

Research Article

Taxonomic review of the Oriental genus *Polyplocia* Lestage, 1921 (Ephemeroptera, Euthyplociidae), with two new records for Thailand

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Abstract

Previously, the euthyplociid mayfly from Thailand was reported as *Polyplocia* spp. without designation to any nominal species. In this study, the genus *Polyplocia* Lestage, 1921 in Thailand was reviewed. Two species are recognized: *Polyplocia orientalis* Nguyen & Bae, 2003 and *P. nebulosa* Gonçalves & Peters, 2016. This is the first report of *P. nebulosa* in Continental Asia. Cytochrome c oxidase subunit I (COI) data and illustrations based on nymphal characters were used to confirm two different species in Thailand. Additional morphological characteristics of the nymph and eggs of the two species from Thailand are also shown in detail. The taxonomic status of *Polyplocia* in the Oriental region is reviewed. Keys to known nymphal and imaginal stages are provided and the distribution of *Polyplocia* in the Oriental region is discussed.

Key words: Burrowing mayfly, Continental Asia, distribution, habitat, Insular Asia, taxonomy



Academic editor: Eduardo Dominguez Received: 31 May 2023 Accepted: 23 August 2023 Published: 11 September 2023

ZooBank: https://zoobank. org/9CCA5878-F22D-4BE9-B78B-A267F4B3542C

Citation: Kwanboon S, Boonsoong B, Suttinun C (2023) Taxonomic review of the Oriental genus *Polyplocia* Lestage, 1921 (Ephemeroptera, Euthyplociidae), with two new records for Thailand. ZooKeys 1179: 197–217. https://doi.org/10.3897/zookeys.1179.107312

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Introduction

Lestage (1921) established the genus *Polyplocia*, and *Polyplocia vitalisi* Lestage, 1921 from Tonkin, Vietnam, was considered the type species based on one male imago. Ulmer (1939) described two new species *P. campylociella* Ulmer, 1939 and *P. crassinervis* Ulmer, 1939 based on a northern Borneo (now Sabah, Malaysia) subimago. Demoulin (1952) revised the imago characters of *Polyplocia* as having a transverse pronotum, forewing with MA fork at same level as Rs fork, and at least one intercalary vein in the cubital field with its base connected to CuP, forceps with only one segment and three caudal filaments on the abdomen (Gonçalves and Peters 2016) and provided new figures of *P. crassinervis* and *P. vitalisi*. Demoulin (1953) later described the female imago, also from Sarawak, and synonymized the species as *P. campylociella* (= *P. crassinervis*) based on wing venation. He later classified a potential nymph of *Polyplocia* for the first time from West Borneo (now Indonesia) as *P. ?crassinervis*, despite the synonymy, and identified the nymphal characters: long tusks with truncated apex, antennae almost the same length as the tusks,

foretibiae with short apical projection, foretarsi with apical projection, and gills I with two lamellae (Demoulin 1966). Thirty-seven years later, Nguyen and Bae (2003a) described a new species, *P. orientalis* Nguyen & Bae, 2003, based on nymphs from Dak Lak, Vietnam and stated that this nymph could be associated with *P. vitalisi*. Gonçalves and Peters (2016) discovered a new species, *P. nebulosa* Gonçalves & Peters, 2016, based on male and female imagos from Malaysia's Sabah State, and provisional nymphs assigned to this species are also described. A description of the structure of the chorion was also provided and recommended as a useful tool for recognising species (Gonçalves and Peters 2016). Lastly, Zheng et al. (2023) reported the first nymph-imago association for the genus in *P. orientalis* from China and expanded the distribution of this species in the Oriental region. Molecular evidence based on cytochrome c oxidase subunit I (COI) revealed an intraspecific distance of 10% between the Vietnamese and Chinese specimens.

To date, the genus Polyplocia is composed of four valid species: P. vitalisi reported only in an adult stage, P. campylociella reported in adult stage with possible nymphal stage, P. orientalis reported in both stages and P. nebulosa reported in adult stage with a possible nymphal stage. The distribution of this genus encompasses the Oriental Realm (Lestage 1921; Ulmer 1939; Demoulin 1966; Nguyen and Bae 2003a; Gonçalves and Peters 2016; Zheng et al. 2023). Polyplocia spp. nymphs from Thailand were mentioned by Kluge (2004) and Gonçalves and Peters (2016). These nymphs may be related to P. orientalis or belong to another species, as they did not possess paired black anterolateral marks on abdominal sterna. Conversely, the genitalia of the male nymphs of Polyplocia spp. from Thailand also had T-shaped penes, as do the nymphs of P. nebulosa (Gonçalves and Peters 2016). In the present study, two species of the genus Polyplocia based on nymphs from Thailand are recorded using morphological and molecular approaches. Some additional descriptions on the nymphal and chorion structures are mentioned. Keys to known species in both stages are provided. The distribution of this genus in Thailand is studied and information on the biological aspects of *Polyplocia* is provided.

Materials and methods

The specimens were collected by hand-picking from headwater streams in the northern and the western parts of Thailand. (Table 1, GPS map versatile navigator (Garmin eTrex 10)). The specimens were preserved in 100% ethanol for molecular and morphological studies. The mature nymphs were reared using earthenware pots connected to an oxygen pump until emergence of winged stages. Measurements (given in mm) and photographs were taken using a NIKON SMZ800 stereoscopic microscope. For scanning electron microscopy (SEM), eggs were dried in a critical point drier (CPD7501) and coated with gold (Sputter Coater SC7620). The SEM photographs were obtained with a FEI Quanta 450 SEM. Final plates were prepared with Adobe Photoshop CC 2022.

Selected specimens were dissected for DNA extraction. Total DNA was extracted using a genomic DNA purification kit (NucleoSpin, Macherey-Nagel, Germany), following the manufacturer's protocol. The COI amplification was performed using LCO1490 and HCO2198 (Folmer et al. 1994). The polymerase

chain reaction (PCR) conditions and procedure were as described by Kwanboon et al. (2021). The PCR products were purified using a Gel and PCR Clean-up Kit (NucleoSpin, Macherey-Nagel, Germany) and were sequenced by ATGC Co., Ltd (Thailand). Other analysed Polyplocia orientalis sequences were obtained from GenBank (OP347109; OP962407). Potamanthus formosus Eaton, 1892, retrieved from GenBank (MZ453438), was used as an outgroup. The genetic distances between species were determined using Kimura-2-parameter distances (Kimura 1980), calculated with the MEGA11 program (Tamura et al. 2021). Sequence alignment and editing were performed using ClustalW in MEGA11. A phylogenetic tree was analysed by the maximum likelihood (ML) method and the most appropriate evolutionary model was calculate using the Find Best DNA/Protein Models (ML) option test provided with MEGA11. The Tamura-Nei 93 model and a proportional discrete Gamma distribution (TN93+G) was performed with MEGA11 using the likelihood-ratchet method with 1000 bootstrap replicates. The GenBank accession numbers are given in Table 2, nomenclature of gene sequences follows Chakrabarty et al. (2013). Nucleotide sequences obtained in this study have been deposited in GenBank. The distribution map was generated with the SimpleMappr software (https://simplemappr.net; Shorthouse 2010).

