CHECKLIST



An annotated checklist of the planthoppers of Iran (Hemiptera, Auchenorrhyncha, Fulgoromorpha) with distribution data

Fariba Mozaffarian¹, Michael R Wilson²

l Insect Taxonomy Research Department, Iranian Research Institute of Plant Protection, Tehran, 19395, P.O. Box 1454, Iran **2** Department of Biodiversity & Systematic Biology, National Museum of Wales, Cardiff, CF10 3NP, UK

Corresponding author: Fariba Mozaffarian (mozaffarian@iripp.ir, faribamozaffarian@gmail.com)

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Abstract

A list of Hemiptera Fulgoromorpha (planthoppers) of Iran is provided, based primarily on literature records from 1902 to the present. In total 15 families and 235 species are recorded, with taxonomic details. Distribution data in Iran are given. *Iranissus ephedrinus* Dlabola, 1980 is transferred from Issidae to Nogodinidae. To resolve nomenclatural difficulty the following new combinations in Issidae are given: *Iranodus dumetorus* (Dlabola, 1981), *Iranodus khatunus* (Dlabola, 1981) and *Iranodus repandus* (Dlabola, 1981). Due to published generic synonomy the following are new combinations: *Duilius seticulosus* (Lethierry, 1874), *D. tamaricis* (Puton & Lethierry, 1887), *D. tamariciola* (Dubovsky, 1966) and *D. v-atrum* (Dlabola, 1985).

Keywords

Hemiptera, Auchenorrhyncha, Fulgoromorpha, annotated checklist, Iran

Introduction

The infraorder Fulgoromorpha, also known as planthoppers, belong to the suborder Auchenorrhyncha of the order Hemiptera. Planthoppers are phytophagous and feed from the sap of a wide range of plants and in many habitats. They may occur in relatively large numbers in some habitats and under certain conditions.

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The wide-ranging history of entomology in Iran has been well-documented by chapters in Abivardi (2001). The earliest available records of Fulgoromorpha in Iran were by Melichar (1902a, b) who described some species from Iran. Afshar (1937) was the first Iranian entomologist who recorded *Ommatissus binotatus lybicus* (Tropiduchidae) in the list of agricultural pests for the country. Subsequent taxonomic studies in the Persian literature mainly focused on the insect pests in both agricultural and forest ecosystems (Gharib 1966, 1998; Kheyri 1989; Ahmadi et al. 1986; Karimzadeh Esfahani et al. 1998 etc). Field expeditions were made to various parts of Iran during the 1970s by Czech entomologists, including Jiri Dlabola, along with entomologists from the Iranian Research Institute of Plant Protection. This resulted in a number of new discoveries of Iranian planthoppers, which were compiled in publications such as Dlabola (1974a, b, 1977a, b, 1980a, 1981, 1982a, b, 1983a, b, 1985 etc). Dlabola is responsible for the description of almost 50% of the currently known planthopper fauna of Iran.

Fifteen planthopper (Fulgoromorpha) families have been recorded from Iran to date. Table 1 gives details of the numbers of species in each family and the number (and percentage) known only from Iran at present. Large numbers of species have been recorded in the Cixiidae, the Delphacidae, the Flatidae and the Issidae. The fauna is characterized by high endemism in groups such as Issidae, which are flightless and have relatively limited dispersal and a narrow distribution. However, the planthopper fauna may also well be under-recorded, for instance around 100 species of Delphacidae are known from Greece (e.g. Drosopoulos et al. 1983) but only 34 so far recorded in Iran. Dlabola (1981) refers to the biotic zones in which species are found.

In the current study a list of Fulgoromorpha species recorded from Iran has been compiled, which is based on literature records and the addition of some new taxonomic data. Much basic taxonomic work is needed in the region and it is hoped that this checklist will facilitate further studies on Fulgoromorpha of Iran. To resolve nomenclatural difficulty the following new combinations in Issidae are given: *Iranodus dumetorus* (Dlabola, 1981), *Iranodus khatunus* (Dlabola, 1981) and *Iranodus repandus* (Dlabola, 1981). Due to published generic synonomy the following are new combinations: *Duilius seticulosus* (Lethierry, 1874), *D. tamaricis* (Puton & Lethierry, 1887), *D. tamaricicola* (Dubovsky, 1966) *D. v-atrum* (Dlabola, 1985).

Economic importance of planthoppers in Iran

Planthoppers are of high economic importance mainly because they reduce crop yield through feeding and oviposition and they can transfer phytoplasmas and viruses to a wide range of plants. The planthoppers of economic importance in the United States have been reviewed by Wilson and O'Brien (1987) who recorded 150 species of planthoppers from 99 economic plants. At least 8 species in that list have been also recorded from Iran, on agricultural products such as date palm, potato, tomato, wheat, corn, rice (Gharib 1966, 1998; Haghshenas and Khajehali 2000; Khajehali et al. 2001;

Families	No. of species	No. species/ % endemic
Caliscelidae	14	7 (50)
Cixiidae	51	19 (36)
Delphacidae	34	4 (11)
Derbidae	4	3 (75)
Dictyopharidae	19	10 (52)
Flatidae	30	19 (63)
Fulgoridae	3	0 (0)
Issidae	45	38 (84)
Kinnaridae	4	3 (75)
Lophopidae	1	0
Meenoplidae	3	1 (33)
Nogodinidae	5	5 (100)
Ricaniidae	3	1 (33)
Tettigometridae	14	1 (25)
Tropiduchidae	5	3 (60)
Totals	235	114 (48%)

Table 1. Numbers of species in each Fulgoromorpha family

Nematollahi and Khajehali 2000). A further nine species have also been recorded in Iran by Abaii (2000) as pests of forest trees.

Annotated species checklist

The following list is intended as an annotated checklist and not a catalogue. Emphasis is given to citations where Iran is recorded. Synonyms added after the current scientific name contain the original name, next the current name followed by its authority and next those synonyms which have been used for records from Iran. Selected taxonomic papers are cited where needed for clarity. Taxa are given in alphabetical order. Where species have only been found in Iran no further details of a wider distribution are given. Species listed in references, where no further information is given beyond the name, are cited as 'listed'. Locality records are listed for each species. Furthermore a list of locality names including latitude, longitude, codes for the localities and a distribution map are also provided. The codes consist of two parts: an alphabetical one for the name of provinces and a numerical one for the locality (Figs 1, 2).

The extralimital distribution in the Palaearctic region was determined using various publications, including: Asche (1982), Asche and Wilson (1989), Demir and Demirsoy (2009), Demir (2007), Demir (2008), Dlabola (1965b, c, 1972, 1974b, 1977c, 1979d, 1980b, c, 1994), Dlabola and Jankovic (1981), Gnezdilov (2002), Gnezdilov and Wilson (2006), Holzinger et al. (2003), Holzinger (2007), Nast (1972), Novikov et al. (2006), Vilbaste (1980) and Wilson (2010a, b). The United Nations geoscheme was used for recording extralimital distributions in other regions and subregions of the world.



Figure 1. Location of the provinces of Iran (See list of the provinces for codes) A Ardabil AG Äzarbäiejan-e Gharbi ASh Äzarbäiejan-e Sharghi B Bushehr Ch Chahārmahāl–Bakhtiāri E Esfahān F Fārs G Gilān Gh Ghom Gs Golestān Gz Ghazvin H Hormozgān Hd Hamadān I Ilām KB Kohgiluyeh–Boyerahmad Kd Kordestān Kh Khuzestān KhJ Khorāsān-e Jonubi KhR Khorāsān-e Razavi KhSh Khorāsān-e Shomāli Km Kermān L Lorestān M Māzandarān Mz Markazi S Sistān–Baluchestān Sm Semnān T Tehrān Y Yazd Z Zanjān.

Family Caliscelidae

The following species were recorded in the family Issidae, subfamily Caliscelinae in early reports from Iran. This subfamily has however, been more recently given full family status (Tishechkin 1998; Emeljanov 1999; Gnezdilov 2003).

Adenissus baluchestanicus Dlabola, 1980

Adenissus baluchestanicus Dlabola, 1980b: 184; Gnezdilov and Wilson (2006) [listed]. Iran localities: Tang-e Sarheh.

Adenissus isinus Dlabola, 1980

Adenissus isinus Dlabola, 1980b: 185; Gnezdilov and Wilson (2006) [listed]. Iran localities: Isin.



Figure 2. Locality map of Fulgoromorpha recorded from Iran (See List of localities for codes)

Adenissus zabolicus Dlabola, 1980

Adenissus zabolicus Dlabola, 1980b: 181; Gnezdilov and Wilson (2006) [listed]. Iran localities: Nehbandān.

Adenissus zahedanicus Dlabola, 1980

Adenissus zahedanicus Dlabola, 1980b: 182; Gnezdilov and Wilson (2006) [listed]. Iran localities: Zāhedān.

Aphelonema brunneolutea Dlabola 1994

Aphelonema brunneolutea Dlabola 1994: 57 Iran localities: Ziārān.

Ahomocnemia chivensis (Kusnezov, 1929)

Caliscelis chivensis Kusnezov, 1929: 330 Ahomocnemia chivensis; Dlabola (1982a). Iran localities: Jiroft. Extralimital distribution: Central Asia.

Aphelonema registanica (Dlabola, 1961)

Peltonotellus registanicus Dlabola, 1961a: 262
Aphelonema registanica; Dlabola (1981).
Iran localities: Kandovān (Māzandarān), Saādatshahr, Tochāl Mountain.
Extralimital distribution: Central Asia.

Chirodisca astyages (Dlabola, 1982)

Caliscelis astyages Dlabola, 1982a: 114; Mirzayans (1995). Caliscelis dimidiata not Costa, 1863; Dlabola (1981) mis-identification. Chirodisca astyages; comb. n. in Emeljanov (1996a). Iran localities: Ahvāz, Bishāpur, Borāzjān, Dārbahāre, Fasā, Ferdows-e Esfandaghe,

Rafsanjān, Shirāz, Sufiān, Zeidān.

Extralimital distribution: Western Asia.

Ommatidiotus dissimilis (Fallén, 1806)

Issus dissimilis Fallén, 1806: 123
Ommatidiotus falléni Stål 1863: 251; Melichar (1902a); syn. by Dlabola (1987).
Omnatidiotus (sic) dissimilis (Fallén, 1806); Dlabola (1981).
Iran localities: Anārak- Temin, Dasht.
Extralimital distribution: Europe, Turkey, Central and Northern Asia.

Ommatidiotus iranicus Horváth, 1905

Ommatidiotus iranicus Horváth, 1905: 380; Nast (1972) [listed]. Iran localities: Unknown locality. Extralimital distribution: Turkey (Demir 2008).

Perissana bispinata (Dlabola, 1980)

Anissus bispinatus Dlabola 1980b: 179 Perissana bispinata; Gnezdilov and Wilson (2006). Iran localities: Kāzerun, Kushk, Masiri, Saādatshahr. Extralimital distribution: Iraq (Gnezdilov and Wilson 2006).

Perissana dlabolai Gnezdilov & Wilson, 2006

Adenissus circularis Dlabola, 1980b: 186 (Preoccupied). Perissana dlabolai Gnezdilov & Wilson, 2006: 15 (Replacement name for A. circularis).

Iran localities: Bānuchārehar, Bazmān, Dehbakri, Doborji, Estahbān, Fasā, Miānjangal.

Perissana jakowleffi (Puton, 1890)

Issus jakowleffi Puton, 1890; 233 Nast (1972) [listed]. Perissana jakowleffi; Gnezdilov and Wilson (2006). Iran localities: Shāhrud. Extralimital distribution: Azerbaijan.

Reinhardema pasagarda (Dlabola, 1982)

Homocnemia pasagarda Dlabola, 1982a: 116 Reinhardema pasagarda; Gnezdilov (2010a). Iran localities: Shul.

Family Cixiidae

List of subfamilies, genera, and subgenera is according to Holzinger et al. (2002). **Subfamily Cixiinae**

Tribe Duilini

New combinations given are due to generic synonymy given by Holzinger et al. (2002).

Duilius fasciata (Horváth, 1894), comb. n.

Hemitropis fasciata Horváth, 1894: 183
Bitropis fasciata; Dlabola (1985).
Iran localities: Ghāsemābād, Kermānshāh (Yazd province), Nikshahr.
Extralimital distribution: Transcaucasus, Turkey, Afghanistan, central Asia.

Duilius seticulosus (Lethierry, 1874), comb. n.

Haplacha seticulosa Lethierry, 1874: 444 Hemitropis seticulosus; Barkhordari et al. (1981) [listed]. Iran localities: Unknown locality. Extralimital distribution: Saudi Arabia (Dlabola 1979d).

Duilius tamaricis (Puton & Lethierry, 1887), n.comb.

Haplacha tamaricis Puton & Lethierry, 1887: 309 Hemitropis tamaricis; Dlabola (1981). Iran localities: Marand, Robāt-e Tork. Extralimital distribution: Turkey, Central Asia.

Duilius tamaricicola (Dubovsky, 1966), comb. n.

Hemitropis tamaricicola Dubovsky, 1966: 35; Dlabola (1985). Iran localities: Borāzjān, Dārbahāre, Ghāsemābād, Kahkom Kahurak, Kermānshāh (Yazd province), Shushtar.

Extralimital distribution: Uzbekistan.

Duilius v-atrum (Dlabola, 1985), comb. n.

Hemitropis v-atrum Dlabola, 1985: 124; Mirzayans (1995).

Iran localities: Ahram, Bāghak, Bāhukalāt, Bilāi, Geno, Ghāsemābād, Kahkom, Kahnuj, Komehr, Miānjangal, Nikshahr, Tis, Zeidān.

Tribe Cixiini

Cixius (Ceratocixius) adornatus iranicus Dlabola, 1979

Cixius (Ceratocixius) adornatus iranicus Dlabola, 1979b: 233; Mirzayans (1995).

Iran localities: Hashtpar, Khalkhāl, Nāhārkhorān, Rezvāndeh, Sheikh-e Mahalleh, Zirāb.

Cixius (Ceratocixius) cunicularius (Linnaeus, 1767)

Cicada cunicularius Linnaeus, 1767: 711

Cixius cunicularius; Mirzayans (1995).

Iran localities: Parehsar.

Extralimital distribution: North Africa, Europe, Central Asia, Northern and Eastern Asia.

Cixius (Ceratocixius) pallipes Fieber, 1876

Cixius (Ceratocixius) pallipes Fieber, 1876: 191; Dlabola (1981), Mirzayans (1995).

Iran localities: Bandar Anzali, Bidruyeh, Dasht, Ghom, Hafttappeh, Hashtpar, Isin, Karaj, Evin, Lāhijān, Mollāsāni, Parehsar, Rek, Sanandaj.

Extralimital distribution: Southern, Western and Eastern Europe, North parts of Western Asia, Afghanistan.

Comments (M. Asche pers comm.): The occurrence of *pallipes* in Middle East is questionable. The true *pallipes* (described from Italy) is apparently geographically confined to the Central and West Mediterranean. In the East Mediterranean and Middle East there occurs a species described by China (1942) as *Cixius (Ceratocixius) wagneri*. This means that all geographical records of both species need confirmation.

Cixius persicus Distant, 1907

Cixius persicus Distant, 1907: 284; Nast (1972) [listed], Dlabola (1981) [listed]. *Cixius longipennis* Melichar, 1902a: 86 (primary homonym); Melichar (1902a). Iran localities: Nehbandān.

Cixius (Ceratocixius) rufus Logvinenko, 1969

Cixius (Ceratocixius) rufus Logvinenko, 1969: 53; Dlabola (1985). Iran localities: Asālem, Behshahr, Nāhārkhorān, Sabzevār. Extralimital distribution: 'S. Russia' (Nast, 1972).

Cixius (Ceratocixius) simplex (Herrich-Schäffer, 1835)

Flata simplex Herrich-Schäffer, 1835: 64 *Cixius (Ceratocixius) simplex*; Dlabola (1985). Iran localities: Pol-e Tang. Extralimital distribution: North Africa, Europe.

Cixius (Acanthocixius) stigmaticus (Germar, 1818)

Flata stigmaticus Germar, 1818: 199
Cixius (Sciocixius) stigmaticus; Dlabola (1985).
Iran localities: Kandovān (Māzandarān).
Extralimital distribution: Southern, Western and Eastern Europe.

Tachycixius desertorum (Fieber, 1876)

Cixius desertorum Fieber, 1876: 182; Dlabola (1981, 1985). *Tachycixius desertorum*; comb. n. in Wagner (1939).

Iran localities: Sufiān, Eslāmābād, Ziārān.

Extralimital distribution: Southern and Eastern Europe, Turkey, Israel, Central Asia Comments: (M. Asche pers comm.). The records of T. *desertorum* from Iran need confirmation. This species is part of a group of apparently closely related species.

Tribe Oecleini

Myndus genocolus Dlabola, 1985

Myndus genocolus Dlabola, 1985: 97; Mirzayans (1995). Iran localities: Bandar Khamir, Bandar Lengeh, Geno, Isin, Kangān.

Myndus musivus (Germar, 1825)

Flata musivus Germar, 1825: 21; Dlabola (1981), Mirzayans (1995). Iran localities: Orumieh, Orumieh lake. Extralimital distribution: Europe, Central Asia.

Myndus sarbazus Dlabola, 1989

Myndus sarbazus Dlabola, 1989: 33 Iran localities: Sarbāz.

Tribe Pentastirini

Anoculiarus ornatus Dlabola, 1985

Anoculiarus ornatus Dlabola, 1985: 123 Iran localities: Bānuchārehar.

Eumecurus apunctatus Dlabola, 1985

Eumecurus apunctatus Dlabola, 1985: 107 Iran localities: Bānuchārehar, Jiroft, Zāboli.

Eumecurus baluchestanicus Dlabola, 1985

Eumecurus baluchestanicus Dlabola, 1985: 110 Iran localities: Masiri.

Eumecurus (Pseumecurus) frontalis (Melichar, 1904)

Oliarus frontalis Melichar, 1904: 31 Eumecurus (Pseumecurus) frontalis; comb. n. in Dlabola (1985). Iran localities: Isin. Extralimital distribution: Northern Africa.

Eumecurus kabulus (Dlabola, 1957)

Oliarus kabulus Dlabola, 1957a: 267 Eumecurus kabulus; comb. n. in Dlabola (1985). Iran localities: Geno. Extralimital distribution: Afghanistan.

Eumecurus octopus Dlabola, 1985

Eumecurus octopus Dlabola, 1985: 106 Iran localities: Tang-e Sarheh.

Eumecurus superstylus Dlabola, 1985

Eumecurus superstylus Dlabola, 1985: 108 Iran localities: Jiroft.

Eumecurus transpunctatus Dlabola, 1985

Eumecurus transpunctatus Dlabola, 1985: 103 Iran localities: Sarbāz, Sekand.

Eumecurus vilbastei Dlabola, 1985

Eumecurus vilbastei Dlabola, 1985: 109 Iran localities: Isin.

Hyalesthes luteipes Fieber, 1876

Hyalesthes luteipes Fieber, 1876: 197; Dlabola (1981, 1994), Mirzayans (1995)
Iran localities: Ab Ask, Evin, Sirik, Tochal Mountain.
Extralimital distribution: North Africa, Southern, Western and Eastern Europe, Turkey, Israel, Central Asia.

Hyalesthes mlokosiewiczi Signoret, 1879

Hyalesthes mlokosiewiczi Signoret, 1879: 116; Farahbakhsh (1961) [listed], Nast (1972) [listed], Dlabola (1981), Hoch and Remane (1985) [listed], Behdad (1988), Mirzay-ans (1995), Emeljanov (1996b), Abaii (2000).

Iran localities: Āb Āsk, Āmol, Bidhend, Eskandari, Evin, Firuzkuh, Ghezel-Bolāgh, Gorgān, Karaj, Khorramābād, Malārd, Marāgheh, Marand, Robāt-e Tork, Rudbārak, Shāhrud, Shirāz, Tabriz, Tochāl Mountain, Varāmin.

Extralimital distribution: Ukraine, Transcaucasia, Turkey, Central Asia.

Hyalesthes obsoletus Signoret, 1865

Hyalesthes obsoletus Signoret, 1865: 128; Dlabola (1971a, 1972 [listed], 1981, 1994), Behdad (1988), Kheyri (1989), Mirzayans (1995), Abaii (2000), Lashkari et al. (2009).

Iran localities: Āb Āsk, Andimeshk, Behshahr, Evin, Firuzkuh, Gorgān, Hafttappeh, Karaj, Marand, Paskuh, Rezvāndeh, Sanandaj, Sufiān, Tākestān- Karaj Rd, Tochāl Mountain, Yāsuj, Ziārān.

Extralimital distribution: North Africa, Southern, Western and Eastern Europe, Northern parts of Western Asia, Afghanistan and Central Asia.

Hyalesthes philesakis Hoch 1986

Hyalesthes philesakis Hoch 1986: 95; Dlabola (1994) Iran localities: Masiri. Extralimital distribution : Moldova, Ukraine, Turkey.

Hyalesthes restultus Dlabola, 1994

Hyalesthes restultus Dlabola, 1994: 44 Iran localities: Komehr.

Hyalesthes scotti Ferrari, 1882

Hyalesthes scotti Ferrari, 1882: 82; Dlabola (1994).
Hyalesthes luteipes Fieber. var. scotti Ferrari; (Linnavuori, 1962a).
Iran localities: Dasht, Gazanak, Sufiān, Tochāl Mountain.
Extralimital distribution: Western and central Mediterranean region, perhaps also in southern parts of central Europe. (Holzinger et al. 2003).

Hyalesthes zabolicus Dlabola, 1985

Hyalesthes zabolicus Dlabola, 1985: 121; Hoch and Remane 1985 [listed]. Iran localities: Zāboli.

Oliarellus fulvus (Kusnezov, 1935)

Hyalesthes fulvus Kusnezov, 1935: 213
Oliarus fulvus (Kusnezov 1935); Nast (1972) [listed], Lashkari et al. (2009).
Pseudoliarus circularis Dlabola, 1981: Dlabola (1981).
Oliarellus fulvus; Dlabola (1985) [listed].
Iran localities: Estahbān, Ghazvin, Golestān province, Robāt-e Tork.
Extralimital distribution: Central Asia.

Oliarus convergens Melichar, 1902

Oliarus convergens Melichar, 1902a: 87; Dlabola (1981) [listed]. Iran localities: Kermān province. Comment: the status of this species is unclear.

Oliarus nigrofurcatus Signoret, 1884

Oliarus nigrofurcatus Signoret 1884: 71; Melichar (1902a), Farahbakhsh (1961) [listed], Nast (1972) [listed], Dlabola (1972) [listed]. Iran localities: Nehbandān.

Extralimital distribution: Transcaucasus, Iraq, Afghanistan and Central Asia. Comments: The identity of this species is unclear.

Pentastira bahtiaricus (Dlabola, 1981)

Oliarus bahtiaricus Dlabola, 1981: 144; *Pentastira bahtiaricus*; comb. n. in Dlabola (1988). Iran localities: Marg-e-Malek

Pentastira major Kirschbaum, 1868

Pentastira major Kirschbaum, 1868: 44 Oliarus major (Kirschbaum, 1868); Dlabola (1981), Mirzayans (1995). Pentastira major; (Dlabola, 1988).

Iran localities: Akhlamad, Āmol, Behshahr, Dasht-e Moghān, Divāndareh, Gazanak, Ghāemshahr, Gharechaman, Gorgān, Kahurak, Karaj, Nowshahr, Rāmiān, Sanandaj, Shemirān, Silvāneh, Zāhedān, Zoshk.

Extralimital distribution: Southern and Eastern Europe, North parts of Western Asia.

Pentastira superspicata Dlabola, 1985

Pentastira superspicata Dlabola, 1985: 119 Iran localities: Kāzerun, Masiri.

Pentastira shul (Dlabola, 1985)

Reptalus shul Dlabola, 1985: 115; Dlabola (1985). Pentastira shul; comb. n. in Dlabola (1988). Iran localities: Kāzerun, Masiri, Miāneh, Shul, Yāsuj.

Pentastiridius (Pentastiridius) leporinus (Linnaeus, 1761)

Cicada leporina Linnaeus, 1761: 242 Oliarus leporinus; Dlabola (1971b, 1981). Flata pallens Germar, 1821; synonymized by Nast (1986). Oliarus pallens (Germar, 1821); Melichar (1902a), Dlabola (1960a, 1971a, 1972 [listed], 1981), Farahbakhsh (1961) [listed], Nast (1972) [listed]. Reptalus pallens (Germar, 1821); Mirzayan (1995). Pentastiridius leporinus; Nast (1986), Mirzayans (1995). Iran localities: Ābādeh, Albāji, Bampur, Bazmān, Birjand, Chābahār, Dālaki, Evin, Gambuyeh, Gāvbandi, Gharechaman, Hafttappeh, Hamidieh, Hāresābād, Hashtpar, Irānshahr, Kandovān (Māzandarān), Marand, Miāneh- Zanjān Rd, Minushahr, Mollāsāni, Shādegān, Shieh, Susangerd, Suza, Tabriz, Varāmin, Zābol.

Extralimital distribution: North Africa, Europe, Northern parts of Western Asia, Afghanistan, Central and Eastern Asia.

Pseudoliarus fuscofasciatus (Melichar, 1902)

Oliarus fuscofasciatus Melichar, 1902a: 88; Nast (1972) [listed], Dlabola (1972 [listed], 1981), Mirzayans (1995).

Pseudoliarus fuscofasciatus; Dlabola 1981.

Iran localities: Ābādeh, Bazmān, Bidruyeh, Dālaki, Geno, Ghom, Isin, Jahrom, Jiroft, Mollāsāni, Zābol.

Extralimital distribution: North east Africa, Yemen, Iraq, Transcaucasia.

Pseudoliarus palestinensis (Linnavuori, 1962)

Oliarus fuscofasciatus palestinensis Linnavuori, 1962a: 3 Pseudoliarus palestinensis; Dlabola (1985).

Iran localities: Ahvāz, Jāsk, Kahkom. Extralimital distribution: Saudi Arabia (Dlabola 1980c), Turkey (Demir 2008).

Reptalus (Trepalus) cuspidatus (Fieber, 1876)

Oliarus cuspidatus Fieber, 1876: 215; Dlabola (1981).

Reptalus (Trepalus) cuspidatus; new subgenus in Emeljanov (1995).

Iran localities: Jahrom.

Extralimital distribution: Southern, Western and Eastern Europe, Turkey, Central Asia.

Reptalus (Trepalus) rufocarinatus (Kusnezov, 1937)

Oliarus quinquecostatus var. *rufocarinatus* Kusnetzov, 1937: 168 *Oliarus bitinctus* Dlabola, 1961a: 254; Dlabola (1981). *Reptalus bitinctus*; Dlabola (1985), Kheyri (1989), Mirzayans (1995). *Oliarus concolor* Fieber, 1876; Mirzayans et al. (1976).

Iran localities: Dasht, Esfahān, Gholhak, Ghom- Esfahān Rd, Ghom, Karaj.

Extralimital distribution: North Africa, Southern, Western and Eastern Europe, Turkey, Afghanistan, Central and Eastern Asia.

Comments: *Oliarus concolor* Fieber, 1876 is taxonomically unclear, and apparently often interpreted by various author in various manners without having studied the (type material). Dlabola (1988): 64 lists it as a separate species. Also *Oliarus bitinctus* Dlabola has had a mixed history, synonymized with *rufocarinatus* by Emeljanov (1978), revived as separate species in *Setapius* by Dlabola (1988) and now as a synonym of *rufocarinatus* in *Reptalus* (*Trepalus*) by Emeljanov (1995).

Reptalus eremicus Dlabola, 1985

Reptalus eremicus Dlabola, 1985: 114; Mirzayans (1995).

Iran localities: Marand, Mashhad, Masiri, Nagsh-e Rostam, Paskuh, Rafsanjān, Robāt-e Tork, Shirāz, Yāsuj.

Reptalus melanochaetus (Fieber, 1876)

Oliarus melanochaetus Fieber, 1876: 198; Dlabola (1981), Behdad (1988), Mirzayans (1995), Abaii (2000).

Reptalus melanochaetus; comb. n. in Emeljanov (1972a).

Iran localities: Bābol- Ghāemshahr, Karaj, Leshtar, Tonekābon.

Extralimital distribution: Southern, Western, and Eastern Europe, Turkey, Central Asia.

Reptalus quinquecostatus (Dufour, 1833)

Cixius quinquecostatus Dufour 1833: 224

Reptalus quinquecostatus; comb. n. in Emeljanov (1972b).

Oliarus quinquecostatus (Dufour 1833); Behdad (1988), Abaii (2000).

Iran localities: Unknown locality (Northern and central provinces).

Extralimital distribution: Southern, Western and Eastern Europe, Turkey, Central and Eastern Asia.

Comment: The identity of this record should be checked since it is part of a complex of several closely related species.

Reptalus reductus Dlabola, 1994

Reptalus reductus Dlabola, 1994: 53; Dlabola (1994). Iran localities: Bāft.

Reptalus ziaran Dlabola, 1985

Reptalus ziaran Dlabola, 1985: 118 Iran localities: Miāneh, Ziārān.

Setapius barajus (Dlabola, 1957)

Oliarus barajus Dlabola, 1957b: 24; Dlabola (1981), Mirzayans (1995) *Setapius barajus*; comb. n. in Dlabola (1988). Iran localities: Irānshahr, Varāmin.

Extralimital distribution: Transcaucasia, Turkey, Afghanistan.

Setapius lindbergi (Dlabola, 1957)

Oliarus lindbergi Dlabola, 1957b: 23; Linnavuori (1962a) [listed], Nast (1972) [listed], Dlabola (1972 [listed], 1981). *Setapius lindbergi*; comb. n. in Dlabola (1988), Kheyri (1989), Mirzayans (1995). *Reptalus lindbergi* (Dlabola, 1960); Kheyri (1989), Mirzayans (1995). Iran localities: Āb Āsk, Bampur, Fasā, Gorgān, Heirān, Jiroft, Marand, Marg-e-Malek, Sarāvān, Shirāz, Tochāl Mountain, Zābol.

Extralimital distribution: Afghanistan, Cyprus, Iraq, Israel, Turkey, Dagestan, Uzbekistan (Nast 1972) Egypt (Dlabola 1974b), Saudi Arabia (Dlabola 1980c).

Family Delphacidae

Subfamilies and tribes follow Asche (1985)

Subfamily Asiracinae

Asiraca clavicornis (Fabricius, 1794)

Cicada clavicornis Fabricius, 1794: 41; Karimzadeh Esfahani et al. (1998), Behdad (1988), Mirzayans (1995) Abaii (2000).

Asiraca flavicornis (sic); Dlabola (1981, 1984).

Iran localities: Esfahān province, Goleestān province, Karaj, Markazi province. Extralimital distribution: North Africa, Europe, Northern parts of Western Asia, Afghanistan, Central and Eastern Asia.

Subfamily Kelisinae

Kelisia praecox Haupt, 1935

Kelisia praecox Haupt, 1935: 133; Dlabola (1981), Lashkari et al. (2009) Iran localities: Tochāl Mountain, Golestān province. Extralimital distribution: Germany, Eastern Europe, Central and Eastern Asia.

Kelisia ribauti Wagner, 1938

Kelisia ribauti Wagner, 1938: 12; Dlabola (1981), Mirzayans (1995). Iran localities: Evin. Extralimital distribution: Western, Northern and Eastern Europe

Extralimital distribution: Western, Northern and Eastern Europe, North parts of Westsern Asia, Afghanistan, Central Asia, Eastern Asia and Russian Far East.

Subfamily Stenocraninae

Stenocranus major (Kirschbaum, 1868)

Delphax major Kirschbaum, 1868: 211 Stenocranus major; Dlabola (1981). Iran localities: Golestān province. Extralimital distribution: Europe, Afghanistan.

Subfamily Delphacinae

Tribe Tropidocephalini

Tropidocephala prasina Melichar, 1902

Tropidocephala prasina Melichar, 1902a: 90 Tropidocephala prasina lateralis Melichar, 1902a: 90; Nast (1972) [listed], Dlabola (1981) [listed]. Iran localities: Bazmān.

Tribe Delphacini

Bostaera bolivari (Melichar, 1901)

Delphacodes bolivari Melichar, 1901: 56 Pseudaraeopus bolivari (Melichar, 1901); Nast (1972) [listed]. Bostaera bolivari; Dlabola (1960a, 1981 [listed])., Mirzayans (1995). Iran localities: Chābahār, Geno, Irānshahr, Isin, Ziārat. Extralimital distribution: North West Africa, Southern Europe, Israel.

Calligypona revi (Fieber, 1866)

Delphax reyi Fieber, 1866: 527 *Calligypona revi*; Dlabola (1981). Iran localities: Bandar Anzali. Extralimital distribution: Europe, Turkey, Central Asia.

Changeondelphax velichkovskyi (Melichar, 1913)

Euidella velitchkovskyi Melichar, 1913: 6 (type species of Changeondelphax Kwon, 1982) Calligypona oriens Dlabola, 1961a: 276 (synonymized by Anufriev 1977). Chloriona oriens; Dlabola (1981). *Changeondelphax velichkovskyi*; comb. n. in Kwon (1982). Iran localities: Ghāemshahr. Extralimital distribution: South Eastern Europe, North Caucasus Eastern Asia.

Chloriona clavata Dlabola, 1960

Chloriona clavata Dlabola, 1960b: 1; Dlabola (1981). Iran localities: Gorgān, Evin. Extralimital distribution: Kyrgyzia (Novikov et al. 2006).

Chloriona unicolor (Herrich-Schäffer, 1835)

Delphax unicolor Herrich-Schäffer, 1835: 66 Chloriona unicolor; Melichar (1902a), Dlabola (1968 [listed], 1981), Mirzayans (1972), Nast (1972) [listed].

Iran localities: Aliābād (Ghom), Evin, Gambuyeh, Marand, Temin Extralimital distribution: North Africa, Europe, Turkey, Central Asia

Conomelus sp.

Conomelus sp.; Dlabola (1981). Iran localities: Tochāl Mountain.

Delphacodes audrasi Ribaut, 1954

Delphacodes audrasi Ribaut, 1954: 180

Delphacodes linnavuorii Le Quesne, 1960: 160; Dlabola (1981), Mirzayans (1995).

Iran localities: Bidruyeh, Gambuyeh, Minushahr, Sufiān.

Extralimital distribution: Southern parts of Eastern Europe, Southern Europe, Turkey, Israel.

Comments: Asche and Remane (1983) discuss the status of *Delphacodes mulsanti* Fieber and suggested that both *D. audrasi* and *D. linnavuorii* are likely to be synonyms of *mulsanti*.

Delphacodes ornatipennis (Haupt, 1927)

Megamelus ornatipennis Haupt, 1927: 9; Dlabola (1964) [listed], 1965c [listed], 1960a, 1972).

Delphacodes ornatipennis; Asche and Remane (1983).

Iran localities: Irānshahr, Shādegān.

Extralimital distribution: East Mediterranean, Afghanistan.

Comments: Likely to become a synonym of *D. mulsanti* after further investigation (M. Asche pers comm.).

Delphax inermis Ribaut, 1934

Delphax inermis Ribaut, 1934: 281; Dlabola (1977b [listed], 1994), Mirzayans (1995). Iran localities: Borāzjān, Gambuyeh, Konārdān, Minushahr.

Extralimital distribution: North West Africa, Southern and Western Europe, East Mediterranean, Afghanistan.

Dicranotropis hamata (Boheman, 1847)

Delphax hamata Boheman, 1847: 45
Dicranotropis hamata; Dlabola (1981).
Iran localities: Lajrān.
Extralimital distribution: North Africa, Europe, North Asia, Central Asia.

Euides caspiana (Dlabola, 1961)

Euidella caspiana Dlabola, 1961a: 268 *Euides caspiana*; Dlabola (1981). Iran localities: Bandar Anzali. Extralimital distribution: Dagestan (Nast, 1972), Turkey (Demir, 2008).

Falcotoya minuscula (Horváth, 1897)

Delphax minuscula Horváth, 1897b: 622
Calligypona minuscula (Horváth, 1897); Dlabola (1960a).
Toya minuscula (Horváth, 1897): Dlabola (1981), Dlabola (1972) [listed].
Falcotoya minuscula; Barkhordari et al. (1981) [listed].
Iran localities: Bandar Anzali, Irānshahr, Robāt-e Tork.
Extralimital distribution: North East Africa, France, Eastern Europe, Turkey, Af-

ghanistan and Central Asia.

Gravesteiniella boldi (Scott, 1870)

Liburnia boldi Scott, 1870: 68

Gravesteiniella boldi; Dlabola (1981).

Iran localities: Golestān province.

Extralimital distribution: Western, Northern and Estern Europe, Cyprus, Central Northern and Eastern Asia.

Halmyra aeluropodis (Emeljanov, 1964)

Calligypona aeluropodis Emeljanov, 1964: 9 *Halmyra aeluropodis*; Dlabola (1974a, 1981). Iran localities: Robāt-e Tork. Extralimital distribution: Kazakhstan (Nast, 1972), Greece (Drosopoulos et al.1983).

Herbalima eforiae (Dlabola, 1961)

Calligypona eforiae Dlabola, 1961b: 314 *Unkanodes eforiae*; comb. n. in Dlabola (1974a). *Herbalima eforiae*; comb. n. in Emeljanov (1977).
Iran localities: Eynvarzān.
Extralimital distribution: Romania, Kazakhstan (Nast ,1972) Kyrgyzia (Novikov et al. 2006).

