



# Timarete posteria, a new cirratulid species from Korea (Annelida, Polychaeta, Cirratulidae)

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#### **Abstract**

A new cirratulid species, *Timarete posteria* sp. n., is described from the intertidal habitats of the eastern coast of South Korea. The new species is closely related to *Timarete luxuriosa* (Moore, 1904) from southern California based on morphological similarity of the branchial and tentacular filaments and the noto- and neuropodial spines. However, *T. posteria* sp. n. differs from the latter based on the following characteristics: 1) evenly divided peristomium into three annulations; 2) 2–4 neuropodial spines originating in the posterior chaetigers alternated by a few capillaries; and 3) complete shift in branchial filaments located about one-third between the notopodium and the dorsal midline. The new species has a methyl green staining pattern (MGSP) distinct from other *Timarete* species. Detailed description and illustrations of the new species are provided with molecular information based on the partial sequences of the mitochondrial cytochrome c oxidase subunit I (COI) and 16S ribosomal RNA (16S). This study also includes a key and discussion of known *Timarete* species from East Asia.

#### **Keywords**

Korean waters, polychaete, taxonomy, Timarete, COI, 16S

#### Introduction

The genus *Timarete* Kinberg, 1866 is a typical multi-tentaculate genus assigned to the cirratulid polychaetes (Blake 1996). *Timarete* species are clearly distinguished from other multi-tentaculate species by having chaetae including capillaries and acicular

spines, tentacular filaments forming a transverse series arising from two or more chaetigers, and branchiae originating in the segment anterior to tentacular filaments and shifted toward the mid-dorsum of the body in middle and posterior chaetigers (Fauchald 1977, Blake 1996).

Recently, several species belonging to *Timarete* have been taxonomically re-evaluated (Blake 1996, Cinar 2007, Magalháes and Bailey-Brock 2010, Magalháes et al. 2014). Blake (1996) redescribed *Timarete perbranchiata* (Chamberlin, 1918) from California and confirmed the taxonomic state of *T. convergens* (Chamberlin, 1918) as a junior synonym of it. Blake (1996) also determined that Cirriformia luxuriosa (Moore, 1904), which had been misconstrued and redescribed it as a species of *Timarete*. Çinar (2007) redescribed *Tima*rete punctata (Grube, 1859) and designated a lectotype and paralectotypes based on syntypes from the U. S. Virgin Islands (Caribbean Sea) deposited in the Zoological Museum of Copenhagen, and he also examined materials from the Mediterranean Sea and South Africa. Magalhães and Bailey-Brock (2010) redescribed an endemic Hawaiian species, previously been known as Cirriformia hawaiensis Hartman, 1956, as a species of Timarete species, T. hawaiensis (Hartman, 1956). Magalhães et al. (2014) re-examined several multitentaculate cirratulid species collected from the Brazilian coast including four *Timarete* species: T. caribous (Grube & Ørsted in Grube, 1859), T. ceciliae Magalhães, Seixas, Paiva, & Elias, 2014, T. oculata (Treadwell, 1932), and T. punctata species complex. Seixas et al. (2017) find out that the presence of two cryptic species within *T. punctata* species complex, one species widely distributed from Atlantic and Pacific Oceans and another species from Bahia, South Atlantic Ocean, using the genetic data of COI and 16S sequences.

Within Korean waters, *Timarete antarctica* (Monro, 1930) is the only recorded species of *Timarete* known in this region (Paik 1989). However, Paik (1989)'s description was very brief and lacked a review of the specific characters. While studying the polychaetes from Korean waters, a new species belonging to the genus *Timarete* was discovered. Here, we conducted a detailed examination of this species using methyl green staining pattern (MGSP) and several ontogenetic characters including the segmental origins of tentacular filaments, distributions of the neuroacicular and notoacicular spines, and dorsal shift of branchial filaments as used by previous investigators (Çinar 2007, Magalhães and Bailey-Brock 2010, Magalhães et al. 2014). In this study, the description and illustrations of the new species are provided together with molecular data pertaining to the barcoded regions of the mitochondrial cytochrome c oxidase subunit I (COI) and 16S ribosomal RNA (16S). Additionally, we reviewed and discussed *Timarete* species recorded from East Asia.