The material was deposited in the collection of the Zoological Museum at Kasetsart University in Bangkok, Thailand (ZMKU) and the Veterinary Anatomy and Pathology Museum, Chiang Mai University, Thailand (VMCMU).

We followed all guidelines of the Animal Ethics Committee of Kasetsart University (approval no. ACKU61-SCI-029) for collecting the mayfly specimens.

Table 1. GPS coordinates of locations of examined specimens.

Species	Locality	GPS coordinates
Polyplocia orientalis	Chiang Rai (CR)	19°26'53.7"N, 99°41'83.6"E
	Chiang Mai (CM)	19°19'31.1"N, 99°58'84.6"E
Polyplocia nebulosa	Kanchanaburi (KN)	14°33'10.8"N, 98°33'94.3"E
	Phetchaburi (PE)	12°28'53.2"N, 99°15'23.0"E
Polyplocia campylociella	Sarawak, Malaysia	4°02'51.0"N, 114°50'11.9"E

Table 2. Sequenced specimens of the genus *Polyplocia*.

Species	Locality	Code	GenBank #	GenSeq Nomenclature
P. nebulosa	Kanchanaburi, Thailand	PC02KN	OR366860	genseq-4 COI
	Phetchaburi, Thailand	PC01PE	OR366862	genseq-4 COI
	Kanchanaburi, Thailand	PC08PE	OR366863	genseq-4 COI
P. orientalis	Chiang Rai, Thailand	PC04CR	OR366857	genseq-4 COI
	Chiang Rai, Thailand	PC05CR	OR366858	genseq-4 COI
	Chiang Rai, Thailand	PC06CR	OR366861	genseq-4 COI
	Chiang Mai, Thailand	PC01CM	OR366859	genseq-4 COI
	Vietnam		OP347109	
	Yunnan, China		OP962407	
P. campylociella	Sarawak, Malaysia		OR366864	genseq-4 COI

Results

Taxonomic review and additional description of nymphal stage

Nymphs of *Polyplocia* spp. from northern of Thailand of Gonçalves and Peters (2016) are assigned to *P. orientalis*. The additional description of *P. orientalis* is based on material from the northern of Thailand (Chiang Rai and Chiang Mai provinces) and variations regarding these populations are given. Five nymphs of *P. nebulosa* are recorded from western Thailand (Kanchanaburi and Phetchaburi provinces). A map of *Polyplocia* species distribution in Oriental region is given in Fig. 1.

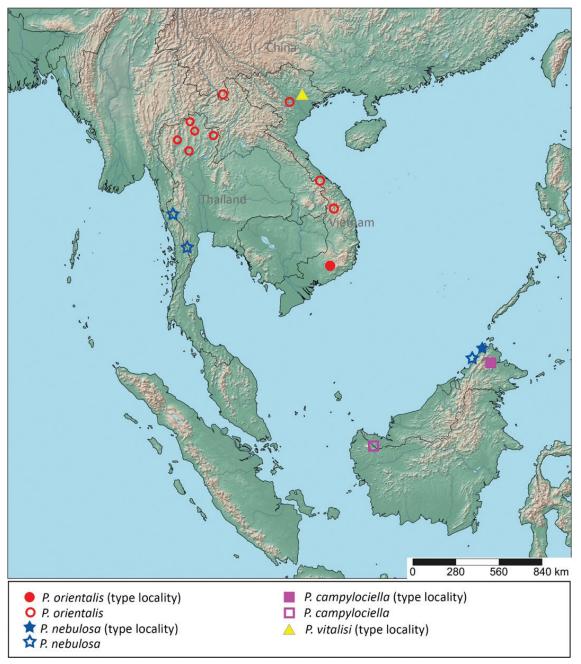


Figure 1. Distribution map of the genus *Polyplocia* in the Oriental region.

Order Ephemeroptera Hyatt & Arms, 1891 Family Euthyplociidae Edmunds & Traver, 1954

Genus Polyplocia Lestage, 1921

Polyplocia Lestage, 1921: 212 (Type: Polyplocia vitalisi); Lestage 1924: 5; Ulmer 1932: 205; Ulmer 1939: 466; Demoulin 1952: 9; Demoulin 1966: 137; Nguyen and Bae 2003a: 280; Gonçalves and Peters 2016: 553; Zheng et al. 2023: 2.

Diagnosis. Imago: i) transverse pronotum, ii) forewing with MA fork at same level as Rs fork, and at least one intercalary vein in the cubital field with its base connected to CuP, iii) forceps with only one segment, and iv) three caudal filaments on abdomen (Demoulin 1952; Gonçalves and Peters 2016). **Nymph:** i) mandibles very long, with apex obliquely truncated, ii) antennae almost the same length as the mandibles, iii) anterior angles of the pronotum protruding into points iv) foretibia terminated by an apophysis provided with an internal brush, v) fore tarsi with a distal unguiform extension, vi) gill I bilamellated, and vii) bare terminal filament (Demoulin 1966; Gonçalves and Peters 2016: 554).

Polyplocia vitalisi Lestage, 1921

Polyplocia vitalisi Lestage, 1921: 212, original description (male imago). Polyplocia vitalisi (Ulmer 1939: 467, figs 7–9, male imago). Polyplocia vitalisi (Demoulin 1952: 16, fig. 3, male imago). Polyplocia vitalisi (Gonçalves and Peters 2016: 558, male imago).

Material examined. None.

Diagnosis. Imago: i) transparent wing, ii) membrane of forewing with purplish colour on C and Sc fields, iii) abdominal sterna without marking, and vi) penis lobe with a smooth apical border (Lestage 1921).

Description. Male imago. See Lestage (1921).

Female imago. Unknown.

Nymph. Unknown.

