



Two new eriophyid mite species associated with Clematis terniflora var. mandshurica in China (Acari, Eriophyidae)

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Abstract

Two new eriophyid mite species associated with *Clematis terniflora* var. *mandshurica*, namely *Aculops jilinensis* **sp. n.** and *Phyllocoptes terniflores* **sp. n.**, are described. Both species infest the tender leaves of host plants, inducing severe curling and blistering.

Keywords

Aculops, Phyllocoptes, plant feeding, taxonomy

Introduction

Clematis terniflora DC. var. mandshurica (Rupr.) Ohwi, called "la liao tie xian lian" in Mandarin, belongs to the family Ranunculaceae, and is native to China (Heilongjiang, Jilin, Liaoning, Shanxi, Inner Mongolia), Korea, Mongolia, and the Russian Far East (website of Flora Republicae Popularis Sinicae – http://frps.eflora.cn/frps/Clematis%20terniflora and Germplasm Resources Information Network – https://npgsweb.ars-grin.gov/gringlobal/taxonomydetail.aspx?id=404246). It is also known in folk medicine and planted as an ornamental plant in the northeast of China.



Figure 1. Damage symptoms associated with *Aculops jilinensis* sp. n. and *Phyllocoptes terniflores* sp. n. on *Clematis terniflora* var. *mandshurica*: **A** leaves have severely curled and blistered, red arrow indicates the magnified curled leaf **B** leaves have moderately curled and blistered, red arrow indicates part of the mite population hidden inside the curled surface.

To date, no eriophyid mite species were reported from *C. terniflora* var. *mandshurica*, although at least nine species are known from other *Clematis* spp. worldwide, namely *Aceria vitalbae* (Canestrini, 1892) (from *C. vitalba* L.), *Calepitrimerus clematisis* Song, Xue & Hong, 2008 (from *Clematis* sp.), *Cupacarus subnotatus* (Nalepa, 1924) (from *C. recta* L.), *Epitrimerus flammulae* Gerber, 1901 (from *C. flammula* L.), *Phyllocoptes atragenes* Liro, 1941 [from *C. alpina* (L.) Mill.], *P. heterogaster* (Nalepa, 1890) (from *C. recta*) (Nalepa, 1891), *P. heteronotus* Nalepa, 1924 (from *C. recta*), *P. monochetus* Nalepa, 1924 (from *C. recta*), and *Platyphytoptus vitalbae* Farkas, 1960 (from *C. vitalba*).

During field surveys in 2015, some leaves were found to be severely curled and blistered (Figure 1). The curled and blistered leaves were checked with the aid of a microscope in the laboratory. Eriophyid mites were found and two new species were identified by the first and third authors.

Materials and methods

Sampling was made on the host plants in the field by the aid of a hand lens (30×), in Jilin Agricultural Science and Technology University, Jilin City, Jilin Province, China. The eriophyid mites gathered with host plant were placed in vials and stored in 75% ethanol, each vial was marked with the collection data. In the laboratory, the samples including mites were poured into a Petri dish and mite specimens were picked up using a fine pin and placed in Keifer's Booster to clear them. They were then slide-mounted with modified Berlese medium (Amrine and Manson 1996).

Specimens were examined with the aid of a Zeiss A2 (Germany) research microscope with phase contrast (A-plan phase contrast objectives: ×10/0.25 (NA), ×20/0.45 (NA); EC plan-NEOFLUAR phase objectives: ×40/0.75 (NA); ×100/1.3 (NA), oil immersion) and schematic drawings were made. Micrographs were taken at ×100 magnification

(×10 eyepiece magnification) with a camera AxioCam MRc attached to the microscope, and connected to a computer, using Axiovision (4.8.2) image analysis software.

The morphological terminology used follows Lindquist (1996) and the generic classification is made according to Amrine et al. (2003). Specimens were measured following de Lillo et al. (2010). For each species, the holotype female measurement precedes the corresponding range for holotype and paratypes (given in parentheses). All measurements are in micrometres (μ m) and represent lengths, when not otherwise specified. All type specimens are deposited as slide-mounted specimens in the Arthropod/Mite Collection of the Department of Entomology, Nanjing Agricultural University (NJAU), Jiangsu Province, China.