### Materials and methods

## Sampling and morphological observations

Samples were collected from the intertidal rocky bottoms. Specimens were sorted using sieves with a pore size of 0.5 mm, fixed initially with a 5% formaldehyde-seawater solu-

tion, and transferred to 85% ethyl alcohol. The characteristics of the whole body were observed with appendages dissected in a petri-dish using a pair of dissection forceps, or surgical knives and needles under a stereomicroscope (SMZ1500; Olympus, Tokyo, Japan). Dissected specimens were mounted onto temporary slides using glycerol or permanent slides using polyvinyl lactophenol solution. Drawings were based on stereomicroscopy and light microscopy (LABOPHOT-2; Nikon, Tokyo, Japan) aided by drawing tubes. Photographs were captured using an image system (LAS V4.7, Leica Microsystems, Heerbrugg, Switzerland). Specimens for scanning electron microscopy (SEM) were dehydrated with a t-BuOH freeze dryer (VFD-21S; Vacuum Device, Ibaraki, Japan). They were mounted on stubs and coated with gold-palladium. SEM observations were conducted using a scanning electron microscope (SU3500; Hitachi, Tokyo, Japan). Type material and additional material examined were deposited at the National Institute of Biological Resources (NIBR) in Incheon, Korea and the National Marine Biodiversity Institute of Korea (MABIK) in Seocheon, Chungcheongnam-do, Korea, respectively.

## Molecular analysis

Genomic DNA was extracted from the posterior segments of three specimens selected from the additional material using a DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. The partial sequences of the mitochondrial cytochrome c oxidase subunit I (COI) and 16S ribosomal RNA (16S) from gDNA were amplified using polymerase chain reaction (PCR) with the following primers: LCO 1490 5'-GGTCAACAAATCATAAAGATATTGG-3' and HCO 2198 5'-TAAACTTCAGGGTGACCAAAAAATCA-3' in COI amplification (Folmer et al. 1994) and 16SarL 5'-CGCCTGTTTATCAAAAACAT-3' and 16SANR 5'- GCTTACGCCGGTCT AACTCAG-3' in 16S amplification (Palumbi 1996; Zanol et al. 2010). PCR amplification was conducted in a total volume of 20  $\mu$ L: 10  $\mu$ L of 2x DyeMIX-Tenuto (Enzynomics), 0.5  $\mu$ L of each primer, 1  $\mu$ L of gDNA, and 8 µL of sterile water. Touchdown-PCR was carried out according to the following cycling program: 94 °C for 5 min, 94 °C for 1 min, 50 °C for 1 min and 72 °C for 1 min, followed by 20 cycles at decreasing annealing temperatures in decrements of 0.5 °C per cycle, followed by 1 min at 94 °C, 15 cycles of 1 min at 40 °C, 1 min at 72 °C, and final extension at 72 °C for 7 min. PCR products were purified with a QIAquick PCR Purification Kit (Qiagen, Chatsworth, CA, USA). Sequences of the new species were obtained using an Applied Biosystems 3730 DNA sequencer. These sequences were aligned with those of other Timarete species using Clustal W (Thompson et al. 1994, Larkin et al. 2007) in Geneious Pro v.9.1.8 (Biomatters, Auckland, New Zealand). The genetic distances of the new species from other species and the Maximum likelihood (ML) tree were determined by MEGA v.6.06 (Tamura et al. 2013).

# Systematic accounts

Family Cirratulidae Ryckholt, 1851 Genus *Timarete* Kinberg, 1866

*Timarete posteria* sp. n. http://zoobank.org/E13A2164-7182-4D18-ACD1-80C4F4C1A9B1 Figs 1–4

Material examined. *Type locality:* South Korea, Gyeongsangbuk-do Province: Pohang-si County, Heunghae-eup, Odo-ri, 36°09'17"N, 129°24'02"E, 13 July 2017, intertidal rocky bottom. *Holotype:* complete specimen (NIBRIV0000829700). *Paratypes:* one complete specimen (MABIKNA00146231); one complete specimen (MABIKNA00146238); one complete specimen (MABIKNA00146239); one complete specimen (MABIKNA00146245). *Non-type material:* 16 specimens (13 complete and 3 incomplete specimens), collection details same as type materials; 11 specimens (all complete), South Korea, Gangwon-do Province: Goseong-gun County, Jugwang-myeon, Munamjin-ri, 35°18'41"N, 129°32'33"E, 10 April 2017, intertidal rocky bottom.

**Diagnosis.** Body with deep ventral groove and distinct segments. Prostomium triangular, without eyespots. Peristomium evenly divided into three annulations. Branchial filaments one pair per segment, beginning from third peristomial annulus, and gradually shifting to mid-dorsum between chaetigers 30–78; completely shifted branchiae at about one-third distance between notopodium and dorsal midline. Grooved tentacular filaments arising from chaetigers 5–6 and occasionally 6–7 or 7–8. Chaetae including capillaries and acicular spines; notopodial spines 1–4, pale brown in color, from chaetigers 16–45; neuropodial spines 2–4, curved distally, thicker than notoacicular spines, dark brown in color, from chaetiger 24–69.

