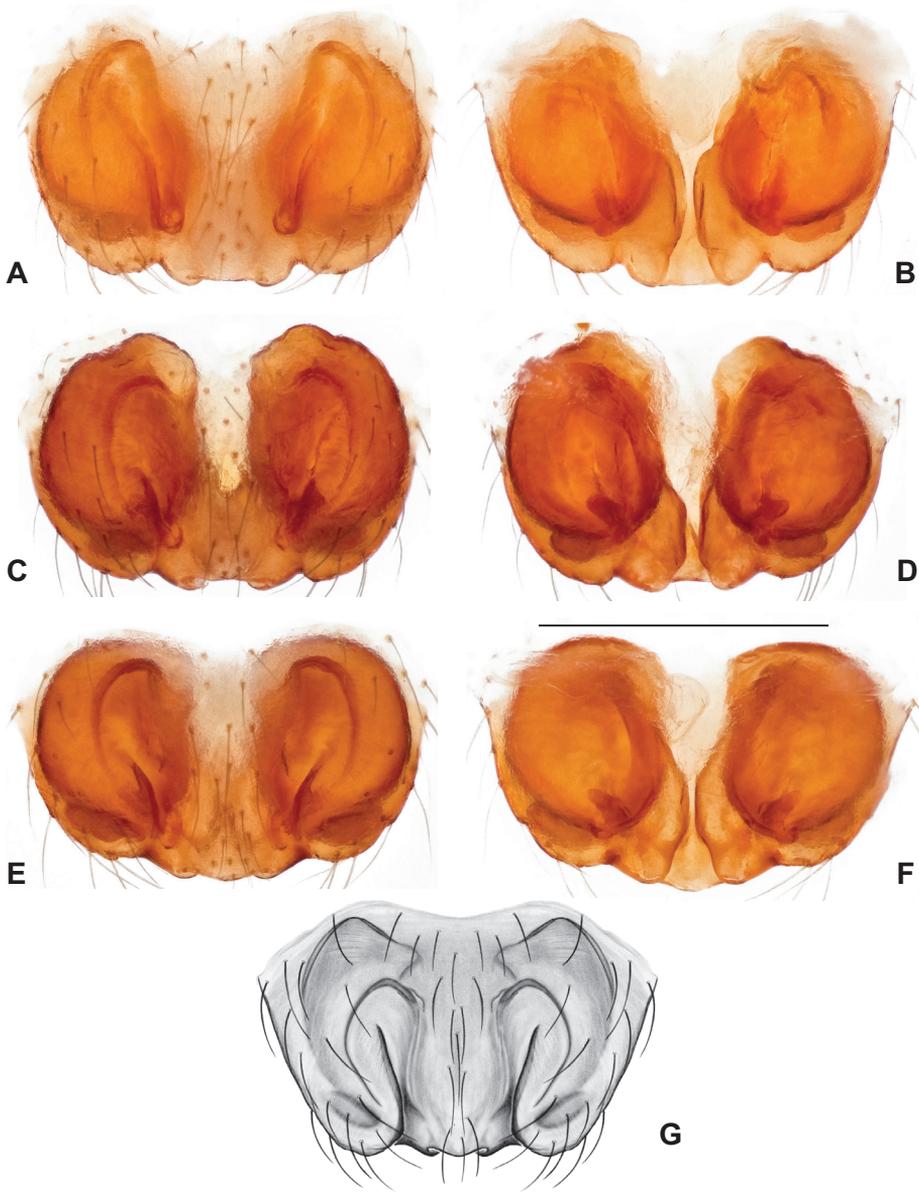


Figure 22. *Nesticus tennesseensis* epigynal variation. Tennessee, Grainger Co., Indian Cave, MCH specimen #1010, ventral (A), dorsal (B). North Carolina, Surry Co., near Fisher Peak, MCH specimen #N1126, ventral (C), dorsal (D). Tennessee, Carter Co., near Watauga Lake, MCH 05_078, ventral (E), dorsal (F). West Virginia, Raleigh Co., Grandview SP, MCH specimen #1362, ventral (G).

Remarks. Gertsch (1984: 24–25) incorrectly identified specimens from the following locations as *Nesticus tennesseensis*: Sensabaugh Saltpeter Cave (3 imm) and “Cave by Clinch River” (one ♀) specimens belong to *N. paynei*. Straley’s Cave No. 1 (2 imm), Harris Cave (1 ♀), and Potter’s Cave (1 ♂, 1 ♀) specimens belong to *N. mimus*.

Nesticus dilutus Gertsch, 1984

Fig. 23A–C

Nesticus dilutus Gertsch, 1984: 27, figs 94–96; Hedin and Dellinger 2005: 10, figs 13, 14.

Material examined. New collections from type locality: USA – Tennessee, Rhea Co. • 2♂, 10♀; Grassy Creek Cave, south of Old Washington; 23 Aug. 1992; M. Hedin, J. Hedin leg.; **Non type material:** – Rhea Co. • ♀; Starve Rock Cave (TRH7); 26 Mar. 2016; K.S. Zigler, M.L. Niemiller, N. Mann leg.; KSZ 15–566.

Diagnosis. A close morphological and genetic relative of *Nesticus tennesseensis*. This species differs most conspicuously from the former in that the basal, dorsal process of the paracymbium is absent (Hedin and Dellinger 2005, fig. 13). The tegular apophysis has a narrow, L-shaped base with a gradually tapering tip, although this condition is found in some northern populations of *N. tennesseensis* (see Fig. 21B). Epigyna very similar to *N. tennesseensis*, but possess more widely separated, pointed medial margins when viewed dorsally (Fig. 23B, C) rather than the extended parallel medial margins in *N. tennesseensis* (Fig. 22B, D, F), and the overall shorter (anterior to posterior) epigynal plate. More troglomorphic (depigmented, lacking median eyes, with proportionately long legs) than all known populations of *N. tennesseensis* (see Hedin and Dellinger 2005).

Variation. The female specimen from Starve Rock Cave has an epigynum very similar to specimens from the type locality.

Distribution and natural history. This troglomorphic taxon was previously known only from the type locality (Grassy Creek Cave), but is now known from two

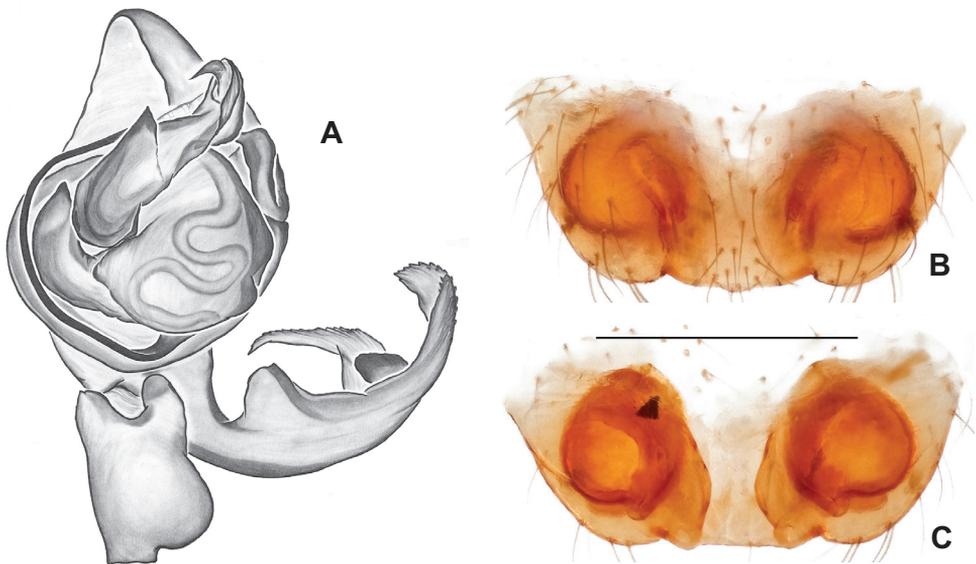


Figure 23. *Nesticus dilutus* ♂ and ♀. Tennessee, Rhea Co., Grassy Creek Cave, ♂ MCH specimen #1307, ventral (A). Tennessee, Rhea Co., Grassy Creek Cave, ♀ MCH specimen #1314, ventral (B), dorsal (C). Scale bar: 0.5 mm.

nearby caves in east-central Tennessee (Fig. 13). Starve Rock Cave is very near Grassy Creek Cave and may share a subterranean connection.

Remarks. Sister to *Nesticus tennesseensis* on UCE trees (Figs 3, 4), but *N. dilutus* sequences are embedded within a clade of *N. tennesseensis* sequences on the mitochondrial gene tree (Fig. 6). This latter pattern is attributed to either deep coalescence or gene tree estimation error.

Nesticus carolinensis (Bishop, 1950)

Figs 24A–D, 25A–F, 26

Ivesia carolinensis Bishop, 1950: 9, pl. 2, figs 1–4.

Nesticus carolinensis: Gertsch 1984: 25, figs 68–70, 88–90; Holler et al. 2020: 230.

Nesticus mimus: Gertsch 1984: 26, figs 85–87 (in part); Holler et al. 2020: 230.

Material examined. Type material: Holotype: USA – North Carolina, **McDowell Co.** • ♂ holotype; Linville Caverns, near Linville Falls; 6 Apr. 1947, S.C. Bishop leg.; AMNH; **Non type material:** – **Avery Co.** • ♂, upper slopes of Mt. Grandfather; 12 Oct. 1923; S.C. Bishop leg.; AMNH; • ♂, 6♀; Edgemont Road at Wilson Creek, 2 mi E Hwy 221; 36.0905°N, -81.8026°W; 24 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_080; • 11♀, 8 imm; Edgemont Road, 1 mile below Hwy 221; 36.0859°N, -81.815°W; 24 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_163; • ♂, ♀, 3 imm; Elk River Cave, 1 mi S Elk River Falls; 36.1892°N, -81.9617°W; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_155; • 2♀, 7 imm; Roseboro Road past first crossing of Rockhouse Creek; 36.0192°N, -81.7813°W; 24 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_164; • 3♂, 9♀; W side of Grandfather Mtn., 1 mi. NE Linville on Hwy 221; 36.0825°N, -81.8568°W; 16 Aug. 1992; M. Hedin leg.; – **Burke Co.** • ♀; Table Rock Mtn; 15 Jun. 1949, no collector information; AMNH; • 3♂, 4♀; Pine Gap Trail, W side of Linville Gorge, S of Linville Falls off Old NC 105; 35.9396°N, -81.9219°W; 16 Aug. 1992; M. Hedin leg.; • ♂, 4♀, 12 imm; Pine Gap Trail, W side of Linville Gorge, S of Linville Falls off Old NC 105; 35.9396°N, -81.9219°W; 25 Aug. 2001; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 01_165; – **Caldwell Co.** • ♀, 1 imm; Burnt Field Branch Cave; 9 May. 1995; C. Holler, C. Holler leg.; • 4♂, 4♀, 3 imm; China Creek at FR 4071 crossing, SW of Blowing Rock; 36.1151°N, -81.6983°W; 24 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_161; – **McDowell Co.** • 4♂, 10♀; Linville Caverns, S of Linville Falls, off Hwy 221N; 35.9189°N, -81.9393°W; 16 Aug. 1992; M. Hedin leg.; • 2♂, 9♀, 5 imm; Hwy 221N, N of Linville Caverns; 35.9268°N, -81.9385°W; 25 Aug. 2001; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 01_166; • 2♀; off Hwy 221N, N Linville Caverns; 35.9317°N, -81.9391°W; 24 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_081; – **Watauga Co.** • 7 imm (identification based on mitochondrial evidence); Green Mountain, Hwy 221, crossing of Green Mountain Creek; 36.1142°N, -81.7782°W; 24 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_162.

Diagnosis. Males may be differentiated from other members of this species group by the combination of palps with a paracymbium with a wide, broad ventral process,

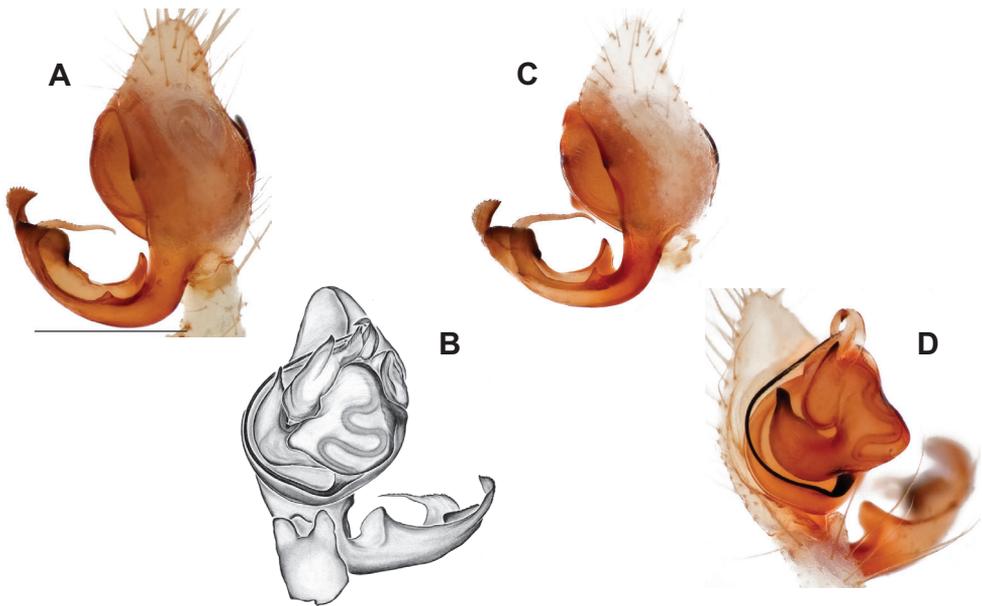


Figure 24. *Nesticus carolinensis* ♂ palps. North Carolina, Avery Co., Elk River Cave, MCH 01_155, dorsal (A), ventral (B). North Carolina, McDowell Co., Linville Caverns, MCH specimen #1225, dorsal (C), ventral (D). Scale bar: 0.5 mm.

the paradistal paracymbial process broad and triangular, a median apophysis that is a thin rectangle with an anterior sclerotized point, and a broad, singularly-pointed tegular apophysis that extends to ~ half the length of the median apophysis (Fig. 24A–D). Females may be differentiated from other members of this species group by epigyna with lateral lobes that are approximately equal to or slightly longer than the median septum, anteriorly-elongated epigynal pockets, and (viewed dorsally) touching parallel medial margins that diverge posteriorly (Fig. 25A–F).

Variation. In males from different sample locations the distal tip of the tegular apophysis varies in shape from blunt (e.g., Grandfather Mtn, Edgemont Rd) to more finger-like (e.g., N Linville Caverns, China Creek, Elk River Cave, etc.). This variation does not obviously follow geographic or phylogeographic (see below) lines. Females from different sample locations are relatively conservative in epigynal morphology (Fig. 25A–F), except for the AMNH specimen from Table Rock Mountain (see further comments below).

Fig. 26 shows an example of variation in adult female body size for specimens from a single collection location (from Edgemont Road, MCH 01_163), illustrating why we have not considered body size variation as particularly taxonomically important in this revision.

Distribution and natural history. Previously known only from caves, but quite common and abundant in suitable near-surface habitats. Mostly from the uplands between the Linville and Grandfather Mountains of western North Carolina, north-east of the Asheville Basin (Fig. 13).

Strong phylogeographic structuring is observed in the mitochondrial data, with a well-supported subclade found east of the Linville Gorge (China Creek, Green Mountain,

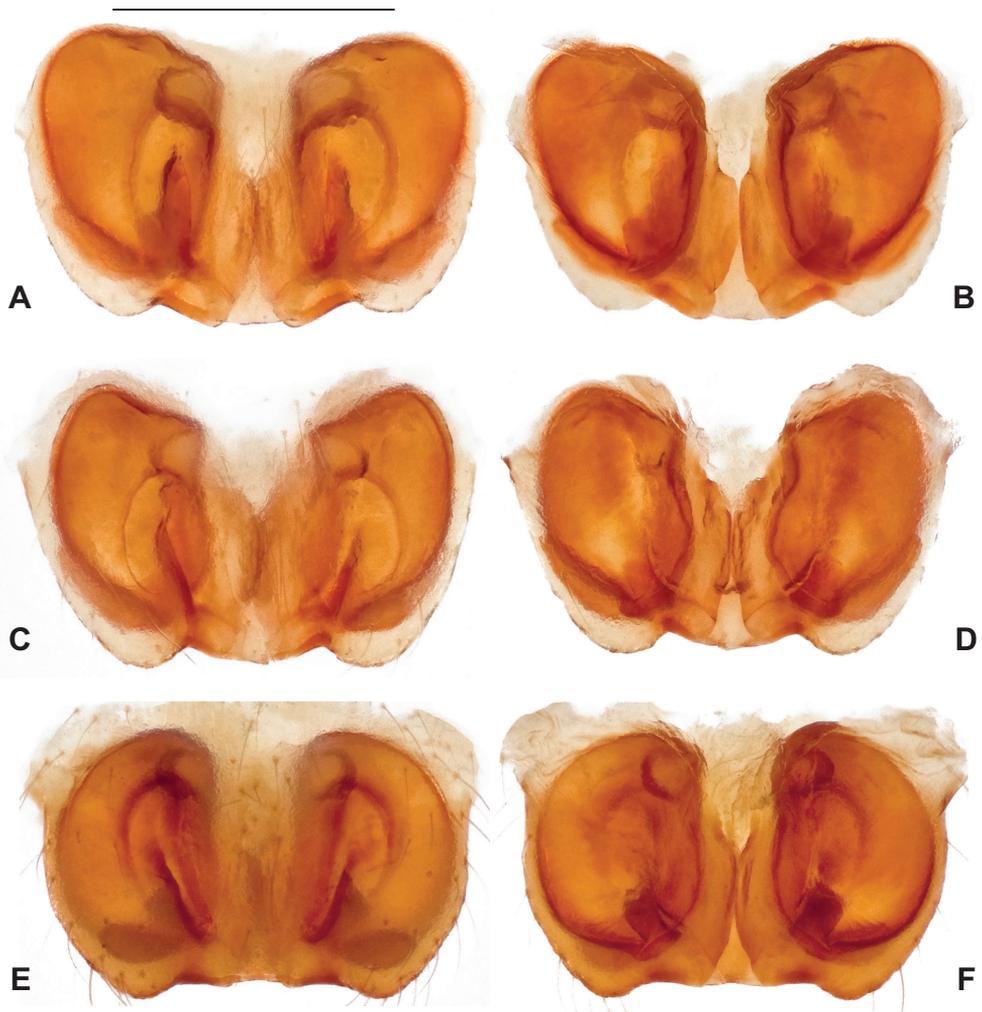


Figure 25. *Nesticus carolinensis* epigynal variation. North Carolina, McDowell Co., Linville Caverns, MCH specimen #1227, ventral (**A**), dorsal (**B**). North Carolina, Avery Co., Elk River Cave, MCH 01_155, ventral (**C**), dorsal (**D**). North Carolina, Burke Co., Table Rock Mtn. (AMNH specimen), ventral (**E**), dorsal (**F**). Scale bar: 0.5 mm.

Elk River Cave, Rockhouse Creek, etc.; Fig. 6). This phylogeographic break also corresponds to a small sampling gap (Fig. 13), so isolation by distance (with incomplete sampling) vs. isolation by geography (e.g., the Linville Gorge) cannot be distinguished.

Remarks. Gertsch (1984) provisionally attributed specimens from two montane locations to *Nesticus mimus*: a single female from Table Rock Mountain (Burke County, NC), which he described and illustrated, and a male specimen from Grandfather Mountain. The Grandfather Mountain male matches *N. carolinensis* specimens from our collections, for which we also collected DNA data. The female from Table Rock Mountain has a divergent epigynal morphology from *N. carolinensis* (wider than tall,



Figure 26. *Nesticus carolinensis* habitus images, both adult females. North Carolina, Avery Co., Edgemont Road, MCH 01_163. Scale bar: 1 mm.

short spermathecae, etc. Fig. 25E, F); we place the specimen here based mostly on geography, adjacent to our other Linville Gorge collections. It remains possible that the specimen is from north of Table Rock, closer to Watauga Lake (and locations for *N. tennesseensis*, see Fig. 22E, F).

Holler et al. (2020) cite new cave records from McDowell County. They also attribute Burnt Field Branch Cave specimens (Caldwell County) to *Nesticus mimus*, but we have examined females from this location and consider them to be *N. carolinensis*, lacking the unique spermathecae of *N. mimus*.

This species is supported as sister to *Nesticus paynei* + *N. roanensis* with a 92% bootstrap and sCF value of 37.5 on the UCE concatenated maximum likelihood tree, and a lower local posterior probability value on the UCE ASTRAL species tree (Figs 3, 4).

***Nesticus paynei* Gertsch, 1984**

Fig. 27A–K

Nesticus paynei Gertsch, 1984: 28, figs 153–155, 159–160.

Nesticus tennesseensis: Gertsch 1984: 26 (in part).

Material examined. Type material: *Holotype*: USA – Tennessee, Anderson Co. • ♂ holotype; Reeder's Cave, 2 mi. N Clinton; 10 Mar. 1965; J.A. Payne leg.; AMNH. **Non**

type material: USA – **North Carolina, Mitchell Co.** • 2♂, 3♀, 2 imm; Pigeonroost Creek, N of Nolichucky River; 36.0983°N, -82.2831°W; 21 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_147; – **Tennessee, Anderson Co.** • 2♀; Norris Dam Cave, 2 mi. N Norris; 20 Sep. 1992; M. Hedin, S. O’Kane leg.; • 3♀; Norris Dam Cave; 6 Oct. 1993; M. Hedin, C. Phillips leg.; • ♂, 3♀, 2 imm; Rieders Lost Creek Cave, TAN36; 30 May. 2016; M.L. Niemiller, E.T. Carter, N.S. Gladstone leg.; MLN 16–027.13; • ♀, 5 imm; Springhill Saltpeter Cave, TAN3; 28 Oct. 2017; M.L. Niemiller, E.T. Carter, N.S. Gladstone, K.D.K. Niemiller, C. Kendall, L. Hayter, M.J. Ravesi leg.; MLN 17–012.5; • 3♀; Weaver Cave, TAN22; 22 Mar. 2016; M.L. Niemiller, C.D.R. Stephen leg.; MLN 16–022.10; • ♂, 7 imm; Weaver Cave, TAN22; 5 Mar. 2017; N.S. Gladstone leg.; NSG 17–TAN22.5; – **Tennessee, Carter Co.** • ♂, 6♀; Grindstaff Cave, near Braemar/Hampton; 18 Sep. 1992; M. Hedin, S. O’Kane leg.; • 5♀; Grindstaff Cave; 11 Oct. 1993; M. Hedin, C. Phillips leg.; • 2♂, 2♀; Grindstaff Cave; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_152; • ♂, ♀; Rockhouse Cave, TCR3; 14 May. 2014; A.S. Engel, A. Paterson, S.W. Jones, et al. leg.; ASE 14–CR3.4; • 15♀, 1 imm; Ingram Branch Road, W of Hwy 19E/37; 36.214°N, -82.1456°W; 9 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_033; • 8♀, 7 imm; Dennis Cove Road, first crossing of Black Mtn branch above Braemer; 36.2774°N, -82.1504°W; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_154; – **Tennessee, Hamilton Co.** • ♀; Clay Cave; 24 Feb. 2013; W.T. Coleman, L. Carver, K.S. Zigler leg.; – **Tennessee, Hancock Co.** • 2♂, 8♀; Cantwell Valley Cave, SW of Sneedville; 7 Oct. 1993; M. Hedin, C. Phillips leg.; • ♀; Hwy 31 on Clinch Mountain; 36.413°N, -83.2237°W; 21 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_068; – **Tennessee, Hawkins Co.** • 4♂, 4♀; Sensabaugh Saltpeter Cave, W of Kingsport; 7 Oct. 1993; M. Hedin, C. Phillips leg.; • 3 imm; Sensabaugh Saltpeter Cave; 15 Apr. 1967; J. Holsinger leg.; AMNH; – **Tennessee, Jefferson Co.** • ♂, 4♀; Tater Cave, TJF8; 3 Aug. 2015; M.L. Niemiller, E.T. Carter, A.S. Engel, L.E. Hayter, K.D. Kendall leg.; MLN 15–016.18; – **Tennessee, Knox Co.** • 3♀, 11 imm; Blowing Hole Cave; 16 May. 2013; K.S. Zigler, M.L. Niemiller leg.; MLN 13–003; • ♂, ♀, 12 imm; Keller Bend Cave; 16 May. 2013; K.S. Zigler, M.L. Niemiller leg.; MLN 13–004; • ♂, 2♀; Kirkpatrick Cave, TKN62; 9 Feb. 2014; M.L. Niemiller, A.S. Engel, S. Engel, C. Kerr leg.; MLN 14–009; • 2♀, 3 imm; Kirkpatrick Cave, TKN62; 6 Jul. 2014; M.L. Niemiller, A.S. Engel, A Paterson leg.; MLN 14–037.10; • ♀; Pedigoe Cave, TKN103; 14 Jul. 2018; N.S. Gladstone leg.; NSG 18–TKN103.8; • 3♂, 7♀; Roaring Springs Cave, W of Copper Ridge; 6 Oct. 1993; M. Hedin, C. Phillips leg.; • ♀; Watercress Cave, TKN153; 13 Jan. 2019; N.S. Gladstone leg.; NSG 19–TKN153.16; – **Tennessee, Loudon Co.** • 2♂, 7♀, 3 imm; Ghost Cave, TLN3; 30 Aug. 2014; M.L. Niemiller, C.D.R. Stephen leg.; MLN 14–043.13; – **Tennessee, Meigs Co.** • 3♀, 2 imm; Blythe Ferry Cave, TME; 26 Jan. 2018; M.L. Niemiller, D. Pelren, J. Traxley, C.L. Barber leg.; MLN 18–004.16; – **Tennessee, Roane Co.** • ♀; “Cave by Clinch River”, AEC controlled area; 31 Jan. 1953, AMNH; – **Tennessee, Sevier Co.** • 2♂, 5♀, 3 imm; Two County Cave, TSV36; 5 Jul. 2014; M.L. Niemiller, A.S. Engel, A. Paterson leg.; MLN 14–036.13; – **Tennessee, Sullivan Co.** • 4♀; Bays Mountain Park, W of Kingsport; 36.507°N, -82.6109°W; 10 Aug.

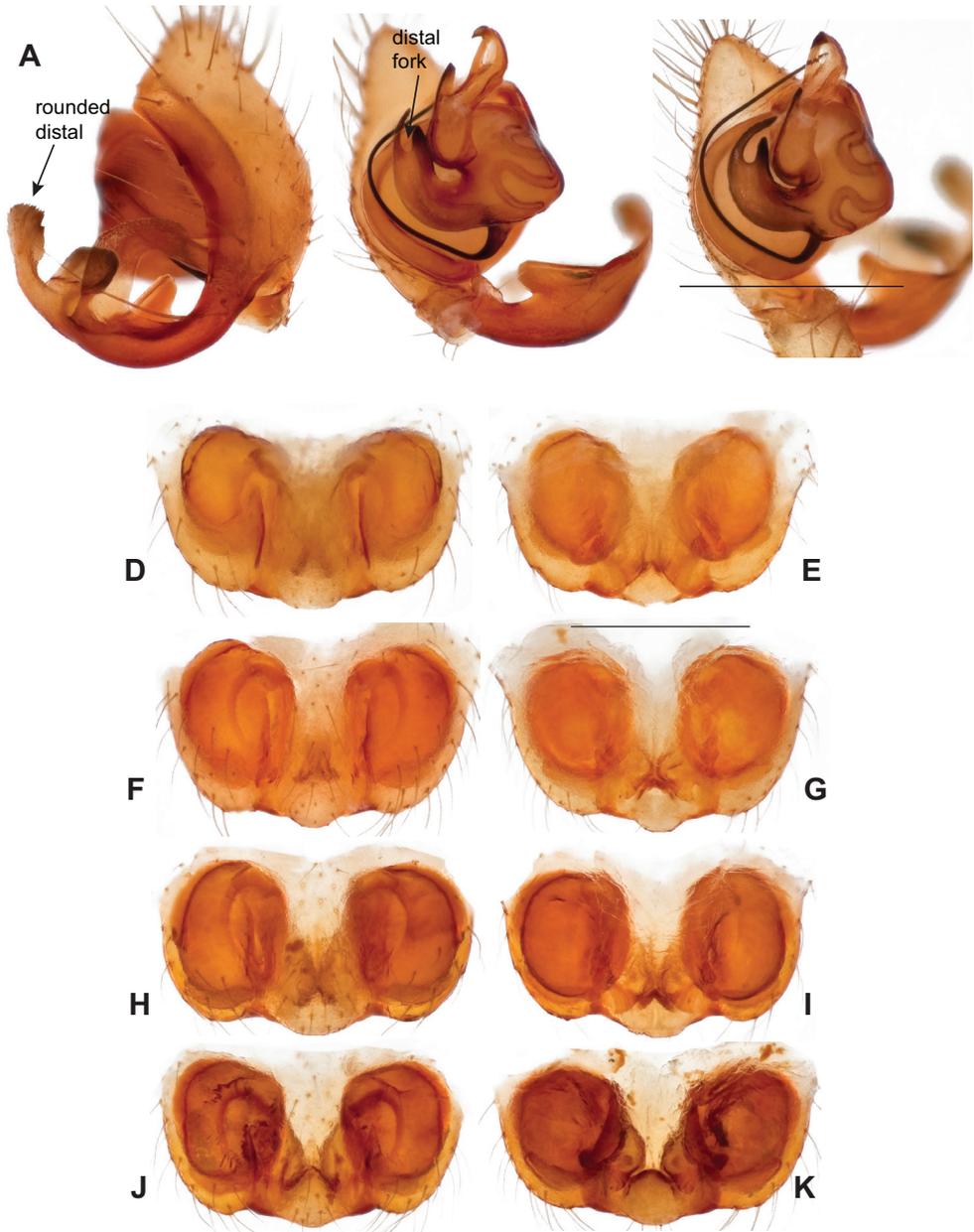


Figure 27. *Nesticus paynei* genitalia. Tennessee, Hawkins Co., Sensabaugh Saltpeter Cave, MCH ♂ specimen #1762, dorsal (A), ventral (B) C Tennessee, Unicoi Co., Rock Creek Recreational Area, MCH 04_032, palp, ventral. Epigynal variation. Tennessee, Roane Co., cave by Clinch River (AMNH specimen), ventral (D), dorsal (E). Tennessee, Hawkins Co., Sensabaugh Saltpeter Cave, MCH specimen #1765, ventral (F), dorsal (G). Tennessee, Unicoi Co., Rock Creek Recreational Area, MCH 04_032, ventral (H), dorsal (I). Tennessee, Carter Co., Ingram Branch Road, MCH 04_033, ventral (J), dorsal (K). Scale bar: 0.5 mm.

2007; M. Hedin, R. Keith leg.; MCH 07_085; • 2♂, 4♀; Eastman Recreation Area, Bays Mountain; 36.5029°N, -82.61°W; 8 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_029; • 4♀; Holston Mountain, Holston Mountain Road, 6 mi E Hwy 19E; 36.4328°N, -82.167°W; 23 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_077; – **Tennessee, Unicoi Co.** • 1 imm (identification based on mitochondrial evidence); road to Unaka Springs, along Nolichucky River, SW of Banner Hill; 36.0982°N, -82.4439°W; 22 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_146; • 2♂, 4♀; Rock Creek Recreational Area, SE of Erwin; 36.1379°N, -82.3482°W; 9 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_032; • 2♂, 3♀; Unaka Mountains, Forest Road 230, NE Unaka Mountain; 36.1396°N, -82.2837°W; 9 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_035; – **Tennessee, Union Co.** • 2♀; Coppock Cave, near Central Valley School, S Ridenour; 25 Sep. 1991; M. Hedin, K. Crandall leg.; • 2♂, 9♀; Coppock Cave; 20 Sep. 1992; M. Hedin, S. O’Kane leg.; – **Virginia, Scott Co.** • ♀; Wininger Cave; D. Hubbard leg.; • 10♀; Wolfe Cave, off Rd 629, W Hwy 23, near Speer’s Ferry; 7 Oct. 1993; M. Hedin, C. Phillips leg.

Diagnosis. Males are diagnosed from closely-related *Nesticus roanensis* by the distally split tegular apophysis (Fig. 27A, B), and from all other members of the species group by distal and dorsal paracymbial processes that are relatively rounded, vs. truncate. Females may be differentiated from other members of this species group (except *N. roanensis*) by epigyna with circular epigynal pockets, and internal plates (viewed dorsally) with touching medial margins (Fig. 27D–K).

Variation. There is minor variation in the depth of the distally bifurcate tegular apophysis (e.g., slightly deeper in neighboring Pigeonroost Creek and Rock Creek Recreation Areas specimens), but in general populations are conspicuously homogeneous despite a large and fragmented geographic distribution (Fig. 13). Female epigynal variation is minimal (Fig. 27D–K).

Distribution and natural history. Most sample locations are from limestone caves in the central part of the upper Tennessee River valley, near Knoxville, Tennessee, and extending northeast and southwest from there (Fig. 13). Some cave populations are highly disjunct, similar to the situation seen in *Nesticus tennesseensis* and *N. carteri*. While all Gertsch (1984) records for *N. paynei* are from caves, many of the new records reported here are from rockpile habitats from the mountains along the North Carolina / Tennessee border near Johnson City, Tennessee (Fig. 13). Montane samples are early branching on the UCE ASTRAL tree (Fig. 4).

Remarks. We identified spiders from Sensabaugh Saltpeter Cave (3 imm) and ‘Cave by Clinch River’ (one ♀) as *Nesticus paynei*. These collections were originally identified by Gertsch (1984: 24–25) as *N. tennesseensis*.

As discussed directly below *Nesticus paynei* is intermixed with *N. roanensis* on mitochondrial trees (Fig. 6), and these taxa are not strictly reciprocally monophyletic on concatenated UCE trees (Figs 3, 4). However, for reasons argued below we consider *N. roanensis* as distinct from *N. paynei*, consistent with our original “morphology first” hypothesis.

Because neither female morphology nor mitochondrial placement can strictly distinguish *Nesticus paynei* from *N. roanensis*, our attribution for some female-only *N. paynei* collections from near Roan Mountain is necessarily tentative. This includes Ingram Branch and Dennis Cove Road collections from north of Roan Mountain (Fig. 13; Suppl. material 1); we provisionally place these as *N. paynei* as they occur at relatively low elevations. Male specimens and/or UCE data will be important to obtain for these locations.

***Nesticus roanensis* sp. nov.**

<https://zoobank.org/AA97398C-1B2D-4E1D-B5DD-09C3DB072EFC>

Fig. 28A–G

Material examined. Type material: Holotype: USA – **North Carolina, Mitchell Co.** • ♂; Roan Mountain, below Roan High Bluff; 36.0931°N, -82.1459°W; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_150 (SDSU_TAC000675); **Paratype:** – **North Carolina, Mitchell Co.** • ♀; Roan Mountain, below Roan High Bluff; 36.0931°N, -82.1459°W; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_150 (SDSU_TAC000676); **Non type material:** USA – **North Carolina, Avery Co.** • ♂, 1 imm; Henson Creek at Henson Creek Baptist Church, on Henson Rd, N of Ingalls; 36.0374°N, -82.042°W; 21 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_138; – **North Carolina, Mitchell Co.** • 2♂, 2♀; Roan Mountain, below Roan High Bluff; 36.0931°N, -82.1459°W; 22 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_150; • ♂, 2♀, 6 imm; upper Roan Valley, Hwy 261; 36.0929°N, -82.0932°W; 21 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_148; – **Tennessee, Carter Co.** • 5♂, 15♀; Hwy 143, NE Roan Mountain, 3 mi. N Carvers Gap; 36.1184°N, -82.0818°W; 9 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_034; • 3♂, 3♀; Hwy 143, NE Roan Mountain, 3 mi. N Carvers Gap; 36.1094°N, -82.0961°W; 31 May. 2016; M. Hedin, S. Derkarabetian, J. Starrett, D. Proud leg.; MCH 16_033.

Diagnosis. Male *Nesticus roanensis* possess a distinctive fork at the base of the tegulum unlike any other species in the species group (Fig. 28A–D). Like the sister species *N. paynei* the distal end of the paracymbial dorsal process is relatively rounded, vs. truncate. Females of *N. roanensis* are very similar to females of sister species *N. paynei*.

Description of ♂ holotype (SDSU_TAC000675). Carapace dusky cream, faint dark pigment behind ocular area. Legs pale yellow / cream. Abdomen dirty pale cream with darker paired lateral pigmentation blotches. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.6, CW 1.5, abdomen length 2.25, total body length 3.85. Leg I total length 11.35 (3.05, 0.75, 3.4, 2.95, 1.2), leg formula 1423, leg I / CW ratio 7.6. Paracymbium with a knob-shaped ventral process with a sclerotized retrolateral keel, a dorsal process with a rounded serrate, distal portion, a rectangular paradistal process, and a translucent, elongate, prolaterally-directed dorsal process. Median apophysis rectangular with an anteriorly directed edge coming to a point, translucent proximal spatulate edge lying above distal regular process. Tegular process with arrowhead-like basal fork, distal

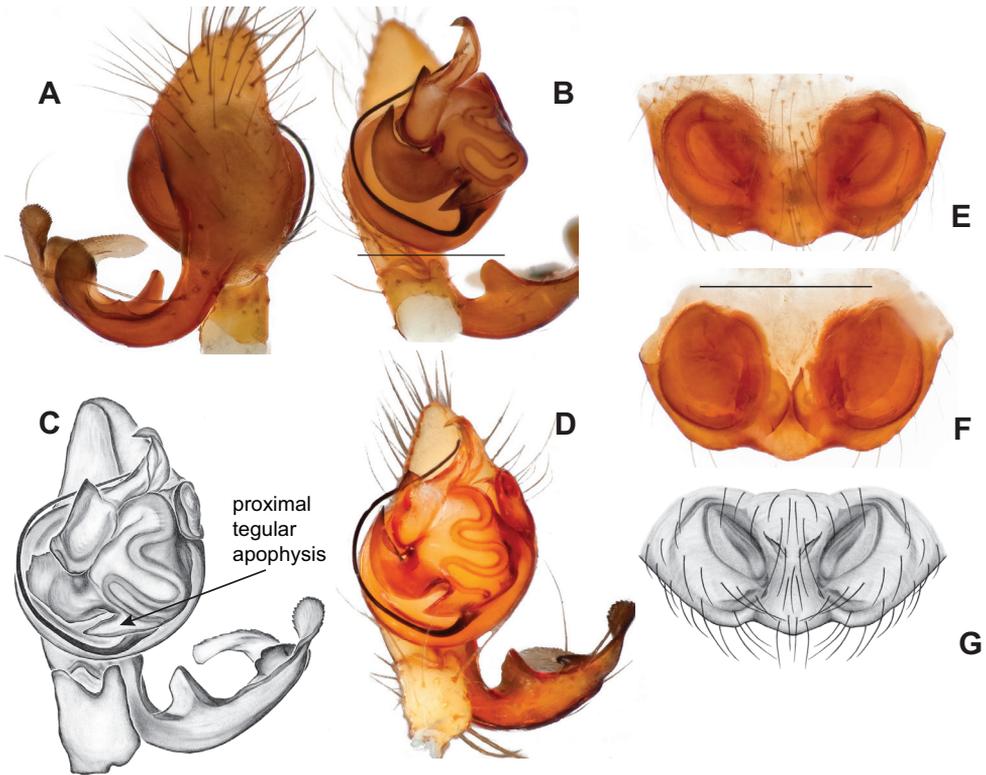


Figure 28. *Nesticus roanensis* sp. nov. genitalia. North Carolina, Mitchell Co., Roan Mountain, below Roan High Bluff, holotype male (SDSU_TAC000675) palp, dorsal (**A**), ventral (**B**) **C** North Carolina, Mitchell Co., upper Roan Valley, MCH 01_148, palp, ventral **D** North Carolina, Avery Co., Henson Creek at Henson Creek Baptist Church, MCH 07_138, palp, ventral. Epigynal variation. North Carolina, Mitchell Co., Roan Mountain, below Roan High Bluff, paratype ♀ (SDSU_TAC000675) epigynum, ventral (**E**), dorsal (**F**) **G** North Carolina, Mitchell Co., upper Roan Valley, MCH 01_148, ventral. Scale bar: 0.5 mm.

process nearly as broad as long with apical point, nose-like bulge at the base of the distal process. Distal tip of conductor bent and directed prolaterally.

♂ **Variation.** Adult males from multiple collection events, including the lower elevation Henson Creek specimens, all closely approximate the holotype male. The distal portion of the tegular apophysis for the male from upper Roan Valley (MCH 01_148) is broken (Fig. 28C).

Description of ♀ paratype (SDSU_TAC000676). Carapace dusky orange, conspicuous faint dark pigment behind ocular area. Legs pale orange. Abdomen dirty pale cream with darker paired lateral pigmentation blotches. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.5, CW 1.4, abdomen length 2.35, total body length 3.85. Leg I total length 9.35 (2.65, 0.7, 2.65, 2.3, 1.05), leg formula 1423, leg I / CW ratio 6.7. Epigynum short, wider than long. Broad proximal median septum, narrowing slightly posteriorly. Lateral to proximal septum lie obliquely oriented, oval-shaped shallow pockets

outlined by circular rings. Short, banana-shaped spermathecae visible lateral to distal septum, approximately perpendicular to septum. Viewed dorsally, circular internal lobes with interior margins bulging inwards and touching along the midline.

♀ **Variation.** The epigyna of females from multiple locations closely approximate the paratype female.

Distribution and natural history. Restricted to Roan Mountain and immediate vicinity at elevations near or above 1800 meters, except for the Henson Creek location (~ 900 meters) on the southeastern flanks of Roan Mountain (Fig. 13). At high elevations spiders were found to be reasonably common under large stones in extensive north-facing talus habitat.

We have collected comprehensively in this region, finding the sister species *Nesticus paynei* to the north and west, other *tennesseensis* group species to the east and southeast (Fig. 13), and other *Nesticus* further southwest. We believe that we have the small geographic distribution of *N. roanensis* well-circumscribed. The lower elevation Henson Creek sample location (~ 900 meters) has important conservation implications for this species, but more extensive regional sampling is needed to fully understand the distribution and abundance of this species.

Etymology. Named after the highlands of Roan Mountain along the North Carolina / Tennessee border.

Remarks. While male morphological evidence clearly supports this species as distinct in a “morphology first” framework (unique forked base of regular apophysis), the UCE phylogenomic evidence is mixed. Concatenated likelihood supports the two sampled *Nesticus roanensis* populations as monophyletic (bootstrap = 100), but nested within a larger *N. paynei* clade (Fig. 3). However, this *N. paynei* paraphyly is weakly supported, with a bootstrap value of 59 and a sCF value of only 30.5. Collapsing this node results in a topology where *N. roanensis* shares a polytomous node with *N. paynei* populations (i.e., there is not strong support for *N. paynei* paraphyly). The ASTRAL topology more clearly favors reciprocal monophyly of *N. roanensis* and *N. paynei*, the former with a posterior probability of 0.99, the latter with a posterior probability of 1.0 (Fig. 4). This recovered monophyly, in combination with morphological diagnosability, would be consistent with our species criteria.

Mitochondrial data fail to support *Nesticus roanensis* as distinct from *N. paynei* (Fig. 6), with *N. roanensis* haplotypes intermixed with *N. paynei* haplotypes, and sometimes sharing nearly identical haplotypes. Because these taxa are closely parapatric it is possible that this reflects mitochondrial introgression at areas of contact on the northern slopes of Roan Mountain (Fig. 13). A combination of introgression and incomplete lineage sorting (or ILS alone) is also a possibility.

Overall, this taxonomic situation illustrates patterns of nuclear vs. mitochondrial vs. morphological discordance as also found elsewhere in Appalachian *Nesticus*. The male morphology of *N. roanensis* is as divergent as any taxon in the species group (Fig. 12C), female morphology and mitochondrial haplotypes are shared with *N. paynei*, while nuclear phylogenomic divergence is mixed. Rates of evolution in these different character classes appear to vary in this group of populations.

nasicus* group, including:Nesticus nasicus* Coyle & McGarity, 1992*Nesticus brimleyi* Gertsch, 1984*Nesticus templetoni* sp. nov.*Nesticus crosbyi* Gertsch, 1984*Nesticus gertschi* Coyle & McGarity, 1992*Nesticus secretus* Gertsch, 1984*Nesticus canei* sp. nov.

A species group strongly supported by nuclear phylogenomics (Figs 3, 4), with *Nesticus nasicus* sister to all other taxa in the group. This group is not recovered as monophyletic with mitochondrial data, with a monophyletic *N. nasicus* separate from a clade that includes remaining group members (Fig. 6). Four taxa, including *N. crosbyi*, *N. secretus*, *N. gertschi* and *N. canei* sp. nov. form a close-knit morphological and phylogenetic subgroup within the more inclusive species group. Mitochondrial relationships within this subgroup are discordant with nuclear and morphological evidence (see below).

Coyle and McGarity (1992) recognized a close morphological relationship between *Nesticus nasicus* and the previously described *N. brimleyi*, citing several shared male and female characters. They also commented on a possible relationship of these two species with *N. gertschi*, citing the “broad, translucent, spatulate, distal paracymbial process” as a possible defining feature for these three taxa (in a group they never formally named). Coyle and McGarity (1992) did not comment on the inclusion of previously described *N. secretus* Gertsch, 1984 or *N. crosbyi* in this group. We agree in recognizing the translucent, spatulate, distal paracymbial process as a defining feature for the entire species group (Fig. 29A–F). Also, the epigynum viewed ventrally is characterized by a protruding nose-like median septum bordered by prominent pockets (Fig. 29G–M).

Species of the *nasicus* group are distributed in the montane southern Blue Ridge, both west (*N. nasicus*) and east (*N. brimleyi*, *N. templetoni*, *N. gertschi*, *N. canei*, *N. crosbyi*) of the Asheville Basin (Fig. 30). The geographic origin of *N. secretus*, perhaps not surprisingly, remains a secret. We hypothesize a possible geographic origin in the English or Green Mountains (see below and Fig. 30).

***Nesticus nasicus* Coyle & McGarity, 1992**

Figs 31A–D, 32A–H

Nesticus nasicus Coyle & McGarity, 1992: 162, figs 1–4, 7–14.

Material examined. Type material: *Holotype*: USA – North Carolina, Jackson Co. • ♂ holotype; 1 mile W of Dillsboro at Cowee Mountain Train Tunnel, rock bank; 28 Oct. 1990; T McGarity leg. AMNH; **New collections from type locality:** – Jackson Co. • ♂,

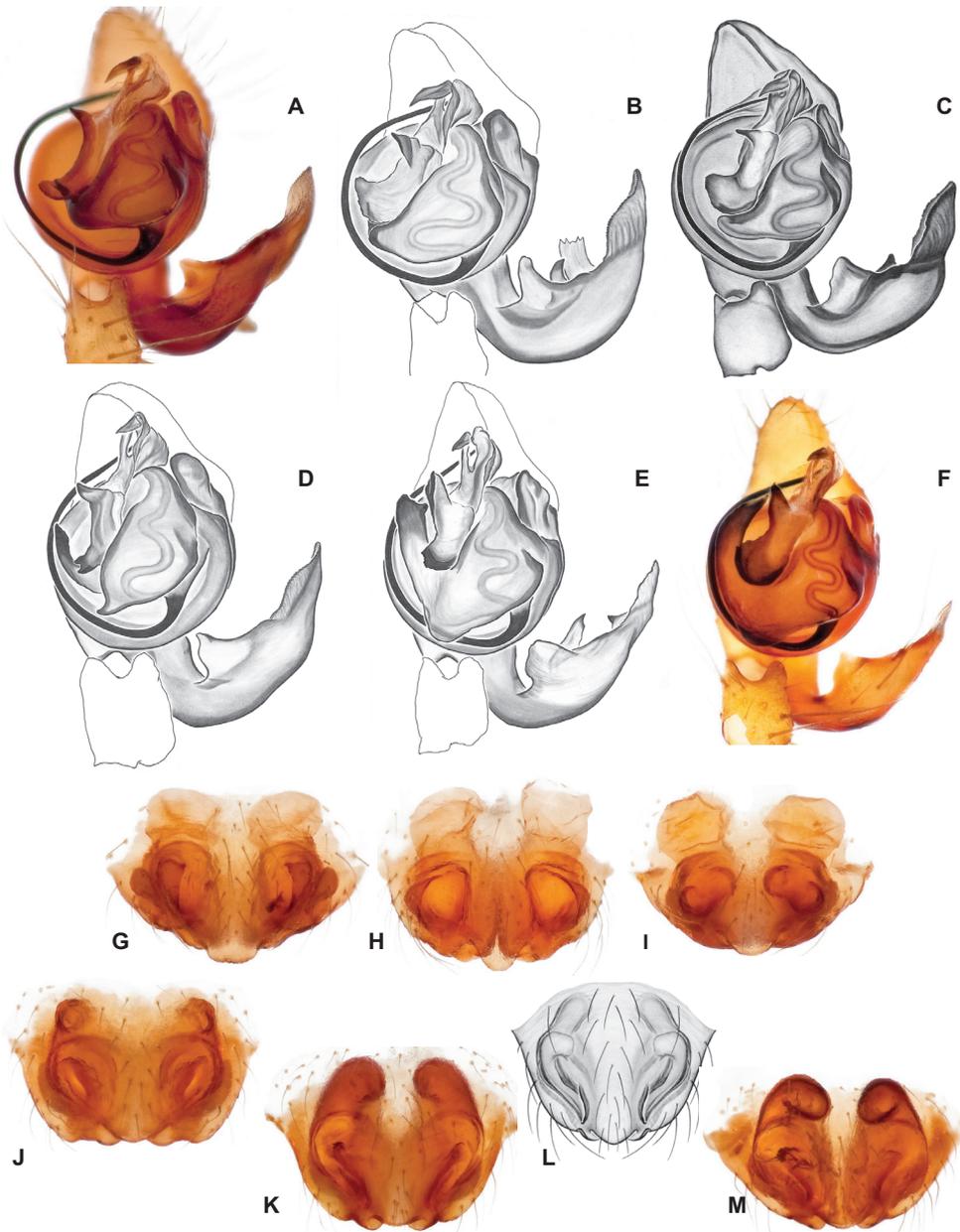


Figure 29. Comparative ♂♀ genitalia of *nasicus* group species; ♂ **A** *Nesticus nasicus* **B** *N. brimleyi* **C** *N. templetoni* **D** *N. crosbyi* **E** *N. gertschi* **F** *N. canei*; ♀ **G** *N. nasicus* **H** *N. brimleyi* **I** *N. templetoni* **J** *N. crosbyi* **K** *N. gertschi* **L** *N. secretus* **M** *N. canei*. All views ventral. See subsequent figures for specimen locations and voucher details.

17♀; Cowee Mountain Train Tunnel, NW of Dillsboro; 35.3768°N, -83.268°W; 14 Aug. 1992; M. Hedin leg. **Non type material:** – **Buncombe Co.** • ♀; NE Mt. Pisgah, Hwy 151, head of McKinney Creek; 35.4448°N, -82.7225°W; 5 Sep. 2002; M. Hedin, M.

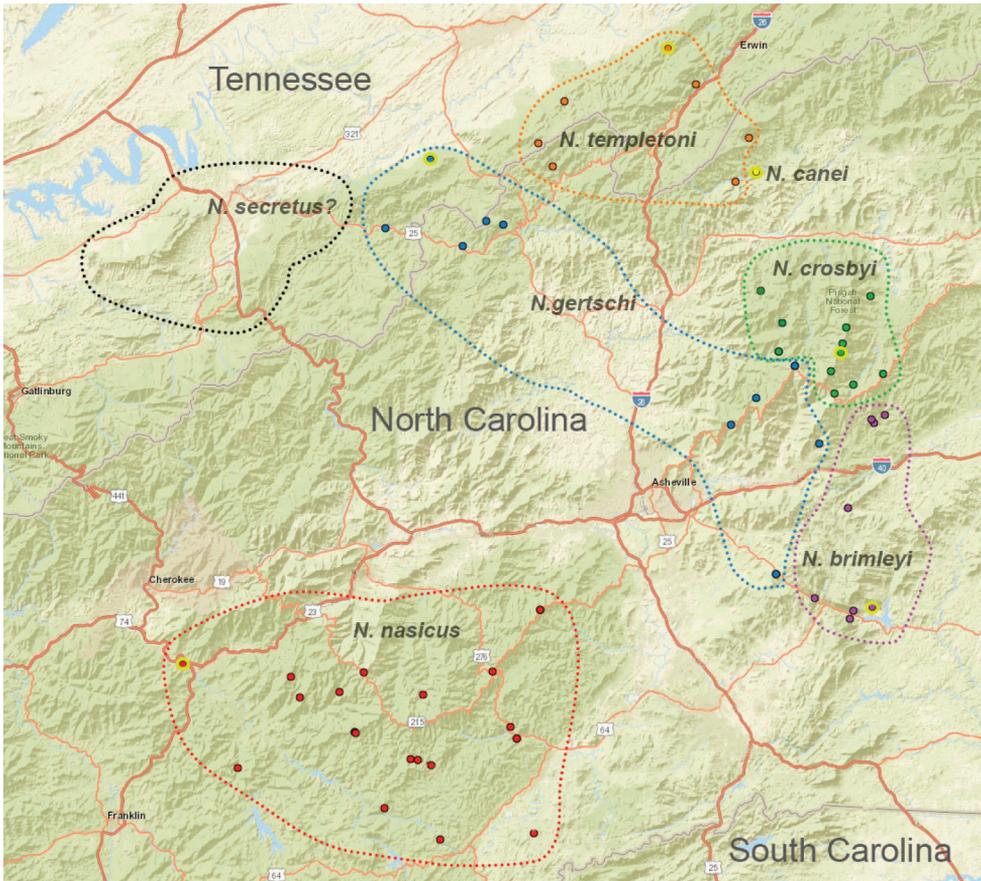


Figure 30. Distribution of *nasicus* group species. Type localities designated with yellow circles. Primary cities shown for geographic context. Dashed lines circumscribe known species distributions; possible distribution of *Nesticus secretus* is tentative, see text for details.

Lower, P. Paquin leg.; MCH 02_194; • ♀; NE Mt. Pisgah, Hwy 151, head of McKinney Creek; 35.4448°N, -82.7225°W; 22 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_074; – **Haywood Co.** • ♀; Blue Ridge Parkway, vicinity Richland Balsam; 35.3666°N, -82.9915°W; 4 Aug. 1992; F. Coyle leg.; • 4♂, 2♀, 7 imm; Hwy 215, along West Fork Pigeon River; 35.339°N, -82.9016°W; 4 Sep. 2002; M. Hedin, F. Coyle, P. Paquin leg.; MCH 02_190; • 2♂, 8♀, 5 imm; Hwy 276, N Pigeon Gap; 35.3677°N, -82.7958°W; 4 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_193; – **Jackson Co.** • 3♀, 1 imm; Coward Mountain near Jackie Spring Gap; 35.3352°N, -83.0897°W; 1 Sep. 2002; M. Hedin, P. Paquin leg.; MCH 02_178; • 4♀, 1 imm; Coward Mountain, E Wolfpen Gap; 35.3606°N, -83.1037°W; 1 Sep. 2002; M. Hedin, M. Lowder leg.; MCH 02_177; • 9♀; Mull Creek on Caney Fork Road, 11 mi. E Hwy 107; 35.3417°N, -83.0292°W; 11 Aug. 1992; M. Hedin leg.; • 2♀; Rich Mountain, SE Sugar Creek Gap; 35.2907°N, -83.004°W; 1 Sep. 2002; M. Hedin, P. Paquin leg.; MCH 02_179; • ♂; SW of Rich Mountain Bald, 0.5 mi E Sugar Creek Gap; 35.2915°N, -83.006°W; 26 Jun.