Taxonomy

Family Eriophyidae Nalepa, 1898 Subfamily Phyllocoptinae Nalepa, 1892 Tribe Anthocoptini Amrine and Stasny, 1994 Genus *Aculops* Keifer, 1966b

Aculops jilinensis sp. n. http://zoobank.org/A3F6DB8B-8143-4DFC-990E-CD2EAB8BC505 Figs 2, 3

Diagnosis. Body fusiform; prodorsal shield with acuminate frontal lobe, median and admedian lines complete and connected at base by a pair of short lines, forming an "arrow", submedian lines connected by a pair of diagonally reaching lines; scapular seta short 15 (14–20) on rear shield margin, projecting posteriorly; opisthosoma dorsally with evenly curved annuli (54–73 dorsally, 71–84 ventrally) and all standard setae for the Eriophyidae; legs with standard setae, empodium simple, 7-rayed; coxigenital region with three pairs of setae and many granules, female genital coverflap with 13 (12–13) longitudinal ridges and two to three transverse lines at base.

Description. FEMALE: (n = 9). Body fusiform, opisthosoma broadest 12 annuli posterior of the prodorsal shield, then tapering regularly until its posterior apex; 179 (179–271), 50 (50–70) wide, 53 (50–60) thick; light yellow. **Gnathosoma** 19 (19–25), projecting obliquely downwards, pedipalp coxal seta (*ep*) 4 (3–4), dorsal pedipalp genual seta (*d*) simple, 6 (6–9), cheliceral stylets 11 (11–20). **Prodorsal shield** 38 (30–42), 45 (35–45) wide, subtriangular; frontal lobe acuminate, 6 (5–7). Median and admedian lines complete and connected at base by a pair of short, almost transversal (slightly oblique) lines, forming an "arrow"; median and admedian lines are also connected at centre by a pair of short, diagonally directed lines, forming an inverted "V"; submedian lines formed by two pairs of incomplete lines (submedian lines I and submedian lines II); submedian lines I reaching about midway, merged with a pair of lines converging posteriorly ('a' in Figure 2AD), themselves joining perpendicularly another

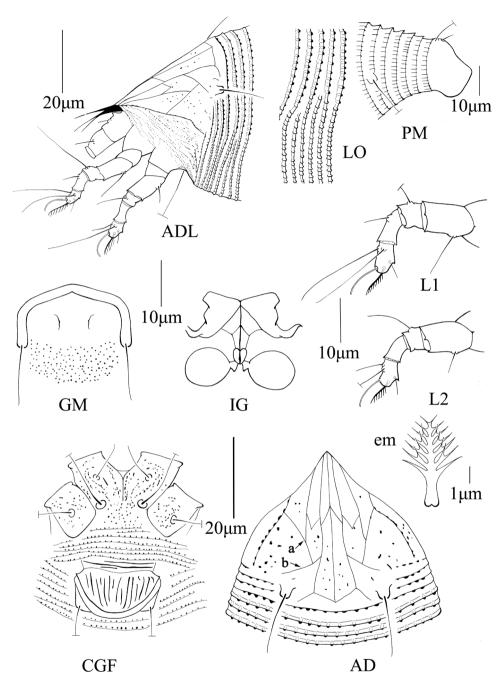


Figure 2. Schematic drawings of *Aculops jilinensis* sp. n.: **ADL** lateral view of anterior body region (slightly rotated dorsad) **LO** lateral view of annuli at mid-region of opisthosoma **PM** lateral view of posteriormost region of opisthosoma **GM** male genital region **IG** female internal genitalia **LI** leg I **L2** leg II **em** empodium **CGF** female coxigenital region **AD** prodorsal shield.

