

Short Communication

# Synonymization of two, monotypic black-coral-commensal scale worm genera, *Antipathipolyeunoa* Pettibone, 1991 and *Parahololepidella* Pettibone, 1969 (Polynoidae, Aphroditiformia)

Brett C. Gonzalez<sup>10</sup>, Victor M. Conde-Vela<sup>10</sup>, Karen J. Osborn<sup>10</sup>

1 Smithsonian Institution, National Museum of Natural History, Department of Invertebrate Zoology, P.O. Box 37012, Washington D.C., USA Corresponding author: Brett C. Gonzalez (gonzalezb@si.edu)

#### Abstract

Parahololepidella Pettibone, 1969 is a polynoid genus commensal with the antipatharian genus *Tanacetipathes* Opresko, 2001. These scale worms are elongate with numerous segments and small elytra. To date, the only other known polynoid associated with *Tanacetipathes* is *Antipathipolyeunoa* Pettibone, 1991. By re-examining the holotype of *Antipathipolyeunoa*, we have identified several overlooked characters that no longer distinguish this genus from *Parahololepidella*. Based on the presence of chaetae on the tentacular segment and elytral irregularity on posterior segments, we propose synonymizing *Antipathipolyeunoa* with *Parahololepidella*.

Key words: Annelida, Antipatharia, black corals, polynoid, Tanacetipathes

## Introduction

Polynoidae Kinberg, 1856 scale worms are one of the most diverse groups of Annelida found in association with other organisms (Martin and Britayev 2018). Members of this hyperdiverse group are commonly found in association with cnidarians (e.g., Barnich et al. 2013), echinoderms (e.g., Sugiyama et al. 2020), decapods (e.g., Pettibone 1993), and even other annelid hosts (e.g., Britayev and Martin 2006). Notably, nearly 50% of known commensal polynoids are associated with black corals and octocorals (Martin and Britayev 1998; Buhl-Mortensen and Mortensen 2004). However, the limited knowledge of the taxonomy and biology of these scale worms restricts our understanding of their adaptations to their hosts (Molodtsova et al. 2016; Martin and Britayev 2018).

Most black-coral-commensal scale worms have elongate bodies with more than average numbers of segments and elytra. Despite these gross similarities, the highly specialized nature of commensal polynoids associated with antipatharian corals is reflected in the erection of separate polynoid genera for each different host (e.g., Pettibone 1969, 1991). In 1969, Pettibone revisited *Polyeunoa* McIntosh, 1885 and *Hololepidella* Willey, 1905—two similar and often confused genera of elongate polynoids (Pettibone 1969). As a result, *Polyeunoa* and *Hololepidella* were revised and three new genera were erected, including *Parahololepidella* Pettibone, 1969, which has been characterized by numerous



Academic editor: Christopher Glasby Received: 11 May 2023 Accepted: 14 August 2023 Published: 1 September 2023

ZooBank: https://zoobank. org/5A97B2F5-9348-4F95-AAC8-D7E1A78E713E

**Citation:** Gonzalez BC, Conde-Vela VM, Osborn KJ (2023) Synonymization of two, monotypic black-coralcommensal scale worm genera, *Antipathipolyeunoa* Pettibone, 1991 and *Parahololepidella* Pettibone, 1969 (Polynoidae, Aphroditiformia). ZooKeys 1178: 61–68. https://doi. org/10.3897/zookeys.1178.106101

**Copyright:** <sup>©</sup> Brett C. Gonzalez et al. This is an open access article distributed under the terms of the CC0 Public Domain Dedication. segments ( $\geq$ 140) and elytra, elytra irregularly arranged posteriorly with different numbers on right and left sides, tentacular segment (segment 1) with chaetae, neuropodia with a digitiform subacicular process, and slightly hooked neurochaetae thicker than notochaetae (Pettibone 1969; Fauchald 1977; Britayev et al. 2014). The only known species, *Parahololepidella greeffi* (Augener, 1918) from West Africa, was originally reported as a free-living species inhabiting mucous tubes incrusted with sand grains and shell fragments. Pettibone (1969) concluded that these tubes were likely associated with another commensal organism and not the worm in question. Britayev et al. (2014) redescribed *Parahololepidella greeffi* from the syntypes and newly collected topotypes from São Tomé and Príncipe (Gulf of Guinea), finding specimens living in association with the antipatharian *Tanacetipathes* cf. *spinescens* and not free-living as originally thought (Rullier 1964; Pettibone 1969). These newly collected worms were camouflaged along the main stems of the coral branches, agreeing with Pettibone's (1969) assessment that this species did not live in mucous tubes.