Distribution. Tonkin (Vietnam).

Remark. This species was described based on one male imago from Tonkin, Vietnam.

Polyplocia campylociella Ulmer, 1939

= P. crassinervis Ulmer, 1939 (Demoulin 1953).

Polyplocia campylociella Ulmer, 1939: 468, figs 10–11, original description (female subimago).

Polyplocia crassinervis Ulmer, 1939: 470, figs 12–15, male subimago).

Polyplocia crassinervis (Demoulin 1952: 18, fig. 4, male imago).

Polyplocia campylociella (Demoulin 1953: 1, fig. 1, female imago = P. crassinervis). Polyplocia campylociella (Demoulin 1966: 137, fig. 1, immature nymph). Polyplocia campylociella (Gonçalves and Peters 2016: 558, male and female imago).

Material examined. MALAYSIA: One immature nymph in alcohol, deposited in ZMKU, Sarawak, Marudi district, Miri division, Gunung Mulu, Sungai Paku, 4°02′51.0″N, 114°50′11.9″E, ~240 m, 13.VI.2023, B. Boonsoong leg.

Diagnosis. The imago of *P. campylociella* can be distinguished from those of other *Polyplocia* species based on the following characteristics: i) wings with dark clouds around cross veins and margins, ii) membrane of forewing with little dark colour on C and Sc fields, iii) styliger plate rounded, and iv) penis V-shaped without dorsal spine (Demoulin 1952).

Description. Male imago. See Demoulin (1952).

Female imago. See Demoulin (1953).

Nymph. See Demoulin (1966). In this study, only one immature nymph was collected. The specimen possessed a pair of anterolateral black marks on abdominal sterna. COI sequences were analysed from the sample.

Distribution. Malaysia (Sarawak), Indonesia (Sambas).

Remarks. The possible nymph of *P. campylociella* was described based on one immature nymph by Demoulin (1966) from Sambas (Indonesia). Additional illustrations of *P. campylociella* are provided in Miller et al. (2018) and Jacobus et al. (2019). The distribution of *P. campylociella* is limited to Insular Asia.

Polyplocia nebulosa Gonçalves & Peters, 2016

Figs 2, 3

Polyplocia nebulosa Gonçalves & Peters, 2016: 554, figs 1–21, original description (male and female imago, female subimago, egg, possible nymph).

Material examined. THAILAND: Two nymph in alcohol, deposited in ZMKU, Kanchanaburi province, Thong Pha Phum district, Pat Sadu Klang, 14°33′10.8″N, 98°33′94.3″E, 349 m, 20.II.2016, B. Boonsoong leg. Three nymphs in alcohol, deposited in ZMKU, Phetchaburi province, Kaeng Krachan district, Ban Krang river, 12°28′53.2″N, 99°15′23.0″E, 386 m, 11.II.2023, A. Vitheepradit leg.

Diagnosis. Imago: i) wings with longitudinal veins light brown and cross veins brown, cross veins with narrow dark brown clouds and margins tinged with brown, ii) sterna II–IX with a pair of blackish brown anterolateral marks, iii) penes broad, T-shaped, fused, with medial groove extending from apex to half-length of penes; large laterally projecting lobes apically rounded with a small dorsolateral spine; basal outer margin of each lobe sclerotized, iv) styliger plate short and straight, not projected posteriorly, and v) eggs 265–267 μm in length and 170–186 μm in width, barrel shaped, without polar caps or other attachment structures, with one visible micropyle and chorion forming an irregular mesh with raised ridges, mesh size from 3.8–8.0 μm (Gonçalves and Peters 2016). **Nymph:** i) larger spines on distal ½ of tusk, and ii) sterna with a pair of anterolateral black marks (Gonçalves and Peters 2016).

Description. Male imago. See Gonçalves and Peters (2016).

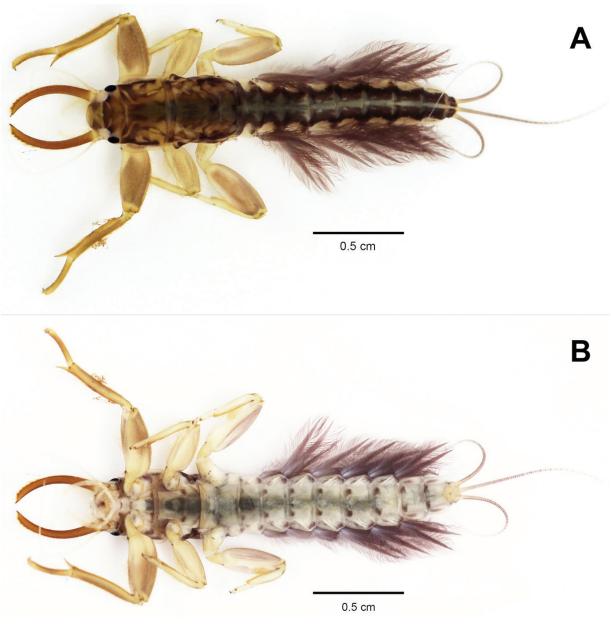


Figure 2. Habitus of nymph of Polyplocia nebulosa: A dorsal view B ventral view.

Female imago. See Gonçalves and Peters (2016).

Nymph. See Gonçalves and Peters (2016).

Additional description. Mandibular tusks (Fig. 3E) strongly arched inward (20.7° in curvature, for angle measurement see Kwanboon et al. (2021)). Labrum (Fig. 3A): median concave with shallow emargination, with long and short setae on sub-basal, subapical and anterior margins. Hypopharynx (Fig. 3B): lingua cordiform, superlingua slightly extended laterally with dense setae on distal margin. Labium (Fig. 3C): paraglossae with dense setae on ventral margin, meet above glossae; glossae drop-shaped; palpi with long setae on outer margin; 3rd segment much broader, acutely rounded at the apex. Maxilla as in Fig. 3D: 1st segment with a few fine setae on outer margin; 2nd segment with long hair-like setae in both inner and outer margin; 3rd segment long, at least 2× longer than 2nd segment, apically pointed with numerous long, hair-like setae.

Distribution. Malaysia (Sabah), Thailand (Kanchanaburi, Phetchaburi).



Figure 3. Polyplocia nebulosa, nymphal morphology: A labrum (dorsal view) B hypopharynx (ventral view) C labium (ventral view) D left maxilla (ventral view) E mandibular tusk. Scale bar: 0.5 mm.