Javesella pellucida (Fabricius, 1794)

Fulgora pellucida Fabricius, 1794: 7
Calligypona marginata (Fabricius, 1794); Dlabola (1960a).
Javesella pellucida; Dlabola (1981), Mirzayans (1995).
Iran localities: Anbarābād, Chābahār, Evin, EynvarzānIrānshahr, Sufiān.
Extralimital distribution: North of Africa, Europe, Turkey, Afghanistan, Central, Eastern and Northern Asia.

Laodelphax striatellus (Fallén, 1826)

Delphax striatellus Fallén, 1826: 75

Laodelphax striatellus (Fallén, 1826); Dlabola (1971a, 1981), Nast (1972) [listed], Ahmadi et al. (1986), Kheyri (1989), Mirzayans (1995), Karimzadeh Esfahani et al. (1998), Haghshenas and Khajehali (2000), Nematollahi and Khajehali (2000), Khajehali et al. (2001), Yarmand et al. (2006), Lashkari et al. (2009).

Laodelphax striatella; Mirzayans et al. (1976).

Iran localities: Ahvāz, Aliābād (Jahrom), Āmol, Asālem, Bājgāh, Bandar Anzali, Behshahr, Behshahr- Sāri, Bidruyeh, Bidzard, Chenārshāhijān, Dārāb, Darband, Dasht, Eskandari, Estahbān, Evin, Eynvarzān, Fasā, Galugāh- Eslāmābād, Gambuyeh, Gazanak, Gholhak, Gorgān, Hashtpar, Jahānābād Sepidān, Jiroft, Karaj, Marand, Miāneh- Gharechaman, Minushahr, Nurābād, Parehsar, Robāt-e gharebil- Dowlatābād, Sanandaj, Sāri-Bābolsar, Sarvestān, Shahdād, Shahr-e Kord, Shirāz, Sufiān, Tabriz, Veresk, Zarghān.

Extralimital distribution: North Africa, Europe, Western Asia, Afghanistan, Central, Northern and Eastern Asia.

Matutinus putoni (Costa, 1888)

Kelisia putoni Costa, 1888: 16 *Calligypona typhae* Lindberg, 1960: 15 (syn. Asche and Remane, 1982); Dlabola (1981) *Matutinus putoni*; Mirzayans (1995).

Iran localities: Anbarieh, Minushahr, Shush. Extralimital distribution: Italy, North Africa, Israel.

Muirodelphax amol Dlabola, 1981

Muirodelphax amol Dlabola, 1981: 154 Iran localities: Āmol.

Muirodelphax aubei (Perris, 1857)

Delphax aubei Perris, 1857: 170 Muirodelphax aubei; Dlabola (1981), Lashkari et al. (2009) Iran localities: Damāvand, Golestān province, Lajrān, Marand, Tochāl Mountain. Extralimital distribution: North Africa, Europe, Turkey, Central and Eastern Asia.

Pseudaraeopus curtulus Dlabola, 1960

Pseudaraeopus curtulus Dlabola, 1960a: 3; Dlabola (1981) [listed], Nast (1972) [listed]. Iran localities: Irānshahr.

Pseudaraeopus iranicus Dlabola, 1960

Pseudaraeopus iranicus Dlabola, 1960a: 4; Dlabola (1981) [listed], Nast (1972) [listed]. Iran localities: Irānshahr.

Ribautodelphax hyrcanus Dlabola, 1981

Ribautodelphax hyrcanus Dlabola, 1981: 157; Mirzayans (1995) Iran localities: Golestān province, Hashtpar.

Sardia rostrata Melichar, 1903

Sardia rostrata Melichar, 1903: 96; Dlabola (1981), Mirzayans (1995) Iran localities: Minushahr. Extralimital distribution: widespread in Asia and Australasia.

Sogatella furcifera (Horváth, 1899)

Delphax furcifera Horváth 1899: 372

Sogatella furcifera; Lashkari et al. (2009).

Iran localities: Golestān province.

Comments: Asche and Wilson (1990) recorded no specimens of this species in Europe or Africa. It is very likely that this record refers to another species.

Extralimital distribution: Eastern Palaearctic (Asche and Wilson, 1990).

Sogatella vibix (Haupt, 1927)

Liburnia vibix Haupt, 1927:13

Calligypona vibix (Haupt, 1927); Dlabola (1960a).

Sogatella suezensis Matsumura 1910; Linnavuori (1964) misdetermination (not *suezensis* Matsumura, 1910); Ahmadi et al. (1986), Kheyri (1989), Mirzayans (1995), Nast (1972) [listed]. *Sogatella vibix*; Dlabola (1965c, 1971a, 1972), Karimzadeh Esfahani et al. (1998), Lashkari et al. (2009).

Iran localities: Albāji, Anbarābād, Bājgāh, Bidruyeh, Dārāb, Esfahān province, Estahbān, Fasā, Firuzābād, Golestān province, Irānshahr, Isin, Jiroft, Minushahr, Nurābād, Sarvestān, Shabestar.

Extralimital distribution: North of Africa, North parts of Western Asia, Afghanistan.

Comments: Asche and Wilson (1990) provide full synonymy and localities.

Toya propinqua (Fieber, 1866)

Delphax propinqua Fieber, 1866: 525

Calligypona propinqua (Fieber, 1866); Dlabola (1960a).

Toya propinqua; Ahmadi et al. (1986), Dlabola (1981), Kheyri (1989), Lashkari et al. (2009); Mirzayans (1995), Nast (1972) [listed].

Iran localities: Āb Āsk, Anbarābād, Andimeshk, Bājgāh, Bandar Anzali, Eskandari, Evin, Eynvarzān, Fasā, Gholhak, Gorgān, Hashtpar, Irānshahr, Isin, Jiroft, Kāzerun, Marand, Minushahr, Rudbārak, Sanandaj, Sepidān, Shirāz, Tochāl Mountain, Zarghān Extralimital distribution: North Africa, Southern, Western and Eastern Europe, Western Asia, Afghanistan, Central Asia and Japan.

Unkanodes latespinosa (Dlabola, 1957)

Calligypona latespinosa Dlabola, 1957a: 275

Unkanodes latespinosa; Dlabola (1971a, 1981), Mirzayans (1995).

Iran localities: Damāvand, Eskandari, Evin, Gholhak, Marand, Miāneh – Gharechaman, Orumieh, Robāt-e gharebil, Sanandaj, Shahdād.

Extralimital distribution: Turkey, Afghanistan and Mongolia.

Unkanodes tanasijevici (Dlabola, 1965)

Elymodelphax tanasijevici Dlabola, 1965a: 658 *Unkanodes tanasijevici*; Ahmadi et al. (1986). Iran localities: Akbar ābād, Bājgāh, Zarghān.

Extralimital distribution: Turkey-Anatolia, Greece, Romania, (Asche,1982), Yugoslavia.

Family Derbidae

Tribe Cenchreini

Malenia isinica Dlabola, 1986

Malenia isinica Dlabola, 1986: 173 Iran localities: Isin.

Malenia masirica Dlabola, 1986

Malenia masirica Dlabola, 1986: 173 Iran localities: Dehbakri, Kermānshāh (Yazd province), Kushk, Masiri, Sheikh-e Mahalleh, Yāsuj.

Malenia sarmatica Anufriev, 1966

Malenia sarmatica Anufriev, 1966: 46; Dlabola (1981), Mirzayans (1995). Iran localities: Hashtpar, Lāhijān, Tonekābon, Zirāb. Extralimital distribution: Ukraine, Transcaucasia.

Tribe Zoraidini

Proutista jezeki Dlabola, 1981

Proutista jezeki Dlabola, 1981: 161 Iran localities: Rāsk.

Family Dictyopharidae

Subfamily Dictyopharinae

Subgenera of Dictyophara follow Emeljanov (2004)

Dictyophara (Ancylocrius) albata Dlabola & Heller, 1962

Dictyophara albata Dlabola & Heller, 1962: 2; Nast (1972) [listed], Dlabola (1981), Behdad (1988), Mirzayans (1995), Abaii (2000), Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Chāhkuh, Estahbān, Geno, Hichān, Irānshahr, Jahrom, Jiroft, Kahurestān, Kāzerun, Nikshahr, Sendarak.

Dictyophara (Ancylocrius) exoptata Dlabola & Heller, 1962

Dictyophara exoptata Dlabola & Heller, 1962: 1; Nast (1972) [listed], Mirzayans et al. (1976) Dlabola (1981), Mirzayans (1995), Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Aliābād (Jahrom), Chāhkuh, Dasht-e arjan, Geno, Irānshahr, Isin, Jiroft, Kahurestān, Kāzerun, Konārdān, NikshharSendarak, Shirāz.

Dictyophara (Chanithus) avocetta Oshanin, 1879

Dictyophara avocetta Oshanin, 1879: 131; Melichar, 1902a, Nast (1972) [listed], Dlabola (1981) [listed], Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Anārak. Extralimital distribution: Azerbaijan, Eastern parts of Central Asia.

Dictyophara (Chanithus) hastata Kusnezov, 1929

Dictyophara hastate Kusnezov, 1929; Mozaffarian and Emeljanov (2010) [listed].

Dictyophara (Chanithus) kazeruna Dlabola, 1986

Dictyophara kazeruna Dlabola, 1986: 179, Mozaffarian and Emeljanov (2010) [listed]. Iran localities: Kāzerun.

Dictyophara (Conopenchus) pazukii (Dlabola, 1984)

Philotheria pazukii Dlabola, 1984: 25 Iran localities: Kamandān.

Dictyophara (Dictyophara) asiatica Melichar, 1912

Dictyophara asiatica Melichar, 1912: 118; Dlabola (1981), Behdad (1988), Abaii (2000), Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Eynvarzān, Gazanak. Extralimital distribution: Northern parts of Western Asia.

Dictyophara (Dictyophara) europaea (Linnaeus, 1767)

Fulgora europaea Linnaeus, 1767: 704

Dictyophara europaea; Mirzayans et al. (1976), Dlabola (1981), Behdad (1988), Mirzayans (1995), Haghshenas and Khajehali (2000), Abaii (2000), Khajehali et al. (2001), Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Ahvāz, Bābol- Ghāemshahr, Bandar Anzali, Behshahr, Chahār mahāl- Bakhtiāri province, Eskandari, Fasā, Feizābād, Hafttappeh, Jahrom, Kāmyārānv, Kandovān (Māzandarān), Khorramābād, Marand, Mollāsāni, Tabriz, Tonekābons, Varāmin, Zirāb.

Extralimital distribution: North Africa, Western, Eastern and Southern Europe, northern parts of Western Asia, Afghanistan, southern parts of Central Asia and Eastern Asia.

Dictyophara (Euthremma) hoberlandti Dlabola, 1974

Dictyophara hoberlandti Dlabola, 1974a: 36; Dlabola (1981), Mirzayans et al. (1976), Behdad (1988), Mirzayans (1995), Abaii (2000), Mozaffarian and Emeljanov (2010) [listed].

Dictyophara (Euthremma) hoberlandti; Emeljanov (2004).

Iran localities: Eynvarzān, Gāvkoshak, Kāzerun, Orumieh, Sepidān, Sisakht.

Raivuna iranica (Linnavuori, 1962)

Dictyophara iranica Linnavuori, 1962a: 7; Nast (1972) [listed], Dlabola (1981), Mirzayans (1995), Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Anbarābād, Chāhkuh, Dālaki, Evin, Ghom Lake, Hafttappeh, Hasanlangi, Irānshahr, Isin, Jiroft, Kandovān (Māzandarān), Marand, Mollāsāni, Nikshahr, Pishin, Rāsk, Sendarak, Sisakht, Yazd, Zābol.

Extralimital distribution: Turkey (Demir, 2008).

Raivuna pallida (Donovan, 1800)

Fulgora pallida Donovan 1800: 1

Chanithus pallidus (= striata Oshanin, 1879); Dlabola (1960a).

Raivuna pallida; Mozaffarian and Emeljanov (2010) [listed].

Iran localities: Anbarābād, Bampur, Irānshahr

Extralimital distribution: North Africa, Southern parts of Western Asia, Transcaucasia, Central and Eastern Asia.

Comments: Linnavuori (1962a) regarded *Chanithus* as a subgenus of *Dictyophara*. Due to problems with the identity of this species literature records may refer to a number of species.

Raivuna striata (Oshanin, 1879)

Raivuna striata (Oshanin, 1879); Mozaffarian and Emeljanov (2010) [listed]. Iran localities: Unknown locality.

Subfamily Orgeriinae

Kumlika mandrita Emeljanov, 1997

Kumlika mandrita Emeljanov, 1997: 89 Iran localities: Sabzevār.

Nymphorgerius convergens Emeljanov, 1972

Nymphorgerius convergens Emeljanov, 1972a: 25; Dlabola (1979b). Iran localities: Shāhkuh-e Pāiin.

Nymphorgerius emeljanovi Dlabola, 1979

Nymphorgerius emeljanovi Dlabola, 1979b: 242 Iran localities: Tochāl Mountain, Ziārān.

Nymphorgerius mullah Dlabola, 1979

Nymphorgerius mullah Dlabola, 1979b: 241 Iran localities: Ziārān.

Nymphorgerius plotnikovi Kusnezov, 1929

Nymphorgerius plotnikovi Kusnezov, 1929: 326; Emeljanov (1997). Iran localities: Bojnurd. Extralimital distribution: Kazakhstan, Turkmenia (Nast, 1972).

Nymphorgerius rostratus Emeljanov, 2009

Nymphorgerius rostratus Emeljanov, 2009: 283 Iran localities: Sangrud.

Tigrahauda ototettigoides (Oshanin, 1913)

Orgerius ototettigoides Oshanin, 1913: 140 Tigrahauda ototettigoides; Emeljanov (1997). Iran localities: Esfarāyen, Kāhe. Extralimital distribution: Central Asia.

Family Flatidae

Bahuflata punctata Dlabola, 1979

Bahuflata punctata Dlabola, 1979a: 230 Iran localities: Bāhukalāt, Sirik.

Derisa atratula Melichar, 1902

Derisa atratula Melichar, 1902a: 103; Dlabola (1960a, 1981), Nast (1972) [listed], Mirzayans (1995), Medler (2003).

Iran localities: Sirjān, Isin, Anbarābād, Bampur, Bazmān, Nikshahr, Bāhukalāt, Chābahār, Tis.

Eurima astuta Melichar, 1902

Eurima astuta Melichar, 1902a: 102; Dlabola (1981) [listed], Nast (1972) [listed], Mirzayans (1995).

Iran localities: Bazmān, Kahurestān, Sendarak. Extralimital distribution: Israel (Nast 1972) Saudi Arabia (Dlabola 1979d).

Mesophantia kanganica Dlabola, 1983

Mesophantia kanganica Dlabola, 1983: 469 in Krampl and Dlabola (1983).

Iran localities: Bānuchārehar, Bojnurd, Hessār, Jiroft, Kangān, Nehbandān, Nosratābad, Sabzevār.

Mesophantia pallens Melichar, 1902

Mesophantia pallens Melichar, 1902b: 18; Abaii (2000), Barkhordari et al. (1981), Dlabola (1981, 1983a), Medler (2003), Mirzayans (1995), Mirzayans et al. (1976), Nast (1972) [listed], Rajabi (1989).

Iran localities: Ābādeh, Bojnurd, Darpahn, Estahbān, Evin, Gāvkoshak, Izadkhāst, Jahrom, Kāshān, Kāzerun, Khosro- Shirin, Mashhad-e Ardehāl, Mohammadābād, Nishābur, Rafsanjān, Sangān, Shāhrud, Sisakht, Yāsuj.

Mesophantia sabzevaranica Dlabola, 1983

Mesophantia sabzevaranica Dlabola, 1983: 466 in Krampl and Dlabola, (1983); Mirzayans (1995).

Iran localities: Denā mt (west slope), Doborji, Estahbān, Ferdows-e Esfandaghe, Gāvkoshi, Geno, Ghaderābād, Jahrom, Jiroft, Kāzerun, Kermān, Kuhenjān, Māhārlu, Miānjangal, Mohammadābād, Nishābur, Posht- e kuh, Sarāvān, Sarbāz, Shul, Sisakht.

Mesophantia tisina Dlabola, 1983

Mesophantia tisina Dlabola, 1983: 468 in Krampl and Dlabola (1983). Iran localities: Dehbakri, Tis.

Persepolia columbaria Dlabola & Safavi, 1972

Persepolia columbaria Dlabola & Safavi, 1972: 2; Dlabola and Safavi (1972), Dlabola (1974a, 1981), Mirzayans et al. (1976), Behdad (1988), Rajabi (1989), Mirzayans (1995), Abaii (2000).

Iran localities: Kāzerun, Nurābād, Shirāz, Yāsuj. Extralimital distribution: Saudi Arabia (Dlabola, 1980c).

Persepolia jasmuriana Dlabola, 1982

Persepolia jasmuriana Dlabola, 1982b: 165

Iran localities: Bāhukalāt, Bārgāh, Bazmān, Geno, Isin, Rudān, Sarāvān, Tang-e Sarheh.

Persepolia secunda Dlabola, 1981

Persepolia secunda Dlabola, 1981: 190 Iran localities: Zāboli.

Persepolia servadeina Dlabola, 1982

Persepolia servadeina Dlabola, 1982b: 163

Iran localities: Bazmān, Zāboli, Pākuh, Sarāvān, Tang-e Sarheh, Zāhedān, Kermānshāh (Yazd province), Khutanābād, Kermān, Māhān, Golbāf.

Phantia borazianica Dlabola, 1989

Phantia borazianica Dlabola, 1989: 44 Iran localities: Borāzjān.

Phantia christophii Rusiecka, 1902

Phantia christophii Rusiecka, 1902: 423; Melichar (1902a), Dlabola (1972), Nast (1972) [listed].

Iran localities: Temin. Extralimital distribution: Afghanistan, Turkmenia.

Phantia crucispina Dlabola, 1989

Phantia crucispina Dlabola, 1989: 42 Iran localities: Zāhedān.

Phantia cylindricornis Melichar, 1902

Phantia cylindricornis Melichar, 1902a: 99; Dlabola (1960a, 1972 [listed], 1989), Nast (1972) [listed], Medler (2003).

Iran localities: Anbarābād, Bāhukalāt, Chābahār, Espakeh, Hasanlangi, Irānshahr, Mināb, Ziārat.

Extralimital distribution: Afghanistan and Turkmenia.

Phantia denasuta Dlabola, 1989

Phantia denasuta Dlabola, 1989: 40 Iran localities: Bāhukalāt, Bampur, Bandar Chārak, Hasanlangi, Jāsk.

Phantia ferganensis Dubovsky, 1966

Phantia ferganensis Dubovsky, 1966: 75; Dlabola (1989). Iran localities: Bāhukalāt, Bampur, Jāsk. Extralimital distribution: Uzbekistan.

Phantia finita Dlabola, 1989

Phantia finita Dlabola, 1989: 49 Iran localities: Miānjangal.

Phantia flavida Rusiecka, 1902

Phantia flavida Rusiecka, 1902: 424; Nast (1972) [listed], Dlabola (1981) [listed]. Iran localities: Unknown locality. Extralimital distribution: Azerbaijan.

Phantia helleri Linnavuori, 1962

Phantia helleri Linnavuori, 1962b: 2; Nast (1972) [listed], Dlabola (1981, 1989), Mirzayans (1995).

Iran localities: Ahram, Bāhukalāt, Bampur, Bandar Abbās, Bandar Lengeh, Bārgāh, Bazmān, Dārāb, Dārbahāre, Darzin, Darpahn, Fāriāb, Gambuyeh, Ganāveh, Gāvkoshi, Geno, Ghasr-e Ghand, Ghom, Ghotbābād, Hājiābād, Hasanlangi, Isin, Jāsk, Jiroft, Kahnuj, Kahurestān, Kamālābād, Khutanābād, Mahārlu, Mijān, Mināb, Nikshahr, Nugh, Rafsanjān, Rāsk, Rudān, Sekand, Sendarak, Shirāz, Shul, Shusf, Sirik, Tang-e Sarheh, Tis, Zeidān.

Phantia lactea Rusiecka, 1902

Phantia lactea Rusiecka, 1902: 424; Nast (1972) [listed], Dlabola (1981) [listed]. Iran localities: Unknown locality.

Phantia ovatospina Dlabola, 1989

Phantia ovatospina Dlabola, 1989: 43 Iran localities: Nehbandān.

Phantia picea Dlabola, 1989

Phantia picea Dlabola, 1989: 46 Iran localities: Bāhukalāt, Bandar Abbās, Jāsk, Nikshahr.

Phantia putoni Rusiecka, 1902

Phantia putoni Rusiecka, 1902: 423; Dlabola (1981) [listed]. Iran localities: Unknown locality.

Phantia rubromarginata Rusiecka, 1902

Phantia rubromarginata Rusiecka, 1902: 424; Nast (1972) [listed], Dlabola (1972 [listed], 1981 [listed]).
Iran localities: Unknown locality.
Extralimital distribution: Afghanistan.

Phantia subquadrata (Herrich Schäffer, 1838)

Poeciloptera subquadrata Herrich Schäffer, 1838: 2 Phantia subquadrata; Dlabola (1989). Iran localities: Yāsuj. Extralimital distribution: Turkey (Demir 2007).

Phantia viridula Puton, 1890

Phantia viridula Puton, 1890: 230; Dlabola (1972) [listed], Nast (1972) [listed], Puton (1890).

Iran localities: Shāhrud. Extralimital distribution: Armenia.

Tisia esfandiarii Dlabola, 1981

Tisia esfandiarii Dlabola, 1981: 192 Iran localities: Tis.

Zarudnya fusca Melichar, 1902

Zarudnya fusca Melichar, 1902a: 101; Dlabola (1960a), Nast (1972) [listed], Medler (2003).

Iran localities: Bampur, Bazmān, Irānshahr. Extralimital distribution: Turkmenia.

Zarudnya interstitialis Melichar, 1902

Zarudnya interstitialis Melichar, 1902a: 101; Dlabola (1960a, 1981 [listed]), Nast (1972) [listed], Medler (2003), Mirzayans (1995)

Iran localities: Anbarābād, Bampur, Bazmān, Irānshahr, Jiroft, Kharposht, Sendarak.

Extralimital distribution: North Africa.

Family Fulgoridae

The genus *Dorysarthrus* was transferred from Dictyopharidae to Fulgoridae by Emeljanov (1979).

Dorysarthrus mobilicornis Puton, 1895

Dorysarthrus mobilicornis Puton, 1895: 88; Dlabola (1981), Mirzayans (1995). Iran localities: Bāhukalāt, Ghasr-e Ghand, Nikshahr, Sirik. Extralimital distribution: Israel.

Dorysarthrus simonyi Melichar, 1912

Dorysarthrus simonyi Melichar, 1912; Dlabola 1984, Mirzayans (1995). Iran localities: Mināb. Extralimital distribution: Saudi Arabia (Dlabola. 1979d).

Dorysarthrus sumakovi Oshanin, 1908

Dorysarthrus sumakovi Oshanin, 1908: 471; Nast (1972) [listed]. Iran localities: Unknown locality. Extralimital distribution: Turkmenia.

Family Issidae Spinola, 1839

Anatalodus karabachicus (Logvinenko, 1975)

Aeluropsis karabachica Logvinenko, 1975: 59
Hysteropterum ignavum Dlabola, 1981: 179; Dlabola (1981).
Anatolodus ignavus; Dlabola (1982a), Mirzayans (1995).
Anatalodus karabachicus; Gnezdilov (2010b) [listed].
Iran localities: Ghezel-Bolāgh, Māku, Khoy, Marand, Nikshahr.
Extralimital distribution: Turkey.

Cavatorium ardakanum Dlabola, 1980

Cavatorium ardakanum Dlabola, 1980b: 210 Iran localities: Shul.

Cavatorium bispinatum Dlabola, 1980

Cavatorium bispinatum Dlabola, 1980b: 209 Iran localities: Sisakht, Yāsuj.

Cavatorium quadrispinatum Dlabola, 1980

Cavatorium quadrispinatum Dlabola, 1980b: 211 Iran localities: Doborji, Fasā.

Cavatorium sarbaz Dlabola, 1980

Cavatorium sarbaz Dlabola, 1980b: 212 Iran localities: Sekand.

Eusarima (Nepalius) iranica Gnezdilov & Mozaffarian, 2011

Eusarima (Nepalius) iranica Gnezdilov & Mozaffarian, 2011: 457 Iran localitities: Dārābād, Tehrān.

Inflatodus astyages Dlabola, 1982

Inflatodus astyages Dlabola, 1982a: 124 Iran localities: Ziārān.

Inflatodus kyaxares Dlabola, 1982

Inflatodus kyaxares Dlabola, 1982a: 122 Iran localities: Ziārān.

Inflatodus persicus (Dlabola, 1981)

Hysteropterum persicum Dlabola, 1981: 181 Inflatodus persicus; Mirzayans (1995). Iran localities: Ābyek, Ghazvin, Tochāl Mountain, Zanjān.

Inflatodus viridans (Dlabola, 1974)

Hysteropterum viridatum Dlabola, 1971a: 380 [Preoccupied] *Hysteropterum viridans* Dlabola, 1974a: 44 (nom. nov. for *H. viridatum* Dlabola, 1971: 380 nec Caldwell, 1945); Dlabola (1974a [listed], 1981 [listed]). *Inflatodus viridans*; Mirzayans (1995).

Iran localities: Ghezel-Bolāgh, Varāmin.

Iranodus amygdalinus Dlabola, 1980

Iranodus amygdalinus Dlabola, 1980b: 205; Behdad (1988), Mirzayans (1995), Abaii (2000).

Iran localities: Bānuchārehar, Chābahār, Doborji, Ferdows-e Esfandaghe, Gotbābād, Hoseinābād, Isin, Jiroft, Kāzerun, Miānjangal, Mohammadābād.

Iranodus dumetorus (Dlabola, 1981), comb. n.

Hysteropterum dumetorum Dlabola 1979 nomen nudum. Species not listed or described *Iranodus dumetorus* (Dlabola, 1980b); 205 nomen nudum. Species listed but not described.

Hysteropterum dumetorum Dlabola, 1981: 185

Iran localities: Khānehkhoreh, Shirāz, Dehbid.

Comments: The species was validly described as *Hysteropterum dumetorum* in Dlabola (1981). However, the name appeared as a new combination in Dlabola (1980b) based on the species being described in Dlabola (1979), where in fact it was not mentioned. On this basis the two earlier citations of the name must be nomina nuda. But the new combination given as *Iranodus dumetorus* was 2 years before the species was described. Accordingly we validate the name here in the combination *Iranodus dumetorus* (Dlabola, 1981).

Iranodus khatunus (Dlabola, 1981), comb. n.

Hysteropterum khatunum Dlabola, 1979 nomen nudum. Species not listed or described *Iranodus khatunus* Dlabola, 1979: Mirzayans (1995).

Iranodus khatunus (Dlabola, 1980b); 205 nomen nudum. Species listed but not described Mirzayans (1995).

Hysteropterum khatunum Dlabola, 1981: 187

Iran localities: Bānuchārehar, Dehpābid.

Comments: The species was validly described as *Hysteropterum khatunum* in Dlabola (1981). However, the name appeared as a new combination in Dlabola (1980b) based on the species being described in Dlabola (1979), where in fact it was not mentioned. On this basis the two earlier citations of the name must be *nomina nuda*. But the new combination given as *Iranodus khatunus* was 2 years before the species was described. Accordingly we validate the name here in the combination *Iranodus khatunus* (Dlabola, 1981).

Iranodus nishabur Dlabola, 1982

Iranodus nishabur Dlabola, 1982a: 128 Iran localities: Birjand, Nishābur.

Iranodus repandus (Dlabola, 1981), comb. n.

Hysteropterum repandum Dlabola, 1979 nomen nudum. Species not listed or described. *Iranodus repandus* Dlabola, 1979; Mirzayans (1995).

Iranodus repandum (Dlabola, 1980b): 207 nomen nudum. Species listed but not described. *Hysteropterum repandum* Dlabola, 1981: 183; Abaii (2000), Behdad (1988).

Iran localities: Jahrom, Kāzerun, Shirāz.

Comments: The species was validly described as *Hysteropterum repandus* in Dlabola (1981). However, the name appeared as a new combination in Dlabola (1980b) based on the species being described in Dlabola (1979), where in fact it was not

mentioned. On this basis the two earlier citations of the name must be nomina nuda. But the new combination given as *Iranodus repandus* was 2 years before the species was described. Accordingly we validate the name here in the combination *Iranodus repandus* (Dlabola 1981).

Iranodus transversalis Dlabola, 1980

Iranodus transversalis Dlabola, 1980b: 204; Mirzayans (1995) Iran localities: Bāft, Lalezār.

Mycterodus

This large issid genus is now often treated species placed in subgenera such as *Aconosimus*. But not all *Mycterodus* species have yet been placed in subgenera, so those species found in Iran are listed under *Mycterodus* only.

Mycterodus astragalicus Dlabola, 1974

Mycterodus astragalicus Dlabola, 1974a: 42; Behdad (1988); Abaii (2000). *Aconosimus astragalicus*; Dlabola (1997).

Iran localities: Kuhrang.

Mycterodus demavendinus Dlabola, 1981

Mycterodus demavendinus Dlabola, 1981: 174; Behdad (1988), Abaii (2000). Aconosimus demavendinus; Dlabola (1997). Iran localities: Damāvand, Tochāl Mountain.

Mycterodus elbursicus (Logvinenko, 1974)

Mycterodus elbursicus Logvinenko, 1974: 838 Aconosimus elbursicus; Dlabola (1983b), Dlabola (1997). Iran localities: Shāhkuh-e Pāiin.

Mycterodus fagetophilus Dlabola, 1980

Mycterodus fagetophilus Dlabola, 1980a: 62, Dlabola (1997). Iran localities: Chālus.

Mycterodus guilanicus Dlabola, 1981

Mycterodus guilanicus Dlabola, 1981: 176, Dlabola (1997). Iran localities: Lāhijān.

Mycterodus hezarmeshedi Dlabola, 1980

Mycterodus hezarmeshedi Dlabola, 1980a: 66 Aconosimus hezarmeshedi; Dlabola (1997). Iran localities: Kuh-e Hezār.

Mycterodus inassuetus Dlabola, 1981

Mycterodus inassuetus Dlabola, 1981: 176 Aconosimus inassuetus; Dlabola (1997). Iran localities: Gazanak.

Mycterodus kandavanicus Dlabola, 1980

Mycterodus kandavanicus Dlabola, 1980a: 63, Dlabola (1997). Iran localities: Ghazvin, Kandovān (Māzandarān).

Mycterodus krameri Dlabola, 1974

Mycterodus krameri Dlabola, 1974b: 297; Logvinenko (1974), Dlabola (1981), Dlabola (1997) [listed], Abaii (2000), Gnezdilov et al. (2004), Lashkari et al. (2009). Iran localities: Behshahr, Bojnurd, Dasht, Gorgān, Rostamābād.

Mycterodus lanceatus Dlabola 1997

Aconosimus lanceatus Dlabola 1997: 307 Mycterodus lanceatus Dlabola 1991, nomen nudum [The species appears not to be formally described in 1991]. Iran localities: Kelärdacht, Budhär

Iran localities: Kelārdasht, Rudbār.

Mycterodus peterseni Dlabola, 1980

Mycterodus peterseni Dlabola, 1980a: 68, Dlabola (1997) Iran localities: Kandovān (Māzandarān).

Mycterodus sexpunctatus Dlabola, 1980

Mycterodus sexpunctatus Dlabola, 1980a: 67; Gnezdilov et al. (2004). *Aconosimus sexpunctatus*; Dlabola (1997). Iran localities: Azādbar, Kandovān (Māzandarān).

Mycterodus shahrudicus Dlabola, 1980

Mycterodus shahrudicus Dlabola, 1980a: 65 *Aconosimus shahrudicus*; Dlabola (1997). Iran localities: Ghazvin.

Pentissus bamicus Dlabola, 1980

Pentissus bamicus Dlabola, 1980b: 207; Mirzayans (1995). Iran localities: Dehbakri, Dehpābid, Jiroft, Khutanābād, Taftān, Tamandān.

Phasmena adyoungi Dlabola, 1982

Phasmena adyoungi Dlabola, 1982a: 131 Iran localities: Ghasr-e Ghand.

Phasmena nasuta Melichar, 1902

Phasmena nasuta Melichar, 1902a: 93; Melichar (1906), Nast (1972) [listed], Dlabola (1981) [listed].
Iran localities: Temin.

Extralimital distribution: Turkey.

Phasmena telifera Melichar, 1902

Phasmena telifera Melichar, 1902a: 92; Melichar (1906), Nast (1972) [listed], Dlabola (1981) [listed], Mirzayans (1995).
Iran localities: Sekand, Temin.

Quadriva aurita (Dlabola, 1982)

Hysterodus auritus Dlabola, 1982a: 162 *Quadriva aurita* (Dlabola, 1982a); comb. n. in Gnezdilov et al. (2004). Iran localities: Yāsuj.

Quadriva dehbakrina (Dlabola, 1980)

Hysterodus dehbakrinus Dlabola, 1980b: 196 *Quadriva dehbakrina*; comb. n. in Gnezdilov et al. (2004). Iran localities: Dehbakri.

Quadriva lassa (Dlabola, 1981)

Hysteropterum lassum Dlabola, 1981: 178 Quadriva lassa (Dlabola, 1981): comb. n. in Gnezdilov et al. 2004. Iran localities: Khānehkhoreh, Marg-e Malek, Shirāz.

Quadriva ochaninei (Puton, 1890)

Conosimus ochaninei Puton, 1890: 232 Conosimus oshanini; Nast (1972) [listed]. Quadriva ochaninei; comb. n. in Gnezdilov (2010b). Iran localities: Unknown locality. Extralimital distribution: Central Asia. Comment: Although the species had been listed in Nast (1972) from Iran, according to Gnezdilov (2010b) the species has only been collected from Central Asia.

Quadriva proxima (Dlabola, 1980)

Hysterodus proximus Dlabola, 1980b: 198 *Quadriva proxima*; comb. n. in Gnezdilov et al. (2004). Iran localities: Dehbakri.

Quadriva sabzevarana (Dlabola, 1980)

Hysterodus sabzevaranus Dlabola, 1980b: 193 *Quadriva sabzevarana*; comb. n. in Gnezdilov et al. (2004). Iran localities: Jiroft.

Quadriva taftanica (Dlabola, 1980)

Hysterodus taftanicus Dlabola, 1980b: 197 *Quadriva taftanica*; comb. n. in Gnezdilov et al. (2004). Iran localities: Taftān.

Quadriva tangesarhena (Dlabola, 1980)

Hysterodus tangesarhenus Dlabola, 1980b: 193 Hysterodus tangesarhensis; Mirzayans (1995). Quadriva tangesarhena; comb. n. in Gnezdilov et al. (2004). Iran localities: Semirom, Sepidān, Tang-e Sarheh.

Scorlupaster asiaticum (Lethierry, 1878)

Hysteropterum asiaticum Lethierry, 1878: 27; Dlabola (1981). *Hysteropterum tshurtshurnum* Linnavuori 1957: 96 *Scorlupaster asiaticum*; comb. n. in Emeljanov (1972b).

Iran localities: Shirāz.

Extralimital distribution: Syria, Transcaucasia, Afghanistan, Central and Northern Asia.

Scorlupaster emersum (Dlabola, 1981)

Hysteropterum emersum Dlabola, 1981: 184 Scorlupella emersum: Mirzayans (1995). Iran localities: Robāt-e gharebil.

Scorlupella montana (Becker, 1865)

Issus montana Becker, 1865

Scorlupella montana; Mozaffarian and Gnezdilov (in press).

Iran localities: Ālmeh, Dasht, Kandovān (Āzarbāijān-e Sharghi), Matnagh, Tāzekand, Yāichi.

Extralimital distribution: South Europe, Caucasus, Central Asia, Turkey, East Mediterranean (except North Africa).

Tautoprosopa transcaspia (Emeljanov, 1978)

Brachyprosopa transcaspia Emeljanov, 1978: 332

Verticisium pictifrons (Melichar, 1906), sensu Dlabola (1979c), misidentification (see Gnezdilov 2002).

Tautoprosopa transcaspia; Gnezdilov (2002), Gnezdilov et al. (2004).

Iran localities: Dareh gaz, Khalkānlu, Khargh, Māzandarān, Shurlukh. Extralimital distribution: Kazakhstan, Turkmenia (Gnezdilov 2002).

Family Kinnaridae

Perloma boroumandi (Dlabola, 1981)

Adolenda boroumandi Dlabola, 1981: 138; Mirzayans (1995). Perloma boroumandi; Emeljanov (1984). Iran localities: Bāhukalāt, Geno, Ghasr-e Ghand, Nikshahr. Comment: Listed as Cixiidae in Dlabola (1981).

Perloma brunnescens (Emeljanov, 1984)

Propleroma brunnescens Emeljanov, 1984: 471
Perloma brunnescens; Wilson 2010a.
Iran localities: Kāravāndar.
Extralimital distribution: UAE (Wilson, 2010a).

Perloma satrapa (Dlabola, 1981)

Adolenda satrapa Dlabola, 1981: 140 Perloma satrapa; Emeljanov (1984). Iran localities: Ghasr-e Ghand, Kāzerun. Comment: Listed as Cixiidae in Dlabola (1981).

Perloma zarudnyi (Emeljanov, 1984)

Propleroma zarudnyi Emeljanov, 1984: 474 Perloma zarudnyi; Wilson (2010a). Iran localities: Kāravāndar.