pair of lines oriented anteromesally ('b' in Figure 2AD); submedian lines II flanking lateral edges of shield, joining with lines 'a'; submedian lines II, together with lines 'a' and 'b', forming a triangular cell, opened posterolaterally; submedian lines I connected with admedian lines at center by a pair of "V" shaped lines; many granules distributed in the 'triangular' cell and between median, admedian and submedian lines. Scapular tubercles on rear shield margin, 24 (23–26) apart, scapular seta (sc) 15 (14–20), projecting posterior. Coxigenital region with 7 (5-10) microtuberculated annuli. Coxal plates with granules throughout; anterolateral seta on coxisternal plate I (1b) 9 (7–10), 13 (13–14) apart, proximal seta on coxisternal plate I (1a) 43 (35–45), 9 (8–11) apart, proximal seta on coxisternal plate II (2a) 48 (40-50), 25 (23-25) apart. Prosternal apodeme 6 (6-8). Leg I 37 (29-37), femur 9 (9-12), basiventral femoral seta (bv) 10 (9–11); genu 7 (5–7), antaxial genual seta (*l*") 21 (20–25); tibia 9 (7–9), paraxial tibial seta (l) 6 (6-8), located at 1/3 from dorsal base; tarsus 6 (6-7), paraxial, fastigial, tarsal seta (ft') 19 (18-20), antaxial, fastigial, tarsal seta (ft') 22 (22-26), paraxial, unguinal, tarsal seta (u) 4 (4-5); empodium (em) 9 (7-9), simple, 7-rayed, tarsal solenidion (ω) 9 (8–10), rod-like. *Leg II* 26 (23–26), femur 9 (9–13), basiventral femoral seta (bv) 9 (8-12); genu 4 (4-5), antaxial genual seta (l") 10 (8-11); tibia 7 (6-8); tarsus 6 (5-6), paraxial, fastigial, tarsal seta (ft) 7 (6-9), antaxial, fastigial, tarsal seta (ft') 25 (21–25), paraxial, unguinal, tarsal seta (u) 5 (4–5); empodium (em) 7 (7–8), simple, 7-rayed, tarsal solenidion (ω) 9 (9–10), rod-like. **Opisthosoma** dorsally arched, with 56 (54-73) dorsal semiannuli bearing rounded microtubercles except last 7-9th semiannuli with elongated microtubercles; ventrally with 72 (71-84) semiannuli, with (longitudinally) elongated microtubercles. Seta c2 30 (30–35) on ventral semiannulus 12 (11–13), 45 (43–50) apart; seta d 73 (60–73) on ventral semiannulus 24 (20–28), 33 (30-45) apart; seta e 17 (15-20) on ventral semiannulus 42 (40-50), 17 (17-25) apart; seta f 25 (22-26) on 6th ventral semiannulus from rear, 18 (16-18) apart. Seta *h1* 5 (4–5), seta *h*2 70 (60–72). *Female genitalia* 14 (14–17), 21 (21–24) wide, coverflap with 13 (12-13) longitudinal ridges and two to three transverse lines at base, seta 3a 34 (34-45), 16 (16-20) apart.

MALE: (n = 1, dorsal view). Body fusiform, 270, 50 wide; light yellow. *Gnathosoma* 25, projecting obliquely downwards, pedipalp coxal seta (ep) 3, dorsal pedipalp genual seta (a) simple, 5, cheliceral stylets 20. *Prodorsal shield* 32, 45 wide, subtriangular, frontal lobe acuminate, 5; shield design similar to that of female. Scapular tubercles on rear shield margin, 26 apart, scapular seta (sc) 15, projecting posteriorly. *Coxigenital region* with 7 microtuberculated annuli. Coxal plates with granules, anterolateral seta on coxisternal plate I (1b) 10, 14 apart, proximal seta on coxisternal plate I (1a) 43, 10 apart, proximal seta on coxisternal plate II (1a) 43, 10 apart, proximal seta on coxisternal plate II (1a) 40, 27 apart. Prosternal apodeme 7. *Leg I* 35, femur 12, basiventral femoral seta (1a) 10; genu 7, antaxial genual seta (1a) 23; tibia 9, paraxial tibial seta (1a) 7, located at 1a from dorsal base; tarsus 7, paraxial, fastigial, tarsal seta (1a) 28, antaxial, fastigial, tarsal seta (1a) 23, paraxial, unguinal, tarsal seta (1a) 5; empodium (1a) 9, simple, 7-rayed, tarsal solenidion (1a) 10, rod-like. *Leg II* 30, femur 8, basiventral femoral seta (1a) 12; genu 5, antaxial genual seta (1a)

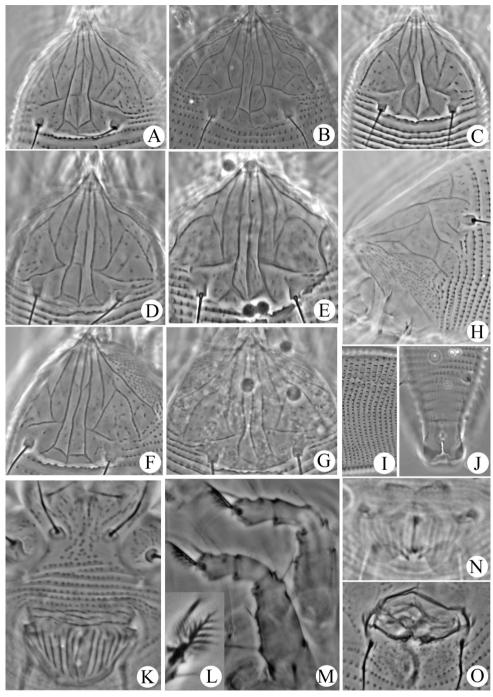


Figure 3. Micrographs of *Aculops jilinensis* sp. n.: **A–G** prodorsal shield **H** lateral view of anterior body region **I** lateral view of annuli at mid-region of opisthosoma **J** ventral view of posteriormost region of opisthosoma **K** female coxigenital region **L** empodium **M** legs I (right) and II (left) **N** female internal genitalia **O** male genital region.