In 1991, Pettibone erected three monotypic genera commensal with antipatharian corals, Antipathipolyeunoa, Bayerpolynoe and Tottonpolynoe, and included a key to all related taxa (Pettibone 1991). Among these new genera, Antipathipolyeunoa Pettibone, 1991 was described from specimens removed from the stems of Tanacetipathes tanacetum (Pourtalès, 1880) [as Antipathes] in Barbados (Pettibone 1991). The only known species, Antipathipolyeunoa nuttingi Pettibone, 1991, was characterized by numerous segments and elytra, tentacular segment (segment 1) without chaetae, neuropodia with projecting subacicular process, and falcate neurochaetae stouter than notochaetae. Pettibone (1991) remarked that Antipathipolyeunoa closely resembled Polyeunoa (sensu Pettibone 1969), but differed based on the presence of a prominent subacicular process on the prechaetal acicular lobe of the neuropodium. Surprisingly, the included key and remarks both overlooked her previous work on Parahololepidella despite its striking resemblance to Antipathipolyeunoa, specifically, in the presence of a prominent subacicular process (Pettibone 1969, 1991). This omission could be attributed to the fact that, at that time, Parahololepidella greeffi was not known to be associated with black corals (Britayev et al. 2014).

When recently comparing the original diagnoses of *Parahololepidella* and *Antipathipolyeunoa*, it became evident that the only distinguishing feature between these two genera was the presence/absence of chaetae on segment 1. Pettibone (1969) indicated that the posterior elytral pattern may have "some irregularity" in *Parahololepidella*, but did not describe this condition further, and made no reference to this condition for *Antipathipolyeunoa*. Pettibone (1969) noted that the exact elytral arrangement was often omitted in elongate polynoids given their variability, but appeared constant for the anterior regions. The later redescription of *Parahololepidella greeffi* (see Britayev et al. 2014) described in detail the irregular and asymmetrical arrangement of elytra, sometimes occurs from segment 32, where a single dorsal cirrus and an elytron may occur on opposing sides of the same segment.

Upon re-examination of the holotype (USNM 80097) and the three paratypes (USNM 136587) of *Antipathipolyeunoa nuttingi*, several morphological features were observed that were missed during the original description. We found that all specimens of *A. nuttingi* have chaetae on the tentacular segment (segment 1), and elytral irregularities are present in the posterior region of all three paratypes. The lack of elytral variation in the holotype suggests that this condition may have been overlooked because it is the only specimen complete with dorsal cirri and elytra still attached.

Based on the presence of chaetae on the tentacular segment and the irregularities in the posterior segments found in the type material, we conclude that no significant differences exist between the monotypic genera *Parahololepidella* and *Antipathipolyeunoa*. Therefore, we propose the synonymy of *Antipathipolyeunoa* with *Parahololepidella* and accordingly include *Antipathipolyeunoa nuttingi* as a member of *Parahololepidella*; providing an updated systematic account for *Parahololepidella* as well as additional morphological details for *Parahololepidella nuttingi* comb. nov.

# **Systematics**

Suborder Aphroditiformia Levinsen, 1883 Family Polynoidae Kinberg, 1856 Subfamily Arctonoinae Hanley, 1989

## Parahololepidella Pettibone, 1969

- Parahololepidella Pettibone, 1969: 54 [type species: Hololepidella greeffi Augener, 1918, by original designation].—Britayev et al. 2014: 28 [diagnosis emended; type species redescribed using syntypes (ZHM 5692) and topotypes (NNMN 24481) due to the poor state of the original syntypes].
- Antipathipolyeunoa Pettibone, 1991: 715 [type species: Antipathipolyeunoa nuttingi Pettibone, 1991, by original designation].

**Remarks.** The type specimens of *Parahololepidella* (ZHM 5692) were examined and illustrated by Pettibone for her 1969 revision of *Hololepidella*, and subsequently re-examined by Britayev et al. (2014) when they redescribed *Parahololepidella greeffi* (NNMN 24481 and MNCN 16.01/13708). Only original notes and illustrations by Pettibone were examined herein.