1992; B. Dellinger leg.; • ♀; SW of Rich Mountain Bald, 0.5 mi E Sugar Creek Gap; 35.2915°N, -83.006°W; 17 Apr. 1994; M. Hedin, B. Dellinger leg.; • 7♀; Wolf Creek at Cullowhee Creek, off Cullowhee Mountain Road; 35.2468°N, -83.1843°W; 11 Aug. 1992; M. Hedin leg.; – **Transylvania Co.** • ♀; along West Fork French Broad River, Silverstein Road, 2 mi. N Hwy 64; 35.1573°N, -82.8758°W; 19 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_128; • ♀; below Conneestee Falls, off Hwy 276 S Brevard; 35.1647°N, -82.7319°W; 2 Oct. 1992; B. Dellinger leg.; • ♂, 2♀; Hwy 215, 4.7 mi. NW Balsam Grove along Bald Knob branch; 35.2568°N, -82.9098°W; 13 Aug. 1992; M. Hedin leg.; • 5♀; Hwy 215, S Pinhook Gap; 35.2575°N, -82.9204°W; 22 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_073; • 2♀, 1 imm; Hwy 276 at Davidson River, opposite Stillwater Branch; 35.284°N, -82.7591°W; 28 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_180; • 4♀; Hwy 276 at Davidson River, opposite Stillwater Branch; 35.284°N, -82.7591°W; 4 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_192; • ♀; Hwy 276, Looking Glass Creek just N Looking Glass Falls; 35.2978°N, -82.7676°W; 19 Aug. 2007; M. McCormack, S. Derkarabetian leg.; MCH 07_129; • ♀; Hwy 281, E Owens Gap; 35.1957°N, -82.9608°W; 20 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_071; • ♀, 1 imm; N Fork French Broad, FR 140 off Hwy 215; 35.2503°N, -82.889°W; 4 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_189.

Diagnosis. Males may be distinguished from other members of the species group by the palp with a uniquely shaped tegular apophysis (except in comparison to *Nesticus brimleyi*), combined with a narrow-based paracymbial dorsal process (Fig. 31A, C) which differs greatly from *N. brimleyi*. The *N. nasicus* epigynum is very

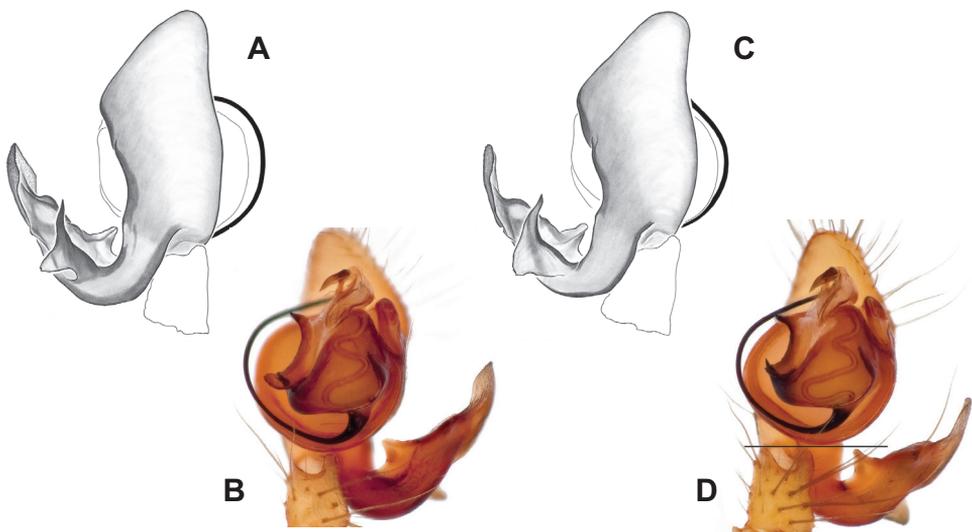


Figure 31. *Nesticus nasicus* ♂ palps. North Carolina, Transylvania Co., Hwy 215, NW of Balsam Grove, MCH specimen #1155, dorsal (A), ventral (B). North Carolina, Haywood Co., along West Fork Pigeon River, MCH 02_190, dorsal (C), ventral (D). Scale bar: 0.5 mm.

similar to *N. brimleyi* and *N. templetoni*. When viewed dorsally all possess “crinkled” sac-shaped structures above (anterior to) the main epigynal plate, which we hypothesize are homologous to vulval pockets (Vp) as seen in Japanese *Nesticus* (Suzuki and Ballarin 2020). Further diagnostic features for *N. nasicus* are discussed in Coyle and McGarity (1992).

Variation. Notable variation exists in the shape of the dorsal process of the paracymbium, which is sometimes narrow and finger-like (Coyle and McGarity 1992: figs 1, 2), or fishtailed (Fig. 31A–D), with variation in the shape of the end of the process. Both the distomedial and dorsomedial paracymbial processes vary in presence across populations, with a distomedial process found in West Fork Pigeon River and Cowee Mountain males (Coyle and McGarity 1992: fig. 2), and a dorsomedial process found only in Cowee Mountain males; these processes are lacking in males from other populations. As discussed below similar population-level variation in these processes is observed in *Nesticus templetoni*. The shape of both the lateral and apical processes of the median apophysis also varies across populations (Fig. 31A–D).

Epigyna vary across sample locations in the length of the projection of the median septum, the shape of the epigynal pockets (though generally spherical), the width of epigynal pocket lateral hoods, and the length of the spermathecae (Fig. 32A–H).

Distribution and natural history. Previously known only from two locations but now known to be reasonably widespread in the Great Balsam and Pisgah Mountains southwest of Asheville North Carolina, west of the Asheville Basin (Fig. 30). The southeastern Connestee Falls population, found east of the French Broad River, is a geographic outlier.

Remarks. No obvious phylogeographic trends are apparent in the mitochondrial data, with geographically separate locations seemingly less genetically divergent than in other similarly widespread taxa (Fig. 6). This is particularly striking considering the notable male morphological variation observed across populations.

Nesticus brimleyi Gertsch, 1984

Figs 33A–C, 34A–H

Nesticus brimleyi Gertsch, 1984: 30, figs 126–128, 138–140; Coyle and McGarity 1992: figs 5, 6; Holler et al. 2020: 230.

Material examined. Type material: *Holotype*: USA – North Carolina, Rutherford Co. • ♂ holotype; Rumbling Bald Cave, Lake Lure, Rumbling Bald Mountain; 2 Jul. 1977; P. Hertl leg; AMNH; **New collections from near type locality.** – Rutherford Co. • 2♂, 6♀; SE side of Rumbling Bald Mountain, N of Lake Lure; 35.4487°N, -82.2167°W; 18 Aug. 1992; M. Hedin leg. **Non type material:** – Henderson Co. • ♀; Hwy 74 along Hickory Creek, E of Bearwallow; 35.4591°N, -82.3035°W; 20 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_134; – McDowell Co. • ♀; headwaters of Crooked Creek, Mt. Hebron Road, N of Cross Mountain;

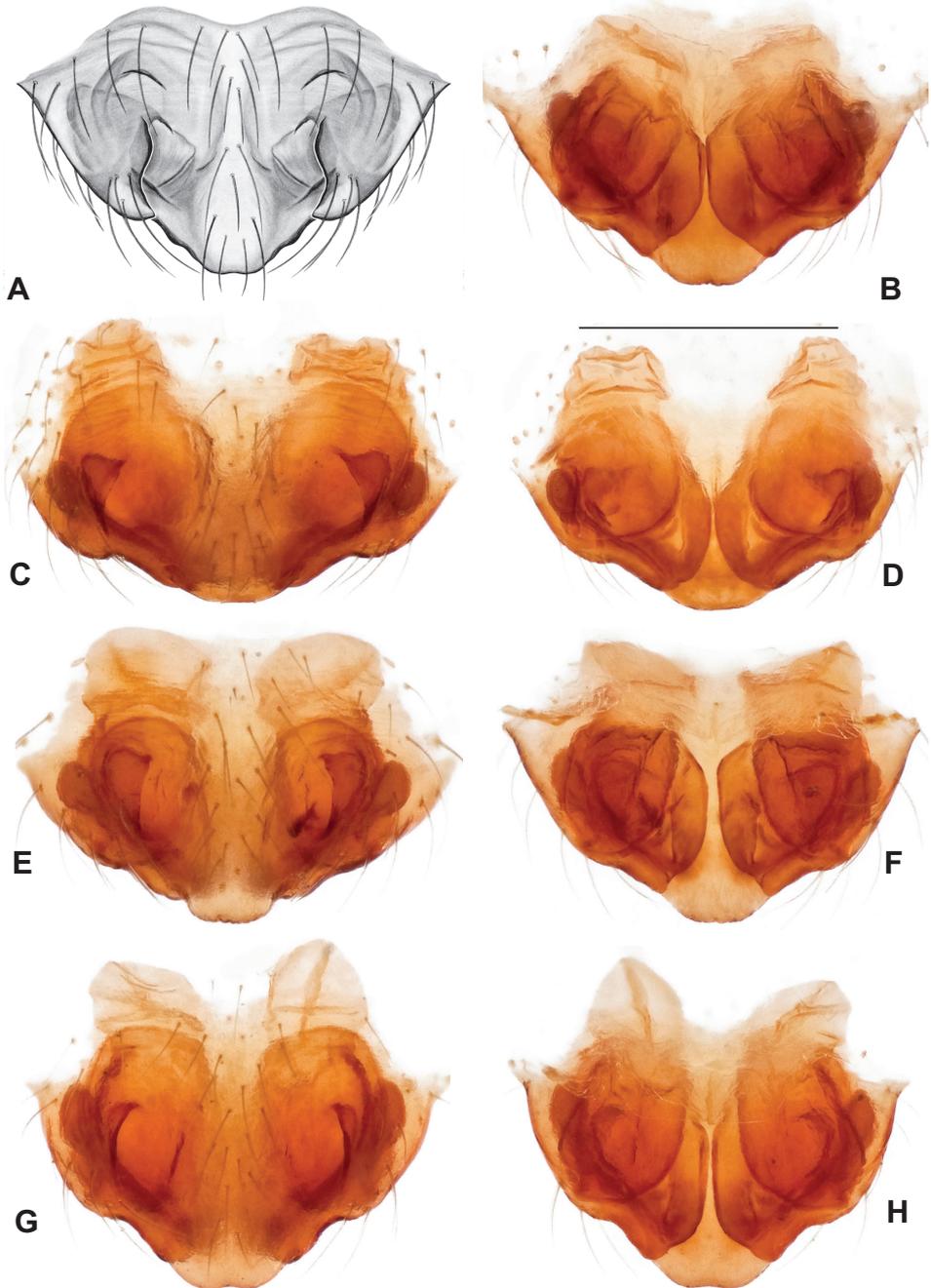


Figure 32. *Nesticus nasicus* epigynal variation. North Carolina, Haywood Co., along West Fork Pigeon River, MCH 02_190, ventral (A), dorsal (B). North Carolina, Jackson Co., Coward Mountain, E of Wolfpen Gap, MCH 02_177, ventral (C), dorsal (D). North Carolina, Transylvania Co., Hwy 276 at Davidson River, MCH 02_192, ventral (E), dorsal (F). North Carolina, Transylvania Co., Hwy 215, NW of Balsam Grove, MCH specimen #1157, ventral (G), dorsal (H). Scale bar: 0.5 mm.

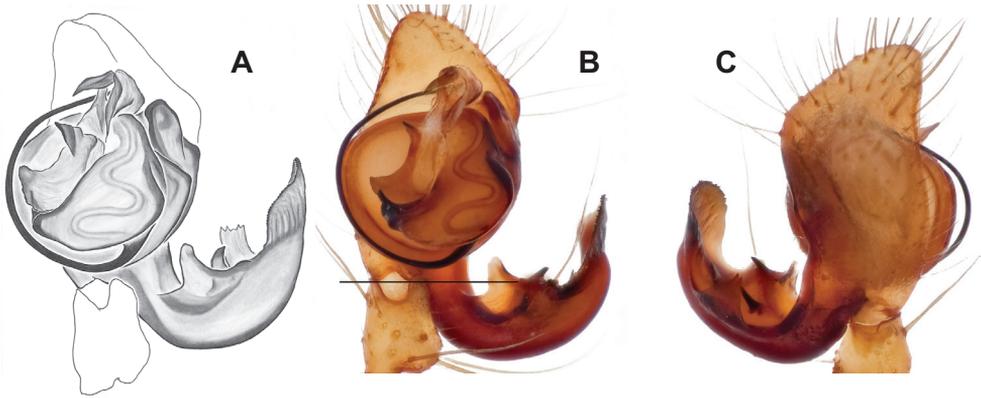


Figure 33. *Nesticus brimleyi* ♂ palps **A** North Carolina, Rutherford Co., SE side of Rumbling Bald Mountain, MCH specimen #1247, ventral. North Carolina, Henderson Co., Newberry Creek, MCH 04_075, palp, ventral (**B**), dorsal (**C**). Scale bar: 0.5 mm.

35.5726°N, -82.2532°W; 20 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_135; • ♀; near Curtis Creek campground, FR 482, N of Old Fort; 35.6889°N, -82.1976°W; 20 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_136; • 2♀; Newberry Creek above Horse branch, N of Old Fort; 35.6825°N, -82.217°W; 20 Aug. 2001; M. Hedin, M. Lowder, R. McClanahan leg.; MCH 01_141; • ♂, 11♀; Newberry Creek, N of Old Fort; 35.6789°N, -82.214°W; 22 Aug. 2004; M. Hedin, R. Keith J. Starrett, S. Thomas leg.; MCH 04_075; – **Rutherford Co.** • ♂, ♀; Chimney Rock Park, Moonshiner’s Cave; 5 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_014; • ♀; S side Round Top Mountain, just N of Chimney Rock; 35.4439°N, -82.2451°W; 5 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_015.

Diagnosis. Male paracymbium with three medial processes that lie between the ventral and dorsal processes, including ventromedial, distomedial, and dorsomedial processes (Fig. 2D.; see also Coyle and McGarity 1992: figs 4, 5). We have not seen populations of other species that simultaneously include all three processes. Also, a distally-thin tegular apophysis projects beneath the median apophysis (Fig. 33A–C). The epigynum is very similar to that of *Nesticus templetoni* (compare Fig. 34A–H to Fig. 36A–J).

Variation. In the northern Newberry Creek population the male distomedial process is reduced (but present as low spikes), and the base of the dorsal paracymbial processes is wider then narrows to a forked tip (Fig. 33A–C). Epigynal variation is limited, even across northern vs. southern disjunct populations (Fig. 34A–H). Described by Gertsch as a “*pale cavernicole*”, but many populations are from boulderfield void spaces, and most specimens are not pale.

Distribution and natural history. Previously known only from fissure caves, including those summarized by Holler et al. (2020) from Polk and Rutherford counties. Included here are many new records from near surface populations, including new

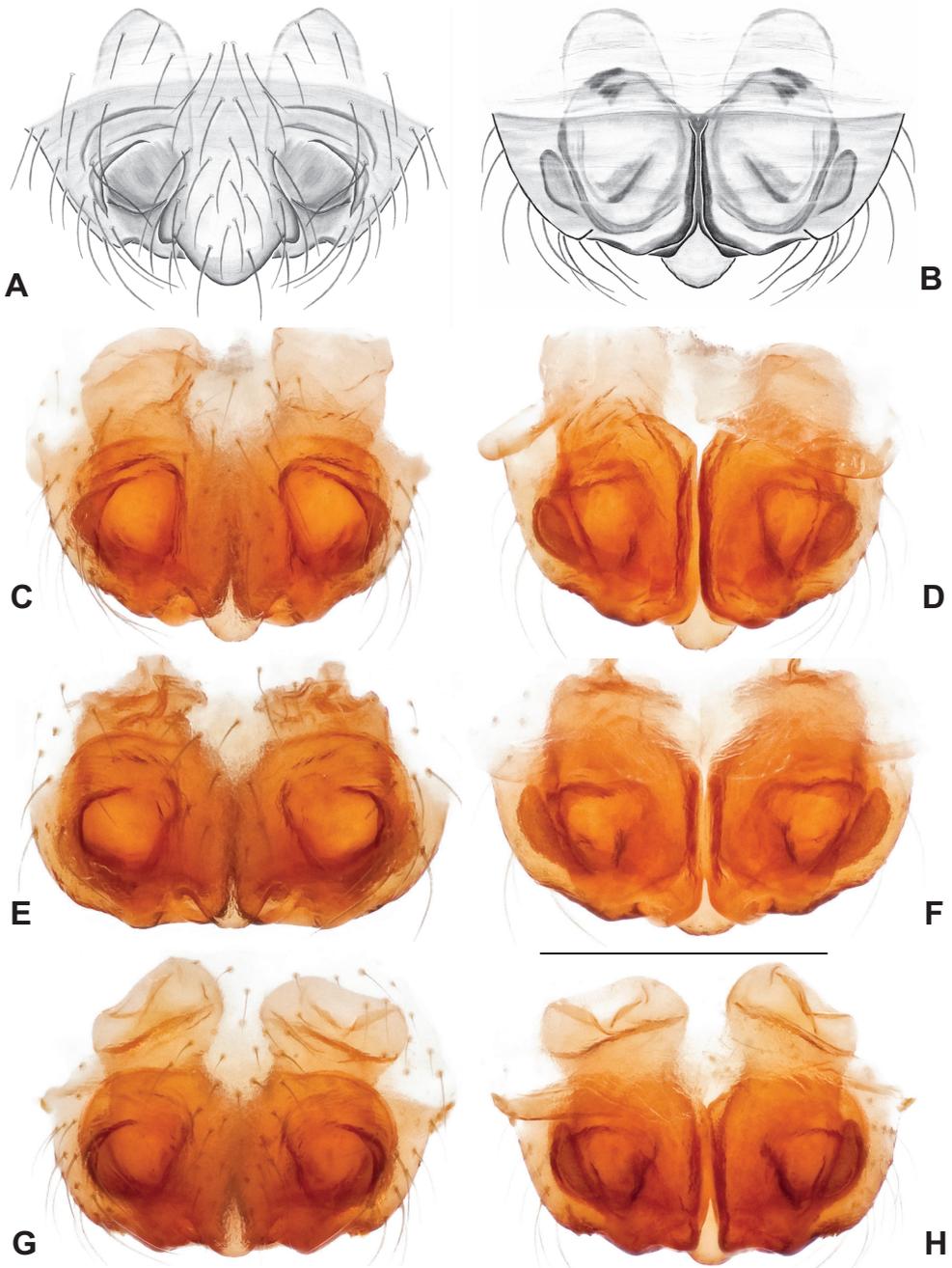


Figure 34. *Nesticus brimleyi* epigynal variation. North Carolina, Rutherford Co., SE side of Rumbling Bald Mountain, MCH specimen #1254, ventral (**A**), dorsal (**B**). North Carolina, Henderson Co., W of Bat Cave, MCH 07_134, ventral (**C**), dorsal (**D**). North Carolina, McDowell Co., headwaters of Crooked Creek, MCH 07_135, ventral (**E**), dorsal (**F**). North Carolina, Henderson Co., Newberry Creek, MCH 04_075, ventral (**G**), dorsal (**H**). Scale bar: 0.5 mm.

northern records from Henderson and McDowell Counties (Fig. 30). For example, at Newberry Creek (MCH 04_075), spiders were “common in a ... well shaded hemlock/rhododendron” boulderfield. The northwards distributional extension, and demonstration of an overall larger geographic and microhabitat distribution, has important conservation implications for this species.

Remarks. *Nesticus brimleyi* is strongly supported by nuclear phylogenomics as sister to *N. templetoni* but is geographically separated from this species by highlands occupied by other members of the species group (*N. gertschi*, *N. crosbyi*, and *N. canei*; Fig. 30). The mitochondrial gene tree includes two strongly supported geographic subclades (Fig. 6), corresponding to southern (Broad River drainage) versus northern *N. brimleyi* populations (Fig. 30).

***Nesticus templetoni* sp. nov.**

<https://zoobank.org/5AB0E873-4666-4B6A-A213-72A3B24D978C>

Figs 35A–H, 36A–J

Material examined. Type material: *Holotype*: USA – **Tennessee, Unicoi Co.** • ♂ holotype; Rich Mountain, Clarks Creek; 36.1457°N, -82.5278°W; 10 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_036 (SDSU_TAC000669); ***Paratypes***.– **Tennessee, Unicoi Co.** • ♂, ♀; Rich Mountain, Clarks Creek; 36.1457°N, -82.5278°W; 10 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_036; **Non type material:** – **North Carolina, Madison Co.** • ♂, 2♀; East Prong Hickory Fork Creek, off Hwy 212; 35.999°N, -82.7033°W; 21 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_144; – **North Carolina, Yancey Co.** • ♀; E Spivey Gap, Hwy 19W, along Big Creek, NW of Sioux; 36.0342°N, -82.4043°W; 21 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_146; • 4♂, 2♀; Scronce Creek Road, W of Bee Log; 35.9805°N, -82.4245°W; 22 Oct. 2012; M. Hedin, J. Bond, F. Coyle leg.; MCH 12_141; – **Tennessee, Greene Co.** • 2♂, 6♀; Bald Mountain Road, NW Camp Creek Bald; 36.0284°N, -82.7253°W; 10 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_038; • 7♀, 10 imm; Bald Mountains, E Greystone Mountain, Round Knob Road; 36.0799°N, -82.6859°W; 10 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_037; – **Tennessee, Unicoi Co.** • ♂, 2♀; along Mill Creek, Mill Creek Road on Rich Mountain, NE of Ernestville; 36.1018°N, -82.4859°W; 22 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_147; • 13♀, 3 imm; Rich Mountain, Clarks Creek; 36.1457°N, -82.5278°W; 10 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_036.

Diagnosis. In comparison to its sister species *Nesticus brimleyi* (see above), males of *N. templetoni* can be diagnosed by a shortened tegular apophysis (of variable shape) with a small, sclerotized extension lying behind the lateral process of the median apophysis, and never possessing all three medial paracymbial processes (Fig. 35A–H). The epigynum is very similar to that of *N. brimleyi*, with epigynal pockets in the latter generally more circular with stronger lateral hoods (Fig. 34A–H).

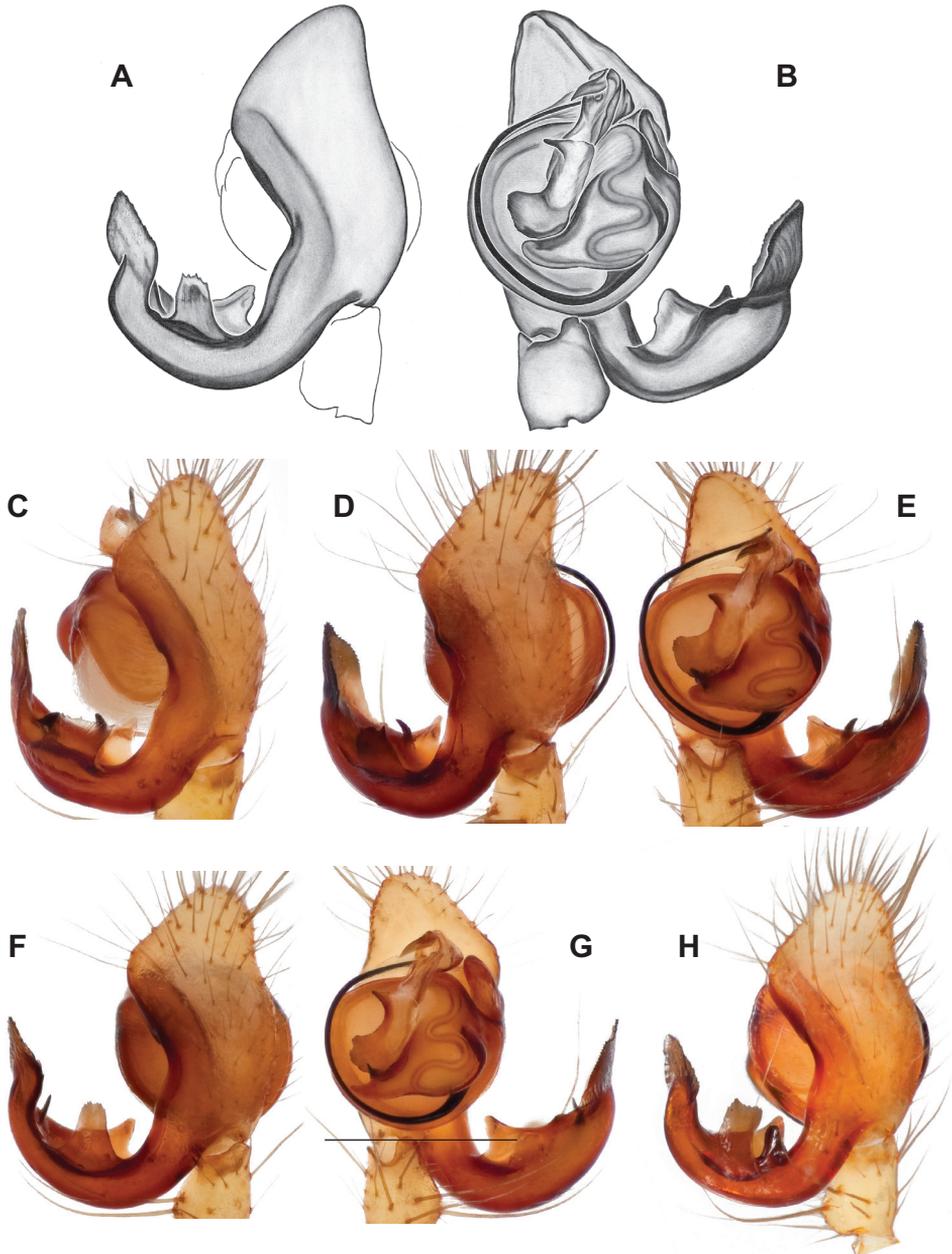


Figure 35. *Nesticus templetoni* sp. nov. ♂ palps. North Carolina, Unicoi Co., Rich Mountain, Clark Creek, MCH 04_036 (SDSU_TAC000669), dorsal (**A**), ventral (**B**) **C** Tennessee, Greene Co., Bald Mountain Road, MCH 04_038, dorsal. Tennessee, Unicoi Co., along Mill Creek, MCH 07_147, dorsal (**D**), ventral (**E**). North Carolina, Madison Co., East Prong Hickory Fork Creek, MCH 01_144, dorsal (**F**), ventral (**G**). North Carolina, Yancey Co., Scronce Creek Road, MCH 12_141, dorsal (**H**). Scale bar: 0.5 mm.

Description of ♂ holotype (SDSU_TAC000669). Carapace cream-colored, very faint pigment in ocular area. Legs pale yellow to cream. Abdomen mostly pale cream, faint paired lateral pigmentation blotches. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.25, CW 1.1, abdomen length 1.6, total body length 2.85. Leg I total length 9.85 (2.75, 0.5, 3, 2.6, 1), leg formula 1423, leg I / CW ratio 9.0. Palp with shoe-shaped tegular apophysis, with small dark sclerotized extension lying behind lateral process of median apophysis. Lateral process of median apophysis itself concave, broadening and well-sclerotized along edge, distal process drawn into thin tip. Ventral process of paracymbium translucent and triangular, distal process spatulate (consistent with species group), dorsal process wide at base, translucent, relatively short. Short, dark, conspicuous ventromedial process (Fig. 35A).

♂ **Variation.** Extensive population-level variation is seen in the male palps across relatively short geographic distances in this species. This includes variation in the shape of the shoe-shaped tegular apophysis and the sclerotized extension, the presence and shape of the paracymbial ventromedial and distomedial processes, and the shape of the dorsal paracymbial process (Fig. 35A–H). Mill Creek males approximate type males (Fig. 35D, E). Western Bald Mountain Road males possess a dorsal process that is nearly square in shape and includes a unique distal spike, with both ventromedial and distomedial paracymbial processes (Fig. 35C). Northwestern Hickory Fork Creek males only possess a distomedial paracymbial process (Fig. 35F, G). Southern Scronce Creek males lack ventro- and distomedial processes altogether and possess a dorsal process that is particularly wide at the base with a unique basal sclerotized extension (Fig. 35H), perhaps representing a dorsomedial process that has migrated to the edge of the paracymbium.

Description of ♀ paratype (SDSU_TAC000670). Carapace color as in male. Legs pale yellow to cream. Abdomen with paired, lateral darker markings on dirty gray background. Eye development as in male, eyes with rings of dark pigment. CL 1.3, CW 1.25, abdomen length 1.8, total body length 3.1. Leg I total length 10.75 (3, 0.75, 3.1, 2.7, 1.2), leg formula 1423, leg I / CW ratio 8.6. Epigynum, viewed laterally, with a prominent nose-like cream-colored median septum, like other members of the species group. Viewed ventrally, oval-shaped epigynal pockets lateral to median septum, angled outwards from top to bottom. Dorsal view showing spermathecae below epigynal pockets, angled upwards obliquely, approximately avocado-shaped. With sac-shaped structures anterior to epigynal pockets, hypothesized as vulval pockets (Vp). Epigynal plates meeting along midline, parallel from top to bottom.

♀ **Variation.** Variation exists in the shape of the lateral epigynal pockets (ventral view), but the overall vulval pocket morphology, spermathecal shape, and parallel epigynal plates is fairly conserved across populations (Fig. 36A–J).

Distribution and natural history. Populations have been collected from the larger Bald Mountains area along the North Carolina / Tennessee border, southwest of Erwin, Tennessee (Fig. 30). Most populations have been collected from boulderfield

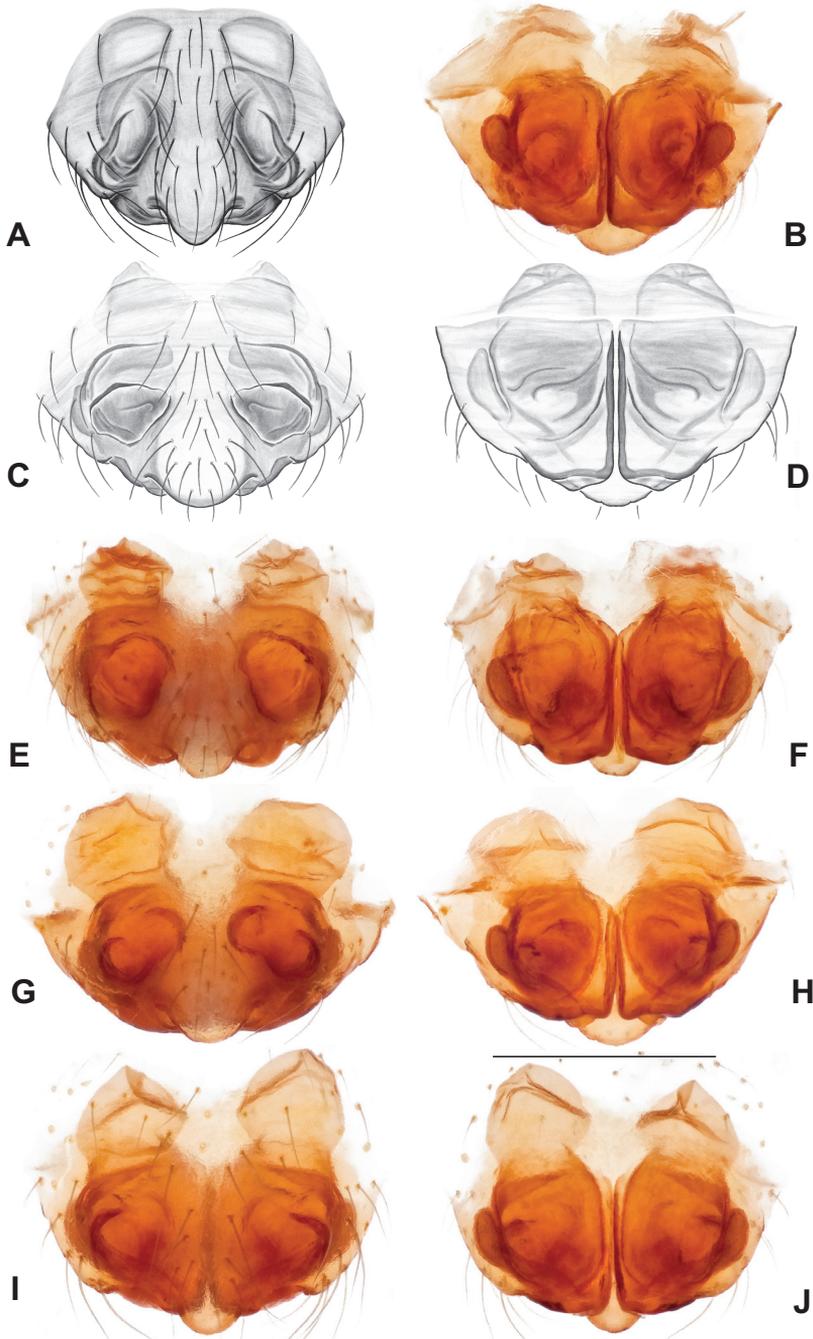


Figure 36. *Nesticus templetoni* sp. nov. epigynal variation. North Carolina, Unicoi Co., Rich Mountain, Clark Creek, MCH 04_036 (SDSU_TAC000670), ventral (**A**), dorsal (**B**). North Carolina, Madison Co., East Prong Hickory Fork Creek, MCH 01_144, ventral (**C**), dorsal (**D**). Tennessee, Greene Co., Bald Mountains, MCH 04_037, ventral (**E**), dorsal (**F**). Tennessee, Unicoi Co., along Mill Creek, MCH 07_147, ventral (**G**), dorsal (**H**). North Carolina, Yancey Co., E of Spivey Gap, MCH 01_146, ventral (**I**), dorsal (**J**). Scale bar: 0.5 mm.

void spaces. For example, at the type locality, spiders were found to be “common” in a “deep, moist boulderfield”. At Bald Mountain Road (MCH 04_038) spiders were found near and under rock accumulations adjacent to a small stream.

Etymology. This species is named to recognize and honor Dr. Alan Templeton, Charles Rebstock Professor Emeritus of Biology, Washington University. A brilliant evolutionary, speciation, and conservation biologist, with a deep love for all biodiversity. PhD dissertation advisor of MH, honored here for his inspiration and support during the first author’s formative years as an evolutionary biologist.

Remarks. Two strongly supported geographic subclades are recovered with mitochondrial data (Fig. 6), corresponding to eastern / southern (Clarks Creek, Mill Creek, Scronce Creek) versus western (Bald, Bald Mtn Road, Hickory Fork) sample locations (Fig. 30). Increased nuclear phylogenomic sampling might ultimately reveal these geographic populations as reciprocally monophyletic.

Nesticus crosbyi Gertsch, 1984

Figs 37A–D, 38A–H

Nesticus crosbyi Gertsch, 1984: 33, figs 173, 174.

Material examined. Type material: *Holotype*: USA – North Carolina, Yancey Co. • ♀ holotype; Commissary Ridge Trail, 100 yards west of main peak of Mt. Mitchell; 22 Aug. 1960; T.C. Barr leg.; AMNH; **New collections from near type locality: – Yancey Co.** • 2♂, 5♀; Mt. Mitchell SP, just NE summit parking lot; 35.7671°N, -82.2641°W; 15 Aug. 1992; M. Hedin leg.; • ♂, 2♀; Mt. Mitchell SP, just NE summit parking lot; 35.7671°N, -82.2641°W; 4 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_012; • 2♀; Mt. Mitchell SP, just NE summit parking lot; 35.7671°N, -82.2641°W; 25 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_083; **Non type material: – Buncombe Co.** • 2♀, 3 imm; Walker branch of Dillingham Creek, drainage N of Walker Falls branch, Little Andy Creek; 35.7677°N, -82.3594°W; 25 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_168; • ♂, 2♀; Walker branch of Dillingham Creek, drainage N of Walker Falls branch, Little Andy Creek; 35.7677°N, -82.3594°W; 5 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_196; • ♀, 1 imm; SW of Cane River Gap, Hwy 197, 5 mi ENE Barnardsville; 35.8036°N, -82.3536°W; 25 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_167; – **Yancey Co.** • ♂, ♀; Black Mountains, near Cattail Peak; 35.7977°N, -82.2564°W; 4 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_012a; • 1 imm (identification based on geography and mitochondrial evidence); Blue Ridge Parkway at Bald Knob Ridge Trail, near entrance to Mt. Mitchell SP; 35.715°N, -82.2736°W; 21 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_141; • 2♂, 2♀; FR 472 along South Toe River, below Chestnut knob; 35.7265°N, -82.2452°W; 20 Aug. 2001; M. Hedin, M. Lowder, R. McClanahan leg.; MCH 01_143; • 2♀; Mt. Mitchell SP, near Mt Craig; 35.7776°N, -82.2616°W; 4 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_012a; • ♀, 1 imm; Mt. Mitchell SP, off Hwy 128, between Mt Gibbes and

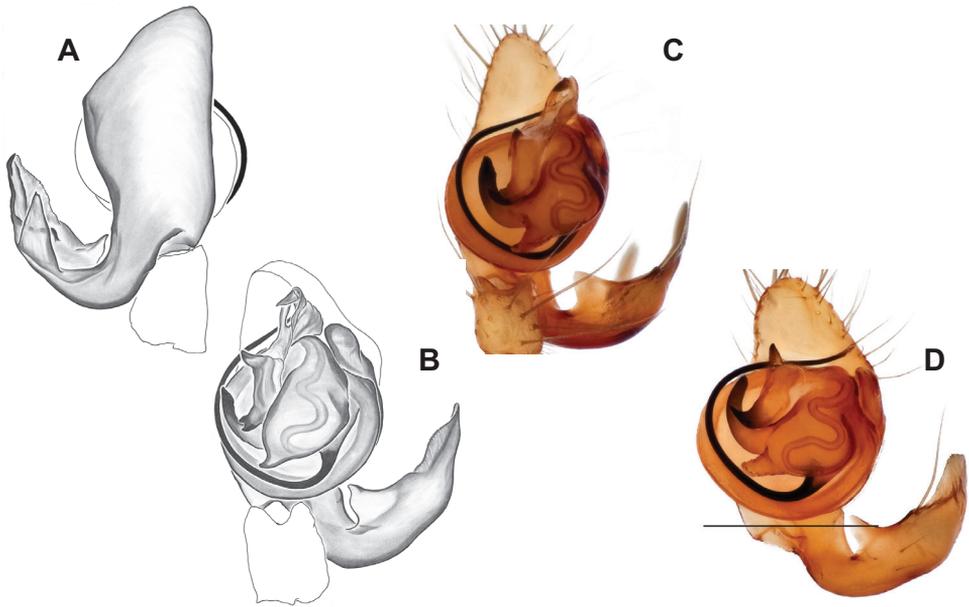


Figure 37. *Nesticus crosbyi* ♂ palps. North Carolina, Yancey Co., Mt. Mitchell SP, just NE of summit parking lot, MCH specimen #1201, dorsal (A), ventral (B) C North Carolina, Buncombe Co., Prices Creek Road at Price Creek, MCH 07_149, ventral D North Carolina, Buncombe Co., Walker branch of Dillingham Creek, MCH 02_196, ventral. Scale bar: 0.5 mm.

Steps Gap; 35.7432°N, -82.2788°W; 26 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_170; • 1 imm (identification based on UCE and mitochondrial evidence); Shuford Creek, off Whiteoak Rd., SW of Celso; 35.8382°N, -82.2193°W; 21 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_139; • 2♀; south of Big Laurel Mountain, N off Blue Ridge Parkway; 35.7401°N, -82.1991°W; 20 Aug. 2001; M. Hedin, M. Lowder, R. McClanahan leg.; MCH 01_142; • ♂, 2♀, 1 imm; Prices Creek Road at Price Creek; 35.8448°N, -82.3869°W; 22 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_149.

Diagnosis. Male palps differ in many ways from other members of the species group (including closest relatives), with a forked base of the tegulum, a narrow, curved regular apophysis, a beak-like basal process of the median apophysis, and a translucent dorsal paracymbial process with a relatively wide base (Fig. 37A–D). Females have genitalia similar to members of the close-knit morphological and phylogenetic subgroup, also including *Nesticus gertschi*, *N. secretus*, and *N. canei*, but can be diagnosed by epigynal internal anterior plates/lobes that differ in shape (Fig. 38A–H) and appear to lack the hypothesized vulval pockets (Vp) seen in other members of the species group (Fig. 29G–M).

Description of ♂ from near type locality (MCH specimen #1201). Carapace dusky cream to orange, conspicuous faint dark pigment behind ocular area and along carapace margins bleeding inwards. Legs pale yellow to cream. Abdomen mostly pale

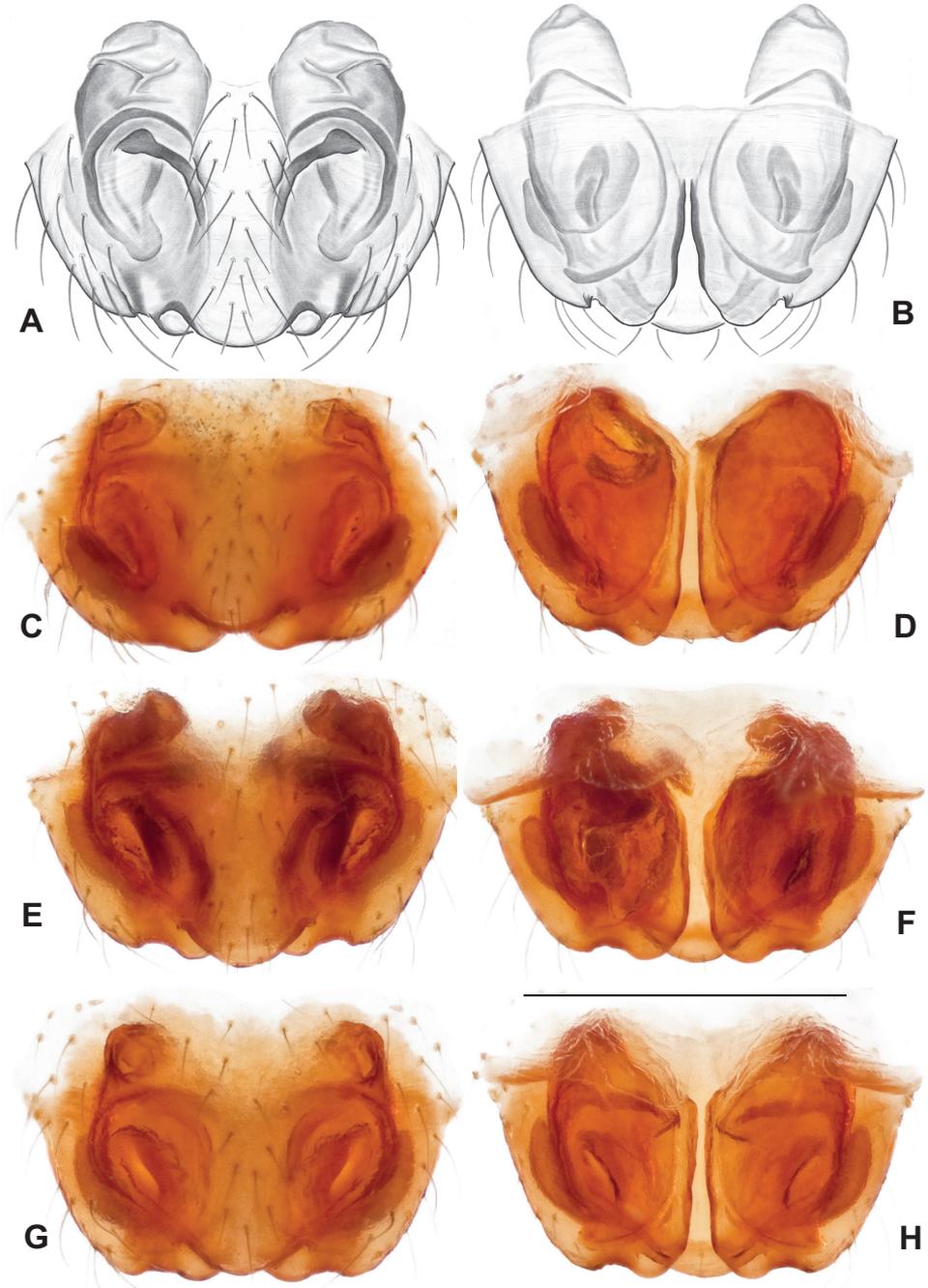


Figure 38. *Nesticus crosbyi* epigynal variation. North Carolina, Yancey Co., Mt. Mitchell SP, MCH specimen #1204, ventral (A), dorsal (B). North Carolina, Buncombe Co., SW of Cane River Gap, MCH 01_167, ventral (C), dorsal (D). North Carolina, Buncombe Co., Prices Creek Road at Price Creek, MCH 07_149, ventral (E), dorsal (F). North Carolina, Buncombe Co., Walker branch of Dillingham Creek, MCH 02_196, ventral (G), dorsal (H). Scale bar: 0.5 mm.

cream with crisp paired lateral pigmentation blotches. All eyes approximately equal in size, except for AMEs, $\sim 1/4$ width of ALEs. Eyes with rings of dark pigment. CL 1.6, CW 1.45, abdomen length 2.15, total body length 3.75. Leg I total length 9.85 (2.65, 0.65, 2.95, 2.5, 1.1), Leg formula 1423, leg I / CW ratio 6.8. Palp with forked base of the tegulum, including a short basal branch and a narrow, curved, dark, thin regular apophysis. Median apophysis with lateral process well-sclerotized and beak-like, thin apical process. Paracymbium with well-developed triangular translucent ventral process, distal process typical for the species group, paradistal process reduced to a sclerotized low ridge, and a translucent dorsal process with a relatively wide base, mostly lacking distal serrations.

Variation. Male variation was observed in the shape of the median apophysis lateral process, the paracymbial ventral process, and the proximal fork of the tegulum (Fig. 37A–D). Female variation was observed in the shape of the epigynal internal anterior lobes (Fig. 38A–H).

Distribution and natural history. Previously known only from the type location (Mt. Mitchell), corresponding to the highest uplands east of the Mississippi River in North America, above 2000 meters in elevation. Our new records indicate that this species is more widespread in the Black Mountains (both to the north and southeast), and we include here new records from west of the Blacks, in the Great Craggy Mountains (Fig. 30). This demonstration of an overall larger geographic distribution, with populations also at lower elevations (e.g., Prices Creek at 930 m), has important conservation implications for this species.

Most collections have resulted in a relatively modest number of specimens taken. For example, at an apparently pristine boulderfield along the South Toe River (MCH 01_143), three persons each searching for 30 minutes collected four total adult specimens.

Remarks. Strongly supported as a clade by UCE data (Figs 3, 4), but not recovered as monophyletic on the mitochondrial gene tree (Fig. 6), where sequences are intermixed with sequences from close relatives *Nesticus gertschi* and *N. canei*. Mitochondrial introgression and/or incomplete lineage sorting could explain this result, as all three species occur in the same geographic region (Fig. 30), making lineage contact and introgression possible.

Nesticus gertschi Coyle & McGarity, 1992

Fig. 39A–F

Nesticus gertschi Coyle & McGarity, 1992: figs 15–20.

Material examined. Type material: Holotype: USA – Tennessee, Greene Co. • ♂ holotype; Cedar Creek Cave, 100 m into cave; 16 Mar. 1991; T. McGarity leg; AMNH; **New collections from type locality:** – Tennessee, Greene Co. • 2♂, 9♀; Cedar Creek Cave, 1 mi. S Cedar Creek; 21 Sep. 1992; M. Hedin, S. O’Kane leg. **Non type material:** – North Carolina, Buncombe Co. • ♂; 0.1 mi. NW Hickory Nut Gap Hwy 74, NW of Gerton; 35.4898°N, -82.3627°W; 5 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_013; • ♂, ♀, 1 imm; 0.1 mi. NW Hickory Nut Gap Hwy

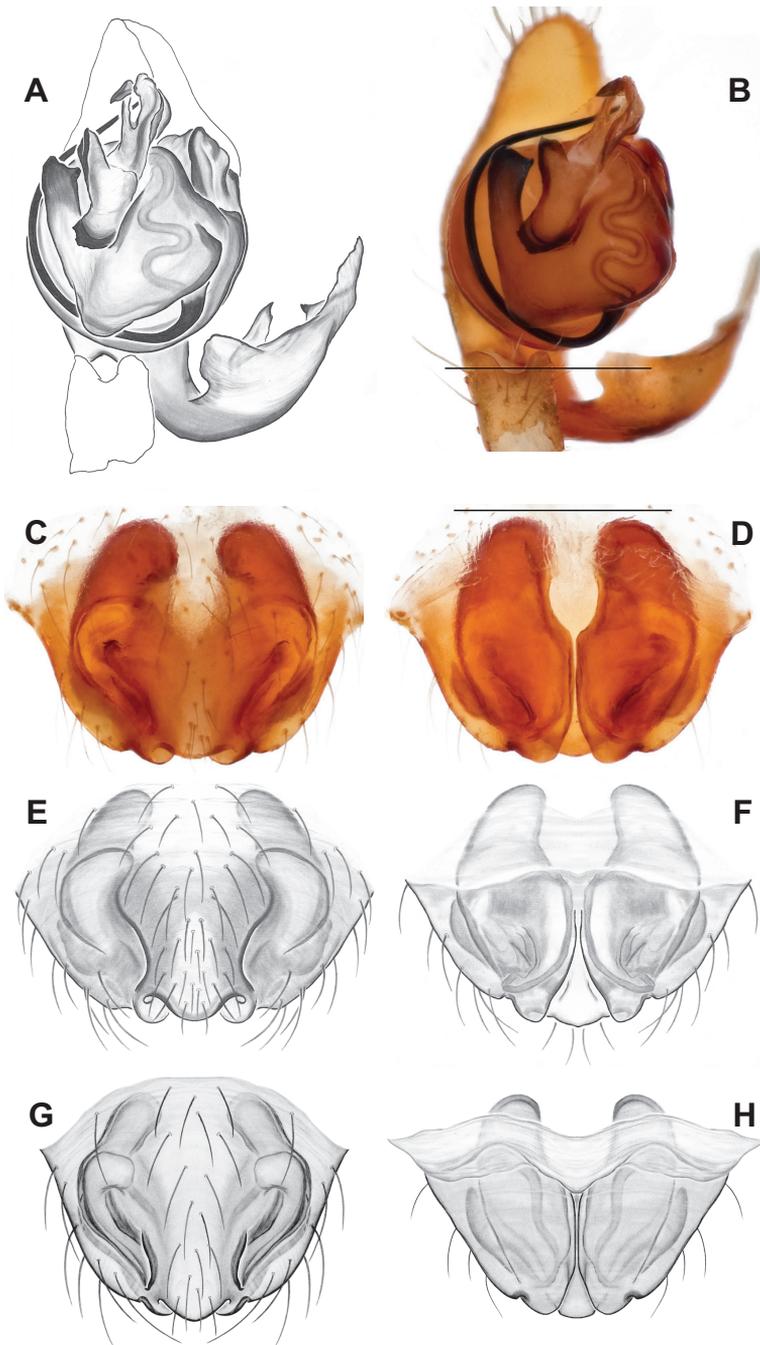


Figure 39. *Nesticus gertschi* and *N. secretus* genitalia. *N. gertschi* **A** North Carolina, Yancey Co., Blue Ridge Parkway at Balsam Gap, MCH 02_195, ♂ palp, ventral **B** North Carolina, Buncombe Co., NW of Hickory Nut Gap, Hwy 74, MCH 01_173, ♂ palp, ventral. North Carolina, Buncombe Co., NW of Hickory Nut Gap, Hwy 74, MCH 01_173, epigynum, ventral (**C**), dorsal (**D**). North Carolina, Yancey Co., Blue Ridge Parkway at Balsam Gap, MCH 02_195, epigynum, ventral (**E**), dorsal (**F**). Scale bar: 0.5 mm. *N. secretus* Gertsch 1984 epigynum. Tennessee, Great Smoky Mountains National Park, ♀ holotype, ventral (**G**), dorsal (**H**).

74, NW of Gerton; 35.4898°N, -82.3627°W; 27 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_173; • 4♂, 7♀; Blue Ridge Parkway, Mile 370, 3 mi. SW Craggy Gardens turnoff; 35.6768°N, -82.4322°W; 15 Aug. 1992; M. Hedin leg.; • 2♂, 8♀; Flat Creek at NE edge of Montreat; 35.6528°N, -82.2972°W; 12 Aug. 1992; M. Hedin leg.; • 6♀; FR 63 along Mineral Creek, S of Dillingham; 35.7093°N, -82.3939°W; 21 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_143; – **North Carolina, Madison Co.** • ♂, ♀; Anthodite Cave; 5 Jan. 2002; J.D. Mayes leg.; • ♂, 4♀; FR 467 to Rich Mountain, 0.5 mi. to jct w/ Hwy 25/70, W of Hurricane; 35.9274°N, -82.7792°W; 22 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_144; • 4♀, 1 imm; Rich Mountain, 0.5 mi. N Rich Mountain lookout; 35.9313°N, -82.806°W; 19 Aug. 2001; M. Hedin, M. Lowder leg.; MCH 01_140; – **North Carolina, Yancey Co.** • 3♀, 2 imm; Blue Ridge Parkway at Balsam Gap, just down Big Butt trail; 35.7495°N, -82.3343°W; 26 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_169; • 2♂, 3♀; Blue Ridge Parkway at Balsam Gap, just down Big Butt trail; 35.7495°N, -82.3343°W; 5 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_195; – **Tennessee, Cocke Co.** • 5♀; along French Broad River, north of Wolf Creek Bridge, FR 209; 35.9228°N, -82.9585°W; 12 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_044.

Diagnosis. See Coyle and McGarity (1992) for diagnosis comparing *Nesticus gertschi* to other members of the species group; here revised to recognize the close relationship to *N. canei* sp. nov. Males can be distinguished from the latter by the tegular apophysis tip (beyond bend) broad and truncate (Fig. 39A, B), and paracymbial distal process with subdistal processes. Females are very similar to *N. secretus* and *N. canei* sp. nov., with internal anterior plates of epigyna not projecting inwards and ventrally as strongly as in the latter species (Fig. 39C–F).

Variation. This species shows surprisingly little genitalic variation despite a relatively large geographic distribution (e.g., compare ♂ Fig. 39A, B. to Coyle and McGarity (1992) figs 15–17), and obvious lowland geographic barriers (Fig. 30). Specimens from surface-dwelling populations are generally smaller in body size than cave-dwelling specimens from the type locality. Two female specimens (of five total) from surface collections along the French Broad River (MCH 04_044) lack eye pigmentation, while cave-dwelling specimens from both Cedar Creek Cave and Anthodite Cave possess eye pigmentation.

Distribution and natural history. Previously known only from the type locality (Cedar Creek Cave), this species is a fairly widespread surface-dwelling species (Fig. 30). The geographic distribution is apparently fragmented with northern, central, and southern populations, with all but one known population from east of the French Broad River (Anthodite Cave being the exception).

Surface collections are mostly from shaded boulderfields, with field notes suggesting spiders to be “fairly common” under rocks in void spaces. Montreat specimens were found in dark cracks and crevices of a man-made rock wall within 3 meters of a stream.

Remarks. As discussed below, possibly synonymous with *Nesticus secretus*.

Strongly supported as a clade by UCE data, with nuclear subclades corresponding to northern vs. central + southern collection locations (Figs 3, 4), separated by the

Mars Hill lowland gap. Not recovered as monophyletic on the mitochondrial gene tree (Fig. 6), where sequences are intermixed with those of closely related *Nesticus crosbyi* and *N. canei* sp. nov.

Gertsch (1984, p. 30) cites a record for *Nesticus reclusus* as (“*McDowell County, Montreat, 16 October, 1923, female*”). However, our 1992 collections from Montreat (now in Buncombe County) only include *N. gertschi*, which is the locally prevalent species (Fig. 30). Also, members of the *reclusus* group are not known from east of the Asheville Basin (Fig. 53). We have not seen the 1923 specimen but suspect either mislabeling or misidentification.

***Nesticus secretus* Gertsch, 1984**

Fig. 39G, H

Nesticus secretus Gertsch, 1984: 33, figs 173, 174.

Material examined. Type material: *Holotype*: USA – Tennessee • ♀ holotype; Great Smoky Mountains National Park; 8 Jul. 1933; W.J. Gertsch leg.; AMNH.

Remarks. Gertsch cites the type data for this “*small, dusky epigean species with short legs*” as “female holotype from Little Pigeon River, Great Smoky Mountains National Park, Sevier County, Tennessee, 8 July 1933 (W.J. Gertsch)”. However, the label associated with the holotype female (see above) includes neither specific locality nor county information.

The type female is clearly a representative of the *nasicus* group, and is potentially synonymous with *Nesticus gertschi* (see Fig. 39C, D). However, essentially all eastern *nasicus* group populations are known from east of the French Broad River, while the Great Smoky Mountains National Park lies west of this (Fig. 30). Also, extensive collections from the eastern portion of the Great Smoky Mountains National Park have only ever resulted in the collection of members of the *tennesseensis* group (*N. cherokeensis* and *N. silvanus*, Fig. 13), or members of the *reclusus* group (*N. binfordae* and *N. reclusus*, Fig. 53). Both *N. binfordae* and *N. reclusus* have been collected from along the Little Pigeon River.

A possible region to search for *Nesticus secretus* would be the English or Green Mountains, west of the French Broad River, but not too distant from records for *N. gertschi* (Fig. 30). Because of this possibility we retain *N. secretus* as a valid taxon, pending further focused collection efforts.