13; tibia 7; tarsus 6, paraxial, fastigial, tarsal seta (ft) 9, antaxial, fastigial, tarsal seta (ft) 23, paraxial, unguinal, tarsal seta (u) 5; empodium (em) 7, simple, 7-rayed, tarsal solenidion (ω) 10, rod-like. **Opisthosoma** dorsally arched, with 70 semiannuli, with rounded microtubercles on the posterior margin, last 7-9th semiannuli with elongated microtubercles, ventrally with 80 semiannuli, with elongated microtubercles. Seta c2 38 on ventral semiannulus 16, 55 apart; seta d 65 on ventral semiannulus 28, 45 apart; seta e 16 on ventral semiannulus 46, 26 apart; seta f 29 on f ventral semiannulus from rear, 24 apart. Seta f 5, seta f 5. **Male genitalia** 24 wide, seta f 40, 20 apart.

Type material. Holotype female (slide number NJAUAcariEriJ8A.1; marked Holotype), from *C. terniflora* var. *mandshurica* (Ranunculaceae), Jilin Agricultural Science and Technology University, Jilin City, Jilin Province, China, 43°57'16"N, 126°28'58"E, elevation 221m, 19 July 2015, coll. Yan-Mei Sun. **Paratypes** 8 females and 1 male on 9 microscope slides (slide number NJAUAcariEriJ8A.2–8A.10), same collection data and repository as holotype.

Relationship to host. Infesting the tender upper leaves; making leaves severely curled and blistered (Figure 1A); hiding inside the curled surfaces (Figure 1B).

Etymology. The specific designation *jilinensis* is derived from the name of location, Jilin City, where the new species was collected.

Remarks. Up to now, no eriophyoid mite species in the genus Aculops was reported from the host plant family Ranunculaceae. The new species is similar to the other species in the genus Aculops but can be easily distinguished by characters of specific prodorsal shield design. However, it is mostly similar to A. alachuae Keifer, 1966b, which also has dorsal annuli with rounded microtubercles, coxal plates with granules, female genital coverflap with longitudinal ridges and prodorsal shield with lined design and many granules. The new species can be separated from A. alachuae by its 7-rayed empodium (4-rayed in A. alachuae), median and admedian lines connected at the base by a pair of transverse lines forming an "arrow" (an "arrow" is present at the base of median line, but not connected with admedian lines in A. alachuae), submedian lines connected by diagonally reaching lines (submedian lines separated in A. alachuae), scapular seta sc short, 15 (14-20) (seta sc 27 in A. alachuae). Aculops alachuae was reported infesting Rhus copallinum L. var. leucantha (Jacq.) DC. (Anacardiaceae) from Florida, USA, galling host plant leaves (Keifer, 1966b). The new species is also similar to A. euphorbicolus (Keifer, 1964), which also have annuli with rounded microtubercles (53 dorsal annuli), coxal plates with granules, female genital coverflap with longitudinal ridges and prodorsal shield with lined design, 7-rayed empodium, but can be differentiated by prodorsal shield with many granules between lines (prodorsal shield without granules in A. euphorbicolus), median line complete (median line incomplete in A. euphorbicolus), opisthosoma with 72 (71-84) ventral annuli (opisthosoma with 60 ventral annuli in A. euphorbicolus), female genital coverflap with 13 (12-13) longitudinal ridges and two to three transverse lines at base (female genital coverflap with 8-10 ridges and two rows of granules in A. euphorbicolus). Aculops euphorbicolus was reported from Euphorbia corollata L. (Euphorbiaceae) from Virginia (USA), making deformed flower clusters or galls (Keifer, 1964).

Some intraspecific differences in the design of the prodorsal shield were observed, especially between the median and admedian lines. The median line is complete in all specimens examined except the specimen illustrated in Figure 3C (median line interrupted at centre). Besides connected at base, median and admedian lines are always separated in specimens in Figures 3E and G. Median and admedian lines connected at base, basal 2/3 and 1/3 in Figures 3A, B, D, and F.

