# First records of Gastrotricha from South Africa, with description of a new species of Halichaetonotus (Chaetonotida, Chaetonotidae) 

M. Antonio Todaro ${ }^{1, \dagger}$, Matteo Dal Zotto ${ }^{1, \ddagger}$, Sarah J. Bownes ${ }^{2,8,}$, Renzo Perissinotto ${ }^{2,1}$<br>I Dipartimento di Biologia, Università di Modena e Reggio Emilia, via Campi, 231/D, I-41125 Modena, Italy 2 School of Biological \& Conservation Sciences, University of KwaZulu-Natal, Westville Campus, Private Bag X54001, Durban 4000, South Africa<br>$\dagger$ urn:lsid:zoobank.org:author:1F7357F2-5A2D-4914-9DAD-145669A8536A<br>$\ddagger$ urn:lsid:zoobank.org:author:5A61631A-A41D-4685-8B8E-35EB1CB4ABE1<br>§ urn:lsid:zoobank.org:author:0F25F311-06C0-469B-861E-64C11B649B34<br>| urn:lsid:zoobank.org:author:641CADAE-2E9B-4449-8590-8238C1265598<br>Corresponding author: M. Antonio Todaro (antonio.todaro@unimore.it)

Academic editor: R. Hochberg | Received 8 September 2011 | Accepted 5 October 2011 | Published 31 October 2011
urn:lsid:zoobank.org:pub:4B5BC54C-0CE0-4490-8E38-7844A61A3740
Citation: Todaro MA, Dal Zotto M, Bownes SJ, Perissinotto R (2011) First records of Gastrotricha from South Africa, with description of a new species of Halichaetonotus (Chaetonotida, Chaetonotidae). ZooKeys 142: 1-13. doi: 10.3897/ zookeys.142.2036


#### Abstract

During a survey of the biota of the St. Lucia Estuary in the iSimangaliso Wetland Park, South Africa, a number of Gastrotricha were found among samples of meiofauna. Fresh, marine sediment yielded several specimens belonging to a total of seven species. Of these, two are already known from other regions (i.e., Dactylopodola australiensis and Heteroxenotrichula squamosa), one is described as new to science (Halichaetonotus sanctaeluciae sp. n.), while the remaining four (Pseudostomella sp., Halichaetonotus sp.1, Halichaetonotus sp. 2, Xenotrichula sp.) require further collections and analysis, in order to establish the extent of their affiliation to species already described. General appearance, shape of hydrofoil scale and the occurrence of three long spines on the dorsal side make the new species most closely related to $H$. australis and H. marivagus. The key differences from these taxa and between Halichaetonotus sanctaeluciae sp. n. and $H$. aculifer are discussed.


## Keywords

Gastrotricha, meiofauna, new species, South Africa, St. Lucia Estuary, iSimangaliso Wetland Park

[^0]
## Introduction

Marine Gastrotricha from Africa are poorly known. Apart from the pioneering research carried out in Algeria by d'Hondt (1974) and in Somalia by Valbonesi and Luporini (1984, 1987), only the study of Hummon (2011) has been undertaken recently in that region. The latter work includes records of marine gastrotrichs from the Mediterranean and Red Sea coasts of Egypt. In addition, scattered records of species can be found for Tunisia (e.g., Westheide 1972; Todaro et al. 2011) and Kenya (see Balsamo et al. 1992, p. 496). Macrodasys africanus, described long ago from Namibia, is the only gastrotrich species known from southern Africa (Remane 1950).

This study was initiated after several gastrotrich specimens were recovered from formalin-fixed samples collected during an ongoing investigation of the biodiversity and ecology of meiofauna inhabiting the St Lucia Estuary (KwaZulu-Natal, South Africa). As identification to species of soft-bodied meiofaunal taxa, like Gastrotricha, is best achieved when the taxonomic characteristics are observed on fresh specimens, the collection of a series of samples was undertaken for a dedicated in vivo analysis.

It should be highlighted that a census of the biota populating this area bears special relevance, as the St Lucia Estuary is the largest estuarine lake in Africa, a Ramsar Wetland of International Importance and a crucial ecosystem within the iSimangaliso (formerly Greater St Lucia) Wetland Park, South Africa’s first UNESCO World Heritage Site (Taylor 2006).

## Methods

All samples containing gastrotrichs were collected from St Lucia beach ( $28^{\circ} 23^{\prime} \mathrm{S} ; 32^{\circ}$ $25^{\prime} \mathrm{E}$ ), on the ocean side of the sand berm currently closing the estuary. Sediment samples were collected subtidally (about 40 cm water depth) over a neap tide and spring low tide on 22 February and 7 October 2010, respectively. On each occasion, several samples were collected by gently shovelling 600 ml plastic jars through the top 5-10 cm of sediment until full. Within 1-2 days of collection, samples were sent to Modena (Italy) via courier service. In the lab, gastrotrichs were extracted from the sediment by the narcotization-decantation technique, using an isosmotic (7\%) magnesium chloride solution; the fauna-containing supernatant was then poured directly into a 5 cm diameter Petri dish and scanned for specimens under a Wild M8 dissecting microscope, set at 50x magnification (Todaro and Hummon 2008). For taxonomic surveying, the gastrotrichs were removed with a micropipette from the Petri dish, fresh-mounted on slides and observed using a Nikon Eclipse 90i microscope, equipped with Differential Interference Contrast optics (Nomarski) and a DS-5M Nikon digital camera. During the observation, animals were measured with the Nikon ACT-2U software. The description of the new species follows the convention of Hummon et al. (1992, 1993), whereas the position of morphological characters along the body are given in percentage units $(\mathrm{U})$ of total body length measured from anterior to posterior end.

Granulometric analysis of the substrata was carried out according to Todaro et al. (2006). Mean grain size, sorting coefficient, kurtosis, and skewness were calculated by a computerized program based on the equation of Seward-Thompson and Hail (1973).

Abbreviations are as follows: TL, total body length; PhL, pharynx length; FuL, furca length; PhIJ, pharyngeointestinal junction; TbA, adhesive tubes of the anterior series; TbL , adhesive tubes of the lateral series; TbP , adhesive tubes of the posterior series.

## Results

Altogether, 24 samples collected on four different occasions were analysed. Gastrotrichs were only found in sandy material collected in February and October 2010 (Table 1). In total, seven species belonging to five genera and four families representing both orders Macrodasyida ( 2 spp ) and Chaetonotida ( 5 spp ) were found. Two species were identified as known taxa (Dactylopodola australiensis and Heteroxenotrichula squamosa), one is described here as new to science (Halichaetonotus sanctaeluciae sp. n.), while for the remaining four taxa the data gathered so far are not sufficient to exclude their affiliation to species already described. They have, therefore, been provisionally named as follows: Pseudostomella sp., Halichaetonotus sp. 1; Halichaetonotus sp. 2 and Xenotrichula sp. (Table 1).

## Taxonomic account

Order Macrodasyida Remane, 1925 [Rao \& Clausen, 1970]
Family Dactylopodolidae Strand, 1929
Genus Dactylopodola Strand, 1929
Dactylopodola australiensis Hochberg, 2003
http://species-id.net/wiki/Dactylopodola_australiensis
Fig. 1A
Material. 1 specimen, South Africa, KwaZulu-Natal, St Lucia beach, 7 October 2010, SJ Bownes legit.

Morphometry. TL, $319 \mu \mathrm{~m}$; PhL, $92 \mu \mathrm{~m}$; PhIJ at U26; TbA, 4 per side; TbL, $1+1+1+2$ per side; TbP, 5 per side; Ocellar granules absent.

Remarks. the single specimen found is a young adult at the male phase. Among the nine species of Dactylopodola described so far (Hummon and Todaro 2010), the body shape of the specimen from South Africa most resembles D. australiensis Hochberg, 2003, D. indica (Rao \& Ganapati, 1968), D. mesotyphle Hummon, Todaro, Tongiorgi \& Balsamo, 1998 and D. typhle (Remane, 1927). By virtue of its body size (considering the age), number and arrangement of the adhesive tubes our specimen

Table I. Gastrotrich taxa found at St. Lucia beach and granulometric characteristics of the microhabitat at the time of sampling during 2010.

| Taxon | Sampling date |  |
| :--- | :---: | :---: |
|  | 22 February | 7 October |
| Dactylopodola australiensis | - | + |
| Pseudostomella sp. | + | - |
| Halichaetonotus sanctaeluciae sp. n. | + | + |
| Halichaetonotus sp. 1 | + | - |
| Halichaetonotus sp. 2 | + | - |
| Heteroxenotrichula squamosa | + | - |
| Xenotrichula sp. | + | - |
| Granulometric parameters |  | 1.95 |
| Mean particle size (phi) | 0.80 | 0.77 |
| Sorting (phi) | 3.58 | 2.55 |
| Kurtosis | -0.99 | -0.10 |
| Skewness |  | + |

+, species present; -, species absent.


Figure I. Gastrotricha from St. Lucia beach, South Africa DIC photomicrographs. A-B Macrodasyida, C-F Chaetonotida. A, Dactylopodola australiensis, ventral view B, Pseudostomella sp., twisted juvenile C Halichaetonotus sp.1, dorsal view D Halichaetonotus sp. 2, ventral view E Heteroxenotrichula squamosa, ventral view $\mathbf{F}$ Xenotrichula sp., dorsal view.
best approaches the morphometric traits of $D$. indica and, especially, of $D$. australiensis. While $D$. indica is reported (Rao and Ganapati 1968) to have only 2 TbA and 4 TbP per side (vs 4 and 5, respectively), $D$. australiensis seems to differ from the South African specimen solely in the length of the pharynx ( $131 \mathrm{vs} 92 \mu \mathrm{~m}$ ) and in the position of the pharyngeo-intestinal junction (U34-U35 vs U26) (see Hochberg 2003); it is possible that dissimilarities are due to the early age of the African specimen.

## Family Thaumastodermatidae Remane, 1927

Subfamily Thaumastodermatinae Remane, 1927
Genus Pseudostomella Swedmark, 1956

## Pseudostomella sp.

Fig. 1B

Material. 1 juvenile specimen, South Africa, KwaZulu-Natal, St Lucia beach, 22 February 2010, NAF Miranda legit.

Morphometry. TL, $208.6 \mu \mathrm{~m}$; PhL, $92 \mu \mathrm{~m}$; PhIJ at U26; oral palps, $36.4 \mu \mathrm{~m}$ in length, showing 5 dorsal and 6 ventral papillae; cuticular covering made up of relatively large tetrancres; $\mathrm{TbA}, 3$ per side; $\mathrm{TbL}, 5$ per side; $\mathrm{TbP}, 4$ per side, 3 at the end of the caudal pedicle and 1 near its base (inner side).

Remarks. The anatomical traits of the animal from South Africa do not seem to match those of any other known species of Pseudostomella. However, specimen was a juvenile (i.e., not reproductively mature) and so could not be fully described. Should an adult be found in the future, useful comparisons could be restricted to species bearing a cuticular covering made up of tetrancres and relatively long caudal pedicles.

Order Chaetonotida Remane, 1925 [Rao \& Clausen, 1970]
Suborder Paucitubulatina d'Hondt, 1971
Family Chaetonotidae Gosse, 1864
Genus Halichaetonotus Remane, 1936
Halichaetonotus sanctaeluciae sp. n.
urn:lsid:zoobank.org:act:61F9C96F-6E24-493A-9CE8-7AC2F06F2CF3
http://species-id.net/wiki/Halichaetonotus_sanctaeluciae
Figs 2-3

Type locality. South Africa, KwaZulu-Natal, St. Lucia beach (Lat. 28²3'S; Lon. $32^{\circ} 25^{\prime} \mathrm{E}$ ); among medium, moderately siliceous grains on a high-energy sandy beach at mid-tide level.

Type specimens. Holotype, the $146.5 \mu \mathrm{~m}$ long adult specimen shown in Figure 3.


Figure 2. Gastrotricha from St. Lucia beach, South Africa. Halichaetonotus sanctaeluciae sp. n., schematic drawings $\mathbf{A}$ dorsal view $\mathbf{B}$ ventral view (locomotor cilia omitted).

Material examined. Four specimens, two adults (including the holotype) plus one subadult collected on 22 February 2010 (NAF Miranda legit) and 1 adult collected on 7 October 2010 (SJ Bownes legit)

Diagnosis. Medium-sized Halichaetonotus (LT to $146.6 \mu \mathrm{~m}$ ), head, neck and trunk well defined; head rounded, lacking hypostomion but with a small cephalion; medium-long furca projecting from the posterior of the trunk. Body enveloped by 15 columns ( 7 dorsal, 2 lateral +2 ventrolateral hydrofoil scales, $2+2$ ventral small


Figure 3. Gastrotricha from St. Lucia beach, South Africa. Halichaetonotus sanctaeluciae sp. n., habitus A dorsal view $\mathbf{B}$ ventral view. DIC photomicrographs.
scales) of alternating keeled scales each with 17-19 scales. Scales, round on head and neck becoming oval to semi-elliptical on the trunk; in general, keel extending beyond the edge of the scales as short spiny processes; on three posterior scales, one median and two lateral, keels forming long and robust spines extending beyond end of trunk. Two small spiny scales on dorsal and several keeled scales on ventral base of furca. Laterally and ventrolaterally, $2+2$ columns of hydrofoil scales of varying length; ventrally, $2+2$ additional columns of smaller scales; locomotory cilia arranged in two longitudinal bands, interciliary ventral field naked except for two pairs of perianal ovoid keeled/spiny scales. Almost circular mouth opening into cylindrical pharynx with 2 teeth, then sack-like intestine and terminal ventral anus. All specimens parthenogenetic, sometimes with single large egg in position dorsal to mid intestine.

Etymology. The specific name alludes to the geographic locality where the new species has been found.

Description. The description is mainly based on an adult specimen, $146.5 \mu \mathrm{~m}$ in total length. Head rounded, slightly elongated along anterior/posterior axis, bearing a shallow cephalion but no pleural lobes or visible hypostomion; neck narrower than head, trunk sac-like, terminating in a furcate caudum. Body widths at the head/neck/ trunk/caudum are $31 / 22.5 / 34.5 / 22 \mu \mathrm{~m}$, at $\mathrm{U} 11 / 27 / 58 / 81$, respectively. Caudum of medium length ( $26.6 \mu \mathrm{~m}$ ), paired laterally divergent adhesive tubes ( $20 \mu \mathrm{~m}$ in length) with a slightly swollen base $(6.5 \mu \mathrm{~m})$, covered by scales.

Cuticular armature. Head, neck, and trunk covered dorsally and lateroventrally by alternating columns ( 7 dorsal, $1+1$ lateral and $1+1$ ventrolateral hydrofoil, $2+2$ ventral) of 17-19 keeled scales, barely overlapping. On dorsal side, head and neck scales are round (3-5 $\mu \mathrm{m}$ in diameter), while trunk scales are oval to semi-elliptical ( $9.5 \times 5.5-12.7 \times$ $6.6 \mu \mathrm{~m}$ ). In general, keel on dorsal scales extends beyond the edge of scales as short spiny process; however, on two lateral and one median trunk scales, at U63 and U71, respectively, keels form robust, very long spines projecting $26 \mu \mathrm{~m}$ beyond scales. On posterior trunk region are two oval, double keeled scales $(5 \times 4 \mu \mathrm{~m})$ each anchoring a sensorial bristle at U79.5 and a couple of oval spiny scales ( $4 \times 3.5 \mu \mathrm{~m}$ ) bearing spines ( $4-5 \mu \mathrm{~m}$ long) protruding into the furcal indentation. Lateral and ventrolateral spines of hydrofoil scales bearing flattened lamellae, most of which taper into a long hairy process; lamellae bearing spines of the lateral scales are longer than related ventrolateral ones (up to 25 vs up to 19), while lamellae of a column are longest at mid trunk. On ventral side, up to five keeled scales, 3-4 $\mu \mathrm{m}$ long, cover the fleshy portion of each furcal branch; the interciliary field appears naked except for two pairs of oval keeled scales in the perianal region; scales of anterior pair are larger $(9.5 \times 4.5 \mu \mathrm{~m})$ than posterior ones $(6.0 \times 3.5 \mu \mathrm{~m})$.

Ventral ciliation. paired longitudinal bands extending from U03 to approximately U77; each broadly club-shaped anteriorly, but narrowing considerably from the posterior pharyngeal region; bands approach each other immediately behind the mouth, but remain separate throughout their entire length; individual cilia are about $11 \mu \mathrm{~m}$ in length.

Digestive tract: Mouth of medium size (ca. $6 \mu \mathrm{~m}$ in diameter), projecting very slightly ventrally and leading progressively into a $32 \mu \mathrm{~m}$ long pharynx; pharynx muscular, roughly cylindrical ( $8 \mu \mathrm{~m}$ in diameter), showing a bulb anteriorly ( $12 \mu \mathrm{~m}$ in diameter); two cuticular teeth are visible within the bulb; pharynx connected to sacklike intestine at pharyngeo-intestinal junction at U25; intestine straight, narrowing posteriorly, anus ventral at U77.

Reproductive tract. Three specimens were in parthenogenic phase, two of which with a large egg filling much of the trunk.

Taxonomic affinities. Highly variable cuticular armature distinguishes the 30 species of Halichaetonotus described so far (Hummon and Todaro 2010, Hummon 2010). The new species most closely resembles $H$. marivagus, Balsamo, Todaro \& Tongiorgi, 1992, and H. australis Nichols \& Todaro, 2005, in that all three species are characterised by three dorsal spines close to the posterior end of the trunk. Spines are longest in $H$. australis (up to $46 \mu \mathrm{~m}$ ), intermediate in $H$. sanctaeluciae sp. n. (up to $26 \mu \mathrm{~m}$ ) and shortest in $H$. marivagus (up to $15 \mu \mathrm{~m}$ ).

Halichaetonotus marivagus known from the Mediterranean, can easily be distinguished from the new species also on the basis of its wide hypostomion, which is absent in $H$. sanctaeluciae sp. n., and for exhibiting a large cephalion that covers much of the dorsal side of its head (Balsamo et al. 1992).

Halichaetonotus australis described from the east coast of Australia, is unique in that the large median dorsal spine precedes the lateral ones (the opposite is true for $H$. sanctaeluciae sp. n.). Moreover, the keel of the dorsal scales does not extend beyond the edge of the scales (Nicholas and Todaro 2005), whereas in H. santaeluciae sp. n. keels form a spiny process.

The new species also resembles $H$. aculifer (Gerlach, 1953) in terms of size and, most importantly, the shape of the hydrofoil scales. However, the presence of three long spines on the posterior trunk and the absence of ventral interciliary field scales in $H$. sanctaeluciae sp. n. are features that can easily differentiate this species from $H$. aculifer (see Gerlach 1953).

## Halichaetonotus sp. 1

Fig. 1C

Material. 1 adult specimen, South Africa, KwaZulu-Natal, St Lucia beach, 22 February 2010, NAF Miranda legit.

Morphometry. TL, $106.2 \mu \mathrm{~m}$; PhL, $27.2 \mu \mathrm{~m}$; PhIJ at U29.5; FuL, 21 (adhesive tube $17 \mu \mathrm{~m}$ ); dorsal cuticular covering made up of seven columns of 17 overlapping, keeled scales. Scales round (up to $4 \mu \mathrm{~m}$ in diameter) on the head and neck region becoming ovoid (up to $8 \times 4$ ) over trunk. With exception of medial one, scales of posterior-most row bear keel extending into a $14 \mu \mathrm{~m}$ - long spine; $1+1$ columns of hydrofoil scales ventrolaterally.

Remarks. by virtue of the six spiny scales on the trunk rear, the animal appears different from any other species described so far. Unfortunately, a break of the slide occurred during the observation prevented the examination of the ventral side. Without detailed observations of the ventral scales' shape, size and arrangement, we choose to avoid making a formal description of the species until additional specimens are observed.

## Halichaetonotus sp. 2

Fig. 1D

Material. 2 adult specimens and 2 juveniles, South Africa, KwaZulu-Natal, St Lucia beach, 22 February 2010, NAF Miranda legit.

Morphometry. TL, up to $104 \mu \mathrm{~m}$; PhL, up to $25 \mu \mathrm{~m}$; PhIJ at U29; FuL, up to 17.5 (adhesive tube $13 \mu \mathrm{~m}$ ); dorsal cuticular covering made up of nine columns of 17 slightly overlapping, keeled scales. Scales round (up to $2-3 \mu \mathrm{~m}$ in diameter) on head
and neck region becoming elliptical (up to $5 \times 2 \mu \mathrm{~m}$ ) over trunk. Ventrolaterally, $1+1$ columns of hydrofoil scales bearing large lamellae.

Remarks. Morphometry and general appearance of the specimens found at St Lucia match the metric and meristic characteristics of the cosmopolitan Halichaetonotus decipiens (Remane, 1929). Unfortunately, the detritus attached to the ventral side of one of the two adults and the large egg inside the second, prevented the observation of the cuticular details on the ventral side. Consequently, that identification to species cannot be made without reasonable doubts.

## Family Xenotrichulidae Remane, 1927 <br> Genus Heteroxenotrichula Wilke, 1954

## Heteroxenotrichula squamosa Wilke, 1954

http://species-id.net/wiki/Heteroxenotrichula_squamosa Fig. 1E

Material. 1 adult specimen, South Africa, KwaZulu-Natal, St Lucia beach, 22 February 2010, NAF Miranda legit.

Morphometry. TL, $179 \mu \mathrm{~m}$; PhL, $41.5 \mu \mathrm{~m}$; PhIJ at U25.6; FuL, $52.3 \mu \mathrm{~m}$ (adhesive tube $20.5 \mu \mathrm{~m}$ ); Dorsal cuticular covering made up of seven columns of 43-45 overlapping, scales extending laterally as hydrofoil scales. Ten flat scales on the inner margin of each furcal branch. Two pairs of head sensory cirri, 17-28 $\mu \mathrm{m}$ in length, 1 pair of head tentacles, $11.5 \mu \mathrm{~m}$ in length.

Remarks. morphometry and general appearance of the specimen from St. Lucia is in general accordance with data reported for the cosmopolitan H. squamosa; in particular, the South African specimen appears of a size intermediate between individuals of the Mediterranean populations described by Wilke (1954) and Luporini et al. (1973) and specimens described from the Atlantic coast of France by Ruppert (1979).

## Genus Xenotrichula Remane, 1927

## Xenotrichula sp.

Fig. 1F

Material. 1 adult specimen. South Africa, KwaZulu-Natal, St Lucia beach, 22 February 2010, NAF Miranda legit.

Morphometry. TL, $196 \mu \mathrm{~m}$; PhL, $49.5 \mu \mathrm{~m}$; PhIJ at U29; FuL, $27 \mu \mathrm{~m}$ (adhesive tube $10.2 \mu \mathrm{~m}$ ); dorsal cuticular covering made up of 15 columns of 48 pedunculated scales (median column).

Remarks. Morphometry and general appearance of the specimens found at St Lucia fall within the taxonomic range of the cosmopolitan Xenotrichula intermedia

Remane, 1934 and of an as yet unnamed Xenotrichula sp. from Kuwait. The latter two species are siblings, sharing almost identical external morphology but very different organization of the muscular system (see Leasi and Todaro 2009). Technical reasons (several specimens are necessary) prevented the use of confocal laser scanning microscopy to study the organization of the muscular system in the only specimen available. Consequently, identification of the animal found at St. Lucia could not be made with sufficient confidence.