**Description.** Holotype: complete, 5.5 cm in length (4.8–13.2 cm in paratypes) and 5.7 mm in maximum width (4.0–6.0 mm in paratypes), with approximately 261 segments.

*Body* elongated, rounded dorsally, flattened ventrally, with distinct ventral groove throughout and tapering posterior end. All segments distinct, narrow, crowded throughout body with distinct lateral shoulders. Body color in alcohol pale grey to dark grey, branchiae and tentacular filaments yellowish grey; live specimens with body dark red and branchiae and tentacular filaments light orange. No separate pigmentation on body (Fig. 1).

*Prostomium* short, triangular, blunt distally, and as long as three anterior chaetigers. Nuchal organs round, present on posterior-lateral prostomial region. Eyespots absent (Figs 2A, B, 3A).

*Peristomium* with three annulations nearly equal in length, longer than prostomium and as long as four anterior chaetigers; second and third annulations with 2–3 lateral wrinkles (Figs 2A, B, 3A).

Branchial filaments one pair per segment, from posterior margin of third peristomial annulus, continuing on most segments except about last ten segments; branchial



Figure 1. Timarete posteria sp. n., paratype (MABIKNA00146236), lateral view. Scale bar: 1.0 mm.

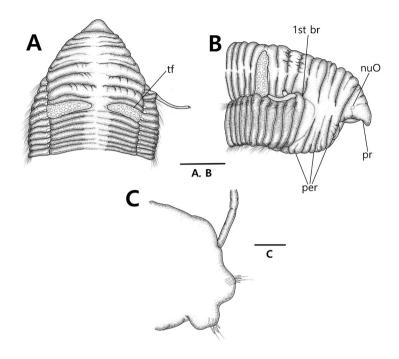
filaments located just above notopodial ridges in anterior 42 chaetigers (29–77 chaetigers in all specimens examined); then shifting gradually to mid-dorsum forming lateral bulge over notopodia from chaetiger 43 (30–78 in all specimens examined) to near posterior end; fully shifted branchiae located about one-third distance between notopodium and dorsal midline. (Figs 2A–C, 3B).

Tentacular filaments formed two transverse groups separated by median gap and arising on dorsum of chaetigers 5–6 (6–7 or 7–8 in some specimens examined); each group with about 18–21 filaments arranged in 2–3 transverse rows (Figs 2A, B, 3A).

*Parapodia*, notopodia forming lateral shoulders dorsally; noto- and neuropodium widely separated throughout (Figs 2A–C, 3A, B).

Chaetae including capillaries with serrated edge observed under light microscopy (400x) and SEM observation and acicular spines. Capillary chaetae about 8–10 capillaries arranged in two longitudinal rows in anterior parapodia. Notopodial spines nearly straight, pale brown in color, present from chaetiger 40 (16–45 in all specimens examined); 1–3 spines per segment accompanied by 1–3 companion capillaries from chaetiger 40 to posterior end. Neuropodial spines curved distally, slightly thicker than notopodial spines, dark brown in color, from chaetiger 30 (24–69 in all specimens examined); 2–3 spines per segment with 1–2 companion capillaries from chaetiger 30 to very posterior end (Figs 3C, D, 4A–C).

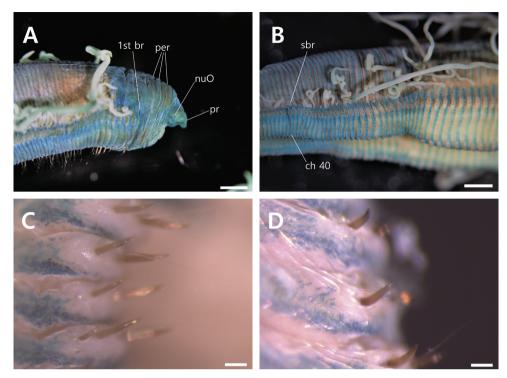
Pygidium with terminal anus (Fig. 4D).



**Figure 2.** *Timarete posteria* sp. n., **A–B** holotype (NIBRIV0000829700) **C** paratype (MABIK-NA00146239) **A** anterior end, dorsal view **B** anterior end, lateral view **C** parapodium of mid-body. Scale bars: 1.0 mm (**A, B**), 0.5 mm (**C**). Abbreviations: branchia (br), chaetiger (ch), nuchal organ (nuO), peristomium (per), prostomium (pr), tentacular filaments (tf).

**Methyl green staining pattern (MGSP).** Body stained with transverse bands on posterior half of each segment forming complete rings. Branchial and tentacular filaments not stained. Prostomium, peristomium, and dorsum of first 3 or 4 chaetigers intensely stained with dark green. Noto- and neuropodial ridges not stained (Fig. 3A, B).