Polyplocia orientalis Nguyen & Bae, 2003

Figs 4-6

Polyplocia orientalis Nguyen & Bae, 2003a: 280, figs 1–2, original description (nymph).

Polyplocia orientalis (Gonçalves and Peters 2016: 558, nymph).

Polyplocia spp. (Gonçalves and Peters 2016: 558, possible nymph (Thailand)). Polyplocia orientalis (Zheng et al. 2023: 2, figs 1–9, male and female imago, egg, nymph).

Material examined. THAILAND: Eleven nymphs in alcohol, deposited in ZMKU, Chiang Rai province, Phan district, Pu Kaeng waterfall, 19°26'53.7"N, 99°41'83.6"E, 540 m, 5.III.2021, S. Kwanboon leg. Three nymphs in alcohol, deposited in VMCMU, Chiang Rai province, Phan district, Pu Kaeng waterfall, 19°26'53.7"N, 99°41'83.6"E, 540 m, 29.I.2023, S. Chanaporn leg. Two nymphs in alcohol, deposited in ZMKU, Thailand, Chiang Mai province, Chiang Dao district, Huay Mae Mae, 19°19'31.1"N, 99°58'84.6"E, 809 m, 20.XI.2018, C. Damrong leg.

Diagnosis. Imago: i) membrane of wings transparent and colourless, and ii) T-shaped male penis with apical depression on both lobes (Zheng et al. 2023). **Nymph:** i) large body size (25.0–46.4 mm), ii) yellowish abdominal sterna without anterolateral black mark, iii) spines on 1/3 of tusk length distally, and iv) apically expanded dorsal lobe of gill I (Nguyen and Bae 2003a; Zheng et al. 2023).

Description. Male imago. See Zheng et al. (2023).

Female imago. See Zheng et al. (2023).

Nymph. See also Nguyen and Bae (2003a) and Zheng et al. (2023).

Variability and additional description (Thai specimens). Nymph. Male body length 17.22 mm; cerci length 11.78 mm; median filament length 10.5 mm. Female body length 27.9 mm; cerci length 18.5 mm; median filament length 16.2 mm.

Head. Length 2× of maximum width; narrower than pronotum. Compound eye black on dorsolateral margin. Antenna 8.8 mm in length; scape with at least three short setae.

Mouthpart. Labrum (Fig. 5A) shallow, concave at anteromedian margin, with a tuft of dense long, simple setae; anterior margin with a row of 14-16 long, simple setae; dorsal surface with long, fine setae and short, simple setae scattered over area. Mandibular tusks (Fig. 5E) strongly arched inward (18.6° in curvature); a row of simple setae on base of tusks; ventrally almost bare. Right mandible without prostheca. Left mandible with prostheca as long as incisors, truncate, broader apically. Maxillary palp (Fig. 5D) 1st segment with a few of long, fine setae on outer margin; 2nd segment with lateral long, hair-like setae in both inner and outer margin; 3rd segment long, at least 6 × longer than wide, apically pointed, with numerous of long, hair-like setae on lateral inner margin and scattered over half of segment apically. Hypopharynx (Fig. 5B) lingua cordiform; superlingua extended laterally. Labium (Fig. 5C) paraglossae articulate above glossae; labial palpi 1st segment with long, hair-like setae on outer margin and fine, simple setae on inner margin; 2nd segment with long, hair-like setae on outer margin; 3rd segment much broader than 2nd segment, apically truncated almost straight with a tuft of stout, simple setae, with long, hair-like setae on outer margin and fine, simple setae on inner margin.

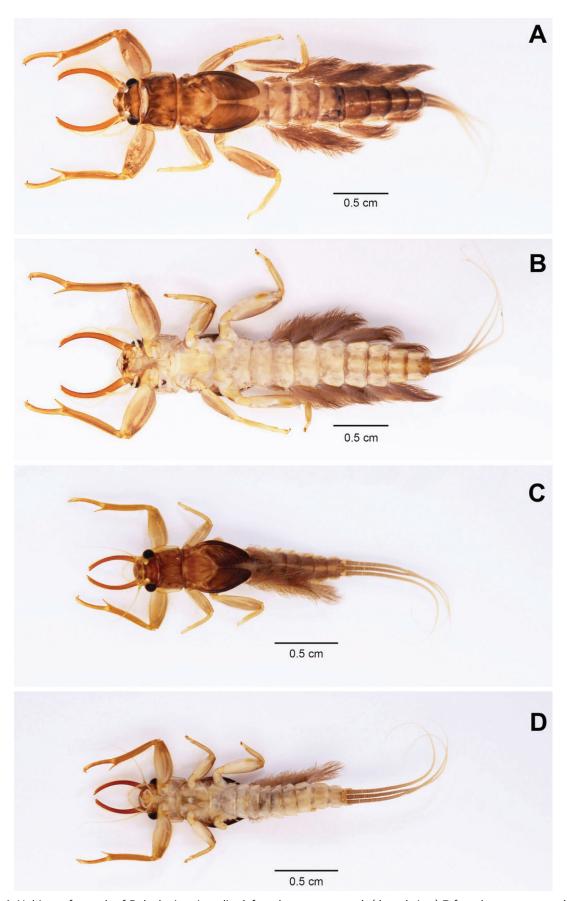


Figure 4. Habitus of nymph of *Polyplocia orientalis*: **A** female mature nymph (dorsal view) **B** female mature nymph (ventral view) **C** male mature nymph (dorsal view) **D** male mature nymph (ventral view).