## Conclusion

The first report of gastrotrichs from South Africa shows that, despite the relatively low abundance of taxa retrieved so far, the potential for the discovery of new species unknown to science is remarkable. Soft-bodied organisms such as gastrotrichs must be processed fresh and live, in order to optimise observation of their key taxonomic characteristics. The unavoidable time delay ( $\geq 7$ days) incurred between sample collections and analysis, due to the remote distance between the study site (St Lucia, South Africa) and the laboratory of analysis (Modena, Italy), has probably led to the loss of a number of potentially critical specimens in the samples (e.g. adult stages) and may account for the absence of gastrotrichs from some of the samples. It is, therefore, necessary to consider for the future the option of completing both sampling and analysis on site. This would also allow the collection of sufficient material to complement the traditional morphological studies with more advanced techniques, such as molecular analysis and confocal microscopy. This may prove invaluable towards the resolution of what is currently known in biogeography as the "meiofauna paradox" (Giere 2009). Indeed, it is necessary at this stage to ascertain whether the taxa that could not be conclusively identified to species level are actually already known cosmopolitan species, with broad geographic distribution, or rather new and localized species but "cryptic" in the sense that they cannot be distinguished only on the basis of external morphological characters. This will provide a valuable contribution towards the knowledge of the biodiversity of the St Lucia Estuary and the broader ecosystem of the World Heritage Site of which this wetland is an integral part.

## Acknowledgements

Funding for this project was provided by the National Research Foundation (NRF, Pretoria), the University of KwaZulu-Natal (Durban) and the Ministero Università e Ricerca (Rome) (grant to MAT: PRIN-2007 'Approccio integrato all'identificazione dei Gastrotrichi marini'). We wish to thank the iSimangaliso Wetland Park Authority and the staff and management of EKZN Wildlife, for providing permits and logistic support during the collections. Nelson A.F. Miranda is thanked for helping with field
collections, particularly during February 2010. We are thankful to M. Balsamo and R. Hochberg for their helpful comments of an early draft of this article.

## References

Balsamo M, Todaro MA, Tongiorgi P (1992) Marine gastrotrichs from the Tuscan Archipelago (Tyrrhenian Sea): II. Chaetonotida, with description of three new species. Bollettino di Zoologia 59: 487-498. doi: 10.1080/11250009209386710
Gerlach SA (1953) Gastrotrichen aus dem Kuestengrundwasser des Mittelmeeres. Zoologisher Anzeiger 150: 203-211.
Giere O (2009) Meiobenthology. The Microscopic Motile Fauna of Aquatic Sediments. Springer, Berlin, 526.
d'Hondt JL (1974) Contribution a l'étude de la microfaune interstitielle des plages de l'ouest Algerien. Vie et Milieu 23 (for 1972-73): 227-241.
Hummon WD (2010) Marine Gastrotricha of San Juan Island, Washington, USA, with notes on some species from Oregon and California. Meiofauna Marina 18:11-40.
Hummon WD (2011) Marine Gastrotricha of the Near East: 1. Fourteen new species of Macrodasyida and a redescription of Dactylopodola agadasys Hochberg, 2003. ZooKeys 94: 1-59. doi: 10.3897/zookeys. 94.794
Hummon WD, Balsamo M, Todaro MA (1992) Italian marine Gastrotricha: I, Six new and one redescribed species of Chaetonotida. Bollettino di Zoologia 59: 499-516. doi: 10.1080/11250009209386711

Hummon WD, Todaro MA, Tongiorgi P (1993) Italian marine Gastrotricha: II. One new genus and ten new species of Macrodasyida. Bollettino di Zoologia 60: 109-127. doi: 10.1080/11250009309355798

Hummon WD, Todaro MA (2010) Analytic taxonomy and notes on marine, brackish-water and estuarine Gastrotricha. Zootaxa 2392: 1-32.
Leasi F, Todaro MA (2009) Meiofaunal cryptic species revealed by confocal microscopy: the case of Xenotrichula intermedia (Gastrotricha). Marine Biology 156: 1335-1346. doi: 10.1007/s00227-009-1175-4

Luporini P, Magagnini G, Tongiorgi P (1973) Chaetonotoid gastrotrichs of the Tuscan coast. Bollettino di Zoologia 40: 31-40. doi: 10.1080/11250007309427479
Nicholas W, Todaro MA (2005) Observations on Gastrotricha from a sandy beach in southeastern Australia with a description of Halichaetonotus australis sp. nov. (Gastrotricha: Chaetonotida). New Zealand Journal of Marine and Freshwater Research 39: 973-980. doi: 10.1080/00288330.2005.9517367

Remane A (1950) Macrodasys africanus nov. spec., ein Gastrotrich von der Küste SüdwestAfrikas. Kieler Meeresforschung 7: 35-37.
Ruppert EE (1979) Morphology and systematics of the Xenotrichulidae (Gastrotricha, Chaetonotida). Mikrofauna des Meeresbodens 76: 1-56.

Seward-Thompson BL, Hails JR (1973) An appraisal of the computation of statistical parameters in grain size analysis. Sedimentology 20: 161-169. doi: 10.1111/j.1365-3091.1973. tb01612.x
Taylor RH (2006) Ecological responses to changes in the physical environment of the St Lucia estuary. Unpublished PhD thesis 2006, Aas, Norway: Norwegian University of Life Sciences, 172 pages.
Todaro MA, Hummon WD (2008) An overview and a dichotomous key to genera of the phylum Gastrotricha. Meiofauna Marina 16: 3-20.
Todaro MA, Leasi F, Bizzarri N, Tongiorgi P (2006) Meiofauna densities and gastrotrich community composition in a Mediterranean sea cave. Marine Biology 149: 1079-1091. doi: 10.1007/s00227-006-0299-z

Todaro MA, Kanneby T, Dal Zotto M, Jondelius U (2011) Phylogeny of Thaumastodermatidae (Gastrotricha: Macrodasyida) Inferred from Nuclear and Mitochondrial Sequence Data. PLoS ONE 6: e17892. doi: 10.1371/journal.pone. 0017892
Valbonesi A, Luporini P (1984) Researches on the coast of Somalia. Gastrotricha Macrodasyoidea. Monitore Zoologico Italiano (Supplemento) 1: 1-34.
Valbonesi A, Luporini P (1987) Researches on the coast of Somalia. Gastrotricha Chaetonotoidea. Monitore Zoologico Italiano (Supplemento) 14: 235-261.
Westheide W (1972) Räumliche und zeitliche Differenzierungen im Verteilungsmuster der marinen Interstitialfauna. Verhandlungsbericht der Deutschen Zoologischen Gesellschaft 65: 23-32.
Wilke U (1954) Mediterrane Gastrotrichen. Zoologische Jahrbücher Abteilung Systematik Geographie und Biologie der Tiere (Jena) 82: 497-550.

# Taxonomic contribution to the Aleiodes melanopterus (Erichson) species-group (Hymenoptera, Braconidae, Rogadinae) from Brazil 

Eduardo Mitio Shimboril ${ }^{1, t}$, Angélica Maria Penteado-Dias ${ }^{2, \ddagger}$<br>I Embrapa Agropecuária Oeste, BR 163, km 253.6, Dourados, MS, Brazil 2 Universidade Federal de Säo Carlos, Rodovia Washington Luiz, km 235, CEP 13 565-905, São Carlos, SP, Brazil<br>$\dagger$ urn:lsid:zoobank.org:author:3A171087-3715-4AA2-8731-DB3BEC7DECDA<br>$\ddagger$ urn:lsid:zoobank.org:author:A401BE09-6822-4933-8C09-CB5D8C0CE07E<br>Corresponding author: Eduardo Shimbori (shimbori@gmail.com), Angélica Penteado-Dias (angelica@ufscar.br)

Academic editor: C. van Achterberg | Received 13 June 2011 | Accepted 20 October 2011 | Published 31 October 2011
urn:lsid:zoobank.org:pub:D03235E6-60F1-4841-AD72-F4441FA7F9E4
Citation: Shimbori EM, Penteado-Dias AM (2011) Taxonomic contribution to the Aleiodes melanopterus (Erichson) species-group (Hymenoptera, Braconidae, Rogadinae) from Brazil. ZooKeys 142: 15-25. doi: 10.3897/zookeys.142.1705


#### Abstract

The Aleiodes melanopterus (Erichson, 1848) species-group includes 21 species, of which seven are known from the Neotropical region: A. flavistigma Shaw, 1993, A. lucidus (Szépligeti, 1906), A. melanopterus, A. mexicanus Cresson, 1869, A. politiceps (Gahan, 1917), and the new species $A$. shaworum sp. n. and $A$. vassununga sp. n. Distribution ranges of $A$. melanopterus, A. flavistigma and $A$. lucidus are extended and the female of $A$. lucidus is described. A key to the Neotropical species of this species-group is presented.


## Keywords

Brazil, new species, Eucystomastax, distribution

## Introduction

The Neotropical species of Aleiodes melanopterus (Erichson, 1848) species-group has been treated as several different genera until Shaw (1993) clarified its relationship with others Aleiodes species-groups. The Neotropical lineage was then proposed to be a subgenus within Aleiodes, namely Eucystomastax Brues, 1912, including five nominal spe-

[^1]cies: flavistigma Shaw, 1993, lucidus (Szépligeti, 1906), melanopterus (Erichson, 1848), mexicanus Cresson, 1869 and politiceps (Gahan, 1917) (Yu et al. 2005). In a broad sense the melanopterus group is present in Palaearctic, Nearctic, and Neotropical regions (Marsh and Shaw 1999), and includes 21 species, composing a monophyletic group defined by the large oral space and narrow clypeus (Fortier and Shaw 1999). All Neotropical species plus politiceps (Gahan, 1917) comprises a derived subgroup within the melanopterus group, defined by having pectinate tarsal claws and strongly protruding clypeal carina (Shaw 1993; Fortier and Shaw 1999).

## Material and methods

A portion of examined specimens, deposited at DCBU (Universidade Federal de São Carlos), comes from several different surveys throughout Brazil. Additional specimens were loaned from several entomological collections in Brazil, deposited temporally at DCBU: Museu Paraense Emilio Goeldi (MPEG), Instituto Nacional de Pesquisas da Amazônia (INPA), Museu Nacional do Rio de Janeiro (MNRJ), Museu de Zoologia da Universidade de Sáo Paulo (MZUSP), Coleção Entomológica Padre Jesus S. Moure Departamento de Zoologia da Universidade Federal do Paraná (DZUP). We examined 194 specimens of the Aleiodes melanopterus species-group, all collected in Brazil.

Terminology used mostly follows Shaw (1993), exception made for some microsculpture characters that follows Marsh and Shaw (1999). Abbreviations used in the descriptions follows strictly that used by Shaw (1993), namely: BL= body length, excluding antenna and ovipositor; FWL= fore wing length; F\# = flagellum \#; MS= malar space; $\mathrm{EH}=$ maximum eye height; $\mathrm{EW}=$ maximum eye width; TW= temple width; OS= oral space, maximum width in anterior view; OOD= ocellar-ocular distance (shortest distance from eye margin to lateral ocellus); $\mathrm{OD}=$ ocellus diameter (maximum width of lateral ocellus); T\#= tergum \#; OL= ovipositor length; HBTL= hind basitarsus length; HTS = hind tibial spur length (longest spur); R\#= radius segment \#. Colour pictures were taken by stereomicroscope. Greyscale pictures were taken at SEM in low vacuum.

## Results

We identified all previously known South American species among examined material, i.e. Aleiodes flavistigma Shaw, 1993, Aleiodes lucidus (Szépligeti, 1906), Aleiodes melanopterus (Erichson, 1848); plus two new species described bellow: Aleiodes shaworum sp. n. and Aleiodes vassununga sp. n. One of the specimens examined constitutes the first recorded female of $A$. lucidus, and also the first record of this species from Brazil. The distribution range of A. flavistigma is extended to Minas Gerais State in Brazil; this is the first record of this species outside of Santa Catarina State. Some morphological features of the Brazilian $A$. melanopterus specimens are described and discussed.

Key to Neotropical species of Aleiodes melanopterus (Eucystomastax) speciesgroup (modified from Shaw 1993)

1 Metasoma orange to reddish brown (Fig. 2)............................................... 2

- Metasoma black apically or mostly black (Fig. 1) ......................................... 4

2 First and second metasomal terga striate; body bicoloured, head and legs black; malar space narrow, about $1 / 2$ basal width of mandible $\qquad$ A. mexicanus Cresson

- First and second metasomal terga strongly costate (Fig. 14); body unicoloured (Fig. 2); malar space about equal to basal width of mandible 3

3 Pterostigma yellow (Fig. 2) .........................................A. vassununga sp. n.

- Pterostigma black....................................................... A. politiceps (Gahan)

4 Notauli absent or very shallow anteriorly, mesonotum entirely smooth (Fig. 10); epicnemial carina effaced dorsally or completely absent (Fig. 11)
A. lucidus (Szépligeti)

- Notauli distinct, although smooth (Fig 8); epicnemial carina entirely present (Fig. 12) 5
5 Pterostigma yellow ........................................................A. Alavistigma Shaw
- Pterostigma black. 6
6 Clypeus strongly protruding (Fig. 7); hind coxa smooth dorsally; mesonotum orange, pronotum and fore coxa often orange .....A. melanopterus (Erichson)
- Clypeus not strongly protruding (Fig. 6); hind coxa striated dorsally (Fig. 9); pronotum, mesonotum and fore coxa black (Fig. 1).......A. shaworum sp. n.


## Aleiodes flavistigma Shaw, 1993

http://species-id.net/wiki/Aleiodes_flavistigma

Material examined. Brazil: 3 females, Nova Teutônia, SC, X.1967, F. Plaumann col.; 1 female, Extrema, MG, 25.XII.1990, E. Mariano col.

Distribution. Brazil, Minas Gerais and Santa Catarina States.

## Aleiodes lucidus (Szépligeti, 1906)

http://species-id.net/wiki/Aleiodes_lucidus
Figs 10, 11
Macrostomion lucidus Szépligeti 1906: 609

Material. Female. Fazenda São João, Diamantina, MT, Brazil, 450m, 5.II.1981, Ekis \& Young col.

Description. Female. Body length 12 mm , fore wing length 11.6 mm .


Figures $\mathbf{I}, \mathbf{2}$ (scale line $=4 \mathrm{~mm}$ ). Habitus left: $\mathbf{I}$ Aleiodes shaworum sp. n. $\mathbf{2}$ A. vassununga sp. n.

Head. Flagellum broken at F40, F1 2 times longer than wide, F2 1.8 times longer than wide, flagellomeres beyond second about as long as wide; malar space $3 / 4$ basal width of mandible; MS/EH 0.33; TW/EW 0.67; occipital carina weak ventrally, not meeting the hypostomal carina; oral opening width 1.16 times clypeo-antennal distance; clypeus height 0.3 times its width, protruding and bordered by carina; OS/ MS 2.08; OOD/OD 1.33; face smooth, swollen medially; temple smooth; maxillary palpal segments 2-4 swollen.

Mesosoma. Entirely smooth; notauli very shallow anteriorly, otherwise absent (Fig. 10); epicnemial carina nearly absent (Fig. 11); precoxal sulcus absent; propodeum with median carina complete. Legs: tarsal claws pectinate; hind coxa smooth dorsally. Wings: dusky; R1/R2 0.42; R1/recurrent vein 0.62 ; 1CUa/1cu-a 1.85 ; basella/mediella 0.4.

Metasoma. T1 length/width 1.15; T2 length/width 0.7; T3 length/width 0.58 ; all terga smooth except for medio-longitudinal carina on T1 and T2; OL/HBTL 0.22; HTS/HBTL 0.3.

Colour. Body colour black, except mesothorax orange to infuscate orange.
Distribution. Bolivia, Mapiri and Santa Cruz; Brazil, Mato Grosso.
Comments. This species was known only from male until the present study. The female is very similar to male, but has a larger body size, and differs in some wing vein proportions and the smoother face. The diagnostic characters of the species (e.g. notauli virtually absent, whole body with smooth sculpturing, reduced epicnemial carina) are present in the female specimen examined.

## Aleiodes melanopterus (Erichson, 1848)

http://species-id.net/wiki/Aleiodes_melanopterus
Figs 3, 7
Rogas melanopterus Erichson, 1848: 588
Macrostomion peruvianum Szépligeti, 1904: 193
Rhogas rufithorax Cameron, 1911: 313
Rhogas fortipalpus Cameron, 1911:314
Rhogas forticarinatus Cameron, 1911:314
Eucystomastax bicolor Brues, 1912: 223

Material examined. 74 females and 108 males. Brazil. Acre (AC): Cruzeiro do Sul, Rio Branco; Amazonas (AM): Serra dos Porcos; Espírito Santo (ES): Fundão, Santa Teresa; Goiás (GO): Anápolis, Goiânia - "Campinas", Jataí; Mato Grosso (MT): Aripuanã, Alta Floresta, Barra do Tapirapé, Chapada dos Guimarāes, Itiquira, Jaçanã - P.N. Xingu, Rondonópolis; Mato Grosso do Sul (MS): Campo Grande, Dourados; Minas Gerais (MG): Araxá, Corinto, Passos; Pará (PA): Barreirinha, Belém, Canindé, Parauapebas -


Figures 3-8. $\mathbf{3}$ Aleiodes melanopterus face $\mathbf{4}$ Aleiodes shaworum sp. n. face $\mathbf{5}$ Aleiodes vassununga sp. n. face $\mathbf{6}$ Aleiodes shaworum sp. n. head left $\mathbf{7}$ Aleiodes melanopterus head left arrow at end of occipital carina 8 Aleiodes shaworum sp. n. thorax dorsal.

Serra Norte, Peixe-Boi, Redenção - Gorotire, São Félix do Xingu, Serra Norte; Paraná (PR): Curitiba, Maringá, Ponta Grossa, Rolândia, São José dos Pinhais; Rio de Janeiro (RJ): Santa Maria Madalena; Rio Grande do Sul (RS): Santa Maria, São Leopoldo;

Rondônia (RO): Ouro Preto d'oeste, Porto Velho, Vilhena; Santa Catarina (SC): Nova Teutônia; São Paulo (SP): Barueri, Cananéia - Ilha do Cardoso, Castilho, Caraguatatuba, Juquitiba, Luís Antônio, Monte Alegre, Nova Europa, Onda Verde, Rio Claro, Salesópolis, São Carlos, São Paulo, Tabatinga, Ubatuba, Vargem Grande Paulista.

Morphological notes on Brazilian specimens. Head. Occipital carina of all examined specimens is absent ventrally, thus occipital and hypostomal carina do not meet (Fig. 7). This is probably a misinterpreted character in Shaw (1993) corrected by Fortier and Shaw (1999) (character 20, state 1 for melanopterus); face sculpture variable, smooth to rugose, rugosity concentrated near raised median area when present (Fig. 3).

Colour variation. The examined specimens present distinct colour pattern variation. Propodeum always orange; metasoma black in virtually all specimens ( $97 \%$ ); hind coxa black ( $99 \%$ of specimens); mid coxa somewhat lighter than hind; fore coxa bright orange in $97 \%$ of the specimens, contrasting with remainder black leg; one of the examined specimens has dark pronotum and propleuron.

Distribution. South America, from Suriname to Northern Argentina (North to South), and from South-eastern Brazil to Eastern Peru (East to West). Not recorded in the East of Andean Cordillera, Central America (Shaw 1993), and Northeast Brazilian region. South American countries with records: Argentina, Bolivia, Brazil, Ecuador, Paraguay, Peru and Suriname.

## Aleiodes shaworum sp. n.

urn:Isid:zoobank.org:act:27156FC8-61DA-4E1C-9065-5B68F29B3522
http://species-id.net/wiki/Aleiodes_shaworum
Figs 1, 4, 6, 8, 9, 12, 13, 15

Material. Type locality: Brazil, São Paulo, Juquitiba, Sítio Sonho do Vovô, Atlantic forest.

Type specimens. Holotype, female, pinned. Original label: "Sítio Sonho do Vovô, Juquitiba - SP - 22-IV-1988 - (V) L.A. Joaquim, col." DCBU / UFScar, São Carlos.

Paratypes (DCBU): 1 female, same as holotype; 2 males, Brazil, Barueri, SP, 11.V. 1966 and 22.I.1966; 1 male: Brazil, Estação Florestal, Caraguatatuba, SP, 40 m, VII. 1965.

Diagnosis. This species is similar to $A$. melanopterus. It can be distinguished by colour pattern: mesonotum black, contrasting with orange mesopleuron, metapleuron, and propodeum; smaller oral opening and clypeus not strongly protruding; striated sculpture on dorsal part of hind coxa; fore wing vein R1 relatively shorter.

Description. Female. Body length 9-9.2 mm, fore wing length 8.2 mm .
Head (Fig. 4). Flagellum with 60 flagellomeres, F1 2.5-3 times longer than wide, F2 about twice longer than wide, remaining flagellomeres almost twice as long as wide; malar space $1 / 2$ basal width of mandible; MS/EH 0.2 ; TW/EW 0.62 ; occipital carina absent ventrally, not meeting hypostomal carina (Fig. 6); oral opening width slightly greater than clypeo-antennal distance; clypeus height/width $1 / 3$; clypeus not protrud-


Figures 9-14.9 Aleiodes shaworum sp. n.: part of first metasomal terga and hind coxa dorsal surface arrow at microsculpture striation on hind coxa. I0,II Aleiodes lucidus: $\mathbf{1 0}$ head and mesoscutum dorsal II head and mesonotum left arrow indicating absence of epicnemial carina. $\mathbf{I 2}$ Aleiodes shaworum sp. n. mesopleuron left arrow at epicnemial carina. I3 14 Metasoma dorsal: 13 Aleiodes shaworum sp. n. 14 Aleiodes vassununga sp. n..
ing, without carinate boarder; OS/MS 3.1-3.2; OOD/OD 0.87; face rugo-striate; frons and vertex smooth; temples smooth scattered with punctuations near occipital carina; maxillary palpus swollen, especially segments 2 and 3.

Mesosoma (Fig. 8). Pronotum smooth laterally and middorsally, granular-rugulose anteriorly; propleuron smooth; mesonotum weakly coriaceous; mesoscutum with short median posterior carina; notauli weakly scrobiculate; mesopleuron smooth and shinny, precoxal sulcus absent; epicnemial carina complete; propodeum smooth to weakly coriaceous dorsally, carinate-rugose basally, median carina complete. Legs: tarsal claws pectinate; hind coxa dorsally striate, same sculpture occasionally present on mid and fore coxa (Fig. 9). Wings (Fig. 15): dusky; R1/R2 0.29-0.33; R1/recurrent vein $0.38-0.4 ; 1 \mathrm{CUa} / 1 \mathrm{cu}-\mathrm{a} 1.7-2.5$; basella/mediella $0.41-0.47$.

Metasoma (Fig. 13): T1 length/width 1.19-1.25; T2 length/width 0.73-0.75; T3 length/width $0.48-051$; T1 and T2 weakly rugose, apically smooth, raised triangular smooth area present; reminder metasomal terga smooth; OL/HBTL 0.21; HTS/ HBTL 0.27-0.36.