***Nesticus canei* sp. nov.**

<https://zoobank.org/F17F97DD-6F1D-4A48-B9B7-122075D9EAE6>

Fig. 40A–D

Material examined. Type material: *Holotype*: USA – North Carolina, Yancey Co.

• holotype ♂; Hwy 19W along Cane River, near Egypt-Ramseytown Fire Station,

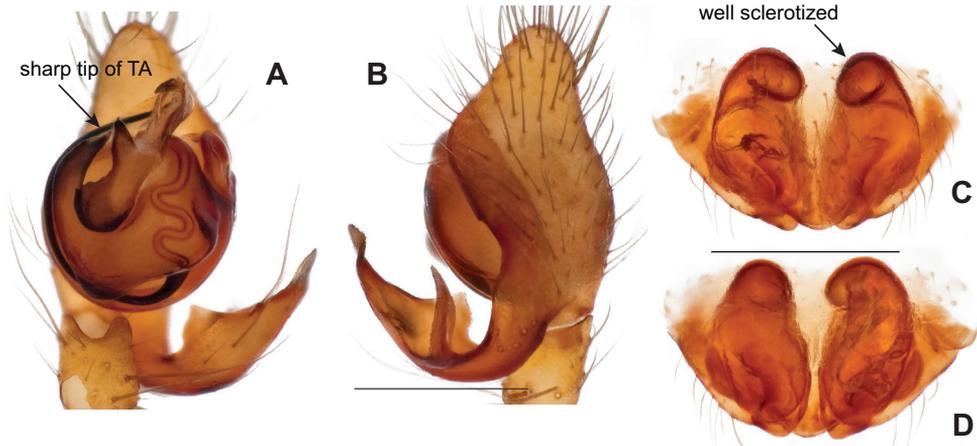


Figure 40. *Nesticus canei* sp. nov. genitalia. North Carolina, Yancey Co., Hwy 19W, along Cane River, near Egypt–Ramseytown Fire Station, MCH 04_043 (SDSU_TAC000671), ♂ palp, ventral (A), dorsal (B). North Carolina, Yancey Co., Hwy 19W, along Cane River, near Egypt–Ramseytown Fire Station, MCH 04_043 (SDSU_TAC000672), epigynum, ventral (C), dorsal (D). Scale bar: 0.5 mm.

near Lewisburg; 35.9921°N, -82.3927°W; 11 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_043 (SDSU_TAC000671); **Paratypes:** – **Yancey Co.** • ♂, ♀; Hwy 19W along Cane River, near Egypt–Ramseytown Fire Station, near Lewisburg; 35.9921°N, -82.3927°W; 11 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_043; **Non type material:** – **Yancey Co.** • 9♀, 17 imm; Hwy 19W along Cane River, near Egypt–Ramseytown Fire Station, near Lewisburg; 35.9921°N, -82.3927°W; 11 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_043.

Diagnosis. The male palp is like that of *Nesticus gertschi* (Fig. 39A, B) but the distal end of the regular apophysis is acute rather than blunt, and the basal portion of median apophysis is relatively more expanded (Fig. 40A, B). The distal paracymbial process lacks the subdistal processes as found in *N. gertschi*. Female with dorsal portion of internal anterior lobes/plates well sclerotized, rounded anteriorly and curving ventrally (Fig. 40C, D).

Description of ♂ holotype (SDSU_TAC000671). Carapace dusky cream to orange, conspicuous faint dark pigment behind ocular area, along carapace margin bleeding inwards. Legs pale yellow to cream. Abdomen mostly pale cream, with crisp paired lateral pigmentation blotches. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.6, CW 1.3, abdomen length 2, total body length 3.6. Leg I total length 10.45 (3, 0.65, 3.1, 2.55, 1.15), leg formula 1423, leg I / CW ratio 8.0. Palp regular apophysis with a 90-degree bend, distal end acute and blade-like. Lateral process of median apophysis concave, broadening and well-sclerotized along edge, distal process drawn into thin tip that closely parallels regular apophysis tip. Paracymbium with strong ventral process,

distal process consistent with species group (spatulate) and without other processes, dorsal process translucent blade of medium width, reaching above ventral process, weakly serrated at tip.

♂ **Variation.** The palp of the paratype male is similar to the holotype.

Description of ♀ paratype (SDSU_TAC000672). Carapace dusky cream, very faint dark pigment behind ocular area, along carapace margin bleeding inwards. Legs pale yellow to cream. Abdomen with paired, lateral darker marking on a dirty gray background. Eyes approximately equal in size, except for AMEs, $\sim 1/4$ width of ALEs. Eyes with rings of dark pigment. CL = 1.2, CW 1.05, abdomen length 1.55, total body length 2.75. Leg I total length 8.1 (2.3, 0.55, 2.3, 2, 0.95), leg formula 1423, leg I / CW ratio 7.7. Epigynum, viewed laterally, with a prominent nose-shaped, cream-colored median septum, like other members of the species group. Viewed dorsally, dorsal-projecting portion of internal anterior lobes well sclerotized, rounded anteriorly and curving ventrally. Sclerotization making these appear as dark circles sitting above epigynum when viewed ventrally. Viewed dorsally, spermathecae below epigynal pocket, angled obliquely upwards, approximately banana-shaped.

♀ **Variation.** Adult females from the type locality vary in body size and in carapace and abdomen color (dark vs. light) but share a similar epigynum.

Distribution and natural history. Known only from the type locality from along the Cane River, a tributary of the Nolichucky River. Adjacent collections have thus far only resulted in the collection of non-sister *Nesticus templetoni* (Fig. 30) and *N. paynei* further east (Fig. 13). We hypothesize that *N. canei* has a very small geographic distribution. More collecting effort in the immediate vicinity of the type locality is needed to understand the geographic extent of this apparently microendemic species.

Specimens from the type collection were found to be relatively common in void spaces beneath rocks in a small shaded boulderfield in roadside forest, at approximately 700 meters in elevation.

Etymology. Named after the Cane River, a small north-flowing river found only in Yancey County, North Carolina.

Remarks. Morphologically very similar to *Nesticus gertschi* and sister to this taxon on UCE trees (Figs 3, 4). This species is of conservation importance because of an apparently naturally small geographic distribution.

***barrowsi* group, including:**

Nesticus bondi sp. nov.

Nesticus barrowsi Gertsch, 1984

Nesticus lowderi sp. nov.

This small species group is strongly supported as monophyletic on both concatenated and coalescent phylogenomic trees (Figs 3, 4). Furthermore, the three species within this group are each strongly supported by nuclear phylogenomic data, with

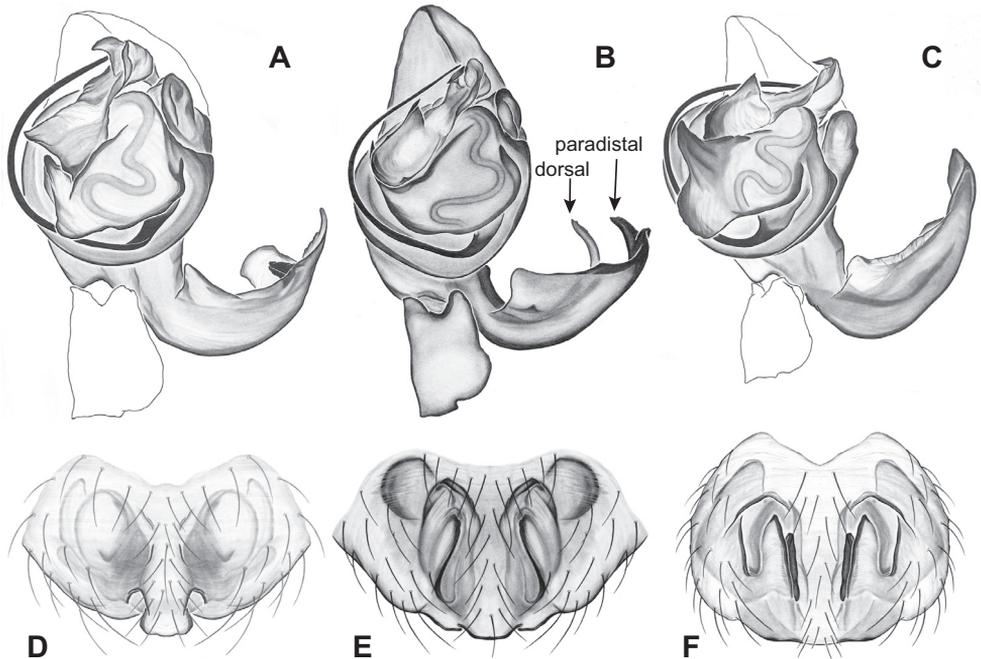


Figure 41. Comparative ♂♀ genitalia of *barrowsi* group; ♂ **A** *Nesticus bondi* **B** *N. barrowsi* **C** *N. lowderi*; ♀ **D** *N. bondi* **E** *N. barrowsi* **F** *N. lowderi*. All views ventral. See subsequent figures for specimen locations and voucher details.

high gene and site CF values suggesting minimal gene tree variance within the group (Fig. 5). Each species is recovered with high support on the mitochondrial tree (Fig. 6), but the species group itself is polyphyletic, fragmented into three distantly related mitochondrial clades (Fig. 6).

Consistent with phylogenomic data, each species in this group is morphologically distinctive, easily separated by diagnostic features of both male and female genital morphology (Fig. 41). Overall, species delimitation within this group is straightforward, likely reflecting a relatively more ancient history (and extinction of intervening lineages) within the group. This situation parallels the *archeri* group, but is unique for a montane lineage of Appalachian *Nesticus*.

We do not identify diagnostic morphological features for the entire species group, as many aspects of both male and female morphology occur elsewhere in the combined lineages sister to the *barrowsi* group (i.e., larger clade including *barri*, *carteri*, and *reclusus* groups; Figs 3, 4).

Each of the species in this species group occupies a relatively small geographic distribution in three disjunct pockets of the far western Blue Ridge (Fig. 42). These disjunct pockets are separated by montane habitats occupied by members of the *reclusus* group (Fig. 53). As discussed below, all species in the *barrowsi* group appear to be naturally rare, which again parallels the *archeri* group.

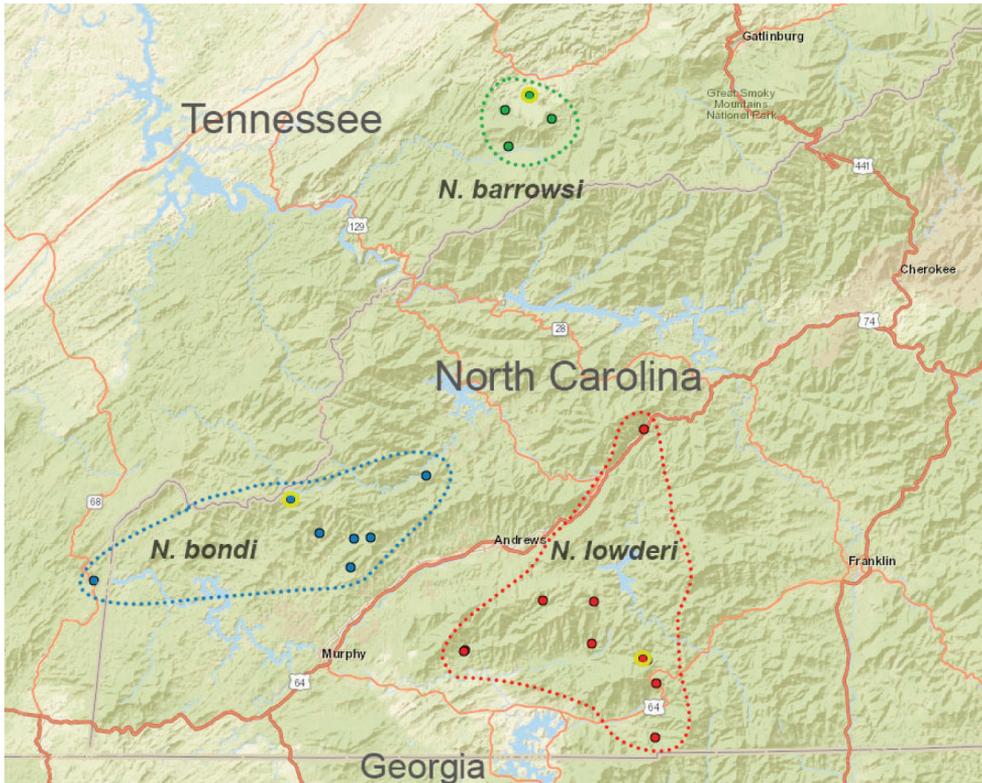


Figure 42. Distribution of the *barrowsi* group. Type localities designated with yellow circles. State boundaries and major cities shown for geographic context. Dashed lines circumscribe known species distributions.

***Nesticus bondi* sp. nov.**

<https://zoobank.org/4E1DA202-2E5A-41DC-9936-323CFA01BE53>

Fig. 43A–J

Material examined. Type material: *Holotype*: USA – **North Carolina, Cherokee Co.** • holotype ♂; along Tipton Creek, 1.2 mi. S NC/TN state line; 35.2503°N, -84.0724°W; 26 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_158; SDSU_TAC000665; ***Paratypes*.** – **North Carolina, Cherokee Co.** • 6♀; along Tipton Creek, 1.2 mi. S NC/TN state line; 35.2503°N, -84.0724°W; 26 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_158; **Non type material:** – **North Carolina, Cherokee Co.** • 5 imm; along Tipton Creek, 1.2 mi. S NC/TN state line; 35.2503°N, -84.0724°W; 26 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_158; • ♂, 5♀, 2 imm; Davis Creek Road, along Davis Creek, Snowbird Mountains, N of Grandview; 35.2151°N, -84.0368°W; 16 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_110; • ♂, 2♀, 1 imm; Dinkin Cove Road, N of Hanging Dog Mountain; 35.1809°N, -83.9988°W; 16 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_109; • ♂, 5♀; Hanging Dog Creek,

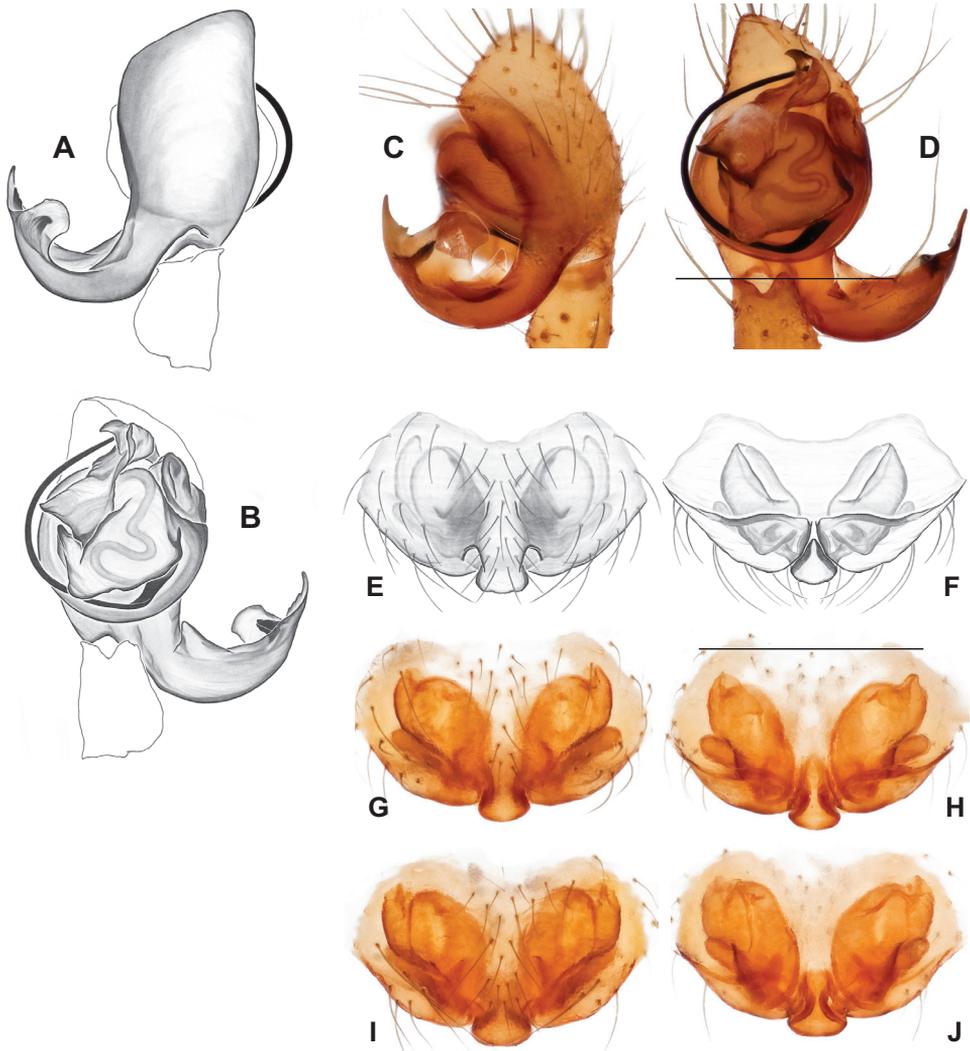


Figure 43. *Nesticus bondi* sp. nov. ♂ palp and ♀ epigynal variation. North Carolina, Cherokee Co., along Tipton Creek, MCH 02_158 (SDSU_TAC000665), ♂ palp, dorsal (A), ventral (B). North Carolina, Graham Co., along Snowbird Creek, MCH 02_161, ♂ palp, dorsal (C), ventral (D). Scale bar: 0.5 mm. North Carolina, Cherokee Co., along Tipton Creek, MCH 02_158 (SDSU_TAC000666), epigynum, ventral (E), dorsal (F). North Carolina, Cherokee Co., Dinkin Cove Road, MCH 07_109, epigynum, ventral (G), dorsal (H). North Carolina, Cherokee Co., Davis Creek Road, MCH 07_110, epigynum, ventral (I), dorsal (J). Scale bar: 0.5 mm.

below Hanging Gap; 35.2112°N, -83.9739°W; 17 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_055; • ♀; Hanging Dog Creek, E Boiling Springs; 35.2094°N, -83.9945°W; 17 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_056; USA – **North Carolina, Graham Co.** • ♂, 2 imm; along

Snowbird Creek, near Wilson Cabin; 35.2733°N, -83.9051°W; 27 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_161; – **Tennessee, Polk Co.** • 1 imm (identification based on UCE and mitochondrial evidence); Hwy 68, vic Apalachia, just S Hiwassee River; 35.1676°N, -84.3159°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_111.

Diagnosis. Males are easily distinguished from other members of the species group by the unique shape of the median apophysis, the shape of the tegular apophysis and tegular keel, the shape of the dorsal paracymbial process, and possession of a thorn-shaped distomedial paracymbial process (Fig. 43A–D). Epigynal morphology, particularly the shape of the posterior extension of the median septum, is distinctive for the entire Appalachian clade (Fig. 43E–J).

Description of ♂ holotype (SDSU_TAC000665). Carapace dusky cream to orange, faint gray pigmentation behind ocular area leading to midline and around edges. Legs pale yellow / cream. Abdomen with paired dark gray blotches on a light gray background. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.3, CW 1.17, abdomen length 1.35, total body length 2.65. Leg I total length 9.8 (2.72, 0.53, 2.87, 2.58, 1.1), leg formula 1423, leg I / CW ratio 8.4. Paracymbium possesses a well-sclerotized thorn-shaped distomedial process. Paracymbial dorsal process a large transparent lobe that lacks a basal process, approximately contiguous with the distal paracymbial process, itself conspicuously weakly sclerotized, narrow, pointed, and weakly serrate along dorsal edge. Ventral paracymbial process triangular. Median apophysis somewhat triangular with a sclerotized point directed prolaterally. Tegulum with posterior keel; tegular process short, beak-like, narrows distally, and directed anteriorly. Distal tip of conductor bent and directed prolaterally.

♂ **Variation.** Males from different geographic locations show very minor variation in the width (at base) of the dorsal paracymbial process and depth of indentation between dorsal and distal processes (Fig. 43A–D).

Description of ♀ paratype (SDSU_TAC000666). Carapace dusky cream to orange, gray pigmentation behind ocular area leading to midline and around edges. Legs pale yellow / cream. Abdomen with paired dark gray / black blotches on a light gray background. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.4, CW 1.24, abdomen length 1.75, total body length 3.15. Leg I total length 8.32 (2.38, 0.54, 2.39, 2.01, 1), leg formula 1423, leg I / CW ratio 6.7. Epigynum with well-defined orifices lateral to a posterior extension of the median septum, itself widening posteriorly with a flattened posterior edge. Spermathecae elongated and directed anterolaterally. Posterolateral edges of epigynum folded over dorsally to form dorsal posterior flaps. Viewed dorsally, large, internal lobes extend anterolaterally with sclerotized rims.

♀ **Variation.** Females from different geographic locations show very minor variation in the shape of the anterior internal sclerotized epigynal lobes (Fig. 43E–J).

Distribution and natural history. Most populations are from the southwestern flanks of the Snowbird Mountains of western North Carolina (Fig. 42). A single

immature specimen is known from further west at Apalachia (placement based on UCE and mitochondrial evidence), suggesting that additional populations likely reside in the intervening montane habitats (Fig. 42).

At the type locality of Tipton Creek, *Nesticus bondi* (♂, 6♀) was found in syntopy with *N. sheari* (4♀); field notes read “*Nesticus* in boulderfield above road, north-facing, concentrated in small drainage”. Because we did not identify specimens directly in the field, it remains unclear if these different species were found side-by-side or were perhaps somehow segregated by microhabitat at this location. At Davis Creek (MCH 07_110), *Nesticus* were found “under rocks at streamside – many from webs under a large rock shelter cave”.

Etymology. Named after Dr. Jason Bond, Professor and Schlinger Chair of Insect Systematics at the University of California Davis. Jason was born in the southern Appalachians, schooled in the mountains of western North Carolina, and perhaps sometimes paddled in the Snowbird Mountains. Jason has been a longtime close friend and arachnological colleague of MH and is for him forever a source of scientific (and life) inspiration.

Remarks. The immature specimens from Tipton Creek are here attributed to *Nesticus bondi*, but some (or all) could be *N. sheari*.

Nesticus barrowsi Gertsch, 1984

Figs 44A, B, 45A–H

Nesticus barrowsi Gertsch, 1984: 35, figs 103–105, 118–120; Reeves 2000: 338.

Material examined. Type material: Holotype: USA – Tennessee, Blount Co. • ♂ holotype; Tuckaleechee Caverns, Tuckaleechee Cove; 1 Nov. 1938; W.B. Jones leg.; AMNH. **New collections from type locality:** – Blount Co. • ♀; Tuckaleechee Caverns, Tuckaleechee Cove; 22 Sep. 1992; M. Hedin, S. O’Kane leg. **Non type material:** – Blount Co. • ♂, 7♀; Great Smoky Mountains NP, Gregory Cave, Cades Cove; 21 Aug. 1992; M. Hedin leg.; • 2♀; Great Smoky Mountains NP, Rich Mountain Blowhole, Calf Cave; 2 Aug. 2000; M. Hedin, J. Cokendolpher, W. Reeves leg.; MCH 00_147; • ♀; Great Smoky Mountains NP, White Oak Sinks, Rainbow Cave; 21 Aug. 1992; M. Hedin leg.

Diagnosis. The diagnosis of Gertsch (1984) is revised here to recognize the phylogenetic affinities within the *barrowsi* group. *Nesticus barrowsi* is troglomorphic (long-legged, pale, approximately eyeless, relatively large-bodied), unlike other species in the species group. The male tegular apophysis curves to lie behind a quadrate median apophysis, is sharply tipped, without a basal keel. The paracymbial dorsal process is translucent, skinny and finger-like, while the distal process includes a well-sclerotized pointed tip and a small ventral keel (Fig. 44A, B). Female *N. barrowsi* differ from other members of the species group in overall morphology of the epigynum, including the pear-shaped spermathecae (Fig. 45A–H).

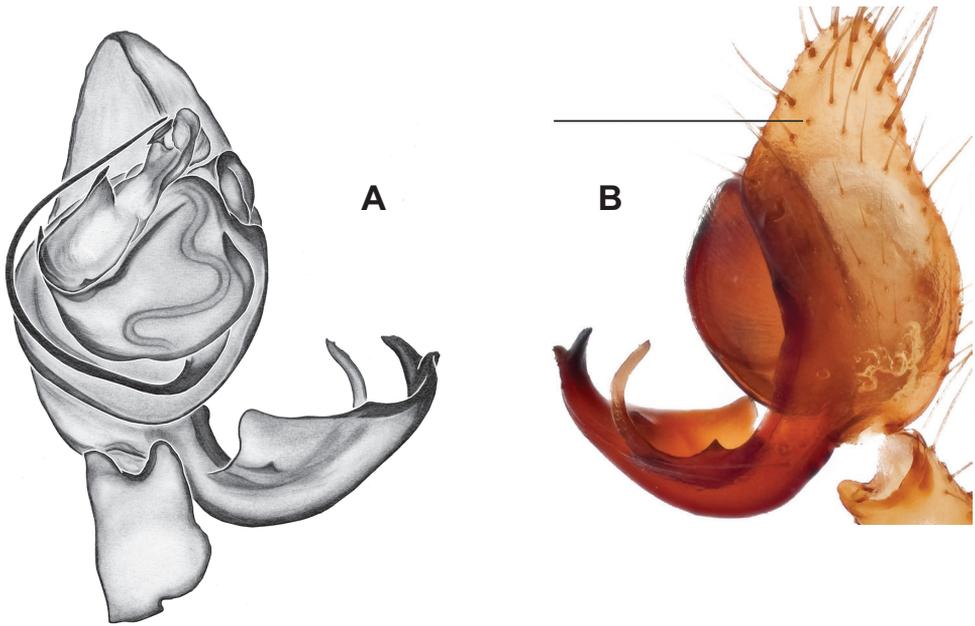


Figure 44. *Nesticus barrowsi* ♂ palps. Tennessee, Blount Co., Great Smoky Mountains NP, Gregory Cave, MCH specimen #1295, ventral (A), dorsal (B). Scale bar: 0.5 mm.

Variation. Minor variation is observed in the height and width of the paired epigynal plates across geographic locations (Fig. 45A–H).

Distribution and natural history. This troglomorphic species is only known from caves in karst windows along the northwestern edge of Great Smoky Mountains National Park (Cades Cove, Tuckaleechee Cove; Fig. 42). Reeves (2000) reported *Nesticus barrowsi* in sympatry with *N. stupkai* at two cave locations in Great Smoky Mountains National Park. We also collected these species in near syntopy at White Oak Sinks, with *N. barrowsi* found in the dark zone of caves and *N. stupkai* found closer to cave entrances (twilight zone).

***Nesticus lowderi* sp. nov.**

<https://zoobank.org/1F0FB7CB-C0AD-4284-B1CA-F44AC6C9E160>

Figs 46A–D, 47A–H

Material examined. Type material: *Holotype*: USA – North Carolina, Clay Co. • ♂ holotype; Chunky Gal Mountain, Chestnut Branch of Barnard’s Creek; 35.0857°N, -83.6327°W; 6 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_016 (SDSU_TAC000667); ***Paratypes*:** – Clay Co. • 3♀; data as for holotype; **Non type material:** – Clay Co. • ♂; along Barnard’s Creek, N side of Chunky Gal Mountain; 35.0868°N, -83.6372°W; 24 Apr. 1992; B. Dellinger leg.; • ♀; Eagle Fork Creek (Dave Barrett) SE

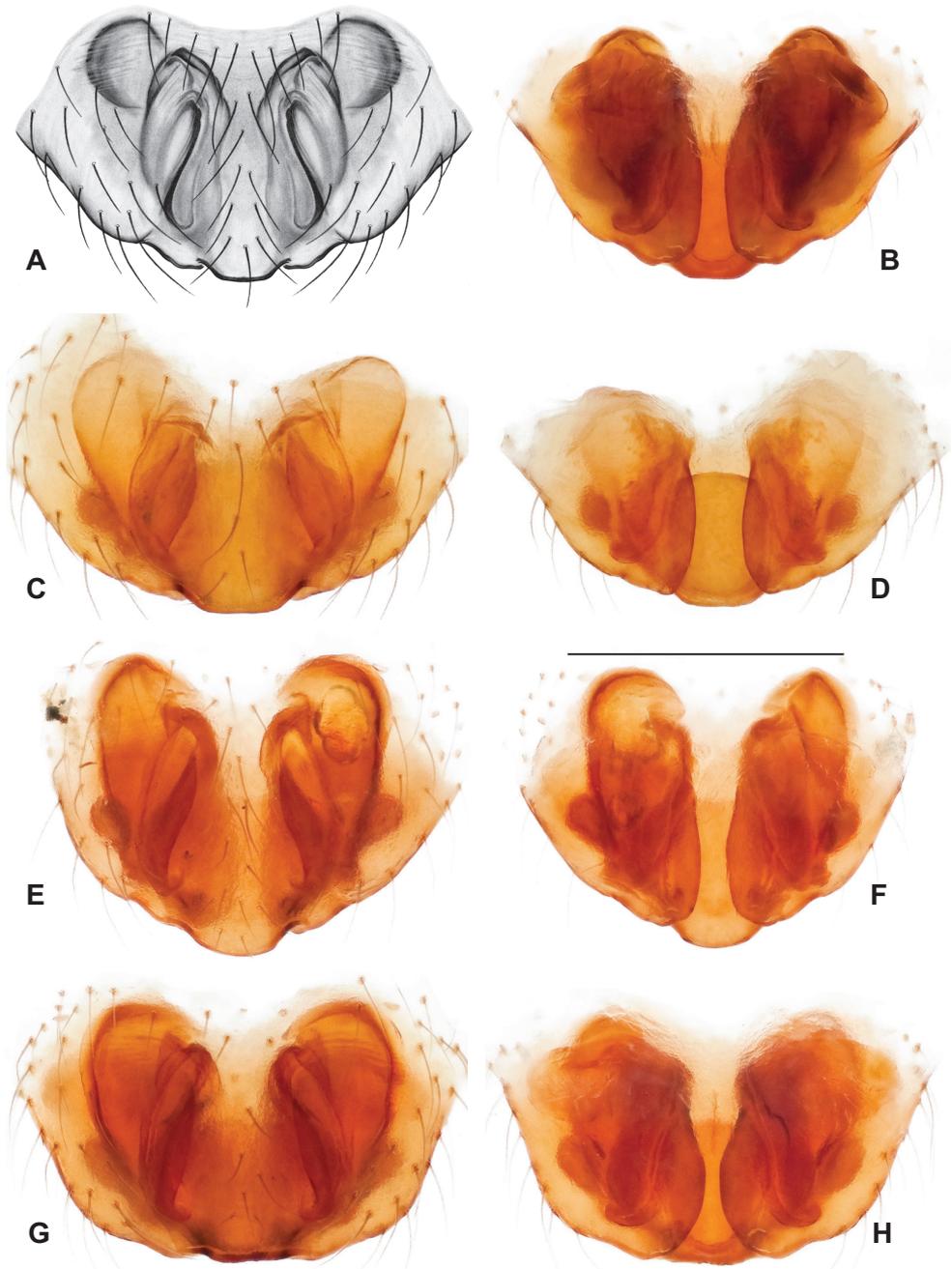


Figure 45. *Nesticus barrowsi* epigynal variation. Tennessee, Blount Co., Great Smoky Mountains NP, Calf Cave, MCH 00_147, ventral (A), dorsal (B). Tennessee, Blount Co., Great Smoky Mountains NP, White Oak Sinks, Rainbow Cave, MCH specimen #1303, ventral (C), dorsal (D). Tennessee, Blount Co., Tuckaleechee Caverns, MCH specimen #1568, ventral (E), dorsal (F). Tennessee, Blount Co., Great Smoky Mountains NP, Gregory Cave, MCH specimen #1297, ventral (G), dorsal (H). Scale bar: 0.5 mm.

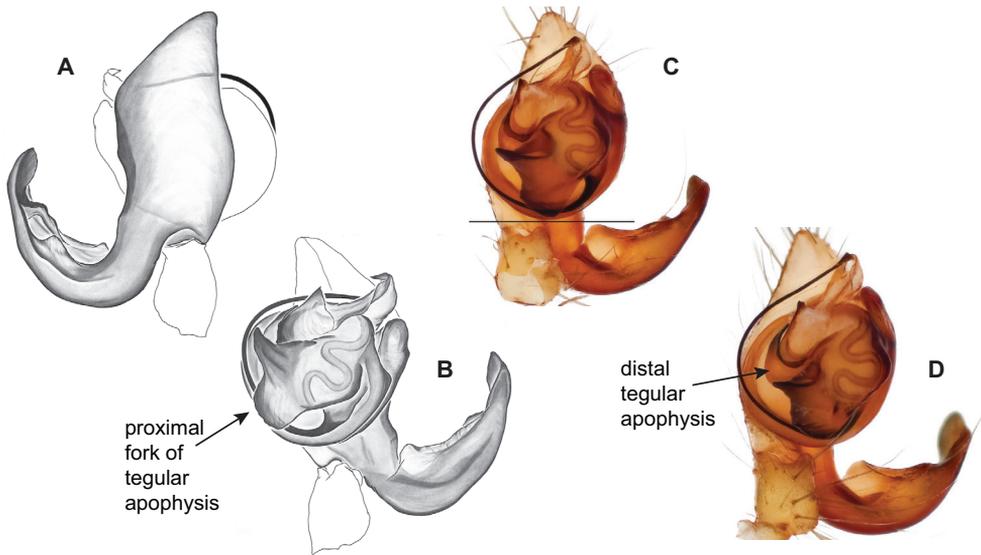


Figure 46. *Nesticus lowderi* sp. nov. ♂ palps. North Carolina, Clay Co., Chunky Gal Mountain, Chestnut Branch of Barnard's Creek, MCH 99_016 (SDSU_TAC000667), dorsal (A), ventral (B) C North Carolina, Clay Co., Big Tuni Creek, MCH 02_171, ventral D North Carolina, Clay Co., Fires Creek Road, Picnic Area along Fires Creek, MCH 07_108, ventral. Scale bar: 0.5 mm.

of Shooting Creek N of Hightower Bald; 35.0075°N, -83.6225°W; 20 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_143; • ♂, 6♀; Fires Creek Road, Picnic Area along Fires Creek; 35.0955°N, -83.8586°W; 16 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_108; • 3♂, 5♀; Fires Creek, Long Branch, just up from Short Branch; 35.1467°N, -83.7618°W; 21 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_144; • 2♀, 1 imm; Fires Creek, near Leatherwood Falls, just NE Fires Creek Picnic Area; 35.0961°N, -83.8566°W; 18 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_060; • 2♀; FR 440, along Big Tuni Creek, 2 mi. N Woods Road; 35.1025°N, -83.7007°W; 16 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_107; • ♂, 7♀, 7 imm; FR 440, Big Tuni Creek, E Tusquitee Bald near Bob Allison Picnic Area; 35.1463°N, -83.6974°W; 30 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_171; • ♂; W side Chunky Gal Mountain, Hwy 64, near scenic overlook; 35.0627°N, -83.6204°W; 6 May. 1999; M. Hedin, B. Dellinger leg.; MCH 99_019; – Swain Co. • 2♀; Nantahala River Gorge, Blowing Springs Cave; 10 Sep. 2001; J.D. Mayes leg.

Diagnosis. Several male features distinguish *Nesticus lowderi* from other members of the species group (and Appalachian clade), including the distinctive shape of the posterior keel of the forked tegular apophysis and the low sinuous paradistal process (Fig. 46A–D). Median bars, extending V-shaped upwards from the median septum and interrupting the epigynal pockets, diagnose *N. lowderi* females from other members of the species group and other common regional taxa (e.g., *N. reclusus*).

Description of ♂ holotype (SDSU_TAC000667). Carapace cream-colored, faint gray pigmentation behind ocular area leading to midline. Legs pale yellow / cream. Abdomen with many dark gray blotches on a pale cream background. All eyes approximately equal in size, except for AMEs, $\sim 1/2$ width of ALEs. Eyes with rings of dark pigment. CL 1.32, CW 1.2, abdomen length 1.48, total body length 2.8. Leg I total length 9.88 (2.71, 0.58, 2.9, 2.59, 1.1), leg formula 1423, leg I / CW ratio 8.2. Ventral paracymbial process consists of a large, basal lobe that broadens down length of paracymbium. Distal process somewhat spoon-shaped, dorsal process a low lobe, and the paradistal process consists of a sinuous, prolaterally directed extension with a heavily sclerotized anterior edge. Median apophysis rectangular with an anteriorly directed point and a sclerotized prolateral edge. Tegulum forked, with strong posterior keel including a wide lobe with a flattened edge. Distal tegular process crescent-shaped with a heavily sclerotized point directed anterolaterally, closely appressed to median apophysis. Distal tip of conductor bent and directed prolaterally.

♂ **Variation.** Males from different locations varied slightly in the shape of the basal fork of the tegular apophysis (Fig. 46A–D).

Description of ♀ paratype (SDSU_TAC000668). Carapace dusky cream to orange, with faint gray pigmentation behind ocular area leading to midline and around edges. Leg pale yellow / cream. Abdomen with paired dark gray blotches on a light gray background. All eyes approximately equal in size, except for AMEs, $\sim 1/2$ width of ALEs. Eyes with rings of dark pigment. CL 1.25, CW 1.11, abdomen length 1.46, total body length 2.71. Leg I total length 8.34 (2.41, 0.51, 2.39, 2.04, 0.99), leg formula 1423, leg I / CW ratio 7.5. Epigynal pockets interrupted by median bars that extend upwards V-shaped from base of median septum to nearly the top of the larger pocket (giving an overall appearance of an anchor, Fig. 47A–H). The presence of these bars forms septal grooves that lie directly adjacent to the median septum, and smaller pockets lateral to the V-shaped bars. Median septum slightly protruding posteriorly past lateral lobes. Spermathecae elongated and curved along lateral borders of epigynum, approximately banana-shaped. Ventrolateral sides of epigynal plate bulging (convex), as viewed from the side. Viewed dorsally, large internal lobes extend anteriorly and possess sclerotized rims. Interior margins directed inward diagonally towards the center of the epigynum.

♀ **Variation.** Epigynal structure fairly uniform across collecting locations (Fig. 47A–H). Blowing Springs Cave females are concolorous and relatively long-legged.

Distribution and natural history. Most populations are from the Chunky Gal, Tusquitee, and Valley River Mountains of western North Carolina (Fig. 42). Holler et al. (2020) attributed female specimens from Blowing Springs Cave to *Nesticus cooperi* (= *N. reclusus*); we instead have identified these as *N. lowderi* based on epigynal morphology, including the inward curve of the internal plates (Fig. 47A–H). This would represent a disjunct northern-most record for *N. lowderi* (Fig. 42) and should be confirmed with the collection of males and /or nuclear DNA data from this location. Another possibility is sympatry at this location.

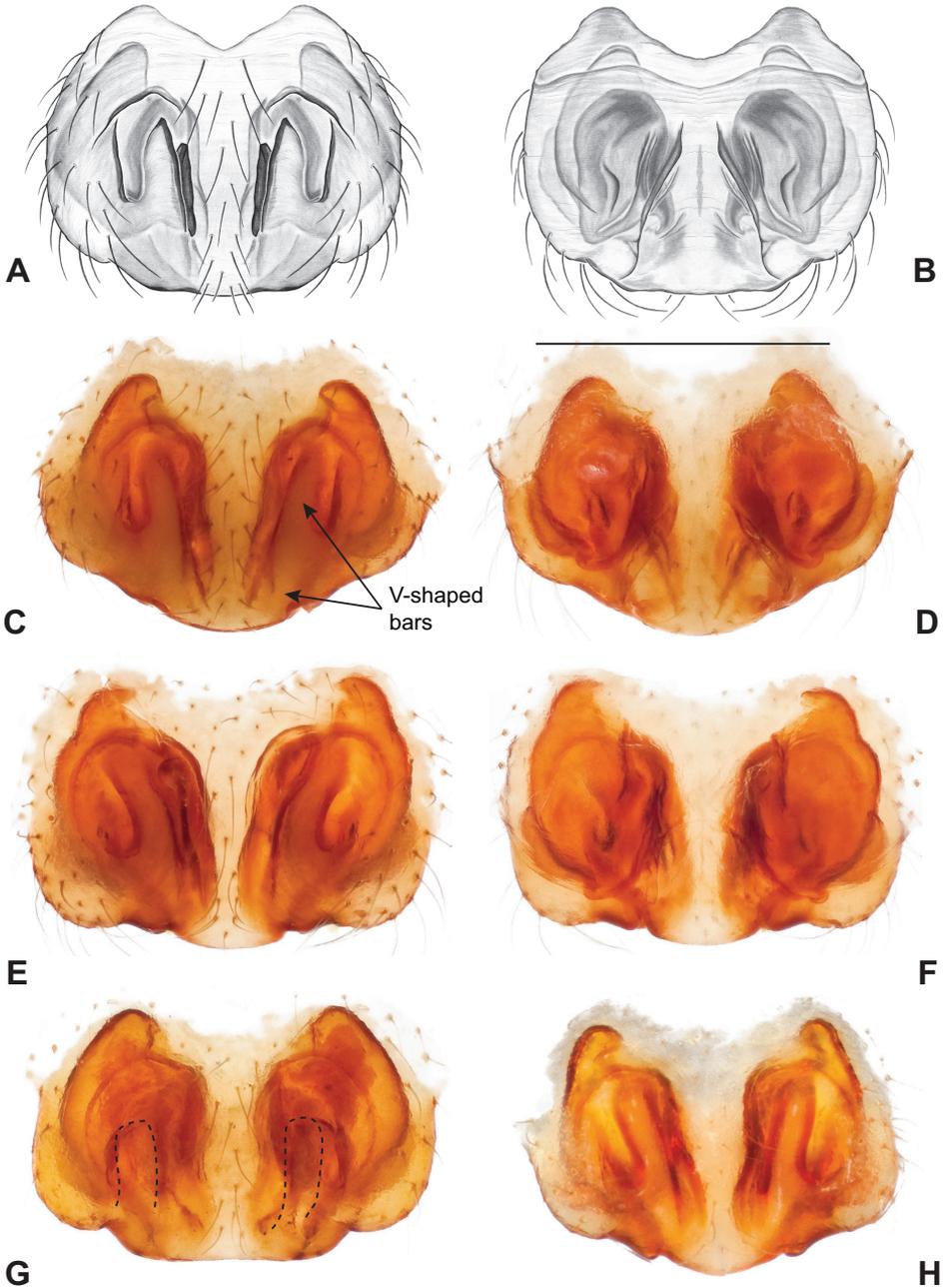


Figure 47. *Nesticus lowderi* sp. nov. epigynal variation. North Carolina, Clay Co., Chunky Gal Mountain, Chestnut Branch of Barnard's Creek, MCH 99_016 (SDSU_TAC000668) ventral (**A**), dorsal (**B**). North Carolina, Clay Co., Big Tuni Creek, MCH 02_171, ventral (**C**), dorsal (**D**). Swain Co., Blowing Springs Cave, ventral (**E**), dorsal (**F**). North Carolina, Clay Co., Fires Creek Road, Picnic Area along Fires Creek, MCH 07_108, ventral (**G**). North Carolina, Clay Co., Dave Barrett Fork of Eagle Fork Creek, MCH 02_143, ventral (**H**). Scale bar: 0.5 mm. Septal bars outlined in image **G** to better reflect actual specimen.

At Fires Creek (MCH 02_144), *Nesticus lowderi* (3♂, 5♀) was found in syntopy with *N. reclusus* (♂, 4♀); field notes read “30-minute survey, 3 persons, S-facing and N-facing rock fields”. Because we did not identify specimens directly in the field it remains unclear if these different species were truly syntopic or were segregated somehow at this location. Also, at least 15 immatures were collected at this location but were not identified to species because of sympatry.

Collection records suggest that this species is less common in the Chunky Gal Mountains than in the more westerly Tusquitee and Valley River Mountains.

Etymology. This species is named to recognize and honor Michael Lowder, faculty member at Stanly Community College, native North Carolinian, fan of western North Carolina, and collector of many Appalachian *Nesticus*. Michael was the first graduate student of MH, who remains forever grateful for our continued friendship and reflects on our early lab and field time together with great fondness.

Remarks. The extent of mitochondrial divergence observed in this taxon over a small geographic region (including only Chunky Gal, Tusquitee, and Valley River Mountains) is notable (Fig. 6).

***barri* group, including:**

Nesticus barri Gertsch, 1984

Nesticus furtivus Gertsch, 1984

This group includes the sister species *Nesticus barri* and *N. furtivus*, a clade supported by nuclear phylogenomics (Figs 3, 4) but not mitochondrial evidence (Fig. 6). Each species is phylogenomically distinctive, with minimal UCE gene tree conflict as evidenced by high gene and site CF values (Fig. 5).

The unique morphology of each species is discussed below. We do not attempt to identify diagnostic morphological features for this small species group.

Nesticus barri and *N. furtivus* are cave-dwelling species from the Tennessee / Alabama / Georgia (TAG) region, the former conspicuously widespread for a cave-restricted species, while *N. furtivus* is narrowly endemic to limestone caves from a single mountain (Fig. 49).

***Nesticus barri* Gertsch, 1984**

Fig. 48A, C

Nesticus barri Gertsch, 1984: 36, figs 121–123, 161–163; Hedin and Dellinger 2005: 3, figs 2–10; Snowman et al. 2010: fig. 1; Carver et al. 2016: fig. 2.

Nesticus valentinei Gertsch, 1984: 29, figs 150–152.

Material examined. Non type material: USA – Alabama, Jackson Co. • ♀; Fern Cave, AJK597; 1 Dec. 2018; M.L. Niemiller, M.E. Slay, T. Inebnit, J. Pinkley, J.

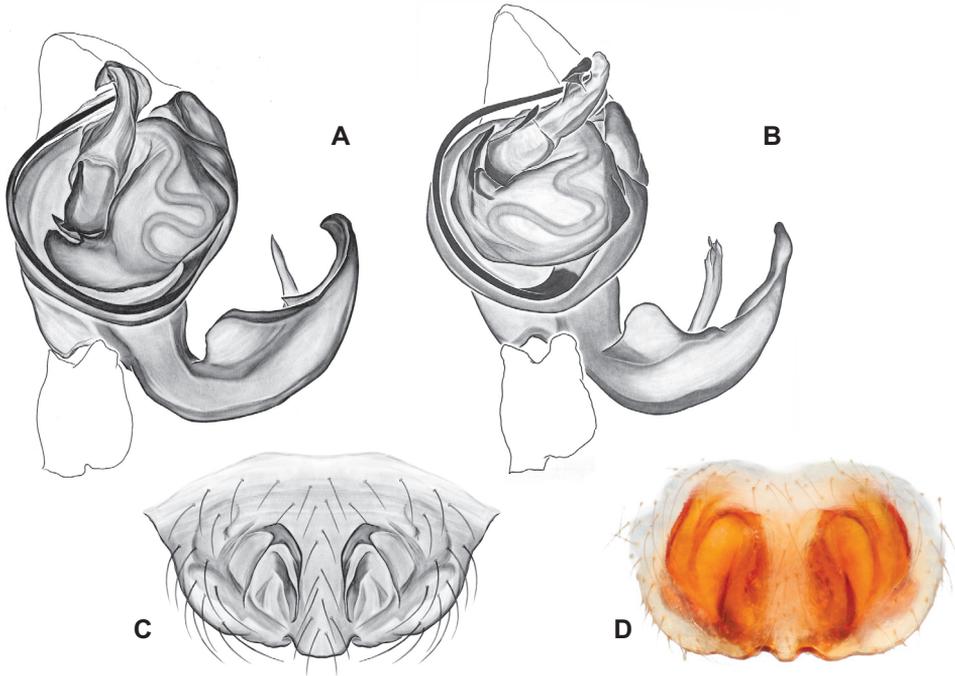


Figure 48. *Nesticus barri* and *N. furtivus* genitalia. *N. barri* – Tennessee, Marion Co., Tate Spring Cave, MCH 04_050, ♂ palp ventral (A), epigynum ventral (C). *N. furtivus* – Tennessee, Hamilton Co., Raccoon Mountain Caverns, SE Chattanooga, MCH 00_137, ♂ palp ventral (B), epigynum ventral (#1660) (D).

Lamb, P. Pattavina, K. Sapkota, B. Miller, N. Mann leg.; MLN 18–051.8; • ♀, 1 imm; Fern Cave, AJK597, bottom of cave; 1 Aug. 2008; J. Pinkley leg.; JP 08–AJK597.1; • ♀; Fern Cave, AJK597, Johnston entrance; 2 Jun. 2018; M.L. Niemiller, M.E. Slay, T. Inebnit, B. Miller, et al. leg.; MLN 18–020.8; • ♀; Fern Cave, AJK597, Morgue – past first Bat Room; 23 Jun. 2018; A. Hinkle, S. Pitts leg.; AH 18–001.2; • ♀; Fern Cave, AJK597, upper formation passage; 2 Jun. 2018; M.L. Niemiller, M.E. Slay, T. Inebnit, B. Miller, et al. leg.; MLN 18–020.26; • ♀; Fern Cave, AJK597, upper north passage; 3 Jun. 2018; M.L. Niemiller, M.E. Slay, T. Inebnit, B. Miller, et al. leg.; MLN 18–021.1; • 8 ♀; Guess Creek Cave, E Trenton; 25 Sep. 1992; M. Hedin, J. Hedin, S O’Kane leg.; • 2 ♀, 1 imm; Moody Cave, AJK1189; 18 Mar. 2019; M.L. Niemiller, J. Lamb, A. Hinkle leg.; MLN 19–014.20; • ♀, 1 imm; Tumbling Rock Cave, AJK171; 8 Mar. 2014; M.L. Niemiller, C.D.R. Stephen, K.S. Zigler, R. Miller, C. Borer, C. Maddux, J. Clark, V. Leray leg.; MLN 14–011.10; – **Alabama, Marshall Co.** • 9 ♀; Bishop Cave, N of Guntersville Dam; 25 Sep. 1992; M. Hedin, J. Hedin, S O’Kane leg.; • ♂, 2 ♀; Bishop Cave; 17 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_056; – **Tennessee, Franklin Co.** • ♂, 8 ♀; Keith Cave, S of Cowan; 24 Mar. 1995; M. Hedin, J. Hedin leg.; • 2 ♀, 2 imm; Little Crow Creek Cave, TFR15; 20 Sep. 2008; M.L. Niemiller, BT Miller, J. Miller, N. Mann leg.; MLN 08–041; •

♂, 3♀; Lost Cove Cave, N/NE of Sherwood; 23 Sep. 1992; M. Hedin, J. Hedin, S O’Kane leg.; • 2♂, 7♀; Salt River Cave, W of Gonce, Alabama; 24 Mar. 1995; M. Hedin, J. Hedin leg.; • 2♀, 1 imm; Sinking Cove Cave, TFR25; 15 Oct. 2016; N.S. Gladstone leg.; NSG 16–TFR25.10; – **Tennessee, Marion Co.** • ♂, 2♀; Tate Spring Cave, SE of Monteagle; 15 Aug. 2004; M. Hedin, L. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_050.

Diagnosis. The diagnosis of Gertsch (1984) is here modified to reflect a close phylogenetic relationship to *Nesticus furtivus*. These taxa share an overall similarity in features of the male paracymbium and shape of the median apophysis but differ in the shape of the tegular apophysis (Fig. 48A–D). In *N. barri* the tegulum is forked, with a basal projection shaped like a curved blade, and with a distal crescent-shaped tegular process lying close behind the quadrate median apophysis. The epigynum of *N. barri* is similar to that of distant relative *N. lowderi* in general structure (Fig. 47A–H), but with internal plates (viewed dorsally) not as long. Females are distinctly different from *N. furtivus*, as discussed below in the diagnosis for this latter species.

Variation. The shape of the basal tegular fork varies notably across cave locations. One male from Salt River Cave (MCH #2105) completely lacked a dorsal paracymbial process, without evidence that this was broken off. Variation in *Nesticus barri* epigynal morphology was illustrated in Hedin and Dellinger (2005), figs 2–8.

Distribution and natural history. Known from possibly hundreds of caves in northwest Alabama and south-central Tennessee (Fig. 49; Hedin and Dellinger 2005: fig. 1; Snowman et al. 2010: fig. 1; Carver et al. 2016: fig. 2). Carver et al. (2016) reported on the reproductive biology of this species.

Remarks. Based on consideration of morphology Hedin and Dellinger (2005) hypothesized that troglomorphic spiders from Tate Spring Cave, Tennessee represented a northern population of *Nesticus barri*. The mitochondrial data included here further support this hypothesis (Fig. 6).

Nesticus furtivus Gertsch, 1984

Fig. 48B, D

Nesticus furtivus Gertsch, 1984: 27, figs 97–99; Hedin and Dellinger 2005: 12, figs 15, 16.

Material examined. New collections from type locality: USA – **Tennessee, Hamilton Co.** • ♀; Raccoon Mountain Caverns, se Chattanooga; 28 Mar. 1993; M. Hedin, M. Wolinsky leg.; • ♂; Raccoon Mountain Caverns; 25 Jul. 2000; M. Hedin, D. Wood, B. Dellinger, S. Perlack leg.; MCH 00_137; • ♀; Raccoon Mountain Caverns; 19 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_063; **Non type material:** – **Marion Co.** • ♀; Hugden Branch Cave (TMN 127); 17 Apr. 2016; K.S. Zigler, P.R. Heald leg.; KSZ 15–570.

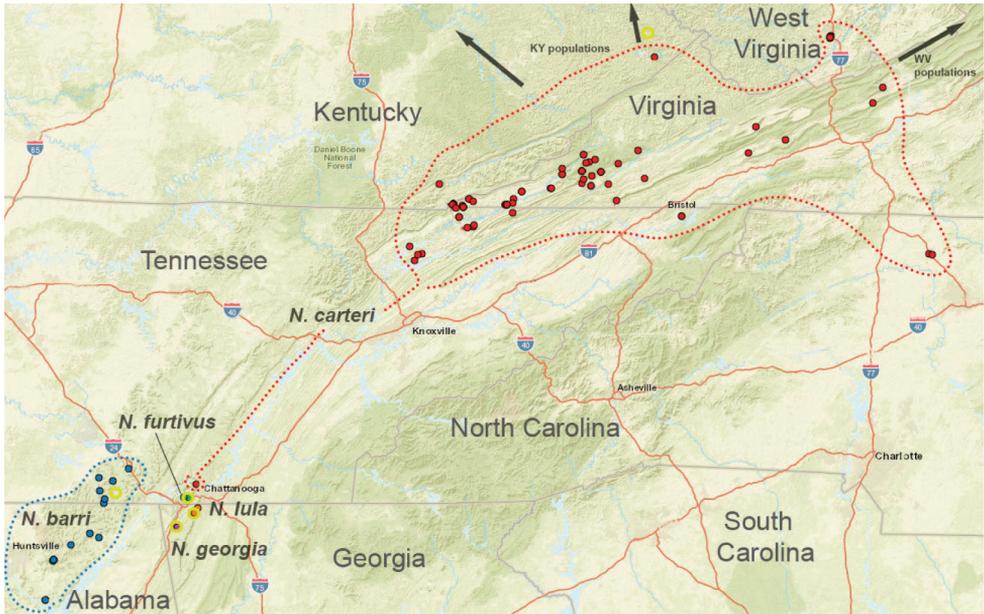


Figure 49. Distribution of *barri* and *carteri* groups. Type localities designated with yellow circles. State boundaries and major cities shown for geographic context. Dashed lines circumscribe known species distributions.

Diagnosis. Closely related to *Nesticus barri*, but the males differ in that the tip of the *N. furtivus* paracymbial dorsal process is finely forked, the shape of the basal tegular fork is broader (rather than blade-like), and the apical tegular fork is reduced and lacking a distinct tip (Fig. 48B). Female *N. furtivus* have a distinctly wide median septum that narrows to a conspicuous tip posteriorly (Fig. 48D).

Variation. The Hugden Branch Cave female specimen, representing the second known location for this species, is troglomorphic with an epigynum that closely matches females from the type locality.

Distribution and natural history. This troglomorphic species is known from two nearby caves from a single mountain in southeastern Tennessee, near Chattanooga (Fig. 49; Hedin and Dellinger 2005: fig. 1; Carver et al. 2016: fig. 2).

Carver et al. (2016) provide important natural history, reproductive biology, and abundance data for this rare species, extending earlier observations of Hedin and Dellinger (2005).

***carteri* group, including:**

Nesticus carteri Emerton, 1875

Nesticus georgia Gertsch, 1984

Nesticus lula Zigler & Milne, 2022

This small species group is strongly supported as monophyletic on both concatenated and coalescent phylogenomic trees (Figs 3, 4). The sister species *Nesticus georgia* and *N. lula* are recovered together with high support on the mitochondrial tree (Fig. 6), but they are separate from *N. carteri* populations, the latter fragmented into three distantly related mitochondrial clades (Fig. 6). This is one of the most notable examples of mitonuclear discordance in the Appalachian clade, here hypothesized to result from a lack of phylogenetic signal in the mitochondrial data at greater phylogenetic depths.