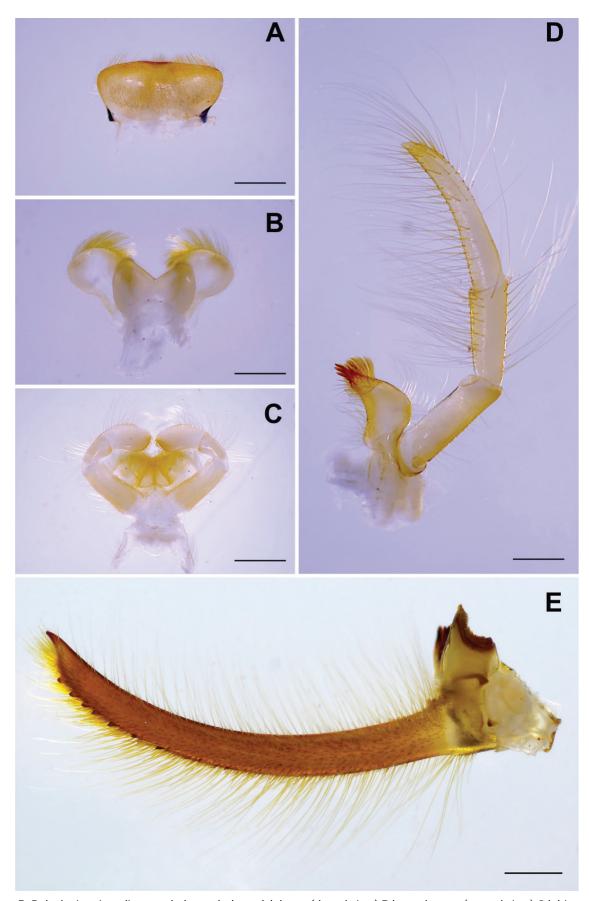


Figure 5. Polyplocia orientalis, nymphal morphology: A labrum (dorsal view) B hypopharynx (ventral view) C labium (ventral view) D left maxilla (ventral view) E mandibular tusk. Scale bar: 0.5 mm.

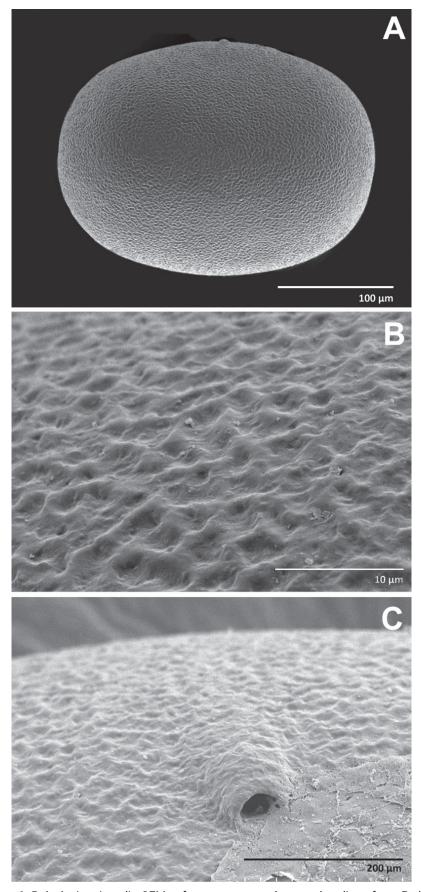


Figure 6. Polyplocia orientalis, SEMs of egg structures: $\bf A$ general outline of egg $\bf B$ chorion surface $\bf C$ micropyle.

Legs. Ratio of forelegs: midlegs: hindlegs 1:0.7:0.7; ratio of foreleg segments 1:1.2:0.7:0.4; ratio of midlegs segments 1:1:0.4:0.2 with moderately developed setae; ratio of hindleg segments 1:0.6:0.2:0.1, lack setae on femora.

Female subimago. *Egg.* (Fig. 6) Dissected from female subimago; length 272 μ m, width 214 μ m; oval-shaped; no polar caps or other attachment structures; rough chorionic surface, mesh-like with irregular raised ridges (Fig. 6B); two visible linear micropyles formed with micropyle canal on the surface (Fig. 6C).

In this study, Thai *Polyplocia orientalis* showed some variation in these characters combined: i) Labrum slightly concave on anterior margin, nearly straight, ii) tusks cylindrical pointed apically, strongly arched inward (18.6° curvature) and outer margin with 7–9 larger spines on 1/3 of tusk length distally, and iii) eggs oval-shaped with two visible micropores.

Distribution. China (Yunnan), Vietnam (Dak Lak, Lam Dong, Thua Thien Hue), Thailand (Chiang Rai, Chiang Mai, Phrae, Phayao, Nan) (Nguyen and Bae 2003a, Gonçalves and Peters 2016, Zheng et al. 2023, this study).

Key to mature nymphs of Polyplocia

1	Abdominal sterna without a pair of anterolateral black marks (Fig. 4B, D).
	P. orientalis Nguyeun & Bae, 2003
_	Abdominal sterna with a pair of anterolateral black marks (Fig. 2B)
	P nobulase Concelves & Paters 2016 / P compulacialle Illmer 1030*

*The details of nymph of *P. campylociella* are insufficient because of the limited description by Demoulin (1966) and immaturity of the specimen in this study. However, ratio of arrangement of spines on mandibular tusk vs. tusk length of mature nymph (Jacobus et al. 2019) seems different from *P. nebulosa*.

Key to imagos of *Polyplocia* (modified from Gonçalves and Peters 2016 and Zheng et al. 2023)

1	Veins with dark brown clouds around cross veins and margins of wings with brownish tinge; membrane of forewing without purplish color on C and Sc fields (Sc field may be a little darker); abdominal sterna with a pair of blackish marks on anterolateral margins
_	Veins without dark clouds around cross veins and wing margins translucent; membrane of forewing colored only on C and Sc fields, purplish; abdominal sterna without markings
2	Styliger plate rounded and projected, penis V-shaped, penes apparently without dorsal spine
-	Styliger plate not projected, short and straight, penis T-shaped, each lobe of penes with small dorsal spine laterally directed
3	Both penis lobes have an apical depression
	P. orientalis Nguyen & Bae, 2003
_	Penis lobes with smooth apical margin

Biological aspects

The genus *Polyplocia* was collected at an altitude of 300–800 metres above sea level; *Polyplocia nebulosa* was found in a headwater stream in the forest, while *P. orientalis* was found in a headwater stream and a limestone waterfall quite disturbed by tourist activities. The nymphs were found on the underside of cobbles in slow-flowing waters at the margins of the stream (Figs 7, 8). The riverbed was covered in cobble, gravel and sand. The nymphs were frequently collected alongside the potamanthiid mayfly *Rhoenanthus magnificus*.

Molecular analysis

The partial sequences of the mitochondrial COI gene (658 bp) of the two species found in Thailand and P. campylociella from Malaysia were obtained from specimens of each locality (Table 2) to investigate the species delineation. The K2P analysis revealed that the intraspecific genetic distances of P. orientalis from Thailand vary between 0 and 10.7% and the intraspecific genetic distance of P. nebulosa from Thailand is 0.15-2.02%. The interspecific distances between Thai P. orientalis and P. nebulosa are high, ranging from 17.7-23.6% (Table 3). The sequences of four specimens of P. orientalis from Thailand and two sequences of P. orientalis from Vietnam and China present a K2P intraspecific distance ranging between 6.25% and 10.6%, respectively. The intraspecific distances of P. orientalis (Thailand) and P. orientalis (Vietnam) range between 9.51% and 10.6%, while P. orientalis (Thailand) and P. orientalis (China) range between 6.23% and 7.41%. The interspecific distances of all P. orientalis and P. nebulosa are 17.7%-23.6% and the interspecific distances of all P. orientalis and P. campylociella are 19.1%-21.7%, while the interspecific distances of P. nebulosa and P. campylociella are very high (20.7%–23.1%) (Table 3).