Tribe Phyllocoptini Nalepa, 1892 Genus *Phyllocoptes* Nalepa, 1887

Phyllocoptes terniflores sp. n. http://zoobank.org/BA0B864A-96A5-48C4-A9BC-C2F3F499364E Figs 4, 5

Diagnosis. Body fusiform; prodorsal shield with broad frontal lobe, scapular setae ahead of rear shield margin, projecting upward-centrally, median, admedian and submedian lines formed by granules aligned and making a network; opisthosoma dorsally with three ridges, middorsal ridge fade as long as lateral ridges, with 55 (55–60) dorsal and 100 (92–100) ventral annuli, all standard setae of the Eriophyidae present; legs with standard setae, empodium simple, 5-rayed; coxigenital region with three pairs of setae and many granules, female genital coverflap with 12 (10–12) longitudinal ridges and two transverse lines at the base.

Description. FEMALE: (n = 10). Body fusiform, 256 (200–304), 73 (68–78) wide, opisthosoma broadest 12–13 annuli posterior of the prodorsal shield, then tapering regularly until its posterior apex; light yellow. Gnathosoma 25 (20-26), projecting obliquely downwards, pedipalp coxal seta (ep) 3 (3-4), dorsal pedipalp genual seta (d) simple, 9 (7-9), cheliceral stylets 15 (15-22). *Prodorsal shield* 50 (50-55), 57 (55-65) wide, semicircular; frontal lobe broad, 7 (6-7). Shield pattern composed of granules aligned and connected by faint lines. Median line: largely broken at centre; anterior part originated on the frontal lobe and ended at about 1/5 of the anterior prodorsal shield, thereafter, connected with admedian lines by a pair of transverse lines; posterior part originated at about 4/5 of prodorsal shield, vanished at rear of prodorsal shield, connected with admedian lines by a pair of transverse line at anterior. Admedian lines complete and sinuous and connected with submedian lines by two pairs of transverse lines at basal 3/4 and center of prodorsal shield. Submedian lines flanking lateral edges of shield branched into two curled lines, forming a large open semicircle at lateral side of prodorsal shield; many aligned granules distributed between submedian lines. Scapular tubercles ahead of rear shield margin, 22 (22-26) apart, scapular seta (sc) 12 (12-14), projecting upward-centrally. Coxigenital region with 13 (11-13) microtuberculated annuli. Coxal plates with granules and irregular lines throughout, anterolateral seta on coxisternal plate I (1b) 15 (13–15), 15 (15–17)

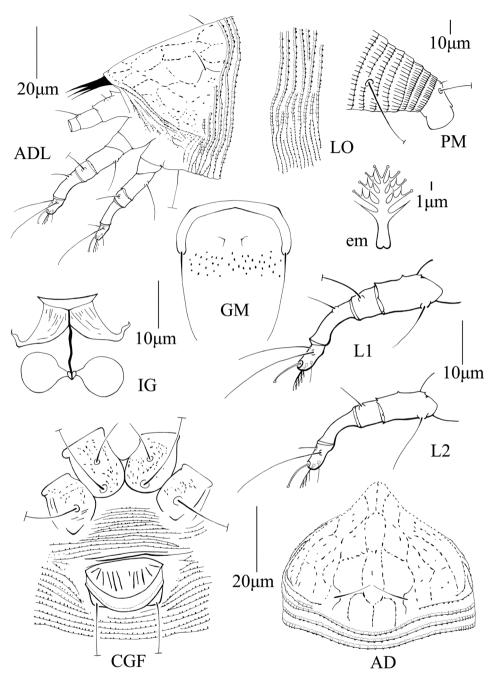


Figure 4. Schematic drawings of *Phyllocoptes terniflores* sp. n.: **ADL** lateral view of anterior body region (slightly rotated dorsad) **LO** lateral view annuli **PM** lateral view of posterior opisthosoma **em** empodium **GM** male genital region **IG** female internal genitalia **LI** leg I **L2** leg II **CGF** female coxigenital region **AD** prodorsal shield.