Colour (Fig. 1): Black; mesopleuron, metapleuron and propodeum bright orange.
Male. Essentially as the female, but with slightly larger eyes; body length 10 mm ; flagellum 59-63F.

Etymology. The species is named in honour to Scott Shaw, for his contribution to the knowledge on this group.

Distribution. Brazil, State of São Paulo, Brazilian Atlantic Forest.
Comments. Despite the superficial resemblance with melanopterus, this species have quite distinctive characters, including the lack of a strongly protruding clypeus, which has been considered one of the synapomorphies for the Neotropical melanopterus species-group clade (Shaw 1993; Fortier and Shaw 1999).

## Aleiodes vassununga sp. n.

urn:lsid:zoobank.org:act:25462065-6F95-4324-9189-9D82350575B1
http://species-id.net/wiki/Aleiodes_vassununga
Figs 2, 5, 15

Material. Type locality: Brazil, São Paulo, Santa Rita do Passa Quatro, Parque Estadual de Vassununga - Mata Praxedes, $21^{\circ} 40^{\prime} 56^{\prime \prime}$ S, $47^{\circ} 37^{\prime} 13^{\prime \prime} \mathrm{W}$, Semi-deciduous Atlantic forest.

Type specimens: Holotype, female, pinned. Original label: "Sta. Rita P. Quatro, SP, Brasil. Pq. Est. De Vassununga - Mata Praxedes - S21º40'56", W47³7'13" 31.III. 2006 - Armadilha Malaise - A.M.P. Dias col." DCBU / UFSCar, São Carlos.

Paratype (DCBU): 1 Male, Brazil, SP, Barueri, 6.XII.1966, K. Lenko col.
Diagnosis. This species resembles the North American species A. politiceps and melanopodus Marsh and Shaw, 1999, both on body colour and the coarse sculpture on the


Figure 15. Aleiodes shaworum sp. n. right wings.
metasoma, but it can be readily distinguished by the bright yellow spot on pterostigmal area of fore wing, as in flavistigma. In the key to North American species it will run to A. melanopterus, from which can be distinguished by the shorter ovipositor and smooth face sculpturing. It is also the only South American species without a black metasoma.

Description. Female (Holotype). Body Length 10.8 mm , fore wing length 8.8 mm .
Head (Fig 5). Flagellum with 68 flagellomeres, F1 width 0.4 times its length, F15 0.8 times wider than long; malar space 0.3 times eye height, 0.8 times basal width of mandible; TW/EW 0.55 ; occipital carina not meeting hypostomal carina; oral opening width 1.26 times clypeo-antennal distance, OS/MS 2.5; OOD/OD 1.15; head entirely smooth; clypeus strongly protruding and margined by carina; maxillary palpus slightly swollen.

Mesosoma. Almost entirely smooth, pronotum sparsely rugose laterally; mesoscutum with short postero-median carina; notauli smooth, with some weak crenulae anteriorly; precoxal sulcus absent; epicnemial carina complete; propodeum smooth dorsally, carinate-rugose basally; median carina complete. Legs: tarsal claws pectinate; hind coxa dorsally smooth. Wings: dusky with yellow spot on pterostigmal area (Fig. 2); R1/R2 0.33; R1/recurrent vein 0.42 ; 1CUa/1cu-a 1.11 ; basella/mediella 0.39 .

Metasoma (Fig. 14). T1 length/width 1.21; T2 length/width 0.76; T3 length/width 0.41 ; T1, T2 and basal half of T3 strongly costate; OL/HBTL 0.25 ; HTS/HBTL 0.25.

Colour (Fig. 2): Body entirely reddish brown, including scapus; antenna, ocelli, eyes, labial palpi, maxillary palpus segments $2-5$ and ovipositor sheaths black; legs darkening apically from the tibia.

Male. Similar to female but face with some transverse rugositie; body length 11 mm ; fore wing length 8.8 mm ; flagellum with 63 flagellomeres.

Etymology. The name of species refers to locality of collection of material for study. Distribution. Brazil, State of São Paulo.

## Acknowledgements

To CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico), FAPESP (Fundação de Amparo à Pesquisa do Estado de São Paulo), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior), INCT HYMPAR SUDESTE (Instituto Nacional de Ciência e Tecnologia dos Hymenoptera Parasitoides da Regiáo Sudeste Brasileira) for financial support; to Instituto Florestal do Estado de Sảo Paulo and managers of Parque Estadual de Vassununga for allowing collection of insects. The manuscript was improved thanks to the helpful comments of Scott Shaw and one anonymous reviewer, and the editor, Kees van Achterberg.

## References

Brues CT (1912) Brazilian Ichneumonidae and Braconidae obtained by the Stanford expedition. Annals of the Entomological Society of America 5: 193-228.
Cameron P (1911) On the Hymenoptera of the Georgetown Museum, British Guiana: Part II. Timehri 1: 306-330.
Erichson WF (1848) Insecten. In: Schomburgh R(Ed) Reisen in Britisch-Guiana in den Jahren 1840-1844: Dritter Theil. Verlagsbuchhandlung von JJ Weber, Leipzig, 553-617.
Fortier JC, Shaw SR (1999) Cladistics of the Aleiodes Lineage of the Subfamily Rogadinae (Hymenoptera: Braconidae). Journal of Hymenoptera Research 8(2): 204-237.
Shaw SR (1993) Systematic status of Eucystomastax Brues and characterization of the Neotropical species (Hymenoptera: Braconidae: Rogadinae). Journal of Hymenoptera Research 2 (1): $1-11$.

Marsh PM, Shaw SR (1999) Revision of the North American Aleiodes Wesmael (Part 5): The melanopterus (Erichson) Species-Group (Hymenoptera: Braconidae, Rogadinae). Journal of Hymenoptera Research 8 (1): 98-108.
Szépligeti GV (1904) Sudamerikanische Braconiden. Annales Historico-Naturales Musei Nationalis Hungarici 2: 173-197.
Szépligeti GV (1906) Braconiden aus der Sammlung des ungarischen National-Museums, 1. Annales Historico-Naturales Musei Nationalis Hungarici 4: 547-618.
Yu D, van Achterberg K, Horstmann K (2005) World Ichneumonoidea 2004: Taxonomy, Biology, Morphology and Distribution. CD. Taxapad, Vancouver, Canada.

# Revision of the new world genus Crassomicrodus Ashmead (Hymenoptera, Braconidae, Agathidinae), with an identification key to species 

José Isaac Figueroa ${ }^{1, \dagger}$, Michael Joseph Sharkey ${ }^{2, \ddagger}$, Jesus Romero Nápoles ${ }^{3, \S}$, José Antonio Sánchez García ${ }^{4,1}$, Ana Mabel Martínez ${ }^{1, \pi}$, Victor López-Martínez ${ }^{5, \#}$, Samuel Pineda ${ }^{3, \dagger \dagger}$


#### Abstract

I Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolás de Hidalgo, Km. 9.5 carretera Morelia-Zinapecuaro, Tarimbaro, Michoacán, 58880, México 2 Department of Entomology, University of Kentucky, S-225 Ag. Science Center North, Lexington, Kentucky 40546-0091, USA 3 Instituto de Fitosanidad, Colegio de Postgraduados, Km. 36.5 Carretera México-Texcoco, Montecillo Edo. de México, 56230, México 4 CIIDIR-IPN-Unidad Oaxaca, Area de Control biológico, Hornos \#1003, Santa Cruz xoxocotlán, Oaxaca, C.P. 71230, México 5 Facultad de Ciencias Agropecuarias, Universidad Autónoma del Estado de Morelos, Av. Universidad 1001, Col. Chamilpa, Cuernavaca, Morelos, C.P. 62210, México


$\dagger$ urn:lsid:zoobank.org:author:655ECBB7-0185-475B-8E63-A05B4ABF305D
$\ddagger$ urn:lsid:zoobank.org:author:77B8EC3A-442C-4A7A-AF85-A31C27E257F2
§ urn:lsid:zoobank.org:author:060B5596-538B-44BE-B5B7-DFFAE6D483DB
| urn:lsid:zoobank.org:author:0FFD7283-CF7A-4583-8645-D934A5F392AD
II urn:lsid:zoobank.org:author:C94E75F1-C242-4DD6-99BF-F0766C887CA7
\# urn:lsid:zoobank.org:author:05FA91F9-17D4-48CF-AFAC-47096CE8E306
$\dagger \dagger$ urn:lsid:zoobank.org:author:FFDFCAF3-2468-4461-85E2-0FFB240FC2E5
Corresponding author: José Isaac Figueroa (figueroaji@yahoo.com.mx)

Academic editor: C. Achterberg \| Received 13 June 2011 | Accepted 7 October 2011 | Published 31 October 2011
urn:lsid:zoobank.org:pub:0C860B63-6279-43FC-92F1-374772A962CD
Citation: Figueroa JI, Sharkey MJ, Nápoles JR, García JAS, Martínez AM, Martínez VL, Pineda S (2011) Revision of the new world genus Crassomicrodus Ashmead (Hymenoptera, Braconidae, Agathidinae), with an identification key to species. ZooKeys 142: 27-75. doi: 10.3897/zookeys.142.1709


#### Abstract

A key to species and descriptions are presented for 14 species of the New World genus Crassomicrodus Ashmead. Seven new species, C. azteca, C. clypealis, C. costaricensis, C. jalisciensis, C. mariae, C. oaxaquensis, and C. olgae are described. C. fenestratus (Viereck) is synonymized with C. nigriceps (Cresson). C. melanopleurus (Ashmead) is recognized as a valid species.


[^2]
## Keywords

insecta, taxonomy, parasitoid wasps, new species, Ichneumonoidea

## Introduction

The agathidine wasp genus Crassomicrodus was erected by Ashmead (1900), with the type species Microdus fulvescens (Cresson, 1865). A few years later Bradley (1916) pointed out that Microdus divisus (Cresson, 1873), the type species of Epimicrodus (Ashmead, 1900), shared many characters with the designated type of Crassomicrodus, and therefore synonymized Epimicrodus Ashmead under Crassomicrodus. Members of Crassomicrodus are characterized by a short ovipositor, tarsal claws without basal lobes, and the lack of pegs near the apex of the lateral surface of the hind tibia. Crassomicrodus is closely related with the genus Agathirsia Westwood (Pucci and Sharkey 2004). The original placement of Crassomicrodus and Agathirsia in the tribe Agathidini was proposed by Sharkey (1992) but recently Sharkey et al. (2006), using both molecular and morphological data sets, proposed the tentative inclusion of these genera in the tribe Earinini.

Prior to this publication Crassomicrodus contained eight species recognized (Muesebeck et al. 1951, Shenefelt 1970, Marsh and Carlson 1979, Figueroa et al. 2008). Previous to 2008 there were nine nominal species but Figueroa et al. (2008) synonymized C. medius Cresson under C. fulvescens. Six species were included in the Hymenoptera of America North of Mexico (Muesebeck 1927). One species was described from Puerto Rico (Viereck 1913), and Marsh (1960) added one species collected in the U.S.A. Other nominal species, such as C. nigricaudos (Viereck 1905) and C. pumilus = Epimicrodus pumilus (Szépligeti 1913, Brues 1926) were included in the genus, but these were misidentified. C. nigricaudos was transfered to Agathirsia by Muesebeck (1927), and Epimicrodus pumilus was transferred to the genus Bassus by Pucci and Sharkey (2004). According to Sharkey (1997) species of Crassomicrodus are restricted to the New World, and can be found from southern of Canada to Costa Rica, although in this study we found one species from Colombia. Autographa californica (Speyer), a lepidopteran, is the unique host record for Crassomicrodus, i.e., C. fulvescens (Cresson) (Sharkey 1997, Figueroa et al. 2008).

In this revision, we redescribe seven species, describe seven new species of Crassomicrodus, and synonymize C. nigriceps under C. fenestratus. We also recognize C. melanopleurus as a valid species and provide an identification key for all species of Crassomicrodus.

## Materials and methods

Species treatments. Descriptions of all included species are based on all material examined. All measurements were performed using a micrometer adapted to an Iroscope microscope and are given in millimeters. Terminology used for the species descriptions follows Sharkey and Wharton (1997) and Sharkey (2006) and for microsculpture of
surface we follow Eady (1968) and Sharkey and Wharton (1997). Data labels were transcribed to a database in the Program Paradox Version 4.5 and the information is presented in a standardized format organized by country and state or province. All photographs were taken using a Leica MZ 16 stereoscope equipped with JVC KY-F75 3CCD digital camera and were prepared using an Auto-Montage imaging system.

Specimens sources. For this revisión were borrowed the types of C. apicipennis Muesebeck, C. divisus (Cresson), C. fenestratus Viereck, C. fulvescens (Cresson), C. medius (Cresson), C. muesebecki Marsh, C. nigriceps (Cresson), C. nigrithorax Muesebeck, C. pallens (Cresson), Microdus melanopleurus Ashmead and Orgilus rileyi Ashmead. Paratypes, homotypes and additional specimens were provided from the following institutions: American Entomological Institute Collection, Florida (AEIC); American Museum of Natural History, New York (AMNH); Academy of Natural Sciences, Philadelphia, Pennsylvania (ANSP); California Academy of Sciences, San Francisco (CAS); Universidad Autónoma de Nuevo León, Nuevo León (CIBE-UANL); Canadian National Collection, Ottawa (CNC); Cornell University Insect Collections, New York (CUIC); Essig Museum of Entomology, University of California, California (EMEC); University of Wyoming, Wyoming (ESUW); Florida State Collection of Arthropods, Florida (FSCA); Hymenoptera Institute Collection, University of Kentucky, Kentucky (HIC); Instituto de Biología, Universidad Autónoma de México (IBUNAM); Fundación e Instituto Miguel Lillo, Universidad Nacional de Tucumán, Argentina (IMLA); Illinois Natural History Survey, Illinois (INHS); Instituto Nacional de Investigaciones Forestales Agrícolas y Pecuarias, Guanajuato (INIFAP); University of Wisconsin, Wisconsin (IRCW); Kansas State University Collection, Kansas (KSUC); Museum of Comparative Zoology, Harvard University (MCZ); Michigan State University Collection, Michigan (MSUC); Museo de la Universidad de Costa Rica (MUCR); Ohio State University, Ohio (OSU); Texas A \& M University, Texas (TAMU); Universidad Autónoma de Yucatán (UADY); The Bohart Museum of Entomology, University of California-Davis, California (UCDC); University of Colorado Museum, Colorado (UCMC); University of California-Riverside, California (UCR); Enns Entomology Museum, University of Missouri-Columbia, Missouri (UMRM); University of Minnesota-St. Paul, Minnesota (UMSP); Smithsonian National Museum of Natural History, Washington (USNM).

## Descriptions and keys

Genus Crassomicrodus Ashmead, 1900
http://species-id.net/wiki/Crassomicrodus
Crassomicrodus Ashmead 1900. Type species Microdus fulvescens Cresson 1865, designated by Ashmead 1900 [Examined].
Epimicrodus Ashmead 1900. Type species Microdus divisus Cresson 1873, designated by Ashmead 1900 [Examined].

Crassomicrodus Ashmead $=($ Epimicrodus Ashmead $)$ synonymized by Bradley 1916.
Diagnosis. Crassomicrodus species can be distinguished from other agathidines with the following combination of characters: simple tarsal claws, without basal lobes, apicolateral pegs of the hind tibia are hair-like, labio-maxillary complex not elongate; mandible with two teeth; and metasomal tergum 1 smooth.

Description. Head. Transverse or triangular; area between antennal sockets with a median pyramidal-shaped elevation or transverse; gena not bulging to distinctly bulging; labio-maxillary complex not elongate, mandible with two teeth, antenna with 25 to 43 flagellomeres.

Mesosoma. Pronotum surface smooth or punctuate; notauli from lacking to impressed; anterolateral edges of scutellum with or without a small acute projection; lateral scutellar depression from smooth to crenulate; dorsal surface of propodeum from rugulose to reticulate rugose; subalar lobe separated from mesopleuron by a wide or narrow groove; metapleuron from smooth to reticulate rugose; inner spur of hind tibia from 0.47 to 0.78 times longer than basitarsus; tarsal claw without basal lobe; outer apex of the hind tibia without flattened pegs; forewing vein R1 0.47-0.70 times longer than RS; crossvein $r$ arising before or beyond middle of stigma. Metasoma. Metasomal median tergite 1 smooth; apical width 1.78-3.92 times longer than basal width; ovipositor sheaths and ovipositor variable in length; metasoma 1.00-1.42 times longer than mesosoma.

## Key to the New World species of Crassomicrodus Ashmead

1 Head triangular in frontal view (Figs 2a, 4a, 5a, 6a, 10a, 12a, 14a); gena not bulging; area between antennal sockets with a median pyramidal-shaped elevation (Figs 2a, 4a, 5a, 6a, 10a, 12a, 14a); length of ventrolateral margin of clypeus similar to diameter of tentorial pit (Fig. 1a, 8a) 2

- Head transverse in frontal view (Figs 1a, 3a, 7a, 8a, 9a, 11a, 13a), if somewhat triangular then length of ventrolateral margin of clypeus longer than diameter of tentorial pit (Fig. 3a); gena bulging or at least slightly bulging (Figs 1a, 3a, 7a, 8a, 9a, 11a, 13a); area between antennal sockets variable in shape 10
2(1) Malar space at least 0.8 times as long as eye height (Figs 5a, 14a)................ 3
- Malar space at most 0.6 times as long as eye height (Figs 2a, 4a, 6a, 10a, 12a) 4
3(2) Scutellar sulcus with 3 or 4 carinae; second submarginal cell quadrangular; fore and middle legs black; body length $6.95-8.60 \mathrm{~mm} . . C$. divisus (Cresson)
- $\quad$ Scutellar sulcus with 1 carina; second submarginal cell triangular; fore and middle legs yellowish-orange; body length $4.20-6.48 \mathrm{~mm}$
C. pallens (Cresson)

4(2) Forewing vein R1 at most half the length of RS; head yellowish-orange (Fig. 14a)
C. pallens (Cresson)


13(12) Inner spur of hind tibia distinctly longer than half of basitarsus length (0.600.72 ); gena bulging (Fig. 8a); lateral depression of scutellum smooth, only its ventral edge with small punctures $\qquad$ C. melanopleurus Ashmead

- Inner spur of hind tibia less than half length of basitarsus (a few specimens with the spur at most reaching 0.54 times); gena distinctly bulging; lateral depression of scutellum rugose and fovelate.
C. fulvescens (Cresson)

14(10) Head and mesosoma black; anterolateral edges of scutellum lacking small acute projection 15

- Head and mesosoma at least with some areas yellowish-orange or yellowishred (Figs lacde); anterolateral edges of scutellum with small acute projection (Fig. 1df)
C. apicipennis Muesebeck

15(14) Notauli not impressed over most of mesoscutum (Fig. 9c); metasoma black; setae at base of mandible similar in size to setae on rest of body surface ( 9 ad ); length of setae in scutellar disk 0.18 to 0.20 mm ; ovipositor sheaths of female at least 1.83 mm in length C. muesebecki Marsh

- $\quad$ Notauli impressed over most of mesoscutum (Fig. 11c, 13d); metasoma yel-lowish-orange (Figs 11e, 13e); setae at base of mandible distinctly longer than setae on rest of body surface (11ad, 13ac); length of setae in scutellar disk less than 0.17 mm ; ovipositor sheaths of female at most 0.22 mm in length ... $\mathbf{1 6}$
16(15) Area between antennal sockets with a median elevation in trapezoidal shape (Fig. 13a); gena distinctly bulging (Fig. 13a); malar space 0.54-0.59 times longer than eye height; 31-34 flagellomeres; body length 6.70-7.08 mm.....
C. olgae sp. n .
- Area between antennal sockets with a median transverse elevation (Fig. 11a); gena bulging (Fig. 11a); malar space 0.38-0.47 times longer than eye height; 28-31 flagellomeres; body length $3.95-5.35 \mathrm{~mm} . . . C$. nigrithorax Muesebeck


## Species descriptions of Crassomicrodus

Crassomicrodus apicipennis Muesebeck, 1927
http://species-id.net/wiki/Crassomicrodus_apicipennis
Fig. 1a-f
Crassomicrodus apicipennis Muesebeck 1927: 18-19

Holotype female. Mount Hood, Oregon [USA]. Cat. No. 28695 (USNM).
Description female. Body. Length. 4.90-5.97 mm. Color (Fig. 1e). Integument black except yellowish-orange as follows, basal area of mandible, pronotum, mesonotum, subalar lobe, tegula, metasoma, femora, basal area of hind tibia, anterior and middle tibia and tarsomeres; mandible apex, basal area of hind tibia; tarsomeres blackish; wing veins dark brown; forewing infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Specimens range from black to yellowish-


Figure I. Crassomicrodus apicipennis. Female a anterior view of head, arrow indicates a median transverse elevation with two weakly defined lateral tubercles, ventrolateral margin of clypeus, and tentorial pit b dorsal view of head, arrow indicates groove between lateral ocelli with small foveolae c lateral view of mesosoma, arrows indicate pronotum and metapleuron $\mathbf{d}$ dorsal view of mesosoma, arrows indicate anterolateral edge of scutellum and propodeum $\mathbf{e}$ female habitus $\mathbf{f}$ dorsal view of scutellum, arrow indicates anterolateral edge of scutellum with a small acute projection.
orange on the head, propleuron, metanotum, propodeum, mesopleuron, metapleuron and hind coxa. Head (Fig. 1ab). Transverse in frontal view; face dorsomedially with weak longitudinal ridge in most specimens; eye height/width $=1.31-1.35$; eye height $0.65-0.68 \times$ inter-ocular distance; area between antennal sockets with a median transverse elevation and two weakly defined tubercles; frons deeply excavated and rugulose with small foveolae; posterior surface of antennal sockets smooth; groove between lateral ocelli with small foveolae; median ocellus separated from lateral ocellus by groove with small foveolae; gena bulging; malar space $0.48-0.55 \times$ as long as eye height; clypeus (anterior view) 2.29-2.33× wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 29-32 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. $1 \mathrm{c}-\mathrm{f})$. Pronotum smooth; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum with a small acute projection; scutellar disc convex with sparse setae from 0.16 to 0.17 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression rugose and foveolate; carinae of central metanotal area forming a triangular cell; propodeum reticulate rugose, more pronounced on lateral margins; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly of shorter than subalar lobe; metapleuron rugose with reticulate-foveolae. Legs. Inner spur of middle tibia 0.76-0.83× length of basitarsus; inner spur of hind tibia $0.59-0.69 \times$ length of basitarsus; metabasitarsus $1.18-1.25 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.00-2.21× longer than basitarsus; hind femur length $3.38-4.12 \times$ its maximum width. Wings. Forewing length/ width $=2.46-2.51$; stigma $2.69-3.50 \times$ longer than maximum width; forewing vein R1 0.60-0.66× as long as vein RS; vein RS sinuate; vein r arising slightly before middle of stigma; second submarginal cell triangular, with petiole $0.07-0.13 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.37-3.52$; hind wing vein $1 \mathrm{M} 1.65-1.80 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $4-5$ hamuli. Metasoma. Apical width of petiole (tergum 1) $2.90-3.21 \times$ wider than basal width; minimum width of petiole $0.58-0.60 \times$ apical width; length of ovipositor sheath $0.17-0.30 \mathrm{~mm}$.