Parahololepidella now includes two species commensal with the antipatharian genus *Tanacetipathes* Opresko, 2001. Both *Parahololepidella* species show cryptic coloration patterns and are found nestled along the stems of the coral branches.

## Parahololepidella nuttingi (Pettibone, 1991), comb. nov.

Fig. 1

Antipathipolyeunoa nuttingi Pettibone, 1991: 716-719, figs 1, 2.

**Material examined.** *Holotype.* **BARBADOS** • 1; Sta. 65, off Payne's Bay Church; 91 m; 1918; collector CC Nutting; Barbados-Antigua Expedition; on *Antipathes tanacetum* (now *Tanacetipathes*); USNM 80097. *Paratypes.* **VENEZUELA** • 3; Sta. 736, W of Tortuga Island; 10.95, -65.8667; 69–155 m; 22 July 1968; *R/V* Pillsbury; on *Antipathes tanacetum* (now *Tanacetipathes*); USNM 136587.



Figure 1. Parahololepidella nuttingi (Pettibone, 1991) comb. nov., holotype of Antipathipolyeunoa nuttingi USNM 80097 (A, D, F-J); paratype USNM 136587 (B, C, E) A, B whole specimens, dorsal view C dorsal view of middle segments showing variation in elytral distribution; white circles represent dorsal cirrophores and white triangles represent elytrophores D dorsolateral view of anterior with detail (inset) of left tentacular segment (segment 1) showing single chaeta E anterior end, dorsal view A, C-E specimens stained with Shirlastain A to observe morphology in detail F cirrigerous parapodia, ventral view, ventral cirri missing G chaetae from cirrigerous parapodium H notochaetae I distal end of supraacicular neurochaeta with tooth-like protuberance indicated with black arrow. Abbreviations: dCp, dorsal cirrophores; dSt, dorsal style; neC, neurochaetae; noac, notoacicula; noC, notochaetae; SbP, subacicular process. Scale bars: 1 mm (A-C); 0.5 mm (D, E); 0.2 mm (F); 50  $\mu$ m (G); 20  $\mu$ m (H, I); 5  $\mu$ m (J).

**Redescription (based on the holotype).** Body with numerous segments, >80 (Fig. 1A, B). Elytra numerous, >40 pairs. Paired elytra on segments 2, 4, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 26, 29, 32, 33, 35, then alternating dorsal cirri and elytra until the end (holotype; Fig. 1A). Elytral variation present in posterior third

of paratypes (see "Variation" below); elytron and dorsal cirrus may occur on same segment (Fig. 1C). Elytra oval; margins and surface smooth; cover dorsum anteriorly; dorsum exposed on middle and posterior segments (Fig. 1A). Elytrophores short; less pronounced than dorsal cirrophores. Dorsal tubercles inconspicuous. Dorsal cirrophores not extending beyond notopodia (Fig. 1F). Dorsal cirrostyles smooth, 5–6 times longer than parapodium.

Prostomium bilobed; anterior notch shallow (Fig. 1D, E). Cephalic peaks prominent, subtriangular, as continuous extension of the prostomium, equal width to median antenna ceratophore (Fig. 1D, E). Median antenna in anterior notch (Fig. 1D, E); ceratophore short, bulbous with smooth ceratostyle, roughly three times the width of the prostomium. Lateral antennae inserted subterminally; ceratophores distinct (Fig. 1D); styles shorter than palps. Palps short, stout, ~1.5 times the width of the prostomium (Fig. 1B, D, E). Two pairs of eyes; large, laterally positioned. Tentacular segment (segment 1) not visible dorsally (Fig. 1E). Dorsal tentacular cirri longer than ventral tentacular cirri. Tentaculophores lateral to prostomium, with single chaeta (Fig. 1D, inset); tentacular styles smooth. Facial tubercle oblong, with smooth rounded margins. Segment 2 (buccal segment) with first pair of elytra and biramous parapodia; nuchal fold absent. Buccal cirri longer than following ventral cirri. Dorsal cirri from segment 3.