apart, proximal seta on coxisternal plate I (1a) 45 (35-45), 12 (11-14) apart, proximal seta on coxisternal plate II (2a) 65 (63-65), 32 (31-35) apart. Prosternal apodeme 6 (6-7). Leg I 36 (36-38), femur 13 (13-15), basiventral femoral seta (bv) 13 (13-14); genu 7 (6–8), antaxial genual seta (l') 38 (35–40); tibia 12 (10–12), paraxial tibial seta (1) 7 (5-7), located at 1/3 from dorsal base; tarsus 7 (7-8), paraxial, fastigial, tarsal seta (ft') 17 (17-20), antaxial, fastigial, tarsal seta (ft') 25 (25-28), paraxial, unguinal, tarsal seta (u') 5 (4–5); empodium (em) 8 (7–8), simple, 5-rayed, tarsal solenidion (ω) 8 (7–8), knobbed. *Leg II* 35 (33–35), femur 14 (13–15), basiventral femoral seta (bv) 15 (12–15); genu 6 (5–6), antaxial genual seta (*l*") 9 (6–9); tibia 10 (8–10); tarsus 6 (6-7), paraxial, fastigial, tarsal seta (ft') 5 (5-6), antaxial, fastigial, tarsal seta (ft'') 24 (24-28), paraxial, unguinal, tarsal seta (u) 5 (4-5); empodium (em) 8 (7-8), simple, 5-rayed, tarsal solenidion (ω) 9 (9–10), knobbed. **Opisthosoma** dorsally with 55 (55– 60) semiannuli, with rounded microtubercles on the posterior margin, except last 6th semiannuli with elongated microtubercles; ventrally with 100 (92-100) semiannuli, with nearly rounded microtubercles on central part; moreover, with elongated microtubercles in side area and the ventral semiannulus between seta e and f; last 6 ventral semiannuli with elongated and linear microtubercles. Seta c2 40 (40-43) on ventral semiannulus 21 (20-22), 67 (65-74) apart; seta d 54 (43-45) on ventral semiannulus 43 (39–43), 48 (47–50) apart; seta e 32 (29–32) on ventral semiannulus 69 (63–69), 26 (24–26) apart; seta f37 (36–37) on 6th ventral semiannulus from rear, 28 (25–28) apart. Seta h1 5 (4-5), seta h2 87 (83-87). Female genitalia 17 (17-20), 24 (24-27) wide, coverflap with 12 (10-12) longitudinal ridges and two transverse lines at the base, seta 3a 27 (23–28), 19 (19–21) apart.

MALE: (n = 1). Body fusiform, 233, 72 wide; white. Gnathosoma 20, projecting obliquely downwards, pedipalp coxal seta (ep) 3, dorsal pedipalp genual seta (d) simple, 7, cheliceral stylets 15. Prodorsal shield 50, included the frontal lobe, 55 wide, with a broad based frontal lobe broad, 8, shield design similar to that of female. Scapular tubercles ahead of rear shield margin, 22 apart, scapular seta (sc) 11, projecting centrad. Coxigenital region with 14 microtuberculated annuli. Coxal plates with irregular lines, anterolateral seta on coxisternal plate I (1b) 9, 14 apart, proximal seta on coxisternal plate I (1a) 33, 10 apart, proximal seta on coxisternal plate II (2a) 50, 28 apart. Prosternal apodeme 10. *Leg I* 34, femur 12, basiventral femoral seta (*bv*) 9; genu 5, antaxial genual seta (l') 27; tibia 10, paraxial tibial seta (l) 6, located at 1/3 from dorsal base; tarsus 7, paraxial, fastigial, tarsal seta (ft) 16, antaxial, fastigial, tarsal seta (ft") 24, paraxial, unguinal, tarsal seta (u') 4; empodium (em) 7, simple, 5-rayed, tarsal solenidion (ω) 8, knobbed. *Leg II* 27, femur 11, basiventral femoral seta (bv) 10; genu 10, antaxial genual seta (l') 6; tibia 9; tarsus 6, paraxial, fastigial, tarsal seta (ft) 5, antaxial, fastigial, tarsal seta (ft") 24, paraxial, unguinal, tarsal seta (u) 5; empodium (em) 6, simple, 5-rayed, tarsal solenidion (ω) 8, knobbed. **Opisthosoma** dorsally with 45 semiannuli, with rounded microtubercles on the posterior margin, last 5th semiannuli with elongated microtubercles; ventrally with 82 semiannuli, with nearly rounded microtubercles on central part; moreover, with elongated microtubercles in side area and the ventral semiannulus between seta e and f; last six ventral semiannuli with elon-