Male. Similar to female.
Host. Unknown.
Distribution. Canada, Mexico, and USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median transverse elevation, anterolateral edges of scutellum with a small acute projection, groove between lateral ocelli with small foveolae, and head and mesosoma black with some areas yellowish-orange.

Remark. This species is near to C. nigrithorax, but differs in that C. nigrithorax has the mesosoma black; anterolateral edges of scutellum lacking small acute projection; eye height $0.69-0.70 \times$ inter-ocular distance; malar space $0.38-0.47 \times$ longer than eye height; inner spur of middle tibia $0.89-0.95 \times$ length of basitarsus; and scutellar disc convex with sparse setae from 0.14 to 0.15 mm in length.

Material examined. Holotype $q$ : USA, Oregon: Mt. Hood. Collection Ashmead (USNM). Allotype $\delta^{\lambda}$ : same data as type (USNM). Homotype $q$ : USA, Iowa: Iowa Co.,

10／VIII／1934，Jaques H．E．（CNC）．Other specimens examined．－CANADA，Alberta： Alta． 1 Q；Scandia Alta．， 1 q 26／VII／1956，Peck O．（CNC）．British Columbia：Hed－ ley，Nickel Plate City， 1 \＆17／VIII／1953， 1524 m，Martin J．E．H．；Kamloops 1 đ 9／ VII／1939，Spencer G．J．；Richter Pass，Osoyoos 1 q 28／VI／1959，Kelton L．A．（CNC）． Manitoba：Aweme，N Criddle， 1 q 9／VII／1924（CNC）．MEXICO，Durango： 5 mi ． W Durango 1 § 24／VI／1964，Howden H．F．（HIC）．Sonora：Nogales 1 § 4／IX／1965， Michelbacher A．E．（EMEC）；Tamaulipas：Santa Teresa 1 ठ 15／V／1952，Cazier M．，W． Gertsch \＆R．Schrammel（AMNH）．USA：Iowa Co．， 1 q 19／VI／1935，Huizinga H． （USNM）．Arizona：Springerville， 1 ठ 26／VII／1956，Butler－Gerhardt（USNM）； 9 km SE Camp Verde，Clear Creek，Yavapai Co．， 1 § 11／IX／1986，Parker F．\＆T．Griswold （CNC）．Colorado： $1 \delta^{\top}$ ；Boulder Canyon， 1 q 8／VIII／1960， 2377 m，Dreisbach R．K． （USNM）； 3 mi．NW Wiggins，Morgan Co．， 1 § 8／VIII／1974，Favreau M．\＆T．M．； Pagosa Springs， 1 中， 1 đ̋ 22－24／VI／1919， 2195 m，37．16 N 107．0 W（AMNH）；Craig， 1 § 25／VI／1949，Bryant L．（CAS）；Fort Morgan， 1 § 5／VIII／1960，Dreisbach R．K．； Limon， 1 ठ 16／VIII／1949，Dreisbach R．R．\＆R．K．Shwab（MSUC）；Limon， 1 ठ 16／ VIII／1949，Dreisbach R．R．\＆R．K．Shwab（AEIC）；Nunn， 1 q 7／VI／1976，Lavigne R． （ESUW）；Prowers Co．， 1 \＆5／VI／1962，Marston N．（KSUC）；Running Creek Field Sta．Elbert Co．T9S，R65W，Sec 35 NE nativo， 2 q 2118 m，Brown F．M．（UCMC）． Kansas：Manhattan， 1 q， 1 § 16／VII／1950，Evans H．E．；Manhattan， 1 đ 24／V／1935， Wilbur D．A．（KSUC）；Sitka，Clark Co．， 1 ＋12／VI／1960，Eickwort G．C．（MSUC）． Minnesota：Albert Lea，Freeborn Co．， 1 q， 1 § 11／VI／1961，Levi H．（MCZ）；Browns Valley， 1 đ 4／VIII／1935，Denning D．G．；Lincoln Co．， 1 § 4／VI／1938，Mickel C．E．； Ortonville： 1 đ̋ 5／VIII／1935，Deggy R．H．； 6 đ 5／VIII／1935，Denning D．G．；Polk Co．， 1 Q 16／VIII／1936，Daggy R．H．；Yellow，Medicine Co．， 1 § 13／VIII／1936，Mickel C．E． （UMSP）．Missouri：Diehlstadt， 1 đ 5／VIII／1938，Wingo C．（UMRM）；Columbia， 1 § 7／VIII／1958，Blickenstaff C．C．（USNM）．Nebraska：Valentine Refuge， 1 ठ 7／VI／1972， Townes H．\＆M．Townes（AEIC）； 1.5 mi．N Mullen（Middle Loup River），Hooker Co．， 1 \＆2－4／VII／1983，Grissell \＆Menke（HIC）；Thomas Co．， 1 o 21／VIII／1951，Dre－ isbach R．R．（MSUC）；Halsey： 1 § 12／VIII／1925，Dawson R．W．； 1 § 15／VIII／1925， Dawson R．W．；McCool， 1 q 22／VIII／1940，Milliron H．E．（UMSP）；Morril Co．， 1 ठ 22／VIII／1951，Dreisbach R．R．（USNM）．Oklahoma：Lake Carl Etling，Black Mesa State Park， 25 mi．by road NW Boise City，Cimarron Co．， 1 § 14／VIII／1967，Leech H．（CAS）；Pond Creek，Grant Co．， 2 Q 11／VI／1960，Eickwort G．C．（MSUC）．Texas： Spring， 1 § 21／VI／1947，Rockefeller D．（AMNH）； 30 mi ．N Uvalde，Uvalde Co．， 1 q 21／VI／1983，Pulawski W．J．（CAS）；East Hwy．side of Road way park， 7 mi．S Beeville， 1中 2／VI／1964，Hull F．M．\＆M．C．（CNC）；Sam Houston，Bexar Co．， 1 \＆31／III／1953， Adelson B．J．（EMEC）； 3 mi．W Estelline，Hall Co．， 1 \＆3／VI／1979，Michener C．D． （HIC）；Alice： 1 § 17／VII／1954，Dreisbach R．R．（MSUC）； 1 § 17／VII／1954，Dreis－ bach R．R．（USNM）；Bangs， 1 q 6／VIII／1938（USNM）；Bentsen Rio Grande Valley State Park．Hidalgo Co．： 1 \＆11／VIII／1983，Bars M．Kaul（CNC）； 1 đ 10／VI／1982，
 25／V／1977， 1 §̉；27／VI／1983， 1 ð 28／VI／1983， 3 中 4／VI／1982， 1 ð 9／VI／1982，Por－ ter C．（FSCA）；Bentsen Rio Grande Valley State Park．Nr．Mission，Hidalgo Co．： 1 中，

1 đ 13/VI/1983, 1 中, 1 § 20/VI/1983, 1 ð 22/III/1984, 1 ð 26/VII/1984, 1 q 30/ VIII/1983, 4 ㅇ, 2 §̉; 4/VIII/1983, Porter C. (FSCA); Dumas, Moore Co., 3 § y 1 § 14/VI/1960, Fischer R.L. (MSUC); Brownsville, 1 § 6/VII/1895; Kerrville, 1 q 12/IV/1907, Pratt F.C.; College State, 2 đ 29/III/1954, Lewis W.J. (USNM). Utah: Goblin Vly, sand dunes, Emery Co., 1 Q 20/VI/1980, Parker F.D.; Wild Horse cr. W Goblin Vly., Emery Co., 1 \& 26/VII/1983, 1463 m, Parker F. \& T. Griswold (CNC). Bonanza, Uintah Co., 1 ¢ 27/VI/1978, Bohart G. (UCMC). Wyoming: Glendo, 1 đ 24/VI/1960, Lavigne R.J. (USNM).

## Crassomicrodus azteca Figueroa, Romero \& Sharkey, sp. n. urn:lsid:zoobank.org:act:D1F7CD97-570A-42DB-B164-312E6869C770 http://species-id.net/wiki/Crassomicrodus_azteca

Fig. 2a-d

Description female. Body. Length. 5.20-5.50 mm. Color (Fig. 2d). Integument black except yellowish-orange as follows, medial area of mandible, tegula, femora, anterior and middle tibiae and tarsomeres, and metasoma; eyes silver; ocelli translucent honey yellow; wing veins dark brown; forewing slightly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. In some specimens the first metasomal tergite blackish. Head (Fig. 2ab). Triangular in frontal view; face with weak longitudinal ridge dorsomedially; eye height/width $=1.43-1.45$; eye height $0.59-0.63 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation; frons excavated with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior surface of antennal sockets slightly rugulose; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.60-0.63 \times$ as long as eye height; clypeus $2.27-2.35 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 31-33 flagellomeres; setae at base of mandible slightly longer than setae on rest of body surface. Mesosoma (Fig. 2bcd). Pronotum punctulate with setae; lateral pronotal margins with a shallow, crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.13 to 0.14 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth, sometimes with microfoveolate grooves in its ventral and dorsal margins; carinae of central metanotal area almost circular shaped, sometimes triangular; propodeum reticulate rugose with abundant setae in its lateral areas; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly of smaller size than the subalar lobe; metapleuron reticulate rugulose in its ventral half and smooth in its dorsal half. Legs. Inner spur of middle tibia $0.65-0.70 \times$ length of basitarsus; inner spur of hind tibia $0.58-0.64 \times$ length of basitarsus; metabasitarsus $1.05-1.16 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.41-2.67× longer than basitarsus; hind femur length $3.50-4.00 \times$ its maximum width. Wings. Forewing length $/$ width $=2.50$ 2.55; stigma 3.00-3.23× longer than maximum width; forewing vein R1 $0.61-0.68 \times$ as


Figure 2. Crassomicrodus azteca sp. n. Female $\mathbf{a}$ anterior view of head, arrow indicates a median pyram-idal-shaped elevation $\mathbf{b}$ dorsal view of head and mesosoma, arrow indicates posterior surface of antennal sockets slightly rugulose $\mathbf{c}$ lateral view of mesosoma, arrow indicates subalar lobe separated from mesopleuron by narrow groove $\mathbf{d}$ female habitus, arrow indicates triangular-shaped second submarginal cell.
long as vein RS; vein RS slightly straight; vein r arising before middle of stigma; second submarginal cell triangular, with petiole $0.07-0.12 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ weakly pigmented in 0.75 of its basal length; hind wing length/width $=3.37-3.38$; hind wing vein 1M 1.44-1.70× longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $4-5$ hamuli. Metasoma. Apical width of petiole (tergum 1) 3.46-3.71× wider than basal width; minimum width of petiole $0.53-0.59 \times$ apical width; length of ovipositor sheath $0.13-0.28 \mathrm{~mm}$.

Male. Similar to female, but the posterior surface of antennal sockets smooth.

Host. Copitarsia sp. (Lepidoptera: Noctuidae) in Brassica oleraceae L. (cauliflower).
Distribution. Mexico.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, malar space $0.60-0.63 \times$ as long as eye height, setae at base of mandible slightly longer than setae on rest of body surface, body length $5.20-5.50 \mathrm{~mm}$, head and mesosoma black, and forewing slightly infumate.

Remarks. This species is similar to C. nigrithorax, but differs in that C. nigrithorax has gena bulging; area between antennal sockets with a median transverse elevation; groove between lateral ocelli with small foveolae; and malar space $0.38-0.47 \times$ longer than eye height.

Etymology. The specific epithet is a noun in apposition to Crassomicrodus, which is reference to some of the indigenous people of Mexico.

Material examined. Holotype : MEXICO , México: Chapingo, 31/VII/1967, J.L. Carrillo Sánchez. Copitarsia sp. Allotype $\delta^{\imath}$ : same data as holotype. Holotype and Allotype deposited in Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolás de Hidalgo, Tarímbaro, Michoacán, México (IIAF-UMSNH). Paratypes: MEXICO, Guanajuato: 10 mi . NW Leon, 1 ô 19/VIII/1954, 2042 m , Chillcott J.G. (CNC); Guanajuato, 1 Q 15/VIII/1953, Vaurie C. \& P. Vaurie (AMNH). Hidalgo: Pachuca, Junction Rt. 85, 2 ¢ $24 / \mathrm{IV} / 1965$, 2438-2591 m, Weems H.V. Jr. (FSCA). Jalisco: Lagos de Moreno, 1 ¢ 12/VIII/1954, 1920 m, Dreisbach R.R. (USNM); Rancho La Quinta Teocaltiche, 1 § $25 / \mathrm{VIII} / 1979,1707 \mathrm{~m}$, Villegas B. (UCDC). México state: Chapingo: 1 § 1950, F. Pacheco M. (USNM); 2 q 1/VIII/1967, 7/VIII/1967, J.L. Carrillo Sánchez. Copitarsia sp., (IIAF); Chimalhuacán, 1 đ 28/VIII/1967, J.L. Carrillo Sánchez, Copitarsia sp. (IIAF); Pirámides de Teotihuacan, 3 \& 7/VII/1951, Hurd P.D. (USNM); Texcoco, 1 § 12/VIII/1954, 2134 m , Chillcott J.G. (CNC). Nuevo Leon: 32 km W Linares, San Pedro Iturbide, 1 §, 2 q 5/X/1962, Townes H. \& M. Townes (AEIC). Puebla: 14 km NE Cañada Morelos, 1 § 10/VII/1974, Chemsak J., E.G. Linsley \& J. Linsley (EMEC). Zacatecas: 15 km E Sombrerete, 1 § 30/VII/1951, Hurd P.D. (USNM); 5 km NE Huejucar, 1 Q 13/IX/1984, Pulawski W.J., 22.21 N 103.13 W (CAS); 5 mi. N Zacatecas, 1 q 19/IX/1970, Bohart G.E. \& R.M. Bohart (CNC); 9 mi. SE Fresnillo, 1 § 7-14/VIII/1954, Linsley E.G., J.W. MacSwain \& R.F. Smith (EMEC); Fresnillo, 1 ठ 15/VIII/1947, 2134 m, Rockefeller D. (AMNH).

## Crassomicrodus clypealis Figueroa, Sharkey \& Romero, sp. n. urn:Isid:zoobank.org:act:1D3A68FB-656B-44F1-BC9E-B7E343BF4A90 <br> http://species-id.net/wiki/Crassomicrodus_clypealis

Fig. 3a-e
Description female. Body. Length. $7.38-7.50 \mathrm{~mm}$. Color (Fig. 3c). Integument yellowish-orange except black as follows, mandible apex, head, antenna, propleuron, mesopleuron, metapleuron, metanotum, propodeum, coxa and trochanters; ocelli translucent honey yellow; apical area of hind tibia, middle and hind tarsomeres black-


Figure 3. Crassomicrodus clypealis sp. n. Female a anterior view of head, arrows indicate a median pyram-idal-shaped elevation with two weakly defined tubercles, ventrolateral margin of clypeus, and tentorial pit $\mathbf{b}$ dorsal view of head, arrow indicates frons deeply excavated with a pair of microfoveolate grooves $\mathbf{c}$ female habitus $\mathbf{d}$ dorsal view of mesosoma, arrows indicate impressed notauli $\mathbf{e}$ lateral view of mesosoma, arrow indicates subalar lobe separated from mesopleuron by narrow groove.
ish; wing veins dark brown; forewing strongly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig. 3ab). Transverse in frontal view; face with weak longitudinal ridge dorsomedially; eye height/width $=$ 1.38-1.43; eye height $0.68-0.70 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles; frons deeply excavated with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena slightly bulging; malar space $0.46-0.48 \times$ as long as eye height; clypeus $2.40-2.42 \times$ wider than high; length of ventrolateral margin of clypeus distinctly longer than diameter of ten-
torial pit; antenna with 38-39 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 3cde). Pronotum smooth; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.07 to 0.08 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression with punctures and foveolae; carinae of central metanotal area in triangular shaped; propodeum reticulate rugose; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; metapleuron reticulate rugulose or reticulate punctures. Legs. Inner spur of middle tibia $0.69-0.72 \times$ length of basitarsus; inner spur of hind tibia $0.55-0.58 \times$ length of basitarsus; metabasitarsus $1.15-1.23 \times$ length of tarsomeres III, IV, and V combined; hind tibia $2.25-2.30 \times$ longer than basitarsus; hind femur length $3.56-3.78 \times$ its maximum width. Wings. Forewing length/width $=2.42-2.50$; stigma $2.88-3.00 \times$ longer than maximum width; forewing vein R1 $0.58-0.62 \times$ as long as vein RS; vein RS sinuate; vein $r$ arising slightly before middle of stigma; second submarginal cell triangular, with petiole $0.06-0.09 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=2.91-3.15$; hind wing vein $1 \mathrm{M} 1.35-1.43 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 6-7 hamuli. Metasoma. Apical width of petiole (tergum 1) 2.82-3.14× wider than basal width; minimum width of petiole $0.56-0.64 \times$ apical width; length of ovipositor sheath $0.35-0.37 \mathrm{~mm}$.

Male. Similar to female, except antenna with 36 to 41 flagellomeres and pronotum may be slightly melanic.

Host. Unkown.
Distribution. USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, length of ventrolateral margin of clypeus distinctly longer than diameter of tentorial pit, and gena slightly bulging.

Remarks. This species is near to male of C. fulvescens, but differs in that C. fulvescens has the length of ventrolateral margin of clypeus similar to the diameter of the tentorial pit; gena distinctly bulging; and forewing infumate with a large hyaline spot in first submarginal cell.

Etymology. Named "clypealis" to emphasize that the ventral margin of the clypeus is longer than the diameter of each tentorial pit.

Material examined. Holotype $Q$ : USA, Colorado: nr. Roggen, 4/IX/1972, Lanham U.N. \& C.C. Lanham. Allotype ō: Roggen, 29/VIII/1930, Rodeck H.G.. Holotype and Allotype deposited in UCMC. Paratypes: $1 \delta^{\top}$; USA, Colorado: 3 mi . NW Roggen, Weld Co., 4/IX/1974, Lanham U.N. (CNC); 1 q same data as holotype; 1 § same data as allotype (UCMC); Sandhills N of Roggen Weld Co.: 1 § 12/IX/1934, Rodeck H.G. (HIC); 1 § 12/IX/1934, Rodeck H.G. (USNM); 2 § 12/IX/1934, Rodeck H.G.; 1 § 16/VIII/1990, Bowers M.D. (UCMC). Kansas: Ness Co., 1 ठ $7 /$
 VIII/1925, 1 § 15/VIII/1925, Dawson R.W. (UMSP).

Crassomicrodus costaricensis Figueroa, Sharkey \& Romero, sp. n. urn:lsid:zoobank.org:act:B9743BFC-4824-494E-BD02-B19BCBF66134<br>http://species-id.net/wiki/Crassomicrodus_costaricensis<br>Fig. 4a-e

Description female. Body. Length. 8.10-8.70 mm. Color (Fig. 4e). Integument black except eye silver, ocelli translucent yellow (Fig. 1d); medial area of mandible yellowreddish; metasoma dark brown; forewing strongly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig. 4ab). Triangular in frontal view; face with longitudinal ridge dorsomedially; eye height/width = $1.41-1.42$; eye height (lateral view) $0.75-0.77 \times$ inter-ocular distance (anterior view); area between antennal sockets with a median pyramidal-shaped elevation; frons excavated with a little longitudinal groove; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.47-0.51 \times$ as long as eye height; clypeus 2.13-2.32× wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 41 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 4cde). Pronotum punctulate with setae; lateral pronotal margins with a shallow, crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.12 to 0.14 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth with punctures or rugosities in its ventral border; carinae of central metanotal area almost circular shaped; propodeum reticulate rugulose, more pronounced on lateral margins; subalar lobe separated from mesopleuron by wide rugulose groove, width almost of similar size to subalar lobe; metapleuron smooth, only reticulate rugulose one-fourth of ventral area. Legs. Inner spur of middle tibia $0.76-0.89 \times$ length of basitarsus; inner spur of hind tibia $0.62-0.72 \times$ length of basitarsus; metabasitarsus $1.02-1.18 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.04-2.38× longer than basitarsus; hind femur length $4.07-4.17 \times$ its maximum width. Wings. Forewing length/width $=2.58$; stigma $4.54-5.00 \times$ longer than maximum width; forewing vein R1 $0.60-0.64 \times$ as long as vein RS; vein RS sinuate; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.13-0.20 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/ width $=3.48-3.65$; hind wing vein $1 \mathrm{M} 1.01-1.08 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 6-7 hamuli. Metasoma. Apical width of petiole (tergum 1) 2.86-2.91× wider than basal width; minimum width of petiole $0.49-0.59 \times$ apical width; length of ovipositor sheath $0.28-0.41 \mathrm{~mm}$.

Male. Unknown.
Host. Unknown.
Distribution. Costa Rica and El Salvador.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, eye height $0.75-0.77 \times$ inter-ocular distance, antenna with 41 flagellomeres,


Figure 4. Crassomicrodus costaricensis sp. n. Female a anterior view of head, arrow indicates a median pyramidal-shaped elevation $\mathbf{b}$ dorsal view of head $\mathbf{c}$ lateral view of mesosoma, arrow indicates subalar lobe separated from mesopleuron by wide groove $\mathbf{d}$ dorsal view of mesosoma $\mathbf{e}$ female habitus.
 and mesosoma black, metasoma dark brown, and wings strongly infumate.

Remark. Specimens from Costa Rica are homogeneous in their measurements, but the specimen from El Salvador differs significantly. Nonetheless it is considered conspecific here due to similarity in other characters. More specimens and molecular data should easily test this hypothesis in the future.

Etymology. The specific name is a noun in apposition to Crassomicrodus and is chosen because of the locality where the holotype was collected, Costa Rica.

Material examined. Holotype $q$ : COSTA RICA, Guanacaste: Barra Honda NP, VI/1988, 200 m. , Gauld \& Mitchel, deposited in MUCR. Paratypes: 1 q same data as holotype (MUCR); Guanacaste: Scrub forest (7yr), Open site, 1 \& 22/VI/1985, Gauld \& Janzen, 300 m. (HIC). EL SALVADOR, [La Unión]: Vol. Conchagua, 1 ; 27-29/V/1958, Cartwright O.L. (MUCR).

## Crassomicrodus divisus (Cresson, 1873)

http://species-id.net/wiki/Crassomicrodus_divisus
Fig. 5a-e
Crassomicrodus divisus (Cresson): Bradley 1916: 139-140; Muesebeck 1927: 21.
Microdus divisus Cresson 1873: 52 [Examined].
Orgilus rileyi Ashmead 1888: 640 [Examined].
Epimicrodus divisus Ashmead 1900: 129.