Family Lophopidae

Lophops pallidus Melichar, 1902

Lophops pallidus Melichar, 1902a: 90 Lophops pallida; Nast (1972) [listed], Dlabola (1981) [listed]. Iran localities: Anārak. Extralimital distribution: Saudi Arabia (Dlabola, 1980c).

Family Meenoplidae

Anigrus farsicus Dlabola, 1986

Anigrus farsicus Dlabola, 1986: 171 Iran localities: Borāzjān, Rāsk.

Meenoplus albosignatus Fieber, 1866

Meenoplus albosignatus Fieber, 1866: 510; Dlabola (1984), Mirzayans (1995).
 Iran localities: Sepidān.
 Extralimital distribution: Southern Europe, Caucasus, North parts of Western Asia.

Nisia nervosa (Motschulsky, 1863)

Meenoplus atrovenosus Lethierry, 1888: 466 Nisia nervosa; Mirzayans (1995).

Iran localities: Isin.

Extralimital distribution: widely distributed from subtropical and tropical regions of Old World. North of Africa, North parts of Western Asia, UAE and Eastern Asia.

Family Nogodinidae

Hadjia nerii Dlabola, 1981

Hadjia nerii Dlabola 1981: 197 Iran localities: Isin.

Hadjia quadrifasciata Dlabola, 1981

Hadjia quadrifasciata Dlabola 1981: 195 Iran localities: Doborji, Geno, Ghotbābād, Hājiābād.

Iranissus ephedrinus Dlabola, 1980, new placement

Iranissus ephedrinus Dlabola, 1980b: 201

Iran localities: Bānuchārehar, Ferdows-e Esfandaghe, Geno, Hājiābād, Jiroft, Mohammadābād, Tis.

Comment: This species was described and placed in Issidae, We here transfer the species to Nogodinidae. (Suggested transfer by V. Gnezdilov pers. comm. and confirmed by MRW by examination of specimens).

Morsina persica Melichar, 1902

Morsina persica Melichar, 1902a: 98; Nast (1972) [listed], Dlabola (1981), Mirzayans (1995).

Iran localities: Bazmān, Geno, Gorgān, Jāsk, Jiroft, Mumān, Sendarak.

Philbyella glarea Dlabola & Heller, 1962

Philbyella glarea Dlabola & Heller, 1962: 2; Dlabola (1981), Mirzayans (1995), Nast (1972) [listed].

Iran localities: Bandar Abbās, Chābahār, Fāriāb, Kahnuj, Sirjān, Tang-e Sarheh.

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Family Ricaniidae

Pochazia umbrata Melichar, 1896

Pochazia umbrata Melichar, 1896: 385; Nast (1972) [listed]. Iran localities: Unknown locality. Extralimital distribution: Turkmenia, Russia (Siberia) (Nast, 1972).

Ricania hedenborgi Stål, 1865

Ricania hedenborgi Stål, 1865: 162; Mirzayans et al. (1976), Dlabola (1981, 1983a [listed], 1984), Mirzayans (1995), Haghshenas and Khajehali (2000)

Iran localities: Chahārmahāl- Bakhtiāri province, Kāzerun, Shirāz, Sarvestān, Aliābād (Jahrom), Hamadān, Asadābād, Izeh.

Extralimital distribution: Greece, Turkey, Armenia.

Ricania soraya Dlabola, 1983

Ricania soraya Dlabola, 1983a: 93 Iran localities: Ghaderābād, Kāzerun, Masiri.

Family Tettigometridae

Eurychila pantherina (Horváth, 1891)

Tettigometra pantherina Horváth, 1891: 81 *Eurychila pantherina*; Dlabola (1981, 1984), Mirzayans (1995). Iran localities: Meshkinshahr, Minushahr. Extralimital distribution: Transcaucasus, Afghnistan and Central Asia.

Mitricephalus macrocephalus (Fieber, 1865)

Tettigometra macrocephalus Fieber, 1865: 569 *Mitricephalus macrocephalus*; Dlabola (1981). Iran localities: Kandovān (Māzandarān), Lajrān, Tochāl Mountain. Extralimital distribution: Europe, Turkey, Afghanistan, Central and Northern Asia.

Tettigometra angulata Lindberg, 1948

Tettigometra angulata Lindberg, 1948: 19; Dlabola (1972) [listed], Nast (1972) [listed], Mirzayans (1995).

Iran localities: Evin, Kāshān, Rafsanjān.

Extralimital distribution: Transcaucasia, Turkey, Israel, Afghanistan and Central Asia.

Tettigometra costulata Fieber, 1865

Tettigometra costulata Fieber, 1865: 572; Melichar (1902a), Dlabola (1971a, 1972, 1981), Nast (1972) [listed], Behdad (1988), Rajabi (1989), Mirzayans (1995), Abaii (2000).

Iran localities: Aliābād (Ghom), Bazmān, Evin, Gazanak, Geno, Ghom, Karaj, Kuhrang, Māku, Marand, Mohammadābād.

Extralimital distribution: North Africa, Southern Europe, Yugoslavia, Transcaucasia, Northern parts of Western Asia, Afghanistan, Central Asia.

Tettigometra demavenda Dlabola, 1981

Tettigometra demavenda Dlabola, 1981: 170; Dlabola (1981). Iran localities: Gazanak, Lajrān.

Tettigometra depressa Fieber, 1865

Tettigometra depressa Fieber, 1865: 563; Nast (1972) [listed].

Iran localities: Unknown locality.

Extralimital distribution: North Africa, Southern, Western and Eastern Europe, North West of Asia.

Tettigometra eremi Lindberg, 1948

Tettigometra eremi Lindberg, 1948: 27; Nast (1972) [listed], Dlabola (1972, 1981), Mirzayans (1995).

Iran localities: Ābyek, Āmol, Eynvarzān, Gazanak, Kandovān (Māzandarān), Lajrān, Robāt-e gharebil.

Extralimital distribution: Ukraine, Transcaucasia, Turkey, Afghnistan and Central Asia.

Tettigometra hexaspina Kolenati, 1857

Tettigometra hexaspina Kolenati 1857: 428; Dlabola (1984) Iran localities: Kāzerun, Khoy.

Tettigometra pseudovitellina Mitjaev, 1971

Tettigometra pseudovitellina Mitjaev, 1971: 75; Dlabola (1981) Iran localities: Āb Āsk, Marand. Extralimital distribution: Kazakhstan (Nast, 1972).

Tettigometra sordida Fieber, 1865

Tettigometra sordida Fieber, 1865: 571; Nast (1972) [listed]. Iran localities: Unknown locality. Extralimital distribution: Western and Eastern Europe.

Tettigometra sororcula Horváth, 1897

Tettigometra sororcula Horváth 1897a: 90; Dlabola 1994 [listed]. Iran localities: unknown locality. Extralimital distribution: Southern Europe.

Tettigometra sulphurea Mulsant & Rey, 1855

Tettigometra sulphurea Mulsant & Rey, 1855: 209; Mirzayans et al. (1976), Dlabola (1981), Mirzayans (1995).

Iran localities: Ābyek, Dārān, Delijān, Evin, Gāvkoshak, Geno, Ghazvin, Ghom-Esfahān Rd, Kandovān (Āzarbāijān-e Sharghi), Karaj, Rudehen.

Extralimital distribution: Southern, Western and Eastern Europe, North parts of Western Asia, Afghanistan and Central Asia.

Tettigometra varia Fieber, 1865

Tettigometra varia Fieber, 1865: 565; Dlabola (1981), Mirzayans (1995). Iran localities: Ghom, Robāt-e gharebil. Extralimital distribution: Eastern Europe, Jordan, Afghanistan and Central Asia.

Tettigometra vitellina Fieber, 1865

Tettigometra vitellina Fieber, 1865: 566; Barkhordari et al. (1981) [listed], Dlabola (1981), Mirzayans (1995), Mirzayans et al. (1976), Nast (1972) [listed].

Iran localities: Damāvand, Dasht-e arjan, Evin, Firuzkuh, Gazanak, Ghazvin, Kandovān (Māzandarān), Karaj, Māku, Marand, Robāt-e Tork, Saādatshahr, Tehrān.

Extralimital distribution: Yugoslavia, Transcaucasia, Northern parts of Western Asia, Afghanistan and Central Asia.

Family Tropiduchidae

Kazerunia leguaniforma Dlabola, 1977

Kazerunia leguaniforma Dlabola, 1977a: 164; Dlabola (1977a, 1981[listed]). Iran localities: Chābahār.

Kazerunia ochreata Dlabola, 1974

Kazerunia ochreata Dlabola, 1974a: 41; Dlabola (1977a). Iran localities: Fasā, Kāzerun.

Kazerunia undulata Dlabola, 1977

Kazerunia undulata Dlabola, 1977a: 166 Iran localities: Ghasr-e Ghand, Sarbāz.

Ommatissus lybicus Bergevin, 1930

Ommatissus binotatus Fieber, 1876; Afshar (1937), Gardenhire (1959) [listed], Farahbakhsh (1961), Gharib (1998), Dlabola (1994), Mirzayans (1995), Abaii (2000). *Ommatissus binotatus lybicus* Bergevin 1930: 20; Gharib (1966), Behdad (1988, 1991).

Iran localities: Bāfgh Kahkom, Bam, Bandar Abbās, Geno, Isin, Jahrom, Jandagh, Khorramshahr, Khur va Biābānak, Mehrān, Shahdād, Tabas.

Comments: Asche and Wilson (1989) recognized the variety *lybicus* as a distinct species.

Extralimital distribution: Middle East (Asche and Wilson 1989), Saudi Arabia (Dlabola, 1979d), UAE (Wilson 2010b).

Trypetimorpha fenestrata Costa, 1862

Trypetimorpha fenestrata Costa, 1862: 60; Dlabola (1981), Mirzayans (1995). Specimens from Iran do not conform exactly to the male genitalia structure illustrated

in Huang and Bourgoin (1993. Figs 40-47) but fall into the variation described for the species.

Iran localities: Dasht.

Extralimital distribution: Cyprus, Yugoslavia, Algeria, Israel, Italy (Huang and Bourgoin 1993), Kyrgizia (Novikov et al. 2006).

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References

- Abaii M (2000) Pests of forest trees and shrubs of Iran. Ministry of Agriculture. Agricultural Research, Education & Extension Organization, Iran, 178 pp.
- Abivardi C (2001) Iranian Entomology- An Introduction. Vol. 1 Faunal Studies. pp 1–444. Vol. 2 Applied Entomology, Springer Verlag, Berlin, 445–100.
- Afshar DJ (1937) Les insectes nuisibles aux arbres fruitiers en Iran. Ministry of Agriculture, Iran, 112 pp.
- Ahmadi AA, Izadpanah K, Jafari A (1986) Report of four delphacid species from cereal fields in the Fars province. In: Proceedings of the eighth plant protection congress of Iran (Isfahan), August- September 1986. Published by College of Agriculture, Isfahan University, Isfahan, 20 p.

- Anufriev GA (1966) Novye vidy ciakadovykh roda *Malenia* Haupt, 1929 (Homoptera, Derbidae) iz SSSR. In Tezisy dokladov konferencii molodykh mauchnykh rabotnikov Gorkovskogo universiteta. Sec biol nauk Gorkiy 45–47 [New species of planthoppers of the genus *Malenia* Haupt, 1924 (Homoptera, Derbidae) in the USSR.]. Abstracts of papers by young scientific workers in the Department of Biological Sciences, Gorkiy, 45–47.
- Anufriev GA (1977) Delphacids (Homoptera, Auchenorrhyncha) of the Kurile Islands fauna. Zoologicheskii Zhurnal 56(6): 855–869.
- Asche M (1982) Beitrage zur Delphaciden-Fauna der Turkei (Anatolien) (Homoptera Cicadinea Delphacidae). Marburger Entomologische Publikationen 1(7): 71–98
- Asche M (1985) Zur Phylogenie der Delphacidae Leach, 1815 (Homoptera Cicadina Fulgoromorpha). Marburger Entomologische Publikationen 2: 1–912.
- Asche M, Remane R (1982) Wer sind Kelisia putoni A. Costa, 1888 und Delphax nigrifrons Matsumura, 1910? (Homoptera Cicadina Delphacidae). Marburger Entomologische Publikationen 1 (7): 13–18.
- Asche M, Remane R (1983) Zur Problematik von *Delphacodes mulsanti* (Fieber, 1866) und zur Kenntnis einiger benachbarter Taxa (Homoptera Auchenorrhyncha Fulgoromorpha Delphacidae) (Vorläufige Mitteilung). Marburger Entomologische Publikationen 1(9): 25–56.
- Asche M, Wilson MR (1989) The palm–feeding planthopper genus *Ommatissus* (Homoptera: Fulgoroidea: Tropiduchidae). Systematic Entomology 14: 127–147. doi: 10.1111/j.1365-3113.1989.tb00271.x
- Asche M, Wilson MR (1990) The delphacid genus *Sogatella* and related groups: a revision with special reference to rice- associated species (Homoptera: Fulgoroidea). Systematic Entomology 15: 1–42. doi: 10.1111/j.1365-3113.1990.tb00301.x
- Barkhordari M, Samet Kh, Farzaneh A (1981) Étude préliminaire sur la faune des *Tamarix*. Journal of Entomological Society of Iran 6(1, 2): 3–8.
- Behdad E (1988) Pests and diseases of forest trees and shrubs and ornamental plants of Iran. Neshat Publisher, Esfahan, 824 pp.
- Behdad E (1991) Pests of fruit crops in Iran. Markaz-e Nashr-e Bahman, Iran, 822pp.
- Bergevin E de (1930) Note sur trois espèces d'Hémiptères recueillis en Egypte et description d'une nouvelle espèce d'*Urentius* (Hémiptère Tingitidae) et d'une nouvelle variété d'*Ommatissus binotatus* Fieb. (Homoptère Cixiidae). Bulletin de la Société Entomologique d'Égypte 14: 17–20.
- Boheman CH (1847) Nya Svenska Homoptera. Svenska vetenskaps-akademiens handlingar. Kungl. Svenska vetenskapsakademien 1847: 23–67.
- Costa E (1862) Di in nuovo genere di Emitteri Omotteri. Genere Trypetimorpha. Nob. Annuario Museo Zoologico della R. Univata di Napoli 1 (art 5): 60–65.
- Costa A (1888) Notizie ed osservazioni sulla Geo-Fauna Sarda. Memoria terza Risultamento della ricerche fatte in Sardegna nella estate del 1883. Atti della Reale Accademia delle Scienze Fisiche e Matematiche, Napoli, (2) 1(9): 1–64.
- Demir E (2007) Auchenorrhyncha (Homoptera) data from Ankara with two new records to Turkey. Munis Entomology & Zoology 2(2): 481–492.
- Demir E (2008) The Fugoromorpha and Cicadomorpha of Turkey, Part I: Mediterranean region (Hemiptera). Munis Entomology & Zoology 3(1): 447–522.

- Demir E, Demirsoy A (2009) Preliminary report on the Fulgoromorpha (Hemiptera) fauna of Kemliye (Erzincan) with a new record for Turkey. Munis Entomology & Zoology 4 (1): 280–286.
- Distant WL (1907) Rhynchotal notes XLI. Annals & Magazine of Natural History (7) 19: 277–295. doi: 10.1080/00222930709487267
- Dlabola J (1957a) Die Zikaden Afghanistans (Homopt.-Auchenorrhyncha) nach den Ergebnissen der von Herrn J. Klapperich in den Jahren 1952–1953 nach Afghanistan unternommenen Expedition. Mitteilungen der Münchner Entomologischen Gesellschaft 47: 265–303.
- Dlabola J (1957b) Results of the Zoological expedition of the National Museum in Prague to Turkey. 20. Homoptera, Auchenorrhyncha. Acta Entomologica Musei Nationalis Pragae 31(469): 19–68.
- Dlabola J (1960a) Iranische Zikaden (Homoptera: Auchenorrhyncha). (Ergebnisse der entomologischen Reisen Willy Richter, Stuttgart, in Iran, 1954 und 1956, N: 31). Stuttgarter Beiträge zur Naturkunde 41: 1–24.
- Dlabola J (1960b) Einige neue Zikaden aus Dagestan und Zentralasien (Homoptera). Stuttgarter Beiträge zur Naturkunde 40: 1–5.
- Dlabola J (1961a) Die Zikaden von Zentralasien, Dagestan und Transkaukasien (Homopt. Auchenorrhyncha). Acta Entomologica Musei Nationalis Pragae 34 (587): 241–358.
- Dlabola J (1961b) Neue und bisher unbeschriebene Zikaden–Arten aus Rumänien und Italien (Hom, Auchenorrh.). Acta Societatis Entomologicae Czechosloveniae 58(4): 310–323.
- Dlabola J (1964) Die Zikaden Afghanistans (Homoptera, Aucherorrhyncha), II. Ergebnisse der Sammelreisen von Dr. H. G. Amsel, G. Ebert, Dr. Erichson, J. Klapperich und Dr. K. Lindberg. Mitteilungen der Münchner Entomologischen Gesellschaft 54: 237–255.
- Dlabola J (1965a) Neue Zikadenarten aus Südeuropa (Homoptera–Auchenorrhyncha). Acta Entomologica Musei Nationalis Pragae 36: 657–669.
- Dlabola J (1965b) Ergebnisse der zoologischen Forschungen von Dr. Z. Kaszab in der Mongolei. 54; Homoptera, Auchenorrhyncha. Acta Faunistica Entomologica Musei Nationalis Pragae 11: 79–136.
- Dlabola J (1965c) Jordanische Zikaden (Homptera Auchenorrhyncha) (Bearbeitung der von. J. Klapperich im Jahre 1956–9 in Jordanien, Libanon und Syrien gesammelten Ausbeute). Acta Faunistica Entomologica Musei Nationalis Pragae 36: 419–450.
- Dlabola J (1968) Ergebnisse der zoologischen Forschungen von Dr. Z. Kaszab in der Mongolei. Nr. 169; Homoptera, Auchenorrhyncha. Acta Faunistica Entomologica Musei Nationalis Pragae 13: 23–36.
- Dlabola J (1971a) Taxonomische und chorologische Ergänzungen der Zikadenfauna von Anatolien, Iran, Afghanistan und Pakistan (Homoptera: Auchenorrhyncha). Acta entomologica Bohemoslovaca 68 (6): 377–396.
- Dlabola J (1971b) Taxonomische und chorologische Ergänzungen zur Türkischen und Iranischen Zikadenfauna (Homopt, Auchenorrhyncha). Acta faunistica Entomologica Musei Nationalis Pragae 14(163): 115–137.
- Dlabola J (1972) Beiträge zur Kenntnis der Fauna Afghanistans. Homoptera Auchenorrhyncha. Acta Musei Moraviae Scientiae Biologicae 16–17: 189–248.
- Dlabola J (1974a) Ergebnisse der Tschechoslowakisch- Iranischen Entomologischen Expedition nach dem Iran, 1970, No. 3 (Homoptera: Auchenorrhyncha) (1 Teil). Acta Entomologica Musei Nationalis Pragae, Supplement 6: 29–73.

- Dlabola J (1974b) Zur Taxonomie und Chorologie einiger Mediterraner Zikaden (Homoptera Auchenorrhyncha). Acta Zoologica Academiae Scientiarum Hungaricae 20(3–4): 289–308.
- Dlabola J (1977a) Die Tropiduchiden- Gattung *Kazerunia* in Iran (Homoptera: Auchenorrhyncha). Reichenbachia 16 (15): 163–167.
- Dlabola J (1977b) Neue iranische Cicadelliden und Arten mit faunistischen Erstfunden (Homoptera: Auchenorrhyncha). Acta Entomologica Bohemoslovaca 74 (4): 242–262.
- Dlabola J (1977c), Chorologische Ergänzungen zur Zikadenfauna des Mittelmeergebietes (Homoptera, Auchenorrhyncha). Acta Musei Nationalis Pragae 33 B: 1–2, 21–39.
- Dlabola J (1979a): *Bahuflata* gen.n neue Membraciden und Cicadiden-Arten aus dem Iran (Homoptera: Auchenorrhyncha). Reichenbachia 17 (28): 229–241.
- Dlabola J (1979b) Neue Zikaden aus Anatolien, Iran und aus südeuropäischen Ländern (Homoptera: Auchenorrhyncha). Acta Zoologica Academiae Scientiarum Hungaricae 21 (3– 4): 235–257.
- Dlabola J (1979c) *Tshurtshurnella*, *Bubastia* und andere verwandte Taxone (Auchenorrhyncha, Issidae). Acta Entomologica Bohemoslovaca 76(4): 266–286.
- Dlabola J (1979d) Insects of Saudi Arabia. Homoptera. Fauna of Saudi Arabia 1: 115–139.
- Dlabola J (1980a) Neue Zikadenarten der Gattungen Siculus gen. n, Mycterodus und Adarrus aus Südeuropa und 6 neue Mycterodus aus Iran (Homoptera: Auchenorrhyncha). Acta faunistica Entomologica Musei Nationalis Pragae 16(184): 55–71.
- Dlabola J (1980b) Tribus-Einteilung, neue Gattungen und Arten der Subf. Issinae in der eremischen Zone (Homoptera, Auchenorrhyncha). Acta Musei Nationalis Pragae 47(4): 173–245.
- Dlabola J (1980c) Insects of Saudi Arabia. Homoptera: Auchenorrhyncha (Part 2). Fauna of Saudi Arabia 2: 74–94.
- Dlabola J (1981) Ergebnisse der tschechoslovakisch-iranischen entomologischen Expeditionen nach dem Iran (Mit Angaben über einige Sammelresultate in Anatolien) (1970 und 1973) (Homoptera, Auchenorrhycha).- II Teil, Acta Musei Nationalis Pragae 40: 127–311.
- Dlabola J (1982a) Fortsetzung der Ergänzungen zur Issiden Taxonomie von Anatolien, Iran und Griechenland (Homoptera: Auchenorrhyncha). Acta Musei Nationalis Pragae 38 B (3): 113–169.
- Dlabola J (1982b) Übersicht der Gattung *Persepolia* aus dem Iran. Memorie Della Societa Entomologica Italiana 60(2): 163–168.
- Dlabola J (1983a) Ergebnisse der tschecholosvakisch-iranischen entomologischen Expeditionen 1970 und 1973 nach dem Iran, neue eremische Ricaniden und Flatiden (Homoptera; Auchenorrhyncha). Acta Musei Nationalis Pragae 41: 91–97.
- Dlabola J (1983b) Neue mediterrane, meistens anatolische Issiden (Homoptera, Auchenorrhyncha). Acta Entomologica Bohemoslovaca 80(2): 114–136.
- Dlabola J (1984) Neue Zikadenarten von dem Mediterraneum-Raum und Iran mit weiteren Beiträgen zur iranischen Fauna (Hom, Auchenorrhyncha). Acta Musei Nationalis Pragae 40: 21–63.
- Dlabola J (1985) Neue Cixiiden vom Iran, Nachbarländern und anderen Mediterrangebieten (Homoptera; Auchenorrhyncha). Acta Entomologica Bohemoslovaca 82: 92–128.
- Dlabola J (1986) Neue Arten der Fulgoromorphen Zikadenfamilien vom Mittelmeergebiet und Nahen Osten. (Homoptera, Auchenorrhyncha: Cixiidae, Meenoplidae, Derbidae, Dictyopharidae, Lophopidae und Issidae). Acta Musei Nationalis Pragae 42 (3–4): 169–196.

- Dlabola J (1987) Neue taxonomische Erkenntnisse über die Gattungen *Ommatidiotus* und *Conossimus* (Homoptera, Issidae). Acta Entomologica Musei Nationalis Pragae 42: 73–82.
- Dlabola J (1988) Reklassifikation der Gattungen der Pentastirini und neue Taxone der Cixiidae (Homoptera, Auchenorrhyncha). Acta Entomologica Bohemoslovaca 85(1): 49–70.
- Dlabola J (1989) Neue Issiden und andere Zikadenarten des Mediterraneums und vom zuliegenden Eremial (Homoptera, Auchenorrhyncha). Acta Musei Nationalis Pragae 45(1): 21–59.
- Dlabola J (1994) Ergänzungen zur iranischen, israelischen und benachbarten Zikadenfaunen mit Beschreibungen 30 neuer Taxone (Homoptera, Auchenorrhyncha). Acta Musei Nationalis Pragae 49 (1–4): 41–110.
- Dlabola J (1997) *Mycterodus* verwandte Taxone und sieben neue Zikadenarten (Homoptera, Auchenorrhyncha). Acta Entomologica Musei Nationalis Pragae 44 (1995): 301–319.
- Dlabola J, Heller F (1962) Iranische Zikaden II, (Ergebnisse der entomologischen Reisen Willi Richter, Stuttgart, im Iran 1954 und 1956, Nr. 42). Stuttgarter Beiträge zur Naturkunde 90: 1–8.
- Dlabola J, Jankovic L (1981) Drei neue *Erythria*-Arten und einige Ergänzungen der jugoslawischen Zikadenfauna. Bulletin de l'Academie Serbe des Sciences et des Arts Classe des Sciences Mathematiques et Naturelles Sciences Naturelles 75 (21): 67–79.
- Dlabola J, Safavi M (1972) *Persepolia*, eine neue Zikadengattung aus Iran (Homoptera: Auchenorrhyncha). Applied Entomology and Phytopathology 33(1–3): 1–4.
- Donovan E (1800) An epitome of the natural history of the insects of India, and the islands in the Indian Seas: comprising upwards of two hundred and fifty figures and descriptions of the most singular and beautiful species. T. Bensley, London, 70 pp.
- Dubovsky GK (1966) Tsikadovye (Auchenorrhyncha) Ferganskoi doliny. [Cicadinea (Auchenorrhyncha) of the Fergana Valley]. Fan, Tashkent, 256 pp.
- Drosopoulos S, Asche M, Hoch H (1983) Contribution to the planthopper fauna of Greece (Homoptera, Auchenorrhyncha, Fulgoromorpha, Delphacidae). Annales de l'Institut Phytopathologique Benaki (N.S.) 14: 19–68.
- Dufour L (1833) Recherches anatomiques et physiologiques sur les Hémiptères, accompagnées de considérations relatives à l'histoire naturelle et à la classification de ces insects. Mémoires des Savants étrangers de l'Académie Royale des Sciences de l'Institut de France 4: 129–462.
- Emeljanov AF (1964) Novye cikadovye iz Kazarkstana (Homoptera, Auchenorrhyncha). Trudy Zool Institute, Leningrad, 34: 1–51.
- Emeljanov AF (1972a) New species of the subfamily Orgeriinae of Palaearctic fauna (Homoptera, Dictyopharidae). Trudy VEO. Proceedings of the All-Union Entomological Society 55: 14–38.
- Emeljanov AF (1972b) New genera of leafhoppers of the families Cixiidae and lssidae (Homoptera, Auchenorrhyncha) in the USSR. Entomological Review Washington 50: 350–354.
- Emeljanov AF (1977) Leaf-hoppers (Homoptera, Auchenorrhyncha) from the Mongolian People's Republic based mainly on materials of the Soviet Mongolian zoological expeditions (1967–1969)]. Nasekomye Mongol 5: 96–195.
- Emeljanov AF (1978) New genera and species of leafhoppers (Homoptera, Auchenorrhyncha) from the USSR and Mongolia. Entomologicheskoye Obozreniye 57(2): 316–332

- Emeljanov AF (1979) The problem of Family distinction between the Fulgoridae and the Dictyopharidae (Homoptera, Auchenorrhyncha), Trudy ZIN AN SSSR [Proceedings of the Zoological Institute of the Academy of Science USSR] 82: 3–22.
- Emeljanov AF (1997) New species and new data on the distribution of the subfamily Orgeriinae in the Palaearctic Region (Homoptera: Dictyopharidae). Zoosystematica Rossica 6(1/2): 83–90.
- Emeljanov AF (1984) To the knowledge of the families Kinnaridae and Meenoplidae (Homoptera, Fulgoroidea). Entomologicheskoye Obozrenie, [Entomological Review], 1984, 63, 3: 468–483.
- Emeljanov AF (1995) The new taxa of the tribe Pentastirini (Homoptera, Cixiidae) from Palaearctic. Zoologicheskii Zhurnal 74 (9): 73–89.
- Emeljanov AF (1996a) A new genus, *Chirodisca* gen. n, and new subgenera of the genus Aphelonema Uhl. (Homoptera, Fulgoroidea, Issidae). Entomologicheskoye Obozrenie 75(4): 834–835.
- Emeljanov AF (1996b) First record of *Hyalesthes mlokosiewiczi* Signoret from Middle Asia and some other new data on its distribution (Homoptera: Cixiidae). Zoosystematica Rossica 5(1): 28.
- Emeljanov AF (1999) Notes on delimitation of families of the Issidae group with description of a new species of Caliscelidae belonging to a new genus and tribe (Homoptera, Fulgoroidea). Zoosystematica Rossica 8: 61–72.
- Emeljanov AF (2004) The subgeneric division of the genus *Dicytophara* Germar, 1833 (Homoptera; Dicytopharidae). Russian Entomological Journal 12(4): 357–358.
- Emeljanov AF (2009) A New Species of the Planthopper Genus Nymphorgerius Oshanin (Homoptera, Dictyopharidae) from Iran. Entomological Review 89 (4): 426–427. doi: 10.1134/S001387380904006X
- Fabricius JC (1794) Ryngota. Entomologia systematica emendata et aucta: secundun classes, ordines, genera, species, adjectis synonimis, locis, observationibus, descriptionibus. 4: 1–472.
- Fallén CF (1806) Försök till Svenska Cicad-Arternas uppställning och beskrifning. [Concluded]. Kongliga Vetenskaps Academiens Handlingar 27: 113–130.
- Fallén CF (1826) Cicadariae, earumque Familiae, affines. Hemiptera Svaciæ. Cimicides eorumque familiæ affines. ex officina Berlingiana . Londini Gothorum, 2: 1–80.
- Farahbakhsh Gh (1961) A checklist of economically important insects and other enemies of plants and agricultural products in Iran. Plant organization Press, Iran, 153pp.
- Fieber FX (1865) Synopse der europ. Arten Tettigometra. Verhandlungen der Kaiserlich-Königlichen Zoologisch-botanischen Gesellschaft in Wien 15: 561–572.
- Fieber FX (1866) Grundzüge zur generischen Theilung der Delphacini. Verhandlungen der Kaiserlich-Königlichen Zoologisch-botanischen Gesellschaft in Wien 16: 517–534
- Fieber FX (1876) Les Cicadines d'Europe d'après les originaux et les publications les plus récentes. Deuxième partie: Descriptions des espèces. Traduit de l'allemand par Ferd. Reiber [sic]. Revue et Magasin de Zoologie pure et appliquée 3 (4): 11–268.
- Gardenhire RQ (1959) Summary of Insect conditions in Iran. Applied Entomology and Phytopathology 18: 51–61.

- Germar EF (1818) Bermerkungen über einige Gattungen der Cicadarien. Magazin der Entomologie 3: 177–227.
- Germar EF (1821) Bemerkungen über einige Gattungen der Cicadarien. Magazin der Entomologie 4: 1–106.
- Germar EF (1825) *Flata stigmatica* Germ, *Flata musiva* n. sp, *Coelidia scutata* Germ. Agusti Ahrensii Fauna Insectorum Europae 11: 20–22.
- Gharib A (1966) Palm leafhopper *Ommatissus binotatus* Fieb. var. lybicus Berg. (Homoptera: Tropiduchidae). Applied Entomology and Phytopathology 24: 37–47.
- Gharib A (1998) Important pests of palm. Agricultural Research, Education & Extension Organization, Tehran, 41 pp.
- Gnezdilov VM (2002) On the identity and systematic position of *Hysteropterum pictifrons* Melichar, 1906 (Homoptera: Cicadina, Issidae). Acta Zoologica Academiae Scientiarum Hungaricae 43(3): 213–217.
- Gnezdilov VM (2003) A new tribe of the family Issidae (Homoptera, Cicadina) with comments on the family as a whole. Zoosystematica Rossica 11: 305–309.
- Gnezdilov VM (2010a) *Reinhardema* nov. gen, a new genus of the family Caliscelidae from Iran (Hemiptera: Fulgoroidea). Cicadina 11: 13–15.
- Gnezdilov VM (2010b) New synonyms, combinations and faunistic records of Western Palaearctic planthoppers of the family Issidae (Homoptera, Fulgoroidea). Entomologicheskoe Obozrenie 89(2): 413–422.
- Gnezdilov VM, Drosopoulos S, Wilson MR (2004) New data on taxonomy and distribution of some Fulgoroidea (Homoptera, Cicadina). Zoosystematica Rossica 12: 217–223.
- Gnezdilov VM, Mozaffarian F (2011) A new species of the genus *Eusarima* Yang (Hemiptera: Fulgoroidea: Issidae) from Iran. Acta entomologica Musei nationalis Pragae 51 (2): 457-462.
- Gnezdilov VM, Wilson MR (2006) Systematic notes on tribes in the family Caliscelidae (Hemiptera: Fulgoroidea) with the description of new taxa from Palaearctic and Oriental Regions. Zootaxa 1359: 1–30.
- Haghshenas A, Khajeali J (2000) Leafhoppers (Homoptera: Auchenorrhyncha) fauna of potato fields in Chagar Mahal and Bakhtiari provice. In: Proceedings of the fourteenth Iranian Plant Protection Congress (Isfahan), September 2000. Published by Isfahan University of Technology, Isfahan, 249.
- Haupt H (1927) Homoptera Palestinae I. Bulletin. The Zionist Organization. Institute of Agriculture and Natural History. Agricultural Experiment Station 8: 5–43.
- Haupt H (1935) Unterordnung: Gleichflugler, Homoptera. In: Brohmer P, Ehrmann, P, Ulmer, G (Eds) Die Tierwelt Mitteleuropas 4 (10): 115–262.
- Herrich-Schäffer GAW (1835) Homoptera. Nomenclator entomologicus: Verzeichnis der europäischen Insecten, zur Erleichterung des Tauschverkehrs mit Preisen versehen. Friedrich Pustet, overall work published 1835–1840. Regensburg. Vol. 1 (1–3): 1–116.
- Herrich-Schäffer GAW (1838) Jassus ornatus, Poecilloptera subquadrata, Flata pallida, Jassus intricatus. Deutschlands Insecten 154: 1–5.
- Hoch H (1986) Acht neue Arten der Gattung *Hyalesthes* Signoret, 1865 (Homoptera Fulgoroidea Cixiidae). Marburger Entomologische Publicationen 2 (3): 87–122.

- Hoch H, Remane R (1985) Evolution und Speziation der Zikaden-Gattung *Hyalesthes* Signoret, 1865 (Homoptera Auchenorrhyncha Fulgoroidea Cixiidae). Marburger Entomologische Publicationen 2: 1–427.
- Holzinger WE (2007) Redescriptions of Ordalonema faciepilosa, Peltonotellus melichari and P. raniformis, with a key to Western Palaearctic genera of Caliscelidae (Hemiptera: Fulgoromorpha). European Journal of Entomology 104: 277–283.
- Holzinger WE, Emeljanov AF, Kammerlander L (2002) The family Cixiidae Spinola 1839 (Hemiptera: Fulgoromorpha) - a Review. Denisia 04, Zugleich Kataloge des OÖ Landesmuseums, Neue Folge 176: 113–138.
- Holzinger WE, Kammerlander I, Nickel H (2003) Auchenorrhyncha of central Europe, Fulgoromorpha, Cicadomorpha excl. Cicadellidae. Koninklijke Brill, 673 pp.
- Horváth G (1891) Hémiptères recueillis dans l'Arménie Russe avec la description d'espèces et variétés nouvelles. Revue d'Entomologie. Publiée par la Société Française d'Entomologie 10: 68–81.
- Horváth G (1894) Hémiptères recueillis dans la Russie méridionale et en Transcaucasie. Revue d'Entomologie. Publiée par la Société Française d'Entomologie 13: 169–189.
- Horváth G (1897a) Description d'Hémiptères nouveaux et notes diverses. Revue d'Entomologie 16: 81–97.
- Horváth G (1897b) Homoptera nova ex Hungaria. Természetrajzi Füzetek. Kiadja a Magyar Nemzeti Muzeum Budapest 20: 620–643.
- Horváth G (1899) Hémiptères de l'Île de Yesso (Japon). Természetatrajzi Füzetek 22: 365–374.
- Horváth G (1905) Species generis Ommatidiotus Spin. Annales historico-naturales musei nationalis Hungarici 3: 378–387.
- Huang J, Bourgoin T (1993) The planthopper genus *Trypetimorpha*: systematics and phylogenetic relationships (Hemiptera: Fulgoromorpha: Tropiduchidae). Journal of Natural History 27: 609–629. doi: 10.1080/00222939300770351
- Karimzadeh Esfahani J, Kharrazi Pakdel A, Kheyri M (1998) Leafhopper (Homoptera: Auchenorrhyncha) fauna of sugarbeet fields in Isfahan province. In: Proceedings of the thirteenth Iranian Plant Protection Congress (Karaj), August 1998. Published by Karaj Junior College of Agriculture, Karaj, 69.
- Khajehali J, Seyedoleslami H, Kamali K (2001) A contribution to study of hoppers (Hom.: Auchenorrhyncha) of potato fields in Isfahan and Daran. Applied Entomology and Phytopathology 68 (1&2): 43–71.
- Kheyri M. (1989) An inventory of pests attacking sugar-beet in Iran. Applied Entomology and Phytopathology 56 (1&2): 75–91.
- Kirschbaum CL (1868) Die Cicadinen der Gegend von Wiesbaden und Frankfurt A. M. nebst einer Anzahl neuer oder schwer zu unterscheidender Arten aus anderen Gegenden Europa's. Tabellarisch Beschrieben. Jahrbücher des Vereins für Naturkunde im Herzogthum Nassau 21–22: 1–202.
- Krampl F, Dlabola J (1983) A new genus and species of Epipyropid moth from Iran ectoparasitic on a new *Mesophantia* species with a revision of the host genus (Lepidoptera, Epipyropidae; Homoptera, Flatidae). Acta Entomologica Bohemoslovaca 80(6): 451–472.