Parapodia subbiramous. Notopodia reduced, with subconical lobe (Fig. 1F). Neuropodia broad; subconical prechaetal lobe longer than postchaetal lobe, with digitiform subacicular process (Fig. 1F, G). Noto- and neuroacicula penetrating epidermis on some segments (Fig. 1F). Notochaetae few (9–2) (Fig. 1F), decreasing posteriorly, less stout than neurochaetae (Fig. 1G, H). Neurochaetae few (12–6) (Fig. 1F, G), more numerous in middle segments; shafts smooth (Fig. 1G); faint spinous rows distally (Fig. 1I); tips falcate, 4–7 times longer than wide (Fig. 1G, I), occasionally with small, tooth-like protuberance (Fig. 1J). Ventral cirri from segment 3, smooth, longer than neuropodium in anterior segments, becoming subequal in length posteriorly. Pygidium rounded. Anus terminal. Anal cirri long, equal in length of last five segments. Nephridial papillae present from segment 6.

**Measurements.** Fixed holotype 21.5 mm long, 2.2 mm wide excluding chaetae, 83 segments. The three paratypes (USNM 136587) consist of one posteriorly incomplete specimen, two additional anterior ends, and several middle and posterior fragments. It was only possible to trace one of the two shorter anterior fragments to their respective remaining body fragments. Longest anterior paratype fragment, 24.5 mm long, 2 mm wide excluding chaetae, 85 segments. Reconstructed paratype, 25 mm long, 2.3 mm wide excluding chaetae, 102 segments. Shortest anterior paratype fragment, 5.7 mm long, 1.5 mm wide excluding chaetae, 25 segments.

**Variation.** Pigmentation is present in all specimens, present along the midline of the dorsum, with wider bands of pigment present in the paratypes (Fig. 1B). Additional pigmentation occurs on the cirrophores (Fig. 1B), but it is otherwise completely lacking (Fig. 1B, C). Of the two nearly complete paratypes, the ely-tral distribution patterns are as follows:

#### Paratype 1.

- R: 2, 4, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 26, 29, 32, 33, 35, 37, 38, 41, 43, 45, 47, 49, 52, 54, 56, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, 79, 81, 83, 85
- L: 2, 4, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 26, 29, 32, 34, 36, 38, 41, 43, 45, 47, 49, 51, 53, 55, 56, 58, 59, 60, 62, 63, 65, 67, 69, 71, 73, 75, 77, 79, 81, 83, 85

Paratype 2.

- R: 2, 4, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 26, 29, 32, 34, 36, 38, 40, 42, 44, 46, 47, 50, 52, 54, 56, 58, 60, 62, 64, 66, 86, 70, 72, 74, 76, 80, 82, 84, 86, 88, 90, 92, 94, 96, 98, 100, 102.
- L: 2, 4, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 26, 29, 32, 33, 35, 37, 39, 41, 43, 45, 47, 49, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 80, 82, 84, 86, 88, 90, 92, 94, 96, 98, 100, 102.

**Remarks.** The current diagnosis mostly agrees with that of Pettibone (1991) but differs on the presence of chaetae on the tentacular segment (for both holotype and paratypes), and for the irregular distribution pattern of elytra found posteriorly in the paratypes. The holotype only varies from the paratypes in the elytral variation and pigmentation.

Parahololepidella nuttingi comb. nov. is very similar to P. greeffi, but can be differentiated as follows. In P. nuttingi, the dorsal cirrophores are shorter than the notopodium in middle segments, whereas in P. greeffi, the dorsal cirrophores surpass the notopodium (see Pettibone (1991, fig. 1G) and (1969, fig. 4C), respectively). In P. nuttingi, the neurochaetae have falcate tips 4–7 times longer than wide, whereas in P. greeffi, the falcate tips are only 2–3 times longer than wide. And finally, in P. nuttingi, when present, the tooth-like protuberance on the falcate tips of the neurochaetae are small (easily overlooked), whereas in P. greeffi, the tooth-like structures are larger when present, giving them almost a bidentate appearance. Britayev et al. (2014) cautioned that the neurochaetal tooth-like structures in P. greeffi (ZMH 5692) were artifacts due to poor preservation and dehydration over time. However, given that similar structures were found in all specimens of P. nuttingi, we feel that this character is valid and illustrates the importance of detailed microscopical examination and reexamination.

# Acknowledgements

Together, we would like to thank the American Women's History Initiative Pool (AWHIP) for their generosity in funding this project.