Holotype female. Illinois [USA]. No. 1726.1 (ANSP).
Description female. Body. Length. $6.95-8.60 \mathrm{~mm}$. Color (Fig. 5e). Integument black except reddish yellow as follows, pronotum, mesonotum, subalar lobe, tegulae, hind femora, and metasoma; mandible and wing veins dark brown; eye silver or blackish; ocelli translucent yellow; forewing infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Rarely, central area of mesopleuron or hind coxa or propodeum and metapleuron reddish yellow. Head (Fig. 5ab). Triangular in frontal view; face with longitudinal ridge dorsomedially; eye height/width $=1.38-1.40$; eye height $0.57-0.58 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation, sometimes with two weakly defined tubercles; frons excavated with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.78-0.83 \times$ as long as eye height; clypeus $1.85-1.95 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 32-35 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 5cde). Pronotum reticulate rugulose, sometimes strigose; lateral pronotal margins with a shallow, crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc slightly convex with sparse setae from 0.10 to 0.11 mm in length; scutellar disc sloped posteriorly and flattened; lateral scutellar depression reticulate rugulose and foveolae; carinae of central metanotal area forming a triangular cell; propodeum reticulate rugose; subalar lobe separated from mesopleuron by wide rugose groove with reticulate foveolae, width almost of similar size to subalar lobe; metapleuron reticulate-rugose. Legs. Inner spur of middle tibia $0.67-0.71 \times$ length of basitarsus; inner spur of hind tibia $0.59-0.68 \times$ length of basitarsus; metabasitarsus $1.21-1.29 \times$ length of tarsomeres III, IV, and V combined; hind tibia $2.17-2.27 \times$ longer than basitarsus; hind femur length $3.91-4.22 \times$ its maximum width. Wings. Forewing length/width $=2.56-2.60$; stigma 3.27-3.70× longer than maximum width; forewing vein R1 $0.59-0.64 \times$ as long as vein RS; vein RS not sinuate; vein $r$ arising before middle of stigma; second submarginal cell quadrangular, with petiole $0.12-0.21 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.25-3.51$; hind wing vein 1 M $1.52-1.60 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $6-7$ hamuli. Metasoma. Apical width of petiole (tergum 1) 3.10-3.13× wider than basal width; minimum width of petiole $0.61-0.63 \times$ apical width; length of ovipositor sheath $0.33-0.41 \mathrm{~mm}$.


Figure 5. Crassomicrodus divisus. Female a anterior view of head, arrow indicates a median pyramidalshaped elevation $\mathbf{b}$ dorsal view of head $\mathbf{c}$. dorsal view of mesosoma $\mathbf{d}$ lateral view of mesosoma, arrow indicates subalar lobe separated from mesopleuron by wide groove $\mathbf{e}$. female habitus.

Male. Unknown.
Host. Unknown.
Distribution. Canada, Mexico, and USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, malar space $0.78-0.83 \times$ as long as eye height, scutellar disc sloped posteriorly and flattened, head black and mesosoma mostly black with some areas reddish yellow.

Remarks. Males of C. divisus were not found in this revision, although Muesebeck (1927) recorded them. We carefully examined long series from diverse localities to find them but without success. Therefore we speculate that males of this species are absent or very rare.

Material examined. Type: M. divisus 1 q, Ill. (ANSP), O. riley 1 $q$ (USNM). Homotype: 1 Q, Texas: Davis Ranch, NW Blanco Co., 22/IV/1959, Mason W.R.M. (CNC). Other specimens examined.- 1 q (AMNH). 9 q (INHS). 1 q (MSUC). 1 ㅇ (USNM). 1 q; 19/VI/1898 (AEIC). 1 q; VII/1917 (USNM). 1 q; CANADA, Ontario: Constance Bay, Carleton Co., 24-31/VI/1977, Sanborne M. (CNC). 1 q; Ottawa, 3/X/1912, Beaulne J.I. (CNC). 1 ; Cana (USNM). 1 ; ; MEXICO, Durango: 10 miles N Durango, 12/VII/1954, McSwain (EMEC). 1 ; Guanajuato: San Miguel Allende, 12/VIII/1953, Vaurie C. \& P. Vaurie (AMNH). 1 ; ; Jalisco: 4 miles W Mazamitla, 16/X/1950, 2073 m., Smith Ray F. (AMNH). 1 \& Guadalajara, 17/ IX/1957, Dreisbach R. \& K. Dreisbach (MSUC). 1 q; Guadalajara, 17-20/VII/1965, Evans H.E. (MCZ). 1 q; Nuevo León: 4 miles W El Cercado, 6/VI/1951, Hurd P.D. (EMEC). 1 +; 9 miles W Iturbide, 3/VII/1974, Clark, Murray, Ashe, Schaffner (TAMU). 1 中; Puebla: Puebla, 3/VII/1952, Gilbert E.E. \& C.D. MacNeil (EMEC). 1 Y; Zacatecas: Trancosa, 3/VII/1961, Dreisbach R. \& K. Dreisbach (MSUC). 4 P; USA, Colo. (USNM). 1 q; Detroit, 7/VI (USNM). 1 q; Arizona: Canelo, 10/ VII/1957, Butler G.D. (HIC). 1 q; Illinois: Algonquin, 30/VIII/94-99 (INHS). 1 ○; Algonquin, Nason (INHS). 1 q; Grand Tower, 28/VII/1905 (INHS). 1 q; Illinois, VIII/1899 (MCZ). 1 q; Meredosia, 20/VIII/1917 (INHS). 1 中; Iowa: Ames, 25/V/1925 (UCDC). 1 q; Sioux City, Ainslie C.N. (USNM). 1 q; Kansas: Baldwin, V/190?, Bridwell J.C. (USNM). 1 q; Baldwin, VIII, Bridwell J.C. (UCDC). 1 q; Douglas Co. (MCZ). 1 q; Manhattan, 10/VI/1950, Evans H.E. (KSUC). 1 \& Manhattan, 10/VII/1949, Kring James B. (KSUC). 1 q; Onaga, 9/VII/1922, Crevecoeur (KSUC). 1 q; Riley Co., 16/VI, Smith R.C. (KSUC). 1 q; Riley Co., 9/VII, Popenoe (KSUC). 1 q; Riley Co., V, Marlatt (KSUC). 1 q; Kentucky: Lexington (USNM). 1 O; Michigan: White Pigeon, St. Joseph Co., 14/VI/1959, Fischer R.L. (MSUC). 1 Q; Minnesota: Lake Pepin E Frontenac, 24/V/1941, Sun V.P. (UMSP). 1 q; Ortonville, 5/VIII/1935, Denning D.G. (UMSP). 1 q; Olmsted Co., VIII/1896, Linslie C.N. (USNM). 1 q; Missouri: Columbia, Boone Co., 16/VIII/1968, Parker F.D., malaise trap (HIC). 1 q; Clayton, 3/VI/1939, Pickel B.H. (AEIC). 1 \&; Columbia, 1/ VIII/1968, Parker F.D. malaise trap (UCDC). 1 q; Columbia, 10/VIII/1967, Parker F.D. malaise trap (USNM). 1 ; Columbia, 14/IX/1939, Crajo W.S. (UMRM). 1 ; Columbia, 17/VIII/1966, Huber S.F. (USNM). 1 q; Columbia, 2/VIII/1968, Parker F.D. malaise trap (UCDC). 1 q; Columbia, 23/VIII/1967, Parker F.D. malaise trap (USNM). 2 q; Columbia, 26/VIII/1967, Parker F.D. malaise trap (USNM). 1 ; Columbia, 10/VII/1967, Parker F.D., malaise trap (USNM). 1 q; Columbia, 22/ VII/1967, Parker F.D., malaise trap (USNM). 1 q; Columbia, 20/IX/1967, Parker F.D., malaise trap (USNM). 1 ; Columbia, 6/IX/1967, Parker F.D. malaise trap (USNM). 1 q; Columbia, 7/IX/1966, Huber S.F. (USNM). 1 q; Columbia, 16/ VIII/1966, Huber S.F. (USNM). 2 q; Columbia, Boone Co., 22/VIII/1968, Parker F.D. malaise trap (USNM). 2 ; Jefferson City, 6/VII/1941, Adams C.F. (USNM). 1 ; Sapp, 10/VII/1954 (UMRM). 1 Q ; Nebraska: Maxwell, 25/VII/1967 (IRCW). 1 Q; New Jersey: Camden, 1892? (MSUC). 1 q; Trenton, Abbatt (MCZ). 1 q; New York: Boston, 1/VIII/1909, M.C.V. (UCDC). 1 q; Lancaster, 10/VIII/1891, E.P.V.
(CAS). 1 Q; Ohio: Champaign Co., 23/VII/1941, Guillaspy J.F. (OSU). 1 + Champaign Co., 23/VII/1941, Guillaspy J.F. (OSU). 1 q; Dayton O., VIII/1927, Basker C.A. (OSU). 1 q; W Jefferson, Franklin Co. (USNM). 1 q; South Dakota: Vermillion, 30/VI/1960, Walgenbach D.D. (IRCW). 1 q; Davis Ranch, NW Blanco Co., 22/IV/1959, Mason W.R.M. (CNC). 1 q; Edna, 19/VII/1908, Mitchell J.D. (USNM). 1 q; Texas: Sonora, Sutton Co., 19/V/1973, Menke \& Miller (USNM). 1 q; Virginia?: Va. (USNM). 1 ; Wisconsin: Columbia Co., 23/VII/1961, Carney Don (IRCW). 1 q; Dane Co., VIII/1899 (AEIC). 1 q; Dane Co., 10/VIII/1899 (AEIC). 1 + St. Croix Co., 2/VIII/1916, McNeel W. (AEIC).

## Crassomicrodus fulvescens (Cresson, 1865)

http://species-id.net/wiki/Crassomicrodus_fulvescens
Microdus fulvescens Cresson 1865: 297 [Examined].
Microdus medius Cresson 1865: 298 [Examined].

Holotype female. Col. No. 1727.1 (ANSP).
This species was recently investigated by Figueroa et al. (2008), who found that $C$. medius, based on males only, is conspecific with C. fulvescens, which was based entirely on females.

## Crassomicrodus jalisciensis Figueroa, Romero \& Sharkey, sp. n.

 urn:lsid:zoobank.org:act:E141B22C-E1E6-458A-A817-9550A6ED2FDA http://species-id.net/wiki/Crassomicrodus_jalisciensisFig. 6a-e

Description female. Body. Length. 7.35-7.50 mm. Color (Fig. 6e). Integument yellowish orange except black as follows, face, frons, gena temple, vertex, antenna, mandible apex, propleuron, ventral area of mesopleuron, apical area of hind tibia and tarsomeres; eye silver or blackish, ocelli translucent yellow; blackish; wing veins dark brown; forewing infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Sometimes trochanters blackish and/or propleuron yellowish orange. Head (Fig. 6ab). Triangular in frontal view; face without longitudinal ridge dorsomedially; eye height/width $=1.34-1.45$; eye height $0.59-0.61 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation; frons deeply excavated and crenulate with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior surface of antennal sockets rugulose; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.58-0.63 \times$ as long as eye height; clypeus $2.40-2.50 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 38-40 flagellomeres; setae at base of mandible slightly longer than setae on rest of body surface; face setose. Mesosoma (Fig.


Figure 6. Crassomicrodus jalisciensis. Female a anterior view of head, arrow indicates a median pyramidalshaped elevation b dorsal view of head, arrow indicates posterior surface of antennal sockets rugulose c lateral view of mesosoma, arrows indicate pronotum and subalar lobe separated from mesopleuron by wide groove $\mathbf{d}$ dorsal view of mesosoma $\mathbf{e}$ female habitus.

6cde). Pronotum strigose; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.13 to 0.15 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression punctulate; carinae of central metanotal area almost pentagonal shaped with the top inverted; propodeum reticulate rugulose; subalar lobe separated from mesopleuron by wide rugulose groove, width almost of similar size to subalar lobe; metapleuron reticulate-rugulose. Legs. Inner spur of middle tibia $0.69-0.78 \times$
length of basitarsus; inner spur of hind tibia $0.61-0.78 \times$ length of basitarsus; metabasitarsus $1.24-1.26 \times$ length of tarsomeres III, IV, and V combined; hind tibia $2.50-2.63 \times$ longer than basitarsus; hind femur length $4.54-4.76 \times$ its maximum width. Wings. Forewing length/width $=2.72-3.02$; stigma $3.45-3.57 \times$ longer than maximum width; forewing vein R1 $0.63-0.69 \times$ as long as vein RS; vein RS sinuate; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.09-0.11 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.76-4.11$; hind wing vein 1M 1.56-1.64× longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 5-6 hamuli. Metasoma. Apical width of petiole (tergum 1) 3.07-3.23× wider than basal width; minimum width of petiole $0.54-0.58 \times$ apical width; length of ovipositor sheath $0.30-0.33 \mathrm{~mm}$.

Male. Similar to female except color as follows: head, propleuron, pronotum, scutellum, metanotum, propodeum, mesopleuron, subalar lobe, metapleuron, coxae and trochanters black; inner spur of middle tibia almost half length of basitarsus ( $0.55 \times$ ).

Host. Unknown
Distribution. Mexico.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, posterior surface of antennal sockets rugulose, face setose, setae at base of mandible slightly longer than setae on rest of body surface, subalar lobe separated from mesopleuron by wide rugulose groove, and mesosoma mostly yellowish orange with wings infumate.

Remark. This species is near C. oaxaquensis, but differs in that C. oaxaquensis has the mesosoma black; wings hyaline; face with a weak longitudinal ridge dorsomedially; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles. One specimen of C. jalisciensis has the head and mesosoma black, but differs from C. oaxaquensis by leg and wing coloration.

Etymology. C. jalisciensis refers to the state of Jalisco, where all specimens have been found.

Material examined. Holotype $q$ : MEXICO, Jalisco: 9 miles W Tepatitlán, El Refugio, 3/VII/1953, C. Vaurie \& P. Vaurie. Allotype ot: same data as holotype. Paratypes: 2 q same data as holotype; Guadalajara, 1 \& 23-28/VII/1965, H.E. Evans (MCZ); 8 miles S Guadalajara, 1 Q 10/VII/1963, Parker F.D. \& L.A. Stange (USNM); Guadalajara, $2 \circlearrowleft^{\lambda}$ 16/VII/1951, $2 \circlearrowleft^{\top} 1$ q 17/VII/1951, Evans H.E. (AEIC). Holotype and allotype and paratypes with same data deposited in AMNH.

## Crassomicrodus mariae Figueroa, Sharkey \& Romero, sp. n. urn:lsid:zoobank.org:act:D958480B-3DC0-4377-95BB-AB49F0B381F8 <br> http://species-id.net/wiki/Crassomicrodus_mariae

Fig. 7a-e

Description female. Body. Length. 5.13-5.38 mm. Color (Fig. 7e). Integument black except yellowish-orange as follows, basal area of mandible, tegulae, femora, fore and middle tibia; medial area of hind tibia pale yellow; ocelli translucent yellow; apical
area of mandible reddish; tarsomeres and apical area of hind tibia blackish; tergum dark brown; sternum and wing veins brown; forewing lightly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig. 7ab). Transverse in frontal view; face with longitudinal ridge dorsomedially; eye height/width $=1.35-1.42$; eye height $0.67-0.72 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation; frons not excavated; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena bulging; malar space $0.38-0.43 \times$ as long as eye height; clypeus $2.53-2.67 \times$ wider than high; length of ventrolateral margin of clypeus distinctly longer than diameter of tentorial pit; antenna with 25 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 1cde). Pronotum punctuate; lateral pronotal margins with weakly crenulate groove; notauli not impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.09 to 0.10 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth centrally and microfoveolate on the margins; carinae of central metanotal area almost circular shaped; propodeum reticulate rugulose, more pronounced on lateral margins; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; ventral one-fourth of metapleuron reticulate punctuate, remainder smooth. Legs. Inner spur of middle tibia $0.67-0.74 \times$ length of basitarsus; inner spur of hind tibia $0.52-0.53 \times$ length of basitarsus; metabasitarsus $1.35-1.42 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.21-2.30× longer than basitarsus; hind femur length $3.14-3.23 \times$ its maximum width. Wings. Forewing length/width $=2.07-2.17$; stigma $2.65-2.75 \times$ longer than maximum width; forewing vein R1 $0.48-0.50 \times$ as long as vein RS; vein RS slightly sinuate; vein $r$ arising at middle of stigma; second submarginal cell triangular, with petiole $0.03-0.06 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ not pigmented throughout; hind wing length/width $=2.96-3.30$; hind wing vein $1 \mathrm{M} 2.00-2.35 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 4 hamuli. Metasoma. Apical width of petiole (tergum 1) 2.17-2.38× wider than basal width; minimum width of petiole $0.70-0.77 \times$ apical width; length of ovipositor sheath $0.76-0.78 \mathrm{~mm}$.

Male. Similar to female except male has 26-28 flagellomeres, carinae of central metanotal area almost pentagonal shaped, 4-5 hamuli.

## Host. Unknown

## Distribution. USA.

Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, frons not excavated, antenna with 25 flagellomeres, notauli not impressed, length of ovipositor sheath $0.76-0.78 \mathrm{~mm}$, forewing vein R1 $0.48-0.50 \times$ as long as vein RS, head and mesosoma black, and wings lightly infumate.

Remarks. This species is near to C. muesebecki, but differs in that C. muesebecki measures 6.08 a 6.95 mm , area between antennal sockets with a median transverse elevation and two weakly defined lateral tubercles, frons deeply excavated, antenna with 28-29 flagellomeres, pronotum more smooth, sparse setae on scutellar disc from 0.18


Figure 7. Crassomicrodus mariae. Female a anterior view of head, arrow indicates a median pyramidalshaped elevation $\mathbf{b}$ dorsal view of head, arrow indicates frons not excavated $\mathbf{c}$ lateral view of mesosoma, arrow indicates punctate pronotum d dorsal view of mesosoma, arrow indicates notauli not impressed e female habitus.
to 0.20 mm in length, length of ovipositor sheath $1.83-2.33 \mathrm{~mm}$, and coloration of metasoma is black.

Etymology. This species is named in honor of María Espinosa Morales, wife of the first author.

Material examined. Holotype $q$ : USA, California: 5 miles W Llano, 2/V/1937, Timberlake, deposited in USNM. Allotype ō: USA, Nevada: Patrick, Washoe Co., 22/

VI/1971, Bohart R.M., deposited in UCDC. Paratypes: USA, California: Apple Valley, 1 ơ 20/V/1955, Mason W.R.M. (CNC); Colton, 1 q Eddy F.A. (USNM); Hwy 76 at junc. to Mt. Palomar, San Diego Co., 1 ふ 26/VI/1976, Coville R.E. (EMEC); Mojave Desert Love Joy Butte, 1 q 10/V/1944, Melander A.L. (UCR); Sagehen Creek nr. Hobart Mills, Nevada Co., 1 ठ 24/VI/1964, Froebe J.A. (UCDC). Colorado: Limon, 1 § 16/VIII/1949, Dreisbach R.R. \& R.K. Schwab (USNM). Nevada: 15 miles E Reno, Nevada Co., 1 § 4/VI/1963, Irwin M.E.; 4 § same data as Allotype; 1 § same data as Allotype but collected by Grissell E.E. (UCDC). 2 miles Nixon Washoe Co., 1 \& 22/VI/1961, Parker F.D.; 12 miles NE Stillwater Churchill, 2 \& 3/VI/1961, Parker F.D.; Winnemucca, Humboldt Co., 1 q 15/VI/1960, Parker F.D.; Winnemucca, 1 \$ 30/V/960, Haig T.R. (USNM). Utah: Wild Horse Cr., N Goblin Valley, Emery Co., 1 q 3/VI/1982, 1494 m, Parker F.D. \& Griswold T. (CNC).

## Crassomicrodus melanopleurus (Ashmead, 1894)

http://species-id.net/wiki/Crassomicrodus_melanopleurus
Fig. 8a-e
Microdus melanopleurus Ashmead 1894: 125 [Examined]. Syn. n.

Holotype male. San Jose del Cabo. Cat. No. 223 (CAS)
Description female. Body. Length. $6.10-7.75 \mathrm{~mm}$. Color (Fig. 8e). Integument yellowish orange except ocelli translucent yellow reddish; antenna brown or black; eye, apical area of mandible and sometimes apical area of hind tibia and tarsomeres blackish; sometimes head and propleuron black; wing veins dark brown; forewing infumate with a hyaline spot on the first submarginal cell that is bigger than parastigma, sometimes forewing slightly infumate without distinguished the hyaline spot. Head (Fig. 8ab). Transverse in frontal view; face without longitudinal ridge dorsomedially; eye height/width $=1.38-1.39$; eye height $0.67-0.69 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles; frons excavated with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior area of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena bulging; malar space $0.48-0.57 \times$ as long as eye height; clypeus $2.40-2.55 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 35-38 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 8cde). Pronotum smooth; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.09 to 0.11 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth with punctures on the ventral margin; carinae of central metanotal area forming a triangular cell; propodeum reticulate rugulose, sometimes rugose; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter


Figure 8. Crassomicrodus melanopleurus. Female a anterior view of head, arrows indicate a median py-ramidal-shaped elevation with two weakly defined tubercles ventrolateral margin of clypeus, and tentorial pit $\mathbf{b}$ dorsal view of head, arrow indicates frons excavated with a pair of microfoveolate grooves $\mathbf{c}$ lateral view of mesosoma $\mathbf{d}$ dorsal view of mesosoma, arrow indicates impressed notauli $\mathbf{e}$ female habitus, arrow indicates inner spur of hind tibia.
than the subalar lobe; ventral three-fourths of metapleuron reticulate rugulose, remainder with punctures. Legs. Inner spur of middle tibia $0.77-0.86 \times$ length of basitarsus; inner spur of hind tibia $0.60-0.72 \times$ length of basitarsus; metabasitarsus $1.11-1.19 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.17-2.38× longer than basitarsus; hind femur length $3.50-3.85 \times$ its maximum width. Wings. Forewing length/ width $=2.72-2.76$; stigma $3.43-3.55 \times$ longer than maximum width; forewing vein R1 $0.61-0.67 \times$ as long as vein RS; vein RS not sinuate; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.07-0.15 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.60-4.10$; hind wing vein $1 \mathrm{M} 1.55-1.85 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $5-8$ hamuli. Metasoma. Apical width of petiole (tergum 1) 3.00-3.70× wider than basal width; minimum width of petiole $0.54-0.56 \times$ apical width; length of ovipositor sheath $0.20-0.33 \mathrm{~mm}$.

Male. Similar to female except color as follows: head, propleuron, mesopleuron, metapleuron, propodeum, coxa and trochanters black; sometimes coloration similar to female.

Host. Unknown.
Distribution. Mexico and USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, head transverse in frontal view, gena bulging, inner spur of middle tibia $0.77-0.86 \times$ length of basitarsus; inner spur of hind tibia $0.60-0.72 \times$ length of basitarsus, and body mostly yellowish orange with wings infumate.

Remarks. This species is difficult to circumscribe, the head shape and general coloration have a wide range of variation. We found some specimens with the head triangular in frontal view but they have the forewing infumate with a hyaline spot on the first submarginal cell, that occupies most of the space of the cell. Muesebeck (1927) suggested that the species could represent males of $C$. fulvescens. However the type of $M$. melanopleurus does not correspond to the characters of $C$. fulvescens, therefore we consider it to be a valid species. Crassomicrodus melanopleurus is very similar to C. fulvescens, but differs in that C. fulvescens has the gena distinctly bulging; frons deeply excavated; lateral scutellar depression rugose and foveolate; carinae of central metanotal defining an almost circular cell; inner spur of middle tibia $0.54-0.58 \times$ length of basitarsus; inner spur of hind tibia $0.48-0.54 \times$ length of basitarsus; and metapleuron completely reticulate-rugose.