The unique morphology of each species is discussed below; we otherwise do not attempt to identify diagnostic morphological features for this small species group.

This group includes the geographically widespread *Nesticus carteri*, and short-range endemic sister species from caves of northwestern Georgia (*N. georgia*, *N. lula*). An intriguing southern population of *N. carteri* (Pitchfork Cave), which is highly disjunct from any other known *N. carteri* population, might bridge this biogeographic gap (Fig. 49).

***Nesticus carteri* Emerton, 1875**

Fig. 50A–J

Nesticus carteri Emerton, 1875: 279, pl. 1, fig. 28; Gertsch 1984: 34, figs 124, 125, 175–177.

Material examined. New collections from near type locality: – **Kentucky, Carter Co.** • 3♂, 13♀; Laurel Cave, Carter Caves State Park, 10 mi NE Olive Hill; 15 Sep. 1992; M. Hedin, S. O’Kane leg.; **Non type material:** – **Indiana, Crawford Co.** • ♂, ♀; Heron Cave, ca. 7 mi. S of Leavenworth; 12 Sep. 1996; J. J. Lewis leg.; • 9♀; Wallier Cave; 26 Apr. 1997; J. J. Lewis leg.; – **Kentucky, Pike Co.** • ♂, ♀; Lick Creek County Park, N of Hwy 460, NE of Belcher; 37.3996°N, -82.3057°W; 26 Jun. 2014; M. Hedin leg.; MCH 14_008; – **North Carolina, Surry Co.** • ♂; just E of Pilot Mtn State Park, Pilot Knob Park Rd; 36.3415°N, -80.4538°W; 1 Jun. 2016; M. Hedin, S. Derkarabetian, J. Starrett, M. Lowder leg.; MCH 16_036; • ♂, 6♀; Pilot Mtn State Park, near campground; 36.3479°N, -80.4732°W; 31 May. 2016; M. Hedin, S. Derkarabetian, J. Starrett, M. Lowder leg.; MCH 16_035; – **Tennessee, Claiborne Co.** • 2♀; Sour Kraut Cave, TCB46; 1 Jun. 2015; M.L. Niemiller, E.T. Carter, L.E. Hayter leg.; MLN 15–009.10; • ♀, 1 imm; Station Creek Cave, CGNHP; 6 Jun. 2019; K.S. Zigler, L.E. Trumbore leg.; KSZ 19–102; • 2♀; English Cave, 0.9 mi. S Hamilton School; 25 Sep. 1991; M. Hedin, K. Crandall leg.; • 10♀; English Cave, 20 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♀; Kings Saltpeter Cave, TCB52; 30 May. 2015; M.L. Niemiller, C.D.R. Stephen, E.T. Carter, A.S. Engel, S. Engel, P.B. Hart leg.; MLN 15–008.34; • 2♀, 2 imm; Coonsies Creek Cave, TCB57; 23 Mar. 2016; M.L. Niemiller, C.D.R. Stephen leg.; MLN 16–023.13; – **Tennessee, Hamilton Co.** • 3♂, 7♀; N of Tiftonia, near Pitchfork Cave; 22 Sep. 1992; M. Hedin leg.; – **Tennessee, Hancock Co.** • ♂, 6♀; Hwy 63, S Mulberry Gap; 36.5659°N, -83.2465°W; 21 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_070;

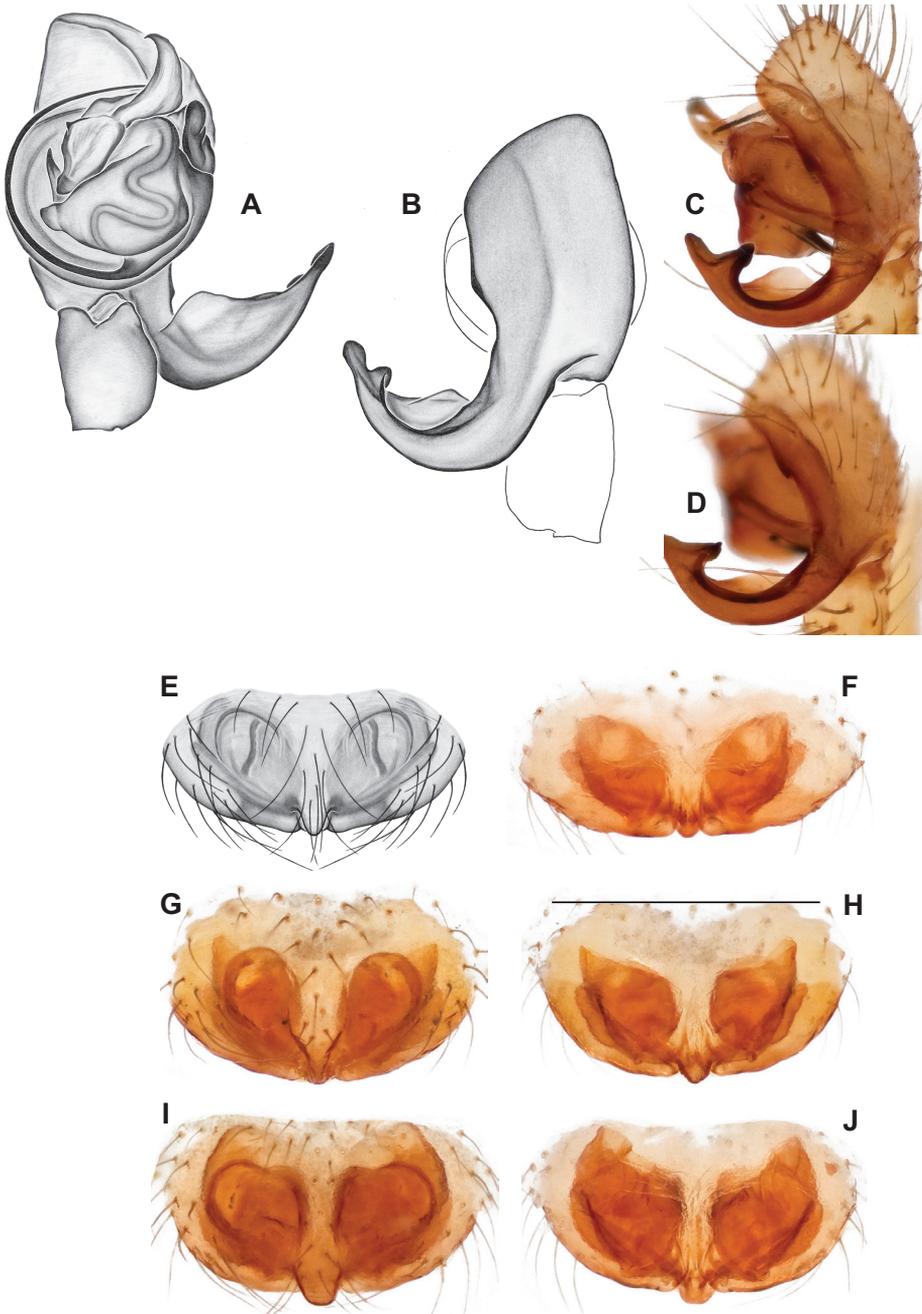


Figure 50. *Nesticus carteri* ♂ palps. Tennessee, Hamilton Co., near Pitchfork Cave, MCH specimen #1571, ventral (A), dorsal (B). C Virginia, Scott Co., Cliff Mountain, MCH 04_028, dorsal D Tennessee, Hancock Co., S of Mulberry Gap, MCH 05_070, dorsal. *N. carteri* epigynal variation. Tennessee, Hamilton Co., near Pitchfork Cave, MCH specimen #1580, ventral (E), dorsal (F). Tennessee, Hancock Co., S of Mulberry Gap, MCH 05_070, ventral (G), dorsal (H). Virginia, Scott Co., Cliff Mountain, MCH 04_028, ventral (I), dorsal (J). Scale bar: 0.5 mm.

– **Tennessee, Sullivan Co.** • 12♀; Bristol Caverns, SE of Bristol; 18 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♀, 1 imm; Bristol Caverns, TSL1; 17 Oct. 2017; N.S. Gladstone leg.; NSG 17–TSL1.9; – **Tennessee, Union Co.** • 2♀; Big Cave, TUN10; 22 Mar. 2015; M.L. Niemiller, C.D.R. Stephen leg.; MLN 15–005.18; • 5♀; Oaks Cave, TUN5; 23 Mar. 2015; M.L. Niemiller, C.D.R. Stephen, E.T. Carter, LE Hayter leg.; MLN 15–007.3; • ♂, 3♀; Rogers Hollow Cave, TUN23; 22 Mar. 2015; M.L. Niemiller, C.D.R. Stephen leg.; MLN 15–002.6; • 3♀; Wright Cave, TUN9; 21 Mar. 2015; M.L. Niemiller, C.D.R. Stephen, E.T. Carter, JP McClendon leg.; MLN 15–001.10; – **Virginia, Giles Co.** • 2♀; Salamander Cave, CGNHP; 26 Jul. 2019; K.S. Zigler, L.E. Trumbore leg.; KSZ 19–169; • ♀, 10 imm; Sugar Run Cave System, Birthday Entry; 27 Aug. 2018; T. Malabad leg.; – **Virginia, Lee Co.** • 3♀, 5 imm; Bacon Cave; 8 Mar. 2017; T. Malabad leg.; • ♀, 5 imm; Bacon Cave; 21 Mar. 2018; T. Malabad leg.; • ♀; Bacon Cave; 15 Nov. 2019; T. Malabad leg.; • 2♀; Bacon Cave; 22 Oct. 2019; T. Malabad, K. Kosič Ficco leg.; • ♂; Bacon Cave; 3 Mar. 2020; T. Malabad, K. Kosič Ficco, R. Blackwell, L. Young leg.; • ♀; Bacon Cave; 10 Mar. 2021; T. Malabad, W. Orndorff, Z. Orndorff leg.; • 2♂, 15♀; Bowling Cave, SW of Pineville; 19 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♂1 imm; Burja Cave; 29 Apr. 2017; T. Malabad leg.; • 1 imm; Burja Cave; 1 Jul. 2017; T. Malabad leg.; • ♂; Burja Cave; 19 Aug. 2017; T. Malabad leg.; • ♂; Burja Cave; 1 Dec. 2018; T. Malabad leg.; • 6♀; Cave Spring Recreational Area, NE of Dryden; 36.8033°N, -82.921°W; 21 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_071; • 2♂, 7♀; Cumberland Gap National Historic Park, Skylight Cave; 20 Sep. 1992; M. Hedin, S. O’Kane leg.; • 2♀, 2 imm; Gallohan No. 2 Cave; 29 Jan. 2018; T. Malabad leg.; • ♂1 imm; Gap Cave, CGNHP; 31 Aug. 2019; K.S. Zigler, et al. leg.; KSZ 19–235; • 2♀; Gibson Frazier Cave, 8 miles southwest of Jonesville, VA; 20 Nov. 2019; T. Malabad, R. Blackwell leg.; • 4♀, 2 imm; Indian Burial Cave; 30 Jan. 2018; T. Malabad leg.; • ♂, ♀, 3 imm; Indian Cave, CGNHP; 5 Jun. 2019; K.S. Zigler, L.E. Trumbore leg.; KSZ 19–165; • ♀, 2 imm; Indian Cave, CGNHP; 9 Jul. 2019; K.S. Zigler, L.E. Trumbore leg.; KSZ 19–78; • 2♀, 2 imm; Little Saltpeter Cave, CGNHP; 11 Jul. 2019; K.S. Zigler, L.E. Trumbore leg.; KSZ 19–13; • 2♀; Litton Cave No. 1, 4.8 miles west of Sticklelyville, VA; 10 Mar. 2021; T. Malabad, W. Orndorff, Z. Orndorff leg.; • ♂, 5♀; Litton Cave No. 2, 6.3 miles east of Jonesville, VA; 24 Mar. 2021; T. Malabad, W. Orndorff leg.; • 2♀, 2 imm; Pack Rat Cave, CGNHP; 10 Jul. 2019; K.S. Zigler, LE Trumbore leg.; KSZ 19–60; • ♀; Robertson Cave No. 1, 1.75 miles northeast of Wheeler, VA; 17 Sep. 2020; T. Malabad, K. Kosič Ficco, M. Ficco leg.; • ♀; Robertson Cave No. 2, 1.75 miles northeast of Wheeler, VA; 28 Apr. 2021; T. Malabad, K. Kosič Ficco, W. Orndorff, M. Ficco leg.; • 8♀; Secret Cave, 1.3 miles southeast of Dryden, VA; 11 Mar. 2021; T. Malabad, W. Orndorff, Z. Orndorff leg.; • ♂, 3♀; Secret Cave; 22 Apr. 2021; T. Malabad, W. Orndorff, Z. Orndorff, J. Lewis, L. Young leg.; • 2♀, 3 imm; Skylight Cave, CGNHP; 5 Jun. 2019; K.S. Zigler, LE Trumbore leg.; KSZ 19–132; • 6 imm; Spangler Cave, west of Jonesville, VA; 30 Jan. 2018; T. Malabad leg.; • 5♀; Spangler Cave; 27 Jan. 2020; T. Malabad, R. Blackwell, Rick Reynolds leg.; • ♀, 1 imm; Young–Fugate Cave, southwest of Wheeler, VA; 26 Aug. 2015; W. Orndorff leg.; • 2 imm;

Young–Fugate Cave; 14 Sep. 2016; T. Malabad leg.; • ♀; Young–Fugate Cave; 22 Oct. 2018; T. Malabad leg.; • ♂, 3♀; Young–Fugate Cave; 28 Oct. 2019; T. Malabad, K. Kosič Ficco leg.; • ♂, 3♀; Young–Fugate Cave, Fugate entrance; 24 Jun. 2020; T. Malabad, A. Malabad leg.; – **Virginia, Rockbridge Co.** • 3♂, 13♀; Dollhouse Cave, Natural Bridge, E of Springfield; 16 Sep. 1992; M. Hedin, S. O’Kane leg.; – **Virginia, Scott Co.** • 4♀; Alley Cave (entrance sink), E of Natural Tunnel State Park; 19 Sep. 1992; M. Hedin, S. O’Kane leg.; • 2♀; Alley Cave (entrance sink), E of Natural Tunnel State Park; 22 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_072; • ♀; Big Entrance Crawl Cave; 3 May. 2017; T. Malabad leg.; • ♂, 9♀; Cliff Mountain, Dry Branch, County Road 655, NE of Duffield; 36.7495°N, -82.7787°W; 7 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_028; • ♂, 2♀, 7 imm; Grisby Cave; 7 Mar. 2017; T. Malabad leg.; • ♂; Hill Cave, 5.2 miles northeast of Duffield, VA; 3 Mar. 2020; T. Malabad, K. Kosič Ficco, R. Blackwell, L. Young leg.; • ♂, 14♀, 2 imm; Hwy 23/58/421 at Moccasin Gap, near Weber City; 36.6338°N, -82.555°W; 22 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_073; • ♂, 2 imm; Kerns Cave; 16 Sep. 2015; W. Orndorff leg.; • 2♀, 1 imm; Kerns No. 1 Cave, northwest of Fort Blackmore, VA; 7 Mar. 2017; W. Orndorff leg.; • 4♀; Kerns No. 1 Cave; 4 Mar. 2020; T. Malabad, K. Kosič Ficco, R. Blackwell, L. Young leg.; • ♀; Spurlock Cave, northeast of Duffield, VA; 17 Dec. 2020; T. Malabad, K. Kosič Ficco, M. Ficco leg.; • 2♀; Summer Shaft, west of Dungannon, VA; 10 Sep. 2020; T. Malabad, K. Kosič Ficco, M. Ficco leg.; – **Virginia, Smyth Co.** • 3♂, 14♀; Atwell’s Tunnel Cave, N of Nebo; 17 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♀; Beaver Creek Cave; 9 Dec. 2014; E. Koertge leg.; – **Virginia, Tazewell Co.** • ♀; Whitt Cave, southwest of Tazewell, VA; 6 May. 2021; T. Malabad, K. Kosič Ficco, M. Ficco leg.; – **Virginia, Wise Co.** • ♀; above Guest River, County Road 660, 3 mi. S of County Road 658, SE of Coeburn; 36.9009°N, -82.4146°W; 7 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_027; • ♀; Cloud Hole Cave, SW of East Stone Gap, VA; 18 Dec. 2020; T. Malabad, K. Kosič Ficco, M. Ficco leg.; • ♀; Getting Warmer Cave, NE of Big Stone Gap, VA; 24 May. 2020; T. Malabad, K. Kosič Ficco, M. Ficco, Sara Fleetwood, P. Schuchardt leg.; • ♀; Parsons Cave, southeast of East Stone Gap, VA; 29 Jan. 2020; T. Malabad, K. Kosič Ficco, R. Blackwell, Rick Reynolds, L. Young leg.; • ♀; Space Turtles Cave, NE of Big Stone Gap, VA; 13 Jun. 2020; T. Malabad, K. Kosič Ficco, M. Ficco, P. Schuchardt leg.; • ♀, 8 imm; Wildcat Caverns; 14 Sep. 2016; W. Orndorff leg.; – **West Virginia, Kanawha Co.** • 2♀; Kanawha SF, Davis Creek campground; 38.2474°N, -81.6586°W; 24 Jun. 2014; M. Hedin leg.; MCH 14_003; – **West Virginia, Mercer Co.** • 3♂, ♀, 1 imm; Camp Creek State Park, along Mash Fork; 37.5039°N, -81.1343°W; 4 Jun. 2016; M. Hedin, S. Derkarabetian, J. Starrett leg.; MCH 16_050; • ♂, 19♀; Camp Creek State Park, vic Campbell Falls trailhead; 37.5092°N, -81.1337°W; 15 Sep. 1992; M. Hedin, S. O’Kane leg.; • 4♂, 5♀; Camp Creek State Park, vicinity Blue Jay campground; 37.5137°N, -81.1309°W; 25 Jun. 2014; M. Hedin leg.; MCH 14_007; • 4♀; Camp Creek State Park, near campground; 37.5019°N, -81.1357°W; 13 Aug. 2007; M. Hedin, R. Keith leg.; MCH 07_095.

Diagnosis. Male palp with a distinctive elongate conductor, with a tip that lacks the strong distal fold found in other Appalachian taxa. Strongly concave median apophysis with medial point, tegular apophysis with a shallow fork, basal branch just a small lobe (Fig. 50A–D). Paracymbium simple with a well-sclerotized, short paradistal process of various shape. Epigynum distinctive, wider than long with broad lateral pockets and an obviously pointed median septum (Fig. 50E–J).

Variation. The paradistal dorsal process of the paracymbium varies in shape from very low and inconspicuous (Fig. 50A–D), to spoon-like, to rectangular (Gertsch fig. 124). The distal (highly sclerotized) fork of the tegular apophysis also varies in length and shape, from nearly straight to more curved. Males from the disjunct Pilot Mountain and Pitchfork Cave locations (Fig. 49) fall within this range of variation.

Minor variation was observed in the shape of the epigynal median septum (sometimes with a median bulge, then narrowing distally, viewed ventrally, Fig. 50E–J), but no obvious geographic trends were apparent.

Distribution and natural history. This species has the largest known geographic distribution of any Appalachian *Nesticus* species, ranging from southern Tennessee (near Chattanooga) to southern Indiana, east to West Virginia, and southeast towards Winston-Salem (Fig. 49). Because we have collected this species from near surface habitats at relatively low elevations, we hypothesize that this taxon can withstand slightly drier situations, perhaps explaining this relatively broad distribution.

The southern Pitchfork Cave population is highly disjunct from all other more northerly records; this is possibly an artifact of insufficient collecting effort on the eastern edge of the Cumberland Plateau in east-central Tennessee (Fig. 49). UCE data indicate that the Pitchfork Cave population is genetically divergent (on a relatively long branch), and sister to all remaining *Nesticus carteri* populations (Figs 3, 4).

This species is known from both caves (both deeper and twilight situations) and dark, relatively moist near-surface habitats (mostly void spaces in rock piles). As noted above, *Nesticus carteri* has been collected in near syntopy with *N. holsingeri* at Alley Cave, Virginia, where the former is found in a talus sink leading to the cave entrance, the latter collected from the dark zone of the cave.

Remarks. This species is not recovered as monophyletic on mitochondrial gene trees but is instead fragmented into three separate clades (Fig. 6).

Nesticus georgia Gertsch, 1984

Fig. 51A–C

Nesticus georgia Gertsch, 1984: 39, figs 156–158, 164–166.

Material examined. Type material: *Holotype*: USA – **Georgia, Dade Co.** • ♂ holotype; Sitton's Cave, near Trenton; 28 Nov. 1952, E.J. Kuenzler leg.; AMNH. **New collections from type locality:** – **Dade Co.** • ♂, ♀; Sitton's Cave, 1 mi E of Trenton; 30 Sep. 1991; M. Hedin, K. Crandall leg.; • 2♂, 3♀; Sitton's Cave; 23 Sep. 1992; M. Hedin, S. O'Kane leg.

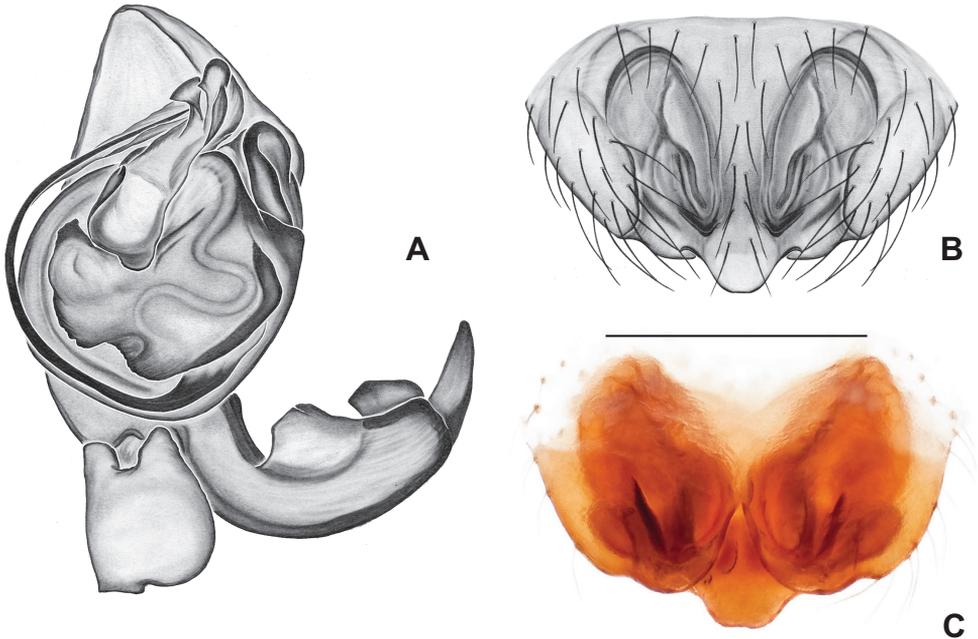


Figure 51. *Nesticus georgia* genitalia **A** Georgia, Dade Co., Sitton's Cave, MCH specimen #1015, ♂ palp, ventral. Georgia, Dade Co., Sitton's Cave, MCH specimen #1014, epigynum, ventral (**B**), dorsal (**C**). Scale bar: 0.5 mm.

Diagnosis. Nearly eyeless, long-legged taxon. Male palp most similar to that of *Nesticus lula*, but with a spatulate tegular apophysis and details of the distal edge of the paracymbial ventral process with a sclerotized process projecting dorsally (Fig. 51A). Ventral epigynum very close to similarly troglomorphic *N. lula*, difficult to separate based on epigynal morphology alone (compare Fig. 51B, C to Zigler and Milne 2022: figs 2, 3). Both *N. georgia* and *N. lula* are also similar in overall epigynal morphology to the more distantly related (but geographically proximate) *N. barri* (Fig. 48C).

Distribution and natural history. Known only from a handful of limestone caves from three adjacent counties in northwest Georgia (Fig. 49; Hedin and Dellinger 2005: fig. 1; Carver et al. 2016: fig. 2). Reeves (1999) summarized natural history information (microhabitat preference, fecundity, prey items, etc.) for topotypic *Nesticus georgia*.

Nesticus lula Zigler & Milne, 2022

Fig. 49

Nesticus lula Zigler & Milne, 2022: 293, figs 1A, C, 2, 3, 7.

Material examined. Type material: Holotype: USA – Georgia, Walker Co. • ♂; Lula Falls Cave (GWK617); 15 Apr. 2014; K.S. Zigler, L. Carver, L. Lyles leg.; KSZ 13–169

(SDSU_G2084). **Non type material:** – Walker Co. • ♂; Bee Rock Cave (GWK123); 31 May. 2015; K.S. Zigler, T. Lichtefeld, M. Abercrombie leg.; KSZ 15–388.

Diagnosis. Morphological diagnosis as in Zigler and Milne (2022).

Distribution and natural history. This troglomorphic taxon is currently known from only two caves in northwestern Georgia (Fig. 49; Zigler and Milne 2022: fig. 7).

***reclusus* group, including:**

Nesticus sheari Gertsch, 1984

Nesticus dellingeri sp. nov.

Nesticus jonesi Gertsch, 1984

Nesticus binfordae sp. nov.

Nesticus dykemanae sp. nov.

Nesticus bishopi Gertsch, 1984

Nesticus stupkai Gertsch, 1984

Nesticus reclusus Gertsch, 1984

Phylogenomic structure indicates three subclades within this larger group, including a distinctive *Nesticus sheari* sister to all other species, and a close-knit *N. dellingeri* subgroup sister to a *N. reclusus* subgroup. This overall structure is strongly supported by both concatenated and coalescent UCE analyses (Figs 3, 4). Relationships within the *N. reclusus* subgroup are particularly challenging, where concatenated versus coalescent UCE analyses differ in resolution, and both differ from morphology and mitochondrial evidence.

The mitochondrial data do not support the overall *reclusus* group as monophyletic, and although several species are recovered as monophyletic, their interrelationships vary strongly from that suggested by nuclear data (Fig. 6). Also, some species supported by nuclear evidence are intermixed on mitochondrial trees, suggesting a role for mitochondrial introgression. These patterns are more fully discussed in the species accounts below.

Male genital morphology suggests common ancestry for this complex of eight species (Fig. 52). The male tegular apophysis is approximately S-shaped (with modifications), and the ventral paracymbial process includes an associated ventromedial process that varies in shape (although this is mostly lacking in *Nesticus dykemanae*). In our discussion of this group below we refer to a male paracymbial paradistal process; it is possible that this represents a distally migrated dorsal process (Fig. 52).

Species in this group are distributed in the montane southern Blue Ridge west of the Asheville Basin, except for the geographically disjunct *Nesticus jonesi* known from a single cave in northern Alabama (Fig. 53). Groups of taxa show interesting parallel geographic distributions in the southern Blue Ridge. This includes a southeastern *N. dellingeri* sister to taxa from the Great Smoky Mountains, a southern *N. bishopi* related to *N. stupkai* also from the Great Smoky Mountains, and *N. reclusus* which also spans from south to north (Fig. 53).

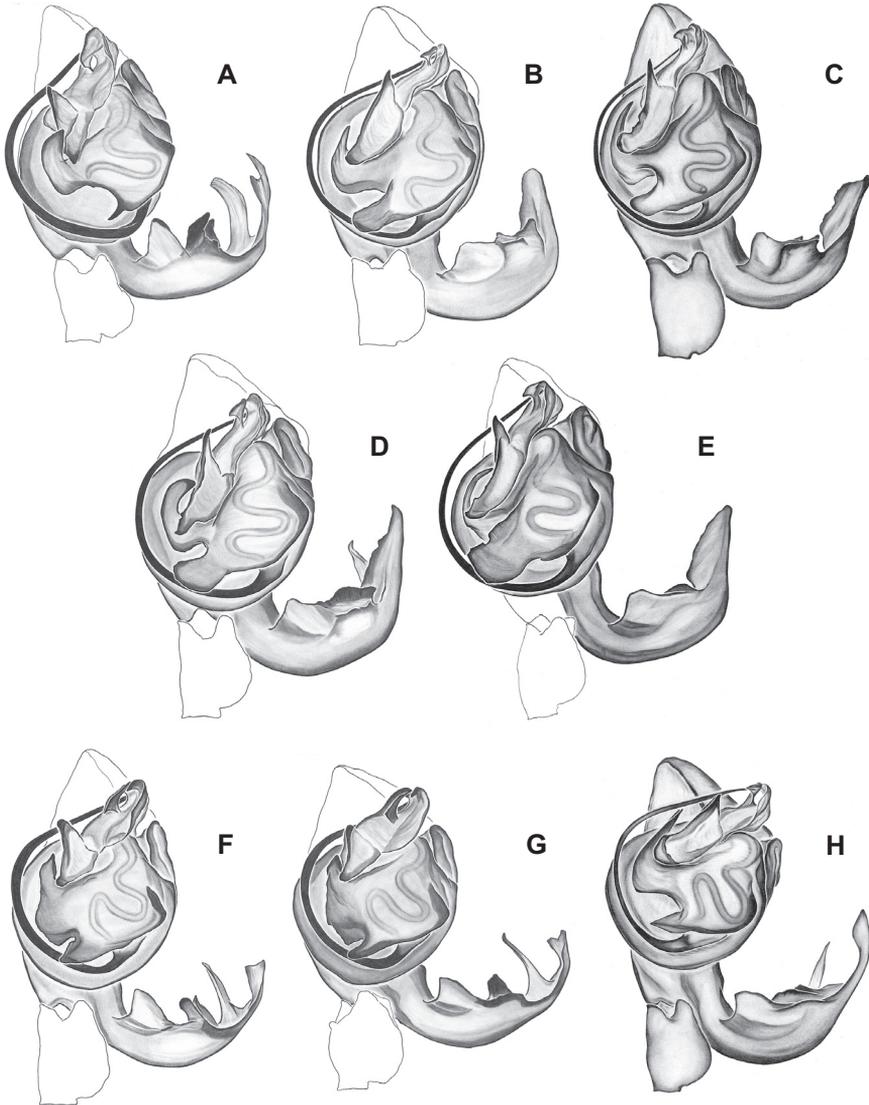


Figure 52. Comparative ♂ morphology of *reclusus* group species **A** *Nesticus sheari* **B** *N. dellingeri* **C** *N. jonesi* **D** *N. binfordae* **E** *N. dykemanae* **F** *N. bishopi* **G** *N. stupkai* **H** *N. reclusus*. All views ventral. See subsequent figures for specimen locations and voucher details.

***Nesticus sheari* Gertsch, 1984**

Fig. 54A–F

Nesticus sheari Gertsch, 1984: 32, figs 79–81, 135–137.

Material examined. Type material: Holotype: USA – North Carolina, Graham Co. • ♂ holotype; Joyce Kilmer Memorial Forest, Poplar Cove; 30 May 1975; W.A. Shear

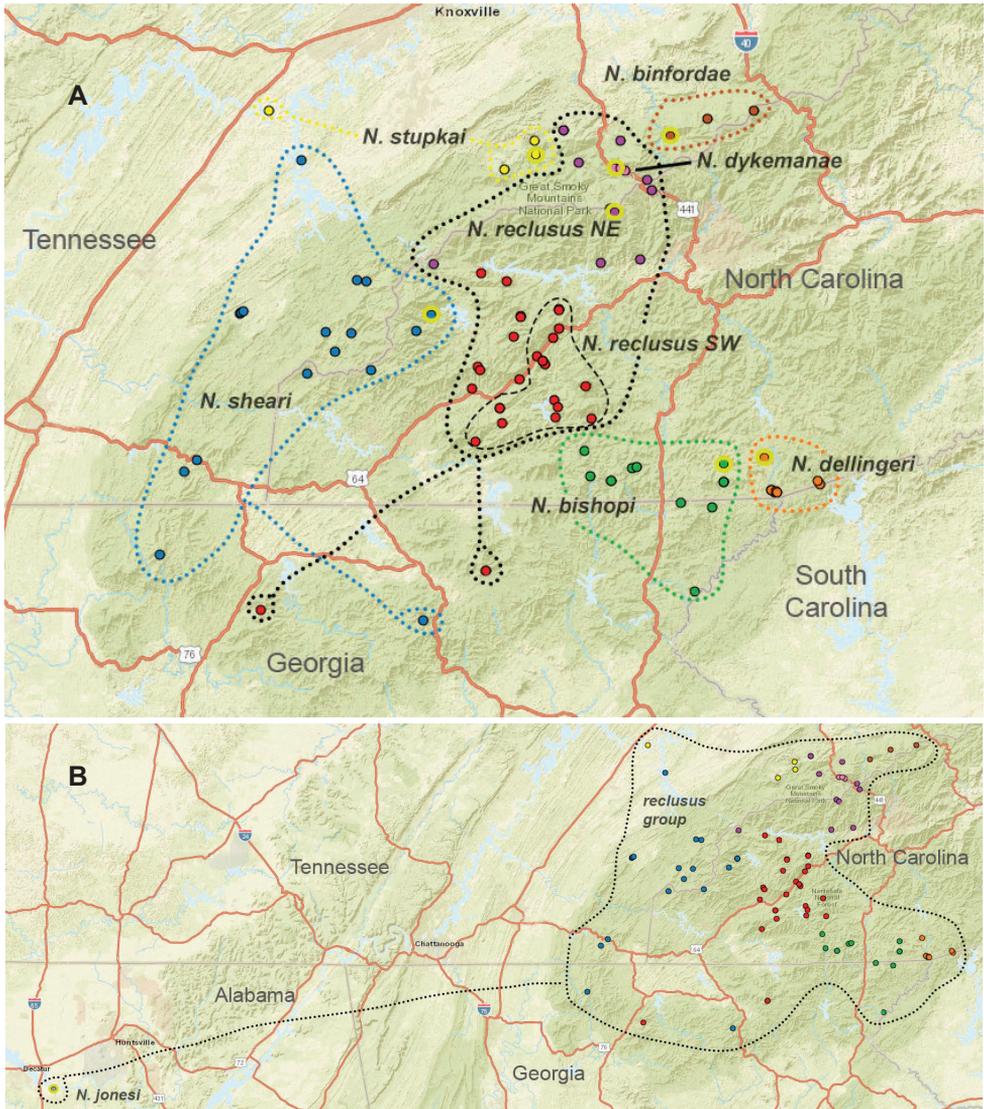


Figure 53. A distribution of *reclusus* group species **B** distribution of *Nesticus jonesi*. Type localities designated with yellow circles. State boundaries and major cities shown for geographic context. Dashed lines circumscribe known species distributions. The geographic distribution of “*N. cooperi*-like” populations is circled; this includes some female-only locations which are inside this distribution and included here only for purposes of graphical convenience.

leg.; AMNH; **New collections from type locality:** – **North Carolina, Graham Co.** • 3♂, 7♀; Joyce Kilmer Memorial Forest, NW of Robbinsville; 35.3585°N, -83.9291°W; 1 Aug. 1992; M. Hedin leg.; • 4♀; Joyce Kilmer Memorial Forest, NW of Robbinsville; 35.3585°N, -83.9291°W; 28 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_164; • 2♀; Joyce Kilmer Memorial Forest, NW of Robbinsville; 35.3585°N,

-83.9291°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_115; **Non type material.** – **Georgia, Fannin Co.** • 19♀; Cohutta Wilderness, Cowpen Trail, NW Three Forks Mountain trailhead; 34.8905°N, -84.5715°W; 22 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_151; **Georgia, Union Co.** • 4♀; Sosebee Cove State Natural Area, off Hwy 180, 2 mi W jct Hwy 19, S Blairsville; 34.7617°N, -83.9482°W; 18 Apr. 1994; M. Hedin leg.; – **North Carolina, Cherokee Co.** • 4♀; along Tipton Creek, 1.2 mi. SNC/TN stateline; 35.2503°N, -84.0724°W; 26 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_158; • 2♀; FR 50 Shuler Creek, below Wolf Ridge; 35.2424°N, -84.2227°W; 17 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_054; – **North Carolina, Graham Co.** • ♂, 2♀; along Wright Creek, S Santeetlah Creek, W of Seven Springs Gap; 35.3258°N, -83.9647°W; 19 Aug. 1991; B. Dellinger, D. Loch leg.; – **Tennessee, Loudon Co.** • ♂, ♀; Blankenship Cave, TLN1; 25 Jan. 2014; M.L. Niemiller, E.T. Carter leg.; MLN 14-006; – **Tennessee, Monroe Co.** • ♂, 2♀, 2 imm; Alans Hideaway Cave, TMO9; 16 Nov. 2013; M.L. Niemiller, E.T. Carter, M. Finkle leg.; MLN 13-079.1; • 2♀; along North River, FR 217, Unicoi Mountains; 35.3215°N, -84.1199°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_112; • ♂, 16♀, 4 imm; along Tellico River, near Bald River Falls; 35.3248°N, -84.1787°W; 26 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_157; • 7♀; Bald River, FR 126, E of Holly Flats campground; 35.2855°N, -84.1586°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_113; • 2♂, 8♀; Citico Creek near confluence with Flat Creek; 35.4252°N, -84.1047°W; 27 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_160; • ♀; Doublecamp Creek, 0.5 mi. E confluence with Citico Creek, Unicoi Mountains; 35.4224°N, -84.0847°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_114; • ♀, 4 imm; Gay Cave, TMO3; 16 Nov. 2013; M.L. Niemiller, C.D.R. Stephen, M. Finkle leg.; MLN 13-077; • 3♀, 5 imm; Lick Creek Cave, TMO8; 16 Nov. 2013; M.L. Niemiller, C.D.R. Stephen, E.T. Carter, M. Finkle leg.; MLN 13-078; – **Tennessee, Polk Co.** • ♂, 10♀; FR 221, N of Peavine Mountain, vicinity Big Frog Mountain Wilderness; 35.0531°N, -84.5139°W; 23 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_152; • ♂; S Ocoee River at Thunder Rock Road, off Hwy 64; 35.0743°N, -84.4852°W; 17 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_052.

Diagnosis. The diagnosis of Gertsch (1984) is here modified to recognize relationships to other *Nesticus reclusus* group members. Male paracymbium with the combination of translucent blade-like paradistal process, distal process with twisted, tubular tip; well-sclerotized, toothlike dorsomedial process adjacent to small flange-like ventromedial process (Fig. 54A, B). These paracymbium characters are similar to those found in *N. bishopi* and *N. stupkai*. Male tegular apophysis with a general S-curve, distal process a truncate curving blade, basal process nipple-like. Acute distal median apophysis. In females the posterior end of the epigynal median septum is “squared-off” on three sides, like a chisel (Fig. 54C–F), projecting inwards towards the abdomen.

Variation. In males from non-type locations the ventromedial process is more confluent with the ventral process (less displaced medially) and more elongate.

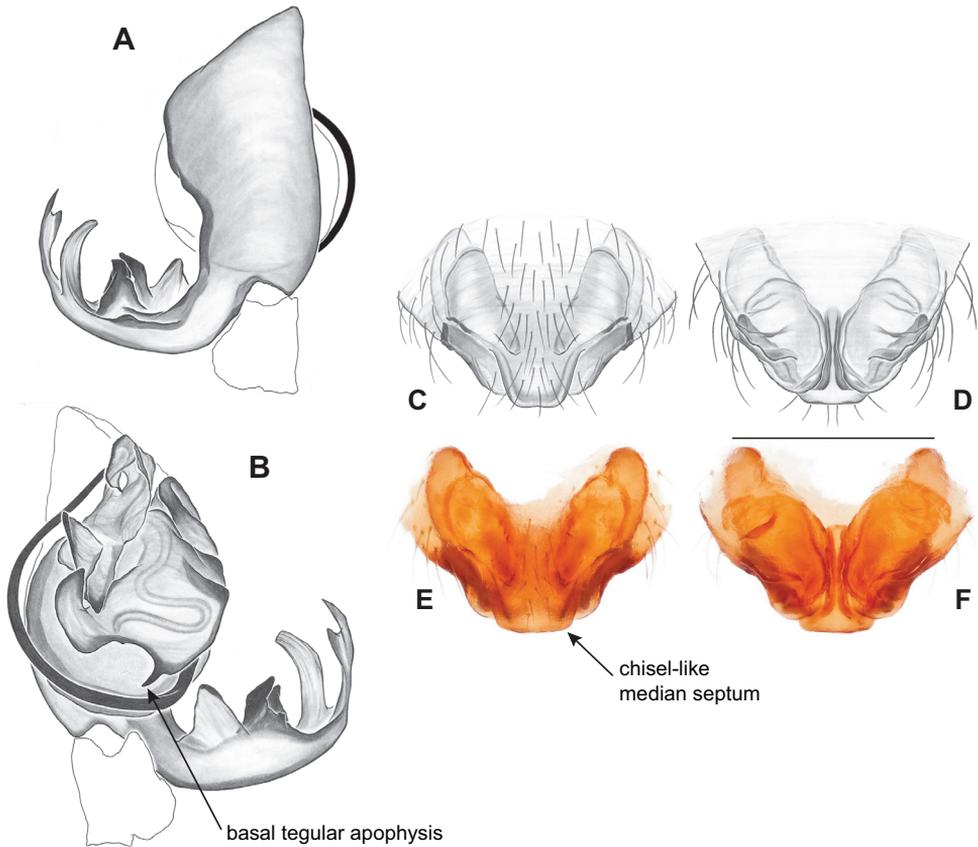


Figure 54. *Nesticus sheari* genitalia. Tennessee, Polk Co., N of Peavine Mountain, vicinity Big Frog Mountain Wilderness, MCH 02_152, ♂ palp, dorsal (A), ventral (B). Tennessee, Polk Co., vicinity Big Frog Mountain Wilderness, MCH 02_152, epigynum, ventral (C), dorsal (D). Georgia, Union Co., Sosebee Cove State Natural Area, MCH specimen #1995 epigynum, ventral (E), dorsal (F). Scale bar: 0.5 mm.

Different populations exhibit very little epigynal variation despite a large and fragmented geographic distribution. Some adult females from Holly Flats campground are approximately one-half the size of other adult females.

Distribution and natural history. Previously known only from the type locality at Joyce Kilmer Memorial Forest, which now represents one of the easternmost known records for the species. The species distribution almost forms a circle in the montane uplands that surround the Ducktown lowlands (lacking the eastern edge), with disjunct Cohutta, Sosebee Cove, and Blankenship Cave populations (Fig. 53). It appears to surround the geographic distribution of the more narrowly distributed *Nesticus bondi*. Previously thought to be a strictly montane taxon, but we report here several important new cave records.

As an example of natural history we include here field notes for Cohutta Wilderness (MCH 02_151) where we collected 19♀. Notes read “small drainage in pine

forest, rocks in drainage, moist but not running water”, where spiders were collected from beneath rocks.

As discussed above *Nesticus sheari* (4♀) was found in sympatry with *N. bondi* (♂, 6♀) at Tipton Creek. Because we did not identify specimens directly in the field, it remains unclear if these different species were found side-by-side or were perhaps somehow segregated by microhabitat. *Nesticus sheari* was also collected in sympatry with the nesticid *Eidmanella* Roewer, 1935 at Doublecamp Creek (07_114); *Nesticus* is otherwise rarely found in sympatry with members of this genus.

Remarks. Monophyletic on mitochondrial and nuclear trees, with high gene and site CF values for the latter. Phylogenomic evidence strongly supports *Nesticus sheari* as sister to remaining members of the *reclusus* group (Figs 3, 4).

***Nesticus dellingeri* sp. nov.**

<https://zoobank.org/BDC1A285-4B27-4549-901B-FB2BBDE32B64>

Fig. 55A–G

Material examined. Type material: *Holotype*: USA – **North Carolina, Macon Co.** • holotype ♂; vicinity Whiteside Mountain, off Hwy 64, SW of Cashiers; 35.0793°N, -83.1415°W; 8 Aug. 1992; M. Hedin, I.-M. Tso leg.; MCH specimen #1047; ***Paratypes*:** – **North Carolina, Macon Co.** • 5♂, 9♀; vic Whiteside Mountain, off Hwy 64, SW of Cashiers; 35.0793°N, -83.1415°W; 8 Aug. 1992; M. Hedin, I.–M. Tso leg.; ***Non type material*:** – **North Carolina, Jackson Co.** • 2♀; along Chattooga River, NE side near mouth Scotsman Creek; 35.013°N, -83.1123°W; 16 Aug. 1991; B. Dellinger leg.; • ♂; along Chattooga River, NE side, 0.2 mi. W mouth Scotsman Creek; 35.0136°N, -83.1135°W; 17 Aug. 1991; B. Dellinger leg.; • ♂, 5♀; Whitewater River, below Upper Falls; 35.0337°N, -83.0141°W; 2 Sep. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_183; – **North Carolina, Macon Co.** • ♂, 5♀, 5 imm; Chattooga River, vic BullPen bridge crossing; 35.0172°N, -83.1262°W; 2 Sep. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_180; • 7♀; Chattooga River, vic BullPen bridge crossing, SE of Highlands; 35.0172°N, -83.1262°W; 8 Aug. 1992; M. Hedin leg.; – **South Carolina, Oconee Co.** • ♂, 3♀; along Whitewater River, just S NC/SC stateline; 35.0271°N, -83.0094°W; 13 Apr. 1992; B. Dellinger leg.

Diagnosis. Sister to other members of a phylogenomic subclade including *Nesticus binfordae*, *N. dykemanai* and *N. jonesi*, and morphologically most similar to these geographically disjunct taxa (in particular, sharing the spade-like basal regular apophysis; Fig. 52). Males of *N. dellingeri* differ from males of these other taxa in the shape of the distal regular apophysis (broad vs. skinny), the shape of the median apophysis, the shape of the paracymbial distal process, and in details of the paracymbial ventral process cusps. This species shares a very similar epigynal morphology with other members of the subclade (see descriptions below).

Description of ♂ holotype (MCH specimen #1047). Carapace dusky cream to orange, with faint dark pigment behind ocular area and along carapace margin. Legs

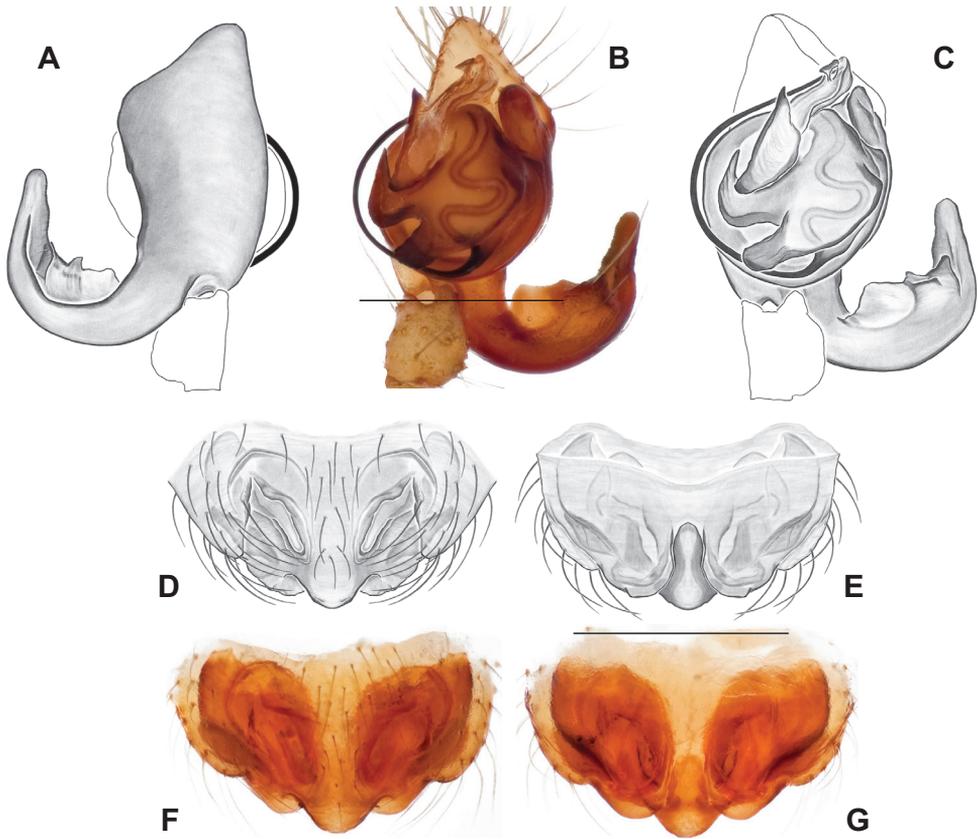


Figure 55. *Nesticus dellingeri* sp. nov. genitalia. North Carolina, Macon Co., vicinity Whiteside Mountain, MCH specimen #1047, ♂ palp, dorsal (A), ventral (B) C North Carolina, Jackson Co., Whitewater River, below Upper Falls, MCH 02_183, palp, ventral. North Carolina, Macon Co., vicinity Whiteside Mountain, MCH specimen #1057, epigynum, ventral (D), dorsal (E). North Carolina, Jackson Co., Whitewater River, below Upper Falls, MCH 02_183, ventral (F), dorsal (G). Scale bar: 0.5 mm.

approximately concolorous pale. Abdomen with paired, lateral darker markings on dirty gray background. All eyes approximately equal in size, AMEs barely visible. Eyes with light rings of dark pigment. CL 1.4, CW 1.1, abdomen length 2, total body length 3.4. Leg I total length 9.9 (2.75, 0.6, 3, 2.5, 1.05), leg formula 1423, leg I / CW ratio 9.0. Palp with broadly S-shaped tegular apophysis, distal part a short, curved blade with acute tip, basal fork of apophysis a short, square sclerotized spade (Fig. 55A–C). Median apophysis anvil-shaped, distal end a sharp tip. Conductor tip bent, surrounded by small funnel-shaped cuticular sheath. Paracymbium lacking a paradistal process, distal process finger-like with slight serration along paradistal edge. Lacking a dorsomedial process. Distal part of ventral paracymbial process thickened, with small cusps.

♂ **Variation.** Males from four non-type localities match topotypic males very closely (Fig. 55A–C).

Description of ♀ paratype (MCH specimen #1057). Carapace color as in male, slightly darker orange. Legs approximately concolorous pale. Abdomen with paired, lateral darker maculations on dirty gray background. All eyes approximately equal in size, AMEs miniscule but visible. Eyes with rings of dark pigment. CL 1.2, CW 1.1, abdomen length 1.65, total body length 2.85. Leg I total 7.3 (2.1, 0.55, 2.05, 1.7, 0.9), leg formula 1423, leg I / CW ratio 6.6. Epigynum generally wider than tall, median septum relatively wide with directly adjacent lateral pockets (Fig. 55D–G). Posterior end of septum with lateral bars oriented obliquely upwards, dark spermathecae lying beneath these bars and approximately following the upwards oblique path. Median septum narrowing past these bars and projecting inwards towards the abdomen. Viewed dorsally, dorsal internal pockets lying slightly above sclerotization of the lateral pockets.

♀ **Variation.** Females from different locations share a very similar epigynal morphology (Fig. 55D–G).

Distribution and natural history. Known only from a very small area in the upper Chattooga River and upper Whitewater River drainages (Fig. 53), along the south face of the Blue Ridge Escarpment in the North and South Carolina borderlands. Except for the type locality, most collections are relatively small in total animals collected, suggesting a natural rarity for this species. At the type locality (Whiteside Mountain) a total of 14 adults and 8 immature specimens was collected from a crevice cave at the base of rocky cliffs.

Nesticus bishopi has also been collected from nearby locations in the Chattooga River Gorge (locations near Scotsman Creek; Fig. 53), suggesting that these species might somewhere be syntopic in this area.

Etymology. This species is named to recognize and honor Bob Dellinger, a special naturalist from western North Carolina. Bob's knowledge of the flora and fauna of southern Appalachia is remarkable, and he personally collected or helped to collect (with first author MH) many *Nesticus* from this region.

Remarks. *Nesticus dellingeri* is geographically disjunct from phylogenetic relatives *N. binfordae*, *N. dykemanae* and *N. jonesi* (Fig. 53). The regions separating these taxa have been extensively sampled for *Nesticus* and are occupied by species from other species groups (*N. nasicus*, *tennesseensis* group members), or more distant *reclusus* group members.

Nesticus jonesi Gertsch, 1984

Fig. 56A–C

Nesticus jonesi Gertsch, 1984: 38, figs 153–155, 167–169.

Material examined. Type material: Holotype: USA – Alabama, Morgan Co. • ♂ holotype; Cave Spring Cave; 2 May 1959; W.B. Jones, Royer, Steeves, T.C. Barr leg; AMNH; **New collections from type locality:** – Morgan Co. • 4♂, 14♀; Wheeler NWR, Cave Spring Cave, E of Decatur; 14 Nov. 1992; M. Hedin, J. Hedin leg.

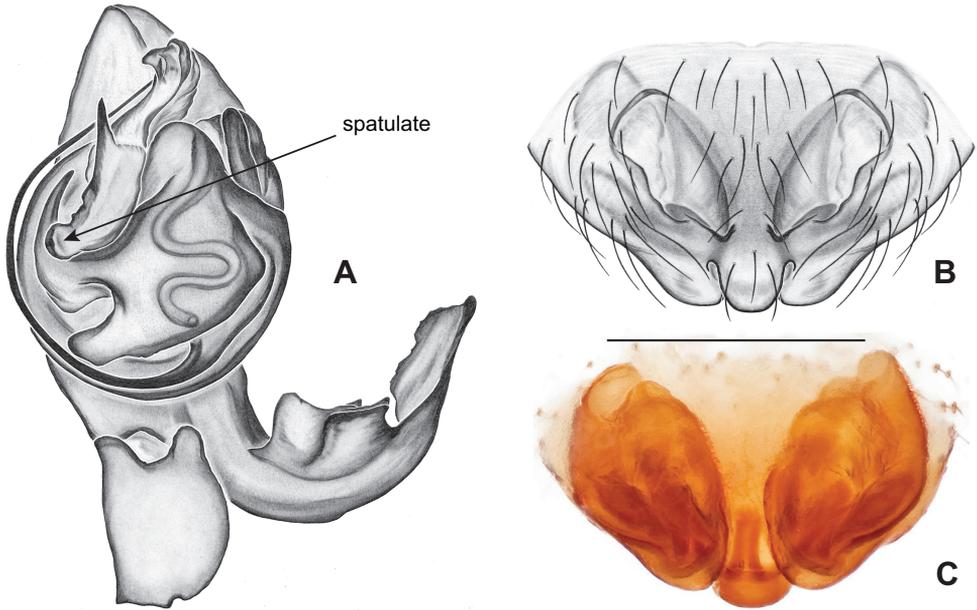


Figure 56. *Nesticus jonesi* genitalia **A** Alabama, Morgan Co., Cave Spring Cave, MCH specimen #1644, ♂ palp, ventral. Alabama, Morgan Co., Cave Spring Cave, MCH specimen #1656, epigynum, ventral (**B**), dorsal (**C**). Scale bar: 0.5 mm.

Diagnosis. Similar to regional congener *Nesticus barri*, this species is long-legged and nearly eyeless, but is otherwise morphologically and genetically allied with members of the *reclusus* group from montane western North Carolina. Very similar in male and female genital morphology to close phylogenomic kin *N. dellingeri* (Fig. 55A–G), *N. dykemanae* (Fig. 59A–C) and *N. binfordae* (57A–C), but geographically disjunct, troglomorphic, and larger in body size. Also differing from these taxa in the shape of the regular apophyses (both basal and distal), and the shape of the basal edge of the median apophysis (Fig. 56A).

Distribution and natural history. This species is known only from the type locality south of the Tennessee River in north-central Alabama (Fig. 53). Geographically far-flung from phylogenetic relatives, perhaps similar to the biogeographic situation observed in *Nesticus paynei* and/or *N. carteri*, both of which also include disjunct populations towards the southern end of the Tennessee River valley.

Collections in 1992 revealed a very large spider population in Cave Spring Cave, perhaps up to 1,000 individuals. This cave is home to a protected bat colony and located in a US National Wildlife Refuge. The extraordinary size of the *Nesticus jonesi* population is perhaps related to the high productivity associated with the large bat colony and/or the protected status of this cave.

Remarks. Part of a near phylogenomic trichotomy with *Nesticus dykemanae* and *N. binfordae* (Figs 3, 4), with sCF values near a lower limit.