**Variations.** Several morphological characters in cirratulids are highly variable ontogenetically and a few of them are clearly considered size-dependent in *Timarete* species (Blake 1996, Magalhães and Bailey-Brock 2010, Magalhães et al. 2014). We examined the relationships between the ontogenetic characteristics including the segmental origin of noto- and neuropodial spines and the shift of branchial filaments, and the total number of chaetigers in the new species according to the correlation analyses based on 31 complete specimens (Fig. 5). In *Timarete posteria* sp. n., the segmental origin of neuropodial spines ranged from chaetigers 16 to 45, strongly size-dependent (N = 31, r = 0.81). The segmental origin of notopodial spines varied from chaetigers 24 to 69, with weak size-dependent characteristics if compared with those of neuropodial spines (N = 31, r = 0.67). The dorsal shift of branchial filaments occurred between chaetigers 30 and 78 regardless of the total number of chaetigers in the new species (N = 31, r = 0.40) (Fig. 5). It is known that the appearance of tentacular filaments is generally variable in a few *Timarete* species (Magalhães et al. 2014). In *T. posteria* sp. n., the tentacular fila-



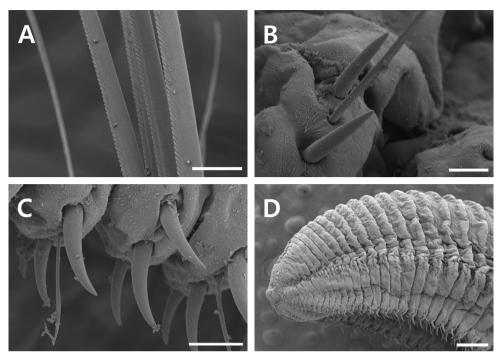
**Figure 3.** *Timarete posteria* sp. n., **A–D** holotype (NIBRIV0000829700) **A** anterior end, lateral view with MGSP **B** mid-body segments, lateral view with MGSP **C** notopodial spines of mid-body segments **D** neuropodial spines of mid-body segments. Scale bars: 1.0 mm (**A, B**), 0.1 mm (**C, D**). Abbreviations: branchia (br), chaetiger (ch), nuchal organ (nuO), peristomium (per), prostomium (pr), shift of branchial filament (sbr).

ments always originated in the dorsum of two consecutive chaetigers although their locations were variable: usually on chaetigers 5–6 (in 28 specimens) and occasionally 6–7 (in two specimens) or 7–8 (in four specimens) in all the 34 specimens examined. This variation appears to be somewhat related to body size because the tentacular filaments on chaetigers 7–8 occur in specimens containing more than 300 segments (almost similar to specimens with less than 300 segments on chaetigers 5–6). Further studies with a larger population and more temporal samples are needed to determine a more accurate relationship between the segmental origin of the tentacular filaments and the body size.

**Etymology.** The epithet of the specific name, *posteria*, is derived from the Latin *posterior*, meaning 'hind'. This name refers to the shift in the appearance of the branchial filaments from relatively posterior chaetigers. The gender of the genus name, *Timarete*, is feminine and the specific name of this new species is designated as feminine.

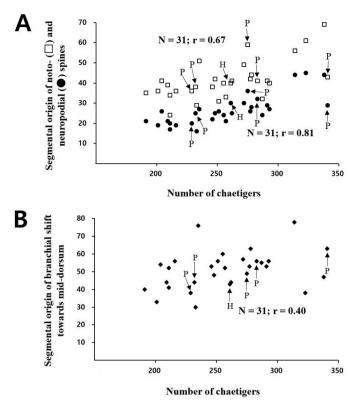
**Habitat and distribution.** This species is a common inhabitant of seagrass beds in the intertidal rocky bottoms and distributed in the East Sea (or the Sea of Japan) of South Korea.

**Molecular information.** In the present study, partial COI sequences each measuring 658 bp in size from five specimens and partial 16S sequence of 519 bp in size



**Figure 4.** SEM observation of *Timarete posteria* sp. n., **A–D** paratype (MABIKNA00146238) **A** capillary chaetae of anterior chaetiger **B** notopodial spines of posterior chaetiger **C** neuropodial spines of posterior chaetiger **F** pygidium, lateral view. Scale bars: 0.02 mm (**A**), 0.025 mm (**B**), 0.05 mm (**C**), 0.25 mm (**D**).