Material examined. Holotype ${ }^{\lambda}$ : MEXICO, San José del Cabo (CAS). Other specimens examined.- MEXICO, San Luis Potosí: K398, 25 miles N Tamazuncha, 1 § 21/VIII/1960, Howden H. (CNC). USA, California: 15 miles E Baker, Cronese Wash, San Bernardino Co., 1 q 3 万 17/IV/1981, Pulawski W.J. (CAS); 15 miles W Baker, San Bernardino Co., 7 ¢ y 7 § 17/IV/1981, Bohart R.M. (UCDC). Banning, Riverside Co., 2 ठ 2/VII/1952, Grigarick A.A.; $1 \circlearrowleft^{\top}$ 2/VII/1952, Mathis H.L.; 2 q 27/VI/1952, Evans E.M.; 1 q 27/VII/1952, Barcus D.E.; 1 q 27/VI/1952, Miyagawa
S.; 1 Ø 28/VI/1952, Miyagawa S. (UCDC); 1 Ø 28/VI/1952, Nakata J.H.; 1 Ø 16/ VII/1950, Adelson B. (USNM). Cronese Valley, San Bernardino Co., 1 § 3/IV/1953, MacSwain J.W. (EMEC); 3 q 4 ठ 25/IV/1978, Smith N.J. (UCDC); 1 § 25/IV/1978, Smith N.J. (USNM). Del Puerto Cyn, Stanislaus Co., 1 ठ 12/VI/1978, Bohart R.M. (UCDC). Palm Springs Sta. Riverside Co., 5 đ 22/VII/1952, Menn J.J.; 1 ¢ y 3 ठ 21/VII/1952, MacSwain J.W.; 1 đ̃ 6/VII/1975, Linsley E.G. \& J.M. Linsley (EMEC); 1 đ 22/VII/1952, Menn J.J. (HIC); 1 § 19/V/1941, Knull D.J. \& J.N. Knull (OSU); 4 ¢ 22/VII/1952, Barcus D.E. (UCDC); 1 q y 2 ở 22/VII/1952, Menn J.J.; 1 ỏ 21/ VII/1952, Thompson D.S.; 1 o 22/VII/1952, Barcus D.E. (USNM). Tanbark Flat, Los Angeles Co., 2 \& 26/VI/1952, Anderson R.L.; Tanbark Flat, Los Angeles Co., 1 ふ 12/VII/1952, Grigarick A.A. (UCDC); 1 đ 23/VII/1952, Evans E.M. (USNM). Tracy, San Joaquin Co., 1 § 27/VII/1950, MacSwain J.W. (EMEC).

## Crassomicrodus muesebecki Marsh, 1960

http://species-id.net/wiki/Crassomicrodus_muesebecki
Fig. 9a-e
Crassomicrodus muesebecki Marsh 1960: 153-154 [Examined].

Holotype female. 7 miles Southwest of Trimmer, Fresno County, California [USA]. June 2, 1951. Cat. No. 64876 (USNM)

Description female. Body. Length. 6.08-6.95 mm. Color (Fig. 9e). Integument black except yellowish orange as follows, femora, one-fourth basal area of hind tibia, fore and middle tibia with its tarsomeres; medial areas of mandible yellow reddish; eyes silver or blackish; ocelli translucent yellow; medial area of hind tibia pale yellow, apical area of hind tibia with its tarsomeres blackish; wing veins dark brown; forewing slightly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig. 9ab). Transverse in frontal view; face with weak longitudinal ridge dorsomedially; eye height/width $=1.41-1.45$; eye height $0.61-0.62 \times$ inter-ocular distance; area between antennal sockets with a median transverse elevation and two weakly defined lateral tubercles, sometimes tubercles not defined; frons deeply excavated with two or more foveolae on center, posteriorly continue a pair of smooth groove that diverge towards the ocellar area, sometimes microfoveolate; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena distinctly bulging; malar space $0.46-0.50 \times$ as long as eye height; clypeus 2.30-2.50× wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 28-29 flagellomeres; setae at base of mandible similar on size than setae on rest of body surface. Mesosoma (Fig. 9cde). Pronotum smooth with abundant setae on pronotal groove and lateral pronotal area; lateral pronotal margins with weakly crenulate groove; notauli not impressed; anterolateral edges of scutellum lacking small acute projection; scutellar


Figure 9. Crassomicrodus muesebecki. Female a anterior view of head, arrows indicate a median transverse elevation with two weakly defined tubercles $\mathbf{b}$ dorsal view of head $\mathbf{c}$ dorsal view of mesosoma, arrow indicates notauli not impressed $\mathbf{d}$ lateral view of mesosoma $\mathbf{e}$ female habitus.
disc convex with sparse setae from 0.18 to 0.20 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression microfoveolate centrally, with rugosities and foveolae on the margins; carinae of central metanotal area almost circular shaped; propodeum reticulate rugulose, more pronounced on lateral margins; subalar lobe separated from mesopleuron by narrow rugose reticulate groove, width distinctly shorter than the subalar lobe; three-fourth dorsal area of metapleuron smooth, rest reticulate-punctuate. Legs. Inner spur of middle tibia $0.72-0.86 \times$ length of basitarsus; inner spur of hind tibia $0.59-0.67 \times$ length of basitarsus; metabasitarsus $1.03-1.18 \times$ length of tarsomeres III, IV, and $V$ combined; hind tibia $2.63-2.81 \times$ longer than basitarsus; hind femur length $3.09-3.33 \times$ its maximum width. Wings. Forewing length/width $=2.51-2.53$; stigma $2.62-3.00 \times$ longer than maximum width; forewing vein R1 0.59-0.64× as long as vein RS; vein RS not sinuate; vein $r$ arising at middle of stigma; second submarginal cell triangular, with
petiole $0.04-0.11 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ not pigmented throughout; hind wing length/width $=3.26-3.44$; hind wing vein $1 \mathrm{M} 1.66-1.76 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $4-5$ hamuli. Metasoma. Apical width of petiole (tergum 1) 1.78-2.11× wider than basal width; minimum width of petiole $0.67-0.70 \times$ apical width; length of ovipositor sheath $1.83-2.33 \mathrm{~mm}$.

Male. Similar to female.
Host. Unknown.
Distribution. USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median transverse elevation, gena distinctly bulging, setae at base of mandible similar on size than setae on rest of body surface, notauli not impressed, scutellar disc convex with sparse setae from 0.18 to 0.20 mm in length, length of ovipositor sheath $1.83-2.33 \mathrm{~mm}$, and body black with wings slightly infumate.

Remarks. C. muesebecki Marsh was described with observations on 14 specimens. In this revision, we included 13 of these, one homotype and 14 additional specimens. With the inclusion of these new specimens we confirm Marsh's (1960) observation that specimens from central and northern California have infumate wings, and in almost all the hind tibiae are yellowish orange, whereas the more southern specimens have hyaline wings and the medial areas of hind tibiae pale yellow. The species status of these two groups is tentative.

Material examined. Holotype $Q_{\text {: }}$ USA, California: 7 mi . SW Trimmer, Fresco Co. 2/VI/1951, Mac Neill C.D. (USNM). Paratypes revised: USA, California: Keen Camp, San Jacinto Mts., 2 O 31/V/1939, Bush W.C. (EMEC); 1 O 31/V/1939, Smith Ray F. (USNM). Ribbonwood, San Jacinto Mts., 1 q 21/V/1940, Michener C.D. (EMEC); 1 q 21/V/1940, Michener C.D. (USNM). Tuolumne County, 1 q 9/VI/1938, 1067 m., Hardman N.W. (EMEC); 1 q 9/ VI/1938, 1067 m., Hardman N.W. (USNM). Bucks Lake, Plumas County, 1 q 23/VI/1949, Cox D. (CAS). Bass Lake, Madera County, 1 ¢ 6/VI/1938, Bohart R.M.; Rucker Lake, Nevada Co., 1 q 5/VII/1949, Schlinger E.I.; Rumsey, Yolo Co., 1 Y 30/V/1956, Bohart R.M. (UCDC). Idyllwild, San Jacinto Mountains, 1 § 19/VI/1951, Bechtel G.C. (USNM). Homotype $q$ : Pinon Flat, San Jacinto Mts., 15/V/1941, Van Dyke E.C. (CNC). Other specimens examined.- California: Genoa, 3 ¢ 2 § 26/VI/1948, Townes H.M.G. \& D. Townes (AEIC); Keen Camp, 2 miles W San Jacinto Mts., 1 Q 31/V/1939, Laningham T.E.; Tuolumne County, 1 Ø 9/VI/1938, 1067 m. (EMEC); Ribbonwood, San Jacinto Mountains, 1 O 21/V/1940, Michener C.D. (HIC); Kelso Dunes, MNP, San Bernardino Co., 1 § 19/V/2001, 770 m., Hawks D. 34.53.20 N 115.43.02 W; Kelso Dunes, San Bernardino, 2 30/V/1999, Ballmer G.R. (UCR); Keen Camp, San Jacinto Mountains, 1 O 31/V/1939, Smith Ray F.; Westwood Hills, Los Angeles, 1 ठ 27/VII/1970, Linsley E.G. (USNM). Utah: Leeds Cyn. Washington Co., 1 § 30/V/1973, Torchio P. \& F. Parker (CNC).

## Crassomicrodus nigriceps (Cresson, 1872)

http://species-id.net/wiki/Crassomicrodus_nigriceps
Fig. 10a-e
Crassomicrodus nigriceps (Cresson): Muesebeck 1927: 21-22 [Examined]. Microdus nigriceps Cresson 1872: 182.
Crassomicrodus fenestratus Viereck 1913: 558-559 [Examined]. Syn. n.
Holotype female. Collection Belfrage. Cat. No. 1637 (USNM)
Description female. Body. Length. 6.05-9.50 mm. Color (Fig. 10e). Coloration of this species has a wide variation, there are specimens with the body totally dark to some areas yellowish-orange or yellow reddish; forewing infumate with a hyaline spot on the first submarginal cell, that occupies most of the space of the cell. Head (Fig. 10ab). Triangular in frontal view; face with weak longitudinal ridge dorsomedially; eye height/width $=1.48-1.50$; eye height $0.67-0.68 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles; frons excavated; posterior surface of antennal sockets smooth; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging, sometimes slightly bulging; malar space $0.51-0.64 \times$ as long as eye height; clypeus $1.88-2.21 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 37-43 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 10cde). Pronotum smooth, sometimes strigose; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.11 to 0.12 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth, sometimes with punctures on the ventral margins; carinae of central metanotal area forming a triangular cell; propodeum reticulate rugose; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; metapleuron reticulate rugulose. Legs. Inner spur of middle tibia $0.75-0.81 \times$ length of basitarsus; inner spur of hind tibia $0.59-0.62 \times$ length of basitarsus; metabasitarsus 1.21-1.29× length of tarsomeres III, IV, and $V$ combined; hind tibia 2.04-2.22× longer than basitarsus; hind femur length $4.17-4.35 \times$ its maximum width. Wings. Forewing length/ width $=2.70-2.76$; stigma $3.57-3.85 \times$ longer than maximum width; forewing vein R1 0.62-0.69× as long as vein RS; vein RS straight; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.07-0.22 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.88-4.09$; hind wing vein 1 M $1.48-1.71 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $6-8$ hamuli. Metasoma. Apical width of petiole (tergum 1) 3.33-3.43× wider than basal width; minimum width of petiole $0.49-0.60 \times$ apical width; length of ovipositor sheath $0.17-0.26 \mathrm{~mm}$.

Male. Similar to female.
Host. Unknown.


Figure 10. Crassomicrodus nigriceps. Female a anterior view of head, arrows indicate a median pyramidalshaped elevation with two weakly defined tubercles $\mathbf{b}$ dorsal view of head, arrow indicates posterior surface of antennal sockets smooth $\mathbf{c}$ lateral view of mesosoma, arrow indicates pronotum smooth $\mathbf{d}$ dorsal view of mesosoma $\mathbf{e}$ female habitus.

Distribution. Colombia, Costa Rica, Cuba, El Salvador, Guatemala, Haiti, Honduras, Mexico, Puerto Rico, Dominican Republic, and the USA.

Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, head triangular in frontal view, face with a few sparce setae, setae at base of
mandible distinctly longer than setae on rest of body surface, antenna with 37-43 flagellomeres, body usually with some areas yellowish-orange and wings infumate.

Remarks. This species was described as Microdus nigriceps by Cresson (1872). Muesebeck (1927) considered it as valid species in the genus Crassomicrodus; however due to the wide range of color variation among specimens of this species, it is described to C. fenestratus as a different species, but the types of both species have affinity of characters. C. nigriceps is the most variable species, on coloration and measurements, that all species of Crassomicrodus. Our careful examination of each specimens allowed to group them in three sub-groups; the first sub-group formed by specimens from Texas (USA), of size 7.30 to 9.50 mm and the body color yellowish-orange or yellow reddish except head, propleuron, metapleuron, propodeum, ventral area of mesopleuron, and legs black; the coloration in mesosoma of this sub-group could vary to only propleuron black and some areas of legs yellowish-orange. The second sub-group is formed by specimens from the Yucatan peninsula (Mexico), of size 6.05 to 7.95 mm and the coloration blackish on all body, although the coloration could vary to only mesoscutum yellowish-orange or yellow reddish. The last sub-group is formed by specimens from Loggerhead Key island (Florida, USA), of size 6.80 to 7.30 mm and the coloration yellowish-orange or yellow reddish except head, propleuron, metapleuron and propodeum black, ventral area of mesopleuron and legs blackish, although the coloration on mesopleuron and legs could vary. The last sub-group is where the majority of the specimens that come from other places have more affinity.

Material examined. Type $P$ : M. nigriceps, collection Belfrage (USNM). Type $q_{q}$ : C. fenestratus Porto Rico, C.W. Hooker (USNM). Homotypes $q$ : HONDURAS, [Siguatepeque]: 30 km SE Siguatepeque, 11-12/VIII/1978, Chemsak J.A., E.G. Linsley \& J.M. Linsley (CNC). ${ }^{\top}$ : MEXICO, Oaxaca: 23 miles S Matías Romero, 6/ IV/1962, Parker F.D. (CNC). $\uparrow$, ${ }^{\top}: 27$ miles SW Salina Cruz, 14/VII/1987, Wharton R. (TAMU). ${ }^{\text {on }}: 63$ miles E Tehuantepec, 16/IX/1967, 152 m., Painter E.M. \& R.H. Painter (CNC). $\uparrow$ : NICARAGUA, 1 miles SW Managua, 4/IX/1967, 2200 m., Painter R.H. \& E.M. Painter (AEIC). Other specimens examined.- COLOMBIA, Tolima: Armero, 1 q 30I-5II/1977, Peyton E.L. (USNM); 1 q 26-30/1977, 1 § 26-30/1977, Peyton \& Suarez (HIC, USNM). COSTA RICA, [Limón]: Limon, 1 § 15/II/1964, Evans H.E. (MCZ). Guanacaste: 10 km E Paseo Tempisque, 1 \& 30/VII/1990, Chamberlain W.F. (TAMU); Sector Santa Rosa; open field nr. road to Playa Naranjo, 1 \& 8-18/VI/1995, Dadelahi-Price \& Zitani (ESUW). CUBA, Santiago de las Vegas 1 đ (USNM). [Ciego de Avila]: Baraguá, 1 ¢ 19/X/1927, Scaramuzza L.C. (USNM). [Cienfuegos]: Ciego Montero, Cruces, L.K., $1 \overbrace{}^{\top}$ XII/1917, Alayo P. (HIC); Baraguá, 1 q 27/VIII/1927, Scaramuzza L.C. (MCZ). Pinar del Rio: San Vicente, 1 q 26VII5VIII/1939, Parsons C.T. (MCZ). DOMINICAN REPUBLIC, [San Cristóbal]: San Cristobal Prov., 1 § 5/VIII/1967 (TAMU). EL SALVADOR, [San Salvador]: 6 miles W Quezaltepeque, 1 \& 2 § 12/VIII/1963, 500 m., Cavagnaro D.Q. \& M.E. Irwin (CAS); 6 miles W Quezaltepeque, 1 \& 15/VII/1963, 500 m., Cavagnaro D.Q. \& M.E. Irwin (CAS); Sonzacate, 1 đ̋ 25/VI/1958, Bottimer L.J. (HIC). GUATEMALA. [Chiquimula]: Mocá, Guatalón, 1 \& III-IV/1931, 1000 m., Bequaert J. (MCZ).
[Guatemala]: Amatitlán, 1 \& 1 ô 6/VII/1947, 1219 m., Vaurie C. \& P. Vaurie (AMNH); 6 miles N Amatitlan, 1 ô 29/XI/1972, Dasch B. \& C. Dasch (AEIC). Zacapa: Rio Hondo, 3 Q VIII/1987, Sharkey M.J. (CNC, HIC). HAITI, Fond Parisien, 2 § 11-18/II/1922, 18 m. (AMNH); Trou Caiman, 1 q 1 § 4/IX/1934, Bates M. (MCZ). HONDURAS: Nr. Corozal Brit., 1 q 18/VII/1963, Porter C. (MCZ). [Colón]: Puerto Castilla, 1 ô 21/III/1924 (USNM). El Paraíso: Galeras, 1 § 31/ VII/1992, Stange L.; Yuscarán, 1 ¢ 4/VIII/1992, 840 m., Porter C. \& L. Stange (FSCA). La Paz: La Paz, 1 đ̋ 23/VI/1979, Chemsak J.A., A. Michelbacher, M. Michelbacher \&W.W. Middlekauff (EMEC). MEXICO, Baja California Sur: 51 km W La Paz (km 51), 1 ¢ 26/VIII/1977, 275 m., Fisher E. \& R. Westcott; 25 miles W La Paz, 1 đ 30/VIII/1959, Radford K.W. \& Werner F.G. (CAS). Campeche: Champoton, 1 ठ 8/VII/1964, Pallister J.C. \& D. Pallister (AMNH). Chiapas: N. Chiapas, 3 km S Oaxaca Rte. 190, 1 ふ 12/VIII/1962, 1524-1829 m., Milliron H.E. (CNC); 7 miles SE Soyalo, 1 đ 27/III/1953, Bechtel R.C. \& E.I. Schlinger; Santo Domingo, 15 miles SE Simojovel, 1 \& 8-15/VII/1958, Smith R.F.; Suchiapa, 1 q 18/VII/1957, Hurd P.D. (EMEC); Km 47 Tuxtla Gutierez-Venustiano Carranza, 1 § 10/VII/1988, 480 m., Cadena A. \& L. Cervantes (IBUNAM). Coahuila: 6 miles W Saltillo, 1 § 14/ VII/1972, 5200 ft., Dasch B. \& C. Dasch (AEIC). Colima: Armeria, 1 ð 1/VIII/1954, Cazier M. y W. Gertsch Bradts (AMNH); 21 miles NW Manzanillo, 2 đ 30/VIII/1970, Wasbauer M.S. \& J.S. Wasbauer, malaise trap; Playa de Oro Rd. NW Manzanillo, 1 q 1 đ 31/VIII/1970, Wasbauer J.S. (EMEC). Guerrero: Xalitla, 1 ठ 20/III/1959, 457 m., Evans H.E. \& Anderson D.M. (CUIC). Jalisco: Chamela, 1 \& 26-30/IX/1985, Parker F.D. \& T.L. Griswold; El Tuito, 1 § 2/X/1985, Parker F.D. \& T.L. Griswold; Guadalajara, 1 Q VIII/1962, Hull Frank M. (CNC); 22 km E El Grullo, 2 q 17/ VII/1989, Griswold T., 1200 m.; Chamela Res. Sta., 1 \& 24/VII/1986, Sanchez MM.T. (HIC); Rio Santiago, 15 miles N Guadalajara, 1 ठ 22/VII/1965, Evans H.E. (MCZ); 6 miles N Autlan, 1 万7/VII/1984, Shaffner, Woolley, Carroll \& Friedlander (TAMU); Tequila, 1 q 6/VII/1956, Dreisbach R. \& K. Dreisbach (USNM). Michoacán: 11 miles E Apatzingan, 3 Q 20/VIII/1954, Linsley E.G., J.W. MacSwain \& Smith R.F. (EMEC). Morelos: 3 miles N Alpuyeca, 1 q 4/III/1959, 1036 m., Evans H.E. \& Anderson D.M.; Las Estacas, 1 ठ 6/IV/1959, 914 m., Evans H.E. (CUIC). Nuevo León: Apodaca, 1 Q 27/VII/1963, Howden H. \& A. Howden (CNC); Carr. Miguel Aleman km 15, Apodaca, $1 \delta^{\lambda} 10 / \mathrm{VIII} / 1984$, Vera A. (CIBE-UANL). Oaxaca: Tehuantepec, 1 ठ 23/XI/1972, Dasch B. \& C. Dasch (AEIC); 23 miles S Matias Romero, 2 § 14/VIII/1963, Parker F.D. \& L.A. Stange (HIC, USNM); 23 miles S Matias Romero, 1 ठ 22/IV/1962, Parker F.D.; El Camaron, 1 ठ 24/IV/1962, Parker F.D. (USNM); Puebla: 12 miles N Chapulco, 1 q 29/VII/1963, Caltagirone L.E. (EMEC). Quintana Roo: Felipe Carrillo Puerto, 4 q 10-14/X/1986, 19.35 N 88.03 W, malaise trap; 1 ô 10-14/X/1986, Parker F.D., 19.35 N 88.03 W; 1 q 2 ô 12-14/ X/1986, Griswold T., 19.35 N 88.03 W (CNC). 25 km W Felipe Carrillo Puerto, 1 q 15/X/1986, Parker F.D., 19.35 N 88.17 W; 3 km SW Puerto Morelos, 1 q 1 ő 7/X/1986, Parker F.D.; Coba, 2 § 5/X/1986, Griswold T., 20.36 N 87.35 W (CNC). Vallarta, 17 km W Puerto Morelos, 5 đ 6-8/X/1986, Griswold T., 2 đ 6-8/X/1986,