***Nesticus binfordae* sp. nov.**

<https://zoobank.org/70333742-4E8D-46B5-BA00-455CEEAA4E32>

Figs 57A–C, 58A–F

Material examined. Type material: *Holotype*: USA – Tennessee, Sevier Co. • ♂ holotype; Great Smoky Mountains NP, Greenbrier Cove, Middle Prong Little Pigeon River, 1.3 mi. upstream Greenbrier Picnic Area; 35.7042°N, -83.3653°W; 20 Aug. 1992; M. Hedin leg; MCH specimen #1290; ***Paratypes*:** – Sevier Co. • ♀ paratype; data as for holotype; MCH specimen #1287; • 5♂, 14♀; data as for holotype; **Non type material:** – Coker Co. • ♀; Great Smoky Mountains NP, N side Indian Camp Creek on Maddron Bald Trail; 35.7378°N, -83.2777°W; 16 Apr. 1994; M. Hedin, B. Dellinger leg.; • 2♀; Great Smoky Mountains NP, trail from Low Gap to Mt. Cammerer; 35.754°N, -83.1658°W; 1 Aug. 2000; M. Hedin leg.; MCH 00_146.

Diagnosis. Most similar to close phylogenomic kin *Nesticus dykemanae* (Fig. 59A–C) and *N. jonesi* (Fig. 56A). Differing from the latter in having a sharp-tipped median apophysis, the shape of the basal tegular apophysis, and having a whip-like paradistal paracymbial process. Very similar to *N. dykemanae*, sharing the double-tipped median apophysis, but differing in the shape of the basal tegular apophysis and possessing a whip-like paradistal process. Sharing an almost identical epigynal morphology with *N. jonesi*.

Description of ♂ holotype (MCH specimen #1290). Carapace dirty light orange, dusky lines leading from fovea to eye group. Legs colored as carapace, without markings. Abdomen background color as carapace, six pairs of lateral faint darker markings. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.5, CW 1.3, abdomen length 1.7, total body length 3.2. Leg I total length 14.2 (3.9, 0.7, 4.35, 3.85, 1.4), leg formula 1423, leg I / CW ratio 10.9. Palp with broadly S-shaped tegular apophysis, distal part a short skinny curved

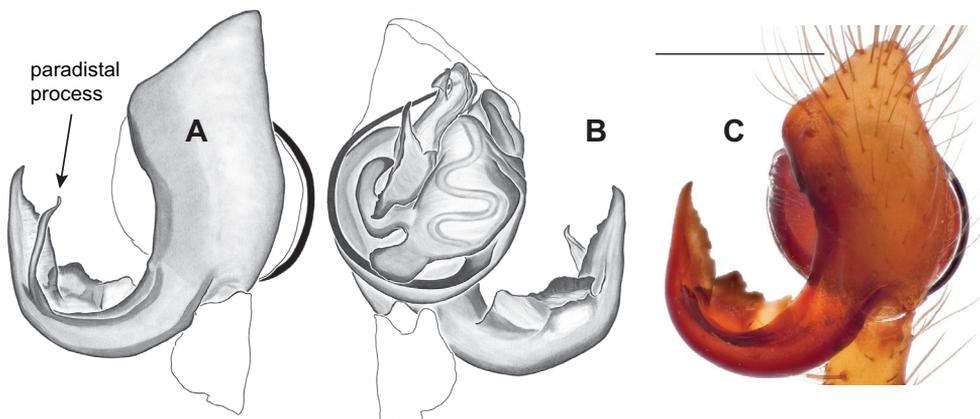


Figure 57. *Nesticus binfordae* sp. nov. ♂ palps. Tennessee, Sevier Co., Great Smoky Mountains NP, Middle Prong Little Pigeon River, MCH specimen #1290, dorsal (A), ventral (B). Middle Prong Little Pigeon River, MCH specimen #1289, dorsal (C). Scale bar: 0.5 mm.

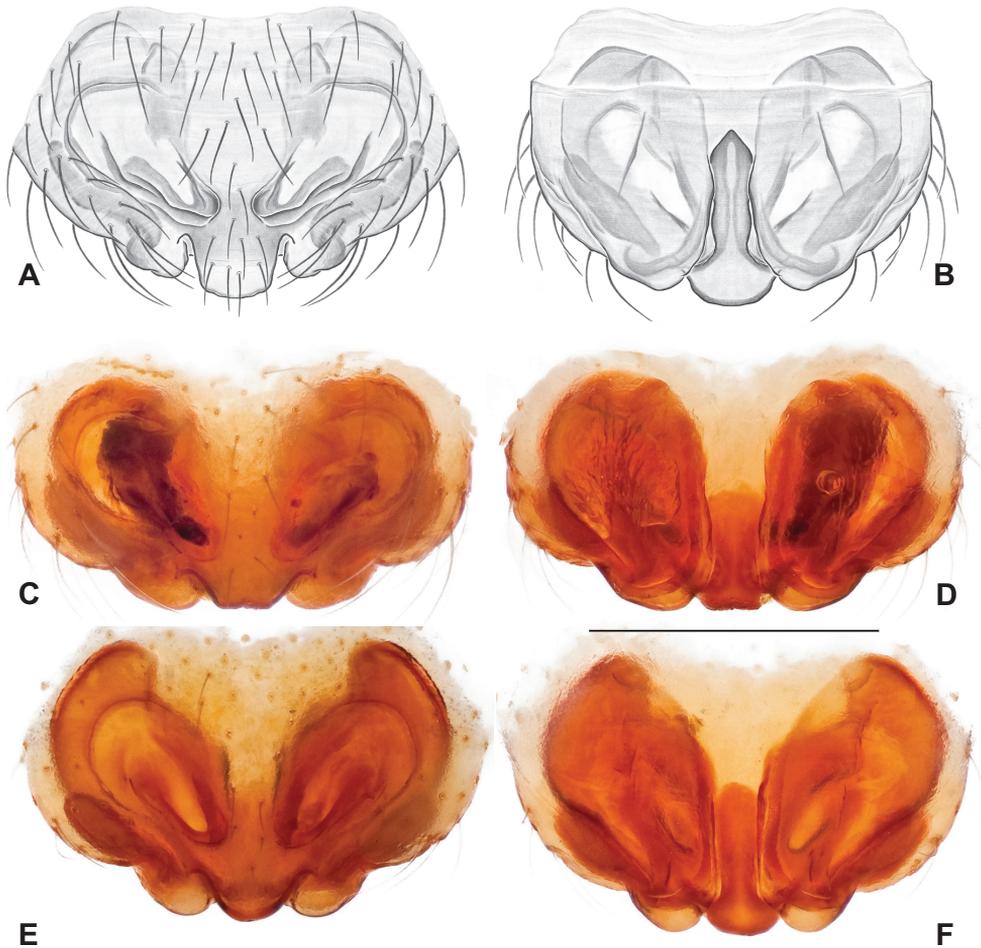


Figure 58. *Nesticus binfordae* sp. nov. epigynal variation. Tennessee, Sevier Co., Great Smoky Mountains NP, Middle Prong Little Pigeon River, MCH specimen #1283, ventral (**A**), dorsal (**B**). Tennessee, Cocke Co., Great Smoky Mountains NP, N side of Indian Camp Creek, MCH specimen #1981, ventral (**C**), dorsal (**D**). Tennessee, Cocke Co., Great Smoky Mountains NP, trail from Low Gap to Mt. Cammerer, MCH 00_146, ventral (**E**), dorsal (**F**). Scale bar: 0.5 mm.

blade with tapered tip, basal fork of apophysis a squat sclerotized spade with rounded edges (Fig. 57A–C). Median apophysis anvil-shaped, both ends with sharp tips. Conductor tip bent, surrounded by small funnel-shaped cuticular sheath. Paracymbium with a skinny whip-like paradistal process, and distal process finger-like with slight serration along paradistal edge. Lacking a dorsomedial process. Distal part of ventral paracymbial process thickened, forming a small blade without cusps (Fig. 57A–C).

♂ **Variation.** Males are only known from the type locality and all match the holotype male, except for MCH specimen #1289 which lacks the paracymbial paradistal process (Fig. 57C). Close examination of this specimen suggests that this process was broken off (process base is evident).

Description of ♀ paratype (MCH specimen #1287). Carapace subdued burnt orange, distinct darker markings leading from fovea forward, dusky ring to edge of carapace. Legs light orange, with faint dusky dark markings. Abdomen slightly paler than carapace, with fused distal lateral dark markings. Posterior eyes approximately equal in size, ALE slightly smaller than PLEs, AMEs ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.65, CW 1.4, abdomen length 2.35, total body length 4. Leg I total length 13.15 (3.75, 0.75, 4, 3.25, 1.4), leg formula 1423, leg I / CW ratio 9.4. Epigynum generally wider than tall, median septum relatively wide at top with adjacent heart-shaped lateral pockets (considering both sides). Septum narrows towards posterior end where lateral bars extend obliquely upwards, dark spermathecae lying beneath these bars and approximately following the upwards oblique path. Median septum extending past these bars and dipping inwards towards the abdomen. Viewed dorsally, dorsal internal plates lying slightly above sclerotization of the lateral pockets.

♀ **Variation.** Females from different locations share a very similar epigynal morphology (Fig. 58A–F).

Distribution and natural history. Known only from three parallel north-flowing drainages in the Great Smoky Mountains National Park, including the Middle Prong of the Little Pigeon River, and more easterly draining Indian Camp and Cosby Creeks.

At the type locality in 1992 spiders were “very abundant in rock crevices, low to the ground, close to the river”.

Along the Maddron Bald and Mt. Cammerer trails we collected both *Nesticus cherokeensis* and *N. binfordae*, indicating that these species are syntopic or nearly so at these locations. At both locations multiple collections were taken along an elevational transect and unfortunately lumped into a single collecting event, so it is not possible to discern if different species were collected at the exact same location (truly syntopic) or were closely parapatric along these elevational transects.

Etymology. Named to honor Dr. Greta Binford. Friend, arachnologist, and Past President of the American Arachnological Society (AAS), here recognized for her inspirational spider research and her leadership in making the AAS a more diverse and welcoming society. We suspect that Dr. Binford would also greatly appreciate the beauty of the habitats that this spider calls home.

Remarks. Part of a near phylogenomic trichotomy with *Nesticus dykemanae* and *N. jonesi* (Figs 3, 4), with sCF values near a lower limit.

This species was called “N novsp2” (from site 48) in Hedin (1997b) and lumped with *Nesticus dykemanae* despite having distinctive (non-sister) ND1/16S sequences.

Nesticus dykemanae sp. nov.

<https://zoobank.org/6A313CD5-EA60-4F68-8CBF-37887618A8B1>

Fig. 59A–I

Material examined. Type material: Holotype: USA – Tennessee, Sevier Co. • ♂ holotype; Great Smoky Mountains NP, Hwy 441 near Chimney Tops trailhead; 35.6364°N, -83.4709°W; 31 Jul. 2000; M. Hedin, J. Cokendolpher leg.; MCH

00_143 (SDSU_TAC000673); **Type material:** *Paratypes*: – **Sevier Co.** • ♂, 4♀; Great Smoky Mountains NP, Hwy 441 near Chimney Tops trailhead; 35.6364°N, -83.4709°W; 31 Jul. 2000; M. Hedin, J. Cokendolpher leg.; MCH 00_143; **Non type material:** – **Sevier Co.** • ♂, 8♀; Great Smoky Mountains NP, Hwy 441 half way between tunnel and Chimney picnic area; 35.6414°N, -83.4819°W; 16 Apr. 1994; M. Hedin, F. Coyle, B. Dellinger leg.; • 5♀; Great Smoky Mountains NP, Hwy 441 N Chimneys campground; 35.6406°N, -83.4949°W; 27 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_098.

Diagnosis. This species is included in a phylogenomic subclade with *Nesticus jonesi* and *N. binfordae*. Males share the double-tipped median apophysis with the latter species but differ in the shape of both distal and proximal tegular apophyses, lack a whip-like paracymbial paradistal process, and have a less modified ventral paracymbial process (Fig. 59A–C). The epigynum is most distinctive in the larger phylogenomic subclade (including *N. dellingeri*), with lateral bars that extend from the median septum obliquely upwards at approximately 45-degree angles, interrupting the lateral pockets (Fig. 59D–I). Also, dark spermathecae lie below the septum bars, extending obliquely outwards.

Description of ♂ holotype (SDSU_TAC000673). Carapace dusky cream to orange, with conspicuous faint dark pigment behind ocular area, and along carapace margin bleeding inwards. Legs approximately concolorous pale. Abdomen with strong paired, lateral darker markings on a dirty orange/gray background. All eyes approximately equal in size, except for AMEs, ~ 1/4 width of ALEs. Eyes with rings of dark pigment. CL 1.5, CW 1.3, abdomen length 1.5, total body length 3. Leg I total length 14.25 (3.95, 0.75, 4.35, 3.8, 1.4), leg formula 1423, leg I / CW ratio 11.0. Palp with broadly S-shaped tegular apophysis, distal part a particularly skinny curved blade with sharp tip, basal fork of apophysis a squat sclerotized spade with saw-like leading edge (Fig. 59A–C). Median apophysis anvil-shaped, both ends with sharp tips, apical end more tongue-shaped. Conductor tip bent, surrounded by small funnel-shaped cuticular sheath. Paracymbium lacking paradistal process, distal process finger-like with slight serration along paradistal edge. Lacking a dorsomedial process. Distal part of ventral paracymbial process only slightly thickened, lobe-like (Fig. 59A–C).

♂ **Variation.** Other than the holotype male only two other males are known, and these closely match the holotype. MCH specimen #1977 (Fig. 59C) appears slightly unusual because this male had recently molted.

Description of ♀ paratype (SDSU_TAC000674). Carapace color as in male, dark pigment not as strong. Legs approximately concolorous pale, very faint pigmentation. Abdomen with strong paired, lateral darker markings on a slightly lighter background. Eye development as in male. Eyes with rings of dark pigment. CL 1.45, CW 1.25, abdomen length 1.95, total body length 3.4. Leg I total length 11.3 (3.25, 0.6, 3.35, 2.9, 1.2), leg formula 1423, leg I / CW ratio 9.0. Epigynum generally wider than tall, median septum with adjacent heart-shaped lateral pockets (considering both sides). Septum towards posterior end with lateral bars that extend obliquely upwards

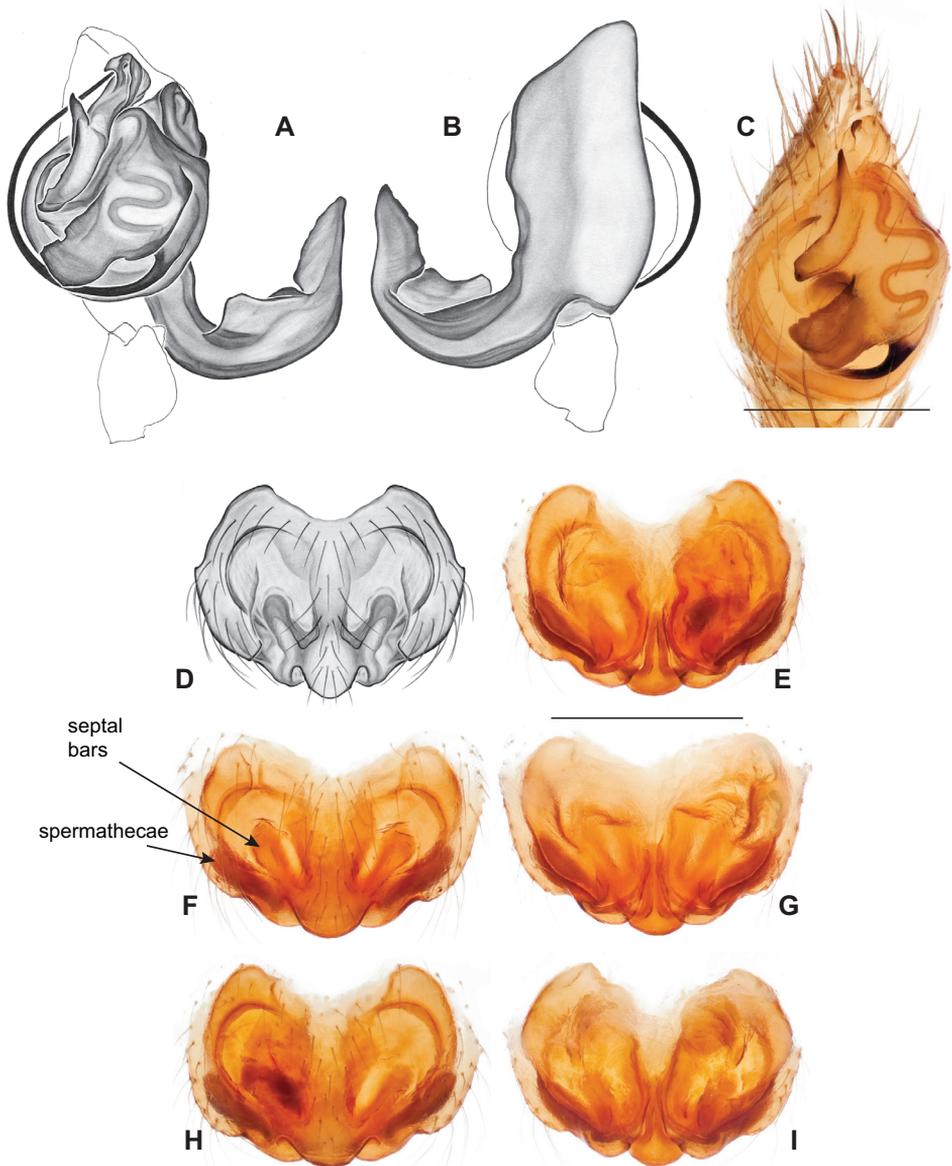


Figure 59. *Nesticus dykemanai* sp. nov. genitalia. ♂ palps – Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441 near Chimney Tops trailhead, MCH 00_143 (SDSU_TAC000673), ♂ palp ventral (A), dorsal (B) C Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441, between tunnel and Chimney picnic area, MCH specimen #1977, ♂ palp ventral. Scale bar: 0.5 mm. epigynal variation – Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441 near Chimney Tops trailhead, MCH 00_143 (SDSU_TAC000674), epigynum ventral (D), dorsal (E). Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441, between tunnel and Chimney picnic area, MCH specimen #1973, epigynum ventral (F), dorsal (G). Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441, N of Chimneys campground, epigynum ventral (H), dorsal (I). Scale bar: 0.5 mm.

at approximately 45-degree angles, interrupting lateral pockets. Dark spermathecae lying below septum bars, extending obliquely outwards. Median septum broadening slightly past these bars and dipping inwards towards the abdomen. Viewed dorsally, dorsal internal plates lying distinctly above the sclerotized ring of the lateral pockets.

♀ **Variation.** Females from adjacent locations share a very similar epigynal morphology (Fig. 59D–I).

Distribution and natural history. Known from three closely adjacent locations from near the headwaters of the West Prong of the Little Pigeon River, Great Smoky Mountains National Park, on the southwest slopes of Mt. Leconte (Fig. 53). All nearby surrounding collections have resulted in the collection of *Nesticus reclusus* (Fig. 53), suggesting that this microendemism is real (rather than a collecting artifact). Gertsch (1984) also includes records of *N. reclusus* from the “top of Mt. Leconte”.

1994 collections from near the Chimney Picnic Area resulting in collections of a male and eight females were from a “large talus breakdown in a south-facing cove” in rich hardwood forest.

Etymology. Named to honor Wilma Dykeman (1920–2006), a writer, speaker, teacher, historian, and environmentalist who spent most of her life in western North Carolina and eastern Tennessee. Mrs. Dykeman was devoted to social justice and environmental integrity, discussing Appalachian water pollution in her classic 1955 book ‘The French Broad’, and sharing a social justice award in 1957 for her co-authored book ‘Neither Black Nor White’.

Remarks. Part of a near phylogenomic trichotomy with *Nesticus binfordae* and *N. jonesi* (Figs 3, 4), with sCF values near a lower limit.

This species was called “*N. novsp2*” (from site 49) in Hedin (1997b) and lumped with *N. binfordae* despite having distinctive (non-sister) ND1/16S sequences.

Nesticus bishopi Gertsch, 1984

Figs 60A–D, 61A–H

Nesticus bishopi Gertsch, 1984: 33, figs 147–149.

Material examined. Type material: Holotype: USA – North Carolina, Macon Co. • ♀ holotype; Highlands; 6 Apr. 1929; S.C. Bishop leg.; AMNH; **New collections from near type locality: – North Carolina, Macon Co.** • ♂, 8♀; below Glenn Falls, SW of Highlands; 35.0312°N, -83.2383°W; 2 Aug. 1992; M. Hedin leg.; • ♂, ♀; below Glenn Falls, SW of Highlands; 35.0312°N, -83.2383°W; 31 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_175; **Non type material: – Georgia, Rabun Co.** • 2♀; Holcomb Branch of Holcomb Creek, off Hale Ridge Road, NE of Rabun Bald; 34.9831°N, -83.2661°W; 14 Apr. 1992; T. McGarity leg.; • 2♂, 3♀; Hwy 246/106 along Mud Creek, NE of Dillard; 34.9924°N, -83.3385°W; 19 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_126; • 2♂, ♀; Hwy 76 at Chattooga River crossing, confluence with Pole Creek; 34.8172°N, -83.3061°W; 2 Sep. 2002;

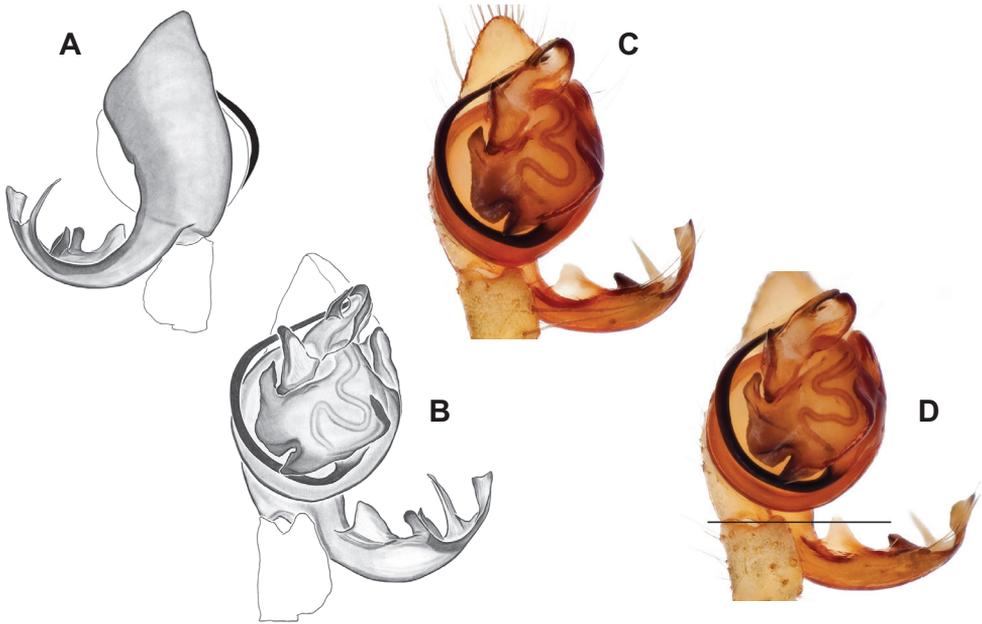


Figure 60. *Nesticus bishopi* ♂ palps. North Carolina, Macon Co., below Glenn Falls, MCH specimen #1078, dorsal (A), ventral (B) C Georgia, Rabun Co., Chattooga River at confluence with Pole Creek, MCH 02_181, ventral D North Carolina, Macon Co., along Black Creek, MCH 02_141, ventral. Scale bar: 0.5 mm.

M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_181; – **North Carolina, Jackson Co.** • ♀; along Chattooga River, NE side between Scotsman and Glade Creek; 35.0123°N, -83.1164°W; 13 Jul. 1992; B. Dellinger leg.; – **North Carolina, Macon Co.** • ♂, 8♀; 4.3 mile S Standing Indian campground, along Nantahala River; 35.0347°N, -83.5057°W; 10 Aug. 1992; M. Hedin leg.; • ♀, 5 imm; 4.3 mile S Standing Indian campground, along Nantahala River; 35.0347°N, -83.5057°W; 20 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_140; • 2♂, 5♀, 2 imm; along Black Creek, NE side Chunky Gal Mountain; 35.092°N, -83.5663°W; 20 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_141; • 2♀; Coweeta Hydrological Lab, along Shope Fork of Coweeta Creek, FR 751; 35.0611°N, -83.4447°W; 19 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_125; • ♂; Coweeta Hydrological Lab, along Shope Fork of Coweeta Creek, W of Otto; 35.0601°N, -83.4547°W; 23 Oct. 2012; M. Hedin, J. Bond leg.; MCH 12_043; • 2♀; FR 710, 2 mi. N Deep Gap; 35.0425°N, -83.555°W; 20 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_142; • 9♀; near Dry Falls, Cullasaja River, off Hwy 64 NW of Highlands; 35.069°N, -83.239°W; 11 Aug. 1992; M. Hedin leg.

Diagnosis. Compared to other members of the challenging *reclusus* subgroup, *Nesticus bishopi* is similar in detail in all aspects of male and female morphology to

N. stupkai (compare Figs 60A–D, 61A–H, 62A–H). Shared male features include a distal tegular apophysis shoe-shaped with a beak-like tip, base of distal part with a sclerotized and blade-like shoulder, proximal fork of tegular apophysis arrowhead-like, median apophysis short and triangular, paracymbium with translucent bladelike paradistal process, distal process with twisted tip, and toothlike distomedial process directly adjacent to small flange-like ventromedial process. Although we cannot find morphological characters that distinguish *N. bishopi* from *N. stupkai*, we retain both as distinct based on diagnostic DNA characters and an allopatric distribution (see further arguments in the Discussion). Both species are easily distinguished from closely-related *N. reclusus* by the shape of the male median and tegular apophyses (Figs 63A–F, 64A–F).

Description of ♂ from near type locality (MCH specimen #1078). Carapace dusky cream to orange, conspicuous faint dark pigment behind ocular area. Legs pale yellow to cream. Abdomen dirty pale cream, faint paired lateral pigmentation blotches. Eyes approximately equal in size, except for AMEs, ~ 1/3 width of ALEs. Eyes with rings of dark pigment. CL 1.3, CW 1.1, abdomen length 1.75, total body length 3.05. Leg I total length 9.75 (2.65, 0.55, 2.95, 2.5, 1.1), leg formula 1423, leg I / CW ratio 8.9. Palp with broadly S-shaped tegular apophysis, distal part shoe-shaped with a beak-like tip, base of distal part with a sclerotized and blade-like shoulder. Basal fork of tegular apophysis like a sclerotized broad-based arrowhead (Fig. 60A–D). Median apophysis short and triangular. Conductor tip bent, surrounded by small funnel-shaped cuticular sheath. Paracymbium with translucent bladelike paradistal process, distal process with twisted, tubular tip; well-sclerotized, toothlike distomedial process directly adjacent to small flange-like ventromedial process (Fig. 60A–D).

Variation. Males and females from both sides of the Little Tennessee River barrier (see below) share very similar genitalic morphologies (Figs 60A–D, 61A–H).

Distribution and natural history. From montane habitats in southern North Carolina and northern Georgia. Populations are found both east (Coweeta Mountains, including the type locality) and west (Nantahala Mountains) of the Little Tennessee River, a known dispersal barrier in other arachnid taxa (e.g., Thomas and Hedin 2008; Keith and Hedin 2012; Hedin and McCormack 2017). Western populations (Black Creek Road, Standing Indian, Coweeta, Deep Gap Road) indeed form a subclade on mitochondrial trees (Fig. 6).

As an example of natural history, 1992 collections near Standing Indian Campground were made in a northwest-facing rocky ravine, where many specimens were collected “in dark ravine, wet, deep litter, rocks, *Rhododendron*”.

Nesticus bishopi has been collected from locations very near *N. dellingeri* in the Chattooga River gorge (locations near Scotsman Creek; Fig. 53), suggesting that these species might somewhere be syntopic in this area.

Remarks. The species is obviously morphologically very similar to a disjunct *Nesticus stupkai* and is arguably conspecific from a morphological perspective. We have retained *N. bishopi* as distinct at the species level because this taxon is monophyletic on both UCE and mitochondrial trees (Figs 3, 4, 6), and is geographically disjunct from sister species *N. stupkai*. One complication is that *N. stupkai* is paraphyletic with respect to *N. bishopi* on nuclear trees (Figs 3, 4), as further discussed below.

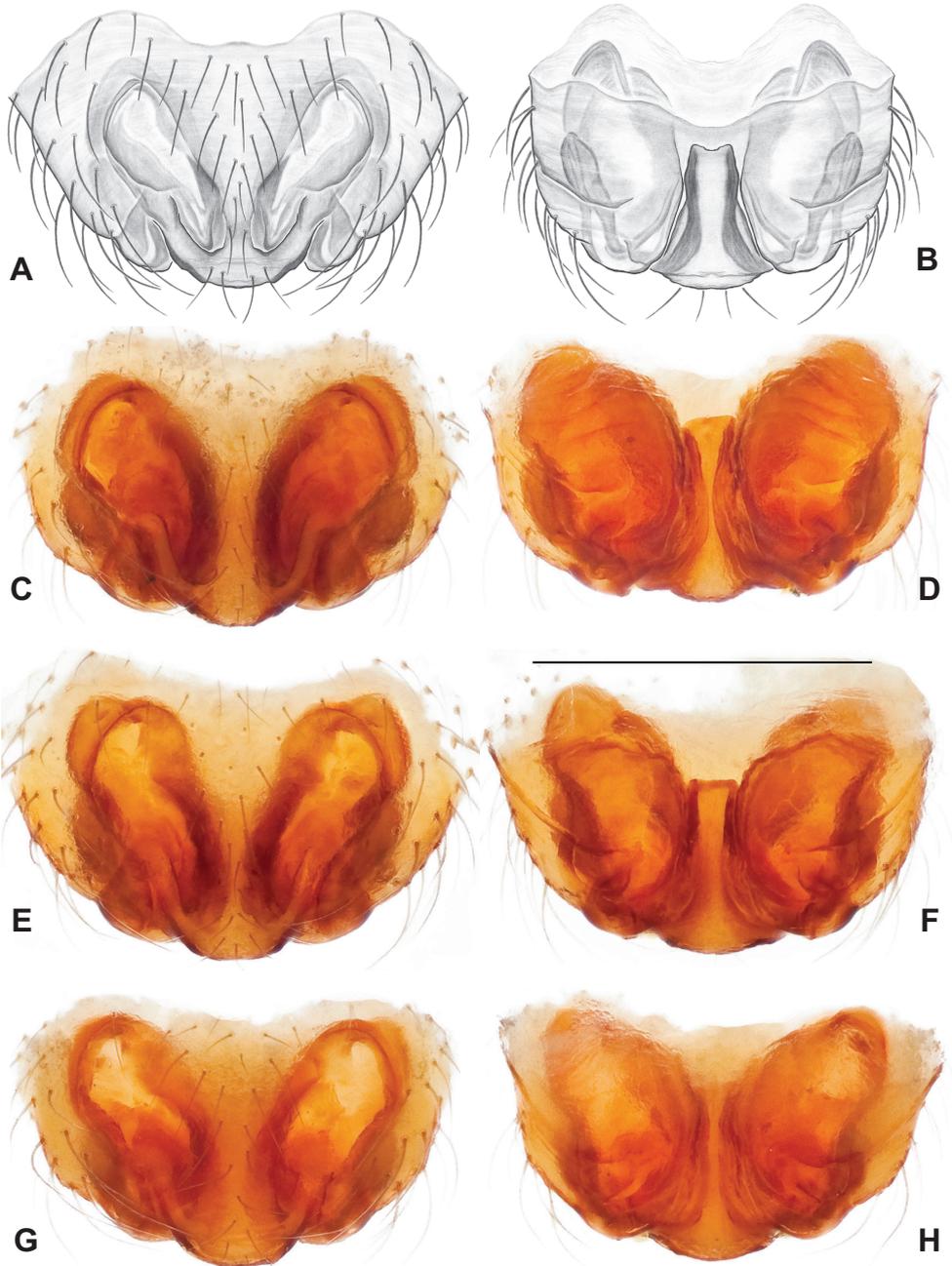


Figure 61. *Nesticus bishopi* epigynal variation. North Carolina, Macon Co., below Glenn Falls, MCH specimen #1071, ventral (**A**), dorsal (**B**). Georgia, Rabun Co., Chattooga River at confluence with Pole Creek, MCH 02_181, ventral (**C**), dorsal (**D**). North Carolina, Macon Co., along Black Creek, MCH 02_141, ventral (**E**), dorsal (**F**). North Carolina, Jackson Co., along Chattooga River, between Scotsman and Glade Creek, MCH specimen #2016, ventral (**G**), dorsal (**H**). Scale bar: 0.5 mm.

We have made extensive collections of other *Nesticus* taxa in the region that separates *N. bishopi* from *N. stupkai*, finding only other *Nesticus* species (e.g., *N. silvanus*, *N. cherokeensis*, etc.; Fig. 53). As such, we view the probability of on-going gene flow as low.

Nesticus stupkai Gertsch, 1984

Fig. 62A–I

Nesticus stupkai Gertsch, 1984: 31, figs 71–74, 106–108; Reeves 2000: 338.

Material examined. Type material: *Holotype*: USA – Tennessee, **Blount Co.** • ♂ holotype; White Oak Sinks, Great Smoky Mountains National Park; 21 Jul. 1937; A. Stupka leg; AMNH; **New collections from near type locality:** – **Blount Co.** • ♀; Great Smoky Mountains NP, White Oak Sinks, Blowhole; 21 Aug. 1992; M. Hedin leg. **Non type material:** – **Blount Co.** • 6♂, 9♀; Blowing Cave, NE Townsend, off Hwy 321; 22 Sep. 1992; M. Hedin, S. O’Kane leg.; • ♀; Great Smoky Mountains NP, Little River at Mile 40 of Hwy 73; 35.6688°N, -83.6827°W; 22 Aug. 1992; M. Hedin leg.; – **Loudon Co.** • ♂, 4♀, 16 imm; Benjos Cave, TLN11; 30 Aug. 2014; M.L. Niemiller, C.D.R. Stephen, E.T. Carter leg.; MLN 14–044.17.

Diagnosis. See Diagnosis of *Nesticus bishopi* for details on shared male morphology. Females are likewise similar to *N. bishopi*, with a narrowing median septum with posterior bars that form an anchor shape, directed upwards and outwards, spermathecae lying lateral to these bars at approximately the same angle (compare Fig. 61A–H. to Fig. 62D–I). Without knowledge of geographic origin, we cannot distinguish epigynal morphologies of these two species.

Females of *Nesticus stupkai* and *N. bishopi* can be distinguished from the closely related *N. reclusus* by the outwards oriented dorsal epigynal plates in the former (Figs 61A–H, 62D–I), vs. the inwards oriented dorsal epigynal plates in the latter (Fig. 65A–F).

Variation. Males and females from different populations share very similar genital morphologies (Fig. 62A–C).

Distribution and natural history. With a distribution similar to *Nesticus barrowsi*, from cave entrances in karst windows along the northwestern edge of Great Smoky Mountains National Park (Fig. 53), and nearby surface (boulderfield) habitats. We include a new important western record from Benjos Cave, southwest of Knoxville in the Tennessee River Valley.

As an example of natural history, specimens from Blowing Cave were collected from a cave entrance, while those from Little River were collected from beneath rock-piles directly adjacent to a stream.

Reeves (2000) reported *Nesticus stupkai* in sympatry with *N. barrowsi* at two cave locations in Great Smoky Mountains National Park, and we collected this taxon pair in near sympatry in the White Oak Sinks, with *N. barrowsi* found in the dark zone of caves, and *N. stupkai* found closer to cave entrances (twilight zone). Reeves (2000) also recorded *N. stupkai* from Myhr Cave. We have confirmed male *N. reclusus* from this

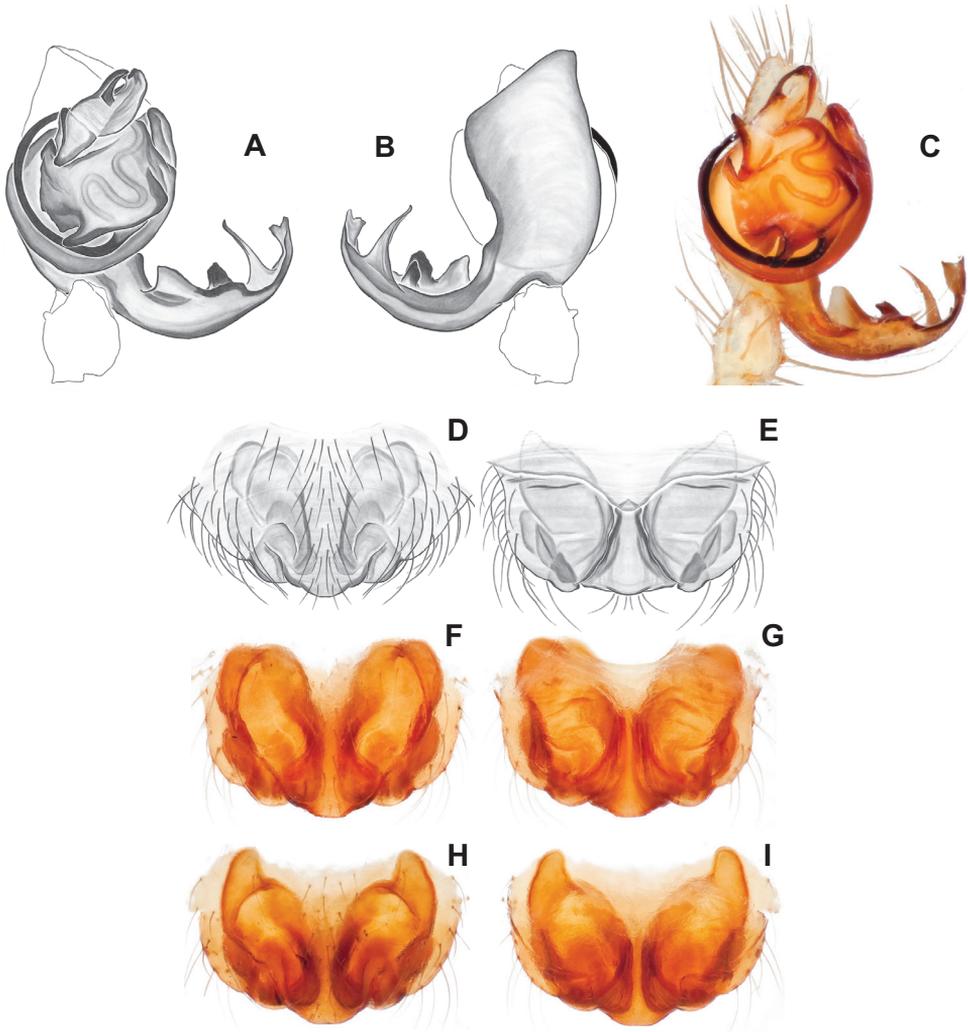


Figure 62. *Nesticus stupkai* genitalia. Tennessee, Blount Co., Blowing Cave, NE of Townsend, MCH specimen #1555, ♂ palp, ventral (A), dorsal (B) C Tennessee, Blount Co., Great Smoky Mountains NP, White Oak Sinks, holotype ♂ palp, ventral. Tennessee, Blount Co., Blowing Cave, MCH specimen #1567, epigynum, ventral (D), dorsal (E). Tennessee, Blount Co., Great Smoky Mountains NP, Little River at Mile 40 of Hwy 73, MCH specimen #1305, epigynum, ventral (F), dorsal (G). Tennessee, Blount Co., Great Smoky Mountains NP, White Oak Sinks, MCH specimen #1304, epigynum, ventral (H), dorsal (I). Scale bar: 0.5 mm.

location (Fig. 63A). This is either a case of sympatry or an original misidentification, as females of these species can be difficult to distinguish.

Remarks. *Nesticus bishopi* plus *N. stupkai* together form a strongly supported nuclear clade (Figs 3, 4). Within this clade however *N. stupkai* is not monophyletic on UCE trees, with the White Oak Sinks (type) population strongly supported as more

closely related to a geographically distant *N. bishopi* clade than to the geographically adjacent Little River plus Blowing Cave *N. stupkai* clade (Figs 3, 4).

One possibility is that the latter clade (Little River, Blowing Cave) is not *Nesticus stupkai*, but a separate lineage. We have closely compared males and females from the White Oak Sinks (type) population to males and females from Blowing Cave and detect no morphological differences (Fig. 62A–C). Despite paraphyly, and as an expectation to the species criteria used in this revision, we retain *N. stupkai* as a distinct species.

***Nesticus reclusus* Gertsch, 1984**

Figs 63A–G, 64A–L, 65A–F, 66A–K

Nesticus reclusus Gertsch, 1984: 29, figs 75–78, 109–111.

Nesticus cooperi Gertsch, 1984: 30, figs 132–134, 144–146. syn. nov.

Material examined. NORTHEASTERN LOCATIONS: **Type material:** *Holotype:* USA – **North Carolina, Swain Co.** • ♂ holotype; Andrew's Bald, Great Smoky Mountains National Park; no date given; W.M. Barrows leg.; AMNH; **Non type material:** – **North Carolina, Swain Co.** • 8♂, 4♀; Great Smoky Mountains NP, Clingman's Dome, vicinity Forney Ridge parking area; 35.5558°N, -83.496°W; 20 Aug. 1992; M. Hedin leg.; • 2♀; Great Smoky Mountains NP, Deep Creek, 0.25 mi. above Deep Creek CG, N Bryson City; 35.4644°N, -83.4344°W; 14 Aug. 1992; M. Hedin leg.; • ♀, 1 imm; Great Smoky Mountains NP, Hwy 441 E Thomas Ridge, 6.4 mi. N Smoke-mont CG turnoff; 35.6°N, -83.4091°W; 26 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_091; • 9♀; Great Smoky Mountains NP, Noland Creek at Laurel Branch, off Fontana Road, W of Bryson City; 35.4582°N, -83.5293°W; 26 Aug. 2005; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 05_093; • ♂; Hwy 129, NE of Cheoah Dam along Cheoah Reservoir; 35.4554°N, -83.9254°W; 17 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_116; – **Tennessee, Sevier Co.** • ♂; Great Smoky Mountains NP, Elkmont Area; 35.6536°N, -83.5802°W; 31 Jul. 2000; M. Hedin, J. Cokendolpher leg.; MCH 00_144; • 2♂, 5♀; Great Smoky Mountains NP, Hwy 441 0.8 mi. N Newfound Gap; 35.62°N, -83.4197°W; 20 Aug. 1992; M. Hedin leg.; • ♂; Great Smoky Mountains NP, Lower Baskins Creek; 35.6957°N, -83.4823°W; B. Dellinger leg.; • ♂; Great Smoky Mountains NP, N side of Mt Buckley, W of Clingman's Dome; 35.5626°N, -83.5058°W; 21 Oct. 2012; M. Hedin, J. Bond, F. Coyle, S. Cameron leg.; MCH 12_039; • 3♀; Wear Cove, Myhr Cave; 2 Aug. 2000; M. Hedin, J. Cokendolpher, W. Reeves leg.; MCH 00_148; • 2♂, ♀; Wear Cove, Myhr Cave; 29 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_182.

SOUTHWESTERN LOCATIONS: USA – **North Carolina:** *Swain County*, Lost Nantahala Cave, near Nantahala, 17 May. 1979, coll. P.T. Hertl, S.P. Plantani, C.O. Holler (♂ holotype of *Nesticus cooperi*). – **Georgia, Gilmer Co.** • ♂, 2♀; Rock Creek Road, N of Rich Mountain Wilderness, 3 mi. E Cherry Log at Hwy 76; 34.7811°N,

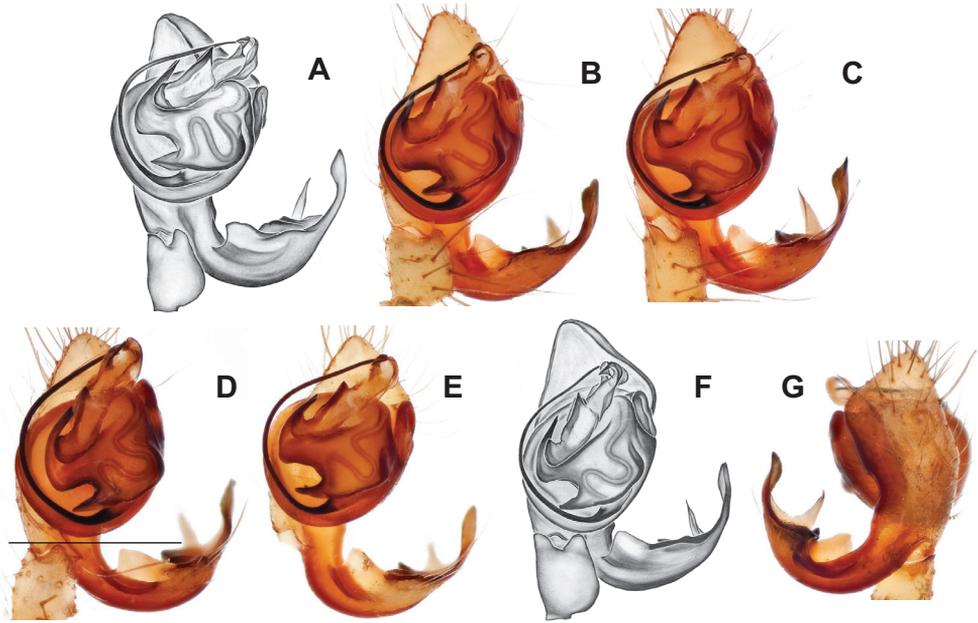


Figure 63. northeastern *Nesticus reclusus* ♂ palps, ventral view (*except for **G**) **A** Tennessee, Sevier Co., Wear Cove, Myhr Cave, MCH 01_182 (*right palp, inverted in Photoshop) **B** Tennessee, Sevier Co., Great Smoky Mountains NP, Lower Baskins Creek **C** Tennessee, Sevier Co., Great Smoky Mountains NP, Hwy 441, N of Newfound Gap **D** Tennessee, Sevier Co., Great Smoky Mountains NP, Elkmont Area, MCH 00_144 **E** North Carolina, Swain Co., Great Smoky Mountains NP, south of Clingman's Dome, near Forney Ridge parking area **F** North Carolina, Swain Co., Great Smoky Mountains National Park, Andrew's Bald (male holotype, AMNH) **G** North Carolina, Swain Co., Cheoah Dam along Cheoah Reservoir (MCH 07_116), dorsal view. Scale bar: 0.5 mm.

-84.3339°W; 15 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_102; – **Georgia, Towns Co.** • 12♀, 3 imm; 180 spur to Brasstown Bald; 34.8593°N, -83.8008°W; 21 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_147; • 2♀; 180 spur to Brasstown Bald; 34.8593°N, -83.8008°W; 15 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_104; – **North Carolina, Cherokee Co.** • 3♂, 4♀; Beaver Creek Road, along Beaver Creek, N of Andrews; 35.2152°N, -83.8327°W; 18 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_057; • ♂, 3♀; Junaluska Road along Junaluska Creek, SE of Andrews; 35.176°N, -83.768°W; 18 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_059; • ♂; Junaluska Road along Junaluska Creek, SE of Andrews; 35.176°N, -83.768°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_122; • 2♂, 4♀, 2 imm; Tatham Gap Road, S of Tatham Gap, N of Andrews; 35.2495°N, -83.8154°W; 18 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_058; • ♂, 2♀; Watkins Creek Road, off Hwy 19, SW of Topton; 35.2312°N, -83.7204°W; 19 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_066; – **North Carolina, Clay Co.** • ♂, 8♀, 3 imm; along Fires Creek,

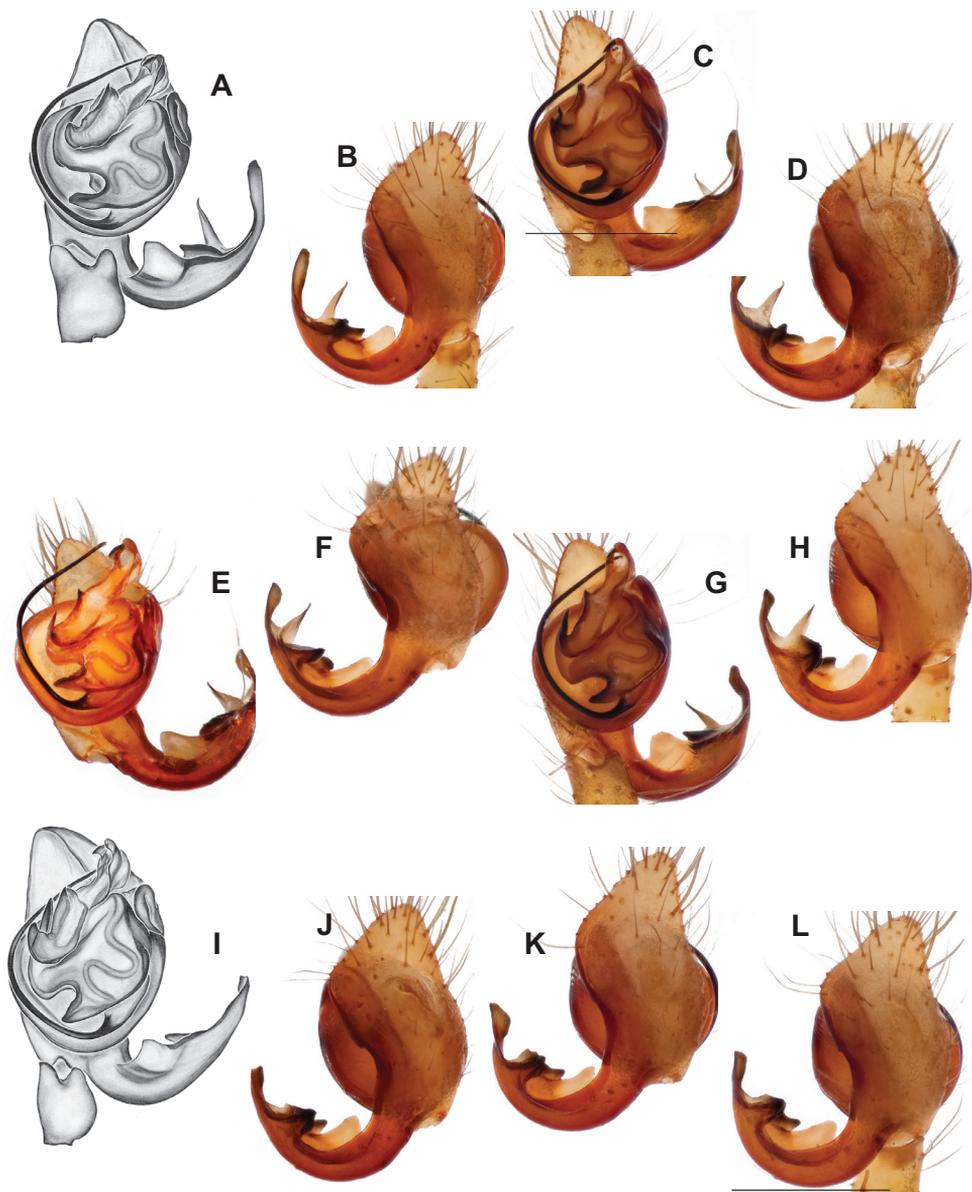


Figure 64. southwestern *Nesticus reclusus* ♂ palps. North Carolina, Graham Co., Appalachian Trail, S of Stecoah Gap, MCH 02_165, ventral (A), dorsal (B) North Carolina, Graham Co., ENE of Fontana Village, MCH 07_120, ventral (C), dorsal (D). Georgia, Gilmer Co., Rock Creek Road, N of Rich Mountain Wilderness, MCH 07_102, ventral (E), dorsal (F). North Carolina, Cherokee Co., Beaver Creek Road, MCH 04_057, ventral (G). North Carolina, Cherokee Co., S of Tatham Gap, MCH 04_058, dorsal (H). North Carolina, Clay Co., Tusquitee Mountains, Long Branch of Fires Creek, MCH 02_144, ventral (I), dorsal (J). North Carolina, Swain Co., Nantahala River Gorge, Nantahala River Gorge, across from Talc Mountain quarry, dorsal (K). North Carolina, Macon Co., S Wayah Bald, MCH 02_169, dorsal (L). Scale bar: 0.5 mm.

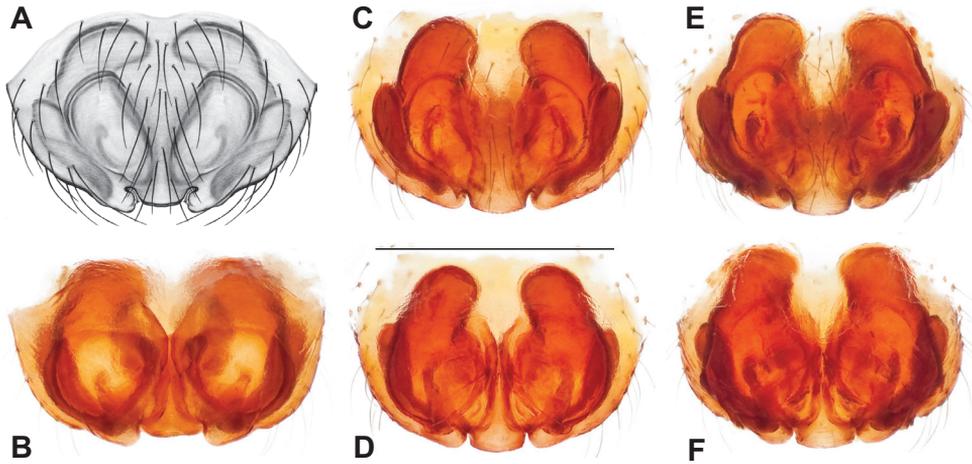


Figure 65. northeastern *Nesticus reclusus* epigynal variation. North Carolina, Swain Co., Great Smoky Mountains NP, south of Clingman's Dome, vicinity Forney Ridge parking area, MCH specimen #1973, ventral (**A**), dorsal (**B**). North Carolina, Swain Co., Great Smoky Mountains NP, N of Smokemont Campground turnoff, MCH specimen #N1019, ventral (**C**), dorsal (**D**). North Carolina, Swain Co., Great Smoky Mountains NP, Noland Creek at Laurel Branch, MCH specimen #N1051, ventral (**E**), dorsal (**F**). Scale bar: 0.5 mm.