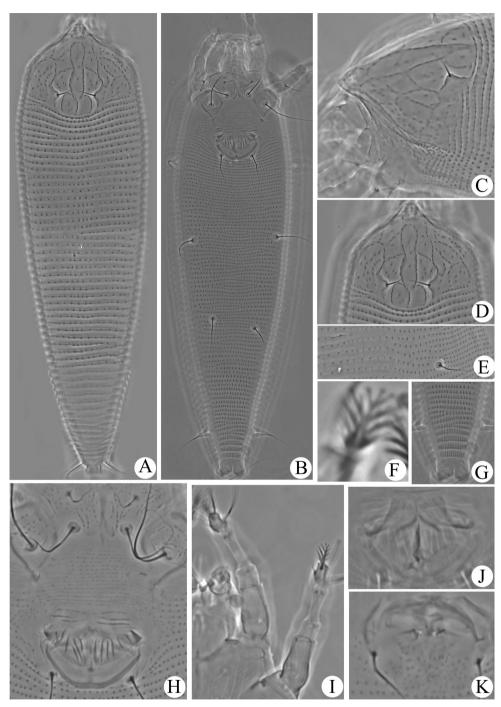


Figure 5. Micrographs of *Phyllocoptes terniflores* sp. n.: **A** dorsal view of female **B** ventral view of female **C** lateral view of anterior body region **D** prodorsal shield **E** lateral view annuli **F** empodium **G** ventral view of posterior opisthosoma **H** coxigenital region and female genitalia **I** leg I and leg II **J** female internal genitalia **K** male genital region.

gated and linear microtubercles. Seta c2 30 on ventral semiannulus 18, 62 apart; seta d 30 on ventral semiannulus 30, 40 apart; seta e 20 on ventral semiannulus 50, 23 apart; seta f 26 on 6th ventral semiannulus from rear, 23 apart. Seta h 15, seta h 260. *Male genitalia* 19 wide, seta h 3a 14.

Type material. Holotype female (slide number NJAUAcariEriJ8B.1; marked Holotype), from *C. terniflora* var. *mandshurica* (Ranunculaceae), Jilin Agricultural Science and Technology University, Jilin City, Jilin Province, China, 43°57'16"N, 126°28'58"E, elevation 221m, 19 July 2015, coll. Yan-Mei Sun. **Paratypes** 9 females and 1 male on ten microscope slides (slide number NJAUAcariEriJ8B.2–8B.11), same collection data and repository as holotype.

Relationship to host. Infesting the tender upper leaves and making leaves severely curled and blistered (Figure 1A); hiding inside the curled surfaces (Figure 1B).

Etymology. The specific designation *terniflores* is derived from the species name of the host plant, *terniflora*.

Remarks. The new species was compared with others in the genus *Phyllocoptes* infesting *Clematis* sp. This species is similar to *P. atragenes* [from *C. alpina*, infesting host plant as curled leaves], but can be differentiated from the latter by its shield pattern: the median and admedian lines are discontinuous (median and admedian lines continuous in *P. atragenes*), dorsal opisthosoma with 55 (55–60) annuli (dorsal opisthosoma with 48 annuli in *P. atragenes*) and dorsal annuli with rounded microtubercles (dorsal annuli smooth in *P. atragenes*). This species is also similar to *P. heterogaster* (Nalepa, 1891) [from *C. recta*, infesting host plant as abnormal hair], but can be differentiated from the latter by having its coxal plates with granules and short lines (coxal plates smooth in *P. heterogaster*), empodium 5-rayed (empodium 4-rayed in *P. heterogaster*), median line present on anterior of prodorsal shield (median line absent from anterior of prodorsal shield in *P. heterogaster*).

Besides species from *Clematis* sp., the new species is also similar to *P. calirubi* Keifer, 1938 [from Rubus ursinus Cham. & Schltdl. (Rosaceae)], P. exochordae Keifer, 1972 [from Exochorda racemosa (Lindl.) Rehder (Rosaceae)] and P. neenachensis Keifer, 1966a [from Oenothera deltoides Torr. & Frém. (Onagraceae)] by dorsal and ventral annuli with rounded microtubercles, female genital coverflap with longitudinal ridges and especially prodorsal shield design formed by granules aligned (besides with the generic characters of *Phyllocoptes*). The new species can be differentiated from the later three species by large size of body, 256 (200-304) (140-155 in P. calirubi, 200-215 in P. exochordae and 145-195 in P. neenachensis), median line present at anterior of dorsal shield (median lines absent from anterior of dorsal shield in all three species), coxal area with many granules and short lines (coxal area with few short lines in P. calirubi; coxal area I with short lines, coxal area II smooth in P. exochordae; coxal area with granules and short lines in P. neenachensis), solenidion knobbed (solenidion unknobbed in all three species), empodium 5-rayed (empodium 5-rayed in P. calirubi, empodium 6-rayed in P. exochordae and empodium 4-rayed in P. neenachensis) and short scapular seta 7 (6-7) (scapular seta 11 in P. calirubi and P. exochordae, scapular seta 10 in P. neenachensis).