Parker F．D．，20．50 N 87．00 W（CNC）； 1 q 6－8／X／1986，Griswold T．（HIC）．Sinaloa： 34 miles N Los Mochis， 1 § 27／VIII／1963，Parker F．D．\＆L．A．Stange（HIC）；Maza－ tlán， 8 § 15－20／VIII／1962，Evans H．E．（MCZ）；Santa Ana， 1 q 2／VIII／1985，Ekis G． （UCDC）；Concordia， 1 §／VII／1963，Parker F．D．\＆L．A．Stange（USNM）．Sonora： Santa Ana，riv．， 1 § 4／VIII／1985，Ekis G．（UCDC）； 19.4 miles S Estacion Llano， 2 q 25／VIII／1964，Schlinger E．I．， $700 \mathrm{~m} .((\mathrm{USNM})$ ．Veracruz：Lake Catemaco， 1 ठ 8－16／VIII／1960，Howden H．F．（CNC）；Puente Nacional， 6 miles SE Rinconada， $1 \sigma^{\top}$ 30／IX／1975，Chemsak J．（EMEC）；Veracruz， 1 \＆ 1 § 28VII－11VIII／1956，Dreisbach R．\＆K．Dreisbach（MSUC）．Yucatán：Uxmal， 1 q 16－18／VI／1959，Vaurie P．\＆C． Vaurie（AMNH）；Mérida， 2 \＆22－25／VII／1962，Evans H．E．（ESUW）； 8 miles E Me－ rida， 1 § 28／VI／1966（HIC）；Chichén Itzá， 1 § 19／VII／1962，Evans H．E．；Mérida， 2 ㅇ 1 § 22－25／VII／1962，Evans H．E．；Uxmal， 1 q 22／VII／1962，Evans H．E．（MCZ）； 9 miles N Uxmal， 1 § 1／VIII／1980，Schaffner，Weaver，Friedlander；Xmatkuil，Méri－ da， 1 q 25－28／V／1996，Wharton \＆León，malaise trap（TAMU）． 9 km N Teya Pueb－ lo， 1 q 1／IX／1999， 1 ő 12／X／1999， 2 đ 14／IX／1999， 1 q 2／IX／1999， 1 q 3 o 3／ VIII／1999，Suárez C．（UADY）．Tamaulipas：Mesa de Llera，ca．Ciudad Victoria， 1 q 1／VI／1977，Porter C．\＆A．Cerbone（FSCA）．PUERTO RICO，Coamo Springs， 1 § 1／VII／1915；Mayaguez， 1 中 21－23／VI／1915（AMNH）；Lake Guanica，P．R．， 5 o 30／ VI／1936，Dozier H．L．；Guanica， 2 q 29／VI／1914，Smith；Lajas， 1 ō X－XI／1960， Cotte R．（USNM）．USA，Arizona： 1 § 12／VIII／1974，Townes H．\＆M．Townes；Por－ tal， 1 \＆17／VIII／1974，Townes H．\＆M．Townes（AEIC）；Phoenix， 1 \＆2／VI／1942 （CAS）； 5 miles E Nogales，Santa Cruz Co．， 1 q 1／IX／1970，Bohart G．E．\＆R．M．Bo－ hart（CNC）； 10 miles SW Patagonia，Santa Cruz Co．， 1 q 13／IX／1958，Cazier M．A． （EMEC）．Madera Cyn．，Santa Cruz Co， 1 q 31／VII／1966， 1487 m．，Kovacic C．R．； Box Cyn，Santa Cruz Co．， 1 q26／VIII／1978，Meyer R．P．（UCDC）．Wild Morning Glory， 2 ¢ 1957，Lochiel；W sl．Patagonia Mountains， 4 q 9／VIII／1908，Butler G．D． \＆F．G．Werner（USNM）．Florida：S．Miami 1 Q（MCZ）．Isla Loggerhead Key，Dry Tortugas， 10 § 1／IX／1961， 1 ¢ 4 ふ 2／IX／1961，Weems H．V．Jr．； 2 miles NW Orange Spg．，Putnam Co．， 1 Q 13X－5XI／1975，Wiley J．，malaise trap；Boca Chica Key，Mon－ roe Co．， 2 §11／VII／1971，Pierce W．H．；Clermont，Lake Co．， 1 q 29／VIII／1983，No－ lfo V．（FSCA）．Highlands Hamm．St．Pk．Fla．， 1 ठ 26／III／1963，Zeiger C．F．；Key Largo， 1 \＆2／V／1957，Weems H．V．（HIC）．Georgia：St．Catherines Island，Liberty Co．， 1 § 22／V／1973，Rozen J．G．（AMNH）．Louisiana：Gilliam， 61 §／IX／1907， Bishopp F．C．（USNM）．North Carolina：Jacksonville，Onslow Co．， 1 \＆3／IX／1963， Bohart G．E．（CNC）．Texas：Austin， 1 § 26／VI／1922（AMNH）； 1 Q X／1899（MCZ）； 1 đ 26／VI／1922，Cazier M．A．（KSUC）； 1 中 IX／1926，Bishopp F．C．（USNM）；Bent－ sen Rio Grande State Park，Hidalgo Co．， 1 ¢ 6－8／VI／1983，Pulawski W．J．（CAS）． Brownsville，Cameron Co．， 1 §11／IV／1976，Bruce Tilden；Sullivan City，Hidalgo Co．， 1 § 10／IV／1976（CAS）；Salado Creek，Bexar Co．， 1 § 13／III／1952，Wawbauer M． （EMEC）．Fleming Key，Monroe Co．， 1 q 7／VIII／1979， 1 § 8／VIII／1979，Acree John A．\＆H．V．Weems Jr．， 1 q 8－9／III／1980，Williams H．E．\＆H．V．Weems Jr．（FSCA）． Valley Botanical Garden，McAllen，Hidalgo Co．， 1 ठ 12／VII／1980， 1 ð 23／VIII／1980， Porter C．C．（FSCA）．Valley State Park，Bentsen Rio Grande，Hidalgo Co．， 1 § 4／

VI／1982， 1 § 1／VI／1979， 3 § 11／VI／1982， 3 § $14 / \mathrm{VI} / 1982,4$ § 16／VI／1983， 1 q $2 /$ VIII／1980， 1 中 1 ठ 30／V／1979， 2 中 3 ठ 31／V／1979， 1 ð 4／VI／1983， 1 中 1 ठ 6／ VI／1982，Porter C．（FSCA）．Valley State Park，near Mission，Bentsen Rio Grande， Hidalgo Co．， 2 中 6 đ 1－25／VIII／1980， 2 đ 10／VI／1981， 1 q 1 ठ 12／VI／1981， 1 ठ 12／VIII／1983， 1 Q 2 § 17／VI／1983， 1 § 4／VI／1981， 2 § 6／VI／1981， 1 § 9／VI／1981， Porter C．（FSCA）．McAllen Botanical Garden，Hidalgo Co．， 1 ¢ 31／V／1982， 1 ő 31／ VI／1982，Porter C．（FSCA）．McAllen Botanical Garden，McAllen，Hidalgo Co．， 1 q 1／VII／1985， 1 q 15／VI／1985， 1 q 2／VII／1985， 1 q 20／VI／1984， 1 q 30／VI／1985， 1 \＆6／VII／1983，Porter C．（FSCA）．San Antonio， 1 q 11／IV／1942，Melander A．L．； Wharton，Wharton Co．， 1 ð 24／VI／1917（MCZ）．Dallas， 1 q 14／IX／1905，Jones C．R．；Galveston［Galveston Co．］， 1 ふ 29／VII／1924，Tretter；Garrett， 1 ð 21／VII／1908， Tucker E．S．；Plano［Collin Co．］， 2 \＆ $1 \delta$ X／1907，E．S．Tucker．Paris［Lamar Co．］， 1 q 27／VIII／1905， 1 đ 26／VIII／1905，F．C．Bishopp．Wolfe City， 1 \＆20／V／1907， 1 ठ 4／ VI／1909，F．C．Bishopp；Victoria， 1 đ 25／IX／1906，Crawford J．C．；Victoria，Victoria Co．， 1 ठ 25／VI／1917；Waco， 1 ¢ 17／II／1939，Jones C．R．（USNM）．

## Crassomicrodus nigrithorax Muesebeck， 1927

http：／／species－id．net／wiki／Crassomicrodus＿nigrithorax
Fig．11a－e
Crassomicrodus nigrithorax Muesebeck 1927：17－18［Examined］．

Holotype female．Colorado［USA］．Cat．No． 28694 （USNM）
Description female．Body．Length．3．95－5．35 mm．Color（Fig．11e）．Integument black except yellowish as follows，three－quarter of the basal area of mandible，tegulae， femora，three－quarter of the basal area of hind tibia，fore and middle tibia，fore and middle tarsomeres，and metasoma；ocelli translucent yellow；eyes silver or blackish； wing veins clear brown；forewing slightly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma．Sometimes hind coxa and trochanters yellowish－orange and／or fore and middle femora，apical area of tibia，and fore and middle tarsomeres blackish，rarely tegula blackish．Head（Fig．11ab）．Trans－ verse in frontal view；face with longitudinal ridge dorsomedially；eye height／width＝ $1.30-1.35$ ；eye height $0.69-0.70 \times$ inter－ocular distance；area between antennal sock－ ets with a median transverse elevation and two weakly defined lateral tubercles；frons excavated with a central groove almost foveolate，sometimes a pair of microfoveolate groove that diverge towards the ocellar area；posterior surface of antennal sockets smooth；groove between lateral ocelli microfoveolate；median ocellus separated from lateral ocellus by microfoveolate groove；gena bulging；malar space $0.38-0.47 \times$ as long as eye height；clypeus $2.67-2.91 \times$ wider than high；length of ventrolateral margin of clypeus similar to diameter of tentorial pit；antenna with 28－31 flagellomeres；setae at base of mandible slightly longer than setae on rest of body surface．Mesosoma（Fig． 11 cde ）．Pronotum smooth except near of subpronope rugulose；lateral pronotal mar－


Figure II. Crassomicrodus nigrithorax. Female a anterior view of head, arrows indicate a median transverse elevation with two weakly defined tubercles $\mathbf{b}$ dorsal view of head $\mathbf{c}$ dorsal view of mesosoma, arrow indicates notauli impressed $\mathbf{d}$ lateral view of mesosoma $\mathbf{e}$ female habitus.
gins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection, sometimes slightly the projection; scutellar disc convex with sparse setae from 0.14 to 0.15 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression rugose and foveolate on the margins and microfoveolate centrally; carinae of central metanotal area almost circular shaped; propodeum reticulate rugulose more pronounced on lateral margins, anterolateral areas with abundant setae; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; metapleuron reticulate rugulose or punctulate, more pronounced on ventral half. Legs. Inner spur of middle tibia $0.89-0.95 \times$ length of basitarsus; inner spur of hind tibia $0.63-0.76 \times$ length of basitarsus; metabasitarsus 1.15-1.26x length of tarsomeres III, IV, and V combined; hind
tibia $1.92-2.16 \times$ longer than basitarsus; hind femur length $3.23-3.38 \times$ its maximum width. Wings. Forewing length/width $=2.46-2.60$; stigma $2.95-3.08 \times$ longer than maximum width; forewing vein R1 $0.65-0.68 \times$ as long as vein RS; vein RS sinuate; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.03-0.12 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/ width $=3.45-3.58$; hind wing vein $1 \mathrm{M} 1.83-1.97 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 3-4 hamuli. Metasoma. Apical width of petiole (tergum 1) $2.80-2.82 \times$ wider than basal width; minimum width of petiole $0.64-0.70 \times$ apical width; length of ovipositor sheath $0.19-0.22 \mathrm{~mm}$.

Male. Similar to female.
Host. Unknown.
Distribution. Mexico and USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median transverse elevation, head transverse in frontal view, grooves between ocelli microfoveolate, gena bulging, antenna with 28-31 flagellomeres, anterolateral edges of scutellum usually lacking small acute projection, head and mesosoma black, and metasoma yellowish.

Remarks. This species is near to C. apicipennis, but differs in that C. apicipennis has areas of mesosoma yellowish-orange; anterolateral edges of scutellum has small acute projection; eye height $0.65-0.68 \times$ inter-ocular distance; malar space $0.48-0.55 \times$ as long as eye height; inner spur of middle tibia $0.76-0.83 \times$ length of basitarsus; and scutellar disc convex with sparse setae from $0.16-0.17 \mathrm{~mm}$ in length.

Material examined. Holotype $q$ : Colo (USNM). Allotype ${ }^{\top}$ : Colo (USNM). Homotype $q$ : MEXICO, Guerrero: 4 miles S Taxco, 8/VIII/1954, 1463 m., Chillcott J.G. (CNC). Other specimens examined.- MEXICO, Baja California Sur: Las Animas, Sierra Laguna, 1 ठ 1 ¢ 12/X/1941, Ross \& Bohart (CAS). Guerrero: 17 miles NW San Marcos, 1 § 13/VII/1966, Wagner P.M. \& P.K. Wagner (TAMU). Jalisco: Chamela, PT, 1 § 4-9/VII/1993, Sharkey M.J. (CNC). Morelos: Yautepec, 1 q 23/ VII/1963, Parker F.D. \& L.A. Stange (HIC). Nayarit: 18 miles NW Ixtlán del Río, 1 § 25/VII/1966, Wagner P.M. \& P.K. Wagner (TAMU). Nuevo Leon: 5 miles S Monterrey, 1 § 1/VII/1963, Howden H. \& A. Howden (CNC). Oaxaca: 27 miles SW Salina Cruz, 3 \& 14/VII/1987, Wharton R. (TAMU); 3 miles W Oaxaca, 1 § 21/XI/1972, Dasch B. \& C. Dasch (AEIC). Puebla: 11 miles SE Acatlan, 1 o 10/VII/1952, Gilbert E.E. \& C.D. McNeil; 5 miles E Tepexco, 1 ゐ 24/VIII/1977, 1250 m., Schlinger E.I. (EMEC); 3 miles N Petalcingo, 1 ¢ 2 § 3/VIII/1963, Parker F.D. \& L.A. Stange; Petalcingo, 1 Y/VIII/1963, Parker F.D. \& L.A. Stange (USNM). San Luis Potosí: 6 miles S Ciudad de Valles, 1 § 21/VII/1954, 61 m., Chillcott J.G. (CNC); Valles, 1 Ø 21/VII/1946, Pallister J. \& D. Pallister (AMNH). Sinaloa: Cutiacan, 1 § 11/ IX/1970, Bohart G.E. \& R.M. Bohart (HIC). Tamaulipas: Est. Carboneros Guemez, $1 \overbrace{}^{\top}$ 8/IX/1988, Loyola J.C. (CIBE-UANL); 44 miles W Tampico, 1 q 22/VIII/1967, Hevel Gary F. (USNM). Tlaxcala: 21 miles W Apizaco, 1 \& 20/VIII/1958, Howden H.F. (CNC). Veracruz: 10 miles SW Perote, 1 § 27/VII/1974, Clark-Murria-Ashe $\&$ Schaffner (TAMU). USA, Arizona: 13 miles SW Apache, Cochise Co., 1 § 13/

VIII/1970, Rozen J.G. (AMNH); Santa Cruz Sycamore Cyn. 9 miles W Peña Blanca Lk., 7 ¢ 12 ठ 12/VIII/1983, 1250 m., Anderson R. (CNC); Tucson Mountains, 1 q $1 \circlearrowleft^{\top} 16 / \mathrm{VIII} / 1955$, Butler G.D. (USNM). California: Big Flat, Coffee Creek, Trinity Co. 1 q 21/VI/1934, Van Dyke E.C. (CAS); 6 miles W Tragedy Spr., Amador Co., 1 O16/VII/1960, Rice R.E.; Donner Pass, 1 § 1/VIII/1948, Townes H.M.G. \& D. Townes (HIC); Westgard Pass, Inyo Co., 2 Q 16/V/1979, Bohart R.M.; 5 miles E Woodland, Yolo Co., 1 ¢ 10/IX/1970 (UCDC); Yuba, Sierra Co. 1 đ 6/VII/1962, Irwin M.E.; Quatal Canyon, NW corner Ventura Co., 1 q 1 § 9/V/1959, Powell J.; Davis, 1 § 10/V/1960, Parker F.D. (USNM). Colorado: Colo 1 q (HIC); New Mexico: Hatch, 1 \& 27/VIII/1974, Townes H. \& M. Townes (CNC); Hatch, 1 \& 30/ VIII/1974, Townes H. \& M. Townes (AEIC). Oregon: Cave Jct., 1 § 27/V/1978, Townes H. \& M. Townes (AEIC). Utah: Strbry Daniel Pass, 1 \& 18/VI/1948, Townes H.M.G. \& D. Townes (AEIC).

## Crassomicrodus oaxaquensis Figueroa, Romero \& Sharkey, sp. n.

 urn:lsid:zoobank.org:act:EF0FBCCA-7C2F-4AA6-8C0F-A4DAE53AEBF2 http://species-id.net/wiki/Crassomicrodus_oaxaquensisFig. 12a-e

Description female. Body. Length. $7.10-7.65 \mathrm{~mm}$. Color (Fig. 12e). Integument black except yellowish-orange as follows, ocelli, fore tibia, two-thirds apical areas of fore and middle femur, two-thirds basal areas of middle tibia and metasoma; medial area of mandible yellow reddish; eyes silver; wing veins dark brown; forewing almost hyaline. Sometimes first metasomal tergite black, and the yellowish-orange of legs is reduced to only the apical area of fore and middle femora and apical area of fore tibia. Head (Fig. 12ab). Triangular in frontal view; face with weak longitudinal ridge dorsomedially; eye height/width $=1.34-1.45$; eye height $0.60-0.62 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles; frons excavated with a pair of microfoveolate grooves that diverge towards the ocellar area; posterior surface of antennal sockets rugulose; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.55-0.58 \times$ as long as eye height; clypeus $2.25-2.44 \times$ wider than high; length of ventrolateral margin of clypeus almost similar to diameter of tentorial pit; antenna with 38-39 flagellomeres; setae at base of mandible slightly longer than setae on rest of body surface; face very setose. Mesosoma (Fig. 12cde). Pronotum punctate; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc slightly convex with sparse setae from 0.15 to 0.16 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression rugulose and punctate; carinae of central metanotal area forming a triangular cell; propodeum reticulate rugulose with abundant sparse setae on lateral areas; subalar lobe separated from mesopleuron by wide rugose groove, width almost of similar size to subalar lobe; metapleuron reticulate rugulose in its ventral half


Figure 12. Crassomicrodus oaxaquensis sp. n. Female a anterior view of head, arrows indicate a median pyramidal-shaped elevation with two weakly defined tubercles $\mathbf{b}$ dorsal view of head, arrow indicates posterior surface of antennal sockets rugulose $\mathbf{c}$ lateral view of mesosoma, arrow indicates subalar lobe separated from mesopleuron by wide groove $\mathbf{d}$ dorsal view of mesosoma $\mathbf{e}$ female habitus.
and smooth or punctuate in its dorsal half. Legs. Inner spur of middle tibia 0.68-0.73x length of basitarsus; inner spur of hind tibia $0.58-0.66 \times$ length of basitarsus; metabasitarsus $1.25-1.32 \times$ length of tarsomeres III, IV, and $V$ combined; hind tibia 2.33-2.38x longer than basitarsus; hind femur length $4.64-4.76 \times$ its maximum width. Wings. Forewing length/width $=2.57-2.76$; stigma $2.91-3.33 \times$ longer than maximum width; forewing vein R1 $0.65-0.70 \times$ as long as vein RS; vein RS sinuate; vein $r$ arising before middle of stigma; second submarginal cell triangular, with petiole $0.08-0.09 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.70-3.73$; hind wing vein 1 M 1.79-1.82× longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 5 hamuli. Metasoma.

Apical width of petiole (tergum 1) 3.00-3.41× wider than basal width; minimum width of petiole $0.46-0.47 \times$ apical width; length of ovipositor sheath $0.22-0.26 \mathrm{~mm}$.

Male. Similar to female except metasomal tergite has the color dark brown, antenna with 39 or 40 flagellomeres, and hind wing with 4 or 5 hamuli.

Host. Unknown.
Distribution. Mexico.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, head triangular in frontal view, posterior surface of antennal sockets rugulose, face very setose, setae at base of mandible slightly longer than setae on rest of body surface, subalar lobe separated from mesopleuron by wide rugose groove, head and mesosoma black, and wings almost hyaline.

Remarks. This species is near to C. jalisciensis, but differs in that C. jalisciensis has areas of mesosoma yellowish orange, wings infumate, face without longitudinal ridge dorsomedially, and a median elevation between antennal sockets without defined lateral tubercles.

Etymology. C. oaxaquensis is after Oaxaca, in reference to the known geographical distribution of the species.

Material examined. Holotype $q$ : MEXICO, Oaxaca: Llano de las Flores, 15 miles NE Ixtlán de Juárez, 21/VII/1985, Woolley \& Zolnerowich. Allotype ${ }^{\text {J }}$ : same data as holotype. Paratypes 1 , $2 \widehat{\sigma}^{\lambda}$ : same data as holotype. All types deposited in TAMU.

## Crassomicrodus olgae Figueroa, Sharkey \& Romero, sp. n.

urn:lsid:zoobank.org:act:B4527E95-134E-4DD0-B8B3-00637C850D46
http://species-id.net/wiki/Crassomicrodus_olgae
Fig. 13a-e

Description female. Body. Length. 6.70-7.08 mm. Color (Fig. 13e). Integument black except yellowish-orange as follows, medial area of mandible, femora, fore tibia, basal half of middle and hind tibia, and metasoma; ocelli translucent yellow; wing veins brown; forewing slightly infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig. 13ab). Transverse in frontal view; face with longitudinal ridge dorsomedially; eye height/width $=1.38-1.39$; eye height $0.68-0.74 \times$ inter-ocular distance; area between antennal sockets with a median trapezoidal-shape elevation and two weakly defined tubercles; frons deeply excavated; posterior surface of antennal sockets smooth; groove between lateral ocelli slightly microfoveolate; median ocellus separated from lateral ocellus by smooth groove; gena distinctly bulging; malar space $0.54-0.59 \times$ as long as eye height; clypeus $2.08-2.19 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 32 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 13cde). Pronotum with the pronotal groove reticulate rugulose and lateral areas smooth; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum with


Figure I3. Crassomicrodus olgae sp. n. Female a anterior view of head, arrows indicate a median elevation in trapezoidal shape with two weakly defined tubercles $\mathbf{b}$ dorsal view of head $\mathbf{c}$ lateral view of mesosoma $\mathbf{d}$ dorsal view of mesosoma, arrow indicates notauli impressed $\mathbf{e}$ female habitus.
slight acute projection, sometimes without projection; scutellar disc convex with sparse setae from 0.16 to 0.17 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression with punctures centrally and foveolae in its margins; carinae of central metanotal area almost circular shaped; propodeum reticulate rugose, more pronounced on lateral margins; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; metapleuron reticulate rugulose or foveolate in its ventral half and smooth in its dorsal half. Legs. Inner spur of middle tibia $0.85-0.92 \times$ length of basitarsus; inner spur of hind tibia $0.58-0.65 \times$ length of basitarsus; metabasitarsus 1.11-1.32× length of tarsomeres III, IV, and $V$ combined; hind tibia $2.00-2.17 \times$ long-
er than basitarsus; hind femur length $3.50-3.85 \times$ its maximum width. Wings. Forewing length/width $=2.45-2.71$; stigma 3.17-3.58× longer than maximum width; forewing vein R1 $0.61-0.63 \times$ as long as vein RS; vein RS slightly sinuate; vein $r$ arising slightly before middle of stigma; second submarginal cell triangular, with petiole $0.11-0.13 \mathrm{~mm}$ long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width $=3.36-3.76$; hind wing vein $1 \mathrm{M} 1.83-1.85 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with 4 hamuli. Metasoma. Apical width of petiole (tergum 1) $2.88-3.18 \times$ wider than basal width; minimum width of petiole $0.48-0.55 \times$ apical width; length of ovipositor sheath $0.17-0.20 \mathrm{~mm}$.

Male. Similar to female except that male has 31-34 flagellomeres, fore and middle femora and tibia yellowish-orange, hamuli with 4 or 5 hooks; sometimes the petiole color blackish.