NE Omphus Ridge; 35.1099°N, -83.8267°W; 21 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_145; • ♂, 4♀; Tusquitee Mountains, Fires Creek, Long Branch, just up from Short Branch; 35.1467°N, -83.7618°W; 21 Aug. 2002; M. Hedin, F. Coyle, M. Lowder, P. Paquin leg.; MCH 02_144; – **North Carolina, Graham Co.** • 3♂, 13♀, 11 imm; 0.25 mi. S Stecoah Gap on Appalachian Trail, off Hwy 143, Cheoah Mountains, NE of Cheoah; 35.353°N, -83.7187°W; 28 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_165; • 3♂, 8♀, 7 imm; along Panther Creek at Cook Branch confluence, N of Grassy Gap; 35.3677°N, -83.6272°W; 28 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_167; • ♀; Franks Creek, along Franks Creek Road, E of Sweetgum; 35.3158°N, -83.7361°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_121; • ♂, 2♀, 3 imm; Hwy 28, 0.6 mi. E entrance to Cable Cove campground; 35.4234°N, -83.7514°W; 28 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_166; • 3♂, 4♀; Hwy 28, ENE of Fontana Village, N side Yellow Creek Mountains; 35.4387°N, -83.8122°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_120; • 5♀; Panther Creek, FT 405; 35.3683°N, -83.6267°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_119; • ♂, 8♀, 4 imm; Snowbird Mountains, N Tatham Gap, head of Long Creek on FR 423; 35.2579°N, -83.8196°W; 27 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_162; • ♂, ♀; south of Stecoah Gap on Appalachian Trail, Cheoah Mountains, NE of Cheoah; 35.3546°N, -83.7186°W; 18 Jul. 1991; B. Dellinger leg.; – **North Carolina, Macon Co.** • 29♀, 10 imm; Ball Road, SE of Beechertown; 35.2687°N, -83.6672°W; 30 Aug. 2002; M. Hedin, M. Lowder,

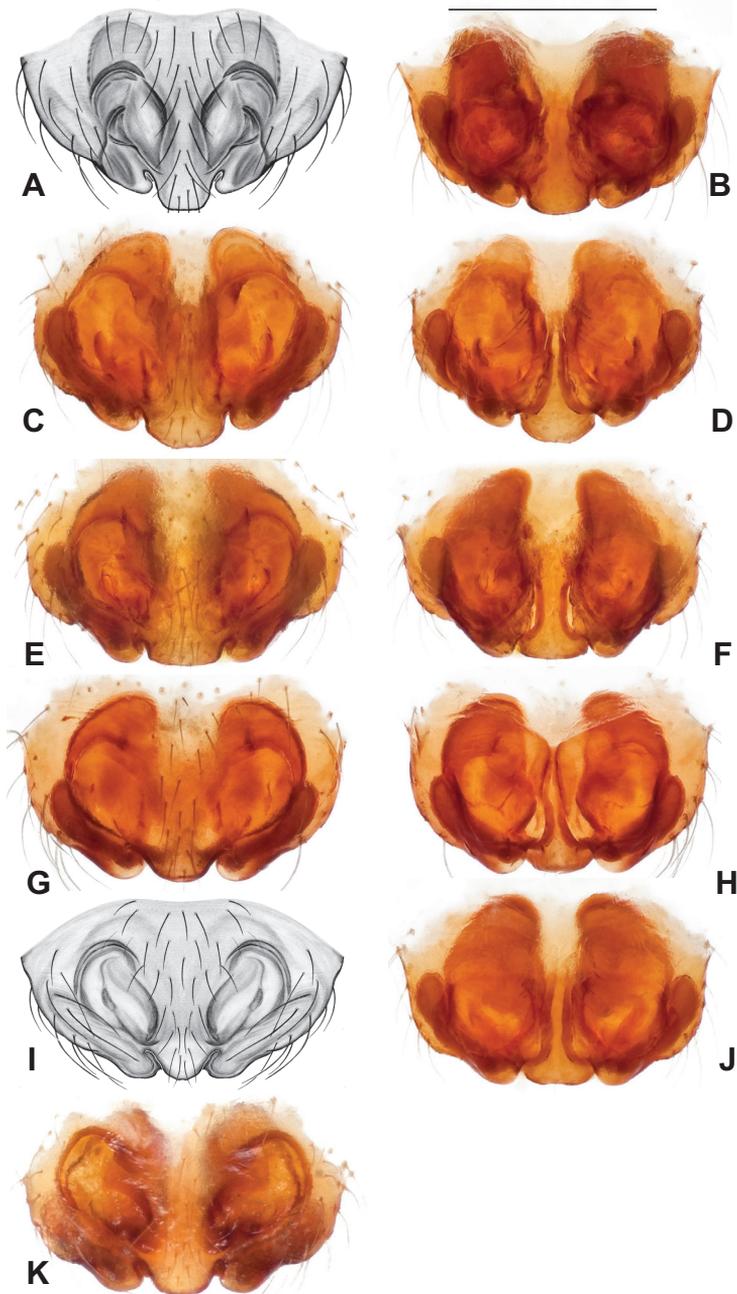


Figure 66. southwestern *Nesticus reclusus* epigynal variation. North Carolina, Graham Co., Appalachian Trail, S of Stecoah Gap, MCH 02_165, ventral (A), dorsal (B). North Carolina, Graham Co., ENE of Fontana Village, MCH 07_120, ventral (C), dorsal (D). North Carolina, Cherokee Co., Beaver Creek Road, MCH 04_057, ventral (E), dorsal (F). Georgia, Gilmer Co., Rock Creek Road, N of Rich Mountain Wilderness, MCH 07_102, MCH specimen #N1160, ventral (G), dorsal (H). North Carolina, Clay Co., Tusquitee Mountains, Long Branch of Fires Creek, MCH 02_144, ventral (I), dorsal (J). North Carolina, Macon Co., Ball Road, SE of Beechertown, MCH 04_072, ventral (K). Scale bar: 0.5 mm.

P. Paquin leg.; MCH 02_172; • ♂, 3♀; Ball Road, SE of Beechertown; 35.2687°N, -83.6672°W; 21 Aug. 2004; M. Hedin, R. Keith, J. Starrett, S. Thomas leg.; MCH 04_072; • 2♀; Jarrett Creek, W of Wayah Gap; 35.1587°N, -83.6349°W; 18 Aug. 2007; M. Hedin, M. McCormack, S. Derkarabetian leg.; MCH 07_123; • ♀; just N Jarrett Bald, above Wine Spring Creek; 35.1777°N, -83.6302°W; 1 May. 1993; B. Dellinger leg.; • 6♀; Nantahala River Gorge, SE of Hwy 74 19W, on Ball Road (also called Wayah Road); 35.2613°N, -83.6608°W; 10 Aug. 1992; M. Hedin leg.; • ♀; Nantahala River Gorge, vicinity Patton's Run Overlook; 35.278°N, -83.681°W; 29 Aug. 2001; M. Hedin, M. Lowder, P. Paquin leg.; MCH 01_183; • 2♂, ♀, 8 imm; S Burnington Gap, head of Ben Creek; 35.2185°N, -83.5639°W; 30 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_170; • ♂, 2♀, 12 imm; S of Wayah Bald on FR 388, 0.9 mi. S Wayah Road; 35.1559°N, -83.5512°W; 30 Aug. 2002; M. Hedin, M. Lowder, P. Paquin leg.; MCH 02_169; • ♀; Wine Spring Creek, E Nantahala Lake off Wayah Bald Road, S of Aquone; 35.1913°N, -83.6381°W; 25 Mar. 1993; B. Dellinger leg.; – **North Carolina, Swain Co.** • 2♀; Nantahala River Gorge, 0.25 mi. downstream from Blowing Spring, Hwy 19W; 35.3307°N, -83.6272°W; 18 Apr. 1994; M. Hedin leg.; • ♂, 2♀; Nantahala River Gorge, E side of River along Hwy 19W, across from Talc Mountain quarry, NE of Hewitt; 35.312°N, -83.6406°W; 8 Apr. 1993; B. Dellinger leg.

Diagnosis. Male palps of *Nesticus reclusus* are easily distinguished from close phylogenetic relatives *N. stupkai* and *N. bishopi*. In *N. reclusus* the distal tegular apophysis is shaped differently and has a blunt or forked tip, the space separating the distal from basal parts of the tegular apophysis is itself wide, and the median apophysis is shaped differently, with a spatulate basal end and a blade-like distal tip (Figs 63A–G, 64A–L). Females of these three species are challenging to diagnose; see comments above regarding the orientation of the dorsal epigynal plates.

Variation. We here discuss and distinguish *Nesticus reclusus* populations as “northeastern” vs. “southwestern”, separated by the Little Tennessee River, including the Little Tennessee River Gorge and Fontana Lake (Fig. 53). The type locality for *N. reclusus* is in the northeast, at Andrew's Bald in Great Smoky Mountains National Park. Southwestern populations surround and include the type locality of *N. cooperi* in the Nantahala River Gorge. We hypothesize that the Little Tennessee River might act as a dispersal barrier and promote divergence, although as discussed below combined evidence does not support this hypothesis.

In the northeast we examined males from seven locations in addition to the type locality, noting minimal palpal variation (Fig. 63A–G). One male from Lower Baskins Creek possessed a palp with a translucent bladelike paradistal process slightly wider at the base, and mostly lacking a ventromedial paracymbial process. Females from the northeast have conspicuously dark spermathecae and (viewed dorsally) the dorsal-projecting internal anterior plates are well sclerotized (Fig. 65A–F).

In the southwest we examined males from eighteen separate locations. All southwestern males approximated character conditions seen in northeastern males for all but one character. Males from eight locations possessed a paracymbium with the paradistal process lacking (and distomedial process moving towards the edge; Fig. 64I–L), like the condition seen in type *N. cooperi* (Gertsch 1984, figs 132–134). These locations

included Fires Creek (MCH 02_144, MCH 02_145), Ben Creek (MCH 02_170), Wayah Bald (02_169), Ball Road (MCH 04_072), Panther Creek (02_167, 07_119), Junaluska Road (MCH 04_059, MCH 07_122) and Nantahala River Gorge (1993 collection, very near the type locality of *N. cooperi*). These locations are geographically contiguous, found mostly along the western flanks of the Nantahala Mountains including the Nantahala River Gorge (Fig. 53).

Females from southwestern populations vary slightly (Fig. 66A–K), but those from sample locations with “*N. cooperi*-like” males (Fig. 66I–K) are not obviously different from other populations. That is, we could not discern a distinctive “*N. cooperi*-like” female morphology.

Distribution and natural history. This relatively wide-ranging montane species occurs from the northern side of the Great Smoky Mountains National Park, southwestward across the Little Tennessee River to the Yellow Creek, Cheoah, Snowbird, Nantahala, Valley River, and Tusquitee Mountains (Fig. 53). Two conspicuously disjunct populations occur even further south, in northern Georgia at Brasstown Bald and Rock Creek Road (Fig. 53). We comment more on the Rock Creek Road specimens below.

We hypothesize that the geographic gap north and northeast of Fontana Lake in the Great Smoky Mountains is an artifact of poor sampling, as this region is mostly roadless (Fig. 53). As such, northeastern vs. southwestern populations should be approximately contiguous, except for the river barrier itself. This differs from the situation in *Nesticus bishopi* versus *N. stupkai*, where we view the geographic disjunction as real (Fig. 53).

As an example of natural history, at Ball Road (MCH 02_172) a team collected 29 females and ten immatures in a 30-minute devoted survey from beneath rocks in a south-facing boulderfield. As mentioned above, *Nesticus reclusus* (♂, 4♀) was found in syntopy with *N. lowderi* (3♂, 5♀) at Fires Creek (MCH 02_144).

See comments above regarding the unlikely Gertsch (1984) record of *N. reclusus* from “*McDowell County, Montreat*”. Despite extensive collections we have never found members of the *reclusus* group from east of the Asheville Basin (Fig. 53).

Remarks. Gertsch (1984) described both *Nesticus reclusus* and *N. cooperi*, distinguishing males by the shape of the basal tegular apophysis and the shape / presence of a paradistal paracymbial process (Gertsch referred to this as a dorsal process; see our comments above). Importantly, although Gertsch examined many records for montane *N. reclusus*, he only had *N. cooperi* specimens from two adjacent Nantahala River Gorge populations. Our geographic sampling has greatly expanded the distribution for southwestern *N. reclusus*, including many locations surrounding the type locality of *N. cooperi*. With this greater sampling we found that male morphology varies slightly with geography, particularly in the presence of the paradistal paracymbial process. We could not discern the shape differences in the basal tegular apophysis that Gertsch (1984) noted (Figs 63, 64). From a morphological perspective we view this as a single species with a relatively broad montane distribution, with minor male morphological variation across this distribution.

Nuclear phylogenomic data is mostly consistent with this single species hypothesis, except for the southern disjunct Rock Creek Road population, further discussed below.

Only one “*Nesticus cooperi-like*” population was sampled for nuclear data (Nantahala River Gorge) and is embedded within a paraphyletic grade including both northeastern and other southwestern *N. reclusus* (Figs 3, 4). The nuclear data within this complex are notable for many low gene and site CF values, and low local posterior probability values (Figs 3, 4), suggesting extensive gene tree discordance.

The mitochondrial evidence is similarly challenging to interpret in this complex, as mitochondrial data do not support the larger *reclusus* group as monophyletic, and species interrelationships diverge strongly from that suggested by the nuclear data (Fig. 6). Within *Nesticus reclusus* itself Noland and Clingmans sequences are recovered with *N. stupkai* sequences, separate from Smokemont and Newfound sequences. We hypothesize that this discordance is a result of mitochondrial introgression from *N. stupkai* into certain *N. reclusus* populations, where these taxa occur in geographic proximity. For example, sympatry in Myhr Cave is a potential conduit for mitochondrial gene exchange. Six sampled locations with a “*N. cooperi-like*” paracymbium do not form a clade on mitochondrial trees (Fig. 6).

The southern disjunct Rock Creek Road sample (Fig. 53) adds further intrigue to this complex. Mitochondrial sequences are highly divergent, falling with *Nesticus sheari* (Fig. 6), while nuclear sequences are sister to a clade including *N. stupkai*, *N. bishopi*, and remaining *N. reclusus* (Figs 3, 4). At the same time, males from this location possess unremarkable palps, identical in detail to other southwestern *N. reclusus* palps (Fig. 64E, F), and females are similarly morphologically unremarkable (Fig. 66G, H). We suspect that gene flow across species boundaries (perhaps involving *N. sheari*?) might be impacting results in this part of the *Nesticus* phylogeny. More geographic and UCE sampling in this geographic region will be needed to resolve this tricky taxonomic issue.

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Supplementary material 1

***Nesticus* morphological material examined**

Authors: Marshal Hedin, Marc A. Milne

Data type: occurrences (excel file)

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Link: <https://doi.org/10.3897/zookeys.1145.96724.suppl1>

Supplementary material 2

***Nesticus* immature specimen records**

Authors: Marshal Hedin, Marc A. Milne

Data type: occurrences (excel file)

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Supplementary material 3

Specimens used in molecular work

Authors: Marshal Hedin, Marc A. Milne

Data type: DNA records (excel file)

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Supplementary material 4

Input alignments, analysis log files, and output tree files

Authors: Marshal Hedin, Marc A. Milne

Data type: zip. archive

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A new species of *Dipsas* (Serpentes, Dipsadidae) from central Panama

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Abstract

A new species of *Dipsas* Laurenti, 1768, from Central Panama is described based on molecular analyses, hemipenial morphology, and external characters. This is the sixth species of *Dipsas* to be described for the country; the snake has been suspected to exist since 1977 and has not been thoroughly studied until now. Additionally, morphological comparisons including scale counts are done with other species within the genus, and the current geographic distribution of *Dipsas temporalis* (Werner, 1909), the sister species, is updated. Finally, a key to the species of *Dipsas* currently known from Middle America is presented.

Resumen

Describimos una nueva especie de *Dipsas* Laurenti, 1768 de la región central de Panamá en base a análisis moleculares, morfología hemipenial y caracteres de morfología externa. Esta es la sexta especie del género *Dipsas* descrita para el país. Se sospechaba su existencia desde 1977 pero no había sido estudiada

exhaustivamente hasta ahora. Adicionalmente, presentamos comparaciones morfológicas (incluyendo lepidosis) con otras especies del género y actualizamos la distribución geográfica de su especie hermana *Dipsas temporalis* (Werner, 1909). Finalmente, presentamos una clave para las especies de *Dipsas* distribuidas en Centroamérica.

Keywords

Dipsadini, *Dipsas temporalis*, new species, phylogeny, snail-eating snake, systematics

Introduction

The Neotropical snake genus *Dipsas* Laurenti, 1768, belongs to the tribe Dipsadini, a group of primarily arboreal snakes that includes the genera *Dipsas*, *Plesiodipsas* Harvey et al., 2008, *Sibon* Fitzinger, 1826, and *Tropidodipsas* Günther, 1858 (Harvey et al. 2008; Zaher et al. 2009; Grazziotin et al. 2012; Arteaga et al. 2018). The “snail-eating” snakes (Mertens 1952; Peters 1956) or “snail-suckers” (Peters 1956) are part of a larger group of neotropical snakes called the “goo-eaters” (Cadle and Greene 1993) because of their proclivity for feeding on soft and often slimy invertebrates. According to Cadle and Greene (1993), the “goo-eaters” also include the mainly earthworm- and slug-eating species in the genera *Adelphicos* Jan, 1862, *Atractus* Wagler, 1828, *Geophis* Wagler, 1830, and *Ninia* Baird & Girard, 1853, and possibly also *Chersodromus* Reinhardt, 1860 and *Cryophis* Bogert & Duellman, 1963 (see Sheehy 2012); though, *Cryophis* is also known for preying on salamanders (Mulcahy 2007). The discovery of a broader diet for some snail-eating snakes of the genera *Sibon* and *Dipsas* to include additional invertebrates and anuran eggs further refined our understanding of the diet of these snakes (Ryan and Lips 2004; Montgomery et al. 2007; Ray et al. 2012).

The genus *Dipsas* currently contains 53 small- to moderately-sized species that can be distinguished from the other genera of the tribe by external features, such as body often strongly compressed (in arboreal taxa), head distinct from neck, usually more than 10 infralabials, vertebral scale row usually enlarged, preoculars 0–2, supralabials and infralabials not notably enlarged, mental groove very weak to absent, and often two or more pairs of infralabials in contact behind mental (Peters 1960; Harvey and Embert 2008; Uetz et al. 2022). Harvey and Embert (2008) also describe internal characteristics, including a well-developed tracheal lung and characteristics of the hemipenes. Species of *Dipsas* are Neotropical and range from central Mexico to southern South America (Peters 1960; Solórzano 2004; Ray 2009), and five species are currently recognized in Panamanian territory: *D. articulata* Cope, 1868, *D. nicholsi* (Dunn, 1933), *D. temporalis* (Werner, 1909), *D. tenuissima* Taylor, 1954, and *D. viguieri* (Bocourt, 1884). Detailed reviews of Panamanian *Dipsas* are provided by Peters (1960), Savage (2002), Cadle and Myers (2003), and Ray (2017). Of these, *D. tenuissima* is at the southern and easternmost extent of its range and *D. viguieri* is at the northern and westernmost extent of its range in Panama (Ray 2017). Based on current information, *D. nicholsi* is endemic to the country, with most records found east of the Panama

Canal and one record west of it (Myers et al. 2007). *Dipsas temporalis* was, until now, one of the most widespread of the species of *Dipsas* in Panama (Ray 2017).

The most complete, recent taxonomic review of the genus was by Peters (1960), who principally used color pattern to recognize species. This has led to several remaining taxonomic issues. The availability of new material has resulted in species and groups of species within the genus being revised frequently in subsequent years (Cadle and Myers 2003; Passos et al. 2004, 2005; Cadle 2005; Harvey 2008; Harvey and Embert 2008; Sheehy 2012; Arteaga et al. 2018). Phylogenetic relationships among species of *Dipsas* and closely related genera remain unclear, since most phylogenetic studies published regarding snake systematics (Zaher et al. 2009; Vidal et al. 2010; Grazziotin et al. 2012; Pyron et al. 2013; Figueroa et al. 2016) have not sampled a sufficient set of species in these genera. However, all these studies corroborated paraphyly of the genus *Dipsas* with respect to *Sibynomorphus* (see Sheehy 2012). A recent study focused on the systematics of South American *Dipsas* and *Sibon* described several new species, and synonymized *Sibynomorphus* with *Dipsas* (Arteaga et al. 2018).

Between 1997 and 2015, one of us (JMR) regularly studied reptiles and amphibians in Parque Nacional General de División Omar Torrijos Herrera (PNGDOTH), near the community of El Copé de La Pintada, Coclé Province, Republic of Panama. In 1977, before the area was established as a national park, it was visited by the late Charles W. Myers, who suggested that at least one undescribed species of *Dipsas* occurred at the site (Myers et al. 2007). More recently, other researchers have agreed with that assessment (Ray et al. 2012). After examination of specimens collected in 2006–2009 and 2011 and after analysis of molecular data, including the updated phylogeny constructed for this paper, we confirm the existence of at least one new species of *Dipsas* at this site, which we herein describe. Additionally, we have confirmed the presence of this species at other sites. We also confirm that *Dipsas temporalis*, the species to which this snake was believed to belong, is still found in Panama; thus, we update the range of *D. temporalis*. Finally, we provide a key to the Central American species of the genus *Dipsas*.

Materials and methods

Ethics statement

This study was carried out in strict accordance with the guidelines for use of live amphibians and reptiles in field research (Beaupre et al. 2004) compiled by the American Society of Ichthyologists and Herpetologists (**ASIH**), the Herpetologists' League (**HL**), and the Society for the Study of Amphibians and Reptiles (**SSAR**). All procedures with animals (see below) were reviewed by the Ministerio del Ambiente, Agua y Transición Ecológica (**MAATE**) in Ecuador and by UNARGEN-Ministerio de Ambiente in Panamá, and specifically approved as part of obtaining the following field permits for research and collection: MAE-DNB-CM-2018-0105 and MAATE-DBI-

CM-2022-0245 (granted to Universidad San Francisco de Quito) and SC/A-8-09, SC/A-28-09, SC/A-37-11, SC/A-33-12, SE/A-60-16, and SE/A-33-18 (granted to Museo Herpetológico de Chiriquí). Specimens were euthanized with 20% benzocaine, fixed in 10% formalin or 90% ethanol, and stored in 70% ethanol. Museum vouchers were deposited at the Smithsonian National Museum (**USNM**), Museo Herpetológico de Chiriquí (**MHCH**), the Senckenberg Forschungsinstitut Frankfurt (**SMF**), and at Museo de Zoología de la Universidad San Francisco de Quito (**ZSFQ**).

Common names

Criteria for common name designation are as proposed by Caramaschi et al. (2006) and Coloma and Guayasamin (2011–2017), reviewed by Arteaga et al. (2019). These are as follows (in order of importance): (i) the etymological intention (implicit or explicit) that the authors used when naming the species (specific epithet); (ii) a common name that is already widely used in the scientific literature; (iii) a common name that has an important ancestral or cultural meaning; (iv) a common name based on any distinctive aspect of the species (distribution, morphology, behavior, etc.).

Material examined

We examined 31 specimens suspected to be a new species from 15 locations in Panama. Of these, we examined 23 specimens collected at Parque Nacional General de División Omar Torrijos Herrera (**PNGDOTH**), located 7.5 km north of the community of El Copé de La Pintada, Coclé Province, Republic of Panama (8.670383, -80.592343, 763 m a.s.l.) between 650 and 850 m. Specimens from eight other species of *Dipsas* also were examined for comparison purposes (Appendix 1).

We gathered additional data for the Central American species of *Dipsas* from Peters (1960), Savage (2002), Cadle and Myers (2003), Solórzano (2004), and Ray (2017). We follow Dowling (1951) for the method of counting ventrals and subcaudals and Savage (1973) for the terminology of scales in the loreal region of the head. We follow Peters (1960) and Harvey and Embert (2008) for terminology for cephalic shields. Sex was determined by probe or by subcaudal incision unless hemipenes were everted. Head and scale measurements were made to the nearest 0.1 mm using digital calipers held under a dissecting microscope. Snout-vent length and tail length measurements were taken to the nearest 1.0 mm using a squeeze box (Quinn and Parker 1976) or tape measure.

Terminology for measurements is abbreviated as: snout-vent length, **SVL**; tail length, **TL**; total length, **TOL**; head length, **HL**; jaw length, **JL**; and head width, **HW**. Eye length equals the horizontal distance across eye at widest point. Scale dimensions were measured at the longest or widest points along the longitudinal or perpendicular axis of the body, respectively. Drawings of the head were made using digital photography and a dissecting microscope by Shannon Christensen. Hemipenial preparation

follows Zaher (1999) and Zaher and Prudente (2003). Once prepared, the hemipenes were stained with alizarin in 70% ethanol to facilitate the visualization of calcified structures (Harvey and Embert 2008; Nunes et al. 2012).

Molecular phylogenetics

A subset of molecular data is presented here for 19 species of *Dipsas* (Appendix 3), taken from the thesis of CMS (Sheehy 2012), which included 175 total taxa representing most other genera in the subfamily Dipsadinae. Five loci were used: (1) a 714 base pair fragment of the mitochondrial NADH dehydrogenase subunit 4 (ND4), (2) a 199 base pair fragment of tRNAs His, Ser and Leu, (3) a 1071 base pair fragment of the mitochondrial cytochrome-b gene (cyt-b), (4) a 525 base pair fragment of the nuclear protein-coding neurotrophin-3 (NT3) gene, and (5) a 732 base pair fragment of the nuclear protein-coding dynein, axonemal, heavy chain 3 (DNAH3) gene (see Appendix 2 for primers used). Genomic DNA was isolated from tissues using a Qia-gen DNeasy kit (Qiagen, Valencia, California, USA). All amplification reactions used GoTaq Green Master Mix, 2X (Promega Corporation, Madison, Wisconsin, USA). Thermal cycling followed standard protocols and are detailed in Sheehy (2012). Successfully amplified PCR products were prepared for sequencing by using the ExoSAP-IT kit (United States Biochemical). A BigDye Terminator Cycle Sequencing Kit (Applied Biosystems Inc.) was used for sequencing following the manufacturer's protocol and using PCR primers. The sequenced products were precipitated using an ethanol/sodium acetate method and rehydrated in HPLC purified formamide (HIDI). The sample was then analyzed on an ABI PRISM 3100xl Genetic Analyzer in the Genomics Core Facility at the University of Texas at Arlington, USA.

Alignments were constructed using the program Sequencher 4.8 (Gene Codes, Ann Arbor, Michigan, USA), and edited by eye using the program MacClade 4.08 (Maddison and Maddison 2005). The tRNAs were aligned using an annotated mitochondrial genome for *Sibon nebulatus* (GenBank accession number EU728583; Mulcahy and Macey 2009) as a template sequence.

Phylogenetic analyses were conducted using Maximum Likelihood (ML) and Bayesian Index (BI) on the data matrix consisting of 194 taxa and up to 3241 base pairs. Various models of molecular evolution were tested using the software package MEGA 5 (Tamura et al. 2011) on the complete alignment partitioned by gene fragment (seven partitions: ND4, cyt-b, tRNA His, tRNA Ser, tRNA Leu, NT3, and DNAH3). The model test results identified GTR+I+G and GTR+G as among the best-fit models of nucleotide substitution for each gene fragment based on corrected Akaike Information Criterion (AICc), although they did not always receive the best scores. The ML analyses employing the rapid bootstrapping algorithm were conducted using the program RAxML 7.3.0 (Stamatakis 2006) on the CIPRIS Science Gateway server v. 3.2 (Miller et al. 2010) using the model GTR+G instead of GTR+I+G because the 25 discrete rate categories appear to better estimate invariant sites (Stamatakis 2006).

The multiple alignment was partitioned by gene region (five partitions: ND4, cytb, tRNAs, NT3, DNAH3), which allowed RAxML to calculate and apply the most appropriate gamma distribution parameter to each partition separately. Nodal support for ML was provided by rapid bootstrapping (1000 pseudoreplicates), with bootstrap values ≥ 0.70 considered strong support (Hillis and Bull 1993).

Bayesian analyses were conducted with the computer program MrBayes (Huelsenbeck and Ronquist 2001) on a partitioned alignment using the reversible-jump Markov chain Monte Carlo algorithm (mixed model), which avoids the risk of acquiring misleadingly high posterior probabilities at the nodes of hard or nearly hard polytomies due to their arbitrary resolution (Lewis et al. 2005). Each of the four protein coding genes in the alignment was partitioned by codon position with one partition including the first and second positions and another including the third position for a total of nine partition schemes (the three tRNAs were not partitioned).

Two independent runs were conducted simultaneously with four Markov chains (three heated and one cold) per run, and average standard deviation of the split frequencies below 0.01 was considered acceptable. Stationarity was determined to be reached visually using Tracer v. 1.5 (Rambaut and Drummond 2009). The analysis ran for 17,000,000 generations while sampling trees every 1000 generations. Stationarity was reached after approximately 11,500,000 generations, after which the standard deviation of the split frequencies dropped to 0.008. Therefore, we sampled the resulting 5000 trees from the last five million generations (12–17 million generations), which should be a good representation of the posterior distribution of trees. The initial 12 million generations were discarded as burn-in, and a 50% majority rule consensus tree with estimates of Bayesian support was constructed using the remaining sampled trees. Posterior probabilities (PP) provided nodal support for Bayesian analyses, with PP values ≥ 0.95 considered strong support (Alfaro et al. 2003; Huelsenbeck and Rannala 2004; Mulcahy et al. 2011).

Distribution maps and ecological niche models

We present ranges of occurrence for two species of *Dipsas*, *D. temporalis* and a new species herein described. Presence localities are derived from museum vouchers (Appendix 1), photographic records (iNaturalist), and the literature. For each species, a binary environmental niche model (ENM) accompanies the dot maps. These models estimate potential areas of distribution based on observed presences and a set of environmental predictors (Elith and Leathwick 2009). To delimit the occupancy areas and the potential species distribution, we used the BAM diagram proposal (Soberón and Peterson 2005; Peterson et al. 2011). To create the models, we used presence localities as described above, 19 bioclimatic variables from Worldclim 1.4 (Hijmans et al. 2005), and Maxent 3.4.1k, an algorithm based on the principle of maximum entropy (Phillips et al. 2006; Elith et al. 2011; Renner and Warton 2013).

For the first explorative exercise, we used the 19 climate layers from the WorldClim project and assessed which variables were the most important for the model, according to the Jackknife test calculated in MaxEnt (Royle et al. 2012). Correlated environmental

variables ($r < 0.8$) were identified using the PEARSON correlation test of PAST 3. In a second modelling exercise, we used the locality records for each species and the variables identified in the first approach to generate the species distribution. 5,000 iterations were specified to the program with clamping and no extrapolation. All other parameters in MaxEnt were maintained at default settings. To create the binary environmental niche models, suitable areas were distinguished from unsuitable areas by setting a minimum training presence threshold value. The logistic format was used to obtain the values for habitat suitability (continuous probability from 0 to 1), which were subsequently converted to binary presence-absence values on the basis of the established threshold value, defined herein as the minimum training presence. The convergence threshold was set to 10^{-5} , maximum iterations to 500, and the regularization parameter to “auto.”

Results

Systematics

The ML and Bayesian analyses were largely congruent, particularly with respect to the well-supported clades. The ML phylogeny of a well-supported clade containing most species of *Dipsas* sampled (except “*D.*” *gaigeae*; see Sheehy 2012) is here presented, with Bayesian posterior-probabilities superimposed on well-supported nodes (Fig. 1). The specimens from PNGDOTH formed a clearly divergent, strongly supported lineage separate from the other Central American species and is sister to *Dipsas temporalis*, to which it differs by ~ 7% (uncorrected pairwise-distance) for the ND4 locus. Based on this genetic distinctiveness, along with discontinuous morphological variation in scalation and unique hemipenes morphology (see below), we determine that it does, indeed, represent a new species as previously hypothesized.

Dipsas aparatiritos sp. nov.

<https://zoobank.org/E96CAB59-FBB7-451B-9D11-4372182F9809>

Figs 2–8, Appendix 3

Proposed standard English name: Hidden Snail-eating Snake

Proposed standard Spanish name: Caracolera Escondida

Type material. Holotype. PANAMA • ♀; PNGDOTH, ca. 7.5 km N of El Copé de La Pintada, Coclé Province, 8.670383°N, 80.592343°W, 763 m a.s.l.; 30 Jul 2010; S. Gotte, J. Jacobs, D. Mulcahy and R. Reynolds; USNM 579828 (Biol. Survey Field Series 4608) (Figs 3, 4).

Paratype. PANAMA • ♀; PNGDOTH, ca. 7.5 km N of El Copé de La Pintada, Coclé Province, 8.670383°N, 80.592343°W, 763 m a.s.l.; 30 Jul 2010; S. Gotte, J. Jacobs, D. Mulcahy and R. Reynolds; USNM 579829 (Biol Survey Field Series 4609) (Figs 5, 6).

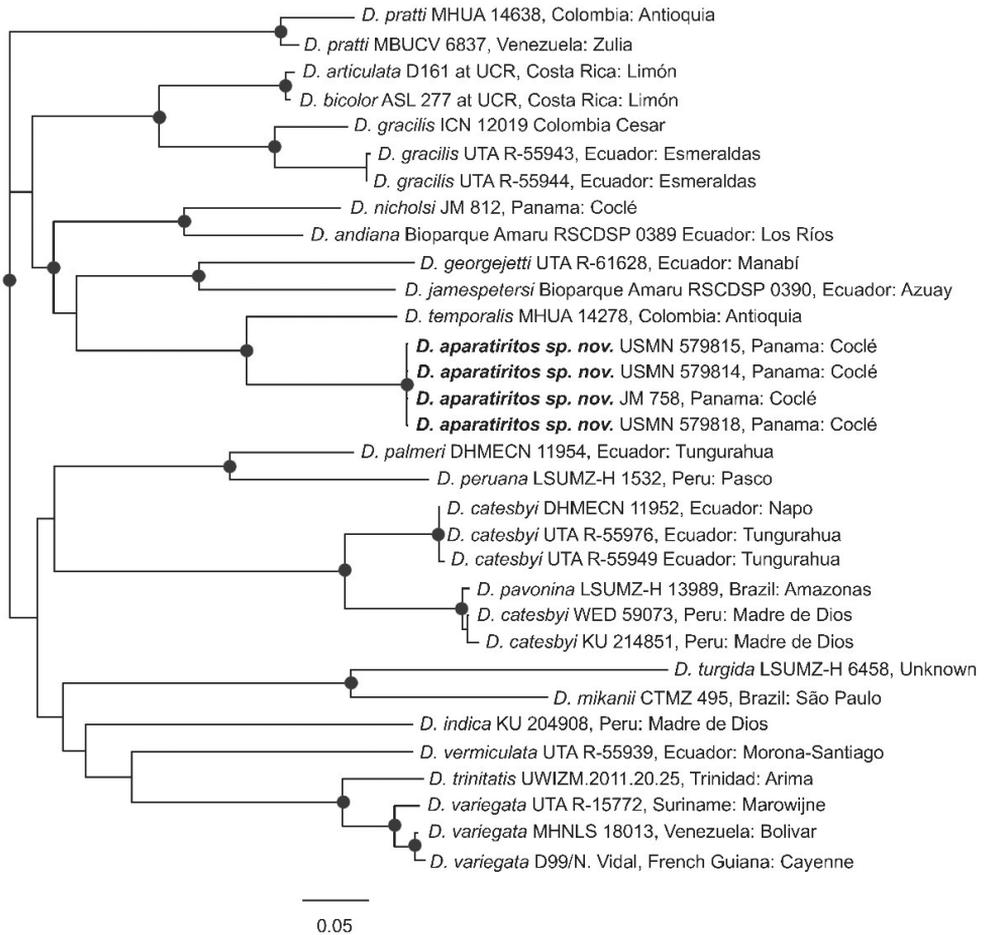


Figure 1. Phylogeny of 20 species of *Dipsas* using the best ML tree. Black circles denote strong nodal support (≥ 0.95 PP and ≥ 0.70 ML bootstrap). See Sheehy (2012) for further details on the outgroup taxa.

Diagnosis. *Dipsas aparatiritos* sp. nov. is placed in the genus *Dipsas* based on phylogenetic evidence (Fig. 1) and the absence of a labial that is noticeably higher than other labials. The species is diagnosed based on the following combination of characters: (1) 15/15/15 smooth dorsals with enlarged vertebral row (1.5–2.4× as wide as adjacent rows); (2) loreal and a preocular in contact with orbit; (3) 7 supralabials with 4th and 5th contacting orbit, 1st supralabial fused with nasal scale; (4) 8–9 infralabials with 3rd to 6th in contact with chin shields, first pair of infralabials not in contact behind symphysial due to presence of two postmentals; (5) 191–196 ventrals in males, 177–197 in females; (6) 122–136 divided subcaudals in males, 111–126 in females; (7) dorsal and ventral color consisting of 17–20 dark brown to black white-bordered body bands (10–12 dorsal scales long anteriorly to 3–5 dorsal scales long posteriorly) separated from each other by white to pale yellow (anteriorly) to pale brown (posteriorly) interspaces measuring 2–6 dorsal scales long, ventral surfaces white with encroachment from the



Figure 2. Live individual of *Dipsas aparatiritos* sp. nov. in Parque Nacional General de División Omar Torrijos Herrera photographed in the wild and not collected. Photography by Kevin Enge.

dorsal dark blotches and with smaller blackish marks in-between the blotches, dorsal aspect of head dark reddish brown with small blotches on the labial and temporal scales as well as a pale nuchal collar, throat white with small dark brown to blackish markings, iris pale brown with minute black speckles; (8) 310–465 mm SVL in males, 169–424 mm females; (9) 122–260 mm TL in males, 65–247 mm in females.

Description of the holotype. An adult female; SVL 424 mm; TL 211 mm (49.7% SVL); head broadly distinct from body; head length 13.2 mm (3.10% SVL); head width 7.3 mm (55% head length); snout-orbit distance 3.3 mm; eye diameter 2.5 mm; rostral broader than high, triangular in frontal view, not visible from above; internasals broader than long; prefrontals broader than long and do not enter the orbit; from above, the triangular shape of the top of the preocular is visible; supraocular longer than broad; frontal longer than broad, with a triangular shape in dorsal view; parietals longer than broad; nasal entire and fused with the first supralabial on both sides; loreal longer than high, enters the orbit; one upper preocular; two postoculars; temporals 2+3 left side, 2+2 right side, where the upper primary and secondary scales are fused; 7 supralabials, 4 and 5 contacting orbit (first supralabial is fused with the nasal) symphyisial contacting the first pair of chin shields; 9 infralabials; four pairs of irregular chin shields, the first pair is smaller, second pair is longer than broad, the third pair is slightly longer than broad, but its scales are not in contact, the last pair is broader than long. Dorsals smooth in 15-15-15 rows; mid-vertebral scales moderately enlarged; 178 ventral scales; 118 paired subcaudals; cloacal scale single.



Figure 3. Holotype (USNM 579828) of *Dipsas aparatiritos* sp. nov. showing **a** dorsum and **b** venter. Ruler units in cm. Photographs by James Poindexter.

In preservative, dorsal ground color of head uniformly brown except for some small dark-brown blotches on the occipital areas; laterals with small pale brown and dark blotches; white supralabials with evident pale brown and dark blotches; ground color of infralabial and gular region cream colored with dark-brown blotches and pale-brown spots; dorsal color of body pale brown with dark-brown blotches and pale interspaces; on the anterior portion of the body the blotches are dark-brown and long (between 10 and 13 scales) contacting the opposite one in the vertebral row, the interspaces are pale brown with small and scarce dark-brown spots on the dorsal, and white on the lateral; on the middle of the body, the dark-brown blotches diminish their length (between 8 and 9 scales), and they lose the dorsal continuation between them in the vertebral row, the interspaces get a pale-brown color with some small dark-brown spots; on the posterior portion, the blotches are shorter (5–7 scales), rounded, and they are margined by a white edge with many small dark-brown spots; ground color of the belly cream-colored, with irregular blotches of different sizes along the ventral line of the interspaces; tail resembles the body in color pattern; body with 16 blotches, and tail with 12. Color in preservative (70% ethanol) similar to color in life.

Description of the paratype. An adult female; SVL 328 mm; TL 170 mm (51.8% SVL); head broadly distinct from body; head length 12.2 (3.7% SVL); head width 6.6mm (54% head length); snout-orbit distance 2.9 mm; eye diameter 2.3 mm; rostral broader than high, triangular in frontal view, not visible from above; internasals, broader than long; prefrontals long as wide, no enter the orbit; from above, the triangular shape of the top of the preocular is visible; supraocular longer than broad; frontal longer than broad, with a triangular shape in dorsal view; parietals longer than broad; nasal entire; loreal longer than high, enters the orbit; one upper preocular; two postoculars; temporals 2+3 left side, 3+4 right side; 8 supralabials, 4 and 5 contacting orbit; symphyisial contacting the first pair of chin shields; 9 infralabials; three pairs of irregular chin shields, the first pair is the smaller, second pair is longer than broad; the third pair is slightly broader than long. Dorsals smooth in 15-15-15; vertebral scale moderately enlarged; 183 ventral scales; 124 paired subcaudals; cloacal scale single. In preservative, dorsal ground color of head uniformly brown except for some small dark-brown blotches on the occipital areas; laterals with small blotches pale brown and dark; white supralabials with evident pale brown and dark blotches; ground color of infralabial and gular region cream with dark-brown blotches and pale-brown spots; dorsal color of body pale-brown with dark-brown blotches and pale interspaces; on the anterior and middle portion of the body the blotches are dark-brown and long (12–14 scales) contacting the opposite one in the vertebral row, the interspaces are pale brown with small and scarce dark-brown spots on the dorsal, and white on the lateral; on the posterior portion, the blotches are shorter (between 5 and 7 scales), rounded, they are margined by a white edge with many dark-brown small spots, and they lose the dorsal continuation between them in the vertebral row, the interspaces get a pale-brown color with some small dark-brown spots; ground color of the belly cream, with irregular blotches of different sizes along the ventral line of the interspaces; tail resembles the body; body with 19 blotches, and tail with 15. Color in preservative (70% ethanol) similar to color in life.

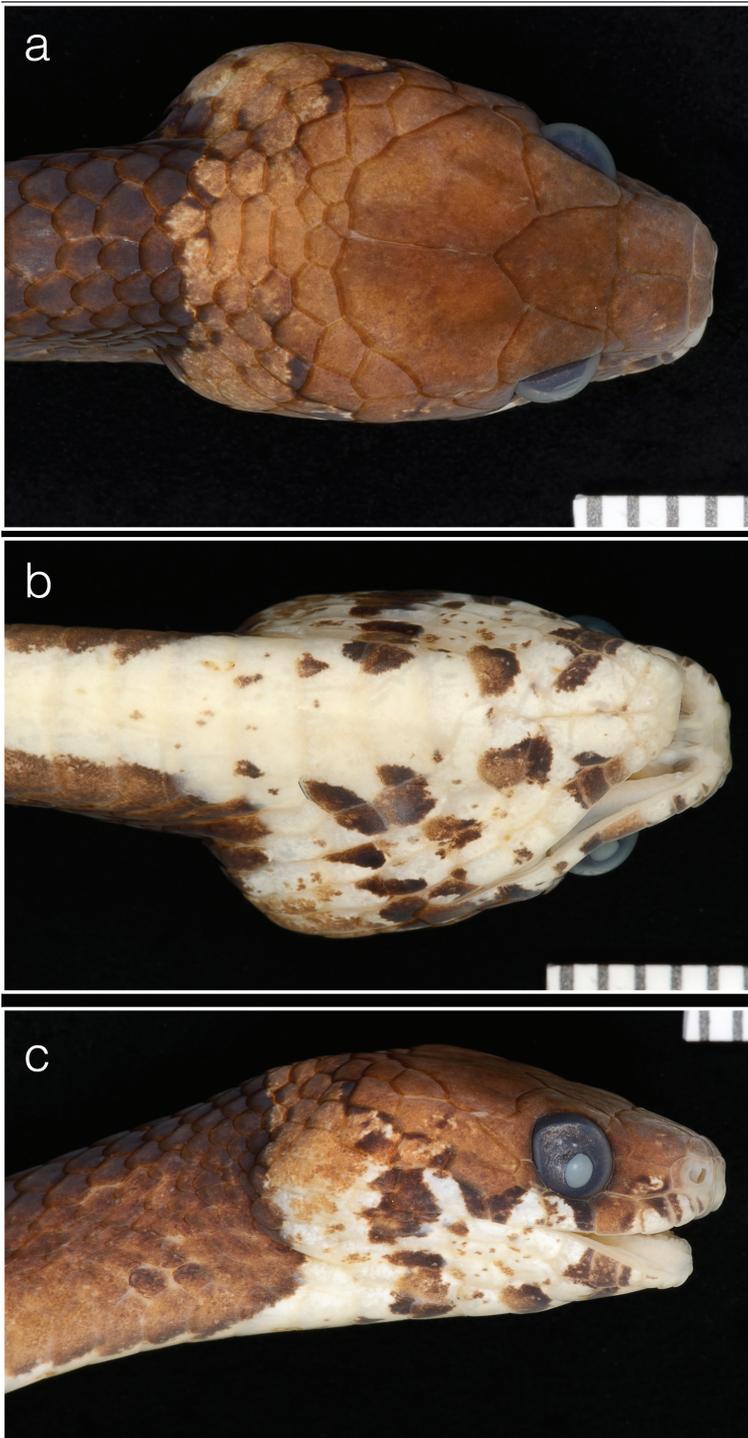


Figure 4. Holotype (USNM 579828) of *Dipsas apantiritos* sp. nov. showing **a** dorsum of head and **b** chin shields and **c** lateral view. Ruler notches denote mm. Photographs by James Poindexter.



Figure 5. Paratype (USNM 579829) of *Dipsas aparatiritos* sp. nov. showing **a** dorsum and **b** venter. Ruler units in cm. Photographs by James Poindexter.

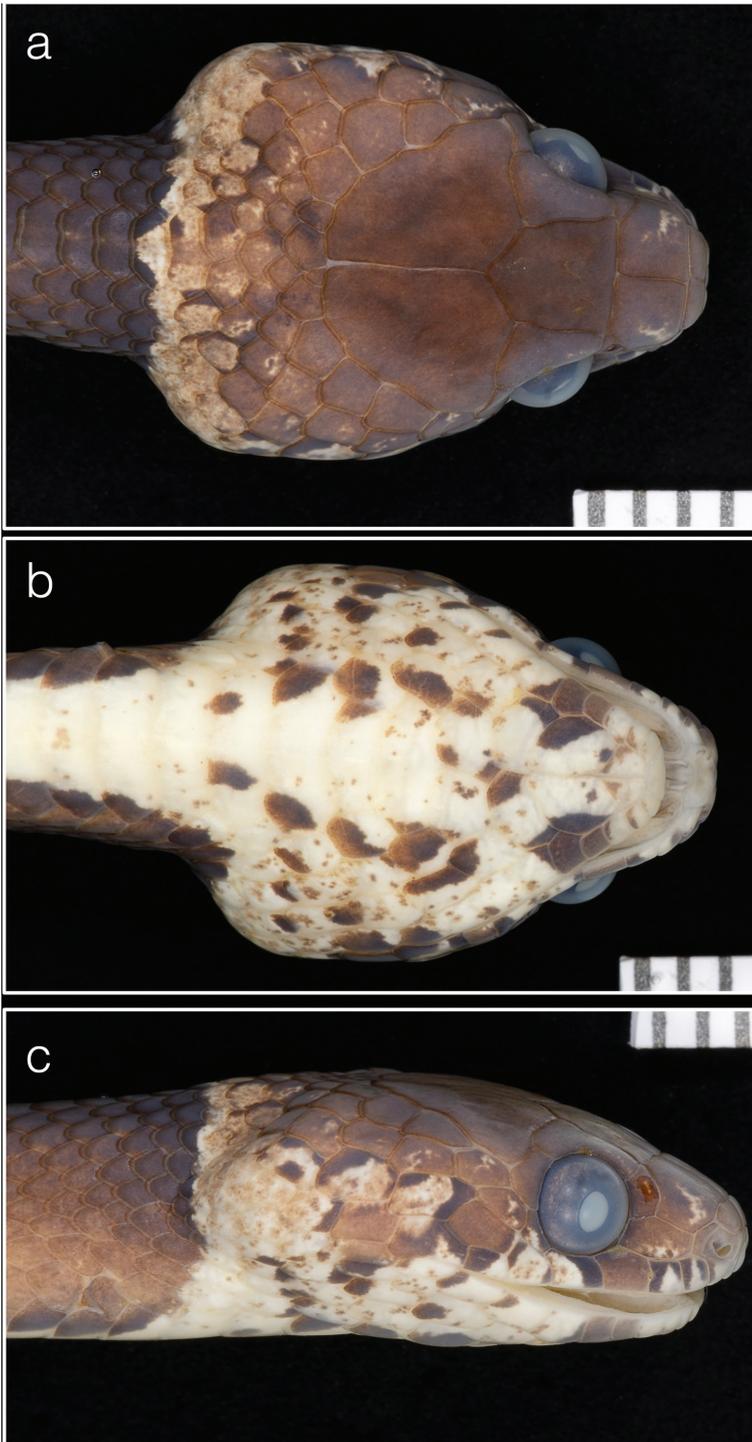


Figure 6. Paratype (USNM 579829) of *Dipsas aparatiritos* sp. nov. showing **a** dorsum of head and **b** chin shields and **c** lateral view. Ruler notches denote mm. Photographs by James Poindexter.

Referred specimens. MHCH 2311, juvenile male collected by Sebastian Lotzkat and Andreas Hertz on 18 August 2010 at Cerro Mariposa, Veraguas province, Panama (8.51166°N, 81.12163°W; 940 m), SMF 89551–53, adult males collected by Leonhard Stadler and Nadim Hamad between 8 May and 7 July 2008 at the type locality. SMF 90036, adult male collected by Arcadio Carrizo on 28 July 2008 at Cerro Negro, Veraguas province, Panama (8.56901°N, 81.09894°W; 700 m). SMF 97346, adult male collected by Abel Batista on 25 January 2013 at Donoso, Coclé province, Panama. SMF 89953–54, juvenile and adult of undetermined sex, respectively, collected by Leonhard Stadler and Nadim Hamad on 8 May 2008 at the type locality. SMF 89769, juvenile of undetermined sex collected by Sebastian Lotzkat and Andreas Hertz on 3 April 2009 at Cerro Negro, Veraguas province, Panama (8.56901°N, 81.09894°W; 700 m). MHCH 3123, adult female collected by Marcos Ponce and Roger Morales on 30 May 2018 at Cerro Campana, Panama province, Panama (8.69378°N, 79.92098°W; 730 m).

Additionally, a series of individuals was collected from Parque Nacional General de División Omar Torrijos Herrera between 2006 and 2009 that included 15 females and 12 males. There was variation between sexes and among individuals (Tables 1–3). A summary of the most commonly measured characteristics includes the range of 173–192 ventrals in females ($n = 11$) and 187–191 in males ($n = 12$), subcaudals 116–131 in females ($n = 13$) and 129–136 in males ($n = 8$). All individuals had either 7 or 8 supralabials on both sides ($n = 26$) except one female USNM 579810 with only 6 on the left. Individuals ($n = 25$) had 8 or 9 left infralabials with two individuals having 10. However, the right infralabials ranged from 7–9 with the same individual as above (USNM 579810) having 6 (Fig. 7).

Hemipenial morphology. Description based on the hemipenes fully everted, but not completely expanded, for the specimen USNM 579815 (Fig. 8). Distal end of retractor muscle divided, hemipenis unilobed, unicapitate and unicalyculate; capitulum with papillate and spinulate calyces, it covers approximately the distal half of the organ in the sulcate face, and the distal one-third in the asulcate; the inferior capitular edge of the sulcate face is V-shape, and in the asulcate face the capitular arch is present. In both faces, the hemipenial body is covered by a few small spines, and mostly by medium-sized spines which have curved and robust tips. The base of the organ also is covered by dispersed little spinules on both faces; there is not an evident nude pocket, and there are two spines of similar size on the asulcate side. The sulcus spermaticus bifurcates at the base of the capitulum; both branches diverge and extend diagonally oriented, and end at the distal edge of the lateral face of the organ.

Comparisons. *Dipsas aparatiritos* sp. nov. can be distinguished from all other similar or related species by the following combination of characters: 15 dorsal scale rows; one upper preoculars; two or three postoculars; temporals 1+2; seven or eight supralabials, fourth and fifth contacting the orbit; eight or nine infralabials, no infralabials in contact behind mental; vertebral row moderately enlarged; 191–196 ventrals in males, and 177–197 in females; 129–136 subcaudals in males, and 111–131 in females; by the alternating dark brown and tan brown bands running the length of the body, including the tail.

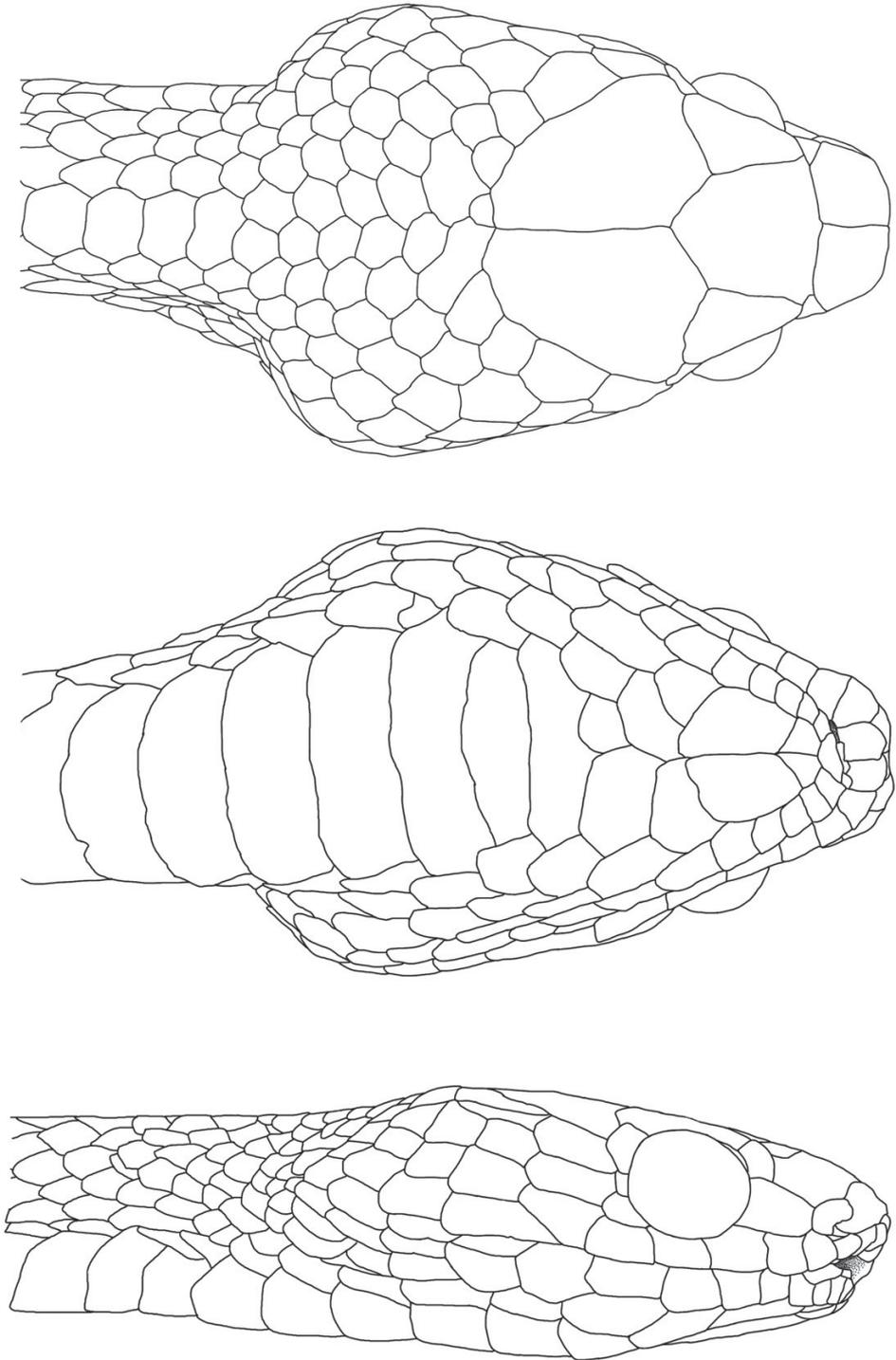


Figure 7. Illustration of the head scales of a *Dipsas aparatiritos* sp. nov. (USNM 579810). Drawings by Shannon Bowley Christensen.

Table 1. Measurements of body and head of *Dipsas aparatiritos* to nearest mm. * = Holotype, ** = Paratype.

| Catalogue number | Sex | Svl (mm) | Tail length (mm) | Vertebral scale width (mm) | Dorsal scale width (mm) | Eye length (mm) | Rostral to eye length (mm) | Head width (mm) | Head length (mm) |
|------------------|-----|----------|------------------|----------------------------|-------------------------|-----------------|----------------------------|-----------------|------------------|
| USNM 579820 | F | 169 | 65 | 1.11 | 0.82 | 1.98 | 2.19 | 4.27 | 8.90 |
| USNM 579822 | F | 197 | 95 | 1.17 | 0.78 | 2.10 | 1.93 | 4.29 | 8.83 |
| USNM 579827 | F | 205 | | 1.35 | 0.99 | 2.19 | 1.93 | 4.52 | 8.80 |
| USNM 579826 | F | 242 | 124 | 1.13 | 1.04 | 2.18 | 2.43 | 4.86 | 9.75 |
| USNM 579813 | F | 265 | 133 | 1.17 | 1.25 | 2.38 | 2.25 | 4.75 | 9.78 |
| USNM 579825 | F | 312 | 175 | 1.53 | 1.51 | 2.51 | 2.69 | 4.91 | 11.85 |
| USNM 579808 | F | 319 | 162 | 1.34 | 1.46 | 2.34 | 2.71 | 5.00 | 11.18 |
| USNM 579829** | F | 328 | 170 | 2.19 | 1.75 | 2.34 | 2.96 | 6.61 | 12.28 |
| USNM 579824 | F | 333 | 179 | 1.86 | 1.70 | 2.38 | 2.51 | 5.09 | 11.49 |
| USNM 579807 | F | 346 | 165 | 1.64 | 1.82 | 2.26 | 2.99 | 5.63 | 12.68 |
| USNM 579823 | F | 357 | 189 | 1.94 | 1.82 | 2.54 | 2.81 | 5.73 | 11.98 |
| USNM 579810 | F | 395 | 219 | 2.19 | 1.65 | 2.60 | 2.97 | 5.97 | 13.80 |
| USNM 579814 | F | 400 | 221 | 2.06 | 2.03 | 2.40 | 3.54 | 5.81 | 13.32 |
| USNM 579809 | F | 420 | 221 | 1.73 | 2.40 | 2.58 | 3.11 | 6.29 | 14.00 |
| USNM 579828* | F | 424 | 211 | 2.70 | 2.22 | 2.56 | 3.31 | 7.35 | 13.26 |
| USNM 579816 | M | 310 | 122 | 1.12 | 1.33 | 2.47 | 2.39 | 5.16 | 10.57 |
| USNM 579815 | M | 415 | 244 | 1.72 | 1.83 | 2.64 | 3.14 | 5.91 | 12.75 |
| USNM 579812 | M | 420 | 236 | 1.63 | 1.77 | 2.88 | 3.00 | 5.81 | 12.94 |
| USNM 579819 | M | 450 | 251 | 1.83 | 2.15 | 2.83 | 3.44 | 5.86 | 12.06 |
| USNM 579811 | M | 465 | 260 | 1.90 | 1.69 | 3.05 | 3.30 | 6.05 | 13.77 |
| USNM 579817 | M | 465 | 241 | 2.18 | 2.11 | 2.70 | 3.27 | 5.91 | 13.17 |

**Figure 8.** Hemipenes of *Dipsas aparatiritos* sp. nov. USNM 579815 **a** sulcate face **b, d** lateral faces **c** asulcate face. Photographs by James Poindexter.