- Kusnezov V (1929) Materialien zur Kenntnis der Homopteren-Fauna Turkestans. Zoologischer Anzeiger 79: 305–334.
- Kusenezov V (1937) Neue asiatische Oliarus-Arten (Homoptera-Cixiidae). Entomologisches Nachrichtenblatt 10:161–168.
- Kwon YJ (1982) New and little known planthoppers of the family Delphacidae (Homoptera: Auchenorrhyncha). Korean Journal of Entomology 12 (1): 1–11
- Lashkari M, Nouri Ganbalani G, Mozaffarian F, Ghorbani Kh, Fathi A (2009) Faunistic study of planthoppers infraorder Fulgoromorpha (Hem. Auchenorrhyncha) in different climatic regions of Gorgan, Iran. Journal of Entomological Research 1(2): 119–133.
- Le Quesne WJ (1960) Some further taxonomic observations on Delphacidae (Hom.), including a synonymic change and a new European species. The Entomologist 93: 186–188
- Lethierry LF (1874) Hémiptères nouveaux. Petites Nouvelles Entomologiques, 449 pp.
- Lethierry LF (1878) Homoptères nouveaux d'Europe et des contrées voisines. Deuxième partie. Compte rendu des séances de la Société Entomologique de Belgique 21: 25–31.
- Lethierry LF (1888) Liste des Hémiptères recueillis à Sumatra et dans l'île Nias par Mr. E. Modigliani. Annali del Museo civico di storia naturale di Genova (2) 6: 460–470; separate paged 1–11.
- Lindberg H (1948) Materialien zu einer Monographie der Gattung *Tettigometra* (Hom. Cicad.). Notulae entomologica 28: 1–40.
- Lindberg H (1960) Supplementum Hemipterorum Insularum Canariensium. Societas Scientarum Fennica Commentationes Biologicae 22(6): 3–20.
- Linnavuori R (1957) Remarks on some Mediterranean Homoptera. Annales entomologici Fenneci, Helsinki 23: 94–97.
- Linnavuori R (1962a) Hemiptera of Israel III. Annales Zoologici Sogietatis Zoologica Botanica Fennica "Vanamo" 24(3): 1–108.
- Linnavuori R (1962b) Studien an der Gattung *Phantia* Fieb. (Homoptera, Flatidae). Beiträge zur Naturkunde aus dem Staatlichen Museum fur Naturkunde in Stuttgart 89: 1–4.
- Linnavuori R (1964) Hemiptera of Egypt, with remarks on some species of the adjacent Eremian region. Annales Zoologici Fennici 1: 306–356.
- Linnaeus C (1761) II. Hemiptera. Cicada. Fauna Svecica: sistens animalia sveciae regni: Mammalia, Aves, Amphibia, Pisces, Insecta, Vermes, distributa per classes & ordines, genera & species, cum differentiis specierum, synonymis auctorum. Editio altera, auctior. insectorum. L. Salvii, Stockholmiae, 578 pp.
- Linnaeus C (1767) II. Hemiptera. Systema naturae: per regna tria naturae, secundum classes, ordines, genera, species cum characteribus, differentiis, synonymis, locis. Editio duodecima, reformata, L. Salvii , [Stock]holmiae. 1 (2): 533–1327.
- Logvinenko VN (1969) [New and not popular representatives of the Cixidae family (Homoptera, Auchenorrhyncha) of the Caucasus and Transcaucasia] Transcaucasia scientific Expedition of the Institute of Zoology Academy of Sciences, Ukrainian SSR in 1966–1967. Vestnik Zoologogii 1: 53–59.
- Logvinenko VN (1974) A review of species of the genus *Mycterodus* Spin. (Homoptera, Issidae) of the fauna of the USSR. Entomologicheskoe Obozrenie 53(4): 830–852.
- Logvinenko VN (1975) A review of leafhoppers of genus *Aeluropsis* Em. (Auchenorrhyncha, Issidae). Vestnik Zoologogii 1: 56–61.

- Matsumura S (1910) Neue Cicadinen aus Europa und dem Mittelmeergebiet (Schluss). Journal of the College of Science, Imperial University of Tokyo 27(18): 1–38.
- Medler JT (2003) Types of Flatidae (Homoptera) XXIII. Lectotype designations and taxonomic notes on Melichar species in the Moravian museum, Brno, Czech Republic. Oriental Insects 37: 343–357.
- Melichar L (1896) Cicadinen (Hemiptera-Homoptera) von Mittel-Europa. F. L. Dames. Berlin. i-xxvii: 1–364.
- Melichar L (1901) Eine neue Homopteren-Gattung und Art aus der Gruppe Delphacini. Wiener Entomologische Zeitung 20: 55–56.
- Melichar L (1902a) Homopteren aus West- China, Persien und dem Süd-Ussuri- Gebiete, St. Petersburg Museum Zool Annals 7: 76–146.
- Melichar L (1902b) Monographie der Acanaloniiden und Flatiden (Homoptera) (Fortsetzung.). Annalen des Naturhistorischen Hofmuseums, Wien.17: 1–253.
- Melichar L (1903) Homopteren-Fauna von Ceylon. Verlag von Felix L. Dames, Berlin, i–iv, 248 pp.
- Melichar L (1904) Neue Homopteren aus Süd-Schoa, Galla und den Somal-Ländern. Verhandlungen der Kaiserlich-Königlichen Zoologisch-botanischen Gesellschaft in Wien 54: 25–48.
- Melichar L (1906) Monographie der Issiden. (Homoptera). Abhandlungen der K. K. Zoologisch-botanischen Gesellschaft in Wien 3: 1–327.
- Melichar L (1912) Monographie der Dictyophoriden (Homoptera). Abhandlungen der K. K. Zoologisch-botanischen Gesellschaft in Wien 7 (1): 1–221.
- Melichar L (1913) Cicadina. Faune du district de Walouyki du gouvernement de Woronège (Russie) par Vladimir Velitchkovsky 7: 1–11.
- Mirzayans H (1995) Insects of Iran, The list of Homoptera: Auchenorrhyncha in the Insect Collection of plant Pests & Diseases Research Institute. Ministry of Agriculture, Agricultural Research, Iran, 63 pp.
- Mirzayans A, Borumand H, Zairi M, Rajabi Gh (1976) Insect fauna from province of Fars (Iran) (1). Journal of Entomological Society of Iran 3(1,2): 109–135.
- Mitjaev ID (1971) Leafhoppers of Kazakhstan (Homoptera Cicadinea). Science of Kazakh SSR, 211 pp.
- Mozaffarian F, Emeljanov AF (2010) A faunestic study of the subfamily Dictyopharinae Spinola, 1839 (Hem.: Dictyopharidae) in Iran. In: Manzari, S (Ed) Proceedings of the ninetheenth Iranian Plant Protection Congress, Tehran, July-August 2010. Published by Iranian Research Institute of Plant Protection, Tehran, 128.
- Mozaffarian F, Gnezdilov VM (in press) First record of *Scorlupella montana* (Becker) (Hemiptera: Fulgoroidea: Issidae) from Iran. Journal of Entomological Society of Iran.
- Mulsant ME, Rey C (1855) Description de quelques Hémiptères-Homoptères nouveaux ou peu connus. Annales de la Société Linnéenne de Lyon (2) 2: 197–249, 426.
- Nast J (1972) Palaearctic Auchenorrhyncha (Homoptera) An annotated check list. Polish Scientific Publishers, Warsaw, Poland, 550 pp.
- Nast J (1986) Notes on some Auchenorrhyncha (Homoptera), 6–10. Annales Zoologici (Warsaw) 40 (2–5): 297–307.

- Nematollahi MR, Khajehali J (2000) Major leafhoppers and planthoppers (Hom.: Auchenorrhyncha) of corn fields in Esfahan. In: Proceedings of the fourtheenth Iranian Plant Protection Congress (Isfahan), September 2000. Published by Isfahan University of Technology, Isfahan, 235.
- Novikov DV, Novikova1 NV, Anufriev GA, Dietrich CH (2006) Auchenorrhyncha (Hemiptera) of Kyrgyz Grasslands, Russian Entomological Journal 15(3): 303–310.
- Oshanin VT (1879) Materials for the study of the fauna of the Hemiptera of Turkestan. Annales de la Section du Turkestan de la Société des Amis des Sciences Naturelles, de l'Anthropologie et de l'Ethnographie 1: 99–163.
- Oshanin VT (1908) Verzeichnis der palaearktischen Hemipteren. I. Neue Gattungen und Arten von Homopteren aus Russisch-Asien. Annuaire du Musée Zoologique de l'Académie Impériale des Sciences de St.-Pétersbourg 12: 463–477.
- Oshanin VT (1913) Synopsis der Tribus Orgeriaria der russischen Fauna. (Hemiptera-Homoptera). Revue Russe d'Entomologie. Publiée par la Société Entomologique de Russie 13: 135–147.
- Perris E (1857) Nouvelles excursions dans les Grandes Landes. Annales de la Société Linnéenne de Lyon 4: 83–180.
- Puton A (1890) Une douzaine d'Hémiptères nouveaux et notes diverses. Revue d'Entomologie. Publiée par la Société Française d'Entomologie 9: 227–236.
- Puton A (1895) Hémiptères nouveaux. Revue d'Entomologie. Publiée par la Société Française d'Entomologie 14: 83–91.
- Puton A, Lethierry L (1887) Hémiptères nouveaux de l'Algérie. Revue d'Entomologie. Publiée par la Société Française d'Entomologie 6: 298–311.
- Ribaut H (1934). Nouveaux delphacides (Homoptera-Fulgoroidea.). Bulletin de la Société d'Histoire Naturelle de Toulouse 66: 281–301.
- Ribaut H (1954) *Delphacodes audrasi* n. sp. (Homoptera, Araeopidae). Bulletin de la Societe Linnéenne de Lyon, 23(7): 180–181
- Rajabi Gh (1989) Insects attacking Rosaceous fruit trees in Iran. Third volume: Homoptera. Ministry of Agriculture, Iran, 256 pp.
- Rusiecka M de (1902) Monographie du genre *Phantia* Fieb. Mittheilungen der Schweizerischen Entomologischen Gesellschaft 10: 420–424.
- Scott J (1870) On certain British Hemiptera-Homoptera. Descriptions of new species of the genus *Liburnia*, Stål. Entomologists monthly Magazine 7: 67–75.
- Signoret V (1865) Descriptions de quelques Hémiptères nouveaux Annales de la Société Entomologique de France 4 (5): 115–130.
- Signoret V (1879) Une nouvelle espèce de cixide du genre *Hyalesthes*. Bulletin de la Société Entomologique de France 5 (9): 66.
- Signoret V (1884) Descriptions de deux espèces nouvelles de Cixiides. Bulletin de la Société Entomologique de France 6 (4): 71–72.
- Stål C (1863) Beitrag zur Kenntnis der Fulgoriden. Entomologische Zeitung. Herausgegeben von dem entomologischen Vereine zu Stettin 24: 230–251.
- Stål C (1865) Homoptera nova vel minus cognita. Öfversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar 22: 145–165.

- Tishechkin D Yu (1998) Acoustic signals of Issidae (Homoptera, Cicadinea, Fulgoroidea) compared with signals of some other Fulgoroidea with notes on taxonomic status of the subfamily Caliscelinae. Zoologichesky Zhurnal 77: 1257–1265.
- Vilbaste J (1980) On the Homoptera-Cicadinea of Kamchatka. Annales Zoologici 35(24): 367–418.
- Wagner W (1938) (Homopt. Delphacidae). Bombus 4: 12.
- Wagner W (1939) Die Zikaden des mainzer Beckens, zugleich eine Revsion der Kirshbaumschen Arten aus der Umgebung von Wiesbaden. Jahrbücher des Vereins für Naturkunde im Herzogthum Nassau 86: 77–212.
- Wilson MR (2010a) Order Hemiptera, families Meenoplidae and Kinnaridae. Arthropod fauna of the UAE 3: 126–131.
- Wilson MR (2010b) Order Hemiptera, family Tropiduchidae. Arthropod fauna of the UAE 3: 132–134
- Wilson SW, O'Brien LB (1987) A survey of planthopper pests of economically important plants (Homoptera: Fulgoroidea). In: Wilson MR, Nault LR, (Eds) Proceedings of 2nd International Workshop on Leafhoppers and Planthoppers of Economic Importance, Brigham Young University, Provo, Utah, July- August 1986. CABI, 343–360.
- Yarmand H, Sadeghi SE, Mohammadi M, Ebrahimi SS, Seif Allahi AR (2006) Survery of arthropods associated with range medicinal plants in Tehran, Ghazvin and Isfahan provinces. Iranian Journal of Forest and Range Protection Reseach 3(2): 107–227.

List of the localities, their coordinates and the codes (Fig. 2)

Åbadeh 31°09'N, 52°39'E (F2) Ābāsk 35°53'N, 52°10'E (M7) Åbyek 36°02'N, 50°31'E (Gz2) Ahram 28°53'N, 51°16'E (B4) Ahvāz 31°19'N, 48°40'E (Kh10) Akbarābād 29°14'N, 52°46'E (F11) Akhlamad 36°36'N, 58°57'E (KhR4) Albāji 31°29'N, 48°37'E (Kh8) Aliābād (Ghom) 35°08'N, 50°59'E (Gh2) Aliābād (Jahrom) 29°30'N, 52°35'E (F17) Ålmeh 37°23'N, 56°8'E (Gs4) Āmol 36°28'N, 52°21'E (M8) Anārak 26°44'N, 62°15'E (S16) Anbarābād 28°25'N, 57°50'E (Km10) Anbarieh 32°19'N, 48°08'E (Kh1) Andimeshk 32°27'N, 48°20'E (Kh1) Asadābād 34°47'N, 48°07'E (Hd2) Asālem 37°43'N, 48°54'E (G2) Azādbar 36°07'N, 51°10'E (T2)

Bābol 36°34'N, 52°42'E (M9) Bābolsar 36°41'N, 52°38'E (M9) Bafgh 31°35'N, 55°24'E (Y4) Baft 29°14'N, 56°36'E (Km7) Bāghak 28°54'N, 51°07'E (B4) Bāhukalāt 25°43'N, 61°25'E (S18) Bājgāh 29°43'N, 52°35'E (F9) Bam 29°06'N, 58°21'E (Km11) Bampur 27°11'N, 60°27'E (S9) Bandar Abbās 27°11'N, 56°16'E (H7) Bandar Anzali 37°28'N, 49°27'E (G4) Bandar Chārak 26°43'N, 54°16'E (H15) Bandar Khamir 26°56'N, 55°35'E (H5) Bandar Lengeh 26°33'N, 54°52'E (H3) Bānuchārehar 28°30'N, 57°00'E (Km8) Bārgāh 27°38'N, 57°42'E (Km12) Bazmān 27°49'N, 60°11'E (S5) Behshahr 36°41'N, 53°32'E (M11) Bidhand 33°14'N, 50°18'E (E2) Bidruyeh 32°46'N, 48°13'E (Kh1) Bidzard 29°21'N, 51°51'E (F6) Bilāi 26°30'N, 57°06'E (H12) Birjand 32°53'N, 59°12'E (KhJ1) Bishāpur 29°43'N, 51°24'E (F5) Bojnurd 37°28'N, 57°19'E (KhSh2) Borāzjān 29°16'N; 51°12'E (B3) Chābahār 25°18'N, 60°37'E (S19) Chāhkuh 26°37'N, 54°58'E (H2) Chālus 36°39'N, 51°25'E (M3) Chenārshāhijān 29°51'N, 51°34'E (F5) Dālaki 29°25'N, 51°17'E (B2) Damāvand 35°44'N, 52°03'E (T7) Dārāb 28°45'N, 54°32'E (F18) Dārābād 35°48'N; 51°26'E (T4) Dārān 32°58'N, 50°24'E (E6) Dārbahāre 30°26'N, 49°54'E (Kh14) Darband 36°01'N, 51°28'E (T4) Dareh gaz 27°51'N, 56°17'E (H7) Darpahn 26°44'N, 57°28'E (H12) Darzin 27°58'N, 55°58'E (H6) Dasht 37°17'N, 56°07'E (Gs4) Dasht-e arjan 29°39'N, 51°58'E (F8) Dasht-e Moghān (A)

Dehbakri 29°03'N, 57°55'E (Km9) Dehbid 30°38'N, 53°13'E (F10) Dehpābid 28°37'N, 60°46'E (S4) Delijān 33°59'N, 50°40'E (Mz1) Denā mt (west slpe) 30°49'N, 51°35'E (KB1) Divandareh 35°55'N, 47°01'E (Kd1) Doborji 28°27'N, 55°06'E (F18) Dowlatābād 28°20'N, 56°38'E (Km8) Esfahān 32°40'N, 51°40'E (E3) Esfarāyen 37°05'N, 57°29'E (KhSh4) Eskandari 32°48'N, 50°25'E (E7) Eslāmābād 37°42'N, 56°5'E (Gs5) Espakeh 26°47'N, 60°14'E (S8) Estahbān 29°07'N, 54°02'E (F16) Evin 35°40'N, 51°26'E (T4) Eynvarzān 35°39'N, 52°12'E (T8) Fāriāb 27°28'N, 57°06'E (H10) Fasā 28°55'N, 53°38'E (F15) Feizābād 36°52'N, 54°33'E (Gs1) Ferdows-e Esfadaghe 28°44'N, 57°15'E (Km10) Firuzābād 28°50'N, 52°34'E (F13) Firuzkuh 35°45'N, 52°45'E (T9) Galugāh 36°43'N, 53°48'E (M11) Gambuyeh 31°23'N, 48°30'E (Kh7) Ganāveh 29°34'N, 50°31'E (B1) Gāvbandi 27°12'N, 53°03'E (H1) Gavkoshak 29°38'N, 51°48'E (F7) Gāvkoshi 28°38'N, 57°12'E (Km10) Gazanak 35°54'N, 52°12'E (M6) Geno 27°24'N, 56°12'E (H6) Ghaderābād 30°20'N, 53°21'E (F2) Ghāemshahr 36°28'N, 52°53'E (M9) Gharechaman 37°36'N, 47°08'E (Ash4) Ghāsemābād 27°10'N, 60°20'E (S8) Ghasr-e Ghand 26°12'N, 60°45'E (S13) Ghazvin 36°16'N, 50°00'E (Gz1) Ghezel-Bolāgh 39°22'N, 44°12'E (AG1) Gholhak 35°46'N, 51°26'E (T4) Ghom 34°38'N, 50°53'E (Gh1) Ghom Lake (Gh1) Ghotbābād 27°46'N, 56°06'E (H6) Golbāf 29°51'N, 57°44'E (Km5) Gorgān 36°50'N, 54°25'E (Gs2)

Hafttappeh 32°02'N, 48°23'E (Kh2) Hājiābād 28°08'N, 55°52'E (H6) Hājiābād 28°18'N, 55°54'E (F18) Hamadān 34°47'N, 48°30'E (Hd1) Hamidieh 31°28'N, 48°26'E (Kh6) Hāresābād 36°07'N, 57°36'E (KhR2) Hasanlangi 27°21'N, 56°52'E (H9) Hashtpar 37°48'N, 48°54'E (G1) Heirān 38°22'N, 48°34'E (A2) Hessār 36°02'N, 59°20'E (KhR6) Hezār mt (Peak) 29°31'N, 57°15'E (Km6) Hichān 26°20'N, 60°03'E (S11) Hoseinābād 28°51'N, 54°14'E (F18) Irānshahr 27°12'N, 60°41'E (S9) Isin 27°19'N, 56°17'E (H7) Izadkhāst 31°30'N, 52°07'E (F1) Izeh 31°49'N, 49°51'E (Kh4) Jahānābād 29°42'N, 53°51'E (F14) Jahrom 28°30'N, 53°33'E (F17) Jandagh 34°03'N, 54°24'E (E4) Jask 25°38'N, 57°46'E (H14) Jiroft 28°40'N, 57°43'E (Km10) Kāhe 35°06'N, 58°09'E (KhR8) Kahkom 28°12'N, 55°46'E (Y3) Kahnuj 27°33'N, 57°42'E (Km12) Kahurak 29°26'N, 59°39'E (S3) Kahurestān 27°12'N, 55°34'E (H4) Kamālābd 29°17'N, 53°02'E (F12) Kamandān 33°18'N, 49°27'E (L2) Kāmyārān 34°47'N, 46°56'E (Kd3) Kandovān (Āzarbāijān-e Sharghi) 37°54'N, 46°05'E (Ash2) Kandovān (Māzandarān) 36°08'N, 51°17'E (M6) Kangān 27°50'N, 52°04'E (B6) Karaj 35°49'N, 50°58'E (T2) Kāravāndar 27°57'N, 60°43'E (S4) Kāshān, 33°59'N, 51°26'E (E1) Kāzerun 29°36'N, 51°39'E (F6) Kelārdasht 32°32'N, 50°20'E (M5) Kermān 30°14'N, 56°58'E (Km3) Kermānshāh (Yazd province) 31°20'N, 54°54'E (Y2) Khalkānlu 37°05'N, 58°44'E (KhR1) Khalkhāl 37°37'N, 48°31'E (A3) Khānehkhoreh 30°51'N, 53°08'E (F2)

Khargh 36°57'N, 58°00'E (KhSh3) Kharposht 32°49'N, 59°02'E (KhJ1) Khātunābād 29°28'N, 57°48'E (Km9) Khorramābād 36°45'N, 50°53'E (M2) Khorramshahr 30°25'N, 48°11'E (Kh12) Khosro- Shirin 30°53'N, 52°00'E (F2) Khoy 38°21'N, 44°50'E (AG5) Khur va Biābānak 33°46'N, 55°05'E (E5) Komehr 30°26'N, 51°52'E (F3) Konārdān 28°37'N, 53°20'E (F17) Kuhenjān 29°19'N, 52°54'E (F11) Kuhrang 32°32'N, 50°20'E (Ch3) Kushk 30°31'N, 51°30'E (F4) Lāhijān 37°12'N, 50°00'E (G5) Lajrān 35°16'N, 52°08'E (T7) Lalezār 29°31'N, 56°48'E (Km6) Leshtar 32°02'N, 50°16'E (Ch1) Māhān 30°05'N, 58°18'E (Km5) Mahārlu 29°21'N, 52°49'E (F11) Māku 39°17'N, 44°31'E (AG2) Malard 35°40'N, 50°58'E (T3) Marāgheh 37°22'N, 46°14'E (Ash6) Marand 38°25'N, 45°46'E (Ash1) Marg-e-Malek 32°29'N, 50°29'E (Ch1) Mashhad 36°17'N, 59°35'E (KhR5) Mashhad-e Ardehāl 34°1'N, 51°2'E (E1) Masiri 30°15'N, 51°30'E (F4) Matnagh 37°50'N, 46°31'E (Ash2) Mehrān 33°06'N, 46°09'E (I1) Meshkinshahr 38°23' 56N; 47°40' 55E (A1) Miāneh 37°25'N, 47°42'E (Ash7) Miānjangal 29°06'N, 53°42'E (F15) Mijān 28°42'N, 57°56'E (Km10) Mināb 27°08'N, 57°05'E (H11) Minushahr 30°20'N, 48°13'E (Kh13) Mohammadābād 28°57'N, 57°55'E (Km9) Mollāsāni 31°35'N, 48°52'E (Kh9) Mumān 25°35'N, 60°29'E (S19) Nagsh-e Rostam 29°59'N, 52°54'E (F10) Nāhārkhorān 36°45'N, 54°27'E (Gs2) Nehbandān 31°41'N, 60°02'E (KhJ2) Nikshahr 26°13'N, 60°13'E (S12) Nishābur 36°03'N, 59°06'E (KhR3)

Nosratābad 29°03'N, 57°56'E (Km8) Nowshahr 36°38'N, 51°30'E (M4) Nugh 34°17'N, 58°09'E (KhR8) Nurābād 30°06'N, 51°31'E (F4) Orumieh 37°33'N, 45°4'E (AG4) Orumieh lake (AG) Pākuh 27°55'N, 61°30'E (S6) Parehsar 37°37'N, 49°02'E (G3) Paskuh 27°33'N, 61°38'E (S6) Pishin 26°04'N, 61°25'E (S17) Pol-e Tang 32°51'N, 48°02'E (L1) Posht- e kuh 29°25'N, 56°50'E (Km6) Rafsanjān 30°24'N, 55°58'E (Km1) Rāmiān 37°01'N, 55°08'E (Gs1) Rāsk 26°14'N, 61°23'E (S15) Rek 32°15'N, 59°7'E (KhJ1) Rezvandeh 37°33'N, 49°09'E (G3) Robāt-e gharebil 37°21'N, 56°16'E (KhSh1) Robāt-e Tork 33°44'N, 50°50'E (Mz2) Rostamābād 36°54'N, 49°27'E (G7) Rudān 27°32'N, 57°25'E (H10) Rudbār 36°48'N, 49°23'E (G7) Rudbārak 36°04'N, 52°32'E (M5) Rudehen 35°44'N, 51°51'E (T6) Saādatshahr 30°06'N, 53°07'E (F10) Sabzevār 36°12'N, 57°40'E (KhR2) Sanandaj 35°19'N, 46°59'E (Kd2) Sangān 28°35'N, 61°19'E (S4) Sangrud 36°40'N, 49°38'E (G7) Sarāvān 27°08'N, 62°14'E (S7) Sarbāz 26°39'N, 61°14'E (S14) Sāri 36°33'N, 53°02'E (M9) Sarvestān 29°16'N, 53°12'E (F12) Sekand 26°43'N, 61°31'E (S14) Semirom 31°24'N, 51°34'E (E8) Sendarak 26°50'N, 57°25'E (H12) Sepidān 29°58'N, 52°23'E (F3) Shabestar 38°10'N, 45°41'E (Ash3) Shādegān 30°39'N, 48°40'E (Kh11) Shahdād 30°25'N, 57°42'E (Km4) Shāhkuh-e Pāiin 36°34'N, 54°26'E (Gs3) Shahr-e Kord 32°19'N, 50°51'E (Ch2) Shāhrud 36°25'N, 55°01'E (Sm1)

Shemirān 35°49'N, 51°24'E (T4) Shieh 35°05'N, 50°10'E (G6) Shirāz 29°37'N, 52°31'E (F9) Shul 30°02'N, 52°09'E (F9) Shurlukh 36°20'N, 60°35'E (KhR7) Shusf 32°00'N, 59°53'E (KhJ2) Shush 32°11'N, 48°14'E (Kh2) Shushtar 32°02'N, 48°51'E (Kh3) Silvāneh 37°25'N, 44°51'E (AG3) Sirik 26°30'N, 57°06'E (H13) Siriān 29°26'N, 55°40'E (Km2) Sisakht 30°47'N, 51°33'E (KB1) Sufiān 38°16'N, 45°58'E (Ash3) Susangerd 31°33'N, 48°11'E (Kh5) Suza 26°46'N, 56°03'E (H8) Tabas 33°35'N, 56°54'E (Y1) Tabriz 38°05'N, 46°17'E (Ash2) Taftān 28°36'N, 61°07'E (S4) Tākestān 36°03'N, 49°42'E (Gz2) Tamandān 28°36'N, 61°04'E (S4) Tang-e Sorkheh 26°29'N, 60°02'E (S11) Tazekand 38°16'N, 47°14'E (Ash5) Tehrān 35°41'N, 51°24'E (T4) Temin 28°40'N, 61°08'E (S4) Tis 25°21'N, 60°37'E (S19) Tochāl Mountain 35°53'N, 51°26'E (T4) Tonekābon 36°49'N, 50°52'E (M1) Varāmin 35°19'N, 51°39'E (T5) Veresk 35°56'N, 52°58'E (M12) Yāichi 37°35'N, 46°11'E (Ash2) Yāsuj 30°39'N, 51°35'E (KB2) Yazd 31°54'N, 54°20'E (Y2) Zābol 31°02'N, 61°29'E (S1) Zāboli 27°07'N, 61°40'E (S10) Zāhedān 29°29'N, 60°51'E (S2) Zanjān 36°40'N, 48°28'E (Z1) Zarghān 29°46'N, 52°43'E (F10) Zeidān 27°58'N, 51°31'E (B5) Ziārān 36°06'N, 50°31'E (T1) Ziārat 27°41'N, 57°05'E (H11) Zirāb 36°10'N, 52°59'E (M10) Zoshk 36°20'N, 59°10'E (KhR4)

RESEARCH ARTICLE



Synopsis of Falsocis Pic (Coleoptera, Ciidae), new species, new records and an identification key

Cristiano Lopes-Andrade^{1,†}, John F. Lawrence^{2,‡}

l Departamento de Biologia Animal, Universidade Federal de Viçosa, 36570-000, Viçosa, Minas Gerais, Brazil **2** Australian National Insect Collection, CSIRO Ecosystem Sciences, GPO Box 1700, Canberra, ACT 2601, Australia

t urn:lsid:zoobank.org:author:6A3F4101-8599-44D9-9210-BCD5DD8564E9 t urn:lsid:zoobank.org:author:5E0EBD0E-5EA3-4304-9914-0D0A4DC52B4E

Corresponding author: Cristiano Lopes-Andrade (ciidae@gmail.com)

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Abstract

Three new species of *Falsocis* Pic are described: *F. aquilonius* **sp. n.** from Panamá, Costa Rica and Colombia, *F. egregius* **sp. n.** from a single locality in northern Brazil and *F. occultus* **sp. n.** from two localities in southeastern and southern Brazil. New records, comparative notes and an identification key for male and female specimens of *Falsocis* species are also provided.

Keywords

Minute tree-fungus beetles, Ciinae, Neotropical, Tenebrionoidea

Introduction

Falsocis Pic, 1916 is a distinctive but relatively uniform genus of Neotropical Ciidae, recently redescribed by Lopes-Andrade (2007) and currently including two species: *E opacus* Pic, 1916, from French Guyana and northern Brazil; and *E brasiliensis* Lopes-Andrade, 2007, an endangered species occurring in small remnants of the Brazilian

Atlantic Forest in the states of Minas Gerais, Espírito Santo and Bahia. *Falsocis* species are not frequently collected and few specimens are known from museum collections.

The aims of this work are to provide new records and comparative notes on *Falsocis*, descriptions of three additional species of the genus and an updated identification key for the species.

Material and methods

Slide preparations of male terminalia, and methods for photographing slide preparations and specimens, followed mostly Lopes-Andrade (2011), except for the stereosmicroscope and camera used, a Zeiss Stemi 2000-C and a Canon EOS 1000D attached, respectively. Terms for external morphology of ciids are explained and discussed by Lopes-Andrade and Lawrence (2005) and Lopes-Andrade (2008). It must be emphasized that "tegmen", as used here, is synonymous with "paramere", "parameral piece" or "apicale" and not with the same term as used in Cucujoidea, Chrysomeloidea and Curculionoidea, which is essentially synonymous with phallobase or basal piece (see Lawrence and Lopes-Andrade 2010). The terms "low", "medium" and "high" (or "minor", "intermediate" and "major"; Scott 1926) refers to the conspicuousness of secondary sexual characters in male head and pronotum of ciids. Low males are the ones with the weakest horns and/or tubercles, being barely discernible from females. In medium males, these features are conspicuous but not developed at their most. High males have the most prominent horns and/or tubercles. Such variation in male development is common in ciids and usually seen as two categorical forms, low and high, in species known from few localities and/or small series (e.g. Cis pickeri Lopes-Andrade et al. and Xylographus seychellensis Scott; see Lopes-Andrade et al. 2009; Scott 1926) and seen as a continuum in widely distributed and frequently collected ciids (e.g. Ceracis cornifer Mellié; Lopes-Andrade pers. obs.). The new species described here were compared to named specimens of both described *Falsocis* species, and all type specimens were examined. Images of all described and new species are provided, but images of male terminalia of *F* opacus and *F* brasiliensis are available in Lopes-Andrade (2007) and not repeated here. The identification key was elaborated to work for both male and female specimens. However, it is not advisable to identify any Falsocis based on female specimens alone, since secondary sexual characters and male terminalia are important features for recognizing species.

Range, mean and standard deviation are given for measurements and ratios. Measurements of antennomeres were taken from the holotypes. The following abbreviations are used for measurements and ratios: CL, length of the antennal club; EL, elytral length (median length from base of scutellum to elytral apex); EW, greatest elytral width; FL, length of the antennal funicle; GD, greatest depth of the body; PL, pronotal length along midline (including the anterior pronotal plate in males); PW, greatest pronotal width; TL, total length (EL+PL; head not included). The ratio GD/EW was taken as an indication of degree of convexity; TL/EW indicates degree of body elongation. Values for TL and TL/EW of the type series of *F. brasiliensis* and the unique available male *F. opacus* are also provided here, since in Lopes-Andrade (2007) TL included the length of the head seen from above. Each description is based on the holotype, which is a fully pigmented male. Differences among paratypes are given in the section on "Variation", together with standard measurements and ratios of the type series.

The distribution map (Fig. 41) was created using latitude and longitude coordinates estimated by tracking localities in the freeware Google Earth 6.0.3 (Google Earth 2011) and plotting them in a map using the freeware DIVA-GIS 7.3.0 (Hijmans et al. 2001).

The following acronyms are used in this paper:

ANIC	Australian National Insect Collection, CSIRO Entomology (Canberra,
	Australia)
CMN	Canadian Museum of Nature (Ottawa, Ontario, Canada)
CMNH	Carnegie Museum of Natural History (Pittsburgh, Pennsylvania, USA)
CNCI	Canadian National Collection of Insects (Ottawa, Ontario, Canada)
FMNH	Field Museum of Natural History (Chicago, Illinois, USA)
LAPC	Cristiano Lopes-Andrade Private Collection (Viçosa, MG, Brazil)
MNHN	Muséum National d'Histoire Naturelle (Paris, France)
MTD	Museum für Tierkunde Dresden (Dresden, Germany)

Taxonomy

Falsocis aquilonius Lopes-Andrade & Lawrence, sp. n. urn:lsid:zoobank.org:act:01613AEF-7593-43F1-B83C-CB7F8A13AF21 http://species-id.net/wiki/Falsocis_aquilonius Figs 1–9, 41

Type locality. Cerro Campana, in the province of Panamá, Panama (8°26'N, 81°17'W).

Etymology. The specific epithet is from the Latin "aquilonius" (adjective), which means "from the North", in reference to its occurrence at the northernmost locality for *Falsocis* species.

Diagnosis. Epipleura enlarged posteriorly, but just slightly explanate (Figs 2–3), not or barely visible from above, with external margin simple (not crenulate). Pronotum with lateral margins visible for their entire lengths from above. Male abdominal sex patch small with a diameter of one-fifth the length of the first abdominal ventrite.

Description. Male holotype (Figs 1–3), measurements in mm: TL 2.21; PL 0.95; PW 1.11; EL 1.26; EW 1.16; GD 0.89. Ratios: PL/PW 0.86; EL/EW 1.09; EL/PL 1.33; GD/EW 0.77; TL/EW 1.91. Body oblong, strongly convex, mostly yellowish brown; coxae and femora pale yellow; mesoventrite, metaventrite and first abdominal ventrite whitish. Head not visible from above; frontoclypeal ridge slightly raised,



Figures 1–6. *Falsocis aquilonius* Lopes-Andrade & Lawrence, sp. n., male holotype 1–3, shown in the same scale, female paratype from Puntarenas, Costa Rica 4 and slide preparation of male terminalia of a paratype 5–6. I Dorsal view 2 Lateral view 3 Ventral view 4 Dorsal view 5 Eighth sternite 6 Aedeagus (left) and gross outline of aedeagus (right) showing tegmen (continuous line), penis (dashed line) and basal piece (dotted line).

bearing two very short, barely pronounced tubercles; disc slightly convex, closely and coarsely punctate, glabrous; in between punctures finely granulate. Eyes coarsely facetted; greatest eye width 0.15mm. Each antenna (left antenna measured; FL 0.18mm; CL 0.18mm; CL/FL 1.00) with length of antennomeres (in mm) as follows: 0.08; 0.06; 0.06; 0.05; 0.03; 0.02; 0.02; 0.05; 0.05; 0.08. Pronotum with single and coarse punctation; punctures very close to each other on disc and near the lateral and posterior margins, but somewhat shallower and separated by a distance of about one puncture-width at the anterior projection; in between punctures finely granulate; vestiture gravish, dual (seen in lateral view under a magnification of 100×), consisting of conspicuous stout erect bristles (~0.05mm) and very small decumbent setae (a bit less than 0.02mm); anterior angles rounded, produced forward; anterior edge explanate and produced forward forming a plate that slightly curves upward and tapers toward a rounded apex; lateral margins slightly explanate and visible for their entire lengths from above, irregularly crenulate. Hindwings fully developed. Scutellum subtriangular, densely covered by stout decumbent bristles (better seen in lateral view); basal width 0.15mm. Sides of elytra parallel at basal three-fourths, broadly rounded posteriorly (seen from above) and converging to a blunt apex; punctation single and confused; punctures coarse and separated by a distance of one puncture-width or less; in between punctures dull; vestiture dual, similar to that of pronotum; lateral and apical margins not visible from above; epipleura tapering from base to middle, then enlarging to apex, with external margin simple (not crenulate). Ventral surface of thorax and abdomen finely granulate, with vestiture of decumbent slender setae. Prosternum flat; prosternal process laminate, three-fourths the length of procoxae. First abdominal ventrite twice as long as the second at midline; setose sex patch circular and margined, located near the posterior margin and with a diameter of one-fifth the length of the ventrite at midline.