# **Additional information**

## **Conflict of interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## **Ethical statement**

No ethical statement was reported.

## Funding

This project was funded by American Women's History Initiative Pool (AWHIP).

#### **Author contributions**

Conceptualization: BCG. Funding acquisition: KJO. Investigation: BCG. Resources: KJO. Supervision: KJO. Visualization: VMCV. Writing – original draft: BCG. Writing – review and editing: VMCV, KJO.

## **Author ORCIDs**

Brett C. Gonzalez https://orcid.org/0000-0001-6968-2677 Victor M. Conde-Vela https://orcid.org/0000-0002-3964-5426 Karen J. Osborn https://orcid.org/0000-0002-4226-9257

#### Data availability

All of the data that support the findings of this study are available in the main text.

# References

- Barnich R, Beuck L, Freiwald A (2013) Scale worms (Polychaeta: Aphroditiformia) associated with cold-water corals in the eastern Gulf of Mexico. Journal of the Marine Biological Association of the United Kingdom 93(8): 2129–2143. https://doi. org/10.1017/S002531541300088X
- Britayev TA, Martin D (2006) Scale-worms (Polychaeta, Polynoidae) associated with chaetopterid worms (Polychaeta, Chaetopteridae), with description of a new genus and species. Journal of Natural History 39(48): 4081–4099. https://doi.org/10.1080/00222930600556229
- Britayev TA, Gil J, Altuna Á, Calvo M, Martín D (2014) New symbiotic associations involving polynoids (Polychaeta, Polynoidae) from Atlantic waters, with redescriptions of *Parahololepidella greeffi* (Augener, 1918) and *Gorgoniapolynoe caeciliae* (Fauvel, 1913). Memoirs of the Museum of Victoria 71: 27–43. https://doi.org/10.24199/j. mmv.2014.71.04
- Buhl-Mortensen L, Mortensen PB (2004) Symbiosis in deep-water corals. Symbiosis 37: 33–61.
- Fauchald K (1977) The polychaete worms. Definitions and keys to the orders, families and genera. Natural History Museum of Los Angeles City, Science Series 12: 1–188.
- Martin D, Britayev TA (1998) Symbiotic Polychaetes: Review of Known Species. Oceanography and Marine Biology: an Annual Review 36. CRC Press, London, 217–340. https://doi.org/10.1201/b12646
- Martin D, Britayev TA (2018) Symbiotic polychaetes revisted: an update of the known species and relationships (1998–2017). In: Hawkins SJ, Evans AJ, Dale AC, Firth LB, Smith IP (Eds) Oceanography and Marine Biology: An Annual Review. Taylor & Francis, 371–448. https://doi.org/10.1201/9780429454455
- Molodtsova TN, Britayev TA, Martin D (2016) Cnidarians and their polychaete symbionts. In: Goffredo S, Dubinsky Z (Eds) The Cnidaria, Past, Present and Future: The World of Medusa and Her Sisters. Springer International Publishing, Cham, 387–413. https://doi.org/10.1007/978-3-319-31305-4\_25
- Pettibone MH (1969) The genera *Polyeunoa* McIntosh, *Hololepidella* Willey, and three new genera (Polychaeta, Polynoidae). Proceedings of the Biological Society of Washington 82: 43–62.
- Pettibone MH (1991) Polynoid polychaetes commensal with antipatharian corals. Proceedings of the Biological Society of Washington 104: 714–726.
- Pettibone MH (1993) Scaled polychaetes (Polynoidae) associated with ophiuroids and other invertebrates and review of species referred to *Malmgrenia* McIntosh and replaced by *Malmgreniella* Hartman, with descriptions of new taxa. Proceedings of the Biological Society of Washington 538: 1–92. https://doi.org/10.5479/ si.00810282.538

- Rullier F (1964) Résultats scientifiques des campagnes de la *Calypso* Campagne de la *Calypso* aux îles du Cap Vert (1959) (suite): 5. Annélides polychètes. Annales de l'Institut Océanographique 41: 113–218.
- Sugiyama T, Jimi N, Goto R (2020) Widening the host range of the ectosymbiotic scaleworm *Asterophilia culcitae* (Annelida: Polynoidae) to three enchinoderm classes, with data on its body color variation. Plankton & Benthos Research 15(4): 289–295. https://doi.org/10.3800/pbr.15.289