from a single specimen were obtained for the genetic analysis of Timarete posteria sp. n. They were deposited in GenBank under the accession number MH708229-MH708233 (COI) and MH822840 (16S). The intra-specific genetic distance between five COI sequences was measured according to the Kimura-2-parameter (K2P) model and ranged from 0 to 0.4 %. We carried out the genetic comparison of the new species with three Timarete species available, including T. caribous (Grube and Ørsted in Grube, 1859), T. ceciliae Magalhães, Seixas, Paiva, and Elias, 2014, and T. punctata (Grube, 1859) from the Brazilian coast, with COI and 16S sequences previously announced from GenBank (Magalhães et al. 2014). Based on entire genetic data uploaded in GenBank, the inter-specific genetic distances of COI and 16S sequences between the new species and other *Timarete* species were 23.7–26.2 % and 22.2–26.5 %, respectively (K2P distance). We examined the molecular phylogenetic relationship based on the Maximum likelihood (ML) tree using the genetic data available from GenBank on several cirratulids belonging to the multi-tentaculate genera, Cirriformia, Cirratulus, and Timarete, with the new species (Rousset et al. 2007, Hardy et al. 2011, Magalhães et al. 2014, Weidhase et al. 2014, Lado et al. 2016, Weidhase et al. 2016). The GenBank accession numbers of them are represented on Table 1. In ML tree (Fig. 6), all cirratulid species showed the specific validity by the molecular data of the present study. In generic level, the *Timarete* species including *T. posteria* sp.



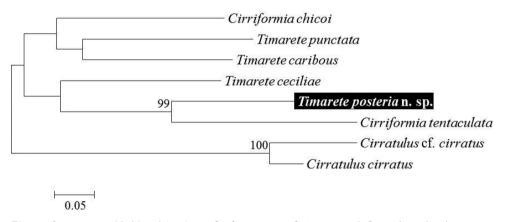
**Figure 5.** Variation of ontogenetic characters in *Timarete posteria* sp. n. **A** relationship between total number of chaetigers and segmental origin of noto- and neuropodial spines **B** relationship between total number of chaetigers and segmental origin of branchial shift toward mid-dorsum. Abbreviations: holotype (H), paratypes (P).

n. formed a clade with two *Cirriformia* species, *C. chicoi* Magalhães, Seixas, Paiva, & Elias, 2014 and *C. tentaculata* (Montagu, 1808), showing a similar result to the phylogenetic tree of Magalhães et al. (2017). This result suggests that both of the genera *Timarete* and *Cirriformia* are not monophyletic and they are closely related to each other. However, the reality of phylogenetic relationship between *Timarete* and *Cirriformia* still merits further study with more morphological and molecular information of the multi-tentaculate genera.

**Remarks.** The major characteristics of *Timarete posteria* sp. n. are mostly similar to those of *Timarete luxuriosa* (Moore, 1904), originally described from Southern California (Blake 1996). Both species share the following morphological features: 1) branchial filaments a single pair per segment, gradually shifting toward mid-dorsum from relatively posterior chaetigers compared to its congeners (the dorsal shift of branchial filaments occurred at about chaetiger 35 when the body with 350 segments in *T. luxuriosa* and occurred at chaetigers 38–78 with more than 300 segments and at chaetigers 30–76 with less than 300 segments in the new species, while that occurred at chaetigers 7–26 in other *Timarete* species); 2) tentacular filaments arising from the

Species	GenBank accession number		ъ.
	COI	16S	Data source
Timarete posteria sp. n.	MH708229	MH822840	Present study
Timarete caribous	KM192177	KM192193	Magalhães et al. 2014
Timarete ceciliae	KM192179	KM192195	Magalhães et al. 2014
Timarete punctata	KM192188	KM192205	Magalhães et al. 2014
Cirriformia tentaculata	KR916808	KT033725	Lado et al. 2016 (COI)
			Weidhase et al. 2016 (16S)
Cirriformia chicoi	KM192165	KM192189	Magalhães et al. 2014
Cirratulus cirratus	HM417794	DQ779609	Rousset et al. 2007 (16S)
			Hardy et al. 2011 (COI)
Cirratulus cf. cirratus	KM083601	KT033724	Weidhase et al. 2014 (COI)
			Weidhase et al. 2016 (16S)

**Table 1.** GenBank accession numbers for sequences obtained in the present study.



**Figure 6.** Maximum likelihood (ML) tree for four species of *Timarete* with four other related species in the multi-tentaculate cirratulid genera, *Cirriformia* and *Cirratulus*, inferred from combined dataset with COI and 16S. Numbers above the branch indicate ML bootstrap values from 1000 replication.