## Host. Unknown.

## Distribution. USA.

Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median trapezoidal-shape elevation, head transverse in frontal view, gena distinctly bulging, groove between lateral ocelli slightly microfoveolate, anterolateral edges of scutellum usually with slightly acute projection, head and mesosoma black and wings slightly infumate.

Remarks. This species is near to C. oaxaquensis, but differs in that C. oaxaquensis has a triangular-shaped head, area between antennal sockets with a median pyramidalshaped elevation, gena not bulging, and antenna with 38-39 flagellomeres.

Etymology. This species is named in honor of Olga Margot De la Rosa Reyes, mother of the first author.

Material examined. Holotype $q$ : USA, California: Shingle, El Dorado Co., 22/V/1955, Burdick D.J. Allotype ỏ: same data as holotype. Holotype and allotype deposited in EMEC. Paratypes: California: Tahoe National Forest, Pineland Drive 3.2 km S Tahoe City, Placer County, 1 § 20/VII/1983, 1900 m., Davies Thomas W. (CAS); 4 miles S Railway Flat, Calaveras Co., 2 § 19/V/1969, 853 m., Linsley E.G.; Shingle, El Dorado Co., 1 § 22/V/1955, Burdick D.J. (EMEC); River Pines, Amador Co., 1 § 26/IV/1975 (UCDC); Volcano, Amador Co., 1 § 5/V/1957, Moore C.G. (USNM). Utah: 23 miles NE Logan, Cache Co., $1 \not \subset$ 20/VI/1963, Toschi C.A. (EMEC).

## Crassomicrodus pallens (Cresson, 1873)

http://species-id.net/wiki/Crassomicrodus_pallens
Fig. 14a-e
Crassomicrodus pallens (Cresson): Muesebeck 1927: 20.
Microdus pallens Cresson 1873: 53.

Holotype female. Illinois [USA]. Cat. No. 2746 (ANSP).
Description female. Body. Length. $4.20-6.48 \mathrm{~mm}$. Color (Fig. 14e). Integument yellowish-orange except ocelli translucent yellow; eyes black or silver; mandible
apex and apical area of hind tibia and tarsomeres blackish. Sometimes propleuron black with metapleuron and propodeum blackish, rarely head and mesopleuron blackish. Wing veins dark brown; forewing infumate with a hyaline spot on the first submarginal cell that is similar in size to the parastigma. Head (Fig.14ab). Triangular in frontal view; face without longitudinal ridge dorsomedially; eye height/width = 1.36; eye height $0.55-0.58 \times$ inter-ocular distance; area between antennal sockets with a median pyramidal-shaped elevation and two weakly defined tubercles; frons excavated with a pair of microfoveolate groove that diverge towards the ocellar area; posterior surface of antennal sockets smooth, rarely rugulose; groove between lateral ocelli smooth; median ocellus separated from lateral ocellus by smooth groove; gena not bulging; malar space $0.77-0.86 \times$ as long as eye height; clypeus $1.63-2.00 \times$ wider than high; length of ventrolateral margin of clypeus similar to diameter of tentorial pit; antenna with 29-34 flagellomeres; setae at base of mandible distinctly longer than setae on rest of body surface. Mesosoma (Fig. 14cde). Pronotum strigulose or reticulate rugulose; lateral pronotal margins with weakly crenulate groove; notauli impressed; anterolateral edges of scutellum lacking small acute projection; scutellar disc convex with sparse setae from 0.09 to 0.11 mm in length; scutellar disc sloped posteriorly and rounded; lateral scutellar depression smooth, rarely with punctures on the ventral margins; carinae of central metanotal area almost triangular shaped; propodeum reticulate rugulose; subalar lobe separated from mesopleuron by narrow rugulose groove, width distinctly shorter than the subalar lobe; metapleuron reticu-late-rugulose. Legs. Inner spur of middle tibia $0.74-0.81 \times$ length of basitarsus; inner spur of hind tibia $0.61-0.72 \times$ length of basitarsus; metabasitarsus $1.02-1.12 \times$ length of tarsomeres III, IV, and V combined; hind tibia 2.22-2.39× longer than basitarsus; hind femur length $3.44-3.85 \times$ its maximum width. Wings. Forewing length/ width $=2.46-2.55$; stigma $3.00-3.44 \times$ longer than maximum width; forewing vein R1 $0.47-0.57 \times$ as long as vein RS; vein RS not sinuate; vein $r$ arising slightly before middle of stigma; second submarginal cell triangular, with petiole $0.05-0.09$ mm long; vein $\mathrm{M}+\mathrm{CU}$ distinctly pigmented throughout; hind wing length/width = 3.10-3.39; hind wing vein $1 \mathrm{M} 1.41-1.53 \times$ longer than $1 \mathrm{r}-\mathrm{m}$; hind wing with $4-5$ hamuli. Metasoma. Apical width of petiole (tergum 1) 3.60-3.92× wider than basal width; minimum width of petiole $0.58-0.63 \times$ apical width; length of ovipositor sheath $0.20-0.33 \mathrm{~mm}$.

Male. Similar to female.
Host. Unknown.
Distribution. Mexico and USA.
Diagnosis. Distinguished from other Crassomicrodus species by the following combination of characters: area between antennal sockets with a median pyramidal-shaped elevation, head triangular in frontal view, malar space $0.77-0.86 \times$ as long as eye height, body length $4.20-6.48 \mathrm{~mm}$, forewing vein R1 $0.47-0.57 \times$ as long as vein RS, body usually yellowish-orange.

Remarks. Crassomicrodus pallens resembles C. divisus in the shape of the head, but differs by the characters in the key. A few specimens of this species have the malar space


Figure 14. Crassomicrodus pallens. Female a anterior view of head, arrows indicate a median pyramidalshaped elevation with two weakly defined tubercles $\mathbf{b}$ dorsal view of head $\mathbf{c}$ lateral view of mesosoma $\mathbf{d}$ dorsal view of mesosoma $\mathbf{e}$ female habitus.
shorter than eye height ( 0.77 times), specimens with this variation also have R1 less than 0.57 times as long as vein RS.

Material examined. Type: 1 : Ill. (ANSP). Homotype: 1 q, USA, South Carolina: Hilton Head Island, 17/VII/1965, Howden H. \& A. Howden (CNC).

Other specimens examined.- MEXICO, Colima: 21 miles N Manzanillo, 1 q 25/VIII/1970, Wasbauer M.S. \& J.S. Wasbauer (CNC). Nayarit: Arroyo Santiago, Nr. Jesus Maria, 2 ¢ 5/VII/1955 (CNC, EMEC). Sinaloa: 2.5 miles N Mazatlán, 1 \& 12/VIII/1970, Wasbauer M., malaise trap (CNC); 35 miles S Es-
cuinapa， 1 § 24／IV／1961，Howden \＆Martin（HIC）．Sonora：Alamos， 2 q， 1 ふ 5／IX／1970，Bohart R．M（UCDC）；La aduana W Alamos， 1 \＆18／VIII／1964， Irwin M．E．（USNM）； 5 miles E Navojoa， 1 q 9／IX／1970，Bohart R．M．（UCDC）． Veracruz：Minatitlán， 1 ¢ 1／II／1992，Osborn H．（USNM）．USA，Alabama：Ala－ bama， 2 q 1980；Auburn，Lee Co．， 1 q 9／VI／1917；Tuskegee， 1 ð 22／VII／1930， Beamer R．H．（USNM）．Arkansas：Washington Co． 1 q 11／X／1955，Baker T．A． at light（USNM）；Shoal Bay Rec．Area， 1 ô 20／V／1981，Dasch B．\＆C．Dasch （AEIC）．Connecticut：East Hartford， 1 q 13／VI／1947，Evans Howard E．；Riv－ erbank，East Hartford， 1 q 20／IX／1946，Evans Howard E．（CNC）．Florida： $1 \sigma^{\Uparrow}$ （CNC）．Tarpon Sprs．， 1 § 20／III／1950，Townes H．K．；Cedar Key， 1 § 28／III／1985， Townes H．\＆M．Townes；Elfes， 1 \＆4／IV／1937，Franclemont J．G．； 5 miles NE Bronson，Levy Co．， 1 ő 29／III／1986，LaSalle John（AEIC）；De Funiak Springs， 1 § 17－19／X／1914；Crestview， 1 § 15－16／X／1914（AMNH）；Pine Hill Estates， Gainesville， 1 § 4／X／1973，Weems H．V．Jr．，malaise trap（CNC）；Archbold Biol． Sta．，Highlands Co．， 1 q 28VII10VIII／1987，Wahl D．B．（HIC）；Fleming Key， Monroe Co．， 2 \＆14／VIII／1979，Acree John A．\＆H．V．Weems Jr．insect flight trap； 1 miles W Interlachen，Putnam， 1 q， 5 § 16／IV／1984，Stange L．A．； 2 miles N Holt，Okaloosa Co．， 1 ő 28／X／1983，Stange L．A；Gainesville，Alachua Co．， 1 ô 1／VIII／1977， 1 ¢ 31／VII／1977，Davis L．R．Jr．；Interlachen，Putnam Co．， 2 ふ 4／V／1986，Stange L．A．；Monteoca，Alachua Co．， 1 \＆26／IX／1977， 1 \＆3／X／1977， Butler F．Jerry；Nokomis，Sarasota Co．， 1 § 9／I／1967；Palmdale，Glades County， 1 đ̃ 25／VI／1972，Pierce W．H．；Pine Hill Estates，Gainesville， 1 q 2／X／1973， 1 q 20／IX／1973， 1 \＆29／IX／1973， 1 q 30／IX／1973，Weems H．V．Jr．，malaise trap；San Felasco Hammock，Alachua Co．， 1 q 17／III／1977，Fairchild G．B．\＆Weems H．V． Jr．；Suwanne Riv．St．Pk．，Suwannee Co．， 1 \＆13－25／IV／1977，Wiley J．R．（FSCA）； San Felasco Hammock，Alachua Co．， 1 q 9－14／III／1977，Fairchild G．B．（UCDC）； Arcadia， 1 ふ 2／VII／1962， 1 đ 3／VII／1962，Krombein Karl V．；Alachua Co．， 1 ふ̋ 4／VII／1955，Weems H．V．；Gainesville， 1 \＆28／VIII／1960，Stange L．A．（USNM）． Georgia： 2 \＆（USNM）；Pine Mtn．Rabun Co．， $1 \overbrace{}^{\top} 1 / \mathrm{VIII} / 1957,457$ m．，Chillcott J．G．（CNC）； 10 miles N Waycross，Ware Co．， 1 § 27／VIII／1960，Marsh P．M．； Lakeland， 1 ठ 18／IV／1940，Fattig P．W．（USNM）．Illinois： $1 \jmath^{\lambda}$ VIII（USNM）； Havana， 1 § 18／VIII／1912，Devil＇s Hole；Meredosia， 1 § 22／VIII／1917；St．Anne， 1 ठ 22／VII／1935，Ross \＆DeLong（INHS）；Carbondale，Jackson Co．， 1 q 28／ IX／1956，Downey J．C．（USNM）．Indiana：Forest Nursery，Jackson Co． 1 § 23／ IX／1938，Schnell R．L．（USNM）．Iowa：Sioux City， 1 § 15／VII／1927，Ainslie C．N． （USNM）．Kansas：Lawrence， 1 ¢ 12／VI／1960，Menke A．S．（UCDC）； 1 § Bald－ win，V，Bridwell J．C．；Lawrence vicinity，Douglas Co．， 1 ô 29／VI／1962，Roberts R．（USNM）．Maryland：Beltsville， 1 đ 23／V，Krombein K．V．；College Park， 1 đ 17／VIII／1914（USNM）；Crownsville， 1 \＆14／VII／1956，Krombein Karl V．（HIC）． Massachusetts $2 \circlearrowleft^{\lambda}$（USNM）；Truro 1 中， $1 \circlearrowleft^{\lambda} 4 / I X / 1904$ ，Morse A．P．（MCZ）； Cabo Cod， 1 q 6／IX／1939，Dreisbach R．R．（MSUC）；Nantucket， 1 § 7／IX／1909； Nantucket， 1 § 8／IX／1926，Johnson C．W．（USNM）．Michigan：Newago Co．， 1 ふ

13／VI／1940，Dreisbach R．R．（AEIC）；Newago Co．， 2 § 30／VII／1944，Dreisbach R．R．（MSUC，USNM）．Minnesota：Ft．Snelling， 1 Q 2／VIII／1923，Hertig A．T．； Ft．Snelling， 2 28／VII／1922，Nichol A．A．；John Latsch St．Pk．，S Minneiska， 1 ㅇ 1／V／1951；Jordan， 1 q 15／IX／1930，Talford H．S．；Jordan，sand area， 1 q 13／ VII／1923，Hertig A．T．（UMSP）．Missouri：Columbia， 1 ठ 29／IX／1964，Bayer L．G． （AEIC）；Williamsville， 1 q 1－16／VI／1969，Becker J．T．，malaise trap（CNC）；Co－ lumbia， 1 ¢ 20／VII／1967，Parker F．D．（UCDC）；Taney Co．， 1 中， 1 ô13／IX／1944， Portman R．W．（UMRM）；Columbia， $1 \not \subset 22 / V I I I / 1967$ ，Parker F．D．，malaise trap； Columbia， 1 Q 26／X／1931，Craig W．S．（USNM）．Nebraska：Valentine Refuge， 1 q 4／VI／1972， 1 § 6／VI／1972，Townes H．\＆M．Townes（AEIC）；Custer Co．， 1 § 21／ VIII／1951，Dreisbach R．R．（MSUC）．New Jersey：Palmyra， 1 q 29／VIII／1933，Ca－ zier M．A．（AMNH）；Bergenfield， 1 ठ VIII／1918？，Schott F．M．；Lucaston， 1 ठ 12／ IX／1902，Daecke E．（USNM）．New Mexico：Hatch， 1 q 29／VIII／1974， 1 \＆30／ VIII／1974，Townes H．\＆M．Townes（AEIC）．Riverton， 2 § 5／IX／1948；Westville， 1 ¢ 12／IX／1897（USNM）．New York：Cold Springs Harb， 1 ő 25／VII，Melander A．L．（AEIC）．North Carolina：Clinton， 1 § 24／V／1951，Townes H．\＆M．Townes； Nags Head， 1 §， 1 中 25／V／1948，Krombein K．V．；Smokemont， 1 ठ 15／VIII／1947， Bullock \＆Dreisbach（AEIC）；Fort Bragg，Cumberland Co．， 1 中 27IX／3X／1967， Birchim Jim D．（CAS）；Cabo Hatteras mt．Buxton， 1 ō 14／VIII／1961，Howden H．；Highlands， 1 § 4／VI／1957， 1158 m．，Vockeroth J．R．（CNC）．Jacksonville， Onslow Co．， 1 đ IX／1963；Kill Devil Hills， 2 đ̋ 26／VI／1954， 1 q 2／VII／1954， Krombein Karl V．；（HIC）；Kill Devil Hills， 2 q， 6 ठ 1／VII／1954， 1 q 2／VII／1954， 1 ¢ 22／VI／1954， 2 ð 23／VI／1954， 1 中， 7 ふ 26／VI／1954， 1 ふ 28／VI／1954， 1 ふ 3／VII／1954， 1 中， 8 § 30／VI／1954，Krombein Karl V．；Raleigh， 1 ð 20／V／1937 （UMSP）；Salvo，Dare Co．， 1 § 6／VIII／1958，Krombein（USNM）；Southern Pines， 1 ใ 3／V／1951，Howden H．\＆A．Howden； 1 ¢ 16／IX／1956，Krombein Karl V．； $1 ठ^{\top}$ 16／VIII（AEIC，USNM）．North Dakota： 7 miles SE Sheldon，Ransom Co．， 1 § 5／ VIII／1973，Powers J．R．； 11 miles W Walcott，Richland Co．， 2 \＆23／V／1988，Pow－ ers J．R．（EMEC）．Oklahoma：Lake Texoma 2 miles E Willis， 1 § VI／1965，Bohart R．M．（UCDC）；Cimarron River near Freedom，Woods Co． 1 \＆11／V／1984，Hevel G．F．\＆J．F．Hevel（USNM）．Pennsylvania：Dauphin， 1 ठ 6／VI／1909，Daecke E． （USNM）；Philadelphia， 1 đ，1991？，Fox（USNM）．Rhode Island：Providence $10^{\Uparrow}$ （USNM）．South Carolina：Greenville， 1 ठ 2／X／1941， 1 ठ 8／X／1941，Townes H． \＆M．Townes（AEIC）．Tennessee：Jefferson City， 2 ¢ 7／VII／1947，Valentine B．D． （CNC）．Texas：Junction，Kimble Co．， 1 ¢ 5／V／1986，Pulawski W．J．（CAS）；Seago－ ville cerca Dallas， 2 ¢ XI／1944，Weyrauch（IMLA）；Austin 1 〇（MCZ）；Hunts－ ville，Chartman $1 \delta^{\top}$ ；New Boston， 1 § 17／X／1905，Bishopp F．C．；Pierce， 1 § 22／ IV／1907，alfalfa，Mitchell J．D．（USNM）．Virginia：Ft．Humphreys， 1 đ 6／IX／1928， Mickel C．E．（UMSP）；Barcroft， 1 § 2／IX／1934，Bridwell J．C．；Falls Church， 1 q 30／VIII／1923，Greene C．T．；Nelson Co．， 1 \＆17／VIII／1927，Robinson W．；Ross－ lyn， 1 ¢ V／1929；Tazewell， 1 ¢ 9／VI／1915，Jackson L．O．（USNM）．Wisconsin： Griffith St．Nursery，Wood Co．， 1 § 8／VI／1948，Shenefelt R．D．（AEIC）．

## Acknowledgments

We wish to thank to the curators of the following institutions for the generous loan of specimens: AEIC, AMNH, ANSP, CAS, CIBE-UANL, CNC, CUIC, EMEC, ESUW, FSCA, HIC, IBUNAM, IMLA, INHS, INIFAP, IRCW, KSUC, MCZ, MSUC, MUCR, OSU, TAMU, UADY, UCDC, UCMC, UCR, UMRM, UMSP, USNM. We acknowledge SNI-CONACYT support to JIF (41093). This work was supported also by Coordinación de la Investigación Científica of the Universidad Michoacana de San Nicolás de Hidalgo and CONACYT-Ciencia Básica. Support to MJS was provided by NSF DEB 0205982.

## References

Ashmead WH (1894) Some parasitic Hymenoptera from lower California. Proceedings of the California Academy of Science 4(2): 125.
Ashmead WH (1900) Classification of the ichneumon-flies or the superfamily Ichneumonoidea. Proceedings of the United States National Museum 23: 110-152. doi: 10.5479/ si.00963801.23-1206.1
Bradley JC (1916) Taxonomic notes on Agathinae (Hymenoptera-Braconidae). Psyche 23: 139-140.
Brues CT (1926) Studies on Ethiopian Braconidae, with a catalogue of the African species. Proceedings of the American Academy of Arts and Sciences 61(8): 287. doi: 10.2307/20026158
Cresson ET (1865) Catalogue of Hymenoptera in the collection of the Entomological Society of Philadelphia, from Colorado territory. Proceedings of the Entomological Society of Philadelphia 4: 297-299.
Cresson ET (1872) Texan Hymenoptera. Transactions of the American Entomological Society Supplement 4: 182.
Cresson ET (1873) Descriptions of North American Hymenoptera, No.5. The Canadian Entomologist 4: 52-53.
Eady RD (1968) Some illustrations of microsculpture in the Hymenoptera. Proceeding of the Royal Entomological Society London (A) 43(4-6): 66-72.
Figueroa JI, Sharkey MJ, Romero-Nápoles J, López-Martínez V, Sánchez García JA, Martínez AM, Pineda S (2008) Redescription of Crassomicrodus fulvescens (Cresson) (Hymenoptera: Braconidae: Agathidinae), with new distributional data and revised taxonomic status. Zootaxa 1934: 63-68.
Marsh PM (1960) A new species of Crassomicrodus Ashmead (Hymenoptera: Braconidae). The Pan-Pacific Entomologist 36(3): 153-154.
Marsh PM, Carlson RW (1979) Superfamily Ichneumonoidea. In: Krombein KV, Hurd PD, Smith DR, Burks BD (Eds) Catalog of Hymenoptera in America North of Mexico. Vol. 1. Symphyta and Apocrita (parasitica). Smithsonian Institution Press. Washington, D.C., 196.

Muesebeck CFW (1927) A revision of the parasitic wasps of the subfamily Braconinae occurring in America North of Mexico. Proceedings of the United States National Museum 69(16): 1-73. doi: 10.5479/si.00963801.69-2642.1
Muesebeck CFW, Krombein KV, Townes HK (1951) Hymenoptera of America North of Mexico Synoptic Catalog. United States Department of Agriculture. Agriculture Monograph No. 2. United States Department of Agriculture, Washington, D.C., 1420 pp.
Pucci T, Sharkey MJ (2004) A revision of Agathirsia Westwood (Hymenoptera: Braconidae: Agathidinae) with notes on mouthpart morphology. Journal of Hymenoptera Research 13: 64-107.
Sharkey MJ (1992) Cladistics and tribal classification of the Agathidinae (Hymenoptera: Braconidae). Journal of Natural History 26: 425-447. doi: 10.1080/00222939200770251
Sharkey MJ (1997) Subfamily Agathidinae. In: Wharton RA, Marsh PM, Sharkey MJ (Eds). Manual of the new world genera of the family Braconidae (Hymenoptera). Special Publication of the International Society of Hymenopterists 1, 69-83.
Sharkey MJ (2006) Two new genera of Agathidinae (Hymenoptera: Braconidae) with a key to the genera of the New World. Zootaxa 1185: 37-51.
Sharkey MJ, Wharton RA (1997) Morphology and Terminology. In: Wharton RA, Marsh PM, Sharkey MJ (Eds) Manual of the New World Genera of the Family Braconidae (Hymenoptera). International Society of Hymenopterists. Special Publication No. 1, 20-37.
Sharkey MJ, Laurenne NM, Sharanowski B, Quicke DLJ, Murray D (2006) Revision of the Agathidinae (Hymenoptera: Braconidae) with comparisons of static and dynamic alignments. Cladistics 22: 546-567. doi: 10.1111/j.1096-0031.2006.00121.x
Shenefelt RD (1970) Braconidae 3. Agathidinae. In: Ferriere Ch et van der Vecht J (Ed) Hymenopterorum Catalogus. Uitgeverij Dr. W Junk NV The Hague, Holland. Pars 6. p. 380.
Szépligeti GV (1913) Afrikanische Braconiden des Deutschen Entomologischen Museums (Hym.). Entomologische Mitteilungen 2(12): 385.
Viereck HL (1905) Notes and descriptions of Hymenoptera from the Western United States, in the collection of the University of Kansas. Transactions of the Kansas Academy of Sciences 19: 288.
Viereck HL (1913) Descriptions of ten new genera and twenty-three new species of Ichneu-mon-flies. Proceedings of the United States National Museum 44(1968): 558-559.


[^0]:    Copyright M. A. Todaro et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

[^1]:    Copyright E.M. Shimbori, A.M. Penteado-Dias. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

[^2]:    Copyright J.I. Figueroa et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