Dipsas aparatiritos sp. nov. differs from the majority of its congeners by having the nasal scale fused with the first supralabial, anterior infralabials separated by a pair of (rarely fused) small postmentals, and temporals usually entering the orbit.

Table 2. Scale counts for dorsals, ventrals, labials and loreals, along with dorsal blotch counts for a series of 31 *Dipsas aparatiritos* sp. nov. collected from Parque Nacional General de Division Omar Torrijos Herrera. Also included are available data for specimens collected at other sites. s = single; w = wide loreal; co = contacting the orbit; irr l = irregular loreal. *holotype and **paratype

| Catalogue number | Sex | Dorsal scale rows | Ventrals | Sub-caudals | Anal plate | Right supralabials | Left supralabials | Right infralabials | Infralabials contact behind mental | Left infra-labials | Right loreal | Left loreal | Dorsal blotches |
|------------------|-----|-------------------|----------|-------------|------------|--------------------|-------------------|--------------------|------------------------------------|--------------------|--------------|-------------|-----------------|
| SMF 89554 | – | 15-15-15 | | 116 | s | 7(4-6) | | 8 | | | | | |
| SMF 89769 | Juv | 15-15-15 | 181 | 181 | s | 7(4-5) | | 9 | | | | | |
| SMF 89953 | Juv | 15-15-15 | 175 | 110 | s | 7(4-5) | | 8 | | | | | |
| MHCH 3123 | F | 15-15-15 | 194 | 126 | s | 7(4-5) | | 9(2-5) | | | | | |
| USNM 579820 | F | 15-15-15 | | | s | 7-4.5 | 9-5.6 | 9 | 0 | 8 | w co | w co | 20 |
| USNM 579822 | F | 15-15-15 | | 131 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 17 |
| USNM 579827 | F | 15-15-15 | | | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 18 |
| USNM 579826 | F | 15-15-15 | 197 | 129 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 18 |
| USNM 579813 | F | 15-15-15 | 188 | 125 | s | 7-4.5 | 7-4.5 | 8 | 0 | 8 | w co | w co | 20 |
| USNM 579825 | F | 15-15-15 | 177 | 119 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 19 |
| USNM 579808 | F | 15-15-15 | 184 | 118 | s | 7-4.5 | 7-4.5 | 8 | 0 | 8 | w co | w co | 18 |
| USNM 579829** | F | 15-15-15 | 183 | 124 | s | 8-4.5 | 8-4.5 | 9 | 0 | 9 | w co | w co | 19 |
| USNM 579824 | F | 15-15-15 | | 121 | s | 7-4.5.6 | 7-4.5.8 | 8 | 0 | 8 | w co | w co | 19 |
| USNM 579807 | F | 15-15-15 | 178 | 111 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 18 |
| USNM 579823 | F | 15-15-15 | 185 | 122 | s | 7-4.5 | 7-4.5 | 9 | 0 | 8 | w co | w co | 19 |
| USNM 579810 | F | 15-15-15 | 182 | 116 | s | 7-4.5 | 6-4.5 | 8 | 0 | 8 | w co | w co | |
| USNM 579814 | F | 15-15-15 | 182 | 124 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 17 |
| USNM 579809 | F | 15-15-15 | 180 | 118 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 18 |
| USNM 579828* | F | 15-15-15 | 178 | 118 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 16 |
| USNM 579816 | M | 15-15-15 | 192 | | s | 7-4.5 | 7-4.5 | 9 | 0 | | w co | w co | 17 |
| USNM 579815 | M | 15-15-15 | 196 | 135 | s | 7-4.5 | 8-5.6 | 9 | 0 | 9 | w co | w co | 17 |
| USNM 579812 | M | 15-15-15 | 195 | 131 | s | 7-4.5 | 7-4.5 | 10 | 0 | 9 | w co | w co | 19 |
| USNM 579819 | M | 15-15-15 | 194 | 136 | s | 7-4.5 | 7-4.5 | 8 | 0 | 8 | irr l | irr l co | 19 |
| USNM 579811 | M | 15-15-15 | 195 | 129 | s | 7-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 18 |
| USNM 579817 | M | 15-15-15 | 191 | | s | 8-4.5 | 7-4.5 | 9 | 0 | 9 | w co | w co | 19 |
| MHCH 2311 | M | 15-15-15 | 194 | 130 | s | 7(4-5) | | 9(3-6) | | | | | |
| SMF 89551 | M | 15-15-15 | 191 | 130 | s | 7(4-5) | | 9 | | | | | |
| SMF 89552 | M | 15-15-15 | 192 | 122 | s | 7(4-5) | | 9/8 | | | | | |
| SMF 89553 | M | 15-15-15 | 190 | 130 | s | 7(4-5) | | 9 | | | | | |
| SMF 90036 | M | 15-15-15 | 192 | 122 | s | 7(4-5) | | 9/8 | | | | | |
| SMF 97346 | M | 15-15-15 | 192 | | s | – | | 9 | | | | | |

Dipsas aparatiritos sp. nov. shares with the other Central American species of the genus the number of dorsal scales rows (15-15-15), except with *D. gaigeae* Oliver (13-13-13); number of temporals (1+2+ 2); absence of preoculars, except *D. brevifacies* Cope (1, 2 or 3); and number of postoculars (2,3), except *D. temporalis* Werner (3,4). The number of infralabials (9–10) is in the range of all Panamanian species, but the infralabial scales in contact behind mental (0) differs from all species, except with *D. temporalis*. The number of supralabials (7–8) is within the variation found in *D. gaigeae* (7–8), *D. nicholsi* (7–9), *D. temporalis* (6–8), and *D. tenuissima* Taylor (8), but differs from *D. articulata* Cope, *D. bicolor* Günther, *D. brevifacies*, and *D. viguieri* Bocourt (9–10); the supralabials scales in contact with the eye (4–5) also

Table 3. Scale counts related to the ocular region of the series of 31 *Dipsas aparatiritos* sp. nov. specimens. upp = upper; low = lower. * = holotype, ** = paratype.

| Catalogue number | Sex | Right preocular | Left Preocular | Right presubocular | Left presubocular | Right postocular | Left postocular | Right post-subocular | Left post-subocular |
|------------------|-----|-----------------|----------------|--------------------|-------------------|------------------|-----------------|----------------------|---------------------|
| SMF 89554 | – | | | | | 2 | | | |
| SMF 89769 | Juv | | | | | 3 | | | |
| SMF 89953 | Juv | | | | | 2 | | | |
| MHCH 3123 | F | | | | | 2 | | | |
| USNM 579820 | F | 1 upp | 1 upp | 0 | 0 | 3 | 3 | 0 | 0 |
| USNM 579822 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579827 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579826 | F | 1 upp | 1 upp | 0 | 0 | 3 | 3 | 0 | 0 |
| USNM 579813 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579825 | F | 1 upp | 1 upp | 0 | 0 | 3 | 3 | 0 | 0 |
| USNM 579808 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579829** | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579824 | F | 1 upp | 1 upp | 0 | 0 | 1 upp/ 1 low | 1 upp/ 1 low | 0 | 0 |
| USNM 579807 | F | 0 | 0 | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579823 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579810 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579814 | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579809 | F | 1 upp | 1 upp | 0 | 0 | 2 | 1 upp/ 1 low | 0 | 0 |
| USNM 579828* | F | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579816 | M | 1 upp | 1 upp | 0 | 0 | 3 | 3 | 0 | 0 |
| USNM 579815 | M | 1 upp/ 1 low | 1 upp/ 1 low | 0 | 0 | 3 | 3 | 0 | 0 |
| USNM 579812 | M | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579819 | M | 2 | 1 upp/ 1 low | 0 | 0 | 3 | 2 | 0 | 0 |
| USNM 579811 | M | 1 upp | 1 upp | 0 | 0 | 2 | 2 | 0 | 0 |
| USNM 579817 | M | 1 upp | 1 upp | 0 | 0 | 2 | 1 upp/ 1 low | 0 | 0 |
| MHCH 2311 | M | | | | | 4 | | | |
| SMF 89551 | M | | | | | 2 | | | |
| SMF 89552 | M | | | | | 3 | | | |
| SMF 89553 | M | | | | | 3 | | | |
| SMF 90036 | M | | | | | 3 | | | |
| SMF 97346 | M | | | | | 3 | | | |

are in the variation found in the other species (Table 4). The vertebral row is enlarged moderately as in *D. nicholsi* and *D. temporalis*, and it different from the other species where it is scarcely enlarged. The number of ventral scales of males and females of *Dipsas aparatiritos* sp. nov. is larger than *D. brevifacies* and *D. gaigeae* and fewer than *D. articulata*, *D. tenuissima* and the males of *D. temporalis*, while overlapping with *D. bicolor*, *D. nicholsi*, *D. viguieri*, and the females of *D. temporalis* (Table 2). The number of subcaudal scales of males and females is larger than *D. brevifacies*, *D. gaigeae*, *D. nicholsi*, and *D. tenuissima*, while overlapping with *D. articulata*, *D. bicolor*, *D. temporalis*, and *D. viguieri* (Table 4).

The new species is sister to *Dipsas temporalis*, from which it differs on the following characters of coloration and lepidosis. In *D. aparatiritos* sp. nov., the first dorsal band extends far onto the ventrals (restricted to the dorsum or barely entering ventrals in *D. temporalis*) and the posterior body bands form elliptical blotches usually broken along the vertebral line (bands complete over dorsum or elliptical blotches joined along

Table 4. Scale counts, measurements and degree of enlargement of the vertebral row of the species of *Dipsas* known to occur in Central America, combining data from the examined specimens listed in Appendix 1 and from references listed in Materials and methods. The values of the ventral and subcaudal counts are minimum and maximum.

| | <i>D. articulata</i> | <i>D. bicolor</i> | <i>D. breviplicata</i> | <i>D. gaigeae</i> | <i>D. nicholsi</i> | <i>D. aparatiritos</i> | <i>D. temporalis</i> | <i>D. tenuissima</i> | <i>D. viguieri</i> |
|-------------------------------------|------------------------|------------------------|------------------------|------------------------|-------------------------------|------------------------|-------------------------------|----------------------|------------------------|
| Dorsals | 15-15-15 | 15-15-15 | 15-15-15 | 13-13-13 | 15-15-15 | 15-15-15 | 15-15-15 | 15-15-15 | 15-15-15 |
| Ventrals | M 198–217 F 195–210 | M 195–199 F 185–199 | M 167–181 F 166–174 | M 162–166 F 163–167 | M 192–210 F 186–201 | M 190–196F 177–197 | M 197–208 F 184–192 | M 225 F 227 | M 196–211 F 190–206 |
| Subcaudals | M 115–135 F 108–118 | M 129–132 F 111–129 | M 71–102 F 69–87 | M 64–72 F 53–62 | M 81–100 F 84–97 | M 122–136F 111–131 | M 120–132 F 120–123 | M 99 Fno data | M 113–129 F 102–126 |
| Preoculars | 0 | 0 | 1, 2, 3 | 0 | 0 | 0 | 0 | 0 | 0 |
| Postoculars | 2–3 | 2–3 | 3 | 2 | 2 | 2–3 | 3–4 | 3 | 2–3 |
| Supralabials | 9–10 | 10 | 9–10 | 7–8 | 7–9 | 7–8 | 6–8 | 8 | 9–10 |
| Supralabials contacting eye | [4,5] [5,6] | [4,5,6,7] | [4,5] | [3,4] | [3,4] [4,5] | [4,5] | [3,4] [4,5] | [4,5] | [4,5] [5,6] |
| Infralabials | 10–13 | 10–12 | 9–13 | 7–9 | 10–13 | 9–10 | 8–13 | 9–10 | 9–12 |
| Infralabials in contact | [1,1] | [1,1] | [2,2] | [1,1] | [1,1] [2,2] | 0 | 0 | [1,1] | [1,1] |
| Temporals | 2+3+4 | 1+2+3 | 2+3+4 | 2+3+4 | 2+3+4 | | 2+3+4 | 2+3+3 | 2+3+4 |
| TOL of largest specimen (mm) | M 715 F 655 | M no data F 627 | M 596 F 536 | M 652 F 726 | M 861 F 798 | M 725 F 713 | M 697 F 645 | M 554 F 572 | M 719 F 547 |
| Vertebral row | Scarcely enlarged | Scarcely enlarged | Scarcely enlarged | Not enlarged | Moderately to broadlyenlarged | Moderately enlarged | Moderately to broadlyenlarged | Scarcely enlarged | Scarcely enlarged |
| TL/TOL | M 32% F 31% | M 33% F% F 26% | M 30% F 26% | M 23% F 28% | M 25% F 24% | M 35% F 34% | M 38% F 33% | M 29%F no data | M 33%F 30% |

the vertebral line in *D. temporalis*). The color of the anterior interspaces is white or bright pale yellow in *D. aparatiritos* sp. nov. and pale brown in *D. temporalis*. Overall, *D. temporalis* compared to *D. aparatiritos* sp. nov. have a greater number of ventral scales in males ($\bar{x} = 198$) vs. ($\bar{x} = 192$) and females ($\bar{x} = 192$) vs. ($\bar{x} = 184$) respectively, although there is overlap in the counts (Table 5, Fig. 10).

Etymology. The species name is an adjective formed from the Greek word *aparatiritos* (απαρατηρητος), which means unnoticed. The snake has hidden in plain sight for more than forty years at a very well-studied field site for herpetological research. We suggest the common name “Hidden Snail-eater” (“Caracolera Escondida” in Spanish).

Distribution. *Dipsas aparatiritos* sp. nov. is found in both the Atlantic and Pacific slopes of the Cordillera Central in western Panama, with an additional population on the Parque Nacional Chagres. The species occurs over an estimated 9,630 km² area and has been recorded at elevations 597–1002 m above sea level, which makes it the most wide-spread species of *Dipsas* in Panama. A series of individuals were collected from PNGDOTH. This is a mid-elevation, premontane cloud-forest with mature secondary forest and many streams branching from Río Guabal (McCaffery and Lips 2013). The mean annual rainfall is 3500 mm and mean annual temperature range is 19–31 °C (Lips et al. 2006). Two localities (Donoso, Colón province, and Quebrada Las Tres

Table 5. Differences in coloration, scale counts and size between *Dipsas temporalis* and *D. aparatiritos* sp. nov. The range of each continuous variable is from our own sample, Harvey (2008), and Lotzkat (2015). The numbers in parentheses represent the sample size.

| Variable | <i>Dipsas temporalis</i> | | <i>Dipsas aparatiritos</i> sp. nov. | |
|---|---|----------------------------|---|-----------------------------|
| First dorsal band extends far onto the ventrals | No | | Yes | |
| Condition of posterior body bands | Complete over dorsum or elliptical blotches joined along the vertebral line | | Forming elliptical blotches usually broken along the vertebral line | |
| Color of anterior interspaces | Pale brown | | White or bright pale yellow | |
| Infralabials | 8–9 | | 9–10 | |
| Sex | Males (<i>n</i> = 5) | Females (<i>n</i> = 8) | Males (<i>n</i> = 12) | Females (<i>n</i> = 16) |
| Maximum TOL | 694 mm | 630 mm | 688 mm | 713 mm |
| Ventral scales | 183–210 | 184–203 | 177–197 | 190–196 |
| Subcaudal scales | 112–132 | 111–134 | 122–136 | 111–131 |

Honeras, Panama province) are in valleys 134–197 m above sea level. Since these localities are much lower in elevation than all other reported localities, it is likely that the specimens collected there (SMF 97346 and MCZ 50214) were actually found in the neighboring mountain ridges (Fig. 9).

Natural history notes. The holotype was encountered at 21:58 h in mature secondary (40+ years) premontane forest on the Atlantic versant, but only ca. 100 m from the Continental Divide. The trail is known as “the old logging road” as described by Myers et al. (2007). The Tropical Amphibian Declines in Streams (TADS) project, which has been working in the area since 1997, refers to the trail as “Rocky Road,” while the park calls it “La Salida” to Sendero La Rana. The snake was elongate and crawling on small tree 0.75 m off the ground. The paratype was encountered at 21:59 h in mature secondary (40+ years) premontane forest on the Atlantic versant, but only ca. 100 m from the Continental Divide on the same trail as the holotype. The snake was elongate and crawling on small tree 0.75 m off the ground. Lotzkat (2015) found specimens of *Dipsas aparatiritos* sp. nov. foraging at night on vegetation 30–200 cm above the ground. JMR found this species to be more common in forest and along streams rather than around ponds. In PNGDOTH, JMR examined the fecal samples of this species and found that one (2% of the sample) contained the operculum of a snail and 49 (98%) contained oligochaete chaetae.

Despite being a new species, it is relatively common at the PNGDOTH site and has been documented for years, thus providing much data on the natural history. Specimens have been found in vegetation, at times over one meter in height, but at other times just centimeters off the ground where it blended in well with leaf litter, as proven by one individual found on the ground (Fig. 2). Gravid females were found in all months except February, March, October, and December with the highest frequency in June and July (JMR pers. obs.). Females had either one or two ova. Breeding events were not observed, although one night four different *Dipsas aparatiritos* sp. nov. were observed intertwined on a single branch (Fig. 11).

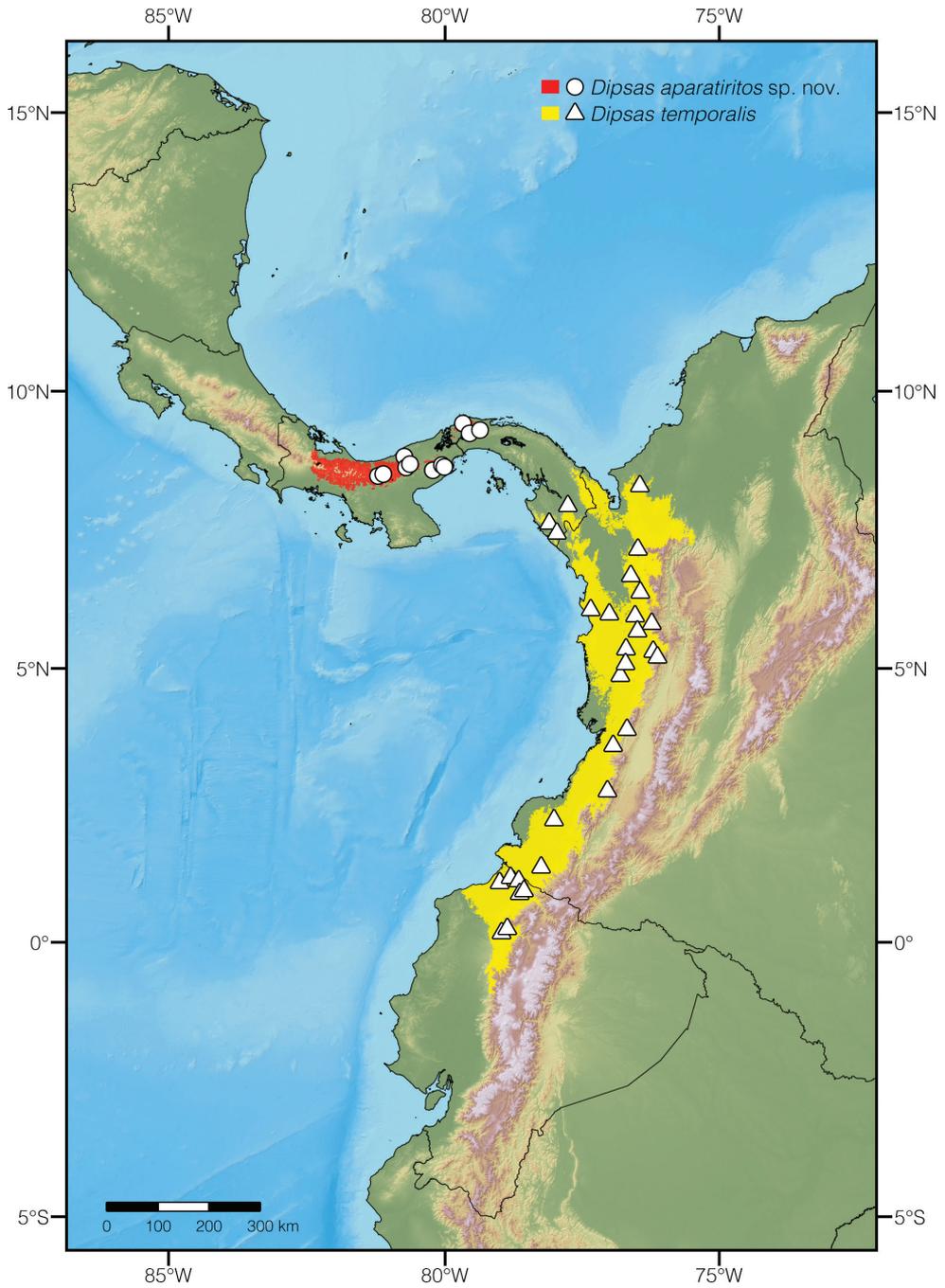


Figure 9. Map of locality data for *Dipsas aparatiritos* sp. nov. (red showing range, circles marking specimens included in this paper) and updated range data for *D. temporalis* (yellow showing range, triangles marking specimens included in this paper).



Figure 10. Photographs of species of *Dipsas* previously subsumed under *D. temporalis* **a** *D. aparatiritos* sp. nov. from Cerro Gaital, Antón, Coclé province, Panama **b** *D. temporalis* ZSFQ 5063 from Durango, Esmeraldas province, Ecuador **c** *D. temporalis* ZSFQ 5062 from Durango, Esmeraldas province, Ecuador.



Figure 11. Four individuals of *Dipsas aparatiritos* sp. nov. intertwined on one plant at Parque Nacional General de División Omar Torrijos Herrera. Photograph by Noah Carl.

Near the area where the holotype of *Dipsas aparatiritos* sp. nov. was found in PNGDOTH, JMR has recorded the following species of amphibians and reptiles: salamanders including *Oedipina collaris* (Stejneger, 1907), and *Bolitoglossa colonnea* (Dunn, 1924), frogs, including *Diasporus diastema* (Cope, 1875), *Espadarana prosoblepon* (Boettger, 1892), lizards including *Anolis humilis* Peters, 1863, and *Enyalioides heterolepis* (Bocourt, 1874), and snakes including *Bothrops asper* (Garman, 1883), *Bothriechis schlegelii* (Berthold, 1846), *D. nicholsi*, *Imantodes cenchoa* (Linnaeus, 1758), *Oxybelis brevirostris* (Cope, 1861), *Sibon annulatus*, and *S. nebulatus* (Linnaeus, 1758).

Conservation. We consider *Dipsas aparatiritos* sp. nov. to be included in the Near Threatened category following the IUCN Red List categories and criteria, v. 3.1, second edition (IUCN 2012) because, although the species' estimated extent of occurrence is less than 10,000 km² and nearly 44% of this area has already been deforested (CATHALAC 2011), the species occurs in at least four major national parks (Santa Fe, PNGDOTH, Altos de Campana, and Chagres) and satellite images show that there is forest connectivity between populations. At PNGDOTH, the occurrence rate of *D. aparatiritos* sp. nov. has actually increased by a factor of three in the period between 2006 and 2012 (Zipkin et al. 2020). Also, the body condition of the individuals in this locality increased following the collapse of amphibian populations due to chytridiomycosis (Zipkin et al. 2020). However, the causes for these changes are enigmatic given that amphibians presumably

do not comprise an important part of the diet of this species. The status and trend of other populations should be evaluated carefully given that *D. aparatiritos* sp. nov. is endemic to Panama and probably highly dependent on old-growth forests.

Other *Dipsas* species at the site. In addition to the new species there are two other species of *Dipsas* known from the site: *Dipsas nicholsi* (Myers et al. 2007 [see edit to proof]) and *Dipsas articulata* (Vecchiet et al. 2014). This adds one more confirmed species, bringing the total to three. Furthermore, also known to occur at the site are at least four species of *Sibon* (*S. argus*, *S. canopy*, *S. longifrenis*, and *S. nebulatus*), which are closely related phylogenetically (Peters 1960; Sheehy 2012) and ecologically (Ray et al. 2011). *Sibon lamari* also may be present at the site (JMR unpubl. data). *Dipsas aparatiritos* sp. nov. was found throughout the general survey area, both on metered-transects and within the adjacent forest between transects.

Key to Central American *Dipsas*

- 1 Dorsals 13-13-13, loreal longer than high contacting the orbit; preoculars absent; seven supralabials, third and fourth contacting the orbit; 7 or 8 infralabials, one pair in contact behind the mental; vertebral scale not enlarged; ventrals M 162–166, F 163–167; subcaudals M 64–72, F 53–62 *Dipsas gaigeae*
- Dorsals 15-15-15 **2**
- 2 Ventrals > 220; square loreal contacting the orbit; one preocular; eight supralabials, fourth and fifth contacting the orbit; 9 or 10 infralabials, one pair in contact behind the mental; vertebral scale slightly enlarged; ventrals M 225, F 227; subcaudals M 99 *Dipsas tenuissima*
- Ventrals < 220 **3**
- 3 Black horseshoe pattern present on the dorsum of head; irregular or square-shaped loreal contacting the orbit; preoculars absent; 8 or 9 supralabials, fourth and fifth contacting the orbit; 12 infralabials, one pair in contact behind the mental; vertebral scale slightly enlarged; ventrals M 192–210, F 186–201; subcaudals M 81–100, F 84–97; beige with dark brown saddles *Dipsas nicholsi*
- Lack of black horseshoe pattern on the dorsum of head; typically, dark brown alternating with paler brown or tan; white outline may be present **4**
- 4 Alternating brown with pale beige or white with rose/pink/red on white spots of dorsum **5**
- Lacking rose/pink/red on white spots of dorsum **6**
- 5 Single chin shields; irregular or square shape loreal contacting the orbit; one preocular; 10 supralabials, fourth, fifth, and sixth contacting the orbit; 11 or 12 infralabials, one pair in contact behind the mental; vertebral scale not enlarged; ventrals M 195–199, F 185–199; subcaudals M 129–132, F 111–129 *Dipsas bicolor*
- Paired chin shields; loreal longer than high or square loreal contacting the orbit; preoculars absent; eight supralabials, fourth and fifth contacting the orbit; 10 or 11 infralabials, one pair in contact behind the mental; vertebral scale slightly enlarged; ventrals M 198–217, F 195–210; subcaudals M 115–135, F 108–118 *Dipsas articulata*

- 6 Supralabials 6–8.....7
 – Supralabials 9.....8
 7 Loreal longer than high contacting the orbit; one preocular; seven or six supralabials, fourth and fifth or third and fourth contacting the orbit; 8–10 infralabials, none in contact behind the mental; vertebral scales slightly enlarged; ventrals M 197–208, F 184–200; subcaudals M 120–132, F 120–123 *Dipsas temporalis*
 – Loreal longer than high contacting the orbit; one preocular; 7 or 8 supralabials, fourth and fifth contacting the orbit; 9 or 10 infralabials, none in contact behind the mental; vertebral scales moderately enlarged; ventrals M 191–196, F 177–197; subcaudals M 129–136, F 111–131, Head pale brown *Dipsas aparatiritos*
 8 Irregular or square shape loreal contacting the orbit; preoculars absent; 9 supralabials, fourth and fifth or sixth contacting the orbit; 9–11 infralabials, one pair in contact behind the mental; vertebral scales slightly enlarged; ventrals M 196–211, F 190–206; subcaudals M 113–129, F 102–126; Head reddish-brown *Dipsas viguieri*
 – Loreal longer than high contacting the orbit; preoculars one; nine supralabials, fourth and fifth contacting the orbit; 10–12 infralabials, two pairs in contact behind the mental; vertebral scale slight enlarged; ventrals M 167–181, F 166–174; subcaudals M 71–102, F 69–87 *Dipsas brevifacies*

Discussion

In the past decade, a significant number of species have been added to the fauna of Panama, either as range extensions across political borders or as newly described species to science. The former includes *Ninia sebae* (Duméril, Bibron, & Duméril, 1854) and *Porthidium volcanicum* Solórzano, 1995 in the western part of the country, and *Leptophis cupreus* (Cope, 1868) (Batista and Wilson 2017) and *Micrurus dumerilii* Jan, 1858 (Prairie et al. 2015) in the east. The latter includes dipsadine species such as *Sibon perissostichon* (Köhler et al. 2010) and *S. noalamina* (Lotzkat et al. 2012), along with the colubrine *Tantilla berguido* (Batista et al. 2016). Additionally, the number of the very rare *Geophis bellus* Myers, 2003 (Dipsadinae) specimens has increased significantly (Lara et al. 2015 and an additional, complete specimen of *Atractus imperfectus* Myers, 2003 (Dipsadinae) was found (Ray 2017). According to our assessment, the range of *Dipsas temporalis* in Panama has been reduced to the eastern portion of the Darien. However, this species is still currently found in Panama.

Interestingly, *Dipsas aparatiritos* sp. nov. has been known at the PNGDOTH site since the late 1970s when Charles Myers visited and mentioned the potential presence of at least one new species of *Dipsas*. Given how similar it is to the previously documented *D. temporalis*, and that the very rare *D. nicholsi* also was found in this remote area, suggests that other species of *Dipsas* may be found in other isolated, mountainous areas around the country. There is a need for continued research, especially in remote areas, to fully document the serpent fauna of Panama.

Dipsas aparatiritos sp. nov. is sister to *D. temporalis*. We have decided to name in our phylogeny the specimen MHUA 14278 as *D. temporalis* following the work of Sheehy (2012) and Arteaga et al. (2018) and to not follow the suggestion by Barros et al. (2012) of identifying the sample as *D. sanctijoannis*. The sample in question was identified before as *D. pratti* (Daza et al. 2009, see GenBank) but Sheehy (2012) incorporated samples of *D. pratti* from the type locality and Venezuela, which clearly represents a different species. Sheehy presents the sample in question as *D. temporalis*. Later, Arteaga et al. (2018) presented a near topotypic sequence of *D. temporalis*, QCAZR5050, from San Lorenzo, Esmeraldas, 866 m. This near topotypic sequence forms a tight clade with the sample in question, MHUA 14278, in their phylogeny. *Dipsas temporalis* is typically a lowland Chocoan species inhabiting from Ecuador to Panama, below 100 m elevation. *Dipsas pratti* is a highland Andean species inhabiting the Cordillera Central and the Cordillera Oriental of Colombia and Venezuela, as shown by Barros et al. (2012). *Dipsas sanctijoannis* is a highland species distributed along the Cordillera Occidental and Cordillera Central of Colombia, and known from elevations between 1585 and 2400 (Boulenger 1911; Harvey et al. 2008). The lowest record of *D. sanctijoannis* that we know about is the type, from the town of Pueblo Rico, above the Río San Juan, near the Risaralda-Choco border at 1585 m (Boulenger 1911). Harvey et al. (2008) reports on a specimen of *D. temporalis* from near the type locality and along the San Juan drainage but, from much lower elevation, ca 60 m elevation (USNM 267244). This specimen is less than 90 km away from the type locality of *D. sanctijoannis*. The specimen MHUA 14278, originates from the lowlands of the northwestern branch of the Department of Antioquia, at 233 m elevation. Both the previous phylogenetic analyses and the lowland affinity of *D. temporalis* as compared to the highland *D. pratti* and *D. sanctijoannis* support our taxonomic decision.

Despite being a newly described species, *Dipsas aparatiritos* sp. nov. is quite common at the type locality. Fortunately, this area is a protected national park. Regardless, during the ten years JMR spent studying at the site, there was a reduction in number of park rangers (already very few for such a large, protected area), and there was a decline in the care of the trails near the ranger station. The site was logged in the past and unpermitted collection of rare butterflies was observed at the site, suggesting that other unpermitted collectors could arrive in the future. In 2015, the community began to pave the road leading into the park in an effort to pave to the town of La Rica inside the park boundaries. This advancement will greatly increase the ease with which tourists and poachers alike are able to reach the site. In the past, the site was only accessible with high-clearance four-wheel-drive vehicles. Finally, chytridiomycosis reached the site in 2004, but Ray et al. (2012) showed that *D. aparatiritos* sp. nov. (*Dipsas* sp. in that publication) feeds primarily on oligochaetes. There may be a desire of horticulturists and invertebrate enthusiasts to collect bromeliads where both the bromeligenous oligochaetes and *D. aparatiritos* sp. nov. spend considerable time. It is hoped that the area will remain protected and *D. aparatiritos* sp. nov. can continue to thrive.

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Appendix I

Specimens examined

The numbers with an asterisk (*) correspond to holotypes.

Dipsas aparatiritos sp. nov. **Panama**, Coclé, Donoso: SMF 97346; El Copé, ca. 5.5 km N of Parque Nacional General de División Omar Torrijos Herrera: USNM 579807–579829; Panamá Oeste, Cerro Campana: MHCH 3123; Veraguas, Cerro Mariposa: MHCH 2311, SMF 89551–53, SMF 89953–54; Cerro Negro: SMF 89769, SMF 90036.

Dipsas articulata. **Nicaragua**, Río San Juan, Elev. 13m, Río Indio Lodge: MVZ 269222–269223; **Panama**, Bocas del Toro: AMNH 124125, Cocuyas de Veragua: ANSP 10113*; Isla Bastimentos, Old Point: USNM 297917; Laguna de Tierra Oscura, 3.7 km S of Tiger Key: USNM 348490–348491; **Costa Rica**, Limón, La Castilla, Lower Reventazon: ANSP 22380; Pandora: UMMZ 125236.

Dipsas bicolor. **Honduras**, Gracias a Dios, Bachi Kiamp: USNM 578015; Cabeceras de Río Rus Rus: USNM 559619; Urus Tingni Kiamp: USNM 561921; Warunta Tingni Kiamp: USNM 561922; Olancho, Nueva Esperanza: USNM 559618.

Dipsas brevifacies. **Mexico**, Yucatán: FMNH 20634, 36397, 36401, 36406, USNM 6562; Citilpeck: ANSP 10129.

Dipsas gaigeae. **Mexico**, Colima: AMNH 82017; Colima: USNM 160938; Hacienda Paso del Rio/Periquillo: UMMZ 80221*; Jalisco, Barra de Navidad: USNM 196499.

Dipsas nicholsi. **Panama**, Canal Zone, Madden Forest Preserve: KU 110310–314; Coclé, Parque Nacional General de División Omar Torrijos Herrera: USNM 579806; Panamá: FMNH 217310, Chagres Village: ANSP 21907.

Dipsas temporalis. **Colombia**, Antioquia, unknown: UV-C 5388; Choco, Agua Clara, Río Tamana: USNM 267244; Chocó, Condoto: NHMUK; N slope Alto de Buey: LACM 72747; Valle de Cauca, Tamboral: CPZ-UV 04568; Valle del Cauca, Quebrada la Batea: CZI-R 080; **Panama**, Comarca Emberá-Wounaan, Serranía de Jingurudo: MHCH 2878; Darién, S slope Cerro Cituro, Serranía de Pirre: KU 110301, 110303–304, 110307–309; Ridge between Río Jaqué and Río Imamadó: KU 110295; Panamá, S slope Cerro La Campana: KU 110293; Rancho Frío Field Station: MHCH 2881. **Ecuador**, Esmeraldas, Durango: ZSFQ 5062 and ZSFQ 5063; Junto al Río Chuchubí: QCAZ 5050; 16 km W of Lita: MHNG 2521.083; Tundaloma Lodge: MZUTI 3331.

Dipsas tenuissima. **Costa Rica**, San José, 15 mi NW San Isidro del General: KU 31961*; **Panama**, Chiriquí, Pto. Armuelles: ANSP 24255; Panama Isthmus, MZUSP 2049.

Dipsas vignieri. **Colombia**, Chocó: FMNH 74376; **Panama**, Darién: AMNH 36200; Río Tuira at Río Mono: KU 110316; Canal Zone, Madden Forest Preserve: UF 44291, KU 110317; Madden Forest Road, 2.0 mi S. Trans Isthmus Highway: UF 44290; Pipeline Road: UMMZ 155717.

Appendix 2

Table A1. Primers used in this study, gene, name, direction, sequence (5'–3' direction), and reference.

| Primers | | | | Reference |
|---------|----------|-----|----------------------------------|-----------------------------|
| cyt-b | S20596F | (F) | AACCACTCTTGTTAATCAACTACA | Ingrasci 2011 |
| cyt-b | S21790R | (R) | ACCCATGTTTGGTTTACAAAAACAATGCT | Ingrasci 2011 |
| cyt-b | GLUDG | (F) | TGACTTGAARAACCAAYCGTTG | Parkinson et al. 2002 |
| cyt-b | AtrCB3 | (R) | TGAGAAGTTTTCYGGGTGRTT | Parkinson et al. 2002 |
| ND4 | ND4 | (F) | CACCTATGACTACCAAAAGCTCATGTAGAAGC | Arévalo et al. 1994 |
| ND4 | LEU | (R) | CATTACTTTTACTTGGATTGACACCA | Arévalo et al. 1994 |
| ND4 | 605F | (F) | GTCTCCATCTATGACTCCCA | Ingrasci 2011 |
| ND4 | L68R | (R) | TACCACTTGGATTGACACCA | Ingrasci 2011 |
| NT3 | NT3-F3 | (F) | ATATTTCTGGCTTTTCTCTGTGGC | Noonan and Chippindale 2006 |
| NT3 | NT3-R4 | (R) | GCGTTTCATAAAAAATATTGTTTGACCGG | Noonan and Chippindale 2006 |
| DNAH3 | DNAH3-fl | (F) | GGTAAATGATAGAAGAYTACTG | Townsend et al. 2008 |
| DNAH3 | DNAH3-r6 | (R) | CTKGAGTTRGAHACAATKATGCCAT | Townsend et al. 2008 |

Appendix 3

Table A2. Specimens used in genetic analyses and respective GenBank numbers.

| Taxa | Voucher museum number | Field or tissue number | Locality | ND4 | cyt-b | NT3 | DNAH3 |
|------------------------|-----------------------------|--------------------------|--------------------------|----------|----------|----------|----------|
| <i>D. andiana</i> | Bioparque Amaru RSCDSP 0389 | JM 79 (J. M. Daza) | Ecuador: Los Ríos | JX398453 | JX398607 | JX398744 | JX293843 |
| <i>D. aparatiritos</i> | USMN 579815 | JM 664 | Panama: Coclé | JX398476 | JX398626 | | |
| <i>D. aparatiritos</i> | USMN 579814 | JM 663 | Panama: Coclé | JX398475 | JX398625 | | |
| <i>D. aparatiritos</i> | USNM HerpTissue 113 | JM 758 | Panama: Coclé | JX398477 | JX398627 | JX398752 | |
| <i>D. aparatiritos</i> | USMN 579818 | JM 795 | Panama: Coclé | JX398478 | JX398628 | JX398753 | |
| <i>D. articulata</i> | | D161; MSM/ASL at UCR | Costa Rica: Limón | JX398454 | | JX398740 | |
| <i>D. bicolor</i> | | ASL 277 at UCR | Costa Rica: Limón | JX398455 | | JX398741 | JX293844 |
| <i>D. catesbyi</i> | DHMECN 11952 | ENS 13477 | Ecuador: Napo | JX398456 | JX398608 | | |
| <i>D. catesbyi</i> | UTA R-55949 | ENS 12341 | Ecuador: Tungurahua | JX398457 | JX398609 | JX398742 | JX293845 |
| <i>D. catesbyi</i> | UTA R-55974 | ENS 12204 | Ecuador: Tungurahua | JX398458 | JX398610 | JX398743 | JX293846 |
| <i>D. catesbyi</i> | KU 214851 | WED 57932 | Peru: Madre de Dios | EF078537 | EF078585 | | |
| <i>D. catesbyi</i> | | WED 59073 | Peru: Madre de Dios | JX398459 | JX398611 | JX398745 | JX293847 |
| <i>D. georgejetti</i> | UTA R-61628 | ENS 12817 | Ecuador: Manabí | JX398554 | JX398694 | JX398817 | JX293897 |
| <i>D. gracilis</i> | ICN 12019 | RAM 315 | Colombia: Cesar | JX398465 | JX398615 | JX398746 | JX293852 |
| <i>D. gracilis</i> | UTA R-55943 | ENS 12671 | Ecuador: Esmeraldas | JX398466 | JX398616 | JX398747 | JX293853 |
| <i>D. gracilis</i> | UTA R-55944 | ENS 12672 | Ecuador: Esmeraldas | JX398467 | JX398617 | JX398748 | |
| <i>D. indica</i> | KU 204908 | WED 56989 | Peru: Madre de Dios | JX398468 | JX398618 | JX398734 | JX293854 |
| <i>D. jamespetersi</i> | Bioparque Amaru RSCDSP 0390 | JM 72 (J. M. Daza) | Ecuador: Azuay | JX398555 | JX398695 | JX398818 | JX293898 |
| <i>D. mikanii</i> | | CTMZ 495 | Brazil: São Paulo | | JX398693 | JX398816 | JX293896 |
| <i>D. nicholsi</i> | | JM 812 | Panama: Coclé | JX398469 | JX398619 | | |
| <i>D. pavonina</i> | | LSUMZ-H 13989 | Brazil: Amazonas | JX398470 | JX398620 | JX398749 | JX293855 |
| <i>D. palmeri</i> | DHMECN 11954 | ENS 12421 | Ecuador: Tungurahua | JX398471 | JX398621 | | |
| <i>D. peruana</i> | | LSUMZ-H 1532 | Peru: Pasco | JX398472 | JX398622 | JX398750 | JX293856 |
| <i>D. pratti</i> | MBUCV 6837 | TB 149H | Venezuela: Zulia | JX398473 | JX398624 | JX398751 | |
| <i>D. pratti</i> | MHUA 14638 | | Colombia: Antioquia | JX398474 | JX398623 | | |
| <i>D. temporalis</i> | MHUA 14278 | | Colombia: Antioquia | GQ334583 | GQ334482 | GQ334667 | GQ334560 |
| <i>D. tungida</i> | LSUMZ 36734 | LSUMZ-H 6458 | Bolivia: Unknown | JX398556 | JX398696 | JX398819 | JX293899 |
| <i>D. trinitatis</i> | UWIZM.2011.20.25 | | Trinidad: Arima | JX398479 | JX398629 | | |
| <i>D. variegata</i> | | D99; Vidal et al. (2000) | French Guiana: Cayenne | JX398480 | JX398630 | JX398737 | JX293857 |
| <i>D. variegata</i> | MHNLS 18013 | ENS 11187 | Venezuela: Bolivar | JX398481 | JX398631 | | |
| <i>D. variegata</i> | UTA R-15772 | WWL 3152 | Suriname: Marowijne | JX398482 | JX398601 | JX398736 | JX293858 |
| <i>D. vermiculata</i> | UTA R-55939 | ENS 12353 | Ecuador: Morona-Santiago | JX398483 | JX398632 | JX398754 | JX293859 |

First record of the genus *Touranella* Attems, 1937 (Diplopoda, Polydesmida, Paradoxosomatidae) from Laos, with a description of a new species

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Abstract

The paradoxosomatid genus *Touranella* Attems, 1937 is recorded from Laos for the first time, with a new species, *Touranella champasak* **sp. nov.**, described here. The taxonomy of the genus is discussed, an identification key is provided, and the current distribution of all species is mapped.

Keywords

Bioinventory, Champasak, diversity, millipede, new species, taxonomy

Introduction

The genus *Touranella* Attems, 1937 was established for a single species, *Touranella gracilis* Attems, 1937. Attems (1937) distinguished this genus from other paradoxosomatids recorded in Vietnam at that time by the following features: gonopod femorite strongly reduced or completely absent, and solenomere (= Rinnenast) arising from the prefemorite. No other records had been reported until Golovatch (1994: 186) described the second species of the genus, *Touranella himalayaensis* Golovatch, 1994,

from Nepal. The type locality of *T. himalayaensis* lies approximately 2,500 km north of Vietnam, indicating a significant biogeographical gap among species in the same genus. Six additional species of *Touranella* were described between 2009–2018, five from Vietnam and one from Nepal. To date, eight *Touranella* species have been described (Sierwald and Spelda 2021), which are listed below:

1. *Touranella cattiensis* Golovatch & Semenyuk, 2010 from Cat Tien National Park, Dong Nai, Vietnam.
2. *Touranella gracilis* Attems, 1937 from Da Nang, Vietnam.
3. *Touranella himalayaensis* Golovatch, 1994 from Panchthar, Nepal.
4. *Touranella hirsuta* Golovatch, 2009 from Bi Doup–Nui Ba National Park, Lam Dong, Vietnam.
5. *Touranella moniliformis* Golovatch & Semenyuk, 2018 from Cat Tien National Park, Dong Nai, Vietnam.
6. *Touranella peculiaris* Golovatch, 2009 from Bi Doup–Nui Ba National Park, Lam Dong, Vietnam.
7. *Touranella pilosa* Golovatch, 2016 from Sankhua Sabha, Nepal.
8. *Touranella trichosa* Golovatch & Semenyuk, 2018 from Kon Ka Kinh National Park, Gia Lai, Vietnam.

This work reports the first record of *Touranella* in Laos, with a description of a new species. With this discovery, the geographical gap in the distribution of this genus is slightly narrowed (Fig. 1).

Material and methods

Examined material was collected by M. Thayer and her colleagues during their field expedition to Laos in 2008 and is currently housed in the Field Museum of Natural History (FMNH).

The specimen was examined under a Leica M205 microscope. Line drawings were made using a camera lucida attached to the Leica M205 microscope. Colour images were taken using the Nikon 5100 imaging system with varying lens sizes under normal and ultraviolet (UV) light. Images were photographed in different layers and stacked using Helicon Focus v. 6.0, then grouped into plates in Photoshop v. 6.0. A gonopod was dissected for morphological observation and mounted on an aluminum stub, coated with gold for SEM imaging. SEM images were taken using a Leo Scanning Electron Microscope (Carl Zeiss SMT, Peabody, MA) at FMNH. A distribution map was created using Google Map.

Abbreviations

- FMNH** Field Museum of Natural History;
INS Insect Division;
NP National Park.

Taxonomic part

Order Polydesmida Pocock, 1887

Family Paradoxosomatidae Daday, 1889

Genus *Touranella* Attems, 1937

Touranella Attems, 1937: 231.

Touranella—Attems 1938: 233; Hoffman 1963: 591 (placed in the newly described tribe Alogolykini); Jeekel 1968: 64, “*incertae sedis*”; Hoffman 1980: 172, “unassigned tribal position”; Golovatch 1994: 187 (placed in the Alogolykini); Golovatch 2009a: 6; Golovatch 2009b: 120; Nguyen and Sierwald 2013: 1179; Golovatch 2016: 139; Golovatch and Semenyuk 2018: 16.

Type species. *Touranella gracilis* Attems, 1937, by original designation.

Touranella champasak sp. nov.

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Figs 2–6

Material examined. Holotype: LAOS • male; Champasak Province, Bolaven Plateau, Ban Thongvay (=Xekatam), vic. old logging road, N of village; 15°14.288'N, 106°31.891'E; 1,095 m elev.; 8–16 June 2008; A. Newton & M. Thayer leg.; selectively logged forest, FMHD#2008-037, flight intercept trap, ANMT site 1231; **FMNH INS 3716303**.

Diagnosis. The new species can be recognized by a submoniliform body; poorly developed paraterga; sparsely setose metaterga; the presence of a highly elevated, setose, trapeziform, sternal process between male coxae 4; a strongly reduced gonofemorite devoid of a femoral process; a somewhat twisted solenophore that distally sheaths a rod-shaped solenomere; and well-developed lamina medialis and lamina lateralis.

The species is most similar to *Touranella moniliformis* Golovatch & Semenyuk, 2018 from Cat Tien NP (Vietnam) by having a (sub-)moniliform body, poorly developed paraterga, and sparsely setose metaterga. The two species can be distinguished by the gonopod conformation, and the presence of a gonofemoral process in *T. moniliformis* (absent from the new species).

Regarding the absence of a gonofemoral process, the new species is similar to *T. peculiaris* Golovatch, 2009, but can be distinguished by a strongly reduced gonofemorite (vs considerably elongated in *T. peculiaris*).

Etymology. The species epithet, “*champasak*”, is a noun in apposition and refers to the province name where the type was collected.

Description. Holotype length ca 21.6 mm, width of midbody pro- and metazona about 1.5 mm and 1.9 mm, respectively.

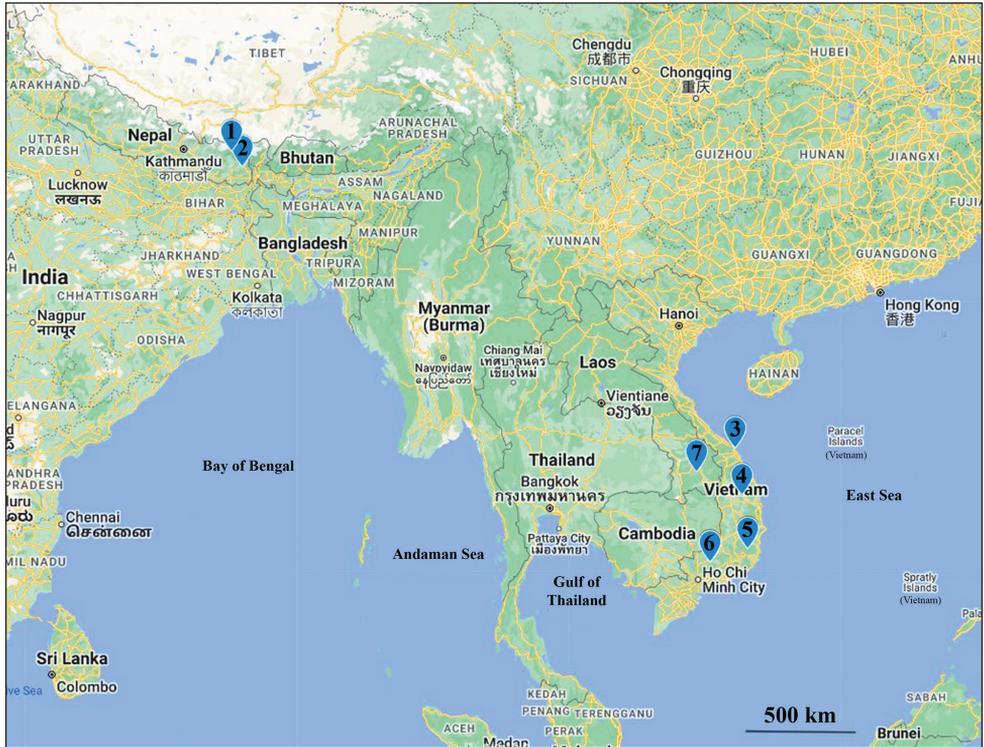


Figure 1. Distribution of the genus *Touranella* Attems, 1937. 1 = *Touranella pilosa* Golovatch, 2016, 2 = *Touranella himalayaensis* Golovatch, 1994, 3 = *Touranella gracilis* Attems, 1937, 4 = *Touranella trichosa* Golovatch & Semenyuk, 2018, 5 = *Touranella hirsuta* Golovatch, 2009 and *Touranella peculiaris* Golovatch, 2009, 6 = *Touranella cattiensis* Golovatch & Semenyuk, 2010 and *Touranella moniliformis* Golovatch & Semenyuk, 2018, 7 = *Touranella champasak* sp. nov.

Body brown and darkish brown, except several antennomeres; legs and sterna brownish yellow or yellow; posterior margins of prozonae and metazonae, anterior margins of metazonae, and transverse sulcus black; metaterga with a yellow axial band running from collum to telson (Fig. 2D).

Antenna long and slender, approximately reaching to segment 5 when extended back; antennomere 1 very short and robust (Fig. 2B); antennomere 2=3=4=5=6> 7 in length. Tip with four sensory cones. Antennomere 2 strongly constricted at base (Fig. 2A, B).

Collum smooth and shiny, suboval, with two rows of setae: 3+3 anterior and 2+2 posterior. Paraterga small, broadly rounded lobe (Fig. 2A).

Body submoniliform. Prozonae and metazonae smooth, shiny (Figs 2A, C–D, 3). Metatergal transverse sulcus present from segment 5, but completely developed starting on bodyring 6 (Fig. 2D). Metaterga with traces of two setal rows: 2+2 anterior and 2+2 posterior. Pleurosternal carinae present as full crests on segments 2–4, becoming less developed on subsequent segments, completely missing on segments 18–19. Stricture between pro- and metazonae very distinct, fully striolate at bottom on both dorsal and lateral sides (Fig. 3D). Axial line thin, distinct.

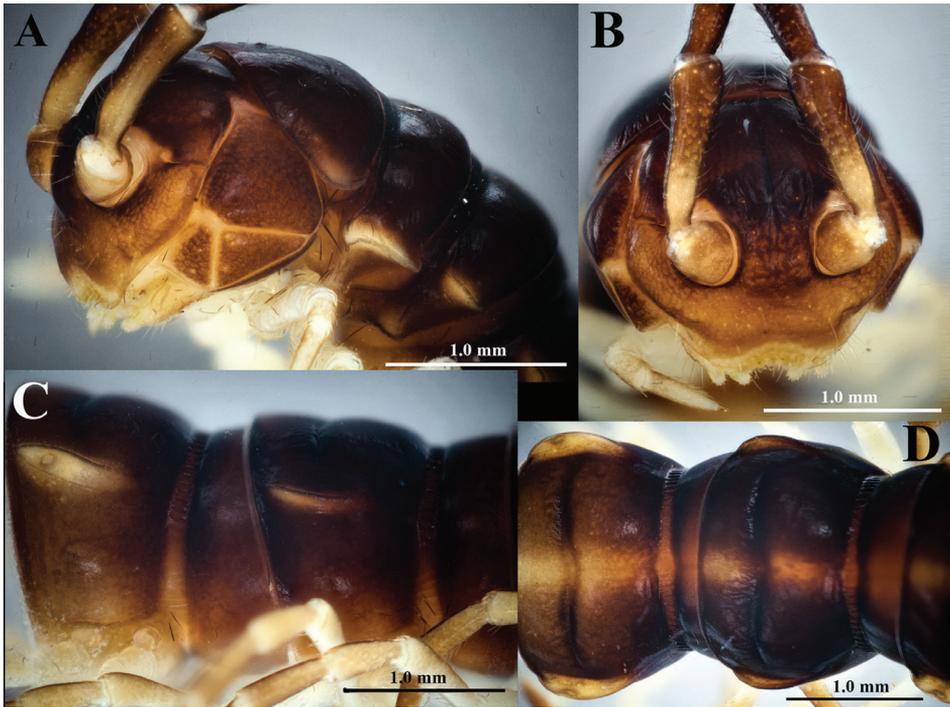


Figure 2. *Touranella champasak* sp. nov., holotype **A, B** head, lateral and anterior views, respectively **C, D** segments 8 and 9, lateral and dorsal views, respectively.

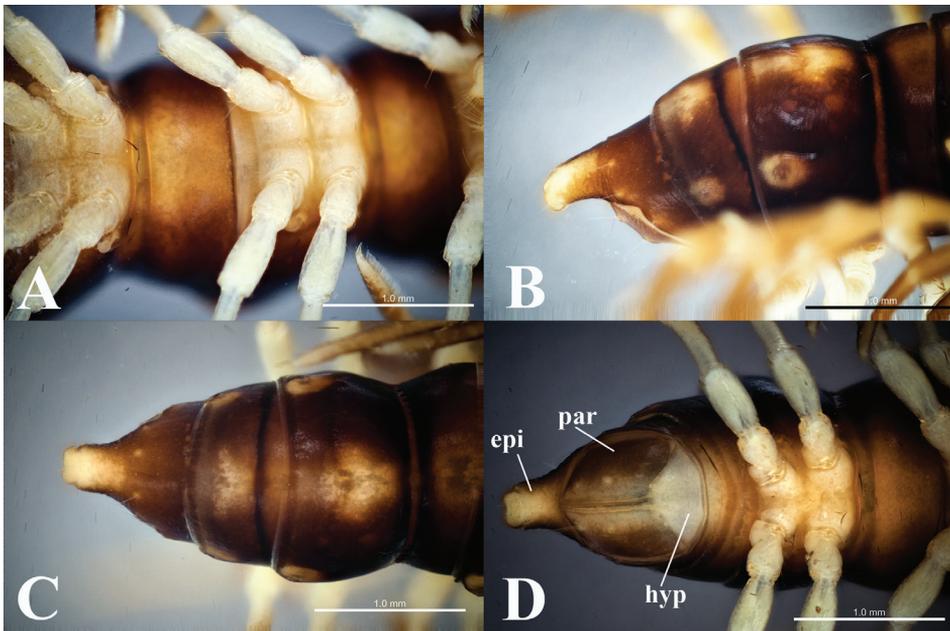


Figure 3. *Touranella champasak* sp. nov., holotype **A** segments 8 and 9, ventral view **B–D** caudal part of body, lateral, dorsal, and ventral views, respectively. Abbreviations: epi = epiproct; par = paraproct; hyp = hypoproct.