Male terminalia in paratypes (Figs 5–6). Eighth sternite (Fig. 5) with posterior margin straight; posterior angles slightly produced forming two small lateral prominences. Basal piece (Fig. 6) nearly one-third the length of tegmen. Tegmen (Fig. 6) with anterior portion subtriangular; lateral margins straight, slightly diverging to apex; posterior portion with a median V-shaped emargination of about one-third the length of tegmen, forming two lateral lobes. Penis (Fig. 6) as long as tegmen, subcylindrical; lateral margins straight for half of their lengths then slightly curved inwards to a narrowly rounded apex.

Females (Fig. 4). Similar to males, but frontoclypeal ridge straight, barely sinuous, with rounded angles. Anterior pronotal margin broadly rounded. Abdomen devoid of sex patch.

Variation. Males, measurements in mm (n = 6, including the holotype): TL 2.00– 2.84 (2.45 ± 0.32); PL 0.74–1.32 (1.02 ± 0.20); PW 1.00–1.37 (1.16 ± 0.12); EL 1.21–1.47 (1.37 ± 0.11); EW 1.05–1.42 (1.23 ± 0.12); GD 0.84–1.16 (0.97 ± 0.13). Ratios: PL/PW 0.74–0.96 (0.87 ± 0.09); EL/EW 1.04–1.17 (1.12 ± 0.05); EL/PL 1.12–1.64 (1.37 ± 0.18); GD/EW 0.74–0.88 (0.79 ± 0.05); TL/EW 1.90–2.17 (1.99 ± 0.10). The anterior plate (projected anterior margin) of pronotum in males varies from very small (Fig. 7), medium size (Fig. 8, similar to that of holotype) to extremely projected (Fig. 9); but the apex of this plate is rounded or blunt and devoid of tufts. Females, measurements in mm (n = 5): TL 2.21–2.53 (2.38 ± 0.14); PL 0.79–0.89 (0.83 ± 0.04); PW 1.11–1.37 (1.24 ± 0.10); EL 1.37–1.63 (1.47 ± 0.11); EW 1.16–1.47



Figures 7–9. Falsocis aquilonius Lopes-Andrade & Lawrence, sp. n., male paratypes, dorsal view, shown in the same scale. 7 Low male from Cerro Campana (Panama) 8 Medium male from La Lola (Costa Rica)
9 High male from Puntarenas (Costa Rica).

(1.33 ± 0.13); GD 0.95–1.16 (1.05 ± 0.08). Ratios: PL/PW 0.60–0.81 (0.67 ± 0.08); EL/EW 1.04–1.27 (1.12 ± 0.09); EL/PL 1.63–1.94 (1.78 ± 0.15); GD/EW 0.74–0.95 (0.80 ± 0.09); TL/EW 1.68–2.14 (1.81 ± 0.19).

Type series. *Holotype.* (ANIC) **Panama:** \R.P.: Panama Cerro Campana Feb. 22, 1975 Lawrence, Erwin \ *Phellinus* sp. \ J.F. Lawrence Lot [printed] 3799 [handwritten] \ *Falsocis aquilonius* Lopes-Andrade & Lawrence HOLOTYPUS [printed on red paper]\. *Panatypes.* **Panama:** 1 male and 1 female (ANIC), same data as holotype; 3 males (1 ANIC, 2 LAPC), 4 females (2 ANIC, 1 LAPC) \2.5mi S.W.Rincon Puntarenas, Costa Rica, Mar. 1–7, '67 \ J.F. Lawrence Lot. [printed] 2175 [handwritten] \ R. Andrews Collector\. **Costa Rica.** 1 male (ANIC) \La Lola, C.R. III-8–1958 M.J. Stelzer [handwritten] \ MS 58.4 [handwritten]\; 1 female (ANIC) \Puerto Viejo COSTA RICA VIII-4–65 \ J.F. Lawrence Lot [printed] 1611 [handwritten]\. **Colombia.** 1 male (Marseul Collection, MNHN) \Dupont Colomb [18]41 [handwritten in circular green paper] \ MARSEUL [handwritten in rectangular small green paper] \ [circular blue paper, without information] \ FALSOCIS SPP. [handwritten] det. J.F. Lawrence 19 [printed]\. All paratypes distinguished labeled *Falsocis aquilonius* Lopes-Andrade & Lawrence PARATYPUS [printed on yellow paper]\.

Comments and comparative notes. One specimen collected in an unidentified *Phellinus*. This is the only *Falsocis* species known from Central America, but it also occurs at the northernmost South America (Fig. 41). Recent field collections in Colombia were unsuccessful in recapturing the species, so the country record is based only on a single old male without precise locality data. In *F. opacus*, the epipleura is much more enlarged, easily visible from above, with external margin crenulate; the male eighth

sternite is similar, but basal piece is strongly developed and is almost half the length of aedeagus; tegmen is elongate, four times as long as wide. In the remaining *Falsocis*, the epipleura is narrower near the apex.

Falsocis egregius Lopes-Andrade & Lawrence, sp. n. urn:lsid:zoobank.org:act:E6BFE936-A896-449F-8D6A-ADE8528F8728 http://species-id.net/wiki/Falsocis_egregius Figs 10–16, 41

Type locality. Santarém, in the state of Pará, northern Brazil (2°26'S, 54°42'W).

Etymology. The specific epithet is from the Latin "egregius" (adjective), which means "singular", "extraordinary", in a reference to the outstanding head morphology of males.

Diagnosis. Head of male with sides of vertex produced laterally to form a pair of triangular plates, each with acute apex (Fig. 13, arrows); frontoclypeal horns wide at base and tapering to apex, slightly arcuate. Pronotum with anterior angles broadly rounded, barely produced forward; males with a row of long setae along the apex of the anterior projection but not forming distinct tufts (Figs 10, 12, arrows).

Description. Male holotype (Figs 10-13), measurements in mm: TL 2.85; PL 1.25; PW 1.50; EL 1.55; EW 1.55; GD 1.35. Ratios: PL/PW 0.83; EL/EW 1.00; EL/ PL 1.24; GD/EW 0.87; TL/EW 1.84. Body oblong, strongly convex, mostly light yellowish brown; mouthparts and apices of femora reddish brown; ventral vestiture consisting of very fine decumbent setae. Head strongly developed, the anterior and lateral margins easily visible from above; dorsum concave, slightly tumid on disc; punctation relatively fine, shallow, sparse; in between punctures finely granulate; frontoclypeal ridge explanate, each side produced upward forming a conspicuous horn that is slightly arcuate, bearing two small tubercles between them; each side of vertex explanate and produced laterally forming a conspicuous triangular plate with an acute apex (Fig. 13, arrows). Eyes coarsely facetted; greatest width 0.18mm. Each antenna (left antenna measured; FL 0.22mm; CL 0.23mm; CL/FL 1.05) with length of antennomeres (in mm) as follows: 0.12; 0.07; 0.08; 0.04; 0.04; 0.03; 0.03; 0.06; 0.06; 0.11. Pronotum with shallow, coarse, single punctation; punctures separated by a distance about a puncture-width; in between punctures smooth at disc and finely granulate at the anterior projection; vestiture indistinctly dual, consisting of conspicuous yellowish stout erect bristles (~0.05mm) and very fine decumbent setae (<0.02mm); anterior angles broadly rounded, just slightly produced forward; anterior margin (beyond anterior angles) explanate, produced forward forming a plate that slightly curves downward and narrows toward a slightly arcuate apex with a row of very long slender setae (>0.35mm; Figs 10, 12, arrows). Scutellum subtriangular; punctation finer than those of pronotum and elytra; vestiture consisting of stout decumbent bristles (better seen in lateral view); basal width 0.21mm. Hindwings fully developed. Elytra subparallel at basal two-thirds, with posterolateral angles broadly rounded (as seen from above) and then converging to a blunt apex; punctation single and confused, a bit sparser than that of



Figures 10–16. *Falsocis egregius* Lopes-Andrade & Lawrence, sp. n., male holotype 10–13 and female paratype 14 shown in the same scale, and slide preparation of male terminalia of a paratype 15–16. 10 Dorsal view (pronotal tuft of long setae, arrow) 11 Lateral view 12 Ventral view (pronotal tuft of long setae, arrow) 13 Head and pronotum view from above (triangular plate behind eyes, arrows) 14 Female 15 Eighth sternite 16 Aedeagus (left) and gross outline of aedeagus (right) showing tegmen (continuous line), penis (dashed line) and basal piece (dotted line).

pronotum; in between punctures smooth; vestiture indistinctly dual, consisting of conspicuous stout erect bristles (~0.07mm) and very minute decumbent setae (<0.02mm); lateral and apical margins not visible from above; epipleura tapering from base to apex. Surface of the ventral thoracic and abdominal sclerites granulate, somewhat rugose. Prosternum flat; prosternal process laminate, almost half the length of procoxae. First abdominal ventrite more than twice as long as the second at midline; setose sex patch circular and margined, located at the middle of the ventrite and with a diameter of one-third the length of the ventrite at midline.

Male terminalia in paratypes (Figs 15–16). Eighth sternite (Fig. 15) with posterior margin curved inward; angles not produced. Basal piece (Fig. 16) almost half the length of tegmen. Tegmen (Fig. 16) with anterior portion broadly rounded; lateral margins almost straight; posterior portion bearing a deep V-shaped emargination reaching the middle of the structure and forming two lateral lobes. Penis (Fig. 16) subcylindrical, lateral margins straight for most of their lengths and a bit expanded to a rounded apex; the penis is turned and lays laterally in the slide preparation.

Female (Fig. 14). Similar to males, but frontoclypeal ridge straight, barely sinuous, with rounded angles. Anterior pronotal margin broadly rounded. Abdomen devoid of sex patch.

Variation. Males, measurements in mm (n = 5, including the holotype): TL 2.75–2.95 (2.87 \pm 0.08); PL 1.20–1.35 (1.26 \pm 0.05); PW 1.50–1.55 (1.52 \pm 0.03); EL 1.50–1.65 (1.56 \pm 0.05); EW 1.55–1.65 (1.57 \pm 0.04); GD 1.30–1.40 (1.35 \pm 0.04). Ratios: PL/PW 0.80–0.87 (0.83 \pm 0.03); EL/EW 0.97–1.00 (0.99 \pm 0.01); EL/PL 1.15–1.32 (1.24 \pm 0.06); GD/EW 0.84–0.87 (0.86 \pm 0.02); TL/EW 1.77–1.87 (1.83 \pm 0.05).

Female, measurements in mm (n = 1): TL 2.50; PL 0.95; PW 1.45; EL 1.55; EW 1.50; GD 1.05. Ratios: PL/PW 0.66; EL/EW 1.03; EL/PL 1.63; GD/EW 0.70; TL/EW 1.67.

Type series. *Holotype.* (CMNH) **Brazil:** \Santarem Brazil Acc. No.2966 \ CM \ *Falsocis egregius* Lopes-Andrade & Lawrence HOLOTYPUS [printed on red paper]\. *Paratypes.* **Brazil:** 4 males (1 ANIC; 2 CMNH; 1 LAPC, dissected) and 1 female (CMNH) \Santarem Brazil Acc. No.2966 \ CM\, same data as holotype. All paratypes distinguished labeled *Falsocis egregius* Lopes-Andrade & Lawrence PARATYPUS [printed on yellow paper]\.

Comments and comparative notes. The species is currently known from a single collection in Santarém, northern Brazil (Fig. 41). Only one female was available for examination. The series is very small, but species sufficiently distinct to allow description. Differs from *F. aquilonius* sp. n. and *F. opacus* in having the epipleura narrow posteriorly. *Falsocis occultus* sp. n. has deeper and closer pronotal punctation. Very similar to *F. brasiliensis*, but differing mainly in features mentioned in the diagnosis and in the conspicuous basal piece of male terminalia. In *F. brasiliensis*, the eighth sternite has the anterior margin only slightly curved inward; the basal piece was not observed in the available slide preparations and is possibly membranous (Lopes-Andrade 2007); the tegmen has a subtriangular posterior portion, subparallel lateral margins and anterior V-shape emargination is only one-fourth to one-third deep.

Falsocis occultus Lopes-Andrade & Lawrence, sp. n.

urn:lsid:zoobank.org:act:EF30CDE6-0A8A-444A-9A89-0ECB68E9CB17 http://species-id.net/wiki/Falsocis_occultus Figs 17–25, 41

Type locality. Linhares, in the state of Espírito Santo, southeastern Brazil (19°23'S, 40°04'W).

Etymology. The specific epithet is from the Latin "occultus" (adjective), which means "hidden", in reference to the fact that the population from the type locality stayed hidden among fungi forgotten in a field-base for near four years.

Diagnosis. Pronotum with anterolateral angles not produced forward; lateral margins not visible from above; male with anterior projection ending in an acute apex (Figs 19, 23-25, arrows) bordered by medium-size bristles.

Description. Male holotype (Figs 17-19), measurements in mm: TL 3.55; PL 1.70; PW 1.75; EL 1.80; EW 1.75; GD 1.40. Ratios: PL/PW 0.97; EL/EW 1.03; EL/PL 1.06; GD/EW 0.80; TL/EW 2.03. Body oblong, strongly convex, mostly dark brown; mouthparts, antennae and tarsi dark yellowish brown; femora and tibiae dark reddish brown. Head concealed by the anterior pronotal projection (seen from above) except for its anterolateral angles; dorsum concave with disc slightly tumid; punctation coarse, shallow; in between punctures finely granulate; frontoclypeal ridge explanate and produced forming a broad acute triangular plate at each anterior angle, with two small tubercles between them. Eyes coarsely facetted; greatest eye width 0.21mm. Each antenna (left antenna measured; FL 0.30mm; CL 0.28mm; CL/FL 0.93) with length of antennomeres (in mm) as follows: 0.15; 0.09; 0.08; 0.08; 0.06; 0.05; 0.03; 0.08; 0.08; 0.12. Pronotum with single, coarse, relatively deep punctation; punctures very close to each other, separated by a distance of one puncture-width or less; in between punctures smooth but not shining; vestiture yellowish, indistinctly dual (seen under a magnification of 100x), consisting of stout erect bristles (~0.05mm) and small decumbent setae (~0.03mm), both better seen in lateral view; anterior angles not produced forward; anterior margin (beyond anterior angles) explanate, strongly produced forward forming a plate that slightly curves downward and narrows toward an acute apex (Fig. 19, arrow) ornamented by a row of increasingly stout bristles in either side (Figs 17-19); lateral margins slightly crenulate, not visible from above, bearing a row of stout bristles. Scutellum subtriangular, its margins indistinct so that it seems to be contiguous with elytra; punctation conspicuous but slightly finer than those of pronotum and elytra; vestiture consisting of stout decumbent bristles (better seen in lateral view); basal width 0.20mm. Hindwings fully developed. Elytra parallel at basal threefourths, posteriorly broadly rounded (as seen from above) and converging to a blunt apex; punctation single and confused, slightly finer than that of pronotum, consisting of relatively deep punctures separated by a distance near a puncture-width; in between punctures smooth, dull; vestiture distinctly dual, the erect bristles about 0.1mm long



Figures 17–22. *Falsocis occultus* Lopes-Andrade & Lawrence, sp. n., male holotype 17–19 and female paratype 20 shown in the same scale, and slide preparation of male terminalia of a paratype 21–22. 17 Dorsal view 18 Lateral view 19 Ventral view (acute pronotal apex, arrow) 20 Dorsal view 21 Eighth sternite 22 Aedeagus (left) and gross outline of aedeagus (right) showing tegmen (continuous line), penis (dashed line) and basal piece (dotted line).



Figures 23–25. *Falsocis occultus* Lopes-Andrade & Lawrence, sp. n., male paratypes **23–24** shown in the same scale, dorsal view (acute pronotal apex, arrows). **23** Low male from Linhares (in the state of Espírito Santo, southeastern Brazil) **24** Medium male from Linhares **25** The unique specimen known from Nova Teutônia (in the state of Santa Catarina, southern Brazil).

and the decumbent setae about 0.03mm long; lateral and apical margins not visible from above; epipleura tapering from base to the basal one-sixth, then continuing as a narrow line to the apex. Ventral surfaces of thorax and abdomen finely granulate; vestiture consisting of slender decumbent setae. Prosternum flat; prosternal process laminate, two-thirds the length of the procoxae. First abdominal ventrite more than twice as long as the second at midline; setose sex patch suboval and margined, located at the middle of the ventrite and with a diameter of one-third the length of the ventrite at midline.

Male terminalia in paratypes (Figs 21–22). Eighth sternite (Fig. 21) with posterior margin almost straight; angles not produced. Basal piece (Fig. 22) nearly one-third the length of tegmen. Tegmen (Fig. 22) with anterior portion mostly rounded but apex acute; lateral margins slightly sinuous and diverging; posterior portion bearing a deep V-shaped emargination reaching the middle of the structure, forming two lateral lobes. Penis (Fig. 22) subcylindrical; lateral margins subparallel for most of their lengths; apical third subtriangular, membranous.

Females (Fig. 20). Similar to males, but frontoclypeal ridge straight, barely sinuous, with rounded angles. Anterior pronotal margin broadly rounded. Abdomen devoid of sex patch.

Variation. Males, measurements in mm (n = 15, including the holotype): TL 2.11– 3.55 (2.84 \pm 0.42); PL 0.84–1.70 (1.28 \pm 0.25); PW 1.16–1.75 (1.48 \pm 0.17); EL 1.25– 1.80 (1.53 \pm 0.19); EW 1.21–1.84 (1.52 \pm 0.17); GD 0.95–1.47 (1.22 \pm 0.16). Ratios: PL/PW 0.73–0.97 (0.86 \pm 0.07); EL/EW 0.86–1.07 (1.01 \pm 0.05); EL/PL 1.00–1.50 (1.22 ± 0.15) ; GD/EW 0.72–0.93 (0.80 ± 0.05); TL/EW 1.73–2.03 (1.86 ± 0.09). In the unique specimen from Nova Teutonia, a teneral male, pronotal and elytral bristles are larger (0.08mm and 0.15mm, respectively) than in specimens from Linhares. In small males, the anterior pronotal plate is barely projected (Fig. 23). However, all males have an acute pronotal apex (Figs 19, 23–25, arrows). Females, measurements in mm (n = 15): TL 2.30–3.15 (2.63 ± 0.27); PL 0.80–1.16 (0.97 ± 0.11); PW 1.05–1.75 (1.42 ± 0.19); EL 1.40–2.00 (1.64 ± 0.18); EW 1.10–1.85 (1.50 ± 0.19); GD 1.05–1.50 (1.23 ± 0.15). Ratios: PL/PW 0.62–0.81 (0.69 ± 0.05); EL/EW 1.00–1.41 (1.11 ± 0.10); EL/PL 1.50–2.00 (1.71 ± 0.15); GD/EW 0.79–0.95 (0.83 ± 0.04); TL/EW 1.64–2.23 (1.77 ± 0.14).

Type series. *Holotype.* (LAPC) **Brazil:** \BRASIL: ES Linhares 11–21.x.2004 P.C. Grossi leg. \ *Falsocis occultus* Lopes-Andrade & Lawrence HOLOTYPUS [printed on red paper]\. *Paratypes.* **Brazil:** 56 (27 males, 1 dissected, and 19 females, LAPC; 5 males and 5 females, ANIC), same data as holotype; 1 male (FMNH) \Nova Teutonia, Sta. Catharina, BRAZ. 300–500m alt. Fritz Plaumann leg [printed] XI:1940 [handwritten] \ [circular red paper, without information] \ Falsocis sp. 115 \. All paratypes distinguished labeled \ *Falsocis occultus* Lopes-Andrade & Lawrence PARATYPUS [printed on yellow paper]\.

Other specimens examined. 26 specimens, gender not determined (2 CNCI for molecular analysis, 4 LAPC in absolute alcohol, 20 MTD), same data as holotype.

Comments and comparative notes. Known from two localities, in southeastern and southern Brazil (Fig. 41). The specimens from Linhares (in the state of Espírito Santo, southeastern Brazil) were collected in *Hexagonia papyracea* Berk. (Polyporaceae) and bred in the laboratory until December 2009 in the original basidiomes, without addition of either water or nutrients. In male *F. brasiliensis* and *F. egregius* sp. n., the apex of the pronotal projection bears a row of setae that are comparatively longer (Figs 10–12, 26–29, 33–34). Differs from *F. aquilonius* sp. n. and *F. opacus* in having the epipleura narrow posteriorly.

New distributional data, additional comments and comparative notes

Falsocis brasiliensis Lopes-Andrade, 2007 http://species-id.net/wiki/Falsocis_brasiliensis Figs 26–34, 41

Additional records. Brazil: 1 female (LAPC) \BRASIL: ES Santa Maria de Jetibá 08.xii.2003 leg. Furieri & Nunes\; 3 males (1 LAPC, in absolute alcohol; 2 CNCI, for molecular analysis) and 1 female (LAPC, in absolute alcohol) \BRASIL: MG Viçosa "Belvedere" 03.ii.2011 leg. L.S. Araújo, C.A. Carvalho\; 2 males and 1 female (LAPC) \BRASIL: MG Viçosa Mata da Biologia, ponto 37 16.iv.2010; leg. T. Mariani & C. Lopes-Andrade; ex *Hymenochaete luteobadia*\; 2 males (LAPC) \BRASIL: MG Viçosa Mata da Biologia, ponto 37 16.iv.2010; leg. T. Mariani & C. Lopes-Andrade; ex *Hymenochaete luteobadia*\; 2 males (LAPC) \BRASIL: MG Viçosa Mata da Biologia, ponto 37 16.iv.2010; leg. T. Mariani & C. Lopes-Andrade; ex *Hymenochaete luteobadia*\.

Comments. The specimens from Viçosa (in the state of Minas Gerais, southeastern Brazil; Figs 26–31) were all collected in *Hymenochaete luteobadia* (Fr.) Höhn. & Litsch.



Figures 26–31. *Falsocis brasiliensis* Lopes-Andrade, 2007, specimens from Viçosa (in the state of Minas Gerais, southeastern Brazil), all shown in the same scale. 26 Dorsal view (produced angles, large arrows; two tufts of long setae, small arrows) 27 Lateral view 28 Ventral view (produced angles, large arrows; two tufts of long setae, small arrows) 29 Head and pronotum view from above 30 Low male 31 Female.

(Hymenochaetacea). This fungus was previously misidentified as *Phellinus* sp. (Lopes-Andrade 2007, Graf-Peters et al. 2011). After its description, the species was recollected in Viçosa and a single female was found in Santa Maria de Jetibá (in the state of Espírito Santo, southeastern Brazil; Fig. 32). The species is restricted to forests, known only from small remnants, and was never found in open areas. Specimens from Venda Nova do Imigrante (Espírito Santo; Fig. 33) are reddish, and the ones from Jussari (in the state of Bahia; Fig. 34) are dark brown. In low males (Fig. 30), the frontoclypeal and pronotal projections are very weak and the pronotum narrower anteriorly. TL (in mm, not including the head) and TL/EW of the type series are as follows: Males (n = 10, including the holotype), TL 2.00–2.80 (2.25 \pm 0.22), TL/EW 1.54–1.81 (1.64 \pm 0.07); Females (n = 5), TL 2.00–2.35 (2.14 \pm 0.14), TL/EW 1.48–1.68 (1.60 \pm 0.08).


Figures 32–34. *Falsocis brasiliensis* Lopes-Andrade, 2007, dorsal view, shown in the same scale. **32** The unique female specimen from Santa Maria de Jetibá, in the state of Espírito Santo, southeastern Brazil **33** Male paratype from Venda Nova do Imigrante, Espírito Santo (two tufts of long setae, arrows) **34** Male paratype from Jussari (in the state of Bahia).

Falsocis opacus Pic, 1916

http://species-id.net/wiki/Falsocis_opacus Figs 35–37, 41

Comments. The subspecies *F. opacus flavus* Pic, 1922 was based on a single female collected in Bas-Maroni (French Guyana, near Cayenne), and it is merely a teneral specimen of *F. opacus* (J. F. Lawrence pers. obs.). The type of *F. opacus* (MNHN) is 2.10mm long, rather than 3mm as mentioned in the description (Pic 1916), but other measurements were not taken by the time it was examined. In the unique male *F. opacus* measured by Lopes-Andrade (2007), TL is 3.05mm and TL/EW 1.91, not including the head.

Falsocis sp.

http://species-id.net/wiki/Falsocis Figs 38–41

Record. 1 female (MNHN) **Peru:** \Mission de Sarayacu (Riv. Oucayale) [Ucayali] [handwritten in green paper] \ CO 47 [handwritten in a circular yellowish paper] \ MUSEUM PARIS PÉROU PAMPAS [sic] DEL SACRAMENTO DE CASTELNAU 1847 [written in light brown paper] \ [circular blue paper, without information]\.

Comments and comparative notes. The specimen was collected at Sarayacu Mission (6°47'S, 75°07'W, Fig. 41) during the French Scientific Expedition to the Pampa del Sacramento, Peru, in 1847. It resembles females of *F. brasiliensis*, *F. egregius* sp. n.



Figures 35–40. *Falsocis opacus* Pic, 1916, from Altamira, in the state of Pará, northern Brazil 35–37; arrows pointing out the explanate outer apical margins of epipleura and *Falsocis* sp., female from Peru 38–40, all shown in the same scale. 35, 38 Dorsal view 36, 39 Lateral view 37, 40 Ventral view.

and *F. occultus* sp. n. in having the epipleura narrow posteriorly. However, the shallow punctation of pronotum resembles mostly that of the former two species. Female *F. brasiliensis* from Jussari (Brazil) are similar in color, but body is more elongate. It is very large, with a length of near 3mm. Only females of *F. occultus* sp. n. reach such a length. A subquadrate body is also observed in a few *F. brasiliensis*, but the largest females of this species are only 2.4mm long. The very transverse pronotum, twice as long as wide, does not occur in any other known *Falsocis* species. This female from Peru possibly belongs to another new species, but description is not possible without examining a male. Measurements (in mm) and ratios are as follows: TL 3.05; PL 1.00; PW 2.00; EL 2.00; EW 2.11; GD 1.68; PL/PW 0.50; EL/EW 0.95; EL/PL 2.00; GD/EW 0.80; TL/EW 1.45.



Figure 41. Distribution map for *Falsocis* Pic species. *Falsocis aquilonius* sp. n. (full circle), *F. brasiliensis* Lopes-Andrade (full squares), *F. egregius* sp. n. (open triangle), *F. occultus* sp. n. (open circles), *F. opacus* Pic (x) and *Falsocis* sp. (open star inside full circle). The question mark indicates an imprecise location of *F. aquilonius* sp. n. in Colombia.

Identification key to the species of Falsocis Pic

1	Epipleura enlarged posteriorly, forming a slightly or strongly explanate poste-
	rolateral margin in elytra, conspicuously or barely visible from above2
1'	Epipleura narrowing to apex, forming a narrow, not explanate, posterolateral
	margin in elytra, not visible from above
2(1)	Pronotum with lateral margins visible from above. Posterior elytral margin
	(outer margin of epipleura) simple (not crenulate), barely visible from above.
	Known from Panamá, Costa Rica and Colombia Falsocis aquilonius sp. n.
2'	Pronotum with lateral margins not visible from above. Posterior elytral mar-
	gin (outer margin of epipleura) crenulate, easily visible from below and above
	(Figs 35-37, arrows). Known from Brazil (Altamira, in the state of Pará) and
	French Guyana Falsocis opacus Pic, 1916
3(1')	Pronotal punctation deep and coarse, with punctures usually separated by
	half a puncture-width or less at disc. Anterior plate of pronotum in males

	devoid of tufts of very long setae and acute at apex (Figs 19, 23-25)
3'	Pronotal punctation shallow and fine, with punctures usually separated by
	about a puncture-width at disc. Anterior plate of pronotum in males with
	slender setae usually as long as to twice as long (or more) than an eye-width,
	with apex straight, barely emarginated or slightly rounded4
4(3')	Anterior pronotal angles distinctly produced forward and somewhat acute
	(Figs 26, 28, large arrows). Males with head devoid of conspicuous projection
	behind each eye (Fig. 29); long setae of anterior pronotal plate organized in
	two tufts (Figs 26, 28, 33, small arrows)
4'	Anterior pronotal angles barely produced forward and rounded (Figs 10-12).
	Males with head bearing a conspicuous triangular plate, projected outward, be-
	hind each eye (Fig. 13, arrows); long setae of anterior pronotal plate organized
	as a row, not forming tufts (Figs 10, 12, arrows)
	as a row, not forming tufts (Figs 10, 12, arrows)

Discussion

The phylogenetic position of *Falsocis* is uncertain. In the available phylogenetic analysis of the family, based on molecular data, *F. brasiliensis* was not part of any defined clade (Buder et al. 2008). It is morphologically related to Acanthocis Miyatake from Japan, with a unique combination of features among Ciini (Lopes-Andrade 2007): prominent outer apical angle of protibiae, forming an acute tooth; laminate prosternal process; and subconical procoxae. However, these features are also observed in Porculus Lawrence, in which species also have a very elongate female ovipositor similar to that of *Falsocis*, male terminalia with tegmen bearing a deep V-shaped emargination and subcylindrical penis (Lopes-Andrade pers. obs). Porculus differs from Falsocis and Acanthocis in the single punctation of pronotum and elytra, and dorsal vestiture consisting of minute setae (Lawrence 1987, Lopes-Andrade 2007). A strongly sclerotized female ovipositor devoid of gonostyli was observed in *P. grossus* Lawrence (Lawrence and Lopes-Andrade 2010), but in *P. brunneus* (Mellié) the structure is complete and less sclerotized, similar to that of Falsocis species. Falsocis and Porculus are restricted to the Neotropical region (sensu Morrone 2002), and Acanthocis is known only from Japan (Lawrence and Lopes-Andrade 2010). These three genera possibly constitute an independent lineage of Ciinae.

Species of *Falsocis* are not frequently collected and all are allopatric. They are known only from forests or forest remnants and usually one or few specimens are found together in the field. The greatest series is that for *F. occultus* sp. n., which was reared in laboratory. Accurate data on host fungi are scarce. Available information on *F. brasiliensis* suggests the species is monophagous (Graf-Peters et al. 2011). It was considered an endangered species by Lopes-Andrade (2007). However, not only this species but possibly all *Falsocis* are seriously threatened or endangered. For instance, besides great collection efforts in the state of Rio Grande do Sul, southern Brazil (Graf-Peters et al. 2011) and in an unpublished

survey in Pará, northern Brazil (Lopes-Andrade pers. obs.), no *Falsocis* was collected. Most of the recent records (from 2000 up to know) are from the Brazilian Atlantic Forest, from small remnants in southeastern and the most southern portion of northeastern Brazil. The single old record of *F. occultus* sp. n. from Nova Teutônia, in the state of Santa Catarina, suggests the species would have had a very broad distribution throughout the Atlantic Forest. This biome has been seriously affected and fragmented by land use and urbanization, and less than 8% of the original forests still remain (Colombo and Joly 2010).

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References

- Buder G, Grossmann C, Hundsdoerfer A, Klass K-D (2008) A contribution to the phylogeny of the Ciidae and its relationships with other cucujoid and tenebrionoid beetles (Coleoptera: Cucujiformia). Arthropod Systematics & Phylogeny 66 (2): 165–190.
- Colombo AF, Joly CA (2010) Brazilian Atlantic Forest lato sensu: the most ancient Brazilian forest, and a biodiversity hotspot, is highly threatened by climate change. Brazilian Journal of Biology 70(3): 697–708. doi: 10.1590/S1519-69842010000400002
- Google Earth (2011) Google Earth, 6.0.3. Google Software. [Available from: http://earth.google.com/]
- Graf-Peters LV, Lopes-Andrade C, Da Silveira RMB, Moura LD, Reck MA, De Sá FN (2011) Host fungi and feeding habits of Ciidae (Coleoptera) in a subtropical rainforest in southern Brazil, with an overview of host fungi of Neotropical ciids. Florida Entomologist 94(3): 553–566. doi: 10.1653/024.094.0320
- Hijmans RJ, Guarino L, Cruz M, Rojas E (2001) Computer tools for spatial analysis of plant genetic resources data: 1. DIVA-GIS. Plant Genetic Resources Newsletter 127: 15–19.
 [Program available from: http://www.diva-gis.org; downloaded in 19/January/2011]

- Lawrence JF (1987) A new genus of Ciidae (Coleoptera, Tenebrionoidea) from the Neotropical region. Revista Brasileira de Entomologia 31(1): 41–47.
- Lawrence JF, Lopes-Andrade C (2010) Ciidae Leach in Samouelle, 1819. In: Leschen RAB, Beutel RG, Lawrence JF (Eds) Handbuch der Zoologie/Handbook of Zoology. Band/Volume IV Arthropoda: Insecta. Teilband/Part 38. Coleoptera, Beetles. Volume 2. Morphology and Systematics (Polyphaga partim). Walter de Gruyter, Berlin, 504–514.
- Lopes-Andrade C (2007) Notes on *Falsocis* Pic (Coleoptera: Tenebrionoidea: Ciidae), with the description of an endangered Brazilian species. Zootaxa 1544: 41–58.
- Lopes-Andrade C (2008) An essay on the tribe Xylographellini (Coleoptera: Tenebrionoidea: Ciidae). Zootaxa 1832: 1–110.
- Lopes-Andrade C (2011) The first *Strigocis* Dury (Coleoptera, Ciidae) from the southern Neotropical region and a provisional key to world species. Zookeys 81: 27–37. doi: 10.3897/ zookeys.81.940
- Lopes-Andrade C, Lawrence JF (2005) *Phellinocis*, a new genus of Neotropical Ciidae (Coleoptera: Tenebrionoidea). Zootaxa 1034: 43–60.
- Lopes-Andrade C, Matushkina N, Buder G, Klass K-D (2009) *Cis pickeri* sp. n. (Coleoptera: Ciidae) from Southern Africa. Zootaxa 2117: 56–64.
- Morrone JJ (2002) Biogeographical regions under track and cladistic scrutiny. Journal of Biogeography 29: 149–152. doi: 10.1046/j.1365-2699.2002.00662.x
- Pic M (1916) Diagnoses génériques et spécifiques. Mélanges Exotico-Entomologiques, Moulins 18: 1–20.
- Pic M (1922) Nouveautés diverses. Mélanges Exotico-Entomologiques, Moulins 36: 1–32.
- Scott H (1926) Coleoptera, Ciidae. Reports of the Percy Sladen Trust Expedition to the Indian Ocean in 1905. Vol. 8, No. 1. Transactions of the Linnean Society of London, Series 2, Zoology 19 (1): 1–41.

RESEARCH ARTICLE



Rainforest understory beetles of the Neotropics, Mizotrechus Bates 1872, a generic synopsis with descriptions of new species from Central America and northern South America (Coleoptera, Carabidae, Perigonini)

Terry L. Erwin[†]

Hyper-diversity Group, Department of Entomology, MRC-187, National Museum of Natural History, Smithsonian, Institution, Washington, P.O. Box 37012, DC 20013-7012, USA

turn:lsid:zoobank.org:author:A7923396-E5AB-4C31-842A-12B8D8C0F315

Corresponding author: Terry L. Erwin (erwint@si.edu)

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Abstract

Information on the single previously described species, *Mizotrechus novemstriatus* Bates 1872 (type locality: Brazil – Amazonas, Tefé), is updated and 17 new species for the genus from Nicaragua, Costa Rica, Panamá, Colombia, Venezuela, and Guyane are described. The species records in the literature and on determined specimens in some collections of *M. novemstriatus* Bates from Central America are not that species; currently, *M. novemstriatus* is known only from its type locality in Amazonian Brazil. For the new species described, their known general distributions are as follows: *Mizotrechus batesi* **sp. n.** (Guyane), *Mizotrechus bellorum* **sp. n.** (Guyane), *Mizotrechus brulei* **sp. n.** (Guyane), *Mizotrechus belevedere* **sp. n.** (Guyane), *Mizotrechus costaricensis* **sp. n.** (Costa Rica), *Mizotrechus dalensi* **sp. n.** (Guyane), *Mizotrechus gorgona.* **sp. n.** (Colombia), *Mizotrechus grossus* **sp. n.** (Guyane), *Mizotrechus jefe* **sp. n.** (Panamá), *Mizotrechus gorgona.* **sp. n.** (Guyane), *Mizotrechus minutus* **sp. n.** (Guyane), *Mizotrechus neblinensis* **sp. n.** (Guyane), *Mizotrechus poirieri* **sp. n.** (Guyane), and *Mizotrechus voldai* **sp. n.** (Panamá). Long-term use of flight intercept traps in Guyane provided so many new species that apparently the use of FITs is the way to collect adults of this taxon, previously known from very few specimens. Many more species of this genus can be expected to be discovered throughout the

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Neotropics; the present contribution is a preliminary synopsis with identification key and adult images of all known species. Likely numerous species are yet to be discovered throughout tropical climes.