dorsum of chaetigers 5–6 (usually 5–6 and sometimes 6–7 or 7–8 in the new species); 3) notopodial spines originating from chaetiger 50 (24–69 in the new species) and pale brown in color; and 4) neuropodial spines originating from chaetiger 31 (16–45 in the new species) and dark brown in color (Blake 1996). However, *T. posteria* sp. n. clearly differs from *T. luxuriosa* in peristomium, notopodial spines, and shifting branchial filaments as follows: 1) the peristomium is evenly divided into three annulations in *T. posteria* sp. n., while that of *T. luxuriosa* comprises one large and three smaller annulations; 2) *T. posteria* sp. n. bears 2–4 neuropodial spines accompanied by a few capillaries in the posterior chaetigers, however, *T. luxuriosa* contains a single neuropodial spine without capillaries in the posterior chaetigers after around chaetiger 90; 3) completely shifted branchial filaments are located at about one-third distance between notopodium and dorsal midline in *T. posteria* sp. n., whereas those of *T. luxuriosa* are positioned at about two-thirds distance (Blake 1996).

The methyl green staining pattern (MGSP), which may be of diagnostic value, is unavailable for many *Timarete* species except for a few species recently described (Imajima and Hartman 1964, Blake 1996, Çinar 2007, Magalhães and Bailey-Brock 2010, Magalhães et al. 2014). Nevertheless, the MGSP of the new species is distinct from the previously described patterns from five *Timarete* species, including *T. caribous* (Grube, 1859), *T. ceciliae* Magalhães, Seixas, Paiva & Elias, 2014, *T. hawaiensis* (Hartman, 1956), *T. oculata* (Treadwell, 1932), and *T. punctata* (Grube, 1859), by a combination of the following features: 1) intense staining of the prostomium, peristomium, and dorsum of the first three chaetigers; 2) the presence of transverse bands forming complete rings in the posterior half of each segment (Magalhães and Bailey-Brock 2010, Magalhães et al. 2014). MGSP is a useful diagnostic feature in *Timarete* species, and additional MGSP information for several *Timarete* species is still required.

#### **Discussion**

Timarete antarctica (Monro, 1930) was originally reported from South Georgia in the Antarctic region (Monro 1930, Hartman 1966). It also has been widely reported from Korean and Japanese waters (Imajima and Hartman 1964, Paik 1989). However, this species is clearly distinguished from other *Timarete* species including *Timarete posteria* sp. n. based on having single capillary chaetae and longitudinal rows of tentacular filaments, while other *Timarete* species contain both capillary chaetae and acicular spines, and tentacular filaments arranged in transverse rows (Monro 1930, Imajima and Hartman 1964, Hartman 1966, Paik 1989, Blake 1996, Magalhães et al. 2014). Therefore, generically *T. antarctica* should be referred to the genus *Protocirrineris* Czerniavsky, 1881 based on the chaetal composition and arrangement of tentacular filaments (Blake 1996).

Blake (1996) suggested that the shift of the branchial filaments toward the middorsum in middle and posterior chaetigers is a significant generic diagnostic feature of *Timarete*. However, some *Timarete* species, including *T. japonica* Zachs, 1933, *T. dasylophius* (Marenzeller, 1879), and *T. gibbosa* (Moore, 1903) from East Asia, have been recorded or combined without considering the shift in branchial filaments (Zachs 1933, Imajima and Hartman 1964). Zachs (1933) defined *T. japonica* from the Sea of Japan based on a very brief record without description and illustration. Although the detailed diagnostic features of the species were inadequately dealt with, *T. japonica* is distinctly different to *Timarete* species in the diagnostic features suggested by Blake (1996). Zachs (1933) suggested that the lateral branchial filaments attached almost at the bases of notopodia are a diagnostic feature of *T. japonica*. We think that *T. japonica* may be a species of *Cirriformia* Hartman, 1936, because of the absence of shifting branchial filaments.

Imajima and Hartman (1964) redefined two cirratulid species originally recorded from Japanese waters, which were described as species of the genus *Cirratulus* Lamarck, 1818, as *Timarete* species, *T. dasylophius* and *T. gibbosa*. The single diagnostic feature of *Timarete* species included the tentacular filaments present on the dorsum of two or

more chaetigers, which determined the taxonomic status of these two species (Imajima and Hartman 1964). Among these two species, T. gibbosa has the shift of branchial filaments, which indicates that a pair of branchiae per segment are located at about midway between the parapodium and the dorsal mid-line except those on anterior segments (Moore 1903). Despite of having shifted branchial filaments, T. gibbosa is suspected to be a species of Cirratulus based on Imajima and Hartman's (1964) description indicating that this species has the transverse series of eyespots on the prostomium and the neuropodial spines from chaetiger 1. These characteristic features are commonly observed in Cirratulus (Blake 1996, Bottero et al. 2017). Moreover, the shift of branchial filaments, which is one of the representative characteristic features of this species, is also found from *Cirratulus* as well as *Timarete* (Blake 1996, Bottero et al. 2017). The affiliation of *C. dasylophius* to *Timarete* is also questionable because of the lack of reference to the shift of branchial filaments in the original description as well as Imajima and Hartman's study (Marenzeller 1879, Imajima and Hartman 1964). Furthermore, T. dasylophius has the branchial filaments from chaetiger 2 (Imajima and Hartman 1964) while *Timarete* species generally bear those from the last peristomial annulation or chaetiger 1 (Blake 1996, Magalhães et al. 2014). We suppose that Imajima and Hartman (1964) may have overlooked the presence of the scars of branchial filaments present on the peristomium or chaetiger 1.