Figure 7. Habitat of Polyplocia nebulosa.



Figure 8. Habitat of *Polyplocia orientalis*: **A** Pu Kaeng waterfall (limestone waterfall) **B** microhabitat.

Table 3. Genetic distances (COI) between sequenced specimens using the Kimura 2-parameter.

	Species	1	2	3	4	5
1	P. orientalis (Thailand)	0.00-10.7				
2	P. orientalis (China)	6.23-7.41	_			
3	P. orientalis (Vietnam)	9.51-10.6	10.0	-		
4	P. nebulosa (Thailand)	17.7-23.6	20.7-21.9	16.7-20.1	0.15-2.02	
5	P. campylociella (Malaysia)	19.1-20.6	19.8	21.7	20.7-23.1	_
6	Potamanthus formosus	22.8-24.1	23.8	23.0	23.0-24.3	25.2

COI sequences analysis was built by maximum likelihood (ML) using MEGA11 (Fig. 9). Ten sequences of *Polyplocia* were separated into two main distinct clades: the first is the *P. nebulosa* and *P. campylociella* clade which is further separated into two clades of *P. nebulosa* (Thailand) and *P. campylociella* (Malaysia), while the second is a *P. orientalis* clade which is further separated into two clades of *P. orientalis* (Vietnam) and *P. orientalis* (Thailand, China).

Discussion

Specimens of the *Polyplocia* from Thailand were studied based on the following nymphal characters defined by Demoulin (1966). Of the nominal species that are newly reported to the country, *P. orientalis* is distributed in northern Thailand (Chiang Rai and Chiang Mai provinces). Some variations in the morphological characters of the Thai *P. orientalis* were observed in this study (Table 4): Labrum slightly concave on anterior margin, nearly straight; the mandibular tusks are cylindrical, pointed apically and strongly arched inward (18.6° in curvature), with 7–9 spines on 1/3 of the tusk length distally. The second species, *P. nebulosa*, is distributed in western Thailand (Kanchanaburi and Phetchaburi provinces). Nymphs of these two species from Thailand can be easily separated by the presence of anterolateral black marks ventrally on *P. nebulosa* abdomen and their absence in *P. orientalis*. As we have only a very limited description of *P. campylociella* nymphs, their differences are not discussed in this study. The combinations of characters used to differentiate nymphs of known species are listed in Table 4.

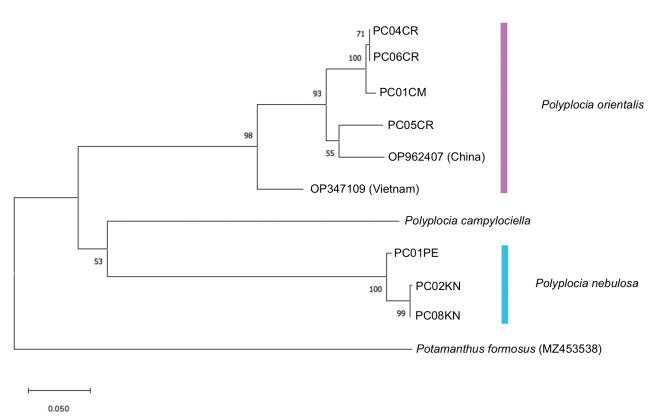


Figure 9. The COI phylogenetic construction based on the maximum likelihood (ML) analysis of three *Polyplocia* species, *Potamanthus formosus* was used as the outgroup. Scale bars refer to average of base substitution per site.

Table 4. Selected nymphal characters of known species of the genus Polyplocia.

Characters		P. orientalis Nguyen & Bae, 2003	P. nebulosa Gonçalves & Peters, 2016	P. campylociella Ulmer, 1939	
Head	antenna length vs. tusk length	longer	longer	slightly longer	
Labrum	anterior margin	medially slightly concave with shallow emargination	medially concave with shallow emargination	medially concave (from fig. 1b)	
Mandibular	length of tusks vs. length of head	2×-3×	2×	?	
tusks	inward curvature	18.6°-21.3°	20.7° (in this study)	?	
	number of spines	3-12	8–9 (fig. 18 Gonçalves and Peters 2016; this study)	?	
	presence of large spines	Apical 1/3	Apical 1/2	Apical 1/3 (Jacobus et al. 2019)	
Legs	length of apical spine of the foretibiae vs. length of the foretarsi	1/3-1/4	less than 1/2, 1/3 (in this study)	?	
Abdomen	sterna	whitish	a pair of anterolateral black marks	a pair of anterolateral black marks	
Winged	associated	yes	possible nymph	possible nymph	
stage	eggs shape (subimago)	oval; barrel	barrel	?	
	eggs length vs. width	1.3×	1.6×	?	
	number of micropores	2	1	?	
Distribution		Vietnam, China, Northern Thailand	Borneo, Malaysia, Western Thailand	Borneo, Malaysia	
References		Nguyen and Bae (2003a); Zheng et al. (2023); this study	Gonçalves and Peters (2016); this study	Demoulin (1966)	

The egg structure of all *Polyplocia* species has a similar chorionic surface pattern that forms an irregular mesh with raised ridges; however, we found small differences between the eggs of Thai and Chinese specimens of *P. orientalis*. The specimen from Thailand is oval-shaped, with two visible micropores, while *P. orientalis* from China and Vietnam has a barrel-shaped egg with no mention of a micropore (Nguyen and Bae 2003a; Zheng et al. 2023). The egg of the Thai *P. orientalis* shows different characters than *P. nebulosa* in terms of shape; the length vs width ratio, and the number of visible micropores. Gonçalves and Peters (2016) mention that the structure of the chorion should be considered as a foundation for future study of the genus. The present study confirmed that the structure of the chorion is useful for recognising the species.