Keywords

Flight Intercept Traps (FITs), light traps, Neotropics, Nicaragua, Costa Rica, Panamá, Colombia, Venezuela, Guyane, Brazil

Resumen

Se actualiza la información sobre la única especie descrita anteriormente, Mizotrechus novemstriatus Bates 1872 (localidad tipo: Brasil - Amazonas, Tefé) y se describen 17 nuevas especies dentro del género provenientes de Nicaragua, Costa Rica, Panamá, Colombia, Venezuela y Guayana Francesa. Los registros para la especie en la literatura y en especímenes identificados en algunas colecciones de M. novemstriatus Bates de Centroamérica no son esa especie; actualmente, M. novemstriatus se conoce solamente de su localidad tipo en el área amazónica de Brasil. Para las nuevas especies que se describen, sus distribuciones conocidas son las siguientes: Mizotrechus batesi sp. n. (Guayana Francesa), Mizotrechus bellorum sp. n. (Guayana Francesa), Mizotrechus brulei sp. n. (Guayana Francesa), Mizotrechus belvedere sp. n. (Guayana Francesa), Mizotrechus costaricensis sp. n. (Costa Rica), Mizotrechus dalensi sp. n. (Guayana Francesa), Mizotrechus edithpiafae sp. n. (procedencia desconocida), Mizotrechus fortunensis sp. n. (Panamá), Mizotrechus gorgona. sp. n. (Colombia), Mizotrechus grossus sp. n. (Guayana Francesa), Mizotrechus jefe sp. n. (Panamá), Mizotrechus marielaforetae sp. n. (Guayana Francesa), Mizotrechus minutus sp. n. (Guayana Francesa), Mizotrechus neblinensis sp. n. (Guayana Francesa, Venezuela), Mizotrechus poirieri sp. n. (Guayana Francesa), y Mizotrechus woldai sp. n. (Panamá). El uso prolongado de trampas de intersección de vuelo en Guayana Francesa proveyó tantas nuevas especies que aparentemente el uso de estas trampas es la única manera para colectar adultos de este taxón, previamente conocido de muy pocos especímenes. Muchas más especies de este género se esperan descubrir a través de los Neotrópicos; esta contribución es una sinopsis preliminar con clave de identificación e imágenes de los adultos de todas las especies conocidas. Probablemente numerosas más especies están por encontrarse en los trópicos.

Palabras clave

Trampas de intersección de vuelo, trampas de luz, Neotrópicos, Nicaragua, Costa Rica, Panamá, Colombia, Venezuela, Guayana Francesa, Brasil

Introduction

Bates (1872) proposed the genus *Mizotrechus* for several species including, *Mizotrechus novemstriatus* Bates 1872. The other species were later transferred by him (Bates, 1883) to *Perigona* Laporte de Castelnau 1835. Thus, *Mizotrechus* has been thought to be monobasic since Bates' action in 1883. Bates (1883) also regarded a specimen from Chontales, Nicaragua collected by Janson to be the same species he described from Ega (now Tefé), Brazil. Careful study reported herein of male genitalia and body proportions indicates Bates' determination to be incorrect. Now that many new species have emerged from Guyane (French Guiana and "Cayenne" of earlier authors) and Central American collections and I have been able to study Bates' Ega type, I regard all Central American specimens identified as *Mizotrechus novemstriatus* Bates to be misidentifications. Given the uniformity of shape and color of species in this genus (Figs 1–18) and the previous paucity of specimens to study, it is understandable that mistakes could have been made. With the incredible new collections from Guyane, some species in ample series, it is time to set this genus on the road to a better understanding; that is the purpose of this paper which is predictably just the beginning.

Methods and specimens

Methods and species concepts follow those previously described (Erwin and Kavanaugh 1981; Kavanaugh and Erwin 1991). The species validation and diagnosis format follows as closely as possible that suggested in Erwin and Johnson (2000). Measurements of length (ABL, SBL) and width (TW) follow those of Ball (1972) and Kavanaugh (1979): ABL (apparent body length), measured from apex of labrum to apex of longer elytron (in adults of this genus, the abdomen often protrudes beyond the elytral apex, thus the ABL often is much larger that the SBL; SBL (standardized body length), equals the sum of the lengths of the head (measured from apex of clypeus to a point on midline at level of the posterior edge of compound eyes); PL (pronotum length) is measured from apical to basal margin along midline; LE (elytron length) is measured from apex of scutellum to apex of the longer elytron; and TW (total width) measured across both elytra at their widest point with suture closed. Attributes of the elytron include the basic elytral structure occurring between the elytral intervals that frequently has been expressed as a "stria," or "row of punctures," etc. I refer to this structure as an interneur (Erwin 1974). Attributes of the abdominal ventral sterna are referred to using the numbering system generally accepted in Carabid studies, i.e., the sternum divided medially by the hind coxae is sternum II (the first being hidden) and the last visible is sternum VII (Liu et al., 2011). In describing attributes of the male genitalia, the proximal end of the median lobe has traditionally been referred to as the "basal bulb" in studies of Carabidae. However, in many more basal groups, the proximal end is not a scleritized bulb as it is in most higher groups such as the Lebiini. In my revision of the genus Pericompsus (Erwin, 1974), I encountered the same problem with the term "stria" for features of the elytra. The result was the use of the term "interneur" to apply to the attribute lying between intervals. Through use of this term, one could describe the feature as interneur striate, punctate, striatiopunctate, etc. The same holds true for the proximal end of the median lobe. In Snodgrass (1935), the term "phallobase" is used, and I have adopted it here. So, by extension, in Carabidae we can say phallobase hooded (Lebiini), phallobase of two parallel sclerotized struts (basal trechines and Andinodontis), phallobase of two uneven struts (Bembidion), etc. Kavanaugh (pers. comm.) points out that with struts there is still a connecting membrane surrounding the struts forming a "bulb."

Included in this study are a total of 56 specimens: ten from the National Museum of Natural History, Washington, DC (NMNH) in my charge, 41 specimens received from SEAG (Société Entomologique Antilles-Guyane) in Guyane, a single specimen from INBIO collected in Costa Rica and sent to me by Angel Solis, and a single specimen from the University of Alberta, Edmonton, Canada (UASM) collected in Colom-

bia and sent to me by George E. Ball. Also studied were the lectotype of *Mizotrechus novemstriatus* Bates from the Muséum National d'Histoire Naturelle, Paris (MHNP, Azadeh Taghavian, Collection Manager) along with an unidentified specimen determined by Bates simply as "Mizotrechus/G^{er}Bates," and the specimen from Nicaragua that Bates identified incorrectly came from the Natural History Museum in London (BMNH, Beulah Garner, Collection Manager).

The habitus images of the adult beetles portray most of the character states referred to in the key provided. Illustrations of male genitalia are standard for descriptive taxonomy of carabid beetles in both preparation and aspects presented, as is the presentation of the female genitalia. The habitus images of the adults were made with a Visionary DigitalTM high resolution imaging system rendered using Photoshop to become "Digital Photo-illustrations." Figure captions include an ADP number, which is a unique identification number for the specimen that was illustrated or imaged and links the specimen and associated illustrations and/or images to additional information in electronic databases at the NMNH. Representative specimens will be transferred to Guyane when they have a museum in which to keep them secure.

Geographical data are presented for species based on all known specimens available at the time of manuscript preparation, including those in the literature. Georeferences have been determined from locality information provided on specimen labels; only those exact georeferences reported in decimal degrees that are provided on the label are placed in quotes, otherwise I have estimated these as closely as possible from places, mileage, etc., listed on the label and searched with Google Earth. Latitude and longitude are reported in decimal degrees. Distribution maps are provided for the species (Figs 39–41). Here, vernacular names in English are proposed, as common names are becoming increasingly needed in conservation and/or agricultural and forestry applications. These names are based on criteria set forth in Erwin (2011b) and applied in Erwin (2011a).

Taxonomy

Mizotrechus Bates 1872 Trough beetles http://species-id.net/wiki/Mizotrechus

Mizotrechus Bates, 1872:199

Type species. Mizotrechus novemstriatus Bates, 1872:199

Number of species. Previously one, now 18.

Taxonomy. Stable herein as of this publication, yet many new species expected in Central and South America, particularly south of the main Amazon channel. Adelphotaxon: probably *Perigona* Laporte de Castelnau 1835.

Proposed English vernacular name. Trough beetles. An attribute of adults of the Tribe Perigonini is the peculiar setiferous trough-like margin at the latero-apical por-

tion of the elytron. While Bates' use of "*mizo*" as part of the genus name is unclear, I found that the French translation of the Japanese word, *mizo*, is ditch or pit in English. Perhaps Bates (who published much on Japanese beetles) was referring to the long marginal setiferous "trough" near the apex of the elytron.

Distribution. Presently known from Central and (northern) South America and here predicted to be found throughout the Amazon Basin and possibly north into México.

Habitat. Tropical rainforest, dry forest, and cloud forest possibly in leaf litter, or more likely subcortical (under loose bark). I suggest that these beetles are subcortical based on their coloration, robust exoskeleton, markedly expanded distal antennomeres, prolonged sharp mandibles, serrate or undulating humerus in some species, and somewhat depressed form. Some of these attributes are found in the adults of genera *Pachyteles, Holmalomorpha, Catapiesis, Inpa*, and *Morion*, among others, whose adult members are known to be subcortical inhabitants for part of their life history.

References. Bates (1872, 1883)

Diagnostic combination. Differing in adult attributes from those of its probable adelphotaxon, Perigona Laporte de Castelnau 1835 and from Diploharpus Chaudoir 1850 by the following: 1) adult *Mizotrechus* possess markedly expanded distal antennomeres (neither adults of Perigona nor Diploharpus adults have such); 2) both male and female Mizotrechus adults have the foreleg femur modified in some manner (except *M. batesi*, sp. n.) such as dentate, serrate, swollen or ridged (lacking in adults of both Perigona and Diploharpus); 3) adult Mizotrechus lack discal setae of the elytra (present in *Perigona* at apex of interneur 5 and in *Diploharpus* at middle and apical third in interneur 3); 4) adult Mizotrechus have umbilicate setae at apical third off-set and surrounded by a dense patch of short setae (both Perigona and Diploharpus have the umbilicate setae in-line and with few if any short setae); 5) adult *Mizotrechus* have a dense patch of setae in the apical trough of the elytron (neither Perigona nor Diploharpus have such a patch, although some species have a few setae); 6) sternum III of both sexes of Mizotrechus species have a short row of reclinate setae medially set between two narrowly spaced ambulatory setae (neither adults of Perigona nor Diploharpus nor a new genus (Erwin, in prep) have such).

Distribution. Lowland tropical rainforests in Central and northern South America, as well as midland cloud and dry forests in Central America.

Included species. The species list below, as well as arrangement of descriptions that follow is ordered alphabetically.

<i>Mizotrechus batesi</i> sp. n.	Guyane
<i>Mizotrechus bellorum</i> sp. n.	Guyane
<i>Mizotrechus belvedere</i> sp. n.	Guyane
Mizotrechus brulei sp. n.	Guyane
Mizotrechus chontalesensis sp. n.	Nicaragua
Mizotrechus costaricensis sp. n.	Costa Rica
Mizotrechus dalensi sp. n.	Guyane

Mizotrechus edithpiafae sp. n.	?
<i>Mizotrechus fortunensis</i> sp. n.	Panamá
Mizotrechus gorgona. sp. n.	Colombia
Mizotrechus grossus sp. n.	Guyane
<i>Mizotrechus jefe</i> sp. n.	Panamá
<i>Mizotrechus marielaforetae</i> sp. n.	Guyane
Mizotrechus minutus sp. n.	Guyane
<i>Mizotrechus neblinensis</i> sp. n.	Guyane, Venezuela
Mizotrechus novemstriatus Bates 1872	Brazil
<i>Mizotrechus poirieri</i> sp. n.	Guyane
<i>Mizotrechus woldai</i> sp. n.	Panamá

Key to the species of *Mizotrechus* Bates 1872

1	Elytron with striatopunctate interneurs, punctures small, evident, regularly
	spaced. Foreleg femur (Fig. 19) unmodified posteriorly. Size very small (ABL
	= 4.4–4.6 mm) <i>Mizotrechus minutus</i> sp. n.
1'	Elytron with smooth or uneven interneurs, punctures if evident not regularly
	spaced. Foreleg femur modified posteriorly, either slightly swollen, ridged,
	dentate, or serrate. Size small to large (ABL = 5.9–8.9 mm)2
2(1')	Form narrow, parallel-sided (Fig. 1), pronotum nearly square; size small (ABL
	= 5.9 mm, narrow body) <i>Mizotrechus batesi</i> sp. n.
2'	Form broad, pronotum sub-cordiform; size larger (ABL = 5.9 (broad body) –
	8.9 mm)
3(2')	Elytral margin behind humerus evidently serrate, the narrowly explanate mar-
	gin dentate with minute seta at point of each tooth4
3'	Elytral margin behind humerus not evidently serrate, smooth or rough, if
	rough explanate margin interrupted by minute setae and interval between
	setae blunt
4(3)	Elytron with all interneurs deep and evenly impressed throughout. Foreleg
	femur (Fig. 23) slightly swollen along base of postero-ventral margin in form
	of long ridge <i>Mizotrechus brulei</i> sp. n.
4'	Elytron with discal interneurs moderately deep, lateral interneurs much less
	impressed. Foreleg femur (as in Fig. 24) serrate along basal third to half of
	postero-ventral margin5
5(4')	Size larger (ABL = 8.4 mm)
5'	Size smaller (ABL = 6.6–7.4 mm)7
6(5)	Pronotum markedly convex; lateral margin straight before hind angle, a small
	acute tooth behind posterior lateral seta (ABL = 8.4 mm)
6'	Pronotum barely convex; lateral margin sinuate before right hind angle, (ABL
	= 8.4 mm) <i>Mizotrechus costaricensis</i> sp. n.
	-

7(5')	Pronotum barely convex, narrow; hind angle about right (ABL = 6.6 mm)
	<i>Mizotrechus jefe</i> sp. n.
7'	Pronotum markedly convex, broad; hind angle a small tooth (ABL = 7.4
	mm) Mizotrechus woldai sp. n.
8(3')	Elytron with all interneurs deep and evenly impressed throughout9
8'	Elytron with discal interneurs moderately deep, lateral interneurs much less
	impressed12
9(8)	Size large (ABL = 8.4 mm). Pronotum markedly convex, large with markedly
	arcuate side margins. Foreleg femur (Fig. 22) dentate on postero-ventral mar-
	gin Mizotrechus edithpiafae sp. n.
9'	Size smaller (ABL 6.5–7.1 mm). Pronotum barely convex, longer than wide,
	nearly quadrate. Foreleg femur (as in Fig. 21) with a short ridged on postero-
	ventral margin10
10(9')	Pronotum with hind angle sharp, about a right angle, not produced as a small
	denticle Mizotrechus chontalesensis sp. n.
10'	Pronotum with hind angle sharp, produced as a small denticle11
11(10')	Pronotum nearly quadrate (W/L = 1.356)
	Mizotrechus novemstriatus Bates
11'	Pronotum much wider than long ($W/L = 1.996-2.228$)
12(8')	Pronotum with posterior lateral margin moderately lobed just anterior to
	dentiform hind angle
12'	Pronotum with posterior lateral margin straight just anterior to dentiform
	hind angle or right hind angle13
13(12')	Size large (ABL 7.6–9.4 mm)14
13'	Size smaller (ABL 6.0–6.8 mm)16
14(13)	Form elongate and narrow. Elytron with margin behind humerus rough, un-
	dulating between microsetae. Foreleg femur (as in Fig. 24) serrate on postero-
	ventral margin
14'	Form broader and shorter. Elytron with margin behind humerus smooth, not
	undulating between microsetae. Foreleg femur (Fig. 20) dentate on postero-
	ventral margin15
15(14')	Elytra very convex, at least interneurs 1-3 deep and complete. Pronotum
	with margin anterior to hind angle slightly arcuate Mizotrechus dalensi n.
	sp
15'	Elytra rather depressed, only interneur 1 deep and complete. Pronotum with
	margin anterior to hind angle straight Mizotrechus grossus sp. n.
16(13')	Pronotum with margin anterior to hind angle slightly arcuate
16'	Pronotum with margin anterior to hind angle straight17
17(16')	Form elongate and narrow. Elytron with at least 4 interneurs deep and com-
	plete, 2-4 about equal to 1 in depth. Foreleg femur (as in Fig. 21) ridged on
	postero-ventral margin

Accounts of taxa

Mizotrechus batesi sp. n. Bates' trough beetle urn:lsid:zoobank.org:act:897D9EB5-D50F-4021-9818-A79D5C86175A http://species-id.net/wiki/Mizotrechus_batesi Figs 1, 40

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 30 November 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124884, male).

Derivation of specific epithet. The epithet "*batesi*" is an eponym, based on the family name of Henry Walter Bates, whose eleven years of collecting beetles in South America capturing adults of many new species, and genera such as this, would qualify him for the Hall of Fame of beetle collectors and describers, if there was such a Hall.

Proposed English vernacular name. Bates' trough beetle.

Diagnosis. With the attributes of the genus as described above and small sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose and punctulate. Occiput punctulate. Pronotum quadrate with lateral margin shallowly emarginate just anterior to hind angle; base densely punctulate. Elytra narrow and elongate, about the width of pronotum across anterior third, and with 8 well-impressed interneurs, intervals flat; margin behind humerus shallowly serrulate. Foreleg femur with slightly produced ridge on postero-ventral margin.

Description. (Fig. 1). *Size*: See Appendix 1. Small for genus, ABL = 5.9 mm, SBL = 4.9 mm, TW = 1.8 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum and legs shiny. *Head*: Labrum quadrate and medially notched apically. Eye small, moderately convex. Gena long, straight. Frons, occiput and gena glabrous. *Prothorax*: Broad, narrowed slightly toward base, margin beaded, not explanate except at hind angle; surface punctulate, punctures widespread, glabrous. *Pterothorax*: Elytron moderately convex, intervals slightly costate, interneurs striate, not punctate, apex slightly oblique and slightly rounded, sutural apex narrowly truncate. Metasternum sparsely setiferous in male. *Legs*: Normal in male; foreleg femur (as in Fig. 21) with slightly produced long ridge on postero-ventral margin and with a very short secondary ridge basally and above end of long ridge, not dentate; posterior trochanter tapered to point, length nearly half that of femur. *Abdomen*: Abdominal sterna moderately setiferous; sternum VII medially notched in male. *Male genitalia*: Aedeagus and parameres missing from sin-



Plate 1. Digital Photo-illustrations, habitus, dorsal aspect: **I** *Mizotrechus batesi* sp. n., ABL = 5.9 mm, male holotype, ADP124884; type locality. **2** *Mizotrechus bellorum* sp. n., ABL = 6.9 mm, male holotype, ADP124890; type locality. **3** *Mizotrechus belvedere* sp. n., ABL = 7.2 mm, male holotype, ADP129201; type locality. **4** *Mizotrechus brulei* sp. n., ABL = 6.7 mm, female paratype, ADP124926; type locality.

gle known male holotype, although the ring sclerite is present. *Female genitalia*: Un-known.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. The adult holotype was active in November, at the end of the dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 40). This species is currently known only from the type locality in the lowlands of Guyane.

Mizotrechus bellorum sp. n.

Bells' trough beetle urn:lsid:zoobank.org:act:E336EDFB-B152-4B87-8C8E-1AE5A4B8B377 http://species-id.net/wiki/Mizotrechus_bellorum Figs 2, 25, 40

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 13 September 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124890, male)

Derivation of specific epithet. The epithet "*bellorum*" is an eponym, based on the family name of Ross and Joyce Bell, with a special thanks for a career that brought light to the rhysodine clade of very interesting beetles with colors and tough cuticle also found in adults of *Mizotrechus* species.

Proposed English vernacular name. Bells' trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons at sides with evident rugae, punctulate. Occiput at sides with evident rugae, punctulate. Pronotum nearly quadrate, narrowed behind, with straight margins to hind angle, hind angle dentate, tooth small; base sparsely rugulose. Elytra broad and short, much wider than the width of pronotum across anterior third, and with only interneurs 1–3 moderately engraved, 4–8 evident yet shallowly impressed, more so toward margin, not punctate; margin behind humerus uneven, yet not serrate. Foreleg femur subdentate at the base of postero-ventral margin.

Description. (Figs 2, 25). *Size*: See Appendix 1. Medium sized for the genus, ABL = 6.9 mm, SBL = 5.87 mm, TW = 2.55 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny, elytra slightly duller due to shallowly engraved slightly stretched microsculpture. *Head*: Labrum quadrate and apico-medially emarginate. Eye large, moderately convex. Gena moderately short, straight. Frons, occiput and gena glabrous. *Prothorax*: Narrow, quadrate, narrowed slightly toward base, margins not emarginate before hind angle, angle dentate, tooth small, margin moder-

ately explanate except wider at hind angle; surface punctulate, punctures widespread, glabrous. *Pterothorax*: Elytron moderately convex, disk flat, intervals flat, interneurs not punctate, apex moderately oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. *Legs*: Normal in male; foreleg femur (as in Fig. 20) with a small obtuse tooth at basal third of postero-ventral margin; posterior trochanter narrowly acute at apex, about half the length of the femur. *Abdomen*: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae. *Male genitalia*: Median lobe (Fig. 25) elongate and robust with ostium moderately elongate, over half the length of the median lobe; apex a losp. n.tulate distal end less bent ventrad than in *M. dalensi* and thicker in cross section, moderately curved in lateral aspect, ventral margin proximal to apex straight then evenly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded, not crested, opening more or less 30 degrees off axis with that of shaft. Parameres large, left a half longer than the right, both broadly rounded, asetose. *Female genitalia*: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. Adults are active in September, near the end of the dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 40). This species is currently known only from the type locality in the lowlands of Guyane.

Mizotrechus belvedere sp. n.

Brule's trough beetle

urn:lsid:zoobank.org:act:8239C95E-35C9-450D-93A0-6FF8B48ED39F http://species-id.net/wiki/Mizotrechus_belvedere Figs 3, 26, 40

Holotype. GUYANE, Saül, Commune de Saül, Belvédère de Saül, 283–325 m, 3.6223°N, 53.2159°W, 17 February 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP129201, male).

Derivation of specific epithet. The epithet "*belvedere*" is a singular Latinized masculine noun in apposition, based on the name of the area in which these beetles are found.

Proposed English vernacular name. Belvédère trough beetle.

Diagnosis. With the attributes of the genus as described above and moderately large sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose above and behind eye in an arc and moderately punctulate. Occiput moderately punctulate. Pronotum longer than wide with lateral margins moderately explanate and basally notched; base moderately microrugose. Elytra moderately broad and short, apex prolonged, wider than the width of pronotum

across anterior third, and with 5 well-impressed irregularly punctulate interneurs, intervals not convex; margin behind humerus rough, intervals between microsetae blunt. Foreleg femur with slightly produced postero-ventral margin.

Description. (Figs 3, 26). Size: See Appendix 1. Moderately small for genus, ABL = 7.2 mm, SBL = 6.14 mm, TW = 2.48 mm. Color: see diagnosis, above. Luster: Head, pronotum and legs shiny, elytra matte. *Head*: Labrum quadrate, apico-medially slightly V-notched. Eye moderately large and convex. Gena with very slight bulge. Frons, occiput and gena glabrous. Prothorax: Moderately narrow, narrowed slightly toward base, margin moderately explanate, wider before hind angle; surface punctulate, punctures moderately dense, glabrous. Pterothorax: Elytron barely convex, intervals nearly flat, interneurs with well-impressed irregularly-spaced punctulate, apex slightly obliquely prolonged and slightly rounded at extreme sutural apex. Metasternum sparsely setiferous in male. Legs: Normal in male; foreleg femur (as in Fig. 23) with slightly produced ridge along postero-ventral margin, not dentate, without short dorsal ridge at base of longer ridge; posterior trochanter tapered to rounded point in male, length half that of femur. Abdomen: Abdominal sterna moderately setiferous, densely so medially on II and III; sternum III of male with short dense patch of decumbent setae medially set between two ambulatory setae; sternum VII shallowly and medially notched in male. Male genitalia: Median lobe (Fig. 26) short and robust with ostium moderately elongate, over half the length of the median lobe; apex a short blunt distal end that is less prominent than in *M. brulei*, slightly curved in lateral aspect, ventral margin proximal to apex evenly curved; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less 20 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. Female genitalia: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. The holotype was active in February, the rainy season.

Other specimens examined. None.

Geographic distribution. (Fig. 40). This species is currently known only from the type locality in the lowlands of Guyane.

Mizotrechus brulei sp. n.

Brule's trough beetle

urn:lsid:zoobank.org:act:59CE9DC7-C3BF-4DAE-9CE1-D72FF093AAA6 http://species-id.net/wiki/Mizotrechus_brulei Figs 4, 27, 40

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 13 September 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124886, male).

Derivation of specific epithet. The epithet "*brulei*" is an eponym, based on the family name of Stephané Brule, whose team in Guyane has been collecting beetles using Flight Intercept Traps and capturing adults of many new species, such as this one.

Proposed English vernacular name. Brule's trough beetle.

Diagnosis. With the attributes of the genus as described above and small sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose only at extreme anterior angles and punctulate. Occiput punctulate. Pronotum quadrate with lateral margins basally shallowly emarginate; base moderately microrugose. Elytra moderately narrow and elongate, wider than the width of pronotum across anterior third, and with 8 well-impressed interneurs, intervals not convex; margin behind humerus moderately serrulate. Foreleg femur with slight swelling near the base on the postero-ventral margin.

Description. (Figs 4, 27). Size: See Appendix 1. Moderately small for genus, ABL = 6.0-6.8 mm, SBL = 5.53-6.41 mm, TW = 2.03-2.22 mm. Color: see diagnosis, above. Luster: Head, pronotum and legs shiny. Head: Labrum quadrate, apico-medially barely emarginate. Eye small, moderately convex. Gena long, straight. Frons, occiput, and gena glabrous. Prothorax: Broad, narrowed slightly toward base, margin beaded, not explanate except at hind angle; surface punctulate, punctures widespread, glabrous. Pterothorax: Elytron barely convex, intervals nearly flat, interneurs striate, not punctate, apex slightly oblique and slightly rounded, sutural apex narrowly truncate. Metasternum sparsely setiferous in male. *Legs*: Normal in both sexes; foreleg femur (Fig. 23) with slightly produced ridge along postero-ventral margin, not dentate, without short dorsal ridge at base of longer ridge; posterior trochanter tapered to acute point in male, acuminate in female, length half that of femur. Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae; sternum VII medially notched in male, slightly emarginated in female. Male genitalia: Median lobe (Fig. 27) short and robust with ostium moderately elongate, over half the length of the median lobe; apex short, blunt, distal end more prominent than in *M. belvedere*, slightly curved in lateral aspect, ventral margin proximal to apex evenly curved; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less in line with axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. Female genitalia: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult specimens were collected in flight intercept traps in the rainforest understory. Adults are active in April and September, in both the rainy and dry seasons.

Other specimens examined. Paratypes: GUYANE, Montagne des Chevaux, Commune de Roura, RN2 PK22, 90 m, 4.7127°N, 52.3966°W, 14 April 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124960, female), 9 January 2011 (NMNH: ADP124924, female), 6 March 2010 (NMNH: ADP127159, male), 8 August 2010 (NMNH: ADP129205, male), 22 August 2010 (NMNH: ADP128729, male, ADP128727, female), 3 October 2010 (NMNH: ADP128731, male), Saül, Commune de Saül, Belvédère de Saül, 283–325 m, 3.6223° N, 53.2159° W, 29 October 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124926, female).

Geographic distribution. (Fig. 40). This species is currently known only from three lowland localities in Guyane.

Mizotrechus chontalesensis sp. n. Chontales trough beetle

urn:lsid:zoobank.org:act:638541F2-014B-45D7-8A7E-C0D80817DEA5 http://species-id.net/wiki/Mizotrechus_chontalesensis Figs 5, 21, 39

Holotype. NICARAGUA, Chontales, ? nr. Santo Domingo, 514 m, 12.262°N, 85.083°W, (EM Janson)(BMNH: ADP127181, female).

Derivation of specific epithet. The epithet "*chontalesensis*" is a Latinized noun in apposition, based on the name of the area in which these beetles are found.

Proposed English vernacular name. Chontales trough beetle.

Diagnosis. With the attributes of the genus as described above and moderately large sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons and occiput quite smooth, without regular rugae; micropunctation widespread and hardly evident. Pronotum longer than wide with lateral margins narrowly explanate and basally shallowly emarginate with sharp, not denticulate, hind angles; base smooth, without regular microrugosity. Elytra moderately broad and short, apices not prolonged, wider than the width of pronotum across anterior third, and each with 8 well-impressed irregularly punctulate interneurs, intervals not convex; margins behind humeri rough, intervals between microsetae blunt. Foreleg femur with slightly produced ridge on postero-ventral margin.

Description. (Figs 5, 21). *Size*: See Appendix 1. Moderately sized for genus, ABL = 7.1 mm, SBL = 5.87 mm, TW = 2.29 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, elytra, and legs shiny. *Head*: Labrum quadrate, apico-medially barely emarginate. Eye small and barely convex. Gena straight. Frons, occiput, and gena glabrous. *Prothorax*: Moderately narrow, narrowed slightly toward base, margin narrowly explanate, slightly wider before hind angle; surface punctulate, punctures widespread, very fine, glabrous. *Pterothorax*: Elytron barely convex, intervals nearly flat, all interneurs with well-impressed irregularly-spaced punctures, apex not prolonged, slightly rounded at extreme sutural apex. *Legs*: Normal in female; foreleg femur (Fig. 21) with slightly produced, short, arcuate ridge on postero-ventral margin at basal forth, not dentate; posterior trochanter tapered to rounded point in female, length half that of femur. *Abdomen*: Abdominal sterna moderately setiferous, densely so medially on II and III;



Plate 2. Digital Photo-illustrations, habitus, dorsal aspect: **5** *Mizotrechus chontalesensis* sp. n., ABL = 7.1 mm, female holotype, ADP127181; type locality. **6** *Mizotrechus costaricensis* sp. n., ABL = 8.4 mm, male holotype, ADP128620; type locality. **7** *Mizotrechus dalensi* sp. n., ABL = 8.2 mm, male holotype, ADP124894; type locality. **8** *Mizotrechus edithpiafae* sp. n., ABL = 8.4 mm, female holotype, ADP124948; locality unknown.

sternum VII barely medially emarginate in female. *Male genitalia*: Unknown. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. Unknown.

Other specimens examined. None.

Geographic distribution. (Fig. 39). This species is currently known only from the type locality in the dry tropical forested lowlands of Nicaragua.

Mizotrechus costaricensis sp. n.

Costa rica trough beetle

urn:lsid:zoobank.org:act:FE064F4E-01E2-456D-8B59-6374BDECCF4F http://species-id.net/wiki/Mizotrechus_costaricensis Figs 6, 28, 39

Holotype. COSTA RICA, Alajuela, San Ramón, Reserva Biologica Alberto Brenes, Rio San Lorenzo, 850 m, 10.2283°N, 84.5857°W, 30 June – 5 July 1999 (J Rodriguez) (INBIO: ADP128620, INB0003057334, male).

Derivation of specific epithet. The epithet "*costaricensis*" is a Latinized noun in apposition, based on a geographic name formed from "Costa Rica," the country in which an adult of this species has been found, and the Latin suffix "*-ensis*," meaning of, or from that place.

Proposed English vernacular name. Costa Rica trough beetle.

Diagnosis. With the attributes of the genus as described above and large sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons and occiput shallowly rugose above and behind eye in an arc asp. n.rsely and finely punctulate. Pronotum subcordiform with lateral margins moderately explanate and basally emarginate before right hind angles; base shallowly and regularly microrugose. Elytra moderately narrow and short, apex prolonged, wider than the width of pronotum across anterior third, and with 5 well-impressed irregularly punctulate interneurs, intervals not convex; margin behind humerus rough, intervals between microsetae blunt. Foreleg femur with markedly serrate postero-ventral margin.

Description. (Figs 6, 28). *Size*: See Appendix 1. Large for genus, ABL = 8.4 mm, SBL = 6.88 mm, TW = 2.88 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum and legs shiny, elytra matte. *Head*: Labrum quadrate, apico-medially slightly produced. Eye moderately small and convex. Gena straight. Frons, occiput, and gena glabrous. *Prothorax*: Moderately narrow, narrowed slightly toward base, margin narrowly explanate, wider before hind angle; surface sparsely punctulate, punctures very fine, glabrous. *Pterothorax*: Elytron barely convex, intervals nearly flat, 5 interneurs with well-impressed irregularly spaced punctures, apex not prolonged, slightly rounded at extreme sutural apex. Metasternum sparsely setiferous in male. *Legs*: Normal in

male; foreleg femur (as in Fig. 24) with markedly serrate postero-ventral margin, not dentate; posterior trochanter tapered to acute point in male, length half that of femur. *Abdomen*: Abdominal sterna moderately setiferous, densely so medially on II and III; sternum III of male with short row of decumbent setae medially set between two ambulatory setae; sternum VII shallowly and medially notched in male. *Male genitalia*: Median lobe (Fig. 28) short and robust with ostium moderately elongate, over half the length of the median lobe; apex with a very short and narrow distal end, slightly rounded in lateral aspect, more twisted laterally than in *M. fortunensis*, ventral margin proximal to apex straight then briefly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less 30 degrees off axis of shaft. Parameres large, left a twice longer than the right, both broadly rounded, asetose. *Female genitalia*: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The holotype was collected in an aerial net. Adults are active in early July, the area's dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 39). This species is currently known only from the type locality in premontane rainforests of Costa Rica.

Mizotrechus dalensi sp. n.

Dalens' trough beetle

urn:lsid:zoobank.org:act:1FAB3B90-4F18-4690-825D-B13F8510E82E http://species-id.net/wiki/Mizotrechus_dalensi Figs 7, 20, 29, 40, Plate 11

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 30 November 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124894, male).

Derivation of specific epithet. The epithet "*dalensi*" is an eponym, based on the family name of P.H. Dalens, whose team in Guyane has been collecting beetles using Flight Intercept Traps and capturing adults of many new species, such as this one.

Proposed English vernacular name. Dalens' trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose and punctulate. Occiput shallowly rugose at sides and punctulate. Pronotum quadrate with lateral margins basally straight to slightly toothed hind angle; base moderately rugose. Elytra broad and moderately short, much wider than width of pronotum across anterior third, and with two well- impressed interneurs, the remainder shallower toward margin; margins behind humeri not serrulate. Foreleg femur dentate basally on postero-ventral margin.

Description. (Figs 7, 20, 29, Plate 11). Size: See Appendix 1. Medium-sized for genus, ABL = 7.6-8.2 mm, SBL = 6.40-7.09 mm, TW = 2.64-3.11 mm. Color: see diagnosis, above. Luster: Head, pronotum and legs shiny; elytra dull due to marked slightly stretched well-impressed microsculpture. Head: Labrum quadrate, apico-medially barely emarginate. Eye moderately large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. Prothorax: Broad, narrowed slightly toward base, margin beaded, not explanate except at hind angle; surface punctulate, punctures widespread, and with shallow yet evident transverse microsculpture, surface glabrous. Pterothorax: Elytron moderately convex, intervals flat, interneurs striate, not punctate, becoming shallower toward margin, apex slightly oblique, straight, sutural apex narrowly and slightly rounded. Metasternum sparsely setiferous in male. Legs: Normal in both sexes; foreleg femur (Fig. 20) dentate at basal third on postero-ventral margin; posterior trochanter tapered to acute point in male, slightly acuminate in female, length half that of femur. Abdomen: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae; sternum VII medially V-notched in male, U-notched in female. Male genitalia: Median lobe (Fig. 29) elongate and robust with ostium moderately elongate, over half the length of the median lobe; apex a losp. n.tulate distal end less bent ventrad than in *M. poirieri*, moderately curved in lateral aspect, ventral margin proximal to apex straight then evenly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded and crested, opening more or less 40 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. *Female genitalia*: (Plate 11). Ovipositor with broad laterotergite (lt) and two gonocoxites (gc 1, gc 2); gonocoxite 1 apico-laterally setose; gonocoxite 2 falcate, base (b) medium-size, narrow, blade (bl) elongate, with two dorsal ensiform setae (des), and one ventral ensiform seta (ves), all ensiform setae moderately long and robust; with ventral preapical nematiform setae (evident on A, hidden on B). Reproductive tract (Plate 11A) proximally with short, broad bursa copulatrix (bc), continuous at its distal end with common oviduct (co) and long robust spermatheca (sp), latter slightly narrowed distally; spermathecal gland (sg) bulbous; spermathecal gland duct (sgd) long, slender, attached to spermatheca at base of its broadened portion.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adults at four localities were collected in flight intercept traps in the rainforest understory. Adults are active in March and November, in both the rainy and dry seasons.

Other specimens examined. Paratypes: Guyane, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 23 November 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124896, male); Mount Itoupe, 570 m, 3.0148°N, 53.0721°W, 17 March 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP126485, male); Montagne des Chevaux, Commune de Roura, RN2 PK22, 90 m, 4.7127°N, 52.3966°W, 8 August 2010 (NMNH: ADP129203, female), 1 November 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124956, female), Saül, Com-

mune de Saül, Belvédère de Saül, 283–325 m, 3.6223°N, 53.2159°W, 20 December 2010 (NMNH: ADP127167, male), 4 January 2011 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124922, female), 36 km W Regina, Petite Montague Tortue, P17, 94 m, 4.3204°N, 52.2404°W, 23 June 2010 (G Lamarre)(NMNH: ADP127161, male).

Geographic distribution. (Fig. 40). This species is currently known from five localities in the lowland and midland rainforests of Guyane.

Mizotrechus edithpiafae sp. n.