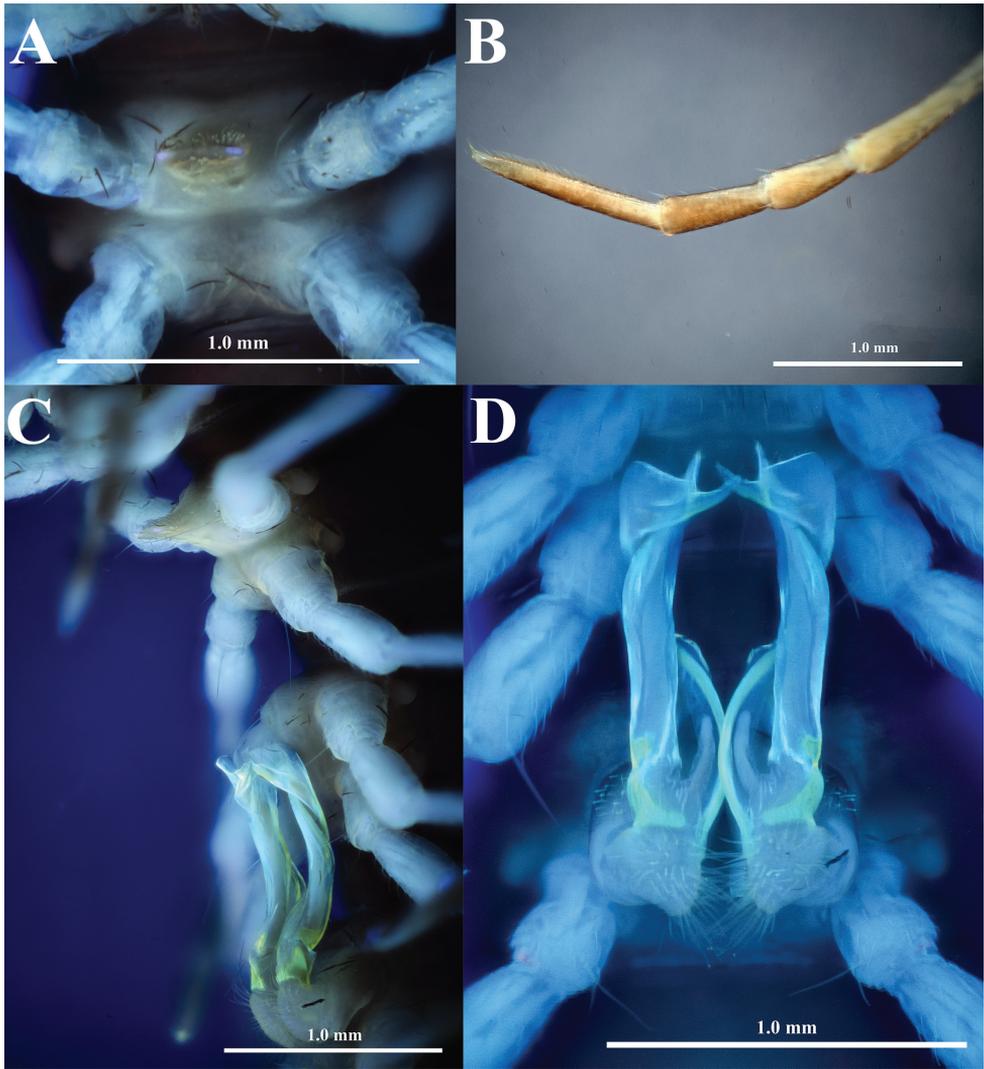


Figure 4. *Touranella champasak* sp. nov., holotype **A** sternum 5, ventral view, UV light **B** posterior leg on segment 9, anterior view, normal light **C** gonopods and sternum 5, lateral view, UV light **D** gonopods, ventral view, UV light.

Paraterga (Figs 2C, D, 3B, C) yellowish, small as complete crests from lateral side, but more obvious on pore-bearing segments, slightly directed caudally upwards.

Epiproct (**epi**) (Fig. 3B–D) long, broadly truncated, flattened dorsoventrally, lateral tubercles minute; tip with four spinnerets. Hypoproct (**hyp**) (Fig. 3D) subtrapeziform, with two separated, distolateral, setiferous knobs. Paraprocts (**par**) sub-semicircular with two distinct setiferous knobs.

Legs long and slender, about 1.7–1.8 times as long as midbody height. Prefemora not swollen. Femora without modification. Tarsal brushes (Fig. 4B) present on legs until segment 16.

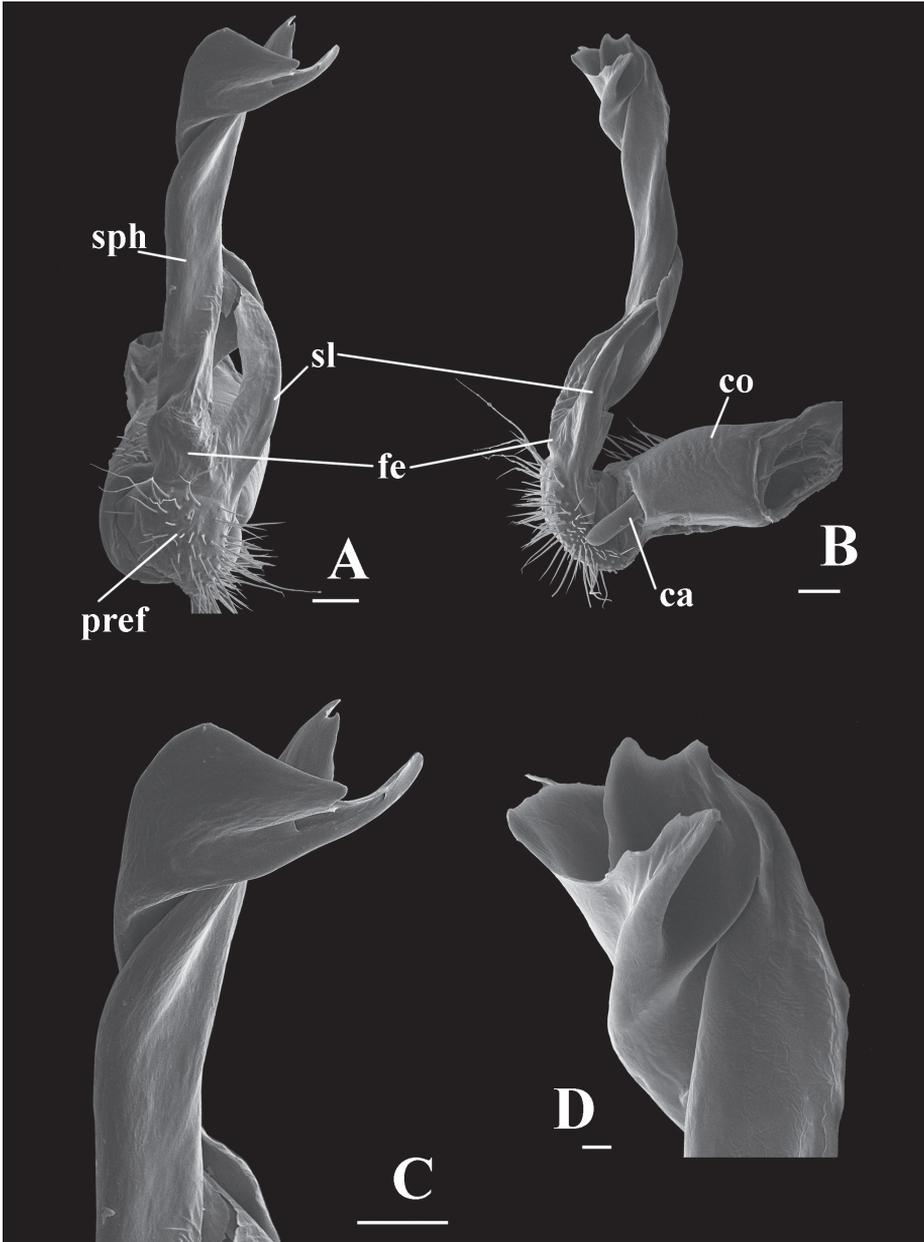


Figure 5. *Touranella champasak* sp. nov., holotype **A, B** right gonopod, ventral view and mesal view, respectively **C, D** distal part of gonopod, ventral view and mesal view, respectively. Abbreviations: co = gonocoxite; pref = gonoprefemorite; fe = gonofemorite; sph = solenophore; sl = solenomere; ca = cannula. Scale bars: 0.1 mm (**A-C**), 0.02 mm (**D**).

Sterna (Fig. 3A, D) with distinct cross-impression, without modifications except a highly elevated, setose trapeziform process between coxae 4 (Fig. 4A, C). This process carrying a setal brush on anterior side and two pores at base.

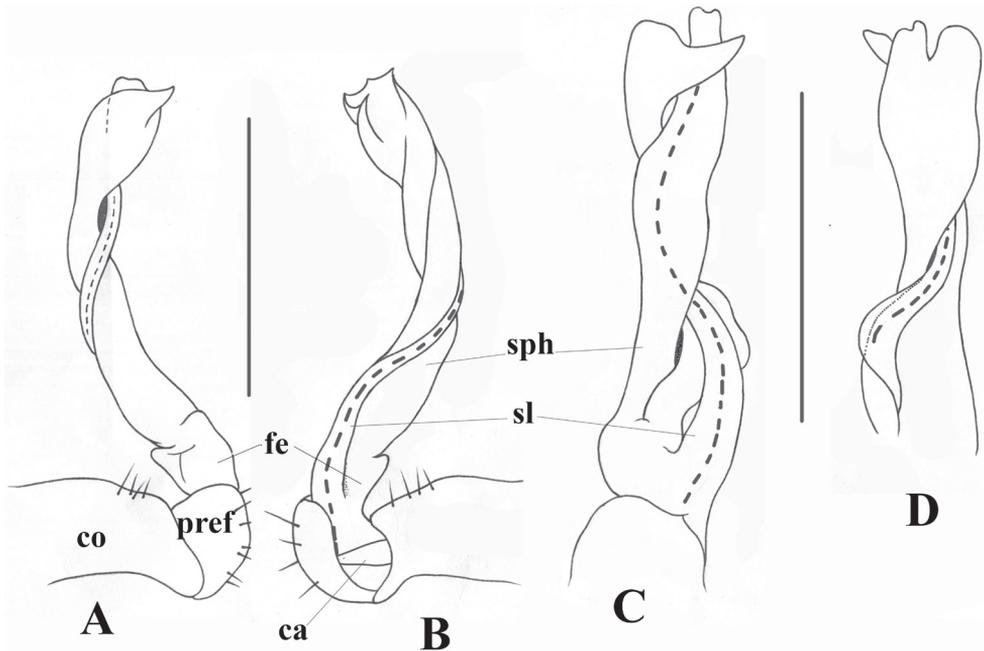


Figure 6. *Touranella champasak* sp. nov., holotype **A–D** right gonopod, lateral, mesal, dorsal, and ventral views, respectively. Abbreviations: co = gonocoxite; pref = gonoprefemorite; fe = gonofemorite; sph = solenophore; sl = solenomere; ca = cannula. Scale bars: 1 mm.

Gonopods (Figs 4C, D, 5, 6) simple. Coxite (**co**) subcylindrical, as long as about $\frac{1}{2}$ telopodite, distoventral part sparsely setose. Prefemorite (**pref**) short, densely setose. Femorite (**fe**) strongly reduced, without femoral process. Postfemoral region extremely long, consisting of only solenomere (**sl**) and solenophore (**sph**). Solenomere rod-shaped, arising from prefemorite, distal part sheathed by solenophore, which is suberect, slightly twisted at distal part; lamina lateralis with an apical spine and a well-developed, rounded lobe. Tip of gonopod serrated with three distinct denticles.

Remarks. Even though the distributional gap is slightly narrowed by the occurrence of this genus in Laos, more species most probably have yet to be discovered, at least in and between southern Vietnam and Nepal, including Laos, northern Thailand, and Myanmar (Fig. 1).

An identification key to *Touranella* species

Since the recent key provided by Golovatch (2016), three more species have been discovered; therefore, the key is updated.

- 1 Metaterga smooth, without setae or with two setal rows.....2
- Metaterga with three setal rows or densely setose.....3
- 2 Gonopod femoral process present *T. moniliformis*
- Gonopod femoral process absent..... *T. champasak* sp. nov.

- 3 Gonopod femoral process absent..... 4
- Gonopod femoral process present 7
- 4 Metaterga beset with long setae placed inside minute pores/knobs. Solenophore with vestigial parabasal lobe, distinct, acuminate, apical uncus and a couple of characteristic subapical outgrowths *T. trichosa*
- Metaterga with transverse rows of setae, instead of long hairs. Solenophore with or without a shoulder near base, and in a different shape..... 5
- 5 Metaterga with six rows of setae borne on small bosses..... *T. hirsuta*
- Metaterga with three rows of setae 6
- 6 Gonofemorite short. Solenophore without a basal shoulder *T. cattiensis*
- Gonofemorite considerably elongated. Solenophore with a basal shoulder
..... *T. peculiaris*
- 7 Gonofemorite carrying three processes..... *T. pilosa*
- Gonofemorite carrying only a single process 8
- 8 Femoral process long. Basal shoulder of solenophore well developed.....
..... *T. himalayaensis*
- Femoral process short. Basal shoulder of solenophore less developed *T. gracilis*

Discussion

Attems (1937) distinguished the monotypic genus *Touranella* by a greatly shortened gonofemorite, the presence of a femoral process, and the densely setose metaterga. This diagnosis was supported by the discovery of the second species, *Touranella himalayaensis* Golovatch, 1994. However, other *Touranella* species recently found in Vietnam have revealed new diagnostic characters as in Golovatch (2009a, 2009b, 2016) or Golovatch and Semenyuk (2010, 2018). Briefly, the genus can be recognized by having a submoniliform body, poorly developed paraterga, legs with neither modifications nor adenostyles, the presence of a sternal process between coxae 4, the gonofemorite either strongly reduced or very short as compared to the solenophore, the solenomere mostly rod-shaped or subflagelliform, sheathed by the solenophore distally, and both lamina medialis and lamina lateralis well developed.

Morphologically, the genus *Touranella* can be divided into two groups based on the presence or absence of the gonofemoral process. The first group includes the types species, *T. gracilis*, and four others, *T. himalayaensis*, *T. pilosa*, *T. trichosa*, and *T. moniliformis*. These species are characterized by the absence of the gonofemorite, or having it strongly reduced with a femoral process. They are also characterized by a solenophore with or without a lateral basal shoulder. The second group contains *T. peculiaris*, *T. cattiensis*, *T. hirsuta*, and *T. champasak* sp. nov., which are characterized by a very short or considerably elongated gonofemorite, without a femoral process. Given the absence of the femoral process and/or short gonofemorite, this second group is relatively close to the genus *Yuennanina* Attems, 1936. However, *Touranella* can be differentiated from *Yuennanina* using the first leg pair in males (femoral tubercles are absent from *Touranella* males, but present in *Yuennanina* males) and coxa (a thumb-

like process is evident anteriorly in *Yuennanina*, but absent from *Touranella*). The relationship between *Touranella* with *Yuennanina* remains uncertain at this time.

The genus *Touranella* belongs to the tribe Alogolykini, created by Hoffman (1963: 591) for the genera *Tetracentrosternus* Pocock, 1895, *Alogolykus* Attems, 1936, and *Touranella*. He stated that members of this tribe could be recognized by having an extremely shortened gonofemorite; presence of a femoral process arising from the prefemorite; a slender solenophore that completely or partially sheaths the solenomere; and the first male leg pair without femoral tubercles. Jeekel (1965) noted the presence of femoral tubercles in the legs of *Tetracentrosternus* males and suggested a closer relationship between *Tetracentrosternus* and *Yuennanina*. Subsequently, Jeekel (1968: 127) retained these three genera (*Tetracentrosternus*, *Alogolykus*, and *Yuennanina*) in the tribe Alogolykini. Instead, he considered *Touranella* as *incertae sedis*, and stated that: “It is true that, as in all other Alogolykini, the gonopod femorite is reduced as in *Touranella*, but this is not a reason to postulate a close relationship. As a matter of fact, the tibiotarsus and its relation to the solenomerite rather strongly suggest the conditions in, e.g. the Orthomorphini, etc.” (Jeekel 1968: 65). This exclusion was still retained by Hoffman (1980: 171) and Jeekel (1980: 174). However, the genus was re-assigned to the tribe Alogolykini by Golovatch (1994: 187) and Nguyen and Sierwald (2013: 1179). This assignment was supported by additional newly described species (Golovatch 2009a, 2009b; Golovatch and Semenyuk 2010, 2018; Golovatch 2016).

According to Likhitrakarn et al. (2013) and Golovatch et al. (2021), the tribe Alogolykini can be distinguished from its close relative, Polydrepanini, as members have a strong, rod-shaped solenomere (vs a thin, flagelliform solenomere in Polydrepanini). Both these tribes are the only components of the subfamily Alogolykinae. This tribe Alogolykini currently consists of seven genera: a monotypic *Alogolykus* (from Myanmar), *Yuennanina* (three species from southern China), *Tetracentrosternus* (four species from Myanmar, Thailand, and southern China), and *Touranella* (eight species from Nepal, Laos, and Vietnam), *Singhalorthomorpha* Attems, 1914 (three species from Sri Lanka), a monotypic *Curiosoma* Golovatch, 1984 (from India), and finally a monotypic *Carlogonopus* Golovatch, Aswathy, Bhagirathan & Sudhikumar, 2021 (also from India) (Golovatch et al. 2021). However, a revision of this tribe is beyond the scope of this paper, and it is suggested that phylogenetic analyses employing morphological and molecular data are needed to elucidate relationships among these genera.

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Generic placement of the African flea beetle *Polyclada maculipennis* Bryant and the possible occurrence of the genus *Procalus* Clark in sub-Saharan Africa (Coleoptera, Chrysomelidae, Galerucinae, Alticini)

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Abstract

Polyclada Chevrolat and *Procalus* Clark are flea beetle genera (Coleoptera, Chrysomelidae, Galerucinae, Alticini). *Polyclada* is endemic to the Afrotropical region, while *Procalus* has never been described outside of the Neotropical region. The new combination *Procalus maculipennis* (Bryant, 1942), **comb. nov.** is proposed for *Polyclada maculipennis* Bryant, 1942. Its plausible type locality is Venezuela, and not Cameroon, as recorded on the labels of the type material, and hence the occurrence of *P. maculipennis* in Africa is questionable.

Keywords

Afrotropical region, Neotropical region, taxonomy, new combination, lectotype designation

Introduction

Polyclada Chevrolat, 1836 is an Afrotropical flea beetle genus occurring in sub-Saharan Africa, Saudi Arabia, and Yemen. It comprises 16 described species currently under revision (Biondi and D'Alessandro 2010, 2012; Biondi et al. 2022). The genus

is associated with Anacardiaceae (*Sclerocarya birrea* (A. Rich) Hochst.) and Burseraceae (*Commiphora* spp.), in a variety of woodland and savannah ecosystems (Chaboo et al. 2007; Iannella et al. 2021). *Polyclada* is one of the five Afrotropical genera belonging to the *Blepharida*-group *sensu* Furth and Lee (2000) and Prathapan and Chaboo (2011), along with *Diamphtidia* Gerstaecker, *Xanthophysca* Fairmaire, and the recently re-evaluated *Blepharidina* Bechyné and *Calotheca* Heyden (Biondi et al. 2017, 2019; D'Alessandro et al. 2018, 2019, 2020, 2021). The *Blepharida*-group currently comprises 21 genera from the Afrotropical, Nearctic, Neotropical, and Oriental regions (Medvedev 1999; Furth and Lee 2000; Prathapan and Chaboo 2011; Biondi et al. 2017). Furth and Lee (2000) provided a morphological synthesis of the group based on adult characters (tarsal claws, procoxal cavities, head, pronotum, hind femora, eye, proepimeron, and metatibia) and larval characters (antenna, mandible, labrum, stemmata, endocarina, coronal suture, and frontal suture). However, some characters are shared by most (but not all) of the genera (Furth and Lee 2000), and a more comprehensive analysis based on the whole set of genera and representative species is badly needed to more rigorously test the monophyly of the group.

During revisionary studies of the genus *Polyclada*, we examined the type material of *P. maculipennis* Bryant, 1942, in the general collection of the Natural History Museum in London (NHMUK) and noticed that it belongs to the Neotropical genus *Procalus* Clark, 1865. Bryant (1942) based the description of this species on three specimens from Cameroon and believed it to be allied to *Polyclada bohemani* (Baly, 1861).

Procalus comprises an unknown number of species, but including *P. mutans* (Blanchard, 1851), *P. viridis* (Philippi & Philippi, 1864), *P. lenzi* (Harold, 1876), *P. reduplicatus* Bechyné, 1951, *P. malaisei* Bechyné, 1951, and *P. silvai* Jerez, 1995 (Jerez 1992, 1995). Three more species were reported by Artigas and Solar (2015): *P. artigasi* Jerez, *P. ortizi* Jerez, and *P. vilosensis* Jerez, which are also cited by other authors (Jerez 2003; Prathapan and Chaboo 2011). However, they were described in an unpublished doctoral thesis (Jerez 1999a, cited by Jerez 2003), therefore, according to Article 9.12 of the International Code of Zoological Nomenclature (ICZN 2020), these three names cannot be considered as available.

In this paper, we revise the taxonomic status of *Polyclada maculipennis* and discuss the possible occurrence of the genus *Procalus* in sub-Saharan Africa.

Materials and methods

Examined material consisted of dried, pinned specimens preserved in the institutions listed below. The specimens were examined and dissected under a Leica M205C stereomicroscope. Photographs were taken using a Leica DMC5400 camera and were compiled using Zerene Stacker v. 1.04. Scanning electron micrographs were taken using a Hitachi TM-1000. Abbreviations of the depositories follow Evenhuis (2022).

Depositories

| | |
|--------------|---|
| BAQ | collection of M. Biondi, University of L'Aquila, Italy; |
| MSNG | Museo Civico di Storia Naturale "Giacomo Doria", Genova, Italy; |
| NHMB | Naturhistorisches Museum, Basel, Switzerland; |
| NHMUK | The Natural History Museum, London, United Kingdom; |
| NMPC | National Museum (Natural History), Prague, Czech Republic. |

Results

Procalus maculipennis (Bryant, 1942), **comb. nov.**

Figs 1–3, 5–8

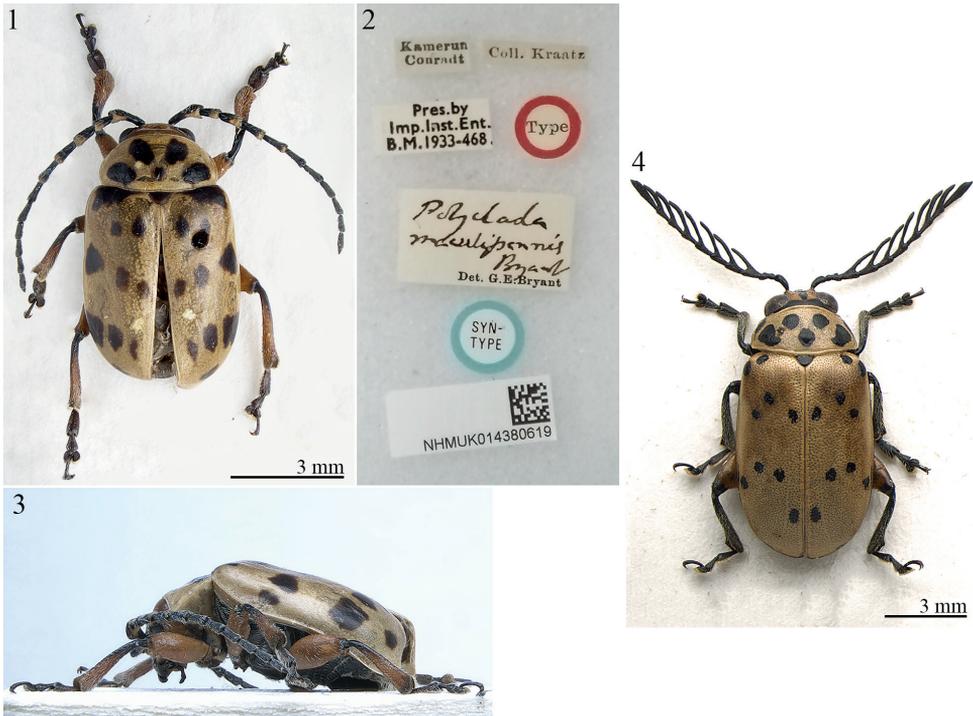
Polyclada maculipennis Bryant, 1942: 164.

Type material examined. Lectotype of *Polyclada maculipennis* ♂: "Kamerun Conradt // Coll. Kraatz // Pres. By Imp. Inst. Ent. B.N. 1933-468 // *Polyclada maculipennis* Bryant / Det. G.E. Bryant" (NHMUK) (here designated by M. Biondi and P. D'Alessandro) (Figs 1–3). Paralectotypes of *Polyclada maculipennis*, 2♀♀: same data as for lectotype (NHMUK).

Material of *Procalus* species examined for comparison. Chile: *Procalus reduplicatus*, syntype, 1♀, "Chile // Coll. Nickerl / Mus. Pragense // *Procalus mutans* Blchd. // Typus // *Procalus reduplicatus* n. sp. Type ♀ / 1948 Det. J. Bechyně" (NMPC); *Procalus* cf. *viridis*, 1 specimen, El Tabo, Valparaiso, 29 Sept. 1986 (NHMB); *Procalus* cf. *lenzi*, 1 specimen, Santiago, Pudahuel, 20 Nov. 1986 (NHMB); *Procalus* cf. *malaisei*, 2 specimens, D. Ed. Varas Arangua leg., 1921 (MSNG); *Procalus* cf. *lenzi*, 1 specimen, ibid (MSNG); *Procalus* cf. *viridis*, 1 specimen, Viña del Mar, Valparaiso, May 1899, F. Silvestri leg. (MSNG); *Procalus* cf. *silvai*, 1 specimen, Concepcion, 1903, P. Herbst leg. (MSNG); *Procalus* cf. *reduplicatus*, 1 specimen, ibid (MSNG); *Procalus* cf. *mutans*, 1 specimen, Concepcion, Sept. 1903, P. Herbst leg. (MSNG); *Procalus* sp., 2 specimens, ibid (MSNG); *Procalus* cf. *reduplicatus*, 1 specimen, Concepcion, 1904, P. Herbst leg. (MSNG); *Procalus* sp., 1 specimen, ibid (MSNG).

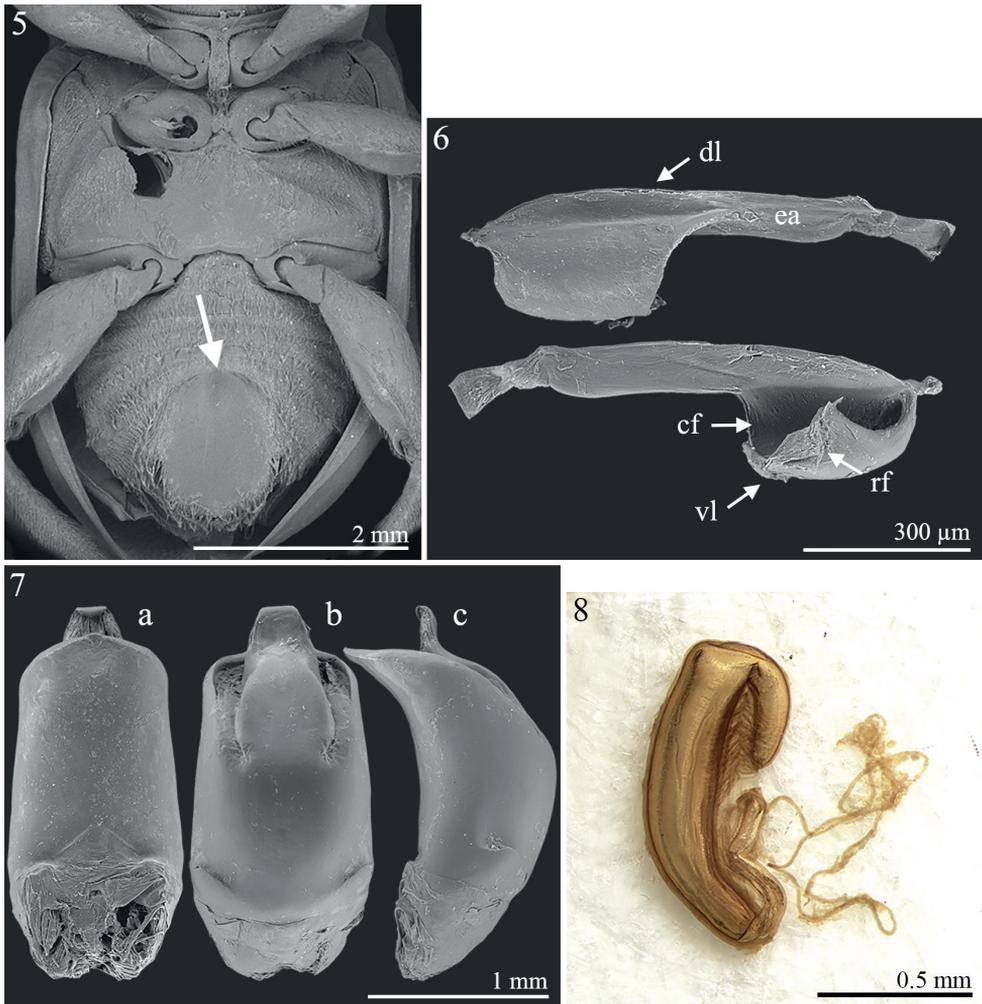
Remarks. *Polyclada* can be immediately distinguished from similar genera by the antennae, longer than half body length, with antennomeres 4–10 pectinate or flabellate in male and serrate in female (Fig. 4). Characters of the antenna along with other morphological features, such as procoxal cavities open posteriorly, antennomere 4 at least double length of antennomere 3, as well as elytral punctation always confused, densely and uniformly impressed, permits identification of the genus within the *Blepharida* group in the Afrotropical region (Biondi and D'Alessandro 2012; Biondi et al. 2017).

The type material of *Polyclada maculipennis* has all the diagnostic characters of *Procalus* (Clark 1865; Jerez 1992), none of which occur in any *Polyclada* species: antennae



Figures 1–4. Lectotype of *Polyclada maculipennis* Bryant, 1942 **1** habitus, dorsal view **2** ibid., labels **3** ibid., habitus, lateral view **4** *Polyclada bohemani* Baly, male, Kenya (BAQ), habitus.

with antennomeres 1–5 flattened compared to 6–11, especially in male, and antennomere 1 clearly distally enlarged, and strikingly serrated in male (Fig. 1; Clark 1865; Jerez 1992: figs 4A, 5A, 6A, 7A, 8A, 1995: figs 1, 2); fifth abdominal sternite in male with a wide, deep, oval depression (Fig. 5; Jerez 1992: figs 5E, 6D, 7C, 8E, 1995: fig. 2); metafemoral extensor tendon simplified, very slender (Fig. 6; Furth and Suzuki 1994: fig. 6b; Jerez 1992: figs 2, 3D); dorsal lobe straight, with very elongate extended arm; central furrow very wide; ventral lobe subtriangular; recurved flange short, poorly sclerotized. Additionally, the median lobe of aedeagus and spermatheca (Figs 7, 8) are typical of the *Procalus* species (median lobe of aedeagus: Furth and Suzuki 1994: fig. 6a; Jerez 1992: figs 4E, 5C, 6E, 7B, 8G, 1995: fig. 10; spermatheca: Furth and Suzuki 1994: fig. 6c; Jerez 1992: figs 4D, 5D, 6B, 7D, 8E, 1995: fig. 11). Median lobe of the aedeagus of the lectotype here designated (Fig. 7): thickset and smooth; in ventral view tapering towards the apex, and slightly narrowing subapically; apex subtriangular, widely obtuse, with a small median tooth; in lateral view, clearly curved. Spermatheca of the paralectotypes (Fig. 8): subcylindrical and elongate basally, curved towards the ductus attachment; distal part distinctly bent and about 1/3 the basal part in length; ductus basally inserted, short, uncoiled, moderately thickset. We therefore propose the new combination *Procalus maculipennis* (Bryant, 1942), comb. nov.



Figures 5–8. *Procalus maculipennis* Bryant, 1942 comb. nov. **5** ventral parts in male **6** metafemoral extensor tendon **7** median lobe of aedeagus in ventral, dorsal, and lateral views (**a–c**, respectively) **8** spermatheca. Abbreviations: cf: central furrow; dl: dorsal lobe; ea: extended arm; rf: recurved flange; vl: ventral lobe.

Discussion

Clark (1865) reported the genus *Procalus* as abundant and widely distributed in Chile, and also present in Brazil and Bolivia. Based on Scherer (1983), this genus occurs in Chile, Bolivia, Brazil, and Argentina. Jerez (1992, 1995) considered *Procalus* as occurring only in Chile, based on her examined material. Later, she (Jerez 1999b) stated that, based on both public and personal collections, the genus is distributed between latitudes of 30°50'S and 40°50'S, and cited undetermined *Procalus* material from southern Argentina preserved in the Museum National d'Histoire Naturelle de Paris. Endemic to Chile or not, it has never been described outside of the Neotropical region.

What about *Procalus maculipennis* comb. nov. from Cameroon? We can only speculate about some different hypotheses. Hypothesis A: the genus *Procalus* has a Gondwanan distribution, even though it is dramatically more abundant in the Neotropical region. The Gondwanan distributions of terrestrial taxa generally refer to genera with clearly differentiated species, or higher taxa (cf. Gómez-Zurita and Cardoso 2021). Among flea beetles, the genera with Afrotropical–Neotropical disjunct distributions occur in the two regions with clearly differentiated species: *Terpnochlorus* Fairmaire from the Afrotropical region, Venezuela, and Mexico, and, if the synonymy is confirmed, the Malagasy *Abrarius* Fairmaire, is possibly a senior synonym of *Gioia* Bechyné from South America (Biondi and D'Alessandro 2012). A second Afrotropical–Neotropical disjunct distribution is observed at a higher taxonomic level; for example, *Zomba* Bryant is the only representative of the subtribe Monoplatina in the Afrotropical region. This subtribe occurs almost exclusively in the Neotropical and southern part of the Nearctic regions and is present in the Australian region only with the genus *Opisthopygme* Blackburn, 1896. Based on the diagnostic characters reported by Jerez (1992), *Procalus maculipennis* comb. nov. is so similar to *P. reduplicatus* Bechyné that one could evaluate to establish a synonymy in a possible future revision of the genus. For Hypothesis A to be true, these two taxa would have remained so similar despite approximately 135 million years of independent evolution (cf. Donateli Gatti et al. 2021), even though the diversification of *Procalus* in South America occurred in a much shorter time, likely during the Pliocene (Jerez 1999b). Hypothesis B: the species was imported to Africa via host plants. *Procalus* species are associated with *Lithraea* Miers ex Hook. & Arn. and *Schinus* L. (Anacardiaceae) (Jerez 1992, 1999b). The genus *Lithraea*, native to South America, is reported as introduced only in California and Tunisia, not in Cameroon or other sub-Saharan countries (POWO 2021). *Schinus* is also native to South America, is used ornamentally around the world, and was imported into several African countries, but not into Cameroon and adjacent areas (POWO 2021).

Hypothesis C: the types are mislabelled. Starting from the assumption that Leopold Conradt was the collector, it is possible that the entomological material he collected or somehow acquired in Venezuela (where he stayed for some time before 1889) was brought to Cameroon, where he subsequently collected in 1896 (Rohlfien 1975). In fact, material from Venezuela reached the Deutschen Entomologischen Instituts—now Senckenberg Deutsches Entomologisches Institut (SDEI)—via Gustav Kraatz in 1905 along with material from Togo and Cameroon (Rohlfien 1975). It is plausible that the samples from Venezuela were mixed up with, and then erroneously labelled as being from Cameroon. A similar interpretation about the putative and unlikely disjunct distributions of some Hymenoptera species have been offered by Liston et al. (2017). Similarly, Furth (1998) highlighted that *Blepharida semisulcata* Achard originally described from Cayenne (French Guiana) is a mislabelled specimen from the Afrotropical Region. For us, this is the most plausible hypothesis to explain the alleged presence of *Procalus* in Africa. However, fieldwork in Cameroon and Venezuela in search of the original habitat and host plants of *Procalus maculipennis* comb. nov. can put the issue to rest.

Acknowledgements

We are grateful to the collection managers and curators from the institutions that enabled us to study their material: NHMUK, NHMB, MSNG, and NMPC. Thanks to Dr Josephine Paris (University of L'Aquila) for the English-language editing. We finally thank David Furth, Chi-Feng Lee, an anonymous reviewer, and the editor, Caroline Chaboo, for improving the manuscript.

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A key to genera of Dikraneurini from China, with description of a new species of *Cornicola* Ohara & Hayashi (Hemiptera, Cicadellidae, Typhlocybinae)

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Abstract

The leafhopper genus *Cornicola* Ohara & Hayashi, previously recorded from Japan, is recorded from China for the first time and a new species, *C. maculatus* Xu, Dietrich & Qin, **sp. nov.**, is described and illustrated, including its color polymorphism. This genus has male genitalia and hind wing venation similar to those found in Emposcini but it is more appropriately placed in Dikraneurini. A key to species of *Cornicola* is given together with a key to the genera of Dikraneurini from China.

Keywords

Auchenorrhyncha, distribution, Homoptera, leafhopper, morphology, polymorphism, taxonomy

Introduction

The tribe Dikraneurini is a diverse group and differs from other Typhlocybinae leafhoppers in lacking an appendix in the forewing and in usually having the hind wing submarginal vein complete and extended past vein RA or RP basad along the costal margin (Dietrich 2005). However, some genera included in this tribe either lack the hind wing submarginal vein (*Typhlocybella* Baker) or have this vein reduced or obsolete at the apex

of the costal margin and thus resemble species of *Empoascini* (Viraktamath and Dietrich 2011; Dietrich 2013; Ohara and Hayashi 2022). One such genus in the latter category is *Cornicola* Ohara & Hayashi, 2022, with *C. mizuki* Ohara & Hyashi, from Japan, as its type species. In this paper, a second species of *Cornicola* is described as new from south-west China, together with a key to Chinese Dikraneurini genera. To date, Dikraneurini contain 74 genera and 497 valid species distributed throughout the world (Dmitriev et al. 2022) of which 25 genera and more than 60 species occur in China and have been studied by Matsumura (1931), Anufriev and Emeljanov (1988), Dworakowska (1972, 1979, 1993a), Chou and Ma (1981), Zhang and Chou (1988), Zhang (1990), Zhang and Kang (2007), Kang and Zhang (2012, 2013), Yang et al., (2012), Kang et al. (2013), Jiao and Yang (2015, 2020), Huang et al. (2018), Kang et al. (2018), Qin et al. (2020).

Materials and methods

The specimens examined in this study were preserved in 95% ethanol stored for three years resulting in loss of the original color; they are now deposited in the insect collection of Illinois Natural History Survey, Champaign, Illinois (INHS). Morphological terminology used in this work follows Xu et al. (2021).

Taxonomy

Family Cicadellidae Latreille, 1825

Subfamily Typhlocybinae Kirschbaum, 1868

Tribe Dikraneurini McAtee, 1926

Genus *Cornicola* Ohara & Hayashi, 2022

Type species. *Cornicola mizuki* Ohara & Hayashi, 2022, by original designation.

Diagnosis. *Cornicola* is easily distinguishable from all other known Typhlocybinae in having the following combination of characters: (1) crown of head much narrower than pronotum and strongly elevated above anterior margin of pronotum (Figs 3, 6); (2) forewing with vein R2 and RM arising from r cell and MCu from m cell (Fig. 9); (3) hind wing with submarginal vein obsolete along costal margin and anal vein branched (Fig. 10); (4) male pygofer with dorsal margin almost straight, with short preapical fingerlike process, folded mesad subapically, ventral appendage absent (Figs 16, 17); and (5) subgenital plates fused in proximal 1/3, with lateral macrosetal row (Fig. 19).

Notes. Ohara and Hayashi (2022) recognized that *Cornicola* is related to *Igutettix* Matsumura, 1932 and therefore placed the genus in Dikraneurini; and also compared the genus to *Vilbasteana* Anufriev, 1970, *Koreoneura* Hossain & Kwon, 2021 and *Sweta* Viraktamath & Dietrich, 2011. However, the hind wing venation of *Cornicola* differs from the above-mentioned genera and instead resembles that of the Southeast Asian

dikraneurine genera *Rakta* Dietrich, 2013 and *Albodikra* Dietrich, 2013 in having the submarginal vein obsolete or reduced apically along the costal margin of the hind wing (Fig. 10; fig. 2b, d in Dietrich 2013) and thus resembling that of Emposcini. *Cornicola* differs from these two genera in having an anteclypeus only slightly convex in both sexes (Figs 5, 8) (strongly swollen and broad in males of *Rakta* and *Albodikra*). Despite a strong resemblance of the hind wing venation of the new genus to the common pattern in Emposcini and some additional similarities in the male genitalia (e.g., elongate style), *Cornicola* is clearly more closely related to Dikraneurini and may represent a transitional form between Dikraneurini and Emposcini.

Distribution. Japan (Hokkaido, Honshu, Shikoku) and China (Chongqing).

Key to species of *Cornicola* Ohara & Hayashi (males)

- 1 Male 2S apodemes extended nearly to posterior margin of segment V; aedeagus with shaft not widened at apex in lateral view, distal processes not forked at base, evenly curved in posterior view.....
*Cornicola mizuki* Ohara & Hayashi
- Male 2S apodemes reaching to end of segment IV (Fig. 15); aedeagus with shaft apex widened in lateral view (Fig. 23) distal processes forked near base, longer fork bent at acute angle in posterior view (Fig. 24).....
*Cornicola maculatus* Xu, Dietrich & Qin, sp. nov.

Cornicola maculatus Xu, Dietrich & Qin, sp. nov.

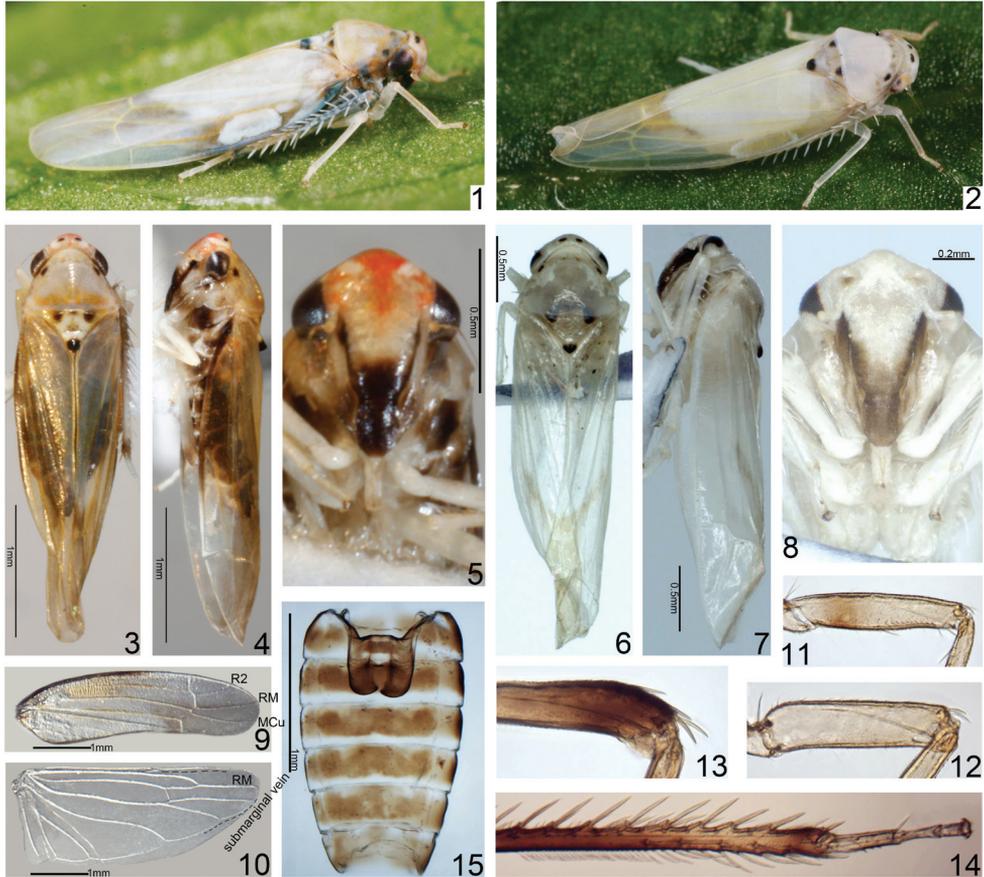
<https://zoobank.org/8BADF99F-7AA7-45AA-B06D-484E0D6565D0>

Figs 1–24

Type materials. *Holotype*. ♂ (INHS) S.W. CHINA, Chongqing Jinyan Shan, 29.829630°N, 106.376380°E, 872 m, 10 Sep. 2016, CH Dietrich, sweep roadside, CN-16-08. *Paratypes*. 4♂5♀(INHS) same data as holotype.

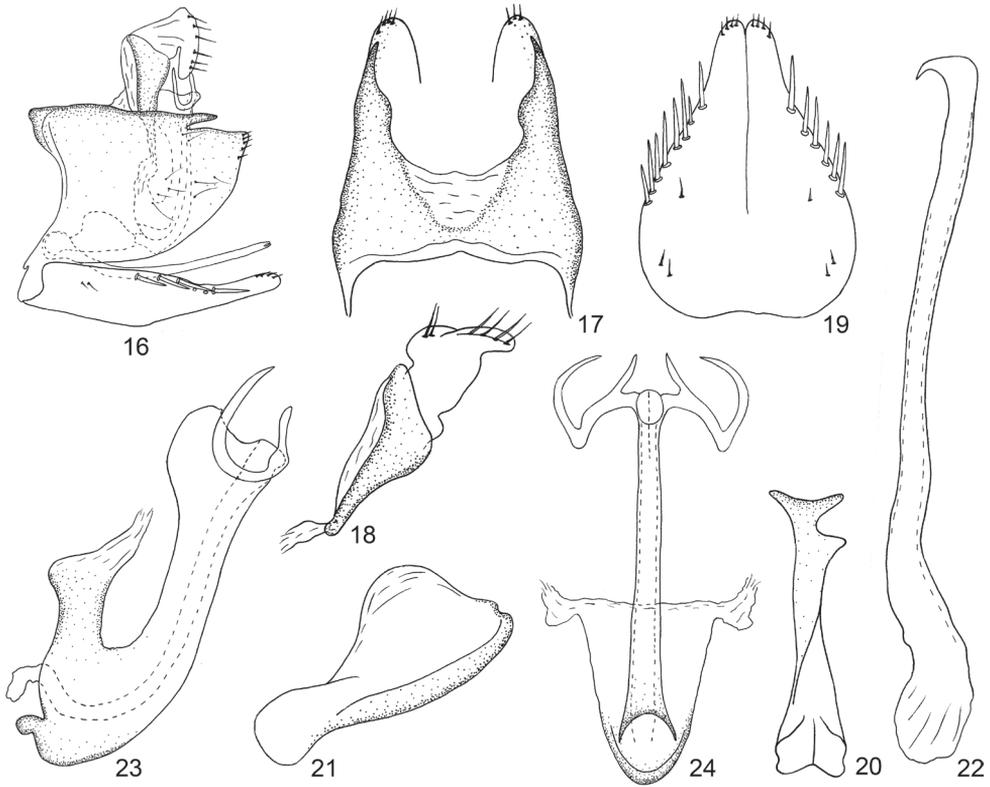
Description. Body length: male 3.1–3.5 mm, female 3.2–3.5 mm.

Adults of this species are polymorphic with two color forms, one being mostly white (Figs 6, 7) and other with extensive reddish-brown coloration (Figs 3, 4) in both males and females (Figs 1–8). Reddish-brown morph (Figs 1, 3–5): Crown beige, with two small black oval patches and two irregular whitish patches surrounding dark red coronal suture, frontoclypeus with lateral dark brown band in lower half extending to base of antenna, lorum orange to dark (Figs 1, 3, 5). Eyes dark, ocelli irregularity with whitish spots (Figs 1, 3–5). Pronotum mostly orange red, with two small oval patches behind eyes, mesonotum with suborbicular spots, otherwise whitish except heart-shaped black patch medially in scutellum (Figs 1, 3–5). Forewing orange to reddish, veins brown; hind wing hyaline, veins white (Figs 1, 3, 4, 9). Front and middle legs almost hyaline, whitish except tarsus brown, hind legs brown (Fig. 1). White morph (Figs 2, 6–8): white overall with black spots and maculate, as in reddish-brown morph.



Figures 1–15. *Cornicola maculatus* Xu, Dietrich & Qin sp. nov. **1, 2** Adults showing different body coloration **3** female adult, dorsal view **4** female adult, left lateral view **5, 8** face **6** male adult (abdomen removed), dorsal view **7** male adult (abdomen removed), left lateral view **9** forewing **10** hind wing **11** left femur and base of tibia, anterior view **12** left middle femur, anterior view **13** left hind femur apex and base of tibia, anterior view **14** distal part of hind tibia and tarsus, anterior view **15** sternal apodemes.

Basal sternal abdominal apodemes parallel sided, reaching end of segment IV (Fig. 15). Male pygofer almost triangular in lateral view, dorsal margin with finger-like process arising near distal third of dorsal margin and extended posterad, not reaching apex; distal lobe bearing 6 or 7 microsetae, ventral margin with 8 or 9 feeble microsetae, dorsal bridge occupying more than one-third length of pygofer (Figs 16, 17). Anal tube gradually narrowed apically (Fig. 18). Subgenital plate longer than pygofer lobe in lateral view, broad basally, fused in basal two-thirds, tapered distally, apex rounded and strongly narrowing, with sparse scattered microsetae, 6–8 macrosetae arranged in single row along each dorsolateral margin near midlength (Fig. 19).



Figures 16–24. *Cornicola maculatus* Xu, Dietrich & Qin sp. nov. **16** male genital capsule, left lateral view **17** male pygofer, dorsal view **18** anal tube, left lateral view **19** subgenital plates **20** connective, lateral view **21** connective, dorsal view **22** style **23** aedeagus, left lateral view **24** aedeagus, ventral view.

Connective widest medially with subapical angular projection in lateral view, apical margin emarginate medially (Figs 20, 21). Style apodeme much shorter than apophysis, preapical lobe absent, without conspicuous setae, slightly broadened preapically, apex smooth, slightly broadened then tapered to hooklike tip, curved laterad (Fig. 22). Aedeagus with shaft broad at base, narrowed near middle and with broad dorsal distal lobe in lateral view; pair of slender distal processes extended laterad from adjacent gonopore, each with short dorsomedially directed spine and elbow-like bend near midlength with distal part curved dorsomesad in posterior view and anterodorsad in lateral view (Figs 23, 24).

Notes. This new species differs from *Cornicola mizuki* by the characters noted in the key.

Distribution. China (Chongqing).

Etymology. The species name is derived from the Latin words ‘*maculatus*’, referring to the black spots on the crown and thorax.

Key to genera of Chinese Dikraneurini

- 1 Subgenital plates fused basally (Fig. 19) 2
 – Subgenital plates separate..... 6
- 2 Hind wing with submarginal vein obsolete along costal margin (Fig. 10).....
 *Cornicola* Ohara & Hayashi, 2022
- Hind wing with submarginal vein complete, extending along costal margin,
 around apex 3
- 3 Hind wing with MP and CuA fused for short distance.....
 *Karachiota* Ahmed, 1969
- Hind wing with MP and CuA separate, connected by a short cross-vein
 (Fig. 10)..... 4
- 4 Forewing with veins R2 and RM confluent preapically
 *Motschulskyia* Kirkaldy, 1905
- Forewing with veins R2 and RM separate, connected by a cross-vein..... 5
- 5 Connective about twice longer than wide *Cuanta* Dworakowska, 1993
- Connective nearly as long as wide *Platfusa* Dworakowska, 1993
- 6 Hind wing with veins R and RA free, connected by a cross-vein
 *Urvana* Dworakowska, 1993
- Hind wing with veins R and RA confluent distally 7
- 7 Forewing with veins R2, RM and MCu confluent preapically..... 8
- Forewing with veins R2 and MCu separate preapically 9
- 8 Male pygofer with dense setae distally; 2S apodemes surpassing segment III.
 *Flatseta* Jiao & Yang, 2015
- Male pygofer without dense setae distally; 2S apodemes not reaching segment
 III *Takagioma* Thapa, 1989
- 9 Pygofer ventral appendage present *Golwala* Dworakowska, 1993
- Pygofer ventral appendage absent..... 10
- 10 Style moderately long and thin, longer than subgenital plate.....
 *Uniformus* Jiao & Yang, 2020
- Style shorter than subgenital plate..... 11
- 11 Forewing with vein MCu reduced, not extending to wing margin 12
- Forewing with vein MCu complete..... 13
- 12 Subgenital plates triangular, narrowed apicad..... *Naratettix* Matsumura, 1931
- Subgenital plates nearly oblong, truncated apicad
 *Dicraneurula* Vilbaste, 1968
- 13 Connective absent..... *Forcipata* DeLong & Caldwell, 1942
- Connective present 14
- 14 Connective immovably attached or fused with base of aedeagus..... 15
- Connective movably articulated with base of aedeagus..... 16
- 15 Male pygofer with upper appendage, without articulated caudal sclerite; sub-
 genital plate with few macrosetae *Togaricrania* Matsumura, 1931
- Male pygofer without upper appendage, with articulated caudal sclerite; sub-
 genital plate without macrosetae *Trifida* Thapa & Sohi, 1986

| | | |
|----|---|---|
| 16 | Forewing with vein R2 and RM confluent preapically | 17 |
| – | Forewing with veins R2 and RM separate, connected by cross-vein | 21 |
| 17 | Aedeagus with pair of processes on shaft | 18 |
| – | Aedeagus without processes | 20 |
| 18 | Subgenital plate with more than eight macrosetae, arranged roughly in two rows | <i>Michalowskiya Dworakowska</i> , 1972 |
| – | Subgenital plate with fewer than five macrosetae, arranged in single row..... | 19 |
| 19 | Male pygofer nearly rectangular, without process; subgenital plate with few basal setae | <i>Iniesta Dworakowska</i> , 1993 |
| – | Male pygofer variable in shape, with process; subgenital plate without basal setae..... | <i>Anaka Dworakowska & Viraktamath</i> , 1975 |
| 20 | Aedeagus with gonopore apical, with circle of unpigmented cuticular outgrowths; male 2S apodemes surpassing segment V | <i>Uzeldikera Dworakowska</i> , 1971 |
| – | Aedeagus with gonopore not as above; male 2S apodemes surpassing segment IV | <i>Igutettix Matsumura</i> , 1932 |
| 21 | Subgenital plates not surpassing pygofer lobe..... | 22 |
| – | Subgenital plates surpassing pygofer lobe | 23 |
| 22 | Style with well-developed preapical lobe | <i>Dikraneura Hardy</i> , 1850 |
| – | Style without preapical lobe | <i>Ayubiana Ahmed</i> , 1969 |
| 23 | Aedeagus with dorsal apodeme absent..... | <i>Riyavaroa Dworakowska</i> , 1993 |
| – | Aedeagus with dorsal apodeme present | 24 |
| 24 | Aedeagus with paired shafts | <i>Notus Fieber</i> , 1866 |
| – | Aedeagus with single shaft | 25 |
| 25 | Male pygofer triangular; aedeagus without processes | <i>Wagneriala Anufriev</i> , 1970 |
| – | Male pygofer nearly round; aedeagus with few processes..... | <i>Erythria Fieber</i> , 1866 |

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