The first molecular study of the genus *Polyplocia* was conducted by Zheng et al. (2023). The intraspecific distance of *P. orientalis* from Vietnam and China was mentioned to be as high as 10%. In our study, the maximum intraspecific distance between the Thai *P. orientalis* was relatively high, at 11%. When we compared the intraspecific distance of specimens from Thailand and Vietnam (10–11%) and Thailand and China (6–7%), the result from the genetic point of view also supported assigning the *Polyplocia* from northern Thailand to *P. orientalis*, as the maximum intraspecific distance of all specimens of *P. orientalis* is 11%. The genetic distances between the *P. orientalis* from northern Thailand and the sequence of *P. nebulosa* from western Thailand support the separation of this genus in Thailand into two species due to the high interspecific distance that ranges between 18% and 24%. The genetic distances of all known sequences of *P. orientalis* (sequences from Vietnam and China were added) and *P. nebulosa*

from western Thailand also confirmed the separation into two species, with an interspecific distance of 18–24%. The result of interspecific distances between *P. campylociella* from Malaysia and two species from the Continental Asia are relatively high at the maximum 22% for *P. orientalis* and 23% for *P. nebulosa*. Kwanboon et al. (2021) also reported a variation in the interspecific distance as high as 14–20% for congeneric burrowing mayflies in Southeast Asia. The ML tree consists of two main clades; the *P. nebulosa* and *P. campylociella* clades and a *P. orientalis* clade (Fig. 9). The *P. nebulosa* and *P. campylociella* clades could represent two closely related species since they share some morphological characteristics, such as an abdominal sternum with two blackish marks on the anterolateral edges and wings with dark clouds around the cross veins and margins (Table 4). We recommend that molecular studies (i.e., the COI gene) be included as a basis for future studies of all burrowing mayflies, based on our findings and the studies of other burrowing mayflies.

Polyplocia nymphs from Thailand were mentioned by Kluge (2004) as Polyplocia sp. (locality and character are unknown) and by Gonçalves and Peters (2016) as Polyplocia spp. from many localities in northern Thailand. In this study, we collected specimens from one of the localities in which Polyplocia spp. were found by Gonçalves and Peters (2016) (Chiang Rai Province, Doi Luang National Park, Namtok Pu Keang). Therefore, the Polyplocia spp. of Gonçalves and Peters (2016) are proposed to be P. orientalis like the northern Thailand Polyplocia from this study. Although, the Polyplocia sp. of Kluge (2004) is not assigned to either P. nebulosa or P. orientalis, as the data are still lacking.

The distribution of the genus Polyplocia Lestage, 1921 in the Oriental region is shown in Fig. 1. Polyplocia orientalis was reported in Vietnam (Dak Lak, Lam Dong and Thua Thien Hue) and China (Yunnan) and, in this study, in the northern part of Thailand (Chiang Rai, Chiang Mai, Phrae, Phayao and Nan). Therefore, this species has the potential to be assigned as the dominent species of Polyplocia in mainland Asia in the Oriental region. Surprisingly, the discovery of P. nebulosa in Thailand is the first report of this species from Continental Asia, as all other specimens were reported from East Malaysia (Sabah) or Insular Asia. This wide range of distribution of mayflies between Continental and Insular Asia has been reported in many families. For example, Potamanthellus caenoides Ulmer, 1939 (Neoephemeridae) is known as a widespread mayfly in the Oriental region. This species has also been recorded from the Insular Asia, in Indonesia (Sumatra, Java, Bali, Lombok and Flores), Malaysia (Sabah and Sarawak) and the Philippines (Mindanao), and in Continental Asia, in Malaysia (the Malay peninsula), Thailand (Chiang Mai), India (western Ghat) and Vietnam (Dak Lak) (Bae and McCafferty 1998; Nguyen and Bae 2003b; Sartori et al. 2003; Selvakumar et al. 2015; Banazair and Christopher 2019). Such wide distribution is also observed for the burrowing mayfly Rhoenanthus (Rhoenanthus) speciosus Eaton, 1881 of the family Potamanthidae. This species was reported from the Insular Asia, in Indonesia (Sumatra) and East Malaysia (Sabah) and from Continental Asia, in west Malaysia (Pahang) and Thailand (Songkla and Narathiwat) (Bae and McCafferty 1991; Parnrong et al. 2002; Kwanboon et al. 2021). The most diverse family in the Oriental region, Baetidae, also has a widespread species, Labiobaetis moriharai Müller-Liebenau, 1984. This species was recorded in Insular Asia in Brunei (Temborong) and Malaysia (Sabah) and in Continental Asia in Malaysia (Selangor) and Vietnam (Dong Nai) (Kaltenbach

and Gattolliat 2020). The distribution of *P. campylociella* is limited to eastern Malaysia and the distribution of the type species *P. vitalisi* is limited to the type locality (Tonkin, Vietnam); the nymph remains a mystery.

Our study allows us to conclude that the genus *Polyplocia* in Thailand is represented by two species, *P. orientalis* and *P. nebulosa*, based on a combination of different morphological characters, egg characters and molecular evidence. We have reported the first nominal species records of this family in Thailand and expanded the distribution of these two species. The distribution of *P. orientalis* in Thailand is limited to the northern parts and mainland Asia, while the distribution of *P. nebulosa* extends from Insular Asia to Continental Asia, in western Thailand in our study. We expect to find a broader distribution of this genus in Thailand, especially in the southern and the eastern parts, as we gain a more in-depth understanding of the distribution pattern of this genus in the Oriental region.

Acknowledgements

This research was partially supported by Chiang Mai University, the Faculty of Veterinary Medicine, Chiang Mai University, and the Centre of Excellence on Biodiversity (BDC) Office of Higher Education Commission. We are most grateful to Assistant Professor Dr. Akekawat Vitheepradit (Department of Entomology, Faculty of Agriculture, Kasetsart University) who kindly provide specimens for this study. We are thankful to Dr. Chonlakarn Auychinda (Department of Biology, Faculty of Science, Silpakorn University) for his support with lab work and preparation of the COI barcode. Sarawak Forestry Permission to conduct research on biological resources - Permit SFC.810-4/6/1 (2023) - 023. We are most grateful to our colleagues for assistance during field trips. Further, we would like to thank the Department of Veterinary Biosciences and Veterinary Public Health, Faculty of Veterinary Medicine, Chiang Mai University and Department of Zoology, the Faculty of Science at Kasetsart University in Bangkok for their assistance and use of their facilities. The authors are also grateful to the reviewers for their valuable recommendations and comments on the manuscript.

Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

Funding

This research was partially supported by Chiang Mai University for research funding through the Research Center for Veterinary Biosciences and Veterinary Public Health, Faculty of Veterinary Medicine.