Piaf's trough beetle urn:lsid:zoobank.org:act:6D88FB32-8C21-49DA-BEAD-719E3ABB3AB6 http://species-id.net/wiki/Mizotrechus_edithpiafae Figs 8, 22, 30

Holotype. Locality unknown. Specimen, (BMNH: ADP128624, male), labeled (Bates' handwriting) "Mizotrechus / G^{er} Bates."

Derivation of specific epithet. The epithet "*edithpiafae*" is an eponym, based on the full name of Edith Piaf, the famous French singer, 19 December 1915–11 October 1963, whose voice had an incredible range of diversity, as is that found in the carabid species richness of Guyane, and who sang a variety of "torch songs", and here I play on the word "torch", the same word that applies to what is used to ignite the trees of the unique tropical rainforests of South America, an Armageddon in our own times.

Proposed English vernacular name. Piaf's trough beetle.

Diagnosis. With the attributes of the genus as described above and large sized for the genus as it is presently understood. Adults with darkly infuscate integument, elytra slightly paler, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture black. Frons and occiput smooth above and behind eye asp. n.rsely and finely punctulate. Pronotum quadrate and highly convex with lateral margins narrowly explanate and basally straight before slightly denticulate hind angles; base shallowly and regularly microrugose. Elytra very broad and short, apex not prolonged, wider than the width of pronotum across anterior third, and with 8 well- impressed irregularly punctulate interneurs, intervals not convex; margin behind humerus rough, intervals between microsetae blunt. Foreleg femur with markedly dentate ventral margin.

Description. (Figs 8, 22). *Size*: See Appendix 1. Large for genus, ABL = 8.4 mm, SBL = 7.16 mm, TW = 2.94 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny, elytra slightly matte. *Head*: Labrum quadrate, apico-medially moderately v-notched. Eye moderately small and convex. Gena straight. Frons, occiput, and gena glabrous. *Prothorax*: Broad, barely narrowed toward base, margin narrowly explanate, wider before hind angle; surface sparsely punctulate, punctures very fine, glabrous. *Pterothorax*: Elytron barely convex, intervals nearly flat, 8 interneurs with well-impressed irregularly-spaced punctures, apex not prolonged, slightly rounded at extreme sutural apex. Metasternum sparsely setiferous in male. *Legs*: Normal in male; foreleg femur (Fig. 22) ridged along

the posterio-ventral margin and markedly produced at middle of ridge ; posterior trochanter tapered to rounded point in male, length half that of femur. *Abdomen*: Abdominal sterna moderately setiferous, densely so medially on II and III; sternum III of male with short dense row of decumbent setae medially set between two ambulatory setae; sternum VII shallowly and medially notched in male. *Male genitalia*: Median lobe (Fig. 30) short and robust with ostium moderately elongate, over half the length of the median lobe; apex with a moderately long narrow distal end, slightly curved in lateral aspect, ventral margin proximal to apex evenly curved, ventral margin proximal to apex straight; endophallus (everted) with complexly folded tracheal fields and a narrow sclerotized rod; phallobase not fully hooded, opening 20 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. *Female genitalia*: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. Unknown. Other specimens examined. None. Geographic distribution. Unknown.

Mizotrechus fortunensis sp. n.

Fortuna trough beetle

urn:lsid:zoobank.org:act:CCCF2C24-8587-42A0-83A5-BD953EF41585 http://species-id.net/wiki/Mizotrechus_fortunensis Figs 9, 31, 39

Holotype. PANAMÁ, Chiriquí Province, Fortuna, 1050 m, 8.7341°N, 82.2701°W, 13 August 1978 (H. Wolda)(NMNH: ADP124968, male).

Derivation of specific epithet. The epithet "*fortunensis*" is a Latinized noun in apposition, based on a geographic name formed from "Fortuna" (Panamá) the place in which adults of this species have been found, and the Latin suffix "*-ensis*," meaning of, or from that place.

Proposed English vernacular name. Fortuna trough beetle.

Diagnosis. With the attributes of the genus as described above and large sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons smooth, not or barely rugose near eye, surface punctulate. Occiput smooth, punctulate. Pronotum quadrate with lateral margins basally shallowly emarginated, hind angles slightly toothed; base densely microrugose. Elytra broad and short, much broader than width of pronotum across anterior third, and with 3 well-impressed interneurs, others shallower toward margin; margins behind humeri shallowly serrulate. Foreleg femur serrate on postero-ventral margin.

Description. (Figs 9, 31). *Size*: See Appendix 1. Large-sized for genus, ABL = 8.5– 8.7 mm, SBL = 7.02–7.66 mm, TW = 2.93 –3.03 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny; elytra dull due to marked slightly stretched



Plate 3. Digital Photo-illustrations, habitus, dorsal aspect: **9** *Mizotrechus fortunensis* sp. n., ABL = 8.7 mm, male holotype, ADP124968; type locality. **10** *Mizotrechus gorgona.* sp. n., ABL = 8.4 mm, female Holotype, ADP128622; type locality. **11** *Mizotrechus grossus* sp. n., dorsal aspect, ABL = 8.9 mm, male paratype, ADP127169; type locality. **12** *Mizotrechus jefe* sp. n., ABL = 6.6 mm, male paratype, ADP011173; type locality.

well-impressed microsculpture. Head: Labrum quadrate, entire apically. Eye moderately large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. Prothorax: Quadrate, narrowed slightly toward base, margin beaded, not explanate except at hind angle, hind angle feebly dentate; surface punctulate, punctures widespread, and with microsculpture nearly effaced, surface glabrous. Pterothorax: Elytron moderately convex, disk flat, intervals flat, interneurs striate, not punctate, shallower toward margin, apex slightly oblique, straight, sutural apex narrowly and slightly rounded. Metasternum sparsely setiferous in male. Legs: Normal in both sexes; foreleg femur (as in Fig. 24) serrate on postero-ventral margin; posterior trochanter narrowly rounded in both sexes, length half that of femur. Abdominal sterna moderately setiferous; sternum IV of male without medial patch of decumbent setae; sternum VII medially and shallowly V-notched in male. Male genitalia: Median lobe (Fig. 31) short and robust with ostium moderately elongate, over half the length of the median lobe; apex with a very short and narrow distal end, slightly rounded in lateral aspect, less twisted laterally than in *M. costaricensis*, ventral margin proximal to apex straight then briefly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less 20 degrees off axis of shaft. Parameres large, left twice longer than the right, both broadly rounded, asetose. Female genitalia: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The six adults were collected at UV light trap in premontane forest understory. Adults are active in May, July, and August, in the rainy and early dry seasons.

Other specimens examined. Paratypes: Panamá, Chiriqui Province, Fortuna, 1050 m, 8.7341°N, 82.2701°W, 15 May 1978 (H. Wolda)(NMNH: ADP124964, male), 7 July 1978 (H. Wolda)(NMNH: ADP124964, male), 19 July 1978 (H. Wolda)(NMNH: ADP124952, male), 27 July 1977 (H. Wolda)(NMNH: ADP070275, male), 13 August 1978 (H. Wolda)(NMNH: ADP124954, male).

Geographic distribution. (Fig. 39). This species is currently known only from the type locality in the premontane forested midlands of Panamá.

Mizotrechus gorgona sp. n.

Isla Gorgona trough beetle

urn:lsid:zoobank.org:act:14F5ED42-8D4B-4720-829E-5F1145C10EA9 http://species-id.net/wiki/Mizotrechus_gorgona Figs 10, 24, 39

Holotype. COLOMBIA, Cacua, Isla Gorgona Mancora, 60 m, 2.96°N, 78.18°W, 26 June – 18 July 2000 (H Torres)(UASM: ADP128622, female).

Derivation of specific epithet. The epithet "*gorgona*" is a Latinized noun in apposition and is the name of the island off Colombia where these beetles are found.

Proposed English vernacular name. Isla Gorgona trough beetle.

Diagnosis. With the attributes of the genus as described above and moderately large sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose above and behind eye in an arc, not extended on occiput, and moderately punctulate. Occiput moderately punctulate. Pronotum longer than wide with lateral margins narrowly explanate and basally with sharp, not denticulate, hind angles; base smooth, without regular microrugose. Elytra moderately narrow and elongate, apex prolonged, wider than the width of pronotum across anterior third, and with 7 well-impressed irregularly punctulate interneurs, intervals not convex; margins behind humeri serrate, intervals between microsetae dentate. Foreleg femur with markedly serrate postero-ventral margin.

Description. (Figs 10, 24). *Size*: See Appendix 1. Moderately large for genus, ABL = 8.4 mm, SBL = 7.15 mm, TW = 2.86 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny; elytra matte. *Head*: Labrum quadrate, apico-medially barely emarginate. Eye small and barely convex. Gena straight. Frons, occiput, and gena glabrous. *Prothorax*: Moderately broad, narrowed slightly toward base, margin narrowly explanate, slightly wider before hind angle; surface punctulate, punctures widespread, very fine, glabrous. *Pterothorax*: Elytron barely convex, intervals nearly flat, all interneurs with well-impressed irregularly-spaced punctures, apex prolonged, slightly rounded at extreme sutural apex. *Legs*: Normal in female; foreleg femur (Fig. 24) with markedly serrate postero-ventral margin, not dentate; posterior trochanter tapered to rounded point in female, length half that of femur. *Abdomen*: Abdominal sterna moderately setiferous, densely medially on II and III; sternum VII barely medially notched in female. *Male genitalia*: Unknown. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The type specimen was collected in a Malaise trap in the rainforest understory at the type locality. Adults are active in late June, or early July during the "relatively" dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 39). This species is currently known only from the type locality on an island off the Pacific coast of Colombia.

Mizotrechus grossus sp. n.

Gross trough beetle

urn:lsid:zoobank.org:act:2C262CC2-85AE-4CA2-8BEB-07BEE80B4ADF http://species-id.net/wiki/Mizotrechus_grossus Figs 11, 32, 40, Plate 12

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 27 March 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP126301, male).

Derivation of specific epithet. The epithet "*grossus*" is a Latin adjective that adequately describes this species with large robust adults.

Proposed English vernacular name. Gross trough beetle.

Diagnosis. With the attributes of the genus as described above and very large sized for the genus as it is presently understood; adults have dark castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons shallowly rugose at sides and overall punctulate. Occiput punctulate. Pronotum cordiform with lateral margins straight to well-developed hind angle which is dentate; base densely microrugose. Elytra broad and short, moderately wider than the width of pronotum across anterior third, and with 3 well- impressed interneurs, outer interneurs more shallow toward margin; margins behind humeri uneven, but not evidently serrulate. Foreleg femur dentate on postero-ventral margin near base.

Description. (Figs 11, 32, Plate 12). Size: See Appendix 1. Large for genus, ABL = 8.1-8.9 mm, SBL = 7.07-7.71 mm, TW = 2.72-3.26 mm. Color: see diagnosis, above. Luster: Head, pronotum and legs shiny; elytron shiny but with evident shallow and slightly stretched microsculpture. Head: Labrum quadrate and medially emarginate. Eye moderately large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. Prothorax: Broadly cordiform, narrowed moderately toward base, margin narrowly explanate, more broadly near base, hind angle dentate; surface punctulate, punctures widespread, glabrous, microsculpture nearly effaced. Pterothorax: Elytron moderately convex, intervals flat, interneurs striate, not punctate, apex oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. Legs: Normal in both sexes; foreleg femur (as in Fig. 20) markedly dentate on postero-ventral margin at basal third; trochanter acute in both sexes, about half the length of the femur. Abdomen: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae. Defense gland system (Plate 12) with two very large, medially slightly concave, bean-shaped pygidial gland reservoirs, each with a markedly long and narrow annulated collecting canal connected to the efferent duct near the base of the gland. Male genitalia: Median lobe (Fig. 32) elongate and robust with ostium moderately elongate, over half the length of the median lobe; apex with a losp. n.tulate distal end slightly more bent ventrad than in M. poirieri, moderately curved in lateral aspect, ventral margin proximal to apex straight then evenly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded and slightly crested, opening more or less 40 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. Female genitalia: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The type series was collected in a flight intercept trap in the rainforest understory at four locations in Guyane. Adults are active in March, April, June, August, October, November, and December, during both the rainy and dry seasons.

Other specimens examined. Paratypes. Guyane, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 26 April 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP127064, female), 15 June 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP127062, ADP127062, females), 24 August 2009 (S Brule,

PH Dalens, & E Poirier)(NMNH: ADP124892, female), 10 October 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124898, male), 30 November 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124900, female); Mount Itoupe, 570 m, 3.0148°N, 53.0721°W, 17 March 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP126484, 126486, males); Montagne des Chevaux, Commune de Roura, RN2 PK22, 90 m, 4.7127°N, 52.3966°W, 9 August 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124958, male), 13 December 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124962, male); Reserve Tresor, Commune de Roura, Route de KAW km 18, 250 m, 4.6104°N, 52.2790°W, 21 November 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124972, male), Saül, Commune de Saül, Belvédère de Saül, 283–325 m, 3.6223°N, 53.2159°W, 17 November 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP127169, male), Commune de Regina, Reserve Naturelle des Nouragues Inselberg, Petit Plateau, 144 m, 4.0833°N, 52.6833°W, 5 June 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP127169, male).

Geographic distribution. (Fig. 40). This species is currently known from six localities in the lowlands and midlands of Guyane.

Mizotrechus jefe sp. n.

Jefe trough beetle urn:lsid:zoobank.org:act:3CE7E44F-2EBC-4B71-8251-F041B37D6081 http://species-id.net/wiki/Mizotrechus_jefe Figs 12, 33, 39

Holotype. PANAMÁ, Panamá Province, Cerro Jefe, 700–750 m, 9.2311°N, 79.3496°W, 20 May 1972 (RT Allen)(NMNH: ADP011173, female).

Derivation of specific epithet. The epithet "*jefe*" is a place name, based on the area where the type specimen was collected.

Proposed English vernacular name. Jefe trough beetle.

Diagnosis. With the attributes of the genus as described above and moderately small sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons with rugae nearly effaced, punctulate. Occiput punctulate. Pronotum narrowly cordiform with lateral margins shallowly emarginated; base rugulose. Elytra narrow and short, slightly wider than the width of pronotum across anterior third, and with 8 well-impressed interneurs; margins behind humeri moderately serrulate. Foreleg femur with slightly produced ridge on postero-ventral margin.

Description. (Fig. 12). *Size*: See Appendix 1. Moderately small for genus, ABL = 6.6 mm, SBL = 5.84 mm, TW = 2.27 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum and legs shiny; elytra shiny yet with evident isodiametric microsculpture. *Head*: Labrum quadrate and medio- apically slightly emarginate. Eye moderately large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous.

Prothorax: Moderately narrow, narrowed slightly toward base, margin narrowly explanate throughout, hind angle about right, not dentate; surface punctulate, punctures widespread, glabrous. *Pterothorax*: Elytron slightly convex, disk flat, intervals flat, interneurs striate, not punctate, apex moderately oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in female. *Legs*: Normal in female; foreleg femur (as in Fig. 21) with slightly produced, short, slightly arcuate ridge on postero-ventral margin at basal forth, not dentate; trochanter narrowly rounded. *Ab-domen*: Abdominal sterna moderately setiferous; sternum IV of female with median patch of setae that is more dense than elsewhere. *Male genitalia*: Unknown. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected along a road in the cloud forest understory. Adults are active in May, during the rainy season.

Other specimens examined. None.

Geographic distribution. (Fig. 39). This species is currently known only from the type locality in the midlands of Panamá.

Mizotrechus marielaforetae sp. n.

Laforêt's trough beetle

urn:lsid:zoobank.org:act:552D9468-9A77-49AF-A12D-5774041E1875 http://species-id.net/wiki/Mizotrechus_marielaforetae Figs 13, 30, 40

Holotype. GUYANE, Montagne des Chevaux, Commune de Roura, RN2 PK22, 90 m, 4.7127°N, 52.3966°W, 28 November 2010 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP127183, male).

Derivation of specific epithet. The epithet "*marielaforetae*" is an eponym, based on the full stage name of Marie Laforêt, the famous French actress and singer, 5 October 1939 – present, whose voice has a range of diversity like that found in the carabid species richness of Guyane, and who sang a variety of "torch songs", and here I play on the word "torch", the same word that applies to that being used to ignite the forests of the unique tropical rainforests of South America, an Armageddon in our own times.

Proposed English vernacular name. Laforêt's trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons without evident rugae, punctulate. Occiput at sides with evident rugae, punctulate. Pronotum nearly quadrate, quite narrowed behind, with lateral margins straight to hind angle, hind angle dentate, tooth small; base densely punctate. Elytra moderately broad and short, slightly wider than the width of pronotum across anterior third, and with only interneur 1 deeply engraved, 2–5 readily evident yet more shallowly



Plate 4. Digital Photo-illustrations, habitus, dorsal aspect: 13 *Mizotrechus marielaforetae* sp. n., dorsal aspect, ABL = 5.9 mm, male paratype, ADP127183, type locality. 14 *Mizotrechus minutus* sp. n., ABL = 4.4 mm, female paratype, ADP124920; Guyane, Commune de Saül, Belvédère de Saül. 15 *Mizotrechus neblinensis* sp. n., dorsal aspect, ABL = 6.9 mm, female holotype, ADP124944; type locality.
16 *Mizotrechus novemstriatus* Bates 1872, ABL = 7.2 mm, female lectotype, ADP127163; type locality.

impressed, 6–8 traceable, none punctate; margins behind humeri entire, slightly emarginate. Foreleg femur with slightly produced ridge on postero-ventral margin.

Description. (Figs 13, 30). Size: See Appendix 1. Moderately small sized for the genus, ABL = 5.9 mm, SBL = 5.12 mm, TW = 2.18 mm. Color: see diagnosis, above. Luster: Head, pronotum, and legs shiny, elytra duller due to shallowly engraved slightly stretched microsculpture. *Head*: Labrum quadrate and apico-medially emarginate. Eye large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. Prothorax: Narrow, quadrate, narrowed moderately toward base, margins straight before hind angle, angle dentate, tooth small, margin moderately explanate except wider at hind angle; surface punctulate, punctures widespread, glabrous. Pterothorax: Elytron moderately convex, intervals flat, proximal interneurs not punctate, distal interneurs somewhat punctate, apex moderately oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. Legs: Normal in male; foreleg femur (as in Fig. 23) with slightly produced ridge on postero-ventral margin, not dentate; posterior trochanter narrowly rounded at apex, about half the length of the femur. Abdomen: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae. Male genitalia: Median lobe (Fig. 30) short and robust with ostium moderately elongate, over half the length of the median lobe; apex with a short narrowly rounded distal end, ventral margin proximal to apex straight, then abruptly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less 40 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. Female genitalia: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. Adults are active in November, at the end of the dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 40). This species is currently known only from the type locality in the lowlands of Guyane.

Mizotrechus minutus sp. n.

Minute trough beetle

urn:lsid:zoobank.org:act:90504CC6-C4AD-4077-98F4-D5E501F72EB7 http://species-id.net/wiki/ Figs 14, 19, 34, 40

Holotype. GUYANE, Saül, Commune de Saül, Belvédère de Saül, 283–325 m, 3.6223°N, 53.2159°W, 10 December 2010 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124966, male).

Derivation of specific epithet. The epithet "*minutus*" is a Latin adjective that adequately describes this species with very small adults.

Proposed English vernacular name. Minute trough beetle.

Diagnosis. With the attributes of the genus as described above and very small sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons with rugae nearly effaced, punctulate. Occiput punctulate. Pronotum quadrate with lateral margins shallowly emarginated; base nearly smooth. Elytra broad and short, slightly wider than the width of pronotum across anterior third, and with 8 well-impressed punctate interneurs; margins behind humeri not serrulate.

Description. (Figs 14, 19, 34). Size: See Appendix 1. Very small, ABL = 4.4-4.6 mm, SBL = 3.77-3.88 mm, TW = 1.49-1.58 mm. Color: see diagnosis, above. Luster: Head, pronotum, elytra, and legs shiny; microsculpture effaced. Head: Labrum quadrate and apico-medially emarginate. Eye moderately large, moderately convex. Gena moderately long, straight. Frons, occiput, and genae glabrous. Prothorax: Broad, narrowed slightly toward base, margin slightly explanate to hind angle, angle about right, not dentate; surface punctulate, punctures widespread, glabrous. Pterothorax: Elytron moderately convex, intervals slightly convex, interneurs striato- punctate, apex moderately oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. Legs: Normal in both sexes; foreleg femur unmodified (Fig. 19); trochanter narrowly rounded. Abdomen: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae. Male genitalia: Median lobe (Fig. 34) elongate and narrow with ostium moderately elongate, over half the length of the median lobe; apex with a long narrow distal end, straight in lateral aspect and narrowly rounded, ventral margin proximal to apex straight; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less 15 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. Female genitalia: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. Adults are active in December, at the end of the dry season.

Other specimens examined. Paratypes: Guyane, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 11 December 2009 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124902, male), Saül, Commune de Saül, Belvédère de Saül, 283–325 m, 3.6223°N, 53.2159°W, 10 December 2010 (S Brule, PH Dalens, & E Poirier)(NMNH: ADP124920, female).

Geographic distribution. (Fig. 40). This species is currently known only from two localities in the lowlands of Guyane.

Mizotrechus neblinensis sp. n.

Tipui trough beetle urn:lsid:zoobank.org:act:11B2AD1D-46FC-45BD-8F93-4BA20D3CA2CF http://species-id.net/wiki/Mizotrechus_neblinensis Figs 15, 35, 40, 41

Holotype. VENEZUELA, Amazonas, Cerro de la Neblina (Smithsonian Basecamp), 140 m, 0.0148°N, 66.1604°W, 10–20 February 1985 (PJ Spangler, PM Spangler, RE Faitoute & WE Steiner)(NMNH: ADP124944, female).

Derivation of specific epithet. The epithet "*neblinensis*" is a Latinized noun in apposition, based on a geographic name formed from Pico de Neblina, a tipui on the border of Venezuela and Brazil, the area in which adults of this species have been found, and the Latin suffix *"-ensis,*" meaning of, or from that place.

Proposed English vernacular name. Tipui trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons with nearly effaced rugae, punctulate. Occiput punctulate. Pronotum nearly quadrate with lateral margins straight to hind angle, hind angle a very small tooth; base densely rugulose. Elytra broad and short, slightly wider than the width of pronotum across anterior third, and with 6 evident shallowly impressed interneurs, 7–8 nearly effaced; margins behind humeri moderately serrulate. Foreleg femur with markedly developed and produced ridge on postero-ventral margin.

Description. (Figs 15, 35). Size: See Appendix 1. Medium sized for the genus, ABL = 6.4–6.9 mm, SBL = 5.54–6.12 mm, TW = 2.25–2.46 mm. *Color*: see diagnosis, above. Luster: Head, pronotum, elytra and legs shiny, microsculpture absent. Head: Labrum quadrate and apico- medially emarginate. Eye large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. Prothorax: Broad, narrowed slightly toward base, margin narrowly explanate except wider at hind angle, hind angle a very small tooth; surface punctulate, punctures widespread, glabrous. Pterothorax: Elytron moderately convex, disk flat, intervals flat, interneurs striato-punctate, punctures small, shallow, apex oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. Legs: Normal in both sexes; foreleg femur (as in Fig. 22) ridged along the posterio-ventral margin and markedly produced at middle of ridge; posterior trochanter narrowly rounded at apex, about half the length of the femur. Abdomen: Abdominal sterna moderately setiferous; sternum IV of female with denser patch of setae at midline. Male genitalia: Median lobe (Fig. 35) short and robust with ostium moderately elongate, over half the length of the median lobe; apex a short blunt distal end, slightly curved in lateral aspect, ventral margin proximal to apex evenly arched and barely curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded, opening more or less five degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.
Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. Adults are active in February, in the rainy season.

Other specimens examined. Paratypes: Guyane, Montagne des Chevaux, Commune de Roura, RN2 PK22, 90 m, 4.7127°N, 52.3966°W, 19 September 2009 (NMNH: ADP124970, male), 31 October 2010 (NMNH: ADP128733, male). Venezuela, Amazonas, Cerro de la Neblina (Smithsonian Basecamp), 140 m, 0.0148°N, 66.1604°W, 10–20 February 1985 (PJ Spangler, PM Spangler, RE Faitoute & WE Steiner)(NMNH: ADP124948, female).

Geographic distribution. (Figs 40, 41). This species is currently known from low-land localities, one each in Guyane and Venezuela.

Mizotrechus novemstriatus Bates, 1872

Nine-lined trough beetle

http://species-id.net/wiki/Mizotrechus_novemstriatus Figs 16, 41

Lectotype. (labeled by me in Paris, early 1970's): BRAZIL: Amazonas, Ega (Tefé) approximately 3.35°S, 64.71°W, 46 m (HW Bates)(MNHP: female). ADP127163.

Derivation of specific epithet. The epithet "*novemstriatus*" is descriptive Latin, meaning 9-lined and referring to the elytral interneurs, including the scutellar striole.

Proposed English vernacular name. Nine-lined trough beetle.

Diagnosis. With the attributes of the genus as described above and moderate sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons smooth, scattered rugulae not organized, moderately punctulate. Occiput smooth, moderately punctulate. Pronotum subcordiform with lateral margins narrowly explanate and basally with sharp, not denticulate, hind angles; base with regular rugulae. Elytra broad and short, wider than the width of pronotum across anterior third, and with 8 well-impressed irregularly punctulate interneurs, intervals not convex, apex not prolonged, margins behind humeri rough, intervals between microsetae blunt. Foreleg femur with evident ridge on postero-ventral margin.

Description. (Fig. 16). *Size*: See Appendix 1. Moderate sized for genus, ABL = 7.2 mm, SBL = 6.05 mm, TW = 2.48 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, elytra, and legs shiny. *Head*: Labrum quadrate, apico-medially v-notched. Eye small and moderately convex. Gena straight. Frons, occiput, and gena glabrous. *Prothorax*: Moderately broad, narrowed slightly toward base, margin narrowly explanate, slightly wider before hind angle; surface punctulate, punctures widespread, very fine, glabrous. *Pterothorax*: Elytron barely convex, intervals nearly flat, all interneurs with well-impressed irregularly-spaced punctulate, apex not prolonged, slightly rounded at extreme sutural apex. *Legs*: Normal in female; foreleg fe-

mur (as in Fig. 22) ridged along the postero-ventral margin and markedly produced at middle of ridge; posterior trochanter tapered to rounded point in female, length half that of femur. *Abdomen*: Abdominal sterna moderately setiferous, densely medially on II and III; sternum VII barely medially notched in female. *Male genitalia*: Unknown. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. Unknown.

Other specimens examined. None.

Geographic distribution. (Fig. 41). This species is currently known only from the type locality in the lowlands of Brazil.

Mizotrechus poirieri sp. n.

Poirier's trough beetle

urn:lsid:zoobank.org:act:FA89B987-1B34-4AAA-B5A4-DE4971821A76 http://species-id.net/wiki/Mizotrechus_poirieri Figs 17, 36, 40

Holotype. GUYANE, Saut Pararé, Arataie River, Nouragues Field Station, 51 m, 4.0378°N, 52.6725°W, 30 November 2009 (S Brule, PH Dalens, & E Poirier) (NMNH: ADP124888, male).

Derivation of specific epithet. The epithet "*poirieri*" is an eponym, based on the family name of E. Poirieri, whose team in Guyane has been collecting beetles using Flight Intercept Traps and capturing adults of many new species, such as this one.

Proposed English vernacular name. Poirier's trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons with evident rugae, punctulate. Occiput with evident rugae, punctulate. Pronotum nearly quadrate with lateral margins slightly emarginate to hind angle, hind angle about right, not dentate; base sparsely rugulose. Elytra broad and short, much wider than the width of pronotum across anterior third, and with only interneur 1 deeply engraved, 2–8 evident yet shallowly impressed, not punctate; margins behind humeri entire. Foreleg femur with slightly produced swelling on postero-ventral margin.

Description. (Figs 17, 36). *Size*: See Appendix 1. Medium sized for the genus, ABL = 6.6 mm, SBL = 5.65 mm, TW = 2.42 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny, elytra duller due to shallowly engraved slightly stretched microsculpture. *Head*: Labrum quadrate and apico-medially emarginate. Eye large, moderately convex. Gena moderately long, straight. Frons, occiput, and gena glabrous. *Prothorax*: Narrow, quadrate, narrowed slightly toward base, margins slightly emarginated before hind angle, angle about right, not dentate, margin moderately ex-



Plate 5. Digital Photo-illustrations, habitus, dorsal aspect: **17** *Mizotrechus poirieri* sp. n., ABL = 6.6 mm, male holotype, ADP124888; type locality. **18** *Mizotrechus woldai* sp. n., dorsal aspect, ABL = 7.4 mm, female holotype, ADP124946; type locality.

planate except wider at hind angle; surface punctulate, punctures widespread, glabrous. *Pterothorax*: Elytron moderately convex, disk flat, intervals flat, interneurs not punctate, apex markedly oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. *Legs*: Normal in male; foreleg femur (as in Fig. 20) with a minute setose tooth on postero-ventral margin, subdentate; posterior trochanter narrowly acute at apex, about half the length of the femur. *Abdomen*: Abdominal sterna moderately setiferous; sternum IV of male with narrow and dense patch of decumbent setae. *Male genitalia*: Median lobe (Fig. 36) elongate and robust with ostium moderately elongate, over half the length of the median lobe; apex a losp. n.tulate distal end that is more bent ventrad than in *M. dalensi*, moderately curved in lateral aspect, ventral margin proximal to apex straight then evenly curved to apex; endophallus with complexly folded tracheal fields; phallobase hooded and crested, opening more or less 20 degrees off axis of shaft. Parameres large, left a third longer than the right, both broadly rounded, asetose. *Female genitalia*: Unknown.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.

Way of life. The adult holotype was collected in a flight intercept trap in the rainforest understory. Adults are active in November, at the end of the dry season.

Other specimens examined. None.

Geographic distribution. (Fig. 40). This species is currently known only from the type locality in the lowlands of Guyane.

Mizotrechus woldai sp. n. Wolda's trough beetle urn:lsid:zoobank.org:act:2806BBDA-1FDF-4030-B08F-4BAF13DBBC60 http://species-id.net/wiki/Mizotrechus_woldai Figs 18, 39

Holotype. PANAMÁ, Canal Zone, Barro Colorado Island, 99 m, 9.1628°N, 79.8395°W, 5 May 1978 (H Wolda)(NMNH: ADP 124946, female).

Derivation of specific epithet. The epithet "*woldai*" is an eponym, based on the family name of H. Wolda, long time scientist with the Smithsonian Tropical Research Institute in Panamá, whose persistent light trapping produced many new and interesting species, such as one adult of this new species.

Proposed English vernacular name. Wolda's trough beetle.

Diagnosis. With the attributes of the genus as described above and medium sized for the genus as it is presently understood; adults have castaneous integument, except anterior parts of mandible, baso-lateral corner of labrum, and clypeal suture piceous. Frons with evident rugae, punctulate. Occiput without rugae, punctulate. Pronotum nearly quadrate with lateral margins slightly emarginate to hind angle, hind angle dentate, tooth small; base sparsely rugulose. Elytra broad and short, slightly wider than the width of pronotum across anterior third, and with only interneurs 8 deeply engraved, not punctate; margins behind humeri serrate.

Description. (Fig. 18). *Size*: See Appendix 1. Medium sized for the genus, ABL = 7.4 mm, SBL = 6.04 mm, TW = 2.29 mm. *Color*: see diagnosis, above. *Luster*: Head, pronotum, and legs shiny, elytra duller due to shallowly engraved slightly stretched microsculpture. *Head*: Labrum quadrate and apico-medially emarginate. Eye large, moderately convex. Gena moderately short, straight. Frons, occiput, and gena glabrous. *Prothorax*: Very broad, quadrate, narrowed slightly toward base, margins slightly emarginated before hind angle, angle dentate, margin beaded, moderately explanate at hind angle; surface punctulate, punctures widespread, glabrous. *Pterothorax*: Elytron flat, disk flat, intervals flat, interneurs not punctate, apex moderately oblique and straight, sutural apex narrowly rounded. Metasternum sparsely setiferous in male. *Legs*: Normal in female; foreleg femur (as in Fig. 21) with slightly produced, short, arcuate ridge on postero-ventral margin at basal forth, not dentate. *Abdomen*: Abdominal sterna moderately setiferous. *Male genitalia*: Unknown. *Female genitalia*: Not investigated; however, it is likely similar to that illustrated on Plate 11.

Dispersal potential. These beetles are macropterous and capable of flight. They are moderately swift and agile runners.



Plate 6. Digital Photo-illustrations, anterior femur, ventral aspect: 19 Mizotrechus minutus sp. n., ADP124966; type locality. 20 Mizotrechus dalensi sp. n., ADP124896; type locality. 21 Mizotrechus chon-talesensis sp. n., ADP127181; type locality. 22 Mizotrechus edithpiafae sp. n., ADP124948; type locality.
23 Mizotrechus brulei sp. n., ADP129205; type locality. 24 Mizotrechus gorgona. sp. n., ADP128622; type locality.

Way of life. The adult holotype was collected in a ground level UV light trap in the rainforest understory. Adults are active in May during the heavy rainy season in Panamá.

Other specimens examined. None.

Geographic distribution. (Fig. 39). This species is currently known from a single lowland locality in Panamá.

Note. Considering the hundreds of specimens of carabids collected by Wolda in a series of suspended light traps from canopy level to ground level, it is curious that no other specimens of this species were collected (see *M. fortunensis* above, where many specimens were collected by Wolda at light traps).



Plate 7. Digital Photo-illustrations, male genitalia, median lobe and parameres, left lateral, dorsal, and right lateral aspects: **25** *Mizotrechus bellorum* sp. n., ADP124890; type locality. **26** *Mizotrechus belvedere* sp. n., ADP129201; type locality. **27** *Mizotrechus brulei* sp. n., ADP127159, Guyane, Montagne des Chevaux, Commune de Roura.



Plate 8. Digital Photo-illustration, male genitalia, median lobe and parameres, left lateral, dorsal, and right lateral aspects: **28** *Mizotrechus costaricensis* sp. n., ADP128620: type locality. **29** *Mizotrechus dalensi* sp. n., ADP129203, Guyane, Saut Pararé, Arataie River, Nouragues Field Station. **30** *Mizotrechus edithpiafae* sp. n., ADP129203; G^{er} ?locality (see text).



Plate 9. Digital Photo-illustration, male genitalia, median lobe, left lateral, dorsal, and right lateral aspects: **31** *Mizotrechus fortunensis* sp. n., ADP124590; type locality. **32** *Mizotrechus grossus* sp. n., ADP127165; type locality. **33** *Mizotrechus marielaforetae* sp. n., ADP127183; type locality.



Plate 10. Digital Photo-illustration, male genitalia, median lobe, left lateral, dorsal, and right lateral aspects: **34** *Mizotrechus minutus* sp. n., ADP124902; type locality. **35** *Mizotrechus neblinensis* sp. n., ADP124944; type locality. **36** *Mizotrechus poirieri* sp. n., ADP124888; type locality.



Plate 11. Digital Photo-illustration, female genitalia: *Mizotrechus dalensi* sp. n., ADP127161; Petite Montague Tortue, Guyane. A. Dorsal aspect. Legend, **bc**, bursa copulatrix; **co**, common oviduct; **sg**, spermathecal gland; **sgd**, spermathecal gland duct; sp, spermatheca. dorsal aspect; **vc**, villous canal; **lt**, laterotergite; **gc1**, gonocoxite 1; **gc2**, gonocoxite 2. B. Gonocoxite 2, dorsal aspect: Legend, **b**, base of gonocoxite 2; **bl**, blade of gonocoxite 2; **des**, dorsal ensiform seta; **ves**, ventral ensiform setae.



Plate 12. Digital Photo-illustration, pygidial (defense) gland system: *Mizotrechus grossus* sp. n., dorsal aspect, ADP127064; type locality. Legend: cc, collecting canal; ed, efferent duct; edbl, efferent duct, basal lobe; gldr, pygidial gland reservoir. Ovipositor sclerites, Legend: gc1, gonocoxite 1; gc2, gonocoxite 2; lt, laterotergite.



Figure 39. Distribution symbol map for known localities of *M. chontalesensis*, *M. costaricensis*, *M. fortunensis*, *M. gorgona*, *M. jefe*, *M. woldai*, spp. n.



Figure 40. Distribution symbol map for known localities of *M. batesi*, *M. bellorum*, *M. belvedere*, *M. brulei*, *M. dalensi*, *M. grossus*, *M. marielaforetae*, *M. minutus*, *M. poirieri*, *M. neblinensis*, spp. n.



Figure 41. Distribution symbol map for known localities of M. neblinensis, sp. n., M. novemstriatus Bates.

Discussion

For 108 years, adults of *Mizotrechus* Bates 1872 have flown "under the radar," so to speak. Then, along came the FITs (Flight Intercept Traps) used by the SEAG group in Guyane. Since collection of the unique holotype of *M. novemstriatus* Bates from Ega (now Tefé), Brazil, and a misidentified (by Bates) specimen collected by Janson in Nicaragua, this genus has remained an enigma and rarely collected. All the very few specimens that have resided in collections have been determined as *M. novemstriatus* Bates, because the genus was thought to be monobasic. The synopsis presented in this paper is an opening salvo, only. There is no question that the use of FITs, in the rest of the Oronoco and Amazon Basins and in Middle America, will produce many new spe-

cies in addition to those described herein. The type locality of the type species is south of the Amazon River; and this suggests that there may be many more species residing on the Brazilian Shield, just as there are on the Guyana Shield, as revealed by this study.