Consequently, we suggest that three *Timarete* species previously recorded from East Asia, *T. japonica*, *T. dasylophius*, and *T. gibbosa*, are not valid species within the genus yet. Further study with the type materials is needed to verify their generic affiliation. Under a modern view of cirratulid taxonomy, meanwhile, three species among presently known *Timarete* species, *T. anchylochaeta* (Schmarda, 1861), *T. norvegica* (Quatrefages, 1865), and *T. polytricha* (Schmarda, 1861), are still remaining to be designated as valid members of *Timarete* because previous records of them have only poor information with brief descriptions and simple drawings (Schmarda 1861, Quatrefages 1865, Blake 1996, Magalhães et al. 2014). We herein provide a key to the species regarded as valid members of *Timarete*.

# Key to valid species of the genus Timarete

1	Dorsal branchiae abruptly shifted2
_	Dorsal branchiae gradually shifted
2	Neuropodial spine on posterior segments single
_	Neuropodial spine on posterior segments more than two
3	Number of tentacular filaments 7–9 <i>T. hawaiensis</i> (Hartman, 1956)
_	Number of tentacular filaments 15–20 <i>T. filigera</i> (Delle Chiaje, 1828)
4	Branchial filaments one pair per segment
_	Branchiae filaments 2–5 pair per segment

)	Branchiae and tentacular filaments with black lateral stripes				
		59)			
_	Branchiae and tentacular filaments without pigmentation patterns	6			
6	Lateral bulge over notopodia formed in shift of branchiae	7			
_	Lateral bulge over notopodia not formed in shift of branchiae	9			
7	Shift of branchiae arising beyond chaetiger 30	8			
_	Shift of branchiae arising in chaetigers 12–14 <i>T. nasuta</i> Ehlers, 18				
8	Posterior chaetigers with 2–4 neuropodial spines				
_	Posterior chaetigers with single neuropodial spine				
		04)			
9	Notopodial spines originated from chaetigers 11–23				
		014			
_	Notopodial spines originated from chaetigers 57–58				
	T oculata (Treadwell 19				

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#### References

- Blake JA (1996) 8. Family Cirratulidae Ryckholdt, 1851. Including a revision of the genera and species from the eastern North Pacific. In: Blake JA, Hilbig B, Scott PH (Eds) Taxonomic Atlas of the Benthic Fauna of the Santa Maria Basin and the Western Santa Barbara Channel. Volume 6 The Annelida Part 3, Polychaeta: Orbiniidae to Cossuridae. Santa Barbara Museum of Natural History Press, Santa Barbara, California, 263–384.
- Bottero MAS, Elias R, Magalháes WF (2017) Taxonomic revision of *Cirratulus* (Polychaeta: Cirratulidae) from the coasts of Argentina, with description of a new species. Journal of the Marine Biological Association of the United Kingdom 97: 889–896. https://doi.org/10.1017/S0025315417000807
- Çinar ME (2007) Re-description of *Timarete punctata* (Polychaeta: Cirratulidae) and its occurrence in the Mediterranean Sea. Scientia Marina 71: 755–764. https://doi.org/10.3989/scimar.2007.71n4755
- Fauchald K (1977) The Polychaete Worms. Definitions and keys to the orders, families and genera. Natural History Museum of Los Angeles County. Science Series 28: 1–190.

- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294–299.
- Hardy SM, Carr CM, Hardman M, Steinke D, Corstorphine E, Mah C (2011) Biodiversity and phylogeography of Arctic marine fauna: insights from molecular tools. Marine Biodiversity 41: 195–210. https://doi.org/10.1007/s12526-010-0056-x
- Hartman O (1966) Polychaeta Myzostomidae and Sedentaria of Antarctica. Antarctic Research Series 7: 1–158. https://doi.org/10.1029/AR007
- Imajima M, Hartman O (1964) The polychaetous annelids of Japan. Allan Hancock Foundation Publications. Occasional Papers 26: 1–452.
- Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallance IM, Wilm A, Lopez R, Thopson JD, Gibson TJ, Higgins DG (2007) Clustal W and Clustal X version 2.0. Bioinformatics 23: 2947–2948. https://doi.org/10.1093/bioinformatics/btm404
- Lobo J, Teixeira MA, Borges LM, Ferreira MS, Hollatz C, Gomes PA, Sousa R, Ravara A, Costa MH, Costa FO (2016) Starting a DNA barcode reference library for shallow water polychaetes from the southern European Atlantic coast. Molecular Ecology Resources 16(1): 298–313. https://doi.org/10.1111/1755-0998.12441
- Magalhães WF, Bailey-Brock JH (2010) Redescription of *Cirriformia crassicollis* (Kinberg, 1866) and *Timarete hawaiensis* (Hartman, 1956) new combination, (Polychaeta: Cirratulidae), endemic polychaetes to the Hawaiian Islands. Zootaxa 2625: 53–62.
- Magalháes WF, Seixas VC, Paiva PC, Elias R (2014) The Multitentaculate Cirratulidae of the Genera *Cirriformia* and *Timarete* (Annelida: Polychaeta) from Shallow Waters of Brazil. PLoS ONE 9(11): e112727. https://doi.org/10.1371/journal.pone.0112727
- Magalháes WF, Linse K, Wiklund H (2017) A new species of Raricirrus (Annelida: Cirratuliformia) from deep-water sunken wood off California. Zootaxa 4353: 51–68. https://doi.org/10.11646/zootaxa.4353.1.3
- Marenzeller E (1879) Südjapanische Anneliden. Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-naturwissenschaftliche Classe, Wien 41: 109–154.
- Monro CCA (1930) Polychaete worms. Discovery Reports, Cambridge 2: 1–222.
- Moore JP (1903) Polychaeta from the coastal slope of Japan and from Kamchatka and Bering Sea. Proceedings of the Academy of Natural Sciences of Philadelphia 55: 401–490.
- Paik EI (1989) Illustrated encyclopedia of fauna and flora of Korea, Vol. 31. Polychaeta. Ministry of Education, Seoul, 764 pp.
- Palumbi SR (1996) Nucleic acid II: the polymerase chain reaction. In: Hillis DM, Moritz C, Mable BK (Eds) Molecular Systematics. Sinauer Associates, Sunderland, MA, 205–247.
- Quatrefages A (1865) Histoire naturelle des Annelés marins et d'eau douce. Annélides et Géphyriens. Volume 1. Librarie Encyclopédique de Roret, Paris, 588 pp. https://doi.org/10.5962/bhl.title.122818
- Rousset V, Pleijel F, Rouse GW, Erséus C, Siddall ME (2007) A molecular phylogeny of annelids. Cladistics 23(1): 41–63. https://doi.org/10.1111/j.1096-0031.2006.00128.x
- Seixas VC, Zanol J, Magalháes WF, Paiva PC (2017) Genetic diversity of *Timarete punctata* (Annelida: Cirratulidae): Detection of pseudo-cryptic species and a potential biological

- invader. Estuarine, Coastal and Shelf Science 197: 214–220. https://doi.org/10.1016/j. ecss.2017.08.039
- Schmarda LK (1861) Neue wirbellose Thiere beobachtet und gesammelt auf einer Reise um die Erdr 1853 bis 1857. Turbellarien, Rotatorien und Anneliden. Zweite Hälfte. Wilhelm Engelmann, Leipzig, 164 pp.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. Molecular Biology and Evolution 30: 2725–2729. https://doi.org/10.1093/molbev/mst197
- Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Research 22(22): 4673–4680. https://doi.org/10.1093/nar/22.22.4673
- Weidhase M, Bleidorn C, Helm C (2014) Structure and anterior regeneration of musculature and nervous system in *Cirratulus* cf. *cirratus* (Cirratulidae, Annelida). Journal of Morphology 275: 1418–1430. https://doi.org/10.1002/jmor.20316
- Weidhase M, Bleidorn C, Simon CA (2016) On the taxonomy and phylogeny of *Ctenodrilus* (Annelida: Cirratulidae) with a first report from South Africa. Marine Biodiversity 46(1): 243–252. https://doi.org/10.1007/s12526-015-0355-3
- Zachs I (1933) K. faune kol'chatykh chervi Severo-Yaponskogo morya (Polychaeta). Gosudarstvennyi Gidrologicheskii Institut, Issledovaniia Morei SSSR. Leningrad 14: 125–137.
- Zanol J, Halanych K, Struck T, Fauchald K (2010) Phylogeny of the bristle worm family Eunicidae (Eunicida, Annelida) and the phylogenetic utility of noncongruent 16S, COI and 18S in combined analyses. Molecular Phylogenetics and Evolution 55: 660–676. https://doi.org/10.1016/j.ympev.2009.12.024