Author contributions

Sedtawut Kwanboon contributed in the process of Collected the data; Contributed data or analysis tools; Wrote the paper. Boonsatien Boonsoong contributed in the process of

Conceived and designed the analysis; Collected the data; Wrote the paper. Chanaporn Suttinun contributed in the process of Collected the data; Contributed data or analysis tools; Performed the analysis; Wrote the paper.

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Data availability

All of the data that support the findings of this study are available in the main text.

References

- Bae YJ, McCafferty WP (1991) Phylogenetic Systematics of the Potamanthidae (Ephemeroptera). Transactions of the American Entomological Society 117(3/4): 1–143.
- Bae YJ, McCafferty WP (1998) Phylogenetic systematics and biogeography of the Neoephemeridae (Ephemeroptera: Pannota). Aquatic Insects 20(1): 35–68. https://doi. org/10.1076/aqin.20.1.35.4489
- Banazair OA, Christopher G (2019) A new record of *Potamanthellus caenoides* Ulmer, 1939 (Ephemeroptera: Neoephemeridae) from the Chalakudi River, Southern Western Ghats of India. International Journal of Aquatic Biology 7(1): 35–37. https://doi.org/10.22034/ijab.v7i1.555
- Chakrabarty P, Warren M, Page LM, Baldwin CC (2013) GenSeq: An updated nomenclature and ranking for genetic sequences from type and non-type sources. ZooKeys 346: 29–41. https://doi.org/10.3897/zookeys.346.5753
- Demoulin G (1952) Contibution à l'étude des Ephoronidae Euthyplociinae (Insectes Ephéméroptères). Bulletin de l'Institut Royal des Sciences Naturelles de Belgique 28(45): 1–22.
- Demoulin G (1953) A propos des *Polyplocia* de Borneo (Insectes Ephéméroptères). Bulletin de l'Institut Royal des Sciences Naturelles de Belgique 29(19): 1–12.
- Demoulin G (1966) Contribution à l'étude des Euthyplociidae III (Insectes Ephéméroptères). Zoölogische Mededeelingen 41(7): 137–141.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome C oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294–299.
- Gonçalves IC, Peters JG (2016) A new species of *Polyplocia* Lestage from Malaysia with comments on the genus (Ephemeroptera, Euthyplociidae, Euthyplociinae). Zootaxa 4184(3): 553–560. https://doi.org/10.11646/zootaxa.4184.3.9
- Jacobus LM, Macadam CR, Sartori M (2019) Mayflies (Ephemeroptera) and Their Contributions to Ecosystem Services. Insects 10(6): 170. https://doi.org/10.3390/insects10060170
- Kaltenbach T, Gattolliat JL (2020) *Labiobaetis* Novikova & Kluge in Borneo (Ephemeroptera, Baetidae). ZooKeys 914: 43–79. https://doi.org/10.3897/zookeys.914.47067
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. Journal of Molecular Evolution 16(2): 111–120. https://doi.org/10.1007/BF01731581
- Kluge NJ (2004) *The phylogenetic system of Ephemeroptera*. Kluwer Academic Publishers, Dordrecht-Netherlands, 456 pp. https://doi.org/10.1007/978-94-007-0872-3

- Kwanboon S, Sartori M, Boonsoong B (2021) Behningiidae and Potamanthidae (Insecta, Ephemeroptera) in Thailand. ZooKeys 1067: 57–82. https://doi.org/10.3897/zookeys.1067.72779
- Lestage JA (1921) Les Ephéméres indo-chinoises. Annales de la Société Entomologique de Belgique 61: 211–222.
- Lestage JA (1924) Faune entomologique de l'Indochine française. Les Éphémères de l'Indochine Française. Opuscules de l'Institut Scientifique de l'Indochine 3: 79–93.
- Miller DB, Bartlett S, Sartori M, Breinholt JW, Ogden TH (2018) Anchored phylogenomics of burrowing mayflies (Ephemeroptera) and the evolution of tusks. Systematic Entomology 43(4): 692–701. https://doi.org/10.1111/syen.12298
- Nguyen VV, Bae YJ (2003a) A new euthyplociid burrowing mayfly (Ephemeroptera: Euthyplociinae, Polymitarcyidae) from Vietnam. Korean Journal of Biological Sciences 7(3): 279–282. https://doi.org/10.1080/12265071.2003.9647716
- Nguyen VV, Bae YJ (2003b) Taxonomic review of the Vietnamese Neoephemeridae (Ephemeroptera) with description of *Potamanthellus unicutibius*, new species. The Pan-Pacific Entomologist 79: 230–236.
- Parnrong S, Buathong M, Sites RW (2002) New records of Behningiidae, Potamanthidae, and Prosopistomatidae (Ephemeroptera) from Thailand. Science Asia 28: 407–409.
- Sartori M, Derleth P, Gattolliat JL (2003) New data about the mayflies (Ephemeroptera) from Borneo. In: Gaino E (Ed.) Research Update on Ephemeroptera and Plecoptera, University of Perugia, Italy, Perugia, 403–406.
- Selvakumar C, Sivaramakrishnan KG, Janarthanan S (2015) A new record of *Potamanthellus caenoides* Ulmer 1939 (Ephemeroptera: Neoephemeridae) from the southern Western Ghats of India. Biodiversity Data Journal 3: e5021. https://doi.org/10.3897/BDJ.3.e5021
- Shorthouse DP (2010) SimpleMappr, an online tool to produce publication-quality point maps. [Retrieved from] https://www.simplemappr.net [Accessed April 30, 2023]
- Tamura K, Stecher G, Kumar S (2021) MEGA11: Molecular Evolutionary Genetics Analysis Version 11. Molecular Biology and Evolution 38(7): 3022–3027. https://doi.org/10.1093/molbev/msab120
- Ulmer G (1932) Bemerkungen über die seit 1920 neu aufgestellten Gattungen der Ephemeropteren. Entomologische Zeitung Stettin 93: 204–219.
- Ulmer G (1939) Eintagsfliegen (Ephemeropteren) von den Sunda-Inseln. Archiv für Hydrobiologie 16(Supplement): 443–692.
- Zheng X, Qiang X, Zhou C (2023) First nymph-imago association in *Polyploica* confirming the distribution of Euthyplociidae (Ephemeroptera) in China. Deutsche Entomologische Zeitschrift 70(1): 1–11. https://doi.org/10.3897/dez.70.96986