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References

- Amrine JW Jr, Manson DCM (1996) Preparation, mounting and descriptive study of eriophyoid mites. In: Lindquist EE, Sabelis MW, Bruin J (Eds) Eriophyoid Mites: Their Biology, Natural Enemies and Control. Elsevier, World Crop Pests 6: 383–396. doi: 10.1016/S1572-4379(96)80023-6
- Amrine JW Jr, Stasny TA (1994) Catalog of the Eriophyoidea (Acarina: Prostigmata) of the world. Indira Publishing House, West Bloomfield, Michigan, 798 pp.
- Amrine JW Jr, Stasny TA, Flechtmann CHW (2003) Revised keys to world genera of Eriophyoidea (Acari: Prostigmata). Indira Publishing House, West Bloomfield, Michigan, 244 pp.
- Canestrini G (1892) Prospetto dell'Acarofauna Italiana. Parte Va. Famiglia dei Phytoptini (Phytoptidae). Atti della Societá Veneto-Trentina di Sciense naturali, Padova, ser. 2(1): 541–722.
- de Lillo E, Craemer C, Amrine JW Jr, Nuzzaci EG (2010) Recommended procedures and techniques for morphological studies of Eriophyoidea (Acari: Prostigmata). Experimental and Applied Acarology 51: 283–307. doi: 10.1007/S10493-009-9311-x
- Farkas HK (1960) Über die Eriophyiden (Acarina) Ungarns. I. Beschreibung neuer und wenig bekannter Arten. Acta Zoologica Academiae Scientarium Hungaricae 6: 315–339.
- Gerber C (1901) Zoocécidies Provençales. Comptes Rendus Assocociation Française pour l'avancement des Scienxes, Congres Ajaccio 1901: 524–550.
- Keifer HH (1938) Eriophyid Studies II. Bulletin of the California Department of Agriculture, 27: 301–323.
- Keifer HH (1964) Eriophyid Studies B-11. California Department of Agricultural, Sacramento, 20 pp.
- Keifer HH (1966a) Eriophyid Studies B-19. California Department of Agricultural, Sacramento, 20 pp.
- Keifer HH (1966b) Eriophyid Studies B-21. California Department of Agricultural, Sacramento, 20 pp.
- Keifer HH (1972) Eriophyoid Studies C-7. U.S. Department of Agriculture, Agricultural Research Service, 24 pp.
- Lindquist EE (1996) External anatomy and notation of structures. In: Lindquist EE, Sabelis MW, Bruin J (Eds) Eriophyoid mites: their biology, natural enemies and control. World Crop Pests, 6, Elsevier Science Publishers, Amsterdam, Netherlands, 3–31. doi: 10.1016/S1572-4379(96)80003-0

- Liro J (1941) Über neue und seltene Eriophyiden (Acarina). Societas zoologica-botanica Fennica Vanamo 8(7): 1–53.
- Nalepa A (1887) Die Anatomie der Phytopten. Sitzungsberichte der kaiserlichen Akademie der Wissenschaften. Mathematisch-naturwissenschaftliche Klasse, Wien. Abtheilung 1 96(4): 115–165.
- Nalepa A (1890) Zur Systematik der Gallmilben. Sitzungsberichte der kaiserlichen Akademie der Wissenschaften. Mathematisch-naturwissenschaftliche Klasse, Wien. Abtheilung 1 99(2): 40–69.
- Nalepa A (1891) Neue Gallmilben. Nova Acta Academiae Caesareae Leopoldino-Carolinae Germanicae Naturae Curiosorum Verhandlungen der kaiserlichen Leopoldinische-Carolinische Deutschen Akademie der Naturforscher 55(6): 363–395.
- Nalepa A (1892) Neue Arten der Gattung *Phytoptus* Duj. und *Cecidophyes* Nal. Denkschriften der kaiserlichen Akademie der Wissenschaften. Mathematisch-naturwissenschaftliche Klass (Wien) 59: 525–540.
- Nalepa A (1898) Eriophyidae (Phytoptidae). Das Tierreich. Eine Zusammenstellung und Kennzeichnung der rezenten Tierformen. Berlin. 4. Lief., Acarina, 74 pp.
- Nalepa A (1924) Polymorphe Eriophyiden. Marcellia 20(4-6): 87-96.
- Song ZW, Xue XF, Hong XY (2008) Eriophyoid mite fauna (Acari: Eriophyoidea) of Gansu Province, northwestern China with descriptions of twelve new species. Zootaxa 1756: 1–48.