With the increase in our knowledge from two specimens to 56 specimens and from one species to 18 species, this poorly known genus now has become recognized as a likely dominate subcortical (or leaf litter?) inhabitant of the Neotropics. The Tribe Perigonini is little known overall and has never been a group seriously studied in the Western Hemisphere. It is very likely that the other two described genera, *Perigona* Laporte de Castelnau 1835 and *Diploharpus* Chaudoir 1850, and a third as yet undescribed genus (Erwin, in prep) will also be replete with new species, as is *Mizotrechus*, once they are studied. For example, *Diploharpus* is now known to have 13 described species assigned to it throughout its entire range from México south to Bolivia and Brazil. In the Guyane FIT samples alone, I have sorted out 27 morphospecies of *Diploharpus*.

While writing descriptions and creating the key, it became apparent that certain specific attributes have begun to form biogeographic patterns. The serrate humerus is found in the Central America/Colombia species and one Guyane species, and the serrate femur in the same species in Central America/Colombia only; the dentate femora only in the Guyane species. When more species are discovered and documented/described, this genus has the potential of contributing to a better understanding of the biogeography of the Central American/northern South American connections.

As I have documented with my fogging assemblages, a novel collecting technique opens new and exciting faunal components to scientific investigation and knowledge. Unfortunately, the opening of new taxonomic vistas with these new collecting techniques, and discovery of the new and/or rare species found, comes at a time when taxonomists themselves are becoming rare (Pearson et al. 2010; Cardosa et al. 2011). While these authors provide some solutions, the question remains: Will those solutions be enough and in time, while biodiversity is still here, to help us fully grasp what global species richness means (Erwin et al. 2005)?

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References

- Bates HW (1872) Notes on Carabidae, and descriptions of new species (No. 12). Entomologist's Monthly Magazine 8: 199–202.
- Bates HW (1883) Coleoptera, Carabidae, Cicindelidae, vol. 1, pt. 1, Supplement, 153–255.In: Godman FD, Salvin O (Eds) Biologia Centrali-Americana. Taylor & Francis, London.
- Cardosa P, Erwin TL, Borgas AV, New TR (2011) The seven impediments in invertebrate conservation and how to overcome them. Biological Conservation 144 (2011): 2647–2655. doi: 10.1016/j.biocon.2011.07.024
- Chaudoir M de (1850) Mémoire sur la familla des carabiques. Bulletin de la Société Impériale des naturalistas de Moscou 23: 349–460.
- Erwin TL (1974) Studies of the subtribe Tachyina (Coleoptera: Carabidae: Bembidiini), Part II: A revision of the New World-Australian Genus *Pericompsus* LeConte. Smithsonian Contributions to Zoology No. 162, 96 pp.
- Erwin TL (2011a) Halocoryza Alluaud 1919, sea-side beetles of the Atlantic, Pacific, and Indian Oceans: a generic synopsis and description of a remarkable new species from Baja California (Coleoptera, Carabidae, Scaritini, Clivinina). ZooKeys 127: 1–13. doi: 10.3897/ zookeys.127.1748
- Erwin TL (2011b) A Treatise on the Western Hemisphere Caraboidea (Coleoptera): Their classification, distributions, and ways of life Volume III (Carabidae – LOXOMERIFORMES, MELAENIFORMES), Pensoft, Sofia-Moscow, 412 pp., 66 plates.
- Erwin TL, Johnson PJ (2000) Naming species, a new paradigm for crisis management in taxonomy: Rapid journal validation of scientific names enhanced with more complete descriptions on the internet. The Coleopterists Bulletin 54(3): 269–278. doi: 10.1649/0010-065X(2000)054[0269:NSANPF]2.0.CO;2
- Erwin TL, Kavanaugh DH (1981) Systematics and zoogeography of *Bembidion* Latreille: I. The *carlhi* and *erasum* groups of western North America (Coleoptera: Carabidae, Bembidiini). Entomologica Scandinavica, Supplement 15: 33–72.
- Erwin TL, Pimienta MC, Murillo OE, Aschero V (2005). Mapping patterns of β-diversity for beetles across the western Amazon Basin: A preliminary case for improving conservation strategies. Proceedings of the California Academy of Sciences, ser. 4, 56(Suppl. I) No. 7: 72–85.
- Kavanaugh DH (1979) Studies on the Nebriini (Coleoptera: Carabidae), III. New Nearctic Nebria species and subspecies, nomenclatural notes, and lectotype designations. Proceedings of the California Academy of Sciences ser. 4, 42: 87–133.
- Laporte de Castelnau FL (1835) In: Études entomologiques, ou descriptions d'insectes nouveaux et observations sur la synonymie. Méquignon-Marvis, Paris, 95–159.
- Liebherr JK, Will KW (1998) Inferring phylogenetic relationships within the Carabidae (Insecta, Coleoptera) from characters of the female reproductive tract. In: Ball GE, Casale A, Vigna-Taglianti A (Eds) Phylogeny and classification of Caraboidea (Coleoptera, Adephaga): Proceedings of a Symposium (28 August 1996, Florence, Italy) XX International Congress of Entomology. Atti, Museo Regionali di Scienze naturali, Torino, Italy, 107–170.

- Liu Y, Kavanaugh DH, Shi HL, Liang HB (2011) A key to species of subgenus *Lithochlaenius* (Coleoptera, Carabidae, Chlaeniini, *Chlaenius*), with descriptions of three new species. ZooKeys 128: 15–52. doi: 10.3897/zookeys.128.1804
- Pearson DL, Hamilton AL, Erwin TL (2011) Recovery Plan for the Endangered Taxonomy Profession. Bioscience 61(1): 58–63. 10.1525/bio.2011.61.1.11
- Snodgrass, RE (1935) Principles of Insect Morphology. McGraw-Hill Book Company, Inc. New York.
- Will KW, Attygalle AB, Herath K (2000) New defensive chemical data for ground beetles (Coleoptera: Carabidae): interpretations in a phylogenetic framework. Biological Journal of the Linnaean Society 71: 459–481.

Appendix I.

Tables with measurements and ratios for adults of species of *Mizotrechus* Bates 1872. All measurements in millimeters. Apparent body length (ABL) provided in the descriptions.

		TOTAL I	ENGTH (SB	L)			
		Males		Females			
Species	Ν	Range	Mean	N	Range	Mean	
batesi	1	4.897					
bellorum	1	5.874					
belvedere	1	6.142					
brulei	2	5.535-5.644	5.59	7	6.408-7.096	6.759	
chontalesensis				1	5.886		
costaricensis	1	6.888					
dalensi	4	6.404–7.101	6.736	4	6.408-7.096	6.759	
edithpiafae	1	7.162					
fortunensis	6	7.02–7.66	7.256				
gorgona				1	7.15		
grossus	10	7.068-7.711	7.301	4	6.986–7.643	7.257	
jefe				1	5.836		
marielaforetae	1	5.123					
minutus	1	3.767		2	3.7341-3.879	3.807	
neblenaensis	2	5.665-5.69	5.678	2	5.539-6.118	5.829	
novemstriatus				1	6.045		
poirieri	1	5.649	5.649				
woldai				1	6.404		
		MAXIM	IUM WIDTH	ł			
		Males		Females			
Species	Ν	Range	Mean	Ν	Range	Mean	
batesi	1	1.792					
bellorum	1	2.556					
belvedere	1	2.484					
brulei	2	2.036-2.046	2.041	7	1.986-2.224	2.123	
chontalesensis				1	2.286		
costaricensis	1	2.888					
dalensi	4	2.646-2.964	2.813	4	2.724-3.114	2.833	
edithpiafae	1	2.942					
fortunaensis	6	2.928-3.032	2.972				
gorgona				1	2.86		
grossus	10	2.72-3.114	2.974	4	2.846-3.262	2.994	
jefe				1	2.272		
marielaforetae	1	2.178					
minutus	1	1.49		2	1.51-1.576	1.543	
neblenaensis	2	2.256-2.358	2.307	2	2.3–2.464	2.382	

novemstriatus			1	2.48	
poirieri	1	2.418			
woldai			1	2.286	

		Males			Females			
Species	N	Range	Mean	N	Range	Mean		
batesi	1	1.434						
bellorum	1	1.191						
belvedere	1	1.196						
brulei	2	1.459-1.495	1.477	7	1.213-1.434	1.307		
chontalesensis				1	1.358			
costaricensis	1	1.139						
dalensi	4	1.199–1.274	1.234	4	1.170-1.287	1.229		
edithpiafae	1	1.218						
fortunaensis	6	1.057-1.151	1.109					
gorgona				1	1.192			
grossus	10	1.201–1.324	1.238	4	1.186-1.281	1.227		
jefe				1	1.3			
marielaforetae	1	1.198						
minutus	1	1.27		2	1.280-1.282	1.281		
neblenaensis	2	1.207-1.238	1.223	2	1.276-1.306	1.291		
novemstriatus				1	1.325			
poirieri	1	1.191						
woldai				1	1.358			

PRONOTUM: WIDTH (at widest part)/LENGTH

		Males		Females			
Species	N	Range	Mean	N	Range	Mean	
batesi	1	1.179					
bellorum	1	1.283					
belvedere	1	1.208					
brulei	2	1.192-1.236	1.214	7	1.204-1.308	1.248	
chontalesensis				1	1.226		
costaricensis	1	1.236					
dalensi	4	1.333-1.362	1.347	4	1.305-1.414	1.362	
edithpiafae	1	1.281					
fortunaensis	6	1.179-1.249	1.215				
gorgona				1	1.194		
grossus	10	1.303-1.383	1.344	4	1.295-1.360	1.324	
jefe				1	1.206		
marielaforetae	1	1.222					
minutus	1	1.311		2	1.315-1.316	1.315	
neblenaensis	2	1.262-1.328	1.295	2	1.281-1.347	1.314	
novemstriatus				1	1.356		
poirieri	1	1.308					
woldai				1	1.168		

LENGTH OF PRONOTUM / LENGTH OF HEAD								
		Males			Females			
Species	N	Range	Mean	N	Range	Mean		
batesi	1	1.938						
bellorum	1	1.897						
belvedere	1	2.29						
brulei	2	2.059-2.121	2.09	7	1.965-2.201	2.073		
chontalesensis				1	2.243			
costaricensis	1	2.4						
dalensi	4	2.061-2.231	2.134	4	1.95-2.186	2.06		
edithpiafae	1	2.456						
fortunaensis	6	2.144-2.347	2.206					
gorgona				1	2.275			
grossus	10	1.942-2.265	2.08	4	1.938-2.288	2.087		
jefe				1	2.075			
marielaforetae	1	2.263						
minutus	1	2.113		2	2.097-2.233	2.165		
neblenaensis	2	2.001-2.257	2.129	2	1.996-2.228	2.112		
novemstriatus				1	2.188			
poirieri	1	1.892						
woldai				1	2.812			

RESEARCH ARTICLE



Two new species of *Harpactea* (Araneae, Dysderidae) from Turkey

Kadir Boğaç Kunt^{1,†}, Mert Elverici^{2,‡}, Recep Sulhi Özkütük^{3,§}, Ersen Aydın Yağmur^{4,|}

I Poligon Sitesi 71/27-B TR-06810 Dodurga, Çayyolu, Ankara, Turkey 2 Department of Biological Sciences, Faculty of Arts and Sciences, Middle East Technical University, TR-06531 Ankara, Turkey 3 Department of Biology, Faculty of Science, Anadolu University, TR-26470 Eskişehir, Turkey 4 Ege University, Science Faculty, Biology Department, Zoology Section, TR-35100 İzmir, Turkey

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 urn:lsid:zoobank.org:author:7A21C546-989F-417F-BCC3-8D682CCF2B62
 urn:lsid:zoobank.org:author:8DB0B243-5B2F-4428-B457-035A8274500C

Corresponding author: Kadir Boğaç Kunt (chaetopelma@gmail.com)

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Abstract

Two new species, *Harpactea arnedoi* sp. n. and *H. kencei* sp. n., are described on the basis of both sexes from the eastern Mediterranean and Aegean regions of Turkey. *H. kencei* sp. n. can be easily distinguished from all other Turkish and European representatives of the genus by the structure of the flattened, massive embolus on the male copulatory organ. Although resembling *Stalagtia* in palpal morphology, we describe one of the new species as *Harpactea arnedoi* sp. n. For both Turkish species, detailed morphological descriptions and diagnoses are presented together with figures of the copulatory organs.

Keywords

Dysderidae, Harpacteinae, Eastern Mediterranean, Aegean

Introduction

The first records of the genus *Harpactea* from Turkey were *H. babori* from İstanbul Province and *H. sturanyi* from Konya Province (Nosek 1905). However, the majority of Turkish *Harpactea* species were described by Brignoli (1978a-b, 1979). In recent years, Lazarov and Deltshev (2008) and Lazarov (2010) re-described *H. sanctaeinsulae* and *H. babori* respectively, which were previously only known from males. Bayram et al. (2009) described *H. christodeltshevi* from the south-eastern Anatolia and Kunt et al. (2010) described *H. erseni*, from the Aegean region. Currently 20 species are known from Turkey (Bayram et al. 2011).

During our surveys of the Turkish spider fauna, we have encountered two new species of *Harpactea* from the eastern Mediterranean and Aegean regions. The purpose of this study is to describe and diagnose these two new species on the basis of both sexes.

Materials and methods

Specimens were collected by means of pitfall traps, sifter and hand collecting from two different provinces of Turkey (Fig. 1). Digital images of the pedipalps and vulvae were taken with a Leica DFC295 digital camera attached to a Leica S8AP0 stereomicroscope, with 5–15 photographs taken in different focal planes and combined using image stacking software. Photographic images were edited using PHOTOSHOP CS2 and COREL-DRAW X3 was used to create the plates. All measurements are in mm. Terminology for the body measurements and copulatory organs follows Chatzaki and Arnedo (2006).

The following abbreviations are used in the text and figures: Carapace and abdomen: AL, abdominal length; CL, carapace length; CWmax, maximum carapace width; CWmin, minimum carapace width. Eyes: AME, anterior median eyes; PLE, posterior lateral eyes; PME, posterior median eyes; AMEd, diameter of anterior median eyes; PLEd, diameter of posterior lateral eyes; PMEd, diameter of posterior median eyes. Chelicera: ChF, length of cheliceral fang; ChG, length of cheliceral groove; ChL, total length of chelicera (lateral external view). Legs: Ta, tarsus; Me, metatarsus, Ti, tibia; Pa, patella; Fe, femur; Tr, trochanter; C, coxa; D, dorsal; Pl, prolateral; Rl, retrolateral; V, ventral. Male bulb: AA, accessory apophysis; Co, conductor; E, embolus. Vulva: aba, anterior basal arc; As, anterior spermatheca; btas, basal transverse part of the anterior spermatheca; dc, distal crest; des, distal expansion of the spermatheca; pd, posterior diverticulum; rsas, rod-shaped part of the anterior spermatheca; tb, transverse bar. Depository: AUZM, Anadolu University, Zoology Museum, Eskişehir, Turkey; cKBK, Personal collection of Kadir Boğaç Kunt, Ankara, Turkey; SMF, Senckenberg Museum, Frankfurt am Main, Germany.



Figure 1. Type localities of the new species. ★ *H. arnedoi* sp. n. ■ *H. kencei* sp. n.

Taxonomy

Family Dysderidae C. L. Koch, 1837 Genus *Harpactea* Bristowe, 1939

Harpactea arnedoi sp. n.

urn:lsid:zoobank.org:act:8BC9D2B7-4E3D-4A17-8017-6DAB08F82D8F http://species-id.net/wiki/Harpactea_arnedoi Figs 2–8

Material examined. Holotype (AUZM), Turkey, Gaziantep Province, Kuşçubeli Pass [37°6'50.20"N; 36°36'34.20"E], 13.XI.2010, under leaf litter, leg. E.A.Yağmur. **Paratypes** 1 \bigcirc , 1 \bigcirc (AUZM); 2 $\bigcirc \bigcirc$, 2 $\bigcirc \bigcirc$, 9 juveniles (cKBK & SMF), same data as holotype.

Derivatio nominis. The new species is dedicated to the Spanish arachnologist, Dr. Miquel Arnedo (Barcelona University, Barcelona, Spain), who has made important contributions to the taxonomy of the family Dysderidae.

Diagnosis. *H. arnedoi* sp. n. differs from all other *Harpactea* species in the structure of the copulatory organs. However, the general morphology of the male palp resembles that of *H. zoiai* Gasparo, 1999 known from Greece (see Gasparo 1999). The male of *H. arnedoi* sp. n. differs in having an uneven spherical shape of the palpal bulb; the region between the bulb and the distal continuation is elongated and has a funnel-like appearance; the embolus is shorter, hook-like and bends downwards towards the tip. The vulva is apparent and characterized by a peripherally sclerotized posterior diverticulum and folded distal expansion.



Figures 2–6. *Harpactea arnedoi* sp. n. 2 Cheliceral teeth 3 left palp, retrolateral view 4 ditto, nearly anterior view 5 ditto, nearly retrolateral view 6 ditto, distal view. Scale line: 0.5 mm.

Notes. While describing *H. arnedoi* sp. n., it was difficult to decide whether it would be best assigned to *Harpactea* or to *Stalagtia*. By the characteristic structure of the male copulatory organ (globular tegulum, long embolus, poorly developed conductor), the new species is similar to members of *Stalagtia*. However, the male palp possesses a poorly developed conductor (Fig. 4), and the female copulatory organs are similar to those of *Harpactea* species in having a short posterior diverticulum and anterior spermathecae. Thus, we feel the new species is correctly placed in *Harpactea*. Further evidence in support of this generic placement is the absence of ventral spines on the anterior tibiae and metatarsi in the new species, particularly as Deeleman-Reinhold (1993) stated that their presence was diagnostic of *Stalagtia*. However, the only known Turkish species of *Stalagtia*, *S. thaleriana* Chatzaki & Arnedo, 2006, does not possess ventral spines on the anterior tibiae and metatarsi, as was mentioned earlier by Chatzaki and Arnedo (2006) and Kunt et al. (2009).

Description. Measurements: [Holotype ♂ / Paratype ♀]: AL 2.26 / 2.50; CL 1.75 / 1.73; CWmax 1.45 / 1.35; CWmin 0.63 / 0.65; AMEd 0.10 / 0.11; PLEd 0.09 /

0.09; PMEd 0.08 / 0.08; ChF 0.37 / 0.38; ChG 0.18 / 0.19; ChL 0.62 / 0.64 mm. Leg measurements are given in Table 1.

MALE: Small sized spider. Carapace greenish - light brown, with smooth surface and distinct fovea. AME, PLE and PME closely grouped; AME separated. Sternum, labium, gnathocoxae and chelicerae light brown. Sternum dark brown laterally, with thin, long hair near margins. Cheliceral groove with four teeth: retromargin with two tiny teeth; promargin with two strong teeth, largest tooth closest to base of the cheliceral groove (Fig. 2). Top of the labium and gnathocoxae with short, strong hair, sparsely distributed. Abdomen greyish-light brown, with short, thin blackish hair over the entire surface. Legs yellowish-light brown with sparse blackish setae. Ventral surface of coxae with long, thin, black sparse hair. Leg IV > Leg I > Leg II > Leg III. Tarsi with three claws. Tarsi III and IV with fine scopulae. Legs III and IV with fine metatarsal scopulae covering slightly less than the distal half of the segment (ventral surface only). Prolateral part of coxae III and IV with 1 spine. Dorsal parts of femora, tibiae and metatarsi with spines. Number of spines can vary among individuals. Detailed leg spination of *H. arnedoi* sp. n. is given in Table 2.

Palpal tarsus of the male covered with thin and elongated setae. Tegulum yellow, lighter than the legs. Bulb almost spherical. The anteroventral region of the bulb has a chitinized edge. Between bulb and distal appendages there is a neck-shaped transition region (Figs 3–5). This region is lightly chitinized and dusky in patches. From the base of the embolus, conductor and accessory apophysis have dark brown tips. Embolus is slender and cylindrical up to its hook-like tip and it is not so heavily sclerotized. At the periphery it is membranous and projects downwards, parallel to the palpal tibia. Conductor and accessory apophysis are developed at the end of the neck-shaped transition region, on the opposite tips of an ear-shaped structure. Both are triangular in shape, whereas the accessory apophysis is longer and jagged (Fig. 6). The relative positions of the distal appendages are most easily seen at a 90 degree angle in ventral view. Embolus separated from conductor, but later bends with a sharp curve and accompanies the accessory apophysis.

FEMALE: No differences found between male and female, in terms of body colour and morphology. Vulva sclerotized almost uniformly. Distal crest long and butt-ended at the tip. Distal expansion of spermatheca convoluted. Rod-shaped part of the anterior spermatheca short and cylindrical. Basal transverse part of the anterior spermatheca separates from rod-shaped part laterally at an acute angle and forms a triangular shape. Anterior basal arc lies linearly through the centre and periphery, but widens at the edges. Transverse bar short and crescent-shaped with tips turned downwards. Posterior diverticulum shaped as a membranous sac and is sclerotized at the periphery (Figs 7, 8).

Ecology. Specimens were collected in Kuşçubeli Pass, located in the Amanos Mountains, from habitats covered by Turkish Pine (*Pinus brutia*) and scrub type oak forests (*Quercus infectoria*). A variety of herbaceous plants and low shrubs such as *Ruscus aculeatus* are also widely represented in those forests. Sampling was done by sifting leaf litter during the early winter.

Distribution. H. arnedoi sp. n. is known from the type locality only.

(Holotype $\stackrel{?}{\circ}$ / Paratype $\stackrel{\bigcirc}{+}$)	Fe	Pa	Ti	Me	Ta
Leg I	1.42 / 1.23	0.80 / 0.73	1.15 / 0.98	1.05 / 0.78	0.38 / 0.32
Leg II	1.10 / 1.17	0.80 / 0.75	1.12 / 0.93	1.05 / 0.85	0.37 / 0.31
Leg III	1.05 / 1.00	0.55 / 0.55	0.83 / 0.63	1.02 / 0.95	0.37 / 0.32
Leg IV	1.63 / 1.50	0.83 / 0.78	1.40 / 1.25	1.50 / 1.35	0.40 / 0.33

Table 1. Leg measurements of *H. arnedoi* sp. n.



Figures 7–8. Vulva of *Harpactea arnedoi* sp. n. 7 dorsal view 8 ventral view. Scale lines: 0.1 mm.

් (Holotype)	Leg I	Leg II	Leg III	Leg IV
С	0	0	1 Pl	1 Pl
Fe	2 Pl	1-2 Pl	1, 1 Pl 1, 1, 1 D	2 Pl 1 D
Pa	0	0	1 RI	0
Ti	0	0	1, 1 Pl 2, 1, 1 Rl 1, 1, 2 V	1, 1 Pl 1 D 2, 1, 1 Rl 1, 1, 2 V
Me	0	0	1, 1 Pl 1, 1, 1 Rl 1, 1, 2 V	1, 1, 1 Pl 1, 1, 1 D 4 Rl 1, 1, 2 V
$\stackrel{\bigcirc}{_+}$ (Paratype)				
С	0	0	1 Pl	1 Pl
Fe	2 Pl	1 Pl	1 Pl 1, 1 D	2 Pl 2 D
Pa	0	0	1 RI	0
Ti	0	0	1, 1 Pl 2, 1, 1 Rl 1, 1, 2 V	1, 1, 1 Pl 1 D 2, 1, 1 Rl 1, 1, 2 V
Me	0	0	1, 1, 1 Pl 1, 1 Rl 1, 1, 2 V	1, 1, 2 Pl 1, 1 D 4 Rl 1, 1, 2 V

Table 2. Leg spination of *H. arnedoi* sp. n.

Harpactea kencei sp. n.

urn:lsid:zoobank.org:act:557FE296-4BF3-4E21-A8D6-4750C38795E6 http://species-id.net/wiki/Harpactea_kencei Figs 9–15

Material examined. Holotype \Im (AUZM), Turkey, Muğla Province, Milas District, Kıyıkışlacık Village [37°16'38.80"N; 27°33'47.97"E], 13.XI.2010, under stones, leg. M.Elverici. **Paratypes** 1 \Im (AUZM), 1 \Im 5 \Im \Im (cKBK & SMF), same data as holotype.

Derivatio nominis. The new species is named in honor of the well-known Turkish biologist Prof. Dr. Aykut Kence (Middle East Technical University, Ankara, Turkey) for his important contributions to Turkish biology.

Diagnosis. *H. kencei* sp. n. can be easily distinguished from all other Turkish *Harpactea* by the unique structure of the male palp and broad, grooved sides, crescent like anterior basal arc-shaped structure of the female vulva. In general appearance the embolus is well sclerotized, thorn-like and with a hook-shaped tip. Towards the tip, the embolus flattens like a spoon and ends with a thorn-like tip inflecting upon itself. On the embolar base there is a small ear-shaped conductor relatively well sclerotized at its peripheries. This is attached to the embolus at an angle of 90 degrees. Unique embolus (not a simple thornlike structure) and the presence of the conductor differentiate *H. kencei* sp. n. from *H. diraoi*, *H. isaurica*, *H. sanctaeinsulae* and any other similar Turkish species, which all have simple spiniform embolus structures (see Brignoli 1978b).

Description. Measurements: [Holotype \Diamond / Paratype Q]: AL 2.13 / 2.25; CL 1.63 / 1.96; CWmax 1.35 / 1.53; CWmin 0.63 / 0.75; AMEd 0.07 / 0.08; PLEd 0.05 / 0.07; PMEd 0.05 / 0.06; ChF 0.34 / 0.37; ChG 0.21 / 0.27; ChL 0.64 / 0.71 mm. Leg measurements are given in Table 3.

MALE: Small to medium sized spider. Carapace reddish dark brown, with smooth surface and distinct fovea. AME, PLE and PME closely grouped; AME separated. Difference between width of eyes region and thoracic region of cara-



Figures 9–13. *Harpactea kencei* sp. n. 9 Cheliceral teeth 10 left palp, retrolateral view 11 ditto, nearly anterior view 12 ditto, prolateral view 13 ditto, distal view. Scale line: 0.5 mm.

pace remarkable. Sternum, labium, gnathocoxae and chelicerae reddish-brown. Sternum with long, thin hair near the margin, while centrally smooth and shiny. Cheliceral groove with four teeth: retromargin bears a tiny tooth at the base of the groove and a strong tooth at the top; promargin with two strong teeth of almost equal sizes (Fig. 9). Labium with short, strong hair, sparsely distributed along the surface; hair cover denser at the top. Gnathocoxae also with moderately strong hair, denser and slightly longer at the margins. Abdomen yellowish-light brown, with short, thin blackish hair over the entire surface. Legs yellowish-light brown with sparse blackish setae. Anterior legs slightly darker than posterior legs. Leg IV > Leg I > Leg II > Leg III. Tarsi with three claws. Tarsi III and IV with fine scopulae. Legs III and IV with fine metatarsal scopulae covering slightly less than the distal half of the segment (ventral surface only). Prolateral part of coxae III and IV with 0-5 spines. Detailed leg spination of *Harpactea kencei* sp. n. is given in Table 4.

Tegulum yellowish brown, longer than wide, cylindrical. Embolar base broad, embolus and conductor reciprocally located at peripherals. Accessory apophysis absent. Embolus blackish brown, flattens like a spoon through the tip and ends with a thorn-

(Holotype $\stackrel{\wedge}{\bigcirc}$ / Paratype $\stackrel{\bigcirc}{+}$)	Fe	Pa	Ti	Me	Ta
Leg I	1.57 / 1.61	0.95 / 0.96	1.28 / 1.29	1.25 / 1.27	0.48 / 0.48
Leg II	1.48 / 1.50	0.80 / 0.82	1.23 / 1.25	1.20 / 1.22	0.40 / 0.40
Leg III	1.13 / 1.17	0.56 / 0.63	0.80 / 0.88	0.90 / 1.13	0.40 / 0.40
Leg IV	1.62 / 1.63	0.82 / 0.87	1.45 / 1.57	1.52 / 1.75	0.51 / 0.53

Table 3. Leg measurements of Harpactea kencei sp. n.



Figures 14–15. Vulva of Harpactea kencei sp. n. 14 dorsal view 15 ventral view. Scale lines: 0.1 mm.

් (Holotype)	Leg I	Leg II	Leg III	Leg IV
С	0	0	1 Pl	5 Pl
Fe	2, 1 Pl	1, 1, 1 Pl	1, 1 D 1, 1 Rl	3, 1, 1 D
Pa	0	0	1 RI	0
Ti	0	0	1, 2 Pl 2, 1, 2 Rl 1, 1, 2 V	2, 1, 1 Pl 2, 1, 1 Rl l 1, 1, 2 V
Me	0	0	1, 1, 1 Pl 5 Rl 2 V	1, 1, 1 Pl 1, 1, 1 Rl 3, 2 V
$\stackrel{\bigcirc}{_{+}}$ (Paratype)				
С	0	0	0	1 Pl
Fe	2 Pl	2 Pl	1, 1 D 1, 1 Rl	2, 1, 1 D
Pa	0	0	1 RI	0
Ti	0	0	1, 1 Pl 2, 1, 1 Rl 1, 1, 2 V	2, 1, 1 Pl 2, 1, 1 Rl 1, 1, 2 V
Me	0	0	1, 1 Pl 6 Rl 2, 2 V	1, 1, 1 Pl 1, 1, 1 Rl 2, 1, 2 V

Table 4. Leg spination of Harpactea kencei sp. n.

like tip inflecting upon itself. Conductor small, flattened and ear-shaped, separated from embolus at the base.

FEMALE: No differences found between male and female, in terms of body colour and morphology. Distal crest of vulva short, with conical tip. Distal expansion of spermatheca spherical, with triangular shape. Basal transverse part of the anterior spermatheca short, linear and peripherally sclerotized. Rod-shaped part of the anterior spermatheca elongated. Anterior basal arc crescent-like and grooved. Tips membranous, in the shape of half of a heart. Transverse bar smooth, well sclerotized at the center. Posterior diverticulum membranous (Figs 14–15).

Ecology. Specimens were collected in the course of field studies aiming to determine the spider fauna of olive groves found in Kıyıkışlacık Village. The area was surveyed periodically over one year using pitfall traps, sweep nets, active searching at ground level and under stones both during the day and at night, sifting tree litter and by shaking tree branches. The altitude of the study area varied from sea level up to 100 m at its maximum.

Individuals of the new species were found under stones and collected from pitfall traps in olive groves and shrub forest associations dominated by *Quercus* trees. The first adult specimen was collected in November and additional specimens were found during March and April. It is likely that this species is most active during the early spring.

Distribution. *H. kencei* sp. n. is known from the type locality only.

Comments. According to the classification established by Deeleman-Reinhold (1993), *H. arnedoi* sp. n. and *H. kencei* sp. n. can be placed within the species group *rubicunda* (D) because they possess the following characteristics:

- 1. Posterior diverticulum of the female vulva membranous in both species.
- 2. Embolus of *H. arnedoi* sp. n. is spiniform, while it is massive-spiniform and flattened in *H. kencei* sp. n.
- 3. Conductor massive in both species.
- 4. Patellae and coxae with spines in both species.

Discussion

The genus *Harpactea* is represented in Turkey by 20 described species. Of these, *H. agnolettii*, *H. colchidis*, *H. galatica*, *H. lazonum*, *H. lyciae*, *H. medeae*, *H. pisidica*, *H. sbordonii* and *H. vignai* were described on the basis of females only; while *H. christodeltshevi*, *H. erseni* and *H. korgei* were described on the basis of males. Due to the inadequacy of the existing descriptions, and the unavailability of many of the type specimens for further investigations, it is currently not possible to draw accurate conclusions about the status of the genus *Harpactea* in Turkey. Nevertheless, the relevant literature shows that most of the recorded species belong in the species group *rubicunda* (D) and *Harpactea* species belonging to this group have often been described from Thrace, Aegean and the Mediterranean coast (see Brignoli 1978a-b, 1979; Lazarov and Deltshev 2008; Lazarov 2010).

With the descriptions of *H. arnedoi* sp. n. and *H. kencei* sp. n. presented herein, the total number of Harpactea species reported from Turkey is now 22. With the exception of H. babori, H. mithridatis and H. sturanyi, all other species are only known only from their type localities or proximate vicinities. H. babori, with the type locality in Büyükada (Insel Prinkipo), has a distribution pattern ranging from Belgrad Forest (Silva Belgradensis) which is one of the last remaining relict forests in İstanbul, and lies along the Istranca Mountain range (both Bulgarian and Turkish sides). As the predominant harpacteid spider species of the Istranca Mountains, H. babori has been recorded from Shumen Town in Bulgaria as the northernmost record and from Stara Planina as the westernmost record of its known distribution range. H. mithridatis has been recorded from Ordu province (East Black Sea Region) in Turkey and from Adzharia region (Khulo) in SW Georgia; while *H. sturanyi*, in addition to its type locality in Konya (Central Anatolia), has also been recorded from the Aegean coast of Turkey (Denizli, İzmir and Muğla provinces; senior author, pers. obs.) and from the Bulgarian side of the Istranca Mountain (see Drensky 1936, 1938). What we would like to emphasize here is the fact that the native Harpactea species, which were also recorded from other countries, are principally distributed through zones with similar ecological conditions. Moreover, it is a known fact that other than some exceptional species like H. hombergi, H. lepida and H. rubicunda, species in the genus Harpactea show narrow distribution patterns in general. Furthermore, the geological, floristic and climatic conditions of Turkey appear to have been conducive to creating opportunities for isolation and diversification of species within the genus Harpactea. With our ongoing research on the spider fauna of Turkey, we expect there will be many more additions to the genus in the near future.

Chatzaki and Arnedo (2006) expressed the taxonomic situation of the genus *Harpactea* in their remarkable revision on the epigean harpacteid spiders of Crete with such words: "*Harpactea* is one of the most ill-defined genera in the whole family and a major taxonomic revision of this genus is urgently called for". We agree with this and believe that the *Harpactea* fauna of Turkey will play a key role in any future revision of the genus because Anatolia constitutes a transition region between Europe, the Balkans, and the Caucasus and the Middle East.

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References

- Alicata P (1974) Su una nuova specie di *Harpactea* (Araneae, Dysderidae) della Turchia. Animalia 1: 69–72.
- Bayram A, Kunt KB, Yağmur EA (2009) A new spider species of the genus *Harpactea* (Araneae; Dysderidae) from southeast Anatolian region of Turkey. Turkish Journal of Arachnology 2 (2): 1–4.
- Bayram A, Kunt KB, Danışman T (2011) The checklist of the spiders of Turkey, Version 10.1.0. Ankara: Turkish Arachnological Society. http://www.spidersofturkey.com
- Brignoli PM (1978a) Ragni di Turchia IV. Leptonetidae, Dysderidae ed Agelenidae nuovi o interessanti di grotte della Turchia meridionale (Araneae). Quaderni del Museo di Speleologia 3: 37–54.
- Brignoli PM (1978b) Ragni di Turchia V. Specie nuove o interessanti, cavernicole ed epigee, di varie famiglie (Araneae). Revue Suisse de Zooogie 85: 461–541.
- Brignoli PM (1979) Spiders from Turkey, VI. Four new species from the coast of the Black Sea (Araneae). Bulletin of the British Arachnological Society 4: 310–313.
- Chatzaki M, Arnedo M (2006) Taxonomic revision of the epigean representatives of the spider subfamily Harpacteinae (Araneae: Dysderidae) on the island of Crete. Zootaxa 1169: 1–32.
- Deeleman-Reinhold CL (1993) The genus *Rhode* and the harpacteine genera *Stalagtia*, *Folkia*, *Minotauria*, and *Kaemis* (Araneae, Dysderidae) of Yugoslavia nad Crete, with remark on the genus *Harpactea*. Revue Arachnologique 10 (6): 105–135.
- Drensky P (1936) Katalog der echten Spinnen (Araneae) der Balkanhalbinsel. Opis na Paiatzite ot Balkanikia polouostrow. Spisanie na Bulgarskata Akademiia na Naukite 32: 1–223.
- Drensky P (1938) Faounata na paiatzite (Araneae) w Beulgaria. II. Podrazred Arachnomorphae; I klon Tetrastica; semeystwa Filistatidae; Dysderidae i Oonopidae. Bulletin des Institutions Royales D'histoire Naturelle 11: 81–106.
- Gasparo F (1999) Descrizione di *Harpactea zoiai* n. sp. del Peloponneso meridionale (Araneae, Dysderidae). Bollettino Museo Regionale di Scienze Naturali Torino 16: 303–308.

- Kunt KB, Yağmur, EA, Kürşat, A (2009) A new record for spider fauna of Turkey, Stalagtia thaleriana Chatzaki & Arnedo, 2006 (Araneae; Dysderidae). Biological Diversity and conservation 2: 45–49.
- Kunt KB, Özkütük RS, Kaya RS (2010) A new species of *Harpactea* (Araneae, Dysderidae) from Aegean region of Turkey. ZooKeys 59: 39–45. doi: 10.3897/zookeys.59.483
- Lazarov S, Deltshev, C (2008) New data for *Harpactea sanctaeinsulae* Brignoli, 1978 and description of the unknown female (Araneae, Dysderidae). Turkish Journal of Arachnology 1: 67–69.
- Lazarov S (2010) New faunistic data for *Harpactea babori* (Nosek, 1905) and description of the unknown female (Araneae, Dysderidae). Turkish Journal of Arachnology 3: 9–12.
- Nosek A (1905) Araneiden, Opilionen und Chernetiden. In: Penther A, Zederbauer E. Ergebnisse einer naturwissenschaftlichen Reise zum Erdschias-Dagh (Kleinasien). Annales des naturhistorischer Museum, Wien 20: 114–154.